Gestational hypothyroidism-induced changes in L-type calcium channels of rat aorta smooth muscle and their impact on the responses to vasoconstrictors

Katayoun Sedaghat 1, 2, Saleh Zahediasl 1, 2, Asghar Ghasemi 1, 2 *

1 Endocrine Physiology Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran
2 Endocrine Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Article type: Original article

Article history: Received: May 5, 2014 Accepted: Nov 6, 2014

Keywords: Aorta smooth muscle Gestational hypothyroidism L-type calcium channel

ABSTRACT

Objective(s): Thyroid hormones play an essential role in fetal growth and maternal hypothyroidism which leads to cardiovascular deficiency in their offspring. Considering this, we intended to investigate the impact of gestational hypothyroidism on offspring vascular contractility and possible underlying mechanisms.

Materials and Methods: Hypothyroidism was induced in female rats by administration of 6-n-propyl-2-thiouracil in drinking water (0.02%) till delivery. The offspring aorta smooth muscle (without endothelium) contractile response to KCl (10-100 mM), KCl in the presence of nifedipine (10^{-6}-10^{-4} \mu M), phenylephrine (10^{-5}-10^{-2} M) and finally, phenylephrine and caffeine 100 mM in Ca^{2+}-free Krebs were measured.

Results: KCl and phenylephrine-induced contractions were considerably lower in gestational hypothyroid (GH) than euthyroid offspring. GH responded to nifedipine with less sensitivity than control. The GH and control groups produced almost equal contraction in respond to phenylephrine and caffeine in Ca^{2+}-free Krebs.

Conclusion: This study suggests that in hypothyroid offspring L-type Ca^{2+} channels are less functional, while intracellular Ca^{2+} handling systems are less modified by low levels of maternal thyroid hormones.

Introduction

Physiological functioning of the body systems is highly influenced by intrauterine conditions in which the mammalian fetus develops. Suboptimal maternal environment, such as insufficient availability of nutrients, oxygen, and hormones can change the developmental regulatory planning of fetal tissue, lead to diseases, such as cardiovascular and metabolic diseases in adult life (1-3). Thyroid hormones play an essential role in fetal normal development (4, 5). T3 reduces the vascular resistance and therefore, causes relaxation especially in arteries. Cardiovascular system response to hypothyroidism is low cardiac output, pulse pressure and increased vascular resistance, while the opposite symptoms are in effect in thyroid overactivation (5-7). Furthermore, either in hyperthyroidism or hypothyroidism, the response of the vessels is changed to the vasoconstrictors and vasodilators agents (8, 9). Most studies have focused on measuring the responses to vasoconstrictors in adult models of thyroid deficiencies, while there is not much evidence available on the maternal or gestational models of these diseases on the offspring’s vascular status. In many studies, it has been demonstrated that adult models of hypothyroidism develop a defective vascular smooth muscle response to α-adrenergic agonists and KCl or barium chloride in comparison to euthyroids (10-16). However, one of the few studies that investigated the effects of prenatal hypothyroid model in rats on the density of adrenoceptors, reported a decrease in the myocardial α1-adrenergic receptor density within 15-28 days after birth, indicating the modulation of adrenergic receptor proteins by thyroid hormones during fetal development (17). More recently a study showed that an isolated aorta smooth muscle (without endothelium) of congenitally hypothyroid adult male offspring of a female rat, who received 6-Propyl-2-thiouracil (PTU) during pregnancy, demonstrated markedly less contractile response to vasoconstrictors in comparison to euthyroids (8). The contraction response to KCl and α1-adrenergic agonists in the vascular smooth
muscle is produced by extracellular Ca\(^{2+}\) influx, activated by voltage gated calcium channels (VGCC), non-voltage dependent Ca\(^{2+}\) influx (15, 18), and intracellular Ca\(^{2+}\) release from internal stores, such as sarcoplasmic reticulum (SR) (12, 15, 18). The markedly lower responses of hypothyroids to KCl and \(\alpha_1\)-agonists compared to controls, reported in previous studies suggest some impairments in function of calcium entering/releasing at the plasma membrane (through channels) (15, 19, 20) or in the intracellular systems (15, 21), respectively. There is evidence from previous studies showing that the expression and function of the VGCCs, including L-type Ca\(^{2+}\) channels, are under the control of thyroid hormones, via genomic and nongenomic mechanisms (20, 22-24). In this study we used a model of gestational hypothyroidism produced by dams who consumed PTU during pregnancy. Using PTU and methimazole as agents that induce thyroid deficiency in dams are valid models of inducing hypothyroid state in offspring, which has been used in several studies related to developmental pathophysiology of thyroid hormones (8, 25-31).

This study aimed to investigate whether gestational hypothyroidism in a rat model may modulate L-type Ca\(^{2+}\) channels functions in later adult life, determined by developing some deficiencies in aorta smooth muscle responses to the KCl and selective \(\alpha_1\)-adrenergic agonist, phenylephrine. Also, gestational thyroid hormones may induce changes in offspring intracellular Ca\(^{2+}\) handling machinery by stimulating aorta smooth muscle cells with the internal Ca\(^{2+}\) releasing vasoconstrictors, such as phenylephrine and caffeine, in a Ca\(^{2+}\)-free environment.

**Materials and Methods**

**Animals**

Female Wistar rats, 155-250 g (inbred in the Endocrine Physiology Research Center Animal Facilities) were used for mating in this study. The animals were kept in 12 hr light/dark cycle, 22±3°C temperatures and had free access to rat chow (Pars Co., Tehran) and water. Animal handling and experiments were carried out in accordance with the local ethics committee of the Research Institute for Endocrine Sciences, Shahid Beheshi University of Medical Sciences. Maximum efforts were made to minimize the animal's possible discomfort and stress.

**Induction of gestational hypothyroidism**

Hypothyroidism was induced in pregnant rats by adding 200 mg PTU (Sigma Aldrich, Germany) to 1000 ml of drinking water, from the first day of pregnancy until delivery (8, 31, 32). While the control group only received tap water, upon delivery the drug was removed from drinking water.

**Experimental groups**

Male offspring were weighed on days 0, 15, 30, 45, 60 and 150 of birth. Five months-old (150 days) rats were placed into the control (n=9) and gestational hypothyroid (GH) (n=10) groups and assessed for the contractile force measurement of the aorta.

**Thyroid hormone measurement**

To confirm the PTU-induced hypothyroidism, blood samples were drawn from the dam and neonates on delivery day and from the abdominal aorta of adult male offspring, five-months after birth. All the samples were centrifuged (3000 g, for 10 min in 4°C) and kept at -80°C until the time of the hormonal assay. T\(_3\) and T\(_4\) were measured using enzyme linked immunosorbent assay kits (Pishtaz Teb Zaman Co., Iran). Intra- and interassay coefficients of variation were 3.7 and 4.3% for T\(_3\) and 5.3 and 5.9% for T\(_4\), respectively.

**Aortic ring preparation**

Adult male offspring of GH and control groups were anesthetized by IP injection of 50 mg/Kg ketamine and 10 mg/Kg xylazine (16). The thorax was cut open and a section of the thoracic aorta ring was dissected and cleaned of the connective tissue and vessels in ice cold (4°C) Krebs-Henseleit solution (composition in mmol/l: NaCl 118, KCl 4.7, KH\(_2\)PO\(_4\) 1.2, MgSO\(_4\) 1.2, CaCl\(_2\) 2.5, Glucose 10, and NaHCO\(_3\) 25, (Merck, Germany)), pH adjusted to 7.4 and temperature to 37°C and the ring was cut into 5 mm sections in length. Endothelium was removed by gently rotating the ring around a metal wire inserted into the lumen (33). The Krebs solution inside the chamber, where the ring was hocked to an isometric sensitive force transducer (MLT0202, ADInstruments, Spain), was constantly gassed with a combination of 95% O\(_2\) and 5% CO\(_2\). Contraction force data were recorded by the sensitive force transducer coupled to the PowerLab data acquisition system (ML866 PowerLab 4/30, ADInstruments, Australia). Before the experiment began, the aorta ring was placed under 2.0 g resting tension and allowed to equilibrate for a period of 60 min, while the solution inside the chamber was renewed every 15 min. To determine the removal of the endothelium, acetylcholine (10\(^{-5}\) M) was added to pre-contracted aorta ring with phenylephrine (10\(^{-6}\) M). The ability of the tissue to relax in response to acetylcholine indicated the presence of endothelium (33, 34).

**Isolated aorta contraction experiments**

Range of KCl (10-100 mM) and phenylephrine (10\(^{-9}\), 10\(^{-6}\) M) concentrations, separated by refreshing
Table 1. Comparing the weights of male offspring in gestational hypothyroid and control groups in 0-150 days after birth

| Post-natal days | GH weight (g) | Control weight (g) | P-value |
|-----------------|--------------|--------------------|---------|
| 0               | 6.656 ± 0.579 | 7.957 ± 0.425      | 0.1026  |
| 15              | 18.48 ± 1.297 | 24.65 ± 1.176      | 0.0054*** |
| 30              | 47.44 ± 3.761 | 66.26 ± 2.416      | 0.0002*** |
| 45              | 99.00 ± 9.771 | 128.2 ± 3.690      | 0.0016*** |
| 60              | 139.1 ± 9.525 | 175.6 ± 5.674      | 0.0021*** |
| 150             | 296.87 ± 3.567| 311.33 ± 2.341     | 0.4866  |

Values are mean±SEM. ***P-values<0.001 vs compared with control. Number of animals per group is 15; GH: Gestational hypothyroidism.

Krebs solution and single concentrations of nifedipine (10⁻⁴, 10⁻³ μM) prior to KCl (10-100 mM) were applied to the ring tissue inside the chamber. In the second set of experiments, the Krebs solution was replaced by Ca²⁺-free Krebs (CaCl₂ was replaced by MgCl₂ 1.2 mmol/l). The range of phenylephrine concentrations (10⁻⁶, 10⁻⁵ M) was used to test the contractile force in Ca²⁺-free environment. Finally, after washing with Ca²⁺-free Krebs solution, caffeine 100 mM was added and the contractile force was recorded for 30 min in response to that. Nifedipine was dissolved in ethyl-alcohol (96%) with the stock concentration of (3×10⁻³ M). Caffeine (100 mM) was dissolved, while heating and stirring, in Ca²⁺-free Krebs (50 ml). Before applying the solution temperature was decreased to 37°C.

At the end of experiments the length and weight of the aorta ring were measured for calculating the tension produced by each of vasoconstrictor agents in the experiment normalized to cross-sectional area with the average tension displayed as g/mm², using the formula below (8):

Cross-sectional area (mm²) = weight (mg) / length (mm) × density (for the vascular smooth muscle is 1.05 mg/mm²)

Statistical analysis

Values were presented as mean±SEM. Unpaired Student t-test was performed to analyze the differences between the tensions produced in hypothyroid vs control group. Paired Student t-test compared the differences between the data obtained from groups before and after nifedipine administration and data related to contractions recorded with Ca²⁺-containing and Ca²⁺-free Krebs solutions. Statistical analysis and graph plotting was performed by GraphPad Prism version 4.00 for Window’s (GraphPad software, San Diego, California, USA).

Results

Weight differences

Male offspring weights in the GH and control groups were significantly different on days 15 (P-value<0.01), 30, 45 and 60 (P-value<0.001) after birth (Table 1).

Thyroid hormone measurement

serum levels of thyroid hormones, T₃ and T₄, in dams and neonate and adult offspring indicated a significant decrease in dams who had consumed PTU during pregnancy and GH neonates in comparison to their control groups (P-value<0.001). No differences were observed in GH adult offspring compared to controls (Table 2).

Aortic smooth muscle contraction in response to vasoconstrictors

KCl-induced contraction

KCl concentrations (10-100 mM) were used to produce a dose-response curve. In the GH group the amplitude of contractile responses to KCl at 60, 80, and 100 mM were significantly lower than controls (for 60 and 80 mM, P-value<0.01 and for 100 mM KCl, P-value<0.001). Average maximum tension was

Table 2. Levels of thyroid hormones (ng/dl) in dam, neonate (at birth) and adult offspring (at 5 months age)

| Hormones                  | Dam          | Offspring       |             |             |
|---------------------------|--------------|-----------------|-------------|-------------|
|                           | Control      | PTU consumed    | Neonate     | GH          | Control      | Adult         | GH          |
| 3,5,3 ’-Triiodothyronine (ng/dl) | 93.4 ± 3.8   | 51.7 ± 4.9***   | 62.9 ± 3.4  | 39.5 ± 4.3*** | 95.7 ± 4.3  | 87.7 ± 5.3    |
| Thryroxine (ng/dl)        | 2400 ± 0.2   | 520 ± 0.04***   | 730 ± 0.06  | 370 ± 0.04*** | 3800 ± 0.1  | 3400 ± 0.2    |

Thyroid hormones, 3,5,3 ’triiodothyronine (T₃) and thyroxine (T₄) (ng/dl) in dam and neonate and adult (5 months old) offspring. Values are mean±SEM (*** P-value<0.0001) compared with control; GH: Gestational hypothyroidism.
Group compared to control (0.717±0.06 for GH vs 0.981±0.02 for control. Mean±SEM, *P*-value <0.001) (Figure 1a).

**Effect of Nifedipine on KCl-induced contraction**
To investigate the effect of thyroid hormones on Ca²⁺ conductance through the membrane channels, nifedipine was used within the range of 10⁻¹⁰-10⁻⁷M, as separate doses against KCl (10-100 mM). Nifedipine, in a concentration dependent manner, reduced the contractile force of aorta smooth muscle in both GH and control groups (Figure 1b-e).

Generally, in control group, nifedipine with concentrations 10⁻⁹ M (*P*-value<0.05) and 10⁻⁸-10⁻⁷ M (*P*-value<0.001) inhibited the response to KCl 100 mM. In GH group, nifedipine with concentrations 10⁻⁸-10⁻⁷ M, inhibited the response to KCl 100 mM (*P*-value<0.001) (Figure 1f).

**Phenylephrine-induced contraction**
Phenylephrine (10⁻⁹-10⁻⁶M)-induced contractions were used to produce a dose-response curve. GH responses to phenylephrine (10⁻⁷, 10⁻⁸M) were noticeably lower than control (*P*-value<0.05 and *P*-value<0.01, respectively) (Figure 2a). However, in Ca²⁺-free Krebs solution, the contractile responses of both groups to phenylephrine were almost the same (Figure 2b). The responses of aorta smooth muscle to phenylephrine was significantly reduced, when...
shifting from the Ca$^{2+}$-containing to Ca$^{2+}$-free Krebs solutions ($P$-value=0.001). However, responses were the same for the control and GH groups in Ca$^{2+}$-free Krebs (Figure 2c).

**Caffeine-induced contraction**

Caffeine (100 mM) was added in a Ca$^{2+}$-free Krebs environment. Recording contraction from the aorta smooth muscle of control and GH groups for 30 min revealed no specific difference in contractile force between the two groups (0.317 ± 0.04, 0.271 ± 0.05; mean±SEM for control and GH groups, respectively; $P$-value=0.514) (Figure 3).

**Discussion**

In this study we tried to determine the role of thyroid hormones in developing functional calcium handling system in vasculature during fetal development to produce suitable physiological responses in adulthood. The aortic rings from the offspring of PTU consumed dams with denuded endothelial layer were prepared and their responses to vasoconstrictors were examined against those from euthyroid mothers. Our results from the initial GH and control groups responses to KCl and selective $\alpha_1$-adrenoceptor agonist, phenylephrine, were almost similar to those reported in previous studies of hypothyroid adult rats who received anti-thyroid agent or thyroidectomy against euthyroids (14, 16, 35, 36). In all the studies, the hypothyroid group (preparations with denuded endothelium) reaction to KCl and $\alpha_1$-adrenoceptor agonists was markedly reduced in comparison to euthyroids (8, 14, 35). In our study, the marked lower responses of GH to KCl and phenylephrine, relative to controls, suggests some defects in the functioning of Ca$^{2+}$ entering through Ca$^{2+}$ channels or release from intracellular resources.

In an attempt to determine the possible sites of Ca$^{2+}$ handling dysfunction in the GH aorta smooth muscle cells, we first used nifedipine, as a VGCC (L-type Ca$^{2+}$ channel) blocker. Before nifedipine administration, the response of the control group to KCl was markedly higher than the GH response. Adding increasing concentrations of nifedipine made the contractile force induced by KCl in control group closer to GH, until both groups reached to almost equal contractile forces at the nifedipine dose of $10^{-7}$M. The stronger response to KCl and nifedipine inhibitory effect in euthyroids compared to GH suggest some displacement or dysfunction in the plasma membrane L-type Ca$^{2+}$ channels of the latter group, which may possibly be related to the reduced thyroid hormone levels in fetal life. Many previous studies have shown that the
expression and function of the VGCCs are controlled by thyroid hormones, via genomic and nongenomic mechanisms (20, 22-24). Investigating the effect of T₃ on the slow Ca²⁺ channel function in cultured chick ventricular cells demonstrated an increase in slow channel Ca²⁺ influx and Ca²⁺ channel antagonist binding sites under the influence of T₃ in comparison to cells grown in environment without T₃ (22). There are some other reports indicating the increase in the Ca²⁺ currents by VGCC in aortic ring tissues (12) and rat ventricular cells along with reduction in expression of α₁C-subunit of VGCC (genomic regulation) (20), or increase in L-type Ca²⁺ channel current and mRNA expression in hyperthyroid rabbit myocytes (24), while not affecting the Ca²⁺ release from SR in all of them. In contrast to previous reports, another study conducted on rat cardiac and vascular tissue demonstrated a decrease in the Ca²⁺ channel density in the hyperthyroid versus hypothyroid rat model (23). Most of the previous results related to the smooth muscle of cardiac vessels indicated that increase in thyroid hormones above the physiological levels would increase influx through the L-type Ca²⁺ channels, possibly by increasing the number of protein channels or the Ca²⁺ current; our data revealed the same event, through a reverse mechanism. Gestational hypothyroidism caused markedly weaker KCl-induced contractile force compared to euthyroid offspring and lower response to KCl in the presence of maximum concentration of nifedipine, indicating that it is likely a scarcity of thyroid hormones during fetal life causing the reverse mechanistic profile to hyperthyroidism, most likely through the genomic regulatory mechanism, affecting the Ca²⁺ channel number or conductivity.

High concentrations of selective α₁-agonist, phenylephrine (10⁻⁷-10⁻⁶ M), produced significantly weaker contractions in the GH compared to control. This was similar to previous results obtained from comparable data of hypothyroid vs euthyroid adult rats that received α₁ agonist agents (14, 16, 35, 36). In our study, while the contraction responses to higher concentrations of phenylephrine were considerably higher for controls compared to GH, both groups’ responses significantly reduced in Ca²⁺ -free Krebs solution and became almost equal.

Experimentally, the activation of α₁-adrenoceptors produces an increase in the intracellular [Ca²⁺], via two routes; opening the plasma membrane Ca²⁺ channels and releasing Ca²⁺ from internal sources. Plasma membrane Ca²⁺ channels are either VGCCs or non-voltage dependent Ca²⁺ channels, including receptor-operated calcium channels (ROCCs), such as G-protein-coupled receptors GPCRs, like, α₁-adrenoceptor-operated calcium channels and store-operated channels (SOCs) (34, 36, 37) (both are usually blocked by Ni²⁺) (15, 19). It has been shown that aorta smooth muscle contraction induced by α₁-adrenoceptors is highly dependent on Ca²⁺ entry from extracellular environment. A study showed that nimodipine, strongly inhibit contraction force produced by noradrenaline, almost similar to the inhibitory effect of prazocine on aorta smooth muscle contraction (15), implying the joint effect of α₁-adrenoceptor to voltage and non-voltage dependent Ca²⁺ channels for replenishing intracellular [Ca²⁺] and producing contraction.

In the scarcity of extracellular Ca²⁺, an α₁-agonist, such as phenylephrine depletes the internal Ca²⁺ sources, which the contractile force would be entirely dependent on. In our study, while there was a significant difference in control and GH responses to phenylephrine in the presence of ample extracellular Ca²⁺, their responses to the same stimulant became similar in Ca²⁺-free Krebs. The change in the responses relative to the presence and the absence of extracellular [Ca²⁺], suggests that low thyroid hormone levels during fetal life does not modify the intracellular Ca²⁺ releasing systems in GH, while strongly affects gene expression, protein structure or function of L-type Ca²⁺ channels. Therefore, while GH aorta cells are still dependent on normal extracellular [Ca²⁺] to produce contraction in response to α₁-adrenergic agonist vasoconstrictors, they are less sensitive (35, 36) in their responses to them in comparison to euthyroids. Furthermore, in Ca²⁺-free Krebs, the caffeine-induced contraction force was almost the same for both groups. In Ca²⁺-free environment, caffeine acts on ryanodine receptors to release Ca²⁺ from internal sources (11, 15, 38), a result which likely supports those mentioned above in the way that internal Ca²⁺ - releasing machinery is least changed by low thyroid hormones during fetal life.

Another point to consider in this study was the normal levels of thyroid hormones (T₁ and T₃) in the GH adult rats compared to controls. Therefore, the differences in the contractile responses observed between adult GH and euthyroids may have been stemmed from genetic modifications in the structure and function of the Ca²⁺ channels. These modifications are initiated in the prenatal or early postnatal periods of life and developed in adulthood, during which thyroid hormones began to reach to their normal physiological levels.

This study provides new insights towards deeper understanding of the changes in the vascular reactions in gestational hypothyroidism, by emphasizing on the role of modified calcium channels in producing contractions. However, it still lacks the mechanistic view of the underlying reasons, which can be gained through precise measurements of changes at the genes or/protein levels of the Ca²⁺ channels, which warrants to be considered in the future.
Conclusion

The contractile responses to vasoconstrictors in adult hypothyroid rats as well as fetal or GHs are considerably reduced in comparison to euthyroids. Our results suggest that lower levels of thyroid hormones during pregnancy affects Ca\textsuperscript{2+} handling system by reducing the Ca\textsuperscript{2+} conductance through plasma membrane with least modification in internal Ca\textsuperscript{2+} releasing/storing systems. Obtained results which may have some valuable clinical relevance in the future on treatment options for cardiovascular problems of offspring born to hypothyroid mothers.

Acknowledgment

This study was supported by a grant (No. 497) from Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Authors would like to thanks from Mahyar Baveisi and Fatemeh Bagheripour for their technical help.

Conflict of interest

The authors declare that they have no conflict of interest related to this manuscript.

References

1. Bertram CE. Animal models and programming of the metabolic syndrome. Br Med Bull 2001; 60:103-121.
2. Fowden AL, Forhead AJ. Endocrine mechanisms of intrauterine programming. Reproduction 2004; 127:515-26.
3. Fowden AL, Giussani DA, Forhead AJ. Intrauterine programming of physiological systems: Causes and consequences. Physiology 2006; 21:29-37.
4. Zahedi S. Importance of thyroid hormones in intrauterine programming. Int J Endocrinol Metab 2010; 8:186-7.
5. Patel J, Landers K, Li H, Mortimer RH, Richard K. Delivery of maternal thyroid hormones to the fetus. Trends Endocrinol Metab 2011; 22:164-70.
6. Danzi S, Klein I. Thyroid hormone and the cardiovascular system. Med Clin North Am 2012; 96:257-68.
7. van Tuyl M, Blommaart PE, de Boer PA, Wert SE, Ruijter JM, Islam S, et al. Prenatal exposure to thyroid hormone is necessary for normal postnatal development of murine heart and lungs. Dev Biol 2004; 272:104-17.
8. Khalalsari M, Shafiee M, Ghasemi A, Asl SZ. Effect of orally administered propylthiouracil in pregnant and lactating rats on isolated aorta contractility of their adult male offspring. Med Sci Monit 2009; 15:BR123-7.
9. Sutendar M, Garcia-Bournissen F, Koren G. Hypothyroidism in pregnancy. J Obstet Gynaecol Can 2007; 29:354-6.
10. Sabio JM, Rodriguez-Maresca M, Luna JD, Garcia del Rio C, Vargas F. Vascular reactivity to vasoconstrictors in aorta and renal vasculature of hyperthyroid and hypothyroid rats. Pharmacology 1994; 49:257-64.
11. Sanders KM. Invited review: mechanisms of calcium handling in smooth muscles. J Appl Physiol 2001; 91:1438-49.
12. Stratton DB, Morrow RJ. Calcium mobilization and sensitivity in intact and triton skinned aorta from thyropathologic rats. Life Sci 1991; 48:2207-14.
13. Rahmani MA, Cheema IR, Sen S, Peoples B, Riley SR. The effect of hyperthyroidism and hypothyroidism on alpha 1- and alpha 2-adrenergic responsiveness in rat aortic smooth muscle. Artery 1987; 14:362-83.
14. Pantos C, Mourouzis C, Katramadou M, Saranteas T, Mourazis I, Karageorgiou H, et al. Decreased vascular reactivity to alpha1 adrenergic stimulation in the presence of hypothyroid state: a part of an adaptive response? Int Angiol 2006; 25:216-20.
15. Gisbert R, Perez-Vizcaino F, Cogolludo AL, Noguera MA, Ivorra MD, Tamargo J, et al. Cytosolic Ca\textsuperscript{2+} and phosphoinositide hydrolysis linked to constitutively active alpha 1D-adreceptors in vascular smooth muscle. J Pharmacol Exp Ther 2003; 305:1006-14.
16. Brown L, Nankervis R, Kerr D, Sernia C. Adrenoceptor-mediated cardiac and vascular responses in hypothyroid rats. Biochem Pharmacol 1994; 47:281-8.
17. Noguchi A, Whitsett JA. Ontogeny of alpha 1-adrenergic receptors in the rat myocardium: effects of hyperthyroidism. Eur J Pharmacol 1982; 86:43-50.
18. Zhong H, Minneman KP. Alpha1-adrenergic subtypes. Eur J Pharmacol 1999; 375:261-76.
19. Guibert C, Ducret T, Savineau JP. Voltage-independent calcium influx in smooth muscle. Prog Biophys Mol Biol 2008; 90:10-23.
20. Watanabe H, Washizuka T, Komura S, Yoshida T, Hosaka Y, Hatada K, et al. Genomic and non-genomic regulation of L-type calcium channels in rat ventricle by thyroid hormone. Endocr Res 2005; 31:59-70.
21. Vetter R, Rehfeld U, Reissfelder C, Fechner H, Seppet E, Kreutz R. Decreased cardiac SERCA2 expression, SR Ca uptake, and contractile function in hypothyroidism are attenuated in SERCA2 overexpressing transgenic rats. Am J Physiol Heart Circ Physiol 2011; 300:H943-50.
22. Kim D. Effect of Thyroid Hormone on slow Calcium channel function in cultured chick ventricular cells. J Clin Invest 1987; 80:88-94.
23. Hawthorn MH. Effect of thyroid status on beta-adreceptors and calcium channels in rat cardiac and vascular tissue. Naunyn Schmiedebergs Arch Pharmacol. 1988; 337:539-544.
24. Yu Z, Wang T, Xu L, Huang CX. Thyroid hormone increased L-type calcium channel mRNA expression and L-type calcium current of myocytes in rabbits. Biomed Mater Eng 2012; 22:49-55.
25. Narayanan CH, Narayanan V, Browne RC. Effects of induced thyroid deficiency on the development of sucking behavior in rats. Physiol Behav 1982; 29:361-70.
26. Koromilas C, Liapi C, Zarros A, Stolakis V, Tsaigiani A, Skandali N, et al. Effects of experimentally-induced maternal hypothyroidism on crucial offspring rat brain enzyme activities. Int J Dev Neurosci 2014; 35C:1-6.
27. Goodman JH, Gilbert ME. Modest thyroid hormone insufficiency during development induces a
cellular malformation in the corpus callosum: a model of cortical dysplasia. Endocrinology 2007; 148:2593-7.
28. Gilbert ME, Sui L, Walker MJ, Anderson W, Thomas S, Smoller SN, et al. Thyroid hormone insufficiency during brain development reduces parvalbumin immunoreactivity and inhibitory function in the hippocampus. Endocrinology 2007; 148:92-102.
29. Ahmed OM, Abd El-Tawab SM, Ahmed RG. Effects of experimentally induced maternal hypothyroidism and hyperthyroidism on the development of rat offspring: I. The development of the thyroid hormones-neurotransmitters and adenosinergic system interactions. Int J Dev Neurosci 2010; 28:437-54.
30. Amerion M, Tahajjodi S, Hushmand Z, Mahdavi Shahri N, Nikravesh MR, Jalali M. The effect of maternal thyroid disorders (hypothyroidism and hyperthyroidism) during pregnancy and lactation on skin development in wistar rat newborns. Iran J Basic Med Sci 2013; 16:665-74.
31. Karbalaei N, Ghasemi A, Hedayati M, Godini A, Zahediasl S. The possible mechanisms by which maternal hypothyroidism impairs insulin secretion in adult male offspring in rats. Exp Physiol 2014; 99:701-14.
32. Ghasemi A, Mehrazin F, Zahediasl S. Effect of nitrate and L-arginine therapy on nitric oxide levels in serum, heart, and aorta of fetal hypothyroid rats. J Physiol Biochem 2013; 69:751-9.
33. Weber LP, Chow WL, Abebe W, MacLeod KM. Enhanced contractile responses of arteries from streptozotocin diabetic rats to sodium fluoride. Br J Pharmacol 1996; 118:115-22.
34. McAllister RM, Grossenbarg VG, Delp MD, Laughlin MH. Effects of hyperthyroidism on vascular contractile and relaxation responses. Am J Physiol 1998; 274:E946-53.
35. Gunasekera RD, Kuriyama H. The influence of thyroid states upon responses of the rat aorta to catecholamines. Br J Pharmacol 1990; 99:541-7.
36. Vargas F, Moreno JM, Rodriguez-Gomez I, Wangensteen R, Osum A, Alvarez-Guerra M, et al. Vascular and renal function in experimental thyroid disorders. Eur J Endocrinol 2006; 154:197-212.
37. McFadzean I, Gibson A. The developing relationship between receptor-operated and store-operated calcium channels in smooth muscle. Br J Pharmacol 2002; 135:1-13.
38. Karaki H, Ozaki H, Hori M, Mitsui-Saito M, Amano K, Harada K, et al. Calcium movements, distribution, and functions in smooth muscle. Pharmacol Rev 1997; 49:157-230.