Development and validation of a CT-based radiomics nomogram for preoperative prediction of tumor histologic grade in gastric adenocarcinoma

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Abstract

Objectives: To develop and validate a radiomics nomogram for preoperative prediction of tumor histologic grade in gastric adenocarcinoma (GA).

Methods: This retrospective study enrolled 592 patients with clinicopathologically confirmed GA (low-grade: n=154; high-grade: n=438) from January 2008 to March 2018 who were divided into training (n=450) and validation (n=142) sets according to the time of computed tomography (CT) examination. Radiomic features were extracted from the portal venous phase CT images. The Mann-Whitney U test and the least absolute shrinkage and selection operator (LASSO) regression model were used for feature selection, data dimension reduction and radiomics signature construction. Multivariable logistic regression analysis was applied to develop the prediction model. The radiomics signature and independent clinicopathologic risk factors were incorporated and presented as a radiomics nomogram. The performance of the nomogram was assessed with respect to its calibration and discrimination.

Results: A radiomics signature containing 12 selected features was significantly associated with the histologic grade of GA (P<0.001 for both training and validation sets). A nomogram including the radiomics signature and tumor location as predictors was developed. The model showed both good calibration and good discrimination, in which C-index in the training set, 0.752 [95% confidence interval (95% CI): 0.701–0.803]; C-index in the validation set, 0.793 (95% CI: 0.711–0.874).

Conclusions: This study developed a radiomics nomogram that incorporates tumor location and radiomics signatures, which can be useful in facilitating preoperative individualized prediction of histologic grade of GA.

Keywords: Adenocarcinoma; histologic grade; nomograms; stomach neoplasm; X-ray computed tomography

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Introduction

Although the incidence of gastric cancer has decreased due to improved primary prevention, the mortality rate of gastric cancer still ranks third globally (1,2). In 2012, there were an estimated 952,000 new cases, of which nearly
three-quarters occurred in Asia, and more than two-fifths occurred in China (3). Gastric adenocarcinoma (GA) accounts for 95% of gastric malignancies (4). Referring to the World Health Organization (WHO) classification (5th edition), well and moderately differentiated GAs are classified as low-grade, and poorly differentiated GAs are classified as high-grade for better prognostic significance and reproducibility (5,6). The histologic grade of tumor differentiation has been proven to be associated with clinical outcomes (7-13). Patients with high-grade disease face a higher risk of lymph node metastasis and recurrence than those with low-grade disease, which leads to a poorer prognosis after curative surgery (14,15). Recent studies have demonstrated that patients with limited metastatic disease exhibit favorable overall survival after receiving neoadjuvant chemotherapy (16,17). However, the risk of lymphopenia and hemoglobinopathy is also significantly increased (18). To alleviate the side effects brought by excessive preoperative treatments, it is worth predicting the histologic grade of GA. Furthermore, predicting the preoperative histologic grade can also help identify patients with a higher risk of recurrence and help select individualized and proper therapeutic strategies, which will ultimately improve the outcome.

Currently, preoperative endoscopic biopsy is used as the gold standard for the diagnosis of GA and its histologic grade (19). However, as an invasive procedure, endoscopic biopsy carries a risk of iatrogenic perforation and postendoscopic infection (20,21). Additionally, intratumor heterogeneity may inevitably produce sampling bias, and the histological results may be inconsistent with those obtained during surgery (22,23). For patients with “indefinite for neoplasm/dysplasia” lesions, the diagnostic accuracy for adenocarcinoma is even lower than 70% (24). Therefore, an auxiliary method in addition to biopsy for GA grading may be beneficial.

In clinical practice, contrast-enhanced computed tomography (CT) is used for preoperative staging of GA (19). Liu et al. leveraged the textures of CT images to predict gastric cancer differentiation degree, Lauren classification, and vascular invasion status (25). However, subjective selection of CT textures may cause intra- and inter-observer variability. Li et al. studied iodine concentration in spectral CT, which demonstrated potential in the diagnosis of gastric cancer and its histologic types (22). However, spectral CT is still not widely used. Recently, radiomics has been proposed to convert medical images into high-dimensional features, which allows high-throughput data mining and quantitative feature measurements (26,27). In addition, a number of studies have shown that radiomic feature extraction and comprehensive feature analysis are conductive to individualized management for patients (28-31). Although a previous study used radiomics signatures to predict adverse histopathological status (including WHO grade) of gastric cancer (32), an optimal prediction model combining radiomics signatures and clinical features has yet to be developed.

In this research, we aimed to associate the radiomics signature with clinical characteristics to construct and validate a radiomics nomogram to assist in predicting the histologic grade of GAs preoperatively.

Materials and methods

Patients

This study was approved by the institutional research board of Guangdong Provincial People’s Hospital. The requirement for informed consent was waived because this was a retrospective study. The records and images of 738 patients with GA in Guangdong Provincial People’s Hospital from January 2008 to May 2018 were collected and obtained. All these patients with GA satisfied the following inclusion criteria: 1) contrast-enhanced CT examination ≤1 month preoperatively; 2) visible tumor lesions on CT images; 3) postoperative specimen with pathologic diagnosis of histologic differentiation grade of GA; and 4) received no preoperative neoadjuvant chemotherapy or radiotherapy. A total of 146 cases were excluded from the 738 cases collected. More details can be found in Figure 1.

Figure 1 Flowchart of patient enrollment.
The 592 enrolled patients with GA (mean age, $58.83\pm12.31$; range, 26–90) years were separated into training and validation sets based on the CT examination time. The training set consisted of 450 patients, and the validation set consisted of 142 patients.

We collected baseline clinicopathologic characteristics, including age, sex, clinical stage, CT-reported tumor location, preoperative carcinoembryonic antigen level (CEA, 0–5 ng/mL: normal; >5 ng/mL: abnormal), carbohydrate antigen (CA19-9, 0–27 U/mL: normal; >27 U/mL: abnormal), time of preoperative CT scan and date of surgery, preoperative and postoperative histologic grade, clinical stage, CT-reported main tumor location and T stage from the institution archives. There were no missing data for any of the clinical messages described above except for 77 missing preoperative histologic grades. The relative location of the tumor inside the stomach was also taken into consideration. According to the third edition of the Japanese classification of gastric carcinoma (33), tumors of the stomach were anatomically designated into three parts according to the main tumor location on CT: upper, middle and lower portions. The gastric stump was listed as the fourth location since it does not belong to any of the aforementioned locations. According to the main tumor location on CT: upper, middle and lower portions. The gastric stump was listed as the fourth location since it does not belong to any of the aforementioned locations. The clinical stage and CT-reported T stage were diagnosed based on the American Joint Committee on Cancer (AJCC) staging system (8th edition) (34).

**Histologic grade assessment**

Pathohistologic examination of each pre- and postoperative specimen was performed by two pathologists blinded to the CT findings. In concordance with the 5th edition of the WHO tumor classification of the digestive system (5) and with the consent of the two pathologists, the histologic results were divided into low- and high-grade.

**CT image acquisition protocol**

Before the CT scan, each patient fasted for more than 5 h and was asked to drink 600–1,000 mL of water. Then, routine non-enhanced CT was performed with multidetector row CT (MDCT) scanners, which covered the whole stomach region. Then, iodinated contrast (Ultravist 370, Bayer Schering Pharma, Berlin, Germany) was injected into the antecubital vein through an automatic power pump injector (Ulrich CT Plus 150, Ulrich Medical, Ulm, Germany) at a dose of 1.5 mL/kg and a rate of 3.5 mL/s. After 30- and 60-second delaying intervals, arterial phase and portal venous phase images were obtained, respectively. The CT scanning parameters are shown in Supplementary Table S1.

**Radiomics feature extraction and feature analysis methodology**

Since most gastric tumor lesions showed significant enhancement in the portal venous phase and could be well delineated from the adjacent tissues, CT images of this phase were obtained from the picture archiving and communication system (PACS, Carestream, Canada) for further image feature extraction. ITK-SNAP software (Version 3.6.0, http://www.itksnap.org, USA) was used for manual delineation of visible tumor contours and obtaining regions of interest (ROIs) for radiomics analysis by two radiologists with 2 and 5 years of experience in the diagnosis of gastrointestinal images (33). Intragastric air, necrosis area, enlarged lymph nodes, and perigastric adipose tissue were carefully excluded from the contours (Figure 2).

Then, a volume of interest (VOI) was constructed by integrating the sequences of ROIs. Before feature extraction, the CT image sequence was rescaled with one pixel size of $1 \text{mm} \times 1 \text{mm} \times 1.25 \text{mm}$ by linear interpolation. The pixels from −300 Housefield units (HU) to 700 HU were normalized to the range of [1, 100] by applying min-max normalization. After that, the radiomic features of VOIs were extracted and analyzed using the embedded algorithms provided by MATLAB R2019b (MathWorks, Natick, MA, USA). Extracted features consisted of first-order intensities, texture features, wavelet features, and shape- and size-based features.

**Selection of radiomics features and construction of radiomics signature**

To investigate intra- and inter-observer reproducibility, 25

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**Figure 2** An example of manual segmentation in gastric adenocarcinoma. (A) A diffusely infiltrating mass with enhancement is shown on the portal venous phase CT image; (B) The manually segmented area is shown on the same axial slice.
cases were chosen randomly from the training set, and VOI
delineation and radiomics feature extractions were repeated
by the two radiologists. To determine the more robust
features, the reproducibility and stability of the extracted
features were assessed by intra- and inter-class correlation
coefficients (ICCs), which were calculated based on feature
extractions by reader 1 twice and once by each of the two
readers, respectively. Only stable features with ICC>0.90
were reserved (36).

For data standardization, features of the training and
validation sets were first normalized by the mean ($\bar{X}$) and
standard deviation (SD) of the training set with z-score
according to the following formula:

$$X' = \frac{X - \bar{X}}{SD}.$$

To reduce the feature dimension and to filter out
redundant features, the Wilcoxon rank-sum test was
introduced to further determine the features with a
statistically significant difference (P<0.05) in two labels
from the preselected features. The least absolute shrinkage
and selection operator regression (LASSO) algorithm was
utilized for final dimensionality reduction. Ten-fold cross-
validation was used to select the tuning parameter ($\lambda$) by
using the minimum criteria and 1 standard error of the
minimum criteria (the 1-SE criteria). To lower the risk of
overfitting, the log ($\lambda$) corresponding to 1-SE was chosen,
where there are 12 nonzero coefficients. The details are
provided in Figure 3. The features with the most
significance and the best reproducibility were ultimately
preserved. After that, the coefficients of each feature were
calculated by a generalized linear model. The radiomics
score, i.e., the Rad-score, of every single patient was
calculated by a weighted linear combination of the features
and their corresponding coefficients.

**Assessment of radiomics signature predictive performance**

The discriminative power of the radiomics signature was
tested by the receiver operating characteristic (ROC) curve.
The maximum positive likelihood ratio (true positive
value/false positive value) was used to determine the
optimal cutoff value. Note that the cutoff value of the
validation set was directly drawn from the training set to
guarantee independence. The specificity, sensitivity and
accuracy were also computed using the same cutoff value.
The area under the receiver operating characteristic curve
(AUC) was also introduced to evaluate the effectiveness of
the predictive model in both the training and validation
sets.

**Radiomics nomogram construction**

Multivariable logistic regression analysis was performed to
establish a prediction model by combining the radiomics
signature and independent clinicopathologic predictors

![Figure 3](image-url)

**Figure 3** Feature selection with LASSO binary logistic regression model. (A) Tuning parameter ($\lambda$) selection in the LASSO model using 10-fold cross-validation. AUC was plotted vs. log ($\lambda$). Using the minimum criteria and the 1 standard error of the minimum criteria (the 1-SE criteria), dotted vertical lines were drawn at the best value. The 1-SE criteria were chosen according to 10-fold cross-validation, whereas 12 radiomics features were chosen; (B) LASSO coefficient profiles of features. A coefficient profile plot was plotted vs. log ($\lambda$). As $\lambda$ becomes larger, the coefficients of more features shrank to 0. Each colored line represents the coefficient of each feature. The vertical grey line was drawn at the selected $\lambda$, where 12 features had nonzero coefficients. LASSO, least absolute shrinkage and selection operator; AUC, area under the receiver operating characteristic curve.
with P values less than 0.05 in the univariable analysis. We visualized the model as a radiomics nomogram in the training set based on multivariable logistic analysis to promote the clinical application value of the prediction model.

**Assessment of radiomics nomogram predictive performance**

The ROC curve was applied in the assessment of the discriminative performance of the radiomics nomogram. DeLong’s test was executed to calculate the statistical significance in comparing differences in AUCs between the radiomics nomogram and the clinical model and between the radiomics nomogram and biopsy results. The radiomics nomogram was assessed by a calibration curve.

**Statistical analysis**

Statistical analysis was performed using IBM SPSS Statistics (Version 26.0; IBM Corp., New York, USA) and R software (Version 3.5.1; R Foundation for Statistical Computing, Vienna, Austria). Student’s t test and Pearson Chi-square test were used to compare continuous and categorical data, respectively. All P values were two sided, and P<0.05 was considered statistically significant.

**Results**

**Clinical characteristics**

In the total 592 cases included, 515 accepted preoperative biopsies, among which 97 with preoperative diagnosis of low-grade GA and 346 patients with preoperative diagnosis of high-grade GA had identical postoperative results as the preoperative ones. Forty of the patients who were preoperatively diagnosed with low-grade GA were postoperatively diagnosed with high-grade GA; among them, 28 patients’ radiomic results were consistent with their postoperative results. Thirty two of the patients who were preoperatively diagnosed with high-grade GA had postoperative low-grade results; among them, 23 patients’ radiomic results were consistent with their postoperative low-grade results. The training set consisted of 300 males and 150 females, with average age of 58.54±12.29 (range: 26−90) years, and the validation set consisted of 88 males and 54 females, with average age of 59.77±12.38 (range: 30−87) years. No statistically significant difference was found in age or sex between the training and validation sets (P=0.296; P=0.305, respectively).

Table 1 demonstrates the clinical characteristics of patients in training and validation sets.

| Characteristics | Training set [n (%)] (N=450) | Validation set [n (%)] (N=142) |
|-----------------|------------------------------|---------------------------------|
| Age (±s) (year) | Low grade | High grade | P* | Low grade | High grade | P* |
| Male | 84 (78.5) | 216 (63.0) | 0.003 |
| Female | 23 (21.5) | 127 (37.0) | |
| Gender | 61.55±9.91 | 57.59±12.81 | 0.001 |
| CEA level | Normal | 81 (75.7) | 291 (84.8) | 0.029 |
| Abnormal | 26 (24.3) | 52 (15.2) | |
| CEA | Normal | 77 (72.0) | 274 (79.9) | 0.084 |
| Abnormal | 30 (28.0) | 69 (20.1) | |
| Tumor location** | Upper-third | 47 (43.9) | 57 (16.6) | <0.001 |
| Middle-third | 16 (15.0) | 106 (30.9) | |
| Lower-third | 43 (40.2) | 178 (51.9) | |
| Gastric stump | 1 (0.9) | 2 (0.6) | |
| Rad-score (95% CI) | 0.283 (0.222−0.363) | 0.215 (0.139−0.282) | <0.001 |

CEA, carcinoembryonic antigen; CA 19-9, carbohydrate antigen 19-9; 95% CI, 95% confidence interval; *, Continuous variables were compared using student t test while categorical variables were compared using Chi-square test; **, Tumors of the stomach were categorized into four parts based on the main tumor location reported by CT.
patients included in the study. Age, sex and tumor location exhibited significant differences between patients with low- and high-grade GAs in both the training and validation sets (P=0.001 and P=0.004; P=0.003 and P<0.001; P<0.001 and P<0.001). In contrast, CEA and CA19-9 levels showed no significant difference between them in either the training or validation sets (P>0.05).

**Radiomics features reproducibility**

For each patient, the VOI was extended into a total of 16,384 three-dimensional radiomics features, of which 9,449 features were strongly reproducible (ICC>0.90). For further selection, the Wilcoxon rank-sum test was conducted to filter the features with statistically significant differences between GA of different grades and drew 427 filtered features.

**Radiomics features selection and signature construction**

The LASSO logistic regression model was used to select the final 12 features with relatively high predictive outcome from the 427 robust features in the training set (Figure 3A,B). The radiomics score (Rad-score) of each case with GA was calculated by a weighted linear combination of these twelve features and their corresponding coefficients.

**Radiomics signature predictive performance**

For low- and high-grade GA patients, the median Rad-scores were 0.283 and 0.215 in the training set and 0.302 and 0.189 in the validation set, respectively. Regardless of the training set or the validation set, the median Rad-scores in the low-grade group were higher than those in the high-grade group (P<0.001 for both). The ROC curves shown in Figure 4 yield AUCs of 0.705 [95% confidence interval (95% CI): 0.650–0.760] for the training set and 0.720 (95% CI: 0.630–0.810) for the validation set, demonstrating the discriminative power of the radiomics signature. The statistical results, including sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV) and accuracy, are demonstrated in Table 2.

**Radiomics nomogram construction**

In the univariate analysis, sex, age, radiomics score, CEA

![Figure 4](https://www.cjcrcn.org/Chin J Cancer Res 2021;33(1):69-78)
level and tumor location were associated with the histologic grade of GA. Multivariable logistic regression analysis identified the radiomics score and tumor location as independent predictors (Table 3). The tumor location was integrated into the nomogram with the radiomics signature (Figure 5).

Radiomics nomogram predictive performance

Figure 4 demonstrates the ROC curves of the radiomics nomogram, radiomics signature and tumor location alone. The radiomics nomogram demonstrated relatively good discriminative power for histologic grade prediction among the three indicators, with improvements in the AUC from 0.662 for the clinical model to 0.752 (P<0.05, DeLong’s test) in the training set and from 0.716 for the clinical model to 0.793 (P=0.03, DeLong’s test) in the validation set. In our datasets, biopsy outperformed the nomogram, with AUCs of 0.824 in the training set (P<0.05, DeLong’s test) and 0.825 in the validation set (P<0.05, DeLong’s test). The calibration curve of the radiomics nomogram shown in Figure 4 depicted good agreement between the observed outcome and the prediction.

Discussion

In this study, a radiomics nomogram that preoperatively predicts the histologic grade of GA based on tumor location and radiomics signatures extracted from CT images was developed and validated. It demonstrated AUCs of 0.752 and 0.793 in the training and validation sets, respectively.

In practice, surgeons conventionally use endoscopic biopsy for the diagnosis of GA and its histologic grade. However, the risks of gastric perforation, bleeding, abdominal pain, postendoscopic infection, and an overall discordance rate of 21.5% between pathohistologic results of endoscopic biopsy and postoperative specimens in mucosal GA indicates a rather limited value of endoscopic biopsy (19-21,37). The comparison between the preoperative and postoperative pathohistologic results of

Table 2 Predictive performance of radiomics nomogram, clinical characteristics and radiomics signature in training and validation sets

| Predictive performance | AUC (95% CI) | Sensitivity | Specificity | PPV     | NPV     | Accuracy |
|------------------------|-------------|------------|------------|---------|---------|----------|
| Training-nomogram      | 0.752 (0.701–0.803) | 0.697 | 0.720 | 0.888 | 0.425 | 0.702 |
| Validation-nomogram    | 0.793 (0.711–0.874)  | 0.947 | 0.511 | 0.796 | 0.828 | 0.803 |
| Training-clinic        | 0.662 (0.606–0.718)  | 0.828 | 0.449 | 0.828 | 0.449 | 0.738 |
| Validation-clinic      | 0.716 (0.624–0.809)  | 0.916 | 0.511 | 0.791 | 0.750 | 0.782 |
| Training-radiomic      | 0.705 (0.650–0.760)  | 0.659 | 0.645 | 0.856 | 0.371 | 0.656 |
| Validation-radiomic    | 0.720 (0.630–0.810)  | 0.842 | 0.553 | 0.792 | 0.634 | 0.746 |

AUC, area under the receiver operating characteristic curve; 95% CI, 95% confidence interval; PPV, positive predictive value; NPV, negative predictive value.

Table 3 Univariable and multivariable analysis of risk factors

| Risk factors  | Univariable analysis | Multivariable analysis |
|---------------|----------------------|------------------------|
|               | OR                   | 95% CI                 | P          | OR                   | 95% CI                 | P          |
| Gender        | 2.147                | 1.289–5.578            | 0.003      | 1.364                | 0.781–2.383            | 0.276      |
| Age           | 0.973                | 0.955–0.991            | 0.004      | 0.980                | 0.959–1.001            | 0.066      |
| Radiomics score | 0                   | 0–0.010               | <0.001     | 0.003                | 0–0.028               | <0.001     |
| CEA level     | 0.557                | 0.327–0.947            | 0.031      | 0.718                | 0.401–1.284            | 0.264      |
| CA19-9 level  | 0.646                | 0.393–1.063            | 0.086      |                     |                       |            |
| Location      |                      |                        |            |                      |                       |            |
| Upper-third   | Ref                  |                        |            | Ref                  |                        |            |
| Middle-third  | 5.463                | 2.845–10.488           | <0.001     | 3.380                | 1.682–6.791           | <0.001     |
| Lower-third   | 3.413                | 2.050–5.684            | <0.001     | 2.526                | 0.211–6.615           | <0.001     |
| Gastric stump | 1.649                | 0.145–18.757           | 0.687      | 1.619                | 0.033–8.194           | 0.813      |

CEA, carcinoembryonic antigen; CA 19-9, carbohydrate antigen 19-9; OR, odds ratio; 95% CI, 95% confidence interval.
our study also exhibited a difference of 13.98%, which further verifies that the existence of intra-tumor heterogeneity could lead to a substantial discordance rate while affecting clinical decisions. Thus, researchers are now investigating auxiliary noninvasive approaches to precisely predict the histologic grade of GA preoperatively. A previous study analyzed CT texture to predict pathohistological characteristics, including the differentiation degree of gastric cancer, with a sensitivity of 78.7% and a specificity of 83.3% (25). However, CT texture is subjectively assessed and may lead to intra- and inter-observer variability. Therefore, we controlled this variability in our study by constructing a radiomics signature with more reproducible radiomics features (ICC>0.90). Lin et al. explored and discovered the potential diagnostic significance of preoperative CEA/CA19-9 levels (38), so we included these tumor markers in our study, in addition to other clinical and semantic imaging data, including age, sex and CT-reported tumor location, which were analyzed by univariable analysis. Then, a radiomics nomogram was modeled by combining the radiomics features with independent predictive factors drawn by multivariable analysis. In 2018, a published study showed that normalized arterial phase iodine concentration in spectral CT demonstrated a high efficiency in diagnosing poorly differentiated GA (22). Currently, spectral CT is not widely used in daily clinical practice. Li et al. applied machine learning-based computational models to predict adverse histopathological status, including WHO grade, of gastric cancer, with AUCs of 0.65 and 0.63 in the training and validation sets, respectively (32), while the AUCs of the radiomics nomogram developed in our study were higher (0.752 in the training set and 0.793 in the validation set). Although biopsy outperformed the radiomics model in our study, as a noninvasive method, radiomics still has the potential to aid in diagnosis when the patient cannot tolerate an endoscopic examination due to coagulation disorders or critically ill status or refuses to undergo endoscopy. Since contrast-enhanced CT is routinely performed for the staging and management of GA, CT-based radiomics nomograms may be more applicable and more popular in clinical practice. The nomogram may facilitate preoperative identification of patients with a higher risk of recurrence, worse clinicopathological features and poorer prognosis.

There are some limitations in this study. The radiomics features used in the study were extracted only from the portal venous phase CT images because the differentiation between the tumor and the adjacent normal gastric tissues was maximal in this phase. Nonenhanced scans, arterial phase, and venous phase images with extracted radiomics features should be investigated in further studies. Because of our retrospective collection of cases, the quality of CT images acquired by different CT machines may be unstable. Prospective studies are expected to avoid confounding factors and to assure discriminative power in GA histologic grades of radiomics signatures.

**Conclusions**

A radiomics nomogram in discriminating low-grade GA from high-grade GA before treatment was developed and validated. As a quantitative and noninvasive approach, the radiomics nomogram may play a part in facilitating personalized management and improving outcomes of patients with GA.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

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Table S1 CT scanning parameters for patients

| Scanner                                                      | kV (mAs)* | Rotation time (s) | Detector collimation (mm) | Field of view (mm²) | Matrix       | Reconstruction section thickness (mm) | Acquisition time (s) | Arterial phase | Portal venous phase |
|---------------------------------------------------------------|-----------|-------------------|---------------------------|---------------------|--------------|---------------------------------------|----------------------|----------------|-------------------|
| 256-slice Brilliance iCT (Philips Healthcare, Cleveland, Ohio, USA) | 120       | 0.5               | 128x0.625                 | 360x360             | 512x512      | 1.00                                  | 30                    | 60             |                   |
| 64-slice LightSpeed VCT (GE Medical systems, Milwaukee, WI, USA) | 120       | 0.4               | 64x0.625                  | 360x360             | 512x512      | 1.25                                  | 30                    | 60             |                   |

*, The tube current is automatically selected based on the attenuation of each scanning object and ranges from 100 to 300 mAs.