Bone marrow infiltrated natural killer cells predicted the anti-leukemia activity of MCL1 or BCL2 inhibitors in acute myeloid leukemia

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Abstract
Acute myeloid leukemia (AML) is still incurable due to its heterogeneity and complexity of tumor microenvironment. It is imperative therefore to understand the molecular pathogenesis of AML and identify leukemia-associated biomarkers to formulate effective treatment strategies. Here, we systematically analyzed the clinical characters and natural killer (NK) cells portion in seventy newly-diagnosis (ND) AML patients. We found that the proportion of NK cells in the bone marrow of ND-AML patients could predict the prognosis of patients by analyzing the types and expression abundance of NK related ligands in tumor cells. Furthermore, MCL1 inhibitor but not BCL2 inhibitor combined with NK cell-based immunotherapy could effectively improve the therapeutic efficiency via inhibiting proliferation and inducing apoptosis of AML primary cells as well as cell lines in vitro. There results provide valuable insights that could help for exploring new therapeutic strategies for leukemia treatment.

Keywords: NK cells, MCL1 inhibitor, BCL2 inhibitor, AML, Immunotherapy

Main text
Natural killer (NK) cells are a type of cytotoxic immune cells that can recognize and kill cancer cells rapidly and efficiently. In recent years, more and more studies have revealed the biological characteristics of NK cells and their ability to recognize cancer cells directly [1]. Although immunotherapy has made a great breakthrough in stimulating the immune system against hematologic malignancies, there are few studies on NK cell-based immunotherapy [2].

Killer cell immunoglobulin-like receptors (KIR) are a type of receptor mainly expressed on the surface of human NK cells and partially activated T cells [3]. Functionally, KIR genes could be divided into inhibitory and activated types, which can specifically recognize and bind HLA class I molecules on the surface of target cells. They regulate the killing function of active cells in an effective switch system and play an important role in anti-infection and anti-tumor [4].

In this study, we elucidated the clinical relevance of KIRs and NK cells in bone marrow (BM) of acute myeloid leukemia (AML) patients. Our data indicated that NK cell ratio can predict the prognosis of patients, and can synergistically kill leukemia cells with MCL1 inhibitors to improve treatment efficiency.
Fig. 1 (See legend on next page.)
Results and discussions

Expression level and prognosis of KIRs in AML

Here we reported a study on the transcriptional levels of KIRs in cancer and normal samples by analyzing the data from Oncomine and ENCORI (The Encyclopedia of RNA Interactomes) (Additional file 1 and Table S1). These results indicated that KIRs played an important role in solid tumors such as kidney renal clear cell carcinoma and lung cancer. In AML patients, the expression levels of KIRs were much higher than that in normal samples (Additional file 2). Notably, the expression of KIR2DL group (KIR2DL1, KIR2DL3 and KIR2DL4) was significantly downregulated in patients with FLT3 mutations, whereas KIR2DS and KIR3DL group (except KIR3DL3) were upregulated (Fig. 1a). Interestingly, RAS activation status was not related to the expression of KIRDS in AML (Additional file 3). Next, we further explored the critical efficiency of KIRs for predicting the survival of patients with AML. The Kaplan-Meier curve and log rank test analyses revealed that the increased KIR2DL1 ($p = 0.0043$), KIR2DL3 ($p = 0.0028$), KIR2DL4 ($p = 0.0092$), KIR3DL1 ($p = 0.013$) and KIR3DL2 ($p = 0.0088$) mRNA levels were significantly related to poor prognosis for overall survival (OS) of AML patients (Fig. 1b). Whereas, the KIR2DS4 mRNA level had no tendency to indicate prognosis ($p = 0.33$). Furthermore, when FLT3 mutation status was combined, the prognostic values of the KIRs factors were consistent with the above results (Fig. 1c).

NK cells in BM indicated poor prognosis in AML

The immune dysfunction recently has been considered as a risk factor in AML and predicted poor prognosis [5]. Next, we utilized Cibersort to deconvolute the gene-expression data of 713 newly diagnosed AML (ND-AML) patients (140 patients as Training cohort and 573 patients as Validation cohort) and generated a gene matrix with a signature of more than 10 immune cell subtypes (Table S2 and S3). Patients were divided into “low” and “high” subgroups (according to the cutoff value of conversion score of total NK cells, activated NK cells and resting NK cells, respectively). The prognostic analysis indicated that the low NK cells or low resting NK cells predict poor prognosis, while the low activated NK cells indicated a favorable prognosis in AML patients (Fig. 1d and Additional file 4). Notably, the differential expression of KIRs were only in total NK cells but not in activated or resting NK cells (Fig. 1e). Thus, we proposed a model to describe the cellular and molecular basis for the potential prognostic value of monitoring the proportion of immune cells in patients (Fig. 1f).

Further, the single-cell RNA sequence data of AML patients at diagnosis and matched samples after chemotherapy were used for immune cell subtype analysis [6]. UMAP (Uniform Manifold Approximation and Projection) analysis indicated the proportion of NK cells in total BM cells of AML samples at diagnosis was much lower than those in matched samples after chemotherapy (Fig. 2a). We further validated the NK cells proportion (CD45$^+$CD3$^+$CD56$^+$CD16$^+$) of lymphocytes in BM cells from patients with hematological malignancies (30 lymphoma cases without BM infiltration as control; 95 ND-AML cases and 25 refractory/relapse (R/R) AML cases) (Fig. 2b). The proportion of lymphocytes, especially NK cells in BM, was significantly decreased in ND-AML samples compared with normal samples, and the ratio reached the lowest in R/R AML cases (Fig. 2c). These data were consistent with theory that NK cells might be one of important mediators of anti-leukemia immunity and indicated that NK cells of BM might play an anti-leukemia effect in leukemogenesis [7, 8]. In addition, KIRs expressions were much higher in R/R AML samples among these three groups (Fig. 2d and Table S4). When we divided ND-AML samples into high NK cells group and low NK cells group, according to the proportion of NK cells in BM. We found there was no significant differences in the clinical characteristics of the two groups (Table 1). However, KIRs were mainly expressed in samples with low NK cell group (Fig. 2e and Table S4). The expression data further validated survival data of NK cells and KIRs expression patterns which showed above.

NK cells could synergy with MCL1 inhibitor to kill leukemia cells

Previous studies showed that BCL-2 and MCL-1 proteins could control the survival of NK cells in vivo [9]. Thus, we explored the relationship between the expression of these two molecules and NK cells in BM. We found MCL1 expression level has a significant negative
Fig. 2 NK cells enhanced the anti-leukemia activity synergized with MCL1 inhibitor. a UMAP analysis of NK cells in AML samples at diagnosis and after chemotherapy by using single-cell RNA sequence data. The percentage of NK cells among them were further compared. b The flow cytometry analysis of BM infiltrated NK cells by CD45+CD3− CD56+CD16+ gating. c Proportions of lymphocytes and NK cells subsets in BM of patients with lymphoma, ND-AML and R/R AML. d The expression levels of KIRs in BM of patients with lymphoma, ND-AML and R/R AML. e Comparison of KIRs expression levels between high NK group and low NK group in ND-AML patients. f The cell viability test of OCI-AML3 and MOLM13 treated with venetoclax or maritoclax by co-cultured with UCB-NK cells. g Protein validation of the knockdown efficiency of MCL1 siRNAs in OCI-AML3 and MOLM13 cells. h The cell viability of OCI-AML3 and MOLM13 treated with MCL1 siRNA or scramble siRNA (h) and co-cultured with UCB-NK cells (i). j RNA and protein expression levels of BCL2 and MCL1 in normal controls and ND-AML samples with low or high NK cells. k Scheme of cell viability analysis by using primary samples of AML patients. l Cell viability of BM cells with high or low NK cells with venetoclax or maritoclax treatment. m-n Apoptosis induced by inhibitors among AML patient cells with high or low NK cells. Mean ± SEM values are shown. *P < 0.05, **P < 0.01, ***P < 0.001
Table 1 Basic characteristics of ND-AML patients between NK% high and NK% low in BM

| Variable                        | ND-AML NK%<sup>high</sup> (n=34) | ND-AML NK%<sup>low</sup> (n=35) | P value |
|--------------------------------|----------------------------------|---------------------------------|---------|
| Age, years                     | 47                               | 19–74                           | ns      |
| Median                         |                                  |                                 |         |
| Range                          | 15–78                            | 19–74                           |         |
| Gender, no. (%)                |                                  |                                 | ns      |
| Male                           | 20 (58.8%)                       | 16 (45.7%)                      |         |
| Female                         | 14 (41.2%)                       | 19 (54.3%)                      |         |
| Hepatic or renal function, no. |                                  |                                 | ns      |
| Normal                         | 32                               | 32                              |         |
| Abnormal                       | 2                                | 3                               |         |
| WBC, 10<sup>9</sup>/L          |                                  |                                 | ns      |
| Median                         | 10.665                           | 13.79                           |         |
| Range                          | 1–249.98                         | 1.4–371.9                       |         |
| Platelet, 10<sup>9</sup>/L      |                                  |                                 | ns      |
| Median                         | 34.5                             | 28                              |         |
| Range                          | 1.87–401                         | 3–167                           | 56      |
| Hb, g/L                        |                                  |                                 | ns      |
| Median                         | 81                               | 74                              |         |
| Range                          | 20–116                           | 52–149                          |         |
| PB NK, %                       |                                  |                                 | <0.001  |
| Median                         | 14.1                             | 5.4                             |         |
| Range                          | 4.2–42.9                         | 2.3–21.7                        |         |
| BM blasts, %                   |                                  |                                 | ns      |
| Median                         | 53.7                             | 53.1                            |         |
| Range                          | 6.6–94.1                         | 16.3–95.1                       |         |
| BM Lymph, %                    |                                  |                                 | ns      |
| Median                         | 11.3                             | 6.2                             |         |
| Range                          | 2.2–49                           | 1.7–49.2                        |         |
| BM NK, %                       |                                  |                                 | <0.001  |
| Median                         | 19.65                            | 7.6                             |         |
| Range                          | 12.5–43.6                        | 2–11.9                          |         |
| BM T, %                        |                                  |                                 | <0.001  |
| Median                         | 66.5                             | 77.7                            |         |
| Range                          | 45.6–80.4                        | 46–86.2                         |         |
| Cytogenetics, no. (%)          |                                  |                                 |         |
| t(15;17)/PML-RARA              | 0                                | 0                               |         |
| t(8;21)/AML1-ETO               | 1                                | 4                               |         |
| inv(16;16)/CBFb-MYH11          | 2                                | 1                               |         |
| CN-AML<sup>b</sup>             | 23                               | 22                              |         |
| Unfavorable<sup>c</sup>        | 8                                | 8                               |         |
| Mutations, no. (%)             |                                  |                                 |         |
| FLT3 mutations                 | 15                               | 5                               |         |
| RAS mutations                  | 6                                | 6                               |         |

ALT Alanine aminotransferase, AST Aspartate aminotransferase, WBC White blood cell, RBC Red blood cell, Hb Hemoglobin, BM Bone marrow, ns Not significance

P values were calculated by means of nonparametric Wilcoxon rank-sum test for continuous variables and x-square test for categorical variables

<sup>a</sup>Hepatic abnormality as defined by ALT 2.5× normal value or AST 2.5× normal value, while renal abnormality as defined by creatinine 2.5× normal value

<sup>b</sup>CN-AML: cases having no cytogenetically identifiable abnormalities

<sup>c</sup>Unfavorable: inv. (3)/t (3;3), t (9;22), 11q23 abnormalities, 25, 27, del (5q),del (7p), and complex karyotype
correlation with NK cells, while BCL2 expression level
has no correlation with NK cells both in training and
validation cohorts (Additional file 5). As we known,
BCL2 and MCL1 inhibitors are currently considered as
new therapeutics for leukemia, and showed promising
response in patients [10]. Therefore, we quest whether
NK cells could synergize with these new inhibitors to
achieve better therapeutic effects. To test our hypothesis,
we treated AML cell lines OCI-AML3 and MOLM13
with BCL2 inhibitor (Venetoclax) or MCL1 inhibitor
(Maritoclax), then co-cultured with gradient numbers of
NK cells derived from umbilical cord blood (UCB-NK).
The cell viability suggested that UCB-NK could cooper-
ate with maritoclax but not venetoclax to kill leukemia
cells (Fig. 2f). Moreover, we applied the MCL1 siRNA to
to knockdown the expression of MCL1 in OCI-AML3 and
MOLM13 cells and then cocultured with UCB-NK
(Fig. 2g-i). The cell viability was significantly decreased
in both OCI-AML3 and MOLM13 cells treated with
MCL1 siRNA compared with those treated with scramble siRNA (Fig. 2h). In addition, the cell viability of OCI-
AML3 and MOLM13 was significantly impaired after
treatment with MCL1 siRNA and UCB-NK cells (Fig. 2i).
This result was consistent with the data on the pharma-
cological effects of maritoclax (Fig. 2f). Next, we sought
to validate effects by using mononuclear cells isolated
from the BM of ND-AML patients. Samples were di-
vided into two groups according to the NK proportion
in BM (Fig. 2k and Table S4), and the RNA/protein ex-
pression levels of BCL2 and MCL1 were examined.
MCL1 was much more expressed in the low NK cell
group of ND-AML samples both at transcriptional and
translational level (Fig. 2j). Consistent with the results
from cell line studies, patient samples with low NK
treated with maritoclax were less viable than samples
with high NK (Fig. 2k). In addition, maritoclax induced
 cell apoptosis was significant enhanced in samples with
low NK proportion (Fig. 2m and n). While there was no
significant difference in cell viability and apoptosis be-
tween samples with high NK and low NK with veneto-
class treatment (Fig. 2k-n).

Conclusions
The importance of NK ratio in BM of AML patients has
generated tremendous interest in understanding its role
in disease management. Immune dysfunction could pre-
dict therapeutic reactivity and unfavorable prognosis [5].
By analyzing the types and expression abundance of li-
gands related to NK function expressed in tumor cells,
we first proposed that the proportion of NK cells in the
BM of AML patients could predict the prognosis of pa-
ients. The lower the proportion of NK cells, the worse
of the prognosis. Therefore, increasing the number of
NK cells through small molecule compounds or
cytokines, such as interleukin 15 or interleukin 2 etc.,
may be a new method of anti-leukemia. Next, we plan to
perform experiments to verify the effect of NK on pro-
gnosis in different conditions, and conduct prospective
controlled cohort clinical trials to verify this hypothesis
in the future. Second, our data indicated that NK cell-
based immunotherapy combined with MCL1 inhibitor
but not BCL2 inhibitor could effectively improve the
therapeutic efficiency of AML. These findings might
provide new insights and theoretical basis for exploring
new targets for leukemia treatment.

Supplementary Information
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Abbreviations
NK: Natural killer; KIR: Killer cell immunoglobulin-like receptors; HLA: Human
leukocyte antigen; ENCORI: The Encyclopedia of RNA Interactomes;
AML: Acute Myeloid Leukemia; ND-AML: Newly diagnosed AML;
UMAP: Uniform Manifold Approximation and Projection; R/R AML: Refractory/
relapse AML; UCB-NK: NK cells derived from umbilical cord blood

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Authors’ contributions
Contribution: Y.J.D. designed and performed all cell culture experiments,
interpreted data, created a graphical summary, and wrote the paper; F.H. and
S.L.C performed bioinformatics analyses; S.Y.H performed single-cell sequen-
cing analysis interpreted data; J.M.Z wrote the paper; F.H. and
H.M.S. assisted cell culture experiments and edited the paper; D.W.W. con-
ceived and oversaw the study, interpreted data, and wrote the paper. The
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Availability of data and materials
All the data obtained and/or analyzed in the current study were available
from the corresponding authors on reasonable request.
Ethics approval and consent to participate
This study was approved by Ethics Committee of Sun Yat-sen University Cancer Center and Ruijin Hospital affiliated to Shanghai Jiao Tong University School of Medicine.

Consent for publication
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Competing interests
The authors declare no competing financial interests.

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