Integrin-Mediated Tumorigenesis and Its Therapeutic Applications

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Integrins, a family of adhesion molecules generally exist on the cell surface, are essential for regulating cell growth and its function. As a bi-directional signaling molecule, they mediate cell-cell and cell-extracellular matrix interaction. The recognitions of their key roles in many human pathologies, including autoimmunity, thrombosis and neoplasia, have revealed their great potential as a therapeutic target. This paper focuses on the activation of integrins, the role of integrins in tumorigenesis and progression, and advances of integrin-dependent tumor therapeutics in recent years. It is expected that understanding function and signaling transmission will fully exploit potentialities of integrin as a novel target for tumors.

Keywords: integrin, cancer, signaling transduction, talin, FAK

INTRODUCTION

Integrins are a type I transmembrane protein and the main ligands for cell adhesion. There are altogether 18 α and 8 β subunits known in mammals, generating 24 kinds of heterodimers (1). Each subunit has a large ectodomain, a single transmembrane domain (TMD) and a comparatively short cytoplasmic tail. The transmembrane region is the key link of information transmission and interaction between TMD and cytoplasmic tail, regulating the affinity between integrins and their ligands. Though they vary in size, the classic α subunit is made up of around 1,000 amino acids, compared with 750 for the β subunit (2).

As unique adhesion molecules, integrin can signal in both directions across the plasma membrane. Intracellular activators like talins trigger the conformational changes of integrins and recruit multivalent protein complexes (“clustering”) that bind directly or indirectly to the integrin cytoplasmic tail (3–5). These combinations represent a complex, highly dynamic system that relates to ligand-binding affinity, which is responsible for regulating various aspects of cellular fate like cell migration and extracellular matrix (ECM) assembly and remodeling (6). Events introduced above are called “inside-out” signaling. Integrins also enable human cells to respond to changes in the extracellular environment through outside-in signaling. Outside information communicates to cells via intracellular means, bringing about changes in cell polarity, cytoskeletal structure, gene expression, cell survival and proliferation (7).

Integrin heterodimers are often classified by the special sequences they can recognize. Those sequences are generally known as RGD or LDV tripeptides, or some complex peptide like GFOGER. Researchers conventionally classified integrins into 4 types: RGD receptors, collagen receptors, laminin receptors and leukocyte-specific receptors (8). For example, Integrin αvβ3 binds to a
spectrum of ECM molecules using the RGD triple-peptide motif (9), which includes von Willebrand factor, fibronectin, fibrinogen, proteolyzed forms of collagen and laminin, and vitronectin. Other integrins, like α5β1, can only selectively bind to fibronectin (10).

The binding of integrins and ligands are not only located in the classical extracellular matrix (ECM). Integrin interacts with various proteins on the surfaces of cells, even on fungal cells and viruses. Those proteins include hormones, growth factors, and polyphenols (11). Notably, many growth factors bind to the ECM, and the spatial arrangement of integrin and growth factor binding sites in the ECM enables simultaneous engagement of their cognate receptors on the plasma membrane (12). Integrin involves in proliferative signaling, tumor invasion and metastasis, evasion of apoptosis, and stimulation of angiogenesis. This was achieved by cooperating with growth factor receptors like epidermal growth factor receptor (EGFR), ErbB-2 to amplify downstream pathways such as PI3K, AKT, MAPK and the Rho family small GTPases (13). Tejeshwar et al. found that EGFR regulates integrin tension and the spatial organization of focal adhesions, and that the mechanical tension threshold for outside-in integrin activation is tunable by EGFR (14). There are also plenty of non-ECM molecules that interact with integrins, making integrins essential mediators of cell biology.

**ACTIVATION AND SIGNAL TRANSMISSION OF INTEGRIN**

Each integrin exists either in the “bent” state of low-affinity or in an extended high-affinity conformation (15–17). The transition from a “bent” to an extended conformation is called “activation,” which is reversible and rapid. This process involves two key mechanisms: the extension of the head and the separation of the legs, which are triggered by “inside-out” or “outside-in” signals (18, 19). However, recent work clearly illustrated that integrins are vertically positioned on the cell membrane and exist in three main conformations: bent-closed (inactive), extended-closed (active, low affinity) and extended-open (active, high affinity) conformations (20). (Figure 1) There are two common models for activation of integrins: the “switchblade” and the “deadbolt” models, which describe a transition state from the curved one to the extended conformation (21–23). (Figure 2)

As we know, one of the fully studied integrin pathways is the focal adhesion kinase (FAK) signaling pathway. Upon binding to its specific ligand, it leads to maximal FAK activation. The FAK-Src complex has multiple downstream effectors (24). FAK-Src complex promotes the activity of a GTPase which belongs to the Ras superfamily, which is generally known as Rac1 (Ras-related C3 botulinum toxin substrate 1). Rac1 activation is involved in spreading and in the early stages of migration (25). At later stages of cell spreading or for instance, by constitutive activation of αβ3 via ligand binding, RhoA activity leads to the formation of stressfibers and promotes migration (26). In addition, phosphorylation of FAK leads to the Ras-mediated activation of the MAP-kinase pathway (MAPK/ERK pathway), which is associated with proliferation and tumorigenic behavior. Through this pathway, several transcription factors such as the oncogene C-myc and C-jun are activated via phosphorylation. Therefore, the activation of the MAPK pathway leads to the transcription of genes that are important for cell proliferation and cell cycle progression. This pathway can be activated by cell adhesion (e.g., binding of α5β1 to fibronectin) or growth factors (such as epidermal growth factor (EGF)) (27, 28). Moreover, phosphorylated FAK connects with PI3K, which leads to the activation of AKT via PDK1 (29). The AKT signaling pathway can also lead to the phosphorylation of YAP which acts as an apoptotic suppressor (30). The activation of YAP represents a cross-talk with a newer signaling pathway known as Hippo pathway. This pathway controls organ size by regulating cell proliferation and apoptosis (31).

Dynamic remodeling of adhesions is an important mechanism employed by cells to regulate integrin–ECM interactions and cellular signaling. This is done through rapid endocytic and exocytic trafficking of integrin receptors during cell migration, invasion and cytokerinosis. Integrin traffic is relevant in several pathological processes, especially in cancer. Importantly, conceptual progress in the field has identified well-known cancer oncogenes and mutations as being crucial regulators of integrin traffic. To support their proliferation rate, cancer cells exploit active integrin-mediated ECM endocytosis to directly acquire nutrients from the extracellular environment (32).

Integrin activation is a process of conformational changes which allows integrins to bind their ligands. This process is well modulated through the interaction between the integrin αβ cytoplasmic tails (CTs) and their binding partners. Many researchers believe that the change of cytoplasmic tail is the main cause of conformational change (33). Evidence suggests that talins and kindlins are the proteins that bind to cytoplasmic domain and mediate this process (34). In “inside-out” signaling, intracellular activators such as talins or kindlins, binding to the CTs of β subunit leads to the separation of the α and β tails and induces conformational changes in the ectodomain, thereby increasing its affinity for ligands, also known as the “activation” of integrin (35, 36). Conformational changes and clustering of a single integrin can affect affinity to its ligands (15). The affinity of integrin can also be regulated by ligand in vitro to induce conformational changes in the extracellular domain of integrin. Studies suggest that intracellular tensile forces can also lead to integrin activation that is ultrasensitive to lower levels of forces compared with cytoskeletal adaptor binding alone (37). In general, the bi-directional signaling reactions are regulated by the dynamic interaction of integrins and proteins on both sides of the membrane.

Talin is one of the most well-known integrin activators that mediates integrin adherence to the extracellular matrix. Talins activate integrins by binding to the CTs of β-integrin via its typical 4.1-protein/ezrin/radixin/moesin (FERM) domain. The membrane-proximal NPXY of β-tail has been identified as the talin-binding site, and the membrane-distal NPXY specifically interacts with kindlins. By binding integrins to actin, talin increases the affinity to the corresponding ligands (integrin activation) as well as recruits a large number of proteins to form the core of the integrin adhesion complex, which in turn activates...
adhesion plaque kinases (FAK) and Src family kinases (SFKs) (Figure 1). For example, loss of talin-1 leads to diminished \textit{in vivo} metastasis of prostate cancer cells via FAK–Src complexes and AKT kinase signaling (38). Downregulation of talin-1 has also been shown to promote hepatocellular carcinoma progression (39). In platelets, talin-1 is the principal direct effector of Rap1 GT Pases that regulates platelet integrin activation in hemostasis (40). Researchers now have established a pipeline approach to evaluate the effect of talin-1 mutations. Through a series of computational methods, biochemical and cell biological analysis, results suggested cancer-related point mutations in talin-1 can affect cell behaviour and so may contribute to cancer progression (41).

Another family of FERM-containing proteins is kindlins, which are a recently discovered integrin interaction partners that play a synergistic role in talin activation of integrin. Although the molecular details of talin-mediated integrin activation are known, the mechanism of kindlin involvement in this process remains elusive. In the knockout and overexpression experiments, kindlin-1, kindlin-2, and kindlin-3 could regulate specific integrin activation, but only in accordance with the interaction between talin-1 and the cytoplasmic tail of integrin. Activation of integrin αIIbβ3 was enhanced by co-expression of kindlin-1 or kindlin-2 and decreased by knocking out endogenous Si-RNA of kindlin-2. The ligand binding to integrin αIIbβ3 is activated due to an overexpressed N-terminal head domain of talin (42–44). Ussar S, et al. found that deletion of kindlin-1 in intestinal epithelial cells or colon cancer cell lines reduced talin-dependent integrin β1 activation or directly reduced integrin-mediated cell adhesion (45). Interrupting kindlins’ dimer formation impairs kindlin-mediated integrin activation (46). Zainab H. used all-atomic microsecond-scale molecular dynamics simulations of integrin αIIbβ3 TM/CT structure in an explicit lipid-water environment and then found that kindlin-2 cooperates with talin-1 to facilitate integrin αIIbβ3 activation by enhancing talin-1 interaction with the membrane proximal (MP) region of β3-integrin (47). Both talins and kindlins are essential for integrin conformational

\begin{figure}
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\caption{Signal transduction of the integrin family. From an inactive conformation to a low affinity, intermediate state that may arise from talin and/or kindlin binding. And in the active state, integrin subunits were separated, forming a 45 degree angle. Integrins are connected to the actin cytoskeleton and can initiate cytoskeletal remodeling (Left). Integrin-controlled cell migration is largely mediated by signaling pathways involving members of the focal adhesion kinase (FAK)-SRC family kinase. Integrins are ligated and initiate multiple downstream effectors.}
\end{figure}
activation, to which they seem to contribute differently by allowing the vinculin-mediated perception of mechanical forces (talins) and triggering biochemical signaling pathways (kindlins) (48), e.g. through paxillin and focal adhesion kinase (FAK) (49–51). Though they cooperatively support integrin activation, the functional significance of post-translational modifications of kindlins controlling integrin signaling has been gradually recognized (52).

When it comes to integrin activation, some accelerants such as paxillin (51, 53), ADAP (54) and migfilin (55) must be mentioned. There are also some inhibitors, such as ICAP-1 (56), filamin (57, 58) and sharpin (59), allowing for fine-tuning of the integrin activation process. Findings showed that sharpin may complex with both kindlin-1 and the integrin-β1 cytoplasmic tail to restrict the talin head domain binding, thus inhibiting β1-integrin activation. Besides, integrins also interact with many cytoplasmic proteins, such as Filamina, Dok1 and 14-3-3 proteins, etc. (60).

**EFFECTS OF INTEGRINS ON SELF-RENEWAL AND PROLIFERATION OF TUMOR STEM CELLS**

In cancer, the strict control of proliferation is lost due to extrinsic factors such as the presence of mitogenic compounds (growth factors, cytokines or exogenous substances) or intrinsic factors such as activation of oncogenes, converting cancer cells in a self-sufficient entity. In this context, integrins play a crucial role by directly promoting proliferation or by indirectly interacting with growth factor receptors. Interactions between growth factor receptors and integrins in cancer are involved in proliferation. Three types of interactions can be distinguished (1): direct interaction (2), modulation of expression levels and (3) reciprocal activation (61). Integrin signaling has been shown to drive many stem cell functions. Plaks et al. found that specialized extracellular matrix niches and integrin signaling support the function of normal stem cells and their tumor derivatives (62). It has been found that integrin β1 mediates the adhesion of basal keratinocytes to the basement membrane in epithelium and controls the stem cell renewal by regulating the polarity axis of asymmetric cell and cell cycle progression (63). The highly expressed laminin binding to integrin α6β1 of tumor stem cells not only promotes adhesion to the surface of the endothelial basement membrane near the lumen, but also transmits self-renewal signals through FAK (64). Integrin αVβ5 could play as a functional cancer stem cell marker essential for glioblastoma maintenance and ZIKV infection, providing potential brain tumor therapy (65). A recent study found arsenic and BaP co-exposure human bronchial epithelial cells have a high expression of integrin α4, leading to activation of the Hedgehog pathway and PI3K/Akt pathway, enhancing arsenic and BaP co-exposure-induced cancer stem cell (CSC)-like property and tumorigenesis (66).
ROLE OF INTEGRINS IN ADHESION AND TUMOR Invasion

Extensive evidence shows that the expression of integrin is significantly different in tumor cells compared to normal ones. Integrin signaling in cancer cells is dysfunctional, which is of significance to understand how tumor cells use integrin activity to regulate invasion and movement and to study the regulatory mechanism of integrin function.

It is well known that the transition from carcinoma in situ to invasive cancer is driven by a series of adhesion changes. By remodeling or dissolving E-cadherin-dependent junctions and integrin-mediated adhesion, unmatched cancer cells or groups of cancer cells would separate from adjacent normal cells and the basement membrane below. Through FAK and SFKs, integrins directly phosphorylate E-cadherin–β-catenin complex to remodeling E-cadherin-dependent junctions, promoting the migration and invasion of cancer cells (67). Integrin-mediated adhesion of fibronectin triggers a negative feedback signal that blocks the formation of E-cadherin mediated cell-to-cell adhesion (68). Putting integrin β1 into β1-deficient epithelial cells resulted in loss of cell contact and dispersion of cells (69), suggesting that integrin-extracellular matrix adhesion plays an inhibitory role in the regulation of cell-cell junctions. Therefore, the internal and external signals of integrins can disrupt intercellular adhesion by increasing myosins’ contractibility and E-cadherin junction stability through FAK and SRC signals (70). Integrin and integrin-dependent processes are implicated in almost every step of cancer development, including tumor growth, invasion and perfusion into the vascular system, survival of circulating tumor cells, extravasation into secondary sites, and metastasis and colonization of new tissues. Integrins expressed on the cell surface is to adhere to the ECM. Ligation provides traction that is essential tumor cell survival and invasion. A recent study has indicated that hypoxia selectively enhances the expression of integrin α5β1 receptor in breast cancer to promote metastasis (71). The expression and potential roles of thrombospondins (TSP-4) in the crosstalk between CAFs and gallbladder cancer (GBC) cells has remained unclear. Research showed that a complex TSP-4/integrin α2/HSF1/TGF-β cascade mediates reciprocal interactions between GBC cells and CAFs, providing a promising therapeutic target for gallbladder cancer patients (72).

For most solid tumors, the basement membrane first needs to be breached. This process is thought to require proteolysis, and integrins play their roles by upregulating the expression of matrix metalloproteinases (MMP) and promoting the activation and function of proteases at the extracellular matrix. Integrins control cell migration and invasion by influencing the activity and localization of matrix-degrading proteases, such as urokinase-type plasminogen activator (uPA) and MMP2 (73, 74) Invasive cancers penetrate the stroma through a variety of different integrin-dependent mechanisms and migrate to surrounding tissues in the form of a single cell or groups of cells (75). Furthermore, tumor-associated fibroblasts (CAFs) can promote cancer progression through several integrin-related mechanisms. Invasion is caused by deposition or regulation of fibroblast arrangement or by direct physical pulling of cancer cells from the primary tumor (76–79). In order to metastasize smoothly, tumor cells must attach to vasculature in distant organs and penetrate into perivascular tissues. Thrombosis is thought to support cancer metastasis through the recruitment of fibronectin to activate integrins. After extravasation, the contact of integrin with the extracellular matrix in perivascular tissue could determine whether the inoculated tumor cell would continue to proliferate or become dormant state (80–82). Integrin trafficking is also crucial for collective cell migration or morphogenetic movements of cell sheets. Rab-coupling protein (RCP)-dependent integrin recycling pathway was employed by invasive cancer cells for effective migration (83, 84).

EFFECTS OF MULTIPLE INTEGRIN SIGNALS ON TUMOR MICROENVIRONMENT

Generally, tissue has a strictly regulated, specific optimum hardness (85), which is perceived by cells through integrins and their cytoskeletons. Hence, integrins are important mechanical receptors, and together with other adherent proteins such as integrin-activated proteins, talin, nucin and CRK-related substrates, convert mechanical signals into biochemical signals (86, 87). Several studies have discussed the role of integrin in angiogenesis, especially the integrin αv. Evidence suggests that integrin αv promotes tumor angiogenesis, depending on environments (88). Integrin αvβ4 may also exert a similar environment-dependent pro-angiogenesis effect (89). In contrast, integrin α3β1 signaling in endothelial cells negatively regulates tumor angiogenesis by decreasing VEGFR2 expression (90). Signals from integrins also influence other behaviors in the tumor microenvironment. Studies show that TNFα pro-apoptotic signaling is regulated by the ECM and the integrin that is engaged, and Integrin α6β1 is inhibitory for the pro-apoptotic signal of TNF (91).

Integrins play bidirectional regulatory roles between cancer cells and cancer-associated fibroblasts (CAFs). CAFs that express IL-32 contain an RGD cell attachment sequence that binds to integrin B3-positive cancer cells to promote breast cancer cell invasion and metastasis (92). CAF-derived extracellular vesicles that express annexin A6 plays a pivotal role in gastric cancer drug resistance via activation of β1 integrin-FAK-YAP signaling (93). Colorectal cancer cells express integrin αvβ6 activated CAFs through TGF-β, which subsequently secrete stromal cell-derived factor-1 (SDF-1) and promote colorectal cancer cell metastasis (94). These research studies reveal that integrins act as receptors that regulate the interactions between CAFs and cancer cells in tumor progression and drug resistance. Studies in the future may reveal more about the integrin signaling mechanisms involved about remodeling the tumor microenvironment.
microenvironment during tumor development. Factors secreted by cancer cells profoundly alter the biology and composition of the stroma by inducing immune cells, triggering angiogenesis, and inducing the activation of CAFs, which generates a lot of tumor-promoting signals (76).

**CLINICAL APPLICATION OF INTEGRIN**

Integrins have been seen as potential therapeutic targets since they were discovered to promote pathogenic processes. The inhibition of integrins has led to several marketed drugs, and many others are being investigated preclinically in both academic and industry settings. Since 2015, there have been at least 130 clinical trials of integrin-targeted therapies (95). Unfortunately, there are still a few unsuccessful inhibitors (Table 1). Efalizumab, which targeted αL integrins, was withdrawn from the market because of multiple cases of progressive multifocal leukoencephalopathy (PML), said to be integrins, was withdrawn from the market because of multiple cases of progressive multifocal leukoencephalopathy (PML), said to be involved with inhibition of α4-containing integrins and αLβ2 (96).

Previous studies have found that α4 and β2 integrins are receptors mediating the neutrophil adhesion to the endothelium. Researchers evaluated the α4 and β2 integrins’ expression and functions in human primary neutrophils obtained from patients having chronic non-healing wounds and undergoing a prolonged hyperbaric oxygen therapy (150 kPa per 90 minutes). Cell adhesion function of both neutrophilic integrins α4β1 and β2 was significantly reduced, which could be of great importance for the design of novel therapeutic protocols focused on anti-inflammatory agents (97). Integrin αVβ3 is highly expressed on activated endothelial cells of tumor neovasculature and thus is key to tumor angiogenesis. RGD-binding integrins, mainly the αv integrin subfamily and important to the whole integrin family, are introduced about their expression in different human cancers and their pre-clinical antagonists. (Table 2) New molecules that target αv-containing integrins are now entering clinical trials for fibrotic diseases, including idiopathic pulmonary fibrosis (IPF) and nonalcoholic steatohepatitis (NASH), which have high and increasingly unmet medical need (95, 98, 99).

Integrins can also be used in diagnostic imaging. Integrin-inhibiting peptide Apticitide (TC-99M-P280), a gpIIbIIIa imaging technique for the diagnosis of acute deep venous thrombosis, is now available. [99mTc]3PRGD2 imaging is valuable for the diagnosis and staging of esophageal cancer. It may be less sensitive than [18F]FDG imaging for detecting metastatic lesions in small lymph nodes. The T/B value was correlated with the expression of integrin αVβ3 (100). Integrin αVβ3 in imaging is in the PH2 trial phase. It is reported that other imaging agents are in the early stage of development (101, 102). As a PET tracer 18F-Alfatide II has been recently proven to possess good diagnostic value in distinguishing between breast cancer and benign breast lesions (103). Nell et. al found that Ga-68-Trivehexin is a promising probe for imaging of αVβ6-integrin expression in human cancers because of its high expression density at the boundary of tumor and healthy tissue (104). Recent studies also show that it may be possible to develop next-generation nanomedicine based on the combined derivatives of resveratrol and tetrac targeting the Integrin αvβ3 (105).

**CONCLUSION**

Integrins have attracted much attention in recent years and are closely related to the development of cancers. We discussed much about the significance of integrin in cell migration and cell adhesion, which are important processes in tumor growth. Integrin-mediated cancer signals are also initiated by several integrin-binding proteins, which include talins, kindlins, MMPs, osteopontin, actinin and so on. Integrins interact with the actin cytoskeleton through these signaling molecules. And because of the polymerization and contraction generated by actin, the main signaling occurs while integrin activates. However, when integrin is misregulated, various mechanisms unfreeze the regulation of integrin signaling in cancer, enabling tumor cells to proliferate unrestrictedly and invade some tissue boundaries, allowing them to survive in microenvironments. The diversification of integrin and their roles in many diseases indicate the great potential of this superfamily as a drug target. Nowadays, designing drugs specific to integrin activation is possible as the structure of integrin has been recognized. By studying the mechanism of integrin and its related signaling pathways, we consider by regulating the expression of integrin or blocking the downstream signaling...

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**TABLE 1** | Integrin-targeting drugs once came out.

| Inhibitor Name | Target | Mechanism | Application | In Market |
|----------------|--------|-----------|-------------|-----------|
| Lifitagrast     | α4β2   | prevents  | Dry eye disease | 2016     |
|                 |        | lymphocyte adhesion |             |          |
| Vedolizumab     | α4β7   | inhibits binding | Ulcerative colitis | 2014     |
|                 |        | to MADCAM1 | and Crohn’s disease | | |
| Natalizumab     | Pan-α4 | inhibits ligand | Multiple sclerosis | 2004     |
|                 |        | binding to α4β7 and α4β1 | and Crohn’s disease | | |
| Efalizumab      | αL     | preventing | Plaque psoriasis | 2003 (withdrawn 2009) |
|                 |        | lymphocyte activation and migration | | |
| Tirofiban       | αIIbβ3 | inhibits binding to fibrinogen | Coronary syndrome and CVD | 1998     |
| Eptifibatide    | αIIbβ3 | inhibits binding to fibrinogen | Coronary syndrome and CVD | 1998     |
pathways of integrin to make its function. Although integrins have been discovered for more than 100 years, only a few of their inhibitors have been used in clinical applications, and no specific therapeutic inhibitors have been developed for cancer. Therefore, selectively blocking this acquired migration and invasion ability by targeting key metastatic molecules or regulatory proteins like integrin would be an attractive therapeutic strategy.

**AUTHOR CONTRIBUTIONS**

QL and TL reviewed the literature and drafted the article. JX, YL, DZ, and BS finalized the paper and provided suggestions to improve it. All authors participated in designing the concept of this manuscript. All authors contributed to the article and approved the submitted version.

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**TABLE 2 | αv-integrins expressed in different human cancers and their pre-clinical antagonists.**

| Integrin | Cancer Type | Main Expression Feature | Drug | Drug Targeted Cancer Type | Clinical Trial |
|----------|-------------|-------------------------|------|--------------------------|---------------|
| αvβ3     | Gastric cancer | Stroma and endothelia ↑, correlates with survival | Etaracizumab (Abegrin) | Colorectal/melanoma/prostate/ thyroid cancer | Phase II |
|          |             | Correlates with grade | Intetumumab (CNTO 95) | Colorectal/melanoma/prostate/ thyroid cancer | Phase II |
|          | Lung cancer brain metastasis | Endothelia ↑ tumor cells ↓ | Abciximab (c7E3) | Melanoma/breast cancer | Pre-clinical |
|          | Non-small cell lung cancer | Endothelia ↑ tumor cells ↓ | Vittaxin (MEDI-532) | Melanoma/breast cancer | Phase II |
|          | Oral cancer | Intratumoral endothelia ↑ | Cilengitide | Melanoma/breast cancer | Phase II |
|          | Pancreatic cancer | Involved in lymph node metastasis | | Lung/liver/stomach cancer | Phase I |
|          | Prostate cancer | Peritumor ↑ | | Melanoma/gastric/hepatic/breast carcinoma | Pre-clinical |
| αvβ5     | Gastric cancer | Tumor, stroma and endothelial cells↑ independent prognostic factor in intestinal-type | Intetumumab (CNTO 95) | Melanoma/Prostate cancer | Phase II |
|          | Lung cancer brain metastasis | Endothelia ↑ tumor cells ↓ | Cilengitide | Melanoma/breast cancer | Pre-clinical |
|          | Non-small cell lung cancer | Tumor and stroma cells ↑ no correlation with survival | | | |
|          | Prostate cancer | Tumor and stroma cells ↑ no correlation with survival | | | |
| α5β1     | Oral cancer | Stroma ↑ | Volociximab (M200) | Melanoma/prostate cancer | Phase II |
|          | Ovarian cancer | Correlates with survival | ATN-161 | Glioblastoma | Phase II |
| αvβ6     | Gastric cancer | Potential prognostic marker in early stage | Intetumumab (CNTO 95) | Prostate cancer/melanoma | Phase II |
| Basal cell carcinoma | Infiltrative subtype ↑ | | | | |
| Non-small cell lung cancer | Intratumoral | | | | |
|          | | | | | |

↑ means up-regulation; ↓ means down-regulation.
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