SUPPLEMENTAL METHODS

Plasmid Construction

Slug and c-Kit were amplified by PCR using Phusion™ high-Fidelity DNA Polymerase (New England Biolabs), and then cloned into pMIGR1 vector. To construct pMig-Slug-2A-c-Kit (Slug/c-Kit), Slug and c-Kit genes were in-frame linked by T2A sequence and then cloned into pMIGR1 vector.

To construct the c-Kit-LUC luciferase reporter, ~0.8 kb of c-Kit promoter region was amplified by PCR using specific primers and cloned in to pGL4.10 vector (Promega). To generate the mutant variants of c-Kit-Luc, point mutations in the four recognition sites were introduced by PCR using specific primers. To construct the Slug-Luc luciferase reporter, ~1.5 kb of Slug promoter region was amplified by PCR using specific primers and cloned in to pGL4.10 vector. To construct shRNA targeting mouse c-Kit, we first constructed pLKO.1maxGFP2aPuro vector by cloning maxGFP2aPuro fragment into BamH1 and Nsil sites of pLKO.1 vector. We cloned shRNA control and c-Kit shRNA #1, #2, and #3 into AgeI and EcoR1 sites of pLKO.1maxGFP2aPuro vector. All primers were listed in Table S3.

Retroviral and Lentiviral Transduction of Hematopoietic Cells

For generation of retrovirus, 293T cells were transfected with a mixture of DNA containing 1.5 µg of pCL-Eco (IMGENEX), 2.5 µg of pMIGR1 vector only or pMIGR1 expression vectors expressing Slug, c-Kit, or Slug/c-Kit by Fugene HD (Roche) according to the manufacturer's instruction. For preparation of the lentiviruses, 293T cells were transfected with a mixture of DNA containing 2.5 µg of shRNA lentiviral vectors (scramble shRNA control, c-Kit shRNAs or c-Myc shRNAs) and 2.5 µg of the packaging mixture (GeneCopoeia, Rockville, MD) by Fugene HD. All Media containing retroviruses or lentiviruses were collected 24 hours after transfection,
and filtered through a 0.45 μm pore-size filter. All shRNAs excluding c-Kit shRNAs were purchased from Sigma and listed in Table S4.

To transduce HSPCs, cells were enriched from BM of mice following 5-Fu injection (150 mg/kg body weight). Enriched cells were cultured overnight and were spun at 200 X g. The supernatant was aspirated and replaced with retroviral or lentiviral particles-containing supernatant supplemented with 5 μg/ml polybrene (Sigma), followed by centrifugation (900 x g for 45 min). After two days of transduction, lentivirus-transduced cells were selected by 1 μg/ml puromycin, and then performed for the further experiments.

Flow Cytometric Analysis

BM MNCs were isolated from mouse BM and purified by density gradient centrifugation through Ficoll. For analysis of Lin-Sca-1+c-Kit+ (LSK) cells, one million of BM MNCs were stained with a mixture of biotinylated antibodies against mouse CD11b, CD3e, CD45R (B220), Ly-6G (Gr-1), and TER-119. Subsequently, the cells were co-stained with streptavidin-FITC, anti-Sca-1-APC, and anti-c-Kit-PE/Cy7.

For sorting and analyzing SP cells, BM MNCs were incubated with 5 µg/ml Hoechst 33342 (Life Technologies) at 37 ºC for 90 mins. For sorting and analyzing SLAM cells, BM MNCs were co-stained with anti-CD150-APC and anti-CD48-PE.

For analysis of peripheral blood, the samples were collected in PBS containing 50 mM EDTA solution via lateral tail vein incision. The peripheral blood MNCs were purified by density gradient centrifugation through Ficoll and then co-stained with anti-CD45.1-PE and anti-CD45.2-Alexa647 antibodies. All antibodies, unless stated, were purchased from Biolegend and eBioscience (Table S5). Flow cytometry was performed on a BD LSRFortessa or FACSAria, and all flow cytometric data were analyzed with FlowJo software (TreeStar).

BM Transplantation
6-8-week-old C57BL/6.SJL (CD45.1) mice were given acidified antibiotic water (Sulfamethoxazole and Trimethoprim) for 7 days, and then lethally irradiated with 10 Gy total body irradiation before BM transplantation. HSCs were transduced with retroviral or lentiviral particles for two days, and then transplanted into mice by retro-orbital injection. Adult splenocytes (1.5x10^6 cell/ per CD45.1 mouse) were injected as helper cells along with HSCs. Recipient mice were continuously fed with acidified antibiotic water for 1 month to reduce the chance of spontaneous infection. For serial transplantation, 6x10^5 BM MNCs from the primary recipients were injected into lethally irradiated secondary recipients.

**Luciferase Reporter Assay**

293T cells or primary BM MNCs were cultured in 24-well plate overnight, and then transfected with 50 ng of luciferase reporter, 400 ng of pMig or pMig-Slug, and 50 ng of pCMV-LacZ using Fugene HD. After 72 hours of transfection, cells were lysed in 250 µl of the passive lysis buffer (Promega) and assayed with a luciferase assay kit (Promega) as directed by the manufacturer. Luciferase activities were expressed as relative luciferase/LacZ activities and normalized to those of control transfections in each experiment.

**RNA Extraction, RT-PCR, and Real-time PCR**

Total RNA samples were extracted using Quick-RNA MicroPrep Kit (Zymo Research). For real-time PCR, 100 ng of total RNA was reverse-transcribed using SuperScirpt III™ cDNA Synthesis Kit (Invitrogen). *Hprt* or *Gapdh* expression was used as internal control to normalize relative expression of each gene. The list of primers is included in the supplemental Information (Table S3).
1. Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC. Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. *The Journal of experimental medicine* 1996 Apr 1; **183**(4): 1797-1806.

2. Stern P, Astrof S, Erkeland SJ, Schustak J, Sharp PA, Hynes RO. A system for Cre-regulated RNA interference in vivo. *Proceedings of the National Academy of Sciences of the United States of America* 2008 Sep 16; **105**(37): 13895-13900.
**Figure S1. Slug Deficiency Leads to a Higher Expression Level of c-Kit in HSCs**

(A) qPCR analysis of c-Kit mRNA expression levels in Lin⁻Sca-1⁺ HSC subpopulation. Lin⁻Sca-1⁺ HSC subpopulation was sorted by FACs from bone marrow of Slug⁺/+ and Slug⁻/- mice after 5-FU treatment (n = 3) and followed by RNA extraction and qPCR. Expression levels of c-Kit were normalized to Hprt levels.

(B) qPCR analysis of c-Kit mRNA expression levels in Lin⁻ subset and SLAM subset. Expression levels of c-Kit were normalized to Hprt levels.

(C, D) Flow cytometric analysis of c-Kit expression in distinct hematopoietic subsets in BM cells from Slug⁺/+ and Slug⁻/- mice (n = 4). The levels of c-Kit were evaluated by flow cytometry. MFI, mean fluorescence intensity. Data are representative of two independent experiments. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (*P < 0.05; **P < 0.01).
Figure S2. Semiquantitative-PCR and qPCR Analysis of c-Kit Transcripts in Primitive Hematopoietic Cells

(A-B) HSPCs were transduced with pMig or pMig-Slug retroviruses, cultured for three days, and followed by RNA extraction and PCR (A) and qPCR (B) analysis. Data are representative of two independent experiments. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (**p < 0.01).
Figure S3. Expression profiles of Snail Family Members in Different Hematopoietic Cell Lineages. The indicated subsets of hematopoietic cells were sorted by FACS and analyzed by PCR. HSC: hematopoietic stem cell, CLP: common lymphoid progenitor, CMP: common myeloid progenitor, Pro-T: progenitor T-cell, Pro-B: progenitor B-cell, GMP: granulocyte/macrophage progenitor, MEP: megakaryocyte/erythrocyte progenitor.
Figure S4. The sequence of Mouse c-Kit Promoter and its Luciferase Report’s constructs
(A) The DNA Sequences of Mouse c-Kit Promoter. Potential E-boxes for Slug binding sites (5’-caggtg-3’) in mouse c-Kit promoter region. The start code (ATG) for c-Kit gene is indicated.
(B) Construction diagram of the luciferase reporters for the mutant c-Kit promoters in E-boxes. E, E-box; X, mutant.
Figure S5. c-Kit luciferase reporter assays in 293T cells and HSPCs

293T cells or primary HSPCs were transfected with the reporter plasmid c-Kit-Luc (A), or its mutants (B, C, D) together with pMig (vector control) or pMig-Slug, then cultured for 72 h before luciferase activity assay. pCMV-LacZ was included in each transfections as an internal control to normalized luciferase activity. Data are representative of three independent experiments. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (*P < 0.05; **p < 0.01; N.S., not significant, P > 0.05).
Figure S6. c-Kit Knockdown by shRNAs
(A) Semi-quantitative PCR analysis of c-Kit knockdown in primary HSPCs. Transcriptional level of c-Kit was examined by gene-specific, semi-quantitative RT-PCR.
(B) Flow cytometric analysis of c-Kit protein expression in HSPC infected with lentivirus-expressing c-Kit shRNA #2.
Figure S7. The Percentage of Donor-derived Cells (CD45.2+) in PB of Recipients

(A) Percentage of PBMCs in recipients reconstituted with Slug\(^{+/+}\) or Slug\(^{-/-}\) infected with shRNA CTR or c-Kit shRNA \#2 were evaluated by flow cytometric analysis at 4 weeks after BM transplantation (n = 3 mice).

(B, C) Percentage of PBMCs in recipients reconstituted with Slug\(^{+/+}\) or Slug\(^{-/-}\) infected with shRNA CTR or c-Kit shRNA \#3 were evaluated by flow cytometric analysis at 2 (B) and 8 (C) weeks after BM transplantation (n = 3-4 mice). Data are representative of two independent experiments. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (*P < 0.05; **p < 0.01; N.S., not significant, P > 0.05).
Figure S8. Analysis of Donor-derived (CD45.2) Hematopoietic Cells in PB of Recipients. The percentage of myeloid (CD11b+ and Gr-1+) cells (A, D, G), T cells (CD3ε+) (B, E, H), and B cells (B220+) (C, F, I) in recipients reconstituted with Slug+/+ or Slug-/- infected with shRNA CTR or c-Kit shRNA#2 were evaluated by flow cytometric analysis at 2 weeks (A-C), 4 weeks (D-F), and 8 weeks (G-I) after BM transplantation (n = 4 mice), respectively. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (not significant, P > 0.05).
Analysis of Donor Cells in Peripheral Blood of Recipients. PBMCs were analyzed by flow cytometry in primary pMig-c-Kit reconstituted mice (n = 4). Data are representative of three independent experiments.

**Figure S9**
Figure S10. Analysis of Donor-Derived Hematopoietic Cells in PB of Recipients. Percentage of myeloid (CD11b+ and Gr-1+) cells (A, D, G), T cells (CD3ε+) (B, E, H), and B cells (B220+) (C, F, I) in primary pMig, pMig-Slug, pMig-cKit, and pMig-Slug/cKit reconstituted mice were evaluated by flow cytometric analysis at 2 weeks (A-C), 4 weeks (D-F), and 8 weeks (G-I) after BM transplantation (n = 5 mice), respectively. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (*P < 0.05; not significant, P > 0.05).
Figure S11. SCF Induces the Transcription Level of Slug. qPCR analysis of endogenous Slug transcripts in side population HSCs. The cells were treated with different doses of SCF (A) or treated with SCF for different times (B), and then were proceeded to qPCR analysis and normalized to Hrpt level. **p < 0.01.
Figure S12. High dose of 5-FU enriches the compartment of HSPCs
Mice were injected with 5-FU [150 or 300 mg/kg body weight (BW)]. Total BM cells were harvested 7 days after injection and analyzed by flow cytometric analysis.
Figure S13. DNA Sequence of Slug Promoter

c-Myc binding site (5’-CANNTG-3’) and FOXM1 binding site (5’-(C/T)AAA(C/T)A-3’) are indicated. The start code (ATG) for Slug gene is indicated.
Figure S14. Slug Luciferase Reporter Assay

K562 cells were transfected with the reporter plasmid Slug-Luc together with pMig (vector control), pMig-c-Myc, or pMig-FoxM1, then cultured for 72 h before luciferase activity assay. pCMV-LacZ was included in each transfections as an internal control to normalized luciferase activity. Data are representative of three independent experiments. All data represent mean ± SD. Two-tailed Student’s t tests were used to assess statistical significance (**p < 0.01).
Figure S15. Key Role of c-Myc and FoxM1 in SCF/c-Kit-Slug Feedback Loop

(A) Semi-quantitative PCR analysis of endogenous c-Myc knockdown by shRNAs in primary BM cells. Lin- cells were infected with lentiviruses expressing shRNA targeting c-Myc or control shRNA and selected with puromycin. Transcriptional level of c-Myc was examined by semiquantitative RT-PCR. Density of amplified DNA bands was quantified by Image J (NIH) and normalized by Gapdh. Data are representative of two independent experiments.

(B) Semi-quantitative PCR analysis of endogenous c-Myc transcripts in HSPCs treated with SCF. HSPCs were treated with or without SCF (100 ng/ml) for 12 hrs and then analyzed by PCR using c-Myc specific primers. Data are representative of two independent experiments.

(C) qPCR analysis of Slug transcripts in wild-type LSK subpopulation following treatment with or without SCF (100 ng/ml) and FoxM1-binding inhibitor FDI-6 (40 µM) for 12 hrs. Slug transcripts was analyzed by qPCR and normalized to Gapdh level. Data are representative of two independent experiments. **p < 0.01.

(D) qPCR analysis of FoxM1 transcripts in HSPCs after treatment with SCF. HSPCs were enriched from wild-type mice and treated with or without SCF (100 ng/ml) for 12 hours, followed by qPCR analysis. FoxM1 expression was normalized to Gapdh level. Data are representative of two independent experiments. **p < 0.01.
Table S1. Gene expression profiles from microarray data analysis

| Gene Symbol (or Genomic Position) | Fold change (Slug\textsuperscript{+/+} vs. Slug\textsuperscript{-/-}) | ANOVA p-value |
|----------------------------------|-------------------------------------------------|----------------|
| Adam8                           | 5.55                                           | 0.034662       |
| Bmp7                            | 3.52                                           | 0.02986        |
| Faim3                           | 4.35                                           | 0.003286       |
| Gm10717                         | 2.79                                           | 0.040442       |
| St3gal6                         | 3.26                                           | 0.023531       |
| Cd96                            | 3.1                                            | 0.001186       |
| Mapk11                          | 2.68                                           | 0.007001       |
| Inpp4b                          | 3.11                                           | 0.007794       |
| Bhlhe40                         | 2.83                                           | 0.011203       |
| Cxcr6                           | 2.88                                           | 0.038504       |
| Arg1                            | 4.46                                           | 0.023874       |
| Gm16271                         | 3.7                                            | 0.03781        |
| Phlda1                          | 3.06                                           | 0.02399        |
| Ccdo184                         | 3.56                                           | 0.031538       |
| Il4                             | 3.17                                           | 0.009127       |
| Rasgrp1                         | 3.21                                           | 0.048914       |
| Gm26216                         | 2.79                                           | 0.024533       |
| Lpcat2                          | 2.84                                           | 0.005914       |
| Mboat1                          | 2.6                                            | 0.023953       |
| Stab2                           | 16.01                                          | 0.024117       |
| Ccr2                            | 5.03                                           | 0.046822       |
| Cs2f                            | 8.13                                           | 0.034921       |
| Ighv5-12; Igh-VJ558             | -33.28                                         | 0.061425       |
| Igkv3-9                         | -11.83                                         | 0.018412       |
| Dhrs3                           | -3.88                                          | 0.033815       |
| Selp                            | -2.79                                          | 0.002956       |
| Plac8                           | -2.59                                          | 0.023045       |
| Kit                             | -2.58                                          | 0.020692       |
| LOC102643064                    | 8.73                                           | 0.031955       |
| Ikzf2                           | 2.73                                           | 0.040157       |
| Flrt2                           | 3.31                                           | 0.004743       |
| 1700113H08Rik                    | 3.42                                           | 0.012626       |
| Ccr9; Gm17200                    | 3.34                                           | 0.021881       |
| Ptger2                          | 2.68                                           | 0.04142        |
| Tlr1                            | 3.08                                           | 0.047968       |
| Tnfsf14                         | 3.07                                           | 0.037037       |
| Mir342                          | 3.47                                           | 0.035325       |
| Tm4sf5                          | 2.69                                           | 0.033192       |
| 177000956 - 177013459            | 2.73                                           | 0.000436       |
| Gm26497                         | 3.69                                           | 0.049349       |
| Il5                             | 3.9                                            | 0.036871       |
| Mmp14                           | -3.51                                          | 0.003193       |
|                |     |      |
|----------------|-----|------|
| **Tbc1d8**     | -3.53 | 0.037839 |
| **Scd1**       | -2.89 | 0.003648 |
| **Sema6b**     | -3.05 | 0.021768 |
| **H2-Eb2**     | -2.82 | 0.043391 |
| **Itga2b**     | -2.97 | 0.04893 |
| **Gm24630**    | 3.68  | 0.024501 |
| **Gm25967**    | 2.6   | 0.035973 |
| **Gm25631**    | -3.09 | 0.015075 |
| **Gm8906**     | -3.26 | 0.015109 |
| **Rhag**       | -3.8  | 0.040231 |
### Table S2. The prediction of Slug binding sites in the selected target candidates

| Gene Name | Ensembl ID | CHR | Biotype       | Description         | Best CRM Score | Ave CRM Score |
|-----------|------------|-----|---------------|---------------------|----------------|---------------|
| Sema6b    | ENSMUSG0000001227 | 17  | protein_coding | sema domain         | 1.0000         | 1.0000        |
| Kit       | ENSMUSG0000005672 | 5   | protein_coding | kit oncogene [Source:MG I Symbol;Acc:MGI:96677] | 1.0000         | 0.9836        |
| Selp      | ENSMUSG0000026580 | 1   | protein_coding | selectin            | 0.9673         | 0.9673        |

Transcription factor binding sites (TFBSs) and gene structure diagram

#### Sema6b

![Gene Structure Diagram for Sema6b](image)

#### Kit

![Gene Structure Diagram for Kit](image)

#### Selp

![Gene Structure Diagram for Selp](image)
| Primer                  | Sequence (5' to 3')                                                                 | Application |
|------------------------|------------------------------------------------------------------------------------|-------------|
| c-Kit-F2               | ctctctcagggcgcaattagatctctctagaccATGAGAGGCGCTCGGCgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgccgc
| shNRA           | Supplier | Catalog number  |
|-----------------|----------|----------------|
| c-Myc-shRNA #1  | Sigma    | TRCN0000234924 |
| c-Myc-shRNA #2  | Sigma    | TRCN0000234925 |
| c-Myc-shRNA #3  | Sigma    | TRCN0000234926 |
| c-Myc-shRNA #4  | Sigma    | TRCN0000054853 |
| c-Myc-shRNA #5  | Sigma    | TRCN0000054854 |
| Antibodies                                                   | Supplier     | Catalog number |
|--------------------------------------------------------------|--------------|----------------|
| Mouse hematopoietic lineage biotin panel                     | eBioscience  | 88-7774-75     |
| APC anti-mouse Ly-6A/E(Sca-1)                                | Biolegend    | 108112         |
| PE/Cy7 anti-mouse CD117(c-Kit)                               | Biolegend    | 105814         |
| APC anti-mouse CD150 (SLAM)                                  | Biolegend    | 115910         |
| PE anti-mouse CD48                                           | Biolegend    | 103406         |
| FITC Streptavidin                                            | Biolegend    | 405202         |
| PE anti-mouse CD45.1                                         | Biolegend    | 110708         |
| Alexa Fluor® 647 anti-mouse CD45.2                          | Biolegend    | 109818         |
| Pacific Blue™ anti-mouse/human CD11b                        | Biolegend    | 101223         |
| PE/Cy7 anti-mouse CD3e                                       | Biolegend    | 100319         |
| Alexa Fluor® 700 anti-mouse/human CD45R/B220                | Biolegend    | 103231         |
| PerCP/Cy5.5 anti-mouse Ly-6G/Ly-6C (Gr-1)                    | Biolegend    | 108427         |
| Anti-Flag Tag Monoclonal antibody                            | Thermo Scientific | MA1-91878  |

Table S5. List of antibodies