The origin of the skewed amplitude distribution of spontaneous excitatory junction potentials in poorly coupled smooth muscle cells
Young, JS; Brain, Keith; Cunnane, TC

DOI:
10.1016/j.neuroscience.2006.11.054

License:
Creative Commons: Attribution (CC BY)

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (Harvard):
Young, JS, Brain, K & Cunnane, TC 2007, 'The origin of the skewed amplitude distribution of spontaneous excitatory junction potentials in poorly coupled smooth muscle cells', Neuroscience, vol. 145, no. 1, pp. 153-161. https://doi.org/10.1016/j.neuroscience.2006.11.054

Link to publication on Research at Birmingham portal

Publisher Rights Statement:
Elsevier Open access article (supported by a grant from the Wellcome Trust, administered through the University of Oxford). This version is published in Neuroscience, [Volume 145, Issue 1, 2 March 2007, Pages 153–161] DOI: http://dx.doi.org/10.1016/j.neuroscience.2006.11.054. This article is licensed under a CC-BY licence.

General rights
Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

• Users may freely distribute the URL that is used to identify this publication.
• Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
• User may use extracts from the document in line with the concept of ‘fair dealing’ under the Copyright, Designs and Patents Act 1988 (?).
• Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy
While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 11. Mar. 2020
THE ORIGIN OF THE SKEWED AMPLITUDE DISTRIBUTION OF SPONTANEOUS EXCITATORY JUNCTION POTENTIALS IN POORLY COUPLED SMOOTH MUSCLE CELLS

J. S. YOUNG,* K. L. BRAIN AND T. C. CUNNANE
Department of Pharmacology, University of Oxford, Mansfield Road, Oxford OX1 3QT, UK

Abstract—The skewed amplitude distribution of spontaneous excitatory junction potentials (sEJPs) in the mouse vas deferens and other electrically-coupled smooth muscle syncytia has been attributed to electrically-attenuated depolarizations resulting from the spontaneous release of quantized packets of ATP acting on remote smooth muscle cells (SMCs). However, in the present investigation surface SMCs of the mouse isolated vas deferens were poorly electrically coupled, with input resistances (176±18 MΩ, range: 141–221 MΩ, n=4) similar to those of dissociated cells. Furthermore, the amplitude of evoked EJPs was more variable in surface compared with deeper SMCs (F test, F=17.4, P=0.0001). Using simultaneous electrophysiology and confocal microscopy to investigate these poorly-coupled cells, it is shown that α-latrotoxin-stimulated sEJPs correlate, in timing (median delay ranged from ~30 to ~57 ms, P=0.05 in all experiments, n=5) and amplitude (Pearson product moment correlation, ρ>0.55 and P<0.001), with purinergic neuroeffector Ca\(^{2+}\) transients (NCTs) in SMCs. The temporal correlation between sEJPs of widely ranging amplitude with NCTs in the impaled SMC demonstrates that all sEJPs could arise from neurotransmitter action on the impaled cell and that the skewed distribution of sEJPs can be explained by the variable effect of packets of ATP on a single SMC. The amplitude correlation of sEJPs and NCTs argues against the attenuation of electrical signal amplitude along the length of a single SMC. The skewed sEJP amplitude distribution arising from neurotransmitter release on single SMCs is consistent with a broad neurotransmitter packet size distribution at sympathetic neuroeffector junctions. © 2006 IBRO. Published by Elsevier Ltd. All rights reserved.

Key words: calcium imaging, confocal microscopy, neurotransmission, smooth muscle, ATP, sEJP.

The quantal basis of neurotransmission at the skeletal neuromuscular junction was identified by del Castillo and Katz (1954), in their reanalysis of experiments by Fatt and Katz (1952). They concluded that 'transmission at a nerve-muscle junction takes place in all-or-none ‘quanta’ whose sizes are indicated by the spontaneously occurring miniature (end-plate potential) discharges.' The narrow, Gaussian amplitude distribution of the miniature end-plate potentials (their Fig. 7), the invariance of this distribution as the Ca\(^{2+}\) concentration changed and the existence of a multimodal histogram of excitatory postjunctional potentials imply that evoked neurotransmitter release causes integer multiples of a consistent unitary electrical response. The distribution of packet sizes released from sympathetic postganglionic terminals has been much harder to study because the electrical syncytium that arises through the coupling of smooth muscle cells (SMCs) makes it difficult to distinguish local from distant neurotransmitter release sites during intracellular recording. To tackle this problem, several different approaches have been tried. By differentiating the excitatory junction potential (EJP) it is possible to identify intermittent ‘discrete events’ buried within the depolarization phase (Blakeley and Cunnane, 1979), which indicate the intermittent release of neurotransmitter packets. Some of these discrete events had sizes comparable to the amplitude of the derivative of spontaneous excitatory junction potentials (sEJPs), which argues that the same types of packets contribute to both events, but does not indicate that evoked release is an integral multiple of unitary packets. A further approach is to measure local extracellular potential changes arising following both spontaneous and evoked neurotransmitter release: the excitatory junction currents (sEJCs and EJCs; Brock and Cunnane, 1987). This approach demonstrates intermittent neurotransmitter release with a broad packet size. A development of this approach to study visualized varicosities under conditions of low release probability with small electrode tips (Macleod et al., 1994) demonstrated that, at 1 mM Ca\(^{2+}\), the truncated amplitude distribution of EJPs was broad (their Fig. 3B), arguing against packets of uniform size; that packet size was not uniform was implicit in the model they used to fit the results at higher external Ca\(^{2+}\) concentrations, where the basic packet size varied as a gamma variate (Robinson, 1976). Further evidence for a broad amplitude distribution of the fundamental packet size at the autonomic neuroeffector junction has come from the use of laser-scanning confocal microscopy to image purinergic neuroeffector Ca\(^{2+}\) transients (NCTs) on an impulse-to-impulse basis at individual neuroeffector junctions (Brain et al., 2002, 2003). NCTs arise when intermittently-released packets of ATP activate local P2X receptors, causing a local increase in smooth muscle Ca\(^{2+}\) concentration. The finding that these local Ca\(^{2+}\) transients are amplified by Ca\(^{2+}\)-induced Ca\(^{2+}\) release in the SMC,
however, means that the broad amplitude distribution of NCTs at a single junction cannot be used to imply a broad distribution of basic packets size (Brain et al., 2003).

The mouse vas deferens, unlike that of the guinea pig, provides a good model to study quantal neurotransmission because there is already good evidence that the SMCs are poorly electrically coupled; in particular, it has not been possible to detect electronic potentials spreading from cell-to-cell or across the mouse vas deferens (Holman et al., 1977).

By combining confocal imaging with simultaneous intracellular electrophysiological recording in the mouse isolated vas deferens, it is possible to establish whether a correlation occurs between a traditional electrophysiological approach to monitoring neurotransmitter release, the EJP, and recently-developed optical approaches. Moreover, the combined techniques permit a study of the effective neurotransmitter packet size at the autonomic neuromuscular junction.

**EXPERIMENTAL PROCEDURES**

**Ca$^{2+}$ indicator loading**

Eight- to 12-week-old Balb/c mice (Harlan, Bicester, Oxfordshire, UK) were killed by cervical fracture and both vasa deferentia removed. Efforts were made to minimize the number of animals used and their suffering; all experiments were carried out in accordance with the UK Animals (Scientific Procedures) Act 1986 and European Communities Council Directive 86/609/EEC. The connective tissue around each vas deferens was carefully dissected in order to obtain clear images of SMCs and to remove any ganglia located close to the prostatic end.

Each vas deferens was then exposed to 10 μM Oregon Green 488 1,2-bis(o-aminophenoxy)ethane-N,N,N',N'-tetraacetic acid–1 acetoxymethyl ester (BAPTA-1 AM) (Invitrogen, Paisley, Renfrewshire, UK) in 1% dimethyl sulfoxide/0.2% pluronic F-127 (Sigma-Aldrich, St. Louis, MO, USA) in physiological salt solution (PSS) for 2 h at 36 °C. Each tissue was then cut longitudinally to create a flat sheet and rinsed in PSS, bubbled with 95% O$_2$/5% CO$_2$, for at least 10 min. Tissues were pinned flat, serosal side up in a Sylgard-lined organ bath, and mounted on the stage of an upright confocal microscope. The PSS contained (mM): NaCl 118.4, NaHCO$_3$ 25.0, NaH$_2$PO$_4$ 1.13, KCl 4.7, CaCl$_2$ 1.8, MgCl$_2$ 1.3 and glucose 11.1. The pH was maintained at 7.4 and the solution oxygenated by continuous bubbling with 95% O$_2$/5% CO$_2$.

**Confocal microscopy**

The vas deferens was placed in a chamber that was continuously superfused with standard PSS (bath temperature 33–34 °C). Images were acquired with a Leica SP2 upright confocal microscope (Leica Microsystems, Milton Keynes, UK). A series of 100 or 200 frames was captured at approximately 5 or 13.5 Hz to generate one image set. Such sets were acquired once every minute. Between 8 and 12 such sets were acquired for each SMC. Surface SMCs do not lie perfectly orthogonal to the optical axis of the microscope, and therefore measurement of SMC lengths required this finding to be taken into consideration. When measuring cell lengths, a series of high-resolution (1024×1024 pixels) images was taken at intervals along the z-axis. An average was then taken of the resultant series (‘stack’) of images, and the x-y projected length was calculated using a polygon tool function of Canvas (version 9, ACD Systems, Miami, FL, USA). The z-component was calculated by measuring the focal plane position at each end of the cell, so that a more precise length in three-dimensional space could be calculated using Pythagoras’ theorem.

**Image analysis**

Image analysis was performed with the stack profile function of the Leica LCS software or using an Image J (http://rsb.info.nih.gov/ij/download.html) plug-in written by R. J. Amos. In the first frame of the image series a region of interest was established which encompassed the portion of a SMC visible within the confocal plane. The fluorescent signal in this region was measured over time throughout the image set. Data were exported to Excel (Microsoft, Redmond, WA, USA) for formatting and then to Spike 2 (Cambridge Electronic Design, Cambridge, UK) for analysis in conjunction with electrophysiological recordings.

**Electrophysiology**

Conventional intracellular recording techniques were used to record sEJPs in SMCs (see Brock and Cunnane, 1992). Each vas deferens was superfused with PSS and drugs were applied by swapping the perfusion solution to one containing the drug at the required final bath concentration. Preparations were perfused with prazosin (100 nM; Sigma-Aldrich) and α-latrotoxin (LTX) (25 pM; Sigma-Aldrich) for 30 min prior to recordings. The low concentration of latrotoxin was used to induce a mildly elevated rate of spontaneous neurotransmitter release so that a sufficient number of events could be detected within a fixed imaging period, but not so high that the majority of events could not be independently measured. Microelectrodes (tip resistances of 140–160 MΩ) were filled with Texas Red (2 mM filtered in 5 M potassium acetate; weight 625 Da; Invitrogen). The membrane potential was measured with an Axoclamp 2B (Axon Instruments, Sunnyvale, CA, USA) in bridge mode, in conjunction with a frame-coupled TTL output from the microscope to allow temporal correlation of electrophysiological and confocal recordings.

Recordings of electrically-evoked EJPs were achieved by applying rectangular pulses (0.6 ms duration; voltage amplitude at twice the threshold for eliciting EJPs, typically around 20 V) delivered through Ag/AgCl electrodes positioned around the prostatic end of the vas deferens. In some experiments, ‘non-surface’ SMCs were impaled using an almost vertical electrode approach (in the absence of confocal microscopy) with no attention paid to remaining close to the serosal surface.

In additional experiments, surface SMCs were voltage-clamped at various values of alternating depolarization and hyperpolarization with respect to their resting membrane potential (mean ~ 65.1±2 mV) for 200 ms, with approximately 10 s between steps. The input resistance was calculated for the linear part of the voltage–current relationship (around the resting membrane potential). Both inward and outward currents were identified during depolarizing steps (in two of four cells), and in one cell, an inward current was observed following a hyperpolarizing step (data not shown). These phenomena were not further investigated as part of the present study.

The voltage and current were digitized (at 5 kHz and 1 kHz, respectively) with a PowerLab system (ADInstruments, Chalgrove, UK). Recordings of sEJPs were exported from Chart 4.2 (ADInstruments) for analysis with Spike 2 (ADInstruments). The microelectrode was always within the field of view of the confocal image. On completion of each experiment, the SMC recorded from was labeled by injecting Texas Red by iontophoresis (current injection, +2 nA, 15 s).

**Analysis of correlations between NCTs and sEJPs**

Analysis of the amplitudes of sEJPs and NCTs was performed on data where confocal images were acquired at 5 Hz. To obtain a...
more accurate measurement of the temporal relationship between sEJPs and NCTs, a faster acquisition rate of approximately 13.5 Hz (74 ms frame duration) was used. The peak amplitudes and timings of both sEJPs and NCTs were calculated using a custom-written script for Spike 2. The occurrence of sEJPs was defined by their amplitude ($V_i$). The threshold for determining NCTs varied between recordings from different SMCs. The threshold was adjusted until it provided a specificity comparable to manual counting ($F_i$ range of 0.05–0.1).

To investigate the temporal correlation between changes in the fluorescent signal and sEJPs, the time from each change in fluorescent signal to the nearest sEJP was measured (referred to as delay). To determine whether these events displayed a significant temporal correlation, the distribution of delays was compared (using chi-square tests) with the expected distribution under the assumption that sEJPs and NCTs were uncorrelated. The expected distribution of delays is the double exponential or Laplace distribution,

$$P = \frac{1}{2\nu} \ln\left(\frac{1}{1-2nd}\right)$$

for $0 < nd < 0.5$. As the Laplace distribution is an even function (i.e. symmetrical around the y axis),

$$t_{n+} = t_n$$

To test the validity of using the Laplace distribution to model expected temporal delays between sEJPs and NCTs, the electrophysiological record in each experiment was shifted by a random increment relative to the fluorescence trace (at least 5 s away from the raw trace; a circular boundary condition was applied) and both the median delay and the chi-square value were recalculated. As such a shift is significantly greater than $1/\nu$ then a shift should remove any temporal correlation. In all cases, there was no resulting significant difference between the resulting observed delays (where sEJP had been shifted) and the expected delays as predicted by the Laplace distribution ($P > 0.2$, $n = 5$).

Occasionally, sEJPs occurred with a high local frequency. Correlating the amplitude and timing of these sEJPs with the corresponding $Ca^{2+}$ fluorescence was difficult due to the long time course of NCTs, such that several sEJPs were often observed to correlate with what appeared to be a single NCT. For this reason, such events were not included in the analysis.

**Statistical analysis**

For other statistical tests, the normality and homogeneity of variance were tested prior to statistical analysis. Fitting of a third-order polynomial to the I:V relationship of surface SMCs (Fig. 1B) was performed using GraphPad Prism 4.0b (GraphPad Software Inc., San Diego, CA, USA). To test the variance of evoked EJP amplitudes in surface and deeper cells using the $F$ test, the EJP amplitudes were normalized to the mean amplitude per cell. The term $n$, used in the presentation of statistical analyses throughout, refers to the number of animals.

**RESULTS**

**EJPs and electrical coupling**

Surface SMCs were voltage-clamped to determine their input resistance, to give an indirect measure of their elec-

Fig. 1. Voltage-clamp of surface SMCs. (A) Cells were voltage-clamped at various values with respect to their resting membrane potential. Data were down-sampled to 100 Hz for presentation. (B) The resulting current–voltage relationship ($n = 4$) is centered around the mean resting membrane potential, which was $-65.1 \pm 2$ mV. Points plotted are mean ± S.E.M. The fitted line is a third-order polynomial.

Fig. 2. The amplitude of electrically-evoked EJPs is more variable in surface SMCs, than non-surface SMCs. Within each surface SMC, electrically-evoked EJPs are more variable in amplitude ($n=6$) than those from non-surface (i.e. deeper) SMCs ($n=6$). For more traditional amplitude frequency histograms, see Supplementary Fig. 1.
trical coupling. The mean resistance was 176±18 MΩ (mean±S.E.M., range: 141–221 MΩ; n=4; Fig. 1).

Another test of electrical coupling is the variability of EJP amplitude. Electrically-evoked EJPs were recorded from surface and non-surface SMCs (Fig. 2). Electrically-evoked EJPs from surface SMCs were more variable in amplitude (S.D. =13.4 mV; n=6) than those recorded from non-surface (i.e. deeper) SMCs (S.D. =8.2 mV; n=6) (F test, F_{244,542} =17.4, P<0.0001).

LTX-stimulated sEJPs and NCTs

An extract of the black widow spider venom, LTX was used to increase sEJP and NCT frequency, so that a sufficient number of events could be recorded before the Ca^{2+} indicator was photobleached. LTX (25 pM)-stimulated sEJPs in surface SMCs had a right-skewed distribution (Fig. 3), ranging from 2 mV (the threshold for detection) to 37 mV (n=4). Superfusion of LTX (25 pM) for 30 min, in the presence of the α₁-adrenoceptor antagonist prazosin (100 nM), increased the frequency of spontaneous NCTs to 220±70% of controls (P<0.05, n=6, one-tailed paired t-test; NCT frequency measurement according to Brain et al., 2003) but did not affect their amplitudes (amplitudes 111±14% of controls, P=NS, n=6, two-tailed paired t-test). NCTs have previously been shown to require the activation of P2X₁ receptors (Brain et al., 2002). After applying the P2X₁ antagonist 4,4',4''-carbonylbis[iminoo-5,1,3-benzenetriyl bis(carbonyl-imino)]tetrakis(benzene-1,3-disulfonic acid) (NF449) (1 μM; Tocris, UK) for 1 h, the amplitude and frequency of latrotoxin-stimulated spontaneous NCTs were reduced by 59±5% and 77±8%, respectively (amplitude: P<0.05; frequency: P<0.01; n=6, one-tailed paired t-test for both comparisons). The presence of residual NCTs following superfusion of NF449 prompted a study of the effect of NF449 on EJPs. Even after 1 h exposure to a higher concentration of NF449 (10 μM), the amplitude of EJPs was only reduced by 69±9% of controls (P<0.01, n=6, two-tailed paired t-test).

To determine whether latrotoxin might have a direct effect on smooth muscle membrane conductance, the resting membrane potential of surface SMCs was measured. During a control period the membrane potential was −66±6 mV (mean±S.D., n=4), and following superfusion of LTX, the resting membrane potential (measured in between depolarizations) was −68.6±2 mV (n=4).
Simultaneous electrophysiology and confocal imaging was used to determine the relationship between the well-established method for monitoring neurotransmitter release, electrophysiology, with NCTs. Focal increases in intracellular Ca\(^{2+}\) concentration (Fig. 4A) were coincident with sEJPs (Fig. 4B, C). The time from the peak of each NCT to the nearest sEJP was measured; the median time between the peak of an NCT and the peak of the nearest sEJP ranged from \(-30\) to \(-57\) ms (n=5; Table 1). The frequency distribution of temporal relationships between sEJPs and NCTs was significantly different from an expected distribution, as modeled using the Laplace distribution (chi-square, \(P<0.05\) in all experiments, n=5; Fig. 5, Table 1).

Many, but not all, sEJPs were associated with NCTs in the monitored region (174 NCTs with 413 sEJPs were sampled from n=4 experiments). However, the entire SMC could not be simultaneously viewed within one confocal optical section (mean field: 94 \(\mu m\), range 81–100 \(\mu m\); mean SMC length: 204±9 \(\mu m\); 28 sites from n=6). Correcting for the average SMC length gives an estimated 0.91 NCTs per sEJP. The number of NCTs recorded (174) is not significantly different from that expected under the hypothesis that all NCTs arise from the cell recorded from (95% confidence interval, 163–217 NCTs; making no assumption about the distribution, but applying the Central Limit Theorem for large n), i.e. these measurements are consistent with each sEJP being coincident with a NCT occurring somewhere within the SMC.

If the surface SMCs are electrically-coupled, there should be a temporal correlation between sEJPs and NCTs in the adjacent cell. There was no temporal correlation between the occurrence of NCTs in adjacent SMCs (see Fig. 6 for an example). Simultaneous electrophysiology and confocal imaging revealed a good temporal correlation between sEJPs and NCTs in the impaled cell (Fig. 6Aii, B) but no correlation between NCTs in a neighboring cell (Fig. 6Ai) and either NCTs or sEJPs of the impaled cell (Fig. 6Aii, B). It was rare to observe coincident NCTs in adjacent cells. Such occurrences were often at sites where a nerve branch ran between two SMCs and have previously been suggested to arise when ATP released from a varicosity binds to P2X\(_1\) receptors on more than one SMC (Brain et al., 2002).

There was a correlation between the amplitudes of NCTs and sEJPs, both for large and small amplitude events (Pearson product moment correlation, \(\rho>0.55\) and 0.71; Table 1). The temporal relationship between NCTs and sEJPs (n=5) was not normally distributed, as modeled by the Laplace distribution (chi-square, \(P<0.05\) in all experiments, n=5; Table 1).

### Table 1. The temporal relationship between NCTs and sEJPs (n=5)

| Characteristic | i   | ii  | iii | iv  | v   |
|----------------|-----|-----|-----|-----|-----|
| No. of NCTs   | 36  | 36  | 24  | 12  | 54  |
| Median peak   | -57 | -46 | -42 | -37 | -30 |
| NCT vs. peak sEJP (ms) | 25, 75% Quartiles (ms) | <0.001 |
| Chi-square test, \(P\) | -122, 77 | <0.001 | -91, 13 | <0.0001 | -74, 10 | <0.05 | -71, 24 | <0.0001 |

Fig. 5. The frequency distribution of temporal delays between sEJPs and NCTs. Data are presented for a typical experiment (filled bars) and expected values (open bars), as modeled by the Laplace distribution. The boundaries between intervals are shown at one frame width (74 ms) for presentation; for analysis, they were set to ensure the expected number of events remained constant in all bins.

Fig. 6. NCTs present in a single SMC do not correlate with sEJPs or NCTs recorded from an adjacent cell. (A) Ca\(^{2+}\) imaging of two adjacent SMCs (i and ii, inset). (B) Intracellular recording corresponding to SMC ii. Fluorescent images were acquired at 13.5 Hz.
NCT amplitudes were binned and the mean amplitude of coincident normalized to a percentage of the maximum value in each experiment. The amplitudes were arise from the same event and that the variability in sEJP amplitude where both events could be simultaneously measured, suggests that they arise from the same event and that the variability in sEJP amplitude represents variability in the local action of ATP. The amplitudes were normalized to a percentage of the maximum value in each experiment. NCT amplitudes were binned and the mean amplitude of coincident sEJPs for each bin was plotted. sEJP amplitudes are mean ± S.E.M. for each preparation. Lines were fitted to the binned data; the solid lines relate to the filled symbols and the dashed lines to the open symbols.

\( P < 0.001 \) for each of \( n = 4 \) experiments, unbinned data) (Fig. 7; for unbinned data see Supplementary Fig. 2).

**DISCUSSION**

The hypothesis that surface SMCs are poorly coupled is supported by several observations. First, input resistances of surface SMCs (176 ± 18 MΩ, range: 141–221 MΩ) are comparable to dispersed cells (331 ± 43 MΩ; Blakeley et al., 1989). Input impedance for syncytial smooth muscle lies in the range of 5–30 MΩ (Bennett, 1967; Holman et al., 1977; Manchanda and Venkateswarlu, 1999). Second, the amplitude of evoked EJPs in surface SMCs is shown to be more variable (S.D. = 13.4 mV) than those from non-surface (i.e. deeper) SMCs (S.D. = 8.2 mV). An increase of EJP variability has been previously reported in the guinea-pig vas deferens when SMCs are uncoupled with heptanol (Manchanda and Venkateswarlu, 1999). Third, the resting membrane potential (−66 mV) is more positive than that of SMCs recorded elsewhere, being similar to values recorded from dissociated cells (−45 mV; Blakeley et al., 1989).

The variable amplitude of sEJPs has been attributed to the electronic attenuation of depolarizations originating from distant release sites i.e. on neighboring cells (Tomita, 1970; Purves, 1976). Evidence for quantal neurotransmission from sympathetic nerves without the confounding effects of electrotonic attenuation comes from the work of Hirst and Neild (1980). Using short segments of guinea-pig isolated gut submucosal arterioles, they reported that the distribution of sEJPs was monophasic but highly right-skewed, particularly after deconvolving to account for recording noise (see their Fig. 3). In our experience, such a distribution is difficult to distinguish from an exponential distribution when sEJPs are counted manually, as it is difficult to identify small amplitude events close to the noise. We did not observe a similar distribution of sEJPs in the surface cells of the vas deferens, despite the fact that the expected number of close contact varicosities should be smaller (i.e. those contacting just one SMC, rather than the 100–200 varicosities in the short arteriole segments). It is possible that the signal-to-noise ratio in the present work was not sufficient to identify a peak in the amplitude-frequency histogram, but our experiments do confirm a highly skewed rather than Gaussian distribution of sEJPs.

In this sense there is agreement that neurotransmitter release cannot be the sum of units of constant size. Hirst and Neild (1980) also argued that the EJP amplitude distribution could be constructed as an arbitrarily weighted sum of integral multiples of the sEJP distribution, but they themselves noted that such a construction was not possible if the raw data were deconvolved to correct for electrical noise, and they also noted that the arbitrary weighting in the sum did not yield the expected binary distribution of amplitude coefficients. These two observations imply that even in their work the EJP could not be explained as the sum of independently released neurotransmitter packets.

It is unlikely that the skewed frequency distribution of sEJP amplitudes can be explained by variable distances between varicosities and the electrode; this would result in a weak correlation or no correlation between the amplitudes of NCTs and sEJPs, rather than the strong positive correlation reported. An alternative model for neurotransmission (the modular hypothesis; Edwards et al., 1990) where neurotransmitter packets act on small clusters of readily-saturated receptors, is not supported by the findings of a broad sEJP amplitude distribution and no evidence of a multimodal EJP amplitude distribution exists. Bennett et al. (1995) similarly concluded, from their own Monte Carlo simulations, that the modular hypothesis is not tenable at this junction.

In addition to temporal coupling, NCTs and sEJPs were also correlated in amplitude throughout the amplitude range. It has been suggested that spontaneous NCTs only arise from the release of the contents of giant dense-cored vesicles (Blair et al., 2003) rather than all vesicle types. This now seems unlikely given the good temporal (and hence frequency) correlation between sEJPs and NCTs described in the present work. It is improbable that both sEJPs and NCTs reflect release from only one population of large vesicles. Others have shown that more than one basic mechanism of ATP release occurs from terminals in the mouse vas deferens by using peptide fragments of α-soluble N-ethylmaleimide-sensitive factor attachment protein (α-SNAP) (Bennett et al., 2001), and it will be of interest to see whether it is this SNAP-dependent pathway that generates NCTs. The present observations do not exclude the possibility of fractional release of the neurotransmitter contents of

![Fig. 7. Amplitudes of sEJPs and NCTs are positively correlated (Pearson product moment correlation, \( P < 0.001 \) for each of \( n = 4 \) experiments, unbinned data). The amplitude correlation between sEJPs and NCTs, where both events could be simultaneously measured, suggests that they arise from the same event and that the variability in sEJP amplitude represents variability in the local action of ATP. The amplitudes were normalized to a percentage of the maximum value in each experiment. NCT amplitudes were binned and the mean amplitude of coincident sEJPs for each bin was plotted. sEJP amplitudes are mean ± S.E.M. for each preparation. Lines were fitted to the binned data; the solid lines relate to the filled symbols and the dashed lines to the open symbols.](image-url)
demonstrates that focal increases in intracellular Ca
confocal microscopy acquisition rate) the present study
for poor electrical coupling between surface SMCs.
porally coupled. These findings provide further evidence
for poor electrical coupling between surface SMCs.
Within the limits of our recording system (i.e. 13.5 Hz
confocal microscopy acquisition rate) the present study
demonstrates that focal increases in intracellular Ca
are tightly temporally coupled to the electrical sign of sponta-
neous neurotransmitter release in this tissue; i.e. the sEJP.
Hence, the present work validates the use of NCTs to
monitor neurotransmitter release, with the proviso that
some NCTs may arise from the superposition of bursts of spontaneously-released neurotransmitter packets. The
rate at which images were sampled limits the temporal
resolution with which sEJP recordings can be compared
with NCTs. Using line scanning confocal microscopy it has
been reported that electrically-evoked NCTs can be de-
tected 6 ms after stimulation (Brain et al., 2002), suggest-
ing that the real delay between the sEJP and NCT is much
briefer than the range of values, ∼30 to ∼57 ms, reported
here (n=5; Table 1). NCTs were not generated by the
activation of α1-adrenoceptors because they were ob-
served in the presence of the α1-adrenoceptor antagonist
prazosin (100 nM) as previously described (Brain et al.,
2002). The observation that NF449 greatly reduced NCT
amplitude and frequency (presumably as the postjunc-
tional response fell below the detection threshold), is con-
sistent with previous reports that NCTs are abolished or
greatly reduced by α,β-methylene ATP (Brain et al., 2002,
2003), showing that they reflect the action of ATP at P2X
receptors. The observation that NCTs remain in the pres-
ence of the P2X, antagonist NF449 (1 μM) is consistent
with the observation that the desensitizing agonist α,β-
methylene ATP does not abolish all excitatory junctional
potentials (or currents; Alcorn et al., 1986; Liang et al.
2000) or NCTs (Brain et al., 2003), and more importantly
that NF449 did not abolish EJPs after 1 h of exposure (the
present study). Given that EJPs do not occur in P2X1
knockout mice (Mulryan et al., 2000) it may be that there is
a proportion of sites that are either insensitive to these
drugs or inaccessible within the incubation time used.
The time course of NCT recovery (1.8 s; 174 NCTs,
n=4) was significantly slower than previously reported,
when the fluorescent signal was measured over a smaller
area (280 ms; Brain et al., 2002). This dependence of the
time course on the area in which the signal is measured is
consistent with most models of near-membrane Ca\(^{2+}\)
kinetics in SMCs (Kargacin and Fay, 1991). Also, the dura-
tion of the NCT is consistent with observations from the
urinary bladder (Heppner et al., 2005). Occasional bursts
of sEJPs led to a fused local Ca\(^{2+}\) transient that could
easily be interpreted as one NCT. This latter finding sug-
gests that the amplitude distribution of spontaneous NCTs
cannot be relied upon to reflect the response of single
packets of neurotransmitter unless simultaneous electrophysiological recording is used or the temporal resolution
of the optical detection system is improved.

The amplitude of NCTs at a given junction can vary by
more than ninefold (Brain et al., 2002) but given that a
significant proportion of the amplitude is due to (potentially
highly variable) amplification by Ca\(^{2+}\)-induced Ca\(^{2+}\)
release (Brain et al., 2003), it is not possible to attribute
variation in the NCT amplitude solely to variability in the
neurotransmitter packet size, or even to variation in the
number of P2X receptors opened. For this reason, mea-
suring the amplitude distribution of sEJPs is a more direct
measure of variation in neurotransmitter packet size than
the measurement of NCT amplitudes.

### Latrotoxin as a tool to study spontaneous neurotransmitter release

Both crude black widow spider venom and purified LTX
have been extensively used to study molecular neuro-
transmission in vertebrates (e.g. Ceccarelli and Hurlbut,
1980; Hurlbut et al., 1990; Auger and Marty, 1997) with
the established LTX concentration used often in the low nano-
molar range (1–2 nM; Hurlbut et al., 1994; Davletov et al.,
1998; Silva et al., 2005). Yet low picomolar concentrations
(25 pM, the present study; 130 pM, McMahon et al., 1990)
have been shown to increase marginally the rate of neu-
rotransmitter release. In brain synaptosomes (Nicholls
et al., 1982) LTX increased the rate of neurotransmitter re-
lease even in the presence of the Na\(^+\) channel blocker
tetrodotoxin, implying that its action is independent of so-
dium-dependent nerve action potentials. Despite these
very low concentrations it is possible that LTX-stimulated
eJPs do not have the same properties as true sEJPs.
While there was no effect of this low concentration of LTX
on the resting membrane potential, and no direct SMC
effects have been reported at this concentration, it is pos-
sible that latrotoxin caused a small conductance change
which was well balanced by other conductances. This
possibility was not investigated.

### Variability in the electrical properties of surface SMCs

Despite being depolarized to greater than 0 mV during
voltage clamp, only two of four cells exhibited active
currents. This variability in the cells’ responses to depo-
larizing steps contrasts to the opening of both inward
and outward currents in response to depolarizing voltage
steps reported by Blakeley and colleagues (1989) but is
consistent with the observations of Holman and colleagues
1977; compare their Fig. 3A and 3B). Variability in the electrical properties of surface SMCs is also
demonstrated in recorded resting membrane potentials:
−40 to −80 mV (Blakeley et al., 1989) and −60 to −72
mV (the present study). A likely explanation for the
variance in resting membrane potential, and responses

to depolarizing voltage steps of surface SMCs, is a varying degree of electrical coupling. Moreover, ‘coupled’ and ‘uncoupled’ represent two ends of a spectrum; surface cells are somewhere toward the ‘electrically-uncoupled’ extreme.

Simultaneous electrophysiological recording and confocal microscopy now reveals that NCTs are coupled in timing and in amplitude with sEJPs, implying that NCTs provide a sensitive, high-resolution technique to monitor spontaneous neurotransmitter release. The observation of a wide and skewed amplitude distribution of sEJPs in cells that are very poorly coupled weakens the argument for neurotransmission of a uniform packet size at autonomic neuroeffector junctions. It is likely that each packet corresponds to the release of the neurotransmitter contents of one vesicle, but whether the variability in the postjunctional response to neurotransmitter packets results from variable neurotransmitter release, or from variable postjunctional receptor responses to consistent neurotransmitter release, remains to be established.

Acknowledgments—The Wellcome Trust provided financial support (069768 to T.C.C.; Research Fellowship to K.L.B.).

REFERENCES

Alcorn RJ, Cunnane TC, Kirkpatrick K (1986) Actions of alpha, beta-methylene ATP and 6-hydroxydopamine on sympathetic neurotransmission in the vas deferens of the guinea-pig, rat and mouse: support for cotransmission. Br J Pharmacol 89:647–659.

Auger C, Marty A (1997) Heterogeneity of functional synaptic parameters among single release sites. Neuron 19:139–150.

Bennett MR (1967) The effect of intracellular current pulses in smooth muscle cells of the guinea-pig vas deferens at rest and during neurotransmission. J Gen Physiol 50:2459–2470.

Bennett MR, Farnell L, Gibson WG, Karunanithi S (1995) Quantal transmission at purinergic junctions: stochastic interaction between ATP and its receptors. Biophys J 68:925–935.

Bennett MR, Farnell L, Gibson WG, Lin YQ, Blair DH (2001) Quantal and non-quantal current and potential fields around individual sympathetic varicocities on release of ATP. Biophys J 80:1311–1328.

Blair DH, Lin YQ, Bennett MR (2003) Differential sensitivity to calcium and osmotic pressure of fast and slow ATP currents at sympathetic varicocities in mouse vas deferens. Auton Neurosci 105:45–52.

Blakeley AG, Cunnane TC (1979) The packeted release of transmitter from the sympathetic nerves of the guinea-pig vas deferens: an electrophysiological study. J Physiol 296:85–96.

Blakeley AG, Dunn PH, Petersen SA (1989) Properties of excitatory junction potentials and currents in smooth muscle cells of the mouse vas deferens. J Auton Nerv Syst 27:47–56.

Brain KL, Jackson VM, Trout SJ, Cunnane TC (2002) Intermittent ATP release from nerve terminals elicits focal smooth muscle Ca

2

transients in mouse vas deferens. J Physiol 541:849–862.

Brain KL, Cuprian AM, Williams DJ, Cunnane TC (2003) The sources and sequestration of Ca

2

contribution to neuroeffector Ca

2

transients in the mouse vas deferens. J Physiol 553:627–635.

Brock JA, Cunnane TC (1987) Relationship between the nerve action potential and transmitter release from sympathetic postganglionic nerve terminals. Nature 326:605–607.

Brock JA, Cunnane TC (1992) Electrophysiology of neuroeffector transmission in smooth muscle. In: Autonomic neuroeffector mechanisms (Burnstock G, Hoyle CHV, eds), pp 121–213. Melbourne, Australia: Harwood Academic Publishers.

Ceccarelli B, Hurbut WP, Mauro P (1972) Depletion of vesicles from frog neuromuscular junctions by prolonged tetanic stimulation. J Cell Biol 54:30–38.

Ceccarelli B, Hurbut WP (1980) Ca

2

-dependent recycling of synaptic vesicles at the frog neuromuscular junction. J Cell Biol 87:297–303.

Davletov BA, Meunier FA, Ashton AC, Matsushita H, Hirst WD, Lelianova VG, Wilkin GP, Dolly JO, Ushkaryov YA (1998) Vesicle exocytosis mediated by alpha-latrotoxin is mediated by latrophilin and requires both external and stored Ca

2

. EMBO J 17:3909–3920.

del Castillo J, Katz B (1954) Quantal components of the endplate potential. J Physiol 124:560–573.

Edwards FA, Konnerth A, Sakmann B (1990) Quantal analysis of inhibitory synaptic transmission in the dentate gyrus of rat hippocampal slices: a patch-clamp study. J Physiol 430:213–249.

Fatt P, Katz B (1952) Spontaneous subthreshold activity at motor nerve endings. J Physiol 117:109–128.

Harata NC, Choi S, Pyle JL, Aravanis AM, Tsien RW (2006) Frequency-dependent kinetics and prevalence of kiss-and-reuse and reuse at hippocampal synapses studied with novel quenching methods. Neuron 49:243–256.

Heppner TJ, Bone AD, Nelson MT (2005) Elementary purinergic Ca

2

transients evoked by nerve stimulation in rat urinary bladder smooth muscle. J Physiol 564:201–212.

Hirst GD, Neild TO (1989) Some properties of spontaneous excitatory junction potentials recorded from arterioles of guinea-pigs. J Physiol 303:43–60.

Holman ME, Taylor GS, Tomita T (1977) Some properties of the smooth muscle of mouse vas deferens. J Physiol 266:751–764.

Hurbut WP, Chiergatti E, Valtorta F, Haimann C (1994) Alpha-latrotoxin channels in neuroblastoma cells. J Membr Biol 138:91–102.

Hurbut WP, Lezzi N, Fesce R, Ceccarelli B (1990) Correlation between quantal secretion and vesicle loss at the frog neuromuscular junction. J Physiol 425:501–526.

Kargacin G, Fay FS (1991) Ca

2

movement in smooth muscle cells studied with one- and two-dimensional diffusion models. Biophys J 60:1088–1100.

Liang SX, D’Arbe M, Phillips WD, Lavidis NA (2000) Development of fast purinergic transmission in the mouse vas deferens. Synapse 37:283–291.

Macleod GT, Lavidis NA, Bennett MR (1994) Calcium dependence of quantal secretion from visualized sympathetic nerve varicocities on the mouse vas deferens. J Physiol 480:61–70.

Manchanda R, Venkateswarlu K (1999) Quantal evoked depolarizations underlying the excitatory junction potential of the guinea-pig isolated vas deferens. J Physiol 520:527–537.

McMahon HT, Rosenthal L, Meldolesi J, Nicholls DG (1990) α-Latrotoxin releases both vesicular and cytoplasmic glutamate from isolated nerve terminals. J Neurochem 55:2039–2047.

Mulryan K, Gitterman DP, Lewis CJ, Vial C, Leckie BJ, Cobb AL, Brown JE, Conley EC, Buell G, Pritchard CA, Evans RJ (2000) Reduced vas deferens contraction and male infertility in mice lacking P2X1 receptors. Nature 403:86–89.

Nicholls DG, Rugolo M, Scott IG, Meldolesi J (1982) Alpha-latrotoxin of the black widow spider venom depolarizes the plasma membrane, induces massive calcium influx, and stimulates transmitter release in guinea pig brain synaptosomes. Proc Natl Acad Sci U S A 79:7924–7928.

Purves RD (1976) Current flow and potential in a three-dimensional syncytium. J Theor Biol 60:147–162.

Robinson J (1976) Estimation of parameters for a model of transmitter release at synapses. Biometrics 32:61–68.
Silva AM, Liu-Gentry J, Dickey AS, Barnett DW, Misler S (2005) Alpha-latrotoxin increases spontaneous and depolarization-evoked exocytosis from pancreatic islet beta-cells. J Physiol 565: 783–799.

Tomita T (1970) Electrical properties of mammalian smooth muscle. In: Smooth muscle (Bülbring E, Brading AF, Jones A, Tomita T, eds), pp 197–243. London: Edward Arnold.

APPENDIX

Supplementary data

Supplementary data associated with this article can be found in the online version, at doi: 10.1016/j.neuroscience.2006.11.054.

(Accepted 28 November 2006)
(Available online 5 January 2007)