Determination of Volatility Parameters of Secondary Organic Aerosol Components via Thermal Analysis

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Abstract: To date, there are limited data on the thermal properties of secondary organic aerosol (SOA) components. In this study, we employed an experimental method to evaluate the physical properties of some atmospherically relevant compounds. We estimated the thermodynamic properties of SOA components, in particularly some carboxylic acids. The molar heat capacity, melting point and enthalpy, and vaporization enthalpy of the samples were determined via differential scanning calorimetry and thermogravimetric analysis, and their vaporization enthalpy (\(\Delta H_{\text{vap}}\)) was estimated using Clausius–Clapeyron and Langmuir equations based on their thermogravimetric profiles. The thermodynamic properties of benzoic acid as a reference compound agree well with the reported values. The obtained specific heat capacities of benzoic acid, phthalic acid, ketopinic acid, DTAA, and 3-methylbutane-1,2,3-tricarboxylic acid (3-MBTC) are 118.1, 169.4, 223.9, 246.1, and 223.2 J mol\(^{-1}\)K\(^{-1}\), respectively. The \(\Delta H_{\text{vap}}\) of benzoic acid, phthalic acid, ketopinic acid, DTAA, and 3-methylbutane-1,2,3-tricarboxylic acid (3-MBTC) are 93.2 ± 0.4, 131.6, 113.8, and 124.4 kJ mol\(^{-1}\), respectively. The melting and vaporization enthalpies of the SOA components range from 7.3 to 29.7 kJ mol\(^{-1}\).

Keywords: volatility; specific heat capacity; vaporization enthalpy; melting temperature and enthalpy; thermal gravimetric analysis; differential scanning calorimetry; secondary organic aerosols (SOA)

1. Introduction

Atmospheric aerosol is a complex mixture of organic and inorganic compounds and has significant effects on human health, visibility, and climate [1–3]. Primary aerosol is composed of particles directly emitted to the atmosphere from various sources, such as biomass burning and vehicle exhausts [4,5]. Secondary aerosols are formed in the air via chemical reactions. Secondary organic aerosol (SOA) is formed by the oxidation of volatile organic compounds with O\(_3\), OH, and NO\(_3\) [6,7]. SOA consists of hundreds of semi-volatile compounds constantly partitioned between the particle and gas phases as a function of temperature, relative humidity, and their concentrations [8]. The physical properties of SOA are related to the contribution of the individual compounds [9]. Monoterpene (e.g., \(\alpha\)- and \(\beta\)-pinene) is a typical biogenic precursor for SOA [10,11]. SOA products from the photooxidation and ozonolysis of \(\alpha\)-pinene have been reported [12–14]. Abundant \(\alpha\)-pinene SOA constituents, such as pinic acid, oxopinonic acid, hydroxyl-pinonic acid, terpenyl acid, and pinalic acid, account for approximately 58–72% of the mass of \(\alpha\)-pinene SOA [15]. Similarly, 3-methylbutane-1,2,3-tricarboxylic acid (3-MBTC) and terpenyl
acid are products of further reactions of pinonaldehyde and nopinone at a later stage in α- and β-pinene photooxidation [16]. Diaterpenyl acid acetate (DTAA) has been observed in SOA obtained from the ozonolysis and photooxidation of both α- and β-pinene [17]. Many of these compounds are also found in the SOA of other monoterpenes species, such as d-limonene [18,19]. Polyaromatic hydrocarbons (PAHs) are common anthropogenic organics released from combustion and vehicle exhausts [20]. Aromatic carboxylic acids might be formed by the photochemical oxidation of PAHs via an oxidative ring-cleavage and can be used as a tracer of anthropogenic SOA [21,22]. Carboxylic acids contribute a substantial portion of total SOA mass [23]. Michoud et al. (2021) [24] reported that PM$_{2.5}$ from the western Mediterranean Basin (France) in 2013 contained 49% and 15% of dicarboxylic acids and tricarboxylic acids of biogenic origin, respectively. Carboxylic acids are highly soluble in water under high relative humidity, affecting the chemical and radiative properties of ambient particles [25,26].

In the past few decades, significant efforts have been made to predict ambient aerosol concentration using air quality models. These models have uncertainties in predicting atmospheric aerosol concentrations. One of the major sources of the uncertainties comes from the poor understanding of important aerosol properties such as vapor pressure and vaporization enthalpy. Chemical transport models (CTMs) such as Community Multiscale Air Quality (CMAQ), Goddard Earth Observing System (GEOS-CHEM) and Particulate Matter Comprehensive Air Quality Models (PMCAMx) are implemented with thermodynamic properties and absorptive partitioning framework [27–31]. Phase partitioning between the condensed and vapor phases governs organic aerosol concentrations in the atmosphere. Thermodynamic properties (i.e., saturation vapor pressure and vaporization enthalpy) play an important role in the temporal and spatial distribution of atmospheric aerosol between gas and particle phases [32,33]. The enthalpy of vaporization (amount of energy required to change one mole of a substance from solid to the gaseous state) and saturation vapor pressure are temperature-dependent properties. Vaporization enthalpy varies inversely, whereas saturation vapor pressure changes directly with temperature [34]. Volatility parameters of multicomponent organic aerosols need to consider the wide range of volatility associated with the multicomponent composition. However, the estimation of volatility parameters for individual SOA components is relatively easy. Gas-particle partitioning can be predicted using the composition and volatility data of individual components (such as the volatility basis set) in atmospheric OA. The partition coefficient $K_{om,i}$ in the models is inversely related to effective saturation concentration $C_{i}^{*}$ of specie $i$ (Equation (S1)). Effective saturation concentrations as a function of vaporization enthalpy and temperature can be estimated in a modified form of Clausius–Clapeyron equation (Equation (S2)) [35,36]. The functionality of carboxylic acid dramatically reduces the vapor pressure [23].

Over the last few decades, some modeling and experimental techniques have been developed to probe the volatility properties of low-volatility organic species present in atmospheric aerosols [32]. Pankow et al. (2007) [37] developed SIMPOL.1 that is a group contribution method to predict liquid vapor pressure and vaporization enthalpy of organic compounds based on their chemical structures. Similarly, Acree et al. (2003) [38] has developed a database for phase change enthalpy of organic compounds based on their associated functional groups. Ribeiro et al. (1999) [39] employed the Knudsen mass-loss effusion method to estimate the vapor pressures and enthalpies and entropies of vaporization of dicarboxylic acids. Salo et al. (2010) [9] estimated the enthalpy of vaporization for various cyclic and straight-chain dicarboxylic acids using a volatility tandem differential mobility analyzer (VTDMA). Saleh et al. (2010b) [8] estimated the enthalpy of vaporization of adipic acid and pimelic acid at 25 °C using the integrated volume method (IVM). In addition, the combination of thermal gravimetric analysis (TGA) and differential scanning calorimetry (DSC) is a well-established technique for estimating the thermal properties of chemical species [40–42]. In TGA and DSC analyses, changes in mass and heat are measured simultaneously as functions of temperature. Thermogravimetry is a convenient and rapid technique for estimating vapor pressure curves and vaporization enthalpies.
even with a small sample quantity. Booth et al. (2011) [43] reported the enthalpy of vaporization for some carboxylic acids using the Clausius–Clapeyron equation based on thermogravimetric data.

Although many studies have reported the composition of α-pinene SOA, there are limited experimental data on the thermal properties. In this study, we estimated the molar heat capacity, melting point, saturation vapor pressure, and enthalpy of melting and vaporization for biogenic SOA components, in particular, straight-chain dicarboxylic acids, cyclic dicarboxylic acids, and anthropogenic SOA components such as aromatic organic acids using an integrated TGA-DSC analysis system. This provides the thermal properties of biogenic and anthropogenic SOA species, which can serve as a good reference for yield and partitioning in chemical transport models.

2. Materials and Methods

2.1. Material

Five dicarboxylic acids (malonic acid (>99%, C$_3$H$_4$O$_4$, Sigma-Aldrich), succinic acid (≥99%, C$_4$H$_6$O$_4$, Sigma-Aldrich), adipic acid (99%, C$_6$H$_10$O$_4$, Sigma-Aldrich), and suberic acid (≥99%, C$_8$H$_{14}$O$_4$, Sigma-Aldrich)), three aromatic organic acids (benzoic acid (98%, C$_7$H$_6$O$_2$, Sigma-Aldrich), phthalic acid (98%, C$_8$H$_6$O$_4$, Sigma-Aldrich), 4-hydroxybenzoic acid (>99%, C$_7$H$_8$O$_3$, Sigma-Aldrich)), phenol (catechol (98%, C$_6$H$_4$O$_2$, Sigma-Aldrich)), six α-pinene SOA species (pinic acid (99%, C$_9$H$_{14}$O$_4$, Sigma-Aldrich), ketopinic acid (99%, C$_{10}$H$_{14}$O$_3$, Sigma-Aldrich), cis-pinonic acid (≥98%, C$_{10}$H$_{16}$O$_3$, Sigma-Aldrich), 3-methylbutan-1,2,3-tricarboxylic acid (3-MBTCA) (98%, C$_8$H$_{12}$O$_6$, Synthesized), terpenylic acid (72%, C$_8$H$_{14}$O$_5$, Synthesized), and DTAA (77%, C$_{10}$H$_{16}$O$_6$, Synthesized)) were used in the thermogravimetric analysis. All commercial compounds with high-purity reagents were purchased from Sigma-Aldrich, St. Louis, MO, USA. All the synthesized SOA species were prepared in the laboratory of authors. For the synthesis 3-MBTCA, a solution of dimethyl maleate was mixed with a solution of methyl isobutyrate followed by acidification with tetrahydrofuran to form trimethyl 3-methylbutane-1,2,3-tricarboxylate (triester). Then, triester was mixed with a solution of NaOH in water and MeOH followed by acidification with HCl and drying to yield 3-MBTCA (triacid). For the synthesis of terpenylic acid, cis-pinonic acid was dissolved in H$_2$SO$_4$ solution and saturated with (NH$_4$)$_2$SO$_4$ followed by drying to yield homoterpenyl methyl ketone. The homoterpenyl methyl ketone was dissolved in KOH and KMnO$_4$ solution and filtered to remove precipitates. The filtrate was acidified with H$_2$SO$_4$ solution and extracted with diethyl ether followed by drying over Na$_2$SO$_4$, and recrystallization to form terpenylic acid. To synthesize DTAA, CH$_3$MgBr solution was added to ethyl cyclopent-3-ene-1-carboxylate and NH$_4$Cl, and processed to form 2-(cyclopent-3-en-1-yl)propan-2-ol (tertiary alcohol). 4-Dimethylaminopyridine was added to tertiary alcohol in pyridine and acetic anhydride, and processed to yield 2-(cyclopent-3-en-1-yl)propan-2-yl acetate. Oxone and sodium periodate were added to acetate solution in acetonitrile/H$_2$O. The mixture was filtered, extracted with diethyl ether, and dried over Na$_2$SO$_4$ followed by evaporation and recrystallization to form DTAA. The laboratory-scale synthesis procedure of the aforementioned SOA species was discussed in detail by Babar et al. (2020b) [11].

2.2. Heat Capacity Analysis

Heat capacity measurements were performed using a TA Instrument SDT Q600, New Castle, DE, USA. A sapphire disc (α-aluminum oxide) was taken as a reference disc for isothermal flow as used in various references (ASTM E1269-1). DSC data were acquired in the heating mode over the temperature range of −30 °C to 120 °C with a heating rate of 20 °C/min. The heat flow of the blank and sapphire samples was measured in consecutive runs.

2.3. TGA/DSC Analysis

TGA-DSC analysis was performed using a TA Instrument SDT Q600, New Castle, DE, USA. For all SOA species, the initial mass was approximately 20 mg. The ramping
temperature was from 25 °C to 300 °C at a rate of 2.5 °C/min. The purge gas flow of N\textsubscript{2} was maintained at 100 mL/min. Before the experiment, the instrument for heat flow was calibrated from 100 °C to 388 °C using aluminum as a standard.

2.4. Estimation of Specific Heat Capacity

Specific heat capacity is the amount of energy required to raise the temperature of 1 mole of a substance by 1 K. ASTM E1269-11 \cite{44} provides a detailed method for estimating the specific heat of a substance using DSC \cite{24}. The specific heat capacity of a sample is calculated using Equation (1).

\[ C_{p(s)} = C_{p(st)} \frac{D(s)W_{st}}{D_{st}W_{s}}, \]  

where \( C_{p(s)} \) is the specific heat capacity of the specimen (J g\(^{-1}\) K\(^{-1}\)), \( C_{p(st)} \) the specific heat capacity of the sapphire standard (J g\(^{-1}\) K\(^{-1}\)), \( D(s) \) the vertical displacement between the specimen holder and the specimen DSC thermal curves at a given temperature (mW), \( D_{st} \) the vertical displacement between the specimen holder and the sapphire DSC thermal curves at a given temperature (mW), \( W_{s} \) the mass of the specimen (mg), and \( W_{st} \) the mass of sapphire standard (mg). Bernardes et al. (2020) \cite{40} developed a more accurate method to correct the deviation of heat capacities measured using different instruments. From the heat flow curves of DSC data, heat capacity, \( C_{p,m}(T) \), at a given temperature, \( T \), can be calculated as follows;

\[ C_{p,m}(T) = k(T) \frac{M}{m} \beta \Delta \phi(T), \]  

where \( m \) and \( M \) are the mass (g) and molar mass (g mol\(^{-1}\)) of the compound, respectively; \( \beta \) the heating rate (C min\(^{-1}\)), and \( \Delta \phi(T) \) the difference in heat flow rate (mW) between the sample and blank runs at a given temperature. \( k(T) \) is a temperature-dependent correction factor, expressed as;

\[ k(T) = \frac{C_{p,m}(\alpha-Al_{2}O_{3}, T)_{Ref}}{C_{p,m}(\alpha-Al_{2}O_{3}, T)_{Obs}}, \]  

where \( C_{p,m}(\alpha-Al_{2}O_{3}, T)_{Ref} \) represents the recommended heat capacity of sapphire at temperature \( T \) \( \) \( (\) provided by ASTM E1269-11 \) \cite{44} \( ) \) and \( C_{p,m}(\alpha-Al_{2}O_{3}, T)_{Obs} \) is the corresponding experimental value calculated using Equation (2). To estimate \( C_{p,m}(\alpha-Al_{2}O_{3}, T)_{Ref} \) as a function of temperature, an empirical equation is used as follows;

\[ C_{p,m} = A + BT + CT^2 + DT^3 + ET^{-2} \]  

where \( A, B, C, D, \) and \( E \) are constants and equal to 102.43, 38.75, \(-15.91, 2.63, \) and \(-3.00, \) respectively. Equation (4) is just an empirical equation with no physical application.

2.5. Estimation of the Melting Point and Enthalpy

For all samples, the melting temperature \( T_{m} \) was determined using the respective DSC curves. During a phase change, the heat supplied by the instrument is absorbed by the sample as latent heat of fusion. This results in an endothermic dip in the DSC heat curve of the sample at a certain temperature. According to the International Confederation for Thermal Analysis and Calorimetry standard, the melting point for pure compounds is the onset of the melting peaks. Similarly, for compounds with high impurities or amorphous structures (like polymers), the melting point is the minima of the melting peaks. Molar enthalpy of fusion \( \Delta H_{fus} \) (kJ mol\(^{-1}\)) at \( T_{m} \) was determined to be the peak area
of the endothermic heat curve per number of moles corresponding to mass in the melting range [45]. The melting enthalpy at 298 K was estimated using Sidwicks equation [46]:

\[
\Delta H_{\text{fus}}(298K) = \Delta H_{\text{fus}}(T_{\text{fus}}) + 0.0544 \left( 298 - T_{\text{fus}} \right)
\] (5)

2.6. Estimation of Saturation Vapor Pressure and Vaporization Enthalpy

The saturation vapor pressure of all SOA species was estimated using the Langmuir equation [47]:

\[
\frac{1}{a} \frac{dm}{dt} = p \alpha \sqrt{ \frac{M}{2\pi RT} }
\] (6)

where \( \frac{dm}{dt} \) \((1/a)\) is the rate of mass loss per unit area \((\text{kg s}^{-1} \text{m}^{-2})\), \( p \) the vapor pressure \((\text{Pa})\), \( M \) the molecular mass of sample \((\text{kg mol}^{-1})\), \( R \) the gas constant \((\text{kJ mol}^{-1} \text{K}^{-1})\), \( \alpha \) the vaporization coefficient, and \( T \) the absolute temperature \((\text{K})\). Rearranging the Langmuir equation, we obtain

\[
 p = kv
\] (7)

where \( k = \sqrt{2\pi R/\alpha} \) and \( v = \left( \frac{1}{a} \right) \left( \frac{dm}{dt} \right) \sqrt{T/M} \). The values of \( k \) and \( v \) depend on the instrument and sample, respectively. The vapor pressure of a reference compound is estimated using Antoine’s equation:

\[
\log p = A - B T + C
\] (8)

where \( p \) \((\text{Pa})\) and \( T \) \((\text{K})\) are the vapor pressure and absolute temperature, respectively, and \( A, B, \) and \( C \) are Antoine’s constants. To calculate the instrument-dependent \( k \) values, a linear relationship between \( p \) and \( v \) was plotted using Equation (7). For SOA species with unknown Antoine constants, the vapor pressure values were calculated using the instrument-dependent \( k \) values, which were estimated using the reference compound (benzoic acid in this study). The enthalpy of vaporization was calculated from the vapor pressure and temperature data using the Clausius–Clapeyron equation:

\[
\ln p = A - \frac{\Delta H_{\text{Vap}}}{RT},
\] (9)

\[
-\Delta H_{\text{Vap}} = RT \left( (\ln p) - A \right)
\] (10)

where \( p \) \((\text{Pa})\), \( \Delta H_{\text{Vap}} \) \((\text{kJ mol}^{-1})\), \( R \) \((\text{kg m}^2 \text{K}^{-1} \text{mol}^{-1} \text{K}^{-1})\), \( T \) \((\text{K})\), and \( A \) are the estimated vapor pressure, enthalpy of vaporization, gas constant, absolute temperature, and y-intercept, respectively. \( \Delta H_{\text{Vap}} \) is the slope of the line when \( \ln(p) \) against \( 1/T \). In this study, replicate analyses were carried out for benzoic acid and catechol with standard deviations less than 3% for the estimation of \( \Delta H_{\text{Vap}} \).

3. Results and Discussion

3.1. Specific Heat Capacity

Figure 1a shows the heat flow of pan, sapphire, and benzoic acid as a function of temperature. In the first and last four minutes of \( C_p \) measurement, the isothermal condition was maintained at the lowest and highest temperatures of \(-30^\circ \text{C}\) and \(115^\circ \text{C} \), respectively.
Table 1. Heat capacities of biogenic and anthropogenic SOA species.

| SOA Species | $T_m$ (°C) | $C_p, 298 K$ (J mol$^{-1}$ K$^{-1}$) | $C_p(T) = A + BT + CT^2 + DT^3 + ET^{-2}$ | References |
|-------------|------------|----------------------------------|---------------------------------------|------------|
| Malonic acid | 132.8      | 137.1                            | $-1.91 \times 10^5$ $1.17 \times 10^3$ $-2.69 \times 10^0$ $2.20 \times 10^{-3}$ $2.02 \times 10^{-6}$ | This study |
|             |            |                                  |                                       | Fukai et al. (1991) [49] |
| Suberic acid | 141.1      | 221.5                            | $-1.33 \times 10^5$ $8.02 \times 10^2$ $-1.80 \times 10^0$ $1.44 \times 10^{-3}$ $1.45 \times 10^{-6}$ | This study |
| Benzoic acid | 122.7      | 118.1                            | $7.01 \times 10^2$ $-1.98 \times 10^0$ $1.56 \times 10^{-3}$ $1.62 \times 10^{-6}$ $-1.53 \times 10^{-9}$ | This study |
|             | 121.3      | 125.1                            | $-1.16 \times 10^4$ $6.60 \times 10^1$ $-1.37 \times 10^{-1}$ $1.01 \times 10^{-4}$ $1.46 \times 10^{-8}$ | Santos et al. (2010) [48] |
| Phthalic acid | 193.9      | 169.4                            | $-1.16 \times 10^4$ $6.60 \times 10^1$ $-1.37 \times 10^{-1}$ $1.01 \times 10^{-4}$ $1.46 \times 10^{-8}$ | This study |
| Catechol    | 108.1      | 135.9                            | $1.51 \times 10^3$ $-7.72 \times 10^0$ $1.45 \times 10^{-1}$ $-9.37 \times 10^{-5}$ $-2.15 \times 10^{-8}$ | Santos et al. (2010) [48] |
|             | 140.2      | 137.1                            | $1.32 \times 10^2$ $2.80 \times 10^{-1}$ $5.40 \times 10^{-4}$ $2.05 \times 10^{-3}$ $-2.15 \times 10^{-9}$ | This study |
| Pinic acid  | 94.9       | 189.9                            | $-1.30 \times 10^4$ $7.40 \times 10^1$ $-1.53 \times 10^{-1}$ $1.14 \times 10^{-4}$ $1.64 \times 10^{-8}$ | Bret-Dibat et al. (1989) [51] |
| Ketopinic acid | 103.1     | 223.9                            | $-1.94 \times 10^3$ $1.19 \times 10^0$ $-2.74 \times 10^{-3}$ $2.23 \times 10^{-6}$ $2.06 \times 10^{-8}$ | This study |
| cis-Pinonic acid | 107.1   | 246.1                            | $-7.53 \times 10^3$ $4.64 \times 10^{-1}$ $-1.07 \times 10^{-3}$ $8.84 \times 10^{-8}$ $7.94 \times 10^{-7}$ | This study |
| Terpenylic acid | 81.5      | 223.2                            | $-3.87 \times 10^3$ $2.74 \times 10^3$ $-7.35 \times 10^{-3}$ $7.04 \times 10^{-6}$ $3.17 \times 10^{-8}$ | This study |
| DTAA *      | 130.2      | 524.1                            | $-1.08 \times 10^5$ $6.24 \times 10^2$ $-1.34 \times 10^0$ $1.03 \times 10^{-3}$ $1.31 \times 10^{-7}$ | This study |

Table 1. Heat capacities of biogenic and anthropogenic SOA species.

* DTAA: Diaterylpenyclic acid acetate.

The specific heat of benzoic acid was calculated using Equation (2) and plotted as a function of temperature (Figure 1b). The heat capacity of benzoic acid at room temperature (298.15 K) was 118.1 J mol$^{-1}$ K$^{-1}$, which is consistent with the reported value (125.1 J mol$^{-1}$ K$^{-1}$) [48]. The specific heat capacity of SOA species was calculated using the abovementioned method and summarized in Table 1. In most cases, the specific capacity increased by less than 1% with a 1 °C change in temperature. This indicates that the specific heat capacity of SOA species varies with temperature. This temperature-dependence of specific heat capacity has been reported with Equation (4). Herein, for SOA species, the coefficients in Equation (4) were estimated using linear regression and are listed in Table 1.
Bret-Dibat et al. (1989) [51] estimated the thermodynamic properties of some aromatic species using Kirchhoff’s equation (Equation (11)).

\[
\Delta H(T_o) = \Delta H(T) + \int_{T_o}^{T} \Delta C_p(T) \,dT,
\]

where \( T_o \) is a common reference temperature (usually 298.15 K), and \( \Delta C_p \) is \( C_p(\text{vapor}) - C_p(\text{solid}) \) for vaporization or \( C_p(\text{vapor}) - C_p(\text{liquid}) \) for evaporation. For catechol, \( C_p \) of 135.9 J mol\(^{-1}\) K\(^{-1}\) is obtained, which agrees with the reported value (140.2 J mol\(^{-1}\) K\(^{-1}\)) at 298 K. To date, the specific heat capacities of suberic acid, pinic acid, ketopinic acid, cis-pinonic acid, terpenylic acid, and DTAA have not been reported. Thus, the values obtained here would be good references for the thermodynamic properties of these SOA species.

3.2. Melting Point

TGA and DSC profiles of the SOA species in a temperature range of 25 °C to 300 °C at a heating rate of 2.5 °C min\(^{-1}\) are shown in the Supplementary Materials Figure S1. For all the charts, the blue solid and red striped lines represent the mass and heat curves of TGA/DSC, respectively. In most cases, a strong endothermic peak on the heat curve indicates the melting of SOA species. Herein, the rate of evaporation of the SOA species increased after fusion.

The change in calorific value as a function of temperature was steady but very rapid when a phase change occurred. The DSC profiles of adipic acid, catechol, cis-pinonic acid, ketopinic acid, benzoic acid, succinic acid, terpenylic acid, and 4-hydroxybenzoic acids show only two endothermic peaks. The first peaks at 151.4 °C, 108.1 °C, 103.2 °C, 122.7 °C, 183.8 °C, 81.5 °C, and 214.2 °C, correspond to the respective \( T_m \), and they are consistent with the reported values. For all the SOA species, a second endothermic peak occurred when the samples were completely vaporized (i.e., sample pan emptied), except for 4-hydroxybenzoic acid, which showed a second peak when 13% of the initial mass was remaining. All the SOA species showed typical TGA profiles with insignificant mass loss till \( T_m \), indicating a vaporization phase (i.e., mass transfer from solid to gas phase). In addition, a rapid mass loss was observed after \( T_m \), which is identified as the evaporation phase of molten samples.

For the case of malonic acid, pinic acid, and suberic acid, the first peaks were observed at 89.8 °C, 32.2 °C, and 133.9 °C, respectively, which are attributed to the thermal decomposition of the respective species. The second peaks at 132.8 °C, 94.9 °C, and 141.1 °C, respectively, corresponded to \( T_m \). Similarly, for 3-MTBCA, the first endothermic peak with a little mass loss appeared at 38.4 °C, corresponding to the evaporation of impurity or volatile fractions due to decomposition. The second peak of 3-MTBCA corresponding to \( T_m \) was detected at 136.4 °C. For DTAA, three endothermic peaks were observed. The first peak at 110.0 °C indicates evaporation of volatile reagents used in the synthesis of DTAA. As DTAA was synthesized using multiple chemical species, an increase in temperature results in the decomposition of volatile fractions. The second peak corresponds to the melting of DTAA with an onset melting point \( (T_m) \) of 147.1 °C. The third endothermic peak between 205–260 °C corresponds to the temperature at which SOA species was completely evaporated. The thermogram of phthalic acid shows two endothermic peaks very close to each other. The first peak has an onset \( (T_m) \) of 193.2 °C and minima of 214.1 °C, and the second endothermic peak at 218.7 °C indicates complete vaporization. The TG curve shows that phthalic acid started to evaporate as it melted. The evaporation phase occurred at 26 °C. However, pure evaporation occurred only from 217 °C to 223 °C. All the samples were sublimated during the heating process. The melting points and enthalpies of the samples are listed in Table 2.
Table 2. Melting temperatures and melting enthalpies at melting and room temperature of biogenic and anthropogenic SOA species.

| SOA Species          | $T_m$ (°C) | $\Delta H_{fus,Tm}$ (kJ mol$^{-1}$) | $\Delta H_{fus,298}$ K (kJ mol$^{-1}$) | Method * | References                  |
|----------------------|------------|------------------------------------|---------------------------------------|----------|-----------------------------|
| Malonic acid         | 132.8      | 22.7                               | 16.8                                  | TGA–DSC  | This study                  |
|                      | 132.9      | 18.7                               |                                       | KME      | Booth et al. (2010) [53]    |
| Succinic acid        | 183.8      | 63.5                               | 54.9                                  | TGA–DSC  | This study                  |
|                      | 187.8      |                                    |                                       | TDMA     | Bilde et al. (2003) [54]    |
| Adipic acid          | 151.4      | 35.0                               | 28.1                                  | TGA–DSC  | This study                  |
|                      | 152.9      |                                    |                                       | TDMA     | Acree et al. (1991) [55]    |
|                      | 153.2      |                                    |                                       |          |                             |
| Suberic acid         | 141.1      | 28.2                               | 21.9                                  | TGA–DSC  | This study                  |
|                      | 142.2      |                                    |                                       | TGA–DSC  | Acree et al. (1991) [55]    |
|                      | 140.1      |                                    |                                       | TDMA     | Roux et al. (2005) [56]     |
| Benzoic acid         | 122.7      | 14.2 ± 1.5                         | 8.6 ± 1.5                             | TGA–DSC  | This study                  |
|                      | 121.4      |                                    |                                       | TGA–DSC  | Sharma et al. (2003) [57]   |
|                      | 122.4      |                                    |                                       | TGA–DSC  | Ramos et al. (2017) [42]    |
|                      | 123.7      |                                    |                                       | TGA–DSC  | Brittain et al. (2009) [58] |
| 4-Hydroxybenzoic acid| 215.5      | 30.6                               | 20.2                                  | TGA–DSC  | This study                  |
|                      | 214.8      |                                    |                                       | TGA–DSC  | Gracin et al. (2002) [59]   |
|                      | 214        |                                    |                                       | TDMA     | Nordström et al. (2006) [60]|
| Phthalic acid        | 193.9      | 102.4                              | 93.2                                  | TGA–DSC  | This study                  |
|                      | 190.35     |                                    |                                       | Calorimetric | Sabbah et al. (1999) [61]  |
| Catechol             | 108.1      | 22.1 ± 0.8                         | 17.6 ± 0.8                            | TGA–DSC  | This study                  |
|                      | 104.5      |                                    |                                       | SLE      | Lee et al. (1997) [62]      |
|                      | 103.5      |                                    |                                       | TGA–DSC  | Verevkin et al. (2008) [63]|
| Pinic acid           | 94.9       | 7.3                                | 3.5                                   | TGA–DSC  | This study                  |
|                      | 91.1       |                                    |                                       | TGA–DSC  | Kołodziejczyk et al. (2019c) [12]|
| Ketopinic acid       | 103.1      | 13.9                               | 9.7                                   | TGA–DSC  | This study                  |
| cis-Pinonic acid     | 107.1      | 29.7                               | 25.5                                  | TGA–DSC  | This study                  |
|                      | 103.2      |                                    |                                       | TGA–DSC  | Kołodziejczyk et al. (2019c) [12]|
| Terpenylic acid      | 81.5       | 7.9                                | 4.8                                   | TGA–DSC  | This study                  |
| DTAA e               | 147.1      | 20.5                               | 12.8                                  | TGA–DSC  | This study                  |
|                      | 121.1      |                                    |                                       | TGA–DSC  | Kołodziejczyk et al. (2020) [64]|
| 3-MBTCA f            | 136.4      | 130.0                              | 123.9                                 | TGA–DSC  | This study                  |
|                      | 149.6      |                                    |                                       | TGA–DSC  | Kołodziejczyk et al. (2020) [64]|

* TGA–DSC: thermal gravimetric analysis—differential scanning calorimetry; TDMA: tandem differential mobility analyzer; SLE: solid–liquid equilibrium; KME: Knudsen mass effusion; DTAA: Diaterpenylic acid acetate; 3-MBTCA: 3-methylbutane-1,2,3-tricarboxylic acid.

3.3. Vaporization Enthalpy

In this study, benzoic acid was used as a reference compound to estimate the instrument-dependent $k$ value. The thermogravimetric data, calorimetric data, and Antoine’s constants of benzoic acid are available in the literature [47,65–68]. Antoine’s constants $A$, $B$, and $C$ in the temperature range of 132 °C–250 °C for benzoic acid are 7.80, 2776.12, and $-43.97$, respectively [69]. For each organic compound, the rate of mass loss (dm/dt) was calculated from the respective TGA curves. The vapor pressure of benzoic acid was estimated as a function of temperature using Antoine’s equation (Equation 8). A linear plot of $p$ versus $v$ in a temperature range of 135–165 °C (vaporization phase) is shown in Figure S2. From the plot, the mean $k$-value $k_{avg}$ was calculated equal to 4,453,444.50 kg$^{0.5}$ s$^{-1}$ m K$^{-0.5}$ mol$^{-0.5}$. Table S1 lists $k$, $k_{avg}$, and coefficient of determination ($R^2$) for all samples at various temperature ranges.
The vapor pressure of benzoic acid was estimated as the product \( k \) and sample-dependent mass change \( v \) (observed in the TG profile). Antoine’s constant for SOA species is only available for the vaporization phase; thus, the TGA/DSC method cannot be used to accurately estimate the vapor pressure in the sublimation period. Therefore, we used the same \( k \)-value to estimate the vapor pressure of the SOA species for the sublimation and vaporization. Wright et al. (2002) \[47\] employed the same approach to estimate the enthalpies of sublimation and vaporization.

Figure 2a compares the estimated vapor pressure of benzoic acid (the product of \( k_{\text{avg}} \) and \( v \)) in this study and the reported values obtained using Antoine’s constant in the NIST \[69\] database. The values are in good agreement, showing a linear regression of 98.3% for the evaporation regime. This indicates that the vapor pressure estimated beyond the fusion point of benzoic agrees well with the reported value. Figure S3 shows that the vapor pressure as a function of temperature for adipic acid and suberic acid are also in good agreement. In SOA study vapor pressure at room temperature might be appropriate and useful. The estimation of vapor pressure (\( P_{298K}^\circ \)) at room temperature using TGA/DSC data of this study is too uncertain because samples in our TGA/DSC analysis began to evaporate at a much higher temperature above room temperature. We obtained some preliminary data by extrapolation, but we determined not to report the data because of large uncertainties.

![Figure 2](image-url)

**Figure 2.** For benzoic acid (a) Saturation vapor pressure as a function of temperature, (b) correlation of saturation vapor pressure estimated in this study and that reported by NIST, and (c) estimated enthalpies of sublimation, melting, and vaporization as a function of temperature.

The molar enthalpy of vaporization was calculated from the slope of the linear curve of \( \ln(p) \) versus \( T^{-1} \). Table 3 compares the enthalpies of vaporization obtained in this study and the reported values in the literature. For the molar enthalpy of vaporization, the effect of temperature was negligible due to an insignificant mass-loss rate during vaporization (Figure S2).

**Table 3.** Vaporization enthalpies of biogenic and anthropogenic SOA species.

| SOA Species | \( \Delta H_{vap} \) (kJ mol\(^{-1}\)) | Method * | References |
|-------------|------------------------------------|----------|------------|
| Malonic acid | 132.7 | TGA–DSC \( a \) | This study |
| | 132.1 ± 5 | TPD \( f \) | Cappa et al. (2008) \[70\] |
| | 107 ± 4 | EDB \( b \) | Soonsin et al. (2010) \[71\] |
| | 111.4 | KME \( i \) | Ribeiro et al. (1999) \[39\] |
| Succinic acid | 116.1 | TGA–DSC \( a \) | This study |
| | 112 ± 12 | VTDMA \( c \) | Salo et al. (2010) \[9\] |
| | 125 ± 8 | EDB \( b \) | Soonsin et al. (2010) \[71\] |
| | 119.5 | TPTD \( g \) | Chattopadhyay et al. (2005) \[72\] |
| Adipic acid | 99.0 | TGA–DSC \( a \) | This study |
| | 119 ± 18 | KME \( i \) | Booth et al. (2010) \[53\] |
| | 97 ± 8 | VTDMA \( c \) | Salo et al. (2010) \[9\] |
The molar enthalpy values of vaporization for dicarboxylic acids, including malonic acid, succinic acid, adipic acid, and suberic acid were estimated as 132.7, 116.1, 99.0, and 94.12 kJ mol\(^{-1}\), respectively. For benzoic acid, phthalic acid, catechol, and 4-hydroxybenzoic acid, \(\Delta H_{vap}\) was estimated to be 93.2 ± 0.4, 131.6, 93.0 ± 2.5, and 119.5 kJ mol\(^{-1}\), respectively, and that of ketopinic acid, DTAA, and 3-MBTCA was 113.8, 130.2, and 135.4 ± 3.7 kJ mol\(^{-1}\), respectively. \(\Delta H_{vap}\) was not reported for pinic acid and cis-pinoic acid. Cappa et al. (2008) [70] used the temperature-programmed desorption method to estimate the temperature-dependent vaporization of malonic acid and reported \(\Delta H_{vap}\) of 132.1 ± 5 kJ mol\(^{-1}\), which agrees well value obtained here (132.7 kJ mol\(^{-1}\)). Using VTDMA, Salo et al. (2010) [9] estimated \(\Delta H_{vap}\) for succinic, adipic, and suberic acids as 112 ± 12, 97 ± 8, and 101 ± 10 kJ mol\(^{-1}\), respectively, which are consistent values obtained here (116.1, 99.0, and 94.1 92 kJ mol\(^{-1}\), respectively). Soonsin et al. (2010) [71] reported \(\Delta H_{vap}\) of 101 ± 10 and 101 ± 10 kJ mol\(^{-1}\) for malonic and succinic acids, respectively, which are approximately 15% and 5% lower than the values obtained here. The variation in the evaporation rate in the electrodynamic balance method is attributed to the transition of crystals to the amorphous state and vice versa. Ortiz-Montalvo et al. (2012) [78] has estimated \(\Delta H_{vap}\) of succinic acid to be 71.3 kJ mol\(^{-1}\) using SIMPOL.1. It is significantly lower than the experimental \(\Delta H_{vap}\) of 116.1, 112 ± 12, 125 ± 8, 119.5 kJ mol\(^{-1}\) of this study and earlier studies [9,71,72]. For adipic acid, Booth et al. (2010) [53] has reported \(\Delta H_{vap}\) of 119 ± 18 kJ mol\(^{-1}\) based on DSC, which is approximately 30% higher than the 99 kJ mol\(^{-1}\) obtained in this study. This difference is attributed to the difference in the heating rate, \(N_2\) purging flow rate, and sample structure [47]. Furthermore, the variation in \(\Delta H_{vap}\) for adipic acid could also be attributed to its thermal decomposition to valeric acid [76]. Benzoic acid is used as a reference compound to estimate the thermodynamic properties of organic and

### Table 3. Cont.

| SOA Species         | \(\Delta H_{vap}\) (kJ mol\(^{-1}\)) | Method \(^a\) | References                  |
|---------------------|------------------------------------|--------------|-----------------------------|
| Suberic acid        | 94.1 \(\pm\) 10                    | TGA–DSC \(^a\) | This study \[73\]            |
|                     |                                    | VTDMA \(^e\) | Salo et al. (2010) [9]       |
|                     | 130                                | TPD \(^f\)   | Cappa et al. (2008) [70]     |
| Benzoic acid        | 93.2 \(\pm\) 0.4                   | TGA–DSC \(^a\) | This study                  |
|                     | 91.4                               | TGA–DSC \(^a\) | Murray et al. (1980) [65]    |
| 4-Hydroxybenzoic acid | 119.5                            | TGA–DSC \(^a\) | Heath et al. (1992) [75]     |
| Phthalic acid       | 131.6                              | TGA–DSC \(^a\) | This study                  |
|                     | 129.8                              | Calorimetric | Sabbah et al. (1999) [61]    |
|                     | 135.9 \(\pm\) 3.1                  | IVM \(^i\)   | Babar et al. (2020a) [76]    |
| Catechol            | 93.0 \(\pm\) 2.5                   | TGA–DSC \(^a\) | This study                  |
|                     | 80.0                               | KME \(^j\)   | Chen et al. (2006) [77]      |
| Ketopinic acid      | 113.8                              | TGA–DSC \(^a\) | This study                  |
|                     | 136.6 \(\pm\) 3.5                  | IVM \(^c\)   | Babar et al. (2020a) [76]    |
| DTAA \(^k\)         | 130.2                              | TGA–DSC \(^a\) | This study                  |
| 3-MBTCA \(^k\)      | 135.4 \(\pm\) 3.7                  | IVM \(^c\)   | Babar et al. (2020b) [11]    |

\(^a\) TGA–DSC: thermal gravimetric analysis—differential scanning calorimetry; \(^b\) EDB: electrodynamic balance; \(^c\) IVM: integrated volume method; \(^d\) TDMA: tandem differential mobility analyzer; \(^e\) VTDMA: volatility tandem differential mobility analyzer; \(^f\) TPD: temperature-programmed desorption; \(^g\) TPTD: temperature-programmed thermal desorption; \(^h\) DMT: dynamic mass transfer; \(^i\) KME: Knudsen mass effusion; \(^j\) DTAA: Diaterpenylic acid acetate; \(^k\) 3-MBTCA: 3-methylbutane-1,2,3-tricarboxylic acid.
SOA species. Using the DSC method, Murray et al. (1980) [65], and Gau-Yi et al. (2010) [74] estimated $\Delta H_{vap}$ of benzoic acid as 89.7 ± 3 and 91.4 kJ mol$^{-1}$, respectively, which are consistent with the value (93.2 ± 0.4 kJ mol$^{-1}$) estimated in this study. For phthalic acid, $\Delta H_{vap}$ of 129.8 and 135.9 ± 3.1 kJ mol$^{-1}$ reported by Babar et al. (2020a) [76], and Sabbah et al. (1999) [61] respectively, agree well with the value obtained here. For ketopinic acid, $\Delta H_{vap}$ of 113.8 kJ mol$^{-1}$ in this study is comparable to 136.6 ± 3.5 kJ mol$^{-1}$ of Babar et al. (2020a) using IVM [76]. For 3-MBTA, the $\Delta H_{vap}$ of 124.4 kJ mol$^{-1}$ of this study is approximately 3% and 20% lower than the value of 128.4 ± 4.7 and 150 ± 15 kJ mol$^{-1}$ reported by Babar et al. (2020b) [11], and Kostenidou et al. (2018) [64], respectively. The difference with the value obtained using the DMT method is because the accommodation coefficient $\alpha$ was assumed to be 1 by Kostenidou et al. (2018) [64], but it should range from 0.1 to 0.8 [79]. For DTAA, Babar et al. (2020b) [11] reported $\Delta H_{vap}$ of 135.4 ± 3.7 kJ mol$^{-1}$, which is comparable to 130.2 kJ mol$^{-1}$ of this study. For cis-pinonic acid and pinic acid, a negligible evaporation rate was measured, indicating the low volatility of the compounds. $\Delta H_{vap}$ for aromatic and straight-chain carboxylic acids are comparable with modeled and experimental data. A little discrepancy in the estimation of vaporization enthalpy in IVM, TDMA, TGA-DSC and VTDMA might be attributed to the uncertainties in the measurement. As mentioned, replicate analyses of benzoic acid and catechol might imply minimal uncertainties in the estimation of thermodynamic properties of other examined compounds in this study.

Atmospheric modeling tools, such as volatility basis set, have provided a much better understanding of atmospheric chemistry by considering the decrease in OA volatility with aging. However, they underestimate OA concentrations due to the lack of understanding of the emission inventory and thermodynamic properties of OA [80]. A more accurate database for gas–particle partitioning parameters (such as $C^*$ and $\Delta H_{vap}$) may reduce the uncertainties in atmospheric models, such as CMAQ and PMCAMx [27,28,81–83]. Our experimental vaporization enthalpies are in the range of 93–135 kJ mol$^{-1}$ for biogenic and anthropogenic SOA species. Kostenidou et al. (2018) [64] reported experimental $\Delta H_{vap}$ of 150 ± 15 kJ mol$^{-1}$ for 3-MBTA. Zhang et al. (2022) [28] employed this value in CMAQ model for SOA formation in southwest China during June–August 2012. Boyd et al. (2017) [84] measured a very high $\Delta H_{vap}$ of 237 kJ mol$^{-1}$ for α-pinene SOA that is around 3 times higher than the 40–70 kJ mol$^{-1}$ typically used in the atmospheric model [85]. Therefore, it needs more experimental data for the improvement of CTM in SOA formation. Our experimental data can improve the prediction of vapor pressure and SOA formation. There is still a need for more experimental data for accurate prediction of the gas–particle partitioning of OA.

4. Conclusions

In this study, we obtained a thermogravimetric profile to estimate the specific heat capacity ($C_p$), melting point ($T_m$), melting enthalpy ($\Delta H_{fus}$), and vaporization enthalpy ($\Delta H_{vap}$) of atmospherically relevant compounds. The $C_p$ of malonic acid, suberic acid, benzoic acid, phthalic acid, catechol, pinic acid, ketopinic acid, cis-pinonic acid, terpenylic acid, and DTAA are 137.1, 221.5, 118.1, 169.4, 135.9, 189.9, 223.9, 246.1, 223.2, and 524.1 J mol$^{-1}$ K$^{-1}$, respectively. $T_m$ of malonic acid, succinic acid, adipic acid, suberic acid, benzoic acid, phthalic acid, catechol, pinic acid, ketopinic acid, cis-pinonic acid, terpenylic acid, DTAA, and 3-MBTA are 132.8 °C, 183.8 °C, 151.4 °C, 141.1 °C, 122.7 °C, 193.9 °C, 108.1 °C, 94.9 °C, 103.1 °C, 107.1 °C, 81.5 °C, 131.1 °C, and 136.4 °C, and their $\Delta H_{fus}$ is 22.7, 63.5, 35.0, 28.2, 14.2 ± 1.5, 102.4, 22.1 ± 0.8, 7.3, 13.9, 29.7, 7.9, 20.5, and 130.0 kJ mol$^{-1}$, respectively. $\Delta H_{vap}$ of malonic acid, succinic acid, adipic acid and suberic acid, benzoic acid, phthalic acid, catechol and 4-hydroxybenzoic acid, ketopinic acid, DTAA, and 3-MBTA are 132.7, 116.1, 99.0, 94.12, 93.2 ± 0.4, 119.5, 131.6, 93.0 ± 2.5, 113.8, 130.2, and 124.4 kJ mol$^{-1}$, respectively.
The obtained thermodynamic properties of the dicarboxylic acids agree well with the reported data. The difference between the estimated $\Delta H_{\text{vap}}$ and the reported values is attributed to the difference in the experimental methods. A further investigation is needed to answer the insufficient evaporation rate for cyclic dicarboxylic acids. This study paves the way for understanding SOA in the atmosphere and serves as a good reference for developing new methods or techniques for predicting the variation of $\Delta H_{\text{vap}}$ with temperature in the tropospheric temperature range.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/atmos13050709/s1, Figure S1: TGA and DSC profiles for (a) Malonic acid, (b) Succinic acid, (c) Adipic acid, (d) Suberic acid, (e) Benzoic acid, (f) Phthalic acid, (g) Catechol, (h) Pinic acid, (i) Ketopinic acid, (j) cis-Pinonic acid, (k) 3-MBTCA, (l) Terpenylic acid, (m) DTAA, and (n) 4-Hydroxybenzoic acid.; Figure S2: Estimated $k_{\text{avg}}$ for benzoic acid at 135 $^\circ$C to 160 $^\circ$C (vaporization phase) with a step change of 5 $^\circ$C.; Table S1: Estimated ki-values and coefficient of determination at corresponding temperature ranges using benzoic acid as a reference compound; Figure S3: Correlation of the estimated saturation vapor pressure with that reported in NIST for (a) adipic acid. (b) suberic acid. (c) Catechol.; Figure S4: A Saturation vapor pressure as a function of time for (a) Malonic acid, (b) Succinic acid, (c) Adipic acid, (d) Suberic acid, (e) Phthalic acid, (f) Catechol, (g) Pinic acid, (h) Ketopinic acid, (i) cis-Pinonic acid, (j) 3-MBTCA, (k) Terpenylic acid, and (l) DTAA.

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