Pax6-dependent, but β-catenin-independent, function of Bcl9 proteins in mouse lens development

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Bcl9 and Bcl9l (Bcl9/9l) encode Wnt signaling components that mediate the interaction between β-catenin and Pygopus (Pygo) via two evolutionarily conserved domains, HD1 and HD2, respectively. We generated mouse strains lacking these domains to probe the β-catenin-dependent and β-catenin-independent roles of Bcl9/9l and Pygo during mouse development. While lens development is critically dependent on the presence of the HD1 domain, it is not affected by the lack of the HD2 domain, indicating that Bcl9/9l act in this context in a β-catenin-independent manner. Furthermore, we uncover a new regulatory circuit in which Pax6, the master regulator of eye development, directly activates Bcl9/9l transcription.

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Pygopus (Pygo) and Legless (Lgs) were discovered as dedicated Wnt signaling components in Drosophila (Kramps et al. 2002, Parker et al. 2002, Thompson et al. 2002) and Xenopus (Belenkaya et al. 2002). In Drosophila, pygo and lgs are classified as segment polarity genes, and mutations in their coding sequences lead to a dramatic developmental arrest, reminiscent of wingless mutations (Kramps et al. 2002, Parker et al. 2002, Thompson et al. 2002). They are considered to serve as dedicated and essential β-catenin transcriptional coactivators. Lgs simultaneously binds Pygo and Arm/β-catenin via two evolutionarily conserved homology domains, HD1 and HD2, respectively (Kramps et al. 2002). The prevailing model is that Pan/TCF > Arm/β-catenin > Lgs > Pygo serially recruit each other to the DNA in order to efficiently activate Wnt target gene expression, forming a “chain of adaptors.” In this model, the sole function of Lgs is to recruit Pygo to β-catenin (Städeli and Basler 2005).

Vertebrates feature two pygo genes, PYGO1 and PYGO2, and two lgs homologs, BCL9 and BCL9l (BCL9/9l). In contrast to β-catenin loss-of-function mutants, Pygo1/Pygo2 double-knockout mice proceed normally throughout gastrulation and die later during embryonic development from a series of tissue-specific defects. Pygo1 knockout mice are viable and fertile, with no apparent phenotype. Compound Pygo1/Pygo2 knockout mutants are indistinguishable from Pygo2 knockout, suggesting that Pygo2 plays the more important role during development (Li et al. 2007, Schwab et al. 2007, M. A. Aguet, unpubl.). Unlike in Drosophila, in mice, Pygo1 and Pygo2 are now considered to be tissue-specific Wnt pathway components.

Bcl9/9l are necessary for the expression of Wnt targets in normal intestinal epithelia (Deka et al. 2010) and the myogenic progenitors during muscle regeneration, where a reduction of Bcl9/9l inhibits the Wnt-driven myogenic differentiation (Brack et al. 2009). This indicates that Bcl9/9l are required for the expression of key subsets of β-catenin target genes. Consistent with this, mutating β-catenin such that it cannot bind Bcl9/9l [β-catenin-D164A] results in embryonic lethality at mid-gestation, around embryonic day 10.5 (Valenta et al. 2011). In addition, a growing body of evidence indicates that Bcl9/9l and their interaction with β-catenin are important not only for normal cell functions but also in different types of tumors (Mani et al. 2009, Deka et al. 2010, Brembeck et al. 2011, Takada et al. 2012). When their interaction is abrogated in these tumors, an attenuated migratory potential is observed, accompanied by a decrease in the expression of the Lgr5-associated stem cell genes (de la Roche et al. 2012, Kawamoto et al. 2012, A. Moor, P. Anderle, C. Cantu, N. Wiedemann, F. Baruthio, P. Rodriguez, J. Deka, S. André, T. Valenta, B. Gyorffy, et al., in prep.). The conclusion from these studies was that, in mammals, Bcl9/9l retains a critical global function for β-catenin-mediated target gene control, at least in a tissue-specific manner. Based on comparison with the Pygo2 knockout situation, this role can be considered, at least in part, Pygo-independent. However, the molecular function of Bcl9/9l during mammalian development and tissue homeostasis remains largely unexplored.

Here we present an analysis of a series of Bcl9/9l knockout mouse strains in which, via the deletion of the HD1 or HD2 domains, Bcl9/9l lose the ability to bind Pygo or β-catenin, respectively. We found that both interactions are relevant for development, and their individual disruption leads to embryonic lethality. Unexpectedly, however, we found that Bcl9/9l contribute in a Pygo-dependent, but β-catenin-independent, fashion to eye lens formation, revealing for the first time that Bcl9/9l have functions that are separate from canonical Wnt signaling. Moreover, we provide molecular evidence for a novel genetic circuit containing Bcl9/9l and Pygo that does not involve β-catenin: Pax6, the master regulator of eye differentiation, directly activates Bcl9 and Bcl9l transcription. Pygo2 is required for the function of Bcl9/9l, and the Bcl9/9l–Pygo2 complex appears to sustain Pax6 expression. This reveals that, independently of canonical Wnt signaling, Bcl9/9l and Pygo2 constitute a critical molecular unit that responds to tissue-specific regulators, such as, in the lens, the Pax6 transcription factor.
Results and Discussion

Pygo2 is assumed to be recruited by Bcl9/9l to promote the output of the Wnt signaling pathway. However, clear evidence that the Bcl9/9l–Pygo2 interaction is relevant during mammalian development is lacking. To specifically investigate this, we generated constitutive knock-in Bc19 and Bc19l alleles that carry an in-frame deletion of the conserved HD1 [Fig. 1A; Supplemental Fig. S1], the domain responsible for binding to Pygo proteins [Kramps et al. 2002; Städeli and Basler 2005; Fiedler et al. 2008]. This deletion abrogates the recruitment of Pygo proteins to the β-catenin transcriptional complex [Fig. 1A; Mosimann et al. 2009]. Indeed, via GST pull-down assays, we confirmed that a Bc19 protein lacking the HD1, when incubated with total protein extracts obtained from 12.5-dpc postcoitum [dpc] embryos, fails to bind Pygo2 but still binds β-catenin [Fig. 1B], supporting the validity of the rationale and indicating that a deletion of this domain does not lead to improper folding of the protein. Pygo1 was not detectable in our experiments, consistent with the previous finding that it is only weakly expressed later during embryonic development [Li et al. 2004]. For this reason, in our study, we only considered the contribution of Pygo2.

Double-heterozygous knock-in mice (Bc19HD1/+; Bc19lHD1/+; Bc19HD1/HD1) develop normally and do not display any obvious defect. Interestingly, mice homozygous mutant only in either Bc19 or Bc19l [that is, Bc19HD1/HD1, Bc19lHD1/HD1 and Bc19lHD1/HD1] are born in ~50% of the expected Mendelian ratios. This indicates a substantial redundancy between Bc19 and Bc19l, at least concerning the functions mediated by the HD1. In contrast, no double-homozygous mutants (Bc19HD1/HD1; Bc19lHD1/HD1) were ever found later than 13.5 dpc [Fig. 1D], indicating embryonic lethality at this stage. Throughout the study, to simplify the analysis and strengthen the validity of the conclusion drawn, we investigate the effects of the simultaneous deletion in both genes and refer to these mice as Bc19/9l–HD1.

Of note, the complete loss of Bc19/9l (Bc19/9l knockout) causes an earlier lethality, occurring between 9.5 and 10.5 dpc [Supplemental Fig. S2; M Aguet, unpubl.], this suggests the interesting notion that, between 9.5 and 13.5 dpc, Bc19/9l have Pygo-independent functions. Bc19/9l–HD1 mutant embryos at 13.5 dpc are slightly smaller than the control embryos [Fig. 1C], the cause of the lethality is currently under investigation. Remarkably, the timing of the embryonic lethality of Bc19/9l–HD1 mice is similar to that of Pygo1/Pygo2 knockout embryos [Supplemental Fig. S3]. The time point differs slightly from that previously reported [Li et al. 2007; Schwab et al. 2007]. This minor difference may be due to an influence of the genetic background or a difference between the independently generated genomic manipulations. These results indicate that an important aspect of Bc19/9l function is mediated by the HD1 domain. With the caveat that other, so far unrecognized proteins may also interact with Bc19/9l via the HD1 domain, the results show that the Pygo2–Bc19/9l interaction is required for mouse development as of 13.5 dpc.

An aspect in which the Bc19/9l–HD1 embryos are identical to the Pygo2 knockout ones is that they display an obvious eye defect: The lens is absent [Fig. 2]. Of note, the developing eye is one of the head structures displaying a marked expression of both Bc19 and Bc19l [Supplemental Fig. S4]. The lens defect also often appears in triple-mutant mice, which retain only one wild-type allele of both Bc19 and Bc19l [Supplemental Fig. S4]. The cause of the lethality is currently under investigation. Remarkably, the timing of the embryonic lethality of Bc19/9l–HD1 mice is similar to that of Pygo1/Pygo2 knockout embryos [Supplemental Fig. S3]. The time point differs slightly from that previously reported [Li et al. 2007; Schwab et al. 2007]. This minor difference may be due to an influence of the genetic background or a difference between the independently generated genomic manipulations. These results indicate that an important aspect of Bc19/9l function is mediated by the HD1 domain. With the caveat that other, so far unrecognized proteins may also interact with Bc19/9l via the HD1 domain, the results show that the Pygo2–Bc19/9l interaction is required for mouse development as of 13.5 dpc.

During eye development, the surface head ectoderm thickens to form the lens placode and at 10.5 dpc invaginates to generate the lens pit [Lang 2004]. In Bc19/9l–HD1 embryos, lens development is blocked at this stage: The surface head ectoderm does not form the eye placode, the first morphological sign of lens development [Fig. 2, bottom panels], indicating that the Bc19/9l–Pygo2 complex is required early at the onset of lens formation. Also, in Bc19/9l knockout embryos, lens development is arrested at this stage [Supplemental Fig. S2E].

Interestingly, a possible role of Bc19/9l in lens formation is in contradiction with the notion that Wnt signaling must be turned off during lens development [Smith et al. 2005; Machon et al. 2010]. Therefore, we reasoned...
that Bcl9/9l might not act as Wnt signaling components in this tissue. This hypothesis predicts that, if Bcl9/9l can still bind Pygo2 but not β-catenin, the lens will develop correctly. To assess this possibility, we created a knock-in absence of the HD2 domain, Bcl9/9l can still bind to S5): We and others have previously shown that, in the other words, a truncated “chain of adaptors” lacking Bcl9/9l–Pygo2 complex functions independently of the physical interaction with β-catenin.

In addition, we incorporated the lens-cre transgene, a widely used lens-specific Cre recombinase driver [Ashery-Padan et al. 2000], to generate Bc9/9lHD1/flox, Bc9/9lHD2/flox, lens-creTg/+ [lens-ΔHD1] and Bc9/9lHD2/flox, lens-creTg/+ [lens-ΔHD2] mice. This is in agreement with data from Song et al. [2007], who found that lens-cre-driven Pygo2 deletion does not entirely recapitulate the lens loss induced by constitutive deletion of Pygo2. Importantly, however, lens-ΔHD2 pups have normally shaped lenses and are in all cases indistinguishable from control littermates [Supplemental Fig. S6]. This indicates that the Bc9/9l–Pygo2 complex does not require β-catenin to mediate its role during lens development.

To our knowledge, this is the first report demonstrating that Bc9/9l act outside the canonical Wnt signaling cascade. A growing body of evidence indicates that, in
mice, Pygo2 has tissue-specific, Wnt-independent functions in the testis [Nair et al. 2008; Cantù et al. 2013] and lens [Song et al. 2007]. Also, in Drosophila, Pygo was recently reported to act in a β-catenin-independent manner during heart development [Tang et al. 2013, 2014]. It will be critical to establish whether other Wnt-independent functions of Pygo2 require the interaction with Bcl9/9l and whether these functions are conserved among species.

Our finding that the Bcl9/9l-Pygo2 complex functions independently of β-catenin in eye development implies that, in this tissue, these factors also act independently from upstream canonical Wnt signals. The abrogation of the Bcl9/9l-Pygo2 interaction causes a lens arrest very reminiscent of Pax6 loss in this tissue [Ashery-Padan et al. 2000]. Therefore, we reasoned that Pax6 might lie upstream of Bcl9/9l function. To test whether Pax6 controls Bcl9, Bcl9l, and Pygo2 expression, we exploited the mouse-derived lens cell line αTN4, a widely used model for lens cell differentiation. This cell line expresses Pax6 and its direct transcriptional target, αA-crystallin, encoded by the Cryaa gene [Yang and Cvekl 2005]; importantly, Bcl9/9l and Pygo2 are also expressed (Fig. 4A), making this cell line a suitable model for studying their regulation.

We identified in silico Pax6 consensus sequences between −5 kb and +1 kb of the transcriptional start site (TSS) of Bcl9, Bcl9l, and Pygo2. To determine whether any of these serve as bona fide Pax6-binding sites, we performed chromatin immunoprecipitation (ChIP) experiments. We designed PCR primers to amplify the Pax6-binding sites located within evolutionarily conserved regions (Supplemental Fig. S7). Indicative of Pax6 binding, we found that specific regions upstream of Bcl9 and Bcl9l are enriched in our ChIP assay when using an anti-Pax6 antibody (Fig. 4B). Note that the Cryaa promoter, our positive control, displays a comparable enrichment, while no enrichment is observed in several other genomic regions [e.g., Prm2 promoter] (Fig. 4B). To probe the relevance of Pax6 binding within these regions, we treated αTN4 cells with siRNA against Pax6 (siPax6): Pax6 mRNA levels were reduced to <5% of normal expression, causing a down-regulation of its target αA-crystallin [Fig. 4C]. Of note, Pax6 depletion also resulted in the down-regulation of both Bcl9 and Bcl9l transcripts.

To test whether Pax6 also affected Bcl9/9l expression in vivo, we monitored Bcl9/9l expression in the developing eye in the absence of Pax6 function. To this end, we exploited Small eye (Sey) mice that bear a Pax6-truncating mutation, a good model to investigate Pax6 loss of function in vivo [Hogan et al. 1986; Hill et al. 1991]. Homozygous Pax6Sey/Sey embryos can be distinguished as early as 10.5 dpc by the phenotype that, at this stage, the ectoderm does not invaginate to form a lens pit [Hogan et al. 1986]. In wild-type mice, we found Bcl9l [Fig. 4D] and Bcl9l [Supplemental Fig. S4] proteins to be highly expressed in the invaginating surface head ectoderm, overlapping with the Pax6 expression domain (Callaerts et al. 1997). In Pax6Sey/Sey embryos, the ectodermal Bcl9l expression appears to be strongly reduced compared with wild-type controls (Fig. 4D, cf. top and
Consequently, by activating Bcl9/9l–Pygo2 complex is to sustain Pax6 expression. One possibility is that the main function of the lens differentiation is an interesting line of future investigation. The molecular mechanism by which the Bcl9/9l–Pygo2 complex achieves its tissue-specific function in a positive feedback loop, a very common mechanism in biological systems, instrumental to the development (Fig. 5).

In Pygo2-null mice, Pax6 levels are reduced in the surface head ectoderm at 10.5 dpc, on the other hand, when Pax6 function is lacking, Pygo2 expression is unaffected [Fig. 4F, indicating that Bcl9/9l not only are Pax6 targets but also act together with Pygo2 upstream of Pax6. The effect on Pax6 expression is not observed in the Bcl9/9l-AHD2 developing lens, lending further support to the notion that, in this context, Bcl9/9l act in a β-catenin-independent fashion. The epistatic hierarchy between Bcl9/9l–Pygo2, Pax6, and Bcl9/9l raises the prediction that, if Pygo2 expression was altered in lens cells, Pax6 and Bcl9/9l expression would be affected accordingly. To test this, we treated αTN4 cells with siRNA targeting Pygo2 (siPygo2): Consistently, Pygo2 down-regulation causes a decrease in Pax6 mRNA level and diminished levels of its target genes, Cryaa, Bcl9, and Bcl9l [Fig. 4E]. Taken together, our results reveal the existence of a novel genetic regulatory circuit in which Bcl9/9l–Pygo2 act upstream of Pax6, which in turn activates Bcl9 and Bcl9l. Thus, in a β-catenin-independent manner, a Bcl9/9l–Pygo2 complex sustains correct lens formation during mouse development (Fig. 5).

Bcl9 and Bcl9l, in a manner requiring Pygo2 binding, thus emerge as new downstream effectors of Pax6 during lens development. In this context, their action does not appear to be modulated by upstream canonical Wnt signals but by a tissue-specific transcription factor, Pax6, and the master regulator of the lens-specific differentiation program. The molecular mechanism by which the Bcl9/9l–Pygo2 complex achieves its tissue-specific function in lens differentiation is an interesting line of future investigation. One possibility is that the main function of the Bcl9/9l–Pygo2 complex is to sustain Pax6 expression. Consequently, by activating Bcl9/9l, Pax6 would buttress its own expression, creating a positive feedback loop, a very common mechanism in biological systems, instrumental to produce an all-or-none response to a signal (Brandman et al. 2005).

It is currently widely accepted that, in the lens, the Wnt pathway must be turned off (Smith et al. 2005), and a recent study discovered that this inhibition is mediated at least in part by Pax6 [Machon et al. 2010]. It is therefore tantalizing to speculate that Bcl9/9l and Pygo2 not only act independently from β-catenin but, together with Pax6, are involved in the inhibition of the Wnt-induced genetic program, thus ensuring that lens development proceeds correctly. To test this speculation, a more detailed understanding of the timing and nature of Wnt signaling and its targets in the developing lens is needed. Our results provide a novel circuit and a novel paradigm (a β-catenin-independent Bcl9/9l–Pygo2 module) that need to be integrated into the emergent picture.

Materials and methods

Generation of Bcl9/9l knock-in mouse strain

Knock-in mutants in Bcl9 and Bcl9l were generated by standard techniques (inGenious Targeting Laboratory). Briefly, the targeting vector was electroporated into BA1 (C57BL/6 × 129/SvEv) hybrid embryonic stem cells. After selection with the antibiotic G418, surviving clones were expanded for PCR and Southern blotting analyses to confirm recombinant embryonic stem cell clones. Mouse embryonic stem cells harboring the knock-in allele were microinjected into C57BL/6 blastocysts. Resulting chimeras were bred to wild-type C57BL/6N mice to generate F1 heterozygous offspring. Neo cassette excision was obtained by crossing heterozygous knock-in animals with mice expressing Flp recombinase. All mouse experiments were performed in accordance with Swiss guidelines and approved by the Veterinarian Office of the Kanton of Zurich, Switzerland.

Cell culture and quantitative RT–PCR

The αTN4 cell line was cultured in DMEM supplemented with 10% FBS at 37°C. Total RNA was extracted using TRizol (Invitrogen). Quantitative real-time SYBR Green-based PCR reactions were performed in triplicate and monitored with the ABI Prism 7900HT system (Applied Biosystem). All primers are listed in the Supplemental Material.

ChIP

αTN4 were fixed with 1% formaldehyde for 10 min at room temperature and lysed in a nuclei lysis buffer (1% SDS, 50 mM Tris, 10 mM EDTA, protease inhibitors). Chromatin was sonicated to a size between 200 and 1000 base pairs using a Covaris focused ultrasonicator (10 times for 1 min, 10% duty cycle, 5% intensity, 100 cycles per burst). The antibodies and the primers used are listed in the Supplemental Material.

Immunofluorescence

Previously fixed cryosections were blocked with 10% goat serum and 0.3% Triton-X-100 in PBS and incubated overnight at 4°C with the following antibodies: α-Pax6 (Millipore), α-Bcl9 (Abcam), α-Bcl9l (Abnova), and α-β-catenin (Cl. 14, BD Transduction Laboratories). Slides were then incubated with a fluorescently labeled secondary antibody (Alexa 488 goat anti-mouse or Alexa 555 goat anti-rabbit, 1:500). Nuclei were stained with DAPI (1:1000, Sigma).

Figure 5. Our data indicate that Pax6 is required for the transcription of Bcl9/9l. Bcl9/9l then assemble with Pygo2 to ensure a correct lens development; the lens development is arrested when the Bcl9/9l–Pygo2 interaction is abolished. The complex Bcl9/9l–Pygo2 also lies genetically upstream of Pax6, possibly with the function of sustaining its expression.
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References

Ashery-Padan R, Marquardt T, Zhou X, Gruss P. 2000. Pax6 activity in the lens primordium is required for lens formation and for correct placement of a mouse. *Genes Dev* 14: 2701–2711.

Belenkaya TY, Han C, Standley HJ, Lin X, Houston DW, Heasman J, Lin X. 2002. pygopus encodes a nuclear protein essential for wingless/Wnt signaling. *Development* 129: 4089–4101.

Brack AS, Murphy-Seiler E, Hanifi J, Deka J, Eyckerman S, Keller C, Aguet M, Rando TA. 2009. BCL9 is an essential component of canonical Wnt signaling that mediates the differentiation of myogenic progenitors during muscle regeneration. *Dev Biol* 335: 93–105.

Brandman O, Ferrell JE, Li R, Meyer T. 2005. Interlinked fast and slow positive feedback loops drive reliable cell decisions. *Science* 310: 496–498.

Brembeck FH, Wiese M, Zatula N, Grigoryan T, Dai Y, Fritzmann J, Machon O, Kreslova J, Ruzickova J, Vacik T, Klimova L, Fujimura N, Tronnier J, Kollros P, Halder G, Gehring WJ. 1997. PAX-6 in development and evolution. *Development* 124: 1359–1370.

Callaerts P, Halder G, Gehring WJ. 1997. PAX6 in development and evolution. *Annu Rev Neurosci* 20: 483–532.

Cantu C, Valenta T, Hausmann G, Vilain N, Aguet M, Basler K. 2013. The Pygo2-H3K4me2/3 interaction is dispensable for mouse development and Wnt signaling-dependent transcription. *Development* 140: 2377–2386.

Deka J, Wiedemann A, Nudler P, Murphy-Seiler F, Rulten J, Eyckerman S, Stelte J-C, Andre S, Vilain N, Zilman O, et al. 2010. Bcl9/Bcl3 are critical for Wnt-mediated regulation of stem cell traits in colon epithelium and adenoarcinomas. *Cancer Res* 70: 6619–6628.

de la Roche M, Rutherford TJ, Gupta D, Vrechekse DB, Saxty B, Freund SM, Bienz M. 2012. An intrinsically labile nuclear component of the Wnt signalling pathway. *Nat Commun* 3: 680.

Friedler M, Sánchez-Barrena MJ, Nekrasov M, Mieszczanek J, Rybin V, Müller J, Evans P, Bienz M. 2008. Decoding of methylated histone H3 tail by the Pygo-BCL9 complex is dispensable for mouse development and Wnt signaling-dependent transcription. *Mol Cell* 30: 507–518.

Hill RE, Favor J, Hogan BL, Ton CC, Saunders GF, Hanson IM, Prosser J, Jordan T, Hastie ND, van Heyningen V. 1991. Mouse small eye results from some 2 which affects the differentiation of both lens and nasal placodes in the mouse. *J Embryol Exp Morphol* 97: 115–117.

Kawamoto SA, Coleska A, Ran X, Yi H, Yang C-Y, Wang S. 2012. BCL9-2 promotes early stages of intestinal tumor progression. *BMC Biol* 10: 15.

Kim AN, Müller L-AD, Song N, Taketo MM, Lang RA. 2005. The duality of β-catenin function: a requirement in lens morphogenesis and signaling suppression of lens fate in periocular ectoderm. *Dev Biol* 285: 477–489.

Koike H, Diaz-Griffero F, Pena-Cruz V, Berzagolli M, et al. 2009. BCL9 promotes tumor progression by conferring enhanced proliferative, metastatic, and angiogenic properties to cancer cells. *Cancer Res* 69: 7577–7586.

Kosimann C, Hausmann G, Basler K. 2009. β-Catenin hits chromatin: regulation of Wnt target gene activation. *Nat Rev Mol Cell Biol* 10: 276–286.

Kosimann C, Hausmann G, Basler K. 2009. β-Catenin hits chromatin: regulation of Wnt target gene activation. *Nat Rev Mol Cell Biol* 10: 276–286.

Li B, Mackay DR, Ma J, Dai X. 2004. Cloning and developmental expression of mouse pygopus 2, a putative Wnt signaling component. *Genomics* 84: 398–405.

Li B, Rie C, Teng A, Bilanchone V, Munguia JE, Hu M, Jessen S, Piccolo S, Waterman ML, Dai X. 2007. Developmental phenotypes and reduced Wnt signaling in mice deficient for pygopus 2. S. *Genesis* 45: 318–325.

Mech Dev 122: 1171–1182.

Mellman J, Naganomi I, Sun P, Mishra DP, Rhaume C, Li B, Sassone-Corsi P, Dai X. 2008. Nuclear regulator Pyg02 controls spermiogenesis and histone H3 acetylation. *Dev Biol* 320: 446–455.

Parker DS, Jemison J, Cadigan KM. 2002. Pygopus, a nuclear PHD-finger protein required for Wingless signaling in *Drosophila*. *Development* 129: 2565–2576.

Schwab KR, Patterson LT, Hartman HA, Song N, Lang RA, Lin X, Potter SS. 2007. Pygo1 and Pygo2 roles in Wnt signaling in mammalian kidney development. *BMC Biol* 5: 15.

Smith AN, Müller L-AD, Song N, Taketo MM, Lang RA. 2005. The duality of β-catenin function: a requirement in lens morphogenesis and signaling suppression of lens fate in periocular ectoderm. *Dev Biol* 285: 477–489.

Song N, Schwab KR, Patterson LT, Yamaguchi T, Lin X, Potter SS, Lang RA. 2007. pygopus 2 has a crucial, Wnt pathway-independent function in lens induction. *Development* 134: 1873–1885.

Staedeli R, Basler K. 2005. Dissecting nuclear Wingless signaling: recruitment of the transcriptional co-activator Pygopus by a chain of adaptor proteins. *Mech Dev* 122: 1171–1182.

Takada K, Zhu D, Bird GH, Sukhdeo K, Zhao J-J, Mani M, Lemieux M, Carrasco DE, Ryan J, Horst D, et al. 2012. Targeted disruption of the BCL9/β-catenin complex inhibits oncocigenic Wnt signaling. *Sci Transl Med* 4: 148ra17.

Tang M, Yuan W, Fan X, Liu M, Bodmer R, Occor K, Wu X. 2013. Pygopus maintains heart function in aging *Drosophila* independently of canonical Wnt signaling. *Circ Cardiovasc Genet* 6: 472–480.

Tang M, Yuan W, Bodmer R, Wu X, Occor K. 2014. The role of pygopus in the differentiation of intracardiac valves in *Drosophila*. *Genes Dev* 52: 19–28.

Thompson B, Townsley F, Rosin-Arbesfeld R, Musihi H, Bienz M. 2002. A new nuclear component of the Wnt signalling pathway. *Nat Cell Biol* 4: 367–373.

Valenta T, Gay M, Steiner S, Draganova K, Zemke M, Hoffmans R, Cinelli P, Aguet M, Sommer L, Basler K. 2011. Probing transcription-specific outputs of β-catenin in vivo. *Genes Dev* 25: 2631–2643.

Xie Q, Yang Y, Huang J, Ninkovic J, Walcher T, Wolf L, Vitenzon A, Zheng D, Götz M, Beebe DC, et al. 2013. Pygo6 interactions with chromatin and identification of its novel direct target genes in lens and forebrain. *PloS ONE* 8: e64507.

Yang Y, Cvekl A. 2005. Tissue-specific regulation of the mouse α-crystallin gene in lens via recruitment of Pax6 and c-Maf to its promoter. *J Mol Biol* 351: 453–469.

Mani M, Carrasco DE, Zhang Y, Takada K, Gatt ME, Dutta-Simmons J, Ikeda H, Diaz-Griffero F, Pena-Cruz V, Bertagnolli M, et al. 2009.