Background: Psoriatic plaques have been shown to contain increased levels of proinflammatory cytokines. Serum levels of interleukin (IL)-6, IL-7, IL-8, and interferon (IFN-γ) have been reported elevated in psoriatic patients.

Aim: To evaluate serum cytokine profiles in psoriasis patients by improved enzyme-linked immunosorbent assay (ELISA) technique and to correlate these levels with disease severity.

Methods: We analyzed single serum samples from 10 patients with active untreated psoriasis, two patients with active treated psoriasis, and five healthy volunteers for major T helper type 1 and T helper type 2 cytokines using the LINCOplex ELISA multi-analyte detection system that permits simultaneous detection of multiple cytokines from a single sample. The disease severity, including erythema, induration, scale, and surface area, was assessed.

Results: IFN-γ was markedly elevated in all sera from psoriasis patients, 33.8 ± 1.3 pg/ml (mean ± standard error) versus 8 ± 1.5 pg/ml for normal controls (p < 0.01) and positively correlated with all indices of disease severity (Spearman r > 0.6). IL-8 was also increased in psoriasis patients (24.4 ± 1.8 pg/ml) versus normal controls (3.6 ± 1.2 pg/ml) (p < 0.05) and positively correlated with the degree of erythema (Spearman r > 0.6). Mean IL-12 levels were decreased in sera from psoriasis patients (8.5 ± 1.2 pg/ml) compared with normal controls (42.2 ± 5.3 pg/ml) (p < 0.01). Also, serum IL-10 levels were below detection levels in psoriatics compared with controls (6.4 ± 1.3 pg/ml).

Conclusions: This new ELISA system allowed rapid and reliable detection of numerous cytokines in single serum samples from patients with psoriasis. We observed that IFN-γ and IL-8 cytokines were elevated in psoriatics and correlated with parameters of disease severity while IL-10 and IL-12 were decreased.

Introduction

Psoriasis vulgaris is a persistent chronic relapsing, inflammatory and hyperproliferative skin disorder with genetic predisposition. In the United States, psoriasis affects approximately 2% of the population. There is currently no good experimental model for psoriasis. Psoriatic plaques have been shown to contain increased levels of cytokines, including tumor necrosis factor-alpha (TNF-α), interleukin (IL)-6, IL-1α, IL-1β, IL-8, and macrophage migration inhibitory factor, whereas IL-10 expression has been shown to be lower than in perilesional skin.

While TNF-α appears to be playing a critical role in propagation of lesional psoriatic plaques, serum TNF-α has been reported to be within normal limits. On the contrary, serum IL-6, IL-8, and interferon-gamma (IFN-γ) have been reported significantly elevated in the sera of psoriatic patients with active disease. It has been reported that IL-6 elevation positively correlates with Psoriatic Assessment and Severity Index scores. A negative correlation has been seen with IL-1α levels and disease activity. In patients with psoriatic arthritis IL-2R, IL-1ra, and IL-10 were found to be elevated in the sera during periods of disease activity. Of special note, sIL-2R has been shown to have a significant association with the severity of skin involvement, whereas IL-1ra correlates with the number of tender/swollen joints. The current state of knowledge suggests that over-expression of proinflammatory, type 1, cytokines and the under-expression of type 2 cytokines are of pathological significance in psoriasis. As the psoriatic story unfolds, we need more reliable and rapid tests to elucidate the biological relevance of Th1 versus Th2 cytokines in the pathogenesis of psoriasis. Moreover, the measurement of these cytokines may allow for better monitoring and prediction of disease activity.
Methods

Twelve psoriatic patients were admitted to dermatology inpatient ward at Cedars Medical Center/University of Miami for Goeckerman therapy for recalcitrant psoriasis. Of the 12 patients selected for enrollment in this University of Miami Institutional Review Board-approved study, nine had plaque psoriasis, two had guttate and one had erythroderma. Out of nine plaque psoriasis patients, seven had received no systemic therapy and two were on methotrexate. The guttate and erythrodermic patients had not received systemic therapy at the time of the inclusion in the study.

At the time of admission, the patients were assessed by the same physician and assigned scores for four indices of disease severity. Independent scores for body surface area (BSA) involvement, degree of erythema, thickness and scale of a target plaque that denoted the severity of the patients disease state were given. Score for percent BSA involvement was from 1 (0–20%), 2 (20–40%), 3 (40–60%), 4 (60–80%) to 5 (80–100%). A target lesion was selected for assessment; thickness was scored from 0 (flat), 1 (minimal thickness), 2 (mild), 3 (moderate) to 4 (severe). The target lesion was assessed for erythema as follows: 0 (none), 1 (minimal), 2 (mild), 3 (moderate), and 4 (severe). Scaling of the target lesion was assessed as: 0 (none), 1 (minimal), 2 (mild), 3 (moderate), and 4 (severe). On admission the patient’s vital signs, weight and baseline laboratory data were drawn. Each patient consented to the use of residual blood from a routine laboratory draw to measure serum cytokine levels. In addition five healthy non-psoriatic volunteers with no family history of psoriasis consented to have their blood drawn for the purpose of measuring serum cytokine levels.

For the purpose of the assay, 25 μl of sera was used to measure IL-1β, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IFN-γ, granulocyte–monocyte colony-stimulating factor (GM-CSF), and TNF-α. Measurements were made by LINCoPlex simultaneous multi-analyte detection system (Linco Research Inc., St Charles, Missouri, USA) following manufacturer’s instruction. All cytokines are simultaneously measured from a single specimen. The LINCoPlex assay was shown to produce results comparable with standard single cytokine assays. Statistical analysis of the results was performed with the aid of Statistical Software (StatSoft, Tulsa, Oklahoma, USA).

Results

Serum cytokine levels of major Th1 and Th2 type cytokines were simultaneously measured from the single specimen prior to initiation of therapy in patients with recalcitrant psoriasis. Results are presented in Table 1. Th2 type cytokine, IL-10, was undetectable in psoriatic patients, of any type, compared with controls (6.4±1.5 pg/ml). The IL-5, another of the Th2 type cytokines, was not detected in normal nor psoriatic patients. The remaining two tested Th2 type cytokines, IL-4 and IL-6, were present at similar low level in psoriasis patients and normal controls, with the exception that in two psoriasis patients both were significantly increased.

When compared with normal sera, the level of Th1 type cytokine IFN-γ was markedly elevated in all sera from psoriasis patients, 33.8±1.3 pg/ml (mean±standard error) versus 7.6±2.2 pg/ml for normal controls (p < 0.01). Besides, when IFN-γ was compared with clinical indices of disease severity, it showed positive correlation (Spearman rank order test: BSA, r = 0.64; thickness, r = 0.62; erythema, r = 0.7; scale, r = 0.67; p < 0.05). The IL-2, another Th-1 type cytokine, did not show elevation in the serum of psoriasis patients, whereas IL-12 was decreased in psoriatics (8.5±1.2 pg/ml) compared with normal controls (42.2±5.3 pg/ml, p < 0.01). From three tested cytokines that cannot be clearly defined as Th1 or Th2, IL-8, TNF-β, and GM-CSF, only IL-8 was significantly increased in serum of psoriatic patients (24.4±1.8 pg/ml), compared with normal controls (3.6±1.2 pg/ml, p < 0.05) and positively correlated with the degree of erythema (Spearman r > 0.6; p < 0.05). Fig. 1 illustrates the correlation of the erythema score with serum level of IFN-γ and IL-8.

Discussion

Our study shows for the first time that 11 major human cytokines, including the major Th1 and Th2 cytokines, were simultaneously quantitatively determined from a single sample in the serum of psoriatic patients. In prior studies solitary cytokines were measured in separate ELISA studies that can potentially cause variation in results due to the variation between samples and individual experiments. In our testing using LINCoPlex kit and Luminex technology (Linco Research Inc.) 11 captured antibodies for specific cytokines were coupled to specific bead sets with different fluorophores. After analytes were bound to the capture antibody on the beads, a detector antibody coupled to phycoerythrin is used as a reporter. The assay was read using a dual laser system. Using multiple bead sets, 11 different assays were simultaneously performed and detected on the individual serum samples. Performing cytokine testing that way means variation between test samples and individual experiments is eliminated. In addition, the cost of testing is significantly decreased.
Table 1. Summary of demographics, clinical indices of disease severity and serum cytokine levels (pg/ml)

| Patient | Sex | Age (years) | Psoriatic type | BSA | Thickness | Erythema | Scale | Th1 | Th2 |
|---------|-----|-------------|----------------|-----|-----------|----------|-------|-----|-----|
| 1       | Male | 59          | Plaque         | 2   | 4         | 3        | 3     | <3  | <3 |
| 2       | Male | 37          | Plaque         | 2   | 3         | 2        | 3     | 6   | <3 |
| 3       | Female | 75         | Plaque         | 4   | 3         | 3        | 3     | 6   | <3 |
| 4       | Male | 53          | Plaque         | 1   | 3         | 3        | 3     | 6   | <3 |
| 5       | Male | 59          | Plaque         | 1   | 4         | 3        | 3     | 4   | 9  |
| 6       | Female | 25         | Plaque         | 4   | 3         | 3        | 3     | 4   | 9  |
| 7       | Female | 75         | Plaque         | 1   | 4         | 3        | 3     | 4   | 9  |
| 8       | Female | 36         | Plaque-MTX     | 3   | 4         | 4        | 3     | 6   | <3 |
| 9       | Male | 42          | Plaque-MTX     | 3   | 4         | 4        | 3     | 6   | <3 |
| 10      | Female | 64         | Erythrodermic  | 5   | 4         | 4        | 3     | 6   | <3 |
| 11      | Female | 38         | Guttate        | 3   | 2         | 3        | 3     | 4   | 9  |
| 12      | Male | 48          | Guttate        | 4   | 3         | 4        | 3     | 6   | <3 |
| Control 1 | Female | 30       | None           | 0   | 0         | 0        | 0     | <3  | 3  |
| Control 2 | Male | 28          | None           | 0   | 0         | 0        | 0     | <3  | 3  |
| Control 3 | Male | 38          | None           | 0   | 0         | 0        | 0     | <3  | 3  |
| Control 4 | Female | 42        | None           | 0   | 0         | 0        | 0     | <3  | 3  |
| Control 5 | Female | 39        | None           | 0   | 0         | 0        | 0     | <3  | 3  |

Serum cytokines in psoriasis and disease severity

Over-expression of proinflammatory, type 1, cytokines is believed to be of pathogenic significance in psoriasis. Our data confirm previously published data that the TH1 type cytokine IFN-\(\gamma\) is elevated in the serum of psoriatic patients. In addition to this, our data suggest that the serum levels of IFN-\(\gamma\) in psoriatic patients are increased in a proportion of patients compared to normal controls. Among the tested TH1 type cytokines, IL-2 and IL-12 were found to be elevated in patients with active psoriasis, while IL-10 was decreased. Elevation of IL-10 in psoriatic patients has been associated with disease improvement. The administration of recombinant IL-12 provided clinically significant improvement of psoriatic disease to regulate the inflammatory process. Other attempts of these patients with both skin and articular disease to regulate the inflammatory process. 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correlate with BSA score. We do not have a good explanation for that observation.

As per previous reports, we found that patients with psoriatic skin disease had normal serum levels of TNF-α, significantly higher IFN-γ and moderately increased IL-8.4,6,7 We were able to furthermore demonstrate an association between serum level of IFN-γ and clinical indices of severity.

Among the tested cytokines there were three (TNF-α, GM-CSF, IL-8) that cannot be clearly classified as Th1 or Th2. TNF-α, while significantly elevated in the lesional psoriatic plaques of patients, is known to be at normal levels in the psoriatic patient's sera. Our data support previous findings. Even in the patients with an erythroderma the TNF-α level was normal. Of note, TNF-α levels have been noted to remain unchanged after standard psoriasis therapy.7 GM-CSF was not elevated in our patient population. To our knowledge GM-CSF in psoriasis has not been previously reported.

We found IL-8 to be elevated in our patients. The level of IL-8 was found to positively correlate with degree of erythema. IL-8 is a known chemotactic factor for neutrophils and, by an indirect mechanism, induces neutrophil degranulation. As neutrophils are present in the epidermis of the psoriatic lesions, it is possible that IL-8 aids in their recruitment and thus contributes to the erythema. While the kinetics of IL-8 in the serum of patients with psoriasis are not known, one could presume that IL-8 could be a good index for the inflammatory process and a crucial element in the cellular activation of psoriasis.

Among the tested psoriatic patients, two were on methotrexate therapy. This number of patients is not adequate to draw definitive conclusive data.

We realize the limitations of our data, since we had a small number of patients. Yet, with the mounting body of scientific evidence in the literature, and our congruent results we are able to conclude that psoriasis is not, as many times inferred, a Th1 versus Th2 disease. Rather it is a complex disease in which the cytokine network is dysregulated with only some components of the Th1–Th2 cytokine network being involved. The new technique that we utilized in our study is rapid, flexible, and inexpensive compared with the traditional ELISA, and requires a very small amount of serum for testing. It could be modified in the future, to test specific cytokines relevant in psoriasis, to better understand the pathophysiology of psoriasis, and to possibly serve as an objective tool to assess severity, progression and prognosis. Furthermore, the manipulation of these cytokines may allow for new and more specific therapeutic alternatives.

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