Identification of key immune-related genes and immune infiltration in atrial fibrillation with valvular heart disease based on bioinformatics analysis

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Background: Atrial fibrillation (AF) is the most common persistent arrhythmia. Valvular heart disease (VHD) and AF frequently coexist. In our study, from performing bioinformatics analysis, we sought to identify immune-related genes (IRGs) and explore the role of immune cell infiltration in AF-VHD in depth, aiming at investigating the potential molecular mechanism and developing new therapeutic targets for AF, including AF-VHD.

Methods: The gene expression of the GSE41177 and GSE79768 datasets were downloaded from the Gene Expression Omnibus database. Differentially expressed genes (DEGs) were analyzed via the limma package in Bioconductor with R software. Differentially expressed immune-related genes (DEIRGs) were selected via combination ImmPort database with DEGs, and the enrichment function and pathway analysis were explored. A protein-protein interaction (PPI) network was built with a Search Tool for the Retrieval of Interacting Genes/Proteins plugin in Cytoscape. The CIBERSORT algorithm was used to evaluate immune infiltration in the left atrial (LA) tissues between AF-VHD and sinus rhythm (SR) patients. Finally, a correlation analysis between key DEIRGs and infiltrating immune cells was performed.

Results: A total of 130 DEIRGs were detected. Enrichment function of DEIRGs demonstrated that they are significant in immune and inflammatory responses. The key DEIRGs assessed by the PPI network and involved in both the immune and inflammatory responses were the C-X-C motif chemokine ligand (CXCL) 1, pro-platelet basic protein (PPBP), CXCL12, and C-C motif chemokine ligand 4 (CCL4). The immune infiltration findings indicated that, compared with the LA tissues from SR patients, the tissues from AF-VHD patients contained a higher proportion of gamma delta T cells, but a lower proportion of CD8 and regulatory T cells. The results of correlation analysis demonstrated that CXCL1 was positively correlated with activated mast cells and significantly negatively correlated with resting mast cells. PPBP, CXCL12, and CCL4 were positively correlated with the infiltration of various immune cells, such as neutrophils, plasma cells, and resting dendritic cells.

Conclusions: The key immune-related genes and the differences in immune infiltration in LA tissues play an essential role in the occurrence and progression of AF-VHD.

Keywords: Atrial fibrillation (AF); valvular heart disease (VHD); immune-related genes; immune cells; Gene Expression Omnibus; CIBERSORT

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Introduction

Atrial fibrillation (AF) is the most common persistent arrhythmia (1). Patients who develop AF have an increased risk of heart failure, stroke, cardioembolic hospitalization, and mortality (1). Patients with valvular heart disease (VHD) frequently develop AF, because VHD can lead to the disruption of the atrial architecture due to increased left atrial (LA) pressure. It can also cause atrial dilation and alter wall stress (2). VHD patients, especially those with rheumatic mitral stenosis, have a significant risk of stroke and thromboembolism. On average, the rate of stroke among patients with AF is approximately 6 times that of individuals without AF. The relative risk is even higher in patients with both AF and mitral stenosis (about 15 times that of people without AF). However, the underlying mechanisms of AF-VHD are not well understood. Therefore, understanding the AF-VHD biology is vital for the diagnosis and treatment of this complicated disease.

Previous studies have shown that immune and inflammatory responses may be essential in the pathophysiological mechanism of AF, also in AF-VHD, one of the main types of AF (3,4). Several studies have found that inflammatory biomarkers, such as C-reactive protein, interleukin (IL)-6, IL-10, and tumor necrosis factor (TNF)-α, have been found to be related to AF and its outcome (5-8). And some researches indicated that the elevated levels of serum hs-CRP, IL-6, and sCD-40L were strongly associated with AF-VHD (4,9,10). In addition, several immune-related genes (IRGs), such as C-X-C motif chemokine receptor (CXCR) 2, may play an important functional role in atrial remodeling and AF inducibility. The results of the above studies suggest that IRGs may be a new therapeutic strategy for AF (11,12). The infiltration of immune cells has also been demonstrated to influence the development of AF. Hohmann et al. reported the significant increase of inflammatory CD3+ T-cell counts over the course of sinus rhythm (SR) development into paroxysmal AF and persistent AF in LA tissue samples from AF-VHD and SR patients (13). Furthermore, to better understand the underlying molecular immune mechanism of AF-VHD, we investigated the relationship between key IRGs and infiltrating immune cells. We present the following article in accordance with the MDAR reporting checklist (available at http://dx.doi.org/10.21037/jtd-21-168).

Methods

Patients and datasets

The GSE41177 and GSE79768 datasets were downloaded from the Gene Expression Omnibus (GEO) database (GPL570 platform, HG-U133_Plus_2, Affymetrix Human Genome U133 Plus 2.0 Array). Only human LA samples from AF-VHD patients and VHD patients with SR (as the control group) were selected in each dataset. The GSE41177 contained 23 samples, including 16 LA tissue samples from patients with chronic AF (AF-VHD group) and 3 from patients with SR. The GSE79768 contained 7 atrial tissue samples from AF-VHD patients and 6 atrial tissue samples from patients with SR.

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Identifying differentially expressed IRGs (DEIRGs)

Gene expression matrices were created with R software (version 3.6.3; https://www.r-project.org/), and the R package of “Bioconductor” (http://www.bioconductor.org/) was applied for data analyses. The SVA package (https://www.bioconductor.org/packages/release/bioc/html/sva.html) was used for the normalization of the GSE41177 and GSE79768 datasets (19). Using the limma package (https://
www.bioconductor.org/packages/release/bioc/html/limma.html), differentially expressed genes (DEGs) were screened. Only genes that satisfied the cut-off value of $P<0.05$ and fold change (FC) $\geq 1.5$ ($\log_{2} FC > 0.5$) were selected as DEGs. DEIRGs were identified by matching 1,793 IRGs to DEGs using the IRG dataset from the ImmPort database (https://www.immport.org/shared/genelists). With the “pheatmap” package in R software (https://bioconductor.org/packages/release/bioc/html/heatmaps.html), heatmaps for the DEGs and DEIRGs was created. Using the “ggplot2” package, a volcano map of DEIRGs was created to show the differential expression of DEIRGs.

**Functional and pathway enrichment analyses of DEIRGs**

Gene ontology (GO) enrichment and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway of DEIRGs was performed by using Database for Annotation, Visualization, and Integrated Discovery (https://david.ncifcrf.gov/) (20,21). $P<0.05$ illustrated statistical significance.

**Protein-protein interaction (PPI) network and potential crucial genes analyses**

The Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) plugin was applied to investigate the potential interaction between DEIRGs (confidence score: 0.9, maximum number of interactions: 0) and visualized the PPI network using Cytoscape software (version 3.8.1; https://www.cytoscape.org/) (22).

**Evaluation of immune cell infiltration**

Using CIBERSORT in R, we filtered out samples with $P<0.05$ and got an immune cell infiltration matrix with 22 types of immune cells. The proportions of immune cells were also obtained. The “ggplot2” package was conducted to carry out principal component analysis (PCA) and clustering analysis on the immune cell infiltration matrix data and created a 2D PCA map and violin diagrams to visualize the differences in immune cell infiltration. Spearman correlation analysis in R was applied to the 22 types of infiltrating immune cells, and the “Corrplot” package was conducted to create a correlation heatmap to visualize the result (23). The immune infiltration levels for each immune cell in the 2 groups were represented with violin diagrams using the “vioplot” package in R.

**Correlation analysis between key IRGs and infiltrating immune cells**

The Spearman correlation analysis in R was used to explore the potential relations between the key IRGs and infiltrating immune cells, which was visualized by applying the package of “ggpubr”.

**Results**

**DEIRG screening**

After batch correction and standardization of the merged GSE41177 and GSE79768 datasets, a total of 919 DEGs, comprising 88 upregulated and 831 downregulated genes, were obtained (Figure 1A). Next, by matching the IRGs to the DEGs, 130 DEIRGs were selected, including 7 upregulated and 123 downregulated genes (Figure 1B,C).

**GO analysis and pathway enrichment**

The results of the GO analysis showed that DEIRGs were mostly related to immune and inflammatory response under biological processes (BP), the plasma membrane and extracellular region under cellular components (CC), and receptor binding and receptor activity under molecular functions (MF) (Figure 2). The KEGG analysis results suggested that the DEIRGs were mostly enriched in cytokine-cytokine receptor interactions, antigen processing and presentation, rheumatoid arthritis (RA), the chemokine signaling pathway, and Leishmaniasis (Figure 3).

**PPI network analysis**

The PPI network of DEIRGs is shown in Figure 4. Degree (number of adjacent connected nodes) $\geq 15$ was set as the cut-off criterion for hub genes (Figure 4B). Angiotensinogen (AGT), major histocompatibility complex class IE (HLA-E), HLA class IB (HLA-B), angiotensin II receptor type 2 (AGTR2), C-X-C motif chemokine ligand (CXCL) 1, HLA class II DR alpha (HLA-DR), pro-platelet basic protein (PPBP), CXCL12, C-C motif chemokine ligand 4 (CCL4), HLA class II DP beta 1 (HLA-DPB1), interferon regulatory factor 9 (IRF9), HLA class II DQ alpha 1 (HLA-DQA1), HLA class II DP alpha 1 (HLA-DPA1), CXCR4, and 2’-5’-oligoadenylate synthetase 1 (OAS1) were identified as hub genes. The hub genes involved in both the immune response and inflammatory response in BP were selected as
Figure 1 Heatmap and volcano plot of DEG and DEIRG expressions for AF-VHD and sinus rhythm. (A) DEG expression heatmap for left atrial tissues (top 50 upregulated and downregulated genes). (B) Heatmap of DEIRG expression for left atrial tissues (all upregulated and downregulated genes). (C) Volcano plot of DEIRGs [the differences are set as P<0.05 and |log FC| >0.5]. DEG, differentially expressed gene; DEIRG, differentially expressed immune-related gene; AF-VHD, atrial fibrillation-valvular heart disease; FC, fold change.
key genes, specifically **CXCL1**, **PPBP**, **CXCL12**, and **CCL4**.

**Immune infiltration analyses**

We first investigated the difference between AF-VHD and SR LA tissues in immune infiltration with 22 subpopulations of immune cells by using the CIBERSORT algorithm. The proportions of immune cells obtained from the 9 SR patients and 22 AF-VHD patients are summarized in *Figure 5*. Using PCA, the immune cells in the LA tissues of AF-VHD patients and SR revealed distinct group-bias clustering and individual differences (*Figure 6A*). The correlation heatmap showed that resting CD4 memory T cells and CD8 T cells, M2 macrophages, gamma delta T cells, activated mast cells, and resting mast cells all had a significant negative correlation. In contrast, naive CD4 T cells and resting natural killer cells had a significant positive correlation (correlation coefficients ≥0.5 or ≤−0.5 with P<0.05 were supposed to be the statistically significant positive or negative correlations) (*Figure 6B*). Compared with SR, LA tissues from AF-VHD patients generally contained a lower proportion of CD8 T cells and regulatory T cells (Tregs), whereas the proportions of gamma delta T cells were relatively higher (P<0.05) (*Figure 6C*).

**Correlation analysis between hub genes and infiltrating immune cells**

The results of the correlation analysis revealed that **CXCL1** was positively correlated with activated mast cells (r=0.441, P=0.013) and significantly negatively correlated with resting mast cells (r=−0.638, P=0.0001). **PPBP** was correlated significantly positively with neutrophils (r=0.603, P=0.0004). **CXCL12** was positively correlated with plasma cells (r=0.476, P=0.007). **CCL4** was positively correlated with plasma cells (r=0.476, P=0.007), neutrophils (r=0.483, P=0.007), and resting dendritic cells (r=0.494, P=0.005), and correlated negatively with CD8 T cells (r=0.483, P=0.019) (*Figure 7*).

**Discussion**

AF is one of the most prevalent arrhythmias (24). VHD and AF frequently coexist. When associated with VHD, the incidence of thromboembolism in AF significantly
increases (25). Heart failure and stroke caused by thromboembolism are complications of AF that result in a high rate of disability and mortality, making AF a big challenge for public health (26). Treatments for AF in symptomatic patients include oral anticoagulants and LAA closure for stroke prevention, drugs for rate control, and anti-arrhythmic drug therapy and catheter ablation for conversion and the prevention of recurrence (2,27). However, the long-term efficacy and safety of these treatment regimens are still not well understood. The pathogenesis and biomarkers of AF still require further investigation. With better understanding of the underlying mechanisms of AF and AF-VHD, AF-VHD patients will receive better management and therapy. In recent years, increasing evidence has revealed that the pathogenesis of AF may be related to immune and inflammatory responses (28-31). Therefore, we sought to identify IRGs and explore the role of immune cell infiltration in AF-VHD in depth. A total of 130 DEIRGs were detected as candidate biomarkers in the pathogenesis of AF-VHD, and the enrichment function identified the underlying mechanisms of DEIRGs. The results of the GO enrichment analysis indicated that DEIRGs were significantly correlated with immune and inflammatory responses. Further, these genes were involved in cytokine-cytokine receptor interactions, the chemokine signaling pathway, tuberculosis, influenza A, RA, and other pathways via the KEGG analysis. Higher levels of cytokines, such as IL-6, IL-8, and TNF-α, have been reported in patients with AF compared with the levels in SR patients, and these inflammatory markers can predict the outcome of AF ablation (8,32-36). Similar with AF, the expression of some cytokines, such as IL-1β, TNF-α, TGF-β1 have increased in AF-VHD (37,38). Among them, the increase of TGFβ1 can promote the occurrence of atrial fibrosis, which is a key role in the mechanism of the occurrence and maintenance in atrial fibrillation (38-41). Recently, a Danish cohort study found that, compared with the risk among the general population, there is a 40% increase in the risk of AF in patients with RA (42). These findings indicate that DEIRGs in AF-VHD are involved in the immune and inflammatory processes for the disease.

The PPI was further analyzed by the STRING plugin in Cytoscape 3.8.1, and 15 hub genes (AGT, HLA-E, HLA-B, AGTR2, CXCL1, HLA-DRA, PPBP, CXCL12, CCL4, HLA-DPB1, IRF9, HLA-DQA1, HLA-DP41, CXCR4, and OASI) were selected in the present study. Of these, CXCL1, PPBP, CXCL12, and CCL4 were judged to be the core genes for the immune and inflammatory responses, according to the...
Figure 4 PPI network of DEIRGs in AF-VHD and sinus rhythm patients. (A) By using Cytoscape, the PPI network for DEIRGs was constructed. (B) Degree (number of adjacent connected nodes) ≥15 was set as the cut-off criterion for hub genes and of adjacent connected nodes for the hub genes, as shown. PPI, protein-protein interaction; DEIRGs, differentially expressed immune-related genes; AF-VHD, atrial fibrillation-valvular heart disease; AGT, Angiotensinogen. HLA, human leukocyte antigen; CXCL, C-X-C motif chemokine ligand; PPBP, pro-platelet basic protein; CCL, C-C motif chemokine ligand; IRF, Interferon regulatory factor; CXCR, C-X-C motif chemokine receptor; OAS, ligoadenylate synthetase.
results of GO analysis.

CXCL1, a member of the CXC subfamily of chemokines, is a secreted growth factor that signals through the G-protein coupled receptor CXCR2 and modulates inflammation, angiogenesis, wound healing, tumorigenesis, and cell motility by regulating CXCL1–CXCL2 (43). Recent studies have indicated that CXCL1–CXCR2 signaling may play a crucial role in regulating monocyte infiltration and in the pathophysiology of hypertension and cardiac remodeling (44-46). It was also reported that the CXCL1/CXCL2 single axis may participate in AF induced by a high dose of angiotensin II or prolonged hypertension (47). PPBP (also known as CXCL7), a platelet-derived growth factor, belongs to the CXC chemokine family. It is released from the activated platelets and participates in the response to vascular injury (48). It can also stimulate a variety of processes, such as mitogenesis, glucose metabolism, and the synthesis of the extracellular matrix and plasminogen activator (49,50). Maneerat et al. identified PPBP as one of the significant biomarkers correlated with the development of coronary heart disease (CHD) (51).

CXCL12 (also known as SDF-1) belongs to the CXC family, a subclass of the chemokine family, and regulates cellular activity by binding to CXCR4 or CXCR7, which are receptor proteins located in the cytomembrane, contributing to the activation of various intracellular signaling pathways (52-56). Previously published studies have suggested that there may be associations between CXCL12 and cardiovascular disease, leading to the elevation of serum CXCL12 levels, which in turn increases the risk of coronary artery disease (57-59). CCL4 (also known as macrophage inflammatory protein (MIP)-1beta, MIP-1/β) is a crucial chemotactic mediator for the recruitment of monocytes/macrophages (60). It is associated with the pathogenesis of several diseases, including psoriasis vulgaris, sarcoidosis, cystic fibrosis, and multiple sclerosis (61-64). In obese patients, as well as in patients with a history of cardiac surgery, CCL4 levels have been found to be increased, which may be associated with new-onset AF (65,66). Moreover, it is also considered to play an important role in metabolic inflammation (67). Further research is required to explore the involvement of these 4 key IRGs in the mechanism of AF.

To further explore the role of immune cell infiltration in AF-VHD, CIBERSORT was used to evaluate AF-VHD immune infiltration. In this program, we found reduced infiltration of CD8 T cells and Tregs, as well as increased infiltration of gamma delta T cells, which may be related

Figure 5 From GSE41177 and GSE49468 datasets, the relative percentage of 22 subpopulations of immune cells in 31 left atrial samples are shown. NK, natural killer.
Figure 6 Visualization and evaluation of immune cell infiltration. (A) Principal component analysis cluster plot of immune cell infiltration in AF-VHD and SR tissues. (B) Correlation heat map for immune cells. (C) Differences in immune infiltration between AF-VHD and SR. SR group is marked in blue and the AF-VHD group is marked in red. P<0.05 was supposed to be statistically significant. AF-VHD, atrial fibrillation-valvular heart disease; SR, sinus rhythm.
to the pathogenesis of AF-VHD. Several studies have shown that only a few CD8+ T cells are present in the left appendages of AF patients (14). Chen et al. reported that, in peripheral whole blood and right atrial tissue of AF patients, the proportion of Treg cells was significantly lower (68). Nevertheless, to the best of our knowledge, there has been no research on the role of gamma delta T cells in AF-VHD and further experimental data are needed.

We further investigated the association between CXCL1, PPBP, CXCL12, CCL4, and immune infiltration. We found that CCL4 was positively correlated with plasma cells, neutrophils, and resting dendritic cells, and negatively correlated with CD8 T cells. CXCL1 was positively correlated with activated mast cells and significantly negatively correlated with resting mast cells. CXCL12 was positively correlated with plasma cells, and PPBP was significantly positively correlated with neutrophils. These findings require further researches to elucidate the underlying complicated interactions between IRGs and immune cells. Based on these results, CXCL1, PPBP, CXCL12, and CCL4 appear to play key roles in AF-VHD via the regulation of immune infiltration.

Conclusions

In summary, we found that the IRGs CXCL1, PPBP,
CXCL12, and CCL4 may be key biomarkers of AF-VHD. We also found that some immune cells may contribute to the occurrence and progression of AF-VHD. Moreover, the correlations between CXCL1, PPBP, CXCL12, CCL4, and immune cells may play an important role in the pathogenesis of AF-VHD. Further researches of these IRGs and immune cells are required to study if they may be a novel agent to the molecular targeted therapies for AF-VHD patients.

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Footnote

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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/jtd-21-168). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). GEO belongs to public databases. The patients involved in the database have obtained ethical approval. Users can download relevant data for free for researches and publish relevant articles. Our study is based on this open source data, so there are no ethical issues and other conflicts of interest.

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