Stay-green traits to improve wheat adaptation in well-watered and water-limited environments

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Abstract

A stay-green phenotype enables crops to retain green leaves longer after anthesis compared with senescent types, potentially improving yield. Measuring the normalized difference vegetative index (NDVI) during the whole senescence period allows quantification of component stay-green traits contributing to a stay-green phenotype. These objective and standardized traits can be compared across genotypes and environments. Traits examined include maximum NDVI near anthesis (Nmax), senescence rate (SR), a trait integrating senescence (SGint), plus time from anthesis to onset (OnS), mid-point (MidS), and near completion (EndS) of senescence. The correlation between stay-green traits and yield was studied in eight contrasting environments ranging from well watered to severely water limited. Environments were each classified into one of the four major drought environment types (ETs) previously identified for the Australian wheat cropping system. SGint, OnS, and MidS tended to have higher values in higher yielding environments for a given genotype, as well as for higher yielding genotypes within a given environment. Correlation between specific stay-green traits and yield varied with ET. In the studied population, SGint, OnS, and MidS strongly correlated with yield in three of the four ETs which included well-watered environments (0.43–0.86), but less so in environments with only moderate water-stress after anthesis (−0.03 to 0.31). In contrast, Nmax was most highly correlated with yield under moderate post-anthesis water stress (0.31–0.43). Selection for particular stay-green traits, combinations of traits, and/or molecular markers associated with the traits could enhance genetic progress toward stay-green wheats with higher, more stable yield in both well-watered and water-limited conditions.

Key-words: Crop adaptation, crop improvement, drought, genotype×environment interaction, leaf senescence, phenotyping, stay-green, water limitation, wheat.

Introduction

Developing cultivars with superior adaptation to water-limited environments has been impeded by complex interactions between genotype and environment (G×E), leading to changes in the yield rankings of genotypes in different water-limited environments (Cooper et al., 2001; Richards et al., 2002). To improve the rate of yield gain in the face of...
G×E, researchers have sought physiological and morphological traits linked with high yield in the target population of environments (TPE) that are less susceptible to environmental influences than yield per se (Jackson et al., 1996; Hammer et al., 2002, 2005; Tardieu 2003; Casadebaig et al., 2016). The stay-green phenotype has been linked to improved yield stability in a number of cereal crop species including wheat and sorghum, particularly under terminal drought stress (recently reviewed in Gregersen et al., 2013). Plants exhibiting the stay-green phenotype are able to maintain green leaf area for longer after anthesis than senescent lines, allowing maintenance of photosynthesis for longer during the grain-filling period (Thomas and Smart, 1993; Thomas and Howarth, 2000). Thus, selection for stay-green has been targeted to improve crop adaptation to water-stressed environments in a number of crops including sorghum and wheat (Christopher et al., 2008, 2014; Borrell et al., 2012, 2014a, b; Jordan et al., 2012; Lopes and Reynolds, 2012; Gregersen et al., 2013).

The stay-green phenotype has long been recognized as having potential for crop improvement (Thomas and Smart, 1993). This phenotype can be either functional, where photosynthesis and accumulation of assimilates to harvested tissues are prolonged, or non-functional, where plants appear green but there is no benefit in terms of yield (e.g. due to a lesion in the chlorophyll recycling process, or disrupted transfer of nitrogen from leaf to grain). Only functional stay-green is of interest for crop improvement. Functional stay-green can be achieved by varying leaf-greenness dynamics in a number of different ways (Thomas and Howarth, 2000). Plants may be greener around anthesis before the onset of senescence, commence senescence later, or senesce more slowly (Thomas and Howarth, 2000; Harris et al., 2007; Christopher et al., 2014).

Stay-green has previously been assessed in the field using various techniques. Rapid evaluation by visual assessments have been performed by rating whole-plant senescence (Rosenow et al., 1983; Henzell et al., 1992; Jordan et al., 2012), or assessing green leaf number per culm (Haussman et al., 1999), the greenness of all fertile shoots (Foulkes et al., 2007), or greenness of the flag leaf and peduncle (Joshi et al., 2007). More objective measures of greenness have been taken for individual leaves with the Minolta SPAD meter (Borrell et al., 1996; Harris et al., 2007; Christopher et al., 2008) and, more recently, the canopy with normalized difference vegetative index (NDVI)-based methods (e.g. Lopez and Reynolds, 2012; Christopher et al., 2014). Those methods commonly relied on measurements from one or a few time points late in the crop cycle, with linear regressions fitted to model senescence dynamics (e.g. Harris et al., 2007; Lopes and Reynolds, 2012). However, traits such as onset of leaf senescence vary between genotypes, and the date of measurements can impact the results (Christopher et al., 2014). In addition, the dynamics of senescence appear to follow a non-linear pattern (Borrell et al., 2000a; Christopher et al., 2008, 2014; Vijayalakshmi et al., 2010).

A method has recently been proposed to assess quantitative, component stay-green traits in field trials (Christopher et al., 2014). The dynamics of canopy senescence were shown to fit closely a logistic model fitted to periodic NDVI measured using the NTech Greenseeker®. This information was then used to estimate a number of component stay-green traits including the timing from anthesis to (i) senescence onset (OnS); (ii) mid-senescence (MidS); and (iii) near completed senescence (EndS); as well as (iv) the initial NDVI level (near anthesis; Nmax); (v) an indicator of the senescence rate (SR); and (vi) a parameter derived by integrating NDVI over the senescence period that provides a measure somewhat analogous to green leaf area duration (SGint; Table 1). This approach provides a more detailed understanding of genotypic variation in stay-green phenotype by examining contributing traits during the whole senescence period for each genotype. Traits are estimated based on objective NDVI measurements which are fitted to a logistical model standardized to thermal time with respect to anthesis for each genotype. This enables genotypes to be compared both within and across environments. The method also allows hundreds of genotypes to be characterized in multiple environments, allowing investigations of G×E and characterization of populations for genetic studies.

Recent advances have also occurred in our understanding of the patterns of seasonal water deficit in the target population of environments (TPE) of Australian rain-fed wheat crops (Chenu et al., 2011, 2013). Changes in the timing of water deficit with respect to crop development lead to different impacts of water-stress by affecting different processes. Chenu et al. (2013) determined that seasonal water-stress patterns encountered by Australian wheat crops can be classified into four main environment types (ETs), dependent upon the

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**Table 1. Abbreviations and descriptions of stay-green traits**

| Abbreviation | Stay-green trait | Description |
|--------------|-----------------|-------------|
| Nmax         | Maximum leaf greenness | Maximum NDVI value usually near anthesis |
| OnS<sup>a</sup> | Onset of leaf senescence | Thermal time from anthesis to 90% of N<sub>green max</sub> <sup>a</sup> |
| MidS<sup>a</sup> | Mid-point of leaf senescence | Thermal time from anthesis to 50% of N<sub>green max</sub> <sup>a</sup> |
| EndS<sup>a</sup> | Near completion of leaf senescence | Thermal time from anthesis to 10% of N<sub>green max</sub> <sup>a</sup> |
| SR           | Indicator of the rate of senescence | Indicator of the rate of NDVI decrease at MidS |
| SGint        | Stay-green integral (senescence integral) | Cumulative NDVI from anthesis to after senescence completion at 1500 °Cd after anthesis. |

<sup>a</sup> N<sub>green max</sub> is the difference between the Nmax and the final NDVI value at 1500 °Cd.

<sup>b</sup> OnS, MidS, and EndS have previously been labelled TFN90, TFN50, and TFN10, respectively, in Christopher et al. (2014)
tuning and severity of water-stress from least stressed ET1 to the most stressed ET4. Knowledge of the classification of environments encountered in multi-environment field trials can aid breeders, in particular, by providing information about whether specific trials are more or less representative of the environments encountered in the TPE (Chenu et al., 2011; Chenu, 2015). Thus, breeders could potentially ‘weight’ trial results according to the relevance of the particular ET in the TPE. They can also aid researchers to choose and/or manipulate environmental conditions to generate environments that will be representative of the TPE; and to interpret the value of traits, including stay-green traits, across environments (Rebetzke et al., 2013; Chenu, 2015).

The aim of the current study was to determine the potential of the recently proposed component stay-green traits for crop improvement. Can stay-green traits be useful to select for high-yielding genotypes in a broad range of water-stress environments or, conversely, for adaptation to specific classes of water-stress environments? What is the magnitude of the effect of individual stay-green traits in different environments? A mapping population of doubled-haploid lines segregating for stay-green traits was studied over 3 years in a range of environments as well as soil N and P levels. Nematode densities were below known damage thresholds for sensitive wheat cultivars. Non-limiting levels of nutrients were applied using 120 kg ha−1 urea prior to sowing and 40 kg ha−1 of Starter Z+ containing 10.5% N, 19.5% P, 2.2% S, and 2.2% Zn at sowing. Weeds and diseases were controlled as necessary.

Materials and methods

Plant material

To reduce variation for height and maturity date, 184 lines were selected from a much larger doubled-haploid population derived from the bread wheat (Triticum aestivum) cultivars SeriM82 and Hartog (Christopher et al., 2013, 2014). The parental lines contrast for yield and stay-green. SeriM82 is a high-yielding drought-tolerant, stay-green line (Sivapalan et al., 2000, 2001; Olivares-Villegas et al., 2007), while Hartog is a senescent cultivar adapted to subtropical Australia (Manschadi et al., 2006, 2010; Christopher et al., 2008).

Field trials

Field trials were established during three seasons (2010, 2011, and 2012) at three sites in southern Queensland, Australia: Gatton (GAT; 27.54°S, 152.34°E, 89 m a.s.l.) in 2010, and Kingsthorpe (KTP; 27.51°S, 151.78°E; 442 m a.s.l.) and Warwick (WAR; 28.2°S, 152.0°E; 480 m a.s.l.) in 2011 and 2012. Crops were grown either in natural rain-fed conditions (‘rf’ treatment), in irrigated conditions (‘ir’) where irrigation was used to control water-stress throughout the season, or under a rain-out shelter (‘ro’) which excluded rainfall from anthesis onwards. Some rain-fed treatments were also watered to near field capacity immediately after sowing to ensure uniform establishment. Experiment names are derived from a combination of location, year, and treatment such that the rain-fed trial at Warwick in 2011 is designated ‘WAR11rf’ (Table 2). Heavy, alkaline cracking clay soils with high water-holding capacity predominated at all sites.

Crops were sown in 2 m×6 m plots with a row spacing of 25 cm and a target population density of 100 plants m−2. Soil tests were performed prior to planting to estimate parasitic nematode densities as well as soil N and P levels. Nematode densities were below known damage thresholds for sensitive wheat cultivars. Non-limiting levels of nutrients were applied using 120 kg ha−1 urea prior to sowing and 40 kg ha−1 of Starter Z+ containing 10.5% N, 19.5% P, 2.2% S, and 2.2% Zn at sowing. Weeds and diseases were controlled as necessary.

Trial design and statistical analysis

All trials were designed as partially replicated row–column experiments (GAT10rf 39%, GAT10ir 38%, WAR11rf 38%, GAT11rf 37%, KTP11rf 42%, WAR12rf 32%, GAT12rf 36%, and WAR12ro 21%; Cullis et al., 2006). The designs included an underlying component for autocorrelation in the column and row direction, and entries were latinized along rows and columns. All designs were generated using ‘immdesign’ (Butler et al., 2008). Up to 184 SeriM82×Hartog double-haploids plus the parents were tested in each trial (Table 2).

The statistical analysis comprised a set of individual analyses and a set of bivariate analyses. Individual analyses were performed for each stay-green trait and yield in each trial. A linear mixed model was fitted to each separate data set. Each model included a random term for replicate blocks, a separable autoregressive structure for both rows and columns at the residual level, and terms to account

### Table 2. Trial identifier (Trial ID), sowing date, days from sowing to anthesis for the reference parent Hartog (DTA), days from sowing to maturity of Hartog (DTM), plant-available soil water capacity (PAWC, mm), plant-available soil water at sowing (PAW, mm), irrigation at sowing (Irri init, mm), irrigation immediately prior to anthesis (Irri anth, mm), cumulative in-crop rainfall (ICR, mm), water potentially available (WPA), average daily maximum temperature from sowing to maturity (Avg Temp, °C), cumulative radiation from sowing to maturity (Cum Radn, MJ m−2), number of genotypes tested (No. genos), environment mean yield (Yld, g m−2), and environment type as depicted in Fig. 1 (ET)

| Trial ID | Sowing date | DTA | DTM | PAWC | PAW | Irri init | Irri anth | ICR | WPAa | Avg Temp | Cum Radn | No. genos | Yld | ET |
|----------|-------------|-----|-----|------|-----|-----------|-----------|-----|------|----------|----------|-----------|-----|----|
| GAT10ir  | 26 May      | 94  | 134 | 285  | 185 | 25        | 42        | 216 | 468  | 15.7     | 1808     | 143       | 464 | ET1|
| GAT10rf  | 26 May      | 89  | 130 | 285  | 185 | 25        | –         | 214 | 424  | 15.6     | 1741     | 101       | 498 | ET1|
| WAR11rf  | 24 June     | 98  | 141 | 258  | 258 | 25        | –         | 187 | 445  | 13.2     | 2447     | 183       | 567 | ET1|
| GAT11rf  | 9 June      | 89  | 131 | 312  | 312 | 25        | –         | 157 | 494  | 15.0     | 2157     | 151       | 659 | ET2|
| KTP11rf  | 10 June     | 107 | 149 | 290  | 290 | –         | –         | 188 | 478  | 13.2     | 2543     | 182       | 579 | ET2|
| GAT12rf  | 10 July     | 79  | 115 | 285  | 284 | –         | –         | 95  | 379  | 16.8     | 2202     | 191       | 442 | ET3|
| WAR12rf  | 22 June     | 105 | 144 | 295  | 255 | –         | –         | 109 | 364  | 13.8     | 2649     | 189       | 415 | ET3|
| WAR12ro  | 22 June     | 96  | 136 | 216  | 176 | –         | –         | 0   | 176  | 13.5     | 2485     | 76        | 208 | ET4|

a Trial identifiers indicate the sites as Gatton (GAT), Kingsthorpe (KTP), or Warwick (WAR); the year from 2010 to 2012; and the treatment as irrigated (ir), rain-fed (rf), or rain-out shelter (ro).

b Water potentially available (WPA)=PAW+Irri init+Irri anth+ICR.
for spatial field trend following the method of Gilmour et al. (1999). Genotypes were fitted as random to estimate the genetic effects of each trait within each trial. Best linear unbiased predictors (BLUPs) were calculated from each separate analysis. The BLUPs from each individual trial analysis of each trait were applied to a principal components analysis (PCA) using the ‘princomp’ function in the R software package ‘stats’ (R Development Core Team, 2013). To standardize the data across traits, the PCA was applied to a correlation matrix of the traits, implemented through the ‘cor’ command in princomp. This analysis projects the data onto a reduced set of components, where the first two components explain the majority of the variability in the data. The results were summarized as a biplot where scores for each genotype and the loading for each trait are plotted for these first two components (Gabriel, 1971).

A series of bivariate linear mixed models were performed to estimate the genetic variance of the stay-green trait, the genetic variance of yield, and the correlation between the two at each trial, as previously described in Christopher et al. (2014). Each model included the relevant random and spatial terms established in the individual trial analyses. The slope of the genetic regression was calculated between yield and each trait using the estimated genetic variances and correlation. An approximate standard error for each slope was calculated using Taylor expansion. The genetic correlations provide a predictive quantity of the potential for the rate of change in yield based on the unit change in the trait within the range of the data.

Individual and bivariate analyses were performed in ASReml-R (Butler et al., 2009) using R software. Best linear unbiased estimates (BLUEs) were also calculated.

Characterization of the water-stress environments

Crops in each trial were simulated with the computer crop simulation model, the Agricultural Production Systems sIMulatoR APSIM-wheat v7.0 (Holzworth et al., 2014), using climatic and soil data collected at each site. The water deficit patterns experienced by the parent cultivar Hartog were determined for each trial based on the water-stress index computed by APSIM. The water-stress index corresponds to a water supply/demand ratio that integrates the crop demand and the water available to the roots. A ratio of 1 indicates no water-stress, while a ratio of 0 corresponds to a full stress, with no water available to the crop (Chenu et al., 2013). To ensure best estimation of the stress pattern for each trial, simulated anthesis dates from APSIM were adjusted to concur with the observed date of anthesis, by adapting the thermal time duration between emergence and floral initiation. Simulations of yield closely or slightly underpredicted grain yield (Supplementary Fig. S1 at JXB online), suggesting that transpiration patterns and water-deficit responses were well approximated. Data from all trials were centred at anthesis, and averaged every 100 degree Celsius days (°Cd) between emergence and 450 °Cd after anthesis. The trials were classified according to their similarity to the four previously identified main ETs from the Australian TPE (Fig. 1; Chenu et al., 2011, 2013).

The four ETs are defined as ET1–ET4, which rank roughly in ascending order of water-stress such that ET1 represents environments where crops experience moderate to severe water-stress of short duration mainly after anthesis, ET3 where crops mainly experience moderate to severe water-stress from the lead up to anthesis and the early grain-filling period, and ET4 where severe water-stress leading up to anthesis is not relieved (Chenu et al., 2013).

Crop measurements

Emergence counts were taken to ensure plots were well established. For each plot, Zadoks stages were recorded weekly to determine anthesis date (Zadoks code 65, Z65; Zadoks et al., 1974). NDVI was measured weekly for each plot starting from awn emergence (Z49) until after maturity using a hand-held Greenseeker model 505.
Stay-green traits (Table 1) were estimated from a logistic function fitted to NDVI data centred at anthesis for each plot (for more details, see Christopher et al., 2014).

Results

Each of the four major water-stress environment types in the TPE were represented in the multi-environment trials

Using a combination of natural environmental variation and applied treatments, each of the major water-stress ETs described for wheat crops in the Australian TPE were sampled (Table 2; Fig. 1; Chenu et al., 2013).

All sites had a significant amount of plant-available water (PAW) stored in the soil prior to sowing (Table 2). In-crop rainfall varied from 0 mm at W AR12ro to 216 mm at GAT10ir. The environments produced a wide range of yields, with trial mean yields for all genotypes ranging from 208 g m$^{-2}$ in the rain-out shelter at WAR12ro to 659 g m$^{-2}$ at GAT11rf (Table 2).

The seasonal water-stress patterns for each of the eight environments were classified into one of the four water-stress ETs previously identified (Table 2; Fig. 1). GAT10ir, GAT10rf, and WAR11rf only exhibited minor water deficits throughout the season, and so were classified in ET1 (Fig. 1). For crops at GAT11rf and KTP11rf, water deficit built up to reach a maximum near or following anthesis, but this stress was relieved by rainfall soon thereafter, allowing grain filling to finish with little water-stress, corresponding to ET2 (Fig. 1). Greater stress affected crops at GAT12rf and WAR12rf, where water deficit became moderate to severe well before anthesis and continued well into the grain-filling period (ET3; Fig. 1). The most severe water-stress was observed at WAR12ro, where a severe water deficit from before anthesis continued through to maturity (ET4; Fig. 1).

Yield tended to decrease with increasing water-stress from ET1 to ET4

As anticipated, water availability was a major yield constraint in the studied environments. There was a general trend to decreased yield as the water-stress pattern became more severe from the least stressed ET1 through to the most stressed ET4 (Table 2; Fig. 2). Although environment mean grain yield of the eight environments was not well correlated with in-crop rainfall (Table 2), it was strongly, positively correlated with the amount of potentially available moisture, which was estimated as PAW at sowing plus in-crop rainfall and irrigation, particularly in 2011–2012 trials (Table 2). Other factors probably influenced yield in certain environments. For example, environments at Gatton 2010 had the highest in-crop rainfall and potentially available water, but yield was lower than environments in 2011 (Table 2). The high in-crop rainfall at Gatton 2010 was accompanied by increased cloud cover, which decreased incident radiation. Cumulative radiation for environments at GAT10ir (1808 MJ m$^{-2}$) and GAT10rf (1741 MJ m$^{-2}$) were well below those for other environments, which ranged from 2157 MJ m$^{-2}$ to 2649 MJ m$^{-2}$ (Table 2), particularly during the grain-filling period (Supplementary Fig. S2b).

In addition, crops in some environments with a weekly mean of the daily maximum temperatures reaching or exceeding 30 °C probably experienced heat stress around anthesis and during the grain-filling period (Supplementary Fig. S2). Prolonged temperatures above 30 °C pre- and post-anthesis are known to affect wheat productivity (e.g. Tashiro and Wardlow, 1990; Reynolds et al., 2001). Heat stress could potentially have affected yield in all environments classified in ET3 and ET4, as well as GAT11rf in ET2 (Supplementary Fig. S2, solid lines). Environments in ET1 as well as KTP11rf in ET2 did not reach these temperatures during the early- or mid-grain-filling period, or reached them only late in development when starch deposition would have been nearly complete (Supplementary Fig. S2). Thus, heat stress is less likely to have affected yield in these environments.

Yield rankings for genotypes varied between environments, indicating crossover G×E. Genetic correlations between pairs of environments varied from −0.45 to +0.87, and the mean correlation for all eight environments was +0.3. In the overall analysis, the variance component due to G×E was 1.6 times greater than that for genotype. These results are not uncommon for multi-environment trials in the Australian TPE and highlight the need to seek adaptation indicators with higher heritability and lower G×E than yield per se.

Stay-green traits were strongly affected by changes in water-stress between environments

Variation between genotypes for yield tended to decrease with increased water-stress and lower yield from ET1 to ET4 (Fig. 2a). The trend towards lower yield with increasing water-stress was accompanied by a marked shortening of the period from anthesis to the onset of senescence (OnS), mid-point of senescence (MidS), and near completion of senescence (EndS; Fig. 2f–h). Smaller relative decreases from ET1 to ET4 were also observed for the indicator of the maximum senescence rate (SR; except for the two light-constrained environments of GAT10), the maximum NDVI near anthesis (Nmax), and the stay-green integral (SGint).

Water-stress tended to shorten the thermal time period from sowing to anthesis, but only in some environments (Fig. 2b). There was a clear trend towards shorter plants in more water-stressed environments, ranging from an environment mean of 104 cm at GAT10ir in ET1 to 70 cm at WAR12ro in ET4 (Fig. 2c).

Higher values of Nmax, OnS, MidS, EndS, and SGint are correlated with higher yielding environments

There is a clear positive correlation between environment mean yield of the standard genotype Hartog and higher
Christopher et al. environment mean values for Nmax, OnS, MidS, EndS, SR, and SGint (Fig. 3), with less water-stressed environments generally having higher yield and higher stay-green trait values in the order ET1–ET2> ET3> ET4 (Figs 2, 3). The slope of these regression lines provides an estimate of the apparent effect of stay-green traits on the yield of Hartog across environments. For example, yield increases by 0.73 g m$^{-2}$ for each °Cd increase in OnS (Fig. 3b). If we consider a typical day (18 °Cd) during the early grain-filling period in the TPE, this corresponds to an ~13 g m$^{-2}$ increase for a 1 d delay in senescence onset, representing a 3.2% yield increase for a trial with a mean yield of 400 g m$^{-2}$. Change in MidS had an impact of a similar order of magnitude on yield, and EndS slightly higher (Fig. 3b). Thus, delays of only a few days in OnS, MidS, and EndS in the different environments were correlated with considerable increases in the yield of Hartog. Substantial impacts
of Nmax, SR, and SGint were also observed (Fig. 3a, c, d), but it is important to remember that these stay-green traits can be closely correlated to each other. Somewhat counter-intuitively, the positive correlation between SR and yield in several environments indicates that the higher yield is correlated with a faster rate, as discussed below.

Genetic correlations between stay-green traits and yield varied with the water-stress environment type

High correlations between stay-green traits and yield were observed across genotypes within most trials (Fig. 4). The degree of genetic correlation varied with the water-stress ET. The timings of senescence onset (OnS) and of the mid-point of senescence (MidS) were significantly correlated with yield in ET1, ET3, and ET4 (P≤0.05 or ≤0.01) but not in ET2 (P>0.05; Fig. 4). The SGint was also significantly correlated with yield in ET1, in one ET2 (GAT11rf), and in ET3 (P≤0.05). In ET4, the correlation between SGint and yield was close (0.73), but not statistically significant (P>0.05). The lower number of genotypes tested in ET4 (76) and reduced yield variation due to severe stress (Fig. 2a) may have reduced the likelihood of detecting a significant correlation in this environment. Finally, the genetic correlation between mean estimated EndS and yield was significant for only two environments, GAT10rf and GAT12rf (0.42 and 0.39, respectively; P≤0.05; not shown).

In contrast to OnS, MidS, and SGint, correlations between yield and SR or Nmax were more varied across water-stress environments. The maximum leaf canopy greenness (Nmax) was correlated with yield for both environments classified as ET2 and one in ET3 (WAR12rf). The SR was significantly correlated with yield in both ET3 environments, but in other ETs only at WAR11rf (ET1). Height and time to anthesis were not significantly correlated with yield in most environments, probably reflecting the previous selection of the population to reduce variation for these traits (Christopher et al., 2013). The range of BLUPs for anthesis dates among the doubled-haploids was from 4.0 d at GAT12rf to 4.6 d at GAT10rf, 4.9 d at HRS12rf, 7.1 d at GAT11rf, 7.3 d at KTP1rf, 8.0 d at GAT10ir, and 10.7 d at WAR11rf. However, yield was significantly positively correlated with time to anthesis at one of the radiation-limited environments (GAT10rf, P<0.05) and significantly negatively correlated with height at the other radiation-limited environment (GAT10ir, P<0.05) and at one ET2 environment (KTP11rf, P<0.001). Including these traits as covariates in the statistical model for the yield prediction (BLUPs) did not significantly affect the results for any environment, so the analysis without covariates was used.

No clear pattern in the genetic correlations between stay-green traits and yield was observed in sites affected by heat stress (i.e. all ET3 and ET4 environments, as well as GAT11rf in ET2) compared with others (Fig. 4; Supplementary Fig. S2). Similarly, there was no clear difference in the pattern...
of genetic correlations for the potentially radiation-limited environments at GAT10ir and GAT10rf compared with the other likely non-radiation-limited environments (Fig. 4; Supplementary Fig. S2).

Greater genotype mean values for Nmax, OnS, MidS, EndS, and SGint are correlated with higher yield within environments.

The ability to retain green leaf area had significant effects on yield of individual genotypes which varied with environment type. The slope of regression lines used to estimate the correlations between stay-green traits and yield at each site (Fig. 4) give an indication of the magnitude of the apparent effect of stay-green traits on yield in various environments (Table 3). For example, an increase in OnS by 1 °Cd led to a yield increase from little or none at KTP11rf up to 0.96 g m⁻² at GAT10rf (Table 3). For a typical day of 18 °Cd during early grain filling, this equates to a yield increase of 17.3 g m⁻² d⁻¹ at GAT10rf, representing ~3.5% of the trial mean yield of 498 g m⁻² for each day’s delay in OnS. Similarly, for MidS, the gains range up to 0.66 g m⁻² per °Cd delay at GAT10rf, equating to 9.9 g m⁻² for each 18 °Cd day, or ~2.4% of the trial mean yield. Thus, a delay of just a few days in OnS or MidS, or an increase in Nmax or SGint was associated with considerable yield differences between genotypes. The apparent magnitude of impact for delayed OnS is similar to that observed for changes between environments in the mean yield of Hartog discussed above (Fig. 3).

The apparent effects on yield of SGint, OnS, and MidS were greatest in ET1 and ET3. They were low in the mild, late-stressed ET2. They were also low in the worst-stressed ET4, possibly resulting from reduced variation in yield in this environment (Fig. 2). There seems to have been little effect of SR on yield in most environments, except GAT10ir (Table 3). The effect of Nmax was varied, ranging from negative to positive in ET1 and ET3. However, there was a consistent positive effect of Nmax on yield in the mild, later-stressed ET2.

To examine further the differences in stay-green traits among genotypes within environments, we plotted the average NDVI of the 5% highest yielding genotypes against the 5% lowest yielding genotypes for each environment (Fig. 5). Clear differences were observed between the highest and lowest yielding genotypes in most environments, with the higher yielding genotypes retaining green leaf area for longer (Fig. 5). The differences appear greatest in the two radiation-limited ET1

![Fig. 4](https://example.com/fig4.png)

**Fig. 4.** Genetic correlations between yield and stay-green traits in eight field environments in southern Queensland, Australia. Stay-green traits, maximum NDVI around anthesis (Nmax), indicator of maximum senescence rate (SR); stay-green integral (SGint), thermal time from anthesis to senescence onset (OnS), and mid-senescence (MidS) are defined in Table 1, and environment types (ET1–ET4), as indicated at the right hand side, are illustrated in Fig. 1. Significance levels for bivariate comparisons are indicated at the levels $P \leq 0.01 (**), P \leq 0.05 (*)$. A bar is absent when there was little variation for the trait in the particular environment or when the correlation between the trait and yield was close to zero. Slopes of regressions fitted to these correlations are given in Table 3. (This figure is available in colour at JXB online.)

**Table 3.** Slope and the SE of the slope of bivariate regressions between yield and stay-green traits Nmax, OnS, MidS, SR, and SGint for the eight studied environments

|          | Nmax | OnS | MidS | SR  | SGint |
|----------|------|-----|------|-----|-------|
|          | Slope| SE  | Slope| SE  | Slope | SE  | Slope| SE  | Slope| SE  | Slope | SE  | ET  |
| GAT10ir  | 17.74| 1.57| 0.49 | 0.16| 0.38  | 0.10| 0.41 | 0.39| 0.53 | 0.15| ET1  |
| GAT10rf  | 0.96 | 0.40| 0.66 | 0.24| 0.35  | 0.18| 0.24 | 0.17| 0.02 | 0.06| ET2  |
| WAR11rf  | −20.08| 7.01| 0.34 | 0.11| 0.60  | 0.16| 0.16 | 0.07| 0.03| 2.25| ET1  |
| GAT11rf  | 12.65| 0.39| 0.35 | 0.18| 0.24  | 0.17| 0.17 | 0.02| 0.06| 0.58 | 0.19| ET2  |
| KTP11rf  | 9.79 | 0.43| −0.04| 0.18| 0.00  | 0.28| 2.06 | 0.04| 0.38 | 0.47| ET2  |
| WAR12rf  | 4.56 | 0.16| 0.26 | 0.09| 0.44  | 0.13| 0.11 | 0.05| 0.54 | 0.17| ET3  |
| GAT12rf  | −9.47| 1.94| 0.37 | 0.08| 0.49  | 0.09| 0.09 | 0.38| 0.92 | 0.23| ET3  |
| WAR12ro  | 0.15 | 0.07| 0.16 | 0.07| 0.07  | 0.05| 0.07 | 0.29| 0.18| ET4  |
environments at Gatton 2010 (GAT10ir, GAT10rf; Fig. 5), where variation for yield was also greatest (Fig. 2). The smallest differences in NDVI between the high- and low-yielding groups were in the two ET2 environments (GAT11rf and KTP11rf, Fig. 5), where significant correlation between yield and stay-green traits OnS, MidS, and SR was mostly absent (Fig. 3). These results are in accordance with the genetic correlations observed between yield and stay-green traits (Fig. 4).

The graphs in Fig. 5 also illustrate why the correlation between SR and yield is positive in some environments, which could at first seem counter-intuitive (Fig. 3). In all environments, the high-yielding genotypes commence senescence later than the low yielders, but NDVI difference narrows as senescence progresses, such that they attain full senescence at a similar time after anthesis (Fig. 5). Commencing senescence later, but completing senescence at a similar time, requires an increased rate of senescence but it results in an increase in overall green leaf area retention (measured by SGint) and increased yield.

It is unlikely that yield contrast between high- and low-yielding groups result from differences in anthesis date. Differences between groups in the mean period from sowing to anthesis were small (ranging from −35 °Cd to +38 °Cd) and non-significant for all environments (Student’s t-test; P > 0.05).

Relationships between stay-green traits varied between environments
PCA was used to examine relationships, not only between stay-green traits and yield, but also between the various

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**Fig. 5.** Logistic regressions of mean normalized difference vegetative index (NDVI) of the highest yielding 5% of genotypes (solid lines) and the lowest yielding 5% of genotypes (dashed lines) at the eight studied environments plotted over thermal time relative to anthesis. Note that differences between high- and low-yielding groups for anthesis were small (from −35 to +38 °Cd) and non-significant for all environments (Student’s t-test; P > 0.05).
stay-green traits (Fig. 6). As anticipated, some stay-green traits can be closely correlated, as discussed above, considering the relationships between Nmax, OnS, MidS, and EndS and their influence on SR and SGint.

A close positive relationship was observed between SGint and yield, as the vector for SGint in the first two principal components had similar direction and length to the yield vector in most environments, and almost overlays it in some instances (Fig. 6). Vectors for OnS and MidS were generally in a similar direction to yield, but not as close as SGint, indicating that SGint was more closely related to yield. The exception was in ET2 environments (KTP11rf and GAT11rf), where Nmax was more closely aligned to yield than OnS and MidS, and even SGint in KTP11rf. This supports the high genetic correlations observed between yield and Nmax in these environments (Fig. 4). The Nmax vector aligned poorly with yield in most ETs other than ET2 (except WAR12rf; ET3), and tends to be close to perpendicular to the yield vector in some cases, suggesting that the two are relatively unrelated in such environments. The vector for SR was also generally close to perpendicular with that of yield in all environments, suggesting a weak relationship. The vectors for SR and Nmax were generally pointing in opposite directions, suggesting a negative relationship. This agrees with the fact that a higher SR is required to reach EndS at a similar time after anthesis from a higher Nmax. Accordingly, vectors for SR were generally on the same side of the yield vector as those for OnS, MidS, and SGint, but the opposite side to the Nmax vector. Overall, yield

![Fig. 6. Biplots of results from principal components analyses (PCA) indicating correlations between yield and stay-green traits for (a) an irrigated trial at Gatton in 2010 (GAT10ir; ET1), and rain-fed trials in (b) Gatton in 2010 (GAT10rf; ET1), (c) Warwick in 2011 (WAR11rf; ET1), (d) Gatton in 2011 (GAT11rf; ET2), (e) Kingsthorpe in 2011 (KRP11rf; ET2), (f) Gatton in 2012 (GAT12rf; ET3), and (g) Warwick in 2012 (WAR12rf; ET3), as well as a trial with rain exclud ed using a rain-out shelter at (h) Warwick in 2012 (WAR12ro; ET4). The direction and magnitude of the vectors represent the effects, in relation to the first two principal components for each analysis for yield and stay-green traits (maximum NDVI around anthesis [Nmax], indicator for maximum senescence rate [SR], stay-green integral [SGint]; thermal time from anthesis to senescence onset [OnS], and mid-senescence [MidS]). Each point corresponds to data for a single genotype. The proportion of variation explained by components one and two are given in parentheses. (This figure is available in colour at JXB online.)](image-url)
was more closely related to SGint > MidS > OnS > SR > Nmax in all environments except in ET2, where Nmax was more closely related to yield (SGint, Nmax > MidS > OnS > SR).

Discussion

The study aim was to determine the potential of recently described stay-green traits to improve knowledge about crop adaptation and to select for adaptation to particular TPE.

Stay-green traits related to whole-of-senescence dynamics are strongly influenced by changes in the water-stress environment

Standardized estimates of component stay-green traits allow comparison of genotype adaptation between different locations and growing seasons. Traits were characterized by fitting a logistic model to NDVI measurements taken at intervals during crop development for each trial plot of each genotype (Christopher et al., 2014), quantifying important characteristics of the stay-green dynamics which differ between genotypes. As discussed above, functional stay-green can be achieved by varying leaf-greenness dynamics in a number of ways (Thomas and Howarth, 2000). Crops may be greener before the onset of senescence (corresponding to higher Nmax), senescence may begin later after anthesis (greater OnS and MidS values), or be slower (lower SR) and/or later finishing (greater EndS) (Thomas and Howarth, 2000; Christopher et al., 2014). Each of these component traits of the stay-green phenotype can contribute to overall green leaf area retention, which is captured by SGint.

Stay-green traits were useful indicators of crop performance in various water-stress environments, despite variation in other important factors affecting yield that could potentially have confounded interpretations including heat stress and radiation limitation. Overall, correlations between stay-green traits and yield were relatively robust and strongly affected by water stress. It is nevertheless important to note that while small differences in phenology were accounted for in the method (using thermal time after flowering), the studied population was also pre-selected to reduce variation for height and anthesis date (Christopher et al., 2013), as these traits are known to affect yield. Some variation in height and anthesis date remained (Fig. 2), but significant correlations with yield occurred in only two of the eight environments for height, and in only one environment for anthesis date.

Overall, Nmax, OnS, MidS, EndS, and SGint all decreased, while SR increased, as water-stress increased from environments in ET1 through to ET4 (Figs 2, 3). All of these traits were highly responsive to water stress.

To enable selection for yield, traits correlated with yield in the relevant environments are preferred.

Genetic variability for stay-green traits correlates with yield in a broad range of environments including those with little water-stress

Stay-green traits OnS, MidS, and SGint could be used as proxy traits to select for adaptation in a broad range of environments. Stay-green phenotype and related traits have frequently been linked to improved yield in crops experiencing a terminal water stress (in wheat, Christopher et al., 2008; Lopes and Reynolds, 2012; Kipp et al., 2014; in sorghum, Borrell et al., 2000b, 2012, 2014a, b; Jordan et al., 2012; maize, Kamara et al., 2003; Wang et al., 2012; rice, Jiang et al., 2004; Hoang and Kobata, 2009; and a range of crops, Thomas and Smart, 1993; Thomas and Howarth, 2000; Gregersen et al., 2013). Stay-green is also widely recognized as a key drought adaptation mechanism in cereals (Passioura, 2006; Richards, 2006; Cattivelli et al., 2008; Gregersen et al., 2013; Thomas and Oughum, 2014). In the absence of water-stress, however, stay-green is not always correlated with yield (in wheat, Lopes and Reynolds, 2012; Gregersen et al., 2013; and in sorghum, Jordan et al., 2012) and can even be associated with reduced yield. For instance, in irrigated wheat and in rice in China, stay-green has been associated with slow export of leaf carbohydrate to the grain, increased lodging, and harvest difficulties due to delayed ripening, all of which can contribute to reduced yield (Gong et al., 2005; Yang and Zhang, 2005). However, even when water is not limiting, increased leaf area duration can lead to prolonged radiation interception and maintenance of photosynthetic capacity, ultimately enhancing potential biomass and grain yield, as shown in sorghum (Borrell et al., 2000b). In the current study, the positive correlations between yield and the stay-green traits SGint, OnS, and MidS suggest that in some wheat populations there is little, if any, physiological cost associated with stay-green including under well-watered conditions (ET1). Furthermore, higher values of stay-green traits OnS, MidS, and SGint appear to be beneficial to yield, as illustrated by the positive and consistent correlation with yield in six out of the eight studied environments, including the three least stressed environments Gat10ir, Gat10rf, and WAR11rf (ET1; Fig. 4), as well as a number of water-stressed environments (ET3 and ET4; Fig. 4).

Compared with some other stay-green traits, SR was not as strongly or consistently correlated with yield (Figs 4, 6). For EndS, although higher values were correlated with higher yield across environments (Figs 2h, 3b), EndS was correlated with yield for genotypes within only two environments in the current study. This suggests that EndS is less useful for genetic selection than several other stay-green traits, at least in this genetic material in this TPE using the current method. This result contrasts with results in sorghum, where estimates of senescence late in crop growth are generally highly correlated with grain yield under post-anthesis drought (Borrell et al., 2000b, 2014a, b; Jordan et al., 2012).

Finally, Nmax was found more promising as an indicator of adaptation to intermediate water-stress environments classified as ET2. In ET2, where post-anthesis water-stress was largely relieved later in the season, OnS, MidS, and SGint appeared less related to high yield than in ET1, ET3, and ET4. Thus, for the SeriM82×Hartog population, OnS and MidS could be useful to select for adaptation to either well-watered environments (ET1) or more severely stressed environments (ET3 and 4), but not ET2 environments. In contrast, Nmax appeared more promising in the ET2 environments examined.
Overall, SGint appears the most useful trait to characterize the stay-green phenotype, as it was highly or significantly correlated with yield in all but one of the ET2 environments (KTP11rf; Fig. 4). This is consistent with expectations, since SGint is an integrator of delayed senescence (area under the logistical curve), measuring leaf area duration and hence maintenance of photosynthetic capacity during grain filling.

**Which trait to select where, and how?**

SGint had the closest relationship with yield in most environments and will probably be useful for selection in a broad range of environments (Figs 4, 6). However, SGint may be complemented by the quantification of traits such as OnS and MidS, especially in studies looking for physiological processes associated with the stay-green phenotype. Change in SGint can arise from changes in different sets of stay-green traits. For example, a genotype with an earlier onset of senescence (lower OnS) might have a similar SGint to a genotype with later onset, if the rate of senescence is slower and EndS greater (see, for example, fig. 6 in Christopher et al., 2014). In the Serim82×Hartog population studied, traits associated with delayed onset of senescence and mid-senescence appeared to be the major determinants of SGint and yield adaptation. In conditions where there was little genetic variation in the timing of senescence completion (EndS) between genotypes, this would mean that a higher, rather than a lower, rate of senescence (SR) was associated with higher SGint and higher yield. While higher EndS and lower SR do not appear so useful for adaptation in this study, stay-green due to a slower rate of senescence has been reported in other populations of wheat (Lopez and Reynolds, 2012) and in other species including sorghum (Borrell et al., 2000a; Thomas and Howarth, 2000; Harris et al., 2007). Lower SR and greater EndS may prove suitable selection traits in such situations. Similarly, leaf greenness at spike maturity (often measured when the peduncle has senesced) has been proposed as another measure of stay-green associated with improved adaptation in wheat (Lopez and Reynolds, 2012) and sorghum (Borrell et al., 2000b). We found little correlation between yield and leaf greenness (NDVI) near the completion of senescence in most environments (Christopher et al., 2014), since in the northern eastern Australian TPE, the leaf canopy of wheat is usually fully senesced at the time of spike maturity (Christopher et al., 2008). We did, however, observe an exception at Gatton in 2010 (GAT10ir and GAT10rf), where exceptionally high rainfall and low radiation during the late grain-filling period resulted in the green-leaf area remaining after crop maturity, possibly associated with sink limitation.

The eight environments tested in this study reflect the variability in water-stress environment types (ET1–ET4) encountered across the Australian cropping region in current and future climates, not only in subtropical northern Australia but also in temperate and Mediterranean climatic regions (Chenu et al., 2011, 2013; Watson et al., 2015). The combination of ET1, and the more stressed ET3 and ET4, represents the majority of environments encountered in many regions of Australia (Chenu et al., 2013). Selection for greater SGint, OnS, and MidS should increase adaptation to these environment types. However, these traits did not significantly relate to yield in ET2 (Fig. 4), which occurs less frequently in the TPE. Overall these traits did not have negative effects in any Australian environment types, and appear promising for adaptation of Australian wheats.

Technologies to determine NDVI for a large number of genotypes over the full senescence period are improving rapidly with development of drones and ‘phenomobiles’ (e.g. Chapman et al., 2014; Deery et al., 2014). Measurements of NDVI using a drone to estimate phenology have recently been demonstrated to improve predictions of wheat yield and quality (Magney et al., 2015). Furthermore, there is potential to identify molecular markers that could be used to enrich germplasm for stay-green traits in early generations, before the need for field phenotyping. Thus, the potential to use dynamic stay-green traits to accelerate selection for adaptation to water-stressed environments is predicted to increase rapidly in the near future. We believe that further research using a broader range of genotypes and environments is warranted and would help determine the most suitable stay-green trait(s) for breeding in other environments in internationally important TPE. Further study of the physiological traits underlying changes in stay-green traits is also required to better understand crop senescence and identify associated genetic controls. Comparative genomics will also be helpful, for example, to determine if the stay-green physiological mechanisms (Borrell et al., 2014a, b) and associated gene networks (Borrell et al., 2015) in sorghum are similar to those in wheat.

**Conclusion**

Increased knowledge about the extent of genetic variation in the component stay-green traits will increase our understanding of crop adaptation in relation to water availability. Combining the use of stay-green traits and environmental water-stress characterization, both standardized for thermal time relative to anthesis, provides a powerful method both to characterize, and to select for, adaptation to well-watered and water-stressed environments.

Stay-green traits used singly, or in combination with other traits and/or markers, have great potential for selecting either broad or specific water-stress adaptation. Stay-green traits SGint, OnS, and MidS were positively correlated with high yield in major water-stress environment types encountered in Australian cropping systems, including those environments with little water stress. There appears to be little, if any, yield penalty associated with these traits in any of the tested environments. Nmax could be useful to select for adaptation to moderately post-anthesis stressed environments in TPE where these are important. Overall, these traits have potential to increase the rate of progress towards higher yield with greater yield stability of wheat in a range of environments. The development of molecular markers to select for these traits would be highly desirable, enabling selection in early generations. Understanding the physiology underlying these stay-green traits will also aid in identification of better and
more stable markers and/or genes for adaptation to water-stressed environments.

**Supplementary data**

Supplementary data are available at *JXB* online.

**Figure S1.** Yield of wheat cultivar Hartog simulated in the Agricultural Production Systems sIMulator (APSIM-wheat v7) and plotted against adjusted mean yield (best linear unbiased predictors; BLUPs) estimated from measurements at eight trials in south east Queensland during 2010, 2011, and 2012.

**Figure S2.** Weekly average of daily maximum temperatures and cumulative incident radiation throughout the growing season plotted at dates relative to anthesis of the reference cultivar Hartog for the eight studied environments.

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**References**

Borrell AK, Hammer GL, Douglas A. 2000a. Does maintaining green leaf area in sorghum improve yield under drought? Leaf growth and senescence. Crop Science 40, 1026–1037.

Borrell AK, Hammer GL, Henzell G. 2000b. Does maintaining green leaf area in sorghum improve yield under drought? Dry matter production and yield. Crop Science 40, 1037–1048.

Borrell AK, Henzell RG, Douglass ACL. 1996. Visual rating of green leaf retention is highly correlated with measured green leaf area in sorghum. In: Froiss MA, Henzell RG, Kneipp JF, eds. Proceedings of the Third Australian Sorghum Conference, Tamworth, 20–22 February1996. Occasional Publication No. 93. Melbourne: Australian Institute of Agricultural Science, 323–326.

Borrell A, Jordan D, Mullet J, Klein P, Klein R, Nguyen H, Rosenow D, Hammer G, Henzell R. 2012. Diving for genes drought adaptation in sorghum. In: Acquaah G, ed. Principles of plant genetics and breeding, 2nd edn. Oxford: Wiley-Blackwell, John Wiley & Sons, 285–289.

Borrell AK, Mullet JE, George-Jaeggli B, van Oosterom EJ, Hammer GL, Klein PE, Jordan D. 2014a. Drought adaptation of stay-green cereals associated with canopy development, leaf anatomy, root growth and water uptake. Journal of Experimental Botany 65, 6251–6263.

Borrell AK, van Oosterom EJ, Mullet JE, George-Jaeggli B, Jordan DR, Klein PE Hammer GL. 2014b. Stay-green alleles enhance grain yield in sorghum under drought by modifying canopy development and enhancing water uptake. New Phytopathology 203, 817–830.

Borrell A, van Oosterom E, Mullet J, George-Jaeggli B, Jordan D, Klein P, Hammer G. 2015. Identifying the function of sorghum’s drought tolerance stay-green QTL. 23rd Plant and Animal Genome Conference, San Diego, USA, 10–14 January 2015.

Butler DG, Cullis BR, Gilmour AR, Gogel BJ. 2009. ASReml-R reference manual, release 3. Technical report, Queensland Department of Primary Industries, Brisbane.

Butler DG, Eccleston JA, Cullis BR. 2008. On an approximate optimality criterion for the design of field experiments under spatial dependence. Australian and New Zealand Journal of Statistics 50, 295–307.

Casadebaig P, Zheng B, Chapman S, Huth N, Faivre R, Chen K. 2016. Assessment of the potential impacts of wheat plant traits across environments by combining crop modelling and global sensitivity analysis. Plos One 11, e0146385.

Cattivelli L, Rizza F, Badeck FW, Mazzucotelli E, Mastrangelo AM, Francia E, Maré C, Tondelli A, Stanca AM. 2008. Drought tolerance improvement in crop plants: an integrated view from breeding to genomics. Field Crops Research 105, 1–14.

Chapman SC, Merz T, Chan A, Jackway P, Hraban S, Drecer MF, Holland E, Zheng B, Ling TJ, Jimenez-Berni J. 2014. Phyto-copter: a low-altitude, autonomous remote-sensing robotic helicopter for high-throughput field-based phenotyping. Agronomy 4, 279–301.

Chen K. 2015. Characterising the crop environment—nature, significance and applications. In: Saddars VO, Calderini DF, eds. Crop physiology: applications for genetic improvement and agronomy. London: Academic Press, 321–348.

Chen K, Deilimfard R, Chapman S. 2013. Large-scale characterization of drought pattern: a continent-wide modelling approach applied to Australian wheat-belt spatial and temporal trends. New Phytologist 198, 801–820.

Chen K, Cooper M, Hammer G, Mathews K, Drecer F, Chapman S. 2011. Environment characterization as an aid to wheat improvement: interpreting genotype-environment interactions by modelling water-deficit patterns in North-Eastern Australia. Journal of Experimental Botany 62, 1743–1755.

Christopher JT, Christopher MJ, Jennings R, Jones S, Fletcher S, Borrell AK, Manschadi AM, Jordan D, Mace E, Hammer GL. 2013. QTL for root angle and number in a population developed from bread wheats (Triticum aestivum) with contrasting adaptation to water-limited environments. Theoretical and Applied Genetics 126, 1563–1574.

Christopher JT, Manschadi AM, Hammer GL, Borrell AK. 2008. Developmental and physiological traits associated with high yield and stay-green phenotype in wheat. Australian Journal of Agricultural Research 59, 354–364.

Christopher JT, Veyradier M, Borrell AK, Harvey G, Fletcher S, Chen K. 2014. Phenotyping novel stay-green traits to capture genetic variation in senescence dynamics. Functional Plant Biology 41, 1035–1048.

Cooper M, Woodruff DR, Phillips IG, Basford KE, Gilmour AR. 2001. Genotype-by-management interactions for grain yield and grain protein concentration of wheat. Field Crops Research 69, 47–67.

Cullis BR, Smith AB, Coombes NE. 2006. On the design of early generation variety trials with correlated data. Journal of Agricultural, Biological and Environmental Statistics 11, 381–393.

Deery D, Jimenez-Berni J, Jones H, Sirault X, Furbank R. 2014. Proximal remote sensing buggies and potential applications for field-based phenotyping. Agronomy 4, 349.

Foulkes MJ, Sylvester-Bradley R, Weightman R, Snape J. 2007. Identifying physiological traits associated with improved drought resistance in winter wheat. Field Crops Research 103, 11–24.

Gabriel KR. 1971. The biplot graphical display of matrices with application to principal component analysis. Biometrika 58, 453–467.

Gilmour AR, Cullis BR, Welham SJ, Thompson R. 1999. ASRemL reference manual. Biometrics Bulletin 3, NSW Agriculture, ORANGE, 2800, Australia.

Gong YH, Zhang J, Gao JF, Lu JY, Wang JR. 2005. Slow export of photosynthesized from stay-green leaves during grain filling stage in hybrid winter wheat (Triticum aestivum L.). Journal of Agronomy and Crop Science 191, 292–299.

Gregersen PL, Culetic A, Boschian L, Krupinska K. 2013. Plant senescence and crop productivity. Plant Molecular Biology 82, 603–622.

Hammer GL, Kropf MJ, Sinclair TR, Porter JR. 2002. Future contributions of crop modelling—from heuristics and supporting decision making to understanding genetic regulation and aiding crop improvement. European Journal of Agronomy 18, 15–31.

Hammer GL, Chapman S, van Oosterom E, Podlich DW. 2005. Trait physiology and crop modelling as a framework to link phenotypic complexity to underlying genetic systems. Australian Journal of Agricultural Research 56, 14.

Harris K, Subudhi P, Borrell A, Jordan D, Rosenow D, Nguyen H, Klein P, Muller J. 2007. Sorghum stay-green QTL individually reduce post-anthesis drought-induced leaf senescence. Journal of Experimental Botany 58, 327–338.

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Haussmann BIG, Obilana AB, Aiyecho PO, Blum A, Schipprack W, Geiger HH. 1999. Quantitative genetic parameters of sorghum (Sorghum bicolor (L.) Moench) grown in semi-arid Kenya. Euphytica 105, 109–118.

Henzell RG, Brengman RL, Fletcher DS, McCosker AN. 1992. Relationships between yield and non-senescent (stay-green) in some grain sorghum hybrids grown under terminal drought stress. In: Foale MA, Henzell RG, Vance PN, eds. Proceedings of the Second Australian Sorghum Conference. Gatton, Australia. 4–6 February 1992. Occasional Publication no. 68. Melbourne: Australian Institute of Agricultural Science, 355–356.

Hoang TB, Kobata T. 2009. Stay-green in rice (Oryza sativa L.) of drought-prone areas in desiccated soils. Plant Production Science 12, 397–408.

Holzworth DP, Huth NI, deVoil PG, et al. 2014. APSIM—evolution towards a new generation of agricultural systems simulation. Environmental Modelling and Software 62, 327–350.

Jackson P, Robertson M, Cooper M, Hammer G. 1996. The role of physiological understanding in plant breeding; from a breeding perspective. Field Crops Research 49, 11–37.

Jiang GH, He YQ, Xu CG, Li XH, Zhang Q. 2004. The genetic basis of stay-green in rice analyzed in a population of doubled haploid lines derived from an indica by japonica cross. Theoretical and Applied Genetics 108, 688–698.

Jordan DR, Hunt CH, Cruickshank AW, Borrell AK, Henzell RG. 2012. The value of the stay-green trait in sorghum. Crop Science 52, 1153–1161.

Joshu AK, Kumbari M, Singh VP, Reddy CM, Kumar S, Rane J, Chand R. 2007. Stay green trait: variation, inheritance and its association with spot blight resistance in spring wheat (Triticum aestivum L.). Euphytica 153, 59–71.

Kamara AY, Menkir A, Badu-Apaku B, Ibikunle O. 2003. Reproductive and stay-green trait responses of maize hybrids, improved open-pollinated varieties and farmers’ local varieties to terminal drought stress. Maydica 48, 29–37.

Kipp S, Mistele B, Schmidhalter U. 2014. Identification of stay-green and early senescence phenotypes in high-yielding winter wheat, and their relationship to grain yield and grain protein concentration using high-throughput phenotyping techniques. Functional Plant Biology 41, 227–235.

Lopes MS, Reynolds MP. 2012. Stay-green in spring wheat can be determined by spectral reflectance measurements (normalized difference vegetation index) independently from phenoology. Journal of Experimental Botany 63, 3789–3798.

Magney TS, Eitel JUH, Huggins DR, Viering LA. 2015. Proximal NDVI derived phenology improves in-season predictions of wheat quality and quantity. Agriculture and Forestry Methodology 217, 46–60.

Manschadi AM, Christopher JT, deVoil, Hammer GL. 2006. The role of root architectural traits in adaptation of wheat to water-limited environments. Functional Plant Biology 33, 823–837.

Manschadi AM, Christopher JT, Hammer GL, deVoil P. 2010. Experimental and modelling studies of drought-adaptive root architectural traits in wheat (Triticum aestivum L.). Plant Biosystems, 144, 458–462.

Olivares-Villegas JJ, Reynolds MP, McDonald GK. 2007. Drought-adaptive attributes in the Seri/Babax hexaploid wheat populations. Functional Plant Biology 34, 189–203.

Passiourea JB. 2006. Increasing crop productivity when water is scarce—from breeding to field management. Agricultural Water Management 80, 176–196.

R Development Core Team. 2013. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org/.

Rebetzke GJ, Chenu K, Biddulph B, Moeller C, Deerey DM, Rattey AR, Bennett D, Barrett-Lennard EG, Mayer JE. 2013. A multisite managed environment facility for targeted trait and germplasm phenotyping. Functional Plant Biology 40, 1–13.

Reynolds MP, Nagarajan S, Razzaque MA, Ageeb OAA. 2001. In: Reynolds MP, Ortiz-Monsasterio JI, Mc Nab A, eds. Application of physiology in wheat breeding. Mexico, D.F.: CIMMYT, 124–135.

Richards RA. 2006. Physiological traits used in the breeding of new cultivars for water-scarce environments. Agricultural Water Management 80, 197–211.

Richards RA, Rebetzke GJ, Condon AG, van Herwaarden AF. 2002. Breeding opportunities for increasing the efficiency of water use and crop yield in temperate cereals. Crop Science 42, 111–121.

Rosenow DT, Quisenberry JE, Wendt CW, Clark LE. 1983. Drought tolerant sorghum and cotton germplasm. Agricultural Water Management 7, 207–222.

Sivapalan S, O’Brien L, Ortiz-Ferrara G, Hollamby GJ, Barclay I, Martin PJ. 2000. An adaptation analysis of Australian and CIMMYT/ICARDA developed wheat genotypes in the West Asia North Africa (WANA) region. Australian Journal of Agricultural Research 51, 903–915.

Sivapalan S, O’Brien L, Ortiz-Ferrara G, Hollamby GJ, Barclay I, Martin PJ. 2001. Yield performance and adaptation of some Australian and CIMMYT/ICARDA developed wheat genotypes in the West Asia North Africa (WANA) region. Australian Journal of Agricultural Research 52, 61–670.

Smith AB, Gullis BR, Thompson R. 2001. Analyzing variety by environment data using multiplicative mixed models and adjustments for spatial field trends. Biometrics 57, 1138–1147.

Tardieu F. 2003. Virtual plants: modelling as a tool for the genomics of tolerance to water deficit. Trends in Plant Science 8, 9–14.

Tashiro T, Wardlaw IF. 1990. The response to high temperature shock and humidity changes prior to and during the early stages of grain development in wheat. Australian Journal of Plant Physiology 17, 551–561.

Thomas H, Howarth CJ. 2000. Five ways to stay green. Journal of Experimental Botany 51, 329–333.

Thomas H, Smart CM. 1993. Crops that stay green. Annals of Applied Biology 123, 193–219.

Thomas H, Ougham H. 2014. The stay-green trait. Journal of Experimental Botany 65, 3889–3900.

Vijayalakshmi K, Fritz AK, Paulsen GM, Bai GH, Pandravada S, Gill BS. 2010. Modelling and mapping QTL for senescence-related traits in winter wheat under high temperature. Molecular Breeding 26, 163–175.

Wang A-Y, Li Y, Zhang C. 2012. QTL mapping for stay-green in maize (Zea mays) Canadian Journal of Plant Science 92, 249–256.

Watson J, Zheng B, Chapman SC, Chenu K. 2013. Impact of projected climates on drought occurrence in the Australian wheatbelt. In: 17th Australian Agronomy Conference. 20–24 September, Hobart, Australia, 163–167.

Yang J, Zhang J. 2005. Grain filling of cereals under soil drying. New Phytologist 169, 223–236.

Zadok JC, Chang TT, Konzak FC. 1974. A decimal code for growth stages of cereals. Weed Research 14, 415–421.