Inflammation-related plasma protein levels and association with adiposity measurements in young adults

Susanna Klevebro1,2, Sophia Björkander3, Sandra Ekström3,4, Simon K. Merid1, Olena Gruzieva3,4, Anders Mälarstig5, Åsa Johansson6, Inger Kull1,2, Anna Bergström3,4 & Erik Melén1,2

Obesity-related inflammation is associated with cardiovascular, metabolic, and pulmonary diseases. The aim of this study was to demonstrate associations between adiposity measurements and levels of inflammation-related plasma proteins in a population of young adults. Subjects from a population-based birth cohort with a mean age of 22.5 years were included in the study population (n = 2074). Protein levels were analyzed using the Olink Proseek Multiplex Inflammation panel. Percentage body fat (%BF) and visceral fat rating (VFR) measurements were collected using Tanita MC 780 body composition monitor. Linear regression of standardized values was used to investigate associations. Potential effect modifications by sex and BMI category were assessed. Of 71 investigated proteins, 54 were significantly associated with all adiposity measurements [ %BF, body mass index (BMI), VFR and waist circumference]. Among proteins associated with %BF, seven showed a larger or unique association in overweight/obese subjects and three showed a significant effect modification by sex. Fourteen proteins more strongly associated with VFR in females compared to males. Adipose-associated systemic inflammation was observed in this young adult population. Sex and adiposity localization influenced some of the associations. Our results highlight specific proteins as suitable biomarkers related to adiposity.

Abbreviations
%BF Percentage body fat
BMI Body mass index
CDCP CUB domain-containing protein
FDR False discovery rate
FGF Fibroblast growth factor
GDNF Glial cell line-derived neurotrophic factor
HGF Hepatocyte growth factor
IL Interleukin
LAPTGF Latency-associated peptide transforming growth factor
LIFR Leukemia inhibitory factor receptor
LOD Limit of detection
MCP Monocyte chemotactic protein
NPX Normalized protein expression
SCF Stem cell factor
VFR Visceral fat rating

1Department of Clinical Science and Education, Södersjukhuset, Karolinska Institutet, Sjukhusbacken 10, 118 83 Stockholm, Sweden. 2Sachs’ Children and Youth Hospital, Södersjukhuset, Stockholm, Sweden. 3Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden. 4Centre for Occupational and Environmental Medicine, Region Stockholm, Stockholm, Sweden. 5Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden. 6Department of Immunology, Genetics and Pathology, Science for Life Laboratory, Uppsala University, Uppsala, Sweden. *email: susanna.klevebro@ki.se
Obesity has been demonstrated to increase the risk of cardiovascular, metabolic and pulmonary disease\textsuperscript{1, 2}, and chronic inflammation is believed to drive disease development\textsuperscript{3, 4}. Adipose tissue is a complex and highly active metabolic endocrine organ. A variety of immune cells infiltrate and become resident in adipose tissue\textsuperscript{5}, where they, along with adipocytes, secrete inflammatory factors\textsuperscript{6}. Adipose tissue expansion induces an innate and adaptive immune response, and affect glucose metabolism and inflammation\textsuperscript{7}. Visceral adipose tissue is wrapped around major abdominal organs and is an independent risk factor for cardiovascular and metabolic disease\textsuperscript{8}. Visceral and subcutaneous adipose tissues differ in composition of infiltrated cells, and in their production of adipose-derived secreted factors\textsuperscript{9–11}. Associations between adipose tissue and selected pro-inflammatory factors have been demonstrated in children, adolescents and adults\textsuperscript{12–14}. Weight change and body mass index (BMI) have also been associated with several inflammation-related proteins in studies utilizing proteomic methods in cohorts of overweight and obese participants\textsuperscript{15–17}.

Fat deposition differs between females and males. At comparable BMI, females have a higher percentage body fat while males have more lean mass. The fat more likely accumulate around hips and thighs in females and around the trunk and abdomen in males\textsuperscript{18}. Animal models indicate that sex hormones, for example the estrogen to androgen ratio, influence adipose tissue deposition\textsuperscript{19}. Sex differences in CRP levels have been correlated to differences in visceral- and subcutaneous adipose tissue\textsuperscript{20}. Further, there are immunological differences between the sexes, as females mount stronger acute inflammatory responses to infectious agents and vaccines, but are also more vulnerable to chronic inflammatory conditions\textsuperscript{21}. How adiposity measurements associate with a large panel of inflammation-related biomarkers in a young population including both normal and overweight subjects has not been previously studied.

The primary aim of this study was to demonstrate associations between adiposity measurements and levels of inflammation-related plasma proteins in a population of normal- and overweight/obese young adults. The secondary aim was to assess interaction between body fat and sex regarding association with inflammation-related protein levels.

Results

Characteristics of the study population. In total, 2074 subjects (1147 females and 927 males) were included in the final study population (Fig. 1). In comparison with subjects from the original population-based cohort, the final study population showed a higher proportion of females (Supplementary Table S1). As expected, males and females differed in their anthropometric measurements (Table 1). Males had higher BMI but lower body fat % and reported a higher level of physical activity compared to females. More females were smokers and more males used e-cigarettes and snuff. Protein levels differed between the sexes for 54 of the 71 proteins (i.e. nominal p-value < 0.05 for association). For most of the proteins, the association between sex and protein level
was not related to differences in body composition, as demonstrated by including %BF, BMI and VFR as covariates in separate regression models (Supplementary Table S2). Median values were lower in females compared to males for 47 of the proteins. Mean, standard deviation, median and 25th–75th percentile of NPX values are demonstrated in Supplementary Table S2.

Adiposity measurements and inflammation-related proteins. All adiposity measurements in this study were associated with the level of most of the inflammation-related plasma proteins in the Olink panel. Figure 2 demonstrates positive and negative associations with %BF, BMI, VFR, and waist circumference based on the results from regression analyses adjusted for sex, smoking, e-cigarette use, snuff use, and age at follow-up (complete results presented in Supplementary Table S3). Of the 71 proteins, 54 were associated with all four adiposity measurements and most associations were in a positive direction of effect. In our study population, association with %BF was apparent for 58 of the proteins at an FDR of 5%.

Effect modification by sex on associations between %BF and inflammation-related proteins. Three of the 58 proteins associated with %BF demonstrated effect modification by sex at an FDR of 5% (Table 2). Glial cell line-derived neurotrophic factor (GDNF) was negatively associated with %BF in females but not in males, whereas Stem Cell Factor (SCF) had a strong negative association with %BF in males but not in females. These examples are illustrated in scatterplots of raw data with regression lines of the association in females and males respectively in Fig. 3. Interleukin-18 receptor 1 (IL18R1) had a stronger positive association with %BF in males compared to females. The effect modification was not related to differences in BMI between the sexes. All six proteins shown in Table 2 had a p-value for the interaction term < 0.05 also in a model with BMI included as a covariate (data not shown).

Effect modification by BMI on associations between %BF and inflammation-related proteins. To examine if the association between %BF and inflammation-related proteins differed in normal weight versus overweight/obese subjects (BMI ≥ 25) an interaction term between BMI category and %BF was introduced in the model. Seven proteins (DCPD1, FGF23, HGF, IL-6, LAPTGFβ1, MCP-1, and MCP-4) had significant effect modification of BMI category, showing a stronger or unique association with %BF in the overweight/obese group compared to the normal weight group (Table 3). For example, Interleukin-6 (IL-6) was positively associated with %BF in both groups, although the effect was larger in the overweight/obese group (Fig. 4). Due to small numbers, analyses stratified in three groups of normal weight, overweight and obese subjects resulted in large confidence intervals. It was not possible to show statistically significant differences in association with %BF between overweight and obese subjects. CDCP1, FGF23, HGF and IL-6 still had more prominent

| Variable                      | Females (n = 1147) | Males (n = 927) | p-value* |
|-------------------------------|-------------------|----------------|----------|
| Age (y)                       | 1147              | 927            | 0.320    |
| Weight (kg)                   | 1147              | 927            | 0.001    |
| Height (m)                    | 1147              | 927            | 0.001    |
| BMI (kg/m²)                   | 1147              | 927            | 0.001    |
| Body fat (%)                  | 1147              | 927            | 0.001    |
| VFR (score)                   | 1147              | 927            | 0.001    |
| Waist circ (cm)               | 1141              | 923            | < 0.001  |
| Smoking habits                | 1146              | 924            | 0.014    |
| Do not smoke                  | n = 742           | n = 645        | 64.8%    |
| Used to smoke                 | n = 140           | n = 117        | 12.2%    |
| Sometimes                     | n = 163           | n = 108        | 14.2%    |
| Every day                     | n = 101           | n = 54         | 8.8%     |
| E cigarette use               | n = 29            | n = 48         | 2.5%     |
| Snuff use                     | n = 75            | n = 207        | 6.5%     |
| PA level                      | 948               | 783            | 0.001    |
| Low                           | n = 159           | n = 102        | 16.8%    |
| Moderate                      | n = 294           | n = 181        | 31.0%    |
| High                          | n = 495           | n = 500        | 52.2%    |
| BMI category at 24            | 1147              | 927            | 0.007    |
| Underweight                   | n = 70            | n = 51         | 6.1%     |
| Normal weight                 | n = 854           | n = 637        | 74.5%    |
| Overweight                    | n = 175           | n = 191        | 15.3%    |
| Obesity                       | n = 48            | n = 48         | 4.2%     |

Table 1. Descriptive characteristics of the study population (n = 2074). *Mann–Whitney U-test or Chi².
positive associations with %BF in overweight compared to normal weight subjects and the coefficients did not indicate that the associations were driven by obesity, whereas MCP-1 had a clear positive association with %BF only in the obese group (Supplementary Table S4).

Physical activity and association between %BF and inflammation-related proteins. Of the 2074 participants, 1728 had information regarding physical activity. Including physical activity as a covariate did not alter the main results, Supplementary Table S5. Effect modification of physical activity (as a binary variable of low/moderate vs high) was examined by including an interaction term in the adjusted model of %BF. At a 5% FDR the interaction term was not significant for any of the proteins.

VFR and inflammation-related proteins. Of the 71 proteins analyzed, 55 were associated with VFR in our study population, Supplementary Table S3. Sex modified the association between VFR and protein level in 15 of the 55 proteins, Table 4. The effect was larger in females compared to males for 14 of the proteins. As with %BF, SCF demonstrated a negative association with VFR only in males. GDNF, that differed in association with
%BF between females and males, was not associated with VFR at all. The proteins that were modified by BMI category in the association with %BF did not demonstrate differences in association with VFR at a 5% FDR of the interaction term. Leukemia inhibitory factor receptor (LIFR) was negatively associated with all adiposity measurements, Fig. 2, Supplementary Table S3, and showed a significant negative association with VFR only in the normal weight BMI group, Table 5.

Discussion

In our population-based study of young adults, we show that adiposity reflects in a systemic inflammatory protein profile. Some of the proteins demonstrated a more marked positive association with %BF in overweight/obese subjects, suggestive of an accelerated release of these factors with increasing BMI, possibly promoting a chronic inflammatory state that has previously been related to increased risk of cardiometabolic disease development. For most proteins, the association between protein level and total %BF was similar between the sexes. Observed differences in protein levels between females and males may be related to biological differences but also partly to differences in adiposity. Our results show that some protein levels associated differently to adiposity depending on sex. For example, there was a more prominent positive association with VFR in females compared to males for several proteins, indicating that the relation of visceral fat and the inflammatory profile differs between females and males.

The positive association between inflammatory protein levels and %BF in this study could indicate a spill-over of a local inflammation or be the result of a feed-back loop of increasing inflammation, in the adipose tissue. The effects of increased circulating inflammatory protein levels are likely to have systemic effects and implications for health. Obesity in childhood is likely to persist into adulthood, and elevated levels of inflammatory factors in childhood and adolescence have been shown to track into adulthood. Even moderate overweight has been associated with an increased risk of cardiovascular disease. Increased risk of airway obstruction with increasing BMI has previously been demonstrated in this cohort. The same study also demonstrated effect modification by sex in some of the associations. Longitudinal, large scale analysis of plasma proteins, that have been performed in predominantly select groups of patients, have shown that inflammatory patterns are influenced by weight loss and weight maintenance, indicating potential benefit of a weight loss intervention. Similar patterns related to body fat and distribution have also been demonstrated in a population based study of elderly. Physical activity has been shown to influence weight and inflammatory biomarkers. In our cross-sectional study however, we found no significant interaction between %BF and physical activity in the association with protein levels. Dietary interventions might also be of importance and for example plant based diets have been associated with reduced levels of inflammatory biomarkers such as IL-6.

IL-6 is one of the major pro-inflammatory factors released from adipose tissue and higher levels have also previously been associated with obesity as well as diseases like diabetes and asthma. However, IL-6 plays a highly complex role in metabolic regulation and can be secreted from adipocytes, adipose tissue macrophages and other adipose cell types. Adipocyte-derived IL-6 was shown to accumulate adipose tissue macrophages without influencing glucose or insulin tolerance, while myeloid-cell derived IL-6 suppressed the polarization of M1 macrophages and improved tolerance. In the current study, IL-6 was positively associated with %BF in both females and males and accelerated in overweight/obese subjects. The positive association of IL-6 and %BF did not differ significantly between females and males; however, IL-6 showed a stronger association with VFR in females. Several other proteins also demonstrated stronger positive association with VFR in females compared to males. Females have a higher %BF and a lower visceral fat mass compared to males and sex hormones are involved in fat mass regulation and distribution. A previous study showed a larger effect of visceral fat on the risk of cardiovascular diseases and type 2 diabetes in females compared to males. The results from our study support the theory that the association between visceral fat and inflammation may differ depending on sex.

Obesity-related risk of metabolic disease is affected by age, sex, total body fat content, and body fat distribution. Especially an excess amount of visceral fat associates with metabolic syndrome and cardiovascular disease. Abundance and function of adipocytes and macrophages differ depending on fat mass localization. Macrophages are the most abundant immune cell in adipose tissue and can constitute 50% of immune cells in obese conditions, to be compared with 10% in lean adipose tissue. MCPs (Monocyte

---

Table 2. Association between %BF and protein levels in females and males for proteins with effect modification by sex. Stratified analyses in men and women from linear regression of transformed protein levels adjusted for smoking, e-cigarette use, snuff use, and age at follow-up. The table includes all proteins with unadjusted p-value for the interaction term %BF×Sex of < 0.05. p-values at 0.05 FDR. *p-value for the interaction term without FDR correction.

| Protein | Females Coef | 95% CI | p-value | Males Coef | 95% CI | p-value | p-value interact | p-value interact* |
|---------|--------------|--------|---------|------------|--------|---------|-----------------|-----------------|
| CD2C1F  | 0.43         | 0.33 to 0.53 | < 0.001 | 0.58       | 0.48 to 0.67 | < 0.001 | 0.264 | 0.027 |
| GD1F    | −0.21        | −0.31 to −0.11 | < 0.001 | 0.00       | −0.08 to 0.08 | 0.947 | 0.047 | 0.001 |
| IL10    | 0.05         | 0.04 to 0.15 | 0.240   | 0.20       | 0.10 to 0.30 | < 0.001 | 0.264 | 0.024 |
| IL1R1   | 0.41         | 0.31 to 0.51 | < 0.001 | 0.60       | 0.51 to 0.69 | < 0.001 | 0.048 | 0.002 |
| IL6     | 0.67         | 0.59 to 0.74 | < 0.001 | 0.52       | 0.43 to 0.61 | < 0.001 | 0.264 | 0.025 |
| SCF     | −0.09        | −0.19 to 0.00 | 0.064   | −0.31      | −0.41 to −0.21 | < 0.001 | 0.048 | 0.002 |
chemotactic proteins) are key factors in the regulation of monocyte/macrophage migration and infiltration. Higher levels of MCP have been associated with obesity and adipose tissue localization\(^{10,39,40}\). MCP-1 and MCP-4 showed significant positive correlation with %BF only in overweight/obese subjects in our study. Further, MCP-1 was positively associated with VFR in this group.

Several proteins demonstrated significant association with %BF in the normal weight population, with similar effect estimates in overweight/obese subjects. These proteins might be less relevant as indicators of adiposity-related inflammation and might reflect a general increase in body size. A few proteins were negatively associated with %BF in this study. SCF was negatively associated with %BF in males, and GDNF in females. SCF promotes brown adipocyte differentiation, contributes to mitochondrial function and energy expenditure\(^{41,42}\) and low levels of SCF has been associated with increased incidence of cardiovascular events\(^{43}\). The browning capacity

---

**Figure 3.** Examples of effect modification by sex. GDNF (A) and SCF (B) and association with %BF in females and males. Scatterplot and regression line of protein levels (in NPX) and %BF in females and males respectively.
of adipose tissue and differentiation of precursor cells to beige adipocytes have been associated with metabolic conditions. GDNF is involved in neuron survival and regeneration and has been described in several inflammatory conditions. Studies in rodents have demonstrated a protective effect against obesity of GDNF.

All adiposity measurement had a negative association with LIFR. LIF, one of the ligands for LIFR, influence adipocyte differentiation. Hence, we speculate that the LIF-LIFR signaling might be impaired in obesity. Indeed, a negative association between LIFR and VFR was not observed in the overweight/obese group.

A limitation to this study was that the plasma protein levels we have measured reflect the combined proteome from many cell types and tissues. We are not able to trace the origin of the proteins and differentiate proteins secreted by macrophages, adipocytes, or other cells. Also, tissue levels and tissue effects are unknown.

Table 3. Association between %BF and protein levels stratified by BMI categories. Stratified analyses in BMI categories from linear regression of transformed protein levels adjusted for sex, smoking, e-cigarette use, snuff use, and age at follow-up. The table includes all proteins with unadjusted p-value for the interaction term %BF*BMI_category of < 0.05. p-values at 0.05 FDR. *p-value for the interaction term without FDR correction.

| Protein  | Coef | 95% CI  | p-value | Coef | 95% CI  | p-value | p-value interact | p-value interact* |
|----------|------|---------|---------|------|---------|---------|----------------|------------------|
| CCL3     | 0.34 | 0.23 to 0.46 | <0.001  | 0.60 | 0.43 to 0.77 | <0.001  | 0.159  | 0.041            |
| CCL20    | 0.09 | −0.03 to 0.20 | 0.232  | 0.37 | 0.19 to 0.55 | <0.001  | 0.120  | 0.023            |
| CDCP1    | 0.27 | 0.16 to 0.38 | <0.001  | 0.88 | 0.71 to 1.05 | <0.001  | 0.005  | <0.001           |
| DNER     | −0.19 | −0.31 to −0.08 | 0.007  | −0.30 | −0.47 to −0.12 | 0.002  | 0.161  | 0.044            |
| FGF21    | 0.30 | 0.19 to 0.42 | <0.001  | 0.74 | 0.58 to 0.91 | <0.001  | 0.111  | 0.019            |
| FGF23    | 0.07 | −0.05 to 0.19 | 0.343  | 0.56 | 0.39 to 0.72 | <0.001  | 0.030  | 0.003            |
| HGF      | 0.12 | 0.01 to 0.23 | 0.082  | 0.72 | 0.57 to 0.87 | <0.001  | <0.001 | <0.001           |
| IL6      | 0.50 | 0.39 to 0.61 | <0.001  | 0.86 | 0.72 to 1.00 | <0.001  | <0.001 | <0.001           |
| IL18     | 0.14 | 0.02 to 0.26 | 0.057  | 0.46 | 0.30 to 0.63 | <0.001  | 0.157  | 0.035            |
| LAPTGFbeta1 | −0.03 | −0.14 to 0.09 | 0.733  | 0.33 | 0.15 to 0.50 | 0.001  | 0.032  | 0.004            |
| MCP1     | −0.10 | −0.21 to 0.01 | 0.153  | 0.36 | 0.20 to 0.53 | <0.001  | 0.030  | 0.003            |
| MCP4     | −0.08 | −0.20 to 0.04 | 0.255  | 0.27 | 0.11 to 0.44 | 0.003  | 0.005  | <0.001           |
| OSM      | 0.02 | −0.10 to 0.13 | 0.843  | 0.35 | 0.18 to 0.52 | <0.001  | 0.111  | 0.018            |
| TNFSF14  | 0.15 | 0.04 to 0.26 | 0.034  | 0.38 | 0.21 to 0.54 | <0.001  | 0.121  | 0.025            |

Figure 4. Example of effect modification by BMI category. IL-6 and association with %BF in normal weight and overweight/obese subjects. Scatterplot and regression line of protein level (in NPX) and %BF in normal weight and overweight/obese BMI groups respectively.
factors measured in this study are “inflammation-related” in the sense that they may be up- or downregulated during infection/inflammation, still their presence may not be a sign of chronic inflammation. The participants in this study were clinically healthy at the time of the study visit but we were not able to objectively rule out presence of inflammatory conditions. Another limitation is that protein expression was measured only on one occasion, limiting the inference of dynamic relationships between body composition and systemic inflammation.

There are several methods that measure adiposity and body composition. In this study we used a combination of bioimpedance and anthropometric measurements. Anthropometric measurements are easy to use in large cohort studies. Bioimpedance gives additional objective information regarding actual adiposity and fat mass location. Bioimpedance is a non-invasive, relatively cheap method and measurements of %BF correlates well with dual-energy X-ray absorptiometry. The measurement of visceral fat by bioimpedance is not as accurate as measurement by MRI or DT, which are not feasible methods in cohorts of this size.

Our results show that adiposity is associated with the levels of inflammation-related markers in a young adult population with a normal distribution of BMI. Overweight/obesity strongly correlate with the levels of specific inflammatory markers, including IL-6. We also demonstrate that sex and adiposity localization influence these associations. The results highlight differences of importance when using inflammation-related plasma proteins.
as biomarkers associated with adiposity. Our study show that adiposity-driven inflammation can be observed in young adults before potential development of obesity-related diseases. The findings might have implications for targeted interventions aiming to reduce the inflammatory load in early adulthood.

**Methods**

**Study design and study population.** The study population was based on participants in the BAMSE (Swedish abbreviation for Child (Barn), Allergy, Milieu, Stockholm, Epidemiological) cohort, a Swedish population-based cohort of 4089 children born in Stockholm 1994–1996. The children have been followed through repeated administration of questionnaires and have been invited to undergo clinical examinations at ages 4, 8, 16 and 24 years. Participants of the clinical examination at the 24-year follow-up, who had complete data regarding biomarkers and bioimpedance measurements, were included in the present study. Pregnancy was the only exclusion criterion. In total, 2270 subjects participated in the clinical examination and 2074 subjects with a median age of 22.5 years (range 20.9–25.2 years) were included in the final study population (Fig. 1).

**Clinical investigation.** Venous blood was collected in EDTA tubes (BD Vacutainer®) at the 24-year follow-up. Fasting prior to sampling was not required. Participants were asked to re-schedule the follow-up visit if not feeling well but no test to evaluate presence of acute inflammation was performed. Plasma was obtained by centrifugation, aliquoted and stored at −80 °C until analyzed. Height was measured twice to the nearest 0.5 cm using a wall-mounted stadiometer, and the mean value was used for analyses. Waist circumference was measured at the end of an expiration below elbow level. Weight and bioimpedance measurements were taken using Tanita MC 780 body composition monitor according to instructions from the manufacturer. In the present study, we included body mass index (BMI), percentage body fat (%BF) and visceral fat rating (VFR) measurements. The measurement of visceral fat was expressed as a rating from 1 to 60 developed by the manufacturing company. BMI was used as a continuous variable in analyses of association with protein level. In stratified analyses, BMI was categorized in two categories, normal weight (18.5 to < 25 kg/m²) and overweight including obesity (≥ 25 kg/m²). Level of physical activity was defined based on time spent on moderate and vigorous intensity activities reported in the 24-year questionnaire. The answers were categorized according to IPAQ as high (≥ 7 h per week of moderate to vigorous activity or ≥ 3.5 h per week of vigorous activity), moderate (≥ 2.5 h per week of moderate to vigorous activity) or low (< 2.5 h per week of moderate to vigorous activity) level of physical activity. Smoking was categorized into daily smoking, occasional smoking, and no current smoking.

**Proseek multiplex inflammation panel.** The expression of 92 inflammation-related protein biomarkers in plasma were analyzed by the Proseek Multiplex Inflammation Panel (version 95302) from Olink Biosciences, Uppsala, Sweden. Assay characteristics and validations are available from the manufacturer’s webpage (https://www.olink.com/resources-support/document-download-center/). In brief, antibodies labelled with complementary oligonucleotide sequences were allowed to bind pairwise to the target protein. Upon DNA-polymerization, the paired oligonucleotides form a reporter sequence that was amplified by qRT-PCR. Data are expressed as Normalized Protein Expression (NPX) units on a log2 scale calculated from normalized Ct values. Samples that deviated more than 0.3 NPX from the median value of an internal control were excluded. The lower limit of detection (LOD) was defined as three standard deviations above background. 71 proteins with >75% of samples above LOD were included in the analyses and, in accordance with recommendations by the company, values below LOD were not replaced by arbitrary values. The full names of the proteins are given in Supplementary Table S6.

**Statistical methods.** All statistical analyses were performed using Stata version 16 (StataCorp LP, College Station, TX, USA). Circos plots were constructed using the circclize package in R version 3.6.1 (R Foundation for Statistical Computing, Vienna, Austria). Median, 25th, and 75th percentiles are presented for continuous variables, number and percentage for categorical variables, and comparison between groups were tested using Mann–Whitney U-test or Chi-2. Linear regression with robust standard errors was used to investigate associations between adiposity measurements and protein levels. Protein values were standardized using rank-based inverse normal transformation. Significance was based on a false discovery rate (FDR) of 5% using the Benjamini–Hochberg procedure. Based on principal component regression analysis, covariates considered as potential confounders included sex, smoking, e-cigarette use, snuff use, age at follow-up, and level of physical activity. Information regarding physical activity was not available from all study subjects and therefore not included in the main regression model. A sensitivity analysis of the primary outcome that included the level of physical activity was performed. Potential effect modifications by sex as well as BMI category were examined by introducing an interaction term in the regression model. Effect modification was considered significant based on a 5% FDR. Stratified results are presented for all nominally significant associations.

**Ethics statement.** The study was conducted in accordance with the Declaration of Helsinki and approved by the Regional Ethics Committee in Stockholm (DNR 2016/1380-31/2). All participants in this study were over the age of 18 years and provided written informed consent. At previous follow-ups of the BAMSE study, informed consent has also been collected from a parent or legal guardian.
References

1. Bhaskaran, K., Doo-Santos-Silva, I., Leon, D. A., Douglas, I. J. & Smeeth, L. Association of BMI with overall and cause-specific mortality: A population-based cohort study of 3.6 million adults in the UK. *Lancet Diabetes Endocrinol.* 6, 944–953 (2018).

2. Gurka, M. I., Filipp, S. L., Musani, S. K., Sims, M. & DeBoer, M. D. Use of BMI as the marker of adiposity in a metabolic syndrome severity score: Derivation and validation in predicting long-term disease outcomes. *Metabolism* 83, 68–74 (2018).

3. Aday, A. W. & Ridker, P. M. Targeting residual inflammatory risk: A shifting paradigm for atherosclerotic disease. *Front. Cardiovasc. Med.* 6, 16 (2019).

4. Reinheir, T. & Roth, C. L. Inflammation markers in type 2 diabetes and the metabolic syndrome in the pediatric population. *Curr. Diabetes Rep.* 18, 131 (2018).

5. Ferrante, A. W. The immune cells in adipose tissue. *Diabetes Obes. Metab.* 15(Suppl 3), 34–38 (2013).

6. Ouchi, N., Parker, J. L., Lugus, J. J. & Walsh, K. Adipokines in inflammation and metabolic disease. *Nat. Rev. Immunol.* 11, 85–97 (2011).

7. McLaughlin, T., Ackerman, S. E., Shen, L. & Engleman, E. Role of innate and adaptive immunity in obesity-associated metabolic disease. *J. Clin. Investig.* 127, 5–13 (2017).

8. Neeland, I. J. et al. Visceral and ectopic fat, atherosclerosis, and cardiometabolic disease: A position statement. *Lancet Diabetes Endocrinol.* 7, 715–725 (2019).

9. Fried, S. K., Bunkin, D. A. & Greenberg, A. S. Omertal and subcutaneous adipose tissues of obese subjects release interleukin-6: Depot difference and regulation by glucocorticoid. *J. Clin. Endocrinol. Metab.* 83, 847–850 (1998).

10. Bruun, J. M., Lihn, A. S., Pedersen, S. B. & Richelsen, B. Monocyte chemoattractant protein-1 release is higher in visceral than subcutaneous human adipose tissue (AT): Implication of macrophages residing in the AT. *J. Clin. Endocrinol. Metab.* 90, 2282–2289 (2005).

11. Michaud, A., Drolet, R., Noél, S., Paris, G. & Tchernof, A. Visceral fat accumulation is an indicator of adipose tissue macrophage infiltration in women. *Metabolism* 61, 689–698 (2012).

12. Singer, K., Eng, D. S., Lumeng, C. N., Gebremariam, A. & Lee, M. J. The relationship between body fat mass percentiles and inflammatory markers before and during weight loss: Results from randomized trial of dietary intervention. *Sci. Rep.* 10, 7913 (2020).

13. Fried, S. K., Lee, M.-J. & Karastergiou, K. Shaping fat distribution: New insights into the molecular determinants of depot- and sex-dependent adipose biology. *Obes. Silver Spring Md.* 23, 1345–1352 (2015).

14. Chang, E., Varghese, M. & Singer, K. Gender and sex differences in adipose tissue. *Curr. Diabetes Rep.* 18, 69 (2018).

15. Cartier, A. et al. Sex differences in inflammatory markers: What is the contribution of visceral adiposity?. *Am. J. Clin. Nutr.* 89, 1307–1314 (2009).

16. Klein, S. L. & Flanagan, K. L. Sex differences in immune responses. *Nat. Rev. Immunol.* 16, 626–638 (2016).

17. Singh, A. S., Mulder, C., Twisk, J. W. R., van Mechelen, W. & Chinapaw, M. J. M. Tracking of childhood overweight into adulthood: A systematic review of the literature. *Obes. Rev. Off. J. Int. Assoc. Study Obes.* 9, 474–488 (2008).

18. Juonala, M. et al. Childhood C-reactive protein in predicting CRP and carotid intima-media thickness in adulthood: The Cardiovascular Risk in Young Finns Study. *Artheroscler. Thromb. Vasc. Biol.* 26, 1883–1888 (2006).

19. Bogers, R. P. et al. Association of overweight with increased risk of coronary heart disease partly independent of blood pressure and cholesterol levels: A meta-analysis of 21 cohort studies including more than 300,000 persons. *Arch. Intern. Med.* 167, 1720–1728 (2007).

20. Ekstroim, S. et al. Body mass index status and peripheral airway obstruction in school-age children: A population-based cohort study. *Thorax* 73, 538–545 (2018).

21. Lind, L. et al. Changes in proteomic profiles are related to changes in BMI and fat distribution during 10 years of aging. *Obes. Silver Spring Md.* 28, 178–186 (2020).

22. Liberman, K., Forti, L. N., Beyer, I. & Bautmans, I. The effects of exercise on muscle strength, body composition, physical functioning and the inflammatory profile of older adults: A systematic review. *Curr. Opin. Clin. Nutr. Metab. Care* 20, 30–53 (2017).

23. Eichelmann, F., Schwinghaschlk, L., Fedirko, V. & Aleksandrova, K. Effect of plant-based diets on obesity-related inflammatory profiles: A systematic review and meta-analysis of intervention trials. *Obes. Rev. Off. J. Int. Assoc. Study Obes.* 17, 1067–1079 (2016).

24. Rodrigues, K. E. et al. IL-6, TNF-a, and IL-10 levels/polymerisms and their association with type 2 diabetes mellitus and obesity in Brazilian individuals. *Arch. Endocrinol. Metab.* 61, 438–446 (2017).

25. Chen, Y. et al. Correlation between serum interleukin-6 level and type 1 diabetes mellitus: A systematic review and meta-analysis. *Cytokine* 94, 14–20 (2017).

26. Peters, M. C. et al. Plasma interleukin-6 concentrations, metabolic dysfunction, and asthma severity: A cross-sectional analysis of two cohorts. *Lancet Respir. Med.* 4, 574–584 (2016).

27. Mauer, J., Denson, J. L. & Brüning, J. C. Versatile functions for IL-6 in metabolism and cancer. *Trends Immunol.* 36, 92–101 (2015).

28. Han, M. S. et al. Regulation of adipose tissue inflammation by interleukin 6. *Proc. Natl. Acad. Sci. U. S. A.* 117, 2751–2760 (2020).

29. Stefan, N. Causes, consequences, and treatment of metabolically unhealthy fat distribution. *Lancet Diabetes Endocrinol.* 8, 616–627 (2020).

30. Karlsson, T. et al. Contribution of genetics to visceral adiposity and its relation to cardiovascular and metabolic disease. *Nat. Med.* 25, 1390–1395 (2019).

31. Iacobini, C., Pugliese, G., Blasetti Fantuzzi, C., Federici, M. & Menini, S. Metabolically healthy versus metabolically unhealthy obesity. *Metabolism* 92, 51–60 (2019).

32. Sato, F. et al. Association of epicardial, visceral, and subcutaneous fat with cardiometabolic diseases. *Circ. J. Off. Jpn. Circ. Soc.* 82, 502–508 (2018).

33. Lu, J., Zhao, J., Meng, H. & Zhang, X. Adipose tissue-resident immune cells in obesity and type 2 diabetes. *Front. Immunol.* 10, 1713 (2019).

34. Sartipy, P. & Loskutoff, D. J. Monocyte chemoattractant protein 1 in obesity and insulin resistance. *Proc. Natl. Acad. Sci. U. S. A.* 100, 7265–7270 (2003).

35. Kim, C.-S. et al. Circulating levels of MCP-1 and IL-8 are elevated in human obese subjects and associated with obesity-related parameters. *Int. J. Obes.* 2005(30), 1347–1355 (2006).
Acknowledgements
We thank the children and parents participating in the BAMSE cohort and all staff involved in the study through the years.

Author contributions
S.K.: Conceptualization, methodology, formal analysis, writing—original draft. S.B.: Conceptualization, methodology, writing—original draft. S.E.: Validation, data curation, project administration, writing—original draft. S.K.M.: Formal analysis, visualization, writing—review & editing. O.G.: Investigation, writing—review & editing. A.M.: Methodology, writing—review & editing. Å.J.: Methodology, writing—review & editing. I.K.: Investigation, project administration, writing—review & editing. A.B.: Investigation, project administration, writing—review & editing. E.M.: Conceptualization, methodology, supervision, funding acquisition, writing—review & editing.

Funding
Open access funding provided by Karolinska Institute. This study was supported by grants from the Swedish Research Council, the Swedish Research Council for Health, Working Life and Welfare, Formas, the Swedish Heart–Lung Foundation, the European Research Council (TRIBAL, Grant agreement 757919), the Swedish Asthma and Allergy Research Foundation and Region Stockholm (ALF project, and for cohort and database maintenance). S Klevebro was supported by Region Stockholm (clinical postdoctoral appointment) and the Swedish Heart–Lung Foundation.

Competing interests
The authors declare no competing interests.

Additional information
Supplementary Information The online version contains supplementary material available at https://doi.org/10.1038/s41598-021-90843-x.

Correspondence and requests for materials should be addressed to S.K.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2021