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Fluorescent Imaging of Antigen Released by a Skin-Invading Helminth Reveals Differential Uptake and Activation Profiles by Antigen Presenting Cells

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Abstract

Infection of the mammalian host by the parasitic helminth Schistosoma mansoni is accompanied by the release of excretory/secretory molecules (ES) from cercariae which aid penetration of the skin. These ES molecules are potent stimulants of innate immune cells leading to activation of acquired immunity. At present however, it is not known which cells take up parasite antigen, nor its intracellular fate. Here, we develop a technique to label live infectious cercariae which permits the imaging of released antigens into macrophages (MΦ) and dendritic cells (DCs) both in vitro and in vivo. The amine reactive tracer CFDA-SE was used to efficiently label the acetabular gland contents of cercariae which are released upon skin penetration. These ES products, termed ‘0-3hRP’, were phagocytosed by MHC-II+ cells in a Ca++ and actin-dependent manner. Imaging of a labelled cercaria as it penetrates the host skin over 2 hours reveals the progressive release of ES material. Recovery of cells from the skin shows that CFDA-SE labelled ES was initially (3 hrs) taken up by Gr1+MHC-II+ neutrophils, followed (24 hrs) by skin-derived F4/80+MHC-IIlo MΦ and CD11c+ MHC-IIhi DC. Subsequently (48 hrs), MΦ and DC positive for CFDA-SE were detected in the skin-draining lymph nodes reflecting the time taken for antigen-laden cells to reach sites of immune priming. Comparison of in vitro-derived MΦ and DC revealed that MΦ were slower to process 0-3hRP, released higher quantities of IL-10, and expressed a greater quantity of arginase-1 transcript. Combined, our observations on differential uptake of cercarial ES by MΦ and DC suggest the development of a dynamic but ultimately balanced response that can be potentially pushed towards immune priming (via DC) or immune regulation (via MΦ).

Introduction

Trematode parasites {e.g. Schistosoma sp, Fasciola sp, and Trichobilharzia sp} are important parasites of mammalian hosts in the developing, as well as the developed world, and cumulatively are a major health burden to humans and domestic animals. Infective schistosome larvae gain entry to the host as free-swimming cercariae which penetrate the host via a percutaneous route. The precise mechanism by which Schistosoma larvae penetrate the skin to facilitate their onward migration is a matter of debate [1–3]. Infection of mouse skin by S. mansoni cercariae occurs rapidly but many of the larvae are still in the skin by 40 hours [4,5]. Excretory/secretory (ES) molecules released by invading larvae aid penetration of the skin but also lead to the stimulation, and down-regulation, of the dermal inflammatory response [6]. Indeed, the extended contact between ES molecules released by invading larvae and innate immune cells in the skin, particularly following exposure to protective radiation-attenuated (RA) larvae [7], indicates that the innate response may be critical in limiting the success of initial infection. Therefore, the innate immune system in the skin could provide a target for manipulation in the pursuit of anti-schistosome vaccines and/or drugs but the cellular target(s) and mechanisms by which larval ES molecules act on the innate immune response are poorly understood.

The skin is populated with a range of innate immune cells [8], and pro- and anti-inflammatory innate responses occur quickly following cercarial penetration [9]. An initial neutrophil-rich cutaneous response resolves shortly after the majority of larvae have left the skin [4,10]. The cutaneous response also involves macrophages (MΦ) [7], dendritic cells (DC) [7] and Langerhans’ cells (LC) [11] which form cellular foci around the sites of parasite entry [12,13]. Activation of cells with antigen presenting function in the skin also directs their emigration to CD4+ rich areas of the skin draining lymph node (sdLN), where DCs and LCs have been observed to accumulate following exposure to RA schistosomes [11].

The ES products released in the first 3 hours after transformation of S. mansoni cercariae into schistosomula (termed 0-3hRP) stimulate cytokine production by MΦ in a MyD88-dependent fashion implying the involvement of one or more Toll-like receptors (TLR) [14]. Moreover, 0-3hRP stimulates DC that in turn drive strong Th2 responses both in vitro and in vivo [15], likely resulting from its capacity to limit the maturation and hence stimulatory capacity of the DC population [16]. Several studies have characterised the composition of ES material released by...
Author Summary

Schistosomiasis is caused by the parasitic worm Schistosoma with over 200 million people infected across 76 countries. The parasitic larvae (called cercariae) infect mammalian hosts via the skin, but the exact mechanisms by which dermal cells interact with molecules released by invading larvae are unclear. A better understanding of the infection process and stimulation of the early immune response would thus enable a targeted approach towards the development of drugs and vaccines. Here, we have used the fluorescent tracer CFDA-SE to label infectious cercariae and, together with confocal microscopy, have for the first time tracked in real time the parasite infecting via the epidermis and depositing excretory/secretory material in its wake. Phagocytic macrophages and dendritic cells in the skin internalised excretory/secretory molecules released by the larvae, and both cell types were subsequently located in the draining lymph nodes where priming of the acquired immune response occurs. In vitro studies determined that macrophages were slower to process released parasite material than dendritic cells; they also secreted lower levels of pro-inflammatory cytokines but greater quantities of regulatory IL-10. The relative abundance of macrophages versus dendritic cells in the skin infection site and their differential rates of antigen processing may be crucial in determining the success of adaptive immune priming in response to infection.

Insights into how schistosome infection may impact upon phagocytic cells of the innate immune response in the skin, and how this may affect the priming of the adaptive immune response in the skin-draining lymph nodes (sDLN).

Materials and Methods

Animals

Female C57BL/6 mice (8–12 weeks old) were bred and maintained at the University of York and housed under specific pathogen free (SPF) conditions in filter topped cages. All experiments were carried out within the guidelines of the United Kingdom Animal’s Scientific Procedures Act 1986. All the research that involved the use of animals was approved by the University of York Ethics committee.

Parasites

A Puerto Rican strain of S. mansoni was maintained by routine passage through outbred NMR-1 mice and Biomphalaria glabrata snails. Cercariae were shed from snails harbouring patent schistosome infections by exposure to light for up to 2 hours. Isolated cercariae were washed ×3 by pulse centrifugation at 200 g in 10 ml of sterile aged tap water (ATW) and re-suspended.

Fluorescent labelling of cercariae and 0-3hRP

Cercariae (~1–5×10⁴/ml) were incubated with various concentrations of the amine reactive tracer Vybrant CFDA-SE (Invitrogen Ltd, Paisley, UK) diluted with ATW at 28°C for 60 mins. Cercariae were concentrated by pulse centrifugation at 200 g followed by 3× washes in ATW prior to re-suspension in ATW and incubation for a further 60 mins to allow unconjugated dye to diffuse out of the parasite. Parasites were again washed 3× prior to measurement of fluorescence, or were fixed with 2% paraformaldehyde for 20 mins prior to imaging.

For collection of labelled 0-3hRP released from transforming cercariae, the protocol of Jenkins et al. [15] was modified. Suspensions of CFDA-SE labelled cercariae were mechanically-transformed [30] to separate heads from tails and then cultured in serum free RPMI 1640 (Invitrogen Ltd) containing 200 U ml⁻¹ penicillin and 100 μg ml⁻¹ streptomycin (Invitrogen Ltd) for 5 hrs. The supernatant containing 0-3hRP was concentrated in Vivaspin 15 tube (Sartorius Stedim Ltd, Epsom, UK) with a 5-kDa membrane. The protein concentration was determined using a Coomassie Plus-200 assay (Perbio Science Ltd, Cheshire, UK).

Measurement of labelling efficiency and imaging of CFDA-SE labelled cercariae

Aliquots of CFDA-SE labelled and unlabelled cercariae were placed in black 96 well clear bottom plates (Camlab Ltd, Cambridge, UK). Fluorescence was measured on a POLARStar OPTIMA microplate reader (BMG Labtech, Saitama City, Japan) (492±5 nm excitation; 520±5 nm emission). A manual count of cercariae per well was performed and data expressed as relative fluorescent units (RFU) per live cercaria.

Confocal or fluorescent microscopy was performed on both live and fixed parasites, or fixed cells, using a Zeiss confocal LSM 510 meta (Carl Zeiss Ltd, Welwyn Garden City, UK) or a Nikon Labophot fluorescent microscope equipped with a Nikon Coolpix 995 (Nikon Corp, Tokyo, Japan). All images were captured at ×10, ×20 or ×100 using identical laser settings at 488 nm excitation; 520 nm emission wavelengths with a pinhole setting of between 2–50 μm and parasites were manually kept in focus during skin penetration or movement. Photographic analysis was performed using Adobe photoshop or LSM image browser 4.2.
(Carl Zeiss Ltd, UK) and 3D images reconstruction was performed and analysed using Volocity 4.3.2 (Improvision, Coventry, UK) from LSM Z stacks.

Analysis of cercarial penetration efficiency

Anesthetised mice were infected via the pinnae [31] with 500 unlabelled or CFDA-SE labelled cercariae. After 30 mins, the remaining parasite suspension was collected and the number of non-penetrating cercariae established.

Production of inflammatory MΦ, BMMΦ, and BMDC

Peritoneal exudate cells (PEC) were extracted from mice by peritoneal lavage, 5 days post-injection with 0.5 ml sterile 3% Brewers thioglycollate medium (Sigma–Aldrich) [14]. PEC were separated into adherent and non-adherent populations by adherence to plastic after culture for 2 hours at 37°C, 5% CO₂.

BMMΦ were derived as follows. Femurs of naïve mice were removed, flushed with chilled PBS and the resulting cell suspension washed and re-suspended in Dulbecco’s Modified Eagle’s Medium (DMEM, Invitrogen Ltd) supplemented with 10% of heat-inactivated low endotoxin foetal bovine serum (Invitrogen Ltd) and 20% L929 cell-conditioned medium (Gift P. Kaye, University of York). Cells were plated at 1×10⁶ per well in 24 well plates (VWR, Luttworth, UK) and incubated at 37°C in 5% CO₂ for 7 days. BMDC were obtained as previously described [15] following 7 days culture in the presence of 20 μg/ml GM-CSF (Peprotech, London, UK).

Adherent PEC, non-adherent PEC, BMMΦ, and BMDC were then cultured with the following: 1000 unlabelled or CFDA-SE labelled cercariae, or 40 μg/ml unlabelled or CFDA-SE labelled 0-3hRP. Supernatants from cell cultures were removed and stored at −20°C prior to cytokine analysis. The remaining cells were removed using chilled PBS, washed and re-suspended in chilled culture media prior to labelling with antibodies and analysis by flow cytometry.

Flow cytometry and reagents

Flow cytometric analysis of in vivo cultured cells, or those recovered ex vivo, was performed on a DakoCytomation Cyan ADP analyser (Dako, Ely, UK). Cells were initially blocked for 30 mins with anti-CD16/32 mAb in PBS containing 1% FCS, and 2 mM EDTA. Subsequently, cells were labelled with directly conjugated antibodies; F4/80 Pacific Blue (#M5/114.15.2) (Insight Biotechnology Ltd, Wembley, UK). Biotin conjugated antibodies against CD11c (#N418) and GR-1 (#Ly-6C) were probed with streptavidin allophycocyanin or streptavidin Pacific Blue (Invitrogen Ltd). All antibody concentrations were optimised and all analyses performed alongside irrelevant isotype controls. Data was analysed using Summit v4.3 (Dako, UK).

Cytokine detection

Cytokine levels were measured by ELISA. IL-6 was captured with anti-IL-6 mAb (#MP5-20F3) and probed with biotinylated anti-IL-6 mAb (#MP5-32C110) detected with streptavidin peroxidase conjugate (BD Pharmingen, Oxford, UK). IL-12p40, IL-10 and TNF-α were measured using kits (Invitrogen Ltd) according to the manufacturer’s protocol. The lower sensitivity of the assays were 15 pg/ml (TNF-α), 20 pg/ml (IL-6), and 32 pg/ml (IL-12p40, IL-10).

Real-time PCR

Cell samples were re-suspended in Trizol (Invitrogen Ltd) and RNA extracted following the manufacturer’s protocol. Extracted RNA was reverse transcribed into cDNA using Superscript II Reverse Transcriptase (Invitrogen Ltd), checked for quality and genomic DNA contamination, and 10 ng (5 μl) of each resulting cDNA sample analysed by real time PCR on an ABI PRISM 7500 sequence detection system (Applied Biosystems, Warrington, UK). Relative quantities of RNA were determined using Taqman probes (Sigma–Aldrich, UK). The specific primer pairs and probes were; Arginase, 5’- TGACCTTGACCTTTGATGC, 3’-GTA-GAGGACCCGTGGCTTG, Probe: 5’- GTGTTGGAAGGCC-TACTTTACAAGAGAAGGCTCTC, INOS 5’- CTGCAATG-GACGGATATAAGG, 3’- CTAAACATGACAGAGATTT-CCTG, Probe: 5’- AGTCCTGCCATTGCTG. The relative expression of each gene was normalised to the values for the GAPDH housekeeping gene before statistical analysis. GAPDH 5’- CCATGTTTGTGATGGGTGTG, 3’- CCGTTCCAAATGCG-CAAAGTT Probe: CATCCCTGCACACCAACTGTGGCTTTAGG.

Infection of mice with CFDA-SE labelled cercariae and ex vivo cell recovery

Mice were infected with 1000 unlabelled or CFDA-SE labelled cercariae for 30 mins on each ear [31]. Pinnae from naive and infected (unlabelled or CFDA-SE labelled cercariae) mice were collected at 3, 24, 48 and 72 hours. Pinnae were split and then placed on 50 μg/ml Liberace (Roche Products Ltd, Welwyn Garden City, UK) in RPMI 1640 and incubated at 37°C for 30 mins. Pinnae were then torn into large pieces using tweezers and incubated with shaking for a further 30 mins. Auricular lymph nodes (sILN) that drain the pinnae were also removed from the infected mice. They were then cut into small pieces and incubated with 0.2 mg/ml DNAse (Sigma–Aldrich, UK) and 0.5 mg/ml collagenase D (Roche Products Ltd) for 20 mins. Pinnae and sILN cell suspensions were filtered through 100 μm metal gauze, washed in PBS pH 7.2 and enumerated prior to being labelled with antibodies and analysed by flow cytometry.

Intracellular staining of endosome compartments and co-localisation

BMMΦ and BMDC were cultured as previously described and seeded at 0.2×10⁶ onto circular cover slips. Cells were then stimulated for up to 18 hrs with CFDA-SE labelled 0-3hRP, and compared to the uptake of Alexa Fluor488 or 594-labelled E. coli bioparticles (1 μm; Invitrogen Ltd) representing a control microbial material and known to be a classical pro-Th1 stimulant. Cells attached to the coverslips were then fixed in 2% paraformaldehyde and permeabilised using 0.2% saponin (Sigma–Aldrich, UK) and 0.5 mg/ml collagenase D (Roche Products Ltd) for 20 mins. Pinnae and sILN cell suspensions were filtered through 100 μm metal gauze, washed in PBS pH 7.2 and enumerated prior to being labelled with antibodies and analysed by flow cytometry.

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Co-localisation of CFDA-SE labelled 0-3hRP or Alexa 488 E. coli bioparticles with the intracellular markers were analysed using Volocity 4.3.2 software to generate a co-localisation coefficient Mx.

$$Mx = \frac{\sum X_{i}^{coloc}}{\sum X_{i}}$$

The coefficient ranges from 0 to 1, with 1 indicating that the entire signal from one channel is co-localised with the other and 0 representing no co-localisation between channels. The threshold for each channel was generated automatically to exclude voxels for which Pearson’s correlation between the channels is less than or equal to 0, based on a technique from Costes et al [32].

Statistics
Changes in CFDA-SE labelled material uptake, cytokine production and differences in co-localisation were evaluated using Students t-test or one-way ANOVA (***, $P < 0.001$; **, $P < 0.01$; *, $P < 0.05$). Differences were considered significant when $P < 0.05$.

Results
Labelling of live S. mansoni cercariae with CFDA-SE
CFDA-SE dye preferentially labels material localised within the cercarial pre- and post-acetabular glands and their associated ducts as shown by the fluorescent 2D and 3D confocal images (Fig. 1A–B and supplementary Video S1). The relative absence of labelling on the outer surface of the cercaria, or within the body and tail, indicates that CFDA-SE does not cross the surrounding glycocalyx and suggests that the dye travels up the ducts to enter the acetabular glands which are rich in proteases [17,18]. Optimal concentrations and incubation conditions for labelling live cercariae with CFDA-SE were established (see supplementary Figure S1 A–E). Furthermore, labelling parasites with CFDA-SE under these optimal conditions did not adversely affect the infective potential or viability of cercariae compared to unlabelled parasites since both sets of cercariae had almost identical penetration efficiencies of approximately 70% (see supplementary Figure S1); it also does not affect their ability to mature into adults, or lay eggs (data not shown).

Transformation of cercariae initiates the release of CFDA-SE labelled acetabular gland contents
Transformation of CFDA-SE labelled cercariae into schistosomula leads to a decrease in their fluorescence (Fig. 2A). This results from the release of CFDA-SE labelled acetabular gland contents (ES material) which accompanies the transformation of cercariae. The contents of the acetabular glands contain numerous proteases that conventionally facilitate penetration of host skin [17,18]; this ES material was detected in the culture media, in which the parasites had transformed over 3 hours, as an increase in fluorescence (Fig 2A). CFDA-SE labelled ES material is clearly visible as discreet vesicles being released from the acetabular gland duct openings (Fig. 2B & C). CFDA-SE labelled material was only evident within the contents of the vesicle and not the surrounding material (Fig. 2C).

Uptake of released CFDA-SE labelled material by adherent cells
Live CFDA-SE labelled cercariae were cultured with PEC to examine the ability of phagocytic cells to internalise larval ES products. The majority of PEC obtained at this time point (day 5) were CD11b+ MΦ rather than neutrophils (data not shown).
frequency of MHC-II population, ES material released by live cercariae enhanced the exhibited even distribution (Fig. 3B). Within the adherent exposed to an equivalent concentration of CFDA-SE dye alone within distinct intracellular components, whereas cells directly uptake was a specific process is that CFDA-SE label localises indeed been phagocytosed. The lack of uptake by non-adherent uptake of CFDA-SE labelled 0-3hRP by 77.43% and 67.89% Moreover, over 40% of MHC-II labelled 0-3hRP was significantly greater (P (BMM (5 mM) or cytochalasin D (10
m
of CFDA-SE demonstrating that cells with potential antigen contents are released in order to facilitate parasite migration. As the parasite continues to migrate, fluorescence associated with the larval head progressively declines in the acutabular glands, compatible with the notion that the gland contents are released in order to facilitate parasite migration. Moreover, the migration path of the parasite is revealed as a trace of CFDA-SE material left in its wake.

Skin cells extracted from the pinnae of mice infected with labelled cercariae, and analysed by flow cytometry revealed that up to ~7% of CD45 cells were CFDA-SE+. 3 hrs after infection (Fig. 5B); By 48 hrs, there was a significant decrease (P<0.01) in the percentage of CD45 cells that were CFDA-SE+, which was followed by a further decline by 72 hrs when the majority of parasites should have left the skin [4]. Phenotypic analysis of the
cells showed that the labelled ES material was initially taken up by GR-1⁺ MHC-II⁺ cells (neutrophils), but by 48 hrs far fewer CFDA-SE⁺ GR-1⁺ MHC-II⁺ cells were detected (Fig. 5C). Both F4/80⁺ MHC-II⁺ and CD11c⁺ MHC-II⁺ cells, predicted to be skin-derived MΦ and DC respectively, were also CFDA-SE⁺ demonstrating that antigen presenting cells (APC) in the skin had taken up ES material released in vivo by invading larvae, or had taken up apoptosing neutrophils that had previously taken up CFDA-SE material. The number of DC and MΦ recovered from the skin that were CFDA-SE⁺ peaked at 24 hrs but declined thereafter (P<0.01) possibly reflecting their onward migration to draining lymphoid tissues. Our data also show that the proportions of MΦ and DC in the skin that were CFDA-SE⁺ were similar at each time point (P>0.05; Fig. 5C). However, as the total number of MΦ in the pinnae after infection (CFDA-SE⁺ and CFDA-SE⁺ cells combined) is approximately twice that of DC (5.37±0.39×10⁵ cf. 3.01±0.27×10⁵ at 24 hrs), we infer that MΦ are less efficient than DC at taking up CFDA-SE material.

CFDA-SE⁺ cells were also detected in the sdLN that drain the infection site. Although only negligible numbers of CFDA-SE⁺ cells were recorded in the sdLN by 24 hrs (Fig 5D), virtually no CFDA-SE⁺ GR-1⁺ MHC-II⁺ cells were detected in the sdLN at any time (data not shown) implying the lack of recruitment of neutrophils to this location, or that they had rapidly been removed following apoptosis. The vast majority of CFDA-SE⁺ cells in the sdLN were either F4/80⁺ MHC-II⁺ or CD11c⁺ MHC-II⁺ (Fig. 5E) indicating that MΦ and DCs which

Figure 3. Material released by CFDA-SE-labelled cercariae is phagocytosed by adherent MHC-II⁺ cells. A: Uptake of the labelled material in adherent cells is significantly greater than non-adherent cells; (p<0.001). B: Representative fluorescent, and merged with brightfield, images of adherent PECs exposed to unlabelled or CFDA-SE labelled cercariae, or directly to free CFDA-SE. C: Cells that take up ES material expressed elevated levels of MHC-II and 40.4% of these were also positive for CFDA-SE. All results for PEC are representative of 5 independent experiments. D: Fluorescence of BMMΦ stimulated with unlabelled or CFDA-SE labelled cercariae, or unlabelled or CFDA-SE labelled 0-3hRP with flow cytometry histogram plots displaying MFI (Unl Cerc = unlabelled cercariae, Unt CFDA-SE Cerc = untransformed labelled cercariae, Tr CFDA-SE Cerc = transformed labelled cercariae, RPMIc = concentrated RPMI, 0-3hRP = 0-3 hour released preparation). E: Addition of EGTA (5 mg/ml) or Cytochalasin D (10 µg/ml) significantly decreased uptake of CFDA-SE 0-3hRP. Representative plots are shown, bars are means±SEM where n = 6 mice.
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had taken up labelled parasite molecules in the skin had migrated to the sdLN. Alternatively, CFDA-SE labelled parasite antigen released by cercariae within the first 2 hrs as they penetrate (Fig. 5A and supplementary Video S5) may have drained freely to the sdLN and was processed by cells in situ. However, the lack of CFDA-SE+ cells at the earliest time point (3 hrs) in the sdLN would suggest that the incorporation of freely draining CFDA-SE released parasite material in the sdLN does not occur, although free fluorescein isothiocyanate painted directly on the skin could be detected in the sdLN by 3 hrs (data not shown). Rather, as the peak numbers of CFDA-SE+ MΦ and DC in the sdLN was reached at 48 hrs, we believe that this reflects the migration of antigen laden cells to the sdLN.

BMDC are more highly activated by CFDA-SE labelled cercariae and 0-3hRP than BMMΦ

As cells expressing surface markers characteristic of MΦ and DC were both observed to take up CFDA-SE labelled molecules in...
Figure 5. In vivo infection with CFDA-SE-labelled cercariae results in the release of labelled material in skin and its uptake by cells in the skin and sdLN. A; Stills from a time-lapse video (see supplementary Video S4) showing confocal images of a CFDA-SE-labelled infective cercaria penetrating and migrating through a mouse pinna, (scale bar = 100 µM). Attachment of the cercaria to the stratum corneum at 00:01:00, the loss of its tail by 00:18:00, penetration through outer layers of epidermis and deposition of gland material through 00:10:00 and onward migration up to 2 hours post-infection 02:00:00. B; Pinnae from mice infected with CFDA-SE labelled cercariae were digested and skin cell suspensions enumerated for CFDA-SE+ cells by flow cytometry (significant difference compared to 3 hrs). C; Phenotype of CFDA-SE+ cells from digested skin of pinnae exposed to CFDA-SE labelled cercariae showing uptake by GR-1+ MHC-II+, F4/80+ MHC-II+, and CD11c+ MHC-II+ cells. D; Phenotype of CFDA-SE+ cells from the sdLN of the same mice above showing uptake by F4/80+ MHC-II+, and CD11c+ MHC-II+ cells with a significant increased level of CFDA-SE+ cells present at 48 hrs (** denotes p = <0.01). Data in B–D is represents data from 6 different mice.

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the skin of infected mice, the relative reactivity of these two cell types to stimulation with molecules released by live cercariae was compared using parallel cultures of BMMD and bone marrow derived dendritic cells (BMDC).

A similar number of BMDC and BMMD internalised ES material released from CFDA-SE labelled cercariae but significantly more BMDC internalised CFDA-SE labelled 0-3hRP (Fig. 6A; P<0.05). BMDC also internalised greater amounts of both CFDA-SE ES and 0-3hRP as reflected in the significantly greater MFI (Fig. 6B). In addition, BMDC incorporated 0-3hRP initially at a faster rate than BMMD, and a greater proportion of BMDC had taken up CFDA-SE 0-3hRP at all time points (e.g. 50.27%±2.4 versus 32.47%±3.8 at 2 hrs; Fig. 6C). BMDC also expressed much higher MFI levels of CD40 CD86 and MHC II

![Image](image-url)

**Figure 6.** Comparison of the relative ability of BMMD and BMDC to internalise and be activated by CFDA-SE labelled material released from cercariae, or CFDA-SE-labelled 0-3hRP. A; BMDC internalise greater quantities of released material from cercariae or 0-3hRP compared to BMMD and B; express higher MFI±SEM. C; BMDC internalise 0-3hRP at a faster rate than BMMD. D; Expression of CD40, CD86 and MHC II of BMDC (red) and BMMD (blue) after stimulation with cercariae (dotted line) and 0-3hRP (solid line), values shown in insert are MFI±SEM. E; Levels of IL-12p40, TNF-α, IL-6 and IL-10 in BMMD (open bars) and BMDC (hatched bars). F; Levels of mRNA measured by qPCR for arginase 1 and iNOS (relative to GAPDH) in BMMD and BMDC. Cultures of BMMD and BMDC were derived from the same mouse, and data shows the mean±SEM for separate animals. Data represent cells obtained from 6 different mice.

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expression in response to both cercariae and 0-3hRP (Fig. 6D). Inflammatory cytokine (i.e. IL-6, IL-12p40 and TNF-α) output from BMDC in response to 0-3hRP was much greater than from BMMφ whereas the production of regulatory IL-10 was significantly lower ($P<0.01$; Fig. 6E). This suggests that Mφ exhibit a more regulatory phenotype than DC exposed to ES products released by transforming cercariae. To examine this question, we used qPCR to reveal that BMDC expressed significantly higher levels of inducible nitric oxide synthase (iNOS) transcript than BMMφ ($P<0.01$; Fig. 6F). This marker of ‘classical activation’ indicates that BMMφ are less active than BMDC at responding to 0-3hRP. In contrast, BMMφ had significantly elevated levels of arginase 1 mRNA ($P<0.01$) which converts L-arginine via an alternative pathway that does not yield toxic nitrogen products. This supports the idea that the cercarial ES promote the development of BMMφ that are regulatory.

Translocation of CFDA-SE labelled 0-3hRP to the LAMP-1+ phagosome is faster in BMDC than BMMφ

Both BMMφ and BMDC initially internalised CFDA-SE labelled 0-3hRP into the EEA-1+ compartment but BMDC translocated 0-3hRP into the LAMP-1+ phagosome faster than BMMφ (Fig. 7A). The rate of translocation of AF594 E. coli bioparticles was faster compared to CFDA-SE 0-3hRP for both BMDC and BMMφ but did not significantly differ between the two cell types (Fig. 7B). The reduced translocation rate by BMMφ for 0-3hRP was visualised by co-localisation with LAMP-1+ phagosomes at 15 mins in BMDC, whilst it was still present in EEA-1+ compartments in BMMφ (see supplementary Figure S3 A). By 30 mins, almost all the CFDA-SE labelled 0-3hRP was observed in LAMP-1+ compartments in BMDC but in BMMφ, it was still found in the EEA-1+ compartments (see supplementary Figure S3 B). Co-culture of AF594 E. coli bioparticles and CFDA-SE 0-3hRP revealed that the two antigens were internalised into different phagosomes within the same cell (Fig. 7C) and did not affect each others translocation from the early phagosome to the phagolysosome (data not shown).

Discussion

The skin and its associated immune cells is the first barrier to schistosome cercariae during infection but the impact of the dermal innate immune response on parasite survival and the development of adaptive immunity is largely unknown. In this study, cercariae were labelled with a fluorescent tracer in order to facilitate the visualisation of the parasite in the skin, the release of ES products, and uptake of parasite material by Mφ and DC in vitro and in vivo. Intriguingly, although both Mφ and DC take up labelled ES material released by cercariae (aka 0-3hRP), which is known to favour the development of pro-Th2 DC [15], it is processed at a slower rate than a typical pro-Th1 agent, and it is processed slower by Mφ than by DC. This suggests that the processing of parasite material in the skin by these two cell types might have significant effect on the balance of the immune environment to favour immune priming or immune regulation.

The amine reactive tracer CFDA-SE specifically labelled the contents of the cercarial acetabular glands which contain an abundance of tissue degrading protease enabling penetration of the skin [17,19]. Indeed, the presence of esterases or proteases in these glands presumably facilitates the efficient cleaving of the dye from its inactive to its fluorescent state [26]. Importantly, we determined that CFDA-SE did not alter the ability of acetabular gland products to activate innate immune MHC-II+ cells. Indeed, there was no change in the expression of MHC-II, CD40 and CD86, or the production of IL-6, IL-12, IL-10 and TNF-α in response to labelled compared with unlabelled cercariae or 0-3hRP. Therefore, we conclude that CFDA-SE is
an ideal fluorescent label with which to track the fate of parasite released molecules in relation to cells of the innate immune response.

CFDA-SE labelled material was released by cercariae only upon transformation into schistosomula. It was released as membrane-less vesicles [35] implying that the contents of the glands which contain numerous antigenic proteins and glycoproteins [10] are effectively labelled by CFDA-SE whilst still in the acutabular glands. As there is no de novo synthesis of protein by cercariae [30], the contents of the acutabular glands are pre-formed; as such it is the first antigenic material detected by the host’s innate immune response. However, the lack of labeling on the outer portion of the vesicle suggests that other non-protein molecules such as lipids and/or glycosans surround the protein rich contents as they are expelled from the sucker during skin penetration, and consequently may represent additional source of ligands for innate immune cells [36].

Fluorescent labelling of the parasites enabled for the first time detailed real time imaging of parasites as they penetrate into and through the skin. Cercariae were observed to firstly attach to the outer stratum corneum and then burrow into the epidermis. In every case, the cercarial tail detached from the parasite body at penetration and moved out of the field of view propelled by their continued movement. Cercarial tails (see supplementary Video S4 and Video S5) never entered the skin and therefore do not provide a source of material to modulate the immune response as suggested by others [37].

Shortly after attachment (~10 min), the acutabular gland contents were released supporting the idea that gland material is used not only as an aid for attachment but also that it is required for entry into the stratum corneum [38]. Others have postulated that the stratum corneum offers little barrier to cercariae since in aqueous conditions its structural integrity is lost and the parasites simply push through [1,3]. However, evidence shown here supports the view that acutabular gland material is released at the point of infection to aid penetration shown as a thick ring of fluorescent material. Whether this material is used to lyse cells, or the extra cellular matrix, is not clear. The progressive reduction in the fluorescence of the glands as the parasite migrates through the epidermis is revealed as a trace of fluorescence marking a ‘penetration tunnel’ [35,39]. By 2 hrs, most of the acutabular gland content appears to be spent [40,41]. However, some CFDA-SE material persists indicating that gland contents remain available for digesting the epidermal basement membrane [4,10], the dermis [19], or even a blood or lymphatic vessel, facilitating the parasite’s onward migration. Moreover, the presence CFDA-SE material in the skin until at least 48 hrs shows that some ES material persists in the skin, possibly as material just released by the invading larvae.

The uptake of CFDA-SE labelled material released from cercariae is largely an active actin and Ca2+ dependent phagocytic process, as uptake was inhibited by EGTA and cytochalasin D. Although the receptors responsible for uptake of cercarial ES are presently unknown, they are likely to include CD206 and CD209 which could recognise mannose- and fucose-rich glycoproteins abundant in 0-3hRP [22]. As the cells that phagocytosed labelled ES material were MHCII+, we conclude that the ultimate fate of phagocytosed gland contents is to be processed and then presented to the adaptive immune system. Cercarial ES and 0-3hRP were effective at activating these MHC-II+ cells through increased expression of MHC-II, co-stimulatory markers (CD40, CD86) and pro-inflammatory cytokines (IL-12p40/23, IL-6 and TNF-α) which would all promote their phagocytic activity, migratory capacity, and ability to act as effective APC.

The phagocytic machinery to internalise foreign particles results in the formation of a phagosome that matures and plays a key role in initiation of the immune system. However, the endosomal processing pathway for pro-Th2 0-3hRP was retarded compared to a typical pro-Th1 stimulus E. coli. Some microbes aid their survival [42] by disrupting the TLR signalling pathway involved in phagosome development [33,43]. For example, Mycobacterium tuberculosis arrests phagosome maturation by retaining EEA-1 on the phagosome [44]. Enhanced phagosome maturation (i.e. in response to E. coli) leads to increased processing of antigen to MHC-II molecules through the engagement of TLRs [45]. In the case of 0-3hRP, the reduced rate of phagosome maturation compared to E. coli could suggest that it has a reduced stimulatory response by being less efficient at binding and activating TLRs. Alternatively, 0-3hRP may trigger a different signalling pathway which does not efficiently promote phagosome maturation. For example, schistosome egg antigen (SEA) which also induces potent pro-Th2 DC [46] is reported to stimulate DC independent of TLR2, TLR4 and MyD88 [47,48], whilst filarial ES-62 appears to use a non-conventional signalling pathway [49,50]. Evidence that 0-3hRP and E. coli bioparticles do not colocalise within the same LAMP-1+ phagolysosome, supports the hypothesis that each phagosome is independent of each other [45]. A similar phenomenon occurs in BMDC co-pulsed with SEA (pro-Th2) and Propionibacterium acnes (pro-Th1) whereby the two stimulants occupy different locations within the same cell and induce contrasting Th subsets [51]. However, increased concentrations of P. acnes are suggested to enhance antigen processing and induce weak Th1-specific SEA specific responses [51]. The delayed transfer of 0-3hRP to the mature phagosome is one possible explanation for the limited maturation phenotype of DC stimulated with 0-3hRP shown by proteomic analysis [16]. Moreover, these DC exhibited limited expression of MHC-II and other co-stimulatory molecules, but were potent inducers of Th2 responses in vitro and in vivo [15].

By infecting the pinnae with CFDA-SE labelled cercariae, we determined that dermal-derived MΦ, DC, and neutrophils phagocytosed labelled ES in vivo. Eosinophils are rare in skin exposed to a single dose of cercariae as in this study but are highly abundant following multiple infections and appear to have an important role in defining an IL-4/IL-13 rich cytokine environment of the infection site (PC Cook & AP Mountford; manuscript in preparation). Neutrophils quickly influx into the infection site [7,10,25] and were the most abundant (~50%) cell type to have phagocytosed CFDA-SE labelled ES at 3 hrs. Neutrophils are an important source of chemokines which attract monocytes, MΦ and DC to the site of infection [52]. Indeed CCL3 and CCL4 are present at increased levels immediately after cercarial penetration [7]. The decline in CFDA-SE+ neutrophils after 24 hrs is likely to reflect rapid degradation of labelled ES material due to their potent proteolytic activity [53] followed by their rapid clearance from the skin; none were observed in the sdLN. The other dominant CFDA-SE+ cell populations in the skin at 3 hrs were MHC-II+ MΦ and DC which each accounted for ~25% of the total CFDA-SE+ cell population. As MΦ are more populous than DC in naive mouse skin [8], and in our study on infected pinnae are also much more abundant, we appear to show that DC take up labelled material more efficiently than MΦ.

As the numbers of CFDA-SE+ MΦ and DC in the skin peak at 24 hrs but declined thereafter, we infer that both cell types migrate to the sdLN. Indeed, LC emigrate from the epidermis to the sdLN following exposure to schistosome larvae [11], although their migration can be delayed by up to 48 hours in response to parasite-derived prostaglandin D2 [13]. A similar interpretation
could be argued for the data presented here as only a very small number of CFDA-SE^+ cells were detected in the local sdLN up to 48 hrs. Delayed migration could affect MΦ as well as DC, although it may also reflect differences in the temporal migration rates of the two types of cell. This delayed cell migration could aid parasite escape from the skin.

Although both MΦ and DC internalised CFDA-SE labeled 0-3hRP, DC phagocytosed greater amounts of antigen and ant at a faster rate. DC also expressed higher levels of activation markers, increased levels of IL-6, TNF-α and IL-12p40/23, and had significantly greater expression of iNOS. On the other hand, MΦ secreted significantly increased levels of regulatory IL-10 and had far more transcripts for arginase 1. In this context, schistosome larvae are known to induce the production of many different mediators with immunoregulatory function which serve to protect the parasite from immune attack but also to limit damage to the host caused by inflammation [6]. The production of IL-10 by the skin and skin-derived cells in response to schistosomiasis is critical in limiting IL-12 driven pathology in the skin [7,12,13,54]. Prostaglandin E2 which is released by cercariae upon transformation [54] is a potent inducer of IL-10 secretion from MΦ [55] and could be important in our model. The observation that BMΜΦ but not BMDC produce abundant IL-10 in response to cercarial ES may implicate MΦ as the possible source of this cytokine in vivo which in turn could mediate the actions of DC. In fact was recently reported that dermal-derived MΦ in the sdLN can produce IL-10 which directly suppresses the activity of DC [56]. High levels of IL-10 can cause reduced phagosomes maturation [57] and may help explain the limited maturation of MΦ in response to our ES material. The elevated levels of arginase-1 in our studies are also indicative of the MΦ having an ‘alternatively activated’ phenotype which is a feature of many helminth infections [58–60]. The balance of arginase/iNOS production is central in controlling the function of MΦ with arginase countering the pro-inflammatory cascade and production of NO [61]. Arginase-1 production by MΦ is also important in wound healing [62] and is a feature of tissue remodeling after repeated infection of the skin by schistosome cercariae (PC Cook & AP Mountford; manuscript in preparation). Our data here indicate that cercarial ES products directly drive MΦ to take on an ‘alternatively activated’ phenotype independent of other host derived immune mediators (e.g. IL-4 and IL-13).

Finally, the increased kinetics of antigen translocation through the endosomal pathway of BMDC is indicative of a higher activation rate and increased activation of these cells [33,45]. As DCs secrete higher quantities of IL-12 compared to MΦ in other infection models [63,64] and are more potent APC [65], our data suggest that DC favour pro-inflammatory responses but that MΦ have the capacity to regulate this response. The greater uptake of ES material by DC relative to MΦ in vitro and the greater proportion of DC that were CFDA-SE^+ in the skin is evidence that DC are more important than MΦ as APC. However, as the skin comprises both cell types, the relative abundance of MΦ versus DC within the inflammatory foci which form around schistosome larvae in the skin may explain why there is a balanced immune phenotype of stimulation and regulation [9]. It would be instructive to determine whether MΦ and DC in the skin of schistosome infected mice differ in their expression of various TLRs and C-type lectins that might explain their differential rates of processing of schistosome ES products and thus their function as APC. Manipulation of the skin’s immune response to promote the development of anti-parasite immune responses must therefore take account of DC populations to maximise presentation of parasite antigens but also to consider the regulatory role of skin-derived MΦ.

Supporting Information

Figure S1 Optimisation of labelling conditions with the amine reactive tracer CFDA-SE. A; The uptake of various concentrations of CFDA-SE label measured using a fluorometer and expressed as RFU per cercariae (n = 7 separate experiments and a minimum of 300 parasites examined for each experiment). B; The percentage of live motile cercariae was determined visually by light microscopy (n = 5 experiments, minimum 300 parasites observed in each experiment). C; Representative fluorescent images with bright field images shown insert of cercariae labelled with various concentrations of CFDA-SE (scale bars = 50 μm). D; Representative bright field and fluorescent images of cercariae labelled with CFDA-SE using different incubation times (scale bars = 50 μm). E; The persistence of CFDA-SE within un-transformed cercariae was determined over 24 hours (mean 300 cercariae±SEM). F; Penetration efficiency of unlabelled and CFDA-SE labelled cercariae into mouse pinnae (n = 5). Explanatory text: To optimise labelling of live cercariae with CFDA-SE, parasites freshly shed from the intermediate snail host were incubated with various concentrations of the amine reactive tracer. The fluorescence of cercariae, measured by fluorometry and expressed as relative fluorescent units (RFU) / cercaria, progressively increased with concentrations of up to 20 μM CFDA-SE (S2A). Above this, no increase in fluorescence was observed but the viability of cercariae, as judged by visual detection of body motility, opacity of parasite and flame cell movement, was dramatically reduced resulting in increased numbers of transformed (tail-less) and dead larvae (S2B). The fluorescence of individual cercaria as revealed by microscopy (S2C) confirmed that parasite labelling progressively increased up to 20 μM. The optimal duration for labelling using 20 μM CFDA-SE was 60 mins, with a high intensity of tracer within the parasite head (S2D). After 60 mins, increasing rates of parasite transformation and death were recorded (data not shown). The persistence of CFDA-SE within un-transformed cercariae was demonstrated as most of the label was retained by cercariae over a 24 hr period and did not decay, or leech out of the intact parasites (S2E). Labelling parasites with CFDA-SE did not adversely affect the infective potential of cercariae compared to unlabelled parasites since both sets of cercariae had almost identical penetration efficiencies of approximately 70% (S2F).

Found at: doi:10.1371/journal.pntd.0000528.s001 (4.31 MB TIF)

Figure S2 CFDA-SE does not affect the ability of parasite derived material to activate BMΜΦ. A; Expression of MHC-II, CD40 and CD86 in response to unlabelled and labelled cercariae (left hand panels) and 0-3hRP (right hand panels). B; The production of inflammatory (IL-12p40, IL-6 & TNF-alpha) and regulatory cytokines (IL-10) in response to unlabelled and labelled cercariae and 0-3hRP, and the controls LPS and RPMIc. Explanatory text: Both cercariae and 0-3hRP activate BMΜΦ as judged by increased expression of MHC-II, and the co-stimulatory molecules CD40 and CD86, regardless of whether they were labelled with CFDA-SE (S3A). CFDA-SE labelled cercariae and 0-3hRP induced the production of IL-12/23p40, IL-6, TNF-alpha and IL-10 at levels similar to those induced by equivalent numbers of unlabelled parasites or quantities of 0-3hRP (S3B). Therefore, the use of CFDA-SE to label cercariae or 0-3hRP does not affect their capacity to activate host phagocytic cells.

Found at: doi:10.1371/journal.pntd.0000528.s002 (2.34 MB TIF)

Figure S3 Confocal images on the prolonged translocation of 0-3hRP in BMΜΦ compared to BMDC. Confocal images of the translocation of 0-3hRP(green) within BMDC and BMΜΦ from the EEA-1^+ early phagosome (purple) to LAMP-1^+ phagolysosome
(red) at 15 mins (A) and 30 mins (B). Explanatory text: Both BMDC and BMMΦ translocate CFDA-SE labelled 0-3hRP from the early phagosome labelled with EEA-1 (Purple) to a phagolysosome labelled with LAMP-1. However this occurs at an increased rate with BMDC which can be deduced by the increased co-localisation of CFDA-SE 0-3hRP with LAMP-1 (red) at 30 minutes.

**Video S1** Labelling of cercariae with the amine reactive tracer CFDA-SE.

Found at: doi:10.1371/journal.pntd.0000528.s004 (0.17 MB MOV)

**Video S2** Co-localisation of 0-3hRP (green) with EEA-1 (purple) and LAMP-1 (red) within BMM

Found at: doi:10.1371/journal.pntd.0000528.s006 (0.09 MB (green) with EEA-1 (purple) and LAMP-1 (red) after 30 mins.

**Video S3** Co-localisation of Alexa Fluor 488 E. coli bioparticles (green) with EEA-1 (purple) and LAMP-1 (red) after 30 mins.

Found at: doi:10.1371/journal.pntd.0000528.s008 (0.06 MB MOV)

**References**

1. McKerrow JH, Salter J (2002) Invasion of skin by Schistosoma cercariae. Trends Parasitol 18: 193–195.

2. Curwen RS, Wilson RA (2003) Invasion of skin by schistosome cercariae: some neglected facts. Trends Parasitol 19: 63–66. discussion 66–68.

3. McKerrow JH (2003) Invasion of skin by schistosome cercariae: some neglected facts. Trends Parasitol 19: 66–68.

4. Wheeler PR, Wilson RA (1979) Schistosoma mansoni: a histological study of migration in the laboratory mouse. Parasitology 79: 49–62.

5. He YN, Salakub K, Ramaocaus K (2005) Comparison of skin invasion among three major species of Schistosoma. Trends Parasitol 21: 201–203.

6. Jenkins SJ, Hewinton JP, Jenkins GR, Mountford AP (2005) Modulation of the host’s immune response by schistosome larvae. Parasite Immunol 27: 385–393.

7. Hogg KG, Kumkate S, Andersson S, Mountford AP (2003) Interleukin-12 p40 secretion by cutaneous CD11c+ and F4/80+ cells is a major feature of the innate immune response in mice that develop Th1-mediated protective immunity to Schistosoma mansoni. Infect Immun 71: 3536–3547.

8. Dupasquier M, Stottner P, van Oudenaren A, Romani N, Leenen PJ (2004) Macrophages and dendritic cells constitute a major subpopulation of cells in the mouse dermis. J Invest Dermat 123: 876-879.

9. Mountford AP, Trotein F (2004) Schistosomes in the skin: a balance between immune priming and regulation. Trends Parasitol 20: 221–226.

10. Incani RN, McLaren DJ (1984) Histopathological and ultrastructural studies of cutaneous reactions elicited in naive and chronically infected mice by invading schistosomula of Schistosoma mansoni. Int J Parasitol 14: 259–270.

11. Kumkate S, Jenkins GR, Pageley RA, Hogg KG, Mountford AP (2006) CD207+ Langerhans cells constitute a minor population of skin-derived antigen presenting cells in the draining lymph node following exposure to Schistosoma mansoni. Int J Parasitol Submitted.

12. Hogg KG, Kumkate S, Mountford AP (2005) IL-10 regulates early IL-12-mediated immune responses induced by the radiation-attenuated schistosome vaccine. Int Immunol 15: 1451–1459.

13. Angeli V, Faveere C, Ruy O, Fontaine J, Teissier E, et al. (2001) Role of the parasite-derived prostaglandin D2 in the inhibition of epidermal Langerhans cell migration during schistosomiasis infection. J Exp Med 193: 1135–1147.

14. Jenkins SJ, Hewinton JP, Ferret-Bernard S, Mountford AP (2005) Schistosome larvae stimulate macrophage cytokine production through TLR4-dependent and -independent pathways. Int Immunol 17: 1409–1418.

15. Jenkins SJ, Mountford AP (2005) Dendritic cells activated with products released by schistosome larvae drive Th2-type immune responses, which can be inhibited by manipulation of CD10 costimulation. Infect Immun 73: 395–402.

16. Ferret-Bernard S, Curwen RS, Mountford AP (2005) Proteomic profiling reveals that Th2-inducing dendritic cells stimulated with helminth antigens have a ‘limited maturation’ phenotype. Proteomics 5: 980–993.

17. Curwen RS, Ashton PD, Sundaralingam S, Wilson RA (2006) Identification of novel proteins, and immunomodulators and the secretions of schistosome cercariae that facilitate host entry. Mol Cell Proteomics 5: 835–844.

18. Knudsen GM, Medzihradzky KF, Lim KC, Hansell E, McKerrow JH (2005) Proteomic analysis of Schistosoma mansoni cercariae secretions. Mol Cell Proteomics 4: 1862–1873.

19. Salter JP, Lim KC, Hansell E, Hinch I, McKerrow JH (2000) Schistosome invasion of human skin and degradation of dermal elastin are mediated by a single serine protease. J Biol Chem 275: 38667–38673.

**Video S4** Infection of murine pinnae by CFDA-SE cercariae and deposition of released labelled ES material over 120 mins.

Found at: doi:10.1371/journal.pntd.0000528.s007 (0.75 MB MOV)

**Video S5** Initial infection of murine pinnae by CFDA-SE cercariae and release of tail imaged by time lapse confocal microscopy over 60 mins.

Found at: doi:10.1371/journal.pntd.0000528.s008 (0.06 MB MPEG)

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**Author Contributions**

Conceived and designed the experiments: RAP APM. Performed the experiments: RAP SAA. Analyzed the data: RAP SAA. Contributed reagents/materials/analysis tools: PCC. Wrote the paper: RAP PCC JDT APM.

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38. Stirewalt MA (1974) Schistosoma mansoni: cercaria to schistosomule. Adv Parasitol 12: 115–182.
39. Wang L, Li YL, Fishelson Z, Kusel JR, Ruppel A (2005) Schistosoma japonicum migration through mouse skin compared histologically and immunologically with S. mansoni. Parasitol Res 95: 218–223.
40. Brink LH, McLaren DJ, Smither SR (1977) Schistosoma mansoni: a comparative study of artificially transformed schistosomula and schistosomula recovered after cercarial penetration of isolated skin. Paraenetologia 74: 73–86.
41. Cousin GE, Stirewalt MA, Dorsey CH (1981) Schistosoma mansoni: ultrastructure of early transformation of skin- and shear-pressure-derived schistosomula. Exp Parasitol 51: 341–365.
42. Kinchum JM, Kavichandran KS (2008) Phagosome maturation: going through the acid test. Nat Rev Mol Cell Biol 9: 791–795.
43. Shiratsuchi A, Watanabe I, Takeuchi O, Akira S, Nakashima Y (2004) Inhibitory effect of Toll-like receptor 4 on fusion between phagosomes and endosomes/lysosomes in macrophages. J Immunol 172: 2039–2047.
44. Via LE, Deretic D, Ulmer RJ, Hibler NS, Huber LA, et al. (1997) Arrest of mycobacterial phagosome maturation is caused by a block in vesicle fusion between stages controlled by rah5 and rah7. J Biol Chem 272: 13326–13331.
45. Blander JM, Medzhitov R (2006) On regulation of phagosome maturation and antigen presentation. Nat Immunol 7: 1029–1035.
46. MacDonald AS, Maizels RM (2008) Alarming dendritic cells for Th2 induction. J Exp Med 205: 13–17.
47. Kane CM, Jung E, Pearce EJ (2008) Schistosoma mansoni egg antigen-mediated modulation of Toll-like receptor (TLR)-induced activation occurs independently of TLR2, TLR4, and MyD88. Infect Immun 76: 5754–5759.
48. Marshall FA, Pearce EJ (2008) Uncoupling of induced protein processing from maturation in dendritic cells exposed to a highly antigenic preparation from a helminth parasite. J Immunol 181: 7562–7570.
49. Marshall FA, Grierson AM, Garside P, Harnett W, Harnett MM (2005) ES-62, an immunomodulator secreted by filarial nematodes, suppresses clonal expansion and modifies effector function of heterologous antigen-specific T cells in vivo. J Immunol 175: 5817–5826.
50. Goodridge HS, Stepek G, Harnett W, Harnett MM (2005) Signalling mechanisms underlying subversion of the immune response by the filarial nematode secreted product ES-62. Immunology 115: 296–304.
51. Cerri L, MacDonald AS, Kane C, Dziarszinska F, Pearce EJ (2004) Cutting edge: dendritic cells copulsed with microbial and helminth antigens undergo modified maturation, segregate the antigens to distinct intracellular compartments, and concurrently induce microbe-specific Th1 and helminth-specific Th2 responses. J Immunol 172: 2016–2020.
52. Nathan C (2006) Neutrophils and immunity: challenges and opportunities. Nat Rev Immunol 6: 173–182.
53. Savina A, Amigorena S (2007) Phagocytosis and antigen presentation in dendritic cells. Immunol Rev 219: 145–156.
54. Ramaswamy K, Kumar P, He YX (2000) A role for parasite-induced PGE2 in IL-10-mediated host immunoregulation by skin stage schistosomula of Schistosoma mansoni. J Immunol 165: 4567–4574.
55. Strassmann G, Patil-Rootta V, Finkelman F, Fong M, Kambayashi T (1994) Evidence for the involvement of interleukin 10 in the differential deactivation of murine peritoneal macrophages by prostaglandin E2. J Exp Med 180: 2363–2370.
56. Toichi E, Lu KQ, Swick AR, McCormick TS, Cooper KD (2008) Skin-infiltrating monocytes/macrophages migrate to draining lymph nodes and produce IL-10 after contact sensitizer exposure to UV-irradiated skin. J Invest Dermatol 129: 2705–2715.
57. Via LE, Fratti RA, McFalone M, Pagans-Ramos E, Deretic D, et al. (1998) Effects of cytokines on mycobacterial phagosome maturation. J Cell Sci 111(Pt 7): 987–997.
58. Herbert DR, Holcher C, Mohrs M, Arendse B, Schwemmman A, et al. (2004) Alternative macrophage activation is essential for survival during schistosomiasis and downmodulates T helper 1 responses and immunopathology. Immunity 20: 623–635.
59. Anthony RM, Urban JJ Jr, Alem F, Hamed HA, Rozo CT, et al. (2006) Memory Th1/2 cells induce alternatively activated macrophages to mediate protection against nematode parasites. Nat Med 12: 955–960.
60. Nair MG, Cochrane DW, Allen JE (2003) Macrophages in chronic type 2 inflammation have a novel phenotype characterized by the abundant expression of Ym1 and Fizz1 that can be partly replicated in vitro. Immun Lett 85: 173–180.
61. Munder M, Eichmann K, Modell M (1998) Alternative metabolic states in murine macrophages reflected by the nitric oxide synthase/arginase balance: competitive regulation by CD14+ T cells correlates with Th1/Th2 phenotype. J Immunol 160: 3347–3354.
62. Shearer JD, Richards JR, Mills CD, Caldwel MD (1997) Differential regulation of macrophage arginine metabolism: a proposed role in wound healing. Am J Physiol 272: E181–190.
63. Siegemund S, Schutze N, Freudenberg MA, Lutz MB, Strabinger RK, et al. (2007) Production of IL-12, IL-23 and IL-27p28 by bone marrow-derived conventional dendritic cells rather than macrophages after LPS/TLR4-dependent induction by Salmonella Enteritidis. Immunobiology 212: 739–750.
64. Liu CH, Fan YT, Dias A, Esper L, Corn RA, et al. (2006) Cutting edge: dendritic cells are essential for in vivo IL-12 production and development of resistance to Toxoplasma gondii infection in mice. J Immunol 177: 31–35.
65. Steinman RM (1991) The dendritic cell system and its role in immunogenicity. Annu Rev Immunol 9: 271–296.