Components of the Engulfment Machinery Have Distinct Roles in Corpse Processing

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Abstract

Billions of cells die in our bodies on a daily basis and are engulfed by phagocytes. Engulfment, or phagocytosis, can be broken down into five basic steps: attraction of the phagocyte, recognition of the dying cell, internalization, phagosome maturation, and acidification. In this study, we focus on the last two steps, which can collectively be considered corpse processing, in which the engulfed material is degraded. We use the Drosophila ovarian follicle cells as a model for engulfment of apoptotic cells by epithelial cells. We show that engulfed material is processed using the canonical corpse processing pathway involving the small GTPases Rab5 and Rab7. The phagocytic receptor Draper is present on the phagocytic cup and on nascent, phosphatidylinositol 3-phosphate (PI(3)P)- and Rab7-positive phagosomes, whereas integrins are maintained on the cell surface during engulfment. Due to the difference in subcellular localization, we investigated the role of Draper, integrins, and downstream signaling components in corpse processing. We found that some proteins were required for internalization only, while others had defects in corpse processing as well. This suggests that several of the core engulfment proteins are required for distinct steps of engulfment. We also performed double mutant analysis and found that combined loss of draper and αPS3 still resulted in a small number of engulfed vesicles. Therefore, we investigated another known engulfment receptor, Crq. We found that loss of all three receptors did not inhibit engulfment any further, suggesting that Crq does not play a role in engulfment by the follicle cells. A more complete understanding of how the engulfment and corpse processing machinery interact may enable better understanding and treatment of diseases associated with defects in engulfment by epithelial cells.

Introduction

Engulfment by epithelial cells is essential for the health and maintenance of several organs including the retina, lungs, and kidney [1–4]. Improper clearance can result in or exacerbate serious conditions such as retinitis pigmentosa, age-related macular degeneration, and asthma [3–5]. Despite the importance of epithelial cells in engulfment, the molecular changes within these cells that occur during engulfment are only now beginning to be elucidated. Much of the
recent progress has identified the core proteins that are required for engulfment in different cell types and across species.

Several proteins required for engulfment have been identified and investigated in *C. elegans*, *Drosophila*, and mammals. Two well-known engulfment receptors are Draper/Ced-1 and integrins. In *C. elegans*, these receptors act in two partially parallel pathways: Ced-1 working with Ced-6 and Ced-7 and integrins working upstream of Ced-2/5/12 [6, 7], both of which can activate the small GTPase Rac1 [8]. In *Drosophila*, Rac1 is activated by Ced-5/Ced-12, but it is not known what acts upstream of Ced-5/Ced-12 [9, 10]. In *Drosophila* hemocytes, integrins have recently been shown to be required for engulfment [11–14] and Draper has been shown to work in parallel to integrins in these cells [11]. Ced-12 has been shown to activate Rac1 in conjunction with another GEF, DRK/DOS/SOS, both acting downstream of Draper in phagocytic glia [10]. Conversely, in *Drosophila* hemocytes, Draper and Ced-12 were found to act in parallel pathways [15], suggesting that the engulfment pathways may differ between cell types. In mammals, the activation of Rac1 by the Ced-12 ortholog Dock180/ELMO1 has been studied extensively, although the Dock180/ELMO1 complex is usually activated by another engulfment receptor, Bai1 [16, 17]. Although Bai1 orthologs have not been reported in *Drosophila*, other conserved engulfment receptors include the CD36 family members Crq and Debris buster [18–21]. Determining the specific roles of these proteins during engulfment is an active area of investigation.

Once the engulfed material has been internalized, it is degraded through the corpse processing pathway. The corpse processing pathway has been well-characterized in *C. elegans* and mammals, indicating that the machinery is conserved across species. In *C. elegans*, phagocytic cup formation is dependent on the receptor Ced-1 and large GTPase Dynamin [22, 23]. The nascent phagosomes fuse to early and late endosomes for phagosome maturation, using the small GTPases Rab5 and Rab7, respectively [23, 24]. The late endosomes then fuse to lysosomes, mixing their contents together and degrading the engulfed material [25–28]. These events are conserved in mammals, although there are a large number of receptors in addition to Ced-1 homologues. In this study, we refer to “engulfment” as the complete process, from recognition through acidification. We refer to “corpse processing” as the process including only phagosome maturation and acidification, after internalization and formation of the phagosome. Lastly, we refer to “phagosome maturation” as the process encompassing vesicle association with Rab5- and/or Rab7-GFP.

The studies on phagosome maturation have largely been centered around the function of Rab GTPases. However, recent work has also uncovered a role for specific lipid composition changes on the phagosome surface. Phosphatidylinositol 3-phosphate (PI(3)P) serves as a signaling molecule for several different cellular processes including phagosome maturation in mammals and *C. elegans* [29, 30]. Interestingly, in *C. elegans*, PI has been shown to be phosphorylated and de-phosphorylated in cyclic waves for proper phagosome maturation, requiring two kinases and one phosphatase [29]. Phosphatidylinositol 4,5-bisphosphate (PI(4,5)P2) associates with the unsealed phagocytic cup while PI(3)P associates with newly sealed phagosomes [31]. Lipid composition, specifically the presence of PI(3)P or PI(4,5)P2, can serve as a localization cue for proteins required for phagosome maturation [31]. In *Drosophila*, phagosome maturation has been characterized in hemocytes processing engulfed bacteria [32] and epidermal cells processing degraded dendrites [20]. We have recently demonstrated that phagosome maturation markers can be detected during engulfment of dead cells by epithelial follicle cells in the *Drosophila* ovary [33]. Throughout this study, we refer to PI(3)P, Rab5, and Rab7 as markers of the canonical corpse processing pathway.

While much is known about the canonical corpse processing machinery, how the core engulfment machinery (phagocytic receptors and downstream proteins) interacts with the
corpse processing machinery is not as well understood. The phagocytic receptor Draper/Ced-1 has been shown to play different roles depending on the cell type. In *C. elegans* and some cell types in *Drosophila*, Draper/Ced-1 is required for internalization and corpse processing [34–37]. However, in hemocytes and epidermal cells of *Drosophila*, Draper is required only for corpse processing [20, 38]. Another phagocytic receptor, Croquemort, is required for internalization in hemocytes [18] and corpse processing in epidermal cells of *Drosophila* [20]. This indicates that the engulfment machinery does interface with the corpse processing machinery, and that these interactions can differ by cell type.

Here, we investigate how the engulfment machinery interacts with the corpse processing machinery in the epithelial follicle cells of the *Drosophila* ovary. The *Drosophila* ovary serves as an excellent model for studying cell death and engulfment by epithelial cells. The *Drosophila* ovary is made of chains of progressively developing egg chambers. Each mid-stage egg chamber consists of the germline-derived nurse cells and oocyte and surrounding epithelial follicle cells. Apoptotic cell death in mid-oogenesis can be induced easily by starvation [39–45]. When flies are deprived of nutrients, the germline-derived nurse cells undergo apoptosis and the surrounding somatically-derived follicle cells enlarge and engulf the dying material [39, 45]. We have uncovered some of the molecular changes required for follicle cells to engulf the dying germline [14, 45]. We found that the phagocytic receptor Draper activates the JNK pathway, working in a positive feedback loop [45]. Recently, we have found that the integrin heterodimer, αPS3/βPS, is also required for engulfment but not JNK activation [14].

In this study, we show that the epithelial follicle cells utilize the canonical corpse processing pathway to degrade the dying germline. We find that Draper is present on nascent phagosomes whereas integrins are not. Moreover, Draper functions in both internalization and corpse processing in the follicle cells, whereas integrins are required only for internalization and activation of downstream signaling molecules. Surprisingly, we found three distinct categories of mutant phenotypes: those with defects in internalization only, those with defects in internalization and phagosome maturation, and those with defects in internalization, phagosome maturation, and acidification. We also found that combined loss of two phagocytic receptors, *draper* and *αPS3*, still resulted in a small number of engulfed vesicles, which was not further affected by the loss of another phagocytic receptor, *Crq*. However, we found that *Crq* may be required to promote nurse cell death. This suggests that Draper and integrins are the major phagocytic receptors on the follicle cells. Our findings also suggest that several engulfment genes may have dual roles for internalization and corpse processing, while others are only required for internalization. This work also indicates a possible explanation for why an engulfing cell utilizes multiple engulfment receptors. Each receptor may have specific, non-overlapping functions that are crucial for successful engulfment.

**Materials and Methods**

**Fly strains and manipulations**

All strains were reared on standard cornmeal molasses fly food at 25°C unless otherwise indicated. For starvation experiments, adult flies were placed in new vials containing fly food supplemented with freshly made yeast paste for 1.5–2 days and transferred to apple juice agar vials overnight [14]. All strains were obtained from Harvard TRiP [46], the Bloomington Stock Center or Vienna *Drosophila* Resource Center unless otherwise indicated (Table 1). UAS-Rab5GFP and UAS-Rab7GFP were recombined with the FC specific driver GR1-GAL4 (Trudi Schüpbach, Princeton University, NJ, USA; [33, 45, 47] and were crossed to dsRNA lines for corpse processing analysis. Some dsRNA lines were lethal, so GR1-GAL4 was combined with tubulin-GAL80° and flies were reared at 18°C. Progeny with tubulin-GAL80° were transferred to 29°C
for 2 days to inactivate GAL80. Crosses using αPS3 dsRNA were also transferred to 29°C for 2 days as described previously [14]. All other lines were reared, conditioned, and starved at 25°C.

drprΔ [14, 45, 48] was provided by Estee Kurant (Technion-Israel Institute of Technology, Haifa, Israel), and crqKO is described in [20]. The UAS-mCherry::2XFYVE line [49] was provided by Amy Kiger (University of California, San Diego, CA, USA).

Antibody staining and microscopy
For antibody staining, flies were dissected in Grace’s medium and ovaries were fixed and stained as described previously [44]. For LysoTracker staining, flies were dissected in PBS, incubated in 1:600 LysoTracker for 5 minutes, rinsed and washed in PBS, and finally fixed in PBS, heptane, and paraformaldehyde for 20 minutes. At this point, samples were treated as with standard antibody staining. All samples were mounted in VectaShield with DAPI (Vector Labs). Primary antibodies used were: cleaved α-Dcp-1 (1:100, Cell Signaling), α-Dlg (1:100, Developmental Studies Hybridoma Bank (DSHB)), α-Draper (1:50, DSHB), α-αPS3 (1:1000, [14]), and α-β-Gal (1:400, Promega). Secondary antibodies used were goat-α-rabbit Cy3 and goat-α-mouse Alexa Fluor 647 (Jackson ImmunoResearch), each at 1:100. Egg chambers were imaged on an Olympus FV10i confocal microscope, images were processed using ImageJ and Adobe Photoshop, and figures were made using Adobe Illustrator and Graphpad Prism.

Table 1. Candidate dsRNA Screen Results.

| Bloomington or Vienna # | Allele   | Gene  | Germline engulfment defects | Other defects                          |
|------------------------|----------|-------|-----------------------------|---------------------------------------|
| 28556                  | HM05042  | Ced-12* | ***                        |                                       |
| 3799                   | GD1529   | shi*   | ***                        | Excessive FC growth at terminal end of egg chamber |
| 105971                 | KK101444 | shi    | ***                        | n.d.                                  |
| 34832                  | HMS00147 | Rab5*  | ***                        |                                       |
| 33734                  | GD10124  | dor*   | ***                        |                                       |
| 27519                  | JF02669  | Rab8   | ***                        | n.d.                                  |
| 34373                  | HMS01363 | Rab8   | ***                        |                                       |
| 28021                  | JF02855  | Cdc42  | ***                        | Double layer at the posterior         |
| 27722                  | JF02804  | kayak  | **                         |                                       |
| 100708                 | KK108017 | Src42A*| **                         |                                       |
| 27051                  | JF02377  | Rab7*  | *                          |                                       |
| 33617                  | HMS00010 | Fak56D | *                          |                                       |
| 28342                  | JF02978  | Rab35  | —                          |                                       |
| 28701                  | JF03117  | Rab2   | —                          |                                       |
| 34922                  | HMS01271 | Rab2   | —                          |                                       |
| 31688                  | JF01861  | Rab9   | —                          |                                       |
| 42942                  | HMS02635 | Rab9   | —                          |                                       |
| 28708                  | JF03135  | Rab14  | —                          |                                       |
| 34654                  | HMS01130 | Rab14  | —                          |                                       |
| 28513                  | JF03133  | shi    | —                          |                                       |
| 36921                  | HMS00154 | shi    | —                          |                                       |

Genes disrupted, Bloomington and Vienna (v) stock numbers, and any associated TRiP numbers are listed. The phenotypes are based on a 3-star system, where 3 stars (*** is the most severe, 2 stars (**) is moderate, and 1 star (*) is the least severe, and is usually associated with weak and/or variable lines. Two dashes (—) indicate no phenotype, n.d. is not determined. A pound sign (#) indicates the lines that are analyzed in this paper.

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Engulfment quantification

We quantified vesicle uptake by Dcp-1 staining as previously described [14]. A central slice was counted for each egg chamber. At least three egg chambers were analyzed for each phase (phases 1–4) and genotype, unless otherwise indicated. The number of Rab7GFP-positive or LysoTracker-positive vesicles were counted in the same way. We quantified Rab7GFP-positive vesicles using the Dcp-1/Rab7GFP merge, to verify that the vesicles quantified contained engulfed germline. To determine if phagosome maturation was occurring properly, the ratios were then calculated by dividing the average number of Rab7-positive by the average number of Dcp-1-positive vesicles. \( P \) values were determined using two-tailed \( t \) tests.

Results

Vesicles engulfed by the follicle cells are processed using the canonical corpse processing pathway

When flies are deprived of protein, the germline cells in some egg chambers in mid-oogenesis undergo apoptosis and are subsequently engulfed by the surrounding epithelial follicle cells [14, 39, 45, 50]. We previously analyzed dying egg chambers based on the state of nurse cell chromatin and characterized engulfment into five distinct phases [14, 45]. Throughout this paper, we show phase 0 (or healthy), phase 3 (mid-dying), and phase 5 (terminal) egg chambers. The nurse cell chromatin in phase 0 (healthy) egg chambers is dispersed (Fig 1A, 1D and 1G), whereas the nurse cell chromatin in phase 3 egg chambers is highly condensed (Fig 1B, 1E and 1H). By phase 5, little to no lingering nurse cell nuclear fragments remain.

To visualize phagosome maturation, we used existing transgenic lines expressing GFP fusions to Rab5 [51] and Rab7 [52] in conjunction with an antibody raised against cleaved caspase Dcp-1. We have found that this antibody against active Dcp-1 marks the dying germline and the subsequent engulfed material [14, 33, 50]. Studies in \textit{C. elegans} and \textit{Drosophila} have shown that early phagosomes are labeled with Rab5 whereas late phagosomes are labeled with Rab7 [20, 23]. We found that both Rab5 and Rab7-labeled endosomes accumulated near the apical surface of the follicle cells in healthy and early dying egg chambers (Fig 1A–1A’, 1D–1D’). In dying egg chambers, Rab5 and Rab7 were detected surrounding several of the engulfed Dcp-1-positive vesicles. As Rab5 and Rab7 are known to label maturing phagosomes, this indicates maturation of the Dcp-1-positive phagosomes. Rab5GFP was seen as puncta around Dcp-1-positive vesicles (Fig 1B–1C’), whereas Rab7GFP completely surrounded the Dcp-1-positive vesicles (Fig 1E–1F). The last step of corpse processing is fusion with lysosomes, which can be visualized using LysoTracker, an indicator for compartments of high acidification. We found that there were no large LysoTracker-positive vesicles in healthy egg chambers (Fig 1G–1G’), but there were several LysoTracker-positive vesicles seen by phase 3 (Fig 1H–1I). Our results are consistent with the previous studies performed in \textit{C. elegans} and \textit{Drosophila}, indicating that the dying germline is internalized and processed using the canonical corpse processing pathway.

Engulfment receptors have distinct expression patterns during corpse processing

Previously, we showed that Draper is enriched, internalized, and required within engulfing follicle cells [45]. Recently, we also found that loss of Draper resulted in a larger number of engulfed vesicles by phase 4 of engulfment than loss of integrins [14]. The \textit{C. elegans} ortholog of Draper, CED-1, has previously been shown to be present not only on the phagocytic cup, but also on phagosomes within the engulfing cell [53], indicating that CED-1 becomes
Engulfment Machinery in Corpse Processing
internalized with the engulfed material. The increase in internalized vesicles in the *Drosophila* ovary in *draper* mutants could be due to an increase in internalization by a Draper-independent mechanism, or because vesicles accumulate over time because of a defect in corpse processing. To distinguish between these two possibilities, we first investigated the expression patterns of Draper and integrins with respect to the phagosome maturation markers phosphatidylinositol 3-phosphate (PI(3)P) and Rab7. We obtained transgenic lines expressing mCherry fused to FYVE [49], which targets mCherry to membranes enriched with PI(3)P. PI(3)P co-localized with the engulfed vesicles near the apical surface, suggesting that they have begun to mature. Our data also provide further information regarding differences in how Draper and integrins function within an engulfing cell. These two possibilities are investigated further below.

To further investigate the engulfment pathways, we used double mutant analysis. First we generated double mutants with *draper* and follicle cell-specific RNAi against *αPS3*. Engulfment was visualized and quantified (Fig 3A–3G) using the cleaved Dcp-1 antibody to mark engulfed vesicles, and anti-Discs Large to monitor the growth of follicle cell membranes. Loss
of *draper* and *αPS3* simultaneously resulted in stronger engulfment defects in phase 3 compared to single mutants (Fig 3A–3G). Loss of *draper* normally results in some accumulation of vesicles beginning in phase 3, and the double mutant phenotypes suggest this could partially result from *draper*-independent engulfment mediated by the integrin pathway.

We also wanted to determine whether the engulfment genes *Ced-12* and *Src42A* functioned in the same pathway as *αPS3*. To analyze these genes in combination with *αPS3* by RNAi, it was necessary to recombine *UAS-αPS3-dsRNA* and *GR1-GAL4* onto the same chromosome. While this recombinant did have engulfment defects, we found that it showed a weaker phenotype in the earlier phases of engulfment (Fig 3H and 3I) compared to the trans-heterozygote (*UAS-αPS3-dsRNA*/*GR1-GAL4*, Fig 3A and 3B, [14]). Previously, we showed that *αPS3* is required for engulfment starting in phase 2 [14], however the recombinant did not show engulfment defects until phase 3 (Fig 3N and 3O). To determine if the *GR1-GAL4, αPS3* dsRNA recombinant was effectively knocking down *αPS3*, we used antibody staining to compare *αPS3* levels between the recombinant and the trans-heterozygote. Both showed reduced *αPS3* staining compared to wild-type (S1 Fig). These results suggest that there may be compensation by another pathway when *αPS3* is continuously knocked down when maintaining a permanent stock, or the recombinant may have a less efficient knockdown of *αPS3*.

Double mutants were generated by crossing the *GR1-GAL4, UAS-αPS3-dsRNA* recombinant to either a control dsRNA line (*luciferase, Fig 3H and 3I*) or dsRNA lines against genes of...
Engulfment Machinery in Corpse Processing

Interest (Ced-12 and Src42A). Combined loss of Src42A and αPS3 resulted in poor viability, but the egg chambers did not have added defects compared to single knockdowns (Fig 3G, 3H and 3I), suggesting that much like C. elegans, integrins may act through Src42A for engulfment.

Combined loss of Ced-12 and αPS3 resulted in considerably fewer vesicles taken up during phases 1 and 2 compared to single knockdowns, although there was only a moderate difference in phase 3 and no difference in phase 4 (Fig 3J, 3M and 3O). This suggests that Ced-12 may be activated in an integrin-independent manner initially, but integrins act through Ced-12 later in engulfment, unlike the pathway described in C. elegans in which integrins work through Ced-12 from the initiation of engulfment. One possibility is that Ced-12 is initially activated by another engulfment protein, perhaps Draper, which has been shown to activate Ced-12 previously (Lu, 2014), and acts in early phases of engulfment, as our data suggest. Together, these results suggest that the role of integrins may be to bind to the apoptotic cell and activate downstream signaling (via Src42A and Ced-12) while Draper may be required for internalization and processing as in other systems [37, 38].

Despite the importance of Draper and αPS3 during engulfment [11, 12, 14, 45, 48, 54, 55], loss of both receptors did not result in a complete block in internalization (Fig 3E, 3F and 3K). Another phagocytic receptor has been studied in Drosophila, Croquemort (Crq) [18–20]. Indeed, we found that crq becomes up-regulated during mid- to late engulfment (Fig 4A-4C), suggesting it could be involved. To determine whether Crq could be acting as a third engulfment receptor in the follicle cells, we examined the ovaries of crqKO mutants. The ovaries showed abnormalities in some egg chambers, where the follicle cells died prematurely, occasionally resulting in egg chambers completely devoid of follicle cells and undead germline (Fig 4D–4F) similar to what is seen when apoptotic genes are disrupted in the germline [42, 56, 57]. This severe phenotype did not occur in every egg chamber, and several healthy egg chambers were found with healthy follicle cells remaining. Also, when the nurse cells attempted to die, the chromatin did not fragment in the stereotypical fashion in phase 2 egg chambers. Often, the nurse cell nuclei formed a line, distinct from the disorganized chromatin normally seen in phase 1 and the tightly condensed chromatin seen in phase 3. This suggests that Crq may play a role in promoting nurse cell nuclear breakdown during nurse cell death.

We stained crqKO egg chambers with cleaved Dcp-1 and found that complete loss of crq resulted in no defects in vesicle uptake when compared to controls (Fig 5A–5D and 5K). As CD36 receptors can function together with other receptors, we performed experiments with double and triple mutants with αPS3 and Draper. Loss of either crq and draper, or crq and...
αPS3, resulted in no stronger defect than loss of either gene individually (Fig 5E–5H and 5K). Even the triple mutant with draper and crq null mutations combined with a knockdown of αPS3 resulted in no additional defects (Fig 5I–5K). These results suggest that crq is not required in the follicle cells for engulfment, although it may be required for follicle cell survival or for promoting nurse cell chromatin changes during cell death. The residual engulfment that occurs in draper, αPS3 double mutants may either indicate that another unknown engulfment receptor still acts in follicle cells, that there is an incomplete knockdown of αPS3, or that there is some engulfment that is not dependent on an engulfment receptor.

Because we did not find engulfment defects in crq mutants, we investigated other CD36 family receptors. We obtained knockdown or knockout lines of several CD36 receptors that are reported to be expressed in the ovary (FlyBase.org). We found that loss of emp or santa-maria did not have noticeable phenotypes. However, ovaries of peste mutants were grossly abnormal and resulted in significantly more “undead” egg chambers than seen in crq mutant ovaries, making it challenging to analyze engulfment. Another CD36 receptor previously reported to be required for engulfment, debris buster, is not expressed in the ovary, suggesting it is not required during oogenesis.
Integrins and Draper function independently of each other during corpse processing

As Draper becomes internalized and may function in a partially parallel pathway to integrins, we next asked if these and other core engulfment proteins are required for phagosome maturation or acidification. Since Rab7 association was more discrete than Rab5 association (Fig 1),

Fig 5. Croquemort is not required for engulfment by the follicle cells. (A-J) Mid-stage healthy and dying egg chambers from the indicated genotypes stained with DAPI (cyan) and antibodies against cleaved Dcp-1 (yellow) and Discs large (red). The Discs large channel was brightened in C-D and G-H to better visualize follicle cell enlargement. (A-B) Control (w^{1118}) egg chambers show normal follicle cell enlargement and engulfment. (C-D) Loss of crq alone (crq^{KO}) shows no defects in follicle cell enlargement or engulfment. (E-F) Loss of crq and draper (crq^{KO}; draper^{Δ5}) results in strong engulfment defects, similar to loss of draper alone. (G-H) Loss of crq and αPS3 (crq^{KO}; GR1-GAL4/UAS-αPS3^{dsRNA}) results in strong engulfment defects, but not stronger than loss of αPS3 alone. (I-J) Loss of crq, draper, and αPS3 (crq^{KO}; draper^{Δ5} UAS-αPS3^{dsRNA}/draper^{Δ5}; GR1-GAL4) results in strong engulfment defects, but not stronger than loss of draper and αPS3. (K) Average number of Dcp-1-positive vesicles engulfed per central slice for each phase of death. Scale bar is 50μm. All data are mean ± s.e.m. At least three egg chambers were quantified for each genotype and phase in K-M, except those noted here. For K, 19 egg chambers were quantified for w^{1118}; 32 for crq^{KO}; 44 for αPS3^{dsRNA}; 46 for draper^{Δ5}; 85 for draper^{Δ5} αPS3^{dsRNA}; 20 for crq^{KO}; draper^{Δ5}; 7 for crq^{KO} αPS3^{dsRNA}; 86 for crq^{KO}; draper^{Δ5} αPS3^{dsRNA}; 45 for crq^{KO}; draper^{Δ5} αPS3^{dsRNA}; 46 for crq^{KO}; draper^{Δ5} αPS3^{dsRNA}. Phase 2 for crq^{KO}; draper^{Δ5} and crq^{KO}; αPS3^{dsRNA} and phase 3 of crq^{KO}; αPS3^{dsRNA} have less than 3 egg chambers quantified. The data for αPS3^{dsRNA}, draper^{Δ5}, and draper^{Δ5} αPS3^{dsRNA} were first shown in Fig 3. Two-tailed t-tests were performed: ****P<0.005, ***P<0.01, **P<0.05.

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we used Rab7 to analyze potential defects in phagosome maturation in the engulfment machinery mutants (Fig 6). As all mutants had defects in internalization, we calculated the ratio of Rab7- to Dcp-1-positive vesicles to determine if fewer vesicles became Rab7-positive, indicating defects in phagosome maturation (Fig 6). This is based on the hypothesis that nascent phagosomes (Dcp-1+, Rab7-) mature to Dcp-1+, Rab7+ phagosomes. To analyze defects in acidification, we used LysoTracker, which marks acidified compartments. We also calculated the ratio of LysoTracker- to Dcp-1-positive vesicles to determine if fewer vesicles became acidified, indicating defects in acidification. Surprisingly, we found that engulfment defective mutants fell into three general categories in terms of defects in corpse processing. Loss of αPS3 or kayak (JNK pathway transcription factor) resulted in defects in internalization, but no defects in the ratio of Rab7-positive vesicles (phagosome maturation) or the ratio of LysoTracker-positive vesicles (acidification) (Fig 6A–6B, 6G–6I). For example, loss of αPS3 resulted in fewer Dcp-1-positive vesicles throughout engulfment (Fig 6G). However, the same ratio (approximately 70%) of those vesicles still became positive for Rab7, indicating that the vesicles taken up still progressed as in the control (Fig 6H). This holds true for acidification as well. The same ratio (approximately 180%) of those internalized vesicles became positive for LysoTracker, indicating that the vesicles taken up are still acidified as in the control (Fig 6I). Surprisingly, loss of Ced-12 or Src42A resulted in not only defects in internalization, but also phagosome maturation (Fig 6C–6C′, 6G and 6H). However, loss of Ced-12 or Src42A resulted in milder defects in acidification compared to draper (Fig 6D–6D′, 6I). Noticeably, the effect of the Src42A knockdown is weaker than that of Ced-12. This could be due to inefficient knockdowns or may speak to different requirements for these proteins. As seen in other systems, loss of draper resulted in defects in internalization, phagosome maturation, and acidification (Fig 6E–6F, 6G–6I). Indeed, LysoTracker labeling was almost completely abolished in draperΔ5 egg chambers (Fig 6F and 6I). Together, this suggests that some engulfment genes, such as draper, may be required for multiple steps during corpse processing while others, such as Ced-12, may be required for the efficiency of certain steps during corpse processing.

We also examined loss of known corpse processing genes shibire (dynamin–known to affect internalization), Rab5 (early endosomes), Rab7 (late endosomes), and deep orange (encoding VPS18, a lysosome biogenesis and fusion gene [58]). Loss of shibire and Rab5 resulted in a defect very similar to loss of draper. Egg chambers expressing shibiredsRNA or Rab5dsRNA took up little to no vesicles, and those that were engulfed were not processed (Rab7-positive) or acidified (Fig 7A–7B′, 7E–7G). Loss of Rab7 (shown to be an effective RNAi knockdown in [59]) had no defects in internalization, but had a reduction in acidification (Fig 7E and 7G). Interestingly, loss of dor resulted in an elevated numbers of internalized vesicles, which were processed (became Rab7-positive) in the same proportions as controls. These vesicles also became acidified, but were often extremely small, resulting in a high number of small acidified vesicles within the follicle cells (Fig 7C–7G). However, the ratio of LysoTracker- to Dcp-1-positive vesicles was lower than that of control (Fig 7G). This suggests that loss of dor may result in a decrease in acidification, either resulting from defects in lysosome biogenesis or fusion to late endosomes.

**Discussion**

Degradation of the cell corpse is crucial during engulfment. Here, we show that the epithelial follicle cells utilize the canonical corpse processing pathway to process Dcp-1-positive engulfed vesicles. Moreover, we show that engulfment receptors have distinct expression patterns during engulfment; Draper is present on the nascent, PI(3)P-positive phagosomes, whereas integrins are not. Using double mutant analysis, our data suggest that integrins may act through Src42A
Engulfment Machinery in Corpse Processing

Fig 6. Engulfment mutants have distinct effects on corpse processing. (A-F) Dying egg chambers from the indicated genotypes stained with DAPI (cyan) and antibodies against cleaved Dcp-1 or Dig (magenta). (A-A') Loss of aps3 (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-aps3dsRNA) results in reduced vesicle uptake, but some of the vesicles become Rab7-positive. (B-B') Loss of aps3 (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-aps3dsRNA) results in reduced vesicle uptake and normal acidification. (C-C') Loss of Ced-12 (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-Ced-12dsRNA) results in reduced vesicle uptake, reduced Rab7 association, and Rab7 aggregates. (D-D') Loss of Ced-12 (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-Ced-12dsRNA) results in reduced acidification, with little to no vesicles acidified at phase 3. (E-E') Loss of draper (UAS-GAL4/UAS-draperdsRNA; GR1-GAL4, UAS-Rab7GFP/+)) results in reduced vesicle uptake, and very few vesicles are Rab7 positive. (F-F') Loss of draper (draper'Δ5) results in a severe delay in acidification, with little to no vesicles acidified at phase 3. (G) Average number of Dcp-1-positive vesicles engulfed per central slice. (H) Quantified for each genotype, phase, and quantification method in G-I, except those noted here. For G, 47 egg chambers were quantified for the control ratio of Rab7-positive to Dcp-1-positive vesicles in phase 2 egg chambers and (I) ratio of LysoTracker-positive vesicles to Dcp-1-positive vesicles in phase 3 egg chambers. Egg chamber scale bar is 50 μm. Zoom scale bar is 10 μm. All data are mean ± s.e.m. At least three egg chambers were quantified for each genotype, phase, and quantification method in G-I, except those noted here. For G, 47 egg chambers were quantified for the control (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-luciferase dsRNA and UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-luciferase dsRNA); 215 for aps3dsRNA, 69 for kayakdsRNA, 82 for Ced-12dsRNA, 32 for Ssrc42AΔ5dsRNA, and 73 for draperdsRNA. For H, 8 egg chambers were quantified for control (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-luciferase dsRNA); 41 for aps3dsRNA, 6 for kayakdsRNA, 4 for Ced-12dsRNA, 5 for Ssrc42AΔ5dsRNA, and 22 for draperdsRNA. For I, 7 egg chambers were quantified for control (the mixture from G); 21 for aps3dsRNA, 13 for kayakdsRNA, 13 for Ced-12dsRNA, 29 for Ssrc42AΔ5dsRNA, and 13 for draperΔ5. Phase 1 for Ssrc42A has less than 3 egg chambers quantified. The data for control, aps3dsRNA, Ced-12dsRNA, and Ssrc42AΔ5dsRNA were first shown in Fig 3. Two-tailed t-tests were performed: ***P<0.005, **P<0.01, *P<0.05.

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and Ced-12 in a pathway partially parallel to Draper during later phases of engulfment. However, our results suggest that Ced-12 is not initially activated by integrins, but rather may be activated by Draper. These conclusions about Ssrc42A and Ced-12 could also be affected by inefficient knockdowns in the GR1-GAL4, UAS-aps3-dsRNA recombinant or the double RNAi knockdowns.

Double aps3 draper mutants had a very strong inhibition of engulfment, but it was not completely blocked. Surprisingly, the addition of the crq mutation to the draper and aps3 mutants did not result in a further block in engulfment. These results suggest that Draper and integrins are the main phagocytic receptors in the follicle cells. These results also suggest that another, yet unknown, engulfment receptor, may be required for engulfment.

Since loss of Ced-12 leads to defects in phagosome maturation but loss of integrins does not, this suggests that the role for Ced-12 in corpse processing is independent of integrins. Consistent with other systems, we found that draper mutants have defects in internalization, phagosome maturation, and acidification [37, 38]. In contrast, knockdowns of the JNK pathway transcription factor, Kayak, only have defects in internalization. Our data suggest that there may be three categories of defects: 1) internalization only (aps3, kayak); 2) internalization and phagosome maturation (Ced-12, Ssrc42A); and 3) internalization, phagosome maturation, and acidification (draper). However, given that we quantified the acidification defects in a draper null mutant and used RNAi to knockdown Ced-12 and Ssrc42A, it is possible that one or both of these genes are also required for acidification and this phenotype would only be apparent with a null mutant. The defect we see in kayak RNAi egg chambers suggests that the role of the JNK pathway is in signal amplification, and not in corpse processing during engulfment. This is consistent with our findings that loss of kayak and basket (JNK [45]) only result in significant defects by phase 3 of engulfment. To our knowledge, we provide the first systematic study of corpse processing in an epithelial layer and an investigation into the cross-talk between several core engulfment machinery components and corpse processing.

We propose the following model (Fig 8) for cross-talk between engulfment and corpse processing machinery in the follicular epithelium: in early dying phase 1 egg chambers, Draper is present on the phagocytic cup and nascent phagosomes and is required for internalization, phagosome maturation, acidification, as well as signaling to other engulfment machinery. As Draper is present on healthy follicle cells, it is responsible for the majority of the internalization in phase 1. Also in phase 1 egg chambers, we propose that Ced-12 is activated by Draper and is required for phagosome maturation. Consistent with this, integrin mutants do not show engulfment defects in phase 1 egg chambers [14]. By phase 2, integrins are present on the apical
Engulfment Machinery in Corpse Processing

Fig 7. Loss of the canonical corpse processing genes, shi and dor, result in opposite defects. (A–D) Dying egg chambers from the indicated genotypes stained with DAPI (cyan) and antibodies against cleaved Dcp-1 or Dmg (magenta). (A–A') Loss of shi (UAS-GAL4/tub-GAL80); GR1-GAL4, UAS-Rab7GFP/UAS-shiRNA results in reduced vesicle uptake, and the few vesicles that are engulfed are not Rab7-positive. (B–B') Loss of shi (UAS-GAL4/tub-GAL80); GR1-GAL4, UAS-Rab7GFP/UAS-shiRNA results in little to no LysoTracker-positive vesicles. (C–C') Loss of dor (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-dorRNA) results in elevated numbers of vesicles, and many are Rab7-positive. (D–D') Loss of dor (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-dorRNA) results in elevated numbers of LysoTracker-positive vesicles. (E) Average number of Dcp-1-positive vesicles engulfed per central slice. (F) Ratio of Rab7-positive to Dcp-1-positive vesicles in phase 2 egg chambers and (G) ratio of LysoTracker-positive vesicles to Dcp-1-positive vesicles in phase 3 egg chambers. Egg chamber scale bar is 50μm. Zoom scale bar is 10μm. At least three egg chambers were quantified for each genotype, phase, and quantification method in E–G, except those noted here. For E, 47 egg chambers were quantified for the control (UAS-GAL4/+; GR1-GAL4, UAS-Rab5GFP/UAS-luciferase and UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-luciferase); 33 for shiRNA, 15 for Rab5ΔRNA, 41 for Rab7ΔRNA; and 48 for dorΔRNA. For F, 8 egg chambers were quantified for control (UAS-GAL4/+; GR1-GAL4, UAS-Rab7GFP/UAS-luciferase); 2 for shiΔRNA, 2 for Rab5ΔRNA, and 6 for dorΔRNA. For G, 7 egg chambers were quantified for control (the mixture from E); 8 for shiΔRNA, 1 for Rab5ΔRNA, 18 for Rab7ΔRNA, and 8 for dorΔRNA. Phase 2 for shiΔRNA and Rab5ΔRNA and phase 4 for Rab7ΔRNA have less than 3 egg chambers quantified. Phase 3 for Rab5ΔRNA, LT only, has less than 3 egg chambers quantified. The data for control were first shown in Fig 3. Two-tailed t-tests were performed: ***,P<0.005, **,P<0.01, *,P<0.05.

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surface and may play a role in anchoring the engulfing cell to the dying cell, as shown in RPE cells [60, 61]. By phase 3, Draper and integrins are both active and appear to work in concert to regulate internalization and engulfment. During this phase of engulfment, JNK signaling is activated and has amplified the engulfment signal, at least in part by producing more Draper for internalization. Consistent with other systems, integrins may work through Src42A and Ced-12 during mid- to late engulfment.

We found it striking that the core engulfment machinery has distinct temporal regulation and different functions within the engulfing cell. One possibility is that different categories of proteins (receptors, kinases, adaptor proteins, etc.) inherently play different roles within engulfment. However, two engulfment receptors (Draper and integrins) have very different functions in terms of corpse processing while Src42A and Ced-12 (a tyrosine kinase and adaptor protein, respectively) are both required for efficient phagosome maturation. Another possibility is that the proteins required for corpse processing are localized to phagosomes. While loss of Ced-12 and Src42A resulted in less efficient phagosome maturation, we do not know if these proteins localize to maturing phagosomes. This will only become clear with further investigation.

The regulation of these proteins is another point of interest. As mentioned above, we do not know if Ced-12 and Src42A directly localize to phagosomes or not. Phosphatidylinositol (PI) can be differentially phosphorylated, which may serve as a regulator for interaction with some, or all, of these proteins. As we have seen, Draper is present on several phagosomes that are PI (3)P-positive, suggesting that PI(3)P may be required to maintain Draper on nascent phagosomes. However, Draper is also present on some PI(3)P-negative phagosomes, suggesting either that Draper is present on less mature phagosomes or Draper-positive, Rab7- and PI(3)P-negative vesicles are recycling endosomes, allowing for Draper to be recycled back to the apical surface. In C. elegans, the phagocytic cup is marked by PI(4,5)P2 while the sealed and nascent phagosomes are marked by PI(3)P [31]. Sorting nexins have been shown to associate with certain phosphorylated forms of PI, but not others, and their presence on phagosomes affects their maturation [35]. Association of K-Ras with endosomes has been shown to be dependent on endosomal acidic phospholipids, specifically phosphatidylserine [62]. Lipid composition may be a more general method of recruiting proteins to the appropriate vesicle membrane for efficient phagosome maturation.

Engulfment by epithelial cells is crucial for the health and maintenance of several organs throughout an organism; however, very little is known about how epithelial cells degrade the engulfed material. In this study, we show that most, but not all, of the engulfment machinery is required for efficient corpse processing. Draper is present on the phagocytic cup and PI(3)P-
positive vesicles, suggesting a possible method of recruitment of proteins, such as Src42A and Ced-12, to the proper phagosome membrane for efficient fusion with endosomes. This work suggests that the core engulfment machinery may play several roles within a cell, including internalization, corpse processing, acidification, and downstream signaling. We have established the Drosophila ovary as a powerful model for investigating the cross-talk between engulfment and corpse processing machinery in an in vivo system. There are striking similarities between the Drosophila epithelial follicle cells and mammalian retinal pigment epithelial cells [14], suggesting that the information gained here may be useful for diseases such as retinitis pigmentosa and age-related macular degeneration.

Supporting Information

S1 Fig. αPS3 is knocked down in all lines used for analysis. Dying mid-stage egg chambers from the indicated genotypes stained with DAPI (cyan) and an antibody against αPS3 (green). (A-A') Egg chambers from wild-type (w1118) flies show enrichment of αPS3 (white) on the apical surface of the follicle cells. (B-B') Knockdown of αPS3 (GR1-GAL4/UAS-αPS3dsRNA) in the follicle cells show no enrichment of αPS3. (C-C') A GR1-GAL4, UAS-αPS3dsRNA recombinant (UAS-GAL4/+; GR1-GAL4, UAS-αPS3dsRNA/UAS-luciferase) also shows no enrichment. Scale bar is 50 μm. (TIF)

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Author Contributions

Conceived and designed the experiments: TLM AKT CSH JSP KM. Performed the experiments: TLM TFJ JDT CSH JSP SE. Analyzed the data: TLM JSP KM. Contributed reagents/ materials/analysis tools: NCF. Wrote the paper: TLM KM.

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