Detection of human mammaglobin mRNA in breast cancer cells among Vietnamese women

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Background: Detecting circulating tumor cells (CTCs) is a promising approach for evaluating the progression of occult metastases as well as the efficacy of treatment therapies among patients with breast cancer. A real-time reverse transcriptase polymerase chain reaction (RT-PCR) technique has been proposed for detecting CTCs due to its high sensitivity. In this study, we aimed to validate the RT-PCR technique for human mammaglobin (hMAM) mRNA detection among Vietnamese women with breast cancer.

Patients and methods: Peripheral blood samples and breast cancer tissues from 43 patients suffering from breast cancer and 21 patients with fibroids were obtained. Real-time RT-PCR and gene sequencing techniques were employed to detect hMAM gene in CTCs of breast cancer cell lines and cancer tissues.

Results: hMAM mRNA transcription was detected in 36 out of 43 (83.7%) breast cancer tissues and in blood of 23 out of 43 (53.5%) breast cancer patients, while it was detected in only 9.5% out of tissues and 0% of the blood of fibrosis patients. hMAM mRNA in the peripheral blood of breast cancer patients increased with tumor size, stage of cancer and distant metastasis (P<0.05). No difference in the expression of hMAM mRNA was found in breast cancer tissue regarding age, distant metastasis, lymph node, stages of cancer and histopathology (P>0.05).

Conclusion: The study highlighted the expression of hMAM mRNA in breast cancer cells and tissues. This reveals the overall picture of the replication of hMAM mRNA in breast cancer, suggesting the role of hMAM mRNA in breast cancer molecular pathology.

Keywords: human mammaglobin mRNA, hMAM mRNA, breast cancer, circulating tumor cell, CTC, women, Vietnam

Introduction
Breast cancer is a common health issue among women in Vietnam. A recent report estimates that the age-standardized incidence rate of breast cancer in Vietnamese women increased significantly from 13.8 to 29.9 per 100,000 women between 2000 and 2010, which means that there would be 12,533 new breast cancer cases annually nationwide.¹ Early screening, detection and treatment are thus vital for the reduction of this disease burden.² Recently, detecting circulating tumor cells (CTCs) has been found to be a promising approach for evaluating the progression of occult metastases as well as the efficacy of treatment therapies among patients with breast cancer.³ CTCs flow in the patients’ peripheral blood stream and are believed to be a mediator of distant metastases of breast cancer to other organs.⁴⁻⁶ Thus, identifying CTCs in the peripheral blood of breast cancer patients can have critical implications in assisting oncologists to determine appropriate therapeutic methods.
A number of methods have been tested to detect CTCs, such as cytogenetics, morphology and flow cytometry, but they have shown limited diagnostic values.7 Recently, a real-time reverse transcriptase polymerase chain reaction (RT-PCR) method has been proposed for the detection of CTCs due to its high sensitivity.8-10 This technique uses specific expression of marker genes for different tissues or tumors in order to detect the existence of these tumor cells.3 It has been previously used to identify circulating breast cancer cells among patients with metastases.10,11

Markers for RT-PCR may vary across clinical settings, of which human mammaglobin (hMAM) mRNA is one of the most specific markers for detecting CTCs via peripheral blood in breast cancer patients.12,13 hMAM is a glycoprotein with 93 amino acids and a member of secretoglobin–uteroglobin family.3,14 Its expression and overexpression are primary in mammary tissues and breast cancer tissues, respectively.1,14 However, to date, there is a lack of data about the expression of hMAM mRNA in the Vietnamese population. Our hypothesis is that there is a significant difference in the expression of hMAM mRNA between breast cancer tissue and fibrosis tissue, as well as between breast cancer tissue and peripheral blood of breast cancer patients. In this study, we aimed to validate the RT-PCR technique for hMAM mRNA among Vietnamese women with breast cancer.

Patients and methods

Study design and sampling

A total of 43 patients with confirmed breast cancer in all stages were recruited at the time of diagnosis at the K Hospital – the largest oncology hospital in Vietnam. They were excluded if they had tumors in other organs, were treated by any therapies and did not accept to participate. In addition, 21 fibroid patients in the same hospital were also invited to participate in the study. A convenient sampling technique was used to recruit patients.

Blood samples

Five milliliters of peripheral blood in both breast cancer patients and fibroid patients was obtained and stored in EDTA. The blood samples were centrifuged at 4,000 rpm for 20 minutes. After that, the plasma was removed and the white blood cells were collected to enrich the cells, including breast cancer cells. Samples were stored at 4°C and processed immediately after being drawn.

Tissue samples

About 20–30 mg of tissue samples were obtained from those patients from whom blood was drawn, after they had a surgery to remove the tumor. Samples were then stored in sterile vials at –80°C until RNA separation.

Breast cancer cells

MCF-7, BT-474, KPL-4 and MDA-MB231 cells were cultured in DMEM with high glucose (Thermo Fisher Scientific, Waltham, MA, USA) +10% FBS (Thermo Fisher Scientific) +1% penicillin/streptomycin (Thermo Fisher Scientific) +2 mM l-glutamine. These cells were counted before the experiment. These cell lines were purchased from Vietnam Academy of Science and Technology. They were counted using the following steps:

- We took 90 µL of cell culture fluid +10 µL of trypan blue which were incubated at room temperature for about 3–4 minutes. Then, we took 10 µL of cells after dyeing with trypan blue and placed them in a Neubauer counting chamber.
- We counted the number of living cells in each 1 mm². Living cells were not stained and the total number of cells was calculated according to the following formula:
  1. Cell concentration in the initial fluid = total number of cells counted (in 1 mm² cells)/count times ×10⁴×
     2. Total cell count = cell concentration in the initial fluid × number of cell fluids (cells)

RT-PCR procedure

Tumor cells in the blood were obtained by centrifuging the blood samples at 4,000 rpm for 20 minutes. Cells in breast cancer tissues were taken after the tumor removal surgery. Samples were stored at 4°C and sent immediately to the laboratory for total RNA extraction. Total RNA from breast cancer cells in the peripheral blood and tissues was processed using the Mini Kit (Qiagen Kit Rneasy; Qiagen, Hilden, Germany). The purity and concentration of total RNA were checked by using RNA absorption spectrum on NanoDrope-1000. The purity depends on the ratio of OD at 260 nm to OD at 280 nm, and the ratio ranging from 1.7 to 2.0 is acceptable for considering purity.

cDNA creation (RT-PCR) was done using the Invitrogen™ SuperScript™ II Reverse Transcriptase with up to 1 pg of total RNA. Because the total RNA samples extracted from the tissues and the blood samples were not similar, it was necessary to adjust the process from total RNA to cDNA in order to ensure that the imports for real-time PCR had equivalent total RNA (about 100 ng). First, we used about 100 ng of total RNA and mixed it with RNase-free water to obtain a volume of 6 µL per tube. One microliter of the reaction mix and 1 µL
or random hexamers were also added and incubated at 65°C for 5 minutes; then, this combination was kept on ice for 1 minute. Finally, 10 µL of buffer and 2 µL of enzyme were added to reach a volume of 20 µL per tube.

cDNA was synthesized in three thermal cycles: 25°C/10 minutes, 50°C/50 minutes, and 85°C/5 minutes. The cDNA product was stored at −20°C according to the manufacturer’s guideline (Thermo Fisher Scientific). RT-PCR was performed with the primers for hMAM, GAPDH. GAPDH is a standard gene. We implemented PCR reaction with GAPDH F/R primer to test the quality of synthesized cDNA. After confirming that good-quality cDNA was synthesized, PCR was conducted with hMAM primer. A standard curve was built by using breast cancer samples with a certain number of cells (2×10⁴). We performed PCR for cDNA, which was synthesized from sample and breast cancer cells. hMAM primers (Table 1) were used to check this process.

**PCR and sequencing**

PCR from cDNA was processed with hMAM and GAPDH primers. PCR product was checked by electrophoresis on 1% agarose gel. Several samples were sequenced to confirm that the cloned genes were hMAM genes.

**Real-time PCR**

Real-time PCR with hMAM primer was conducted using Roche’s SYBR Green kit. Copies obtained from PCR products were computed as follows:

\[
X (\text{g})/\mu\text{L DNA}/[\text{length RNA} \times 2 \times 340] \times 6.022 \times 10^{23} = Y \text{ copies/\muL}
\]

where 340 is the molecular weight of a nucleotide and 6.022×10²³ is the number of molecules in 1 mole of the substrate.

Number of original cDNA = number of copies obtained from PCR/2ⁿ (n is the number of PCR cycles)

Standard curves were built by calculating the number of cDNA in each tube after diluting at the rate of 10/100/1.000/10.000. SYBR Green, the amplified response could be seen through the emitted fluorescence signal, which is attached to the double-stranded DNA from the start to the end of the process, by using a camera system that can track the fluorescence signal in each tube. The results were applied to construct the standard curves and then were used for calculating the cDNA copies.

**Statistical analysis**

Data were analyzed using SPSS 16.0 software (SPSS, Inc., Chicago, IL, USA). Chi-squared test was used to identify the differences in hMAM (+) expression in various groups. A P-value <0.05 was considered statistical significance.

**Ethical consideration**

The study protocol was approved by the Institutional Review Board of the Ministry of Health and Hanoi Medical University (3018/QD-BYT). Informed consent was obtained from all patients before the experiment. This study was conducted in accordance with the Declaration of Helsinki.

**Results**

After cloning the cDNA of hMAM by PCR from four breast cancer cell lines, KPL4, MCF7, BT474 cell lines were positive and the MDA-MB231 cell line was negative (Figure 1) for the presence of the gene.

Standard curves were developed using 20,000 BT474 cell lines. Concentrations of PCR were obtained at ~679 ng. According to the formula, the number of cDNA copies of hMAM generated from the breast cancer cell line BT474 was 10⁷. Based on this standard, it is possible to calculate the number of copies of hMAM mRNA (Figure 2).

Figure 3 shows the sequencing of the PCR products to confirm that the amplified gene sample was a copy of the hMAM gene, which was published in the gene bank.

Standard curves were made by measuring crossing point (CP) values (Figure 4) in six reaction tubes. Tube hMAM cDNA 1× was generated from 20,000 BT474 cells, which was equivalent to 10⁷ copies. At a dilution of 100 times, which equaled to 200 breast cancer cells, the copies were detected.

**Table 1 Sequences of primers used in this study**

| Primer | Sequences | Temperature (°C) | Length (bp) |
|--------|-----------|-----------------|-------------|
| GAPDH F | 5'-CGG AGT CAA CGG ATT TGG TCG TAT-3' | 65.3 | 307 |
| GAPDH R | 5'-AGC CTT CTC CAT GGT GGT GAA GAC-3' | 67 |  |
| hMAM F | 5'-CTC CCA GCA CTG CTA CGC AGG CTC-3' | 67.2 |  |
| hMAM R | 5'-CAC CTC AAC ATT GCT CAG AGT TTC ATC CG-3' | 66.3 | 202 |

**Abbreviation:** hMAM, human mammaglobin mRNA.
with the CP at 33.14 and the number of copies at 893. Less CP value had higher number of cDNA copies.

Table 2 shows that hMAM mRNA transcription was detected in 83.7% of the breast cancer tissues and in blood of 53.5% of breast cancer patients, while it was detected in only 9.5% of the fibrosis tissues and 0% in blood of fibrosis patients.

Table 3 reveals that hMAM mRNA in the peripheral blood of breast cancer patients increased with tumor size, stage of cancer and distant metastasis (P<0.05). We did not find any differences in hMAM mRNA in the breast cancer tissue.

Discussion

In this study, RT-PCR technology allows to investigate low levels of mRNA that cannot be detected by other classical methods such as the northern blot.15 RT-PCR is often considered the most sensitive test for detecting molecular markers of tumors.16 RT-PCR also provides a high specificity with the design of specific genes of interest.

The synthesis of cDNA based on RNA was performed by reverse enzymes. After cDNA synthesis, cDNA quality was tested by the amplification of GAPDH (internal standard) gene with specific primers. GAPDH is present in every cell of the body and is often used to test the quality of cDNA, eliminating false-negative cases due to poor quality of cDNA. After the amplification reaction, if the quality of the cDNA is poor or cDNA is contaminated, the electrophoretic band on agarose will be blurred and other sub-bands will appear. Our results showed that the PCR product with GADPH F/R primers showed a dark light band with the size of about 307 bp, as expected. This demonstrates that RNA quality and cDNA syntheses were successful, and the cDNA produced met the conditions that were required for subsequent hMAM gene replication.

Initial trials were performed on four breast cancer cell lines, MCF-7, KPL-4, BT-474 and MDA-MB231. hMAM was expressed in three out of the four breast cancer cell lines, except MDA-MB231. It has been argued that none of the genes are expressed in all breast cancer cell lines, but the cancer cell line remains the fundamental for cancer research.17,18

hMAM is one of 23 members of the secretoglobin–uteroglobin family. With northern blot and RT-PCR techniques, hMAM was detected in 90% malignant breast cancer tissues, while it was not detected in benign breast tissue or healthy breast tissue or was detectable at very low levels.19 Of 43 breast cancer samples, 36 cases (83.7%) showed hMAM mRNA replication. This result is in line with previous studies. According to Raica et al, using RT-PCR, the detection rate for hMAM in primary breast cancer was 78.7%.20 According to Al Joudi, the detection rate of hMAM mRNA in breast cancer tissue was 90%.19 With a very high detection rate in breast cancer, hMAM mRNA is the perfect marker for breast cancer diagnosis. Detection of hMAM in breast cancer tissue is an important positive finding for the expression of hMAM mRNA in CTCs. Meanwhile, in the non-cancer control group in the study (ie, fibrous tissue), after performing the RT-PCR technique, only two samples (9.5%) showed hMAM mRNA replication, which was much lower than that in the breast cancer patients.

The rate of expression of hMAM mRNA in the breast cancer tissue was very high. Among cases with very small tumor (ie, <2 cm – T1), 90% cases were found to have hMAM mRNA copies. hMAM mRNA copies were found in 87.5% cases among patients in Phase I (very early stage) and 76.7%
of patients with the absence of lymph node metastases. This suggests that *hMAM* replication is present in the tumor tissue at the early stage and the *hMAM* mRNA in tissues may be used for early diagnosis of breast cancer. Results showed no statistically significant difference in the incidence of *hMAM* mRNA expression with tumor size, lymph node metastasis, distant metastasis and stage of disease. Roncella et al, studying 148 breast cancer samples, showed that the incidence of *hMAM* mRNA in breast cancer patients was 63%, 56% and 100% among patients having tumor size of <2, 2–5 and >5 cm, respectively. The authors did not find any differences in the presence of *hMAM* mRNA in age, histopathologic and lymph node status. Al-Joudi et al found that the detection rate of *hMAM* mRNA in breast cancer was not related to tumor size,
marital status, family history of breast cancer and the presence of PR, ER, HER endocrine receptors. Thus, hMAM mRNA was detected at an early stage of breast cancer with high rates. This result implies the potential use of hMAM mRNA as a new marker contributing to the diagnosis of breast cancer.

On comparing the detection rate of hMAM mRNA in the peripheral blood of patients with breast cancer, we found that the expression of hMAM mRNA in the blood was related to the disease stage, tumor size and distant metastasis. Research shows that by detecting hMAM mRNA replication, breast cancer cells in the blood can be found in the early stage when the tumor is small, without clinical metastases. According to Lee et al, the rate of hMAM mRNA detection in peripheral blood in the first and second stages was 23.4% and in the third and fourth stages was 82.9%. The hMAM mRNA expression rate in the blood was related to lymph node metastasis, distant metastasis and endocrine receptors. Nakagawa et al detected CTCs in 29% cases with stage I, 45% cases with stage II and 77% cases with stage III. Grünewald et al demonstrated that hMAM mRNA level was associated with tumor size and lymph node metastases, while no association was found for other genes.

In this study, RT-PCR was conducted with breast cancer cell lines, breast cancer tissues, peripheral blood of breast cancer patients and breast tissues of patients with breast fibroids in order to detect the differences in hMAM mRNA in both groups of patients. On comparing to Gene Bank, we found that hMAM mRNA expression levels in both groups were similar. The similarity was 100% with hMAM gene published in the International GenBank with codes AY893203, AY888136 and U33147. We supposed that the expression of hMAM mRNA found in breast cancer patients is an over-

![Figure 3](image)

**Figure 3** Sequences of hMAM gene in PCR positive.
Abbreviation: hMAM, human mammaglobin mRNA; PCR, polymerase chain reaction.

![Figure 4](image)

**Figure 4** Real-time PCR for hMAM cDNA determined the detection level in BT474 cell lines.
Abbreviation: CP, crossing point; hMAM, human mammaglobin mRNA; PCR, polymerase chain reaction.

| Setting  | Channel | 530 |
|----------|---------|-----|
| Color compensation | Off |
| Program | Cycles |
| Method | Automated (F<sub>max</sub>) |
| Units |

| Results | Inc | Pos | Name | Type | CP | Concentration | Standard |
|---------|-----|-----|------|------|----|---------------|----------|
| 1 | 1 | H2O | Unknown | 26.72 | 8.93E4 | 1.00E5 |
| 2 | 1x | Standard | 29.46 | 1.25E4 | 1.00E4 |
| 3 | 10x | Standard | 33.14 | 8.93E2 | 1.00E3 |
| 4 | 100x | Standard | >40.00 | <8.50E0 | 1.00E2 |
| 5 | 1000x | Standard | >40.00 | <8.50E0 | 1.00E1 |

**Table 2** hMAM mRNA transcripion in the study

| Samples | Samples (n) | hMAM mRNA (+), n/N (%) |
|---------|------------|------------------------|
| Breast cancer tissue | 43 | 36/43 (83.7%) |
| Fibrosis tissue | 21 | 2/21 (9.5%) |
| Peripheral blood in breast cancer patients | 43 | 23/43 (53.5%) |
| Peripheral blood in fibrosis patients | 21 | 0/21 (0%) |

Abbreviation: hMAM, human mammaglobin mRNA.
replication of the mRNA, resulting in increased synthesis of mammaglobin protein, which might be the cause of tumor cell growth. In fact, using RT-PCR, Watson et al did not find hMAM expression in different benign tissues or in tumors that are only found in breast cancer. Therefore, we assumed that the hMAM mRNA abnormalities in women with breast cancer were not inherited, even after they migrated to other societies. However, further studies are needed to understand clearly the mechanism of this gene in next generations in order to develop appropriate prevention strategies.

Standard curves were developed based on the breast cancer cell lines with enhanced gene copying. According to the RT-PCR results, BT474 cell lines detected hMAM mRNA replication, which can be used to construct the curve. After counting the number of specific cells per milliliter, we performed total RNA synthesis and generated cDNA using PCR with hMAM primer. The product mass measurement obtained from the PCR reaction was the amplified hMAM gene. Based on the formula to transform the product mass to the number of copies of the hMAM gene after amplification, we divided it by $2^{12}$ (25 was the number of PCR cycles) to compute the number of copies of the hMAM cDNA extracted from 20,000 BT474 cells at the first stage. At the 100-fold dilution equivalent to 200 breast cancer cells and CP equal to 33, 14,893 copies were identified as the lowest threshold that could detect hMAM replication. According to Zehentner et al, by using RT-PCR technique to mix BT474 breast cancer cell lines with 5 mL of blood from healthy people, the detection threshold was 100 BT474 cells/mL of blood, and the detectable copy threshold was greater than 1,000.

The strengths of this study included the development of a procedure to detect hMAM mRNA from the breast cancer tissue and peripheral blood of breast cancer patients. The results show that hMAM could be detected in the blood at a very early stage. This could be the initial step to monitor treatment of breast cancer along with other markers, actively contributing to the diagnosis and treatment of breast cancer. Meanwhile, small sample size in this study could reduce the representativeness of the population to the entire population of women with breast cancer in Vietnam.

### Conclusion

In summary, the results of the study provide an overall picture of the replication of hMAM mRNA in breast cancer, suggesting the role of hMAM mRNA in breast cancer molecular pathology.

### Table 3 hMAM expression in the breast cancer tissue and blood according to different characteristics

| Characteristics               | Cases (n) | hMAM (+) in breast cancer tissues | hMAM (+) in peripheral blood of breast cancer patients |
|------------------------------|-----------|----------------------------------|-------------------------------------------------------|
|                              |           | n  | %   | P-value | n  | %   | P-value |
| Age (years)                  |           |    |     |         |    |     |         |
| ≤50                          | 21        | 16 | 76.2 | 0.19    | 10 | 47.6 | 0.45    |
| >50                          | 22        | 20 | 90.9 | 0.90    | 13 | 59.1 | 0.45    |
| Tumor size                   |           |    |     |         |    |     |         |
| T1                           | 10        | 9  | 90.0 | 0.50    | 3  | 30.0 | 0.02    |
| T2                           | 17        | 13 | 76.5 | 0.50    | 7  | 41.2 | 0.02    |
| T3 and T4                    | 16        | 14 | 87.5 | 0.50    | 13 | 81.2 | 0.50    |
| Distant metastasis           |           |    |     |         |    |     |         |
| M0                           | 38        | 31 | 81.6 | 0.29    | 18 | 47.4 | 0.03    |
| M1                           | 5         | 5  | 100.0| 0.01    | 5  | 100.0| 0.01    |
| Lymph node                   |           |    |     |         |    |     |         |
| No                           | 13        | 10 | 76.9 | 0.40    | 5  | 38.5 | 0.19    |
| Yes                          | 30        | 26 | 86.7 | 0.01    | 18 | 60.0 | 0.01    |
| Stage of cancer              |           |    |     |         |    |     |         |
| I                            | 8         | 7  | 87.5 | 0.75    | 2  | 25.0 | 0.01    |
| II                           | 19        | 15 | 78.9 | 0.75    | 8  | 42.1 | 0.01    |
| III and IV                   | 16        | 14 | 87.5 | 0.75    | 13 | 81.2 | 0.75    |
| Histopathology type          |           |    |     |         |    |     |         |
| Ductal                       | 29        | 27 | 93.1 | 0.06    | 16 | 55.2 | 0.52    |
| Lobular                      | 8         | 5  | 62.5 | 0.06    | 3  | 37.5 | 0.52    |
| Mucus                        | 6         | 4  | 66.7 | 0.06    | 4  | 66.7 | 0.06    |
| CA15-3 (U/mL)                |           |    |     |         |    |     |         |
| ≤32                          | 33        | 29 | 84.4 | 0.50    | 16 | 48.5 | 0.23    |
| >32                          | 10        | 7  | 70.0 | 0.50    | 7  | 70.0 | 0.50    |

**Abbreviation:** hMAM, human mammaglobin mRNA.
Disclosure
The authors report no conflicts of interest in this work.

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