An overview of the solid state fermentation in the production of fungal protease enzymes

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Abstract

Enzymes are among the most important products obtained for human needs through microbial sources. A large number of industrial processes in the area of industrial, environmental and food technology utilize enzymes at some stage or another; Solid State Fermentation (SSF) holds tremendous potential for the production of enzymes, especially in those processes where the crude fermented product may be used directly as the enzyme source. Fungal proteases are used in many industrial processes for the production of foods and metabolites, production of enzymes from fungi offered many advantages which include low cost and high productivity. Hence because of the higher yielding capacity of SSF and the demand for proteases it highly imperative to search for novel microorganisms from possible environment and subject them to SSF for protease investigation to add up to the nation need of the enzymes and boast economy.

Keywords: Proteases; Fungi; Solid State Fermentation and Substrates

1. Introduction

The process of solid state fermentation for food application is one of the oldest knowledge available to humans. To many communities it is a part of their traditional knowledge to prepare the fermented food [1]. The traditional fermentations such as Koji, Tofu, Miso, sausages, pickles, ensiling are the extension of the traditional fermentation knowledge exploiting the generally registered as safe (GRAS) strains of fungi and bacteria to carry out the fermentation, the process provide extracellular fungal enzymes and have been the basis to initiate the microbial enzymes production by SSF in industrial environment the Koji process undoubtedly influenced the first production of microbial enzyme by SSF on industrial scale, and the emergence of modern SSF based enzyme industry may be attributed to the entrepreneurship of Jokichi Takamine and later by underkofler’s efforts in producing mold bran enzymes for saccharification of grain [2].

Solid state fermentation however as described by Hoda [3] is the one in which micro-organisms secrete the necessary enzymes for degradation of the available substrate molecule in order to meet their nutritional requirements. In this system the fermented mass consist of non-utilized solid substrate containing microbial cells, spores and products for which a number of co-metabolites are formed during the course of fermentation. Equally, SSF is the fermentation in the absence of free liquid and recovery of fermentation products requires it’s extraction from the solid fermented medium with a suitable solvent or solutions [4]. A common extract or solvents is distilled or deionized water [5, 6]. Malathi and Chakraborty [7] also used distilled water for extraction of alkaline protease from solid wheat bran fermentation. Tunga et al., [4] also reported that fermented mass was soaked with water for two hours at room temperature to extract proteolytic enzyme. Other extractants have also been used to extract other enzymes e.g. [8] used sodium chloride...
solution, [9] extracted glycoamylase by sodium chloride solution, [10] used 0.002M succinate buffer for lipase and protease, while Castilho [11] extracted pectinases from wheat bran by acetate buffer at pH 4.4. Equally, [3] extracted milk clotting enzyme from wheat bran fermented by Aspergillus oryzae using distilled water, tap water and different salt concentrations of potassium chloride, magnesium chloride, calcium chloride and sodium chloride.

2. Substrate used for the Production of Enzymes in Solid State Fermentation

Agro industrial residues are generally considered the best substrates for the SSF processes. A number of such substrates have been employed for the cultivation of microorganisms to produce host of enzymes, some of such substrates includes sugarcane bagasse, wheat bran, rice bran, rice husk, soy – hull, corncobs, banana waste, tea waste, cassava waste, palm oil mill waste, Aspen pulp, Sugar beet pulp, sweet sorghum pulp, apple pomace, peanut meal, coconut oil cake, mustard oil cake, cassava flour, wheat flour, steamed rice, steam pre-treated willow, starch e.t.c. [12, 13, 14, 15]. It is interesting to note that although a number of substrates have been employed for culturing different microorganisms, wheat bran has been the preferred choice in most of the studies. Malathi and Chakraborty [7] evaluated a number of carbon sources (bran) for alkaline protease production and reported wheat to be the best for cultivation of A. flavus IMI 32.7634. A strain of A. niger Tieghem 331221 produced large quantities of an extracellular acid protease when grown in SSF system using wheat bran as the sole substrate [16, 17] indicated that wheat bran and rice husk supports the maximal growth and β–galactosidase production by A. oryzae.

2.1. Advantages of SSF

- Process is simple
- Cost effective
- Less effluent release, reduce pollution
- High titers (high product yield)
- Aeration process is easy
- Resembles the natural habitat of some fungi and bacteria
- Easier downstream processing

2.2. Factors Which Influence SSF

- Selection of microorganism: - this is one of the key factors that can improve yield of the product. Bacteria yeast and filamentous fungi can be used. Filamentous fungi has shown better result growing in the solid substrate
- Substrate: - substrate also play an important role in determining the growth of microorganisms there by increasing the product yield, substrate is chosen in such a way that it should provide physical support as well as nutrients to the growing culture.
- Process optimization: - this includes the optimization of physico–chemical and biochemical parameters such as size, initial moisture, pH and pre-treatment of the substrate relative humidity, temperature of in emanation, agitation and accretion, age and size of the inoculum, nutrient supplementation such as N, P and trace element, supplementation of additional carbon source and inducers. Extraction of product and its purification.
- However there are certain problems that are associated with SSF such includes heat transfer which could make it difficult to control the temperature during the fermentation process. Heat is also generative during the metabolic activities of microorganism, since the substrate used has low thermal conductivity heat removal will be slow, and when the heat generated goes beyond certain level it will result in product denaturation and will affect growth of microbes ultimately ending up in reduction in yield and quality of the product.

2.3. Application of SSF

The process of solid state fermentation could be employed in the Agro-food industry e.g. for traditional food fermentations e.g. Koji, fermented cheese e.t.c. and also for bioconversion of byproducts e.g. Sugar Pulp Baggasse composting. SSF is also applicable in the Agricultural Industry for biocontrol, bioinsecticides and for plant growth hormones/enhancers equally in industrial fermentation. Solid state fermentation is applicable for production of enzymes, antibiotics, organic acid as well as ethanol.
3. Fungi as Source of Proteases

Fungi are an attractive source of protease owing to the limited space required for their cultivation and their readily susceptibility to genetic manipulation. So, it is a group of great practical and scientific interest to biotechnologists. Many moulds synthesize and excrete large quantities of proteases into the surrounding media. A number of fungal genera such as Aspergillus, Rhizopus, Mucor, Penicillium, Fusarium, Cephalosporium, Conidiobolus and Neurospora are known to produce acid, alkaline and neutral proteases [18, 19, 20] Those proteases are active over a wide PH (PH4 – 11) and temperature range (30-70°C) [21].

Acid protease of commercial importance are prepared exclusively from fungal sources and are tentatively divided into two sub-groups by their physiological characteristics i.e. pepsin – like acid proteinases and rennin – like acid proteinases. Pepsin – like acid proteinases have usually been reported in the group of black aspergilla such Aspergillus niger A. awamori, A. usamii and A. saitoi, but also occur in species of penicillium and Rhizopus [22, 23].

Rennin – like acid protease are produced by strain of Mucormihei, Mucorpusillus and Trametes sanguinea [24]. Studies on milk clotting enzymes have been carried out by many workers because of their industrial importance for use as rennin substitutes in cheese making. For this purpose various fungal strains including species from Aspergillus, Penicillium and Mucor have been exploited [25]. Production of milk clotting enzymes from microbial sources for use as rennin substitutes has been receiving increasing attention, for long time, calf, rennet has been the traditional source of milk clotting enzymes for cheese manufacturing but decreasing worldwide rennet supplies accompanied by ever increasing cheese production and consumption made necessary the utilization of rennet substitutes [26]. The substitutes are used extensively in the dairy industry to produce a suitable curd with good flavor. The specialized nature of the enzyme is due to its specificity in cleaving a single peptide bond in k-casein to generate insoluble para K-casein and C-terminal glycopeptides. [27] Stressed that for alternative sources of milk clotting enzymes of microbial origin, studies were mainly focused on fungal aspartic proteases, although a few reports on bacterial enzymes can be found. Equally [28] added that filamentous fungi are the major producers of microbial aspartic proteases, again it was observed that fungal substitudes for animal proteases include those from Rhizo mucormiehei [29] Rhizopus oryzae [18] Aspergillus oryzae [30] penicillium oxalicum [31] Amylomyces rouxi and Fussarium subglutinans ((30] among the several microorganisms studied, Rhizomucor miehei and Rhizomucor pusillus have gained industrial acceptance as producers of milk – clotting enzymes [28].

Filamentous fungi are used in many industrial processes for the production of enzymes and metabolites. Among the many advantages offered by the production of enzymes by fungi are low material costs coupled with high productivity, faster production and the ease with which the enzymes can be modified. Further the enzymes being normally extracellular are easily recoverable from the medium [30].

Although, certain rennet preparation are available in market, some from those Thermophilic fungi R. mehei and R. Pasillus, but their uses for cheese production is limited due to their higher thermal stability and low ratio of milk clotting activity to proteolytic activity which may lead to undesirable changes in the organoleptic quality of the final product. A possible solution to alleviate the technical challenges described is the identification of milk clotting enzyme with enhanced specificty and reduced thermo tolerance, perform a pioneering study of the ability of mesophile mucor specie strains to secret milk clotting enzymes, where by a collection of 41 strains out of which 28 strains turnout to be active producers of such enzymes, equally [32] proved the production of aspartic proteases from other mesophile mucor specie strains (Mucor bacilliformis and Mucor circinelloides) which like bovine chymosin have less heat stability than those from Thermophilic fungi, production of milk clotting enzymes with such quality was also reported in penicillium oxalicicum [31] and Nocadiopsis specie by [32].

4. Function of Rennin and Peptidase (Proteases)

The function of rennin enzyme is to curdle milk and separate it into semi-solid curds and liquid whey. Curdling of the milk is necessary if the milk is to be retained in the stomach long enough for the milk proteins to be digested properly. Young mammals would drive no benefit from the milk they drink if it passes through the stomach too quickly which is what would happen if it was allowed to remain in its non-curdle state. Due to its importance in curdling milk, rennin enzyme is widely used in the food industry, notably in the production of cheese [33].

Peptide hydrolyzing enzymes (Peptidases) play an important role in food industry, so the peptidase producing organism is used as starters for the production of cheese. The enzymes rennet can be used to catalyse the conversion of casein in milk to para-casein by removing a glycopeptide from the soluble casein. Para-casein a further clot i.e. coagulates in the
presence of calcium ions to form white, creamy lumps called the curd leaving behind the supernatant called the whey [21]. The major application of peptidases or proteases in diary industry is in cheese manufacturing where the primary function of enzymes is to hydrolyze the specific peptide bond to generate Casein and macro peptides [34]. In general peptidases or proteases have a good number of applications in the food industry e.g degradation of the turbidity complex resulting from protein in fruit juices and alcoholic liquors, improvement of quality of protein rich foods, soy protein hydrolysis, gelatin hydrolysis, casein and whey protein hydrolysis meat protein recovery and meat tenderization [35].

5. Conclusion

Today only 20-30% of the world demand for milk clotting preparation was been covered by calf rennet many successful attempts had been done for producing milk clotting enzymes from other sources such as plants or microorganisms. Various microorganisms are known as producers of rennet substitutes such as Aspergillus oryzae, Endothia parasitica, Mucor circinelloides, Mucor miehei and Rhizomucor pusillus. Hence because of the higher yielding capacity of SSF and the demand for proteases it highly imperative to search for novel microorganisms from possible environment and subject them to SSF for protease investigation to add up to the nation need of the enzymes and boast economy.

Compliance with ethical standards

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Disclosure of conflict of interest

Authors declare No conflict of interest for this paper.

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