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CD47 plays a critical role in T-cell recruitment by regulation of LFA-1 and VLA-4 integrin adhesive functions

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ABSTRACT CD47 plays an important but incompletely understood role in the innate and adaptive immune responses. CD47, also called integrin-associated protein, has been demonstrated to associate in cis with β1 and β3 integrins. Here we test the hypothesis that CD47 regulates adhesive functions of T-cell α4β1 (VLA-4) and αLβ2 (LFA-1) in vivo and in vitro models of inflammation. Intravital microscopy studies reveal that CD47−/− Th1 cells exhibit reduced interactions with wild-type (WT) inflamed cremaster muscle microvessels. Similarly, murine CD47−/− Th1 cells, as compared with WT, showed defects in adhesion and transmigration across tumor necrosis factor-α (TNF-α)–activated murine endothelium and in adhesion to immobilized intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1) under flow conditions. Human Jurkat T-cells lacking CD47 also showed reduced adhesion to TNF-α–activated endothelium and ICAM-1 and VCAM-1. In cis interactions between Jurkat T-cell β2 integrins and CD47 were detected by fluorescence lifetime imaging microscopy. Unexpectedly, Jurkat CD47 null cells exhibited a striking defect in β1 and β2 integrin activation in response to Mn\textsuperscript{2+} or Mg\textsuperscript{2+}/ethylene glycol tetraacetic acid treatment. Our results demonstrate that CD47 associates with β2 integrins and is necessary to induce high-affinity conformations of LFA-1 and VLA-4 that recognize their endothelial cell ligands and support leukocyte adhesion and transendothelial migration.

INTRODUCTION

CD47 is a ubiquitously expressed 50-kDa transmembrane glycoprotein with a single immunoglobulin G (Ig)-like domain, a hydrophobic, five transmembrane-spanning segment, and a short hydrophobic cytoplasmic tail (Brown et al., 1990). CD47 has also been called integrin-associated protein because of its demonstrated ability to interact in cis with αvβ3, αvβ5, α2β1, and α4β1 integrins in nonleukocyte cell types (reviewed in Brown and Frazier, 2001). CD47 also interacts in trans with signal regulatory proteins (SIRPs) and thrombospondin (TSP; reviewed in Barclay, 2009). CD47 is involved in a broad range of important physiological processes, including leukocyte phagocytosis, recognition of “self,” immune cell homeostasis, cell migration and regulation, leukocyte transendothelial and transepithelial...
migration, platelet adhesion and activation, and nitric oxide signaling (Brown and Frazier, 2001; Isenberg et al., 2008).

Previous studies in CD47−/− mice established that CD47 plays a role in neutrophil emigration in a bacteria-induced peritonitis model (Lindberg et al., 1996), a lipopolysaccharide-induced, neutrophil-mediated acute lung injury and a bacterial pneumonia model (Su et al., 2008), and in vitro models of human neutrophil and monocyte transmigration across endothelium (Cooper et al., 1995; de Vries et al., 2002) and epithelium (Parkos et al., 1996). CD47 also was implicated in dendritic cell recruitment in a trinitrobenzenesulfonic acid–induced colitis model of hapten-stimulated inflammation (Fortin et al., 2009) and T-cell activation in the myelin oligodendrocyte glycoprotein–induced experimental autoimmune encephalomyelitis model (Han et al., 2012). We recently reported that human endothelial CD47 interacts with T-cell–expressed SIRPγ during T-cell transendothelial migration (TEM) under flow conditions in vitro (Stefanidakis et al., 2008). We also reported that CD47−/− mice showed reduced recruitment of blood T-cells, neutrophils, and monocytes in a dermal air pouch model of tumor necrosis factor-α (TNF-α)–induced inflammation and both endothelial- and leukocyte-expressed CD47s were required (Azcutia et al., 2012). On the basis of our findings that T-cells show significantly reduced recruitment in murine models in vivo and in human and murine in vitro models of inflammation, we investigated whether the defect is related to loss of CD47-dependent integrin adhesive functions.

The results of our study indicate that CD47 associates with T-cell β2 integrins as assessed by fluorescence lifetime imaging microscopy (FLIM) and CD47 is necessary for induction of VLA-4 and LFA-1 integrin high-affinity conformations that bind to their ligands vascular cell adhesion protein 1 (VCAM-1) and intercellular adhesion molecule 1 (ICAM-1), respectively. Our results suggest that in cis CD47–integrin associations, together with the previously reported in trans CD47–SIRP interactions (Stefanidakis et al., 2008), are important for both adhesion and transmigration across the vascular endothelium in vivo and in vitro models of inflammation.

RESULTS
CD47−/− Th1 cells exhibit defective adhesive interactions with TNF-α–activated endothelium in the murine cremaster microvasculature
To evaluate the role of CD47 in leukocyte adhesive interactions with endothelium in vivo, we performed intravital microscopy studies in the murine cremaster microcirculation after intracranial injection of TNF as described earlier (Alcaide et al., 2012). Equal numbers of in vitro–generated wild-type (WT) and CD47−/− Th1 effector cells, each labeled with a different color fluorescent dye, were coinjected into WT recipient animals to enable comparison of WT and CD47−/− Th1 cells in the same postcapillary venules. The microvessel parameters for WT and CD47−/− mice are listed in Table 1. The behavior of transferred Th1 cells was monitored for 2–5 min postinjection. CD47−/− Th1 cells exhibited significantly reduced tethering adhesive interactions relative to WT Th1 cells (Figure 1A). The reciprocal experiment of simultaneous injection of differentially fluorescent-labeled WT and CD47−/− Th1 cells into CD47−/− recipient mice revealed that CD47−/− Th1 cells interacted less than did WT cells. Tethered Th1 cells become stably bound to the inflamed vessel wall for more than three consecutive video frames (~2 s). Few, if any, of the transferred T-cells arrest for more than several seconds. These results indicate expression of CD47 in T-cells is required for optimal Th1 effector cell adhesive interactions in this model. There was no difference between WT and CD47−/− T-cells in the rolling velocities, indicating that CD47 was not involved in Th1 cell rolling on inflamed venules (Figure 1B). This adhesion defect is not explained by reduced expression of adhesion molecules in CD47−/− mice because WT and CD47−/− Th1 cells express identical levels of LFA-1, VLA-4, and P-selectin glycoprotein ligand-1 (PSGL-1) and similar levels of intracellular IFN-γ, the Th1 signature cytokine (Figure 1, C and D). We routinely verified the absence of CD47 expression in CD47−/− T-cells (Figure 1C).

CD47 regulates Th1 effector cell adhesion and transmigration in vitro
To further delineate the adhesion defect in CD47−/− T-cells, we monitored adhesion and TEM of WT and CD47−/− Th1 cells on WT and CD47−/− murine heart endothelial cell (MHEC) monolayers by live-cell videomicroscopy in an in vitro flow model (Alcaide et al., 2012). WT Th1 cells arrested, and subsequently ~30% of adherent cells transmigrated across TNF-α–activated WT monolayers (Figure 2, A and B). As we would predict from the in vivo studies, CD47−/− Th1 cells exhibited reduced adhesion and TEM across WT endothelium compared with WT Th1 cells. CD47−/− T-cells also showed reduced adhesion and TEM across CD47−/− MHEC monolayers. Of interest, WT Th1 cells also exhibited significantly reduced adhesion and TEM across CD47−/− MHECs, indicating that endothelial cell CD47 also plays a role in TEM. Analysis of MHECs isolated from CD47−/− and WT mice showed essentially identical levels of surface-expressed PECAM-1 and ICAM-2 at baseline and similar levels of ICAM-1, VCAM-1, and E-selectin expression at baseline and after 4 h of TNF-α treatment (Figure 2C). These in vivo and in vitro studies indicate that expression of CD47 in both T-cells and endothelium is required for normal Th1 cell adhesion and TEM of TNF-α–inflamed endothelium. Here we focus on the role that CD47 expressed on T-cells plays in leukocyte recruitment.

CD47 regulates Th1 cell adhesion to immobilized ICAM-1 and VCAM-1
CD47 associates with and regulates the adhesive functions of reticulocyte-expressed α4β1 integrins (Brittain et al., 2004), but no study examined whether CD47 regulates αLβ2 (LFA-1) integrin adhesive interactions with its endothelial ligand ICAM-1. We therefore studied the adhesion of WT and CD47−/− Th1 cells to ICAM-1-Fc immobilized with CXCL12 (SDF-1α) chemokine under flow conditions (Alcaide et al., 2012). WT Th1 cells arrested on ICAM-1, whereas CD47−/− Th1 cells showed a significant reduction in arrest (Figure 3A). Of note, the greatest relative reduction occurred at the highest shear stress level examined. This assay was repeated using immobilized VCAM-1-Fc molecules without chemokine because T-cells constitutively express VLA-4 integrins capable of binding VCAM-1 under flow conditions (Chen et al., 2012). A significant defect in CD47−/− Th1 cell arrest to VCAM-1 was observed at high shear stress levels but not at the lowest shear stress tested (Figure 3B). There were no differences between WT and CD47−/− Th1 cell accumulation on immobilized E-selectin, suggesting that induction of Th1 selectin ligand expression was not affected (Figure 3C). Taken together, the data suggest that CD47 in Th1 cells plays an important role in the function of LFA-1 and VLA-4 integrins required for adhesion to their endothelial cell ligands.

CD47 does not regulate LFA-1 adhesion-strengthening ICAM-1
Because CD47−/− Th1 cells showed a larger defect in adhesion to ICAM-1 compared with VCAM-1, we focused on LFA-1 and determined whether adhesion strengthening to ICAM-1, termed avidity regulation, was affected. Accordingly, we compared the ability of adherent WT and CD47−/− Th1 cells to resist detachment
difference in the rate of cell detachment was detected upon applying the shear flow regime (Figure 3D). This finding demonstrates that CD47 is not necessary for postadhesion LFA-1 adhesion strengthening to ICAM-1.

**CD47-null human T-cells phenocopy the defects in murine CD47−/− Th1-cell adhesion**

An earlier study reported that human Jurkat T-cell clone JINB8 lacking CD47 (CD47−) showed reduced binding to a TNF-α-stimulated human endothelial-like cell line as compared with the parental clone expressing CD47 (CD47+; Ticchioni et al., 2001). Accordingly, we evaluated their adhesion to TNF-α–activated human umbilical vein endothelial cells (HUVECs) and immobilized ICAM-1 and VCAM-1. Both Jurkat clones express similar surface levels of LFA-1 and VLA-4 α-subunits and the respective common β2 and β1 integrin subunits (Figure 4A). CD47+ Jurkat cells showed significantly greater adhesion to 4-h TNF-α–activated HUVEC monolayers than did CD47− cells at each shear stress tested (Figure 4B). Of interest, the dominant integrin responsible for T-cell adhesion was VLA-4 as determined by function-blocking monoclonal antibody (mAb) studies (Figure 4C), which is consistent with earlier studies (Chen et al., 1999; Ticchioni et al., 2001). Blocking LFA-1 on CD47+ Jurkat cells reduced adhesion to similar levels as unblocked CD47− Jurkat cells. Blockade of LFA-1 on CD47− Jurkat cells did not further reduce adhesion to HUVECs. The lesser role of LFA-1 in Jurkat T-cell adhesion to HUVECs is likely the reason that Jurkat CD47− T-cell adhesion is not reduced to a greater level, as we would have predicted (Figure 4C).

We next evaluated Jurkat T-cell adhesion to immobilized ICAM-1, VCAM-1, and E-selectin under the same conditions as in Figure 3. CD47− T-cells showed significantly reduced stable arrest to similar levels as unblocked CD47+ Jurkat cells. Blockade of LFA-1 on CD47− Jurkat cells did not further reduce adhesion to HUVECs. The lesser role of LFA-1 in Jurkat T-cell adhesion to HUVECs is likely the reason that Jurkat CD47− T-cell adhesion is not reduced to a greater level, as we would have predicted (Figure 4C).

from ICAM-1 immobilized with CXCL12 under increasing flow rates (Sircar et al., 2007). Whereas the number of CD47− Th1 cells initially bound under the lowest shear flow setting was 31% lower than with WT (264 ± 86 cells/mm² for WT adhesion vs. 182 ± 60 cells/mm² for CD47−, p < 0.05, Student’s t test), no effects in arrest were most pronounced on ICAM-1 compared with VCAM-1, whereas there was no defect in adhesion to E-selectin (Figure 5C). As a control, we created stably transfected CD47− Jurkat cells expressing full-length CD47 tagged with green fluorescent protein (GFP; CD47+GFP) or GFP alone (CD47−GFPcont).
CD47 regulates β1 and β2 high-affinity conformations in response to Mn2+ or Mg2+/ethylene glycol tetraacetic acid

Integrin affinity regulation was evaluated on Jurkat CD47+ and CD47− cells by reporter mAbs that detect activated conformations of β1 and β2 integrins. Kim127 mAb detects and stabilizes an extended and closed conformation of β2 integrins (intermediate affinity), and mAb 24 detects and stabilizes open “active” conformation of β2 integrins (high affinity; Salas et al., 2004). Jurkat T-cells express predominantly LFA-1 (αLβ2) and not the other α-subunit chains that associate with β2 integrins, and hence these mAbs to β2 integrins detect primarily the intermediate- and high-affinity conformations of LFA-1 (Salas et al., 2004). HUTS21 detects and stabilizes activated β1 integrins (Luque et al., 1996), and we showed previously that expression of this epitope correlated with robust adhesion of human memory T-cells to VCAM-1 (Lim et al., 2000). As expected, incubation of CD47+ T-cells with Mn2+ or Mg2+/ethylene glycol tetraacetic acid (EGTA), global activators of integrins, triggered robust expression of both intermediate- and high-affinity LFA-1 (Figure 6, A and B). In contrast, CD47− T-cells showed a significantly reduced expression of intermediate- and high-affinity conformations of LFA-1 in response to Mn2+ or Mg2+/EGTA. It is unlikely that this assay did not detect transient LFA-1 activation because the mAbs stabilize the active conformation and were present throughout the assay. In addition, failure to detect mAb expression was not due to a change (loss) in total surface expression of β1 or β2 integrins upon exposure to Mn2+ or Mg2+/EGTA (Figure 6C). CD47+ T-cells also showed reduced induction of high-affinity β1 integrins versus CD47− T-cells after treatment with Mn2+ or Mg2+/EGTA buffers (Figure 6D). Consistent with the failure of Mn2+ or Mg2+/EGTA treatments to induce intermediate- and high-affinity LFA-1 conformations in the absence of CD47, CD47− T-cells exhibited little binding of soluble ICAM-1-Fc versus CD47+ T-cells (Figure 6E). In addition, treatment of CD47− cells with Mn2+ did not restore binding comparable to CD47+ T-cell adhesion to either immobilized ICAM-1 or VCAM-1 under flow (Supplemental Figure S1, A and B). Taken together, the results indicate that CD47 expression is required for expression of intermediate- and high-affinity conformations of LFA-1 and for binding soluble ICAM-1, as well as for induction of activated β1 integrins.

To determine whether β2 integrins in CD47− cells could be activated when extracted from the membrane milieu of CD47− Jurkat T-cells, we examined the effect of 1.0 mM Mn2+ or β2 integrins in the soluble phase. When detergent lysates of CD47+ and CD47− T-cells were incubated in the presence of Mn2+ and mAb24 to immunoprecipitate activated β2 integrins, an equal and robust

The level of CD47 expression was similar to that for the parental clone (data not shown) and restored arrest to ICAM-1 to a level similar to that for the parent clone, whereas CD47−GFP-containing cell adhesion did not improve (Figure 6D). Finally, we examined the adhesion-strengthening capability of CD47+ and CD47− Jurkat T-cells, using a detachment assay. CD47+ Jurkat T-cells also exhibited 35% reduction in initial binding under the lowest shear stress conditions (211 ± 24 cells/mm² for CD47+ adhesion vs. 137 ± 24 cells/mm² for CD47−; p < 0.05) but no defect in the rate of detachment versus CD47+ cells (Figure 5E), which is consistent with the behavior of murine CD47−/− Th1 cells (Figure 3D).

Figure 2: Th1 effector T-cell interactions with TNF-α-activated murine endothelium in an in vitro flow chamber. (A) The numbers of accumulated and (B) transmigrated T-cells in the videos were quantified by ImageJ software (National Institutes of Health, Bethesda, MD). Data are mean ± SEM. *p ≤ 0.05 and **p ≤ 0.01 vs. WT Th1 cell on WT MHEC. *p ≤ 0.05 are WT Th1 vs. CD47−/− on CD47−/− MHEC (Student’s t test), n = 3 separate experiments. (C) MHECs were treated with medium or medium containing murine TNF-α (100 ng/ml) for 4 h, and CD47, VCAM-1, ICAM-1, E-selectin, ICAM-2, and PECAM-1 expression levels were detected by unlabeled primary mAb followed by staining with a PE-labeled goat anti-rat secondary mAb. Cell fluorescence was determined by FACSCalibur flow cytometry (BD, Franklin Lakes, NJ). Representative histograms of surface expression of molecules are shown from 10 separate experiments.
applied fluorescence lifetime imaging microscopy (FLIM), a quantitative method for determining Förster resonance energy transfer (FRET; Table 2 and Figure 7A), to study whether CD47 is required for full activation of β2 integrins in reticulocytes from sickle cell patients (Brittain et al., 2004). To evaluate whether CD47 associates with β2 integrins in T-cells, we applied fluorescence lifetime imaging microscopy (FLIM), a quantitative method for determining Förster resonance energy transfer (FRET; Table 2 and Figure 7A), to study whether β2 integrin and CD47 are sufficiently close to imply a physical interaction. Epifluorescence images were captured as quality control of the staining (Figure 7B). The lifetime of the donor molecule (τ1; picoseconds), in this case β2 integrin, labeled with Alexa Fluor 488–conjugated secondary antibody was determined first in the absence of an acceptor fluorophore (Table 2 and Figure 7A, donor only). FRET between the donor fluorophore (β2 integrin) and acceptor (CD47 directly labeled with Alexa Fluor 594) was defined by the lifetime of the donor (τm) for β2 integrin–CD47 associates with and regulates α4β1 integrins in reticulocytes (Brittain et al., 2004). On the basis of this report and our findings that mouse T-cell recruitment requires CD47 expression in a dermal air pouch model of inflammation (Azcutia et al., 2012), we used molecular imaging analysis, in vitro flow chamber adhesion assays, and in

\[ \text{signal was detected in the absence and presence of CD47 (Figure 6F, lanes 3 and 6). mAb 24 did not immunoprecipitate β2 integrins in either cell type in the absence of Mn}^{2+} \text{ (lanes 2 and 5). As a control, immunoprecipitation with mAb TS1/18, which recognizes all} \beta_2 \text{ integrins, is shown in lanes 1 and 4. These data indicate that CD47 is required for full activation of β2 integrins in the plasma membrane milieu of human T-cells but not with detergent-solubilized β2 integrins.} \]

\[ \text{CD47 interacts with β2 integrin} \]

Purified CD47–β2 integrin–CD47, and the α1 value indicates that 31.7 ± 4.1% of β2 integrin molecules interact with CD47 on the cellular membrane of Jurkat T-cells (Table 2). For comparison purposes the noninteracting molecules β2 integrin and PSGL-1 were costained as donor and acceptor molecules, respectively. A decrease of τ1 was found when PSGL-1 was used as acceptor, but the interacting fraction was only 13.3 ± 1.5% and approached the lower levels of sensitivity. Furthermore, the τm of the donor was unchanged in the presence of the acceptor, indicating that β2 integrin and PSGL-1 were not in sufficient proximity to support a physical interaction (Supplemental Table S1 and Supplemental Figure S2A). As a positive control a decrease in τ1 was observed between LFA-1 α-chain (αL integrin) and the common β2 chain (Supplemental Table S1 and Supplemental Figure 2A). Representative histograms illustrate the τm of the donor β2 integrin in the different conditions (Supplemental Figure S3, A–F). These results support a direct association of CD47 with β2 integrin.

We next asked whether the CD47–β2 integrin association was altered upon incubation of T-cells with Mg\(^{2+}\)/EGTA–containing medium, using the same donor and acceptor antibodies. Of interest, there was no change in τ1 (and τm) for β2 integrin–CD47, still indicating a close association between β2 integrin and CD47. The α1 values, however, indicate that significantly fewer β2 integrin molecules interact with CD47 (Table 2). This decrease in association was not due to a change (loss) in total surface expression of β1 or β2 integrins upon exposure to Mn\(^{2+}\) or Mg\(^{2+}\)/EGTA (Figure 6C). We next explored whether the extended or fully activated β2 integrin conformations interact with CD47 by staining cells with mAb 24 and KIM127 incubated in medium containing Mg\(^{2+}\)/EGTA or medium alone. Under conditions of medium with Mg\(^{2+}\)/EGTA or medium alone, we were unable to detect a signal above background for the extended conformation (KIM127). Although the signal for the fully activated β2 integrin (mAb 24) was also too low to detect in medium alone, the signal was clearly detected in cells incubated in medium containing Mg\(^{2+}\)/EGTA. Reduced τ1 (and τm) for β2 integrin–CD47 indicates close association between activated β2 integrins and CD47, and the α1 value indicates that 28 ± 1% of activated β2 integrin molecules interact with CD47 on the cellular membrane of Jurkat T-cells (Table 3). These data indicate that the fully activated β2 integrins do interact with CD47, but the results do not distinguish whether the fraction of fully activated integrins associated with CD47 increases or decreases upon cation-induced integrin activation.

**DISCUSSION**

CD47 associates with and regulates α4β1 integrins in reticulocytes (Brittain et al., 2004). On the basis of this report and our findings that mouse T-cell recruitment requires CD47 expression in a dermal air pouch model of inflammation (Azcutia et al., 2012), we used molecular imaging analysis, in vitro flow chamber adhesion assays, and in

![Image](https://example.com/image.png)
CD47 regulates β2 integrin affinity

CD47 is required for LFA-1 and VLA-4 high-affinity conformation expression

LFA-1 is predicted to assume at least three distinct conformational states: a bent structure with low affinity for ligand, an extended and closed headpiece structure with intermediate affinity, and an extended and open headpiece structure with high affinity (Hogg et al., 2011; Lefort et al., 2012). Recent studies report that the high-affinity LFA-1 conformation is required for T-cell arrest and adhesion to ICAM-1 expressed by endothelium or binding soluble ICAM-1 (Constantin et al., 2000; Salas et al., 2004; Shamri et al., 2005; Smith et al., 2005). Mn²⁺ or Mg²⁺/EGTA treatments bypass inside-out signaling and trigger high-affinity β1 and β2 conformations and rapid ligand binding (Shimaoka et al., 2002). Indeed, CD47⁺ T-cells exhibit a robust response to these stimuli and avidly bind integrin activation reporter mAbs. In contrast to the defects in arrest, the assay revealed no differences in detachment of adherent T-cells that lacked CD47. This suggests that the absence of CD47 affects avidity regulation of LFA-1 but not avidity regulation.

CD47 regulates LFA-1 and VLA-4 adhesive functions under shear flow conditions

Effector T-cell arrest, migration, and transendothelial migration require VLA-4 and LFA-1 integrin activation and binding to their endothelial ligands VCAM-1 and ICAM-1. Our present data show that murine CD47⁻/− Th1 effector cells, as compared with WT, have reduced adhesive interactions with inflamed cremaster microvessels and reduced adhesion and transmigration across TNF-α-activated endothelium. These defects can be explained by our data showing that CD47⁻/− Th1 cells have reduced adhesion to immobilized VCAM-1 and ICAM-1 but not E-selectin. This result implies that CD47⁻/− Th1 cells have impaired β1 and β2 integrin adhesive functions. Accordingly, the use of human Jurkat T-cells that lack CD47 showed striking defects in integrin-dependent adhesion identical to murine CD47⁻/− Th1 cells. Thus we conclude that CD47 expression plays a critical role in LFA-1 and VLA-4 binding to ICAM-1 and VCAM-1 under flow conditions in vivo and in vitro. The role of CD47 in regulation of VLA-4 adhesion under shear flow conditions was examined previously (Ticchioni et al., 2001), using the same Jurkat T-cell lines used here. These authors reported that CD47⁻ Jurkat T-cells had reduced adhesion to TNF-activated endothelium and to immobilized VCAM-1. Our present data extend this finding to include a striking defect in LFA-1 adhesive function.

We also performed detachment assays to quantify LFA-1 adhesion strengthening to ICAM-1 in CD47⁻ cells. Adhesion strengthening occurs after integrin activation and initial ligand binding and involves avidity regulation, defined as clustering or increased mobility of integrins in the plasma membrane of adherent cells (Kim et al., 2004). In contrast to the defects in arrest, the assay revealed no differences in detachment of adherent T-cells that lacked CD47. This suggests that the absence of CD47 affects avidity regulation of LFA-1 but not avidity regulation.
from sickle cell disease patients, CD47 coimmunoprecipitated with β1 integrins and regulated VLA-4-dependent adhesion to VCAM-1 and TSP-1 (Brittain et al., 2004). Other reports used various CD47 chimeric molecules expressed in Jurkat CD47- cells to demonstrate that only the extracellular IgV and first transmembrane domains of CD47 were required to restore Jurkat T-cell arrest on VCAM-1 (Ticchioni et al., 2001). A similar strategy in a CD47 (null) T-cells have impaired adhesion to immobilized ICAM-1 and VCAM-1 but not E-selectin under shear flow conditions. (A–C) Jurkat T-cells were drawn across immobilized ICAM-1 (A), VCAM-1 (B), or E-selectin (C) proteins at various shear stress levels, and adhesion was measured as described in Materials and Methods. (D) Transfection of CD47 Jurkat cells with a plasmid containing CD47 tagged with GFP (CD47 + GFP) rescued cell adhesion to ICAM-1. In contrast, transfection of CD47- cells with a plasmid containing only GFP (CD47-GFPcontrol) did not. (E) Detachment of Jurkat T-cells prebound to immobilized ICAM-1 is not affected by the absence of CD47. Data are means ± SEM of at least three independent experiments. *p ≤ 0.05, **p ≤ 0.01 (Student’s t test).

In summary, our results indicate that CD47 in cis interactions regulate LFA-1 and VLA-4 integrin affinity, and in turn, this process plays a substantial role in the adhesion and diapedesis of T-cells in models of inflammation. From our present results we infer the existence of a distinct and perhaps novel pathway that regulates T-cell recruitment in vivo to sites of inflammation, and we identify a potential therapeutic target for the treatment of immune-mediated diseases.

MATERIALS AND METHODS

Materials
Recombinant human and mouse E-selectin, VCAM-1, and ICAM-1 Fc-chimeras were from R&D Systems (Minneapolis, MN). Recombinant mouse interleukin-2 (IL-2), IL-12, and TNF-α were purchased from BioLegend (San Diego, CA). The following hybridoma clones were purchased from the American Type Culture Collection, Manassas, VA) and used as purified IgG: mAbs to β2 integrins include KIM127 (Andrew et al., 1993) and TS1/18 (Miller et al., 1986); TS1/22 recognizes LFA-1 (Miller et al., 1986); B6H12 and 2D3 are functional blocking and nonblocking, respectively, and recognize human CD47 (Brown et al., 1990). 13A9 recognizes a functional epitope in PSLG-1 (Snapp et al., 1997); mAb 24 was provided by Nancy Hogg (Cancer Research Institute, London, United Kingdom; Dransfield and Hogg, 1989); HUTS 21, LIA1/2.1, and HP2/1 were provided by Francisco Sanchez-Madrid (Hospital de la Princesa, Madrid, Spain; Lim et al., 2000); rabbit polyclonal anti-CD18 was reported previously (Lindberg et al., 1996; de Vries et al., 2002; Su et al., 2008; Azcutia et al., 2012).

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Materials
Recombinant human and mouse E-selectin, VCAM-1, and ICAM-1 Fc-chimeras were from R&D Systems (Minneapolis, MN). Recombinant mouse interleukin-2 (IL-2), IL-12, and TNF-α were purchased from BioLegend (San Diego, CA). The following hybridoma clones were purchased from the American Type Culture Collection, Manassas, VA) and used as purified IgG: mAbs to β2 integrins include KIM127 (Andrew et al., 1993) and TS1/18 (Miller et al., 1986); TS1/22 recognizes LFA-1 (Miller et al., 1986); B6H12 and 2D3 are functional blocking and nonblocking, respectively, and recognize human CD47 (Brown et al., 1990). 13A9 recognizes a functional epitope in PSLG-1 (Snapp et al., 1997); mAb 24 was provided by Nancy Hogg (Cancer Research Institute, London, United Kingdom; Dransfield and Hogg, 1989); HUTS 21, LIA1/2.1, and HP2/1 were provided by Francisco Sanchez-Madrid (Hospital de la Princesa, Madrid, Spain; Lim et al., 2000); rabbit polyclonal anti-CD18 was reported previously (Lindberg et al., 1996; de Vries et al., 2002; Su et al., 2008; Azcutia et al., 2012).
interferon (IFN)-γ, ICAM-1, ICAM-2, VCAM-1, PECAM-1, E-selectin, PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA). Vibrant CFSE and Alexa Fluor 488, and goat anti-mouse and anti-rabbit Alexa Fluor 680 cell tracker, goat anti-PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA). Vibrant CFSE and Alexa 680 cell tracker, goat anti-PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA). Vibrant CFSE and Alexa 680 cell tracker, goat anti-PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA). Vibrant CFSE and Alexa 680 cell tracker, goat anti-PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA). Vibrant CFSE and Alexa 680 cell tracker, goat anti-PSGL-1, LFA-1, and VLA-4 were purchased from BD PharMingen (San Jose, CA).

Mice
CD47-deficient (CD47−/−) mice (C57BL/6 strain) were obtained from Eric Brown (Genentech, South San Francisco, CA; Lindberg et al., 1996). WT C57BL/6 mice (Charles River Laboratories, Wilmington, MA) were used to establish a breeding colony in our facility for use as WT control animals. All mice used were bred in the same pathogen-free facility at Harvard Medical School New Research Building in accordance with the guidelines of the Committee of Animal Research at the Harvard Medical School and the National Institutes of Health animal research guidelines.

Cells
HUVECs were isolated and passaged as previously described and used at passage 2 for in vitro flow chamber assays (Stefanidakis et al., 2008). MHECs were prepared as described (Alcaide et al., 2012) from 8- to 12-d-old animals. Murine CD4+ Th1 effector cells were derived from naive T-cells by CD3 and CD28 stimulation in the presence of IL-12 and IFN-γ polarizing conditions as previously described (Alcaide et al., 2012). The human Jurkat T-cell E6.1 clone expressing CD47 (TIB-152) was from the American Type Culture Collection, and Jurkat T-cell clone E6.1 lacking CD47 (JINB8) was described previously (Ticchioni et al., 2001). Jurkat T-cells were maintained in RPMI-1640 supplemented with 10% fetal bovine serum (FBS), antibiotics, and L-Glutamax.

Flow cytometry
Flow cytometry analysis of intracellular IFN-γ was performed to corroborate the differentiation of Th1 cells and monitor expression of murine CD47, LFA-1, VLA-4, and PSGL-1 and also on human Jurkat T-cells using standard immunofluorescence staining and flow cytometry methods.

Intravital microscopy
In vitro polarized WT and CD47−/− Th1 cells were labeled with different fluorescent dyes (CFSE or Alexa 680), and then 3 × 10^6 of each cell type were mixed together and coinjected retrograde using a femoral artery catheter into WT or CD47−/− recipient mice 2 h after TNF-α injection as described (Alcaide et al., 2012). Leukocyte–endothelial adhesive interactions were obtained with an Olympus FV 1000 intravital microscope (Center Valley, PA) equipped with a LumPlan 40x/0.8 numerical aperture (NA) water immersion objective and digitally recorded with an Olympus DP71 charge-coupled device video camera and Olympus Fluoview 1000 imaging software. Rolling Th1 cells were identified as the visible cells passing through and transiently interacting with vessel surface in a plane perpendicular to the vessel axis. Rolling velocities were calculated as previously detailed (Alcaide et al., 2012). Vcrit was calculated as described previously (Ley et al., 1995; Yang et al., 1999).

T-cell adhesion to immobilized Fc chimera adhesion molecules and MHECs under defined laminar shear flow conditions in vitro
T-cell adhesion and transendothelial migration and interactions with immobilized adhesion molecules were performed as described

FIGURE 6: Mn^{2+} or Mg^{2+}/EGTA fails to induce strong LFA-1 and β1 high-affinity conformation expression or binding of soluble ICAM-1-Fc chimera in CD47−(null) Jurkat T-cells. (A–D) Mn^{2+} or Mg^{2+}/EGTA–induced expression of LFA-1 and open “activated” conformations of β1 integrins by 0.5 mM Mn^{2+} or 10 mM Mg^{2+}/1 mM EGTA treatments was significantly reduced in CD47− Jurkat T-cells. (E) High-affinity β1 integrins induced by Mn^{2+} or Mg^{2+}/EGTA buffer were measured by mAb HUTS21. (F) CD47− T-cells exhibited little binding of soluble ICAM-1 as compared with CD47+ T-cells induced by Mn^{2+} and Mg^{2+}/EGTA treatments. Results are normalized to isotype-matched, control nonbinding mAb. Expression of these active conformations of integrins in 0.5 mM Mn^{2+} or 10 mM Mg^{2+}/1 mM EGTA treatments was significantly reduced in CD47− Jurkat T-cells. Lysates of Jurkat cells were subjected to immunoprecipitation with mAb TS1/18 (lanes 1 and 4) served as a positive control to ensure the presence of β2 integrin in the lysate.

| Cells |
| --- |
| Murine CD47 | LFA-1, VLA-4, and PSGL-1 |
| Human Jurkat | E6.1 clone |

| Flow cytometry |
| --- |
| Intracellular IFN-γ |

| Intravital microscopy |
| --- |
| Rolling Th1 cells |

| T-cell adhesion |
| --- |
| to immobilized Fc chimera adhesion molecules |

| Volume 24 November 1, 2013 CD47 regulates β2 integrin affinity | 3365 |
TABLE 2: FLIM analysis of total β2 integrin–CD47 interactions.

| Condition | τ₁ (ps) | a₁ (%) | τₘ (ps) |
|-----------|---------|--------|---------|
| β2 integrin (donor only) | 2836 ± 11 | 100 ± 0 | 2836 ± 11 |
| β2 integrin + CD47 (B6H12) | 842.7 ± 137ᵃ | 32 ± 4 | 2218 ± 136ᵃ |
| β2 integrin (donor only) + Mg²⁺/EGTA | 2889 ± 21.3 | 100 ± 0 | 2889 ± 21 |
| β2 integrin + CD47 (B6H12) + Mg²⁺/EGTA | 804 ± 101ᵇᵃ | 23 ± 1ᶜ | 2442 ± 50ᵇᵃ |

Values represent mean ± SEM for n = 20 for each condition.
ᵃ p < 0.001 β2 integrin–CD47 vs. β2 integrin donor only.
ᵇ p < 0.001 β2 integrin–CD47 + Mg²⁺/EGTA vs. β2 integrin donor only + Mg²⁺/EGTA.
ᶜ p < 0.05 β2 integrin–CD47 + Mg²⁺/EGTA vs. β2 integrin + CD47 unstimulated.
ᶠn.s., β2 integrin–CD47 + Mg²⁺/EGTA vs. β2 integrin + CD47 unstimulated.

CD47− and CD47+ Jurkat cells were solubilized in cold lysis buffer (10 mM HEPES, pH 7.4, 150 mM NaCl, 1 mM phenylmethylsulfonyl fluoride, and 1× protease inhibitor cocktail [P8340; Sigma-Aldrich, St. Louis, MO]) and centrifuged at 100,000 × g for 30 min at 4°C. Lysates were divided and immunoprecipitated overnight at 4°C with protein A/G–Sepharose and mAb TS1/18, mAb 24, or mAb 24 and 1 mM Mn²⁺, and proteins were separated by SDS–PAGE. The β2 integrin was detected by Western blot with goat anti-human β2 integrin polyclonal antibody (R&D Systems, Minneapolis, MN).

Immunoprecipitation and blotting

Immunoprecipitation and blotting

Immunoprecipitation and blotting

Immunoprecipitation and blotting

Epifluorescence and time-correlated single-photon counting FLIM analysis

Aliquots of Jurkat CD47+ cells (3 × 10⁵) were fixed in 4% paraformaldehyde and transferred to glass slides by Cytospin centrifugation (Shandon, Astmoor, England) and then

CD47 upon integrin activation with Mg²⁺/EGTA. The color scale for τₘ ranges from 10 to 3500 ps. The β2 integrin was identified with the donor fluorophore (Alexa Fluor 488) and CD47 with the acceptor fluorophore (Alexa Fluor 594). (B) Localization of β2 integrin and CD47 by epifluorescence. Fixed cells were stained with (a, c) anti-β2 integrin polyclonal antibody alone (Quinn et al., 2001), (b, d) anti-β2 integrin antibody and anti-CD47 (B6H12) antibody labeled with Alexa 594 (unstimulated or with Mg²⁺/EGTA stimulation, respectively), (e) anti–activated-β2 integrin (mAb 24) alone upon Mg²⁺/EGTA stimulation, and (f) activated-β2 integrin (mAb 24) and anti-CD47 antibody also upon Mg²⁺/EGTA stimulation. Nucleus stained with DAPI.

FIGURE 7: CD47 and β2 integrin interact on the cellular membrane of Jurkat T-cells. (A) Representation of interacting fraction τₘ by pseudocolor images of the FLIM-FRET analysis of the interaction between β2 integrins with CD47 in unstimulated conditions and upon Mg²⁺/EGTA activation, and the interaction between activated β2 integrin detected by mAb 24 and
blocked with 5% FBS in PBS at room temperature. A murine mAb to CD47 (B6H12) directly conjugated with Alexa 594 and a polyclonal Ab to β2 integrins were diluted in PBS–5% FBS and incubated overnight at 4°C. As controls mAbs to PSGL-1 or αL integrin were used. Unlabeled primary Abs were detected using Alexa Fluor 488– or Alexa Fluor 594–conjugated secondary Abs when they corresponded. Protein interactions were defined by time-correlated single-photon counting FLIM as previously described (Mandal et al., 2008; Bair et al., 2012). The fluorescence lifetime baseline lifetime of Alexa Fluor 488 (donor fluorophore, αL or β2 integrin) was calculated by single-exponential-decay fitting of fluorescence emission in the absence of Alexa Fluor 594 (acceptor fluorophore). For samples stained for both donor and acceptor, lifetimes were fitted to a biesponential decay with lifetime of one component fixed to the donor-only lifetime. The lifetime for the interacting component, τi, and the fractional contributions for the percentage of interacting fluorophores, ai, were determined. Four or more separate experiments were performed, with n reported as total number of cells analyzed, and within each cell at least 10 different areas were used to determine the mean value. A Plan APO VC 60x oil DC N2 objective 1.4 NA, mounted on a Nikon Ti-E inverted microscope equipped with filter cubes used for DAPI, fluorescein isothiocyanate, and tetramethylrhodamine isothiocyanate fluorophores (Nikon), was used for epifluorescence and FLIM as described (Mandal et al., 2008; Bair et al., 2012). Nikon Elements 3.10 imaging software was used to collect epifluorescence data. For FLIM acquisition, Becker and Hickl (Berlin, Germany) a BDLS-480-SCM Picoscience Diode Laser and both long-pass (HQ500LP) and bandpass (HQ435/50) emission filters were used in combination with a hybrid detector (HPM-100-40 GaAsP hybrid detector) integrated with a Hamamatsu (Hamamatsu, Japan) R10467-40 hybrid photomultiplier tube. Becker and Hickl SPCM software with detector controller card was used to acquire FLIM data, and SPCImage 3.0 (Becker and Hickl, Berlin, Germany) software was used for FLIM analysis.

Statistical analysis

Data are expressed as the mean ± SEM unless otherwise stated. Statistical analyses by Student’s t test or by analysis of variance followed by the Newman–Keuls test were performed with Prism software (GraphPad, La Jolla, CA) and considered statistically significant at p < 0.05.

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References

Alcaide P, Maganto-García E, Newton G, Travers R, Croce KJ, Bu Dx, Luscinskas FW, Lichtman AH (2012). Difference in Th1 and Th17 lymphocyte adhesion to endothelium. J Immunol 188, 1421–1430.
Andrew D, Shock A, Ball E, Ortepp S, Bell J, Robinson M (1993). KIM185, a monoclonal antibody to CD18 which induces a change in the conformation of CD18 and promotes both LFA-1 and CR3-dependent adhesion. Eur J Immunol 23, 2217–2222.
Azcuita V, Stefanakis M, Tsuibo N, Mayadas T, Croce KJ, Fukuda D, Akaiwa M, Newton G, Luscinskas FW (2012). Endothelial CD47 promotes vascular endothelial-cadherin tyrosine phosphorylation and participates in T cell recruitment at sites of inflammation in vivo. J Immunol 188, 2553–2563.
Bair AM, Turman MV, Vaine CA, Panettieri RAJR, Soberman RJ (2012). The nuclear membrane leukotriene synthetic complex is a signal integrator and transducer. Mol Biol Cell 23, 4456–4464.
Barclay AN (2009). Signal regulatory protein alpha (SIRP[alpha]/CD47) interaction and function. Curr Opin Immunol 21, 47–52.
Brittain JE, Han J, Atagi KI, Orringer EP, Parise LV (2004). Mechanism of CD47-induced alpha4beta1 integrin activation and adhesion in sickle reticulocytes. J Biol Chem 279, 42393–42402.
Brown EJ, Frazier WA (2001). Integrin-associated protein (CD47) and its ligands. Trends Cell Biol 11, 130–135.
Brown E, Hooper L, Ho T, Gresham H (1990). Integrin-associated protein: a 50-kD plasma membrane antigen physically and functionally associated with integrins. J Cell Biol 111, 2785–2794.
Chen C, Mobley JL, Dwir O, Shimron F, Grabovsky V, Lobb RR, Shimizu Y, Alon R (1999). High affinity very late antigen-4 subsets expressed on T cells are mandatory for spontaneous adhesion strengthening but not for rolling on VCAM-1 in shear flow. J Immunol 162, 1084–1095.
Constantin G, Majeed M, Giagulli C, Piccio L, Kim JY, Butcher EC, Laudanna C (2000). Chemokines trigger immediate beta2 integrin affinity and mobility changes: differential regulation and roles in lymphocyte arrest under flow. Immunity 13, 759–769.
Cooper D, Lindberg FP, Gamble JR, Brown EJ, Vadas MA (1995). Transendothelial migration of neutrophils involves integrin-associated protein (CD47). Proc Natl Acad Sci USA 92, 3978–3982.
de Vries HE, Hendriks JJA, Honing H, de Lavalleire CR, van der Pol SMA, Hooijberg E, Dijkstra CD, van den Berg TK (2002). Signal-regulatory protein alpha CD47 interactions are required for the transmigration of monocytes across cerebral endothelium. J Immunol 166, 5832–5839.
Dransfield I, Hogg N (1989). Regulated expression of Mg2+ binding epotope on leucocyte integrin alpha subunits. EMBO J 8, 3759–3765.
Fortin G, Raymond M, Van VQ, Rubio M, Gautier P, Sarfati M, Franchimont D (1995). Transendothelial rolling in vivo. J Exp Med 181, 669–675.
Hogg N, Patzak I, Willenbrock F (2011). The insider’s guide to leukocyte integrin signalling and function. Nat Rev Immunol 11, 416–426.
Isenberg JS, Roberts DD, Frazier WA (2008). CD47: a new target in cardiovascular therapy. Arterioscler Thromb Vasc Biol 28, 615–621.
Kim M, Carman CV, Yang W, Salas A, Springer TA (2004). The primacy of affinity over clustering in regulation of adhesiveness of the integrin alphaMbeta2. J Cell Biol 167, 1241–1253.
Lefort CT, Rossaint J, Moser M, Petrich BG, Zarbock A, Monley SJ, Critchley DR, Ginsberg MH, Fassler R, Levy K (2012). Distinct roles for talin-1 and kindlin-3 in LFA-1 extension and affinity regulation. Blood 119, 427–428.
Ley K, Bullard DC, Arbones ML, Bosse R, Vestweber D, Tedder TF, Beaudet AL (1995). Sequential contribution of L- and P-selectin to leucocyte rolling in vivo. J Exp Med 181, 669–675.
Lim YC, Wakelin MW, Henault L, Goetz DJ, Yednock T, Cabana C, Sanchez-Madrid F, Lichtman AH, Luscinfskas FW (2000). Alpha4beta1-integrin activation is necessary for high-efficiency T-cell subset interactions with VCAM-1 under flow. Microcirculation 7, 201–214.

Lindberg FP, Bullard DC, Caver TE, Gresham HD, Beaudet AL, Brown EJ (1996). Decreased resistance to bacterial infection and granulocyte defects in IAP-deficient mice. Science 274, 795–798.

Luque A, Gomez M, Puzon W, Takada Y, Sanchez-Madrid F, Cabana C (1996). Activated conformations of very late activation integrins detected by a group of antibodies (HUTS) specific for a novel regulatory region (355–425) of the common beta1 chain. J Biol Chem 271, 11067–11075.

Mandal AK et al. (2008). The nuclear membrane organization of leukotriene synthesis. Proc Natl Acad Sci USA 105, 20434–20439.

McDonald JF, Zheleznyak A, Frazier WA (2004). Cholesterol-independent interactions with CD47 enhance alpha v beta3 avidity. J Biol Chem 279, 17301–17311.

Miller L, Schwarting R, Springer TA (1986). Regulated expression of the Mac-1, LFA-1, p150,95 glycoprotein family during leukocyte differentiation. J Immunol 137, 2891–2900.

Parkos CA, Colgan SP, Liang TW, Nusrat A, Bacarra AE, Carnes DK, Madara JL (1996). CD47 Mediates post-adhesive events required for neutrophil migration across polarized intestinal epithelium. J Cell Biol 132, 437–450.

Quinn MT, Swain SD, Parkos CA, Jutlana KL, Siemsen DW, Kurk SL, Jesaitis AJ, Jutlina MA (2001). A carbohydrate neoeptope that is up-regulated on human mononuclear leukocytes by neuraminidase treatment or by cellular activation. Immunology 104, 185–197.

Salas A, Shimaoka M, Kogan AN, Hanwood C, von Andrian UH, Springer TA (2004). Rolling adhesion through an extended conformation of integrin alphaLbeta2 and relation to a I and a I-like domain interaction. Immunity 20, 393–406.

Shamir R, Grabovsky V, Gauguet JM, Feigelson S, Manevich E, Kolanus W, Robinson MK, Staunton DE, von Andrian UH, Alon R (2003). Lymphocyte arrest requires instantaneous induction of an extended LFA-1 conformation mediated by endothelium-bound chemokines. Nat Immunol 6, 497–506.

Shimaoka M, Takagi J, Springer TA (2002). Conformational regulation of integrin structure and function. Annu Rev Biophys Biomol Struct 31, 485–516.

Sircar M et al. (2007). Neutrophil transmigration under shear flow conditions in vitro is junctional adhesion molecule-C independent. J Immunol 178, 5879–5887.

Smith A, Carrasco YR, Stanley P, Kieffer N, Batista FD, Brown EJ (2005). A talin-dependent LFA-1 focal zone is formed by rapidly migrating T lymphocytes. J Cell Biol 170, 141–151.

Snapp KR, Wagers AJ, Craig R, Stoolman LM, Kansas GS (1997). P-selectin glycoprotein ligand-1 is essential for adhesion to P-selectin but not E-selectin in stably transfected hematopoietic cell lines. Blood 89, 896–901.

Stefanidakis M, Newton G, Lee WY, Parkos CA, Luscinfskas FW (2008). Endothelial CD47 interaction with SIRPg is required for human T-cell transendothelial migration under shear flow conditions in vitro. Blood 112, 1280–1289.

Su X, Johansen M, Looney MR, Brown EJ, Matthay MA (2008). CD47 deficiency protects mice from lipopolysaccharide-induced acute lung injury and Escherichia coli pneumonia. J Immunol 180, 6947–6953.

Ticchioni M, Raimondi V, Lamy L, Wijdenes J, Lindberg FP, Brown EJ, Bernard A (2001). Integrin-associated protein (CD47/IAP) contributes to T cell arrest on inflammatory vascular endothelium under flow. FASEB J 15, 341–350.

Yang J, Hirata T, Croce K, Merrif-Skloof G, Tchernychev B, Williams E, Flamenhaf R, Furie BC, Furie B (1999). Targeted gene disruption demonstrates that P-selectin glycoprotein ligand 1 (PSGL-1) is required for P-selectin-mediated but not E-selectin-mediated neutrophil rolling and migration. J Exp Med 190, 1769–1782.
Azcutia et al.
**Supplemental Table S1.** FLIM analysis of αL integrin/β2 integrin and β2 integrin/PSGL-1 interactions.

| Condition                  | $\tau_1$ (ps) | $a_1$ (%) | $\tau_m$(ps) |
|----------------------------|---------------|-----------|--------------|
| αL integrin (donor only)   | 2762±38       | 100±0     | 2613±49      |
| αL integrin – β2 integrin  | 334.6±38a     | 46±6      | 1617±113a    |
| β2 integrin (donor only)   | 2915±12       | 100±0     | 2888±10      |
| β2 integrin + PSGL-1       | 2233±99b      | 13±2      | 2894±11      |

Values represent means ± SEM for n = 10 for each condition.  

$^a$ P<0.001 vs αL integrin donor only.  

$^b$ P<0.001 vs β2 integrin donor only.
**Supplemental Figure S1.** Human Jurkat CD47- (null) T cell adhesion to immobilized ICAM-1 and VCAM-1 is reduced upon Mn\(^{2+}\) activation in an in vitro flow chamber model. Jurkat T cells were activated with Mn\(^{2+}\) (0.5 mM) for 15 min at 37\(^\circ\)C and drawn across immobilized ICAM-1-Fc (A) or VCAM-1-Fc (B) at various shear stress levels as described in Methods. Treatment of CD47- cells with Mn\(^{2+}\) did not restore binding comparable to CD47+ T cell adhesion. Data are Means ± SEM, n = 3. *p ≤ 0.05, **p ≤ 0.01 (Student t test).
**Supplemental Figure S2.** β2 integrin interacts with αL integrin but not with PSGL-1 on the cellular membrane of Jurkat T cells. (A) Representation of interacting fraction $\tau_m$ by pseudocolor images of the FLIM-FRET analysis of the interaction between αL and β2 integrins and β2 integrin and PSGL-1. The color scale for the $\tau_m$ lifetimes ranges from 10 to 3500 picoseconds (ps). As positive control αL integrin was identified with the donor fluorophore (Alexa Fluor 488) and β2 integrin with the acceptor fluorophore (Alexa Fluor 594). As a non-interacting control, β2 integrin, was used as a donor and PSGL-1 as an acceptor. (B) Localization of β2 integrin, αL integrin and PSGL-1. Fixed cells were costained with (a) anti-αL integrin antibody alone (clone TS1/22) (b) anti-αL integrin antibody and anti-β2 integrin polyclonal antibody (Quinn et al., 2001) followed by an anti-mouse Alexa Fluor 488-conjugated and anti-rabbit Alexa Fluor 594-conjugated secondary antibodies, respectively. (c) anti-β2 integrin antibody alone and (d) anti-β2 integrin antibody and anti-PSGL-1(clone 13A9) followed by an anti-rabbit Alexa Fluor 488-conjugated and anti-mouseAlexa Fluor 594-conjugated secondary antibodies, respectively. The nucleus was stained with DAPI.
Supplemental Figure S3. (A-D) Mean lifetime ($\tau_m$) histograms of $\beta2$ integrin as the donor molecule: (A) with no acceptor (Donor only) in unstimulated conditions, (B) with CD47 (clone B6H12) as the acceptor molecule in unstimulated conditions, (C) donor only upon Mg$^{2+}$/EGTA stimulation, (D) with CD47 as the acceptor molecule upon Mg$^{2+}$/EGTA stimulation. (E,F) Activated $\beta2$ integrin as the donor molecule (E) without or (F) with CD47 as the acceptor molecule, respectively, upon Mg$^{2+}$/EGTA stimulation. (G,H) Mean lifetime $\tau_m$ histograms of $\beta2$ integrin as the donor molecule: (G) with no acceptor (Donor only), and (H) with PSGL-1 as the acceptor molecule. (I,J) $\alpha$L integrin as the donor molecule: (I) with no acceptor (Donor only), and (J) with $\beta2$ integrin as the acceptor molecule. Histograms are representative of 20 different cells.