Differentiating T2 hyperintensity in neonatal white matter by two-compartment model of diffusional kurtosis imaging

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In conventional neonatal MRI, the T2 hyperintensity (T2h) in cerebral white matter (WM) at term-equivalent age due to immaturity or impairment is still difficult to identify. To clarify such issue, this study used the metrics derived from a two-compartment WM model of diffusional kurtosis imaging (WM-DKI), including intra-axonal, extra-axonal axial and radial diffusivities (De, Da,⊥ and De,⊥), to compare WM differences between the simple T2h and normal control for both preterm and full-term neonates, and between simple T2h and complex T2h with hypoxic-ischemic encephalopathy (HIE).

Results indicated that compared with control, the simple T2h showed significantly increased De,⊥ and Da, but no significant change in Da in multiple premyelination regions, indicative of expanding extra-axonal diffusion microenvironment; while myelinated regions showed no changes. However, compared with simple T2h, the complex T2h with HIE had decreased Da, increased De,⊥ in both premyelination and myelinated regions, indicative of both intra- and extra-axonal diffusion alterations. While diffusion tensor imaging (DTI) failed to distinguish simple T2h from complex T2h with HIE. In conclusion, superior to DTI-metrics, WM-DKI metrics showed more specificity for WM microstructural changes to distinguish simple T2h from complex T2h with HIE.

As a higher signal intensity in periventricular and subcortical white matter (WM) than in normal unmyelinated WM on T2 weighted image (T2WI), T2 hyperintensity (T2h) has a high incidence of up to 80% in preterm infants at term-equivalent age (TEA)1,2, and originally coined as diffuse and excessive high signal intensity (DEHSI). Besides of preterm neonates, it is also a common finding in full-term neonates. Such kind of WM abnormality, particularly combined with multiple parenchymal lesions may affect the following neurodevelopment outcomes2,6. One plausible hypothesis is that T2h represents a prematurity-related developmental phenomenon for its high incidence1,2 in preterm infants at TEA and disappearance after a postmenstrual age (PMA) of 50 weeks6. Conversely, other studies pointed out that it may also represent an early stage of WM injury that has been closely linked with abnormal WM microstructure at term7,8 and cognitive impairments up to 9 years of age1,9. From this, it is worthy to clarify the WM microstructure differences between the pure T2h and T2h combined with multiple parenchymal lesions2,6,10-12 (e.g. hypoxic-ischemic encephalopathy, HIE).

Recent advances in magnetic resonance imaging (MRI) have provided more important information about the development in the neonatal WM13. Through structural and diffusion tensor imaging (DTI), the myelination process presented three successive stages14,15: first, fibers organization in fascicles which lead to an increased fractional anisotropy (FA) on DTI but without obvious change on conventional MRI; second, "premyelination" stage by showing a shortening signal intensity on T1 weighted image (T1WI) and a decreased diffusivity,

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including the mean diffusivity (MD), axial diffusivity (AD) and radial diffusivity (RD), but without significant change in FA due to the oligodendrocytes and membranes proliferation; Third, “true” myelination stage by showing a shortening signal intensity on T2WI, decreased RD and increased FA due to the oligodendrocytes spiral ensheathment around the axon and compact packaged fibers. Since oligodendrocyte precursors (pre-OLs) are particularly vulnerable to a variety of chemical mediators including reactive oxygen species, glutamate, cytokines, and adenosine16–18, the myelination process often delays due to pre-OLs loss in the developing brain, especially in the premyelination regions19–22. With respect to axon, a recent histological study further reported that the axon degeneration was restricted to necrotic WM injury rather than non-necrotic WM injury 22. Thus, it can be concluded that the axon damage may be the critical factor for identifying the severe WM injury.

With respect to the T2h, diffusion weighted imaging (DWI) and DTI have revealed that such WM abnormality is associated with significant region-specific changes, such as decreased FA, and increased apparent diffusion coefficient (ADC), AD and RD7,23–25. However, it remains the difficulty for DTI in identifying whether these abnormal diffusivities are due to intra- or extra-axonal microstructural changes. As an alternative, diffusional kurtosis imaging (DKI) is a clinically feasible extension of DTI that examines the additional contribution of non-Gaussian diffusion effects as a result of brain microstructural complexity26. Recently, a two-compartment non-exchange diffusion model of the WM has been proposed that is suitable for DKI analysis and provides analytical expressions for the intra- and extra-axonal diffusion tensors27. Notably, the derived metrics, including intra-axonal diffusivity (D\textsubscript{a}) and extra-axonal axial and radial diffusivities (D\textsubscript{e/||} and D\textsubscript{e/\perp}), have been demonstrated to improve the understanding of WM alterations in various clinical diseases, such as Alzheimer’s disease28,29, schizophrenia30 and stroke31.

Taken together, a key aspect of differentiating the varying T2h may lie in the intra- and extra-axonal microstructure changes of WM. From this, we propose an hypothesis that the simple T2h (i.e. pure T2h) may be due to delayed myelination only, which meant only diffusivity changes in extra-axonal space; being different from simple T2h, the complex T2h (i.e. T2h combined with HIE) may be due to axonal damage which indicates the different microstructural changes (as exhibited in Fig. 1). Therefore, aiming at verifying such hypothesis by using the DKI-WM metrics, the case-control neonate study was conducted to clarify the intra- and extra-axonal microstructure changes between normal control and simple T2h; between simple T2h and complex T2h.

**Results**

**Subjects.** The study design flowchart is shown in Fig. 2. Of 423 neonates reviewed, 132 neonates with the PMA of 37–42 weeks had received conventional MRI and DKI. Further 13 neonates were excluded from the
analysis due to congenital metabolic disorder (n = 2), congenital infection (n = 3) or motion artifact (n = 8). Thus, 92 full-term neonates with gestational age (GA) of 39.18 ± 1.28 weeks and PMA at MRI scan of 40.55 ± 1.45 weeks and 27 preterm neonates with GA of 35.08 ± 2.07 weeks and PMA at MRI scan of 38.58 ± 1.73 weeks were enrolled for MRI interpretation. Following discussion and consultation, 21, 41 and 27 full-term neonates, and 8, 11 and 8 preterm neonates, were allocated respectively to the normal control group, simple T2h group and complex T2h with other abnormality group (detailed gestational ages and PMA at MRI scan in each group were shown on Table 1); 5 full-term neonates with GA of 39.20 ± 1.87 weeks and PMA at MRI scan 40.51 ± 2.01 weeks from complex T2h with other abnormality group were further allocated to the complex T2h with HIE group. The rest 22 full-term neonates and 8 preterm neonates in complex T2h with other abnormality group, and 3 full-term neonates presenting with punctate WM lesions without T2h were excluded from DKI data analysis.

In preterm and full-term neonates, the proportions of T2h were 70.4% (19 of 27) and 73.9% (68 of 92), and the proportions of simple T2h were 40.7% (11 of 27) and 44.6% (41 of 92), respectively. The proportions of T2h and simple T2h did not differ significantly between preterm and full-term neonates (P = 0.81, 0.83, respectively). The demographic parameters and clinical details of neonates in control group, simple T2h group and complex T2h with HIE group are shown in Table 1. There were no significant differences in the GA, PMA, birth weight (BW), age at the time of MR scan, presence of small size for GA, gender, or multiple births between the preterm simple T2h and control groups, full-term simple T2h and control groups, or full-term simple T2h and complex T2h with HIE groups. Compared with the full-term simple T2h group, the preterm simple T2h group had significantly lower mean GA, PMA and BW, and a significantly higher age at the time of MR scan.

Table 1. Participant demographics. Note: SGA, small size for gestational age. Note: *Data presented as means ± SDs; ‡data presented as medians and ranges; †data presented as frequencies and percentages.
Observer agreement in the interpretation of T2h in MRI. The first observer allocated 38, 42 and 36 neonates, respectively, to the control, simple T2h and complex T2h groups, while the second observer allocated 31, 53 and 32 neonates, respectively. On the second review by the first observer, 33, 48 and 35 participants were allocated to the control, simple T2h and complex T2h groups. Thus, the Kappa values for inter- and intra-observer agreements in the classification of MR findings were 0.74 and 0.82, respectively, corresponding to substantial and almost perfect agreement, respectively.

Tract-based spatial statistics (TBSS) analysis. Part I. Differences in WM-DKI metrics between the simple T2h and control groups. As shown in Fig. 3, the preterm simple T2h group showed significantly decreased FA, significantly increased MD, AD, RD, $D_a$, and $D_{a,\perp}$ in the superior corona radiata (SCR) and subcortical deep WM regions of superior frontal gyrus, superior occipital gyrus, precentral gyrus and postcentral gyrus, etc., and no change in $D_y$ compared with the control group. In comparisons between the full-term simple T2h and control groups, besides of above regions, similar results were observed in more widespread WM regions, including posterior thalamic radiation (PTR), corpus callosum (CC) and external capsule (EC), etc. However, for both preterm and full-term simple T2h groups compared with corresponding control groups, no differences were observed mainly in the myelinated WM regions at TEA, such as the cerebral peduncle and posterior limb of the internal capsule (PLIC). The metric changes in the simple T2h group reflect increased diffusivity from the extra-axonal space, which was compatible with delayed myelination (illustrated in Fig. 1). When these diffusional metrics were compared directly between the preterm and full-term simple

Figure 3. Tract-based spatial statistics for comparisons between the preterm simple T2h and control groups, and the full-term simple T2h and control groups. Regions colored light blue-blue represent significantly decreased voxels ($P < 0.05$) in the simple T2h group compared with the control group, while regions colored yellow-red represent significantly increased voxels ($P < 0.05$) in the simple T2h group compared with the control group. These have been overlaid on the mean FA template with a mean skeleton (green). Compared with the control group, the simple T2h group showed decreased FA and increased MD, AD, RD, $D_a$, and $D_{a,\perp}$ in multiple premyelination regions in both preterm and full-term neonates, but no differences were observed in myelinated regions (such as the cerebral peduncle and posterior limb of the internal capsule). Moreover, $D_y$ showed no significant change.
T2h groups (with GA, PMA and BW as covariates for correction), no differences were found (TBSS analysis not shown), indicating no essential differences between preterm and full-term neonates with the simple T2h.

**Part II. Differences in WM-DKI metrics between the simple T2h and complex T2h with HIE groups.** Compared with the full-term simple T2h group, the complex T2h with HIE group exhibited significantly decreased Da and FA in the CP, PLIC, PTR, SCR, CC and EC, with mainly increases in MD, RD, and De,⊥ (Fig. 4). The involvement of the CP and PLIC in these changes indicates the extensive axonal damage occurring in both myelinated and premyelination WM regions in the complex T2h with HIE group compared with the full-term simple T2h group, and MD, RD and De,⊥ were increased to differing extents.

**ROI analysis.** The PLIC, SCR, CC and EC, which respectively represented projection fibers, commissural fibers and association fibers respectively, were selected as target ROI regions. The results of the ROI analysis (Tables 2–5) were almost entirely consistent with those of the TBSS analysis. Da was identified as distinguishing between the simple T2h and complex T2h with HIE groups in regions that represented myelinated (the PLIC) and premyelination fibers (the SCR). In addition, all parameters for the PLIC showed no significant differences between the simple T2h and control groups in both preterm and full-term neonates.

**Discussion**

In this study, an advanced technique employing a two-compartment WM-DKI model was used to explore the diffusion variation underlying T2h in the neonatal WM. The results provide the novel insights into the possible abnormal changes underlying simple T2h, which were similar between preterm and full-term neonates. Furthermore, above findings of WM-DKI metrics suggest that simple T2h and complex T2h with HIE may originate from differing WM microstructural changes.

With respect to simple T2h, previous DTI studies22–25 have also observed lower FA and higher MD, AD and RD in neonates. However, due to the non-specificity of DTI measures for the intra- and extra-axonal diffusion, it is still difficult to determine what structure alters on earth. Our results regarding the simple T2h group observed the changes of DKI-WM metrics mainly in multiple premyelination regions hinting that the elevated diffusivity was from the extra-axonal space rather than the intra-axonal space; besides, such abnormality was absent in highly anisotropic regions (such as the PLIC and CP etc) where show almost complete myelination at the TEA32. A more recent DTI study25 about T2h (DEHSI) also observed similar patterns of spatial distribution. Specifically, higher diffusivity values and lower FA were found in centrum semiovale and OR, while CP and CST showed no difference between those with or without DEHSI at TEA. It is well known that the normal premyelination process is closely linked to a decrease in brain water content and the proliferation of oligodendrocyte lineage precursors, which showed an overall decrease in diffusivity and increase in FA with age34. Hence, the extra-axonal diffusional changes in the opposite way (increase in extra-axonal diffusivity and decrease in FA) observed in simple T2h were just compatible with the delayed myelination, which might result from a sparsity of oligodendrocytes and
Preterm neonates

|                        | FA (×10⁻³ mm/s²) | MD (×10⁻³ mm/s²) | AD (×10⁻³ mm/s²) | RD (×10⁻³ mm/s²) | D_A (×10⁻³ mm/s²) | D_AD (×10⁻³ mm/s²) |
|------------------------|------------------|------------------|------------------|------------------|------------------|-------------------|
| Control group          | 0.38 ± 0.02      | 1.16 ± 0.07      | 1.58 ± 0.08      | 0.90 ± 0.06      | 1.06 ± 0.17      | 1.93 ± 0.12       |
| Simple T2h group       | 0.37 ± 0.03      | 1.18 ± 0.07      | 1.70 ± 0.07      | 0.92 ± 0.08      | 1.21 ± 0.19      | 1.95 ± 0.10       |
| Full-term neonates     |                  |                  |                  |                  |                  |                   |
| Control group          | 0.39 ± 0.02      | 1.14 ± 0.05      | 1.67 ± 0.05      | 0.87 ± 0.05      | 1.04 ± 0.09      | 1.93 ± 0.07       |
| Simple T2h group       | 0.38 ± 0.03      | 1.16 ± 0.05      | 1.68 ± 0.05      | 0.90 ± 0.06      | 1.07 ± 0.17      | 1.93 ± 0.08       |
| Complex T2h with HIE   | 0.33 ± 0.04      | 1.21 ± 0.06      | 1.68 ± 0.08      | 0.97 ± 0.07      | 0.87 ± 0.09      | 1.94 ± 0.08       |
| P value                | 0.03             | 0.04             | 0.05             | 0.03             | 0.04             |                   |

Table 2. Comparisons of DKI-WM metrics of the posterior limb of the internal capsule among study groups. Note: data presented as means ± SDs. AD, axial diffusivity; D_A, intra-axonal diffusivity; D_AD, extra-axonal axial diffusivity; D_A/R, extra-axonal radial diffusivity; FA, fractional anisotropy; MD, mean diffusivity; RD, radial diffusivity.

Table 3. Comparisons of DKI-WM metrics of the superior corona radiata among study groups. Note: data presented as means ± SDs. AD, axial diffusivity; D_A, intra-axonal diffusivity; D_A/R, extra-axonal axial diffusivity; D_A/R, extra-axonal radial diffusivity; FA, fractional anisotropy; MD, mean diffusivity; RD, radial diffusivity.

a relatively elevated free water content in the extracellular matrix (as illustrated Fig. 1). This speculation was also supported by an animal experiment23, in which they found non-necrotic WM injury mainly led to myelination failure but without axonal degeneration. Additionally, we found no obvious differences in the microstructural changes of preterm and full-term neonates with simple T2h, suggesting that these two populations possess similar microstructural alterations.

Targeting the complex T2h with HIE, lower FA and slightly higher MD and RD were found than those with simple T2h. Although these, it was not sufficiently specific to differentiate these two groups since similar diffusivity changes were observed between simple T2h and controls. Notably, being different from simple T2h, D_A was markedly decreased in both myelinated and premyelination WM regions in complex T2h with HIE. These observations may hint an extensive breakdown of axons due to these severe WM damage, which was also demonstrated by previous histological studies22,33. All these may suggest D_A to be a specific biomarker for distinguishing these two abnormalities. A similar change in D_A has been reported in a study of acute and subacute ischemic lesions31, supporting D_A as a sensitive and specific biomarker of axonal abnormalities. Furthermore, these findings suggest that the microstructural changes can be more severe in complex T2h, partly explaining the varied neurodevelopmental outcomes observed previously34.
### Table 4. Comparisons of DKI-WM metrics of the corpus callosum among study groups.

|                      | FA       | MD   | AD   | RD   | $D_a$ | $D_o$ | $D_{a,o}$ |
|----------------------|----------|------|------|------|-------|-------|-----------|
| **Preterm neonates** |          |      |      |      |       |       |           |
| Control group        | 0.26 ± 0.02 | 1.71 ± 0.06 | 2.18 ± 0.06 | 1.47 ± 0.06 | 1.33 ± 0.15 | 2.51 ± 0.11 | 1.74 ± 0.08 |
| Simple T2h group     | 0.25 ± 0.01 | 1.75 ± 0.06 | 2.23 ± 0.07 | 1.51 ± 0.07 | 1.48 ± 0.24 | 2.56 ± 0.13 | 1.75 ± 0.08 |
| **Full-term neonates** |          |      |      |      |       |       |           |
| Control group        | 0.28 ± 0.02 | 1.62 ± 0.06 | 2.12 ± 0.07 | 1.38 ± 0.06 | 1.30 ± 0.11 | 2.43 ± 0.09 | 1.64 ± 0.07 |
| Simple T2h group     | 0.26 ± 0.02 | 1.68 ± 0.07 | 2.15 ± 0.07 | 1.44 ± 0.07 | 1.36 ± 0.17 | 2.43 ± 0.11 | 1.68 ± 0.08 |
| Complex T2h with HIE group | 0.23 ± 0.03 | 1.75 ± 0.08 | 2.18 ± 0.08 | 1.53 ± 0.09 | 1.17 ± 0.15 | 2.46 ± 0.08 | 1.78 ± 0.09 |

P value

| Preterm simple T2h vs. control group | 0.253 | 0.398 | 0.153 | 0.618 | 0.310 | 0.122 | 0.640 |
| Full-term simple T2h vs. control group | 0.001 | 0.007 | 0.149 | 0.002 | 0.150 | 0.780 | 0.041 |
| Preterm simple T2h vs. full-term simple T2h group | 0.402 | 0.125 | 0.021 | 0.304 | 0.099 | 0.001 | 0.062 |
| Full-term simple T2h vs. complex T2h with HIE group | 0.005 | 0.028 | 0.388 | 0.010 | 0.024 | 0.643 | 0.016 |

### Table 5. Comparisons of DKI-WM metrics of the external capsule among study groups.

|                      | FA       | MD   | AD   | RD   | $D_a$ | $D_o$ | $D_{a,o}$ |
|----------------------|----------|------|------|------|-------|-------|-----------|
| **Preterm neonates** |          |      |      |      |       |       |           |
| Control group        | 0.21 ± 0.01 | 1.29 ± 0.08 | 1.57 ± 0.07 | 1.15 ± 0.08 | 1.21 ± 0.20 | 1.73 ± 0.08 | 1.27 ± 0.09 |
| Simple T2h group     | 0.20 ± 0.02 | 1.38 ± 0.08 | 1.67 ± 0.08 | 1.24 ± 0.09 | 1.39 ± 0.25 | 1.83 ± 0.10 | 1.35 ± 0.10 |
| **Full-term neonates** |          |      |      |      |       |       |           |
| Control group        | 0.23 ± 0.01 | 1.24 ± 0.05 | 1.54 ± 0.05 | 1.09 ± 0.05 | 1.18 ± 0.12 | 1.70 ± 0.06 | 1.22 ± 0.05 |
| Simple T2h group     | 0.21 ± 0.02 | 1.29 ± 0.07 | 1.57 ± 0.06 | 1.15 ± 0.08 | 1.23 ± 0.22 | 1.73 ± 0.07 | 1.27 ± 0.09 |
| Complex T2h with HIE group | 0.19 ± 0.02 | 1.36 ± 0.05 | 1.62 ± 0.04 | 1.23 ± 0.05 | 1.08 ± 0.09 | 1.79 ± 0.04 | 1.37 ± 0.06 |

P value

| Preterm simple T2h vs. control group | 0.343 | 0.041 | 0.028 | 0.053 | 0.293 | 0.047 | 0.074 |
| Full-term simple T2h vs. control group | 0.007 | 0.036 | 0.137 | 0.021 | 0.495 | 0.295 | 0.061 |
| Preterm simple T2h vs. full-term simple T2h group | 0.714 | 0.296 | 0.112 | 0.430 | 0.122 | 0.133 | 0.741 |
| Full-term simple T2h vs. complex T2h with HIE group | 0.004 | 0.007 | 0.022 | 0.005 | 0.104 | 0.030 | 0.005 |

this population may reflect selection bias rather than the real situation. Third, the absence of follow-up for our enrolled neonates necessitates further validation of our hypothesis.

In conclusion, through using two-compartment WM-DKI metrics, this study provided series of interesting findings about the underlying microstructural changes of T2h: (1) for both preterm and full-term neonates with simple T2h, increased extra-axonal diffusivity and unchanged intra-axonal diffusivity in multiple premyelination WM regions may hint the delayed myelination; (2) while, T2h with HIE showed markedly decreased intra-axonal diffusivity which may be related to axonal damage; (3) superior than conventional DTI metrics, WM-DKI metrics are more specific for identifying the WM microstructural changes (e.g. intra- and extra-axonal diffusivity) in developmental brain.

**Materials and Methods**

This single-center retrospective study was approved by the Ethics Committee of the First Affiliated Hospital of Xi’an Jiaotong University, and the written parental consent was obtained prior to scanning. This study was conducted in accordance with the Declaration of Helsinki.

**Subjects.** All neonates were consecutively collected from our institution’s neonatal intensive care unit, from December 2010 to February 2014. The neonates that had received conventional MR imaging and DKI at the PMA of 37–42 weeks were included. Any neonates who diagnosed as congenital metabolic disorder, malformation or infection, or had poor MR image quality were excluded from this study. Demographic parameters and clinical details of all the neonates were reviewed and recorded by an experienced neonatal radiologist (Q.L.S. with 10 years of related experience).
**MR data acquisition.** All MR images were obtained using a 3.0 Tesla scanner (Signa HDxt, General Electric Medical System, Milwaukee, WI, USA) equipped with an 8-channel phased array radio-frequency head coil. Chloral hydrate (50 mg/kg) was administered orally for sedation in 30 minutes before the scan. The neonate was kept warm with swaddle and the head immobilized by molded foam. Micro-earplugs and earmuffs were used to protect the hearing. During imaging, the neonate was monitored by the attending pediatrician, and vital signs were monitored continuously.

Three-dimensional fast spoiled gradient-recalled echo T1WIs (repetition time/echo time, 4200/116.4–118.9 ms) and fast spin-echo T2WIs (repetition time/echo time, 2000/500–1000 ms) were obtained. DKI was acquired using the following parameters: 18 directions; b value = 0, 500, 1000 and 2000 s/mm²; repetition time/echo time = 8000/100.2–117.7 ms; field of view, 180 × 180 mm²; matrix, 256 × 256; and acquisition time of 8 minutes 42 seconds. The total scan time was less than 30 minutes.

**MR interpretation.** MR abnormalities (i.e. T2h and other abnormalities) were screened according to the clinical MR standards. Wherein, T2h was defined as visually higher signal intensity in periventricular and/or subcortical WM than in normal unmyelinated WM on T2WIs; while the “anterior caps” and “posterior arrowheads” would be excluded due to its common high signal intensity. The other abnormalities included cystic encephalomalacia, punctate lesions, a loss of volume of the periventricular WM and corpus callosum, the change of gray matter signal intensity, the widening of the subarachnoid space and intraventricular hemorrhage.

Neonates without any MR abnormalities and no evidences of any clinical episodes that might cause cerebral damage or delayed maturation were allocated to the control group. Neonates with T2h but no above abnormalities were allocated to the simple T2h group. Meanwhile, neonates with both T2h and above abnormalities were allocated to the complex T2h group. Furthermore, in the complex T2h group, the full-term neonates fulfilled both MR35 and clinical diagnostic criteria for HIE36 would be allocated to the complex T2h with HIE group. Specifically, the MR diagnostic criteria refers to the focal or diffuse abnormal signal intensities in the bilateral globus pallidus, putamen or thalamus on conventional MRI35. And the clinical diagnostic criteria for HIE36 includes: A pH ≤ 7.0 or a base deficit ≥ 16 mmol/L on umbilical cord blood or any postnatal blood sample within 1 hour of age; or history of an acute perinatal event and either no blood gas available, or a pH from 7.01 to 7.15 or a base deficit from 10 to 15.9 mmol/L, with a 10-minute Apgar score ≤ 5, or assisted ventilation initiated at birth and continued for at least 10 minutes. The rest neonates in the complex T2h group, and neonates without T2h but with any other abnormalities were excluded.

Two experienced pediatric radiologists (B.L.Y and J.G., respectively with 35 and 7 years of related experience), blinded to the neonatal/perinatal history, evaluated all MR images independently. They reviewed the published written and visual descriptions of the T2h appearances and agreed on its appearances prior to image analysis. The MR images were anonymized and reviewed on the same workstation, with the same window width and level. One month after the initial review, the MR images were reviewed for a second time by one of the observers (J.G.). Disagreements regarding image findings were resolved by discussion and mutual agreement. In the end, the intra- and inter-observer agreements were evaluated.

**MR data processing.** Rigid registration and distortion correction were performed after brain extraction37,38. Artifact-corrupted DWIs were excluded by using an automated method39. Diffusion tensor and kurtosis tensor were estimated by using constrained weighted linear least squares (CWLLS)40,41, according to the following equation42:

\[
S(b) = \exp \left( -bD + \frac{1}{6}b^2D^2K + O(b^3) \right).
\]

where \(S(b)\) was the diffusion weighted signal at a particular b value, and \(S(0)\) the signal without applying any diffusion gradient. \(D, K\) were the apparent diffusion coefficient and diffusional kurtosis.

Parametric maps of FA, MD, AD, and RD were derived from the diffusion tensor43.

The kurtosis tensor (KT) was transformed by the three eigenvectors of the diffusion tensor (DT)43:

\[
\hat{W}_{iii} = \sum_{i'=1}^{3} \sum_{j'=1}^{3} \sum_{k'=1}^{3} c_{i'j'k'} \hat{c}_{ijk} W_{ijk} / \lambda_i^{3}.
\]

The kurtosis along an individual DT eigenvector can be computed from the transformed KT43:

\[
K_i = \frac{MD^2}{\lambda_i^3} \cdot \hat{W}_{iii}.
\]

where \(\lambda_i\) was the eigenvalues of the DT. RK can be derived by using the eigenvectors on the radial directions43.

In the WM model for DKI, let \(D_a\) and \(D_e\) represent diffusion tensors in the intra-axonal and extra-axonal spaces. The axonal water fraction (AWF) was denoted by the symbol \(f\). The DW signal was described as a function of the b value by the equation27:

\[
\frac{S(b)}{S(0)} = f \exp(-bD_a n) + (1 - f) \exp(-bD_e n).
\]
The AWF was estimated by the equation:

\[
f = \frac{K_{\text{max}}}{K_{\text{max}} + 3}.
\]  

(5)

The diffusion coefficients in the intra-axonal and extra-axonal spaces were calculated by the equations:

\[
D_{a,i} = D_0 \left[ 1 - \sqrt{\frac{K_e(1-f)}{3f}} \right]
\]  

(6)

\[
D_{e,i} = D_0 \left[ 1 + \sqrt{\frac{KF_e}{3(1-f)}} \right]
\]  

(7)

The intra-axonal diffusivity:

\[
D_a = \text{tr}(D_a)
\]  

(8)

The axial diffusivity in the extra-axonal space:

\[
D_{e,i} = \lambda_{e,1}
\]  

(9)

where \(\lambda_{e,1}\) was the primary eigenvalue of \(D_e\).

The radial diffusivity in the extra-axonal space:

\[
D_{r,i} = \frac{\lambda_{e,2} + \lambda_{e,3}}{2}
\]  

(10)

where \(\lambda_{e,2}, \lambda_{e,3}\) were the 2nd, 3rd eigenvalues of \(D_e\).

Artifacts rejection and tensor estimation were performed by using the in-house software implemented in MATLAB version 7.11.0 (MathWorks, Natick, MA, USA).

**TBSS and Region of interest (ROI) analysis.** TBSS was performed by using the optimized pipeline for neonates. All the FA images were normalized to the neonatal Johns Hopkins template. The aligned FA image of each subject was projected onto the mean FA skeleton (threshold = 0.15). Inter-group comparisons of the above metrics were tested with adding covariates (including GA, PMA and BW) in the general linear model. The number of permutations was set at 5000. All tests were taken to be significant at \(P < 0.05\) after family-wise error rate (FWE) correction with threshold-free cluster enhancement (TFCE). ROI analysis was also performed based on the Johns Hopkins University WM label atlas. The PLIC, SCR, CC and EC, which respectively represented projection fibers, commissural fibers and association fibers respectively, were selected as target regions.

**Statistical analysis.** Statistical analysis was conducted using SPSS for Windows version 17.0 (SPSS, Chicago, IL, USA). Measurement data are reported as means ± standard deviations or medians with ranges, and categorical data as frequencies and percentages. Student's t-test, one-way analysis of variance, the rank-sum test, χ² test, as appropriate, were used for comparisons of demographic data among groups. All parameters from ROI measurements were compared using a general linear model with GA, PMA and BW as covariates. Observer agreement was evaluated by Kappa analysis, and assessment criteria were as follows: 0.00, poor; 0.01–0.20, slight; 0.21–0.40, fair; 0.41–0.6, moderate; 0.61–0.8, substantial; and 0.81–1.0, almost perfect.

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Author Contributions
J.Y. conceived the study. J.G. and Y.Y.L. performed the experiments. J.G., Y.Y.L., X.J.L., L.X.Z., Q.L.S. and B.L.Y. analyzed the data. J.Y. supervised the experiments. J.G., X.J.L., C.J., D.X. and J.Y. wrote the manuscript. All of the authors discussed the results and reviewed the manuscript.

Additional Information
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