Neurodegeneration with Brain Iron Accumulation

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Abstract

The term NBIA encompasses a heterogeneous group of inherited disorders characterized clinically by progressive extra pyramidal syndrome and pathologically by excessive iron deposition in brain, primarily affecting the basal ganglia (globus pallidus mainly). The hallmark of this syndrome is the age specific phenotypic presentation and intraphenotypic heterogeneity. NBIA are at present include ten subtypes with genes identified in nine subtypes. They form an important differential diagnosis for the phenotype of global developmental delay in infancy/childhood to dystonia-parkinsonism or isolated parkinsonism at all ages and also for the isolated cranio cervical dystonia of adult onset. There needs to be a high index of clinical suspicion for this syndrome and the evaluation includes MRI brain T2* weighted imaging which reveal symmetrical iron deposition in bilateral globus pallidi and other basal ganglia. The T2 * imaging pattern of iron deposition varies amongst the different subtypes and the combination of clinical phenotype and MRI signature makes it easier to confidently make a diagnosis of NBIA and to recommend genetic testing. The treatment to date is mostly symptomatic with targeted therapies on the horizon.

Keywords: NBIA, global developmental delay, dystonia parkinsonism, parkinsonism

INTRODUCTION

Pathological accumulation of iron in basal ganglia or other areas of brain has been known to be associated with a progressive disorder of nervous system since 1920s.[1] Hallervorden–Spatz disease was in common usage for several decades. Due to advances in science that have unmasked several underlying genetic mutations in patients with brain iron accumulation[3] and also because of the links of Julius Hallervorden and Hugo Spatz Nazi Germany in 1920s, this has been replaced by neurodegeneration with brain iron accumulation (NBIA).[2–4] NBIA are a heterogeneous group of genetically determined disorders that are characterized by iron deposition in several brain areas including basal ganglia manifesting with a progressive, extrapyramidal symptoms[1] (dystonia, parkinsonism, chorea), sometimes with pyramidal signs, cognitive dysfunction, or ocular abnormalities.

NBIA demonstrate wide heterogeneity in the phenotype, not only between subtypes but also among a given subtype. NBIA are “ultrarare” with a prevalence of <1/1,000,000 affected[5] but remain a differential diagnoses in both pediatric and adult-onset neurodegenerative diseases, and most neurologists or pediatric consultants will possibly encounter a handful of cases in their lifetime. It can be a difficult condition to diagnose and this article reviews current understanding of NBIA to help clinicians with diagnosis and management of NBIA.

HOW TO DIAGNOSE?

The diagnosis of NBIA requires the following: (1) identification of the phenotypes that are associated with NBIA, (2) magnetic resonance imaging (MRI) brain [with iron detecting sequences T2*/susceptibility-weighted imaging (SWI)/gradient recalled echo] demonstrating deposition of iron in basal ganglia with or without white matter changes on T2-weighted imaging, and (3) genetic testing for identification of specific subtype of the disease.

GENETICS OF NBIA

Several clearly defined and undefined genetic defects/mutations can lead to NBIA [Table 1]. Historically, PANK2 was first to be identified,[6] but over the past 2 decades, several conditions have been identified that manifest with NBIA. Since 2004, aceruloplasminemia (described in 1987, gene identified in 2000) and neuroferritinopathy (identified in 2001) have been included in NBIA.[5] Seitelberger disease, also called infantile neuroaxonal dystrophy, was later renamed as PLA2G6-associated neurodegeneration (PLAN) is caused by mutations in phospholipase A2 gene.[6] Causative pathogenic mutation for Woodhouse–Sakati syndrome was identified as c2orf37 mutation (also called as DCAF17 mutation) in 2008.[7] The invention of high-field MRI in 1980s and later addition of sequences that could identify iron deposition in...
The common genetic causes of NBIA are summarized in Table 1, pathological features in Table 2, and pathogenesis in Table 3.

It can be debated whether accumulation of iron in the brain is the primary process responsible for the degenerative changes, or secondary to neurodegeneration. Iron deposition in brain is known to be part of normal aging and also commonly seen in MRI scans in other neurodegenerative disorders such as Parkinson's disease, multiple sclerosis, and multisystem atrophy. Contemporary understanding favors that the following factors play an important role in causing cell loss and iron accumulation: (9) (1) mitochondrial dysfunction – mitochondria are the iron sink in cells and iron recycling in them is through mitophagy and lysosomal degradation of iron-containing proteins. Altered mitochondrial fitness may hence affect iron turnover; (2) cell degeneration mediated through reactive oxygen species (specifically PANK2, PL2G6), (3) altered lipid metabolism (PANK2, COASY, PL2G6, FA2H, c19orf12), and (4) altered mitophagy and autophagy (WDR 45, ATP13A2).

Differences in the relative contribution of each of the above processes can explain the heterogeneous pathological picture and possibly explain the varying phenotypes. The clinicopathologic variability is well known in NBIA. The 10 subtypes can be further regrouped as follows: (9)

A. NBIA caused by defects in genes coding for proteins involved in lipid metabolism in lipid membrane homeostasis: pantothenate kinase-associated neurodegeneration (PKAN), PLAN, fatty acid 2 hydroxylase–associated neurodegeneration (FAHN), mitochondrial membrane protein–associated neurodegeneration (MPAN), COASY, protein–associated neurodegeneration (COPAN).

B. NBIA caused by defects in gene coding for proteins for iron metabolism: aceruloplasminemia, neuroferritinopathy.

C. Other forms of NBIA – Beta-propeller-protein-associated neurodegeneration (BPAN), Kufor–Rakeb syndrome, Woodhouse–Sakati syndrome.

**Phenotype–genotype Correlation of NBIA**

**NNBIA 1: PKAN**

PKAN is one of the most common forms of NBIA encountered in clinical practice. PKAN can have heterogeneous phenotypes and it can be of classic and atypical variants. (12)

**Classic form of NBIA** has onset in the first decade with gait or postural difficulty secondary to dystonia as the presenting symptom. Earlier in the course, focal (cranial or limb musculature) dystonia is common. Axial dystonia predominates as disease advances. Oromandibular dystonia is common and jaw opening dystonia can be associated with a characteristic geste “antagoniste” that resembles a praying mantis. This geste with the patient touching the chin with both hands characteristically clenched into a fist with flexion at the elbows has been called “mantis sign,” a pathognomic of PKAN. (13) Dys tonic opisthotonus or back arching has also been described as a characteristic feature of NBIA related to PANK2 and PL2G6 mutations. (14) There can be mild developmental delay and corticospinal tract involvement. Retinitis pigmentosa and oculomotor abnormalities suggestive of midbrain degeneration (like square wave jerks and poor convergence) may be present. (12) Seizures, chorea, and parkinsonism are rare. (15) Recurrent episodes of status dystonicus though rare are described in the classic variant. (12,15) The patient usually loses ambulation within 10–15 years in the classic form.

**Atypical form** is characterized by onset in the second or third decade of life (rarely in seventh and eighth decades). (16) The atypical disease has less severe dystonia and rigidity with slower progression to parkinsonism when compared with classic presentation.

Within atypical variant, four predominant movement disorder phenotypes have been described.
The dystonic phenotype has early focal limb dystonia, dystonic tremor,[17] action-induced dystonia, and oromandibular dystonia with progression to generalized dystonia. Varying degrees of spasticity are seen. Although rare, PKAN can be seen in late-onset focal dystonia, and a patient with isolated blepharospasm at the age of 55 years has been described with PKAN.[16]

The parkinsonism phenotype should be easy to pick up in young patients who have bradykinesia, rigidity, and rest tremor or gait impairment. Speech abnormalities of stuttering, palilalia, hyophonia, and spasmodic dysphonia are common and these might be the sole presenting feature or part of the early disease in a teenager or young adult. The parkinsonism can be levodopa responsive.[12,17]

The other phenotypes that have been reported include isolated drug responsive freezing of gait,[18] a choreic phenotype with senile chorea-like presentation,[19] motor tics and vocal tics,[20] altered sleep architecture,[21] and HARP syndrome (hypobetalipoproteinemia, acanthocytosis, retinitis pigmentosa, and pallidal degeneration).[22,23]

| Gene        | Gross morphology                                                                 | Iron                                                                 | Axonal spheroids          | Lewy body pathology | Tau pathology                                      | Gliosis                  |
|-------------|----------------------------------------------------------------------------------|----------------------------------------------------------------------|---------------------------|---------------------|---------------------------------------------------|--------------------------|
| PANK2       | Neuronal loss in GP, reduced myelin, normal SN                                   | GP neurons, glia, microglia macrophages, perivascular and iron dusting | GP Ubq, APP, NF           | No                  | Occasional tangles and threads                     | GP and widespread        |
| COASY       | Proposed similar to PANK2                                                        |                                                                      |                           |                     |                                                   |                          |
| C9orf12     | Neuronal loss in GP, reduced myelin, GP, and SN atrophy                          | GP neurons, macrophages, less in astrocytes                          | Ubq in cortex, GP, SN, caudate, putamen, brainstem | Severe Lewy bodies and Lewy neurites, GP, SN, cortex, striatum | Rare tau                | Widespread               |
| PLA2G6      | Cerebellar, cortical, GP and brain stem atrophy, SNPr particularly affected. Cerebellar granular cells >purkinje cells | GP and sparse in SNPr                                                  | Severe p-NF, Ubq, APP, alpha-syn | Severe Lewy bodies and Lewy neurites. Ubq, alpha-syn | Early-onset hyperphosphorylated tau inclusions, threads, and tangles in cortex | Variable                 |
| FA2H        | Proposed brainstem atrophy and demyelination                                     | Proposed white matter lesions and enlarged axons                      |                           |                     |                                                   |                          |
| WDR45       | SN >GP, neuronal loss, cerebellar atrophy, purkinje and granule layer, cortical atrophy | Strongest in SN, also GP and glia                                     | GP, SN plus thalamus      | No                  | Tau tangles, hippocampus, cortex, putamen, few in atrophied SN and GP |                          |
| ATP13A2     | Proposed similar to PANK2                                                        | Neuronal loss in cerebellum, GP >SN, cortex, perivascular and astrocytic terminals | Iron-laden globular structures in glia and variable in neurons | Unknown             | Unknown                                           | Yes                      |
| DCAF17/c2orf37 | Proposed similar to PANK2                                                        | Neuronal loss in cerebellum, GP >SN, cortex, perivascular and astrocytic terminals | Yes, GP, Ubq, and NF. Ferritin and iron-laden inclusions in glia >neurons GP, putamen, thalamus, and cerebellar cortex | Unknown             | Few                                               | Yes, but some atrophy too |
| CP          | Severe neuronal loss in putamen, dentate nucleus. Neuronal atrophy and iron in visceral organs | Neuronal loss in cerebellum, GP >SN, cortex, perivascular and astrocytic terminals | Iron-laden globular structures in glia and variable in neurons | Unknown             | Unknown                                           | Yes                      |
| FTL         | Mild atrophy in the cerebellum, cortex, putamen. GPe and SN mildly affected      | Cerebellum and putamen, glia, perivascular, and perineuronal          | Yes, GP, Ubq, and NF. Ferritin and iron-laden inclusions in glia >neurons GP, putamen, thalamus, and cerebellar cortex | Unknown             | Few                                               | Yes, but some atrophy too |

APP: Amyloid precursor protein; GPe: Globus pallidus externa; GPi: Globus pallidus interna; NF: Neurofilament; SNc: Substantia nigra pars compacta; SNPr: Substantia nigra pars reticulata; Ubq: Ubiquitin; alpha syn: Alpha synuclein
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| Genes coding for proteins helping lipid metabolism and membrane homeostasis | Effect of gene mutation |
|--------------------------------------------------------------------------|------------------------|
| PKAN                                                                     | Defective enzymatic activity, accumulation of N-pantothenyl cysteine, free cysteine, chelates iron which accumulates.[9,10] |
| COPAN                                                                    | Decreased/absent activity of this enzyme hampers synthesis of coenzyme A and lipid synthesis, mitochondrial dysfunction.[9] |
| PLAN                                                                     | Defective PLA2G6 activity, inability to repair the oxidized and damaged phospholipids. So axonal damage ensues leads to axonal degeneration.[16] |
| FAHN                                                                     | Hydroxy fatty acids are precursor to ceramide synthesis, ferritin is known to associate with myelin, and faulty myelin causes iron accumulation.[20] |

| Genes coding for proteins of iron metabolism                              | Effect of gene mutation |
|--------------------------------------------------------------------------|------------------------|
| Mutation in the gene encoding ceruloplasmin                              | Absent ceruloplasmin, reduced export of iron, so accumulation of extracellular transferrin free iron, oxidative damage and degeneration. |
| Mutation in the ferritin light chain                                     | Variant ferritin causes unsafe iron storage, increased cytosolic free iron and reactive oxygen species formation, degeneration |

| Other forms of NBIA                                                       | Effect of gene mutation |
|--------------------------------------------------------------------------|------------------------|
| WDR45                                                                    | Encoding of a truncated or destabilized WDR 45, disrupted autophagy, impaired elimination of abnormal and toxic protein, cellular stress and death. |
| ATP13A2 is a lysosomal P-type ATPase divalent cation transporter Mutation in DCAF17 (also called c2orf37)[7] | Its mutation leads to the disruption of autophagy.[11] |
|                                                                           | It is a nucleoprotein and exact function is not known. |

**Table 3: Pathogenesis of NBIAs**

**NBIA 2: PLAN**

This syndrome has a continuum of three age distinct but overlapping phenotypes[24-26] of classic infantile neuroaxonal dystrophy, atypical neuroaxonal dystrophy of childhood onset including Karak syndrome and PLA2G6-related dystonia parkinsonism (PARK 14) of adult onset.

**Classic infantile neuroaxonal dystrophy (NBIA 2a)** is characterized by dystrophic axons found on nerve biopsy and is a devastating syndrome of neurodevelopmental regression. It accounts for 85% of the PLAN cases.[26] The onset of disease is between 6 months and 2 years of age, with axial hypotonia, psychomotor retardation which progresses to gait disturbances (due to ataxia or postural instability).[27] The axial hypotonia is unique to this variant of PLAN and is not seen in other variants. Strabismus, nystagmus and optic atrophy, and progressive pyramidal dysfunction with bulbar involvement[27,28] are seen as disease advances. Generalized seizures have been reported.[29] Many patients die in the first decade.

**Atypical neuroaxonal dystrophy (NBIA 2b)** has a childhood onset between 1 and 6 years of age.[15] The presentation is with gait instability (ataxia or postural instability) with dyspraxia and speech regression with two-thirds of patient showing optic atrophy. The disease progresses with spastic tetraparesis, nystagmus, seizures, dystonia, dysarthria, and cognitive disturbances with neuropsychiatric features.

**Karak syndrome** described by Mubaidin et al.[30] in 2003 is similar to atypical childhood-onset PLAN and is due to homozygous missense mutation in PLA2G6 in 2006,[31] so it is grouped under atypical neuroaxonal dystrophy (NAD) category in 2009.

**Adult-onset PLAN/PLA2G6-associated dystonia parkinsonism/ PARK 14I.** This is an adolescent or adult-onset condition with subacute levodopa dystonia (blepharospasm, foot dystonia as presenting symptoms) parkinsonism (rigidity and bradykinesia seen in majority), cognitive decline, oculomotor abnormalities, psychiatric features, and pyramidal signs.[32] Cerebellar symptoms and signs are generally absent. Autonomic dysfunction (bladder or bowel disturbances) if present can be an important clue to the diagnosis of PLAN as this is not a feature of other NBIA subtypes. Neuropsychiatric features can be a presenting feature. The oculomotor disturbances like supranuclear gaze palsy, slow saccades, fragmented saccades, nystagmus, and apraxia of eyelid opening can be seen.[33] The parkinsonism is levodopa responsive and patients can develop levodopa-induced dyskinesia.[27,34,35]

**NBIA 3: Neuroferritinopathy**

This is the only autosomal dominantly inherited subtype of NBIA caused by mutation in ferritin light chain.[36] Unlike other common NBIA, the onset of neuroferritinopathy is in midlife.[37] The phenotype is of slowly progressive movement disorder with subtle cognitive deficits. The movement disorder has a focal onset chorea (39.7%), lower limb dystonia (38.5%), parkinsonism (6%), tics (1.2%), tremor (7.2%), and cerebellar ataxia (4.2%).[38] There is striking asymmetry throughout the disease.[37] The triad of oromandibular dyskinesia (65%) and impairment of voice and speech (dysarthrophonia)
with action-specific facial dystonia (63%) is specific to neuroferritinopathy. Pyramidal dysfunction, oculomotor abnormalities (slow saccades, apraxia of eyelid opening, limitation of vertical eye movements), sleep disturbances, vibration, and proprioception loss can develop later. Frontal executive dysfunction may develop early or late. A case of adult-onset isolated craniocervical dystonia with later development of parkinsonism identified as NFT has also been described.\[6\]

**NBIA 4: MPAN**

This is an autosomal recessive NBIA described by Hartig et al. caused by mutations in c19orf12 gene.\[40\] The mean age of onset is 11 years.\[41\] The condition is characterized by progressive spastic paraplegia, parkinsonism unresponsive to L-DOPA treatment, and psychiatric or behavioral symptoms, with variable optic atrophy and motor axonal neuropathy. The onset of the disease is similar to that seen in PKAN and PLAN with gait or speech abnormalities, focal dystonia restricted to feet and hands, and oromandibular dystonia, with early pyramidal dysfunction (lower limbs more affected than upper limbs).

Motor axonal neuropathy has been observed in 39%-44% of the patients.\[40,41\] In advanced disease, progressive muscle atrophy makes the differentiation from atypical PKAN and PLAN easier. Optic atrophy has been reported in 74% of patients.\[41\] Cognitive decline is very common with inattention, hyperactivity, emotional lability, depression, and stereotypic hand movements.

**NBIA 5: BPAN**

This X-linked dominant disease previously called as SENDA has been renamed as BPAN after the causative gene mutation in WDR45 gene was identified in 2012.\[42\] There are two phases of disease progression in SENDA. In childhood, there is generally delayed speech and motor development with ataxic gait with preserved gait stability till adolescence/adulthood usually in late adolescence.

In advanced disease, there is marked neurocognitive status with rapid development of parkinsonism, dystonia, and cognitive decline. Seizures can be seen in childhood but are rare later in the course. The parkinsonism is levodopa responsive with early development of motor fluctuations and disabling dyskinesias. The dystonia starts in the upper limbs with possible development of camptocormia. There is rapid progression of disease with spastic paraparesis and dysphagia. Hypersomnia with shortened sleep onset latency on multiple sleep latency test (MSLT) and abnormal rapid eye movement (REM) sleep can be seen. A characteristic dance-like movement of extremities with onset of sleep has been noted. A Rett’s syndrome-like phenotype with repetitive midline hand wringing movements has been reported and is one of the important differentials for atypical Rett’s phenotype. In advanced stages, patients are bed-bound and profoundly demented. Colobomata, astigmatism, myopia, and loss of pupillary ruff are the ocular abnormalities reported.

**NBIA 6: COPAN**

A homozygous mutation of COASY, which encodes for coenzyme A synthesis, causes this condition.\[44\] The cases reported in literature had normal development till the age of 2 years but later developed gait abnormality which progressed to spastic quadriplepsis with dystonia parkinsonism. Oromandibular dystonia, cognitive decline, and mild axonal neuropathy have been described in the patients with this NBIA.

**Aceruloplasminemia**

Aceruloplasminemia is caused by mutation in ceruloplasmin gene with an average age at diagnosis of 51 years. A clinical triad of retinal degeneration, diabetes mellitus, and neurologic symptoms/signs and anemia predates the development of diabetes and neurological symptoms. In a recent review,\[46\] the most common presenting symptom was cerebellar dysfunction in 71%, hyperkinetic movement disorder in 64% (dystonia-blepharospasm and oromandibular dystonia, tremor, chorea), parkinsonism in 20%, and cognitive dysfunction in 60% (apathy and memory impairment).

**Fatty acid 2 hydroxylase–associated neurodegeneration**

The mutation in the gene encoding for fatty acid 2 hydroxylase was identified in 2010.\[47\] The phenotype is similar to that of infantile PLAN and many other leukodystrophies with onset in the first decade of gait impairment and falls progressing to spastic quadriplepsis, dystonia, cerebellar dysfunction, variable optic atrophy, divergent strabismus, and seizures. Most of the patients became wheelchair-bound by adolescence.\[47\] Axonal neuropathy was reported in a single family.\[48\]

**Kufor–Rakeb syndrome**

This NBIA disorder was described in 1994 from Jordan.\[49\] The chromosome 1p locus was designated as PARK9 locus for the monogenic parkinsonian phenotype. The causative compound heterozygous gene mutation was identified in 2006 in lysosomal ATP13A2. It is a syndrome of juvenile-onset parkinsonism, spasticity, supranuclear gaze palsy,\[25\] and cognitive features (visual hallucinations and dementia). Oculogyric dystonic spasms, facial-faucial-finger mini-myoclonus and autonomic dysfunction are variably associated.

Parkinsonism is initially responsive to levodopa with early development of disabling dyskinesias and hallucinations. Dystonia develops in 50% of the cases later in the disease course.\[48\] Cerebellar features are rare, but there is marked olfactory dysfunction. In the first ever case of Kufor–Rakeb syndrome reported from India recently, a novel nonsense mutation in ATP13A2, clinically he had no cognitive dysfunction, myoclonus, or hallucinations and a slow clinical progression.\[50\]

**Woodhouse–Sakati syndrome**

This is a rare neuroendocrine disorder described in 1983. The causative gene mutation was identified in c2orf37 gene in 2008.\[7\] The clinical presentation is dysmorphic facies (alopecia, high forehead, malocclusion), hypogonadism, diabetes mellitus, mental retardation, sensorineural deafness,
and extrapyramidal features. Seizures, polyneuropathy, thyroid dysfunction, keratoconus, and syndactyly of hand or feet have been described in some of the cases. Thrombocytopenia has been reported in a family of three affected siblings from India.\textsuperscript{[53]} The extrapyramidal features and diabetes may not develop till late teens or early adulthood. The movement disorder phenotype is reported only in 50% cases as a combination of focal onset chorea and dystonia with progression and later gait difficulty and immobilization. Eye movements are normal. Deafness and cognitive decline or mental retardation is seen in 75% of patients. This syndrome forms an important differential diagnosis for deafness dystonia phenotype from Mohr–Tranebaerg disease.

**Leukoencephalopathy with dystonia and motor neuropathy – SCP2 mutations**

A new NBIA phenotype?

There are two case reports of this condition in the literature to date. The causative mutation was identified to be in the gene encoding sterol carrier protein leading to its deficiency. It was first described in 2006 in a 45-year-old male with a 28-year history of spasmodic torticollis, spinocerebellar ataxia, and motor neuropathy.\textsuperscript{[52]} MRI brain showed no iron deposition of basal ganglia, but a leukoencephalopathy and hyperintense thalamus and pons (butterfly-like lesions) with no iron deposition. In 2015, the second case was reported of a 55-year-old male with a 20-year history of spinocerebellar ataxia, gait disturbance, and deafness with no extrapyramidal disturbances,\textsuperscript{[53]} but MRI brain demonstrated iron deposition and no leukoencephalopathy.

**GTPBP2 mutations and NBIA**

A family of three siblings\textsuperscript{[54]} have been described with this mutation presenting with delayed developmental milestones and later developed action dystonia of hands and feet, ataxia motor neuronopathy, and cognitive decline (low IQ and neuropsychiatric disturbances). MRI brain of all the three siblings demonstrated hypointense globus pallidi and

| NBIA subtype                          | Genetic mutation | Radiology (MRI brain with T2/T2*/SWI sequences)                                                                 |
|---------------------------------------|------------------|---------------------------------------------------------------------------------------------------------------|
| PKAN                                  | PANK2            | Eye of the tiger appearance in GP, mid hypointensity in SN [Figure 1a]                                        |
|                                       |                  | The eye of the tiger appearance may fade as disease advances. It is neither pathognomonic nor specific to PKAN as it is found in COPAN, neuroferritinopathy, and non-NBIA disorders such as multiple sclerosis, multisystem atrophy, and carbon mono oxide poisoning\textsuperscript{[57]} |
| PLAN                                  | PLA2G6           | Cerbellar and vermal atrophy with callosal thinning and vertical orientation, calval hypertrophy               |
| Infantile neuroaxonal dystrophy       |                  | Inconsistent iron deposition in GP, SN\textsuperscript{[33]}                                                                                      |
| Atypical neuroaxonal dystrophy        |                  | Cerbellar atrophy and callosal changes in some cases                                                         |
| PLAN DP/PARK 14                       |                  | Inconsistent iron deposition in GP, SN. Cerebellar T2 white matter hyperintensities                           |
| Neuroferritinopathy                   | Ferritin light chain | Hypointense caudate, putamen, thalamus, GP, SN, and red nucleus                                               |
| MPAN                                  | C19orf12         | GP and SN hypointense with hyperintense streaking of medial medullary lamina of GP                           |
| BPAN                                  | WDR45            | Cortical and cerebellar atrophy                                                                               |
| COPAN                                 | COASY            | “Halo” in SN (T1 hyperintensity) with T2 GP and cerebral peduncles                                             |
| Aceruloplasminemia                    | Ceruloplasmin    | Eye of the tiger                                                                                              |
|                                       |                  | Hypointense SN, GP with swelling of caudate and putamen                                                        |
| FAHN                                  | Fatty acid 2 hydroxylase | Hypointense caudate, putamen, thalamus, GP, SN, and red nucleus                                           |
|                                       |                  | Hypointensities in cerebellum                                                                                  |
| Kufor-Rakeb syndrome                  | ATP13A2          | GP more hypointense than SN                                                                                   |
|                                       |                  | Confluent subcortical and periventricular cerebral T2 white matter hyperintensities                        |
|                                       |                  | Atrophy of cerebellum, medulla and spinal cord\textsuperscript{[3]}                                          |
| Woodhouse-Sakati syndrome             | DCAF17/c2orf37   | Hypointense GP, CAUDATE, and putamen                                                                          |
|                                       |                  | Cerebellar brainstem and pyramidal atrophy                                                                    |
| Leukoencephalopathy with dystonia and motor neuropathy | SCP2          | Hypointense GP and SN with widespread and confluent cerebral T2 white matter hyperintensities         |
| New NBIA                              | GTPBP2           | Hypointense GP and SN with cerebellar vermain atrophy                                                         |

NBIA: Neurodegeneration with brain iron accumulation; MRI: Magnetic resonance imaging; SWI: Susceptibility-weighted imaging; PKAN: Pantothenate kinase-associated neurodegeneration; GP: Globus pallidus; SN: Substantia nigra; PLAN: PLA2G6-associated neurodegeneration; MPAN: Mitochondrial membrane protein-associated neurodegeneration; BPAN: Beta-propeller-protein-associated neurodegeneration; COPAN: COASY protein-associated neurodegeneration; FAHN: Fatty acid 2 hydroxylase-associated neurodegeneration.
substantia nigra on T2* imaging with cerebellar vermiculotomy. The causative mutation was in the GTPBP gene encoding for GTP-binding protein 2 (GTP superfamily are enzymes involved in cell proliferation and differentiation, intracellular transport).

**Radiological features of NBIA**

MRI brain (T1, T2, T2*, and FSE) sequences are very useful for the diagnosis and to differentiate between various NBIA syndromes [55][56] [Table 4].

Within the brain parenchyma, there are certain areas that are inherently rich in iron – globus pallidus, substantia nigra, red nucleus, dentate nucleus, putamen, and thalamus. With normal aging, there is an increased iron deposition within basal ganglia and dentate nuclei.

In NBIA, there is symmetric distribution of excess iron accumulation in the iron-rich areas of the brain mentioned above. Iron appears isointense on T1-weighted and hypointense on T2-weighted imaging. Calcium also appears isointense on T1 and hypointense on T2, but computed tomography (CT) scan differentiates the two as calcium is hyperintense than the surrounding brain parenchyma on CT and iron is isointense.

The MRI workup of NBIA is incomplete without T2*-weighted acquisitions and/or SWI imaging. They enhance this degree of hypointensity (described as “blooming”). All NBIA subtypes reveal iron deposition in globus pallidi on MRI but differ in the co-occurrence of other findings.

**Approach to a patient with suspected NBIA**

Usually in a pediatric or neurology/movement disorders clinic, a clinician encounters a patient with a mixed bag of neurological symptoms/signs and an abnormal MRI brain showing iron deposition [Flowcharts 1 and 2]. Sometimes the report from a radiologist can state NBIA with classic features like “eye of a tiger sign” that can ease the diagnostic burden. More commonly, we as clinicians will have to unjumble the puzzle with a revisit and look for more clinical clues that facilitate diagnosis. Genetic diagnosis is not always possible as most of the mutations have been described in a small group of patients. A detailed clinical history and examination can be very helpful. It is not always easy to get the correct age of onset, course, and progression. One should look for associated features such as retinitis pigmentosa, optic atrophy, peripheral neuropathy, seizures, cognitive decline/neuropsychiatric features, cerebellar features, and endocrine abnormalities.

The next step is to investigate toward the specific subtype with laboratory, electrophysiological, and MRI of brain with iron-specific sequences.

Laboratory investigations include hemogram for anemia, serum and urine copper levels, serum ceruloplasmin, serum ferritin, iron levels, blood sugar, and hormone level testing for hypogonadism where appropriate. Neuroferritinopathy has low serum ferritin levels with all other parameters being normal, whereas aceruloplasminemia has low or absent serum ceruloplasmin, elevated ferritin, low iron, and low serum copper but normal urinary copper and raised blood sugar levels. Serum hormonal assays are to be done in suspected cases of Woodhouse–Sakati syndrome.

Electroencephalogram can be helpful if seizures are in question as in PLAN Infantile neuroaxonal dystrophy (INAD) and atypical NAD (but not in PLAN-DP), FAHN, BPAN, and Woodhouse–Sakati syndrome. Electromyography studies should be done to look for sensorimotor neuropathy in case neuropathy is suspected as in PLAN and motor neuronopathy/axonopathy in MPAN. Sleep analysis can help in investigating and managing reduced sleep time insomnia and sleep apnea. Abnormal REM sleep with decreased sleep-onset latency
Flowchart 2: Approach to an adult with suspected NBIA

Genetic testing can confirm the diagnosis, but not all NBIA cases will have either of the mutations described and there are still some unknown NBIA cases. The advancement of genetic diagnosis and whole genome sequencing has meant that sometimes one may see a patient in clinic for the first time who comes with a genetic diagnosis. This is not easy to interpret genetic results in an asymptomatic patient or in someone with minimal signs. It is important to understand that not all mutations in the known genes causing NBIA are pathogenic, so one must be careful in interpreting genetic results. OMIM usually lists commonly known mutations but one might want to seek help from an expert neurogeneticist who has a better understanding of these mutations and have access to larger genetic databases to confirm in case of a doubt about pathogenicity of a newly found mutation with minimal or no clinical signs. MRI can help as well to look for changes in asymptomatic patients with these mutations.

**Management**

Although there are some trials currently underway for specific treatments, the management of NBIA to date remains
symptomatic. A multidisciplinary approach to therapy is recommended with a combination of medical, surgical, and good supportive/nursing care/physiotherapy. Medical therapy for dystonia with trihexyphenidyl and spasticity with benzodiazepines and baclofen can be tried, but the response is variable and not sustained.[15] Botulinum toxin therapy is particularly useful for drug-resistant dystonia especially oromandibular dystonia and salivary gland toxin injections for drooling in advanced cases. In patients with NBIA and parkinsonism, levodopa trial is always justified. Parkinsonism in PKAN is not levodopa responsive; whereas in PLAN (childhood-onset and dystonia parkinsonism), MPAN, BPAN, and Kufor–Rakeb syndrome respond initially but early development of disabling dyskinesias and hallucinations can be seen. Dopamine agonists should be used very cautiously in NBIA, especially with neuropsychiatric or cognitive problems as evidence of success is limited.

A preliminary study of deferiprone, an oral iron chelator, showed robust reduction in brain iron on brain MRI in patients with PKAN, but no measurable benefit in clinical disease outcomes.[69]

Intrathecal baclofen pump can be used for symptomatic management of dystonia/spasticity where oral medications have failed or are not advisable in view of dysphagia. Of recent interest is a novel intraventricular baclofen delivery in nine children which included one PKAN patient,[60] but additional studies are required.

Deep brain stimulation has been offered to very few patients with NBIA and there are insufficient data in literature to guide usage of deep brain stimulation (DBS). This can be reviewed with local teams with expertise in DBS management of dystonia and Parkinson’s on a case-to-case basis. In a patient with PKAN, some improvement was noted after bilateral globus pallidus interna (gpi) DBS (improvement in Burke-Fahn-Marsden Dystonia Rating Scale (BFMDRS)). It was not persistent and the benefit diminished with time.[19] The dystonic storm of patients with classical PKAN is resistant to drugs and Gpi DBS has been reported to be life-saving.[61]

Good supportive care with a multidisciplinary team including physiotherapy, occupational therapy psychology support, speech and language therapy, and nursing care remains the mainstay of management of patients with NBIA.

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Conflicts of interest
There are no conflicts of interest.

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