Immunochemical Assays for Determination of Organophosphorus Pesticides in Milk

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Abstract

The detection of pesticide residues is an important task in ensuring the safety of milk. In the last decade, organophosphorus insecticides have been used, but they also pose a serious risk because of their high toxicity. For fast screening of milk, immunochemical methods of analysis are appropriate. Enzyme-linked immunosorbent assay is the most common assay mode and base of many commercialized assay kits. Residues of some organophosphorus compounds in milk and dairy products determined by ELISA are shown. For fast screening of pesticides, the lateral flow strips are also described. The advantages of a new developed method MNPs-based immunoassay for determination of phosphorus pesticides in milk was described and some publications were represented. In recent years, simultaneous identification of more than one pesticide is increasingly perceptual, that way the examples for multi-immunoassays for determination of organophosphorus pesticides was presented.

Keywords: Organophosphorus pesticides; Milk; Immunoassay; ELISA; Lateral test; MNPs; Multi-assays

Abbreviations: DDT: dichloro diphenyl trichloroethane; ELISA: enzyme-linked immunosorbent assay; FLISA: fluorescent-linked immunosorbent assay; FPIA: Fluorescence polarization immuno-analysis; HPLC: high performance liquid chromatography; IFA: immuno-fluorescent assay; MNPs: magnetic nanoparticles; MRL: maximum residue limits; OPP: organophosphorus pesticides

Introduction

Milk and dairy products are widely used food products which are being consumed by people of different age groups. In this regard, the quality and safety of raw milk is an important issue [1,2]. The detection of pesticide residues is an important task in ensuring the safety of milk. The use of organochloride pesticide like dichloro diphenyl trichloroethane (DDT) and hexachlorocylohexane (HCH) was banned. However, the residual effects are still causing problems [3-5]. The residues of DDT persist in the environment for long periods and enter the food chain. In the last decade, organophosphorus insecticides (OPP) and carbamyl derivatives have been used that are relatively easy to degrade in the environment but also pose a serious risk because of their high toxicity. There are variety publications reporting organic compounds exceeding established maximum residue limits (MRL), for different OPP pesticides, like fenthion, dichlorvos, methyl parathion, chlorpyrifos, and malathion in milk [6-8]. Melgar et al. [7] found that the frequency of total samples containing detectable levels of OPP residues was 6.73 % in total milk and 8.67 % in raw milk. The highest percentage incidence measured was for dichlorvos (5.78 %), followed by coumaphos (2.06 %), and parathion methyl (0.83 %). Salas et al. [8] described that approximately 39.6% of the pasteurized Mexican milk samples contained detectable levels of OP pesticide residues. Eight samples contained residues exceeding established maximum residue limits (MRL). They found that the OP pesticides present in these samples were dichlorvos, phorate, chlorpyrifos, and chlorfenvinphos. OP pesticides are strong cholinesterase inhibitors and they are like neurotransmitters for acetylcholinesterase, butyrylcholinesterase and pseudocholinesterase. When these enzymes are inhibited, acetylcholine accumulates in the neurons of the muscle nodes or synapses and changes the natural path of the nervous impulse. Acetylcholine accumulation causes a number of diseases like Alzheimer’s disease and Parkinsonian disorders, and also many other degenerative diseases [9,10].

Pesticide analysis is performed by liquid, gas and high performance liquid chromatography (HPLC), and HPLC along with Mass Spectrometry [11,12]. These methods are very suitable for monitoring toxic pesticides but have a number of disadvantages: expensive, time-consuming to prepare samples, require well-trained personnel, and are not adapted to conduct on-site and (on time) real-time analysis. For fast screening of milk, immunochemical methods of analysis [13] are appropriate. They
are simple, fast, cheap, sensitive methods, provide real-time and time-based analysis, and do not require costly instrumentation. In addition, the immunoassay is also suitable for the analysis of milk, since milk samples can be analyzed after simple pretreatment such as dilution or protein precipitation.

**Immunochemical Methods**

Immunochemical methods are based on the reaction between the antigen (the pesticides studied) and the respectively antibody specific, which is very selective and rapid [14]. Since pesticides are small molecules, they cannot be detected by sandwich immunoassay. A competitive method should be used whereby the antigen in the sample competes with the labeled pesticide to bind to the antibody [15]. From immunochemical methods, the largest application for the determination of organophosphorus pesticides is the enzyme-linked immunosorbent assay (ELISA) and the immunofluorescent assay (IFA).

**ELISA and FLISA**

A significant number of ELISA assays were developed to identify different organophosphorus pesticides [16]. Analyzes are sensitive and detect very low concentrations of pesticides. Enzyme-linked immunosorbent assay (ELISA) is the most common assay mode and base of many commercialized assay kits. The preparation of monoclonal and polyclonal antibody against organophosphorus pesticides, and the antigen-enzyme conjugate synthesis, is the very important task [17-20]. The most common markers for those conjugates are the enzymes horseradish peroxidase and alkaline phosphatase. Quantification of the target analytes is based on corresponding substrate color change. Residues of some organophosphorus compounds in milk and dairy products determined by ELISA are shown in Table 1.

**Table 1:** Representative examples of ELISA for determination of Organophosphorus pesticide.

| Pesticides | Antibody | Linear Range      | Reference |
|------------|----------|-------------------|-----------|
| Parathion  | p        | 0.1-5 µg/L        | [21]      |
| Parathion  | p        | 0.2 µg/L          | [22]      |
| Paraion    | m        | 10-100 µg/mL      | [23]      |
| Paraion    | p        | 28-275 µg/L       | [24]      |
| Paraion    | p        | 0.03 µg/L-275mg/L | [25]      |
| Achetate   | P        | 5-140 ng/mL       | [26]      |
| dichlorvos | p        | 0.01-5 ng/mL      | [26]      |
| paraion    | p        | 0.01-5 ng/mL      | [26]      |

In recent years, a number of analyzes for pesticides in milk have been done based on fluorescent-linked immunosorbent assay (FLISA) [21-24]. This method is highly sensitive, because combined selectivity of antibody and sensitivity of fluorescent marker [25,26]. Different fluorescent markers are used: organic fluorescent dyes, fluorescent marker proteins, green fluorescent proteins, quantum dots [14]. ELISA and FLISA are simple and low cost methods, but analysis is multi-stage, requires multiple washing and takes a long time (Figure 1).

**Lateral Flow Strips**

For fast screening of pesticides, the lateral flow strips are also used, but they are less sensitive and inaccurate. Lateral flow tests are also known as lateral flow immunochromatographic assays. The lateral flow strips are based on a competitive analysis. Unlabeled antigen and competitive antigen-marker are transported by the flow onto a nitrocellulose membrane, and they are competing for coupling with antibody immobilized onto the membrane [27]. The label most commonly used in immunochromatographic assay is colloidal gold, enzyme, magnetic nanoparticles [28]. The lateral flow strips are able to detect low pesticide concentrations. Their low detection limit is in the range of ng/mL, but often have false results. Residues of some organophosphorus compounds determined by lateral flow tests are shown in Table 2 [29-32].
**FPIA and MNPs based IFA**

The IFA for pesticide determining does not draw back from the sensitivity of the ELISA method but does not require repeated washing and is a faster method [33]. Fluorescence polarization immuno-analysis (FPIA) is quick and applicable method. There many publications about OP pesticide detection. Anna Yu. Kolosova, et al. [34] was developed and optimized monoclonal antibody for the detection of parathion-methyl (PM). PM concentration determinable by the FPIA ranged from 25 to 10000 ppb. The detection limit was 15 ppb. Xu et al. [35] make the FPIA which detect 5 OPs simultaneously with a limit of detection below 10ng/mL. Heterogeneous IFA is preferable to homogeneous as it provides a more stable test; due to the immobilization of antibodies onto solid supports [36]. Application of nanoparticles as carriers increases the degree of immobilization of the antibody enhances the sensitivity of the immunoassay and preserves the activity of the immobilized antibody over time [37]. Particularly suitable for this purpose are magnetic nanoparticles (MNPs), since due to the applied magnets separation accelerates and facilitates the conduct of the assay, reducing the interference of the interfering components in the milk sample and preserves the activity of the immobilized antibody over time [38]. MNPs have a high specific surface area and provide good contact between the immobilized antibody and the antigen and correspondingly high rate of immunoreaction. The principle of the method is based on competitive analysis. The target antigen and antigen-fluorescent dye conjugate compete for coupling to antibody immobilized onto MNPs (Table 3).

| Pesticide        | Marker     | Samples     | LOD     | Reference |
|------------------|------------|-------------|---------|-----------|
| Methyl parathion | Er³⁺       | Food        | 2 ng/mL | [29]      |
| Endosulfan       | Gold nanoparticles | Agricultural products | 10 μg/L | [30]      |
| Methyl parathion | Er³⁺       | Food        | 2 ng/mL | [29]      |
| Parathion        | Er³⁺       | Food        | 2 ng/mL | [29]      |
| Endosulfan       | HRP        | Agricultural products | 1 μg/L | [30]      |
| Methyl parathion | Gold nanoparticles | -            | 8 ng/mL | [31]      |
| Chlorpyrifos     | Gold nanoparticles | Agricultural products | 10 ng/mL | [29]      |
| Methyl paraoxon  | Fe₃O₄      | Food        | 1.7 ng/mL | [32]      |

At the first step, OPP containing sample is added to MNPs with immobilized antibody. Then, fluorescent conjugate (OPP-fluorescent dye) is added and the conjugate is coupled to the non-occupied binding sites of the antibody. The MNP are collected with a permanent magnet. After that, the fluorescence intensity of the uncoupled conjugate (OPP-fluorescent dye) in the supernatant is measured. The excess of the conjugate is directly proportional to the OPP concentration in the sample. On this principle have been developed assays for the detection of ochratoxin, aflatoxin M1, enterotoxin A, and antibiotics in milk [38-41]. There are few publications about MNP-based immunoassay for the determination of phosphorus pesticides in milk [42-44].

**Multi-Immuoassays**

Testing several pesticides simultaneously in one sample is a very perceptual analysis. In recent years, simultaneous identification of more than one pesticide [45] is increasingly perceptual. To achieve this, it is possible to increase the number of antibodies that recognize individual antigens in one sample. A more economical alternative, however, is to create a single antibody that is able to identify several analytes in a single test and which is termed a broad-specific antibody. First, multi-hapten antigen was prepared by sequentially conjugating haptons of four different pesticides (chlorpyrifos, triazophos, carbofuran and parathion methyl) to the carrier protein bovine serum albumin (BSA) [46]. An antibody with wide cross-reactivity is then obtained.

For example, Liang et al. [47,48] synthesized a common hapten of several 0, 0-dimethyl organophosphorus pesticides to produce a broad-spectrum antibody. The hapten is conjugated to bovine albumin (BSA) to produce an immunogen. Rabbits were immunized with the resulting immunogen and polyclonal antiserum was produced against each of the antigens.

A common, broadly specific enzyme-linked immunosorbent assay for malathion, dimethoate, phenate, phosmet, methidathion, fenitrothion, methyl parathion and fenithion has been developed. The following IC50 values were determined under optimal conditions: 30 μg/L for malathion, 28.9 μg/L for dimethoate, 88.3 μg/L for fenate, 159.7 μg/L for phosmet, 191.7 μg/L for fenithion.
methidathion, 324.0μg / L for fenitrothion, 483.9μg / L for methyl parathion and 788.9μg / L for fenothion. Antibodies produced using the multi-hapten strategy have much lower affinities to analytes compared to antibodies derived from a single hapten. Probably this is due to a lower hapten-protein molecular ratio for each individual hapten in the multi-hapten immunogen compared with those immunogens that contained only a single hapten [46]. Mixing several single antigens together based on a certain proportion to form a multi-hapten immunogen was also used for the preparation of mixed antibodies [49,50]. Each hapten is coupled to a carrier protein individually and then the mixture is used as immunogen. In this case, a lower titer of multivariate was established as compared to the obtained antibodies with only one individual hapten.

Conclusion

The immunochemical assays for determination of organophosphorus pesticides are simple, fast, cheap, sensitive methods, provide real-time and time-based analysis, and do not require costly instrumentation. In addition, the immunoassay is also suitable for the analysis of milk, since milk samples can be analyzed after simple pretreatment such as dilution or protein precipitation. The developed MNPs based IFA and multi-assays are very perspective assays, suitable for determination of organophosphorus pesticides in milk.

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