Review

Substance Use Disorder in the COVID-19 Pandemic: A Systematic Review of Vulnerabilities and Complications

Yufeng Wei 1,* and Rameen Shah 1

1 Department of Chemistry, New Jersey City University, Jersey City, NJ 07305, USA
* Correspondence: ywei@ywei@njcu.edu; Tel.: +1-201-200-2309

Abstract: As the world endures the coronavirus disease 2019 (COVID-19) pandemic, conditions of 35 million vulnerable individuals struggling with substance use disorders (SUDs) worldwide have not received sufficient attention for their special health and medical needs. Many of these individuals are complicated by underlying health conditions, such as cardiovascular and lung diseases and undermined immune systems. During the pandemic, access to the healthcare systems and support groups is greatly diminished. Current research on COVID-19 has not addressed the unique challenges facing individuals with SUDs, including the heightened vulnerability and susceptibility to the disease. In this systematic review, we will discuss the pathogenesis and pathology of COVID-19, and highlight potential risk factors and complications to these individuals. We will also provide insights and considerations for COVID-19 treatment and prevention in patients with SUDs.

Keywords: COVID-19; SARS-CoV-2; Coronavirus; Substance Use Disorder (SUD); Immunology; Neuroinflammation; Blood-Brain Barrier (BBB); hypothalamic–pituitary–adrenal (HPA) axis

1. Introduction

The novel coronavirus 2019, now officially termed as SARS-CoV-2, causes the coronavirus disease 2019 (COVID-19) by infecting the respiratory system [1]. The disease was first detected and reported in Wuhan, China, in December 2019 [2], and has now spread to over 150 countries of all continents except Antarctica. According to the World Health Organization (WHO) Coronavirus Disease Dashboard, the global tally of coronavirus cases has passed 10 million with the death toll approaching 500,000 [3]. The United States alone has counted over 2.4 million infections and over 120,000 deaths, contributing more than a quarter of both overall infections and deaths globally (John Hopkins University Coronavirus Resource Center) [4]. The WHO characterized the COVID-19 as a pandemic on March 11, 2020 [5]. The fatality rate of the disease is particularly high among patients who are older and who have underlying health issues, such as cancer, diabetes, and compromised lung function or lung disease.

The United Nations (UN) reported that 35 million people worldwide suffer from substance use disorders (SUDs) [6]. In the U.S., the number of individuals experiencing SUDs is 20.3 million [7]. A large number of individuals with SUDs have underlying health conditions, particularly cardiovascular and lung diseases and hepatitis C or HIV-1 infections. Together with complicated socioeconomic issues, these populations are particularly vulnerable to COVID-19 [8]. Despite the fact that researchers and clinicians around the world have collected and disseminated tremendous amounts of data on COVID-19, we have very little knowledge of the interactions and comorbidity of COVID-19 and SUD. In this review, we will analyze relevant COVID-19 and SUD literatures, and highlight the susceptibilities and vulnerabilities for individuals with SUD.
SARS-CoV-2 is known to attack the respiratory tract and could lead to severe lung damage or pulmonary fibrosis [2]. Smokers of tobacco and marijuana, and possibly people who vape, are susceptible to chronic obstructive pulmonary disease (COPD) [9,10], which could cause severe complications of COVID-19 and lead to a higher fatality rate [11,12]. Other substances of abuse, such as opioids and methamphetamine, function increasingly through the brain and immune system to indirectly affect the respiratory system. Opioid use disorder (OUD) tends to slow breath rate and decrease blood oxygen content (hypoxemia). An extended period of hypoxemia is one of the major causes of overdose fatality. COVID-19, which shrinks lung capacity, could heighten the condition caused by opioid overdose [13,14]. Methamphetamine may contribute to pulmonary hypertension and edema through cardiomyopathy and restricted blood circulation [15]. Many substances of abuse also impair the bidirectional interactions between the brain and immune responses, resulting in an increase in the infection rates among individuals with SUDs. Some drugs exert proinflammatory effects in the central nervous system (CNS), leading to neuroinflammation. The buildup of proinflammatory cytokines and chemokines in the CNS may exaggerate the already excessive inflammatory response in the peripheral tissues of COVID-19 patients.

2. How does SARS-CoV-2 infect human cells and cause COVID-19?

Coronaviruses are named for their crown-like spikes protruding from the surface of the virion, and can be classified into four genera, known as α, β, γ, and δ. There are seven known coronaviruses that can infect humans. Four of them, α-coronaviruses 229E and NL63, and β-coronaviruses OC63 and HKU1, infect people on a regular basis and cause common cold symptoms. The other three, all belonging to the β-coronavirus subfamily, are believed to have originated from bats and evolved through jumping animal species to become new human coronaviruses [16,17]. These include the SARS-CoV, the coronavirus that caused severe acute respiratory syndrome (SARS) outbreak in 2003, the MERS-CoV, the coronavirus that caused Middle East Respiratory Syndrome (MERS) outbreak in 2012, and the SARS-CoV-2, the novel coronavirus that is causing the current COVID-19 pandemic razing the whole world.

Like all other coronaviruses, SARS-CoV-2 contains a single-strand (ss) RNA genome. The sequence of the 29,903 nucleotides (nt) long SARS-CoV-2 genome was first reported by Chinese scientists and made publicly available on GenBank with accession number MN908947 [18]. The genome organization of SARS-CoV-2 is similar to that of other representative β-coronaviruses. It is comprised of a 5'-untranslated region (UTR), and at least ten open reading frames (ORFs) that encode non-structural and structural viral proteins. The first ORF is the 21,291-nt replicase gene, ORF1ab, encoding 16 non-structural proteins (NSP1-16). The subsequent ORFs encode four structural proteins, spike (S), envelope (E), membrane (M), and nucleocapsid (N), as well as several accessory proteins that do not participate in viral replication and transcription. Structural proteins are important in maintaining viral structure and stability. The S protein is responsible for infection and transmission. Non-structural proteins, such as the RNA-dependent RNA polymerase (NSP12), have functions in viral genome transcription and replication. The SARS-CoV-2 genome was found to have 79.6% sequence similarity to SARS-CoV, and shares 96% identity at the whole-genome level to a bat coronavirus (Bat CoV RaTG13) detected in Rhinolophus affinis [17].

Similar to SARS-CoV, SARS-CoV-2 recognizes the angiotensin converting enzyme 2 (ACE2) receptor by its S protein and utilizes it for cell entry [17,19]. The heavily glycosylated S protein triggers virus cell entry by fusing the receptor binding domain (RBD) on the S1 subunit to the host ACE2 receptor, engaging the transition of S2 subunit to a stable post-fusion conformation [20]. Cryo-electron microscopy (EM) structures of the pre-fusion [20] and post-fusion structures [21] of the S protein have been reported. The SARS-CoV-2 S protein has been shown to have a much higher binding affinity to the ACE2 than the SARS-CoV S protein [20,22]. The S protein contains 22 N-linked glycans, and the complex glycosylation is likely to play a role in shielding and camouflaging for immune evasion of the virus [23,24]. The S protein is activated by type II transmembrane serine protease (TMPRSS2), a host protease co-expressed with ACE2 on the cell surface [21,25]. In cells not
expressing TMPRSS2, other proteases, such as cathepsin B/L, may activate the S protein and facilitate viral entry [26].

Upon cell entry, SARS-CoV-2 has a similar life cycle and pathogenesis as other β-coronaviruses, including SARS-CoV and MERS-CoV [27]. Upon ACE2 receptor binding, the virus fuses its membrane with the host cell plasma membrane, releasing its genomic RNA into the cytoplasm. Since the viral RNA is similar to the human messenger RNA (mRNA), it triggers the host ribosome to start translating the viral RNA and producing viral proteins. The viral replicate ORF is translated into two overlapping polyproteins, PP1a and PP1ab, which require extensive processing. The 33.8-kDa main viral protease (M(pro)), also referred to as the 3-chymotrypsin-like protease (3CL(pro)), performs the function by autolytic cleavage of the protease itself, and then subsequently digests the polyproteins into 16 non-structural proteins (NSP1 – NSP16). One of these 16 NSPs, NSP12, known as the RNA-dependent RNA polymerase (RdRp), together with NSP7 and NSP8, carries out the critical process of the viral RNA synthesis, and is central to the viral replication and transcription cycle. The N-terminal non-structural protein, NSP1, has been shown to bind to the 40S small ribosomal subunit, shutting down all host cell protein production by blocking the mRNA entry tunnel. NSP1 binding to ribosomes and blocking host cell translation effectively inhibits type-I interferon (IFN-I)-induced innate immune response by turning off the retinoic acid-inducible gene (RIG)-I antiviral sensor [28]. The inhibition of the IFN-I-induced innate immunity allows the assembly of viral particles inside the host cell. The newly produced structural proteins, S, M, and E, are inserted into the endoplasmic reticulum (ER) or Golgi membrane, while the N protein associates with the newly synthesized viral RNA to stabilize the genome. The viral particles are assembled into the ER-Golgi intermediate compartment (ERGIC), fuse with the plasma membrane, and bud off the host cell. The released virions will further infect more cells. The functions of other NSPs are not fully understood. A comparative structural genomics study revealed a possible functional intra-viral and human-virus interaction network of NSPs [29]. Recurrent mutations in the SARS-CoV-2 genome have been identified in some NSPs and the S protein, suggesting ongoing adaptations of the coronavirus through transmission [30]. Particularly, the D614G mutation in the S protein makes it more stable, and the virus becomes more infectious and transmissible [31,32]. This mutated virus is the dominant form in Europe and North America since March 2020 [33].

3. Why are individuals with SUDs particularly vulnerable to COVID-19?

SARS-CoV-2 can attack and damage human organs through two major events: direct viral attacks against target organs and abnormal immune responses and inflammation [34]. Initial evidence focuses on the damage to the respiratory system and the lung, and is correlated with clinical symptoms of the patients [2,11,17,18]. The identified viral entry receptor, ACE2, is abundantly expressed in the epithelial cells along the respiratory tract and the lung alveoli [26,35,36]. High level expression of ACE2 receptors is also reported in organs and tissues outside the respiratory system, including the heart, kidney, and intestine [37]. Therefore, these organs are the potential targets of and could be damaged by SARS-CoV-2. As mentioned earlier, smoking of tobacco and marijuana directly impairs respiratory system. Other substances of abuse can cause cardiovascular diseases, which amplify respiratory and pulmonary complications. We will discuss these complications in the next section.

More severely, ACE2 is abundantly expressed in vascular endothelial cells [37]. Several clinical cases have been reported to indicate direct involvement of vascular endothelial cells in COVID-19 pathology at different organs, suggesting that the damages to the lung, heart, kidney, liver, small intestine, and bowel, are actually caused by endotheliitis (endothelialitis). Direct viral infection of endothelial cells induces inflammation and inflammatory cell death at the endothelium [38]. Comparing lung tissues from deceased patients of acute respiratory distress syndrome (ARDS) associated with influenza and COVID-19, the lungs from COVID-19 patients displayed distinctive vascular impairments of the pulmonary vessels. Most significantly, viruses were found inside the endothelial cells of the lung tissues from COVID-19 patients, which disrupted cell membrane, caused prevalent thrombosis with microangiopathy, and induced elevated intussusceptive angiogenesis [39].
A greater number of ACE2+ endothelial cells were found in COVID-19 patients, correlating to changes in endothelial morphology, including disruption of endothelial cell junctions, cell swelling, and detachment from the basal membrane. The presence of the SARS-CoV-2 virus inside the endothelial cells, together with the induced inflammation, may directly contribute to the endothelial injury [38,39]. Although there are no reported cases yet, one potential target of SARS-CoV-2 infection is the brain microvascular endothelial cell (BMVEC). BMVECs line up the microcapillary beds and form the blood-brain barrier (BBB) together with brain astrocytes and pericytes. The BBB prevents pathogens and toxins from trespassing into the brain side. Tight-junction (TJ) protein complexes, composed of occludin, claudins, junctional adhesion molecules (JAMs) and membrane-directed scaffolding protein zonulae occcludentes (ZO), form a physical inter-endothelial barrier that strictly controls migration of molecular and cellular contents from the circulation into the brain [40,41]. High expression of efflux pumps and stereospecific solute transporters at the endothelium additionally limits molecules from crossing the barrier [42-44]. The BBB plays an essential role in protecting the brain from pathogen invasion. Viral infection of BMVECs could result in endothelial dysfunction, leakage, and even rupture, and is detrimental to the BBB integrity. The damaged BBB allows the virus to migrate into the brain side, and infect neuronal tissues [45]. Another possible route of CNS invasion could be through invading the peripheral nerve terminals and then entering the CNS via trans-synaptic transfer [46]. The first case of meningitis associated with SARS-CoV-2 has been reported, in which viral RNA was detected in the cerebrospinal fluid (CSF) of the patient [47]. Clinical evidence from Wuhan, China showed that more than 1/3 of COVID-19 patients had neurological symptoms, and CNS involvement was linked to the prognosis and severity of the disease [48,49]. Substances of abuse have been shown to severely compromise the endothelial barrier at the BBB, leading to increased BBB permeability and possibly intensified brain damage in COVID-19. We will describe the detrimental effects on the BBB in the next section.

The other aspect of COVID-19 pathology involves abnormal immune responses, which could exaggerate into a cytokine storm [50,51]. Evidence has shown that COVID-19 elevates proinflammatory cytokines and chemokines, including tumor necrosis factor (TNF)-α, interleukin (IL)-1β and IL-6, granulocyte-colony stimulating factor (G-CSF), interferon gamma (IFN-γ)-induced protein-10 (IP-10), monocyte chemoattractant protein-1 (MCP-1), and macrophage inflammatory proteins-1α (MIP-1α) [52-54]. Although there has been no reported evidence, it is possible that pattern recognition receptors, such as toll-like receptors (TLR3, TLR7, and TLR8), RIG-I, and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRP1, NLRP3, and NLRP12), are also activated by COVID-19 through innate immunity [55]. It has been reported that IL-6 was significantly increased in severe COVID-19 cases, and its level was closely correlated with the severity of patients [56]. Human bronchial epithelial cells infected with SARS-CoV-2 showed elevated expression of type I and type III IFNs and IL-6 [56]. Furthermore, type I interferon, IFN-α, stimulates the expression of ACE2, the molecular target of SARS-CoV-2, in primary human nasal epithelial cells [35]. Type III interferon, IFN-λ, has been shown to disrupt the lung epithelial barrier by direct inhibition of lung epithelial proliferation and repair, contributing to COVID-19 pathogenesis in the lower airways [57]. The upregulation of IL-6 and other proinflammatory cytokines was also observed in SARS cases [58] and influenza infection [59]. Substances of abuse can induce high level of expression of proinflammatory cytokines and chemokines in the CNS and cause neuroinflammation, which can worsen inflammatory responses in COVID-19. Details will be discussed in the next section.

The bidirectional communication between the brain and the immune system plays a critical role in COVID-19 pathogenesis. It has been well established that brain-immune interactions are widespread and significant. For instance, the immune system produces hormones and neurotransmitters [60-62], while the anterior pituitary cells in CNS produce proinflammatory cytokines, such as IL-6 [63]. Microglial cells are immune effectors in the CNS, which produce and secrete cytokines and neurotrophic or neuron survival factors upon inflammation and injury [64]. For pathogen infections, the innate immunity provides the first line of defense through recognition of pathogen-associated molecular patterns (PAMPs), initiating nonspecific cellular and humoral responses and rapidly activating nonspecific neural responses, including systemic hormonal
responses through the hypothalamic-pituitary-adrenal (HPA) axis [65]. Consisting of the hypothalamus of the brain, and endocrine organs, the pituitary and cortex of the adrenal glands, the HPA axis is responsible for systematic inflammation control. The paraventricular nucleus (PVN) of the hypothalamus plays the main governing role of the HPA axis, releasing a wide range of neuropeptides, including the corticotrophin-releasing hormone (CRH) and arginin-vasopressin (AVP). These neuropeptides reach the anterior pituitary (AP) to activate corticotrope cells to secrete adrenocorticotropic hormone (ACTH). ACTH subsequently enables the synthesis and secretion of glucocorticoids in the zona fasciculata of the adrenal cortex through melanocortin type 2 receptors [66]. The physiological feedback loop involves releasing immune mediators and cytokines, such as IL-1, IL-6, and tumor-necrosis factors (TNFs), by the innate immune system to activate neural responses, which amplify local inflammation to contain and eliminate pathogen invasions. Upon pathogen clearance, the brain responds by activating the HPA axis and releasing anti-inflammatory molecules, glucocorticoids, from the adrenal cortex. The release of this final product of the HPA axis sends a signal to the immune system to terminate the inflammatory responses, completing the hormonal negative-feedback loop. Glucocorticoids also negatively regulate the HPA axis itself, restoring host homeostasis, including CNS and cardiovascular system, as well as metabolic balances. The interplay between the nervous system and the immune system plays a critical role in forming a cohesive and integrated early host response for pathogen clearance through an optimized innate inflammatory response. Impairment of the HPA axis by various substances of abuse could render the host highly susceptible to inflammation and even increased mortality from septic shock from exposure to infectious and proinflammatory triggers, including COVID-19. Inappropriate and excessive CNS responses could predispose the host to extreme inflammation, including cytokine storm that has been observed clinically in influenza [59,67], SARS [68], and COVID-19 [51].

4. What are the effects of commonly abused substances on COVID-19?

For individuals with SUDs, both the infection of vascular endothelial cells and the proinflammatory immune responses of COVID-19 could pose severe risks. It has been well studied that substances of abuse can cause irreversible BBB damage [69] and impair the HPA axis and immune responses [66]. To understand the vulnerabilities of SUDs in COVID-19 infection at the molecular level, we will examine the brain-immune interactions and the HPA axis attenuation, neuroinflammation, immunosuppression, and BBB impairment induced by most commonly abused substances in the comorbidity of COVID-19.

4.1. Nicotine

Nicotine is available in the form of tobacco, which also contains many other types of chemicals. Tobacco smoking is associated with arterial thrombosis and atherosclerosis in the heart, abdomen, and neck [70], and is associated with increased risk of stroke, pulmonary disease, and emphysema [71]. As nicotine damages the lung directly, smoking is one of the leading causes of COPD. Smokers and COPD patients are at high risk in developing severe disease and have a higher mortality rate in COVID-19 [12]. Smoking can significantly worsen COVID-19 progression, with more critical conditions and higher fatality [72]. Cigarette smoking increases the number of alveolar macrophages (AMs), the innate immune cells in the lung by several fold. These cells secrete lysosomal enzyme, elastase, which can damage connective tissue and parenchymal cells of the lung, one of the contributors to the COPD pathogenesis [73].

Nicotine is also an important immune modulator. It significantly reduces antibody responses and T-cell proliferation [74]. The immune suppression by nicotine, particularly the decrease in CD8+ T-cells that facilitate the rapid resolution of acute viral infections, increases the susceptibility of smokers for viral infections [75]. Analysis of clinical data has indicated that smokers are twice as much as non-smokers to contract the virus, have a more severe disease progression, and have higher mortality rates [76].

Nicotine induces BBB leakage and increases BBB permeability by diminishing the expression of tight junction proteins, including occludin, claudin-3, ZO-1, and JAMs [77-80]. Nicotine alters actin
cytoskeleton arrangement in the BMVEC, which also greatly increases BBB permeability, resulting in a surge of bacterial invasion to the brain [81]. Nicotine induces oxidative stress, which can progressively compromise the BBB integrity [82]. Nicotine increases gene expression of proinflammatory cytokines, TNFα, IL-1β, and IL-18, and chemokines, CCL2, CCL8, and CXC3CL1, and suppresses anti-inflammatory factors, Bcl6, IL-10, and CCL25 in the brain microvessels [83]. Nicotine’s detrimental effects on the BBB and induced neuroinflammation are serious concerns for COVID-19.

4.2. Alcohol

This history of fermentation production and the use of alcohol could date back to 10,000 BC. Although light-to-moderate consumption may arguably have positive health benefits, particularly in lowering cardiovascular risks [84,85], high dose alcohol can have severe neurotoxic effects and cause dementia [86]. Alcohol has been linked to liver damage, inflammation of the pancreas and stomach, and neurodegenerative disorders. High alcohol consumption changes gene expression of immune response genes in the brain region frontal cortex [87]. Proinflammatory signaling is also connected to high alcohol intake [88]. Binge alcohol (blood alcohol content B.A.C ≥ 0.08 %) elevates expression of proinflammatory cytokines, IL-1β and IL-6, and chemokine CCL-2 (MCP-1) [89]. Binge alcohol can also damage various organs, including the gut, liver, and brain [90], and lead to spleen atrophy in a hippocampus-mediated fashion [91]. As suggested in a comparative risk assessment using the margin of exposure (MOE) benchmark, alcohol is considered to have the highest risk of mortality [92]. The alcohol-induced inflammatory cytokine release could expose COVID-19 patients to excessive inflammatory responses. The spleen damage by alcohol could weaken the immune response in COVID-19 by reducing the production of antibodies and lymphocytes against the virus [93].

Alcohol can induce BBB disruption by decreasing the expression of tight junction proteins and increasing mitogen-activated protein kinase (MAPK) activities [94]. Alcohol may also stimulate inositol 1,4,5-triphosphate receptor (IP3R)-operated intracellular Ca²⁺ release, activating myosin light chain kinase (MLCK). The heightened kinase activities lead to the phosphorylation of cytoskeletal and tight junction proteins, compromising the BBB integrity [95]. Alcohol-mediated oxidative stress in BMVEC can also activate MLCK that alters cytoskeleton and tight junction protein structures, causing BBB leakage [96]. Disruption of the BBB associated with chronic alcohol use could increase the possibilities of invading pathogens, including SARS-CoV-2, to infiltrate the brain.

4.3. Marijuana

Marijuana, also known as cannabis, is the most frequently used illicit substance of abuse in the U.S. Currently, 11 states and the District of Columbia have legalized the recreational use of marijuana, and 16 states have decriminalized marijuana use and possession. The use of marijuana for medical purposes is legal in 33 states. It is, however, illegal under federal law to use and possess marijuana. It is classified as a Schedule I substance by the US Drug Enforcement Administration (DEA), indicating a high potential for abuse and no accepted medical use, despite the fact that medical marijuana has been well established [97,98]. Two cannabinoids, Δ⁹-tetrahydrocannabinol (THC) and cannabidiol (CBD), are the main active ingredients in marijuana. Cannabinoids act on the endocannabinoid system (ECS), which is composed of two cannabinoid receptors, CB1, expressed in most neuronal cells, and CB2, expressed predominantly in immune cells [99]. THC is the component that produces psychotropic effects through stimulating the CB1 receptor and mediating the inhibition of neurotransmitter release [100]. CBD is considered to regulate immune response, such as cytokine release, and blood pressure, with little to no psychotropic side effects, as CBD only binds to CB2 but not to CB1 receptors [100].

Both THC and CBD are lipophilic, allowing them to readily pass through the BBB and enter the CNS. THC is an immunosuppressor. It has been reported to suppress antibody response and T lymphocytes activities. It prevents macrophage and macrophage-like cells, such as microglia, from migrating towards the nodes of microbial invasions. It also suppresses proinflammatory factors and...
promotes anti-inflammatory activities [101]. THC downregulates proinflammatory cytokines, IL-1α, IL-1β, and TNFα. By dampening immune responses to invading pathogens, THC could make the host more susceptible to viral infections, such as HIV-1 [101] and possibly COVID-19. Other harmful effects of marijuana include cardiovascular, cerebrovascular, and neurological complications, such as stroke, cognitive dysfunction, and behavioral problems [102]. Some evidence suggested a link between smoking marijuana and risk of COPD [103], one of the major risk factors for COVID-19 complications, but the risk is only significant if the individuals also smoked tobacco [104]. On the contrary, a study, which has not been peer-reviewed at the time of preparation of this article, showed the beneficial effect of CBD in preventing COVID-19 by modulating ACE2 expression and downregulating serine protease TMPRSS2 [105].

4.4. Opioids

Opioids, including illegal drug heroin, synthetic drug fentanyl, prescription pain-killing drugs oxycodone (OxyContin®), hydrocodone (Vicodin®), codeine, and morphine, are effectors on the endocrine system [106]. They act as immune suppressors that impair the function of macrophages, natural killer (NK) cells, and T-cells, and are associated with higher risks of infectious diseases, such as pneumonia [107]. The endogenous opioid system (EOS), comprising three naturally occurring opioid peptides (β-endorphins, dynorphins, and enkephalins) and three classes of opioid receptors, μ (MOR), δ (DOR), and κ (KOR), are tightly linked to substance abuse and the development of addiction, and are responsible for systemic infection [108]. Activation of opioid receptors in the brain stem could lead to respiratory depression and overdose fatality [109]. Respiratory depression is a leading factor to hypoxemia in COVID-19 complications. Morphine desensitizes the HPA axis and inhibits the release of anti-inflammatory glucocorticoids through potentiating proinflammatory cytokine, IL-1β [110]. Inhibition of endogenous glucocorticoid and activation of IL-1β by opioids could significantly increase the severity and inflammatory response of COVID-19 in patients with OUD. It has been shown that individuals with OUD are more susceptible to opportunistic infections [111], dispositioning these individuals at a higher chance of contracting the virus in COVID-19.

4.5. Cocaine

Cocaine is a potent stimulant, and is the second most abused illicit drug after marijuana. Cocaine can be abused in several forms, such as chewing the leaves of Erythroxylum coca tree, injecting the water-soluble hydrochloride salt form, and smoking or snorting pure, freebase form, called “crack”. One of the most significant pathophysiological effects of cocaine is its cardiotoxicity, which has been well documented and extensively reviewed [112-116]. Cardiac arrhythmias and acute myocardial ischemia or infarction (MI) are the leading causes of cocaine-induced sudden cardiac death. Other cardiovascular diseases associated with cocaine include heart failure, cardiomyopathies, aortic dissection, and endocarditis. Cocaine blocks voltage-dependent Na⁺ and K⁺ channels in the sinoatrial node and the myocardium, depressing cardiovascular contractility [117]. Due to its function as an ion channel blocker, cocaine has been effectively used as a local anesthetic [118]. cocaine can also induce binding and opening of L-type Ca²⁺ channels, causing the influx of Ca²⁺ in cardiomyocytes and elevation of intracellular Ca²⁺ concentration. This second messenger pathway may also lead to cardiac arrhythmia [119]. Clinically, cocaine increases myocardial oxygen demand by increasing heart rate and hypertension, while it decreases the oxygen supply due to coronary vasoconstriction [120-122]. Cocaine impairs endothelial functions [123], sensitizes constrictor effects of catecholamines [124], and causes microvascular diseases and thrombosis [125,126]. The cocaine-mediated oxygen imbalance can be particularly detrimental in COVID-19, in which the coronavirus can cause hypoxemia because of the diminishing of lung capacity.

Cocaine exerts its effect through binding to three monoamine transporters on nerve terminals: the serotonin transporter (SERT), the dopamine transporter (DAT), and the norepinephrine transporter (NET), with $K_i$ of 0.14, 0.64, and 1.6 μM, respectively [127]. Upon binding to these transporters, cocaine inhibits the reuptake of the neurotransmitters from the synaptic cleft, leading to prolonged synapses and activation of postsynaptic receptors. Cocaine also binds directly to two
classes of neurotransmitter receptors, muscarinic acetylcholine and sigma receptors [128-130]. The interactions with the transporters and the receptors form the molecular basis for cocaine neurotoxicity. Cocaine stimulates the HPA axis, increasing the secretion of neuronal peptide CRH, which leads to subsequent releasing of β-endorphin and ACTH. Through general circulation, ACTH reaches the adrenal glands, and promotes the biosynthesis of glucocorticoids [131]. Cocaine-induced stimulation of the HPA axis and immune suppression can alter antibody formation, lymphocyte subset profile, and lymphocyte proliferation. Cocaine suppresses responses to the proinflammatory cytokine, IL-6, and dampens cytotoxic activation of macrophages, natural killer cells, and cytotoxic T lymphocytes [132]. Due to compromised immune responses, cocaine abusers have considerably high incidences of viral infections, including human immunodeficiency virus (HIV), influenza, and potentially SARS-CoV-2.

Cocaine can induce BBB dysfunction, disrupt neurovascular capillaries and basement membrane [133], and increase BBB permeability [134-136]. The detrimental effects of cocaine on the BBB are partially attributed to the loss of tight junction protein complexes, including ZO-1 and JAM-2 [136-139]. Cocaine also increases the expression of matrix metallopeptidase (MMP)-1, which contributes to the rearrangement of the cytoskeleton structure of the basement membrane [137,138]. The adverse effects of decreased tight junction protein complexes and remodeling of the basement membrane fibers cause the BBB leakage and make it open to peripheral toxins and viruses, including SARS-CoV-2. An increase in proinflammatory cytokine, TNFα, has also been reported in BMVECs exposed to cocaine [140], which could be a concern for endothelial health in COVID-19.

4.6. Amphetamine, methamphetamine (METH), and 3,4-methylenedioxymethamphetamine (MDMA, Ecstasy)

D-Amphetamine and its synthetic derivatives, METH and MDMA, are addictive psychostimulants associated with neuropsychiatric complications, including deficits in attention, memory, and executive functioning [141,142]. Amphetamines mediate neurodegenerative changes in the brain, including persistent loss of dopamine (DA) transporters [143-147] and receptors [148], loss of serotonin (5-HT) transporters [149], and decrease in dopamine and serotonin level and its metabolites [150-152]. METH has been linked to various cardiac pathologies, including hypertension, tachycardia, and congestive heart failure or cardiomyopathy [15]. Clinically, METH abusers commonly showed dilated and poorly contracting left ventricles (LV) and substantially lowered left ventricular ejection fractions (LVEF) in comparison to non-users [153]. METH-associated cardiac pathologies are a major cause of pulmonary edema. The reduced lung capacity due to the fluid collection in the lungs and constriction of blood vessels can severely complicate COVID-19 symptoms and negative prognosis.

Amphetamines stimulate the HPA axis and increase the plasma glucocorticoids through a CRH-dependent mechanism involving serotonin [154-157]. The stimulation increases the production of CRH and AVP in the PVN neurons, which, in turn, activates the production of ACTH in corticotropic cells in the anterior pituitary gland. ACTH circulates through the systemic blood stream to reach and activate the adrenal cortex to release glucocorticoids. The HPA axis is an essential component of the response to pathogen infections. However, chronic activation of the HPA axis and markedly increased glucocorticoid due to the recreational use of amphetamine, METH, and MDMA can be harmful to the brain. Amphetamines modify brain expression of the genes and proteins associated with the HPA axis, including the glucocorticoid receptor (GR) and the mineralocorticoid receptor (MR). The remodeling of brain cells and disruption of the HPA axis are the hallmarks of depression and anxiety/despair states associated with drug use [158,159]. As mentioned earlier, the disruption of the HPA axis and the associated immune suppression could pose the drug abusers to a higher risk of viral infections. SARS-CoV-2, known for its high infection rate, will be particularly harmful to individuals with defective immune systems related to the use of addictive drugs.

Amphetamine-like psychostimulants can trigger inflammatory processes, compromise neurogenesis in the brain, and damage the BBB integrity [160]. METH, for example, is strongly associated with ischemic stroke and hypoxia [161,162]. Binge use of METH causes a sustained reduction in global and cerebral blood flow [163,164]. METH is also known to damage the central
nervous system (CNS) by compromising the integrity of the blood-brain barrier (BBB) [165-167]. METH and MDMA can decrease the expression of tight junction proteins, including ZO-1, occludin, and claudin-5. These drugs activate microglia and astrocyte to secrete proinflammatory cytokines and chemokines, as well as vasoactive factors, and elevate expression of peptidases, such as MMP-1 and MMP-9, to degrade tight junction proteins and modify BBB basement membrane structure [168,169]. METH abuse has been shown to increase brain infection of peripheral bacteria and viruses [170-172]. Similarly, MDMA has been shown to cause BBB dysfunction with increased BBB permeability [173], and to excessively activate astrocytes and microglia [174]. It may lead to edema [173]. With BBB dysfunction, it is almost certain that the risks of SARS-CoV-2 invasion into the brain will be immensely heightened among individuals using or abusing these psychostimulants, and complications in these patients are vastly expected.

Neuroinflammation induced by METH and MDMA also plays a major role in BBB damage and may deteriorate COVID-19 conditions. COVID-19 patients exhibit abnormal immune responses related to high levels of proinflammatory cytokines, including TNFα and IL-6. METH significantly increases the expression of TNFα and IL-6 in the hippocampus, frontal cortex, and striatum [175]. The expression of these proinflammatory cytokines is linked to METH-induced microglial activation [176,177]. MDMA also elevates the expression of proinflammatory cytokines, such as IL-1β, in the brain [178]. The excessive expression of proinflammatory cytokines in brain tissues could further damage the BBB and cause oxidative stress [69,179]. Neuroinflammation poses a major risk for individuals with COVID-19.

4.7. Summary of roles of substances of abuse in COVID-19

Substances of abuse may lead to COVID-19 complication and severity in several ways. Smoking tobacco and marijuana could cause direct damage to the respiratory system, such as COPD. Other substances mostly work through modulating brain and immune functions, including the promotion of proinflammatory factors, suppression of immune responses, and impairment of the BBB. Neuroinflammation induced by several substances of abuse and inflammatory activities caused by COVID-19 in the peripheral tissues may mutually intensify the adverse effects of one another, leading to negative progression of the disease. With impaired HPA axis and immune imbalance, the patients are highly susceptible to SARS-CoV-2 infections. Compromised BBB may pose a high risk of viral infection in the brain tissue. Figure 1 sketches the possible pathological effects of commonly abused substances on various tissues and systems and their connection to COVID-19 complications. Table 1 summarizes the adverse effects of these substances on the CNS, the immune system, the respiratory system, and the cardiovascular system, and their relations to the severity and negative prognosis of COVID-19.

**Figure 1.** Pathological effects of substances of abuse on various tissues and their implied complications in COVID-19.

**Table 1.** Connection of substances of abuse to COVID-19
| Substance                  | Target system       | Pathology                                                                                                                                                                                                 | COVID-19                                                                 |
|---------------------------|---------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------|
| Tobacco (Nicotine)        | Respiratory system  | Main cause of COPD                                                                                                                                                                                        | Increased severity and mortality                                         |
|                           | Immune system       | Immune suppression, Decreased CD8+ T-cells                                                                                                                                                              | Higher infection rate                                                   |
|                           |                     | Increased inflammatory cytokines (TNFα, IL-1β, IL-18) and chemokines (CCL2, CCL8, and CX3CL1); decreased anti-inflammatory factors, Bcl6, IL-10, and CCL25 | Increased inflammatory cytokines and chemokines, TNFα, IL-1β, IL-6       |
|                           | CNS                 | BBB leakage through loss of tight junction proteins                                                                                                                                                     | Endotheliitis and CNS infection                                          |
| Alcohol                   | Immune system       | Increased proinflammatory cytokines, IL-1β and IL-6, and chemokine CCL-2                                                                                                                               | Increased inflammatory cytokines and chemokines, TNFα, IL-1β, IL-6       |
|                           |                     | Spleen atrophy                                                                                                                                                                                           | Impaired production of antibodies and lymphocytes                       |
|                           | CNS                 | Increased BBB permeability through cytoskeletal and tight junction remodeling                                                                                                                            | Endotheliitis and CNS infection                                          |
| Marijuana (THC, CBD)      | Respiratory system  | Enhanced COPD with tobacco                                                                                                                                                                                | Increased severity and mortality                                         |
|                           | Immune system       | Immunosuppression; reduced antibody response and T lymphocyte activities; reduced migration of macrophage                                                                                              | Increased infection and reduced viral response and clearance            |
| Opioids (heroin, fentanyl, morphine) | Respiratory system  | Respiratory depression                                                                                                                                                                                   | Increased severity and mortality                                         |
|                           | Immune system       | Desensitizing HPA axis; inhibiting glucocorticoid release, increased IL-1β; neuroinflammation                                                                                                            | Increased opportunistic infections, excessive inflammatory response     |
| Cocaine                   | Cardiovascular system | Cardiac arrhythmias and acute MI; oxygen imbalance; microvascular diseases and thrombosis                                                                                                                  | Increased severity and mortality                                         |
|                           | Immune system       | Stimulating HPA axis; immunosuppression; defects in antibody formation, lymphocyte proliferation, macrophage and NK activation                                                                          | High incidence of viral infection                                       |
|                           | CNS                 | Increased BBB permeability due to loss of tight junction proteins; rearrangement of cytoskeleton structure                                                                                              | Endotheliitis and CNS infection                                          |
| Amphetamine, METH, MDMA   | Cardiovascular system | Hypertension, tachycardia, and cardiomyopathy leading to pulmonary edema;                                                                                                                                 | Increased severity and mortality                                         |
5. What are the treatment and prevention strategies for individuals with SUDs?

5.1. General approaches for treatments and vaccines

The therapeutic strategies for COVID-19 have been focused on repurposing existing drugs against this novel coronavirus [180-182]. Among these repurposed drugs, the antimalaria drugs chloroquine and hydroxychloroquine [183,184] are extremely controversial, including a retracted study [185] and terminated solidarity clinical trial by the WHO [186]. The U.S. Food and Drug Administration (FDA) recently revoked its Emergency Use Authorization (EUA) to treat COVID-19 [187]. Other drugs include the anti-HIV drugs Lopinavir-Ritonavir in combination with Ribavirin [188-190], tumor chemotherapy drugs, doxorubicin and paclitaxel [191], traditional herbal medicines [192], broad spectrum antiviral drug Niclosamide [193], Janus-associated kinase (JAK) 1 and 2 inhibitor, Ruxolitinib [194], anti-influenza drug Favipiravir [195], antiviral drug Remdesivir [196-198], and most recently, a commonly used steroid, dexamethasone [199]. Many of these drugs primarily target the RdRp (NSP12) or the main protease, M<sup>pro</sup>. A cryo-EM structure of NSP12 in complex with its cofactors NSP7 and NSP8 has been reported [200]. Remdesivir, the only proven effective drug against COVID-19 so far, is found to potently inhibit RdRp in MERS-CoV [201] and SARS-CoV2 [202]. Ribavirin and Favipiravir also function as RdRp inhibitors. The crystal structure of the M<sup>pro</sup> with its inhibitor has been recently solved and reported [203,204]. Due to its importance in viral production, and the lack of similar proteins in human cells, the M<sup>pro</sup> is considered an important drug target in treating COVID-19 [203,205,206]. Niclosamide and the anti-HIV drugs, Lopinavir-Ritonavir, are inhibitors of the main protease. Structure-based design has led to the development of new inhibitors of the M<sup>pro</sup> with desirable pharmacokinetic properties and low toxicity [207]. The SARS-CoV-2 entry point, ACE2, and the associated protease, TMPRSS2, are also considered potential drug targets [25].

Tremendous research efforts have been put into identifying, isolating, and developing neutralizing antibodies against SARS-CoV-2. The S protein, which plays a key role in recognizing and binding to the ACE2 receptor to gain entry into the host cell, is the main focus for developing neutralizing antibodies and vaccines against SARS-CoV-2 [21,22]. Neutralizing monoclonal antibodies isolated from convalescent COVID-19 patients were found to block the RBD surface of the S protein from binding to ACE2, and these antibodies showed effectiveness in reducing viral infection in animal models [208-210]. The crystal structure of the RBD-bound antibody provided a clear picture on inhibition of viral interaction with ACE2 [211]. The neutralizing antibodies isolated from SARS-CoV patients seemed to cross-neutralize SARS-CoV-2 [212]. The RBDF is not the only site that neutralizing antibodies may block. A monoclonal antibody isolated from convalescent COVID-19 patients exhibits high neutralization potency against SARS-CoV-2. This antibody does not bind to the RBD, but it tightly associates with the N-terminal domain of the S protein [213]. A recombinant antibody fused with the human ACE2 extracellular domain displayed desired neutralizing properties in vivo and in mice [214]. Single-domain camelid antibodies from a llama were found to cross-react and neutralize MERS-CoV, SARS-CoV, and SARS-CoV-2 S pseudotyped viruses [215]. Cocktails of antibodies that simultaneously bind to different epitopes of the RBD may provide more potent neutralizing power and significantly reduce virus escaping through mutations [216-218].

Vaccines are crucial in combating the COVID-19 pandemic. The RBD of the S protein is, again, the main target for vaccine development [219]. According to WHO “Draft landscape of COVID-19
candidate vaccines”, there are 13 candidates currently in clinical trials, and 128 candidates in preclinical stage [220]. These vaccine candidates cover almost all technology platforms, including more traditional non-replicating or replicating viral vector, inactivated or live attenuated virus, recombinant protein subunit, to more recently developed nucleic acid (DNA or RNA), peptide, and viral-like particle [221]. There have been some encouraging results from clinical trials, including the first mRNA vaccine, mRNA-1273 [222], and an adenovirus type-5 (Ad-5) vectored vaccine [223].

5.2. Challenges for individuals with SUDs

With the race to therapeutics and vaccines towards COVID-19, rarely have the efforts specifically been directed at SUD complications in COVID-19. As we analyzed earlier, SUDs complicate and impair the respiratory system directly, and can intensify the severity of the disease through cardiovascular damage and immune abnormality. There are increasing concerns on capillary endothelial damage or endotheliitis by COVID-19 and SUDs, including the harmful effects on the BBB integrity. If the coronavirus is allowed to migrate across the BBB and to infect the brain, long-term neurological degeneracy is expected, and the treatment will be deemed challenging. SUDs can weaken the immune system, alter and disrupt the HPA axis, and stimulate neuroinflammation with heightened expression of TNFα, IL-1β, and IL-6 in the CNS. Anti-TNFα therapy has been used in severe cases of autoimmune inflammatory disease to control inflammation by downregulating IL-6 and IL-1β [224]. Anti-IL-6 antibody may also be beneficial in inflammation control [225]. These treatments could be helpful for COVID-19 patients with SUDs.

Current antiviral drugs are designed to interfere with viral replication or viral protein processing. For example, the most effective drug, Remdesivir, which can shorten the recovery time by 30% [196], inhibits viral RNA replication [202]. Neutralizing antibodies bind to RBD of the S protein, so the virus cannot bind to ACE2. However, clearance of the virus is heavily dependent on the individual’s immune system, including activating phagocytes and natural killer cells. As discussed earlier, several substances of abuse, such as nicotine, marijuana, cocaine, and amphetamines, have shown to suppress immune and antibody responses. Clinical cases indicated that antibody-secreting cells (ASCs), T follicular helper (Tfh) cells, as well as activated CD4+ and CD8+ T-cells, are critical to symptomatic recovery from COVID-19 [226]. As a result, SUD-induced reduction of T-cell activation will dampen the ability to efficiently clear the virus from the body. For vaccines to work efficiently, robust immune responses are required. CD4+ and CD8+ T-cell responses are essential in protective antiviral immunity by vaccination [227,228]. Due to the immunosuppression and reduction of T-cell responses, individuals with SUDs may not develop sufficient protective antibodies against the virus. A clinical study suggested that SARS-CoV-2 specific immunoglobulin G (IgG) antibodies may last only 2-3 months before a steep decline in both asymptomatic and mildly symptomatic COVID-19 patients [229]. Considering their substantially undermined immune systems, the protective antibodies in patients with SUDs are very likely to decline much quicker, making vaccines considerably less effective in these patients. There is a potential risk, although no reported cases yet, that antibodies or vaccines may promote COVID-19 pathogenesis through antibody-dependent enhancement (ADE). While SUDs compromise the immune system, virus-specific antibodies are likely to promote viral entry into various immune cells, including monocytes, macrophages, and B cells [230,231], which may further deteriorate the immune response towards the virus.

It is now widely accepted that COVID-19 may stay with us for some extended periods. People with mild or no symptoms can transmit the pathogen as effectively as those with severe symptoms [232,233]. The complications and compromised immune systems associated with SUD make drug abusers particularly vulnerable to COVID-19. Therapeutics and vaccines currently under development do not address the specific concerns and risk factors for these individuals. There are even greater challenges for people with SUD under COVID-19 pandemic, as they will experience a higher infectious rate, limited access to healthcare system and support groups, inadequate food and housing, and increased likelihood of homelessness and incarceration. They may also face excessive discrimination, and have higher chances of relapses and overdose death. The research community should heed to the challenges and difficulties these individuals may experience in the pandemic,
uncover scientific evidence to link COVID-19 severity and mortality with substance use, and advance effective treatment and prevention strategies for people with SUDs.

**Author Contributions:** Conceptualization, Y.W.; methodology, Y.W. and R.S.; formal analysis, Y.W. and R.S.; resources, Y.W. and R.S.; data curation, Y.W. and R.S.; writing—original draft preparation, Y.W. and R.S.; writing—review and editing, Y.W. and R.S.; visualization, Y.W. and R.S.; supervision, Y.W.; project administration, Y.W.; funding acquisition, Y.W. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Institute on Drug Abuse (NIDA) of the National Institutes of Health (NIH), grant number R21DA046223.

**Acknowledgments:** We thank Ammer Abdin, Madellyne Sanchez, and Song Zhang, for collecting and analyzing relevant literatures and intellectual discussions for this article.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Coronaviridae Study Group of the International Committee on Taxonomy of, V. The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. *Nat Microbiol* 2020, 5, 536-544, doi:10.1038/s41564-020-0695-z.
2. Zhu, N.; Zhang, D.; Wang, W.; Li, X.; Yang, B.; Song, J.; Zhao, X.; Huang, B.; Shi, W.; Lu, R., et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *New England Journal of Medicine* 2020, 382, 727-733, doi:10.1056/NEJMoa2001017.
3. World Health Organization. WHO Coronavirus Disease (COVID-19) Dashboard. Available online: [https://covid19.who.int/?gclid=CjwKCAjw-D3BBIEiwAjVMv7lZxWSCeKBbqAB9-TFuuZjmGDDMDed6ZzGzjKeqBgN3kIA6kRI9XEHoCG9kQAvDBwE](https://covid19.who.int/?gclid=CjwKCAjw-D3BBIEiwAjVMv7lZxWSCeKBbqAB9-TFuuZjmGDDMDed6ZzGzjKeqBgN3kIA6kRI9XEHoCG9kQAvDBwE) (accessed on June 28, 2020).
4. Johns Hopkins University & Medicine. Coronavirus Resource Center. Available online: [https://coronavirus.jhu.edu/](https://coronavirus.jhu.edu/) (accessed on June 28, 2020).
5. World Health Organization. WHO Director-General’s opening remarks at the media briefing on COVID-19 – 11 March 2020. Available online: [https://www.who.int/dg/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19---11-march-2020](https://www.who.int/dg/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19---11-march-2020) (accessed on March 17, 2020).
6. United Nations Office on Drugs and Crime. World Drug Report 2019. Available online: [https://wdr.unodc.org/wdr2019/](https://wdr.unodc.org/wdr2019/) (accessed on June 25).
7. Substance Abuse and Mental Health Services Administration. *Key Substance Use and Mental Health Indicators in the United States: Results from the 2018 National Survey on Drug Use and Health;* 2019.
8. Volkow, N.D. Collision of the COVID-19 and Addiction Epidemics. *Ann Intern Med* 2020, 10.7326/M20-1212, M20-1212, doi:10.7326/M20-1212.
9. Sundar, I.K.; Nevid, M.Z.; Friedman, A.E.; Rahman, I. Cigarette smoke induces distinct histone modifications in lung cells: implications for the pathogenesis of COPD and lung cancer. *J Proteome Res* 2014, 13, 982-996, doi:10.1021/pr400998n.
10. Aldington, S.; Williams, M.; Nowitz, M.; Weatherall, M.; Pritchard, A.; McNaughton, A.; Robinson, G.; Beasley, R. Effects of cannabis on pulmonary structure, function and symptoms. *Thorax* 2007, 62, 1058-1063, doi:10.1136/thx.2006.070781.
11. Wu, Z.; McGoogan, J.M. Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72 314 Cases From the Chinese Center for Disease Control and Prevention. *JAMA* 2020, 10.1001/jama.2020.2648, doi:10.1001/jama.2020.2648.
12. Alqahtani, J.S.; Oyelade, T.; Aldhaher, A.M.; Alghamdi, S.M.; Almehmadi, M.; Alqahtani, A.S.; Quaderi, S.; Mandal, S.; Hurst, J.R. Prevalence, Severity and Mortality associated with COPD and Smoking in patients with COVID-19: A Rapid Systematic Review and Meta-Analysis. *PLOS ONE* **2020**, *15*, e0233147, doi:10.1371/journal.pone.0233147.

13. Alexander, G.C.; Stoller, K.B.; Haffajee, R.L.; Saloner, B. An Epidemic in the Midst of a Pandemic: Opioid Use Disorder and COVID-19. *Ann Intern Med* **2020**, 10.7326/M20-1141, M20-1141, doi:10.7326/M20-1141.

14. Becker, W.C.; Fiellin, D.A. When Epidemics Collide: Coronavirus Disease 2019 (COVID-19) and the Opioid Crisis. *Ann Intern Med* **2020**, 10.7326/M20-1210, M20-1210, doi:10.7326/M20-1210.

15. Zhao, S.X.; Kwok, C.; Swaminathan, A.; Gohil, A.; Crawford, M.H. Clinical Characteristics and Outcome of Methamphetamine-Associated Pulmonary Arterial Hypertension and Dilated Cardiomyopathy. *JACC Heart Fail* **2018**, *6*, 209-218, doi:10.1016/j.jchf.2017.10.006.

16. Cui, J.; Li, F.; Shi, Z-L. Origin and evolution of pathogenic coronaviruses. *Nature Reviews Microbiology* **2019**, *17*, 181-192, doi:10.1038/s41579-018-0118-9.

17. Zhou, P.; Yang, X-L.; Wang, X-G.; Hu, B.; Zhang, L.; Zhang, W.; Si, H-R.; Zhu, Y.; Li, B.; Huang, C-L., et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* **2020**, *579*, 270-273, doi:10.1038/s41586-020-2012-7.

18. Wu, F.; Zhao, S.; Yu, B.; Chen, Y.M.; Wang, W.; Song, Z.G.; Hu, Y.; Tao, Z.W.; Tian, J.H.; Pei, Y.Y., et al. A new coronavirus associated with human respiratory disease in China. *Nature* **2020**, *579*, 265-269, doi:10.1038/s41586-020-2008-3.

19. Letko, M.; Marzi, A.; Munster, V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. *Nat Microbiol* **2020**, *5*, 562-569, doi:10.1038/s41564-020-0688-y.

20. Wrapp, D.; Wang, N.; Corbett, K.S.; Goldsmith, J.A.; Hsieh, C.L.; Abiona, O.; Graham, B.S.; McLellan, J.S. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science* **2020**, *367*, 1260-1263, doi:10.1126/science.abb2507.

21. Walls, A.C.; Park, Y.J.; Tortorici, M.A.; Wall, A.; McGuire, A.T.; Veesler, D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell* **2020**, *181*, 281-292 e286, doi:10.1016/j.cell.2020.02.058.

22. Tai, W.; He, L.; Zhang, X.; Pu, J.; Voronin, D.; Jiang, S.; Zhou, Y.; Du, L. Characterization of the receptor-binding domain (RBD) of 2019 novel coronavirus: implication for development of RBD protein as a viral attachment inhibitor and vaccine. *Cellular & Molecular Immunology* **2020**, 10.1038/s41423-020-0400-4, doi:10.1038/s41423-020-0400-4.

23. Vankadari, N.; Wilce, J.A. Emerging WuHan (COVID-19) coronavirus: glycan shield and structure prediction of spike glycoprotein and its interaction with human CD26. *Emerging Microbes & Infections* **2020**, *9*, 601-604, doi:10.1080/22221751.2020.1739565.

24. Watanabe, Y.; Allen, J.D.; Wrapp, D.; McLellan, J.S.; Crispin, M. Site-specific glycan analysis of the SARS-CoV-2 spike. *Science* **2020**, 10.1126/science.abb9983, eabb9983, doi:10.1126/science.abb9983.

25. Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Krüger, N.; Herrler, T.; Erichsen, S.; Schiergens, T.S.; Herrler, G.; Wu, N.H.; Nitsche, A., et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* **2020**, *181*, 271-280.e278, doi:10.1016/j.cell.2020.02.052.

26. Sungnai, W.; Huang, N.; Bécavin, C.; Berg, M.; Queen, R.; Litvinukova, M.; Talavera-López, C.; Maatz, H.; Reichard, D.; Sampaziotis, F., et al. SARS-CoV-2 entry factors are highly expressed in nasal epithelial
cells together with innate immune genes. *Nature Medicine* 2020, 26, 681-687, doi:10.1038/s41591-020-0868-6.

27. Li, X.; Geng, M.; Peng, Y.; Meng, L.; Lu, S. Molecular immune pathogenesis and diagnosis of COVID-19. *J Pharm Anal* 2020, 10, 102-108, doi:10.1016/j.jpha.2020.03.001.

28. Thoms, M.; Buschauer, R.; Ameismeier, M.; Koepke, L.; Denk, T.; Hirschenberger, M.; Kratzat, H.; Hayn, M.; Mackens-Kiani, T.; Cheng, J., et al. Structural basis for translational shutdown and immune evasion by the Nsp1 protein of SARS-CoV-2. *bioRxiv* 2020, 10.1101/2020.05.18.102467, 2020.2005.2018.102467, doi:10.1101/2020.05.18.102467.

29. Srinivasan, S.; Cui, H.; Gao, Z.; Liu, M.; Lu, S.; Mkandawire, W.; Narykov, O.; Sun, M.; Korkin, D. Structural Genomics of SARS-CoV-2 Indicates Evolutionary Conserved Functional Regions of Viral Proteins. *Viruses* 2020, 12, doi:10.3390/v12040360.

30. van Dorp, L.; Acman, M.; Richard, D.; Shaw, L.P.; Ford, C.E.; Ormond, L.; Owen, C.J.; Pang, J.; Tan, C.C.S.; Boshier, F.A.T., et al. Emergence of genomic diversity and recurrent mutations in SARS-CoV-2. *Infect Genet Evol* 2020, 10.1016/j.meegid.2020.104351, 104351, doi:10.1016/j.meegid.2020.104351.

31. Hu, J.; He, C.L.; Gao, Q.; Zhang, G.J.; Cao, X.X.; Long, Q.X.; Deng, H.J.; Huang, L.Y.; Chen, J.; Wang, K., et al. The D614G mutation of SARS-CoV-2 spike protein enhances viral infectivity and decreases neutralization sensitivity to individual convalescent sera. *bioRxiv* 2020, 10.1101/2020.06.20.161323, 2020.2006.20161323, doi:10.1101/2020.06.20.161323.

32. Zhang, L.; Jackson, C.B.; Mou, H.; Ojha, A.; Rangarajan, E.S.; Izard, T.; Farzan, M.; Choe, H. The D614G mutation in the SARS-CoV-2 spike protein reduces S1 shedding and increases infectivity. *bioRxiv* 2020, 10.1101/2020.06.12.148726, 2020.2006.12.148726, doi:10.1101/2020.06.12.148726.

33. Korber, B.; Fischer, W.; Gnanakaran, S.; Yoon, H.; Theiler, J.; Abfalterer, W.; Foley, B.; Giorgi, E.; Bhattacharya, T.; Parker, M., et al. Spike mutation pipeline reveals the emergence of a more transmissible form of SARS-CoV-2. *bioRxiv* 2020, 10.1101/2020.04.29.069054, 2020.2004.29.069054, doi:10.1101/2020.04.29.069054.

34. Cao, W.; Li, T. COVID-19: towards understanding of pathogenesis. *Cell Research* 2020, 30, 367-369, doi:10.1038/s41422-020-0327-4.

35. Ziegler, C.G.K.; Allon, S.J.; Nyquist, S.K.; Mbano, I.M.; Miao, V.N.; Tzouanas, C.N.; Cao, Y.; Yousif, A.S.; Bals, J.; Hauser, B.M., et al. SARS-CoV-2 Receptor ACE2 Is an Interferon-Stimulated Gene in Human Airway Epithelial Cells and Is Detected in Specific Cell Subsets across Tissues. *Cell* 2020, 181, 1016-1035 e1019, doi:10.1016/j.cell.2020.04.035.

36. Ravindra, N.G.; Alfajaro, M.M.; Gasque, V.; Wei, J.; Filler, R.B.; Huston, N.C.; Wan, H.; Szegeti-Buck, K.; Wang, B.; Montgomery, R.R., et al. Single-cell longitudinal analysis of SARS-CoV-2 infection in human bronchial epithelial cells. *bioRxiv* 2020, 10.1101/2020.05.06.081695, 2020.2005.06.081695, doi:10.1101/2020.05.06.081695.

37. Hamming, I.; Timens, W.; Bulthuis, M.L.; Lely, A.T.; Navis, G.; van Goor, H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J Pathol* 2004, 203, 631-637, doi:10.1002/path.1570.

38. Varga, Z.; Flammer, A.J.; Steiger, P.; Haberecker, M.; Andermatt, R.; Zinkernagel, A.S.; Mehra, M.R.; Schuepbach, R.A.; Ruschitzka, F.; Moch, H. Endothelial cell infection and endothelitis in COVID-19. *The Lancet* 2020, 395, 1417-1418, doi:10.1016/S0140-6736(20)30937-5.
39. Ackermann, M.; Verleden, S.E.; Kuehnel, M.; Haverich, A.; Welte, T.; Laenger, F.; Vanstapel, A.; Werlein, C.; Stark, H.; Tzankov, A., et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. *New England Journal of Medicine* 2020, 10.1056/NEJMoa2015432, doi:10.1056/NEJMoa2015432.

40. Bauer, H.C.; Krizbai, I.A.; Bauer, H.; Traweger, A. "You Shall Not Pass"-tight junctions of the blood brain barrier. *Front Neurosci* 2014, 8, 392, doi:10.3389/fnins.2014.00392.

41. Luissant, A.C.; Artus, C.; Glacial, F.; Ganeshamoorthy, K.; Couraud, P.O. Tight junctions at the blood brain barrier: physiological architecture and disease-associated dysregulation. *Fluids Barriers CNS* 2012, 9, 23, doi:10.1186/2045-8118-9-23.

42. Löscher, W.; Potschka, H. Drug resistance in brain diseases and the role of drug efflux transporters. *Nat Rev Neurosci* 2005, 6, 591-602, doi:10.1038/nrn1728.

43. Miller, D.S.; Cannon, R.E. Signaling pathways that regulate basal ABC transporter activity at the blood-brain barrier. *Curr Pharm Des* 2014, 20, 1463-1471, doi:10.2174/1381612813199990457.

44. Stamatovic, S.M.; Johnson, A.M.; Keep, R.F.; Andjelkovic, A.V. Junctional proteins of the blood-brain barrier: New insights into function and dysfunction. *Tissue Barriers* 2016, 4, e1154641-e1154641, doi:10.1080/21688370.2016.1154641.

45. Baig, A.M.; Khaleeq, A.; Ali, U.; Syeda, H. Evidence of the COVID-19 Virus Targeting the CNS: Tissue Distribution, Host-Virus Interaction, and Proposed Neurotropic Mechanisms. *ACS Chem Neurosci* 2020, 11, 995-998, doi:10.1021/acschemneuro.0c00122.

46. Li, Y.C.; Bai, W.Z.; Hashikawa, T. The neuroinvasive potential of SARS-CoV2 may play a role in the respiratory failure of COVID-19 patients. *J Med Virol* 2020, 92, 552-555, doi:10.1002/jmv.25728.

47. Moriguchi, T.; Harii, N.; Goto, J.; Harada, D.; Sugawara, H.; Takamino, J.; Ueno, M.; Sakata, H.; Kondo, K.; Myose, N., et al. A first case of meningitis/encephalitis associated with SARS-CoV-2. *Int J Infect Dis* 2020, 94, 55-58, doi:10.1016/j.ijid.2020.03.062.

48. Li, H.; Xue, Q.; Xu, X. Involvement of the Nervous System in SARS-CoV-2 Infection. *Neurotoxicity Research* 2020, 38, 1-7, doi:10.1007/s12640-020-00219-8.

49. Mao, L.; Jin, H.; Wang, M.; Hu, Y.; Chen, S.; He, Q.; Chang, J.; Hong, C.; Zhou, Y.; Wang, D., et al. Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China. *JAMA Neurology* 2020, 77, 683-690, doi:10.1001/jamaneurol.2020.1127.

50. Li, H.; Liu, L.; Zhang, D.; Xu, J.; Dai, H.; Tang, N.; Su, X.; Cao, B. SARS-CoV-2 and viral sepsis: observations and hypotheses. *The Lancet* 2020, 395, 1517-1520, doi:10.1016/S0140-6736(20)30920-X.

51. Coperchini, F.; Chiovato, L.; Croce, L.; Magri, F.; Rotondi, M. The cytokine storm in COVID-19: An overview of the involvement of the chemokine/chemokine-receptor system. *Cytokine Growth Factor Rev* 2020, 53, 25-32, doi:10.1016/j.cytogfr.2020.05.003.

52. Huang, C.; Wang, Y.; Li, X.; Ren, L.; Zhao, J.; Hu, Y.; Zhang, L.; Fan, G.; Xu, J.; Gu, X., et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *The Lancet* 2020, 395, 497-506, doi:10.1016/S0140-6736(20)30183-5.

53. Liu, J.; Li, S.; Liu, J.; Liang, B.; Wang, X.; Wang, H.; Li, W.; Tong, Q.; Yi, J.; Zhao, L., et al. Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients. *EBioMedicine* 2020, 55, 102763, doi:10.1016/j.ebiom.2020.102763.

54. Gong, J.; Dong, H.; Xia, S.Q.; Huang, Y.Z.; Wang, D.; Zhao, Y.; Liu, W.; Tu, S.; Zhang, M.; Wang, Q., et al. Correlation Analysis Between Disease Severity and Inflammation-related Parameters in Patients with COVID-19 Pneumonia. *medRxiv* 2020, 10.1101/2020.02.25.20025643, 2020.2002.2025.20025643, doi:10.1101/2020.02.25.20025643.
55. Iwasaki, A.; Pillai, P.S. Innate immunity to influenza virus infection. Nat Rev Immunol 2014, 14, 315-328, doi:10.1038/nri3665.

56. Liu, T.; Zhang, J.; Yang, Y.; Ma, H.; Li, Z.; Zhang, J.; Cheng, J.; Zhang, X.; Zhao, Y.; Xia, Z., et al. The potential role of IL-6 in monitoring severe case of coronavirus disease 2019. medRxiv 2020, 10.1101/2020.03.01.20029769, 2020.2003.2001.20029769, doi:10.1101/2020.03.01.20029769.

57. Broggi, A.; Ghosh, S.; Sposito, B.; Speafico, R.; Balzarini, F.; Lo Cascio, A.; Clementi, N.; De Santis, M.; Mancini, N.; Granucci, F., et al. Type III interferons disrupt the lung epithelial barrier upon viral recognition. Science 2020, 10.1126/science.abc3545, eabc3545, doi:10.1126/science.abc3545.

58. Wang, W.; Ye, L.; Ye, L.; Li, B.; Gao, B.; Zeng, Y.; Kong, L.; Fang, X.; Zheng, H.; Wu, Z., et al. Up-regulation of IL-6 and TNF-alpha induced by SARS-coronavirus spike protein in murine macrophages via NF-kappaB pathway. Virus Res 2007, 128, 1-8, doi:10.1016/j.virusres.2007.02.007.

59. Liu, Q.; Zhou, Y.-h.; Yang, Z.-q. The cytokine storm of severe influenza and development of immunomodulatory therapy. Cellular & molecular immunology 2016, 13, 3-10, doi:10.1038/cmi.2015.74.

60. Aller, M.A.; Arias, J.L.; Lorente, L.; Nava, M.P.; Duran, H.J.; Arias, J. Neuro-immune-endocrine functional system and vascular pathology. Med Hypotheses 2001, 57, 561-569, doi:10.1054/mehy.2001.1408.

61. Meazza, C.; Pagani, S.; Travaglino, P.; Bozzola, M. Effect of growth hormone (GH) on the immune system. Pediatr Endocrinol Rev 2004, 1 Suppl 3, 490-495.

62. Hosoi, T.; Nomura, Y. Functional role of acetylcholine in the immune system. Front Biosci 2004, 9, 2414-2419, doi:10.2741/1404.

63. Fuchs, F.; Damm, J.; Gerstberger, R.; Roth, J.; Rummel, C. Activation of the inflammatory transcription factor nuclear factor interleukin-6 during inflammatory and psychological stress in the brain. Journal of Neuroinflammation 2013, 10, 905, doi:10.1186/1742-2094-10-140.

64. Pivneva, T.A. Microglia in normal condition and pathology. Fiziol Zh 2008, 54, 81-89.

65. Sternberg, E.M. Neural regulation of innate immunity: a coordinated nonspecific host response to pathogens. Nature Reviews Immunology 2006, 6, 318-328, doi:10.1038/nri1810.

66. Armario, A. Activation of the hypothalamic-pituitary-adrenal axis by addictive drugs: different pathways, common outcome. Trends Pharmacol Sci 2010, 31, 318-325, doi:10.1016/j.tips.2010.04.005.

67. Tisoncik, J.R.; Korth, M.J.; Simmons, C.P.; Farrar, J.; Martin, T.R.; Katze, M.G. Into the eye of the cytokine storm. Microbiol Mol Biol Rev 2012, 76, 16-32, doi:10.1128/MMBR.05015-11.

68. Huang, K.J.; Su, I.J.; Theron, M.; Wu, Y.C.; Lai, S.K.; Liu, C.C.; Lei, H.Y. An interferon-gamma-related cytokine storm in SARS patients. J Med Virol 2005, 75, 185-194, doi:10.1002/jmv.20255.

69. Sajja, R.K.; Rahman, S.; Cucullo, L. Drugs of abuse and blood-brain barrier endothelial dysfunction: A focus on the role of oxidative stress. J Cereb Blood Flow Metab 2016, 36, 539-554, doi:10.1177/0271678X15616978.

70. Howard, G.; Wagenknecht, L.E.; Burke, G.L.; Diez-Roux, A.; Evans, G.W.; McGovern, P.; Nieto, F.J.; Tell, G.S. Cigarette smoking and progression of atherosclerosis: The Atherosclerosis Risk in Communities (ARIC) Study. Jama 1998, 279, 119-124, doi:10.1001/jama.279.2.119.

71. Bhat, V.M.; Cole, J.W.; Sorkin, J.D.; Wozniak, M.A.; Malarcher, A.M.; Giles, W.H.; Stern, B.J.; Kittner, S.J. Dose-Response Relationship Between Cigarette Smoking and Risk of Ischemic Stroke in Young Women. Stroke 2008, 39, 2439-2443, doi:10.1161/STROKEAHA.107.510073.

72. Patanavanich, R.; Glantz, S.A. Smoking Is Associated With COVID-19 Progression: A Meta-analysis. Nicotine & Tobacco Research 2020, 10.1093/ntt/ntaa082, doi:10.1093/ntt/ntaa082.
73. Sopori, M. Effects of cigarette smoke on the immune system. Nat Rev Immunol 2002, 2, 372-377, doi:10.1038/nri803.

74. Geng, Y.; Savage, S.M.; Razani-Boroujerdi, S.; Sopori, M.L. Effects of nicotine on the immune response. II. Chronic nicotine treatment induces T cell anergy. J Immunol 1996, 156, 2384-2390.

75. Zhou, Z.; Chen, P.; Peng, H. Are healthy smokers really healthy? Tob Induc Dis 2016, 14, 35, doi:10.1186/s12971-016-0101-z.

76. Vardavas, C.I.; Nikitara, K. COVID-19 and smoking: A systematic review of the evidence. Tob Induc Dis 2020, 18, 20-20, doi:10.18332/tid/119324.

77. Abbruscato, T.J.; Lopez, S.P.; Mark, K.S.; Hawkins, B.T.; Davis, T.P. Nicotine and Cotinine Modulate Cerebral Microvascular Permeability and Protein Expression of ZO-1 through Nicotinic Acetylcholine Receptors Expressed on Brain Endothelial Cells. Journal of Pharmaceutical Sciences 2002, 91, 2525-2538, doi:10.1002/jps.10256.

78. Hutamekalin, P.; Farkas, A.E.; Orbók, A.; Wilhelm, I.; Nagyössi, P.; Veszélka, S.; Deli, M.A.; Buzás, K.; Hunyadi-Gulyás, É.; Medzihradszky, K.F., et al. Effect of nicotine and polyaromatic hydrocarbons on cerebral endothelial cells. Cell Biology International 2008, 32, 198-209, doi:10.1016/j.cellbi.2007.08.026.

79. Rodriguez-Gaztelumendi, A.; Alvehus, M.; Andersson, T.; Jacobsson, S.O. Comparison of the effects of nicotine upon the transcellular electrical resistance and sucrose permeability of human ECV304/rat C6 co-cultures and human CaCo(2) cells. Toxicol Lett 2011, 207, 1-6, doi:10.1016/j.toxlet.2011.08.014.

80. Hawkins, B.T.; Abbruscato, T.J.; Egleton, R.D.; Brown, R.C.; Huber, J.D.; Campos, C.R.; Davis, T.P. Nicotine increases in vivo blood-brain barrier permeability and alters cerebral microvascular tight junction protein distribution. Brain Res 2004, 1027, 48-58, doi:10.1016/j.brainres.2004.08.043.

81. Chen, Y.-H.; Chen, S.H.-M.; Jong, A.; Zhou, Z.Y.; Li, W.; Suzuki, K.; Huang, S.-H. Enhanced Escherichia coli invasion of human brain microvascular endothelial cells is associated with alterations in cytoskeleton induced by nicotine. Cellular Microbiology 2002, 4, 503-514, doi:10.1046/j.1462-5822.2002.00209.x.

82. Mazzone, P.; Tierney, W.; Hossain, M.; Puverna, V.; Janigro, D.; Cucullo, L. Pathophysiological impact of cigarette smoke exposure on the cerebrovascular system with a focus on the blood-brain barrier: expanding the awareness of smoking toxicity in an underappreciated area. Int J Environ Res Public Health 2010, 7, 4111-4126, doi:10.3390/ijerph7124111.

83. Bradford, S.T.; Stamatovic, S.M.; Dondeci, R.S.; Keep, R.F.; Andjelkovic, A.V. Nicotine aggravates the brain postischemic inflammatory response. American Journal of Physiology-Heart and Circulatory Physiology 2011, 300, H1518-H1529, doi:10.1152/ajpheart.00928.2010.

84. Blomster, J.I.; Zoungas, S.; Chalmers, J.; Li, Q.; Chow, C.K.; Woodward, M.; Mancia, G.; Poulter, N.; Williams, B.; Harrap, S., et al. The Relationship Between Alcohol Consumption and Vascular Complications and Mortality in Individuals With Type 2 Diabetes. Diabetes Care 2014, 37, 1353-1359, doi:10.2337/dc13-2727.

85. Jones, A.; McMillan, M.R.; Jones, R.W.; Kowalik, G.T.; Steeden, J.A.; Pruessner, J.C.; Taylor, A.M.; Deanfield, J.E.; Muthurangu, V. Habitual alcohol consumption is associated with lower cardiovascular stress responses – a novel explanation for the known cardiovascular benefits of alcohol? Stress 2013, 16, 369-376, doi:10.3109/10253890.2013.777833.

86. Perry, P.J.; Argo, T.R.; Barnett, M.J.; Liesveld, J.L.; Liskow, B.; Hernandez, J.M.; Trnka, M.G.; Brabson, M.A. The Association of Alcohol-Induced Blackouts and Grayouts to Blood Alcohol Concentrations. Journal of Forensic Sciences 2006, 51, 896-899, doi:10.1111/j.1556-4029.2006.00161.x.
87. Liu, J.; Lewohl, J.M.; Harris, R.A.; Iyer, V.R.; Dodd, P.R.; Randall, P.K.; Mayfield, R.D. Patterns of gene expression in the frontal cortex discriminate alcoholic from nonalcoholic individuals. *Neuropsychopharmacology* 2006, 31, 1574-1582, doi:10.1038/sj.npp.1300947.

88. Blednov, Y.A.; Benavidez, J.M.; Geil, C.; Perra, S.; Morikawa, H.; Harris, R.A. Activation of inflammatory signaling by lipopolysaccharide produces a prolonged increase of voluntary alcohol intake in mice. *Brain Behav Immun* 2011, 25 Suppl 1, S92-S105, doi:10.1016/j.bbi.2011.01.008.

89. Liu, X.; Mao, X.; Chang, S.L. Altered gene expression in the spleen of adolescent rats following high ethanol concentration binge drinking. *Int J Clin Exp Med* 2011, 4, 252-257.

90. Wang, H.J.; Zakhari, S.; Jung, M.K. Alcohol, inflammation, and gut-liver-brain interactions in tissue damage and disease development. *World J Gastroenterol* 2010, 16, 1304-1313, doi:10.3748/wjg.v16.i11.1304.

91. Liu, X.; Connaghan, K.P.; Wei, Y.; Yang, Z.; Li, M.D.; Chang, S.L. Involvement of the Hippocampus in Binge Ethanol-Induced Spleen Atrophy in Adolescent Rats. *Alcoholism, clinical and experimental research* 2016, 40, 1489-1500, doi:10.1111/acer.13109.

92. Lachenmeier, D.W.; Rehm, J. Comparative risk assessment of alcohol, tobacco, cannabis and other illicit drugs using the margin of exposure approach. *Scientific Reports* 2015, 5, 8126, doi:10.1038/srep08126.

93. Lewis, S.M.; Williams, A.; Eisenbarth, S.C. Structure and function of the immune system in the spleen. *Science Immunology* 2019, 4, eaau6085, doi:10.1126/sciimmunol.aaau6085.

94. Singh, A.K.; Jiang, Y.; Gupta, S.; Benlhabib, E. Effects of chronic ethanol drinking on the blood–brain barrier and ensuing neuronal toxicity in alcohol-prefering rats subjected to intraperitoneal LPS injection. *Alcohol and Alcoholism* 2007, 42, 385-399, doi:10.1093/alcalc/agl120.

95. Haorah, J.; Knipe, B.; Gorantla, S.; Zheng, J.; Persidsky, Y. Alcohol-induced blood-brain barrier dysfunction is mediated via inositol 1,4,5-triphosphate receptor (IP3R)-gated intracellular calcium release. *J Neurochem* 2007, 100, 324-336, doi:10.1111/j.1471-4159.2006.04245.x.

96. Haorah, J.; Knipe, B.; Leibhart, J.; Ghorpade, A.; Persidsky, Y. Alcohol-induced oxidative stress in brain endothelial cells causes blood-brain barrier dysfunction. *Journal of Leukocyte Biology* 2005, 78, 1223-1232, doi:10.1189/jlb.0605340.

97. Haorah, J.; Knipe, B.; Gorantla, S.; Zheng, J.; Persidsky, Y. Alcohol-induced blood-brain barrier dysfunction is mediated via inositol 1,4,5-triphosphate receptor (IP3R)-gated intracellular calcium release. *J Neurochem* 2007, 100, 324-336, doi:10.1111/j.1471-4159.2006.04245.x.

98. Aggarwal, S.K.; Carter, G.T.; Sullivan, M.D.; ZumBrunnen, C.; Morrill, R.; Mayer, J.D. Medicinal use of cannabis in the United States: historical perspectives, current trends, and future directions. *J Opioid Manag* 2009, 5, 153-168, doi:10.5055/jom.2009.0016.

99. Pertwee, R.G. Pharmacological actions of cannabinoids. *Handb Exp Pharmacol* 2005, 10.1007/3-540-26573-2_1, 1-51, doi:10.1007/3-540-26573-2_1.

100. Miller, A.M.; Stella, N. CB2 receptor-mediated migration of immune cells: it can go either way. *British Journal of Pharmacology* 2008, 153, 299-308, doi:10.1038/sj.bjp.0707523.

101. Cabral, G.A.; Jamerson, M. Marijuana use and brain immune mechanisms. *International review of neurobiology* 2014, 118, 199-230, doi:10.1016/b978-0-12-801284-0.00008-7.

102. Archie, S.R.; Cucullo, L. Harmful Effects of Smoking Cannabis: A Cerebrovascular and Neurological Perspective. *Frontiers in Pharmacology* 2019, 10, doi:10.3389/fphar.2019.01481.

103. Tan, W.C.; Lo, C.; Jong, A.; Xing, L.; Fitzgerald, M.J.; Vollmer, W.M.; Buist, S.A.; Sin, D.D. Marijuana and chronic obstructive lung disease: a population-based study. *CMAJ* 2009, 180, 814-820, doi:10.1503/cmaj.081040.
104. Tashkin, D.P. Does smoking marijuana increase the risk of chronic obstructive pulmonary disease? *CMAJ* 2009, 180, 797-798, doi:10.1503/cmaj.090142.

105. Wang, B.; Kovalchuk, A.; Li, D.; Ilnytsky, Y.; Kovalchuk, I.; Kovalchuk, O. In Search of Preventative Strategies: Novel Anti-Inflammatory High-CBD Cannabis Sativa Extracts Modulate ACE2 Expression in COVID-19 Gateway Tissues. 2020, 10.20944/preprints202004.0315.v1, doi:10.20944/preprints202004.0315.v1.

106. Katz, N.; Mazer, N.A. The impact of opioids on the endocrine system. *Clin J Pain* 2009, 25, 170-175, doi:10.1097/AJP.0b013e3181850df6.

107. Plein, L.M.; Rittner, H.L. Opioids and the immune system - friend or foe. *Br J Pharmacol* 2018, 175, 2717-2725, doi:10.1111/bph.13750.

108. Holden, J.E.; Jeong, Y.; Forrest, J.M. The endogenous opioid system and clinical pain management. *AACN Clin Issues* 2005, 16, 291-301, doi:10.1097/00044067-200507000-00003.

109. Trigo, J.M.; Martin-Garcia, E.; Berrendero, F.; Robledo, P.; Maldonado, R. The endogenous opioid system: a common substrate in drug addiction. *Drug Alcohol Depend* 2010, 108, 183-194, doi:10.1016/j.drugalcdep.2009.10.011.

110. Ocasio, F.M.; Jiang, Y.; House, S.D.; Chang, S.L. Chronic morphine potentiates the inflammatory response by disrupting interleukin-1beta modulation of the hypothalamic-pituitary-adrenal axis. *J Neuroimmunol* 2001, 118, 277-285, doi:10.1016/s0165-5728(01)00337-x.

111. Pramanik, P. Cocaine Cardiac Toxicity : Revisited. 2018; 10.5772/intechopen.79929.

112. Wood, D.M.; Dargan, P.I.; Hoffman, R.S. Management of cocaine-induced cardiac arrhythmias due to cardiac ion channel dysfunction. *Clinical Toxicology* 2009, 47, 14-23, doi:10.1080/15563650802339373.

113. Calatayud, J., M.D., D.D.S., Ph.D.; González, Á., M.D., D.D.S., Ph.D. History of the Development and Evolution of Local Anesthesia Since the Coca Leaf. *Anesthesiology: The Journal of the American Society of Anesthesiologists* 2003, 98, 1503-1508.

114. Premkumar, L.S. Selective Potentiation of L-Type Calcium Channel Currents by Cocaine in Cardiac Myocytes. *Molecular Pharmacology* 1999, 56, 1138-1142, doi:10.1124/mol.56.6.1138.

115. Lange, R.A.; Cigarroa, R.G.; Yancey, C.W., Jr.; Willard, J.E.; Popma, J.J.; Sills, M.N.; McBride, W.; Kim, A.S.; Hillis, L.D. Cocaine-induced coronary-artery vasoconstriction. *N Engl J Med* 1989, 321, 1557-1562, doi:10.1056/nejm198912073212301.
121. Fischman, M.W.; Schuster, C.R.; Resnekov, L.; Shick, J.F.; Krasnegor, N.A.; Fennell, W.; Freedman, D.X. Cardiovascular and subjective effects of intravenous cocaine administration in humans. *Arch Gen Psychiatry* 1976, 33, 983-989, doi:10.1001/archpsyc.1976.01770080101010.

122. Flores, E.D.; Lange, R.A.; Cigarroa, R.G.; Hillis, L.D. Effect of cocaine on coronary artery dimensions in atherosclerotic coronary artery disease: enhanced vasoconstriction at sites of significant stenoses. *J Am Coll Cardiol* 1990, 16, 74-79, doi:10.1016/0735-1097(90)90459-3.

123. Havranek, E.P.; Nademanee, K.; Grayburn, P.A.; Eichhorn, E.J. Endothelium-dependent vasorelaxation is impaired in cocaine arteriopathy. *J Am Coll Cardiol* 1996, 28, 1168-1174, doi:10.1016/s0735-1097(96)00299-9.

124. Vita, J.A.; Treasure, C.B.; Fish, R.D.; Yeung, A.C.; Vekshtein, V.I.; Ganz, P.; Selwyn, A.P. Endothelial dysfunction leads to increased coronary constriction to catecholamines in patients with early atherosclerosis. *Journal of the American College of Cardiology* 1990, 15, A158, doi:10.1016/0735-1097(90)92348-6.

125. Kelly, R.F.; Sompalli, V.; Sattar, P.; Khankari, K. Increased TIMI frame counts in cocaine users: a case for increased microvascular resistance in the absence of epicardial coronary disease or spasm. *Clin Cardiol* 2003, 26, 319-322, doi:10.1002/clc.4950260705.

126. Weber, J.E.; Hollander, J.E.; Murphy, S.A.; Braunwald, E.; Gibson, C.M. Quantitative comparison of coronary artery flow and myocardial perfusion in patients with acute myocardial infarction in the presence and absence of recent cocaine use. *J Thromb Thrombolysis* 2002, 14, 239-245, doi:10.1023/a:1025056912284.

127. Ritz, M.C.; Lamb, R.J.; Goldberg, S.R.; Kuhar, M.J. Cocaine receptors on dopamine transporters are related to self-administration of cocaine. *Science* 1987, 237, 1219-1223, doi:10.1126/science.2820058.

128. Sharkey, J.; Ritz, M.C.; Schenden, J.A.; Hanson, R.C.; Kuhar, M.J. Cocaine inhibits muscarinic cholinergic receptors in heart and brain. *J Pharmacol Exp Ther* 1988, 246, 1048-1052.

129. Flynn, D.D.; Vaishnav, A.A.; Mash, D.C. Interactions of cocaine with primary and secondary recognition sites on muscarinic receptors. *Mol Pharmacol* 1992, 41, 736-742.

130. Sharkey, J.; Glen, K.A.; Wolfe, S.; Kuhar, M.J. Cocaine binding at sigma receptors. *Eur J Pharmacol* 1988, 149, 171-174, doi:10.1016/0014-2999(88)90058-1.

131. Goeders, N.E. The HPA axis and cocaine reinforcement. *Psychoneuroendocrinology* 2002, 27, 13-33, doi:10.1016/s0306-4530(01)00034-8.

132. Halpern, J.H.; Sholar, M.B.; Glowacki, J.; Mello, N.K.; Mendelson, J.H.; Siegel, A.J. Diminished interleukin-6 response to proinflammatory challenge in men and women after intravenous cocaine administration. *J Clin Endocrinol Metab* 2003, 88, 1188-1193, doi:10.1210/jc.2002-020804.

133. Barroso-Moguel, R.; Villeda-Hernandez, J.; Mendez-Armenta, M.; Rios, C. Brain capillary lesions produced by cocaine in rats. *Toxicol Lett* 1997, 92, 9-14, doi:10.1016/s0378-4274(97)00027-1.

134. Sharma, H.S.; Muresanu, D.; Sharma, A.; Patnaik, R. Chapter 11 - Cocaine-Induced Breakdown of the Blood–Brain Barrier and Neurotoxicity. In *International review of neurobiology*, Academic Press: 2009; Vol. 88, pp. 297-334.

135. Yang, Y.; Yao, H.; Lu, Y.; Wang, C.; Buch, S. Cocaine Potentiates Astrocyte Toxicity Mediated by Human Immunodeficiency Virus (HIV-1) Protein gp120. *PLOS ONE* 2010, 5, e13427, doi:10.1371/journal.pone.0013427.

136. Yao, H.; Kim, K.; Duan, M.; Hayashi, T.; Guo, M.; Morgello, S.; Prat, A.; Wang, J.; Su, T.-P.; Buch, S. Cocaine Hijacks σ1 Receptor to Initiate Induction of Activated Leukocyte Cell Adhesion Molecule:
Implication for Increased Monocyte Adhesion and Migration in the CNS. *The Journal of Neuroscience* 2011, 31, 5942-5955, doi:10.1523/jneurosci.5618-10.2011.

137. Fiala, M.; Eshleman, A.J.; Cashman, J.; Lin, J.; Lossinsky, A.S.; Suarez, V.; Yang, W.; Zhang, J.; Popik, W.; Singer, E., et al. Cocaine increases human immunodeficiency virus type 1 neuroinvasion through remodeling brain microvascular endothelial cells. *Journal of NeuroVirology* 2005, 11, 281-291, doi:10.1080/135502805952835.

138. Dhillon, N.K.; Peng, F.; Bokhari, S.; Callen, S.; Shin, S.-H.; Zhu, X.; Kim, K.-J.; Buch, S.J. Cocaine-mediated Alteration in Tight Junction Protein Expression and Modulation of CCL2/CCR2 Axis Across the Blood-Brain Barrier: Implications for HIV-Dementia. *Journal of Neuroimmune Pharmacology* 2008, 3, 52-56, doi:10.1007/s11481-007-9091-1.

139. Gandhi, N.; Saiyed, Z.M.; Napuri, J.; Samikkannu, T.; Reddy, P.V.B.; Agudelo, M.; Khattak, P.; Saxena, S.K.; Nair, M.P.N. Interactive role of human immunodeficiency virus type 1 (HIV-1)-clade-specific Tat protein and cocaine in blood-brain barrier dysfunction: Implications for HIV-1-associated neurocognitive disorder. *Journal of NeuroVirology* 2010, 16, 294-305, doi:10.3109/13550284.2010.499891.

140. Lee, Y.W.; Hennig, B.; Fiala, M.; Kim, K.S.; Toborek, M. Cocaine activates redox-regulated transcription factors and induces TNF-alpha expression in human brain endothelial cells. *Brain Res* 2001, 920, 125-133, doi:10.1016/s0006-8993(01)03047-5.

141. Cadet, J.L.; Krasnova, I.N. Chapter 5 - Molecular Bases of Methamphetamine-Induced Neurodegeneration. In *International Review of Neurobiology*, Hari Shanker, S., Ed. Academic Press: 2009; Vol. Volume 88, pp. 101-119.

142. Cadet, J.L.; Krasnova, I.N. Interactions of HIV and Methamphetamine: Cellular and Molecular Mechanisms of Toxicity Potentiation. *Neurotoxicity Research* 2007, 12, 181-204.

143. Sekine, Y.; Minabe, Y.; Ouchi, Y.; Takei, N.; Iyo, M.; Nakamura, K.; Suzuki, K.; Tsukada, H.; Okada, H.; Yoshikawa, E., et al. Association of dopamine transporter loss in the orbitofrontal and dorsolateral prefrontal cortices with methamphetamine-related psychiatric symptoms. *The American journal of psychiatry* 2003, 160, 1699-1701.

144. Volkow, N.D.; Chang, L.; Wang, G.J.; Fowler, J.S.; Leonido-Yee, M.; Franceschi, D.; Sedler, M.J.; Gatley, S.J.; Hitzemann, R.; Ding, Y.S., et al. Association of dopamine transporter reduction with psychomotor impairment in methamphetamine abusers. *The American journal of psychiatry* 2001, 158, 377-382.

145. Zahniser, N.R.; Sorkin, A. Trafficking of dopamine transporters in psychostimulant actions. *Semin Cell Dev Biol* 2009, 20, 411-417, doi:10.1016/j.semcdb.2009.01.004.

146. Kahlig, K.M.; Lute, B.J.; Wei, Y.; Loland, C.J.; Gether, U.; Javitch, J.A.; Galli, A. Regulation of dopamine transporter trafficking by intracellular amphetamine. *Mol Pharmacol* 2006, 70, 542-548, doi:10.1124/mol.106.023952.

147. Rothman, R.B.; Baumann, M.H. Monoamine transporters and psychostimulant drugs. *Eur J Pharmacol* 2003, 479, 23-40, doi:10.1016/j.ejphar.2003.08.054.

148. Volkow, N.D.; Chang, L.; Wang, G.J.; Fowler, J.S.; Ding, Y.S.; Sedler, M.; Logan, J.; Franceschi, D.; Gatley, J.; Hitzemann, R., et al. Low level of brain dopamine D2 receptors in methamphetamine abusers: association with metabolism in the orbitofrontal cortex. *The American journal of psychiatry* 2001, 158, 2015-2021.

149. Sekine, Y.; Ouchi, Y.; Takei, N.; Yoshikawa, E.; Nakamura, K.; Futatabashi, M.; Okada, H.; Minabe, Y.; Suzuki, K.; Iwata, Y., et al. Brain serotonin transporter density and aggression in abstinent methamphetamine abusers. *Archives of general psychiatry* 2006, 63, 90-100, doi:10.1001/archpsyc.63.1.90.
150. Moszczynska, A.; Fitzmaurice, P.; Ang, L.; Kalasinsky, K.S.; Schmunk, G.A.; Peretti, F.J.; Aiken, S.S.; Wickham, D.J.; Kish, S.J. Why is parkinsonism not a feature of human methamphetamine users? *Brain : a journal of neurology* **2004**, *127*, 363-370, doi:10.1093/brain/awh046.

151. Wilson, J.M.; Kalasinsky, K.S.; Levey, A.I.; Bergeron, C.; Reiber, G.; Anthony, R.M.; Schmunk, G.A.; Shannak, K.; Haycock, J.W.; Kish, S.J. Striatal dopamine nerve terminal markers in human, chronic methamphetamine users. *Nature medicine* **1996**, *2*, 699-703.

152. Sulzer, D.; Sanders, M.S.; Poulsen, N.W.; Galli, A. Mechanisms of neurotransmitter release by amphetamines: a review. *Prog Neurobiol* **2005**, *75*, 406-433, doi:10.1016/j.pneurobio.2005.04.003.

153. Won, S.; Hong, R.A.; Shohet, R.V.; Seto, T.B.; Parikh, N.I. Methamphetamine-associated cardiomyopathy. *Clinical cardiology* **2013**, *36*, 737-742, doi:10.1002/clc.22195.

154. Gagliano, H.; Andero, R.; Armario, A.; Nadal, R. Repeated amphetamine administration in rats revealed consistency across days and a complete dissociation between locomotor and hypothalamic-pituitary-adrenal axis effects of the drug. *Psychopharmacology (Berl)* **2009**, *207*, 447-459, doi:10.1007/s00213-009-1676-3.

155. Rotllant, D.; Nadal, R.; Armario, A. Differential effects of stress and amphetamine administration on Fos-like protein expression in corticotropin releasing factor-neurons of the rat brain. *Dev Neurobiol* **2007**, *67*, 702-714, doi:10.1002/dneu.20345.

156. Williams, M.T.; Schaefer, T.L.; Furay, A.R.; Ehman, L.A.; Vorhees, C.V. Ontogeny of the adrenal response to (+)-methamphetamine in neonatal rats: the effect of prior drug exposure. *Stress* **2006**, *9*, 153-163, doi:10.1080/10253890600902842.

157. Doyle, J.R.; Yamamoto, B.K. Serotonin 2 receptor modulation of hyperthermia, corticosterone, and hippocampal serotonin depletions following serial exposure to chronic stress and methamphetamine. *Psychoneuroendocrinology* **2010**, *35*, 629-633, doi:10.1016/j.psyneuen.2009.10.001.

158. Zuloaga, D.G.; Jacobskind, J.S.; Raber, J. Methamphetamine and the hypothalamic-pituitary-adrenal axis. *Front Neurosci* **2015**, *9*, 178, doi:10.3389/fnins.2015.00178.

159. Parrott, A.C.; Montgomery, C.; Wetherell, M.A.; Downey, C.; Scholey, A.B. MDMA, cortisol, and heightened stress in recreational ecstasy users. *Behav Pharmacol* **2014**, *25*, 458-472, doi:10.1097/fbp.0000000000000660.

160. Silva, A.P.; Martins, T.; Baptista, S.; Gonçalves, J.; Agasse, F.; Malva, J.O. Brain injury associated with widely abused amphetamines: neuroinflammation, neurogenesis and blood-brain barrier. *Curr Drug Abuse Rev* **2010**, *3*, 239-254, doi:10.2174/187447371003040239.

161. Ohta, K.; Mori, M.; Yoritaka, A.; Okamoto, K.; Kishida, S. Delayed ischemic stroke associated with methamphetamine use. *J Emerg Med* **2005**, *28*, 165-167, doi:10.1016/j.jemermed.2004.06.015.

162. Rothrock, J.F.; Rubenstein, R.; Lyden, P.D. Ischemic stroke associated with methamphetamine inhalation. *Neurology* **1988**, *38*, 589-592, doi:10.1212/wnl.38.4.589.

163. Polesskaya, O.; Silva, J.; Sanfilippo, C.; Desrosiers, T.; Sun, A.; Shen, J.; Feng, C.; Poleskkiy, A.; Deane, R.; Zlokovic, B., et al. Methamphetamine causes sustained depression in cerebral blood flow. *Brain Res* **2011**, *1373*, 91-100, doi:10.1016/j.brainres.2010.12.017.

164. Chung, Y.A.; Peterson, B.S.; Yoon, S.J.; Cho, S.N.; Chai, S.; Jeong, J.; Kim, D.J. In vivo evidence for long-term CNS toxicity, associated with chronic binge use of methamphetamine. *Drug Alcohol Depend* **2010**, *111*, 155-160, doi:10.1016/j.drugalcdep.2010.04.005.

165. Dietrich, J.B. Alteration of blood-brain barrier function by methamphetamine and cocaine. *Cell and Tissue Research* **2009**, *336*, 385-392, doi:10.1007/s00441-009-0777-y.
166. Reiner, B.C.; Keblesh, J.P.; Xiong, H. Methamphetamine abuse, HIV infection, and neurotoxicity. *Int J Physiol Pathophysiol Pharmacol* **2009**, *1*, 162-179.

167. Turowski, P.; Kenny, B.-A. The blood-brain barrier and methamphetamine: open sesame? *Frontiers in neuroscience* **2015**, *9*, 156-156, doi:10.3389/fnins.2015.00156.

168. Asanuma, M.; Miyazaki, I.; Higashi, Y.; Tsuji, T.; Ogawa, N. Specific gene expression and possible involvement of inflammation in methamphetamine-induced neurotoxicity. *Ann NY Acad Sci* **2004**, *1025*, 69-75, doi:10.1196/annals.1316.009.

169. Kousik, S.M.; Napier, T.C.; Carvey, P.M. The effects of psychostimulant drugs on blood brain barrier function and neuroinflammation. *Front Pharmacol* **2012**, *3*, 121, doi:10.3389/fphar.2012.00121.

170. Nath, A.; Maragos, W.F.; Avison, M.J.; Schmitt, F.A.; Berger, J.R. Acceleration of HIV dementia with methamphetamine and cocaine. *J Neurovirol* **2001**, *7*, 66-71, doi:10.1080/135502801300069737.

171. Letendre, S.L.; Cherner, M.; Ellis, R.J.; Marquie-Beck, J.; Gragg, B.; Marcotte, T.; Heaton, R.K.; McCutchan, J.A.; Grant, I. The effects of hepatitis C, HIV, and methamphetamine dependence on neuropsychological performance: biological correlates of disease. *Aids* **2005**, *19* Suppl 3, S72-78, doi:10.1097/01.aids.00001922073.18691.ff.

172. Schep, L.J.; Slaughter, R.J.; Beasley, D.M. The clinical toxicology of metamfetamine. *Clin Toxicol (Phila)* **2010**, *48*, 675-694, doi:10.3109/15563650.2010.516752.

173. Sharma, H.S.; Ali, S.F. Acute Administration of 3,4-Methylenedioxymethamphetamine Induces Profound Hyperthermia, Blood–Brain Barrier Disruption, Brain Edema Formation, and Cell Injury. *Annals of the New York Academy of Sciences* **2008**, *1139*, 122-128, doi:10.1196/annals.1432.052.

174. Monks, T.J.; Jones, D.C.; Bai, F.; Lau, S.S. The role of metabolism in 3,4-(+)-methylenedioxyamphetamine and 3,4-(+)-methylenedioxymethamphetamine (ecstasy) toxicity. *Ther Drug Monit* **2004**, *26*, 132-136, doi:10.1097/00007691-200404000-00008.

175. Gonçalves, J.; Martins, T.; Ferreira, R.; Milhazes, N.; Borges, F.; Ribeiro, C.F.; Malva, J.O.; Macedo, T.R.; Silva, A.P. Methamphetamine-Induced Early Increase of IL-6 and TNF-α mRNA Expression in the Mouse Brain. *Annals of the New York Academy of Sciences* **2008**, *1139*, 103-111, doi:10.1196/annals.1432.043.

176. Lai, Y.-T.; Tsai, Y.-P.N.; Cherrng, C.G.; Ke, J.-J.; Ho, M.-C.; Tsai, C.-W.; Yu, L. Lipopolysaccharide mitigates methamphetamine-induced striatal dopamine depletion via modulating local TNF-α and dopamine transporter expression. *Journal of Neural Transmission* **2009**, *116*, 405-415, doi:10.1007/s00702-009-0204-2.

177. Gonçalves, J.; Baptista, S.; Martins, T.; Milhazes, N.; Borges, F.; Ribeiro, C.F.; Malva, J.O.; Silva, A.P. Methamphetamine-induced neuroinflammation and neuronal dysfunction in the mice hippocampus: preventive effect of indomethacin. *European Journal of Neuroscience* **2010**, *31*, 315-326, doi:10.1111/j.1460-9568.2009.07059.x.

178. Torres, E.; Gutierrez-Lopez, M.D.; Mayado, A.; Rubio, A.; O’Shea, E.; Colado, M.I. Changes in interleukin-1 signal modulators induced by 3,4-methylenedioxymethamphetamine (MDMA): regulation by CB2 receptors and implications for neurotoxicity. *Journal of Neuroinflammation* **2011**, *8*, 53, doi:10.1186/1742-2094-8-53.

179. Yamamoto, B.K.; Bankson, M.G. Amphetamine Neurotoxicity: Cause and Consequence of Oxidative Stress. *2005*, *17*, 87-118, doi:10.1615/CritRevNeurobiol.v17.i2.30.

180. Sanders, J.M.; Monogue, M.L.; Jodlowski, T.Z.; Cutrell, J.B. Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19): A Review. *Jama* **2020**, 10.1001/jama.2020.6019, doi:10.1001/jama.2020.6019.
181. Liu, C.; Zhou, Q.; Li, Y.; Garner, L.V.; Watkins, S.P.; Carter, L.J.; Smoot, J.; Gregg, A.C.; Daniels, A.D.; Jervey, S., et al. Research and Development on Therapeutic Agents and Vaccines for COVID-19 and Related Human Coronavirus Diseases. *ACS Cent Sci* 2020, 6, 315-331, doi:10.1021/acscentsci.0c00272.

182. Zhou, Y.; Hou, Y.; Shen, J.; Huang, Y.; Martin, W.; Cheng, F. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2. *Cell Discovery* 2020, 6, 14, doi:10.1038/s41212-020-0153-3.

183. Mahévas, M.; Tran, V.-T.; Roumier, M.; Chabrol, A.; Paule, R.; Guillaud, C.; Fois, E.; Lepeule, R.; Szwebel, T.-A.; Lesure, F.-X., et al. Clinical efficacy of hydroxychloroquine in patients with covid-19 pneumonia who require oxygen: observational comparative study using routine care data. *BMJ* 2020, 369, m1844, doi:10.1136/bmj.m1844.

184. Wang, M.; Cao, R.; Zhang, L.; Yang, X.; Liu, J.; Xu, M.; Shi, Z.; Hu, Z.; Zhong, W.; Xiao, G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Research* 2020, 30, 269-271, doi:10.1038/s41422-020-0282-0.

185. Mehra, M.R.; Desai, S.S.; Ruschitzka, F.; Patel, A.N. RETRACTED: Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis. *The Lancet* 2020, 10.1016/S0140-6736(20)31180-6, doi:10.1016/S0140-6736(20)31180-6.

186. World Health Organization. “Solidarity” clinical trial for COVID-19 treatments. Available online: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/global-research-on-novel-coronavirus-2019-ncov/solidarity-clinical-trial-for-covid-19-treatments (accessed on June 29, 2020).

187. U.S. Food & Drug Administration. Coronavirus (COVID-19) Update: FDA Revokes Emergency Use Authorization for Chloroquine and Hydroxychloroquine. Available online: https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-revokes-emergency-use-authorization-chloroquine-and (accessed on June 29, 2020).

188. Hung, I.F.-N.; Lung, K.-C.; Tso, E.Y.-K.; Liu, R.; Chung, T.W.-H.; Chu, M.-Y.; Ng, Y.-Y.; Lo, J.; Chan, J.; Tam, A.R., et al. Triple combination of interferon beta-1b, lopinavir–ritonavir, and ribavirin in the treatment of patients admitted to hospital with COVID-19: an open-label, randomised, phase 2 trial. *The Lancet* 2020, 10.1016/S0140-6736(20)31042-4, doi:10.1016/S0140-6736(20)31042-4.

189. Cao, B.; Wang, Y.; Wen, D.; Liu, W.; Wang, J.; Fan, G.; Ruan, L.; Song, B.; Cai, Y.; Wei, M., et al. A Trial of Lopinavir–Ritonavir in Adults Hospitalized with Severe Covid-19. *New England Journal of Medicine* 2020, 382, 1787-1799, doi:10.1056/NEJMoa2001282.

190. Khalili, J.S.; Zhu, H.; Mak, N.S.A.; Yan, Y.; Zhu, Y. Novel coronavirus treatment with ribavirin: Groundwork for an evaluation concerning COVID-19. *J Med Virol* 2020, 92, 740-746, doi:10.1002/jmv.25798.

191. Al-Motawa, M.; Abbas, H.; Wijten, P.; de la Fuente, A.; Xue, M.; Rabbani, N.; Thornalley, P.J. Vulnerabilities of the SARS-CoV-2 virus to proteotoxicity – opportunity for repurposed chemotherapy of COVID-19 infection. *bioRxiv* 2020, 10.1101/2020.04.07.029488, doi:10.1101/2020.04.07.029488.

192. Hu, K.; Guan, W.J.; Bi, Y.; Zhang, W.; Li, L.; Zhang, B.; Liu, Q.; Song, Y.; Li, X.; Duan, Z., et al. Efficacy and safety of Lianhuawingwen capsules, a repurposed Chinese herb, in patients with coronavirus disease 2019: A multicenter, prospective, randomized controlled trial. *Phytotherapy Research* 2020, 10.1016/j.phymed.2020.153242, 153242, doi:10.1016/j.phymed.2020.153242.

193. Xu, J.; Shi, P.-Y.; Li, H.; Zhou, J. Broad Spectrum Antiviral Agent Niclosamide and Its Therapeutic Potential. *ACS Infectious Diseases* 2020, 10.1021/acsinfecdis.0c00052, doi:10.1021/acsinfecdis.0c00052.
194. Cao, Y.; Wei, J.; Zou, L.; Jiang, T.; Wang, G.; Chen, L.; Huang, L.; Meng, F.; Huang, L.; Wang, N., et al. Ruxolitinib in treatment of severe coronavirus disease 2019 (COVID-19): A multicenter, single-blind, randomized controlled trial. *J Allergy Clin Immunol* 2020, 10.1016/j.jaci.2020.05.019, doi:10.1016/j.jaci.2020.05.019.

195. Coomes, E.A.; Haghbayan, H. Favipiravir, an antiviral for COVID-19? *Journal of Antimicrobial Chemotherapy* 2020, 75, 2013-2014, doi:10.1093/jac/dkaa171.

196. Beigel, J.H.; Tomashek, K.M.; Dodd, L.E.; Mehta, A.K.; Zingman, B.S.; Kalil, A.C.; Hohmann, E.; Chu, H.Y.; Luetkemeyer, A.; Kline, S., et al. Remdesivir for the Treatment of Covid-19 — Preliminary Report. *New England Journal of Medicine* 2020, 10.1056/NEJMoa2007764, doi:10.1056/NEJMoa2007764.

197. Grein, J.; Ohmagari, N.; Shin, D.; Diaz, G.; Asperges, E.; Castagna, A.; Feldt, T.; Green, G.; Green, M.L.; Lescure, F.X., et al. Compassionate Use of Remdesivir for Patients with Severe Covid-19. *N Engl J Med* 2020, 10.1056/NEJMoa2007016, doi:10.1056/NEJMoa2007016.

198. Wang, Y.; Zhang, D.; Du, G.; Du, R.; Zhao, J.; Jin, Y.; Fu, S.; Gao, L.; Cheng, Z.; Lu, Q., et al. Remdesivir in adults with severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. *The Lancet* 2020, 395, 1569-1578, doi:10.1016/S0140-6736(20)31022-9.

199. Ledford, H. Coronavirus breakthrough: dexamethasone is first drug shown to save lives. *Nature* 2020, 10.1038/d41586-020-01824-5, doi:10.1038/d41586-020-01824-5.

200. Gao, Y.; Yan, L.; Huang, Y.; Liu, F.; Zhao, Y.; Cao, L.; Wang, T.; Sun, Q.; Ming, Z.; Zhang, L., et al. Structure of the RNA-dependent RNA polymerase from COVID-19 virus. *Science* 2020, 10.1126/science.abb7498, eabb7498, doi:10.1126/science.abb7498.

201. Gordon, C.J.; Tchesnokov, E.P.; Feng, J.Y.; Porter, D.P.; Gotte, M. The antiviral compound remdesivir potently inhibits RNA-dependent RNA polymerase from Middle East respiratory syndrome coronavirus. *J Biol Chem* 2020, 295, 4773-4779, doi:10.1074/jbc.AC120.013056.

202. Yin, W.; Mao, C.; Luan, X.; Shen, D.D.; Shen, Q.; Su, H.; Wang, X.; Zhou, F.; Zhao, W.; Gao, M., et al. Structural basis for inhibition of the RNA-dependent RNA polymerase from SARS-CoV-2 by remdesivir. *Science* 2020, 10.1126/science.abc1560, 2020.2004.2008.32763, doi:10.1126/science.abc1560.

203. Jin, Z.; Du, X.; Xu, Y.; Deng, Y.; Liu, M.; Zhao, Y.; Zhang, B.; Li, X.; Zhang, L.; Peng, C., et al. Structure of M(pro) from COVID-19 virus and discovery of its inhibitors. *Nature* 2020, 10.1038/s41586-020-2223-y, doi:10.1038/s41586-020-2223-y.

204. Zhang, L.; Lin, D.; Sun, X.; Curth, U.; Drosten, C.; Sauerhering, L.; Becker, S.; Rox, K.; Hilgenfeld, R. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-ketoamide inhibitors. *Science* 2020, 368, 409-412, doi:10.1126/science.abb3405.

205. ul Qamar, M.T.; Alqahtani, S.M.; Alamri, M.A.; Chen, L.-L. Structural basis of SARS-CoV-2 3CLpro and anti-COVID-19 drug discovery from medicinal plants. *Journal of Pharmaceutical Analysis* 2020, 10.1016/j.jpha.2020.03.009, doi:10.1016/j.jpha.2020.03.009.

206. Gentile, D.; Patamia, V.; Scala, A.; Sciorintino, M.T.; Piperno, A.; Rescifina, A. Putative Inhibitors of SARS-CoV-2 Main Protease from A Library of Marine Natural Products: A Virtual Screening and Molecular Modeling Study. *Marine Drugs* 2020, 18, 225.

207. Dai, W.; Zhang, B.; Jiang, X.-M.; Su, H.; Li, J.; Zhao, Y.; Xie, X.; Jin, Z.; Peng, J.; Liu, F., et al. Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease. *Science* 2020, 368, 1331-1335, doi:10.1126/science.abb4489.
208. Wu, Y.; Wang, F.; Shen, C.; Peng, W.; Li, D.; Zhao, C.; Li, Z.; Li, S.; Bi, Y.; Yang, Y., et al. A non-competing pair of human neutralizing antibodies block COVID-19 virus binding to its receptor ACE2. *medRxiv* 2020, 10.1101/2020.05.01.20077743, 2020.05.01.20077743, doi:10.1101/2020.05.01.20077743.

209. Shi, R.; Shan, C.; Duan, X.; Chen, Z.; Liu, P.; Song, J.; Song, T.; Bi, X.; Han, C.; Wu, L., et al. A human neutralizing antibody targets the receptor binding site of SARS-CoV-2. *Nature* 2020, 10.1103/s41586-020-2381-y, doi:10.1103/s41586-020-2381-y.

210. Rogers, T.F.; Zhao, F.; Huang, D.; Beutler, N.; Burns, A.; He, W.-t.; Limbo, O.; Smith, C.; Song, G.; Woehl, J., et al. Isolation of potent SARS-CoV-2 neutralizing antibodies and protection from disease in a small animal model. *Science* 2020, 10.1126/science.abc7520, eabc7520, doi:10.1126/science.abc7520.

211. Ju, B.; Zhang, Q.; Ge, J.; Wang, R.; Sun, J.; Ge, X.; Yu, J.; Shan, S.; Zhou, B.; Song, S., et al. Human neutralizing antibodies elicited by SARS-CoV-2 infection. *Nature* 2020, 10.1038/s41586-020-2380-z, doi:10.1038/s41586-020-2380-z.

212. Wec, A.Z.; Wrapp, D.; Herbert, A.S.; Maurer, D.P.; Haslwanter, D.; Sakharkar, M.; Jangra, R.K.; Dieterle, M.E.; Lilov, A.; Huang, D., et al. Broad neutralization of SARS-related viruses by human monoclonal antibodies. *Science* 2020, 10.1126/science.abc7424, eabc7424, doi:10.1126/science.abc7424.

213. Chi, X.; Yan, R.; Zhang, J.; Zhang, G.; Zhang, Y.; Hao, M.; Zhang, Z.; Fan, P.; Dong, Y.; Yang, Y., et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. *Science* 2020, 10.1126/science.abc6952, 2020.05.08.3964, doi:10.1126/science.abc6952.

214. Lei, C.; Qian, K.; Li, T.; Zhang, S.; Fu, W.; Ding, M.; Hu, S. Neutralization of SARS-CoV-2 spike pseudotyped virus by recombinant ACE2-Ig. *Nat Commun* 2020, 11, doi:10.1038/s41467-020-16048-4.

215. Wrapp, D.; De Vlieger, D.; Corbett, K.S.; Torres, G.M.; Wang, N.; Van Breedam, W.; Roose, K.; van Schie, L.; Team, V.-C.C.-R.; Hoffmann, M., et al. Structural Basis for Potent Neutralization of Betacoronaviruses by Single-Domain Camelid Antibodies. *Cell* 2020, 181, 1004-1015 e1015, doi:10.1016/j.cell.2020.04.031.

216. Hansen, J.; Baum, A.; Pascal, K.E.; Russo, V.; Giordano, S.; Wloga, E.; Fulton, B.O.; Yan, Y.; Koon, K.; Patel, K., et al. Studies in humanized mice and convalescent humans yield a SARS-CoV-2 antibody cocktail. *Science* 2020, 10.1126/science.abb0827, eabb0827, doi:10.1126/science.abb0827.

217. Brouwer, P.J.M.; Caniels, T.G.; van der Straten, K.; Snitselaar, J.L.; Aldon, Y.; Bangaru, S.; Torres, J.L.; Okba, N.M.A.; Claireaux, M.; Kerster, G., et al. Potent neutralizing antibodies from COVID-19 patients define multiple targets of vulnerability. *Science* 2020, 10.1126/science.abc5902, eabc5902, doi:10.1126/science.abc5902.

218. Baum, A.; Fulton, B.O.; Wloga, E.; Copin, R.; Pascal, K.E.; Russo, V.; Giordano, S.; Lanza, K.; Negron, N.; Ni, M., et al. Antibody cocktail to SARS-CoV-2 spike protein prevents rapid mutational escape seen with individual antibodies. *Science* 2020, 10.1126/science.abb0831, eabb0831, doi:10.1126/science.abb0831.

219. Hotez, P.J.; Corry, D.B.; Bottazzi, M.E. COVID-19 vaccine design: the Janus face of immune enhancement. *Nat Rev Immunol* 2020, 20, 347-348, doi:10.1038/s41577-020-0323-4.

220. World Health Organization. Draft landscape of COVID-19 candidate vaccines. Available online: [https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines](https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines) (accessed on June 19, 2020).
221. Thanh Le, T.; Andreadakis, Z.; Kumar, A.; Gómez Román, R.; Tollefsen, S.; Saville, M.; Mayhew, S. The COVID-19 vaccine development landscape. *Nat Rev Drug Discov* **2020**, *19*, 305-306, doi:10.1038/d41573-020-00073-5.

222. Wang, F.; Kream, R.M.; Stefano, G.B. An Evidence Based Perspective on mRNA-SARS-CoV-2 Vaccine Development. *Med Sci Monit* **2020**, *26*, e924700, doi:10.12659/msm.924700.

223. Zhu, F.-C.; Li, Y.-H.; Guan, X.-H.; Hou, L.-H.; Wang, W.-J.; Li, J.-X.; Wu, S.-P.; Wang, B.-S.; Wang, Z.; Wang, L., et al. Safety, tolerability, and immunogenicity of a recombinant adenovirus type-5 vectored COVID-19 vaccine: a dose-escalation, open-label, non-randomised, first-in-human trial. *The Lancet* **2020**, 10.1016/S0140-6736(20)31208-3, doi:10.1016/S0140-6736(20)31208-3.

224. Feldmann, M.; Maini, R.N.; Woody, J.N.; Holgate, S.T.; Winter, G.; Rowland, M.; Richards, D.; Hussell, T. Trials of anti-tumour necrosis factor therapy for COVID-19 are urgently needed. *The Lancet* **2020**, *395*, 1407-1409, doi:10.1016/S0140-6736(20)30858-8.

225. Conti, P.; Ronconi, G.; Caraffa, A.; Gallenga, C.E.; Ross, R.; Frydas, I.; Kritas, S.K. Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by Coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies. *J Biol Regul Homeost Agents* **2020**, *34*, doi:10.23812/CONTI-E.

226. Thevarajan, I.; Nguyen, T.H.O.; Koutsakos, M.; Druce, J.; Caly, L.; van de Sandt, C.E.; Jia, X.; Nicholson, S.; Catton, M.; Cowie, B., et al. Breadth of concomitant immune responses prior to patient recovery: a case report of non-severe COVID-19. *Nature Medicine* **2020**, *26*, 453-455, doi:10.1038/s41591-020-0819-2.

227. Grifoni, A.; Weiskopf, D.; Ramirez, S.I.; Mateus, J.; Dan, J.M.; Moderbacher, C.R.; Rawlings, S.A.; Sutherland, A.; Premkumar, L.; Jadi, R.S., et al. Targets of T cell responses to SARS-CoV-2 coronavirus in humans with COVID-19 disease and unexposed individuals. *Cell* **2020**, 10.1016/j.cell.2020.05.015, doi:10.1016/j.cell.2020.05.015.

228. Braun, J.; Loyal, L.; Frentsch, M.; Wendisch, D.; Georg, P.; Kurth, F.; Hippenstiel, S.; Dingeldey, M.; Kruse, B.; Fauchere, F., et al. Presence of SARS-CoV-2 reactive T cells in COVID-19 patients and healthy donors. *medRxiv* **2020**, 10.1101/2020.04.17.20061440, 2020.2004.2017.20061440, doi:10.1101/2020.04.17.20061440.

229. Long, Q.-X.; Tang, X.-J.; Shi, Q.-L.; Li, Q.; Deng, H.-J.; Yuan, J.; Hu, J.-L.; Xu, W.; Zhang, Y.; Lv, F.-J., et al. Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections. *Nature Medicine* **2020**, 10.1038/s41591-020-0965-6, doi:10.1038/s41591-020-0965-6.

230. Iwasaki, A.; Yang, Y. The potential danger of suboptimal antibody responses in COVID-19. *Nature Reviews Immunology* **2020**, *20*, 339-341, doi:10.1038/s41577-020-0321-6.

231. Kulikarni, R. Antibody-Dependent Enhancement of Viral Infections. *Dynamics of Immune Activation in Viral Diseases* **2019**, 10.1007/978-981-15-1045-8_2, 9-41, doi:10.1007/978-981-15-1045-8_2.

232. Kissler, S.M.; Tedijanto, C.; Goldstein, E.; Grad, Y.H.; Lipsitch, M. Projecting the transmission dynamics of SARS-CoV-2 through the postpandemic period. *Science* **2020**, 10.1126/science.abb5793, eabb5793, doi:10.1126/science.abb5793.

233. Qiu, J. Covert coronavirus infections could be seeding new outbreaks. *Nature* **2020**, 10.1038/d41586-020-00822-x, doi:10.1038/d41586-020-00822-x.