A Primer on a Comprehensive Genetic Approach to Vascular Anomalies

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The field of vascular anomalies has grown tremendously in the last few decades with the identification of key molecular pathways and genetic mutations that drive the formation and progression of vascular anomalies. Understanding these pathways is critical for the classification of vascular anomalies, patient care, and development of novel therapeutics. The goal of this review is to provide a basic understanding of the classification of vascular anomalies and knowledge of their underlying molecular pathways. Here we provide an organizational framework for phenotype/genotype correlation and subsequent development of a diagnostic and treatment roadmap. With the increasing importance of genetics in the diagnosis and treatment of vascular anomalies, we highlight the importance of clinical geneticists as part of a comprehensive multidisciplinary vascular anomalies team.

Keywords: vascular malformations, genetic, mutation, anomalies, somatic

INTRODUCTION

Vascular anomalies encompass both vascular malformations and vascular tumors (1). Over the past few decades, our knowledge of the genetic changes underlying these tumors and malformations has grown tremendously. This has led to updates in the classification of some lesions as well as new therapeutic options. The goal of this review is to provide teams taking care of children and adults with vascular anomalies an overview of the molecular pathways involved in disease pathogenesis and an organizational framework based on the current classification system that encompasses knowledge of their genetic and molecular etiologies. We also aim to provide a roadmap for evaluation of vascular anomalies based on both clinical and genetic features and to highlight the critical role of genetic diagnosis in the classification and treatment of vascular anomalies.

Developmental Origin and Classification of Vascular Anomalies

Vascular malformations form due to errors in vascular morphogenesis, which occurs during early embryonic life, sometime between 4 and 10 weeks of gestation. Mesenchymal cells form primitive groups, called hemangioblasts, that differentiate into the four types of circulatory vessels (arterial, venous, capillary, lymphatic) through a series of remodeling, pruning, and continued growth. This is a process referred to as angiogenesis. The molecular mechanisms that govern angiogenesis are complex, with a series of genes controlling each critical step, from blood island formation, to endothelial cell differentiation, sprouting, migration, and remodeling (2, 3). Identifying and understanding these molecular mechanisms is key to the development of targeted therapeutics for children with vascular anomalies.
The care of patients with vascular anomalies has been plagued by misdiagnosis, which can delay appropriate treatment and misinform patients and their families. Comprehensive diagnostic evaluation requires careful attention to patient and family history, clinical and radiologic features of the lesion, and thoughtful classification of the lesion. Many centers utilize a multidisciplinary team, with both medical and surgical providers with experience and expertise in vascular anomalies, to provide comprehensive care for these patients. Increasingly, geneticists are becoming key members of such multidisciplinary teams. The important role of genetics in the care of patients with vascular anomalies is highlighted by updates to the classification system.

In 1996, the International Society for the Study of Vascular Anomalies (ISSVA) developed a standardized classification guideline for children and adults with vascular Anomalies (4–6). In 2018, the ISSVA classification system was updated to capture our expanding knowledge base and report known genetic causes of vascular anomalies (6, 7).

**Inherited vs. Sporadic Vascular Anomalies**

Understanding the types of genetic changes that drive the formation of vascular anomalies is key to appropriate testing and diagnosis. These pathogenic genetic variants can be categorized as either germline (familial or heritable mutations) or somatic (acquired mutation in somatic “body” tissues). The majority of vascular anomalies arise from somatic mutations, in which there is a post-zygotic mutation in a single cell that then perpetuates itself and is present in groups of cells throughout the body (called mosaicism). In this case, the mutation is not present in every cell. Genetic testing in patients with high suspicion for somatic driver mutations is presently best completed on directly affected tissue, which often requires surgical or skin biopsy from the vascular anomaly (Figure 1). Conversely, germline mutations are present in all of the individual’s cells including the egg and sperm. Genetic testing in patients with high suspicion for germline driver mutations can be completed on any tissue, skin, blood, hair, or saliva.

Familial or inherited vascular malformations or syndromes are less common but provide unique insight regarding the molecular mechanisms that control vascular morphogenesis. The majority are inherited in an autosomal dominant fashion and involve mutations that lead to a loss-of-function of the affected gene. There is generally not 100% penetrance of familial vascular disorders and some of the phenotypes may not be readily apparent until later in life, suggesting that a second hit may be needed for tumor or malformation to present. In addition, a wide spectrum may be noted with respect to type, location, and presence of the malformation. When evaluating a patient with a vascular anomaly, family history including a 3-generation pedigree, may determine whether one might consider a germline mutation. Family and personal history of cutaneous findings, macrocephaly, history of malignancies, and developmental or learning disabilities; for example, implicates PTEN Hamartoma Tumor Syndrome (PHTS) and prompts germline testing to confirm. This has immediate implications for management with surveillance for thyroid cancer starting in childhood and additional malignancies in adulthood (8).

Historically, a low frequency of identified germline mutations led clinicians to seek tissue biopsy and many key somatic mutations were identified in this manner. Identification of tissue specific somatic mutations expands our understanding of the phenotypic spectrum of these disorders and our knowledge of their pathophysiology. As we continue to define the specific molecular changes and learn the breadth of clinical findings, we improve the inaccuracies in clinical classification and may be able to pursue genotype-phenotype correlation (9). Molecular diagnosis provides objective classification and serves to direct personalized therapies based on molecular phenotype (10). The identification of genes involved in these developmental defects of
the vasculature is critical to our understanding of their etiology, as well as providing targeted therapeutic interventions (3, 11, 12).

Role of Genetics
Genetic testing is now a key component of comprehensive care for patients with vascular anomalies. Accurate identification of a genetic variant can aid in definitive diagnosis, allow for necessary preventive screening, and directly impact therapeutic decision making. Elucidation of the genetic changes underlying vascular anomalies has also played an invaluable role in understanding normal vascular developmental programming and biology. The landscape has changed dramatically with the recognition of an expanding repertoire of germline and somatic mutations recognized to drive the development of vascular malformations and tumors. The majority of identified mutations in vascular anomalies occur within two key intracellular signaling pathways—the RAS/MAPK and PI3K/AKT/mTOR pathways (Figure 2). The PI3K/AKT/mTOR pathway is crucial for many cellular processes, including cell cycle regulation, proliferation, and migration, and is often termed the “anti-apoptosis pathway.” Several activating mutations within this pathway are associated with vascular anomalies and complex vascular syndromes. The RAS/MAPK pathway is also crucial for cell cycle regulation, proliferation, and migration, and is often referred to as the “proliferation pathway.” Several vascular anomalies are associated with mutations in this pathway and frequently termed “rasopathies.” There is also crosstalk between these 2 main pathways. Finally, TGF-β signaling, which is ubiquitous and key for regulation of numerous biological processes, has been implicated in the pathophysiology of hereditary hemorrhagic telangiectasia. As we learn more about the mutations involved in the development of various vascular malformations and tumors, it becomes clear that activating mutations in these key intracellular signaling pathways are frequently the cause of endothelial cell dysfunction and malformation development (13). The types of malformations or tumors that result are dependent on the specific mutation, tissue mosaicism, impact on protein function, and interaction with other genes. For example, mammalian target of rapamycin (mTOR) signaling is initiated in the phosphoinositide 3-kinase (PI3K)/AKT pathway involved in cell cycle regulation and often implicated in patients with vascular anomalies and overgrowth who have somatic mutations in PIK3CA. This finding has led to targeting of the overactive PI3K/AKT/mTOR pathway with sirolimus (an mTOR inhibitor), which has demonstrated clinical benefit in several vascular malformations and tumors (10, 11). As we continue to identify the specific driver mutations that cause mosaic or sporadic vascular anomalies, our understanding of the pathophysiology will continue to grow and offer insight into therapeutic strategies. It is very likely that additional gene mutations will be identified and become future targets for therapy. We offer here the current information regarding protein function for the known genetic

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**FIGURE 2 | Genetic pathways implicated in vascular anomalies.** The majority of identified mutations in vascular anomalies occur within two key intracellular signaling pathways—the RAS/MAPK and PI3K/AKT/mTOR pathways. The PI3K/AKT/mTOR pathway is crucial for many cellular processes, including cell cycle regulation, proliferation, and migration, and is often termed the “anti-apoptosis pathway.” Several activating mutations within this pathway are associated with vascular anomalies and complex vascular syndromes. The RAS/MAPK pathway is also crucial for cell cycle regulation, proliferation and migration, and is often referred to as the “proliferation pathway.” Several vascular anomalies are associated with mutations in this pathway and frequently termed “rasopathies.” There is also crosstalk between these 2 main pathways. The TGF-β Signaling Pathway is also key for regulation of numerous biological processes, has been implicated in the pathophysiology of hereditary hemorrhagic telangiectasia. Each pathway and their overlap are demonstrated pictorially here. The known associations with vascular malformations and syndromes are highlighted in red.
mutations in vascular malformations and tumors. However, this information is growing rapidly and we are still at the very early stages of understanding this biology and identifying future targets for therapy that will continue to shape the management of vascular anomalies.

**VASCULAR MALFORMATIONS**

Using the 2018 International Society for the Study of Vascular Anomalies classification of vascular malformation by simple subtype (6, 7), we provide an outline of the inherited/germline and somatic mutations identified in various malformations and recommended clinical evaluation for each malformation or syndrome. Examples of some of the key vascular malformations, syndromes, and tumors are found in Figure 3. We also provide a pathway for initial evaluation and recommended "next steps" for each type of vascular malformation or tumor (Figure 4). When guided by a careful history and physical exam, we believe this tool can be beneficial to clinicians as they navigate initial evaluation and management, recognizing that ultimately a multidisciplinary approach and opinion will be invaluable.

**Capillary Malformations**

Capillary malformations (CMs), commonly known as port-wine stains, are common in the general population with an incidence of 0.3% (14). Pathologically they are defined by dilated venule-type channels in the superficial dermis with characteristically slow-flow on imaging. Capillary malformations are often associated with underlying soft tissue or bony hypertrophy, though whether this is causative or associated with the increased blood flow is unclear. They are typically present at the time of birth and may change in color over time. With age, vascular stasis in the abnormally formed capillaries results in vascular ectasia and soft tissue thickening. Although most isolated capillary malformations and associated syndromes are sporadic (Table 1), familial inheritance has been described (Table 2). Sturge-Weber syndrome (SWS) represents the most well-known cause of somatic capillary malformation. The capillary malformations seen in SWS and the more common isolated, uncomplicated capillary malformations are frequently found to have mutations in GNAQ or GNA11, which can lead to constitutive activation of the RAS/MAPK pathway (Figure 1) (24, 25). Inherited capillary malformation syndromes are most commonly due to mutations in the RAS/MAPK signaling pathway and are inherited in an autosomal dominant fashion with incomplete penetrance and variable expression.

**Venous Malformations**

Venous malformations (VMs) are slow-flow lesions that can either be mass-like within tissues and unrelated to named veins, or they can represent anomalous development of anatomic veins. They are often considered the most common subtype of vascular malformation, with an incidence of between 1 in...
FIGURE 4 | Diagnostic algorithm for vascular anomalies. This figure gives a suggested diagnostic algorithm for the initial evaluation and next steps when evaluating a patient with a vascular malformation or tumor. It is not all inclusive, but gives suggestions based on initial characteristics of the lesion and highlights key/critical steps that should not be missed. GLA, generalized lymphatic anomaly; KLA, Kaposiform lymphatic anomaly; CCLA, central conducting lymphatic anomaly; GSD, Gorham Stout Disease; LIC, localized intravascular coagulation; KTS, Klippel Trenaunay Syndrome; PROS, PIK3CA related overgrowth spectrum; BRBN, Blue Rubber Bleb Nevus; IH, infantile hemangioma; CH, congenital hemangioma.

5,000 to 1 in 10,000 births. They can range in size from very small to extensive and are purple to bluish in color due to stagnant blood flow within the lesion, that contributes to pain, swelling, and thrombotic complications (4, 5, 26–28). The majority of lesions are noted at birth, but deeper lesions are sometimes asymptomatic until the time of puberty when they tend to increase in size and become symptomatic. More than 90% of venous malformations are sporadic and isolated in nature (Table 3), but may rarely present with a familial pattern (Table 4) (15). TIE2/TEK gene mutations are thought to be causative in most isolated venous malformations. However, more recently, somatic activating mutations in PIK3CA have also been identified in venous malformations (41). TIE2/TEK is an endothelial cell specific tyrosine kinase receptor that functions via the PI3K/AKT/mTOR signaling pathway (Figure 1) and plays a significant role in regulating angiogenesis, proliferation, cell migration, and vessels stability (3, 15). Blue rubber bleb nevus syndrome (BRBNS) is a rare, often severe disorder known to be caused by TIE2/TEK somatic mutations (30).

Careful documentation of birthmarks on family pedigree can help identity hereditary cases. Glomuvenous malformations and mucocutaneous venous malformation (VMMC) have demonstrated autosomal dominant inheritance but are thought to require a second hit for lesion formation (26).

Lymphatic Malformations

Lymphatic malformations (LM) encompass a range of developmental or functional defects affecting lymphatic vessels. These can be focal or diffuse disorders with marked heterogeneity (Table 5) (46). Lymphatic anomalies can be associated with significant comorbidities such as pain, infection, visceral, and bone involvement and disfigurement. Most lymphatic malformations are due to sporadic mutations in genes that regulate lymphangiogenesis. However, certain genetic syndromes are associated with developmental abnormalities of the lymphatics, including Down syndrome, Turner syndrome, Noonan syndrome and Cardiofaciocutaneous syndrome (47, 48). These are important to be aware of in the general evaluation
of children with these disorders. Lymphedema is clinically distinct from lymphatic malformations in that it is due to functional defects in initial or collecting lymphatics. Some forms of lymphedema and related syndromes appear to have familial inheritance (Table 6) (3). Sporadic lymphatic malformations and related syndromes are believed to involve mutations in VEGFC/VEGFR3 and PI3K/AKT/mTOR pathways, but a clear pathogenetic mechanisms has not been defined (3, 15).

### Arteriovenous Malformations
Arteriovenous malformations (AVMs) occur due to a direct connection between an artery and vein, bypassing the normal capillary bed, leading to arterIALIZATION of the veins and a high-flow lesion with propensity to grow and bleed (54). Sporadic AVMs (Table 7) are commonly associated with mutations in the RAS/MAPK pathway (Figure 1). RASA1 mutations are associated with CM-AVM and may be causative in some
TABLE 2 | Inherited/germline conditions involving capillary malformations (CM).

| Associated malformation or syndrome                      | Inheritance | Clinical phenotype                                                                 | Candidate gene | Protein function or suspected role (if known) | Clinical evaluation                                                                 | References |
|----------------------------------------------------------|-------------|------------------------------------------------------------------------------------|----------------|---------------------------------------------|-------------------------------------------------------------------------------------|------------|
| Capillary malformation–arteriovenous malformation (CM–AVM)* | AD          | • Circumscribed, small diffusely distributed CMs (6% single lesion); halo surrounding  | RASA1, EPHB4   | • Loss of function; encodes Ras GTPase activating protein p120RasGAP                  | • Brain/spine imaging • Genetic testing and counseling • Screening echo • X-rays for LLD • ENT-epistaxis. • US fast-flow lesion | (3, 9, 11, 15, 20, 21) |
| Generalized essential telangiectasia                      | AD          | • Generalized telangiectasias that progress peripheral to central                    | None identified | • Unknown                                    | • Rule out gastric bleeding                                                        | (3, 22)   |
| Angioma serpiginosum                                      | X-linked    | • Classical pinpoint, dilated, think walled capillaries, located sub-epidermally along the lines of Blaschko | Unknown reports of Xp11.23 deletion containing PORCN (but contradicted by some) | • Unknown                                    | • EGD to rule out esophageal papillomas • Skin and nail exam                      | (3, 23)   |

*Note that CM-AVM is classified under both capillary and arteriovenous malformation. LLD, leg length discrepancy.

sporadic AVMs (3, 15). Other known molecular changes found in sporadic AVMs are found in MAP2K/MEK and several groups are starting to use MEK inhibitors for off-label management (29, 57). The most common inherited AVM syndrome (Table 8) is hereditary hemorrhagic telangiectasia (HHT), also known as Osler-Weber-Rendu syndrome. HHT is a disorder of vascular dysplasia characterized by multiple AVMs. Small telangiectasias may manifest as superficial skin and mucosal membrane lesions with potential to rupture presenting as epistaxis or gastrointestinal bleeding. Larger AVMs can be found within the lungs, central nervous system, and liver. HHT is inherited in an autosomal dominant manner with clinical heterogeneity and an age-dependent natural history. Pathologic mutations to genes involved in TGF-beta signaling pathway (Figure 1), important for angiogenesis, have been described including ENG, ALK1/ACVRL1, GDF2, SMAD4/MADH4. An appropriate pediatric HHT screening evaluation must include family history, physical examination, and a low threshold to proceed with non-invasive screening and imaging (58, 59). Heterozygous ENG and ALK mutations, such as found in HHT, have also been found in sporadic AVMs.

Complex Syndromes Associated With Vascular Anomalies
There are several complex syndromes that include overgrowth of soft tissues in combination with vascular malformations. The majority of these syndromes are caused by somatic mutations, primarily isolated to the PI3K/akt/mTOR signaling pathway (Figure 1) (11, 15). These syndromes and the candidate genes involved in their development are summarized in Table 9. Recognition of these syndromes is important due to the association of other complications, such as malignancy in CLOVES and Maffucci syndromes, and thrombosis in Klippel-Trenaunay and Proteus syndromes.

Vascular Tumors
Vascular tumors include both benign, locally aggressive, and malignant tumors comprised of one or more vascular components (7, 62). The molecular etiology of many of these
### TABLE 3 | Somatic disorders involving venous malformations.

| Associated malformation or syndrome | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical evaluation | References |
|------------------------------------|--------------------|----------------|-----------------------------------------------|--------------------|------------|
| Sporadic VMs | • Solitary, localized, and non-familial  
• Soft, mass or sponge-like lesion with blue or purple coloration | TIE2/TEK  
PIK3CA | • Endothelial cell-specific tyrosine kinase receptor. Important for angiogenesis (angiopoietin receptor). Gain of function mutations, probably affects endothelial cell behavior, may involve paracrine signaling between endothelial and smooth muscle cells, down-regulation of PDGF-beta production, only mucocutaneous veins are affected  
• Catalytic alpha subunit of PI3K. Somatic activating mutation that increases PI3K/AKT/mTOR signaling | • US or MRI of lesion depending on location and symptoms  
• Screening for localized intravascular coagulopathy (PT, PTT, fibrinogen, d-dimer)  
• Consider brain MRI | (3, 10–12, 15) |
| Multifocal VM | • Rare, multifocal variant  
• Tend to be mosaic for first mutation with 2nd hit | TIE2/TEK | • Endothelial cell-specific tyrosine kinase receptor. Important for angiogenesis (angiopoietin receptor) | • Consider whole body MRI | (11, 12) |
| Verrucous VM | • Raised reddish-purple hyperkeratotic lesion (extremities)  
• Enlarge over time, more hyperkeratotic with increased bleeding | MAP3K3 | • Somatic activating mutation that may increase RAS/MAPK signaling | • Skin exam  
• Consider evaluation for underlying bleeding diathesis | (3, 10, 12, 29) |
| Fibroadipose vascular anomaly (FAVA) | • Within a muscle; increased fibroadipose tissue and smaller, non-spongiform vessels  
• Often single limb, appears slightly enlarged | PIK3CA | • Catalytic alpha subunit of PI3K. Somatic activating mutation that increases PI3K/AKT/mTOR signaling | MRI or US of the lesion | (3, 10, 12) |
| Blue rubber bleb nevus syndrome | • Sporadic, some AD inheritance  
• Multiple cutaneous and internal VM  
• Intestinal and hepatic  
• Tens to hundreds of blue or purple, compressible, and hyperkeratotic | TEK/TIE2 | • Endothelial cell-specific tyrosine kinase receptor. Important for angiogenesis (angiopoietin receptor) | • MRI or ultrasound, consider full body MRI consider biopsy  
• Screening for localized intravascular coagulopathy (D-dimer, fibrinogen, PT, PTT)  
• Urinalysis for hematuria  
• Screening for iron deficiency anemia  
• Endoscopy/colonoscopy | (3, 10–12, 15, 30, 31) |
| Cerebral cavernous malformation (CCM) | • Immature vessels reflecting abnormal angiogenesis presents with seizures, headaches, hemorrhage, and neurologic defects | KRIT1 (CCM1)  
CCM2  
CCM3  
Sporadic or AD | • Adaptor protein Integrin β1 pathway involved in arterial specification, cell adhesion, endothelial cell junctions, and migration  
• Adaptor protein, scaffold protein, overlaps with KRIT1  
• Apoptosis, VEGF signaling. Overlaps with KRIT1  
• The CCM proteins interact together and dysfunction of the CCM signaling complex leads to altered vascular integrity and endothelial cell organization. They suppress RhoA GTPase signaling | • Brain MRI with gradient echo or susceptibility weighted imaging | (3, 9–12, 15) |
| Associated malformation or syndrome | Clinical phenotype | Candidate gene/inheritance | Protein function or suspected role (if known) | Clinical evaluation | References |
|-------------------------------------|--------------------|---------------------------|-----------------------------------------------|--------------------|------------|
| PTEN-associated venous anomalies    | • Hamartoma of soft tissue, intramuscular vascular lesions, with fast-flow lesions in 86% • Increased risk of cancer | PTEN AD | • Tumor suppressor | • Head circumference • Skin exam • Brain MRI with contrast • Cancer screening (thyroid, breast, uterine, colon) | (3, 32–34) |
| Mucocutaneous venous malformations (VMCM) | • Small, multifocal bluish mucocutaneous lesions | TIE2/TEK AD | • Endothelial cell-specific tyrosine kinase receptor. Important for angiogenesis (angiopoietin receptor) | • Physical exam, consider imaging • Consider cardiac work up | (3, 15, 35) |
| Cerebral cavernous malformation, familial | • Cerebral lesions dilated channels with endothelial cell layers that have defective tight junctions in the brain, retina and spinal cord • Majority present in between the second and fifth decades with seizures, focal neurologic findings, headaches, and cerebral hemorrhage • Cutaneous vascular lesions in 9% and retinal vascular lesions in 5% | KRT1 CCM2 PDCD10 AD | • Loss-of-function mutations affecting subendothelial matrix, vascular structure, and adhesion | • Brain MRI with gradient echo or susceptibility weighted imaging | (3, 36) |
| Glomuvenous malformations or Glomangiomas | • Superficial, multiple raised or plaque-like lesions • Cobblestone appearance, painful on palpation | GLMN AD, 100% penetrance with variable expressivity | • Phosphorylated protein that is a member of the Skp1-Cullin-F-box-like complex • Essential for normal development of the vasculature • Loss of function mutation, likely requires a somatic second hit | • Detailed skin exam • Genetic testing and counseling • Biopsy vs. removal if concern about diagnosis • No routine imaging unless clinical concern | (3, 15, 26, 37) |
| Hyperkeratotic cutaneous capillary-venous malformation (HCCVM) | • Crimson-colored irregularly shaped lesions that extend into the dermis and hypodermis • Composed of dilated capillaries and blood-filled venous-like channels • Associated with cerebral capillary malformations, which may present with headaches, seizures, and intracranial hemorrhage | KRT1 (also known as CCM1) CCM2 CCM3 Possible AD | • RAS antagonist, may be involved in cellular adhesion and vascular integrity. The CCM proteins interact together and dysfunction of the CCM signaling complex leads to altered vascular integrity and endothelial cell organization | • Detailed skin exam • Brain imaging for cerebral capillary malformations | (38, 39) |
| Varicose veins | • Twice as common in females as males | FOXC2 AD, with reduced penetrance | • Unknown | • Physical exam with referral to vascular surgeon if symptomatic concerns | (3, 40) |
TABLE 5 | Somatic disorders involving lymphatic malformations.

| Associated malformation or syndrome | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical work-up | References |
|-------------------------------------|--------------------|----------------|-----------------------------------------------|------------------|------------|
| Sporadic LMs                        | • Localized or extensive malformed lymphatic vessels in the skin or deep soft tissues  
  • Predilection for head/neck areas | PIK3CA | • Catalytic alpha subunit of PI3K.  
  Somatic activating mutation that increases PI3K/AKT/mTOR signaling | • Imaging of involved site with MRI  
  • Evaluation for other sites of disease if clinically indicated | (3, 10–12) |
| Generalized lymphatic anomaly (GLA) | • Previously known as lymphangiomatosis  
  • Diffuse or multifocal LMs throughout the skin, soft tissues, abdominal, and thoracic viscera  
  • Pericardial and pleural effusions may be noted  
  • May have significant bone involvement | PIK3CA | • Catalytic alpha subunit of PI3K.  
  Somatic activating mutation that increases PI3K/AKT/mTOR signaling | • Full body imaging with CT and/or MRI  
  • Screening for coagulopathy (PT, PTT, fibrinogen, d-dimer)  
  • Biopsy not necessary  
  • Echo | (10, 11, 42) |
| Gorham stout syndrome, aka “disappearing bone disease” | • Progressive osteolysis with replacement of bone by soft tissue and vascular channels, primarily lymphatic in origin  
  • Defining characteristic is disappearance of bone rather than a mass lesion in bone | Unknown | • Likely involvement of PI3K/AKT/mTOR signaling pathway | | |
| Kaposiform lymphangiomatosis (KLA)  | • Systemic and frequently aggressive lymphatic anomaly  
  • More extensive thoracic involvement than GLA  
  • Frequently associated with coagulopathy, effusions, and intralesional hemorrhage  
  • Systemic involvement is common | NRAS, possible | • Intracellular RAS signaling. A proto-oncogene that encodes a small GTPase that regulates cell proliferation in the RAS/MAPK and PI3K/AKT/mTOR signaling pathways | • Full body imaging with CT and/or MRI  
  • Consider echocardiography  
  • Screening for coagulopathy (PT, PTT, fibrinogen, d-dimer)  
  • Consider angiopeptin and VEGFR3 screening  
  • Biopsy not necessary | (3, 10, 11, 44, 45) |
| Congenital pulmonary LM              | • LMs in the lungs, heart, pancreas, kidneys, and mesentery  
  • Respiratory distress at birth due to pulmonary hypoplasia secondary to chyloous pleural effusions  
  • May overlap with Noonan, Turner, and Down syndromes | CCBE1 | • Extracellular guidance molecule for migrating lymphatic endothelial cells, enhances lymphangiogenic activity of VEGF-C | • Full body imaging with CT and/or MRI  
  • Consider echocardiography  
  • Developmental and neurologic screening | (3, 15) |
| Associated malformation or syndrome | Inheritance | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical evaluation | References |
|-------------------------------------|-------------|--------------------|----------------|-----------------------------------------------|--------------------|------------|
| Primary lymphedema (Nonne-Milroy disease) | AD          | • Lower-limb lymphedema, present as pedal edema at birth or soon after  
• Usually bilateral  
• Hydrocele (males)  
• Prominent veins  
• Uplifting toenails  
• Papillomatosis  
• Urethral anomalies in males  
• Upslanting toenails  
• Papillomatosis  
• Urethral anomalies in males | FLT4  
VEGFR3 | • Endothelial cell tyrosine kinase receptor, important for lymphangiogenesis (VEGF-C receptor)  
• Transcription factor, regulates PDGFβ | • MR lymphangiography or NM lymphoscintigraphy (lack of uptake radioactive colloid in ilioinguinal lymph nodes)  
• PT/OT evaluation  
• Compression fitting  
• Good skin hygiene  
• Genetic testing and counseling | (3, 10, 11, 15) |
| Late-onset lymphedema (Meige Disease) | Probable AD | • Most common subtype of primary lymphedema (~80% all cases)  
• Present at birth but onset of symptoms at puberty  
• Edema begins in feet and ankles and progresses to legs and knees | FOXC2  
GJC2 | • Transcription factor, regulates PDGFβ  
• Gap junction protein | • Evaluation as per primary lymphedema plus  
• Evaluation to rule out secondary lymphedema (malignancy, thrombosis, etc.) | (3) |
| Microcephaly-lymphedema-chorioretinal dysplasia | AD, or sporadic | • Patients have persistence of fetal lymphedema  
• Hypotonia,  
• Microcephaly  
• Intellectual disability  
• Chorioretinopathy | KIF11 | • Spindle motor protein, may affect microtubule function | • Evaluation as per primary lymphedema plus  
• Ophthalmology exam  
• Echocardiogram  
• Consider brain MRI (lissencephaly reported) | (3, 15) |
| Hypotrichosis-lymphedema-telangiectasia syndrome | AR          | • Absent eyebrows and eyelashes  
• Eyelid edema from birth  
• Lymphedema in the first of second decade of life  
• Telangiectasias (palms, soles, scalp, scrotum, legs) | SOX18 | • Spindle motor protein, may affect microtubule function | • Evaluation as per primary lymphedema plus  
• Serial echocardiogram (dilation of the ascending aorta reported)  
• Skin and nail exam | (3, 15) |
| Lymphedema with distichiasis | AD          | • Late onset lymphedema with distichiasis (fine hairs from Meibomian glands on inner eyelid)  
• Variable cardiac/renal/palate defects, varicose veins  
• Neck webbing | FOXC2 | • Transcription factor, regulates PDGFβ | • Evaluation as per primary lymphedema plus  
• Ophthalmology exam (corneal irritation, recurrent conjunctivitis, and photophobia, ptosis)  
• Echocardiogram to rule out congenital heart disease  
• Spine MRI to rule out extradural arachnoid cysts | (3, 49) |
### TABLE 6 | Continued

| Associated malformation or syndrome | Inheritance | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical evaluation | References |
|-------------------------------------|-------------|--------------------|---------------|-----------------------------------------------|--------------------|-----------|
| Lymphedema with choanal atresia     | AR          | • Exceedingly rare, 1 family with lymphedema and choanal atresia | PTPN14 | • Loss of function mutation in protein tyrosine phosphatase | • Evaluation as per primary lymphedema | (3)       |
| Cholestasis-lymphedema (Aagenaes Syndrome) | AR          | • Jaundice at birth and recurrent through life | Unknown gene on 15q | • Unknown | • Evaluation as per primary lymphedema plus • Hepatic/GI evaluation including CMP, serum bile acids (elevated), and lipid panel (hyperlipidemia) | (3)       |
| Lymphedema-intestinal LM-mental retardation (Hennekam Syndrome) | AR          | • Severe progressive lymphedema including genitalia and face | CCBE1 FAT4 ADAMTS3 | • Function in migration of lymphatic endothelial cells | • Evaluation as per primary lymphedema plus • Neurologic evaluation • Hearing screen • Abdominal US • Nutritional assessment • Thyroid studies • Screening for anemia • Spine X-rays | (3, 50–52) |
| Lymphedema-myelodysplasia (Emberger syndrome) | AD          | • Lymphedema of limbs and genitalia in early childhood • Myelodysplasia with development of AML • Craniofacial anomalies • Congenital deafness | GATA2 | • Transcription factor involved in gene regulation during vascular development and hematopoietic differentiation | • Evaluation per primary lymphedema plus • Hematologic evaluation including bone marrow biopsy • Neurologic and hearing assessments | (3, 53) |

### TABLE 7 | Somatic disorders involving arteriovenous malformations.

| Associated malformation or syndrome | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical work-up | References |
|-------------------------------------|--------------------|---------------|-----------------------------------------------|-----------------|-----------|
| Sporadic AVM                        | • Abnormal connections between arteries and veins without a normal capillary bed | MAP2K1/MEK1 KRAS | • Stimulates enzymatic activity of MAP kinases | • Imaging with MRI and/or angiography | (3, 10, 11, 54– 56) |
|                                     | • Results in arterialization of venous system, pain, tissue destruction, and bleeding complications | NRAS BRAF | • Intracellular RAS/MAPK signaling | • Thorough exam/history to exclude HHT or CM-AVM |           |
TABLE 8 | Inherited/germline conditions involving arteriovenous malformations.

| Associated malformation or syndrome | Inheritance | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical work-up | References |
|------------------------------------|-------------|--------------------|----------------|-----------------------------------------------|-----------------|-----------|
| Hereditary Hemorrhagic Telangiectasia (HHT) aka Osler-Weber-Rendu Syndrome | AD          | Telangiectasias (lip, tongue, buccal mucosa, face, chest, and fingers) | ENG, ALK1/ACVRL1, GDF2 SMAD4/MADH4 | TGF-β signaling pathway | CBC/iron studies, Sitting and supine pulse oximetry every 1–2 years during childhood, Contrast echo every 5 years by age 10, Periodic endoscopy/colonoscopy to rule out GI polyps and malignant change of juvenile polyps, ENT evaluation, Brain MRI/MRA in infancy and after puberty, Liver US, Genetic testing and counseling, Antibiotic prophylaxis for dental and invasive procedure, Filter on IV lines to prevent bubbles if pulmonary AVM is present | (3, 58, 59) |
| CM-AVM1*                           | AD or sporadic | Small, multifocal CMs often accompanied by a pale halo | RASA1 | Loss of function; encodes Ras GTPase activating protein p120RasGAP | Brain/Spine imaging, Genetic testing and counseling, Screening echocardiography, X-rays for leg length discrepancy, ENT evaluation if epistaxis, US shows fast-flow lesion | (3, 9, 11, 15, 20, 21) |
| CM-AVM2*                           | Unknown, likely AD | Small telangiectasias around the lips and on upper thorax | EPHB4 | Loss of function mutation. Transmembrane receptor preferentially expressed in endothelial cells, acts via RAS/MAPK pathway | Brain/Spine imaging, Genetic testing and counseling, Screening echocardiography, X-rays for leg length discrepancy, ENT evaluation if epistaxis | (12, 20, 21) |

*Note that CM-AVM is classified under both capillary and arteriovenous malformation.

tumors is unknown, but some candidate genes have recently been identified (Table 10). Infantile hemangiomas are the most common vascular tumor, and are generally benign, but growth can be disruptive of function or cosmesis, and occasionally life-threatening. The role of the pediatrician is key to determining which lesions require referral for additional management. Vascular tumors other than hemangiomas often require multi-modal therapy, including chemotherapy,
| Syndrome                        | Clinical phenotype                                      | Candidate gene | Protein function or suspected role (if known) | Clinical work-up                                                                 | References |
|--------------------------------|-------------------------------------------------------|----------------|-----------------------------------------------|---------------------------------------------------------------------------------|------------|
| CLOVES syndrome                | • Characterized by congenital lipomatous overgrowth   | PIK3CA         | • Catalytic alpha subunit of PI3K. Somatic activating mutation that increases PI3K/AKT/mTOR signaling | • Brain MRI • X-rays for leg length discrepancy • Scoliosis screen • Echocardiogram • EEG • Renal US to rule out anomalies and q3-6 months to screen for Wilms tumor • Screening for paraspinal high-flow lesions with spinal cord ischemia • Thrombophilia evaluation • Developmental • Feeding assessment | (3, 15)   |
| Klippel-Trenaunay syndrome     | • Characterized by slow-flow capillary-lymphatic-venous malformations and soft tissue overgrowth of an extremity and/or trunk • Often involves the pelvis as well. Can be unilateral or bilateral | PIK3CA, AGGF1 | • Catalytic alpha subunit of PI3K. Somatic activating mutation that increases PI3K/AKT/mTOR signaling • Angiogenic factor with G patch and FHA domains, increases angiogenesis in vitro | • MRI imaging of affected area • Thrombophilia evaluation and labs for localized intravascular coagulation | (3, 15)   |
| Megalencephaly-capillary malformation syndrome (MCAP) | • Congenital megalencephaly or hemimegalencephaly • Reticulate capillary stains • CM of the lip/philtrum • Asymmetry, focal, or generalized overgrowth • Hypotonia • Seizures • Mild to severe intellectual disability • Syndactyly | PIK3CA, from brain tissue | • Catalytic alpha subunit of PI3K. Somatic activating mutation that increases PI3K/AKT/mTOR signaling | • Brain MRI q 6 months for the first 2 years then yearly till 8 years to rule out neurological complications • X-rays for leg length discrepancy • Scoliosis screen • EEG • Echocardiogram • Renal US to rule out anomalies and q 3 months to screen for Wilms tumor • Developmental and feeding assessment • Sleep evaluation • Thrombophilia evaluation | (3, 10, 15) |
| Maffucci syndrome              | • Multiple spindle cell hemangiomomas associated with multiple enchondromas • Vascular lesions often do not appear until puberty • Patients have increased risk for malignancy | IDH1 and IDH2 | • Mutant enzymes catalyze the reduction of alfa-ketoglutarate to D-2-hydroxy glutarate, cause downstream genomic hypermethylation | • Screening of lesions due to malignancy potential • Risk of many types of malignancy reported (chondrosarcomas, gliomas, ovarian tumors, and other sarcomas) • Limb length x-rays and imaging of the extremities for other malformations | (3, 10, 60) |
| Proteus syndrome               | • Bony and soft tissue overgrowth that develops and progresses rapidly in the toddler period and tends to plateau after adolescence • Increased risk of malignancy • Pulmonary complications • Increased risk thrombus | AKT1           | • Intracellular PI3K/AKT/mTOR signaling/apoptosis | • Scoliosis screen • Skin exam • Thrombophilia evaluation and monitoring for DVT and PE • Developmental assessment • Monitoring for bullous pulmonary disease | (3, 10, 11) |

(Continued)
and consultation with a pediatric hematologist-oncologist is indicated.

**DISCUSSION**

It is critical for teams treating patients with vascular anomalies to have a basic understanding of the genetic changes underlying vascular anomalies. Genetic results have implications for patient management, screening, and ultimately treatment.

As we begin to understand more about the genetic underpinnings of vascular anomalies, we identify potential therapeutic targets that will ultimately change treatment paradigms. Prior to the discovery of the underlying molecular pathways involved in the pathogenesis of vascular malformations and tumors, medical management was based primarily on symptom management with antiplatelet and anticoagulant agents to control localized intravascular coagulation and platelet activation, and pain medications to alleviate symptoms (1, 5, 8, 9). Over the years, multiple medications, primarily based on adaptation of oncologic agents, have been utilized in patients with complex and life-threatening vascular anomalies. This includes drugs such as bevacizumab, interferon-alpha, and cyclophosphamide, that have come with their own significant risks and side effects and generally can be considered sledgehammer rather than target approaches (68–70). Even propranolol’s efficacy in infantile hemangiomas was identified serendipitously in children who received the drug for the cardiac complications they developed due to their massive infantile hemangiomas (71). Despite its successful use, the mechanism and relationship of propranolol to vascular growth pathways remains to be elucidated (72–74).

In the early 2000s, several groups began to report on the importance of PI3K/AKT/MAPK, and TGF-β signaling in the pathogenesis of vascular malformations (Figure 1) (75–78). Based on the efficacy of sirolimus, an mTOR inhibitor, in angiomylipomas associated with tuberous sclerosis complex (79, 80) and Kaposi sarcoma (81), sirolimus was first used in a child with Kaposiform hemangioendothelioma in 2010 (82). Shortly after, a small case series of children with complex vascular malformations and tumors was published, in which all patients demonstrated a significant response and improvement with sirolimus (68). This prompted the first prospective clinical trial that was published in 2016 and confirmed both the efficacy and safety of Sirolimus in the treatment of complex vascular anomalies (83). With the identification of the PI3K/AKT/mTOR pathway as an important driver of somatic overgrowth syndromes (84) and the identification of somatic PI3K mutations within certain vascular malformations (85, 86), PI3K inhibition has become the next major therapeutic target for vascular anomalies (87–89). Future targets will likely be aimed at the RAS/MAPK pathway, where somatic mutations are known to be important in AVMs, capillary malformations, and other aggressive vascular syndromes (10, 44, 55). A list of targeted therapies currently under investigation or in use for vascular anomalies is in Table 11. These discoveries will likely continue to grow at a rapid pace and ultimately inform our classification and management of patients in the clinic. For this reason, it is key for burgeoning vascular anomalies clinical teams to include geneticists as key team members for discovery as well as patient management.

Trained geneticists and genetic counselors are crucial members of a comprehensive multidisciplinary vascular anomalies team. Detailed discussion of family history, consideration of genetic testing, and review of results is an important part of the comprehensive care of patients with vascular malformations and tumors. Understanding which lesions are likely to be associated with somatic vs. germline changes is key to developing a thoughtful approach to genetic testing. A family history suggestive of an inherited mutation will

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### Table 11

| Syndrome | Clinical phenotype | Candidate gene | Protein function or suspected role (if known) | Clinical work-up | References |
|----------|-------------------|---------------|---------------------------------------------|-----------------|-----------|
| Parkes Weber syndrome | Similar to CM-AVM but with overgrowth of affected limb | RAS1, loss of function mutation | Intracellular signaling, RasGTPase | Brain/spine MRI | (5, 9, 11, 15) |
| Familial intraosseous vascular malformation | Extensive vascular lesions in the intraosseous spaces of the craniofacial bones associated with other midline defects | ELMO2 | Translation of extracellular signals to cellular movements | US shows fast-flow lesion | (5, 61) |

**Figure 1** pathogenesis of vascular malformations (It is critical for teams treating patients with vascular anomalies to have a basic understanding of the genetic changes underlying vascular anomalies. Genetic results have implications for patient management, screening, and ultimately treatment.)
| Tumor                          | Clinical phenotype                                                                                                           | Candidate gene | Protein function or suspected role (if known)                                                                 | Clinical work-up                                                                 | References |
|-------------------------------|------------------------------------------------------------------------------------------------------------------------------|----------------|----------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------|------------|
| Infantile hemangioma          | • Most common tumor of infancy • Comprised of capillaries and proliferating endothelial cells • Grow rapidly during first few months of life then regress in early childhood | Possible variants in VEGFR2 and TEM8 | • Altered VEGF-A and VEGFR2 signaling • Constitutive activation of VEGF-dependent VEGFR2 signaling • Sequestration of integrin 1B, inhibition of NFAT transcription and reduction of VEGFR1 | • Consider liver US if > 5 cutaneous hemangiomas • Consider evaluation for PHACES if facial segmental hemangioma (brain MRI/MRA, Echocardiogram, eye exam) | (3, 10, 12, 62) |
| Congenital hemangioma         | • Fully formed at birth • Can be rapidly involuting (RICH), partially involuting (PICH), or non-involuting (NICH)           | GNAQ, GNA11    | • Organization of the formation and remodeling of blood vessels. GNAQ mediates signals between G-protein-coupled receptors and downstream effectors. Increased RAS/MAPK signaling | • None unless clinically indicated • Large lesions can have transient Kasabach-Merritt phenomenon (screen with CBC, fibrinogen, PT, PTT) | (3, 10, 11) |
| Pyogenic granuloma            | • Post-natal lesion with mean onset around age 6 years • Benign, pedunculated, fragile, and frequently bleed                  | KRAS, NRAS, HRAS GNAQ, BRAF | • Upregulated RAS/MAPK/ERK signaling                                                                                   | • None unless clinically indicated                                                                                                           | (3, 10, 11) |
| Kaposiform hemangioendothelioma and Tufted angioma | • Vascular neoplasm generally present at birth and enlarges during infancy • Locally aggressive. Associated with Kasabach-Merritt phenomenon | GNA14          | • G-protein related signal transduction                                                                                   | • Evaluation for Kasabach Merritt phenomenon (CBC, fibrinogen, PT, PTT) • Imaging of site (MRI) • Oncology consult | (3, 10, 11) |
| Angiosarcoma                  | • High-grade malignant neoplasm of endothelial cell origins • Can arise in skin, deep soft tissues, or visceral organs • Can be radiation or lymphedema-associated | PTPRB, PLCG, KDR/VEGFR2 mutations, FLT4/VEGFR3 amplifications MYC amplification (in XRT induced) | • Vascular endothelial growth factor receptors • Proto-oncogene, increased expression of genes involved in cell proliferation | • Imaging of site and for metastatic disease, including brain • PET/CT • Oncology consult | (3, 10, 63, 64) |
| Epithelioid hemangioendothelioma | • Malignant endothelial tumor with variable clinical behavior • Multifocal lesions can be stable, grow slowly, or rapidly progress and metastasize | WWTR1-CAMTA1 translocation YAP1-TFE3 translocation | • Transcription factor signaling in the hippo pathway | • Imaging of site and for metastatic disease, including brain • Consider PET/CT • Oncology consult | (3, 10, 63, 65) |
| Familial infantile myofibromatosis | • Fibrous tumor of early childhood • Solitary lesions can regress • Multifocal or generalized lesions can be life threatening | PDGFRβ         | • Receptor tyrosine kinase and mitogen for mesenchyme-derived cells, signaling in embryonic development, including recruitment of vascular smooth muscle cells | • Imaging of site and for metastatic disease • Oncology consult | (3, 10, 66, 67) |
prompt consideration of germline testing, which can readily be performed by routine phlebotomy. Somatic mutation testing requires sampling the affected tissue, which for most patients requires a skin biopsy with appropriate tissue handling after biopsy is performed. The decision about which genes to screen for and which laboratory to use is made based on a combination of experience, cost, and insurance approval. Current technology available in the field of cancer genomics is likely to be useful as we attempt to further define the genetics of vascular anomalies (93). Members of the genetics and genetic counseling teams are key to providing input on which testing to send as well as interpretation of results. Family should be prepared to receive either definitive positive or negative results, or commonly a variant of uncertain significance with a more nuanced interpretation. Help with interpretation and contextualization of variants of uncertain significance by a geneticist is key as we begin to understand more of the molecular mechanisms and genetic changes in these malformations and tumors. The role of the geneticist and the importance of genetic testing will become increasingly important as the field of vascular anomalies continues to grow, offering new insights into the etiology of these malformations and tumors, and new avenues for improvement in patient care.

**AUTHOR CONTRIBUTIONS**

JD conceived and wrote the manuscript together with AB, DA, FB, and TN contributed critical revision of the article, patient photos for the figure, and final approval of the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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