Multiple Sites in αB-Crystallin Modulate Its Interactions with Desmin Filaments Assembled In Vitro

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Abstract

The β3- and β8-strands and C-terminal residues 155–165 of αB-crystallin were identified by pin arrays as interaction sites for various client proteins including the intermediate filament protein desmin. Here we present data using 5 well-characterised αB-crystallin protein constructs with substituted β3- and β8-strands and with the C-terminal residues 155–165 deleted to demonstrate the importance of these sequences to the interaction of αB-crystallin with desmin filaments. We used electron microscopy of negatively stained samples to visualize increased interactions followed by sedimentation assays to quantify our observations. A low-speed sedimentation assay measured the ability of αB-crystallin to prevent the self-association of desmin filaments. A high-speed sedimentation assay measured αB-crystallin cosedimentation with desmin filaments. Swapping the β3-strand of αB-crystallin or deleting residues 155–165 increased the cosedimentation of αB-crystallin with desmin filaments, but this coincided with increased filament-filament interactions. In contrast, substitution of the β3-strand with the equivalent αA-crystallin sequences improved the ability of αB-crystallin to prevent desmin filament-filament interactions with no significant change in its cosedimentation properties. These data suggest that all three sequences (β3-strand, β8-strand and C-terminal residues 155–165) contribute to the interaction of αB-crystallin with desmin filaments. The data also suggest that the cosedimentation of αB-crystallin with desmin filaments does not necessarily correlate with preventing desmin filament-filament interactions. This important observation is relevant not only to the formation of the protein aggregates that contain both desmin and αB-crystallin and typify desmin related myopathies, but also to the interaction of αB-crystallin with other filamentous protein polymers.

Introduction

Human αB-crystallin is a small heat shock protein (αHSP) that interacts with a variety of important cellular proteins with the capacity to polymerise into either filaments [1], or tubules [2] or fibrils [3]. The fact that αB-crystallin is often part of the histopathological signature used to characterise a variety of human diseases [4] highlights the potentially important role that αB-crystallin plays in their etiology. This became apparent when it was discovered that mutations in both αB-crystallin [R120G; [5]] and desmin [6] can cause cardiomyopathy, typified by aggregates containing both proteins [4]. The R120G mutation in αB-crystallin also induced the aggregation of desmin filaments in transfected cells [7]. The dissociation constant was increased two fold for the R120G mutant compared to the wild-type αB-crystallin [7], which appeared to encourage the increased interaction of desmin filaments leading to their aggregation in transfected cells and in the muscles of affected individuals. Previously it had been established that αB-crystallin modulated the assembly of intermediate filaments [8] and reduced the extent of filament-filament interactions in vitro [9]. Over-expression of wild-type αB-crystallin is capable of reversing intermediate filament aggregation in transfected cells suggesting that αB-crystallin was involved in regulating the local associations of intermediate filaments [10]. The fact that mutations in αB-crystallin caused the aggregation of desmin filaments [5,7] also supports this view. It is therefore important to identify the sequences in αB-crystallin that are responsible for the effects on intermediate filaments and particularly desmin because mutations in either can be the genetic basis of myopathy [11,12,13].

Pin array studies identified sequences in αB-crystallin involved in the recognition of a variety of different client proteins including desmin and GFAP, two examples of intermediate filament proteins [1]. These sequences were not unique to the interaction of αB-crystallin with either desmin or GFAP [1], evidence of the ability of αB-crystallin to recognise a wide range of potential protein clients [14,15,16,17,18,19,20,21,22,23]. The five sequences in αB-crystallin with the strongest binding to desmin were spread throughout the primary sequence from the N- to the C-terminus and included some that were involved in αB-crystallin oligomerisation. The pin-array assays did not consider the assembly status of the desmin or GFAP, a potentially important factor in the mechanism of αB-crystallin activity. Indeed, the interaction between the αB-crystallin peptides and desmin was inversely
correlated with temperature in the pin array studies [1]. In contrast, the fraction of \(\alpha B\)-crystallin that pelleted with desmin filaments in the sedimentation assays increased with temperature [7]. Therefore it is important to verify that the sequences identified using the pin arrays are involved in the interaction of \(\alpha B\)-crystallin with desmin filaments.

Three \(\alpha B\)-crystallin peptide regions (\(\beta 3\)-strand, residues 73–85; \(\beta 8\)-strand, residues 131–138 and the C-terminal sequences 155–165) gave some of the strongest interactions with desmin using the pin array approach [1] were selected for our studies. Recent crystallisation [24] and solution structural [25,26] studies confirmed that all three regions are surface exposed on the \(\alpha B\)-crystallin subunit and were potentially available to bind client proteins such as desmin (see Fig. 1). Substituting the \(\beta 3\)- and \(\beta 8\)-strands with the equivalent sequences from \(\alpha A\)-crystallin and \(C.\) elegans HSP12.2 produced \(\alpha B\)-crystallin protein constructs that have been well-characterised previously in terms of structural changes and client protein interactions [16,17]. Exchange of either the \(\beta 3\)-strand from \(\alpha A\)-crystallin or \(C.\) elegans HSP12.2 with the equivalent \(\alpha B\)-crystallin sequence was shown to have minimal effect on the secondary, tertiary and quaternary structure of \(\alpha B\)-crystallin [17]. Replacement of the \(\beta 8\)-strand of \(\alpha B\)-crystallin with those from \(\alpha A\)-crystallin and \(C.\) elegans HSP12.2 also did not affect secondary structure, but oligomer size was increased [16]. Likewise deleting the C-terminal sequences 155–165 altered protein oligomerisation but without significant effects upon protein secondary structure. Interestingly chaperone activity was decreased for all but one of the client proteins tested (\(\beta L\)-crystallin, alcohol dehydrogenase and citrate synthase) [14,16,17] for these five different \(\alpha B\)-crystallin protein constructs demonstrating that

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**Figure 1. Location of IF interactive domains in \(\beta 3\)-strand (red), \(\beta 8\)-strand (yellow), and C-terminal 155–165 residues (blue) of human wild-type \(\alpha B\)-crystallin.** The primary sequences for human wild type \(\alpha B\)-crystallin, human wild type \(\alpha A\)-crystallin, and \(C.\) elegans wild type HSP12.2 were aligned using the residue numbers for human \(\alpha B\)-crystallin in ClustalX. The boxes and colors in the \(\alpha B\)-crystallin sequence correspond with the interactive sequences labeled on the surface of the 3D model. The amino acid substitutions in the \(\alpha B\)-crystallin protein constructs are indicated by **bold-italics**. The \(\beta 3\)-strand of \(\alpha B\)-crystallin, \(\beta 8\)-strand of \(\alpha B\)-crystallin, and \(\beta 8\)-strand of \(\alpha A\)-crystallin, \(\beta 3\)-strand of \(\alpha A\)-crystallin, \(\alpha B\)-crystallin, and \(C.\) elegans HSP12.2 produced \(\alpha B\)-crystallin protein constructs that have been well-characterised previously in terms of structural changes and client protein interactions [16,17]. Exchange of either the \(\beta 3\)-strand from \(\alpha A\)-crystallin or \(C.\) elegans HSP12.2 with the equivalent \(\alpha B\)-crystallin sequence was shown to have minimal effect on the secondary, tertiary and quaternary structure of \(\alpha B\)-crystallin [17]. Replacement of the \(\beta 8\)-strand of \(\alpha B\)-crystallin with those from \(\alpha A\)-crystallin and \(C.\) elegans HSP12.2 also did not affect secondary structure, but oligomer size was increased [16]. Likewise deleting the C-terminal sequences 155–165 altered protein oligomerisation but without significant effects upon protein secondary structure. Interestingly chaperone activity was decreased for all but one of the client proteins tested (\(\beta L\)-crystallin, alcohol dehydrogenase and citrate synthase) [14,16,17] for these five different \(\alpha B\)-crystallin protein constructs demonstrating that...
all three sequences are intimately involved in and optimised for client protein recognition in αB-crystallin. Recent studies, found that both the β3-strand and C-terminal sequences, but not the β3-strand in αB-crystallin, were responsible for regulating microtubule dynamics and preventing tubulin polymerisation [2,27]. Perhaps therefore, there are differences in the availability of the β3-strand, β8-strand and the C-terminal sequences 153–165 in αB-crystallin when a protein polymer is the client rather than individual protein subunits. For these reasons, we have determined the effect of the selected β3- and β8-strand substitutions as well as the C-terminal 153–165 deletion on the interaction of αB-crystallin with desmin filaments.

In the present study, the data demonstrate that the substituted β3-strand, β8-strand and C-terminal sequences in αB-crystallin can alter the interaction of αB-crystallin with desmin filaments and therefore all three sequences contribute to the interaction of αB-crystallin with desmin filaments. The sequence substitutions in αB-crystallin involving the C. elegans β3-strand and β8-strand as well as the C-terminal 153–165 deletion all caused increased desmin filament-filament interactions. In contrast, the αA-crystallin β3-strand substitution in αB-crystallin prevented desmin filament-filament interactions even more effectively than wild type αB-crystallin, but in a temperature specific manner. The data suggest that the interaction of αB-crystallin with desmin filaments does not always lead to the prevention of desmin filament aggregation, which we discuss with respect to desmin filament aggregation as a histopathological characteristic of desmin related myopathies.

Materials and Methods

Sequence alignment and molecular modeling

The amino acid sequence of human αB-crystallin was aligned with the sequences of human αA-crystallin and C. elegans HSP12.2 using ClustalX [28] and mapped to a 3D model (Fig. 1) for human αB-crystallin, which is in good agreement with all reported X-ray and NMR structures [18,24,26,28].

Mutagenesis of human αB-crystallin

Constructs selected for this study had either the β3 strand or β3 strand substituted by the equivalent sequences from αA-crystallin [16,17] and C. elegans HSP12.2 [16,17] as well as a previously characterised C-terminal deletion, which removed residues 153–165 [14]. The mutant proteins were similar in secondary structure, molecular weight and solubility to the wild-type protein. Mutagenesis of human αB-crystallin was performed using the QuikChange site-directed mutagenesis kit (Qiagen, Valencia, CA, USA) as previously described [14,16,17]. Two chimeric mutants in the β3 domain of αB-crystallin were created, in which the β3-strand (DRFSVNLDVKHFSG) was swapped with the corresponding sequence from either human αA-crystallin, DKKVIFLDVHKFS (αAβ3), or C. elegans HSP12.2, KEKFVGLDVQFT (CEβ3) [17]. Two chimeric mutants were created by substituting the β3-strand of human αB-crystallin, LTTTLSSLSST, with the corresponding sequences from human αA-crystallin, SALCSCLSS (αAβ8), or C. elegans HSP12.2, STVKSCHL (CEβ8) [16]. A fifth protein construct was created by deleting a C-terminal desmin interactive site (Δ155–165) [14].

Purification of proteins

Wild type and mutant human αB-crystallin were purified from bacterial lysates as previously described using ion exchange chromatography [29]. The wild type and the other five αB-crystallin protein constructs were soluble. Human desmin was purified from bacterial lysates as previously described using ion exchange chromatography [30,31]. Wild-type αB-crystallin, the five αB-crystallin protein constructs (αAβ3, CEβ3, αAβ8, CEβ8, and Δ155–165), and desmin were purified to >97% purity as determined by SDS-PAGE.

Assembly of desmin filaments

Assembly of desmin was performed as previously described [9,30]. Purified Desmin at 0.2 g/l in 6 M urea, 20 mM Tris-HCl pH8, 1 mM DTT, 1 mM EDTA, 0.2 mM PMSF (in the presence or absence of αB-crystallin at 0.08 g/l) was dialysed out of urea in a stepwise fashion by reducing the urea concentration to 4 M, then 2 M, then 0 M over a period of 24 h at 22°C. Desmin assembly was then initiated by dialysis into 20 mM Tris-HCl pH7.4, 50 mM NaCl for 16 h at either 22°C, 37°C or 44°C to ensure that assembly equilibrium has been reached.

Analysis of desmin αB-crystallin interactions by electron microscopy

Desmin, αB-crystallin, mixtures of both were diluted into assembly buffer to 100 μg/ml. A carbon film that had been coated onto freshly cleaved mica was then floated onto the surface of the sample prior to being negatively stained with 1% (w/v) uranyl acetate (Agar Scientific, UK) and retrieved with 400 mesh copper grids (Agar Scientific, UK). Grids were examined in an Hitachi H-7600 transmission electron microscope (Hitachi High-Technologies Corporation, Japan), using an accelerating voltage of 100 kV. Images were acquired using a CCD camera (Advanced microcopy Technology, Danvers, MA) and assembled into montages using Adobe® Photoshop CS (Adobe System, San Jose, CA).

Evidence of the association between desmin filaments and αB-crystallin particles in the EM images was statistically examined using likelihood ratio tests (LRT). For each combination of desmin and αB-crystallin protein construct, two representative images were selected for our analysis. Using ImageJ, a grid square was overlaid randomly over the image with a grid cell size equivalent to 11400 nm². Forty cells from the 125 total were then randomly selected and the number of filaments and particles within each cell counted. We proposed that the relation between the mean number of particles in a cell, μ, and the number of filaments in a cell, x, could be well described by,

\[ \mu = \beta_0 + \beta_1 (1 - e^{-\kappa x}). \]

The parameter β0 describes the background density of particles, \( \beta_1 \) is the maximum additional number of particles associated with filaments in the cell, and \( \kappa \) describes how quickly each additional filament contributes particles. We also proposed that the variation in αB-crystallin particle numbers was negative-binomial distributed (NBD) to correctly account for potential variation among cells caused by unknown sources. Richards (2005) [32] provides details on how to calculate the likelihood under the assumption of a NBD. The null, which states that there is no association, is obtained by setting \( \kappa = \beta_1 = 0 \). The test-statistic is \( G = 2(LL_1 - LL_0) \), where \( LL_1 \) and \( LL_0 \) are the maximum log-likelihood associated with the general model and the null model, respectively. Under the null hypothesis, G is chi-square distributed [33] with 2 degrees of freedom as the general model has two additional free parameters: \( \beta_1 \) and \( \kappa \).

Analysis of desmin αB-crystallin interactions using centrifugation

To investigate the interactions between desmin and the various αB-crystallin protein constructs, two separate sedimentation assays
(Fig. 2) were used to separate desmin filaments and their associated αB-crystallin from un-associated αB-crystallin [9,30]. In the high speed sedimentation assay, a 200 µl sample was layered onto a 100 µl 0.85 M sucrose cushion containing 10 mM Tris-HCl pH 7.0, 50 mM NaCl, 1 mM DTT, 200 µM PMSF. Samples were then centrifuged at 30,584 rpm (RCFmax = 80,000 x g) for 30 min at 4°C using a Beckman Coulter TLS-55 rotor (k factor = 50) to give pellet (Calculated size of pelletted particles ≈100 S) and supernatant fractions. The supernatant was carefully removed and samples prepared preserving volume equivalence so that a direct comparison could be made between pellet and supernatant fractions. The samples were then separated on 12% (w/v) polyacrylamide gels by SDS-PAGE and the separated proteins visualised by Coomassie Brilliant Blue staining. Destained gels were imaged using a Fujifilm LAS-100. Stained bands were quantified using Fujifilm Image Gauge V4.0 software. The protein content in the pellet (P) and supernatant (S) fractions at each temperature was measured based on Coomassie Brilliant Blue staining densities after SDS-PAGE and then plotted as bar charts to summarise the complete dataset and provide an overview. The total protein was the sum of the densities in the S and P fractions for each sample. The amount of desmin or αB-crystallin in each fraction was the density of the selected band in that fraction divided by the total protein. Low-speed centrifugation will pellet only desmin filaments that have become associated with each other by filament-filament interactions as well as any associated αB-crystallin (see Fig. 2 and [9,30]). This assay was initially developed to study interactions between keratin filaments [34]. Individual desmin filaments and unassociated αB-crystallin will not be pelleted under these sedimentation conditions. Immediately following assembly, samples were centrifuged at 4,900 rpm (RCFmax = 2,500 x g) for 10 min at 20°C using an Eppendorf 5417R benchtop centrifuge and standard fixed angle rotor (F-45-30-11, k-factor = 377). The supernatant was carefully removed from the pellet (Calculated S-value of pelleted material ≈2258S) and as with the high-speed assay, both fractions prepared for SDS-PAGE in a way to preserve the relative protein levels in each fraction so as to allow direct comparisons to be made when viewing the stained SDS-PAGE gels. Band intensities were quantified as described above.

### Results

Some αB-crystallin protein constructs appear to increase filament-filament interactions as seen by electron microscopy

Negative staining with uranyl acetate followed by electron microscopy was used to visualise the interactions between αB-crystallin particles and desmin filaments in samples prior to the sedimentation assays (Fig. 3). The wild type αB-crystallin formed 15–20 nm particles in agreement with observations by ourselves [8,31] and others [35], which were seen at all three temperatures (Fig. 3; WT αB). Similarly the desmin filaments (Fig. 3; Des) had a consistent morphology at the three different temperatures typically forming 10 nm filaments many microns long. When mixed together (Fig. 3; Des + WT aB), both individual desmin filaments and αB-crystallin particles were readily apparent for all the various desmin-αB-crystallin combinations (Fig. 3), but now some of the αB-crystallin particles were observed to be associated with the filaments at the three different temperatures eg (Fig. 3; [Des + WT

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**Figure 2. Desmin sedimentation assays.** (LEFT) Schematic of the 80,000 g (high-speed) sedimentation assay [8,9,30]. Desmin was assembled at 22, 37, or 44°C and centrifuged at 80,000 g. The pellet (P) will contain desmin filaments and any aggregates formed as a result of filament-filament interactions. Any αB-crystallin that associates with these filaments or their aggregates will also be cosedimented. The supernatant (S) will contain soluble αB-crystallin and also any assembly intermediates or unassembled desmin. Therefore this assay measures filament assembly and αB-crystallin binding to assembled filaments. (RIGHT) Schematic of the 2,500 g (low speed) centrifugation assay. Individual desmin filaments will not be sedimented by these sedimentation conditions, neither will αB-crystallin. Only when the assembled desmin filaments self-assemble into filament aggregates, will these sediment. Therefore this assay measures filament-filament interaction. If αB-crystallin binds to these aggregates, then it too will be cosedimented. Unlike the high-speed assay, it is the aggregate-associated αB-crystallin which will sediment into the pellet fraction (P) rather than the individual filaments and their associated αB-crystallin. The supernatant (S) will contain the free desmin filaments, their associated αB-crystallin, desmin assembly intermediates and the unassociated αB-crystallin particles.

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Temperature dependent increase in the association of wild type αB-crystallin with desmin by low- and high-speed sedimentation assay

Desmin filament assembly was conducted at 22°C, 37°C or 44°C in the presence or absence of the various αB-crystallin protein constructs. Pellet and supernatant fractions were analysed by SDS-PAGE and a representative experimental series is shown (Fig. 5). These data were combined with two other data sets and the % in each pellet (Figs. 5 and 6) and corresponding supernatant (data not shown) fractions calculated along with the standard error of the mean. The % of protein in the pellet fractions from both the low- and high-speed sedimentation assays from each sample are presented on the same bar in the charts (Figs. 5 and 6), with the proportion corresponding to the low speed assay being represented by the lower portion of each bar. This method of data presentation facilitates the comparison of both (high-speed and low-speed) sedimentation assays for each combination of αB-crystallin protein construct and desmin, which is needed to assess the interaction of αB-crystallin with desmin.

Desmin alone assembled efficiently and 94–98% sedimented into the pellet (P) fractions (Fig. 5A and 6). This was determined by the high-speed assay and there was no significant difference between the three different temperatures. In contrast, the low-speed centrifugation assay revealed that there was a temperature dependent increase in the proportion of desmin pelleted corresponding to 24%, 45% and 61% at 22°C, 37°C and 44°C respectively (Fig. 6). This assay measured filament-filament interactions and the data therefore suggest that these interactions were temperature dependent. We excluded the possibility that desmin failed to assemble equally efficiently at the three different temperatures, because the high-speed assay revealed a similar % of desmin in the pellet fractions at the three different temperatures (Fig. 6).

In the presence of αB-crystallin, 93–100% desmin sedimented into the high-speed pellet at the three temperatures indicating that wild type αB-crystallin did not alter the extent of desmin assembly (Fig. 5, HIGH SPEED; cf Des and Des + WT αB). Fig. 6; cf Des and Des + WT αB). In contrast, the low-speed sedimentation assay showed that the presence of αB-crystallin prevented filament-filament associations and only 12% of the desmin filaments sedimented at 22°C and 37°C (Fig. 5, 6. LOW SPEED; cf Des and Des + WT αB). At 44°C, the preventative effect was lost and there was no statistically significant difference in the proportion of pelletable desmin in the presence or absence of wild type αB-crystallin (Fig. 6, Des).

These experiments also measured the proportion of αB-crystallin that co-sedimented with the desmin filaments (Fig. 7A). In the absence of desmin, wild type αB-crystallin remains almost entirely in the supernatant fractions of both the high- and low-speed centrifugation assays (Figs. 4 and 6B). In the presence of desmin, however, there is a temperature dependent increase in the proportion of wild type αB-crystallin in the pellet fractions from the high-speed sedimentation assay (Figs. 4 and 6A). This corresponded to 9%, 17%, and 23% of the wild type αB-crystallin at 22°C, 37°C and 44°C respectively. These data confirm observations made in a previous study [7].

These data form the baseline for assessing the effects of changing the β3-strand, β3-strand sequences and deleting residues 155–165 in wild type αB-crystallin on the association of αB-crystallin with the desmin filaments and the subsequent effects on filament-filament interactions. All the various αB-crystallin protein constructs were soluble as determined by both sedimentation assays (Fig. 7B) and formed mono-disperse particles as judged by electron microscopy (Fig. S1). Only the Δ155–165 αB-crystallin protein construct showed an increased tendency to pellet and then only at 44°C (Fig. 7B, d155 αB).

Substituting the β8 strand of αB-crystallin and deleting the C-terminal residues 155–165 can promote desmin filament-filament interactions rather than inhibiting them

By comparison to the β3-strand substitutions, changing the β8-strand produced αB-crystallin protein constructs that showed either no significant improvement (αA-crystallin β8 chimera αB-crystallin...
Figure 4. Analysis of αB-crystallin interaction with desmin filaments using a likelihood ratio test (LRT). Samples from the shown combinations of desmin and αB-crystallin were analyzed by the described LRT. Examples of some of the selected images are shown. Strong statistical evidence (G2 log-likelihood scores 16.8–64.2, P values < 0.001%) was found that the αB-crystallin particles of the samples tested were positively associated with the desmin filaments (see summary table) in these samples. Bar = 100 nm.

| Desmin - αB-crystallin COMBINATION | G2 (log-likelihood) | P value |
|------------------------------------|---------------------|---------|
| Desmin + wild type αB-crystallin @22°C | 26.0                | < 0.001 |
| Desmin + Δ155 αB-crystallin @22°C  | 20.5                | < 0.001 |
| Desmin + αAβ3 αB-crystallin @37°C | 16.8                | < 0.001 |
| Desmin + CEβ8 αB-crystallin @44°C | 64.2                | < 0.001 |

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Figure 4. Analysis of αB-crystallin interaction with desmin filaments using a likelihood ratio test (LRT). Samples from the shown combinations of desmin and αB-crystallin were analyzed by the described LRT. Examples of some of the selected images are shown. Strong statistical evidence (G2 log-likelihood scores 16.8–64.2, P values < 0.001%) was found that the αB-crystallin particles of the samples tested were positively associated with the desmin filaments (see summary table) in these samples. Bar = 100 nm.
see Figs. 4, 5 and 6A) or a very obvious increase in desmin filament-filament interactions (C. elegans HSP12.2 β8 chimeraζB-crystallin (CEβ8)), the most dramatic observed at 44°C (Fig. 5B and 6, LOW SPEED; [Des + CEβ8 aB]). In fact, reduction in the ability of this C. elegans HSP12.2 β8 chimeraζB-crystallin to prevent desmin filament-filament interactions was mirrored by the deletion of the C-terminal residues 155–165 (Figs. 4B and 5, LOW SPEED; [Des + d155 aB]). It was also apparent from the high-speed centrifugation assay that a very striking shift in the pelletable portion of both these ζB-crystallin constructs when in the presence of desmin filaments (Figs. 4B and 6A. HIGH SPEED; [Des + CEβ8 aB] and [Des + d155 aB]). Quantification of the proportion of ζB-crystallin in the pellet fractions (Fig. 7A; [Des + CEβ8 aB] and [Des + d155 aB]) demonstrated that in excess of 95% of these ζB-crystallin proteins had indeed co-sedimented with the desmin filaments. For the Δ155–165 protein construct this might be partly

Figure 5. Gel electrophoretic analysis of the low- and high-speed sedimentation properties of desmin and wild type and mutant ζB-crystallins. (A) The low- and high-speed sedimentation properties of each individual protein was determined at 3 different temperatures. The pellet (P) and supernatant (S) fractions were analysed by SDS-PAGE and the proportion of each protein in each fraction determined. By high-speed sedimentation assay, which measures the efficiency of desmin assembly, virtually all the desmin had pelleted at 22°, 37° and 44°C. By low-speed sedimentation assay, there was a temperature dependent increase in the proportion of desmin sedimented. The ζB-crystallin remained largely in the supernatant fractions of both sedimentation assays. (B) Analysis of desmin pelleted by high- and low-speed sedimentation assay in the presence of either wild-type or the various ζB-crystallin protein constructs at three different temperatures. ζAβ3 ζB-crystallin (aAb3 aB) reduced the proportion of desmin filaments sedimenting at low-speed at 44°C. Conversely, the CEβ8 and the A155–165 ζB-crystallin protein constructs induced the complete low-speed sedimentation of desmin at 44°C. For each sedimentation assay, the band intensities were quantified and then combined with two other data sets to determine statistical significance and summarized in Figs. 5 and 6.

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The substitution of the β3 strand in the wild type αB-crystallin with that from αA-crystallin caused a dramatic decrease in the proportion of sedimentable desmin at 44°C in the low-speed assay (Fig. 6 [Des + aAb3 aB] dark blue bar cf [Des + WT aB], light blue bar). The effect was very evident by the gel analysis of the 44°C samples (Fig. 5B, LOW SPEED; cf Des + aAb3 aB and Des + WT aB). Quantification of these gel data (Fig. 6, Des + aAb3 aB) showed that there was a 5 fold difference in the sedimentable desmin by low-speed sedimentation assay (Fig. 6 [Des + aAb3 aB] dark blue bar cf [Des + WT aB], light blue bar). The results indicated a significant reduction in the extent of the filament-filament interactions in the presence of the αAβ3-αB-crystallin protein construct. The high-speed sedimentation assays showed no change in the level of assembled desmin at 44°C when coassembled with the αAβ3 αB-crystallin protein construct (Fig. 5B and 6, HIGH SPEED, Des + aAb3 aB) compared to desmin assembled in the presence of wild type αB-crystallin (Fig. 5B and 6, HIGH SPEED, Des + WT aB) in support of this conclusion.

Although the effect of substituting the β3-strand from C. elegans HSP12.2 (CEB3) was not as dramatic compared to that with the equivalent sequences from αA-crystallin (αAβ3), there was still a significant reduction (~2 fold) in the level of desmin sedimented in the low-speed centrifugation assay at 37°C (Fig. 5B and 6; LOW SPEED [Des + CEB3 aB] cf LOW SPEED [Des + WT aB]). At both 22°C and 44°C, no additional effects for the CEβ3 strand substitution into αB-crystallin were apparent (Figs. 5B and 6, LOW SPEED; cf [Des + CEB3 aB] and [Des + WT aB]). Once again this was not because of any effects of the αB-crystallin protein constructs upon desmin assembly per se as there was no significant difference in the pelletable desmin by high-speed sedimentation assays (Figs. 5B and 6 HIGH SPEED). Electron
Figure 7. Cosedimentation of αB-crystallin with and without desmin filaments. A. Cosedimentation of αB-crystallin with desmin filaments. Summary of the low-speed (light blue and dark blue) and high-speed (light red and dark red) sedimentation data for various αB-crystallin protein constructs coassembled with desmin, as quantified by gel densitometry. The percentage of αB-crystallin in the pellet fractions at 22°C, 37°C and 44°C was determined after both low- and high-speed sedimentation to quantify the association of αB-crystallin with the sedimented desmin filaments. The most striking observation is that both the CEβ8 (CEβ8 aB + Des) and Δ155–165 (d155 aB + Des) αB-crystallin protein constructs showed significant increases in desmin binding at 44°C as shown by the high speed assay (44°C, dark red bars). Conversely, αAβ3 αB-crystallin (αAβ3 aB + Des) showed significantly decreased association at 44°C at high speed. The CEβ3 (CEβ3 aB) and αAβ8 (αAβ8 aB) protein constructs showed similar sedimentation properties to wild type αB-crystallin. B. Aggregation of wild type and mutant αB-crystallins as measured by low- and high-speed sedimentation. Summary of the low-speed (blue) and high-speed (red) sedimentation data for the various αB-crystallin protein constructs as quantified by gel densitometry. The percentage of αB-crystallin in the pellet fractions at 22°C, 37°C and 44°C was determined after both low- and high-speed sedimentation assay to quantify the aggregation of the αB-crystallins. All the other protein constructs showed similar sedimentation properties to the wild type (WT aB) αB-crystallin, except Δ155–165 (d155 aB) αB-crystallin at 44°C, which showed increased aggregation by both low- (darker blue) and high-speed (darker red) sedimentation assay.

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microscopy confirmed that the morphology of the desmin filaments and the αB-crystallin particles were similar to those seen in samples containing the wild type αB-crystallin and desmin (Fig. 3 cf [Des + WT] with [Des + αB]).

The combined data show that substituting the β3-strand of αB-crystallin can have beneficial effects for the ability of αB-crystallin to prevent desmin filament-filament interactions. The effects are temperature specific, but these data demonstrate that it is possible to improve this activity by changing the β3-strand, confirming that this domain plays an important role in the association of αB-crystallin with desmin filaments.

The role of αB-crystallin binding in the prevention of desmin filament-filament interactions

The proportion of αAb3 that cosedimented with desmin filaments in the high-speed centrifugation assay at 44°C was significantly lower than that seen for the wild type αB-crystallin (Fig. 7A, 44°C [αAb3 αB + Des] and [WT αB + Des]). We interpret this to indicate reduced binding to the filaments. This was in addition to the ability of αAb3 to prevent desmin filament-filament associations at 44°C, which was increased some 5 fold compared to wild type (Fig. 6, 44°C [Des + αAb3 αB] and [Des + WT αB]). Conversely, the proportion of CEβ3 that cosedimented with desmin filaments in the high-speed centrifugation assay at 44°C was significantly higher than that seen for the wild type αB-crystallin (Fig. 7A, 44°C [CEβ3 αB + Des] and [WT αB + Des]), indicating increased binding to the filaments. The ability, however, of CEβ3 αB-crystallin to prevent desmin filament-filament interactions at 44°C was decreased compared to wild type αB-crystallin (Fig. 6, 44°C blue bars [Des + αB] and [Des + WT αB]). Therefore the ability to bind to desmin filaments does not necessarily correlate with the prevention of desmin filament-filament interactions. More specifically, increased αB-crystallin binding was not necessarily correlated with increased effectiveness in the prevention of filament-filament interactions.

Discussion

Implications for αB-crystallin interactions with desmin filament

The data presented here are the first analysis of the regions in αB-crystallin previously identified as capable of interacting with desmin filaments. The strand-swapping approach used here has confirmed that the β3-strand is an important interaction site in αB-crystallin for desmin filaments. A 5 fold improvement at 44°C in preventing filament-filament interactions for the αAb3 protein construct is a direct measure that this sequence is indeed important. Equally dramatic results were also obtained for one of the β strand-swapped protein constructs (CEβ8) and for the C-terminal deletion of residues 155–165. For these two protein constructs (CEβ8 and d155), a significant decrease in the ability of αB-crystallin to prevent desmin filament-filament interactions was observed (Fig. 6). The results emphasise the importance of all three regions to the interactions between αB-crystallin and self-assembling desmin filaments and subsequent filament-filament interactions (Fig. 5).

We also conclude from our data that it is reasonable to expect that changes to any interacting region can potentially produce positive as well as negative effects upon the observed activities of

**Figure 8. Summary of the influences of αB-crystallin on desmin filaments.** The β3- and β8-strands and the Δ155–165 sequences (C-terminal domain) in αB-crystallin were identified from peptide array studies as being desmin interaction sequences. In wild type αB-crystallin these sequences contribute to the interaction of the αB-crystallin oligomers with desmin filaments to prevent their self-association and the formation of filament-filament aggregates. This activity can be increased by substituting the β3-strand from other small heat shock proteins (αA-crystallin and C. elegans HSP12.2). Substituting the β8-strand in αB-crystallin or removing the 155–165 residues appears to lead to the loss of this activity, but increases the binding of αB-crystallin to desmin filaments. This in turn will encourage increased filament-filament interactions, which in the case of the many point mutations in αB-crystallin linked to inherited myopathies, then leads to protein inclusion formation and the appearance of the histopathological feature of desmin-related myopathies – protein inclusions containing both desmin and αB-crystallin.
αB-crystallin toward desmin filaments (Fig. 8). Indeed summarizing the activity of the β3-strand, β8-strand and Δ155–165 αB-crystallin protein constructs toward other client proteins shows that both improvement and deterioration should be expected with such changes to client protein binding sequences (Table 1). Currently the data presented here do not distinguish between a direct or indirect interaction of these regions with the desmin filaments. Nevertheless the fact that αB-crystallin particles are seen to decorate desmin filaments (Fig. 3) and the various αB-crystallin protein constructs cosedimented to either greater or lesser extents (Figs. 4 and 5), we interpret to mean that αB-crystallin binds directly to the desmin filament and involves the β3-strand, β8-strand and Δ155–165 sequences of αB-crystallin.

These results therefore confirmed the pin array studies that found multiple sites were responsible for the interaction of αB-crystallin with desmin [1]. Indeed the analysis of the effects of the cardiomyopathy causing mutant R120G αB-crystallin adds to this argument as this residue is outside of the β8-strand studied here, but is part of the β7-strand identified from the pin-arrays to be involved also in binding to desmin [1]. In patients, the R120G mutation leads to desmin filament aggregation and the formation of characteristic inclusions that were also enriched in αB-crystallin [5]. It was subsequently shown that the mutation also increased the binding affinity of αB-crystallin to desmin filaments by increasing the Kd by some two fold [7].

Relevance of desmin filament binding to histopathological aggregates of desmin and αB-crystallin

Another feature to emerge from the data presented here is that a significant increase in the binding of αB-crystallin to desmin filaments does not necessarily result in a similar increase in the ability of αB-crystallin to prevent the self-association of desmin filaments. This observation was first made with R120G αB-crystallin and GFAP filaments using a simplified viscometry assay [36] and was later confirmed for desmin filaments using the high-speed sedimentation assay [7]. Both the CEβ8 and Δ155–165 αB-crystallin protein constructs studied here showed increased cosedimentation with desmin filaments by high-speed sedimentation assay (Fig. 7A). Electron microscopy (Fig. 3) revealed this was direct binding to the desmin filaments, which coincided with an increase in desmin filament-filament associations as measured by the low-speed centrifugation assay (Fig. 6). In contrast, the αAβ3 αB-crystallin construct significantly inhibited desmin filament-filament associations, but the cosedimentation of this protein construct with desmin filaments was also significantly decreased at 44°C (Figs. 4–5). Compare these data to the detailed analysis of the Q151X myopathy-causing mutation in αB-crystallin where increased desmin cosedimentation of Q151X αB-crystallin was accompanied by a very significant decrease in desmin filament-filament associations [31]. The current study therefore confirms the importance of the β3- and β8-strands in interactions with desmin filaments. It also adds to previous observations that increased binding of αB-crystallin to desmin filaments does not necessarily correlate with the prevention of desmin filament-filament associations. This is reminiscent of the situation in desmin related myopathies where the characteristic histopathological feature of the disease is protein aggregates containing both αB-crystallin and desmin [11,12,13].

A role of αB-crystallin in modulating interactions between biopolymers?

Further quantitative studies are required to define the relationship between binding of αB-crystallin and the polymerisation and subcellular distribution of important biopolymers such as intermediate filaments, microtubules and actin filaments. This poses an obvious question concerning the selection or hierarchy in the interaction of αB-crystallin with the different polymers and how this involves the different interaction sequences.

For the client protein T4 lysozyme, αB-crystallin has both high and low affinity binding sites [37,38]. Binding to the high affinity site appeared to induce structural changes in the client protein itself. For desmin, the measured dissociation constant [7] is equivalent to the low affinity site on T4 lysozyme, but how this might affect the subunit geometry within the filament has not yet been determined. The pin array studies show that the β3-strand, β8-strand and 155–165 regions are all involved in binding to all three cytoskeletal proteins [1]. Refinement of these studies has shown that peptides derived from the β3-strand and the C-terminal 15–165 region could inhibit tubulin assembly whereas the sequences in the β7-strand and including R120 actually promoted tubulin assembly [2]. Interestingly the β3-strand was not involved in the binding to microtubules, but the studies here have identified

Table 1. Comparison of the chaperone activities of WT and the various αB-crystallin protein constructs used in this study with different client proteins.

| Insulin (DTT-induced) | β Crystallin | ADH | CS | Median # of Subunits | Far-UVCD | Near-UVCD |
|------------------------|-------------|-----|----|----------------------|---------|-----------|
| 22°C | 37°C | 44°C | 50°C | 50°C | 50°C | 22°C | 37°C | 50°C | 37°C | 50°C |
| no αB | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | x | x | x | x |
| WT | 0.85 | 0.91 | 0.94 | 0.16 | 0.13 | 0.59 | 24 | - | - | - |
| αAβ3 | 0.91 | 0.73 | 0.77 | 0.25 | 0.22 | 0.51 | 27 | UC | UC | UC |
| CEβ3 | 0.95 | 1.03 | 1.09 | 0.49 | 0.42 | 0.69 | 27 | UC | UC | UC |
| αAβ8 | 0.95 | 0.86 | 0.95 | 0.43 | 0.74 | 0.61 | 31 | UC | UC | UC |
| CEβ8 | 0.93 | 0.87 | 0.87 | 6.09 | 0.99 | 0.62 | 31 | UC | UC | UC |
| Δ155 | 1.02 | 1.33 | 1.28 | 11.33 | 2.75 | 0.81 | 25 | UC | UC | UC |

Numbers shown are normalised values of protein aggregation as determined by light scattering. Aggregation was calculated as (light scattering in the presence of chaperone)/light scattering in the absence of chaperone) for the relevant client proteins. Where normalised values became greater than 1.00, this indicates increased protein aggregation. The results summarized here for the various αB-crystallin protein constructs are from previously published work (αAβ3 and CEβ3 see reference [17]; αAβ8 and CEβ8 see reference [16]; Δ155 see reference [14]). Median number of subunits was determined by size exclusion chromatography. Data from far- and near-UVCD spectroscopy were used to analyze secondary and tertiary structure of the αB-crystallin protein constructs. All αB-crystallin proteins consisted of large polydisperse oligomers and had far- and near-UVCD spectra unchanged (UC) from wild type (WT) αB-crystallin at 37°C and 50°C.

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were stained with 1% (w/v) uranyl acetate and then processed for crystallin proteins used in this study. The

Supporting Information

Figure S1 Electron microscopy characterisation of the 2B-crystallin proteins used in this study. The 2B-crystallin samples were stained with 1 % (w/v) uranyl acetate and then processed for electron microscopy. All proteins appeared as monodisperse particles. Scale bar represents 100 nm. (TIF)

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