Reaction to *Plasmodiophora brassicae* Pathotype 6 in Lines of Brassica Vegetables, Wisconsin Fast Plants, and Canola

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Additional index words: clubroot, host resistance, napa cabbage, Rapid Cycling Brassica Collection

Abstract. Field trials were conducted from 2008 to 2010 to assess the disease reaction to clubroot, caused by *Plasmodiophora brassicae* Woronin, in selected lines of *Brassica* spp., including short-season vegetable crops [Shanghai pak choy (*B. rapa* subsp. *Chinensis* var. *communis*), Chinese flowering cabbage (*B. rapa* subsp. *Chinensis* var. *utilis*), and napa cabbage (*B. rapa* subsp. *Pekinesis*), the Rapid Cycling Brassica Collection (RCBC), also known as Wisconsin Fast Plants, and spring canola (*B. napus*). The trials were conducted on naturally infested soil with *P. brassicae* at the Muck Crops Research Station in Ontario, Canada, where pathotype 6 is predominant. Clubroot incidence and severity were higher in 2008 and 2010 compared with 2009. The lines of Shanghai pak choy and Chinese flowering cabbage were highly susceptible to clubroot, but each of the clubroot-resistant cultivars of napa cabbage, ‘Deneko’, ‘Biilk’, and ‘Yuki’, was highly resistant to pathotype 6. Among the RCBC lines, *B. carinata* and *B. juncea* were highly susceptible and could be used as susceptible models for further studies. Two RCBC lines, *B. napus* and *B. sativus*, were resistant to pathotype 6. Two of the canola cultivars, 46A76 and 46A65, were susceptible, but two others, ‘45H21’ and ‘Invigor 5020LL’, were highly resistant to pathotype 6. This difference in response can be exploited in future studies of clubroot reaction in canola.

Clubroot, caused by the soilborne protist *Plasmodiophora brassicae* Woronin, has long been recognized as an important disease of Brassica vegetable crops worldwide, and most *Brassica* spp. are highly susceptible. Management of clubroot is a challenge where Brassica crops are grown frequently because resting spores can survive for many years and are readily disseminated through movement of infested soil (Karling, 1968; Wallenhammar, 1996).

Development and proliferation of *P. brassicae* in susceptible hosts cause disorganized growth (hyperplasia and hypertrophy) of the root tissues that disrupts uptake and transport of water and nutrients (Mithen and Magrath, 1992). Plants become wilted and stunted, and yield can be reduced when symptoms are severe. Clubroot is estimated to cause 10% to 15% loss in Brassica crops throughout the world (Dixon, 2006).

Management strategies to reduce clubroot severity include extended crop rotation with nonhosts (Wallenhammar, 1996), liming (Dobson and Gabrielson, 1983; Murakami et al., 2002; Myers and Campbell, 1985), manipulation of seeding date to avoid warm periods critical for infection and symptom development (Adhikari, 2010; Gossen et al., 2011; McDonald and Westerveld, 2008), and application of fungicides (Mitani et al., 2002; Suzuki et al., 1995) and biofungicides (Narisawa et al., 2000; Peng et al., 2011). However, each of these approaches has limitations, so they need to be used together as components of an integrated management system.

Clubroot-resistant cultivars of several Brassica vegetable crops have been developed in recent years. For example, genes for clubroot resistance in European fodder turnips (*B. rapa* L.) have been used to develop clubroot-resistant Chinese cabbage (Hirai, 2006), and resistant cultivars of cabbage have been released by Syngenta (Donald and Porter, 2009). However, the resistance of most of these cultivars has not proven to be durable because the pathogen is genetically diverse.

Materials and Methods

A selection of Brassica species and cultivars was evaluated under field conditions for clubroot incidence and severity on an organic soil (Hemisol pH 6.3–6.7, 69% to 75% organic matter) naturally infested with *P. brassicae* pathotype 6. The trials were conducted at the Muck Crops Research Station (MCRS) of the University of Guelph, Holland Marsh, Ontario, Canada (lat. 44°15’ N; long. 79°35’ W).
in 2008, 2009, and 2010 at sites within 30 m of each other.

The study examined host reaction to clubroot in selected lines of RCBC, Asian vegetables, and canola. In 2008, the RCBC lines (RCBC, Madison, WI) selected for the study were: *Brassica carinata* (L.) A. Braun (genome designation, BCCbc; haploid chromosome number, *bc* = 17), *B. juncea* (L.) Czern (ABAabb, *ab* = 18), *B. napus* L. (ACAacc, *ac* = 19), *B. nigra* (L.) W.D.J. Koch (Bb, *b* = 8), *B. oleracea* L. (Ccc, *c* = 9), *B. rapa* L. (Aaa, *a* = 10) astrotab, *B. rapa* standard rapid cycling, *B. rapa* atrazine resistant, and *Raphanus sativus* L. (radish). The RCBC lines were selected to represent the six most economically important and widely grown Brassica crops. The three lines of *B. rapa* were included because some of the phenotype/genotype characters are different among these lines and there was a report of resistance to clubroot associated with triazine-resistant canola (Vigier et al., 1989). *Raphanus sativus*, which is a close relative of the genus *Brassica* (Williams and Hill, 1986), was included because transfer of genes for clubroot resistance from *R. sativus* to a *Brassica* spp. through somatic hybridization has been reported (Hagimori et al., 1992). The Asian vegetables included two lines of Shanghai pak choy: a generic line (Chan Man Hop Seeds Co., Hong Kong) and cv. Mei Qing Choi (Stokes Seeds Ltd., Ontario, Canada), two lines of Chinese flowering cabbage: generic (Chan Man Hop Seeds Co.) and cv. Tsoi-sim (Stokes Seeds Ltd.), and three cultivars of napa cabbage: ‘Deneko’ and ‘Bilko’ (Bejo Seeds Inc., New York, NY) and ‘Yuki’ (Stokes Seeds Ltd.). Shanghai pak choy, which is highly susceptible to *P. brassicae* (McDonald and Westerveld, 2008), was used as the susceptible control in these trials. The trials also included three canola cultivars from Pioneer Hi-Bred, Ontario, Canada: ‘46A76 IMI’ (imidazolinone-tolerant), ‘46A65’ (conventional), and ‘45H21 RR’ (Roundup Ready hybrid), and one from Bayer Crop Science, Ontario, Canada: ‘Invigor 5020 LL’ (Liberty Link). In 2009, the generic lines of Shanghai pak choy and Chinese flowering cabbage were dropped from the study because of the scarcity of the seeds, and they also exhibited a similar reaction to *P. brassicae* as the respective named cultivars Mei Qing Choi and Tsoi-sim. In addition, two of the napa cabbage cultivars (Bilko and Yuki) were not included in 2009 because they were resistant to clubroot and similar to ‘Deneko’. Several additional lines were not included in the study in 2010 because their interaction with *P. brassicae* was similar to that of other lines in the trial and consistent over the previous 2 years: RCBC lines *B. nigra*, *B. rapa* astrotab plant and atrazine-resistant, and *R. sativus*, and canola ‘46A65’ and ‘Invigor 5020 LL’. Trials conducted in 2008 and 2009 confirmed that all of the napa cabbage cultivars were resistant, so they were not included in the trial in 2010. ‘Tsoi-sim’ was also dropped in 2010 because of poor seed germination compared with ‘Mei Qing Choi’. Also, trials in previous years concluded that ‘Mei Qing Choi’ was more susceptible than ‘Tsoi-sim’.

The trials were established by direct seeding on 9 July 2008, 9 July 2009, and 3 Aug. 2010. These seeding dates were selected to ensure that seedling germination and establishment occurred during the warmest portion of summer, when there is a high risk of clubroot development (McDonald and Westerveld, 2008). The plots were arranged in a randomized complete block design with four replications. In 2008, the amount of available seed of the RCBC lines was limited, so they were seeded in two 1.5-m-long rows with three replications. The other lines were seeded in single 5.4-m-long rows with 40 cm between rows. In 2009 and 2010, each plot consisted of two 3-m-long rows.

Plants of the RCBC lines, Shanghai pak choy, Chinese flowering cabbage, and canola cultivars, were dug or pulled 6 weeks after seeding. Although still immature, the canola plants were assessed at this time because this study was mainly focused on incidence and severity of clubroot during vegetative crop growth. However, the napa cabbage cultivars were harvested 10 weeks after seeding, at their optimum harvest maturity. All of the plants in each plot (23–86 plants) were harvested for clubroot assessment in 2008 and 2009. In 2010, 25 plants per plot were assessed. The roots were cleaned and then assessed for clubroot incidence and severity. Clubroot severity was rated using a 0–3 scale (Kugimiya et al., 1999), where: 0 = no galling, 1 = a few small galls (small galls on less than one-third of the root), 2 = moderate galling (small to medium-sized galls on one-third to two-thirds of the root), and 3 = severe galling (medium to large-sized galls on greater than two-thirds of the root). A disease severity index (DSI) was calculated using the following equation (Kobriger and Hagedorn, 1983):

\[
DSI = \frac{\sum \left(\frac{\text{class no.}}{\text{(no. plants in each class)}}\right) \times 100}{\text{(total no. plants per sample) \times (no. classes – 1)}}
\]

Weather parameters were measured at a weather station (Campbell Scientific, Edmonton, Alberta, Canada) located within 100 m of the experimental plots. Air temperatures were measured using a HMP35C probe, soil temperatures at 5-cm depth using a Model 107 probe, and rainfall data using a tipping-bucket rain gauge. Temperatures (air and soil) and rainfall data were recorded every hour using a CR21X data logger. Daily maximum, minimum, and mean temperatures and total rainfall were calculated for the growing period of the crops in each trial.

Statistical analyses were performed using SAS Version 9.1 (SAS Institute, Cary, NC). A mixed-model analysis of variance of the data in each trial was conducted using PROC MIXED with species/lines as the fixed effect and year and replication as random effects.

The data set for each trial was tested for normality using the Shapiro-Wilk test of residuals, and outliers were identified using Lund’s test of standardized residuals (Lund, 1975). No outliers were found in any data set. Mean separation was conducted using Tukey’s multiple mean comparison test. Differences were significant at *P* ≤ 0.05 unless otherwise noted.

Results and Discussion

The RCBC lines of *R. sativus* and *B. napus* were resistant to *P. brassicae* at this site and *B. oleracea* was moderately susceptible to clubroot (Table 1). Other RCBC lines were highly susceptible, especially *B. carinata* and *B. juncea*, which had levels of clubroot similar to the highly susceptible control, Shanghai pak choy (Table 1). There were no differences in clubroot incidence and severity among the lines of *B. rapa*, which indicates that resistance to atrazine is not consistently related to resistance to clubroot, as reported previously (Vigier et al., 1989). To our knowledge, this is the first study to evaluate the reaction of RCBC lines to *P. brassicae*. However, there was a study on resistance gene selection to *Albugo candida* in rapid cycling *B. campestris* lines (Edwards and Williams, 1987). Use of these highly susceptible short-generation RCBC lines as model crops could reduce the duration of many studies of this host-pathogen interaction relative to standard lines of the same crop. Their small stature would also facilitate studies in situations in which space is restricted such as containment facilities. The RCBC seeds are quite expensive ($10/100 seeds) but this cost is small compared with the daily cost of using growth cabinets and containment facilities.

Two lines of Shanghai pak choy, a generic line and ‘Mei Qing Choi’, were included in the trial in 2008 because seed of the generic line that had been used in previous studies (e.g., McDonald and Westerveld, 2008) had become scarce, but seed of a named cultivar had become available. The same situation with seed availability also occurred for the lines of Chinese flowering cabbage (generic and ‘Tsoi-sim’). For both crops, plant phenology and development were similar in the two lines (data not shown), and both lines were highly susceptible to *P. brassicae* (Table 1). Therefore, the generic cultivars were dropped from the trial in 2009. We conclude that the named cultivars have a similar reaction to *P. brassicae* pathotype 6 to that of the generic lines used in previous studies and so can be used as replacements for the generic lines in subsequent studies. Shanghai pak choy has potential as a susceptible model crop for studies on clubroot because of its small size and susceptibility, but it does not go to seed as rapidly as the RCBC lines.

All three cultivars of napa cabbage, each of which is marketed as resistant to clubroot, were highly resistant at this site in 2008. ‘Bilko’ and ‘Yuki’ were dropped from the study in 2009 because they had the same
Asian vegetables

Rapid cycling Brassica crops

| Crops/species and line | 2008 CI | DSI | 2009 CI | DSI | 2010 CI | DSI |
|------------------------|--------|-----|--------|-----|--------|-----|
| **Brassica carinata**  | 97 f   | 97 f | 45 c   | 19 c| 85 c   | 82 c|
| **B. juncea**          | 96 f   | 95 f | 30 bc  | 16 bc| 83 c   | 75 c|
| **B. nigra**           | 89 ef  | 66 def| 3 a   | 1 a | —      | —   |
| **B. rapa, astropsych**| 84 ef  | 69 ef| 2 a   | 1 a | —      | —   |
| **B. rapa, standard**  | 82 ef  | 74 ef| 19 abc| 8 abc| 87 c   | 77 c|
| **B. rapa, atrazine-resistant** | 75 def | 67 ef| 8 ab | 7 ab| —      | —   |
| **B. oleracea**        | 57 c–f | 37 b–e| 10 ab | 3 ab| 58 b   | 39 b|
| **Raphanus sativus**   | 10 ab  | 7 ab | 0 a   | 0 a | —      | —   |
| **B. napus**           | 4 ab   | 1 a | 1 a   | 0.4 a| 4 a    | 3 a |

Asian vegetables

Shanghai pak choy (*B. rapa* subsp. *Chinensis* var. *communis*)
- Generic: 89 ef, 71 ef
- Mei Qing Choi: 80 ef, 56 cde, 9 ab, 3 ab, 89 c, 64 bc

Chinese flowering cabbage (*B. rapa* subsp. *Chinensis* var. *utilis*)
- Generic: 38 bcd, 23 abc
- Tsoi-sim: 37 bcd, 23 abc, 4 ab, 2 ab

Napa cabbage (*B. rapa* subsp. *Pekinensis*)
- Deneko: 6 ab, 2 a, 0 a
- Bilko: 3 ab, 3 a
- Yuki: 1 ab, 1 a

Canola (*B. napus*)
- 46A76: 52 cde, 44 b–e, 11 ab, 4 ab, 81 c, 73 c
- 46A65: 33 abc, 26 a–d, 4 a, 1 a
- Invigor 5020 LL: 0 a, 0 a, 0 a
- 45H21: 0 a, 0 a, 0 a, 7 a, 3 a

*Means within a column followed by the same letter do not differ based on Tukey’s test at P = 0.05.*

In conclusion, the current study examined the reaction to clubroot as ‘Deneko’. This observation supports a previous report of assessment of cv. Bilko under controlled conditions, in which this cultivar was highly resistant to each of the pathotypes of *P. brassicae* that occur in Canada (Hasan, 2010). There was no clubroot development in ‘Deneko’ in 2009, which confirmed the result from 2008 when there was a little disease development. Therefore, ‘Deneko’ was dropped from the study in 2010.

Among the four canola cultivars, 46A76 was highly susceptible to pathotype 6 in all 3 years of the study. Cultivar 46A65 had slightly lower clubroot incidence than ‘46A76’ in 2008 but generally did not differ from ‘46A76’ and so was dropped from the study in 2010. Cultivars Invigor 5020 LL and 45H21 were both highly resistant in 2008 and 2009, so ‘Invigor 5020 LL’ was dropped from the study in 2010. It is interesting to note that a low level of clubroot (both incidence and severity) developed on ‘45H21’ in 2010 (Table 1). This indicates that even a highly resistant line can develop low levels of clubroot when conditions are highly conducive for infection and pathogen development. Also, the canola cultivars Invigor 5020 LL and 45H21, which are susceptible to pathotype 3 and ECD 16/15/12 (Strelkov et al., 2006) were resistant to pathotype 6, so either line can be used to differentiate pathotype 6 from pathotype 3.

Clubroot incidence and severity were higher in 2008 and 2010 than in 2009. There was a small line-by-year interaction, but the lines mostly exhibited a similar pattern of response across all 3 years (Table 1). Rainfall during the growing season over the 3 years of the study was quite consistent and adequate for crop growth, but temperatures during the early growing period of the crop were lower in 2009 (17.9 °C) than 2008 (20.4 °C) and 2012 (21.1 °C). The cool conditions in 2009 may have contributed to lower clubroot incidence and severity (Gossen et al., 2011). However, the low levels of clubroot in 2009 may also have resulted from low concentrations of resting spores of *P. brassicae* in that specific portion of the study site, although all of the trials were conducted within 30 m of each other. Later on, the overall spore concentration of *P. brassicae* in the research site was found to be 1 x 10⁷ to 1.3 x 10⁹/g of soil (M.T. Tesfaendrias, personal communication). Previous studies have demonstrated that root hair infection (Naiki et al., 1978) and clubroot severity (Hildebrand and McRae, 1998) increase with increasing spore load above a minimum threshold level for symptom development (Faggian and Strelkov, 2009). The distribution of clubroot within fields can be very patchy (Strelkov et al., 2007), and existing tests to assess the density of resting spores are too labor-intensive and time-consuming (Dhingra and Sinclair, 1985; Murakami et al., 2000) to be feasible when intensive sampling is required. A rapid and inexpensive test that could be used to quantify inoculum concentration at numerous points in the field would be very useful for identifying uniform areas for field trials as well as for various epidemiological studies.

In conclusion, the current study examined the reaction to *P. brassicae* pathotype 6 in a range of Brassica lines. Use of these lines as model crops could expedite a broad range of clubroot research on Brassica vegetables and canola. The RCBC lines of *B. carinata* and *B. juncea* were highly susceptible to pathotype 6, and lines of *R. sativus* and *B. napus* were highly resistant. These lines may be useful as model crops for studies such as host-pathogen interaction, seed yield in relation to disease severity, and for disease management trials. Additional studies with other pathotypes would be useful. This study confirmed that the cultivars of napa cabbage, ‘Deneko’, ‘Bilko’, and ‘Yuki’, were highly resistant to *P. brassicae* pathotype 6. The susceptible canola cultivars 46A76 and 46A65 and resistant cultivars Invigor 5020 LL and 45H21 can also be used for further studies on host-pathogen interaction.

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