CASE REPORT

Asthmatic adult with marked blood eosinophilia: is it truly asthma?

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SUMMARY
A middle-aged woman presented with symptoms suggestive of allergic asthma but with markedly elevated peripheral eosinophilia. She did not respond to inhaled corticosteroids, thereby prompting further investigations. Chest radiograph was normal. CT of the chest revealed bi-apical ground glass opacities. Bronchoalveolar lavage revealed predominantly eosinophilic yield. Autoimmune screen was negative. Bone marrow biopsy showed a normocellular marrow with increased eosinophils. A diagnosis of chronic eosinophilic pneumonia (CEP) was made after exclusion of other causes of eosinophilia. Treatment of her CEP with systemic corticosteroids (prednisolone 0.5 mg/kg/day) resulted in dramatic improvement in symptoms and peripheral eosinophilia.

BACKGROUND
Peripheral blood eosinophilia is frequently encountered in the general medical and respiratory clinics. Asthma is often associated with peripheral eosinophilia, however, marked levels of eosinophilia are uncommon and should warrant further investigation. The patient described in this article presented with an initial diagnosis of allergic asthma but failed to respond to inhaled corticosteroids. Marked blood eosinophilia prompted the investigation for other eosinophilic lung diseases, which eventually yielded a diagnosis of chronic eosinophilic pneumonia (CEP). This article illustrates the importance of a high index of suspicion for eosinophilic lung diseases should patients previously diagnosed as allergic asthma fail to respond to therapy, especially if this was associated with marked blood eosinophilia.

CASE PRESENTATION
A 47-year-old woman who was a lifelong non-smoker was referred to the pulmonary clinic for non-productive cough and exertional dyspnoea for the past 6 months. She worked as a manager in a trading company. Her medical, social, family, medication and travel histories were non-contributory. She had initially consulted another institution for similar symptoms 6 months prior. Full blood count (FBC) then showed mildly raised absolute eosinophil count of 0.98 × 10^9/L (differential eosinophil count 13.6%). Chest radiograph was normal (figure 1A). Spirometry was normal with no bronchodilator response and the flow-volume loops were also normal. She did not have symptoms of allergic rhinitis or gastro-oesophageal reflux disease or atopy. She was treated for possible asthma with budesonide 160 µg and formoterol 4.5 µg combination turbuhaler, two puffs twice daily. In view of her lack of response to treatment, she was referred for further evaluation at our centre. Further history obtained during her visit to our clinic revealed that she had no fever, weight loss or night sweats. Clinical examination showed no clubbing or cervical lymphadenopathy.

INVESTIGATIONS
Repeat FBC count showed markedly raised absolute eosinophil count of 9.38 × 10^9/L (differential eosinophil count 56.6%). A repeat chest radiograph was normal (figure 1B). Autoimmune workup including antidouble-stranded DNA antibody, extractable nuclear antigens (ENA) screen, rheumatoid factor, antineutrophil cytoplasmic antibody and antinuclear antibody was negative. Stool examination for ova, cysts, parasites, leukocytes and culture was also negative. Contrast CT scan of the chest (figure 2A) showed ground glass changes with thickening of the interlobar septa at the apices of both lungs, more marked over the apical segment of the left upper lobe. These changes were notably absent in other parts of the lung (figure 2B–D). Bronchoalveolar lavage (BAL) with differential cell counts was performed from bilateral upper lobe apicosterior segments, revealing predominantly eosinophilic yield of 83.3% (figure 3, table 1). Bacterial, fungal and tuberculosis cultures from the BAL were negative. Transbronchial lung biopsy performed in the left upper lobe apicosterior segment showed large collections of eosinophils predominantly in the interstitium and within the lumens of vessels (figure 4A,B). Specifically, allergic granulomas suggestive of granulomatosis with polyangiitis, typically consisting of histiocytes and multinucleated giant cells surrounding a central necrotic zone, was absent. A bone marrow biopsy was also performed which showed normal cellularity for age, approximately 50%. Lymphocytes (about 10%) showed normal morphology and feature a diffuse interstitial infiltrate of mainly singly disposed CD3+ T cells, accompanied by occasional CD20+ B cells. A reactive lymphoid aggregate was present. Myeloid cell maturation was normal and no significantly increased numbers of immature myeloid precursors were seen (<2%) despite immunostaining for CD34 and CD117. The FIP1L1-PDGFR- alpha fusion and Bcr-Abl transcripts were not detected in the bone marrow. These findings were in keeping with a normocellular marrow with increased eosinophils.
Reminder of important clinical lesson

but without evidence of chronic myeloproliferative disorders or lymphomatous involvement. Flow cytometry was normal.

DIFFERENTIAL DIAGNOSIS
Possible differential diagnosis can be divided into two groups: primary causes of blood eosinophilia ad secondary causes of blood eosinophilia.

Primary causes of blood eosinophilia include:
1. Haematological malignancies (such as acute leukaemia or a chronic myeloid disorder).
2. Idiopathic hypereosinophilia.

Secondary causes of blood eosinophilia include:
2. Allergy-related causes such as allergic asthma.
3. Autoimmune conditions (such as eosinophilic granulomatosis with polyangiitis, granulomatosis with polyangiitis, sarcoidosis and chronic eosinophilic pneumonia).
4. Drug-related causes (such as carbamazepine, sulfa drugs, non-steroidal anti-inflammatory drugs, nitrofurantoin, etc).
5. Malignancy (such as lymphoma, metastatic cancers).

In this patient, bone marrow biopsy and flow cytometry excluded myeloproliferative, lymphoproliferative disorders and malignancy. Extensive history obtained excluded drug-induced causes of peripheral eosinophilia. Several stool investigations, autoimmune screening tests and lack of systemic manifestations of autoimmune diseases also excluded parasitic and autoimmune causes. Investigations to exclude fungal infections such as coccidioidomycosis or parasitic infections (eg, Strongyloides antibody) should be considered in the appropriate endemic areas or in the presence of relevant travel history. Aspergillus antibody should be considered when clinical features of allergic bronchopulmonary aspergillosis is present (eg, mucus plugging, eosinophilia, elevated serum total IgE).

A diagnosis of CEP was made due to the presence of respiratory symptoms, BAL eosinophilia with consistent imaging findings and exclusion of other known causes of eosinophilia as above.

TREATMENT
Our patient was started on oral prednisolone 30 mg daily after the diagnosis of CEP was made. Repeat FBC after 2 weeks of starting steroid therapy showed a dramatic decrease in peripheral blood eosinophilia. The absolute eosinophil count decreased from 9.38×10⁹/L (56.6%) to 1.07×10⁹/L (13.5%). Her symptoms of cough and breathlessness had also improved.

OUTCOME AND FOLLOW-UP
Following initial steroid therapy at 0.5 mg/kg/day, our patient continued to improve, and the steroid dose was tailed down over the course of a year. However, she did have occasional relapses with worsened exertional dyspnoea and required uptitration of steroid doses each time.

DISCUSSION
The concurrent appearance of blood eosinophilia and pulmonary lung infiltration is described as pulmonary eosinophilia or pulmonary infiltration with eosinophils syndrome. The defining characteristics of pulmonary eosinophilia include: peripheral blood eosinophilia with abnormalities on pulmonary imaging, lung tissue eosinophilia and increased eosinophils in BAL fluid.1–3

CEP was first described by Carrington in 1969, where he described nine patients who presented with a syndrome of chronic and life-threatening illness with high fever, night sweats, weight loss and severe dyspnoea.4 It is a rare disorder with a reported incidence of 0.23 per 100,000 population per year between 1990 and 20044 and account for up to 2.5% of all interstitial lung disease cases in Europe.5 The cause of CEP is currently unknown but may involve selective migration of T-helper 2 cells to the lungs and release of interleukin 5 and related cytokines, resulting in eosinophilic accumulation in lungs and production of toxic eosinophilic products.5

To our knowledge, there are no specific agreed diagnostic criteria for idiopathic CEP. Diagnosis is usually based on the

Figure 1  (A) Chest radiograph at time of initial presentation. (B) Chest radiograph 6 months later.
association of: respiratory symptoms of more than 2 weeks’ duration, alveolar and/or blood eosinophilia (BAL differential eosinophil count >25%, but typically ≥40%, blood eosinophilia ≥1000/μL), pulmonary infiltrates in a predominantly peripheral pattern on chest imaging, as well as exclusion of any known causes of eosinophilia. CEP typically affects patients in their 30s to 40s, and a history of atopy is found in up to 60% of these cases. The onset of disease is typically insidious and a mean time to diagnosis from presentation can take up to 5 months. Common presenting symptoms include cough, dyspnoea, fever, night sweats and weight loss. Asthma may be present in up to 50% of patients and may occur before, during or after the diagnosis of CEP. No laboratory studies are specific for CEP. Peripheral blood eosinophils are typically >1000/μL, accompanied by a high erythrocyte sedimentation rate and C reactive protein. Serum IgE is elevated in nearly 50% (mean of 1214 ng/mL). Spirometry in CEP may be obstructive,

Table 1

| Cell type          | Absolute cell count | Percentage |
|--------------------|---------------------|------------|
| Macrophages        | 41.5                | 10.4       |
| Lymphocytes        | 7                   | 1.8        |
| Neutrophils        | 9.5                 | 2.4        |
| Eosinophils        | 333.5               | 83.3       |
| Mast cells         | 0                   | 0          |
| Ciliated epithelial cells | 2               | 0.5        |
| Squamous epithelial cells | 6.5              | 1.6        |

Figure 2  (A) Contrasted CT of the chest in the axial plane showing bilateral apical ground glass changes with thickening of the interlobular septa at the apices of both lungs, more marked over the apical segment of the left upper lobe. (B) Contrasted CT of the chest in the axial plane showing absence of ground glass changes and septal thickening in the midlung. (C) Contrasted CT of the chest in the axial plane (soft tissue window) showing lack of adenopathy. (D) Contrasted CT of the chest in the coronal plane showing upper lobe predominance of ground glass opacities.

Figure 3  Bronchoalveolar lavage at high magnification (×40) showing predominantly eosinophilic yield (Diff-Quick stain).
administration is typically dramatic, with response seen within 48 hours. An alternative diagnosis should be sought if patients do not improve quickly with steroids. Clinical response can be measured by improvement in symptoms, decline in pulmonary or peripheral blood eosinophilia, marked reduction or clearing of radiographic abnormalities, as well as physiological improvement on spirometry or diffusion coefficient.

Both symptomatic and radiographic relapse is common in CEP after cessation of therapy or during tailing of steroid doses. Time of relapse can occur any time from months to years after the initial presentation. Management of relapse includes increasing steroid dose up to 0.5 mg/kg/day for the next 1 to 2 weeks. Although the total dose and length of treatment may vary among patients, one study showed that up to three-quarters of patients require prolonged steroid therapy, with a mean duration of 19 months. In the study by Marchand et al., up to 69% of patient were still on oral corticosteroid therapy over the mean follow-up period of 6.2 years. Some studies also recommend the use of inhaled corticosteroids to lower the rate of CEP relapse.7

Figure 4  Transbronchial lung biopsy under low power (×10) magnification with H&E stain showing large collections of eosinophils predominantly in the interstitium. (B) Transbronchial lung biopsy under high power (×40) magnification with H&E stain showing large collections of eosinophils predominantly in the alveolar spaces and interstitium.

A restrictive or normal. A case series of 19 patients revealed an obstructive pattern in 21%, a restrictive pattern in 47% and a normal pattern in 32%. Diffusion studies usually reveal reduced carbon monoxide transfer factor and carbon monoxide transfer coefficient. Radiological findings typically show bilateral peripheral, non-segmental, consolidative opacities described as the 'photographic negative' of pulmonary oedema. However, the 'photographic negative' pattern on the chest radiograph is present in only one-fourth of patients and is not specific for CEP. Bronchoscopy in CEP typically shows BAL differential eosinophil counts of >25%. Only rarely is eosinophilia absent in BAL, in which case lung biopsies are performed. Otherwise, lung biopsy is required to make a diagnosis of CEP. If lung biopsies are performed, transbronchial lung biopsy is usually sufficient and surgical lung biopsies are rarely required. Also, bone marrow biopsy may not be necessary in the initial workup of the typical case of chronic eosinophilic pneumonia but may considered should the eosinophilia not respond to treatment with steroids.

The mainstay of treatment in CEP includes the use of steroids. However, to the best of our knowledge, there has been no consensus recommendation on the duration and dose of steroid therapy. In some studies, acutely unwell patients with rapidly progressive disease received 3 to 5 days of high-dose steroids (intravenous methylprednisolone 250 mg every 6 hours), after which they were converted to oral prednisolone once they are stabilised. All other patients were started on oral prednisolone, typically at a dose of 0.5 mg/kg/day and tapered down after 2 weeks of treatment. Clinical improvement following steroid...
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