Microvascular density and hypoxia-inducible factor in intraepithelial vocal fold lesions

Anna Rzepakowska1 · Michał Żurek2 · Jakub Grzybowski3 · Paweł Pihowicz3 · Barbara Górnicka3 · Kazimierz Niemczyk1 · Ewa Osuch-Wójcikiewicz1

Received: 13 October 2018 / Accepted: 20 February 2019 / Published online: 6 March 2019 © The Author(s) 2019

Abstract

Objective The promotion of neovascularisation is a crucial aspect of carcinogenesis. The study evaluates the microvascular density (MVD) and expression of hypoxia-induced factor (HIF-1α) in hypertrophic vocal fold (VF) lesions of different histopathological states including non-dysplastic, low-grade, high-grade dysplasia and invasive glottic cancer.

Materials and methods Histological specimens collected from patients diagnosed and treated in a single centre with different histological grades were immunohistochemically stained with CD31, CD34 and HIF-1α. Of the total number of 77 analysed VF specimens, 20 were non-dysplastic, 20 had low-grade dysplasia, 17 high-grade dysplasia and 20 were invasive cancers.

Results The highest mean value for MVD evaluated with expression of CD31 (MVD CD31) was 21.23 ± 14.46 and identified in the low-grade dysplasia group. The average MVD CD31 was 13.74 ± 5.56 and 20.11 ± 9.28 in the high-grade dysplasia and invasive cancer group, respectively. The highest MVD evaluated with CD34 (MVD CD34) was revealed for invasive cancer 35.64 ± 17.21. The MVD CD34 was higher for low-grade than in high-grade dysplasia (25.87 ± 12.30 vs 24.65 ± 15.92, respectively). The expression of HIF-1α was strong or very strong in 60% of non-dysplastic lesions, 100% of low-grade dysplasia cases, 53% of high-grade dysplasia cases and 50% of invasive cancer cases. The comparison of MVD CD31 with MVD CD34 revealed a strong positive correlation (ρ value 0.727). The comparison of both MVD CD31 and MVD CD34 with HIF-1α resulted in no linear relationship (ρ value of 0.143 and 0.165, respectively).

Conclusion The stage of low-grade dysplasia in intraepithelial vocal fold lesions is related to significant advancement of angiogenesis together with the highest hypoxia level.

Keywords Intraepithelial lesions · Dysplasia · Glottic cancer · Larynx · Microvascular density · Hypoxia · Angiogenesis

Introduction

The model of the genetic changes responsible for progression of hypertrophic laryngeal lesions to dysplastic and invasive cancer is still incomplete and lacking [1]. The architectural and cytological disorder found within epithelial cells can be histopathologically classified as squamous hyperplasia, dysplasia or invasion. This pathology is controlled by a multistep process of genetic changes, which not only influence the potential for cellular multiplication at this level, but also inevitably alter the surrounding microenvironment indirectly, providing for the most optimal conditions for tumour expansion [2]. Neoangiogenesis, defined as the growth of new vessels, is essential for neoplastic lesions during their process of progression and is, therefore, one of the fundamental events in the carcinogenesis [3]. The formation of neoplastic microvessels is stimulated by several other...
growth factors including vascular endothelial growth factor (VEGF), transforming growth factor (TGF) and various cytokines secreted by the altered cells [3]. However other triggering conditions of the microenvironment, including the state of hypoxia, are also critical for these processes of tumour formation [4].

Hypertrophy of vocal fold (VF) epithelium is most often caused by smoking, particularly when combined with the consumption and abuse of high-percentage alcohol. Active gastroesophageal reflux disease (GERD) in patients is another risk factor for the emerging hypertrophy. Exposure to the mentioned agents is responsible for promotion of both non-dysplastic and dysplastic lesions, as well as invasive cancer [1]. Despite there being new and emerging literature on human papilloma virus (HPV) as related to neoplasia within the larynx, the published data remain controversial and necessitate further investigation [5]. The clinically identified hyperplasia or leukoplakia of the vocal folds can provide for a wide range of histopathological diagnoses. During the last few decades, there was no consensus on a unified pathological classification of precancerous laryngeal lesions and three different rating systems were valid and in use [5]. Fortunately, the newest 4th edition of the World Health Organization Classification of Head and Neck Tumours from 2017 was able to consolidate the terminology and grading systems into a more cohesive and uniform framework. Currently under this novel classification system, precancerous lesions are classified according to their grade into either low- or high-grade dysplasia, or invasive cancer [5]. Carcinoma in situ can be distinguished from high-grade dysplasia, through the presence of severe nuclear and cytologic atypia, but is only done so if it influences the treatment decision [5]. Morphologically, the criteria for low-grade dysplasia include pathologies that range from squamous hyperplasia to basal cell enlargement, display no or minimal cellular and nuclear atypia of the epithelium, and is limited to the lower one-half of its thickness. High-grade dysplasia, on the other hand, is diagnosed when atypical epithelial cells are present and involve the lower one-half of the epithelium, with the potential to reach full thickness. Infiltration of atypical cells through the basal membrane is the definition of invasive cancer [5].

VF epithelium is a layer composed of stratified squamous cells, where only the deepest layer of basal cells has contact with the stromal tissue. In light of current research, the previous theory that the epithelial surface becomes deprived of blood vessels has been undermined [4]. The microarchitecture of stromal blood vessels provides numerous sprouts that penetrate with a thin layer of stromal tissue upwards between epithelial cells [4]. A study by Arens et al. confirmed the thickness of the epithelial layer increases in accordance with the grade of premalignant VF lesions [6]. Such epithelial proliferation translates into an increased demand for nutrients. Neoangiogenesis is, therefore, inevitably preceded by a level of ischemia [4]. Hypoxia not only promotes vascular growth, but also favours the synthesis of procarcinogenic factors, including hypoxia-inducible factor (HIF-1α), which as a consequence may initiate malignant transformations [7].

Our study was designed to investigate the relation of microvascular density and hypoxia in premalignant and malignant vocal fold lesions and enable for a better understanding of the mechanisms involved in the carcinogenesis of the laryngeal epithelium. This can be further utilised in the future development of more targeted and individualized therapies, particularly during the course of early treatment and prevention, where changes to intervention can influence patient outcomes.

Materials and methods

The protocol of the study was approved by the Institutional Ethic Review Board of the local Medical University.

This retrospective study was performed in 2018 by searching archived medical records and paraffin-embedded tissue blocks of patients treated with laryngeal microsurgery in our department from 2017 to 2018 due to suspicious hypertrophic lesions of the VFs. The design of the study assumed inclusion of 80 patients with different histopathological (HP) grades of VF lesions: non-dysplastic (stromal edema, inflammatory changes), low-grade dysplasia, high-grade dysplasia (with carcinoma in situ) and invasive cancer. The exclusion criteria included a history of radiotherapy to the neck area, papillomatosis related to HPV infection, and a history of immunodeficiency or prior systemic chemotherapy. A total of 79 patients (19 in the high-grade dysplasia group and 20 cases for each of the remaining three groups) were collected. The laryngeal specimens obtained during microsurgical resection or tissue biopsy were re-examined by two histopathologists, blinded to the previous diagnosis, to confirm the histopathology result. In one case of high-grade dysplasia, there was uncertainty regarding the invasion into the basal membrane since it could not be resolved due to the vestigial stroma as this was thermally damaged with the laser during resection. Another case of high-grade dysplasia showed findings suspicious of a coexisting HPV infection and was, therefore, rejected. Ultimately, a total number of 77 cases with a confirmed histopathological diagnosis were included for further analysis.

Immunohistochemistry

Four serial sections of 3–5 µm thickness were obtained from 77 formalin-fixed paraffin-embedded tissue blocks with a microtome (HM 340E Electronic Rotary Microtome, Thermo Shandon). All sections were stained
with hematoxylin and eosin (H&E) in an automatic tissue processor (ASP 6026, Leica, USA). Corresponding sections were stained with anti-CD31 antibody (clone: JC70A; DAKO/Agilent, USA), anti-CD34 antibody (clone: QBEnd10; DAKO/Agilent, USA) and anti-HIF-1α antibody (clone: EP1215Y; Abcam, UK).

Immunostaining was performed using EnVision FLEX, High pH (Link) (DAKO/Agilent, USA) and Autostainer Link 48 (DAKO/Agilent, USA).

In brief, the tissue sections were deparaffinized by incubation in a wash buffer, pH 9.0, at 98 °C for 20 min in PT Link station (DAKO/Agilent, USA). After 10 min of incubation in the wash buffer at room temperature, the endogenous peroxidase activity was blocked through immersion in a 3% hydrogen peroxide solution for 10 min. The slides were then incubated for 20 min with the primary antibodies, followed by the addition of the secondary antibody. In the case of anti-CD31 antibody, the reaction was strengthened through the addition of Mouse LINKER according to the producer’s recommendations. Visualization was achieved by the application of 3,3′-diaminobenzidine. Finally, cell nuclei were counterstained with hematoxylin. The tissue sections were dehydrated with graded concentrations of ethanol, cleared with xylene and covered with tape cover slippers (Klinipath, The Netherlands).

The assessment of the immunostainings was carried out by two examiners using light microscopy and the results were averaged.

**Evaluation of microvessel density**

First, each slide was examined at low magnification (40×), selecting regions with the highest density of vessels (hot spots) before being viewed at high magnification (200×). CD31-positive or CD34-positive vessels were counted in three or five hot spots, respectively. Only continuous, membranous staining was considered as positive. Any large microvessel with a lumen or any single, separated endothelial cell was given a count of one. The vessels were counted within the epithelium and at the epithelium/stroma edge. Figure 1 presents examples of H&E and immunostaining for both CD31 and CD34 in each grade of intraepithelial VF lesions at 100× magnification.

**Evaluation of HIF-1α expression**

There are no standardized levels of HIF-1α staining. The evaluation of this staining was only quantitative and a specimen was considered as positive if more than 1% of cell nuclei were stained. Membranous or cytoplasmic staining was not assessed in this study. Using the methodology found in majority of publications with positively stained samples as a basis, the following classification was adopted: 1–25% positive nuclei—weakly positive expression, 26–50% positive nuclei—moderately positive expression, 51–80% positive nuclei—strong positive expression, more than 80% positive nuclei—very strong positive expression. Figure 2 presents four categories of expression of HIF-1α immunostaining in intraepithelial VF lesions at 100× magnification.
Statistical analysis

All parameters were evaluated using SPSS 18.0 and Statistica 13 for the analyses and a $p$ value lower than 0.05 was considered statistically significant. The differences between groups with different histopathological diagnosis and expression of CD31, CD34 and HIF-1α were compared using nonparametric Kruskal–Wallis and Mann–Whitney $U$ tests. Box plots were used for a graphical representation of the received results. The Spearman rank correlation coefficients were used to calculate the correlation between microvessel density (MVD) and the expression of CD31 and CD34 alike, as well as the relationship between MVD and HIF-1α levels.

Results

The immunohistochemical analysis was finally performed on sections derived from 77 VF lesions. The patient age ranged from 23 to 89 years, with a median of 61.96 years and standard deviation of 11.62 years. There were 54 males and 23 females among the studied samples. The groups with non-dysplastic lesions, low-grade dysplasia and invasive cancer consisted of 20 patients each. The group with high-grade dysplasia included 17 patients. Detailed demographic data of each group are presented in Table 1.

Microvascular density evaluated with expression of CD31 (MVD CD31) in endothelial cells revealed statistically significant differences ($p = 0.0115$) in microvascular count between the analysed groups. The highest mean value of MVD CD31 expression ($21.23 \pm 14.46$) was identified in lesions with low-grade dysplasia. Interestingly, the average MVD CD31 was $13.74 \pm 5.56$ for the high-grade dysplasia group and $20.11 \pm 9.28$ for the invasive cancer group. Detailed statistics of MVD CD31 expression for each group are presented in Table 2. Figure 3 presents a graphical representation of data for MVD CD31 expression in the analysed groups.

CD34 expression in endothelial cells was used for the comparative assessment of microvascular density (MVD CD34). Statistical analysis confirmed a significant difference ($p = 0.0001$) in microvascular count between the analysed groups for this method as well. However, the highest mean value for MVD CD34 was revealed this time in lesions with invasive cancer $35.64 \pm 17.21$, but was also found to be higher in low-grade than in high-grade dysplasia ($25.87 \pm 12.3$ vs $24.65 \pm 15.92$, respectively). The statistical results of MVD CD34 are presented in Table 3. Figure 4 presents the statistical data graphically for MVD CD34 in the analysed groups.

On immunohistochemical staining of laryngeal specimens for HIF-1α, strong or very strong expression was identified in 60% of non-dysplastic lesions, 100% of low-grade dysplasia, 53% of high-grade dysplasia and 50% of invasive cancers. The details concerning expression of HIF-1α in different grades of histopathological diagnoses are presented in Table 4.

The comparison of MVD evaluated with CD31 and CD34 revealed a strong positive correlation of both methods ($\rho$ value of 0.727; range 1 to $-1$). However, individual comparisons of both MVD CD31 and MVD CD34 with HIF-1α resulted in no linear relationship ($\rho$ values of 0.143 and 0.165, respectively).

Table 1. Characteristic of patients with vocal fold lesions

| Category             | Number | Female/male | Mean age ± SD; range (years) |
|----------------------|--------|-------------|------------------------------|
| Non-dysplasia        | 20     | 12/8        | 55.15 ± 12.38 (range 23–71)  |
| Low-grade dysplasia  | 20     | 3/17        | 61.55 ± 10.65 (range 35–85)  |
| High-grade dysplasia | 17     | 4/13        | 64.94 ± 11.25 (range 37–89)  |
| Invasive cancer      | 20     | 4/16        | 66.65 ± 9.31 (range 56–88)   |
| All patients         | 77     | 23/54       | 61.96 ± 11.62 (range 23–89)  |
Table 2 The evaluated microvessel density (MVD) with CD31 in different grades of histopathological diagnosis

|                      | CD31 No. of specimens | CD31 MVD Mean | CD31 MVD 95% confidence interval (CI) | CD31 MVD Standard deviation (± SD) | CD31 MVD Standard error | CD31 MVD Range | p value       |
|----------------------|------------------------|---------------|--------------------------------------|-----------------------------------|-------------------------|----------------|--------------|
| Non-dysplasia        | 20                     | 12.47         | 8.53–16.40                           | 8.41                              | 1.88                    | 3.00–33.33     | 0.02*         |
| Low-grade dysplasia  | 20                     | 21.23         | 14.47–28.00                          | 14.46                             | 3.23                    | 5.33–56.00     | 0.12#         |
| High-grade dysplasia | 17                     | 13.74         | 10.89–16.60                          | 5.56                              | 1.35                    | 5.67–23.67     | 0.03*         |
| Invasive cancer      | 20                     | 20.11         | 15.77–24.46                          | 9.28                              | 2.07                    | 6.00–45.33     |              |
| All patients         | 77                     | 17.01         | 14.60–19.43                          | 10.63                             | 1.21                    | 3.00–56.00     | 0.01†         |

*Statistically significant p value (< 0.05)

Fig. 3 The boxplot diagram presenting microvascular density with CD31 (mean, standard deviation) for non-dysplastic, low-grade dysplasia, high-grade dysplasia and invasive cancer of vocal folds

Table 3 The evaluated microvessel density (MVD) with CD34 in different grades of histopathological diagnosis

|                      | CD34 MVD No. of specimens | CD34 MVD Mean | CD34 MVD 95% confidence interval (CI) | CD34 MVD Standard deviation (± SD) | CD34 MVD Standard error | CD34 MVD Range | p value       |
|----------------------|---------------------------|---------------|--------------------------------------|-----------------------------------|-------------------------|----------------|--------------|
| Non-dysplasia        | 20                        | 13.96         | 9.69–18.23                           | 9.12                              | 2.04                    | 2.00–31.80     | 0.004*        |
| Low-grade dysplasia  | 20                        | 25.87         | 20.11–31.63                          | 12.30                             | 2.75                    | 2.80–49.40     | 0.61*         |
| High-grade dysplasia | 17                        | 24.65         | 16.46–32.83                          | 15.92                             | 3.86                    | 4.00–57.00     | 0.03*         |
| Invasive cancer      | 20                        | 35.64         | 27.59–43.69                          | 17.21                             | 3.85                    | 10.00–92.60    |              |
| All patients         | 77                        | 25.04         | 21.47–28.62                          | 15.75                             | 1.80                    | 2.00–96.60     | 0.0001†        |

*Statistically significant p value (< 0.05)

⁎p value for the Mann–Whitney test for the comparison of values between the two groups
†p value for Kruskal–Wallis for overall assessment of differences between groups
Laryngeal lesions, of premalignant and malignant nature, have been a topic of discussion and debate focusing on laryngeal pathology. It has been proven that 90% of laryngeal cancers are preceded by precancerous lesions of leukoplakia or mucosa hypertrophy characterized histopathologically as epithelial dysplasia [7]. Therefore, identification and management of intraepithelial lesions are imperative in the prevention of invasive neoplasia. Understanding the mechanisms of the processes involved in malignant transformation forms the basis of proper treatment. The period from first diagnosis of laryngeal dysplasia to malignant transformation correlates with the grade of dysplasia [8]. Identification of biochemical factors that correlate with the progression of intraepithelial dysplasia progression to invasive cancer would enable for individualisation of treatment strategies in addition to early intervention. Current therapy for non-dysplastic and low-grade dysplasia includes two options: “watchful waiting” or microsurgical excision. For high-grade dysplasia, a more radical excision is proposed (subepithelial cordectomy of type II). Present day recommendations for invasive early glottic cancer include surgical excision or radiotherapy as equivalent methods. Publications concerning
angiogenesis or hypoxia in intraepithelial laryngeal lesions are very scarce, however. Kojc et al. found that in contrast to invasive cancer, laryngeal intraepithelial lesions stained positively for CD34 stromal cells but there was no reaction with myofibroblast antibodies in the stroma [9]. Szafrański et al. reported a significant increase in microvessel density identified with CD34 and CD105 in laryngeal cancer and dysplasia when compared to the healthy control group [10]. Prior research on microvessel density in intraepithelial lesions of the oesophagus and cervix confirmed an increased vessel number associated with the transformation from intraepithelial lesions to invasive cancers [11, 12]. CD31 and CD34 are most commonly utilised as endothelial markers in the literature. While CD31 is detected in matured endothelial cells, CD34 is expressed in both precursor and differentiated endothelial cells, making this marker less specific. Anti-CD34 antibodies also immunostain lymphatic endothelial cells, fibroblasts and adipocytes, and may falsely elevate the values for CD34 MVD in comparison to the CD31 MVD count.

Studies centred on angiogenesis and vessel formation most often correlate MVD with inflammatory states, notably regarding the number of macrophages. We did not find any works comparing hypoxia with MVD in intraepithelial lesions. During the study design, it was expected to observe an increase in MVD with the rise in severity of histopathological grade in laryngeal lesions. An increase in hypoxia factor seemed to correlate with increased neovascularization. It is, however, noteworthy that mean vessel number was higher in the low-grade as opposed to high-grade dysplasia, as was witnessed in both methods of MVD evaluation. Even more intriguing is the pattern of HIF-1α expression in low-grade dysplasia, where 100% of lesions had a positive reaction in more than 51% of cell nuclei found within lesions. Tobacco exposure and low-pH states chronically irritate the epithelium, putting it under much oxidative stress. The pathological analysis of epithelial thickness in different stages of intraepithelial lesions performed by Arens et al. confirmed the epithelium becomes thicker with the progression of HP status [6]. This strong expression of HIF-1α during the early stage of dysplasia can be associated with an increase in nutritional demands of proliferating cells. This elevated hypoxia factor may not only promote further neangiogenesis, but also influence transpiration at this stage and development of cellular atypia and invasiveness.

Using prior research on angiogenesis in precancerous lesions and laryngeal cancer as a basis, the analysis of MVD and hypoxia may result in treatment individualisation in the future. However, a consensus about the influence of vascular density on laryngeal cancer prognosis has yet to be reached. A high expression of CD31 and VEGF was affirmed by recent studies to correlate with poorer clinical outcomes, including reoccurrence and survival rates, especially concerning laryngeal cancer at the early stage [13–15]. Pignatoro et al. suggested that angiogenesis may be the crucial aspect in early stages of laryngeal carcinogenesis [15]. Kwon et al. reported on enhanced expression of HIF-1α in early laryngeal cancer and its association with radio-resistance [16]. It was suggested for patients with early-stage laryngeal carcinomas displaying a high expression of HIF-1α, treatment modalities alternative to primary radiotherapy should be considered in these cases [16].

Sun et al. compared MVD CD34 in laryngeal cancer (26.5 ± 6.4) with the peritumoural tissue (12.2 ± 4.0) and found a significant difference (p < 0.05) among them, but also confirmed a higher MVD CD34 in patients with lymph node metastasis than that in the N0 group [17]. They suggested that MVD CD34 has an important role in promoting metastases in laryngeal cancer and can serve as a prognostic marker [17]. Popov et al. reported a mean MVD with CD34 in a study group of laryngeal cancer of 14.27 ± 12.92 and confirmed a significantly higher MVD in patients with metastasis than in those without [18]. Pietruszewiska et al. found a statistically significant correlation between MVD CD34 and tumour size, as well as nodal metastasis [19].

The number of papers considering the involvement of hypoxia factors in laryngeal cancer is even smaller than those evaluating angiogenesis markers. Starska et al. found a statistically significant relationship (p < 0.05) between HIF-1α with pTNM, tumour recurrence and overall survival in patients with laryngeal cancer [20]. There are further reports that higher levels of HIF-1α can be associated with hypoxia-mediated multi-drug resistance in patients undergoing chemotherapy via the inhibition of drug-induced apoptosis of laryngeal cancer cell, leading to a decrease in drug accumulation within tumour cells [21].

Gong et al. performed a systematic review of 28 studies evaluating the association between HIF-1α factors and survival in head and neck cancer [22]. The analysis revealed HIF-1α overexpression significantly correlated with worse survival outcomes in oral carcinoma, nasopharyngeal carcinoma and oropharynx carcinoma, but not in laryngeal cancer [22].

The above-mentioned studies analyse the influence of angiogenesis and hypoxia in invasive laryngeal cancer. The identification of biochemical factors affecting the progression of intraepithelial atypia and dysplasia in vocal fold lesions may be pivotal in the evolution of treatment methods and prevention of invasive laryngeal cancer development. The advancement of angiogenesis and the level of hypoxia in low-grade dysplasia suggest the occurrence of significant molecular changes already at play in the early stages and may influence further processes, ultimately leading to neoplastic transformation.
The possibility of novel treatment options comprising of antibody therapy that inhibit neoangiogenesis (bevacizumab) encourages the need for further research on neovascularization in intraepithelial vocal fold lesions. Such therapies, in the case of intraepithelial lesions, could be administered topically or as intralesional antibody therapy. Reoxygenation of the laryngeal epithelium at the early stage of hypertrophic pathology or undergoing low-grade dysplasia can potentially inhibit progression of dysplasia, atypia and invasion.

Conclusions

The stage of low-grade dysplasia in intraepithelial vocal fold lesions is related to significant advancement of angiogenesis together with the highest hypoxia level. Neoangiogenesis and hypoxia during the early stages of vocal fold intraepithelial dysplasia may influence progression of cellular atypia and the further stages of carcinogenesis.

Funding

None declared.

Compliance with ethical standards

Conflict of interest

None declared.

Ethical approval

This article does not contain any studies with animals performed by any of the authors. All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Informed consent

Informed consent was obtained from all individual participants included in the study.

Open Access

This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. Gale N, Gnepp DR, Poljak M, Strojan P, Cardesa A, Helliswell T, Šifrer R, Volavček M, Sandison A, Zidar N (2016) Laryngeal squamous intraepithelial lesions: an updated review on etiology, classification, molecular changes, and treatment. Adv Anat Pathol 23(2):84–91
2. Trivedi S, Rosen CA, Ferris RL (2016) Current understanding of the tumor microenvironment of laryngeal dysplasia and progression to invasive cancer. Curr Opin Otolaryngol Head Neck Surg 24(2):121–127
3. Lionello M, Staffieri A, Marioni G (2012) Potential prognostic and therapeutic role for angiogenesis markers in laryngeal carcinoma. Acta Otolaryngol 132(6):574–582
4. Karseladze AI (2016) Intraepithelial ischemia is a principal factor promoting cancerization of the covering epithelial tissues. Med Hypotheses 94:154–160
5. Gale N, Poljak M, Zidar N (2017) Update from the 4th edition of the World Health Organization Classification of Head and Neck Tumours: what is new in the 2017 WHO blue book for tumours of the hypopharynx, larynx, trachea and parapharyngeal space. Head Neck Pathol 11(1):23–32
6. Arens C, Glanz H, Wömckhaus J, Hersemeyer K, Kraft M (2007) Histologic assessment of epithelial thickness in early laryngeal cancer or precursor lesions and its impact on endoscopic imaging. Eur Arch Otorhinolaryngol 264(6):645–649
7. Dispensa F, De Stefano A, Marchese D, Martines F, Dispensa C (2012) Management of laryngeal precancerous lesions. Auris Nasus Larynx 39:280–283
8. Gallo A, de Vincentis M, Della Rappa C, Moi R, Simonelli M, Matti A et al (2001) Evolution of precancerous laryngeal lesion: a clinicopathologic study with long-term follow-up on 259 patients. Head Neck 23:40–47
9. Kojc N, Zidar N, Vodopivec B, Gale N (2005) Expression of CD34, alpha-smooth muscle actin, and transforming growth factor beta1 in squamous intraepithelial lesions and squamous cell carcinoma of the larynx and hypopharynx. Hum Pathol 36(1):16–21
10. Szaferowski T, Sierzchinski J, Szczepanski MJ, Whiteside TL, Ludwig N, Krzeski A (2018) Microvessel density in head and neck squamous cell carcinoma. Eur Arch Otorhinolaryngol 275(7):1845–1851
11. Jiang S, Yang Y, Fang M, Li X, Yuan X, Yuan J (2016) Co-evolution of tumor-associated macrophages and tumor neo-vessels during cervical cancer invasion. Oncol Lett 12(4):2625–2631
12. Konda VJ, Hart J, Lin S, Tretiakova M, Gordon IO, Campbell L, Kulkarni A, Bissonnette M, Seewald S, Waxman I (2013) Evaluation of microvascular density in Barrett’s associated neoplasia. Mod Pathol 26(1):125–130
13. Bolzoni Villaret A, Barbieri D, Peretti G et al (2013) Angiogenesis and lymphangiogenesis in early-stage laryngeal carcinoma: prognostic implications. Head Neck 35:1132–1137
14. Schütter A, Weller P, Kanaan O, Niel I, Heusgen L, Höing B, Hallkamp P, Zander S, Mandapathil M, Dominas N, Arnoldis J, Stuck BA, Lang S, Bankfalvi A, Brandau S (2018) CD31 and VEGF are prognostic biomarkers in early-stage, but not in late-stage, laryngeal squamous cell carcinoma. BMC Cancer 18(1):272. https://doi.org/10.1186/s12885-018-4180-5
15. Pignataro L, Carloni N, Midolo V et al (2001) Clinical relevance of microvessel density in laryngeal squamous cell carcinomas. Int J Cancer 92:666–670
16. Kwon OJ, Park JJ, Ko GH, Seo JH, Jeong BK, Kang KM, Woo SH, Kim JP, Hwa JS, Carey TE (2015) HIF-1α and CA-IX as predictors of locoregional control for determining the optimal treatment modality for early-stage laryngeal carcinoma. Head Neck 37(4):505–510
17. Sun S, Pan X, Zhao L, Zhou J, Wang H, Sun Y (2016) The expression and relationship of CD68-tumor-associated macrophages and microvascular density with the prognosis of patients with laryngeal squamous cell carcinoma. Clin Exp Otorhinolaryngol 9(3):270–277
18. Popov TM, Tikov D, Goranova TE, Stancheva G, Kaneva RP, Todorov S, Stoyanov O, Rangachev J, Marinov T, Popova DP, Mitev VI, Konov D (2015) Macrophageal infiltration and microvessel density in laryngeal carcinoma: study of 52 cases. Acta Otorhinolaryngol Ital 35(5):321–324

Springer
19. Pietruszewska W, Kobos J, Gryczyński M (2003) Microvessel density and endothelial area in assessment of angiogenesis in patients with laryngeal cancer. Otolaryngol Pol 57(1):5–15

20. Starska K, Forma E, Jóźwiak P, Bryś M, Lewy-Trenda I, Brzezińska-Błaszczyk E, Krześlak A (2015) Gene and protein expression of glucose transporter 1 and glucose transporter 3 in human laryngeal cancer-the relationship with regulatory hypoxia-inducible factor-1α expression, tumor invasiveness, and patient prognosis. Tumour Biol 36(4):2309–2321

21. Li DW, Dong P, Wang F, Chen XW, Xu CZ, Zhou L (2013) Hypoxia induced multidrug resistance of laryngeal cancer cells via hypoxia-inducible factor-1α. Asian Pac J Cancer Prev 14(8):4853–4858

22. Gong L, Zhang W, Zhou J, Lu J, Xiong H, Shi X, Chen J (2013) Prognostic value of HIFs expression in head and neck cancer: a systematic review. PLoS One 8(9):e75094

Publisher’s Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.