Safety of hydrothermally treated kernels from edible Jatropha curcas L. (Chuta) as a novel food pursuant to Regulation (EU) 2015/2283

EFSA Panel on Nutrition, Novel Foods, Food Allergens (NDA), Dominique Turck, Torsten Bohn, Jacqueline Castenmiller, Stefaan De Henauw, Karen Ildico Hirsch-Ernst, Alexandre Maciuk, Inge Mangelsdorf, Harry J Mc Ardle, Androniki Naska, Carmen Pe laez, Kristina Pentieva, Alfonso Siani, Frank Thies, Sophia Tsabouri, Marco Vinceti, Francesco Cubadda, Thomas Frenzel, Marina Heinonen, Rosangela Marchelli, Monika Neu häuser-Berthold, Morten Poulsen, Miguel Prieto Maradona, Josef Rudolf Schlatter, Henk van Loveren, Paolo Colombo and Helle Katrine Knutsen

Abstract

Following a request from the European Commission, the EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA) was asked to deliver an opinion on hydrothermally treated kernels from edible Jatropha curcas (Chuta) as a novel food (NF) pursuant to Regulation (EU) 2015/2283. Although Jatropha curcas is generally considered a toxic plant due to the presence of phorbol esters (PEs), edible varieties exist in Central America. The applicant has developed a breeding programme for an edible cultivar and proposes the kernels from this cultivar as an NF as whole kernels or fragments thereof to be used as a snack or as a food ingredient. Procedures are in place to avoid commingling with non-edible kernels, with the last steps being the analytical control of PEs concentrations in all produced batches. The Panel considers that the production process of the NF is sufficiently described and that the information provided on the composition of the NF is sufficient for its characterisation. Components of the NF were tested for genotoxicity applying the standard in vitro test battery and no genotoxic concerns have been identified. In a conservative scenario for exposure to PEs from the NF, it was assumed that all kernels contain PEs at the level of detection of the analytical method. When comparing the estimated maximum exposure to PEs with a reference point from a subchronic study in pigs, a margin of exposure ≥ 900 is obtained, which is considered sufficiently large. The presence of anti-nutritional factors does not pose safety concerns as they are within the ranges found in vegetables. The Panel concludes that the NF is safe under the proposed conditions of use.

© 2022 European Food Safety Authority. EFSA Journal published by John Wiley and Sons Ltd on behalf of European Food Safety Authority.

Keywords: edible Jatropha curcas kernels, Chuta, phorbol esters, hydrothermal treatment, anti-nutritional factors, novel food, safety

Requestor: European Commission
Question number: EFSA-Q-2018-00542
Correspondence: nda@efsa.europa.eu
Panel members: Dominique Turck, Torsten Bohn, Jacqueline Castenmiller, Stefaan De Henauw, Karen Ildico Hirsch-Ernst, Helle Katrine Knutsen, Alexandre Maciuk, Inge Mangelsdorf, Harry J McArdle, Androniki Naska, Carmen Pelaez, Kristina Pentieva, Alfonso Siani, Frank Thies, Sophia Tsabouri and Marco Vinceti.

Declarations of interest: The declarations of interest of all scientific experts active in EFSA’s work are available at https://ess.efsa.europa.eu/doi/doiweb/doisearch.

Acknowledgements: The Panel wishes to thank the Working Group on Compendium of Botanicals, Eirini Kouloura and Petra Gergelova for the support provided to this scientific output.

Suggested citation: EFSA NDA Panel (EFSA Panel on Nutrition, Novel Foods, Food Allergens), Turck D, Bohn T, Castenmiller J, De Henauw S, Hirsch-Ernst KI, Maciuk A, Mangelsdorf I, McArdle HJ, Naska A, Pelaez C, Pentieva K, Siani A, Thies F, Tsabouri S, Vinceti M, Cubadda F, Frenzel T, Heinonen M, Marchelli R, Neuhäuser-Berthold M, Poulsen M, Prieto Maradona M, Schlatter JR, van Loveren H, Colombo P and Knutsen HK, 2022. Scientific Opinion on the safety of hydrothermally treated kernels from edible Jatropha curcas L. (Chuta) as a novel food pursuant to Regulation (EU) 2015/2283. EFSA Journal 2022;20(1):6998, 26 pp. https://doi.org/10.2903/j.efsa.2022.6998

ISSN: 1831-4732

© 2022 European Food Safety Authority. EFSA Journal published by John Wiley and Sons Ltd on behalf of European Food Safety Authority.

This is an open access article under the terms of the Creative Commons Attribution-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited and no modifications or adaptations are made.
Table of contents

Abstract ................................................................................................................................................ 1
1. Introduction ........................................................................................................................................ 4
1.1. Background and Terms of Reference as provided by the requestor .............................................. 4
2. Data and methodologies ..................................................................................................................... 4
2.1. Data ................................................................................................................................................ 4
2.2. Methodologies ............................................................................................................................... 4
3. Assessment ......................................................................................................................................... 5
3.1. Introduction ..................................................................................................................................... 5
3.2. Identity of the NF ............................................................................................................................ 5
3.3. Production process .......................................................................................................................... 5
3.4. Compositional data ......................................................................................................................... 6
3.4.1. Proximate analysis ....................................................................................................................... 6
3.4.2. Phorbol esters, contaminants, microbiological and process contaminants .................................. 7
3.5. Stability .......................................................................................................................................... 9
3.6. Specifications .................................................................................................................................. 9
3.7. History of use of the NF and of its source ....................................................................................... 10
3.7.1. History of use of the source ........................................................................................................ 10
3.7.2. History of use of the NF ............................................................................................................ 11
3.8. Uses and use levels and anticipated intake .................................................................................... 11
3.8.1. Target population ....................................................................................................................... 11
3.8.2. Proposed uses and use levels ..................................................................................................... 12
3.8.3. Anticipated intake of the novel food .......................................................................................... 12
3.8.4. Estimate of exposure to undesirable substances ........................................................................ 12
3.9. Absorption, distribution, metabolism and excretion (ADME) ....................................................... 13
3.10. Nutritional information ............................................................................................................... 13
3.10.1. Fatty acids and amino acids composition, sugars and minerals ............................................... 13
3.10.2. Anti-nutritional factors ........................................................................................................... 14
3.11. Toxicological information ........................................................................................................... 15
3.11.1. Genotoxicity: studies conducted with the NF and Jatropha kernels ...................................... 16
3.11.2. Subacute toxicity .................................................................................................................... 16
3.11.3. Subchronic toxicity ................................................................................................................ 17
3.12. Human data .................................................................................................................................. 18
3.13. Allergenicity ................................................................................................................................. 18
4. Discussion ......................................................................................................................................... 19
5. Conclusions ....................................................................................................................................... 20
5.1. Protection of Proprietary data in accordance with Article 26 of Regulation (EU) 2015/2283 ......... 20
6. Steps taken by EFSA ....................................................................................................................... 20
References ........................................................................................................................................... 21
Abbreviations ......................................................................................................................................... 24
Appendix A ............................................................................................................................................ 26
1. Introduction

1.1. Background and Terms of Reference as provided by the requestor

On 29 August 2016, the company JatroSolutions GmbH submitted a request to the German Federal Office of Consumer protection and Food Safety (BVL) in accordance with Article 4 of Regulation (EC) 258/1997 to place on the EU market edible *Jatropha curcas* L. kernels (Chuta®) as a novel food ingredient.

Pursuant to article 35(1) of Regulation (EU) 2015/2283 any request for placing a novel food on the market within the Union submitted to a Member State in accordance with Article 4 of Regulation (EC) 258/1997 of the European Parliament and of the Council concerning novel foods and novel foods ingredients and for which the final decision has not been taken before 1 January 2018, shall be treated as an application submitted under Regulation (EU) 2015/2283.

In accordance with Article 10(3) of Regulation (EU) 2015/2283, the European Commission asks the European Food Safety Authority to provide a scientific opinion by carrying out the assessment for edible *Jatropha curcas* L. kernels (Chuta®) as a novel food.

2. Data and methodologies

2.1. Data

The safety assessment of this novel food (NF) is based on data supplied in the application, information provided by the EFSA Working Group on Compendium of Botanicals and information submitted by the applicant following EFSA requests for supplementary information.

During the assessment, the Panel identified additional data which were not included in the application (Panigrahi et al., 1984; Makkar and Becker, 1997; Garcia Estepa et al., 1999; Haas et al., 2002; Aregheore et al., 2003; Chivandi et al., 2006; Goel et al., 2007; Makkar et al., 2008; Rakshit et al., 2008; Martin-Cabrejas et al., 2009; Schlemmer et al., 2009; Shah and Sanmukhani, 2010; Chomchai et al., 2011; Baldini et al., 2014a; Langrand et al., 2015; Li et al., 2015; Gupta et al., 2016; Pal et al., 2017; Sabolová et al., 2017; Suvari et al., 2017; Vagadia et al., 2017; White, 2017; Faria-Machado et al., 2019; Nematollahi et al., 2020).

Administrative and scientific requirements for NF applications referred to in Article 10 of Regulation (EU) 2015/2283 are listed in the Commission Implementing Regulation (EU) 2017/2469.

A common and structured format on the presentation of NF applications is described in the EFSA guidance on the preparation and presentation of an NF application (EFSA NDA Panel, 2016). As indicated in this guidance, it is the duty of the applicant to provide all of the available (proprietary, confidential and published) scientific data (including both data in favour and not in favour) that are pertinent to the safety of the proposed NF.

This NF application includes a request for protection of proprietary data in accordance with Article 26 of Regulation (EU) 2015/2283. The data requested by the applicant to be protected comprise: compositional data including nutritional information and allergens, biological and process contaminants, management of Chuta cultivation, shelf-life of the NF, phorbol esters analytical methods, procedures for the verification of phorbol esters content, hydrothermal treatment, molecular markers, anticipated intake and toxicological information.

2.2. Methodologies

The assessment follows the methodology set out in the EFSA guidance on NF applications (EFSA NDA Panel, 2016) and the principles described in the relevant existing guidance documents from the EFSA Scientific Committee. The legal provisions for the assessment are laid down in Article 11 of Regulation (EU) 2015/2283 and in Article 7 of the Commission Implementing Regulation (EU) 2017/2469.

---

1 Regulation (EC) 258/97 of the European Parliament and of the Council of 27 January 1997 concerning novel foods and novel food ingredients. OJ L 43, 14.2.1997, pp. 1–6.
2 Regulation (EU) 2015/2283 of the European Parliament and of the Council on novel foods, amending Regulation (EU) No 1169/2011 of the European Parliament and of the Council and repealing Regulation (EC) No 258/97 of the European Parliament and of the Council and Commission Regulation (EC) No 1852/2001 (2013/0435 (COD), OJ L 327, 11.12.2015, pp. 1–22.
3 Commission Implementing Regulation (EU) 2017/2469 of 20 December 2017 laying down administrative and scientific requirements for applications referred to in Article 10 of Regulation (EU) 2015/2283 of the European Parliament and of the Council on novel foods. OJ L 351, 30.12.2017, pp. 64–71.
The information provided by the EFSA Working Group on Compendium of Botanicals is based on an extensive literature search on *Jatropha curcas*, following a search strategy and standard operating procedure as described by the University of Chemistry and Technology (UCT) of Prague (Dibusz and Vejvodova, 2020).

This assessment concerns only the risk that might be associated with consumption of the NF under the proposed conditions of use and is not an assessment of the efficacy of the NF with regard to any claimed benefit.

Dietary intake to the NF was estimated based on the proposed uses and use levels (see Section 3.8).

### 3. Assessment

#### 3.1. Introduction

The NF which is the subject of the application is the hydrothermally treated *Jatropha curcas* L. kernels (Chuta®), hereinafter referred as ‘Chuta’ or ‘Chuta kernel’. Chuta kernels are obtained from edible *Jatropha curcas* cultivars (called ‘EdibleNut’ cultivars) developed in the context of JatroSolutions breeding programme. The applicant intends to market the NF as a whole food (consumed as a ‘snack’ similar to other nuts) or as a food ingredient (whole kernels or fragments thereof, excluding flour) in selected food categories. The proposed target population is the general population (adolescents and adults).

According to the definition as reported in Article 3.2 of NF regulation (EU) 2015/2283 for *Jatropha curcas* L. kernels (Chuta) the following applies:

‘food consisting of, isolated from or produced from plants and their parts’ (iv).

#### 3.2. Identity of the NF

The NF is obtained from edible *Jatropha curcas* cultivars. *Jatropha curcas* L. is a plant belonging to the Euphorbiaceae family (order Malpighiales) and has seeds rich in oil. It is distributed in wild or semi-cultivated stands in many tropical regions across Africa, Southeast Asia and especially Central and North America (Aregheore et al., 1998; Martinez-Herrera et al., 2012a; Vera-Castillo et al., 2014). The *Jatropha curcas* fruits are round to oval fruits that contain one to four black oval seeds (approximately 2 cm long and 1 cm wide). After removal of the black shell, one white/beige kernel per seed is obtained with a weight of 0.4–0.5 g (Makkar et al., 1998a,b, 2011).

Although *Jatropha curcas* kernels are toxic because of the presence of phorbol esters (PEs), the possibility of using the protein and oil-rich kernels as animal feed following detoxification procedures has been studied (Devappa et al., 2010b; He et al., 2011; Francis et al., 2013; Montes et al., 2014; Vera-Castillo et al., 2014). After extraction of the oil for biodiesel production, kernel meals intended for the diet of farm animals and in aquaculture are detoxified by means of thermal and chemical treatments to decrease anti-nutrients and PEs content (Makkar and Becker, 1999; Goel et al., 2007; Kumar et al., 2011).

An edible variety of *Jatropha curcas* is also available and known since centuries by local populations in Mexico (see Section 3.7). From a phenotypical and compositional point of view, this edible cultivar is similar to the non-edible varieties (Makkar and Becker, 1997; Makkar et al., 2011; Valdes-Rodriguez et al., 2013) but contains PEs at concentrations below limits of detection (LOD) by different analytical methods (LOD as low as 0.026 μg PEs/g kernel by HPLC-UV) (Makkar et al., 1997; Martinez-Herrera et al., 2010; Devappa et al., 2013; Baldini et al., 2014b; Faria-Machado et al., 2019).

The NF in the present assessment is the hydrothermally treated kernel from an edible cultivar of *Jatropha curcas* L. (Chuta).

#### 3.3. Production process

The applicant reported that quality control checkpoints and principles of Hazard Analysis and Critical Control Point (HACCP) are applied in the production process.

Chuta kernels are obtained from *J. curcas* plants grown from ‘EdibleNut’ planting material developed within the breeding programme of JatroSolutions GmbH. Farmers located in tropical or subtropical areas characterised by adequate climatic factors for Chuta production are using ‘EdibleNut’ planting material that is verified for quality, identity and genetic purity based on the use of molecular markers. The molecular marker used by the applicant targets a mutation in a single genetic locus that...
has been identified as coding for PEs biosynthesis (King et al., 2013). The mutation in this genetic region prevents PEs biosynthesis as suggested by PEs concentrations below the LOD of the analytical method applied (0.1 μg PEs/g kernel by HPLC-UV) (He et al., 2011; King et al., 2013).

Once the plantlets reach the appropriate size, they are planted at adequate density into prepared soils. Fertilisation is done considering the nutritional conditions of the soil and the nutritional requirements of the plant. Plant protection is carried out including the concepts of integrated pest management and good agricultural practices. Fruits are allowed to ripen naturally and are harvested manually or semi-mechanically once they have reached a suitable maturity condition. To ensure traceability of the product, information on location, farm, parcel and block, where Chuta fruits were harvested, is included in the accompanying documents and/or the label. Farmers are requested to respect strict procedures to avoid any possibility of commingling with other *Jatropha* plants or seeds.

During post-harvest handling, the fruits are cleaned and mechanically de-husked by machines calibrated according to fruit size and moisture content to extract the seeds. The shelled seeds are then dried generally by solar or forced air-drying. Afterwards, Chuta seeds are sorted by size and cleaned to remove debris and other residues. In case the subsequent processing steps are not performed immediately, the seeds are packed in hermetically sealed bags and stored in a dry, cool, aerated and closed place. The applicant stated that traceability is ensured in every step and that to implement the quality management measures in the farms a ‘harvesting’ and a ‘post-harvest’ officer per farm are appointed.

Before proceeding further with the production process, analytical controls are carried out. Specifically, particular attention is paid and specific measures are in place to prevent possible commingling with non-edible (i.e. due to the presence of PEs) *Jatropha curcas* plants and kernels (see Section 3.4.2). The applicant stated that every batch of Chuta kernels will undergo, among others, PEs analytical control according to a specific procedure for appropriate sampling (see Appendix A) and applying a validated analytical method. From each batch of seeds, a number of incremental samples (from 10 up to 100) have to be taken depending on the size of the lots (from 0.1 up to 500 tons) to form an aggregate sample (with a minimum weight of 3.5 kg that may increase proportionally to the number of incremental samples to be taken). Five laboratory samples extracted from each aggregate sample will be de-shelled, ground and analysed for PEs according to the validated method. Only batches in which PEs are undetectable in all five samples will be further processed. The seeds from suitable batches are de-shelled mechanically. Whole kernels and broken kernel fragments are separated. Afterwards, kernels are processed through a hydrothermal treatment (at temperature > 120°C) to reduce anti-nutrient content and the microbiological load, and according to the applicant at the same time improving flavour and texture of the kernels. After the hydrothermal treatment, Chuta kernels are packed in airtight non-transparent polypropylene sacks.

The applicant stated that the de-shelling of seeds and hydrothermal treatments of kernels will be performed exclusively in European processing facilities.

The Panel considers that the production process is sufficiently described.

### 3.4. Compositional data

#### 3.4.1. Proximate analysis

The NF is the hydrothermally treated kernels of edible *Jatropha curcas*. Kernels are primarily constituted of fats (approximately 60%) and proteins (25%), the remaining components being fibre, ash and carbohydrates (see Table 1). The moisture content is generally limited to < 1%.

The applicant provided results of the analysis of a total of six batches of the NF obtained from kernels received from three different countries (Cameroon, Paraguay and Mexico) to also account for possible variability due to different environmental and agronomic conditions (Senger et al., 2017).

| Parameter      | Batch 1 (Cameroon) | Batch 2 (Paraguay) | Batch 3 (Mexico) | Batch 4 (Mexico) | Batch 5 (Mexico) | Batch 6 (Paraguay) | Method of analysis |
|----------------|---------------------|--------------------|------------------|-------------------|------------------|------------------|-------------------|
| Moisture (%)   | 0.3                 | 0.6                | 0.3              | 1.1               | 1.1              | 0.9              | ASU L 06.00-3     |
| Total fat (%)  | 58.9                | 61.1               | 57.7             | 60.5              | 60.5             | 53.8             | ASU L 06.00-6, gravimetric after extraction |
Analyses were conducted in an accredited laboratory according to standard methods (i.e. ASU, ISO). The Panel considers that the information provided on proximate analysis is sufficient for characterising the NF.

In addition to the proximate analysis, the applicant provided detailed analyses of fatty acid and amino acid composition, sugars, minerals and anti-nutritional factors in the NF that are reported in Section 3.10.

3.4.2. Phorbol esters, contaminants, microbiological and process contaminants

Phorbol esters

PEs represent the potential major hazard in the use of *Jatropha* kernels (Makkar et al., 1998b; Martinez-Herrera et al., 2012a; Vera-Castillo et al., 2014). Data on concentrations of PEs from three batches of *Jatropha curcas* kernels and three batches of Chuta kernels have been submitted in the original dossier. A non-validated HPLC-UV method (modified from Devappa et al., 2010b; refined by Montes et al., 2013) has been used initially by the University of Hohenheim with an LOD of 0.05 mg PEs/g (expressed as 12-O-tetradecanoylphorbol-13-acetate (TPA) equivalent). While the three Chuta batches showed PEs content below the LOD, the three non-edible *Jatropha* kernel batches contained values from 1.8 to 9.2 mg PE/g. From literature data, in flour obtained from edible *Jatropha* seeds collected from a few regions in the Mexican area (e.g. Veracruz) known for the presence of edible *Jatropha* plants, PEs were not detectable. According to Makkar et al. (1998a,b), concentrations of PEs in non-edible *Jatropha* kernels across the world (e.g. Nicaragua, Nigeria) ranged from 2.2 to 2.7 mg/g kernel, while for the edible Mexican variety concentrations were not detectable (< 0.01 mg/g LOD expressed as TPA equivalent) or ranged from 0.01 up to 0.11 mg/g, depending on the location. Makkar et al. (2011) reported values of 2.79 mg/g as an average for non-edible kernels while undetectable levels for the edible varieties were noted (< 3 μg/g sample as TPA equivalent). Martinez-Herrera et al. (2010) have noted undetectable levels in the edible varieties, with levels ranging from 0.6 to 4.1 mg/g for the non-edible variety instead (same analytical method than Makkar et al., 1998b). A content of 2.45 mg PEs/g of seed (Baldini et al., 2014a) and 2–6 mg/g of kernel (Devappa et al., 2010b) has been also reported for non-edible kernels.

Upon request from EFSA, a more sensitive analytical method was developed by the applicant and validated by an accredited laboratory for the detection of PEs in Chuta kernels. According to these new UHPLC-UV and UHPLC-MS methods (the latter used for identification of PEs peaks, if any), the applicant stated that an LOD of 0.75 μg PEs/g kernel (as TPA equivalent) was obtained. Concentration of PEs in analysed Chuta kernels was below the LOD. In addition, the applicant performed a sensitivity test for the detection of PEs by intentionally commingling of *Jatropha* kernels (containing 1.5 mg PEs/g) in Chuta kernels at a proportion of 1:1,000 using this analytical method. Based on the sampling protocol, which varies according to the size of the batch, a total of five samples of material obtained from ground aggregate samples were analysed. The presence of PEs using this methodology was evident in all five samples. Intentional commingling at a

| Parameter | Batch 1 (Cameroon) | Batch 2 (Paraguay) | Batch 3 (Mexico) | Batch 4 (Mexico) | Batch 5 (Paraguay) | Batch 6 (Paraguay) | Method of analysis |
|-----------|--------------------|--------------------|------------------|------------------|--------------------|--------------------|--------------------|
| Saturated FA (%) | 12.0 | 12.0 | 9.7 | 12.2 | 12.0 | 10.5 | ISO 5508/5509 GC-FID |
| Carbohydrates (%) | 3.3 | 2.1 | 1.4 | 3.6 | 3.9 | 7.1 | Calculated parameter |
| Total sugars (%) | 2.0 | 2.1 | 1.4 | 2.4 | 2.5 | 1.7 | Enzymatic methods |
| Total fibre (%) | 7.4 | 8.1 | 8.5 | 8.2 | 8.5 | 7.6 | ASU L 00.00-18, gravimetric |
| Total protein (%) | 26 | 25 | 28 | 21.6 | 21.1 | 25.9 | ASU L 06.00-7, Kjeldahl |
| Ash (%) | 4.1 | 4.4 | 5.4 | 5.0 | 4.9 | 4.7 | ASU L 06.00-4, gravimetric |

FA: fatty acids; ASU: Official collection of analytical methods according to § 64 LFGB (German Food, Commodities and Feed Code); ISO: International Organization for Standardization; GC-FID: gas chromatography-flame ionisation detection.
proportion corresponding to half of a *Jatropha* kernel in 1,000 Chuta kernels (i.e. 1:2,000) did not result in a fully reliable detection of PEs after applying the same sampling protocol as described above. On the basis of these experiments and accounting for the uncertainty in the LOD, the Panel uses a twofold higher concentration than the LOD when estimating exposure to PEs from the NF (see Section 3.8.4).

**Contaminants and microbiological parameters**

Additional parameters including contaminants (heavy metals), mycotoxins and microbiological quality are reported in Table 2.

**Table 2:** Batch to batch analysis of the NF: contaminants and microbiological quality

| Parameter | Batch 1 | Batch 2 | Batch 3 | Method of analysis |
|-----------|---------|---------|---------|--------------------|
| **Contaminants** | | | | |
| Arsenic (mg/kg) | < 0.05 | < 0.05 | 0.06 | ASU L 00.00-135 - ICP-MS |
| Lead (mg/kg) | < 0.02 | < 0.02 | < 0.02 | ASU L 00.00-135 - ICP-MS |
| Cadmium (mg/kg) | < 0.02 | < 0.02 | < 0.02 | ASU L 00.00-135 - ICP-MS |
| Mercury (mg/kg) | < 0.005 | < 0.005 | 0.009 | ASU 00.00-19/4 |
| Aflatoxin B1 (µg/kg) | < 0.2 | < 0.2 | < 0.2 | DIN EN 14123 - HPLC-PCD |
| Aflatoxin B2 (µg/kg) | < 0.2 | < 0.2 | < 0.2 | DIN EN 14123 - HPLC-PCD |
| Aflatoxin G1 (µg/kg) | < 0.2 | < 0.2 | < 0.2 | DIN EN 14123 - HPLC-PCD |
| Aflatoxin G2 (µg/kg) | < 0.2 | < 0.2 | < 0.2 | DIN EN 14123 - HPLC-PCD |
| Sum B1, B2, G1, G2 (µg/kg) | < 0.8 | < 0.8 | < 0.8 | DIN EN 14123 - HPLC-PCD |
| Deoxynivalenol (DON) (µg/kg) | < 20 | < 20 | < 20 | PV-18-Fusarium (LC-MS/MS) |
| HT-2 Toxin (µg/kg) | < 10 | < 10 | < 10 | PV-18-Fusarium (LC-MS/MS) |
| T-2 Toxin (µg/kg) | < 10 | < 10 | < 10 | PV-18-Fusarium (LC-MS/MS) |
| Zearalenone (ZEAs) (µg/kg) | < 10 | < 10 | < 10 | PV-18-Fusarium (LC-MS/MS) |
| Ochratoxin A (OTA) (µg/kg) | < 0.5 | < 0.5 | < 0.5 | DIN EN 14132 |
| **Microbiology (CFU/g)** | | | | |
| Mesophilic total aerobic count (CFU/g) | < 10 | < 10 | < 10 | ISO 4833 |
| Yeasts (CFU/g) | < 10 | < 10 | < 10 | ISO 7954 |
| Moulds (CFU/g) | < 10 | < 10 | < 10 | ISO 7954 |
| Enterobacteriaceae (CFU/g) | < 10 | < 10 | < 10 | ISO 21528-2 |
| Coagulase-positive *Staphylococcus* (CFU/g) | < 10 | < 10 | < 10 | ISO 6888-1 |
| Bacillus cereus, presumptive (CFU/g) | < 10 | < 10 | < 10 | ASU L 00.00-33 |
| *Salmonella* spp. (in 25 g) | ND | ND | ND | ISO 6579 |
| *Listeria monocytogenes* (CFU/g) | < 10 | < 10 | < 10 | ISO 11290-2 |

ND: not detected; HPLC-PCD: high-performance liquid chromatography-post-column derivatisation; ICP-MS: inductively coupled plasma-mass spectrometry; CFU: colony forming unit; LC-MS/MS: liquid chromatography with tandem mass spectrometry; ISO: International Organization for Standardisation; DIN: German Institute for Standardisation; ASU: official collection of analytical methods according to § 64 LFGB (German Food, Commodities and Feed Code).

Results referring to the mycotoxins and microbiological contaminants have been obtained from three representative NF batches collected after a 6-month storage period (vacuum packaging at 20°C in a cabinet).

Analyses on chemical contaminants were performed according to requirements of the Regulation (EC) No 1881/2006. Certificates are available and analyses were performed in an accredited laboratory according to standard validated methods.

Pesticide residues were analysed according to a multiscreen method for dry foodstuffs with high carbohydrate/protein content according to the accredited method ASU L 00.00-115 QuEChERS (Quick Easy Cheap Effective Rugged Safe) with LC-MS/MS (liquid chromatography-tandem mass spectrometry) and GC-MS/MS (gas chromatography-tandem mass spectrometry). All values were below the limit of quantification.

---

4 Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuffs. OJ L 364, 20.12.2006, pp. 1-33.
Process contaminants

The hydrothermal treatment (with temperature above 120°C) may lead to the production of harmful by-products. The applicant provided experimental data on acrylamide, furan, chloropropanols (i.e. 3-MCPD) and glycidyl fatty acid esters (FAE) concentrations from three batches of the NF.

Concentrations found in the NF were in many occasions below limits of detection. Results obtained in an additional single batch of untreated kernels are also reported for comparative purposes (Table 3).

The Panel notes that for 3-monochloropropane diol (3-MCPD) and glycidyl FAE, no data from comparable foods are currently available. When considering 3-MCPD, the maximum level (ML) for ‘hydrolysed vegetable protein’ and ‘soy sauce’ is set at 20 µg/kg. For the sum of 3-MCPD and 3-MCPD fatty acid esters, expressed as 3-MCPD, an ML in ‘vegetable fats as an ingredient in food’ is set at 1.25 mg/kg (Regulation (EC) No 1881/2006).

The 3-MCPD FAE concentrations were up to about 0.2 mg/kg fat in the NF, while for glycidyl FAE levels lower than 0.1 mg/kg fat were recorded. The concentrations were similar in the batch with raw kernels.

Levels of acrylamide according to Commission Recommendation (EU) 2019/18885 should be monitored in roasted nuts. When considering 3-MCPD, the maximum level (ML) for ‘hydrolysed vegetable protein’ and ‘soy sauce’ is set at 20 µg/kg. For the sum of 3-MCPD and 3-MCPD fatty acid esters, expressed as 3-MCPD, an ML in ‘vegetable fats as an ingredient in food’ is set at 1.25 mg/kg (Regulation (EC) No 1881/2006).

The 3-MCPD FAE concentrations were up to about 0.2 mg/kg fat in the NF, while for glycidyl FAE levels lower than 0.1 mg/kg fat were recorded. The concentrations were similar in the batch with raw kernels.

Levels of acrylamide according to Commission Recommendation (EU) 2019/18885 should be monitored in roasted nuts. Concentrations of acrylamide in the NF were rather low (maximum of 31 µg/kg) in comparison to other nuts: concentrations in roasted nuts and seeds have been reported to range from 33 to 251 µg/kg (Nematollahi et al., 2020) or from 21 to 271 µg/kg with levels depending on temperature (Suvvari et al., 2017).

Furans were not detected (<200 µg/kg NF) in any of the analysed batches.

To summarise, although data are limited, there are no indications of substantial formation of process contaminants.

The Panel considers that the information provided on PEs, contaminants, mycotoxins and microbiological quality is sufficient for characterising the NF.

3.5. Stability

The applicant provided experimental data after 6-month storage of the NF. The Panel notes that the applicant did not provide data from the same batches over time (no results at time 0). Data from a total of four batches (three hydrothermally treated batches and one untreated) after 6-month storage in vacuum packaging at 20°C have been provided. From the results recorded, no safety concerns have been identified.

In addition, in consideration of the relevant fat content and upon request from EFSA, the applicant provided data from an accelerated stability test to determine oxidative resistance (Oxipres) performed at high temperatures (70–110°C) and oxygen pressure (5–6 bar) to stimulate lipid oxidation and assess oxidative resistance (Trojáková et al., 2001; Sabolová et al., 2017). The results indicated that the NF showed a high resistance towards fat oxidation. According to the measured induction points of

---

**Table 3:** Heat-induced contaminants in raw and hydrothermally treated Chuta kernels

|                   | mg/kg fat | µg/kg | Acrylamide(b) | Furan(c) |
|-------------------|-----------|-------|---------------|----------|
| **3-MCPD and glycidyl FAE (expressed as 3-MCPD)**<sup>a</sup> | <0.20 | 0.12 | <0.10 | 31 | <200 |
| **3-MCPD FAE (expressed as 3-MCPD)**<sup>a</sup> | <0.20 | 0.11 | <0.10 | 26 | <200 |
| **glycidyl FAE (expressed as glycidol)**<sup>a</sup> | <0.20 | <0.10 | <0.10 | <0.10 | <200 |
| **NF batch A** | <0.20 | 0.18 | <0.10 | <0.10 | <200 |
| **NF batch B** | <0.20 | 0.18 | <0.10 | <0.10 | <200 |
| **NF batch C** | <0.20 | 0.18 | <0.10 | <0.10 | <200 |
| **Raw Chuta batch** | <0.20 | 0.18 | <0.10 | <0.10 | <200 |

(a): DGF C-VI18 (10) (GC-MS).
(b): PV-212-Acryl (LC-MS/MS).
(c): Validated in house GC-MS.
the Oxipres and applying the Arrhenius equation, the applicant estimated the oxidative stability to be at least 23 months at temperatures ≤ 25°C.

The Panel considers that, also given the low moisture content of the NF (around 1% and always < 3%), formation of possible degradation products or microbial activity is unlikely. Data after 6 months of storage assessing microbiological activity support this statement (see Section 3.4.2).

A shelf-life of 1 year was proposed by the applicant.

The Panel considers that the data provided sufficient information with respect to the proposed stability of the NF.

### 3.6. Specifications

The specifications of the NF are indicated in Table 4. The applicant stated that each batch of the NF will be subjected to analytical quality controls to ensure that all product specifications are met, including physico-chemical parameters, organoleptic characteristics, microbial counts, concentrations of heavy metals and pesticide residues.

To exclude health risk from exposure to PEs from commingling of the NF with non-edible *Jatropha* kernels, an accurate procedure for sampling and analysis of PEs as described in the production process (Sections 3.3 and 3.4.2) must be followed for each NF batch. Only batches with PEs levels below the LOD can be fully processed and considered for food use.

**Table 4: Specifications of the NF**

| Parameter                          | Specification |
|------------------------------------|---------------|
| **Physico-chemical parameters**    |               |
| Moisture (%)                       | ≤ 3           |
| Total fat (%)                      | 54–61         |
| Total protein (%)                  | 21–32         |
| Carbohydrates (%)                  | 1–7           |
| Total fibre (%)                    | 6–10          |
| Ash (%)                            | 3–5           |
| **Contaminants**                   |               |
| Phorbol esters (PEs) (µg TPA eq/g kernel)\(^a\) | ≤ 0.75        |
| Lead (mg/kg)                       | ≤ 0.20        |
| Cadmium (mg/kg)                    | ≤ 0.20        |
| Sum aflatoxins B1, B2, G1, G2 (µg/kg) | ≤ 4           |
| **Microbiological**                |               |
| Total aerobic microbial count (CFU/g) | < 1,000     |
| Total yeast/moulds count (CFU/g)   | < 100         |
| Enterobacteriaceae (CFU/g)         | < 10          |
| *Salmonella* spp.                  | Not detected in 25 g |
| Listeria monocytogenes (CFU/g)     | ≤ 100         |

An accurate procedure for sampling and analysis of PEs as described in the production process (Sections 3.3, 3.4.2 and Appendix A) must be followed for each NF batch. Only batches with concentrations of PEs below the LOD can be fully processed. TPAeq: 12-O-tetradecanoylphorbol-13-acetate equivalent; CFU: colony forming unit; UHPLC-UV-MS: ultra-high-performance liquid chromatography coupled to ultraviolet spectrophotometry and mass spectrometry.

\(^a\): Validated UHPLC-UV-MS method for detection of PEs peaks.

The Panel notes that *Listeria monocytogenes* should be part of the specification parameters to contribute to the safety of the NF as foreseen by Regulation (EC) No 2073/2005\(^6\).

The Panel considers that the information provided on the specifications of the NF is sufficient and does not raise safety concerns.

---

\(^6\) Commission Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. OJ L 338, 22.12.2005, pp. 1–30.
3.7. History of use of the NF and of its source

3.7.1. History of use of the source

*Jatropha curcas* is a tropical and subtropical plant that grows in the wild and is widely distributed. The non-edible variety is also semicultivated in Central and South America, Africa and Southeast Asia. In the last decades, the non-edible *Jatropha* kernels were used to extract oil to be used as a biofuel, while exhausted cakes were considered as possible source of proteins for use in feed after decontamination of PEs and reduction of anti-nutritional factors by different procedures (Makkar et al., 2008, 2012; Kumar et al., 2011; Wang et al., 2011; Contran et al., 2013). The Panel noted that *Jatropha* seeds are listed as a ‘harmful botanical impurity’ in Directive 2002/32/EC on undesirable substances in animal feed where it is stated that *Jatropha* seeds and fruits and their processed derivatives may only be present in feed materials and compound feed ‘in trace amounts not quantitatively determinable’.

EFSA has assessed the human and animal health risk related to use of detoxified *Jatropha* kernel meal (EFSA CONTAM Panel, 2015).

3.7.2. History of use of the NF

A *Jatropha* variety considered to be ‘PEs-free’ is grown or cultivated in some regions of Mexico (e.g. state of Veracruz) and used to prepare a variety of traditional dishes. It is consumed always after seed roasting or cooking as such or more commonly ground and used as a constituent of dishes. Roasting or heating procedures are needed to make kernels more tasteful and digestible and particularly to decrease the content of some heat-labile anti-nutritional components (see Section 3.10).

Specifically in the Totonaca culture, that started about 1500 B.C., the consumption of roasted seeds of the edible *Jatropha* variety (named ‘pinon manso’ and its seed ‘xuta’) is in its cooking tradition (Aregheore et al., 1998; Makkar et al., 2011; Martinez-Herrera et al., 2012a; Valdes-Rodriguez et al., 2013; Vera-Castillo et al., 2014; Osuna-Canizalez et al., 2015). However, although even recipes are available, only very limited information on the level of intake is available.

The NF has no history of use in Europe.

3.8. Uses and use levels and anticipated intake

3.8.1. Target population

The target population proposed by the applicant for the consumption of the NF is the general population (adolescents and adults). However, as the NF is intended to be used as an ingredient in standard food categories, it cannot be excluded that the NF can be consumed by other groups of the population. Therefore, the safety data and the exposure assessment shall cover all population groups (Commission Implementing Regulation (EU) 2017/2469, article 5(6)).

3.8.2. Proposed uses and use levels

The NF is proposed to be used as a snack or as an ingredient in several food products (e.g. mixed breakfast cereals, dried fruits and cereal bars). These food products, defined using the FoodEx2 hierarchy, and the maximum use levels are reported in Table 5. The NF in the European market is intended to be used as a whole food similar to the way peanuts are consumed.

### Table 5: Food categories and maximum use levels intended by the applicant

| FoodEx2 level | FoodEx2 CODE | Food category            | Maximum use level (g NF/100 g) |
|---------------|--------------|--------------------------|-------------------------------|
| 4             | A0DBS        | Peanuts and similar      | 100                           |
| 4             | A00FA        | Cereal bars mixed        | 5                             |
| 4             | A00EL        | Mixed breakfast cereals  | 5                             |

7 Directive 2002/32/EC of the European Parliament and of the Council of 7 May 2002 on undesirable substances in animal feed. OJ L 140, 30.05.2002, pp. 10-22.

8 FoodEx2 is a standardised food classification and description system [https://www.efsa.europa.eu/en/data/data-standardisation](https://www.efsa.europa.eu/en/data/data-standardisation).
3.8.3. Anticipated intake of the novel food

EFSA performed an assessment of the anticipated daily intake of the NF based on the applicant’s proposed uses and maximum proposed use levels (Table 5), using individual data from the EFSA Comprehensive European Food Consumption Database (EFSA, 2011). The lowest and highest mean and 95th percentile anticipated daily intakes of the NF (on an mg/kg body weight (bw) basis), among the EU dietary surveys, are presented in Table 6. The estimated daily intake of the NF for each population group from each EU dietary survey is available in the Excel file annexed to this scientific opinion (under supporting information).

Table 6: Intake estimate resulting from the use of the NF as an ingredient in the intended food categories at the maximum proposed use levels

| Population group | Age (years) | Mean intake (mg/kg bw per day) | P95th intake (mg/kg bw per day) |
|------------------|-------------|--------------------------------|--------------------------------|
|                  |             | Lowest(a) | Highest(a) | Lowest(b) | Highest(b) |
| Infants          | < 1         | 0         | 2          | 0         | 0          |
| Young children   | 1 to < 3    | 0         | 13         | 0         | 76         |
| Other children   | 3 to < 10   | 0         | 21         | 0         | 153        |
| Adolescents      | 10 to < 18  | 0         | 29         | 0         | 127        |
| Adults           | ≥ 18        | 3         | 44         | 0         | 301        |

bw: body weight.
(a): Intakes are assessed for all EU dietary surveys available in the food comprehensive database on 03/08/2020. The data relate to a period in which UK was still a Union Member State. The lowest and the highest averages observed among all EU surveys are reported in these columns.
(b): Intakes are assessed for all EU dietary surveys available in the food comprehensive database on 03/08/2020. The data relate to a period in which UK was still a Union Member State. The lowest and the highest P95th observed among all EU surveys are reported in these columns (P95th based on < 60 individuals are not considered).
(c): Includes elderly, very elderly, pregnant and lactating women.
(d): Referred as 'toddlers' in the EFSA food consumption comprehensive database (EFSA, 2011).

The Panel notes that the highest estimated 95th percentile intake (i.e. 301 mg/kg bw per day in adults) on the basis of 22 dietary surveys covered by the EFSA Food Consumption Database is corresponding to a regular daily intake of about 21 g of kernels (equivalent to 40–50 kernels) for an adult of 70 kg bw (EFSA Scientific Committee, 2012).

Based on the characteristics of the NF, the Panel assumes that the NF consumption when used as a snack is similar to that of peanuts.

3.8.4. Estimate of exposure to undesirable substances

The presence of PEs represents the potential major hazard for the consumption of Chuta kernels (see Section 3.4.2). The Panel notes that a possible exposure to the PEs can be related to the accidental presence of non-edible Jatropha kernels as a result of commingling with the Chuta kernels (see Sections 3.3, 3.4.2 and 4) or can be due to a minimal amount of PEs (below the LOD of the analytical method) present in the NF.

Assuming that all the kernels contain an amount of PEs that is at the LOD, an estimate of the theoretical exposure to the PEs can be made (Table 7). To account for uncertainties in the analytical method in these estimations, it is considered appropriate to correct the LOD by a factor of two (1.5 instead of the 0.75 µg PEs/g de-shelled kernel (as TPA equivalent) as reported in the specifications) (see Section 3.4.2).
3.9. Absorption, distribution, metabolism and excretion (ADME)

The applicant did not provide any studies or information pertaining to the ADME characteristics of the NF.

3.10. Nutritional information

The major components of the NF are lipids and proteins. Total fat comprises approximately 60% of the NF with the relative proportion of unsaturated fatty acids accounting for around 80% of total fat, while proteins account for about 25% (see Section 3.4.1).

3.10.1. Fatty acids and amino acids composition, sugars and minerals

The applicant has provided detailed analyses of fatty acids and amino acid composition, sugars and minerals in the NF. The fatty acid composition (Table 8) showed that mono- and- poly-unsaturated fatty acids are the major components.

Table 8: Fatty acid composition (fatty acid % of total fat) of three batches of raw Chuta kernels

| Fatty acid               | Mean | Range          |
|-------------------------|------|----------------|
| Myristic acid (C14:0)   | 0.24 | 0.18–0.29      |
| Palmitic acid (C16:0)   | 11.52| 11.19–11.73    |
| Palmitoleic acid (C16:1)| 0.53 | 0.48–0.57      |
| Stearic acid (C18:0)    | 6.44 | 5.26–7.98      |
| Oleic acid (C18:1)      | 34.83| 30.70–38.10    |
| Linoleic acid (C18:2)   | 45.89| 41.57–50.21    |
| α-Linolenic acid (C18:3)| 0.21 | 0.18–0.23      |
| Arachidic acid (C20:0)  | 0.19 | 0.18–0.20      |
| Total saturated fatty acids | 18.39| 17.48–19.55 |
| Total unsaturated fatty acids | 81.46| 80.33–82.37 |
| Total mono-unsaturated fatty acids | 35.36| 31.25–38.58 |
| Total poly-unsaturated fatty acids | 46.10| 41.75–50.43 |

Data from three raw batches of Chuta – internal method.

The amino acid profile was assessed for three batches (Table 9).

Table 9: Amino acid composition of three batches of the NF

| Amino acid | Results (g/100 g) | Amino acid | Results (g/100 g) |
|------------|-------------------|------------|-------------------|
|            | Mean  | Range          | Mean  | Range          |
| Isoleucine | 1.26  | 1.11–1.50      | Glycine | 1.79  | 1.01–3.10 |
| Histidine  | 0.85  | 0.56–1.40      | Serine | 2.51  | 1.29–4.70 |
Carbohydrates do not constitute a major component of the kernels (approximately 4%, ranging from 1.4 to 7.1 g/100 g kernel), the most represented sugar being sucrose. A similar or higher (i.e. phosphorus) content of minerals (Table 10) is noted in comparison to other nuts (Ros, 2010; White, 2017).

Hazelnuts, Brazil nuts and macadamia nuts have similar fat levels while only peanuts possess similar protein values (Ros, 2010). The Panel noted that the fatty acid (Chuta kernels) and amino acid profiles (NF) are similar to those of other nuts (Maguire et al., 2004; Senger et al., 2017).

### 3.10.2. Anti-nutritional factors

Non-edible and edible varieties of *Jatropha curcas* possess similar levels of anti-nutrients (Makkar et al., 1998b; Devappa et al., 2010b; Francis et al., 2013; Vera-Castillo et al., 2014). The applicant submitted data from three batches of raw Chuta kernels and three hydrothermally treated kernels (the NF) documenting the concentration of anti-nutrient compounds, such as phytic acid (myoinositol hexaphosphoric acid, IP6), trypsin inhibitors (TI) and lectin (i.e. curcin). The Panel noted that results are referring to different batches and not to the same batch before and after the hydrothermal treatment.

Anti-nutrients are widely reported to be present at various concentrations in cereals and legumes. For phytic acid, a wide variability of contents in vegetables is reported. In Chuta kernels batches (raw and hydrothermally treated), the phytates (analysed with HPLC according to Zeller et al. (2015) and expressed as IP6) showed values ranging from 3.7% to 4.6% irrespective of heat treatment. Values around 2.2% and up to 5.8% in whole wheat flour and in wheat brans, respectively, are reported (Garcia Estepa et al., 1999). Other authors reported ranges from 1.1% to 8.7% in wheat bran and germs and rice bran, while phytic acid was ranging from 1% to 5.4% in oilseeds (e.g. sunflowers and soybean), from 0.61% to 2.38% in beans, from 0.17% to 4.47% in peanuts and from 0.35% to 9.42% in almonds (Schlemmer et al., 2009; Gupta et al., 2015; EFSA NDA Panel, 2019). Since phytates are heat stable, the content is not influenced by thermal treatments (Martínez-Herrera et al., 2006; Pal et al., 2017).

### Table 10: Content of minerals from three batches of the NF

| Parameter   | mg/100 g NF | Mean | Range          |
|-------------|-------------|------|----------------|
| Calcium     |             | 215  | 202–232        |
| Phosphorus  |             | 1141 | 986–1363       |
| Magnesium   |             | 548  | 488–600        |
| Potassium   |             | 890  | 814–1017       |
| Sodium      |             | 3    | 2–4            |
| Zinc        |             | 5    | 4–6            |

According to standard accredited methods, rounded figures.

Hazelnuts, Brazil nuts and macadamia nuts have similar fat levels while only peanuts possess similar protein values (Ros, 2010). The Panel noted that the fatty acid (Chuta kernels) and amino acid profiles (NF) are similar to those of other nuts (Maguire et al., 2004; Senger et al., 2017).

### Table 10: Content of minerals from three batches of the NF

| Amino acid   | Results (g/100 g) | Amino acid   | Results (g/100 g) |
|--------------|-------------------|--------------|-------------------|
|              | Mean              | Range        | Mean              | Range         |
| Leucine      | 1.75              | 1.57–2.00    | Threonine         | 1.04          | 0.80–1.50     |
| Lysine       | 0.78              | 0.46–1.30    | Tyrosine          | 0.88          | 0.64–1.20     |
| Methionine   | 0.36              | 0.33–0.40    | Proline           | 1.13          | 0.97–1.40     |
| Phenylalanine| 1.43              | 1.09–2.00    | Aminobutyric acid | < 0.05        | < 0.05        |
| Valine       | 1.54              | 1.36–1.90    | Carnosine         | < 0.05        | < 0.05        |
| Alanine      | 1.54              | 1.16–2.20    | Hydroxylysine     | < 0.05        | < 0.05        |
| Arginine     | 2.37              | 1.97–3.20    | Hydroxyproline    | < 0.05        | < 0.05        |
| Aspartic acid| 2.61              | 2.11–3.50    | Ornithine         | < 0.5         | < 0.5–1.10    |
| Cystine      | < 0.05            | < 0.05       | Taurine           | < 0.05        | < 0.05        |
| Glutamic acid| 3.98              | 3.55–4.60    |                   |               |               |

Mean of three batches, LC-MS/MS.
For trypsin inhibitors, the inhibitory activity (TIA, mg/g dry matter) is measured. In the available literature, a maximum of about 42 mg/g dry matter in raw lentil flours (Pal et al., 2017), a range of 16–27 mg/g in whole soybean (Vagadia et al., 2017) or up to 12.5 and 48.2 mg/g in peas and raw soybeans, respectively (Gilani et al., 2012), were reported. TIA was ranging from 18.4 to 27.5 mg/g dry matter in a variety of Jatropha kernels from different origin (non-edible kernels included) (Makkar et al., 1997). It is known that TIA can be decreased by heat treatment (Schlemmer et al., 2009; Martinez-Herrera et al., 2010; Gilani et al., 2012; Mada et al., 2012; Vagadia et al., 2017). The applicant has provided data from three raw Chuta kernels and three ‘roasted’ Chuta kernels (considered representative of the NF), in duplicate (according to Smith et al., 1980). Results provided by the applicant indicated that heat-treated kernels have at least halved TIA levels when compared to the raw ones (average of about 14.3 vs. 34.0 mg trypsin inhibited/g defatted kernel).

Lectins are carbohydrate-binding glycoproteins; specifically, in Jatropha kernels, the ribosome-inactivating protein (RIP) type 1, namely curcin, is found (Makkar et al., 2012). Type 1 RIP is characterised by low toxicity, especially when compared to type 2 RIPs (e.g. ricin) (EFSA CONTAM Panel, 2008; Devappa et al., 2010b; Wu and Sun, 2012). Curcin concentrations are assessed by the applicant according to a specific qualitative method based on SDS-gel electrophoresis followed by mass spectrometry (SDS-gel electrophoresis and nano-LC-ESI-MS/MS). Heat treatments removed curcin peptides from the NF in the five tested batches since no levels were detected in hydrothermally treated kernels (< 0.05 mg curcin/g protein), while in other five untreated Chuta kernels, an average of 4.83 mg curcin/g protein (4.35–5.58) was found. This is in agreement with the published literature that reports lectin activity in vegetables and legumes (e.g. peanuts, lentils, beans) disappearing after heating or boiling (Aregheore et al., 1998; Martin-Cabrejas et al., 2009; Embaby, 2011; Martinez-Herrera et al., 2012a).

Saponins are present in a variety of plants as well as in Jatropha kernels. It is reported that in both non-edible and edible Jatropha cultivars levels are similar, ranging from 1.8 and 3.4% in kernel meal (Makkar et al., 1997; Devappa et al., 2010b) to 2.1–2.9% in defatted Jatropha seeds (Martinez-Herrera et al., 2006). Saponins found in Jatropha kernels are reported to be non-haemolytic and with limited toxicity (Devappa et al., 2010). Heat treatment only marginally decreased saponin levels (Makkar et al., 1998b; Mada et al., 2012). Saponin content is reported to be higher in soybean meal than in Jatropha meals (Martinez-Herrera et al., 2006). The applicant did not perform any additional investigation on saponins.

It is anticipated that the hydrothermal treatment of Chuta kernels will minimise or inactivate the content of specific anti-nutritional components. The presence of anti-nutritional substances does not appear to be at levels higher than those recorded in cereals, legumes or nuts. In addition, the mean estimated chronic intake of the NF is limited to a few grams per day (Section 3.8.3). The Panel considers that consumption of the NF is not nutritionally disadvantageous.

### 3.11. Toxicological information

The applicant provided eight in vitro genotoxicity studies on the NF and Jatropha kernels. These studies which were claimed proprietary by the applicant are listed in Table 11.

| Test material                  | Reference                          | Type of study                        |
|-------------------------------|------------------------------------|-------------------------------------|
| NF oil                        | Unpublished study report (2021a)    | Bacterial reverse mutation test (Ames test) |
| Jatropha kernel oil            | Unpublished study report (2021b)    |                                      |
| NF defatted meal               | Unpublished study report (2021c)    |                                      |
| Jatropha kernel defatted meal  | Unpublished study report (2021d)    |                                      |
| NF oil                        | Unpublished study report (2021e)    | *In vitro* mammalian cell micronucleus test |
| Jatropha kernel oil            | Unpublished study report (2021f)    |                                      |
| NF defatted meal               | Unpublished study report (2021g)    |                                      |
| Jatropha kernel defatted meal  | Unpublished study report (2021h)    |                                      |

At least six PEs are present in *Jatropha* seeds (Haas et al., 2002; Goel et al., 2007). In a previous assessment performed by the CONTAM Panel (EFSA CONTAM Panel, 2015), a read-across comparison
with the structural analogue TPA, a well-known non-genotoxic tumour promoter, was performed. The analysis suggested that PEs present in *Jatropha* seeds have similar, but also additional, structural alerts relevant to genotoxicity when compared to TPA. The phorbol esters, like TPA, are reported to mimic the action of diacyl glycerol, activator of protein kinase C (PKC), which regulates different signal transduction pathways. Interference with the activity of PKC affects a number of processes including phospholipid and protein synthesis, enzyme activities, DNA synthesis, phosphorylation of proteins, cell differentiation and gene expression (Goel at al., 2007; Devappa et al., 2010b; EFSA CONTAM Panel, 2015).

### 3.11.1. Genotoxicity: studies conducted with the NF and Jatropha kernels

The potential genotoxicity of the NF was investigated in a bacterial reverse mutation test and an *in vitro* mammalian cell micronucleus test (Unpublished study report, 2021a,c,e,g). For comparative purposes, the same investigations were also performed with non-edible *Jatropha* kernels (Unpublished study report, 2021b,d,f,h) (Table 11). Oil extracted from kernels and the remaining defatted meals from both Chuta and *Jatropha* were considered. These studies were conducted in compliance with OECD principles of Good Laboratory Practice (GLP) (OECD, 1998a) and in accordance with the OECD test guidelines No 471 and 487 of 2020 and 2016, respectively.

The assessment of the mutagenic potential of the NF (unpublished study reports, 2021a–d) was performed with histidine-dependent auxotrophic mutants of *Salmonella typhimurium* strains TA98, TA100, TA102, TA1535 and TA1537 that were exposed to the NF and *Jatropha* oil emulsified in acetone at concentrations up to 5 µL/plate (stock emulsion of 50 ± 5 mL/L) either in the presence or absence of liver microsomal fractions (S9). No reproducible or dose-related increases in revertant colony numbers over control counts were observed with any of the strains following exposure to NF oil or *Jatropha* oil at any concentration (irrespective of the presence or absence of S9). No evidence of toxicity towards the bacterial test strains was obtained following exposure to the NF oil. Therefore, under the experimental conditions applied, the NF oil and the *Jatropha* oil were non-mutagenic at concentrations up to 5 µL/plate, in the absence or presence of metabolic activation.

The supernatant obtained from the centrifugation of a stock suspension of the defatted kernel meals (concentration of 50 g/L in sterile demineralised water followed by dilutions) caused the formation of a slimy coating on the plates that prevented the counting of the colonies, at any dilution. Therefore, due to interferences of the test substance with the test system, no evaluation of revertant colonies and no mutagenicity assessment was performed on the defatted kernel meals (from both Chuta and *Jatropha*).

In the *in vitro* mammalian cell micronucleus test (unpublished study reports, 2021e–h), taking into consideration characteristics of the test item (i.e. the oil) that was insoluble in all tested solvents, concentrations up to 1.25 µL/mL as emulsion in acetone were tested in cultured human peripheral blood lymphocytes in the presence or absence of metabolic activation (S9 fraction). No cytotoxicity and no solid precipitates were observed (0.31, 0.63 and 1.25 µL/mL), however, the presence of an oily film at all concentrations tested was noted. No statistically significant increases in the number of binucleate cells containing micronuclei both after 4-h treatment in the presence of S9 mix or following 23-h treatment in the absence of S9 were recorded. The NF oil and the *Jatropha* oil did not show any evidence of clastogenicity or aneugenicity in the absence and presence of metabolic activation.

When the defatted kernel meals were assessed, the same approach as in the bacterial reverse mutation test was followed. For the NF, as well as for *Jatropha* defatted meals, neither a statistically significant nor a biologically relevant increase in the number of binucleated cells containing micronuclei in human lymphocytes at the three evaluated concentrations (2.5%, 5% and 10%) was observed. Under the experimental conditions applied, the NF did not show any evidence of clastogenicity or aneugenicity.

Based on the results of these studies, the Panel considers that there are no concerns regarding genotoxicity of the NF.

### 3.11.2. Subacute toxicity

The applicant did not perform any study with the NF. However, publications with descriptions of studies conducted in a few animal species (e.g. rats and fish) with test items derived from edible *Jatropha curcas* (i.e. defatted meals) or from a non-edible variety of *Jatropha curcas* after detoxification treatment (i.e. defatted and detoxified meals) have been provided and are summarised below.

- Studies conducted with edible *Jatropha* kernels
Studies performed with edible *Jatropha curcas* defatted meals were carried out in rats (Panigrahi et al., 1984; Makkar and Becker, 1999; Martinez-Herrera et al., 2012b). Although with limitations (e.g. test item not fully representative of the NF), these studies are considered as supportive for the current assessment and are summarised in Table 12.

### Table 12: In vivo studies with edible *Jatropha curcas* meals

| Reference                      | Type of study               | Strain/species                  | Dose and exposure/route of administration                                                                 |
|--------------------------------|-----------------------------|--------------------------------|----------------------------------------------------------------------------------------------------------|
| Makkar and Becker (1999)       | Nutritional quality study   | Sprague Dawley (7 male rats/group) | Diets with 10% protein (casein, non-heated and heated defatted edible *Jatropha* meal)                     |
| Martinez-Herrera et al. (2012b)| Nutritional quality study   | Wistar rats (10/group, 5 males and 5 females) | Rats were fed ad libitum (10% protein) and feed intake was recorded daily: Diet 1) Protein free, 2) with casein, 3) Edible *J. curcas* defatted flour heat-treated, 4) Diet 3 + 1% lysine added, 5) Diet 3 + 500 phytase units |
| Panigrahi et al. (1984)        | Nutritional quality study   | Wistar rats (4 sex/group)       | Arm with oil 15% of diet (pure maize oil, pure *Jatropha* oil, mix 1:1 and 4:1) Arm with raw and roasted defatted *Jatropha* meals providing 48% of crude protein in the diet |

In the 7-day feeding study in SD rats (7 male rats/group), casein, unheated and heated defatted *Jatropha* meal were used in diets to provide 10% protein. At the end of the 7 days, protein efficiency ratio (PER)\(^9\) for the unheated *Jatropha* diet was clearly lower than the one with casein (1.29 vs. 3.52) and also feed intake and body weight were affected (21 and 23% lower, respectively). Heated *Jatropha* diet only marginally affected PER (3.02 vs. 3.52), feed intake and body weight (7% lower, considered related to a reduced protein utilisation) (Makkar and Becker, 1999).

The 28-day rat study (Martinez-Herrera et al., 2012b) was conducted using a set of diets containing heat-treated (121°C for 15 min) defatted flour of edible *Jatropha* kernels with different combinations of enzymes (see Table 12). Rats were fed ad libitum and feed intake was recorded daily. The PER in the *Jatropha* diets was lower than that of casein diet (2.07), especially in the pure *Jatropha* diet (1.37, diet 3), however still within standard levels for cereals and legumes (1.2 and 1.4 for maize and soybeans, respectively). The authors reported no adverse effects at the end of the 28-day study.

Finally, one study in Wistar rats investigated the toxicity of kernel oil (mixed with maize oil in various proportions and added to the feed) and kernel meal (roasted and non-roasted) from a non-toxic genotype of *Jatropha* (grown in the Veracruz, region of Mexico). Food consumption and body weight were lower, especially in male rats fed with meals. This was considered to be a consequence of the low palatability of the feed. No indications of toxicity were found when a diet containing the *Jatropha* oil or defatted meals providing 48% of crude protein was fed to rats for 5–7 weeks (Panigrahi et al., 1984).

Studies conducted with non-edible *Jatropha* kernels (detoxified and defatted meals)

Some studies were carried out in different animal species using the detoxified non-edible *Jatropha* meal after oil extraction. Those studies were performed in fish, shrimp, rat and pig and all were of short duration (< 28 days) and rather focused on growth performance and feed utilisation generally with limited assessment of toxicological endpoints (Aregheore et al., 2003; Chivandi et al., 2006; Rakshit et al., 2008; Kumar et al., 2011; Wang et al., 2011; Makkar et al., 2012). In these studies, detoxified *Jatropha* meals obtained after solvent extraction of oil from non-edible *Jatropha curcas* plants intended for biofuel production were used. This approach resulted in defatted feed with approximately 60% of protein content, limited amount of fat and some residual levels of PEs. The Panel was of the view that the test material used in these studies, although based on *Jatropha curcas* kernels, was not representative of the NF because of the different final composition and detoxification treatments that have taken place. In addition, the duration of the studies was short. Therefore, these studies were not used in the assessment of the NF.

#### 3.11.3. Subchronic toxicity

No subchronic toxicity studies performed with edible *Jatropha* kernels were reported.

\(^9\) PER = fresh body mass gain (g)/crude protein fed (g).
The Panel considers that the available data set for the assessment of the NF is limited. However, data referring to the toxicity of PEs from *Jatropha* are of relevance.

With reference to the toxicity of PEs, the Panel noted that an NOAEL (no observed adverse effect level) of 0.4 mg/kg bw per day was identified in a subchronic study in pigs by EFSA CONTAM Panel (2015). This study (described by Li et al., 2015) was specifically designed to investigate the use of defatted *Jatropha* kernel meal to replace soybean meal in the diet of growing pigs. The main goal was to measure growth performances (feed intake, average daily body weight gain, gain-to-feed ratio); however, several safety endpoints were also considered (haematological and biochemical investigations, weight of selected organs and histopathology of liver and kidney).

A total of 144 pigs were used, divided in six groups of 12 pigs/sex each. Post-mortem investigations were performed on two pigs sex/group. The study duration was 79 days, feed consumption was measured daily and body weight every 2 weeks. Clinical pathology investigations were performed on study days 14, 28, 56 and 79. While haematology was rather complete (data available only for day 79), biochemical investigations were limited to two hepatic parameters, ALP and ALT, since liver has been identified as the main target.

The *Jatropha* kernel meals were obtained after oil extraction, steam treatment and solvent removal. The final meal contained 0.11 mg/g of PEs and was used to replace from 0% (control) up to 75% of soybean meal protein in the diet. In the six treatment groups, the corresponding amounts of PEs in the diet were 0, 2.75, 5.50, 8.25, 11.00 and 13.75 mg/kg diet for 0%, 15%, 30%, 45%, 60% and 75% protein replacement, respectively.

Growth performances were clearly impaired at concentrations of PEs of ≥ 8.25 mg/kg diet (decreased food consumption and growth) while some changes in haematological parameters (mainly lower haemoglobin concentration) at 11.00 and 13.75 mg/kg diet were recorded. In the same two dose groups, an increase in ALP and a decrease in ALT were noted throughout the study.

No changes in organ weights (heart, liver, spleen, lung, kidney and pancreas) were reported. At histopathology, no changes were noted in kidneys. Signs of steatosis or hepatic lipidosis and leucocyte infiltration were recorded at concentrations of ≥ 5.50 mg/kg diet (reported as 'mild') and signs of 'degeneration and necrosis' at concentrations of ≥ 8.25 mg/kg diet.

An NOAEL was not mentioned by the authors, however, at the concentration of 5.50 mg PEs/kg diet no effects on growth performance were noted. The next concentration of 8.25 mg PEs/kg diet 'did not alter haemato-immunological and pathological parameters but reduced growth performance'. The concentration of 5.50 mg PEs/kg diet was identified as an NOAEL by the CONTAM Panel, corresponding approximately to 0.4 mg PEs/kg bw per day (EFSA CONTAM Panel, 2015).

### 3.12. Human data

The applicant provided no human data with the NF that were relevant for the safety assessment. If a non-edible kernel is ingested, clinical symptoms include burning and pain in the mouth or throat and vomiting and this intoxication is generally self-limiting. However, it is reported that sometimes dizziness, nausea, abdominal pain and severe diarrhoea may occur and a few cases of more severe symptoms are also reported, mainly in children due to ingestion of a number of kernels (Devappa et al., 2010a; Shah and Sanmukhani, 2010; Chomchai et al., 2011; EFSA CONTAM Panel, 2015; Langrand et al., 2015; Gupta et al., 2016).

### 3.13. Allergenicity

This NF has a protein content of up to 32%, according to the proposed specifications. There is little information available on the allergenic potential of *Jatropha curcas* from the literature. The applicant indicated the absence of any report regarding allergic reactions from Mexican areas where people are consuming edible *Jatropha* kernels. However, Maciel et al. (2009) studied the allergenic properties of a non-edible type of *J. curcas* and identified a potentially allergenic 2S albumin, named Jat c 1. This potential allergen was shown to cross-react with the 2S albumin from castor bean. Subsequently, Crespo et al. (2016) characterised the IgE-binding regions of this specific allergen. It is also noted that *Jatropha* kernels contain trypsin inhibitors that are part of the prolamin superfamily that includes the major allergens from tree nuts (EFSA NDA Panel 2014).
Finally, the applicant provided ELISA (peanuts, almonds, pistachios, macadamia nuts and Brazil nuts) and PCR (cashew, hazelnuts and pecans) analysis for the detection of food allergens\textsuperscript{10} potentially present in the NF and all gave negative results. However, while the ELISA data are informative on IgE binding of allergens and potential cross-reactive proteins, the PCR analyses do not allow to assess potential cross-reactivities between the NF and the common allergenic foods tested.

The Panel considers that, given the protein content of the NF (20–30 g/100 g), allergic reactions (cross-reactivity and/or primary sensitisation) to the NF are possible.

4. Discussion

The NF is the hydrothermally treated kernels (Chuta), from edible \textit{Jatropha curcas} cultivars (EdibleNut). The applicant intends to market the NF as a whole food, consumed as a snack similar to other nuts or added as an ingredient as kernel or fragments thereof in foods such as cereal bars, breakfast cereals or mixed with dried fruits.

The target population proposed by the applicant for the consumption of the NF is the general population (adolescents and adults).

The potential presence of PEs is the major hazard in the consumption of \textit{Jatropha} kernels (see Sections 3.4.2 and 3.11). The Panel noted that the edible cultivar used in production of the NF is phenotypically indistinguishable from the \textit{Jatropha curcas} varieties that are extensively used for biofuel production and that contain variable levels of PEs. Therefore, the Panel considers it particularly crucial that the process ensures that only the selected cultivar (‘non-PE’ producing plants) within the applicant’s breeding programme is used. The entire production process must ensure that mixing with non-edible kernels does not occur. To control this, all batches must be tested for concentrations of PEs with appropriate sensitive methods prior to being marketed.

The analytical method and the procedures intended for detection of PEs were demonstrated to be able to recognise at least a single non-edible kernel with a content of 1.5 mg PEs/g kernel (PEs content in kernels from non-edible varieties can range from 0.6 up to about 10 mg/g) when mixed with 1000 edible kernels. It is noted that only whole kernels or their fractions are consumed (not flour).

The applicant demonstrated the validity of molecular markers used to check that seeds to be used by the farmers are of the genotype of the edible \textit{Jatropha curcas} cultivars. Although it is plausible that PEs biosynthesis does not occur in these edible plants, low PE levels (below the LOD) might be still present. The Panel made a theoretical exposure calculation to PEs possibly present in the NF (Section 3.8.4) and took into account uncertainties in the analytical method, by considering an LOD corrected with a factor of two (1.5 instead of the 0.75 \( \mu \)g PEs/g de-shelled kernel (as TPA equivalent) as reported in the specifications). Margins of exposure to the NOAEL (no observed adverse effect level) of 0.4 mg PEs/kg bw per day as identified by EFSA CONTAM Panel (2015) are reported in Table 13.

| Age group       | Possible PEs intake from P95 consumption of the NF (\( \mu \)g/kg bw per day) | MOE  |
|-----------------|--------------------------------------------------------------------------|------|
| Young children  | 0.11                                                                     | 3,600|
| Other children  | 0.23                                                                     | 1,700|
| Adolescents     | 0.19                                                                     | 2,100|
| Adults          | 0.45                                                                     | 900  |

In its scientific opinion in 2015, the CONTAM Panel stated that due to limitations in the toxicological data set and the characteristic of PEs (protein kinase C activation and structural alert for genotoxicity), an MOE (margin of exposure) of 400 would not be sufficient to conclude that the human risk is low. On the basis of the new data generated in the genotoxicity tests performed with the NF and with the

\textsuperscript{10} Regulation (EU) No 1169/2011 of the European Parliament and of the Council of 25 October 2011 on the provision of food information to consumers, amending Regulations (EC) No 1924/2006 and (EC) No 1925/2006 of the European Parliament and of the Council, and repealing Commission Directive 87/250/EEC, Council Directive 90/496/EEC, Commission Directive 1999/10/EC, Directive 2000/13/EC of the European Parliament and of the Council, Commission Directives 2002/67/EC and 2008/5/EC and Commission Regulation (EC) No 608/2004; J L 304, 22.11.2011, p. 18.
non-edible *Jatropha* kernels, the Panel concludes that no genotoxic concerns have been identified. In addition, the calculated MOEs according to a conservative exposure scenario were 900 or higher. The Panel considers that these margins are sufficient to ensure a safe consumption of the NF.

The presence of anti-nutrient components in the NF is within the ranges found in cereals, legumes and nuts and do not pose safety concerns. The Panel considers that, taking into account the composition of the NF and the proposed conditions of use, its consumption is not nutritionally disadvantageous.

Due to the protein content and possible cross-reactivity, allergic reactions to the NF are possible.

5. Conclusions

The Panel concludes that the NF, the hydrothermally treated *Jatropha curcas* L. kernels from an edible cultivar (Chuta), is safe under the proposed conditions of use.

5.1. Protection of Proprietary data in accordance with Article 26 of Regulation (EU) 2015/2283

The Panel could not have reached the conclusion on the safety of the NF under the proposed conditions of use without the data claimed as proprietary by the applicant: management of Chuta cultivation and use of molecular markers, compositional data including nutritional information and allergens, biological and process contaminants information, analytical methods for PEs detection, procedures for verification of PEs content, toxicological information (*in vitro* genotoxicity studies).

6. Steps taken by EFSA

1) On 19/10/2018 EFSA received a letter from the European Commission with the request for a scientific opinion on the safety of edible *Jatropha curcas* (Chuta) as a novel food. Ref. Ares (2018)5373912.
2) On 19/10/2018, a valid application on edible *Jatropha curcas* (Chuta) as a novel food, which was submitted by JatroSolutions GmbH, was made available to EFSA by the European Commission through the Commission e-submission portal (NF 2018/0177) and the scientific evaluation procedure was initiated.
3) On 18/01/2019, EFSA requested the applicant to provide additional information to accompany the application and the scientific evaluation was suspended.
4) On 18/06/2019, additional information was provided by the applicant through the Commission e-submission portal and the scientific evaluation was restarted.
5) On 26/07/2019, EFSA requested the applicant to provide additional information to accompany the application and the scientific evaluation was suspended.
6) On 15/05/2020, additional information was provided by the applicant through the Commission e-submission portal and the scientific evaluation was restarted.
7) On 08/06/2020, EFSA requested the applicant to provide further clarifications to the additional information provided.
8) On 16/07/2020, additional clarifications were provided by the applicant through the Commission e-submission portal.
9) On 06/08/2020, EFSA requested the applicant to provide further clarifications to the additional information provided.
10) On 16/10/2020, additional clarifications were provided by the applicant through the Commission e-submission portal.
11) On 06/11/2020, EFSA requested the applicant to provide further clarifications to the additional information provided.
12) On 27/11/2020, additional clarifications were provided by the applicant through the Commission e-submission portal and the scientific evaluation was restarted.
13) On 12/02/2021, EFSA requested the applicant to provide additional information to accompany the application and the scientific evaluation was suspended.
14) On 31/08/2021, additional information was provided by the applicant through the Commission e-submission portal.
15) On 20/09/2021, EFSA requested the applicant to provide further clarifications to the additional information provided.
16) On 29/09/2021 additional information was provided by the applicant through the Commission e-submission portal and the scientific evaluation was restarted.

17) During its meeting on 24/11/2021, the NDA Panel, having evaluated the data, adopted a scientific opinion on the safety of hydrothermally treated kernels from edible Jatropha curcas (Chuta) as a novel food as a NF pursuant to Regulation (EU) 2015/2283.

References

Aregheore EM, Makkar HPS and Becker K, 1998. Assessment of lectin activity in a toxic and a non-toxic variety of J. curcas using latex agglutination and haemagglutination methods and inactivation of lectin by heat treatments. Journal of the Science of Food and Agriculture, 77, 349–352.

Aregheore EM, Becker K and Makkar HPS, 2003. Detoxification of a toxic variety of Jatropha curcas using heat and chemical treatments, and preliminary nutritional evaluation with rats. The South Pacific Journal of Natural and Applied Sciences, 21, 51–56.

Baldini M, Bortolomeazzi R, Verardo G, Pascali J, Piasentier E and Franceschi L, 2014b. Determination of phorbol esters in seeds and leaves of Jatropha curcas and in animal tissue by high-performance liquid chromatography tandem mass spectrometry. Industrial Crops and Products, 59, 268–276.

Chivandi E, Erlwanger KH, Makuza SM, Read JS and Mtimuni JP, 2006. Effects of dietary Jatropha curcas meal on percent packed cell volume, serum glucose, cholesterol and triglyceride concentration and alpha-amylase activity of weaned fattening pigs. Research Journal of Animal and Veterinary Sciences, 1, 18–24.

Chomchai C, Kriengsunthornkij W, Sirisamut T, Nimsomboon T, Rungrueng W and Silpasupagornwong U, 2011. Toxicity from ingestion of Jatropha curcas (‘saboo dum’) seeds in Thai children. Southeast Asian Journal of Tropical Medicine and Public Health, 42, 946–950.

Contran N, Chessa L, Lubino M, Bellavite D, Roggero DP and Enne G, 2013. State-of-the-art of the J. curcas productive chain: from sowing to biodiesel and by-products. Industrial Crops and Products, 42, 202–215.

Crespo LM, Deus de Oliveira N, Damatta RA, Veiga do Nascimento V, Pacheco Soares T and Tavares Machado OL, 2016. Identification of IgE-binding peptide and critical amino acids of J. curcas allergen involved in allergic response. SpringerPlus, 5, 454. https://doi.org/10.1186/s40064-016-2036-5

Devappa RK, Makkar HPS and Becker K, 2010a. Jatropha toxicity-a review. Journal of Toxicology and Environmental Health, Part B, 13, 476–507.

Devappa RK, Makkar HPS and Becker K, 2010b. Optimization of conditions for the extraction of phorbol esters from Jatropha oil. Biomass and Bioenergy, 34, 1125–1133.

Devappa RK, Bingham JP and Khanal SK, 2013. High performance liquid chromatography method for rapid quantification of phorbol esters in Jatropha curcas seed. Industrial Crops and Products, 49, 211–219.

Dibusz K and Vejvodova P, 2020. Systematic literature search to assist EFSA in the preparatory work for the safety assessment of Novel Food applications and Traditional Food notifications. EFSA Supporting Publication 2020:EN-1774, 72 pp. https://doi.org/10.2903/sp.efsa.2019.EN-1774

EFSA CONTAM Panel, 2008. Ricin (from Ricinus communis) as undesirable substances in animal feed [1] – Scientific Opinion of the Panel on Contaminants in the Food Chain. https://doi.org/10.2903/j.efsa.2008.726

EFSA CONTAM Panel (EFSA Panel on Contaminants in the Food Chain), 2015. Scientific Opinion on risks for human and animal health related to the presence of phorbol esters in Jatropha kernel meal. EFSA Journal 2015;13(12):4321, 80 pp. https://doi.org/10.2903/j.efsa.2015.4321

EFSA (European Food Safety Authority), 2011. Use of the EFSA Comprehensive European Food Consumption Database in Exposure Assessment. EFSA Journal 2011;9(3):2097, 34 pp. https://doi.org/10.2903/j.efsa.2011.2097

EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies), 2014. Scientific Opinion on the evaluation of allergenic foods and food ingredients for labelling purposes. EFSA Journal 2014;12(11):3894, 286 pp. https://doi.org/10.2903/j.efsa.2014.3894

EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies), 2016. Guidance on the preparation and presentation of an application for authorisation of a novel food in the context of Regulation (EU) 2015/2283. EFSA Journal 2016;14(11):4594, 24 pp. https://doi.org/10.2903/j.efsa.2016.4594

EFSA NDA Panel (EFSA Panel on Nutrition, Novel Foods and Food Allergens), 2019. Scientific Opinion on the safety of chia seeds (Salvia hispanica L.) powders, as novel foods, pursuant to Regulation (EU) 2015/2283. EFSA Journal 2019;17(6):5716, 16 pp. https://doi.org/10.2903/j.efsa.2019.5716

EFSA Scientific Committee, 2012. Guidance on selected default values to be used by the EFSA Scientific Committee, Scientific Panels and Units in the absence of actual measured data. EFSA Journal 2012;10(3):2579, 32 pp. https://doi.org/10.2903/j.efsa.2012.2579

Embaby HES, 2011. Effect of heat treatments on certain antinutrients and in vitro protein digestibility of peanut and sesame seeds. Food Science and Technology Research, 17, 31–38.
Faria-Machado AF, Licurgo FMS, Pires JMF, da Silveira CR, Wilhelm AE, de Souza MdLM and Antoniassi R, 2019. Method validation for analysis of phorbol esters from Jatropha curcas. Industrial Crops and Products, 140, 111627. https://doi.org/10.1016/j.indcrop.2019.111627

FAOUN (Food and Agriculture Organization of the United Nations), 2012. Rome. Available online: www.fao.org/3/i3009e/i3009e00.htm

Francis G, Oliver J and Sujatha M, 2013. Non-toxic Jatropha plants as a potential multipurpose multi-use oilseed crop. Industrial Crops and Products, 42, 397–401.

Garcia Estepa RM, Guerra-Hernández E and García-Villanova B, 1999. Phytic acid content in milled cereal products and breads. Food Research International, 32, 217–221.

Gilani GS, Wu Xiao C and Cockell KA, 2012. Impact of antinutritional factors in food proteins on the digestibility of protein and bioavailability of amino acids and on protein quantity. British Journal of Nutrition, 108, 315–332.

Goel G, Makkar HPS, Francis G and Becker K, 2007. Phorbol esters: structure, biological activity, and toxicity in animals. International Journal of Toxicology, 26, 279–288.

Gupta A, Kumar A, Agarwal A, Osawa M and Verma A, 2016. Acute accidental mass poisoning by Jatropha curcas in Agra, North India. Egyptian Journal of Forensic Sciences, 6, 496–500.

Gupta RK, Gangoliya SS and Singh NK, 2015. Reduction of phytic acid and enhancement of bioavailable micronutrients in food grains. Journal of Food Science and Technology, 52, 676–684.

Haas W, Sterk H and Mittelbach M, 2002. Novel 12-deoxy-16-hydroxyphorbol diesters isolated from the seed oil of Jatropha curcas. Journal of Natural Products, 65, 1434–1440. https://doi.org/10.1021/np020060d

He W, King AJ, Khan MA, Cuevas JA, Ramíramanan D and Graham IA, 2011. Analysis of seed phorbol-ester and curcin content together with genetic variation in multiple provenances of Jatropha curcas L. from Madagascar and Mexico. Plant Physiology and Biochemistry, 49, 1183–1190.

King AJ, Montes LR, Clarke JG, Afleck J, Li Y, Witsenboer H, van der Vossen E, van der Linde P, Tripathi Y and Tavares E, 2013. Linkage mapping in the oilseed crop Jatropha curcas L. reveals a locus controlling the biosynthesis of phorbol esters which cause seed toxicity. Plant Biotechnology Journal, 11, 986–996.

Kumar V, Makkar HPS and Becker K, 2011. Detoxified Jatropha curcas kernel meal as dietary protein source: growth performance, nutrient utilisation and digestive enzymes in common carp (Cyprinus carpio L.) fingerlings. Aquaculture Nutrition, 17, 313–326.

Langrand J, Médermacht C, Schmitt C, Blanc-Brisset I, Villa AF, de Haro L and Garnier R, 2015. Intoxications par pigions d’Inde (Jatropha curcas): 24 observations rapportées aux centres antipoison de Paris et Marseille. Bulletin De La Société De Pathologie Exotique, 108, 139–143. https://doi.org/10.1007/s13149-014-0412-3

Li Y, Chen L, Lin Y, Fang ZF, Che LQ, Xu SY and Wu D, 2015. Effects of replacing soybean meal with detoxified Jatropha curcas kernel meal in the diet on growth performance and histopathological parameters of growing pigs. Animal Feed Science and Technology, 204, 18–27.

Maciel FM, Laberty MA, Deus Oliveira N, Pinto Felix S, dos Santos Soares AM, Vericimo MA and Tavares Machado OL, 2009. A new 2S albumin from J. curcas L. seeds and assessment of its allergenic properties. Peptides, 30, 2103–2107.

Mada SB, Garba A, Mohammed A, Muhammad A, Olajunjo A and Mohammed HA, 2012. Effects of boiling and roasting on antinutrients and proximate composition of local and some improved varieties of Arachis hypogaea L. (groundnut). International Journal of Food Nutrition and Safety, 1, 45–53.

Maguire LS, O’Sullivan SM, Galvin K, O’Connor TP and O’Brien NM, 2004. Fatty acid profile, tocopherol, squalene and phytosterol content of walnuts, almonds, peanuts, hazelnuts and the macadamia nut. International Journal of Food Sciences and Nutrition, 55, 171–178.

Makkar HPS, Becker K, Sporer F and Wink M, 1997. Studies on nutritive potential and toxic constituents of different provenances of Jatropha curcas. Journal of Agricultural and Food Chemistry, 45, 3152–3157.

Makkar HPS and Becker K, 1997. Potential of J. curcas seed meal as a protein supplement to livestock feed, constraints to its utilisation and possible strategies to overcome constraints. In: Gütlich GM, Mittelbach M and Trabi M (eds.). Biofuels and Industrial Products from Jatropha curcas. Dbv-Verlag für die Technische Universität Graz, Graz, Austria. pp. 190–205.

Makkar HP, Becker K and Schmook B, 1998a. Edible provenances of Jatropha curcas from Quintana Roo state of Mexico and effect of roasting on antinutrient and toxic factors in seeds. Plant Foods for Human Nutrition, 52, 31–36.

Makkar HPS, Adenibigbe AO and Becker K, 1998b. Comparative evaluation of non-toxic and toxic varieties of Jatropha curcas for chemical composition, digestibility, protein degradability and toxic factors. Food Chemistry, 62, 207–215.

Makkar HPS and Becker K, 1999. Nutritional studies on rats and fish (carp Cyprinus carpio) fed diets containing unheated and heated J. curcas meal of a non-toxic provenance. Plant Foods for Human Nutrition, 53, 183–192.

Makkar HPS, Francis G and Becker K, 2008. Protein concentrate from Jatropha curcas screw-pressed seed cake and toxic and antinutritional factors in protein concentrate. Journal of the Science of Food and Agriculture, 88, 1542–1548.

Makkar HPS, Kumar V, Oyeleye OO, Akinleye AO, Angulo-Escalante MA and Becker K, 2011. Jatropha platyphylla, a new non-toxic Jatropha species: physical properties and chemical constituents including toxic and antinutritional factors of seeds. Food Chemistry, 125, 63–71.
Makkar HPS, Kumar V and Becker K, 2012. Use of detoxified *Jatropha* kernel meal and protein isolate in diets of farm animals. In: Biofuel co-products as livestock feed-opportunities and challenges, Chapter 21. pp. 351–378.

Martín-Cabrejas MA, Aguilera Y, Pedrosa MM, Cuadrado C, Hernández T, Díaz S and Esteban RM, 2009. The impact of dehydration process on antinutrients and protein digestibility of some legume flours. Food Chemistry, 114, 1063–1068.

Martínez-Herrera J, Siddhuraju P, Francis G, Davila-Ortiz G and Becker K, 2006. Chemical composition, toxic/antimetabolite constituents, and effects of different treatments on their levels, in four provenances of *Jatropha curcas* L. from Mexico. Food Chemistry, 96, 80–89.

Martínez-Herrera J, Martínez Ayala A, Makkar HPS, Francis G and Becker K, 2010. Agroclimatic conditions, chemical and nutritional characterization of different provenances of *Jatropha curcas* L. from Mexico. European Journal of Scientific Research, 39, 396–407.

Martínez-Herrera J, Jimenez Martinez C and Guemes VN, 2012a. Use of *Jatropha curcas* (non-toxic variety) as traditional food and generation of new products in Mexico. In: *Jatropha, challenges for a new energy crop* Volume 1: Farming, Economics and Biofuel. Carels, Sujatha and Bahadur Editors. https://doi.org/10.1007/978-1-4614-4806-8

Martínez-Herrera J, Jimenez Martinez C, Martínez Ayala A, Garduno Siciliano L, Mora Escobedo R, Davila Ortiz G, Chamorro Cevallos M, Makkar H, Francis G and Becker K, 2012b. Evaluation of the nutritional quality of nontoxic kernel flour from *Jatropha curcas* L. in rats. Journal of Food Quality, 35, 152–158.

Montes JM, Technow F, Bohlinger B and Becker K, 2013. Grain quality determination by means of near infrared spectroscopy in *Jatropha curcas* L. Industrial Crops and Products, 43, 301–305.

Montes JM, Technow F, Martin M and Becker K, 2014. Genetic diversity in *Jatropha curcas* assessed with SSR and SNP markers. Diversity, 6, 551–566.

Nematomali A, Kamankesh M, Hosseini H, Hadian Z, Ghasemi J and Mohammadi A, 2020. Investigation and determination of acrylamide in 24 types of roasted nuts and seeds using microextraction method coupled with gas chromatography–mass spectrometry: central composite design. Food Measure, 14, 1249–1260. https://doi.org/10.1007/s11694-020-00373-9

OECD (Organisation for Economic Co-operation and Development), 1998. OECD Principles of good laboratory practice (as revised in 1997). OECD series on principles of good laboratory practice and compliance monitoring number 1, ENV/MC/HEM(98)17, 41 pp.

Osesa-Canizalez FJ, Atkinson CJ, Vázquez-Alvarado JMP, Barrios-Gómez EJ, Hernández-Arenas M, Rangel-Estrada SE and Cruz-Cruz E, 2015. State of the art on science and technology for production and processing of non-toxic *Jatropha*. Special Publication No. 60. 100 pp. https://doi.org/10.13140/RG.2.1.1153.1123

Pal RS, Bhartiya A, Yadav P, Kant L, Mishra KK, Aditya JP and Pattanayak A, 2017. Effect of dehulling, germination and cooking on nutrients, anti-nutrients, fatty acid composition and antioxidant properties in lentil (*Lens culinaris*). Journal of Food Science and Technology, 54, 909–920. https://doi.org/10.1007/s13197-016-2351-4

Panigrahi S, Francis BJ, Cano LA and Burbage MB, 1984. Toxicity of *Jatropha curcas* seeds from Mexico to rats and mice. Nutrition Reports International, 29, 1089–1099.

Rakshit KD, Darukheswara J, Raj KR, Narasimhamurthy K, Saibaba P and Bhagya S, 2008. Toxicity studies of detoxified *Jatropha* meal (*Jatropha curcas*) in rats. Food and Chemical Toxicology, 46, 3621–3625.

Ros E, 2010. Health benefits of nut consumption. Nutrients, 2, 652–682. https://doi.org/10.3390/nu2070652

Sabolova M, Johanidesová A, Hasalíková E, Fíšnar J, Doležal M and Rěbová Z, 2017. Relationship between the composition of fats and oils and their oxidative stability at different temperatures, determined using the Oxipres apparatus. European Journal of Lipid Science and Technology, 119, 1600454. https://doi.org/10.1002/ejlt.201600454

Schlemmer U, Frölich W, Prieto RM and Grases F, 2009. Phytate in foods and significance for humans: food sources, intake, processing, bioavailability, protective role and analysis. Molecular Nutrition & Food Research, 53, S330–S375.

Senger E, Bohlinger B, Esgaib S, Hernández-Cubero LC, Montes JM and Becker K, 2017. Chuta (edible *Jatropha curcas* L.), the newcomer among underutilized crops: a rich source of vegetable oil and protein for human consumption. European Food Research and Technology, 243, 987–997. https://doi.org/10.1007/s00217-016-2814-x

Senger E, Martin M and Montes JM, 2015. Classification of *Jatropha curcas* L. genotypes into germplasm groups associated with the presence of phorbol esters by means of seed characteristics. Industrial Crops and Products, 78, 9–12.

Shah V and Sanmukhani J, 2010. Five cases of *Jatropha curcas* poisoning. The Journal of the Association of Physicians of India, 58, 245–246.

Smith C, Van Mengen W, Twaalffhoven L and Hitchcock C, 1980. The determination of trypsin inhibitor levels in foodstuffs. Journal of the Science of Food and Agriculture, 31, 341–350.

Suvani M, Tirpanco Sivri G and Oksüz Ö, 2017. Effect of different roasting temperatures on acrylamide formation of some different nuts. IOSR Journal of Environmental Science, Toxicology and Food Technology, 11, 38–43.

Trojakova L, Rěbová Z and Pokorny J, 2001. Determination of oxidative stability in mixtures of edible oil with non-lipidic substances. Czech Journal of Food Sciences, 19, 19–23.

Unpublished study reports, 2021a. Study No. 21030901G803; Laus GmbH.

Unpublished study reports, 2021b. Study No. 21030902G803; Laus GmbH.
Unpublished study reports, 2021c. Study No. 21030903G803; Laus GmbH.
Unpublished study reports, 2021d. Study No. 21030904G803; Laus GmbH.
Unpublished study reports, 2021e. Study No. 21030901G860; Laus GmbH.
Unpublished study reports, 2021f. Study No. 21030902G860; Laus GmbH.
Unpublished study reports, 2021g. Study No. 21030903G860; Laus GmbH.
Unpublished study reports, 2021h. Study No. 21030904G860; Laus GmbH.

Vagadia BH, Vanga SK and Raghvan V, 2017. Inactivation methods of soybean trypsin inhibitor - a review. Trends in Food Science and Technology, 64, 115–125.

Valdés-Rodríguez OA, Sánchez-Sánchez O, Pérez-Vazquez A and Caplan J, 2013. The Mexican non-toxic Jatropha curcas L., food resource or biofuel? Ethnobotany Research and Applications, 11, 001–007.

Vera-Castillo YB, Cuevas JA, Valenzuela-Zapata AG, Urbano B and Gonzalez-Andres F, 2014. Biodiversity and indigenous management of the endangered non-toxic germplasm of Jatropha curcas L. in the Totonacapan (Mexico), and the implications for its conservation. Genetic Resources and Crop Evolution, 61, 1–16. https://doi.org/10.1007/s10722-014-0109-2

Wang H, Chen YI, Zhao Y, Liu H, Liu J, Makkar HPS and Becker K, 2011. Effects of replacing soybean meal by detoxified Jatropha curcas kernel meal in the diet of growing pigs on their growth, serum biochemical parameters and visceral organs. Animal Feed Science and Technology, 170, 141–146.

White J, 2017. Crunch on this... a fresh look at nuts for renal nutrition. Journal of Renal Nutrition, 27, e7–e9.

Wu W and Sun R, 2012. Toxicological studies on plant proteins: a review. Journal of Applied Toxicology, 32, 377–386.

Zeller E, Schollenberger M, Kühn I and Rodehutscord M, 2015. Hydrolysis of phytate and formation of inositol phosphate isomers without or with supplemented phytases in different segments of the digestive tract of broilers. Journal of Nutritional Science, 4, 1–12. https://doi.org/10.1017/jns.2014.62

Abbreviations

ADME absorption, distribution, metabolism and excretion
ALP alkaline phosphatase
ALT alanine transaminase
ASU official collection of analytical methods according to § 64 LFGB (German Food, Commodities and Feed Code)
B.C. before Christ
BVL German Federal Office of Consumer Protection and Food safety
bw body weight
CFU colony forming units
CONTAM Panel EFSA Panel on Contaminants
DIN German Institute for Standardisation
DNA deoxyribonucleic acid
DON deoxynivalenol
ELISA enzyme-linked immunosorbent assay
ESI electrospray ionisation
FA fatty acids
FAE fatty acid esters
GC-FID gas chromatography - flame ionisation detection
GC-MS/MS gas chromatography - tandem mass spectrometry
GLP good laboratory practice
HACCP hazard analysis and critical control points
HPLC-UV high-performance liquid chromatography - ultraviolet
ICP-MS inductively coupled plasma-mass spectrometry
IP6 inositol hexaphosphate
ISO International Organization for Standardisation
LC-MS/MS liquid chromatography - tandem mass spectrometry
LFD lateral flow device
LFGB German Food, Commodities, and Feed Code
LOD limit of detection
MCPD monochloropropane diol
ML maximum level
MOE margin of exposure
ND not detected
NDA Panel EFSA Panel on Nutrition, Novel Foods and Food Allergens
| Acronym | Description |
|---------|-------------|
| NOAEL  | no observed adverse effect level |
| NF     | Novel food |
| OECD TG| Organisation for Economic Co-operation and Development Test Guideline |
| OTA    | ochratoxin A |
| PCD    | post-column derivatisation |
| PCR    | polymerase chain reaction |
| PEs    | phorbol esters |
| PER    | protein efficiency ratio |
| PKC    | protein kinase C |
| QuEChERS | Quick Easy Cheap Effective Rugged Safe |
| RIP    | ribosome-inactivating proteins |
| SD     | Sprague Dawley |
| SDS    | sodium dodecyl sulfate |
| TI     | trypsin inhibitor |
| TIA    | trypsin inhibitory activity |
| TPA    | 12-O-tetradecanoylphorbol-13-acetate |
| UCT    | University of Chemistry and Technology (Prague) |
| UHPLC-MS | ultra high-performance liquid chromatography - mass spectrometry |
| UHPLC-UV | ultra high-performance liquid chromatography - ultraviolet |
| ZEA    | Zearalenone |
Appendix A – Phorbol esters analytical control: procedures for appropriate samplings

| Batch weight (tons) | Weight or number of sublots | Number of incremental samples |
|---------------------|-----------------------------|-----------------------------|
| ≥ 500               | 100 tons                    | 100                         |
| > 100 and ≤ 500     | 5 sublots                   | 100                         |
| > 10 and ≤ 100      | 5 sublots                   | 100                         |
| > 5.0 and ≤ 10      | –                           | 80                          |
| > 1.0 and ≤ 5.0     | –                           | 60                          |
| > 0.1 and ≤ 1.0     | –                           | 30                          |
| ≤ 0.1               | –                           | 10                          |

Each subplot shall be sampled separately. Aggregate samples are composed by a minimum of 10 incremental samples. The minimum amount of an aggregate sample shall be 3.5 kg. This amount may increase proportionally according to the number of incremental samples taken.