Case Report

Hepatic YAP1-TFE3 Rearranged Epithelioid Hemangioendothelioma

Mira M. Lotfalla, Andrew L. Folpe, Karen J. Fritchie, Patricia T. Greipp, Gretchen G. Galliano, Kevin C. Halling, Taofic Mounajjed, Jorge Torres-Mora, and Rondell P. Graham

1Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA
2Anatomic Pathology, Ochsner Health System, New Orleans, LA, USA

Correspondence should be addressed to Rondell P. Graham; graham.rondell@mayo.edu

Received 26 February 2019; Revised 10 May 2019; Accepted 29 May 2019; Published 23 June 2019

Academic Editor: Yoshifumi Nakayama

Copyright © 2019 Mira M. Lotfalla et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Epithelioid hemangioendothelioma (EHE) is an uncommon low-grade malignant vascular tumor that may arise in soft tissue/bone or visceral sites such as the liver and lung. As this tumor exhibits epithelioid morphology, it frequently causes diagnostic confusion with other epithelioid vascular neoplasms as well as carcinomas. While 90% of classic EHE are driven by a WWTR1-CAMTA1 fusion gene, a histologically distinctive subset of EHE has been recently shown to harbor a different fusion gene, YAP1-TFE3. This variant is likely underrecognized given its rarity and only recent description. Notably, EHE frequently involves the liver but only one case of hepatic YAP1-TFE3 rearranged EHE has been reported to date [2]. We present the second case of YAP1-TFE3 rearranged EHE affecting a 65-year-old woman and presenting as multiple liver masses, with characterization of the fusion gene at the transcriptomic and genomic levels. There are several educational points noted from this case. YAP1-TFE3 rearranged EHE shows distinctly vasoformative foci, unlike classic EHE and mimicking angiosarcoma or epithelioid hemangioma. The tumors cells show a histiocytoid appearance with voluminous cytoplasm, similar to other TFE3-rearranged tumors. Finally, in the liver, this tumor may in part mimic focal nodular hyperplasia of the liver which is an underrecognized diagnostic pitfall. This report highlights the key diagnostic and genetic features of this newly recognized variant of hepatic EHE to aid pathologists in appropriately classifying these tumors.

1. Introduction

Epithelioid hemangioendothelioma (EHE) is an uncommon low-grade malignant vascular tumor that may arise in soft tissue/bone or visceral sites such as the liver and lung. As this tumor exhibits epithelioid morphology, it frequently causes diagnostic confusion with other epithelioid vascular neoplasms as well as carcinomas. While 90% of classic EHE are driven by a WWTR1-CAMTA1 fusion gene, a histologically distinctive subset of EHE has been recently shown to harbor a different fusion gene, YAP1-TFE3 [1]. This variant is likely underrecognized given its rarity and only recent description. Notably, EHE frequently involves the liver but only one case of hepatic YAP1-TFE3 rearranged EHE has been reported to date [2]. We present an illustrative second case presenting as liver masses which shows the distinctive morphology and molecular features of this entity as well as the potential diagnostic pitfalls.

2. Case Presentation

A 65-year-old woman with multiple liver masses presented for transplantation following a diagnosis of EHE made on biopsy. Preoperative imaging revealed an enlarged liver with multifocal disease throughout the hepatic parenchyma. The lesions showed peripheral enhancement with central hypooptenuation on arterial phase and appeared isodense to the liver parenchyma on delayed phase imaging. Some of the masses showed calcifications (Figure 1). There was no significant interval change in size in the 10 months from initial diagnosis to transplantation. No chest or pelvic
masses were identified elsewhere during the preoperative workup.

Pathologic examination of the liver explant disclosed multiple tan discrete nodules ranging from 0.5 to 3.0 cm in greatest dimension and involving the bilateral lobes. Low power examination revealed 2 main histologic patterns. A subset of the nodules contained well-formed vascular channels and scattered cords of tumor cells embedded in an abundantly sclerotic hyalinized matrix (Figure 2(a)), while other areas of tumor were characterized by small, irregular central hyalinized scars containing small subtle foci of necrotic cells and a mild bile ductular reaction (Figure 2(b)). On high magnification, the tumor cells in both components exhibited mild cytologic atypia and were characterized by abundant pale cytoplasm, nuclei bearing open chromatin and visible though not large nucleoli. Scattered intracytoplasmic vacuoles were noted in the tumor cells arranged in a linear pattern (Figure 2(c)). Mitotic figures were not observed; however, focal vascular invasion by single tumor cells was noted. There was stromal lymphocytic inflammation, and focal calcification was noted in one of the lesions. The surrounding liver parenchyma had a nodular architecture, reminiscent of focal nodular hyperplasia (Figure 2(b)).

Immunohistochemically, the tumor cells showed an endothelial phenotype with strong expression of CD31 (clone JC70A; 1/350; Dako) and ERG (clone EPR3864; predilute; Ventana) (Figure 2(d)). The tumor nuclei were positive for TFE3 (clone MRQ-37; 1/50; Cell Marque) (Figure 2(a)), but CAMTA1 (polyclonal; 1/300; Novus Biologicals) was negative (Figure 2(e)). Glutamine synthetase (clone GS-6; 1/2000; Millipore) showed a map-like pattern of distribution in the hepatocytes surrounding small irregular areas of scar mimicking focal nodular hyperplasia (Figure 2(f)).

RNA sequencing with fusion gene analysis from formalin-fixed paraffin embedded tissue revealed an in-frame YAP1-TFE3 fusion gene joining exon 1 of YAPI to exon 4 of TFE3 and a reciprocal TFE3-YAPI joining exon 3 of TFE3 to exon 2 of YAPI, similar to the findings of Antonescu and colleagues [1], using a published lab-developed protocol [3]. No other fusion genes, including WWTR1-CAMTA1, were identified. Fluorescence in situ hybridization (FISH) was performed with lab-developed split apart probes for the TFE3 locus (bacterial artificial chromosome clones RP11-416B14, CTD-2311N12, CTD-3009K20, and CTD-2173N15; all from Abbott Molecular, IL) and dual fusion probes for YAPI-TFE3 loci (bacterial artificial chromosome clones RP11-635D24, RP11-451F15, RP11-90M3, and RP11-750P5 along with the previously mentioned clones for TFE3; all from Abbott Molecular, IL) using standard techniques as previously reported by our group [4]. Interphase FISH revealed rearrangement of the TFE3 locus in all tumor cells and YAPI-TFE3 fusion at the genomic level (Figure 3(b)). An image of intact CAMTA1 split apart FISH performed using the same method and probes to the CAMTA1 locus is also shown (Figure 3(c)). Based on the combination of morphologic, immunophenotypic, and genetic findings, this case was classified as a YAPI-TFE3 rearranged EHE.

3. Discussion

EHE, a malignant vascular neoplasm first described by Enzinger and Weiss in 1982 [5], may arise at a variety of anatomic locations including bone, soft tissue, and viscera. Although this entity is well-recognized, EHE still represents a diagnostic challenge in routine practice due to its rarity and morphologic overlap with other epithelioid tumors, including other benign and malignant epithelioid vascular neoplasms and carcinomas, especially when these tumors present at visceral sites such as the liver. The vast majority (90%) of EHE harbor the WWTR1-CAMTA1 fusion event [6, 7], which can be detected with molecular genetic methods or with the CAMTA1 immunostain, a reliable surrogate [8].

Recently, Antonescu and colleagues described a distinctive vascular neoplasm showing some features of EHE, but characterized by an alternate YAPI-TFE3 fusion [1]. It is not yet entirely certain whether this unusual tumor represents a variant of EHE or an altogether unique tumor [1]. Although both tumors share epithelioid morphology and a corded growth pattern, tumors harboring the YAPI-TFE3 fusion differ from conventional EHE inasmuch as they display overt vasoformation, consist of cells with voluminous cytoplasm, and lack the distinctive myxochondroid matrix of the latter tumor [1]. However, the natural history of conventional and YAPI-TFE3 EHE seems to be similar, with significant potential for late distant metastasis [1].

Antonescu et al’s seminal description of YAPI-TFE3 EHE did not identify any tumors primary to the liver [1]. To the best of our knowledge, the only bona fide report of this rare tumor occurring in the liver is that of Kuo et al., who reported a morphologically typical, TFE3-rearranged EHE occurring
in a 39-year-old woman [2]. Although another primary hepatic vascular tumor has been reported by Lee and coworkers as representing EHE showing “concomitant YAPI-TFE3 and WWTR1-CAMTA1 fusions,” the morphologic and molecular genetic features of this lesion are not fully convincing, in our opinion [9].

Our case report highlights several diagnostic challenges of this variant of EHE. YAPI-TFE3 rearranged EHE harbors areas with well-formed blood vessels, potentially mimicking a hemangioma, an entity much more common in the liver. The biphasic appearance, however, with a second component of tumor cells arranged in cords and containing scattered intracytoplasmic lumina should trigger pathologists to consider this variant of EHE. The epithelioid morphology of the neoplastic population may also raise the possibility of other epithelioid vascular tumors such as epithelioid hemangioma,
epithelioid angiosarcoma, and the conventional form of EHE with WWTR1-CAMTA1 fusion. Although the tumor cells of epithelioid hemangioendothelioma are plump and hobnailed, they lack significant atypia. Furthermore, these tumors are uniformly vasoformative and often containing numerous eosinophils. The latter feature was not seen in this case. Additionally, epithelioid hemangioendothelioma is characterized by frequent FOSB gene rearrangements rather than TFE3 fusion genes [10]. Epithelioid angiosarcoma exhibits more significant cytologic atypia and mitotic activity, including atypical mitoses. As previously mentioned, the conventional form of EHE lacks well-formed vascular channels and produces distinctive myxochondroid matrix. Finally, a combination of immunohistochemical studies including CAMTA1 and TFE3 and molecular studies assessing for WWTR1-CAMTA1 and YAPI-TFE3 fusions aids in the correct diagnosis.

The epithelioid morphology of EHE may also mimic involvement by primary or metastatic carcinoma. Recognizing vasoformative areas is a helpful clue to endothelial lineage, but these areas may not always be present on biopsy material. The degree of atypia in YAPI-TFE3 rearranged EHE is usually relatively mild, and pathologists should consider performing a battery of immunostains for endothelial markers (CD31, CD34, FLI-1, and ERG) on any epithelioid neoplasm lacking keratin expression or with a uniform cytologic appearance.

Finally, low magnification examination of the explanted specimen in our patient showed some masses with a striking resemblance to focal nodular hyperplasia (FNH). This low power architecture may be deceptive, potentially resulting in a failure to recognize the underlying vascular neoplasm as the tumor cells in our case were sparsely distributed within small irregular areas of fibrous tissue and in nearby sinusoids and vessels. It has been suggested that FNH and peritumoral hyperplasia, as seen in this case, are reactive processes unified by blood flow abnormalities, and that marked angiogenesis may produce the FNH phenotype, an appealing hypothesis [11]. In this case, the absence of an abnormal artery and the presence of atypical cells percolating through the small central area of fibrous tissue and the adjacent liver parenchyma on higher magnification aided distinction from FNH. It should also be emphasized that “map-like” glutamine synthetase expression, a potential pitfall illustrated by this case, should always be interpreted in the context of other clinical and morphological findings.

YAPI and WWTR1 are orthologues and are believed to function similarly in their respective fusion genes. A review of the biology of these genes is beyond the scope of this report but YAPI-TFE3 and WWTR1-CAMTA1 are believed to be oncogenic drivers. In brief, YAPI and WWTR1 function via TEAD transcription factors to drive expression of proproliferative and antiapoptotic genes [12]. TFE3 is one of the MIT family of transcription factors which bind to specific recognition sequences of DNA to alter gene expression. The available data indicate that, in YAPI-TFE3, the promoter of
YAPI is juxtaposed to the key domains of TFE3 to drive the development of tumors [1]. The specificity of the YAPI-TFE3 and WWTR1-CAMTA1 fusion genes makes them useful biomarkers for diagnosis as shown in this case.

4. Conclusion

In conclusion, we describe a second case of YAPI-TFE3 hepatic EHE with complete characterization of the fusion gene at the transcriptomic and genomic levels. This case also exhibited findings reminiscent of focal nodular hyperplasia, which is an underrecognized potential pitfall. Pathologists should be aware of the morphologic features of this rare and recently described variant to avoid misclassification as other vascular tumors, carcinoma and focal nodular hyperplasia. Further clinicopathologic studies are necessary to determine the behavior of this variant, especially those arising in the liver.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

[1] C. R. Antonescu, F. Le Loarer, J.-M. Mosquera et al., “Novel YAPI-TFE3 fusion defines a distinct subset of epithelioid hemangiendothelioma,” Genes, Chromosomes and Cancer, vol. 52, no. 8, pp. 775–784, 2013.

[2] F.-Y. Kuo, H.-Y. Huang, C.-L. Chen et al., “TFE3-rearranged hepatic epithelioid hemangiendothelioma—a case report with immunohistochemical and molecular study,” APMIS-Acta Pathologica, Microbiologica et Immunologica Scandinavica, vol. 125, no. 9, pp. 849–853, 2017.

[3] J. L. Winters, J. I. Davila, A. M. McDonald et al., “Development and verification of an RNA sequencing (RNA-Seq) assay for the detection of gene fusions in tumors,” The Journal of Molecular Diagnostics, vol. 20, no. 4, pp. 495–511, 2018.

[4] R. P. Graham, A. A. Nair, J. I. Davila et al., “Gastroblastoma harbors a recurrent somatic MALAT1-GLI1 fusion gene,” Modern Pathology, vol. 30, no. 10, pp. 1443–1452, 2017.

[5] S. W. Weiss and F. M. Enzinger, “Epitheloid hemangiendothelioma: a vascular tumor often mistaken for a carcinoma,” Cancer, vol. 50, no. 5, pp. 970–981, 1982.

[6] C. Errani, L. Zhang, Y. S. Sung et al., “A novel WWTRI-CAMTA1 gene fusion is a consistent abnormality in epithelioid hemangiendothelioma of different anatomic sites,” Genes, Chromosomes and Cancer, vol. 50, no. 8, pp. 644–653, 2011.

[7] M. R. Tanas, A. Sboner, A. M. Oliveira et al., “Identification of a disease-defining gene fusion in epithelioid hemangiendothelioma,” Science Translational Medicine, vol. 3, no. 98, p. 98ra82, 2011.

[8] L. A. Doyle, C. D. M. Fletcher, and J. L. Hornick, “Nuclear expression of CAMTA1 distinguishes epithelioid hemangiendothelioma from histologic mimics,” The American Journal of Surgical Pathology, vol. 40, no. 1, pp. 94–102, 2016.

[9] S. J. Lee, W. I. Yang, W.-S. Chung et al., “Epithelioid hemangiendotheliomas with TFE3 gene translocations are compossible with CAMTA1 gene rearrangements,” Oncotarget, vol. 7, no. 7, pp. 7480–7488, 2016.

[10] S. C. Huang, L. Zhang, Y. S. Sung et al., “Frequent FOS gene rearrangements in epithelioid hemangioendothelioma: a molecular study of 58 cases with morphologic reappraisal,” The American Journal of Surgical Pathology, vol. 39, no. 10, pp. 1313–1321, 2015.

[11] I. R. Wanless, “Epithelioid hemangioendothelioma, multiple focal nodular hyperplasias, and cavernous hemangiomas of the liver,” Archives of Pathology & Laboratory Medicine, vol. 124, no. 8, pp. 1105–1107, 2000.

[12] B. Zhao, X. Ye, J. Yu et al., “TEAD mediates YAP-dependent gene induction and growth control,” Genes & Development, vol. 22, no. 14, pp. 1962–1971, 2008.
