Methionine Enhances the Contractile Activity of Human Colon Circular Smooth Muscle In Vitro

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INTRODUCTION

Chronic constipation is a major health problem. It is chronic in nature and dramatically affects the patient’s quality of life, causing tremendous impact on both the individual patient and society as a whole (1). To cope with this problem, many kinds of prokinetic drugs with different mechanisms have been used. However, the effectiveness of the available drugs has been limited. Cholinergic agents such as bethanechol and neostigmine have a limited role in the management of constipation because they have broad muscarinic effects and need continuous monitoring of electrocardiography, blood pressure and oxygen saturation during medication use (2). Dopamine receptor antagonists such as metoclopramide (3, 4), and domperidone (5-7) have not shown clinically significant effects on large bowel motility (8). Tegaserod (9, 10) and cisapride (11), which are serotonin receptor modulators (12), emerged as the most promising drugs for chronic constipation. However, production of these drugs has been suspended due to their severe cardiac side effects (4, 13, 14). Successive results have shown that development of new effective drugs to manage chronic constipation has been unsatisfactory.

We have previously demonstrated that the sulfur-containing amino acid methionine acts as a specific blocker of the stretch-dependent potassium channels and nitrergic responses in the mouse colon (15). As such, methionine has the effect of depolarizing the resting membrane potential of the murine colon muscles and enhancing spontaneous contractions. Based on this data, we sought to determine whether methionine has any effect on the human colon. Human colon tissues were obtained from the specimens of colon resection. Microelectrode recording was performed and contractile activity of muscle strips and the propagation of the contractions in the colon segment were measured. At 10 μM, methionine depolarized the resting membrane potential (RMP) of circular muscle (CM) cells. In the CM strip, methionine increased the amplitude and area under the curve (AUC) of contractions. In the whole segment of colon, methionine increased the amplitude and AUC of the high amplitude contractions in the CM. These effects on contraction were maximal at 10 μM and were not observed in longitudinal muscles in both the strip and the colon segment. Methionine reversed the effects of pretreatment with sodium nitroprusside, tetrodotoxin and Nω-oxide-L-arginine, resulting in depolarization of the RMP, and increased amplitude and AUC of contractions in the muscle strip. Methionine treatment affected the wave pattern of the colon segment by evoking small sized amplitude contractions superimposed on preexisting wave patterns. Our results indicate that a compound mimicking methionine may provide prokinetic functions in the human colon.

Key Words: Colon; Methionine; Gastrointestinal Motility; Humans

MATERIALS AND METHODS

Preparation of tissues and protocol for recordings

Human colon tissues were obtained from the specimens of patients undergoing elective colon resections for non-obstructive neoplasms. The tissues were immediately stored in oxygenated Krebs-Ringer’sbicarbonate (KRB) solution.

For intracellular recording of circular smooth muscle, the tissues were transferred to a Petri dish coated with Sylgard (Dow Corning Co., Midland, MI, USA) and first pinned downed with the mucosa side facing upward. The mucosal layer was removed by sharp dissection leaving the submucosal layer intact. For the final electrophysiologic recording of the inner circular muscle,
the muscles were pinned with the submucosa facing down in the electrophysiologic chamber and longitudinal and outer circular muscles were finely removed by sharp dissection. For the intracellular recording from the longitudinal smooth muscle, thin strips (1.5 mm thick and 10 mm long) were cut parallel to the longitudinal muscle with a double-bladed scalpel, and pinned down to expose a cross section of the entire muscle layer. The electrophysiologic chamber was constantly perfused with pre-warmed, preoxygenated Krebs solution, maintaining the temperature at 37.5 ± 0.5°C. The muscles were equilibrated for at least 2 hr before experiments. A conventional microelectrode recording was performed using a sharp microelectrode filled with 3 M/L KCl. Membrane potentials were measured with a high-input resistance electrometer, and outputs were displayed on oscilloscope. Agar bridge reference electrode (Warner Instruments, Harvard Apparatus company, Hamden, CT, USA) was used because the sulfur compound in methionine reacts with Ag. Resting membrane potentials were measured using pClamp software (version 9.0. Axon Instruments, Foster City, CA, USA) and Origin software (MicroCal Software, Northampton, MA, USA) programs. Resting membrane potential (mV), amplitude (mV) and frequency (/min) of the slow waves were analyzed.

For preparation of the muscle strip, mucosa and submucosal layers were removed and longitudinal and circular muscle bundles were obtained by sharp dissection. The size of the muscle strips were 2 mm in width and 1 cm in length. For contractile activity recording, the muscle strips were attached with a suture to a force transducer in an organ bath and a resting force of 1 g was applied. The mechanical signals were digitized and recorded on Acknowledge software (Biopac Systems Inc., Goleta, CA, USA) for data analysis. Frequency (/min), amplitude (mN), and area under the curve (AUC, sec × mN/min) of contractions were analyzed. In the muscle strip recording, the area under the curve was defined as the integrated area under the waves occurring in one minute (AUC for one minute).

The colonic segment with the mucosa and submucosa intact was prepared by cutting the whole layer of the colon segment parallel to the longitudinal muscle. The size of the colon segment was 4 cm in length and 2 cm in width and included the taenia coli. A stainless steel rod was placed parallel to the longitudinal muscle and placed at the organ bath. To record the propagation pattern within the colonic segment, circular smooth muscle (CM) tension was recorded at three (oral, middle, aboral) sites, and longitudinal smooth muscle (LM) tension was recorded by perpendicular traction. The contractile activity was measured as described above. In colonic segment recordings, the area under the curve was defined as the integrated area under a single wave (AUC for one wave).

**Drugs and solutions**
The KRB used in this study contained (in mM/L) 120.4 NaCl, 5.9 KCl, 15.5 NaHCO3, 11.5 glucose, 1.2 MgCl2, 1.2 NaH2PO4, and 2.5 CaCl2. This solution had a pH of 7.3-7.4 at 37.5°C when bubbled to equilibrium with 97% O2-3% CO2.

L-methionine, sodium nitroprusside (SNP), tetrodotoxin (TTX) and Nω-oxide-L-arginine (L-NA) were obtained from Sigma Chemical Company (St Louis, MO, USA).

**Statistical analysis**
All data are expressed as mean ± standard deviation. Student’s paired t-tests were used to determine if data sets differed, and P values of less than 0.05 were taken to indicate significant differences between sets of observations. The n values reported in the text refer to the number of recordings from the muscle strips, which is equivalent to the number of patients used unless otherwise stated.

**Ethics statement**
Obtainment and the use of the human colon specimen were approved by the institutional review board of the Seoul National University Hospital (IRB approval number: H-0603-071-170). Informed consent was waived by the board.

**RESULTS**

**Intracellular recording of membrane potential**
Electrical activity in the normal human colonic circular smooth muscle

**Fig. 1.** Effect of methionine on the membrane potential of human colon circular muscle. (A) Methionine at 10 µM concentration caused depolarization of the resting membrane potential. (B) Various concentrations of methionine (5, 10, 100, 500 µM) were used to treat circular muscle. 10 µM of methionine treatment resulted in maximal depolarization in the resting membrane potential of circular muscle. The figure shows the resting membrane potential according to the concentration of methionine.
muscle (CM) is characterized by slow waves originating from interstitial cells of Cajal (ICC) in the submucosal layer (Fig. 1).

First, circular smooth muscle (CM) was treated with up scaling concentrations of methionine at 5, 10, 100, and 500 μM to 3, 12, 6, and 3 tissues, respectively. Methionine depolarized the resting membrane potential (RMP) of CM, and this was most significant when treated with 10 μM methionine (control -50.5 ± 6.4 vs 10 μM methionine -48.9 ± 7.0 mV; P = 0.018; n = 12) (Fig. 1). 10 μM methionine did not have a significant effect on the amplitude (control 29.87 ± 6.37 mV vs methionine 27.64 ± 10.31 mV, P = 0.164) and frequency (control 9.09 ± 6.70/min vs methionine 11.3 ± 13.31/min, P = 0.427) of the slow waves in the CM.

The NO donor SNP (100 μM) induced hyperpolarization of CM from -51.00 ± 12.29 mV to -55.45 ± 13.42 mV and subsequent application of 10 μM methionine induced depolarization to -48.43 ± 16.59 mV. Methionine seems to have a tendency (P = 0.064, n = 6) to reverse the effect of SNP (Fig. 2A) in CM.

TTX (1 μM) depolarized the membrane potential of CM from -62.10 ± 8.83 mV to -56.48 ± 8.75 mV and subsequent addition of 10 μM methionine caused significant (P = 0.033, n = 7) additional depolarization to -51.81 ± 18.05 mV (Fig. 2B).

Most GI muscles have tonic inhibitory drive due to the spontaneous production and release of NO. 100 μM L-NA, an inhibitor of nitric oxide synthase, caused depolarization of CM from -66.80 ± 11.89 mV to -65.36 ± 13.00 mV, suggesting a substantial basal production of NO in human colonic muscles. In the presence of L-NA, treatment with 10 μM methionine caused small but significant (P = 0.038, n = 6) additional depolarization to -64.65 ± 11.82 mV (Fig. 2C).

Fig. 2. Effect of methionine on the membrane potential of human colon circular muscle in the presence of pretreated drugs. Pretreatment drug was perfused at 15 min intervals to 10 μM methionine. (A) Pretreating with 100 μM SNP hyperpolarized the RMP; 10 μM of methionine depolarized the RMP and overcame the effect of SNP. (B) Pretreating with 1 μM TTX depolarized the RMP; 10 μM methionine depolarized the RMP and had additional effect over TTX. (C) Pretreating with 100 μM NOLA depolarized the RMP; 10 μM methionine depolarized the RMP and had additional effect over NOLA. *The horizontal line is the reference to compare the RMP after drug treatment.

Fig. 3. Effect of methionine on the contraction of human colon CM strip. (A) After 10 μM methionine treatment on CM, amplitude and AUC for one minute increased. The black vertical line indicates the starting point of 10 μM methionine perfusion. (B) First KRB solution, then 1 μM TTX and 10 μM methionine were perfused at 45 min time intervals. TTX had a neurogenic blocking effect and inhibited the contraction in the CM strip. Note that after perfusing 10 μM methionine in the presence of TTX, the amplitude and area under the curve for one minute increased, indicating that 10 μM of methionine treatment had overcome the blocking effect of TTX and enhanced the contraction of the CM strip. The data in the open circle is magnified at the second row. (C) First KRB solution, then 100 μM NOLA and 10 μM methionine was perfused at 45 min time intervals. NOLA enhanced the contraction in the CM strip. Note that after perfusing 10 μM methionine, the amplitude and AUC increased further. 10 μM of methionine treatment had additional effect over NOLA.
Contraction Effect of Methionine on Colon Smooth Muscle

Muscle strip tension recording

In the circular muscle (CM) strip, 10 μM methionine increased the amplitude (control 17.2 ± 13.2 vs methionine 27.8 ± 11.2 mN; P = 0.04; n = 6) and the area under the curve (AUC, control 266.4 ± 159.3 vs methionine 420.3 ± 151.5 mN X sec/min; P = 0.006; n = 6) of contractions. The frequency (control 2.24 ± 1.39/min vs methionine 2.48 ± 1.85/min; P = 0.342; n = 6) of contractions increased but it was not significant (Fig. 3A).

10 μM methionine was applied to 7 longitudinal smooth muscle strips, but had no significant effect on frequency (control 1.30 ± 0.99 vs methionine 1.05 ± 0.97/min; P = 0.556), amplitude (22.74 ± 10.97 vs methionine 19.76 ± 10.34 mN; P = 0.163) and AUC (394.14 ± 235.66 vs 278.45 ± 167.35 mN X sec/min; P = 0.117).

In CM, 1 μM TTX attenuated the contractile activity and subsequent addition of 10 μM methionine reversed this effect and enhanced the contraction. 10 μM methionine overcame the neurogenic block effect of 1 μM TTX and increased the frequency (control 9.59 ± 5.44, TTX 3.65 ± 3.16, TTX + methionine 11.08 ± 5.84 mN; P = 0.011; n = 6) and AUC (control 84.45 ± 25.48, TTX 21.40 ± 16.73, TTX + methionine 99.66 ± 5.89 mN X sec/min; P = 0.011), significantly (Fig. 3B).

Whole colon segment tension recording

In the contractile activity of the whole colonic segment, two different wave patterns were mixed in CM with variable ratios. We defined the one with higher amplitude as the dominant wave (DW) which was observed both in the CM and LM layer, and smaller amplitude waves were defined as non-dominant (non-DW) waves observed only in the CM layer.

First, the whole colonic segment was treated with upscaling concentrations of 5, 10, 100, and 500 μM methionine to 3, 9, 6, and 3 tissues, respectively. The methionine effect was maximal at the 10 μM concentration and increased the amplitude (control 18.61 ± 9.81 vs 10 μM methionine 22.92 ± 10.52 mN; P = 0.027; n = 9) and AUC in the DW (control 665.08 ± 87.23 vs 10 μM methionine 805.78 ± 649.69 mN × sec/wave; P = 0.026) of the CM (Fig. 4).

10 μM methionine did not have a significant effect on the frequency, amplitude and AUC in the DW of the longitudinal smooth muscle (P = 0.427, 0.696, 0.915, respectively).

Methionine treatment also had an effect on the wave patterns of the colon segment by evoking small sized amplitude contractions (non-DWs), superimposed on preexisting wave patterns (DW). In 3 out of 11 samples, methionine treatment evoked non-DWs which were not present in the control state (Fig. 5). Significance of this finding is not clear at the moment, but since non-DWs are presumed to have a retrograde propagation function (our unpublished data), methionine may also enhance mixing of colonic contents.

DISCUSSION

Prokinetic agents are drugs that enhance motor activity of the
smooth muscle in the gastrointestinal tract. These drugs have been well used in upper gastrointestinal tract motility disorders such as gastroesophageal reflux disease, diabetic gastropathy, irritable bowel syndrome and bile acid reflux gastritis (11, 16-18). A clear mechanism of how these drugs work is not well known, but it is believed that these drugs enhance the motility agonists or antagonize inhibitory transmitters and thus improve motility (19, 20).

On the contrary, agents that could be used in colonic motility disorders have not been sufficiently investigated. Lower GI motility disorders that seem to have benefit from prokinetics are postoperative ileus, acute colonic pseudoobstruction, chronic intestinal pseudoobstruction, idiopathic constipation, and constipation secondary to a primary disease (such as dementia, multiple sclerosis, diabetic neuropathy, spinal cord injury, Hirschsprung’s disease, Chagas’ disease, myopathic intestinal pseudoobstruction) (21). Prokinetic agents used clinically are cholinergic agents, dopamine, receptor antagonists, and serotonin receptor modulators. Since cholinergic agents such as acetylcholine act on muscarinic cholinergic receptors, they also cause non-specific and broad muscarinic effects (2). Domperidone and metoclopramide, known dopamine D2 receptor antagonists, act most effectively on the upper GI tract. The only agents that are effective on colonic motility are cisapride (11) and tegaserod maleate (9, 10) which are serotonin receptor modulators (12). Cisapride is an antagonist to 5-HT (hydroxytryptamine) and acts as a 5-HT3 agonist to powerfully enhance GI motility. However, this agent caused serious cardiac arrhythmias by acting on the HERG channel and was suspended from the market. Tegaserod maleate is a non-specific 5-HT4 receptor agonist, and has been widely used in Europe for managing irritable bowel syndrome and bile acid reflux gastritis (11, 16-18). A clear mechanism of how these drugs work is not well known, but it is believed that these drugs enhance the motility agonists or antagonize inhibitory transmitters and thus improve motility (19, 20).

The sulfur containing amino acid methionine is an essential amino acid, which has been known to enhance the contractile activity in coronary smooth muscle by increasing the N-methylation of phospholipids which comprise the cell membrane and control the intracellular calcium reaction (24-26). However, its effect in human gastrointestinal smooth muscle has not been studied in detail. In this paper, we sought to examine the effects of methionine, which has proved to have a prokinetic effect in murine colon. We have shown that methionine depolarized the resting membrane potential of the human colon CM in the electrophysiologic study. In contractile studies, methionine increased the amplitude and area of contractions in the circular muscle strip. The increased contractile effect of methionine was also evident in the whole colonic segment by increased amplitude and area of contractions in the circular muscle layer. The enhanced effect on the contractile activity was observed only in circular smooth muscle but not in the longitudinal smooth muscle.

In the normal state, gastrointestinal smooth muscle cells are kept in the inhibitory state through various inhibitory neuronal inputs, of which NO is the most prominent inhibitory input. NO is released by non-adrenergic, non-cholinergic (NANC) inhibitory neurons in gastrointestinal (GI) smooth muscles and serves as the primary enteric inhibitory neurotransmitter in GI muscles, and nitricergic neurons regulate gut tone, phasic contractile amplitude and frequency, and inhibitory reflexes (27, 28). NO induces hyperpolarization of membrane potential which results in reduced smooth muscle excitability. A recent study in mice suggests that part of the hyperpolarizing effects of NO may be mediated by stretch-dependent potassium (SDK) channels that are expressed in GI smooth muscles (29). In the mouse, the two-pore K+ channel, TREK-1, encodes SDK channels in murine GI smooth muscle cells, especially in the colon (29, 30). It has been noted that sulfur-containing amino acids block SDK channels in colonic myocytes, and that compounds of this class are effective in blocking NO dependent responses in intact muscles. In murine mice, methionine blocked SDK channels in colonic myocytes, depolarized membrane potentials and increased the frequency of contractions in intact muscles. These compounds also inhibited responses to the NO donor sodium nitroprusside (SNP) and neurally released NO (15).

In this paper, we tried to determine whether a similar phenomenon occurs in human colonic muscles. We pretreated with
TTX (a sodium channel blocker which blocks all neurogenic control on gastrointestinal smooth muscle), L-NA (NO synthase inhibitor which enhances gastrointestinal smooth muscle contractility) and SNP (a spontaneous NO releasing compound which hyperpolarizes the resting membrane potential). Our results from this study indicated that methionine had the effect of overcoming the spontaneous neuronal effect and release of NO and reversed the effect of nitricergic stimulation in the human colonic muscle strip. In the whole colon segment, L-NA enhanced the contractions which were potentiated with the addition of methionine. However, effects of methionine after pretreatment with SNP and TTX in whole colon segments were variable (data not shown). This could be due to complex interactions of the intact intrinsic nervous system in the whole colon segment.

The sulfur containing amino acid methionine seems to be a compound that could be a candidate for prokinetic drugs in the human colon. However, methionine is transformed into homocysteine during bio-metabolism. Surplus accumulation of homocysteine in the human body induces atherosclerosis, and increases the risk of Alzheimer’s disease and coronary artery disease (31-33). For this reason, methionine cannot be directly used to enhance colonic motility in humans. But based on our findings, development of a new compound that mimics methionine, devoid of cardiac side effects, may have a role as an effective prokinetic agent for the human colon.

It is concluded that methionine enhances the basal contractile activity of the CM in both the muscle strip and the segment of the human colon. Methionine also has the effect of overcoming the spontaneous neuronal effect and release of NO and reverses the effect of nitricergic stimulation in the muscle strip. A compound mimicking methionine may provide prokinetic functions in the human colon.

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