**Introduction**

Plants possess an effective innate immune system that is activated upon recognition of microbial pathogens. Plants recognize microorganisms by detecting microbe-associated molecular patterns (MAMPs), such as bacterial flagellin, which leads to MAMP-triggered immunity (MTI) [1]. Successful pathogens evolved ingenious mechanisms to evade or suppress host immunity [2–4]. An important class of immune suppressive proteins produced by pathogenic microbes are the so-called effector proteins, which are often able to suppress MTI responses in susceptible hosts. Effectors are pathogen-derived proteins that facilitate infection by altering host cellular processes [5]. Plant pathogenic bacteria, fungi and oomycetes all use effector proteins during infection [6]. These effector proteins enable pathogens to colonize their host and cause disease. Numerous effector proteins are recognized by disease resistance (R) proteins, leading to effector triggered immunity (ETI). The outcome of a specific plant-pathogen interaction depends on the specific repertoires of R genes and effectors in the plant and pathogen.

Oomycetes are fungus-like eukaryotic microorganisms related to brown algae [7]. Oomycete plant pathogens cause some of the most destructive plant diseases in the world. The potato late blight pathogen *Phytophthora infestans*, for example, is responsible for a loss in crop yields of over €1 billion a year in the European Union alone. Other oomycetes, such as *Phytophthora sojae* and *Peronosclerospora soehngenii*, also cause severe damage to the economically important crops soybean and sorghum, respectively [8–10]. Research on oomycete pathogens gained new impetus in the early-90s with genomic research and the cloning of oomycete avirulence (*Avr*) effectors defined initially as recognized by specific host *Avr* genes) and pathogenicity genes [11–14]. The discovery of conserved motifs in oomycete *Avr* effector proteins, coupled with whole genome sequencing of several oomycete pathogens resulted in the identification of hundreds of putative *Avr/effector* proteins [15–18]. Oomycete effector proteins can broadly be divided into two groups; effectors that are released into the plant apoplast and effectors that are delivered into the host cytoplasm. Host-translocated effectors were initially identified because of their avirulence function, which causes them to trigger a hypersensitive response in plants that carry the matching *R* gene. With the aid of evolutionary genomics approaches, two main types of oomycete
host-translocated effectors are currently recognized, Crinklers and RXLR-effectors [19–23].

In the last decade, a large number of RXLR effector proteins have been identified [24]. ATR1 and ATR13 are two effectors that are produced by the downy mildew pathogen *Hyaloperonospora arabidopsidis*, a pathogenic oomycete that infects *Arabidopsis thaliana* [19,22]. ATR1 and ATR13 each have an N-terminal signal peptide followed by the highly conserved RXLR amino acid sequence motif. This motif codes for an arginine (R), a random amino acid (X), a leucine (L) and another arginine and can be found in effector proteins of different oomycetes, suggesting that it is important for their function. The RXLR motif is sometimes followed by the less conserved dEER motif which consists of two glutamic acid residues and an arginine residue, preceded by an optional aspartic acid residue [22]. Apart from its presence in oomycetes, an RXLR-like motif has been found in proteins identified in malaria parasites (*Plasmodium* species), which are also translocated into host cells. In *Plasmodium* species this motif is called the HT/PEXEL (host-targeting/Plasmodium export element) motif and was demonstrated to be essential for translocating the HT/PEXEL proteins into host blood cells [25,26]. This, together with the finding that matching R proteins often reside in the host cytoplasm, led to the hypothesis that the RXLR motif has a role in protein delivery into host cells. This was confirmed by the finding that the RXLR-EER motif of the *P. infestans* effector protein Avr3a could target *Plasmodium* proteins into the erythrocyte cytoplasm [27]. Furthermore, substitutions of the RXLR and dEER motifs of effectors Avr1b and Avr3a with other residues blocked translocation of these proteins into plant cells [28,29], indicating that the RXLR motif is essential for cytoplasmic delivery of these effectors. Kale et al. [30] proposed that the RXLR domain binds to the phospholipid phosphatidylinositol-3-phosphate (PIP), which is then followed by endocytosis. However, Vaeno et al. [31] showed that a positively charged patch in the effector domain of Avr3a, and not the RXLR domain, is involved in PIP binding [23,24,30,31]. Hence, the exact role of the RXLR motif in protein translocation remains unclear [32].

Another important aspect of oomycete effector biology is the identification of effector host targets. With over 130 putative RXLR effector genes in the *H. arabidopsidis* genome, over 350 predicted RXLR effectors in *P. ramorum* and *P. sojae*, and more than 530 RXLR effector sequences in the genome of *P. infestans*, assigning functions to all of them is an enormous challenge [16–18,22]. A number of screens to determine virulence and/or avirulence functions of predicted RXLR effectors have already been undertaken. Screening of large numbers of putative RXLR effector proteins for their ability to trigger specific cell death responses resulted in the identification of only a small number of RXLR proteins with an avirulence function [33–37]. In contrast, the search for RXLR effectors that contribute to virulence resulted in more positive candidates. In a screen of 169 putative effectors of *P. sojae*, most were able to suppress programmed cell death responses [33]. In another study a set of 64 RXLR effector candidates of *H. arabidopsidis* isolate Emoy2 were tested for their ability to suppress callose deposition and growth of the bacterial pathogen *Pseudomonas syringae* in 12 A. thaliana accessions. A total of 43 RXLR proteins were found to enhance bacterial growth and 35 suppressed callose deposition [36], confirming the notion that RXLR effector proteins function by modulating host immunity. An example of this is RXL44 of *H. Arabidopsidis* Emoy2 that is able to downregulate salicylic acid-triggered defense responses by targeting the host’s Mediator subunit 19A for degradation [30].

Cabral et al. [39] recently described a set of 18 RXLR-containing proteins that were produced by *H. arabidopsidis* isolate Waco9 during infection of *A. thaliana*. These putative effectors were identified from Expressed Sequence Tags (ESTs), which were obtained from leaves of the highly susceptible *A. thaliana* *Ws eds1-1* mutant infected with the *H. arabidopsidis* strain Waco9. Additional sequencing of alleles of the 18 RXLR genes in other *H. arabidopsidis* isolates revealed signs of diversifying selection, supporting a putative effector role of the identified RXLR proteins. Furthermore, Cabral et al. [39] showed that one of the identified RXLRs, RXLR29, is able to suppress MTI and enhances disease susceptibility to *P. syringae* in *A. thaliana*. In order to identify potential functions of the other RXLRs we undertook intensive screening of 13 of the 18 RXLR genes described by Cabral et al. [39], and assessed their effect on host immunity. Transgenic *A. thaliana* plants expressing *H. arabidopsidis* RXLR genes were generated and screened for enhanced susceptibility to several different pathogens. In addition, the RXLR expressing lines were checked for altered MTI responses. Furthermore, we used the EDN-system, which exploits the bacterial type III secretion system to secrete proteins into host cells, to deliver the RXLR proteins to *A. thaliana* leaf cells in order to confirm actions of selected RXLRs in suppressing MTI.

### Materials and Methods

#### Sequence analysis

To assess sequence conservation in the 18 RXLR effector proteins identified by Cabral et al. [39], protein sequences were aligned using CLC Main Workbench software (www.clcbio.com), resulting in the identification of 4 distinct groups. Subsequently, the members within each group were aligned. A similarity-score per position of the multiple sequence alignment was derived based on the similarity of each amino acid and the five amino acids upstream and downstream to the consensus sequence of that group. The conservation-score per site was converted into a graphical representation via Matrix2png [40].

To identify (for Emoy2 RXLR proteins) and confirm (for Waco9 RXLR proteins) clusters of similar N-terminal RXLR effector regions, 130 Emoy2 RXLR proteins were retrieved alongside 18 Waco9 RXLR sequences from Baxter et al. [18] and Cabral et al. [39], respectively, from which the N-terminal 60 amino acids were extracted. The similarity between these sequences was established using BLASTp [41] with an e-value cutoff of 1e-5. Clusters of similar N-terminal regions were formed based on the similarity using the MCL clustering algorithm [42,43] with an inflation value of 2.

#### Cultivation of plants

*A. thaliana* accession Col-0 and Col-0 mutants *npr1-1* [44], fli2 (SALK_141277) [45] and *eds1-2* [46] were used. Seeds of *A. thaliana* lines were sown on quartz sand. Two-week-old seedlings were transferred to 60-mL pots containing a sand-potting soil mixture that had been autoclaved twice for 20 min with a 24-h interval. *A. thaliana* used for *H. arabidopsidis* and *Phytophthora capsici* inoculations were sown directly on this sand-potting soil mixture. Plants were cultivated in a growth chamber with a 10-h day (200 μEm s⁻²m⁻²) at 21°C and a 14-h night (20°C) cycle at 70% relative air humidity. Plants were supplied with modified half-strength Hoagland nutrient solution once a week [47], as described [48].

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Construction of transgenic \textit{A. thaliana} lines

The open reading frames of \textit{H. arabidopsis} isolate Waco9 RXLR genes without the predicted signal peptide [39] were cloned in pENTR/D-TOPO (Invitrogen) and transferred by LR recombination (Invitrogen) into the Gateway destination binary vectors pAMPT-GW with \textit{Cauliflower mosaic virus} 35S promoter driven expression. Constructs were transferred to \textit{Agrobacterium tumefaciens} strain GV3101 (pMP90RK) and transformed into \textit{A. thaliana} Col-0 using the floral-dip method [49]. Transformants were selected by spraying T1 progeny with BASTA Finale SL14 (Bayer CropScience BV, Mijndrecht, The Netherlands) according to the manufacturers instruction. From 1:3 segregating T2 lines, homozygous T3 lines were obtained for further testing.

The \textit{in planta} expression of the Waco9 RXLR genes was verified using semi-quantitative RT-PCR. Three-week-old plants were harvested and total RNA was isolated as described by Van Wees et al. [50]. Fermentas RevertAid H minus Reverse Transcriptase (Fermentas, St. Leon-Rot, Germany) was used to synthesize cDNA. The cDNA was amplified for 20, 25 or 30 cycles using gene-specific primer pairs (Table S1).

Cultivation of pathogens and pathogen inoculation

\textit{H. arabidopsidis} isolate Waco9 was maintained on susceptible Col-0 plants as described [51]. Sporangia were obtained by washing seedlings that were densely covered by sporangiophores in water. The obtained suspension was filtered using Miracloth and diluted to a concentration of 50 spores·μL⁻¹. \textit{A. thaliana} seedlings of 14 days old were sprayed with the spore suspension and dried for 2 h. Subsequently, the plants were placed at 20 °C, 9-h day (100 μE·m⁻²·s⁻¹), 15-h night and 100% relative humidity for 6 days. Disease was scored by determining the number of sporangiofophores per plant [52].

\textit{P. capsici} LT3112 [53] was grown on V8 agar plates for one week at 21 °C, 10-h light and 14-h dark. To collect zoospores, overgrown agar slices were placed in 10 mL sterile dH₂O for 1 h at 20 °C. Subsequently, the dH₂O was replaced with fresh dH₂O and the agar slices were kept at 24°C overnight followed by 1 h at 4°C. The dH₂O was checked for zoospores and diluted to a concentration of 50 zoospores·μL⁻¹. Plants were sprayed with the zoospore suspension and kept at 21°C, 100% air humidity and complete darkness for 24 h after which the plants were transferred to standard growing conditions with 100% relative humidity. Six days after inoculation disease severity was determined by scoring the percentage of diseased leaves per plant. Leaves were scored as diseased when macroscopically visible necrotic lesions were present [53].

\textit{P. syringae} pv. \textit{tomato} DC3000 strains were cultivated overnight in King’s medium B at 28°C. For growth of \textit{P. syringae} pv. \textit{tomato} DC3000 ΔCEL [54], 50 μg·mL⁻¹ rifampicin and 100 μg·mL⁻¹ spectinomycin was added to the medium. Bacterial cells were collected by centrifugation (4000 rpm, 10 min) and for dipping experiments the bacteria were resuspended and diluted to a final concentration of 2.5·10⁻⁷ μL⁻¹ in 10 mM MgSO₄ containing 0.015% Silwet L-77. Leaves of 5-week-old \textit{A. thaliana} plants were dipped in this bacterial suspension, after which the plants were placed at 100% relative air humidity. Disease levels were assessed at 3 or 4 d post inoculation by determining the percentage of leaves showing symptoms. Leaves were scored as diseased when necrotic or water-soaked lesions surrounded by chlorosis were visible. Disease index was determined as described [48].

Flg22-mediated growth reduction assay

Sterile \textit{A. thaliana} seeds were placed in 24-well plates containing 1 mL of MS medium (4.4 g·L⁻¹ MS, 10 g·L⁻¹ sucrose, pH 5.7). Ten seeds were placed in each well and 3 wells were used per line per treatment. Depending on the treatment, flg22 was added to a final concentration of 50 nM or 500 nM. After 2 days at 4°C the plates were placed at standard plant growth conditions (as described under cultivation of plants) for 10 days. Subsequently, the fresh weight of the total plant biomass per well was measured and the number of plants in each well was determined as described [2].

Statistical analysis

Redundancy analyses (RDA) were applied to the phenotypic data obtained in the different experiments of this study (Hellinger transformed within treatment) to perform a multifactorial test on the effects of the RXLR overexpressors on different components of the plant immune system (coded as binary variables). The significance of the models and of each explanatory variable included in the models was tested using 1000 permutations. Phenotype scores from the significant (Pr(>F) < 0.05) RDA axes (RDA1 and RDA2) were used in RDA ordination to perform hierarchical cluster analysis according to the ‘Ward method’, and the resulting dendrogram was projected in the RDA ordination space. This allows identification of the main discontinuities among groups and/or genotypes described by all descriptors [55]. Identification of groups was done using the pvrect function in the \textit{pvclust} package in the software package R. All multivariate analyses were performed with the software package R 2.15.2 “Trick or Treat” [56] using the \textit{vegan} [57] and \textit{pvclust} [58] packages. For the heatmap, the relative averages of the different experiments were placed in a matrix and this matrix was converted into a heatmap using Matrix2png [40].

Callose staining and microscopic analysis

Analysis for callose deposition was performed mainly as described [39,59]. In brief, leaves of 5-week-old \textit{A. thaliana} accession Col-0 plants were pressure-infiltrated with a 2-ml syringe containing a bacterial suspension consisting of 1·10⁸ cfu·mL⁻¹ \textit{P. syringae} pv. \textit{tomato} DC3000 ΔCEL in 10 mM MgSO₄. A total of 80 leaf samples were taken for callose staining 12 to 14 h after infiltration. Leaves were cleared with 100% ethanol, re-hydrated and stained with aniline blue (0.05% in phosphate buffer pH 8.0) for 24 h. Samples were analyzed with an Olympus AX70 microscope using an UV filter. Callose spots were counted using the ImageJ software [http://rsb.info.nih.gov/ij/] [60].

Results

RXLR effectors can be separated into groups based on their conserved N-termini

RXLR proteins consist of an N-terminal signal peptide followed by an RXLR domain and a C-terminal effector domain. Based on their amino acid sequences, the 18 RXLR proteins identified by Cabral et al. [39] could be divided in four groups of two or three RXLR proteins each, and eight RXLR proteins that showed no similarity to the other RXLRs. Strikingly, within each of the four groups the N-terminus is highly conserved while the C-terminus is very divergent (Figure 1A). To investigate whether amino acid sequence conservation in the N-terminus of RXLR proteins is a common phenomenon, we aligned all 134 identified RXLR sequences in the genome of the sequenced \textit{H. arabidopsidis} isolate Emo2 [18]. Based on the first 60 amino acids of each RXLR protein, around 60 percent of the RXLRs can be placed in a
group with at least one other RXLR protein and in most cases the similarity between proteins within one group is limited to the N-terminus (data not shown). In Figure 1B the amino acid sequence conservation pattern of the six *H. arabidopsidis* RXLRs of isolate Emoy2 [18] that group with RXLR13 and RXLR23 from Waco9 is shown. The N-termini show an amino acid sequence similarity of 60%-80%, while the sequence similarity in the C-terminal parts is relatively low. Thus, conservation of the N-terminal seems to be common for RXLR proteins in *H. arabidopsidis*.

**In planta** expression of Waco9 RXLR effectors

The RXLR gene transcripts of *H. arabidopsidis* identified by Cabral et al. [39] are expressed during infection, suggesting a role in pathogen virulence. While for other oomycete pathogens transformation protocols have been established [12,61,62], it is currently still not possible to transform *H. arabidopsidis*. Hence, in order to investigate the function of *H. arabidopsis* effector proteins in the infection process, *A. thaliana* plants were transformed to constitutively express a single RXLR effector gene. Of the 18 RXLR effector genes identified by Cabral et al. [39], the coding region without the signal peptide of 13 was successfully cloned behind the constitutive 35S CaMV promoter and transformed into *A. thaliana* (Table 1). Independent lines of RXLR transgenes that were segregating for a single transgene and had different levels of expression were selected (Figure 2). To check if expression of the RXLR effector genes affected plant growth and development, the rosette diameters and leaf morphology were monitored during a growth period of five weeks. In none of the transgenic lines did expression of the RXLR gene lead to a noticeably altered plant phenotype (data not shown).

Expression of single Waco9 RXLR effector genes does not affect basal resistance against *H. arabidopsidis*

To test whether overexpression of any of the single Waco9 RXLR genes in *A. thaliana* results in increased susceptibility to infection by the downy mildew pathogen, two-week-old RXLR-expressing seedlings were infected with *H. arabidopsidis* Waco9. After six days, disease levels were scored by counting the number of sporangiophores on each seedling. This was done in four independent experiments, in which two independent lines of each RXLR gene were tested twice. In all experiments the highly susceptible mutant Col-0 edsl-2 [46] was included as a control. In Figure 3, the number of sporangiophores in the RXLR transgenic lines compared to wild-type Col-0 is shown for all four experiments. In all four experiments, edsl-2 showed an enhanced susceptibility to *H. arabidopsidis* compared to Col-0. Only in

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**Figure 1. N-terminal sequence conservation of *H. arabidopsidis* RXLR proteins.** (A) The amino acid sequences of the 18 RLXR proteins of *H. arabidopsidis* isolate Waco9 identified by Cabral et al. [38] were grouped using BLASTp on the N-terminal 60 amino acids of these proteins. Color schemes show for each amino acid the similarity within the 11 amino acid region surrounding the specific amino acid. (B) N-terminal 60-amino acid regions of 130 Emoy2 RXLR proteins [18] were checked for similarity using BLASTp, leading to the identification of 23 groups containing at least two Emoy2 RXLRs. An alignment of the members of one of these identified groups is shown as example. RXL4, RXL5, RXL15, RXL26, RXL58, RXL76 are aligned and this group includes the homologs of RXLR13 (RXL76) and RXLR23 (RXL4) of *H. arabidopsidis* Waco9. Alignments are depicted as in (A). doi:10.1371/journal.pone.0110624.g001
| RXLR     | Emoy2 \(^1\) | Remark | Size (AA) | RXLR dis. \(^2\) | EER dis. \(^2\) | Homologs           |
|----------|--------------|--------|-----------|------------------|----------------|-------------------|
| RXLR3    |              |        | 129       | 29               | -              | H. arabidopsisidis |
| RXLR4    |              |        | 134       | 29               | -              | H. arabidopsisidis |
| RXLR6    | HaRXL60      |        | 129       | 27               | 44             | H. arabidopsisidis |
| RXLR9    | HaRXL78      |        | 150       | 28               | -              | H. arabidopsisidis |
| RXLR13   | HaRXL76      |        | 286       | 32               | 49             | H. arabidopsisidis |
| RXLR16   | HaRXL30; HaRXL79 |    | 198       | 28               | 41             | H. arabidopsisidis |
| RXLR17   | HaRXL42      |        | 135       | 28               | 41             | H. arabidopsisidis |
| RXLR19   |              |        | 299       | 31               | -              | H. arabidopsisidis |
| RXLR20   | HaRXL10      |        | 241       | 23               | -              | H. arabidopsisidis |
| RXLR21   | HaRXL37; HaRXL75 |    | 115       | 29               | 44             | Oomycetes         |
| RXLR22   |              |        | 137       | 29               | -              | Waco9 specific    |
| RXLR23   | HaRXL4       |        | 304       | 32               | 49             | H. arabidopsisidis |
| RXLR29   |              |        | 132       | 28               | -              | H. arabidopsisidis |

\(^1\) Gene ID of RXLRs from Emoy2 [18].

\(^2\) Number of amino acid to signal peptide cleavage site.

\(^3\) When present, number of amino acid to signal peptide cleavage site.

\(^4\) Homologs present in this group, adapted from Cabral et al. [18].

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experiment 1 and 4 did one or two of the 13 RXLR overexpressing lines permit enhanced H. arabidopsidis growth compared to Col-0. However, none of the RXLR-overexpressing lines showed a consistently altered level of disease resistance in more than one experiment. We therefore concluded that none of the 13 tested H. arabidopsidis RXLR effector proteins has a strong effect on the level of susceptibility to H. arabidopsidis when ectopically expressed in planta.

Waco9 RXLRs can alter P. capsici resistance levels in A. thaliana

The RXLR overexpressing lines were also tested for resistance against P. capsici. Initially, this oomycete has a similar biotrophic lifestyle as H. arabidopsidis, i.e. growing intercellularly and forming haustoria that penetrate host cell walls. In contrast to H. arabidopsidis, P. capsici switches to a necrotrophic lifestyle in later stages of infection and produces a largely different set of RXLR effector proteins [17,18,53]. Two-week-old seedlings were infected with P. capsici and 6 days later the plants were scored for disease severity. Again, disease assays were performed in four independent experiments, in which two independent lines of each RXLR gene were tested twice. In all experiments the highly susceptible mutant eds1-2 was included as a control. On wild-type Col-0, disease symptoms developed from small necrotic lesions to completely dead plants at day 6 after inoculation. In all experiments, disease caused by P. capsici developed faster on eds1-2, resulting in more severe symptoms at day 6 after inoculation (Figure 4). Strikingly, eleven out of thirteen RXLR overexpressing lines showed an altered disease phenotype upon inoculation with P. capsici. RXLR9- and RXLR23-overexpressing lines exhibited a lower level of disease than Col-0 in two of the four experiments, while the RXLR29 overexpressor developed significantly fewer disease symptoms in three of the four experiments. In contrast, overexpression of RXLR20 resulted in a significant increase in disease severity in one of the four experiments. Overall, RXLR9, RXLR23 and RXLR29 were able to consistently alter plant immune responses to P. capsici infection, suggesting a role for these effector proteins in modulating host immunity.

Waco9 RXLRs can suppress basal resistance against P. syringae

To test whether any of the RXLR proteins are able to suppress defenses that are not oomycete specific, the RXLR-overexpressing A. thaliana lines were inoculated with the virulent bacterial plant pathogen P. syringae pv. tomato DC3000. Disease assays were

![Figure 2. Expression levels of H. arabidopsidis RXLR genes in A. thaliana.](image)

Semi-quantitative analysis of the expression levels of the RXLR transgenes from H. arabidopsidis Waco9 in A. thaliana accession Col-0. Expression levels were assessed in independent transgenic lines using RXLR gene-specific primers. Depicted are ethidium bromide-stained agarose gels with PCR products after 20, 25 or 30 cycles of PCR amplification. The PCR product of the A. thaliana actin gene (At3g18780) was used as internal control (25 cycles of PCR amplification).

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![Figure 3. Effect of ectopic expression of Waco9 RXLR genes on the level of resistance against H. arabidopsidis.](image)

In four experiments, two independent overexpressing lines of each of the 13 Waco9 RXLR genes was tested twice for the level of resistance against H. arabidopsidis Waco9. Two-week-old plants were spray inoculated and 6 days later the number of conidiophores per plant was determined. In each experiment the number of conidiophores on Col-0 is set at 100%. Subsequently, the number of conidiophores in all other lines is given relative to Col-0 in the same experiment. The enhanced susceptible mutant eds1-2 was included as a positive control. Results represent mean ± SEM (n = 18) and asterisks indicate significant differences (ANOVA and Fisher’s LSD corrected for type I errors; p<0.05).

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performed with two independent lines of each of the 13 Waco9 RXLR overexpressors and the level of disease severity was compared to wild-type Col-0 and the enhanced susceptible mutant npr1-1 [44]. In Figure 5, the level of P. syringae disease severity of all RXLR overexpressing lines compared to Col-0 is shown. Seven of the RXLR overexpressors displayed altered resistance against P. syringae at least once, with RXLR6, RXLR16, RXLR19 and RXLR29 showing enhanced susceptibility to P. syringae infection in two experiments, and RXLR9 in three experiments. Overall, these results suggest that the Waco9 RXLR effectors RXLR6, RXLR9, RXLR16, RXLR19 and RXLR29 suppress immune responses to the bacterial pathogen P. syringae.

Effects of Waco9 RXLRs on flg22-induced basal immune responses

In the disease assays described above, we looked at the influence of the Waco9 RXLR proteins on immune responses triggered by different pathogens. In order to investigate the effect of the RXLRs specifically on MTI responses, we monitored reactions of the 13 Waco9 RXLR overexpressors to the well-characterized MAMP flg22, a 22-amino acid derivative of bacterial flagellin. Typically, treatment of A. thaliana seedlings with flg22 activates MTI, resulting in a growth reduction due to reallocation of resources and toxicity of defense related products [63]. The reduction in seedling growth can be used as a measure for MTI response activation [2,64]. RXLR-overexpressing seedlings were grown in liquid MS medium containing 50 or 500 nM of flg22 and...
in each experiment mock-treated and flg22-treated plants of the flagellin receptor mutant fls2 were included as controls. After 10 days of growth, the fresh weight per plant was measured and the relative change in fresh weight in flg22- versus non-treated plants was determined. Figure 6A and 6B show that treatment of Col-0 seedlings with flg22 resulted in a significant decrease in fresh weight in four independent experiments, whereas the fls2 mutant was not responsive to flg22. In order to assess the effect of RXLR overexpression on flg22-mediated growth reduction, the average fresh weights of the RXLR overexpressors were first normalized to that of the untreated plants in the same experiment (set at 100%). Subsequently, the average fresh weights of the flg22-treated RXLR overexpressors were compared to that of flg22-treated Col-0 plants (Figures 6C and 6D). In response to treatment with 50 nM flg22, RXLR3, RXLR21, RXLR23 and RXLR29 overexpressing lines showed a significant difference in growth reduction in only one or two out of four experiments. After treatment with 500 nM flg22, RXLR4 and RXLR29 showed a significant difference in growth reduction in one out of four experiments. Surprisingly, in all statistically significant cases, the RXLR-overexpressing lines produced stronger rather than weaker flg22-induced growth inhibition compared to Col-0. Figures 6E and 6F show the averages of the four different experiments from Figures 6C and 6D, respectively. Notably, the pattern of the fresh weight averages in flg22-treated plants of each RXLR overexpressor are highly similar between plants treated with 50 nM or 500 nM flg22 and tend to be lower in the RXLR overexpressing lines than in wild-type Col-0. Taken together, these results suggest that several RXLR-overexpressing plants are slightly more sensitive to flg22-induced MTI, resulting in enhanced levels of growth reduction in flg22-treated seedlings.

Multifactorial analysis of Waco9 RXLR-overexpressing plants

The above-described results show that a number of Waco9 RXLR proteins, when constitutively expressed in Col-0, have relatively mild effects on different components of the plant immune system. To obtain a clearer picture of the relative importance of the effects of each of the tested RXLRs in the combined dataset, we performed a redundancy analysis (RDA) followed by a hierarchical clustering of the combined data of all the experiments performed with the RXLR overexpressors. To this end, data from all the experiments were transformed to a value between −1 and 1, with −1 being the minimum value, 1 the maximum value and 0 the score obtained for Col-0 for each treatment. Then, a heatmap of all the results from the different experiments was produced and the RXLR overexpressors were clustered using hierarchical clustering (Figure 7A). This clustering led to identification of five distinct groups of RXLR proteins (p<0.05; Figure 7A). Only A. thaliana constitutively expressing RXLR20 cluster together with Col-0 and thus RXLR20 does not appear to influence the tested defense responses. All other RXLRs cluster with one or three other RXLRs. The ordination biplot generated by RDA confirms the hierarchical clustering (Figure 7B). Eigenvectors derived from the RDA (Figure 7C) indicate that 45% of the variation is explained by RDA1 with growth reduction after treatment with 50 nM flg22 as a positive contributor, and H. arabidopsidis infection as the major negative contributor. RDA2 explains another 41% of the phenotypic variation, with P. capsici infection as the main contributor. Taken together, the combined results of the five different treatments show that all tested Waco9 RXLRs, except for RXLR20, have an effect on one or more components of the plant immune system.

RXLR9 suppresses callose deposition

To further test the above observed RXLR trends independently of constitutive in planta expression, we used the bacterial effector detector vector (EDV) system. This system has been used successfully to deliver the H. arabidopsidis RXLR effector ATR13 into A. thaliana leaf cells [39]. It is based on the fusion of a candidate effector protein to the N-terminus of the type III secreted bacterial effector, AvrRps4, allowing delivery of the effector into A. thaliana leaf cells by bacteria such as P. syringae pv. tomatum DC3000 mutant ΔCEL, which strongly triggers MTI [54,65]. Immune suppressive effects of the delivered putative effector can be tested by quantifying its effect on P. syringae ΔCEL-triggered callose deposition [39]. Waco9 RXLR29 has previously been shown to suppress pathogen-induced callose deposition [39]. Here, a different RXLR protein, RXLR9, was selected for analysis. RXLR9 was cloned in the EDV effector delivery system and expressed in the P. syringae ΔCEL mutant strain and delivered to A. thaliana by pressure infiltrating P. syringae ΔCEL (RXLR9) into leaves of Col-0 plants. As a negative control the same EDV system was used to deliver the YFP protein into plant cells, and EDV-ATR13 was used as a positive control [39,59]. At 12 h after pathogen infiltration, the immune-suppressive effect of RXLR9 was evaluated by quantifying callose deposition in the infiltrated leaf tissue. Infiltration of Col-0 leaves with P. syringae ΔCEL (YFP) resulted in strong callose deposition at the site of tissue infiltration (Figure 8), confirming previous findings that P. syringae ΔCEL triggers a strong MTI response [39,54,65]. Also, infiltration of Col-0 with bacteria delivering ATR13 reduced the number of callose deposition sites, confirming the immune suppressive effect of ATR13 [59]. Infiltration with P. syringae ΔCEL delivering RXLR9 led to a reduction in callose production that was similar to that of ATR13. We therefore concluded that RXLR9 is able to suppress MTI.

Discussion

When the genome sequences of different phytopathogenic oomycetes were unraveled, motif searches revealed that these pathogens possess large repertoires of putative effector genes. Different classes of effector candidate proteins have been identified, such as apoplastic effectors and host-translocated Crinklers and RXLR-effectors [23]. The last effector group is represented by proteins containing the RXLR motif in their amino acid sequence, which can be easily retrieved from the genome sequence [16–18,66]. H. arabidopsidis strain Emoy2 has 134 predicted RXLR effectors, but for most of these information on expression and function is lacking. Cabral et al. [39] identified 18 RXLR proteins of the H. arabidopsidis strain Waco9 that are expressed during infection of A. thaliana. Here we describe the screening of these putative effectors for their impact on different components of the plant immune system.

Conserved N-termini of RXLR proteins

Based on amino acid sequences, the set of 18 Waco9 RXLR proteins used in this study can be divided in several groups. Closer scrutiny of these groups revealed that the N-termini of the different RXLR proteins within one group are highly similar, while in most cases the C-termini show little similarity (Figure 1A). The set of 134 RXLRs identified in the H. arabidopsidis Emoy2 genome [18] display similar clustering, based on amino acid sequence similarity in the N-terminal protein domains (Figure 1B). Current evidence suggests that the C-termini of RXLR proteins is important for their effector functions, while the N-terminus is involved in protein translocation [28,29]. Many effector proteins are under selective
Figure 6. Effects of Waco9 RXLRs on flg22-induced reduction of seedling growth. In four independent experiments seedlings of Col-0, the flagellin receptor mutant fls2, and two independent lines of each of the 13 Waco9 RXLR overexpressors were grown on liquid MS medium in the presence or absence of 50 nM (A), (C) and (E) or 500 nM (B), (D) and (F) of flg22. After 10 days of growth, the fresh weight (FW) of a pool of 10 plants per line was determined. In (A) and (B) the relative FWs of Col-0 and fls2 are depicted for the 4 independent experiments, in which the FW of the untreated plants was set at 100%. Results represent mean ± SEM (n = 3) and asterisks indicate significant differences between treated and non-treated plants (Students t-test; p < 0.05). In (C) and (D) the relative FWs of the flg22-treated RXLR overexpressors are depicted for the 4 experiments. These relative FWs are normalized to the relative FW of untreated Col-0 (set at 100%). The dotted line shows the average relative FW of flg22-treated Col-0. Results represent mean ± SEM (n = 3) and asterisks indicate significant differences in relative FW compared to Col-0 (ANOVA and Fisher's LSD corrected for type I errors; p < 0.05). In (E) and (F) the averages of the relative FWs of the flg22-treated RXLR overexpressors are depicted (i.e. the averages of the results in (C) and (D)), again the dotted line represents the average relative FW of flg22-treated Col-0.

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pressure to evade detection by host immune receptors. During effector production and secretion in the *Plasmodium* parasite, the RXLR-related PEXEL motif is cleaved behind the leucine residue after which the C-terminal part of the protein is translocated into erythrocytes [67]. If the translocation system of *H. arabidopsidis* RXLR proteins works in a similar manner, the N-terminal part of the protein is not under diversifying selection driven by the host immune system. This might explain why RXLR proteins exhibit a relatively high level of amino acid conservation in the N-terminal part of the protein, while the C-terminal part following the RXLR motif shows relatively low level of sequence conservation.

**RXLR effectors modulate host defense responses**

Transgenic plants expressing single RXLR genes did not show strongly enhanced susceptibility when infected with *H. arabidopsidis* (Figure 3). It is likely that the production of RXLRs by *H. arabidopsidis*, which include those that are ectopically expressed, is sufficient to render additional expression of particular effectors largely ineffective. The only Waco9 RXLR gene in our screen that
has a homolog in *Phytophthora* species is RXLR21 (Table 1). Since this RXLR gene is present in *Phytophthora* as well as *Hyaloperonospora* species, it is likely that *P. capsici* also possesses an RXLR21 ortholog. Overexpression of RXLR21 in *A. thaliana* did not influence the level of *P. capsici* disease. Whether this is due to production of a related RXLR protein by the pathogen itself remains unclear. However, overexpression of certain Waco9 RXLR proteins did affect the level of resistance against *P. capsici*. For example, overexpression of RXLR9, RXLR23 and RXLR29 reduced symptom development caused by *P. capsici* infection compared to wild-type Col-0 plants (Figure 4, Table 1). This is unexpected because RXLRs normally suppress host immune responses, resulting in enhanced symptom development. This paradox might be explained by the lifestyle of *P. capsici*, which consists of an initial biotrophic phase followed by a switch to necrotrophy. Disease was scored by determining the number of necrotic lesions on each plant at day 6 after inoculation and thus reflects the necrotrophic phase of the *P. capsici* infection. *H. arabidopsidis* is an obligate biotroph and suppressing host cell death is likely important for its survival. The RXLRs used in this study might therefore contribute to cell death suppression, which in the bioassays with the hemi-biotroph *P. capsici* would reduce necrotic lesion formation.

To test the influence of the Waco9 RXLRs on basal immune responses, RXLR transgenic lines were inoculated with a virulent bacterial strain of *P. syringae pv. tomato*. Overexpression of five of the 13 Waco9 RXLRs enhanced the susceptibility of *A. thaliana* to *P. syringae* infection in at least two independent experiments. Fabro et al. [36] tested 64 Emoy2 RXLRs and found that a similarly high proportion of the RXLRs altered the level of susceptibility of *A. thaliana* to *P. syringae* infection. Homologs of 4 Waco9 RXLRs were tested in this screening as well (RXLR6, RXLR16, RXLR21 and RXLR23). However, for none of these RXLRs a consistent effect on bacterial growth in Col-0 could be observed. Fabro et al. also found that, of the RXLRs that led to increased growth of *P. syringae*, 77% suppressed *P. syringae* ΔCEL-induced callose deposition. Callose deposition is a typical MTI response, suggesting that many RXLR effectors interfere with basal host immunity. In this study, flg22-induced growth reduction was used to monitor the effects of the different putative effectors on MTI. In some experiments, overexpression of Waco9 RXLRs resulted in an enhanced effect of flg22 treatment on growth reduction (RXLR3 and RXLR23 in the 50 nM treatment, RXLR4 and RXLR29 in the 500 nM treatment). Also, while the growth reduction generally did not differ between the Waco9 RXLR overexpressors and Col-0, a number of lines (RXLR3, RXLR4, RXLR16, RXLR17, RXLR21, RXLR23, RXLR29) displayed a trend towards an enhanced flg22 response (average growth reduction of fls22-treated plants larger than that in Col-0; Figure 6C and 6D). The enhanced flg22-mediated growth reduction is unexpected since repression of host basal immunity should attenuate flg22-mediated seedling growth inhibition. The cause of this puzzling finding remains unresolved, but one explanation might be that suppression of one defense signaling pathway by an RXLR effector leads to a compensatory upregulation of another pathway through cross-talk within the defense network [68]. Although not statistically significant, RXLR9 overexpressing plants consistently displayed a smaller reduction in fresh weight than Col-0 after treatment with 500 nM flg22 (Figure 6D). This is in line with our observations in the *P. syringae* bioassays that RXLR9 overexpressing plants have enhanced susceptibility to *P. syringae* infection (Figure 5) and that *P. syringae* ΔCEL delivering RXLR9 triggers less callose deposition (Figure 8). In contrast, plants expressing RXLR29, which has previously been shown to be able to suppress callose deposition [39], appear to have a stronger growth reduction after flg22 treatment. This might be explained by the fact that the different experiments, although both related to MTI, measure very different responses that are very likely regulated through different pathways.

Multiple weak effectors might act strongly together

A number of RXLR proteins have previously been shown to severely alter host defense responses [36,38]. In our study a number of RXLRs have clearly measurable effects on certain plant immune system outputs (e.g. RXLR9 and RXLR29 significantly affect susceptibility to *P. syringae* and *P. capsici*, respectively) but most RXLR effectors had no significant effect on the tested defense responses. Additionally, in some cases the variation between repeats was as important as the difference between the tested line and the control, making it difficult to draw conclusions based on the separate experiments. However, when data from all
experiments were combined in a multifactorial analysis, significant effects on host immunity were revealed for a relatively high number (12) of the 13 Waco9 RXLRs tested (Figure 7). Five functional effector groups were identified, but these did not correspond to the groups identified based on N-terminal protein sequence relationships. Also, Wang et al. [33] showed that ~75% of 169 putative RXLR effectors of P. sojae influence host immunity, illustrated by their ability to suppress programmed cell death responses induced by the pro-apoptotic protein BAX. In addition, Fabro et al. [36] found that 72% of the tested H. arabidopsidis Emoy2 RXLRs suppress host immunity when delivered by the EDV system, resulting in enhanced growth of P. syringae in A. thaliana. In both studies, the RXLR effectors were identified in the pathogen genomes without prior information on expression patterns for these genes. In our study the Waco9 RXLRs tested were identified in an EST library of the A. thaliana – H. arabidopsidis Waco9 interaction [39] and are therefore likely to play a role in this plant-pathogen interaction. This might explain the relatively high level of RXLR proteins with some effect on host immunity in our screen. That no effects were observed for RXLR2 could mean that we should look at other defense responses of the plant to find an activity. However, it could also suggest that this protein targets other processes of its host, like nutrient transport.

In conclusion, the data presented in this paper suggest that a large number of the Waco9 RXLR genes that are expressed during infection of the host are likely to contribute to pathogen virulence. Further, the data obtained in the independent experiments show that many RXLR proteins have only weak effects on certain components of the plant immune system, which could only be revealed by combining the data of the different experiments. H. arabidopsidis contains 134 genes in its genome encoding potentially secreted proteins with an RXLR domain [18]. If several have minor activities on one or more components of the plant immune system, their concerted action might substantially modulate the host immune response. In this scenario, each RXLR protein with a minor activity might be disposed of or mutated without major loss of pathogenicity, offering a low risk evolution-ary strategy.

Supporting Information
Table S1 Sequences of the primers used in this study. (DOCX)

Author Contributions
Conceived and designed the experiments: MJCP AC BJMR MFS JEP GVDA CMJP. Performed the experiments: MJCP PCAW AC BJMR MFS JB. Analyzed the data: MJCP PCAW AC BJMR MFS JB JEP GVDA CMJP. Contributed reagents/materials/analysis tools: MJCP AC BJMR MFS JEP GVDA CMJP. Contributed to the writing of the manuscript: MJCP AC BJMR MFS JEP GVDA CMJP.

References
1. Felix G, Duran JD, Volko S, Boller T (1999) Plants have a sensitive perception system for the most conserved domain of bacterial flagellin. Plant J 18: 265–276.
2. Bardoo BV, Van der Ent S, Pel MJ, Tommassen J, Pieterse CMJ et al. (2011) Pseudomonas syringae: immune recognition of flagellin in both mammals and plants. PLoS Pathog 7: e1002206.
3. Pel MJ, Pieterse CMJ (2013) Microbial recognition and evasion of host immunity. J Exp Bot 64: 1237–1248.
4. De Jonge R, Van Esse HP, Kombrink A, Shinya T, Desaki Y, et al. (2010) Conserved fungal LysM effector Ecp6 prevents chitin-triggered immunity in plants. Science 329: 953–957.
5. Hogenhout SA, Van der Hoorn RAL, Terzuoli R, Kamoun S (2009) Emerging concepts in effector biology of plant-associated organisms. Mol Plant-Microbe Interact 21: 115–122.
6. Jones JDG, Dong J (2006) The plant immune system. Nature 444: 323–329.
7. Ariztia EV, Andersen RA, Sogin ML (1991) A new phylogeny for chromophyte algae, including Mallomonas papillosa (Synurophyceae) and Tribonema nucleatum (Xantophyceae). J Phycol 27: 428–436.
8. Tyler BM, 2007. Physophaora sojae: root rot pathogen of soybean and model oomycete. Mol Plant Pathol 8: 1–8.
9. Haverkort AF, Boonkamp PM, Hutten R, Jacobsen E, Lotz LAP, et al. (2008) Societal costs of late blight in potato and prospects of durable resistance through cisgenic modification. Potato Res 51: 47–57.
10. Perumal R, Nimmakayala P, Erratamuthu SR, No EG, Reddy UK, et al. (2008) Simple sequence repeat markers useful for sorghum downy mildew (Peronosclerospora sorghi) and related species. BMC Genet 9: 77.
11. Pieterse CMJ, Raverof EP, Davide LC (1991) An in planta induced gene of Physophaora sojae. Physophaora sojae codes for ubiquitin. Plant Mol Biol 17: 799–811.
12. Juddson HS, Dudler R, Pieterse CMJ, Unkles SE, Michelmore RW (1993) Expression and antisense inhibition of transgenes in Physophaora sojae is modulated by choice of promoter and position genes. Gene 133: 63–69.
13. Pieterse CMJ, Bergsen A-C, Folders J, Govers F (1994) Expression of the Phytophthora sojae rppI resistance gene in transgenic tobacco plants. Mol Gen Genet 244: 269–277.
14. Kamoun S (2007) Groovy times: filamentous pathogen effectors revealed. Curr Opin Plant Biol 10: 530–535.
15. Schornack S, Huisman E, Cano LM, Boukhrot TO, Oliva R, et al. (2009) Ten things to know about oomycete effectors. Mol Plant Pathol 10: 795–803.
16. Haas BJ, Kamoun S, Zody MC, Jiang RHY, Handsaker RE, et al. (2009) Genome sequence and analysis of the Irish potato famine pathogen, Phytophthora infestans. Nature 461: 393–398.
17. Tyler BM, Tripathy S, Zhang X, Dehal P, Jiang RHY, et al. (2006) Phytophthora genome sequences uncover evolutionary origins and mechanisms of pathogenesis. Nature 431: 1261–1266.
18. Baxter I, Tripathy S, Ishaque N, Bost N, Culbard A, et al. (2010) Signatures of adaptation to obligate biotrophy in the Phytophthora parasitica genome. Science 330: 1549–1551.
19. Allen R, Binner-Eddy P, Grenvoin-Biggle L, Meitz J, Rehman A, et al. (2004) Host-parasite coevolutionary conflict between Arabidopsis and downy mildew. Science 306: 1957–1960.
20. Shan W, Cao M, Dan L, Tyler B (2004) The Avr1b locus of Phytophthora sojae encodes an elicitor and a regulator required for avirulence on soybean plants carrying resistance gene Rps1b. Mol Plant-Microbe Interact 17: 394–403.
21. Armstrong M, Whisson S, Pritchard L, Bos J, Venter E, et al. (2005) An ancestral oomycete oocyte locus contains late blight avirulence gene Avr3a, encoding a protein that is recognized in the host cytoplasm. Proc Natl Acad Sci U S A 102: 7766–7771.
22. Rehman A, Gordon A, Rose J, Allen R, Armstrong M, et al. (2005) Differential recognition of highly divergent downy mildew avirulence gene alleles by RPP1 resistance genes from two Arabidopsis lines. Plant Cell 17: 1839–1850.
23. Stassen JH, Van den Ackerveld G (2011) How do oomycete effectors interfere with plant life? Curr Opin Plant Biol 14: 407–414.
24. Boukhrot TO, Schornack S, Banfield MJ, Kamoun S (2012) Oomycete effectors, and all that jazz. Curr Opin Plant Biol 15: 483–492.
25. Miller NJ, Bhattacharjee S, van Ooij C, Lelieveld K, Harris T, et al. (2004) A host-targeting signal in virulence proteins reveals a secretome in malarial infection. Science 306: 1934–1937.
26. Martin M, Good R, Rug M, Kneuper E, Cosman A (2004) Targeting malaria virulence and remodeling models to the host erythrocyte. Science 306: 1930–1933.
27. Bhattacharjee S, Miller NJ, Lelieveld K, van Wijk J, Kampergat T, et al. (2006) The malarial host-targeting signal is conserved in the Irish potato fungus pathogen. Plant Pathol 2: 453–463.
28. Whisson SC, Bevinck PC, Moleleki L, Avovoa AO, Morales JG, et al. (2007) A translation signal for delivery of oomycete effector proteins into host plant cells. Nature 450: 115–118.
29. Dou D, Kale SD, Wang X, Jiang RHY, Bruce NA, et al. (2008) RXLR-mediated entry of Phytophthora sojae effector Avr1b into soybean cells does not require pathogen-encoded machinery. Plant Cell 20: 1930–1947.
30. Kale SD, Gu B, Capelluto DGS, Dou D, Feldman E, et al. (2010) External lipid PEM mediates entry of eukaryotic pathogen effectors into plant and animal host cells. PloS Pathog 2: 1–9.
31. Yaeno T, Li H, Chaparro-Garcia A, Schornack S, Koshiba S, et al. (2011) Phosphatidylinositol monophosphate-binding interface in the oomycete RXLR effector AVR5A is required for its stability in host cells to modulate plant immunity. Proc Natl Acad Sci U S A 108: 14682–14687.
32. Pete B, Kamoun S (2014) How do filamentous pathogens deliver effector proteins into plant cells? PLoS Biol 12: e1001801.
33. Wang Q, Han C, Terrahe AO, Yu X, Ye W, et al. (2011) Transcriptional programming and functional interactions within the Phytophthora sojae RXLR effector repertoire. Plant Cell 23: 2046–2089.
34. Oh S, Young C, Lee M, Oliva R, Boukhrot TO, et al. (2009) In planta expression screens of Phytophthora infestans RXLR effectors reveal diverse phenotypes.

PLOS ONE | www.plosone.org 12 November 2014 | Volume 9 | Issue 11 | e110624
including activation of the Solanum bulbocastanum disease resistance protein Rpa-hb2. Plant Cell 21: 2929–2947.

35. Vleeshouwers VGAA, Rietman H, Kernek P, Champounet N, Young C, et al. (2008) Effector genomics accelerates discovery and functional profiling of potato disease resistance and Phytophthora infestans avirulence genes. PLoS ONE 3: e2875.

36. Fabro G, Steinbrenner J, Coates M, Ishuga N, Baxter L, et al. (2011) Multiple candidate effectors from the oomycete pathogen Hyaloperonospora arabidopsidis suppress host plant immunity. PLoS Pathog 7: e1002548.

37. Gorisiching S, Krasleva KV, Dahlbeck D, Staskawicz BJ (2012) Computational prediction and molecular characterization of an oomycete effector and the cognate Arabidopsis resistance gene. PLoS Genet. 8: e1002502.

38. Caillaud MCC, Asai S, Rallapalli G, Piquerez S, Fabro G, et al. (2013) A downy mildew effector attenuates salicylic acid-triggered immunity in Arabidopsis by interacting with the host mediator complex. PLoS Biol 11: e1001732.

39. Cabral A, Staessen JHM, Stassen J, Bautor J, Parker JE, et al. (2011) Identification of Hyaloperonospora arabidopsidis transcript sequences expressed during infection reveals isolate-specific effectors. PLoS ONE 6: e19328.

40. Pavlidis P, Noble WS (2003) Matrix2png: a utility for visualizing matrix data. Bioinformatics 19: 295–296.

41. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ (1990) Basic Local Alignment Search Tool. J Mol Biol 215: 403–410.

42. Enright A, Van Dongen S, Ouzounis C (2002) An efficient algorithm for large-scale detection of protein families. Nucleic Acids Res 30: 1573–1580.

43. Van Dongen S (2000) Graph clustering via a discrete unconvolving process. Siam J Matri Appl Anal 30: 121–141.

44. Cao H, Bowling SA, Gordon AS, Dong X (1994) Characterization of an Arabidopsis mutant that is nonresponsive to inducers of systemic acquired resistance. Plant Cell 6: 1503–1592.

45. Shan LB, He P, Li JM, Heese A, Peck SC, et al. (2008) Bacterial effectors target genes but stimulates the expression of the jasmonate-inducible gene Atvsp upon challenge. Plant Mol Biol 41: 367–373.

46. Fabro G, Steinbrenner J, Coates M, Ishuga N, Baxter L, et al. (2011) A downy mildew effector attenuates salicylic acid-triggered immunity in Arabidopsis by interacting with the host mediator complex. PLoS Biol 11: e1001732.

47. Coates M, Rallapalli G, Piquerez S, Fabro G, et al. (2013) Multiple candidate effectors from the oomycete pathogen Hyaloperonospora arabidopsidis suppress host plant immunity. PLoS Pathog 7: e1002548.

48. Van Wees SCM, Luijendijk M, Smoorenburg I, Van Loon LC, Pieterse CMJ (1996) Arabidopsis is susceptible to infection by a downy mildew fungus. Plant Cell 2: 437–445.

49. Fabro G, Steinbrenner J, Coates M, Ishuga N, Baxter L, et al. (2011) Multiple candidate effectors from the oomycete pathogen Hyaloperonospora arabidopsidis suppress host plant immunity. PLoS Pathog 7: e1002548.

50. Koch E, Slusarenko A (1990) Arabidopsis is susceptible to infection by a downy mildew fungus. Plant Cell 2: 437–445.

51. Koch E, Slusarenko A (1990) Arabidopsis is susceptible to infection by a downy mildew fungus. Plant Cell 2: 437–445.

52. Van Damme M, Zeilmaker E, Elbers E, Andel A, De Saen-van der Velden, et al. (2009) Downy mildew resistance in Arabidopsis by mutation of HOMO- SERINE KINASE. Plant Cell 21: 2179–2189.

53. Wang Y, Boumeester K, Van de Mortel JE, Shan W, Gover F (2013) A novel Arabidopsis-oomycete pathosystem: differential interactions with Phytophthora capsici reveal a role for camalexin, indole glucosinolates and salicylic acid in defense. Plant Cell Environ 36: 1192–1203.

54. Alfano J, Charkowski A, Deng W, Bader J, Petnicki-Oocwieja T, et al. (2000) The Pseudomonas syringae Hop pathogenicity island has a tripartite mosaic structure composed of a cluster of type III secretion genes bounded by exchangeable effector and conserved effector loci that contribute to parasitic fitness and pathogenicity in plants. Proc Natl Acad Sci U S A 97: 4856–4861.

55. Kassey VE, Chiplis G, Gilbert D, Butler A, Toussaint M, et al. (2011) Experimental climate effect on seasonal variability of polyphenol/phenoloxidase interplay along a narrow four-bog ecological gradient in Sphagnum falkhus. Global Change Biol 17: 2945–2957.

56. R Core Team (2012) R: A language and environment for statistical computing. R Foundation for Statistical Computing ISBN 3-900051-07-0: http://www.R-project.org.

57. Oksanen J, Blanchet FG, Kindlir P, Legendre P, Minchin PR, et al. (2012) vegan: community ecology package. R package version 2.0-3: http://CRAN.R-project.org/package=vegan.

58. Suzuki R, Shimodaira H (2011) pvclust: hierarchical clustering with P-values via multiscale bootstrap resampling. R package version 1.2-2: http://CRAN.R-project.org/package=pvclust.

59. Sohn KH, Lei R, Nemri A, Jones JDG (2007) The downy mildew effector proteins ATR1 and ATR13 promote disease susceptibility in Arabidopsis thaliana. Plant Cell 19: 4077–4090.

60. Abramoff MD, Magalhães PJ, Ram SJ (2004) Image processing with ImageJ. Biophotonics International 11: 36–42.

61. Judelson HS, Tyler BM, Michelmore RW (1991) Transformation of the necrotrophic plant pathogen Hyaloperonospora arabidopsidis transcript sequences expressed during infection reveals isolate-specific effectors. PLoS ONE 6: e19328.

62. Judelson HS, Tyler BM, Michelmore RW (1991) Transformation of the necrotrophic plant pathogen Hyaloperonospora arabidopsidis transcript sequences expressed during infection reveals isolate-specific effectors. PLoS ONE 6: e19328.

63. Gomez-Gomez L, Boller T (2002) Flagellin perception: a paradigm for innate immunity. Trends Plant Sci 7: 251–256.

64. Fincham JRS (1989) Transformation in fungi. Microbiol Rev 53: 148–170.

65. Gomez-Gomez L, Boller T (2002) Flagellin perception: a paradigm for innate immunity. Trends Plant Sci 7: 251–256.

66. Fincham JRS (1989) Transformation in fungi. Microbiol Rev 53: 148–170.

67. Chang HH, Falick AM, Carlton PM, Sedat JW, DeRisi JL, et al. (2008) N-Alignment Search Tool. J Mol Biol 215: 403–410.

68. Levesque CA, Brouwer H, Cano L, Hamilton JP, Holt C, et al. (2010) Genome project. Parasitol 160: 107–115.

69. Levesque CA, Brouwer H, Cano L, Hamilton JP, Holt C, et al. (2010) Genome project. Parasitol 160: 107–115.

70. Levesque CA, Brouwer H, Cano L, Hamilton JP, Holt C, et al. (2010) Genome project. Parasitol 160: 107–115.