Iatrogenic CJD due to pituitary-derived growth hormone with genetically determined incubation times of up to 40 years

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Patients with iatrogenic Creutzfeldt-Jakob disease due to administration of cadaver-sourced growth hormone during childhood are still being seen in the UK 30 years after cessation of this treatment. Of the 77 patients who have developed iatrogenic Creutzfeldt-Jakob disease, 56 have been genotyped. There has been a marked change in genotype profile at polymorphic codon 129 of the prion protein gene (PRNP) from predominantly valine homozygous to a mixed picture of methionine homozygous and methionine-valine heterozygous over time. The incubation period of iatrogenic Creutzfeldt-Jakob disease is significantly different between all three genotypes. This experience is a striking contrast with that in France and the USA, which may relate to contamination of different growth hormone batches with different strains of human prions. We describe the clinical, imaging, molecular and autopsy features in 22 of 24 patients who have developed iatrogenic Creutzfeldt-Jakob disease in the UK since 2003. Mean age at onset of symptoms was 42.7 years. Gait ataxia and lower limb dysaesthesiae were the most frequent presenting symptoms. All had cerebellar signs, and the majority had myoclonus and lower limb pyramidal signs, with relatively preserved cognitive function, when first seen. There was a progressive decline in neurological and cognitive function leading to death after 5–32 (mean 14) months. Despite incubation periods approaching 40 years, the clinical duration in methionine homozygote patients appeared to be shorter than that seen in heterozygote patients. MRI showed restricted diffusion in the basal ganglia, thalamus, hippocampus, frontal and the paracentral motor cortex and cerebellar vermis. The electroencephalogram was abnormal in 15 patients and cerebrospinal fluid 14-3-3 protein was positive in half the patients. Neuropathological examination was conducted in nine patients. All but one showed synaptic prion deposition with numerous kuru type plaques in the basal ganglia, anterior frontal and parietal cortex, thalamus, basal ganglia and cerebellum. The patient with the shortest clinical duration had an atypical synaptic deposition of abnormal prion protein and no kuru plaques. Taken together, these data provide a remarkable example of the interplay between the strain of the pathogen and host prion protein genotype. Based on extensive modelling of human prion transmission barriers in transgenic mice expressing human prion protein on a mouse prion protein null background, the temporal distribution of codon 129 genotypes within the cohort of patients with iatrogenic Creutzfeldt-Jakob disease in the UK suggests that there was a point source of infecting prion contamination of growth hormone derived from a patient with Creutzfeldt-Jakob disease expressing prion protein valine 129.
Introduction

Prion diseases are neurodegenerative conditions of humans and animals caused by the templated misfolding and aggregation of cellular prion protein (Collinge, 2001). It is becoming increasingly clear that the more common degenerative brain diseases, such as Alzheimer's and Parkinson's disease, involve the accumulation of aggregates of misfolded host proteins by a process of seeded protein polymerization (Jucker and Walker, 2013). In this way, prion diseases can be considered a paradigm for the other protein misfolding diseases and prion-like mechanisms are now one of the most rapidly developing research areas in neurodegeneration research internationally. The remarkable and apparently unique phenomenon of zoonotic and horizontal transmission of prions, affords special opportunities to understand the host determinants of clinical phenotype, infection and susceptibility. Genetic variation in the prion protein gene (PRNP), specifically a common polymorphism at codon 129 encoding either methionine or valine, is a key risk factor and determinant of susceptibility, clinical phenotype and incubation time (Collinge et al., 1991, 2006; Palmer et al., 1991; Pocchiari et al., 2004; Mead et al., 2006, 2009a).

Treatment of persons of short stature with cadaver-sourced growth hormone was first given in 1958 and 1 year later in the UK as a clinical trial (Raben, 1958). In 1985, four cases of Creutzfeldt-Jakob disease (CJD) were reported in patients who had received cadaver-sourced growth hormone in the UK (Powell Jackson et al., 1985). Further cases were reported from the USA, Europe and Australia and treatment with human growth hormone was stopped in the UK in May 1985 by which time recombinant growth hormone was becoming available. Of 1849 persons who received growth hormone in the UK between 1959 and 1985, 38 were known to have developed CJD by 2000 and the estimated risk of developing iatrogenic CJD was at that time 4.5%, this risk being greatest in those patients who received treatment at ages 8–10 years with a peak incubation period of 20 years (Swerdlow et al., 2003). One preparation (Hartree modified Wilhelmi preparation) was common to all patients who developed iatrogenic CJD.

Materials and methods

Study design

A national referral system for prion diseases was set up in the UK in 2004. UK neurologists were asked by the Chief Medical Officer to refer all patients with suspected prion disease jointly to the National CJD Research and Surveillance Unit (Edinburgh, UK) and to the NHS National Prion Clinic (London, UK). This enables epidemiological surveillance, provision of specialist clinical care and also participation in clinical research and the National Prion Monitoring Cohort study; recruitment of patients to this study began in 2008. All but 2 of 18 patients with CJD in this study who had received cadaver-derived growth hormone were recruited into the Cohort study. In addition, similar patients seen by clinicians in the MRC Prion Unit/NPC as part of the PRION-1 trial of quinacrine (Collinge et al., 2009), between 2001 and 2007, were included in the current study.

After the initial visit, repeated follow-up visits were made to assess progress of the disorder at ∼6-week intervals until death. Progression was assessed by the MRC Prion Disease
Rating Scale (MRC Scale), systematic neurological and neuropsychological assessments, and a range of other rating scales (Thompson et al., 2013). Ethical approval for these studies was obtained from the Local Research Ethics Committee of UCL Institute of Neurology/National Hospital for Neurology and Neurosurgery.

**Magnetic resonance imaging**

MRI studies performed at either 1.5 or 3 T were reviewed in 20 of the patients. In six of the MRI studies, diffusion weighted images (DWI) were not available and cortical involvement could not be assessed in a comparable way. The images were reviewed by a consultant neuro-radiologist and a consultant neurologist, both with 9 years’ experience in prion disease imaging, and agreement was achieved via consensus review. Signal abnormality was assessed in the caudate, putamen and thalamus on T2-weighted, fluid attenuation inversion recovery (FLAIR) and/or DWI sequences. Cortical signal abnormality was assessed on DWI sequences where available in the following areas: frontal, parietal, temporal, occipital, cingulate, insula, hippocampus and cerebellum. Any other relevant findings were also noted.

**Neuropathology**

Autopsies were performed either at the local hospital or the National Hospital for Neurology and Neurosurgery. Autopsies were carried out in a post-mortem room designated for high risk autopsies. Informed consent to use the tissue for research was obtained in all cases. The anterior frontal, temporal, parietal and occipital cortex and the cerebellum (at the level of dentate nucleus) were dissected during the post-mortem procedure. The remaining tissue was stored permanently in 10% formalin to allow for additional tissue sampling. In cases where retention of the entire brain was not consented, additional tissue sampling of posterior frontal cortex including motor strip, basal ganglia, thalamus, hippocampus and cerebellar vermis was carried out during post-mortem dissection. Tissue samples were immersed in 10% buffered formalin and prion infectivity was inactivated by immersion into 98% formic acid for 1 h. Tissue samples were processed to paraffin wax and tissue sections were routinely stained with haematoxylin and eosin and immunostained for abnormal prion protein [anti-PrP monoclonal antibody ICSM35 (D-Gen Ltd); 0.1 μg/ml solution] according to published protocols (Wadsworth et al., 2008c). The extent of neuropil vacuolation and pattern and intensity of prion protein deposition was assessed semiquantitatively.

**Molecular strain typing of disease-related PrP**

Frozen brain tissue was available from seven patients for disease-related prion protein (PrPSc) molecular strain typing (Collinge et al., 1996; Hill et al., 2003). Biochemical studies were carried out in a microbiological containment level 3 facility with strict adherence to safety protocols. Ten per cent (w/v) brain homogenates (grey matter; frontal cortex) were prepared in Dulbecco’s phosphate-buffered saline lacking Ca2+ or Mg2+ ions using tissue grinders (Wadsworth et al., 2008c). Proteinase K digestion (50 μg/ml final protease concentration, 1 h, 37°C), electrophoresis [on 16% Tris-glycine gels (Invitrogen)] and immunoblotting using anti-PrP monoclonal antibody 3F4 (Cambridge Bioscience) was performed as described previously (Wadsworth et al., 2001, 2008c; Hill et al., 2003). Internal typing controls of defined PrPSc types were analysed on each gel when assigning PrPSc type to an unknown sample. Different human PrPSc isoforms, referred to as molecular strain types, can be identified in the brain of patients with phenotypically distinct forms of CJD and are classified by both the fragment size and ratio of the three principal PrP bands seen after protease digestion. To date we have characterized four major types of human PrPSc that can be commonly identified in sporadic and acquired human prion diseases (Collinge et al., 1996; Collinge, 2001; Hill et al., 2003; Wadsworth et al., 2003, 2008a, b), although much greater heterogeneity seems likely (Hill et al., 2003; Wadsworth and Collinge, 2011). Sporadic and iatrogenic CJD (classical CJD) and kuru are associated with PrPSc types 1–3, whereas type 4 PrPSc is uniquely associated with variant CJD. An earlier classification of PrPSc types seen in classical CJD described only two banding patterns (Parchi et al., 1996, 1999) with PrPSc types 1 and 2 that we describe corresponding with the type 1 pattern of Gambetti and Parchi and colleagues, and our type 3 fragment size corresponding to their type 2 pattern. More recently Parchi et al. (2011) have demonstrated that their PrPSc types 1 and 2 can be further classified by molecular criteria and have integrated these data with clinical-pathological variations that are seen in patients with classical CJD. Currently the number of distinct prion strains that comprise classical CJD remains unknown and a unified internationally accepted classification system of PrPSc subtypes remains an important goal (Brandner, 2011).

**Genetic analyses**

In addition to the detailed analysis of the clinical data in the 22 patients, which is the main focus of this paper, data were retrieved from the UK iatrogenic CJD database, principally to determine the genotypes in all cases of the polymorphism at codon 129 in the PRNP gene. The database contains details of growth hormone preparations received by the patients and duration of disease assessed by the local clinician. Polymorphism at codon 129 was determined in 50 of 77 patients by the MRC Prion Unit or obtained from a previous publication (Swerdlow et al., 2003); another six were assessed by other laboratories. Twenty-one patients did not have gene sequencing or screening for PRNP codon 129 polymorphism.

Whole exome sequencing was done using the Agilent SureSelect Human All Exon v2 target enrichment kit. Sequencing was performed on an Illumina HiSeq2000 and achieved an average 30-fold depth-of-coverage of target sequence.

**Results**

Details of the individual patient data are shown in Supplementary Table 1. The following is a summary of these data.
Patients

Twenty-two patients (five female) aged 27–51 (mean 42.8) years were studied sequentially from 2003. All have died. Average follow-up period until death was 7.3 months and the maximum follow-up period was 24.9 months. In a subset of patients, progression was assessed by serial measurements of the MRC Prion Disease Rating Scale (MRC Scale) (Fig. 1). There were between two and 15 follow-up assessments for each patient (average of 6.7 per person); three were assessed twice, one three times, one four times, two five times, one six times and three seven times, one nine times, two 10 times and one 15 times. The mean and median duration of illness was 16.0 (4–32) months and 14.0 months, respectively.

The precise dates of starting and stopping treatment with any type of growth hormone were known for all but one patient; details of treatment with one product (Hartree modified Wilhelmi preparation), which is most implicated as the causative agent of iatrogenic CJD in the UK, were incomplete in two patients. The range of minimum possible incubation times, calculated as the time from last injection of any type of growth hormone to onset of symptoms was 18.3–33.6 (mean 25.9) years and the range of maximum incubation periods calculated from time of first injection to onset of symptoms was 23.2–43.3 (mean 32.8) years, assuming the infection occurred at the first or last exposure. The range of incubation times from midpoint of treatment, ignoring missed injections, with any type of growth hormone to onset of symptoms was 20.6–37.6 (mean 29.3) years. Similar calculations were done for the Hartree modified Wilhelmi preparation product, the only preparation received by every patient, and the most probable source of infection. The range of minimum incubation periods was 22.1–32.8 (mean 27.9) years, maximum incubation period was 22.6–39.8 (mean 31.3) years and the range of incubation periods from the midpoint of injections was 22.5–35.8 (mean 29.9) years. The average and median age at treatment with growth hormone was 9.3 (2–16) years and 9.0 years. The cumulative dose of growth hormone given to each patient is not available but the total duration of treatment is known and has been used as a surrogate measure of total exposure (Supplementary Table 1). There was no relation between duration of dosage of any product and incubation period (r = 0.102, \(P = 0.53\) for Hartree modified Wilhelmi preparation; \(r = 0.08, P = 0.61\) for all preparations).

Symptoms

Initial symptoms are summarized in Table 1. Ataxia of gait was the first symptom in 11 of 22 patients and nearly all developed this symptom within the first 2 months. Other initial symptoms were tremor, leg pain, daytime somnolence, dizziness, headaches, myoclonus and cognitive impairment. There were two patients who described unsteadiness induced by a moving stimulus such as waves; one could not stand in the sea due to poor balance and another became unsteady when getting out of shallow water.

During the evolution of the disease all patients ultimately had unsteadiness of gait and other cerebellar symptoms such as limb dysmetria (lower limb more so than upper limb). Thirteen patients described pain in the lower limbs, including hypersensitivity to light touch in the legs and a variety of dysesthesiae such as an ‘itchy sensation’, numbness, patchy tingling and burning in the lower limbs. Two had hip pain, three had knee pain, one had both and one patient had calf pain. Ten patients had a sleep disturbance, including daytime somnolence, insomnia and early waking. Two of these patients had excessive sleepiness during the day, falling asleep in public places. There was no pattern of these symptoms occurring at any particular stage throughout the course of the illness.

Neurological signs

The neurological findings when first seen in the National Prion Clinic are summarized in Table 1. All patients had ataxia of gait, a lesser ataxia of the limbs and most had myoclonus. Half had a cerebellar dysarthria. Seventeen had evidence of pyramidal lesions with either brisk reflexes or extensor plantar responses. Eleven had reduced power in lower limbs.

Relative to the marked ataxia, cognition was initially less affected in most patients. Nineteen patients had a Mini-Mental State Examination assessed at the first visit. Fifteen of these patients scored \(\geq 18/30\) (mean 23.7 range 18–29/30).
losing points on orientation and recall. One patient could not be tested adequately as she had a history of stable long term cognitive problems. Of the other six patients three were too impaired to test and three scored 8, 7 and 4/30, respectively. There was no correlation between when cognitive function was first assessed and the duration of symptoms. To delineate further the earliest cognitive deficits a short cognitive examination, specifically designed for patients with prion disease, was done in the nine patients well enough to cooperate. Executive function assessed by letter fluency (1 min) was mildly impaired in all when first tested 3–19 (mean 8.4) months after the first symptoms. Verbal or visual recognition memory was mildly impaired in the three patients seen more than 1 year after onset of symptoms. All had preserved visual perception. Four patients had more extensive psychometric testing confirming mild memory impairment in all and mildly impaired executive function in three. Subsequently there was progressive decline in cognitive function; increasing dysarthria limited assessment later in the course of the disease.

Over a period of months the ataxia increased resulting in loss of ambulation and inability to groom, dress and feed without assistance. Leg weakness increased and contributed to the loss of ambulation. The dysarthria progressively increased and speech became unintelligible. Speech output then gradually declined making cognitive assessment difficult. Myoclonus usually persisted but the pain in the limbs lessened in most patients. Incontinence of urine and faeces developed in all. Ultimately the patient entered an akinetic mute state.

### Magnetic resonance imaging

MRI brain scans were available in 20 patients (five female) mean age 42.7 years (range 27–51 years). Eighteen of the patients scanned showed caudate, putamen and thalamic signal abnormality (Fig. 2 and Table 2). The thalamic signal abnormality was seen in all the patients where there was basal ganglia signal abnormality and was mostly diffuse with clear involvement of all the thalamic nuclei. Of the cortical areas, the cingulate gyrus most commonly showed signal abnormality (15 of 17 patients who underwent DWI). The frontal cortex was next most commonly involved (14 of 17 patients), with focal involvement of the medial aspect of the precentral gyrus and paracentral lobule bilaterally seen in 10 of 16 (Fig. 2) with relative sparing of the postcentral gyrus. In five of these patients, the abnormality was confined to the precentral gyrus. The next most commonly affected cortical area was the superior cerebellar vermis, seen in 7 of 16 patients (Fig. 3). The parietal, temporal and insula cortices were rarely involved and the occipital cortex was not involved in any of the cases (Table 2). Involvement of the hippocampus, particularly the tail, was seen in nine patients. Additional MRI findings included one patient with septo-optic dysplasia and partial agenesis of the corpus callosum.

### Prion protein gene sequencing

All patients were analysed for the entire PRNP open reading frame by Sanger sequencing. None had a mutation.
At polymorphic codon 129 there were 17 heterozygotes (MV), four methionine homozygotes (MM) and one valine homozygote (VV).

Although there were insufficient data to compare statistically survival times by 129 polymorphism, MM patients had a mean duration of 7.8 months, the VV patient 17 months and those with the MV polymorphism a mean of 18.6 months (10–32 months). In addition, the duration of disease from first symptom was significantly longer in the heterozygotes ($P = 0.02$ two-tailed $t$-test). Although there were only four patients who were MM three of these had the most rapid progression ($P = 0.04$, Mann Whitney U-test).

The average incubation time from the midpoint of growth hormone administration was 28.6 years for MV and 31.8 years for MM (four patients). The patient who carried the VV polymorphism developed symptoms after 20.6 years.

Combining the data with those already published (cases up to 2000) there were 33 heterozygotes (MV), and 23 homozygotes, 15 of whom were VV and eight MM. There was a highly significant difference in the distribution of the three polymorphisms at codon 129 over time and incubation period, with each pairwise comparison of genotypes being statistically significant in post hoc analyses (MM mean 30.8 years (95% confidence interval 26.9–32.6), MV mean 23.4 years (9.0–36.7), VV mean 14.3 years (7.7–20.2), ANOVA $P < 10^{-7}$). Fourteen of 15 valine homozygotes occurred before 1998 and seven of eight methionine homozygotes occurred after 2004 (Fig 4).

### Whole exome sequencing

Twenty-seven iatrogenic CJD cases, including the 22 in this report and one from outside the UK, were exome sequenced and allele frequencies compared with internal and publicly available control frequencies. The top-ranked single variant association was in *RRP9* (ribosomal RNA processing 9 small subunit gene), for which association testing of rs145537768, p.R143W, achieved $P = 10^{-7}$, Fisher’s exact test. While this was the strongest statistical association in the exome study it does not surpass standard statistical thresholds for a genome-wide finding ($P < 5 \times 10^{-8}$) and requires replication study in other national cohorts of iatrogenic CJD to be recognized as a confirmed association. The variant was detected in five growth hormone-related iatrogenic CJD cases ($n = 27$), confirmed in all cases by Sanger sequencing, but was not found in 574 internal non-prion disease controls.

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**Figure 2** MRI findings in a 46-year-old male patient with iatrogenic CJD due to growth hormone. Basal ganglia and thalamic hyperintensity is seen on T2-weighted images (A), DWI at $b = 1000s/mm^2$ (B) and (C) DWI at $b = 3000s/mm^2$. Cortical hyperintensity in the paracentral lobule bilaterally and right precentral gyrus is not visualized on T2-weighted images (D) but is seen on DWI $b = 1000s/mm^2$ (E) and (F) is more conspicuous at DWI at $b = 3000s/mm^2$ (Patient 12).
### Table 2 Summary of MRI findings in each patient

| Case | MRI | Caudate | Putamen | Globus Pallidus | Thalamus | Frontal Lobule | Parietal | Temporal | Occipital | Cingulate | Insula | Hippocampus | Cerebellum |
|------|-----|---------|---------|----------------|----------|---------------|----------|----------|------------|-----------|--------|-------------|-----------|
| 1    | Y limited | +       | +       | -              | +        | n/a           | n/a      | n/a      | n/a        | n/a       | n/a    | n/a         | n/a       |
| 2    | Y limited | -       | -       | -              | +        | -             | -        | -        | +          | -         | + tail | +           | +         |
| 3    | Y       | +       | +       | -              | +        | +             | -        | -        | +          | -         | +      | -           | -         |
| 4    | Y limited | +       | +       | -              | +        | n/a           | n/a      | n/a      | n/a        | n/a       | n/a    | n/a         | n/a       |
| 5    | N       |         |         |               |          |               |          |          |            |           |        |             |           |
| 6    | Y limited | +       | +       | -              | +        | n/a           | n/a      | n/a      | n/a        | n/a       | n/a    | n/a         | n/a       |
| 7    | Y       | +       | -       | +              | +        | -             | -        | -        | +          | -         | +      | +           | +         |
| 8    | Y       | +       | +       | -              | +        | -             | -        | -        | +          | +         | + tail | +           | +         |
| 9    | Y       | +       | +       | -              | +        | +             | -        | -        | -          | -         | -      | + tail +/- | -         |
| 10   | Y limited | +       | +       | -              | +        | n/a           | +        | -        | +          | -         | -      | -           | +         |
| 11   | Y       | +       | +       | +              | +        | -             | -        | -        | -          | -         | +      | -           | +         |
| 12   | Y       | +       | +       | +              | +        | -             | -        | -        | -          | -         | -      | +/-         | +         |
| 13   | Y       | +       | +       | -              | +        | -             | -        | -        | -          | -         | +      | -           | -         |
| 14   | Y       | +       | +       | +              | +        | -             | -        | -        | -          | -         | -      | -           | +         |
| 15   | Y limited | +       | +       | +              | +        | +             | -        | -        | +          | -         | -      | + tail -   | -         |
| 16   | Y       | +/-     | +/-     | +/-           | -        | +             | -        | -        | -          | -         | -      | +/-         | +/-       |
| 17   | N       |         |         |               |          |               |          |          |            |           |        |             |           |
| 18   | Y       | +       | +       | -              | +        | +             | -        | +        | +          | -         | + head | +           | +         |
| 19   | Y       | +       | +       | -              | +        | +             | -        | -        | -          | -         | +      | +           | +         |
| 20   | Y       | +       | +       | -              | +        | +             | -        | -        | +          | +         | + tail | +           | +         |
| 21   | Y       | +       | +       | +              | +        | -             | -        | -        | +          | -         | +/-    | +/-         | +/-       |
| 22   | Y       | +       | +       | +              | +        | +             | -        | -        | +          | -         | +      | -           | -         |

Y = MRI obtained; N = MRI not obtained; Y limited = limited or no DWI sequences available; + = feature present; - = feature absent; +/- = equivocal abnormality. n/a = not available.
changes a conserved amino acid residue, is predicted to be damaging by both \textit{in silico} prediction software PolyPhen-2 and SIFT, and is rare in control populations with a reported allele frequency of 0.0014 in non-Finnish Europeans (122/121370 in the entire series, http://exac.broadinstitute.org/gene/ENSG00000114767).

\section*{Electroencephalography}

Twenty-one patients had an EEG, six of which were initially normal. The other 15 had generalized slow activity and one of which had short runs of periodic slow wave complexes.

\section*{Nerve conduction studies}

No systematic peripheral nerve conduction studies were undertaken. However, four patients had been investigated by the local clinician prior to referral to the National Prion Clinic. One patient with wasting of the small hand muscles and fasciculation had neurophysiological evidence of denervation leading to an initial diagnosis of motor neuron disease. A second patient had clinical evidence of a length dependent sensorimotor axonal neuropathy, greater in the lower limbs, assessed 1 year before referral, while two other patients also had some evidence of a similar sensory motor neuropathy on limited assessment.

\section*{Cerebrospinal fluid}

Sixteen patients had CSF examination. The fluid was acellular in all. Five had a raised protein concentration (>0.41 g/l). 14-3-3 protein was sought in 14 patients and detected in six.

\section*{Neuropathology}

Nine autopsies were conducted between 2003 and 2013; the whole brain was available in eight. We report an overview here; a full morphological study will be reported separately. In all cases there was variably prominent microvacuolar degeneration in the hemispheric cortex, deep grey nuclei and cerebellar cortex. Immunostaining for the abnormal PrP revealed synaptic labelling in all grey matter areas examined and in all but one case, there were also microplaques in all grey matter structures (Fig. 5). Variability in the intensity of
Figure 5 Neuropathological findings in a representative iatrogenic CJD case. Sections (A–L) stained with haematoxylin and eosin. Sections (A¹–L¹) immunostained for abnormal prion protein deposition with anti-PrP monoclonal antibody ICSM35. Throughout the grey matter there is variably prominent microvacuolar degeneration and abnormal prion protein deposition in a diffuse synaptic manner and as micro-plaques. In the anterior frontal cortex (A and B, haematoxylin and eosin) there is mild patchy microvacuolar degeneration and abnormal prion protein deposits (A¹ and B¹, ICSM35) are restricted to the deep cortical layers. In the posterior frontal lobe, including motor cortex (C and D, (continued)
the immunoreactivity for the abnormal prion protein was evident but detailed comparison between the cases and separately within each case was not feasible as prolonged formalin-fixation in some cases significantly attenuated the immunoreactivity.

**Disease-related prion protein analysis by immunoblotting**

PrPSc typing (molecular strain typing) (Collinge et al., 1996; Hill et al., 2003) was performed with frontal cortex samples from seven patients with iatrogenic CJD where frozen tissue was available. A representative immunoblot is shown in Fig. 6. All seven cases showed PrPSc types that are congruent with those seen in sporadic CJD. Five were 129MV with type 3 PrPSc and two were 129MM with type 2 PrPSc using the London classification (Hill et al., 2003).

**Figure 6** PrP immunoblot of patient brain. Proteinase K (PK)-digested 10 % (w/v) brain homogenates from patients with sporadic CJD or iatrogenic CJD were analysed by enhanced chemiluminescence using anti-PrP monoclonal antibody 3F4. The provenance of each brain sample is designated above each lane and the type of PrPSc (London classification; Hill et al., 2003) detected in each sample and the PRNP codon 129 genotype of the patient (M, methionine; V, valine) is designated below. Control samples of human PrPSc types 2 and 3 from patients with sporadic CJD (lanes 1 and 2) are compared with the PrPSc molecular strain types seen in Patients 4, 10, 11 and 12 with iatrogenic CJD (lanes 3–6).

**Discussion**

This study, covering the period from 2000–14, shows that iatrogenic CJD due to cadaver-sourced pituitary growth hormone, a treatment that was discontinued in 1985 in the UK, continues to occur in the UK at a frequency of 0–6 cases per annum. Incubation periods are now extraordinarily long, with estimates ranging from 18–40 years, the uncertainty based on clinical onsets and lengths of treatment with potentially infected batches. In this paper we have reviewed the clinical features, progression, imaging abnormalities, prion protein genotype, PrPSc type by western blot and preliminary neuropathology data.

**Clinical features and imaging correlations**

Typically patients with growth hormone induced iatrogenic CJD present with gait ataxia, cerebellar dysarthria and lower limb pain with cognitive function much less affected. Later sleep disturbance, cognitive decline and pyramidal signs with weakness in the lower limbs develops. Ultimately, the patient enters an akinetic mute state typical of all types of CJD with death at 4–32 (mean 16.0) months after the initial presentation. There are several points of special interest in this clinical presentation.

First, gait ataxia, with much less limb ataxia, especially in the upper limbs, is typical of superior cerebellar vermis damage. The MRI and pathological examination of the cerebellum demonstrated extensive damage of the vermis with lesser involvement of the cerebellar hemispheres consistent with these clinical findings. Interestingly, in addition to gait ataxia, two of the patients presented with abnormal visual input imbalance characterized by unsteadiness induced by moving stimuli (waves when paddling and looking at moving water) suggesting abnormal visual vestibular interaction partly dependent on cerebellar pathways.

Second, pain in the lower limbs was of a quality suggesting abnormal spino-thalamic function and MRI confirmed a diffuse abnormality of the thalamus which, although maximal medially, did extend laterally to the ventro-posterior components, part of the pain and thermal pathways. Of note is the similarity of this pain to that experienced by many patients...
with variant CJD and kuru, which are also acquired prion diseases associated with peripheral infection with prions where thalamic involvement, especially posteriorly, occurs. However, a contribution of the more peripheral components of the spino-thalamic system cannot be excluded.

Third, later in the course of the disease cognitive dysfunction with prominent memory decline emerged. The extensive involvement of many areas of the deep nuclei, thalamus and cortex could explain this, but of note is the prominent involvement of the hippocampus on MRI and pathologically. This, together with the thalamic changes, is an adequate explanation of the amnesia. Thalamic involvement would also explain the sleep disturbance similar to that occurring in fatal familial insomnia (FFI) and frequently seen in sporadic CJD.

Finally, weakness in the lower limbs due to pyramidal involvement correlated with cortical ribboning involving the dorsal and medial motor strip demonstrated on MRI in most patients. Such involvement of the motor cortex is rare in other forms of prion disease. More pronounced involvement of the motor cortex and parietal lobe when compared with the anterior frontal or occipital cortex was also confirmed morphologically with extensive micro-vacuolar change and intense deposition of synaptic abnormal prion protein.

Neuropathology
A detailed quantitative analysis of the pathology in the nine autopsied cases will be the subject of a separate communication. However, a striking feature was the presence of PrP plaques in all but one case. This latter non-plaque case suggests that the different pathological appearances might be caused by different prion strains. Similarly, iatrogenic CJD due to dural grafts, a condition particularly prevalent in Japan, may show two distinct pathologies, plaque-type and non-plaque-type, which have been linked with differing transmission properties in transgenic mice (Kobayashi et al., 2014). It is possible that future studies using material from the original batches of pituitary-derived growth hormone and transgenic mice could clarify the pathogenesis of the disease in our cases and comparison with the literature on dural graft iatrogenic CJD.

PRNP codon 129 polymorphism determines incubation time
A remarkable feature of iatrogenic CJD cases in the UK is the distribution of the codon 129 polymorphism. An excess of VV homozygotes was reported in early UK cases and then thought to be a marker of susceptibility (Collinge et al., 1991). MV heterozygosity was reported to confer relative resistance to sporadic CJD (Palmer et al., 1991). In the initial study of 27 UK patients 52% were homozygous for valine and only 4% (one case) was homozygous for methionine; the remainder were heterozygotes. This contrasts with the French experience in 77 patients where 48% were homozygous for methionine and 22% homozygous for valine (Brandel et al., 2003). Similar proportions were also found in the few cases reported from the USA (Brown et al., 2012). Furthermore in sporadic CJD, kuru and variant CJD, methionine homozygosity predominates. This is thought to result from the relative resistance afforded by heterozygosity at polymorphic PrP residue 129 and from conformational selection whereby different prion strains are preferentially propagated by prion proteins of different primary sequence (Collinge, 1999; Collinge and Clarke, 2007). For example the bovine spongiform encephalopathy-related prion strain causing variant CJD appears only to be compatible with methionine 129 human PrP (Wadsworth et al., 2004).

One of the new findings of our study of UK patients is that there has been a change in the distribution of the 129 polymorphism in the past 12 years. The VV genotype has now greatly decreased and MM genotype increased while the frequency of heterozygotes has remained relatively constant. There are similarities and differences from the studies of kuru (Cervenakova et al., 1999; Collinge et al., 2006; Mead et al., 2009b). In kuru both homozygous genotypes predominated in young cases with presumably a shorter incubation time, but in later and older cases heterozygotes occurred more frequently.

A possible explanation for these superficially contradictory distributions of codon 129 in different outbreaks could be the compatibility of host genotype and strain of the infecting prion, in keeping with the conformational selection model of prion transmission (Collinge, 1999; Collinge and Clarke, 2007). This model suggests that transmission is facilitated, with shorter incubation times, if the host prion protein can readily adopt the preferred conformation associated with the strain of the infecting prion. In the case of the UK, it is likely that one particular preparation (Hartree modified Wilhelmi preparation) was responsible for the outbreak as all patients to date received this preparation.

Swerdlov et al. (2003) estimated that ~400 000 pituitary hormones were harvested for growth hormone production, but even this may be an underestimate. Attempts were made to exclude patients who had neurological diseases but protocols for harvesting were not strictly monitored (Swerdlow et al., 2003). It is likely that some of these pituitary glands were sourced from cases with CJD; in the 1970s, the time when the majority of pituitary sourced growth hormone was obtained, it is estimated from UK mortality data that 1 in 7000 deaths would have been due to sporadic CJD. Two-thirds of cases of sporadic CJD worldwide are of the 129MM genotype and therefore it is not surprising that most cases of iatrogenic CJD outside the UK were of this genotype. However, in the present series, the pattern was different with the early patients, which were predominantly of the VV genotype. As only ~24% of cases of sporadic CJD are 129VV the question arises as to why the UK patients differ from the rest of the world in this regard.

One possibility is that the screening of donors was more successful than suggested above and by chance only one or
two cases of sporadic CJD were the source of the growth hormone and these had an atypical phenotype that occurred more frequently in VV or MV cases. Alternatively many donors were included, however, a single VV or MV case had a particularly high titre of infective material in the pituitary. The profound influence of codon 129 in controlling human prion transmission barriers has been extensively modelled in transgenic mice expressing human PrP on a mouse PrP null background (Wadsworth et al., 2010). These studies have shown that transmission barriers to infection with classical CJD prions are asymmetric, dependent upon the codon 129 genotype of the prion source and the recipient (Wadsworth et al., 2010). Whereas, transgenic mice expressing human PrP 129 valine are highly susceptible to classical CJD prions regardless of the PrPSc type or codon 129 genotype of the inoculum (Collinge et al., 1995, 1996; Telling et al., 1995; Hill et al., 1997; Korth et al., 2003; Kobayashi et al., 2007; Wadsworth et al., 2008b), the absence of a transmission barrier to classical CJD prions is not uniformly observed in mice expressing human PrP 129 methionine in the absence of mouse PrP. Here, mismatch at residue 129 between the inoculum and host can significantly affect transmission. Thus while there appears to be no barrier to transmission of classical CJD prions from codon 129 methionine homozygous patients (Asante et al., 2002; Korth et al., 2003; Beringue et al., 2008; Kong et al., 2008), transmission of classical CJD prions from valine homozygous patients is often associated with more prolonged and variable incubation periods and reduced attack rates (Asante et al., 2002; Korth et al., 2003; Kobayashi et al., 2007, 2015). In the case of classical CJD prions from codon 129 heterozygous patients the transmission efficiency in transgenic mice expressing human PrP 129 methionine varies dependent upon PrPSc type and whether prions are propagated on human PrP with methionine or valine at residue 129 (Asante et al., 2002; Korth et al., 2003; Bishop et al., 2010; Kobayashi et al., 2015). These data provide an experimental background with which to interpret the temporal distribution of codon 129 genotypes within the cohort of iatrogenic CJD patients in the UK and suggest that the infecting prion contamination of growth hormone was from a VV or MV individual.

Finally, the significance of the polymorphism in RRP9 found on exome sequencing is unclear. The sample is small and the finding requires replication in an independent cohort. Any association could be related to either the initial condition for which the patients were treated or iatrogenic disease or both.

Mismatch of incubation period and rapidity of symptomatic progression

A further seemingly paradoxical observation is that those recent clinical cases with particularly long incubation times have had the shortest clinical durations once symptomatic. Rates of clinical decline in our series of iatrogenic CJD have been most in keeping with observations of sporadic CJD with the MM genotype being more rapid than MV (Pocchiari et al., 2004). A plausible explanation for these observations is that the generation of a host prion strain compatible with host genotype occurs in the periphery during the prolonged incubation time; however, following CNS invasion the rapidity of disease progression [which is thought to be determined by the rates of production of toxic PrP species (designated PrP\textsuperscript{L} for lethal); Hill et al., 2000; Collinge and Clarke, 2007; Sandberg et al., 2011, 2014] is more rapid in MM versus MV individuals owing to higher levels of homotypic substrate PrP available for conversion.

Conclusion

This study is the first to clearly define the clinical, imaging and neuro-pathological characteristics of patients with iatrogenic CJD due to cadaver-sourced growth hormone. It demonstrates that cases continue to occur at a low but steady rate in the UK and that the incubation period can be up to four decades. We have shown that all three common genotypes at PRNP are susceptible albeit with markedly different incubation periods, a phenomenon also seen in kuru. Whether similar susceptibility, with differing mean incubation times, will be seen in the UK related to the transmission of bovine spongiform encephalopathy prions remains to be seen. Further, we have demonstrated a dissociation between the incubation period and the rapidity of decline once a person develops symptoms.

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Collinge J. Variant Creutzfeldt-Jakob disease. Lancet 1999; 354:
Cervenakova L, Goldfarb L, Garruto R, Lee HS, Gajdusek CD, Brown
Brown P, Brandel JP, Sato T, Nakamura Y, Mackenzie J, Will RG,
Unaltered susceptibility to BSE in transgenic mice expressing human
prion protein. Nature 1995; 378: 779–83.
Collinge J, Sidle KC, Meads J, Ironside J, Hill AF. Molecular analysis
of prion strain variation and the aetiology of ‘new variant’ CJD. Nature
1996; 383: 685–90.
Collinge J, Whitfield J, McKintosh E, Beck J, Mead S, Thomas DJ
et al. Kuru in the 21st century—an acquired human prion disease
with very long incubation periods. Lancet 2006; 367: 2068–74.
Hill AF, Desbruslais M, Joiner S, Sidle KC, Bell JE, Budka H, et al.
Molecular classification of sporadic Creutzfeldt-Jakob disease. Brain
2003; 126: 1333–46.
Jucker M, Walker LC. Self-propagation of pathogenic protein aggre-
gates in neurodegenerative diseases. Nature 2013; 501: 45–51.
Kobayashi A, Asano M, Mohri S, Kitamoto T. Cross-sequence trans-
mission of sporadic Creutzfeldt-Jakob disease creates a new prion
strain. J Biol Chem 2007; 282: 30022–28.
Kobayashi A, Matsuura Y, Mohri S, Kitamoto T. Distinct origins of
dura mater graft-associated Creutzfeldt-Jakob disease: past and
future problems. Acta Neuropathol Commun 2014; 2: 32.
Kobayashi A, Teruya K, Matsuura Y, Shirai T, Nakamura Y, Yamada
M, et al. The influence of PRNP polymorphisms on human prion
disease susceptibility: an update. Acta Neuropathol 2015; 130:
159–70.
Kong Q, Zheng M, Casalone C, Qing L, Huang S, Chakraborty B,
et al. Evaluation of the human transmission risk of an atypical
bovine spongiform encephalopathy prion strain. J Virol 2008; 82:
3697–701.
Korth C, Kaneko K, Groth D, Heye N, Telling G, Mastronardi, et al.
Abbreviated incubation times for human prions in mice expressing a
chimeric mouse-human prion protein transgene. Proc Natl Acad Sci
USA 2003; 100: 4784–89.
Mead S, Poulter M, Beck J, Webb T, Campbell T, Linehan J, et al.
Inherited prion disease with six octapeptide repeat insertion mutation–
molecular analysis of phenotypic heterogeneity. Brain 2006;
129: 2297–317.
Mead S, Poulter M, Uphill J, Beck J, Whitfield J, Webb TE, et al.
Genetic risk factors for variant Creutzfeldt-Jakob disease: a
genome-wide association study. Lancet Neurol 2009a; 8: 57–66.
Mead S, Whitfield J, Poulter M, Shah P, Uphill J, Campbell T, et al.
A novel protective prion protein variant that colocalizes with kuru
exposure. N Engl J Med 2009b; 361: 2056–65.
Palmer MS, Dryden AJ, Hughes JT, Collinge J. Homozygous prion
protein genotype predisposes to sporadic Creutzfeldt-Jakob disease.
Nature 1991; 352: 340–42.
Parchi P, Castellani R, Capellari S, Ghetti B, Young K, Chen SG, et al.
Molecular basis of phenotypic variability in sporadic Creutzfeldt-
Jakob disease. Ann Neurol 1996; 39: 767–78.
Parchi P, Giese A, Capellari S, Brown P, Schulz-Schaeffer W, Windl O,
et al. Classification of sporadic Creutzfeldt-Jakob disease based on
molecular and phenotypic analysis of 300 subjects. Ann Neurol
1999; 46: 224–33.

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Conflict of interest
J.C. is a Director and J.C. and J.D.F.W. are shareholders of
D-Gen Limited which owns one of the antibodies used in
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Supplementary material
Supplementary material is available at Brain online.

References
Asante EA, Linehan J, Desbruslais M, Joiner S, Gowland I, Wood A,
et al. BSE prions propagate as either variant CJD-like or sporadic
CJD-like prion strains in transgenic mice expressing human prion
protein. EMBO J 2002; 21: 6358–66.
Beringue V, Le Dur A, Tixador P, Reine F, Lepourry L, Perret-Liaudet
et al. Prominent and persistent extraneural infection in human
PrP transgenic mice infected with variant CJD. PLoS One 2008; 3:
e1419.
Bishop MT, Will RG, Manson JC. Defining sporadic Creutzfeldt-
Jakob disease strains and their transmission properties. Proc Natl
Acad Sci USA 2010; 107: 12005–10.
Brandel JP, Preece M, Brown P, Croes E, Laplanche JL, Agid Y, et al.
Distribution of codon 129 genotype in human growth hormone-
treated CJD patients in France and the UK. Lancet 2003; 362:
128–30.
Brandner S, Korth C, Kaneko K, Groth D, Heye N, Telling G, Mastrianni, et al.
Unaltered susceptibility to BSE in transgenic mice expressing human
prion protein. Nature 1995; 378: 779–83.
Brown P, Brandel JP, Sato T, Nakamura Y, Mackenzie J, Will RG,
et al. Iatrogenic Creutzfeldt-Jakob disease, final assessment. Emerg
Infect Dis 2012; 18: 901–7.
Cervenakova L, Goldfarb L, Garruto R, Lee HS, Gajdusek CD, Brown
P. Phenotype-genotype studies in kuru: Implications for new variant
Creutzfeldt-Jakob disease. Proc Natl Acad Sci USA 1999; 95:
13239–41.
Collinge J. Variant Creutzfeldt-Jakob disease. Lancet 1999; 354:
317–23.
Collinge J. Prion diseases of humans and animals: their causes and
molecular basis. Annu Rev Neurosci 2001; 24: 519–50.
Collinge J, Gorham M, Hudson F, Kennedy A, Keogh G, Pal S, et al.
Safety and efficacy of quinacrine in human prion disease (PRION-1
study): a patient-preference trial. Lancet Neurol 2009; 2009:
334–44.
Collinge J, Palmer MS, Dryden AJ. Genetic predisposition to iatro-
genic Creutzfeldt-Jakob disease. Lancet 1991; 337: 1441–42.
Collinge J, Clarke A. A general model of prion strains and their patho-
genicity. Science 2007; 318: 930–36.
Collinge J, Palmer MS, Sidle KCL, Hill AF, Gowland I, Meads J et al.
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3398 | BRAIN 2015: 138; 3386–3399 P. Rudge et al.
Parchi P, Strammiello R, Giese A, Kretzschmar H. Phenotypic variability of sporadic human prion disease and its molecular basis: past, present, and future. Acta Neuropathol 2011; 121: 91–112.

Pocchiari M, Puopolo M, Croes EA, Budka H, Gelpi E, Collins S, et al. Predictors of survival in sporadic Creutzfeldt-Jakob disease and other human transmissible spongiform encephalopathies. Brain 2004; 127: 2348–59.

Powell Jackson J, Weller RO, Kennedy P, Preece MA, Whitcombe EM, Newsom Davis J. Creutzfeldt-Jakob disease after administration of human growth hormone. Lancet 1985; 2: 244–46.

Raben MS. Treatment of a pituitary dwarf with human growth hormone. J Clin Endocrinol Metab 1958; 18: 901–3.

Sandberg MK, Al Doujaily H, Sharps B, Clarke AR, Collinge J. Prion propagation and toxicity in vivo occur in two distinct mechanistic phases. Nature 2011; 470: 540–2.

Sandberg MK, Al Doujaily H, Sharps B, De Oliveira MW, Schmidt C, Richard-Londt A, et al. Prion neuropathology follows the accumulation of alternate prion protein isoforms after infective titre has peaked. Nat Commun 2014; 5: 4347.

Swerdlow AJ, Higgins CD, Adlard P, Jones ME, Preece MA. Creutzfeldt-Jakob disease in United Kingdom patients treated with human pituitary growth hormone. Neurology 2003; 61: 783–91.

Telling GC, Scott M, Mastronardi J, Gabizon R, Torchia M, Cohen FE, et al. Prion propagation in mice expressing human and chimeric PrP transgenes implicates the interaction of cellular PrP with another protein. Cell 1995; 83: 79–90.

Thompson AG, Lowe J, Fox Z, Lukic A, Porter MC, Ford L, et al. The Medical Research Council Prion Disease Rating Scale: a new outcome measure for prion disease therapeutic trials developed and validated using systematic observational studies. Brain 2013; 136: 1116–27.

Wadsworth JD, Asante EA, Collinge J. Contribution of transgenic models to understanding human prion disease. Neuropathol Appl Neurobiol 2010; 36: 576–97.

Wadsworth JD, Asante EA, Desbruslais M, Linehan J, Joiner S, Gowland I, et al. Human prion protein with valine 129 prevents expression of variant CJD phenotype. Science 2004; 306: 1793–96.

Wadsworth JD, Collinge J. Molecular pathology of human prion disease. Acta Neuropathol 2011; 121: 69–77.

Wadsworth JD, Hill AF, Beck JA, Collinge J. Molecular and clinical classification of human prion disease. Br Med Bull 2003; 66: 241–54.

Wadsworth JD, Joiner S, Hill AF, Campbell TA, Desbruslais M, Luthert PJ, et al. Tissue distribution of protease resistant prion protein in variant CJD using a highly sensitive immuno-blotting assay. Lancet 2001; 358: 171–80.

Wadsworth JD, Joiner S, Linehan JM, Asante EA, Brandner S, Collinge J. Review. The origin of the prion agent of kuru: molecular and biological strain typing. Philos Trans R Soc Lond B Biol Sci 2008a; 363: 3747–53.

Wadsworth JD, Joiner S, Linehan JM, Desbruslais M, Fox K, Cooper S, et al. Kuru prions and sporadic Creutzfeldt-Jakob disease prions have equivalent transmission properties in transgenic and wild-type mice. Proc Natl Acad Sci USA 2008b; 105: 3885–90.

Wadsworth JD, Powell C, Beck JA, Joiner S, Linehan JM, Brandner S, et al. Molecular diagnosis of human prion disease. Methods Mol Biol 2008c; 459: 197–227.