Innate immune responses are effective for insect survival to defend against entomopathogens including a fungal pathogen, *Metarhizium rileyi*, that infects a lepidopteran *Spodoptera exigua*. In particular, the fungal virulence was attenuated by cellular immune responses, in which the conidia were phagocytosed by hemocytes (insect blood cells) and hyphal growth was inhibited by hemocyte encapsulation. However, the chemokine signal to drive hemocytes to the infection foci was little understood. The hemocyte behaviors appeared to be guided by a Ca^{2+} signal stimulating cell aggregation to the infection foci. The induction of the Ca^{2+} signal was significantly inhibited by the cyclooxygenase (COX) inhibitor. Under the inhibitory condition, the addition of thromboxane A2 or B2 (TXA2 or TXB2) among COX products was the most effective to recover the Ca^{2+} signal and hemocyte aggregation. TXB2 alone induced a microaggregation behavior of hemocytes under *in vitro* conditions. Indeed, TXB2 titer was significantly increased in the plasma of the infected larvae. The elevated TXB2 level was further supported by the induction of phospholipase A2 (PLA2) activity in the hemocytes and subsequent up-regulation of COX-like peroxinectins (*SePOX*-F and *SePOX*-H) in response to the fungal infection. Finally, the expression of a thromboxane synthase (*Se-TXAS*) gene was highly expressed in the hemocytes. RNA interference (RNAi) of *Se-TXAS* expression inhibited the Ca^{2+} signal and hemocyte aggregation around fungal hyphae, which were rescued by the addition of TXB2. Without any ortholog to mammalian thromboxane receptors, a prostaglandin receptor was essential to mediate TXB2 signal to elevate the Ca^{2+} signal and mediate hemocyte aggregation behavior. Specific inhibitor assays suggest that the downstream signal after binding TXB2 to the receptor follows the Ca^{2+}-induced Ca^{2+} release pathway from the endoplasmic reticulum of the hemocytes. These results suggest that hemocyte aggregation induced by the fungal infection is triggered by TXB2 via a Ca^{2+} signal through a PG receptor.

Keywords: insect, fungi, thromboxane, *Spodoptera exigua*, hemocyte
INTRODUCTION

Insect innate immunity includes cellular and humoral immune responses to prokaryotic and eukaryotic pathogens and parasites (1). Cellular responses are acutely induced and performed by mesodermal blood cells called hemocytes. Cellular actions include phagocytosis, encapsulation, and nodulation, depending on the type of invading pathogens (2, 3). In humoral immunity, antimicrobial peptides (AMPs) are produced and secreted by fat body and some hemocytes into hemolymph circulation to remove the residual pathogens after the cellular immune defense (4). Immune mediators such as cytokines and eicosanoids induce cellular and humoral immune responses to effectively defend against entomopathogens in insects (5, 6).

Eicosanoids are a group of oxygenated C20 polyunsaturated fatty acids including prostaglandins (PGs) that mediate cellular and humoral responses against various pathogens in as well as mammals (7). PGs are usually derived from arachidonic acid (AA: 5,8,11,14-eicosatetraenoic acid) by cyclooxygenase (COX) (8). However, terrestrial insects lack AA in phospholipids and thus transform C18 linoleic acid to AA by elongase and desaturase (9, 10). AA is then oxygenated by the dioxygenase activity of cyclooxygenase (COX) to form PGH2, which is isomerized by various cell-specific PG synthases to form prostanoids (11). More specifically, a prostanoid, thromboxane A2 (TXA2) is formed by TXA2 synthase (TXAS), a member of the cytochrome P450 epxoygenase superfamily. TXA2 exerts its biological activity through a G protein-coupled receptor called TP (12).

Thromboxanes act in blood clotting in mammals by reducing blood flow through vasoconstriction and driving platelets near to the clotting site (13). A similar blood clotting process has been reported in Drosophila melanogaster by demonstrating the presence coagulation factors commonly found in humans (14). Eicosanoids mediate the wound healing process since a Drosophila line mutant in PLA2 suffers hemolymph coagulation failures (15). Although this study did not identify specific eicosanoids, prostanoids like thromboxanes might be associated with wound healing in Drosophila. Two thromboxanes (TXA2 and TXB2) mediate immune responses in a model lepidopteran insect, Spodoptera exigua (16). Chemical identification of TXB2 in fat body and the presence of its biosynthetic gene (Se-TXAS) supported the physiological role of these thromboxanes.

Various roles of PGs in immune mediation have been unraveled in selected insect species. PGs mobilize sessile hemocytes to increase the number of circulating hemocytes in plasma in S. exigua (17). In Manduca sexta, PGs promote the migration of hemocytes to infection foci (18). Anopheles mosquito midgut cells produce and release PGE2, which attracts hemocytes and establishes a systemic cellular immune response to the malarial pathogen (19). In the infection site, PGs activate hemocytes to extend cytoplasm to induce hemocyte-spreadng behavior, which is required for phagocytosis (20), nodulation, and encapsulation (21, 22). PGs also induce the release of phenoloxidase (PPO) from oenocytes (a type of hemocytes) into hemolymph, where it is activated to phenoloxidase (PO) for melanin formation around nodules and encapsulated parasitoids (23). PGs mediate the synthesis of various AMPs among insect species (20, 24). Despite the multi-functional roles of PGs, we began to understand their respective physiological functions because thromboxanes do not mediate female reproduction unlike other PGs (PGD2 and PGE2) that mediate oocyte development in Drosophila melanogaster and S. exigua (25–27). These suggest that prostanoids may mediate their specific physiological processes within general immune responses, such as nodulation.

Based on the roles of thromboxanes in platelet aggregation in the process of mammalian blood clotting, we tested the physiological role of thromboxanes in recruiting hemocytes to infection foci in insects using S. exigua. To explain the cellular behavior with respect to intracellular processes, we assessed Ca2+ signaling in response to thromboxanes in S. exigua.

MATERIALS AND METHODS

Insect Rearing

Larvae of two lepidopteran species (S. exigua and Plutella xylostella) were collected from Welsh onion (Allium fistulosum L.) and Chinese cabbage (Vigna angularis) fields in Andong, Korea, respectively. Coleopteran (Tenebrio molitor) larvae were provided by Bio Utility (Andong, Korea). Insects were reared in a laboratory under our standard conditions of 25 ± 2°C temperature, 16:8 h (L:D) photoperiod, and 60 ± 5% relative humidity (RH). S. exigua larvae were reared on an artificial diet (28) and underwent five larval instars (L1-L5). Larvae of P. xylostella were reared on fresh cabbage leaves and underwent four larval instars (L1-L4). Approximately 3.5 cm body length T. molitor larvae were used for pathogenicity testing.

Chemicals

Arachidonic acid (AA: 5,8,11,14-eicosatetraenoic acid), dexamethasone [DEX: (11β, 16α)-9-fluoro-11,17,21-trihydroxy-16-methylprega-1,4-diene-3], naproxen [NAP: (S)- (+)-2-(6-methoxy-2-naphthyl)propionic acid], and esculentin (ESC: 6-hydroxy-7-methoxycoumarin) were purchased from Sigma-Aldrich Korea (Seoul, Korea) and dissolved in dimethyl sulfoxide (DMSO). Fura-8AM was purchased from AAT Bioquest (Sunnyvale, CA, USA) and dissolved in DMSO. Thromboxane B2 (TXB2: 9α,11,15S-trihydroxythromboxa-5Z,13E-dien-1-oic acid) and thromboxane A2 (TXA2: 9α,11α-methylene-15S-hydroxy-11α-deoxy-11α-methylene-thromboxa-5Z,13E-dien-1-oic acid) were purchased from Cayman Chemicals (Ann Arbor, MI, USA). DAN (dantrolene sodium: 1-[(5-(p-nitrophenyl) furfurylidene)amino] hydantoin sodium salt) were purchased from Sigma-Aldrich Seoul, Seoul, Korea. Prostaglandin E2 (PGE2: 5Z,11α,13E,15S)-11,15-dihydroxy-9-oxoprosta-5,13-dienoic acid), prostaglandin D2 (PGD2: 9α,15S-dihydroxy-11-oxoprosta-5Z,13E-dien-1-oic acid), prostaglandin I2 (PGI2: 6,9α-epoxy-11α,15S-dihydroxy-prosta-5Z,13E-dien-1-oic acid), 2-aminoethoxydiphenylborate (2-APB), thapsigargin (TPG), U-73122 (1-[(6-[17β]-3-methoxyestra-1,3,5[10]-trien-17-yl)amino]hexyl)-1H-pyrrole-2,5-dione), terutroban (TTB), and dazoxiben (DAZ) were purchased from Sigma-Aldrich.
Korea and dissolved in DMSO. Phosphate-buffered saline (PBS) was prepared with 100 mM phosphoric acid and adjusted to pH 7.4. Anticoagulant buffer (ACB) was prepared with 186 mM NaCl, 17 mM Na₂EDTA, and 41 mM citric acid.

**Bioinformatics for Prediction of G Protein-Coupled Receptors**

Based on GPCR sequences of *Bombbyx mori* (GenBank accession number: NP_001370303.1), *S. litura* (XP_022813850, XP_022814039, XP_022816105, XP_022817498, XP_022817499, XP_022821197, XP_022822728, XP_022822897, XP_022823006, XP_022826743, XP_022827237, XP_022827354, XP_022827490, XP_022827519, XP_022827626, XP_022834024, XP_022834058, XP_022834098, XP_022838045), *Heliothis virescens* (CDF56917.1), and *Trichoplusia ni* (XP_026733990), their orthologs of *S. exigua* were inferred from Transcriptome Shotgun Assembly (GGRZ01048721.1, GGRZ01057920.1, GGRZ01138956.1, GGRZ01102221.1, GGRZ01084708.1, GGRZ01092489.1, GGRZ01018271.1, GRRG01043383.1, GGRZ01038487.1, GGRZ01037013.1, GARL01065754.1, GARL01040814.1, GGRZ01082913.1, GGRZ01068498.1, GGRZ01117535.1, GGRZ01069701.1, GGRZ01224538.1, GGRZ01247645.1, GGRZ01215736.1, GGRZ01095571.1, GGRZ01137651.1, GGRZ01106723.1, GGRZ01113408.1, GGRZ01069261.1) using the BlastN program (https://www.ncbi.nlm.nih.gov/orffinder). The resulting sequences were subjected to open reading frame (ORF) analysis by using ORFfinder (https://www.ncbi.nlm.nih.gov/orffinder/). The ORF sequences were deposited to GenBank with accession numbers (AE027700.1, AXY04240.1, AXY04245.1, AXY04246.1, AXY04251.1, AXY04254.1, AXY04275.1, AXY04297.1, AXY04299.1, AZA07970.1). Alternatively, the protein coding sequences of the GPCRs were inferred by mapping the protein sequences of the GPCR sequences at NCBI and newly identified ORF sequences against a reference whole genome assembly in *S. exigua* (GCA_011316535) using exonarate-2.2.0 with protein2genome model. Phylogenetic analysis was performed with the Neighbor-Joining method and a Poisson correction model using MEGA6.06 software (www.megasoftware.net). Bootstrapping values were obtained with 1,000 replications to test supports on each node in the resulting phylogenetic tree.

**Entomopathogenic Fungi Source and Culturing**

Larvae of *S. exigua* infesting Welsh onion in Andong, Korea suffered from green muscardine disease symptoms and were collected for diagnosis. Fungal spores were collected in tubes containing sterile water by scraping from infected insect larvae. After stirring, 250 µL of the fungal suspension was spread on potato dextrose agar (PDA) and incubated at 25 ± 1°C and 70 ± 5% RH. The cultured spores were sub-cultured on PDA.

**Morphological Identification of EPF Isolate**

The cultured fungal colonies were transferred onto slides with PVA mounting medium (PVA MTNG) (BioQuip Products, Gladwick Street, CA, USA) and incubated at 50°C for 48 h. The slides were observed under an optical microscope (DM500, Leica, Wetzlar, Germany) with 400× magnification. Fungal samples were collected from PDA cultured for 14 days and subjected to Au-coating using a Sputter Coating machine (Jeol Korea, Seoul, Korea). The treated samples were observed under a scanning electron microscope (SEM; JSM-6300, Jeol Korea) at 200 × magnification. The resulting morphological characters were used for the identification of the entomopathogenic fungi according to a taxonomy of key characters such as colony, conidiophore, and conidial shapes (29).

**Molecular Identification of EPF Isolate**

Genomic DNA (gDNA) was extracted from a mixture of isolated spores and hyphae with phenol extraction and ethanol precipitation, as described by Park and Kim (17). Internal transcribed spacer (ITS) region was amplified using ITS forward primer (5’-TTGATTACGTCCGTGCCCC-3’) and ITS reverse primer (5’-TTCACCTGCGCTTTACTAAGG-3’) as reported by Vrain et al. (30). For PCR amplification of the ITS region, the extracted gDNA was used as a template with 35 cycles under the following conditions: 1 min at 94°C for denaturation, 1 min at 56°C for annealing, and 1 min at 72°C for extension. A second PCR was conducted using M13 universal primer-conjugated primers and sequenced by Macrogen (Seoul, Korea). The obtained nucleotide sequence was analyzed using the BlastN program of the National Center for Biotechnology Information (NCBI, www.ncbi.nlm.nih.gov). The evolutionary relationship was inferred using a Neighbor-Joining phylogenetic tree with MEGA6.06 (31). Bootstrap values on the branches were estimated with 1,000 replications.

**Pathogenicity of Fungal Isolate**

A conidial suspension of *M. rileyi* was prepared by scraping the fungal culture into 1.5 mL tubes containing autoclaved Triton X-100 (0.1%) solution (Duksan Pure Chemicals, Ansan, Korea). After disentangling the conidial clumps by vortexing for 3 min, conidia in the suspension were counted using a Neubauer hemocytometer (Marienfeld-Superior, Lauda-Königshofen, Germany) under 40× magnification.

The isolated fungal were fed to L3 larvae of *S. exigua* or *P. xylostella*, or 3.5 cm-length *T. molitor* larvae. A piece (1 × 1 cm) of cabbage containing 1,000 conidia was completely consumed by each larva. Each treatment was replicated 3 times and each replication consisted of 10 insects, in which each Petri dish held a single insect. The Petri dishes were kept in desiccators (4202-0000, Bel-Art Products, Pequannock, NJ, USA) maintaining a constant temperature of 25 ± 1°C and 75 ± 5% RH, following Winston and Bates (32) using a saturated solution of NaCl. Dead insects were counted every 24 h up to 7 days. For topical or injection application, 1,000 conidia per larva were used. Newly (< 30 min) molted fourth instar larvae were considered ‘unsclerotized’ while the larvae at 1 day post molting were considered ‘sclerotized’ cuticles. To remove the epicuticle layer of the integument, the abdominal tergum was rubbed with a brush soaked in acetone for 2 min.
Ca\textsuperscript{2+} Signaling in Response to Fungal Challenge

To observe the Ca\textsuperscript{2+} signal and hemocyte aggregation behavior in response to fungal challenges (33), L5 larvae of *S. exigua* were injected with 2 µL of fungal conidia and hyphae were injected. At selected time points post-injection (PI), hemolymph was collected and fixed on a slide glass by using 2.5% paraformaldehyde. The Fura-positive cells were observed under a fluorescence microscope (DM500, Leica, Wetzlar, Germany) at 400 × magnifications. Fluorescence change over time was analyzed using ImageJ software (https://imagej.nih.gov/ij). Each time point was replicated three times.

Inhibition of Ca\textsuperscript{2+} Signaling

To inhibit Ca\textsuperscript{2+} flux, L5 larvae were co-injected with 1 µL of DEX (10 µg), TTB (1 µg/µL), or DAZ (1 µg/µL) along with Fura-8 (1 mM). After 30 min, 2 µL of fungal conidia and hyphae were injected with TXB\textsubscript{2} (1 µg). After another 10 min, hemolymph was bled and the hemocytes were fixed on a slide glass by using 2.5% paraformaldehyde. Aggregated hemocyte percentage and Fura-8-positive cells were determined as described above.

Fluorescence Labeling of Conidia and Phagocytosis Assay

*M. rileyi* conidia were obtained from the 10-day culture on PDA medium. The conidia were resuspended in a sterile bicarbonate buffer (9.5 mL of 0.2 M Na\textsubscript{2}CO\textsubscript{3} mixed with 41.5 mL of 0.2 M NaHCO\textsubscript{3}, pH 9.4). Then 1 µL of 10 mg/mL FITC fluorescein isothiocyanate (FITC; Sigma-Aldrich Korea) was added to the fungal pellet and placed on a shaker (170 rpm) for 30 min at room temperature under darkness (34). The conidia were washed four times with ice-cold PBS containing 0.02% EDTA and resuspended with 1 mL of TC100 insect medium (Welgen, Gyeongsan, Korea) and stored at -20°C.

For the phagocytosis assay, L5 larvae were surface-sterilized with 70% ethanol and injected with 2 µL of FITC-labeled conidia. At 30, 60, and 90 min PI, the larvae were bled through a clipped proleg onto a slide glass containing 5 µL of ACB. Phagocytic rates were determined by assessing the ratio of 100 hemocytes with or without ingested conidia under a fluorescence microscope (Leica, Wetzlar, Germany).

Nodulation Assay

After surface sterilization with 70% ethanol, L5 larvae were anesthetized with ice and injected with 2 µL of conidial suspension (1×10\textsuperscript{5} conidia/mL) through the inter-segmental membrane between the last two abdominal segments using a Hamilton micro-syringe (Reno, NV, USA) equipped with a 26-gauge needle. After 8 h of incubation at room temperature, the hemocytic nodules were counted by dissecting the larvae under a stereomicroscope (Stemi SV 11, Zeiss, Jena, Germany) at 50 × magnifications. To assess the effects of DEX on nodule formation in larvae infected with *M. rileyi*, larvae were co-injected with 10 µg of DEX along with the conidia. For the time-course experiment, the number of nodules formed was counted at selected time points (1–8 h) PI, 3 larvae/time point.

**RNA Interference**

For RNAi, double-stranded RNAs (dsRNAs) encoding TXA\textsubscript{2} synthase (dsTXAS), SePGE\textsubscript{2}R (dsPGE\textsubscript{2}R), and green fluorescence protein (dsCON) were prepared as described by Vatanparast et al. (38) using Megascript RNAi Kit (Ambion, Austin, TX, USA). dsRNAs were mixed with a transfection reagent Metafectene PRO (Biontex, Plannegg, Germany) at a 1:1 (v/v) ratio and incubated at room temperature for 30 min to form liposomes. dsRNA (1 µg) was injected into L5 larval hemocoe...
with a microsyringe. The RNAi efficiency was evaluated by RT-qPCR at the selected time points. Each treatment was replicated three times using independent RNA preparations.

**PLA2 Activity Determination**

Secretory PLA2 (sPLA2) activity in *S. exigua* larval plasma was determined at three different time points during incubation at 25°C using a commercial assay kit (sPLA2 Assay Kit, Cayman Chemical, Ann Arbor, MI, USA) with diheptanoyl-thio-phosphatidyl choline as the enzyme substrate following Vatanparast et al. (38). Cellular PLA2 (cPLA2) activity measurement in fat body preparations used the same kit, with a different substrate, arachidonyl-thio-phosphatidyl choline. A spectrofluorometer (VICTOR multi-label Plate reader, PerkinElmer, Waltham, MA, USA) was used to measure enzyme activity. Each treatment was replicated with three biologically independent enzyme preparations. Specific enzyme activity (µmol/min/µg) was calculated by dividing absorbance change by the amount of total protein. Protein concentrations were determined following Bradford (39).

**Total Hemocyte Count**

Hemolymph was collected from L5 larvae into ACB by cutting an abdominal proleg and aspirating the exuded hemolymph with glass capillaries (TW100-4, World Precision Instrument, Sarasota, FL, USA). Hemocytes were counted with a hemocytometer (Neubauer improved bright line, Superior Marienfeld, Lauda-Königshofen, Germany) under a phase contrast microscope (BX41, Olympus, Tokyo, Japan). Each treatment was independently replicated three times. To evaluate the effect of TXB2 on THC, test larvae were injected with TXB2 (1 µg per larva). Hemolymph was collected at each min after TXB2 treatment up to 10 min and assessed for THC as described.

**Effect of TXB2 on Hemocyte Micro-Aggregation**

Micro-aggregation is defined as the aggregation of four or more hemocytes. L5 larvae were injected with different PGs and bled at different time points. The hemolymph (20 µL) was mixed with 10 µL of TC100 insect cell culture medium on a cavity well microscope slide (Globe Scientific, Mahwah, NJ, USA) to observe micro-aggregation behavior. Each treatment was replicated three times. To observe hemocyte migration on glass slides, 1 µL of TXB2 (1 µg) was added to 10 µL of hemocyte suspension obtained from naive larvae. The hemocyte behavior was monitored by fluorescence microscopy (Leica, Wetzlar, Germany) at 200×.

**Sample Preparation for TXB2 Analysis Using LC-MS/MS**

Larvae immunity was challenged with 1,000 conidia injected into hemocoels of L5 larvae and incubated at 25°C for 16 h. Control larvae were injected with sterile PBS. Fat body samples were collected into 15 mL tubes from 70 larvae and washed with cold (4°C) PBS. Each sample was homogenized three times (10 min per cycle) in PBS with an ultrasonicator (Bandelin Sonoplus, Berlin, Germany) at 75% power, and subsequently adjusted to pH 4.0 using 1 N HCl. Prostanoids were extracted with 500 µL of ethyl acetate. The combined ethyl acetate extracts were dried under nitrogen to approximately 50 µL and applied to a small silicic acid column (2 × 90 mm containing 30 mg of Type 60A, 100-200 mesh silicic acid, Sigma-Aldrich Korea). Extracts were sequentially eluted with 300 µL of polar solvents starting with 100% ethyl acetate, followed by ethyl acetate: acetonitrile (1:1, v:v), 100% acetonitrile, acetonitrile: methanol (1:1, v:v), and 100% methanol. The acetonitrile:methanol fraction was used to quantify TXB2. Each treatment was replicated three times using independent sample collections.

**LC-MS/MS Analysis**

LC-MS/MS was performed using a QTrap 4500 (AB Sciex, Framingham, MA, USA) equipped with an autosampler, a binary pump, and a column oven. The analytical column was an Osaka Soda C18 column (2.1 mm x 150 mm, 2.7 µm) maintained at 40°C (Osaka, Japan). The mobile phases consisted of 0.1% formic acid in water (A) and 0.1% formic acid in acetonitrile (B). The linear gradient was as follows: 30% B at 0 min, 30% B at 2 min, 65% B at 12 min, 95% B at 12.5 min, 95% B at 25.0 min, 30% B at 28.0 min, and 30% B at 30 min. The flow rate was 0.40 mL/min. The autosampler was set at 5°C and the injection volume was 10 µL. The LC-MS/MS and LC-MS/MS was equipped with an electrospray ionization (ESI) source. ESI was performed in negative ion mode. After optimization, the source parameters were: source temperature at 400°C, curtain gas flow rate at 32 µL/min, ion source gas flow rate at 60 µL/min, and spray voltage adjusted to ~4,500 V. Analyses were performed in Multiple Reaction Monitoring (MRM) detection mode using nitrogen as the collision gas. Peak detection, integration, and quantitative analysis were done using MassView1.1 software (AB Scxiex).

**Statistical Analysis**

The data for continuous dependent variables were subjected to one-way analysis of variance (ANOVA) using PROG GLM in the SAS program (40). Mortality assay by the leaf dipping method was analyzed by repeated measure ANOVA. All experiments were conducted in three biologically independent replicates and the means ± standard errors (SE) were plotted using Sigma Plot (Systat Software, Point Richmond, CA, USA). The means were compared with the least significant difference (LSD) test at a Type I error of 0.05.

**RESULTS**

**Identification and Pathogenicity of Fungal Isolates**

Fungal spore colonies on the PDA medium began to grow slowly with a white velvety appearance (early stage) and irregular borders at 4 days, then turned to pale green or malachite green with sporulation at 7 days (Figure S1A). The vegetative and reproductive structures developed hyaline hyphae and conidiophores with smooth and septate walls. The branches, which formed near the septa, developed in clusters on the same point with 2~4 phalids exhibiting a short, rounded, and
considerable variation in the fungal virulence (lethal median time (LT50) was much faster against 100%) clustered with another formation following fungal infection (Figure 1B) suppressed phagocytosis ((spores with DEX led to substantially increased mortality increased in newly molted larvae, before cuticular sclerotization (increased in newly molted larvae, before cuticular sclerotization (64% following spore injections (Figure 1A). M. rileyi virulence increased in newly molted larvae, before cuticular sclerotization (Figure S2A) or to integument treated with an organic solvent to remove the epicuticular layer (Figure S2B). Co-injections of spores with DEX led to substantially increased mortality (Figure 1A). The DEX treatment led to significantly suppressed phagocytosis (Figure 1B) and hemocyte nodule formation following fungal infection (Figure 1C). sPLA2 and iPLA2 activities significantly increased within 5 min post-fungal infection (PFI; Figure 1D), with parallel increases in mRNAs encoding sPLA2 and iPLA2B, but not iPLA2A (Figure 1E).

Eicosanoids Induce Intracellular Ca2+ Mobilization
Hemolymph collected from M. rileyi-challenged larvae had labeled spores and hyphae connected to hemocytes (Figure 2A). Proportions of Fura-positive hemocytes increased with time, up to 10 min PFI, in correlation (r = 0.943; P < 0.0001) with numbers of hemocyte aggregates (Figure 2B). Fura-staining intensity also increased with time up to 10 min. Treating S. exigua larvae with the PLA2 inhibitor, DEX, led to reduced proportions of Fura-positive hemocytes and aggregated hemocytes (Figure 2C).

Prostanoids Influence Ca2+ Mobilization and Hemocyte Microaggregation
Treating larvae with a nonspecific cyclooxygenase inhibitor, NAP, prior to the conidial challenge led to substantially suppressed proportions of Fura-positive hemocytes (down by about 40%) and hemocyte microaggregates (down by about 35%), both of which were reversed in larvae co-treated with NAP+PGF2 (Figure 3A). Treatment with the lipooxygenase inhibitor ESC led to small but statistically significant reductions in both parameters, with Fura-positive proportions down by about 8% and aggregated hemocytes down by about 24%. Treating larvae with DEX led to reductions in proportions of Fura-positive hemocytes (down by about 20%) and aggregated hemocytes (down by about 50%). Figure 3B shows the influences of selected prostanoids on possible reversals of DEX treatments before the conidial challenge. Co-injecting DEX along with one of three PGs led to increases in both parameters (Figures S3A–E). PGE2 treatment led to increased Fura-positive hemocyte proportions by about 56%; PGD2 treatment increased by about 50%; PGI2 treatment did not reverse the DEX effect. TXA2. TXB2 treatments led to very high proportions of Fura-positive hemocytes, up to just over 60% for TXA2 and nearly 80% for TXB2. Parallel experiments with proportions of aggregated hemocytes led to similar results. Figure 3C shows the outcomes of treating larvae with specific inhibitors of thromboxane synthesis, DAZ, and a thromboxane receptor antagonist, TTB. Compared to controls, co-injecting conidia+DAZ and, in a separate group of experimental larvae, conidia+TTB led to significant decreases in proportions of Fura-positive hemocytes (down by about 10% after DAZ treatment and by about 20% after TTB treatment) and aggregated hemocytes (down by about 20% after DAZ treatment and by about 35% after TTB treatment).

Figure 4A outlines the biosynthesis of TXA2/TXB2 from AA in S. exigua. Two POXs, SePOX-F and SePOX-H, convert AA into PGH2, which is converted into TXA2. TXA2 can convert into TXB2 spontaneously. Figure 4B shows that the M. rileyi challenge did not influence the time-dependent accumulation of mRNAs encoding Se-PG-XO-A (upper left), while accumulations of mRNAs encoding Se-PX-F and SePOX-H were significantly elevated beginning 16 h PFI (upper right and lower left panels). Figure 4B (lower right) shows that the accumulation of mRNAs encoding Se-TXAS, the enzyme that converts AA into TXA2, was also significantly elevated at 16 h PFI.

We estimated concentrations of TXB2 in fat body (1S larvae) by LC-MS/MS. Chemical identification of TXB2 was confirmed by two specific ion peaks, which matched the ion peaks in an authentic TXB2 chemical standard (Figure 5A). TXB2 was detected at 0.28 ± 0.02 ng/g in naive larvae, which was significantly increased to 0.61 ± 0.05 ng/g in M. rileyi-challenged larvae (Figure 5A). Se-TXAS expression was silenced by injecting dsRNA specific to the gene into experimental larvae (Figure 5B). Injecting M. rileyi into dsRNA-treated larvae led to significantly enhanced pathogenicity, which was reversed in larvae co-injected with conidia+TXB2 (Figure 5C).

The Influence of TXB2 on Hemocyte Behavior
Figure 6A reports the influence of TXB2 injections (1 μg/larva) on total hemocyte numbers over 10 h PI. The blue line represents DMSO-treated control larvae and the red line shows total hemocyte counts increase over time PI. Figure 6B shows reports that DEX injections led to no visible changes in DIC, nor Ca2+ signaling. Co-injecting DEX+TXB2, visible DIC and Fura staining. Figure 6C shows that separate treatments with the indicated prostanoids led to substantial increases in proportions of microaggregated hemocytes with the exception of PGI2.
treatments, which did not influence microaggregation. The thromboxane injections led to statistically significant higher proportions of microaggregated hemocytes, compared to controls and the three PG treatments. Figure 6D reports proportions of microaggregated hemocytes steadily increase as a function of time over 60 min after TXB2 injections to a maximal level at about 60% of total hemocytes. This stimulatory activity of TXB2 exhibited a dose-dependency.

Thromboxanes Influence Hemocyte Behavior via a Specific Receptor

Within the S. exigua genome there are 37 predicted GPCR candidates (Table S3), which were used to be compared with mammalian PG receptors. In particular, Figure 7A shows that a S. exigua PGE2 receptor (Se-PGE2R) is clustered with human, house mouse, and zebrafish thromboxane receptors. Injecting a dsRNA construct designed to the SePGE2R sequence led to severe reductions in mRNAs encoding Se-PGE2R, down by about 80% (Figure 7B). Figure 7C shows that silencing Se-PGE2R led to reduced accumulations of mRNAs encoding PGE2R and reduced Ca2+ signaling, seen in microphotographs as severe reductions in Fura-positive hemocytes and the accompanying histogram. Co-injections of dsPGE2R+TXA2 and, separately, dsPGE2R+TXB2, did not restore FURA staining. Similar images and the accompanying histogram document the same outcomes with proportions of microaggregated hemocytes.

Thromboxane Up-regulates Ca2+ via Ca2+-Induced Ca2+ Release Pathway

The signaling pathway to up-regulate Ca2+ signal in hemocytes in response to TXB2 was monitored by treating larvae with selected compounds that influence Ca2+ signaling in mammalian cells (Figure 8). The micrographs in Figure 8A show that phospholipase C inhibitor (‘U-73122’), IP3 receptor inhibitor (2-APB), and ryanodine receptor inhibitor (‘DAN’) significantly suppressed Ca2+ signaling and hemocyte aggregation following the M. rileyi challenge. The accompanying histograms show the quantitative values. In contrast, a sarco/endoplasmic reticulum Ca2+-ATPase (SERCA) inhibitor (‘TPG’) did not suppress the
Ca\textsuperscript{2+} signal. The micrographs in Figure 8B and their accompanying histograms similarly show that co-injections with TXB\textsubscript{2} did not reverse the influences of the drug treatments.

**DISCUSSION**

Entomopathogenic fungi are effective biological control agents deployed in a range of ecological settings and they are used in bioremediating environmental toxins (41). Their market values are rapidly growing (42), in parallel with their use. Nonetheless, the understanding insect physiological and molecular mechanisms of host defense against invading fungi remains incomplete. Here, we contribute a new understanding of the biochemical signaling mechanisms responsible for host cellular immune responses to infections by an entomopathogen, *M. rileyi*, which is known to be a pathogenic fungus to *S. exigua* (17). Specifically, we identified two eicosanoids, the prostanoids TXA\textsubscript{2} and TXB\textsubscript{2}, as key signals responsible for hemocyte migration to infection foci, phagocytosis of fungal conidia, and hemocyte microaggregation reactions to infection.

The entomopathogenicity of *M. rileyi* was enhanced by the addition of DEX, a pharmaceutical PLA\textsubscript{2} inhibitor. PLA\textsubscript{2} is the first step in the biosynthesis of all eicosanoids and inhibiting this enzyme effectively eliminates eicosanoid signaling. Here, DEX treatments led to increased mortality and decreased phagocytosis, nodulation, hemocyte aggregation, activity of sPLA\textsubscript{2} and cPLA\textsubscript{2}, and intracellular Ca\textsuperscript{2+} signaling. We infer eicosanoids, particularly TXA\textsubscript{2} and TXB\textsubscript{2}, are central actors in the immune response to fungal infections.
Among different eicosanoids, PGs mediated the Ca$^{2+}$ signal in response to the fungal infection. Inhibiting the lipoxygenase pathways with ESC treatments did not influence Ca$^{2+}$ signaling nor hemocyte aggregation, while separate experiments with the cyclooxygenase inhibitor naproxen (NAP) led to sharp reductions in both parameters, with Ca$^{2+}$ signaling down by about 30% and aggregation down by about 45%. The reductions were reversed following co-injection of NAP+PGE$_2$. Hence, PGs, but not lipoxygenase products, operate in the two hemocyte defense parameters. We found that separate co-injections of DEX + two PGs, PGE$_2$ and PGD$_2$, but not DEX+PGI$_2$, reversed the DEX inhibitory effects on Ca$^{2+}$ signaling and hemocyte aggregation, to some extent. DEX+TXA$_2$ and DEX+TXB$_2$ treatments led to steep increases in Ca$^{2+}$ signaling, higher than controls, while separate DEX+TXB$_2$ co-injections returned hemocyte aggregations to control levels. Experiments with two thromboxane-specific compounds, DAZ and TTB, emphasize the point. DAZ specifically inhibits TXB biosynthesis and TTB is a TBX receptor antagonist. Treatments with these compounds led to significantly reduced
Ca²⁺ signaling and hemocyte aggregation. Our interpretation is that prostanoids, particularly the two thromboxanes, mediate these hemocytic reactions to fungal infections.

Physiological activities take place over time and analysis of the influence of time reveals subtle aspects of thromboxane signaling in insect responses to the fungal challenge. Fungal infections led to increased phagocytosis, recorded as about 15% at 30 min PFI, which increased to 22% by 60 min, and approximately doubled over the next hour. We infer that phagocytosis accelerated over the 3 h time frame. Nodulation reactions to infection were undetectable until 3 h PFI, then increased from about 2 nodules/larva to nearly 30/larvae over the next 6 h, increasing in a non-linear manner. Accumulation of mRNAs encoding PLA₂ took place on a different time-scale, with increases recorded from 3 to 10 min PFI. Two cellular parameters, Ca²⁺ signaling (recorded as changes in Fura intensities) and aggregated hemocytes, increased over the first 10 min PFI, then declined significantly over the next 20 min. This is also apparent in micrographs taken over the same time frames. We note, also, that hemocyte microaggregation proportions steadily increased with time for 60 min PI. Our broad point is that insect immune reactions take place over time, and our appreciation of the time dimension enriches our understanding of immunity.

Research with eicosanoids is challenging because these compounds are produced and operate in very small amounts. Physiological quantities of PGs and other eicosanoids are often determined indirectly using radioimmunoassays and bioassays. Sensitive mass spectrometry now enables accurate direct determinations of eicosanoid quantities at physiological levels. Our data show TXB₂ occurred in the fat body of naïve larvae at approximately 0.28 ng/g, which increased by 2.2-fold to about...
0.61 ng/g at 16 h PFI. This chemical determination strongly bolsters our view that TBX2 is present and operates in larval tissues. Injecting a dsRNA construct specific to SeTXAS led to reduced accumulations of mRNAs encoding SeTXAS at 24, 48, and 72 h PI. Treatments with the gene silencing construct led to increased larval mortality at 5 days PFI, which was significantly, albeit not completely, reversed in larvae co-injected with dsTXAS+TXB2.

One eicosanoid action in insect cellular immunity is the activation of Ca²⁺ signaling within hemocytes in mealworms, Tenebrio molitor. Roy and Kim (33) reported that bacterial infections stimulate Ca²⁺ activation, recorded as Fura-positive cells. One role of Ca²⁺ is activating hemocyte spread by F-actin extension, documented by co-localization of Ca²⁺ and F-actin. In this report, we consider Ca²⁺ mobilization in more detail by applying selected pharmaceuticals. U-73122 is a phospholipase C inhibitor that inhibits Ca²⁺ release from endoplasmic reticulum (ER) stores (43). 2-APB influences a wide range of channels, including Ca²⁺ channels, possibly in an indirect manner by cytoplasmic acidification (44). DAN inhibits ryanodine receptors (45), a class of intracellular Ca²⁺ channels located in ER and responsible for releasing Ca²⁺ from intracellular stores (46). SERCA operates in Ca²⁺ uptake by transferring Ca²⁺ from the cell cytoplasm into the lumen of the sarcoplasmic reticulum or endoplasmic reticulum. TPG treatments did not influence intracellular Ca²⁺ signaling, nor hemocyte aggregation.

Upon fungal infection, hemocytes physically attach to the fungal hyphae or conidia. The hemocytes aggregating around the fungi exhibited intensive Ca²⁺ signaling. The increased Ca²⁺ signal was positively associated with hemocyte aggregation behavior. However, DEX treatment suppressed the Ca²⁺ signal and inhibited hemocyte aggregation. This inhibition was rescued by adding AA. We infer that eicosanoids induce Ca²⁺ signaling to activate hemocyte aggregation. Ca²⁺ is required for hemocyte behavior. In M. sexta, plasmacytoid endoparasitoid wasp against D. melanogaster encodes a Ca²⁺ blocker mimicking SERCA to shut down Ca²⁺ bursts, which results in the host immunosuppression (48). Among eicosanoids, thromboxane treatments (TXA₂ and TXB₂) highly activated the intracellular Ca²⁺ signal, which led to hemocyte aggregation in response to fungal infection.
Among other PGs known in *S. exigua*, PGD₂ and PGE₂ also activated the hemocyte behavior while PGI₂ did not because it acts as an anti-inflammatory mediator (49). TXA₂ mediates blood clotting in mammals by reducing blood flow to the site of a clot through vasoconstriction and by aggregating platelets to the site (13). In insects, the wound healing process along with coagulation factors is mediated by eicosanoids, shown by using a PLA₂-mutant line of *Drosophila* (15). This study shows increased TXB₂ titer in response to the fungal infection. We also observed that the total hemocyte numbers in the hemolymph and their migratory behavior were increased by TXB₂. We infer that thromboxanes contribute to cellular immune responses, which include the wound healing process by stimulating hemocyte aggregation to the infection foci.

Terutroban has been used to inhibit the mammalian thromboxane receptor (TP) (50). It inhibits TXA₂ or TXB₂ action to mediate the Ca²⁺ signal and hemocyte aggregation in *S. exigua*. This suggested that the thromboxane actions are mediated through TP-like receptors in insects. In humans, TP receptors exist in two alternative splicing variants, TPα and TPβ, in which TPα is the dominant isoform translated in platelets and vascular cells, and the TPβ isoform is present in vascular smooth muscle cells (51, 52). TPs

![FIGURE 6](image-url)
are classified into seven transmembrane GPCRs. Despite no TP ortholog in *S. exigua* genome, a PG receptor, *Se*-PGE$_2$R, is required for TXB$_2$ to mediate the Ca$^{2+}$ signal and immune responses. Like mammals, insects produce three groups of prostanoids, PGs, prostacyclin, and thromboxane (22, 26, 49). Although different prostanoids act via specific GPCRs in mammals, there are multiple receptors, cross-reactivity, and cross-talks for each prostanoid, in which PGE$_2$ is the most versatile prostanoid because of four different receptor subtypes (53). Two PG receptors were known in *S. exigua*. The first PGE$_2$ is specific to oenocytoid hemocytes and induces the hemocyte cell lysis to release prophenoloxidase for melanization during cellular immune responses (54). The second receptor (*Se-hcPGPCRI*) is expressed in various cell types and mediates immune and reproductive processes (26). The latter *SePGE$_2$R* is closely associated with the mammalian TPs in phylogenetic analysis, and its RNAi treatment prevented the immunological functions of thromboxanes in *S. exigua*. These suggest that *SePGE$_2$R* is shared by two prostanoids. This kind of receptor-ligand interaction is explained by a functional pleiotropy in PRXamide neuropeptides and their receptors, exhibiting differential binding affinities (55). Based on this pleiotropism, differential binding affinities of PGE$_2$
and TXB₂ to common PG receptors may form orchestrated multi-organ physiological outcomes. This hypothesis will be tested through receptor-ligand binding assays.

This study demonstrates the significant role of thromboxanes in mediating hemocyte aggregation upon infection foci of the fungal conidia in insects. Our working model of thromboxane actions at the cellular level is depicted in Figure 9. We expect to refine our model as new details emerge of continued research. Initially, the chemokine-like role of eicosanoids was introduced in another lepidopteran insect, Manduca sexta, in which hemocyte migration...
to the infection foci was inhibited by DEX treatment (18). Our current study suggests that thromboxanes act as chemokine-like factors to mediate hemocyte migration to infection foci during wound healing or other cellular immune responses in insects.

 DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material.

 AUTHOR CONTRIBUTIONS

MCR and YK carried out the experiment. MCR, DS, and YK wrote the manuscript with support from KN and JK. YK conceived the original idea. YK supervised the project. All authors contributed to the article and approved the submitted version.

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 SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2021.791319/full#supplementary-material

REFERENCES

1. Lemaitre B, Hoffmann J. The Host Defense of Drosophila Melanogaster. Annu Rev Immunol (2007) 25:697–743. doi: 10.1146/annurev.immunol.25.022106.141615
2. Kwon H, Bang K, Cho S. Characterization of the Hemocytes in Larvae of Protaetia Brevitarsis Seoulensis: Involvement of Granulocyte-Mediated Phagocytosis. PLoS One (2014) 9:e103620. doi: 10.1371/journal.pone.0103620
3. Lee J, Hwang S, Cho S. Immune Tolerance to an Intestine-Adapted Bacterium, Chryseobacterium Sp., Injected Into the Hemocoel of Protaetia Brevitarsis Seoulensis. Sci Rep (2016) 6:31722. doi: 10.1038/srep31722
4. Kim H, Choi D, Jung J. Kim Y. Eicosanoid Mediation of Immune Responses at Early Bacterial Infection Stage and its Inhibition by Photuridinae Temperata Subsp. Temperata, an Entomopathogenic Bacterium. Arch Insect Biochem Physiol (2018) 99:e21502. doi: 10.1002/arch.21502
5. Shrestha S, Kim Y. Various Eicosanoids Modulate the Cellular and Humoral Immune Responses of the Beetle Armyworm, Spodoptera Exigua. Biosci Biotechnol Biochem (2009) 73:2077–84. doi: 10.1271/bbb.90272
6. Ishii K, Hamamoto H, Kamimura M, Nakamura Y, Noda H, Imamura K, et al. Insect Cytokine Paralytic Peptide (PP) Induces Cellular and Humoral Immune Responses in the Silkworm Bombyx Mori. J Biol Chem (2010) 285:28635–42. doi: 10.1074/jbc.M110.138446
7. Kim Y, Ahmed S, Stanley D, An C. Eicosanoid-Mediated Immunity in Insects. Dev Comp Immunol (2018) 83:130–43. doi: 10.1016/j.dci.2017.12.005
8. Stanley D, Kim Y. Eicosanoid Signaling in Insects: From Discovery to Plant Protection. Crit Rev Plant Sci (2014) 33:20–63. doi: 10.1080/07352689.2014.847631
9. Hasan MA, Ahmed S, Kim Y. Biosynthetic Pathway of Arachidonic Acid in Spodoptera Exigua in Response to Bacterial Challenge. Insect Biochem Mol Biol (2019) 111:103179. doi: 10.1016/j.ibmb.2019.103179

FIGURE 9 | Intracellular immune signaling in S. exigua hemocytes following fungal infection. Fungal hyphae or conidia are recognized by pattern recognition receptor (‘PRR’), which activates the Toll pathway to increase PLA2 activity. The activated PLA2 releases linoleic acid (‘LA’), which is desaturated and elongated into arachidonic acid (‘AA’). AA is then oxygenated by two peroxinectins (‘POX F/H’) into PGH2, which is isomerized into TXA2/TXB2 by TXA2 synthase (‘TXAS’). TXA2/TXB2 is transferred out of the cell to bind with its GPCR (‘PGE2R’) and activate phospholipase C (‘PLC’). PLC increases intracellular inositol triphosphate (‘IP3’) concentrations which binds to its receptor (‘IP3R’) on endoplasmic reticulum, thereby releasing Ca2+ into the cytoplasm Ca2+ triggers calcium-induced calcium release (‘CICR’) from a ryanodine receptor (‘RyR’). The free Ca2+ is a secondary signal to activate small G proteins for actin polymerization to facilitate hemocyte behavior.
10. Kim Y, Stanley D. Eicosanoid Signaling in Insect Immunology: New Genes and Unresolved Issues. Genes (2021) 12:211. doi: 10.3390/genes12020211
11. Feletou M, Huada Y, Varshney PK. Endothelin-Mediated Control of Vascular Tone: COX-1 and COX-2 Products. Br J Pharmacol (2011) 164:894–912. doi: 10.1111/j.1476-5381.2011.01276.x
12. Naramiya S, Sugimoto Y, Ushikubi F. Prostanoid Receptors: Structures, Properties, and Functions. Physiol Rev (1999) 79:1193–226. doi: 10.1152/physrev.1999.79.4.1193
13. Braune S, Kipper JH, Jung F. Effect of Prostanoids on Human Platelet Function: An Overview. Int J Mol Sci (2020) 21:9020. doi: 10.3390/ijms21239020
14. Theopold U, Krautz R, Dushay MS. The Oenocytoids in the Beet Armyworm, Spodoptera Exigua, – Intraspecific Restriction Fragment Length Polymorphism in the Fundus Applied Nematology Group. Fund Appl Nematol (1992) 15:563–74.
15. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. Mol Biol Evol (2013) 30:2725–9. doi: 10.1093/molbev/ms3197
16. Winston PW, Bates DH. Saturated Solutions for the Control of Humidity in Biological Research. Ecology (1960) 41:232–7. doi: 10.2307/1931961
17. Roy MC, Kim Y. Eicosanoid-Induced Calcium Signaling Mediates Cellular Immune Responses of Tenebrio Molitor. Entomol Exp Appl (2021) 169:888–98. doi: 10.1111/eea.13037
18. Rohloff LH, Wiesner A, Gitz P. A Fluorescence Assay Demonstrating Stimulation of Phagocytosis by Haemolymph Molecules of Galleria Mellonella. J Insect Physiol (1994) 40:1045–9. doi: 10.1016/0022-1910(94)90057-4
19. Vatanparast MD, Ahmed S, Lee DH, Hwang SH, Hammock B, Kim Y. EpOMEs Act as Immune Suppressors in a Lepidopteran Insect, Spodoptera Exigua. Sci Rep (2020) 10:20183. doi: 10.1038/s41598-020-77325-2
20. Bustin SA, Benes V, Garson JA, Hellemans J, Huggett J, Kubista M, et al. The MIQE Guidelines: Minimum Information for Publication of Quantitative Real-Time PCR Experiments. Clin Chem (2009) 55:611–22. doi: 10.1373/clinchem.2008.112797
21. Livak KJ, Schmittgen TD. Analysis of Relative Gene Expression Data Using Real Time Quantitative PCR and the 2−ΔΔCT Method. Methods (2001) 25:402–8. doi: 10.1086/meth.2001.1262
22. Vatanparast MD, Ahmed S, Herrero S, Kim Y. A Non-Venomous Spla2 of a Lepidopteran Insect: Its Physiological Functions in Development and Immunity. Dev Comp Immunol (2018) 89:83–92. doi: 10.1016/j.devimm.2018.08.008
23. Bradford MM. A Rapid and Sensitive Method for the Quantification of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. Anal Biochem (1976) 72:248–54. doi: 10.1016/0003-2697(76)90527-3
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