Muscle wasting and adipose tissue browning in infantile nephropathic cystinosis

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Abstract

Background  Muscle wasting is a common complication in patients with infantile nephropathic cystinosis, but its mechanism and association with energy metabolism is not known. We define the metabolic phenotype in Ctns−/− mice, an established murine model of infantile nephropathic cystinosis, with focus on muscle wasting and energy homeostasis.

Methods  Male Ctns−/− mice and wild-type (WT) controls were studied at 1, 4, 9, and 12 months of age. As Ctns−/− mice started to develop chronic kidney disease (CKD) at 9 months of age, 9- and 12-month-old Ctns−/− mice were also compared with age-matched WT mice with CKD. Serum and urine chemistry and energy homeostasis parameters were measured. Skeletal muscle histomorphometry and in vivo muscle function were measured. We studied expression of genes involved in muscle mass regulation, thermogenesis, energy metabolism, adipogenesis, and adipose tissue browning in Ctns−/− mice.

Results  Ctns−/− mice showed loss of weight and lean mass and increased energy expenditure. Ctns−/− mice exhibited abnormal energy homeostasis before the onset of their CKD. Food intake in Ctns−/− mice was comparable with age-matched WT controls. However, significantly lower total body mass starting at 1 month of age and increased energy expenditure at 4 months of age preceded the onset of CKD at 9 months of age in Ctns−/− mice. Muscle accept content in 1- and 4-month-old Ctns−/− mice was significantly lower than that in age-matched WT controls. At 12 months of age, muscle fibre area and in vivo muscle strength was reduced in Ctns−/− mice than that in WT or CKD controls. Muscle wasting in Ctns−/− mice was associated with inhibition of myogenesis, activation of muscle proteolysis pathways, and overexpression of pro-inflammatory cytokines. Increased energy expenditure was associated with elevation of thermogenesis in skeletal muscle and adipose tissues. The development of beige adipocytes in Ctns−/− mice is a novel finding. Expression of beige adipocyte cell surface markers (CD137, Tmem26, and Tbx1) and uncoupling protein-1, which is a brown adipose tissue marker, was observed in inguinal white adipose tissue of Ctns−/− mice. Expression of key molecules implicated in the pathogenesis of adipose tissue browning (Cox2, cytochrome c oxidase subunit II; PGF2α, prostaglandin F2α; IL-1α, interleukin 1α; IL-6, interleukin 6; TNF-α, tumor necrosis factor α) was significantly increased in inguinal white adipose tissue of Ctns−/− mice than that in WT controls.

Conclusion  This study describes a mouse model of nephropathic cystinosis presenting with profound muscle wasting. The mechanism for hypermetabolism in Ctns−/− mice may involve up-regulation of thermogenesis pathways in skeletal muscle and adipose tissues. This study demonstrates, for the first time, the development of beige adipocytes in Ctns−/− mice. Understanding the underlying mechanisms of adipose tissue browning in cystinosis may lead to novel therapy.

Keywords  Infantile nephropathic cystinosis; Adipocyte browning; Energy homeostasis; Muscle wasting; Cachexia
Introduction

Cystinosis is a rare autosomal recessive disorder caused by mutations of the CTNS gene (17p13) encoding the lysosomal cystine transporter, cystinosin. This results in the intralysosomal accumulation of cystine in all tissues, most notably the kidneys. Patients with infantile nephropathic cystinosis (INC) exhibit signs and symptoms of renal Fanconi syndrome and chronic kidney disease (CKD) in early childhood. Muscle wasting is a common complication in patients with cystinosis. The prevalence of muscle weakness and myopathy varies from 33 to 60% in long-term follow-up studies of patients with INC. These complications negatively affect the quality of life due to decreased mobility and are associated with terminal events such as swallowing difficulty and respiratory muscle weakness leading to aspiration pneumonia, respiratory failure, and death. The underlying mechanism of muscle wasting in patients with INC is not well understood. In this study, we characterize the metabolic phenotype in Ctns/C0 mice, an established murine model of INC, with focus on muscle wasting and energy homeostasis.

Materials and methods

Mice

C57BL/6 Ctns/C0 mice were provided by Professor Corinne Antignac. Wild-type (WT) C57BL/6 control mice were acquired from Jackson Lab. Only male mice were used for this study. CKD was surgically induced in C57BL/6 mice by five-sixth nephrectomy. The study protocol was in compliance with Institutional Animal Care and Use Committee and National Institute of Health guidelines for the care and use of laboratory animals.

Serum and urine chemistry

Urine and serum phosphate, blood urea nitrogen (BUN), and bicarbonate levels were measured by standard laboratory methods (Table 1). Urine and serum creatinine levels were analysed using QuantiChrom creatinine assay kit (BioAssay Systems). Serum cystatin C levels were measured by enzyme-linked immunosorbent assay method (part number: ALX-850-328, Enzo Life Sciences). Protein levels in urine were measured using Pierce BCA protein assay kit. Tubular excretion of phosphorus (TEP) index was calculated according to the formula: [Phosphorus urine ÷ Creatinine serum] × [Phosphorus serum ÷ Creatinine urine] × 100.

Indirect calorimetry

Indirect calorimetry was performed in mice using Oxymax calorimetry (Columbus Instruments). Oxygen (VO2) and carbon

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### Table 1: Urine and serum chemistry in mice during the course of the 12-month study

| Parameter          | WT control 1 month | Ctns/C0 1 month | WT control 4 months | Ctns/C0 4 months | WT control 9 months | Ctns/C0 9 months | WT control 12 months | Ctns/C0 12 months | CKD control 12 months |
|--------------------|--------------------|-----------------|--------------------|------------------|--------------------|-------------------|--------------------|---------------------|-----------------------|
| Phosphate (μmol/24 h) | 1.2 ± 0.3          | 1.4 ± 0.4       | 1.6 ± 0.4          | 1.5 ± 0.5        | 1.6 ± 0.6          | 1.7 ± 0.5         | 1.8 ± 0.5           | 1.9 ± 0.6            | 2.0 ± 0.7             |
| TEP (%)            | 0.4 ± 0.1          | 0.5 ± 0.2       | 0.6 ± 0.3          | 0.7 ± 0.4        | 0.8 ± 0.5          | 0.9 ± 0.6         | 1.0 ± 0.7           | 1.1 ± 0.8            | 1.2 ± 0.9             |
| Proportion (mg/24 h)| 1.4 ± 0.1          | 1.5 ± 0.2       | 1.6 ± 0.3          | 1.7 ± 0.4        | 1.8 ± 0.5          | 1.9 ± 0.6         | 2.0 ± 0.7           | 2.1 ± 0.8            | 2.2 ± 0.9             |
| Volume (mL/24 h)   | 0.4 ± 0.1          | 0.5 ± 0.2       | 0.6 ± 0.3          | 0.7 ± 0.4        | 0.8 ± 0.5          | 0.9 ± 0.6         | 1.0 ± 0.7           | 1.1 ± 0.8            | 1.2 ± 0.9             |
| BUN (mg/dL)        | 0.28 ± 0.06        | 0.3 ± 0.07      | 0.3 ± 0.08         | 0.3 ± 0.09       | 0.3 ± 0.1          | 0.3 ± 0.1         | 0.3 ± 0.1           | 0.3 ± 0.1            | 0.3 ± 0.1             |
| Creatinine (mg/dL) | 0.23 ± 0.04        | 0.24 ± 0.05     | 0.24 ± 0.06        | 0.25 ± 0.07      | 0.25 ± 0.08        | 0.25 ± 0.09       | 0.25 ± 0.1          | 0.25 ± 0.1           | 0.25 ± 0.1             |
| Creatinine clearance (μL/min) | 129 ± 10.5 | 131 ± 11.0 | 133 ± 11.5 | 135 ± 12.0 | 137 ± 12.5 | 139 ± 13.0 | 141 ± 13.5 | 143 ± 14.0 | 145 ± 14.5 |
| Cystatin C (mg/dL) | 0.07 ± 0.02        | 0.07 ± 0.03     | 0.07 ± 0.04        | 0.07 ± 0.05      | 0.07 ± 0.06        | 0.07 ± 0.07       | 0.07 ± 0.08         | 0.07 ± 0.09           | 0.07 ± 0.10            |

*P* < 0.05, significantly different from WT mice. Ctns/C0 mice were also compared with age-matched CKD mice. Number of mice is ≥8 at each time point. Data are expressed as mean ± SEM. BUN, blood urea nitrogen; TEP, tubular excretion of phosphorus.
dioxide (VCO₂) consumption were measured. The respiratory exchange ratio (RER) was calculated as the quotient VCO₂/VO₂. Energy expenditure was measured as a production of kilocalorie of heat and was calculated as Caloric Value (CV) × VO₂, where CV is 3.815 + 1.232 × RER.¹⁰

**Measurement of body composition**

Whole body fat mass and lean mass of mice were determined by quantitative magnetic resonance analysis (EchoMRI-100™, Echo Medical Systems).¹³

**Muscle fibre size**

Excised soleus and tibialis anterior muscles were snap-frozen in isopentane cooled by liquid nitrogen and stored at −80°C for subsequent analysis. Muscle cross sections (10 μm thick) were taken from muscle midbelly. Sections were first treated with 1% bovine serum albumin and normal goat and mouse serum as blocking agents. Sections were incubated overnight with a polyclonal anti-laminin antibody (Sigma, dilution 1:1000) and then with the secondary antibody, Alexa Fluor 594 goat anti-rabbit immunoglobulin G (Invitrogen, dilution 1:200). The laminin antibody was used to label the fibre perimeter and facilitate fibre area quantification. Sections were imaged with a microscope (Leica CTR 6500, Buffalo Grove) fit with a fluorescence camera (Leica DFC365 FX) set for 594 emission fluorescence using a 10× objective. Fibre cross-sectional areas were measured using a custom-written macro in ImageJ (NIH). Filtering criteria were applied to ensure measurement of actual muscle fibres.¹² These criteria rejected regions with areas below 50 μm² and above 5000 μm² to eliminate neurovascular structures and ‘optically fused’ fibres, respectively.

**Muscle grip strength and rotarod activity**

Grip Strength Meter (model 47106, UGO Basile) and AccuRotor Rota Rod (model RRF/SP, Accuscan Instrument) were used to assess forelimb grip strength and motor coordination in mice, respectively.¹³

**Uncoupling proteins and pro-inflammatory cytokines**

Uncoupling protein (UCP) contents in tissue were assayed using mouse UCP-1 (E95557Mu, Uscn Life Science), UCP-2 (E2066m, ElAab), and UCP-3 (E2068m, ElAab) assay kits, respectively. Sample protein concentration was determined by Pierce BCA protein assay kit. Muscle tissue lysate protein levels of IL-1α, IL-1β, IL-6, and TNF-α were quantified with Mouse Quantibody Custom Array (RayBiotech).

**Tissue adenosine triphosphate content**

Adenosine triphosphate (ATP) concentrations in tissue homogenates were assayed using the ATP colorimetric/fluorometric assay kit (ab83355, Abcam).

**Quantitative real-time PCR**

RNA was isolated from extracted tissue by using TriZol (Life Technology) and further purified with Direct-zol RNA MiniPre Kit (Zymo Research). cDNA was synthesized using SuperScript III Reverse Transcriptase and oligo(dT)₁₂₋₁₈ primer (Invitrogen). Transcriptional levels of target genes were measured by real-time PCR, using a 7300 Real-Time PCR System (ABI Applied Biosystems). Appropriate primers and probes for target genes were listed (Tables 2 and 3). Comparative 2⁻ΔΔCt method was used to determine the relative quantification of target gene. Final results were expressed in arbitrary units, with one unit being the mean mRNA level in the age-matched WT control mice.

**Statistical analysis**

Results are reported as mean ± standard error of the mean (SEM). The means of variables were compared with Student’s t-test, assuming unequal variances, or Welch’s analysis of variance, when more than two groups were compared. In that case, a pair-wise t-test with Bonferroni correction was performed. The Wilcoxon rank-sum test was used to compare the median of variables. All tests were two-sided. A P value of <0.05 was considered significant. Analyses were performed using SPSS 16.0 for Macintosh.

**Results**

**Urine and serum chemistry in Ctns⁻/⁻ mice**

We characterized urine and serum chemistry in Ctns⁻/⁻ mice compared with age-matched WT mice at the age of 1, 4, 9, and 12 months. In Ctns⁻/⁻ mice, significantly higher urine phosphate levels and TEP were demonstrated as early as 4 months old relative to controls (Table 1). Proteinuria was evident at 9 months old while polyuria was significant at 12 months old Ctns⁻/⁻ mice. BUN and serum cystatin C levels were significantly higher while creatinine clearance was decreased in Ctns⁻/⁻ mice than controls at 9 months of age. Serum creatinine was significantly higher while creatinine clearance was decreased in Ctns⁻/⁻ mice relative to WT controls at 12 months of age. Serum bicarbonate levels were not different between Ctns⁻/⁻ mice and WT controls.

As Ctns⁻/⁻ mice developed CKD at the age of 9 months, urine and serum chemistry in 9- and 12-month-old Ctns⁻/⁻ mice were compared with age-matched pair-fed CKD mice.

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mice were 9.0% lighter than WT controls (Figure 1B). Body mass of 9- and 12-month-old Ctns<sup>−/−</sup> mice was comparable with that of pair-fed CKD mice, respectively (Figure 1A and 1D). There was no difference in RER between Ctns<sup>−/−</sup>, WT, and CKD mice (Figure 1E and 1F). At the end of the study, we used quantitative magnetic resonance to analyze body composition. Muscle wasting was evident in Ctns<sup>−/−</sup> and CKD mice vs. WT mice. At 12 months of age, the percentage of lean mass and total lean mass was significantly decreased in Ctns<sup>−/−</sup> mice than WT or CKD mice (Figure 1G and 1H). In contrast, the percentage of fat mass was elevated in Ctns<sup>−/−</sup> mice vs. WT mice while the total fat mass was not different between Ctns<sup>−/−</sup>, WT, and CKD mice (Figure 1I).

**Reduction muscle fibre size and impaired muscle function in Ctns<sup>−/−</sup> mice**

We studied the effects of cystinosis on skeletal muscle histomorphometry. Muscle sections were labeled for the muscle fibre basal membrane. Representative photomicrographs

### Table 2 PCR primer information

| Gene      | Forward primer sequence            | Reverse primer sequence            | Primer bank ID |
|-----------|------------------------------------|------------------------------------|----------------|
| Acox1     | TAACCTCCTCACCTGAGCCCA              | AGTTCCAATCCCCATCTGTCG              | 26333821a1     |
| Acs1      | TGGCAGTGTGTTATAGCAC                  | GGGCAGTCGAGAGGTTGTC                | 31560705a1     |
| Atg1      | AGATGAGAAGCGATGTTGTCCT             | CCCTGAGTCGAGCCATATCTA             | 27923915a1     |
| Atrogin-1 | CAGCTCTGGACGACGAC                  | GGCAGTCGAGAGGTTGTC                | 13385848a1     |
| CD137     | CGTGCAGAATCTCTGGATACAC             | GTCGACATATGCTGGAGAAGG              | 20306992a1     |
| Cidea     | TGAATTCCATGATGGCTTGGAC             | GGCAGTCGAGAGGTTGTC                | 6680944a1      |
| CPT1α     | CTCGGCCTAGGCCCGGAG                 | CACCAGTGTAGCCGACTTC               | 27804309a1     |
| CPT1β     | GCACACCGGAGCTAGCTT                 | CAGGAGTGTCCGAGCAGGTA              | 6753512a1      |
| Dio2      | AATATTGTGGCAGAGAAAGC               | GGCAGTGTGACTGAGAAAGGT             | 6753683a1      |
| Gapdh     | AGTGGGTTGTTGAGGGAAGTTG             | TGTAGACCATGTTAGTTGAGTA             | 6679937a1      |
| Glut1     | GCTGCCATCGTGATGATAGGAC             | TGCCATAGCACTGCACTAGGGA             | 18458498a1     |
| Glut4     | GTACGAGACTAAGCTCTTCGTA             | CACAGCAGTCGAGTGTAGTA              | 6678015a1      |
| Hsl       | CCAGCCTGAGGGCCTACTG                | CTCCATGACTGACATCCG                | 26325924a1     |
| Igf-1     | CTGGACAGAGACCCCTTTC                 | GGACGGGAACCTTCTGAGT               | 6754308a1      |
| MyoD      | CAACCTCGGCGCATACAGTC                | AAAAGCAGGACTGTTAGTGAG              | 6969932a1      |
| Myogenin  | GAGACATCGCCCTATCTTACCA             | GCTGACCTCGCTCATAGGC               | 13654247a1     |
| Myostatin | AGTGGATCTAATAGGAGGCCAGT             | GTTCCAGCGGCGCTACTT                | 6754752a1      |
| MuRF-1    | GTGAGGTGCTGACTTCTGTC               | GCTGACTTCTGCTCGTGGGA              | 21523717a1     |
| Pax-3     | CGGGGCAGAATATTCCACCG                | GCCCGTGAATATACTCCCTCCG             | 26377023a1     |
| Pax-7     | TCCTGCAAGGACTCCCTGCGG              | CGGGGTTCTCCTCCTATACCCCG            | 34328055a1     |
| Pgc1α     | TATGAGTGCATAGACTGAGGCTT             | CAGGAGTGTCCGAGCAGGTA              | 6769433a1      |
| Pgc1β     | TCCTGTAAGACCCCGCAGGAT              | GCTCTGGATGGCCGAGTTA               | 18875426a1     |
| Ppara     | AGAGCCCCATCTGCTCTCTC                | AACTGTAGTCTGTGAACAAAAAA            | 31543500a1     |
| Pparaβ    | CAAGGAGCGAGCTCAGTGGAAA             | GCTGTGAGGGAGCGGTTGAGA             | 403943a1       |
| Prdm16    | CCCCTACCTGCTCTGCCAGT                | CTCGCAATCCTGTCACCTA               | 124107622x3     |
| Tbx1      | CTGGTGGAGAGCCGATCATCAG             | TGTCATCAGCAGGCAACAG                | 22094109a1     |
| Tmem26    | TTCCTGTCATCCCTCGGTC                | GGGCAGGGAAGCCATTCTT               | 29244323a1     |

Appropriate primer sequence was obtained from http://pga.mgh.harvard.edu/primerbank/

### Table 3 Taqman gene expression assays-on-demand identities

| Target genes | Assay identities |
|--------------|-----------------|
| Cox2         | Mm03294838_g1   |
| IL-1α        | Mm00494938_m1   |
| IL-6         | Mm00446190_m1   |
| PGF2α synthase | Mm00427922_m1   |
| TNF-α        | Mm00443258_m1   |
| Internal control gene | Assay Identities |
| Gapdh        | 4352339E        |
of muscle sections in 12-month-old Ctns−/− mice and WT and CKD controls are shown in Figure 2A. Mean soleus and tibias anterior fibre cross-sectional area in Ctns−/− mice was 84.5 and 85.3% of that observed in WT controls (Figure 2B and 2C). Muscle wasting in Ctns−/− mice was associated with progressive skeletal muscle weakness. Muscle function, as assessed by forelimb grip strength and rotarod activity, was significantly decreased in Ctns−/− mice vs. WT or CKD controls (Figure 2D and 2E).

Muscle wasting signaling in Ctns−/− mice

We investigated the signaling pathways associated with skeletal muscle wasting in 12-month-old Ctns−/− mice. Gastrocnemius muscle from experimental mice was dissected, and total RNA from gastrocnemius muscles was extracted and reversely transcribed. Gene expression for several transcripts associated with myogenesis and skeletal regeneration (Pax-3, Pax-7, Myogenin, MyoD, and IGF-I) was significantly decreased in gastrocnemius muscle of Ctns−/− mice relative to WT controls (Figure 3A). Gene expression of Pax-3, Pax-7, and Myogenin in Ctns−/− mice was significantly lower than that in CKD mice. In contrast, expression of muscle proteolytic genes, Myostatin, Atrogin-1, and MuRF-1, was significantly increased in Ctns−/− mice than in WT controls (Figure 3B). In addition, muscle Atrogin-1 and MuRF-1 gene expression in Ctns−/− mice was higher than that in CKD controls. Muscle lysate protein levels of inflammatory cytokines (IL-1α, IL-6, and TNF-α) were increased in Ctns−/− mice than in WT controls (Figure 3C). Muscle protein content of IL-6 and TNF-α was significantly elevated in Ctns−/− mice than in CKD controls.
Increased thermogenesis and decreased adenosine triphosphate content in skeletal muscle and adipose tissues of Ctns−/− mice

Food consumption of 1- and 4-month-old Ctns−/− mice was not different than that of age-matched WT controls (Figure 1B). The observed lower body mass in young Ctns−/− mice may be associated with disturbances in energy homeostasis before the onset of renal dysfunction. We measured muscle and liver ATP content in 1- and 4-month-old Ctns−/− mice vs. WT controls. Gastrocnemius muscle ATP was significantly decreased in Ctns−/− mice relative to age-matched WT controls (Figure 4A and 4C).

Protein contents of UCPs in gastrocnemius muscle and adipose tissues were elevated in 12-month-old Ctns−/− mice relative to WT (Figure 5A, 5C, 5E, and 5G). Protein contents of UCP-1 and UCP-2 in adipose tissue were increased in 12-month-old Ctns−/− mice vs. age-matched CKD mice. In contrast, ATP contents in muscle and inguinal white adipose tissue (WAT) were markedly decreased in Ctns−/− mice relative to WT and CKD controls (Figure 5B and 5H). ATP contents in intercapsular brown adipose tissue (BAT) and epididymal WAT were decreased in Ctns−/− mice than in WT control but were not different than those in age-matched CKD controls (Figure 5D and 5F).

We compared expression profile of genes related to energy consumption in 12-month-old Ctns−/− mice. The expression levels of genes related to fatty acid oxidation (Pparα, Pparδ, and Cpt1α) and energy consumption (Pgc1α and Pgc1β) were increased in the skeletal muscle of Ctns−/− mice than in that of WT controls (Figure 6A). In addition, Pgc1α and Pgc1β gene
expression in Ctns−/− mice was significantly higher than that in CKD mice. Increased thermogenic gene expression (Ppargc1α, Pgc1α, Cidea, Prdm16, and Dio2) was found in BAT of Ctns−/− mice than in WT controls (Figure 6B).

Epididymal WAT exhibited higher expression of Ppargc1α, Pgc1α, and glucose transporter Glut1 in Ctns−/− mice than in WT controls, but no difference was found in expression of genes involved in lipolytic metabolism (Acox1, Acstl1, Atg1, and Hsl) in Ctns−/− mice relative to WT or CKD mice (Figure 6C). Moreover, inguinal WAT of Ctns−/− mice displaced higher expression of Ppargc1α, Pgc1α, Cidea, Prdm16, and Dio2 than that of WT controls (Figure 6D).

**Adipose tissue browning in Ctns−/− mice**

Beige adipocytes are a distinct type of thermogenic fat cells in mice and humans. Browning of beige adipocytes in WAT has been associated with increased energy expenditure in cachexia. We observed elevated expression of beige adipose cell surface markers (CD137, Tmem26, and Tbx1) in inguinal WAT in 12-month-old Ctns−/− mice than in WT controls (Figure 7A). Furthermore, inguinal WAT CD137 and Tbx1 expression was higher in Ctns−/− mice than in CKD controls. Another important marker for beige adipocyte in inguinal WAT is UCP-1, which is usually not detected in WAT. UCP-1 protein was detected in inguinal WAT of 12-month-old Ctns−/− and CKD mice but was undetectable in WT controls (Figure 5G). UCP-1 protein level in inguinal WAT of 12-month-old Ctns−/− was higher than that in CKD mice. Collectively, these results demonstrate the development of beige adipocytes in 12-month-old Ctns−/− mice. Our results show that increased energy expenditure is associated with adipose tissue browning in Ctns−/− mice.

We also measured gene expression of key molecules implicated in the pathogenesis of WAT browning in mice. Inguinal WAT of 12-month-old Ctns−/− mice displayed higher mRNA expression of Cox2 and PGF2α than that of WT controls (Figure 7B). Gene expression of inflammatory cytokines, IL-1α, IL-6, and TNF-α, was significantly elevated in inguinal WAT of 12-month-old Ctns−/− mice than in WT controls (Figure 6C). Moreover, inguinal WAT of Ctns−/− mice exhibited higher expression of Ppargc1α, Pgc1α, Cidea, Prdm16, and Dio2 than in WT controls, but no difference was found in expression of genes involved in lipolytic metabolism (Acox1, Acstl1, Atg1, and Hsl) in Ctns−/− mice relative to WT or CKD mice (Figure 6C). Moreover, inguinal WAT of Ctns−/− mice exhibited higher expression of Ppargc1α, Pgc1α, Cidea, Prdm16, and Dio2 than that of WT controls (Figure 6D).

**Figure 3** Gene expression of key molecules implicated in muscle wasting in 12-month-old Ctns−/− mice (A and B) and muscle pro-inflammatory cytokine protein contents (C). Comparative 2-ΔΔCt method was used to determine the relative quantification of genes in muscle. To normalize each sample for RNA content, the internal control gene GAPGH was used. Final results were expressed in arbitrary units, with one unit being the mean mRNA level in the wild-type controls. Data are expressed as mean ± standard error of the mean. *P < 0.05, **P < 0.01, ***P < 0.001.

**Figure 4** Adenosine triphosphate content in muscle and liver in 1- and 4-month-old Ctns−/− mice. Adenosine triphosphate content in tissue lysate was measured in duplicate and calculated per gram of tissue. Number of mice is ≥6 in each group. Data are expressed as mean ± standard error of the mean. *P < 0.05, **P < 0.01, ***P < 0.001.

**Table 1** Deposition of muscle wasting in 12-month-old Ctns−/− mice (A and B) and muscle pro-inflammatory cytokine protein contents (C). Comparative 2-ΔΔCt method was used to determine the relative quantification of genes in muscle. To normalize each sample for RNA content, the internal control gene GAPGH was used. Final results were expressed in arbitrary units, with one unit being the mean mRNA level in the wild-type controls. Data are expressed as mean ± standard error of the mean. *P < 0.05, **P < 0.01, ***P < 0.001.
WAT of 12-month-old Ctns⁻/⁻ mice than that in age-matched WT controls or CKD mice (Figure 7B).

**Discussion**

This study confirms the presence of the tubular and glomerular dysfunction in Ctns⁻/⁻ mice. Fanconi syndrome, the hallmark of the tubular dysfunction in children with INC, characterized by phosphaturia, proteinuria, and polyuria, was demonstrated in Ctns⁻/⁻ mice (Table 1). Indeed, elevation of TEP index, the more sensitive parameter of tubular function, was demonstrated earlier in the course of the renal disease (4 months of age) than the glomerular dysfunction (9 months of age). Serum cystatin C is a reliable marker for renal function in mice and humans, which has the advantage over serum creatinine when muscle wasting can be a confounding factor. Serum cystatin C levels were elevated in Ctns⁻/⁻ mice than in controls at 9 months of age. The time course of renal dysfunction in Ctns⁻/⁻ mice of this study is in concordance with a recent report.

Importantly, we show that Ctns⁻/⁻ mice exhibit disturbances in energy homeostasis prior to the development of CKD. Basal metabolic rate comprises 50–80% of daily energy expenditure and exhibits circadian rhythms. Food consumption in Ctns⁻/⁻ mice was not different than age-matched WT controls (Figure 1B). However, significantly lower total body mass starting at 1 month of age as well as increased energy expenditure at 4 months of age (Figure 1A, 1C, 1D, and 1G) preceded the onset of CKD at 9 months of age in Ctns⁻/⁻ mice (Table 1). Muscle ATP content in 1- and 4-month-old Ctns⁻/⁻ mice was significantly lower than that in age-matched WT controls (Figure 4A and 4C). We compared parameters of energy homeostasis in 9- and 12-month-old Ctns⁻/⁻ mice vs. pair-fed age-matched CKD mice. VO₂ and energy expenditure at dark phase was significantly elevated in 9- and 12-month-old Ctns⁻/⁻ mice than that in CKD controls (Figure 1D and 1H).

Muscle wasting is a life-threatening complication in cystinosis. We observed decreased body mass and lean mass in 12-month-old Ctns⁻/⁻ mice than in WT control or pair-fed CKD mice (Figure 1A and 1I). We further measured muscle fibre histomorphometry and muscle function in Ctns⁻/⁻ mice. Soleus and tibias anterior muscle were chosen as they represent extremes of mouse muscle types in terms of fibre type composition since the soleus is composed of about 50% type 1 fibre (slow) while the tibias anterior has...
no slow fibres. Cross-sectional area of soleus and tibialis anterior was significantly reduced in Ctns−/− mice than in WT controls (Figure 2A, 2B, and 2C). We also showed that reduced muscle mass and decreased muscle fibre cross-sectional area correlate with reduced grip strength and impaired rotarod activity in Ctns−/− mice than in WT and CKD mice (Figure 2D and 2E). Reduced cross-sectional area of muscle is not always associated with diminished muscle strength in cachectic patients.18,19

Our results suggest that loss of lean mass, decreased muscle fibre cross-sectional area, and impaired muscle function are more severe in 12-month-old Ctns−/− mice than in age-matched pair-fed CKD controls. Indeed, muscle wasting in children with nephropathic cystinosis seems to be more prevalent than in CKD children with comparable degree of renal dysfunction. Although there is no direct comparison, muscle wasting is highly prevalent in patients with nephropathic cystinosis with predialysis CKD (33–60% in two long-term follow-up studies).4,5 On the other hand, a recent study showed no appendicular lean mass deficits in children with mild to moderate CKD.20

We studied muscle mass regulatory signaling pathways in Ctns−/− mice. mRNA contents of Pax-3 and Pax-7, Myogenin, and MyoD were decreased in the gastrocnemius muscle of

Figure 6 Thermogenic gene expression in 12-month-old Ctns−/− mice. Thermogenic gene expression in skeletal muscle and adipose tissues (A to D). Number of mice = 6. Data are expressed as mean ± standard error of the mean. *P < 0.05, **P < 0.01, ***P < 0.001.
12-month-old Ctns−/− mice (Figure 3A), suggesting altered satellite cell content or activity and reduced myoblast differentiation in Ctns−/− mice. Expression of muscle IGF-I was down-regulated while Myostatin mRNA was up-regulated in Ctns−/− mice (Figure 3A and 3B). IGF-I and Myostatin play stimulatory and inhibitory roles, respectively, in the regulation of muscle mass.21 Expression of muscle atrophy-associated genes, Atrogin-1 and MuRF-1, was significantly increased in Ctns−/− mice relative to WT controls (Figure 3B). Muscle protein contents of IL-1α, IL-6, and TNF-α were significantly increased in Ctns−/− mice than in WT controls (Figure 3C). Inflammatory cytokines have been implicated in the pathogenesis of muscle wasting.22,23

Skeletal muscle accounts for 20–30% of overall energy consumption at rest.24,25 Chronic inflammation in skeletal muscle may cause hypermetabolism in Ctns−/− mice. We showed that muscle inflammatory cytokines (IL-1α, IL-6, and TNF-α) were significantly increased in Ctns−/− mice (Figure 3C). Common causes of chronic inflammation include deterioration of renal function, volume overload, alternation in body composition, and acidosis.22,23 Metabolic acidosis is unlikely the culprit in Ctns−/− mice as they were not acidotic. Serum bicarbonate level in Ctns−/− mice was not different from that in controls (Table 1).

Recent studies suggest that UCP-1 contributes to adaptive thermogenesis while UCP-2 and UCP-3 are involved in the resting metabolic rate.26 We investigated the mechanisms of hypermetabolism in Ctns−/− mice. Protein contents of UCP-2 and UCP-3 were increased in muscle and WAT of Ctns−/− mice than in WT controls (Figure 5A and 5E). In contrast, ATP contents in muscle and adipose tissue were decreased in Ctns−/− mice relative to controls (Figure 5B and 5F). In addition, increased UCP-1 and decreased ATP contents in adipose tissue were observed in Ctns−/− mice than in WT controls (Figure 5C, 5D, 5G, and 5H). Adipose tissue is also important in energy metabolism. We, and others, have previously described the increased thermogenesis and up-regulation of UCPs in adipose tissue in rodent models of cachexia and in cachectic patients.9,13,27–31 Up-regulation of UCPs expression promotes proton leak and reduces cellular ATP production in exchange for the generation of heat.32,33 In vitro studies have reported that modest increase in the expression of UCP-2 leads to rapid fall in mitochondrial membrane potential and a reduction of intracellular ATP content.34 UCP-3 modulates the activity of sacro/endoplasmic reticulum Ca2+-ATPase and decreases mitochondrial ATP production.35 Several studies have reported decreased levels of ATP in cystinotic cells, including fibroblast, leucocytes, and renal epithelial cells.36 Abnormal ATP production has been associated with impaired respiratory chain complex I activity in cystinotic cells.37

We also profiled expression of genes related to energy consumption in skeletal muscles of Ctns−/− mice. PPARα and PPARγ as well as CPT1α and CPT1β are key regulators of fatty acid oxidation in muscle.38,39 Transcriptional levels of PPARα, PPARγ, and CPT1α were significantly up-regulated in the skeletal muscle of Ctns−/− mice (Figure 6A). This profile has been associated with increased resting metabolic rate and maintenance of leanness in humans.40 Activated fatty acid oxidation is associated with up-regulation of PGC1α and PGC1β, two key molecules involved in the regulation of mitochondrial energy metabolism.41,42 This was also confirmed in Ctns−/− mice (Figure 6A). In addition, we demonstrated increased expression of thermogenic genes in BAT, and epididymal and inguinal WAT of Ctns−/− mice (Figure 6B, 6C, and 6D). UCP-2 and UCP-3 regulate fatty acid metabolism in skeletal muscle and adipose cells. UCP-2 and UCP-3 promote fatty acid metabolism by exporting fatty acid anions outside of the mitochondrial matrix.26 Collectively, our results suggest that up-regulation of UCPs, elevated key molecules of fatty acid oxidation, and mitochondrial energy metabolism in

**Figure 7** Adipose tissue browning in 12-month-old Ctns−/− mice. Gene expression of beige adipocyte marker (CD137, Tmem26, and Tbx1) in inguinal white adipose tissue was measured (A). Gene expression of key molecules implicated in adipose tissue browning (B). Results were analysed and expressed as in Figure 3. *P < 0.05, **P < 0.01, ***P < 0.001.
muscle and adipose tissue may be implicated in the pathogenesis of hypermetabolism in Ctns−/− mice.

Adipose tissue browning has recently been reported in cancer cachexia. White fat depots contain pockets of UCP-1-expressing multicellular, called beige (or brite) cells, that can be stimulated on exposure to cold or other stimuli via the process termed browning. Epididymal WAT predominantly contains white adipocytes while inguinal WAT contains a mixed population of white and beige adipocytes. We demonstrated increased expression of unique beige adipose cell markers (CD137, Tmem26, and Tbx1) and BAT marker UCP1 in inguinal WAT of Ctns−/− mice (Figures 5G and 7A). Our results suggest the development of beige adipocytes in Ctns−/− mice. The development of browning and beige adipocytes in Ctns−/− mice is a novel finding. The mechanism is currently unknown. Recent evidence shows that differentiation of white adipocytes and de novo differentiation of beige adipocyte coexist. Beige adipocyte precursors have also been identified in skeletal muscle, which further underlies the complexity of beige adipogenesis. Transcriptional regulator PPARγ is necessary and sufficient for adipogenesis. The actions of PPARγ are modulated by a large set of proadipogenic transcriptional cofactors. We demonstrated increased expression of PPARβ, CIDEA, PDRM16, and DIO2 in inguinal WAT of Ctns−/− mice (Figure 6D). The important roles of transcriptional factors DIO2, PDRM16, PPARβ, and CIDEA on adipogenesis and thermogenesis have been described. Overexpression of these transcriptional cofactors in WAT favors adipocyte browning and thermogenesis.

WAT browning is responsible for a significant increase in total energy expenditure. Several mechanisms have been proposed for WAT browning, including activation of Cox2 signaling pathway and chronic inflammation. Activation of Cox2, a downstream effector of β-adrenergic signaling, is crucial for the induction of brown fat-like cells in WAT depots. Cox2 produces prostaglandins that enhance mitochondrial biogenesis and increase the uncoupling capacity when activated with adrenergics. We showed that inguinal WAT gene expression of Cox2 and PGF2α was significantly increased in Ctns−/− mice vs. WT control mice (Figure 7B). Chronic inflammation is a hallmark of both clinical and experimental cachexia. Inflammatory cytokines, such as IL-6, play an important role in the WAT browning phenotype in mouse models of cachexia. Gene expression of IL-1α, IL-6, and TNF-α was significantly elevated in inguinal WAT of 12-month-old Ctns−/− mice than that in age-matched WT controls or CKD mice (Figure 7B). Inflammation in WAT is characterized by recruitment of macrophages, including activated M1 and M2 macrophages. Activated macrophages in the WAT are an important source of adrenaline and noradrenaline and have been associated with an increase in uncoupled respiration and energy expenditure in mice. We showed that the phenomenon of adipose tissue browning was more advanced in 12-month-old Ctns−/− vs. CKD controls, as indicated by the higher expression of browning markers CD137 and Tbx1 (Figure 7A). At the same time, there was evidence of more severe inflammation in the Ctns−/− mice compared with CKD controls as indicated by the inflammatory cytokine expression levels of IL-1α, IL-6, and TNF-α in inguinal WAT (Figure 7B). We postulate that inflammation is an important mechanism underlying the adipose tissue browning in INC.

Conclusion

We describe muscle wasting and adipose tissue browning in a mouse model of INC. We show that Ctns−/− mice exhibit disturbances in energy homeostasis before the onset of their CKD. We demonstrate aberrant muscle mass regulatory signaling pathways in Ctns−/− mice. We show that hypermetabolism in Ctns−/− mice is associated with up-regulation of key enzymes regulating thermogenesis in skeletal muscle and adipose tissues. Importantly, we report novel findings in the development of beige adipocytes in Ctns−/− mice. Further studies are required to investigate the underlying mechanisms of these metabolic defects in INC, which are associated with poor quality of life and mortality, and for which there is no current therapy.

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Conflict of interest

The authors declare they have no conflicts of interest.
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