Effect of *Nigella sativa* on ischemia-reperfusion induced rat kidney damage

Shahrzad Havakhah 1, Hamid R Sadeghnia 2, Mosa-Al-Reza Hajzadeh 3, Nama Mohammadian Roshan 4, Somayeh Shafiee 1, Hossein Hosseinzadeh 5, Narges Mohareri 6, Abolfazl Khajavi Rad 7*

1 Department of Physiology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
2 Pharmacological Research Center of Medicinal Plants, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
3 Neurocognitive Research Center, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
4 Department of Pathology, Ghaem Hospital, Mashhad University of Medical Sciences, Mashhad, Iran
5 Pharmaceutical Research Center, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran
6 Department of Pharmacodynamics and Toxicology, School of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran
7 Neuroinflammation Research Center, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

**ABSTRACT**

**Objective(s):** There are a few previously reported studies about the effect of *Nigella sativa* oil on renal ischemia-reperfusion injury (IRI). The aim of the present study was to test the hypothesis whether pre- or post-treatment with *N. sativa* hydroalcoholic extract (NSE) would reduce tissue injury and oxidative damages in a clinically relevant rat model of renal IRI.

**Materials and Methods:** IRI was induced by clamping of bilateral renal arteries for 40 min followed by reperfusion for 180 min. NSE was prepared in a Soxhlet extractor and administrated with doses of 150 mg/kg or 300 mg/kg at 1 hr before ischemia induction (P-150 and 300) or at the beginning of reperfusion phase (T-150 and 300). via jugular catheter intravenously. The kidneys were then removed and subjected to biochemical analysis, comet assay or histopathological examination.

**Results:** The kidneys of untreated IRI rats had a higher histopathological score (*P*<0.001), while in P-150, as well as T-150 and T-300 groups tubular lesions significantly decreased (*P*<0.001). Pre-and post-treatment with NSE also resulted in a significant decrease in malondialdehyde (MDA) level (*P*<0.001) and DNA damage (*P*<0.001) that were increased by renal I/R injury. NSE treatment also significantly restore (*P*<0.01) the decrease in renal thiol content caused by IRI.

**Conclusion:** The present study shows *N. sativa* extract has marked protective action against renal IRI, which may be partly due to its antioxidant effects.

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**Introduction**

Renal ischemic-reperfusion injury (IRI) is a complex pathophysiologic process that occurs after cardiac arrest, kidney transplantation, hemi nephrectomy and vascular surgery. IRI causes renal cell death, acute kidney injury (AKI), formerly known as “acute renal failure”, acute tubular necrosis (ATN), acute rejection, delayed graft function, and chronic allograft dysfunction. Basic mechanisms of renal ischemic-reperfusion injury are multifactorial and interrelated including anoxia, free radicals generation especially during reperfusion and inflammation responses (1).

Ethnobotanicals are important for pharmacological research and drug development, not only when plant constituents are used directly as therapeutic agents, but also as founding materials for the synthesis of drugs or as models for pharmacologically active compounds. It is estimated that about 25% of the drugs prescribed worldwide are derived from plants (2).

*Nigella sativa*, which belongs to the Ranunculaceae family, commonly grows in Eastern Europe, Mediterranean regions, the Middle East, and Western Asia. Seeds of *N. sativa*, also known as black cumin (English), black caraway (USA) and shonaiz (Persian), frequently used in folk medicine for promoting health and treatment of many diseases such as asthma, rheumatic diseases, microbial infections and cancer (3). Recent clinical and experimental researches have shown many pharmacological properties, like immu nomodulatory (4), anti-inflammatory (5), antioxidant (6), anticarcinogenic (7), antihypertensive (8), anti-ischemic (9) and renal calculi prevention (10).

Hosseinzadeh and Montahaei showed that acute administration of aqueous or ethanolic extract of *N. sativa* and thymoquinone following 60 min warm renal ischemia, significantly improved oxidative stress markers in kidney tissue (11). It has been shown that pre- and post-treatment with *N. sativa* seed oil significantly protect oxidative damage and prevent tissue damage following renal IRI (12).
Moreover, treatment with *N. sativa* oil 6 hr prior to ischemia-reperfusion and at the beginning of reperfusion reduced renal oxidative stress markers and tubular necrosis score (9). Science there is no previously reported study on the effect of hydroalcoholic extract of *N. sativa* (NSE) on histological and DNA damages induced by renal I/R injury, the present study was designed to test whether pre- or post-treatment with NSE would reduce tissue injury and oxidative damages in a clinically relevant rat model of renal IRI.

**Material and Methods**

**Animals**

Adult male Wistar rats weighting 250-300 g from the Central Animal House of Mashhad University of Medical Sciences (Mashhad, Iran), were used throughout the study. The animals were housed in the same room under a constant temperature (22±2°C) and standard conditions of a 12 hr light/dark cycle with free access to food pellets and tap water, available *ad libitum*.

The experimental protocol was approved by the Animal Care and Use Committee, Mashhad University of Medical Sciences and was performed in accordance with the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals.

**Preparation of hydroalcoholic extract of *N. sativa***

The seeds of *N. sativa* were obtained from Medicinal Plants Division of Imam Reza Pharmacy (herbarium No. 80140). They were washed, dried, and crushed to a powder with an electric microniser. The powdered seeds (100 g) were extracted in a Soxhlet extractor with ethanol (70%). The resulting extract was concentrated under reduced pressure and kept at -20°C until use (yielded 32%).

**Chemicals**

5, 5’-dithiobis-2-nitrobenzoic acid (DTNB), 2-thiobarbituric acid (TBA), trichloroacetic acid (TCA), ethylenediaminetetraacetic acid disodium salt (Na₂EDTA), t-octylphenoxypoly-ethoxyethanol (Triton X-100), tris (hydroxymethyl) aminomethane (Trizma base), sodium lauroylsarcosinate (sarkosyl, SLS), sodium dodecyl sulphate (SDS), dimethylsulfoxide (DMSO), ethanol, sodium hydroxide (NaOH) and potassium chloride (KCl) were obtained from Merck (Darmstadt, Germany). Low melting point (LMP) agarose and normal melting point (NMP) agarose were purchased from Fermentas (Glen Burnie, MD). LMP and NMP agarose were diluted in physiological saline to 0.5% and 1%, respectively.

**Study design and induction of renal IRI**

The rats were randomly assigned to one of the following six experimental groups, each consisting of eight animals: control group (sham-operated animals) underwent all surgical procedures similar to all other groups, except that the renal artery was not clamped; Ischemia-reperfusion injury (IRI) group underwent both renal arteries clamped for 40 min (ischemia phase) and fallowed by reperfusion for 180 min (reperfusion phase); Preventive-150 group (P-150) underwent 40 min ischemia and NSE was injected with a dose of 150 mg/kg at 1 hr before ischemia induction (control phase); Preventive-300 group (P-300) underwent 40 min ischemia and NSE was injected with a dose of 300 mg/kg at the beginning of control phase; Treatment-150 group (T-150) underwent 40 min ischemia and NSE was injected with a dose of 150 mg/kg at the beginning of reperfusion phase and Treatment-300 group (T-300) underwent 40 min ischemia and extract was injected with a dose of 300 mg/kg at the beginning of reperfusion phase.

Rats were anaesthetized with intraperitoneal (IP) injection of sodium pentobarbital (70 mg/kg) under sterile condition. At first the neck exposed in midline and jugular vein and carotid artery dissected from the surrounding tissues and canulated for infusion of fluids and extract and connected to power lab system for monitoring and collecting blood samples, respectively. A midline incision was made, kidneys were exposed and renal arteries dissected from surrounding tissues. Ischemia was induced by bilateral renal artery clamping for 40 min with smooth vascular clamps. After the clamps were removed, the kidneys were inspected for restoration of blood flow. After bilateral nephrectomies were carried out, the half of both kidneys were stored at -70°C for biochemical analysis and comet assay, whereas the other half of kidneys were fixed in 10% formalin for histopathological examination.

**Histopathological examination**

Formalin-fixed tissue samples were embedded in paraffin, cut at 5 μm, and stained with hematoxylin and eosin (H & E). Tubular injury was scored in a blinded manner by estimation of the percentage of tubules in the outer medulla and corticomedullary junction that showed epithelial necrosis, debris or cast as follows: Grade 0, no morphologic changes; grade 1, <25%; grade 2, <50% and grade 3, <75% involved. More than 20 consecutive fields were examined under ×400 magnification and averaged per slide (13).

**Thiobarbituric acid reactive species measurement**

Renal lipid peroxides formation was measured as malondialdehyde (MDA), which is the end product of lipid peroxidation and reacts with thiobarbituric acid (TBA) as a TBA reactive substance (TBARS) to produce a pink colored complex which has peak
absorbance at 535 nm (14). In brief, 1 ml of homogenate sample was mixed with 2 ml of TCA-TBA-HCl reagent (15% TCA, 0.67% TBA, and 0.25 N HCl) and heated for 45 min in a boiling water bath. After cooling, the mixture was centrifuged at 3000 rpm for 10 min. The supernatant was collected, and the absorbance was read against blank, at 535 nm. The amount of MDA produced was calculated, using a molar absorption coefficient of 1.56×10^5 M^-1 cm^-1 and expressed as nmol/g tissue (15).

**Total sulfhydryl (SH) groups measurement**

Total thiol content was estimated based on the Ellman method (16). In this method, SH groups react with chromogenic DTNB and produce a yellow-colored dianion (5-thio-2-nitrobenzoic acid, TNB), which has peak absorbance at 412 nm. Briefly, 1 ml Tris-EDTA buffer (0.1 M Tris, 10 mM EDTA, pH=8.6) was added to 50 μl homogenate sample in 2 ml cuvettes. Sample absorbance was read at 412 nm against Tris-EDTA buffer alone (A1), then 20 μl DTNB reagent (10 mM in methanol) was added again (A2). DTNB reagent absorbance was also read as a blank (B). Total thiol concentration was calculated by the following equation and expressed as nmol/mg tissue (15).

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\text{Total thiol concentration (mM)} = \frac{(A_2-A_1-B)}{(1.07/0.05)} \times 13.6
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**Single cell gel electrophoresis (SCGE, Comet) assay**

The in vivo alkaline SCGE assay was conducted based on the method described previously (17). After weighing, the samples were minced, suspended at a concentration of 0.5 g/ml in chilled homogenizing buffer containing 0.075 M NaCl and 0.024 M Na2EDTA (pH 7.5), and then homogenized gently at 500-800 rpm in ice. To obtain nuclei, the homogenate was centrifuged at 700 g for 10 min at 0°C, and the precipitate was resuspended in chilled homogenizing buffer at 0.5 g/ml and allowed to settle; precipitated clumps were then removed. 80 μl NMP agarose was quickly layered on conventional slides, covered with a cover slip, and then the slides were placed on ice to allow agarose to gel. Five microliters of the nucleus suspension, prepared as above, was mixed with 75 μl LMP agarose and the mixture was quickly layered over the NMP agarose layer after removal of the cover slip. Finally, another layer of LMP agarose was added on top. The slides were immersed immediately in a chilled lysing solution (pH = 10) made up of 2.5 M NaCl, 100 mM Na2EDTA, 10 mM Trizma, 1% sarcosyl, 10% DMSO, and 1% Triton X-100 and kept at 0°C in the dark overnight. Then, the slides were placed on a horizontal gel electrophoresis platform and covered with a prechilled alkaline solution made up of 300 mM NaOH and 1 mM Na2EDTA (pH > 13). They were left in the solution in the dark at 0°C for 40 min and then electrophoresed at 0°C in the dark for 30 min at 25 V and approximately 300 mA. The slides were rinsed gently three times with 400 mM Trizma solution (adjusted to pH 7.5 with HCl) to neutralize the excess alkali, stained with 50 μl of 20 μg/ml ethidium bromide and covered with a cover slip. For comet analysis, 100 nuclei were randomly selected from two replicated slides (50 nuclei on each slide), examined and photographed through a fluorescence microscope (Nikon, Japan), at 400x magnification equipped with an excitation filter of 520-550 nm and a barrier filter of 580 nm. Undamaged cells resemble an intact nucleus without a tail, and damaged cells have the appearance of a comet. The percent of DNA in the comet tail (% tail DNA), which is an estimation of DNA damage, was analyzed using the computerized image analysis (CASP) software.

**Statistical analysis**

Data are presented as mean±SEM. Statistical analysis was performed using one-way ANOVA followed by Tukey-Kramer post-hoc test. The P-values less than 0.05 were considered to be statistically significant.

**Results**

**NSE ameliorates renal histologic damage following IRI**

The histopathologic changes were graded and summarized as % renal tissue damage in Figure 1G. There were significant tubular changes including loss of brush border, dilation of renal tubules, cast formation as well as degeneration and necrosis of renal tubular epithelial cells following IRI compared with the sham group (Figure 1A-G). In contrast, pretreatment with low dose of NSE (P-150), as well as treatment with NSE (T-150 and T-300) significantly ameliorated tubular lesions from grade 2 (<50% renal tissue damage in IRI group) to grade 1 (<25% injury) (Figure 1A-G).

**NSE reduces renal lipid peroxidation following IRI**

The renal MDA content was elevated (about 38%) following induction of renal IRI compared to the normal sham-operated group (Figure 2; P<0.05). However, NSE pretreatment resulted in a significant decrease in MDA level compared to the IRI group (P<0.05). As shown in Figure 2, NSE at doses of 150 mg/kg and 300 mg/kg decreased renal TBARS levels about 44.5% (P<0.001) and 25% (P<0.05) following IRI, respectively. Further however, renal MDA content was not significantly affected by treatment with concentrations up to 300 mg/kg NSE.

**NSE increases renal total thiol content following IRI**

Renal IRI resulted in a significant decrease in renal thiol content when compared to normal sham-
Effect of Nigella sativa on renal ischemia-reperfusion injury

Figure 1A-G. Effect of Nigella sativa hydroalcoholic extract (NSE) on histologic changes induced by renal ischemia-reperfusion injury (IRI). Rat’s kidney sections were stained by hematoxylin and eosin (H & E) and examined using a light microscope (×400). Arrow heads show kidney tubules and G represents renal glomeruli. Rats treated with NSE (150 and 300 mg/kg) 1 hr prior (P-150 and P-300) and right after (T-150 and T-300) the reperfusion. A: Control (sham) group with normal renal morphology, B: IRI group with the distinctive pattern of ischemic renal injury (dilation of renal tubules, cast formation as well as tubular epithelial cells necrosis), C: P-150 group with relatively well-preserved architecture and focal tubular necrosis, D: P-300 group with greater tubular necrosis, E: T-150 group with relatively well-preserved architecture and cast formation, F: T-300 group with lower degree of vacuolization and cast formation, G: Histopathological score in experimental groups, values are mean±SEM (n = 8 per group) and analyzed using one-way ANOVA followed by Tukey-Kramer post hoc test. *P<0.05 and ***P< 0.001 compared to IRI group operated rats (P<0.01; Figure 3). As Figure 3 illustrates, renal thiol content was not significantly increased following NSE pre-treatment in comparison with IRI group (P>0.05). On the other hand, a significant restoration in the total thiol levels was observed in NSE treated rats, as compared to IRI group.

NSE decreases oxidative DNA damage following renal IRI

Figure 4A-D shows represented comet images from different treatment group. A significant increase in the % tail DNA was seen in nuclei of renal ischemic group, as compared to those of saline-treated sham-operated animals (P<0.001; Figure 4F). The ischemic-induced DNA damage in NSE-treated animals was decreased about 3.6 fold (150 mg/kg) and 3.2 fold (300 mg/kg) as compared to that of ischemic animals alone (P<0.001; Figure 4E). On the other hand, pretreatment with NSE did not significantly decrease DNA damage induced by ischemic insult (Figure 4E).

Discussion

The aim of the present study was to test whether pre- and post-treatment with N. sativa hydroalcoholic extract would reduce tissue injury and oxidative damages in a clinically relevant rat model of kidney ischemia/reperfusion injury. The results demonstrated that IRI produced significant changes in the renal histopathologic features, induced lipid peroxidation, reduced the total thiol levels and
caused oxidative DNA damages. Pre- and post-treatment with NSE significantly attenuated renal oxidative stress and preserved the normal morphology of kidney.

IRI is an important cause of acute kidney injury (AKI), formerly known as acute renal failure, in native and transplanted kidneys. AKI is a complex syndrome characterized by vasocostriction, glomerular injury with major reductions in glomerular filtration rate, extensive tubular damages and tubular cell necrosis (18). Most of these tubular and glomerular damages occur during the reperfusion phase following ischemia, and generation of reactive oxygen species (ROS). ROS are capable of reacting with lipids, proteins and nucleic acids leading to lipid peroxidation, impairments of enzymatic processes and DNA damages, thereby, inhibiting transcription and repair. Ahmadiasl et al have shown that in the rat model of renal IRI, there is significant increase in renal oxidative stress that leads to both structural and functional damages (19).

Lipid peroxidation, mediated by ROS, is an essential cause of destruction and damage to cell membranes following renal IRI (18). In the present study there was an increase in MDA content of renal tissue in the IRI group compared with sham-operated rats. These data are in good agreement with the work of Bayrak et al who found high lipid peroxidation after renal I/R injury in rats (12). Our study also showed that NSE could reverse the increase of MDA levels to a considerable extent. Among all endogenous antioxidant mechanisms, thiols constitute the major portion of the total body antioxidants and they play a significant role in defense against ROS. Total thiols composed of both intracellular and extracellular thiols either in the free form as oxidized, reduced glutathione, or thiols bound to proteins, which scavenge oxygen free radicals (20). We observed a significant decrease in thiols content during ischemia/reperfusion injury. Such reduction in total thiol levels has shown to occur in the kidney after a period of IRI leading to oxidative stress and cellular damages (21). Moreover, we showed that thiol groups increased following NSE treatment, thereby confirming its antioxidant role in IRI. These findings are in agreement with those of Hosseinizadeh et al, reporting that warm ischemia for 60 min followed by reperfusion for 90 min causes a significant increase in renal lipid peroxidation along with considerable reduction in tissue thiol level (21). We found that IRI caused renal oxidative stress, oxidative DNA damage and structural injury significantly. Consistent with these findings, Simon et al demonstrated that aortic occlusion-induced kidney ischemia/reperfusion injury increased kidney oxidative DNA base damage which coincided with histopathology and apoptosis (22).

Other reports about modulatory effect of N. sativa against ischemic reperfusion injuries show that it improves rat cardiac musculature, mitochondrial dysfunction and subsequent lipid peroxidation post I/R insult (23). The in vivo and in vitro inhibitory effects of N. sativa against cerebral IRI are also reported (24-26). Ameliorating effects of different extracts of N. sativa (aqueous, hydroalcoholic, chloroform and petroleum ether) in middle cerebral artery occluded rat were shown via reduction in lipid peroxidation and elevation in glutathione and antioxidant enzymes, superoxide dismutase and catalase (27, 28). Hobenaghii et al also found that N. sativa extract decreases hippocampal neurons edema and astrocytes death following global cerebral IRI in rats (24). Similarly, in a rat model of gastric mucosal I/R injury, N. sativa normalizes the level of lipid peroxidation, glutathione, superoxide dismutase and lactate dehydrogenase (29). Moreover, Yildiz et al also showed that N. sativa seed oil prevents histological liver damage induced by IRI via decreasing oxidative stress biomarkers (30). Moreover, NSE reduces tissue...
damages in rat ovaries subjected to IRI by attenuating oxidative and inflammatory responses (31). It was shown that inflammatory responses play critical roles in ischemic renal injury (18). Using 18F-fluoro-deoxy-D-glucose-positron emission tomography (18F-FDG-PET) imaging, Entok et al demonstrated that N. sativa (500 mg/kg) and essential oil (5 ml/kg) treatment caused a significant reduction in lipopolysaccharides-induced lung and liver acute inflammation. This anti-inflammatory effect coincides with decreased tissue levels of MDA, nitric oxide and DNA fragmentation and also conserved antioxidant enzymes, superoxide dismutase and catalase activities (32). Bayrak et al showed that pre- and post-treatment with N. sativa seed oil significantly reduced renal histopathological score, improved renal function and serum and tissue anti-oxidative parameters following IRI (12). In the same manner, Yildiz et al confirmed that pre-treatment with N. sativa oil was effective against renal IRI by reducing serum urea and creatinine levels as well as decreasing the tubular necrosis score and improving blood and renal oxidative and antioxidative status (9). Our study has several differences and advantages to other studies. It is well documented that ischemic acute renal failure (ARF) is associated with high mortality in humans and current therapy is limited to supportive measures and preventive strategies, none of which have been definitively shown to alter mortality (33). We administered NSE intravenously with increasing doses (150 and 300 mg/kg) either 1 hr before induction of ischemia or at the beginning of the reperfusion phase, to better understand the time course of drug action, unlike the study of Bayrak et al (12) which N. sativa essential oil was administrated orally with single dose for 7 consecutive days prior to surgery (0.3 ml) or right after ischemia (0.6 ml). The constituents of N. sativa essential oil are quite different from NSE. It has also been shown that acute renal failure from ischemia-reperfusion is associated with tubule cell apoptosis (34). This study focuses on possible protective effects of NSE on DNA damage using comet assay as an index of apoptotic cell death.

Conclusion

In the present study, N. sativa hydroalcoholic extract had marked protective action against renal IRI. These findings are important as there is no previously reported study on the effect of NSE on histological and DNA damages induced by renal IRI. In conclusion, in view of previous reports regarding potent antioxidant and anti-inflammatory and our data, NSE seems to be a highly promising nephroprotective agent against renal I/R injury.

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