INTRODUCTION

Biliary tract cancer (BTC) is the second most common cancer among hepatobiliary cancers, and it is a heterogeneous group of gastrointestinal tumors [1]. BTC includes gallbladder cancer (GBC), intrahepatic cholangiocarcinoma (iCCA), and extrahepatic cholangiocarcinoma (eCCA) [2]. GBC is the most common BTC and is aggressive. Tumor stage is the strongest prognostic factor for patients with GBC [3]. Cholangiocarcinomas (CCAs) are tumors originating from the epithelium of the bile duct and are further classified as iCCA and eCCA [4]. GBC, iCCA, and eCCA are distinct entities because of their different tumor biology and treatment guidelines [5]. The latest National Comprehensive Cancer Network (NCCN) guidelines for hepatobiliary cancers provide different staging and treatment strategies for GBC, iCCA, and eCCA. Overall, the prognosis of BTC is dismal because 60–70% of cases are diagnosed at the advanced stage of disease [6]. The median overall survival (OS) of patients with advanced BTC does not exceed 12 months [7]. This poor prognosis could be partially attributed to a lack of efficient prognostic markers. Therefore, the identification of new and effective biomarkers that are correlated with BTC prognosis is very important and urgent.

Prognostic role of programmed death-ligand 1 expression in patients with biliary tract cancer: a meta-analysis

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ABSTRACT

Previous studies investigated the prognostic role of programmed death-ligand 1 (PD-L1) expression in patients with biliary tract cancer (BTC); however, the results remained controversial. Therefore, we conducted the current meta-analysis with the aim of clarifying the association between PD-L1 expression and prognosis as well as with several important clinicopathological features of BTC. We searched PubMed, Embase, and Web of Science for relevant studies. Studies that detected PD-L1 expression in tumor cells by using immunohistochemistry (IHC) were selected. Pooled hazard ratios (HRs) and pooled odds ratios (ORs) with 95% confidence intervals (CIs) were calculated to estimate the correlations. In total, 15 independent studies with 1,776 patients were included in this meta-analysis. The pooled data demonstrated that high PD-L1 expression was associated with poor overall survival (n=15, HR=1.79, 95% CI=1.55–2.07, p<0.001). The correlation between PD-L1 expression and disease-free survival was not significant (n=6, HR=1.38, 95% CI=1.00–1.91, p=0.051). In addition, no significant correlation was observed between PD-L1 expression and clinical features in patients with BTC. Our study results showed that PD-L1 expression could play a pivotal role as an effective factor of poor prognosis in patients with BTC.
Further understanding of the tumor immune microenvironment has led to the development of immunotherapy, which has garnered much attention in recent years [8, 9]. Programmed death-ligand 1 (PD-L1) is the main ligand of PD-1, and the interaction between PD-L1 and PD-1 is a major inhibitory pathway of immunosuppression in the tumor microenvironment [10]. In addition, previous studies explored the prognostic significance of PD-L1 in patients with BTC, with conflicting results [11–25]. For example, the results of some studies showed that PD-L1 overexpression in tumor cells predicted poor survival of patients with BTC [13, 21], whereas the results of other studies showed that PD-L1 was not a significant prognostic factor for BTC [14]. However, some researchers reported that high expression of PD-L1 was associated with superior survival outcomes [25]. Therefore, we comprehensively retrieved the relevant studies and conducted this meta-analysis to evaluate the association between PD-L1 expression and the prognosis and clinicopathological factors of patients with BTC. As BTC is a group of heterogeneous diseases, we also conducted subgroup analyses according to the different tumor types.

RESULTS

Search results

The process of literature selection is presented in Figure 1. The initial literature search revealed 235 records, and after removing the duplicate papers, 146 articles were subjected to further screening. After examining the title and abstract, 113 reports were discarded. Subsequently, the full text of the remaining 33 studies were evaluated, and 18 studies were excluded owing to the following reasons: 12 studies did not provide usable data, 2 studies did not employ the IHC method, 2 studies did not provide any survival information, 1 study did not focus on PD-L1, and 1 study was overlapped. Finally, a total of 15 studies [11–25] comprising 1,776 patients were included for the meta-analysis.

Characteristics of included studies

The characteristics of the included studies are shown in Table 1. The included studies were published between 2014 and 2019. All the studies used IHC to detect PD-L1 expression in tumor cells. The patients were enrolled from 6 different countries: China, Japan, Germany, the USA, Thailand, and Korea. Regarding the tumor type, 7 studies included patients with eCCA [11, 13, 15, 16, 18, 20, 25], 4 studies focused on patients with iCCA [12, 19, 21, 24], 3 studies reported on patients with iCCA and eCCA [14, 22, 23], and 1 study included patients with GBC [17]. Regarding the association between PD-L1 expression and prognosis, all 15 studies provided data about the association between PD-L1 and OS [11–25], and 6 studies also provided data on the association between PD-L1 expression and DFS [16, 17, 19–21, 25]. The NOS scores of the included 15 studies ranged from 6 to 8, indicating high-quality studies.

Correlation of PD-L1 expression with OS

The data between PD-L1 expression and OS were extracted from all the 15 included studies with 1,776 patients [11–25]. The heterogeneity was not significant (I²=25.9%, p=0.167); therefore, a fixed-effects model was adopted. As shown in Table 2 and Figure 2A, the pooled data demonstrated that high PD-L1 expression was associated with poor OS (n=15, HR=1.79, 95% CI=1.55–2.07, p<0.001). In addition, subgroup analysis was performed for further investigation. On stratification by tumor type, ethnicity, and sample size, the data showed that PD-L1 remained a significant factor of poor OS in patients with eCCA (n=7, HR=1.73, 95% CI=1.08–2.75, p=0.022) and for patients with iCCA (n=4, HR=1.79, 95% CI=1.42–2.25, p<0.001), but not for patients with GBC (n=1, HR=1.92, 95% CI=0.95–3.88, p=0.069; Table 2 and Figure 2B). Moreover, PD-L1 expression was also a significant prognostic factor of poor OS irrespective of ethnicity (Table 2 and Figure 2C) and sample size (Table 2 and Figure 2D). Elevated PD-L1 expression was also a significant prognostic factor for OS in patients with BTC with different cut-off values of PD-L1 (Table 2 and Supplementary Figure 1A).

Correlation between PD-L1 expression and DFS

Six studies with 662 patients provided the HRs for DFS [16, 17, 19–21, 25]. Because of significant heterogeneity (I²=65%, p=0.016), a random-effects model was applied. The pooled HR indicated that the correlation between PD-L1 expression and DFS was not significant (n=6, HR=1.38, 95% CI=1.00–1.91, p=0.051; Table 2 and Figure 3A). Subgroup analysis revealed that PD-L1 overexpression was associated with worse DFS in patients with iCCA (n=2, HR=1.76, 95% CI=1.28–2.42, p<0.001) and in studies with sample size ≥100 (n=3, HR=1.64, 95% CI=1.27–2.13, p=0.001; Table 2 and Figure 3B). However, high PD-L1 expression was not predictive of poor DFS in Asian and Caucasian patients (Table 2 and Figure 3C), in patients with eCCA and GBC (Table 2 and Figure 3B), and in studies with sample size <100 (Table 2 and Figure 3D). The pooled data also indicated that PD-L1 overexpression remained a significant prognostic factor for DFS using various cut-off values of PD-L1 (Table 2 and Supplementary Figure 1B).
Association between PD-L1 expression and clinicopathological features

As GBC, iCCA, and eCCA are heterogeneous diseases and are considered different entities, we analyzed the correlation between PD-L1 expression and clinicopathological factors in the following 3 categories: iCCA, eCCA, and iCCA+eCCA. GBC was not analyzed because only 1 study was eligible. As shown in Table 3, the pooled ORs and 95% CIs showed that, for eCCA, there was no significant correlation between PD-L1 expression and sex (p=0.710), T stage (p=0.492), N stage (p=0.070), or tumor grade (p=0.126). In addition, for patients with iCCA, there was no significant association between PD-L1 expression and sex (p=0.651), tumor size (p=0.661), N stage (p=0.852), vascular invasion (p=0.116), or perineural invasion (p=0.529). For patients with eCCA and/or iCCA, there was no significant correlation between PD-L1 expression and sex (p=0.290), T stage (p=0.741), or N stage (p=0.174; Table 3).

Figure 1. Flow chart of literature search and study selection.
Table 1. Baseline characteristics of eligible studies for this meta-analysis.

| Study         | Year | Patients, n | Country | Tumor type | Period | Age (years) Median/mean (range) | Ethnicity | Specimen | Detection method | Treatment | Cut-off value | Endpoint | NOS score |
|---------------|------|-------------|---------|------------|--------|---------------------------------|-----------|----------|------------------|-----------|---------------|----------|-----------|
| Tamai         | 2014 | 91          | Japan   | eCCA       | 2000-2008 | NA     | Asian                           | Tissue    | IHC      | Surgical resection | Moderate or intense staining | >5% tumor cells | OS       | 6         |
| Gani          | 2016 | 54          | USA     | iCCA       | 1991-2011 | Mean: 64 | Caucasian                        | Tissue    | IHC      | Surgical resection | >5% tumor cells | >50% tumor cells | OS       | 8         |
| Ma            | 2017 | 70          | China   | eCCA       | 2009-2013 | Mean: 62.5 | Median: 57.5 Range: 45-76     | Asian     | Tissue    | Surgical resection | >1% tumor cells | OS       | 6         |
| Sangkhamanon  | 2017 | 46          | Thailand| iCCA, eCCA | NA      | NA     | Asian                           | Tissue    | IHC      | Surgical resection | Score 3 | OS       | 6         |
| Walter        | 2017 | 69          | Germany | eCCA       | 2007-2015 | NA     | Caucasian                        | Tissue    | IHC      | Surgical resection | Score 3 | OS       | 6         |
| Kim           | 2018 | 34          | USA     | eCCA       | 1990-2015 | Median: 67 | Range: 42-86                   | Caucasian | Tissue    | Surgical resection | >1% tumor cells | OS, DFS | 8         |
| Lin           | 2018 | 66          | China   | GBC        | 2009-2014 | Median: 65 | Median: 71 Range: 44-87     | Asian     | Tissue    | Surgical resection | Score 2 | OS       | 8         |
| Ueno          | 2018 | 117         | Japan   | eCCA       | 1995-2006 | Median: 71 | Median: 29-81 Range: 7-86     | Asian     | Tissue    | Surgical resection | Score 3 | OS       | 8         |
| Zhu           | 2018 | 192         | China   | iCCA       | NA       | NA     | Asian                           | Tissue    | IHC      | Surgical resection | >5% tumor cells | OS, DFS | 7         |
| Ahn           | 2019 | 183         | Korea   | eCCA       | 2003-2013 | Median: 68 | Range: 41-83                   | Asian     | Tissue    | Surgical resection | >1% tumor cells | OS, DFS | 8         |
| Dong          | 2019 | 125         | China   | iCCA       | 2012-2013 | Mean: 49  | Median: 29-81 Range: 44-87     | Asian     | Tissue    | Surgical resection | >5% tumor cells | OS, DFS | 8         |
| Kitano        | 2019 | 177         | Japan   | iCCA, eCCA | NA      | NA     | Asian                           | Tissue    | IHC      | Surgical resection | >25% tumor cells | OS       | 7         |
| Kriegsmann    | 2019 | 170         | Germany | iCCA, eCCA | NA      | NA     | Caucasian                       | Tissue    | IHC      | Surgical resection | >1% tumor cells | OS       | 8         |
| Lu            | 2019 | 320         | China   | iCCA       | 2005-2011 | Median: 58  | NA                          | Asian     | Tissue    | Surgical resection | >5% tumor cells | OS       | 8         |
| Yu            | 2019 | 62          | China   | eCCA       | 2015-2017 | Mean: 60.8 | Range: 22-81                   | Asian     | Tissue    | Surgical resection | Score 3 | OS, DFS | 8         |

NA, not available; iCCA, intrahepatic cholangiocarcinoma; eCCA, extrahepatic cholangiocarcinoma; GBC, gallbladder cancer; IHC, immunohistochemistry; OS, overall survival; DFS, disease-free survival; NOS, Newcastle-Ottawa scale.

Table 2. Subgroup analyses of OS and DFS based on different factors.

| Survival outcome | Subgroup          | Studies, n | Effects model | HR (95%CI)       | p value | Heterogeneity Meta-regression, p value |
|------------------|-------------------|------------|---------------|------------------|---------|--------------------------------------|
| OS               | Total             | 15         | Fixed         | 1.79 (1.55-2.07) | <0.001  | 25.9                                 | 0.167 | 0.820 |
|                  | Tumor type        |            |               |                  |         |                                      |       |       |
|                  | eCCA              | 7          | Random        | 1.73 (1.08-2.75) | 0.022   | 62.7                                 | 0.013 |       |
|                  | iCCA              | 4          | Fixed         | 1.79 (1.42-2.25) | <0.001  | 0                                    | 0.718 |       |
|                  | iCCA+eCCA         | 3          | Fixed         | 1.82 (1.38-2.40) | <0.001  | 0                                    | 0.508 |       |
|                  | GBC               | 1          | -             | 1.92 (0.95-3.88) | 0.069   | -                                    | -     |       |
|                  | Ethnicity         |            |               |                  |         |                                      |       | 0.783 |
|                  | Asian             | 11         | Fixed         | 1.77 (1.51-2.07) | <0.001  | 43.5                                 | 0.06  |       |
|                  | Caucasian         | 4          | Fixed         | 1.89 (1.34-2.67) | <0.001  | 0                                    | 0.787 |       |
|                  | Sample size       |            |               |                  |         |                                      |       | 0.960 |
|                  | <100              | 8          | Fixed         | 1.80 (1.38-2.35) | <0.001  | 42                                   | 0.099 |       |
|                  | ≥100              | 7          | Fixed         | 1.78 (1.50-2.12) | <0.001  | 12                                   | 0.338 |       |
|                  | Cut-off value     |            |               |                  |         |                                      |       | 0.166 |
|                  | Cut-off value 5%  | 5          | Fixed         | 1.80 (1.45-2.24) | <0.001  | 0                                    | 0.847 |       |
Cut-off value 1%  4  Fixed  1.49 (1.12-1.98)  0.001  0  0.917
Other cut-off values  6  Random  1.88 (1.14-3.10)  0.014  65.0  0.014
Total  6  Random  1.38 (1.00-1.91)  0.051  64  0.016

Tumor type  0.271
  eCCA  3  Random  0.97 (0.53-1.78)  0.930  76  0.015
  GBC  1  -  1.87 (0.95-3.68)  0.070  -  -
  iCCA  2  Fixed  1.76 (1.28-2.42)  <0.001  0  0.752

Ethnicity  0.365
  Asian  5  Random  1.42 (0.93-2.18)  0.108  63.6  0.027
  Caucasian  1  -  1.18 (0.95-1.47)  0.139  -  -

DFS  0.288
  Sample size
  <100  3  Random  0.93 (0.40-2.16)  0.869  78.8  0.009
  ≥100  3  Fixed  1.64 (1.27-2.13)  <0.001  0  0.724

Cut-off value  0.425
  Cut-off value 5%  3  Fixed  1.78 (1.33-2.37)  <0.001  0  0.939
  Cut-off value 1%  2  Fixed  1.22 (1.00-1.49)  0.045  0  0.458
  Other cut-off values  1  -  0.16 (0.04-0.65)  0.011  -  -

| Cut-off value 1% | Fixed | 1.49 (1.12-1.98) | 0.001 | 0 | 0.917 |
| Cut-off values   | Random| 1.88 (1.14-3.10) | 0.014 | 65.0 | 0.014 |
| Total            | Random| 1.38 (1.00-1.91) | 0.051 | 64 | 0.016 |
| Tumor type       |       |                  |       |    |        |
| eCCA             | Random| 0.97 (0.53-1.78) | 0.930 | 76 | 0.015 |
| GBC              |       | 1.87 (0.95-3.68) | 0.070 | - | - |
| iCCA             | Fixed | 1.76 (1.28-2.42) | <0.001| 0 | 0.752 |
| Ethnicity        |       |                  |       |    |        |
| Asian            | Random| 1.42 (0.93-2.18) | 0.108 | 63.6| 0.027 |
| Caucasian        |       | 1.18 (0.95-1.47) | 0.139 | - | - |
| DFS              |       |                  |       |    |        |
| Sample size      |       |                  |       |    |        |
| <100             | Random| 0.93 (0.40-2.16) | 0.869 | 78.8| 0.009 |
| ≥100             | Fixed | 1.64 (1.27-2.13) | <0.001| 0 | 0.724 |
| Cut-off value    |       |                  |       |    |        |
| Cut-off value 5% | Fixed | 1.78 (1.33-2.37) | <0.001| 0 | 0.939 |
| Cut-off value 1% | Fixed | 1.22 (1.00-1.49) | 0.045 | 0 | 0.458 |
| Other cut-off values | Fixed | 0.16 (0.04-0.65) | 0.011 | - | - |

iCCA, intrahepatic cholangiocarcinoma; eCCA, extrahepatic cholangiocarcinoma; GBC, gallbladder cancer; OS, overall survival; DFS, disease-free survival.

Sensitivity analysis and meta-regression analysis

Sensitivity analysis was performed to evaluate the stability of pooled HRs for OS and DFS. As shown in Figure 4, the results of the sensitivity analysis demonstrated high credibility of the pooled HRs. Meta-regression analysis showed that tumor type (p=0.820), ethnicity (p=0.783), sample size (p=0.960), and cut-off value (p=0.166) did not significantly contribute to heterogeneity of OS (Table 2). Similarly, meta-regression analysis also indicated that tumor type (p=0.271), ethnicity (p=0.365), sample size (p=0.288), and cut-off value (p=0.425) did not significantly contribute to heterogeneity of DFS (Table 2).

Publication bias

The Begg’s funnel plots and Egger’s test were used to estimate the potential publication bias. The results showed that there was no significant publication bias for OS on the Begg’s test (p=0.921, Figure 5A) and Egger’s test (p=0.581, Figure 5B). Similarly, the Begg’s test (p=0.452, Figure 5C) and Egger’s test (p=0.826, Figure 5D) indicated no significant publication bias for DFS.

DISCUSSION

BTCs are a diverse group of tumors and have poor prognosis owing to the advanced stage at the time of initial diagnosis [5]. BTC is associated with immune-related risk factors, and PD-L1 was an important mediating factor in the tumor immune microenvironment. In the present meta-analysis, survival data from 15 studies with 1,776 patients were integrated. The results demonstrated that PD-L1 overexpression was associated with poor OS but not poor DFS in patients with BTC. There was no significant correlation between PD-L1 expression and clinicopathological features in patients with either iCCA or eCCA. Sensitivity analysis, meta-regression analysis and publication bias tests suggested that the results were stable and credible. According to the 2019 NCCN guideline of hepatobiliary cancers, GBC, iCCA, and eCCA are considered 3 different clinical entities, with different TNM staging and prognosis. BTCs are a heterogenous group of tumors including GBC, iCCA, and eCCA. Therefore, to conform to clinical application, we included patients with BTC, and performed subgroup analyses on GBC, iCCA, and eCCA, separately. This meta-analysis provides important implications for the prognosis of BTC and for all professional practitioners who are referring to NCCN guidelines.

PD-L1 is upregulated by many inflammatory mediators and cytokines within the tumor microenvironment [26]. The binding of PD-1/PD-L1 can inhibit T-cell activation, induce activated T-cell apoptosis, and negatively mediate the immune response [27]. Many clinical trials have evaluated the use of PD-1/PD-L1 inhibitors for treating gastrointestinal malignancies [28]. For patients with advanced-stage BTC, treatment options are limited [1]. Current evidence showed that adjuvant chemotherapy was associated with an
Figure 2. Forest plots for the association between PD-L1 expression and overall survival (OS) categorized by different subgroups: (A) the entire patient group; (B) patients with eCCA, iCCA, iCCA+eCCA, or GBC; (C) patients with Asian ethnicity or Caucasian ethnicity; and (D) studies with sample size $\geq 100$ or sample size $<100$. Note: The right-side means “High PD-L1 predicts poor survival” and the left-side means “High PD-L1 predicts better survival”.

Figure 3. Forest plots for the association between PD-L1 expression and disease-free survival (DFS) categorized by different subgroups: (A) the entire patient group; (B) patients with eCCA, GBC, or iCCA; (C) patients with Asian ethnicity or Caucasian ethnicity; and (D) studies with sample size $\geq 100$ or sample size $<100$. Note: The right-side means “High PD-L1 predicts poor survival” and the left-side means “High PD-L1 predicts better survival”.
Table 3. Meta-analysis of PD-L1 expression and clinicopathological features in BTC patients.

| Tumor type  | Clinicopathological feature                  | Studies, n | Effects model | OR (95%CI)   | P value | Heterogeneity |
|-------------|---------------------------------------------|------------|--------------|--------------|---------|---------------|
| eCCA        | Sex (male vs female)                        | 6          | Fixed        | 1.09(0.69-1.72) | 0.710   | 0             | 0.826         |
|             | T stage (T3-T4 vs T1-T2)                    | 4          | Fixed        | 0.80(0.43-1.50) | 0.492   | 0             | 0.515         |
|             | N stage (N1 vs N0)                          | 6          | Fixed        | 1.48(0.97-2.27) | 0.070   | 42.3          | 0.123         |
|             | Grading (G3 vs G1+G2)                       | 3          | Fixed        | 1.85(0.84-4.07) | 0.126   | 22.2          | 0.277         |
| iCCA        | Sex (male vs female)                        | 4          | Fixed        | 1.11(0.71-1.73) | 0.651   | 0             | 0.931         |
|             | Tumor size (≥5cm vs <5cm)                   | 2          | Random       | 0.73(0.18-2.98) | 0.661   | 80.3          | 0.024         |
|             | N stage (N1 vs N0)                          | 4          | Random       | 1.22(0.15-9.91) | 0.852   | 92.8          | <0.001        |
|             | Vascular invasion (yes vs no)               | 4          | Random       | 3.24(0.75-14.00) | 0.116   | 82.4          | 0.001         |
|             | Perineural invasion (yes vs no)             | 3          | Fixed        | 0.77(0.36-1.72) | 0.529   | 0             | 0.706         |
| iCCA+eCCA   | Sex (male vs female)                        | 2          | Fixed        | 1.36(0.77-2.42) | 0.290   | 9.3           | 0.294         |
|             | T stage (T3-T4 vs T1-T2)                    | 2          | Fixed        | 1.10(0.63-1.91) | 0.741   | 41.9          | 0.190         |
|             | N stage (N1 vs N0)                          | 3          | Fixed        | 1.46(0.85-2.52) | 0.174   | 0             | 0.586         |

Improvement in OS of patients with BTC [29]. More recent studies showed that nivolumab had a manageable safety profile and signs of clinical activity in patients with unresectable or recurrent BTC [30]. Another recent study also showed that the adverse effects after nivolumab for metastatic BTC were controllable [31]. All these results imply that immune checkpoint inhibitors show promising clinical efficacy for patients with unresectable BTC.

The prognostic effect of PD-L1 expression has also been investigated in other types of cancers in a meta-analysis. A recent meta-analysis—based on data from 11 studies involving 1,697 cases—showed that PD-L1 overexpression could predict worse survival outcomes in patients with bladder cancer [32]. Another study [33] also showed that high expression of PD-L1 was associated with inferior OS in patients with colorectal cancer. In addition, elevated PD-L1 expression was positively correlated with lymph node metastasis [33]. A comprehensive meta-analysis of 50 studies with 11,383 patients demonstrated that PD-L1 expression on IHC was associated with poor OS and with several clinicopathological factors in patients with lung cancer [34]. In the present meta-analysis, the pooled data showed that high PD-L1 expression was predictive of poor OS, in line with the results of previous studies on other cancers. However, we did not identify any significant correlation between PD-L1 expression and any clinical factors in patients with BTC, which may be owing to the limited sample size while analyzing eCCA and iCCA.

![Figure 4. Sensitivity analysis for the association between PD-L1 expression levels with BTC.](image-url)
Notably, since included studies used different cut-off values of tumor stage and age to divide patients for the analysis of correlation with PD-L1 expression, we did not perform meta-analysis on those two factors (tumor stage and age). Therefore, we examined each included study separately. A total of 11 included studies [11, 13, 15, 18–25] reported the correlation of PD-L1 expression and age of patients, using different cut-off values: 4 studies used 60 years [13, 20, 21, 25], 3 studies applied 65 years [11, 15, 19], one study selected 58 years [24], 63 years [23], 71 years [18], and mean±Standard Deviation (SD) [22], respectively. Except for one study [22] showed that the age of patients with different expression levels of PD-L1 in cancer cells was significantly different (p=0.03), other 10 studies [11, 13, 15, 18–21, 23–25] reported non-significant association between PD-L1 expression and age of patients. Those results suggested the connection between PD-L1 expression and age may not be significant in patients with BTC. It is noteworthy that the correlation of PD-L1 and aging have been explored in relevant recent studies. A recent study from US Food and Drug Administration (FDA) showed that patient-reported outcomes in patients with advanced lung cancer receiving anti-PD-1/PD-L1 therapy were not significantly different between younger and older patients (cut-offed by 70 years) [35]. Moreover, a meta-analysis including 27 randomized controlled trials (RCTs) with 17,546 patients demonstrated that immune checkpoint inhibitors (ICIs) could not significantly improve OS and PFS compared with controls in cancer patients aged over 75 years [36].

Although we strictly performed this meta-analysis according to the PRISMA guidelines and selected eligible studies with uniform criteria, the study has several limitations. First, although all included studies used IHC to detect PD-L1 expression, the cut-off values defining low/high PD-L1 levels were different, which may result in heterogeneity. Second, in studies that enrolled patients with iCCA+eCCA [14, 22, 23], the researchers did not report PD-L1 expression for iCCA.
and eCCA separately. Therefore, the data were extracted from those studies by including the patients with iCCA and eCCA as a whole group. Third, only one study with patients with GBC [17] was included in the analysis. Although we searched the literature by using keywords containing gallbladder cancer, only one study of GBC was included finally. Therefore, the prognostic impact of PD-L1 in patients with GBC could not be sufficiently explored. Fourth, all included studies were retrospective. Although we did not limit the eligible studies to be retrospective or prospective; the included studies were retrospective after literature selection. More prospective studies on this issue are still needed in the future.

In summary, our meta-analysis demonstrated that PD-L1 overexpression was associated with worse OS but not DFS of patients with BTC. The prognostic value of PD-L1 expression was significant for OS of patients with iCCA and eCCA. However, no significant correlation was observed between PD-L1 expression and clinical features of patients with BTC. These results indicate that PD-L1 could play a pivotal role as an effective factor of poor prognosis in patients with BTC. Nevertheless, as the study had several limitations, further large-scale, well-designed studies are needed to confirm our results.

**MATERIALS AND METHODS**

**Search strategy**

This meta-analysis was performed on the basis of the Preferred Reporting Items for Systematic Review and Meta-Analysis (PRISMA) guidelines [37]. The databases of PubMed, Embase, and Web of Science were thoroughly searched until October 2019 by using the following retrieval keywords: (“PD-L1” OR “B7-H1” OR “CD274” OR “programmed cell death ligand 1”) AND (“cholangiocarcinoma” OR “biliary tract cancer” OR “gallbladder cancer” OR “bile duct cancer” OR “hilar cholangiocarcinoma” OR “distal cholangiocarcinoma” OR “intrahepatic cholangiocarcinoma” OR “extrahepatic cholangiocarcinoma”) AND (“survival” OR “prognostic” OR “prognosis” OR “outcome”). Moreover, the reference lists were manually screened to collect potentially relevant studies. Ethical approval was not needed for this meta-analysis because it does not include individual patient information.

**Inclusion and exclusion criteria**

The inclusion criteria for eligible studies were as follows: (1) BTC was diagnosed on histopathological examination; (2) immunohistochemistry (IHC) analysis of PD-L1 expression was conducted; (3) association between PD-L1 expression and OS and/or disease-free survival (DFS) was presented, or sufficient information was provided to compute the hazard ratio (HR) and 95% confidence interval (CI) [38]; (4) the PD-L1 expression in tumor cells was determined; and (5) the report was in the English language. The exclusion criteria for this meta-analysis were as follows: (1) duplicate studies; (2) reviews, case reports, letters, and meeting abstracts; (3) studies with insufficient data; and (4) non-English reports.

**Date extraction and quality assessment**

Data from candidate studies were evaluated and extracted by two independent investigators (C.L. and X.P.). Any disagreements were resolved through discussion. The following information was extracted from eligible studies: the first author’s name, publication year, number of cases, country, ethnicity, study period, patient age, tumor type, detection method, cut-off value for high expression of PD-L1, treatment method, HR and the corresponding 95% CI for OS and DFS, and clinicopathological characteristics. The quality of the included studies was assessed by using the Newcastle-Ottawa Scale (NOS) [39]. The NOS included the 3 following main categories: selection (0–4 points), comparability (0–2 points), and outcome assessment (0–3 points). The NOS scores ranged from 0 to 9. Studies with NOS scores ≥6 were indicated to be of high quality.

**Statistical analysis**

All statistical analyses were performed using Stata version 12.0 (Stata Corp LP, TX, USA). The HRs and 95% CIs were used to assess the prognostic role of PD-L1 expression on the survival of patients with BTC. An HR >1 with a p-value <0.05 indicated a poor prognosis in patients with PD-L1 overexpression. The odd ratios (ORs) with 95% CIs were calculated to determine the correlations between PD-L1 expression and clinicopathological parameters. Statistical heterogeneity among studies was determined using the Cochran Q-test and I-squared test [40, 41]. I² values > 50% and p-values <0.10 were considered to indicate significant heterogeneity, and the random effects model was applied. Otherwise, a fixed-effect model was implemented. Subgroup analysis—stratified by tumor type, ethnicity, and sample size—was performed. Sensitivity analysis was conducted to assess the reliability of the results. Meta-aggression analysis was conducted to identify the source of heterogeneity. Publication bias was measured by using both the Begg’s test and Egger’s test. P-values <0.05 were considered statistically significant.
Abbreviations

PD-L1: programmed cell death ligand 1; CCA: cholangiocarcinoma; ELISA: enzyme-linked immunosorbent assay; CAF: cancer-associated fibroblast; TAM: tumor-associated macrophage; TIL: tumor-infiltrating lymphocyte; CI: confidence interval; HR: hazard ratio; NOS: Newcastle-Ottawa Scale; OS: overall survival; PFS: progression-free survival; PRISMA: Preferred Reporting Items for Systematic Reviews and Meta-Analyses; FDA: Food and Drug Administration; RCT: randomized controlled trial; ICI: immune checkpoint inhibitor.

AUTHOR CONTRIBUTIONS

C.L., X.P., X.G. and Y.F. collected and analyzed the data, wrote the paper; N.L., L.L., S.W., and J.H. analyzed the data; G.Z. and Z.L. conceived and designed this study, analyzed the data, wrote the paper; and all authors reviewed the paper. All authors read and approved the final manuscript.

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CONFLICTS OF INTEREST

The authors declare no Conflicts of interests.

REFERENCES

1. Ghidini M, Pizzo C, Botticelli A, Hahne JC, Passalacqua R, Tomasello G, Petrelli F. Biliary tract cancer: current challenges and future prospects. Cancer Manag Res. 2018; 11:379–88. https://doi.org/10.2147/CMAR.S157156 PMID:30643463
2. Mondaca S, Nervi B, Pinto M, Abou-Alfa GK. Biliary tract cancer prognostic and predictive genomics. Linchuang Zhongliuxue Zazhi. 2019; 8:42. https://doi.org/10.21037/jco.2019.07.06 PMID:31431036
3. Duffy A, Capanu M, Abou-Alfa GK, Huitzil D, Jarnagin W, Fong Y, D’Angelica M, Dematteo RP, Blumgart LH, O’Reilly EM. Gallbladder cancer (GBC): 10-year experience at Memorial Sloan-Kettering Cancer Centre (MSKCC). J Surg Oncol. 2008; 98:485–89. https://doi.org/10.1002/jso.21141 PMID:18802958
4. Blechacz B. Cholangiocarcinoma: Current Knowledge and New Developments. Gut Liver. 2017; 11:13–26. https://doi.org/10.5009/gnl15568 PMID:27928095
5. Ahn DH, Bekaii-Saab T. Biliary cancer: intrahepatic cholangiocarcinoma vs. extrahepatic cholangiocarcinoma vs. gallbladder cancers: classification and therapeutic implications. J Gastrointest Oncol. 2017; 8:293–301. https://doi.org/10.21037/jgo.2016.10.01 PMID:28480068
6. Valle JW, Lamarca A, Goyal L, Barriuso J, Zhu AX. New Horizons for Precision Medicine in Biliary Tract Cancers. Cancer Discov. 2017; 7:943–62. https://doi.org/10.1158/2159-8290.CD-17-0245 PMID:28818953
7. Tariq NU, McNamara MG, Valle JW. Biliary tract cancers: current knowledge, clinical candidates and future challenges. Cancer Manag Res. 2019; 11:2623–42. https://doi.org/10.2147/CMAR.S157092 PMID:31015767
8. Marin-Acevedo JA, Soyano AE, Dholaria B, Knutson KL, Lou Y. Cancer immunotherapy beyond immune checkpoint inhibitors. J Hematol Oncol. 2018; 11:8. https://doi.org/10.1186/s13045-017-0552-6 PMID:29329556
9. Taube JM, Galon J, Sholl LM, Rodig SJ, Cottrell TR, Giraldo NA, Baras A5, Patel SS, Anders RA, Rimm DL, Cimino-Mathews A. Implications of the tumor immune microenvironment for staging and therapeutics. Mod Pathol. 2018; 31:214–34. https://doi.org/10.1038/modpathol.2017.156 PMID:29192647
10. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. Proc Natl Acad Sci USA. 2002; 99:12293–97. https://doi.org/10.1073/pnas.192461099 PMID:12218188
11. Tamai K, Nakamura M, Mizuma M, Mochizuki M, Yokoyama M, Endo H, Yamaguchi K, Nakagawa T, Shiina M, Unno M, Muramoto K, Satoh I, Satoh K, et al. Suppressive expression of CD274 increases tumorigenesis and cancer stem cell phenotypes in cholangiocarcinoma. Cancer Sci. 2014; 105:667–74. https://doi.org/10.1111/cas.12406 PMID:24673799
12. Gani F, Nagarajan N, Kim Y, Zhu Q, Luan L, Bhajjiej F, Anders RA, Pawlik TM. Program Death 1 Immune
Checkpoint and Tumor Microenvironment: Implications for Patients with Intrahepatic Cholangiocarcinoma. Ann Surg Oncol. 2016; 23:2610–17.  
https://doi.org/10.1245/s10434-016-5101-y  
PMD: 27012989

13. Ma K, Wei X, Dong D, Wu Y, Geng Q, Li E. PD-L1 and PD-1 expression correlate with prognosis in extrahepatic cholangiocarcinoma. Oncol Lett. 2017; 14:250–56. https://doi.org/10.3892/ol.2017.6105  
PMD: 28693161

14. Sangkhamanon S, Jongpairat P, Sookprasert A, Wirasorn K, Titapun A, Pukghem A, Ungareevittaya P, Chindaprasirt J. Programmed Death-Ligand 1 (PD-L1) Expression Associated with a High Neutrophil/Lymphocyte Ratio in Cholangiocarcinoma. Asian Pac J Cancer Prev. 2017; 18:1671–74.  
PMD: 28670887

15. Walter D, Herrmann E, Schnitzbauer AA, Zeuzem S, Hansmann ML, Peveling-Oberhag J, Hartmann S. PD-L1 expression in extrahepatic cholangiocarcinoma. Histopathology. 2017; 71:383–92. https://doi.org/10.1111/his.13238  
PMD: 28419539

16. Kim R, Coppola D, Wang E, Chang YD, Kim Y, Anaya D, Kim DW. Prognostic value of CD8CD45RO tumor infiltrating lymphocytes in patients with extrahepatic cholangiocarcinoma. Oncotarget. 2018; 9:23366 –72. https://doi.org/10.18632/oncotarget.25163  
PMD: 29805739

17. Lin J, Long J, Wan X, Chen J, Bai Y, Wang A, Yang X, Wu Y, Robson SC, Sang X, Zhao H. Classification of gallbladder cancer by assessment of CD8+ TIL and PD-L1 expression. BMC Cancer. 2018; 18:4651-8  
PMD: 3005582

18. Ueno T, Tsuchikawa T, Hatanaka KC, Hatanaka Y, Mitsuhashi T, Nakanishi Y, Noji T, Nakamura T, Okamura K, Matsuno Y, Hirano S. Prognostic impact of programmed cell death ligand 1 (PD-L1) expression and its association with epithelial-mesenchymal transition in extrahepatic cholangiocarcinoma. Oncotarget. 2018; 9:20034–47.  
https://doi.org/10.18632/oncotarget.25050  
PMD: 29732001

19. Zhu Y, Wang XY, Zhang Y, Xu D, Dong J, Zhang Z, Yi CH, Jia HL, Yang X. Programmed death ligand 1 expression in human intrahepatic cholangiocarcinoma and its association with prognosis and CD8+ T-cell immune responses. Cancer Manag Res. 2018; 10:4113–23.  
https://doi.org/10.2147/CMAR.S172719  
PMD: 30323667

20. Ahn S, Lee Y, Kim JW, Lee JC, Hwang JH, Yoon YS, Cho JY, Han HS, Choi Y, Kim H. Programmed cell death ligand-1 (PD-L1) expression in extrahepatic biliary tract cancers: a comparative study using 22C3, SP263 and E1L3N anti-PD-L1 antibodies. Histopathology. 2019; 75:526–36. https://doi.org/10.1111/his.13901  
PMD: 31081949

21. Dong Z, Liao B, Shen W, Sui C, Yang J. Expression of Programmed Death Ligand 1 Is Associated with the Prognosis of Intrahepatic Cholangiocarcinoma. Dig Dis Sci. 2019. [Epub ahead of print]. https://doi.org/10.1007/s10620-019-05787-0  
PMD: 31410753

22. Kitano Y, Yamashita YI, Nakao Y, Itoyoama R, Yusa T, Umezaki N, Tsukamoto M, Yamao T, Miyata T, Nakagawa S, Okabe H, Imai K, Chikamoto A, Ishiko T, Baba H. Clinical Significance of PD-L1 Expression in Both Cancer and Stroma Cells of Cholangiocarcinoma Patients. Ann Surg Oncol. 2019. [Epub ahead of print]. https://doi.org/10.1245/s10434-019-07701-4  
PMD: 31407173

23. Kriegsmann M, Roessler S, Kriegsmann K, Renner M, Longuespée R, Albrecht T, Loeffler M, Singer S, Mehrabi A, Vogel MN, Pathil A, Köhler B, Springfield C, et al. Programmed cell death ligand 1 (PD-L1, CD274) in cholangiocarcinoma - correlation with clinicopathological data and comparison of antibodies. BMC Cancer. 2019; 19:72.  
https://doi.org/10.1186/s12885-018-5254-0  
PMD: 30646854

24. Lu JC, Zeng HY, Sun QM, Meng QN, Huang XY, Zhang PF, Yang X, Peng R, Gao C, Wei CY, Shen YH, Cai JB, Dong RZ, et al. Distinct PD-L1/PD1 Profiles and Clinical Implications in Intrahepatic Cholangiocarcinoma Patients with Different Risk Factors. Theranostics. 2019; 9:4678–87. https://doi.org/10.7150/thno.36276  
PMD: 31367249

25. Yu F, Gong L, Mo Z, Wang W, Wu M, Yang J, Zhang Q, Li L, Yao J, Dong J. Programmed death ligand-1, tumor infiltrating lymphocytes and HLA expression in Chinese extrahepatic cholangiocarcinoma patients: possible immunotherapy implications. Biosci Trends. 2019; 13:58–69. https://doi.org/10.5582/bst.2019.01003  
PMD: 30773525

26. Okazaki T, Honjo T. The PD-1-PD-L pathway in immunological tolerance. Trends Immunol. 2006; 27:195–201. https://doi.org/10.1016/j.it.2006.02.001  
PMD: 16500147

27. Flemming A. Cancer: PD1 makes waves in anticancer immunotherapy. Nat Rev Drug Discov. 2012; 11:601. https://doi.org/10.1038/nrd3806  
PMD: 22850780

28. Stein A, Moehler M, Trojan J, Goekkurt E, Vogel A. Immuno-oncology in GI tumours: clinical evidence and
emerging trials of PD-1/PD-L1 antagonists. Crit Rev Oncol Hematol. 2018; 130:13–26. https://doi.org/10.1016/j.critrevonc.2018.07.001 PMID:30196908

29. Horgan AM, Amir E, Walter T, Knox JJ. Adjuvant therapy in the treatment of biliary tract cancer: a systematic review and meta-analysis. J Clin Oncol. 2012; 30:1934–40. https://doi.org/10.1200/JCO.2011.40.5381 PMID:22529261

30. Ueno M, Ikeda M, Morizane C, Kobayashi S, Ohno I, Kondo S, Okano N, Kimura K, Asada S, Namba Y, Okusaka T, Furuse J. Nivolumab alone or in combination with cisplatin plus gemcitabine in Japanese patients with unresectable or recurrent biliary tract cancer: a non-randomised, multicentre, open-label, phase 1 study. Lancet Gastroenterol Hepatol. 2019; 4:611–21. https://doi.org/10.1016/S2468-1253(19)30086-X PMID:31109808

31. Gou M, Zhang Y, Si H, Dai G. Efficacy and safety of nivolumab for metastatic biliary tract cancer. Oncotargets Ther. 2019; 12:861–67. https://doi.org/10.2147/OTT.S195537 PMID:30774373

32. Zhu L, Sun J, Wang L, Li Z, Wang L, Li Z. Prognostic and Clinicopathological Significance of PD-L1 in Patients With Bladder Cancer: A Meta-Analysis. Front Pharmacol. 2019; 10:962. https://doi.org/10.3389/fphar.2019.00962 PMID:31616289

33. Ni X, Sun X, Wang D, Chen Y, Zhang Y, Li W, Wang L, Suo J. The clinicopathological and prognostic value of programmed death-ligand 1 in colorectal cancer: a meta-analysis. Clin Transl Oncol. 2019; 21:674–86. https://doi.org/10.1007/s12094-018-1970-9 PMID:30392153

34. Li H, Xu Y, Wan B, Song Y, Zhan P, Hu Y, Zhang Q, Zhang F, Liu H, Li T, Sugimura H, Cappuzzo F, Lin D, Lv T, and written on behalf of AME Lung Cancer Collaborative Group. The clinicopathological and prognostic significance of PD-L1 expression assessed by immunohistochemistry in lung cancer: a meta-analysis of 50 studies with 11,383 patients. Transl Lung Cancer Res. 2019; 8:429–49. https://doi.org/10.21037/tlcr.2019.08.04 PMID:3155517

35. King-Kallimanis BL, Kanapur B, Blumenthal GM, Theoret MR, Kluetz PG. Age-related differences in patient-reported outcomes in patients with advanced lung cancer receiving anti-PD-1/PD-L1 therapy. Semin Oncol. 2018; 45:201–09. https://doi.org/10.1053/j.seminoncol.2018.06.003 PMID:30482633

36. Li J, Gu J. Efficacy of immune checkpoint inhibitors in cancer patients of different ages: a meta-analysis. Future Oncol. 2019; 15:3633-46. https://doi.org/10.2217/fon-2019-0279 PMID:31650854

37. Moher D, Liberati A, Tetzlaff J, Altman DG, Grp P, and PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Ann Intern Med. 2009; 151:264–69, W64. https://doi.org/10.7326/0003-4819-151-4-200908180-00135 PMID:19622511

38. Tierney JF, Stewart LA, Ghersi D, Burdett S, Sydes MR. Practical methods for incorporating summary time-to-event data into meta-analysis. Trials. 2007; 8:16. https://doi.org/10.1186/1745-6215-8-16 PMID:17555582

39. Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J Epidemiol. 2010; 25:603–05. https://doi.org/10.1007/s10654-010-9491-z PMID:20652370

40. Cochran W. The combination of estimates from different experiments. Biometrics. 1954; 10:101–29. https://doi.org/10.2307/3001666

41. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. Stat Med. 2002; 21:1539–58. https://doi.org/10.1002/sim.1186 PMID:12111919
Supplementary Figure 1. Forest plots for the association between PD-L1 expression and (A) OS and (B) DFS categorized by different cut-off values: cut-off value 1%, cut-off value 5%, and other cut-off values. Note: The right-side means “High PD-L1 predicts poor survival” and the left-side means “High PD-L1 predicts better survival”.