Flavonoids are a versatile class of natural polyphenolic compounds that represent secondary metabolites from higher plants. Their basic structures consists of fifteen-carbon skeleton consisting of two benzene rings (A and B) linked via a heterocyclic pyrane ring (C) to produce a series of subclass compounds such as flavones, flavonols, flavanones, isoflavones, flavanols or catechins and anthocyanins. Their biological activities are dependent on the structure, chemical nature and degree of hydroxylation, substitutions, conjugation and degree of polymerization. A brief description of flavonoids, its source and classification have been described. Although flavonoids are integral in nutraceutical, pharmaceutical, medicinal, cosmetic and other applications their bioavailability to the target tissues and cells are restricted due to poor water solubility and enzymatic degradation. To increase effectiveness, currently encapsulation of the drug candidate in biological material that are able to enhance the potential health benefits by increasing the water solubility and targeted delivery are being achieved. Biodegradable natural, synthetic and semi-synthetic material/ polymers approved by the US Food and Drug Administration (FDA) for use in the preparation of nanodrugs as well as the applied encapsulation technique are discussed that prevent against oxidation, isomerization and degradation of the flavanoids. The aim of this review is to identify specific flavonoids that exhibit increased pharmacological and biological efficiencies on encapsulation. Thus, these potential drugs may help in preventing many chronic diseases and lead to future research directions.

Keywords: Flavonoids, Encapsulation, Delivery Systems, Biological Activity

1. Introduction

Flavonoids are secondary metabolites ubiquitously present in plants that comprise a large group of polyphenolic compound with benzo-γ-pyrene structure responsible for variety of pharmacological activities [1, 2]. The flavonoids are mainly accumulated in the edible parts of plants particularly in fruits and vegetables, responsible for red and dark blue color of berries as well as orange and yellow color in citrus fruits. They also act as a secondary antioxidant defense system in plant tissue exposed to different abiotic and biotic stress and regulate growth factor in plants such as auxin [3].

In the human body they play similar role as vitamins [4, 5]. Their activities are dependent on the structure and chemical nature, degree of hydroxylation, substitutions, conjugation and degree of polymerization. Flavonoids shows a variety of biological activities such as antioxidants, modulators of cell signaling, anti-inflammatory agents, cardio protectants, inhibitors of neurodegeneration and ability to inhibit the growth of a wide range of microorganisms and viruses [6-8].

The chemical structures of flavonoids are based upon a fifteen-carbon skeleton consisting of two benzene rings (A and B) linked via a heterocyclic pyrane ring (C) shown in Figure 1. The classification of flavonoids depends on the level of oxidation and pattern of substitution on the C ring, while individual compound within a class differ in the pattern of substitution on the A and B rings. The main classes of flavonoids, structures and examples with the position of substituents are shown in Table 1 [9].
Table 1. Main classes of flavonoids, structures, examples with position of substituents [14, 15].

| Class of Flavonoids | Structure | Name | Position of Substituents |
|---------------------|-----------|------|-------------------------|
| Flavanones          |           |      |                         |
|                     |           | Hesperetin | 5,7,3',4'-OH, 4'-OMe |
|                     |           | Naringin  | 5,4'-OH, 7-OR           |
|                     |           | Naringenin | 5,7,4'-OH              |
|                     |           | Eriodictyol | 5,7,3',4'-OH         |
|                     |           | Hesperidin | 5,7,3',4'-OH, 4'-OMe, 7-rutinoside |
|                     |           | Likvirtin | 7-OMe                  |
| Flavan-3-ols        |           | (+)-Catechin | 5,7,3',4'-OH       |
|                     |           | Epigallocatechin | 5,7,3',4',5'-OH  |
|                     |           | Epigallocatechin | 5,7,3',4'-OH       |
|                     |           | Gallate    | 7,4'-OH                |
| Flavones            |           | Chrysin    | 5,7-OH                 |
|                     |           | Apigenin   | 5,7,4'-OH              |
|                     |           | Leteolin   | 5,7,3',4'-OH           |
| Flavonols           |           | Rutin      | 5,7,3',4'-OH, 3-rutinoside |
|                     |           | Kaempferol | 5,7,4'-OH              |
|                     |           | Quercetin  | 5,7,3',4'-OH           |
|                     |           | Galangin   | 5,7-OH                 |
| Isoflavones         |           | Genistein  | 5,7,4'-OH              |
|                     |           | Daidzein   | 7,4'-OH                |
|                     |           | Puerarin   | 7,4'-OH, 8-glucoside   |
|                     |           | Glycitein  | 7,4'-OH, 6-OMe         |
| Flavanonol          |           | Taxifolin  | 3,5,7,3',4'-OH         |

Being phytochemicals, flavonoids cannot be synthesized by human and animal and hence form an integral part of human and animal diet [8, 10]. The main classes of flavonoids, food source, their specification and important biological properties are reported in Table 2.

The physicochemical properties of flavonoids such as molecular size, configuration, lipophilicity, solubility, pKa and structure; viz glycoside or aglycone could play a vital role in the absorption of dietary flavonoids. Liberated from food by chewing, aglycans can be easily absorbed by small intestine, while flavonoid glycosides have to be converted into the aglycan [11].

Flavonoids are poorly absorbed in the intestine in their natural form, and are extensively degraded by intestinal microorganisms and/or enzymes, to produce different metabolite. If these metabolite adsorbed are subjected to the hepatic enzymatic system the new metabolites formed differ in their bioactivity.

After the hydrolysis of sugar moieties in the small intestine or due to bacterial activity in the colon, aglycones are generated and further metabolized before reaching the systemic circulation. Briefly, numerous factors could play a role in limiting the glucuronidated or the sulfated form. As a consequence, flavonoids results in poor bioavailability, poor permeability, instability and extensive first-pass bioavailability of flavonoids [12, 13].

Table 2. Main classes of flavonoids, food source, their specification and important biological properties [16-29].

| Class of Flavonoids | Dietary Source | Specifications | Main biological properties |
|---------------------|----------------|----------------|---------------------------|
| Flavanones          | Fruits and vegetables (grape berries, apple, tomato, onion, broccoli and red lettuce), green tea, black tea and red wine | Flavonols are the most ubiquitous flavonoids in food, sensitive to oxidation, lights and pH aglycones slightly soluble but glycosides soluble in water. | Vitamin P factor protecting capillaries and veins, often |
| Flavones            | Parsley, broccoli, celery, carrots, onion leaves, | Natural pigment, flavones are much less common than flavonols | |
2. Encapsulation of Bioactive Compound

Encapsulation is a “nature made” technique used for product formulation to trap important biological ingredients into a carrier, which protect the trapped biological material against oxidation, isomerization and degradation. This technique increases the shelf life of material over a period of time and control/sustained delivery of functional substances when ingested in the body [30]. It also improves the solubility and pharmacokinetics profiles of insoluble drugs. In many cases, targeted drug delivery is greatly enhanced, bioavailability to the target tissues and cells are significantly improved. It reduces their toxic side effects to normal cells and increases the delivery of such drug to tumor tissue [31].

### Table 3. Encapsulating materials are classified according to their origin as natural, synthetic and semisynthetic (Adapted from review “Critical evaluation of biodegradable polymers used in nanodrugs”, Edgar Marin et al. 2013, International Journal of Nanomedicine 2013: 8 3071–3091) [37].

| Origin        | Sub classification            | Examples                                                                 |
|---------------|-------------------------------|--------------------------------------------------------------------------|
| Synthetic     | Hydrolyzable backbones        | Poly (glycolic acid)                                                     |
| Polyesters    |                               | Poly (lactic acid)                                                       |
|               |                               | Poly (caprolactone)                                                      |
|               |                               | Poly (lactic-co-glycolic acid)                                           |
|               |                               | Poly (butylene succinate)                                                |
|               |                               | Poly (trimethylene carbonate)                                            |
|               |                               | Poly (p-dioxanone)                                                       |
|               |                               | Poly (butylene terephthalate)                                            |
| Poly(ester amide) s |                      | Hybrane® S120043                                                         |
| Polyurethanes |                               | DegraPol®45                                                              |
| Polyanhydrides|                               | Pol [(carboxyphenoxy) propane-sebacic acid]                              |
| Polyphosphoesters|                               | Poly [bis (hydroxyethyl) terephthalate-ethyl orthophosphorylate/terephthaloyl chloride] |
| Carbon backbones (hydrolysis cannot occur) | | Poly (ortho esters) I                                                    |
|               |                               | Poly (ortho esters) II                                                   |
|               |                               | Poly (ortho esters) III                                                  |
|               |                               | Poly (ortho esters) IV                                                   |
| Poly (alkyl cyanoacrylates) | | Poly (butyl cyanoacrylate)                                               |
| Polyether     |                               | Poly (ethylene glycol)                                                   |
| Poly (amino acids) |                             | Tyrosine derived polycarbonate                                           |
| Semisynthetic | Microbial polyesters          | Poly (β-hydroxyalkanoate) s                                              |
| Proteins      |                               | Poly (hydroxybutyrate)                                                   |
| Natural       | Animal source                 | Collagen                                                                 |
|               | Vegetable source Poly saccharides | Albumin                                                                  |
|               | Animal source                 | Chitosan                                                                 |
|               | Vegetable source              | Hyaluronate                                                              |
|               |                               | Cellulose                                                                |
|               |                               | Alginate                                                                 |
|               |                               | Starch                                                                   |

Various newly synthesized chemical entities such as poly (lactic-co-glycolic acid) (PLGA), poly (glycolic acid) (PGA) and poly (lactic acid) (PLA) have been approved by the US Food and Drug Administration (FDA) with a wide therapeutic efficacy and easy availability in the market. Since, ancient times, herbal remedies and natural extract are used to cure...
various diseases as they contain several phytoconstituents which work simultaneously against the disease. Conventional therapy provides non-targetability in tissue and organs due to peak and valley fluctuations and requires a frequent dose of administration. The controlled release of drug delivery system provides drug released at a controlled rate and maintains the overall therapeutic concentration of drug in the body [32-36].

There are various techniques that are used for encapsulation such as spray drying, spray cooling/chilling, extrusion, fluidized bed coating, co-acervation, liposome entrapment, inclusion complexation, centrifugal suspension separation, lyophilization, co-crystallization and emulsion, nanoparticles etc. [38, 39].

Generally three steps are involved in the encapsulation of bioactive agents.
1. The formation of wall around the bioactive compound (core material) to be encapsulated.
2. Ensuring that undesired leakage does not occur.
3. Ensuring that undesired materials are kept outside [40, 41].

The effectiveness of nutraceutical product in preventing disease depends on preserving the bioavailability of the active ingredients. After oral administration only small proportion of the molecules are made available due to insufficient gastric resistance time, low permeability and/or solubility within the gut as well as conditions during food processing and storage (temperature, oxygen, light) or in the gastrointestinal tract (pH, enzymes, presence of other nutrients), all these factor limit the activity and potential health benefits of the nutraceutical component [42]. To increase the activity and health benefits it requires product formulation to provide protective mechanism that can maintain the active chemical form until the time of consumption, and deliver this form to the physiological target within the organism [43].

3. Material Used for Encapsulation of Bioactive Compound

Several encapsulating materials can be broadly classified according to their origin as natural, synthetic and semisynthetic materials as shown in Table 3. These materials are biodegradable, biocompatible, non-toxic, non-immunogenic and enhance the stability, bioavailability and bio efficacy of bioactive compound or materials [44]. List of biodegradable polymers approved by the US Food and Drug Administration for use in the preparation of nanodrugs as shown in Table 4. A summary of most widely used natural, synthetic and semisynthetic encapsulating materials are presented below.

| Ingredient name | Route- dosage form | CAS Number |
|-----------------|--------------------|------------|
| Acrylates copolymer | O and TD- Tablet, chewable, film coated, extended release, orally disintegrating, delayed release, film, extended release and patch | --- |
| Ammonio methacrylate copolymer | O-Tablet, capsule and extended release | --- |
| Ammonio methacrylate copolymer type A | O-Powder, for suspension, tablet, extended release and film coated | 33434241 |
| Ammonio methacrylate copolymer type B | O-Capsule, extended release, tablet, chewable and film coated | 33434241 |
| Ammonium calcium alginate | O-tablet | --- |
| Butyl ester of methyl vinyl ether/maleic anhydride copolymer (125000 MW) | T-solution | 25119680 |
| Butyl methacrylate and methyl methacrylate copolymer (3:1: 150000 MW) | Td-patch | 25608337 |
| C13-14 isoparaffin/laureth-7 /polyacrylamide | T-gel | --- |
| Calcium alginate and ammonium alginate | O-tablet | --- |
| Caprylic/capric/succinic triglyceride | SL-aerosol | 97708731 |
| Caprylocaproyl polyoxyglycerides 8 | O-Capsule, liquid filled and solution | 361459383 |
| Carbomer copolymer type A (allyl pentaerythritol crosslinked) | O and T-Emulsion cream and lotion | 9007209 |
| Carbomer copolymer type B (allyl pentaerythritol crosslinked) | OPH, T and TD-Emulsion, cream, gel, lotion, film and extended release | 9007209 |
| Carbomer copolymer type C (allyl pentaerythritol crosslinked) | T-Gel and metered | 9007209 |
| Carbomer homopolymer | O, R and T-Tablet, extended release, enema, disc, gel, lotion and patch | 9007209 |
| Carbomer homopolymer type A (allyl pentaerythritol crosslinked) | O and T-Capsule, tablet, extended release, gel and lotion | 9007209 |
| Carbomer homopolymer type B (allyl pentaerythritol crosslinked) | OPH, O, T and V- Gel, suspension, suspension/ drops, capsule, granule, for suspension, tablet, extended release, cream, gel, lotion, solution. | 9007209 |
| Carbomer homopolymer type B (allyl sucrose crosslinked) | B, OPH, O, R and T-Tablet, suspension, suspension/ drops, capsule, suspension, extended release, orally disintegrating, enema, cream, augmented, emulsion, gel, lotion, ointment and solution | 9007209 |
| Carbomer homopolymer type C (allyl pentaerythritol crosslinked) | OPH, T and TD- Gel, cream, augmented, emulsion, gel, lotion, ointment, and metered | 9007209 |
| Cellulosic polymers | O-Capsule, delayed release, tablet, extended release and film coated | --- |
| Detosu/triethyleneglycol/triethylene glycol polyglycolide copolymer | SC- Injection | --- |
| Ingredient name                                                                 | Route- dosage form                                                                 | CAS Number     |
|--------------------------------------------------------------------------------|-----------------------------------------------------------------------------------|----------------|
| Dimethiconol/trimethylsiloxysilicate crosspolymer (40/60 w/w; 1000000 pa.s)     | O and TD- Tablet, extended release, film and patch                                 | ---            |
| Dimethylaminoethyl methacrylate - butyl methacrylate - methyl methacrylate copolymer | O and TD-Capsule, extended release, pellet, suspension, tablet, chewable, coated, delayed release, film coated, orally disintegrating and patch | 24938167      |
| Ethyl acrylate and methyl methacrylate copolymer (2:1; 600000 MW)             | O-Tablet and extended release                                                     | 9010882       |
| Ethyl acrylate and methyl methacrylate copolymer (2:1; 750000 MW)             | O-Capsule, pellets, extended release, granule, tablet, coated, film coated, orally disintegrating and delayed release | 9010882       |
| Ethylene-propylene copolymer                                                  | TD-Film, extended release and patch                                              | ---            |
| Ethylene-vinyl acetate copolymer (28% vinyl acetate)                           | SC and V-Implant, insert and ring                                                 | 24937788      |
| Ethylene-vinyl acetate copolymer (9% vinyl acetate)                            | V-Insert and ring                                                                 | 24937788      |
| Ethylene-vinyl acetate copolymers                                             | IU, OPH, SC and TD-Insert, suppository, extended release, implant and film       | 24937788      |
| Glycerin polymer solution I-137                                               | O-tablet                                                                          | ---            |
| Isocetyl acrylate/acrylamide/vinyl acetate copolymer, kollidon VA 64 polymer  | O and T-Tablet, film coated and sponge                                             | ---            |
| Laurroyl PEG-32 glycerides                                                   | O-Capsule and tablet                                                              | 121548047     |
| Laurroyl polyoxylglycerides                                                  | O-Capsule, tablet and film coated                                                 | ---            |
| Maltodextrin                                                                | O-Capsule, film, soluble, granule, for suspension, lozenge, paste, solution, suspension, tablet, chewable, coated, effervescent, extended release, film coated and orally disintegrating | 9050366       |
| Methacrylic acid - ethyl acrylate copolymer (1:1) type A                     | O-capule, coated, coated pellets, delayed release, granule, for suspension, tablet, coated particles, film coated, extended release and orally disintegrating | 25212888      |
| Methacrylic acid - methyl methacrylate copolymer (1:1)                        | O-capule, coated pellets, extended release, suspension, tablet, delayed release and film coated | 25086151      |
| Methacrylic acid - methyl methacrylate copolymer (1:2)                        | O-Capule, delayed release, tablet, extended release and film coated               | 25086151      |
| Methacrylic acid copolymer                                                   | O-Capule, coated, coated pellets, extended release, delayed release, for suspension, suspension, tablet, film coated and orally disintegrating | ---            |
| Methyl acrylate - methyl methacrylate PEG/PPG-4/30 copolymer                 | O-Tablet and extended release                                                     | ---            |
| Pigmented polyethylene/polyester 1501 film                                   | OPH-solution                                                                      | 90003116      |
| Poly (DL-lactic-co-glycolic acid), (50:50; 12000 MW)                          | Td-patch                                                                          | ---            |
| Poly (methyl acrylate-co-methyl methacrylate-co-methacrylic acid 7:3:1; 280000 MW) | O-Capule and extended release                                                     | 26936243      |
| Polyacrylic acid (250000 MW)                                                  | T and TD-Patch, film and extended release                                         | 9000314       |
| Polybutene (1400 MW)                                                         | B, OPH and T- Film, soluble, tablet, gel, solution, suspension/ drops and patch   | 90003978      |
| Polyacrylate k                                                               | O-Tablet, coated, extended release and film coated                                | 68424044      |
| Polydextrose k                                                               | O-Tablet and film coated                                                         | ---            |
| Polyether                                                                   | TD and V-Film, extended release, patch and insert                                 | ---            |
| Polyether polyamine copolymer                                                | TD-Film and extended release                                                     | ---            |
| Polyethylene glycol 1000                                                      | O, R, T, TD and V- Concentrate, solution, tablet, film coated, suppository, aerosol, foam, cream and gel | 25322683      |
| Polyethylene glycol 1450                                                     | O, T and U-Capsule, extended release, solution, suspension, tablet, film coated, ointment and suppository | 25322683      |
| Polyethylene glycol 1600                                                     | D, O, R and T-Gel, paste, tablet, coated, suppository and solution                | 25322683      |
| Polyethylene glycol 200                                                        | AU, O and T-Drops, capsule, solution, tablet, extended release and ointment       | 112607        |
| Polyethylene glycol 20000                                                      | O-Capsule, tablet and delayed release                                            | 25322683      |
| Polyethylene glycol 300                                                        | AU, IM, IV OPH, O and T- Drops, injection, solution, liquid, ointment, tablet, film coated, cream and lotion | 25322683      |
| Polyethylene glycol 3000                                                      | O-Tablet and extended release                                                   | 25322683      |
| Polyethylene glycol 3350                                                     | IA, IL, IM, IS, IV, N, O, R, ST, SC, T and V- Injection, suspension, solution, capsule, extended release, suspension, tablet, chewable, coated, delayed release, film coated, orally disintegrating, suppository, cream and ointment | 25322683      |
| Polyethylene glycol 400                                                        | IM, IV, N, OPH, O, R, T and V-Injection, spray, metered, solution/ drops, cpsule, delayed and extended release, liquid filled, concentrate, suspension, syrup, tablet, coated particles, orally disintegrating, aerosol, powder, cream, emulsion, ointment, sponge, swab and suppository | 25322683      |
| Polyethylene glycol 4000                                                      | D, IA, IM, IS, IV, O, R, SL, T and V-Ointment, injection, suspension, extended release, capsule, delayed release, granule, solution, syrup, tablet, coated, film coated, multilayer, orally disintegrating and suppository, cream | 25322683      |
| Polyethylene glycol 4500                                                      | O-Capsule, extended release, tablet and film coated                              | 25322683      |
| Polyethylene glycol 540                                                       | T-ointment                                                                       | 25322683      |
| Polyethylene glycol 600                                                       | IV, O and T-Injection, solution, capsule, liquid filled, tablet, delayed and extended release | 25322683      |
| Polyethylene glycol 6000                                                      | B, O, R, SL, T and V-Tablet, capsule, delayed and extended release, coated, film coated, multilayer, orally disintegrating, suppository, cream and insert | 25322683      |
3.1. Chitosan

Chitosan is a natural N-deacetylated derivative of chitin polycationic polysaccharide consisting linear repeating unit of 2-acetamido-2-deoxy-D-glucose and β-(1-4)-2-amino-2-deoxy-D-glucose show in Figure 2 [45]. The presence of hydroxyl and amino functions show the modulatory effect on cellular-F actin, tight junction protein ZO-1 and protein kinase C and are rapidly internalized by the cell [46-48]. Chitosan is biodegradable, biocompatible, low toxic, non-immunogenic and mucoadhesive in nature. Therefore, chitosan favors wide range of biomedical applications including tissue engineering, drug delivery, wound dressing antimicrobial activity, anti-inflammatory and antioxidant properties [49-52]. As a drug carrier, chitosan has the capacity to deliver drugs to various organs such as kidney, liver, lung and colon. Due to the polycationic characteristics chitosan can interact with the negatively charged molecules and form an efficient nanostructure drug delivery system of several bio molecules (drugs, flavonoids, proteins, DNA) [53, 54]. In addition being a known coadhesive polymer, chitosan helps to prolong mucus binding time of the drug molecules and transiently open the tight junction between the epithelial cells and help the drug transport in a sustained manner [52]. Chitosan are easily...
digested by the chitosanase enzyme secreted by the microorganism in the intestine. Therefore, a combination of chitosan and its derivatives (O-Carboxymethyl chitosan) enhance drug absorption through small intestine involving clathrin-mediated endocytosis [55]. Hazra et al, reported enhanced and controlled oral dosage form for the delivery of hydrophobic flavonoid quercetin in chitosan-coated alginate microsphere. The formulations demonstrated drug entrapment efficiency of ~80% and scanning electron microscopy (SEM) study confirmed the smoothness of polymeric microsphere with a significant pH sensitive swelling index and drug release at simulated gastrointestinal media. The release of quercetin from microsphere shows the absolute retention in gastric fluid (pH-1.2), where in sustained drug is released at pH 7.4 [56]. In another study quercetin loaded chitosan nanoparticles were prepared by the ionic gelation of cationic chitosan with tripolyphosphate (TPP) anions having particle size ~76.58 nm with uniform, smooth surfaced, ellipsoidal shaped nanospheres. The antioxidant activity of quercetin loaded chitosan nanoparticles also indicated that chitosan nanoparticles are useful in improving quercetin oral bioavailability [57].

3.2. PLGA and PLA Nanoparticle

Poly lactic-co-glycolic acid (PLGA) and poly (D, L-lactic acid) PLA are biodegradable polymers because its hydrolysis leads to metabolite monomers lactic acid and glycolic acids which are finally metabolized to CO₂ and H₂O via Kreb’s cycle. PLGA and PLA are approved by USFDA and European Medicine Agency (EMA) in various drug delivery systems in humans (Figure 3) [60]. Being biocompatible polymers, these polymers have been numerous used for the drug delivery and biomaterial applications. The polymers are synthesized with different molecular and co-polymer compositions with diverse properties like modulated size, drug loading ability, nanoparticle uptake efficiency, controlled drug release, bio-distribution and circulating half life of the nano drug composites [61-63]. Bishayee K et al. studied the anti-proliferative activity of quercetin-gold loaded PLGA nanoparticles on HePG2 hepatacarcinoma cell. The drug action was via interacting with cellular DNA, reduction in deacetylation of histone proteins and arresting cell growth in the sub-G stage [64]. In another study quercetin loaded PEGylated PLGA nanoparticle loaded with folic acid demonstrated enhanced uptake of folic acid by intravenous treatment in IGROV-1 (human ovarian adenocarcinoma cell) and HeLa (Human epithelial cells) in xenograft models [65]. Similarly, quercetin encapsulated with PLA nanoparticles by emulsified nanoprecipitation method was studied. The reduction in breast cancer cell approximately 50% (in 2 days) due to sustained release of drug from PLA-quercetin nanoparticle and 40% (in 5 days) with free quercetin were observed [66].

3.3. Liposomes

Liposomes are composed of lipidic amphiphiles usually phospholipids which can form a bilayer membrane spherical vesicle and organize themselves in water to form an aqueous core surrounded by lipid bilayers as shown in Figure 4. The lipid structures of liposomes have capacity to carry and transport both hydrophilic and hydrophobic therapeutic agent [67, 68].

The physiochemical properties of liposomes like size, surface charge, composition, membrane rigidity etc. and the targeting ligand can modulate bioavailability, uptake, pharmacokinetic and bio distribution and in vitro and in vivo stability making it feasible for controlled and targeted drug delivery system. Liposomes are able to deliver low doses of drugs with reduced toxicity and side effects [69].
3.4. Cyclodextrin (CDs)

Cyclodextrin (CDs) are natural macrocyclic oligosaccharides with toroidal structures having lipophilic cavities and a hydrophilic outer surface, thus capable to form inclusion complexes with hydrophobic molecule to significantly increase water solubility [70, 71]. Cyclodextrin inclusion complexes are capable to protect the active ingredients against oxidation, decomposition, light induced reactions, ocular disturbance, microbiological contamination, drug-additive interactions, hygroscopicity etc. There are three natural Cyclodextrin viz. α, β and γ- consisting of 6, 7 and 8 glucopyranose units linked by α-(1,4) bonds with internal diameter 0.5 to 0.8 nm (Figure 5) [72]. Moreover, a semi-synthetic derivative e.g. α-cyclodextrin and co-polymers of cyclodextrin can enhance the solubility in water, rate of release, inclusion capacity and reduce the side effects [73]. Cyclodextrin and its derivatives are reported as an attractive candidate for biomedical applications including drug delivery; improve water solubility, stability, increase antioxidant activity, bioefficacy and bioavailability. Zheng et al. studied the chemical stability and water solubility of quercetin with three β-cyclodextrin derivatives such as unsubstituted β-cyclodextrin, hydroxylpropyl-β-cyclodextrin (HP-β-CD) and sulfobutyl ether-β-cyclodextrin (SBE-β-CD) at alkaline pH. The study revealed that β-cyclodextrin/ quercetin complex sustainably improved the solubility and stability due to formation of inclusion complex model studied by Nuclear Magnetic Resonance (NMR) spectroscopic analysis [74].

Carlotti et al. reported that the preparation of quercetin inclusion complex with methyl-β-cyclodextrin (Me-β-CD) improved the quercetin solubility without affecting the antioxidant activity and photostability, in vitro accumulation of quercetin in porcine skin studies with Franz diffusion cell [75].

3.5. Miscellaneous Nanoparticles

To improve the delivery of drug in vitro and in vivo Wang et al. reported quercetin co-encapsulated fluorescent silicon quantum dots (SiQDS) in poly (ethylene glycol)–block–polylactide (PEG-PLA) by double emulsion method for simultaneous in vitro imaging and biocompatibility studies (Figure 6). The encapsulated nanoparticles effectively suppress human hepatoma HePG2 cell proliferation than free quercetin and significantly inhibit the hydrogen peroxide-induced DNA damage in HePG2 cells [76]. Barreto et al. proposed a new magnetic nanoparticle (Fe$_3$O$_4$) incorporated to a triblock co-polymer of ethylene dioxide and oxyphenylethylene for quercetin delivery in cancer treatment. The magnetic nanoparticle demonstrated a targeted drug delivery and sustained release of drug (its peak at 14.5% after 96 h) [77]. Several researchers have reported the use of mesoporous silica nanoparticles as a promising drug delivery system due to low in vivo toxicity, stability, targeted drug delivery, high surface area and high drug loading efficiency with better release kinetics [78-83]. In another study Catauro et al. studied the silica-quercetin hybrid material using a sol-gel synthesis method for treatment of peri-implant diseases [84].

Figure 5. Structure of α, β and γ-cyclodextrin (CD).

Figure 6. SEM micrographs of: (A) SiO2/Quercetin hybrids 5 Si/Que5, (B) Si/Que10, and (C) Si/Que15.
4. Biological Activity of Encapsulated Flavonoids

Several biological and pharmacological activities of encapsulated flavonoids are widely known. A summary of the biological studies on encapsulated flavonoids are presented below.

4.1. Anticancer

4.1.1. Cytotoxicity Studies

Encapsulation of naturally occurring flavonoids such as Quercetin, Isoscutellarein, Rutin and Isoscutellarein glycoside into liposomes were tested against the cancer cell line SF268 (central nervous system), H460 (non-small cell lung) and MCF7 (breast) by M. Goniotaki et al. The result showed that the quercetin had growth inhibiting (GI50) activity against the cancer cell lines SF268 (31.75µM), MCF7 (24.19 µM) and H460 (80.0 µM). At higher concentration all the flavonoids were inactive against normal cells (peripheral blood mononuclear cells: resting or activated). The liposomal formulation of quercetin was less active than its free form. The liposomal formulation rutin proved to be more active and showed remarkable growth inhibition activity against H460 and SF268 cell lines and the liposomal formulation of isoscutellarein shows considerable growth inhibition activity for all cell lines and best among the all tested flavonoids. The free liposomes were inactive against all cell lines [85].

In another study Gao et al. stated that the encapsulation of quercetin with biodegradable monomethoxy poly(ethylene glycol)-poly(ε-caprolactone) MPEG-PCL micelles suppressed the growth of established xenograft A2780S ovarian tumors through cell apoptosis and inhibiting angiogenesis in vivo. The quercetin loaded micelles showed 36 nm of mean particle size with 6.9% drug loading [86]. Chitosan nanoparticles of Epigallocatechin-3-gallate(EGCG) nanoformulated by graphene nanosheet showed anticancer effect on human melanoma Mell 928 cells by apoptosis via increase in Bax levels, increased poly (ADP-ribose), polymerase (PARP) cleavage, G2/M phase cell cycle arrest, inhibition of cyclin D and D3 induction of p21 and p27, decrease in Bcl-2, caspases-3 and caspases-9 protein expression, which resulted in reduction of cancer cell viability. The EGCG nanoformulated with chitosan showed anticancer effect on xenograft athymic mouse model of melanoma by suppression of tumor group and proliferation, inhibition of CDK4 and 6 and an increase in apoptosis [87]. In another study EGCG nanoformulated as Ca/Al-NO3 layered double hydroxide induced apoptosis, decreased the cell viability and inhibited colony formation in human prostate cancer PC-3 cells [88].

4.1.2. Colon Cancer

The natural flavonoid fisetin (3, 3’, 4’, 7’-tetrahydroxyflavone) encapsulated with monomethyl poly-(ethylene glycol)-poly (ε-caprolactone) (MPEG-PCL) was used to prepared nanoassemblies of fisetin by a self-assembly procedure. Yishan Chen et al studied the effectiveness of fisetin micelles and a promising source for colon cancer therapy with high antitumor activity and low toxicity. The prepared fisetin micelles with particle size 22.4 ± 3.0 nm, polydispersity index 0.163 ± 0.032, the drug loading (DL) and encapsulation efficiency were 9.88±0.14% and 98.53 ± 0.02%. In vitro release study of fisetin micelles demonstrated a sustained and prolonged release than free fisetin and the cumulative release rate of fisetin micelles was 73.58 ± 3.99% and free fisetin was 92.95 ± 6.51% (P<0.05). The cytotoxicity of fisetin micelles by MTT assay indicate that cell viability of CT26 and L929 cells were upto 72.13 and 66.13 respectively. The results indicated that MPEG-PCL were biocompatible and exhibited low toxicity. Fisetin micelles cellular uptake and apoptosis in CT26 cells was higher than that of free fisetin. The in vivo studies were more efficient in suppressing growth and prolonging survival time of tumors than free fisetin (P<0.05). Tumor were analyzed using histological analysis (H & E), TUNEL assay, immunohistochemical detection of Ki-67 cell proliferation and immunofluorescence detection of micro vessel density (MVD). The fisetin micelles enhanced apoptosis induction, antiproliferation and antiangiogenesis effect than free fisetin in the animal model [89].

Multifunctional solid lipid nanoparticles loaded with a cyanine-type IR-780 acting as a diagnostic agent and a photosensitizer and a flavonoid derivative baicalein or fisetin as a therapeutic cargo were fabricated using a solvent diffusion method. The drug loaded lipid nanoparticle exhibited anticancer effect in colon adenocarcinoma cells with lower cytotoxicity and decrease in tumor growth on loVo and CHO-K1 cell lines. They also showed an increased p53 and MnSOD (Manganese superoxide dismutase) expressions after PDT-SLN-EP (photodynamic therapy-solid lipid nanoparticles-electroporation) [90]. Epigallocatechin-3-gallate (ECCG) nanoformulated by graphene nanosheet showed anticancer effect on colon cancer HT29 and SW48 cells, via photothermal destruction of cell assessed by high efficiency near-infrared photothermal therapy [91].

4.1.3. Lung Cancer

Baicalein nanoparticle with dual-targeted ligands of folate and hyaluronic acid showed the anticancer effect on human lung cancer A549 cells as well as paclitaxel-resistance lung cancer A549/PTX by decreasing cell viability and inhibiting tumor growth [92]. Luteolin nanoformulated with PLA-PEG polymer possesses anticancer effect against lung cancer H292 cells and TU212 head and neck squamous cell. The mode of anticancer effect was observed via inhibition of tumor growth, tumor size and colony formation in xenograft mouse model of head and neck cancer [93].

4.1.4. Breast Cancer

Ming sun et al. studied the quercetin-nanostructured lipid carrier (Q-NLC) synthesized using a phase inversion based process method. The size of NLC was 32 nm, the loading capacity and encapsulation efficiency of Q-NLC were 11% and 95% respectively. Q-NLC enhanced the cytotoxicity and
apoptosis in MCF-7 and MDA-MB-231 breast cancer cells. The void NLC was found to be extremely less toxic for the breast cancer cells [94]. Kadari et. al studied the anticancer activity against MCF-7 breast cancer cells for fisetin (FST) encapsulated into PLGA (poly-lactide-co-glycolic acid) nanoparticles as a complex of HPβCD (Hydroxy propyl β cyclodextrin). In vitro studies with nanoformulation FHIC-PNP (FST-hydroxyl propyl β cyclodextrin inclusion complex into PLGA nanoparticles) showed 3.9 times higher toxicity than pure fisetin against MCF-7 human breast cancer cell lines and enhanced the FST-induced apoptosis and ROS generation. In vivo studies in C57BL6 mice revealed that incorporation of FHIC in FHIC-PNP improved the pharmacokinetics and oral bioavailability of fisetin [95].

EGCG core-shell PLGA-casein nanoparticles in combination with paclitaxel demonstrated the anticancer activity on MCF-7 cells and human MDA-MB-231 breast cancer cells by increasing apoptosis and decreasing NF-kB activation [96]. Luteolin in phytosomes possesses anticancer effect by decreasing the expression at NrF2 and its related downstream gene HO1 on human MDA-MB-231 breast cancer cells [97]. Quercetin nanoformulated as phytosomes had anticancer effect on breast cancer MCF-7 cells by increasing apoptosis and decreasing mRNA expression of Nr2 downstream genes NQO1 and MRPI while no significant changes in Nr2 expression was observed due to free quercetin [98].

4.4. Antidiabetic Activity

The antimicrobial activity of hesperetin-loaded PLGA (poly (d, l-lactic-co-glycolic acid) nanoparticle may be attributed to the following (i) the structural properties of hesperetin (flavonoids nature) [103-105]. (ii) The potential charge on PLGA nanoparticle causing cell membrane depolarization [106-108]. (iii) Solubility of hydrophobic hesperetin increase after encapsulation, and (iv) the sustained release of active quercetin 150 and 300 nm (approximately) and the surface charge varied from -26.3 mV to -38.21 mV. Naringenin encapsulation efficiency and loading capacity of CS/ALG (Chitosan/Alginate) core shell nanoparticle at different weight ratios were varied between 57.34% to 98.36% and 7.41% to 19.87% respectively. Nanoparticles with weight ratio of 3:1 (CS: ALG) having encapsulation efficiency 91.4% and loading capacity 15.9% were further used for in vitro and in vivo studies. At pH 1.2 maximum 15% and at pH 7.4 more than 90% Naringenin was released in a slow sustained manner from the core shell nanoparticle. The in-vivo toxicity assessment showed no significant difference between NC (Rats treated with normal saline orally), NTBN (Rats treated with blank nanoparticles orally, 50 mg/kg body weight) and NTNN (Rats treated with Naringenin loaded core shell nanoparticles orally, 50 mg/kg body weight) groups. Regarding the fasting blood glucose, cholesterol and triglyceride and almost no change were observed in serum ALT (Serum alanine transaminase), AST (Aspartate transaminase) and ALP (Alkaline phosphatase) levels in normal and treated group of rats. The encapsulated Naringenin within nanoparticles helps to normalize the pancreatic abnormalities in diabetes, better than free oral Naringenin. The revival and recovery of hepatic tissue architecture appeared to be better in ND (Diabetic rats fed orally with Naringenin-loaded core shell polymeric nanoparticle 50mg/kg body weight) group then in FD (Diabetic rats fed orally with free Naringenin 50 mg/kg body weight, dissolved in 60% ethanol) group [101].

Chitrakara et al. studied the effect of quercetin loaded in PLGA nanoparticle by emulsion-diffusion-evaporation method. The average particle size of the nanoparticle was 179±11.2 nm, zeta potential -6.06±1-51 mV , polydispersity index 0-128 and ~86% quercetin entrapment efficiency. Surface morphology study confirmed the spherical shaped particles with smooth surface, ensuring the absence of unentrapped or adsorbed quercetin by scanning electron microscope (SEM). The nanoparticles retained the antioxidant property of quercetin due to easy lyophilization using D-trehalose (5%). In vitro release study confirmed a controlled release pattern of quercetin from the nanoparticles. In vivo, pharmacokinetics study revealed that the nanoformulation relatively increased the oral bioavailability (~52.3%) and the plasma quercetin concentration was sustained for 6 days, suggesting a reduced dosing frequency of the nanoformulations. An increased superoxide dismutase and catalase level in pancreas and kidneys after pre-oral treatment of nanoparticles were observed. Thus the nanocarriers could be an effective oral therapy for diabetes and its related complications which reduces dose as well as dosing frequency [102].
Duranoglu et al. studied the effective encapsulation of hesperetin into PLGA nanoparticles by using experimental design method. The formed nanoparticles showed maximum encapsulation efficiency (80.5±4.9%) and minimum particle size (260.2 ± 16.5 nm). The process was optimized as follows; 0.5% polyvinyl alcohol (PVA) concentration, 5:13 water: organic phase ratio and 3.59 mL min⁻¹ flow rate of the emulsified solution into 0.1% PVA. The cytotoxicity study or the biocompatibility of nanoparticles against the growth of L929 fibroblast cells was determined by the MTT method. The result revealed that the hesperetin and hesperetin-loaded nanoparticles were biocompatible with normal cell line L929 fibroblast cells up to 184.83 and 190.88 µg ml⁻¹ for 24 h and up to 133.24 and 134.80 µg ml⁻¹ for 48 h. The antimicrobial activity studies were carried using two different methods against Staphylococcus aureus and Escherichia coli. The MIC (minimal inhibitory concentration) values were 125 µg ml⁻¹ for Escherichia coli and 200 µg ml⁻¹ for Staphylococcus aureus, while the free hesperetin did not demonstrate activity in both strains (MIC value >200 µg ml⁻¹) [111].

4.5. Anti-quorum Sensing Activity

Sedef Ilk et al studied the kaempferol loaded chitosan nanoparticles by anti-quorum sensing mechanism against Chromobacterium violaceum CV026. The chitosan/ TPP nanoparticles were synthesized by ionic gelation method with nanoparticle size and zeta potential of 192.27± 13.6 and +35 mV. The loading and encapsulation efficiency of kaempferol loaded chitosan nanoparticles were 78% and 93%. The antioxidant activity evaluation by DPPH assay method indicates that kaempferol loaded chitosan/TPP nanoparticles show scavenging activity of 37±2.5% than free kaempferol (Scavenging activity 22±1.8%). The anti-QS activity of kaempferol loaded chitosan nanoparticle and free kaempferol by disc diffusion method on Chromobacterium violaceum CV026 at different storage time indicate that the nanoparticle inhibited violacein production up to 76%. Hence kaempferol loaded chitosan nanoparticles can act as strong and prolonged time stable quorum quenchers than corresponding pure kaempferol [112].

4.6. Antifungal Activity

Sedef Ilk et al studied the antifungal activity of kaempferol (KAE) loaded into lecithin/chitosan nanoparticles (Lc NPs) against the phytopathogenic fungus Fusarium oxysporium. The mean particle size, poly disperse index (PDI) and zeta potential were found to be 270 ± 10 nm, PDI≤0.2 and + 56 ± 4 mV respectively. KAE-LC NPs showed slow and sustained released for KAE in PBS + DMSO buffer at 37°C with encapsulation efficiency of 93.8 ± 4.28%. In vitro evaluation of KAE-LC NPs was studied by the release kinetics, antioxidant and antifungal activity in a time dependent manner against free KAE. The results demonstrate that nanoparticles had higher antioxidant and antifungal activity (67%) compared to free KAE (no inhibition) against Fusarium oxysporium by the end of 60 day storage period [113].

**Figure 7.** In vitro release profiles of KAE from lecithin/ chitosan nanoparticles (A) at 37°C, (B) at 25°C temperature.

**Figure 8.** Antioxidant activity of KAE-LC NP and pure KAE evaluated by reducing power (FRAP).
Figure 9. Antifungal activity of KAE-LC NPs and pure KAE against pathogenic fungi F. oxysporium. (A) Hyphal-extension growth inhibition. (B) Microscopic analysis of hyphae treated with KAE-LC NPs.

Figure 10. Effect of KAE-LC NPs and pure KAE on the radial growth of Fusarium oxysporium in time dependent manner. All determinations were performed in triplicate and the results expressed as mean±standard deviation.

4.7. Antioxidant Activity

Quercetin encapsulated nanoemulsions were produced using a low energy method-emulsion inversion point and two surfactant viz: Tween 80 and Brij 30. The average droplet diameters were 180-200 nm in the range. Quercetin loaded nanoemulsion incorporated in chicken pate was capable of protection against lipid oxidation but not against protein oxidation. Inhibition of secondary lipid oxidation was about 60% after 24 week of storage. While the free quercetin exhibited 35.4% inhibition and about 8.4% of inhibition in pates added with synthetic antioxidant such as butylated hydroxytoluene-BHT and sodium nitrite after 24 weeks [114].
Juan Huang et al. studied the quercetin and linseed oil encapsulated into nanostructured lipid carrier (NLC) by high pressure homogenization technique. The sustained release pattern of quercetin from quercetin loaded NLCs and antioxidant study by DPPH assay showed that linseed oil could improve the free radical scavenging activity of quercetin loaded NLCs [115].

5. Conclusions

Flavonoids form an integral part of human and animal diet. Flavonoids exhibit diverse categories of pharmacological and biological activities such as Anti-oxidant, Anti-inflammatory, Antidiabetic, Antimicrobial activity, Anti-quorum sensing activity, Anticancer, Modulators of cell signaling etc. Flavonoids exhibit the low water solubility, low permeability, gastric stability etc. which are the major limiting factor for the potential health benefits. Flavonoids when encapsulated with natural, synthetic and semisynthetic materials such as Chitosan, PLGA and PLA, Liposomes, Cyclodextrins etc. shows much better stability, bioavailability, increase shelf life, controlled and sustained release, protect against oxidation, isomerization and degradation etc.

However, only a small portion has been investigated or studied in both flavonoid encapsulation and its biological activities. There are gaps in the research, which need to be bridged in order to exploit the full medicinal potential of flavonoids or encapsulated flavonoids.

Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

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