“More Than Meets the Eye”: Cryptic Diversity and Contrasting Patterns of Host-Specificity in Feather Mites Inhabiting Seabirds
Laura Stefan, Elena Gómez-Díaz, Sergey Mironov, Jacob González-Solís, Karen McCoy

To cite this version:
Laura Stefan, Elena Gómez-Díaz, Sergey Mironov, Jacob González-Solís, Karen McCoy. “More Than Meets the Eye”: Cryptic Diversity and Contrasting Patterns of Host-Specificity in Feather Mites Inhabiting Seabirds. Frontiers in Ecology and Evolution, Frontiers Media S.A, 2018, 6, pp.97. 10.3389/fevo.2018.00097. hal-01957259

HAL Id: hal-01957259
https://hal.umontpellier.fr/hal-01957259
Submitted on 17 Dec 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
“More Than Meets the Eye”: Cryptic Diversity and Contrasting Patterns of Host-Specificity in Feather Mites Inhabiting Seabirds

Laura M. Stefan1,2†, Elena Gómez-Díaz3*, Sergey V. Mironov4, Jacob González-Solís3 and Karen D. McCoy2

†Departament de Biologia Evolutiva, Ecologia i Ciències Ambientals, Institut de Recerca de la Biodiversitat, Universitat de Barcelona, Barcelona, Spain, 1 MIVEGEC Research Unit, UMR 5290, Centre IRD, Centre National de la Recherche Scientifique-IRD-University of Montpellier, Montpellier, France, 3 Estación Biológica de Doñana, Consejo Superior de Investigaciones Científicas, Sevilla, Spain, 4 Zoological Institute, Russian Academy of Sciences, Saint Petersburg, Russia

Feather mites are useful models for studying speciation due to their high diversity and strong degree of host specialization. However, studies to date have focused on the evolution of higher-level mite taxa while much hidden diversity likely occurs at the level of host genera and species. In this study, we examined the diversity and evolution of feather mites infesting six sympatric seabird species from six genera, breeding in the Cape Verde archipelago. We report 32 feather mite morphospecies categorized into 10 genera and three families, of which nine correspond to new, undescribed species. Molecular data corroborated morphological species descriptions, except for two morphologically-cryptic, but genetically distinct mite lineages related to Zachvatkinia oceanodromae and Laminalloptes simplex. Using these communities, we then applied a co-structure approach to test the contribution of ectosymbiont and host factors in driving feather mite evolution. Most seabird species hosted specific and unique feather mite species, even under sympatric conditions, and in general, feather mite species exhibited strong host-driven genetic structure. However, patterns of genetic differentiation were variable. That is, some mite species are more generalist than others and mite lineages/haplotypes can be shared by related seabird species. Interestingly, host-specific mites (e.g., Zachvatkinia spp.) tend to display much higher intra-specific diversity compared to more generalist mites (e.g., Microspalax and Plicatalloptes spp.). We discuss ectosymbiont and host life-history traits that might generate these patterns, such as host dispersal and breeding behavior and/or mite spatial and trophic specialization. Our findings highlight both the vast and largely unrecognized diversity of avian feather mites on seabirds, and the intrinsic complexity of the ecological processes underlying the evolution of these ectosymbionts.

Keywords: host-symbiont systems, cryptic species, co-structure, host-specialization, adaptation, Procellariiformes, morphology, Analgoidea
INTRODUCTION

By definition, parasites and symbionts depend almost entirely on their host for survival and transmission, and thus have evolved intricate adaptations to optimize their life cycle (de Meuës et al., 1998; Criscione et al., 2005). The degree of host dependency is thought to determine the specificity of the interaction, and we expect to find higher species diversity and stronger patterns of host specialization with increasingly intimate interactions (Poulin and Morand, 2000; Mouillot et al., 2006). This specificity may be due to limited opportunities for dispersal and colonization of new hosts (Clayton et al., 1992; Johnson et al., 2002; Whiteman et al., 2004) or may be determined by specific adaptations to host structures (e.g., feather, hair, immune responses), such that parasites/symbionts are incapable of establishing, surviving and/or reproducing on foreign hosts (Tompkins and Clayton, 1999; Reed et al., 2000).

Comparative studies of parasite co-structures have been shown to be useful in elucidating the proximal factors underlying variation in host-specificity and patterns of parasite/symbiont genetic structure (Johnson et al., 2002; Criscione, 2008; Mazé-Guilmo et al., 2016). This approach consists in comparing population demographic or genetic structures between two or more interacting species: a parasite/symbiont inhabiting multiple host species, or multiple parasite/symbiont species on a single host. Studies of this type have previously been used to improve our understanding of avian ectoparasite diversity (Johnson et al., 2002). However, multiple host and parasite/symbiont life-history traits can influence the specificity of an interaction and the resulting population structure of the parasite/symbiont (i.e., dispersal and drift, host ecology and behavior, symbiont life-cycle complexity and demography) (Blouin et al., 1995; McCoy et al., 2003; Criscione and Blouin, 2004; Bruyndonckx et al., 2009; Mazé-Guilmo et al., 2016). This can lead to conflicting predictions on patterns of structure. In this context, communities of ecologically similar and geographically overlapping host species provide an ideal framework to investigate the proximate factors involved in the divergent evolution of their parasites and symbionts. This scenario also offers the opportunity to test parasite/symbiont structure at hierarchical spatial scales (among host individuals, populations and communities), and make predictions on selection and adaptation processes (McCoy et al., 2001). Not surprisingly, few studies have used this multi-species and multi-scale approach (but see Johnson et al., 2002; Rivera-Parra et al., 2015).

Feather mites (Acari: Astigmata: Analgoidea and Pterolichoidae) are particularly useful models to study divergence and speciation processes. They are common obligate ectosymbionts of birds, with approximately 2,500 species described from extant avian orders (Gaud and Atyeo, 1996; Proctor, 2003; Mironov and Proctor, 2008). Feather mites cannot survive in the off-host environment, and have evolved diverse habitat and trophic specializations to the host environment (Dabert and Mironov, 1999; Doña et al., 2015a; Stefan et al., 2015). This strong dependency has led to the “paradigm” that feather mites are highly host-specific, and co-evolved with their bird hosts. However, previous studies reported varying degrees of co-phylogenetic patterns among mite groups, calling this paradigm into question (Gaud and Atyeo, 1996; Proctor, 2003; Doña et al., 2017a,b). That is, some feather mite species appear restricted to one or a few closely-related host species, others are associated with a wide range of hosts from different genera, and there have even been a couple of records of mite species infesting hosts from different avian families and orders (Doña et al., 2017b). One of the main difficulties in evaluating specificity in feather mite-host associations is the potential presence of cryptic diversity. Indeed, the morphological identification of feather mites is difficult. Immature stages of many taxa and females of closely-related species are often indistinguishable, while male morphological characterization is a laborious task even for experienced taxonomists (Doña et al., 2015b). Molecular studies hold much promise for documenting and understanding diversity in this group, but are still relatively scarce (but see Dabert et al., 2001, 2015; Klimov and O'Connor, 2008; Knowles and Klimov, 2011; Štefka et al., 2011; Doña et al., 2015b).

In feather mites, divergence might be affected by either host-related factors; feather structure and dynamics (i.e., moult), defenses (i.e., preening), and/or ectosymbiont life-history traits; dispersal abilities, population size, generation time (Table 1, Proctor, 2003; Pap et al., 2005; Jovani et al., 2006; Clayton et al., 2010; Fernández-González et al., 2015). Due to the dependency of feather mites on host, as their main environment, and the assumed predominant role of vertical transmission, host factors could act as important selective forces promoting the specialization of mite traits. For example, in our previous study, we showed that competition has likely led to within-host spatial and trophic niche partitioning in two co-occurring seabird mite species (M. brevipes and Z. ovata), which in turn correlates with mite-specific morphological traits (Stefan et al., 2015). These adaptations have been hypothesized to shape mite genetic structure and divergence (Gaud and Atyeo, 1996; Dabert and Mironov, 1999; Proctor and Owens, 2000; Proctor, 2003). However, comparative studies examining multiple feather mite species and considering various host and spatial factors are currently lacking.

In the present study, we explore the diversity and genetic structure of feather mite species inhabiting the community of procellariform seabirds of the Cape Verde Islands. These colonial seabirds represent excellent hosts for investigating divergence processes in feather mites. Seabirds are highly pelagic birds that breed in large colonies on remote oceanic islands. These colonies represent spatially discrete and hierarchical habitats, with extremely high host densities. Most seabird species show strong interannual fidelity and natal philopatry to their breeding sites, features that can favor parasite specialization and host specificity (Brooke, 2004; McCoy et al., 2013). However, seabirds are also long distance migrants and frequently breed sympatrically in mixed species colonies, which in turn can promote dispersal and host switching in their ectosymbionts.

The specific aims of our study were: (1) to characterize the diversity of feather mites inhabiting the community of seabirds breeding in the Cape Verde Archipelago; (2) to compare genetic
structure among feather mite species using different hosts in sympatric and allopatric breeding conditions, and (3) to evaluate the role of different host (breeding and nesting behavior) and mite (dispersal ability and within host habitat use) factors in generating these patterns.

**MATERIALS AND METHODS**

**Study Species and Feather Mite Sampling**

The Cape Verde Archipelago is located in the mid-Atlantic Ocean (16°N 24°W) approximately 570 km off the west coast of Africa. Mite sampling was performed on six seabird species, five from the order Procellariiformes (Cape Verde shearwater, *Calonectris edwardsii*; Boyd’s shearwater, *Puffinus boydi*; Bulwer’s petrel, *Bulweria bulwerii*; band-rumped storm petrel, *Hydrobates castro*; Cape Verde petrel, *Pterodroma feae*; Cape Verde shearwater, *Puffinus castro*; and black-bellied storm petrel, *Pterodroma feae*) and one from the order Phaethontiformes (*Phaethon aethereus*), breeding on five islands and islets of the archipelago: Raso, Ilhéu Cima, Ilhéu Grande, Curral Velho and Fogo (Figure 1). All sampled islands harbor multiple seabird species, except Fogo, which is inhabited by a single procellariiform species. From 2004 to 2012, feather mites were collected from captured adult birds using the dust-ruffling method (Walther and Clayton, 1997) and stored in absolute ethanol for subsequent morphological identification and molecular analyses. A total of 453 birds (*C. edwardsii*: 46, *B. bulwerii*: 83, *P. feae*: 76, *H. castro*: 173 and *P. aethereus*: 15) were sampled; details on the geographic locations and number of individuals sampled per locality and host species are shown in Figure 1. Main life-history characteristics of the feather mite genera and seabird hosts are detailed in Table 2.

**Morphological Identifications**

Initial screening of feather mites used morphological criteria to characterize the feather mite community composition on a host. Mites were cleared in lactic acid for 24 h and mounted on microscope slides in PVA medium (BioQuip Products, Rancho Dominguez, California). The slides were cured on a slidewarmer at 40°C for 4 days and then examined using a Leica DM 5000B light microscope with differential interference contrast (DIC) illumination. Mites were identified using the identification keys for bonnetelines (Mironov, 1989a,b, 2000), xolalgids (Dabert and Ehrnsberger, 1991; Mironov and Palma, 2006) and alloptids (Peterson and Atyeo, 1968; Peterson, 1971; Atyeo and Gaud, 1991; Mironov, 1996; Mironov and Stefan, 2016). Feather mite specimens of each identified morphospecies are deposited in the Zoological Institute of the Russian Academy of Sciences, Saint Petersburg, Russia (ZISP) and in the Museum of the Faculty of Biology (Centro de Recursos de Biodiversidad Animal), University of Barcelona, Spain.

**DNA Isolation, Amplification and Sequencing**

When possible, five specimens of each feather mite morphospecies per seabird host and island and from different host individuals were selected for the molecular analyses. Total numbers of mites were: 50 specimens from the genus *Zachvatkinia* (five species), 37 from the genus *Microspalax* (four species), 66 from the genus *Brephosceles* (nine species), 19 from the genus *Plicatilloptes* (one species), 13 from the genus *Laminalloptes* (three species) and 5 specimens from *Onychalloptes* (one species) (Table S1). The mite genera: *Rhinozachvatkinia*, *Promegninia*, *Ingrassia* and *Opetiopoda*, were not included in the molecular analyses due either to low sample size and/or failure to obtain DNA sequences.

---

**TABLE 1 | Key differences in life-history traits of the feather mite genera.**

| Zachvatkinia | Microspalax | Brephosceles | Plicatilloptes | Laminalloptes | Onychalloptes |
|--------------|-------------|--------------|---------------|--------------|--------------|
| Micro-habitat occupied on the host body | Flight feathers | Flight feathers | Soft body feathers and wing coverts, occasionally on flight feathers | Soft body feathers and wing coverts, occasionally on flight feathers | Flight feathers | Flight feathers |
| Body size | Large (400–700 µm) | Small (330–380 µm) | Small (230–400 µm) | Small (330–380 µm) | Large (500–800 µm) | Relatively large (410–440 µm) |
| Body sclerotization | Heavy | Heavy | Weak | Weak | Heavy | Heavy |
| References | Mironov and Stefan, 2013 | Atyeo and Gaud, 1991 | Peterson, 1971 | Mironov, 1996 | Mironov and Stefan, 2016 | Atyeo, 1988 |

---

**FIGURE 1 | Map of the Cape Verde Archipelago showing the five islands where feather mites were sampled. The name of seabird species and the number of dust-ruffling samples examined per host species and locality are shown in boxes.**
### TABLE 2 | Key differences in life-history traits of the seabird host species.

|                      | Cape Verde shearwater | Boyd’s shearwater | Bulwer’s petrel | Cape Verde petrel | Band-rumped storm-petrel | Red-billed tropicbird |
|----------------------|-----------------------|-------------------|-----------------|-------------------|--------------------------|------------------------|
| Mixed colonies       | Yes, with Boyd’s shearwater (but differ in their breeding season) and Bulwer’s petrel | Yes, with Cape Verde shearwater, Bulwer’s petrel and Band-rumped storm-petrel | Yes, with Cape Verde shearwater, Boyd’s shearwater and Band-rumped storm-petrel | No | Yes, with Cape Verde shearwater, Boyd’s shearwater and Bulwer’s petrel | No |
| Shared nests         | No | Yes, with Bulwer’s petrel and Band-rumped storm-petrel | Yes, with Boyd’s shearwater and Band-rumped storm-petrel | No | Yes, with Boyd’s shearwater and Bulwer’s petrel | No |
| Nesting area         | Hollows in cliffs and offshore rocks, and under large boulders | Burrows in soft soil or in rocky cavities | Crevices or under rocks at the base of cliffs | Burrows between rocks on remote cliffs and steep slopes at more than 800 m of altitude | Burrows or rock crevices and sandy soil | Rocky crevices, or on the ground, preferentially on cliffs |
| Breeding season      | March-October | January-June | April-October | November-May | Protracted breeding period (December-June) | All year, but with reproduction peak reached between January and March |
| Migratory distance   | Medium | Medium | Medium | Resident | Medium | Resident? |
| Wintering areas      | Between the Brazil and Falklands current, in front of the coast of southern Brazil and Uruguay | Short migrations to the Central Atlantic; some individuals remain around the Cape Verde Islands | Mid-equatorial Atlantic Ocean, north of the Saint Peter and Saint Paul archipelago | Remain around the breeding area | Undeclr if undertake systematic migrations; may migrate to the western Atlantic | Remain around the breeding area |
| References           | González-Solís et al., 2009 | Zajková et al., 2017 | Ramos et al., 2015 | Monteiro and Furness, 1998 | Monteiro and Furness, 1998 | Furness and Monteiro, 1995 |
DNA extractions were performed on each individual mite using a DNeasy Tissue Kit (Qiagen, Valencia, CA, USA) and the non-destructive method described by Dabert et al. (2008). After DNA extraction, mite exoskeletons were recovered, mounted on microslides, re-identified morphologically and kept as reference vouchers. Partial sequences of two mitochondrial genes, 12S rRNA (12S) and 16S rRNA (16S), were amplified for each feather mite using the following primers: SR-J-14199 (5’-TACATGTACGACTTAT-3’) and SR-N-14594 (5’-AAACTAGATTAGATAACC-3’) for the 12S gene (Kambhampati and Smith, 1995); and 16S A2 (5’-TTTAATTGTGTTACTTGATGATG-3’) and 16C2 (5’-CGCTGTATTCCCTAGATAT-3’) for the 16S gene (Dabert et al., 2001). Amplifications were carried out in a total volume of 25 μl containing 2.5 μl 10x reaction buffer with 15 mM MgCl₂ (Roche Diagnostics), 2 mM MgCl₂, 0.2 mM of each dNTP, 0.4 μM of each primer, 1.25 U Taq DNA polymerase (Roche Diagnostics) and 2 μl of DNA template. Amplification conditions for the 12S rRNA gene consisted of an initial step of 2 min at 94°C, followed by 10 cycles of denaturation at 94°C for 30 sec, annealing at 40°C for 30 sec, and extension at 68°C for 1 min, and 35 cycles of denaturation at 94°C for 30 sec, annealing at 43°C for 30 sec, and extension at 68°C for 1 min, with a final step of 5 min at 72°C. For the 16S rRNA gene, the PCR conditions followed Black and Piesman (1994). Amplification products were separated by electrophoresis in a 2% agarose gel and visualized under UV light. Successful amplifications were sent for sequencing to Beckman Coulter Genomics (France). Only mite specimens with complete sequence information for the two mitochondrial genes were included in the analyses. DNA sequences were checked and edited using Bioedit version 7.0.5.3 (Hall, 1999) and all variable sites were confirmed by visual inspection of the chromatograms. Sequences were aligned for each genus independently using MAFFT version 7, with default parameters.

**Molecular Diversity and Species Delimitation**

We tested for neutrality for each of the mite genus using Tajima’s D test included in DNASP v.5 (Librado and Rozas, 2009). Genetic statistics (number of polymorphic sites, number of haplotypes, nucleotide and haplotype diversity) for each gene and morphospecies/genetic lineage were assessed using DNASP v.5. Mean genetic distances within and between morphospecies/genetic lineages were calculated with MEGA 4.1 using Kimura’s 2-parameter (K2P) distance model (Tamura et al., 2007).

Following the initial morphological characterization of all mites and to control for the potential presence of cryptic species, we conducted two different species delimitation analyses on the molecular data. First, we applied the general mixed Yule-coalescent (GMYC) method (Pons et al., 2006). This method infers an ultrametric tree and, based on this topology, attempts to detect transitions in the tree where branching patterns switch from being attributed to speciation (one lineage per species) to being attributed to an intra-species coalescent process (multiple lineages per species). The GMYC analysis was conducted using the “splits” package (Species Limits by Threshold Statistics) in R version 3.0.3. The single threshold method was applied in order to find the Maximum Likelihood solution of the GMYC model. The ultrametric tree was generated using BEAST v.1.6.2 (Drummond and Rambaut, 2007) after removing identical sequences. In order to choose the most appropriate tree prior, we considered different parameter combinations: Yule vs. coalescent tree priors which refers to the model used to express the expected branching pattern on the tree, and strict clock vs. relaxed uncorrelated lognormal clock, which is compared to determine the model of molecular evolution. Secondly, for each mite genus, we inferred haplotype networks using the concatenated sequences of the two mitochondrial genes to which we applied the statistical parsimony algorithm implemented in TCS version 1.21 (Clement et al., 2000). This method partitions the data into independent haplotype sub-networks connected by changes that are non-homoplasic with a 95% probability (Templeton, 2001). Although this threshold does not necessarily correspond to species boundaries, the algorithm has been shown to be useful in separating independent lineages, which in most cases correspond to good species (Hart and Sunday, 2007).

**Genetic Structure**

To examine the genetic structure of the mites, we first mapped host and geographic information onto the mitochondrial haplotype network of each mite species. In general, each host species harbored its own set of feather mite species, except for three mite species (M. brevipes, B. puffini, and Plicatalloptes sp.1) shared by the Cape Verde and Boyd’s shearwaters (see results). For these mite species, we evaluated the partitioning of genetic variation within and between feather mite populations (in this case at host species and host individual levels), using one-way analyses of molecular variance (AMOVA) in ARLEQUIN version 3.5 and determined significance using 10,000 permutations. We also calculated overall and pairwise estimates of FST in ARLEQUIN (Excoffier and Lischer, 2010) and then examined genetic structure over geographic space. We ran AMOVA analyses, partitioning genetic variation among islands and among individuals within islands for each mite species that occurred on at least two islands.

**RESULTS**

**Morphological Diversity of Feather Mites**

Thirty-two feather mite species belonging to ten genera and three families were found on the six studied seabird species based only on morphological criteria: Zachvatkinia, Rhinozachvatkinia, Promegninia (Avenzoariidae), Microspalax, Brephosceles, Plicatalloptes, Laminalloptes, Onychalloptes (Alliotidae), Ingrassia, and Opetiopoda (Xolalidae) (Table 3). Many of these genera, such as Zachvatkinia, Microspalax, Brephosceles and Ingrassia, co-occurred in multiple seabird species in the Cape Verde Islands, whereas the genera Rhinozachvatkinia, Promegninia, Opetiopoda, Plicatalloptes, Laminalloptes, and Onychalloptes were restricted to one or two host species.
The number of birds sampled for each species is indicated. The number of birds on which a given mite species was found is indicated in brackets preceded by the "+" symbol.

The procellariiform seabirds harbored distinct acarofauna compared to the phaethontiform species, being Ingrassia the only mite genus shared by the two avian orders. Nine of the recorded mite species corresponded to new undescribed species, and six were recently described species (Rhinozachvatkinia calonectris, Promegninia calonectris, Promegninia bulweriae, Opetiopoda bulweriae, Ingrassia calonectris, and Ingrassia micronota) (Stefan et al., 2013, 2014; Mironov et al., 2015). The Cape Verde shearwater and Bulwer's petrel hosted the greatest number of feather mite species (eight each) with the highest number of new species (three for the Cape Verde shearwater and four for Bulwer’s petrel), whereas the Cape Verde petrel harbored the lowest mite richness with only three species found. Except for three feather mite species (M. brevipes, B. puffini, and Plicatalloptes sp.1) which were shared by Cape Verde and Boyd’s shearwaters, each host species harbored its own feather mite assembly (Table 3).

**Table 3**: Feather mite richness for each seabird species breeding in the Cape Verde Islands.

| Fam. Avenzoariidae | Cape Verde shearwater (46) | Boyd's Shearwater (60) | Bulwer's petrel (83) | Cape Verde petrel (76) | Band-rumped storm-petrel (173) | Red-billed tropicbird (15) | Locality |
|---------------------|-----------------------------|------------------------|----------------------|------------------------|-------------------------------|-----------------------------|----------|
| Zachvatkinia ovata  | + (29)                      |                        |                      |                        |                               |                             | Raso, Curral Velho             |
| Z. sp.1             |                             | + (23)                 |                      |                        |                               |                             | Raso, Ilhéu Cima              |
| Z. sp.2             |                             |                        | + (24)               |                        |                               |                             | Raso, Ilhéu Cima, Ilhéu Grande |
| Z. oceanodromae     |                             |                        |                      | + (29)                 |                               |                             | Raso, Curral Velho, Ilhéu Cima, Ilhéu Grande |
| Z. sp.3             |                             |                        |                      | + (10)                 |                               |                             | Fogo                             |
| Rhinozachvatkinia calonectris | + (8)   |                        |                      |                        |                               |                             | Raso, Curral Velho             |
| Promegninia calonectris | + (1)  |                        |                      |                        |                               |                             | Raso                             |
| P. bulweriae        |                             |                        |                      | + (2)                  |                               |                             | Raso, Ilhéu Cima              |

| Fam. Alloptidae      |                              |                        |                      |                        |                               |                             |                      |
| Microspalax brevipes | + (37)                      | + (13)                 |                      |                        |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| M. bulweriae         | + (9)                       |                        |                      |                        |                               |                             | Raso, Ilhéu Cima, Ilhéu Grande |
| M. cymochoreae       | + (12)                      |                        |                      | + (1)                  |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| M. pterodromae       |                             |                        |                      | + (1)                  |                               |                             | Fogo                             |
| Brehosceles puffini  | + (27)                      | + (22)                 |                      | + (29)                 |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| B. sp.1              |                             | + (2)                  |                      | + (4)                  |                               |                             | Raso, Ilhéu Cima              |
| B. sp.2              |                             |                        |                      | + (2)                  |                               |                             | Ilhéu Cima                     |
| B. sp.3              |                             |                        |                      | + (2)                  |                               |                             | Raso                             |
| B. sp.4              | + (5)                       |                        |                      | + (42)                 |                               |                             | Raso, Curral Velho             |
| B. sp.5              | + (3)                       |                        |                      | + (12)                 |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| B. decapus           |                             |                        |                      | + (42)                 |                               |                             | Fogo                             |
| B. lanceolatus       |                             | + (12)                 |                      | + (12)                 |                               |                             | Fogo                             |
| B. disjunctus        |                             | + (12)                 |                      | + (20)                 |                               |                             | Fogo                             |
| Plicatalloptes sp.1  | + (25)                      | + (12)                 |                      | + (25)                 |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| Laminalloptes phaetontis | + (12)  |                        |                      | + (12)                 |                               |                             | Raso                             |
| L. minor             | + (12)                      |                        |                      | + (12)                 |                               |                             | Raso                             |
| L. simplex           | + (11)                      |                        |                      | + (11)                 |                               |                             | Raso                             |
| Oncialloptes microphaeton | + (14) |                        |                      | + (14)                 |                               |                             | Raso                             |

| Fam. Xolalgidae      |                              |                        |                      |                        |                               |                             |                      |
| Ingrassia calonectris | + (30)         |                        |                      | + (23)                 |                               |                             | Raso, Curral Velho             |
| I. dubinini          |                             | + (23)                 |                      | + (26)                 |                               |                             | Raso, Ilhéu Cima              |
| I. micronota         | + (26)                      |                        | + (26)               | + (26)                 |                               |                             | Raso, Ilhéu Cima, Ilhéu Grande |
| I. oceanodromae      |                             | + (59)                 |                      | + (59)                 |                               |                             | Raso, Curral Velho, Ilhéu Cima |
| I. aquinocinctilis   | + (9)                       |                        | + (9)                | + (9)                  |                               |                             | Raso                             |
| Opetiopoda bulweriae | + (4)                       |                        |                      | + (4)                  |                               |                             | Raso                             |

| Total number of mite species | 8 | 6 | 8 | 3 | 5 | 5 |

The number of birds sampled for each species is indicated. The number of birds on which a given mite species was found is indicated in brackets preceded by the "+" symbol.
Molecular Diversity and Structure of Feather Mites

In total, 190 sequences for each of the two mitochondrial genes were obtained for 23 out of the 32 morphologically distinct species (Table S1).

The number of haplotypes per mite species ranged from 1 to 11 for the 12S gene and from 1 to 6 for the 16S gene. Haplotype diversity ranged from 0 to 1 for both genes, while nucleotide diversity ranged from 0 to 0.0051 once cryptic lineages were accounted for (Table S2). No deviation from neutrality was found for any mite genus or gene (12S and 16S) lineages were accounted for (Table S2). Mean genetic distances between the genetic lineages of Z. oceanodromae A and Z. oceanodromae B was 6.6% for the 12S gene and 7% for the 16S gene, whereas between L. simplex A and L. simplex B it was 11.2 and 16.4%, respectively for the two genes.

The GMYC analysis suggested the presence of 26 putative mite species within Cape Verde Islands when considering the Yule-strict clock combination [N_GMYC = 29 (CI 28–30), L_GMYC = 732.49; where N = number of species including the 3 outgroups and L = likelihood of GMYC model]. For the coalescent strict clock, the GMYC analysis resulted in 25 putative mite species [N_GMYC = 28 (CI 28–30), L_GMYC = 738.21] (Figure S1). For both models (Yule and coalescent), the GMYC method indicated that there are two species within Z. oceanodromae and two within L. simplex. The only disagreement between the two models used to delimit species concerned M. brevipes, which was partitioned into two distinct species corresponding to different host species (Cape Verde and Boyd’s shearwaters) for the Yule-strict clock combination. Neither the coalescent strict clock nor the TCS parsimony network analysis (see below) recognized these as separate lineages. In favor of this split, the Bayes factor of the Yule prior with a strict clock gave a slightly better fit than a coalescent prior (Yule: −5755.31, coalescent: −5741.33).

The mtDNA networks revealed high levels of haplotypic diversity, with 102 distinct haplotypes, based on the concatenated 12S and 16S sequences. These sequences grouped into 25 different networks, which agrees with the 25 genetic lineages identified by the GMYC approach. Furthermore, we a high degree of correspondence between morphological and molecular data (Figures 2A–F). Again, the only exceptions were Z. oceanodromae from band-rumped storm petrels and L. simplex from red-billed tropicbirds for which two different sub-networks were found (Z. oceanodromae A and B; Figure 2A and L. simplex A and B; Figure 2E).

As outlined above, clear patterns of host-associated genetic structure were apparent in most of the feather mite genera analyzed. Twenty out of the 23 mite species examined showed a one-host one-mite species pattern (Figure 2). M. brevipes, B. puffini, and Plicatalloptes sp.1 were shared by two related hosts, Cape Verde and Boyd’s shearwaters (Figure 2). The two shearwater hosts share breeding habitats, but differ in their breeding phenology (Table 2). In accordance with this temporal isolation, AMOVA results showed significant host-associated genetic differentiation for M. brevipes (Φ = 0.669, P = 0) and B. puffini (Φ = 0.225, P = 0.010) (Table 4). This was not the case for Plicatalloptes sp.1 (Φ = 0.048, P = 0.069), in which genetic structure between host species was not statistically significant. In contrast, when testing for the effect of geography, the TCS parsimony networks showed only weak spatial genetic structure in the feather mite species that occurred on more than one island population (Figures 3A–D). Indeed, almost all mite species shared haplotypes among islands. Furthermore, the AMOVA analyses with island as a grouping factor supported this observation, with only Zachvatkinia sp.1 and M. brevipes showing a tendency for island-related genetic structure (Table S4).

DISCUSSION

In studying the diversity of feather mites on procellariform and phaethontiform seabirds from Cape Verde, we uncovered a unique mite fauna composed of 32 morphologically distinct species belonging to ten genera, of which nine are new, undescribed species. All seabird species examined in this study hosted at least three feather mite species, with Cape Verde shearwaters and Bulwer’s petrels presenting the richest mite communities (eight species each). This number is in line with mite richness documented in other groups of birds, including seabirds (Fitzpatrick and Threlfall, 1977; Doña et al., 2016). For instance, other procellariform seabirds are known to harbor more than one mite species: Puffinus gravis (nine), Oceanodroma leucorhoa (eight), and Puffinus tenuirostris (seven) (Bourgeois and Threlfall, 1979; Doña et al., 2016).

The host-feather mite associations found in the present study are largely in agreement with previous records in the literature (Peterson, 1971; Mironov, 1989a; Atyeo and Gaud, 1991; Stefan et al., 2013; Mironov and Stefan, 2016), although some inconsistencies were detected. First, Z. ovata was previously reported from Calonecristis diomedea borealis and different species of Puffinus shearwaters (Mironov, 1989a). Our study clearly shows that these two host genera harbor distinct species of Zachvatkinia. Another disagreement concerns Brephosceles puffini, previously known only from Puffinus hosts (Peterson,
FIGURE 2 | Continued
FIGURE 2 | Continued
Stefan et al. Host-Specificity in Feather Mites

FIGURE 2 | Statistical parsimony haplotype networks for seabird feather mites Zachvatkinia (A), Microspalax (B), Brephosceles (C), Plicatalloptes (D), Laminalloptes (E), and Onychalloptes (F) from the Cape Verde Islands based on the concatenation of two mitochondrial genes (12S and 16S). Haplotypes are colored according to the hosts harboring the mites (green – C. edwardsii, blue – P. boydi, red – B. bulwerii, purple – O. castro, orange – P. feae, and gray – P. aethereus). Circle sizes are proportional to the number of individuals possessing each haplotype, indicated in brackets; when no number is indicated, only one mite individual presented this haplotype. Black dots represent mutational steps. The number of mite individuals analyzed per each bird species is shown in brackets.

TABLE 4 | Analysis of molecular variance (AMOVA) on mitochondrial haplotypes partitioned by seabird host.

| Species               | Partition        | d.f. | Sum of squares | % Variation | ΦST      | P   |
|-----------------------|------------------|------|----------------|-------------|----------|-----|
| Microspalax brevipes  | Among-host       | 1    | 15.800         | 66.91       | 0.669    | 0   |
|                       | Within-host      | 18   | 13.400         | 33.09       | –        | –   |
| Brephosceles puffini  | Among-host       | 1    | 2.150          | 22.54       | 0.225    | 0.010|
|                       | Within-host      | 18   | 9.900          | 77.46       | –        | –   |
| Plicatalloptes sp.1   | Among-host       | 1    | 1.503          | 4.82        | 0.048    | 0.069|
|                       | Within-host      | 17   | 17.665         | 95.18       | –        | –   |

This analysis was conducted on the three feather mite species that are shared by Cape Verde shearwater and Boyd’s shearwater.

1971); the present study revealed that B. puffini also inhabits Calonectris shearwaters. This does not seem to be a spurious result given that B. puffini specimens were found in various localities in which the two seabird genera (Calonectris and Puffinus) are not sympatric.

One of the difficulties in measuring diversity and in studying host-parasite associations is the presence of cryptic species. Cryptic speciation appears to be common in ectosymbionts, such as lice (Malenke et al., 2009) and mites (Miller et al., 2013; Doña et al., 2015b). The strong host-associated adaptive pressures on these ectosymbionts can promote convergence in morphology, rendering traditional morphological methods insufficient for species identification (Doña et al., 2015b). To address this problem, we integrated morphological and molecular data. We found that, in general, morphospecies correlated well with genetic lineages, with the exception of two species (Zachvatkinia oceanodromae, Laminalloptes simplex). Similar examples of cases of hidden diversity were found in the literature in avian skin (Whiteman et al., 2006), nasal and feather mites (Doña et al., 2015b). In the case of Z. oceanodromae, some small morphological differences could be detected on secondary examination (i.e., slightly shorter incision in the interlobar membrane; S. Mironov personal observation). Therefore, in order to confirm the existence of the two cryptic species found
FIGURE 3 | Continued
in the present study, more specimens belonging to each species should be collected and redescribed using multilocus genetic data and more quantitative morphological methods (e.g., geometric morphometrics).

Hosts are the most important driving force generating diversity in symbionts due to the selective pressure imposed by the host environment, i.e. immune and behavioral responses (Magalhães et al., 2007). This selection can result in host adaptation, which in turn, can promote specialization and diversification (Gandon and Michalakis, 2002; Lajeunesse and Forbes, 2002; Malenke et al., 2009). In agreement with the hypothesis of host-driven diversification, this study revealed...
that most seabird species hosted specific and unique mite species, even under sympatry. When mite species were shared, it was between phylogenetically related seabird host taxa (“clade-limited switches”) (Krasnov et al., 2010; Poulin, 2010; Poulin et al., 2011; Braga et al., 2015). For example, Zachvatkinia, Microspalax, Brephosceles and Plicatalloptes, all occurred on multiple host species, but with evident host-associated genetic structure that conforms with the high degree of trophic, spatial and morphological specialization reported in previous studies (Dabert and Mironov, 1999; Dabert et al., 2015; Stefan et al., 2015). Our findings are also in agreement with previous co-phylogenetic studies reporting that host switching is infrequent and likely plays a relatively minor role in the diversification of these ectosymbionts (Doña et al., 2016, 2017b).

Despite the high level of host-specificity, we observed differences in host-driven feather mite genetic structuring. That is, Zachvatkinia appear highly genetically structured by the host. In contrast, M. brevipes and B. puffini occur in two related shearwater species but with low host-associated genetic divergence. More extreme is the case of Plicatalloptes sp.1 which showed no genetic differentiation among mites inhabiting each of the two host species. Accumulating evidence suggests that multiple parasite and host life-history traits are important in determining the adaptive dynamics and population structure of parasites (Mazé-Guilmo et al., 2016). In this context, studying multiple host-symbiont/parasite associations, where hosts and symbionts/parasites differ in key characteristics such as dispersal and behavior, can provide insight on the factors that affect their microevolutionary dynamics. In lice, differences in specific habitat use within the host body has been shown to alter patterns of among-host dispersal, with body lice having reduced dispersal potential compared to wing lice (Johnson et al., 2002). In the case of feather mites, ecological characteristics linked to differential dispersal abilities between wing and body mites are not congruent with host-associated patterns of structure. Zachvatkinia and Microspalax are large vane-dwelling wing mites with heavily sclerotized bodies that occupy exposed areas along the ventral surfaces of flight feathers (Stefan et al., 2015), features that should favor dispersal. In contrast, Brephosceles and Plicatalloptes are small bodied mites with weakly sclerotized bodies that occupy more protected areas, wing coverts and soft body feathers, and are less mobile (Peterson, 1971; Bourgeois and Threlfall, 1979; Dabert et al., 2015; L. Stefan personal observations). These incoherencies suggest that successful host-switching in this group may depend more on host behavior and ecology than mite life histories per se.

Due to high host-specificity, host dispersal is generally assumed to drive parasite dispersal, such that their genetic structure is expected to conform to that of their hosts (McCoy et al., 2003; Barrett et al., 2008). However, empirical studies show contrasting results in relation to the degree of co-structure; some parasites mirroring host spatial genetic structuring whereas others appear completely undifferentiated (Mazé-Guilmo et al., 2016). Dispersal opportunities in otherwise highly host-specific mite lineages may arise by the fact that phylogenetically related seabird host species breed sympatrically and may even share nest sites, promoting host switching (i.e., Cape Verde and Boyd’s shearwaters). However, in the case of the shearwaters in Cape Verde, although they share habitat, they are often temporally isolated during the breeding season. Temporal segregation should “a priori” prevent host-switching given that feather mites are not able to live for long periods outside of the host body (Dubinin, 1951; Proctor, 2003; Doña et al., 2017c). It could be that feather mites survive in molted feathers in the nest substrate for relatively long periods of time, but this explanation seems unlikely given the escape behavior of feather mites during molting (Jovani and Serrano, 2001). That is, it has been shown that feather mites tend to avoid the feathers destined to be molted and leave these feathers shortly before they are dropped (Pap et al., 2005).

In contrast to strong host-associated structure, we found little geographic differentiation between mites of different islands. Direct physical contact among hosts of the same species could occur during mating and chick rearing throughout the breeding period. Mite dispersal between host populations could also be favored by juvenile birds that prospect among breeding colonies, or adult birds that change their breeding locality (Boulinier et al., 2016), or even by phoresy (Harbison et al., 2009). This is supported by previous population genetic studies of the seabird hosts that show weak genetic structuring and high between island population gene flow (Gómez-Díaz et al., 2006, 2009; Friesen et al., 2007). Regardless of the specific mechanism of dispersal, the weak structure found at relatively large geographic scales suggest that mite dispersal regularly occurs among host populations and occasionally between certain seabird host species, i.e., host switch. This supports the hypothesis that host-associated selection, rather than simple isolation, is likely responsible for the strong patterns of host-associated divergence in these ectosymbionts. Variation among some mite species in genetic diversity and geographic structure may then arise through interspecific differences in generation times and population sizes that lead, in turn, to a variable impact of genetic drift (Johnson et al., 2003). Regular population bottlenecks have been proposed to explain strong structure in contact-transmitted wing mites of social bats (Bruyndonckx et al., 2009). Unfortunately, with the genetic markers at hand and the limited knowledge on the biology of seabird mites, the drift hypothesis is difficult to test in our system. At present, we only have mite abundance data on individual hosts for two species, M. brevipes and Z. ovata, and both species show similar infestation levels (Stefan et al., 2015). Mitochondrial DNA is typically useful to resolve taxonomic uncertainties, but records few traces of contemporary events. Future studies employing more powerful SNP or microsatellite markers, combined with temporal samples on the same bird host will be needed to explore these alternatives more in-depth.

Overall, this study highlights the rich and diverse fauna of feather mites exploiting seabirds. Our morphological and molecular data have revealed the presence of nine new species and suggest the existence of at least two putative cryptic species. Our findings also reinforce the hypothesis that host specialization is a major driver of parasite/symbiont diversification with infrequent host-switching between related
seabird host species. Different degrees of host-associated genetic structure among mite species may reflect different transmission probabilities and/or genetic drift effects linked to mite life-history traits and population dynamics. Detailed examination of specific parasite/symbiont traits associated with host specificity combined with field-transplantation experiments, and temporal population genetic studies are now called for to test the relative roles of isolation and adaptation in generating mite diversity.

AVAILABILITY OF DATA AND MATERIALS

Sequence data is available at GenBank: Accession nos. KX372354-KX372371; MH077590-MH077951

ETHICS STATEMENT

Bird captures and mite sampling were performed in accordance with good animal practices as defined by current European legislation and under permission from the government authorities of Cape Verde.

AUTHOR CONTRIBUTIONS

EG-D, LS, JG-S, and KM conceived and designed the study. EG-D and JG-S collected data. LS performed the laboratory work. LS performed data analysis with input from EG-D, SM, and KM. EG-D, JG-S, and KM contributed to the interpretation of the results. EG-D, LS, and KM wrote the manuscript with input from JG-S and SM. EG-D, JG-S, and KM supervised the study.

REFERENCES

Atyeo, W. T., and Gaud, J. (1991). Microspalacinae, a new subfamily of the feather mite family Alloptidae Gaud (Acarina, Analgoidea). Folia Parasitol. 38, 327–343.

Barrett, L. G., Thrall, P. H., Burdon, J. J., and Linde, C. C. (2008). Life history determines genetic structure and evolutionary potential of host-parasite interactions. Trends Ecol. Evol. 23, 678–685. doi: 10.1016/j.tree.2008.06.017

Black, I. V. C., and Piesman, J. (1994). Phylogeny of hard- and soft-tick taxa (Acarina: Ixodida) based on mitochondrial 16S rDNA sequences. Proc. Natl. Acad. Sci. 91, 10034–10038. doi: 10.1073/pnas.91.21.10034

Blouin, M. S., Yowell, C. A., Courtney, C. H., and Dame, J. B. (1995). Host movement and the genetic structure of populations of parasitic nematodes. Genetics 141, 1007–1014.

Boulanger, T., Kada, S., Ponchon, A., Dupraz, M., Dietrich, M., Gambel, A., et al., (2016). Migration, prospecting, dispersal? What host movement matters for infectious agent circulation? Integr. Comp. Biol. 56, 330–342. doi: 10.1093/icb/icw015

Bourgeois, C. E., and Threlfall, W. (1979). Parasites of the greater shearwater (Puffinus gravis) from Newfoundland, Canada. Can. J. Zool. 57, 1355–1357. doi: 10.1139/z79-176

Braga, M. P., Razzolini, E., and Boeger, W. A. (2015). Drivers of parasite sharing among Neotropical freshwater fishes. J. Anim. Ecol. 84, 487–497. doi: 10.1111/1365-2656.12298

Brooke, M. (2004). Albatrosses and Petrels Across The World. Oxford, UK: Oxford University Press.

Bruynöndocx, N., Dubey, S., Ruedi, M., and Christie, P. (2009). Molecular cophylogenetic relationships between European bats and their ectoparasitic mites (Acari, Spinturnicidae). Mol. Phylogenet. Evol. 51, 227–237. doi: 10.1016/j.ympev.2009.02.005

Clayton, D. H., Gregory, R. D., and Price, R. D. (1992). Comparative ecology of Neotropical bird lice (Insecta: Phthiraptera). J. Anim. Ecol. 61, 195–206. doi: 10.2307/5631

Clayton, D. H., Koop, J. A. H., Harbison, C. W., Moyer, B. R., and Bush, S. E. (2010). How birds combat ectoparasites. Open Ornithol. J. 3, 41–71. doi: 10.2174/187445321003010014

Clement, M., Posada, D., and Crandall, K. A. (2000). TCS: a computer program to estimate gene genealogies. Mol. Phylogenet. Evol. 14, 493–498. doi: 10.1013/parasite/2008153439

Crisicone, C. D., and Blouin, M. S. (2004). Life cycles shape parasite evolution: comparative population genetics of salmon trematodes. Evolution 58, 198–202. doi: 10.1111/j.0014-3820.2004.tb01587.x

Crisicone, C. D., Poulin, R., and Blouin, M. S. (2005). Molecular ecology of parasites: elucidating ecological and microevolutionary processes. Mol. Ecol. 14, 2247–2257. doi: 10.1111/j.1365-294X.2005.02587.x

Dabert, J., and Mironov, S. V. (2001). Phylogeney of feather mite subfamily Avenzoariinae (Acari: Analgoidea: Avenzoariidae) inferred from combined analyses of molecular and morphological data. Mol. Phylogenet. Evol. 20, 124–135. doi: 10.1006/mpev.2001.0948

Dabert, J., and Ehrnsberger, R. (1991). Zwei neue Federmilben-Arten aus der Gattung Ingrusia Oudemans, 1905 (Analgoidea: Xolalgidae) in Explora (Romagnesi). Onabrucker Naturw. Mitt. 17, 127–142.

Dabert, J., Ehrnsberger, R., and Dabert, M. (2008). Glaucalgus tytonis sp. n. (Analgoidea, Xolalgidae) from the barn owl Tyto alba (Strigiformes, Tytonidae): compiling morphology with DNA barcode data for taxon descriptions in mites (Acari). Zootaxa 1719, 41–52.
petrel (Pterodroma feae) in Fogo Island, Cape Verde. PLoS ONE 12:e0174803. doi: 10.1371/journal.pone.0174803

Miller, A. D., Skoracka, A., Nava, D., de Mendonca, R. S., Szidlo, W., Schultz, M. B., et al. (2013). Phylogenetic analyses reveal extensive cryptic speciation and host specialization in an economically important mite taxon. Mol. Phylogenet. Evol. 66, 928–940. doi: 10.1016/j.ympev.2012.11.021

Mironov, S. V. (1996). On a validity of the genus Plicatalloptes (Acari: Proctophyllodidae) on the chaffinch Fringilla coelebs. Parazitolog. 34, 457–469.

Mironov, S. V., Stefan, L. M. (2013). Redescription of the feather mite species, Zachvatkinia from the USSR (Analogoeida: Avenzoariidae). Parazitologia. 34, 17–49.

Mironov, S. V., and Stefan, L. M. (2016). On identification of species in the feather mite genus Laminalloptes Dubinin, 1955 (Acari: Alogoidea). Acarina 24, 77–85. doi: 10.21684/0132-8077.2016.24.1.77.85

Mironov, S. V., Stefan, L. M., and González-Solís, J. (2015). Niche partitioning of feather mites within a seabird host, Calonectris borealis. PLoS ONE 10:e0144728. doi: 10.1371/journal.pone.0144728

Proctor, H. C., and Owens, I. (2000). Mites and birds: diversity, parasitism and coevolution. Trends Ecol. Evol. (Amst). 15, 358–364. doi: 10.1016/S0169-5347(00)01924-8

Ramos, R., Ramírez, I., Paiva, V. H., Militão, T., Biscoito, M., Menezes, D., et al. (2016). Global spatial ecology of three closely-related gadfly petrels. Sci. Rep. 6:23447. doi: 10.1038/srep23447

Ramos, R., Sanz, V., Militão, T., Bried, J., Neves, V. C., Biscoito, M., et al. (2015). Leapfrog migration and habitat preferences of a small oceanic seabird, Bulwer’s petrel (Bulweria bulwerii). J. Biogeogr. 42, 1651–1664. doi: 10.1111/jbi.12541

Stefan, L. M., Gómez-Díaz, E., Elguero, E., Proctor, H. C., McCoy, K. D., and González-Solís, J. (2015). Niche partitioning of feather mites within a seabird host, Calonectris borealis. PLoS ONE 10:e0144728. doi: 10.1371/journal.pone.0144728

Walther, B. A., and Clayton, D. H. (1997). Dust-ruffling: a simple method for quantifying ectoparasite loads of live birds. J. Field Ornithol. 68, 509–518.

Whiteman, N. K., Santiago-Alarcon, D., Johnson, K. P., and Parker, P. G. (2004). Phylogenetic analyses reveal extensive cryptic speciation and host specialization in chewing lice. J. Parasitol. 90, 105–120. doi: 10.1116/j.zoootaxa.3682.1.4

Whiteman, N. K., Sánchez, P., Merkel, J., Klompen, H., and Parker, P. G. (2006). Differences in straggling rates between two genera of dove lice (Insecta: Siphonaptera: Pulicidae) from petrels and shearwaters (Procellariiformes: Procellariidae). Zootaxa 3682, 105–120. doi: 10.11646/zootaxa.3682.1.4

Whiteman, N. K., Sánchez, P., Merkel, J., Klompen, H., and Parker, P. G. (2006). Sequence-based species delimitation for the DNA taxonomy of undescribed insects. Syst. Biol. 55, 595–609. doi: 10.1080/10635150600852011

Whiteman, N. K., Sánchez, P., Merkel, J., Klompen, H., and Parker, P. G. (2006). Cryptic host specificity of an avian skin mite (Epidermoptidae) vectored by louseflies (Hippoboscidae) associated with two endemic Galápagos bird species. J. Parasitol. 92, 1218–1228. doi: 10.1645/GE-918R.1

Whiteman, N. K., Sánchez, P., Merkel, J., Klompen, H., and Parker, P. G. (2006). Lineage sorting in multihost parasites: Eidmanniella albescens and Pregatella aurifasciata on seabirds from the Galapagos Islands. Evol. Ecol. 5, 3264–3271. doi: 10.1002/ece3.1587

Whiteman, N. K., Santiago-Alarcon, D., Johnson, K. P., and Parker, P. G. (2016). Global spatial ecology of three closely-related gadfly petrels. PLoS ONE 12:e0174803. doi: 10.1371/journal.pone.0174803

Whiteman, N. K., Sánchez, P., Merkel, J., Klompen, H., and Parker, P. G. (2006). Sequence-based species delimitation for the DNA taxonomy of undescribed insects. Syst. Biol. 55, 595–609. doi: 10.1080/10635150600852011

Copyright © 2018 Stefan, Gómez-Díaz, Mironov, González-Solís and McCoy. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.