Effects of collagen crosslinking on porcine and human tarsal plate

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Abstract

Background: Floppy eyelid syndrome is a disorder in which the tarsal plate is easily distensible and is currently treated with conservative or surgical measures. Human tarsal plate contains type I collagen, which is crosslinked in corneal tissue as a treatment for keratoconus. We hypothesized that collagen crosslinking would similarly stiffen tarsal plate tissue and investigated this in porcine and human tarsal plate specimens.

Methods: Riboflavin-sensitized porcine and human tarsus samples were irradiated with ultraviolet-A light. Porcine experiments were analyzed with gross photographs, anterior segment optical computed tomography (AS-OCT) imaging, and tensile testing. A prospective study of human tarsus was performed on samples from patients undergoing wedge resection for floppy eyelid syndrome and was analyzed with AS-OCT and tensile testing.

Results: 73 porcine adnexa and 9 patients (16 eyelids) who underwent wedge excision were included in the study. Grossly, greater stiffness was observed in crosslinked porcine tissue. AS-OCT imaging in porcine tissue showed a distinct hyperreflective band in crosslinked specimens whose area and intensity increased with longer treatment time (P = 0.003); this band was also visible in crosslinked human specimens. Tensile testing was performed, but results were not statistically significant.

Conclusions: AS-OCT imaging, which has not been previously described for tarsal plate, showed a characteristic change in crosslinked porcine and human specimens. Tissue stiffness was increased grossly, but changes in tensile properties were not statistically significant. Further study is warranted to determine relevance as a potential treatment for floppy eyelid syndrome.

Keywords: Floppy eyelid syndrome, Crosslinking, Riboflavin

Background

Floppy eyelid syndrome (FES) is a disorder in which the upper eyelids are lax and easily distensible and was first described by Culbertson and Ostler in 1981 [1]. Clinically, it is characterized by upper eyelids that easily evert with traction, chronic papillary conjunctivitis, eyelash ptosis, blepharoptosis, and resultant keratopathy [2]. Chronic conjunctivitis is exacerbated by mechanical trauma to the eyelids during sleep, often to a greater degree on a patient's preferred sleeping side [2, 3]. FES most commonly affects middle-aged men and is strongly associated with obesity and obstructive sleep apnea (OSA) [4]. Additional associations have been reported with keratoconus, dermatochalasis, skin and joint hyperextensibility, hypertension, and developmental delay [2].

Many pathologic changes in FES eyelids have been described, and although nocturnal mechanical trauma and eye rubbing are thought to play a role, it remains unclear if these are the primary etiological factors in the development of FES [1, 4]. It has been hypothesized that a genetic abnormality in collagen, elastin, or both may predispose patients to the development of FES [5]. In addition, pressure induced eyelid ischemia and systemic reperfusion oxidation injury during sleep may contribute to the degenerative changes seen [6, 7].

Normal human tarsal plate provides the structural integrity of the upper eyelid and is not typical fibrous connective tissue or cartilage, but displays some features of...
both. It consists of fibroblasts surrounded by extracellular matrix, which contains primarily collagens type I and III, elastic fibers composed of elastin and microfibrils, proteoglycans, and glycoproteins concentrated around glandular acini [8, 9]. Several studies have demonstrated normal collagen structure and periodicity in FES [9–11]. However, there is evidence of type I and III collagen accumulation, which may be an adaptive response to repetitive loading of the tissue [9]. In addition, histopathologic studies of FES eyelids have revealed decreased quantity of elastic fibers, change in elastic fiber phenotype from mature elastic fibers to elaunin and oxytalan fibers, and increased levels of matrix metalloproteinases (MMP) that degrade elastic fibers [2, 9, 12, 13]. These changes may also be induced responses due to repetitive stress to the tissue [9]. Tarsal plate fibroblasts have a higher mechanostat set point and decreased sensitivity to mechanical stress, which is proposed to be an adaptive response to greater tissue stress levels in FES [14].

Current treatments for FES include conservative measures such as artificial tears, taping the eyelids closed or wearing a protective shield during sleep [13]. Some patients have improvement of ocular surface symptoms from FES after initiating continuous airway pressure (CPAP) therapy for OSA [15]. Surgical treatments include horizontal eyelid shortening and stabilization by wedge resection, plication, or lateral tarsal strip [2, 13, 16]. Despite these interventions, patients may have inadequate symptomatic relief and may require repeated surgical interventions [16]. An effective, minimally invasive method of increasing tarsal plate stiffness could prove useful either alone or in combination with current treatment methods.

Keratoconus is a corneal ectatic disorder in which progressive steepening of the cornea occurs, the normal parallel organization of collagen fibrils is disrupted, and elastic fibers are abnormally distributed [17]. Since it was first described in 2003, corneal collagen crosslinking has demonstrated ability to halt the progression of ectasia with promising long-term disease stability. It has been approved by the U.S. Food and Drug Administration for the treatment of keratoconus since 2016 [18]. Riboflavin, a photosensitizer applied directly to the corneal stromal collagen, is exposed to UVA (ultraviolet A) radiation, inducing bond formation between collagen fibers (primarily type I and III) and increasing corneal stiffness [19, 20]. In addition, a recent study demonstrated significant stiffening of sheep tarsal plate tissue following collagen crosslinking [21].

In this study, we investigated the effects of collagen crosslinking on porcine and human tarsal plate samples. Given the presence of type I and III collagen in both tarsal plate and corneal tissue, we hypothesized that collagen crosslinking may increase the stiffness of tarsal plate. If effective, tarsal collagen crosslinking could be further investigated as a minimally invasive treatment for FES, as no treatments currently exist to prevent pathophysiological progression of this disease.

Methods
Porcine studies were conducted using fresh porcine cadaveric adnexal tissue retrieved within 24 h postmortem from a slaughterhouse. The slaughterhouse was aware of the intended use of the porcine orbit specimens for medical research but was not asked to give specific consent for the study parameters nor was this required by the IRB who reviewed the study protocol. For human specimens, a prospective study was conducted of patients undergoing wedge excision for floppy eyelid syndrome between October 1, 2016 and April 1, 2018 in a single surgeon’s practice at the Wilmer Eye Institute. After giving written informed consent, patients undergoing unilateral or bilateral wedge excisions for floppy eyelid syndrome were included. The removal of eyelid tissue was in keeping with standards of clinical care, and the tissue would have otherwise been discarded. Patients with a history of prior upper eyelid surgery or with other indications for wedge excision such as neoplasm or trichiasis were excluded. The Johns Hopkins Institutional Review Board prospectively granted approval for this study (No 00116226), which followed the tenets of the Declaration of Helsinki. Informed consent was obtained from each patient for inclusion in the study, which was HIPAA-compliant.

Porcine collagen crosslinking
A total of 73 fresh porcine adnexa were used in this study. For each experiment, porcine upper eyelid tarsus was dissected free of surrounding tissues. The conjunctiva of porcine eyelids is thicker and less adherent to the tarsal plate than in human eyelids, and was dissected off the tarsal plate completely using blunt and sharp dissection with Stevens scissors. Each porcine adnexa contributed 1 upper eyelid tarsal plate to either the treatment or control group. As porcine adnexa were obtained from the supplier as a unit already removed from the animal, it was not possible to select a contralateral eyelid from the same animal as an internal control. A 30-min riboflavin pre-treatment was then completed for both treatment and control groups, with 1 drop of riboflavin solution applied every 5 min to the anterior surface of the dissected specimens. For the treatment samples, this was followed by collagen crosslinking under UVA light application (370 nm) at irradiance of 8–9 mW/cm² as previously described in prior experiments for corneal tissue for either 30 min or 60 min [22]. The control samples were covered with aluminum foil to block ambient UV light during this time period. During the entire irradiance period, 1 drop of riboflavin solution was
applied every 5 min to both specimen groups. Specimens were then stored at 4 °C in 200 uL of balanced salt solution, and either tensile testing or imaging was then performed as below.

In the conventional Dresden protocol for corneal cross-linking, the corneal epithelium is removed and 0.1% riboflavin in isotonic 20% dextran-T-500 is applied as the photosensitizing solution [18]. A preliminary porcine experiment was performed to determine if variations of the corneal crosslinking protocols would have differing effects on tarsal plate tissue. The Dresden protocol riboflavin solution as well as several variations were tested, including 0.1% riboflavin in hypotonic 20% dextran (Sigma-Aldrich, St. Louis, MO, USA), 0.1% riboflavin in hypotonic 20% dextran with 0.05% benzalkonium chloride (BAK) (Sigma-Aldrich, St. Louis, MO, USA), and 0.1% riboflavin in isotonic 20% dextran with 0.05% BAK (Fig. 1). BAK has been shown to increase riboflavin penetration and absorption through corneal tissue, and hypotonic riboflavin solution has been described for crosslinking in thin corneas [23, 24]. In addition, corneal crosslinking has been described with both epithelium intact or with epithelial debridement (“epithelium-off”), with a greater effect seen for epithelium-off crosslinking [25]. In the preliminary porcine experiment, conjunctiva was left in place for one group (“conjunctiva-on”) and was dissected off for a second group (“conjunctiva-off”) (Fig. 1). The 0.1% riboflavin in isotonic 20% dextran solution and conjunctiva-off procedure was selected for all further porcine experiments and for all human experiments.

**Human collagen crosslinking**

A total of 9 patients (16 eyelids) who underwent wedge excision were included in the study. Collagen crosslinking was performed on the same day as the patient’s surgery within 8 h of the time of specimen collection, with specimens stored in balanced salt solution at 4 °C until cross-linking was performed. The tarsal plate was dissected free from the surrounding tissues, and the skin and orbicularis were completely removed from the anterior surface. The tarsal conjunctiva of human eyelids is thinner and more adherent, so the posterior surface was debrided with a #15 blade to remove the epithelium. When bilateral specimens were collected, the contralateral side was used as the experimental control. Treatment was applied to the anterior surface of the tarsal plate specimens. The riboflavin solution used was 0.1% riboflavin in isotonic 20% dextran. Pre-treatment with riboflavin was performed for both control and treatment specimens with 1 drop riboflavin administered every 5 min for 30 min. The treatment specimens were then placed under the same UVA light as for porcine specimens for 30 min, and the control specimens were covered with aluminum foil. During the 30-min irradiation period, 1 drop of riboflavin was applied to both groups every 5 min. Specimens were stored at 4 °C in 200 uL of balanced salt solution, and either tensile testing or imaging was then performed within 48 h as below.

**Imaging and “flop” analysis**

Porcine tarsal plate specimens were dissected in the manner described above and cut to a standard size of 7 mm wide by 30 mm long. All specimens were stored at 4 °C until the time of dissection, and were kept at room temperature from the time of dissection to photographing which was approximately 1 h. Gross photographs of porcine tissue were taken using a tripod-mounted Nikon D7100 DSLR and 105 mm lens (Nikon, Melville, NY,
USA). Standardization of gross photographs for “flop” measurement was established using a horizontal metal clasp mounted on a wood frame with the axis of the clamp aligned to 180 degrees (Fig. 2a). The clasp end was positioned 2 mm from the end of the specimen. The 60 min treatment protocol was selected as this demonstrated the most significant changes (wider hyperreflective band) on OCT analysis. Photographs were taken before and immediately after treatment for 10 control specimens and 10 extended (60-min) treatment specimens. Angle of flop as degrees below 180 before and after treatment was measured accordingly from the photographs. The change in angle from before treatment to after treatment, or “delta flop,” was compared between control and 60-min treatment groups (Fig. 2b).

Anterior segment optical computed tomography (AS-OCT) of porcine and human tarsal plate specimens was performed within 48 h of crosslinking with the Zeiss Cirrus spectral domain HD-OCT device (Carl Zeiss Meditec, Dublin, CA, USA) using an anterior chamber lens. For porcine specimens, Adobe Photoshop (Adobe Systems, San Jose, CA, USA) was used to measure the area of the observed hyperreflective band with the following protocol (Fig. 3): The image coloring and tonality was standardized and compressed in two steps identically for all images: 1) the negative space of the scan was set to the black point of the image using the Levels tool and 2) the midtone point was set to 0.25. The image was then inverted and the Find Edges filter was applied to highlight the boundary of the hyperreflective band. In the central 150 pixel wide area of each scan, the Cluster Pixel Selection tool (tolerance 20) was used to highlight the areas of the hyperreflective band (Fig. 3, red) and the homogenous layer of the scan (Fig. 3, yellow). The area edges were smoothed with the Selection Smooth function with a factor of 2 and then the Measurement panel was used to calculate the area, average height, greyscale brightness, mean, minimum and maximum for each band. Comparison was then made between the 30- and 60-min treatment groups for the height and area of the hyperreflective band and the greyscale differences within specimens.

![Fig. 2 Gross Photographs and “Flop”. (a) Gross photographs of porcine tarsal plate specimens using a standardized metal clamp mounted in a fixed location to the camera and a registration image. Images were auto-aligned via position in Adobe Photoshop using the registration marks and then exposure corrected for contrast. Pre-crosslinking treatment and post-crosslinking treatment photographs are shown for representative control (C2), treatment specimen with least “delta flop” (T1), and two treatment specimens with greatest “delta flop” (T4 and T9). (b) Composite image of all photographed porcine tarsal plate specimens (10 control, 10 60-min treatment) using a standardized metal clamp mounted in a fixed location to the camera and a registration image, demonstrating a change in “flop” for the post-treatment group. All tested control and treatment specimen images were auto-aligned via position in Adobe Photoshop using the registration marks and then exposure corrected for contrast. The overlay image was created using the Layer Difference Blending Mode. *P < 0.0001](image-url)
for the two measured areas (red and yellow). For human specimen AS-OCT, no measurements or adjustments were made (Fig. 4).

**Tensile testing**

Porcine and human specimens underwent collagen crosslinking with the 30-min treatment protocol as above. Uniaxial tensile testing was then performed using a MTS Criterion 43 Tensile Tester (MTS, Eden Prairie, MN, USA). Pig and human tarsal samples were each cut into uniform 5 mm strips. Tissue thickness and central width for each strip was measured using a Vernier caliper (Mitutoyo, Japan) and entered into the software. The ends of individual tarsal sample strips were then placed between sandpaper and affixed using super glue to minimize slipping during the tensile test. Strips were placed in the metal clamps of the tensile tester with a sample test length of 5 mm. Pre-loading was done at 5% as is standard for soft tissue. Samples were uniaxially stretched at 0.2 mm/s until failure using a 100 N load cell. Load/displacement measurements were recorded and analyzed. Tensile strength as indicated by peak stress and Young’s elastic modulus were measured.

**Statistical analyses**

Statistical analyses were performed using SAS software. Statistical differences of means were calculated using student’s t-test with P-values less than 0.05 considered significant.

**Results**

In the preliminary porcine experiment, several different variations of riboflavin solution were tested as well as conjunctiva-off and conjunctiva-on procedures. Gross photographs taken before and after crosslinking showed a clear change in tissue stiffness, with greater stiffness of treatment specimens as compared to control specimens (Fig. 1). There was apparently greater effect in the conjunctiva-off group.

**Fig. 3** Porcine AS-OCT. Anterior segment optical computed tomography (AS-OCT) of control, 30-min treatment, and 60-min treatment porcine specimens. The entire OCT for each specimen is shown, with unadjusted raw image on the right; black and midtone standardization, image inversion, and Adobe Photoshop Find Edges tool applied on the left; and reinverted image with the two measured areas shown in center, with the hyperreflective band highlighted in red and the deeper tissue highlighted in yellow. The area of the hyperreflective band (center panel, red) was significantly wider in 60-min treatment specimens compared to 30-min treatment specimens ($P = 0.003$).

**Fig. 4** Human AS-OCT. AS-OCT of bilateral upper eyelid human tarsal plate specimens from 3 patients with floppy eyelid syndrome demonstrating a hyperreflective band in the treatment specimens after a 30-min collagen crosslinking protocol. The hyperreflective band is not observed in the un-treated contralateral eyelid tarsal plate specimens, which served as controls.
For further porcine studies and all human studies, the Dresden protocol riboflavin solution of 0.1% riboflavin in isotonic 20% dextran solution was selected, and the conjunctiva-off procedure was used. In standardized gross photographs of control specimens and 60-min treatment specimens mounted on a horizontal clamp (Fig. 2a), the change in specimen angle before and after crosslinking treatment (“delta flop”) was significantly different between control (mean – 7.8 degrees) and treatment (mean 16.4 degrees) specimens ($P < 0.0001$, Fig. 2b).

AS-OCT imaging of 15 porcine specimens demonstrated the appearance of a characteristic hyperreflective band at the anterior surface of 30-min and 60-min treatment specimens that was not present in controls (Fig. 3). Intraspecimen assessment of greyscale intensity within the hyperreflective band relative to the deeper tissue confirmed this band was brighter in maximum intensity ($P = 0.014$) and average intensity ($P = 0.039$) in the 60-min compared to the 30-min treatment groups. The area of the band was significantly greater in the 60-min compared to the 30-min treatment group ($P = 0.003$, Fig. 3). There was a trend towards greater height of the band in the 60-min treatment group (Fig. 3). The measured change in “flop” was significantly different between control and treatment groups, with treatment specimens demonstrating smaller angles of variation from 180 degrees after treatment, presumably due to greater tissue stiffness (Fig. 2b).

We demonstrated validity of our tarsal crosslinking protocol using AS-OCT imaging, which to our knowledge has not been previously described for tarsal plate tissue. We observed a hyperreflective band at the anterior surface of crosslinked porcine specimens (Fig. 3). This band was present in 30-min treatment specimens, and its area was significantly greater ($P = 0.003$) and average intensity significantly brighter ($P = 0.039$) in 60-min treatment specimens, suggesting time-dependent correlation of the hyperreflective band with crosslinking treatment. A similar hyperreflective band was also observed in human tissue (Fig. 4).

For tensile testing in porcine tissue, 13 control and 13 treatment specimens were tested. In human tissue, 4 control specimens and 6 treatment specimens were tested (Table 1). There was no significant difference in the Young’s modulus for either porcine or human tissue ($P = 0.90$ and 0.36, respectively).

**Discussion**

Given the current lack of minimally invasive, curative interventions for FES, our goal was to investigate collagen crosslinking as a putative method for increasing tarsal plate stiffness in both porcine and human specimens.

In our study, the crosslinking protocols used are comparable to accepted protocols for corneal crosslinking. Multiple biomechanical experiments have shown that corneal collagen crosslinking increases intra- and interfibrillar collagen bonds, thereby enhancing corneal stiffness as demonstrated by an increased Young’s elastic modulus (ratio of stress over strain) [18]. Our preliminary studies using ex vivo porcine tarsal plate demonstrated a gross difference in stiffness between control and treatment specimens, with treatment specimens having greater stiffness. However, these macroscopic changes were difficult to quantify, and gross photographs are clearly limited by numerous factors such as a lack of standardization of tissue alignment within the forceps and photography angle. We aimed to standardize the gross photographs using a metal clamp mounted in a fixed location relative to a tripod-mounted camera and a registration image (Fig. 2a). The measured change in “flop” was significantly different between control and treatment groups, with treatment specimens demonstrating smaller angles of variation from 180 degrees after treatment, presumably due to greater tissue stiffness (Fig. 2b).

We demonstrated validity of our tarsal crosslinking protocol using AS-OCT imaging, which to our knowledge has not been previously described for tarsal plate tissue. We observed a hyperreflective band at the anterior surface of crosslinked porcine specimens (Fig. 3). This band was present in 30-min treatment specimens, and its area was significantly greater ($P = 0.003$) and average intensity significantly brighter ($P = 0.039$) in 60-min treatment specimens, suggesting time-dependent correlation of the hyperreflective band with crosslinking treatment. A similar hyperreflective band was also observed in human tissue.

**Table 1 Tensile Testing**

| Stress-strain measures | Control samples | Treatment samples | Two-tailed $P$-value |
|------------------------|-----------------|-------------------|---------------------|
|                        | Mean            | SD                | Mean                | SD                |                     |
| **Porcine tarsal experiments** |                 |                   |                     |                   |                     |
| Peak Load (N)          | 28.53           | 12.01             | 32.75               | 14.25             | 0.42               |
| Peak Stress (MPa)      | 4.19            | 2.00              | 5.31                | 3.04              | 0.28               |
| Strain at break point (in/in) | 1.70          | 1.52              | 3.02                | 2.54              | 0.16               |
| Young’s modulus (MPa)  | 8.60            | 7.51              | 8.99                | 7.72              | 0.90               |
| Number of Samples      | 13              |                   | 13                  |                   |                     |
| **Human tarsal experiments** |                 |                   |                     |                   |                     |
| Peak Load (N)          | 10.43           | 1.42              | 11.13               | 1.96              | 0.54               |
| Peak Stress (MPa)      | 2.15            | 0.47              | 2.82                | 1.27              | 0.28               |
| Strain at break point (in/in) | 0.48          | 0.07              | 0.49                | 0.16              | 0.82               |
| Young’s modulus (MPa)  | 6.32            | 2.23              | 7.82                | 2.61              | 0.36               |
| Number of Samples      | 4               |                   | 6                   |                   |                     |

Tensile testing in porcine and human specimens shows no significant difference in the Young’s modulus or peak stress for either group ($P > 0.05$)
crosslinking would allow a surgical incision to be conjunctiva off. A posterior approach to the eyelid for crease incision would allow crosslinking treatment with surgical approach to the tarsal plate through an eyelid linked with conjunctiva dissected off. In vivo, an anterior size constraints of the available tensile testing apparatus.

Also, tarsal plate specimens in this study were cross-linked with conjunctiva dissected off. In vivo, an anterior floppiness with gross photographs (“flop”), but tensile testing did not demonstrate a significant difference in tensile properties. Further evaluation with tensile testing is necessary with larger sample sizes. Should future studies continue to demonstrate increased biomechanical stiffness as a result of tarsal collagen crosslinking, investigation of this procedure in humans as a minimally invasive treatment for FES may be warranted.

Conclusions

Altogether, our results imply efficacy of tarsal crosslinking in increasing stiffness in both porcine tarsus and human tarsus from FES patients. Tarsal collagen crosslinking in porcine and human tissue has not been previously described, so our study provides proof-of-principle for a novel treatment protocol for FES. We also describe a unique use of AS-OCT for tarsal plate imaging and potentially for monitoring tarsal crosslinking safety and efficacy. Our results demonstrated a clear difference in tissue stiffness with gross photographs (“flop”), but tensile testing did not demonstrate a significant difference in tensile properties. Further evaluation with tensile testing is necessary with larger sample sizes. Should future studies continue to demonstrate increased biomechanical stiffness as a result of tarsal collagen crosslinking, investigation of this procedure in humans as a minimally invasive treatment for FES may be warranted.

Abbreviations

AS-OCT: Anterior segment optical computed tomography; BAK: Benzalkonium chloride; CPAP: Continuous positive airway pressure; FES: Floppy eyelid syndrome; MMP: Matrix metalloproteinases; OSA: Obstructive sleep apnea; SD: Standard deviation; UVA: Ultraviolet A.

Acknowledgements

Not applicable.

Authors’ contributions

SD, AZ, AJ, JE and NM contributed to conception of ideas and design of study. SD and NM collected human specimens. SD, AZ and NM participated in porcine and human crosslinking experiments. SD and NM were major contributors in writing the manuscript and designing figures. SD, NM, SM, and AZ were major contributors in manuscript revision. JT and NM analyzed and interpreted the data from all experiments. SM completed the tensile testing experiments and helped in analyzing data. All authors read and approved the final manuscript.

Funding

This study was funded by personal discretionary research funds of the senior author, NM. Funds were only used to purchase porcine specimens and for disposable supplies.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.
9. Ezra DG, Ellis JS, Gaughan C, Beaconsfield M, Collin R, Bunce C, et al.
8. Milz S, Neufang J, Higashiyama I, Putz R, Benjamin M. An
7. Ezra DG, Beaconsfield M, Sira M, Bunce C, Wormald R, Collin R. The
6. Culbertson WW, Tseng SC. Corneal disorders in floppy eyelid syndrome.
5. Lee WJ, Kim JC, Shyn KH. Clinical evaluation of corneal diseases associated
4. Fowler AM, Dutton JJ. Floppy eyelid syndrome as a subset of lax eyelid
3. Schwartz LK, Gelender H, Forster RK. Chronic conjunctivitis associated with
2. Schlotzer-Schrehardt U, Stojkovic M, Hofmann-Rummelt C, Cursiefen C,
1. Culbertson WW, Ostler HB. The floppy eyelid syndrome. Am J Ophthalmol.

References
1. Culbertson WW, Ostler HB. The floppy eyelid syndrome. Am J Ophthalmol. 1981;92:658–75.
2. Schlotzer-Schrehardt U, Stojkovic M, Hofmann-Rummelt C, Cursiefen C, Kruse FE, Holbach LM. The pathogenesis of floppy eyelid syndrome: involvement of matrix metalloproteinases in elastic fiber degradation. Ophthalmology. 2005;112:694–704.
3. Schwartz LK, Gelender H, Forster RK. Chronic conjunctivitis associated with ‘floppy eyelids’. Arch Ophthalmol. 1983;101:1884–8.
4. Fowler AM, Dutton JJ. Floppy eyelid syndrome as a subset of lax eyelid conditions: relationships and clinical relevance (an ASOPRS thesis). Ophthal Plast Reconstr Surg. 2010;26:195–204.
5. Lee WJ, Kim JC, Shyn KH. Clinical evaluation of corneal diseases associated with floppy eyelid syndrome. Korean J Ophthalmol. 1996;10:116–21.
6. Culbertson WW, Tseng SC. Corneal disorders in floppy eyelid syndrome. Cornea. 1994;13:33–42.
7. Ezra DG, Beaconsfield M, Sira M, Bunce C, Wormald R, Collin R. The associations of floppy eyelid syndrome: a case control study. Ophthalmology. 2010;117:831–8.
8. Mitz S, Neufang J, Higashiyama I, Putz R, Benjamin M. An immunohistochemical study of the extracellular matrix of the tarsal plate in the upper eyelid in human beings. J Anat. 2005;206:37–45.
9. Ezra DG, Ellis JS, Gaughan C, Beaconsfield M, Collin R, Bunce C, et al. Changes in tarsal plate fibrillar collagens and elastic fibre phenotype in floppy eyelid syndrome. Clin Exp Ophthalmol. 2011;39:564–71.
10. Goldberg R, Seiff S, McFarland J, Simons K, Shorr N. Floppy eyelid syndrome and obstructive sleep apnoea. Clin Exp Ophthalmol. 2000;28:125–6.
11. Arocker-Mettinger E, Haddad R, Konrad K, Steinigkogler FJ. Floppy eyelid syndrome: light and electron microscopy studies. Klin Monatsbl Augenhilkd. 1986;188:596–8.
12. Netland PA, Sugrue SP, Albert DM, Shore JW. Histopathologic features of the floppy eyelid syndrome. Involvement of tarsal elastin. Ophthalmology. 1994;101:174–81.
13. Ezra DG, Beaconsfield M, Collin R. Floppy eyelid syndrome: stretching the limits. Surv Ophthalmol. 2010;55:35–46.
14. Ezra DG, Ellis JS, Beaconsfield M, Collin J, Bailly M. Changes in fibroblast mechanostat set point and mechno-sensitivity suggest an adaptive response to mechanical stress in floppy eyelid syndrome. Invest Ophthalmol Vis Sci. 2010;51:1885–63.
15. McNab AA. Reversal of floppy eyelid syndrome with treatment of obstructive sleep apnoea. Clin Exp Ophthalmol. 2000;28:125–6.
16. Ezra DG, Beaconsfield M, Sira M, Bunce C, Shah-Desai S, Verty DH, et al. Long-term outcomes of surgical approaches to the treatment of floppy eyelid syndrome. Ophthalmology. 2010;117:839–46.
17. White TL, Lewis PN, Young RD, Kitazawa K, Inatomi T, Kinoshita S, et al. Elastic microfibril distribution in the cornea: differences between normal and keratoconic stroma. Exp Eye Res. 2017;159:40–8.
18. Wollensak G, Spoerl E, Seiler T. Riboflavin/ultraviolet-a-induced collagen crosslinking for the treatment of keratoconus. Am J Ophthalmol. 2003;135(5):620–7.
19. Sarkin N, Varsano D. Corneal collagen crosslinking: a systematic review. Ophthalmologica. 2014;232:210–27.
20. Snibson GR. Collagen cross-linking: a new treatment paradigm in corneal disease - a review. Clin Exp Ophthalmol. 2010;38:141–53.
21. Smith TM, Suzuki S, Cronin BS, Haghhighatparah M, Petcu EB, Philippa CJ, et al. Photochemically induced crosslinking of tarsal collagen as a treatment for eyelid laxity: assessing potentiality in animal tissue. Ophthalmic Plast Reconstr Surg. 2018;34:477–82.
22. Zhu CY, Viana LM, Borkenstein EM, Ellisseff J, Jun AS. Assessment of a novel corneal-shaping device with simultaneous corneal collagen cross-linking using a porcine eye model. Cornea. 2016;35:114–21.
23. Kissner A, Spoerl E, Jung R, Spek K, Pillunat LE, Raiskup F. Pharmacological modification of the epithelial permeability by benzalkonium chloride in UVA/riboflavin corneal collagen cross-linking. Curr Eye Res. 2010;35:715–21.
24. Rosenblat E, Hersh PS. Intraoperative corneal thickness change and clinical outcomes after corneal collagen crosslinking: standard crosslinking versus hypotonic riboflavin. J Cataract Refract Surg. 2016;42:605–609.
25. Mastropasqua L. Collagen cross-linking: when and how? A review of the state of the art of the technique and new perspectives. Eye Vis (Lond). 2015;2:19.
26. Seiler T, Hafezi F. Corneal cross-linking-induced stromal demarcation line. Cornea. 2006;25:1057–9.
27. Konstantopoulos A, Mehta JS. Conventional versus accelerated collagen cross-linking for keratoconus. Eye Contact Lens. 2015;41:65–71.

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