Spectrophotometric determination of Lorazepam in pharmaceutical tablets using batch and reverse flow injection methods

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A reverse flow injection analysis (RFIA) spectrophotometric method have been developed and compared with batch method for the determination of lorazepam (LOR) at the microgram level in pure and pharmaceutical dosage forms. The methods are based on the reaction of LOR with 3-methyl-2-benzothiazolinone hydrazone hydrochloride (MBTH) in the presence of ferric chloride in aqueous medium. The water soluble green color product was measured at \( \lambda_{\text{max}} = 659 \text{ nm} \). Under the optimum conditions, linearity was observed from 2 to 40 and 25 to 400 \( \mu \text{g/mL} \) LOR with detection limits of 0.61 and 2.29 \( \mu \text{g/mL} \) LOR by batch and RFIA procedures respectively. All the chemical and physical parameters related with reverse flow system were carefully considered and both methods were successfully applied for the determination of LOR in its dosage forms. These simple and high throughput methods could be utilized for pharmaceutical analysis of LOR.

1. Introduction

Lorazepam (7-chloro-5-(2-chlorophenyl)-3-hydroxy-2,3-dihydro1H-1,4-benzodiazepin-2-one) is a benzodiazepine indicated for the treatment of anxiety disorders and preoperative sedation [1]. LOR has the chemical formula \( \text{C}_{15}\text{H}_{10}\text{Cl}_{2}\text{N}_{2}\text{O}_{2} \) (Fig. 1).

It is applied in a wide range of indications and patient categories, because of its sedative and anticonvulsant properties. Within pediatrics, it is administered to children with status epilepticus and for intubated and mechanically ventilated patients [2], Lorazepam is inexpensive, long acting (up to 72 h) [3] and has less risk of seizure recurrence. Lorazepam has a shelf life of 3 years and can be given intravenously (IV), intramuscularly (IM), and intranasal (IN) [4].

Determination of LOR concentration in pharmaceutical preparations was previously done using potentiometry and multi-wavelengths spectrophotometry [5], high-performance liquid chromatography (HPLC) [6–8], gas chromatography/mass spectrometry (GC-MS) [9–11], micellar electrokinetic capillary chromatography [12] and voltammetry [13]. Among various methods available for trace analysis, FI analysis methods provide a fast, simple and inexpensive analysis of trace amounts of different chemicals and pharmaceutical forms [14]. The literatures reported only a few FIAs methods [15,16] available for the determination of LOR, one of them [15] was not specific, only one wavelength in the UV region being chosen for the spectrophotometric detection of LOR. Until now the literature contains no reverse flow injection-spectrophotometric methods for the determination of LOR in pure or dosage forms. This study describes fast and simple batch and reverse FI methods as a direct methods that utilize a colorimetric reaction for the determination of LOR in pharmaceutical preparations.

2. Experimental

2.1. Apparatus

- A digital double beam spectrophotometer a type of Shimadzu UV–VIS 260 (Shimadzu, Kyoto-Japan) had been used for all spectral and absorbance measurements of FI procedures. The absorbance measurements were carried out using 1 cm path length of quartz flow matched cells (Cecil, 50 \( \mu \text{L} \) internal volume), and for the absorbance measurements of the batch method, a matched 1 cm silica cells were used.

- A peristaltic pump of six channels (Ismatec, Labortechnik-Analytic, type CH-8152, Glatbrugg Zurich-Switzerland) used for pumping the solutions of reagents. A 6-ways injection valve with different loops (Rheodyne, Altex 210, Supelco -USA) was used for injected samples.

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Fig. 1. Chemical structure of lorazepam.

A flexible vinyl tubing (0.8 mm i.d.) was used for the peristaltic pump, while a tubes of Teflon material with an internal diameter of 0.5 mm was used for made different lengths of reaction coil (RC).

A two-channel manifold (Fig. 2) was employed for the RFIA spectro-photometric determination of LOR drug. The solution of drug (LOR) was transported through one of the two channels in this setup. On the other hand, the second channel was used to transport ferric chloride solution. The reagent solution of methyl-2-benzothiazolinone hydrazone hydrochloride (MBTH) was mixed with oxidant in the reaction coil. The resultant solutions were propelled by peristaltic pump at a total flow rate of 0.6 mL/min into the detector. A green colored product was formed and absorbance was measured at 659 nm.

2.2. Materials and chemicals

An analytical grade reagents, chemicals and distilled water were used usually through the work. Pharmaceutical grade LOR and excipients were received from state company for Drug Industries and Medical Appliance, SDI, Samara-Iraq. Pharmaceutical dosage forms containing LOR were purchased from local commercial sources. Two generic versions each contains LOR as its active constituent were used for analysis in this study (Lorativan®, 2 mg, Domina pharmaceuticals, Syria and Lorazesam®, 1 mg, SDI, Iraq).

2.2.1. LOR solution (500 µg/mL)

For batch and RFIA methods, LOR standard solution (500 µg/mL) was freshly prepared by dissolving 0.025 g of pure LOR powder in 50 mL ethanol. More dilute solutions can be prepared daily by appropriate dilution using distilled water.

2.2.2. Methylbenzothiazoline-2-one hydrazone hydrochloride (0.05 M)

The acidic solution of this reagent was prepared by dissolving 0.025 g of MBTH (Sigma Aldrich, Missouri, United States) in a 100 mL of 0.2 M HCl (Merck, Germany). More dilute solutions can be prepared daily by appropriate dilution using the same solvent (this solution is stable more than two weeks).

2.2.3. Ferric chloride (2% V/V)

This solution was prepared by diluted 2 mL of ferric chloride solution (60% W/V FeCl₃, BHD, UK) in 100 mL distilled water for batch and RFIA methods. More dilute solutions can be prepared daily by appropriate dilution using the same solvent.

2.2.4. Solutions of pharmaceutical dosage forms

Twenty Lorativan retard tablets (2 mg LOR) or forty Lorazesam tablets (1 mg LOR) were weighed and finally powdered. An amount of resultant powder equivalent to 0.0125 g of LOR was dissolved in 25 mL of ethanol and the resultant solution was shaked and filtered into a 50 mL volumetric flask, the residue was washed and diluted to volume with ethanol to obtain 250 µg/mL of LOR. More dilute solutions can be prepared daily by appropriate dilution using distilled water for batch and RFIA methods.

3. Procedures

3.1. General batch procedure

An aliquot of sample containing 20–400 µg of LOR was transferred into a series of 10 mL standard flasks. A volume of 2.0 mL of 0.02 M MBTH solution, and 1.5 mL of 2% V/V ferric chloride solution were added. The contents of the flasks were diluted to the mark with distilled water, mixed well and kept for 10 min. The absorbance was measured at 659 nm (at room temperature 25°C) against reagent blank containing all materials except LOR. The drug concentration was computed using a standard calibration graph. For the optimization of conditions and in all subsequent experiments, a solution of 25 µg/mL was used.

3.2. General RFIA procedure

Working solutions of LOR in the range 25–400 µg/mL were prepared from stock solutions. A 150 µL portion of 0.04 M of MBTH was injected into the stream of LOR which then mixed with the oxidant solution (ferric chloride of 8% V/V), with a total flow rate of 0.6 mL/min (Fig. 2). The resulting absorbance of the green product was measured at 659 nm and a calibration graph was shown in Table 1. Optimization of conditions was carried out on 100 µg/mL of LOR.

4. Results and discussion

4.1. Reaction mechanism of the method

LOR was found to react directly with MBTH in the presence of ferric chloride as oxidant agent to produce a green colored product with absorbance measured at λmax 659 nm. MBTH has been utilized for the spectrophotometric determination of several other compounds [17]. An electrophilic intermediate was formed when

Table 1

| Parameter | Batch procedure | RFIA procedure |
|-----------|----------------|----------------|
| Regression equation | y = 0.0166x + 0.1268 | y = 0.0015x + 0.1008 |
| Linear range (µg mL⁻¹) | 2–40 | 25–400 |
| Correlation coefficient (R²) | 0.9965 | 0.9996 |
| Limit of detection (s/n = 3) (µg mL⁻¹) | 0.614 | 2.287 |
| Limit of quantification (µg mL⁻¹) | 2.047 | 7.624 |
| Reproducibility % | <0.321 | <0.913 |
| Average of recovery, % | 101.165 | 99.560 |
| Through-put (sample per hour) | 6 | 26 |
MBTH loses two electrons and one proton under oxidation process, which can be substituted on LOR under the reaction conditions to form a green colored product at room temperature. The proposed pathway of the reaction [18] is shown in Fig. 3.

All the variables that may affecting mainly the sensitivity and stability of the colored product were optimized by variable one parameter with the time, and keeping others fixed. A typical spectrum for LOR reaction product against reagent blank is shown in Fig. 4.

4.2. Optimization of reaction conditions

4.2.1. Batch spectrophotometric method

The proposed method was optimized to achieve complete reaction formation, the highest sensitivity and maximum absorbance. The best experimental conditions for the determination of LOR were established for MBTH 0.02 M (from 0.25 to 3 mL), FeCl₃ 2% V/V (from 0.1 to 2.5 mL) by adding various volumes of their solutions to a fixed concentration of LOR and measuring the absorbance at 659 nm. The results obtained (Fig. 5) showed that 2.0 mL of 0.02 M of MBTH and 1.5 mL of 2% V/V of FeCl₃ gave the maximum color intensity and led to the maximum color stability of the dye product for 25 μg mL⁻¹ of LOR.

The color intensity reached a maximum after LOR (25 μg/mL) had been reacted immediately with MBTH and FeCl₃ in aqueous medium and became stable after 10 min, and it remained stable for 50 min at least. The effect of order of addition is also studied using different order of addition and according to the results, it was found that the order of addition of reagents (Drug + MBTH + OX) as shown in Fig. 6, gave the maximum absorbance and stability in measurement.

Three different temperatures (5, 25 and 65 °C) were used to study effect of temperature on the reaction. In practice, a higher absorbance was obtained when the color was developed when the calibrated flasks placed in a water bath at (65 °C) than at room temperature (25 °C) or at (5 °C). Further study shows that the colored product was unstable and dissociation quickly at higher temperature, therefore the room temperature was chosen for carried out all the next experiments. The structure of the product was adopted based on the mole continuous variation method (Job’s method), using 1 × 10⁻³ M solution of equimolar of each LOR and MBTH reagent. The results obtained in Fig. 7 show that 1:1 LOR to reagent was formed.
The regression equation was derived using the least-squares method. The intercept, slope, and correlation coefficient values for the calibration curve are calculated and listed in Table 1 and also compared with the RFIA proposed method.

In order to examine the usefulness of the method, and freedom from interference by tablets additives, a recovery testing for LOR was accomplished in the existence of (10-fold) of excipients. The good percentage recoveries were obtained indicating no interference was observed and the method have good selectivity for analysis the LOR in its dosage forms (Table 2).

4.2.2 RFIA-Spectrophotometric determination

The previous method (batch) for the assay of LOR, was used as the basis for the development of reverse flow injection (RFIA) procedure. MBTH has been frequently utilized for the spectrophotometric determination of several compounds, but this reagent is expensive, therefore, a reverse flow manifold had been adopted for estimation LOR. First of all the selection of the best manifold of flow system is very essential, therefore various manifolds reaction were used to perform different paths and the results obtained showed that the manifold in Fig. 6 gave the best absorbance for reaction, and was chosen for further use. Maximum absorbance intensity was obtained when the 150 µL of 0.04 M of MBTH was injected into the stream of 100 µg/mL LOR which then mixed with the oxidant solution (ferric chloride of 8% V/V) in reaction coil as given in Fig. 2. Each sample was injected three times and different physical or chemical parameters were optimized by changing individually while the others were kept constant.

4.2.3 Optimization of the RFIA system conditions

4.2.3.1 Effect of the chemicals variables. Chemical parameters were also optimized and studied to improve the sensitivity of the adopted reaction. The best concentration of the reagent (MBTH) was studied through injected different concentrations (from 0.005 to 0.05 M) through the injection valve. The results in Fig. 8 indicated that the concentration of 0.04 M gave a high absorbance and good precision and was chosen for further use.

Different concentration of oxidant (ferric chloride) were also studied in the range of concentration (from 1 to 10% V/V) FeCl₃ were investigated; the optimum concentration of 8% V/V gave the best results and minimum blank value as shown in Fig. 9 and was considered as optimum value.

4.2.3.2 Effect of the physical variables. The physical parameters were also studied and different coil lengths using for mixing the reagents and complete the reaction was studied in the range of 50–200 cm. The results obtained showed that a coil length of 80 cm gave the highest absorbance as shown in Fig. 10 and was used then in all experiments. After this length the absorbance of colored product decrease with increase coil length because of increase the dilution resulting from dispersion.

The effect of different flow rate (Fig. 11) on the reaction was studied in the range of 0.6–4.2 mL/min. The results obtained

Table 2

| Excipient (250 µg/mL) | Found LOR, µg/mL | Recovery ± SD, %** |
|----------------------|------------------|-------------------|
| Poly vinyl pyrrolidone | 25.19            | 100.76 ± 0.36    |
| Lactose              | 25.32            | 101.28 ± 0.22    |
| Talc                 | 25.22            | 100.88 ± 0.13    |
| Magnesium stearate   | 25.46            | 101.84 ± 0.08    |
| Total (contains all previous excipients) | 26.09            | 104.36 ± 0.17    |

* Present 25 µg/mL of LOR.
** Average of five determinations.
showed that the absorbance of the total flow rate of 0.6 mL/min gave the highest absorbance and good precision. After this value the absorbance decrease with increase the flow rate because the high speed would never get the reactants a sufficient time for mixing (i.e. the sample zone would form and get out the flow cell quickly) therefore, 0.6 mL/min, (total flow rate) was selected to be the best value.

The volume of the injected sample was investigated using five sample loops (75–200 μL) and the results obtained showed that the injected sample of 150 μL gave the best absorbance and was chosen for further experiments (Fig. 12).

After studied all variables, and under the optimized conditions (Table 3) the sampling frequency or sampling rate can be calculated theoretically by measuring the time calculated from the moment of sample injection to the appearance of the maximum absorbance. The calculated time was 2.13 min; therefore the sampling frequency was 28 sample per hour.

4.3. Validation of the current method

Analytical characteristics such as linearity, detection limit, sampling rate, correlation coefficient and relative standard deviation (RSD) of each method were determined for the above optimized conditions as shown in Table 1. A linear range of the calibration graph was obtained using a series of solutions of LOR (25–400 μg/mL) which prepared from the stock solution. In comparison of the batch with RFIA procedure, the later is more convenient than the former method because of its speed (sample throughput of 26 injections per hour) and wider linear range of calibration graph although the batch method is more sensitive than flow injection method.

4.4. Accuracy and precision

Evaluation of the accuracy and precision of the proposed methods, accomplished by prepared and analyzed solutions containing three different concentrations of LOR were in five replicates. The analytical results of this investigation are summarized in Table 4. The low values of percentage relative standard deviation (% RSD) as precision and percentage relative error as accuracy indicated that the proposed methods show a good accuracy and precision.

4.5. Analytical applications

Batch and RFIA methods were also successfully applied for determination of LOR in pharmaceutical dosage forms (tablets) by the analysis of two different concentrations of pharmaceutical preparations using the analytical procedures directly. The results are given in Table 5.

To examine the efficiency and success of the suggested methods, the obtained results compared with those obtained from applying the USP standard method [19]. The performance of the current method was refereed by calculating the Student’s t-and F-values[20]. Values of calculated t-and F-tests did not exceed the theoretical values as illustrated by Table 6.

| Table 3 | Optimum conditions for the determination of LOR. |
|---------|--------------------------------------------------|
| Variable | Studied range | Optimum values |
| Concentration of FeCl₃ (% v/v) | 1–10 | 8 |
| Concentration of MBTH (M) | 0.005–0.05 | 0.04 |
| Reaction coil (cm) | 50–200 | 80 |
| Total flow rate (mL/min) | 0.6–4.2 | 0.6 |
| Injected reagent volume (μL) | 75–200 | 150 |

| Table 4 | Accuracy and precision of the proposed methods. |
|---------|-------------------------------------------------|
| Method | Conc. of LOR, μg/mL | RE% | Rec.% | RSD% |
| Added | Found | |
| Batch | 15 | 15.18 | 1.20 | 101.20 | 0.32 |
| 30 | 30.34 | 1.13 | 101.13 | 0.31 |
| RFIA | 100 | 99.47 | –0.53 | 99.47 | 1.89 |
| 250 | 249.13 | –0.35 | 99.65 | 0.91 |

* Average of four determinations.
Table 5
Application of the proposed and official methods to the determination of LOR in different dosage forms.

| Dosage form | Proposed methods | Official method [19] |
|-------------|------------------|---------------------|
|             | Batch            | RFIA                |
|             | Conc. (µg/mL)    | Rec. (%) | RSD (%) | Conc. (µg/mL) | Rec. (%) | RSD (%) |
|             | Taken | Found | Taken | Found | Taken | Found |
| Lorazepam® (2 mg) | 10 | 10.28 | 102.80 | 0.11 | 75 | 75.49 | 100.65 | 0.56 |
|             | 15 | 15.13 | 100.87 | 0.16 | 100 | 101.11 | 101.11 | 0.56 |
| Lorazepam® (1 mg) | 10 | 10.12 | 101.20 | 0.49 | 75 | 75.73 | 100.97 | 1.28 |
|             | 15 | 15.30 | 102.00 | 0.25 | 100 | 101.35 | 101.35 | 0.55 |

* Average of five determinations.

Table 6
The statistical comparison between the current methods and standard method.

| Dosage form | %Recovery ± SD |
|-------------|----------------|-----------------|
|             | Batch | RFIA | Official method |
| Lorazepam (pure form) | 101.17 ± 0.05 | 99.56 ± 0.13 | 100.05 ± 0.23 |
| Lorazepam® (2 mg) | 101.84 ± 1.37 | 100.88 ± 0.33 | 102.55 ± 0.35 |
| Lorazepam® (1 mg) | 101.60 ± 0.57 | 101.16 ± 0.27 | 101.73 ± 1.31 |
| t (2.776)** | 0.123 | 1.027 |
| F (19.000)** | 14.094 | 2.224 |

* For five determinations. Conc., concentration; RSD, relative standard deviation. ** Theoretical value.

5. Conclusion

The literatures reported only a few spectrophotometric methods and two FIA methods for the determination of LOR in dosage forms and in biological fluids and these methods either need extraction or using a complicated and expensive techniques and until now the literature contains no reverse flow injection-spectrophotometric method for the determination of LOR in pure or dosage forms. The suggested methods, which are simple and rapid, offers the advantages of sensitivity more than the reported methods, and a wide range of determination without the need for extraction or heating. The wide applicability of the new method for routine quality control is well established by the assay of LOR at microgram concentration level in pharmaceutical formulations.

The reaction can be carried out in batch and RFIA and in this paper both approaches were compared.

References

[1] Lexicomp Online®, Pediatric & Neonatal Lexi-Drugs®, Lexi-Comp Inc, Hudson, Ohio; 2016.
[2] van der Vossen AC, van der Velde I, Smeets OSNM, Postma DJ, Eckhardt M, Vermes A, et al. Formulating a poorly water soluble drug into an oral solution suitable for paediatric patients: lorazepam as a model drug. Eur J Pharm Sci 2017;100:205–10.
[3] Joint Formulary Committee. British National Formulary. London: BMJ Group and Pharmaceutical Press; 2012.
[4] Björnsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson S. Incidence, presentation and outcomes in patients with drug-induced liver injury in the general population of Iceland. Gastroenterology 2013;144:1419–25.
[5] Yazdanipour A. Study of protonation equilibrium oflorazepam by potentiometry and multi-wavelengths spectrophotometry. J Iran Chem Res 2009;2:293–7.
[6] Feng J, Shen R, Zhang D, Dong Z, Zhang DD, Wang H. Quantification of Lorazepam in human plasma using HPLC: application to a pharmacokinetic study. Lat Am J Pharm 2015;34(3):622–6.
[7] Mashayekhi HA, Khalilian F. Development of solid-phase extraction coupled with dispersive liquid-liquid microextraction method for the simultaneous determination of three benzodiazepines in human urine and plasma. J Chroma Sci 2016;54(6):608–1073.
[8] Srikantha D, Raju RR. Development and validation of RP-HPLC method for lorazepam in tablet dosage form. Int J Ayu Pharm Chem 2014;1(1):65–75.
[9] Goldberger BA, Chronister CW, Vermes ML. Quantitation of benzodiazepines in blood and urine using gas chromatography-mass spectrometry (GC-MS). Methods Mol Biol 2010;603:75–87.
[10] Papoutsis II, Athanaselis SA, Nikolaou PD, Pistos CM, Spiliopoulou CA, Maravelias CP. Development and validation of an EI-GC-MS method for the determination of benzodiazepines and their metabolites in blood: applications in clinical and forensic toxicology. J Pharm Biomed Anal 2010;52(4):609–14.
[11] Acikkol M, Mercan S, Karadayi S. Simultaneous determination of benzodiazepines and ketamine from alcoholic and nonalcoholic beverages by GC-MS in drug facilitated crimes. Chromatograph 2009;70:1295–303.
[12] Nevada JIB, Penalvo GC, Calderon MSP. Determination of lorazepam and its main metabolite in serum using micellar electrocapillary chromatography with direct injection and ultraviolet absorbance detection. J Chromatogr B 2002;773:151–6.
[13] Carvalho LMD, Correia D, Garcia SC, Bairros AVD, Nascimento PCD, Bohrer D. A new method for the simultaneous determination of 1,4-benzodiazepines and amphetamine as adulterants in phytotherapeutic formulations by voltammetry, Forensic Sci Int 2010;202:75–81.
[14] Thongsrisomboon P, Laiwruangrath B, Laiwruangrath S, Satien-peraku S. Determination of nitrofurans residues in animal feeds by flow injection chemiluminescence procedure. Food Chem 2010;123:834–9.
[15] Frada D, Lorenzo E, Hernandez M, Hernandez L. Rapid determination of some 1,4-benzodiazepines by flow injection analysis. Microchem J 1988;37:257–62.
[16] Norouzi P, Ganjali MR, Mebodi ASE. A novel adsorptive square wave voltammetric method for pico molar monitoring of lorazepam at gold ultra microelectrode in a flow injection system by application of fast fourier transform analysis. Anal Lett 2008;41:1208–24.
[17] Malikarjuna H, Shrivaprasad KH, Venugopala Reddy KR, Lokesh KS. Spectrophotometric determination of somenosteroid anti-inflammatory drugs by oxidative coupling reaction. Austin J Anal Pharma Chem 2016;3:1–6.
[18] Ramachandra B, Naidu NV. Spectrophotometric method for the determination of furazolidone in pharmaceutical formulations and human blood samples with MBTH. Int J Sci Res 2015;4:1026–31.
[19] USP (United States Pharmacopoeia, 30th revision); 2012.
[20] Miller JN, Miller JC. Statistics and chemometrics for analytical chemistry. Gostep, UK: Ashford Colour Press Ltd.; 2010.