A ‘photorelease, catch and photorelease’ strategy for bioconjugation utilizing a \( p \)-hydroxyphenacyl group†

D. Madea, T. Slanina* and P. Klán*

A bioorthogonal ‘catch and photorelease’ strategy, which combines alkyne–azide cycloaddition between \( p \)-hydroxyphenacyl azide and alkyne derivatives to form a 1,2,3-triazole adduct and subsequent photochemical release of the triazole moiety via a photo-Favorskii rearrangement, is introduced. The first step can also involve photo-release of a strained alkyne and its Cu-free click reaction with azide.

Bioconjugation is a strategy to form covalent links between two molecules of biological interest, which is commonly used in nucleic acid research, pharmaceutical chemistry or microbiology. While the first procedures used non-specific approaches, such as ionic interactions with negatively charged DNA or reactions of nucleophilic amines, carboxylates and thiols with electrophiles, modern methods rely on targeted functionalization, such as biotin–streptavidin conjugates, Staudinger ligation and click reactions.

Many bioconjugation strategies are based on bioorthogonal reactions which allow for chemical transformations of molecules without the need to protect reactive functional groups. They are used for fluorescent labeling, cell detection and many other applications. The most common types of bioorthogonal click reactions are cycloadditions, such as Cu(I)-catalyzed alkyne–azide 1,3-dipolar cycloadditions (CuAAC) or Cu-free click reactions of azides or nitrones with strained alkynes, and inverse electron-demand Diels–Alder reaction of 1,2,4,5-tetrazines with strained alkynes or alkenes. Photochemically triggered bioorthogonal reactions, which enable spatial and temporal control over the bioconjugation and can be utilized in site-specific functionalizations, are based on photolabile diazirines, tetrazoles, cyclopropanones.

The concept of ‘catch and release’, involving selective binding of a species from the reaction media, followed by its controlled liberation, has been used, for example, in combinatorial and solid-phase synthesis of complex natural products and biologically active molecules, DNA nanomachines applications, supramolecular host–guest chemistry or resin purification methods. The majority of these methods are based on non-photochemical processes. Nevertheless, involvement of a photochemical step (release) in this strategy has been demonstrated on photolabile polymers, dendrimers and liposomes.

A reversible version of click reactions would be highly demanded in the regulation and reversible on-off switching of biological processes, advanced surface modifications, or solid-phase synthesis. However, the chemical stability of the products of click reactions makes the reverse process very difficult. Several attempts for reversible bioorthogonal reactions have been reported. Aside from the retracted scientific report, only a few ‘catch and release’ strategies suitable for biological applications have been reported. Catch and release DNA decoys are rare examples of non-natural DNA probes that capture and dissociate from DNA-binding proteins upon irradiation of the 4-nitroindole linker. Such a strategy has been demonstrated earlier by Porter and coworkers who developed a reagent containing azido and biotin groups separated by a photoactivatable linker based on a benzoin photoremovable protecting group for isolation of protein adducts of lipid-derived electrophiles using streptavidin beads. In addition, Popik and coworkers have recently developed a reversible derivatization technique for peptides and proteins based on the reaction of photochemically generated 2-naphthoquinone-3-methides.

Photoremovable protecting groups (PPGs) allow for the precise spatial and temporal release of biologically active molecules. The 4-hydroxyphenacyl photoremovable (pHP) protecting group has been shown to liberate rapidly (\( k_{\text{obs}} = (7–100) \times 10^8 \) s\(^{-1}\)) and efficiently (\( \Phi = 0.1–1.0 \)) a wide range of leaving groups.

In this work, we present a ‘catch and photorelease’ strategy which combines alkyne–azide 1,3-dipolar cycloaddition between \( p \)-hydroxyphenacyl azide and alkyne derivatives to form a 1,2,3-triazole linkage in adduct 3 in the first (‘click’) step, and subsequent selective irreversible photochemical...
release of 1H-1,2,3-triazole 4 and 4-hydroxynaphthalic acid 5 moieties via a photo-Favorskii rearrangement.\textsuperscript{41,42} (Scheme 1). In addition, the first step can involve either a Cu(I)-catalyzed or Cu-free alkyne–azide process; a strained alkyne for the latter approach can be generated from the corresponding cyclopropenone via photochemical decarbonylation.\textsuperscript{22} We show that these strategies can facilitate efficient bioorthogonal sequential connection and splitting of two (bio)molecules.

Two pH photodegradable derivatives 3a and b were synthesized by a CuAAC click (‘catch’) approach from pH azide (1a and b) and acetylene (2a and b) derivatives using CuSO\textsubscript{4} and sodium ascorbate in water in 52\% and 38\% isolated chemical yields, respectively (Scheme 2 and ESI†).\textsuperscript{41} The O-methylglycine group in the 3-position of 1b represents a stable amide linkage between the photocleavable pH group and a peptide chain.

The second step of this strategy (Scheme 1) involves photochemical release of a triazole moiety from the pH cage. Irradiation of 3a in CD\textsubscript{3}CN/D\textsubscript{2}O (1:1, v/v) at \(\lambda = 313\) nm led to the formation of free triazole 4a as the major photoproduct in quantitative yield and 4-hydroxybenzaldehyde (6a, 20\%) as side-products, identified by comparison of their NMR spectra with those reported before\textsuperscript{44,45} and using authentic compounds (Scheme 3; Fig. S1, ESI†). Acid 5a is a characteristic product of the photo-Favorskii rearrangement;\textsuperscript{46,47} but benzyl alcohol (such as 6a) also often accompanies the photolysis of pH derivatives.\textsuperscript{41} The major side-photoproducts 5a and 6a are water-soluble and have negligible molar absorption coefficients at 313 nm; thus they do not act as internal optical filters.

The necessary role of water in the pH photodegradation mechanism\textsuperscript{16,41,48} predetermines this transformation for biothermosensitive aqueous solutions. Acetonitrile in CD\textsubscript{3}CN/D\textsubscript{2}O (1:1, v/v) mixtures was used to increase the solubility of the systems, and it resulted in the formation of considerable amounts of the second byproduct 6, which would not be formed in pure water.\textsuperscript{41}

Compound 3b represents a moiety that can interlink the photoactive pH group with another molecule via a peptide chain. Upon irradiation, this peptide tether, which could bear another molecule or a functional group of interest, remains on the photoproduct(s) formed from the pH chromophore. The m-substitution of the pH group by an amido group was chosen because, unlike substituents in the p- and o-positions,\textsuperscript{47} m-substituents do not significantly affect the photophysical properties of pH.\textsuperscript{49} Exhaustive irradiation of 3b in CD\textsubscript{3}CN/D\textsubscript{2}O (1:1, v/v) at \(\lambda = 313\) nm led to the formation of free triazole 4b in quantitative yield and compounds 5b (\(\approx 23\%\)) and 6b (\(\approx 20\%\)) as the major photo-Favorskii rearrangement products (Fig. S2, ESI†).

In addition, 4-hydroxynaphthaldehyde derivative 7 was identified as another side product (Fig. S4, ESI†). Because this compound appeared only after prolonged irradiation, we assume that it is a secondary product formed by 6b oxidation in the presence of oxygen in air.

It has been shown that poor leaving groups such as alcohols or amines possessing pK\textsubscript{a} values above \(\approx 11\) cannot be efficiently released from the pH group.\textsuperscript{35,38} For example, phenolate (pK\textsubscript{a} \(\approx 10\)) or p-cyanophenolate (pK\textsubscript{a} \(\approx 7.2\)) is released with a quantum yield of 0.04 and 0.11, respectively.\textsuperscript{38} Therefore, because the pK\textsubscript{a} value of 1H-1,2,3-triazole is 9.4,\textsuperscript{50} a rather low release quantum yield from its pH derivative 3a, \(\Phi_{\text{dis}} = 9 \times 10^{-4}\) (Table 1), was anticipated.

To increase the quantum yield of photorelease, two modified click adducts 3c and 3d were synthesized using a SPAAC (strain-promoted azide–alkyne cycloaddition) and a CuAAC procedure, respectively (Scheme 4 and ESI†). Compound 3c possesses two electron withdrawing fluorine atoms in the near proximity of the triazole core, which lower its pK\textsubscript{a} value and make it a better leaving group. Indeed, the \(\Phi_{\text{dis}}\) value of 3c was enhanced by a factor of \(\approx 30\) compared to that of 3a (Table 1). Because the triazoles 3a, 3b and 3d were synthesized by CuAAC in the presence of Cu\textsuperscript{I} ions, which are toxic due to the generation of reactive oxygen species,\textsuperscript{51} the non-catalyzed click (SPAAC)

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**Scheme 1** A “catch and photorelease” strategy.

**Scheme 2** Formation of triazole derivatives in the ‘catch’ step.

**Scheme 3** Release of 1,2,3-triazoles from the pH PPG 3.
4-hydroxyphenacyl fluoride was used as an actinometer. Irradiated at \( \lambda = [313 \pm 1.5] \) nm (3a, 3d) or (320 \pm 10) nm (3c, 3e); 4-hydroxyphenacyl fluoride was used as an actinometer. In a mixture of acetonitrile (10%) and acetate buffer (pH = 5.0). In a mixture of acetonitrile (20%) and acetate buffer (pH = 5.0).

| HP derivative | \( \Phi_{\text{dis}}/% \) |
|--------------|------------------|
| 3a \( ^b \)  | 0.09 \( \pm \) 0.02 |
| 3c \( ^b \)  | 2.9 \( \pm \) 0.4  |
| 3d           | 83 \( \pm \) 4      |
| 3e \( ^c \)  | 0.39 \( \pm \) 0.05 |

\( ^a \) Irradiated at \( \lambda = [313 \pm 1.5] \) nm (3a, 3d) or (320 \pm 10) nm (3c, 3e); 4-hydroxyphenacyl fluoride was used as an actinometer. \( ^b \) In a mixture of acetonitrile (10%) and acetate buffer (pH = 5.0). \( ^c \) In a mixture of acetonitrile (20%) and acetate buffer (pH = 5.0).

In conclusion, we synthesized a series of substituted pH P azides and alkynes which can be interlinked using an alkyne-azide 1,3-dipolar cycloaddition protocol and subsequently be released upon irradiation with UV-light via a photo-Favorskii rearrangement. We show that the reactions can be accomplished in aqueous media favorable for biological applications and that the linker can be used to connect/disconnect two (bio)molecules. A rational design of the leaving group (triazole) structure led to a previously reported 4-step synthesis of cyclopropenone 8. It has been shown that this compound photodecarbonylates with a quantum yield of 0.33 to give a strained alkyne 9, which can undergo non-photochemical SPAAC reaction with azides. Thus, a solution of 8 and pH P azide (1a; 1 equiv., \( c \approx 5 \) mg mL\(^{-1}\)) in acetonitrile was irradiated at a wavelength of 350 nm, at which only 8 absorbs (Fig. S6, ESI†). The generated alkyne 9 underwent in situ non-photochemical SPAAC reaction with 1a to form 3e (quantitative yield determined by \(^1\)H NMR; 76% isolated yield). Upon irradiation at 313 nm, this pH P derivative was converted quantitatively into triazole 4e and p-hydroxy-benzyl derivatives 5a and 6a, which could bear a peptide tether to connect the released moiety with a biomolecule, via the photo-Favorskii rearrangement. The disappearance quantum yield of 3e was higher than that of 3a because 4e is a better leaving group compared to 4a.

Our ‘photorelease, catch and photorelease’ strategy is a one-pot multistep procedure, which is solely controlled by light, and does not require any other reagents besides water. In addition, both photochemical steps are orthogonal. The pH P group does not absorb at a wavelength of 350 nm (Fig. S39, ESI†) used for generation of strained alkyne 9; thus cycloadduct 3e remains unreacted in the solution during the first irradiation step. The pH P chromophore transformations rely on irradiation at wavelengths in the UV region (< \( \approx 320 \) nm), which may limit their bioapplications. However, 2-photon excitation could overcome this issue, as was recently successfully demonstrated by Givens and coworkers on pH P-caged diethyl phosphate and ATP using a 550 nm laser.

In conclusion, we synthesized a series of substituted pH P azides and alkynes which can be interlinked using an alkyne-azide 1,3-dipolar cycloaddition protocol and subsequently be released upon irradiation with UV-light via a photo-Favorskii rearrangement. We show that the reactions can be accomplished in aqueous media favorable for biological applications and that the linker can be used to connect/disconnect two (bio)molecules. A rational design of the leaving group (triazole) structure led to the development of a linker that cleaves upon irradiation with near-unity quantum yield. The second generation of this strategy encompasses chromatically orthogonal photoproduction of a strained alkyne which allows for Cu-free click chemistry. We propose that the systems could be used in bioconjugation or drug-delivery applications.
