Genome assembly, annotation, and comparative analysis of the cattail Typha latifolia

Shane D. Widanagama, Joanna R. Freeland, Xinwei Xu, and Aaron B. A. Shafer

1Department of Computer Science, Trent University, Peterborough, ON K9L 0G2, Canada,
2Department of Biology, Trent University, Peterborough, ON K9L 0G2, Canada,
3Department of Ecology, College of Life Sciences, Wuhan University, Wuhan 430072, China, and
4Department of Forensic Sciences, Trent University, DNA Building, 2140 East Bank Drive, Peterborough, ON, K9L 0G2, Canada

*Corresponding author: Department of Forensic Sciences, Trent University, DNA Building, 2140 East Bank Drive, Peterborough, ON, K9L 0G2, Canada. Email: aaronshafer@trentu.ca

Abstract

Cattails (Typha species) comprise a genus of emergent wetland plants with a global distribution. Typha latifolia and Typha angustifolia are two of the most widespread species, and in areas of sympatry can interbreed to produce the hybrid Typha × glauca. In some regions, the relatively high fitness of Typha × glauca allows it to outcompete and displace both parent species, while simultaneously reducing plant and invertebrate biodiversity, and modifying nutrient and water cycling. We generated a high-quality whole-genome assembly of T. latifolia using PacBio long-read and high coverage Illumina sequences that will facilitate evolutionary and ecological studies in this hybrid zone. Genome size was 287 Mb and consisted of 1158 scaffolds, with an N50 of 8.71 Mb; 43.84% of the genome were identified as repetitive elements. The assembly has a BUSCO score of 96.03%, and 27,432 genes and 2700 RNA sequences were putatively identified. Comparative analysis detected over 9000 shared orthologs with related taxa and phylogenomic analysis supporting T. latifolia as a divergent lineage within Poales. This high-quality scaffold-level reference genome will provide a useful resource for future population genomic analyses and improve our understanding of Typha hybrid dynamics.

Keywords: PacBio long-read sequencing; illumina short-read; de novo; Typhaceae; bulrush; broadleaf cattail; hybrids

Introduction

Cattails (Typha spp.) are aquatic macrophytes that are essential components of wetlands around the world (reviewed in Bansal et al. 2019). These macrophytes often dominate ecosystems through a combination of rapid growth, large size, and sexual and asexual reproduction (Yeo 1964; Andrews and Pratt 1978; Miller and Fujii 2010). Cattails are vital to many wetlands where they cycle nutrients, provide habitat, and aid in bioremediation (Grosshans 2014; Svedarsky et al. 2016; Bonanno and Cirelli 2017). However, cattails can also dominate wetlands and in recent decades invasive cattails have been identified in numerous regions, often following anthropogenic changes that have altered water cycles and increased nutrient loads in wetlands (reviewed in Zedler and Kercher 2004; Bansal et al. 2019). Invasive cattails often form monotypic stands with deleterious effects on local plants and animals, wetland water cycling, and biogeochemical cycles (Yeo 1964; Grace and Harrison 1986; Farrer and Goldberg 2009; Lawrence et al. 2016).

Typha latifolia is likely the most widespread cattail species, occurring on every continent except Antarctica (Smith 1987). Typha angustifolia is also widespread throughout the temperate northern hemisphere (Grace and Harrison 1986), including North America where it was likely introduced from Europe several centuries ago (Ciotir et al. 2013, 2017). In some regions of sympatry T. latifolia and T. angustifolia interbreed to produce the hybrid Typha × glauca (Grace and Harrison 1986; Ciotir et al. 2017; Bansal et al. 2019). In regions surrounding the Laurentian Great Lakes and St Lawrence Seaway in North America, Typha × glauca exhibits heterosis (Bunbury-Blanchette et al. 2015; Zapfe and Freeland 2015), and is more abundant than its parental species (Kirk et al. 2011; Freeland et al. 2013; Pieper et al. 2020). In addition to displacing both parental species, invasive Typha × glauca reduces native plant and invertebrate biodiversity (Tuchman et al. 2009; Lawrence et al. 2016), and alters nutrient cycling and community structure in wetlands (Tuchman et al. 2009; Larkin et al. 2012; Geddes et al. 2014; Lishawa et al. 2014; Lawrence et al. 2017).

Although dominant in some regions of North America, Typha × glauca is uncommon in other regions where the parental species are sympatric, including Europe (Ciotir et al. 2017), eastern Canada (Freeland et al. 2013), and China (Zhou et al. 2016). It is not well understood why hybrids are dominant in some regions but not others, although Tissaw et al. (2020) suggested that hybrids may be limited in coastal wetlands because they have difficulty germinating in salt-rich environments. In addition, although advanced-generation and back-crossed hybrids have been experimentally generated (Pieper et al. 2017) and identified in natural populations (e.g., Kirk et al. 2011; Freeland et al. 2013; Pieper et al. 2020), it has not been possible to differentiate...
advanced-generation and backcrossed hybrids based on the small number of species-specific molecular markers that are currently available (Snow et al. 2010; Kirk et al. 2011). Morphology-based assessments are also unreliable due to overlapping phenotypes (Tangen et al. 2021).

A genome-wide suite of SNPs specific to one or the other parent species would greatly facilitate investigations into the dynamics of the Typha × glauca hybrid zone, which is now expanding westwards from the Great Lakes region into the Prairie Pothole Region of Canada and the USA (Tangen et al. 2021). Genome characterization would also facilitate investigations into local adaptation, introgression, and hybrid dynamics; this in turn may inform future management of T. latifolia, which is predicted to experience a dramatically reduced distribution following climate change (Xu et al. 2013), likely to the detriment of wetlands throughout its widespread native distribution. Here, we report the genome assembly, annotation, and analysis of T. latifolia, the first fully sequenced species in the family Typhaceae.

**Materials and methods**

**Sampling and sequencing**

Leaves of a known T. latifolia plant (Figure 1) were taken from an individual grown at Trent University’s (ON, Canada) greenhouse and ground in liquid nitrogen. DNA was immediately extracted using an E.Z.N.A Plant DNA Kit (Omega Bio-Tek, Inc. GA, USA) following the manufacturer’s instructions for frozen material and eluted in a final volume of 100 µl. DNA quality was assessed on a Tapestation (Agilent Technologies, CA, USA) before being sent to The Centre for Applied Genomics, Toronto, ON, for sequencing. Paired-end reads were sequenced on a single lane of an Illumina HiSeqX system (Illumina, Inc. CA, USA). Long-reads (LR) were sequenced on one SMRT HiFi cell using a PacBio Sequel II system (Pacific Biosciences of California, CA, USA).

**Genome assembly**

Adapters and low-quality bases were trimmed from paired-end short reads (SRs) using Trimomatic -v0.36 (Bolger et al. 2014). The optimal k-mer and genome size from the SRs were estimated using Kmergenie -v1.7051 (Chikhi and Medvedev 2014). We then applied a four-step hybrid assembly: (1) SR assemblies were generated using multiple pipelines: ABySS -v2.2.4 (Jackman et al. 2017) with default settings, 32 threads, a 125 GB total memory; SOAPdenovol -v2.49 (Luo et al. 2012) using default settings, 16 threads, 400 GB of total memory; and Platanus -v1.2.4 (Kajitani et al. 2014) using default settings, 32 threads, a 343 GB memory limit, a total memory of 375 GB. We also downsampled the SR data to 20% because really high coverage can be problematic for de Bruijn graphs (Richards and Murali 2013). The SR assembly with highest quality and completeness was selected by calculating contig N50, total contig length as a percentage of estimated genome size, and the number and percentage of contigs longer than 50 Kb. (2) SR contigs were then aligned to raw LR to create scaffolds using DBG2OLC -v2.0160205 (options: k 17, AdaptiveTh 0.001, KmerCovTh 2, MinOverlap 20, RemoveChimera 1) (Ye et al. 2016). (3) LR contigs were assembled using Canu -v2.0 (Koren et al. 2017) using the expected genome size inferred from Kmergenie, a maximum memory of 124 GB, and a maximum of 32 threads. And (4) the hybrid assembly (Step 2) was merged with Canu LR contigs (Step 3) using QuickMerge -v0.3.0 (Chakravorty et al. 2016) following the 2-step strategy described by Solares et al. (2018). Fion -v1.23.0 (Walker et al. 2014) was used to polish the genome by mapping the Illumina SR back to the genome, thereby correcting base errors and small misassemblies. The final assembly was then submitted and assessed for sequencing and assembly artifacts by NCBI.

**Genome evaluation and annotation**

The T. latifolia hybrid genome was assessed for completeness using Benchmarking Universal Single-copy Orthologs (BUSCO) -v3.0.2 (Simão et al. 2015). BUSCOs of the lineage Liliopsida were assessed from OrthoDB release 10 (Waterhouse et al. 2015). We aligned available chloroplast and mitochondrial genomes (Supplementary Table S1) to our assembly using NUCmer -v3.23 (Kurtz et al. 2004) to identify plastid genomes, scaffolds with long matches (>5000 bp) were extracted and further validated on NCBI Blast. Structural and functional annotation was done in the GenSAS -v6.0 annotation pipeline (Humann et al. 2019). We masked interspersed and simple repetitive elements throughout the genome using a database developed through repeat modeler -v2.0.1 (Smit and Hubley 2008–2015) in conjunction with repeat masker -v4.0.7 with the NCBI/rmblast search engine, and quick sensitivity (Smit et al. 2013–2015). The hardmasked version of the genome sequence was used for feature prediction. Available RNA-seq data from T. angustifolia (SRR15541138) were mapped to the genome using hisat2 -v2.1.0 (Kim et al. 2019) and gene prediction with the RNA-seq alignments was performed by the BRAKER2 -v2.1.1 pipeline, which uses Augustus and GeneMark-ET (Lomsadze et al. 2014; Brńu et al. 2021). ncRNA was predicted using tRNAscan-SE -v2.0 (Lowe and Eddy 1997) and Infernal -v1.1.3 (Nawrocki and Eddy 2013). Refinement of the official gene set was performed with an available T. latifolia transcriptome (Moscou 2017) using PASA -v2.3.3 (Haas et al. 2003). Last, our assembly (N50 scaffolds) was aligned and compared to a recent T. latifolia hybrid scaffold assembly (JAAWWQ010000000) using...
Mummer4 (Marçais et al. 2018) requiring a minimum alignment (l) of 10,000 bp.

**Comparative genomics**

Single-copy orthologous genes from five other species were identified using OrthoVenn2 with an e-value cutoff of 1e-5, an inflation value of 1.5, and the annotation, protein similarity network, and cluster relationship network enabled (Xu et al. 2019). OrthoVenn2 performs all-against-all genome-wide protein comparisons and groups genes into clusters with the Markov Clustering Algorithm, where a cluster is made up of orthologs and paralogs (Wang et al. 2015). Here we selected Oryza sativa *Japonica*, Brachypodium distachyon, and Sorghum bicolor from the family Poaceae, and Ananas comosus from the family Bromeliaceae. *Arabidopsis thaliana* from the Brassicaceae family was chosen as the dicot outgroup. All proteomes were from the Ensemble database release 104 (Howe et al. 2021). The identified orthologous sequences were then aligned using MAFFT -v7.741 (Katoh and Standley 2013) and concatenated into single sequences by species using SeqKit -v0.15.0 (Shen et al. 2016). Maximum likelihood phylogenetic analysis was performed using RAxML -v8.2.12 (Stamatakis 2014) with the PROTGAMMAAUTO substitution model (Darriba et al. 2012). The phylogenetic tree divergence times were estimated using MC-MCtree from the PAML package -v4.9j (Yang 1997, 2007), and was calibrated using the divergence time between Sorghum and Oryza (42–52 Mya) (Kumar et al. 2017).

**Results**

**Sequencing data and genome assembly**

A total of 138.6 Gb of raw 151 bp Illumina reads were sequenced (Supplementary Table S2). Quality filtering and trimming removed 6.5 Gb of sequence data. A recommended k-mer size of 101 bp and estimated genome size of 257 Mb was estimated from the short-read data (Supplementary Figure S1). LR data from the Pacbio Sequel II generated 86.8 Gb of raw data: this included 7,244,218 subreads with a mean length of 11,978.2 bp. All sequencing data were deposited in the NCBI Sequence Read Archive (Accession No PRJNA751759). The ABySS assembly that was used 100% of Illumina reads had the most contiguous genome (Supplementary Table S3). This used 100% of Illumina reads had the most contiguous genome (Supplementary Table S3). The ABySS SR assembly had a N50 of 0.011 Mb, 365,565 contigs, and 1,445 contigs longer than 50 Kb. The DBG2OLC assemblies (ABySS contigs + LR contigs) improved the assembly statistics but resulted in a smaller than expected genome (Table 1); here we calculated N50 of 0.132 Mb, 1,845 contigs, and 1,445 contigs longer than 50 Kb (Supplementary Table S4). The LR Canu assembly had an N50 of 8.706 Mb, and contained 1,189 contigs, 821 of which were longer than 50 Kb (95.54%) (Table 1). The final merged and polished genome assembly—DBG2OLC assembly + Canu—was 286.7 Mb with plastids removed. We estimated an N50 of 8.71 Mb, and consisted of 1,158 scaffolds (Table 1). Step 4 resulted in the merger of three scaffolds and the subsequent removal of 30 short scaffolds by NCBI vetting. The GC content was 38.05% (Table 1). The polished genome has been deposited in the NCBI genome database (JAI0KV0000000000). We observed a high degree of synten gradients between our assembly and similar hybrid assembly of T. latifolia (Supplementary Figure S2), although mapping success of un-related individuals was significantly higher in our genome (Supplementary Figure S3).

**Assembly quality and annotation**

Approximately 96.03% of BUSCOs were identified (3148 of 3278) in the assembly. Of these, 3025 BUSCOs were complete (92.28%), and 123 were fragmented. Of the complete BUSCOs, 2461 were single-copies, and 564 had duplicates. Repeats represented 43.84% of the genome (see breakdown in Figure 2). The chloroplast genome was detected and split on two scaffolds, while the entire mitochondrial genome appeared to be assembled (Supplementary Table S5). Total repeat content of the genome is consistent with other Poales genomes (Kawahara et al. 2013, Ming et al. 2015, Redwan et al. 2016): long interspersed nuclear elements (LINEs) comprised 1.22% of the genome, while short interspersed nuclear elements (SINEs) were not detected in the genome (Table 2). The annotation pipeline produced 27,432 genes, which coded for 34,911 proteins and 34,974 mRNA sequences. 2095 rRNA, 502 tRNA, and 214 miRNA sequences were predicted. 27,432 genes, which coded for 34,911 proteins and 34,974 mRNA sequences. All proteomes were from the NCBI vetting. The GC content was 38.05% (Table 1). The polished genome has been deposited in the NCBI genome database (JAI0KV0000000000). We observed a high degree of synten gradients between our assembly and similar hybrid assembly of T. latifolia (Supplementary Figure S2), although mapping success of un-related individuals was significantly higher in our genome (Supplementary Figure S3).

**Comparative genomic analyses**

Ortholog clustering analysis revealed 9168 gene families were shared among T. latifolia, A. comosus, A. thaliana, B. distachyon, O. sativa, and S. bicolor (Figure 2). In total, 1806 gene families were found to be unique to T. latifolia (Figure 2). We aligned 1900 single-copy gene clusters to conduct phylogenomic analysis (Figure 3). The phylogenomic tree generated supports a divergent Typha position in Poales (Figure 3), with Typha forming a separate clade with pineapples (A. comosus). We estimated the bromeliad lineage (Typha-Ananas) to be approximately 70 million years old.

**Table 1** Genome assembly statistics of our four-step hybrid assembly

| Genome size (Mb) | SR contig assembly—step 1 | DBG2OLC contig assembly—step 2 | LR contig assembly—step 3 | Merged polished scaffolds—step 4 |
|------------------|--------------------------|-------------------------------|---------------------------|-------------------------------|
| 350              | 263.74                   | 193.16                        | 287.63                    | 286.77                        |
| N50/L50          | 365,65                   | 1,340                         | 1,190                     | 1,158                         |
| N50/L50          | 11.43 Kb/5,314           | 152.07 Kb/412                 | 8.71 Mb/13                | 8.71 Mb/13                    |
| Max sequence length | 201 bp/193.74            | 52.95 Kb/1,358                | 58.92 Kb/530              | 58.94 Kb/523                  |
| Scaffolds > 10 Kb | 6,048                    | 1,833                         | 1,140                     | 1,132                         |
| Scaffolds > 25 Kb | 1,759                    | 1,785                         | 1,127                     | 1,120                         |
| Scaffolds > 50 Kb | 362                      | 1,445                         | 821                       | 816                           |
| % of scaffolds > 50 Kb | 9.46                   | 92.34                         | 95.54                     | 95.59                         |
| GC content (%)   | 38.40                    | 38.50                         | 38.11                     | 38.05                         |

Short read (SR) contig assembly consists of contigs assembled from 100% of the Illumina reads using the ABySS assembler. The DBG2OLC assembler combined the 100% ABySS contigs and PacBio long reads. The long read (LR) contig assembly was generated from only PacBio long reads using Canu. The merged polished scaffolds are from merging the DBG2OLC assembly and LR contigs and were polished using Pilon.
Discussion

The Typhaceae family (order Poales) is diverse, comprising over 50 recognized species (Christenhusz and Byng 2016). Species in this family are essential components of marshes and wetlands around the world (reviewed in Bansal et al. 2019), and play a role in both bioremediation and the production of biofuel material (Grosshans 2014; Svedarsky et al. 2016; Bonanno and Cirelli 2017). This is the first published whole-genome assembly from a member of the Typhaceae family and adds to the previously characterized chloroplast genome sequences of Sparganium stoloniferum (Guisinger et al. 2020). The assembly and annotation of the Cattail genome. Types of repeats include DNA transposons, long interspersed nuclear elements (LINEs), long terminal repeats (LTRs), unclassified repeats, and simple repeats. (B) Venn diagram of orthologous gene clusters among the broadleaf cattail (Typha latifolia), pineapple (Ananas comosus), thale cress (Arabidopsis thaliana), stiff brome (Brachypodium distachyon), rice (Oryza sativa japonica), and broom-corn (Sorghum bicolor). Only the numbers of ortholog clusters of adjacent species, those common to all species, and those unique to each species are labeled.

Table 2 Summary of repeats masked in the Typha latifolia genome

| Type      | Length (bp) | Percentage of genome (%) |
|-----------|-------------|--------------------------|
| SINE      | 0           | 0                        |
| LINE      | 3,499,426   | 1.22                     |
| LTR elements | 44,172,983 | 15.35                    |
| DNA elements | 3,735,882  | 1.30                     |
| Unclassified | 68,736,281 | 23.88                    |
| Small RNA | 0           | 0                        |
| Satellites| 0           | 0                        |
| Simple repeats | 5,689,134 | 1.98                     |
| Low complexity | 670,871 | 0.23                     |
| Total     | 126,178,695 | 43.84                    |

Discussion

The Typhaceae family (order Poales) is diverse, comprising over 50 recognized species (Christenhusz and Byng 2016). Species in this family are essential components of marshes and wetlands around the world (reviewed in Bansal et al. 2019), and play a role in both bioremediation and the production of biofuel material (Grosshans 2014; Svedarsky et al. 2016; Bonanno and Cirelli 2017). This is the first published whole-genome assembly from a member of the Typhaceae family and adds to the previously characterized chloroplast genome sequences of T. latifolia, T. orientalis, and Sparganium stoloniferum (Guisinger et al. 2010; Su et al. 2019; Liu et al. 2020). The assembly and annotation of the Cattail genome, which we note was a key recommendation from a diverse team of Typha researchers (Bansal et al. 2019), will likely be an important tool in wetland management.

We explored a variety of SR assemblers given the varying quality of outputs seen in Assemblathon metrics (Bradnam et al. 2013). Of the three SR assemblers used, ABYSS produced longer, fewer contigs and scaffolds, which is consistent with previous comparisons (Bradnam et al. 2013). ABYSS’ de Bruijn approach did not have issues with the high coverage Illumina data (e.g., Richards and Murali 2015). We combined with SR assembly with raw long read data using DBG2OLC (Ye et al. 2016) as previous plant genomes with similar data showed promising assembly statistics (i.e., Daccord et al. 2017; Hatakeyama et al. 2018; Zhou et al. 2019). The long reads in the DBG2OLC assembly captured more lengthy and repetitive regions, resulting in a sharp rise in contig N50 and contigs 50 Kb or longer (Table 1). However, the backbone of the final assembly was the PacBio Canu assembly (Table 1 and Supplementary Table S4), with the benefit of this strategy being the removal of chimeras with polished assemblies having very low error rates (Ye et al. 2016).

Accurate genomes are important as sequencing errors can affect both nucleotide diversity and the discovery of markers (Clark and Whitlam 1992). LR sequencing further facilitates detection of structural variants (Amarasinghe et al. 2020), which might be useful in studying Typha dynamics given the link to hybridization and speciation (Weissensteiner et al. 2020). The final assembly consists of only 1158 scaffolds, with ~95% being 50 Kb or longer; T. latifolia possessed higher N50 and fewer scaffolds compared to other plant genome assemblies that used PacBio sequencing and similar hybrid assembly pipelines (Redwan et al. 2016; Hatakeyama et al. 2018; Reuscher et al. 2018). With 96.03% of BUSCOs detected and 92.28% of them complete in our assembly, this would suggest a high-quality annotation. The topology of the phylogenetic tree that we reconstructed based on whole-genome sequences of six species agrees with previously inferred evolutionary relationships (e.g., Darshetkar et al. 2019) by grouping together the two bromeliad species (T. latifolia and A. comosus) and the three graminid species (O. sativa, S. bicolor, and B. distachyon), and by inferring a more recent divergence date for graminids compared to bromeliads. Our estimated Typha lineage age of approximately 70 million years exceeds an earlier estimate of between 22.64 and 57.60 mya that was based on seven cpDNA markers (Zhou et al. 2018), although is comparable to the estimate of 69.5 Myr that was based on a combination of fossil records and cpDNA sequences (Bremer 2000). We acknowledge that the use of whole-genome sequences to infer divergence...
times is still in its infancy, and caution that factors such as rate heterogeneity may complicate such inferences (reviewed in Smith et al. 2018).

The T. latifolia reference genome now allows for studying Typha and hybridization at the molecular level, for example, by providing insight into the adaptation of T. latifolia populations that may be threatened by climate change, and facilitating research into potentially important processes such as hybrid breakdown in invasive Typha hybrids. In addition, genome-wide markers could provide insight into why the hybrid Typha × glauca is dominant in some areas (Kirk et al. 2011; Freeland et al. 2013) but uncommon in others (Freeland et al. 2013; Ciotir et al. 2017). For example, T. angustifolia in North America is thought to have arrived from Europe several centuries ago (Ciotir et al. 2013), and ancestry assessments could test the hypothesis that historical interspecific hybridization led to genetic introgression into some T. angustifolia populations, which may help to explain regional erosion of species barriers. Locus-specific ancestry estimation analyses, markers, and the identification of potentially adaptive genes can all now be used to find evidence of ancient hybridization in T. angustifolia (Goulet et al. 2017; Taylor and Larson 2019). This high-quality draft genome and its comparisons with Poales species will be an indispensable resource for ongoing research into Typha, a genus that both sustains and threatens wetlands around the world.

Data availability

Raw sequencing data can be found in the SR Archive (SRA) under accession number PRJNA751759. Genome assembly has been submitted to GenBank (JAI0KVO00000000). Code used to generate the data can be found at https://gitlab.com/WiDGeT_TrentU/undergrad-theses/-/tree/master/Widanagama_2021. The T. angustifolia RNSseq data are in the SRA under SRR15541138. The T. latifolia transcriptome used can be found at https://figshare.com/articles/dataset/Typha_latifolia_leaf_transcriptome/5661727/1.

Supplementary material is available at G3 online.

Acknowledgments

The authors thank Vikram Bhargav for sampling and preparing the T. latifolia leaves, and Eric Wootton for extracting DNA from the sample.

Funding

The CanSeq 150 project funded the PacBio sequencing: NSERC Discovery grants to J.R.F. and A.B.A.S. and two NSERC Undergraduate Student Research Awards to S.D.W. supported this research. A Compute Canada Resources for Research Groups award to A.B.A.S. and Compute Canada support staff supported the bioinformatics.

Conflicts of interest

The authors declare that there is no conflict of interest.

Literature cited

Amarasinghe SL, Su S, Dong X, Zappia L, Ritchie ME, et al. 2020. Opportunities and challenges in long-read sequencing data analysis. Genome Biol. 21:1–16.

Andrews NJ, Pratt DC. 1978. Energy potential of cattails (Typha spp.) and productivity in managed stands. J Minn Acad Sci. 44:5–8.

Bansal S, Lishawa SC, Newman S, Tangen BA, Wilcox D, et al. 2019. Typha (Cattail) invasion in North American wetlands: biology, regional problems, impacts, ecosystem services, and management. Wetlands. 39:645–684.
macrophyte *Typha orientalis* (Typhaceae). Mitochondrial DNA B Resour. 5:136–137.

Lomsadze A, Burns BD, Borodovsky M. 2014. Integration of mapped RNA-Seq reads into automatic training of eukaryotic gene finding algorithm. Nucleic Acids Res. 42:e119.

Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25:955–964. doi:10.1093/nar/25.5.955.

Luo R, Liu B, Xie Y, Li Z, Huang W, et al. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. Gigascience 1:18.

Marçais G, Delcher AL, Phillippy AM, Coston R, Salzberg SL, et al. 2018. MUMmer4: a fast and versatile genome alignment system. PLoS Comput Biol. 14:e1005944.

Miller RL, Fujii R. 2010. Plant community, primary productivity, and environmental conditions following wetland re-establishment in the Sacramento-San Joaquin Delta, California. Wetlands Ecol Manage. 18:1–16.

Ming R, VanBuren R, Wai CM, Tang H, Schatz MC, et al. 2016. The pineapple genome and the evolution of CAM photosynthesis. Nat Genet. 47:1435–1442.

Moscou M. 2017. *Typha latifolia* Leaf Transcriptome (Version 1). doi: 10.6084/m9.figshare.5661727.v1.

Nawrocki EP, Eddy SR. 2013. Infernal 1.1: 100-fold faster RNA homology searches. Bioinformatics. 29:2933–2935.

Pieper S, Dorken M, Freeland J. 2020. Genetic structure in hybrids and progenitors provides insight into processes underlying an invasive cattail (*Typha × glauca*) hybrid zone. Hereditas (Edinb). 124:714–725.

Pieper SJ, Nicholls AA, Freeland JR, Dorken ME. 2017. Asymmetric hybridization in cattails (*Typha spp.*) and its implications for the evolutionary maintenance of native *Typha latifolia*. J Hered. 108:479–487.

Redwan RM, Saidin A, Kumar SV. 2016. The draft genome of MD-2 pineapple using hybrid error correction of long reads. DNA Res. 23:427–439.

Reuscher S, Furuta T, Bessho-Uehara K, Cosi M, Jena KK, et al. 2018. Assembling the genome of the African wild rice *Oryza longistamnata* by exploiting synteny in closely related *Oryza* species. Commun Biol. 1:1–10.

Richards S, Murali SC. 2015. Best practices in insect genome sequencing: what works and what doesn’t.Curr Opin Insect Sci. 7:1–7. doi:10.1016/j.cois.2015.02.013.

Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics. 31:3210–3212.

Shen W, Le S, Li Y, Hu F. 2016. SeqKit: a cross-platform and ultrafast toolkit for FASTA/Q file manipulation. PLoS One. 11:e0163962.

Smit AFA, Hubley R. 2008–2015. RepeatModeler (Open-1.0). Institute for Systems Biology. http://www.repeatmasker.org.

Smit AFA, Hubley R, Green P. 2013–2015. RepeatMasker (Open-4.0). Institute for Systems Biology. http://www.repeatmasker.org.

Smith SA, Brown JW, Walker JF. 2018. So many genes, so little time: a practical approach to divergence-time estimation in the genomic era. PLoS One. 13:e0197433.

Snow AA, Travis SE, Wildová R, Fér T, Sweeney PM, et al. 2010. Species-specific SSR alleles for studies of hybrid cattails (*Typha latifolia × T. angustifolia*; Typhaceae) in North America. Am J Bot. 97:2061–2067.

Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 30:1312–1313.

Su T, Yang JX, Lin YG, Kang N, Hu GX. 2019. Characterization of the complete chloroplast genome of *Sparganium stoloniferum* (Poales: typhaceae) and phylogenetic analysis. Mitochondrial DNA B Resour. 4:1402–1403.

Taylor SA, Larson EL. 2019. Insights from genomes into the evolutionary importance and prevalence of hybridization in nature. Nat Ecol Evol. 3:170–177.

Tissaw K, Freeland J, Dorken ME. 2020. Salinity, not genetic incompatibilities, limits the establishment of the invasive hybrid cattail *Typha × glauca* in coastal wetlands. Ecol Evol. 10:12091–12103.

Tuchman NC, Larkin DJ, Geddes P, Wildova R, Jankowski K, et al. 2009. Patterns of environmental change associated with *Typha × glauca* invasion in a Great Lakes Coastal Wetland. Wetlands. 29:964–975.

Wang Y, Coleman-Derr D, Chen G, Gu YQ. 2015. OrthoVenn: a web server for genome wide comparison and annotation of orthologous clusters across multiple species. Nucleic Acids Res. 43:W78–W84.

Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, et al. 2014. Pilot: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One. 9:e112963.

Waterhouse RM, Tegenfeldt F, Li J, Zdobnov EM, Kriventseva EV. 2013. OrthoDB: a hierarchical catalog of animal, fungal and bacterial orthologs. Nucleic Acids Res. 41:D358–D365.

Weissensteiner MH, Bunikis I, Catalán A, Francoijs KJ, Kniief U, et al. 2020. Discovery and population genomics of structural variation in a songbird genus. Nat Commun. 11:1–11.

Xu L, Dong Z, Fang L, Luo Y, Wei Z, et al. 2019. OrthoVenn2: a web server for whole-genome comparison and annotation of orthologous clusters across multiple species. Nucleic Acids Res. 47:W52–W58. doi:10.1093/nar/gkz333.

Yang Z. 2007. PAML 4: phylogenetic analysis by maximum likelihood. Comput Appl Biosci. 13:555–556.

Yang Z. 2007. PAML 4: phylogenetic analysis by maximum likelihood. Mol Biol Evol. 24:1586–1591.

Ye C, Hill CM, Wu S, Ruan J, Ma ZS. 2016. DBG2OLC: efficient assembly of large genomes using long erroneous reads of the third generation sequencing technologies. Sci Rep. 6:31900.

Yeo RR. 1964. Life history of common cattail. Weeds. 12:284–288.
Zapfe L, Freeland JR. 2015. Heterosis in invasive F1 cattail hybrids (Typha × glauca). Aquat Bot. 125:44–47.
Zedler JB, Kercher S. 2004. Causes and consequences of invasive plants in wetlands: opportunities, opportunists, and outcomes. Crit Rev Plant Sci. 23:431–452.
Zhou B, Tu T, Kong F, Wen J, Xu X. 2018. Revised phylogeny and historical biogeography of the cosmopolitan aquatic plant genus Typha (Typhaceae). Sci Reports. 8:1–7.
Zhou B, Yu D, Ding Z, Xu X. 2016. Comparison of genetic diversity in four Typha species (Poales, Typhaceae) from China. Hydrobiologia. 770:117–128.
Zhou Y, Minio A, Massonnet M, Solares E, Lv Y, et al. 2019. The population genetics of structural variants in grapevine domestication. Nat Plants. 5:965–979.

Communicating editor: P. Morrell