Original article

Prevalence of *Anaplasma marginale* in cattle blood samples collected from two important livestock regions in Punjab (Pakistan) with a note on epidemiology and phylogeny of parasite

Shahzadi Noor Ul Ain Zafara, Adil Khanb,⇑, Sadaf Niazc, Munir Aktasd, Sezayi Ozubekd, Muhammad Farooqe, Muhammad Moeen Adile, Zbigniew Zaja˛cf, Furhan Iqbala, Ahmad R. Alhimaidih, Ayman A. Swelumg

a Institute of Pure and Applied Biology, Zoology Division. Bahauddin Zakariya University, Multan, Pakistan
b Department of Zoology, Bacha Khan University, Charsadda, Pakistan
c Department of Zoology, Abdul Wali Khan University, Mardan, Pakistan
d University of Firat, Veterinary Faculty, Department of Parasitology, 23119, Elazig, Turkey
e Department of Zoology, Ghazi University, Dera Ghazi Khan, Pakistan
f Department of Biology and Parasitology, Medical University of Lublin, Radziwiłłowska 11, 20-080 Lublin, Poland
g Department of Animal Production, College of Food and Agriculture Sciences, King Saud University, 2460, Riyadh 11451, Saudi Arabia
h Department of Zoology, College of Science, King Saud University, Riyadh 11451, Saudi Arabia

A R T I C L E  I N F O

Article history:
Received 9 September 2021
Revised 13 October 2021
Accepted 13 November 2021
Available online 19 November 2021

Keywords:
Molecular characterization
Cattle
Punjab
Pakistan

A B S T R A C T

Anaplasmosis, caused by intracellular gram-negative bacteria *Anaplasma marginale* is one of the most frequently reported tick-borne disease (TBDs) in tropical and sub-tropical countries, including Pakistan. In the present study, a total of 428 cattle blood samples were collected to examine the prevalence and phylogenetic origin of *A. marginale* in two important livestock regions of Punjab Province in Pakistan, i.e. Lodhran and Dera Ghazi Khan Districts. In addition, association between occurrence of *A. marginale* in cattle blood and selected epidemiological factors has been also investigated. The presence of *A. marginale* genetic material was confirmed in 9% of the tested blood samples taken from cattle in Lodhran and in 17% from Dera Ghazi Khan. Prevalence of *A. marginale* was significantly higher in cattle from Dera Ghazi Khan. All the cattle breeds from both districts were equally susceptible to *A. marginale* infection. We reported higher prevalence of *A. marginale* in cattle living indoors or with other dairy animals in Dera Ghazi Khan. However, no such relationship was observed in the Lodhran district. Sequencing of the msp1b gene shows 96–99% similarity of *A. marginale* in the study area to those reported from other parts of Pakistan, South Africa, and Israel. We recommend that large scale tick and tick-borne disease control strategies must be implemented in both districts.

© 2021 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Majority of Pakistani population resides in rural areas. Rearing and trading livestock is a major source of income and employment in these societies (Irshad et al., 2010). Majority of livestock owners are small farm holders and they are heavily contributing in meeting the dietary requirements of their countrymen as well as in national economy (Ashraf, 2021). 15 indigenous cattle breeds of zebu (one-humped) have been reported from Pakistan that includes local breeds like Sahiwal and Cholistani (known to be tick and heat resistant) and exotic breeds like Holstein-Friesian and Jersey (that are known for high milk production) (Saeed, 2016). Livestock and dairy industry face a lot of challenges in Pakistan including unawareness of livestock owners regarding feed, tick management artificial insemination, finance constrains and limited available health facilities (Arif, et al., 2011; Ramzan et al., 2018). Parasitism is a crucial problem causing health problems in live-
stock farms. Ticks are one of the most important ectoparasites in this region. This is favored by the climate in Pakistan, which provides optimal conditions for the development and reproduction of ticks. A large variety of ticks (belonging to genera *Rhipicephalus*, *Dermacentor*, *Amblyomma* and *Rhipicephalus*) has been reported from Pakistan that infests variety of domestic, wild animals as well as humans (Ashraf, 2021; Durrani and Shakoori, 2009).

*Anaplasma marginale* is an intra-erythrocyte gram-negative bacterium that causes bovine anaplasmosis (Ashraf, 2013). Ticks of *Dermacentor* and *Amblyomma* and *Rhipicephalus* genera act as important vector of *A. marginale* in Pakistan (Kocan et al., 2004; Hairgrove, 2015). The most characteristic symptoms of anaplasmosis are anemia, anorexia, pyrexia, depression, and jaundice in large ruminants. Infection with *A. marginale* can also result in reduced milk production, can lead to abortion in pregnant animals and, in extreme cases can kill animal (Camus et al., 2010; Kumar, 2015).

Prevalence of *A. marginale* is known to vary with sampling sites, vector species, breeds and breeding system as well as the climatic conditions (Bursakov and Kovalchuk, 2019). In the past a few studies have been conducted in various parts of Pakistan showing the prevalence of *A. marginale* in cattle ranging from 6 to 41% (Hussain, 2017; Farooqi et al., 2017; Turi et al., 2018; Ashraf et al., 2021). These reports are mainly from Punjab and Khyber Pakhtoonkhwa (KPK) provinces and most of the remaining part of Pakistan is still unexplored for tick-borne diseases (TBDs) (Parveen et al., 2021).

Lodhran and Dera Ghazi Khan are among those districts in Punjab where local population is heavily dependent on livestock for their earnings, but the TBDs are not explored in detail in these areas. Most local farmers consider all ticks as one species and is mostly unaware of their specific health effects to various animal species they are rearing. To the best of our knowledge, there are no previous reports, from both the districts, regarding molecular detection of *A. marginale* in cattle breeds. Hence, the present study was designed to document the PCR based prevalence of *A. marginale* in cattle breeds enrolled from Lodhran and Dera Ghazi Khan, with a note on epidemiology and phylogenetic origin of this bacterium with the hypothesis that bacterium prevalence would be higher in Dera Ghazi Khan district due to its poor socio economic and hygiene conditions.

2. Materials and methods

2.1. Study area

Two districts (Dera Ghazi Khan and Lodhran) in South Punjab (Pakistan) were included during present investigation (Fig. 2). Lodhran is located at 29° 32' 34" N, 71° 37' 48" E. Lodhran is located on the northern side of river Satluj. On its north are the districts of Multan, Khanewal and Vehari while Bahawalpur is on its southern side. On the east lie the districts of Vehari and Bahawalpur; while district Multan lies on the western side. The climate of the district is hot and dry in summer and cold in winter. The maximum and minimum temperature ranges between 42°C and 28°C in summer. During winter, the temperature fluctuates between 21°C and 5°C. The entire district is smooth plain. District Dera Ghazi Khan is located at 30° 1’ 59" N, 70° 38’ 24" E. The Dera Ghazi Khan is located in a strip between the river Indus and the Koh-Suleman range of mountains separating it from the Baluchistan Province. It is surrounded by Dera Ismail Khan on the North and Rajanpur on its South. Indus river flows on the East across which lie the districts of Muzafargarh and Layyah. Loralai and Dera Bugti districts of Baluchistan Province lies on the West separated by the Koh-Suleman range of mountains. The overall climate of this district is dry with little rainfall. The winter is mild, lasting from November till January with average temperature 4 °C but summer is extremely hot and extended, starting in May, and lasting till September with average temperature around 42 °C. The different geographic location of these districts was one of the key reasons that they were investigated for the presence of *A. marginale* in cattle.

2.2. Sample collection

Ethical Research Committee of the Institute of Pure and Applied Biology at Bahauddin Zakariya University Multan (Pakistan) approved all the experimental procedures and protocols applied in this study via letter number IP&B/EB/Ethics/2019-17. In total 428 cattle blood samples were collected between June and October 2019 (District Lodhran, n = 218 and Dera Ghazi Khan, n = 210) from cattle breeds: Cholistan, Sahiwal, Crossbreed, Daji, Holstein Friesian, Australian Ayrshire and Jersey. Solvin’s formula was used to estimate sample size that were randomly collected during present study. Solvin’s formula was computed as:

\[ n = N/(1 + N \times e^2) \]

Whereas: n = no. of samples, N = total population, e = margin of error.

Only apparently healthy animals were included in this study. Body of each animal was carefully inspected to detect the presence of tick(s) on them. A questionnaire was filled on sampling site with the help of cattle owners used to assess the epidemiological situation on farms (sex, age, number of animals in herd, presence, or absence of other dairy animals in herds, presence or absence of dogs, history of disease, animals’ living environment, water supply to farm and ticks feed on animals).

2.3. DNA extraction

Blood sample (3–5 ml) was collected through jugular vein puncture. Genomic DNA from the collected blood was extracting following the protocol of Saeed et al. (Malik, 2018). Briefly, blood samples were suspended in 500 μL of lysis buffer (20 mM Tris–HCl, 1 mM EDTA, 0.5% SDS) with 0.4 mg/mL Proteinase K (Fermentas, USA) and incubated at 55 °C overnight. Subsequently, samples were heated at 95 °C for 10 min and equal volume of phenol–chloroform: isoamylalcohol (25:24:1 v/v/v), was added to the lysate, vortexed for 30 s and centrifuged at 12,000g for 10 min. The aqueous phase was transferred to new Eppendorf tube and equal volumes of ice cold isopropanol were added. The DNA was pelleted by centrifugation at 12,000g for 15 min and washed with 70% ethanol and dried at 65 °C for 5 min. The DNA was finally re-suspended in 50 μL sterile double distilled water and stored at −20 °C.

2.4. PCR amplification

Primer sequence R 5’-CTG CTT GGG AGA ATG CACCT 3’ and F 5’GCT CTA GCA GGT TAT GCC TC 3’ were applied to generate 265 bp fragment from *A. marginale’s* msp1b (major surface protein–1b) gene following Ashraf et al. (Ashraf, 2021) A reaction mixture (50 μL) that contained 65 mM KCl, 13 mM Tris–HCl (pH 8.3), 1U of DNA polymerase (Vivantas, USA), 2 mM MgCl₂, 250 μM of dNTP, 0.5 μM of primers and 5 μL of template DNA was prepared for PCR. Thermal profile of PCR included 5 min of denaturation at 94 °C followed by 25 repeated cycles of DNA denaturation for 50 s at 95 °C, primer annealing for 50 s at 56 °C and primer extension for 50 s at 72 °C. A final primer extension for 5 min was carried out at 72 °C (Tay et al., 2014). A. marginale positive sample (previously isolated from equine and confirmed by DNA sequencing: GenBank accession number MK792344) and parasite negative samples (no extraction from the cattle blood samples) were run.
as controls (positive and negative respectively) during each reaction for quality control.

2.5. DNA sequencing followed by phylogenetic analysis

To confirm that amplified DNA is of *A. marginale*, 4 amplified PCR amplicons were sent to a commercial lab (First base Sequencing, Malaysia) for DNA sequencing by using the same primer sequences as used during PCR. The obtained DNA sequences were compared using the Basic Local Alignment Search Tool (BLAST) with the NCBI database (https://blast.ncbi.nlm.nih.gov/Blast.cgi), and similar sequences were obtained from the GenBank database (https://www.ncbi.nlm.nih.gov/), to perform multiple alignments among the sequences using MUSCLE.

MEGA version X was used to perform the phylogenetic analysis, with a bootstrap analysis with 1000 replicates. For msp1b evolu-
that amplified PCR products of msp1b gene in present study were 96–99% similar to msp1b gene sequence of A. marginale from Pakistan, South Africa and USA (Fig. 1).

3.3. Epidemiological factors analysis

Indoor living environment \((p = 0.05)\), living with other dairy animals \((p = 0.03)\) and absence of dogs at dairy farms \((p = 0.04)\) were found to be associated with A. marginale infection in cattle from Dera Ghazi Khan District. On the other hand no such relation was observed in Lodhran district \((p > 0.05)\) (Table 2).

4. Discussion

Tick-borne diseases are one of the most important factors limiting the dairy industry and cause huge economic losses (Ashraf, 2013; Malik, 2018). This relationship is also observed in Pakistan, from where there are numerous reports of ticks collected from animals, including livestock. This is favored by the climate. Pakistan has sub-tropical climatic conditions and due to high air temperature and humidity, ticks and TBDs are common in this area (Ashraf, 2013).

In present study we confirmed the occurrence of genetic material of A. marginale in 9% and 17% of cattle blood samples collected from Lodhran and Dera Ghazi Khan Districts respectively (Table 1). Our results are in line with those of Ashraf et al. (Ashraf, 2021) who had reported 8.6% infection rate of A. marginale in cattle from Layyah District in Pakistan. There are few other studies regarding prevalence of A. marginale in cattle from Pakistan. Farooqi et al. (Farooqi, 2018) had documented 18.3% prevalence of A. marginale in bovine samples collected from three distinct zones of Khyber Pakhtunkhwa (KP) province. In the other study conducted in KP, 41.6% cattle from District Lakhi Marwat and Peshawar were reported to be infected with A. marginale (Turi et al., 2018). Hussain et al. (Hussain, 2017) had reported that 6.1% of apparently healthy cattle (Cholistan breed) from District Bahawalpur in Punjab were infected with A. marginale. Bovine anaplasmosis has been reported from various countries across the globe including Turkey (2.3%) (Aktas et al., 2011); Tunisia (3.7–25.4%) (Belkahia, 2015) and Brazil (5.4%) (Barbosa da Silva, 2014). These variations in prevalence of A. marginale reported from different areas of the world are probably due to variation in tick control and management programs in these regions, suitability of the climatic conditions for tick growth, different farm management techniques and animal husbandry practices that are used in those regions (Belkahia, 2015; Yukari and Umur, 2002).

To report the genetic diversity of A. marginale in studied cattle populations, phylogenetic analysis based on msp1b gene of parasite was conducted. The obtained sequences (MW303432, MW303433 and MN867482) were like sequences previously reported from Pakistan from District Layyah (MK032843-1 and MK032842-1), and USA (AF110808.1, M59845.1 and AF221693.1)). The MW303431 presented a higher similarity with msp1b gene sequence of A. marginale from Pakistan (96–99% similar). Some of the obtained sequences were like the South Africa sequences (KU647715, and KU647715) sequences (Fig. 1). The obtained sequences didn’t group with sequences from Egypt (MN222768.1, MN2227687.1) China (KU586030.1, MF326718.1, MF326719.1, AJ533048.1, MF326770.1) Thailand (KT264188.1) and Mexico (EU283346.1) showing a complex diversity worldwide.

Specific epidemiological factors can influence the host–parasite associations including climate conditions, behavioral traits, host sociality, population density, diet, habitat, age, sex, host immune-competence, supplementary feeding and animal translocations (Hussain et al., 2021). During the present investigation, we have also recorded a number of epidemiological factors explaining the characters of cattle as well those of the farms where they were...
kept in order to report their association with the prevalence of *A. marginale*. In present study the prevalence of *A. marginale* did not differ statistically between cattle breeds from Lohdran and Dera Ghazi Khan districts (Table 2). This observation is contradictory to Ashraf et al. (Ashraf et al., 2021) who had documented highest prevalence of *A. marginale* in exotic Holstein Friesian than in local Sahiwal breed. Similarly, Khan et al. (Khan, 2019) and Tay et al. (Tay et al., 2014) had also confirmed higher *Anaplasma* spp. prevalence in Holstein Friesian breed than in local cattle breeds. They assumed that Holstein Friesian breed has long and thick hairs compared to local breeds that make them a preferred host for ticks to infest. There are two potential reasons for this difference in parasite prevalence in cattle breeds from two studies; first, local breeds are more resistant against rickettsial infection (Ashraf, 2021) and second reason is different number of samples that were examined in two studies. The number of samples tested by Ashraf et al. (Ashraf et al., 2021) were almost double than the sample we have included in present study. This difference can clearly affect the breed specific prevalence ratios of *A. marginale*.

In our study, we have reported that absence of the dogs at the dairy farms and cattle living indoor are important factor associated with *A. marginale* infection. We also reported that presence of other dairy animals on farms with cattle is significantly associated with the *A. marginale* infection in cattle from District Dera Ghazi Khan (Table 2). Our results are also in agreement with those of Ashraf et al. (Ashraf, 2013) who had confirmed that *A. marginale* prevalence was higher in herds that do not had dogs and they had proposed that *A. marginale* infection was probably transmitted from infected to healthy by physical contact. Although, no association was found between the presences of dogs at dairy with the prevalence of *A. marginale* during present study but it is also an established facts that rearing dogs at the livestock farms is a common practice in Pakistan, especially in rural areas. It has been reported that dogs in Pakistan (Abid, 2021) are infested with *Rhipicephalus* spp. and these ticks are also known to infest cattle in Pakistan (Ashraf, 2021; Parveen et al., 2021). There is a possibility that physical contact of dogs with cattle can result in tick transfer from one host to other resulting in spread of tick borne diseases among them.

During the second half of the 20th century, many dairy farmers moved away from pasture-based housing systems and housed their dairy cows indoors year-round. This shift in housing system decreased the exposure of the cattle to the various parasites and their vectors. In addition, development and use of pharmaceutical parasiticides decreased the likelihood of heavy parasite infestations and reduced related production losses (Sorge et al., 2015). At dairy farms where modern scientific procedures are applied, the dairy cattle shows greater economic importance as compared to farms where old traditional methods of animal rearing are in practise, encouraging the farmers to adopt appropriate hygiene practices during milking and reproduction of the herd (Simioni et al., 2013). Most of the farms in district Dera Ghazi Khan are

---

Table 1
Comparison of *A. marginale* prevalence in blood samples of various cattle breeds collected from two Lodhran and Dera Ghazi Khan districts.

| Breed    | Districts          | Number of tested blood samples | *A. marginale* positive samples | *A. marginale* negative samples | p-value | Number of tested blood samples | *A. marginale* positive samples | *A. marginale* negative samples | p-value |
|----------|--------------------|--------------------------------|---------------------------------|--------------------------------|---------|--------------------------------|---------------------------------|---------------------------------|---------|
|          | Lohdran            |                                 |                                 |                                |         | Dera Ghazi Khan                |                                 |                                |         |
| Cholistan| 87                 | 8 (9%)                         | 79 (91%)                        | 0.60                           |         | 51                             | 13 (25%)                        | 38 (75%)                        | 0.20    |
| Sahiwal  | 122                | 12 (10%)                       | 110 (90%)                       |                                |         | 19                             | 5 (26%)                         | 14 (74%)                        |         |
| Crossbreed| 9                  | 0 (0%)                         | 9 (100%)                        |                                |         | 57                             | 6 (11%)                         | 51 (89%)                        |         |
| Dali     | –                  | –                              | –                               |                                |         | 37                             | 5 (14%)                         | 32 (86%)                        |         |
| Holstein Friesian| –     | –                              | –                               |                                |         | 35                             | 6 (17%)                         | 29 (83%)                        |         |
| Australian Ayrshire| –       | –                              | –                               |                                |         | 11                             | 1 (9%)                          | 10 (91%)                        |         |
| Jersey   | –                  | –                              | –                               |                                |         | 210                            | 3 (17%)                         | 174 (83%)                       |         |
| Total    | 218                | 20 (9%)                        | 198 (90%)                       |                                |         | 210                            | 3 (17%)                         | 174 (83%)                       |         |

Table 2
Association of *A. marginale* prevalence and epidemiological parameters from Lodhran and Dera Ghazi Khan districts. Based on age, 5 years old animals were considered young while older than 5 years were mature.

| Parameters                 | Districts          | Lohdran | Dera Ghazi Khan |
|----------------------------|--------------------|---------|-----------------|
|                            | Number of tested   |         |                 |
|                            | blood samples      |         |                 |
|                            | *A. marginale*     |         |                 |
|                            | positive samples   |         |                 |
|                            | negative samples   |         |                 |
|                            | p-value            |         |                 |
| Gender                     | Male               | 2 (8%)  | 18 (24%)        |
|                            | Female             | 18 (9%) | 17 (13%)        |
| Age                        | >5 year            | 12 (10%)| 25 (18%)        |
|                            | <5 year            | 08 (9%) | 11 (15%)        |
| Health status              | Healthy            | 20 (10%)| 189 (91%)       |
|                            | Fever              | 0 (0%)  | 09 (100 %)      |
| Dogs presence              | Present            | 11 (9%) | 118 (90%)       |
|                            | Absent             | 9 (10%) | 80 (90%)        |
| Presence of feeding ticks  | Present            | 5 (10%) | 46 (90%)        |
|                            | Absent             | 15 (9%) | 152 (91%)       |
| Living environment         | Outdoor            | –       | –               |
|                            | Indoor             | –       | –               |
| Presence of other dairy    | Present            | –       | –               |
| animals                    | Absent             | –       | –               |
| Water supply               | Pump               | –       | –               |
|                            | Pool               | –       | –               |
|                            |                   |         |                 |
|                            |                   |         |                 |

---

1519
poorly managed and old traditional techniques for animal rearing and management are still in practice that leads to poor hygiene and increases the risk of tick infestation and prevalence of TBDs which is event from the fact that the prevalence of A. marginale was higher in cattle from Dera Ghazi Khan district as compared to Lodhran. Swai et al. (Swai, 2005) and Atif et al. (Atif et al., 2012) had also documented higher infection rate of A. marginale in farms that were managed by old traditional techniques than the farms where modern scientific techniques were applied.

5. Conclusions

In conclusion, we are reporting that prevalence of A. marginale was higher in cattle from Dera Ghazi Khan than in cattle from Lodhran. No specific cattle breed was found susceptible to A. marginale infection. We observed that poor farm management practices are responsible for increasing prevalence of TBDs in studied regions. As we did not include any cattle suffering from anaplasmosis in this study, so none of the enrolled animals showed the symptoms of anaplasmosis but still A. marginale was detected in their blood. Studies similar to the present one are necessary for the prophylactic detection of this bacterium in cattle, so that they must be diagnosed and treated before the onset of acute form of disease preventing the economic losses.

Author contributions

F.I., Z.Z. and M.F. had designed and supervised this study. S.N.A., Z., S.N. and M.M.A. had collected blood and epidemiological data from cattle. A.P., I.A.E., M.A. and M.M.A. had extracted DNA from the blood samples and performed PCR. A.K. had purified PCR products and analyzed DNA sequences. M.A. A.A. A.S. and S.O. had conducted phylogenetic tree and analyzed the data. All authors reviewed the manuscript.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgement

The authors thank the researchers supporting project for their funding this work number (RSP-2021/232) at King Saud University, Riyadh, Saudi Arabia.

Ethics approval

This study was revised and approved by the Ethical Research Committee of Institute of Pure and Applied Biology at Bahauddin Zakariya University Multan (Pakistan) according to the ethical principles of human and animal research. This study was carried out in compliance with the ARRIVE (Animal Research: Reporting of In Vivo Experiments, PLOS Bio 8(6), e1000412,2010) guidelines.

References

Abid, K. et al., 2021. Molecular detection and prevalence of Theileria ovis and Anaplasma marginale in sheep blood samples collected from district Layyah in Punjab Pakistan. Trop. Anim. Health Prod. 53 (4), 439.
Akar, M., Altay, K., Dumanli, N., 2011. Molecular detection and identification of Anaplasma and Ehrlichia species in cattle from Turkey.Ticks Tick Borne Dis. 2, 62–65.
Arl, M. et al., 2013. Prospects and limitations of dairying in Gujanwala district (Punjab-Pakistan). In: Proceedings of International Workshop on Dairy Sciences. Park Nov 21-23, 2011, Agriculture University Peshawar. Pak. J. Anim. Plant Sci. 23, 34–37.
Ashraf, Q.U.A. et al., 2013. A report on the high prevalence of Anaplasma sp. in buffalo from two provinces in Pakistan. Ticks Tick Borne Dis. 4, 395–398.
Ashraf, S. et al., 2021. A report on molecular detection and phylogenetic evaluation of Anaplasma marginale in ticks and blood samples collected from cattle in Layyah, Punjab, Pakistan (P. T. D. S. 2020).
Ashraf, S., Parveen, A., Asif, M., Alanaazi, A.D., Alouffi, A., Muhammad Awais, M., Khan, A., Aktas, M., Ozbuke, S., Iqbal, F., 2021. First report regarding molecular epidemiology and novel variant identification of Anaplasma centrale in cattle from Pakistan. Saudi J. Biol. Sci. 28 (11), 6488–6494. https://doi.org/10.1016/j.sjbs.2021.07.026.
Atif, F.A., Khan, M.S., Iqbal, H.J., Ali, Z., Ullah, S., 2012. Prevalence of tick control infestation in three districts of the Punjab, Pakistan. Pak. J. Sci. 64, 49–53.
Barbosa da Silva, J. et al., 2014. Molecular and serological prevalence of Anaplasma marginale in water buffaloes in northern Brazil. Ticks Tick Borne Dis. 5, 100–104.
Belkaiha, H. et al., 2015. First molecular survey and novel genetic variants’ identification of Anaplasma marginale, A. centrale and A. bovis in cattle from Tunisia. Inf. Gen. Evol. 34, 361–371.
Bursakov, S.A., Kovalchuk, S.N., 2019. Co-infection with tick-borne disease agents in cattle in Russia. Ticks Tick Borne Dis. 10 (3), 709–713.
Camus, E., Uilenberg, G. Anaplasmosis. In Infectious and Parasitic Diseases of Livestock. Bacterial Diseases, Fungal Diseases, Parasitic Diseases. Lefevre, P.C., Blancou, J., Chermette, R., Uilenberg, G. (Eds.), Lavoisier, 1247–1263 (Paris, 2016).
Durrani, A.Z., Shakoori, A.R., 2009. Study on ecological growth conditions of cattle Hylaomma ticks in Punjab, Pakistan. Iran. J. Parasitol. 4, 19–25.
Faroq, S.H. et al., 2018. Molecular epidemiology of bovine anaplasmosis in Khyber Pakhtunkhwa, Pakistan. Trop. Anim. Health Prod. 50, 1591–1598.
Farooqi, S.H., Naz, M., Saleem, M.H., Rashid, M.I., Omeeb, M., Khan, A., Aqil, A., Mahmood, S., 2017. Distribution of ixodid tick species and associated risk factors in temporal zones of Khyber Pakhtunkhwa Province, Pakistan. Pak. J. Zool. https://doi.org/10.17582/journal/2017/49.6.2017.
Hairgrove, T. et al., 2015. Molecular and serological in-herd prevalence of Anaplasma marginale infection in Texas cattle. Prev. Vet. Med. 119, 1–9.
Hussain, M.F. et al., 2017. Molecular detection of Anaplasma in apparently healthy Cholistan breed of cattle from the Bahawalpur district, Pakistan. Trop. Biomed. 34, 37–44.
Hussain, S., Hussain, A., Ho, J., Li, J., George, D., Rehman, A., Zeb, J., Sparagano, O., 2021. An epidemiological survey regarding ticks and tick-borne diseases among stock owners in Punjab, Pakistan: a one health context. Pathogen. 10, 361.
Irshad, N., Qayyum, M., Hussain, M., Khan, M.Q., 2010. Prevalence of tick infestation and theileriosis in sheep and goats. Pak. J. Vet. 30, 178–180.
Khan, N.U. et al., 2019. Prevalence and risk factors analysis associated with anaplasmosis in symptomatic cattle under field conditions in southern Khyber Pakhtoonkhwa, Pakistan. Pu. Appl. Biol. 8, 2119–2127.
Kimura, M., 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J. Mol. Evol. 16, 111–120.
Kocan, K.M., De La Fuente, J., Blouin, E.F., Garcia-Garcia, J.C., 2004. Anaplasma marginale (Rickettsiales: Anaplasmataceae): recent advances in defining host–pathogen adaptations of a tick-borne rickettsia. Parasitology 129 (1), 5285–5300.
Kumar, T. et al., 2015. Emerging status of anaplasmosis in cattle in Hisar. Vet. World 8, 768–771.
Malik, M.I. et al., 2018. Molecular detection of Ehrlichia canis dogs from three districts in Punjab (Pakistan). Vet. Med. Sci. 4, 126–132.
Parveen, A., Ashraf, S., Khan, A., Asif, M., Iqbal, F., 2021. Epidemiology of tick-borne diseases. In: Kumar, S., Bayugar, R.C., Sharma, A.K., Miranda, E.M., Chaubey, A.K. (Eds.), The Entomological Guide to Rhipicephalus. Nova Science Publishers, New York.
Ramzan, M., Ussar, N.U., Syed, H.M.B., Ghulam, M., Alamgir, A.K., 2018. Knowledge, attitude and practices of herdsmen about ticks and tick-borne diseases in district Multan, Pak. Entomol. 40, 13–18.
Saedz, Z. et al., 2016. Molecular prevalence and hematology of tropical theileriosis in Cholistani cattle from nomadic herds of the Cholistan desert, Pakistan. Kaf. Un. Vet. Fak. Derug. 22, 281–286.
Simioni, F.S., Baretta, C.R.D., Steiner, L.M., Lopes, L.S., Tizziani, T., 2013. Qualidade do leite proveniente de propriedades com diferentes níveis de especialização. Semina: Ciênc. Agr. 34 (4), 1901–1912.
Sorge, U.S., Moon, R.D., Stromberg, B.E., Schrath, S.L., Michels, L., Wolff, L.J., Kelton, D.F., Heins, B.J., 2015. Parasites and parasite management practices of organic and conventional dairy herds in Minnesota. J. Dairy Sci. 98 (5), 3143–3151.
Swai, E.S. et al., 2005. Seroprevalence estimation and risk factors for A. marginale on smallholder dairy farms in Tanzania. Trop. Anim. Health Prod. 37, 599–610.
Taji, S.T., Koh, F.X., Kho, K.L., Ong, B.L., 2014. Molecular survey and sequence analysis of Anaplasma spp. in cattle and ticks in a Malaysian farm. Trop. Biomed. 31, 769–776.
Turi, A.T., Rahman, A., Ali, L., 2018. Comparative analysis of indirect ELISA and real time PCR for the detection of Anaplasma marginale in buffalo, cattle and sheep in district Peshawar and Lakki Marwat, Pakistan. South As. J. Life Sci. 6 (2018), 1–6.
Yukari, B.A., Umar, S., 2002. The prevalence of tick species (koxoides) in cattle, sheep and goats in the Burdur region, Turkey. Turk. J. Vet. Anim. Sci. 26, 1260–1270.