A Mouse Model of *Leishmania braziliensis braziliensis* Infection Produced by Coinjection with Sand Fly Saliva

By John Samuelson, Ethan Lerner, Robert Tesh, and Richard Titus

*From the Department of Tropical Public Health, Harvard School of Public Health, Boston, Massachusetts 02115; the *Department of Pathology, New England Deaconess Hospital, Boston, Massachusetts 02215; the Department of Dermatology, Harvard Medical School, Boston, Massachusetts 02115; and the Yale Arbovirus Research Unit, Yale University School of Medicine, New Haven, Connecticut 06510*

**Summary**

To develop a reliable murine model of *Leishmania braziliensis braziliensis* infection, parasites were injected into BALB/c mice in the presence of phlebotomine sand fly salivary gland lysates, which have previously been shown to greatly increase the infectivity of *L. major* in mice. When injected with salivary gland lysates, *L. braziliensis braziliensis* produced progressively enlarging cutaneous nodules, containing many macrophages filled with *Leishmania* amastigotes. In contrast, *L. braziliensis* injected without gland extracts produced small and rapidly regressing lesions. Isoenzyme analysis, monoclonal antibodies, and the polymerase chain reaction with *L. braziliensis*-specific oligonucleotide primers and probes confirmed that parasites causing the lesions were *L. braziliensis*.

*Leishmania braziliensis* is an intracellular protozoan parasite transmitted by phlebotomine sand flies to persons living in Central and South America. *L. braziliensis* is divided into four subspecies: *L. braziliensis braziliensis*, *L. braziliensis guyanensis*, *L. braziliensis panamensis*, and *L. braziliensis peruviana*. The parasites usually cause a self-healing ulcer at the site of the bite. *L. braziliensis braziliensis* is of particular concern because mucocutaneous disease, with disfiguring lesions of the mouth and nose, may develop (1).

*L. donovani*, the cause of visceral leishmaniasis, and *L. mexicana* and *L. major*, which cause cutaneous leishmaniasis, all produce progressive lesions in susceptible BALB/c mice. These lesions are characterized by the presence of numerous macrophages filled with *Leishmania* amastigotes (2). Therefore, mice have been used extensively to study the immune response to and the effect of drugs on these species of *Leishmania*.

In contrast, *L. braziliensis braziliensis* has not produced lesions in BALB/c mice (3). *L. braziliensis* from Panama, presumably *L. braziliensis panamensis*, and *L. braziliensis peruviana* caused cutaneous lesions of BALB/c mice that were variable (4) and progressive (5), respectively. Because of the difficulties with the mouse model, *L. braziliensis* has most frequently been studied in the Golden hamster (3, 6), which has an immune system much less well characterized than that of the mouse and for which few immunological reagents are available.

Sand flies salivate into the skin while probing for a blood-meal and transmitting *Leishmania*. Sand fly saliva contains several substances with potent pharmacological activities including a potent vasodilator (7). The saliva of sand flies also enhances transmission of *Leishmania* parasites, as *L. major* parasites injected with sand fly salivary gland lysates formed lesions in mice that were much larger than those of control parasites injected without saliva (8). The lesions caused by *L. major* and saliva contained as many as 5,000-fold more parasites than controls, and in the presence of salivary gland material as few as ten *L. major* parasites caused a lesion (8).

Here we report that *L. braziliensis braziliensis* parasites caused progressive cutaneous lesions when injected with sand fly glands into BALB/c mice, while control parasites injected alone produced small and rapidly regressing cutaneous nodules.

**Materials and Methods**

*Parasites.* *Leishmania* were cultured in Schneider's drosophila medium or on blood agar plates. *L. braziliensis braziliensis* strain WR604 was isolated from a single cutaneous lesion of a girl living in the Corte de Pebro region of Brazil. The parasite was typed by isoenzymes as an *L. braziliensis braziliensis* by Dr. Richard Kreutzer (Youngstown State, Youngstown, OH). *L. braziliensis guyanensis* strain 151M, which was used for sequencing conserved regions of the kinetoplast minicircle DNAs (kDNAs), was isolated from a Brazilian man and typed by isoenzymes by Dr. William O. Rogers (Harvard University, Boston, MA). Reference strains used as con-
controls for isoenzyme typing and for the PCR included M2903 for *L. braziliensis braziliensis* and PH8 for *L. mexicana amazonensis*.

**Infection of Mice and Harvest.** Groups of four to five BALB/c mice were injected subcutaneously in the hind footpad with either 10^6 *L. braziliensis braziliensis* (WR604 strain) or 10^6 *L. braziliensis* mixed with the lysate of one half of one salivary gland of the sand fly *Lutzomyia longipalpis*. Lesion development was followed by measuring the thickness of the injected footpad with a vernier caliper compared with the thickness of the contralateral uninfected footpad (8). After 2-4 mo mice were killed and the number of parasites within the lesions was determined by limiting dilution analysis (8). Cultured parasites were typed by isoenzymes, monoclonal antibodies, and the PCR. For histology, the infected footpads and draining LN were removed, fixed in formalin, embedded in paraffin, and stained with hematoxylin and eosin or with Giemsa.

**DNA Probe Analysis.** To obtain primers for the PCR that were specific for *L. braziliensis* minicircle DNA and did not prime *L. mexicana* minicircles, kDNA was isolated from *L. braziliensis guyanensis* strain 151M, cut with *MspI* and cloned into the phagemid vector Bluescript. The conserved regions of three *L. braziliensis* kDNAs were sequenced and compared with the sequences of the conserved regions of five *L. mexicana* kDNAs (9). These *L. braziliensis* kDNA sequences will appear in the EMBL, Genbank, and DDBJ Nucleotide data base with the following accession numbers: X54470, X54471, and X54472. In Fig. 1, conserved regions of single *L. braziliensis* and *L. mexicana* minicircles are aligned and the location of *L. braziliensis*-specific primers and probe and *L. mexicana*-specific primers and probe indicated.

For PCR, cultured *Leishmania* were pelleted and DNA extracted in 100 µl of 10 mM Tris, 10 mM EDTA, 1% Triton-X 100, and 100 µl/m proteinase K for 2 h at 50°C (11). After heat inactivation, *Leishmania* target DNAs were amplified using the PCR, Taq polymerase, and the *L. braziliensis*-specific primers LB1 and LB2 or *L. mexicana*-specific primers LM1 and LM2 (Fig. 1). 30 cycles were performed of denaturation at 94°C for 30 s, annealing at 45°C for 120 s, and elongation 72°C for 12 s. PCR products were identified by dot blotting to BioTran membranes, and hybridization with 32P-kinased oligonucleotide probes using a 50°C temperature for annealing and washing with 2 × SSPE and 0.5% SDS (SSPE is composed of 0.36 M NaCl, 20 mM NaH2P04, 2 mM EDTA, pH 7.4 [reference 11]). Alternatively, dot blots of PCR products were hybridized with radiolabeled kDNAs of *L. braziliensis* and *L. mexicana*, generously donated by Dr. Mark Rodgers (Harvard University (10)).

**Isoenzyme Analysis.** Cultured parasites were lysed and isoenzyme profiles prepared as described by Kreuster and Christensen (12). Reference strains run in parallel were *L. braziliensis* and *L. mexicana*. Malic enzyme (1.1.1.40) was chosen to discriminate New vs. Old World *Leishmania*. 6-Phosphogluconic dehydrogenase (E.C. 1.1.1.44) was chosen to distinguish *L. mexicana* from *L. braziliensis* parasites.

![Figure 1](image.png)  
**Figure 1.** DNA sequence of the conserved region of the *L. braziliensis* (*L.b*) kineoplast minicircle aligned with that of *L. mexicana* (*L.m*). Xs mark identical bases between the two sequences. Blocks and arrows show location and direction of primers and probes specific for *L. braziliensis* or *L. mexicana* kDNAs. The sequences of the kDNA primers and probes are listed in Materials and Methods.

![Figure 2](image.png)  
**Figure 2.** Enhancement of *L. braziliensis braziliensis* infection in BALB/c mouse footpads by co-injection of parasites with salivary glands lysates of *L. longipalpis*. Mice injected with one million *L. braziliensis braziliensis* parasites with the lysate of one half of a salivary gland (■) had lesions which grew larger with time and showed no evidence of resolution. In contrast, mice injected without salivary gland lysates (□) had lesions that were smaller and resolved with time.
Figure 3. Histopathology of cutaneous mouse footpad lesions caused by *L. braziliensis braziliensis* parasites coinjected with sand fly salivary gland lysates. (A) At the site of the injection, there was a nodular infiltrate (N) that involved the dermis and the subcutaneous tissue. This nodule was primarily composed of sheets of macrophages within which were large vacuoles containing *L. braziliensis* amastigotes (B). There were also (C) focal accumulations of lymphocytes (L) and (D) microabscesses filled with polymorphonuclear leukocytes (P). (E) There was a marked sinus histiocytosis in the ipsilateral inguinal LN (S). (F) Within the expanded sinuses were macrophages containing *L. braziliensis* amastigotes. (A and E) × 100. (B, D, and F) ×1,800. (C) ×750.

*mAb Analysis.* Cultured parasites were dried onto glass slides, incubated with 1:100 dilutions of mAbs specific for *L. braziliensis braziliensis* (B-16), *L. braziliensis panmenensis* (B-11), *L. braziliensis guyanensis* (B-19), *L. donovani* (D-2), and *L. mexicana* (M-9), generously donated by Dr. Diane McMahon-Pratt (Yale University, New Haven, CT) (13). Bound antibodies were detected with FITC-conjugated sheep anti-mouse antibodies and read blind with an Orthoplan fluorescence microscope (E. Leitz, Inc., Rockleigh, NJ).
Results and Discussion

Sand Fly Salivary Glands Dramatically Increase the Size of L. braziliensis braziliensis Cutaneous Lesions in Mice and the Number of Parasites Present. To develop a mouse model of L. braziliensis braziliensis, Leishmania parasites were injected into BALB/c mice in association with a salivary gland lysate of L. longipalpis, a sand fly vector for visceral leishmaniasis in South America. Coinjection of the sand fly saliva resembles natural Leishmania infection by means of a sand fly bite and has been shown previously to greatly increase the infectivity of L. major in mice (8). When L. braziliensis were injected into BALB/c mice with saliva, cutaneous lesions appeared after 3 wk and progressed with time (Fig. 2). In contrast, when L. braziliensis were injected without saliva, transient lesions formed that rapidly regressed (Fig. 2). Similarly, the number of parasites recovered from the BALB/c lesions caused by L. braziliensis braziliensis plus salivary glands after 8 wk was 20 ± 7 million (average ± SD), while <100 were recovered from L. braziliensis without saliva. Recently, we have also found that L. longipalpis salivary gland lysates enhance infection of BALB/c mice with L. braziliensis braziliensis strain Ltb111 and with L. mexicana strain PH8. Salivary gland lysates also produce larger lesions of L. mexicana in C3h mice, which are relatively resistant to this parasite.

Histopathology of L. braziliensis braziliensis Lesions in Mice Injected with Sand Fly Glands. L. braziliensis braziliensis injected with salivary gland homogenates produced cutaneous nodules, which involved the dermis and the underlying soft tissue of the BALB/c mouse footpad (Fig. 3 A). These nodules were filled with macrophages, each of which contained multiple L. braziliensis braziliensis amastigotes (Fig. 3 B). In addition there were focal accumulations of lymphocytes (Fig. 3 C) and occasional microabscesses composed of polymorphonuclear cells (Fig. 3 D). The draining inguinal LN were greatly enlarged and showed a marked sinus histiocytosis (Fig. 3 E). Within the sinuses were many macrophages containing L. braziliensis braziliensis parasites (Fig. 3, F). The footpad nodules in mice injected with L. braziliensis braziliensis and salivary glands are reminiscent of those caused by L. mexicana in susceptible mice (2). They are distinct from the lesions caused by L. braziliensis braziliensis in hamsters (6), which contain few parasites.

DNA Probe Analyses Show that the Parasites Recovered from the Mouse Lesions Are L. Braziliensis. To confirm that the Leishmania causing the progressive lesions in mice coinjected with sand fly salivary glands were caused by L. braziliensis and not by a stock of L. braziliensis contaminated with other species of Leishmania, parasites were isolated from mouse lesions and typed by DNA probes. Oligonucleotide primers and probes specific for L. braziliensis and of L. mexicana parasites were identified by comparing 300-bp-long conserved portions of the kinetoplast minicircle DNAs, which are highly repeated in the parasites and have previously been used to speciate Leishmania (Fig. 1 and reference 10). Leishmania from mice injected with or without salivary glands were confirmed to be L. braziliensis, because they gave a PCR product with the L. braziliensis-specific primers that was recognized by the radiolabeled L. braziliensis-specific oligonucleotide probe or by radiolabeled L. braziliensis kDNA (Fig. 4, A and B). Like the reference L. braziliensis strain M2903, the L. braziliensis recovered from the mouse did not produce a PCR product with the L. mexicana-specific primers recognized by pMAT13A oligonucleotide probe or by L. mexicana kDNA (Fig. 4, C and D). In contrast, L. mexicana reference strain PH8 DNA produced a PCR product recognized by the oligonucleotide

![Figure 4](image-url)
References

1. Walton, B.C. 1989. Leishmania. A worldwide problem. Int. J. Dermatol. 28:305.
2. Andrade, Z.A., S.G. Reed, S.B. Roters, and M. Sadigursky. 1984. Immunopathology of experimental cutaneous leishmaniasis. Am. J. Pathol. 114:137.
3. Neal, R.A., and C. Hale. 1983. A comparative study of the susceptibility of inbred and outbred mouse strains compared with hamsters to infection with New World cutaneous leishmaniasis. Parasitology. 87:7.
4. Childs, G.E., L.K. Lightner, L.A. McKinney, M.G. Groves, E.E. Price, and L.D. Hendricks. 1984. Inbred mice as model hosts for cutaneous leishmaniasis. Ann. Trop Med. Parasitol. 78:25.
5. Gorczynski, R.M. 1985. Immunization of susceptible BALB/c mice against Leishmania braziliensis. Cell. Immunol. 94:1.
6. McKinney, L.A., and L.D. Hendricks. 1980. Experimental infection of Mystomys albocaudatus with Leishmania braziliensis: pathology. Am. J. Trop. Med. Hyg. 29:753.
7. Ribeiro, J.M.C., P.A. Rossignol, and A. Spielman. 1986. Blood-finding strategy of a capillary-feeding sandfly, Lutzomyia longipalpis. Comp. Biochem. Physiol. 83:683a (Abstr.).
8. Titus, R.G., and J.M.C. Ribeiro. 1988. Salivary gland lysates from the sand fly Lutzomyia longipalpis enhance Leishmania infectivity. Science (Wash. DC). 239:1306.
9. Rogers, W.O., and D.R. Wirth. 1987. Kinetoplast DNA minicircles: regions of extensive sequence divergence. Proc. Natl. Acad. Sci. USA. 84:565.
10. Rodgers, M.R., S.J. Popper, and D.F. Wirth. 1990. Amplification of kinetoplast DNA as a tool in the detection and diagnosis of Leishmania. Exp. Parasitol. 71:267.
11. Innis, M.A., D.H. Gelfand, J.J. Sninsky, and T.J. White. 1990.

Leishmania recovered from the mice and the control L. braziliensis braziliensis strain M2903 both reacted positively with the mAb specific for L. braziliensis braziliensis, but did not react with mAbs specific for L. braziliensis guianensis, L. braziliensis panamanensis, L. donovani, or L. mexicana. Control L. mexicana organisms did not bind the L. braziliensis braziliensis-specific mAb. These results confirm the DNA probe and isoenzyme identification of the L. braziliensis braziliensis causing the mouse lesions.

Final Comments. The results presented here show that coinfection with sand fly glands dramatically increases the size and longevity of cutaneous L. braziliensis braziliensis lesions in BALB/c mice. To our knowledge, this is the first mouse model of L. braziliensis braziliensis infection. The mechanism by which sand fly saliva produces its effect is not known. Recent experiments suggest that the sand fly gland material may hinder the development of the mouse immune response to the parasites (14). Now that a mouse model showing progressive L. braziliensis braziliensis cutaneous lesions is available, these immunological questions can begin to be addressed. We are also investigating whether parasites isolated from human mucocutaneous lesions will produce mucocutaneous lesions in mice injected with the parasites plus salivary glands. If this is the case, a murine model for mucocutaneous disease will also be available.

This work was supported by National Institutes of Health grant AI-27511, by the John D. and Catherine T. MacArthur Foundation grant AI-24511, and by the United Nations Development Program/World Bank/World Health Organization Special Program for Research in Tropical Diseases. J. Samuelson and E. Lerner were supported by the Burroughs Wellcome Life Science Research Foundation and the Lucille P. Markey Charitable Trust awards, respectively.

Address correspondence to John Samuelson, Department of Tropical Public Health, Harvard School of Public Health, 655 Huntington Avenue, Boston, MA 02115.

Received for publication 13 August 1990.
12. Kreutzer, R.D., and H.A. Christensen. 1980. Characterization of *Leishmania* spp. by isoenzyme electrophoresis. *Am. J. Trop Med. Hyg.* 29:199.

13. Grimaldi, G., Jr., J.R. David, and D. McMahon-Pratt. 1988. Identification and distribution of New World *Leishmania* species characterized by serodeme analysis using monoclonal antibodies. *Am. J. Trop Med. Hyg.* 36:270.

14. Titus, R.G., and J.M.C. Ribeiro. 1990. The role of vector saliva in transmission of arthropod-borne disease. *Parasitol. Today.* 6:157.