Research Article

Cardioprotective Effects of Taurisolo® in Cardiomyoblast H9c2 Cells under High-Glucose and Trimethylamine N-Oxide Treatment via De Novo Sphingolipid Synthesis

Stefania Lama,1 Vincenzo Monda,2 Maria Rosaria Rizzo,3 Marco Dacrema,4 Maria Maisto,4 Giuseppe Annunziata,4 Gian Carlo Tenore,4,5 Ettore Novellino,4 and Paola Stiuso4,1

1Department of Precision Medicine, University of Campania "Luigi Vanvitelli", Naples, Italy
2Department of Experimental Medicine, Section of Human Physiology, University of Campania "Luigi Vanvitelli", Naples, Italy
3Department of Advanced Medical and Surgical Sciences, University of Campania "Luigi Vanvitelli", Naples, Italy
4Department of Pharmacy, University of Naples Federico II, Naples, Italy

Correspondence should be addressed to Gian Carlo Tenore; giancarlo.tenore@unina.it and Paola Stiuso; paola.stiuso@unicampania.it

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1. Introduction

Obesity is a medical condition mainly characterized by increased body fat accumulation, which negative effects on health are well-known. Excessive adipose tissue is among the leading causes of chronic inflammation that, in turn, plays a pivotal role in developing or worsening the outcome of various pathologies such as metabolic, cardiovascular, respiratory, viral, and tumoral [1]. Obesity is a widely spread health problem in the world, and it has been recognized as a risk factor for the development of metabolic disorders, such as type 2 diabetes, cardiovascular disease, or atherosclerosis [2, 3]. Diet-induced obesity (DIO) leads a significant increase of oxidative stress (OS) and reduced antioxidant defense, and it is associated with a typical chronic low-grade inflammation, which results in severe cardiovascular complications [4, 5]. Moreover, in cardiomyocytes, DIO induced many alterations, including mitochondria dysfunction [5], endoplasmic reticulum stress [6], apoptosis, and autophagy. Many studies suggest that enhanced autophagy acts as a protective mechanism against OS in the liver, adipose tissue, and skeletal muscle [7, 8]. Furthermore, when cardiac stresses are
sustained for a prolonged period, e.g., in high blood sugar levels, upregulation of autophagy promotes myocyte cellular architecture remodeling. Sphingolipids (SLs), a heterogeneous lipid class that includes ceramide (Cer), sphingosine-1-phosphate, and dihydroceramide present in all membrane structures, have been associated to mediate distinct autophagic pathways described as protective autophagy and autophagy-associated cell death [1, 9]. A recent study showed that Cer directly interacts with microtubule-associated protein light chain 3 (LC3) on mitochondrial membranes, inducing autophagy [10]. Therefore, Cer has been shown to reduce the nutrient transporters in plasma membrane resulting in autophagy activation. S1P has emerged as a cell-proliferative lipid messenger. It has been found to induce survival-mediated or protective autophagy under nutrient starvation, but it was shown to not be related to Beclin1 protein accumulation or class I PI3K or Akt suppression [11, 12]. The conversion of Cer to S1P simultaneously accumulates protein accumulation or class I PI3K or Akt suppression [11, 12]. The goal of the present study was to investigate the effects and signaling mechanisms of Taurisolo® on proliferation, OS, morphological remodeling, and autophagy, in trimethylamine TMAO and hyperglycemia damaged embryonic rat cardiomyoblast H9c2 cells.

2. Materials and Methods

2.1. Chemicals. All chemicals, standards, and reagents used were either analytical or mass grade reagents. The water was treated in a Milli-Q water purification system (Millipore, Bedford, MA, United States) before use. Bovine serum albumin (BSA) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) and trimethylamine N-oxide (TMAO) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Phosphate-buffered saline (PBS) and trypsin-EDTA were from Lonza (Milano, Italy). Fetal bovine serum (FBS) and Dulbecco’s modified Eagle’s medium (DMEM) were purchased from Gibco (Grand Island, NY, USA). Western blot analysis was performed using the following primary antibodies: polyclonal antibody (polyAb) p-ERK 44/42 and polyAb ERK 44/42 from Cell Signaling Technology (Beverly, MA, USA); monoclonal Ab Mn-SOD was from Santa Cruz Biotechnology (San Diego, CA, USA). Blots were incubated with horseradish peroxidase (HRP)-conjugated goat anti-rabbit or HRP-conjugated goat anti-mouse (Immuno-reagents Inc., Raleigh, NC, USA) secondary antibodies. Anti-mAb was purchased from Santa Cruz Biotechnology and secondary antibodies conjugated to Alexafluor 488 from Life Science, Portland, OR, USA, and 4’,6-diamidino-2-phenylindole (DAPI) from Sigma-Aldrich.

2.2. Taurisolo® Supplement. Taurisolo® is a supplement consisting of a polyphenol extract obtained from Aglifamico cultivar grape, collected during the autumn 2016 harvest. Firstly, the Department of Pharmacy, University of Naples Federico II (Naples, Italy), provided the supplement formulation; then, the large-scale production was accomplished by MBBMed Company (Turin, Italy). For the polyphenol extract production, grapes were extracted with water (50°C), and the solution was filtrated and concentrated and underwent a spray-drying process with maltodextrins as support (40-70%) to obtain a fine microencapsulated powder. As previously reported, the High-Performance Liquid Chromatography-diodearray detector (HPLC-DAD, Jasco Inc., Easton, MD, USA) polyphenol profile of Taurisolo® revealed the presence of: Ferulic acid 10.5 ± 0.70 μg/g, Resveratrol 13.6 ± 0.64 μg/g, Caffeic acid 20.7 ± 0.76 μg/g, Procyanidin B3 dimer 22.05 ± 6.61 μg/g, p-coumaric acid 27.9 ± 0.66 μg/g, Rutin 28.4 ± 0.70 μg/g, Quercetin 40.22 ± 7.11 μg/g, Procyanidin C2 trimer 44.6 ± 0.66 μg/g, Procyanidin B4 dimer 56.6 ± 0.88 μg/g, Procyanidin B1 dimer 62.8 ± 0.59 μg/g, Procyanidin B2 dimer 426.5 ± 5.92 μg/g, Syringic acid 539.2 ± 6.02 μg/g, Epicatechin 886.0 ± 7.82 μg/g, Gallic acid 1463.4 ± 65.5 μg/g, and Catechin 4087.0 ± 64.5 μg/g [18].

2.3. Cell Culture. Rat cardiomyocytes (H9c2) (ATCC, Manassas, VA) cells were cultured in DMEM, at two different glucose concentration 5.5 mM (NH-H9c2) and 44 mM (HG-H9c2), supplemented with 10% fetal bovine serum, 100 U/mL of penicillin, and 100 lg/mL of streptomycin in 150 cm² tissue culture flasks at 37°C in a humidified atmosphere of 5% CO2. The cells were fed every 2–3 days and sub-cultured once they reached 70–80% of confluence. After 4 hr incubation, cells were washed with 1% PBS to remove unattached dead cells and treated with Tau (0.5 μg/mL), TMAO (50 μM), and TMAO/Tau combination.

2.4. Cell Proliferation Assay. The evaluation of cell proliferation was performed on NG-H9c2 and HG-H9c2 cell line after 24 and 72 hr incubation with Taurisolo®, TMAO, and TMAO/Tau combination. The cells were seeded in 96-well plates in a number of 30 × 10⁵ per well. The growth was assessed by MTT viability assay as previously described [19]. Then, MTT assay was carried out by triplicate determination on at least three separate experiments. All data were expressed as mean ± SD. We have determined also the cell number and proliferation by TC10 automated cell counter (Bio-Rad, Milan, Italy).

2.5. Morphological Evaluation of Cardiomyocytes by Confocal Microscopy. After 72 hr incubation with Tau, TMAO, and TMAO/Tau combination, the HG-H9c2 cells were fixed for
20 min with a 3% (w/v) paraformaldehyde (PFA) solution and permeabilized for 10 min with 0.1% (w/v) Triton X-100 in phosphate-buffered saline (PBS) at room temperature. To prevent nonspecific interactions of antibodies, cells were treated for 2 hr in 5% fetal bovine serum (FBS) in PBS; then, cells were incubated with a specific mouse monoclonal anti-body raised against actin (1 : 500 Alexa Fluor®, BD Pharmin-gen™) for 24 hr at 37°C. The slides were mounted on microscope slides by Mowiol. The analyses were performed with a Zeiss LSM 510 microscope equipped with a plan-apochromat objective X 63 (NA 1.4) in oil immersion. Actin fluorescence was collected in a multitrack mode. The nuclei were stained with 4′,6-diamidino-2-phenylindole (DAPI) [20].

2.6. Nitrite Levels. Nitrite was measured by the Griess reaction. Briefly, 50 μL of medium was mixed with an equal volume of the Griess reagent (0.5% sulfanilamide, 2.5% H₃PO₄, and 0.05% naphthylethylene diamine in H₂O) and incubated for 10 min at room temperature. Absorbance was assayed at 550 nm and compared with a standard curve obtained using sodium nitrite. [21]

2.7. Thiobarbituric Acid-Reactive Species (TBARS) Assay. Samples were incubated with 0.5 mL of 20% acetic acid, pH 3.5, and 0.5 mL of 0.78% aqueous solution of thiobarbituric acid. After heating at 95°C for 45 min, samples were centrifuged at 4000 rpm for 5 min. The TBARS were quantified by spectrophotometry at 532 nm. Results were expressed as

![Figure 1: Taurisolo® (Tau) counteracts the H9c2 cell injury induced from TMAO in hyperglycemic condition. H9c2 cells were grown in standard (5.5 mM Glu NG-H9c2) and high glucose (44 mM Glu, HG-H9c2) condition. The HG-H9c2 cells were treated, for 72 h with Tau (0.5 μg/μL), TMAO(50 μM), and TMAO-Tau combination. (a) The cell survival was performed with MTT assay. (b, c) AST and LDH as makers of cell injury were performed by the colorimetric assay. (d) LDH1/LDH2 isoenzymes ratio was evaluated at 72 h by the colorimetric assay. Each experiment was repeated at least three times. Results are expressed as mean ± SD.](image-url)
TBARS μM/g of proteins. Each data point is the average of triplicate measurements, with each experiment performed in triplicate. Results are expressed as mean ± SD.

2.8. AST and LDH Assay. NG-H9c2 and HG-H9c2 cardiomyocytes (1 × 10^5 cells/well) after 24 and 72 hr incubation with Taurisolo®, TMAO, and TMAO/Tau combination were cultured in 6-well plates. The medium was collected for the measurement of the Aspartate transaminase (AST) and Lactate dehydrogenase (LDH) enzymes, including isoform 1 and 2 release. The enzyme activity was measured using an Abbott Aeroset fully automatic biochemical analyzer (Abbott Laboratories, USA). The levels of enzymes were assayed according to the instructions provided with the corresponding enzymatic kits.

2.9. Western Blots. We followed the methods of Vanacore et al. 2018 [22] for evaluation the protein expression by Western blot. Briefly, the cells were cultured at different condition for 72 hr, and then, cell pellets were lysed with 1 mL of lysis buffer (1% Triton, 0.5% sodium deoxycholate, 0.1 M NaCl, 1 mM Ethylenediamine tetra-acetic acid (EDTA), pH 7.5, 10 mM Na_2HPO_4, pH 7.4, 10 mM Phenylmethyl sulfonyl fluoride, 25 mM benzamidine, 1 mM leupeptin, and 0.025 units/mL aprotinin). The lysates were centrifuged at 12,000 rpm for 10 min at 4°C. Equal amounts of protein extracts were separated by SDS-PAGE, electrotransferred to nitrocellulose, and reacted with the different antibodies (ERK, pERK, LC3II, and Mn-SOD). All Western blots were repeated for three times. GAPDH was used as internal control. To quantify the results, the relative amount of each protein was determined.

2.10. Mass Spectrometry. LIPID MAPS Lipidomics Gateway and Human Metabolome Database queries were used to assign putative identities to mass features using based on mass accuracy within ±1 Da. (http://www.lipidmaps.org/data/structure/index.html.)

3. Results and Discussion

The present study was designed to test the ability of Taurisolo® to counteract or reduce the high glucose and TMAO-induced cardiomyocytes damage. All experiments were performed utilizing H9c2 cardiomyoblast cells growth in DMEM with 5.5 mM glucose (NG-H9c2), 44 mM glucose (HG-H9c2); both NG and HG-H9c2 cells were treated with 50 μM TMAO (TNG-H9c2 THG-H9c2). We have set these concentrations of glucose and TMAO, because they produced moderate H9c2 cardiomyocyte toxicity and could be the conditions suitable for mimic in vitro diet-induced obesity (DIO) model.

3.1. Cardioprotective Effects of Taurisolo® in H9c2 Cells under TMAO/High Glucose-Induced Cytotoxicity. To gain an insight about the mechanism through which Taurisolo® sustained the cardiomyocyte viability, we have damaged the H9c2 cardiomioblastic cell line with high glucose concentration (44 mM) and TMAO, a molecule linked to obesity and energy metabolism [23, 24] (Figure 1). H9c2 vitality was detected via MTT assay, while LDH and AST release assays were performed for cellular injury. High glucose treatment of H9c2 (44 mM) significantly reduced cell viability (~25%; \( P = 0.011 \)) and increased both AST and LDH release \( (P = 0.037 \) and \( P = 0.0074 \), respectively) compared with the NG-H9c2 growth in standard condition (5.5 mM glucose). In contrast, treatment with Taurisolo® was able to counteract significantly the decrease of vitality and the AST and LDH release in both HG-H9c2 and THG-H9c2 cells, while at the same concentration, Tau-treated NG-H9c2 did not produce the same effect.

In Figure 1(d), we reported the LDH1 : LDH2 isoenzyme ratio, observing that increased of about 1,8 and 2,5 fold in HG-H9c2 and THG-H9c2, respectively, compared to NG-H9c2 (LDH1 : LDH2 ratio = 1). Interestingly, Taurisolo®...
treatment of THG-H9c2 counteracted the increase of LDH1/LDH2 ratio. To demonstrate that the effect of Taurisolo® on proliferation was related to the reduction of reactive oxygen species (ROS) production, we evaluated both lipid peroxidation by aldehide reactive to thiobarbituric acids (TBARS) assay and endogenous free nitric oxide (as NO$_2^-$) by the Griess assay (Figure 2). We observed a significant increase of TBARS production in THG-H9c2 cells compared to Taurisolo®-treated THG-H9c2 cells ($P = 0.0159$) and an increase of NO production in the medium of Taurisolo®-treated THG-H9c2 ($P = 0.038$) compared with untreated THG-H9c2 cells. Furthermore, Taurisolo® treatment counteracted the higher expression of Manganese Superoxide dismutase (Mn-SOD) in THG-H9c2 cells (Figure 3).

Extracellular signal-regulated protein kinase (ERK) and its phosphorylated form (pERK) are important mediators of various cellular responses, such as proliferation, differentiation, and cell death. It is noting that chronic high glucose induced ERK phosphorylation and cell death. We detected ERK activation by evaluating the pERK/ERK ratio by Western blot analysis using phospho-specific antibodies. ERK activation (Figure 4(a)) was elevated in HG-H9c2 cells compared to NG-H9c2 cells (Figure 4), while it was significantly reduced after TMAO treatment in both NG and HG-H9c2 cells. Of note, treatment with Tau modulated ERK activation in prosurvival manner both HG and THG-H9c2 cells.

Collectively, these data suggested that the association of 72 h-hyperglycemia and 72 h-TMAO treatment of H9c2 cells led to both cardiac cell cytotoxicity and increase in OS, as mainly indicated by the increases of both lipid peroxidation and antioxidant enzyme levels. Taurisolo® treatment mitigated the increases in AST and LDH levels and increased H9c2 cell viability, suggesting its protective effect on cardiomyocytes during both hyperglycemia- and TMAO-caused damage. Furthermore, these results showed that Taurisolo® reduced THG-induced H9c2 injury by reducing OS. Decreased levels of TBARS are combined with increase of free NO production, suggesting a potential involvement of Taurisolo® in regulating the balance between NO and peroxynitrite. A protective effect of NO has also been observed in

Figure 3: Taurisolo® (Tau) increase LC3II expression and counteract high levels of MnSOD. H9c2 cells were grown in standard (5.5 mM Glu NG-H9c2) and high glucose (44 mM Glu, HG9c2) condition. The NG and HG-H9c2 cells were treated, for 72 h with Tau (0.5 μg/μL), TMAO (50 μM), and TMAO-Tau combination. (a) Bar graphs show densitometrically quantified LC3II, MnSOD with respect to house-keeping GAPDH in NG (5.5 mM glucose) and HG-H9c2 cells. Representative immunoblots of Tau treated TNG-H9c2 (b) and THG-H9c2 cells (c). Each data point is the average of triplicate measurements, with each experiment performed in triplicate. Results are expressed as mean ± SD.
endothelial cells, cardiomyocytes [25], and inflammatory cells [26]. When MnSOD is overexpressed, more superoxide radicals are converted to H₂O₂, which acts as a cytotoxic agent, and therefore are removed from the physiological equilibrium, causing an increased production of membrane lipid peroxidation. To demonstrate that the cellular damage effect of high glucose and TMAO was related to increased oxidative stress, we assessed cell vitality after treatment for 24 h with 10 μM N-acetyl cysteine (NAC), widely used as a pharmacological antioxidant and cytoprotective compound. In this experimental condition, we did not observe growth inhibition in both HG and THG-H9c2 cells (data not shown). In conclusion, the protective effect of Taurisolo® on cell viability and cell injury is closely linked to the antioxidant activity of its polyphenol composition. The antioxidant potential of Taurisolo® results in decreased both expression of MnSOD (Figures 3(a)–3(c)) and lipid peroxidation levels. Conversely, the increased release of NO leads to an ERK activation and a consequently pro-survival effect against TMAO damage induced in the H9c2 cells.

3.2. Taurisolo® Increases Autophagic Process and Remodeling α-Actin Distribution under High Glucose- and TMAO-Induced H9c2 Injury. As autophagy is important for the maintenance of mitochondrial homeostasis, it was hypothesized that Taurisolo® may preserve the H9c2 from injury induced by TMAO and hyperglycemia via autophagy activation. To prove this hypothesis, we evaluated the autophagy marker protein microtubule-associated protein LC3II expression by western blot analysis. TMAO treatment of HG-H9c2 induced a downregulation of autophagy, as evidenced by a decrease in LC3II, compared with the HG-H9c2 cells. 72 h-treatment with TMAO/Taurisolo® combination of HG-H9c2 cells was able to reverse the decrease of LC3II expression in THG-H9c2 cells (Figures 3(a)–3(c)). Morphological changes induced by both HG and TMAO in H9c2 cells were detected by α-actin immunofluorescence by confocal microscopy; the representative results were shown in Figure 5. The α-actin protein (green signal) in Taurisolo®-treated HG-H9c2 cells was uniformly distributed in the cytoplasm, and cells presented a typical elongated form with respect to untreated HG-H9c2 cells where the shape appeared enlarged and rounded; in THG-H9c2 cells, α-actin accumulated, forming evident punctuated signals.
Meanwhile, Taurisolo® treatment reduced actin aggregation in THG-H9c2 cells. These results demonstrate that exposure to Taurisolo® in both HG and THG-H9c2 cells induced α-actin spatial organization and a functional cell morphological conformation. The purpose of autophagy is to ensure quality control of organelles and proteins, as well as protection of intracellular homeostasis in stress and nutrient efficiency [27–32]. Autophagy is involved in the maintenance of organelle integrity, protein quality [33], and modulated to participate in the pathogenesis of human diseases, such as DM, neurodegenerative diseases, aging, and vascular disease [7, 34, 35]. It has been reported that antioxidant molecules such as resveratrol by increasing autophagic flux ameliorates diabetic cardiomyopathy [36]. In THG-H9c2 damage cell model cells, we demonstrated that Taurisolo®, restoring the autophagic process, induces a reduction of the actin aggregation, restoring a normal cell morphology.

3.3. Taurisolo® Induces the Ex Novo Synthesis of Sphingomyelin, Ceramides, and Their Metabolites. SLs are molecules implicated in cell survival and autophagy. Bioactive lysolipids, including Cer and sphingosine-1-phosphate, may act as both extracellular and intracellular mediators. We used tandem mass spectrometry to investigate which sphingolipids were secreted in the cell medium in Taurisolo®-treated HG and THG-H9c2 cells. In Figure 6, the positive ion mass spectrometry profile of lipid extract in the medium of the HG-H9c2, THG-
H9c2, and both Taurisolo®-treated HG- and THG-H9c2 cells was reported. In both HG- and THG-H9c2 spectra, we identified the following peaks: d18: 1 sphingoid base and phosphocholine headgroup (449 m/z) [37] (Monounsaturated 18-carbon dihydroxylated sphingoid base linked to one chain of palmitic acid denominated 1-O-tricosahexsphinganine-1-O-[D-mannopyranosyl-

α-(1→2)-myo-inositol-1-phosphate] MIPC (d20:0/26:0) (1111 m/z). We observed in the lipid fingerprint profile of Tau-treated HG- and THG-H9c2 cells that the metabolite peaks were centered between 200 and 1500 m/z as sphingosine, C16-C18 sphinganine-1-phosphate, PE-Cer (d16:2/20:1), SM (d18:0/16:0), PI-Cer (d18:0/16:0), 1-O-steroyl-Cer (d18:1/18:0)/Or 1-O-ecosanoyl-Cer (d18:1/16:0), MIPC (18:2/20:1), PIM2 (16:0/18:1), and Glu/Gal-ceramide. Cer plays a central role in sphingolipid metabolism. Cer consists of sphingoid long-chain base linked to an acyl chain via an amide bond and synthesized de novo in the endoplasmatic reticulum (ER); it can be modified into Golgi in sphingomyelin (SM), sphingosine, and glycosphingolipids (e.g., galactosylceramide) and are transported to the plasma membrane (PM). Cer can then be metabolized into ceramide-1-phosphate (C1P) and sphingosine-1-phosphate (SIP) or be resynthesized back into SM. Sphingosine was associated with growth arrest [38] (whereas SP1 promoted cell proliferation and prevents programmed cell death [39]). In H9c2 cells, higher glucose concentration may block the Cer production and then promote transformation in SM and Sph1P, whereas Taurisolo® treatment of HG and THG-H9c2 cells induced a reprogramming lipid metabolism and increased Cer, SM, and Sph1P productions that may protect the cardiomyocyte from glucose cytotoxicity. Further in vivo researches may serve to license Taurisolo® as a useful nutraceutical approach in the prevention of heart damage in obese subjects.

4. Conclusion

It has been previously reported that polyphenols enhance glucose uptake, mainly in the muscular tissue, via increasing expression and/or translocation of GLUT4 transporters [40–42]. Moreover, studies demonstrated that polyphenols Resveratrol and Quercetin exert antioxidant and anti-inflammatory properties [43–46] and can ameliorate the diabetic complications [47]. However, the beneficial effects of polyphenolic compounds in functional foods and nutraceuticals are affected by several factors compromising their bioavailability. Notably, several strategies have been developed by the pharmaceutical and nutraceutical industries in order to counteract this crucial concern, including microencapsulation in various coating materials. Microencapsulation,
Indeed, has been demonstrated to (i) protect polyphenols against degrading factors met during the transit in the gastrointestinal tract, including pH variations and digestive enzymes (increasing, thus, their bioaccessibility) and (ii) favor their absorption across the intestinal barrier (increasing their bioavailability) [48]. In this sense, Taurisolo® is a nutraceutical formulation based on grape polyphenols microencapsulated in maltodextrins. In our previous studies [16], we demonstrated high bioavailability of Taurisolo® polyphenols, leading to assume that also after oral administration, these bioactive compounds may reach high tissue concentrations.

Annunziata and colleagues [16, 17] have demonstrated the ability of the Taurisolo® to reduce circulating levels of TMAO, a molecule related to metabolic syndrome and cardiovascular risk [49]. Moreover, the TMAO levels are inversely associated with the adherence to the Mediterranean diet [50]. The present study was designed to test the ability of Taurisolo® to counteract or reduce the cardiomyocytes damage induced by high glucose concentration and TMAO. The hyperglycemia in ventricular cardiomyoblasts H9c2 cell induces cytotoxicity, OS, and cellular hypertrophy, whereas TMAO reduces autophagy and causes actin accumulation. Taurisolo® exerted a protective effect on both hyperglycemia and TMAO-induced H9c2 cell injury by (1) modulation of OS through its direct scavenger action, (2) induction of morphological changes by α-actin redistribution, and (3) activation of the autophagic process by means of the ex novo synthesis of Cers and their metabolites [51]. Here, we demonstrated that Taurisolo® improves in vitro HG- and TMAO-induced H9c2 dysfunction, suggesting that Taurisolo®, combined with energy restriction, may represent a useful nutraceutical approach for the prevention of cardiomyopathy in obese subjects.

Abbreviations

Sph: Sphingosine  
Sph1P: Sphingosine 1-phosphate  
Cer: Ceramide  
Cer1P: Ceramide 1-phosphate  
GalCer: Galactosylceramide  
GlCer: Glucosylceramide  
LacCer: Lactosylceramide  
GM, GD, GT: Gangliosides  
SM: Sphingomyelin  
LC-MS/MS: Liquid chromatography tandem mass spectrometry  
Q: Quadrupole  
QQQ: Triple quadrupole  
QTTrap: Quadrupole linear ion trap  
PI-P: Phospho-(1′-myo-inositol)  
LC3: Microtubule-associated protein 1 light chain 3.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

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Supplementary Materials

Assignments of the m/z ratios detected in the positive ion mass spectra of the lipid extracts in the growth medium of HG-H9c2 (44 mM glucose) and THG-H9c2 (44 mM glucose + TMAO 50 μM) cells before and after treatment with Tau 0.5 μg/μL.  

References

[1] G. Messina, R. Polito, V. Monda et al., “Functional role of dietary intervention to improve the outcome of COVID-19: a hypothesis of work,” International Journal of Molecular Sciences, vol. 21, no. 9, article 3104, 2020.

[2] P. Hossain, B. Kawar, and M. El Nahas, “Obesity and diabetes in the developing world—a growing challenge,” The New England Journal of Medicine, vol. 356, no. 3, pp. 213–215, 2007.

[3] J. Stock, “Gut microbiota: an environmental risk factor for cardiovascular disease,” Atherosclerosis, vol. 229, no. 2, pp. 440–442, 2013.

[4] C. Chrysohoou, D. B. Panagiotakos, C. Pitsavos et al., “The implication of obesity on total antioxidant capacity in apparently healthy men and women. The ATTICA study,” Nutrition, Metabolism, and Cardiovascular Diseases, vol. 17, no. 8, pp. 590–597, 2007.

[5] C. Patel, H. Ghanim, S. Ravishankar et al., “Prolonged reactive oxygen species generation and nuclear factor-kappaB activation after a high-fat, high-carbohydrate meal in the obese,” The Journal of Clinical Endocrinology and Metabolism, vol. 92, no. 11, pp. 4476–4479, 2007.

[6] J. D. Malhotra, H. Miao, K. Zhang et al., “Antioxidants reduce endoplasmic reticulum stress and improve protein secretion,” Proceedings of the National Academy of Sciences, vol. 105, no. 47, pp. 18525–18530, 2008.

[7] Q. He, S. Sha, L. Sun, J. Zhang, and M. Dong, “GLP-1 analogue improves hepatic lipid accumulation by inducing autophagy via AMPK/mTOR pathway,” Biochemical and Biophysical Research Communications, vol. 476, no. 4, pp. 196–203, 2016.

[8] J. Yan, Z. Feng, J. Liu et al., “Enhanced autophagy plays a cardinal role in mitochondrial dysfunction in type 2 diabetic Goto-Kakizaki (GK) rats: ameliorating effects of (-)-epigallocatechin-3-gallate,” The Journal of Nutritional Biochemistry, vol. 23, no. 7, pp. 716–724, 2012.

[9] W. Zheng, J. Kollmeyer, H. Symolon et al., “Ceramides and other bioactive sphingolipid backbones in health and disease: lipidomic analysis, metabolism and roles in membrane endoplasmic reticulum stress and improve protein secretion,” Biochemical and Biophysical Research Communications, vol. 476, no. 4, pp. 196–203, 2016.
 structure, dynamics, signaling and autophagy," *Biochimica et Biophysica Acta*, vol. 1758, no. 12, pp. 1864–1884, 2006.

[10] R. D. Sentelle, C. E. Senkal, W. Jiang et al., "Ceramide targets autophagosomes to mitochondria and induces lethal mitophagy," *Nature Chemical Biology*, vol. 8, no. 10, pp. 831–838, 2012.

[11] G. Lavieu, F. Scarlatti, G. Sala et al., "Regulation of autophagy by sphingosine kinase 1 and its role in cell survival during nutrient starvation," *The Journal of Biological Chemistry*, vol. 281, no. 13, pp. 8518–8527, 2006.

[12] Y. Li, S. Li, X. Qin et al., "The pleiotropic roles of sphingolipid signaling in autophagy," *Cell Death & Disease*, vol. 5, no. 5, article e1245, 2014.

[13] O. Cuvillier, G. Pirianov, B. Kleuser et al., "Suppression of ceramide-mediated programmed cell death by sphingosine-1-phosphate," *Nature*, vol. 381, no. 6585, pp. 800–803, 1996.

[14] J. R. Van Broeklyn and J. B. Williams, "The control of the balance between ceramide and sphingosine-1-phosphate by sphingosine kinase: oxidative stress and the seesaw of cell survival and death," *Comparative Biochemistry and Physiology Part B: Biochemistry & Molecular Biology*, vol. 163, no. 1, pp. 26–36, 2012.

[15] R. Obeid, H. M. Awwad, Y. Rabagny, S. Graeber, W. Herrmann, and J. Geisel, "Plasma trimethylamine N-oxide concentration is associated with choline, phospholipids, and methyl metabolism," *The American Journal of Clinical Nutrition*, vol. 103, no. 3, pp. 703–711, 2016.

[16] G. Annunziata, M. Maisto, C. Schisano et al., "Effects of Grape Pomace Polyphenolic Extract (Taurisol®) in reducing TMAO serum levels in humans: preliminary results from a randomized, placebo-controlled, cross-over study," *Nutrients*, vol. 11, no. 1, p. 139, 2019.

[17] G. Annunziata, M. Maisto, C. Schisano et al., "Effect of grape pomace polyphenols with or without pectin on TMAO serum levels assessed by LC/MS-based assay: a preliminary clinical study on overweight/obese subjects," *Frontiers in Pharmacology*, vol. 10, p. 575, 2019.

[18] D. Lapi, M. Stornaiuolo, L. Sabatino et al., "The pomace extract taurisol protects rat brain from ischemia-reperfusion injury," *Frontiers in Cellular Neuroscience*, vol. 14, p. 3, 2020.

[19] I. Gomez-Monterrey, P. Campiglia, C. Aquino et al., "Design, synthesis, and cytotoxic evaluation of acyl derivatives of 3-amnionaphtho[2,3-b]thiophene-4,9-dione, a quinone-based system," *Journal of Medicinal Chemistry*, vol. 54, no. 12, pp. 4077–4091, 2011.

[20] C. G. Tenore, E. Pagano, S. Lama et al., "Intestinal anti-inflammatory effect of a peptide derived from gastrointestinal digestion of buffalo (Bubalus bubalis) mozzarella cheese," *Nutrients*, vol. 11, no. 3, p. 610, 2019.

[21] G. C. Tenore, M. Manfra, P. Stiussi et al., "Antioxidant profile and in vitro cardiac radical-scavenging versus pro-oxidant effects of commercial red grape juices (Vitis vinifera L. cv. Aglianico N.)," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 38, pp. 9680–9687, 2012.

[22] D. Vanacore, G. Messina, S. Lama et al., "Effect of restriction vegan diet's on muscle mass, oxidative status, and myocytes differentiation: a pilot study," *Journal of Cellular Physiology*, vol. 233, no. 12, pp. 9345–9353, 2018.

[23] J. Miao, A. V. Ling, P. V. Manthena et al., "Flavin-containing monoxygenase 3 as a potential player in diabetes-associated atherosclerosis," *Nature Communications*, vol. 6, no. 1, article 6498, 2015.

[24] D. M. Shih, Z. Wang, R. Lee et al., "Flavin containing monoxygenase 3 exerts broad effects on glucose and lipid metabolism and atherosclerosis," *Journal of Lipid Research*, vol. 56, no. 1, pp. 22–37, 2015.

[25] L. Santucci, A. Mencarelli, B. Renga et al., "Nitric oxide modulates proapoptotic and antiapoptotic properties of chemotherapy agents: the case of NO-pegylated epirubin," *The FASEB Journal*, vol. 20, no. 6, pp. 765–767, 2006.

[26] D. Ronchetti, V. Borghi, G. Gaitan, J. F. Herrero, and F. Impagnatiello, "NCX 2057, a novel NO-releasing derivative of ferulic acid, suppresses inflammatory and nociceptive responses in in vitro and in vivo models," *British Journal of Pharmacology*, vol. 158, no. 2, pp. 569–579, 2009.

[27] A. Salminen, K. Kaarinanta, and A. Kauppinen, "AMPK and HIF signaling pathways regulate both longevity and cancer growth: the good news and the bad news about survival mechanisms," *Biogerontology*, vol. 17, no. 4, pp. 655–680, 2016.

[28] S. Kume and D. Koya, "Autophagy: a novel therapeutic target for diabetic nephropathy," *Diabetes and Metabolism Journal*, vol. 39, no. 6, pp. 451–460, 2015.

[29] S. Cetrullo, S. D’Adamo, B. Tantini, R. M. Borzi, and F. Flamigni, "mTOR, AMPK, and Sirt1: key players in metabolic stress management," *Critical Reviews in Eukaryotic Gene Expression*, vol. 25, no. 1, pp. 59–75, 2015.

[30] E. A. Dunlop and A. R. Tee, "mTOR and autophagy: a dynamic relationship governed by nutrients and energy," *Seminars in Cell & Developmental Biology*, vol. 36, pp. 121–129, 2014.

[31] S. Sridharan, K. Jain, and A. Basu, "Regulation of autophagy by kinases," *Cancers*, vol. 3, no. 2, pp. 2630–2654, 2011.

[32] N. Mizushima and M. Komatsu, "Autophagy: renovation of cells and tissues," *Cell*, vol. 147, no. 4, pp. 728–741, 2011.

[33] K. Zientara-Rytter and S. Subramani, "Autophagic degradation of peroxisomes in mammals," *Biochemical Society Transactions*, vol. 44, no. 2, pp. 431–440, 2016.

[34] M. Lippai and Z. Szatmari, "Autophagy from molecular mechanisms to clinical relevance," *Cell Biology and Toxicology*, vol. 33, no. 2, pp. 145–168, 2017.

[35] B. Wang, Q. Yang, Y. Y. Sun et al., "Resveratrol-enhanced autophagic flux ameliorates myocardial oxidative stress injury in diabetic mice," *Journal of Cellular and Molecular Medicine*, vol. 18, no. 8, pp. 1599–1611, 2014.

[36] X. X. Lv, S. S. Liu, and Z. W. Hu, "Autophagy-inducing natural compounds: treasure resource for developing therapeutics against tissue fibrosis," *Journal of Asian Natural Products Research*, vol. 19, no. 2, pp. 101–108, 2017.

[37] M. C. Sullards, Y. Liu, Y. Chen, and A. H. Merrill Jr., "Analysis of mammalian sphingolipids by liquid chromatography tandem mass spectrometry (LC-MS/MS) and tissue imaging mass spectrometry (TIMS)," *Biochimica et Biophysica Acta*, vol. 1811, no. 11, pp. 838–853, 2011.

[38] L. R. Ballou, C. P. Chao, M. A. Holness, S. C. Barker, and R. Raghow, "Interleukin-1-mediated PGE2 production and sphingomyelin metabolism. Evidence for the regulation of cyclooxygenase gene expression by sphingosine and ceramide," *Journal of Biological Chemistry*, vol. 267, no. 28, pp. 20044–20050, 1992.
[39] S. Spiegel and S. Milstien, “Sphingosine-1-phosphate: signaling inside and out,” *FEBS Letters*, vol. 476, no. 1-2, pp. 55–57, 2000.

[40] N. Kim, J. O. Lee, H. J. Lee et al., “AMPK, a metabolic sensor, is involved in isoeugenol-induced glucose uptake in muscle cells,” *The Journal of Endocrinology*, vol. 228, no. 2, pp. 105–114, 2016.

[41] S. Momtaz, A. Salek-Maghsoodi, A. H. Abdolghaffari et al., “Polyphenols targeting diabetes via the AMP-activated protein kinase pathway; future approach to drug discovery,” *Critical Reviews in Clinical Laboratory Sciences*, vol. 56, no. 7, pp. 472–492, 2019.

[42] F. Vlavcheski, D. Baron, I. A. Vlachogiannis, R. MacPherson, and E. Tsiani, “Carnosol increases skeletal muscle cell glucose uptake via AMPK-dependent GLUT4 glucose transporter translocation,” *International Journal of Molecular Sciences*, vol. 19, no. 5, article 1321, 2018.

[43] D. M. Breen, T. Sanli, A. Giacca, and E. Tsiani, “Stimulation of muscle cell glucose uptake by resveratrol through sirtuins and AMPK,” *Biochemical and Biophysical Research Communications*, vol. 374, no. 1, pp. 117–122, 2008.

[44] E. Conte, E. Fagone, M. Fruciano, E. Gili, M. Iemmolo, and C. Vanchieri, “Anti-inflammatory and antifibrotic effects of resveratrol in the lung,” *Histology and Histopathology*, vol. 30, no. 5, pp. 523–529, 2015.

[45] M. Suwalsky, F. Villena, and M. J. Gallardo, “In vitro protective effects of resveratrol against oxidative damage in human erythrocytes,” *Biochimica et Biophysica Acta (BBA) - Biomembranes*, vol. 1848, no. 1, pp. 76–82, 2015.

[46] F. C. Huang, H. C. Kuo, Y. H. Huang, H. R. Yu, S. C. Li, and H. C. Kuo, “Anti-inflammatory effect of resveratrol in human coronary arterial endothelial cells via induction of autophagy: implication for the treatment of Kawasaki disease,” *BMC Pharmacology and Toxicology*, vol. 18, no. 1, p. 3, 2017.

[47] G. J. Shi, Y. Li, Q. H. Cao et al., “In vitro and in vivo evidence that quercetin protects against diabetes and its complications: a systematic review of the literature,” *Biomedicine & Pharmacotherapy*, vol. 109, pp. 1085–1099, 2019.

[48] G. Annunziata, M. Jimenez-Garcia, X. Capo et al., “Microencapsulation as a tool to counteract the typical low bioavailability of polyphenols in the management of diabetes,” *Food and Chemical Toxicology*, vol. 139, article 111248, 2020.

[49] L. Barrea, G. Annunziata, G. Muscogiuri et al., “Trimethylamine-N-oxide (TMAO) as novel potential biomarker of early predictors of metabolic syndrome,” *Nutrients*, vol. 10, no. 12, article 1971, 2018.

[50] L. Barrea, G. Annunziata, G. Muscogiuri et al., “Trimethylamine N-oxide, Mediterranean diet, and nutrition in healthy, normal-weight adults: also a matter of sex?,” *Nutrition*, vol. 62, pp. 7–17, 2019.

[51] C. Bedia, T. Levade, and P. Codogno, “Regulation of autophagy by sphingolipids,” *Anti-Cancer Agents in Medicinal Chemistry*, vol. 11, no. 9, pp. 844–853, 2011.