Apolipoprotein A-I and Cancer

Maryam Zamanian-Daryoush* and Joseph A. DiDonato*

Department of Cellular and Molecular Medicine, and Center for Cardiovascular Diagnostics and Prevention, Cleveland Clinic, Cleveland, OH, USA

High-density lipoprotein (HDL) and apolipoprotein A-I (apoA-I), the predominant protein in plasma HDL, have long been the focus of intense studies in the field of atherosclerosis and cardiovascular disease. ApoA-I, in large part, is responsible for HDL assembly and its main atheroprotective function, that of shuttling excess cholesterol from peripheral tissues to the liver for excretion (reverse cholesterol transport). Recently, a protective role for HDL in cancer was suggested from several large clinical studies where an inverse relationship between plasma HDL-cholesterol (HDL-C) levels and risk of developing cancer was noted. This notion has now been tested and found to be supported in mouse tumor studies, where increasing levels of apoA-I/HDL were discovered to protect against tumor development and provision of human apoA-I was therapeutic against established tumors. This mini-review discusses the emerging role of apoA-I in tumor biology and its potential as cancer therapeutic.

Keywords: macrophage, apoA-I, HDL, cancer, cholesterol homeostasis, peptide mimetics, cancer therapeutic

INTRODUCTION

Apolipoprotein A-I (ApoA-I), the major protein component of high-density lipoprotein (HDL), is synthesized predominantly in the liver and the small intestine, and exists transiently in lipid-poor form. ApoA-I initiates assembly of HDL particles through interaction with ATP-binding cassette transporters at the surface of cells in the periphery extracting cholesterol and phospholipids. The HDL particle is further matured by lecithin cholesterol acyltransferase (LCAT) binding to apoA-I on HDL and converting cholesterol to cholesteryl ester (Sorci-Thomas et al., 2009). Cholesterol is transported within the constantly evolving ‘HDL cargo’ in circulation for excretion in liver through scavenger receptor class B, type 1 (SR-B1), a process known as reverse cholesterol transport (RCT; reviewed in Fisher et al. (2012), Rosenson et al. (2012), and Rader and Hovingh (2014)). Cholesterol accumulation and the ensuing inflammation that develops within the arterial wall are major instigators of atherosclerosis and cardiovascular disease (CVD; Hansson and Libby, 2006; Libby et al., 2009), and RCT is considered the primary atheroprotective function of HDL. A number of non-RCT atheroprotective functions of apoA-I/HDL have been described and include anti-inflammatory (Cockerill et al., 1995; Bhattacharyya et al., 2008; Rye et al., 2009; de Souza et al., 2010; Fuhrman et al., 2010; Camont et al., 2011; Gordon et al., 2011), anti-apoptotic (de Souza et al., 2010; Fuhrman et al., 2010), and anti-oxidant activities (Bhattacharyya et al., 2008). Chronic inflammation, oxidative stress, lipids, and cholesterol, which promote atherosclerosis and CVD, have all been associated with tumorigenesis (Lu et al., 2006; Freed-Pastor et al., 2012; Coussens et al., 2013; Noguti et al., 2013; Kobayashi et al., 2015). Given HDL's beneficial role against these atherogenic processes, it was somewhat intuitive to suggest that HDL may also be protective against cancer.
ApoA-I/HDL IS ANTI-TUMORIGENIC

Clinical Observational Studies Suggest a Protective Role for ApoA-I in Cancer

Analysis of Finnish male smokers in the Alpha-Tocopherol Beta-carotene (ATBC) lung cancer prevention study identified a significant inverse association between HDL-C levels and the risk for cancer of the lung, prostate, liver, and hematopoietic system (Ahn et al., 2009). This observation was further strengthened by a large meta-analysis of randomized controlled trials of lipid-altering therapies suggesting an inverse relationship between plasma HDL cholesterol levels and the incidence of cancer development during the conduct of the trials (Jafri et al., 2010). Specifically, for every 10 mg/dL increase in plasma HDL cholesterol level, a significant 36% lower risk of cancer incidence was noted over 625,000 person-years of follow-up and > 8,000 incidents of cancers cumulatively among the trials included in the meta-analysis (Jafri et al., 2010). Consistent with this finding, apoA-I has been identified as a biomarker with reduced plasma levels in patients with early stage ovarian cancer (OC) compared with normal individuals (Kozak et al., 2003, 2005; Zhang et al., 2004; Moore et al., 2006; Clarke et al., 2011; Kim et al., 2012). Furthermore, higher apoA1 mRNA levels in pre-chemotherapy effusions from patients diagnosed with advanced stage OC was observed to be an independent prognostic marker of longer overall survival (Tuft Stavnes et al., 2014). ApoA-I together with transferrin and transferrin, both HDL-associated proteins, β2-microglobulin, and prealbumin along with CA125 are currently used in a US Food and drug administration approved plasma test for OC, and these combined biomarkers test is known as OVA1®. This test is used to identify patients with early stage OC (Kozak et al., 2005; Su et al., 2007; Nossov et al., 2008; Nosov et al., 2009). A correlation with apoA-I levels and risk of disease has also been observed in other cancers. Serum apoA-I levels were found to be twofold lower in patients undergoing surgery for pancreatic cancer compared with healthy controls (Ehmann et al., 2007). Lower serum apoA1 levels were also associated with higher risk of breast cancer (BC; Chang et al., 2007) as well as BC recurrence (Lane et al., 1995), although other studies examining BC showed either no correlation with HDL-C (Furberg et al., 2004) or an inverse association observed only in pre-menopausal women, but not in post-menopausal women (Moorman et al., 1998; Kucharska-Newton et al., 2008). In contrast, a recent study reported a positive association between serum HDL-C and apoA-I levels with BC risk; however, this direct association was only seen in women who had not used hormone replacement therapy (HRT; Martin et al., 2015). In metastatic nasopharyngeal carcinoma (NPC), higher levels of serum apoA-I measured prior to chemotherapy correlated with better overall survival (Jiang et al., 2014). Additionally, in a large European prospective study, the European Prospective Investigation into Cancer and Nutrition (EPIC) study, an inverse association was found between HDL-C levels and endometrial cancer risk (Cust et al., 2007), and similarly plasma concentrations of HDL-C and apoA-I were found to be inversely associated with the risk of colon cancer (van Duijnhoven et al., 2011). Table 1 lists clinical studies which established an association between plasma levels of HDL-C/apoA-I and risk of developing a broad spectrum of cancers.

ApoA-I is Protective in Mouse Tumor Models

Although the above-described clinical studies suggested, for the most part, lower apoA-I plasma levels correlate with poorer patient outcomes in a variety of cancers, they fall short of inferring causality. The first evidence supporting the notion that increasing levels of apoA-I may be protective against tumor development came from mouse tumor studies. Accordingly, there was an apoA-I gene dosage effect on Lewis lung tumor growth, with tumor size smallest for mice homozygous for the human apoA1 transgene (hA1 Tg+/+), intermediate tumor size in mice heterozygous for the human apoA1 transgene (hA1 Tg+/-), referred to hereon as simply apoA1 transgenic (A-I Tg), and largest tumor size in apoA1 null mice (A-I KO; Zamanian-Daryoush et al., 2013). Furthermore, overall tumor burden and metastasis was strongly suppressed in response to challenge with the aggressive and metastatic syngeneic melanoma cells (B16F10L) in animals expressing the apoA1 transgene relative to apoA1 null or wild-type (WT) animals, culminating in improved overall survival (Zamanian-Daryoush et al., 2013). Similarly, in a mouse model of OC (ID8), ApoA-I transgene expression reduced tumor burden and led to a significant increase in survival (Su et al., 2010).

Infusion of Human ApoA-I Inhibits Tumor Development and is Therapeutic against Established Mouse Tumors

Unequivocal support of an anti-neoplastic role for apoA-I came from animal studies where subcutaneous injections of human apoA-I prior to tumor inoculation prevented tumor development in A-I KO mice (Zamanian-Daryoush et al., 2013). More importantly, in a physiologically relevant setting, provision of apoA-I after tumor establishment not only prevented further development of the tumor but also led to tumor shrinkage (Zamanian-Daryoush et al., 2013). Although these protective effects were seen with the syngeneic mouse melanoma tumor, B16F10L, apoA-I therapy was also shown to be effective against human melanoma (A375) in nude mice (Zamanian-Daryoush et al., 2013). Based on these observations a potential role for apoA-I/HDL as an anti-cancer therapeutic has been proposed (Zamanian-Daryoush et al., 2013).

ApoA-I/HDL MIMETIC PEPTIDES EXHIBIT ANTI-TUMOR ACTIVITY

There has been considerable interest, in the HDL field, surrounding the therapeutic potential of a series of 18 amino acid peptides known as apoA-I/HDL mimetics, against cardiovascular disease. These short peptides have certain functional properties of the repeating amphipathic α-helices of apoA-I without actually sharing any sequence homology (Getz and Reardon, 2014; Reddy et al., 2014). These peptides were originally screened...
Breast Multiple time point measurements of serum lipids and lipoproteins in a colon/prostate broad variety

Analysis of 24 randomized controlled trials of lipid-altering therapy (145,743 persons) to correlate baseline and on-treatment HDL-C levels to risk of developing cancer (2.7–5.2 years follow up; 625,477 person-years)

Three panels of proteins for: (i) early diagnosis of neoplasia (benign or malignant) and (ii) distinguishing benign from malignant

Use of LC-MS/MS followed by immunosassays to identify 5 serum protein biomarkers previously reported (Kozak et al., 2003). New analysis of sera from 43 OC patients and 51 healthy controls

Independent evaluation of ApoA-I as biomarker for OC in 182 patient sera (42 OC, 65 benign tumors, and 76 with digestive diseases) sera collected at Mayo Clinic (1980–1989)

Pre-surgery blood samples (41 early stage (I/II), 51 late stage (III/IV), 40 benign disease, and 99 healthy controls) analyzed by proteomics for seven previously identified biomarkers

Development of multiplexed bead-based immunoassay for detection of known serum biomarkers of cancer (118 OC, 84 benign ovarian disease, 61 healthy controls)

Evaluation of apoA-I and GPX3 transcript level by qPCR in effusions, and solid tumors from patients with OC versus those with breast cancer (BC) as diagnostic tool to differentiate between these two cancers (101 OC and 20 BC effusions; 85 solid OC (43 primary, 42 metastasis))

Protease profiling of serum samples from patients with OC versus those with breast cancer (101 OC, 20 each breast, colon, and prostate cancers with 41 healthy controls for validation by immunosassays)

Breast Lipid profile analysis in fasting patient sera prior to diagnostic biopsies (50 malignant and 50 benign)

Lipid profile analysis in fasting patient sera prior to diagnostic biopsies (50 malignant and 50 benign)

Lipid profile analysis of pre-diagnostic sera from age-matched 200 (100 before age 50 and 100 at age 50 or older) case-control BC patients nested from an original cohort of 95,000 women to examine prospective association of plasma HDL-C and BC incidence

Lipid profile analysis in fasting patient (Taiwanese) sera (150 cancer and 71 healthy controls) and association with BC risk

Multiple time point measurements of serum lipids and lipoproteins in a nested case-control (279 cases and 558 matched control subjects) study within a randomized long-term dietary intervention trial with 4,690 women for an average of 10 years to assess the association of serum lipids with the risk of cancer incidence based on menopausal status and use of hormone replacement therapy (HRT)

TABLE 1 | Clinical studies investigating role of apoA-I/HDL in cancer.

| Cancer type | Clinical Study Objective | Conclusion | Reference |
|-------------|-------------------------|------------|-----------|
| Broad variety | Examined the relationship between serum high-density lipoprotein cholesterol (HDL-C) and risk of overall and site-specific cancers among 29,093 Finnish male smokers in the Alpha-Tocopherol Beta-Carotene (ATBC) study cohort | Inverse association between HDL-C levels and the risk of cancer of the lung, prostate, liver, and hematopoietic system | Ahn et al., 2009 |
| Broad variety | Analysis of 24 randomized controlled trials of lipid-altering therapy (145,743 persons) to correlate baseline and on-treatment HDL-C levels to risk of developing cancer (2.7–5.2 years follow up; 625,477 person-years) | For every 10 mg/dL increase in HDL-C, 36 % lower risk of developing cancer | Jafri et al., 2010 |
| Ovarian | ProteinChip biomarker System and Mass spectrometry-based proteomic profiling (SELDI-TOF-MS) to identify disease associated biomarkers in serum samples from patients with ovarian cancer (OC), benign tumors, and healthy donors (109 OC, 19 benign tumors, and 56 healthy controls) | Three panels of proteins for: (i) early diagnosis of neoplasia (benign or malignant) and (ii) distinguishing benign from malignant | Kozak et al., 2003 |
| Ovarian/breast/colon/prostate | ProteinChip biomarker System and Mass spectrometry-based proteomic profiling (SELDI-TOF-MS) and immunosassays to identify and validate biomarkers in serum samples from patients with early stage OC as compared with healthy individuals and other cancers (195 OC, 166 benign tumors, 142 healthy controls for initial screening followed by 41 OC, 20 each breast, colon, and prostate cancers with 41 healthy controls for validation by immunosassays) | Apolipoprotein A-I (ApoA-I; down-regulated in OC), truncated transthyretin (TT; down-regulated in OC), and a cleavage fragment of inter-α-trypsin heavy chain H4 (up-regulated in OC) identified as biomarkers for OC | Zhang et al., 2004 |
| Ovarian | Use of LC-MS/MS followed by immunosassays to identify 5 serum protein biomarkers previously reported (Kozak et al., 2003). New analysis of sera from 43 OC patients and 51 healthy controls | ApoA-I (down-regulated), TT (down-regulated), transferrin (down-regulated), and hemoglobin (up-regulated) identified as biomarkers for OC with their expression reduced in disease | Kozak et al., 2005 |
| Ovarian | Independent evaluation of ApoA-I as biomarker for OC in 182 patient sera (42 OC, 65 benign tumors, and 76 with digestive diseases) sera collected at Mayo Clinic (1980–1989) | ApoA-I and TT were confirmed as biomarkers for OC with their expression reduced in disease | Moore et al., 2006 |
| Ovarian | Pre-surgery blood samples (41 early stage (I/II), 51 late stage (III/IV), 40 benign disease, and 99 healthy controls) analyzed by proteomics for seven previously identified biomarkers | ApoA-I as well as TT, and connective tissue activating protein III (CTAPIII), were confirmed as a panel of biomarker together with CA125 with increased sensitivity for detection of early stage OC | Clarke et al., 2011 |
| Ovarian | Development of multiplexed bead-based immunoassay for detection of known serum biomarkers of cancer (118 OC, 84 benign ovarian disease, 61 healthy controls) | Combination of transthyretin, and apoA-I with CA125 improved sensitivity and specificity of OC diagnosis | Kim et al., 2012 |
| Ovarian/breast | Evaluation of apoA-I and GPX3 transcript level by qPCR in effusions, and solid tumors from patients with OC versus those with breast cancer (BC) as diagnostic tool to differentiate between these two cancers (101 OC and 20 BC effusions; 85 solid OC (43 primary, 42 metastasis)) | ApoA-I transcript in all anatomic sites was higher in OC compared with BC. Higher apoA-I mRNA levels in primary diagnosis pre-chemotherapy effusions were significantly related to better overall survival | Tuft Stavnes et al., 2014 |
| Pancreatic | ProteinChip biomarker System and Mass spectrometry-based proteomic profiling (SELDI-TOF-MS) to identify biomarkers in patient pre-surgery sera (96 cancer patients, 96 healthy controls) | ApoA-I, apoA-II and transthyretin identified as biomarkers with inverse correlation to pancreatic cancer | Ehmann et al., 2007 |
| Breast | Lipid profile analysis in fasting patient sera prior to diagnostic biopsies (50 malignant and 50 benign) | Lower apoA-I levels predict cancer recurrence | Lane et al., 1995 |
| Breast | Lipid profile analysis of pre-diagnostic sera from age-matched 200 (100 before age 50 and 100 at age 50 or older) case-control BC patients nested from an original cohort of 95,000 women to examine prospective association of plasma HDL-C and BC incidence | Low plasma HDL-C predicts risk of developing BC only in pre-menopausal women. Each 1 mg/dL increase in HDL-C is associated with a 4% reduction in risk of BC | Moorman et al., 1998 |
| Breast | Assessed risk of BC associated with serum HDL-C in 38,823 Norwegian women with a median follow-up of 17.2 years | Low HDL-C, as part of the metabolic syndrome, is associated with increased postmenopausal BC risk | Furberg et al., 2004 |
| Breast | Lipid profile analysis in fasting patient (Taiwanese) sera (150 cancer and 71 healthy controls) and association with BC risk | ApoA-I levels in serum inversely associated with BC risk | Chang et al., 2007 |
| Breast | Evaluate association of baseline HDL-C levels with cancer incidence using data from the Atherosclerosis Risk in Communities Study (ARIC) cohort with follow-up from 1987 through 2000 | Modest direct association of low HDL-C with risk of developing BC only in women who were premenopausal at baseline | Kucharska-Newton et al., 2008 |
| Breast | Multiple time point measurements of serum lipids and lipoproteins in a nested case-control (279 cases and 558 matched control subjects) study within a randomized long-term dietary intervention trial with 4,690 women for an average of 10 years to assess the association of serum lipids with the risk of cancer incidence based on menopausal status and use of hormone replacement therapy (HRT) | HDL-C and apoA-I were positively associated with BC risk only when HRT was not used | Martin et al., 2015 |

(Continued)
TABLE 1 | Continued

| Cancer type          | Clinical Study Objective                                                                                                                                                                                                 | Conclusion                                                                                     | Reference                      |
|----------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------|---------------------------------|
| Nasopharyngeal       | Retrospective analysis of 807 patients with metastatic nasopharyngeal carcinoma (NPC) to assess prognostic value of baseline serum lipids in predicting overall survival                                                                 | Higher values of HDL-C and apoA-I were associated with improved overall survival                | Jiang et al., 2014              |
| Endometrial          | Examined the association of pre-diagnostic plasma levels of lipids, lipoproteins and other metabolic factors in developing cancer in a nested case-control (262 cases and 548 matched control subjects) study of the European Prospective Investigation into Cancer and Nutrition (EPIC; 520,000 participants from 10 western European countries) | HDL-C levels were inversely correlated with the risk of developing cancer                        | Cust et al., 2007               |
| Colorectal           | Examined the association of pre-diagnostic plasma levels of lipids, lipoproteins and other metabolic factors in developing cancer in a nested case-control (1,238 cases, 1,238 matched control subjects) study of the European Prospective Investigation into Cancer and Nutrition (EPIC; 520,000 participants from 10 western European countries) | Higher pre-diagnostic HDL-C and apoA-I were statistically significantly inversely associated with risk of colon cancer, but not rectal cancer | van Duijnhoven et al., 2011     |

Clinical studies which established an association between plasma levels of apoA-I/HDL-C and risk of developing a broad spectrum of cancers.

for their ability to form class A amphipathic helices with lipid binding capability, a salient functional feature of apoA-I (Kanellis et al., 1980; Segrest et al., 1983; Anantharamaiah et al., 1985; Mendez et al., 1994). ApoA-I/HDL mimetics have been successfully used in a number of mouse models of atherosclerosis (Garber et al., 2001; Navab et al., 2010; Getz and Reardon, 2011) and shown to have anti-inflammatory and anti-oxidant activities as well as the ability to promote RCT (D’Souza et al., 2010; Ditiakovski et al., 2013). These peptides have similar binding affinities for non-oxidized lipids but a significantly higher affinity for pro-inflammatory oxidized lipids (Van Lenten et al., 2008) and for lysophosphatidic acid (LPA; Su et al., 2010), relative to full-length apoA-I. LPA is known to promote tumor development (Mills and Moofoza, 2003; Li et al., 2009) and has been identified as a biomarker for OC (Xu et al., 1998; Sutphen et al., 2004). Consequently, due to their anti-inflammatory activity and increased lipid binding properties apoA-I/HDL mimetics, when tested for anti-tumor activity, were shown to have protective activity in multiple mouse tumor models including ovarian (Su et al., 2010; Gao et al., 2011, 2012; Ganapathy et al., 2012), and colon cancer (Su et al., 2012). It was shown for OC that the primary functional activity of the mimetic peptides was due to squelching of the bioactive, tumor-promoting LPA and oxidized lipids that serve as potent tumor growth and angiogenic factors. The mimetic peptides 4F and 5F when injected or administered orally, inhibited tumor growth and reduced plasma levels of LPA in the tumor bearing mice (Su et al., 2010). Likewise, in another study the functional target of the apoA-I/HDL mimetic peptide L-4F was presumed to be LPA as administration of this HDL-mimetic peptide was shown to decrease circulating levels of LPA in mimetic-treated animals and demonstrated protection against colon cancer (Su et al., 2012). ApoA-I mimetics were also reported to inhibit tumor angiogenesis in vivo and abrogate growth factor induced proliferation, migration, invasion, and tube formation of endothelial cells in vitro (Gao et al., 2011). The mimetics significantly decreased LPA-induced vascular endothelial growth factor (VEGF) production by cancer cells through inhibition of hypoxia-inducible factor-1α (HIF-1α) as well as interfering with VEGF-induced signaling in endothelial cells thus mitigating VEGF’s ability to promote angiogenesis (Gao et al., 2011, 2012). Furthermore, the mimetics were shown to inhibit cell viability and proliferation of OC cells by reducing the oxidative stress in cancer cells through induced expression of the tumor suppressor enzyme, manganese-containing superoxide dismutase (MnSOD; Ganapathy et al., 2012). Although these apoA-I/HDL peptide mimetics have demonstrated very interesting anti-tumor properties, their apparent mechanism of action (squelching LPA for the most part) does not appear to coincide with the anti-tumor mechanisms of actual apoA-I/HDL (Zamanian-Daryoush et al., 2013).

MECHANISM OF ApoA-I/HDL ANTI-TUMOR ACTIVITY

ApoA-I/HDL Negatively Impacts Tumor-permissive Features of the Tumor Microenvironment

Although the exact molecular mechanism of apoA-I/HDL anti-tumor activity is not known, studies with the syngeneic B16F10L tumors comparing those from apoA-I transgenic vs. A-I KO mice revealed that the overall net impact of host apoA-I on the tumor microenvironment is profound and manifold. These include but are not limited to the following as depicted in Figure 1: decreased recruitment of myeloid-derived suppressor cells (MDSC) and reduced angiogenesis resulting in decreased tumor volume; decreased matrix metalloproteinase-9 (MMP9) protein levels and enzyme activity; decreased overall metastasis, increased accumulation of tumor-associated macrophages (TAMs) with an M1-like anti-tumor phenotype; increased levels of tumor cell killing macrophages; increased recruitment of CD8T cells and decreased levels of the anti-apoptotic protein survivin within the tumor bed (Zamanian-Daryoush et al., 2013). Unexpectedly, levels of the pro-angiogenic protein, VEGF-A, was higher in tumors from A-I Tg mice relative...
FIGURE 1 | Net functional effects of apoA-I/HDL in the tumor microenvironment. This scheme is based on comparative analyses of B16F10L melanoma tumors resected from mice either deficient in apoA-I or mice expressing human apoA-I (Zamanian-Daryoush et al., 2013). (A) The effect of apoA-I/HDL on tumor-infiltrating myeloid cell population. ApoA-I/HDL promotes the accumulation of TAMs with an M1-like anti-tumor phenotype and inhibits the accumulation of the tumor promoting M2-like TAMS as well as MDSCs.

(B) The impact of apoA-I/HDL on tumor vasculature, metastasis, and tumor survival, all phenomena that are subject to regulation by myeloid cells. Apolipoprotein A-I, ApoA-I; high density lipoprotein, HDL; interleukin-10, IL-10; interleukin-12, IL-12; tumor-associated macrophages, TAM; myeloid-derived suppressor cells, MDSCs; matrix metalloproteinase-9, MMP-9. Solid lines ending in arrow head signify promotion whereas solid lines ending with a perpendicular dotted line signify inhibition of biological pathways.

to A1KO mice (Zamanian-Daryoush et al., 2013). It may be that the VEGF-A protein quantified in these studies was VEGF-Ax, an anti-angiogenic VEGF-A variant recently reported to be a product of translational read-through (Eswarappa et al., 2014) though this remains to be proven. Both the antibody and the oligonucleotide probe sets used to detect VEGF-A in Zamanian-Daryoush et al. (2013) cannot distinguish between the two forms. Unlike the observations with apoA-I/HDL peptide mimetics, which appear to inhibit tumor growth by squelching/decreasing circulating levels of the bioactive tumor promoting lipid LPA; the plasma level of LPA, was found to be similar in B16F10L melanoma tumor-bearing A1 KO vs. A-I Tg animals (Zamanian-Daryoush et al., 2013), thus eliminating titration of this bioactive lipid as a functional mechanism, in this setting, for apoA-I protein (Zamanian-Daryoush et al., 2013).
ApoA-I/HDL Anti-tumor Activity Requires an Intact Innate and Adaptive Immune System for Full Anti-tumor Activity

ApoA-I infusion studies with mice deficient in various aspects of immune system revealed that complete apoA-I anti-tumor activity requires both innate and adaptive arms of immunity. In particular, TAMs isolated from tumors resected from A-I Tg animals were skewed toward an M1-like phenotype and macrophages isolated from tumor-bearing A-I Tg animals exhibited enhanced cytotoxicity toward B16F10L tumor cells in vitro (Zamanian-Daryoush et al., 2013). Macrophages are innate immune cells with pivotal roles in tumor biology. Their ability to destroy tumors or promote their development is a function of their phenotypic plasticity, which is governed, in major part, by growth factors, cytokines, and bioactive lipids present in the tumor microenvironment (Mantovani et al., 2005; Martinez et al., 2009; Solinas et al., 2009; Liu and Yang, 2013). TAMs are in general, alternatively activated (M2-like phenotype) and promote angiogenesis, tumor survival, and metastasis, whereas classical activated macrophages (M1-like phenotype) inhibit these processes leading to tumor inhibition. Conversion of macrophage phenotype from M2 to M1 has been demonstrated to result in tumor inhibition in several murine tumor models (Colombo and Mantovani, 2005; Guiducci et al., 2005). The finding that in the tumor microenvironment, apoA-I/HDL promotes the accumulation of macrophages in pro-inflammatory, classically activated anti-tumor M1-like phenotype is in direct contrast with the traditional anti-inflammatory and immunosuppressive functions described for HDL in a typical inflammatory setting (Murphy et al., 2012). How apoA-I/HDL triggers this uncharacteristic effect is not known but is currently under investigation. The transcription factor activator of transcription factor 3 (ATF3) was recently identified as mediating anti-inflammatory and immune modulatory functions of HDL in macrophages (De Nardo et al., 2014). HDL induced the transcriptional expression of ATF3, which in turn inhibited the expression of some 130 toll-like receptor (TLR)-mediated pro-inflammatory genes with immune response-related functions (De Nardo et al., 2014). TLRs recognize pathogen-associated molecular pattern (PAMPs) molecules from viruses and microorganisms along with endogenously derived ligands such as damage-associated molecular pattern molecules (DAMPs) and can play important roles in both pro- and anti-tumor responses (Pradere et al., 2014). HDL inhibits lipopolysaccharide (LPS)-mediated TLR4 signaling by squelching the ligand itself and although HDL pretreatment was found to decrease cellular cholesterol concentration, it did not inhibit CpG-induced TLR9 signaling per se but reduced downstream gene expression by CpG through induction of ATF3 (De Nardo et al., 2014). These results indicate that apoA1/HDL can influence TLR signaling in normal inflammatory settings via at least two different routes.

HDL is an integral component of host immunity because its cargo, a series of bioactive proteins and lipids, has immunomodulatory activities. These activities include but are not limited to (i) ability to scavenge bacterial outer membrane components, LPS(Gram-negative bacteria) or lipoteichoic acid (LTA; Gram-positive bacteria) limiting their pro-inflammatory toxicity and protecting against sepsis (Levine et al., 1993; Birjmovun et al., 2007; Wendel et al., 2007; Guo et al., 2013), (ii) protection against intracellular bacteria such as mycobacteria (Cruz et al., 2008) and parasites (Vanhollebeke and Pays, 2010), (iii) in concert with ATP-binding cassette transporters ABCA1 and ABCG1, HDL has a role in control of hematopoietic stem and multipotential progenitor cell proliferation (Yvan-Charvet et al., 2010), and (iv) influencing immune cell response by modulating cholesterol content in membrane lipid rafts. Cholesterol accumulation in immune cells increases signaling by stabilizing lipid rafts in the plasma membrane as well as in other cellular membranes. Lipid rafts are cholesterol enriched micro-domains with sphingolipids and serve as docking sites for several receptors with important immunological functions, including TLRs (Fessler and Parks, 2011) and T- and B-cell receptors (TCRs; Gupta and DeFranco, 2007; Kabouridis and Jury, 2008). Cholesterol efflux by apoA-I/HDL via ABCA1 and ABCG1 disrupts lipid rafts and their associated signaling pathways (Triantafilou et al., 2002; Zhu et al., 2010). Thus cholesterol efflux-mediated modification of immune response was noted in: (i) monocytes/macrophages, inhibiting their activation and recruitment (Murphy et al., 2008) as well as skewing macrophages toward an M2-like anti-inflammatory immunosuppressive phenotype (Smythies et al., 2010; Feig et al., 2011; Murphy et al., 2012); (ii) neutrophils, inhibiting their migration and adhesion (Murphy et al., 2011); (iii) dendritic cells (DCs), inhibiting their maturation and their ability to induce T cell activation (Perrin-Cocon et al., 2012; Wang et al., 2012), while the immunoactive lipid sphingosine 1-phosphate (SIP) carried in HDL promotes an anti-inflammatory phenotype in DCs thus inhibiting a Th1 response (Idzko et al., 2002); (iv) T cells, inhibiting their activation and proliferation in peripheral lymph nodes in mice fed a high fat diet (Wilhelm et al., 2009). These observations are clearly supportive of apo A-I/HDL’s role as a physiological modulator of membrane cholesterol and immune function in different pathophysiological conditions.

Deletion of ApoA-I/HDL Receptors Lead to Altered Macrophage Phenotypes

Recently, myeloid-specific or global deletion of Abcg1 in mice was reported to be anti-tumorogenic with macrophages displaying a clear shift from an M2-like to an M1-like phenotype (Sag et al., 2015). The anti-tumor effect was demonstrated to be driven by myeloid cells, and in large part due to the increased accumulation of cholesterol in macrophages. Studies on bone marrow-derived macrophages (BMDMs) from Abcg1−/− mice compared to WT mice after treatment with either M1 or M2 polarizing cytokines and comparative analysis of M1 or M2 cell markers showed that the Abcg1−/− BMDMs were intrinsically biased to an M1 pro-inflammatory phenotype (Sag et al., 2015). These findings underscore the significance of cholesterol homeostasis in tumor immunity. Loss of Abca1 in myeloid cells results in increased accumulation of free cholesterol and enhanced pro-inflammatory responses in macrophages, highlighting the
normal anti-inflammatory function of these transporters (Yvan-Charvet et al., 2008; Zhu et al., 2008; Tang et al., 2009). Correspondingly, macrophages from mice globally deleted for scavenger receptor class B, type 1 (Sr-b1), have also been shown to exhibit hyperinflammatory responses (Cai et al., 2012). However, unlike the situation with Abca1 or Abcg1 deletion, the heightened inflammatory responses are not due to alterations in cell membrane or total cell cholesterol levels (Cai et al., 2012). It is not known what effect loss of ABCA1 or SR-B1 protein expression either globally or myeloid-specifically has on tumorigenesis but by extrapolation from tumor studies in mice with ABCG1 loss, one might predict a protective effect due to the heightened pro-inflammatory phenotype these transporter-/receptor-deleted macrophages are known to exhibit. The emerging paradigm from the studies by Zamanian-Daryoush and that of Sag et al., is that TAMs can be converted from an M2-like to an M1-like phenotype by either a hard-wiring to an M1 anti-tumor phenotype through loss of cholesterol/phospholipid transporter ABCG1 (Sag et al., 2015) and perhaps ABCA1 and SR-B1 or in response to increased apoA1/ HDL levels; which by unknown mechanisms alters immune responses to factors within the tumor microenvironment and promotes the accumulation of TAMs with an M1-like phenotype at the expense of TAMs with an immune suppressive M2-like phenotype.

CONCLUSION

In cancer, lipid and cholesterol homeostasis is often dysregulated to facilitate the cancer cells’ increased demand for these building blocks which are required for proliferation and evasion of apoptosis (Schaffner, 1981; Yoshioka et al., 2000; Kolanjiappan et al., 2003; Platz et al., 2009; Smith and Land, 2012). To this end, tumor cells can manipulate their intracellular cholesterol level by reducing expression of ABCA1 which effluxes cholesterol and increasing the expression of SR-B1 which influxes cholesterol. This phenomenon has been reported in several prostate, colon and BCs (Cao et al., 2004; Mooberry et al., 2010; Su et al., 2010; Smith and Land, 2012; Lee et al., 2013).

The ability to modulate lipid and cholesterol movement is at the core of apoA-I/HDL’s actions resulting in profound physiological and cell phenotypic effects. Changes in cholesterol metabolism or levels of components of cholesterol homeostasis namely apoA-I/HDL, ABCA1, ABCG1, and SR-B1 are known to affect immune responses which in turn impact anti-tumor effects. In fact, apoA-I/HDL anti-tumor effects were observed maximally only in fully immune-competent animals (Zamanian-Daryoush et al., 2013). Anti-tumor effects of apoA-I/HDL could be related to (i) the ability of apoA-I/HDL to modulate cholesterol content in immune or tumor cell membrane lipid rafts thus influencing signaling pathways, (ii) the lipid rafts’ role as a platform for biologically active lipids and proteins that may impact the immune response, (iii) the cross-talk between the tumor and surrounding stromal cells. These possibilities are not mutually exclusive. ApoA-I/HDL peptide mimetics appear to primarily function through titrating out bioactive lipids and molecules which function as potent tumor cell angiogenic and growth factors. Although apoA-I/HDL may also perform similar titrating actions, the extent to which this activity is consequential in cancers other than in ovarian and perhaps colon cancer, is not as apparent. ApoA-I/HDL appears to function as an anti-tumor agent in large part by modulating the anti-tumor immune response. In the context of infection or atherogenesis, apoA-I/HDL modulates macrophages toward an anti-inflammatory M2-like phenotype by effluxing cholesterol but in the tumor microenvironment, apoA-I/HDL promotes the accumulation of M1-like macrophages. At the present time, we do not know the mechanism involved in this process but this apparent dichotomy of apoA-I/HDL functional response in different inflammatory settings underscores the complexity of apoA-I/HDL biology and poses intellectual and experimental challenges toward a better understanding of this multifaceted plasma component.

FUNDING

Funding was provided by National Institutes of Health (NIH) grants P01HL076491 and P01HL098055.

ACKNOWLEDGMENTS

We thank J. Smith for careful review of the manuscript and S. L. Hazen for helpful scientific discussions and support. We are also grateful to Tom Hamilton for critical review of the Figure. “Illustration by David Schumick, BS, CMI. Reprinted with the permission of the Cleveland Clinic Center for Medical Art & Photography © 2015. All Rights Reserved”.

REFERENCES

Ahn, J., Lim, U., Weinstein, S. J., Schatzkin, A., Hayes, R. B., Virtamo, J., et al. (2009). Prediagnostic total and high-density lipoprotein cholesterol and risk of cancer. Cancer Epidemiol. Biomarkers. Prev. 18, 2814–2821. doi:10.1158/1055-9966.EPI-08-1248

Anantharamaiah, G. M., Jones, J. L., Brouillette, C. G., Schmidt, C. F., Chung, B. H., Hughes, T. A., et al. (1985). Studies of synthetic peptide analogs of the amphipathic helix. Structure of complexes with dimyristoyl phosphatidylcholine. J. Biol. Chem. 260, 10248–10255.

Bhattacharyya, T., Nicholls, S. J., Topol, E. J., Zhang, R., Yang, X., Schmitt, D., et al. (2008). Relationship of paraoxonase 1 (PON1) gene polymorphisms and functional activity with systemic oxidative stress and cardiovascular risk. JAMA 299, 1265–1276. doi: 10.1001/jama.2009.11265

Birjimohun, R. S., Van Leuven, S. I., Levels, J. H., Van’t Veer, C., Kuivenhoven, J. A., Meijers, J. C., et al. (2007). High-density lipoprotein attenuates inflammation and coagulation response on endotoxin challenge in humans. Arterioscler. Thromb. Vasc. Biol. 27, 1153–1158. doi: 10.1161/ATVBAHA.106.136325

Cai, L., Wang, Z., Meyer, J. M., Ji, A., and Van Der Westhuyzen, D. R. (2012). Macrophage SR-B1 regulates LPS-induced pro-inflammatory signaling in mice and isolated macrophages. J. Lipid Res. 53, 1472–1481. doi: 10.1194/jlr.M023234

Camont, L., Chapman, M. J., and Kontush, A. (2011). Biological activities of HDL subpopulations and their relevance to cardiovascular disease. Trends Mol. Med. 17, 594–603. doi: 10.1016/j.molmed.2011.05.013
Moorman, P. G., Hulka, B. S., Hiatt, R. A., Krieger, N., Newman, B., Vogelman, J. H., et al. (1998). Association between high-density lipoprotein cholesterol and breast cancer varies by menopausal status. Cancer Epidemiol. Biomarkers Prev. 7, 483–488.

Murphy, A. J., Westerterp, M., Yvan-Charvet, L., and Tall, A. R. (2012). Anti-atherogenic mechanisms of high density lipoprotein: effects on myeloid cells. Biochim. Biophys. Acta 1821, 513–521. doi:10.1016/j.bbalip.2011.08.003

Murphy, A. J., Woollard, K. J., Hoang, A., Mukhamedova, N., Sturzaker, R. A., McCormick, S. P., et al. (2008). High-density lipoprotein reduces the human monocyte inflammatory response. Arterioscler. Thromb. Vasc. Biol. 28, 2071–2077. doi:10.1161/ATVBHA.108.166890

Murphy, A. J., Woollard, K. J., Suhartoyo, A., Sturzaker, R. A., Shaw, J., Sviridov, D., et al. (2011). Neutrophil activation is attenuated by high-density lipoprotein and apolipoprotein A-I in vitro and in vivo models of inflammation. Arterioscler. Thromb. Vasc. Biol. 31, 1333–1341. doi:10.1161/ATVBHA.111.226258

Navab, M., Shechter, I., Anantharamaiah, G. M., Reddy, S. T., Van Lenten, B. J., and Fogelman, A. M. (2010). Structure and function of HDL mimetics. Arterioscler. Thromb. Vasc. Biol. 30, 164–168. doi:10.1161/ATVBHA.109.187518

Noguti, J., Andersen, M. L., Cirelli, C., and Ribeiro, D. A. (2013). oxidative stress; cancer; and sleep deprivation: is there a logical link in this association? Sleep Breath 17, 905–910. doi:10.1007/s11325-012-0797-9

Nosov, V., Su, F., Anmeanes, M., Birrer, M., Robins, T., Kotlerman, J., et al. (2009). Validation of serum biomarkers for detection of early-stage ovarian cancer. Am. J. Obstet. Gynecol. 200:639e1–639e5. doi:10.1016/j.ajog.2008.12.042

Nosov, V., Anmeanes, M., Su, F., Lang, J., Ianco, J. M., Reddy, S. T., et al. (2008). The early detection of ovarian cancer: from traditional methods to proteomics. Can we really do better than serum CA-125? Am. J. Obstet. Gynecol. 199, 215–223. doi:10.1016/j.ajog.2008.04.009

Perrin-Cocoun, L., Diaz, O., Carreras, M., Dollet, S., Su, F., Amneus, M., Birrer, M., Robins, T., Kotlerman, J., et al. (2012). High-density lipoprotein phospholipids interfere with dendritic cell Th1 functional maturation. Immunobiology 217, 91–99. doi:10.1016/j.imbio.2011.07.030

Platz, E. A., Till, C., Goodman, P. J., Parh, H. L., Figg, W. D., Albanes, D., et al. (2009). Men with low serum cholesterol have a lower risk of high-grade prostate cancer in the placebo arm of the prostate cancer prevention trial. Cancer Epidemiol. Biomarkers Prev. 18, 2807–2813. doi:10.1158/1055-9965.EPI-09-0472

Pradere, J. P., Dapito, D. H., and Schwabe, R. F. (2014). The Yin and Yang of Toll-like receptors in cancer. Oncogene 33, 3485–3495. doi:10.1038/onc.2013.302

Rader, D. J., and Hovingh, G. K. (2014). HDL and cardiovascular disease. Lancet 384, 618–625. doi:10.1016/S0140-6736(14)61217-4

Reddy, S. T., Navab, M., Anantharamaiah, G. M., and Fogelman, A. M. (2014). Searching for a successful HDL-based treatment strategy. Biochim. Biophys. Acta 1841, 162–167. doi:10.1016/j.bbalip.2013.10.012

Rosenson, R. S., Brewer, H. B. Jr., Davidson, W. S., Fayad, Z. A., Fuster, V., Goldstein, J., et al. (2012). Cholesterol efflux and atheroprotection: advancing the concept of reverse cholesterol transport. Circulation 125, 1905–1919. doi:10.1161/CIRCULATIONAHA.111.066389

Rye, K. A., Bursill, C. A., Lambert, G., Tabet, F., and Barter, P. J. (2009). The metabolism and anti-atherogenic properties of HDL. J. Lipid Res. 50(Suppl.), S195–S200. doi:10.1194/jlr.R800034-JLR200

Sac, D., Cekic, C., Wu, R., Linden, J., and Hedrick, C. C. (2015). The cholesterol transporter ABCG1 links cholesterol homeostasis and tumour immunity. Nat. Commun. 6:6354. doi:10.1038/ncomms7534

Schaffner, C. P. (1981). Prostatic cholesterol metabolism: regulation and alteration. Prog. Clin. Biol. Res. 75A, 279–324.

Segrest, J. P., Chung, B. H., Brouillette, C. G., Kanelis, P., and McGahan, R. (1983). Studies of synthetic peptide analogs of the amphipathic helix. Competitive displacement of exchangeable apolipoproteins from native lipoproteins. J. Biol. Chem. 258, 2290–2295.

Smith, B., and Land, H. (2012). Anticancer activity of the cholesterol exporter ABCA1 gene. Cell Rep. 2, 580–590. doi:10.1016/j.celrep.2012.08.011

Smtyardis, L. E., White, C. R., Maheshwari, A., Palgunachari, M. N., Anantharamaiah, G. M., Chaddha, M., et al. (2010). Apolipoprotein A-I mimetic 4F alters the function of human monocyte-derived macrophages. Am. J. Physiol. Cell Physiol. 298, C1538–C1548. doi:10.1152/ajpcell.00647.2009

Solinas, G., Germano, G., Mantovani, A., and Allavena, P. (2009). Tumor-associated macrophages (TAM) and major players of the cancer-related inflammation. J. Leukoc. Biol. 86, 1065–1073. doi:10.1189/jlb.0609385

Sorci-Thomas, M. G., Bhat, S., and Thomas, M. J. (2009). Activation of lecithin:cholesterol acyltransferase by HDL ApoAI central helices. Clin. Lipidol. 4, 113–124. doi:10.2217/17584299.4.1.113
Su, F., Grijalva, V., Navab, K., Ganapathy, E., Meriwether, D., Imaizumi, S., et al. (2012). HDL mimetics inhibit tumor development in both induced and spontaneous mouse models of colon cancer. Mol. Cancer Ther. 11, 1311–1319. doi: 10.1158/1355-7136.MCT-11-0993

Su, F., Kozak, K. R., Imaizumi, S., Gao, F., Amneus, M. W., Grijalva, V., et al. (2010). Apolipoprotein A-1 (apoA-I) and apoA-I mimetic peptides inhibit tumor development in a mouse model of ovarian cancer. Proc. Natl. Acad. Sci. U.S.A. 107, 19997–20002. doi: 10.1073/pnas.100910107

Su, F., Lang, J., Kumar, A., Ng, C., Hsieh, B., Suchard, M. A., et al. (2007). Validation of candidate serum ovarian cancer biomarkers for early detection. Biomark Insights 2, 369–375.

Sutphen, R., Xu, Y., Wilbanks, G. D., Fiorica, J., Grendys, E. C. J., LaPolla, J. P., et al. (2004). Lysosphospholipids are potential biomarkers of ovarian cancer. Cancer Epidemiol. Biomarkers Prev. 13, 1185–1191.

Tang, C., Liu, Y., Kessler, P. S., Vaughan, A. M., and Oram, J. F. (2009). The macrophage cholesterol exporter ABCA1 functions as an anti-inflammatory receptor. J. Biol. Chem. 284, 32336–32343. doi: 10.1074/jbc.M109.04772

Triantafilou, M., Miyake, K., Golenbock, D. T., and Triantafilou, K. (2002). Mediators of innate immune recognition of bacteria concentrate in lipid rafts and facilitate lipopolysaccharide-induced cell activation. J. Cell Sci. 115, 2603–2611.

Tuft Stavnes, H., Nymoen, D. A., Hetland Falkenthal, T. E., Kaern, J., Trope, C. G., and Davidson, B. (2014). APOA1 mRNA expression in ovarian serous carcinoma effusions is a marker of longer survival. Am. J. Clin. Pathol. 142, 51–57. doi: 10.1093/ajcpdx/NBHRXQL7

van Duijnhoven, F. J., Bueno-De-Mesquita, H. B., Calligaro, M., Jenab, M., Jansen, E. H., et al. (2011). Blood lipid and lipoprotein concentrations and colorectal cancer risk in the European Prospective Investigation into Cancer and Nutrition. Gut 60, 1094–1102. doi: 10.1136/gut.2010.225011

Vanhollebeke, B., and Pays, E. (2010). The trypanolytic factor of human serum: many ways to enter the parasite, a single way to kill. Mol. Microbiol. 76, 806–814. doi: 10.1111/j.1365-2958.2010.07156.x

VanLenten, B. J., Wagner, A. C., Jung, C. L., Ruchala, P., Waring, A. J., Lehrer, R. I., et al. (2008). Anti-inflammatory apolipoprotein A-I mimetic peptides bind oxidized lipids with much higher affinity than human apolipoprotein A-I. J. Lipid Res. 49, 2302–2311. doi: 10.1194/jlr.M801408200

Wang, S. H., Yuan, S. G., Peng, D. Q., and Zhao, S. P. (2012). HDL and ApoA-I inhibit antigen presentation-mediated T cell activation by disrupting lipid rafts in antigen presenting cells. Atherosclerosis 225, 105–114. doi: 10.1016/j.atherosclerosis.2012.07.029

Wendel, M., Paul, R., and Heller, A. R. (2007). Lipoproteins in inflammation and sepsis. II. Clin. Aspects. Intensive Care Med. 33, 25–35. doi: 10.1007/s00134-006-0433-x

Wilhelm, A. J., Zabalawi, M., Grayson, J. M., Weant, A. E., Major, A. S., Owen, J., et al. (2009). Apolipoprotein A-1 and its role in lymphocyte cholesterol homeostasis and autoimmunity. Arterioscler. Thromb. Vasc. Biol. 29, 843–849. doi: 10.1161/ATVBAHA.108.183442

Xu, Y., Shen, Z., Wiper, D. W., Wu, M., Morton, R. E., Elson, P., et al. (1998). Lysophosphatic acid as a potential biomarker for ovarian and other gynecologic cancers. JAMA 280, 719–723. doi: 10.1001/jama.280.8.719

Yoshioka, Y., Sasaki, J., Yamamoto, M., Saitoh, K., Nakaya, S., and Kubokawa, M. (2000). Quantitation by [(1)H-NMR of dolichol, cholesterol and choline-containing lipids in extracts of normal and pathological thyroid tissue. NMR Biomed. 13, 377–383. doi: 10.1002/1099-1492(200011)13:7<377:AID-NMB658>3.0.CO;2-E

Zamanian-Daryoush, M., Lindner, D., Tallant, T. C., Wang, Z., Buffa, J., Klipfell, E., et al. (2013). The cardioprotective protein apolipoprotein A1 promotes potent anti-tumorigenic effects. J. Biol. Chem. 288, 21237–21252. doi: 10.1074/jbc.M113.468967

Zhang, Z., Bast, R. C. Jr., Yu, Y., Li, J., Sokoll, L. J., Rai, A. J., et al. (2004). Three biomarkers identified from serum proteomic analysis for the detection of early stage ovarian cancer. Cancer Res. 64, 5882–5890. doi: 10.1158/0008-5472.CAN-04-0746

Zhu, X., Lee, J. Y., Timmins, J. M., Brown, J. M., Boudyguina, E., Mulya, A., et al. (2008). Increased cellular free cholesterol in macrophage-specific Abca1 knock-out mice enhances pro-inflammatory response of macrophages. J. Biol. Chem. 283, 22930–22941. doi: 10.1074/jbc.M801408200

Zhu, X., Owen, J. S., Wilson, M. D., Li, H., Griffiths, G. L., Thomas, M. J., et al. (2010). Macrophage ABCA1 reduces MyD88-dependent Toll-like receptor trafficking to lipid rafts by reduction of lipid raft cholesterol. J. Lipid. Res. 51, 3196–3206. doi: 10.1194/jlr.M006486

Conflict of Interest Statement: Drs. Maryam Zamanian-Daryoush and Joseph A. DiDonato report being listed as co-inventor on pending and issued patents held by the Cleveland Clinic.

Copyright © 2015 Zamanian-Daryoush and DiDonato. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.