Epidemiology of carbapenem-resistant *Klebsiella pneumoniae* bloodstream infections after renal transplantation from donation after cardiac death in a Chinese hospital: a case series analysis

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**Abstract**

**Background:** Although the high mortality rates have been extensively reported worldwide, few studies have investigated the epidemiology of CRKP-BSIs in the early stage after kidney transplantation (KTx) from donation after cardiac death (DCD). We sought to describe the epidemiological and clinical characteristics of cases of carbapenem resistant *Klebsiella pneumoniae* bloodstream infections (CRKP-BSIs) in kidney transplantation recipients (KTRs) from DCD in our hospital.

**Methods:** A retrospective analysis of clinical data of CRKP-BSIs in KTRs admitted to a Chinese hospital in Beijing, China, between January 1, 2012 and December 31, 2016 was performed. The annual percentage of patients with CRKP, the annual number of total KTRs and KTRs from DCD were determined. The genetic relatedness of the strains was determined by polymerase chain reaction and pulse field gel electrophoresis (PFGE).

**Results:** During the study period, there were total 947 KTRs in our hospital, including 275 KTRs from DCD. Five incidences of CRKP-BSIs in KTRs were identified, and two of them (Case 1,3) from the same foreign hospital. The incidence of CRKP-BSIs in the early stage (within 3 months) following kidney transplantation (KTx) from DCD was about 1.1% (3/275). In Case 1–3 and 5, the rupture of renal transplant artery was presented on the 40th, 16th, 43th and 74th day after KTx, and in Case 4, the thrombus of renal transplant artery was presented on the 13th day after KTx. Three cases (Case 1,2,5) occurring pneumothorax on the 45th, 51th and 32th day after KTx. Four cases (Case 1–4) received the excision of the transplanted kidney for the treatment. Polymerase chain reaction showed the bands for case 2 were distinctive from other cases. Pulse field gel electrophoresis showed mainly three clusters of the bands for all the isolates. (Continued on next page)
**Background**

Renal transplantation has long been deemed to be the best therapeutic option for the treatment of the end stage of kidney failure. However, the deficiency of donors has been the enormous obstacle for the development of renal transplantation in China [1]. The main channel of renal transplantation basically comes from two resources, corpses and living organs [2]. In recent years, under more favorable conditions from aspects of ethics, law and the resources of transplantation, the renal transplantation from donation after cardiac death (DCD) has been widely performed in China. Along with the increasing demanding of organ donors and the technological advancements in the field of organ transplantation, the renal transplantation from DCD has achieved great progress in China, however many unusual complications following transplantation have been reported to date.

The rupture or thrombus of the renal transplant artery, and the infection of the surrounding tissues caused by bacteria, especially Gram-negative bacteria [3], or fungus have been regard as two of the hard-handled complications in the early stage (within 3 months) following renal transplantation. Donor-transmitted infection (DTI), including bacterial or fungal infections, has also been frequently reported from kidney recipients from DCD, characterized by Gram-negative predominant donor-derived infection, with higher crude mortality and graft loss than those without donor-derived infection [4].

As the frequently reported pathogen among the donor-transmitted infections associated with kidney transplantation from DCD, Carbapenem resistant Klebsiella pneumonia (CRKP) is a Gram-negative bacterial pathogen responsible for severe diseases, such as septicemia, pneumoniae, urinary tract infection and soft tissue infection, associated with community and hospital acquired infection [5], and characterized by high mortality, high transmissibility and multidrug resistance [6]. It has been reported that high mortality rates, ranging from 20 to 40%, were related to patients with Klebsiella pneumonia bloodstream infection (KP-BSI) [7], but this mortality was reported to rise up to 67.6% for intensive care unit (ICU) patients [8]. Through various mechanisms, such as production of carbapenem-hydrolyzing enzymes (carbapenemases) or changes in membrane permeability, carbapenem resistance can occur [9].

The patients who has undergone the renal transplantation are susceptible to CRKP, attributed to multifaceted factors, ranging from the surgery, the use of immunosuppressant therapy, mechanical ventilation, prolonged use of invasive devices, use of antimicrobial agents and the high sequential organ failure assessment (SOFA) score at admission [10–17]. In the early stage of post-transplantation, especially the first 3 months after renal transplantation, the incidence and severity of illness is directly related to the utilization of immunosuppression and its modulation that primarily intend to lessen the risk of rejection after transplantation [15].

Although the high mortality rates have been extensively reported worldwide, to date, few studies have investigated the epidemiology of CRKP-BSIs in the early stage after renal transplantation from DCD. The study was carried out aiming at describing the epidemiological and clinical characteristics of carbapenem resistant *Klebsiella pneumoniae* bloodstream infections after renal transplantation from donation after cardiac death in our hospital.

**Method**

**Study design and population**

This was a retrospective cohort study aiming at describing the epidemiological and clinical characteristics of CRKP-BSI after KTx from DCD in the 309th hospital of People Liberation of Army from January 2012 to December 2016. The 309th hospital of People Liberation of Army is a 1430-bed tertiary care teaching hospital with a 20-bed comprehensive Intensive-care-unit (ICU) and a 10-bed surgical Intensive-care-unit (SICU) and approximately 40,000 hospital admissions per year in Beijing, China. Clinical and microbiological characteristic was retrieved from medical records. Case was defined as kidney transplantation (KTx) recipient who developed CRKP-BSIs in the early stage (within 3 months) after KTx, identified by cultures of the patient's blood specimens and later confirmed by polymerase chain reaction (PCR) during the study period. Patients were followed from KTx to death or automatically discharge or discharge upon recovery.
In the analysis of the profile of utilization of immuno-suppression drugs following renal transplantation, we evaluated age; gender; cold ischemia time (CIT); immuno-suppression drugs (calcineurin inhibitor, adjuvant drug and monoclonal and polyclonal antibodies) and etiology of chronic renal failure (CRF), including systemic arterial hypertension (SAH), diabetes, glomerulonephritis (GNM), IgA nephropathy and charlson comorbidity index (CCI) score.

In the analysis of clinical characteristics of the CRKP-BSIs, we analyzed age; gender; ureteral stent; indwelling catheters; use of continuous renal replacement therapy (CRRT); blood transfusion; infection related to invasive devices; intervention for focus control (< 48 h); polymicrobial infection; acute rejection; cytomegalovirus disease; carbapenem resistant klebsiella pneumonia Bloodstream infection (CRKP-BSI); complication of infection; time (days) from kidney transplantation to the excision of the transplanted kidney; time (days) from kidney transplantation to ICU department; time (days) from kidney transplant to rupture/thrombus of the artery of the transplant kidney; time (days) from kidney transplant to the first bloodstream infection (BSI); time (days) from kidney transplantation to death/automatically discharge/discharge upon recovery; Length of stay (LOS, days) after first BSI onset; ICU-LOS (days); total LOS (days); Pitt bacteria score (PBS) on the onset of first BSI; Sequential Organ Failure Assessment (SOFA) score on the onset of first BSI; serum creatinine (µmol/L) before renal transplant; serum creatinine (µmol/L) after renal transplant; serum creatinine (µmol/L) at first BSI; serum creatinine (µmol/L) at the 7th and 14th day after first BSI; combinational antibiotic therapy; the total medical costs (¥) and prognosis.

The research project has been approved by the Ethics Committee and has therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

Microbiology
With an automated susceptibility testing system (VITEK; bioMérieux, Marcy l’Étoile, France), carbapenem-resistant strains of K. pneumoniae were identified. Minimum inhibitory concentration (MICs) were interpreted according to the Clinical and Laboratory Standards Institute breakpoints [16]. Minocycline/Tigecycline susceptibility was interpreted according to breakpoints approved by the US Food and Drug Administration [17]. All of the isolates were submitted to enterobacterial repetitive intergenic consensus polymerase chain reaction (ERIC-PCR) for the genetic relatedness analysis. ERIC-PCR specific primers (ERIC-1R, 5′-ATGTAAGCCTCCGTGGATTCAAC-3′; ERIC-2, 5′-AAGTAACTGACGTGGGTGAGCG-3′) were based on a report by Versalovic et al. [18]. An amplification reaction was performed in a volume of 50 µl containing 5 µl of 10 × PCR buffer, 10 pmol of each ERIC primer, 1.5 mM MgCl2, 0.2 mM of each deoxynucleoside triphosphate (dATP, dGTP, dCTP, and dTTP), 3 µl dimethylsulfoxide, 3 µl of chromosomal DNA solution, and 1 unit of Taq polymerase (Invitrogen; Thermo Fisher Scientific Corporation, Carlsbad, Canada). The DNA concentration range was about 20 to 50 ng of genomic DNA per microliter. Cycling conditions were as follows: denaturation at 95 °C for 3 min, followed by amplification for 35 cycles of 95 °C for 0.5 min, 51.2 °C for 1 min and 72 °C for 2 min, followed by a final extension step at 72 °C for 5 min. The PCRs were performed in a BIO-RAD T100 thermal cycler. Electrophoresis of the PCR products were carried out in 1% agarose gel. All gels were stained with ethidium bromide (1 µg/ml). Amplified DNA fragments of specific sizes were visualized with an ultraviolet (UV) transilluminator and photographed.

Furthermore, the genetic relatedness of the isolates obtained from 5 CRKP-BSI strains were determined by the pulse field gel electrophoresis (PFGE), and the bands showed on the PFGE underwent the clustering analysis via the BioNumerics software version 6.0 (Applied Maths NV, Sint-Martens-Latem, Belgium).

Definition
The clinical and microbiological outcomes were analyzed. BSI was defined according to the Centers for Disease Control and Prevention guidelines (available at: [http://www.cdc.gov/nhsn/pdfs/pscmanual/17pseudomonasdef_current.pdf]). BSI onset was defined as the collection date of isolate. The probable infectious source was determined on the basis of the microbiological results and the analysis by at least 2 physicians. Crude mortality was defined as death occurring after the collection of the first blood culture positive for K. pneumoniae. Mortality attributable to BSI was defined by clinical evidence of active infection and positive cultures, or when death occurred as the result of organ failure that developed or deteriorated during the onset of infection [19]. Appropriate treatment was defined as ≥48 h of treatment with at least one antimicrobial agent that has been proven in vitro activity against CR-KP strains [20]. Septic shock was defined as sepsis associated with organ dysfunction and persistent hypotension despite volume replacement [21].

Results
Increased percentage of CRKP, especially among ICU patients
Klebsiella pneumoniae (KPN) strains were collected and analyzed from January 1, 2012, to December 31, 2016 in our hospital. Among these strains, carbapenem resistant Klebsiella pneumoniae (CRKP) strains were identified via various cultures, coming to the result of the
percentage of CRKP isolates. During this 5-year period, 853 episodes of *Klebsiella pneumoniae* (KPN) in our hospital were obtained, and 18.8% (160/853) of these strains were CRKP isolates, while 36.8% (74/201) of the *K pneumoniae* strains identified from ICU were CRKP isolates. The annual rate of CRKP in clinical isolates in China, obtained from the CHINET surveillance of bacterial resistance [22, 23], has shown the steadily increased trend of the percentage of CRKP during 2012 to 2016 nationally. As Fig. 1 shown, the percentage of CRKP isolates in our hospital significantly increased from 3.4% in 2012 to 32.5% in 2016, with the highest rate obtained in 2015, up to 32.7%. Compared with the trend of the percentage of CRKP isolates in our hospital, the percentage of CRKP in ICU department initially declined from 8.3% in 2012 to 4% in 2013, then significantly increased from 4% in 2013 to 46.3% in 2016, with the highest rate reached 47.7% in 2015. In contrast to the above trends, the rate of CRKP rising from 10.8% in 2012 to 17.9% in 2016 was observed in data obtained from the CHINET surveillance program.

Owing to the deficiency of donors, the total number of kidney transplants was declined from 208 in 2012 to 149 in 2014 in our hospital. With the widely application of kidney transplantation from DCD, it’s rising from 149 in 2014 to 232 in 2016. As for the kidney transplantations from DCD, although kidney transplants from DCD was in absence in the first 2 years of the study, however, it was rising from 18 in 2014 to 170 in 2016. During the study period, there were only 5 CRKP-BSIs after renal transplantation from DCD, including 1 case (Case 1) in 2015 and 4 cases (Case 2–5) in 2016. Noticeably, both of the renal transplantations for Case 1 and Case 3 performed in the same foreign hospital. Therefore, apart from these renal transplantations that performed in foreign hospital, the incidence of CRKP-BSI after renal transplantation from DCD was about 1.1% (3/275) from January 1, 2012 to December 31, 2016 in our hospital.

**The application of immunosuppression regimen during the perioperative period**

The application of immunosuppressive regimen basically complied with a routine regimen, however varied case by case. According to the routine regimen, Mycophenolate Mofetil (MMF) was orally administered (1 g, tid) preoperatively, combined with the intravenous infusion of Basiliximab (20 mg), followed by the second administration of Basiliximab on the 4th day postoperatively, or the RATG (rabbit anti-thymic globulin) (1.25–2.5 mg/kg, qd) for 5 days. During the preoperative period, the glucocorticoid (1 g) was intravenously given, and glucocorticoid (0.5 g) was continually given for 3 consecutive days postoperatively. Tacrolimus, mycophenolate mofetil and glucocorticoid were taken orally for a long time for induction -or anti-rejection therapy. Based on the cases, the most frequently used immunosuppressive drugs were Tacrolimus, MMF and Basiliximab owing to the superiority of the therapeutic efficacy and relative drug safety [24–26]. Undoubtedly, the application of immunosuppressive drugs had altered the immunological status of the 5 patients, causing the subsequent occurrence of CRKP-BSI after KTx.

**The clinical characteristics of the 5 CRKP-BSIs**

The mean age of these 5 patients was 48.6 years (median: 47, range: 44–54 years), and 3/5 of the patients were male. The etiology of chronic renal failure (CRF) for the 5 CRKP-BSIs included systematic arterial hypertension (SAH), diabetes, glomerulonephritis (GMN) and IgA nephropathy, with the most common being SAH (see Additional file 1: Table S1). With respect to case 1 and 2, SAH was recognized as the etiology of CRF, and
SAH, GMN and IgA nephropathy were viewed as the etiology of CRF for case 3 and 4. In contrast to the above cases, SAH and Diabetes turned out to be the etiology of CRF for case 5.

Of the 4 cases (Case 1–3,5), the rupture of renal transplant artery was presented on the 40th, 16th, 43th and 74th day after KTx respectively. The thrombus of renal transplant artery was presented on the 43th and 13th day after KTx for Case 3 and Case 4. Additionally, on the 45th, 51th and 32th day after KTx, three cases (Case 1,2,5) occurring pneumothorax. Four cases (Case 1–4) received the excision of the transplanted kidney for the treatment. Detailed description of the clinical characteristics of the 5 CRKP-BSIs is shown (see Additional file 2: Table S2 and Additional file 3: Table S4).

Antibiotic susceptibility of 5 CRKP-BSIs following renal transplantation from DCD at the first bloodstream infection

In the whole picture, the antibiotic susceptibility of Case 2 distinguished from other cases in line with the genetic distinction (see Additional file 4: Table S3). With regards to Case 2, The MIC for Imipenem and Amikacin was $4_R$ and $\leq 2_R$, and it did not higher than $4_R$ for other antibiotics, except for Gentamicin ($\geq 16_R$). All the other cases showed similar resistance to antibiotics (MIC no lower than 2_R), except for Minocycline/Tigecycline. Based on the disc diffusion method, the result of antibiotic susceptibility to Minocycline/Tigecycline (Case1,3,5) showed 13_S, 18_S and 24_S, thereby suggesting the possibility of the utilization of the Tigecycline that has similar structure as Minocycline.

ERIC-PCR analysis of CRKP ICU isolates

To further determine the genetic relatedness of CRKP-BSI strains in ICU, we selected 11 CRKP ICU isolates for enterobacterial repetitive intergenic consensus (ERIC) PCR based genetic analysis. The obtained results indicated that these isolates belonged to two different clonal types (Fig. 2). The predominant clonal type was shown to be A type ($n = 9$) with the size about 150 kb. The other clonal type was B type ($n = 2$) with the size about 50 kb, belonging to the isolate from Case 2, indicating that the *Klebsiella pneumoniae* strain from Case 2 was different from the other strains. The result suggested that clone spread may contribute to the nosocomial dissemination of CRKP.

PFGE analysis of CRKP ICU isolates

To further determine the genetic relatedness of CRKP-BSI strains in ICU, we randomly selected 10 CRKP-BSI ICU isolates for PFGE analysis. Through the clustering analysis, the obtained results indicated that these isolates belong to three different clonal types, based on the 85% similarity (Fig. 3). The predominant clonal type was shown to be A type (6 isolates [60%], including A1-subtype [$n = 4$], A2-subtype [$n = 1$] and A3-subtype [$n = 1$]), coming from the Transplantation Center Urinary Surgery, Intensive Care Unit and Transplantation Center Intensive Care Unit, respectively. The B type (2 isolates [20%]) was obtained from Transplantation Center Urinary Surgery and Surgery Intensive Care Unit. The C type (2 isolates [20%]) was collected from Intensive Care Unit.

Discussion

*Klebsiella pneumoniae* (KP), with the characteristics of hypervirulent, highly transmissible and high mortality, has attracted a lot of attention in recent years, together with the worldwide spread of multidrug resistant KP strains. As the most lethal type of infection, *Klebsiella pneumoniae* bloodstream infection (KP-BSI), with the alarming high mortality in ICU, has been placed in the focus of various researches worldwide, showing the complex relationships with various factors, ranging from solid organ or stem cell transplantation, the frequent use of invasive devices,
surgery, hematological malignancy to the previous use of antibiotics. In China, the CHINET data showed that the rate of carbapenem resistance of *K. pneumoniae* isolates rose steadily from 10.8% in 2012 to 17.9% in 2016. The rate of CRKP has also increased dramatically worldwide over the past 10 years [27], and in the areas of high CRKP endemicity, such as United States, Israel, Italy, and Greece, the percentage of CRKP-BSI was shown to be 18–68% [19, 28–33]. Compared with these results, we observed a rapid increase in CRKP-BSI rates in our hospital after 2013, and the obtained data showed that these rates were higher than the average rates in China after 2014, especially among ICU patients, rising up to 47.7% in 2015, but declined to 46.3% in 2016 owing to the preventative measurements for the infection. In this research, the incidence of CRKP-BSI in the early stage (within 3 months) after renal transplantation from DCD was about 1.1% (3/275) in our hospital, approximating to the result of nationwide surveillance study of the incidence of carbapenem-resistant gram negatives (CRGN) in Italian transplant recipients performed by Lanini S et al., which comes to the Incidence rates were 1.23, 0.51 and 0.16 per 1000 recipient-days at 0–30, 31–60, and 61–90 days after solid organ transplantation (SOT), respectively [31].

**Fig. 3** PFGE analysis of various CRKP-BSI isolates. Genomic DNA was digested using Xba I enzyme and subjected to PFGE. The bands were further analyzed by BioNumerick software version 6.0. Abbreviation: ICU Intensive care unit, SICU Surgery Intensive Care Unit, TC Transplantation Center, TC-ICU Transplantation Center Intensive Care Unit, TC-US Transplantation Center Urinary Surgery.
Just as Fig. 4 shown, considering the renal transplantation performed in the foreign hospital and the confirmed infection of CRKP following renal transplantation, there is a possibility that Case 1 plays the role as the exogenous source, associated with the following series of CRKP-BSIs in our hospital in the next year. The result of ERIC-PCR has substantiated the supposition that there is certain level of nosocomial clonal spread, disseminating the pathogen from the exogenous resources, and highlighting the necessity of implementation of thorough surveillance of recipients of renal transplantation before transferring from foreign hospital.

Clinical characteristics associated with CRKP-BSI patients after renal transplantation in our study has been attributed to the various aspects. All of the patients had received the renal transplant from donation after cardiac death (DCD). Undoubtedly, compared with the living donor, as the supplement mean for transplantation, the renal transplants from DCD has satisfied the increasing demanding of the resource of kidney nowadays and accelerated the advancement of technology and conception of renal transplants. However, there is controversy with respect to the infection risk of renal transplantation from DCD worldwide. Sousa SR et al. had come to conclusion that renal transplants from decreased donors and a prolonged cold ischemia time were important risk factors for infections following renal transplantation [32]. Fernández-Ruiz M et al. performed a prospective cohort study of 291 KT patients, drawing the conclusion that uncontrolled DCD policies were safe in terms of the risk of post-transplant infection in spite of higher rate of delayed graft function (DGF) and the use of ATG-containing sequential therapy [33]. In our study, we had observed that in the early stage after renal transplantation (3 months), the CRKP induced rupture/thrombus of the artery of the transplanted kidney occurred. During

Fig. 4 a The total number of kidney transplants and the number of donation after cardiac death (DCD) in the hospital from 2012 to 2016. b The number of isolates of CRKP-BSI after the kidney transplants in the hospital from 2012 to 2016.
the course of treatment, the predominant features of all the cases included the placement of catheters, the use of antibiotics, renal transplantation, prolonged stay of ICU department, and the high dose immunosuppressive therapy, predisposing to the vulnerability of CRKP-BSI. Additionally, the most common site of CRKP isolation, in our study, was organ-space surgical site, followed by urinary tract and pneumoniae. In a study of LTx recipients, in the first month after transplantation, 71% of the KPC-Kp-HAIs had abdominal infections [34]. In other studies of KTx recipients, the most common foci of infection have been found to be the surgical wound and the urinary tract [35]. These studies suggest that various site/type of infection, for instance abdominal effusion, surgical wound, urinary tract and so on, can be the susceptible sites of CRKP-BSIs as the manifestation of the systemic infection on specific site, serving as part of the systemic infection. The peri-renal infection can lead to the rupture or thrombus of the renal transplant artery, with the consequence of the excision of transplanted kidney, and be reckoned as the severe complication after kidney transplantation. In our research, all of the five cases have close relationship with CRKP, with one case (Case 2) associated with the mixed infection of CRKP and aspergillus that confirmed by the pathologic report (Fig. 5), therefore administrating different antibiotic regimen with the wide coverage for aspergillus. Xu MJ et al. reported 5 KTRs occurring the infectious rupture of renal transplant artery, with 1 KTR infected with CRKP, while other KTRs infected with fungus [36]. Wan Q et al. reported 3 of 154 KTRs developed confirmed donor-derived CRKP infections, with 1 CRKP-BSI and 1 CRKP died due to the rupture of renal transplant artery, coming to the conclusion that Klebsiella pneumoniae was the most probable causative microorganism leading to the episode of renal artery rupture [4].

When it comes to the treatment for CRKP-BSI after renal transplantation, a multidisciplinary therapy, combined with the surgical therapy and the combinational antibiotic therapy, especially regimens including a carbapenem, has been proven beneficial for the patient, demonstrated in several retrospective studies of general population [37, 38]. From the cases, we learn that it is critical to obtain the positive result of culture in the early stage of treatment, guiding the following definitive antibiotic therapy. Usually, a combined treatment (Tigecycline + Carbapenem, Tigecycline + Fosfomycin, Fosfomycin + Aminoglycoside, Third generation cephalosporin +β-lactamases inhibitors etc.) can be prescribed for the treatment of CRKP-BSI. If permitting, high dose Tigecycline (100 mg, q12h after 200 mg loading dose) scheme is worth recommending to bring a satisfactory outcome for the patient [39], and in Case 1 and 5, the respective scheme had been applied. Based on the differential result of antibiotic susceptibility and the evidence of growth of aspergillus on the transplanted kidney, the renal transplant artery and external iliac artery, plus with the infection of Candida, the antibiotic
regimen, consisted of Meropenem, Tigecycline, Amikacin, Voriconazole and Amphotericin B, was prescribed for Case 2. Important of all, utilization of properly minimvasive surgical approaches, for example the indwelling catheter drainage for the abdominal fluids, can alleviate the severity of the disease to a large extent and extend the survival, combined with the appropriate combined antibiotic therapy. Last but not least, the cooperation between multiple medical departments is also vital for the treatment, not only demonstrating the necessary coordination between departments for the therapy, but also enhancing the efficacy of treatment and avoiding the incompleteness and shortage of medical knowledge and techniques for one department.

In United states and Europe, there are myriad of clinical reports regarding the application of Ceftazidime, Avibactam and Polymyxin B, obtaining surprising and satisfactory therapeutic effect. Nevertheless, Polymyxin B came into Chinese market in December 2017, while Avibactam had not appeared on the Chinese market yet. In the absence of internationally newly developed drugs, the antibiogram for treating CRKP fundamentally was based on the combined treatment, consisted of Carbapenem/Aminoglycoside/Fosfomycin and Tigecycline, until now in China. In the near future, in light of pharmacokinetic/pharmacodynamic (PK/PD) profile of the CRKP-BSIs after KTx, it is worth adopting the multi-drugs combinational antibiotic strategy to elevate the cure rate for the KTRs infected with CRKP-BSI, taking advantage of the beneficial therapeutic approach including the high-dose prolonged-infusion (3–4 h) regimen to drive the PK/PD to acceptable exposures [40].

Our study has several limitations. Clinical data were obtained retrospectively from medical records, and therefore, there may exist some differences in physician practices or accuracy of information. Additionally, the data for patients who may have had significant BSI symptoms such as septic shock and hyperpyrexia, but were not tested owing to the patient refusal or because their blood culture was negative, were not included. Finally, this was a single-center study, showing all the 5 recorded CRKP-BSIs with detailed clinical analysis. A large-scale of study on the clinical characteristics of CRKP-BSIs after renal transplantation from donation after cardiac death is underway to provide more evidence on the clinical significance of the hard-handled multi-resistant superbug. Most important of all, the investigation of transmission of bacterial or fungal infection to kidney transplant recipients from DCD has also been on the way to clarify the etiology of CRKP-BSI after renal transplantation and help designate an effective strategy to eliminate the possibility of bacterial infection before transplantation for the surgeons and physicians in the ICU and transplant department.

As the recipients of renal transplantation from donation after cardiac death, many factors can interact modifying the risk for infection, such as the immunosuppression profile used, the post-operative care, and the profile of epidemiological exposure to the susceptibility to CRKP [32]. It is critical to perform thorough surveillance of the source of kidney before transplantation, the protection of integrity of skin and mucosal barriers in the critically ill patients or the patients undergoing invasive procedures, and all necessary contact precautions should be employed by hospital staff caring these high-risk patients [41]. In conclusion, the thorough surveillance of sources of kidney before transplantation, early clinical identification of the infection of carbapenem resistant klebsiella pneumonia, the necessary preventative measurements and the use of the appropriate treatment are the reasonable strategy to treat the CRKP-BSI in the early stage after renal transplantation from DCD.

Conclusion
An increase in the occurrence of CRKP-BSIs among KTRs from DCD in our hospital was observed in our hospital during January 1, 2012 to December 31, 2016. Rupture/thrombus of the renal transplant artery was demonstrated to be associated with CRKP-BSI in the early stage after KTx from DCD. Despite the low incidence of CRKP-BSI (1.1%) after KTx from DCD, the high mortality (4/5) had been observed.

Additional files

Additional file 1: Table S1. Demographic characteristics and utilization of immunosuppression drugs of patients following renal transplantation. (XLSX 10 kb)

Additional file 2: Table S2. Summary of carbapenem-resistant Klebsiella pneumoniae (CR-KP) isolation site, episodes of sepsis, antibiotic treatments adopted, outcome, excision of transplanted kidney and cause of death. (XLSX 9 kb)

Additional file 3: Table S3. Analysis of clinical characteristics of 5 kidney recipients infected with Klebsiella pneumoniae carbapenem resistant K. pneumoniae. (XLSX 12 kb)

Additional file 4: Table S4. Antibiotic susceptibility of 5 stains of carbapenem-resistant K. pneumoniae isolated from first bloodstream infections in kidney transplant recipients. (XLSX 9 kb)

Abbreviation
A: Abdominal cavity; AMB: Amphotericin B; AMK: Amikacin; ATP: Adenosine triphosphate; B: Blood; CCI: Charlson comorbidity index; CHINET: China bacterial resistance monitoring network; CIT: Cold ischemia time; CPS: Capsular polysaccharide; CR: Chronic renal failure; CRGN: carbapenem-resistant gram negatives; CR-KP: carbapenem-resistant Klebsiella pneumoniae; CRKP-BSI: Carbapenem-resistant Klebsiella pneumoniae bloodstream infection; CRRT: Continuous renal replacement therapy; CTP: Cytidine triphosphate; DCD: Donation after cardiac death; DGF: Delayed graft function; DNA: Deoxynucleobase acid; DTI: Donor-transmitted infection; E: External iliac artery; ERIC-PCR: Enterobacterial repetitive intergeneric consensus polymerase chain reaction; G: Gallbladder; GMN: Glomerulonephritis; GTP: Guanosine triphosphate; I: Incision; ICU: Intensive care unit; IPC: Infection prevention and control policy; KP: Klebsiella pneumoniae; KP-BSI: Klebsiella pneumoniae;
bloodstream infection; KPC-Kp-HAIs: Klebsiella pneumoniae carbapenemase-producing Klebsiella pneumoniae-Hospital acquired infections; KPN: Klebsiella pneumoniae; KTR: Kidney transplantation recipient; KTx: Kidney transplantation; L: Lung; LOS: length of stay; LTx: Liver transplantation; MEM: Meropenem; MIC: Minimum inhibitory concentration; MCEF: Moxycycline plus moxifloxacin; NA: Non-applicable; NIT: Nitrite; PCR: Polymerase chain reaction; PLAs: Chinese people of liberation of army; SAH: Systemic arterial hypertension; SICU: Surgery intensive care unit; SOFA: Sequential organ failure assessment; SOT: Solid organ transplantation; T: Thoracic cavity; TC: Transplantation center; TC-ICU: Transplantation center intensive care unit; TC-US: Transplantation center urinary surgery; TGC: Tigecycline; Ti: Tip of catheter; TTP: Thymidine triphosphate; U: Urinary tract; UV: Ultraviolet; VOR: Voriconazole

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Availability of data and materials

The raw data can be made available to the interested researchers by the authors of this article if requested.

Authors’ contributions

YZ designed and supervised the study, YW collected and analyzed the medical records, and wrote the manuscript, while YZ monitored the study. YW, JW, CX, JY, LZ and XK provided the clinical data retrieved from patients at first hand. HL collected the isolates, performed the drug susceptibility test and wrote the method for the procedure. All of the authors read and approve the final manuscript and have been met the requirements for authorship.

Ethics approval and consent to participate

Ethical approval was obtained from the Ethics committee board of 309th Hospital of Liberation of Army (PLA), No. 17 Heishanhu Road, Haidian District, Beijing, People of Liberation of Army (PLA), for his assistance of providing the collection of the raw data and materials, and the need for individual patient consent was waived (reference number: 30920180310912).

Competing interests

The authors declare that they have no competing interests.

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