Exosomal transfer of stroma-derived miR21 confers paclitaxel resistance in ovarian cancer cells through targeting APAF1

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Advanced ovarian cancer usually spreads to the visceral adipose tissue of the omentum. However, the omental stromal cell-derived molecular determinants that modulate ovarian cancer growth have not been characterized. Here, using next-generation sequencing technology, we identify significantly higher levels of microRNA-21 (miR21) isomiRNAs in exosomes and tissue lysates isolated from cancer-associated adipocytes (CAAs) and fibroblasts (CAFs) than in those from ovarian cancer cells. Functional studies reveal that miR21 is transferred from CAAs or CAFs to the cancer cells, where it suppresses ovarian cancer apoptosis and confers chemoresistance by binding to its direct novel target, APAF1. These data suggest that the malignant phenotype of metastatic ovarian cancer cells can be altered by miR21 delivered by exosomes derived from neighbouring stromal cells in the omental tumour microenvironment, and that inhibiting the transfer of stromal-derived miR21 is an alternative modality in the treatment of metastatic and recurrent ovarian cancer.
Approximately 22,000 new cases of epithelial ovarian cancer have been diagnosed in the United States in 2015 (ref. 1). Over 16,000 deaths per year occurred, making this cancer the most lethal gynaecologic malignancy. Although cancer patients initially respond to platinum- and taxane-based chemotherapy following surgery, most of them experience recurrence within 12–24 months and die of progressively chemotherapy-resistant disease.

One critically important, yet often overlooked, component of tumour progression is the tumour microenvironment, which is primarily composed of fibroblasts, extracellular matrix proteins, endothelial cells and lymphocytic infiltrate. The tumour microenvironment has been shown to directly affect cell growth, migration and differentiation through secreted proteins, cell–cell interactions and matrix remodelling. As it can promote the tumour initiation of normal epithelial cells and facilitate the progression of malignant cells, the tumour microenvironment presents a unique opportunity to discover ways to better diagnose, understand and treat cancer. Recent studies have shown that in addition to initiation via soluble mediators, cell–cell communication can be initiated via surface interactions between circulating exosomes and transmembrane molecules expressed by target cells. The fusion of exosomes with target cell membranes facilitates the transfer of cell surface molecules and receptors from donor to recipient cells. Furthermore, the endocytosis of exosomes by their target cells results in the intracellular release of vesicular contents, including messenger RNA, microRNA (miRNA), proteins and lipids. Tumour exosomes have been shown to have angiogenic properties. For example, colorectal cancer exosomes transfer miRNAs, which promote endothelial cell proliferation and facilitate angiogenesis, whereas glioblastoma-derived exosomes promote tube formation by recipient endothelial cells.

Furthermore, tumour exosomes secrete factors that suppress natural killer cell activity and induce T-cell apoptosis. In this way, the tumour cells themselves create a ‘tumour-friendly’ environment that promotes cancer metastasis and progression.

The transfer of miRNA by exosomes is particularly interesting, because miRNAs are more stable and can control the expression of multiple target genes in the recipient cells. In addition, miRNAs have been shown to regulate cell differentiation, proliferation and apoptosis, and contribute to the development of multiple tumour types. Although the miRNA signatures of tumour-derived exosomes have been identified in multiple tumour types, including ovarian cancer, exosomal miRNA signatures from cancer-associated stromal cells have not been investigated and the functional roles of these exosomal miRNAs in modulating the malignant phenotypes of recipient cancer cells have not been elucidated.

In this study, we use next-generation sequencing to identify differential miRNA signatures in exosomes isolated from ovarian cancer cells and ovarian cancer-associated fibroblasts (CAFs) and adipocytes (CAAs). We demonstrate that specific miRNAs are directly transferred, through exosomes, from CAFs and CAAs to ovarian cancer cells, and we identify the molecular mechanisms by which miRNAs modulate the malignant phenotypes in ovarian cancer cells.

Results

CAFs and CAAs have higher miR21 copy number. miRNAs that transfer between living cells that are involved in cell–cell communication are frequently encapsulated in exosomes, which facilitate their targeted exchange. To identify miRNAs that are transferred by exosomes secreted from omental stromal cells to ovarian cancer cells, we generated miRNA profiles of CAFs, CAAs and ovarian cancer cells by Ion Torrent next-generation sequencing (GEO #GSE77318). Primary CAFs and CAAs were first isolated from ovarian tumour tissue samples and characterized by α-smooth muscle actin staining and adiponectin mRNA expression, respectively. The α-smooth muscle actin expression level in CAFs that we used was significantly higher than that in the normal ovari fibroblasts (Supplementary Fig. 1a). The difference in expression was consistent with that documented in other cancer types. The adiponectin expression levels in the CAAs that we used were significantly lower than those in normal adipocytes (Supplementary Fig. 1b). Similar findings have been reported in adipocytes associated with several cancer types such as gastric cancer and prostate cancer.

Once we characterized the cultured cells that we obtained, we isolated the exosomes from the conditioned media collected from each cell type. Exosomes were first characterized and quantified by electron microscopy, qNano analysis and western blot analysis. The double-layer membrane nature and size of the exosomes were confirmed by electron microscopy and qNano analysis (Fig. 1a,b). A western blot analysis showed the expression of the exosomal markers CD63 and HSP70 but the absence of the cis-Golgi compartment-specific marker GM130 (Fig. 1c). A qNano analysis of media collected from an equal number of cells also demonstrated that a higher number of exosomes was generated from CAFs than from CAAs and ovarian SKOV3 cancer cells (Supplementary Table 1).

Next-generation sequencing was performed on RNA samples extracted from exosomes collected from ovarian cancer cell lines, normal ovari fibroblasts lines, ovarian CAF lines established from primary tumours, CAAs isolated from omental tumours and normal omental adipocytes isolated from omental tissue in patients with benign gynaecologic diseases. The number of sequencing reads for each miRNA and their variants was normalized by adjusting the counts per 1,000 cells (Fig. 1d and Supplementary Data 1). Data analyses identified a diverse population of variants of miRNAs that are collectively known as isomiRNAs (iso-miRs), which are miRNAs that have variations in the sequences with respect to the reference miRNA sequence. The variations include 3′-trimming, 5′-trimming (collectively as sub), 3′-nucleotide addition (super) and nucleotide substitution (variant). The variations of the sequences are mostly non-random and evolutionarily conserved 3′-nucleotide additions or modifications; other iso-miRs appear to be derived from variability in Dicer 1 and Drosha cleavage sites. As iso-miRs share most of their sequence with the most highly expressed form of miRNAs (usually the miRNAs with the conventional sequence), it is possible that these variants share a common set of targets.

A supervised clustering analysis showed distinct miRNA transcriptome signatures among ovarian cancer cells, fibroblasts and adipocytes (Fig. 1d). Among the miRNAs and isomiRs that were identified in exosomes, miR21 and its variants were the most abundant. Among the 84 different forms of miR-21 iso-miR that we identified in exosomes, 8 (including the conventional mature 5′-miR-21 listed in the miRBase) were expressed in all the cell types that we examined. The mature 5′ super variant iso-miR (5′-TAGCTTATCAGACTGATGTTGACA-3′) and the mature 5′ conventional miR21 (5′-TAGCTTATCAGACTGATGTTGAGA-3′) were the most abundant in exosomes isolated from CAFs and CAAs. The expression of mature 5′-miR-21 was second highest in exosomes derived from CAFs and CAAs following the mature 5′ super variant, which had the highest copy number in the exosomes isolated from both CAFs and CAAs. They also had higher copy numbers in normal ovarian fibroblasts, CAFs, normal adipocytes and CAAs than in cancer cells, with CAAs demonstrating the highest copy numbers (Table 1 and Supplementary Data 2), indicating that CAAs produce the highest numbers of exosomal miR21 iso-miRs.
The expression levels of the two isomiRs with the highest levels in CAFs and CAAs (mature 5′ super variant and precursor variant: 5′-TAGCTTATCAGACTGATGTTGACTA-3′), together with that of the mature 5′ conventional miR21, were validated by quantitative reverse transcriptase–PCR (qRT–PCR) analysis. The results showed high concordance between the miRNA-seq and qRT–PCR data. All three miR21 variants showed higher expression levels in the four stromal cell types compared with those in ovarian cancer cells. The mature 5′-miR21 had the largest fold difference in expression levels between CAAs and ovarian cancer cells, whereas the precursor variant had the largest fold difference between CAFs and ovarian cancer cells (Fig. 1a–f and Supplementary Table 2).

Different levels of miR21 in stromal and tumour tissues. To determine whether the higher mature 5′ conventional miR21 copy numbers identified in CAA- and CAF-derived exosomes were due to significantly higher miR21 expression in CAAs and CAFs than in ovarian cancer cells, we performed a qRT–PCR analysis on RNA isolated from primary normal adipocytes (NAs), CAFs, normal ovarian fibroblasts (NFs), CAAs, and ovarian cancer cells. The results showed that both CAAs and CAFs expressed significantly higher levels of endogenous miR21 than did ovarian cancer cells and normal stromal cells (Fig. 2a). To further confirm the differential miR21 expression in the stromal and epithelial components of ovarian tumour tissue, we performed qRT–PCR
analyses on microdissected normal adipocytes isolated from normal omental tissue, CAAs, normal ovarian fibroblasts, CAFs and ovarian cancer cells derived from primary ovarian and omental sites. The results showed that CAAs expressed the highest levels of miR21 (Fig. 2b). In addition, both normal adipocytes and normal ovarian fibroblasts expressed the lowest levels of miR21 and omental ovarian tumours expressed higher levels of miR21 than did those in primary ovarian tumours (Fig. 2b).

To further evaluate the differential expression patterns of miR21 in different components of normal and malignant ovarian tissue, we performed in situ hybridization on an independent set of tissue samples. The results confirmed that normal omental adipocytes and fibroblasts expressed the lowest levels of miR21 (Fig. 2c,d). In addition, the highest levels of miR21 observed in the tumour cells were in the stromal–epithelial interface and the invasion front (Fig. 2e–g), suggesting that miR21 is either upregulated by mediators secreted from neighbouring stromal cells or directly transferred from neighbouring cells. A pairwise comparison demonstrated that in both the tumour and stroma, the cancer in the omental site expressed significantly higher levels

Figure 2 | Differential miR21 expression in ovarian tumour tissue and cancer cell lines. (a) qRT–PCR analysis of miR21 expression in cultured normal ovarian fibroblasts, CAFs, normal adipocytes, CAAs and ovarian cancer cell lines. (b) qRT–PCR analysis of miR21 expression in microdissected normal adipocytes (n = 5), CAAs (n = 9), normal fibroblasts (n = 5), CAFs (n = 5) and ovarian cancer cells from omental (n = 10) and primary ovarian sites (n = 10). ***P < 0.001, **P < 0.01 and *P < 0.05; Mann–Whitney U-test. (c–g) In situ hybridization of miR21 in (c) normal omental adipocytes, (d) normal ovarian tissue and (e) omental and (f) primary ovarian sites of ovarian carcinoma. Strong miR21 staining in omental cancer cells was observed at the tumour–stroma interface (g; dashed line). Digoxigenin-labelled miR21 probes were detected using anti-digoxigenin-alkaline phosphatase and visualized as dark blue nitro-blue tetrazolium and 5-bromo-4-chloro-3′-indolyphosphate precipitate. Sections were counterstained with Nuclear Fast Red. CAAs, cancer-associated adipocytes; CAFs, cancer-associated fibroblasts; T, tumour. Scale bar, 50 µm. (h) Pair-wise comparisons of miR21 expression in omental and primary ovarian tumour sites from stromal (n = 20; P < 0.000; Wilcoxon signed-rank test) and epithelial (n = 19; P = 0.007; Wilcoxon signed-rank test) components of advanced-stage ovarian cancer cells. Relative miR21 expression was measured by in situ hybridization; each line represents the expression level in the same patient. The solid lines indicate higher miR21 expression in the omental site than in the primary ovarian site; the dashed lines represent lower expression in the omental site. (i) Box plot showing miR21 expression in stromal (n = 14) and epithelial (n = 14) components of primary and recurrent omental ovarian tumour tissue, as quantified by in situ hybridization analyses. ***P < 0.01; Mann–Whitney U-test. (j) qRT–PCR of miR21 expression in ovarian cancer HeyA8 and SKOV3 cells and their paclitaxel-resistant sublines HeyA8-MDR and SKOV3-TR. The results were the average from at least three separate experiments. Mean ± s.d.; **P < 0.01; two-tailed Student’s t-test. (k) qRT–PCR of miR21 expression in the primary PEA1 tumour cells and the cell line established after cisplatin treatment (PEA2). The results were the average from at least three separate experiments. Mean ± s.d.; **P < 0.01; two-tailed Student’s t-test.
of miR21 than did the cancer in the primary site in the same patient \((P<0.001\) and \(P=0.007\), respectively) (Fig. 2h). Moreover, recurrent ovarian cancer showed significantly higher levels of miR21 expression than did primary ovarian cancer in both the stromal and the epithelial components of ovarian cancer tissue (Fig. 2i). In vitro, increased miR21 expression levels were observed in the chemoresistant cell lines HeyA8 MDR and SKOV3TR compared with those in their parental lines HeyA8 and SKOV3, respectively \((P<0.01)\) (Fig. 2j). Furthermore, a higher miR21 expression level was observed in the cell line PEA2, established from the recurrent tumour of a pretreated patient \((P<0.01)\) (Fig. 2k). The increase in miR21 expression levels in recurrent or chemoresistant ovarian tumour cells was much lower in vitro (<1.5-fold) than in vivo (>10-fold), suggesting that the stromal cells in the tumour microenvironment increase miR21 expression levels in ovarian cancer cells through the direct transfer of miR21 or transcriptionally by soluble mediators.

Direct transfer of miR21 from CAF and CAA to cancer cells.

To determine whether miR21 can be directly transferred from CAs or CAFs to ovarian cancer cells, we co-cultured ovarian cancer SKOV3ip cells, which have a comparable miR21 expression level as SKOV3 cells, with CAs and CAFs that were transiently transfected with fluorescein amidite (FAM)-tagged miR21. Confocal microscopy detected fluorescently labelled miR21 in the cancer cells co-cultured with CAs and CAFs but not in those co-cultured with mock-transfected CAs and CAFs, suggesting that miR21 was transferred from the CAs and CAFs to the cancer cells (Fig. 3a). To further delineate whether the transfer of miR21 is mediated by exosomes, we first isolated exosomes from conditioned media collected from CAs and CAFs that had been transiently transfected with FAM-tagged miR21 or the pre-miR negative control. The purified exosomes were then added to SKOV3ip cells. Confocal microscopy detected fluorescently labelled signals in the cancer cells incubated with exosomes obtained from FAM-tagged miR21-transfected cells but not in those incubated with exosomes from mock-transfected cells (Fig. 3b), confirming that miR21 was transferred from CAs and CAFs to ovarian cancer cells via exosomes.

Next, we determined whether CAA-derived exosomes, which contained higher levels of miR21 than did those from CAFs, transferred more miR21 to recipient cells. Ovarian cancer SKOV3 cells were treated with the same number of exosomes isolated from CAs and CAFs. A qRT–PCR analysis of miR21 was performed of the recipient cells. The results showed that miR21 expression levels were higher in SKOV3 cells treated with exosomes isolated from CAs than in those treated with exosomes from CAFs (Supplementary Fig. 2a). These data suggest that the amount of miR21 transferred to the recipient cells depends on the amount of miR21 delivered in the exosomes.

To determine whether CAF-derived miR21 can be transferred to ovarian cancer cells in vivo, we injected red mCherry-labelled ovarian cancer SKOV3ip cells (Fig. 3c) subcutaneously into nude mice. Once the tumour nodules had formed\(^1\), miR21-/-/miR21-/- mouse embryonic...

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**Figure 3 | Exosomal transfer of miR21 from adipocytes and fibroblasts to ovarian cancer cells.** (a) CAs and CAFs transiently transfected with FAM-tagged miR21 (miR21-FAM) or without transfection (Control) were co-cultured with mCherry-labelled ovarian cancer SKOV3ip cells for 24 h. Fluorescence microscopy was used to detect the green and red fluorescent signals in SKOV3ip cells. Scale bar, 2 µm. (b) Exosomes were isolated from conditioned media prepared from CAs and CAFs transfected with FAM-labelled miR21 (miR21-FAM) or without transfection (Control) and added to ovarian cancer SKOV3ip cell cultures. SKOV3ip cells were fixed and the nuclei were stained with 4,6-diamidino-2-phenylindole (DAPI) blue. An SP5 confocal microscope was used to detect the green signals in SKOV3ip cells. Scale bar, 2 µm. (c–h) Red mCherry-labelled ovarian cancer SKOV3ip cells (e) were injected subcutaneously into nude mice, to establish tumours. After 1 week, miR21-/-/miR21-/-MEFs transfected with FAM-tagged miR21 (d) were injected intratumorally. After 24 h, tumours were harvested and frozen sections were prepared. A confocal microscopy analysis showed the blue DAPI signals for the nuclei of the two cell types (e), the red mCherry signals for the SKOV3ip cells (f) and the green FAM signals for miR21 (g). Arrowheads indicate the FAM-miR21 signals in the peripheral stromal cells (S) and in some of the red cancer cells (T) (g,h). Scale bar, 5 µm.
fibroblasts (miR21/−/miR21−/− MEFs) were transiently transfected with FAM-tagged miR21 (Fig. 3d) and injected adjacent to the tumour nodules. Tumours were harvested 24 h after fibroblast injection and frozen sections were prepared. Confocal microscopy showed blue 4,6-diamidino-2-phenylindole signals of the round nuclei, indicating the presence of intact cells (Fig. 3e). Ovarian cancer cells were identified by red mCherry labelling, which appeared as granular structures in non-fixed frozen sections (Fig. 3f). Green FAM labelling signals, which represented individual miR21 molecules, were observed in the peripheral stromal cells as well as in some of the cancer cells (Fig. 3g,h). These results suggest that miR21 can be transferred from stromal fibroblasts to cancer cells in vivo.

As transforming growth factor (TGF-β), a well-known regulator of the cancer–stroma interaction, was reported to regulate miR21 expression in pulmonary artery smooth muscle cells20 and pulmonary fibroblasts21, we determined whether increased miR21 expression in ovarian cancer cells was due to TGF-β produced by the stromal cells, which upregulated endogenous miR21 in ovarian cancer cells rather than exosomal transfer. We treated ovarian cancer SKOV3 and OVCA432 cells and CAFs with TGF-β and quantified the miR21 expression levels in these cells by qRT–PCR analysis. The results showed that there were no significant increases in miR21 expression levels in the ovarian cancer cells (Supplementary Fig. 2b) or CAFs (Supplementary Fig. 2c). Thus, the increase in the miR21 expression level in cancer cells was primarily due to the direct transfer of miR21 from neighbouring stromal cells in the microenvironment via exosomes instead of TGF-β stimulation.

miR21 shows no effect on cancer cell proliferation. To determine whether miR21 stimulates ovarian cancer cell proliferation in vitro, ovarian cancer SKOV3 and OVCA432 cells were incubated for 72 h after transient transfection of miR21 precursor or control miR. The relative cell number of SKOV3 and OVCA432 cells shows no significant changes with or without miR21 expression (Supplementary Fig. 3a).

To further evaluate the effect of miR21 on ovarian cancer cell proliferation in vivo, luciferase-labelled SKOV3ip ovarian cancer cells and miR21+/−/miR21+ MEF (miR21-MEF) or miR21−/−/miR21− MEF (MEF) cells were subcutaneously injected into female BALB/c athymic nude mice, to establish tumours. The tumour volumes were measured and quantified 5 days post injection. The luciferase activity of the tumour established shows no significant changes between the miR21-MEF group and the MEF group (Supplementary Fig. 3b).

miR21 stimulates cancer cell motility and invasion. Besides cell proliferation, we determined whether miR21 modulated other malignant phenotypes such as the motility and invasive potential of ovarian cancer cells. The ovarian cancer cells ALST, OVCA433 and SKOV3 were transfected with miR21 precursor (pre-miR21) or negative control (control miR) and seeded onto a transwell plate coated with type I collagen matrix. The cells were allowed to invade for 24 h. Invading cells were then fluorescently labelled with calcine and quantified. The results showed a significant increase in invasive potential in all three cell lines when they were transfected with miR21 compared with the mock transfectants (Supplementary Fig. 4a). An in vitro scratch assay was also performed to evaluate the effect of miR21 on cell motility. The results showed that all three cell lines demonstrated significantly faster wound closure rates in miR21-transfected cells than in mock-transfected cells, suggesting that miR21 enhanced the motility potential of ovarian cancer cells (Supplementary Fig. 4b).

miR21 lowers chemosensitivity and apoptosis in cancer cells. To determine whether exosomal miR21 confers chemoresistance in ovarian cancer cells, we isolated exosomes from miR21+/−/miR21− MEFs and miR21+/−/miR21+ MEFs. They were quantified by qNano (Fig. 4a) and equal numbers of exosomes were added to two ovarian cancer cell lines, OVCA432 and SKOV3. A significantly higher level of miR21 was confirmed in miR21+/−/miR21+ MEF exosome-treated cells than in miR21+/−/miR21− MEF exosome-treated cells (Fig. 4b). Both OVCA432 and SKOV3 cells were treated with 20 nM paclitaxel. The results showed a significant decrease in cell number in cells treated with miR21+/−/miR21+ MEF exosomes compared with those treated with miR21+/−/miR21− MEF exosomes (Fig. 4c).

To further evaluate the role of miR21 in chemoresistance, we directly transfected OVCA432 and SKOV3 ovarian cancer cells, which have low levels of endogenous miR21 expression, with miR21 precursor (pre-miR21) or the negative control and treated them with paclitaxel. The resulting miR21 expression levels in the transfected cells were comparable to the physiological level (Supplementary Fig. 5). The cells transfected with the miR21 precursor showed significantly lower chemosensitivity than did the mock transfectants (Fig. 4d). In addition, OVCA432 and SKOV3 cells transfected with the miR21 precursor had a lower rate of apoptosis than did the mock transfectants (Fig. 4e). Next, we determined whether overexpression of the other two most abundant miR21 variants that we identified had the same biological effect on ovarian cancer cells. Both miR21 mature 5′ super variant and precursor variant, together with the mature miR21, were transfected into SKOV3 and OVCA432 cells and treated with paclitaxel. All three miRNAs demonstrated a significant increase in IC50 in the transfected cells (Supplementary Fig. 6a), suggesting that the two miR21 variants have similar biological activities to those of the mature miR21.

To demonstrate the role of miR21 in chemoresistance in ovarian cancer cells in vivo, luciferase-labelled ovarian cancer SKOV3ip cells were co-injected with miR21+/−/miR21+ MEFs (miR21-MEF) or miR21−/−/miR21− MEFs (MEFs) into female nude mice, followed by a sub-optimal dose of paclitaxel (7.5 nM). SKOV3ip cells alone were also injected as a negative control. We found that the luciferase activity of both the SKOV3ip cell-alone group and the MEF group were significantly lower than that of the miR21-MEF group after the same paclitaxel treatment, suggesting that tumour cells in the miR21-MEF group are more resistant to paclitaxel than those in the SKOV3ip cell-alone group and the MEF group (Fig. 4f.g).

APAF1 is a direct downstream target of miR21 in cancer cells. To delineate the molecular mechanisms underlying the role of miR21 in conferring chemoresistance in ovarian cancer cells, we performed transcriptome profiling on SKOV3 cells transfected with pre-miR21 or the pre-miR-negative control (GEO#GES51457). Pathway analyses identified a set of chemoresistance-associated genes (Fig. 5a,b and Supplementary Table 3). Among these genes, APAF1, which has been shown to be associated with chemoresistance and apoptosis22–24, was selected for further validation studies. To confirm that APAF1 can be downregulated by miR21 delivered in CAA- or CAF-derived exosomes, we incubated SKOV3 cells with exosomes isolated from CAAs and CAFs. qRT–PCR analyses showed that miR21 expression levels in SKOV3 cells incubated with CAAs and CAFs were significantly lower than in cells incubated with complete media only (Supplementary Fig. 2a). In contrast, APAF1 mRNA levels were significantly lower in SKOV3 cells incubated with exosomes derived from miR21-transfected CAAs and CAFs (P < 0.001) (Fig. 5c). These data suggest that
exosomes derived from miR21-transfected CAAs and CAFs can significantly increase miR21 and lower APAF1 expression in recipient ovarian cancer cells. This finding is further supported by the immunohistochemistry data that showed lower APAF1 expression in the stromal–epithelial interface (Fig. 5d), in some of the cases (n = 5) which expressed high levels of stromal miR21 (Fig. 2e–g).

To determine whether APAF1 is a direct downstream target of miR21, we performed a qRT–PCR analysis of ovarian cancer OVCA432 and SKOV3 cells transfected with the miR21 precursor or the negative control, to evaluate its effect on APAF1 expression. The results showed significantly lower APAF1 expression in both cell lines transfected with the miR21 precursor than in the control (Fig. 5e). A similar effect was also observed in cells transfected with the two miR21 variants (miR21 mature 5′ super variant and precursor variant) (Supplementary Fig. 6b). A western blot analysis was then performed to show downregulation of the APAF1 protein in both cell lines (Fig. 5f). The inverse correlation between miR21 and APAF1 mRNA expression was further confirmed by qRT–PCR analyses of the epithelial components of 38 microdissected ovarian cancer tissue samples (R = −0.493, P = 0.002) (Fig. 5g). In addition, PEA2 recurrent ovarian cancer cells, which showed significantly higher miR21 expression than did PEA1 primary cells (Fig. 2g),
The species was a potential target for miR21 (Fig. 5i). Subsequently, an APAF1 coding sequence (5′-GACATGGAAACTGAAGAG-3′) that had a significant homology among different species was a potential target for miR21 (Fig. 5i). Subsequently, the potential miR21 binding sequence was cloned into the luciferase vector and co-transfected into OVCA432 cells in the presence of different concentrations of pre-miR21. The results showed a significant dose-dependent decrease in luciferase activity in cells transfected with pre-miR21 (Fig. 5j). In addition, the effect of pre-miR21 on luciferase activity was abrogated when cells were transfected with a mutated APAF1 coding sequence (Fig. 5k). These data suggest that miR21 directly controls APAF1.
mRNA expression through binding to the APAF1 coding sequence, and that APAF1 is a direct target of miR21.

APAF1 mediates miR21-induced chemoresistance in cancer cells. To determine whether high levels of APAF1 confer paclitaxel sensitivity to ovarian cancer cells, we transiently transfected SKOV3 and OVCA432 ovarian cancer cells with the full-length APAF1 construct or the control vector pcDNA3.1 and treated them with various concentrations of paclitaxel. Both cell lines demonstrated a significant decrease in IC₅₀ in cells transfected with the APAF1 construct compared with the mock transfectants (Fig. 6a). These data suggest that APAF1 sensitized the ovarian cancer cells to paclitaxel treatment. To determine whether upregulation of APAF1 can abrogate miR21-conferred paclitaxel resistance in ovarian cancer cells, we transfected both SKOV3 and OVCA432 cells with pre-miR21 and APAF1 or with pre-miR21 and the empty vector. APAF1 partially abrogated paclitaxel resistance (Fig. 6b), suggesting that besides downregulation of APAF1, other mechanisms are involved in mediating the effect of miR21 on paclitaxel resistance in ovarian cancer cells. The results were further confirmed in OVCA432 cells that were stably transfected with a doxycycline-inducible APAF1 construct (Supplementary Fig. 7).

To evaluate the effect of APAF1 on paclitaxel resistance in ovarian cancer cells in vivo, we established OVCA432 cells stably expressing doxycycline-inducible APAF1 or the control vector and injected them intraperitoneally into female nude mice, followed by paclitaxel treatment. The results showed that the luciferase activity of the APAF1-overexpressing group was significantly lower than that of the control group after the same paclitaxel treatment (Fig. 6c,d). Elevation of APAF1 expression in the tumour tissues from the APAF1-overexpressing group was confirmed using immunohistochemical analysis (Fig. 6e). These data suggest that APAF1 confers paclitaxel resistance in ovarian cancer cells.

Discussion

In this study, we showed, for the first time, that CAAs and CAFs express significantly higher levels of miR21 isomiRs than do ovarian cancer cells, and that the exosomal transfer of miR21 isomiRs from CAAs or CAFs to neighbouring cancer cells can increase the chemoresistance of these cells to paclitaxel through the downregulation of miR21’s direct target APAF1 (Supplementary Fig. 8).

The bidirectional signalling mediated by secreted proteins that occurs between ovarian cancer cells and the omental fibroblasts and adipocytes not only effectively ‘reprogrammes’ fibroblasts and adipocytes to create an optimal microenvironment for tumour implantation but also confers a more aggressive phenotype to the cancer cells that allows them to adapt to their

![Figure 6](image-url)
new microenvironment. Recent studies have shown that cells can communicate through the exchange of bioactive molecules via microvesicles. These small vesicles include ectosomes (100–1,000 nm) and exosomes (30–100 nm), which are shed or secreted by almost all cell types. Exosomes are endosome-derived microvesicles that are secreted as part of multivesicular bodies into the extracellular space. Secreted exosomes can be engulfed by tissues locally or carried in biological fluids to distant sites in the body. Intercellular communication can be initiated through interactions between circulating exosomes and the transmembrane proteins of target cells. Target cells engulf exosomes through endocytosis, resulting in the intracellular release of bioactive molecules, which include membrane proteins, cytoplasmic proteins and lipids.

Recent studies have shown that exosomes also contain non-coding RNAs such as miRNAs. MiR21 has been identified in exosomes derived from breast cancer cells, dendritic cells, T cells and serum samples. Using Ion Torrent next-generation sequencing of miRNA in exosomes, we identified significantly larger copy numbers of miR21 isoforms, in particular the mature 5’ and 3’ super variants, in exosomes isolated from CAFs and CAAs than in those isolated from ovarian cancer cells. This finding was probably due to higher endogenous miR21 expression in CAAs and CAFs than in cancer cells, as we confirmed by qRT–PCR analyses of microdissected tissue samples and in situ localization of miR21 in tissue sections. Significantly lower miR21 expression levels were detected in both normal fibroblasts and adipocytes than in ovarian cancer cells by a qRT–PCR analysis of microdissected tissue samples and in situ hybridization. However, miR21 expression levels in ovarian cancer cells and normal stromal cells in vitro were found to be comparable. The discrepancy may be due to miR21 being upregulated when quiescent normal stromal cells, in particular fibroblasts isolated from tissue samples, grow in culture. In addition, the lack of miR21 transfer from neighbouring CAFs or CAAs may lead to lower miR21 expression in cultured ovarian cancer cells.

By incubating ovarian cancer cells with purified exosomes from CAFs transfected with labelled miR21, we confirmed that miR21 was successfully transferred to the cancer cells. The transfer of miR21 from CAFs to cancer cells was further confirmed by co-culturing labelled miR21-transfected CAFs with ovarian cancer cells both in vitro and in vivo. Increased miR21 expression in cancer cells in the stroma–cancer interface also suggests that miR21 can be transferred from the stromal cells to the neighbouring cancer cells.

miRNAs are a family of short (20–24 nt) non-coding RNAs that posttranscriptionally regulate gene expression in multicellular organisms by affecting both the stability and translation of mRNAs. MiR21 expression has been linked to resistance to a variety of chemotherapeutic agents in both solid and haematologic tumours. By silencing the expression of tumour suppressor genes including PTEN, PDCD4 and p21, miR21 overexpression promotes cell survival and confers resistance to tamoxifen, faslodex and topotecan in breast cancer. Furthermore, miR21 expression is associated with resistance to gemcitabine, 5-fluorouracil and docetaxel in pancreatic, colorectal and prostate cancers, respectively. MiR21 can be transferred by exosomes from stromal cells to cancer cells and can decrease paclitaxel sensitivity, as shown in this study, suggesting that increased miR21 expression in ovarian cancer cells, in particular those in the stroma–cancer interface, may confer a more aggressive and chemoresistant phenotype in those cells than in those located inside the tumour mass. A transcriptome profiling analysis and subsequent validation studies demonstrated a significant decrease in APAF1 in miR21-transfected ovarian cancer cells. APAF1 binds with cytochrome c and dATP to form an apoptosis, which in turn activates caspases 9 and 3 to initiate apoptosis. APAF1 deficiency has been documented to confer chemoresistance in several cancer types. Thus, downregulation of APAF1 by miR21 may increase paclitaxel resistance in ovarian cancer cells. This was confirmed by co-transfecting ovarian cancer cells with both miR21 and APAF1 in the presence of paclitaxel. However, APAF1 cannot completely rescue the effect of miR21 on paclitaxel resistance, suggesting that other miR21 target genes are involved. APAF1 is not a known direct target of miR21; we also failed to demonstrate any sequences in miR21 that were complementary to the 3’-untranslated region of the APAF1 mRNA. Instead, we showed, for the first time, that an APAF1 coding sequence was a potential target for miR21. Further studies confirmed that miR21 directly controls APAF1 mRNA expression through binding to the APAF1 coding sequence, and that APAF1 is a direct target of miR21.

Our transcriptome analysis of miR21-transfected ovarian cancer cells showed upregulation of MMP1, which may lead to an increase in the invasion potential of ovarian cancer cells, as we observed in our in vitro studies. MMP1 is a member of the MMP family, which is associated with invasion and metastasis. MMP1 is not a known target of miR21; however, a pathway analysis showed that multiple transcription factors that have been shown to regulate MMP1 were up- or downregulated in miR21-transfected SKOV3 cells. Among them, smad7 has been shown to be a direct target of miR21 (ref. 37) and a negative regulator of TGF-induced MMP1 expression, suggesting that miR21 regulates MMP1 expression indirectly by targeting genes in the TGF signalling pathways.

In conclusion, our results show that exosomal miR21 can confer chemoresistance and an aggressive phenotype in ovarian cancer cells through its transfer from neighbouring stromal cells, suggesting that preventing the exosomal transfer of miR21 from stromal cells is a new strategy for suppressing ovarian cancer growth. In addition, the identification of APAF1 as a direct target of miR21 and APAF1 as a mediator of miR21 for conferring chemoresistance in ovarian cancer suggests that strategies based on the upregulation of APAF1 in ovarian cancer cells can be used to sensitize ovarian cancer cells to paclitaxel treatment.

Methods

Microdissection of tissue samples. Ovarian tissue samples were obtained from the ovarian cancer repository of the Department of Gynecologic Oncology and Reproductive Medicine under protocols approved by the institutional review board of The University of Texas MD Anderson Cancer Center (Houston, Texas). Normal omental tissue samples were obtained from patients with benign gynaecologic diseases in the Department of Obstetrics and Gynecology under protocols approved by the institutional review board of Baylor College of Medicine. Informed consent was obtained from all patients. Stromal and epithelial components of normal and malignant ovarian tissues were obtained by microdissection. Microdissection was conducted by fixing tissue sections in 70% ethanol, staining them with 2% methyl green to visualize the histologic features. During dissection, the areas of interest in the sections were carefully outlined. Areas with immune cell and blood vessel infiltration were excluded, to minimize contamination.

Isolation of primary fibroblasts and adipocytes. Normal and CAF primary cultures were derived from normal ovaries from patients with benign gynaecologic diseases and from high-grade serous ovarian cancer tissue, respectively, using the explant technique. In brief, tissue samples were cut into 1-mm3 fragments and seeded into T25 tissue culture flasks with 2-ml tissue culture media. After 1 week, an additional 3 ml of culture media were added. Once the flask became 50% confluent, the explants were removed and the epithelial cells were scraped from the flasks under the microscope using a cell scraper. Cells were then subcultured into two T25 flasks. After two to three passages, the percentage of fibroblasts was determined by immunohistochemistry.

Normal adipocytes and CAA primary cultures were derived from the omental tissues of patients with benign gynaecologic diseases and high-grade serous ovarian...
cancer, respectively. Adipose tissue was first digested with collagenase (Sigma-Aldrich Co., St Louis, MO, USA) in HEPES. The digested tissue was then centrifuged. The pellet containing adipocytes, fibroblasts and red blood cells was removed, while the top fatty layer was washed in fresh DMEM/F12 medium twice, with centrifugation, before being transferred to the cell culture flasks. The flask was then re-inverted so that the bottom of the flask was on top. The floating mature adipocytes were attached to the upper portion of the flasks, while any fibroblast-like cells were sunk to the bottom. After 5–7 days to allow sufficient attachment of the mature adipocytes, the media inside was replaced and replaced with 5 ml of fresh media. The flasks were then re-inverted for normal observation and manipulation. The mature adipocytes isolated were characterized using Oil Red O staining.

**Cell lines and culture conditions.** The human ovarian cancer cell lines A2780 (Sigma-Aldrich Co.), ALST (gifts from Dr Michael Birrer’s laboratory at the Massachusetts General Hospital), OVCAC432, OVCAC433 (gifts from Dr Robert Bast’s laboratory at the MD Anderson Cancer Center), OVCAR5 (gifts from Dr Gordon Mills’s laboratory at the MD Anderson Cancer Center), HeyA8, HeyA8-MDR, SKOV3ip3, SKOV3-TR (gifts from Dr Anil Sood’s laboratory at the MD Anderson Cancer Center) and SKOV3 (American Type Culture Collection) were maintained in RPMI 1640 medium supplemented with 10% FBS, 2 mM glutamine and penicillin/streptomycin (Life Technologies Corp., Grand Island, NY, USA). The human ovarian cancer cell lines PEA1 and PEA2 (European Collection of Authenticated Cell Cultures) were maintained in RPMI 1640 medium supplemented with 10% FBS, 2 mM glutamine, penicillin/streptomycin and 1% sodium pyruvate (Sigma-Aldrich Co.). Normal adipocytes and CAAs were established from the omental tissue removed from patients with benign gynaecologic diseases and ovarian cancer, respectively. They were cultured in DMEM medium supplemented with 10% FBS, normal ovarian fibroblasts and CAFs were established from normal ovaries and ovarian cancer tissue removed from patients with benign gynaecologic diseases and ovarian cancer, respectively. They were cultured in 1:1 MCD105/199 medium (Sigma-Aldrich Co.) supplemented with 10% FBS, 1.0 mg ml⁻¹ epidermal growth factor and penicillin/streptomycin (Life Technologies Corp.). MG63 cells (gift from Dr Yung Li’s laboratory at the University of Louisville) were cultured in RPMI 1640, supplemented with 10% FBS and non-essential amino acid (Life Technologies Corp.). pEXZ-miR21 plasmids (GeneCopeia, Rockville, MD, USA) were stably transfected into miR21-/-MEF cells to overexpress miR21. The miR21 stably overexpressed miR21-/-MEF cells were annotated as miR21-/-MEF. Empty pEXZ vectors were also stably transfected into miR21-/-MEF cells as negative controls. All cells were tested negative for mycoplasma contamination and were authenticated by short tandem repeat repeat profiling in the Characterized Cell Line Core at the University of Texas MD Anderson Cancer Center.

**Isolation of exosomes.** After cell cultures reached 90% confluency, cells were washed with PBS and incubated with freshly prepared complete medium containing exosome-free FBS for 4 h. Exosomes were isolated from conditioned medium by differential centrifugation. In brief, conditioned medium was centrifuged at 300 x g for 20 min at 4 °C to remove cells, followed by filtration through a 0.22-µm filter to remove cell debris. Exosomes were pelleted by ultracentrifugation at 100,000 g for 90 min. They were resuspended in PBS, as described by ultracentrifugation again at 100,000 g for 90 min. The size and concentration of the exosomes were quantified by qNano (Qion Science, New Zealand) and exosomes were ready for RNA/protein extraction or cell treatment.

**Transmission electron microscopy.** For electron microscopy analysis, exosomes were isolated by mixing them with an equal volume of 4% paraformaldehyde. Samples were deposed onto Formvar-carbon-coated electron microscopy grids, washed with PBS and further fixed with 1% glutaraldehyde for 5 min. Samples were contrasted first in a solution of uranyl oxalate, pH 7, for 5 min and then contrasted and embedded in a mixture of 4% uranyl acetate and 2% methyl cellulose for 10 min on ice. The samples were observed using a JEM 1010 transmission electron microscope (JEOL USA, Inc., Peabody, MA) at 80 kV.

**Western blot analysis.** Cell extracts were prepared in RIPA buffer (20 mM sodium phosphate, 150 mM NaCl pH 7.4, 1% Nonidet P-40, 0.1% SDS and 0.5% deoxycholic acid) containing complete protease inhibitor mixture (Roche). Proteins were separated on SDS–polyacrylamide gels and electrophoretically transferred to an Immobilon polyvinylidene fluoride membrane (EMD Millipore, Billerica, MA, USA). The membranes were incubated with primary antibodies overnight at 4 °C, followed by incubation with appropriate horse-radish peroxidase-conjugated secondary antibodies at 1:10,000 dilution (Thermo Fisher Scientific, Grand Island, NY, USA) for 1 h at ambient temperature. Signals were developed using ECL chemiluminescence detection reagents (GE Healthcare Life Sciences, Pittsburgh, PA, USA) and visualized on X-ray film (Fujiﬁlm, Tokyo, Japan). The following primary antibodies were used for western blotting: anti-CDC6 (1:1,000; clone RFA4, EMD Millipore), anti-GM130 (1:1,000; 610822, BD Biosciences, San Jose, CA, USA), anti-HSP70 (1:1,000; MAB3516, EMD Millipore), anti-APAF1 (1:1,000; ab2, Sigma-Aldrich Co.) and anti-β-actin (1:5,000; clone AC-15, Sigma-Aldrich Co.). Original images of all blots are supplied as Supporting Figures 9 and 10.

**RNA sequencing using Ion Torrent.** Exosomes were isolated from three CAF and two CAA primary cultures and four ovarian cancer cell lines (A2780, HeyA8, OVCAC433 and SKOV3) following the procedure as described above. Total RNAs were extracted from purified exosomes using the RNAqueous-Micro Kit (Thermo Fisher Scientific) and the amount and quality of small RNA in the total RNA samples were determined using the Agilent small RNA kit on an Agilent 2100 Bioanalyzer. Five nanograms of small RNAs were subjected to library construction using the Ion Total RNA-Seq kit v2 (Life Technologies Corp.) according to the small RNA protocol. In brief, the RNA was first hybridized and ligated with oligo adaptors at 16 °C for 16 h, followed by a reverse-transcription reaction. Size selection of the complementary DNA was performed with a Novex (Life Technologies Corp.) gel kit. After hybridization, the labelled small RNA was ligated to 200-bp adaptors, the RNA fragments of 60–80 nt were cut from the gel and subjected to PCR amplification for 18 cycles. The amplified product was purified using the PoreLink PCR micro kit (Life Technologies Corp.) and then prepared for sequencing using the Ion sequencer kit protocol (Life Technologies Corp.). The complete reads were loaded on the Ion S5 314 chip and sequenced on the Ion Torrent Personal Genomic Machine for 65 cycles.

Ion Torrent reads were collected using Ion Torrent Suite software, which also sorted the data according to the barcodes. The software scores the quality of the reads by assigning Q17 and Q20 scores according to the quality scoring system. Fastq files from the Ion Torrent server were imported to the CLC Genomics Workbench (version 5.1; Life Technologies Corp.). Sequence reads were trimmed to remove reads with ambiguous nucleotides and to create a sequence length of 15–55 nucleotides. Trimmer sequence reads were aligned to an mRNA database (miRBase, release 18). Mapped reads were annotated and counted. The median number of reads was 182,000 for each sample. Ion Torrent reads were then normalized to the number of cells in each cell line used for exosome collection and presented as the total reads per 1,000 cells.

**Quantitative RT-PCR analysis.** Total RNA was extracted from cultured cells using TRI reagent (Molecular Research Center, Cincinnati, OH, USA) or from tissue samples and purified exosomes using the RNAqueous-Micro Kit (Thermo Fisher Scientific).

For exosomal miR21 expression analysis, 0.5 µg of total RNA was first reverse transcribed using the QuantisMir RT kit (System Biosciences, Camarillo, CA, USA). qRT–PCR analysis was performed using iTaq Universal SYBR green super mix (Bio-Rad Laboratories Inc., Hercules, CA, USA) on a CFX Touch real-time PCR detection system (Bio-Rad Laboratories Inc.). The sequence of mature 5’ conventional miR21 (5’-TAGCTTATCAGACTGATGTTGACTA-3’), mature 5’ modified miR21 (5’-TAGCTTATCAGACTGAGTTGACA3’), 5’-TACCAGCAGTCTGAGTTGACA3’), mature 5’-miR21 precursor variant 5’-TACCAGCAGTCTGAGTTGACA3’), and the 5’ universal reverse primer was provided by the QuantisMir RT kit. For cellular miR21 expression analysis, 5 ng of total RNA was first reverse transcribed using the TaqMan microRNA reverse transcription kit (Thermo Fisher Scientific) and miR21 (000397) and U6 (001973) snRNA-specific primers and probes. The relative standard curve method (ΔCt method) was used to determine relative miR21 expression using U6 snRNA as the reference.

For the miRNA expression analysis, 0.5 µg of total RNA was used to synthesize the first strand of cDNA using the ImProm-II Reverse Transcription System (Promega Corp., Madison, WI, USA). The expression of APAF1 was determined by multiplexing quantitative PCR (TaqMan Gene Expression Assay) using FAM-labelled APAF1 (Hs00559441_m1) together with VIC-labelled glyceraldehyde 3-phosphate dehydrogenase-specific TaqMan probes and primers. The relative standard curve method (2-ΔCt) was used to determine the relative mRNA expression, using glyceraldehyde 3-phosphate dehydrogenase as the reference.

**In situ hybridization of miR21.** In situ hybridization was performed to determine the expression of miR21 using miCURY LNA microRNA Detection, Optimization Kit 2 (mir-21) (Exiqon, Denmark) according to the manufacturer’s protocol. In brief, slides were dewaxed, deparaffinized and digested with 15 µg ml⁻¹ proteinase-K at 37 °C for 10 min. Slides were hybridized with 40 nM miR21 or scramble probes in an incubation
chamber at 53 °C for 1 h, followed by stringent washes with SSC buffers at 53 °C. For immunodetection, slides were blocked with digoxigenin blocking reagent (Roche, Switzerland) in maleic acid buffer containing 2% sheep serum at ambient temperature for 15 min and then incubated with sheep anti-digoxigenin conjugated to alkaline phosphatase (diluted 1:800 in blocking reagent (Roche)) at ambient temperature for 60 min. Nitro-blue tetrazolium and 5-bromo-4-chloro-3’-indolyl-phosphate reaction mixture (Roche) with 0.2 mM levamisole (Sigma-Aldrich Co.) were applied to the slides at room temperature at 30 °C. After 2 h of incubation, the reaction was stopped with KTBT buffer. The slides were then counterstained with FastRed nuclear staining reagent, rinsed with tap water, dehydrated and mounted for microscopy. Five random microscope fields per section were evaluated at ×20 original magnification and the total optical density of every visual field was calculated. For each patient, the same procedure was carried out using a 0.1-nM U6 snRNA probe to confirm the RNA integrity.

**In vitro detection of miR21 transfer.** SKOV3ip cells were stably transfected with red mCherry (pBabe vector). For the co-culture experiment, CAFs and CAAs were grown on the 0.4 μm pore size transwell (Thermo Fisher Scientific) and transfected with 10 nM green fluorescent 3’ FAM-labelled miR-21 oligo (Sigma-Aldrich Co.) for 24 h. Cells were then put together with red fluorescent mCherry-labelled ovarian cancer SKOV3ip cells that had been grown on the coverslip in the bottom well of the transwell. After 24 h, the SKOV3ip cells were fixed with 4% paraformaldehyde. Fluorescent microscopy was used to detect the green signals in the SKOV3ip cells.

For the exosome treatment experiment, CAFs and CAAs were transfected with 10 nM green fluorescent 3’ FAM-labelled miR-21 oligo (Sigma-Aldrich Co.) for 24 h. Cells were washed with PBS and incubated with freshly prepared complete medium containing exosome-free FBS for 48 h. CAF- and CAA-conditioned medium was collected and exosomes were isolated from the conditioned medium by differential centrifugation following the procedure described above. The pellet was suspended in serum-free medium and used to test the transferrability of ovarian cancer SKOV3ip cells grown on cover slips. After 24 h, the SKOV3ip cells were fixed with 4% paraformaldehyde and the nuclei were stained with 4,6-diamidino-2-phenylindole blue. SP5 confocal microscopy was used to detect the green signals in the SKOV3ip cells.

**In vivo detection of miR21 transfer.** Red mCherry-labelled ovarian cancer SKOV3ip cells (2 × 10⁵) were injected subcutaneously into female BALB/c athymic nude mice at the age of 6 weeks, to establish tumours. After 1 week, SKOV3ip cells were transfected with 10 nM green fluorescent 3′ FAM-labelled miR-21 oligo (Sigma-Aldrich Co.). SKOV3ip cells (2 × 10⁵) were injected intratumorally. Mice were killed using carbon dioxide chamber followed by cervical dislocation 24 h after and tumours were harvested and prepared for frozen sectioning. The frozen sections were unfixed and mounted with antifade reagent. Confocal microscopy was used to detect the green signals in the SKOV3ip cells. The described animal procedures have been reviewed and approved by the institutional animal care and use committee of the University of Texas MD Anderson Cancer Center.

**MTT assay.** SKOV3 and OVCA432 cells were seeded in a 96-well plate and then subjected to transient transfection for 24 h. After 72 h of paclitaxel treatment, the cells were incubated with 50 μl of 1 mg ml⁻¹ MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (Sigma-Aldrich Co.) in PBS for 3 h. The formazan that formed was then solubilized by adding 150 μl of dimethyl sulfoxide. Absorbance was read at 570 nm using a FLUOstar Galaxy plate reader (BMG Labtech, Offenburg, Germany).

**Apoptosis assay.** Apoptosis was measured using the Cell Death Detection ELISA PLUS kit (Roche Applied Science, Indianapolis, IN, USA) following the manufacturer’s protocol. In brief, cells were incubated with the lysis buffer at ambient temperature for 30 min, followed by centrifugation at 200 g for 10 min. The supernatant (20 μl) was incubated with 80 μl of anti-histone-biotin and anti-DNA-peroxidase reagent in a streptavidin-coated 96-well microtiter plate for 2 h at ambient temperature with gentle shaking. The amount of peroxidase retained in the immunocomplex was photometrically determined with 2.2’-azino-di-[3-ethylbenzthiazoline sulfonate (6)] diaminobenzidine salt as a substrate. Absorbance was measured at 405 nm using a FLUOstar Galaxy plate reader. Data were expressed as the relative percentage of DNA fragmentation.

**Effect of miR21 and APAF1 on paclitaxel sensitivity in vitro.** SKOV3 and OVCA432 cells were treated with 20 nM paclitaxel for 72 h after incubation with equal number of exosomes from miR21⁺/miR21⁻ MEF (Thermo Fisher Scientific) for 24 h. Transfected cells were then subjected to a 72-h treatment of paclitaxel from 0 to 20 or 5 nM for the subsequent MTT assay or apoptosis assay, respectively. SKOV3 and OVCA432 cells were transiently transfected with full-length Apaf1 in pcDNA3.1 + vector or empty pcDNA3.1 + vector for 24 h. Paclitaxel (0–20 nM) were then added to the transfected cells for 72 h. Cell viability was measured using MTT assay as described above.

**Microarray analysis.** SKOV3 cells were transfected with either 5 nM miR21 precursor or negative control precursor miRNA for 24 h, and total RNA was isolated using TRI reagent (Molecular Research Center). Purified RNA samples were amplified and labelled using the MessageAmp Premier RNA amplification kit (Thermo Fisher Scientific) and then hybridized onto Affymetrix GeneChip Human Genome U133 Plus 2.0 microarrays according to the manufacturer’s protocol. The arrays were washed and stained using the Affymetrix Fluidics Station 450 and GeneChip hybridization, washing and staining kit (Affymetrix Inc., Santa Clara, CA, USA). Scanning of the arrays was performed in the Cancer Genomics Core Laboratory at MD Anderson using the GeneChip Scanner 3000 7G (Affymetrix, Inc.). A heatmap was generated using dChip software (Affymetrix, Inc.). Pathway analyses of the data sets were performed using Ingenuity Pathway Software (Ingenuity Systems, Redwood City, CA, USA).

**Immunohistochemical analysis.** Immunolocalization of APAF1 was performed using the tumour sections obtained from ovarian cancer patients or female BALB/c athymic nude mice injected with ovarian cancer OVCA432 cells that had been stably transfected with the doxycycline-inducible APAF1 gene (OVCA432-APAF1-Luc) or with the control (OVCA432-GFP-Luc). Slides containing the sections were stained with commercially available anti-APAF1 (1:500; HP031373) antibodies (Sigma-Aldrich Co.). Target protein expression was visualized using a Biotz33-diaminobenzidine chromogen kit (Biocare Medical, Concord, CA, USA).

**Construction of APAF1 reporter vectors.** The DNA fragment containing the predicted miR21-binding site was amplified from SKOV3 genomic DNA using the following primers: 5’-ATTTAAGGTTAGGGCTCATTAGAGGCGGCTAGT-3’ (forward) and 5’-TTCATGAGTTGAGGTCCGTCGAATCTC-3’ (reverse). PCR products were subsequently cloned into the HindIII and SpeI sites of the pMIR-REPORT miRNA expression reporter vector (Thermo Fisher Scientific) and the integrity of constructs was confirmed by DNA sequencing. Site-directed mutagenesis was carried out to introduce mutations into the putative miR21-binding site using the QuikChange II site-directed kit (Stratagene, La Jolla, CA, USA). The mutation-containing primers were 5’-CTGGCAGCGAACTGCGAGAAAGAAGGAAGCATACGATCGGAGGTTG-3’ (sense) and 5’-CAAATGCTCAGTGATTTTCTTCTCTTGATCTTTTGACCAG-3’ (antisense).

**Luciferase reporter gene activity assay.** SKOV3 cells were seeded onto 24-well plates 1 day before transfection. The cells were transfected with 0.4 μg of the APAF1 reporter construct containing the predicted miR21-binding sites, together with 5, 20 and 100 nM miR21 precursor using Lipofectamine 2000 (Thermo Fisher Scientific). pRL-TK (1:20), which encodes Renilla luciferase, was included in all transfections to normalize transfection efficiency. The cells were washed and lysed with the passive lysis buffer from the Dual-Luciferase Reporter Assay System (Promega Corp.) 24 h after transfection. Luciferase activity was measured in each cell lysate using a FLUOstar Galaxy plate reader. Relative luciferase activity was first normalized with Renilla luciferase activity and then compared with those of the respective control.
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Author contributions

C.L.A., N.-N.C. and S.C.M. planned the experiments and prepared the manuscript. C.L.A. and N.-N.C. performed most the experiments. C.L.A., N.-N.C, T.T. and T.-L.Y. performed the RNA sequencing and microarray experiments. K.-K.K. performed the in vivo experiments. T.-L.Y. established the miR-21−/− mouse embryonic fibroblast line. C.S.L., E.S.L., K.K., R.S. and K.H.L. provided technical supports.

Additional information

Accession codes: Data files from the Ion Torrent RNA-seq sequencing analysis and the transcriptome profiling analysis were deposited in the Gene Expression Omnibus Data
Bank (GEO; National Cancer for Biotechnology Information, Bethesda, MD, USA), respectively, under the accession codes GSE77318 and GSE51457.

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