Research Article

Antifungal Activity of Crude Extract from the Rhizome and Root of Smilacina japonica A. Gray

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This study aimed to investigate the antifungal activity of hydroalcoholic extract from Smilacina japonica A. Gray (SJA) against different fungi. The minimum inhibitory concentration (MIC) for SJA was determined by the broth microdilution method. The antifungal effects of SJA against Candida albicans were further confirmed by cell growth test and time-kill curve test. The effects of SJA on the fungal morphology and ultrastructure were also evaluated. SJA has a broad-spectrum antifungal activity. The MICs of SJA against different fungi, including fluconazole-sensitive and -resistant Candida albicans, other Candida species, and Cryptococcus neoformans, ranged from 208 μg/ml to 1665 μg/ml. Furthermore, SJA displayed fungicidal activity against varied fungi and obviously inhibited the hyphal growth of fungi. The mechanism study revealed that the antifungal activity of SJA might be associated with its effect on the cell morphology and ultrastructure.

1. Introduction

With increased patients with HIV infection, receiving different immunosuppressant treatments, antineoplastic chemotherapy, transplant recipients, or using catheters or other intravenous devices, invasive fungal infections are severe infections and constantly rising in the world [1–3]. Invasive candidiasis remains the most frequent mycosis and is often cited as the fourth most common cause of bloodstream infections with significant morbidity and mortality [4]. To date, common agents used in clinic to treat invasive fungal infections are azoles (e.g., fluconazole, itraconazole, voriconazole), echinocandins (e.g., caspofungin, anidulafungin, micafungin), and amphotericin B. However, progressive increase in exposing to azoles and echinocandins among Candida isolates commonly leads to resistant strains [5, 6]. Some fungi are intrinsically resistant to certain antifungals, such as Candida krusei (to fluconazole) and Cryptococcus spp (to the echinocandins). Drug resistance in fungi worsened the already significant mortality associated with the invasive fungal infections [7]. To explore novel antifungal agents with new molecular mechanism is a valuable solution to this problem.

Traditional medicinal plants have been an important source to discover new agents or lead compounds in the history of drug research and development. A variety of herbal medicines, as Rosa chinensis Jacq., Indigofera suffruticosa Mill., Wrightia tinctoria R. Br., Mahonia aquifolium Nutt., Glycyrrhiza glabra L. and so on, were reported to exert antifungal activity [8–13]. In our screening study, we found that Maianthemum japonicum (A. Gray) LaFrankie, also called Smilacina japonica A. Gray, displayed potent antifungal and even fungicidal effect. Smilacina japonica A. Gray (SJA), locally called “PianTouQi”, is a perennial herb which distributes widely in China. The tender aerial part is edible as a wild vegetable by local people. The rhizome and root have the ability of promoting blood circulation and alleviating pain. It has long been used as folk herbes for the treatment of internal lesion caused by overexertion, asynodia, headache, rheumatism, menstrual disturbance, mastitis, bruises, carbuncle, and furuncle [14].
Previous studies have reported that SJA had several pharmaceutical effects including antibacterial, antitumor, and antioxidant activities [14–16]. However, there are few studies on its mode of antifungal action. In this context, the antifungal activity of SJA and its possible mechanisms of action were reported.

2. Materials and Methods

2.1. Strains and Media. One clinical isolate of fluconazole (FCZ) resistant C. albicans (103), one international calibration strain (SC5314), one American type culture collection (ATCC)-typed Candida strain (C. parapsilosis ATCC22019), Cryptococcus neoformans 32609, C. krusei ATCC2340, C. glabrata ATCC1182, and C. tropicalis 2718 were utilized. SC5314 was kindly provided by William A. Fonzi (Department of Microbiology and Immunology, Georgetown University, Washington, DC). C. krusei ATCC2340 and C. glabrata ATCC1182 were kindly provided by professor Changzhong Wang (School of integrated traditional and western medicine, Anhui university of traditional Chinese medicine, Hefei, China). The other fungi strains were provided by the Changhai Hospital (Shanghai, China). Strains were cultured at 35°C under constant shaking (200 rpm) in complete liquid medium (yeast extract peptone dextrose-YPD) consisting of 1% (w/v) yeast extract, 2% (w/v) peptone, and 2% (w/v) dextrose.

2.2. Medicinal Material Extraction. The dried rhizome and root of SJA were bought from the fourth chain of Huaiya drug store belong to Meixian De-er-kang herbal medicine corporation in Shaanxi province with supply certificate. Approximate 5 g of dried and smashed SJA was extracted with an excess 100 ml of 75% ethanol, then the extract was filtered. After removing the solvent by rotary evaporation under reduced pressure at 50°C, the residue was dissolved with DMSO to approximately 0.333 g/ml SJA. The final extract was dark brown liquid and stored at -20°C for further use.

2.3. Antifungal Susceptibility Test. The antifungal susceptibility test was performed on all strains according to broth microdilution method as described previously [17]. C. parapsilosis ATCC22019 was used as a quality control strain and tested in each assay. FCZ was used as a positive drug control. The initial concentration of fungi suspended in RPMI 1640 medium was about 10^3 cells/ml, and the initial concentration ranged from 0.125 to 64 μg/ml for FCZ and 6.5 to 3330 μg/ml for SJA. The 96-well plates were incubated at 35°C for 24 h, 48 h or 72 h. The minimum inhibitory concentrations (MICs) were determined by optical density. Each strain was tested in triplicate.

2.4. Checkerboard Microdilution Assay. Assays were performed according to the above test 2.3. The combination concentrations ranged from 0.125 to 64 μg/ml for FCZ and 26 to 208 μg/ml for SJA. The fractional inhibitory concentration (FIC) index (FICI) was calculated by the following equation: FICI = FIC A + FIC B, where FIC A is the MIC of the combination/the MIC of drug A alone, and FIC B is the MIC of the combination/the MIC of drug B alone. Synergy was defined as an FICI value of ≤0.5, while antagonism was defined as an FICI value of >4, and the addition was defined as an FICI value of 0.5 ≤ FICI ≤1. An FICI result between 1 and 4 was considered indifferent.

2.5. Cell Growth Test. Each fungus was prepared at the starting inoculum of 10^6 cells/ml in glass tubes. Different concentrations of SJA (208 μg/ml, 416 μg/ml, 832 μg/ml and 3330 μg/ml) and FCZ were added into tubes. After incubation with agitation at 35°C for 24 h or 48 h, pictures were captured [18].

2.6. Time-Kill Curve Studies. Different fungi strains were prepared at the starting inoculum of 10^5 cells/ml. The concentrations were 104 μg/ml, 208 μg/ml, 416 μg/ml or 832 μg/ml for SJA, and DMSO comprised <1% of the total test volume. At predetermined time points (0 h, 4 h, 8 h, 12 h, 16 h, 24 h and 48 h) after incubation with agitation at 35°C, a 100 μl aliquot was removed from every solution and serially diluted 10-fold in sterile water. A 100 μl aliquot from each dilution was spread on the sabouraud dextrose agar plate. Colony counts were determined after incubation at 35°C for 48 h. Fungicidal activity was defined as a ≥ 3 log_{10} reduction from the starting inoculums [19].

2.7. Hyphal Growth Test. C. albicans SC5314 cells were incubated at 37°C in RPMI 1640 for 3 h in the absence or in the presence of different concentrations of SJA (416 μg/ml, 832 μg/ml) or FCZ (0.5 μg/ml). All cells were viewed by light microscopy to assess hyphal formation [20].

2.8. Cell Membrane Permeability. Membrane permeabilization of C. albicans was detected according to the study [21]. Briefly, C. albicans SC5314 (10^7 cells/ml) was incubated with 5 mM calcein acetoxy-methyl ester (Fanbo biochemicals, China) for 2 h. Then the cells were washed three times and C. albicans (10^6 cells/ml) was transferred into tubes. After treating with or without SJA (416 μg/ml, 832 μg/ml) for 3 h, the cells were washed three times and about 20,000 cells were acquired for flow cytometry analysis (Mafllo Astrios flow cytometer).

2.9. Transmission Electron Microscopy (TEM). After treating with or without SJA (416 μg/ml or 832 μg/ml), C. albicans SC5314 was washed in PBS and fixed in 3 ml fixative solution (sodium cacodylate buffer, pH 7.2, containing 4% polyoxymethylene) for 24 h at 4°C. Then, the samples were washed with saline and postfixed with 1% phosphotungstic acid for 90 minutes. The fixed cells were dehydrated through a graded series of ethanol and embedded with EPON-812. Ultrathin sections were prepared and observed after double staining with uranium and lead under a transmission electron microscope (HITACHI H-800, Japan) [22].

2.10. Statistical Analysis. The two-tailed Student’s t-test was used for analysis of two groups. Statistical significance was set at a p-value in the figures as: ∗, P < 0.05.
Table 1: Antifungal activity of SJA against different fungi (μg/ml) a.

| fungi                          | FCZ b | MIC (μg/ml) | SJA |
|-------------------------------|-------|-------------|-----|
|                               | 24 h  | 48 h        |     |
| FCZ-resistant C. albicans 103 | >64   | >64         | 416-832 |
| C. albicans SC5314           | 0.5-1 | 1           | 416-832 |
| Cryptococcus neoformans       | 1-2   | 2           | 208-416 |
| C. krusei                    | >64   | >64         | 1665 |
| C. parapsilosis               | 2     | 2           | 416  |
| C. glabrata                  | >64   | >64         | 208-416 |
| C. tropicalis                | >64   | >64         | 208-416 |

Table 2: Activity of SJA alone and in combination with FCZ against different fungi (μg/ml) a.

| fungi                          | Alone | Combination | FIC index for combination | Mode of interaction |
|-------------------------------|-------|-------------|----------------------------|--------------------|
|                               | FCZ b | SJA | FCZ b | SJA |                      |
| FCZ-resistant C. albicans 103 | >64   | 416 | 64   | 208 | 1.000                | addition       |
| C. albicans SC5314            | 1     | 416 | 1    | 26  | 1.125                | indifference   |
| Cryptococcus neoformans       | 2     | 208 | 0.5  | 26  | 0.375                | synergism      |
| C. krusei                    | >64   | 1665| 8    | 208 | 0.188                | synergism      |
| C. parapsilosis               | 2     | 416 | 1    | 26  | 0.563                | addition       |
| C. glabrata                  | >64   | 416 | >64  | 208 | 1.500                | indifference   |
| C. tropicalis                | >64   | 416 | 64   | 208 | 1.000                | addition       |

a SJA, Smilacina japonica A. Gray; b FCZ, fluconazole.

3. Results

3.1. Fungi Growth Was Inhibited by SJA. The results of the antifungal susceptibility test were summarized in Table 1. SJA displayed a broad-spectrum antifungal activity. The MICs of SJA against different fungi, including FCZ-sensitive and -resistant C. albicans, other Candida species and Cryptococcus neoformans, ranged from 208 μg/ml to 1665 μg/ml at 24 h or 48 h. To further visualize the activity of SJA against these fungi, we evaluated the growth of fungi after incubation with different concentrations of SJA for 24 h and 48 h. SJA at 416 μg/ml, 832 μg/ml and 3330 μg/ml displayed potent growth inhibition activity against all the tested fungi except C. krusei (Figure 1). In addition, SJA at 208 μg/ml also exhibited significant growth inhibition against part fungi (Cryptococcus neoformans, C. glabrata, C. parapsilosis and C. tropicalis).

To evaluate the interaction of SJA with antifungal drug FCZ, checkerboard microdilution assays were tested. As shown in Table 2, synergisms were observed for Cryptococcus neoformans and C. krusei, while additions were found for FCZ-resistant C. albicans 103, C. parapsilosis and C. tropicalis. Indifferences were observed for C. albicans SC5314 and C. glabrata.

To evaluate whether SJA had fungicidal activity against fungi, time-kill curve studies were performed. As shown in Figure 2, SJA displayed potent fungicidal effect against different fungi except C. krusei, which was consistent with the results of cell growth test. SJA at 208 μg/ml displayed fungicidal effect against FCZ-resistant C. albicans 103 and led to a decrease of 1.68-log_{10} Colony-forming units (CFU) /ml at 4 h compared with the initial inoculum concentration (Figure 2(a)). Moreover, SJA at 208 μg/ml, 416 μg/ml and 832 μg/ml resulted in a complete cell-killing at 8 h. For C. albicans SC5314 (Figure 2(b)). SJA also displayed fungicidal effect. SJA at 208 μg/ml yielded a 6.1-log_{10} CFU/ml reduction compared with the control at 24 h. More specifically, SJA at 208 μg/ml, 416 μg/ml and 832 μg/ml exhibited a complete cell-killing at 48 h. For other fungi (Cryptococcus neoformans, C. glabrata, C. parapsilosis and C. tropicalis), both 416 μg/ml and 832 μg/ml SJA resulted in a complete cell-killing at 24 h, and the densities of fungi cells were all reduced to zero. These results confirmed that SJA had excellent fungicidal activity against varied fungi.

3.2. Hyphal Growth of Fungi Was Inhibited by SJA. Fungi like C. albicans can grow in three types of morphological forms including yeast, pseudohyphae, and hyphae. During the early stage of infection, yeast fungi become hyphae form to penetrate human epithelial and endothelial cells and cause the damage to them [23, 24]. In addition, to resist standard antifungal treatments, fungi can form biofilms which contain hyphae [25]. Therefore, the activity of SJA on the growth of fungal hyphae was determined. In the blank control group (Figure 3), C. albicans showed elongated hyphae forms after 3 h incubation. After treating with SJA at 416 μg/ml or 832 μg/ml, most of the C. albicans did not germinate and appeared as yeast form. More interestingly, C. albicans progressively aggregated and became clumps after treating with 832 μg/ml of SJA for 2 h and 3 h.
3.3. The Cell Size and the Membrane Permeability of Fungi Were Increased by SJA. Flow cytometry analysis (side scatter [SSC]-forward light scatter [FSC]) revealed that *C. albicans* SC5314 treated with SJA (416 μg/ml or 832 μg/ml) exhibited a significant increase in cell size, as evidenced by the increase in forward light scattering (Figure 4).

Calcein AM is a non-fluorescent, hydrophobic compound that easily permeates intact, live cells. Once into the cytoplasm of cells, calcein AM is hydrolyzed by cytoplasmic esterases, producing membrane-impermeable, hydrophilic and strongly fluorescent calcein which can be well-retained in intact cell cytoplasm. After incubation with calcein AM, the cellular fluorescence of calcein was detected by flow cytometry to evaluate the effect of SJA on the cell membrane permeabilization. Result displayed that cellular fluorescence of calcein was markedly decreased by treatment of *Candida* cells with different concentrations of SJA (416 μg/ml or 832 μg/ml) compared with the blank control cells (Figure 5). The results indicated that SJA interrupted the *Candida* cell membrane and increased the cell membrane permeability.

3.4. The Ultrastructure of Fungi Was Damaged by SJA. In order to explore the antifungal mechanism of SJA, the ultrastructure of *C. albicans* was monitored by TEM. As shown in Figure 6, the control *Candida* cells have a uniform central density and an intact cell wall. After treatment of SJA at 416 μg/ml or 832 μg/ml, the outer layer of the cell wall became thinner, and the middle layer became thicker. Some abnormal shapes of the vacuoles were also observed. In addition, reduced cytoplasmic density and damaged membranes could be observed. The TEM results indicated that SJA caused obvious damage on the ultrastructure of *C. albicans* including cell wall and membrane structures.

4. Discussion

Fungal diseases cause considerable morbidity and mortality globally and elevate health care costs [26]. The paucity of effective antifungal agents and emergency of drug resistance prompted researchers to develop novel antifungal agents [7]. Natural products are a valuable source for drug discovery, whether in their nascent form as original templates for structure-optimizing for more effective and safe derivatives. About 80% of marketed antibiotics used clinically are derived from natural products [27, 28]. Current antifungal drugs, like echinocandins and amphotericin B are also developed from natural resources. A variety of natural products, as peptides, saponins, flavonoids, alkaloids, essential oils, and so on, have been found to possess antifungal activity [29–33]. As a traditional Chinese medicine, SJA has various therapeutic effects. However, to the best of our knowledge, there was no investigation on the action of SJA against fungi.

In this study, we demonstrated that SJA had antifungal activity against different fungi including *Candida species* and *Cryptococcus neoformans* (with MICs ranging from 208 μg/ml to 1665 μg/ml). In light of the definitions of fungicidal effect through time-kill curve tests, a fungicidal effect was defined as a reduction in ≥3log_{10} CFU/ml compared to initial inoculum [34]. Therefore, SJA was considered fungicidal. Previously, the antifungal effect of steroidal saponins extracted from the genus *Smilacina* called *Smilacina atropurpurea* (Franch.) Wang et Tang (Convallariaceae) was studied by Ying Zhang et al. [35]. Among eight saponins, they found that astropuroside B and F exhibited fungicidal activity against *C. albicans, C. glabrata, Cryptococcus neoformans* and *Aspergillus fumigates* with MFCs≤20 μg/ml except *C. krusei*, while dioscin was selectively active against *C. albicans* and *C. glabrata* (MFC≤5.0 μg/ml) [35]. Compared to *Smilacina atropurpurea*, the extract of SJA exhibited similar fungicidal effects.
Figure 2: Time kill curves of various fungi treated with different concentrations of SJA and FCZ. FCZ-resistant C. albicans 103 (a), C. albicans SC5314 (b), Cryptococcus neoformans (c), C. krusei (d), C. parapsilosis (e), C. glabrata (f) and C. tropicalis (g) were treated with different concentrations of SJA (104 \( \mu \)g/ml, 208 \( \mu \)g/ml, 416 \( \mu \)g/ml and 832 \( \mu \)g/ml) for 48 h. Aliquots were obtained at the indicated time points and serially dilutions were spread on agar plates. Colony counts were determined after 48 h incubation.

effect, but it had a broader antifungal spectrum. According to previous reports on the constituents of the two plants, there are differences in the chemical structures of saponins isolated from *Smilacina atropurpurea* and SJA [14, 36–38]. In addition, the report by Ying Zhang et al. also suggested that not all saponins had antifungal activity [35]. Therefore, our group will perform more studies to explore the exact antifungal ingredients of SJA in collaboration with the other laboratory.

Hyphae play an important role in the fungal infection and biofilm formation. SJA was able to suppress the hyphal
Figure 3: SJA inhibits the hyphal growth of fungi. The *C. albicans* SC5314 cells were incubated in RPMI 1640 liquid medium at 37°C for 3 h in the absence or presence of different concentrations of SJA (416 μg/ml or 832 μg/ml) or FCZ (0.5 μg/ml). The hyphal development was viewed by light microscopy and photographed after 0 h, 1 h, 2 h and 3 h of incubation.

Figure 4: Change of cell size (forward scatter-side scatter) in the presence of SJA. *C. albicans* SC5314 were treated with or without SJA (416 μg/ml to 832 μg/ml) for 3 h. Then the cells were analyzed by flow cytometry.
Figure 5: The effect of SJA on cell membrane permeability. Candida cells (10^7 cells/ml) were cultured in the presence of 5 μM calcein acetoxymethylester for 2 h. After washing three times, the cells were cultured in the presence or absence of SJA (416 μg/ml to 832 μg/ml) for 3 h. Cellular fluorescence intensities of calcein were analyzed by flow Cytometry. Data are means ± SD of triplicates of one representative experiment of three. *, P < 0.05, Student's t-test.

Figure 6: TEM results of Candida cells treated with or without SJA (416 μg/ml or 832 μg/ml).
growth of *C. albicans*. Besides, through the analysis of cellular fluorescent calcein by flow cytometry, we demonstrated that SJA could increase the membrane permeability. The yeast-to-hypha inhibition and the membrane-perturbation effect of SJA are akin to the other saponins like saponin extract from rhizomes of *Dioscorea panthaica* *Prain et Burk* and tea saponin [30, 39].

TEM results showed that SJA obviously damaged the ultrastructure of fungi including the membrane structures and cell wall. Lesions were observed on membrane structures, including the cytoplasmic membrane and organelle membranes such as nucleus and mitochondria. The membrane damage of SJA may contribute to the increased membrane permeability and cell size of fungi. SJA also exhibited a harmful effect on the cell wall, such as the outer layer of the cell wall became thinner, and the middle layer became thicker. Cell wall plays a crucial role in the fungal commensalism and infection [40]. It offers mechanical strength and acts as a barrier, thus protecting the fungus from the hostile environment. Besides, the cell wall glucan plays an important role in the yeast-to-hypha transition [41]. This suggests the hyphal growth inhibition of SJA may be related to its damage on the ultrastructure like cell wall. The harmful effects on the membrane and cell wall of SJA are similar to the *Solanum chrysotrichum* saponin SC-2 and saponin extract from rhizomes of *Dioscorea panthaica* *Prain et Burk* [30, 42].

5. Conclusion

The present study connoted that SJA has strong fungicidal activity against different fungi such as *Candida* species and *Cryptococcus neoformans*. SJA could also inhibit the yeast-hypha transition and increase membrane permeability and cell size of *C. albicans*. The antifungal effects might be through membrane and cell wall damage. As a traditional Chinese medicine, SJA represents a promising herbal medicine for the discovery of novel antifungal compounds. The exact antifungal ingredients and action mechanisms are under investigation in our laboratories.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Wei Liu and Shengzheng Wang are responsible for drafting the paper. Wei Liu participated in the study design, collection and analysis of data, review and drafting the paper. Baozheng Sun, Manman Yang, Ziyue Zhang, Xiuxiu Zhang and Yansong Pang participated in part data collection, data analysis, and revision of the paper. Shengzheng Wang guided all stages of the study and participated in both review and drafting of the paper.

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References

[1] G. Suleyman and G. J. Alangaden, ”Nosocomial fungal infections,” *Infectious Disease Clinics of North America*, vol. 30, no. 4, pp. 1023–1052, 2016.

[2] G. D. Brown, D. W. Denning, and S. M. Levitz, ”Tackling human fungal infections,” *Science*, vol. 336, no. 6082, article no. 647, 2012.

[3] D. Bitar, O. Lortholary, Y. Le Strat et al., ”Population-based analysis of invasive fungal infections, France, 2001–2010,” *Emerging Infectious Diseases*, vol. 20, no. 7, pp. 1163–1169, 2014.

[4] S. Strollo, M. S. Lionakis, J. Adjemian, C. A. Steiner, and D. R. Prevots, ”Epidemiology of hospitalizations associated with invasive candidiasis, United States, 2002–2012,” *Emerging Infectious Diseases*, vol. 23, no. 1, pp. 7–13, 2016.

[5] M. Baixench, N. Aoun, M. Desnos-Ollivier et al., ”Acquired resistance to echinocandins in Candida albicans: case report and review,” *Journal of Antimicrobial Chemotherapy*, vol. 59, no. 6, pp. 1076–1083, 2007.

[6] C. Biswas, S. Chen, C. Halliday et al., ”Identification of genetic markers of resistance to echinocandins, azoles and 5-fluorocytosine in Candida glabrata by next-generation sequencing: a feasibility study,” *Clinical Microbiology and Infection*, vol. 23, no. 9, pp. 676.e7–676.e10, 2017.

[7] D. W. Denning and M. J. Bromley, ”Infectious disease. how to bolster the antifungal pipeline,” *Science*, vol. 347, no. 6229, pp. 1414–1416, 2015.

[8] L. Zhang, H. Lin, W. Liu et al., ”Antifungal Activity of the ethanol extract from *flos rosae chinensis* with activity against fluconazole-resistant clinical *Candida*,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2017, Article ID 4780746, 10 pages, 2017.

[9] S. P. Leite, J. R. Vieira, P. L. de Medeiros et al., ”Antimicrobial Activity of *Indigofera suffruticosa*, *Evidence-Based Complementary and Alternative Medicine*, vol. 3, no. 2, Article ID 839056, pp. 261–265, 2006.

[10] K. Ponnumsamy, C. Petchiammal, R. Mohankumar, and W. Hopper, ”*In vitro* antifungal activity of indirubin isolated from a South Indian ethnomedicinal plant *Wrightia tinctoria* R. Br,” *Journal of Ethnopharmacology*, vol. 132, no. 1, pp. 349–354, 2010.

[11] A. Volleková, D. Kostalova, V. Kettmann, and J. Tóth, ”Antifungal activity of *Mahonia aquifolium* extract and its major protoberberine alkaloids,” *Phytotherapy Research*, vol. 17, no. 7, pp. 834–837, 2003.

[12] A. Fatima, V. K. Gupta, S. Luqman et al., ”Antifungal activity of *Glycyrrhiza glabra* extracts and its active constituent glabridin,” *Phytotherapy Research*, vol. 23, no. 8, pp. 1190–1193, 2009.
N. I. Mongalo, P. M. Dikhoba, S. O. Soyingbe, and T. J. Makhafola, “Antifungal, anti-oxidant activity and cytotoxicity of South African medicinal plants against mycotoxigenic fungi,” HELIYON, vol. 4, no. 11, Article ID e00973, 2018.

X. Liu, H. Zhang, X. Niu, W. Xin, and L. Qi, “Steroidal saponins from Smilacina japonica,” Fitoterapia, vol. 83, no. 4, pp. 812–816, 2012.

S. Zhao, Z. Han, Y. Li, and L. Yang, “Study on the scavenging hydroxyl radical of the extracts from Smilacina japonica,” Journal of South China Agricultural University, vol. 31, no. 2, pp. 59–62, 2010.

Q. Ren, “Experimental Studies of antibacterial effect of 80 species of herbs from the qin mountains,” Territory & Natural Resources Study, vol. 7853, no. 1, pp. 86–88, 2013.

H. Quan, Y.-Y. Cao, Z. Xu et al., “Potent in vitro synergism of flucunazole and berberine chloride against clinical isolates of Candida albicans resistant to flucunazole,” Antimicrobial Agents and Chemotherapy, vol. 50, no. 3, pp. 1096–1099, 2006.

W. Liu, L. P. Li, J. D. Zhang et al., “Synergistic antifungal effect of glabridin and flucunazole,” PLoS ONE, vol. 9, no. 7, Article ID e0103442, 10 pages, 2014.

E. E. Roling, M. E. Klepser, A. Wasson, R. E. Lewis, E. J. Ernst, and M. A. Pfaffer, “Antifungal activities of a fluconazole, caspofungin (MK0991), and anidulafungin (LY 303366) alone and in combination against Candida spp. and Cryptococcus neoformans via time-kill methods,” Diagnostic Microbiology and Infectious Disease, vol. 43, no. 1, pp. 13–17, 2002.

S. Wang, Y. Wang, W. Liu et al., “Novel carboline derivatives as potent antifungal lead compounds: design, synthesis, and biological evaluation,” ACS Medicinal Chemistry Letters, vol. 5, no. 5, pp. 506–511, 2014.

T. Tanida, T. Okamoto, E. Ueta, T. Yamamoto, and T. Osaki, “Antifungal activity and Chemotherapy resistant to fluconazole,” Antifungal Effects of Saponin Extract from Rhizomes of Dioscorea panthaica Prain et Burk against Candida albicans,” Evidence-Based Complementary and Alternative Medicine, vol. 2018, Article ID 6995307, 13 pages, 2018.

T. M. Souza-Moreira, J. A. Severi, E. R. Rodrigues et al., “Flavonoids from Plinia cauliflora (Mart.) Kausel (Myrtaceae) with antifungal activity,” Natural Product Research (Formerly Natural Product Letters), vol. 33, no. 17, pp. 2579–2582, 2018.

S. S. Seyedjavadi, S. Khani, H. Zare-Zardini et al., “Isoflavones of Glycosylphosphatidyl inositol anchored proteins,” Expert Review of Anti-infective Therapy, vol. 18, no. 2, pp. 148–164, 2011.