The Arabidopsis Information Resource (TAIR): improved gene annotation and new tools

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ABSTRACT

The Arabidopsis Information Resource (TAIR, http://arabidopsis.org) is a genome database for Arabidopsis thaliana, an important reference organism for many fundamental aspects of biology as well as basic and applied plant biology research. TAIR serves as a central access point for Arabidopsis data, annotates gene function and expression patterns using controlled vocabulary terms, and maintains and updates the A. thaliana genome assembly and annotation. TAIR also provides researchers with an extensive set of visualization and analysis tools. Recent developments include several new genome releases (TAIR8, TAIR9 and TAIR10) in which the A. thaliana assembly was updated, pseudogenes and transposon genes were re-annotated, and new data from proteomics and next generation transcriptome sequencing were incorporated into gene models and splice variants. Other highlights include progress on functional annotation of the genome and the release of several new tools including Textpresso for Arabidopsis which provides the capability to carry out full text searches on a large body of research literature.

INTRODUCTION

TAIR (The Arabidopsis Information Resource, http://arabidopsis.org) serves as the community database for Arabidopsis researchers and as an essential information source for the wider plant biology and model organism communities (1,2). TAIR contains genetic and genomic data for Arabidopsis thaliana, a well-studied plant that serves as a reference species for many aspects of plant biology (3–7). Arabidopsis thaliana has also served as a highly productive research organism for exploring many areas of fundamental biology including DNA repair, photobiology, protein degradation, the circadian clock, DNA methylation, RNA silencing and G-protein signaling, many of which have direct application to human health (8–11).

TAIR’s usage continues to increase with over 45 000 unique visitors per month in 2010 based on usage data gathered using Google Analytics and over 1.8 million visits in the past year, an increase of 6% over the previous year. Visits originated from around the world with Asia accounting for 36%, the Americas 31% and Europe 30%. Although registration is not required to view data at TAIR, users must register and log in to order seed and DNA stocks from the Arabidopsis Biological Resource Center (ABRC), enter comments on TAIR pages or submit data to TAIR via our online data submission tool. The number of registered TAIR users as of September 2011 has reached 22 000, with 9400 of these records added or...
modified within the past 5 years, serving as an estimate of the most active set of users.

**TAIR DATA TYPES AND SOURCES**

Data available from TAIR include *A. thaliana* and *A. lyrata* genomic sequences, gene structure and function annotation, *A. thaliana* metabolic pathways, gene expression patterns, DNA and seed stock data, genome maps, genetic and physical markers, ecotypes and natural variation data, publications, and information about the Arabidopsis research community. These data come from a variety of sources including manual curation of published literature and sequence data, computational pipelines for annotating gene structure and function, integration of data from other biological databases and resources (GenBank and ABRC/Arabidopsis Biological Resource Center) and submissions from the research community.

Manual literature curation by TAIR curators generates gene function and gene expression annotations based on experiments reported in the peer-reviewed research literature, using Gene Ontology (GO) terms for molecular function, biological process and cellular component, and Plant Ontology (PO) terms for plant anatomical structures and growth and developmental stages (12,13). Additional information extracted from research literature includes gene symbols and full names, alleles (including allele name, mutagen, inheritance, allele type and description) and germplasm information (parent line, associated alleles and phenotype).

In addition to extracting data from the research literature, TAIR uses several computational pipelines to integrate additional data. Functional annotation pipelines are used to assign GO terms to *A. thaliana* and *A. lyrata* genes based on the presence of protein domains or signal sequences. Gene structure pipelines are used to update gene features such as exons and UTRs (untranslated regions) for *A. thaliana* and add new genes based on new transcript evidence. Mapping pipelines are used to assign a genome position to sequenced objects including ESTs (expressed sequence tags) and cDNAs, T-DNA and transposon insertions, markers, SNPs, etc. Data import pipelines are used to download sequence data from GenBank including new ESTs, cDNAs and insertion mutant flanking sequences, and load ABRC data for seed and DNA stocks.

Community data submissions to TAIR include gene families, gene structures, gene function data, mutant phenotypes, protein–protein interactions, gene expression patterns, SNPs, markers, laboratory protocols, gene symbols, metabolic pathway data and links to other resources. A recent development in community data submission to TAIR is the establishment of a novel TAIR-journal collaboration program to collect gene function information directly from authors at the time of publication and the introduction of an online author submission tool to facilitate data submission.

**ARABIDOPSIS GENE FUNCTION ANNOTATION**

Since joining the GO Consortium in 2001, gene function curators at TAIR have worked to capture the available experimental gene function data from the *A. thaliana* research literature in the form of GO and PO controlled vocabulary annotations. In recent years the main focus of our in-house literature curation effort has been on the annotation of newly characterized genes. An average of 260 research articles are added to TAIR each month based on PubMed searches for ‘Arabidopsis’ in the title, abstract or keywords and ~150 of these are linked to gene names or synonyms within TAIR each month using automated methods. These computationally generated links between articles and genes are manually reviewed and confirmed by curators if correct. During this process, abstracts that discuss a newly characterized gene are flagged as high-priority articles for curation. TAIR curators read the full text of ~40 of these high-priority articles each month and make GO and PO annotations based on the experiments reported in the article as well as extracting other types of data (gene names, allele information) as described earlier.

We have also put considerable effort into encouraging community submissions, most recently through the establishment of collaborations with 10 plant science journals and the development of a new interface for community submission of annotations (Berardini et al., manuscript in preparation). Recently we have also begun to integrate external GO annotations from the UniProt Gene Ontology Annotation group at the European Bioinformatics Institute and the Reference Genome group of the GO Consortium (14). We have also integrated annotations inferred by the GO Consortium from links within the ontology. For example, gene products annotated to the molecular function term ‘sodium ion transmembrane transporter activity’ (GO:0015081) are also annotated to the biological process term ‘sodium ion transmembrane transport’ (GO:0035725) because the molecular function term is linked to the biological process term.

Each GO annotation from the sources described earlier consists of a gene identifier, a GO term, an evidence code and a reference. Although in some cases two or more annotations may contain the same gene and GO term, as long as the evidence code or reference differ these are still considered unique annotations and are retained. In other cases two separate annotations to the same gene may provide two related GO terms differing in specificity, for example ‘chloroplast’ versus ‘chloroplast inner membrane’, often because the method differed between the two experiments. In a few cases different methods or even the same method in different hands may produce a different result (e.g. location of a gene product in ‘chloroplast’ versus ‘cytoplasm’, resulting in two independent annotations, both of which are retained in order to provide a complete picture of all experimental results. We encourage all users of GO annotations to make full use of the evidence codes, evidence descriptions and links to the research article describing the experimental result that are available as part of each annotation in such cases.
To supplement the gene function information we extract manually from research literature and incorporate from the community and other resources, we use computational methods to assign GO terms based on the presence of protein domains and other conserved sequences of known function. For each genome release, we use a combination of InterProScan (15) on the proteome combined with the latest InterPro2GO mapping file (http://www.ebi.ac.uk/GOA/InterPro2GO.html) to create GO annotations for proteins based on the presence of domains with mapped GO terms. We also perform a TargetP analysis (16) with plant-specific parameters to identify proteins that are predicted to be secreted or to localize to the chloroplast or mitochondria and create appropriate GO annotations based on these results. Annotations resulting from these computational methods are loaded into TAIR using the IEA (Inferred from Electronic Annotation) evidence code only if they provide an annotation to a GO aspect (molecular function, cellular component or biological process) not yet obtained from other annotation methods for that gene (e.g. for a gene product with an experimental annotation to 'chloroplast', an IEA annotation to 'cytoplasm' would not be loaded).

**GO annotation for the *A. thaliana* genome**

To date, 20% of all *A. thaliana* genes (excluding pseudogenes and genes encoded by transposable elements) have been annotated to at least one GO term for a biological process based on an experiment done directly on the gene in question or its protein or RNA product, as shown in Table 1. A slightly higher proportion (27%) have been assigned a GO cellular component term based on direct experiment, and only 13% have an experimentally based annotation to a molecular function term. When other types of evidence are included, such as sequence similarity to a gene of known function or presence of a domain with a well-defined function, for each GO aspect over half of *A. thaliana* genes have at least one annotation (Table 1, ‘All evidence types’ column). A total of 77% of all *A. thaliana* genes have at least one GO annotation to one of the three GO aspects.

**‘Unknown’ genes annotated to GO root terms**

The goal of the GO consortium is to assign at least one biological process, molecular function and cellular component term to every gene in an organism. In cases where there is no experimental data and no predicted function based on domains or other computational methods, curators assign the root term (e.g. ‘biological process’ rather than a more specific biological process term such as ‘transcription’ or ‘leaf development’) to indicate that this aspect of the gene function is unknown. The presence of an annotation to the root term serves as a way to distinguish ‘unknown’ genes for which a curator has examined the literature and computational outputs and found no possible GO annotation from ‘uncurated’ genes that lack an annotation because existing publications for that gene have not yet been examined. ‘Unknown’ genes account for 30–34% of the genome within different GO aspects (Table 1). However, because our computational methods for locating all publications relevant to a gene are imperfect (in particular we don’t currently search for gene names in supplemental results files), it’s likely that some genes currently classified as ‘unknown’ should be included in the ‘uncurated’ category.

To improve curation efficiency and reduce the fraction of unannotated genes we have begun using a semi-automated curation process to identify papers with cellular component information and create annotations from them. Such a process has been used successfully by WormBase (17) to streamline and improve their curator’s efficiency in dealing with this type of data. We have worked closely with the WormBase Textpresso team to adapt and improve the software that is used in this process for use in *A. thaliana*. A combination of user submissions, semi-automated curation, integration of annotations from collaborating groups and strategic paper selection from the most recent literature will continue to drive the updates of functional information for *A. thaliana*.

### Table 1. *Arabidopsis thaliana* Gene Ontology Annotations

| GO aspect                      | Experimental (%) | All evidence (%) | Unknown (%) | Not annotated (%) |
|-------------------------------|------------------|------------------|-------------|-------------------|
| Biological process (BP)       | 5826 (20)        | 15644 (54)       | 9764 (34)   | 3367 (12)         |
| Molecular function (MF)       | 3816 (13)        | 16504 (57)       | 8732 (30)   | 3539 (12)         |
| Cellular component (CC)       | 7762 (27)        | 15383 (54)       | 7529 (26)   | 5863 (20)         |
| BP, MF or CC                  | 10595 (37)       | 22047 (77)       | n/a         | 939 (3)*          |

Number of *A. thaliana* genes with annotations to the three GO aspects and their percentages relative to the total number of genes excluding pseudogenes and transposable element genes, based on the TAIR10 genome release. ‘Experimental’ category includes genes annotated with evidence codes IDA (inferred from direct assay), IMP (inferred from mutant phenotype), IGI (inferred from genetic interaction), IPI (inferred from physical interaction) and IEP (inferred from expression profile). ‘All evidence’ includes all evidence codes except ND (no biological data available). ‘Unknown’ includes genes annotated to the GO root term within the indicated category using the ND evidence code. ‘Not annotated’ includes genes with no annotation to date within the indicated GO category. Numbers as of 15 September 2011; n/a not applicable.

*Genes with no GO annotation of any kind.*
To maximize both efficiency and accuracy of the genome annotation process TAIR has made use of a combination of computational methods to identify genes requiring updates and carry out simpler updates, and manual curation using the Apollo gene editing tool (19) to review and carry out more complex updates.

**Figure 1.** Overview of TAIR genome releases. (A) Bar graph displaying the number of annotation updates made in each of the 5 TAIR releases. Colored bars represent four different classes of updates: updated genes (light green), genes with CDS updates (orange), new genes (yellow) and new splice variants (dark green). (B) Table comparing the TAIR genome releases by types of data and prediction tools used, areas of focus and genome sequence updates. The red line separating TAIR8 from TAIR9 indicates that coordinates of most genes shifted in the TAIR9 release, as a consequence of the integration of 341 Indels, and the normalization of previously identified sequence contaminations to a standard length of 100 bp. A liftover tool is available at ftp://ftp.arabidopsis.org/home/tair/Software/UpdateCoord/ for updating coordinates of objects mapped to TAIR8 or earlier releases.

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**Figure 1** illustrates the consecutive steps that we have taken to gradually improve the annotation of the _A. thaliana_ genome. After an initial effort (TAIR6) focused on the annotation of protein-coding genes, more extensive reannotation projects were undertaken in subsequent releases to improve the annotation of UTRs, short protein-coding genes, non-coding RNAs (ncRNAs), transposon genes, pseudogenes and splice variants. As shown in **Figure 1B**, while the genome annotation tool PASA (Program to Assemble Spliced Alignments) (20) was the only gene prediction tool used in the first releases, curators increasingly relied on additional gene prediction tools to make use of newly available transcript profiling (RNA-seq) and peptide datasets generated using high-throughput methods.

**TAIR8 genome release (April 2008)**

As with the previously described TAIR6 and TAIR7 releases (21), PASA was used to incorporate all available _A. thaliana_ ESTs and cDNAs into transcript assemblies and generate lists of suggested updates to existing gene models for the TAIR8 release. These updates were categorized by PASA into different groups depending on the type of change required (i.e. extension of the 3’-UTR, altered protein coding sequence, etc.) All but the most straightforward update categories, such as extension of the 5’- or 3’-UTRs, were individually reviewed by curators using a modified PASA interface and marked for manual curation, computational update, or rejection.

In addition to making genome-wide computational updates based on new transcript evidence, for each TAIR release we have targeted a specific subclass of
genes for review and update. For the TAIR8 release, we integrated a large new set of transposable elements provided by Que sneville and co-authors (22) and used this new information to update the gene type for many genes contained within these newly mapped transposable elements from protein-coding or pseudogene to transpos able element gene (see Supplementary Data for more information). Other novel genes introduced in TAIR8 include conserved uORFs (upstream open reading frames located within the UTR of other, larger genes) (23), and very short protein-coding genes with substantial supporting evidence (24). Datasets from several other groups were also used to annotate new genes and splice variants and make gene structure updates (25–27, T. Tatusova, personal communication).

The TAIR8 release contained 27,235 protein coding genes, 859 pseudogenes, 3,900 transposable element genes and 1,288 ncRNAs (33,282 genes in all, 38,963 gene models). A total of 1,291 new genes and 2009 new gene models were added. Thirteen percent (4,330) of A. thaliana genes had annotated splice variants in this release. Updates were made to 3,811 gene structures of which 625 gene models had coding sequence (CDS) updates; a total of 4,007 exons were modified and 683 new exons incorporated. There were 33 gene splits and 41 gene merges. Overall 23% of all existing TAIR7 genes (7,380 genes) were updated, including updates to gene structure and/or gene type.

The TAIR8 release also included changes to the genome sequence. In 14 regions identified as contaminating sequences from vectors, E. coli or rice, the contaminating sequence was replaced with a run of ‘N’ of the same length to avoid changes to chromosome length. In addition, 1,425 single nucleotide substitutions were made to the assembly sequence based on high-confidence resequencing data provided to TAIR (28). The sequences of 518 genes overlapping these substitutions were also updated. Because all assembly changes for the TAIR8 release were substitutions rather than insertions or deletions, the chromosome lengths and gene coordinates were unchanged from the previous releases.

**TAIR9 genome release (June 2009)**

With the TAIR9 release, the set of data used as evidence for updates to gene structures was expanded to include cross-species alignments and peptide data from two large-scale proteomics experiments (29,30). These proteome datasets were used to reclassify 99 pseudogenes as protein coding and merge nine pseudogenes with existing protein coding genes. In addition, 158 peptides were used to update TAIR gene structures. A set of predicted Augustus gene models based on proteome data (30) were evaluated to identify potential exons missing from TAIR8. Of 591 Augustus models examined, 339 were incorporated into TAIR9 gene models, with 175 new splice variants added, 118 modifications to existing TAIR models and 46 new gene models.

The TAIR9 release also included a genome-wide reannotation of pseudogenes based on output from the PseudoPipe software package, provided by the Gerstein lab (31). Further analysis was undertaken to identify a subset of pseudogene models exhibiting CDS disablements or truncations relative to the parent gene. A total of 168 novel pseudogenes were added for the TAIR9 release.

Alternative genome annotation datasets derived from several different software packages, including Gnomon (http://www.ncbi.nlm.nih.gov/projects/geneguide/gnomon.shtml), (predictions provided by Tatiana Tatusova and Alexandre Souvorov, NCBI), EuGene (27,32) (gene predictions provided by Sébastien Aubourg, Unité de Recherche en Génomique Végétale) and AceView (33), predictions provided by Jean Thierry-Mieg, NCBI) were also used as a source of gene structure updates for the TAIR9 release. An analysis of these gene prediction sets was undertaken to identify a set of exons absent from TAIR8 annotation but supported by transcript, peptide or cross-species evidence, resulting in the addition or modification of over a thousand exons for TAIR9. The full set of alternative gene models submitted to TAIR for all three software packages can be viewed in TAIR’s genome browser as tracks within the Community Alternative Annotation section (http://gbrowse.arabidopsis.org/cgi-bin/gbrowse/arabidopsis).

The TAIR9 release contained 27,379 protein coding genes, 926 pseudogenes, 3,901 transposable element genes and 1,312 ncRNAs (33,518 genes in all, 39,640 gene models). A total of 282 new genes and 739 new splice variants were added. Fourteen percent (4,626) of A. thaliana genes had annotated splice variants in this release. Updates were made to 1,254 gene models of which 774 had CDS updates; a total of 1,144 exons were modified and 1,056 new exons incorporated. There were 13 gene splits and 46 gene merges.

Genome assembly updates made for the TAIR9 release included 227 single nucleotide substitutions based on resequencing data provided to TAIR (28,34). A set of 341 insertions or deletions were made based on resequencing data (34) and EST or cDNA sequences deposited in GenBank that supported the change. In accordance with our reference genome policy (http://arabidopsis.org/doc/portals/geneAnnotation/gene_structural_annotation/ ref_genome_sequence/11413) corrections to the reference assembly were only made if supported by at least two independently derived sequence libraries from the Columbia ecotype. In addition to these changes, the 14 regions previously identified in TAIR8 as either vector, E. coli or rice contamination and substituted with the equivalent number of IUPAC ambiguity code ‘N’ were standardized (via deletion) to a set size of 100 bp for TAIR9. As a consequence of these assembly updates, the coordinates of most genes, as well as other mapped features such as transcripts, polymorphisms, T-DNAs, etc. were modified between the TAIR8 and TAIR9 releases.

**TAIR10 genome release (December 2010)**

For the TAIR10 release, RNA-seq data were incorporated as evidence for gene model updates. Data used for this release included a total of 538 million reads obtained from two groups (28,35). RNA-seq reads were mapped...
to the genome using TopHat (36), HashMatch (http://mocklerlab-tools.cgrb.oregonstate.edu/HashMatch.html) and Supersplat (37). After quality and low complexity filtering, we mapped >200 million reads to the genome, including about nine million spliced reads. Spliced aligned reads can be viewed within TAIR's genome browser, in the ‘Spliced RNA-Seq Reads’ track within the Sequence Similarity section (http://gbrowse.arabidopsis.org/cgi-bin/gbrowse/arabidopsis). These spliced read alignments plus peptide data obtained for the TAIR9 release were used as an input for the Augustus gene prediction package (38) and the resulting gene models were categorized and manually reviewed (see Supplementary Figure S1). Validated gene updates, novel genes and novel splice variants from the Augustus output were incorporated into the TAIR10 release. Spliced reads not incorporated into gene models by Augustus were supplied to TAU (http://mocklerlab-tools.cgrb.oregonstate.edu/TAU.html), and resulting models were reviewed by TAIR curators for the addition of novel splice variants. Transcript assemblies were also generated independently via Cufflinks (39), using both spliced RNA-seq reads and a subset of unspliced reads generated by the Ecker lab. Transcript assemblies were filtered and compared to existing gene models, resulting in the addition of 56 novel genes.

In addition to the updates resulting from incorporation of RNA-seq data, new proteome data provided to TAIR (40) was used to directly update 24 gene models. Also, gene models created using the Gnomon pipeline were provided to TAIR by NCBI and reanalysis of these models resulted in 11 additional novel genes, 67 additional alternative splice variants and 164 updates to existing genes. Finally, a set of 125 updates provided by curators at Swiss-Prot (http://www.uniprot.org/) were reviewed and 104 of these updates were incorporated into this release.

The TAIR10 release is summarized in Table 2. For this release, a total of 126 new genes and 2099 new splice variants were added. Updates were made to 1184 gene models of which 707 had CDS updates. There were 41 gene splits and 37 gene merges. No updates were made to the genome assembly for the TAIR10 release.

| Gene Type          | Protein Coding | pre-rRNA | rRNA | snoRNA | miRNA | Other RNA | Pseudo gene | TE gene | Total |
|--------------------|----------------|----------|------|--------|-------|-----------|-------------|---------|-------|
| Nuclear            | 27206          | 631      | 4    | 13     | 71    | 177       | 394         | 924     | 3903  | 33323 |
| Chloroplast        | 88             | 37       | 8    | 0      | 0     | 0         | 0           | 0       | 0     | 133   |
| Mitochondrial      | 122            | 21       | 3    | 0      | 0     | 0         | 0           | 0       | 0     | 146   |
| Total              | 27416          | 689      | 15   | 13     | 71    | 177       | 394         | 924     | 3903  | 33602 |

Number of genes of each category in the TAIR10 genome release.
information so that users can quickly confirm the usefulness of a particular paper and link directly to the full text, if they have the appropriate subscriptions to the journals in question.

N-Browse is an interactive graphical browser for biological networks (42). Users can launch N-Browse using Java Web Start with or without an initial query gene. N-Browse contains 8626 protein–protein interactions based on experimental data curated by TAIR or the protein interaction databases BioGRID (http://thebiogrid.org) (43) or IntAct (http://www.ebi.ac.uk/intact) (44). N-Browse does not currently contain any predicted protein interaction data. Interaction data is available for download at ftp://ftp.arabidopsis.org/home/tair/Proteins/Protein_interaction_data/.

GBrowse_syn is a GBrowse-based synteny browser designed to display multiple genomes, with a central reference species compared to several additional species (45). This tool uses a central ‘joining’ database that contains information about the multiple sequence alignments as well as additional databases for each species represented in the alignments. GBrowse_syn was built to help researchers study and analyze syntenic regions, homologous genes and other conserved elements between sequences. It can also be used to study genome duplication and evolution. By comparing newly sequenced or less studied genomes to the well annotated A. thaliana genome in GBrowse_syn, scientists can identify novel genes and putative regulatory elements. The first version of the GBrowse_syn tool at TAIR includes the genomes of A. thaliana, A. lyrata and Populus trichocarpa.

Integrated Genome Browser (IGB) is an interactive genome browser tool (46). IGB is different from other genome browsers in that it lets the user open, visualize and analyze their own large-scale data sets (i.e. RNA-Seq, ChIP-Seq, epigenetics, tiling array, etc), displaying these data alongside publicly available data sets, including gene models and the reference sequence itself. Using IGB’s QuickLoad system, users can also use IGB to share data with collaborators and members of the community. IGB runs on the user’s local computer rather than on the TAIR servers.

New plant genomes in GBrowse In addition to GBrowse for A. thaliana, TAIR has made GBrowse viewers available for the following plant genomes: A. lyrata, Brachypodium distachyon, Oryza sativa ssp. japonica, O. sativa ssp. indica, P. trichocarpa, Physcomitrella patens, Sorghum bicolor, Vitis vinifera, Zea mays. Gene models for each species were obtained from Ensembl, while transcript data were retrieved from GenBank and aligned to each genome using the GMAP alignment tool (47). Arabidopsis thaliana gene models were aligned to each plant genome using CAT (48), and the alignments are displayed in a GBrowse ortholog track.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online: Supplementary Methods and Supplementary Figure S1.

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