Serum markers of matrix turnover as predictors for the evolution of colorectal cancer metastasis under chemotherapy

B Hanke1,2,5, A Wein1,5, P Martus3, C Riedel1, M Voelker4, EG Hahn1 and D Schuppan*1

1Department of Medicine I, University of Erlangen-Nuernberg, Germany; 2Department of Gerontology, Humboldt University, Berlin; 3Institute of Medical Informatics, Biometry and Epidemiology, Free University, Berlin; 4Bayer AG, Leverkusen, Germany

Connective tissue turnover plays a prominent role in tumour growth and metastasis. We followed serum levels of seven connective tissue parameters in 37 patients with colorectal cancer metastatic to the liver prior to and during chemotherapy. Serum samples with episodes of tumour control (n = 112) showed an increase of matrix metalloproteinase-2 (MMP-2) (P ≤ 0.001) and a decrease of tissue inhibitor of MMPs (TIMP-1) levels (P ≤ 0.01), while serum samples with episodes of tumour progression displayed the reverse pattern (P ≤ 0.01 and P ≤ 0.05, resp.). The ratio of circulating MMP-2/TIMP-1 was also significantly higher in episodes of tumour control vs tumour progression and prior to treatment (P ≤ 0.0001). We conclude that serum MMP-2 appears to reflect tumour resorption, while serum TIMP-1 may mirror tumour expansion.

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Matrix metalloproteinases (MMPs) are proteolytic enzymes that play a central role in benign and malignant matrix remodelling (Kähäri and Saarialho-Kere, 1999; Nagase and Woessner, 1999). MMP-2 and MMP-9 (gelatinases A and B, resp.) degrade basement membrane collagen as well as denatured collagens (gelatin). MMP activity is tightly regulated (1) on the level of transcription, (2) on the level of proteolytic activation of the pro-MMPs, (3) by the tissue inhibitors of metalloproteinases (TIMP-1 and TIMP-2) and (4) by their strict compartmentalisation in cell membrane domains. Enhanced local expression and activity of MMPs and low levels of inhibitory TIMPs correlate with tumour growth and metastasis (Zeng et al, 1995, 1996; Murray et al, 1996). However, the findings on circulating MMP-2 and TIMP-1 levels in patients with metastatic colorectal tumours are unclear (Holten-Andersen et al, 2000; Pellegrini et al, 2000; Ylisirnio et al, 2000).

CEA and CA 19-9 have been previously shown to predict tumour growth in patients with liver metastasis treated by chemotherapy (Hanke et al, 2001). However, there exist no follow-up data with serum connective tissue markers to predict treatment response or nonresponse in these patients. Colorectal cancer metastatic to the liver is particularly suited to validate novel markers, since the condition is characterised by a large and defined tumour burden.

MATERIALS AND METHODS

We studied 37 consecutive patients (29 men, eight women, age 42–76 years, mean 62 years) with colorectal carcinoma metastatic to the liver. Primary carcinoma was confirmed histologically. Histological confirmation was also obtained for synchronous liver metastasis. In case of metachronous liver metastasis, histological confirmation was only pursued when imaging techniques (spiral computerised tomography (CT) of the abdomen or MRT of the liver) did not show clear results. Patients received first-line chemotherapy, consisting of a weekly 1–2 h infusion of folinic acid (500 mg m⁻²) followed by a 24-h infusion of 5-fluorouracil (2600 mg m⁻²). One cycle comprised six weekly infusions followed by 2 weeks of rest. A total of 16 patients received additional biweekly oxaliplatin (85 mg m⁻²) and three patients additional weekly irinotecan (80 mg m⁻²) (Wein et al, 2001, 2003). Treatment response was monitored every 8 weeks by spiral CT and antitumour activity was evaluated in accordance with WHO criteria. Median treatment duration was 7 months. Patients were stratified as ‘prior to treatment’, ‘tumour control’ (complete/partial remission or stable disease) and ‘tumour progression’. In 23 of 37 patients, first serum was obtained immediately prior to treatment. In all 37 patients, we recorded 112 episodes of tumour control, and in 16 of 37 patients we recorded episodes of tumour progression. The patients had been treated within the framework of two clinical phase II studies after obtaining approval by the local ethics committee (Wein et al, 2001, 2003).

Sera were obtained at 8 weekly intervals, along with CT, stored frozen at −80°C and analysed for six circulating connective tissue markers in a single run. Serum collagen IV and VI, tenascin-C, MMP-2, the MMP-9/TIMP-1 complex and free TIMP-1 were measured on the Bayer Immuno 1 Analyzer using fluorescein- and alkaline-phosphatase-labelled monoclonal antibodies to the target antigen (Schuppan et al, 1995; Ropers et al, 2000). Immune

*Correspondence: Professor Dr D Schuppan, Department of Medicine I, University of Erlangen-Nuernberg, Ulmenweg 18, 91054 Erlangen, Germany; E-mail: detlef.schuppan@med1.imed.uni_erlangen.de

5First authorship is equally shared between B Hanke and A Wein.

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complexes were separated with magnetic particles coated with a monoclonal antifluorescein antibody and quantified after substrate addition. Analysis was restricted to MMP-2 and TIMP-1 (representing both the free and the bound protein), since only these markers correlated well with tumour evolution. Normal values for 100 healthy adults were 641 ng ml\(^{-1}\) (s.d. 5.5%) for MMP-2 and 620 ng ml\(^{-1}\) (s.d. 4.9%) for TIMP-1.

For descriptive analysis all sera referring to ‘prior to treatment’, ‘tumour control’ and ‘tumour progression’ were grouped together. For confirmatory analysis, values were averaged for each patient to yield only one value per category. With only four of the patients who contributed values for initial tumour status showing progression, confirmatory analysis was restricted to the comparison of ‘prior to treatment vs tumour control’ and ‘progression vs tumour control’. Comparisons were performed using the Wilcoxon test for paired samples. \(P\)-values were multiplied by two according to Bonferroni’s correction for multiple testing. The level of significance was 0.05 (two-sided). All analyses were performed using SPSSWIN 9.0.

RESULTS

A total of 37 patients received systemic chemotherapy during an average treatment duration of 7 months. Prior to treatment, serum TIMP-1 was elevated (>2 s.d. above the normal mean) in 57% (13 out of 23), decreasing to 36% (40 out of 112) in episodes with tumour control and increasing to 81% (13 out of 16) in episodes with progression \((P<0.01\) and \(P<0.05\), resp.). Serum MMP-2 was elevated in 9% (two out of 23) prior to treatment, increasing to 35% (40 out of 112) in tumour control and decreasing to 12% (two out of 16) with progression (both differences \(P<0.01\), see Table 1). The ratio MMP-2/TIMP-1 differed significantly between episodes and ranged from 0.73 ‘prior to treatment’, increasing to 1.12 under tumour control and decreasing to 0.71 upon tumour progression \((P\leq0.001)\) (Figure 1).

DISCUSSION

This is the first report of the course of serum connective tissue markers in patients undergoing chemotherapy for colorectal carcinoma metastatic to the liver. Among the six tested markers, only MMP-2 and TIMP-1 correlated with the response to chemotherapy. Changes in elimination do not explain the differences in MMP and TIMP-1 serum levels, since during chemotherapy both serum creatinine and parameters of cholestasis remained unaltered.

| MMP-2 (ng ml\(^{-1}\)) | TIMP-1 (ng ml\(^{-1}\)) | MMP-2/TIMP-1 |
|------------------------|------------------------|--------------|
| **Mean** | **Median** | **s.d.** | **Range** | **Mean** | **Median** | **s.d.** | **Range** | **Mean** | **Median** | **s.d.** | **Range** |
| Prechemotherapy | 670** | 599 | 225 | 442–1439 | 956** | 934 | 230 | 605–1318 | 0.73*** | 0.73 | 0.25 | 0.35–1.30 |
| Tumour control | 861 | 827 | 199 | 492–1396 | 808 | 752 | 228 | 454–1556 | 1.12 | 1.09 | 0.32 | 0.46–1.90 |
| Tumour progress | 744** | 779 | 178 | 534–1130 | 1155* | 1062 | 395 | 665–1949 | 0.71*** | 0.62 | 0.31 | 0.29–1.42 |

Levels of significance compared with tumour control: *\(P\leq0.05\), **\(P\leq0.01\), ***\(P\leq0.001\). Normal values: MMP-2 620 ng ml\(^{-1}\) (s.d. 4.9%), TIMP-1 641 ng ml\(^{-1}\) (s.d. 5.5%).

**Figure 1** Ratio of circulating serum MMP-2/TIMP-1 in patients with metastatic colorectal cancer prior to and during chemotherapy. Circles correspond to extreme values in the statistical analysis. Total numbers of episodes = \(N\) (box plot analysis). Levels of significance compared with tumour control: *\(P\leq0.05\), **\(P\leq0.01\), ***\(P\leq0.001\).
The increase of MMP-2 and the decrease of TIMP-1 under tumour control is unexpected, since a common assumption is that enhanced activity of MMP-2, the classical basement membrane collagenase, leads to tumour growth and metastasis, while TIMP-1 as a crucial inhibitor of many MMPs should rather curtail tumour expansion (Nagase and Woessner, 1999). However, it remains to be proven if and how far tumour growth, which is accompanied by a strictly local control of proteolysis, leads to an excessive release of the involved enzymes or inhibitors (MMPs and TIMPs). Alternatively, an enhanced release of MMP-2, an enzyme which is sequestered on the cell surface and on collagens in the matrix, can be expected when the tumour volume and the associated desmoplastic stroma are reduced under effective chemotherapy. In addition, with enhanced resorption of tumour stroma, free TIMP-1 may be consumed by the various MMPs implicated in this process. The latter interpretation is in line with two cross-sectional studies in patients with advanced pulmonary and colorectal cancer, which showed elevated serum levels in tumours that progressed rapidly (Holten-Andersen et al, 2000; Pellegrini et al, 2000).

Therefore, therapies of advanced tumours that are based on unselective MMP inhibitors as a supplement to chemotherapy may be of little use, if not counterproductive. This assumption is supported by the disappointing outcome of several recent clinical studies using such agents (Bramhall et al, 2001; Coussens et al, 2002; Shepherd et al, 2002).

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REFERENCES

Bramhall SR, Rosemurgy A, Brown PD, Bowry C, Buckels JAC for the marimastat pancreatic cancer study group (2001) Marimastat as first-line therapy for patients with unresectable pancreatic cancer: a randomized trial. J Clin Oncol 19: 3447 – 3455

Coussens LM, Fingleton B, Matrisian LM (2002) Matrix metalloproteinase inhibitors and cancer: trials and tribulations. Science 295: 2387 – 2392

Hanke B, Riedel C, Lampert S, Happich K, Martus P, Parsch H, Himmler B, Hohenberger W, Hahn EG, Wein A (2001) CEA and CA 19-9 measurement as a monitoring parameter in metastatic colorectal cancer (CRC) under palliative first-line chemotherapy with weekly 24-hour infusion of high-dose 5-fluorouracil (5-FU) and folinic acid (FA). Ann Oncol 12: 221 – 226

Holten-Andersen MN, Stephens RW, Nielsen HJ, Murphy G, Christensen IJ, Stetler-Stevenson W, Brunner N (2000) High preoperative plasma tissue inhibitor of metalloproteinase-1 levels are associated with short survival of patients with colorectal cancer. Clin Cancer Res 6: 4292 – 4299

Kähäri VM, Saarialho-Kere U (1999) Matrix metalloproteinases and their inhibitors in tumour growth and invasion. Ann Med 31: 34 – 45

Murray GI, Duncan ME, O’Neill P, Melvin WT, Fothergill JE (1996) Matrix metalloproteinase-1 is associated with poor prognosis in colorectal cancer. J Pathol 174: 241 – 246

Nagase H, Woessner JF Jr (1999) Matrix metalloproteinases. J Biol Chem 274: 21491 – 21494

Pellegrini P, Contasta I, Berghella AM, Gargano E, Mammarella C, Adorno D (2000) Simultaneous measurement of soluble carcinoembryonic antigen and the tissue inhibitor of metalloproteinase TIMP-1 serum levels for use as markers of pre-invasive to invasive colorectal cancer. Cancer Immunol Immunother 48: 388 – 394

Ropers T, Kroll W, Becka M, Voelker M, Burchardt ER, Schuppen D, Gehrmann M (2000) Enzyme immunoassay for the measurement of human tenascin-C on the Bayer Immuno 1 analyzer. Clin Biochem 33: 7 – 13

Shepherd FA, Giaccone G, Seymour L, Debruyne C, Bezjak A, Hirsh V, Smylie M, Rubin S, Martins H, Lamont A, Krzakowski M, Sadura A, Zee B (2002) Prospective, randomized, double-blind placebo-controlled trial of marimastat after response to first-line chemotherapy in patients with small-cell lung cancer: a trial of the National Cancer Institute of Canada-Clinical Trials Group and the European Organization for Research and Treatment of Cancer. J Clin Oncol 20: 4434 – 4439

Schuppan D, Stölzel U, Oesterling C, Somasundaram R (1995) Serum assays for liver fibrosis. J Hepatol 22 (Suppl 2): 82 – 88

Wein A, Riedel C, Köckerling F, Martus P, Baum U, Brueckl WM, Reck T, Ott R, Hänsler J, Bernatik T, Becker D, Schneider T, Hohenberger W, Hahn EG (2001) Impact of surgery on survival in palliative patients with metastatic colorectal cancer after first line treatment with weekly 24-hour infusion of high-dose 5-fluorouracil and folinic acid. Ann Oncol 12: 1721 – 1727

Wein A, Riedel C, Brückl W, Merkel S, Ott R, Hanke B, Baum U, Fuchs F, Günther K, Reck T, Papadopoulos T, Hahn EG, Hohenberger W (2003) Neoadjuvant treatment with weekly high-dose 5-fluorouracil as 24-h infusion, folinic acid and oxaliplatin in patients with primary resectable liver metastases of colorectal cancer. Oncology 64: 131 – 138

Ylisirnio S, Hoyhtya M, Turpeenniemi-Hujanen T (2000) Serum matrix metalloproteinases -2, -9 and tissue inhibitors of metalloproteinases-1, -2 in lung cancer: TIMP-1 as a prognostic marker. Anticancer Res 20: 1311 – 1316

Zeng ZS, Cohen AM, Zhang ZF, Stetler-Stevenson W, Guillem JG (1995) Elevated tissue inhibitor of metalloproteinase 1 RNA in colorectal cancer stroma correlates with lymph node and distant metastases. Clin Cancer Res 1: 899 – 906

Zeng ZS, Huang Y, Cohen AM, Guillem JG (1996) Prediction of colorectal cancer relapse and survival via tissue RNA levels of matrix metalloproteinase-9. J Clin Oncol 14: 3133 – 3140