Abstract

The present article describes a comparative study on the cellular reactions in capillary and venous blood in northerners under general hypothermia in a climatic chamber during different photoperiods. The authors examined 108 relatively healthy people living in Archangelsk (80 women and 28 men aged 21 to 50 years old). It was established that total neutrophil count in venous blood was lower by 8.07±0.41%, monocyte count – by 51.32±1.03% and basophil count – by 50.21±1.24% than in capillary blood, but the lymphocyte count was higher by 25.23±0.41% due to smaller forms that are known to be recirculating. After a 5-minute period in a climatic chamber (“USHZ-25N”, Russia) at -25º, 25 volunteers (27.53%) during a polar night and 16 volunteers during a polar day had elevated levels of neutrophils in the venous blood due to the increase in the levels of TNF-α in blood and decrease in noradrenaline, adrenaline, and irisin. During a summer period, the increase in monocyte count contributed to the reactions of neutrophils. The volunteers that reacted to short-term general hypothermia with an increase in the neutrophil count had the following peculiarities in the venous blood: general neutrophil count and their phagocytic activity, lymphocyte count, including CD71+, ATP in lymphocytes, adrenaline, TNF-α, and irisin levels were higher in blood serum. The content of endothelin-1 did not change significantly.

Keywords: Cold; photoperiod; venous blood; capillary blood; neutrophils; lymphocytes; monocytes; cytokines; irisin.
I. Introduction

Nearly all the known studies on the composition of blood investigated venous blood. Functional activity of blood cells is observed primarily in tissues. There are data that venous and capillary blood is not identical in the rate of circulation, coagulation, erythrocytes aggregation and rheological properties [XLIX, XL, XLII, XLVI]. The data on the leucocyte pool in capillary and venous blood is sparse and controversial. The studies by Mingacheva et al (2009) showed that the level of leucocytes was lower and the level of erythrocytes, thrombocytes, and hemoglobin was higher in venous blood than in capillary blood [XXXV]. On the contrary, the study by Ledinkina et al (2016) showed that the level of leukocytes and neutrophils was 8% higher, platelets – 9% higher, and monocytes – 12% higher in venous blood [XXXII]. Despite numerous studies and published data, the issue on the different content of leucocytes and their certain forms in capillary and venous blood is not solved, and the majority of researchers believe that these differences within the physiological norm are insignificant.

The most expressed differences are observed at low temperatures of the skin [XLIV, XVI]. Local hypothermia leads to the degranulation of tissue basophils and mediator-induced edema of derma, which results in the infiltration of the tissue with mononuclear cells, neutrophils, eosinophils and damage of endothelium with the accumulation of the immune complexes [XLIII]. The first reaction to general hypothermia includes a wide range of changes in general hemodynamics: the transformation of microcirculatory bloodstream with an increase in the tonus of precapillary arterioles and a decrease in the activity of the venous blood output, increase in the tonus of magistral vessels and linear rate of the bloodstream. The data on the influence of general hypothermia on the contractility of the myocardium and parameters of cardiac output that maintain central circulation are controversial: from the maintenance of high systemic blood pressure, heart rate and minute cardiac output to the reduction of the minute cardiac output and decrease in the heart rate. All these changes depend on the duration of the cold factor impact or adaptation of a human to cold conditions, probably, due to individual sensitivity to hypothermia. It is known that a dynamic equilibrium is established between circulating and parietal cells, which is constantly shifting to the increase or decrease of its compounds [XXXIX, XIX, XXXVIII].

Chemotactic signals and specific intercellular interactions that can be disturbed in hypothermic conditions play the main role in the determination of the migration direction and overcoming of the barriers between the blood and tissues. The immune response to general hypothermia includes a decrease in the phagocytic activity of neutrophils and monocytes in blood and the levels of activated cells and antibody production. Under the influence of cold conditions and in the winter period, qualitative and quantitative parameters of the cellular immunity are characterized by a decrease in T-helpers and T-suppressors by 10-15% and a general decrease in the functional activity of T-lymphocytes [XIV, XLI, IV]. Urgent systemic adaptation of a
human to any unfavorable impact includes a reaction of vasomotor amines, catecholamines, that provide the alterations in the vascular permeability, activation of the system of blood circulation, heart rate and vascular tonus.

An extreme factor for people who live in the North is the expressed photoperiodicity. The peak of biological darkness is observed in December and January. November and February are considered biological dusk and the excess of visible light (polar day) is observed in June and July. The rhythm of physiological functions associated with the change in the regimen of day and night alters during the period of light insufficiency: the rate of metabolism, the activity of the thermoregulation, breathing, blood circulation and higher nervous activity. The lack of light excludes a natural pathway for Vitamin D, which leads to the disturbances of phosphor-calcium metabolism. In the North of the RF, a sharp change of the climatic parameters is observed during 292 days per year. High repeatability (up to 316 days per year) of discomfort types of weather is observed. A heat discomfort with a tension of mechanisms of thermoregulation is observed nearly all year round, which leads to a significant decrease in the labor productivity up to 200% and an increase in the morbidity rate. Sharp fluctuations of temperature and photoperiodicity increase the energy consumption necessary for the maintenance of homeostasis, reduces its reserves and periods of active performance [XIV, LI].

The aim of the present work was to perform a comparative study of the cellular reactions in the capillary and venous blood in the northerners under general hypothermia in a climatic chamber. The authors suggested that short-term general hypothermia can alter the perfusion of myeloid cells in tissues and recirculation of lymphocytes by the enhancement of the production of cytokines and (or) vasomotor amines. The practical value of the work is in the application of the criteria of individual sensitivity of a human to general hypothermia.

II. Materials and Methods

The study included 108 relatively healthy people who lived in Archangelsk (80 women and 28 men aged 21 to 50 years old). 63 volunteers were examined during the minimum photoperiod (January, February) and 45 volunteers - during a polar day.

All the participants signed the form of informed consent. The study was performed according to the requirements of the Helsinki Declaration (2000).

The authors took samples of venous and capillary blood from the IV finger of the examined volunteers before and after 5-minute hypothermia at -25°C in climatic chamber USHZ-25N (Russia) during a minimum duration of a photoperiod (January) and polar day (June).

In venous and capillary blood, the authors identified hemogram with an automatic hematologic analyzer XS-500i (Sysmex, Japan) and in blood smears that were Romanowsky-Giemsa stained. Neutrogram was calculated by the method of Todorov (1968), monocytogram – by the method of Grigorova (1956),
lymphocytogram – by the method Kassirskiy (1970), and phagocytic activity of neutrophil granulocytes was estimated in peripheral blood. The concentration of irisin, endothelin-1, cytokine TNF-α, adrenaline and noradrenaline in blood was identified by the method of enzyme-linked immunoassay with an automatic immunoenzymometric analyzer “Evolis” (“Bio-RAD”, France); the content of lymphocyte phenotypes (CD3+, CD4+, CD8+, CD10+, CD16+, CD23+, CD25+, CD71+, CD95+, HLA DR+) – with an indirect immune peroxidase reaction with monoclonal antibodies (“MedBioSpectr”, “Sorbent”, Moscow) and the method of flow cytometry with an apparatus Epics XL (“Beckman Coulter”, USA), chemical reagents “Immunotech a Beckman Coulter Company” (France). The changes in the levels of ATP in lymphocytes were evaluated with a luminometerLUM-1 (“Lumtech”, Russia) and a standard chemical reagent kit “Lumtech”.

Mathematical analysis of the obtained results was performed with a software package «Microsoft Excel 2010» and «Statistica 7.0» («StatSoft», USA). The distribution of the values of immunological parameters was checked with Pierson’s test. Null hypothesis on the equality of all the means in the studied groups was checked with a one-factor dispersion analysis. When the distribution was abnormal, the comparison of two different groups by quantitative parameters was performed by non-parametric Mann-Whitney test. The authors calculated the parameters of the descriptive statistics (М – mean arithmetic, σ–standard deviation, m – standard error of the mean, Md – median, R – range, W – variation coefficient, CI 95%). A critical level of significance (p) was taken as 0.05.

III. Results

A comparative analysis of the cellular composition of venous and capillary blood in the examined volunteers before the climatic chamber showed that venous blood had lower content of neutrophils by 8.07±0.41%, primarily due to segmentonuclear cells with 2-3 segments of nuclei (from 1.02±0.06 to 0.85±0.05×10^9 cells/L) and (from 1.34±0.06 to 1.24±0.06×10^9 cells/L), respectively. The concentration of monocytes in venous blood decreased by 51.32±1.03% due to promonocytes (from 0.34±0.04 to 0.16±0.02×10^9 cells/L), mature monocytes (from 0.30±0.02 to 0.16±0.01×10^9 cells/L) and polymorphonuclear cells (from 0.12±0.01 to 0.05±0.004×10^9 cells/L). A decrease in basophils by 50.21±1.24% was also registered. The content of lymphocytes in venous blood was significantly higher (by 25.32±0.41%), including their smaller forms (from 1.10±0.06 to 1.42±0.08×10^9 cells/L) that are known to be recirculating (Table 1).
Table 1: Comparison data of the cellular composition of venous and capillary blood in northerners before a climatic chamber session (M±m)

| Cells, 10^9 cells/L | Capillary blood | Venous blood |
|---------------------|----------------|--------------|
| 1                   | 2              | 3            |
| Leukocytes          | 5.88±0.19      | 5.80±0.19    |
| Neutrophils         | 3.22±0.13      | 2.96±0.14*   |
| Banded neutrophile  | 0.16±0.01      | 0.23±0.02*** |
| Segmentonuclear neutrophils | 3.06±0.01 | 2.73±0.13*** |
| Monocytes           | 0.76±0.06      | 0.37±0.03*** |
| Eosinophils         | 0.11±0.01      | 0.12±0.01    |
| Basophiles          | 0.04±0.01      | 0.02±0.003** |
| Lymphocytes         | 1.74±0.07      | 2.33±0.11*** |

Note: * - p<0.05, ** - p<0.01, *** - p<0.001, during a comparison of capillary and venous blood in the examined patients.

The authors suggest that the decrease in the content of neutrophils, monocytes, and basophils in venous blood in comparison with capillary blood was observed due to perfusion of cells to tissues, and the increase in the content of lymphocytes in venous blood was observed due to a possible recirculation of lymphocytes. The lack of a clear difference in monocyteograms of capillary and venous blood could indicate the ability of monocytes to recirculate.

After a 5-minute hypothermia session in a climatic chamber at -25ºC, 25 volunteers (27.53%) during a polar night and 16 volunteers during a polar day (16.51%) an increased content of neutrophils in capillary and venous blood (from 3.41±0.20 to 4.26±0.35x10^9 cells/L; p<0.001) and (from 2.70±0.17 to 3.02±0.22x10^9 cells/L; p<0.001), including segmentonuclear cells (from 3.25±0.25 to 4.11±0.33x10^9 cells/L; p<0.001) and (from 2.46±0.16 to 2.76±0.21x10^9 cells/L; p<0.001). No changes were observed in the content of eosinophils (0.11±0.02 and 0.10±0.02x10^9 cells/L) and (0.11±0.01x10^9 cells/L) and basophils (0.01±0.001 and 0.01±0.001x10^9 cells/L) and (0.02±0.01 and 0.03±0.01x10^9 cells/L). Thus, the direction of the reactions of the changes in the content of neutrophils, eosinophils, and basophils to hypothermia in summer and winter was similar: under the influence of hypothermia, the activity of the perfusion of myeloid cells in tissues decreased. A decrease in the level of lymphocyte recirculation (from 1.76±0.13 to...
1.40±0.11x10^9 cells/L; p<0.01) and (from 1.95±0.11 to 1.65±0.10x10^9 cells/L; p<0.01), primarily of smaller forms (from 1.19±0.10 to 1.00±0.09x10^9 cells/L; p<0.01) and (from 1.55±0.11 to 1.18±0.08x10^9 cells/L; p<0.001), were registered only during a polar night.

One of the objectives was to reveal if there were changes in the functional activity of lymphocytes in venous blood after a session in a climatic chamber (Table 2). The obtained data showed that the decrease in the recirculation of lymphocytes was associated with a significant decrease in the functional activity of all the studied phenotypes of lymphocytes.

**Table 2**: Content of lymphocyte phenotypes in the peripheral venous blood in patients that react to general hypothermia before and after climatic chamber during a polar night (M±m)

| Parameters, 10^9 cells/L | Before a session in a climatic chamber | After a session in a climatic chamber |
|--------------------------|----------------------------------------|--------------------------------------|
| Lymphocytes              | 1.95±0.11                              | 1.65±0.10**                          |
| CD3+                     | 0.90±0.04                              | 0.40±0.02***                         |
| CD25+                    | 0.40±0.03                              | 0.24±0.02***                         |
| CD71+                    | 0.40±0.03                              | 0.22±0.02***                         |
| HLA-DR II                | 0.39±0.03                              | 0.24±0.02***                         |
| CD10+                    | 0.36±0.03                              | 0.18±0.02***                         |
| CD4+                     | 0.44±0.04                              | 0.20±0.02***                         |
| CD8+                     | 0.39±0.04                              | 0.17±0.01***                         |
| CD16+                    | 0.38±0.03                              | 0.16±0.02***                         |
| CD23+                    | 0.33±0.03                              | 0.15±0.02***                         |
| CD95+                    | 0.28±0.02                              | 0.14±0.02***                         |

Note: ** - p<0.01, *** - p<0.001 during a comparison of capillary and venous blood in the examined patients.

Volunteers that reacted to a short-term hypothermia, before a session in a climatic chamber, had higher content of neutrophils; their phagocytic activity and the concentration of lymphocytes (2.46±0.13 and 1.70±0.15x10^9 cells/L; p<0.001), including T-cells with receptors to transferrin (0.40±0.03 and 0.29±0.02x10^9 cells/L;
p<0.01), was higher. These volunteers had higher content of ATP in lymphocytes (2.48±0.17 and 0.88±0.13 µmol/ml n cells; p<0.001), TNF-α (2.12±0.34 and 1.23±0.27 pg/ml; p<0.01), irisin (5.60±0.70 and 3.74±0.55 µg/ml), and adrenaline (53.03±7.29 and 33.27±4.07 pg/ml) in blood serum without significant changes in the content of endothelin-1 (0.75±0.11 and 0.77±0.18 fmol/ml).

IV. Discussion

Functional activity of blood cells was observed primarily in tissues, where the content of monocytes was 51.32±1.03%, basophils – 50.21±1.24%, and neutrophil granulocytes 8.07±0.41%. The concentration of lymphocytes in venous blood was higher than in capillary by 25.32±0.41% due to smaller forms that are believed to represent the majority of recirculating cells. Recirculating lymphocytes are small, primarily T-cells of the reserve pool, that are capable of further blast-cell transformation and differentiation [II, IX, XXXVII, XV, XXXVI].

The change in the ratio of circulating and parietal pools is the main signal for hemodynamic reaction. Microvasculature is a system of transport bloodstream, its functional condition changes depending on the status of tissues that provide these areas with blood [V, X, XX, XXIX, XXIX, XLVII, VI, XVIII]. Functional condition of the microvasculature is provided by numerous regulatory mechanisms of endothelial origin with secretion of nitrogen oxide and vasoconstrictor endothelin-1, neurogenic sympathetic activity, myogenic mechanisms (peptide ergic and endogenous), as well as pulse and respiratory oscillations. The fastest mechanism of regulation of microvasculature is endothelial. The deficit of endothelial-dependent vasodilation is caused by a shift in the balance of the synthesis of nitrogen oxide and vasoconstrictors to the domination of vasoconstrictors, primarily, endothelin-1 [VIII, XVII]. The main depot of neutrophils is located in the capillary net of lungs. Lungs have nearly unlimited depositing capacity reserving primarily active granulocytes [VII]. Lymphocytes are different by their capability to recirculation. It is suggested that monocytes are capable of recirculation; the tissue pool of monocytes exceeds the content of tissue neutrophils by 3.5 times [XXXIV].

Migration and perfusion of cells are provided by a significant reduction of the bloodstream rate in the capillary net, which is observed in hypothermic conditions. A cell sticks to the capillary walls with its further migration outside the vascular net [II, IX, XXXVII]. More than that, molecules of adhesion provide a selection of cells that stick to the capillary walls. In problem areas, adhesive molecules of endothelium bind leukocytes and transmit a signal required for the transendothelial migration, which is regulated by the interaction of CD47 and signal regulatory protein (SIRPγ) of leukocytes. Binding of ligands with CD47 initiates signaling from endothelial cells which contributes to transendothelial migration [III]. A change in the activity of endothelial cells under the influence of inflammatory cytokines is accompanied by a formation of gaps and an increase in the permeability of cells [XII].
Hypothermia causes degranulation of tissue basophils and edema of the derma due to the effect of mediators, which leads to the infiltration of the tissue with mononuclear cells, neutrophils, eosinophils and the damage of endothelium with the accumulation of the immune complexes [XLIII]. The products of lipid peroxidation, that are actively involved in numerous metabolic processes, affect vascular endothelial cells causing vasoconstriction and damaging cellular and subcellular membranes, which leads to the disturbances in the processes of capillary trophic and gas exchange [XXX, XXXI]. A number of authors report that local hypothermia leads to the increase in the level of catecholamines, corticosteroids, and histamine [XXIV, XXII, XXVI, XXVII, XXIII, XXV, XXI, L].

In the majority of the volunteers, short-term hypothermia did not influence the content of leukocytes, the activity of their migration to tissues and the level of recirculation of lymphocytes. The reaction of inhibition of migration activity to general hypothermia was registered in 20.52% of volunteers with a significant increase in the rate of reactions in winter (27.53 and 16.51%, respectively). It was established that people with a reaction of inhibition of migration activity of leukocytes had some background peculiarities in the performance of the studied parameters. Volunteers that reacted to short-term hypothermia, had a higher content of neutrophils, their phagocytic activity, concentration of lymphocytes, including T-cells with a receptor to transferrin, before a session in a climatic chamber. These volunteers had a higher content of ATP in lymphocytes, TNF-α, irisin, and adrenaline in blood serum without significant changes in the concentration of endothelin-1. Increase in the energy resources can be provided by the increase in the production of TNF-αand irisin, as well as additional transport of iron to cells. TNF-αplays an important role during the first minutes of the development of the inflammatory reaction because it activates endothelium and contributes to the expression of adhesive molecules, which makes granulocytes stick to the internal surface of a vessel. Under the influence of TNF-α, transendothelial migration of leukocytes to the focus of inflammation is observed. This cytokine activates granulocytes, monocytes, lymphocytes and induces the production of other anti-inflammatory cytokines IL-1, IL-6, IFN, GM-CSF that act as synergists of TNF-α [XXXIII]. There are data that confirm the stimulation of recirculation of lymphocytes with the present cytokine [XIII].

It is known that adrenaline enhances tissue exchange, inhibits the expression of histamine, serotonin, and kinins, which can inhibit the perfusion of cells via membranes. Probably, an increase in the level of adrenaline and a lack of the reaction of endothelin-1 are the main stages in a chain of reactive processes that regulate the reactions of migration of leukocytes to hypothermia.

V. Conclusion

Thus, it is suggested that the mechanism of homeostasis maintenance is based on the principle of simultaneous stimulation and inhibition of regulatory reactions.
The increase in the production of TNF-α, irisin, and ATP, functional activity of neutrophil granulocytes and T-lymphocytes stimulate cellular migration to tissues and recirculation of lymphocytes, and adrenaline and endothelin-1 stabilize this process. Endothelin is one of the most potent vasoconstrictors. Apart from the vasoconstrictive effect, it enhances the production of cytokines [XI, I]. The expression of preendotheline-1 and the release of active peptide stimulate different humoral factors (angiotensin II, interleukin-1, adrenaline, noradrenaline, TNF-αvasopressin, thrombin). Thus, the controversy of the tissue bloodstream response to hypothermia is determined by a different initial condition of the functioning of the microvasculature.

References

I. Agui T., Xin X., Cai Y., Sakai T.; Matsumoto K. Stimulation of interleukin-6 production by endothelin in rat bone marrow-derived stromal cells - Blood. 1994; 84: 2531-2538.

II. AmbrusC.M., AmbrusL.L. Regulation of the leukocyte level - Ann.N.Y.Acad. Sci. 1959; 77: 445-479.

III. Azcutia V., Stefanidakis M., Tsuboi N., Mayadas T., Croce K.J., Fukida D. Aikawa M., Newton G., LuscinskasF.M. Endothelial CD47promotesvascularendothelial-cadherintein phosphorylation andparticipatesinTcellrecruitmentat sites of inflammationinvivo - J. Immunol. 2012; 189 (5): 2553-2562.

IV. BalashovaS.N., PashinskayaK.O. Influence of background neutropenia on the immune reactions in relatively healthy patients after short-term cooling. Journal of Ural Medical Academic Science. 2018; 15 (2): 239-247.

V. Barkhatov I.V. Application of laser Doppler flowmetry for the evaluation of disorders in the system of human blood microcirculation. Kazan Medical Journal. 2014; 95 (1): 63-69.

VI. BergstrandS., LindbergL.G., EkA-C., Lindén M., Lindgren M. Blood flow measurement at different depths using photoplethysmography and laser Doppler techniques - Skin. Res. Technol. 2009; 15: 139-147.

VII. Braynsteiner H. Physiologie und Physiopathologie der weissen Blutzellen. Stuttgart, 1959.

Copyright reserved © J. Mech. Cont. & Math. Sci.
Liliya K. Dobrodeeva et al
VIII. Buckton K.E., Brown W.M.C., Smith P.G. Lymphocyte survival in men treated with X-rays for ankylosing spondylitis - Nature. 1967; 214 (87): 470-473.

IX. Carper H.A., Hoffinan P.L. The intravascular survival of transfused canine pelger-huetneutrophils and eoizinophils - Blood. – 1966; 27: 739-745.

X. Choi C.M., Bennett R.G. Laser Dopplers to determine cutaneous blood flow. Dermatol. Surg. 2003; 29: 272-280.

XI. Chugh A., Eudes F., Shim Y.S. Cell-penetrating peptides: Nanocarrier for macromolecule delivery in living cells - IUBMB Life. 2010; 62 (3): 183-193.

XII. Dalmasso A.P., Goldish D., Benson B.A., Tsai A.K., Wasiluk K.R., Vercellotti G.M. Interleukin-4 induced up-regulation of endothelial cell claudin-5 through activation of FoxO1: Role in protection from complement-mediated injury - J. Biol. Chem. 2014; 289 (2): 838-847.

XIII. Dobrodeeva L.K., Patrakeeva V.P. Influence of migration and proliferative processes in leukocytes on the immune status of people living in the highlands. Ekaterinburg: UrO RAS, 2018.

XIV. Dobrodeeva L.K., Zhilina L.P. Immunological reactivity, health status of the population in Arkhangelsk oblast. Ekaterinburg, UrO RAS, 2004.

XV. Ford W.L., Gowans J.L., The Traffic of Lymphocytes. - Seminars Hemat. 1969; 6: 67-83.

XVI. Golder M., Chen C.L.H., O'Shea S., Corbett K., Chrystiel L., French G. Potential risk of crossinfection during peripheral-venous access by contaminated tourniquets - Lancet. 2000; 355(9197): 44.

XVII. Heller P. The blood and blood forming organs - The Year Book of Medicine.1972: 69-86.

XVIII. Humeau A., Steenbergen W., Nilsson H., Stromberg T. Laser Doppler perfusion monitoring and imaging: novel approaches - Med. Biol. Eng. Comput. 2007; 45: 421-435.

XIX. Kalenova L.F., Sukhovey Y.G., Novikova M.A., Fisher T.A. Influence of experimental local temperature factor on the immunological parameters of an organism. Journal of New Medical Technologies. 2009; 16 (4): 21-24.

XX. Kozlov V.I., Sokolov V.G. The study of the fluctuations of bloodstream in the system of microcirculation. Materials of the 2nd All-Russian symposium.
“Application of laser Doppler flowmetry in medical practice”. Moscow, 1998.

XXI. Kozyreva T.V. Central and peripheral thermoreceptors. Comparative analysis of the effects of prolonged adaptation to cold and noradrenaline - Neuroscience and behavioral physiology. 2007; 37(2): 191-198.

XXII. Kozyreva T.V., Eliseeva L.S. Immune response and content of corticosteroids in patients with different regiments of cooling. Journal of Experimental Biology and Medicine. 2002; 133 (4): 384-387.

XXIII. Kozyreva T.V., Eliseeva L.S. Immune response in cold exposures of different types - Journal of Thermal Biology. 2000; 25 (5): 401-404.

XXIV. Kozyreva T.V., Eliseeva L.S., Vavilin V.A. Influence of the rate and depth of cooling on the immune response and content of corticosteron in blood plasma. Russian Physiological Journal. 2000; 86 (12): 1618-1623.

XXV. Kozyreva T.V., Eliseeva L.S., Tsoi L.V., Khramova G.M. Effect of rapid slight cooling of the skin in various phases of immunogenesis on the immune response - Bulletin of experimental biology and medicine. 2006; 142(4): 409–412.

XXVI. Kozyreva T.V., Tkachenko E.Y., Simonova T.G. Functional alterations during adaptation to cold. Achievements in Physiological Sciences. 2003; 34(2): 94-102.

XXVII. Kozyreva T.V., Tkachenko E.Y., Eliseeva L.S., Khramova G.M., Tuzikov F.V., Kozaruk V.P., Voronova I.P. Influence of Ca2+ thermoregulatory reactions, content of lipoproteids in blood and immune response in patients exposed to cold and in patients with arterial hypertension. Journal of RAS. 2007; 4(126): 138-144.

XXVIII. Krechina E.K., Kozlov V.I., Maslova V.V. Microcirculation in gingival parodont tissue. M.: GEOTAR-Media, 2007.

XXIX. Krupatkin A.I. New possibilities in the evaluation of innervation of cutaneous microvessels with a spectral analysis of the fluctuations of the microhemodynamics. Regional blood and microcirculation. 2004; 4: 52-59.

XXX. Kulakov V.Y., Kaminskii B.F. Meteogeophysical stress and ways of its overcoming. Vladivostok: Medicina DV, 2003.

XXXI. Kulikov V.Y., Kim K.B. Oxygen regimen in the adaptation of humans at the Far North. Novosibirsk: Science, 1987.

XXXII. Ledyankina O.V., Puchkova M.S., Fatiyanova A.S. Hematological laboratory assays: handbook for students. Sverdlovsk, 2016.
XXXIII. Liu Na, Liu Juntian, Ji Yuanyuan, Lu Peipei, Wang Chenjing, Guo Fang. C-reactive protein induces TNF-α secretion by p38 MAPK-TLR4 signal pathway in rat vascular smooth muscle cells - Inflammation. 2011; 34 (4): 283-290.

XXXIV. Meuret G., Hoffman G. Monocyte kinetic studies in normal and disease states - Brit. J. Hematol. 1973; 24: 275-279.

XXXV. Migacheva A.A. Comparative characteristics of cellular composition of venous and capillary blood. Post-graduate Journal of Povolzhie. 2009; 7-8: 174-177.

XXXVI. Miller J.F.A.P., Basten A., Stert I., Cheers C. Interaction between lymphocytes in immune responses - Cell. Immunol. 1971; 2 (5): 469-495.

XXXVII. Neuret G., Friedner T.M. Neutrophyle and monocyte kineties in a case of cyclic neutropenia - Blood. 1974; 43: 565-574.

XXXVIII. NikolaeV.M., Semenova K.E., Naumova Y.I., Vladimirov A.S., Golderova L.N., Kuzmina S.S., Fedorova S.A. Functional activity of leukocytes in blood and nonspecific adaptive reaction of rats to cold. Veterinary. 2016; 1: 44-46.

XXXIX. Normal hematopoiesis and its regulation. Edited by Acad. N.A. Fedorova. M.: «Medicina», 1976.

XL. Procedures for the collection of diagnostic blood specimens by venipuncture - Wayne P.A.: Approved standard, 5th edition. - 2003. - NCCLS document H3-A5.

XLI. Samodova A.V., Dobrodeeva L.K. Association of the content of endothelin-1 and response of the immune reaction in people to short-term cooling. Journal of Ural Medical Academic Science. 2018; 15 (2): 299-308.

XLII. Shulman Z.P., Makhaneck A. A. Influence of temperature and acidification of blood on hemorheological properties. Engineering and Physics Journal. 2005; 78 (5): 180-185.

XLIII. Soter N.A., Lewis R.A., Corey E.J., Austen K.F. Local effects of synthetic leukotriens (LTC4, LTD4, LTE4, and LTB4) in human skin - J. Invest. Dermatol. 1983; 80(2): 115-119.

XLIV. Sutton C.D., White S.A., Edwards R., Lewis M.H. A prospective controlled trial of the efficacy of isopropyl alcohol wipes before venesection in surgical patients - Annals of the Royal College of Surgeons. 1999; 81(3): 183-186.

XLV. Thomson A. The Cytokine Handbook. London, 1992.
XLVI. TikhomirovaI.A., MuravievA.V. Physiological role and mechanisms of erythrocytes aggregation. Russian Physiological Journal. 2007; 93 (12): 1382-1393.

XLVII. Timerbulatov V.M., UrazbakhtinI.M., Fayazov R.R., Khasanov A.G., Dautov S. B., SibaevB.M., Sagitov R. B., Sakhautdinova I.V., Uzbakhtin M. I. The application of laser Doppler flowmetry in endoscopy and endosurgery in patients with acute conditions in the abdominal organs. M.: MED press-inform, 2006.

XLVIII. Totolyan A.A., FreidlinI.S. Cells of the immune system. SPb.: Science, 2000.

XLIX. Van Hove L., Chicano T., Brace L. Anemia diagnosis, classification, and monitoring using cell-dyn technology reviewed for the New Millennium - Laboratory Hematology. 1999; 6: 93-108.

L. Zhai H. Frisch S., Pelosi A., Neibart S., MaibachH.I. Antipruritic and thermal sensation effects of hydrocortisone creams in human skin - Skin. Pharmacol. Appl. Skin. Physiol. 2000; 13: 352-357.

LI. Zubatkinal. S., DobrodeevaL.K., Malakhova M.Y., KryzhanovskiyE.V., ZubatkinaO.V. Entropy as a factor of evaluation of the immune status. Journal of I.I. Mechnikov Northern-Western State Medical University. 2012; 1 (4): 57-61.