Plasmodium falciparum parasitaemia in the first half of pregnancy, uterine and umbilical artery blood flow, and foetal growth: a longitudinal Doppler ultrasound study

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Abstract

Background: During early pregnancy, the placenta develops to meet the metabolic demands of the foetus. The objective of this analysis was to examine the effect of malaria parasitaemia prior to 20 weeks' gestation on subsequent changes in uterine and umbilical artery blood flow and intrauterine growth restriction.

Methods: Data were analysed from 548 antenatal visits after 20 weeks' gestation of 128 women, which included foetal biometric measures and interrogation of uterine and umbilical artery blood flow. Linear mixed effect models estimated the effect of early pregnancy malaria parasitaemia on uterine and umbilical artery resistance indices. Log-binomial models with generalized estimating equations estimated the effect of early pregnancy malaria parasitaemia on the risk of intrauterine growth restriction.

Results: There were differential effects of early pregnancy malaria parasitaemia on uterine artery resistance by nutritional status, with decreased uterine artery resistance among nourished women with early pregnancy malaria and increased uterine artery resistance among undernourished women with early pregnancy malaria. Among primigravidae, early pregnancy malaria parasitaemia decreased umbilical artery resistance in the late third trimester, likely reflecting adaptive villous angiogenesis. In fully adjusted models, primigravidae with early pregnancy malaria parasitaemia had 3.6 times the risk of subsequent intrauterine growth restriction (95% CI: 2.1, 6.2) compared to the referent group of multigravidae with no early pregnancy malaria parasitaemia.

Conclusions: Early pregnancy malaria parasitaemia affects uterine and umbilical artery blood flow, possibly due to alterations in placentation and angiogenesis, respectively. Among primigravidae, early pregnancy malaria parasitaemia increases the risk of intrauterine growth restriction. The findings support the initiation of malaria parasitaemia prevention and control efforts earlier in pregnancy.

Keywords: Doppler, Malaria parasitaemia, Placenta, Pregnancy, Umbilical artery, Uterine artery, Foetal growth

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Background
In malaria endemic areas, the World Health Organization recommends prevention and control strategies for malaria parasitaemia in pregnancy, including case management of malaria parasitaemia and anaemia; insecticide-treated nets (ITNs); and, at least two doses of intermittent preventive treatment in pregnancy (IPTp) with sulphadoxine-pyrimethamine after the awareness of foetal movement [1] (from approximately 17–19 weeks’ gestation [2]). The peak prevalence of malaria parasitaemia in pregnancy occurs from 13 to 20 weeks’ gestation [3], mostly prior to the first dose of IPTp [1].

During this critical period of early pregnancy, the placenta develops to meet the growing metabolic demands of the foetus. Extravillous trophoblast cells invade and remodel the uterine spiral arteries, increasing uterine artery blood flow from the maternal circulation to the maternal side of the placenta. Concurrently, villous angiogenesis leads to increased umbilical artery blood flow from the foetus to the foetal side of the placenta.

Doppler ultrasound allows the non-invasive investigation of utero- and foeto-placental blood flow and resistance. The assessment of uterine artery blood flow reflects the extent of trophoblast invasion of the spiral arteries [4,5]. Abnormal uterine artery resistance is associated with pre-eclampsia, intrauterine growth restriction (IUGR) and adverse pregnancy outcomes [6], while increased umbilical artery resistance is associated with foetal distress and IUGR [7,8]. Ultrasound can assess foetal biometry measurements, which can be used to identify IUGR foetuses by comparing estimated foetal weights to established foetal growth standards.

It has been previously hypothesized that malaria parasitaemia in early pregnancy disrupts trophoblast invasion [9], leading to diminished utero-placental blood flow and, ultimately, intrauterine growth restriction. The objective of this analysis was to examine the hypothesis that early pregnancy malaria parasitaemia leads to subsequent changes in uterine artery and umbilical artery resistance indices, indicating alterations in placentation and villous angiogenesis, respectively. The effect of early pregnancy malaria parasitaemia on the risk of subsequent IUGR was also examined. The effect of early pregnancy malaria parasitaemia on uterine artery resistance was modified by maternal nutritional status, with increased resistance among undernourished women with early malaria infection and decreased resistance among nourished women with early malaria infection. Among primigravidae, early pregnancy malaria parasitaemia decreased umbilical artery resistance in the late third trimester. After controlling for confounders, primigravidae with early pregnancy malaria parasitaemia had nearly four times the risk of subsequent intrauterine growth restriction compared to the referent group of multigravidae with no early pregnancy malaria parasitaemia.

Methods
Study population
The longitudinal cohort consisted of pregnant women presenting for first antenatal care at Binza Maternity Hospital in Kinshasa, Democratic Republic of Congo between May 2005 and May 2006, as previously described [10,11]. In brief, 182 pregnant women ≥18 years, with non-hypertensive, non-anomalous singleton pregnancies; and, ultrasound-derived gestational age of ≤22 weeks were enrolled and followed until delivery. Enrolled women were provided with ITNs and received IPTp with sulphadoxine-pyrimethamine twice during pregnancy (between 16–27 and 28–32 weeks’ gestation) regardless of malaria parasitaemia status. All women with microscopy positive malaria parasitaemia were treated throughout follow-up.

All participants provided written informed consent to participate in the study. This study was approved by the Institutional Review Boards of the University of North Carolina at Chapel Hill and the Kinshasa School of Public Health.

For the current analysis of early pregnancy malaria parasitaemia exposure, five HIV-positive participants and 49 participants with no malaria parasitaemia exposure data ≤20 weeks’ gestation were excluded, leaving an analytic sample of 548 antenatal visits after 20 weeks’ gestation from 128 women, which included foetal biometric measures and interrogation of uterine and umbilical blood flow. Excluded participants did not significantly differ from the analytic population in age, gravidity, foetal sex, or socio-economic status (SES), but were more likely to have a low level of education and to be undernourished (data not shown). There were 6 probable recrudescent malaria episodes which occurred within 14 days of a previous infection, despite receiving treatment. Recrudescent malaria episodes were excluded from all analyses. There was no association between being a recrudescent case and parasite density as reflected by qPCR Ct-value (data not shown). Participants in the analytical sample were enrolled at a median of 18 weeks’ gestation (interquartile range [IQR]: 16, 19) and were followed up for a median of 19 weeks (IQR: 17, 21). Study participants had a median of seven follow-up visits (IQR: 6, 8), with five ultrasound scans (IQR: 4, 5) from enrolment to delivery.

Clinical, laboratory, and ultrasound procedures
Baseline data were collected in an interview (socio-demographics, medical history); medical examination (maternal anthropometrics, blood pressure, pulse, temperature); ultrasound examination (estimation of
gestational age and foetal weight); and laboratory testing (malaria parasitaemia thick and thin smears; and, filter-paper dried blood spot samples of peripheral blood) as previously described [11]. At monthly antenatal care visits, women repeated ultrasound, medical, and laboratory examinations (including malaria parasitaemia thick and thin smears and filter-paper dried blood spot samples). Quantitative real-time polymerase chain reaction (qPCR) was conducted to detect *Plasmodium* species from dried blood spot samples and *Plasmodium* positive samples were speciated as previously described [12].

Colour pulsed-wave Doppler ultrasound was used to interrogate the flow velocity waveforms in the left and right uterine arteries and the umbilical artery using standard techniques. The external iliac artery and uterine artery located medial to it were identified using colour flow settings. Flow velocity waveforms were obtained from the uterine artery near the iliac vessel prior to division of the uterine artery into branches. For left and right uterine artery analyses, Doppler indices were measured from the Doppler signal, with qualitative assessment of early diastolic notching. For the umbilical artery, the flow velocity waveform was obtained from the free flowing portion of the umbilical cord and Doppler indices and presence of absent or reversed end-diastolic flow were recorded. All ultrasound measurements were taken by a single, trained obstetrician-gynaecologist (VL) on the GE Logiqbook Ultrasound System (GE Medical Systems, Milwaukee, WI, USA).

**Variable definitions**

The exposure variable, “early pregnancy malaria parasitaemia”, was a binary, time-independent measure representing whether a woman was ever qPCR positive for peripheral *Plasmodium falciparum* malaria parasitaemia ≤20 weeks’ gestation. Quantitative PCR of peripheral dried blood spots rather than blood microscopy was used due to the higher detection threshold of microscopy and the potential for poor microscopy sensitivity and specificity [13].

Outcome variables for placental blood flow included the continuous, time-dependent mean of the left and right uterine artery resistance index (RI); and, the umbilical artery RI after 20 weeks’ gestation. The RI is defined as (peak systolic velocity − end diastolic velocity)/peak systolic velocity. The resistance index was selected as the outcome for several reasons: RI values are constrained between 0 and 1; demonstrate the least variance of the Doppler indices under identical hemodynamic conditions; are frequently used in clinical settings; and, have a truncated normal distribution, making the RI amenable to parametric statistical analyses [13].

IUGR after 20 weeks’ gestation was also examined. IUGR was a binary, time-dependent outcome. At each visit, women were classified as having an IUGR episode if, at that visit, their foetus was <10th percentile of sonographically estimated foetal weight for gestational age in completed weeks, using Hadlock’s algorithm to estimate foetal weight [15] and the ultrasound-derived, longitudinal, sex-specific Johnsen foetal growth standard [16]. The Johnsen growth standard was utilized because it is a longitudinally-derived, sex-specific nomogram. The Landis growth standard developed in the current study cohort [17] was not utilized due to its self-referential nature. Repeated episodes of IUGR were also examined. “Repeat IUGR” was a binary, time-dependent outcome defined as IUGR (as previously defined), with ≥2 total IUGR episodes during pregnancy.

The continuous time metric, gestational age in weeks, was back-calculated from the first ultrasound using Hadlock’s algorithm [18]. Ultrasound estimated gestational age has been shown to provide the best estimate of gestational age, even when last menstrual period dates are considered certain [19].

Based on previous literature of the association between malaria parasitaemia during pregnancy and foetal growth, the potential modifying effects of primigravidity (vs. multigravidity), female foetal sex (vs. male), and baseline maternal mid-upper arm circumference (MUAC) ≤24.3 cm (i.e., the lowest quartile of the full study population) (vs. “normal” MUAC) were examined. MUAC is a proxy for pre-pregnancy weight [20]. Potential confounders included maternal age, education, foetal sex, and socioeconomic status (SES). “Low SES” was defined as a binary composite variable, with unemployed women (or their partners) living in a home with few material assets (no toilet, no water, no electricity) categorized as “low SES”.

**Statistical analysis**

For linear mixed effect (LME) models, mean differences were reported and population average growth curves were plotted to describe the unadjusted and adjusted effect of early pregnancy malaria parasitaemia on uterine artery RI and umbilical artery RI. To describe the effect of early pregnancy malaria parasitaemia on IUGR, unadjusted and adjusted risk ratios (RRs) and 95% CIs were modelled using log-binomial regression generalized estimating equation (GEE) regression models with an exchangeable working correlation matrix. LME and GEE regression models are appropriate for longitudinal analyses as they account for the correlation between repeated measures in an individual [21,22].

For multivariable modelling of the effect of early pregnancy malaria parasitaemia on uterine artery RI and umbilical artery RI, a fully adjusted LME model with a random intercept was initially modelled. Linearity between gestational age and the outcome variables was
considered by examining polynomial transformations of gestational age using -2 log-likelihood (-2LL) tests for nested models using maximum likelihood estimation (\textit{a priori} cut-off of \( p < 0.10 \)). The addition of additional random effects was tested using -2LL tests with a mixed chi-square distribution (\textit{a priori} cut-off of \( p < 0.05 \)). In the final uterine artery RI and umbilical artery RI models random intercepts and slopes were included for each woman. For log-binomial GEE models, a crude model fitted for the early pregnancy malaria parasitaemia exposure variable was utilized. Next, simple log-binomial models with early pregnancy malaria parasitaemia, a theorized effect measure modifier, and their product interaction term were constructed. If the product interaction term was significant using the Wald test (\textit{a priori} cut-off of 0.10), it was considered an effect measure modifier and was included in the multivariable model. For multivariable modelling, a fully adjusted log-binomial model was initially modelled, including the early pregnancy malaria parasitaemia exposure, the significant product interaction terms and potential confounders of the association between malaria parasitaemia and IUGR. A backward elimination approach was used to assess effect measure modification. In LME models, -2LL tests were used to assess the contribution of the product interaction terms to the model using a maximum likelihood approach; in GEE models, Wald tests were used (\textit{a priori} cut-offs of \( p < 0.10 \)). Potential modifiers that did not modify the association between malaria parasitaemia and the outcome were assessed as potential confounders. Covariates were considered confounders if the change in the co-efficient of the main exposure was greater than 10%, within strata of modifiers. Restricted maximum likelihood was used for final LME model estimates, as is common practice [23]. All analyses were performed using SAS software (SAS, Cary, NC, USA).

**Results**

**Baseline characteristics of the study population**

Among the analytical sample of 128 pregnant women, 30% were ever qPCR positive for malaria parasitaemia and 21% had early pregnancy malaria parasitaemia (i.e. prior to 20 weeks’ gestation) (Table 1). Primigravidae with early pregnancy malaria parasitaemia had a mean qPCR Ct-value of 24.1 (SD 12.9), while multigravidae with early pregnancy malaria parasitaemia had a mean value of 31.4 (SD 10.7), indicating higher levels of \( P. falciparum \) nucleic acid levels in the samples from primigravidae (\( p = 0.17 \); data not shown). Similarly, no primigravidae were found to have sub microscopic parasitaemia, while in 30% of multigravida parasitaemia was submicroscopic. These semi-quantitative findings indicate that while the frequency of malaria infection was similar in primigravidae and multigravidae (Table 1), primigravidae had a higher parasite burden compared to multigravidae. Nearly all malaria infections were subclinical, with only one infection accompanied by fever.

The mean age of the study population was 27.6 years (SD: 5.1; range 18–42). Of the study participants, 27% of women were primigravidae and 22% of women had a low baseline MUAC. More than half of women (55%) were pregnant with a female foetus. Women with and without early pregnancy malaria parasitaemia had approximately balanced gravidity, foetal sex, SES and low baseline MUAC. There was a non-significant trend towards greater risk of early pregnancy malaria parasitaemia among younger women and women with less education.

**Uteroplacental blood flow and foetal growth**

Individual trajectories of the continuous outcomes, uterine artery RI and umbilical artery RI, were plotted in order to examine intra-and inter-individual variability over gestational age in weeks (Figure 1). There is a slightly negative slope for uterine artery RI over gestational age (Figure 1, Panel A). Umbilical artery RI was relatively higher in early pregnancy and decreased more rapidly over the course of pregnancy (Figure 1, Panel B). Of the 128 women, 44 (34%) ever had an IUGR episode after 20 weeks’ gestation, 55% had one episode and 27% had two episodes, with 8 women having three or more episodes. Most IUGR episodes occurred in the late second and early third trimesters.

**Early pregnancy malaria parasitaemia and uterine artery resistance**

There was no crude effect of early pregnancy malaria parasitaemia on uterine artery RI (mean difference: \(-0.017; 95\% \text{ CI: -0.045}, 0.011\) (Figure 2, Panel A). In the adjusted model, the effect of early pregnancy malaria parasitaemia on uterine artery RI varied by maternal nutritional status (Figure 2, Panel B). Compared to the referent group of women with no early pregnancy malaria parasitaemia with normal MUAC, early pregnancy malaria had a significant independent effect on uterine artery RI of \(-0.032 (95\% \text{ CI: -0.062}, -0.0006)\). Low MUAC at baseline, indicating maternal undernutrition, did not have a significant independent effect on uterine artery RI (mean difference: \(-0.011; 95\% \text{ CI: -0.042}, 0.019\)). The joint mean difference in uterine artery RI for women with early pregnancy malaria parasitaemia and low MUAC was \(+0.022 (95\% \text{ CI: -0.031}, 0.076)\), with a significant difference between undernourished and nourished women with early pregnancy malaria (\(-2LL\) test for product interaction term \( p = 0.026 \)). Thus, early pregnancy malaria parasitaemia led to a lasting significant increase in uterine artery RI of 4% among undernourished women, but caused a decrease in uterine
artery RI of 6% among nourished women, compared to the referent group of normally nourished women with no early pregnancy malaria. Foetal sex and gravidity were not found to modify the association between early pregnancy malaria parasitaemia and uterine artery RI.

### Early pregnancy malaria parasitaemia and umbilical artery resistance

There was no crude effect of early pregnancy malaria parasitaemia on umbilical artery RI (Figure 3, Panel A). In the adjusted model, the effect of early pregnancy parasitaemia on umbilical artery RI was not statistically significant (Table S1). However, there was a significant effect of low socio-economic status (SES) on umbilical artery RI, with a higher RI among women with low SES compared to those with higher SES (Figure 2). The effect of low SES on umbilical artery RI was modified by maternal age, with a stronger effect among younger women (Table S2).

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### Table 1 Baseline characteristics of pregnant women by early pregnancy malaria parasitaemia status (n = 128).

Kinshasa, Democratic Republic of Congo, 2005–2006

| Characteristic          | Total (n = 128) | Early pregnancy malaria parasitaemia (n = 27) | No early pregnancy malaria parasitaemia (n = 101) | P* |
|-------------------------|-----------------|---------------------------------------------|--------------------------------------------------|----|
| Maternal age            |                 |                                             |                                                  |    |
| ≥30                     | 48 (38)         | 6 (22)                                      | 42 (42)                                          | 0.13|
| 25-29                   | 40 (31)         | 9 (33)                                      | 31 (31)                                          |    |
| <25                     | 40 (31)         | 12 (44)                                     | 28 (28)                                          |    |
| Gravidity               |                 |                                             |                                                  |    |
| Primigravidae           | 34 (27)         | 6 (18)                                      | 28 (82)                                          | 0.56|
| Multigravidae           | 94 (73)         | 21 (22)                                     | 73 (78)                                          |    |
| Foetal sex              |                 |                                             |                                                  |    |
| Female                  | 70 (55)         | 17 (24)                                     | 53 (76)                                          | 0.36|
| Male                    | 57 (45)         | 10 (18)                                     | 47 (83)                                          |    |
| Low SES                 |                 |                                             |                                                  |    |
| Yes                     | 109 (85)        | 23 (21)                                     | 86 (79)                                          | 0.99|
| No                      | 19 (15)         | 4 (21)                                      | 15 (79)                                          |    |
| Low education           |                 |                                             |                                                  |    |
| Yes                     | 55 (43)         | 16 (29)                                     | 39 (71)                                          | 0.054|
| No                      | 74 (57)         | 11 (15)                                     | 62 (85)                                          |    |
| Baseline MUAC <24.3     |                 |                                             |                                                  |    |
| Yes                     | 28 (22)         | 6 (21)                                      | 22 (79)                                          | 0.96|
| No                      | 100 (78)        | 21 (21)                                     | 79 (79)                                          |    |

SES, socio-economic status; MUAC, mid-upper arm circumference.

*a. P* values from χ² tests.

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### Figure 1

Time plot of mean uterine and umbilical artery resistance index against gestational age. Population average smoothed trend line (in black) (n = 128, with 544 visits). Kinshasa, Democratic Republic of Congo, 2005–2006. A. Uterine artery resistance index against gestational age in weeks. B. Umbilical artery resistance index against gestational age in weeks.
malaria parasitaemia on umbilical artery RI varied over time and by gravidity (Figure 3, Panel B). Compared to the referent group of multigravidae with no early pregnancy malaria parasitaemia, the mean difference in umbilical artery RI for early pregnancy malaria parasitaemia varied from 0.038 (95% CI: 0.0065, 0.069) at 21 weeks’ gestation to −0.0087 (95% CI: -0.04, 0.03) at 39 weeks’ gestation. The mean difference in umbilical artery RI was 0.032 (95% CI: 0.012, 0.052) for primigravidae. The mean difference in umbilical artery RI for the joint effect of early pregnancy malaria parasitaemia and primigravidity varied from 0.012 (95% CI: -0.030, 0.055) at 21 weeks’ gestation to −0.032 (95% CI: -0.079, 0.011) at 39 weeks’ gestation. Thus, multigravidae with early pregnancy malaria parasitaemia had elevated umbilical artery RI that decreased to levels similar to unexposed multigravidae during the third trimester. Among primigravidae, early pregnancy malaria parasitaemia led to decreased umbilical artery RI during the late third trimester, particularly compared to primigravidae with no early pregnancy malaria parasitaemia. Both multigravidae and primigravidae with malaria infection had a greater negative umbilical artery RI slope than uninfected women.

Early pregnancy malaria parasitaemia and intrauterine growth restriction
In the unadjusted model, early pregnancy malaria parasitaemia was associated with 1.8 times the risk of subsequent IUGR during pregnancy (95% CI: 1.1, 2.9) (Table 2). In the adjusted model, the effect of early pregnancy malaria parasitaemia was found to vary by gravidity (Wald test for product interaction term: p = 0.035).
Primigravidae with early pregnancy malaria parasitaemia had 3.6 times the risk of subsequent IUGR compared to the referent group of multigravidae with no early pregnancy malaria parasitaemia (95% CI: 2.1, 6.2). Early pregnancy malaria parasitaemia was associated with a small increased risk of IUGR among multigravidae, but this finding did not reach statistical significance. In the unadjusted model for repeat IUGR, early pregnancy malaria parasitaemia was associated with 2.2 times the risk of repeat IUGR episodes. In the adjusted model, the risk of repeat IUGR episodes among primigravidae with early pregnancy malaria parasitaemia was 5.6 (95% CI: 2.8, 11.3). The increased risk for repeat IUGR with early pregnancy malaria parasitaemia likely reflects the better ability of repeated episodes of <10th percentile sonographically estimated foetal weight to capture truly IUGR foetuses than a single episode.

**Discussion**

This study provides evidence that early pregnancy malaria parasitaemia may alter placentation and lead to intrauterine growth restriction. These observations have important implications for future research directions and could ultimately inform the timing of malaria parasitaemia prevention and control efforts. The effects of early pregnancy malaria parasitaemia could be seen on both uterine and umbilical artery blood flow, indicating alterations in placenta and villous angiogenesis, respectively. Primigravidae with early pregnancy malaria parasitaemia had greater than three times the risk of subsequent IUGR compared to multigravidae with no early pregnancy malaria parasitaemia. The effects of early pregnancy malaria parasitaemia on foetal growth were less pronounced among multigravidae, suggesting that the effects of early pregnancy malaria parasitaemia are worse for primigravidae than multigravidae, as described at term [24]. This difference may also reflect the increased intensity of infection among primigravidae compared to multigravidae, as previously suggested [25]. While the findings of the current study, designed as a pilot for a larger subsequent trial, should be interpreted with caution due to small sample size, the results support recent evidence of early pregnancy malaria parasitaemia and restricted foetal growth from Thailand [26].

Differential effects of early pregnancy malaria parasitaemia on uterine artery RI between women with low and normal MUAC were found, suggesting important interactions between malaria parasitaemia and nutritional status on placental development [4,5]. The effects of malaria parasitaemia on foetal growth have previously been shown to vary by nutritional status [11], but the joint effects of malaria parasitaemia and nutritional status during pregnancy on the foetus are not well understood. Extravillous trophoblast invasion is highly regulated and is critical to establishing physiologic uteroplacental blood flow. Malaria parasitaemia in the placenta could dysregulate trophoblast invasion via relative placental hypoxia [27,28], increases in inflammatory cells and mediators (such as TNFα) [29-31]; functional folate deficiency [32,33]; and/or, increased complement activation [34,35]. Maternal nutritional status at baseline did not have a significant, independent effect on uterine artery resistance in the current study. However, maternal nutritional status has been demonstrated to affect placental development, with differential effects based

### Table 2 The effect of early pregnancy malaria parasitaemia on subsequent intrauterine growth restriction (IUGR)

|                      | Early pregnancy malaria parasitaemia | No early pregnancy malaria parasitaemia |
|----------------------|--------------------------------------|----------------------------------------|
|                      | RR (95% CI)                           | RR (95% CI)                            |
| IUGR                 |                                       |                                       |
| Unadjusted           | 1.8 (1.1, 2.9)                        | 1.0 Ref.                               |
| Adjusted*ab          | 3.6 (2.1, 6.2)                        | 1.1 (0.6, 1.9)                         |
| Primigravidae        | 1.4 (0.8, 2.5)                        | 1.0 Ref.                               |
| Multigravidae        | 2.2 (1.1, 4.2)                        | 1.0 Ref.                               |
| Repeat IUGR          |                                       |                                       |
| Unadjusted           | 5.6 (2.8, 11.3)                       | 1.2 (0.5, 3.0)                         |
| Adjusted*cdd         | 1.4 (0.6, 3.6)                        | 1.0 Ref.                               |

GEE, generalized estimating equation; IUGR, intrauterine growth restriction; RR, risk ratios; MUAC, mid-upper arm circumference.

* Adjusted for foetal sex and low baseline MUAC.

** Interaction term (early pregnancy malaria parasitaemia*primigravidae) p = 0.035.

† Adjusted for foetal sex and low baseline MUAC.

‡ Interaction term (early pregnancy malaria parasitaemia*primigravidae) p = 0.06.

GEE log-binomial model estimated unadjusted and adjusted risk ratios (RR) for the effect of early pregnancy malaria parasitaemia on subsequent IUGR episodes and repeat IUGR episodes (n = 544). Adjusted models stratified by gravidity. Kinshasa, Democratic Republic of Congo, 2005–2006.
on the timing and type of nutritional insult [36-38]. Decreased uterine artery resistance (i.e. increased uterine artery blood flow) with early pregnancy malaria or undernutrition may be an adaptive response to increase oxygen and nutrients to the placenta/foetus [39]. However, among undernourished women with early pregnancy malaria, these adaptations may not occur, leading to increased uterine artery resistance.

Increased uterine artery resistance has been previously shown to be associated with restricted foetal growth [5]. In the fully adjusted IUGR model, early pregnancy malaria was found to be associated with IUGR among primigravidae. However, maternal nutritional status at enrolment was not found to be independently associated with IUGR, as previously demonstrated in the current cohort [40]. Neither was there found to be an interaction between early pregnancy malaria and maternal undernutrition at baseline in the IUGR models. These findings indicate that, in this cohort, early pregnancy malaria had a more important impact on foetal growth than maternal nutritional status at baseline.

Altered uterine artery resistance with early pregnancy malaria parasitaemia indicates that placental malaria parasitaemia may be associated with diseases of trophoblast hypo-invasion, such as pre-eclampsia [41]. Evidence consistent with increased trophoblast invasion among nourished women with early pregnancy malaria parasitaemia was also found. Placental bed biopsies are necessary to determine if the effects of early pregnancy malaria parasitaemia on trophoblast invasion are physiological or pathological. The findings demonstrate that malaria parasitaemia prior to 20 weeks’ gestation affects placentaion and may contribute to poor pregnancy outcomes due to dysregulation of trophoblast invasion.

Among primigravidae, early pregnancy malaria parasitaemia led to decreased umbilical artery resistance, which may indicate increased angiogenesis in the villous tree of the placenta [42]. Adaptive villous branching and capillarisation occur in hypoxic placental environments, including term preeclampsia [43], high altitude [44], smoking [45], and anaemia [46]. Branching angiogenesis is associated with increased vascular endothelial growth factor (VEGF) expression [47], previously shown to be associated with placental malaria parasitaemia and maternal hypertension [41]. Increased villous angiogenesis may explain the previously reported high placental to foetal weight ratio among primigravidae with placental malaria parasitaemia [48].

Primigravidae with early pregnancy malaria parasitaemia had an increased risk of subsequent IUGR, compared to multigravidae with no early pregnancy malaria parasitaemia. Primigravidae are known to have increased severity of malaria infection and worse pregnancy outcomes than multigravidae in malaria endemic areas [24,25]. The findings support limited, but growing, evidence that early pregnancy malaria parasitaemia adversely affects birth weight [49-52] and foetal growth [26]. The adverse effect of early pregnancy malaria parasitaemia on foetal growth in primigravidae appeared to occur despite a blood flow profile consistent with moderate alterations in placentaion and increased angiogenesis. This suggests that the effects of early pregnancy malaria parasitaemia on foetal growth among primigravidae occur at the time of infection and may be mediated via mechanistic pathways other than changes in blood flow, such as inflammatory cells and cytokines [53], anaemia [54], or acute changes in blood flow during active parasitaemia [9]. In the current study, a foetus frequently demonstrated signs of IUGR at one or more study time points, yet did not at other time points, suggestive of differential rates of foetal growth during gestation. Further, there was no association between early pregnancy malaria and low birth weight (data not shown), possibly due to the frequent identification and treatment of subclinical malaria parasitaemia in the study cohort.

Conclusion

The prevalence of malaria parasitaemia peaks in early pregnancy [3] and malaria parasitaemia prevention and control measures are infrequently initiated [1] during this critical period in placental development. The effect of early pregnancy malaria parasitaemia on subsequent uterine artery resistance was modified by maternal nutritional status, with increased resistance among undernourished women with early pregnancy malaria and decreased resistance among normally nourished women with early pregnancy malaria. Among primigravidae, early pregnancy malaria parasitaemia decreased umbilical artery resistance in the late third trimester, due to changes in angiogenesis. Thus, early malaria parasitaemia infection may dysregulate placentaion and angiogenesis, leading to lasting changes in both uterine and umbilical artery blood flows. Among primigravidae, early pregnancy malaria parasitaemia was associated with nearly four times the risk of subsequent intrauterine growth restriction compared to multigravidae with no early malaria infection. These findings support the initiation of malaria parasitaemia prevention and control efforts earlier in pregnancy.

Abbreviations

- ITN: Insecticide-treated net; IPTp: Intermittent preventive treatment in pregnancy; IUGR: Intrauterine growth restriction; SES: Socioeconomic status; IQR: Interquartile range; qPCR: Quantitative real-time polymerase chain reaction; RI: Resistance index; MUAC: Mid upper arm circumference;
- LME: Linear mixed effect; RR: Risk ratio; GEE: Generalized estimating equation; -2LL: -2 log-likelihood.

Competing interests

The authors declare there are no competing interests.
Authors’ contributions

JBG developed the objectives, analysed and interpreted the data, and drafted the manuscript. JBG was supported by the Infectious Disease Epidemiology Training Program grant (S-T32-AI070114-04). VL conducted all Doppler ultrasound measurements and interpreted the Doppler data. SHL designed and implemented the study while at the Department of Epidemiology, University of North Carolina – Chapel Hill and contributed to manuscript revision while at GlaxoSmithKline. AHH assisted with statistical analyses and helped prepare the Methods section of the manuscript. AKT supervised field activities, oversaw quality control at the University of Kinshasa and interpreted the data. JMT, SJR and SRM made substantial contributions to the interpretation of results and manuscript revision. All authors read and approved the final manuscript.

Acknowledgements

The authors would like to thank Travis Thompson and Steve Taylor for conducting the DNA extraction and qPCR for the analyses. We are indebted to the women who participated in the study.

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Received: 8 May 2012 Accepted: 5 September 2012 Published: 10 September 2012

References

1. A Strategic Framework for Malaria Prevention and Control During: Pregnancy in the African Region. Brazzaville: WHO Regional Office for Africa; 2004. URL: http://www.who.int/malaria/publications/atoz af_mal_04_01/en/index.html.
2. O’Dowd MJ, O’Dowd TM. Quickening—a re-evaluation. Br J Obstet Gynaecol 1985, 92:1037–1039.
3. Brabin BJ. An analysis of malaria in pregnancy in Africa. Bull World Health Organ 1983, 61:1005–1016.
4. Prefumo F, Sebire NJ, Thilaganathan B. Decreased endovascular trophoblast invasion in first trimester pregnancies with high-resistance uterine artery Doppler indices. J Clin Pathol 2008, 61:1276–1284.
5. Trudinger BJ, Giles WB, Cook CM. Uteroplacental blood flow velocity-time waveforms in normal and complicated pregnancy. Br J Obstet Gynaecol 1985, 92:39–45.
6. Giles WB, Trudinger BJ, Baird PJ. Fetal umbilical artery flow velocity waveforms and placental resistance: pathological correlation. Br J Obstet Gynaecol 1985, 92:31–38.
7. Grimes JD, Graham CH. Placental malaria is associated with cell-mediated inflammatory responses with selective absence of natural killer cells. J Infect Dis 2001, 183:1100.
8. Conroy AL, McDonald CR, Silver KL, Liles WC, Kain KC. Complement activation: a critical mediator of adverse fetal outcomes in placental malaria? Trends Parasitol 2011, 27:94–299.
9. Vrioni AE, Cowley SJ, Thrasher AJ. The placental barrier: fundamental barrier of the human placenta. J Clin Pathol 2003, 56:18–23.
10. Taylor SM, Juliano JJ, Trotman PA, Griffin JB, Landis SH, Kita P, Tshefu AK, Meshnick SR. High-throughput pooling and real-time PCR-based strategy for malaria detection. J Clin Microbiol 2010, 48:512.
11. Ohta C, Obare P, Nanakorn A, Adhami W, Awuondo K, O’Meara W, Remich S, Martin K, Cook E, Cretien JP-L, Lucas C, Osuga J, McEvoy P, Owaga ML, Odera JS, Ogutu B. Establishing a malaria diagnostics centre of excellence in Kisumu, Kenya. Malar J 2007, 6:79.
12. Moulck D. Spectral Doppler Sonography: waveform analysis and hemodynamic interpretation. In Doppler Ultrasound in Obstetrics and Gynecology. 2nd rev. and enlarged ed. edition. Edited by Moulck DZ, Zalud I. Berlin Heidelberg: Springer-Verlag; 2005:35–56.
13. Italian Society of Gynecology and Obstetrics and Gynecology. 21st ed. 2009:379–386.
14. Harp DP, Harrit RB, Martinez-Poyer J. In utero analysis of fetal growth: a sonographic weight nomogram for a sub Saharan African population: a longitudinal study. Ultrasound Obstet Gynecol 2009, 34:579–86.
15. Stolz J, Srasr A, Collins RR. Developing a computer-assisted analysis of multiple fetal growth parameters. Radiology 1984, 154:297–301.
16. Rezende D, Lieberman E, Esquedal E. Fetal growth restriction. In: Fetal Growth Restriction. Philadelphia: Lippincott Williams & Wilkins; 2014:133–156.
17. Bergqvist M, Diulet J, Landis SH, Ananth CV, Lokomba V, Hartmann KE, Thorp JM Jr, Horton A, Atibu J, Ryder RW, Tshefu A, Meshnick SR. Ultrasound-derived fetal size normograms for a sub Saharan African population: a longitudinal study. Ultrasound Obstet Gynecol 2009, 34:579–86.
18. Liang KY, Zeger SL. Longitudinal data analysis using generalized linear models. Biometrika 1982, 38:963–974.
19. Liang KY, Zeger SL. Longitudinal data analysis using generalized linear models. Biometrika 1986, 73:63.
37. Belkacemi L, Nelson DM, Desai M, Ross MG: Maternal undernutrition influences placental-fetal development. Biol Reprod 2010, 83:325–331.

38. Roberts CT: IFPA Award in Placentology Lecture: Complicated interactions between genes and the environment in placentation, pregnancy outcome and long term health. Placenta 2010, 31:S47–S53.

39. Myatt L: Placental adaptive responses and fetal programming. J Physiol 2006, 572:25–30.

40. Landis SH: A longitudinal ultrasound study of fetal growth and intrauterine growth restriction in Kinshasa, Democratic Republic of Congo. Chapel Hill: The University of North Carolina; 2008.

41. Muehlenbachs A, Mutabingwa TK, Edmonds S, Fried M, Duffy PE: Hypertension and maternal-fetal conflict during placental malaria. PLoS Med 2006, 3:e446.

42. Abramowicz JS, Sheiner E: Ultrasound of the placenta: a systematic approach. Part II: Functional assessment (Doppler). Placenta 2008, 29:921–929.

43. Kingdom JCP, Kaufmann P: Oxygen and placental villous development: Origins of fetal hypoxia. Placenta 1997, 18:513–621.

44. Jackson MR, Mayhew TM, Haas JD: Morphometric studies on villi in human term placentae and the effects of altitude, ethnic grouping and sex of newborn. Placenta 1987, 8:487–495.

45. Pfarre C, Macara L, Leiser R, Kingdom J: Adaptive angiogenesis in placentas of heavy smokers. Lancet 1999, 354:303.

46. Kadriyev N, Kosanke G: Increased fetoplacental angiogenesis during first trimester in anemic women. Lancet 1998, 352:1747–1749.

47. Cao Y, Linden P, Shima D, Browne F, Folkman J: In vivo angiogenic activity and hypoxia induction of heterodimers of placenta growth factor/vascular endothelial growth factor. J Clin Invest 1996, 98:2507.

48. Brabin BJ, Romagosa C, Abdelgalil S, Menendez C, Verhoeff FH, McGready R, Fletcher KA, Ovens S, D’Alessandro U, Nosten F, et al: The sick placenta-the role of malaria. Placenta 2004, 25:359–378.

49. Cottrell G, Mary JY, Barro D, Cot M: The importance of the period of malarial infection during pregnancy on birth weight in tropical Africa. AmTrop Med Hyg 2007, 76:849.

50. Huyyn BT FN, Gbaguidi G, Dechavanne S, Borgella S, Guezo-Mevo B, Massougbodji A, Adam N, Deloron P, Cot M: Influence of the timing of malaria infection during pregnancy on birth weight and on maternal anaemia in Benin. Am Trop Med Hyg 2011, 85:214–220.

51. Kalliani L, Mofolo I, Chaponda M, Rogerson SJ, Meshnick SR: The effect of timing and frequency of Plasmodium falciparum infection during pregnancy on the risk of low birth weight and maternal anemia. Trans R Soc Trop Med Hyg 2010, 104:416–422.

52. Taha TET, Gray RH, Mohamedani AA: Malaria and low birth weight in central Sudan. Am J Epidemiol 1993, 138:318.

53. Fried M, Muga RO, Misore AO, Duffy PE: Malaria elicits type 1 cytokines in the human placenta: IFN-(gamma) and TNF-(alpha) associated with pregnancy outcomes. J Immunol 1998, 160:2523.

54. Menendez C, Ordi J, Ismail MR, Ventura PJ, Aponte JJ, Kahigwa E, Font F, Alonso PL: The impact of placental malaria on gestational age and birth weight. J Infect Dis 2000, 181:1740–1745.

doi:10.1186/1475-2875-11-319
Cite this article as: Griffin et al: Plasmodium falciparum parasitaemia in the first half of pregnancy, uterine and umbilical artery blood flow, and foetal growth: a longitudinal Doppler ultrasound study. Malaria Journal 2012 11:319.
Author/s:
Griffin, JB; Lokomba, V; Landis, SH; Thorp, JM; Herring, AH; Tshefu, AK; Rogerson, SJ; Meshnick, SR

Title:
Plasmodium falciparum parasitaemia in the first half of pregnancy, uterine and umbilical artery blood flow, and foetal growth: a longitudinal Doppler ultrasound study

Date:
2012-09-10

Citation:
Griffin, JB; Lokomba, V; Landis, SH; Thorp, JM; Herring, AH; Tshefu, AK; Rogerson, SJ; Meshnick, SR, Plasmodium falciparum parasitaemia in the first half of pregnancy, uterine and umbilical artery blood flow, and foetal growth: a longitudinal Doppler ultrasound study, MALARIA JOURNAL, 2012, 11

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