Human Luteinizing Hormone and Chorionic Gonadotropin Display Biased Agonism at the LH and LH/CG Receptors

Laura Riccetti1, Romain Yvinec2, Danièle Klett2, Nathalie Gallay2, Yves Combrarnous2, Eric Reiter2, Manuela Simoni1,4,5, Livio Casarini1,4 & Mohammed Akli Ayoub2,3,6

Human luteinizing hormone (LH) and chorionic gonadotropin (hCG) have been considered biologically equivalent because of their structural similarities and their binding to the same receptor; the LH/CGR. However, accumulating evidence suggests that LH/CGR differentially responds to the two hormones triggering differential intracellular signaling and steroidogenesis. The mechanistic basis of such differential responses remains mostly unknown. Here, we compared the abilities of recombinant rhLH and rhCG to elicit cAMP, β-arrestin 2 activation, and steroidogenesis in HEK293 cells and mouse Leydig tumor cells (mLTC-1). For this, BRET and FRET technologies were used allowing quantitative analyses of hormone activities in real-time and in living cells. Our data indicate that rhLH and rhCG differentially promote cell responses mediated by LH/CGR revealing interesting divergences in their potencies, efficacies and kinetics: rhCG was more potent than rhLH in both HEK293 and mLTC-1 cells. Interestingly, partial effects of rhLH were found on β-arrestin recruitment and on progesterone production compared to rhCG. Such a link was further supported by knockdown experiments. These pharmacological differences demonstrate that rhLH and rhCG act as natural biased agonists. The discovery of novel mechanisms associated with gonadotropin-specific action may ultimately help improve and personalize assisted reproduction technologies.

Luteinizing hormone (LH) and human chorionic gonadotropin (hCG) are two heterodimeric glycoprotein hormones playing key roles in human reproduction. They are produced by the pituitary gland (LH) and placenta (hCG) and circulate as mixtures of differentially glycosylated isomers which present different half-lives and bioactivities1–3. Both hCG and hLH bind to luteinizing hormone/choriogonadotropin hormone receptor (LH/CGR) in human and LHR in non-human species which are mainly expressed in the ovary and testis. In normal female cycle, LH is involved in late follicular maturation and ovulation and it also triggers corpus luteum steroidogenic activity. hCG is responsible for maintaining steroidogenic activity by corpus luteum over the first four months of pregnancy in women. hCG also inhibits LH and FSH secretion and triggers steroidogenesis in fetal gonads4, 5. LH/CGR belongs to a subgroup of class A (rhodopsin-like) G protein-coupled receptors (GPCRs) characterized by the presence of multiple leucine-rich repeats (LRRs) in their extracellular amino-terminal domain. Regarding its signaling, LH/CGR is known to mediate the canonical G protein-mediated signaling pathway through coupling to heterotrimeric Gαs protein which activates adenylate cyclase. It results in cAMP accumulation and activation of protein kinase A (PKA), as well as in the exchange protein directly activated by cAMP (EPAC). This, in turn, triggers the activation of multiple downstream kinases that modulate the nuclear activity of CAMP response element-binding protein (CREB) and the expression of the genes involved in the physiological responses to these hormones. Besides, LH/CGR was one of the first GPCRs shown to independently activate two G proteins,

1Unit of Endocrinology, Department of Biomedical, Metabolic and Neural Sciences, University of Modena and Reggio Emilia, Modena, Italy. 2PRC, INRA, CNRS, Université François Rabelais-Tours, 37380, Nouzilly, France. 3LE STUDIUM® Loire Valley Institute for Advanced Studies, 45000, Orléans, France. 4Center for Genome Research, University of Modena and Reggio Emilia, Modena, Italy. 5Azienda, Ospedaliero-Universitaria di Modena, Modena, Italy. 6Biology Department, College of Science, United Arab Emirates University, 15551, Al Ain, United Arab Emirates. Correspondence and requests for materials should be addressed to E.R. (email: Eric.Reiter@inra.fr) or L.C. (email: livio.casarini@unimore.it) or M.A.A. (email: mayoub@uaeu.ac.ae)
leading to both adenyl cyclase and phospholipase C activation through functional coupling to Gαs and Gαq, respectively. More recently, LH/CGR has been also reported to engage a multiplicity of G protein-independent pathways including β-arrestin-dependent pathways. For many years, β-arrestins have been considered exclusively as silencers of the GPCRs signaling, prompting ligand-induced receptor internalization and trafficking. Now it is well recognized that β-arrestins regulate GPCR signaling and trafficking and are also able to engage G protein-independent signaling, the most studied of which being ERK1/2 pathway. The implication of β-arrestins in trafficking and signaling of gonadotropin receptors has also been reported.

For many years, LH and hCG have been assumed to be equivalent, even though distinct physiological, molecular, and pharmacological features were described. Importantly, the phenomenon of biased signaling implies that the binding of a given ligand can stabilize a subset of activated conformations of the receptor thereby leading to a selective modulation of downstream signaling pathways. This has been reported for numerous GPCRs. In line with this emerging concept, it has been recently proposed that LH and hCG produced as multiple glycosylated isoforms could potentially trigger selective transduction mechanisms at the LH/CGR. Recent lines of evidence support this hypothesis, showing that, although their structures are similar and they share the same receptor, LH and hCG elicit divergent signaling in several cell models. A genotoxic deletion resulting in the complete absence of exon 10 of LH/CGR found in a hypogonadal patient with type II Leydig cell hypoplasia impaired LH-, but not hCG-induced FSH production, without affecting ligand binding. More recently, it has been reported in human granulosa cells that hCG displays higher potency than LH on the cAMP/ PKA pathway as well as on steroidogenesis, whereas LH is more potent than hCG on ERK1/2 and Akt phosphorylation as well as on related gene expression. The proliferative and steroidogenic/pro-apoptotic effects mediated by LH and hCG in vitro, respectively, are amplified by FSH co-treatment. In fact, in goat ovarian granulosa cells, prolonged LH treatment promotes growth and proliferation whereas hCG leads to higher levels of cAMP and decreased proliferation.

Despite these recent advances, the potential of hCG and LH to differentially activate Gαs and β-arrestin-dependent pathways at the LH/CGR and LHR has not been evaluated. In addition, the relative contributions of these transduction mechanisms to steroidogenesis are still unknown. In the present study, we used a series of bioluminescence and time-resolved resonance energy transfer (BRET and TR-FRET) and reporter assays to quantitatively assess the signaling promoted by rhCG and rhLH in real-time and living cells as previously reported. The canonical Gαs/cAMP pathway as well as β-arrestin 2 recruitment, HEK293 cells were transiently co-transfected with plasmids coding for hLH/CGR and for Rluc8-ΔAAAGCCUUCUGUGCUGAGAAC-3′. One small RNA duplex with no silencing effect was used as a control.

**Materials and Methods**

**Recombinant gonadotropins.** Human recombinant LH (Luveris®) and rhCG (Ovitrille®) were kindly provided by Merck KGaA (Darmstadt, Germany). Molar concentrations of the recombinant rhCG and rhLH were calculated based on their respective specific activities (6500 and 5360 IU) and molecular weights (37 and 30 kDa).

**Cell Culture and Transfection.** HEK293 cells were grown in complete DMEM medium supplemented with 10% (v/v) fetal bovine serum, 4.5 g/l glucose, 100 U/ml penicillin, 0.1 mg/ml streptomycin, and 1 mM glutamine (Invitrogen, Carlsbad, CA, USA). mLTC-1 cells (ATCC CRL-2065, LCG Standards, Molsheim, France) were grown in complete RPMI medium supplemented with 10% (v/v) fetal bovine serum, 50 μg/ml gentamicin, 10 units/ml penicillin and 10 μg/ml streptomycin. Transient transfections were performed by reverse transfection in 96-well plate using Metafectene PRO (Biontient, München, Germany) following the manufacturer’s protocol and using 105 cells, 100 ng of total plasmids and 0.5 μl of Metafectene PRO per well.

**Small interfering RNA transfection.** The siRNA sequence 5′-AAAGCCUUCUGUGCUGAGAAC-3′ was used to target mouse β-arrestin 1 (position 439–459 relative to the start codon) whereas sequence 5′-AAACCGUGCCUCCGCGAUAG-3′ was used to target mouse β-arrestin 2 (position 175–193 relative to the start codon) 39. One small RNA duplex with no silencing effect was used as a control (5′-UUCCGAAACUGUGACGU-3′). The siRNAs were synthesized by GE Healthcare Dharmaco (Velizy-Villacoublay, France). Early passage mLTC-1 cells at 30% confluency in 100 mm dishes were transiently transfected with GeneSilencer following the manufacturer’s recommendations (Genlantis, San Diego, CA, USA). Forty-eight hours after transfection, cells were seeded into assay plates. All assays were performed three days after transfection.

**BRET sensors.** In order to measure cAMP real-time response in living cells, HEK293 cells were transiently transfected with two plasmids coding for hLH/CGR (kindly provided by A. Ulloa-Aguirre, Universidad Nacional Autónoma de México, Mexico) and the BRET-based cAMP sensor CAMYEL (kindly provided by L.I. Jiang, University of Texas, Texas, USA) as previously reported. For mLTC-1 cells, only CAMYEL plasmid was transfected since they naturally express the endogenous LHR.

For the assessment of β-arrestin 2 recruitment, HEK293 cells were transiently co-transfected with plasmids coding for hLH/CGR C-terminally fused to the BRET donor Rluc8 (kindly provided by Dr. A. Hanyaloglu, Imperial College, London, UK) and for β-arrestin 2 N-terminally fused to the BRET acceptor yPET (kindly provided by Dr. M.G. Scott, Cochin Institute, Paris, France). Upon rhCG and rhLH stimulation, β-arrestin 2 translocates to the receptor, leading to energy transfer between Rluc8 and yPET, and as a consequence, to dose-dependent increases in BRET signals. The conformational rearrangements within β-arrestin 2 were measured using β-arrestin 2 double brilliance sensor. For this, HEK293 cells were transiently co-transfected with the plasmids coding for hLH/CGR and for Rluc8-β-arrestin 2-RGFP fusion protein (kindly provided by Dr. R.
Jockers, Cochin Institute, Paris, France). Then, changes in intramembrane BRET signals within sensor were monitored upon cell stimulation with increasing doses of rhCG and rhLH.

**Cell stimulations and BRET measurements.** For the end-point dose-response experiments, medium was aspirated and cells were re-suspended in 40 μl/well of PBS 1X, 1 mM HEPES. Cells were incubated for 30 minutes at 37 °C in a total volume of 40 μl/well of PBS 1X, 1 mM HEPES containing or not increasing concentrations of rhCG and rhLH. BRET measurements were performed upon addition of 10 μl/well of 5 μM Coelenterazine h (Interchim, Montluçon, France), using Mithras LB 943 plate reader (Berthold Technologies GmbH & Co. Wildbad, Germany). For the real-time kinetics, cells were re-suspended in 60 μl/well of PBS 1X, 1 mM HEPES and, for cAMP kinetics, 200 μM IBMX. BRET measurement was immediately performed upon addition of 20 μl/well of EC50 concentrations of rhCG and rhLH as previously calculated in dose-response experiments, and 20 μl/well of coelenterazine h.

**Cre-dependent reporter assay.** HEK293 cells were transiently transfected with hLH/CGR and the pSOM-Luc plasmid expressing the firefly luciferase reporter gene under the control of the cAMP Responsive Element of the somatostatin promoter region. mLTC-1 cells were transiently transfected with the pSOM-Luc plasmid alone. After 48-hours, cells were split into 96-well plates. The day after, cells were stimulated 6-hours with increasing doses of hormones, then washed twice with ice-cold PBS and lysed in 200 μl of lysis buffer (Promega, Madison, WI, USA). Luciferase activity was measured using the luciferase assay system supplied by Promega. An aliquot (20 μl) of each sample was mixed with 50 μl of luciferase assay reagent and the emitted light was measured in Mithras LB 943 plate reader. Values were expressed in relative luciferase activity units (RLU).

**Steroid measurements.** Progesterone and testosterone levels were measured in the supernatants of mLTC-1 cells cultured in complete RPMI medium. For progesterone, cells were first seeded in 48-well plates (10 000 cells/well) and then re-suspended in serum-free RPMI for one hour and stimulated or not with increasing doses of rhCG and rhLH for 3 hours. Cell supernatants were collected and stored at −20 °C until analysis. Progesterone production was measured with a home-made competitive ELISA assay. Briefly, a 96-wells plate was coated overnight at 4 °C with a goat anti-mouse IgG antibody, 10 ng/well (UP462140, Interchim, Montluçon, France). After three washes in PBS 1X containing 0.1% Tween 20, non-specific sites were saturated 1 hour with 200 μl/well of PBS-Tween 20 supplemented with 0.2% BSA. Standard progesterone (Q2600, Steraloids, USA) in PBS-Tween 20-BSA or mLTC-1 cells supernatants (25 μl per well of 1:50 or 1:100 dilution) were then plated on the empty plate. Progesterone-11-Hemisuccinate-HRP (Interchim) were added, together with 36 ng/well of mouse anti-P4 antibody (AbD Serotec, Biogenesis, Interchim). The plate was incubated for 4 hours at room temperature, washed and 100 μl/well of TMB ELISA substrate standard solution (Interchim) was added. The mixture was incubated for 20 min at room temperature in the dark. The reaction was stopped with 2 N H2SO4 and absorbance was measured at 450 nm using Sunrise™ absorbance reader (Tecan, France).

Testosterone levels were measured by using an HTRF®-based assay kit (CisBio Bioassays, Codolet, France) following the manufacturer's protocol and as previously described. Briefly, cells were seeded in 96-well plate (10 000 cells/well), starved overnight in serum-free RPMI medium and incubated 3 hours at 37 °C in 100 μl/well of serum-free RPMI medium containing or not increasing concentrations of rhCG and rhLH. Then 10 μl of culture supernatants were transferred into 384-well and mixed with 10 μl of a mixture of HTRF mix containing an anti-testosterone antibody and testosterone labeled with Terbium and d2 fluorophores, respectively. TR-FRET signals were detected using Mithras LB 943 plate reader (Berthold Technologies GmbH & Co. Wildbad, Germany).

**cAMP HTRF-based assay.** Intracellular cAMP levels were measured using a homogeneous time-resolved fluorescence (HTRF®) assay kit (CisBio Bioassays) according to the manufacturer's instructions.

**Data analysis.** BRET data were represented as “Induced BRET Changes” by subtracting the 540 nm/480 nm ratio of the non-treated cells from the same ratio of cells stimulated with increasing doses of rhCG and rhLH. The % of responses in BRET and steroid measurement were obtained by taking as 100% the maximal response of rhCG at 10 nM in the different assays. All the results were fitted following the appropriate nonlinear regression equations using GraphPad Prism software (San Diego, CA, USA). Statistical analyses were performed with unpaired t-test (for the Emax and EC50 values) in HEK293 and mLTC-1 cells and Two-way ANOVA (for siRNA β-arrestin data in mLTC-1 cells), both using the GraphPad Prism software.

**Bias calculation.** The bias factor (B.F.) was obtained after statistical fitting of the data to the operational model reported by Black and Left7, as previously reported.

**Results**

**LH/CGR-promoted cAMP Response.** To compare the respective efficacies and potencies of rhCG and rhLH, we first assessed the ability of either hormone to elicit the accumulation of cAMP, the prototypical second messenger produced upon coupling and activation of the heterotrimeric Gs protein by LH/CGR. For this purpose, HEK293 cells transiently co-expressing human LH/CGR and the BRET-based cAMP sensor (CAMYEL) were used as previously reported. Changes in BRET signal were monitored in living cells after 30 minutes of incubation with increasing doses of rhCG and rhLH. As expected, both hormones showed very potent effects on the cAMP signaling pathway with EC50 values in the pM range as previously shown (Fig. 1A) (Table 1). Consistent with previous reports, the rhCG dose-response curve is significantly shifted towards the lower doses compared to that of rhLH. Indeed, the EC50 values of rhCG response was found approximately 16 times lower than that of rhLH (p < 0.0001; n = 7) (Table 1). By contrast, at the saturating doses both hormones promoted similar maximal cAMP responses indicating that both act as full agonists on LH/CGR-promoted cAMP production.
We also investigated cAMP accumulation in real-time kinetics by stimulating cells with different concentrations (0.05, 0.5, and 5 nM) of rhCG and rhLH to cover the EC\textsubscript{50} and sub-maximal doses according to the dose curves shown in Fig. 1A. As shown in Fig. 1B, a relationship between the doses and the kinetics was observed with faster and saturating responses obtained when increasing doses were used. Indeed, when rhCG and rhLH were compared at their respective sub-saturating doses (i.e.: 0.01 nM for rhCG and 0.5 nM for rhLH), both promoted a rapid cAMP response with no difference in their kinetics (t\textsubscript{1/2} around 1 min) (Fig. 1B). The response reached a plateau even after 30 minutes of stimulation (Fig. 1B) and this cannot be due to a saturation of the cAMP response in our system. In fact, at a higher and saturating dose of both hormones (5 nM), the cAMP response was further increased with faster (t\textsubscript{1/2} value around 0.5 min) and equivalent kinetics for both hormones (Fig. 1B).

To investigate the cAMP pathway on endogenously expressed LHR, we used a mouse Leydig tumor cell line (mLTC-1) either transfected with CAMYEL sensor (Fig. 1C) or not (Fig. 1D), as previously reported\textsuperscript{10, 11}. Again, BRET measurements revealed rhCG dose-response curve shifted towards the lower doses compared to rhLH (Fig. 1C). Specifically, the EC\textsubscript{50} value of rhCG was approximately 6 times lower than that of rhLH (p = 0.0067; n = 5) (Table 2). To further confirm these data and to exclude any artifact due to the transfection of CAMYEL sensor in mLTC-1 cells, we also measured cAMP levels in native cells using HTRF-based assay, which does not require any cell transfection. We found a similar pattern of rhCG- and rhLH-induced cAMP responses (Fig. 1D).

**Table 1.** Efficacy (E\textsubscript{max}) and efficiency (EC\textsubscript{50}) of rhCG and rhLH on LH/CGR measured in different assays in HEK293 cells. The E\textsubscript{max} values are represented as hormone-induced BRET changes (for cAMP and ß-arrestin 2) and relative luciferase activity \times 10\textsuperscript{3} (for Cre-reporter assay). Statistical analyses were performed with unpaired t-test (**p < 0.01; *p < 0.05, ns non-significant).
It is worth noting that consistent with HEK293 cell data, rhCG and rhLH promoted similar maximal cAMP responses in mLTC-1 cells despite the differences in their potencies confirming that both are full agonists for cAMP pathway (Fig. 1C and D) (Table 2).

**Table 2.** Efficacy ($E_{\text{max}}$) and efficiency ($EC_{50}$) of rhCG and rhLH on LHR measured in different assays in mLTC-1 cells. The $E_{\text{max}}$ values are represented as hormone-induced BRET changes (for cAMP by BRET), % of rhCG-induced response (for cAMP by HTRF, progesterone and testosterone) and relative luciferase activity (for Cre-reporter assay). Statistical analyses were performed with unpaired t-test (**$p < 0.001$; *$p < 0.01$; *$p < 0.05$, ns non-significant).

| Responses               | rhCG          | rhLH          |
|-------------------------|---------------|---------------|
|                         | $E_{\text{max}}$ | $EC_{50}$    | $E_{\text{max}}$ | $EC_{50}$    |
| cAMP (BRET)             | 0.45 ± 0.02   | 68.82 ± 22.30 pM  | 5 | 0.44 ± 0.02   | 459 ± 105.35 pM**  | 5 |
| cAMP (HTRF)             | 100%          | 97.37 ± 47.15 pM  | 3 | 102 ± 5.8%ns | 1980.33 ± 686.64 pM **  | 3 |
| Cre-reporter assay      | 1910 ± 111    | 24.97 ± 16.20 pM  | 4 | 1971 ± 168** | 317.62 ± pM **  | 4 |
| Progesterone            | 100%          | 19.72 ± 4.28 pM  | 3 | 49.78 ± 2.90% *** | 305.83 ± 25.22 pM **  | 3 |
| Testosterone            | 100%          | 2.19 ± 0.91 pM   | 4 | 93.33 ± 6.5% ** | 29.05 ± 6.90 pM **  | 4 |

**Figure 2.** LH/CGR-promoted β-arrestin 2 recruitment and activation in HEK293 cells. HEK293 cells transiently co-expressing either hLH/CGR-Rluc8 and yPET-β-arrestin 2 (A and B) or wildtype hLHR/CGR and Rluc8-β-arrestin 2-RGFP (C) were stimulated 30 minutes with increasing doses (A and D) or immediately with the indicated doses (B) or 0.25 µM (C) of rhCG and rhLH before BRET was measured. Data are means ± SEM of 3–5 independent experiments. The kinetic curves in panel B are representative of 3 experiments performed in triplicate.

It is worth noting that consistent with HEK293 cell data, rhCG and rhLH promoted similar maximal cAMP responses in mLTC-1 cells despite the differences in their potencies confirming that both are full agonists for cAMP pathway (Fig. 1C and D) (Table 2).

**β-arrestin 2 recruitment and activation.** β-arrestins are known to play important roles not only in desensitization/internalization of GPCRs but also in their signaling. Here, we examined the recruitment of β-arrestin 2 upon exposure to increasing doses of rhCG and rhLH using BRET technology as previously reported. HEK293 cells were transiently co-transfected with plasmids coding for hLH/CGR-Rluc8 and for yPET-β-arrestin 2 fusion proteins and BRET measurements were performed in a dose-dependent manner and in living cells. As shown in Fig. 2A, rhCG and rhLH stimulation significantly promoted β-arrestin 2 recruitment to hLH/CGR in a dose-dependent manner with the $EC_{50}$ value of rhCG 12 times lower than that of rhLH ($p = 0.044$; $n = 5$) (Table 1). The significant difference in the two hormone potencies is consistent with the data on cAMP pathway (Fig. 1) (Table 1). Low potencies were found for both hormones on β-arrestin recruitment compared to cAMP responses consistent with our previous report indicating that a higher receptor occupancy rate is needed.
to engage β-arrestin recruitment. Interestingly, our data reveal that rhLH seems to exhibit partial agonistic activity compared to rhCG ($p = 0.0104; n = 5$) (Fig. 2A) (Table 1). Real-time kinetic analysis of BRET signals in cells stimulated with either EC$_{50}$ (10 nM and 50 nM) or saturating (500 nM) concentrations of both rhCG and rhLH, showed a significant difference in β-arrestin 2 recruitment kinetics between the two hormones with $t_{1/2}$ values of $6.15 \pm 0.17$ min and $13.32 \pm 0.32$ min with 500 nM of rhCG and rhLH, respectively (Fig. 2B). These kinetic data are in keeping with the partial activity of rhLH compared to rhCG on β-arrestin recruitment even at the saturating doses of the hormone (Fig. 2B).

Next, we investigated the impact of hLH/CGR activation by rhCG or rhLH on β-arrestin 2 conformations using a previously reported double brilliance β-arrestin 2 BRET sensor. In this sensor, β-arrestin 2 is fused with both a BRET donor and acceptor. Therefore, any change in the intramolecular BRET signals reflects a conformational change in β-arrestin 2. Interestingly, real-time kinetic analyses with 0.25 µM of hormones showed stronger induction of conformational changes under LH than rhCG treatment within β-arrestin sensor (Fig. 2C). However, no significant difference in the half-time (around 12 minutes) between the two hormones was observed (Fig. 2C). Such a difference between rhCG and rhLH on β-arrestin conformation was observed at different concentrations of the hormones (Fig. 2D). Together, the data with β-arrestins are consistent with the different sensitivity of LH/CGR to rhCG and rhLH suggesting the existence of hormone-specific receptor conformation which could in turn impact β-arrestin conformation.

**LHR-promoted integrated responses.** To further assess the impact of the differences elicited by the two hormones at the transductional level, we sought to measure downstream read-outs in the signaling pathways. In HEK293 cells, we used a cre-dependent reporter gene, pSOM-Luc, as an indicator of LH/CGR-induced transcriptional activation. As shown in Fig. 3A, both hormones showed clear dose-dependent activation of luciferase activity with EC$_{50}$ of rhCG almost 5 times lower than that of rhLH ($p = 0.0004; n = 3$) (Table 1). By contrast, the two hormones showed similar maximal responses expressed in relative luciferase activity (Fig. 3A) (Table 1). These observations are consistent with the cAMP data in HEK293 and mLTC-1 cells as shown in Fig. 1.

Next, we explored more distal responses to rhCG and rhLH in mLTC-1 cells endogenously expressing LHR. For this, we measured pSOM-Luc activation in Cre reporter assay (Fig. 3B), progesterone (Fig. 3C) and testosterone (Fig. 3D) responses upon stimulation with increasing doses of rhCG and rhLH. In the Cre reporter assay, we observed a significant shift ($p = 0.0202; n = 4$) similar to the one observed in HEK293 cells with rhCG being more...
potent than rhLH, (Fig. 3B) (Table 2). In progesterone assay, we found that the EC_{50} of rhCG was approximately 15 times lower than that of rhLH (p = 0.0011; n = 3) (Fig. 3C) (Table 2). This observation is in agreement with cAMP data (Fig. 1), β-арестин data (Fig. 2A), and Cre-luciferase-based cAMP assay (Fig. 3A) data confirming that rhCG is more potent than rhLH on LH/CGR. Interestingly, the maximal progesterone produced by rhLH was only 49.8 ± 2.9% (p < 0.0001; n = 3) of that promoted by rhCG, indicating a partial response of rhLH on progesterone synthesis (Fig. 3C) (Table 2). Such a partial response of rhLH is consistent with that observed for LH/CGR-promoted β-арестин 2 recruitment (Fig. 2A). In the case of testosterone, we also observed a higher potency of rhCG (≈13 folds) compared to rhLH, (p = 0.0084; n = 4) (Fig. 3D) (Table 2), which is consistent with progesterone data (Fig. 3C), cAMP responses in HEK293 and mLTC-1 cells (Fig. 1A and C) as well as β-арестин in HEK293 cells (Fig. 2A). We also noticed that both hormones were clearly more potent at activating testosterone than progesterone production (Table 2). However, by contrast to progesterone response, there was no significant difference in the maximal testosterone response between the two hormones (p = 0.3848; n = 4) (Fig. 3D). Together, this illustrates the complexity of the steroidogenic pathways engaged by LHR, suggesting that distinct signaling pathways might control the production of progesterone and testosterone as previously proposed^{11}.

**Implication of β-арестинs in steroid production.** The similarity in the partial effects of rhLH on β-арестин (Fig. 2A) and progesterone (Fig. 3C) responses compared to rhCG suggests that β-арестинs may be implicated in progesterone production. Therefore, we examined the contribution of β-арестин-dependent transduction on the control of LHR-mediated steroidogenesis using siRNA-mediating depletion of endogenous β-арестин 1 or β-арестин 2 in mLTC-1 cells. For this, control, β-арестин 1 or β-арестин 2-depleted mLTC-1 cells were exposed to increasing doses of rhCG (Fig. 4A-C and E) and rhLH (Fig. 4B-D and F). Progesterone and testosterone were measured in parallel in the same cells. Our data confirm the partial progesterone response mediated by rhLH compared to rhCG as shown in Fig. 3B. However, on testosterone production there was no difference in hormone efficacy but rhCG was more potent that rhLH confirming the data shown in Fig. 3D. Interestingly, we observed that the depletion of both β-арестин 1 and β-арестин 2 led to a partial decrease in the progesterone production induced by rhCG (Fig. 4A) and rhLH (Fig. 4B) compared to control siRNA-transfected mLTC-1 cells. The statistical analysis demonstrates the significance of β-арестин depletion for both rhCG (p < 0.0001 for siRNA β-арестин 1 and p = 0.0233 for siRNA β-арестин 2; n = 3) and rhLH (p = 0.001 for siRNA β-арестин 1 and p = 0.0275 for siRNA β-арестин 2; n = 3) with siRNA β-арестин 1 being more efficient. Moreover, β-арестин depletion showed stronger inhibitory effects on testosterone production mediated by either rhCG (Fig. 4C) or rhLH (Fig. 4D) (p < 0.0001 for siRNA β-арестин 1 and β-арестин 2; n = 3). The specificity of the effects of β-арестин depletion on steroidogenesis was demonstrated by measuring cAMP production in parallel in the siRNA-transfected mLTC-1 cell samples. As anticipated, the depletion of β-арестин 1 or 2 had no effect on rhCG (Fig. 4E) or rhLH (Fig. 4F) mediated cAMP production whatever the dose of the hormones used, thus ruling out toxic or side effects that the siRNA may have exerted on these cells. Together, these data suggest the partial implication of β-арестинs in LHR-mediated progesterone and argue for a significance of β-арестин bias between rhCG and rhLH in HEK293 cells.

**Calculation of biases between rhCG and rhLH.** Biases were calculated using data-fitting of the operational model^37 and the procedure previously detailed^11, 38. Bias factor combines both efficacy and potency to quantify the imbalance between two cell responses for an agonist, in comparison to a reference agonist on the same receptor and within the same cell model. For this, we used rhCG as the reference ligand for the different assays in both HEK293 and mLTC-1 cells (Table 3). A bias exists if the bias factor is significantly different from 1 (unpaired t-test). In HEK293 cells, we found that rhLH, compared to rhCG, was significantly biased towards cre-dependent reporter gene against both β-арестин 1 and cAMP (Table 3). To some extent, rhLH is also biased towards β-арестин 2 against cAMP (Table 3). In other words, in HEK293 cells, rhLH preferentially induces Cre-dependent transcription > β-арестин 2 recruitment > cAMP, compared to rhCG. This finding may be considered counter-intuitive since Cre reporter assay and cAMP are connected by the activation of cAMP response element-binding protein (CREB) which is primarily under the control of cAMP/PKA pathway. However, it is also well documented that CREB activation integrates other signaling pathways such as ERK and p38 MAPKs, p90RSK or CAMKs. In that sense, Cre reporter assay can be viewed as a read out which integrates several signalling pathways, not just cAMP/PKA. In mLTC-1 cells, rhLH-promoted response was significantly biased towards cAMP compared to progesterone and testosterone and a moderate bias appeared towards testosterone compared to progesterone (Table 2). However, by contrast to progesterone response, there was no significant difference in the maximal testosterone response between the two hormones (p = 0.3848; n = 4) (Fig. 3D). Together, these data suggest the partial implication of β-арестинs in LHR-mediated progesterone and argue for a significance of β-арестин bias between rhCG and rhLH in HEK293 cells.

**Discussion**
In the present work, we investigated the differential activity exerted by rhCG and rhLH upon binding to their common receptor, human LH/CGR transiently expressed in HEK293 cells or mouse LHR endogenously expressed in mLTC-1 cells. The attention was focused on comparing their pharmacological profiles on the heterotrimeric G proteins, Gαs/cAMP pathway as well as β-арестинs and their contribution to signal transduction mechanisms leading to the modulation of steroidogenesis. For cAMP, rhCG was more potent than rhLH but both hormones promoted full activation of the receptor at saturating doses in HEK293 as well as mLTC-1 cells. A similar difference in potency was also observed on β-арестин recruitment in HEK293 cells, however an interesting difference was also found in hormone efficacy since rhLH promoted only a partial response. We also examined the action of rhLH and rhCG on progesterone and testosterone production in mLTC-1 cells confirming the higher potency of rhCG compared to rhLH. Interestingly, we found that rhLH exhibited a partial activity for progesterone compared to rhCG while both fully promoted testosterone production. Altogether, our data show...
that rhCG is more potent than rhLH on LH/CGR activity in HEK293 and mLTC-1 cells which is consistent with previous studies in COS-7 and granulosa cells. Moreover, the partial agonism of rhLH on LH/CGR on both β-arrestin recruitment and progesterone production suggests a link between these two events. It can be observed that rhCG is more potent than rhLH on LH/CGR activity in HEK293 and mLTC-1 cells which is consistent with previous studies in COS-7 and granulosa cells. Moreover, the partial agonism of rhLH on LH/CGR on both β-arrestin recruitment and progesterone production suggests a link between these two events. It can be observed
that the minimal doses of rhCG or rLH promoting full testosterone responses trigger only very partial (20–25%) progesterone response. It reflects the physiological role of hCG, which serves to induce progesterone synthesis, while LH is a mediator of testosterone production. Indeed, in Leydig cells, androgen synthesis occurs mainly via the so-called Δ5-pathway, while progesterone is a “parallel co-translation product” falling within the relatively ineffective Δ4-pathway. We could speculate that, differently to hCG, LH is fully active on the Δ5- rather than Δ4-pathway. The results provided by β-arrrestins silencing by siRNA (Fig. 4) agree with this view, since these molecules are required to fully support LH/hCG-mediated testosterone but not progesterone synthesis.

In the present study, we are using immortalized cell lines as model systems. It is important to determine to what extent the data obtained in these cells can be indicative of the physiological mechanisms that occur in the ovary. In fact, the transduction mechanisms occurring directly downstream of the ligand-LH/CGR complex, such as Gαs coupling and cAMP production or β-arrrestins recruitment, have been reported to be mostly reproducible across a wide range of cell models. This is corroborated by previous observation in human and goat primary granulosa cells, in transfected COS-7 and human immortalized granulosa cell lines, as well as in mouse primary Leydig cells. In all these cell systems, hCG treatment results in higher cAMP production than LH. One can rationalize that these mechanisms largely rely on the ligand and receptor amino acid sequence, conformational changes, etc., all properties that are nearly identical in granulosa and Leydig cells, as well as in transfected cell lines. Indeed, we demonstrated that hCG is more potent than LH, in terms of cAMP production, in all these cell models. The amino-acid changes existing between the rodent and the human receptor have been shown to have minimal, if any, effects on hLH or hCG binding rates. According to this principle, downstream outputs, which are located farther from the ligand-receptor complex, may lead to more cell type-dependent variations. This is indeed the case when comparing the mLTC-1 cell line used in the present paper and the primary mouse Leydig cells, which were used in another recently published study. In primary mouse Leydig cells, we demonstrated that LH and hCG treatment results in different cAMP production but equal testosterone dose-response curves, whereas in mLTC-1 cells, hCG is more potent than LH at inducing testosterone production. Taken together, these findings indicate that hCG has a higher steroidogenic potential than LH, even if, in physiological settings, their effects may differ as a consequence of tissue-specific modulatory events.

From the molecular point of view, LH- and hCG-induced signaling is known to be differently modulated by exon 10 deletion in LH/CGR, which results in structural and spatial rearrangements at the hinge region of the receptor. In the presence of this deletion, LH signaling is impaired while hCG signaling remain unchanged, suggesting divergences between hCG- and LH-receptor interactions and actions. In addition, hCG and LH were recently shown to interact differently with the hinge region of the receptor and only hCG is capable of inducing both cis- and trans-activation of human LH/CGR. Moreover, both hCG and LH were reported to have similar association rate on rat LH (3 × 10^9 M^-1.min^-1) but different dissociation rate (25 h for hCG and ~9 h for hLH) constants indicating higher residence time of hCG on its receptor. Such a finding was recently supported by dissociation assays on gonadotropin-promoted cAMP production in mLTC-1, suggesting weaker dissociation of hCG from LHR compared to hLH. Furthermore, our real-time cAMP measurements clearly showed differences in the kinetics of the two hormones with rhCG inducing faster cAMP responses than rhLH. This indicates the relationship between the difference in hormone potencies and the kinetics of their induced responses, suggesting a link between pharmacological bias and response kinetics as recently reported. Collectively, these observations suggest that the slower dissociation rate of rhCG compared to that of rhLH and/or the differences in the conformations of hormone-receptor complexes may explain the differences between rhCG- and rhLH-promoted responses.

Our quantitative pharmacological profiling revealed striking peculiarities when comparing the maximal responses elicited by the two gonadotropins on the different readouts. Indeed, even though identical maximal cAMP responses were reached with either gonadotropin, whatever the cell type (HEK293 and mLTC-1), rLH led to significantly weaker maximal β-arrestin 2 recruitment and progesterone production than rhCG. Thus, it could be hypothesized that β-arrestin is a limiting factor for maximal progesterone production. In other words, rLH like rhCG is full agonist on LH/CGR for cAMP and testosterone production, whereas it is only partial agonist for β-arrestin recruitment and progesterone production. The fact that rLH was full agonist on testosterone and had only a partial efficacy on progesterone further demonstrates that the full action of the hormone, in Leydig cells, is exerted via Δ5-pathway, through 17-OH-pregnenolone production (instead of progesterone) as a precursor of testosterone. This is also supported by our recent study using two FSHR and LH negative allosteric modulators (NAMs) showing differential antagonism of the two NAMs on progesterone and testosterone. Moreover, we observed that both hormones differently affected the conformation of β-arrestin 2 as assessed in double brilliance BRET assay used as conformational sensor of β-arrestins. In this assay, rhCG elicited less BRET changes within the β-arrestin 2 sensor than rLH which is consistent with the idea that the two hormones stabilize different conformations of β-arrestin as shown for other GPCRs depending on the ligands applied.

Together, these observations strongly suggest that rhCG and rLH differentially recruit β-arrestins and activate downstream pathways which may control progesterone production. We hypothesized that β-arrestin-dependent transduction could be involved in the control of the balance between progesterone and testosterone production as recently reported. Supporting this view, we demonstrated that depletion of endogenous β-arrestins in mLTC-1 cells using selective siRNA led indeed to partially but significantly reduced progesterone production. This supports the link between the partial agonism of rLH on both β-arrestin and progesterone production. In contrast, such depletion had no effect on hormone-promoted cAMP production in mLTC-1 excluding a role of β-arrestins in cAMP production as expected. Altogether, our results support to the concept of biased agonism exerted by rhCG and rLH and bear the notion that LH/CGR can discriminate the binding of the two hormones, thereby triggering different transduction mechanisms hence intracellular responses. In addition to the pharmacological profiling of rhCG and rLH, our study and the bias analysis reveal the importance to quantify the balance between the different signaling pathways for each hormone. The fact that both hormones naturally coexist but to
different extent and at different stages during folliculogenesis and pregnancy, raises intriguing prospects. The use of these hormones in assisted reproduction could also be impacted by the present findings. Finally, the impact of our study goes beyond the gonadotropin receptors and reproduction since it describes an interesting example of biased signaling involving two endogenous hormones in assisted reproduction. This further demonstrates the physiological relevance of the signaling bias at GPCRs with strong impact on our understanding of GPCR pharmacology and signaling and potential applications in drug discovery programs.

References
1. Arey, B. J. & Lopez, F. J. Are circulating gonadotropin isoforms naturally occurring biased agonists? Basic and therapeutic implications. Rev Endocr Metab Disord 12, 275–288, doi:10.1007/s11154-011-9188-y (2011).
2. Bousfield, G. R. & Dias, J. A. Synthesis and secretion of gonadotropins including structure-function correlates. Rev Endocr Metab Disord 12, 289–302, doi:10.1007/s11154-011-9191-3 (2011).
3. Leão, R. B. & Esteves, S. C. Gonadotropin therapy in assisted reproduction: an evolutionary perspective from biologics to biotech. Clinics (Sao Paulo) 69, 279–293 (2014).
4. Ascoli, M., Fanelli, F. & Segaloff, D. L. The lutropin/choriogonadotropin receptor, a 2002 perspective. Gynecol Endocrinol 30, 174–181, doi:10.3109/095135590.2013.859670 (2014).
5. Gudermann, T., Birnbaumer, M. & Birnbaumer, L. Evidence for dual coupling of the murine luteinizing hormone receptor to adenylyl cyclase and phosphoinositide breakdown and Ca2+ mobilization. Studies with the cloned murine luteinizing hormone receptor expressed in L cells. J Biol Chem 267, 4479–4488 (1992).
6. Gudermann, T., Nichols, C., Levy, F. O., Birnbaumer, M. & Birnbaumer, L. Ca2+ mobilization by the LH receptor expressed in Xenopus oocytes independent of 3', 5'-cyclic adenosine monophosphate formation: evidence for parallel activation of two signaling pathways. Mol Endocrinol 6, 272–278, doi:10.1210/mend.6.2.1314958 (1992).
7. Ulloa-Aguirre, A., Crepieux, P., Poupou, A., Maurel, M. C. & Reiter, E. Novel pathways in gonadotropin receptor signaling and biased agonism. Rev Endocr Metab Disord 12, 259–274, doi:10.1007/s11154-011-9176-2 (2011).
8. Nakamura, K., Lazari, M. F., Korgaonkar, C. & Ascoli, M. Role of the rate of internalization of the agonist-receptor complex on the agonist-induced down-regulation of the lutropin/choriogonadotropin receptor. Molecular endocrinology 13, 1295–1304, doi:10.1210/mend.13.8.0331 (1999).
9. Ayoub, M. A. et al. Assessing Gonadotropin Receptor Function by Resonance Energy Transfer-Based Assays. Front Endocrinol (Lausanne) 6, 130, doi:10.3389/fendo.2015.00130 (2015).
10. Ayoub, M. A. et al. Profiling of FSHR negative allosteric modulators on LH/CGR reveals biased antagonism with implications in steriodogenesis. Molecular and cellular endocrinology 436, 10–22, doi:10.1016/j.mce.2016.07.013 (2016).
11. Lefkowitz, R. J. & Shenoy, S. K. Transduction of receptor signals by beta-arrestins. Science. 308, 512–517 (2005).
12. Reiter, E. & Lefkowitz, R. J. GRKs and beta-arrestins: roles in receptor silencing, trafficking and signaling. Trends Endocrinol Metab 17, 159–165 (2006).
13. Reiter, E., Ahn, S., Shukla, A. K. & Lefkowitz, R. J. Molecular mechanism of beta-arrestin-biased agonism at seven-transmembrane receptors. Annu Rev Pharmacol Toxicol 52, 179–197, doi:10.1146/annurev-pharmtox-010909.105800 (2012).
14. Casarini, L., Reiter, E. & Simoni, M. beta-arrestins regulate gonadotropin receptor-mediated cell proliferation and apoptosis by controlling different FSHR or LHCGR intracellular signaling in the hGL5 cell line. Molecular and cellular endocrinology 437, 11–21, doi:10.1016/j.mce.2016.08.005 (2016).
15. Galden, C. & Ascoli, M. Arrestin-3 is essential for the activation of Fyn by the luteinizing hormone receptor (LHR) in MA-10 cells. Cellular signalling 20, 1822–1829, doi:10.1016/j.cellsig.2008.06.005 (2008).
16. Krishnamurthy, H., Galet, C. & Ascoli, M. The association of arrestin-3 with the follitropin receptor depends on receptor activation and phosphorylation. Molecular and cellular endocrinology 204, 127–140 (2003).
17. Neill, J. D., Duck, L. W., Musgrove, L. C. & Sellers, J. C. Potential regulatory roles for G protein-coupled receptor kinases and beta-arrestins in gonadotropin-releasing hormone receptor signaling. Endocrinology 139, 1781–1788 (1998).
18. Webb, V. et al. Partially deglycosylated equine LH preferentially activates beta-arrestin-dependent signaling at the follicle-stimulating hormone receptor. Mol Endocrinol 24, 561–573, doi:10.1210/me.2010-0347 (2010).
19. Tranchant, T. et al. Preferential beta-arrestin signalling at low receptor density revealed by functional characterization of the human FSH receptor A189 V mutation. Molecular and cellular endocrinology 331, 109–118, doi:10.1016/j.mce.2010.08.016 (2011).
20. Kara, E. et al. A phosphorylation cluster of five serine and threonine residues in the C-terminus of the follicle-stimulating hormone receptor is important for desensitization but not for beta-arrestin-mediated ERK activation. Molecular endocrinology 20, 3014–3026, doi:10.1210/me.2006-0098 (2006).
21. Khan-Sabir, N., Beshay, V. E. & Carr, B. R. In Endotext Vol. 3 (2008).
22. Huhtianniemi, I. T. & Catt, K. J. Differential binding affinities of rat testis luteinizing hormone (LH) receptors for human chorionic gonadotropin, human LH, and ovine LH. Endocrinology 108, 1931–1938, doi:10.1210/endo-108-5-1931 (1981).
23. Galet, C. & Ascoli, M. The differential binding affinities of the luteinizing hormone (LH)/choriogonadotropin receptor for LH and choriongonadotropin are dictated by different extracellular domain residues. Mol Endocrinol 19, 1263–1276, doi:10.1210/mend.2004-0040 (2005).
24. Grzesik, P. et al. Differences in Signal Activation by LH and hCG are Mediated by the LH/CG Receptor’s Extracellular Hinge Region. Front Endocrinol (Lausanne) 6, 140, doi:10.3389/fendo.2015.00140 (2015).
25. Grzesik, P. et al. Differences between lutropin-activated and choriogonadotropin-activated receptor activation. FEBS J 281, 1479–1492, doi:10.1111/febs.12718 (2014).
26. Choi, J. & Smitz, J. Luteinizing hormone and human chorionic gonadotropin: origins of difference. Molecular and cellular endocrinology 383, 203–213, doi:10.1016/j.mce.2013.12.009 (2014).
27. Galandrini, S., Oligny-Longpre, G. & Bouvier, M. The evasive nature of drug efficacy: implications for drug discovery. Trends Pharmacol Sci 28, 423–430 (2007).
28. Kenakin, T. Functional selectivity through protein and biased agonism: who steers the ship? Mol Pharmacol 72, 1393–1401, doi:10.1124/mol.107.040352 (2007).
29. Landomiel, F. et al. Biased signalling in follicle stimulating hormone action. Molecular and cellular endocrinology 382, 452–459, doi:10.1016/j.mce.2013.09.035 (2014).
30. Gromoll, J., Eiholzer, U., Nieschlag, E. & Simon, M. Male hypogonadism caused by homozygous deletion of exon 10 of the luteinizing hormone (LH) receptor: differential action of human chorionic gonadotropin and LH. J Clin Endocrinol Metab 85, 2281–2286, doi:10.1210/jcem.85.6.66636 (2000).
31. Muller, T., Gromoll, J. & Simon, M. Absence of exon 10 of the human luteinizing hormone (LH) receptor impairs LH, but not human chorionic gonadotropin action. J Clin Endocrinol Metab 88, 2242–2249, doi:10.1210/jc.2002-021946 (2003).
32. Casarini, L. et al. LH and hCG action on the same receptor results in quantitatively and qualitatively different intracellular signalling. PLoS One 7, e46682, doi:10.1371/journal.pone.0046682 (2012).
34. Casarini, L. et al. Follicle-stimulating hormone potentiates the steroidogenic activity of chorionic gonadotropin and the anti-apoptotic activity of luteinizing hormone in human granulosa-lutein cells in vitro. *Molecular and cellular endocrinology* **422**, 103–114, doi:10.1016/j.mce.2015.12.008 (2016).

35. Gupta, C. et al. Differential response to sustained stimulation by hCG & LH on goat ovarian granulosa cells. *The Indian journal of medical research* **135**, 331–340 (2012).

36. Klett, D. et al. Low reversibility of intracellular cAMP accumulation in mouse Leydig tumor cells (MLTC-1) stimulated by human Luteinizing Hormone (hLH) and Chorionic Gonadotropin (hCG). *Molecular and cellular endocrinology* **434**, 144–153, doi:10.1016/j.mce.2016.06.028 (2016).

37. Black, J. W. & Leff, P. Operational models of pharmacological agonism. *Proc R Soc Lond B Biol Sci* **220**, 141–162 (1983).

38. van der Westhuizen, E. T., Breton, B., Christopoulos, A. & Bouvier, M. Quantification of ligand bias for clinically relevant beta2-adrenergic receptor ligands: implications for drug taxonomy. *Molecular Pharmacology* **85**, 492–509, doi:10.1124/mol.113.088880 (2014).

39. Kovacs, J. J. et al. Beta-arrestin-mediated localization of smoothened to the primary cilium. *Science* **320**, 1777–1781, doi:10.1126/science.1157983 (2008).

40. Charest, P. G., Terrillon, S. & Bouvier, M. Monitoring agonist-promoted conformational changes of beta-arrestin in living cells by intramolecular BRET. *EMBO Rep* **6**, 334–340, 7400373 [pii] doi:10.1038/sj.embor.7400373 (2005).

41. Lee, M. H. et al. The conformational signature of beta-arrestin2 predicts its trafficking and signalling functions. *Nature* **531**, 665–668, doi:10.1038/nature17154 (2016).

42. Kamal, M. et al. Improved donor/acceptor BRET couples for monitoring beta-arrestin recruitment to G protein-coupled receptors. *Biotchnol J*., doi:10.1002/biot.200900016 (2009).

43. Troispoux, C. et al. Involvement of G protein-coupled receptor kinases and arrestins in desensitization to follicle-stimulating hormone action. *Molecular endocrinology* **13**, 1599–1614, doi:10.1210/me.2013-1314 (1999).

44. Canepa, S. et al. Validation d’une méthode immunoenzymatique pour le dosage de la progesterone dans le plasma des ovins et des bovins. *Cahiers techniques de l’INRA* **64**, 19–30 (2008).

45. Rebois, R. V. Establishment of gonadotropin-responsive murine leydig tumor cell line. *The Journal of cell biology* **94**, 70–76 (1982).

46. Miller, W. L. & Auchus, R. J. The molecular biology, biochemistry, and physiology of human steroidogenesis and its disorders. *Endocrine Reviews* **32**, 81–151, doi:10.1210/er.2010-0013 (2011).

47. Riccetti, L. et al. Human LH and hCG stimulate differently the early signalling pathways but result in equal testosterone synthesis in mouse Leydig cells in vitro. *Reprod Biol Endocrinol.* **15**, 2, doi:10.1186/s12995-016-0224-3 (2017).

48. Klein Herenbrink, C. et al. The role of kinetic context in apparent biased agonism at GPCRs. *Nat Commun* **7**, 10842, doi:10.1038/ncomms10842 (2016).

49. Shukla, A. K. et al. Distinct conformational changes in beta-arrestin report biased agonism at seven-transmembrane receptors. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 9988–9993, doi:10.1073/pnas.0804246105 (2008).

50. Nuber, S. et al. beta-Arrestin biosensors reveal a rapid, receptor-dependent activation/deactivation cycle. *Nature* **531**, 661–664, doi:10.1038/nature17198 (2016).

51. Kenakin, T. & Christopoulos, A. Signalling bias in new drug discovery: detection, quantification and therapeutic impact. *Nature reviews. Drug discovery* **12**, 205–216, doi:10.1038/nrd3954 (2013).

52. Luttrell, L. M. Minireview: More than just a hammer: ligand “bias” and pharmaceutical discovery. *Molecular endocrinology* **28**, 281–294, doi:10.1210/me.2013-1314 (2014).

Acknowledgements
This work has been funded by Région Centre ARD2020 Biomédicaments. L.R. is funded by Associazione Scientificia in Endocrinologia, Andrologia e Metabolismo and Erasmus+ Grant. M.A.A. is funded by LE STUDIUM® Loire Valley Institute for Advanced Studies and AgreenSkills Plus.

Author Contributions
L.R. and M.A.A. designed and performed most of the BRET and FRET experiments, wrote the main manuscript text, and prepared the figures and tables; R.Y. performed bias calculations; D.K. and Y.C. contributed to steroid measurements; N.G. contributed to siRNA experiments; E.R., M.S., and L.C. designed the experiments and wrote the manuscript. All authors reviewed the manuscript.

Additional Information

Competing Interests: The authors declare that they have no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2017