Sperm-borne phospholipase C zeta-1 ensures monospermic fertilization in mice

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Sperm entry in mammalian oocytes triggers intracellular Ca\(^{2+}\) oscillations that initiate resumption of the meiotic cell cycle and subsequent activations. Here, we show that phospholipase C zeta 1 (PLCζ1) is the long-sought sperm-borne oocyte activation factor (SOAF). Plcz1 gene knockout (KO) mouse spermatozoa fail to induce Ca\(^{2+}\) changes in intracytoplasmic sperm injection (ICSI). In contrast to ICSI, Plcz1 KO spermatozoa induced atypical patterns of Ca\(^{2+}\) changes in normal fertilizations, and most of the fertilized oocytes ceased development at the 1–2-cell stage because of oocyte activation failure or polyspermy. We further discovered that both zona pellucida block to polyspermy (ZPBP) and plasma membrane block to polyspermy (PMBP) were delayed in oocytes fertilized with Plcz1 KO spermatozoa. With the observation that polyspermy is rare in astacin-like metalloendopeptidase (Astl) KO female oocytes that lack ZPBP, we conclude that PMBP plays a more critical role than ZPBP in vivo. Finally, we obtained healthy pups from male mice carrying human infertile PLCζ1 mutation by single sperm ICSI supplementation with Plcz1 mRNA injection. These results suggest that mammalian spermatozoa have a primitive oocyte activation mechanism and that PLCζ1 is a SOAF that ensures oocyte activation steps for monospermic fertilization in mammals.

At mammalian fertilization, sperm entry causes Ca\(^{2+}\) oscillations, repetitive acute increases and decreases in cytosolic Ca\(^{2+}\) levels lasting for several hours in human and mouse eggs, which trigger not only resumption of the meiotic cell cycle but also block polyspermy, the entry of multiple sperm heads into the ooplasm. These events are collectively referred to as oocyte activation and are prerequisite for normal subsequent mitosis and embryonic development.

The mechanism of how sperm generate the Ca\(^{2+}\) transients in oocytes have been studied for decades. Microinjection studies introducing extracts from rabbit, hamster, or boar spermatozoa have strongly implied that, in mammals, protein(s) in sperm soluble fraction can trigger Ca\(^{2+}\) oscillations comparable to those caused by fertilization. Such proteins are termed as sperm-borne oocyte activation factor (SOAF). So far, multiple candidates for SOAF have been raised by microinjection studies. However, injection studies of purified protein or analysis of gene knockout (KO) mice showed discrepancies in identifying the SOAF.

A sperm-specific phospholipase C zeta 1 (PLCζ1) was identified as a candidate for SOAF. In somatic cells, phospholipase C (PLC) mediates the digestion of phosphatidylinositol 4,5-bisphosphate (PIP\(_2\)) to generate inositol triphosphate (IP\(_3\)). These cause the release of Ca\(^{2+}\) from the endoplasmic reticulum. PLCζ1 was also shown that it can induce Ca\(^{2+}\) oscillations in oocytes by releasing Ca\(^{2+}\) from the oocyte's endoplasmic reticulum at levels far above the unstimulated very low basal intracytoplasmic Ca\(^{2+}\) concentrations. However, Plcz1 KO mouse was reported preliminary at a conference as male infertile because of defective spermatogenesis.

In this report, we generated Plcz1 KO mice with the CRISPR/Cas9 system and revealed that PLCζ1 plays an essential role in ICSI fertilization but not in normal fertilization. While no Ca\(^{2+}\) spikes were observed in ICSI fertilized oocytes, atypical Ca\(^{2+}\) spikes were observed in all oocytes from normal fertilization. Very recently, Hachem et al., generated Plcz1 KO mice and reported similar results, but there are significant discrepancies between that report and our data regarding Ca\(^{2+}\) spikes. We further discovered that the PMBP plays a more critical role than ZPBP in vivo. Our findings revealed the existence of a PLCζ1 independent oocyte activation mechanism and clarified the role of PLCζ1 in ensuring monospermic fertilization.

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Results

Generation of Plcz1 KO mice and oocyte activation in micromanipulation. Because conventional gene knockout approaches usually replace one to several exons with a drug-resistant gene cassette, there remains a risk of unexpected effects from the truncated open reading frames and the exogenously introduced promoter15. To exclude such possible artifacts, here we utilized a CRISPR/Cas9 approach that deleted all the coding exons (exons 2–14) to generate Plcz1 KO mice (Supplementary Fig. 1). Absence of the PLCζ1 protein was confirmed by western blotting analysis. Examination of our Plcz1 KO mice did not show any defects in spermatogenesis, sperm morphology, motility, or acrosome reaction rates (Supplementary Fig. 1).

To examine the oocyte activation abilities of Plcz1 KO spermatozoa, we performed intracytoplasmic sperm injection (ICSI) using epididymal spermatozoa. All oocytes injected with wild-type (WT) sperm heads formed pronuclei (PN), and most of them developed to 2-cell embryos (100% and 76.8%, respectively, n = 69; Fig. 1a). In contrast, oocytes injected with Plcz1 KO sperm heads failed to form PN or develop to 2-cell stage embryos (1.6% and 0%, respectively, n = 62; Fig. 1a), similarly to sham ICSI treatments (3.3% and 3.3%, respectively, n = 61; Supplementary Fig. 2a). Recordings of intracellular Ca2+ changes in these ICSI oocytes revealed that Plcz1 KO
sperm heads lost the ability to induce rises in intracellular Ca\(^{2+}\) (Fig. 1b). No spikes were observed even after up to three sperm heads were injected (Supplementary Fig. 2b). Injection of whole spermatozoa also resulted in quiescent oocytes indicating no SOAF activity remained (Fig. 1b). These data strongly support the idea that PLC\(\zeta\)1 is the SOAF.

**Fecundity of Plcz1 KO male and oocyte activation in physiological conditions.** The impact of PLC\(\zeta\)1 deficiency on male fertility was then examined by mating experiments with WT female mice. Copulations and vaginal plug formation were observed at normal rates for Plcz1 KO males. Unexpectedly, Plcz1 KO male mice sired healthy pups consistently, although the number of pups per copulation was significantly reduced (WT 8.9 pups vs. KO 2.3 pups; Fig. 2a). We then collected the oocytes from females mated with WT or Plcz1 KO males and examined fertilization status (Fig. 2b). With WT males, all oocytes showed monospermy with a distinct male PN at 12 h after coitus. With Plcz1 KO males, more than 80% of oocytes (90/107) were fertilized, but fewer than one-half formed 2PN (41/90) (Fig. 2c). The remaining fertilized oocytes were observed to experience activation failure (no PN but with fertilization cone(s) at 12 h, indicated as 0PN; 15/90), abnormal activation (1PN; 10/90), or polyspermy (\(\geq\)3PN; 24/90; Fig. 2c). When we recovered and cultured fertilized oocytes after mating with WT or KO males, increases in PN numbers were observed in some oocytes fertilized by KO sperm, implicating delays in fertilization and/or oocyte activation with Plcz1 KO sperm (Supplementary Table 1). When we further cultivated 2PN oocytes and counted blastocysts at 108 h after coitus, preimplantation developmental...
ability was lower in oocytes fertilized by KO sperm than WT sperm (29/90 = 32.2% and 56/62 = 90.2%, respectively) (Supplementary Table 1). Thus, although Plcz1 KO male mice are not sterile, the combined defects of activation failure and polyspermy may explain the reduced litter sizes, caused by male subfertility in vivo.

To examine further the sperm fertilizing ability and oocyte activation ability, we performed in vitro fertilization (IVF) experiments at various sperm concentrations (2, 10, and 50 × 10⁶ spermatozoa/ml) (Fig. 3a, Supplementary Table 2). Fertilization rates were similar between WT and KO groups and increased along with sperm concentration. Characteristically, activation failure was evident with Plcz1 KO spermatozoa at lower sperm concentrations (WT 0% vs. KO 12.4% at 2 × 10⁵/ml), and the polyspermy increased up to ~80% at higher sperm concentrations (WT 7.6% vs. KO 82.4% at 50 × 10⁵/ml; Fig. 3a). We next used live imaging to monitor intracellular Ca²⁺ concentrations. When we focused on monospermic fertilized oocytes, all the IVF fertilized oocytes with Plcz1 KO spermatozoa demonstrated intracellular Ca²⁺ spikes regardless of PN formation (Fig. 3b, Supplementary Fig. 2c). The Ca²⁺ spike patterns were aberrant in these oocytes (usually with decreased amplitude), and the number of spikes significantly decreased (WT 12.0 ± 5.68 spikes, 30 eggs vs. KO 2.75 ± 0.56 spikes, 28 eggs, P < 0.05). When we analysed the oocytes fertilized by a single spermatozoon retrospectively, the number of spikes observed in the 0PN oocytes were significantly fewer than those in the oocytes with formed PNs (Fig. 3c), implicating the existence of a threshold of intracellular Ca²⁺ spikes to trigger PN formation. Consistently, as more spermatozoa fused, the number of Ca²⁺ spikes increased (Fig. 3d), and more oocytes resumed the cell cycle (Fig. 3e). These findings revealed that mammalian spermatozoa have an oocyte activation ability independent from PLCζ. However, a single spermatozoon is insufficient and rarely triggers the resumption of the meiotic cell cycle, with incomplete oocyte activation leading to polyspermy.

Polyspermy block systems in oocytes fertilized by KO spermatozoa. There are two major mechanisms to prevent polyspermy in mammals, the ZPB and the PMBP. The process of the ZPB is well documented. After fertilization, Ca²⁺ oscillations induce the cortical reaction (CR), an exocytosis event that releases the Astl from the cortical granules into the extracellular space. This process exposes glycosylated proteins on the cell surface that can be visualized using *Lens culinaris* agglutinin–fluorescein isothiocyanate complex (LCA–FITC) labelling. The released Astl cleaves the ZP2 protein and changes the ZP structure to prevent excess sperm entry. When we visualized intracellular Ca²⁺ and the CR by LCA–FITC labelling, a delayed and incomplete CR was observed coincident with a delayed Ca²⁺ spike by ~1 h that also had a lower amplitude in oocytes fertilized with Plcz1 KO spermatozoa (Fig. 4a). Some oocytes failed to induce the CR during the initial Ca²⁺ spike. Immunoblotting indicated that ZP2 cleavage was completed by 4 h after insemination with Plcz1 KO spermatozoa, whereas it was completed in 1 h with WT spermatozoa (Fig. 4b).

Ca²⁺ is also required for the PMBP, but its mechanism remains unknown. Here, we assessed the establishment of PMBP by counting the number of spermatozoa fused with ZP-free oocytes (fusion index) at several time points. While the fusion index with WT spermatozoa plateaued at ~2 sperm/oocyte 60 min after insemination as reported previously (2.0 ± 0.54 at 60 min and 2.0 ± 0.51 at 80 min, respectively), the fusion indexes with Plcz1 KO spermatozoa were significantly higher than with the WT and continued to increase even after 60 min (4.1 ± 1.14 at 60 min and 4.6 ± 1.12 at 80 min, respectively; Fig. 4c). We also examined PMBP status 12–15 h after the 1st insemination. Oocytes fertilized with WT spermatozoa or with Plcz1 KO spermatozoa rarely fused with WT spermatozoa at the 2nd insemination (Supplementary Fig. 3a, b). Therefore, we conclude that both the ZPB and PMBP were delayed but eventually established in the oocytes fertilized with Plcz1-deficient spermatozoa.

The significance of these polyspermy blocking mechanisms was further challenged using Astl KO mice, which lacks the ZP2 cleavage. When Astl KO female mice were mated with WT male mice, the ZP2 protein remained intact after fertilization (Supplementary Fig. 3c), and extra spermatozoa accumulated in the perivitelline space indicating failure of the ZPBP, but the PMBP was still functional and all the oocytes exhibited 2PN (Fig. 4d). This process exposes glycosylated proteins on the cell surface that can be visualized using *Lens culinaris* agglutinin–fluorescein isothiocyanate complex (LCA–FITC) labelling. The released Astl cleaves the ZP2 protein and changes the ZP structure to prevent excess sperm entry. When we visualized intracellular Ca²⁺ and the CR by LCA–FITC labelling, a delayed and incomplete CR was observed coincident with a delayed Ca²⁺ spike by ~1 h that also had a lower amplitude in oocytes fertilized with Plcz1 KO spermatozoa (Fig. 4a). Some oocytes failed to induce the CR during the initial Ca²⁺ spike. Immunoblotting indicated that ZP2 cleavage was completed by 4 h after insemination with Plcz1 KO spermatozoa, whereas it was completed in 1 h with WT spermatozoa (Fig. 4b).

Rescue of the infertility of Plcz1 mutants by PLCζ supplementation. Human genetic diseases are mainly caused by small mutations rather than gene deletions. There are several Plcz1 point mutations reported in infertile men. Also, several artificial mutations were shown to influence oocyte activation ability as determined by injecting specific mRNAs into mouse oocytes (Supplementary Fig. 4a, b). Here, we phenocopied the human infertility-associated mutations in mice and introduced H435P (an equivalent mutation of human H398P) as well as D210R (an enzymatically dead mutation)10 mutations into mice using CRISPR/Cas9 technology (Supplementary Fig. 4c–e). Both mutant mRNAs were transcribed at WT levels in mutant mouse testes (Fig. 5a). Interestingly, when sperm proteins were analysed by immunoblotting, while the D210R protein was detected at ~74 kDa in similar amounts to WT mice, no H435P protein was detected at ~74 kDa (all 5 males examined for each genotype; Fig. 5b, Supplementary Fig. 5a). It should be noted that we occasionally observed ~20 kDa signal only in H435P sperm (3/5 males; Supplementary Fig. 5a), implicating the instability of H435P protein in vivo. Homozygous male mice carrying D210R or H435P mutations phenocopied Plcz1 KO male mice in spermatogenesis, fertility, IVF, and ICSI outcomes (Fig. 5c, Supplementary Fig. 5b–d). By using these point mutant mice as male infertility models, we complemented oocyte activation with WT Plcz1 mRNA injection. The optimized concentrations of human and mouse Plcz1 mRNA for the activation of mouse oocytes were adopted in the study. When we injected mouse Plcz1 mRNA (2 ng/μl × 1–3 pl) 1 h after ICSI using mutant spermatozoa, Ca²⁺ oscillations were induced by successful 2PN formation (Fig. 5d). The injection of 0.2 ng/μl human Plcz1 mRNA also successfully activated oocytes. Healthy pups were obtained after transplantation of the embryos obtained with Plcz1 mRNA injection (Fig. 5e, Table 1).
Figure 3. PLCζ1-independent oocyte activation causes abnormal fertilization. (a) The percentages of oocytes with each number of PN at 12 h after insemination. (b) Intracellular Ca\(^{2+}\) changes in oocytes after fertilization. Ratios of GEM-GECO fluorescence of oocytes with each genotype/number of pronuclei (representatives are shown). The numbers of PN formed (top left), and the mean numbers of Ca\(^{2+}\) spikes observed in oocytes (bottom right) are indicated. The numbers in parentheses indicate the numbers of oocytes observed. (c) Ca\(^{2+}\) spikes for oocytes fertilized by a single spermatozoon from Plcz1 KO males. Numbers of Ca\(^{2+}\) spikes for each number of pronuclei are indicated. Oocytes with one or two PN contained significantly more spikes than 0PN oocytes (*P < 0.005). (d) Ca\(^{2+}\) spikes for oocytes fertilized with differing numbers of Plcz1 KO sperm. Number of Ca\(^{2+}\) spikes for oocytes fertilized with 2 spermatozoa contained significantly more spikes than that fertilized with a single spermatozoon (*P < 0.005). (e) The percentages of oocytes that formed PN. Percentages are based on total numbers of oocytes fertilized with indicated number of Plcz1 KO sperm.
Discussion

Here, we have demonstrated that PLCζ1 is the long-sought SOAF in mice as indicated by the complete deletion of Ca\(^{2+}\) spikes in ICSI (Fig. 1). Further, we found that the existence of a PLCζ1-independent oocyte activation mechanism through normal fertilization where sperm–oocyte membrane interaction occurs. The activity is weak and barely triggers meiotic cell cycle resumption. Thus, this sperm–oocyte membrane interaction is probably a primitive but intrinsic mechanism for oocyte activation.

The results contrast with previous work reported at a conference in which Plcz1 KO male mice were presented to be sterile with defective spermatogenesis\(^{13}\). However, this was likely to be an artifact of the KO strategy that produced a truncated product. Very recently, Hachem \textit{et al}., reported a similar phenotype of Plcz1 knockout mice\(^{14}\), but there is a notable difference. While we observed atypical but clear Ca\(^{2+}\) oscillations in all the oocytes after normal fertilization, Hachem \textit{et al}., only found a single spike in one out of forty oocytes. In the present study, by labelling fused sperm chromosomes with Histon H2B-mCherry (Fig. 3b), we only analysed fertilized oocytes. Although the fertilization rate was not indicated, incorporation of unfertilized oocytes in the denominator may explain the difference. The correlation of Ca\(^{2+}\) spikes with the number of fused spermatozoa suggested that spermatozoa have a PLCζ1-independent Ca\(^{2+}\) spike inducing ability (Fig. 3c–e).

In the previous studies with electropermeabilization, more than four Ca\(^{2+}\) spikes were required to induce successful CR and subsequent cell cycle resumption\(^{24,25}\). However, we recently reported that CR completed within a few Ca\(^{2+}\) spikes during normal fertilization\(^{17}\). In this study, we found a significant difference in the number of Ca\(^{2+}\) spikes observed in (0PN) and activated (1 or 2PN) oocytes (2.30 ± 0.48 vs. 2.96 ± 0.56) with Plcz1 KO sperm. Further analysis will be required, however, our data implicates that the mouse oocyte requires an average of three Ca\(^{2+}\) spikes to be activated.

Figure 4. Delayed PMBP establishment by Plcz1 deletion is a cause of polyspermy. (a) Representative fluorescence plots for R-GECO (intracellular Ca\(^{2+}\); gray) and LCA–FITC staining (CR; black) of oocytes after insemination. Black arrowheads highlight Ca\(^{2+}\) spikes that induced the CR, while white arrowheads indicate Ca\(^{2+}\) spikes that failed to initiate the CR. (b) ZP2 immunoblot of oocytes after insemination. Intact ZP2 and the cleaved C-terminal fragments of ZP2 were 120 kD and 90 kD, respectively. A full-length blot is presented in Supplementary Figure 7a. (c) PMBP establishment estimated with the number of fused sperm of \textit{in vitro} fertilized oocytes. The data reflect the means of separate experiments for each genotype. (d) Oocytes collected from oviduct of Astl heterozygous or KO females mated with WT male. Bright field observation (upper) and nuclear staining (lower) using Hoechst 33342 (blue) at 12 h post coitum are indicated. Oocytes from Astl KO females were denuded to prevent spermatozoa within the perivitelline space (arrowheads) interfering with pronuclei counting. The numbers of 2PN oocytes/total oocytes from five females for each genotype are indicated. No KO oocyte exhibited polyspermy. Scale bar = 50 μm.
There are several mechanisms proposed for Ca\(^{2+}\) dependent oocyte activation through sperm–oocyte interaction\(^{26}\), such as the Ca\(^{2+}\) conduit model found in Caenorhabditis elegans\(^{27}\), and a membrane receptor model found in Xenopus\(^{28}\). Further study with Plcz1 KO mice will help to elucidate which mechanism(s) are used in mammals.

It is interesting to note that newts, birds and insects are known to require polyspermy to ensure oocyte activation but use only a single decondensed sperm head to drive further development and ensure the correct diploid chromosome set in embryogenesis. However, polyspermy compromises further development in mammals. Here, we showed that PLCζ1 plays critical roles in both ZPBP and the PMBP. This ensures monospermic fertilization with successful oocyte activation and the prevention of extra spermatozoa from entering the ooplasm. We also showed that PMBP, rather than ZPBP, plays critical roles in the monospermic fertilization in vivo in mice. This finding will shed light onto development and the roles of these two mechanisms in various organisms.

In infertility clinics, because only a few oocytes are collected during retrieval, conventional IVF protocols have largely been replaced with ICSI protocols to increase the success rate and decrease the superovulatory burden on women\(^{29}\). Although ICSI ensures monospermic fertilization, oocyte activation failures still occur and have been reported as a cause of unexplained infertility\(^{30}\). As we observed polyspermy in Plcz1 mutant mice, such patients might have polyspermy in vivo as well as oocyte activation failure following ICSI. If it is caused by male infertility, this form could possibly be treated by a combination of ICSI and PLCζ1 complementation in vitro.
Methods

Animals.  Wild-type mice (C57BL/6 N × DBA/2) F1, also known as B6D2F1, were purchased from CLEA Japan (Tokyo, Japan) or Japan SLC (Shizuoka, Japan). All animal experiments were approved by the Animal Care and Use Committee of the Research Institute for Microbial Diseases, Osaka University, under ethics approval number Biken-AP-H25-02-4, and were performed in accordance with the relevant guidelines and regulations.

Generation of *Plcz1* knockout mice with the CRISPR/Cas9 system. Two pX330 plasmids expressing single guide (sg)RNAs targeting the 5′ and 3′ regions of *Plcz1* (5′–TAGAGAAAGAGCCCCTATG–3′ and 5′–GTGGCACAATTTGAAACCTTCC–3′) were co-transfected with human codon-optimized CAS9 to remove the coding region of *Plcz1* in EGR-G101 (C57BL/6) ES cells. Embryonic stem (ES) cell clones were selected with puromycin. Correctly targeted ES cell clones and germ-line transmission were determined via polymerase chain reaction (PCR) using primers for the KO allele (primer c: 5′–GACCATCTGTTCGATG–3′ and primer d: 5′–AGCACTGAGAATGCAACCC–3′) or the wild-type (WT) allele (primer a: 5′–ATGACTAGGGAGGAGAC–3′ and primer b: 5′–ATTCCCATGACCACTACTACC–3′). A founder mouse with a 50.8 kbp deletion was used to expand the colony.

Intracytoplasmic sperm injection (ICSI).  ICSI was performed as described. In brief, each sperm head was separated from the tail by applying a few piezo pulses, then injected into a denuded MII oocyte using a piezo manipulator (PrimeTech, Ibaraki, Japan). Whole sperm injection was carried out after immobilization of motile spermatozoa by a piezo pulse.

Fertility testing.  Sexually mature male mice of each genotype were caged with B6D2F1 sexually mature female mice. Copulation was confirmed by checking for vaginal plugs every morning and the numbers of pups were counted after caesarean section at 17.5 days post coitum.

**In vivo fertilization assay.**  WT female mice were injected with pregnant mare serum gonadotropin and human chorionic gonadotropin (hCG) at 48 hours intervals. Twelve hours after hCG injection, females were caged with control or mutant male mice. The female mice that copulated with males within 60 min were used in the study. Oocytes were collected from oviducts 7–8 h after coitus with male mice of each genotype and incubated in potassium-supplemented simplex optimized medium (KSOM) medium. The numbers of pronuclei were counted at 12 and 15 h post coitum. Subsequent embryonic development was observed to the blastocyst stage. For the analysis of *Astf* KO oocytes, heterozygous or homozygous KO female mice were superovulated, mated with WT males, and oocytes were collected as described above. To count the number of pronuclei, oocytes were stained with Hoechst 33542 (1 μg/ml, for 10 min) at 12 h after coitus. Zona removal with collagenase (Wako, Osaka, Japan) treatment at 100 μg/ml for 10 min was carried out for KO oocytes to remove sperm fluorescent signals in the perivitelline space.

**In vitro fertilization (IVF).**  IVF was performed as described. In brief, mature oocytes were collected and placed in 100 μl of Toyoda, Yokoyama, and Hoshi (TYH) medium. Spermatozoa were collected from the epididymides of sexually mature male mice of each genotype, and incubated in TYH medium for 2 h for capacitation. Capacitated sperm were added to the drop containing oocytes at a final concentration of 2–50 × 10^5 sperm/ml. After 5 h of co-incubation, oocytes were washed and transferred to KSOM medium.

**Sperm motility analysis.**  Sperm motility was analysed as described. Cauda epididymal spermatozoa were suspended in TYH medium and 10^3 sperm/ml. IVF was performed as described. In brief, mature oocytes were collected and placed in 100 μl of Hepes-CZB medium, followed by incubation in fresh KSOM medium at 37 °C under 5% CO₂ in humidified air for 3 h. After ICSI, the oocytes were incubated for at least 10 min to allow membrane sealing, followed by quick transfer into glass-bottomed chambers for observation of pronuclear formation and embryo development using spinning-disk confocal microscopy. The second polar body was distinguishable from the first polar body by its positive mCherry signal. The formation of pronuclei was examined both by bright field observation and by the sized/shape of the signals in the perivitelline space.

**Analysis of acrosome reaction.**  Acrosomal exocytosis was analysed as described. Spermatozoa expressing enhanced green fluorescent protein (EGFP) in the acrosome were suspended and incubated in TYH medium. At 30 min and 3 h after incubation, an aliquot of the suspension was stained with propidium iodide (final 10 μg/ml) and subjected to fluorescence-activated cell sorting with an EC800 cell analyzer (Sony, Tokyo, Japan). The viability and acrosomal integrity of sperm were determined using the CEROS sperm analysis system (software version 12.3; Hamilton Thorne Biosciences, Beverly, MA, USA) at 30 min and 3 h after incubation.

**Nuclear/pronuclear imaging.**  MII oocytes were subjected to microinjection in 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES)-CBZ medium with mRNA for histone H2B-mCherry (5 ng/μl) (a kind gift from Dr. Kazuo Yamagata from Kindai University, Wakayama, Japan), followed by incubation in fresh KSOM medium at 37 °C under 5% CO₂ in humidified air for 3 h. After ICSI, the oocytes were incubated for at least 10 min to allow membrane sealing, followed by quick transfer into glass-bottomed chambers for observation of pronuclear formation and embryo development using spinning-disk confocal microscopy. The second polar body was distinguishable from the first polar body by its positive mCherry signal. The formation of pronuclei was examined both by bright field observation and by the sized/shape of the mCherry fluorescence.

**Ca^{2+} imaging.**  Ca^{2+} imaging was performed as described. Metaphase (M) stage II oocytes were subjected to microinjection with a mixture of mRNAs for GEM-GECO (60 ng/μl) and histone H2B-mCherry (5 ng/μl) in Hepes-CZB medium, followed by incubation in fresh KSOM medium at 37 °C under 5% CO₂ in humidified air for 3 h. ICSI was performed as described above within 5 min. IVF was performed as described within 10 min using oocytes subjected to partial zona dissection. After fertilization by either method, the oocytes were quickly...
transferred into glass-bottomed chambers for observation using spinning-disk confocal microscopy. Images of Ca\textsuperscript{2+} dynamics were taken at 20-s intervals for 5 h. After Ca\textsuperscript{2+} imaging, images of nuclei/pronuclei with H2B-mCherry were acquired at 15-min intervals for 24 h for evaluation of the number of pronuclei and fused sperm.

**Imaging of the cortical reaction.** Oocytes were observed as described\textsuperscript{17}. Oocytes microinjected with R-GECO (30 ng/μl) were inseminated with capacitated spermatozoa at a concentration of 1.0 × 10\textsuperscript{5}/ml for 10 min. They were observed in CZB medium containing Lens culinaris agglutinin-fluorescein isothiocyanate (LCA–FITC) (5 ng/ul) (J-Oil Mills, Tokyo, Japan).

**Zone pellucida protein (ZP2) cleavage assay.** ZP2 cleavage assay was performed as described\textsuperscript{17}. Oocytes were collected 0.5, 1, 1.5, 2, 3, and 4 h after IVF and lysed in 3 × Tris-glycine sodium dodecyl sulfate (SDS) loading buffer, separated on 5–20% Tris-glycine gels by SDS–polyacrylamide gel electrophoresis (PAGE), transferred to polyvinylidene fluoride membranes (Invitrogen Life Technologies, Carlsbad, CA, USA), blocked in 7.5% nonfat milk in Tris–Buffered Saline containing 0.05% Tween 20, and probed with primary antibodies for ZP2 (gift from Dr. Jurrien Dean of the National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda, MD, USA), followed by secondary antibodies conjugated with horseradish peroxidase\textsuperscript{38}. Chemiluminescence was performed with ECL Plus (GE Healthcare, Little Chalfont, UK), and signals were acquired with the Luminescent Image Analyzer LAS-3000 (Fujiﬁlm, Tokyo, Japan).

**Plasma membrane block to polyspermy (PMBP) assay.** The PMBP assay was performed using two methods. In the ﬁrst method, the fusion index was counted as described previously\textsuperscript{39}. The ZP of denuded MI oocytes were removed by incubation in TYH medium containing 100 μg/ml collagenase (Type I-S, Merck KGaA, Darmstadt, Germany) for 5 min. ZP-free oocytes were preloaded with 1 μg/ml Hoechst 33342 for 10 min and washed four times. After 20, 40, 60, or 80 min of insemination with WT or KO spermatozoa at a concentration of 0.5 × 10\textsuperscript{5}/ml, the oocytes were ﬁxed in 0.25% glutaraldehyde and observed. Only the fused sperm heads were detected as ﬂuorescent positive and counted. In the second method, oocytes were fertilized by conventional IVF with WT or PLC\textsubscript{ζ}1-deﬁcient spermatozoa at a concentration of 0.5 × 10\textsuperscript{5}/ml, as described above. After 12–15 h of incubation in KSOM medium, the ZP of fertilized oocytes were removed by treatment with acidic Tyrode’s solution\textsuperscript{40} followed by staining with Hoechst 33342 for 10 min and four washes. After 30 min of insemination using WT sperm at a concentration of 0.5 × 10\textsuperscript{5}/ml, the oocytes were ﬁxed with 0.25% glutaraldehyde and observed. In this method, all the nuclei were stained and detected as ﬂuorescent positive, thus fused and non-fused sperm nuclei were distinguished by size and shape.

**Antibodies.** Rabbit polyclonal antibody was produced by immunization with mouse PLC\textsubscript{ζ}1 polypeptide (GYRVRPLFSKSGANLEPSS)\textsuperscript{10}. The monoclonal antibody against IZUMO1 (KS64–125) was obtained as described\textsuperscript{41}. The anti-ZP2 antibody used in immunoblotting analysis was a gift from Dr. Jurrien Dean of the National Institute of Diabetes and Digestive and Kidney Diseases.

**Data Availability.** Mutant Plc\textsubscript{z}1 mice used in this study are available through Riken BioResource Center, Japan (http://en.brc.riken.jp/). The stock numbers (RBRC numbers) for B6D2-Plc\textsubscript{z}1<em>em1Osb</em>, B6D2-Plc\textsubscript{z}1<em>em3(D210R)Osb</em>, and B6D2-B6-Plc\textsubscript{z}1<em>em4(H435P)Osb</em> are 10014, 10093, and 09988, respectively. All other data are available from the authors on reasonable request.

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Author Contributions
Y.S., K.N., and M.I. designed the study; all authors performed research; all authors analysed data; and Y.S., K.N., and M.I. wrote the paper.

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