Modification of a PE/PPE substrate pair reroutes an Esx substrate pair from the mycobacterial ESX-1 type VII secretion system to the ESX-5 system

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Running title: PE/PPE determine system-specificity of Esx substrates

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ABSTRACT

Bacterial type VII secretion systems (T7SSs) secrete a wide range of extracellular proteins that play important roles in bacterial viability and in interactions of pathogenic mycobacteria with their hosts. Mycobacterial T7SSs consist of five subtypes, ESX-1–5, and have four substrate classes, namely, Esx, PE, PPE, and Esp proteins. At least some of these substrates are secreted as heterodimers. Each ESX system mediates the secretion of a specific set of Esx, PE, and PPE proteins, raising the question how these substrates are recognized in a system-specific fashion. For the PE/PPE heterodimers, it has been shown that they interact with their cognate EspG chaperone and that this chaperone determines the designated secretion pathway. However, both structural and pulldown analyses have suggested that EspG cannot interact with the Esx proteins. Therefore, the determining factor for system specificity of the Esx proteins remains unknown. Here, we investigated the secretion specificity of the ESX-1 substrate pair EsxB_1/EsxA_1 in Mycobacterium marinum. While this substrate pair was hardly secreted when homologously expressed, it was secreted when co-expressed together with the PE35/PPE68_1 pair, indicating that this pair could stimulate secretion of the EsxB_1/EsxA_1 pair. Surprisingly, co-expression of EsxB_1/EsxA_1 with a modified PE35/PPE68_1 version that carried the EspG_3 chaperone-binding domain, previously shown to redirect this substrate pair to the ESX-5 system, also resulted in redirection and co-secretion of the Esx pair via ESX-5. Our results suggest a secretion model in which PE35/PPE68_1 determines the system-specific secretion of EsxB_1/EsxA_1.

INTRODUCTION

Mycobacteria possess an unusual hydrophobic cell envelope that protects them from various stresses and contributes to the resilience of pathogenic mycobacteria during infection. Classified as high-GC Gram-positive bacteria, the cell envelope of mycobacteria consists of a relatively standard cell membrane with a surrounding peptidoglycan layer. However, mycobacteria belong to a subgroup of high-GC Gram-positive bacteria that have acquired an extra hydrophobic layer of long-chain fatty acids, called mycolic acids. These specific lipids are covalently linked via an arabinogalactan layer to the peptidoglycan layer, forming a highly rigid and impermeable structure. Mycobacteria employ specialized machineries, called type VII secretion systems (T7SSs) to secrete proteins across their complex cell envelope (1, 2). Mycobacterium tuberculosis possesses five of such T7SSs, named ESX-1 to ESX-5 (1, 2), of which ESX-1, ESX-3 and ESX-5 have been functionally analyzed (3–10). Each of these systems plays a different role in the mycobacterial life cycle. For example, ESX-1 has a key role in virulence of pathogenic mycobacteria, as it mediates phagosomal rupture inside macrophages (11–14) and the subsequent escape of M. tuberculosis from the phagolysosome (3–6, 15, 16). ESX-3 and ESX-5 are necessary for iron and fatty acid uptake, respectively, making these systems essential for bacterial viability (7–10). Besides their roles in
nutrient and metabolite acquisition, ESX-3 and ESX-5 are additionally involved in immune modulation of the host (9, 17, 18).

The substrates that are secreted by these three ESX systems belong to distinctive protein families, \textit{i.e.} Esx, PE, PPE and Esp proteins, most of them belonging to the so-called EsxAB clan (Pfam CL0352) (19). Within this clan, the Esx proteins belong to the WxG100 family of proteins that contain an WxG protein motif and are generally 100 amino acids long. The PE and PPE proteins are named after proline-glutamic acid (PE) and proline-proline-glutamic acid (PPE) motifs in their N-terminal domains, respectively (20). These N-terminal homology domains consist of roughly 110 amino acids for the PE protein family and 180 amino acids for the PPE protein family (20). The C-terminal domains of PE and PPE proteins vary extensively in length and sequence (20).

ESX substrates have been shown to form heterodimers in the cytosol, \textit{i.e.} two Esx proteins pair together and PE proteins pair with a PPE protein, and are thought to be secreted as (partially) folded heterodimers (21–25). Crystal structures have been solved for several heterodimeric substrates of different ESX systems, revealing highly conserved features, in which the interface of Esx heterodimers (26, 27), as well as the interface of PE/PPE heterodimers, is formed by two helix-turn-helix structures oriented antiparallel to each other (22, 23, 28). Interestingly, each ESX system secretes its own subset of Esx, PE and PPE substrates that are most-likely responsible for the various roles of ESX systems in the bacterial life cycle. How these structurally similar proteins are specifically targeted to their corresponding ESX system still remains unclear. A conserved T7SS secretion signal (YxxxD/E) has been identified, which is located directly after the helix-turn-helix domain of one partner protein of the Esx heterodimer and the PE partner of the PE/PPE heterodimer. This signal, although required for secretion, is exchangeable among PE substrates of different ESX systems without changing their initial secretion route (29, 30). Hence, this signal does not determine system-specific secretion of these ESX substrates.

Structural analysis showed that the conserved N-termini of PPE proteins contain, in addition to the helix-turn-helix structure, an extra domain that is not present in Esx and PE proteins. This relatively hydrophobic helical tip domain extends from the characteristic four-helix bundle formed by the PE-PPE interface (22, 23). This domain is recognized by a cytosolic chaperone, called EspG, in a system-specific manner (Fig. S1), which interaction is required for secretion of the PE/PPE pair (22, 23, 31). Subsequently, we previously could establish the redirection of the ESX-1 substrate pair PE35/PPE68_1 to the ESX-5 system by replacing its EspG1 chaperone binding domain with the equivalent domain of the ESX-5 substrate PPE18 (32). This suggests that this domain determines through which system these substrates are transported. The remaining question is how the Esx substrate pairs that lack this extended tip domain, are specifically recognized and targeted to their designated systems.

Here, we investigated the signals that determine the system-specificity of Esx substrates in \textit{M. marinum} using the ESX-1
heterodimer EsxB_1/EsxA_1 as model substrates. Its encoding genes esxB_1/esxA_1 (MMAR_0187/MMAR_0188) are adjacent to pe35/ppe68_1 (MMAR_0185/MMAR_0186), which gene products we have previously used as a model ESX-1 dependent PE/PPE heterodimer (31, 32). We found that EsxB_1/EsxA_1 secretion via the ESX-1 system was severely enhanced by the co-expression and secretion of PE35/PPE68_1. Although previous genetic studies have already reported co-dependence of ESX-1 substrates for secretion (33–37), this observation also indicates that balanced expression levels of both heterodimers are required for optimal secretion via ESX-1. Interestingly, the EsxB_1/EsxA_1 pair could be rerouted to the ESX-5 system by solely exchanging the EspG binding domain in PPE68_1. This does not only confirm the strict dependence of the EsxB_1/EsxA_1 heterodimer on PE35/PPE68_1 for secretion, but also reveals that this PE/PPE pair determines the system-specificity of this Esx pair.

RESULTS

**EsxB_1/EsxA_1 require co-expression of PE35/PPE68_1 for efficient ESX-1 dependent secretion**

To investigate how the system-specific secretion of Esx substrates is achieved, we investigated the secretion requirements of EsxB_1/EsxA_1 in *M. marinum*. The corresponding coding genes (MMAR_0187/MMAR_0188) lie adjacent to the gene pair pe35/ppe68_1 (MMAR_0185/MMAR_0186) and are paralogues of the pe35-ppe68-esxB-esxA gene cluster located in the *esx-1* locus (Fig. 1A). We introduced a multicopy plasmid containing esxB_1/esxA_1, expressed under the constitutive hsp60 promoter, in WT *M. marinum* (38). We also included WT *M. marinum* containing the previously analyzed pe35/ppe68_1 gene pair controlled by the same promoter on an integrative plasmid as an ESX-1 substrate control (32). Secretion was analyzed by immunoblotting using the introduced HA and FLAG epitopes at the C-termini of EsxA_1 and PPE68_1, respectively (Fig. 2A). To assess the possible variation in secretion between different colonies, we consistently checked duplicates of the same transformants throughout this study. The cytosolic protein GroEL2 was not detected in the culture supernatant fractions of all analyzed cultures, confirming the integrity of the bacterial cells. Consistent with published data (29, 32), PPE68_1-FLAG was detected in the culture supernatant fraction of the WT strain as a faint smeary band of ~40 kDa (Fig. 2A, lane 11, 12). Whereas EsxA_1-HA was well-expressed, as judged from the HA signals detected in the pellet fractions (Fig. 2A, lane 4, 5), this protein was not secreted by the WT strain (Fig. 2A, lane 13, 14). Surprisingly, secretion of endogenous EsxA, detected by an EsxA antibody, was severely affected by the overexpression of EsxB_1/EsxA_1 (Fig. 2A, lane 13, 14). To investigate the impact of EsxB_1/EsxA_1 overexpression also on other ESX-1 substrates, secretion of EspE was investigated (Fig. S2). This substrate remains attached to the bacterial surface in *M. marinum* and can be extracted by the mild detergent Genapol X-080. The presence of EspE in the Genapol extracted fraction was completely
abolished in the strain overexpressing EsxB\_1/EsxA\_1, indicating that this homologously expressed pair has a broad effect on ESX-1 secretion (Fig S2, lane 13, 14). Since several T7SS substrates, in particular those of the ESX-1 system, have been shown to be dependent on each other for secretion (32, 34, 39), we hypothesized that the secretion of overexpressed EsxB\_1/EsxA\_1 might require the co-expression of the PE35/PPE68\_1 pair that is putatively located in the same operon. Similarly organized loci containing a pe/ppe pair and an adjacent esx gene pair can be observed in other ESX clusters. We co-electroporated the integrative pMV361::pe35/ppe68\_1-flag and the multicopy pSMT3::esxB\_1/esxA\_1-ha in WT M. marinum. Secretion analysis followed by immunoblotting showed that the co-expression of EsxB\_1/EsxA\_1-HA did not seem to affect the expression or secretion of PPE68\_1-FLAG (Fig. 2A, lane 6, 7 and lane 15, 16). In contrast, EsxA\_1-HA was now efficiently secreted and detected as a smeary band of \~15 kDa in the supernatant of the strain that co-expressed both the PE/PPE and the Esx substrate pairs (Fig. 2A, lane 15, 16). Furthermore, the immunoblot signals in the supernatant fractions using the EsxA antibody, which probably detects both EsxA\_1-HA and endogenous EsxA, became comparable to that of the WT strain without any constructs (Fig. 2A, lane 15, 16). However, the presence of EspE in the Genapol extracted fraction was not restored in the strain overexpressing both the Esx and PE/PPE pair, suggesting that the secretion of endogenous ESX-1 substrates was still blocked (Fig. S2, lane 15, 16). These data show that the efficient secretion of overexpressed EsxA\_1-HA relies on the co-overexpression of PE35/PPE68\_1-FLAG.

Because the integrative pMV361 plasmid and the multicopy pSMT3 plasmid differ in copy numbers, thereby possibly resulting in suboptimal co-secretion of the two substrate pairs, we also introduced the complete pe35/ppe68\_1/esxB\_1/esxA\_1 locus into the pSMT3 plasmid again with a FLAG and HA tag fused to the C-termini of PPE68\_1 and EsxA\_1, respectively. The construct is hereafter referred to as the ALL WT construct. We observed that while the cellular levels of both EsxA\_1-HA and PPE68\_1-FLAG were increased (Fig. 2A, lane 8, 9), the secretion of EsxA\_1-HA was similar to the condition when the two substrate pairs were independently expressed (Fig. 2A, lane 17, 18). Taken together, our data suggest that the co-expression of PE35/PPE68\_1, but not its co-transcription, increases the secretion of homologously expressed EsxB\_1/EsxA\_1 in M. marinum.

We next investigated whether the secretion of overexpressed EsxA\_1 requires not only co-overexpression, but also co-secretion of PPE68\_1. To test this, the C-terminal 15 and 21 amino acids of PE35 and EsxB\_1, respectively, containing the YxxxD/E secretion motif, were deleted, which abolishes their secretion. We analyzed the effect of these truncations on the secretion of PPE68\_1 and EsxA\_1 in the WT and an eccCb\_1 mutant strain (esx-1 mutant), previously described as a non-functional esx-1 mutant (40) (Fig. 2B). In all tested cultures, the supernatants were devoid of GroEL2, indicating the integrity of the cells (Fig. 2B). Secretion of both EsxA-HA and PPE68\_1-FLAG was
abolished in the esx-1 mutant strain, confirming these proteins are secreted via the ESX-1 secretion system (Fig. 2B, lane 12). Importantly, immunoblot analysis showed that the deletion of the C-terminal tail of either PE35 or EsxB_1 disrupted the secretion of both PPE68_1 and EsxA_1 in WT M. marinum (Fig. 2B, lane 13, 14, and 17, 18). Only some minor secretion of PPE68_1 was observed in the WT strain, but not in the esx-1 mutant strain, when the secretion signal in EsxB_1 was deleted (Fig. 2B, lane 17, 18). We therefore conclude that the presence of both the PE35 and EsxB_1 C-terminal tails, containing the T7SS secretion signal, are required for the secretion of both substrates. This indicates that EsxB_1/EsxA_1 and PE35/PPE68_1 are co-dependently secreted via the ESX-1 system.

**WT EsxB_1/EsxA_1 is rerouted to the ESX-5 system by introducing the EspG binding domain in PPE68_1**

Two different domains in ESX substrates have been identified that are important for secretion: the YxxxD/E motif, located in the C-terminal tail directly after the helix-turn-helix domain of some Esx and all PE proteins, and the EspG binding domain in the conserved N-terminal domain of PPE proteins. Although the C-terminal tails of PE substrates, secreted through different ESX systems, can in general be exchanged without changing their predetermined secretion route (29), in a previous study we have been able to redirect PPE68_1 to the ESX-5 system by exchanging its EspG1 chaperone binding domain with the equivalent domain of the ESX-5 substrate PPE18 (32).

Here, we aimed to determine which signals are required for establishing rerouting of EsxA_1 to the ESX-5 system, starting by investigating the role of the EspG binding domain of the co-expressed PE/PPE pair. For this, we constructed pe35/ppe68_1/esxB_1/esxA_1, in which ppe68_1 was engineered to express a PPE68_1 variant that carries the EspG binding domain of the PPE18 protein (Fig. 1B, 3A), as previously described (32). This entire construct was named SINGLE SWAP (Fig. 3A) and the modified PPE68_1 protein was named PPE68_1 SWAP. This construct was introduced in M. marinum WT, esx-1, and esx-5 mutant strains, after which secretion was analyzed and compared to the same strains expressing the already explained ALL WT construct.

Interestingly, we observed that the presence of the SINGLE SWAP construct seemed to induce minor lysis of WT M. marinum cells, as a small amount of GroEL2 was consistently detected in the supernatants of these cultures (Fig. 3C). Nevertheless, the detected amount of GroEL2 was comparable among the strains expressing the different constructs, allowing further analysis. As observed before, the expression of the ALL WT construct resulted in expression and secretion of both PPE68_1-FLAG and EsxA_1-HA (Fig. 3C, lane 1, 2 and 9, 10). The EsxA antibody was included to confirm the total EsxA expression and secretion (Fig. 3C, lane 1-2 and 9, 10). As seen previously for PPE68_1 SWAP (32), we observed that the PPE68_1 hybrid expressed from the SINGLE SWAP construct appeared as a slightly higher band than the PPE68_1 WT and was efficiently secreted in the WT strain (Fig. 3C, lane 3, 4 and 11-12).
Notably, while secretion of PPE68_1 SWAP was more efficient than PPE68_1 WT (32), the amount of EsxA_1 was also higher in the supernatant fractions, as judged by an increased intensity of both HA and EsxA signals (Fig. 3C, lane 11, 12).

We subsequently addressed the involved secretion systems by first introducing the SINGLE SWAP construct in the esx-1 mutant strain. In contrast to WT M. marinum cells, GroEL2 was not detected in the supernatant fractions of this mutant strain, indicating the integrity of the cells in the presence of the constructs (Fig. 3D). As expected, secretion of both PPE68_1-FLAG and EsxA_1-HA of the ALL WT construct was abrogated (Fig. 3D, lane 9, 10). In contrast, the PPE68_1 SWAP protein was still secreted (Fig. 3D, lane 11, 12), confirming our previous observation that the PPE68_1 SWAP was secreted independently of the ESX-1 system (32). Importantly, we also still detected EsxA_1 in the supernatant, using both the HA and the EsxA antibody (Fig. 3D, lane 11, 12), suggesting that this ESX-1 substrate is now secreted in an ESX-1 independent manner as well. This is highly interesting as both EsxA_1 and EsxB_1 are unmodified in the SINGLE SWAP construct. Together, these data suggest that PPE68_1 SWAP determines the ESX-1 independent secretion of EsxA_1.

To confirm that PPE68_1 SWAP and EsxA_1 are secreted by the ESX-5 system, we introduced the SINGLE SWAP construct in the ΔeccC5 strain. EccC5 is an essential component of the ESX-5 machinery (41, 42) and deletion of this component blocks ESX-5-dependent secretion (7). In this strain, the presence of the tested SINGLE SWAP construct consistently caused minor bacterial lysis, indicated by the presence of GroEL2 in the supernatant fractions (Fig. 3E). Since a similar phenotype was observed for the same construct in the WT strain, but not in the esx-1 mutant, the bacterial leakage induced upon homologous expression of these proteins seemed to be linked to a functional ESX-1 system. Expressing the ALL WT construct, both PPE68_1-FLAG and EsxA_1-HA were detected in the supernatant fractions of the ΔeccC5 strain by using the FLAG and HA antibody, respectively (Fig. 3E, lane 13, 14). Notably, more intense EsxA signals could be observed in the supernatant fraction of the esx-5 mutant containing the ALL WT construct (Fig. 3E, lane 13, 14), compared to the empty ΔeccC5 strain (Fig. 3E, lane 3). This indicates that overexpressed EsxA_1 is particularly well secreted in this strain. The PPE68_1 SWAP and EsxA_1 of the SINGLE SWAP were only moderately detected in the supernatant of the ΔeccC5 strain (Fig. 3E, lane 15, 16). Thus, our data show that the secretion of both proteins became mostly dependent on the ESX-5 system when the EspG5 binding domain was present. The observed residual secretion of PPE68_1-FLAG and EsxA_1-HA with the SINGLE SWAP construct indicates that a small amount of these substrate pairs can still be secreted via ESX-1. Interestingly, we previously have shown that secretion of PPE68_1 SWAP was completely blocked in the same esx-5 mutant in the absence of overexpressed EsxB_1/EsxA_1 (32). This indicates that this Esx substrate pair might be able to guide some amount of PPE68_1 SWAP to the ESX-1 system.
Finally, we observed a competitive correlation between the secretion of the rerouted substrates PE35/PPE68_1 SWAP and native substrates of the ESX-5 system, i.e. the PE_PGRS proteins. Using the Genapol extraction method to analyze the surface localization of PE_PGRS proteins, we observed a lower amount of these proteins in the Genapol-extracted fraction in the esx-1 mutant strain expressing the SINGLE SWAP construct, compared to the esx-1 mutant strain expressing the WT construct (Fig. S3). This suggests that, similar to what was reported previously, the redirection of ESX-1 substrates to the ESX-5 system interferes with the export of endogenous ESX-5 substrates (32). Interestingly however, WT M. marinum expressing the SINGLE SWAP construct was still unable to secrete EspE, another ESX-1 dependent substrate (Fig. S2, lane 19, 20). This suggests that redirection of PPE68_1 SWAP and EsxA_1 via ESX-5 does not relieve the ESX-1 secretion block that was observed when overexpressing the WT PE35/PPE68_1-FLAG and EsxB_1/EsxA_1-HA heterodimers in M. marinum WT.

In summary, introducing the EspG₅ binding domain in PPE68_1 resulted in the rerouting of both the PPE86_1 and EsxA_1 substrate to the ESX-5 system. This observation does not only further confirm that these proteins are co-secreted, but also shows that the PPE protein is involved in determining the system-specificity of the Esx substrate.

The redirection of EsxB_1/EsxA_1 via the ESX-5 system can be further improved by exchanging the T7SS secretion signals

Although the C-terminal tails, containing the YxxxD/E secretion motif, of PE proteins are essentially not involved in determining the system-specificity of PE/PPE pairs (29), we investigated next the role of the equivalent domain of Esx proteins in substrate redirection. For this, we constructed an additional pe35/ppe68_1/esxB_1/esxA_1 construct, in which the C-terminal domain EsxB_1, containing the YxxxD/E motif, was replaced by the equivalent region of the ESX-5 substrate EsxM (Fig. 3B). We observed however that exchanging this domain abolished secretion of EsxA_1-HA in WT M. marinum (Fig. S4, lane 16, 17). This is in agreement with a previous study, showing that exchanging the C-terminal domain in PE35 with the equivalent domain of PE31 blocks secretion of PPE68_1 in the same WT strain (32).

We therefore investigated next whether exchanging the T7SS secretion signals in EsxB_1 and PE35 could improve the redirection of our SINGLE SWAP construct, i.e. in which the EspG binding domain of PPE68_1 was exchanged (Fig. 3A). The construct, in which the C-terminal amino acids of EsxB_1 was exchanged with the equivalent residues of EsxM (Fig. 3B), was designated DOUBLE SWAP. In addition, we replaced the 15 amino-acid C-terminal domain of the PE35 by the corresponding region of the ESX-5 substrate PE31 (Fig. 3B), resulting in the construct of TRIPLE SWAP, i.e. this construct carried three exchanged domains (Fig. 3A). The secretion of these variants was again analyzed in M. marinum WT, the esx-1 and esx-5 mutant strain.

The presence of the DOUBLE and TRIPLE SWAP constructs seemed to cause
minor lysis of WT *M. marinum* cells, similar to the SINGLE SWAP construct (Fig. 3C). The presence of the C-terminal tail of EsxM in the DOUBLE SWAP construct did not affect the expression or secretion of PPE68_1 SWAP or EsxA_1, as similar intensities of the detected signals were observed compared to those of the SINGLE SWAP substrates (Fig. 3C, lane 5, 6, and 13, 14). However, when ESX-5 secretion signals were introduced in both PE35 and EsxB_1, the secretion of both the PPE68_1 SWAP and EsxA_1 seemed to reach the highest efficiency (Fig. 3C, lane 15, 16).

In the *esx-1* mutant strain, GroEL2 was not detected in the supernatant fractions, indicating the integrity of the cells in the presence of these two constructs (Fig. 3D). Here, the DOUBLE SWAP construct induced comparable levels of EsxA_1 secretion as the SINGLE SWAP construct (Fig. 3D, lane 13, 14), while the secretion of EsxA_1 appeared most efficient in the presence of both the ESX-5 secretion signals in the TRIPLE SWAP construct (Fig. 3D, lane 15, 16). The competitive correlation between the secretion of the rerouted substrates and the ESX-5-depenent PE_PGRS proteins was also observed in the *esx-1* mutant expressing the DOUBLE SWAP and the TRIPLE SWAP construct (Fig. S3, lane 13-16). Finally, no PPE68_1 SWAP and only a minor amount of EsxA_1 was detected in the supernatants of ΔeccC strains, containing either the DOUBLE or the TRIPLE SWAP construct (Fig. 3E, lane 17-20). From these data, we conclude that the exchange of the EspG binding domain in PPE68_1 is essential and sufficient for redirection of both the PE/PPE and Esx pair, while the general secretion motifs merely improve redirection efficiency to the ESX-5 system.

**DISCUSSION**

Each mycobacterial T7SS secretes their own subset of Esx, PE and PPE proteins, which share sequence similarities and show structural resemblance. This phenomenon raises the question how these substrates are specifically recognized by the T7SS subtypes. Recently, we showed that the system-specificity of the PE/PPE substrates is determined by the EspG chaperone binding domain on the PPE protein (32). However, a similar mechanism for system-specific recognition cannot apply to the Esx heterodimers, since they lack this chaperone binding domain and therefore cannot interact with the EspG chaperones (43). In this study, we show that the PE35/PPE68_1 heterodimer defines the system-specific secretion of EsxB_1/EsxA_1 in *M. marinum*.

We found that ESX-1 dependent secretion of overexpressed EsxB_1/EsxA_1 is severely enhanced when PE35/PPE68_1 were co-overexpressed. Co-expression of PE35/PPE68_1 from the same promoter was not required for EsxA_1 secretion, suggesting that this dependency is not transcriptionally linked. Our previous observation that deletion of *espG* leads to a loss of not only PE/PPE secretion, but also of EsxB/EsxA and other ESX-1 substrates in *M. marinum* (32, 44), already hinted towards the dependency of EsxB/EsxA on PE/PPE substrates for their secretion. Interestingly, in *M. tuberculosis* the dependency of EsxA secretion on PPE68 seems more complex. Although PPE68...
itself is indeed crucial for EsxA secretion, an espG₁ deletion in this species, which is predicted to affect PPE68, does not seem to affect EsxA secretion (4, 39). Since there is only a single copy of esxA in the genome of M. tuberculosis, contrary to the four highly homologous esxA copies M. marinum, it is possible that the two pathogens employ distinct mechanisms to govern the regulation and/or secretion of EsxA. Alternatively, there might be redundancy of different PE/PPE proteins in M. tuberculosis in facilitating the secretion of EsxA, or EspG₁ is less important for ESX-1 secretion in M. tuberculosis.

In this study, we also confirmed that the C-terminal tails of PE35 and EsxB₁, containing the YxxxD/E secretion motifs, are required for the secretion of the corresponding heterodimer, consistent with other studies (29). Moreover, the finding that both secretion motifs are strictly required for the secretion of both heterodimers is highly interesting. These data show that the secretion of EsxA₁ is not only dependent on the co-overexpression, but also on the co-secretion of PE35/PPE68₁. In addition, the observation that secretion of PPE68₁ was diminished when the secretion of EsxB₁/EsxA₁ was blocked, by deleting the secretion signal of EsxB₁, suggests that both heterodimers are mutually dependent on each other for their export. Interestingly, the cytosolic accumulation of EsxB₁/EsxA₁, which occurs without co-overexpression of PE35/PPE68₁ or when the C-terminal tails of PE35 and/or EsxB₁ was deleted, abolishes the secretion of endogenous EsxA and EspE. This is in agreement with our previous observation that the introduction of an ESX-5 secretion signal in PE35 of homologously expressed PE35/PPE68₁ results in a secretion block of both PPE68₁ and endogenous EsxA (32). Possibly, a cytosolic factor that is required for substrate recognition, e.g. EspG₁, is titrated away by these non-secreted substrates.

The finding that WT EsxB₁/EsxA₁ could be rerouted to the ESX-5 system in M. marinum by solely manipulating the EspG₁ binding domain of PE35/PPE68₁ is unexpected. The fact that WT EsxB₁/EsxA₁ could be co-rerouted in this manner underlines the strict dependency of EsxB₁/EsxA₁ secretion on PE35/PPE68₁. In addition, secretion of both overexpressed PPE68₁ WT via ESX-1 and PPE68₁ SWAP via ESX-5 does not seem to be enhanced upon co-expression of EsxB₁/EsxA₁. This hints towards a hierarchy in secretion, where the PE/PPE pair controls and might regulate secretion of EsxB₁/EsxA₁. Co-rerouting of endogenous EsxA by the overexpression of PE35/PPE68₁ SWAP was previously not observed (32), suggesting that expression levels of endogenous Esx and PE/PPE pairs are well-balanced. Interestingly, the ability of the ESX-5 system to secrete the two substrate pairs, while they still carry both ESX-1 secretion signals, reflects its flexibility in secreting a wide range of substrates, as already discussed in other studies (22, 23, 40). Nevertheless, rerouting was most efficient when both PE35 and the EsxB₁ carried the ESX-5 C-terminal secretion signals of PE31 and EsxM, respectively. This suggests that the secretion signals of PE31 and EsxM are more optimal for the recognition and secretion by the ESX-5 machinery.

So far, the reason behind co-dependence among ESX substrates remains unclear. It has
been proposed that binding to or activation of the T7SS membrane complex could explain this phenomenon. Previous studies have shown that EsxB binds to the third nucleotide binding domain (NBD) of the conserved membrane complex component EccCα1 and induces hexamerization of this ATPase in vitro (30, 45, 46, 47). Specifically, the last seven amino acids of EsxB have been shown to be essential for this interaction (30). The C-terminus of other Esx homologs in M. tuberculosis are likely to be structured similarly to the C-terminus of EsxB, but do contain different amino acids sequences, suggesting this domain might be involved in system-specific recognition (30, 46, 48). Importantly, structural analysis of EccC of Thermomonospora curvata have shown that the crucial first NBD is kept in an inactivate state by a specific region in the linker 2 domain that connects the first and second NBD, and binding of EsxB is not able to activate this ATPase activity in vitro (40). It has therefore been suggested that an additional trigger is necessary to activate EccC, which could be the binding of PE/PPE substrates. Interestingly, this hypothesis is supported by a recent study, showing a species-specific role for the linker 2 domain of EccC2 in secretion of PE_PGRS proteins, a large subgroup of PE proteins that are secreted via ESX-5 (49).

From our study, it seems evident that the secretion of the Esx substrates are closely linked to that of PE/PPE heterodimers. Possibly, they bind simultaneously or sequentially in order to activate all three ATPase domains of EccC, after which transport through the membrane complex is achieved. Such a model would explain both the necessity for equal expression levels of both heterodimers, as well as the secretion dependency of the Esx pair on the PE/PPE pair that we observed here. However, PE/PPE proteins can only be found in the genus of Mycobacterium, while the homologues of the Esx substrates and the EccC core component can be found in a more diverse repertoire of Gram-positive species (1, 20, 50). It will be interesting to see the differences in substrate recognition and secretion between these different systems.

This study indicates that the recognition of both PE/PPE and Esx substrates by the various ESX systems depends on the EspG chaperones. In M. marinum, the role of these chaperones in ESX secretion is essential (32, 44) and the EspG-binding is required to keep PE/PPE proteins from aggregating (22). Chaperones with dual roles in preventing aggregation and secretion have also been identified in Type III secretion systems (T3SSs) (51). Conserved T3SS chaperones prevent premature assembly of the injection needle and pore-forming subunits, while the effector proteins are targeted to the secretion machinery by binding of their dedicated chaperone partner. Possibly, the EspG chaperones in T7SS have acquired similar roles in keeping substrates in a secretion competent state.

The homologue of EsxA_1, EsxA, is the most-studied ESX substrate and has been suggested to be responsible for ESX-1-induced phagosomal rupture. EsxA has been found to be associated with membrane lysis when a transposon mutant of essA/essB was unable to lyse cultured lung epithelial cell lines (4, 52). Further genetic studies in M. marinum have shown that several different transposon mutants defective in EssA secretion loose haemolytic
activity and are attenuated in zebrafish (3, 12, 15, 53), supporting the hypothesis that EsxA is a crucial virulent factor of pathogenic mycobacteria. However, secretion of different ESX-1 substrate classes has been shown to be interdependent on each other (33, 34), e.g. loss of EspA or PPE68 secretion leads to secretion defects of EsxA and vice versa (32, 34). Therefore, studying functions of individual ESX-1 substrates during the mycobacterial infection cycle has been a challenge. While protein sequences of EsxB and EsxB_1 are identical, EsxA_1 shares 92% protein sequence identity with EsxA. Given the high similarity, it has been suggested that EsxB_1/EsxA_1 have an equivalent functionality as the esx-1 encoded EsxB/EsxA (54). The observation that WT EsxA_1 can be destined for the ESX-5 system provides a unique platform to investigate exact roles of this protein in host-pathogen interactions. Current research is focusing on the redirection of EsxB/EsxA in order to directly assess the membrane lysis activity of this substrate pair.

EXPERIMENTAL PROCEDURES

Bacterial strains and growth cultures
All mycobacterial strains were grown on Middlebrook 7H10 plates (Difco) containing OADC supplement (oleic acid, albumin, dextrose and catalase; BD Biosciences) or liquid 7H9 medium containing ADC supplement (BD Biosciences) and the appropriate antibiotics (see below). *M. marinum* strains were grown at 30 °C, 90 rpm. All mycobacterial strains and mutants are listed in Table S1. *Escherichia coli* strain DH5α was used for cloning procedures and plasmid accumulation, and was grown on lysogeny broth (LB) plates or liquid broth at 37 °C, 200 rpm. Growth media was supplemented with the appropriate antibiotics at the following concentrations: kanamycin (Roche) 25 µg/ml; hygromycin (Sigma) 50 µg/ml.

**Plasmid construction**
All PCRs were carried out with the Phusion High-Fidelity DNA polymerase (Finnzymes) using primers listed in supplemental Table S2. The restriction sites used for cloning are indicated in this table. All plasmids used in this study are listed in supplemental Table S3.

**Protein secretion and immunoblot analysis**
*M. marinum* strains were grown in 7H9 liquid medium supplemented with ADC, 0.05% Tween 80, and appropriate antibiotics until mid-logarithmic phase, after which the cells were washed and inoculated in 7H9 medium with 0.2% dextrose, 0.05% Tween 80 at an optical density at 600 nm (OD600) of 0.4 and grown for another 16 h. The cells (Pellet) were spun down for 10 min at 6,000 xg, washed with phosphate buffered saline (PBS), and resuspended in SDS loading buffer (containing 100mM DTT and 2% SDS). Supernatants were passed through 0.2 µm-pore-size filter units and proteins were precipitated with trichloroacetic acid (TCA) and resuspended in SDS loading buffer. Alternatively, the cells (Pellet) were resuspended in 0.5% Genapol X-080 and incubated for 1 h at room temperature. Samples were spun down and pellets were resuspended in SDS sample loading buffer (Genapol Pellet), while 5x SDS sample buffer was added to the supernatant containing Genapol X-080 (Genapol
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Supernatant) to obtain a final concentration of 1x SDS buffer. Proteins were separated on SDS-PAGE gels and transferred to a nitrocellulose membrane, and membranes were stained with anti-GroEL2 (monoclonal antibody Cs44; John Belisle, NIH, Bethesda, MD, USA), anti-PE_PGRS (7C4.1F7) (40), anti-ESAT-6 (monoclonal antibody [MAb] Hyb76-8) (55), anti-HA (HA.11; Covance), anti-EspE (polyclonal rabbit antibody; Eric Brown; Genentech), anti-Flag (M2 monoclonal antibody produced in mouse, Sigma.

**Data availability statement:** All data are contained within this manuscript.
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Footnotes

The abbreviations used are: T7SS, Type VII secretion system; NBD, nucleotide binding domain; PBS, phosphate buffered saline; TCA, trichloroacetic acid
Figure 1. Schematic representation of the esx-1 locus and the ESX-1 and ESX-5 secretion system. (A) Genetic organization of the esx-1 locus and the duplicated region containing pe35/ppe68_1/esxB_1/esxA_1. Genes are color-coded according to the localization of their encoded proteins (see color key). The frameshift mutation in the eccCb1 mutant (also named M^VU) used in this study is indicated by an arrow. (B) Current model of substrate recognition by the type VII secretion systems. Shown are the model substrates used in this study, i.e. the ESX-1 substrate pairs EsxB_1/EsxA_1 and PE35_1/PPE68_1, and the ESX-5 substrate pairs EsxM/EsxN and PE31/PPE18. Previously, PPE68_1 has been shown to be rerouted to the ESX-5 system by exchanging the EspG1 binding domain with the equivalent domain of PPE18, indicated by red boxes (32). EspG components recognize cognate PE/PPE substrates, independently of the general secretion signal located in the C-terminal tails of the PE proteins. This recognition is required for secretion via the cognate secretion machinery located in the mycobacterial inner membrane. Recognition of Esx substrates by the T7SS membrane complex occurs through the interaction of the C-terminal tail of one of the Esx partner proteins with the third nucleotide binding domain of EccC (46, 47). How PE/PPE pairs are recognized by this complex remains unclear. Notably, only the conserved N-terminal domains of the PE and PPE proteins are shown in this model.
Figure 2. Homologously expressed EsxA_1/EsxB_1 require co-expression and secretion of PE35/PPE68_1 for efficient secretion via the ESX-1 system (A) A schematic representation of the different constructs used are shown on the left. The four genes encoding the *M. marinum* ESX-1 substrates PE35/PPE68_1 (in blue) and EsxB_1/EsxA_1 (in pink) are either expressed from separate *hsp60* promoters (I and II) or co-expressed under the same *hsp60* promoter (III). Immunoblot analysis of the cell pellet and culture supernatant fractions of WT *M. marinum* using an HA antibody to detect EsxA_1-HA, a FLAG-antibody to detect PPE68_1-FLAG, an EsxA antibody, detecting both endogenous and exogenous EsxA paralogues, and a GroEL2 antibody to detect the intracellular control protein GroEL2. (B) A schematic representation of the different constructs used are shown on the left; deletions are indicated by a cross. Immunoblot analysis using the same antibodies as under A to analyze secretion by WT and the ESX-1 mutant. In all blots, equivalent OD units were loaded; 0.2 OD for pellet and 0.5 OD for supernatant fractions. Numbers indicate two independent *M. marinum* colonies carrying the same construct. E, empty strain.
Figure 3. EsxA_1 is co-routed to ESX-5 by introducing the EspG5 binding domain of ESX-5 substrate PPE18 into PPE68_1. (A) Schematic representation of the different constructs used in the secretion analysis. The introduced sequences of the ESX-5 substrates PE31/PPE18 are in grey. The FLAG-tag (on PPE68_1) and HA-tag (on EsxA_1) are shown in red. (B) Alignment of the swapped sequences containing the C-terminal secretion motifs of the ESX-5 substrates PE31 and EsxM and the ESX-1 substrates PE35 and EsxB_1, respectively. The conserved secretion motif YxxxD/E are highlighted with a red box. (C,D,E) Immunoblot analysis of EsxA_1 as detected with an HA antibody, PPE68_1 as probed with a FLAG antibody, endogenous and exogenous EsxA paralogues using an EsxA antibody and intracellular GroEL2 by a GroEL2 antibody, in pellet and supernatant fractions. Different derivatives of PE35/PPE68_1/EsxB_1/EsxA_1 were tested in WT M. marinum (C), an eccCb1 mutant (esx-1 mutant; D), and an eccC5 knockout strain (esx-5 mutant; E). Equivalent OD units of cell pellets (0.2 OD unit) and culture supernatants (0.5 OD unit) are shown. Numbers indicate two independent M. marinum colonies carrying the same construct.
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