Cold Plasma: Clean Technology to Destroy Pathogenic Micro-organisms

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Received: 12 May 2020 / Revised: 11 June 2020 / Accepted: 13 June 2020 / Published online: 20 June 2020
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Abstract
Atmospheric pressure cold plasma is a promising technology in fighting pathogenic micro-organisms. In times of Covid-19 pandemic, it was decided to modify two types of cold plasma devices to study their effectiveness in the killing of pathogenic micro-organisms. These studies have shown that both the devices are efficient in this purpose. While pencil like microwave based device can destroy Aeromonas bacteria and its bacteriophage from 6 cm distance in 2 min, the larger (~40 cm²) RF plasma based device could do the similar killing ability for the larger possible area in 4 min. Optical Emission Spectroscopy (OES) studies revealed that both these devices produce OH radicals which helped in the destruction of both bacteria and its bacteriophage. With suitable modifications, these devices, especially the larger area device may even be implemented for the elimination of Covid-19 affected wards of hospital without using any sensitive chemical process.

Keywords Cold plasma · Germicidal efficiency · Optical emission spectroscopy

Introduction
Pathogenic micro-organisms are among the oldest known reasons for diseases in humans. In spite of great technological advancement of human race, medical science often rendered helpless in front of a pathogenic outbreak. In recent years, SARS, Ebola and ongoing Covid-19 have all shown an urgent requirement to develop novel approaches to destroy them. Traditional sterilization processes for medical products include steam, ethylene oxide (EtO), ionizing radiation (gamma or E-beam), low-temperature steam and formaldehyde, dry heat (hot air) and H₂O₂. Recent developments in sterilization technologies in the last decade focuses on the following factors: low temperature operation so that the technology is suitable for a variety of materials, shorter cycles of operation for faster sterilization, environmental friendliness, and cost reduction (Bunz et al. 2018). Cold plasma can be a useful tool for destroying all types of pathogenic micro-organisms (Bunz et al. 2018; Nasir et al. 2016; Casas-Junco et al. 2019). Cold plasma can be used in vacuum as well as in atmospheric pressure (Barve et al. 2017; Kar et al. 2018; Sharma et al. 2017, 2018). At present, the spread of Covid-19 and other types of novel pathogens have steered the scientific community worldwide to find quick and effective solutions for the sterilization of various medical equipments as well as disinfection of residential units and hospitals. For sterilization of harmful pathogens, atmospheric pressure cold plasma based technologies has been successfully utilized for the killing of laboratory strain of bacteria (Aeromonas) and its bacteriophage (virus of bacteria).

Two cold plasma devices have been tweaked to suit the application of pathogenic destruction. These devices were also characterized by optical emission spectroscopic (OES) method to determine existence of important active species for destruction of pathogens.

1. Device A Microwave cold plasma jet and
2. Device B RF—Hollow cathode cold plasma device.
While device A was originally designed and used for radioactive decontamination (Kar et al. 2018), this device can also effectively destroy Aeromonas bacteria and its bacteriophage even from 6 cm distance. It can be useful for treating infections on human body including open wounds and also sterilization of medical equipments. Device B was originally used for fabrication of carbon nanostructures (Sharma et al. 2017, 2018). However, by carefully changing the gas composition in the plasma and operating power a mere 3 and 4 min exposure under this plasma can effectively destroy Aeromonas bacteria and its bacteriophage, respectively. These devices can help in sterilization of the pathogens.

Experimental and Device Information

Device A: Microwave Cold Plasma Jet This pencil-like device (Fig. 1a) operates in low power (< 100 watts) at microwave frequency (2.45 GHz). The device has a simplistic design consisting of a stainless steel hollow outer shaft and a solid central electrode along the axis of the hollow shaft. The schematic of experimental setup is shown in the Fig. 1c. Ar, the plasma forming gas is bubbled through a heated H$_2$O$_2$ solution to produce plasma in this case. H$_2$O$_2$ decomposes in plasma environment and produces biocidal hydroxyl and hydroxy free radicals and also H$_2$O$_2$ molecules in excited state. The latter is a source of UV radiation which also helps in sterilization process (Sheng et al. 2014).

Device B: RF-Hollow Cathode Cold Plasma Device It’s 13.56 MHz RF source based hollow-cathode cold plasma device whose detailed description already been given elsewhere (Sharma et al. 2017, 2018). Here, two electrodes are kept equal potential and the distance between these two electrodes are less than the mean free path of electron-neutral collision. Under this condition, an electron will make an oscillatory motion between two electrodes gaining energy and when collides with a neutral will transfer this energy. Now, under conducive experimental conditions, plasma will be generated because of this. In the current configuration, this condition is achieved by making a spiral cut groove (~ 300 μm) inside a metallic sheet and both side of the groove remains at equal potential. Gas flows out through the groove to generate plasma. This electrode is kept live with 13.56 MHz RF supply and plasma is generated in these spiral gaps. The ground electrode, which used to be the substrate holder in the previous cases, is a brass disc here with punctuated holes of 3 mm diameter. Another major improvement was to increase the active plasma area from ~ 13 to ~ 40 cm$^2$

Fig. 1 Photograph of a device A, b device B, schematic of experimental setup with c device A and d device B
facilitating larger area applications. The actual photograph of the device is shown in Fig. 1b. This device is suitable for disinfecting larger area. The estimated costs of device A and device B are ~10,000 INR and 80,000 INR, respectively. These costs are on-time setup costs and these fabrication costs and it can be reduced by 30% for bulk production. The schematic of experimental set up is shown in the Fig. 1d. Here, only Ar gas is sent through the device in a controlled manner to produce plasma. Details of the experiments are mentioned in Table 1. Device A typically works between 20 and 80 watts for this purpose and device B works between 50 and 100 watts. The table also shows that there is a need for higher power for Device B. This is because of larger plasma area of the device. When, Device A has a plasma jet of ~2 cm length and 3 mm² area, Device B has an active plasma area of ~40 cm². However, plasma of device B is not jet-like and remains confined on the electrode surface. Owing to the shorter jet length of Device B, it was operated closer to the biological samples compared to Device A, so that the reactive species can reach the sample surface.

Results and Discussion

Figure 2 shows the results of cold plasma treatment on the laboratory strain of Aeromonas bacteria and its bacteriophage virus by both the devices. Standard control sample for bacteria is shown in Fig. 2a. From Fig. 2b it can be seen that 2 min of plasma treatment form device A can effectively destroy Aeromonas even from 6 cm distance (3rd quadrant

| Instrument | Micro-organisms | Gas Composition | (Forward power − Reflected power) = Net power (watt) | Exposure time (Min.) | Distance from plasma (cm) |
|------------|----------------|-----------------|---------------------------------|-------------------|---------------------|
| Device A   | Aeromonas      | 10 LPM Argon bubbled through 11% H₂O₂ solution, heated at 75 °C | (60–10) = 50       | 1                 | 4.5                 |
|            | Aeromonas      | Same as above   | (60–10) = 50                   | 2                 | 6                   |
|            | Bacteriophage  | Argon at 18 LPM | (90–0) = 90                    | 4                 | 2                   |
| Device B   | Aeromonas      | Argon at 18 LPM |                                   |                    |                     |
|            | Bacteriophage  | Argon at 18 LPM |                                   |                    |                     |

Fig. 2  a Standard control sample for Aeromonas bacteria, b, c Growth after 24 h incubation following cold plasma treatment with device A and device B, respectively. d Standard control sample for bacteriophage for device A, e growth after 24 h incubation following cold plasma treatment results with device A on bacteriophage, f standard control sample for bacteriophage for device B, g growth after 24 h incubation following cold plasma treatment with device B on bacteriophage. The zone of clearance due to phage killing is marked by a circle in (d–g)
in Fig. 2b), while similar type of result can be obtained in device B (Fig. 2c) after 3 min of Ar plasma treatment.

Figure 2d, f shows control sets for bacteriophage for device A and device B, respectively. In this experiment, 5 µl of the known concentration virus (∼10⁶ pfu/ml) (plaque forming unit, pfu) is spotted on the top of previously spread bacteria on the plate to make a lawn. Due to the higher count [100–1000 pfu] in the spotted region, the viruses do not form individual plaques and rather form a zone of confluent clearing. The appearance of a clearing in the spotted region means that the virus is active and growth of bacteria in the area indicates that the virus in inactive. It may be noted that due to the higher count of virus in the spotted region, the viruses do not form individual plaques and rather form a zone of confluent clearance. Figure 2e, g shows control sets for bacteriophage for device A and device B, respectively. In this experiment, 5 µl of the known concentration virus (~ 10⁶ pfu/ml) (plaque forming unit, pfu) is spotted on the top of previously spread bacteria on the plate to make a lawn. Due to the higher count [100–1000 pfu] in the spotted region, the viruses do not form individual plaques and rather form a zone of confluent clearing. The appearance of a clearing in the spotted region means that the virus is active and growth of bacteria in the area indicates that the virus in inactive. It may be noted that due to the higher count of virus in the spotted region, the viruses do not form individual plaques and rather form a zone of confluent clearance. Figure 2e, g shows control sets for bacteriophage for device A and device B, respectively with the aforementioned experimental parameters. Thus, device A was able to contain growth of bacteriophage from 6 cm distance by 2 min of plasma treatment and similar result is obtained in device B with 4 min of treatment from 2 cm distance.

It is to be mentioned that bacteriophage has the outer protective layer made of protein similar to Covid-19 and hence more resistant to external sanitizing agents. This virus has also been reported as stronger than corona virus (Turgeon et al. 2014). Since these devices are capable of destroying this bacteriophage, it might be also effective for destroying Covid-19 type of viruses. At present, this technology might be useful in hospitals for bulk disinfection of personal belongings (e.g., mobile phones, purse, tiffin box, jewellery etc.) and surgical equipments in environment friendly dry manner.

From the application point of view, an approximate running cost of both the devices has been calculated. Device A takes up 0.3–0.4 units of electricity in single shift (8 h) operation and additional Ar gas cost will be ~ INR 200–250 per shift. For device B, it will take 0.7–0.8 units of electricity in one shift and additional Ar gas cost will be ~ INR 400–600. Device A will need very little amount of H₂O₂ and the cost is estimated between INR 50–65 in one shift (for INR 1300/1 lit 3% diluted H₂O₂ solution).

Now, as the billing unit price varies in different places depending on load, electricity provider and usage, INR 6 was taken price per unit for the calculation.

Thus, operating device A for one shift will cost between INR 252–317 and device B for one shift will cost between INR 404–605.

Also, considering one sterilization cycle of device A and device B for 10 min each, an 8-h shift can accommodate (480/10) = 48 runs in each case. Thus, cost of each cycle will approximately be INR 5.25–6.06 for device A and INR 8.41–12.60 for device B. Preliminary investigations indicated that materials placed within a (25 × 15 × 15) cm³ chamber, attached to the device can be disinfected in each cycle.

Device A is a line of sight device in its present capacity but device B can disinfect much larger area (2D) because of the structure. This device has also shown capability of disinfection in 3D and more research are being conducted to understand its potential. A process is ongoing to integrate disinfection chamber with the device. However, these studies are still ongoing and different experiments are being conducted.

Typical OES spectra recorded during operation of both the devices are shown in Fig. 3a, b. Basic spectroscopic notations have been used here. Any symbol of an element followed with roman numeric I means de-excited atomic state, e.g., Ar I means that particular atomic line is originated as Ar atom comes to a particular de-excited state from a higher state. Same is true for OI line. Atomic lines appear very sharp in the spectrum. Emission of molecules like NH, OH occurs in visible region due to electronic de-excitation mechanism. Molecular emission lines always occur in bands. Each molecule can produce different bands and they have different probability to occur in a certain experimental conditions. Some of these bands are named after the inventor or their distinct features, e.g., N₂ Kaplan band etc.

Figure 3a exhibits typical characteristic emission lines of the plasma species during operation of Device A. Here, Ar gas was bubbled through the 11% H₂O₂ solution to carry H₂O₂ vapour
in plasma resulting in the occurrence of characteristic OH peek at 281 and 309 nm (Pearse and Gaydon 1950). This OH radical plays an important role in sterilization process (Sheng et al. 2014). Apart from that, Characteristic Hα line at 656 nm and Ar I and OI lines can also be seen in the spectra. A remarkable fact to notice in the plasma is that although Ar is the major constituent of the plasma, OI line is the strongest among all. Oxygen can get incorporated in the plasma from H2O2 and the atmosphere as well. Although the intensity of the OI line does not imply increased dissociation of H2O2, the presence of OI, Hα and OH lines indicates effectiveness of this device for destruction of pathogens. Figure 3b represents typical characteristic emission lines from plasma during operation of Device B. It is an interesting observation that, in spite of absence of H2O2 in plasma, characteristic emission lines of OH, O I are observed along with NH. Different molecular bands of N2 and NO are also observed here along with Ar I lines. Device B is actually helping in dissociation of atmospheric moisture and air resulting in emission of the above bands. Although the reason for this behavior is not clear yet but it is understood that reason lies in different plasma generation mechanism (2.45 GHz for device A and 13.56 MHz for device B) for two devices. Although these experiments show generation of different gemicidal reactive oxidation species (ROS) like OH and OI but ‘safety cross checking’ is inevitable to qualify the device for practical application. This is presently being done with more systematic studies to fulfill this need.

According to the OES spectra, UV A (400–315 nm), UV B (315–280 nm), and UV C (280–100 nm), all three bands of the UV radiation is getting generated. However, UV B band seems to be the strongest. In plasma, active species like free radicals, ions, and UV radiation, all act in synergy to provide the biocidal effect. For the ultraviolet/near ultraviolet spectral region, exposure to the eye should not exceed 1 milliwatt per square centimetre (1.0 mW/cm2) for periods greater than 1000 s. So, total dose comes around 0.03 mJ/cm2 which is well below the maximum limit. No additional precaution was necessary for operation of the devices.

Obtained results clearly indicate that both these devices can be successfully used to contain the spread of pathogenic micro-organism like Covid-19.

Conclusion

These findings clearly indicate that both the devices are capable of destruction of pathogenic Aeromonas bacteria and its bacteriophage. It is seen that 3–4 min exposure is sufficient for the killing of bacteria and its bacteriophage. It is, therefore, anticipated that the devices surely have the capability of destruction of Covid-19 and similar strain of viruses successfully. The devices show ability of rapid disinfection as compared to presently available plasma sterilization technologies. The study shows that Device B is quite advantageous over Device A in terms of larger application area. Also, it is capable of producing reactive species and UV radiation even without application of O2, H2O2, or H2O inside plasma which also make the device more cost effective in comparison. Hence, in conclusion, device B is more suitable among two for gemicidal applications.

Acknowledgements The authors’ are grateful to the Director, Bhabha Atomic Research Centre for his constant support, motivation and encouragement during this exercise.

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