Synthesis, Spectroscopic, and Anticancerous Properties of Mixed Ligand Palladium(II) and Silver(I) Complexes with 4,6-Diamino-5-hydroxy-2-mercaptopyrimidine and 2,2′-Bipyridyl

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Synthesis of two new water-soluble mixed ligand [Pd(bpy)(dahmp)]Cl and [Ag(bpy)(Hdahmp)]NO3 complexes (dahmp and Hdahmp are the deprotonated monoanion and the protonated neutral 4,6-diamino-5-hydroxy-2-mercaptopyrimidine, resp.) is reported. The composition of the reported complexes was discussed on the bases of IR, 1H NMR, and mass spectra, as well as conductivity and thermal measurements. The reported complexes display a significant anticancer activity against Ehrlich ascites tumor cells (EACs). The higher activity of these complexes with their higher conductivity values corresponds to their complete ionization in aqueous solution.

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1. INTRODUCTION

Currently, cisplatin is being used as an anticancer agent in several human cancers, particularly, testicular and ovarian cancers [1, 2]. Side effects, especially nephrotoxicity, of this drug limit its widespread use in high dose [3]. The need to develop new complexes with reduced nephrotoxicity and higher activity has stimulated the synthesis of many new complexes. Over the past years, a renewed interest in Pd(II) complexes as potential anticancer agents has developed. Though a number of interesting Pd(II) targets have been investigated [4–7], the biological utility of such agents continues to be questioned, this may be due to the poor solubility of common Pd(II) complexes under physiologic conditions. Many studies, including nucleic and amino acid derivatives, showed that 2-mercaptopyrimidine and 2-mercapto-4-aminopyrimidine are able to inhibit the synthesis of t-RNA [8]. Thus, they may act as valuable substrates in the synthesis of antitumor chemotherapeutic agents [9]. Also, the effect of 2-mercaptopyrimidine-5-carboxylic acid and S-analogy of pyrimidinic bases on oral epidermoid human carcinoma (KB) have been reported [10, 11].

As a continuation of our research in 4,6-diamino-5-hydroxy-2-mercaptopyrimidine complexes and their biological activities [12], in this research, we report the complexes obtained from the reaction of 4,6-diamino-5-hydroxy-2-mercaptopyrimidine (Hdahmp) with [Pd(bpy)Cl2] and [Ag(bpy)(H2O)2]NO3. They have been investigated using IR, 1H NMR, and mass spectra, conductivity and thermal measurements. In addition, the anticancer activity of these complexes against Ehrlich ascites tumor cells (EACs) has been reported.

2. EXPERIMENTAL

2.1 Material and methods

All manipulations were performed under aerobic conditions using 4,6-diamino-5-hydroxy-2-mercaptopyrimidine and all other reagents (Merck) as received. [Pd(bpy)Cl2] was synthesized by the literature method [13].
The cells of *Ehrlich ascites* (EACs) tumor were obtained from National Cancer Institute (Cairo, Egypt). After harvesting and preparation of the cells, their total number and viability were determined by counting using Trypan blue [14].

### 2.2. Instrumentation

Microanalyses were determined by the Micro Analytical Unit (Cairo University, Cairo, Egypt). Electronic spectra were recorded using a Unicam UV–100 U.V.-vis. Spectrometer. IR spectra were measured as KBr discs on a Matson 5000 FT-IR spectrometer (Cairo University). \(^1\)H NMR spectra were measured on a Varian Gemini WM-200 spectrometer (Laser Centre, Cairo University). Thermal analysis measurements were carried out at room temperature on a YSI Model 32 conductivity bridge. Mass spectra were recorded on a Matson MS 5988 spectrometer (Micro Analytical Unit, Cairo University).

### 2.3. Synthesis of complexes

#### 2.3.1. \([\text{Pd(bpy)(dahmp)}]\)Cl·H₂O

To a stirred suspension of \([\text{Pd(bpy)Cl}_2]\) (0.17 g, 0.5 mmol) in methanol-benzene (3 : 2, V/V) (15 cm³), was added a methanolic solution of KOH (0.055 g, 1 mmol) containing Hdahmp (0.08 g, 0.5 mmol). The resulting suspension was stirred for two days and a brown complex was obtained. It was filtered off, washed with water and methanol, and then air-dried. Conductivity data (10⁻³ M in DMF): \(\Lambda_M = 97.0 \text{ohm}^{-1} \text{cm}^2 \text{mol}^{-1}\). Elemental Anal. Calc. for C₁₄H₁₄N₇O₄S·Pd: C, 35.53; H, 3.17; N, 17.76; S, 6.77; Cl, 7.51. Found C, 35.72; H, 3.11; N, 17.71; S, 6.85; Cl, 7.63.

#### 2.3.2. \([\text{Ag(bpy)(Hdahmp)}]\)NO₃

Silver nitrate (0.087 g, 0.5 mmol) in water (2 cm³) was added to bpy (0.078 g, 0.5 mmol) in methanol (35 cm³) to produce a colorless solution, to which Hdahmp (0.08 g, 0.5 mmol) was added. The reaction mixture was stirred in dark for 3 hours to produce a pale brown solid. It was filtered off, washed with little water, methanol, and diethyl ether, then dried in vacuo. Conductivity data (10⁻³ M in DMF): \(\Lambda_M = 60.0 \text{ohm}^{-1} \text{cm}^2 \text{mol}^{-1}\). Elemental Anal. Calc. for C₁₄H₁₄N₇O₄SAg: C, 34.72; H, 2.89; N, 20.25; S, 6.61. Found C, 34.54; H, 2.78; N, 20.00; S, 6.42.

### 2.4. Conductivity measurements

The conductivity values for \([\text{Pd(bpy)(dahmp)}]\)Cl and \([\text{Ag(bpy)(Hdahmp)}]\)NO₃ were determined. The compounds were dissolved in water and the measurements were done at concentrations; 1, 0.8, 0.6, 0.4, and 0.2 mM. The conductance values (\(\Lambda_M\)) were calculated and plotted against concentration [15].

### 2.5. Anticancer activity against *Ehrlich ascites* carcinoma in mice

All the compounds were screened for their anticancer activity by dissolving samples in minimum amount of DMSO (Hdahmp) or water (complexes) and diluting with phosphate-buffered saline (PBS; pH = 7.2). The anticancer studies using *Ehrlich ascites* tumor cells (EACs) were carried out by incubating 0.2 mL of cells IP. All the treatments started 24 hours after inoculation for 45 days. The tumor-bearing mice were divided into three groups. Group (1) is the standard one that received the 5-florouracil [16] (5-fu; 20 mg/kg/day of mice) for comparison. Group (2) received Hdahmp, \([\text{Pd(bpy)(dahmp)}]\)Cl, \([\text{Ag(bpy)(Hdahmp)}]\)NO₃ complexes (0.01 mg/mice/day). Group (3) is the control one received physiological saline (0.9% sodium chloride).

### 3. Results and Discussion

#### 3.1. Synthesis of complexes

Table 1 lists two new complexes of 4,6-diamino-5-hydroxy-2-mercaptopryrimidine (Hdahmp). The elemental analyses of the isolated complexes agree with the assigned formula. The conductivities (\(\Lambda_M\)) in DMF at room temperature showed the electrolytic character of these complexes [4, 16].

The complex \([\text{Pd(bpy)(dahmp)}]\)Cl was prepared from \([\text{Pd(bpy)Cl}_2]\) and Hdahmp in methanol-benzene in presence of aqueous base, while \([\text{Ag(bpy)(Hdahmp)}]\)⁺ was made from aqueous AgNO₃ with bpy and Hdahmp in methanol.

The complexes are powder-like, stable in the normal laboratory atmosphere, and soluble in water, DMF, or DMSO. We had hoped to characterize the structure of one of the complexes by single X-ray crystallography, but were thwarted on numerous occasions by very small crystal dimensions. Thus, the characterization of these complexes was based on the physical and spectroscopic techniques.

#### 3.2. Vibration spectra

The characteristic IR bands observed and the vibration assignments of 4,6-diamino-5-hydroxy-2-mercaptopryrimidine (Hdahmp) complexes are reported in Table 1. The spectrum of Hdahmp supports the existence of the thione form in the solid phase (Scheme 1). The spectrum of Hdahmp supports the existence of the thione form in the solid phase (Scheme 1). The spectrum of Hdahmp supports the existence of the thione form in the solid phase (Scheme 1).
Table 1: Spectral data of Hdahmp and its complexes.

| Complexes                        | υ(OH) | υ(NH₂) | δ(NH) | υ(C=O) | υ(NC) | υ(M–O) | υ(M–N) |
|----------------------------------|-------|--------|-------|--------|-------|--------|--------|
| [Pd(bpy)(dahmp)]Cl              | 3397  | 3360   | 3165  | 1640   | 1556  | 1450   | 1362   |
| [Ag(bpy)(Hdahmp)]NO₃          | 3308  | 3396   | 3171  | 1650   | 1543  | 1479   | —      |

*ν(Ag–S).

The spectrum of [Ag(bpy)(Hdahmp)]NO₃ shows new strong band near 1370 cm⁻¹ assigned to the ionic uncoordinated NO₃⁻ [16, 26, 27].

### 3.3. Electronic spectra

The electronic spectrum of [Pd(bpy)(dahmp)]⁺ complex shows bands at 475 and 326 nm due to 1A₁g → 1B₁g and 1A₁g → 1E₁g, transitions in a square-planar configuration [4, 23, 27]. Also, the spectrum of [Ag(bpy)(Hdahmp)]⁺ shows bands at 467, 382, and 277 nm; the latter two may arise from charge transfer of the type ligand (π) → b₁g (Ag⁺) and ligand (σ) → b₁g (Ag⁺), respectively, in a typically distorted square planar environment around the metal ion [4, 28, 29].

### 3.4. ¹H NMR spectra

The ¹H NMR spectrum of Hdahmp in d₆-DMSO shows two singlets at 6.07 and 6.18 ppm arising from N(4)H₂ and N(6)H₂ groups, respectively (Scheme 1). The proton of the hydroxyl group O(5)H appears as a broad singlet at δ9.13 ppm and the N(1)H proton gives a singlet at δ7.43 ppm. In the ¹H NMR spectrum of [Pd(bpy)(dahmp)]⁺⁷, the proton of the hydroxyl group is not observed while the resonance arising from N(6)H₂ is shifted to lower field [24, 30]. Also, the resonances arising from N(4)H₂ and N(1)H are slightly shifted to lower field. This is probably due to the decrease in the electron density caused by the withdrawing of electrons by the metal ions from the pyrimidine ring coordination centers [13, 23].
The $^1H$ NMR spectrum of $[\text{Ag(bpy)}(\text{Hdahm})]^+$ confirms the neutral bidentate behavior of Hdadmp through the thione sulfur and the cyclic N(3) center, as there is a slight shift from the free ligand spectrum (Table 2). Also, the bpy ligand, in the complexes, shows upfield shifts as compared with $[\text{Pd(bpy)}\text{Cl}_2]$ or $[\text{Ag(bpy)}(\text{H}_2\text{O})_2]^+$. This is interpreted in terms of strong binding of dahmp$^-$ to Pd(II) or Ag(I) in comparison to binding of chloride and aquo species, respectively [4–6].

### 3.5. Mass spectra

The mass spectrum of $[\text{Pd(bpy)}(\text{dahmp})]\text{Cl}_2\text{H}_2\text{O}$ shows a signal at $m/e$ 474 (Cacld. 472.9) with 2.60% abundance. The spectrum shows signals at 355, 215, and 129 corresponding the loss of (H$_2$O, C$_2$H$_3$N$_3$S), (Cl, Pd) [4], and (N$_2$, C$_3$H$_2$NO) fragments, respectively.

### 3.6. Thermal measurements

The thermal decomposition of $[\text{Pd(bpy)}(\text{dahmp})]\text{Cl}_2\text{H}_2\text{O}$ and $[\text{Ag(bpy)}(\text{Hdahmp})]\text{NO}_3$ was studied by using the thermogravimetry (TG) technique. The thermogram of $[\text{Pd(bpy)}(\text{dahmp})]\text{Cl}_2\text{H}_2\text{O}$ shows four TG inflections in the ranges 32–148, 150–360, 361–475, and 476–593°C. The first weight loss may arise from the elimination of crystal lattice water (Calcd. 3.81, Found 3.73%) [4]. The second step may arise from the release of half Cl$_2$ and C$_3$H$_5$N$_3$ fragments (Calcd. 25.06, Found 24.48%), the third step is due to the removal of CNS and half bpy (C$_6$H$_4$N) fragments (Calcd. 28.76, Found 29.22%) [27], while the fourth step is attributed to the removal of the other half of bpy species (Calcd. 16.49, Found 16.55%) followed by the formation of PdO at 665°C (Calcd. 25.88, Found 26.36%) [4, 27]. The thermogram of $[\text{Ag(bpy)}(\text{Hdahmp})]\text{NO}_3$ shows the first-step weight loss of 9.29% between 198 and 252°C, which corresponds to the release of NO$_3$ species (Calcd. 9.51%). The second decomposition step occurs between 253 and 352°C, this weight loss is attributed the loss of C$_3$H$_6$N$_3$ fragment (Calcd. 17.36, Found 17.89%). The third TG inflection between 432–520°C, may arise from the elimination of bpy, CS, three quarter species (Calcd. 49.18, Found 51.09 %), leaving Ag$_2$O representing (Found 23.00%) [5, 27].

### 3.7. Conductivity measurements

Figure 1 shows the plots of the conductivities of $[\text{Pd(bpy)}(\text{dahmp})]\text{Cl}$ and $[\text{Ag(bpy)}(\text{Hdahmp})]\text{NO}_3$ against concentrations. It is clear that as the concentration increases, the conductivity increases, indicating the complete ionization of the complexes species [31]. Since the conductivity for Cl$^-$ and NO$_3$ is 76 and 71 ohm cm$^{-2}$, respectively [31, 32], this suggests that the complexes ionized completely in aqueous media [33].

### 3.8. Anticancerous activity

The reliable criteria for judging the efficacy of any anticancer drug are prolongation of life span, improving the clinical, hematological, and biochemical profile, as well as reduction in viable tumor cell count in the host [34, 35]. We have reported that $[\text{PdL(pa)}]^+$, $[\text{PdL'(pa)Cl}]$, and $[\text{Pd(bpy)}(\text{cdhp})]$ ($L = 2,2'$-bipyridyl; $L' = 9,10$-phenanthroline, 2-(2'-pyridyl)quinoxaline); pa = anion of 2-pepiridine carboxylic acid; cdhp = dianion of 5-chloro-2,3-dihydroxypyridine) in water or DMSO exhibit potent cytotoxic activity against Ehrlich ascites tumor cells [4, 5]. It is known that the anticancer available drugs inhibit the hematological and biochemical parameters (hemoglobin (Hb), red blood cells count (RBCs), and white blood cells count (WBCs); blood picture). The ultimate goal of this project is to develop mixed ligand complexes containing nitrogen bases effective against...
that cause increase in the viscosity in comparison to the nor-

cancer without side effects on the hematological and bio-

c hematological status of EAC-bearing mice, a comparison study
was made among three groups of mice (each group con-
tains seven mice) from the second day after inoculation.
Group (1) tumor-bearing mice treated with 5-fu (stan-
dard [36, 37]). Group (2) tumor-bearing mice treated with
Hdahmp, [Pd(bpy)(dahmp)]Cl, [Ag(bpy)(Hdahmp)]NO3.
Group (3) is the control tumor-bearing mice. The antican-
tic activity of Hdahmp, [Pd(bpy)(dahmp)]Cl, and
[Ag(bpy)(Hdahmp)]NO3 shows remarkable efficacy man-
ifested by survival and activity, as well as reduction in
the tumor size. The hematological parameters including
hemoglobin (Hb), red blood cells count (RBCs), and white
blood cells count (WBCs) data are reported in Table 3.
It is clear that the hematological parameters of tumor-
bearing mice treated with Hdahmp, [Pd(bpy)(dahmp)]Cl, and
[Ag(bpy)(Hdahmp)]NO3 exhibit much better significant
figures with the use of small doses of (0.01 mg/mice/day)
compared with the standard (5-fu), the market drug
(~0.4 mg/mice/day).

| Compound | Hb(1) (12–16 g/dl) | RBCs(2) (4.0–6.0 × 10^6 cell/cm³) | HCT(3) (35.0–50.0%) | WBCs(4) (4000–11000 × 10^6 cell/cm³) | EAC Count (×10^6 cell/cm³) | MST/day(5) |
|----------|-------------------|---------------------------------|--------------------|-------------------------------------|-----------------------------|------------|
| Hdahmp   | 10.7              | 6.3                             | 40.3               | 8800                                | 44.8                        | 13.2       |
| [Pd(bpy)(dahmp)]Cl | 11.4            | 6.4                             | 40.3               | 12000                               | 31.8                        | 11.4       |
| [Ag(bpy)(Hdahmp)]NO3 | 11.0          | 6.3                             | 41                 | 9000                                | 30.4                        | 11.2       |
| 5-fu     | 10.2              | 6.0                             | 37.7               | 7600                                | 80                          | 13.6       |
| Control (0.9% NaCl) | 7.8             | 4.72                            | 22.2               | 2400                                | 220                         | 9.0        |

(1) Hb = hemoglobin, (2)RBCs = red blood cells count, (3)HCT = hematocrit value, (4)WBCs = white blood cells count values in normal mice are in paren-
theses, (5) the mean survival time.

In order to detect the influence of the solvent in the cy-
totoxicity of Hdahmp, [Pd(bpy)(dahmp)]+, and [Ag(bpy)–
(Hdahmp)]+. As expected, the water-soluble [Pd(bpy)–
(dahmp)]+ and [Ag(bpy)(Hdahmp)]+ are less kidney toxic.

3.9. Effect of survival time

The mean survival time (MST) of groups 1 and 2 was
compared with that of the control group using the fol-
lowing calculations [42]. Percentage (%) increase in lifes-

span over control = [(MST of treated group × 100/MST
of control group) −100]; MST = days of each mice
in the group/total number of mice. Percentage (%) in-
crease in lifespan over control showed to be high in
mice treated with Hdahmp, [Pd(bpy)(dahmp)]+, and
[Ag(bpy)(Hdahmp)]NO3 (Table 3).

3.10. The side effects and toxicity

The side effects and toxicity of Hdahmp, [Pd(bpy)–
(dahmp)]+, and [Ag(bpy)(Hdahmp)]+ have been detected.
After the first week of the treatment, the mice show flu-like...
attack and in the third week show spot dropping on the hair (alopecia). Fortunately, the solid organs have not been affected.

The study of detailed mechanism and in vivo anticancer screens (phase II & III) using the studied complexes are under way.

4. CONCLUSION

There are reports that complexes containing pyridine ring (cyclic nitrogen) display significant anticancer activity. The anticancer activity of the new water-soluble complexes, \([\text{Pd(bpy)(dahmp)}]\)\text{Cl} and \([\text{Ag(bpy)(Hdahmp)}]\)\text{NO}_3, shows remarkable efficacy against \text{Ehrlich ascites} tumor cells (EACs) manifested by survival and activity, as well as reduction in the tumor size.

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REFERENCES

[1] M. P. Hacker, E. B. Double, and I. H. Krakoff, \textit{Platinum Coordination Complexes in Cancer Chemotherapy}, Martinus Nijhoff, Dordrecht, The Netherlands, 1984.

[2] M. Coluccia, A. Nassi, F. Roseto, et al., “A trans-platinum complex showing higher antitumor activity than the cis con-gens,” \textit{Journal of Medicinal Chemistry}, vol. 36, no. 4, pp. 510–512, 1993.

[3] F. P. T. Harmers, W. H. Gispen, and J. P. Neijt, “Neurotoxic side-effects of cisplatin,” \textit{European Journal of Cancer}, vol. 27, pp. 372–376, 1991.

[4] S. I. Mostafa, “Mixed ligand complexes with 2-piperidine-carboxylic acid as primary ligand and ethylene diamine, 2,2′-bipyridyl, 1,10-phenanthroline and quinoxaline as secondary ligands: preparation, characterization and biological activity,” \textit{Transition Metal Chemistry}, vol. 32, no. 6, pp. 769–775, 2007.

[5] S. I. Mostafa, “Synthesis, characterization and antineoplastic activity of 3-chloro-2,3-dihydroxypyridine transition metal complexes,” \textit{Journal of Coordination Chemistry}, in press.

[6] V. X. Jin and J. D. Ranford, “Complexes of platinum(II) or palladium(II) with 1,10-phenanthroline and amino acids,” \textit{Inorganica Chimica Acta}, vol. 304, no. 1, pp. 38–44, 2000.

[7] G. Cavigiolio, L. Benedetto, E. Boccaleri, D. Colangelo, H. Viano, and D. Osella, “Pt(II) complexes with different N-donor aromatic ligands for specific inhibition of telomerase,” \textit{Inorganica Chimica Acta}, vol. 305, no. 1, pp. 61–68, 2000.

[8] S. K. Hadjikakou, M. A. Demertzis, M. Kubicki, and D. Kovala-Demertzis, “Organotin adducts with pyrimidinethione: crystal structure of dimethyld(pyrimidine-2-thiolato)tin(IV) and diphenylpyrimidinethione(2-thiolato)tin(IV),” \textit{Applied Organometallic Chemistry}, vol. 14, no. 11, pp. 727–734, 2000.

[9] V. N. Krishnamurthy, K. V. Naglowara Rao, P. L. Narasimha Rao, and B. Prapulla, “Some potential antiviral agents,” \textit{British Journal of Pharmacology and Chemotherapy}, vol. 31, no. 1, pp. 1–10, 1967.

[10] M. D. Couse, G. Faraglia, U. Russo, et al., “2-mercaptopyridine derivatives as neutral or ionic donors towards tin tetrahalides,” \textit{New Journal of Chemistry}, vol. 21, no. 10, pp. 1103–1111, 1997.

[11] S. D’Ancona, G. Magnolfi, G. Guidetti, et al., “Effects of 6,6′-dithiodinicotinic acid (CPDS) and its metabolite 6-mercaptopnicotinic acid (6-MNA) on murine and hamster fibroblasts (3T3 and BHK) and murine metastatic melanoma cells (F10),” \textit{Chemioterapia}, vol. 5, no. 4, pp. 219–227, 1986.

[12] S. I. Mostafa and N. Hadjiliadis, “New biologically active transition metal complexes of 2-mercapto-4,6-diamino-5-hydroxyprymidine,” \textit{Inorganic Chemistry: An Indian Journal}, vol. 2, no. 3, pp. 186–192, 2007.

[13] W. P. Griffith and S. I. Mostafa, “Complexes of esculetin with second and third row transition elements,” \textit{Polyhedron}, vol. 11, no. 8, pp. 871–877, 1992.

[14] K. R. Sheeki, G. Kuttan, and R. Kuttan, \textit{Amala Research Bulletin}, vol. 17, pp. 73–76, 1997.

[15] W. J. Geary, \textit{Coordination Chemistry Reviews}, vol. 7, no. 81, 1981.

[16] A. Abdulllah, F. Huq, A. Chowdhury, H. Tayyem, P. Beale, and K. Fisher, “Studies on the synthesis, characterization, binding with DNA and activities of two cis-planaramineplatinum(II) complexes of the form: cis-Pt(NH}_3Cl}_2, where L = 3-hydroxypyridine and 2,3-diaminopyrridine,” \textit{BMC Chemical Biology}, vol. 6, article 3, pp. 1472–1484, 2006.

[17] Z. Popovic, D. M. Calogovic, J. Hasic, and D. V. Topic, “Preparation and spectroscopic properties of the complexes of mercuric thiocyanate with pyridine-2-thione and pyridine-2-carboxylic acid. Crystal and molecular structure of two polymorphs of \text{Hg(SCN)}_2\text{Cl}_2\text{H}_2\text{NS}_2,” \textit{Inorganica Chimica Acta}, vol. 285, no. 2, pp. 208–216, 1999.

[18] M. D. Gutierrez, R. Lopez, M. A. Romero, and J. M. Salas, “Spectroscopic studies of some \text{Pd}(II), \text{Pt}(II), \text{Ag}(I), and \text{Au}(III) complexes of 4,6-diamino-2-thiopyrimidine and 4,6-diamino-2-methylthiopyrimidine. Structure and binding site determination,” \textit{Canadian Journal Chemistry}, vol. 66, no. 2, pp. 249–255, 1988.

[19] E. L. Torres and M. A. Mendiola, “Complexes of a triazine-3-thione ligand with divalent metals crystal structure of \text{[Cd}_2\text{DMF}_2\text{]}_2 \text{DMF}, 14H_2O,” \textit{Polyhedron}, vol. 24, no. 12, pp. 1435–1444, 2005.

[20] G. Glolub, H. Cohen, P. Paoletti, A. Bencini, and D. Meyerstein, “Copper-(I) and -(II) complexes with tertiary linear polyamines of the type \text{Me}_2\text{NCH}_2\text{(CH}_2\text{NMeCH}_2\text{)}_n\text{CH}_2\text{NMMe}_2(n = 1-4),” \textit{Journal of the Chemical Society, Dalton Transactions}, no. 10, pp. 2035–2060, 1996.

[21] S. I. Mostafa and S. A. Abd El-Maksoud, “Synthesis and character-ization of some transition metal complexes of 2-amino-3-hydroxyprymidine and its application in corrosion inhibition,” \textit{Monatshefte f"ur Chemie}, vol. 129, no. 5, pp. 455–466, 1998.

[22] M. Guta and M. N. Srivastava, “Synthesis and characterization of complexes of copper(II), nickel(II), cobalt(II) and zinc(II) with alanine and uracil or 2-thiouracil,” \textit{Synthesis and Reactivity in Inorganic, Metal-Orga-nic, and Nano-Metal Chemistry}, vol. 26, no. 2, pp. 305–320, 1996.

[23] S. I. Mostafa, M. A. Kabil, E. M. Saad, and A. A. El-Asmy, “Ligational and analytical applications of a uracil derivative toward some transition metal ions,” \textit{Journal of Coordination Chemistry}, vol. 59, no. 3, pp. 279–293, 2006.

[24] C.-I. Ma, Y. Shi, Q.-F. Zhang, and Q. Jiang, “Syntheses, characteriza-tion and crystal structures of diorganotin compounds with 2-mercaptopyrimidine and 4-amino-2-mercaptoppyrimidine,” \textit{Polyhedron}, vol. 24, no. 10, pp. 1109–1116, 2005.

[25] R. Castro, J. A. G. Vazquez, J. Romero, A. Sousa, R. Pritchard, and C. A. McAuliffe, “4,6-dimethylpyrimidine-2-thionato
(dmpymt−) complexes of nickel(II) and cadmium(II). Crystal structure of \([Cd(dmpymt)_{2}]\): a compound with a calixarene-like structure, *Journal of the Chemical Society, Dalton Transactions*, no. 7, pp. 1115–1120, 1994.

[26] M. A. Haj, M. Quiros, J. M. Salas, and R. Faure, “Silver complexes with triazolopyrimidine ligands containing an exocyclic oxygen atomml: X-ray evidence for an unusual tautomeric form,” *Journal of the Chemical Society, Dalton Transactions*, no. 11, pp. 1798–1801, 2001.

[27] S. I. Mostafa and M. M. Bekheit, “Synthesis and structure studies of complexes of some second row transition metals with 1-(phenylacetyl and phenoxyacet)-4-phenyl-3-thiosemicarbazide,” *Chemical and Pharmaceutical Bulletin*, vol. 48, no. 2, pp. 266–271, 2000.

[28] F. Sabin, C. K. Ryu, P. Fork, and A. Vogler, “Photophysical properties of hexanuclear copper(I) and silver(I) clusters,” *Inorganic Chemistry*, vol. 31, no. 10, pp. 1941–1946, 1992.

[29] M. A. Omary, T. R. Webb, Z. Assefa, G. E. Shankle, and H. H. Patterson, “Crystal structure, electronic structure, and temperature-dependent Raman spectra of Ti[Ag(CN)\(_2\)]: evidence for ligand-unsupported argentophilic interactions,” *Inorganic Chemistry*, vol. 37, no. 6, pp. 1380–1386, 1998.

[30] M. Calvo, A. M. Lanfredi, L. Oro, et al., “Synthesis and properties of rhodium(I) chloranilate and 2,5-dihydroxy-1,4-benzoquinonate complexes. Crystal structures of the binuclear \([\text{Rh}_2(\mu-\text{CA})(\text{cod})_2]\) and tetranuclear \([\text{Rh}_4(\mu-\text{CA})_2(\text{cod})_4]\) complexes (CA = chloranilate anion),” *Inorganic Chemistry*, vol. 32, no. 7, pp. 1147–1152, 1993.

[31] W. P. Griffith and S. I. Mostafa, “Complexes of 3-hydroxyxopyridin-2-one and 1,2-dimethyl-3-hydroxypyridin-4-one with second and third row elements of groups 6, 7 and 8,” *Polyhedron*, vol. 11, no. 23, pp. 2997–3005, 1992.

[32] D. Aguado, T. Montoya, J. Ferrer, and A. Seco, “Relating ions concentration variations to conductivity variations in a sequencing batch reactor operated for enhanced biological phosphorus removal,” *Environmental Modelling & Software*, vol. 21, no. 6, pp. 845–851, 2006.

[33] G. W. Castellan, *Physical Chemistry*, edited by L. Rogers, The Benjamin, Menlo Park, Calif, USA, 1983.

[34] B. D. Clarkson and J. H. Burchenal, “Preliminary screening of antineoplastic drugs,” *Progress in Clinical Cancer*, vol. 1, pp. 625–629, 1965.

[35] C. Oberling and M. Guerin, “The role of viruses in the production of cancer,” *Advances in Cancer Research*, vol. 2, pp. 353–423, 1954.

[36] B. Ardalan, M. D. Buscagila, and P. S. Schein, “Tumor 5-fluorodeoxyuridylate concentration as a determinant of 5-fluorouracil response,” *Biochemical Pharmacology*, vol. 27, no. 16, pp. 2009–2013, 1978.

[37] K. Yoshisue, Z. Hironaga, S. Yamaguchi, A. Yamamoto, S. Nagayama, and Y. Kawaguchi, “Reduction of 5-fluorouracil (5-FU) gastrointestinal (GI) toxicity resulting from the protection of thymidylate synthase (TS) in GI tissue by repeated simultaneous administration of potassium oxonate (Oxo) in rats,” *Cancer Chemotherapy and Pharmacology*, vol. 46, no. 1, pp. 51–56, 2000.

[38] A. Romerosa, P. Bergamini, V. Bertolasi, et al., “Biologically active platinum complexes containing 8-thiotheophylline and 8-(methylthio)theophylline,” *Inorganic Chemistry*, vol. 43, no. 3, pp. 905–913, 2004.

[39] D. Lebwohl and R. Canerra, “Clinical development of platinum complexes in cancer therapy: an historical perspective and an update,” *European Journal of Cancer*, vol. 34, no. 10, pp. 1522–1534, 1998.