First report on a novel *Nigrospora sphaerica* isolated from *Catharanthus roseus* plant with anticarcinogenic properties

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Summary

This paper reports on the vinca alkaloid produced by a novel *Nigrospora sphaerica* isolated from *Catharanthus roseus*. Through liquid chromatography–mass spectrometry (LCMS), only the crude mycelia extract of this fungus was positive for determination of vinblastine. This vinca alkaloid was then purified by using high-performance liquid chromatography (HPLC) and tested for cytotoxicity activity using MTT assays. The breast cell line cancer (MDA-MB 231) was treated with a purified vinblastine which was intracellularly produced by *N. sphaerica*. The purified vinblastine from extracted leaf of *C. roseus* was used as a standard comparison. A positive result with a value of half maximal inhibitory concentration (IC$_{50}$) of $>32$ µg ml$^{-1}$ was observed compared with standard (IC$_{50}$) of 350 µg ml$^{-1}$ only. It showed that a vinblastine produced by *N. sphaerica* has a high cytotoxicity activity even though the concentration of vinblastine produced by this endophytic fungus was only 0.868 µg ml$^{-1}$.

Introduction

Endophytes are microbes that refer to microorganisms that live inside the tissues of plants without causing any apparent harm or diseases to the host plant (Strobel, 2002). In fact, they promote the host plant's growth and the formation of secondary metabolites related to the plant defence (Petrini, 1991 & Chandra, 2002). In fact, they promote the host plant's growth and the formation of secondary metabolites related to the plant defence (Petrini, 1991 & Chandra, 2002). They could produce valuable bioactive compounds with a varied application in both of research and applied fields (Ravindra et al., 2014). Endophytic fungi spend the whole part of their life cycle living symbiotically within the healthy tissues of the host plant (Tan and Zhou, 2001; Ravindra et al., 2014). It also has been recognized as one of important and novel resources of natural bioactive products (Strobel et al., 2004) as most endophytes are capable of synthesizing bioactive compounds that may provide plants with a defence against pathogens (Guo et al., 2008). Some of these compounds have proven useful for discovering a novel drug (Yan et al., 2011). There are many reports that endophytic fungi isolated from a medicinal plant produce a new drug or compound that similar to the host plants (Table 1). All these findings will help to fill the demands of the drugs. In fact, the manufacturing cost of the drugs from endophytic fungi is cheaper than production from the plants as it takes a shorter period to produce it.

*Catharanthus roseus* or well known as a Madagascar periwinkle is a medicinal plant belonging to the family Apocynaceae (Gajalakshmi et al., 2013). Even though this plant is native to Madagascar, it also can be found in Malaysia. Here, it is called as Kemunting Cina and the flower of this plant was chosen as a logo for the National Cancer Council Malaysia (MAKNA) (Ayob and Simarani, 2016). This plant also well known to produce a lot of important compounds especially vinca alkaloids vinblastine and vincristine (Manganey et al., 1979; Krishnan, 1995). Besides, this plant also produces vindoline and catharanthine which are the major monomer alkaloids as well as a biosynthetic precursor for vinblastine and vincristine (Noble, 1990). In 1960, vinblastine was introduced to treat certain types of cancer including breast cancer, testicular cancer and Hodgkin's disease (Armstrong et al., 1964), while in 1963, vincristine was introduced through oxidization of vinblastine to treat leukaemia (Evans et al., 1963). So far, there are only three reports on these alkaloids produced by the endophytic fungi, which were *Altermaria sp.*., *Fusarium oxysporum* and unidentified fungi from *C. roseus* (Guo et al., 1998; Zhang et al., 2000; Yang et al., 2004). Thus, this research was carried out to find a novel endophytic fungus that could produce vinca alkaloids, vinblastine and vincristine from the host plant *C. roseus*. The purified alkaloids will be tested for cytotoxicity test through MTT assay against breast cell line cancer.
Results and discussion

The crude fungal extract was prepared using the 21 days of fungal cultured (Fig. 1), while the crude leaf extract was prepared using dried leaves of wild grown *C. roseus* (white). Through a liquid chromatography–mass spectrometry (LCMS), the crude leaf extract showed a presence of both vinblastine and vincristine. However, crude mycelia extract only showed a positive result on vinblastine. It was determined by the presence of two fragment ions from a peak in the same compounds and indicated a confirmation of the compound detected (Fig. 2). Meanwhile, no detected peak was observed for both alkaloids in a crude broth fungal extract. This result indicated that the vinblastine was intracellularly produced by this fungus. The crude mycelia extract of *N. sphaerica* and crude leaf extract of *C. roseus* consisted 0.868 and 0.666 µg g⁻¹ of vinblastine respectively (Fig. 3). Hence, in this study, the cytotoxicity test was only carried out using vinblastine due to the absence of vincristine in the extracted fungal mass sample.

Purified vinblastine from crude mycelia extract of this fungus and crude leaf extract were obtained using high-performance liquid chromatography (HPLC). The samples were eluted for 5 min of retention time when loaded on analytical C₁₈ column with a flow rate of 1 ml min⁻¹. It was clearly seen from the chromatograph of HPLC, the retention time for fungal vinblastine peak was similar with vinca alkaloid extracted from leaf of *C. roseus*. Cytotoxicity of purified vinblastine produced by both endophytic fungus *N. sphaerica* and *C. roseus* was tested against human breast carcinoma MDA-MB 231 cell line cancer with various concentrations (6.35–400 µg g⁻¹). The test showed a positive result with a half maximal inhibitory concentration (IC₅₀) value of >32 and 350 µg g⁻¹ for the vinblastine that has been purified from both crude fungal extract and a crude leaf extract respectively (Figs 4 and 5). This happened when the vinca alkaloid was binding to beta-tubulin and disruption of microtubule function during mitosis, which leads to mitosis arrest and cell death (Damen *et al.*, 2010). Jordan and Wilson (2004) in their reports also mentioned that the anticancer activity of these alkaloids was attributed to their ability to disrupt microtubules metaphase arrest in dividing cell.

Currently, the commercial vinblastine in the market is produced by *C. roseus*. However, it takes almost one year before it is ready for harvesting. In the other hand, vinblastine produced by *N. sphaerica* only takes only a month of preparation (cultivation, extraction and purification) before it is ready to be used. In short, the vinblastine could be produced faster with a huge amount using crude mycelia extract of *N. sphaerica* instead of

| Host plants        | Endophytic fungi       | Drugs                  | References                  |
|--------------------|------------------------|------------------------|-----------------------------|
| *P. amarus*        | *Trichothecium* sp.    | Trichothecinol-A       | (Ravinda *et al.*, 2014)    |
| *Eugenia jambolana*| *Cephalotheca faveolata* | Sclerotirin            | (Periyasamy *et al.*, 2012) |
| *Camptotheca acuminata* | *Fusarium solani*        | Camptothecin           | (Lin *et al.*, 2011)        |
| *Taxus brevifolia* | *Taxomyces andreanale* | Taxol                  | (Wani *et al.*, 1971)       |
| *Podophyllum peltatum* | *Phialosphala fortinii* | Podophyllotoxin        | (Kumar *et al.*, 2013)      |
current resource of this alkaloid, *C. roseus*. This new finding will help to fill the demand of this valuable natural product for cancer treatment. This was supported by Ravindra et al. (2011) that among the metabolites produced by the endophytic fungi, attention has attracted to the compound with anticancer properties. So far, there were 100 anticancer substances classified into 19 different chemical classes with an activity against 45 different cancer cell lines that have been isolated from 50 fungal species (Abdulmyanova et al., 2015). However, it was reported that only one fungal produced vinblastine; *Alternaria* sp. which was isolated from the *C. roseus* (Guo et al., 1998).

As a conclusion, the crude mycelia extract of *N. sphaerica* isolated from a medicinal plant *C. roseus* was positive produced vinblastine with a concentration of 0.868 µg ml⁻¹ which is higher than vinblastine produced by the crude leaf extract (0.666 µg ml⁻¹). The
cytotoxicity test using the 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) assay against breast cell line cancer MDA-MB 231 for the vinblastine produced by both extracts showed that the vinblastine that has been purified from a crude mycelia extract of N. sphaerica has a better half maximal inhibitory concentration (IC\textsubscript{50}) > 32 \mu g ml\textsuperscript{-1} compared with the vinblastine purified from a crude leaf extract of C. roseus > 350 \mu g ml\textsuperscript{-1}.

**Experimental procedures**

**Microorganism and growth condition**

The fungal strain used in this study was previously isolated from a C. roseus plant (white), purified and identified as *Nigrospora sphaerica* (Ayob and Simarani, 2016). The strain was maintained on slanting agar and reactivated on the PDA plate prior to use. There were six pieces of the plug (1 cm) of fungal growth culture, which were inoculated into a 250 ml conical flask containing 250 ml of yeast extract sucrose broth (YESB) medium (20 g of yeast extract, 40 g of sucrose and 1 l of distilled water with a pH 5.8). The flasks were then incubated at 25 °C under static condition for 21 days. The grown culture was harvested by filter separation method and used for crude extract preparation.

**Preparation of crude fungal extract and crude leaf extract**

The crude fungal extract (CFE) from cell free filtrate and mycelia biomass were prepared according to Wiyakrutta *et al.* (2004) with minor modification. The cell free filtrate was extracted thrice with a dichloromethane (200 ml) and evaporated to dryness using rotary evaporator. The extracted sample was weighed to constitute the crude filtrate extract (CFE). Meanwhile, the mycelia biomass...
were freeze-dried and extracted twice with a mixture of dichloromethane:methanol (1:1, v/v) for 1 h. The extracted sample was then air-dried and weighed before kept in the airtight container until further used.

Meanwhile, the crude extracts of C. roseus leaves were prepared according to Gupta et al. (2005). Briefly, 5 g of powdered samples was soaked thrice with 90% (v/v) ethanol (3 × 30 ml) for 12 h each at room temperature. The alcohol extract was filtered and concentrated in vacuo to reduce the volume down to 10 ml. The sample was then diluted with 10 ml of distilled water followed by acidified with 10 ml of 3% (v/v) hydrochloric acid and washed with hexane (3 × 30 ml). The aqueous portion was basified with ammonia to pH 8.5 and extracted using chloroform (3 × 30 ml). The chloroform extract was washed with distilled water, dried over sodium sulfate and concentrated under vacuum before dissolving in 10 ml of methanol. Both of crude extracts were then dissolved separately in dimethylsulfoxide (DMSO) to obtain 1 g ml⁻¹ of concentration.

Analysis/determination of vinca alkaloids

The presence of vinca alkaloids in both crude extracts were detected using liquid chromatography–mass spectrometry (AB Sciex 3200Q Trap LCMS/MS; AB Sciex Pte, Ltd., Ontario, Canada), method multiple reaction monitoring (MRM), equipped with a column Agilent Zorbax XDB C18 (150 × 4 × 5 μM) and buffered with (A) pure water, ammonium formate and formic acid, (B) acetonitrile, ammonium formate and formic acid. The experiment was run for 10 min and rapidly screened for the peaks because it should have two fragment ions from the same compounds for further confirmation of the peaks.

The fungal crude extracts were then purified using high-performance liquid chromatography (Agilent 1220 Infinity Gradient LC – G4294B) for vinblastine and vincristine. The 10 μl sample was injected in HPLC column C18 Merck (50 × 2 × 1.5 μM), and isocratic elution was performed using methanol and Nano pure water with 0.01% acetic acid at a flow rate of 1 ml min⁻¹. Dual wavelengths 254 and 260 nm were used to detect the compound eluting from the column. The purified vinblastine was collected using Agilent Fraction Collector (G1364C) and was tested for cytotoxicity test.

Cytotoxicity activity (MTT assay)

Cytotoxicity of vinblastine was assessed using the 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT; Sigma) assay against breast cell line cancer MDA-MB 231. The cancer cell were grown in Dulbecco's
modified Eagle’s medium (DMEM) and washed in phosphate-buffered solution (PBS) before 100,000 cells were placed in each well of 96-well plates and treated with fungal extract. After 72 h incubated at 37 °C and 5% CO2, the MTI reagent was removed from the plate and replaced with a DMSO and gently shaken for 30 min. The absorption was determined at 570 nm. The percentage of inhibition (POI) was calculated using formula:

$$POI = \frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \times 100$$

(1)

The half maximal inhibitory concentration (IC50) was determined from the graph of samples concentration vs. percentage of inhibition.

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**Conflict of interest**

None declared.

**References**

Abdulmyanova, L.I., Teomashko, N.N., Terentyeva, E.O., Ruzieva, D.M., Sattarov, R.S., Azimova, S.S., and Gulyanova, T.G. (2015) Cytotoxicity activity of fungal endophytes from vinca. *Int J Curr Microbiol Appl Sci* 4: 321–329.

Armstrong, J.G., Dyke, R.W., and Fouts, P.J. (1964) Vinblastine sulfate treatment of Hodgkin’s disease during a pregnancy. *Science* 143: 703.

Ayob, F.W., and Simarani, K. (2016) Endophytic filamentous fungi from a *Catharanthus roseus*: identification and its hydrolytic enzymes. *Saudi Pharmaceut J* 24: 273–278.

Chandra, S., Bandopadhay, R., Kumar, V., and Chandra, R. (2010) Acclimatization of tissue cultured plantlets: from laboratory to land. *Biotechnol Letter* 32: 191–1205.

Damen, C.W.N., Rosing, H., Schellens, J.H.M., and Beijnen, J.H. (2010) High performance liquid chromatography coupled with mass spectrometry for the quantitative analysis of vinca alkaloids Vinca alkaloids in biological matrices: a concise survey from the literature. *Biomed Chromatogr* 24: 83–90.

Evans, A.E., Farber, S., Brunet, S., and Mariano, P.J. (1963) Vincristine in the treatment of acute leukemia in children. *Cancer* 16: 1902–1906.

Gajalakshmi, S., Vijayalakshmi, S., and Devi, R.V. (2013) Pharmacological activities of *Catharanthus roseus*: a perspective review. *Int J Pharma Bio Sci* 4: 431–439.

Guo, B., Li, H., and Zhang, L. (1998) Isolation of the fungus producing vinblastine. *J Yunnan Univ (Natural Science Ed)* 20: 214–215.

Guo, B., Wang, Y., Sun, X., and Tang, K. (2008) A review: bioactive natural products from endophytes. *Appl Biochem Microbiol* 44: 136–142.

Gupta, R., Kannan, G.M., Sharma, M.S., and Flora, S.J. (2005) Therapeutic effects of *Moringa oleifera* on arsenic-induced toxicity in rats. *Environ Toxicol Pharmacol* 20: 456–464.

Jordan, M.A., and Wilson, L. (2004) Microtubules as a target for anticancer drugs. *Nat Rev Cancer* 4: 253–265.

Krishnan, R. (1995) Advances in horticulture, medicinal and aromatic plants. Malhotra Publishing House, New Delhi 409–428.

Kumar, A., Patil, D., Rajamohanan, P.R., and Ahmad, A. (2013) Isolation, purification and characterization of vinblastine and vincristine from endophytic fungus *Fusarium oxysporum* isolated from *Catharanthus roseus*. *PLoS One* 8: 2–10.

Lin, T., Wang, G.H., Lin, X., Hu, Z.Y., Chen, Q.C., Xu, Y., *et al.* (2011) Three new oblongolides from *Phomopsis sp* XZ-01, an endophytic fungus from *Camptotecha acuminata*. *Molecules* 16: 3351–3359.

Manganey, P., Andriamialisoa, R.Z., Langlois, Y., Langlois, N., and Potier, P. (1979) Preparation of vinblastine, vincristine and leuroidine, antitumor alkaloids from *Catha ranthus* spp (Apocynaceae). *J Am Chem 101*: 2243–2245.

Noble, R.L. (1990) The discovery of the vinca alkaloids–chemotherapeutic agents against cancer. *Biochem Cell Biol* 68: 1344–1351.

Periyasamy, G., Shilpa, A.V., Amit, K., Mishra, P.D., and Sunil, K.D. (2012) Anticancer activity of sclerotiorin, iso sclerotiorin and characterization of *Cephalotheca taveolata* Yakuchi, Nishim & Udagawa. *Indian J Exp Biol 50*: 464–468.

Petri, O. (1991) Fungal endophytes of tree leaves. In: *Microbial Ecology of Leaves* Andrews, J. and Hirano, S.S. (eds). New York, NY, Springer, pp. 179–197.

Ravindra, N., Khanwar Mishra, A., Gond, S.K., Steirle, A. and Steirle, D. (2011) Anticancer compounds, derived from fungal endophytes: their importance and future challenges. *Nat Prod Rep* 28, 1208–1228.

Ravindra, T., Prasad, A., Deepak, P., Pattuparrambil, R.R., Remya, R., Gowroshankar, S., *et al.* (2014) Isolation, purification and characterization of *Trichothecinol-A* produced by endophytic fungus *Trichothecium* sp and its antifungal, anticancer and antimetastatic activities. *Sustainable Chem Proc* 2: 8.

Strobel, G.A. (2002) Rainforest endophytes and bioactive products. *J Crit Rev Biotechnol* 22: 315–333.

Strobel, G., Daisy, B., Castillo, U., and Harper, J. (2004) Natural products from endophytic microorganisms. *J Nat Prod* 67: 257–268.

Tan, R.X., and Zhou, W.X. (2001) Endophytes: a rich source of functional metabolites. *Nat Prod Rep* 18: 448–459.
Wani, M.C., Taylor, H.L., Wall, M.E., Coggon, P., and McPhail, A.T. (1971) Plant antitumor agents VI: the isolation and structure of taxol, a novel antileukemic and anti-tumor agent from *Taxus brevifolia*. *J Am Chem Soc* 93: 2325–2327.

Wiyakrutta, S., Sriubolmas, N., Panphut, W., Thongon, N., Danwisetkanjana, K., Ruangrungsi, N., and Meevootisom, V. (2004) Endophytic fungi with anti-microbial, anti-cancer and anti-malarial activities isolated from Thai medicinal plants. *World J Microbiol Biotechnol* 20: 265–272.

Yan, X.N., Sikora, I.R., and Zheng, J.W. (2011) Potential use of cucumber (*Cucumis sativus* L) endophytic fungi as seed treatment agents root-knot nematode *Meloidogyne incognita*. *J Zhejiang Univ-Sci B (Biomed & Biotechnol)* 12: 219–225.

Yang, X., Zhang, L., Guo, B., and Guo, S. (2004) Preliminary study of a vincristine producing endophytic fungus isolated from leaves of *Catharanthus roseus*. *China Trad Herbal Drug* 35: 79–81.

Zhang, L., Guo, B., Li, H., Zeng, S., Shao, H., Gu, S., and Wei, R. (2000) Preliminary study on the isolation of endophytic fungus of *Catharanthus roseus* and its fermentation to produce products of therapeutic value. *China Trad Herbal Drug* 31: 805–807.