Structural basis of HapE<sup>P88L</sup>-linked antifungal triazole resistance in Aspergillus fumigatus

Peter Hortschansky<sup>1</sup>, Matthias Misslinger<sup>2</sup>, Jasmin Mörl<sup>2</sup>, Fabio Gsaller<sup>2</sup>, Michael J Bromley<sup>3</sup>, Axel A Brakhage<sup>1</sup>, Michael Groll<sup>4</sup>, Hubertus Haas<sup>2</sup>, Eva M Huber<sup>6</sup>

Azoles are first-line therapeutics for human and plant fungal infections, but their broad use has promoted the development of resistances. Recently, a pan-azole-resistant clinical Aspergillus fumigatus isolate was identified to carry the mutation P88L in subunit HapE of the CCAAT-binding complex (CBC), a conserved eukaryotic transcription factor. Here, we define the mechanistic basis for resistance in this isolate by showing that the HapE<sup>P88L</sup> mutation interferes with the CBC’s ability to bend and sense CCAAT motifs. This failure leads to transcriptional derepression of the cyp51A gene, which encodes the target of azoles, the 14-α sterol demethylase Cyp51A, and ultimately causes drug resistance. In addition, we demonstrate that the CBC-associated transcriptional regulator HapX assists cyp51A repression in low-iron environments and that this iron-dependent effect is lost in the HapE<sup>P88L</sup> mutant. Altogether, these results indicate that the mutation HapE<sup>P88L</sup> confers increased resistance to azoles compared with wt A. fumigatus, particularly in low-iron clinical niches such as the lung.

DOI 10.26508/lsa.202000729 | Received 2 April 2020 | Revised 8 May 2020 | Accepted 12 May 2020 | Published online 28 May 2020

Introduction

The global burden of aspergillosis exceeds 14 million people and mortality rates are especially high in patients with chronic and invasive diseases (Bongomin et al, 2017). The main class of therapeutics used to treat aspergillosis are azoles, in particular triazoles. Sub-optimal, widespread and long-term use of these drugs, however, has promoted the development of resistances. In some European centers, the levels of resistance exceed 20% and the U.S. Centers for Disease Control and Prevention have placed Aspergillus fumigatus, the primary etiological agent responsible for aspergillosis, on their watch list for antibiotic-resistant pathogens (https://www.cdc.gov/drugresistance/pdf/threats-report/2019-ar-threats-report-508.pdf). This worldwide development is of growing concern and demands a thorough understanding of the molecular mechanisms that contribute to drug resistance to support the development of alternative therapeutic strategies.

Recently, patient-acquired azole resistance of the human pathogenic mold A. fumigatus has been linked to the CCAAT-binding complex (CBC) (Arendrup et al, 2010; Camps et al, 2012a; Gsaller et al, 2016), a highly conserved and fundamental eukaryotic transcription factor (Bhattacharya et al, 2003). The core version of the CBC is a heterotrimer of the subunits HapB, HapC, and HapE that binds the CCAAT box, a promoter element present in about 30% of all eukaryotic genes (Bucher, 1990; Marino-Ramirez et al, 2004; Furukawa et al, 2020). The two histone-like subunits HapC and HapE associate with the DNA backbone and bend it in a nucleosome-like manner, whereas HapB with its sensor helix αS and its C-terminal anchor inserts into the minor groove and recognizes the CCAAT box (Huber et al, 2012; Nardini et al, 2013). Depending on the target gene and other transcriptional regulators, the CBC hereby either activates or inhibits gene expression.

In certain fungi such as Aspergillus sp., a subset of genes are controlled by a more sophisticated variant of the CBC, termed CBC–HapX. This complex consists of HapB, HapC, and HapE, as well as two copies of HapX. HapX is a basic region leucine zipper (bZIP) that features an additional DNA-binding site 12 bps downstream of the CCAAT box (Hortschansky et al, 2017; Furukawa et al, 2020). CBC–HapX–controlled target genes are involved in iron homeostasis, storage, and consumption as well as ergosterol biosynthesis (Hortschansky et al, 2007; Gsaller et al, 2014, 2016). Ergosterol is a key component of fungal cell membranes and ensures their integrity as well as fluidity. Its biosynthesis involves the 14-α sterol demethylase Cyp51A, which is the primary target of azole-based antifungal drugs such as voriconazole (Oddo et al, 2003; Becher & Wirsel, 2012; Monk et al, 2020).

Apart from mutations in the Cyp51A enzyme that prevent drug binding (Snelders et al, 2010), azole-resistant phenotypes can be based on efflux transporters (Fraczek et al, 2013) or on alterations of the cyp51A promoter (Snelders et al, 2011). In wild-type (wt) A. fumigatus, the cyp51A promoter contains binding sites for three
counteracting transcription factors: two inducers, the sterol regulatory element–binding protein SrbA (Gsaller et al., 2016) and the ATP-binding cassette transporter regulator AtrR (Paul et al., 2019), as well as a repressor, CBC–HapX (Gsaller et al., 2016). In azole-resistant A. fumigatus, however, duplication of a 34-mer region in the promoter (tandem repeat of 34 bps, TR34) creates additional binding sites for SrbA and AtrR, thereby leading to enhanced expression of the cyp51A gene, overproduction of the Cyp51A enzyme, and eventually to azole resistance (Gsaller et al., 2016; Paul et al., 2017).

Recently, in a patient infected with A. fumigatus, another mechanism of azole insensitivity has been discovered. The mutation leads to the amino acid change P88L in subunit HapE of the CBC, impairs the binding affinity of the complex to its target site, and prevents repression of the cyp51A gene (Camps et al., 2012a; Gsaller et al., 2016). This condition also leads to drug resistance by enhanced production of the Cyp51A enzyme, but how the mutant HapEP88L subunit alters functioning of the CBC remained unknown. We here investigated the molecular mechanism of HapEP88L-mediated CBC dysfunction using in vivo and in vitro experiments. X-ray crystallographic analysis of the mutant CBC provided explanation for the reduced affinity of the CBC to its target DNA and significantly extended our current understanding of HapEP88L-induced azole resistance.

Results

In vivo analysis of hapEP88L-induced effects

Biological and physiological impacts of the HapEP88L-mutant CBC subunit were evaluated first in vivo by probing the ability of isogenic wt and hapEP88L-mutant isolates of A. fumigatus to grow under different iron conditions.

Figure 1. Growth phenotyping, gene expression analysis, siderophore production, and azole resistance of Aspergillus fumigatus CBC mutants. (A) Growth pattern of A. fumigatus wild-type (wt), hapEP88L, hapEP88LΔhapX, ΔhapX, and ΔhapC strains on solid minimal medium containing different iron concentrations. Growth was evaluated after incubation at 37°C for 48 h. (B) Production of biomass in submersed cultures (liquid growth at 37°C for 24 h) during iron starvation (−Fe), iron sufficiency (0.03 mM FeSO₄, +Fe), and iron excess (2.5 mM FeSO₄). (C) Gene expression levels of the CBC and CBC–HapX target genes mirB (siderophore transporter), cccA (vacuolar iron transporter), cycA (cytochrome c), and cyp51A (14-α sterol demethylase Cyp51A) under the indicated iron conditions. Northern blot analyses were performed from liquid cultures grown at 37°C for 24 h under iron starvation (−Fe) or iron sufficiency (0.03 mM FeSO₄). Alternatively, mycelia were shifted for 30 min from −Fe to iron sufficiency (0.01 mM FeSO₄, sFe) to generate short-term iron excess. As a loading control, ribosomal (r)RNA levels are shown below. (D) Siderophore production (triacetylfusarinine C and fusarinine C) in mutant A. fumigatus strains compared with wt in the absence of iron. (E) Iron-dependent azole resistance of A. fumigatus mutants. Voriconazole (10 μg/ml) was spotted on agar plates inoculated with A. fumigatus spores, and the width of the inhibition zone was observed as a measure of drug resistance after 48 h. The narrower the inhibition zone was, the more resistant the strains were. Data information: In (B, D, E), data are presented as the mean and SD of three biological replicates and analyzed by one-way ANOVA with Tukey’s multiple comparison test (*P ≤ 0.05; **P ≤ 0.01; ***P ≤ 0.001; ns, not statistically significant). Source data are available for this figure.
conditions. As the CBC–HapX complex controls fungal adaption to varying iron concentrations (Schrettl et al, 2010; Gsaller et al, 2014), iron depletion, sufficiency, and excess were tested. In all settings, growth of the mutant was clearly impaired compared with wt *A. fumigatus*. Deletion of the gene encoding the HapX subunit in the hapE<sup>PRSL</sup> background further aggravated the phenotype when compared with the respective single mutants (Fig 1A). The phenotype of the hapE<sup>PRSL</sup> mutant was less severe than for an isogenic strain lacking a functional CBC (ΔhapC). Similar results were observed when monitoring the fungal biomass obtained from liquid cultures under various iron concentrations (Fig 1B). In summary, the hapE<sup>PRSL</sup> mutation severely impairs viability parameters of *A. fumigatus* and, in particular, the tolerance to low and high iron stress.

To sequester iron from the surroundings, *A. fumigatus* secretes chelators termed siderophores (Haas, 2014), whose re-uptake is mediated by siderophore transporters such as MirB. During iron starvation (–Fe), the wt CBC–HapX complex stimulates biogenesis of MirB to promote acquisition of the metal. In the hapE<sup>PRSL</sup> mutant, however, the strength of gene induction by the CBC was considerably lower (Fig 1C). Furthermore, the hapE<sup>PRSL</sup> strain produced at least 50% less of the extracellular siderophores triacetylfusarinine C and fusarinine C than wt (Fig 1D). In low-iron environments, wt *A. fumigatus* also down-regulates cycA and cyp51A genes, which encode the iron-dependent proteins cytochrome c and 14α-sterol demethylase Cyp51A, respectively, to restrict nonessential iron use (Fig 1C). Both genes, however, showed significant expression in the hapE<sup>PRSL</sup> mutant, indicating a transcriptional deregulation. Strikingly under short-term iron sufficiency (5Fe), effects of the hapE<sup>PRSL</sup> mutation were less discernible. Transcription of the ccaA gene, coding for the vacuolar iron importer, was not affected in the hapE<sup>PRSL</sup> strain but completely abrogated in ΔhapC backgrounds (Fig 1C). A similar, although weaker, tendency was found for cycA and cyp51A. As the promoters of all three of these genes are strong targets of the wt CBC–HapX complex (Furukawa et al, 2020; Gsaller et al, 2014), HapX might compensate the deleterious effects of the mutant HapE subunit and enable their transcription. In agreement, inactivation of HapX in the hapE<sup>PRSL</sup> mutant decreases viability (Fig 1A). Together, these results suggest that HapX plays a dominant role in stabilizing the DNA–regulator complex and that the mutation hapE<sup>PRSL</sup> affects transcription of a subset of CBC targets within the genome including cyp51A. Considering that the CBC controls expression of about 30% of all eukaryotic genes (Bucher, 1990; Furukawa et al, 2020), the mutant HapE subunit might cause dysregulation of many biochemical pathways and provoke the observed severe growth retardation of *A. fumigatus*.

Next, we tested the resistance of *A. fumigatus* to the broad-spectrum antifungal medication voriconazole. Consistent with iron-controlled expression of the cyp51A gene (Fig 1C), resistance of wt *A. fumigatus* to voriconazole was iron dependent. Furthermore, loss of CBC function (ΔhapC or hapE<sup>PRSL</sup>) or HapX abrogated this effect. During iron starvation, hapE<sup>PRSL</sup> strains were considerably more resistant to the drug than wt, revealing that the transcriptional derepression observed for cyp51A in the mutant *A. fumigatus* isolate correlates with increased resistance. In agreement, in the presence of iron, which stimulates cyp51A expression, the effect was less pronounced (Fig 1E).

### In vitro studies with HapE<sup>PRSL</sup>–mutant CBC

To evaluate whether the mutant CBC is able to bind target DNAs in vitro, we performed surface plasmon resonance (SPR) experiments with purified wt and mutant CBCs, as well as various 25-bp long nucleic acid duplexes. Consistent with results from the cyp51A promoter (*Gsaller*...
et al, 2016), HapE<sub>P88L</sub>-mutant CBCs showed a drastic decrease in affinity for cycA (K<sub>D</sub> increases by factor 140) and cccA (up to factor 1,390) CCAAT sequences (Fig 2). Similarly, the half-lives of the protein–DNA assemblies were strongly reduced (factor 67 for cycA and at least factor 76 for cccA). This result confirms that in the HapE<sub>P88L</sub> context, transcriptional control by the CBC is defective. Furthermore, this effect was the same for the A. fumigatus (Afu) and Aspergillus nidulans (An) CBC (Fig 2), indicating that the mechanism of HapE<sub>P88L</sub> transcriptional deregulation is the same across species. Regarding the wt-like expression profile of the ccaA gene in the hape<sub>P88L</sub> setting during a short-term shift from iron starvation to iron sufficiency (sFe; Fig 1C), we additionally investigated the effect of HapX on the DNA-binding affinity of mutant CBC. SPR co-injection assays revealed that the half-life of the ternary CBC<sub>P88L</sub>–cccA–HapX complex is increased sevenfold compared with the binary CBC<sub>P88L</sub>–cccA particle (Fig 3). Hence, it appears that HapX is at least partially able to restore the ability of the CBC to bind to its recognition site.

**Structural examination of HapE<sub>P88L</sub>-mutant CBC**

To investigate how the P88L mutation in HapE alters the DNA-binding capacity of the CBC and ultimately confers drug resistance, we attempted the crystallization of wt and mutant CBC from A. fumigatus (Fig S1A). Structures of the Afu_CBC could be solved in complex with double-stranded 23-bp-long DNA fragments derived from the promoter sequences of either cycA (2.6 Å resolution, Table 1 and Fig S1B) or cccA (2.3 Å resolution, Table 1 and Fig S1B). In addition to our previously determined An_CBC–cycA crystal structure (Huber et al, 2012), we also obtained data on the An_CBC in complex with the cccA DNA fragment (2.2 Å resolution, Table 1 and Fig S1B). Superpositions of the Afu_CBC–cycA and the Afu_CBC–cccA complexes as well as the corresponding A. nidulans proteins indicated high structural similarity, suggesting that complex arrangement and DNA bending are uniform among these species and independent of the nucleic acid sequence and the target gene promoter site (root-mean-square deviation [rmsd] 0.153 Å for A. fumigatus, 0.246 Å for A. nidulans, 0.306 Å for cycA, and 0.369 Å for cccA complexes; Fig S2A).

Azole-resistant Afu_CBC<sub>P88L</sub> protein preparations, however, did not stably associate with Afu_cccA promoter-derived double-stranded DNA, as confirmed by size exclusion chromatography (SEC) (Fig 4A). The residual complex affinity of 429 nM (Fig 2C) probably was not high enough to counteract the shearing forces during chromatography. In addition, Afu_CBC<sub>P88L</sub> failed to crystallize in the presence of An_cycA promoter DNA (Fig 4B). We, therefore, switched organisms and created the HapE<sub>P88L</sub>-mutant A. nidulans CBC. Despite reduced affinity (116 versus 0.83 nM for wt; Fig 2A), we obtained a SEC-stable complex for this variant with the 23-bp-long cycA promoter fragment (Fig 4C) and elucidated its X-ray structure at 2.3 Å resolution (Table 1). With the previously solved wt An_CBC–cycA complex in hands (PDB ID 4G92), direct comparison with the mutant protein–DNA structure was possible. In contrast to the wt CBC that binds the DNA fragment in a 1:1 stoichiometry (Huber et al, 2012), three CBC<sub>P88L</sub> complexes associate with one DNA double strand encoding a single CCAAT-binding motif (Fig 5A). Notably, the orientations of the three CBCs relative to the CCAAT-binding motif deviate from each other, and all differ from the wt structure (Figs 5A and S3).

**Figure 3. Cooperative CBC–HapX binding stabilizes DNA interaction of the HapE<sub>P88L</sub>-mutant CBC.**

(A, B) Surface plasmon resonance analyses are shown for binding of wt CBC (A) or HapE<sub>P88L</sub>-mutant CBC (B) to DNA (panel 1), HapX to DNA (panel 2), and HapX to preformed CBC–DNA complexes (panel 3). The sequence of the immobilized DNA duplex is derived from the Aspergillus fumigatus cccA promoter. Nucleotides (nts) underlined in black are covered by the CBC (Huber et al, 2012), and nts marked in blue represent the HapX consensus binding site (Gsaller et al, 2016). Binding responses of the indicated CBC or HapX concentrations injected in duplicate (black lines) are overlaid with the best fit derived from a 1:1 interaction model, including a mass transport term (red lines). Binding responses of CBC–DNA–HapX ternary complex formation (panel 3, blue lines) were obtained by concentration-dependent co-injection of HapX on preformed binary CBC–DNA complexes after 200 s within the steady-state phase. Sensorgrams in panel 4 depict the association/dissociation responses of HapX on preformed CBC–DNA and were generated by CBC response subtraction (co-injection of buffer) from HapX co-injection responses. Dissociation constants (K<sub>D</sub>) and half-lives of the complexes are plotted inside the graphs.
Intriguingly, the mutant CBCs do not significantly bend the DNA as it was previously noted for the wt protein–DNA complex (Huber et al., 2012) (Fig 5B). Whereas the wt CBC induces a bending angle of 68° (Huber et al., 2012), the respective parameter for the mutant CBC–DNA complex is 9.3°. Reduced DNA curvature results from an altered binding mode of the CBCP88L complexes to the DNA–sugar–phosphate backbone. Actually, for all three CBCP88L proteins in the asymmetric unit, different interaction patterns with the DNA were observed and most of them are based on hydrogen bonds between protein main chain amides and phosphate moieties of the DNA (Fig S3).

Structural superposition proved that the three copies of the protein complex are identical and comparison with the wt CBC coordinates (PDB ID 4G91 [Huber et al., 2012]) illustrated that the mutation P88L neither disrupts the subunit fold nor complex assembly (rmsd < 0.169 Å). However, in each of the mutant CBCs, only the αN helix of HapB was defined in the 2Fο−Fc electron density map, whereas the αS sensor helix, which usually inserts into the DNA double strand and thereby confers sequence specificity to the CBC (Huber et al., 2012), was disordered (Figs 5A and B and 6).

The site of mutation, Pro88, forms the boundary between loop L0 and helix α1 of subunit HapE, and the succeeding residues 89–94 undergo hydrogen bond interactions with the sugar–phosphate backbone of the nucleic acid in the wt CBC–cycA crystal structure (Huber et al., 2012) (Fig 5B and C). Substitution of Pro88 by Leu leads to N-terminal elongation of helix α1 by approximately a half turn (Figs 5D and 6). Superposition of the wt CBC–cycA complex with the mutant CBCP88L protein indicates that the mutation-induced extension of helix α1 clashes with the bent conformation of the DNA observed for the wt CBC (Fig 5A). Thus, it appears that the mutation P88L prevents histone-like DNA binding by the CBC and insertion of HapB’s αS helix into the DNA.

Figure 4. Reconstitution of CBC–DNA complexes for crystallization. (A, B, C) Size exclusion profiles and SDS–PAGE analysis of HapP88L–mutant CBC–DNA complexes from (A, B) Aspergillus fumigatus and (C) Aspergillus nidulans. The strongest CBC target sites from A. fumigatus cccA (A) and A. nidulans cycA (B, C) promoter sequences were chosen as DNA duplexes. Size exclusion chromatography (SEC) fractions that were subjected to crystallization after a subsequent concentration step are marked in red (samples labeled 1C and 2C in the respective SDS–PAGE gels). SDS–PAGE gels were stained for protein with the GelCode Blue Stain Reagent (upper panels) before DNA staining with SYBR Gold Nucleic Acid Gel Stain (lower panels). Source data are available for this figure.
Figure 5. Structural comparison of wt and HapE<sup>P88L</sup> mutant An_CBC–cycA complexes.

(A) Ribbon illustration of CBC–DNA complexes. The 23-bp cycA DNA fragments and the protein assemblies forming the asymmetric units of the crystals are color-coded, whereas crystallographic symmetry mates are shown in gray. Because of the 5’-AA-TT overhangs, the DNA duplexes arrange as fibers in the crystal lattice. The CCAAT-binding motif is colored yellow, whereas HapB, HapC, and HapE subunits are depicted in green, blue, and red, respectively. The mutant CBC does not induce significant DNA bending and fails to interact sequence-specifically with the DNA. Because of nonspecific DNA-binding events, the stoichiometry of mutant CBC complexes to DNA fragments is increased to 3:1 compared with 1:1 for wt CBC.

(B) In wt An_CBC–cycA, Pro88 (green; black arrow) of HapE (red) is located adjacent to the DNA. The P88L mutation disrupts the interaction with the DNA and its bending. Structures are rotated by 180° along the y-axis compared with panel (A).

(C) Hydrogen bond interactions
CCAAT-binding motif. Altogether, the affinity of mutant CBC complexes for CCAAT-binding sites is severely reduced and this loss of sequence specificity leads to the association of three transcription factors per DNA double strand (Fig 5A). The residual ability to bind DNA, however, is insufficient to properly position the CBC in the promoter region of the cyp51A gene and to repress its transcription. The uncontrolled expression of cyp51A leads to overproduction of the 14-α sterol demethylase Cyp51A and eventually renders commonly applied azole concentrations inactive.

**Discussion**

Infections with azole-resistant *A. fumigatus* are of growing concern in clinics. Azoles are the only orally available antifungals (Lelievre et al, 2013), and alternative agents to treat invasive aspergillosis are scarce. Hence, patients suffering from drug-resistant invasive aspergillosis face mortality rates of up to 100% (Meis et al, 2016). The widespread use of triazole-based fungicides in agriculture and export of crops are likely to have contributed to the emergence and spread of resistance (Snelders et al, 2008; Verweij et al, 2009; Camps et al, 2012b; Chowdhary et al, 2013; Bueid et al, 2010), and alternative agents to treat invasive aspergillosis described in the text. However, because in every second patient, the CBC might influence the strength of DNA binding and hence the level of transcription as well.

In contrast to other azole resistance mechanisms, inactivation of the CBC, either by deletion of *hapC* (Gsaller et al, 2016) or by the point mutation P88L in HapE, significantly attenuates virulence of *A. fumigatus* (Arendrup et al, 2010). The reduced viability and pathogenicity of *hapE*P88L-mutant *A. fumigatus* may be the reason why this mutation has so far only been identified in a clinical isolate and not in the environment. Nonetheless, because in every second patient, with azole resistance, the molecular mechanism is not mediated by Cyp51A and of unknown origin (Bueid et al, 2010), CBC-linked drug resistance may contribute to the emergence and spread of resistance (Snelders et al, 2011). Here, we investigated how the recently discovered point mutation P88L in subunit HapE of the CBC, a ubiquitous transcription factor, confers azole resistance to *A. fumigatus* (Camps et al, 2012a).

We show that azole resistance and iron homeostasis are inextricably linked through the action of the CBC and its accessory subunit HapX. In particular, in wt *A. fumigatus*, azole resistance is decreased under low-iron conditions which is consistent with reduced cyp51A expression. This low-iron–mediated azole sensitivity is abolished in HapX-deficient backgrounds and in the *hapEP88L* mutant. We also found that a *hapEP88L* mutant of *A. fumigatus* is less resistant to iron starvation as well as iron overload because of altered gene regulation by the CBC. In vitro SPR analysis revealed that *HapEP88L*-mutant CBCs poorly bind to CCAAT boxes in general. These results agree with the reduced CBC-affinity reported for the cyp51A gene (Gsaller et al, 2016) and mutagenesis experiments on the human HapE homolog NF-YC, which showed residues 43–45 (corresponding to 87–89 in *A. fumigatus* and *A. nidulans*) to be essential for DNA binding (Zemoumi et al, 1999). For this reason, attempts to crystallize the *HapEP88L*-mutant CBC from *A. fumigatus* in complex with DNA failed. Likewise, in the absence of nucleic acid, *Afu_CBC* did not crystallize. The primary sequences of *Afu_CBC* and *An_CBC* only differ by two Val to Ile replacements in subunit HapE (Fig S1A). As visualized by the wt *Afu_CBC–cccA* and wt *Afu_CBC–cycA complex structures, these conserved amino acid variations cause a slight shift of the N-terminal αN helix of HapE that might enhance mobility and prevent crystallization in the absence of a high-affinity DNA ligand (Fig S2B). We, therefore, focused on the *A. nidulans* CBC. Structures of wt and *HapEP88L*-mutant *An_CBC* in complex with DNA visualized that reduced curvature of the nucleic acid is the primary cause for the low affinity to the *HapEP88L*-mutant CBC. The mutation P88L elongates helix α1 of subunit HapE and sterically interferes with DNA bending (Fig 5A). This observation agrees with the reported propensity of proline to often N-terminally cap α-helices (Richardson & Richardson, 1988; Kim & Kang, 1999; Cochran et al, 2001) and its helix breaker function in soluble proteins (Chou & Fasman, 1974) as well as the tendency of leucine to be part of α-helices (Fujiwara et al, 2012). Because of the altered bending angle of the DNA, the sequence-specific HapB subunit fails to find the CCAAT motif, resulting in its structural disorder and the random positioning of CBC complexes on the DNA via electrostatic interactions with the sugar–phosphate backbone. The crucial importance of Pro88 for DNA curvature and high-affinity binding is underpinned by its strict conservation and the X-ray structure of the human CBC homolog, the NF–Y complex (Nardini et al, 2013). Despite these structural changes in the *HapEP88L*-mutant, in vivo and SPR experiments suggest that in the presence of HapX, the affinity on CBC–HapX target sequences is partially retained. Although the basic region leucine zipper HapX can act as a transcription factor only when bound to the CBC, it has an own DNA recognition motif downstream of the CCAAT box (Gsaller et al, 2014; Hortschansky et al, 2015; Furukawa et al, 2020). It is, therefore, conceivable that in the *hapEP88L*-mutant strain, HapX guides CBC–HapX complexes to the nucleic acid and by binding to its target sequence may enable correct positioning of the CBC near the CCAAT sequence. This way, the HapB subunit may be able to recognize the CCAAT motif and to insert into the DNA double strand (Huber et al, 2012; Nardini et al, 2013). In agreement, expression of the strong CBC–HapX target *cccA* is not affected by the *hapEP88L* mutation, and inactivation of HapX in the *hapEP88L* background further reduced the growth ability. We, therefore, suppose that promoters encoding solely the CCAAT box are more severely affected by the mutant HapE subunit and are more likely to lose their transcriptional control than those featuring in addition a HapX-binding site. However, depending on the promoter sequence, other interaction partners of the CBC might influence the strength of DNA binding and hence the level of transcription as well.

of α1 helices of wt and mutant HapE with adjacent DNA backbones (dotted lines) differ because of the mutation P88L (green). Similarly, the orientation of the DNA as well as the site of interaction with the protein are distinct. Protein–DNA contacts are shown for one mutant CBC complex. Interactions of the remaining two CBCP88L complexes with the DNA are provided in Fig S3 (D) 2Fo–F, electron density maps are shown as gray meshes (contoured to 1σ) for the amino acid residues 84–95 of wt and mutant HapE. Leu88 (green) elongates the N-terminal part of helix α1 by a half turn compared with Pro88 (green).
HapEP88L-mutant CBCs (colored). In the mutant complex, the C-terminal αS helix and the anchor motif of subunit HapB (green) are disordered because of missing interactions with the DNA double strand (Fig 5A). Structural changes induced by P88L (green; arrows) in subunit HapE (red) are depicted.

Materials and Methods

Generation of mutant A. fumigatus strains

Strains AT160P+ (wt), hapEP88L, and ΔhapC have been described previously (Gsaller et al, 2016). To inactivate hapX, a construct containing a pyrithiamine resistance cassette was amplified from genomic DNA of a hapX-deficient strain (strain ΔhapX, background AT577) (Gsaller et al, 2014) with primers aAHapX-1 (AGC GAC TAT AGC CGT ATG) and aAHapX-2 (CCT TGG GTC TTG AAG CTT GCG) and transformed into an AT160P+ wt or hapEP88L recipient strain, respectively. Homologous recombination events yielded strains ΔhapX and hapEP88LΔhapX.

Growth analysis of A. fumigatus

Growth assays were performed in Aspergillus minimal medium (1% [wt/vol] glucose, 20 mM glutamine, salt solution, and iron-free trace elements) according to previous reports (Pontecorvo et al, 1953).

Measurement of siderophore production

Fungal strains were grown in liquid cultures under iron limitation conditions. After 24 h, the culture supernatants were transferred to new reaction tubes and saturated with FeSO₄. Next, 0.2 volumes of phenol-chloroform:isoamyl alcohol (25:24:1, PCI) was added for extraction of total extracellular siderophores (TAFc and FsC). After centrifugation, the PCI phase was mixed with five volumes of diethyl ether and one volume of water. In the last step, the upper diethyl ether containing phase was removed, and the amount of TAFc + FsC in the aqueous phase was quantified spectrophotometrically (NanoDrop2000; Thermo Fisher Scientific) using a molar extinction coefficient of ε = 2,996 M⁻¹ cm⁻¹ at 440 nm.

Overproduction and purification of recombinant CBC complexes

The A. nidulans CBC consisting of Hap8293–294, HapC42–132, and HapE137–164 was produced and purified as described (Gsaller et al, 2014). Briefly, synthetic genes coding for the conserved core domains of HapB, HapC, HapE, or HapEP88L were sequentially cloned in the pnCS vector for expression of a polycistronic transcript (Diebold et al, 2011). The expression plasmids were transformed in Escherichia coli BL21(DE3). After overnight autoinduction and cell lysis, the heterotrimERIC wt CBC and the CBCP88L mutant were purified to homogeneity by subsequent chelate affinity and SEC. The equivalent A. fumigatus wt and HapEP88L mutant CBCs (Hap8290–292, HapC40–130, and HapE137–164) were produced the same way. Size exclusion fractions containing pure CBCs were pooled based on SDS–PAGE analysis, concentrated by ultrafiltration (Amicon Ultra-15 10K centrifugal filter device; Millipore) to 16–18 mg ml⁻¹, aliquoted, flash-frozen in liquid nitrogen, and stored at -80°C.

SPR measurements

Real-time SPR protein–DNA interaction measurements were performed by previously published protocols (Gsaller et al, 2016). Notably, for cooperative CBC–HapX binding analysis measured by SPR co-injection on the A. fumigatus cccA promoter motif, A. fumigatus wt and HapEP88L mutant CBCs consisting of the Hap8290–299, HapC40–130, and HapE137–164 subunits were used. The A. fumigatus HapX137–164 bZIP peptide (covering the CBC-binding domain, basic region, and coiled-coil domain) was produced and purified as previously described (Gsaller et al, 2014).

Preparation of CBC–DNA complexes for crystallization

Oligonucleotides were produced by chemical synthesis of the forward and reverse strands (Biomers). These oligonucleotides were dissolved in annealing buffer (10 mM Tris/HCl and 50 mM NaCl, pH 8.0) at a concentration of 200–600 µM.
concentration of 5 mM and annealed by mixing equal volumes of each strand to yield a final DNA duplex concentration of 2.5 mM. The DNA was heated to 95°C for 5 min and allowed to cool slowly to room temperature. Purified CBCs were added to a 1.2-fold molar excess of the respective DNA duplex. CBC (wt)–DNA mixtures were subjected to crystallization without additional purification steps. CBCP88L–DNA preparations were further purified by SEC in 20 mM Tris/HCl, 150 mM NaCl, 1 mM DTT, pH 7.5, using a Superdex prep grade 75 16/60 column (GE Healthcare). The presence of all three CBC subunits and DNA in the collected main fraction was verified by a dual stain method that allows independent visualization of the protein and nucleic acid species (Pryor et al, 2012). In brief, SDS–PAGE gels were first washed with water followed by staining for protein with GelCode Blue Stain Reagent (Thermo Fisher Scientific). Next, the gels were again washed with water, followed by staining with 1× SYBR Gold Nucleic Acid Gel Stain (Invitrogen). SEC-purified CBCP88L–DNA preparations were concentrated 10-fold by ultrafiltration (Amicon Ultra-15 30K centrifugal filter device; Millipore) and subjected to crystallization.

### Crystallization and structure determination

All complexes were crystallized by the sitting drop vapor diffusion technique at 20°C. Crystal drops (0.4 μl) contained equal volumes of

| Crystal parameter | Afu_CBC–cycA | Afu_CBC–cccA | An_CBC–cccA | An_CBC HapE<sup>PRBL</sup>–cycA |
|-------------------|-------------|-------------|-------------|---------------------------|
| Space group       | P2,2,2,1    | P2,2,2,1    | P2,2,2,1    | P2,2,2,1                  |
| Cell constants    | a = 51.57 Å | a = 51.48 Å | a = 65.68 Å | a = 72.15 Å               |
|                   | b = 75.51 Å | b = 83.74 Å | b = 72.01 Å | b = 103.50 Å              |
|                   | c = 142.94 Å| c = 148.42 Å| c = 148.45 Å| c = 159.71 Å              |
| Subunits/AU<sup>a</sup> | 1 Af_CBC | 1 Af_CBC | 1 An_CBC | 3 An_CBCs |
|                   | 1 DNA duplex | 1 DNA duplex | 1 DNA duplex | 1 DNA duplex |
| Data collection   |             |             |             |                           |
| Beam line         | X06SA, SLS  | X06SA, SLS  | X06SA, SLS  | X06SA, SLS                |
| Wavelength (Å)    | 1.0         | 1.0         | 1.0         | 1.0                       |
| Resolution range (Å)<sup>b</sup> | 48–2.6 (2.7–2.6) | 48–2.3 (2.4–2.3) | 48–2.2 (2.3–2.2) | 49–2.3 (2.4–2.3) |
| No. of observations | 63,420      | 157,892     | 169,021     | 271,549                   |
| No. of unique reflections<sup>c</sup> | 17,526      | 29,239      | 36,047      | 52,851                    |
| Completeness (%)<sup>b</sup> | 98.4 (99.6) | 99.7 (99.8) | 98.7 (99.3) | 98.0 (98.4)               |
| R<sub>merge</sub> (%)<sup>d</sup> | 4.5 (50.9)  | 4.4 (58.9)  | 5.1 (56.5)  | 7.0 (59.8)                |
| I/σ(I)<sup>b</sup> | 19.0 (2.6)  | 212.0 (2.9) | 16.0 (2.2)  | 10.5 (1.7)                |
| Refinement (REFMACS) |             |             |             |                           |
| Resolution range (Å) | 30–2.6 | 30–2.3 | 30–2.2 | 30–2.3 |
| No. of refl. working set | 16,636 | 27,765 | 34,232 | 50,185 |
| No. of refl. test set | 876 | 1,461 | 1,802 | 2,641 |
| No. of non-hydrogen | 3,132 | 3,167 | 3,357 | 6,811 |
| Solvent (H<sub>2</sub>O, Cl<sup>−</sup>) | 25 | 69 | 124 | 61 |
| R<sub>work</sub>/R<sub>free</sub> (%)<sup>e</sup> | 19.2/23.2 | 18.7/21.1 | 19.5/21.8 | 22.1/25.4 |
| rmsd bond/angle (Å)/(°)<sup>f</sup> | 0.002/0.991 | 0.003/1.011 | 0.002/1.033 | 0.007/1.331 |
| Average B-factor (Å<sup>2</sup>) | 64.0 | 64.2 | 55.5 | 68.2 |
| Ramachandran plot (%)<sup>g</sup> | 98.0/2.0/0.0 | 98.3/1.7/0.0 | 98.5/1.5/0.0 | 97.5/2.5/0.0 |
| PDB accession code | 6Y35 | 6Y36 | 6Y37 | 6Y39 |

<sup>a</sup>Asymmetric unit.

<sup>b</sup>The values in parentheses for resolution range, completeness, R<sub>merge</sub>, and I/σ(I) correspond to the highest resolution shell.

<sup>c</sup>Data reduction was carried out with XDS and from a single crystal. Friedel pairs were treated as identical reflections.

<sup>d</sup>R<sub>merge</sub>(I) = Σ<sub>hkL</sub>Σ<sub>j</sub>|I<sub>hkL</sub>|j−<I<sub>hkL</sub>><|/Σ<sub>hkL</sub>Σ<sub>j</sub>I<sub>hkL</sub>|j, where I<sub>hkL</sub> is the jth measurement of the intensity of reflection hkl and <I<sub>hkL</sub>>< is the average intensity.

<sup>e</sup>R = Σ<sub>hkL</sub>||F<sub>obs</sub>|−|F<sub>calc</sub>|/Σ<sub>hkL</sub>|F<sub>obs</sub>|, where R<sub>free</sub> is calculated without a sigma cutoff for a randomly chosen 5% of reflections, which were not used for structure refinement and R<sub>work</sub> is calculated for the remaining reflections.

<sup>f</sup>Deviations from ideal bond lengths/angles.

<sup>g</sup>Percentage of residues in favored region/allowed region/outlier region.
the protein–DNA complex (13–15 mg ml−1) and reservoir solution. All DNA fragments that encoded promoter sequences of either cytochrome c (cycA) or the vacuolar iron transporter (cccA) were 23-bp long and carried 5' AA-TT overhangs.

Crystals of the An_CBC-cccA complex grew from 0.2 M ammonium acetate, 0.1 M 4-(2-hydroxyethyl)-1-piperazineethanesulphonic acid (Hepes), pH 7.5, and 25% (wt/vol) polyethylene glycol (PEG) 3350. The Afu_CBC-cycA structure was obtained from 0.1 M MES, pH 6.5, and 25% (wt/vol) PEG8000 and the Afu_CBC-cccA complex crystallized from 0.1 M MES, pH 6.5, and 25% (wt/vol) PEG6000. Crystals of the An_CBCcccA-cycA complex grew from conditions containing 175 mM di-ammonium phosphate and 19% (wt/vol) PEG3350. All crystals were cryoprotected by the addition of a 1:1 (vol/vol) mixture of mother liquor and 70% (vol/vol) glycerol and subsequently super-cooled in a stream of nitrogen gas at 100 K. Diffraction data were collected at the beamline X06SA, Swiss Light Source at λ = 1.0 Å. Reflection intensities were analyzed with the program package XDS (Kabsch, 1993). Structure determination was performed by Patterson search calculations with PHASER (McCoy et al, 2007) using the coordinates of either wt A. nidulans CBC without DNA (PDB ID 4G91 [Huber et al, 2012]) or bound DNA (PDB ID 4G92 [Huber et al, 2012]). Cyclic refinement and model building steps were performed with REFMACS (Vagin et al, 2004) and Coot (Emsley et al, 2010). Water molecules were placed with ARP/wARP solvent (Ferrarini et al, 1997). Translation/libration/screw refinements finally yielded good values for Rcryst and Rfree as well as r.m.s.d. bond and angle values. The models were proven to fulfill the Ramachandran plot using PROCHECK (Laskowski et al, 1993) (Table 1). The DNA-bending angle was analyzed with the Curves+ and SUMR algorithms (Lavery et al, 2009). Graphical illustrations were created with the UCSF Chimera package from the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco (Pettersen et al, 2004). Coordinates and structure factors have been deposited in the Protein Data Bank (for entry codes see Table 1).

Data Availability

Coordinates and structure factors have been deposited in the Protein Data Bank under the entry codes 6Y35 (Afu_CBC-cccA), 6Y36 (Afu_CBC-cccA), 6Y37 (An_CBC-cccA), and 6Y39 (An_CBC_HapEcccA–cycA).

Supplementary Information

Supplementary Information is available at https://doi.org/10.26508/lsa.202000729.

Acknowledgements

This work was funded by the joint D-A-CH program “Novel molecular mechanisms of iron sensing and homeostasis in filamentous fungi” (Deutsche Forschungsgemeinschaft [DFG] to AA Brakhage [BR 1130/14-1], MJ Bromley [GR 1861/8-1], and P Hortschansky [HO 2596/1-1]) and Austrian Science Foundation [FWF-I1346] to H Haas). MJ Bromley received funding from the Wellcome Trust grant 208396/Z/17/Z and the National Institutes of Health (R01AI143198-01). EM Huber acknowledges funding by the DFG–SFB 1309–325870/75 and the Young Scholar’s Program of the Bavarian Academy of Sciences and Humanities. We thank the staff of the beamline X06SA at the Paul Scherrer Institute, Swiss Light Source, Villigen Switzerland for assistance during data collection, and acknowledge funding from the European Community’s Seventh Framework Programme (FP7/2007-2013) under BioStruct-X (grant agreement no. 283570).

Author Contributions

P Hortschansky: funding acquisition, investigation, visualization, project administration, and writing—review and editing.
M Misslinger: investigation and writing—review and editing.
J Mörl: investigation.
F Gailer: investigation and writing—review and editing.
M) Bromley: resources, supervision, funding acquisition, and writing—review and editing.
AA Brakhage: conceptualization, funding acquisition, and writing—review and editing.
M Groll: conceptualization, funding acquisition, investigation, and writing—review and editing.
H Haas: conceptualization, supervision, funding acquisition, project administration, and writing—review and editing.
EM Huber: funding acquisition, investigation, visualization, project administration, and writing—original draft, review, and editing.

Conflict of Interest Statement

The authors declare that they have no conflict of interest.

References

Arendrup MC, Mavridou E, Mortensen KL, Snelders E, Frimodt-Moller N, Khan H, Melchers WJ, Verweij PE (2010) Development of azole resistance in Aspergillus fumigatus during azole therapy associated with change in virulence. PLoS One 5: e10080. doi:10.1371/journal.pone.0010080
Becher R, Winkel SG (2012) Fungal cytochrome P450 sterol 14α-demethylase (CYP51) and azole resistance in plant and human pathogens. Appl Microbiol Biotechnol 95: 825–840. doi:10.1007/s00253-012-4199-5
Bhattacharya A, Deng JM, Zhang Z, Behringer R, de Crombrugge B, Maity SN (2003) The B subunit of the CCAAT box binding transcription factor complex (CBF/NF-Y) is essential for early mouse development and cell proliferation. Cancer Res 63: 8167–8172.
Bongomin F, Gago S, Oladele RO, Denning DW (2017) Global and multi-national prevalence of fungal diseases-estimate precision. J Fungi (Basel) 3: 57. doi:10.3390/jof3040057
Bowyer P, Denning DW (2014) Environmental fungicides and triazole resistance in Aspergillus. Pest Manag Sci 70: 173–178. doi:10.1002/ps.3567
Buchner P (1990) Weight matrix descriptions of four eukaryotic RNA polymerase II promoter elements derived from 502 unrelated promoter sequences. J Mol Biol 212: 563–578. doi:10.1016/0022-2836(90)90223-9
Bueid A, Howard SJ, Moore CB, Richardson MD, Harrison E, Bowyer P, Denning DW (2010) Azole antifungal resistance in Aspergillus fumigatus: 2008 and 2009. J Antimicrob Chemother 65: 2116–2118. doi:10.1093/jac/dkq279
Camps SMT, Dutilh BE, Andrenec MC, Rjjs AJM, Snelders E, Huynen MA, Verweij PE, Melchers WJG (2012a) Discovery of a HapE mutation that causes azole resistance in Aspergillus fumigatus through whole genome sequencing and sexual crossing. PLoS One 7: e50034. doi:10.1371/journal.pone.0050034
Kabsch W (1993) Automatic processing of rotation diffraction data from crystals of initially unknown symmetry and cell constants. J Appl Crystallogr 26: 795–800. doi:10.1107/S0021889893005588
Kim MK, Kang YK (1999) Positional preference of proline in alpha-helices. Protein Sci 8: 1492–1499. doi:10.1021/ps971492
Laskowski RA, Moss DS, Thornton JM (1993) Main-chain bond lengths and bond angles in protein structures. J Mol Biol 231: 1049–1067. doi:10.1006/jmbi.1993.1351
Lavery R, Moakher M, Maddocks JH, Petkевич D, Zakrzewska K (2009) Conformational analysis of nucleic acids revisited: Curves+. Nucleic Acids Res 37: 5917–5929. doi:10.1093/nar/gkp608
Lelievre L, Groh M, Angebault C, Maherault AC, Didier E, Bougnoux ME (2013) Azole resistant Aspergillus fumigatus: An emerging problem. Med Mycol 43: 139–145. doi:10.1093/medjmed.2013.2.0.10
Marino-Ramirez L, Spouge JL, Kanga GC, Landsman D (2004) Statistical analysis of over-represented words in human promoter sequences. Nucleic Acids Res 32: 949–958. doi:10.1093/nar/gkh246
McCoy AJ, Grosse-Kunstleve RW, Adams PD, Winn MD, Storoni LC, Read RJ (2007) Phaser crystallographic software. J Appl Crystallogr 40: 658–674. doi:10.1107/s0021889807021206
Meis JF, Chowdhary A, Rhodes JL, Fisher MC, Verweij PE (2016) Clinical implications of globally emerging azole resistance in Aspergillus fumigatus. Philos Trans R Soc Lond B Biol Sci 371: 20150460. doi:10.1098/rstb.2015.0460
Monk BC, Sagatova AA, Hosseini P, Ruma YN, Wilson RK, Keniya MV (2020) Fungal lanosterol 14alpha-demethylase: A target for next-generation antifungal design. Biochim Biophys Acta Proteins Proteom 1868: 140206. doi:10.1016/j.bbapap.2019.02.008
Nardini M, Gnesutta N, Donati G, Gatta R, Forni C, Fossati A, Vonrhein C, Moras D, Romier C, Bolognesi M, et al (2013) Sequence-specific transcription factor NF-Y displays histone-like DNA binding and H2B-like ubiquitination. Cell 152: 132–143. doi:10.1016/j.cell.2012.11.047
Odds FC, Brown AJP, Gow NAR (2003) Antifungal agents: Mechanisms of action. Trends Microbiol 11: 272–279. doi:10.1016/S0966-842X(03)00117-3
Paul S, Diekema D, Moye-Rowley WS (2017) Contributions of both ATP-binding cassette transporter and Cyp51A proteins are essential for azole resistance in Aspergillus fumigatus. Antimicrob Agents Chemother 61: e02748–16. doi:10.1128/AAC.02748-16
Paul S, Stamnes M, Thomas GH, Liu H, Hagwaria D, Gomi K, Filler SG, Moye-Rowley WS (2019) ABR is an essential determinant of azole resistance in Aspergillus fumigatus. mBio 10: e02563–18. doi:10.1128/mBio.02563–18
Perrakis A, Sixma TK, Wilson KS, Lanzin VS (1997) wARP: Improvement and extension of crystallographic phases by weighted averaging of multiple-refined dummy atomic models. Acta Crystallogr D Biol Crystallogr 53: 448–455. doi:10.1107/s0907444997005596
Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, Ferrin JE (2004) UCSF Chimera: A visualization system for exploratory research and analysis. J Comput Chem 25: 1605–1622. doi:10.1002/jcc.20084
Pontecorvo G, Roper JA, Hemmons LM, Macdonald KD, Bufton AWJ (1953) The genetics of Aspergillus nidulans. Adv Genet 5: 141–238. doi:10.1016/S0065-2660(08)60408-3
Pryor EJ, Jr, Wozniak DJ, Hollis T (2012) Crystallization of Pseudomonas aeruginosa AmrZ protein: Development of a comprehensive method for obtaining and optimization of protein-DNA crystals. Acta Crystallogr Sect F Struct Biol Cryst Commun 68: 985–993. doi:10.1107/s1744309112025316
Richardson JS, Richardson DC (1988) Amino acid preferences for specific locations at the ends of alpha helices. Science 240: 1648–1652. doi:10.1126/science.3381086
Schrettl M, Beckmann N, Varga J, Heiniekamp T, Jacobsen ID, Jochl C, Moussa TA, Wang S, Gsaller F, Blatzer M, et al (2010) HapX-mediated adaption to iron starvation is crucial for virulence of Aspergillus fumigatus. PLoS Path 6: e10001124. doi:10.1371/journal.ppat.10001124
Snelders E, Karawajczyk A, Schaftenaar G, Verweij PE, Melchers WJG (2010) Azole resistance profile of amino acid changes in Aspergillus fumigatus CYP51A based on protein homology modeling. Antimicrob Agents Chemother 54: 2425–2430. doi:10.1128/aac.01599-09

Snelders E, Karawajczyk A, Verhoeven RJA, Venselaar H, Schaftenaar G, Verweij PE, Melchers WJG (2011) The structure-function relationship of the Aspergillus fumigatus cyp51A L98H conversion by site-directed mutagenesis: The mechanism of L98H azole resistance. Fungal Genet Biol 48: 1062–1070. doi:10.1016/j.fgb.2011.08.002

Snelders E, van der Lee HAL, Kuijpers J, Rijs AJMM, Varga J, Samson RA, Mellado E, Donders AR, Melchers WJG, Verweij PE (2008) Emergence of azole resistance in Aspergillus fumigatus and spread of a single resistance mechanism. PLoS Med 5: e219. doi:10.1371/journal.pmed.0050219

Vagin AA, Steiner RA, Lebedev AA, Potterton I, McNicholas S, Long F, Murshudov GN (2004) REFMAC5 dictionary: Organization of prior chemical knowledge and guidelines for its use. Acta Crystallogr D Biol Crystallogr 60: 2184–2195. doi:10.1107/s0907444904023510

Verweij PE, Snelders E, Kema GHI, Mellado E, Melchers WJG (2009) Azole resistance in Aspergillus fumigatus: A side-effect of environmental fungicide use? Lancet Infect Dis 9: 789–795. doi:10.1016/s1473-3099(09)70265-8

Zemzoumi K, Frontini M, Bellorini M, Mantovani R (1999) NF-Y histone fold alpha1 helices help impart CCAAT specificity. J Mol Biol 286: 327–337. doi:10.1006/jmbi.1998.2496

License: This article is available under a Creative Commons License (Attribution 4.0 International, as described at https://creativecommons.org/licenses/by/4.0/).