RESEARCH ARTICLE

The use of the Gail model, body mass index and SNPs to predict breast cancer among women with abnormal (BI-RADS 4) mammograms

Anne Marie McCarthy, Brad Keller, Despina Kontos, Leigh Boghossian, Erin McGuire, Mirar Bristol, Jinbo Chen, Susan Domchek and Katrina Armstrong

Abstract

Introduction: Mammography screening results in a significant number of false-positives. The use of pretest breast cancer risk factors to guide follow-up of abnormal mammograms could improve the positive predictive value of screening. We evaluated the use of the Gail model, body mass index (BMI), and genetic markers to predict cancer diagnosis among women with abnormal mammograms. We also examined the extent to which pretest risk factors could reclassify women without cancer below the biopsy threshold.

Methods: We recruited a prospective cohort of women referred for biopsy with abnormal (BI-RADS 4) mammograms according to the American College of Radiology’s Breast Imaging-Reporting and Data System (BI-RADS). Breast cancer risk factors were assessed prior to biopsy. A validated panel of 12 single-nucleotide polymorphisms (SNPs) associated with breast cancer were measured. Logistic regression was used to assess the association of Gail risk factors, BMI and SNPs with cancer diagnosis (invasive or ductal carcinoma in situ). Model discrimination was assessed using the area under the receiver operating characteristic curve, and calibration was assessed using the Hosmer-Lemeshow goodness-of-fit test. The distribution of predicted probabilities of a cancer diagnosis were compared for women with or without breast cancer.

Results: In the multivariate model, age (odds ratio (OR) = 1.05; 95% confidence interval (CI), 1.03 to 1.08; P < 0.001), SNP panel relative risk (OR = 2.30; 95% CI, 1.06 to 4.99, P = 0.035) and BMI (≥30 kg/m² versus <25 kg/m²; OR = 2.20; 95% CI, 1.05 to 4.58; P = 0.036) were significantly associated with breast cancer diagnosis. Older women were more likely than younger women to be diagnosed with breast cancer. The SNP panel relative risk remained strongly associated with breast cancer diagnosis after multivariable adjustment. Higher BMI was also strongly associated with increased odds of a breast cancer diagnosis. Obese women (OR = 2.20; 95% CI, 1.05 to 4.58; P = 0.036) had more than twice the odds of cancer diagnosis compared to women with a BMI <25 kg/m². The SNP panel appeared to have predictive ability among both white and black women.

Conclusions: Breast cancer risk factors, including BMI and genetic markers, are predictive of cancer diagnosis among women with BI-RADS 4 mammograms. Using pretest risk factors to guide follow-up of abnormal mammograms could reduce the burden of false-positive mammograms.
Introduction

Though mammography screening reduces breast cancer mortality, it is imperfect like all screening tests. The high burden of false-positive tests relative to the number of cancers detected has contributed to controversy about the routine use of mammography screening among women ages 40 to 50, as well as about biennial rather than annual screening [1]. After 10 years of annual mammography screening beginning at age 40, over 60% of women will have a false-positive result and 7% to 9% will have a biopsy [2]. False-positive mammograms can result in inconvenience, pain and anxiety for patients, as well as increased costs [3,4].

Using pretest probability of disease can improve the positive predictive value of a screening test. However, this approach requires the ability to accurately determine an individual’s risk of disease. The Breast Cancer Risk Assessment Tool (BCRAT), or Gail model, uses age, family history of breast cancer, reproductive history and history of breast biopsy or atypical hyperplasia to estimate a woman’s 5-year or lifetime risk of breast cancer [5]. Although the model is well validated, its discriminatory accuracy is modest [6]. Additional risk factors, such as genetic markers [7-14] and body mass index (BMI) [15-19], have been shown to moderately improve breast cancer risk prediction.

Although many studies have focused on predicting cancer risk in the general population, few have employed risk prediction models to improve decisions about follow-up of abnormal mammograms. Current standards in the United States recommend biopsy of a mammographic abnormality if the radiologist deems the probability of cancer diagnosis to be at least 2% [20-22]. Mammogram results are reported using the American College of Radiology (ACR) Breast Imaging-Reporting and Data System (BI-RADS), which includes six result categories, each tied to follow-up recommendations [20]. The BI-RADS 4 category indicates the presence of a suspicious abnormality that should be followed up with a biopsy. However, the 1-year probability of breast cancer for women with a BI-RADS 4 mammogram is 15% to 30% on average [20,22-29]; therefore, the majority of biopsies of BI-RADS 4 abnormalities are benign. Furthermore, the likelihood of cancer diagnosis varies widely within the BI-RADS 4 category, leading to the subdivision of the category into BI-RADS 4A (2% to 9% risk of malignancy), BI-RADS 4B (10% to 49% risk of malignancy) and BI-RADS 4C (50% to 94% risk of malignancy) [22]. A small pilot study suggested that an experienced radiologist using this stratification scheme could increase the threshold for the biopsy decision without missing invasive cancers [30]. In addition, a recent modeling study suggested that the addition of pretest breast cancer risk factors, including genetic markers, could change biopsy decisions for a small proportion of women with abnormal mammograms [31]. Greater ability to predict cancer outcomes in women with BI-RADS 4 mammograms could reduce the burden of false-positive tests from mammography.

In this study, we assessed the usefulness of the Gail model, BMI and a panel of 12 single-nucleotide polymorphisms (SNPs) to predict cancer diagnosis among women with BI-RADS 4 mammograms. We then evaluated the extent to which these factors could improve decisions about biopsy among this group by reclassifying women without cancer below the biopsy threshold.

Methods

Participants

Women referred for breast biopsies at the Hospital of the University of Pennsylvania following a BI-RADS 4 mammogram between January 2010 and April 2012 were invited to participate in the study. Women were excluded if they were younger than 20 years old, had a personal history of breast or ovarian cancer, mantle radiation or known BRCA1/2 mutation. Women who consented provided a buccal swab for DNA testing prior to their biopsy appointment. Three hundred sixty-three women were enrolled. An additional 119 women with a BI-RADS 4 mammograms from a previous study in which breast imaging modalities were compared at the same institution were also included (2002 to 2006; National Institutes of Health grant P01 CA85484; Principal Investigator: M Schnall). Participants in the breast imaging study were enrolled between July 2003 and August 2007. A blood sample from each patient was collected and stored, which was used for genetic analysis. Of the total sample, five patients were missing follow-up information, eleven had data on fewer than nine SNP markers and two had nonbreast malignancies (tubular adenoma, B-cell lymphoma in the breast). These participants were excluded, resulting in a total population of 464 for analysis. Both studies were approved by the University of Pennsylvania Institutional Review Board, and written informed consent was obtained from each study participant.

Risk factors

Participants completed a health history questionnaire, including information on race, age at menarche, age at first live birth, number of biopsies, presence of atypical hyperplasia and family history of breast and ovarian cancer. Using the BCRAT, we estimated the 5-year absolute risk and relative risk (RRs) of breast cancer using source code version 3.0 from the National Cancer Institute website [32]. BMI was calculated by using the patient’s self-reported weight and height at the time of recruitment, or it was extracted from medical record data prior to recruitment.
Single-nucleotide polymorphism panel

Buccal swabs (N = 347) or blood samples (N = 117) were sent to deCODE genetics (Reykjavik, Iceland) for analysis using Illumina Infinium II whole-genome genotyping (Illumina, San Diego, CA, USA). The deCODE genetics SNP assay included 12 loci that have consistently been associated with breast cancer risk: 2q35 (rs13387042), MRPS30 (rs4415084), FGFR2 (rs1219648), TNRC9/TOX3 (rs3803662), 8q24 (rs13281615), LSP1 (rs3817198), 5q11 (rs889312), NEK10 (rs4973768), 1p11 (rs11249433), RAD51L1 (rs999737), COX11 (rs6504950) and CASP8 (rs1045485) [33-40]. The call rate was 99.8%. The deCODE BreastCancer™ test uses individual allele effect sizes for the 12 SNPs to create a RR estimate for each genotype. For each participant, a combined RR estimate for the 12-SNP panel was calculated by multiplying the RR estimates for all SNPs as described previously [11]. Expected and observed allele frequencies and homozygote odds ratios (ORs) for risk alleles are included in Additional file 1. The combined SNP panel RR estimate has been shown to be independent of BCRAT factors [11].

Statistical analysis

The results of the BIRADS 4 biopsies were obtained from pathology records. Logistic regression was used to assess the association of Gail risk factors, BMI and SNP panel RR with breast cancer (invasive or ductal carcinoma in situ (DCIS)). First, each predictor was tested in an age-adjusted model. SNP panel RRs were examined as a log-transformed continuous variable and as categorized RRs <1.00, 1.01 to 1.49 and ≥1.50. The Gail RR was tested as a log-transformed continuous variable. Gail absolute 5-year risk estimate was categorized as <1.67% and ≥1.67%, as well as for the use of chemopreventive drugs [41,42]. BMI data were missing in 17% of participants, and therefore BMI was entered into models, including a category for missing data, as follows: <25 kg/m², 25 to 29.9 kg/m², ≥30 kg/m² and missing. The multivariate logistic regression model included log-transformed SNP panel RR, all Gail risk factors (age, race/ethnicity, age at menarche, age at first live birth, first-degree family history of breast cancer, breast biopsy, atypical hyperplasia) and BMI. We also examined the predictive ability of the various risk factors. Model calibration was assessed using the Hosmer-Lemeshow goodness-of-fit test to compare observed and predicted outcomes within deciles of predicted risk for each model [43]. Discriminatory accuracy was assessed by calculating area under the receiver operating characteristic curve (AUC). DeLong’s test was used to compare AUCs for various models. In our analysis, the model incorporating age and the Gail RR had poor calibration. The original Gail model incorporated 5-year intervals of age, but we entered age as a continuous predictor to minimize the number of predictors in our models. Because of the poor calibration of the age plus Gail RR model, we also examined a model that entered all Gail risk factors individually, and this model was better calibrated to our data. In addition, we performed tenfold cross-validation of the prediction models in the total study population. Finally, we estimated the predicted probability of cancer using the multivariate model and assessed reclassification below several risk thresholds (2%, 3%, 5% and 10%) for cancer cases and noncancer cases. Statistical analyses were performed using SAS 9.3 (SAS Institute, Cary, NC, USA) and Stata/IC 12 (College Station, TX, USA) software.

Results

The mean age of study participants was 48.7 years (SD, 13.2), and approximately one-half of the study population was over age 50 (Table 1). Over 30% of participants were black or African American. The mean 5-year breast cancer risk estimate derived by using the BCRAT was 1.54, and 33% of participants had a 5-year risk estimate of 1.67% or greater. The mean SNP panel RR was 1.22 (SD, 0.44). Over one-fourth of participants had a SNP panel RR estimate of 1.50 or greater, indicating their risk of breast cancer was 50% greater than that of the general population. Of the 464 participants, 74 women (16%) were diagnosed with cancer, 33 (7%) with DCIS and 41 (9%) with invasive cancer.

Table 2 displays the results of age-adjusted and multivariate logistic regression models used to estimate the OR for cancer diagnosis. The SNP panel RR was significantly associated with cancer diagnosis (OR, 2.15; 95% CI, 1.04 to 2.43; P = 0.038). The ORs estimated in our model for the categorized SNP panel RRs were comparable to the predefined RR estimates obtained from deCODE genetics. The Gail RR estimate was not significantly associated with cancer diagnosis, nor was Gail absolute 5-year risk ≥1.67%. Among the Gail factors, only age was significantly associated with breast cancer diagnosis, though the ORs for race/ethnicity, age at menarche, age at first live birth and family history of breast cancer were consistent with expected associations. Prior breast biopsy and atypical hyperplasia were inversely associated with breast cancer, though these data were not statistically significant. Few participants (4.3%) reported prior atypical hyperplasia.

In the multivariate model, age, SNP panel RR and BMI were significantly associated with breast cancer diagnosis. Older women were more likely than younger women to be diagnosed with breast cancer (OR = 1.05; 95% CI, 1.03 to 1.08; P < 0.001). The SNP panel RR remained strongly associated with breast cancer diagnosis after multivariable adjustment (OR = 2.30; 95% CI, 1.06 to 4.99; P = 0.035). Higher BMI was also strongly associated with increased odds of breast cancer diagnosis. Obese women (OR = 2.20; 95% CI, 1.05 to 4.58; P = 0.036) had more than twice the
Next, we evaluated the association of the SNP panel separately for white (N = 277) and black (N = 145) women (Table 3). Among white women, the SNP panel RR was associated with twofold elevated odds of receiving a cancer diagnosis in both age-adjusted (OR = 2.43; 95% CI, 0.99 to 5.98; P = 0.053) and multivariate (OR = 1.97; 95% CI, 0.76 to 5.10; P = 0.161) models, and OR estimates were similar for the SNP panel RR categories and predefined values. There was evidence that the SNP panel RR was associated with breast cancer diagnosis among black women. Among black women, the OR estimate was 4.50 in the age-adjusted model (OR = 4.50; 95% CI, 0.87 to 23.2; P = 0.073) and 4.21 in the multivariate model adjusted for age, Gail factors and BMI (OR = 4.21; 95% CI, 0.79 to 22.6; P = 0.093), though these estimates did not reach statistical significance. In addition, the OR estimates for the SNP panel RR categories were similar to the predefined RR values. There was no significant interaction between race and the SNP panel RR (P = 0.880).

We compared the predictive accuracy of the Gail factors, BMI and SNP panel RR (Table 4). First, Gail RR, SNP panel RR and BMI were tested separately in models including age. The model with age and Gail RR had the lowest predictive ability (AUC = 0.6646), and the Hosmer-Lemeshow goodness-of-fit test indicated poor model fit (P = 0.0073) and 4.21 in the multivariate model adjusted for age, Gail factors and BMI (OR = 4.21; 95% CI, 0.79 to 22.6; P = 0.093), though these estimates did not reach statistical significance. In addition, the OR estimates for the SNP panel RR categories were similar to the predefined RR values. There was no significant interaction between race and the SNP panel RR (P = 0.880).

We compared the predictive accuracy of the Gail factors, BMI and SNP panel RR (Table 4). First, Gail RR, SNP panel RR and BMI were tested separately in models including age. The model with age and Gail RR had the lowest predictive ability (AUC = 0.6646), and the Hosmer-Lemeshow goodness-of-fit test indicated poor model fit (P = 0.0019). All other models exhibited acceptable model fit (P > 0.05). The predictive accuracy was similar for age and the SNP panel RR (0.6848) and age and BMI (0.6845). Age, BMI and the SNP panel RR together yielded an AUC of 0.7007, which was of borderline significance compared to age alone (P = 0.061).

Predictive accuracy was greater in the model including the individual Gail risk factors (0.7144) compared to a model with age alone (P = 0.044). Adding BMI to the

### Table 1 Characteristics of BIRADS 4 cohort, all ages, N = 464

| Characteristic | Data |
|---------------|------|
| Age, yr, mean ± SD (range) | 48.7 ± 13.2 (20 to 86) |
| Age, yr, categories | |
| <35 | 73 | 15.7 |
| 35 to 40 | 41 | 8.8 |
| 40 to 49 | 114 | 24.6 |
| 50 to 59 | 146 | 31.5 |
| 60+ | 90 | 19.4 |
| Race/ethnicity | |
| White | 277 | 59.7 |
| African American/black | 145 | 31.3 |
| Hispanic | 9 | 1.9 |
| Asian | 16 | 3.5 |
| Other | 17 | 3.7 |
| Age at menarche, yr | |
| <11 | 90 | 19.4 |
| 12 to 13 | 200 | 43.1 |
| ≥14 | 107 | 23.1 |
| Unknown | 67 | 14.4 |
| Age at first live birth, yr | |
| <20 | 69 | 14.9 |
| 20 to 24 | 94 | 20.3 |
| 25 to 29 | 76 | 16.4 |
| ≥30 | 80 | 17.2 |
| Nulliparous | 139 | 30 |
| Missing data | 6 | 1.3 |
| First-degree relatives with breast or ovarian cancer, n | |
| 0 | 350 | 75.4 |
| 1 | 101 | 21.8 |
| >1 | 13 | 2.8 |
| Prior breast biopsy, n | |
| 0 | 266 | 57.3 |
| 1 | 123 | 26.5 |
| >1 | 75 | 16.2 |
| Prior AH | |
| Yes | 20 | 4.3 |
| No | 444 | 95.7 |
| Gail 5-yr risk estimate, mean ± SD | 1.54 ± 1.43 |
| Gail 5-yr risk estimate, % | |
| <1.67 | 309 | 66.6 |
| ≥1.67 | 155 | 33.4 |
| Body mass index, kg/m² | |
| <25 | 182 | 39.2 |
| 25 to 29.9 | 95 | 20.5 |

Notes: AH, Atypical hyperplasia; BI-RADS, Breast Imaging-Reporting and Data System; DCIS, Ductal carcinoma in situ; LCIS, Lobular carcinoma in situ; RR, Relative risk; SD, Standard deviation. Data are number and percent unless otherwise stated.
Gail risk factor model increased the AUC (0.7279), but the difference was not statistically significant ($P = 0.341$). Subsequently adding the SNP panel RR to the model further increased the AUC (0.7377; $P = 0.212$). We repeated analyses stratified by age (35 to 49 years and $\geq 50$ years) and found that the addition of BMI and SNP panel RR improved predictive accuracy compared to the Gail factors alone in both age groups, though the AUC values were greater for women ages 50 and older. The addition of the SNP panel had a greater impact on the AUC in the 35 to 49 age group than in women ages 50 and older. When stratified by race, the AUC values were comparable for black women and white women. For the model including Gail factors, BMI and SNP panel RR, the AUC was 0.7518 for white women and 0.7710 for black women. We repeated our analyses excluding women younger than 40, and the results were similar. We performed tenfold cross-validation on the prediction models in the total study population (Table 5). AUC values were slightly attenuated after cross-validation and were not statistically significant.

| Table 2 Logistic regression, odds of cancer among women with BIRADS 4 mammograms, $N = 464^a$ |
|-----------------------------------------------|
| **Age-adjusted** | **Multivariate$^b$** |
| **OR** | **95% CI** | **P-value** | **OR** | **95% CI** | **P-value** |
| SNP panel RR, log continuous scale | 2.15 | 1.04 to 2.43 | 0.038 | 2.30 | 1.06 to 4.99 | 0.035 |
| SNP panel RR, categories | | | | | | |
| $<1.00$ | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| $1.01$ to $1.49$ | 1.09 | 0.59 to 2.02 | 0.788 | 1.09 | 0.60 to 1.98 | 0.778 |
| $\geq 1.50$ | 1.60 | 0.84 to 3.04 | 0.149 | 1.60 | 0.84 to 3.04 | 0.149 |
| Gail RR, log continuous scale | 1.11 | 0.69 to 1.78 | 0.660 | 1.11 | 0.69 to 1.78 | 0.660 |
| Gail absolute 5-yr risk, % | | | | | | |
| $<1.67$ | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| $\geq 1.67$ | 1.09 | 0.60 to 1.98 | 0.778 | 1.09 | 0.60 to 1.98 | 0.778 |
| Age, log continuous scale | 1.05 | 1.03 to 1.07 | <0.001 | 1.05 | 1.03 to 1.08 | <0.001 |
| Race/ethnicity | | | | | | |
| White | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| African American/black | 0.66 | 0.37 to 1.19 | 0.170 | 0.53 | 0.26 to 1.06 | 0.071 |
| Other | 0.86 | 0.33 to 2.20 | 0.748 | 0.81 | 0.30 to 2.23 | 0.689 |
| Age at menarche, yr | | | | | | |
| $<11$ | 1.44 | 0.65 to 3.21 | 0.368 | 1.33 | 0.57 to 3.09 | 0.510 |
| $12$ to $13$ | 1.67 | 0.85 to 3.30 | 0.139 | 1.50 | 0.73 to 3.06 | 0.266 |
| $\geq 14$ | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| Unknown | 0.77 | 0.29 to 2.07 | 0.608 | 0.83 | 0.28 to 2.41 | 0.729 |
| Age at first live birth, yr | | | | | | |
| $<30$ | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| $\geq 30$ | 1.58 | 0.81 to 3.08 | 0.183 | 1.37 | 0.66 to 2.87 | 0.400 |
| Nulliparous | 1.09 | 0.58 to 2.06 | 0.780 | 1.06 | 0.54 to 2.08 | 0.867 |
| Missing data | 1.60 | 0.17 to 15.0 | 0.680 | 2.26 | 0.22 to 23.6 | 0.497 |
| First-degree relatives with breast cancer, n | 1.48 | 0.86 to 2.57 | 0.160 | 1.62 | 0.90 to 2.90 | 0.106 |
| Prior breast biopsy | 0.73 | 0.43 to 1.24 | 0.242 | 0.82 | 0.47 to 1.46 | 0.508 |
| Prior atypical hyperplasia | 0.41 | 0.09 to 1.86 | 0.247 | 0.51 | 0.10 to 2.54 | 0.410 |
| BMI, kg/m$^2$, mean ± SD | | | | | | |
| $<25$ | 1.00 | Reference | 1.00 | Reference | 1.00 | Reference |
| 25 to 29.9 | 1.68 | 0.81 to 3.47 | 0.161 | 1.86 | 0.86 to 4.05 | 0.116 |
| $\geq 30$ | 1.94 | 0.99 to 3.81 | 0.054 | 2.20 | 1.05 to 4.58 | 0.036 |
| Missing data | 1.85 | 0.87 to 3.93 | 0.111 | 1.80 | 0.81 to 3.99 | 0.147 |

$^a$BIRADS, Breast Imaging-Reporting and Data System; BMI, Body mass index; CI, Confidence interval; OR, Odds ratio; RR, Relative risk; SD, Standard deviation; SNP, Single-nucleotide polymorphism. $^b$Multivariate model includes log SNP RR, age, BMI, race/ethnicity, age at menarche, age of first birth, family history of breast cancer, breast biopsy and atypical hyperplasia.
The highest cross-validated AUC was observed for the model including age, BMI and the SNP panel (AUC = 0.6753).

The predicted probabilities of breast cancer diagnosis for each individual were estimated using the model including age, Gail factors, BMI and the SNP panel RR. Figure 1 displays the distribution of predicted probabilities by breast cancer status. Women diagnosed with cancer (true-positives) had a mean predicted probability of cancer diagnosis of 22.6%, compared to 12.2% for women not diagnosed with cancer (false-positives), though the 95% CIs significantly overlapped (Table 6). However, no women diagnosed with cancer had a predicted probability below 5%. On the basis of our model, nine women (3.4%) with BI-RADS 4 mammograms were reclassified below the <2% threshold, none of whom were diagnosed with cancer. Furthermore, 69 women (14.9%) had a predicted probability of cancer less than 5%, and none of these women were subsequently diagnosed with cancer. The positive predictive value of the BI-RADS 4 categorization alone was 15.9%, compared to 18.7% using the BI-RADS 4 categorization along with the prediction model with a 5% predicted probability.

**Discussion**

Our results suggest that breast cancer risk factors can be used to predict cancer diagnosis among women with BI-RADS 4 mammograms. Age, BMI and the 12-SNP panel were strongly associated with cancer diagnosis. Addition of BMI and the 12-SNP panel to Gail risk factors improved model discrimination. Furthermore, using a predicted probability cutoff of 5% for biopsy would reclassify 15% of women below the biopsy threshold while retaining 100% sensitivity in cancer detection in this sample. Though our results need to be prospectively validated, our work provides proof of concept that the use of pretest risk factors to guide follow-up of BI-RADS 4 mammograms could potentially improve mammography screening outcomes by reducing the number of biopsies among women who do not have cancer.

To our knowledge, our present study is the first in which a panel of genetic markers has been tested in women with abnormal mammograms. The SNP panel RR estimates observed were similar to the RR estimates stated by deCODE genetics in our population of women with BI-RADS 4 mammograms, and the SNP panel RR estimate remained strongly associated with cancer diagnosis.

---

**Table 3 Logistic regression, odds of cancer among women with BI-RADS 4 mammograms, by race**

|                     | White (N = 277) | Black (N = 145) |
|---------------------|----------------|-----------------|
|                     | Age-adjusted   | Multivariate^b  | Age-adjusted   | Multivariate^b  |
| SNP panel RR, log continuous scale | OR (95% CI) | P-value | OR (95% CI) | P-value | OR (95% CI) | P-value | OR (95% CI) | P-value |
| <1.00               | 1.00 (reference) | 1.00 (reference) |
| 1.01 to 1.49        | 1.34 (0.64 to 2.82) | 0.437 | 1.56 (0.90 to 8.18) | 0.599 |
| ≥1.50               | 1.84 (0.83 to 4.11) | 0.135 | 1.77 (0.33 to 9.52) | 0.505 |

^bBI-RADS, Breast Imaging-Reporting and Data System; BMI, Body mass index; CI, Confidence interval; OR, Odds ratio; RR, Relative risk; SNP, Single-nucleotide polymorphism.

^Multivariate model includes BMI, age, race/ethnicity, age at menarche, age at first live birth, family history of breast cancer, breast biopsy, atypical hyperplasia.

**Table 4 Predictive accuracy of models using Gail risk factors, body mass index and single-nucleotide polymorphism panel among women with BI-RADS 4 mammograms**

|                     | Total study population (N = 464) | Age 35 to 49 yr (N = 155) | Age ≥50 yr (N = 236) | White (N = 277) | Black (N = 145) |
|---------------------|---------------------------------|---------------------------|---------------------|-----------------|-----------------|
|                     | AUC | P-value^e | AUC | P-value^e | AUC | P-value^e | AUC | P-value^e | AUC | P-value^e | AUC | P-value^e |
| Age, log Gail RR    | 0.6646 | 0.0019 | 0.839 | 0.5475 | 0.966 | 0.6748 | 0.470 | 0.6654 | 0.373 | 0.7243 | 0.495 |
| Age, BMI            | 0.6845 | 0.3649 | 0.210 | 0.5775 | 0.619 | 0.6914 | 0.613 | 0.6826 | 0.317 | 0.7456 | 0.653 |
| Age, log SNP RR     | 0.6848 | 0.3134 | 0.197 | 0.6068 | 0.445 | 0.6917 | 0.508 | 0.6890 | 0.153 | 0.7385 | 0.836 |
| Age, BMI, log SNP RR| 0.7007 | 0.2927 | 0.061 | 0.6258 | 0.337 | 0.7086 | 0.276 | 0.7078 | 0.060 | 0.7527 | 0.661 |
| Gail factors        | 0.7144 | 0.3586 | 0.044 | Reference | 0.5488 | 0.123 | 0.7115 | 0.267 | 0.7390 | 0.019 | 0.7256 | 0.925 |
| Gail factors, BMI   | 0.7279 | 0.7646 | 0.014 | 0.341 | 0.6964 | 0.072 | 0.7272 | 0.167 | 0.7463 | 0.011 | 0.7485 | 0.602 |
| Gail factors, BMI, log SNP RR | 0.7377 | 0.1924 | 0.007 | 0.212 | 0.7242 | 0.026 | 0.7356 | 0.116 | 0.7518 | 0.007 | 0.7719 | 0.442 |

^AUC, Area under the receiver operating characteristic curve; BI-RADS, Breast Imaging-Reporting and Data System; BMI, Body mass index; OR, Relative risk; SNP, Single-nucleotide polymorphism.

^P-value derived from Hosmer-Lemeshow goodness-of-fit (GOF) test.

^P-value derived from DeLong test compared to a model with age only.

^P-value derived from DeLong test compared to reference model.

^Gail factors include age, race/ethnicity, age at menarche, age at first birth, family history of breast cancer, breast biopsy, atypical hyperplasia.
after adjusting for other breast cancer risk factors. Similar to what has been reported in prior studies [7,8,10-14,44,45], the SNP panel in the present study moderately improved predictive accuracy. However, this small improvement may prove to be more clinically valuable for decisions about biopsies among women with abnormal mammograms than for risk stratification in the general population.

It was not entirely surprising that the Gail risk estimate was not significantly associated with cancer diagnosis in our study, because the Gail model was developed to estimate 5-year or lifetime risk of invasive breast cancer in the general population. In our present study, we attempted to predict the risk of diagnosis of either DCIS or invasive cancer in women with abnormal mammograms. The magnitudes of the exposure–disease relationships are likely different for short-term cancer outcomes in the higher-risk BI-RADS 4 population. In our analysis, the model using age and the Gail RR had poor calibration, and therefore the AUC estimates are not meaningful. The poor calibration of this model could have been due to differences in the study population and outcome used in our study, or it could have been a result of our inclusion of age as a continuous predictor to provide a more parsimonious model, whereas the original Gail model used 5-year age categories. Because of this, we also examined a model that entered all Gail risk factors individually, and this model was better calibrated to our data. We observed an AUC of 0.738 for the model with Gail factors, BMI and the SNP panel, which is higher than the AUC observed in the general population for the Gail model alone (0.596) or the Gail model including breast density (0.634) [46]. Researchers in two prior studies evaluated prediction models in women with BI-RADS 4 mammograms. A prediction model trained on 170 French patients with BI-RADS 4 mammograms using Gail risk, age, presence of a palpable lesion, lesion size, hormone replacement therapy and menopause status demonstrated predictive accuracy similar to our model, with an AUC of 0.716 in the training set and AUC of 0.660 when validated in 188 BI-RADS 4 patients from Texas [47]. Similar to our results, age was the strongest predictor of cancer among approximately 4,000 women with BI-RADS 4 mammograms referred for biopsy between 1997 and 2001 in the Vermont Breast Cancer Surveillance System [48]. The presence of a palpable lump, previous breast biopsy, menopause status and use of postmenopausal hormone therapy were also associated with cancer diagnosis. Genetic risk factors and BMI were not included in these prediction models.

Obese women had more than twice the odds of receiving a cancer diagnosis compared to women of normal weight. One possible explanation for this association is that obese women tend to have less-dense breasts and therefore

Table 5 Cross-validation of prediction models

| Total study population (N = 464) | Tenfold cross-validation |
|----------------------------------|--------------------------|
|                                  | AUC 95% CI | P-value | AUC 95% CI | P-value |
| Age, log Gail RR                 | 0.6646 0.5970 to 0.7321 | 0.839  | 0.6482 0.5797 to 0.7167 | 0.159  |
| Age, BMI                         | 0.6845 0.6188 to 0.7501 | 0.210  | 0.6583 0.5911 to 0.7255 | 0.764  |
| Age, log SNP RR                  | 0.6848 0.6195 to 0.7501 | 0.197  | 0.6735 0.6077 to 0.7393 | 0.188  |
| Age, BMI, log SNP RR             | 0.7007 0.6370 to 0.7645 | 0.061  | 0.6753 0.6099 to 0.7407 | 0.258  |
| Gail factors                     | 0.7144 0.6532 to 0.7755 | 0.044  | 0.6522 0.5855 to 0.7188 | 0.955  |
| Gail factors, BMI                | 0.7279 0.6705 to 0.7854 | 0.014  | 0.6561 0.5919 to 0.7203 | 0.924  |
| Gail factors, BMI, log SNP RR    | 0.7377 0.6808 to 0.7946 | 0.007  | 0.6727 0.6099 to 0.7356 | 0.493  |

*Table 6 Predicted probability of cancer using Gail factors, body mass index and single-nucleotide polymorphism panel (N = 464)*

| Cancer | Noncancer |
|--------|-----------|
| Median % (95% confidence interval) | 22.6% (7.0% to 46.6%) | 12.2% (2.6% to 38.8%) |
| Predicted probability, n (%)     | Cancer | Noncancer |
| <2%     | 0 (0%) | 9 (1.9%) |
| <3%     | 0 (0%) | 35 (7.5%) |
| <5%     | 0 (0%) | 69 (14.9%) |
| <10%    | 10 (2.2%) | 162 (34.9%) |

Figure 1 Distribution of the predicted probability of cancer using Gail factors, body mass index and single-nucleotide polymorphism panel.
potentially easier-to-read mammograms, which facilitates a more accurate interpretation of their mammograms by radiologists, such that obese women with a BI-RADS 4 mammogram are more likely to actually have cancer (and less likely to have a false-positive test) than nonobese women. The association of BMI with cancer diagnosis may also reflect disease etiology, as BMI is associated with increased risk of postmenopausal breast cancer [49]. Although BMI data were missing for 17% of participants, we do not believe the missing data biased the observed association. The distribution of risk factors (except for age at first birth) and percentage diagnosed with cancer did not differ for women with missing BMI data and women with complete BMI data. Additional studies are needed to verify this association and to tease apart the effects of BMI and breast density in women with abnormal mammograms.

This study was a first attempt to validate the 12-SNP panel among black women. The SNP panel variants were identified and validated primarily in white/European populations. Several genome-wide association studies and candidate gene studies [50-60], and authors of meta-analyses [61-66] have assessed the association of these 12 SNPs individually with breast cancer risk among black/African American populations, with mixed results. Six of the twelve SNPs in the panel have been replicated in at least one study of black/African American populations: rs1045485 (CASP8) [59], rs1219648 (FGFR2) [54,58,59], rs13387042 (2q35) [52,58,59], rs3817198 (LSP1) [60], rs4415084 (FGF10) [56] and rs999737 (14q24.1, RAD51B) [59]. Validating breast cancer–associated SNPs among black women is challenging, given the large sample sizes needed to detect small associations, differing linkage disequilibrium patterns among different ancestral groups, and disease heterogeneity. Despite the fact that only half of these SNP associations have been replicated, the 12-SNP panel appeared to have predictive value among black women, though our results need to be validated in larger studies. In addition, in future studies, researchers should assess whether race-specific and tumor subtype–specific SNP panels can further improve breast cancer risk prediction.

Several limitations should be considered when interpreting our results. Because we recruited women referred for biopsy at one academic hospital, our study sample may not be representative of all women with abnormal mammograms referred for biopsy. Our sample size was modest, and therefore our results, particularly those of subgroup analyses, should be interpreted cautiously. We performed cross-validation of our prediction models for the entire study sample; however, prospective validation of our results is needed. Given the limited number of cancers (N = 75), our study did not have statistical power to fit separate models for DCIS and invasive cancer or to assess interactions between risk factors. We utilized a validated panel of 12 breast cancer–associated SNPs. To date, nearly 70 SNPs have been identified that are associated with breast cancer risk [67]. Therefore, our results using 12 SNPs may underestimate the utility of genetic markers, and including a larger number of genetic markers may further improve risk prediction. In future studies, researchers should evaluate the use of genetic markers in women with abnormal mammograms. Also, breast density was not controlled for, and this may partly explain the observed association of BMI with cancer diagnosis.

This study has several strengths. Ours is one of the first studies to develop a cancer prediction model for women with abnormal mammograms. We had rich data on recognized breast cancer risk factors ascertained prior to biopsy. We employed a validated panel of genetic markers associated with breast cancer incidence, with RR estimates independent of traditional breast cancer risk factors. Our study population was diverse in terms of age and race/ethnicity, suggesting that our model could be applied broadly.

Conclusions

Our results suggest that pretest breast cancer risk factors could be utilized to individualize biopsy decisions following abnormal mammograms. We found that age, BMI and a 12-SNP panel were significantly associated with breast cancer diagnosis in women with BI-RADS 4 mammograms. The association of obesity with cancer diagnosis was particularly novel and warrants additional investigation. On the basis of results derived from the model using Gail risk factors, BMI and genetic markers, we were able to identify a predicted probability threshold that could be used to identify women who would not benefit from immediate biopsy. Our study, though preliminary, highlights that improved risk modeling for women with abnormal mammograms could reduce the burden of false-positive tests and therefore increase the benefits of mammography. Future studies are needed to validate these results in larger patient populations.

Additional file

Additional file 1: Expected and observed allele frequencies (N = 464).

Abbreviations

AH: Atypical hyperplasia; AUC: Area under the receiver operating characteristic curve; BCRAT: Breast Cancer Risk Assessment Tool; BI-RADS: Breast Imaging-Reporting and Data System; BMI: Body mass index; CI: Confidence interval; DCIS: Ductal carcinoma in situ; LCIS: Lobular carcinoma in situ; OR: Odds ratio; RR: Relative risk; SD: Standard deviation; SNP: Single-nucleotide polymorphism.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

SD and KA conceived of and designed the study, secured grant funding and interpreted the results. AM assisted in study design, performed statistical analyses, interpreted the results and drafted the manuscript. JC assisted in study design, statistical analysis and interpretation of the results. BK assisted.
in study design and acquisition of data, performed statistical analyses and interpreted the results. DX assisted in acquisition of data and interpretation of the results. LB, EM and MB provided study coordination and assisted in acquisition of data. All authors critically reviewed the manuscript and read and approved the final manuscript.

Authors’ information
Susan Domchek and Katrina Armstrong are both senior authors.

Acknowledgements
This work was supported by the following grants from the National Institutes of Health, National Cancer Institute (NIH/NCI): R01 CA85484, U2C CA148310 and US4 CA163313. Additionally, JC was partially supported by NCI award R01 CA164305. The NIH/NCI did not have any role in the study design, collection, analysis or interpretation of data, writing of the manuscript; or the decision to submit the manuscript for publication.

Author details
1 Department of Medicine, Massachusetts General Hospital, 50 Staniford Street, 940F, Boston, MA 02114, USA. 2 Department of Radiology, University of Pennsylvania, Philadelphia, PA, USA. 3 Abramson Cancer Center, University of Pennsylvania, Philadelphia, PA, USA. 4 Department of General Internal Medicine, University of Pennsylvania, Philadelphia, PA, USA. 5 Department of Clinical Epidemiology and Biostatistics, University of Pennsylvania, Philadelphia, PA, USA.

Received: 13 February 2014 Accepted: 18 December 2014

Published online: 08 January 2015

References
1. US Preventive Services Task Force. Screening for breast cancer: U.S. Preventive Services Task Force recommendation statement. Ann Intern Med. 2009;151:716–26. Published errata appear in Ann Intern Med. 2010;152:199–200, and Ann Intern Med. 2010;152:686.
2. Hubbard RA, Kerkilwoska K, Flowers CJ, Yankaskas BC, Zhu W, Migliorelli DL. Cumulative probability of false-positive recall or biopsy recommendation after 10 years of screening mammography: a cohort study. Ann Intern Med. 2011;155:481–92. A published erratum appears in Ann Intern Med. 2014;160:568.
3. Chubuk J, Boudreau DM, Fishman PA, Elmore JG. Cost of breast-related care in the year following false positive screening mammograms. Med Care. 2010;48:815–20.
4. Brewer NT, Salt T, Lillee S. Systematic review: the long-term effects of false-positive mammograms. Ann Intern Med. 2007;146:502–10.
5. Gail MH, Brinton LA, Byar DP, Corle DK, Green SB, Schairer C, et al. Projecting the cumulative incidence of breast cancer: a model for breast cancer risk assessment. J Natl Cancer Inst. 1990;82:71–83.
6. Rockhill B, Spiegelman D, Byrne C, Hunter DJ, Colditz GA. Validation of the Gail et al. model of breast cancer risk prediction and implications for chemoprevention. J Natl Cancer Inst. 2001;93:358–66.
7. Wacholder S, Hartge P, Prentice R, Garcia-Closas M, Feigelson HS, Diver WR, et al. Performance of common genetic variants in breast-cancer-risk models. N Engl J Med. 2010;363:986–93.
8. Gail MH. Discriminatory accuracy from single-nucleotide polymorphisms in models to predict breast cancer risk. J Natl Cancer Inst. 2008;100:1037–41.
9. Gail MH, Mai PL. Comparing breast cancer risk assessment models. J Natl Cancer Inst. 2010;102:665–6.
10. Gail MH. Value of adding single-nucleotide polymorphism genotypes to a breast cancer risk model. J Natl Cancer Inst. 2010;102:665–6.
11. Melaffke ME, Stokowski RP, Phees BK, Prentice RL, Pettinger M, Hinds DA. Assessment of clinical validity of a breast cancer risk model combining genetic and clinical information. J Natl Cancer Inst. 2010;102:1618–27.
12. Dite GS, Mahmoodi M, Bickerstaffe A, Hammer F, Macinnis RJ, Tsikitis M, et al. Using SNP genotypes to improve the discrimination of a simple breast cancer risk prediction model. Breast Cancer Res Treat. 2013;139:887–96.
13. Darabi H, Czene K, Zhao W, Liu J, Hall P, Humphreys K. Breast cancer risk prediction and individualised screening based on common genetic variation and breast density measurement. Breast Cancer Res. 2012;14:R25.
14. Husing A, Canzian F, Beckmann L, Garcia-Closas M, Diver WR, Thun MJ, et al. Prediction of breast cancer risk by genetic risk factors, overall and by hormone receptor status. J Med Genet. 2012;49:601–8.
15. Barlow WE, White E, Ballard-Barbash R, Voak PM, Titus-Ernstoff L, Carney PA, et al. Prospective breast cancer risk prediction model for women undergoing screening mammography. J Natl Cancer Inst. 2006;98:1204–14.
16. Boyle P, Mezetti M, La Vecchia C, Franceschi S, Decarli A, Robertson C. Contribution of three components to individual cancer risk predicting breast cancer risk in Italy. Eur J Cancer Prev. 2004;13:189–95.
17. Colditz GA, Rosner B. Cumulative risk of breast cancer to age 70 years according to risk factor status: data from the Nurses’ Health Study. Am J Epidemiol. 2000;152:950–64.
18. Rosner B, Colditz GA. Nurses’ health study: log-incidence mathematical model of breast cancer incidence. J Natl Cancer Inst. 1996;88:359–64.
19. Pfeiffer RM, Park Y, Kremier AR, Lacey Jr JV, Pave D, Greenlee RT, et al. Risk prediction for breast, endometrial, and ovarian cancer in white women aged 50 y or older: derivation and validation from population-based cohort studies. PLoS Med. 2013;10:e1001492.
20. Liberman L, Menell JH. Breast imaging-reporting and data system (BI-RADS). Radiol Clin North Am. 2002;40:409–30.
21. Bumsides EA, Chhatwal J, Atagaz O. What is the optimal threshold at which to recommend breast biopsy? PLoS One. 2012;7:e48820.
22. American College of Radiology. Breast imaging-reporting and data system (BI-RADS). 4th ed. Reston, VA: American College of Radiology; 2003.
23. Weaver DL, Rosenberg BD, Barlow WE, Ichikawa L, Carney PA, Kierlikowska K, et al. Pathologic findings from the Breast Cancer Surveillance Consortium: population-based outcomes in women undergoing biopsy after screening mammography. Cancer. 2006;106:732–42.
24. Orel Sic, Kay R, Reynolds C, Sullivan DC. BI-RADS categorization as a predictor of malignancy. Radiology. 1999;211:845–50.
25. Lazarus E, Mainiero MB, Schepps B, Koelker SL, Livingston LS. BI-RADS lexicon for US and mammography: interobserver variability and positive predictive value. Radiology. 2006;239:885–91.
26. Popack SP, Tosteson AN, Grove MR, Wells WA, Carney PA. Mammography in 53,803 women from the New Hampshire mammography network. Radiology. 2000;217:832–40.
27. Lo JY, Marley MK, Baker JA, Yale JF Jr CE. Cross-institutional evaluation of BI-RADS predictive model for mammographic diagnosis of breast cancer. AJR Am J Roentgenol. 2002;178:457–63.
28. Liberman L, Abramson AF, Squires BF, Glassman JR, Morris EA, Dershaw DD. The breast imaging reporting and data system: positive predictive value of mammographic features and final assessment categories. AJR Am J Roentgenol. 1998;171:35–40.
29. Baker JA, Kornmuth PJ, Floyd Jr CE. Breast Imaging Reporting and Data System standardized mammography lexicon: observer variability in lesion description. AJR Am J Roentgenol. 1996;166:773–8.
30. Flowers CI, O’Donoghue C, Moore D, Goss A, Kim D, Kim JH, et al. Reducing false-positive biopsies: a pilot study to reduce benign biopsy rates for BI-RADS 4A/B assessments through testing risk stratification and new thresholds for intervention. Breast Cancer Res Treat. 2013;139:769–77.
31. Armstrong K, Handorf EA, Chen J, Bristol Demeter MN. Breast cancer risk prediction and mammography biopsy decisions: a model-based study. Am J Prev Med. 2013;44:15–22.
32. The Breast Cancer Risk Assessment Tool. National Cancer Institute, Bethesda. 2011. www.cancer.gov/bcrisktool. Accessed 1 Nov 2013.
33. Cox A, Dunning AM, Garcia-Closas M, Balabramanani S, Reed MW, Poolay KA, et al. A common coding variant in CASP8 is associated with breast cancer risk. Nat Genet. 2007;39:352–8. A published erratum appears in Nat Genet. 2007;39:668.
34. Stacey SN, Manolias A, Sulem P, Rafnar T, Gudmundsson J, Gudjonsson SA, et al. Common variants on chromosomes 2q35 and 16q12 confer susceptibility to estrogen receptor-positive breast cancer. PLoS One. 2012;7:e48820.
35. Easton DF, Pooley KA, Dunning AM, Pharoah PD, Thompson D, Ballinger DG, et al. Genome-wide association study identifies novel breast cancer susceptibility loci. Nature. 2007;447:1087–93.
36. Hunter DJ, Kraft P, Jacobs KB, Cox DG, Yeager M, Hankinson SE, et al. A genome-wide association study identifies alleles in FTO associated with risk of sporadic postmenopausal breast cancer. Nat Genet. 2007;39:870–4.
37. Stacey SN, Manolias A, Sulem P, Thorlacius S, Gudjonsson SA, Jonsson GF, et al. Common variants on chromosome 5p12 confer susceptibility to estrogen receptor-positive breast cancer. Nat Genet. 2008;40:703–6.
38. Garcia-Closas M, Hall P, Nevanlinna H, Pooley K, Morrison J, Richesson DA, et al. Heterogeneity of breast cancer associations with five susceptibility loci by clinical and pathological characteristics. PLoS Genet. 2008;4:e1000504.
39. Thomas G, Jacobs KB, Kraft P, Yeager M, Wacholder S, Cox DG, et al. A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 14q24.1 (RADS1/L1). Nat Genet. 2009;41:579–84.
40. Ahmed S, Thomas G, Ghousaini M, Healey CS, Humphreys MK, Platte R, et al. Newly discovered breast cancer susceptibility loci on 3p24 and 17q23.2. Nat Genet. 2009;41:585–90.
41. Fisher B, Costantino JP, Wickerham DL, Redmond CK, Kavanah M, Cronin WM, et al. Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. J Natl Cancer Inst. 1998;90:1371–88.
42. Vogel VG, Costantino JP, Wickerham DL, Cronin WM, Cecchini RS, Atkins JN, et al. Effects of tamoxifen vs raloxifene on the risk of developing invasive breast cancer and other disease outcomes: the NSABP Study of Tamoxifen and Raloxifene (STAR) P-2 trial. JAMA. 2006;295:2727–41.
43. Hosmer DW, Lemeshow S. Applied logistic regression. New York: Wiley; 1989.
44. Park JH, Gail MH, Greene MH, Chatterjee N. Potential usefulness of single nucleotide polymorphisms to identify persons at high risk cancer: an evaluation of seven common cancers. J Clin Oncol. 2012;30:2157–62.
45. Warren Anderssen S, Trentham-Dietz A, Gangnon RE, Hampton JM, Figueroa JD, Skinner HG, et al. The associations between a polygenic score, reproductive and menstrual risk factors and breast cancer risk. Breast Cancer Res Treat. 2013;140:247–34.
46. Chen J, Pee D, Ayyagari R, Graubard B, Schaier C, Byrne C, et al. Projecting absolute invasive breast cancer risk in white women with a model that includes mammographic density. J Natl Cancer Inst. 2006;98:1215–26.
47. Mazouni C, Sniege N, Rouzier R, Balleyguier C, Bevers T, Andre F, et al. A new risk model for breast cancer: validation in a population-based sample of post-menopausal women. Breast Cancer Res Treat. 2008;111:765–76.
48. Mazouni C, Sniege N, Rouzier R, Balleyguier C, Bevers T, Andre F, et al. Prediction of the Breast Cancer Risk Model in a Population-Based Sample of Post-Menopausal Women. J Natl Cancer Inst. 2010;102:220–4.
49. Weaver DL, Vacek PM, Skelly JM, Geller BM. Predicting biopsy outcome after mammography: what is the likelihood the patient has invasive or in situ breast cancer? Ann Surg Oncol. 2005;12:660–73.
50. Suzuki R, Orsini N, Saji S, Key TJ, Wolk A. Body weight and incidence of breast cancer: evidence based on 26 studies. PLoS One. 2013;8:e69056.
51. Chen F, Chen GK, Millikan RC, John EM, Ambrosone CB, Bernstein L, et al. Projecting the breast cancer risk: evidence based on 26 studies. PLoS One. 2013;8:e72526.
52. Weaver DL, Vacek PM, Skelly JM, Geller BM. Predicting biopsy outcome after mammography: what is the likelihood the patient has invasive or in situ breast cancer? Ann Surg Oncol. 2005;12:660–73.
53. Chene J, Pee D, Ayyagari R, Graubard B, Schaier C, Byrne C, et al. Projecting absolute invasive breast cancer risk in white women with a model that includes mammographic density. J Natl Cancer Inst. 2006;98:1215–26.
54. Chen J, Pee D, Ayyagari R, Graubard B, Schaier C, Byrne C, et al. Projecting absolute invasive breast cancer risk in white women with a model that includes mammographic density. J Natl Cancer Inst. 2006;98:1215–26.
55. Ruiz-Narvaez EA, Rosenberg L, Cozier YC, Adams-Campbell LL, et al. Lack of association between common single nucleotide polymorphisms on chromosome 5p12 and breast cancer risk. PLoS One. 2013;8:e72154.
56. Ruiz-Narvaez EA, Rosenberg L, Rotimi CN, Adewunmi AA, Ogundiran TO, Adebamowo C, Dennis J, Milne RL, et al. ULTRAFB: a large-scale genotyping identifies a new low breast cancer risk SNP. Cancer Epidemiol Biomarkers Prev. 2013;22:1250–6.