Giant cell hepatitis with autoimmune hemolytic anemia in a nine month old infant

Jihene Bouguila, Sameh Mabrouk, Samia Tilouche, Dajla Bakir, Amel Trabelsi, Amel Hmila, Lamia Boughammoura

Department of Paediatrics, Hospital Farhat Hached, 4000 Sousse, Tunisia
Department of Paediatrics, Hospital Sahloul, 4011 Sousse, Tunisia
Department of Radiology, Hospital Farhat Hached, 4000 Sousse, Tunisia
Laboratory of Anatomopathology, Hospital Farhat Hached, 4000 Sousse, Tunisia

Author contributions: Bouguila J contributed to this work; Mabrouk S, Tilouche S, Hmila A and Boughammoura L designed the research; Bakir D performed radiological assessment; Trabelsi A analyzed the histological exams.

Correspondence to: Dr. Bouguila Jihene, Department of Paediatrics, Hospital Farhat Hached, City of Ibn El Jazzar, 4000 Sousse, Tunisia. jbouguila@yahoo.fr
Telephone: +216-98-407764 Fax: +216-73-226702
Received: August 8, 2012 Revised: September 25, 2012 Accepted: November 17, 2012 Published online: April 27, 2013

INTRODUCTION

Giant cell hepatitis (GCH) associated with autoimmune hemolytic anemia (AHA) is a rare individualized entity, particularly affecting infants, with an unknown pathogenesis and poor outcome [1,2]. Only 27 cases have been reported in pediatric reviews [2-6]. It usually presents as a severe hepatitis, jaundice and fever that begins about 1 year of age and is associated with AHA with a positive direct Coombs’ test [3]. In this article, we describe a new case in a nine month old infant who presented with pallor, fever and jaundice and whose outcome was severe in spite of early treatment. We also review literature data concerning clinical presentation and therapeutic modalities of this rare entity.

CASE REPORT

A 9-mo old infant with no pathological medical history presented with jaundice, fever and watery diarrhea for the last 4 d. Physical examination showed a eutrophic,
febrile (38.3 °C) infant. Jaundice was obvious with pallor, hepatomegaly (liver span of 11 cm) and enlarged spleen. Laboratory features (Table 1) showed a bicitopenia: normochromic normocytic regenerative anemia and thrombocytopenia, associated with hepatic insufficiency, excretion and cholestasis with elevated conjugated bilirubin and gamma glutamyl transpeptidase. Alpha fetoprotein (AFP) was high as well.

With this association of febrile jaundice with liver injury, infectious causes were first suspected. Serologies of hepatitis A, B and C, of cytomegalovirus, Epstein Barr virus, herpes simplex virus and human immunodeficiency virus were all negative. A metabolic cause, particularly tyrosinemia, was also evoked given the elevated levels of free tyrosine and elevated conjugated bilirubin and gamma glutamyl transpeptidase. Alpha fetoprotein (AFP) was high as well.

Table 1 Biological findings before and after treatment (prednisone + azathioprine)

|                      | J1 of hospitalization | J10 of hospitalization | J17 of treatment | J60 of treatment |
|----------------------|------------------------|------------------------|------------------|------------------|
| Leucocytes (elements/μL) | 12400                  | 19800                  | 23300            | 27800            |
| PNN (%)              | 77                     | 42                     | 60               | 57               |
| Hemoglobin (g/dL)    | 9.3                    | 6.8                    | 11.4             | 10.4             |
| MCV (μmol/L)         | 79.9                   | 86.6                   | 85.4             | 96.6             |
| Reticulocytes (elements/μL) | 114000             | 17600                  | 92000            | 184000           |
| Platelets (elements/μL) | 71000                  | 7000                   | 153000          | 152000           |
| SAT/ALT (U/L)        | 1020/810               | 1430/1886              | 368/600          | 1310/522         |
| Total bilirubin (μmol/L) | 101                   | 425                    | 190              | 425              |
| Direct bilirubin (μmol/L) | 82.6                  | 122                    | 78               | 291              |
| PT (%)               | 55.1                   | 51.7                   | 70               | 20               |
| CRP (mg/L)           | 42                     | -                      | 12               | 20               |
| AFP (ng/mL)          | 179                    | -                      | 29               | 29.63            |
| LDH (U/L)            | 2089                   | 1092                   | 400              | -                |
| Haptoglobin (g/L)    | 0.6                    | 0                      | -                | -                |
| DCT                  | + à IgG/C3d            | + à IgG/C3d            | + à IgG/C3d      | + à IgG/C3d      |
| Ferritinemia (ng/mL) | 163                    | -                      | -                | -                |
| Fibrinogen (g/L)     | 3.3                    | -                      | -                | -                |
| Triglycerides (mmol/L) | 3.99                   | -                      | -                | -                |

PNN: Polynuclear neutrophil; MCV: Mean corpuscular volume; SAT: Serum aspartate transaminase; ALT: Serum alanine aminotransferase; GGT: Gamma glutamyl transpeptidase; PT: Prothrombin time; CRP: C reactive protein; AFP: Alpha fetoprotein; LDH: Lactate deshydrogenase; DCT: Direct Coombs’ test; IgG: Immunoglobulin G.

Detected in both urine and blood cultures. This infection was handled with adequate intravenous antibiotics. Abdominal ultrasonography showed an enlarged hyper-echogenous liver, a homogenous splenomegaly, normal non dilated biliary ducts and the presence of a hyper-echogenous cuff surrounding the hepatic pedicle.

Initially, there was no positive progression as fever persisted and both jaundice and pallor worsened. At day 10 of hospitalization, hemoglobin decreased down to 6.8 g/dL (normocytic normochromic regenerative anemia) and platelets were at 6000 elements/μL. Haptoglobin was null and direct Coombs’ test was positive (IgG and C3d). The association of hepatic insufficiency and AHA in a young infant on one hand and the absence of auto-antibodies on the other hand, made us consider the diagnosis of GCH. Thus, a liver biopsy was performed after blood and platelet transfusions and the patient’s stabilization.

Histological analysis confirmed the diagnosis; there was a diffuse transformation of hepatic cells into giant cells with areas of necrosis (Figure 1).

The patient first received an intravenous immunoglobulin course at the dose of 1g/kg per day during 2 d, then both steroids (prednisone: 2 mg/kg per day) and imunosuppressive [Azathioprine (Imurel®): 1 mg/kg per day] therapies were started.

The immediate course was favorable, with clinical as well as biological improvement since day 17 of steroid therapy (Table 1). At day 30 of hospitalization, the infant was discharged from hospital under the same treatment. He was seen 15 d later with stable biological data.

However, he was hospitalized again 60 d after the onset of the therapy, with fever, edema and ascites. We found a urinary tract infection with identification of \( E. Coli \), hepatic insufficiency [prothrombin time (PT) = 20%] and aggravation of the cytolysis (Table 1). The patient received intravenous antibiotics. Liver insufficiency was handled with symptomatic measures. In spite of this management, fever persisted, renal failure appeared and liver function worsened (PT = 12%). The infant died with septic shock. Follow up had lasted 3 mo since the beginning of symptoms.

Figure 1 Liver biopsy showing a diffuse transformation of hepatocytes into giant cells with necrosis areas.
DISCUSSION

GCH associated with AHA was described in 1981 in young children with a severe presentation and high mortality (39% of reported cases)\[^{[4]}\]. This pathological entity is rare\[^{[1,2,3]}\]. 27 cases have been reported in pediatric reviews\[^{[4,5]}\]. In 2011, an Italian pediatrician’s team reported the biggest series of cases of this pathology. They described 16 children during a 28-year period\[^{[4]}\].

The pathogenesis of this entity is still unknown\[^{[1]}\]. Several authors have suggested an autoimmune origin, especially in the presence of AHA and hypergammaglobulin levels\[^{[1]}\].

According to Maggiore et al.\[^{[4]}\], elements suggesting an autoimmune origin among their 16 patients were a positive family history of autoimmune conditions, such as type 1 diabetes, thyroiditis and psoriasis, positive findings for auto-antibodies in some patients, thrombocytopenia, the improvement under immunosuppressive therapy and the decline when tapering doses. However, this hypothesis is not approved by other authors since auto-antibodies are often absent and there is no typical histological evidence of autoimmune hepatitis\[^{[5]}\]. Another hypothesis suggests that the underlying mechanism could be a non controlled release of cytokines by activated T lymphocytes as well as Kupffer cells\[^{[4]}\]. In the present case, we found neither family nor personal history of autoimmune disease; however, there was an initial positive response to steroids as well as immunosuppressive therapy.

GCH associated with AHA specifically occurs in young infants; the first signs often begin between 2.5 and 24 mo of age\[^{[5]}\]. In our patient, the first signs were seen at 9 mo. Hepatocyte transformations into giant cells can happen during the neonatal period and is considered a non specific reaction of immature hepatocytes to various aggressions\[^{[4]}\].

Each time hepatitis is associated with AHA with positive Coombs’ test, either with IgG or complement, in an infant, liver biopsy must be performed as soon as possible in order to confirm the diagnosis of GCH so that early treatment can be started\[^{[4]}\]. The prognosis of this pathology is often poor\[^{[7]}\]. Our patient had an early relapse and died of septic shock.

Treatment possibilities include steroids and/or azathioprine, cyclosporine, cyclophosphamide, tacrolimus, intravenous immunoglobulins, mercaptopurine, mycophenolate mofetil, vincristine, plasmapheresis and anti CD20 (Rituximab)\[^{[5]}\]. Splenectomy was also proposed in some patients as an alternative for AHA resistant to medicines. Anti CD20 were successfully used in 2 patients and splenectomy was performed in 5 cases/10; only 2 of them got positive results\[^{[4]}\].

In this series, 4 patients died because of severe sepsis and post transplantation, respectively in 3 and 1 cases. The other patients (12 cases) are alive and one underwent liver transplantation\[^{[4]}\]. This series illustrates the severity of this pathology and the difficulties of treatment, given the high risk of relapse as well as therapy resistant AHA after relapse. Liver transplantation was reported in 6 patients in the literature; 3 cases relapsed after transplantation\[^{[4]}\].

In our patient, we started therapy by an intravenous course of immunoglobulins followed by the association of prednisone and azathioprine with partial response (improvement without normalization of transaminases). Relapse was rapid (less than 4 mo) with severe hepatic insufficiency but without recurrence of hematological disorders. Our patient died due to septic shock.

In conclusion, GCH associated with AHA is a severe pathological entity. This diagnosis should be evoked when hepatitis of unknown origin occurs in an infant and will be confirmed by liver biopsy. Early treatment associating corticotheraphy and immunosuppressive drugs with sufficient doses is essential to reach total remission with normal transaminases. Treatment must be maintained as long as possible in order to avoid relapses which are more resistant to therapies.

REFERENCES

1. Vajo P, Migliaro F, Ruggeri C, Di Cosmo N, Crispino G, Caropreso M, Vecchione R. Life saving cyclophosphamide treatment in a girl with giant cell hepatitis and autoimmune haemolytic anaemia: case report and up-to-date on therapeutical options. Dig Liver Dis 2006; 38: 846-850 [PMID: 16266839 DOI: 10.1016/j.dld.2005.09.013]

2. Rovelli A, Corti P, Beretta C, Bovo C, Conter V, Mieli-Vergani G, Alemtuzumab for giant cell hepatitis with autoimmune hemolytic anemia. J Pediatr Gastroenterol Nutr 2007; 45: 596-599 [PMID: 18030239 DOI: 10.1097/ MPG.0b013e318033169f]

3. Bouyahia O, Mrad-Mazigh S, Boukthir S, El Gharbi-Samoud M, Les maladies auto-immunes du foie chez l’enfant. Rev Magh Pediatr 2008: 28: 59-66

4. Maggiore G, Scivere M, Fabre M, Gori L, Pacifico L, Resti M, Choulot JJ, Jacquemin E, Bernard O. Giant cell hepatitis with autoimmune hemolytic anemia. J Pediatr Gastroenterol Nutr 2011; 159: 127-132.e1 [PMID: 21349541]

5. Miloh T, Mannwani D, Morotti R, Sukru E, Shneider B, Kerkar N. Giant cell hepatitis and autoimmune hemolytic anemia successfully treated with rituximab. J Pediatr Gastroenterol Nutr 2007; 44: 634-636 [PMID: 17460499 DOI: 10.1097/MPG.0b013e31802e9739]

6. Kashyap R, Sarangi JN, Choudhry VP. Autoimmune hemolytic anemia in an infant with giant cell hepatitis. Am J Hematol 2006; 81: 199-201 [PMID: 16493610 DOI: 10.1002/ ajh.20414]

7. Hartman C, Berkowitz D, Brik R, Arad A, Elhasid R, Shamir R. Giant cell hepatitis with autoimmune hemolytic anemia and hemophagocytosis. J Pediatr Gastroenterol Nutr 2001; 32: 228
A. [Giant cell hepatitis and autoimmune hemolytic anemia: efficacy of splenectomy on hemolysis]. Arch Pediatr 1996; 3: 789-791 [PMID: 8998533 DOI: 10.1016/0929-693X(96)82162-2]

8 Choulot JJ, Parent Y, Etcharry F, Saint-Martin J, Mensire