Progesterone Induces Mucosal Immunity in a Rodent Model of Human Taeniosis by *Taenia solium*

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Received: 2011.10.01; Accepted: 2011.10.26; Published: 2011.11.10

**Abstract**

More than one quarter of human world's population is exposed to intestinal helminth parasites. The *Taenia solium* tapeworm carrier is the main risk factor in the transmission of both human neurocysticercosis and porcine cysticercosis. Sex steroids play an important role during *T. solium* infection, particularly progesterone has been proposed as a key immune-modulatory hormone involved in susceptibility to human taeniosis in woman and cysticercosis in pregnant pigs. Thus, we evaluated the effect of progesterone administration upon the experimental taeniosis in golden hamsters (*Mesocricetus auratus*). Intact female adult hamsters were randomly divided into 3 groups: progesterone-subcutaneously treated; olive oil-treated as the vehicle group; and untreated controls. Animals were treated every other day during 4 weeks. After 2 weeks of treatment, all hamsters were orally infected with 4 viable *T. solium* cysticerci. After 2 weeks post infection, progesterone-treated hamsters showed reduction in adult worm recovery by 80%, compared to both vehicle-treated and non-manipulated infected animals. In contrast to control and vehicle groups, progesterone treatment diminished tapeworm length by 75% and increased proliferation rate of leukocytes from spleen and mesenteric lymph nodes of infected hamsters by 5-fold. The latter exhibited high expression levels of IL-4, IL-6 and TNF-α at the duodenal mucosa, accompanied with polymorphonuclear leukocytes infiltration. These results support that progesterone protects hamsters from the *T. solium* adult tapeworm establishment by improving the intestinal mucosal immunity, suggesting a potential use of analogues of this hormone as novel inductors of the gut immune response against intestinal helminth infections and probably other bowel-related disorders.

**Key words:** *Taenia solium*, cysticercosis, sex hormones, progesterone, inflammation.

**Introduction**

*Taenia solium* is a cestode parasite that affects both human and pigs [1, 2]. Typically described as a health problem in developing countries [3, 4], *T. solium* infection has received growing attention to be considered as an emerging health problem in the United States and other developed countries [5, 6].
The intermediate stage of *T. solium* causes human neurocysticercosis (hNC) and porcine cysticercosis [7]. hNC represents the most severe clinical manifestation of the human disease, whilst porcine cysticercosis is a source of enormous economic losses due to confiscation of contaminated swine meat [4, 8]. The intestinal adult form of the parasite is receiving rising interest since it is considered as the main risk factor in the propagation of the disease for both organisms [9-11]. The fact that the *T. solium* tapeworm carrier be estimated as a central node in the maintenance of the infection is supported by the presence of hNC in human communities which do not consume swine meat, or even have no contact with pigs [12, 13]. Then, more experimental and clinical research strategies should be directed to control the *T. solium* adult tapeworm establishment and egg production.

Sex steroid hormones have an important role during parasite infections [14-16], either by modulating host immune response [17-18] or having direct effects upon parasites [15]. Interestingly, pregnancy in female pigs and castration in male boars increase the prevalence of naturally acquired cysticercosis [19]. Moreover, *T. solium* infection decreases testosterone levels in non-castrated ranging boars [20]. This evidence leads us to assume that sex hormones can be either permissive or restrictive factors in the establishment of the intermediate stage of *T. solium*. In humans, intestinal taeniosis is more frequent in women than in men [21-23]. Similarly, the inflammatory response associated to the presence of brain located-cysticerci, reexhibits a dimorphic pattern, being more severe in women than in men, presumably due to the higher number of eosinophils, and other proinflammatory effector cells, as well as IL-5 and IL-6 levels, in women cerebral spinal fluid [24].

In the same sense, female mice are more susceptible than males to experimental murine cysticercosis by *T. crassiceps* [25]. Such difference in susceptibility is abolished by gonadectomy of both genders [26]. However, restitution of progesterone to gonadectomized mice from both sexes decreased parasite loads by 100%, presumably due to over-regulation of the intracellular progesterone receptor (PR) at splenic T lymphocytes [27] Similarly, ovariectomy of female mice exacerbates *Trypanosoma cruzi* parasitemia, meanwhile progesterone replacement drops it to similar levels as in control intact groups [28]. In a similar manner, *Schistosoma haematobium*-infected hamsters show a decrease in the number of recovered worms and egg load in response to the administration of medroxyprogesterone acetate, a progesterone analogue indicated as human contraceptive [29]. Pregnancy seems to have a protective role against Trichi-

### Materials and Methods

#### Ethics Statement

Animal care and experimentation practices at the Instituto de Investigaciones Biomédicas are frequently evaluated by the Institute’s Animal Care and Use Committee, according to the official Mexican regulations (NOM-062-ZOO-1999). Mexican regulations are in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (NIH and The
Weatherall Report) of the USA, to ensure compliance with established international regulations and guidelines. The protocol was approved by the Committee on the Ethics of Animal Experiments of the Instituto de Investigaciones Biomédicas (Permit Number: 2009-16). Pigs sacrifice to obtain parasites was performed under sodium pentobarbital anesthesia, and all efforts were made to minimize suffering.

**Parasites**

*T. solium* cysticerci were selected according to the macroscopic criteria reported by León-Cabrera and coworkers [46]. Briefly, parasites were dissected from muscle of naturally infected pigs, which were euthanized at the Veterinary University Nacional Autónoma de México, under consent of the University Animal Care and Use Committee to ensure compliance with international regulations and guidelines. The fibrous capsule surrounding each parasite was carefully separated under a dissection microscope. Once separated, cysticerci were placed in tubes containing sterile PBS (1X) supplemented with 100 U/ml of antibiotics-fungizone (Gibco, Grand Island, NY). Samples were centrifuged at 1200 rpm/4°C for 10 min and the supernatant was discarded. Pellets containing cysticerci were placed in Dulbecco’s Modified Medium (DMEM, Gibco, BRL, Rockville, MD) without fetal serum supplementation. Then, they were washed and centrifuged 3 times at 1200 rpm/4°C for 10 min. After the final wash, complete and translucent reddish cysticerci were incubated on 6-well culture plates containing DMEM medium with 25% pig fresh bile supplementation for infectivity test. When the evagination rate was higher than 90%, then parasites were used for subsequent oral infections.

**Progesterone administration**

Ten female golden hamsters (*Mesocricetus auratus*) of 140-160 g, aging between 8 and 10 weeks, were subcutaneously administered with 2 mg/Kg body weight of water-soluble progesterone (Sigma-Aldrich, USA). Each single dose of progesterone was diluted in 0.4 mL of saline solution (0.9% NaCl, Baker). Control animals (n=10) received 0.4 mL of saline solution as vehicle. A stress-related additional control group (n=10) was included in our experiments, which consisted in ten sham injection animals. Hormone and vehicle administration was carried out each other day during four weeks, in order to maintain the same hormonal serum concentration for the entire time of the experiment. Our results were obtained from two independent experiments performed in similar conditions. Animals were fed with Purine Diet 5015 (Purine, St. Louis, MO) and water ad libitum during all the experiment.

**Oral infection experiments**

Two weeks after the beginning of progesterone or vehicle administration, treated and untreated animals were orally infected with four viable *T. solium* cysticerci, according to previous reports [46]. All animals were euthanized 15 days post infection, using a CO2-saturated chamber. During animal necropsy, the entire small intestine was dissected and placed on a Petri dish containing sterile PBS (1X) (Sigma-Aldrich, USA). Under a stereoscopic microscope, the lumen of all small intestines was carefully exposed by making a longitudinal cut using sterile dissection scissors. Then, duodenum-anchored parasites were counted and measured with a calibrator. Blood samples were individually collected from all animal groups for posterior serum analysis. Ileum attachment zones where *T. solium* scoles were located, were placed in 4% paraformaldehyde (J.T. Baker, México), or Trizol reagent (Invitrogen, Carlsbad, California) for posterior analysis. Immediately after necropsy, spleen weight was individually recorded. Spleen samples and mesenteric lymph nodes from all animal groups were individually obtained and placed in RPMI (Gibco, BRL, Rockville, MD) supplemented with 10% fetal calf serum (Gibco, BRL, Rockville, MD), or 4% paraformaldehyde (J.T. Baker, México), or Trizol reagent (Invitrogen, Carlsbad, California) for posterior analysis.

**Cell culture and lymphoid proliferation**

Total leukocytes and red blood cells were extracted from spleen and mesenteric lymph nodes of all animal groups. After single washing with ACK Lysing Buffer (Invitrogen, USA), total leukocytes were recovered and cultured in 96-well sterile plates (1x10^4 cells/well) containing serum-free RPMI medium (Gibco-BRL), at 37°C in humified 5% CO2 atmosphere for 72 h. After this time, cultured leukocytes from spleen and mesenteric lymph nodes of all animals were exposed to 15 μg/well of freshly extracted *T. solium* total antigen during 48 h. Twenty four h before the end of the experiment, 20 μL of AlamarBlue reagent (Biosource International) were added to each culture well. Then, culture plates were frozen at -30°C under darkness and the absorbance was quantified at 570 and 600 nm, using a microplate reader. The 570-600 nm lecture coefficient was employed to assess proliferation index.

**Cytokines and progesterone receptor expression**

Spleen and anchored tapeworms-related duo-
denum samples were placed in Trizol reagent (Invitrogen, Carlsbad, California). Total RNA extraction was as follows: spleen and duodenum were disrupted in Trizol reagent (1 ml/ 0.1 g organ) and 0.2 ml of chloroform was added per ml of Trizol. The aqueous phase was recovered after 10 min of centrifugation at 13000 rpm, RNA was precipitated with isopropyl alcohol, washed with 75% ethanol and dissolved in RNAase-free water. RNA concentration was determined by absorbance at 260 nm and its purity was verified after electrophoresis on 1.0% denaturing agarose gel in presence of 2.2 M formaldehyde. Immediately, total RNA samples were reverse-transcribed by using M-MLV Retrotranscriptase system and dT primer (Invitrogen, USA). Then, cDNA was specifically amplified by semi-quantitative PCR, using TaqDNA polymerase (Biotecnologias Universitarias, UNAM, México) and hamster-specific primers to detect IFNγ, IL-12, IL-4, TNFα, IL-6, PR and 18S-ribosomal RNA (Table 1). Briefly, the 50 µl PCR reaction included 10 µl of previously synthesized cDNA, 5 µl of 10X PCR-buffer (Perkin-Elmer, USA), 1 mM MgCl2, 0.2 mM of each dNTP, 0.05 µM of each primer, and 2.5 units of TaqDNA polymerase (Biotecnologias Universitarias, Mexico). After an initial denaturation step at 95°C for 5 min, temperature cycling was as follows:

- 95°C for 30 s, from 51°C to 62°C (depending on primer sequence) for 45 s and 72°C for 45 s during 35 cycles. An extra extension step was completed at 72°C/10 min for each gene. The 50 µl of the PCR reaction were electrophoresed on 2% agarose gel, stained with ethidium bromide and visible after electrophoresis. The 50 µl of the PCR reaction was as follows:

- 95°C for 30 s, from 51°C to 62°C
- 5°C for 45 s and 72°C for 45 s during 35

Histological examination of inflammatory infiltrate

Anchored tapeworms-related duodenum samples and several fragments of jejunum and ileum from progesterone and vehicle-treated groups, as well as sham injection-infected animals, were placed in 4% paraformaldehyde for 2 weeks (J.T. Baker, México) and subsequently…processed to paraffin embedding and to obtain 4µm tissue sections which were stained with hematoxylin-eosin (H&E) and toluidine blue to demonstrate mast cells. Specimens were washed twice with 1X PBS solution (Sigma-Aldrich, USA), and then were dehydrated using 70%, 80%, 95% and 100% ethanol (J.T. Baker, México), during 15 min each one. Subsequently, organs were placed into xylene (J.T. Baker, México) for 30 min and embedded in 60°C liquid paraffin during 30 min. After this time, liquid paraffin was replaced with new fresh 60°C paraffin. Once the blocks were solidified after 24 h at room temperature, duodenum tissue was cross-sectioned in thin 4 µM slices, by using a microtome (Microtome Olympus Cut 4060, USA). Sections were stained with hematoxylin-eosin (H&E). Evaluation of inflammatory infiltrate degree on H&E-stained duodenum sections was performed under a light microscope using 400 X magnification.

| Table 1. Primers used for amplification of hamster-specific genes. Primer sequences were designed based on hamster-specific gene sequences reported in the Gene database, NCBI, NIH. Primer sequence as well expected molecular weight of the PCR product is shown. bp=base pairs. |

| Primer definition | Primer sequence | Molecular weight of the PCR product (bp) |
|-------------------|-----------------|----------------------------------------|
| IFN-γ (sense)     | 5’TCAAAAGGCTGTTGACACAA | 326 |
| IFN-γ (antisense) | 5’TCTTGTGAGCCGATTTCC  | 167 |
| IL-12 (sense)     | 5’GGCTAGAACAGCTACAGA | 247 |
| IL-12 (antisense) | 5’GGCTAGAACAGCTACAGA | 302 |
| IL-6 (sense)      | 5’CTGAGACTTCATCATCATTCG | 229 |
| IL-6 (antisense)  | 5’CTGAGACTTCATCATCATTCG | 198 |
| TNFα (sense)      | 5’GGAGGAAGAGTTCCTCCACC | 219 |
| TNFα (antisense)  | 5’GGAGGAAGAGTTCCTCCACC |  |
| PR (sense)        | 5’AGTCGGGAATCTGTTACG |  |
| PR (antisense)    | 5’AGTCGGGAATCTGTTACG |  |
| 18S (sense)       | 5’AGTCGGCATCAGTTATGTC |  |
| 18S (antisense)   | 5’AGTCGGCATCAGTTATGTC |  |

Immunofluorescence

Antibodies anti-mouse IL2, IL1β, IL5 and TGFβ done in rabbit, anti-mouse IFNγ, TNFa, IL12, IL10, IL6 and IL13 done in goat, and anti-mouse IL4 done in rat were used as primary antibodies (Santa Cruz, Biotechnology, USA). Anti-rabbit IgG and anti-goat IgG conjugated with rhodamine (TRITC) and anti- rat IgG conjugated with fluorescein isothiocyanate (FITC) (ZIMED Laboratories Inc., USA) were used as secondary antibodies. Duodenums were fixed in 4% paraformaldehyde for 48 h, washed with PBS and stored in PBS containing 30% sucrose at 4°C overnight. Next day, samples were embedded in tissue-freezing medium (Leica, Nussloch, Germany) and frozen at -70°C (dry ice hexane). Serial sections of 20 µm thicknesses were obtained using a cryomicrotome, placed on slides coated with poly-L-lysine (Sigma)
and air-dried. Sections were treated with 1% Triton X-100 after blocking with 1% albumin (BSA) and incubated with primary antibody overnight at 4°C diluted in 1:1000 BSA-PBS 1X. After rinsing in PBS, the sections were incubated with the secondary antibody during 1 h at room temperature diluted in 1:200 BSA-PBS 1X, washed in PBS and embedded in anti-fading DAKO mounting medium (DAKO, USA). Intestine sections processed without the primary antibody were used as negative controls.

For each group, several microscopic fields of duodenum were captured and analyzed by semi-quantitative immunofluorescence laser confocal microscopy. In brief, TIFF images were acquired with the TCS-SPI software and imported into Image Pro Plus for subsequent measuring of the immunofluorescence intensity. In each image, representative areas were selected and, with the exposure times kept constant, the intensity of fluorescence was quantified and expressed as the mean pixel intensity for each region. For each animal, at least six randomly selected areas were analyzed.

**Experimental design and statistical analysis**

Our results were estimated in 2 independent experiments. All experimental groups consisted in 10 animals each. Dependent variables were number of intestinally attached parasites, adult tapeworm length (mm), proliferation index, IL-4, IL-6, TNF-α and PR expression, as well number of inflammatory foci. Number of mast cells is expressed at mm² of tissue section. The independent variable was progesterone treatment. Data from 2 replicates (n=20) of each experimental group were expressed as an average +/- standard deviation, and analyzed by means of one way-ANOVA, and Tukey test as post hoc test. Differences were considered significant when P< 0.05.

**Results**

After 15 days post infection, progesterone treatment significantly reduced the number of intestinally anchored-*T. solium* tapeworms by 80% (Fig. 1A). Vehicle-treated and sham injection-infected hamsters showed between three and four viable parasites (Fig. 1A). It is important to remark that all found tapeworms were strongly attached to the duodenum zone. As expected, tapeworms from vehicle-treated and sham injection-infected hamsters grew up more than 1.5 mm +/- 0.4 (Fig. 1B). In contrast, parasite from progesterone-treated hamsters did not develop more than 0.2 mm +/- 0.16 in length (Fig. 1B), showing besides poorly differentiated scolices. Thus, steady concentrations of progesterone exerted a protective role against the *T. solium* intestinal infection, diminishing both the number of attached parasites and their development.

To assess the possible mechanism involved in progesterone protective actions during infection, spleens from all animal groups were weighed and splenic leukocytes assayed for antigen-specific proliferation (Fig. 2). Although no significant differences were observed among the spleen weight from progesterone, vehicle-treated animals and sham infection-infected hamsters (Fig. 2A), there was a discrete but interesting tendency in hamsters exposed to progesterone to increase spleen weight respect to their infected control littermates. Furthermore, *in vivo* progesterone treatment clearly increased proliferation *in vitro* of *T. solium* antigen-specific leukocytes by 5-fold compared to both infected control groups (Fig. 2B). This result suggests that progesterone should protect hamsters from *T. solium* infection through promotion of a local mucosal anti-parasite immune response.

To evaluate the immuno-stimulating effects of progesterone, cytokine profile expression from spleen and parasite associated-duodenum, as well as PR expression, were assessed (Fig. 3). In spleen, progesterone did not modify gene expression of IL-12 and IFN-γ (Figs. 3A, 3B) but significantly increased by 8-fold the expression of IL-4, one of the most representative Th2-related cytokines (Fig. 3E). A similar effect was observed in the expression of proinflammatory cytokines IL-6 and TNF-α, which showed an increase by 5-fold and 9-fold, respectively (Figs. 3C, 3D). Of special interest was to determine the cytokines profile of duodenal mucosa from progesterone-treated and untreated hamsters, because this tissue was in direct contact with parasites. Interestingly, both proinflammatory cytokines IL-6 and TNF-α were locally increased by 6-fold each (Figs. 3H, 3I), as well as IL-4 (Fig. 3J), in response to progesterone treatment. As was seen in spleen, duodenal expression of IL-12 and IFN-γ was not modified by any treatment (Fig. 3F, 3G). PR expression in progesterone-treated animals was diminished by 4-fold in the spleen, compared to both control infected groups (Fig. 4A). Moreover, progesterone treatment also down-regulated PR expression by 50% in tapeworms-associated duodenum (Fig. 4B). We speculate that progesterone induces protection on *T. solium*-infected hamsters by locally increasing proinflammatory and Th2-related cytokines expression, probably mediated by its intracellular PR, which showed a classical down-regulation pattern in response to progesterone.
Figure 1. Progesterone decreases both parasite load and tapeworm length in T. solium cysticerci-orally infected hamsters. (A) Administration of progesterone significantly diminished the number of intestinally attached adult tapeworms by 80%, respect to both control and vehicle infected groups. (B) Parasites exposed to constant concentrations of progesterone showed total length reduction of seven-fold. These parasites seemed as undifferentiated scolices with no develop of neck and strobila, compared to those tapeworms from control and vehicle groups with well differentiated structures. Tapeworm length was determined as the longitudinal sum of scolex, neck and strobila. Non-manipulated infected hamsters were denominated as Control, meanwhile comparative lines represent significant differences when P<0.05. Results are presented as mean +/- standard deviation.

Figure 2. Progesterone administration increases proliferation rate of parasite specific-leukocytes from spleen and mesenteric lymph nodes. (A) Although progesterone-treated animals showed a discrete tendency to increase spleen weight, no significant differences in spleen weight were observed among treatments. (B) Spleen and mesenteric lymph nodes leukocytes from progesterone and vehicle-treated, as well non-manipulated infected hamsters were in vitro cultured in presence of T. solium total antigen. As a consequence of the in vivo exposition to progesterone, proliferation index of parasitic antigen specific-leukocytes was augmented by 3.5 and 5-fold, respect to immune cells from both control and vehicle-treated hamsters. Non-manipulated infected hamsters were denominated as control. Results are presented as mean +/- standard deviation. **P<0.05 vs the other groups.
Figure 3. Progesterone administration induces IL-6, TNF-α and IL-4 expression on *T. solium* cysticerci-orally infected hamsters. Progesterone induced expression of proinflammatory and Th2-related cytokines IL-6, TNF-α and IL-4, systemically from spleen (A-E) and locally on parasite-associated duodenum (F-J). No significant effects were observed in the Th1-related cytokines IFN-γ and IL-12 expression. 18S was used as control gene of constitutive expression. Optical densitometry (OD) was obtained as a relation between the genes of interest and 18S, and reported as mean +/- standard deviation. **P<0.05 vs the other groups.

Figure 4. Progesterone receptor (PR) expression is reduced in response to progesterone administration. PR showed a classical down-regulation in (A) spleen and (B) duodenum of progesterone-treated infected hamsters. In the spleen, PR expression was diminished by 4-fold respect to both control and vehicle groups (A), meanwhile it presented a decrease of 5-fold in parasite-associated duodenum (B). Data are reported as mean +/- standard deviation. **P<0.05.
Local inflammation at intestinal sites where tapeworms were found was evaluated (Fig. 5). Histological condition of duodenal mucosa from control infected animals was quietly normal, exhibiting a well-defined intestinal villi formed by tunica mucosa. (Fig. 5A, 5B). Progesterone-treated hamsters presented an exacerbated inflammatory infiltrate located along the lamina propria, clearly related to the parasite presence, mainly characterized by eosinophils (~80%), lymphocytes (~10%) and basophils (~5%) (Fig. 5C, 5D). Unexpectedly, bigger and reddish Peyer’s patches were observed in this experimental group, and when parasites were closely located to these immune structures, they exhibited more damage and poor differentiation. As a consequence of such inflammation intensity (Fig. 5D), several properties of the duodenal tissue were lost, such as lack of microvilli number and enlargement, as well as an important increase in goblet cells and mucus production (Fig. 5C). In vehicle-treated (Fig. 5B) and sham injection-infected animals (Fig. 5A) there was an apparent inflammatory infiltrate in response to the presence of the parasite. However, such inflammation reaction had no the same magnitude than such observed in progesterone-treated animals (Fig. 5D), which probably was strongly related to differences in number of attached parasites, their appearance and length from both progesterone-treated and untreated studied groups.

Figure 5. Progesterone promotes exacerbation of inflammatory infiltrate on parasite-associated duodenum from infected hamsters. (A) Non-manipulated infected hamsters and (B) vehicle infected hamsters showed no exacerbated inflammatory infiltrate and well defined tissue structures, although presence of inflammatory infiltrate associated to parasites was still evident. (C) Progesterone treated and infected hamsters showed an exacerbated inflammatory response at duodenal mucosa with eosinophils, basophils and lymphocytes infiltration. (D) Progesterone treated and infected hamsters showed as well lack of tissue arrangement. A: 6X magnification. B-C: 10X magnification. D: 40X magnification. Scale bar=50 microns.
Figure 6. Mucosal mast cells from small intestine mucosa of experimental T. solium-infected hamsters. Progesterone-treated and infected hamsters (A, B) contained more mast cells with abundant and bigger metachromatic granules in the cytoplasm, than vehicle-treated and infected hamsters (C, D) and non-manipulated infected-hamsters (E, F). These findings were consistently observed in mast cells located either in periglandular lamina propria (B, D, F) as in intestinal villi (A, C, E). Arrows indicate toluidine blue stained mast cells. Scale bar=25µm.

Accordingly, progesterone administration induced infiltration of mast cells mainly located at the lamina propria of intestinal glands (Fig. 6B) and villi (Fig. 6A). In progesterone-treated animals mast cells infiltrate was closer associated to parasite’s attachment zones and Peyer’s patches. Moreover, these cells were bigger than those observed in the small intestine of vehicle (Figs. 6 C,D) and control (Figs. 6 E,F) hamsters, exhibiting high rate of metachromatic granules. A higher number of mononuclear and plasmatic cells were frequently observed in these intestinal specimens. In contrast, vehicle and control animals showed low number of small mast cells with poor degranulatory activity (Figs. 6C-F). Overall, parasite elimination was strongly associated with intestinal tissue damage which in turn showed high correlation with progesterone-induced mast cells degranulation. These data support that progesterone administration is able to exacerbate the host’s mucosa inflammatory response that subsequently could mediate reduction in parasite number and growth.

As the anti-parasite immune response in hamsters was related with local inflammation, we carried out immunohistochemical assays in tissue sections from duodenum, in order to determine and compare the local profile of cytokines in each studied group. The mean intensity of IFN-γ was significantly augmented in duodenum sections from progesterone-treated animals, as compared to vehicle-treated and untreated controls (Table 2). Concomitantly, production of IL-5 and IL-13 was diminished in response to the administration of this hormone. This group also exhibited a decrease in the mean intensity of TNF-α, whilst fluorescence intensity related to IL-10 was clearly increased by 4-fold concerning controls. These results suggest that progesterone is able to modify the immunological profile of cytokines at the intestinal mucosa of hamsters, which could be associated with protection against the T. solium infection.

Discussion

Several efforts have been performed in order to control helminthiasis such as onchocerciasis, filariasis, schistosomiasis and taeniosis, among others [47-50]. Mostly based on drug and vaccine design [11, 46, 47, 51], these strategies focus on inhibiting establishment
and reproduction of adult intestinal worms. Although the adult stage of most of these parasites does not cause any severe pathology, with practically asymptomatic clinical manifestations [52], it is now recognized as the main risk factor in the propagation of the disease to natural and accidental intermediate host [10, 11, 46]. It is the case for human and porcine cysticercosis by T. solium [10, 46]. For this reason and by using the experimental taeniasis model in hamsters, our work centers to evaluate the possible protective role of progesterone upon the establishment of the T. solium adult tapeworm, since this hormone possess immunomodulatory properties and participates in pig cysticercosis [27-30].

In fact, progesterone showed a protective effect upon establishment of the T. solium adult stage in hamsters. It is important to mention that the administration of progesterone significantly reduced parasite load and tapeworm growth, with similar results to those obtained through vaccination [46, 51-53]. Therefore, combined treatment of immunogenic molecules and progesterone could be a hopeful alternative for reach a total protection against intestinal parasites invasion. In regard to this issue, several studies report the use of hormones as vaccine’s adjuvants with prominent results preventing type A influenza, tetanus and tuberculosis [54-57].

Table 2. Cytokine pattern in duodenal tissue from progesterone-treated and control hamsters infected with the adult tapeworm of Taenia solium. *Fluorescence intensity data are presented as mean ± standard deviation. †P<0.05, ‡P<0.01.

| Cytokine Family | Cytokine | Untreated controls* | Vehicle* | Progesterone* |
|-----------------|----------|---------------------|----------|--------------|
| Th1             | IL-2     | 53.8±3.9            | 56.2±4.2 | 54.6±1.1     |
|                 | IL-12    | 50.4±4.6            | 53.4±3.2 | 52.3±3.8     |
|                 | IFN-γ    | 44.1±2.4            | 46.6±4.1 | 52.1±3.3     |
|                 | IL-4     | 51.2±1.8            | 50.5±0.6 | 49.8±3.3     |
| Th2             | IL-5     | 44.6±1.5            | 43.7±2.1 | 32.8±3.3     |
|                 | IL-13    | 47.2±2.7            | 44.0±3.6 | 25.0±3.0     |
|                 | TGF-β1   | 31.5±3.2            | 30.0±3.0 | 34.1±1.5     |
| Proinflammatory | IL-1β    | 45.9±3.6            | 46.5±4.1 | 46.2±2.7     |
|                 | IL-6     | 27.8±1.7            | 28.4±1.2 | 25.7±0.8     |
|                 | TNF-α    | 54.1±2.4            | 55.5±1.8 | 42.6±2.8     |
| Regulatory      | IL-10    | 8.6±0.9             | 8.0±1.1  | 32.6±6.1     |

Progesterone also seems to over-activate immune cells that specifically recognize parasite antigens. Worth of mention is the prominent proliferation rate of T lymphocytes in presence of T. solium antigens, which lead us to suppose that progesterone may act through two main ways: a) by increasing antigen specific-immune cell viability and proliferation or b) inducing a long duration-preactivated status on host immune cells. Although both possible explanations are intriguing, they require further experimental analysis.

On the other side, progesterone was able to locally over-regulate proinflammatory and Th2-related cytokines expression. As we have previously shown, the immunomodulatory effects of progesterone on several peripheral immune cell types have seen widely described [43, 45, 58, 59]. Nevertheless, its regulator role upon mucosal immunity has been scarcely studied [60-62]. Our data show that progesterone treatment has the ability to induce IL-4, IL-6 and TNF-α expression at duodenal mucosa of infected hamsters. Expression of these cytokines has been associated to parasite elimination in this animal model [63]. Then, progesterone could improve protection from T. solium infection by peripherally increasing parasite specific-immune cell proliferation, with concomitant local stimulation of IL-4, IL-6 and TNF-α expression. In turn, these cytokines could promote an exacerbated inflammatory reaction with enormous capacity to affect parasite establishment and growth through the activation of eosinophil, basophil and, importantly, mast cells. Of additional interest is the possible direct role of progesterone upon these immune cell types associated to the mucosa. For in-
stance, during human chronic urticaria several reports suggest that mast cells and eosinophil could be over-activated by pregnancy-related factors such as progesterone and estrogens [64, 65]. Concomitantly, 17β-estradiol is able to promote release of β-hexosaminidase and IgE-induced degranulation from mast cell and basophil cell lines [66], meanwhile on uterine mast cells this sex steroid significantly increases histamine release [67]. This suggests that mucosa-associated-innate immune response cells should be affected by sex steroid hormones [66, 67]. However, further investigation concerning the participation of these hormones in the modulation of mast cell, basophil and eosinophil activity at the intestinal mucosa is required.

Progesterone effects may be mediated by its intracellular PR. Our data show that PR expression was down-regulated by progesterone treatment, which denotes a classical feed-back regulation, previously described in several tissues as a regulatory mechanism of progesterone actions. However, to assess this point is necessary the use of anti-progestins with capacity to competitively bind to PR isoforms in hamsters [68, 69].

In experimental T. solium infection in hamsters, the acute inflammatory response is characterized by the presence of eosinophil, neutrophils, macrophages and lymphocytes, that promote destruction of the surrounding cells [26, 27, 38]. Even though Th1-type cytokines activate neutrophils and macrophages in vitro resulting in parasiticidal activity mediated by nitric oxide [39-41] experiments in vivo have shown that neutrophils and macrophages are not capable of killing taenias. The host’s cellular immune response against parasites is regulated by cytokines, and therefore the Th1/Th2 type polarization can result in protection or susceptibility [43]. Several reports have described the ability of sex hormones to influence all cellular types of the innate and adaptive immune systems, thereby modifying a multitude of immunological functions [44, 45]. Moreover, sex hormones influence the development, maturation and state of activation of T lymphocytes, including the Th1/Th2 balance [47, 48].

It has been reported that in the late phase of T. solium infection in hamsters, when animals recover from an acute immunosuppression, a Th1 response occurred: IL-2 seems to be the major cytokine responsible for protection, whereas Th2-cytokines seem to be associated with susceptibility [46, 49, 50]. Our study sampled a broad profile of the cytokines with ability of influencing the course of infection in hamsters. At protein levels, semiquantitative immunofluorescence analysis from duodenum tissue samples suggested that the treatment with progesterone resulted in low levels of Th2-cytokines, accompanied by polarization of the cellular response toward a Th1-type, which has been related with parasite elimination [46, 49, 50]. In contrast to results obtained by mRNA expression, protein levels of TNF-α are significantly reduced in progesterone-treated animals, while IL-10 is dramatically increased. This apparently controversial result can be explained since gene products are susceptible of post-transcriptional regulation mechanisms, as it is well known. Such mechanisms guarantee a balanced response at protein and cellular levels. Particularly, IL-10 could be overproduced in order to diminish exacerbation of the inflammatory response at the duodenal mucosa, which besides eliminating taenias, could result in serious detriment for the host. Thus, the protective effect of the Th1-immune profile observed in progesterone-treated hamsters supports the notion that a strong Th1 response is associated with parasite elimination. Concomitantly, elevation of regulatory cytokines could be involved in preventing inflammation-related injury at the duodenal mucosa of the host.

Progesterone treatment could also have a direct effect on the parasite. In this regard, T. solium has the ability to modulate the expression of molecules related with virulence upon contact with host components, including hormones. In other parasites, specific hormone receptors have been shown to regulate the expression of diverse proteins [52, 53]. Our preliminary data indicate that a non-classic receptor for progesterone is present in the cysticerci of T. solium [40].

Although the hamster model used here possibly does not reflect exactly all aspects of human taeniosis, a set of interactions are established between the parasite, the immune system and the endocrine system, which define the outcome of the infection caused by T. solium. Evidence presented here shows that progesterone treatment in hamsters reduces establishment and development of T. solium, possibly due to a specific increase in the local mucosal immunity in the duodenum, characterized by a strong inflammatory infiltrate and a Th1 immune response, suggesting that interactions between the immune and endocrine systems play a fundamental role in the establishment, development and outcome of intestinal T. solium infection in hamsters.

It is important to say that the experimental proof exposed in this work does not only concern to intestinal parasite infections, but also to other highly prevalent human gastrointestinal disorders where hormones seem to play a decisive role [70-80] such as irritable bowel syndrome (IBS) characterized by recurrent abdominal pain, bloating and constipation.
which is one of the most frequent chronic pelvic pain around the world [70, 71]. Besides, the evidence that progesterone and estradiol are able to modify the intestinal transit time, visceral sensitivity and gut function [72-74] it has been demonstrated that the administration of progesterone diminishes intestinal production of IL-1β and TNF-α, decreasing in turn bowel inflammation, damage and apoptosis in rats undergoing traumatic brain injury [75]. Concomitantly, several clinical trials suggest that the use of medroxyprogesterone acetate (a progesterone analogue) exerts benefits for IBS treatment [71]. Additionally, Freedman and coworkers showed that higher levels of progesterone in mature women subjected to hormonal replacement therapy correlate with low incidence of esophagus and stomach adenocarcinomas, which may partially explain the higher prevalence rates of gastrointestinal tract cancers in men than in women [76]. In a similar way, a hormonal therapy based on the daily administration of ethinyl estradiol (30 µg) and noretisterone (1.5 mg) (both analogue molecules of estradiol and progesterone, respectively) is able to decrease chronic bleeding associated to gastric antral vascular ectasia in decompensated cirrhotic patients [77]. Moreover, high-turnover type osteoporosis (a common complication in patients with primary biliary cirrhosis) is significantly attenuated by hormonal replacement therapy without increasing risk of cholestasis [78]. Progesterone also has beneficial effects on Helicobacter pylori-associated gastritis in female ovarectomized gerbils, which showed a significant reduction in gastrin-positive cells [79]. In female Sprague-Dawley rats, administration of progesterone (300 µg/rat) reduced cysteamine-induced peptic ulcers apparently due to the property of this hormone to increase gastroduodenal mucus levels involved in long gastric protection [80]. Then, progesterone benefits are not only limited to improve a restrictive immune response against intestinal parasites, but it seems to have a major protective role during several human gastrointestinal pathologies, which highlights the possible use of progesterone and progesterone’s analogues as a promissory alternative treatment for management of gastric and esophagus carcinomas, intestinal inflammatory disease, gastritis, peptic ulcer and parasite-related intestinal infections, among others.

The evidence presented in our work illustrates the importance of immunoendocrine interactions in an immunocompetent host. It strongly suggests an important role for sex steroids, particularly progesterone, in the cytokine network. The complexity of the immunoendocrine interactions suggests that all physiological factors (i.e., sex, age, developmental stage) should be taken into consideration in the design of vaccines and new drugs. Interventions aimed at the hormonal network appear as a possible new therapeutic approach to control several immune confrontations, such as taeniosis/cysticercosis, as well as other related enteric disorders.

Taking into consideration the feasible role of progesterone on mucosal immunity, present results may open an interesting perspective in the possible use of sex steroid hormone analogues as adjuvants for anti-parasite vaccination, with strong immune actions but minimal endocrine effects, which could contribute to design different strategies for control intestinal helminth infections such as taeniosis by T. solium and in a nearer future other human gastrointestinal diseases.

Acknowledgments

Financial support was provided by grant IN 214011-3 from the Programa de Apoyo a Proyectos de Innovación Tecnológica, Dirección General de Asuntos del Personal Académico, Universidad Nacional Autónoma de México to J. Morales-Montor. G. Escobedo thanks financial support from Fundación Mexicana para la Salud, A. C., Grant no. 573-Estímulo Antonio Ariza Cañadilla. S. Leon-Cabrería has a doctoral fellowship from CONACYT at the Programa de Doctorado en Ciencias Biomédicas, UNAM.

Conflict of Interests

The authors have declared that no conflict of interest exists.

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