Hepatoprotective activity of methanolic extract of Syzygium jambos (Linn.) leaf against paracetamol intoxicated Wistar albino rats

N. Thamizh Selvam, V. Venkatakrisnan, R. Dhamodharan, S. Murugesan, S. Damodar Kumar

Research Officer-Biochemistry, National Research Institute for Panchakarma, Cheruthuruthy, Kerala, 1Faculty of Microbiology, Pondicherry University, Pondicherry, 2Associate Professor, Department of Botany, Presidency College, 3Associate Professor, Department of Botany, 4Associate Professor, Department of Chemistry, Pachaiyappa’s College, Chennai, Tamil Nadu, India

Abstract

Hepatoprotective activity of methanolic extract of Syzygium jambos (Alston) (Linn.) leaves against Paracetamol-induced hepatic damage in Wistar albino rats was observed at two different doses, 100 and 200 mg/kg body weight. The healthy control, disease control, and standard drug Silymarin-treated groups were also maintained for the comparison. The liver marker enzymes SGOT, SGPT, ALKP, Serum Bilirubin and other metabolic parameters like total cholesterol, HDL-cholesterol were evaluated in all the experimental groups. The changes in liver function parameters were significant in comparison to disease control group and the observed efficacy was comparable to standard drug. The efficacy of the extract was found to be dose dependent. The histopathology study of liver also supports the presence of hepatoprotective activity in S. jambos by showing improved cytoarchitecture of liver cells in the treated groups. The results obtained in this study indicate necessity for further research on isolation and characterization of functional molecules from the extract.

Key words: Hepatoprotection, liver marker enzymes, paracetamol, Syzygium jambos

Introduction

Liver diseases resulting from liver damage is a global problem. A major causative factor is the increasing alcohol consumption in developed countries.1 Malnutrition, anemia, infection, and availability of over-the-counter hepatotoxic drugs are the most frequent causes of liver damage in developing countries.2 It is well recognized that free radicals are critically involved in various pathological conditions such as cancer, cardiovascular disorders, arthritis, inflammation and liver diseases.3 Chemicals and drugs such as CCl₄ and Paracetamol promote formation of free radicals and consequent lipid peroxidation damages the membranes of liver cells and organelles and causing the swelling and necrosis of hepatocytes. This becomes responsible for the release of cytosolic enzymes such as SGOT, SGPT, and ALKP into the circulating blood.4,5 Indian medicinal plants and many herbal formulations belonging to the traditional systems of medicine like Ayurveda have been investigated as hepatoprotective drugs.5 Less side effects and cost effectiveness are the added advantages of these drugs.

Syzygium jambos popularly known as Jambu is a small tree with spreading branches, leaves, simple, opposite, lanceolate, narrowed into short petioles, secondary nerves joined by a prominent looping intramarginal vein. Flowers greenish white in short terminal racemose cymes, stamens many, yellowish white, fruits pale yellow to pinkish white, globose, seeds 1-2, grey in large cavity of the succulent pulp. The bark is astringent, bitter, hemostatic, depurative, vulnerary, antidiarrheal and anthelmintic.6 S. jambos is used in traditional system of medicine for various clinical conditions like gout, hemorrhages, syphilis, leprosy, colic helminthiasis, wounds, and ulcer.6

The present study has been designed to evaluate the hepatoprotective activity of methanol extract of S. jambos leaves in the experimental animal models of Paracetamol-intoxicated Wistar albino rats.

Materials and Methods

Animals

Adult Wistar Albino Rats of either sex weighing between 150 to 200 g were used for the present study. The animals were housed in polypropylene cages and maintained under standard laboratory conditions (temperature 25 ± 2°C) with dark and
light cycle (14/10 h). They were allowed free access to standard dry pellet feed (Amrut Feeds, Pune) and water ad libitum. The rats were acclimatized to laboratory condition for 30 days before commencement of experiment. All procedure described were reviewed and approved by the Institute Animal Ethics Committee (No. 612/02/A/CPCSEA/IAEC/CRIA/2006-07/02 dated 22-12-2006).

**Chemicals**

Methanol (Qualigens Fine chemicals limited, Mumbai), Paracetamol (GSK), Silymarin (Serum Institute of India, Pune), Sodium chloride (Merck Specialties Private limited, Mumbai), 40% Formaldehyde (Nice chemicals Private limited, Cochin), Serum Glutamate Pyruvate Transaminase (Transasia laboratories, Daman), Serum Glutamate Oxaloacetate Transaminase (Transasia Lab), Alkaline phosphatase and Total protein kits were obtained from Transasia Lab.

**Experimental set up**

The following experimental protocol was followed:[7] Healthy Wistar albino rats were divided into five groups consisting of six animals each. The first group (I) consisted of normal control rats which received single daily dose of distilled water throughout the experiment. The Paracetamol group (II) received single daily dose of distilled water for nine days and single dose of Paracetamol on day 8 (2.5 g/kg). The third group (III) was treated with standard drug Silymarin (100 mg/kg) on all nine days and Paracetamol (2.5 g/kg) on day 8, two hours after administration of Silymarin. The fourth group (IV) was treated with lower dose of methanol extract of S. jambos (Alston) (Linn.) (100 mg/kg) throughout the experiment and single dose of Paracetamol (2.5 g/kg) on day 8, two hours after administration of test extract. The fifth group (V) was treated with higher dose methanol extract of S. jambos (200 mg/kg) on throughout the end and Paracetamol (2.5 g/kg) on day 8, two hours after administration of test extract.

On tenth day of the experiment, rats were anesthetized by light chloroform anesthesia and the blood was withdrawn from retro-orbital plexus. The animals were fasted 12 hours before the collection of blood. After blood collection, the rats were sacrificed by cervical dislocation and their liver, kidney, and heart were excised, rinsed in ice-cold normal saline, and stored in refrigerator.

**Biochemical studies**

Serum was separated by centrifugation at 2500 rpm at 30°C for 15 minutes and utilized for the estimation of various biochemical parameters including Serum Glutamate Pyruvate Transaminase (SGPT),[9] Serum Glutamate Oxaloacetate Transaminase (SGOT),[10] Alkaline Phosphatase (ALKP),[11] Total bilirubin,[12] and Lipid profile.[13]

**Histopathological investigation**

Small pieces of liver tissues of each group of animals were stored in solution of commercial formaldehyde for histopathological studies.

**Statistical analysis**

Each experimental value is expressed as the Mean ± SEM. Statistical calculations of the data were performed using Student’s t-test and ANOVA analysis. A probability of P < 0.05 and P < 0.01 was considered as significant.

**Results**

Administration of Paracetamol (2.5 g/kg, p.o.) resulted in a marked increase in the serum hepatic enzyme levels, SGOT, SGPT, ALKP and serum Bilirubin as compared to the normal controls, which indicates liver damage. Pretreatment of the rats with methanol extract of S. jambos prior to Paracetamol administration caused a significant reduction in the values of SGOT, SGPT, ALKP activity and serum Bilirubin in dose-dependent manner almost comparable to the Silymarin-treated group [Table 1].

The other biochemical parameters like Cholesterol and HDL-Cholesterol were also analyzed for the understanding of extract’s influence on fatty acid metabolism [Table 2] and the parameters were found to be influenced to significant extent.

**Table 1: Efficacy of test extracts on liver function tests in Paracetamol intoxicated Wistar albino rats**

| Groups                      | Liver function tests |
|-----------------------------|----------------------|
|                            | SGOT (U/l) | SGPT (U/l) | ALKP (U/I) |
| Healthy control             | 98.35±2.29   | 48.24±2.61 | 89.52±1.27 |
| Disease control             | 362.44±3.74*  | 254.96±4.20* | 194.36±5.19* |
| Silymarin treated (100 mg/kg)| 153.26±2.76*  | 68.49±2.55** | 97.80±4.14* |
| *S. jambos L.D (100 mg/kg)  | 183.67±3.54** | 133.48±4.11** | 116.85±3.10* |
| *S. jambos H.D (200 mg/kg)  | 162.46±4.95*  | 89.70±4.20* | 92.14±2.79** |

Values are means±SEM, n=6 animals in each group, *P<0.05, **P<0.01 when compared to disease control) P<0.001 when compared to healthy control by student’s ‘t’ test. SGOT: Serum glutamate oxaloacetate transaminase, SGPT: Serum glutamate pyruvate transaminase, ALKP: Alkaline phosphatase

**Table 2: Effect of test extracts on biochemical parameters in experimental rats**

| Groups                      | Biochemical parameters (mg/100 ml) |
|-----------------------------|-----------------------------------|
|                             | Total bilirubin | Direct bilirubin | Cholesterol | HDL-Cholesterol |
| Healthy control             | 1.23±0.10       | 0.21±0.04        | 108.66±3.23 | 49.52±1.10      |
| Disease control             | 4.56±0.18       | 1.65±0.09        | 187.55±3.80* | 29.80±1.23      |
| Silymarin treated (100 mg/kg)| 1.44±0.12**     | 0.47±0.07**      | 114.20±3.07** | 43.25±1.80** |
| *S. jambos L.D (100 mg/kg)  | 2.00±0.09*      | 0.81±0.05*       | 128.50±4.12* | 39.15±1.20**    |
| *S. jambos H.D (200 mg/kg)  | 1.68±0.05**     | 0.52±0.06*       | 110.40±3.60* | 42.60±1.16** |

Values are means±SEM, n=6 animals in each group, *P<0.05, **P<0.01 when compared to disease control) P<0.001 when compared to healthy control by student’s ‘t’ test. HDL: High density lipoprotein
The Paracetamol control group showed increase in Cholesterol level and decreased HDL-Cholesterol level in comparison to the normal control. *S. jambos* -treated groups showed reduced level of total cholesterol (110.40 ± 3.60 mg/deciliter) and the elevated level of HDL-Cholesterol in serum. These effects were found to be statistically significant in comparison to paracetamol control group. The efficacy of the extracts was comparable with that of the standard drug *Silymarin* and activity was found to be dose dependent.

The hepatoprotective effect of the *S. jambos* was confirmed by histopathological examination of the liver tissue. The histological architecture of paracetamol-treated liver sections showed extensive hemorrhage and necrosis in the liver parenchyma; hepatocytes showed vacuolated cytoplasm; and collections of inflammatory cells and siderophages [Figures 1 and 2]. Pretreatment with higher dose of the *S. jambos* extract prevented the toxicant-induced changes to significant extent –the sections were almost normal similar to the effect observed in the Silymarin-treated groups [Figures 3 and 4]. This shows presence of significant hepatoprotective effect in the test extracts. Paracetamol is a well-known antipyretic and analgesic agent, which is safe in therapeutic doses, but can produce fatal hepatic necrosis in man, rats, and mice in toxic doses. It is used as an experimental hepatotoxic agent. It is metabolized in the liver to extractable glucuronide and sulfide conjugates. An obvious sign of hepatic injury is the leaking of cellular enzymes into the plasma due to disturbance caused in the transport functions of hepatocytes. When liver cell cytoplasm is damaged, a variety of enzymes located normally in cytosol are released into the blood, thereby causing increased enzyme levels in the serum. The estimation of enzymes in the serum is a useful quantitative marker of the extent and type of hepatocellular damage.

In the present investigation, the dose of Paracetamol administered has produced (2.5 g/kg) liver injury in rats. The rats treated with an overdose of Paracetamol developed significant hepatic damage, which was observed through a substantial increase in the concentration of serum parameters. The abnormal high levels of liver function test parameters

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**Figure 1:** Healthy control rat showing normal hepatic nuclei and cytoplasm. Central venous system and sinusoidal spaces are normal

**Figure 2:** PCM treated rats: Liver showing extensive area of hemorrhage and necrosis in the liver parenchyma, hepatocytes show vacuolated cytoplasm. Collection of inflammatory cells and siderophages are observed

**Figure 3:** Silymarin-treated rat liver showing comparable normal architecture of liver. Some of the hepatocytes show vacuolated cytoplasm. Portal areas and central veins are normal

**Figure 4:** *Syzygium jambos* 200 mg/kg body weight treated group showing marked improvement over Paracetamol group. Hepatocytes show normal cytoplasm. Kupffer cells are normal
like SGOT, SGPT, ALKP, bilirubin, and protein levels were observed in paracetamol control that indicate the damage to the hepatic cells. Pretreatment of the rats with methanol extract of S. jambos at 100 and 200 mg/kg, p.o., for 5 days before Paracetamol administration resulted in significant reversal of Paracetamol-induced elevation of serum marker enzymes.

The results are in agreement with the commonly accepted view that serum level of transaminase returns to normal with healing of hepatic parenchyma and the regeneration of hepatocytes. Furthermore, the stimulation of hepatic regeneration was known to make the liver more resistant to damage by toxins.

**Conclusion**

The present study revealed that the methanol extract of S. jambos leaves can be considered as a significant hepatoprotective agent in view of observed significant hepatoprotective activity in the model system. The results obtained suggest the necessity of carrying out further research on isolation and characterization of specific functional molecules to understand the exact mode of action.

**References**

1. Nadeem M, Dangiya PC, Pasha KV, Imara M, Balani DK, Vohora SB. Hepatoprotective activity of Solanum nigrum fruits. Fitoterapia 1997;58:245-54.
2. WHO. Bulletin of Regional Health Information, 1980-1990: World Health Organization Regional Office for SE Asia. New Delhi: World Health House; 1992.
3. Quambo X, Koji H, Yasuhiro T, Tadata T, Tsuneo N, Shigetoshi K. Hepatoprotective activity of Phenylehanoids from Cistanche deserticola. Planta Med 1998;64:120-5.
4. Singh B, Saxena AK, Chandan BK, Anand KK, Suri OP, Suri KA, et al. Hepatoprotective activity of verbalbin on experimental liver damage in rodents. Fitoterapia 1998;58:135-40.
5. Jose JK, Kuttan R. Hepatoprotective activity of Emblica officinalis and Cheyavanprash. J Ethnopharmacol 2000;72:135-40.
6. Warner PK, Nanbiar VP. Ramankuty C. Indian medicinal plants. A Compendium of 500 species. Vol. 5. Chennai: Orient Longman Ltd; 1996. p. 229-31.
7. Gupta AK, Misra N. Hepatoprotective activity of aqueous extract of Chamomile capituli in paracetamol intoxicated albino rats. Am J Pharmacol Toxicol 2006;1:1-7.
8. Wolf PL, Williams D, Coplon N, Coulson AS. Low Aspartate transaminase activity in serum of patients undergoing chronic hemodialysis. Clin Chem 1972;18:567-8.
9. Tietz NW. Text book of clinical chemistry; WB Saunders; Philadelphia, 1986. p. 1388-91.
10. Wilkinsons JH, Winsten S. Evaluation of a new system for the kinetic measurement of serum alkaline phosphatase. Clin Chem 1969;15:487-95.
11. Tietz NW. Text book of clinical chemistry; WB Saunders; Philadelphia, 1986. p. 579-80
12. Allain CC, Poon LS, Chan LS, Richmond W, Fu PC. Enzymatic determination of serum total cholesterol. ClinChem1974;20:470-5.
13. Mitchell JR, Jollow DJ, Potter WZ, Davis DC, Gillette JR, Brodie BB. Acetaminophen induced hepatic necrosis. I. Role of drug metabolism. J Pharmacol Exp Ther 1973;187:185-94.
14. Chen X, Sun CK, Han GZ, Peng JY, Li Y, Lu YX, et al. Protective effect of tea polyphenols against paracetamol-induced hepatotoxicity in mice is significantly correlated with cytochrome P450 suppression. World J Gastroenterol 2009;15:1829-35.
15. Torielli MV. Pathological aspects of liver injury produced by drugs. In: Slater TF, editor: Biochemical mechanism of liver injury. London, U.K.: Academic Press; 1978. p. 631.
16. Schmidt LE. Age and paracetamol self-poisoning. Gut 2005;54:686-90.
17. Holt MP, Cheng L, Ju C. Identification and characterization of infiltrating macrophages in an acetaminophen –induced liver injury. J Leukoc Biol 2008;84:1410-21.
18. Zimmerman HJ, Seefl BB. Enzymes in hepatic disease. In: Goodly EL, editor: Diagnostic Enzymology, Philadelphia, USA: Lea and Feibiger; 1970. p. 1-38.
19. Campion SN, Johnson R, Aleksunes LM, Goodkin MJ, Rooijen NV, Scheffer LG, et al. Hepatic Mrp4 induction following acetaminophen exposure is dependent on Kupffer cell function. Am J Physiol Gastrointest Liver Physiol 2008;295:294-304.
20. Satturwar PM, Fulzele, Joshi SB, Dorle AK. Hepatoprotective activity of Haridradighrita on carbon tetrachloride-induced liver damage in rats. Indian J Exp Biol 2003;4:1447-51.

हिन्दी सारांश

यकृत रक्षण हेतु जम्बू पत्र अर्क का पेयासिटामोल के साथ विस्तृत अल्बिनो चूहों में तुलनात्मक अध्ययन

तमिल सेल्वम, बेंकटकृष्णन, दामोदरन, मुरुगेन्द्र, दामोदर कुमार

वर्तमान अध्ययन में, जम्बू पत्र (Syzygium jambos) अर्क की दो अलग अनजान मात्रा, 100 और 200 मिलीग्राम/किग्रा शरीर के वजन वाले Wistar चूहों को प्रशांतित किया गया। स्वस्थ नियंत्रण, रोग नियंत्रण और मानक औषधि Silymarin का समूह भी तुलना के लिए बनाया गया। यकृत पर हानि करने वाले पेयासिटामोल और मानव औषधि Silymarin का समूह भी तुलना के लिए बनाया गया। यकृत रक्षण के लिए प्रतिरोधी मामलों में एसजीओटी, एसजीपीटी, एलक्सप्रोफा और कोलेस्ट्रोल जैसे अन्य वचनप्रभाव उपक्रम को ध्यान में रखा गया। यकृत मानकों में उपर्युक्त, रोग नियंत्रण समूह की तुलना में महत्त्वपूर्ण थे और मानव प्रभावकारिता मानक औषधि Silymarin के बारे में था। यकृत कोशिकाविकृतिविज्ञानी अध्ययन में भी चिकित्सा समूह में यकृत की कोशिकाओं में सुधार sytoarchitecture होना हज़ार पत्र में hepatoprotective गतिविधि की उपस्थिति का समर्थन करता है। इस अध्ययन में प्राय परिणामों और अधिक अनुसंधान की आवश्यकता का संकेत मिलता है।