Case Report

Multiple-System Atrophy in Long-Term Professional Painter: A Case Report

Yusa Nagai,1 Riko Kitazawa,1 Miku Nakagawa,1 Munenori Komoda,1 Takeshi Kondo,2 Ryuma Haraguchi,1 and Sohei Kitazawa1

1Division of Molecular Pathology, Ehime University Graduate School of Medicine, Shitsukawa, Ehime, Toon City, 791-0295, Japan
2Department of Legal Medicine, Kobe University Graduate School of Medicine, 7-5-1 Kasumoki-cho, Chuo-ku, Kobe 650-0017, Japan

Correspondence should be addressed to Sohei Kitazawa, kitazawa@m.ehime-u.ac.jp

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1. Introduction

Multiple system atrophy (MSA) is a rare and severe adult-onset, sporadic, and progressive neurodegenerative disorder. Here, we describe an autopsy case of MSA in a long-term professional painter. Although typical glial cytoplasmic inclusion (GCI) was not observed in a routine histological examination, strong α-synuclein immunostaining in the nucleus confirmed the diagnosis of MSA.

2. Case Presentation

A 48-year-old Japanese man, a professional painter, was admitted to the emergency room. His past history revealed that at age 28, he consulted a general practitioner with the chief complain of tremors and inarticulateness, and followup showed intoxication from organic solvents. At age 42, he was diagnosed with diabetes and received regular outpatient treatment. Mitochondria-related neuropathy was ruled out through careful family history taking and a series of tests.
By then he had started to use a wheelchair because of gait disorders, and complained of amnesia. At 46, the patient was diagnosed with spinocerebellar degeneration through extensive neurological tests. The symptoms gradually progressed to repeated episodes of aspiration pneumonia. The patient was found unconscious with cardiopulmonary arrest attributed to aspiration at supper, and despite attempts at resuscitation, he died of multiple organ failure. The autopsy was done 12 hours thereafter.

Postmortem examination revealed the body of a corpulent man, 167 cm tall, weighing 67.8 kg (BMI 24.8). Traumatic scars were noted in the left temporoparietal area, the precordium, and the right forearm. Marked disuse atrophy was observed in the extremities, especially in the legs. Observed also, besides severe lung congestion (left: 390 g and right: 520 g), were significant amounts of food debris and expectoration in the trachea and bronchi, with rubefaction and hemorrhagic change in the mucous membrane, and reflux esophagitis with numerous erosions in the mucous membrane of the stomach. The clinical diagnosis of repeated aspiration pneumonia was thus confirmed, and the direct cause of death was evidently due to suffocation by mis swallowed. Furthermore, fatty liver (1815 g), pancreatic swelling (105 g), a moderate degree of aortic atherosclerosis, and marked accumulation of visceral fat reflected the longstanding history of type 2 diabetes mellitus. A neurogenic bladder with an expanded and trabeculated wall was also noted.

2.1. General Neuropathological Findings. Gross examination of the brain (1300 g) revealed a highly atrophic brainstem, cerebellum, and spinal cord (Figures 1(B) and 1(C)), while the thickness of the cerebral cortex was almost intact (Figure 1(A)). Severe atrophy of the paleostriatum together with dilatation of the lateral ventricle was observed in coronal sections (Figure 1(A)). Additionally, the cerebellar hemisphere demonstrated fading in the gray matter (Figure 1(B)). On the other hand, no depigmentation of the substantia nigra or of the locus coeruleus was observed in sections of the brainstem (Figures 1(B) and 1(C)).

A histopathological examination showed severe neuronal loss, gliosis, and tissue rarefaction in various areas, predominantly in the paleostriatum, the striate body of the substantia nigra, the pons, and the olivary nucleus of the upper medulla oblongata, intermedio lateral of the spinal gray matter (sacral region) (Figure 2), all of which suggested multiple system atrophy (OPCA+SDS).

The expression and localization of α-synuclein in the cerebral cortex, the midbrain, the medulla oblongata, and the spinal cord was determined by immunohistochemical analysis of the relevant formalin-fixed and paraffin-embedded specimens. Sections (4-μm thick) were deparaffinized in
Figure 2: (A–H) Histological and immunohistochemical findings. Interfascicular oligodendroglia with halos in the cytoplasm and astrocytes are observed along with nerve fibers (A, HE staining). α-synuclein staining reveals numerous α-synuclein-positive glial cytoplasmic inclusions (GCI) in the oligodendroglia in the cerebral cortex (B). In the midbrain, the oligodendroglia and astrocytes show severe cytogenic edema (C, HE staining). α-synuclein-positive GCI in the oligodendroglia is also seen in the midbrain (D). The medulla oblongata shows extensive neuronal loss with scant oligodendrogial cells (E, HE staining), where α-synuclein is positive in the cytoplasma (F). Neuronal loss and gliosis are observed in the spinal cord (G, HE staining). Some α-synuclein-positive GCI in the oligodendroglia and astrocytes (arrows) are seen in the spinal cord (H).
xylene for 20 min (solvent refreshed at 10 min and 5 min), immersed in absolute ethanol for 10 min (solvent refreshed at 5 min), rehydrated in 90, 70% ethanol (5 min each), and finally placed in distilled water for 15 min (solvent refreshed every 5 min). The samples were then inactivated in 1 mM EDTA (pH 8.0) plus distilled water in a microwave oven for 15 min (high temperature for 5 min and low for 10 min) and cooled to approximately 35°C for 1 h. After liquid block treatment, the samples were immersed in phosphate buffered saline (PBS) for 15 min (solvent refreshed every 5 min). To observe the localization of α-synuclein, epitope specific rabbit anti-α-synuclein (spring bioscience, CA) was used as the primary antibody. After adding blocking buffer in a moisture chamber and incubating at room temperature for 1 h, the samples were washed 3 times for 5 min each in PBS, incubated again for 1 h with the secondary antibody (SC-2004, santa cruz biotechnology, inc., CA), diluted at 1:200 with PBS, and then immersed in PBS for 15 min (solvent refreshed every 5 min). The samples were then incubated with 3,3′-diaminobenzidine, tetrahydrochloride (DAB) for 15 min, washed in PBS, counterstained with hematoxylin, and observed and photographed under a microscope. The anti-α-synuclein antibody immunostained the glial and neuronal inclusions extensively, especially the cytoplasmic and nuclear inclusions in the oligodendroglia. α-synuclein-positive GCI in the oligodendroglia was occurred in the cerebral cortex, the midbrain, the medulla oblongata, and the spinal cord (Figures 2(B), 2(D), 2(F), and 2(H)). While gliosis and tissue rarefaction were observed in the midbrain, those in the locus coeruleus were relatively mild in the sections of the midbrain of the cerebellum level (data not shown).

3. Discussion

Patients with atypical parkinsonism are significantly more exposed to environmental toxins than are controls, and demonstrate a higher risk of disease onset associated with occupational exposure to organic solvents, plastic monomers and additives, pesticides, and metals [4]. Moreover, symptoms and neurological diseases are observed at a higher frequency in first relatives of MSA patients than in controls (23% in MSA cases versus 10% in controls) [4]. Epidemiological studies in the French West Indies in 1999, on the other hand, implied an association between atypical parkinsonism and high consumption of tropical plants [5]. The European Study on Atypical parkinsonism in 2001 shows a significantly high risk of MSA in subjects with occupational exposure to various toxins, but a significantly lower risk among smokers, a factor often associated with a decreased risk of Parkinson’s disease [6]. A more recent case-control study in the Aquitaine, France in 2004, shows a history of farming as significantly more frequent in MSA patients than in controls; it did not, however, reveal an association between occupational exposure to pesticides and MSA [7]. To date, on the other hand, no familial MSA case with a definite genetic background has been reported, nor has the genetic predisposition to MSA been established. Therefore, although exposure to environmental toxic substances may increase the risk of MSA, its pathogenesis is still unclear, and is simply defined as one of α-synucleinopathy characterized by the presence of GCI.

In the present case, the apparent risk factors for neurological manifestations were heavy exposure to organic solvents. Interestingly, an in vitro experiment has revealed that α-synuclein, originally in an unfolded form, is rapidly folded with an enhanced propensity to fibrillate in organic solvents [8]. Furthermore, by measuring hydrodynamic radii and adding glucose to solvents causes thorough collapse of α-synuclein [9]. The prolonged and extensive exposure to organic solvents together with the hyperglycemic condition attributed to diabetes may have contributed to the onset and clinical course of the present case.

4. Conclusion

Because most MSA cases lack a clear history of exposure to organic solvents with concurrent diabetes, some unknown genetic factors could define susceptibility to the disease. Accumulation of data from particular cases like the present one would be conducive to revealing clues in future studies.

Consent

Written informed consent was obtained from the patient’s family for publication of this case report and accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Conflict of Interests

The authors declare that they have no competing interests.

Authors’ Contribution

Y. Nagai and S. Kitazawa were involved in whole process. R. Kitazawa and T. Kondo analyzed the autopsy case. M. Nakagawa and M. Komoda assisted clinical aspect. R. Haraguchi was involved in HE staining and immunostaining. All authors read and approved the final version of the paper.

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References

[1] J. G. Graham and D. R. Oppenheimer, “Orthostatic hypotension and nicotine sensitivity in a case of multiple system atrophy,” Journal of Neurology Neurosurgery and Psychiatry, vol. 32, no. 1, pp. 28–34, 1969.
[2] M. I. Papp, J. E. Kahn, and P. L. Lantos, “Glial cytoplasmic inclusions in the CNS of patients with multiple system atrophy (striatonigral degeneration, olivopontocerebellar atrophy and Shy–Drager syndrome),” Journal of the neurological sciences, vol. 94, no. 1-3, pp. 79–100, 1989.
[3] T. Ikeda, H. Mashimoto, K. Iwasaki, I. Shimokawa, and T. Matsumo, “A sequential ultrastructural and histoautoradiographic
study of early neoplastic lesions in ethylnitrosourea-induced rat glioma,” Acta Pathologica Japonica, vol. 39, no. 8, pp. 487–495, 1989.

[4] L. E. Nee, M. R. Gomez, J. Dambrosia, S. Bale, R. Eldridge, and R. J. Polinsky, “Environmental- occupational risk factors and familial associations in multiple system atrophy: a preliminary investigation,” Clinical Autonomic Research, vol. 1, no. 1, pp. 9–13, 1991.

[5] D. Caparros-Lefebvre and A. Elbaz, “Possible relation of atypical parkinsonism in the French West Indies with consumption of tropical plants: a case-control study,” The Lancet, vol. 354, no. 9175, pp. 281–286, 1999.

[6] N. Vanacore, V. Bonifati, G. Fabbrini et al., “Epidemiology of multiple system atrophy,” Neurological Sciences, vol. 22, no. 1, pp. 97–99, 2001.

[7] V. Chrysostome, F. Tison, F. Yekhlef, C. Sourgen, I. Baldi, and J. F. Dartigues, “Epidemiology of multiple system atrophy: a prevalence and pilot risk factor study in Aquitaine, France,” Neuroepidemiology, vol. 23, no. 4, pp. 201–208, 2004.

[8] L. A. Munishkina, C. Phelan, V. N. Uversky, and A. L. Fink, “Conformational behavior and aggregation of α-synuclein in organic solvents: modeling the effects of membranes,” Biochemistry, vol. 42, no. 9, pp. 2720–2730, 2003.

[9] A. S. Morar, A. Olteanu, G. B. Young, and G. J. Pielak, “Solvent-induced collapse of α-synuclein and acid-denatured cytochrome c,” Protein Science, vol. 10, no. 11, pp. 2195–2199, 2001.