White Blood Cell Count and Serum Cytokine Profile in Tropical Hardwood Workers in Kumasi

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Background. Occupational exposure to wood dust particles has long been reported of its associated varying degrees of negative health effects due to different extractive chemicals present in the various timber species. However, tropical hardwood is also reported to have higher levels of extractive chemicals of antihistamine, antioxidant, and anti-inflammatory properties. In Ghana, woodworkers have for years been exposed to wood dust from mixed tropical hardwood species, with little or no protective equipment such as nose masks, yet with less significant respiratory conditions. This study seeks to investigate the serum cytokine profile in tropical hardwood workers in Kumasi to provide a better understanding of the immunoregulatory pattern activated in the woodworkers.

Method. The study was carried out among woodworkers, teachers, and security men located in Kumasi. A cross-sectional sampling of adult male workers was selected to participate in the study (86 woodworkers and 89 nonwoodworkers). Participants donated blood collected by venepuncture into EDTA tubes and spun to separate serum for cytokine assay. Cytokines including IFN-gamma, IL-1β, IL-2, IL-4, IL-6, IL-10, IL-12, IL-13, and IL-17 were assayed using the Human Premixed Multianalyte Kit (R&D System, Inc., Minneapolis, USA) following the manufacturer’s procedure. The cytokine levels were quantified using the Luminex®200 analyser.

Results. The mean concentration levels for the various cytokines were significantly different (p < 0.05) between woodworkers and nonwoodworkers except IL-2. There were significantly increased levels of Th1 and Th2 cytokines expressed in the woodworkers more than the nonwoodworkers.

Conclusions. The results from this study reveal that exposed woodworkers of mixed tropical hardwood species show a high level of Th1 and Th2 cytokines in their serum than nonwoodworkers.

1. Introduction

Cytokines are a diverse group of peptides produced by different cell types including cancer cells and various subpopulations of immune system cells. They exert the effect as cellular mediators after binding to appropriate membrane receptors [1]. They are synthesized by both stationed and moving cells including mast cells, macrophages, lymphocytes, basophils, eosinophils, and neutrophils that have been activated [2]. Cytokines participate in different physiological and pathological processes: including the regulation of complex immune responses to pathogens, as well as inflammatory responses as they regulate their duration and intensity [1]. Cytokine plays the role of balancing communications between diverse cells in complex immune system responses, by transmitting signals from one cell type to another; necessary for cell activation, cell suppression, and a regulatory function [1].

Several classifications of cytokines have been proposed. The simple classification used by most authors are based on the cell type that produces the individual cytokines [1]. Other classifications group cytokines based on the potential to stimulate cells of the immune system, also classified as Th1 cytokines (IL-2, IL-12, IL-18, IL-21, IL-23, and IFN-
cytokines with inhibitory effects on the cell immune system also classified as anti-inflammatory cytokines (TNF-α and IL-10); cytokines that effect the regulatory balance of cells of the immune system (IL-10 and TGF-β); Th2 cytokines are classic mediators of the immune response which recognize extracellular parasite and promote IgE and eosinophilic response in atopy (IL-4, IL-5, IL-10, and IL-13) [1, 3, 4]; proinflammatory cytokines stimulate inflammation and play a significant role in immune- oncology [1].

Infiltration of inflammatory cells such as mast cells, lymphocytes, eosinophils, neutrophils, basophils, and macrophages to the site of inflammation has been identified in inflammatory diseases such as allergic rhinitis asthma and bronchitis in temperate woodworkers [5, 6]. Each of these cell types has a distinct role in the immune system and communicates with other immune cells using secreted cytokines [7]. Some temperate softwood and hardwood studied have detected all the 9 cytokines (IFN-γ, IL-1β, IL-2, IL-4, IL-6, IL-10, IL-12, IL-13, and IL-17) simultaneously in a single sample. Each kit contained Human Standard Cocktail, Human Premixed Microparticle Cocktail, Human Premixed Biotin-Ab Cocktail, Streptavidin–Phytoerythrin (PE) Concentrate, Diluent, Calibrator Diluent, Wash Buffer Concentrate, 96-well Microplate, Mixing Bottles Plate Sealer, and a Certificate of Analysis.

All the reagents and samples were brought to room temperature (RT) and an assay was performed at RT. The 96-well filter-bottomed microplate was prefilted by filling each well with 100 μl of wash buffer and removed using a vacuum manifold designed to accommodate a microplate. In each well, 50 μl of a diluted cocktail of microparticles was added to the prefilted bottom microplate. In addition, 50 μl of the standard cocktail was added to the first 6 wells, while to each of the rest of the wells, 50 μl of each sample was added and all securely covered with a foil plate. It was then incubated for 2 hours at room temperature on the horizontal orbital microplate shaker set at 500 rpm. Using the vacuum manifold device, the microplate was washed by first removing the liquid, refilling each well with 100 μl of wash buffer, and again removing the liquid. The washing procedure was repeated 3 times. After washing, 50 μl of diluted biotin antibody cocktail was added to each well, and the plate was securely covered with a new foil plate sealer and incubated for 1 hour at room temperature on the shaker set at 500 rpm. Again, the liquid was removed, and the washing procedure was repeated three times as described above. After washing, 50 μl of diluted Streptavidin–PE was added to each well, securely covered with a new foil plate sealer, and incubated for 30 minutes at room temperature on the shaker set at 500 rpm.

The liquid was removed, and washing was repeated three times as before. The microparticles were resuspended by adding 100 μl of wash buffer to each well. The microplate was incubated for 2 minutes at room temperature on the shaker set at 500 rpm. It was read using the Lumex®200 (manufacturer: Lumex Corporation, United States of America) analyser within 30 minutes. The procedure was used to measure cytokine levels for all the samples. Cytokine concentrations were determined from standard curves prepared on each plate and expressed in picogram per millilitre (pg/ml).
2.5. Statistical Analysis. At the end of the study, the field data was entered directly into Microsoft Excel 2010 statistical package and analysed using STATA version 14. The level of statistical significance was set at a p value < 0.05. Statistical analysis was conducted using Student’s t-test to compare the inflammatory cell counts and the plasma cytokine levels between the exposed woodworkers and nonexposed workers. Spearman rank correlation was used to determine the correlation between the cytokines and the ages of woodworkers. Also, a correlation between the white blood cells and the cytokines was made. All continuous variables were expressed as means ± standard deviation.

3. Result

3.1. Comparison of Inflammatory Cell Counts between Study Populations. The total white blood cells, as well as the differential counts of lymphocytes, neutrophils, and platelets, were compared between the woodworkers and the nonexposed workers using Student’s t-test. The total white blood cells, neutrophils, and platelets count were significantly higher in the woodworkers than the nonexposed workers, even though the total white blood cells were only marginally significant. However, the difference in the lymphocyte counts between the woodworkers and the nonexposed workers was not significant (Table 1).

3.2. Comparison of Cytokine Levels Expressed in Woodworkers with Nonexposed Workers. The level of cytokines expressed in the woodworkers and nonexposed workers is shown in Table 2. The Student’s t-test was used to compare the mean levels of the various cytokines studied at (p < 0.05). Except for IL-2 which was insignificantly higher, all the other cytokines (IL-1β, IL-4, IL-6, IL-10, IL-12, IL-13, IL-17, and IFN-γ) studied were significantly higher in the woodworkers than the nonexposed workers.

3.3. Determining the Classes of Cytokines Measured. The classes of cytokines measured with a significant difference in the woodworkers are shown in italics in Table 2. The major proinflammatory cytokine (IL-1β) and major CD4+ effect subsets cytokines: Th1 (IFN-γ and IL-12p70), Th2 (IL-4 and IL-13) Treg (IL-10), and Th17 (IL-17) cytokines were all significantly higher in the woodworkers.

3.4. Correlation between Cytokines and Ages of Woodworkers. In investigating the correlations between cytokines and the ages of woodworkers in years, Spearman rank correlation was used at 5% significance level.

Table 3 shows that although there is a weak correlation between the various cytokines and the ages of woodworkers, none of the relationships is statistically significant (p > 0.05).

3.5. Ethical Issue. The study was carried out in accordance with the guidelines and principles of the Helsinki’s Declaration of research involving human participants. The basic ethical principles such as respect for persons, beneficence, and justice were adhered to in the performance of the study. The study received ethical approval from the Committee on Human Research, Publication and Ethics (CHRPE) of the School of Medical Sciences of KNUST and Komfo Anokye Teaching Hospital (KATH) Kumasi (Ref: CHRPE/AP/304/15). A written request indicating the purpose and benefits of the research was sent to the leadership of each of the participating group to seek their permission to conduct research among their members. A durbar was held at each of the sites during which the purpose and benefits of the research was explained to them in English and the Ghanaian Language (Twi). All the participants completed informed consent form and received date to return for commencement of study. The participation in the study was voluntary, and respondents could withdraw at any stage of the data collection process. The anonymity of participants was ensured by using codes for individual answered questionnaire received.

4. Discussion

4.1. Comparison of Inflammatory Cell Counts. Increased inflammatory cell and platelet count in the peripheral blood is an indication of inflammatory reaction in the host. In the present study, the total white blood cell, neutrophil, and platelet counts were higher in the woodworkers. This is consistent with the result of Gripenbäck et al. [14] who found a significantly increased number of neutrophils in the peripheral blood of healthy volunteers who were exposed to pine-wood dust. The lymphocyte count was not significantly high in the woodworkers. This result is consistent with the findings of Gripenbäck et al. [14] who found significant decreased lymphocyte numbers in the peripheral blood of healthy volunteers exposed to wood dust and also found increased lymphocytes in the bronchoalveolar lavage (BAL) fluid. The increased inflammatory cells in the peripheral blood in this study suggest that the wood dust may have induced inflammatory reactions in the woodworkers.

4.2. Comparison of Cytokine Levels among Woodworkers and Nonexposed Workers. Cytokines are inflammatory and immunomodulating mediators that exhibit both negative and positive regulatory effects on various target cells [15]; hence, the serum levels of various cytokines may give information on the presence of inflammatory processes [13]. This study investigated the cytokine profile of the different classes of cytokines in the woodworkers.

Proinflammatory cytokines promote inflammation by activating a variety of proinflammatory genes, including phospholipase A2, cyclooxygenase 2 (COX-2), inducible
Th1 cell lineage [19, 20]. This proinflammatory reaction in the woodworkers may contribute to resisting allergic inflammatory reactions that may be induced due to the increased Th2 level in the woodworkers.

Table 2: Result of compared cytokine levels between woodworkers and nonexposed workers.

| Cytokine     | Woodworker (mean ± SD) | Nonexposed worker (mean ± SD) | Mean difference | 95% CI     | T       | p value  |
|--------------|------------------------|-------------------------------|----------------|------------|---------|----------|
| IL-1β        | 10.26 (±2.59)          | 8.86 (±2.68)                 | 1.40           | 0.60–2.19  | 3.47    | 0.0006***|
| IL-2         | 21.68 (±5.74)          | 20.17 (±5.41)                | 1.51           | -0.15–3.17 | 1.79    | 0.0758   |
| IL-4         | 27.86 (±6.27)          | 19.55 (±5.52)                | 8.31           | 6.54–10.07 | 9.29    | <0.001***|
| IL-6         | 9.34 (±3.46)           | 7.32 (±4.39)                 | 2.01           | 0.78–3.24  | 3.24    | 0.0015** |
| IL-10        | 22.40 (±6.17)          | 18.63 (±6.80)                | 3.77           | 1.82–5.71  | 3.83    | 0.0002***|
| IL-12p70     | 18.29 (±4.53)          | 13.12 (±3.78)                | 5.17           | 3.93–6.42  | 8.22    | <0.001***|
| IL-13        | 7.17 (±1.77)           | 6.25 (±1.76)                 | 0.93           | 0.40–1.45  | 3.47    | 0.0007***|
| IL-17A       | 13.30 (±2.66)          | 10.99 (±2.71)                | 2.30           | 1.50–3.10  | 5.68    | <0.001***|
| INF-γ        | 18.63 (±4.39)          | 13.18 (±5.28)                | 5.45           | 3.99–6.91  | 7.39    | <0.001***|

*p values in parentheses; *p < 0.05, **p < 0.01, ***p < 0.001; SD: standard deviation.

Table 3: Correlation between cytokines and ages of woodworkers.

| Cytokine  | Age (year) | p value  |
|-----------|------------|----------|
| IL-6      | 0.019      | (0.8060) |
| IL-10     | -0.022     | (0.7768) |
| IL-1β     | 0.065      | (0.3976) |
| INF-γ     | 0.115      | (0.1310) |
| IL-4      | 0.085      | (0.2629) |
| IL-17A    | 0.070      | (0.3569) |
| IL-2      | -0.011     | (0.8894) |
| E-selectin| 0.003      | (0.9666) |
| IL-13     | 0.017      | (0.8214) |
| GCSF      | 0.068      | (0.3750) |
| IL-12p70  | 0.135      | (0.0747) |
| CD40L     | -0.019     | (0.8060) |
| Eotaxin   | 0.063      | (0.4118) |

*p values in parentheses; *p < 0.05, **p < 0.01, ***p < 0.001. Values were based on Spearman rank correlation.

Nitric oxide synthase (iNOS), and other cytokines and chemokines to initiate inflammation [16–18]. IL-1β and IL-6 also promote the development of T helper (Th17) cells and Th1 cell lineage [19, 20]. The proinflammatory cytokines (IL-1β and IL-6) were increased in the woodworkers in this study, which suggests that the wood dust induces proinflammatory cytokines and therefore may promote inflammatory reaction in the woodworkers. This observation is similar to the result of Long et al. [9], Bornholdt et al. [8] and Määttä et al. [6] in which wood dust of all the tested wood species induced proinflammatory cytokine expression in the human lung tissue and animal model studies. The present study, therefore, supports the previous observation that the wood dust induces an inflammatory response in the woodworkers.

The CD4+ effector subset Th1, Th2, Th17, and Treg cytokines were higher in the woodworkers, which suggests that the wood dust may have induced Th1, Th2, Th17, and Treg immune response in the woodworkers. However, no significant changes have been reported in previous studies on the expression of Th1, Th2, Th17, and Treg cytokines in woodworkers, human tissue, or animals exposed to wood dust [10, 21]. Again, there were no studies reported on CD4+ effector subset cytokine induction in woodworkers exposed to tropical hardwood.

The Th1 cytokines (INF-γ and IL-12) are known to mediate the killing of intracellular pathogens by activating monocytes and macrophages to increase their cytokine secretion and antigen presentation. However, they also resist the Th2 cell cytokine function and Th2 response (allergic response) [3, 4]. Th1 cytokines also downregulate eosinophil differentiation by suppressing the development of Th2 cells in allergic inflammation [22]. This study, therefore, suggests that the increased Th1 cytokine levels observed in the woodworkers may contribute to resisting allergic inflammatory reactions that may be induced due to the increase Th2 level in the woodworkers.

This is consistent with the findings that altering the cytokine-producing profile of Th2 cells by inducing Th1 responses is protective against Th2-related disorders such as asthma and allergy [23]. This study suggests that the Th1 response induced by the wood dust may contribute to resisting allergic response in woodworkers. This may have contributed to the normal lung function indices observed in the woodworkers in our study (not published).

The Th2 cytokines (IL-4 and IL-13) were higher in the woodworkers compared to the nonexposed workers, suggesting that the wood dust may have contributed to induce of Th2 response in the woodworkers. Th2 cytokines are important in hypersensitivity reactions and allergic immunopathology. They are known to enhance mucus release, class switching of B cells to produce IgE, and fibrosis [14]. The action of the Th2 immune response can result in IgE production, inflammation of the airways, and tissue remodelling [22]. Although IgE was not measured in the present study, the increased Th2 cytokines may support the high level of IgE found in a previous study of exposed tropical hardwood workers in Accra [24]. The increased Th2 cytokines in the woodworkers in the present study may have contributed to allergic respiratory symptoms such as sneezing and catarrh and mucous release woodworkers. This is consistent with the findings of Gripenbäck et al. [14] and Shum et al. [25] that elevated production of the Th2 cytokines contributes to allergic airway inflammation. The high
levels of the Th1 cytokines (IFN-γ and IL-12) observed in the present study may be attributed to the high level of Th2 cytokines (IL-4) to antagonize the development of Th2 response and downregulate the Th2 inflammatory reactions in the woodworkers [4, 25].

The Th17 cytokine IL-17A was higher in the woodworkers suggesting that the wood dust may have induced a Th17 response in the woodworkers. IL-17A is known to stimulate the production of other proinflammatory cytokines and play protective roles in host resistance against pathogens at epithelial and mucosal barriers [26, 27]. Elevated levels of IL-17A have been observed in the airways of asthmatics and are associated with neutrophil influx [25]. In the present study, the IL-17A level was higher in the woodworkers, this indicating that IL-17A may contribute to the induction of inflammation reaction in the woodworkers.

The woodworkers had a higher Treg cytokine (IL-10) level than the nonexposed workers suggesting an increased Treg response in the woodworkers. Activated Treg produces IL-10 which inhibits the synthesis of proinflammatory cytokines or suppresses their activities, hence negatively modulating inflammatory response [28]. It also reduces airway hyperreactivity (AHR), lung eosinophil infiltration, and Th2 cytokine production which are characteristics of allergic airway inflammation. IL-10 together with IL-4 and IL-13 downregulates immunological response in the lungs [3, 4, 29, 30]. The higher level of Treg cytokine in the present study suggests that the increased Treg cytokine in the woodworkers may contribute to the suppression of allergic and inflammatory reactions induced by wood dust to limit the immunopathological occurrence in the woodworkers.

4.3. Correlation between Cytokines and Ages of Woodworkers. In previous studies, cytokines have been linked to the aging process, inflammatory cytokines such IL-1β, IL-6, and TNF-α have been recorded to increase with aging [31–33]. The present study shows a nonsignificant positive correlation of cytokine (IL-1β, IFN-γ, IL-4, IL-6, IL-13, IL-17A, and IL-12p70) levels with the age of the woodworkers and a nonsignificant negative correlation of cytokine (IL-10 and IL-2) levels with the age of the woodworkers. Could this be a result of the wood dust stimulating inflammatory suppressors in the woodworkers?

5. Conclusion

The present study reveals that mixed tropical hardwood dust may contribute to inducing allergic inflammatory responses in the woodworkers which are evident by the elevated inflammatory cells and proinflammatory and Th2 cytokines. Moreover, the wood dust may also contribute to inhibiting allergic responses and inflammatory responses which may be induced in the woodworkers; this is evident by the high level of Th1 and Treg cytokines which are known to antagonize inflammatory and allergic responses and the nonsignificant positive correlation of inflammatory cytokines and age of woodworkers.

Data Availability

The datasets used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no competing interests.

Authors’ Contributions

IEE and MAF made contributions to the conception and design of the study. DD, FAY, and IEE made a substantial contribution to the study design and management of the research activities. IEE and RSM analysed the data and drafted the manuscript. All authors were involved in critical revision for important intellectual content and approved the final manuscript.

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References

[1] V. Jurisic, "Multiomic analysis of cytokines in immuno-oncology," Expert Review of Proteomics, vol. 17, no. 9, pp. 663–674, 2020.
[2] J. B. Calixto, M. M. Campos, M. F. Otuki, and A. R. Santos, “Anti-inflammatory compounds of plant origin. Part II. Modulation of pro-inflammatory cytokines, chemokines and adhesion molecules,” Planta Medica, vol. 70, no. 2, pp. 93–103, 2004.
[3] B. Moldoveanu, P. Otmishi, P. Jani et al., “Inflammatory mechanisms in the lung,” Journal of Inflammation Research, vol. 2, pp. 1–11, 2009.
[4] J. R. Murdoch and C. M. Lloyd, “Chronic inflammation and asthma,” Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis, vol. 690, no. 1-2, pp. 24–39, 2010.
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[5] C. Owen, “Chemokine receptors in airway disease: which receptors to target?,” Pulmonary Pharmacology & Therapeutics, vol. 14, no. 3, pp. 193–202, 2001.

[6] J. Määttä, Cytokine, Chemokine, and Chemokine Receptor Expression in RAW 264.7 Mouse Macrophages and in the Lungs of Mice After Exposure to Wood Dust, University of Kuopio, 2008.

[7] J. B. Lange, Effects of Wood Dust: Inflammation, Genotoxicity and Cancer (Doctoral Dissertation, Museum Tusculanum), National Research Centre for the Working Environment. Faculty of Health Sciences, 2008.

[8] J. Bornholdt, J. Hansen, T. Steiniche et al., “K-ras mutations in sinonasal cancers in relation to wood dust exposure,” BMC Cancer, vol. 8, no. 1, p. 53, 2008.

[9] H. Long, T. Shi, P. J. Born et al., “ROS-mediated TNF-alpha and MIP-2 gene expression in alveolar macrophages exposed to pine dust,” Particle and Fibre Toxicology, vol. 1, no. 1, p. 3, 2004.

[10] J. Määttä, R. Haapakoski, M. Lehto et al., “Immunomodulatory effects of oak dust exposure in a murine model of allergic asthma,” Toxicological Sciences, vol. 99, no. 1, pp. 260–266, 2007.

[11] P. Rathee, H. Chaudhary, S. Rathee, D. Rathee, V. Kumar, and K. Kohli, “Mechanism of action of flavonoids as anti-inflammatory agents: a review,” Inflammation & Allergy-Drug Targets (Formerly Current Drug Targets-Inflammation & Allergy) (Discontinued), vol. 8, no. 3, pp. 229–235, 2009.

[12] A. C. Toledo, C. P. P. Sakoda, A. Perini et al., “Flavonone treatment reverses airway inflammation and remodelling in an asthma murine model,” British Journal of Pharmacology, vol. 168, no. 7, pp. 1736–1749, 2013.

[13] J. M. Cavaillon, “Pro-versus anti-inflammatory cytokines: myth or reality,” Cellular And Molecular Biology-Paris-Wegmann-, vol. 47, no. 4, pp. 695–702, 2001.

[14] S. Gripenbäck, L. Lundgren, A. Eklund et al., “Accumulation of eosinophils and T-lymphocytes in the lungs after exposure to pinewood dust,” European Respiratory Journal, vol. 25, no. 1, pp. 118–124, 2005.

[15] M. Sultani, A. M. Stringer, J. M. Bowen, and R. J. Gibson, “Anti-inflammatory cytokines: important immunoregulatory factors contributing to chemotherapy-induced gastrointestinal mucositis,” Chemotherapy research and practice, vol. 2012, Article ID 490804, 11 pages, 2012.

[16] R. N. Apte and E. Voronov, “Interleukin-1—A Major Pleiotropic Cytokine in Tumor–Host Interactions,” in Seminars in cancer biology, vol. 12, no. 4pp. 277–290, Academic Press, 2002.

[17] C. A. Dinarello, “Proinflammatory cytokines,” Chest, vol. 118, no. 2, pp. 503–508, 2000.

[18] J. P. Mizgerd, “Molecular Mechanisms of Neutrophil Recruitment Elicited by Bacteria in the Lungs,” in Seminars in immunology, vol. 14, no. 2pp. 123–132, Academic Press, 2002.

[19] J. V. Aldrich and S. C. Vigil-Cruz, Chemokine and Cytokine Modulators. Burger’s Medicinal Chemistry and Drug Discovery, John Wiley & Sons, New York, 6th ed. edition, 2003.

[20] L. C. Borish and J. W. Steinke, “2. Cytokines and chemokines,” Journal of Allergy and Clinical Immunology, vol. 111, no. 2, pp. S460–S475, 2003.

[21] J. Määttä, M. Lehto, M. Leino et al., “Mechanisms of particle-induced pulmonary inflammation in a mouse model: exposure to wood dust,” Toxicological Sciences, vol. 93, no. 1, pp. 96–104, 2006.

[22] L. M. Teran, M. Mochizuki, J. Bartels et al., “Th1- and Th2-type cytokines regulate the expression and production of eotaxin and RANTES by human lung fibroblasts,” American Journal of Respiratory Cell and Molecular Biology, vol. 20, no. 4, pp. 777–786, 1999.

[23] L. K. Teixeira, B. P. Fonseca, B. A. Barboza, and J. P. Viola, “The role of interferon-gamma on immune and allergic responses,” Memórias do Instituto Oswaldo Cruz, vol. 100, suppl 1, pp. 137–144, 2005.

[24] E. I. Ennin, The Effect of Wood Dust on the Lung Function of Woodworkers at the Accra Timber Market [PhD Thesis], College of the Health Sciences University of Ghana, 2009.

[25] B. O. Shum, M. S. Rolph, and W. A. Sewell, “Mechanisms in allergic airway inflammation—lessons from studies in the mouse,” Expert Reviews in Molecular Medicine, vol. 10, p. e15, 2008.

[26] W. Jin and C. Dong, “IL-17 cytokines in immunity and inflammation,” Emerging microbes & infections, vol. 2, no. 9, article e60, 2013.

[27] R. M. Onishi and S. L. Gaffen, “Interleukin-17 and its target genes: mechanisms of interleukin-17 function in disease,” Immunology, vol. 129, no. 3, pp. 311–321, 2010.

[28] S. M. Opal and V. A. Depalo, “Anti-inflammatory cytokines,” Chest, vol. 117, no. 4, pp. 1162–1172, 2000.

[29] S. Baldacci, E. Omenaas, and M. P. Oryszczyn, “Allergy markers in respiratory epidemiology,” European Respiratory Journal, vol. 17, no. 4, pp. 773–790, 2001.

[30] J. S. Erjefält, “Mast cells in human airways: the culprit?,” European Respiratory Review, vol. 23, no. 133, pp. 299–307, 2014.

[31] H. Bruunsgaard and B. K. Pedersen, “Age-related inflammatory cytokines and disease,” Immunology and Allergy Clinics of North America, vol. 23, no. 1, pp. 15–39, 2003.

[32] M. Michaud, L. Balardy, G. Moulis et al., “Proinflammatory cytokines, aging, and age-related diseases,” Journal of the American Medical Directors Association, vol. 14, no. 12, pp. 877–882, 2013.

[33] E. J. Morley and N. B. Richard, “Cytokine-related aging process,” The Journals of Gerontology Series A: Biological Sciences and Medical Sciences, vol. 59, no. 9, pp. M924–M929, 2004.

[34] I. E. Ennin, M. A. Frempong, D. Dodoo, F. A. Yeboah, and R. S. E. Maalman, White Blood Cell Count and Serum Cytokine Profile in Tropical Hardwood Workers in Kumasi (Preprint), researchsquare, 2022.