Research Article

Phytochemicals in Leaves and Roots of Selected Kenyan Orange Fleshed Sweet potato (OFSP) Varieties

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1. Introduction

Sweet potato ranks seventh among the food crops in the world [1], and is a major contributor of energy and phytochemicals to the human diets, the extent of which depend on varieties and forms of utilization [2, 3]. For instance, the Orange fleshed sweet potato (OFSP) is a promising biofortified crop for sub-Saharan Africa (SSA) with high levels of β-carotene, a provitamin A carotenoid (pVAC) [4]. Biofortified OFSP has been proven to be affordable, convenient, and sustainable food source of pro-vitamin A carotenoids for combating vitamin A deficiency (VAD) in Kenya and other SSA countries [5, 6].

Apart from their high β-carotene content, OFSP varieties are known to have higher levels of other phytochemicals such as flavonoids, phenolics and anthocyanins [7] that may influence the quality and stability of processed products. These phytochemicals are known to enhance human health by acting antagonistically on incidences of cancers and chronic diseases, including cardiovascular disease (CVD), type II diabetes, and impaired cognitive function [8]. Due to their preventive effects against chronic diseases, they are considered as indispensable components in a variety of nutraceutical, pharmaceutical, medicinal and cosmetic applications [9, 10].

The levels of phytochemicals vary among the plant parts being high on leaves or colored roots [9]. Sweet potato leaves are consumed as vegetables in Islands of the Pacific Ocean, Asia, African countries, and to some extent in the United States of America [11]. These leaves contain both phytochemicals and antinutritional factors such as oxalates, tannins, and...
phytates, and the processing methods also influence their levels in food products [12]. Even though attempts have been made to determine phytochemicals content in sweet potato, the major focus has been on β-carotene with limited data on total phytochemicals and antioxidant activity variations among the OFSP varieties currently promoted in Kenya. Therefore, this study evaluated inherent phytochemicals in the leaves and roots of selected OFSP varieties in Kenya.

2. Materials and Methods

2.1. Acquisition of Sweet potato Leaves and Roots Samples. Seven OFSP and two popular white and yellow fleshed varieties namely; Kabode, Vitaa, Kenspot 5, Kenspot 4, SPK031, SPK004, and K/KA/2004/215. Whitesp and Yellowsp were grown at the Kenya Agricultural & Livestock Research Organization-Kakamega following the standard agronomic practices in 2017 season. Leaves and roots samples were harvested at maturity. The yellow and white fleshed sweet potato varieties were included as local check in the study. Important characteristics of test varieties are shown in Table 1. The leaves and roots were harvested, packaged in plastic net bags and gunny bags, respectively, and transported overnight to the International Livestock Research Institute (ILRI) for laboratory analysis.

2.2. Sample Preparation. Approximately 400 g clean sweet potato leaves of each variety was weighed and divided into portions of 100 g. The leaves were transferred to Kraft paper bags, frozen at −20°C for at least 12 hours and freeze dried (Telstar Lyoques-55, Spain). Similarly, seven roots were randomly selected for each variety, washed with tap water and blot dried, peeled, and diced into about 0.25 cm cubes. About 400 g of these cubes was divided into portions of 100 g, placed in Kraft paper bags, and frozen overnight at −20°C before freeze drying. Freeze-dried samples were ground using a warring laboratory electric blender into powder and stored at −20°C until analysis.

2.3. Analytical Methods

2.3.1. Determination of Dry Matter Content. Moisture content of sweet potato leaves and roots was determined by forced air oven drying at 105°C as described by Abong’ et al. [13].

2.3.2. Determination of Vitamin C. Vitamin C in sweet potato leaves and roots was determined by HPLC as described by Fatariah et al. [14] with slight modification. Briefly, 2 g of fresh sample was weighed into 50 mL polypropylene tube and 30 mL of 3% metaphosphoric acid added and homogenized using a ProScientific homogenizer (ProScientific-200, USA). The mixture was sonicated in ultrasonic bath (Jircus BU-9500Z, Japan) for 5 min, vortexed and centrifuged at 845×g for 5 min. The supernatant was filtered using 0.25 µm membrane for HPLC analysis. Sample separation was achieved using a Shimadzu UPLC system. Chromatographic separation was performed on a Shimadzu (Kyoto-Japan) Nexera X2 UPLC system equipped with a Shimadzu SIL-30AC auto-sampler.

| Variety       | Local name | Shape                                | Leaf colour                        | Mean leaf length (cm) | Mean root length (cm) | Mean root diameter (cm) | Root maturity period (month) |
|---------------|------------|--------------------------------------|------------------------------------|-----------------------|-----------------------|--------------------------|-----------------------------|
| Vitaa         | Vitaa      | Deep, five lobed leaf Green           | 6.0 Obovate Purple Orange          | 21                    | 20                    | 4                        |
| Kabode        | Kabode     | Deep, five lobed leaf Green           | 6.5 Elliptic Purple Deep orange    | 23                    | 21                    | 4                        |
| SPK4          | Kakamega-4 | Deep, five lobed leaf Green           | 5.5 Elliptic Purplish red Orange   | 21                    | 19                    | 4                        |
| SPK031        | SPK031     | Deep, narrow five lobed leaf Green    | 5.5 Obovate Purple Orange          | 11                    | 24                    | 4                        |
| Kenspot 5     | Kenspot 5  | Moderate, five lobed leaf Green, purplish veins on the back | 6.0 Elliptic Purplish red Orange | 12                    | 18                    | 5-7                      |
| Kenspot 4     | Kenspot 4  | Reniform shaped, lateral lobed leaf   | Green                              | 15                    | 18                    | 5-7                      |
| Whitesp       | Nyawoo     | Moderate, five lobed leaf Green       | 5.5 Elliptic Purple White          | 25                    | 24                    | 6                        |
| Yellowsp      | Nyamogo    | Deep, five lobed leaf Green           | 6.0 Elliptic Cream Yellow          | 25                    | 23                    | 6                        |
| K/KA/2004/215 | Jankaroti  | Deep, five lobed leaf Yeung leaf-purplish, old leaf-green with purplish colour at the back | 7.0 Elliptic Cream Light orange | 16                    | 23                    | 4                        |

Leaf and root parameters are mean values of 10 representative samples for each variety.
Shimadzu CTO-30A column oven, LC-30 AD pumps, and SPD-M20A Prominance Diode Array Detector. The analytical column used was C18 (Kinetex, 100 m × 3.0 mm, 2.6 µm). The mobile phase composition consisted of 0.3 mM potassium dihydrogen phosphate in 0.35% (v/v) phosphoric acid at a flow rate of 0.2 mL/min at ambient temperature. Injections of 20 µL were performed with a total run time of 12 min. Data were extracted at a wavelength of 242 nm; compound identification was based on matching of the retention times with pure ascorbic acid (Sigma Aldrich). Compound quantitation was carried out through external calibration using peak area method after integration of chromatographic peaks using Shimadzu LabSolutions software.

2.3.3. Extraction of Phenolics and Flavonoids. Total phenolics and flavonoids in freeze dried OFSP roots and leaves were determined through colorimetric assay adopted to be used with a micro-titer plate and reader. Briefly, 0.15 g and 0.25 g of the freeze-dried leaf and root powder, respectively, were weighed into clean propylene tubes before addition of 10 mL of 80% methanol, vortexed (SI-0166, USA), and shaken on a mechanical shaker (Innova 43, USA) at 8°C and an incubation temperature of 25°C for 12 hours. The mixture was centrifuged at 3226×g for 10 min, and the supernatant aliquot was collected to determine the total phenolics and flavonoid contents.

2.3.4. Determination of Total Phenolics. The total phenolic content was determined using a modified Folin-Ciocalletu procedure [15]. Briefly, 20 µL of the sample blank solution (80% methanol), gallic acid standards (10–100 µg/mL) and samples were pipetted into their respective wells in a microtiter plate followed by addition of 100 µL of 10% Folin–Ciocalteu phenol (Sigma Aldrich) reagent with gentle mixing by priming using a multichannel pipette. After 5 min, 80 µL of 7% of sodium carbonate was added and primed gently before the plate was covered using an aluminum foil and the reaction left to incubate at room temperature for 90 min. Absorbance readings were obtained at 725 nm in a microtiter plate spectrophotometer reader (Synergy HT, USA). External standard calibration technique was used to quantify the concentration of total phenolic compounds in mg/100 g of the dry sample as Catechin Equivalent (mg GAE).

2.3.5. Determination of Total Flavonoids. The total flavonoid content was determined using aluminum chloride colorimetric procedure [16]. Briefly, 20 µL catechin standards (10–100 µg/mL) and samples were pipetted into their respective wells in a microtiter plate followed by addition of 80 µL of deionized distilled water and 10 µL of 5% sodium nitrite, and gently mixed by priming. After 5 min, 10 µL of 10% aluminum chloride was added and primed gently before addition of 80 µL of 2 M sodium hydroxide. The plate was covered with aluminum foil and the reaction left to proceed at room temperature for 30 min. Absorbance readings were obtained at 510 nm in a microtiter plate spectrophotometer reader (Synergy HT, USA). External standard calibration technique was used to quantify the concentration of total flavonoids in mg/100 g of the dry sample as Catechin Equivalent (mg CE).

2.3.6. Determination of Carotenoids

(1) Sample Extraction. All sample preparation and sample analysis were conducted under yellow light to protect carotenoids from UV. The carotenoid analysis was performed according to a method described by Muzhingi et al. [17] with some modifications. Briefly, 0.5 g of freeze-dried powdered sample was mixed with 5 mL of absolute methanol and placed in a water bath (SW23GB, Germany) at 70°C for 10 min. The mixture was vortexed for 1 minute and centrifuged at 800×g (Eppendorf, Centrifuge 5810, Germany) for 10 min. Methanol layer was transferred into a 25 mL volumetric flask and subjected to extraction using 5 mL Tetrahydrofuran (THF), vortexed and centrifuged as previously described. Extraction was repeated three more times using 5 mL of THF each time the supernatant layers being transferred into the volumetric flask. Methanol was added to make the final volume to 25 mL before mixing. To each 2 mL of the extract 0.5 mL of methanol, 4 mL of hexane, and 3 mL of HPLC water were added in a 25 mL glass tube. The mixture was vortexed for 1 minute and centrifuged at 800×g for 3 min. The upper phase was transferred into a 15 mL glass tube and dried completely under nitrogen gas using N-Evap machine (Organamation, Model OA-8125, USA) in a water bath set at a maximum of 40°C. The sample was reconstituted by addition of 2 mL of mixture of methanol and tetrahydrofuran (THF): (85:15 v/v) in a tube. The tube was then vortexed and sonicated (Jircus BU-9500Z, Japan) for 30 s before loading to HPLC vials.

(2) Analysis of Specific Carotenoids. Carotenoid analysis was carried out by use of HPLC (Waters 2695, USA) separation module with photo diode detector (Waters 2996, USA) using previously published methods [18]. The carotenoids were separated on a reverse phase C30 column (YMC Wilmington, NC 150×4.6 mm, 3 µm). The mobile phase composition consisted of eluent A being a mixture of methanol, tert-butyl methyl ether and 1.5% ammonium acetate in water (85:12.3, v/v/v) and eluent B being a mixture of methanol, tert-butyl methyl ether and 1% ammonium acetate in the water (8:90:2, v/v/v). A 40 min linear gradient elution programme was used and was set as follows: 0–1 min, 100% A; 1–10 min 100–90% A; 10–22 min 90–45% A; 22–33 min 45–5% A; 33–37 min 5% A; 37–39 min with a linear gradient to 5–100% A; 39–40 min 100% A. The injection volume was 40 µL while oven temperature was set at ambient (25°C) and the carotenoids were monitored at a wavelength of 450 nm. All carotenoids in the samples were identified by comparing peak retention times and absorption spectra with that of known standards.

2.4. Determination of Antioxidant Activity. The total antioxidant activity of sweet potato leaves and roots was determined using 2, 2 diphenyl-1-picrylhydrazyl (DPPH) procedure and the results expressed as Trolox equivalent. A concentration of 0.002% DPPH [19] was adopted with modification. Briefly 0.15 g and 0.25 g of freeze-dried powdered leaves and roots were respectively, weighed into 50 mL polypropylene tube and 10 mL of 80% methanol added before shaking in mechanical shaker for at least 12 hours
(overnight). The mixture was centrifuged at 2588xg for 15 min and supernatant was used for analysis of antioxidant activity. Approximately 50µL of the blank, standards (5–50 µg/mL Trolox) and samples were pipetted into their respective wells in a microtiter plate followed by addition of 50 µL of 0.002% DPPH with gentle mixing by priming using a multichannel pipette. Absorbance reading was obtained at 515 nm in a microtiter plate spectrophotometer reader (Synergy HT, USA) within 10 min. A standard calibration curve of Trolox was used to calculate the concentration of total antioxidant activity in mg per 100 g of the dry sample and expressed as mg of Trolox Equivalent (mg TE).

2.7. Determination of Soluble Oxalates. Soluble oxalate extraction was carried out as per the procedure described by Nguyen and Savage [21], while chromatographic separation was accomplished based on Wang et al. [22] with minor modifications. For soluble oxalates, 0.5 g freeze-dried sample was weighed into polypropylene tube, 20 mL of deionized water was added with the resultant solution shaken for 15 min and centrifuged at 2588xg for 15 min. The supernatant was filtered through 0.45 nm cellulose nitrate filters into HPLC vials. The obtained extract was analyzed by HPLC. Chromatographic separation was performed on a Shimadzu (Kyoto-Japan) Nexera X2 UPLC system equipped with a Shimadzu SIL-30AC autosampler, Shimadzu CTO-30A column oven, LC-30 AD pumps and SPD-M20A Prominence Diode Array Detector. The analytical column used was C18 column (Kinetex, 100 m x 3.0 mm, 2.6 µm). An isocratic gradient elution program was used using 0.02N sulfuric acid as the mobile phase at a flow rate of 0.2 mL/min and oven temperature set at 40°C. Injection volume of 20 µL was used with a run time of 10 min. Oxalic acid standards were prepared for use in identification and quantitation through external calibration.

3. Results and Discussion

3.1. Leaves and Roots Dry Matter Content. Dry matter content varied significantly ($p < 0.05$) among sweet potato varieties and plant part. The roots had higher percent dry matter compared to leaves (Figure 1). The dry matter content ranged 27.21–38.78% and 20.05–25.53% in roots and leaves, respectively. With exception of Kenspot 4, Kenspot 5, and Vitaa...
that had root dry matter contents (above 30%) comparable to local check white and yellow varieties, other OFSP varieties had lower root dry matter contents. The dry matter contents reported in this study were comparable to those reported in a previous study [23]. For the processing purposes, varieties with low dry matter contents are undesirable since they give low yield and absorb more oils when fried products are produced from them [18].

Apart from the agronomic practices and production environments [24], dry matter content in sweet potato is genetically controlled [25–27], and has been shown to have direct influence on the starch content and β-carotene among other important root phytochemicals [28]. The need for breeding OFSP varieties with high dry matter content remains a critical issue in Kenya where most consumers prefer roots of high dry matter content. For instance, Kabode variety that is preferred by farmers and processors as an OFSP of choice in western Kenya had relatively a low dry matter content thus requiring improvement if Kabode has to be promoted for fried products.

Kenya had relatively a low dry matter content thus requiring improvement if Kabode has to be promoted for fried products. However, the variety with low root dry matter content may be suitable for slurry products such as puree.

3.2. Carotenoid Content. Figures 2 and 3 illustrate HPLC chromatograms of major carotenoids and their variations in sweet potato leaves and roots. Total carotenoid content differed significantly ($p < 0.05$) among varieties and plant parts, the leaves indicating significantly ($p < 0.05$) higher values than the roots (Table 2). No carotenoid was detected in the local white variety. Lutein content was the highest (0.11 mg/100 g) in SPK031 roots, zeaxanthin was the highest

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**Table 2: Variations in carotenoids content (mg/100 g dry basis) among nine Kenyan sweet potato varieties.**

| Variety       | Plant part | Lutein | Zeaxanthin | BX | 13ZBC | AllTBC | 9ZBC |
|---------------|------------|--------|------------|----|-------|--------|------|
| K/KA/2004/215 | Roots      | 0.10 ± 0.01 | 0.10 ± 0.07 | 0.24 ± 0.10 | 0.06 ± 0.01 | 3.82 ± 0.07 | 0.06 ± 0.01 |
|               | Leaves     | 36.68 ± 2.74 | 0.75 ± 0.00 | 0.39 ± 0.08 | 2.71 ± 0.24 | 16.93 ± 2.44 | 2.71 ± 0.24 |
| Kabode        | Roots      | 0.02 ± 0.00 | 0.08 ± 0.02 | 0.18 ± 0.06 | 0.06 ± 0.02 | 4.65 ± 0.20 | 0.06 ± 0.08 |
|               | Leaves     | 32.31 ± 0.35 | 0.42 ± 0.18 | 0.28 ± 0.05 | 2.37 ± 0.11 | 14.29 ± 0.30 | 2.37 ± 0.11 |
| Kenspot 4     | Roots      | 0.03 ± 0.00 | 0.02 ± 0.00 | 0.07 ± 0.01 | 0.10 ± 0.04 | 2.64 ± 0.38 | 0.10 ± 0.06 |
|               | Leaves     | 44.66 ± 1.88 | 0.71 ± 0.04 | 0.20 ± 0.01 | 3.19 ± 0.21 | 19.49 ± 0.90 | 3.19 ± 0.21 |
| Kenspot 5     | Roots      | 0.10 ± 0.00 | 0.26 ± 0.02 | 0.46 ± 0.06 | 0.12 ± 0.01 | 9.28 ± 0.10 | 0.12 ± 0.01 |
|               | Leaves     | 28.57 ± 1.07 | 0.60 ± 0.04 | 0.65 ± 0.24 | 2.58 ± 0.21 | 15.85 ± 0.53 | 2.58 ± 0.21 |
| SPK031        | Roots      | 0.02 ± 0.01 | 0.03 ± 0.02 | 0.45 ± 0.18 | 0.39 ± 0.04 | 18.18 ± 3.74 | 0.39 ± 0.04 |
|               | Leaves     | 35.80 ± 2.52 | 0.33 ± 0.01 | 0.15 ± 0.01 | 2.78 ± 0.17 | 16.81 ± 1.38 | 2.78 ± 0.17 |
| SPK4          | Roots      | 0.11 ± 0.01 | 0.24 ± 0.01 | 0.26 ± 0.05 | 0.05 ± 0.01 | 4.43 ± 0.28 | 0.05 ± 0.01 |
|               | Leaves     | 32.35 ± 1.26 | 0.52 ± 0.09 | 0.39 ± 0.04 | 3.32 ± 0.18 | 21.15 ± 1.12 | 3.32 ± 0.18 |
| Vitaa         | Roots      | 0.01 ± 0.01 | 0.07 ± 0.01 | 0.35 ± 0.01 | 0.21 ± 0.02 | 9.86 ± 0.98 | 0.21 ± 0.04 |
|               | Leaves     | 29.69 ± 0.14 | 0.54 ± 0.06 | 0.26 ± 0.02 | 2.24 ± 0.04 | 13.33 ± 0.16 | 2.24 ± 0.04 |
| Whitesp       | Roots      | ND       | ND         | ND         | ND       | ND       | ND       |
|               | Leaves     | 48.28 ± 2.37 | 1.00 ± 0.01 | 0.39 ± 0.02 | 4.43 ± 0.23 | 27.37 ± 1.36 | 4.43 ± 0.23 |
| Yellowsp      | Roots      | 0.07 ± 0.03 | 0.13 ± 0.03 | 0.07 ± 0.01 | 0.03 ± 0.00 | 0.73 ± 0.01 | 0.03 ± 0.00 |
|               | Leaves     | 51.35 ± 4.4 | 3.53 ± 0.33 | 0.60 ± 0.01 | 4.78 ± 0.43 | 28.07 ± 2.93 | 4.78 ± 0.43 |

BX = β-xanthin, 13ZBC = 13 Cis β-carotene, AllTBC = All trans β-carotene, 9ZBC = β-9 Cis β-carotene, ND = Not detected. Results are means of triplicate samples ± standard deviation; values with same letters in the superscript in the same column are not significantly different at $P < 0.05$. 

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**Figure 2:** Carotenoids chromatogram for sweet potato leaves.
were in agreement with general observation of previous leaves while All trans-β-carotene was the most abundant. The lutein β-carotene content ranged 2.24–4.78 mg/100 g, zeaxanthin ranged 0.33–3.53 mg/100 g, β-cryptoxanthin ranged 0.28–0.65 mg/100 g and 13 cis-β-carotene ranged 2.24–4.78 mg/100 g. All trans-β-carotene ranged 13.33–28.07 mg/100 g while 9 cis-β-carotene content ranged 2.24–4.78 mg/100 g.

Lutein was the most abundant carotenoid in sweet potato leaves while All trans-β-carotene was the most abundant carotenoid present in the roots. The findings of this study were in agreement with general observation of previous research [29–31]. Carotenoids contribute towards root and fruit color, attractiveness, and quality parameters as well as play essential biological functions in humans with β-carotene and β-cryptoxanthin being important provitamin A, while lutein and zeaxanthin are natural antioxidants and important for eye health and cognition [32]. The content of β-carotene in the roots is comparable to 8.65 mg/100 g reported by Odongo et al. [33], 5.9–12.8 mg/100 g reported by Vimala et al. [34], and 0.38–7.38 mg/100 g reported by Alam et al. [35]. The β-carotene content in the leaves were, however, lower compared to an average of 53.32 mg/100 g for Tanzanian sweet potato varieties [36]. The lutein content was higher compared to the range of 19.01–28.85 mg/100 g reported in the same study. Carotenoids content in plants is influenced by the variety, plant part, and environmental factors.
Variations in ascorbic acid, flavonoids and phenolic contents in leaves and roots of nine Kenyan sweet potato varieties are presented in Table 3. Vitamin C varied significantly ($p < 0.05$) among the sweet potato varieties regardless of the plant part. The leaves showed significantly ($p < 0.05$) higher vitamin C levels compared to the roots. The vitamin in roots ranged from 4.53 (Vitaa) to 19.05 mg/100 g (K/KA/2004/205) while in the leaves it ranged from 46.64 (Kenspot 5) to 349.05 mg/100 g (Vitaa).

Flavonoid and phenolic compounds differed significantly ($p < 0.05$) among varieties and with plant part, being higher in leaves than in roots. Flavonoids in roots ranged from not detectable (white fleshed) to 25.8 mg CE/100 g (K/KA/2004/215) while in the leaves it ranged from 4097 (SPK4) to 7316 mg CE/100 g (Kenspot 5). Phenolic content was not detected in white roots but was highest (224 mg GAE/100 g) in SPK031 roots. In the leaves, phenolics content ranged from 4496 to 6801 mg GAE/100 g in SPK031 and Kenspot 5, respectively. Antioxidant activity was significantly ($p < 0.05$) higher in leaves than in roots. It was the lowest (3827.3 mgTE/100 g) in K/KA/2004/215 and the highest (4707.6 mgTE/100 g) in SPK31 leaves while in the roots it ranged from 13.56 to 76.6 mgTE/100 g in SPK4 and K/KA/2004/215, respectively.

The vitamin C content in roots found in this study was comparable to 10 mg/100 g reported by [37], but was lower by genetics and cultural practices and hence vary between locations [5, 24].

3.3. Ascorbic Acid, Flavonoids, Total Phenolic Content and Antioxidant Activity. Variations in ascorbic acid, flavonoids and phenolic contents in leaves and roots of nine Kenyan sweet potato varieties are presented in Table 3. Vitamin C varied significantly ($p < 0.05$) among the sweet potato varieties regardless of the plant part. The leaves showed significantly ($p < 0.05$) higher vitamin C levels compared to the roots. The vitamin in roots ranged from 4.53 (Vitaa) to 19.05 mg/100 g (K/KA/2004/205) while in the leaves it ranged from 46.64 (Kenspot 5) to 349.05 mg/100 g (Vitaa).

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**Table 4: Pearson correlation ($r$) between phytochemicals and antioxidant property.**

| Parameter          | Ascorbic acid | Flavonoids | Phenolics | Total carotenoids | β-Carotene |
|--------------------|---------------|------------|-----------|-------------------|------------|
| Antioxidant property | 0.931         | 0.964      | 0.975     | 0.923             | 0.831      |
| p value            | <0.0001       | <0.0001    | <0.0001   | <0.0001           | <0.0001    |

**Table 5: Tannins and soluble oxalates (dry basis) in leaves and roots of nine Kenyan sweet potato varieties.**

| Variety          | Plant part | Oxalates (mg/100 g) | Tannins (g/100 g) |
|------------------|------------|---------------------|------------------|
| K/KA/2004/215    | Leaves     | 853.83 ± 26.55      | 5.05 ± 0.05      |
|                  | Roots      | 130.58 ± 4.88       | 0.09 ± 0.01      |
| Kenspot 5        | Leaves     | 687.93 ± 59.77      | 4.22 ± 0.33      |
|                  | Roots      | 152.52 ± 23.47      | 0.07 ± 0.00      |
| SPK004           | Leaves     | 827.34 ± 52.59      | 4.51 ± 0.12      |
|                  | Roots      | 87.21 ± 1.22        | 0.05 ± 0.01      |
| Vitaa            | Leaves     | 657.38 ± 112.72     | 4.53 ± 0.17      |
|                  | Roots      | 25.58 ± 1.45        | 0.08 ± 0.01      |
| WhiteSP          | Leaves     | 796.87 ± 9.73       | 3.84 ± 0.23      |
|                  | Roots      | 98.62 ± 13.43       | 0.04 ± 0.00      |
| Kabode           | Leaves     | 1369.09 ± 81.47     | 0.87 ± 0.11      |
|                  | Roots      | 793.31 ± 40.03      | 0.10 ± 0.01      |
| Kenspot 4        | Leaves     | 511.62 ± 20.54      | 4.00 ± 0.39      |
|                  | Roots      | 235.01 ± 48.73      | 0.04 ± 0.00      |
| SPK031           | Leaves     | 741.34 ± 87.07      | 3.57 ± 0.53      |
|                  | Roots      | 180.98 ± 10.96      | 0.13 ± 0.02      |
| Yellowsp         | Leaves     | 1618.71 ± 42.39     | 2.91 ± 0.14      |
|                  | Roots      | 122.83 ± 6.69       | 0.07 ± 0.00      |

Results are means of triplicate samples ± standard deviation; values with similar letters in the same column are not significantly different at $P \leq 0.05$.
Phytates in plant foods vary with variety and plant part due to different genetic and physiological make up. Contrary to the findings of Dako et al. [51] who indicated that yellow varieties had higher phytates compared to white fleshed and orange fleshed sweet potato, in the present study yellow varieties contained phytate in moderate to lowest values. The current phytate values are slightly higher compared to their average of 0.05–0.08 g/100 g, probably due to the larger number of varieties included in this study. Lower values were also reported by Olapade and Ogunade [52] and Abubakar et al. [53]. These values especially in the leaves were, however, comparable to those (2.81–3.01 g/100 g) reported for cereals and other vegetables [54]. Phytic acid binds phosphorus in the food matrix. The findings of the present study showed high accumulation of phosphorus being linked to high phytate content, which means that high phytate varieties may also provide good amounts of phosphorus should processing mechanism significantly reduce phytates. The ratio of phytic phosphorus to total phosphorus in leaves was moderate (15–25%) and in agreement with values of 21–25 reported by Ravindra et al. [55]. However, phytic ratio of the roots had very high range (4–40%) and contrasted these earlier findings. The root phytic ratio was still lower than what Ravindran [55] reported for cereals and legumes, 60–70%, indicating that roots phytates accumulate more phosphorus.

3.4.2. Variations in Tannins and Soluble Oxalates.

Tannins varied significantly (p < 0.05) with variety and plant part being higher in leaves (40 times) than in roots (Table 5). Tannin contents in leaves ranged from 0.87 (Kabode) to 5.05 g/100 g (K/KA/2004/205), while in roots it ranged from 0.003 (Whitesp) to 0.132 g/100 g (SPK031). Root tannin values were not different from the average value (0.03 g/100 g) reported for yellow fleshed varieties by other researchers [51]. It is noted that tannin levels reported in this study are quite lower than levels (779–994%) reported in flours of sweet potato grain mixes [56]. Leaves’ tannin content in this study was comparable to previously reported range of 2.28–4.46 g/100 g [57]. Tannins are complex plant metabolites that form part of polyphenols with considerably good medicinal properties. Tannins can, however, be regarded as antinutrients that bind essential minerals such as iron and significantly reduce their availability and hence the need to minimize them in most foods intended for mineral supplementation [20, 52, 58].

Oxalates were significantly (p < 0.05) higher in leaves than in roots. Within the plant portions, oxalates significantly (p < 0.05) varied with variety. Highest oxalate level (1618.7 mg/100 g) was recorded in leaves of Yellowsp varieties while the lowest (511.62 mg/100 g) was recorded in Kenspot 4. Oxalates in roots ranged from 25.58 to 235 mg/100 g in Vitaa and Kenspot 4, respectively. These values were extremely higher than the range of 5–12 mg/100 g reported earlier by Dako and others [51] and Olapade and Ogunade [52]. Roots values were, however, comparable to 126.9–178.3 mg/100 g range reported in an earlier study [53]. Leaves of all sweet potato varieties may not be suitable for frequent human consumption especially for those with kidney stones problems due to the high levels of oxalates exhibited in these varieties [51]. Oxalates bind calcium and magnesium and interfere with their absorption and metabolism hence the need to limit...
dietary intake. For proper utilization of leaves, therefore, appropriate processing mechanism is needed to reduce the tannins and oxalates.

4. Conclusion

For the first time, this study documents the phytochemical profiles of released Kenyan sweet potato varieties, as well as high variations among varieties for inherent phytochemicals. The leaves were superior in all aspects of phytochemicals that were evaluated. The information generated from this study is useful for ration formulations and dietary recommendations. Study on effects of processing methods on these phytochemicals would, however, give a better picture of the actual amounts being ingested by consumers utilizing different OFSP products.

Data Availability

The data used to support the findings of this study are available from the corresponding authors on request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Conceptualization, George Abong; Sita Ghimire and Tawanda Mzhingi, Michael Okoth; Methodology development, George Abong, Michael Okoth, Fredrick Ng'ang'a, Phyllis Ochieng, Daniel Mbogo, Derick Malavi; Validation and formal analysis, George Abong, Tawanda Mzhingi, Michael Okoth, Fredrick Ng'ang'a, Phyllis Ochieng, Daniel Mbogo, Derick Malavi; Investigation, George Abong, Sita Ghimire, Tawanda Mzhingi and Michael Akhwale; Writing—Original Draft Preparation, George Abong; Writing—Review & Editing, All the authors. Sita Ghimire, Tawanda Muzhingi and Michael Okoth supervised the research work.

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