Structure–function correlates of vision loss in neuromyelitis optica spectrum disorders

Norman K. Gigengack1,2, Frederike C. Oertel1,2,3, Seyedamirhosein Motamedi1,2, Charlotte Bereuter1,2, Ankelien Duchow1,2, Rebekka Rust1,2, Judith Bellmann-Strobl1,2, Klemens Ruprecht4, Tanja Schmitz-Hübsch1,2, Friedemann Paul1,2,4, Alexander U. Brandt1,5,7 & Hanna G. Zimmermann1,2,6,7

Optic neuritis (ON) in neuromyelitis optica spectrum disorders (NMOSD) regularly leads to more profound vision loss compared to multiple sclerosis (MS) and myelin-oligodendrocyte-glycopolypeptide-antibody associated disease (MOGAD). Here we investigate ON-related vision loss in NMOSD compared to MS and MOGAD in order to identify neuroaxonal and retinal contributors to visual dysfunction. In this retrospective study we included patients with aquaporin-4-antibody seropositive NMOSD (n = 28), MOGAD (n = 14), MS (n = 29) and controls (n = 14). We assessed optic nerve damage and fovea morphometry by optical coherence tomography. Visual function was assessed as high (HCVA) and low contrast visual acuity (LCVA), and visual fields’ mean deviation (MD). In all diseases, lower visual function was associated with peripapillary retinal nerve fiber layer (pRNFL) and ganglion cell and inner plexiform layer (GCIP) thinning following a broken stick model, with pRNFL and GCIP cutoff point at ca. 60 µm. HCVA loss per µm pRNFL and GCIP thinning was stronger in NMOSD compared with MOGAD. Foveal inner rim volume contributed to MD and LCVA in NMOSD eyes, only. Together these data supports that visual dysfunction in NMOSD is associated with neuroaxonal damage beyond the effect seen in MS and MOGAD. A primary retinopathy, respectively Müller cell pathology, may contribute to this effect.

Optic neuritis (ON) is a frequent manifestation of neuroinflammatory diseases such as neuromyelitis optica spectrum disorders (NMOSD), myelin oligodendrocyte glycoprotein (MOG)-IgG associated diseases (MOGAD) and multiple sclerosis (MS)1–3. ON causes neuroaxonal damage in the optic nerves and retina, which is associated with visual function loss4. ON in NMOSD regularly leads to more severe visual dysfunction than ON in MOGAD and MS5. This worse outcome can only in part be explained by more severe neuroaxonal damage assessed by macular ganglion cell and inner plexiform layer (GCIP) thinning6.

Pathogenic aquaporin-4 antibodies (AQP4-IgG) are specific for NMOSD and can be detected in the majority of patients6–8. AQP4 is an astrocytic water channel and expressed in retinal Müller cells9 with the highest concentration around the fovea centralis10. Foveal thinning11,12 and foveal shape changes13 have been described in AQP4-IgG seropositive NMOSD. Additionally, Müller cell dysfunction has been demonstrated through a decrease of specific b-wave amplitude in electoretinogram (ERG) tests of AQP4-IgG seropositive NMOSD patients16.

However, it is unclear if AQP4-IgG-driven loss and/or dysfunction of Müller cells in the fovea may contribute to visual impairment in AQP4-IgG seropositive NMOSD associated ON. The aim of this study was to investigate the relationship between visual function, neuroaxonal damage, and foveal structure in AQP4-IgG seropositive NMOSD compared with MOGAD and MS.

1Experimental and Clinical Research Center, A Cooperation Between the Max Delbrück Center for Molecule Medicine and Charité - Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Lindenberger Weg 80, 13125 Berlin, Germany. 2NeuroCure Clinical Research Center, Charité - Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany. 3Department of Neurology, University of California San Francisco, San Francisco, CA, USA. 4Department of Neurology, Charité - Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin and Humboldt-Universität zu Berlin, Berlin, Germany. 5Department of Neurology, University of California, Irvine, CA, USA. 6Einstein Center Digital Future, Berlin, Germany. 7These authors contributed equally: Alexander U. Brandt, Hanna G. Zimmermann. *email: hanna.zimmermann@charite.de
Methods

Patients and controls. For this cross-sectional monocentric study, we retrospectively analyzed data from ongoing cohort studies of patients with autoimmune neuroinflammatory diseases. Inclusion criteria were AQP4-IgG or MOG-IgG seropositivity in at least one assay based on cell based assays\(^3,4\) (CBA) (AQP4-IgG: CBA, Euroimmun, Lübeck, Germany; MOG-IgG: by established cell-based assays using the laboratory’s cut-offs (MOG IFT, EUROIMMUN, Laboratöry Stöcker, Germany; Molecular Neuroimmunology Group, University Heidelberg, Heidelberg, Germany)\(^5\), diagnosis of relapsing–remitting MS, or healthy controls. Exclusion criteria for all subjects were age below 18 or above 70, diagnosis of other relevant ocular diseases such as macular holes, amblyopia, glaucoma or hyperopic/myopic eyes of more than ±5 dpt, and a timeframe of less than 3 months since the last ON episode. We included 28 AQP4-IgG seropositive NMOSD patients, 14 MOGAD patients, 29 MS patients, and 14 HCs. All AQP4-IgG seropositive and MS patients fulfilled the diagnostic criteria for NMOSD or MS, respectively, at the time of inclusion\(^6\). Of the 14 MOGAD patients, 5 fulfilled the clinical criteria for AQP4-IgG seronegative NMOSD\(^7\). In each case both eyes were included except for two eyes of the NMOSD and MOGAD group each, which were excluded for unrelated ocular disease. MS patients were not systematically tested for presence of AQP4- or MOG-IgG. All patients underwent scoring with the expanded disability status scale (EDSS)\(^8\).

Ethics. NMOSD/MOGAD and MS/HC cohort studies were approved by the ethics committee of Charité – Universitätsmedizin Berlin (EA1/041/14 and EA1/163/12) and conducted according to the Declaration of Helsinki in its currently applicable version. All participants gave written informed consent.

Optical coherence tomography. All patients and controls underwent optical coherence tomography (OCT) using Spectralis OCT (Heidelberg Engineering, Heidelberg, Germany) with automatic positioning system. Thickness of the peripapillary RNFL (pRNFL) was measured in 3.5 mm ring scans around the optic nerve head as automatically positioned by the device. Thickness of the combined ganglion cell and inner plexiform layer (GCIP) was acquired from the volume of a macular cylinder centered on the fovea within a 6 mm diameter. In the case of peripapillary ring scans, automated pRNFL segmentation was carried out by the device-internal software (Heidelberg Eye Explorer Version 6.3). Automated segmentation of macular volume scans was performed with the SAMIRIX pipeline\(^9\), based on OCTLayerSegmentation\(^20\), part of AURA Tools on NITRC (https://www.nitrc.org/projects/aura_tools/). Scan quality and automated segmentation was checked\(^21\) and corrected by two trained raters (NG, FCO). All raters were masked for the patients’ clinical and visual function characteristics. Furthermore, we used fovea morphometry previously described in detail to model and extract foveal shape parameters from the corrected macular volume scans\(^22\). We analyzed the inner rim volume, which is calculated in a 0.5 mm radius around the foveal center between inner limiting membrane (ILM) and Bruch’s Membrane (BM). Inner rim volume reflects volume of the outer retina including photoreceptors at the fovea and demonstrated strong differentiation between AQP4-IgG seropositive NMOSD and MS in a previous study\(^13\). OCT methods are reported in line with the APOSTEL recommendations\(^23\).

Visual function. All patients and controls underwent monocular vision assessment for both eyes. Best corrected visual acuity was tested with retro-illuminated Early Treatment Diabetic Retinopathy Study (ETDRS) charts at a distance of 4 m for high contrast visual acuity (HCVA), and Sloan 2.5% contrast charts at a distance of 2 m for low contrast visual acuity (LCVA) (Precision Vision, La Salle, IL). For HCVA, if a patient was not able to identify any letters at 4 m, the distance was reduced stepwise to 0.5 m; for LCVA, the distance of 2 m was not changed. Visual acuity was measured as decimal acuity and then converted to the logarithm of the Minimum Angle of Resolution (logMAR). The worst possible HCVA was 1.9 logMAR. We excluded eyes that could not complete acuity testing due to vision poorer than that and did not assign a logMAR-value for finger counting, hand movement, and light perception. Visual field testing was performed under best correction in a 30-2 field with a Heidelberg Edge Perimeter (Heidelberg Engineering, Heidelberg, Germany) in a darkened room, using the SAP-III 30-2 ASTA protocol. We analyzed visual fields using the mean deviation (MD).

Statistical methods. Statistical analysis was performed with R version 3.5.3\(^{24}\) using the packages ggplot\(^{25}\), lme4\(^{26}\) and MuMIn\(^{27}\). Significance was established at p < 0.05. Group comparisons were done with Fisher’s Exact Test for categorical variables and with Mann-Whitney-U-tests in each group pair for continuous parameters.

We included eyes with and without history of ON into further analysis. The association between visual function and retinal layers was investigated with linear mixed models, accounting for within patient inter-eye-correlations as a random intercept. Non-standardized effect size is given as Beta ± standard error. To model the previously described steep drop in visual function below a certain threshold of neuroaxonal damage\(^28,29\), we used linear spline model\(^30\). In previous studies the threshold was identified to be at an approximate pRNFL thickness of 50–75 µm\(^{29,30–33}\). After visual evaluation of scatterplots of our data we determined that a cutoff at 60 µm for both pRNFL and GCIP thickness would be most appropriate to reflect the linear spline model. We then calculated the linear spline models with the knot at that cutoff for each patient group. In a sensitivity analysis, changing knot placement to 75 µm pRNFL thickness did not affect significance of results and only resulted in minor changes of effect size (data not shown). Furthermore, as there were only three eyes of MS-patients with a pRNFL thickness below 60 µm, we did not include the MS group in those comparisons. Differences in the relationship between visual function and retinal structure between all patient groups were examined using an interaction effect for diagnosis in the mixed models. Likewise, we investigated whether there was a relationship between visual function and foveal inner rim volume, as well as between inner rim volume and pRNFL thickness. As foveal thickness was shown to be lower in women\(^34\), and there was a mismatch of women and men between our AQP4-IgG
We analyzed 54 eyes of 28 AQP4-IgG seropositive NMOSD patients, 27 eyes of 14 MOGAD patients, 58 eyes of 29 MS-patients and 28 eyes of 14 HC (Table 1). Twenty-three eyes of the NMOSD group, 15 eyes of the MOGAD group and 24 eyes of the MS group had prior history of ON. NMOSD patients had a higher proportion of women compared with the MOGAD group and were older than the MS group. NMOSD patients also presented with a higher EDSS and visual functional system score than MS patients. Furthermore, NMOSD and MOGAD patient eyes with history of ON had a higher number of ON compared with MS eyes. There was no difference in time since first and last ON (per eye) for each group.

| Sex Female | ON History (Patients: NON/ON unilateral/ON bilateral) | NMOsD/NaAoG/NaAoS | No. of ON (by eye) | Median [Range] | Eyes | Median [Range] | Visual FS | Median [Range] | EDSS | Median [Range] | Years since first ON (by eye) | Median [Range] | Years since last ON (by eye) | Median [Range] |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| N (%) | Median [20–69] | 46.0 [21–59] | 3.75 [0.00–6.50] | 1.00 [0.00–6.00] | 3.75 [0.00–6.50] | 4.7 [0.3–28.1] | 4.00 [0.00–3.00] | 4.4 [0.3–39.6] | 0.00 [0.00–5.00] | 4.75 [0.00–6.00] | 4.4 [0.3–43.0] | 1.00 [0.00–6.00] | 4.7 [0.3–28.1] |
| 26 (92.9%) | 49.5 [20–69] | 39.4 [25–64] | 2.00 [0.00–4.50] | 0.00 [0.00–3.00] | 10 [71.4%] | 26 (92.9%) | 0.00 [0.00–3.00] | 6.5 [0.3–43.0] | – | 0.3 | 0.48 |
| 7 (50%) | 54 | 26 | 8 | 2 | 1 | 0.3 | 0.02 | – |
| 21 (72.4%) | 29 | 14 | 24 | 2 | 1 | 0.3 | 0.02 | – |
| 10 (71.4%) | 24 | 14 | 24 | 2 | 1 | 0.3 | 0.02 | – |

Table 1. Patient and eye characteristics. AQP4-IgG+ aquaporin-4 antibody seropositive patients, MOG-IgG+ myelin-oligodendrocyte-glycoprotein antibody seropositive patients, MS multiple sclerosis, HC healthy control, SD standard deviation, EDSS expanded disability status scale, FS functional system, ON optic neuritis. P-values from Fischer’s Exact Test (sex ratio) or Mann–Whitney-U test (all other items); statistically significant results are printed in bold.

seropositive and MOGAD cohorts, we additionally analyzed inner rim volume in eyes of only female participants. We did not perform post-hoc correction for multiple testing due to the exploratory nature of the study.

**Results**

**Cohort characteristics.** We analyzed 54 eyes of 28 AQP4-IgG seropositive NMOSD patients, 27 eyes of 14 MOGAD patients, 58 eyes of 29 MS-patients and 28 eyes of 14 HC (Table 1).

**Group comparisons of retinal structural and visual function parameters.** First, we compared visual function and OCT between NMOSD and other patient groups and HC (Table 2) and focusing on ON-eyes (Fig. 1). In summary, HCVA LCV A and MD in AQP4-IgG seropositive NMOSD were worse than in HC. While HCVA was worse in eyes of AQP4-IgG seropositive NMOSD with ON compared with MOGAD and MS, this was not significant for LCVA. MD was worse in ON-eyes of AQP4-IgG seropositive NMOSD compared with HC but not compared to MOGAD and MS patients. pRNFL and GCIP of AQP4-IgG seropositive NMOSD-ON-eyes were thinner compared with HC, but not to MOGAD. pRNFL of NMOSD-ON-eyes was significantly thinner than in MS-ON-eyes. Foveal inner rim volume was lower in ON-eyes of AQP4-IgG seropositive NMOSD compared with MS eyes. There was no difference in time since first and last ON (per eye) for each group.

**Structure–function associations.** We used mixed linear spline models to investigate the impact of changes in metrics of neuro-axonal damage on visual function in the different patient groups (Fig. 2). For a thickness of pRNFL or GCIP above 60 µm there was no significant effect on visual function in either group, except for a very small but formally significant association between pRNFL and LCVA in MS (Beta = −0.004 ± 0.002; p = 0.05). Because of this, Table 3 only summarizes the results of the mixed spline models for a thickness of RNFL or GCIP below 60 µm (see Supplemental Table 1 for other results). Below 60 µm, thinner pRNFL and GCIP were associated with worse HCVA, LCVA and MD in all disease groups, respectively, with the exception of MS eyes with a pRNFL below 60 µm, which were not calculated due to low event number. In order to consider possible differences between patient groups regarding the rate to which structural changes affect visual outcome, we added the underlying diagnosis as an interaction effect in the mixed models. Further loss of pRNFL beyond a thickness of 60 µm caused a stronger impairment of HCVA and visual field MD in NMOSD-eyes compared with MOGAD-eyes, while there was no difference between the groups regarding LCVA.

Accordingly, below 60 µm of GCIP-thickness MOGAD eyes showed a less steep loss of HCVA than NMOSD eyes, with a similar trend for visual fields and no difference regarding loss of LCVA. Interestingly, there was no
Table 2. Structural and visual function parameters. Test statistics from Mann–Whitney-U Test. AQP4-IgG+ aquaporin-4 IgG seropositive patients, MOG-IgG+ myelin-oligodendrocyte-glycoprotein seropositive patients, MS multiple sclerosis, nON eyes without history of optic neuritis, ON eyes with history of optic neuritis, SD standard deviation, HCVA high contrast visual acuity, LCVA low contrast visual acuity, logMAR logarithm of minimum angle of resolution, MD mean deviation, pRNFL peripapillary retinal nerve fiber layer, GCIP combined ganglion cell and inner plexiform layer. Significant values are in bold.

|                    | AQP4-IgG+ | MOG-IgG+ | MS    | HC    | AQP4-IgG+ nON versus MOG-IgG+ nON | AQP4-IgG+ nON versus MS nON | AQP4-IgG+ nON versus HC | AQP4-ON versus MS-ON | AQP4-ON versus HC |
|--------------------|-----------|----------|-------|-------|-----------------------------------|-----------------------------|-------------------------|---------------------|------------------|
| Mean (SD)          | nON       | ON       | nON   | ON    |                                   |                             |                         |                     |                  |
| HCVA [logMAR]      | −0.09 (0.11) | 0.33 (0.59) | −0.11 (0.15) | −0.10 (0.15) | −0.13 (0.11) | −0.004 (0.36) | −0.13 (0.09) | 0.46 | 0.07 | 0.21 | 0.004 | 0.013 | <0.001 |
| LCVA [logMAR]      | 0.29 (0.17) | 0.85 (0.68) | 0.25 (0.19) | 0.65 (0.70) | 0.31 (0.23) | 0.48 (0.34) | 0.26 (0.14) | 0.59 | 0.69 | 0.36 | 0.15 | 0.07 | <0.001 |
| MD [dB]            | −0.95 (2.36) | −9.76 (10.6) | −0.27 (0.85) | −6.10 (5.83) | −1.05 (1.41) | −2.78 (5.63) | −0.30 (1.05) | 0.26 | 0.30 | 0.24 | 0.59 | 0.08 | <0.001 |
| pRNFL [μm]         | 94.9 (11.5) | 59.5 (23.4) | 98.5 (13.0) | 51.7 (13.5) | 91.1 (11.9) | 74.4 (15.6) | 96.4 (10.1) | 0.49 | 0.11 | 0.57 | 0.39 | 0.01 | <0.001 |
| GCIP [μm]          | 65.6 (7.15) | 51.7 (9.95) | 69.2 (4.63) | 50.6 (7.84) | 66.6 (7.44) | 56.0 (7.22) | 68.7 (4.83) | 0.11 | 0.72 | 0.03 | 0.61 | 0.13 | <0.001 |
| Inner rim volume   | 0.090 (0.015) | 0.090 (0.022) | 0.107 (0.024) | 0.096 (0.012) | 0.111 (0.018) | 0.098 (0.016) | 0.096 (0.021) | 0.06 | <0.001 | 0.25 | 0.051 | 0.02 | 0.19 |

Figure 1. Group comparison of selected structural and functional parameters. (a) High contrast visual acuity (HCVA) (b) Low contrast visual acuity (LCVA) (c) mean deviation of visual fields and (d) inner rim volume of the fovea of eyes with history of ON AQP4-IgG: Aquaporin-4 IgG seropositive patients; MOG-IgG: Myelin-Oligodendrocyte-Glycoprotein seropositive patients; MS: Multiple Sclerosis; nON eyes without history of optic neuritis; ON eyes with history of optic neuritis, SD standard deviation, HCVA high contrast visual acuity, LCVA low contrast visual acuity, logMAR logarithm of minimum angle of resolution, MD mean deviation, pRNFL peripapillary retinal nerve fiber layer, GCIP combined ganglion cell and inner plexiform layer. Significant values are in bold.

Mann–Whitney–U Test (ns: *p > 0.05; **: p < 0.05; ***: p < 0.01; ****: p < 0.001; *****: p < 0.0001).
such difference between NMOSD and MS eyes. However, when inspecting the corresponding plot, one must note that the results for MS-eyes are heavily skewed by a single MS-ON eye with far outlying visual function. This eye belongs to a patient with a single severe ON episode leading to blindness. When excluding this eye, MS-eyes would show a trend of a less steep loss of HCV A than NMOSD-eyes ($\text{Beta} = 0.022 \pm 0.012; p = 0.07$).

None of the patient groups showed a relevant association between inner rim volume and any measure of visual function above the cutoff point of $0.1 \, \text{mm}^3$ (Fig. 3). However, the mixed linear spline model showed a significant association of inner rim volume and LCV A ($\text{Beta} = -20.55 \pm 7.72; p = 0.01$), MD of visual fields ($\text{Beta} = 275.26 \pm 123.10; p = 0.03$) and—in trend—HCV A ($\text{Beta} = -11.14 \pm 6.24; p = 0.08$) in AQP4-IgG seropositive NMOSD eyes. MOGAD eyes did not show an identifiable pattern in the relationship between inner rim volume and visual function. MS eyes had a significant association between high contrast acuity and inner rim volume below the $0.1 \, \text{mm}^3$ cutoff ($\text{Beta} = -11.29 \pm 4.86; p = 0.03$), which was again entirely dependent on the outlier described above. When excluding this outlier, no significant association between inner rim volume of MS-eyes
and any measure of visual function was found. In female participants, only in the AQP4-IgG seropositive group significant associations were found (Supplementary Table 2).

### Association between inner rim volume and retinal neuro‑axonal damage.
To further identify the mechanism behind the greater loss of HCVA in NMO-eyes compared with MOGAD, we analyzed the relationship between pRNFL and foveal inner rim volume (Fig. 4). There was a positive association between inner rim volume and pRNFL in AQP4-IgG seropositive NMO-eyes (Beta = 724.92 ± 208.93; p = 0.001, Fig. 4a), suggesting a simultaneous degradation of both over the course of the disease. MOGAD eyes showed a trend to a similar association (Beta = 653.78 ± 329.25; p = 0.06, Fig. 4b) while MS-ON eyes also showed a significant albeit smaller positive association (Beta = 315.02 ± 130.76; p = 0.04, Fig. 4c). However, if only ON-eyes were considered, there was a significant positive association between inner rim volume and pRNFL in NMO-eyes (Beta = 724.24 ± 206.56; p = 0.004) and to a lesser degree in MS (Beta = 507.79 ± 167.89; p = 0.007), but not in MOGAD (Beta = 241.82 ± 280.30; p = 0.41). In a subgroup analysis of only female participants, there was only a minor difference in effect size concerning the association between IRV and pRNFL (not shown).

In addition, inner rim volume was not associated with the number of ON in NMO-eyes (Beta = −0.0009 ± 0.0009; p = 0.36) and MOGAD (Beta = −0.0002 ± 0.0014; p = 0.91) but showed a very subtle association in MS-eyes (Beta = −0.004 ± 0.002; p = 0.04). Disease duration had no significant effect on inner rim volume in either of the disease groups (NMO: Beta = −2.66e−6 ± 1.21e−5; p = 0.83, MOGAD: Beta = −2.66e−5 ± 3.21e−5; p = 0.42, MS: Beta = −7.47e−4 ± 8.67e−4; p = 0.40).

### Discussion
In this study we analyzed the association between retinal structural damage including foveal changes and visual function in AQP4-IgG seropositive NMO patients and compared findings against MOGAD patients, MS patients and healthy controls. We found that (a) patients with AQP4-IgG seropositive NMO present with more impaired HCVA than MOGAD and MS despite similar retinal neuroaxonal damage; (b) in contrast, LCVA and MD are similarly affected in NMO, MOGAD and MS; (c) in all diseases the structure–function association followed a broken-stick model: there was no association between visual function and OCT above a certain threshold, but once a certain threshold was reached, more profound structural damage was associated with worse visual function. Here, HCVA and MD showed a steeper association with structural damage in NMO compared with MOGAD, with no significant difference when it comes to the rate of loss of LCVA; (d) Eyes from patients with NMO show additionally an association between foveal inner rim volume and visual function.

Several studies have investigated the relationship between visual and structural parameters in neuroinflammatory diseases. In MS and MOGAD, loss of visual field sensitivity does not follow a linear association with pRNFL thinning, but shows a drastic deterioration after a large amount of pRNFL is lost.32,33,34. This is similar to how visual function relates to neuroaxonal damage in glaucoma, where this type of association has been termed broken stick model35. Alternatively, a linear association between visual acuity and retinal layer thickness has been proposed in MS,36 as well as AQP4-IgG seropositive NMO and MOGAD.37. In our study, the association of MD, HCVA and LCVA to pRNFL and GCIP thickness followed a broken stick model and not a linear association, with a threshold of approximately 60 µm for both pRNFL and GCIP. There was no such association for either disease group above 60 µm, supporting that neuroaxonal damage in the context of neuroinflammatory disease leads to loss of visual function, but only has a measurable effect once a certain amount of neuroaxonal content is lost. In other studies this cutoff point has been identified as being 50 µm for MOGAD37, 75 µm for MS35,36 or 60 µm.
Figure 3. Association between visual function and foveal inner rim volume. Effect of inner rim volume on (a) high contrast visual acuity, (b) low contrast visual as well as (c) mean deviation of visual fields of patients` eyes. Cutoff for mixed linear spline model was visually identified at 0.1 mm³. AQP4-IgG: Aquaporin-4-IgG seropositive patients; MOG-IgG: Myelin-Oligodendrocyte-Glycoprotein-IgG seropositive patients; MS: Multiple Sclerosis patients; HCVA: High contrast visual acuity; LCVA: Low contrast visual acuity; MD: mean deviation.
for NMOSD33. Considering differences in methods and raters as well as the sizable interindividual differences of retinal layer thickness, these results are comparable.

Although the loss of retinal layers started to affect all patient groups at approximately the same threshold, NMOSD eyes suffered a steeper drop in HCVA and MD than MOGAD eyes. The less pronounced difference in the drop of visual fields is in line with a recent study that showed good HCVA outcome after MOGAD-associated ON, but worse visual fields outcomes31. The finding that each µm of retinal layer lost is worth more visual function in NMOSD than MOGAD is in line with a previous study, in which only eyes of NMOSD but not MOGAD patients showed an association between worse HCVA with lower GCIP5. Another study showed worse HCVA and stronger pRNFL thinning in MOGAD eyes at ON nadir, but also a higher recovery rate of visual function that resulted in comparable long term visual outcome in MOGAD and MS37. Together, our study and these others suggest that neuroaxonal damage alone cannot explain the differences in visual outcome between NMOSD patients and MOGAD or MS patients.

A model that might explain this discrepancy is AQP4-IgG driven Müller-cell dysfunction. AQP4-IgG seropositive NMOSD leads to a primary inflammatory astrocytopathy while demyelination is only a secondary process. MOGAD and MS on the other hand primarily result in inflammatory demyelination38. In the healthy retina, Müller cells are responsible for many trophic and regulatory functions, such as glucose metabolism, regulating

Figure 4. Association between inner rim volume and pRNFL in (a) NMOSD (Beta = 724.92 ± 208.93; p = 0.001), (b) MOGAD (Beta = 653.78 ± 329.25; p = 0.06) and (c) MS eyes (Beta = 315.02 ± 130.76; p = 0.02) AQP4-IgG: Aquaporin-4-IgG seropositive patients; MOG-IgG: Myelin-Oligodendrocyte-Glycoprotein-IgG seropositive patients; MS: Multiple Sclerosis patients.
blood flow as well as ion and water homeostasis. Müller cells are found ubiquitously in the human retina, but are most dense in and around the fovea. In NMOSD, Müller cells are a target of AQP4-IgG, which was demonstrated both in animal studies as well as NMOSD patients. In vitro, AQP4-IgG causes non-inflammatory AQP4 receptor internalization and reduced proliferation in Müller cells; in fact, Müller cell dysfunction has been directly demonstrated through specific ERG-waveforms in eyes of NMOSD patients.

We found a significant association between the fovea’s inner rim volume and pRNFL in AQP4-IgG positive NMOSD eyes. Indeed, inner rim volume had a similar effect on visual function in NMOSD as pRNFL and GCIP thinning. After a certain amount of inner rim volume is lost (below 0.1 mm³), drastic deterioration of visual function begins. Regardless of underlying disease, inner rim volume in eyes with values > 0.1 mm³ were not associated with visual function. In NMOSD-ON eyes, there was also a strong positive association between foveal inner rim volume and pRNFL, which could not be found in MOGAD-ON eyes, suggesting AQP4-IgG transmitted Müller cell damage concomitant to neuroaxonal damage. Our results further suggest that the fovea’s contribution of vision is largest in HCV A, less noticeable in visual fields and seemingly absent in LCV A. This fits nicely to the test principles of each vision test, where HCV A is typically dominated by foveal vision, whereas in low contrast vision testing peripheral rods dominate, and visual field testing comprises an average over central and peripheral retinal areas.

In our study, MS patients’ eyes behaved in a similar fashion as NMOSD eyes in regard to neuroaxonal damage but surprisingly also foveal inner rim volume, yet to a lesser degree. While the association of visual function to pRNFL and GCIP in the MS group was dependent on a single outlying eye, there was also a similar association between inner rim volume and pRNFL, marking a clear difference to the MOGAD and control group. A primary retinopathy has been reported in MS by one study, which could explain this association. Alternatively, the density of the retina’s superficial and deep capillary plexus is reduced in MS patients, making an indirect effect through foveal microcirculation possible.

An important strength of our study is the availability of a comprehensive suite of visual tests, which reports HCV A, LCV A and MD. Our study has several important limitations, most notably the small sample size. This is problematic for foveal parameters, which show large interindividual variation even in healthy eyes.

Another problem is the significant difference in age between the NMOSD and MS group, which we decided to not correct, because the effect of age on retinal layers is subtle and given the low sample size we did not want to overstrain the models. Further, vision assessment was carried out in one single session, but at least two independent measurements of visual performance are recommended to account for daily variation in performance of patients. Another concern may be that, the minimum timeframe after ON of 3 months that we set as an inclusion criterion might be too short, especially when considering MOGAD, in which further changes to pRNFL have been shown to occur within 6 months from clinical attacks other than ipsilateral ON. On the other hand, another study did not show further thinning of pRNFL and GCIP 6 months after ON in MS and NMOSD when compared to the respective thickness after 3 months. A further potential confounder may be different numbers of ON in the different cohorts. While the number of ON was not significantly different between MOGAD and NMOSD patients, we cannot fully rule out that non-linear effects of the number of ON, which have been reported both for NMOSD and MOGAD, may have influenced the result. Because most patients in our study had multiple ON episodes, we were not able to perform an analysis in eyes with only one ON.

Although our results indicating a potential contribution of Müller cells to high contrast vision loss are promising, the findings need to be interpreted with caution considering the low sample size and the other above-mentioned limitations. Important other contributors should be alternatively considered and investigated, most notably optic nerve demyelination and retinal vascular changes. Visual evoked potentials (VEP) could elucidate how optic nerve demyelination affects visual outcome, and VEP latency changes were indeed reported recently in NMOSD even in absence of ON. Alternatively, the size of the foveal avascular zone (FAZ) as measured by OCT angiography is already of interest in several other ophthalmological diseases. The possible importance of foveal microcirculation in the development of visual impairment in NMOSD becomes especially apparent when considering the trophic role of Müller cells and the high expression of AQP4 in foot processes facing blood vessels.

Data availability
As patient consent did not cover publication of individual data, the data used in this manuscript will be shared on reasonable request from corresponding author.

Received: 14 January 2022; Accepted: 6 September 2022
Published online: 20 October 2022

References
1. Reich, D. S. et al. Damage to the optic radiation in multiple sclerosis is associated with retinal injury and visual disability. Arch. Neurol. 66, 998–1006 (2009).
2. Jarius, S., Wildemann, B. & Paul, F. Neuromyelitis optica: Clinical features, immunopathogenesis and treatment. Clin. Exp. Immunol. 176, 149–164 (2014).
3. Tajifrouz, D. A., Bhatti, M. T. & Chen, J. J. Clinical characteristics and treatment of MOG-IgG-associated optic neuritis. Curr. Neurol. Neurosci. Rep. 26, 100 (2019).
4. Brandt, A. U. et al. Frequent retinal ganglion cell damage after acute optic neuritis. Mult. Scler. Relat. Disord. 22, 141–147 (2018).
5. Sotilchonos, E. S. et al. Aquaporin-4 IgG seropositivity is associated with worse visual outcomes after optic neuritis than MOG-IgG seropositivity and multiple sclerosis, independent of macular ganglion cell layer thinning. Mult. Scler. J. 26, 1360–1371 (2020).
6. Bennett, J. L. & Owens, G. P. Neuromyelitis optica: Deciphering a complex immune-mediated astrocytopathy. J. Neuroophthalmol. 37, 291–299 (2017).
7. Wingerchuk, D. M. et al. International consensus diagnostic criteria for neuromyelitis optica spectrum disorders. *Neurology* **85**, 177–189 (2015).
8. Iarius, S. et al. Neuromyelitis optica. *Nat. Rev. Dis. Prim.* **2**, 85 (2020).
9. Nagelhus, E. A. et al. Aquaporin-4 water channel protein in the rat retina and optic nerve: Polarized expression in Muller cells and fibrous astrocytes. *J. Neurosci.* **18**, 2506–2519 (1998).
10. You, Y. et al. Evidence of Müller glial dysfunction in patients with aquaporin-4 immunoglobulin G-positive neuromyelitis optica spectrum disorder. *Ophthalmology* **126**, 801–810 (2019).
11. Jeong, I. H., Kim, H. J., Kim, N.-H., Jeong, K. S. & Park, C. Y. Subclinical primary retinal pathology in neuromyelitis optica spectrum disorder. *J. Neurol.* **263**, 1343–1348 (2016).
12. Oertel, F. C. et al. Microstructural visual system changes in AQ4P-antibody-seropositive NMOSD. *Neurol. Neuroimmunol. Neuroinflamm.* **4**, e334 (2017).
13. Motamedi, S. et al. Altered fovea in AQ4P-IgG-seropositive neuromyelitis optica spectrum disorders. *Neurol. Neuroimmunol. Neuroinflamm.* **7**, e805 (2020).
14. Waters, P. et al. Multicentre comparison of a diagnostic assay: Aquaporin-4 antibodies in neuromyelitis optica. *J. Neurol. Neurosurg. Psychiatry* **87**, 1005–1015 (2016).
15. Reindl, M. et al. International multicenter examination of MOG antibody assays. *Neurol. Neuroimmunol. Neuroinflamm.* **7**, e718 (2020).
16. Iarius, S. et al. MOG encephalomyelitis: International recommendations on diagnosis and antibody testing. *J. Neuroinflamm.* **15**, 134 (2018).
17. Polman, C. H. et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann. Neurol.* **69**, 292–302 (2011).
18. Kurtzke, J. F. Rating neurologic impairment in multiple sclerosis: An expanded disability status scale (EDSS). *Neurology* **33**, 1444–1452 (1983).
19. Motamedi, S. et al. Normative data and minimally detectable change for inner retinal layer thicknesses using a semi-automated OCT image segmentation pipeline. *Front. Neurol.* **10**, 1117 (2019).
20. Lang, A. et al. Retinal layer segmentation of macular OCT images using boundary classification. *Biomed. Opt. Express* **4**, 1133 (2013).
21. Tewarie, P. et al. The OSCAR-IB consensus criteria for retinal OCT quality assessment. *PloS ONE* **7**, e34823 (2012).
22. Yadav, S. K. et al. CuBe: parametric modeling of 3D foveal shape using cubic Bézier. *Biomed. Opt. Express* **8**, 4181–4199 (2017).
23. Ayutlu, A. et al. APOSTEL 2.0 recommendations for reporting quantitative optical coherence tomography studies. *Neurology* **13**, 68–79 (2021).
24. R Core Team. R: A language and environment for statistical computing. (2014).
25. Wickham, H. ggplot2: Elegant Graphics for Data Analysis (Springer-Verlag, 2016).
26. Bates, D., MAchler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**, 1–48 (2015).
27. Barton, K. MuMIn: Multi-model inference. (2019).
28. Cheng, H.,aron, M., Schiffman, J. S., Tang, R. A. & Frishman, L. J. The relationship between visual field and retinal nerve fiber layer measurements in patients with multiple sclerosis. *Investig. Ophthalmol. Vis. Sci.* **48**, 5798 (2007).
29. Costello, F. et al. Quantifying axonal loss after optic neuritis with optical coherence tomography. *Ann. Neurol.* **59**, 963–969 (2006).
30. Schmidt, C. O., Ittermann, T., Schulz, A., Grabe, H. J. & Baumeister, S. E. Linear, nonlinear or categorical: How to treat complex associations? Splines and nonparametric approaches. *Int. J. Public Health* **58**, 161–165 (2013).
31. Deschamps, R. et al. Visual field loss and structure–function relationships in optic neuritis associated with myelin oligodendrocyte glycoprotein antibody. *Mult. Scler.* **27**, 855–863 (2021).
32. Costello, F. et al. Tracking retinal nerve fiber layer loss after optic neuritis: A prospective study using optical coherence tomography. *Mult. Scler.* **14**, 893–905 (2008).
33. Mekhasingharak, N. et al. Visual function and inner retinal structure correlations in aquaporin-4 antibody-positive optic neuritis. *Ipn. J. Ophthalmol.* **62**, 598–604 (2018).
34. Filippatou, A. G. et al. Evidence of subclinical quantitative retinal layer abnormalities in AQ4P-IgG seropositive NMOSD. *Mult. Scler.* **27**, 1738 (2021).
35. Alasul, Y. et al. Correlation of retinal nerve fiber layer thickness and visual fields in glaucoma: A broken stick model. *Am. J. Ophthalmol.* **157**, 953–959.e2 (2014).
36. Fisher, J. B. et al. Relation of visual function to retinal nerve fiber layer thickness in multiple sclerosis. *Ophthalmology* **113**, 324–332 (2006).
37. Vicini, R., Brügger, D., Abegg, M., Salmen, A. & Grabe, H. M. Differences in morphology and visual function of myelin oligodendrocyte glycoprotein antibody and multiple sclerosis associated optic neuritis. *Neur. Neurolog*. **268**, 276–284 (2021).
38. Weber, M. S., Derfuss, T., Metz, I. & Brück, W. Defining distinct features of anti-MOG antibody associated central nervous system demyelination. *Ther. Adv. Neurol. Disord.* **11**, 17528641876208 (2018).
39. Bringmann, A. et al. Müller cells in the healthy and diseased retina. *Prog. Retin. Eye Res.* **25**, 397–424 (2006).
40. Félix, C., Levin, M. H. & Verkman, A. S. Complement-independent retinal pathology produced by intravitreal injection of neuromyelitis optica immunoconjugate G. J. *Neuroinflammation* **13**, 275 (2016).
41. Oertel, F. C., Zimmermann, H., Paul, F. & Brandt, A. U. Optical coherence tomography in neuromyelitis optica spectrum disorders: Potential advantages for individualized monitoring of progression and therapy. *Eur. J. Neurol.* **9**, 21–33 (2018).
42. Netti, V. et al. Aquaporin-4 removal from the plasma membrane of human müller cells by AQP4-IgG from patients with neuromyelitis optica induces changes in cell volume homeostasis: The first step of retinal injury?. *Mol. Neurobiol.* **58**(10), 5178–5193 (2021).
43. Yilmaz, H., Ersoy, A. & Icel, E. Assessments of vessel density and foveal avascular zone metrics in multiple sclerosis: An optical coherence tomography angiography study. *Eye* **34**, 771–778 (2019).
44. Provis, J. M., Dubis, A. M., Maddess, T. & Carroll, J. Adaptation of the central retina for high acuity vision: Cones, the fovea and the avascular zone. *Prog. Retin. Eye Res.* **35**, 63–81 (2013).
45. Oertel, F. C. et al. Optical coherence tomography in myelin-oligodendrocyte-glycoprotein antibody-seropositive patients: A longitudinal study. *J. Neuroinflammation* **16**, 154 (2019).
46. Syc, S. B. et al. Optical coherence tomography segmentation reveals ganglion cell layer pathology after optic neuritis. *Brain* **135**, 521–533 (2012).
47. Pach, E. et al. MOG-IgG in NMO and related disorders: a multicenter study of 50 patients. Part 4: Afferent visual system damage after optic neuritis in MOG-IgG-seropositive versus AQ4P-IgG-seropositive patients. *J. Neuroinflammation* **13**, 282 (2016).
48. Oertel, F. C. et al. Retinal optical coherence tomography in neuromyelitis optica. *Neurol. Neuroimmunol. Neuroinflamm.* **8**, e1068 (2021).
49. Ringelstein, M. et al. Longitudinal optic neuritis-unrelated visual evoked potentials changes in NMO spectrum disorders. *Neurology* **94**, e407–e418 (2020).
50. Zivkovic, M. et al. Foveal avascular zone in normal tension glaucoma measured by optical coherence tomography angiography. *Biomed. Res. Int.* **2017**, 3079141 (2017).
51. Balaratnasingam, C. et al. Visual acuity is correlated with the area of the foveal avascular zone in diabetic retinopathy and retinal vein occlusion. *Ophthalmology* 123, 2352–2367 (2016).
52. Lennon, V. A., Kryzer, T. J., Pittoc, S. J., Verkman, A. S. & Hinson, S. R. IgG marker of optic-spinal multiple sclerosis binds to the aquaporin-4 water channel. *J. Exp. Med.* 202, 473–477 (2005).

**Acknowledgements**

We thank Nocturne GmbH for kindly providing fovea shape analysis.

**Author contributions**

N.G., F.O., F.P., A.B. and H.Z. contributed to conception and design of the work. All authors contributed to data collection. N.G., A.B. and H.Z. contributed to drafting the article. N.G. wrote the main manuscript text. N.G., F.O., K.R., F.P., A.B. and H.Z. contributed to critical revision of the article. All authors contributed to the final approval of the version to be published.

**Funding**

Open Access funding enabled and organized by Projekt DEAL.

**Competing interests**

FO was an employee of Nocturne UG (2018) and received research support by American Academy of Neurology (2021–2023), National Multiple Sclerosis Society (2021–2024) and Deutsche Gesellschaft für Neurologie (2021)—all independent from this project. KR received research support from Novartis Pharma, Merck Serono, German Ministry of Education and Research, European Union (821283-2), Stiftung Charité (BIH Clinical Fellow Program) and Arthur Arnstein Foundation; received travel grants from Guthy Jackson Charitable Foundation J. Bellmann-Strobl has received speaking honoraria and travel grants from Bayer Healthcare, and sanofi-aventis/Genzyme, in addition received compensation for serving on a scientific advisory board of Roche, unrelated to the presented work. F. Paul served on the scientific advisory boards of Novartis and MedImmune; received travel funding and/or speaker honoraria from Bayer, Novartis, Biogen, Teva, Sanofi-Aventis/Genzyme, Merck Serono, Alexion, Chugai, MedImmune, and Shire; is an associate editor of Neurology: Neuroimmunology & Neuroinflammation; is an academic editor of PLoS ONE; consulted for Sanofi Genzyme, Biogen, MedImmune, Shire, and Alexion; received research support from Bayer, Novartis, Biogen, Teva, Sanofi-Aventis/Geynzmex, Alexion, and Merck Serono; and received research support from the German Research Council, Werth Stiftung of the City of Cologne, German Ministry of Education and Research, Arthur Arnstein Stiftung Berlin, EU FP7 Framework Program, Arthur Arnstein Foundation Berlin, Guthy-Jackson Charitable Foundation, and NMSS. AUB is co-founder and shareholder of medical technology companies Nocturne GmbH and Motognosis GmbH. He is named as inventor on several patent applications describing MS biomarkers, visual perceptive computing based motor function analysis, and retinal image analysis. H.G. Zimmermann received research grants from Novartis and speaking honoraria from Bayer Healthcare and Novartis. The other authors report no competing interests.

**Additional information**

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1038/s41598-022-19848-4.

**Correspondence** and requests for materials should be addressed to H.G.Z.

**Reprints and permissions information** is available at www.nature.com/reprints.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© The Author(s) 2022