Review

Recent Updates in Redox Regulation and Free Radical Scavenging Effects by Herbal Products in Experimental Models of Parkinson’s Disease

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Abstract: Parkinson’s disease (PD) is a complex multifactorial disease marked by extensive neuropathology in the brain with selective yet prominent and progressive loss of mid-brain dopaminergic neurons. The etiological factors involved in the development of PD are still elusive, but oxidative stress arising when reactive oxygen species (ROS) exceed amounts required for normal redox signaling is considered one of the major factors. ROS cause oxidative damage to proteins, lipids, and DNA and are one of the most prominent factors related to neurodegeneration. Pre-clinical and clinical studies clearly demonstrate the effectiveness of oxidative stress in the pathogenesis of PD. Therefore, regulation of redox signaling and inhibiting excess ROS would contribute greatly not only to extend longevity but also to ameliorate the progression of dopaminergic cell death seen in patients with PD. Several herbal products are beneficial for maintaining nerve cell function and for treating various neurodegenerative disorders by reducing oxidative stress. Here, we summarize the recent knowledge concerning promising herbs that have shown significant beneficial effects based on regulation of redox status and ROS inhibition in toxin-induced PD models.

Keywords: Parkinson’s disease; oxidative stress; redox signaling; antioxidant herbs; reactive oxygen species; 6-hydroxydopamine; substantia nigra
1. Introduction

Parkinson’s disease (PD) is the second most common neurodegenerative disorder of the central nervous system after Alzheimer’s disease (AD). It has been estimated that 1% of individuals >65 years are diagnosed with this disorder, and that this figure may double by the year 2030. PD might be a serious socioeconomic burden in a future aging society [1–3]. PD particularly affects a region of the basal ganglia called the substantia nigra (SN), which produces dopamine (DA). DA acts as a messenger between the SN and another area of the brain called the corpus striatum. Together, they coordinate smooth flowing muscle movement. Inadequate DA levels result in specific neurological symptoms such as tremors, difficulty walking, rigidity, slowed movements, and a decreased ability to make spontaneous movements, which characterize PD. In advanced stages, mental impairment has also been observed. There is a general agreement that PD may represent the final outcome of interactions among multiple factors, including genetic susceptibility and exposure to environmental toxins [4–6].

A great deal has been learned about PD, yet a cure has not been found. Current initial therapy includes DA replacement strategies with L-dopa, dopaminergic agonists, monoamine oxidase B inhibitors, and drug combinations that provide dramatic improvement and have become the accepted way to control PD symptoms. Other medications include catechol-o-methyltransferase, anticholinergic agents, and amantadine [7], but one of the problems when using these therapies is that they lose effectiveness over time. Other problems are the side effects of nausea, vomiting, headache, fatigue, fainting, increased thirst, and tremors. Thus, new approaches for handling PD are clearly needed.

2. Redox Regulation of Neuronal Survival in PD

Research disciplines focusing on oxidative stress have increased our knowledge of importance of the cell’s redox status and the recognition of oxidative stress as a process with implications for many pathophysiological states. Oxidative stress may be defined as an imbalance between oxidant-producing systems and antioxidant mechanisms (redox balance), which results in excess formation of reactive oxygen species (ROS) [8]. The delicate balance between ROS generation and elimination is maintained by many complex mechanisms, and a dysfunction in any of these mechanisms can lead to alterations in cellular redox status. An increase in ROS production or a decrease in ROS-scavenging capacity due to exogenous stimuli or endogenous metabolic alterations can disrupt redox homeostasis, leading to an overall increase in intracellular ROS levels or oxidative stress [9]. Previous studies have suggested that the regulation of signaling pathways by the redox system relies mostly on direct oxidative modifications of redox-sensitive signaling proteins. Redox regulation of neuronal survival at the transcription level, which includes alterations in transcription factors such as nuclear factor-κB (NF-κB), activating protein-1 (AP-1), nuclear factor erythroid 2 (Nrf2), and hypoxia inducible factor (HIF) may affect overall oxidative status [10]. Furthermore, redox regulation of neuronal survival at the signal-transduction level influences the mitogen-activated protein kinase and PI3K/Akt pathways, as well as several redox sensitive apoptotic effectors such as caspases, Bel-2, and cytochrome c, and their functions can be significantly affected by cellular ROS [11–15]. The consequence of oxidative stress leading to neuronal survival or death is likely dependent on the integration of these redox-sensitive signals.
Oxidative stress has been consistently linked to ageing related diseases and has emerged as a causative factor in the pathology of several neurodegenerative disorders including PD, AD, and amyotrophic lateral sclerosis. Neurodegenerative diseases are characterized by progressive dysfunction and death of neurons. A steady-state balance exists between pro-oxidants and antioxidants under normal conditions. However, when the rate of free radical generation exceeds the capacity of antioxidant defenses, oxidative stress ensues with consequential severe damage to cellular machinery. Oxidative stress is associated with mitochondrial and endoplasmic reticulum dysfunction, inducing apoptosis and protein misfolding in neurons. Decreased activities of antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione (GPx), and glutathione peroxidase in neurodegenerative states signify the role of reduced antioxidant potential during neurodegeneration. Oxidative stress results from a number of endogenous sources, including the production of oxygen-free radicals by mitochondrial oxidative phosphorylation, which is the primary source of high-energy compounds in the cell. Mitochondria are a significant source of ROS, as they are the major consumers of molecular oxygen in cells [16,17]. Mitochondrial dysfunction leads to reduced ATP production, increased oxidative stress, altered mitochondrial morphology, impaired calcium buffering, damage to mitochondrial DNA, and alterations in mitochondrial fission and fusion, eventually leading to cell death [18]. Mitochondrial complex I alterations are a major source of ROS generation in patients with PD. An inhibited mitochondrial complex hampers the mitochondrial respiratory chain, which causes incomplete oxygen reduction, thereby generating reactive species including deleterious O$_2^-$, which is further converted to NO$_3^-$ and, finally, to ´OH through the Fenton reaction [19]. Thus, dysfunctional mitochondria are the primary intracellular ROS source contributing to oxidative stress-mediated neurodegeneration in PD models [20–22].

3. Free Radicals Scavengers and PD

Free radicals are generated continuously via normal physiological events such as respiration and some cell-mediated immune functions. ROS molecules are simply oxygen-containing molecules that are highly reactive. They can be divided into free-radical ROS, which have unpaired electrons in their outer orbits (e.g., superoxide and hydroxyl radicals) and nonradical ROS, which lack unpaired electrons, but are chemically reactive and can be converted to free-radical ROS (e.g., hydrogen peroxide). The generation of large amounts of these species results in attacks on intracellular antioxidant defense causing activation of lipid peroxidation, protein modification, and oxidative DNA damage [23–27]. These reactive species and oxidative stress are particularly involved in neurodegenerative diseases, as the brain uses 20% of the inspired oxygen and 90% of the consumed oxygen to produce energy during oxidative phosphorylation. Neuronal cells in the brain are particularly sensitive and vulnerable to oxidative damage because of their high metabolic activity, low antioxidant capacity, and non-replicative nature [28].

The idea that excessive numbers of free radicals play an important role in the complex etiology of PD has spread in the last few years. Post-mortem investigations have revealed evidence of lipid peroxidation and oxidative damage to brain protein DNA in patients suffering from PD [29]. The increase in oxidative stress is attributed to a less active mitochondrial complex I among other factors. The role of free radicals and oxidative stress in PD has been extensively studied, and the results...
obtained make a compelling argument for the concept that oxidative stress occurs in the brains of patients with PD [30–34]. To a large extent, animal models of PD, in which the neurotoxins 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and 6-hydroxydopamine (6-OHDA) are administered, confirm these findings [35,36]. Furthermore, substantia nigra samples from PD brains also have reduced levels of antioxidants and antioxidant enzymes [27,37–40]. These changes increase the balance in favor of a pro-oxidant environment, which increases oxidative damage. Because ROS accumulation promotes excess or premature neuronal death leading to neurodegenerative diseases, the use of free radical scavengers or antioxidants to prevent neurodegeneration has been established for decades. This treatment involves the use of both supplementations with natural ROS scavengers as well as treatment with exogenous antioxidants. Several radical scavenger antioxidants show neuroprotective effects in toxin-induced PD models. A proposition that is widely accepted is that supplementing antioxidants reduces the risk of suffering from PD or delays its progression [41]. Molecules such as cysteine, melatonin, resveratrol, and ebselen are well demonstrated in experimental models to be neuroprotective based on their antioxidant properties [42–45]. Additionally, regular consumption of diets rich in antioxidants such as those found in fruits and vegetables may reduce the risk of developing neurodegenerative diseases such as PD and AD [46]. Antioxidants are capable of transforming ROS into stable and harmless compounds or by scavenging based on redox mechanism. Antioxidants keep our cells clean by combining with and eliminating very dangerous reactive molecules that can cause random changes in proteins, resulting in degeneration of dopaminergic (DAergic) neurons seen in PD [47,48].

4. Oxidative Stress and Toxin-Induced PD Models

Intracellular redox homeostasis is disturbed by oxidative stress, which manifests itself through dysregulation in the balance between systems that produce oxidant agents and the antioxidant defense mechanisms (the redox balance). Experimental models to produce prototypic oxidative stress and selective neuronal death and induction with selected neurotoxins have been utilized to investigate this mechanism. Neurotoxins have remained the most popular tools to obtain greater insights into oxidative stress and redox signaling mechanisms. Among the neurotoxins used to induce DAergic neurodegeneration, 6-OHDA, MPTP, and, more recently, paraquat and rotenone have received the most attention. Presumably, all of these toxins provoke ROS formation. Rotenone and MPTP are similar in their ability to potently inhibit mitochondrial complex I, although they display significant differences, including their ease of use in animals.

6-OHDA enters both DAergic and noradrenergic neurons and inflicts damage to the catecholaminergic pathways in both the peripheral and central nervous systems. 6-hydroxydopamine, the first animal model of PD associated with substantia nigra pars compacta (SNpc) DAergic neuronal death was introduced more than 30 years ago [49]. It is well accepted that 6-OHDA destroys catecholaminergic structures thorough a combined effect on ROS and quinones [50]. Once dissolved in an aerobic and alkaline milieu, 6-OHDA readily oxidizes yielding hydrogen peroxide and p-quinone. Further information regarding 6-OHDA is reviewed in Przedborski and Ischiropoulos [20].

The human Parkinsonian neurotoxin MPTP has been used in a variety of animals to model PD from nonhuman primates to invertebrates such as worms [51,52]. Neuropathological data in both humans
and monkeys indicate that MPTP causes damage to the nigrostriatal DAergic pathway identical to that seen in PD [53,54]. MPTP causes DAergic neurodegeneration by generating free radicals and leading to oxidative stress as shown by alterations in the states of antioxidant enzymes and molecules [55–57]. MPTP per se is nontoxic to neurons, but the enzyme monoamine oxidase B converts MPTP to 1-methyl-4-phenylpyridinium (MPP+) in the brain, which enters DA terminals via DA uptake sites. Within the DA terminals, MPP+ blocks mitochondrial complex I and causes ATP depletion. This is thought to be the main cause of MPTP-induced terminal degeneration. Additionally, MPP+ produces oxidative stress by generating ROS after blocking mitochondrial complex I and DA oxidation, which may participate in MPTP-induced DAergic toxicity [58–60].

Epidemiological studies suggest that exposure to pesticides may be a risk factor for PD. One of the most important pesticides used widely around the world is rotenone. Due to its toxic effects, which resemble PD, natural rotenone represents one of the most recently used approaches in toxic animal models of PD. Similar to MPTP, rotenone is highly lipophilic and readily gains access to all organs including the brain. Rotenone freely crosses all cellular membranes in the brain and accumulates in subcellular organelles such as mitochondria to inhibit mitochondrial complex I. This inhibition has several potentially damaging consequences, including increased formation of ROS, which cause oxidative damage within the cell. Oxidative damage, rather than a bioenergetic defect, is also seen in an *in vivo* rotenone model [61,62].

The potent herbicide paraquat (N,N-dimethyl-4-4-bipiridinium) is another prototypic toxin that exerts deleterious effects through oxidative stress. Paraquat toxicity appears to be mediated by the formation of superoxide radicals [63]. Furthermore, paraquat toxicity is also mediated by redox cycling with cellular diaphorase such as nitric oxide synthase yielding ROS [63]. Paraquat exhibits a striking structural similarity to the MPTP toxic metabolite MPP+. The actual reduction-oxidation cycling reaction of paraquat has been reviewed by Przedborski and Ischiropoulos [20].

Along with the four most popular toxic models of PD discussed above, several other toxins such as manganese (Mn), kainite, methamphetamine, and isoquinoline are believed to damage DAergic neurons based on oxidative stress mechanisms by producing ROS and disturbing overall redox status [20]. However, each model has its own advantages and shortcomings. Several natural antioxidant products act as neuroprotective agents in PD models by targeting oxidative stress and redox regulation mechanisms. Over 4,000 species of plant-derived antioxidants have protective effects against the neurodegeneration induced by elevated ROS levels [64]. Active constituents such as green tea polyphenols, resveratrol, uric acid, curcumin, quercetin, CoQ10, creatinine and several traditional herbal medicines have proven protective against toxin-induced models of PD [65–70]. The use of several neurotoxins as experimental models to reproduce many of the clinical hallmarks of PD both *in vitro* and *in vivo* has allowed the molecular mechanisms of the disease to be investigated further. Common in all types of model is the oxidative stress mechanism leading to a cascade of events and further neuronal damage. Research on these substances may become more widespread if putative studies continue to deliver promising results.
5. Herbal Products and Neurodegenerative Diseases

Synthetic drugs cause undesirable adverse effects, whereas natural products are generally considered safe and effective [65,66]. Herbal products have been used throughout history and within every culture to prevent and treat various diseases. Herbal medicines are becoming popular to improve quality of life with either no or limited side effects. The concept of developing antioxidant agents from herbs is much older, yet increasing importance is being given to herbal products that include investigating active constituents of plants and phytomedicines with a view to discover new antioxidant potential therapeutic compounds for treating various neurodegenerative diseases [67]. Although several agents such as vitamins with antioxidant properties have been proved to be beneficial in neurodegenerative diseases, the ability of these vitamins to act as pro-oxidants by inducing oxidative damage and oxidative stress was also reported. Antioxidant vitamins such as tocopherols, ascorbate, retinol and folic acid have been shown to have pro-oxidant effects at higher doses or under certain conditions [68,69]. In a recent study, repetitive administration of vitamin C injection induces dopaminergic neurotoxicity through generation of oxidative stress, and that this toxicity is related to the decline of GSH in both the SNpc and striatum [70]. Further a number of clinical studies on vitamins and antioxidant supplements have shown to have limited or no significant effect on health [71,72]. Majority of these studies and their conclusions are often drawn from experiments conducted with single antioxidant supplement. On the other hand natural herbs are a mixture of antioxidants and thus may work with different kinetics and dynamics. Plant materials including vegetables, fruits, roots, leaves, barks, oilseeds, grains and spices have been screened and used as an alternative therapeutics in preventing various forms of neuronal cell loss, including the nigrostriatal degeneration seen in PD based on their antioxidant properties [73–75].

6. Antioxidant Herbs in Toxin-Induced PD Models

Nerve cell death from oxidative stress has been implicated in a variety of pathologies, including PD. Thus, it is reasonable to use antioxidants to prevent or halt the progression of the disease caused by oxidative stress. In the following sections, the available recent literature on the therapeutic benefits of natural herbs, the utilized parts, the type of extract, and doses used for preventing and/or treating of PD with respect to manipulating ROS levels by scavenging free radicals and targeting redox-sensitive signaling molecules at the signal-transduction, transcription, or death-execution levels in toxin-induced PD models both in vitro and in vivo are discussed.

6.1. Chrysanthemum indicum

*C. indicum* Linn is an herb that belongs to the family Asteraceae and is distributed widely in China, Korea, and other Asian countries. *C. indicum* has been traditionally used since ancient times as a folk and Oriental medicine with immense ethnopharmacological benefits. The tea prepared from the *C. indicum* flower is a popular drink in China [76]. *C. indicum* has been used to treat inflammation, headache, ulcerative colitis, vertigo, eye irritation, hypertension, and respiratory diseases [77–80]. But, attention is particularly focused on its neuropharmacological actions, including its memory enhancing activity. The effect of a *C. indicum* ethanol extract against MPP⁺-induced damage in SH-SY5Y cells
was evaluated by our group. The results indicated that *C. indicum* at 1, 10, and 100 µg inhibited cell loss, decreased ROS production, regulated the Bax/Bcl-2 ratio, and inhibited poly (ADP-ribose) polymerase (PARP) proteolysis in MPP\(^+\)-induced SH-SY5Y cells [80]. Our investigation scientifically supported the long history and safe use of *C. indicum* as an important functional food with potential benefits for ameliorating the neurodegeneration seen in PD. In another study, the antioxidant properties of *C. indicum* ethanol extract were characterized *in vitro* using different methods, including 1, 1-diphenyl-2-picrylhydrazyl, 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt, hydroxyl, superoxide and nitrite radical scavenging, reducing (Fe\(^3+\) to Fe\(^2+\)) power, inhibition of linoleic acid oxidation, as well as the prevention of free radical-induced DNA damage [81]. The authors suggested that the *C. indicum* extract was a potent source of natural antioxidants. The protective effects of *C. indicum* might be due to the compounds present in the extract such as flavonoids, terpenoids, and polyphenols, which display various biological activities including antioxidant effects [82,83].

6.2. *Gastrodia elata*

*G. elata* Blume belongs to the family Orchidaceae and has been used as a traditional herbal medicine with numerous therapeutic applications for headaches, dizziness, vertigo, and convulsive illnesses such as epilepsy and tetanus since ancient times [84]. This plant is distributed widely in Nepal, Bhutan, India, Japan, Korea, Siberia, and Taiwan as well as on mainland China. It grows at elevations of 400–3,200 meters at the edge of forests. *G. elata* is effectively used as an anticonvulsant, analgesic, sedative for vertigo, and in patients with general paralysis, epilepsy, and tetanus [85]. *G. elata* inhibits glutamate-induced apoptosis in neurons [86], inhibits neuronal damage in kainite-induced seizure [87], inhibits MPP\(^+\)-induced cytotoxicity in SH-SY5Y cells [88], and protects against neuronal cell damage after transient global brain ischemia [89].

In a recent study we investigated the *G. elata* extract and its major bioactive components on MPP\(^+\)-treated MN9D DAergic cells, a classic *in vitro* PD model. The results indicated that vanillyl alcohol, one of the major active bio compounds in *G. elata*, effectively inhibited cytotoxicity and improved cell viability in MPP\(^+\)-induced MN9D DAergic cells. Vanillyl alcohol attenuated the elevation in ROS levels, decreased the Bax/Bcl-2 ratio, and PARP proteolysis. Vanillyl alcohol (1, 10, and 20 µM) protected DAergic MN9D cells against MPP\(^+\)-induced apoptosis by relieving oxidative stress and modulating the apoptotic process [90]. In another study, Shin *et al.* [91] investigated the effects of *G. elata* against methamphetamine (MA)-induced striatal DAergic toxicity in mice. Treatment with MA (7.5 mg/kg, *i.p.* × 4) resulted in significant decreases in behavioral activity, DA level, tyrosine hydroxylase (TH) activity, and TH protein expression. Additionally, MA treatment resulted in significant increases in lipid peroxidation, protein oxidation, and ROS formation. The authors showed that *G. elata* (500 or 1,000 mg/kg, *p.o.*), significantly attenuated MA-induced behavioral and DAergic impairment, and oxidative stress in a dose-dependent manner thereby exhibiting anti-DAergic effects in response to the MA insult *via*, at least in part, inhibiting oxidative stress in the striatum of the mice [91].
6.3. Ginseng

Ginseng, the root of Panax ginseng C.A. Meyer, belongs to the family Araliaceae and is a valuable herb in traditional Chinese medicine. Ginseng has been utilized for over 2,000 years with the belief that it is a general tonic to promote vitality, health, and longevity. P. ginseng roots may be taken orally for diverse benefits, such as an aphrodisiac, a stimulant, to treat type II diabetes, or for sexual dysfunction in men. Ginseng has been included in small doses in energy drinks and teas [92]. It may be found in cosmetic preparations as well, but has not been shown to be clinically effective. A ginseng water extract has been used to treat many kinds of diseases including anemia, insomnia, ischemia, diabetes mellitus, and gastritis [93–96]. A ginseng extract alleviated scopolamine-induced learning disability and improved spatial working memory in mice [97]. The major effect of ginseng extract resides in its ability to scavenge hydroxyl [98,99] and superoxide radicals [100]. The neurotrophic and neuroprotective effects of ginseng in cell lines and animals have recently been reviewed [96]. Hu et al. [101] investigated the protective effects of a water extract of ginseng against MPP⁺-induced cytotoxicity in SH-SY5Y human neuroblastoma cells. The authors suggested that overproduction of ROS, elevated Bax/Bcl-2 ratio, release of cytochrome c and activation of caspase-3 expression in MPP⁺-treated SH-SY5Y cells were significantly ameliorated by 0.01, 0.1, and 0.2 mg/mL of a ginseng water extract. An in-depth study revealed that the possible mechanism for the neurotoxic protective effect in SH-SY5Y cells was due to its strong antioxidant properties for suppressing excessive ROS generation [101]. Luo et al. [102] studied the protective effect of panaxatriol saponins extracted from Panax notoginseng against MPTP-induced neurotoxicity in vivo. These authors detected an effect of P. notoginseng on MPTP-induced behavioral impairment, and other oxidative parameters in the anatomical region of the substantia nigra pars compacta (SNpc) region of mice brain tissues. They concluded that the protective effects of P. notoginseng (100 mg/kg, twice daily for 7 days) might partly be due to its ability to enhance antioxidant capacity and inhibit mitochondrial-mediated apoptosis [102].

6.4. Polygala tenuifolia

Polygalae radix (PRE) is the root of P. tenuifolia Willd and belongs to the Polygalaceae family. P. tenuifolia is widely distributed in China and other Asian countries and has been commonly used to treat central nervous system disorders in traditional Korean and Chinese medicine. P. tenuifolia is one of the most prescribed herbal remedies for treating various cognitive symptoms associated with aging, senile dementia, and PD [103–105]. In a study by Choi et al. [106] PRE was investigated for its protective effects and possible mechanisms of action against toxin-induced neuronal death in a mouse model of PD. PC12 cells were damaged by 6-OHDA in vitro, and ROS, nitric oxide production and caspase-3 activation were assayed. The authors also evaluated the protective effects of PRE on MPP⁺-induced neurotoxicity in rat primary DAergic neurons and in an in vivo mouse model of PD. The results indicated that PRE significantly inhibits 6-OHDA-induced cell damage at doses of 0.05–1 µg/mL with a maximal effect at 0.1 µg/mL. Caspase-3 activity, ROS and NO production were alleviated at 0.1 µg/mL. PRE (0.1 µg/mL) protected mesencephalic DAergic neurons against MPP⁺-induced toxicity and protected DAergic neurons and fibers from MPTP-induced toxicity in the SNpc and striatum in vivo.
when administered orally at 100 mg/kg/day for 3 days. It was concluded that PRE has protective effects on DAergic neurons via its antioxidant and antiapoptotic effects [106]. In another study, the active constituent tenuigenin was studied for its neuroprotective effects in 6-OHDA-induced cytotoxicity in SH-SY5Y cells [107]. These authors indicated that a 10 µM dose of tenuigenin significantly promotes cell viability and reduces cell death. Tenuigenin also protects the mitochondrial membrane potential against 6-OHDA damage and significantly increases glutathione and SOD expression. The authors suggested that the neuroprotection to DAergic neurons provided by tenuigenin from 6-OHDA-induced damage might partly involve its strong antioxidative effects.

6.5. Bacopa monnieri

*B. monnieri* Linn. is in the family Plantaginaceae and a common herb that grows throughout India, Nepal, Sri Lanka, China, Taiwan, Vietnam, as well as in Florida, Hawaii and other southern states of the US where it grows in damp conditions by ponds or in bogs. *B. monnieri* has been used in Indian Ayurvedic traditional medicine for almost 3,000 years and is classified as one of the most important herbs used to improve memory and cognition as well as a potent nerve tonic [108,109]. *B. monnieri* is well tolerated without side effects [109], and the LD50 in rats is as high as 2.7 g/kg when administrated orally [110]. In a study conducted by Singh *et al.* [111], *B. monnieri* protected a DAergic SK-N-SH cell line against MPP⁺- and paraquat-induced toxicity in various survival assays. *B. monnieri* pretreatment (10 mg/mL) significantly attenuated the generation of intracellular ROS and decreased mitochondrial superoxide levels. Furthermore, *B. monnieri* treatment activated the Nrf2 pathway by modulating Keap1 expression, thereby upregulating endogenous GSH synthesis. The authors concluded that *B. monnieri* might be useful in age-related neurodegeneration including PD by preserving cellular redox homeostasis and mitochondrial activities [111]. In an earlier study Hosamani and Muralidhara [112], investigated a standardized *B. monnieri* powder against rotenone-induced oxidative stress and neurotoxicity in *Drosophila melanogaster* exposed to *B. monnieri* powder (0.05 and 0.1%) for 7 days in the diet. The results showed significant attenuation in the levels of endogenous oxidative markers such as malondialdehyde, hydroperoxide, and protein carbonyl content. Complete protection was offered by *B. monnieri* against rotenone (500 mM)-induced oxidative stress and further markedly inhibited DA depletion (head region, 33%; body region, 44%) in the flies. The authors hypothesized that *B. monnieri* may have the ability to mitigate rotenone-induced oxidative stress [112].

In much more recent studies, Shinomol *et al.* tested the hypothesis that a *B. monnieri* extract could offset neurotoxicant-induced oxidative dysfunction in the developing brain of rotenone and 3-nitropropionic acid mouse models [113–116]. They indicated that rotenone-induced (1.0 mg/kg b.w./day, i.p.) oxidative stress and cell death in vitro was attenuated significantly by pretreatment with *B. monnieri* (2, 4, and 6 µg) in DAergic N27 cell lines. In vivo studies revealed that *B. monnieri* (5 mg/kg b.w. once daily for 7 days, i.p.) normalized the levels of oxidative markers (malondialdehyde, ROS, and hydroperoxides) and restored depleted GSH levels in rotenone-induced (1 mg/kg b.w. /day for 7 days) mice. *B. monnieri* also effectively normalized protein carbonyl content in all brain regions, suggesting an ability to prevent protein oxidation. Furthermore, oxidative stress and mitochondrial dysfunction in DAergic (N27) cells and prepubertal mouse brain induced by 3-nitropropionic acid were significantly attenuated as evidenced by restoration of the ETC enzyme
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activities (NADH: ubiquinone oxidoreductase, NADH: cytochrome c reductase, succinate-ubiquinone oxidoreductase, and cytochrome c oxidase) as well as mitochondrial viability. The activities of antioxidant enzymes (SOD, GPx, glutathione reductase, and thioredoxin reductase), Na (+), K (+)-ATPase, and citric acid cycle enzymes in the striatum that were discernible among 3-NPA mice were restored significantly upon B. monnieri pretreatment. In addition, pretreatment with B. monnieri for 10 days followed by a 3-NPA challenge (75 mg/kg b.w./day, i.p.) completely prevented 3-NPA-induced oxidative dysfunction in the striatum and other brain regions. Elevated oxidative marker levels (malondialdehyde, ROS, hydroperoxide, and protein carbonyls) were predominantly abolished among mice given B. monnieri prophylaxis. The neuroprotective effects of B. monnieri may be wholly or in part related to its propensity to scavenge free radicals, maintain redox status, and upregulate antioxidant machinery in striatal mitochondria. The ability of a B. monnieri ethanol extract to enhance reduced glutathione and antioxidant defenses in brain regions has been hypothesized for its potent neuroprotective action [116].

6.6. **Hyoscyamus niger**

*H. niger* Linn, commonly known as Henbane in English and Prasikayavani in Sanskrit [117], belongs to the family Solanaceae and is widely distributed in Europe and Asia. This plant contains tropane alkaloids (hyoscyamine, hyoscine, and scopolamine) as well as flavonol glycosides (quercitin, rutin, and kaempferol), which are among the oldest drugs used in medicine [118]. In the traditional Indian medical system, Ayurveda, powdered seeds of *H. niger* are one of the four plant ingredients used for treating PD [119,120]. Senugupta *et al.* (2010) recently investigated the efficacy of *H. niger* for its antiparkinsonian effects in an MPTP model of PD in mice. Parkinsonian mice were treated twice daily with the extract (125–500 mg/kg, p.o.) for 2 days, and motor functions and striatal DA levels were assayed. The *H. niger* extract significantly attenuated motor disability and striatal DA loss in the MPTP-treated mice. The *H. niger* seed extract also significantly inhibited monoamine oxidase activity and attenuated MPP⁺-induced hydroxyl radical generation in isolated mitochondria. The authors reported that the *H. niger* extract showed antiparkinsonian effect by its ability to inhibit increased hydroxyl radicals generated in mitochondria [121].

6.7. **Hibiscus asper**

*H. asper*, in the family Malvaceae, is an important medicinal plant widely distributed throughout tropical Africa and Madagascar. A methanol extract of *H. asper* leaves possesses a wide spectrum of biological activities, including *in vitro* antioxidant activity, anxiolytic and antidepressant actions, as well as positive effects on spatial memory formation [122]. Hritcu *et al.* [123] evaluated the relationship between the antioxidant and antiapoptotic actions of a methanol extract of *H. asper* leaves and its neuroprotective properties in a 6-OHDA lesioned rat model of PD. The authors suggested that chronic administration of the methanol extract (50 and 100 mg/kg, i.p., daily for 7 days) significantly increases antioxidant enzyme activities (SOD, GPX and CAT), total GSH content, and reduces lipid peroxidation (MDA level) in rat temporal lobe homogenates, suggesting that antioxidant activity is the major mechanism for its potent neuroprotective effect against 6-OHDA-induced PD in a rat model [123]. In another study, a *H. asper* leaf methanol extract was evaluated against unilateral 6-OHDA lesions in
Wister rats. *H. asper* leaves at 50 and 100 mg/kg showed 2,4-dinitrophenyl-1-picryl hydrazyl (DPPH) radical scavenging activity and β-carotene bleaching inhibition activity [122].

6.8. *Melissa officinalis*

*M. officinalis* belongs to the Laminaceae family and is a perennial herb native to southern Europe and the Mediterranean region. Preparations derived from the aerial part of *M. officinalis* are often used in folk medicine to treat various symptoms from the common cold to cardiac dysfunction and ulcers [124]. Data have also supported a protective role for *M. officinalis* intake against AD [125]. Martins et al. [126] studied the protective effect of a *M. officinalis* aqueous extract against Mn-induced oxidative stress in chronically exposed mice. Mn is an essential element for biological systems; however, occupational exposure to high levels of this metal may lead to neurodegenerative disorders resembling PD. Mice were administered Mn (50 mg/kg/day) during the first 15 days followed by 100 mg/kg/day for an additional 75 days in drinking water. The experiment was continued with Mn and/or *M. officinalis* treatment for an additional 3 months in drinking water. Mn-treated mice showed a significant increase in TBARS levels (a marker of oxidative stress) in both the hippocampus and striatum. Decreases in total thiol content in the hippocampus and a significant increase in antioxidant enzyme activity (SOD and CAT) in the hippocampus, striatum, cortex, and cerebellum was also observed. Co-treatment with a *M. officinalis* aqueous extract significantly inhibited antioxidant enzyme activity and attenuated the oxidative damage (TBARS and decreased total thiol levels). The authors suggested that the *M. officinalis* aqueous extract possesses potent antioxidative properties, validating its efficacy in attenuating Mn-induced oxidative stress in the mouse brain as seen in PD [126].

6.9. *Cassia obtusifolia*

*Cassia semen* (sickle pod) is the seed of the annual plant *C. obtusifolia* Linn in the Leguminosae family. It is widely cultivated in Korea and China and commonly drunk as a roasted tea and used as a medicinal food. *C. obtusifolia* has multiple therapeutic actions related to the prevention of dementia and ischemia [127]. Ju et al. [128] studied the protective effects of *C. obtusifolia* in *in vitro* and *in vivo* PD models. In PC12 cells, *C. obtusifolia* (0.1–10 μg/mL) attenuates cell damage induced by 6-OHDA (100 μM)-induced stress in the MTT assay and inhibits the overproduction of ROS, GSH depletion, mitochondrial membrane depolarization, and caspase-3 activation. In addition, *C. obtusifolia* shows radical scavenging activity in DPPH and 2,2-azinobis-(3-ethyl-benzthiazoline-6-sulphonic acid (ABTS) assays. The authors suggested that *C. obtusifolia* inhibits cell loss against 6-OHDA-induced DA neural toxicity via an antioxidant and antimitochondrial-mediated apoptosis mechanism in PC12 cells [128]. Furthermore, *C. obtusifolia* (0.1–1 μg/mL) extract protected DA cells in a mesencephalic DAergic culture against 6-OHDA- (10 μM) and MPP (10 μM)-induced toxicity. An *in vivo* behavioral test (pole test), revealed that treatment of an MPTP-induced group (30 mg/kg, 5 days) with *C. obtusifolia* (50 mg/kg, 15 days) decreased time to turn and time for locomotion activity, which were longer in the MPTP-alone treated group and significantly protected DA neuronal degeneration in the substantia nigra and striatum in an MPTP-induced mouse model of PD [128]. In another study, an ethanol extract of *C. obtusifolia* seeds was investigated in primary mouse hippocampal cultures exposed to three models of cell death implicated in neurodegeneration such as acute and endpoint
studies related to excitotoxicity induced by NMDA, a model of mitochondrial dysfunction induced by incubation with 3-NP, and amyloid-β-protein-induced cell death resulting from incubation with medium from β-amyloid-producing N2a cells. *C. obtusifolia* at 1 and 10 µg/mL was neuroprotective against the mitochondrial toxin 3-NP (1 mM), and Aβ toxicity (8.2 pg/mL), particularly against mitochondrial dysfunction in hippocampal cultures, pointing to a role in the regulation and maintenance of cellular homeostasis and apoptosis, which are relevant for future therapeutic considerations in the treatment of neurodegenerative disorders [129].

6.10. *Croton celtidifolius*

*C. celtidifolius* Baill, in the family Euphorbiaceae, is a tree frequently found in the Atlantic forests of southern Brazil. The *C. celtidifolius* bark has several neurobiological activities including antioxidant and antiinflammatory properties [130,131]. Moreira *et al.* [132] demonstrated that the proantho-cyanidin-rich fraction (PRF) obtained from *C. celtidifolius* bark possesses a wide spectrum of psychopharmacological properties in rats, and that these effects could be attributed to the presence of various catechin and/or proanthocyanidin antioxidant compounds. The same group investigated the effects of *C. celtidifolius* PRF on the neurochemical and behavioral alterations induced by a single intranasal MPTP administration in rats. Pretreatment with PRF (10 mg/kg, i.p.) during 5 consecutive days prevented the inhibition of mitochondrial complex-I in the striatum and olfactory bulb, as well as decreased TH expression in the olfactory bulb and SN of rats infused with MPTP (1 mg/nostril). Moreover, PRF pretreatment attenuated short-term social memory deficits, depressive-like behavior, and reduced locomotor activity observed at different periods after intranasal MPTP administration in rats. The authors indicated that the beneficial effects of PRF in MPTP-intoxicated rats might be due to the presence of various catechin and/or proanthocyanidin compounds, which have profound antioxidant actions [132].

6.11. *Gynostemma pentaphyllum*

*G. pentaphyllum*, a twisting vine orchid in the family Cucurbitaceae, is indigenous to the southern reaches of China, South Korea, and Japan. Gypenosides, which are saponins extracted from this plant, have various bioactivities such as hepatoprotection, antihyperlipidemia, and anticancer effects [133–136]. This plant is known for its powerful antioxidant and adaptogenic effects purported to increase longevity. Earlier reports indicated that *G. pentaphyllum* protects rat cortical cells against glutamate-induced oxidative injury by preventing GSH depletion and lipid peroxidation [137]. Wang *et al.* [138] explored the neuroprotective effects of *G. pentaphyllum* in an MPTP-induced mouse model of PD. Co-treatment with *G. pentaphyllum* (100, 200, and 400 mg/kg) after acute administration of MPTP (20 mg/kg at 2-h intervals) attenuated the decreased GSH content and reduced SOD activity in the substantia nigra. Furthermore, the loss of nigral DAergic neurons and motor dysfunction, which resulted in oxidative stress, was reversed significantly. The authors suggested that the neuroprotective effect of *G. pentaphyllum* in PD models may be attributed to increased antioxidation capacity [138]. In another study, oral administration of an herbal ethanol extract from *G. pentaphyllum* (10 and 30 mg/kg) starting on day 3 post-lesion with 6-OHDA for 28 days markedly ameliorated the reduction in TH-immunopositive neurons induced by 6-OHDA in the substantia nigra. *G. pentaphyllum* administration also recovered
the levels of DA, 3,4-dihydroxyphenylacetic acid, homovanillic acid, and norepinephrine in post-lesion striatal tissues. Four gypenoside derivatives such as gynosaponin TN-1, gynosaponin TN-2, gypenoside XLV, and gypenoside LXXIV were identified from *G. pentaphyllum*. The authors suggested that *G. pentaphyllum* might be helpful in the prevention of PD [139].

6.12. *Thuja orientalis*

*T. orientalis* Linn, an evergreen coniferous tree in the family Cupressaceae, is native to North America and eastern Asia. The leaves of *T. orientalis* are commonly used in traditional oriental medicine. They exert an antioxidant effect by scavenging free radicals and have a protective effect on DNA oxidation and red blood cell hemolysis [140]. Earlier reports suggested that the active principle factor, 15-methoxypinusolidic acid (15-MPA), a pinusolide derivative isolated from a *T. orientalis* methanol extract, has neuroprotective effects. 15-MPA protects against glutamate-induced neurotoxicity in primary cultured rat cortical cells by reducing nitric oxide overproduction, GSH depletion, and lipid peroxidation [141,142]. As *T. orientalis* and its compounds suppress oxidative stress, Ju *et al.* [143] investigated the protective effects of standardized *T. orientalis* leaves against 6-OHDA-induced neurotoxicity in SH-SY5Y cells. The results indicated that *T. orientalis* (1000 µg/mL) shows strong radical scavenging effects in 2,2-diphenyl-2-picrylhydrazyl and 2,2-azinobis-(3-ethylbenzthiazoline-6-sulphonic acid) assays. Furthermore, *T. orientalis* (10 µg/mL) reduces intracellular ROS production induced by 6-OHDA (200 µg/mL). Additionally, *T. orientalis* blocks the reduction in mitochondrial membrane potential, the release of cytochrome c, and caspase-3 activation. Moreover, *T. orientalis* decreases the phosphorylation of extracellular signal-regulated kinase, which has pro-apoptotic functions. The authors indicated that the protective effects of *T. orientalis* in SH-SY5Y cells induced by 6-OHDA might partly be through downregulation of oxidative stress.

6.13. *Paullinia cupana*

*P. cupana* Mart. var. Sorbilis is a Brazilian plant in the Sapindaceae family. The chemical composition of *P. cupana* seeds is quite complex and includes catechin, epicatechin, ent-epicatechin, procyanidins, and phenolic compounds [144]. Some of these components have nigrostriatal DAergic neuroprotective and antioxidant activities [138,145]. The presence of xanthine bases such as caffeine, theophylline, and theobromine, as well as purine alkaloids, which have been described as having antioxidant properties, is also well known [146–149]. The antioxidant activity of a *P. cupana* ethanol extract has been reported [150]. In a recent study de Oliveira *et al.* [151] investigated the effects of commercial powdered *P. cupana* seeds against rotenone-induced cytotoxicity in SH-SY5Y cells as an in vitro model of DAergic neuron degeneration. The *P. cupana* extract (0.312 and 0.625 mg/mL) significantly and dose-dependently increased viability of SH-SY5Y cells treated with rotenone (300 nM). Furthermore, the authors also measured lactate dehydrogenase (LDH) levels and analyzed nuclear integrity with Hoechst 33258 stain. LDH levels decreased significantly by adding 0.312 mg/mL of *P. cupana*, but, unexpectedly, no changes were observed at the higher concentration. Moreover, chromatin condensation and nuclear fragmentation decreased significantly by adding both extract concentrations. The authors suggested that the compounds with potential antioxidant activity present in
P. cupana might be responsible for such profound protective action against rotenone toxicity in SH-SY5Y cells [151].

6.14. Morus alba

Mulberry (M. alba L.) is a member of the Moraceae family and is native to northern China. This plant has been naturalized and is cultivated widely everywhere. Mulberry fruit is commonly eaten, often dried, or made into wine. In traditional oriental medicine, it has been used to treat premature grey hair, to nourish the blood, to treat constipation and diabetes and to generate body fluids, which generally means enhancing health and promoting longevity [152]. In addition, it also ameliorates inflammation-related hematological parameters in carrageenan-induced arthritic rats [153], promotes recovery from physical stress [154], has neuroprotective effects against cerebral ischemia [155], and has radical-scavenging properties [156]. Mulberry fruit contains not only high amounts of anthocyanins [157,158], a subset of the flavonoids that are important natural antioxidants [159,160], but also nonanthocyanin phenolics including rutin and quercetin that have multi-bioactive functions including neuroprotective effects [155,157,158,161]. Kim et al. [162] investigated the protective effects of mulberry fruit (ME) in a toxin-induced PD model. A 70% ethanol extract of ME (1, 10 and 100 µg/mL) significantly protected SH-SY5Y cells from neurotoxicity induced by 150 µM 6-OHDA in a dose-dependent manner. The protective effect of ME was mediated by its antioxidant and antiapoptotic effects, by regulating ROS and NO generation, Bcl-2 and Bax proteins, mitochondrial membrane depolarization, and caspase-3 activation. Furthermore, ME protected mesencephalic primary cells stressed with 6-OHDA (10 µM) or MPP⁺ (10 µM). In an \textit{in vivo} subacute mouse PD model induced by MPTP (30 mg/kg), ME at 500 mg/mL showed a preventative effect against PD-like symptoms (bradykinesia) in a behavioral test and prevented MPTP-induced DAnergic neuronal damage in an immunocytochemical analysis of the SNpc and ST. The authors concluded that the mulberry fruit extract significantly protected neurons against neurotoxins in \textit{in vitro} and \textit{in vivo} PD models through its antioxidant and anti-apoptotic effects [162]. The antioxidant activity of teas essences prepared from mulberry, \textit{Camellia sinensis} L., and \textit{Cudrania tricuspidata} (Carr.) Burea, have been examined using two antioxidant assays. Selected volatile chemicals identified in these plants were also tested for antioxidant activity. All extracts exhibited antioxidant activity with a clear dose response in aldehyde/carboxylic acid and malonaldehyde/gas chromatography assays [163].

6.15. Ginkgo biloba

EGb761 (Figure 5E) is a well-known plant extract obtained from leaves of the \textit{G. biloba}. This patented extract contains two main groups of active compounds [flavonoids (24%) and terpenoids (6%; known as ginkgolides A, B, C, M, J, and bilobalide)]. EGb761 is a potent antioxidant and free radical scavenger. A study evaluated the neuroprotective effect of EGb761 against oxidative stress induced by MPTP in C57BL/6J mice. MPTP administration resulted in a significant decrease in striatal DA levels and TH immunostaining in the striatum and SNpc, and mice that received EGb761 (40 mg/kg) had significantly attenuated MPTP-induced loss of striatal DA levels and TH immunostaining in the striatum and SNpc. The neuroprotective effect of EGb761 against MPTP neurotoxicity is associated with blockade of lipid peroxidation and reduction of superoxide radical production, both of which are
oxidative stress indices. The presence of elevated lipid peroxidation was detected 18 days after the last injection of MPTP. EGb761 blocked lipid peroxidation in the MPTP-treated groups. Furthermore, EGb761 regulates other antioxidant enzymes including GPx and glutathione reductase and shows potent inhibitory activity against SOD. Moreover, EGb761 improves MPTP-induced impairment of locomotion in a manner that correlates with enhanced striatal DA levels in behavioral experiments. These findings suggest that EGb761 attenuates MPTP-induced neurodegeneration of the nigrostriatal pathway in mice and that the neuroprotective effects are exerted via antioxidant or free radical scavenging action [164].

Several other natural products of plant origin such as a *Uncaria rhynchophylla* Hook extract [165], an isoflavone mixture from *Trifolium pretense* [166], salvianolic acid B isolated from *Salvia miltiorrhiza* [167], 6,7-di-O-glucopyranosyl-esculetin extracted from *Fraxinus sieboldiana* [168], baicalein, a flavonoid, obtained from *Scutellaria baicalensis* [169], a leaf extract of *Withania somnifera* [170], fustin, a flavonoid from *Rhus verniciflua* [171,172], *Acanthopanacis senticosus* [173], a rhizome of *Cyperus rotundus* [174], an ethanol extract of *Delphinium denudatum* [175], and a hydroalcohol extract of *Ilex paraguariensis* [176] have protective effects against PD in cellular and animal models based on their antioxidative abilities. A summary of the natural plant antioxidant products showing beneficial effects in *in vitro* and *in vivo* PD models is illustrated in Table 1.

Table 1. Summary of recent antioxidant herbal products reported to be neuroprotective in experimental models of Parkinson’s disease.

| Plant name/species            | Extract/constituent | Dose                | Toxin/tested subjects | Biological activity and targets                                           | Ref. |
|------------------------------|---------------------|---------------------|-----------------------|-------------------------------------------------------------------------|------|
| *Chrysanthemum indicum*      | Ethanol extract     | 1, 10 and 100 µg    | MPP<sup>+</sup>/SH-SY5Y cells | Regulation of Bax/Bcl-2 ratio & decreasing, inhibition of free radicals and oxidative stress | [80] |
| *Gastrodia elata*             | Vanillyl alcohol    | 1, 10 and 20 µM     | MPP<sup>+</sup>/MN9D cells | Decrease oxidative stress                                               | [90] |
|                              | Methanol extract    | 500 or 1,000 mg/kg  | Methamphetamine mine/C57BL/6J mice | Decrease oxidative stress                                               | [91] |
| *Ginseng*                    | Aqueous extract     | 0.01, 01 and 0.2 mg/mL | MPP<sup>+</sup>/SH-SY5Y cells | Decrease in Cytochrome c, Caspace-3 activation & ROS generation         | [101]|
|                              | Panaxatriol saponins| 100 mg/kg, twice daily for 7 days | MPTP/ Kunning mice | Improvement in behavioral impairments caused by MPTP                   | [102]|
| *Polygala tenuifolia*         | Aqueous extract     | 0.05-µg/mL and 100 mg/kg | 6-OHDA/PC12 cells and MPP<sup>+</sup>/C57BL/6 mice | Decrease in ROS & protection of mesencephalic dopaminergic neurons        | [106]|
|                              | Tenuigenin          | 10 µM               | 6-OHDA/SH-SY5Y cells     | Increased expression of glutathione (GSH) and superoxide dismutase (SOD) | [107]|
| Plant name/species | Extract/constituent | Dose | Toxin/tested subjects | Biological activity and targets | Ref. |
|-------------------|---------------------|------|------------------------|--------------------------------|------|
| *Bacopa monnieri* | Standardized aqueous extract | 10 mg/mL | MPP⁺, paraquat/SK-N-SH cells | Decrease in intracellular ROS & upregulation of Nrf2 | [111] |
| | Standardized aqueous extract | 0.05 and 0.1% | Rotenone/Drosophila melanogaster flies | Ablation of oxidative stress and dopamine depletion | [112] |
| | Ethanolic extract | 2, 4 and 6 µg/1 mg/kg b.w./day, i.p. | Rotenone/N27 cell and prepubertal mice | Decrease oxidative stress & increase Anti-oxidant defense | [116] |
| | Ethanolic extract | 0.5 and 1.0 µg | 3-nitropropionic Acid/prepubertal mice | Scavenge free radicals, maintain redox status, and upregulate antioxidant machinery. | [115] |
| *Hyoscyamus niger* | Seed extract | 125–500 mg/kg, p.o. | MPTP/Male Balb/c mice | Alleviation of motor deficits produced by MPTP and inhibition of hydroxyl radical | [121] |
| *Hibiscus asper* | Methanolic extract | 50 and 100 mg/kg, i.p., daily, for 7 days | 6-HDA/Wistar rats | Increased antioxidant defense system like GSH and SOD & decreased MDA levels | [123] |
| *Melissa officinalis* | Aqueous extract | 100 mg/kg/day in the drinking water | Manganeseese/Male albino mice | Decrease in thiobarbituric acid & Increased SOD and catalase | [126] |
| *Cassia obtusifolia* | Seed extract | 0.1-10 µg/mL/50 mg/kg, 15 days | 6-OHA/PC12 cells and MPTP/Male C57BL/6 mice | Inhibition of GSH depletion caspase-3 activation/Decreased T-turn and T-LA time in pole test | [128] |
| | Seed extract | 1.0 and 10 µg | NMDA, 3-NP, Amyloid β/ Hippocampal cultures from C57BL/6 mice | Reduce excitotoxicity, mitochondrial dysfunction, and Aβ toxicity. regulation and maintenance of cellular homeostasis and apoptosis | [129] |
| *Croton celtidifolius* | Bark | 10 mg/kg, i.p. | MPTP/Wistar rats | Prevention of mitochondrial complex-I inhibition & improvement in memory deficits | [132] |
| *Gynostemma pentaphyllum* | Gypenosides | 100, 200 and 400 mg/kg | MPTP/C57BL/6 mice | Increase in SOD and GSH content & survival of nigral dopaminergic neurons | [177] |
| *Thuja orientalis* | Ethanolic leaf extract | 10 µg/mL | 6-OHDA/SH-SY5Y cells | Decrease in ROS production | [143] |
| *Paullinia cupana* | Seed powder | 0.312 and 0.625 mg/mL | Rotenone/SH-SY5Y cells | Survival of SH-SY5Y cells exposed to rotenone | [151] |
| Plant name/species          | Extract/constituent | Dose                     | Toxin/tested subjects                       | Biological activity and targets                                                                 | Ref.  |
|----------------------------|--------------------|--------------------------|---------------------------------------------|--------------------------------------------------------------------------------------------------|-------|
| Mulberry                   | Ethanol extract    | 1, 10 and 100 µg/mL/500 mg/mL and MPP7/Male C57BL/6 mice | 6-OHDA/SH-SY5Y cells and MPP7/Male C57BL/6 mice | Decrease in ROS, NO & Caspase-3 generation/Improvement in behavioral deficits                       | [162] |
| Ginkgo biloba              | EGb761 extract     | 40 mg/kg                 | MPTP/C57BL/6J mice                          | Decrease in lipid peroxidation, increase in striatal dopamine and locomotor activity               | [164] |
| Uncaria rhynchophylla      | Aqueous extract    | 0.1, 0.5 and 1.0 µg (in vitro) and 5 mg/kg/day for 14 days (in vivo) | 6-OHDA/PC12 cells and 6-OHDA/Sprague–Dawley rats | Reduced cell death, generation of ROS, increased GSH levels, and inhibited caspase-3 activity. Lowered dopaminergic cell loss, antioxidative and anti-apoptotic activities. | [165] |
| Trifolium pretense         | Aqueous extract    | 0.5, 1 and 2 µg/mL       | hydrogen peroxide/HCN 1-A cells             | Antioxidant activity                                                                            | [166] |
| Salvia miltiorrhiza        | Salvianolic acid B | 0.1, 1.0 and 10 µM       | 6-OHDA/SH-SY5Y cells                        | Redox regulation and antioxidant effect                                                           | [167] |
| Hypericum perforatum       | Flavonoid rich extract | 3.125, 12.5 and 25 µg/mL | hydrogen peroxide/PC12 cells                | Elevated the cell viability decreased the levels of LDH release and decreased the occurrence of apoptotic cells. Antioxidant and apoptotic activity | [178] |
| Fraxinus sieboldiana       | 6,7-di-O-glucopyranosylesculetin | 0.1, 1 and 10 µM | 6-OHDA/SH-SY5Y cells                        | Redox regulation and antioxidant properties                                                      | [179] |
| Scutellaria baicalensis    | Baicalein          | 0.05, 0.5 and 5 µg/mL    | 6-OHDA/SH-SY5Y cells and Sprague–Dawley rats | Antioxidant and antiapoptotic activities                                                          | [169] |
| Withania somnifera         | Aqueous root extract | 100 mg/kg                | MPTP/C57BL/6 mice                           | Regulation of redox status and antioxidant effects                                                | [170] |
| Cyperus rotundus           | Aqueous rhizome extract | 50 and 100 µg/mL/PC12 cells | 6-OHDA/PC12 cells                           | Antioxidant and antiapoptotic activities                                                          | [174] |
| Delphinium denudatum       | Ethanolic extract  | 200, 400 and 600 mg/kg   | 6-OHDA/Rats                                 | Regulation of antioxidative enzymes and antioxidant effects                                      | [175] |
| Ilex paraguariensis        | Hydro-alcoholic extract | 250 and 500 mg/kg        | MPTP/C57BL/6 mice                           | Antioxidant activity                                                                            | [176] |
7. Conclusions and Future Perspectives

This study searched the literature for the most recent available data about antioxidant plants/constituents that possess neuroprotective activity in experimental models of PD. Despite the great variety of plants, only a few have had their pharmacological effects investigated for antiparkinsonian activity; thus, there are huge prospects in this field for future studies with plants and their bioactive molecules. Notably, the plants discussed in this review have played important roles for centuries as pharmacologically efficient therapeutics against brain diseases. These medications have been validated by traditional use and are time tested, as compared to modern day supplements. Growing evidence suggests a model for redox homeostasis in which the ROS-antioxidant interactions act as a metabolic interface for signals derived from metabolism and the environment. Several redox regulating transcriptional signaling factors and enzymes are involved during the oxidative stress response. Modulation of these factors by antioxidative herbal products provides important opportunities for neuroprotection based on molecular targeting.

Synthetic antioxidants are adding up their negative or no effects and are recently highlighted to be less beneficial to human health. Thus the search for effective, nontoxic natural molecules with immense antioxidative activity has been intensified. Antioxidants provide essential information on cellular redox state and they influence gene expression associated with oxidative stress responses to maximize defense. Several molecules have been identified to be responsible in regulating redox status and oxidative stress which is one of the key factors in the etiopathogenesis of neurodegenerative diseases including PD. In addition to endogenous antioxidant defense systems, a promising strategy for targeting redox status of cells is to use readily available natural substances from vegetables, fruits and herbs. Therefore, consumption of plant-derived antioxidants may appear to be a suitable alternative.

Mounting evidence in randomized control trials indicated that purified antioxidants have produced inconsistent results suggesting that supplementation by natural antioxidant herbs may modulate neurodegeneration. The behavior and bioavailability of the pure antioxidant may differ from that of the same molecule consumed as part of a complex herb matrix [180]. Evidence showed that antioxidant treatment of the natural herbs discussed in the present review results in neuroprotection in experimental models of PD. These herbs can also be utilized as adjuvant therapy with combination of main therapy to ameliorate the symptoms of PD. Moreover, this combination will reduce the side effects and will provide an alternative approach for treatment of PD [181]. The potential clinical benefits deriving from these natural antioxidant herbs is still under wide debate. Further research on their ability to delay or forestall disease progression by protecting or rescuing the DAnergic neurons or even restoring those that have been lost is required.

Future perspectives may include a logical combination of ROS-modulating herbs and their derived molecules that affect redox-sensitive signaling pathways which may further enhance the therapeutic activity when compared to the currently existing treatment regime. Although the repair of damaged DAergic neurons seen in PD may be beyond the possibilities of natural antioxidants, combinations of antioxidant herbs and supplementation with current drugs may serve a neuroprotective purpose and decrease the risk of condition and progression in PD. Integration of pharmacology, natural product chemistry, medicinal chemistry, clinical and other related disciplines could be the most promising strategy for drug discovery and increase success rate. Studies on improved quality control techniques,
authenticity, standardization of the natural herbal molecules and their ability to cross the blood-brain barrier to reach the target sites is quite necessary. Establishing the characteristic chemical “fingerprints” for herbs and herbal products are also warranted.

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References

1. Dowding, C.H.; Shenton, C.L.; Salek, S.S. A review of the health-related quality of life and economic impact of Parkinson’s disease. Drugs Aging 2006, 23, 693–721.
2. Lokk, J. Caregiver strain in Parkinson’s disease and the impact of disease duration. Eur. J. Phys. Rehabil. Med. 2008, 44, 39–45.
3. Winter, Y.; von Campenhausen, S.; Brozova, H.; Skoupa, J.; Reese, J.P.; Botzel, K.; Eggert, K.; Oertel, W.H.; Dodel, R.; Ruzicka, E. Costs of Parkinson’s disease in eastern Europe: A Czech cohort study. Parkinsonism Relat. Disord. 2010, 16, 51–56.
4. Gao, H.M.; Liu, B.; Zhang, W.; Hong, J.S. Novel anti-inflammatory therapy for Parkinson’s disease. Trends Pharmacol. Sci. 2003, 24, 395–401.
5. Kidd, P.M. Parkinson’s disease as multifactorial oxidative neurodegeneration: Implications for integrative management. Altern. Med. Rev. 2000, 5, 502–529.
6. Olanow, C.W.; Tatton, W.G. Etiology and pathogenesis of Parkinson’s disease. Annu. Rev. Neurosci. 1999, 22, 123–144.
7. Suchowersky, O. Parkinson’s disease: Medical treatment of moderate to advanced disease. Curr. Neurol. Neurosci. Rep. 2002, 2, 310–316.
8. Trachootham, D.; Lu, W.; Ogasawara, M.A.; Nilsa, R.D.; Huang, P. Redox regulation of cell survival. Antioxid. Redox Signal 2008, 10, 1343–1374.
9. Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. Int. J. Biochem. Cell Biol. 2007, 39, 44–84.
10. Haddad, J.J. Antioxidant and prooxidant mechanisms in the regulation of redox(y)-sensitive transcription factors. Cell Signal 2002, 14, 879–897.
11. Bauer, M.K.; Vogt, M.; Los, M.; Siegel, J.; Wesselborg, S.; Schulze-Osthoff, K. Role of reactive oxygen intermediates in activation-induced CD95 (APO-1/Fas) ligand expression. J. Biol. Chem. 1998, 273, 8048–8055.
12. Kowaltowski, A.J.; Vercesi, A.E.; Fiskum, G. Bcl-2 prevents mitochondrial permeability transition and cytochrome c release via maintenance of reduced pyridine nucleotides. Cell Death Differ. 2000, 7, 903–910.
13. Leslie, N.R. The redox regulation of PI 3-kinase-dependent signaling. Antioxid. Redox Signal 2006, 8, 1765–1774.
14. Matsuzawa, A.; Ichijo, H. Stress-responsive protein kinases in redox-regulated apoptosis signaling. Antioxid. Redox Signal 2005, 7, 472–481.
15. Zhao, Y.; Wang, Z.B.; Xu, J.X. Effect of cytochrome c on the generation and elimination of $\text{O}_2^\cdot$ and $\text{H}_2\text{O}_2$ in mitochondria. *J. Biol. Chem.* 2003, 278, 2356–2360.

16. Addabbo, F.; Montagnani, M.; Goligorsky, M.S. Mitochondria and reactive oxygen species. *Hypertension* 2009, 53, 885–892.

17. Archer, S.L.; Gomberg-Maitland, M.; Maitland, M.L.; Rich, S.; Garcia, J.G.; Weir, E.K. Mitochondrial metabolism, Redox signaling, And fusion: A mitochondria-ROS-HIF-1alpha-Kv1.5 O2-sensing pathway at the intersection of pulmonary hypertension and cancer. *Am. J. Physiol. Heart Circ. Physiol.* 2008, 294, H570–H578.

18. Beal, M.F. Therapeutic approaches to mitochondrial dysfunction in Parkinson’s disease. *Parkinsonism Relat. Disord.* 2009, 15, S189–S194.

19. Fasano, M.; Bergamasco, B.; Lopiano, L. Modifications of the iron-neuromelanin system in Parkinson’s disease. *J. Neurochem.* 2006, 96, 909–916.

20. Przedborski, S.; Ischiropoulos, H. Reactive oxygen and nitrogen species: weapons of neuronal destruction in models of Parkinson’s disease. *Antioxid. Redox Signal* 2005, 7, 685–693.

21. Ischiropoulos, H.; Beckman, J.S. Oxidative stress and nitration in neurodegeneration: Cause, Effect, Or association? *J. Clin. Invest.* 2003, 111, 163–169.

22. Anderson, S.; Bankier, A.T.; Barrell, B.G.; de Bruijn, M.H.; Coulson, A.R.; Drouin, J.; Eperon, I.C.; Nierlich, D.P.; Roe, B.A.; Sanger, F.; *et al.* Sequence and organization of the human mitochondrial genome. *Nature* 1981, 290, 457–465.

23. Dexter, D.T.; Wells, F.R.; Lees, A.J.; Agid, F.; Agid, Y.; Jenner, P.; Marsden, C.D. Increased nigral iron content and alterations in other metal ions occurring in brain in Parkinson’s disease. *J. Neurochem.* 1989, 52, 1830–1836.

24. Pappolla, M.A.; Chyan, Y.J.; Poeggeler, B.; Bozner, P.; Ghiso, J.; LeDoux, S.P.; Wilson, G.L. Alzheimer beta protein mediated oxidative damage of mitochondrial DNA: Prevention by melatonin. *J. Pineal Res.* 1999, 27, 226–229.

25. Retz, W.; Gsell, W.; Münch, G.; Rosler, M.; Riederer, P. Free radicals in Alzheimer’s disease. *J. Neural. Transm.* 1998, 221–236.

26. Saggu, H.; Cooksey, J.; Dexter, D.; Wells, F.R.; Lees, A.; Jenner, P.; Marsden, C.D. A selective increase in particulate superoxide dismutase activity in parkinsonian substantia nigra. *J. Neurochem.* 1989, 53, 692–697.

27. Sofic, E.; Lange, K.W.; Jellinger, K.; Riederer, P. Reduced and oxidized glutathione in the substantia nigra of patients with Parkinson’s disease. *Neurosci. Lett.* 1992, 142, 128–130.

28. Przedborski, S.; Levivier, M.; Jiang, H.; Ferreira, M.; Jackson-Lewis, V.; Donaldson, D.; Togasaki, D.M. Dose-dependent lesions of the dopaminergic nigrostriatal pathway induced by intrastratal injection of 6-hydroxydopamine. *Neuroscience* 1995, 67, 631–647.

29. Olanow, C.W.; Jenner, P.; Tatton, N.A.; Tatton, W.G. *Parkinson’s Disease and Movement Disorders*; Williams & Wilkins: Baltimore, MD, USA, 1998; pp. 67–103.

30. Uttara, B.; Singh, A.V.; Zamboni, P.; Mahajan, R.T. Oxidative stress and neurodegenerative diseases: A review of upstream and downstream antioxidant therapeutic options. *Curr. Neuropharmacol.* 2009, 7, 65–74.

31. Zhou, C.; Huang, Y.; Przedborski, S. Oxidative stress in Parkinson’s disease: A mechanism of pathogenic and therapeutic significance. *Ann. NY Acad. Sci.* 2008, 1147, 93–104.
32. Ebadi, M.; Srinivasan, S.K.; Baxi, M.D. Oxidative stress and antioxidant therapy in Parkinson’s disease. *Prog. Neurobiol.* 1996, 48, 1–19.
33. Jenner, P. Oxidative stress in Parkinson’s disease. *Ann. Neurol.* 2003, 53, S26–S36.
34. Jenner, P.; Olanow, C.W. Oxidative stress and the pathogenesis of Parkinson’s disease. *Neurology* 1996, 47, S161–S170.
35. Bove, J.; Prou, D.; Perier, C.; Przedborski, S. Toxin-induced models of Parkinson’s disease. *NeuroRx.* 2005, 2, 484–494.
36. Dauer, W.; Przedborski, S. Parkinson’s disease: Mechanisms and models. *Neuron* 2003, 39, 889–909.
37. Ambani, L.M.; Van Woert, M.H.; Murphy, S. Brain peroxidase and catalase in Parkinson disease. *Arch. Neurol.* 1975, 32, 114–118.
38. Kish, S.J.; Morito, C.; Hornykiewicz, O. Glutathione peroxidase activity in Parkinson’s disease brain. *Neurosci. Lett.* 1985, 58, 343–346.
39. Perry, T.L.; Godin, D.V.; Hansen, S. Parkinson’s disease: A disorder due to nigral glutathione deficiency? *Neurosci. Lett.* 1982, 33, 305–310.
40. Riederer, P.; Sofic, E.; Rausch, W.D.; Schmidt, B.; Reynolds, G.P.; Jellinger, K.; Youdim, M.B. Transition metals, Ferritin, Glutathione, And ascorbic acid in parkinsonian brains. *J. Neurochem.* 1989, 52, 515–520.
41. Prasad, K.N.; Cole, W.C.; Kumar, B. Multiple antioxidants in the prevention and treatment of Parkinson’s disease. *J. Am. Coll. Nutr.* 1999, 18, 413–423.
42. Draczynska-Lusiak, B.; Doung, A.; Sun, A.Y. Oxidized lipoproteins may play a role in neuronal cell death in Alzheimer disease. *Mol. Chem. Neuropathol.* 1998, 33, 139–148.
43. Droge, W.; Kinscherf, R.; Hildebrandt, W.; Schmitt, T. The deficit in low molecular weight thiols as a target for antiageing therapy. *Curr. Drug Targets* 2006, 7, 1505–1512.
44. Poeggeler, B. Melatonin, aging, and age-related diseases: Perspectives for prevention, Intervention, And therapy. *Endocrine* 2005, 27, 201–212.
45. Victor, V.M.; Rocha, M. Targeting antioxidants to mitochondria: A potential new therapeutic strategy for cardiovascular diseases. *Curr. Pharm. Des.* 2007, 13, 845–863.
46. Lau, F.C.; Shukitt-Hale, B.; Joseph, J.A. Nutritional intervention in brain aging: Reducing the effects of inflammation and oxidative stress. *Subcell Biochem.* 2007, 42, 299–318.
47. Sudha, K.; Rao, A.V.; Rao, S.; Rao, A. Free radical toxicity and antioxidants in Parkinson’s disease. *Neurol. India* 2003, 51, 60–62.
48. Weber, C.A.; Ernst, M.E. Antioxidants, Supplements, And Parkinson’s disease. *Ann. Pharmacother.* 2006, 40, 935–938.
49. Ungerstedt, U. 6-Hydroxy-dopamine induced degeneration of central monoamine neurons. *Eur. J. Pharmacol.* 1968, 5, 107–110.
50. Cohen, G. Oxy-radical toxicity in catecholamine neurons. *Neurotoxicology* 1984, 5, 77–82.
51. Kitamura, Y.; Kakimura, J.; Taniguchi, T. Protective effect of talipexole on MPTP-treated planarian, a unique parkinsonian worm model. *Jpn. J. Pharmacol.* 1998, 78, 23–29.
52. Kopin, I.J. MPTP: An industrial chemical and contaminant of illicit narcotics stimulates a new era in research on Parkinson's disease. *Environ. Health Perspect.* 1987, 75, 45–51.
53. Davis, G.C.; Williams, A.C.; Markey, S.P.; Ebert, M.H.; Caine, E.D.; Reichert, C.M.; Kopin, I.J. Chronic Parkinsonism secondary to intravenous injection of meperidine analogues. *Psychiatr. Res.* **1979**, *1*, 249–254.

54. Forno, L.S.; DeLanney, L.E.; Irwin, I.; Langston, J.W. Similarities and differences between MPTP-induced parkinsonism and Parkinson’s disease. Neuropathologic considerations. *Adv. Neurol.* **1993**, *60*, 600–608.

55. Muralikrishnan, D.; Mohanakumar, K.P. Neuroprotection by bromocriptine against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced neurotoxicity in mice. *FASEB J.* **1998**, *12*, 905–912.

56. Muralikrishnan, D.; Mohanakumar, K.P. Neuroprotection by bromocriptine against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced neurotoxicity in mice. *FASEB J.* **1998**, *12*, 905–912.

57. Wullner, U.; Loschmann, P.A.; Schulz, J.B.; Schmid, A.; Dringen, R.; Eblen, F.; Turski, L.; Klockgether, T. Glutathione depletion potentiates MPTP and MPP+ toxicity in nigral dopaminergic neurones. *Neuroreport* **1996**, *7*, 921–923.

58. Ali, S.F.; David, S.N.; Newport, G.D.; Cadet, J.L.; Slikker, W., Jr. MPTP-induced oxidative stress and neurotoxicity are age-dependent: Evidence from measures of reactive oxygen species and striatal dopamine levels. *Synapse* **1994**, *18*, 27–34.

59. Chiba, K.; Trevor, A.; Castagnoli, N., Jr. Metabolism of the neurotoxic tertiary amine, MPTP, By brain monoamine oxidase. *Biochem. Biophys. Res. Commun.* **1984**, *120*, 574–578.

60. Markey, S.P.; Johannessen, J.N.; Chiueh, C.C.; Burns, R.S.; Herkenham, M.A. Intraneuronal generation of a pyridinium metabolite may cause drug-induced parkinsonism. *Nature* **1984**, *311*, 464–467.

61. Bashkatova, V.; Alam, M.; Vanin, A.; Schmidt, W.J. Chronic administration of rotenone increases levels of nitric oxide and lipid peroxidation products in rat brain. *Exp. Neurol.* **2004**, *186*, 235–241.

62. Sherer, T.B.; Betarbet, R.; Testa, C.M.; Seo, B.B.; Richardson, J.R.; Kim, J.H.; Miller, G.W.; Yagi, T.; Matsuno-Yagi, A.; Greenamyre, J.T. Mechanism of toxicity in rotenone models of Parkinson's disease. *J. Neurosci.* **2003**, *23*, 10756–10764.

63. Day, B.J.; Patel, M.; Calavetta, L.; Chang, L.Y.; Stamler, J.S. A mechanism of paraquat toxicity involving nitric oxide synthase. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 12760–12765.

64. Nijveldt, R.J.; van Nood, E.; van Hoorn, D.E.; Boelens, P.G.; van Norren, K.; van Leeuwen, P.A. Flavonoids: A review of probable mechanisms of action and potential applications. *Am. J. Clin. Nutr.* **2001**, *74*, 418–425.

65. Gijtenbeek, J.M.; van den Bent, M.J.; Vecht, C.J. Cyclosporine neurotoxicity: A review. *J. Neurol.* **1999**, *246*, 339–346.

66. Johnson, W.C.; Williford, W.O. Benefits, Morbidity, And mortality associated with long-term administration of oral anticoagulant therapy to patients with peripheral arterial bypass procedures: A prospective randomized study. *J. Vasc. Surg.* **2002**, *35*, 413–421.

67. Heinrich, M.; Gibbons, S. Ethnopharmacology in drug discovery: An analysis of its role and potential contribution. *J. Pharm. Pharmacol.* **2001**, *53*, 425–432.

68. Halliwell, B. Vitamin C: Antioxidant or pro-oxidant in vivo? *Free Radic. Res.* **1996**, *25*, 439–454.

69. Osiecki, M.; Ghanavi, P.; Atkinson, K.; Nielsen, L.K.; Doran, M.R. The ascorbic acid paradox. *Biochem. Biophys. Res. Commun.* **2010**, *400*, 466–470.
70. Kang, M.J.; Lee, S.S.; Koh, H.C. Prooxidant properties of ascorbic acid in the nigrostriatal dopaminergic system of C57BL/6 mice. Toxicology 2012, 294, 1–8.

71. de Rijk, M.C.; Breteler, M.M.; den Breeijen, J.H.; Launer, L.J.; Grobbee, D.E.; van der Meche, F.G.; Hofman, A. Dietary antioxidants and Parkinson disease. The Rotterdam Study. Arch. Neurol. 1997, 54, 762–765.

72. Kim, T.H.; Cho, K.H.; Jung, W.S.; Lee, M.S. Herbal medicines for Parkinson’s disease: A systematic review of randomized controlled trials. PLoS One 2012, 7, e35695.

73. Ramarathnam, N.; Ochi, H.; Takeuchi, M. Antioxidants Defense system in Vegetable Extracts. In Natural Antioxidants: Chemistry, Health Effects and Applications; Shahidi, F., Ed.; Aoca Press: Champaign, IL, USA, 1997.

74. Houghton, P.J.; Howes, M.J. Natural products and derivatives affecting neurotransmission relevant to Alzheimer’s and Parkinson’s disease. Neurosignals 2005, 14, 6–22.

75. Van Kampen, J.; Robertson, H.; Hagg, T.; Drobitch, R. Neuroprotective actions of the ginseng extract G115 in two rodent models of Parkinson’s disease. Exp. Neurol. 2003, 184, 521–529.

76. Shunying, Z.; Yang, Y.; Huaidong, Y.; Yue, Y.; Guolin, Z. Chemical composition and antimicrobial activity of the essential oils of Chrysanthemum indicum. J. Ethnopharmacol. 2005, 96, 151–158.

77. Kato, T.; Noguchi, K.; Miyamoto, Y.; Suekawa, M.; Aburada, M.; Hosoya, E.; Sakanashi, M. Effects of Chrysanthemum indicum Linn. on coronary, Vertebral, Renal and aortic blood flows of the anesthetized dog. Arch. Int. Pharmacodyn. Ther. 1987, 285, 288–300.

78. Yu, D.Q.; Xie, F.Z.; He, W.Y.; Liang, X.T. Application of 2D NMR techniques in the structure determination of chrysanthetriol. Yao Xue Xue Bao 1992, 27, 191–196.

79. Wang, Z.D.; Huang, C.; Li, Z.F.; Yang, J.; Li, B.H.; Liang, R.R.; Dai, Z.J.; Liu, Z.W. Chrysanthemum indicum ethanolic extract inhibits invasion of hepatocellular carcinoma via regulation of MMP/TIMP balance as therapeutic target. Oncol. Rep. 2010, 23, 413–421.

80. Kim, I.S.; Ko, H.M.; Koppula, S.; Kim, B.W.; Choi, D.K. Protective effect of Chrysanthemum indicum Linn against 1-methyl-4-phenylpyridinium ion and lipopolysaccharide-induced cytotoxicity in cellular model of Parkinson’s disease. Food Chem. Toxicol. 2011, 49, 963–973.

81. Debnath, T.; Jin, H.L.; Hasnát, M.A.; Kim, Y.; Samad, N.B.; Park, P.J.; Lim, B.O. Antioxidant potential and oxidative DNA damage preventive activity of Chrysanthemum indicum extracts. J. Food Biochem. 2012, 1–9.
86. Lee, Y.S.; Ha, J.H.; Yong, C.S.; Lee, D.U.; Huh, K.; Kang, Y.S.; Lee, S.H.; Jung, M.W.; Kim, J.A. Inhibitory effects of constituents of Gastrodia elata Bl. on glutamate-induced apoptosis in IMR-32 human neuroblastoma cells. Arch. Pharm. Res. 1999, 22, 404–409.

87. Kim, H.J.; Moon, K.D.; Oh, S.Y.; Kim, S.P.; Lee, S.R. Ether fraction of methanol extracts of Gastrodia elata, A traditional medicinal herb, protects against kainic acid-induced neuronal damage in the mouse hippocampus. Neurosci. Lett. 2001, 314, 65–68.

88. An, H.; Kim, I.S.; Kopppula, S.; Kim, B.W.; Park, P.J.; Lim, B.O.; Choi, W.S.; Lee, K.H.; Choi, D.K. Protective effects of Gastrodia elata Blume on MPP⁺-induced cytotoxicity in human dopaminergic SH-SY5Y cells. J. Ethnopharmacol. 2010, 130, 290–298.

89. Kim, H.J.; Lee, S.R.; Moon, K.D. Ether fraction of methanol extracts of Gastrodia elata, medicinal herb protects against neuronal cell damage after transient global ischemia in gerbils. Phytother. Res. 2003, 17, 909–912.

90. Kim, I.S.; Choi, D.K.; Jung, H.J. Neuroprotective effects of vanillyl alcohol in Gastrodia elata Blume through suppression of oxidative stress and anti-apoptotic activity in toxin-induced dopaminergic MN9D cells. Molecules 2011, 16, 5349–5361.

91. Shin, E.J.; Bach, J.H.; Nguyen, T.T.; Nguyen, X.K.; Jung, B.D.; Oh, K.W.; Kim, M.J.; Ko, S.K.; Jang, C.G.; Ali, S.F.; et al. Gastrodia elata bl attenuates methamphetamine-induced dopaminergic toxicity via inhibiting oxidative burdens. Curr. Neuropharmacol. 2011, 9, 118–121.

92. Clauson, K.A.; K.M. Shields; McQueen, C.E.; Persad, N. Safety issues associated with commercially available energy drinks. J. Am. Pharm. Assoc. 2008, 48, 55–63.

93. Wen, T.C.; Yoshimura, H.; Matsuda, S.; Lim, J.H.; Sakanaka, M. Ginseng root prevents learning disability and neuronal loss in gerbils with 5-minute forebrain ischemia. Acta Neuropathol. 1996, 91, 15–22.

94. Vogler, B.K.; Pittler, M.H.; Ernst, E. The efficacy of ginseng. A systematic review of randomised clinical trials. Eur. J. Clin. Pharmacol. 1999, 55, 567–575.

95. Lee, T.F.; Shiao, Y.J.; Chen, C.F.; Wang, L.C. Effect of ginseng saponins on beta-amyloid-suppressed acetylcholine release from rat hippocampal slices. Planta Med. 2001, 67, 634–637.

96. Xiang, Y.Z.; Shang, H.C.; Gao, X.M.; Zhang, B.L. A comparison of the ancient use of ginseng in traditional Chinese medicine with modern pharmacological experiments and clinical trials. Phytother. Res. 2008, 22, 851–858.

97. Jin, S.H.; Park, J.K.; Nam, K.Y.; Park, S.N.; Jung, N.P. Korean red ginseng saponins with low ratios of protopanaxadiol and protopanaxatriol saponin improve scopolamine-induced learning disability and spatial working memory in mice. J. Ethnopharmacol. 1999, 66, 123–129.

98. Zhang, D.; Yasuda, T.; Yu, Y.; Zheng, P.; Kawabata, T.; Ma, Y.; Okada, S. Ginseng extract scavenges hydroxyl radical and protects unsaturated fatty acids from decomposition caused by iron-mediated lipid peroxidation. Free Radic. Biol. Med. 1996, 20, 145–150.

99. Kitts, D.D.; Wijewickreme, A.N.; Hu, C. Antioxidant properties of a North American ginseng extract. Mol. Cell. Biochem. 2000, 203, 1–10.

100. Keum, Y.S.; Park, K.K.; Lee, J.M.; Chun, K.S.; Park, J.H.; Lee, S.K.; Kwon, H.; Surh, Y.J. Antioxidant and anti-tumor promoting activities of the methanol extract of heat-processed ginseng. Cancer Lett. 2000, 150, 41–48.
101. Hu, S.; Han, R.; Mak, S.; Han, Y. Protection against 1-methyl-4-phenylpyridinium ion (MPP+) induced apoptosis by water extract of ginseng (Panax ginseng C.A. Meyer) in SH-SY5Y cells. J. Ethnopharmacol. 2011, 135, 34–42.

102. Luo, F.C.; Wang, S.D.; Qi, L.; Song, J.Y.; Lv, T.; Bai, J. Protective effect of panaxatriol saponins extracted from Panax notoginseng against MPTP-induced neurotoxicity in vivo. J. Ethnopharmacol. 2011, 133, 448–453.

103. Shin, K.Y.; Lee, J.Y.; Won, B.Y.; Jung, H.Y.; Chang, K.A.; Koppula, S.; Suh, Y.H. BT-11 is effective for enhancing cognitive functions in the elderly humans. Neurosci. Lett. 2009, 465, 157–159.

104. Chen, Y.L.; Hsieh, C.L.; Wu, P.H.; Lin, J.G. Effect of Polygala tenuifolia root on behavioral disorders by lesioning nucleus basalis magnocellularis in rat. J. Ethnopharmacol. 2004, 95, 47–55.

105. Zhang, H.; Han, T.; Zhang, L.; Yu, C.H.; Wan, D.G.; Rahman, K.; Qin, L.P.; Peng, C. Effects of tenuifolin extracted from radix polygalae on learning and memory: A behavioral and biochemical study on aged and amnesic mice. Phytomedicine 2008, 15, 587–594.

106. Choi, J.G.; Kim, H.G.; Kim, M.C.; Yang, W.M.; Huh, Y.; Kim, S.Y.; Oh, M.S. Polygalae radix inhibits toxin-induced neuronal death in the Parkinson’s disease models. J. Ethnopharmacol. 2011, 134, 414–421.

107. Liang, Z.; Shi, F.; Wang, Y.; Lu, L.; Zhang, Z.; Wang, X. Neuroprotective effects of tenuigenin in a SH-SY5Y cell model with 6-OHDA-induced injury. Neurosci. Lett. 2011, 497, 104–109.

108. Howes, M.J.; Houghton, P.J. Plants used in Chinese and Indian traditional medicine for improvement of memory and cognitive function. Pharmacol. Biochem. Behav. 2003, 75, 513–527.

109. Russo, A.; Borrelli, F. Bacopa monniera, A reputed nootropic plant: An overview. Phytomedicine 2005, 12, 305–317.

110. Hota, S.K.; Barhwal, K.; Baitharu, I.; Prasad, D.; Singh, S.B.; Ilavazhagan, G. Bacopa monniera leaf extract ameliorates hypobaric hypoxia induced spatial memory impairment. Neurobiol. Dis. 2009, 34, 23–39.

111. Singh, M.; Murthy, V.; Ramassamy, C. Standardized extracts of Bacopa monniera protect against MPP+- and paraquat-induced toxicity by modulating mitochondrial activities, Proteasomal functions, And redox pathways. Toxicol. Sci. 2012, 125, 219–232.

112. Hosamani, R.; Muralidhara. Neuroprotective efficacy of Bacopa monnieri against rotenone induced oxidative stress and neurotoxicity in Drosophila melanogaster. Neurotoxicology 2009, 30, 977–985.

113. Sachs, C.; Jonsson, G. Mechanisms of action of 6-hydroxydopamine. Biochem. Pharmacol. 1975, 24, 1–8.

114. Shinomol, G.K.; Bharath, M.M.; Muralidhara. Pretreatment with Bacopa monnieri extract offsets 3-nitropropionic acid induced mitochondrial oxidative stress and dysfunctions in the striatum of prepubertal mouse brain. Can. J. Physiol. Pharmacol. 2012, 90, 595–606.

115. Shinomol, G.K.; Bharath, M.M.; Muralidhara. Neuromodulatory propensity of Bacopa monnieri leaf extract against 3-nitropropionic acid-induced oxidative stress: in vitro and in vivo evidences. Neurotox. Res. 2012, 22, 102–114.
116. Shinomol, G.K.; Mythri, R.B.; Srinivas Bharath, M.M.; Muralidhara. *Bacopa monnieri* extract offsets rotenone-induced cytotoxicity in dopaminergic cells and oxidative impairments in mice brain. *Cell Mol. Neurobiol.* 2012, 32, 455–465.

117. Sawant, M.; Isaac, J.C.; Narayanan, S. Analgesic studies on total alkaloids and alcohol extracts of *Eclipta alba* (Linn.) Hassk. *Phytother. Res.* 2004, 18, 111–113.

118. El Jaber-Vazdeks, N.; Gonzalez, C.; Ravelo, A.G.; Zarate, R. Cloning, Characterization and analysis of expression profiles of a cDNA encoding a hyoscyamine 6beta-hydroxylase (H6H) from *Atropa baetica* Willk. *Plant Physiol. Biochem.* 2009, 47, 20–25.

119. Gourie-Devi, M.; Ramu, M.G.; Venkataram, B.S. Treatment of Parkinson’s disease in “Ayurveda” (ancient Indian system of medicine): Discussion paper. *J. R. Soc. Med.* 1991, 84, 491–492.

120. Nagashayana, N.; Sankarankutty, P.; Nampoothiri, M.R.; Mohan, P.K.; Mohanakumar, K.P. Association of L-DOPA with recovery following Ayurveda medication in Parkinson’s disease. *J. Neurol. Sci.* 2000, 176, 124–127.

121. Sengupta, T.; Vinayagam, J.; Nagashayana, N.; Gowda, B.; Jaisankar, P.; Mohanakumar, K.P. Antiparkinsonian effects of aqueous methanolic extract of *Hyoscyamus niger* seeds result from its monoamine oxidase inhibitory and hydroxyl radical scavenging potency. *Neurochem. Res.* 2011, 36, 177–186.

122. Foyet, H.S.; Hritcu, L.; Ciobica, A.; Stefan, M.; Kametchouing, P.; Cojocaru, D. Methanolic extract of *Hibiscus asper* leaves improves spatial memory deficits in the 6-hydroxydopamine-lesion rodent model of Parkinson’s disease. *J. Ethnopharmacol.* 2011, 133, 773–779.

123. Hritcu, L.; Foyet, H.S.; Stefan, M.; Mihasan, M.; Asongalem, A.E.; Kametchouing, P. Neuroprotective effect of the methanolic extract of *Hibiscus asper* leaves in 6-hydroxydopamine-lesioned rat model of Parkinson’s disease. *J. Ethnopharmacol.* 2011, 137, 585–591.

124. Salah, S.M.; Jager, A.K. Screening of traditionally used Lebanese herbs for neurological activities. *J. Ethnopharmacol.* 2005, 97, 145–149.

125. Dos Santos-Neto, L.L.; de Vihena, Toledo, M.A.; Medeiros-Souza, P.; de Souza, G.A. The use of herbal medicine in Alzheimer’s disease—A systematic review. *Evid. Based Complement Alternat. Med.* 2006, 23, 441–445.

126. Martins, E.N.; Pessano, N.T.; Leal, L.; Roos, D.H.; Folmer, V.; Puntel, G.O.; Rocha, J.B.; Aschner, M.; Avila, D.S.; Puntel, R.L. Protective effect of *Melissa officinalis* aqueous extract against Mn-induced oxidative stress in chronically exposed mice. *Brain Res. Bull.* 2012, 87, 74–79.

127. Kim, D.H.; Kim, S.; Jung, W.Y.; Park, S.J.; Park, D.H.; Kim, J.M.; Cheong, J.H.; Ryu, J.H. The neuroprotective effects of the seeds of *Cassia obtusifolia* on transient cerebral global ischemia in mice. *Food Chem. Toxicol.* 2009, 47, 1473–1479.

128. Ju, M.S.; Kim, H.G.; Choi, J.G.; Ryu, J.H.; Hur, J.; Kim, Y.J.; Oh, M.S. Cassiae semen, A seed of *Cassia obtusifolia*, Has neuroprotective effects in Parkinson’s disease models. *Food Chem. Toxicol.* 2010, 48, 2037–2044.

129. Drever, B.D.; Anderson, W.G.; Riedel, G.; Kim, D.H.; Ryu, J.H.; Choi, D.Y.; Platt, B. The seed extract of *Cassia obtusifolia* offers neuroprotection to mouse hippocampal cultures. *J. Pharmacol. Sci.* 2008, 107, 380–392.
130. Nardi, G.M.; Felippi, R.; DalBo, S.; Siqueira-Junior, J.M.; Arruda, D.C.; Delle Monache, F.; Timbola, A.K.; Pizzolatti, M.G.; Ckless, K.; Ribeiro-do-valle, R.M. Anti-inflammatory and antioxidant effects of Croton celtidifolius bark. *Phytotherapy Research* 2003, 10, 176–184.

131. Nardi, G.M.; Siqueira Junior, J.M.; Delle Monache, F.; Pizzolatti, M.G.; Ckless, K.; Ribeiro-do-Valle, R.M. Antioxidant and anti-inflammatory effects of products from Croton celtidifolius Bailon on carrageenan-induced pleurisy in rats. *Phytotherapy Research* 2007, 14, 115–122.

132. Moreira, E.L.; Rial, D.; Aguiar, A.S., Jr.; Figueiredo, C.P.; Siqueira, J.M.; DalBo, S.; Horst, H.; de Oliveira, J.; Mancini, G.; dos Santos, T.S.; et al. Proanthocyanidin-rich fraction from Croton celtidifolius Baill confers neuroprotection in the intranasal 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine rat model of Parkinson’s disease. *J. Neural. Transm.* 2010, 117, 1337–1351.

133. Chen, J.C.; Tsai, C.C.; Chen, L.D.; Chen, H.H.; Wang, W.C. Therapeutic effect of gypenoside on chronic liver injury and fibrosis induced by CCl4 in rats. *Am. J. Chin. Med.* 2000, 28, 175–185.

134. Megalli, S.; Aktan, F.; Davies, N.M.; Roufogalis, B.D. Phytopreventative anti-hyperlipidemic effects of Gynostemma pentaphyllum in rats. *J. Pharma. Pharm. Sci.* 2005, 8, 507–515.

135. Wang, Q.F.; Chen, J.C.; Hsieh, S.J.; Cheng, C.C.; Hsu, S.L. Regulation of Bcl-2 family molecules and activation of caspase cascade involved in gypenosides-induced apoptosis in human hepatoma cells. *Cancer Lett.* 2002, 183, 169–178.

136. la Cour, B.; Molgaard, P.; Yi, Z. Traditional Chinese medicine in treatment of hyperlipidaemia. *J. Ethnopharmacol.* 1995, 46, 125–129.

137. Shang, L.; Liu, J.; Zhu, Q.; Zhao, L.; Feng, Y.; Wang, X.; Cao, W.; Xin, H. Gypenosides protect primary cultures of rat cortical cells against oxidative neurotoxicity. *Brain Res.* 2006, 1102, 163–174.

138. Wang, Y.H.; Samoylenko, V.; Tekwani, B.L.; Khan, I.A.; Miller, L.S.; Chaurasiya, N.D.; Rahman, M.M.; Tripathi, L.M.; Khan, S.I.; Joshi, V.C.; et al. Composition, Standardization and chemical profiling of Banisteriopsis caapi, A plant for the treatment of neurodegenerative disorders relevant to Parkinson’s disease. *J. Ethnopharmacol.* 2010, 128, 662–671.

139. Choi, H.S.; Park, M.S.; Kim, S.H.; Hwang, B.Y.; Lee, C.K.; Lee, M.K. Neuroprotective effects of herbal ethanol extracts from Gynostemma pentaphyllum in the 6-hydroxydopamine-lesioned rat model of Parkinson’s disease. *Molecules* 2010, 15, 2814–2824.

140. Nizam, I.; Mushfiq, M. Antioxidant activity of water and alcohol extracts of Thuja orientalis leaves. *Oriental Pharm. Exp. Med.* 2007, 7, 65–73.

141. Choi, Y.; Moon, A.; Kim, Y.C. A pinusolide derivative, 15-methoxypinusolidic acid from Biota orientalis inhibits inducible nitric oxide synthase in microglial cells: implication for a potential anti-inflammatory effect. *Int. Immunopharmacol.* 2008, 8, 548–555.

142. Koo, K.A.; Kim, S.H.; Lee, M.K.; Kim, Y.C. 15-Methoxypinusolidic acid from Biota orientalis attenuates glutamate-induced neurotoxicity in primary cultured rat cortical cells. *Toxicol. In Vitro* 2006, 20, 936–941.

143. Ju, M.S.; Lee, P.; Kim, H.G.; Lee, K.Y.; Hur, J.; Cho, S.H.; Sung, S.H.; Oh, M.S. Protective effects of standardized Thuja orientalis leaves against 6-hydroxydopamine-induced neurotoxicity in SH-SY5Y cells. *Toxicol. In Vitro* 2010, 24, 759–765.

144. Yamaguti-Sasaki, E.; Ito, L.A.; Canteli, V.C.; Ushirobira, T.M.; Ueda-Nakamura, T.; Dias Filho, B.P.; Nakamura, C.V.; de Mello, J.C. Antioxidant capacity and in vitro prevention of dental decay.
plaque formation by extracts and condensed tannins of *Paullinia cupana*. *Molecules* 2007, 12, 1950–1963.

145. Datla, K.P.; Zbarsky, V.; Rai, D.; Parkar, S.; Osakabe, N.; Ariuoma, O.I.; Dexter, D.T. Short-term supplementation with plant extracts rich in flavonoids protect nigrostriatal dopaminergic neurons in a rat model of Parkinson’s disease. *J. Am. Coll. Nutr.* 2007, 26, 341–349.

146. Shirwaikar, A.; Rajendran, K.; Punitha, I.S. *In vitro* antioxidant studies on the benzyl tetraisoquinoline alkaloid berberine. *Biol. Pharm. Bull.* 2006, 29, 1906–1910.

147. Fragoso, V.; do Nascimento, N.C.; Moura, D.J.; e Silva, A.C.; Richter, M.F.; Saffi, J.; Fett-Neto, A.G. Antioxidant and antimutagenic properties of the monoterpene indole alkaloid psychollatine and the crude foliar extract of *Psychotria umbellata* Vell. *Toxicol. In Vitro* 2008, 22, 559–566.

148. Liu, Y.; Ji, H.; Dong, J.; Zhang, S.; Lee, K.J.; Matthew, S. Antioxidant alkaloid from the South China Sea marine sponge *Iotrochota* sp. *Z. Naturforsch. C.* 2008, 63, 636–638.

149. Nobre, H.V., Jr.; Cunha, G.M.; de Vasconcelos, L.M.; Magalhaes, H.I.; Oliveira Neto, R.N.; Maia, F.D.; de Moraes, M.O.; Leal, L.K.; Viana, G.S. Caffeine and CSC, adenosine A2A antagonists, Offer neuroprotection against 6-OHDA-induced neurotoxicity in rat mesencephalic cells. *Neurochem. Int.* 2010, 56, 51–58.

150. Basile, A.; Ferrara, L.; Pezzo, M.D.; Mele, G.; Sorbo, S.; Bassi, P.; Montesano, D. Antibacterial and antioxidant activities of ethanol extract from *Paullinia cupana* Mart. *J. Ethnopharmacol.* 2005, 102, 32–36.

151. de Oliveira, D.M.; Barreto, G.; Galeano, P.; Romero, J.I.; Holubiec, M.I.; Badorrey, M.S.; Capani, F.; Alvarez, L.D. *Paullinia cupana* Mart. var. Sorbilis protects human dopaminergic neuroblastoma SH-SY5Y cell line against rotenone-induced cytotoxicity. *Hum. Exp. Toxicol.* 2011, 30, 1382–1391.

152. Chen, J.K.; Chen, T.T. Tonic herbs. *Chinese Medical Herbology and Pharmacology*, Crampton, L., Ed.; Art of Medicine Press: City of Industry, CA, USA, 2004.

153. Kim, A.J.; Park, S. Mulberry extract supplements ameliorate the inflammation-related hematological parameters in carrageenan-induced arthritic rats. *J. Med. Food* 2006, 9, 431–435.

154. Hwang, K.H.; Kim, Y.K. Promoting effect and recovery activity from physical stress of the fruit of *Morus alba*. *Biofactors* 2004, 21, 267–271.

155. Kang, T.H.; Hur, J.Y.; Kim, H.B.; Ryu, J.H.; Kim, S.Y. Neuroprotective effects of the cyanidin-3-O-beta-d-glucopyranoside isolated from mulberry fruit against cerebral ischemia. *Neurosci. Lett.* 2006, 391, 122–126.

156. Isabelle, M.; Lee, B.L.; Ong, C.N.; Liu, X.; Huang, D. Peroxyl radical scavenging capacity, Polyphenolics, And lipophilic antioxidant profiles of mulberry fruits cultivated in southern China. *J. Agric. Food Chem.* 2008, 56, 9410–9416.

157. Shih, P.H.; Chan, Y.C.; Liao, J.W.; Wang, M.F.; Yen, G.C. Antioxidant and cognitive promotion effects of anthocyanin-rich mulberry (*Morus atropurpurea* L.) on senescence-accelerated mice and prevention of Alzheimer’s disease. *J. Nutr. Biochem.* 2010, 21, 598–605.

158. Liu, L.K.; Lee, H.J.; Shih, Y.W.; Chyau, C.C.; Wang, C.J. Mulberry anthocyanin extracts inhibit LDL oxidation and macrophage-derived foam cell formation induced by oxidative LDL. *J. Food Sci.* 2008, 73, H113–H121.
159. Dugo, P.; Mondello, L.; Errante, G.; Zappia, G.; Dugo, G. Identification of anthocyanins in berries by narrow-bore high-performance liquid chromatography with electrospray ionization detection. *J. Agric. Food Chem.* **2001**, *49*, 3987–3992.

160. Moyer, R.A.; Hummer, K.E.; Finn, C.E.; Frei, B.; Wrolstad, R.E. Anthocyanins, Phenolics, And antioxidant capacity in diverse small fruits: Vaccinium, Rubus, And ribes. *J. Agric. Food Chem.* **2002**, *50*, 519–525.

161. Zhang, W.; Han, F.; He, J.; Duan, C. HPLC-DAD-ESI-MS/MS analysis and antioxidant activities of nonanthocyanin phenolics in mulberry (*Morus alba* L.). *J. Food Sci.* **2008**, *73*, C512–C518.

162. Kim, H.G.; Ju, M.S.; Shim, J.S.; Kim, M.C.; Lee, S.H.; Huh, Y.; Kim, S.Y. Oh, M.S. Mulberry fruit protects dopaminergic neurons in toxin-induced Parkinson’s disease models. *Br. J. Nutr.* **2010**, *104*, 8–16.

163. Nam, S.; Jang, H.W.; Shibamoto, T. Antioxidant activities of extracts from teas prepared from medicinal plants, *Morus alba* L., *Camellia sinensis* L. and *Cudrania tricuspidata* and their volatile components. *J. Agric. Food Chem.* **2012**, *60*, 9097–9105.

164. Rojas, P.; Serrano-Garcia, N.; Mares-Samano, J.J.; Medina-Campos, O.N.; Pedraza-Chaverri, J.; Ogren, S.O. EGB761 protects against nigrostriatal dopaminergic neurotoxicity in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinsonism in mice: Role of oxidative stress. *Eur. J. Neurosci.* **2008**, *28*, 41–50.

165. Shim, J.S.; Kim, H.G.; Ju, M.S.; Choi, J.G.; Jeong, S.Y.; Oh, M.S. Effects of the hook of *Uncaria rhynchophylla* on neurotoxicity in the 6-hydroxydopamine model of Parkinson’s disease. *J. Ethnopharmacol.* **2009**, *125*, 361–365.

166. Occhiuto, F.; Palumbo, D.R.; Samperi, S.; Zangla, G.; Pino, A.; De Pasquale, R.; Circosta, C. The isoflavones mixture from *Trifolium pratense* L. protects HCN 1-A neurons from oxidative stress. *Phytother. Res.* **2009**, *23*, 192–196.

167. Tian, L.L.; Wang, X.J.; Sun, Y.N.; Li, C.R.; Xing, Y.L.; Zhao, H.B.; Duan, M.; Zhou, Z.; Wang, S.Q. Salvianolic acid B, An antioxidant from *Salvia miltiorrhiza*, Prevents 6-hydroxydopamine induced apoptosis in SH-SY5Y cells. *Int. J. Biochem. Cell Biol.* **2008**, *40*, 409–422.

168. Zhou, L.; Kang, J.; Fan, L.; Ma, X.C.; Zhao, H.Y.; Han, J.; Wang, B.R.; Guo, D.A. Simultaneous analysis of coumarins and secoiridoids in *Cortex Fraxini* by high-performance liquid chromatography-diode array detection-electrospray ionization tandem mass spectrometry. *J. Pharm. Biomed. Anal.* **2008**, *47*, 39–46.

169. Mu, X.; He, G.; Cheng, Y.; Li, X.; Xu, B.; Du, G. Baicalein exerts neuroprotective effects in 6-hydroxydopamine-induced experimental parkinsonism in vivo and in vitro. *Pharmacol. Biochem. Behavior* **2009**, *92*, 642–648.

170. RajaSankar, S.; Manivasagam, T.; Sankar, V.; Prakash, S.; Muthusamy, R.; Krishnamurti, A.; Surendran, S. *Withania somnifera* root extract improves catecholamines and physiological abnormalities seen in a Parkinson’s disease model mouse. *J. Ethnopharmacol.* **2009**, *125*, 369–373.

171. Kim, S.; Park, S.E.; Sapkota, K.; Kim, M.K.; Kim, S.J. Leaf extract of *Rhus verniciflua* Stokes protects dopaminergic neuronal cells in a rotenone model of Parkinson’s disease. *J. Pharm. Pharmacol.* **2011**, *63*, 1358–1367.
172. Park, B.C.; Lee, Y.S.; Park, H.J.; Kwak, M.K.; Yoo, B.K.; Kim, J.Y.; Kim, J.A. Protective effects of fustin, a flavonoid from Rhus verniciflua Stokes, on 6-hydroxydopamine-induced neuronal cell death. *Exp. Mol. Med.* **2007**, *39*, 316–326.

173. An, L.; Liu, S.; Dong, Y.; Tang, B.; Dong, W. Protective effect of effective part of Acanthopanacis senticosus on damage of PC12 cells induced by MPP+. *Zhongguo Zhong Yao Za Zhi* **2010**, *35*, 2021–2026.

174. Lee, C.H.; Hwang, D.S.; Kim, H.G.; Oh, H.; Park, H.; Cho, J.H.; Lee, J.M.; Jang, J.B.; Lee, K.S.; Oh, M.S. Protective effect of Cyperi rhizoma against 6-hydroxydopamine-induced neuronal damage. *J. Med. Food* **2010**, *13*, 564–571.

175. Ahmad, M.; Yousuf, S.; Khan, M.B.; Ahmad, A.S.; Saleem, S.; Hoda, M.N.; Islam, F. Protective effects of ethanolic extract of Delphinium denudatum in a rat model of Parkinson’s disease. *Hum. Exp. Toxicol.* **2006**, *25*, 361–368.

176. Milioli, E.M.; Cologni, P.; Santos, C.C.; Marcos, T.D.; Yunes, V.M.; Fernandes, M.S.; Schoenfelder, T.; Costa-Campos, L. Effect of acute administration of hydroalcohol extract of Ilex paraguariensis St Hilaire (Aquifoliaceae) in animal models of Parkinson’s disease. *Phytother. Res.* **2007**, *21*, 771–776.

177. Wang, H.; Li, C.; Wu, X.; Lou, X. Effects of Gynostemma pentaphyllum (Thunb.) Makino polysaccharides supplementation on exercise tolerance and oxidative stress induced by exhaustive exercise in rats. *Afr. J. Argri. Res.* **2012**, *7*, 2632–2638.

178. Kleparnik, K.; Mala, Z.; Havac, Z.; Blazkova, M.; Holla, L.; Bocek, P. Fast detection of a (CA)18 microsatellite repeat in the IgE receptor gene by capillary electrophoresis with laser-induced fluorescence detection. *Electrophoresis* **1998**, *19*, 249–255.

179. Lin, S.; Liu, M.T.; Wang, S.J.; Li, S.; Yang, Y.C.; Shi, J.G. Coumarins from branch of Fraxinus sieboldiana and their antioxidative activity. *Zhongguo Zhong Yao Za Zhi* **2008**, *33*, 1708–1710.

180. Liu, R.H. Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *Am. J. Clin. Nutr.* **2003**, *78*, 517S-520S.

181. Valkovic, P.; Benetin, J.; Blazicek, P.; Valkovicova, L.; Gmitterova, K.; Kukumberg, P. Reduced plasma homocysteine levels in levodopa/entacapone treated Parkinson patients. *Parkinsonism Relat. Disord.* **2005**, *11*, 253–256.

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