Modified Atmosphere Packaging Failure and Its Effect on the Microbiological Quality of the Product

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Abstract: In the present study, microbiological, physicochemical and sensory analyses have been conducted in order to study the effects of damaged modified atmosphere packaging (MAP) on foods sampled from supermarket refrigerators. The products had not expired and no other lesions were apparent on them, except for the damaged packaging. 80 samples of smoked turkey and 70 samples of smoked pork were analysed for Total Viable Count, Enterobacteriaceae, Staphylococcus sp., Pseudomonas sp., Bacillus sp., Clostridium perfringens, Lactic acid bacteria (LAB), Bronchothrix thermosphacta and yeasts-moulds. Finally, the changes in the color and the odor were investigated. In each case products with unharmed packaging were used as controls. The safety and stability of food depends on the multiplying microorganisms which initially present being unable to overcome various adverse intrinsic and extrinsic factors to the food, as well as to the contact microflora when rupture of the sealed packaging occurs. Models have been developed to predict the consequences of pack film properties such as the size and number of micro-perforation per pack on gas concentration and to define minimum homogeneity requirements for MAP. Our results indicate that potential hazards exist concerning public health safety, even if the slightest damage of the packaging has occurred.

Key words: Modified atmosphere packaging (MAP), public health.

1. Introduction

Meat products and consumption in the West may be traced back for at least 2,500 years. The dominant cultural source was Greco-Roman, with evidence from archaeology, surviving documents and even from the names of meat cuts existing today [1]. In the EU and particularly in Greece, overconsumption of meat products is related to a social welfare despite the risks involved under the modern way of living [2]. It is not only because of the economic importance, but also from a public health perspective, that meat industry and the scientific community aimed towards the development of novel techniques and practices to produce and preserve such products.

In recent years, there is an increased progress in packaging technologies in order to fulfil the consumer’s demand for minimally processed and safe products with extended self lives [3]. Today, the Modified Atmosphere Packaging (MAP) including vacuum is used as an efficient preservative practice across a wide range of products which have earned the consumers confer about their safety. Additionally, the implementation of quality management practices and HACCP system certification by the industry and the retail market have increase the conferment of safe foods, although various studies have shown that more efforts required to achieve that goal [4, 5]. Thus, the consumers are getting more and more reluctant or disregard when comes to their obligation
in food inspection in retail or local markets and they are often unaware of the possible health risks [6]. In Greece, all food products available in retail markets are inspected by proper authorities according to a monitoring scheme. But, it seems that there is an undefined number of products suffering from packaging failure which are available to the consumers because there is a debate between the supermarkets and their suppliers about withdrawal policy. Suppliers do not accept their products back as the expiration date is still valid and no “apparent” signs of spoilage exist. At the same time, supermarket managers claim that their employees are experienced enough to distinguish the suspect products from slight differences in their color or the package feeling in the case of vacuumed products. Therefore, the need of an objective evaluation of meat spoilage is still of great importance [7].

In the present study, microbiological analyses have been conducted to various samples handed from a supermarket chain, in order to study the effects of probable damaged packaging on the microbiological stability of the product.

2. Materials and Methods

2.1 Sampling

All meat products were handed to us from branches of a supermarket chain in Northern Greece within an eight months period. All samples were turkey and pork meat products, packaged in vacuum or under modified atmosphere and according to the stores managers had sings of spoilage as discoloration, odour and the “feeling” of package failure. The expiration date of those products was still valid and their distributors or manufactures were unwilling to replace them. In total, 150 samples were collected of which 80 were turkey and 70 pork smoked or sliced products. Products were transferred chilled to the laboratory and analysed within 24 h. Thirty similar products with no apparent package damage were purchased from the same stores and served as controls.

2.2 Microbiological Analysis

Ten grams of each sample were placed in a sterile stomacher bag containing 90 mL of peptone water and homogenized for 5 min, followed by a series of dilution in peptone water. Total viable count, were determined using Plate Count Agar (Oxoid CM 325). Plates incubated at 30 °C for 24-48 h. Enterobacteriaceae were counted on Violet Red Bile Glucose Agar (Oxoid CM 485) aerobically and incubated at 37 °C for 24-48 h. Pink-red colour colonies with precipitation were taken into consideration. Staphylococci were determined on Baird Parker Agar (Oxoid CM 275) incubated aerobically at 37 °C for 24-48 h. Typical black colonies with zones around and atypical black colonies were considered as *Staphylococcus* sp.. *Pseudomonas* sp. were isolated on Pseudomonas Agar (Oxoid CM 559) incubated aerobically at 30 °C for 24-48 h. Oxidase (+) colonies were taken into consideration. *Bacillus* sp. were isolated on Cereus Selective Agar (Merck 1.05267) aerobically at 30 °C for 24-48 h. Pink-purple, opaque colonies were chosen for further examinations (Gram stain, catalase test, motility test, nitrate reduction, tyrosine decomposition, anaerobic fermentation of glucose, VP reaction, production of acid from mannitol and arabinose). Rose Bengal Chloramphenicol Agar (Oxoid, CM 0549) was used to isolate yeasts and moulds after incubation anaerobically at 30 °C for 4-5 days. To confirm the presence of *C. perfringens*, the LS (Lactose-Sulfite) medium was used. An aliquot of each sample was also heated for 20 min at 80 °C for detection of germinated spore forms. pH modified MRS agar was used for the detection of Lactic Acid Bacteria and STAA Medium (Oxoid CM0881 plus SR0151 supplement) for the detection of *B. thermosphacta* after incubation aerobically at 22 °C for 24 h.

The color of the samples was visually compared with that of the controls. Any differentiation in the odor was also recorded.
2.3 Statistical Analysis

All bacterial counts were logarithmically transformed prior to statistical analysis. Mean counts between the various groups of samples were compared with the Students’ t-test (two samples) or ANOVA. The presence or absence (prevalence) of C. perfringens vegetative and spore forms between the groups of samples was statistically evaluated by using the Chi-square procedure.

3. Results and Discussion

Table 1 presents the results from the microbiological examination of our samples. Total Viable Counts (TVC) were abundant in all samples, either turkey or smoked pork, and ranged from 4.0 to 9.1 log cfu/g with an average of 6.12 and 6.90 log cfu/g respectively. Enterobacteriaceae ranged from 1.3 to 7.5 log cfu/g with average values 2.14 log cfu/g for turkey and 4.95 log cfu/g for pork. Staphylococcus sp. ranged from 1.1 to 3.2 log cfu/g (average 1.76 and 2.19 log cfu/g), Pseudomonas sp. ranged from 4 to 9 log cfu/g (average 6.3 and 7.24 log cfu/g), Bacillus sp. from 1.1 to 3.6 log cfu/g (average 1.84 and 2.42 log cfu/g), B. thermosphacta from 3.8 to 8.9 log cfu/g (average 6.87 and 7.18 log cfu/g), moulds and yeasts ranged from 1.1 to 5.7 log cfu/g (average 2.19 and 2.97 log cfu/g) and Lactic acid bacteria ranged from 5.4 to 7.8 log cfu/g (average 6.09 and 6.55 log cfu/g). The 8.75% of turkey samples were positive in vegetative forms of C. perfringens and 27% in spore forms, while in pork samples the respective values were 30% and 28.6%. L. monocytogenes was not detected in any of the samples.

Statistically significant differences were observed

| Bacterial species          | Turkey meat products | Pork meat products |
|----------------------------|----------------------|-------------------|
|                            | Mean | St Dev | Min | Max | Mean | St Dev | Min | Max |
| Total viable count         | 6.12† | 0.93 | 4 | 8 | 6.9† | 1.26 | 4.1 | 9.1 |
| Controls                   | 4.65 | 0.80 | 3.5 | 6.1 | 4.4 | 0.35 | 4.1 | 5.1 |
| Enterobacteriaceae         | 2.14† | 0.50 | 1.3 | 3.9 | 4.95† | 1.52 | 1.8 | 7.5 |
| Controls                   | 2.05 | 0.54 | 1.1 | 3.1 | 2.35 | 0.61 | 1.6 | 3.5 |
| Staphylococcus sp.         | 1.76† | 0.46 | 1.1 | 3 | 2.19† | 0.56 | 1.1 | 3.2 |
| Controls                   | 2.12 | 0.46 | 1.2 | 3.1 | 1.89 | 0.56 | 1.1 | 3 |
| Pseudomonas sp.            | 1.84† | 0.44 | 4.4 | 7.9 | 7.24† | 1.25 | 4 | 9 |
| Controls                   | 4.47 | 0.68 | 3.6 | 6.3 | 4.39 | 0.42 | 3.9 | 5.2 |
| Bacillus sp.               | 1.84† | 0.44 | 1.1 | 3 | 2.42† | 0.64 | 1.1 | 3.6 |
| Controls                   | 0.53 | 0.79 | 0 | 2.1 | 0.87 | 0.88 | 0 | 2.1 |
| B. thermosphacta           | 6.87† | 0.81 | 4.8 | 8.8 | 7.18† | 1.25 | 3.8 | 8.9 |
| Controls                   | 3.83 | 0.42 | 3.2 | 4.7 | 3.84 | 0.25 | 3.4 | 4.2 |
| Yeasts-moulds              | 2.19† | 0.50 | 1.1 | 4 | 2.97† | 0.91 | 1.8 | 5.7 |
| Controls                   | 2.33 | 0.25 | 2 | 2.8 | 3.59 | 0.28 | 3.1 | 4.1 |
| L. monocytogenes           | nd   | nd   | nd | nd | nd   | nd   | nd | nd |
| Lactic acid bacteria       | 6.09† | 0.38 | 5.4 | 7 | 6.55† | 0.62 | 5.5 | 7.8 |
| Controls                   | 3.44 | 0.49 | 2.9 | 4.8 | 3.07 | 0.28 | 2.4 | 3.5 |
| C. perfringens-vegetative  | 8.75%† | 30.0%† |
| Controls                   | 20.0%* | 6.67%* |
| C. perfringens-spores      | 27.0%* | 28.6%* |
| Controls                   | 33.3%* | 20.0%* |

† Denotes statistically significant differences with its corresponding control group; ‡ Denotes statistically significant differences between the same bacterial species for turkey and pork meat products; * Percentage of positive samples; nd: not detected.
between the turkey and pork mean bacterial counts regarding all species \( (t: < -3, P < 0.05) \), except \( B. \) thermosphacta \( (t: -1.9, P > 0.05) \) and the occurrence of \( C. \) perfringens spore forms \( (X^2: 0.46, P > 0.05) \). Pork meat was found to have the highest number of organisms present compared to turkey (Fig. 1). \( Pseudomonas \) sp. by 17.5% and \( B. \) thermosphacta by 46.25% of pork samples exceeded seven logs which are considered as a spoilage indicator [8]. \( Enterobacteriaceae \) were also recovered in relatively large numbers, especially from pork samples, indicating that the sanitary status of the product was compromised [9]. Unusual color and/or odours during the examination were observed only in a small fraction of the samples (6% and 2% respectively).

Sour odor indicates an aerobic spoilage due to the packaging failure [10]. Besides the occurrence of bacterial species those samples did not exhibit profound spoilage such as tense discoloration or slimy appearance, probably as a result of the low storage temperature or due to the presence of preservatives [11]. Our results indicate that not all suspicious samples could be considered as spoiled given that no other physicochemical properties were examined in them. But the microbiological profile in the majority could raise questions about their safety.

4. Conclusion

It is apparent that there is an occurrence of elevated bacterial counts in meat products suffering from a possible packaging failure and therefore an increased risk for the consumers’ exits. Meat products companies and resellers should cooperate and withdraw those products with signs of packaging failure without relying solely on the predefined expiration date.

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