Understanding the correlation between structure and dynamics of clocortolone pivalate by solid state NMR measurement

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Structural characteristics of clocortolone pivalate are unique in the topical corticosteroid field having high penetration power through the stratum corneum of skin as well as low corticosteroid-related adverse effects. The molecule was thoroughly studied by $^{13}$C 2DPASS CP MAS NMR and spin–lattice relaxation time measurements. Molecular correlation time at different carbon nuclei positions was calculated by assuming that the chemical shift anisotropy interaction and heteronuclear dipole–dipole interaction play vital roles in the $^{13}$C spin–lattice relaxation mechanism. The CSA parameters are substantially varied at different carbon nuclei sites. This suggests that the electronic distribution surrounding the carbon nuclei varies widely when the same carbon atom is placed in different chemical surroundings of the molecule. The spinning CSA sideband pattern for C11, C17, C26 nuclei is axially symmetric. The asymmetry parameter is very small ($\leq 0.3$) for C2, C5, C10, C22, C23, C24 nuclei, and it is reasonably high ($\geq 0.9$) for C3, C4, C6, C18, C19, C21 nuclei. The anisotropy parameter is very high for double bonded C14, C15, C18, C19, C21, C22, and C23 nuclei. Spin–lattice relaxation time and molecular correlation time are also varied substantially for carbon nuclei situated at various positions of the molecule. The spin–lattice relaxation time is slow for carbon nuclei residing at the carbon ring, and it is very fast for C12, C17, C16, C26 carbon nuclei situated at the side portion of the molecule. Molecular correlation time is of the order of $10^{-4}$ s for those carbon nuclei attached with neighbouring carbon or oxygen atoms by double bonds like C14, C15, C18, C19, C21, C22, and C23. It implies that the molecular correlation time is very high for those carbon nuclei associated with high values of the chemical shift anisotropy parameter. In contrast, the molecular correlation time is of the order of $10^{-8}$ s for C12, C16, and C17 carbon nuclei. From these studies, it is clear that the various portions of the molecule exhibit different degrees of motion and the dynamics is related with the structural characteristics of the molecule. These investigations on important drug clocortolone pivalate by solid state NMR will help researchers to understand the structure and dynamics of the molecule, which will give a direction to develop advance corticosteroids.

1. Introduction

Introduction of topical glucocorticoids has led marked improvement in the outcomes of several skin diseases. For many years, they have played major roles in the treatment of different dermatoses sensitive to corticosteroids. Skin diseases like atopic dermatitis, contact dermatitis, psoriasis vulgaris, seborrheic dermatitis, etc., can be treated. They are also used for the treatment of different eczematous skin disorders from different age groups of the human population, including children. Topical corticosteroids are classified based on various potency values from class 1, most potent to least potent, class 7 based on their different skin permeation abilities, activity at the target and rate of removal from the site of application. The selection of a specific class of topical corticosteroid is influenced by its sensitivity to the disease and its penetration power to reach the site of the disease. The high potency or super high potency corticosteroids are mainly used for the treatment of Chronic Plaque Psoriasis (CPP) and lichenified eczematous plaques. The mid potency (class 4 to 5) topical corticosteroids are applied for the treatment of various common dermatological disorders. Clocortolone pivalate is one of the widely used mid-potency (class 4) topical corticosteroid used in the treatment of different skin diseases. The use of corticosteroids in skin diseases is determined by their distinctive biological properties like anti-inflammatory, anti-miotic, immunosuppressive action and their ability to inhibit the production of connective tissue molecules. Although topical corticosteroids lack antipruritic activity, they can function by reducing the inflammation of skin disorders and thus, they decrease indirectly the itching of different skin disorders.
In general, the basic structure of topical corticosteroid molecules consists of 21-carbon atoms and an additional 2-carbon side chain at C5. There are three 6-carbon rings and one 5-carbon ring in its core structure. The basic structure of corticosteroid is frequently modified to enhance the glucocorticoid receptor binding affinity, lipophilicity and ability of cutaneous penetration.\textsuperscript{8} Clocortolone pivalate is a nice example where different structural modifications have improved its properties.\textsuperscript{13,14} Clocortolone pivalate is 4-chloro-13α-fluoro-11β, 21-dihydroxy-16α methylpregna-1,4-diene-3, 20-dione 21-pivalate. In the molecule, esterification was introduced at C20 position with a pivalate group to increase lipophilicity and decrease metabolic breakdown. Methylation at the C8 position is done to decrease various allergic reactions of corticosteroids.\textsuperscript{10} Moreover, halogenation by fluorine and chlorine at C13 and C4 position respectively, increases the efficacy and safety profile of this mid potency topical corticosteroid.\textsuperscript{19} Due to its unique structure having high efficacy and low side effects it is widely used as a 0.1% cream for the treatment of many skin diseases. Several clinical trials have been performed on clocortolone pivalate 0.1% cream to prove its efficacy and safety and it is eminently suitable for human use with a relative lack of adverse effects.\textsuperscript{13–16} One of these trials includes 559 adult and paediatric volunteers, who have undergone active treatment of clocortolone pivalate cream. Overall no systemic adverse effects were reported from the study. Hence, it can be concluded that clocortolone pivalate is highly bio-available for local activity in the skin after its topical application.\textsuperscript{17–24}

Chemical shift anisotropy (CSA) is a sensitive probe of detecting the local electronic environment, three-dimensional structural details surrounding the nucleus, and the degree of motion of the molecule. The broadening of spectral line-width due to the presence of various anisotropic interactions makes it complicated to determine the values of CSA parameters from static solid state NMR spectra. In magic angle spinning (MAS) experiment all the information encoded in anisotropic interactions are lost. However, the principal values of CSA tensor can be reconstructed by - two-dimensional MAS/CSA NMR experiment;\textsuperscript{25} SUPER (separation of undistorted powder patterns by effortless recoupling) MAS NMR;\textsuperscript{27} ROCSA (recoupling of chemical shift anisotropy) pulse sequence at MAS frequencies 11 to 20 kHz;\textsuperscript{28} RNCSA (γ-encoded RF\textsuperscript{δ}-symmetry based chemical shift anisotropy) schemes for the system associated with weak homonuclear dipole–dipole interactions;\textsuperscript{29} 2DMAF (two-dimensional magic angle flipping) experiment;\textsuperscript{30–32} 2DMAT (two-dimensional magic angle turning) experiment;\textsuperscript{33} 2DPASS (two-dimensional phase adjusted spinning side-band) MAS (magic angle spinning) SSNMR experiment.\textsuperscript{34–36} The CSA parameters of glass compounds,\textsuperscript{36} biopolymers,\textsuperscript{37–40} antifungal drug itraconazole\textsuperscript{41} were extracted by applying 2DPASS MAS NMR experiments. The purpose of the current work is to elucidate the structural details and spin dynamics of clocortolone pivalate by extracting principal values of CSA parameters by two-dimensional phase-adjusted spinning side-band (2DPASS) magic-angle-spinning (MAS) nuclear magnetic resonance (NMR) experiment, to determine the site-specific spin-lattice relaxation time by Torchia CP method,\textsuperscript{25} and to calculate of molecular correlation time at chemically distinct carbon sites by assuming that the chemical shift anisotropy interaction and heteronuclear dipole–dipole coupling play predominant role in spin–lattice relaxation mechanism. This type of spectroscopic studies is necessary for confounding structure–dynamic relationships of steroids, which has enormous applications in pharmaceutical industry.

2. Experiment section

2.1 NMR-measurements

An active ingredient of clocortolone pivalate from Sigma Aldrich was used for solid state NMR experiments. All solid state NMR experiments were carried out on a JEOL ECX 500 NMR Spectrometer associated with a 3.2 mm JEOL double resonance MAS probe, operating at a resonance frequencies of 500 MHz for \textsuperscript{1}H and 125.721 MHz for \textsuperscript{13}C. The magic angle spinning (MAS) speed was 10 kHz for \textsuperscript{13}C CP-MAS and Torchia CP experiments. The cross-polarization (CP) condition was maintained at contact time 2 ms, and SPINAL-64 \textsuperscript{1}H decoupling. \textsuperscript{13}C-spin lattice relaxation experiment was performed by following Torchia CP method.\textsuperscript{25}

2.2 CSA measurements

2DPASS MAS SSNMR experiment generates an isotropic spectrum in direct-dimension and an anisotropic spectrum in indirect dimension. It can correlate the isotropic dimension with anisotropic dimension.\textsuperscript{16,41–50,52} As the parameters of second rank CSA tensor corresponding to two distinct chemical environments are different, hence, with the help of this experiment, one can also identify two overlapping signals.\textsuperscript{52} Herzfeld and Berger\textsuperscript{41} had shown that the sideband intensities of spinning CSA sideband pattern of different nuclei are related with the CSA parameters and they had introduced a graphical method for evaluating the CSA eigenvalues.

In 1995, Antzutkin et al.\textsuperscript{45} had established the pulse scheme of 2DPASS MAS NMR experiment using sequence of five π pulses. The total time duration of all the PASS sequences containing five π pulses is independent of the sideband phase shift. The time evolution of five π pulses is varied during the 2D PASS experiment according to the PASS equations. The 2DPASS experiments were carried out at two different values of MAS frequencies 600 Hz and 2000 Hz.

3. Results and discussion

3.1 Spin–lattice relaxation measurements

Fig. 1(a) shows the structure of clocortolone pivalate is associated with three 6-carbon rings and one 5-carbon ring. Fig. 1(b) represents \textsuperscript{13}C-CP-MAS-NMR spectrum of clocortolone pivalate. Fig. 2 shows \textsuperscript{13}C spin–lattice relaxation decay curves of clocortolone pivalate at (a) C1, (b) C9, (c) C13, and (d) C27 carbon nuclei positions. Table 1 shows the spin–lattice relaxation time of numerous carbon nuclei is varying substantially. For C5 it is 184 s and for C12, C17, C16, C26 carbon nuclei it values reduces to 1 s. At the position of C20, where the esterification was
incorporated with the molecule by introducing a pivalate group for increasing lipophilicity and decreasing metabolic breakdown, the spin–lattice relaxation time is 175 s. At C8 position, where methylation is introduced to decrease various allergic reactions of corticosteroids, the value of spin–lattice relaxation is 138 s. At the positions of C4 and C13 halogenations was done by chlorine and fluorine atoms to increase the efficacy and safety profile of the drug. The spin–lattice relaxation time of C4

Fig. 1 (a) Molecular structure of clocortolone pivalate. (b) $^{13}$C CP MAS NMR spectrum of clocortolone pivalate.
and C13 are respectively 94 s and 171 s. Spin–lattice relaxation time is considerable high for those carbon nuclei reside at the carbon rings (there are three 6-carbon rings and one 5-carbon ring in the molecule) and it is comparatively lower for carbon nuclei attached at the side of the molecule like C12, C17, C16, C26 (as shown in Fig. 1(a)). The dipole–dipole interaction of those carbon nuclei resides at the side of the molecule is highly influenced by the internal motion and rotational motion of

Fig. 2. $^{13}$C spin–lattice relaxation decay curve of (a) C1, (b) C9, (c) C13, (d) C27. (e) shows the spin–lattice relaxation time is different at chemically distinct carbon sites of clocortolone pivalate.
molecule about its symmetry axis. That’s why the degree of motion is higher for those carbon nuclei.

3.2 Chemical shift anisotropy

Chemical-shift anisotropy interaction has a great role in nuclear spin relaxation mechanism. Hence, the accurate determination of CSA tensor can provide a plenty of information about molecular structure, molecular conformation and spin dynamics. In principal axis system (PAS) off-diagonal components of CSA tensor are cancel out and three diagonal terms survive. The expression of those diagonal components of chemical shift anisotropy tensor (δ11, δ22 and δ33) are given by

\[ \delta_{33} = \frac{e^2 c^2}{2m} \left( \frac{1}{r^3} \right) \delta \left( \frac{\mathbf{L} \cdot \mathbf{r}}{E_n - E_0} \right) \]

\[ \times \sum_n \left\{ \left( \frac{0}{E_n - E_0} \right) \mathbf{L} \cdot \mathbf{r} \right\}^2 \]

\[ \delta_{11} = \frac{e^2 c^2}{2m} \left( \frac{1}{r^3} \right) \delta \left( \frac{0}{E_n - E_0} \right) \]

\[ \times \sum_n \left\{ \left( \frac{0}{E_n - E_0} \right) \mathbf{L} \cdot \mathbf{r} \right\}^2 \]

\[ \delta_{22} = \frac{e^2 c^2}{2m} \left( \frac{1}{r^3} \right) \delta \left( \frac{0}{E_n - E_0} \right) \]

\[ \times \sum_n \left\{ \left( \frac{0}{E_n - E_0} \right) \mathbf{L} \cdot \mathbf{r} \right\}^2 \]

where \( L = L_x \hat{i} + L_y \hat{j} + L_z \hat{k} \) is the angular momentum. The first part of the above three equations regarding principal components of CSA tensor is generated by electrons resides in the spherically symmetric ‘s’ orbital. The second term arise when the electrons are in ‘p’ or ‘d’ orbital. The isotropic component of chemical shift \( (\delta_{iso} = \delta_{11} + \delta_{22} + \delta_{33}) \) represents the average values of the principal components of CSA tensor along three orthogonal directions. Basically, it is the position of the centre of gravity of the spinning CSA sideband pattern. The breadth of the CSA tensor is getting affected due to the change of the centre of gravity of the spinning CSA pattern. Small changes in centre of gravity of CSA pattern is generally associated with larger change in chemical shift anisotropy. According to Haebenerl convention \( \eta = \frac{\delta_{22} - \delta_{11}}{\delta_{33} - \delta_{11}} \) The deviation of the spinning CSA side band pattern from its axially symmetric shape is measured by the asymmetry parameter. The sign of asymmetry parameter indicates the direction of largest separation with respect to the centre of gravity of CSA pattern. ‘Skew’ \( \left( k = \frac{3(\delta_{22} - \delta_{iso})}{\Omega} \right) \) says the amount of orientation of the asymmetry pattern. The width of the sideband pattern is called ‘span’ \( (\Omega = \delta_{11} - \delta_{33}) \).

The spinning CSA sideband pattern becomes axially symmetric, when \( \delta_{22} \) is equal to \( \delta_{11} \) or \( \delta_{33} \). Table 2 says that the asymmetry parameter is zero for C11, C17, C26, it is very small (≈0.3) for C2, C5, C10, C22, C23, C24, and very large (≈0.9) for C3, C4, C6, C18, C19, C21. When it is zero, the CSA pattern is called axially symmetric. On the contrary, the CSA pattern is highly asymmetric, when the asymmetry parameter is very large.

Fig. 3 shows \(^{13}\text{C} 2\text{DPASS CP MAS NMR spectrum of clocortolone pivalate. The direct-dimension and indirect-dimension of two-dimensional spectrum respectively represents the isotropic spectrum and anisotropic spectrum. The spinning CSA sideband patterns of (a) C14, (b) C22, (c) C26, (d) C13, (e) C6, (f) C4, (g) C20, (h) C18, (i) C9 carbon nuclei are also shown in Fig. 3. The different values of CSA parameters at
the magnetic
quently, there originate an anisotropic susceptibility parallel to
along three directions in principal axes system (PAS). Conse-
of magnetic anisotropy for carbonyl group is according to Haeberlen convention
behind the large value of anisotropy of carbonyl group carbon.

Another reason is magnetic anisotropy, which arise due to three
and the molecular conformation and orientation are dissimilar
distribution of electron density surrounding the carbon nuclei,
neighbouring atoms by double bond. For carbonyl group
C14, C15, C18, C19, C21, C22, and C23 nuclei, connected with
miscellaneous values of magnetic susceptibilities
Another reason is magnetic anisotropy, which arise due to three
different directions. This is equally considered as reason
field is getting influenced by the action of the polar-
ization. The polarization is induced on the non spherical
distribution of charges surrounding the carbon nuclei due to
the electrostatic interaction. The strength of the induced
magnetic field is getting influenced by the action of the polar-
Consequently, the induced magnetic field is varied
different directions. This is equally considered as reason
behind the large value of anisotropy of carbonyl group carbon.
Another reason is magnetic anisotropy, which arise due to three
miscellaneous values of magnetic susceptibilities ($\chi_x$, $\chi_y$, $\chi_z$)
along three directions in principal axes system (PAS). Conse-
sequently, there originate an anisotropic susceptibility parallel to
the magnetic field ($\Delta \chi || = \chi_z - \chi_x$) and perpendicular to
the magnetic field ($\Delta \chi \perp = \chi_x - \chi_z$). The McConnell equation
of magnetic anisotropy for carbonyl group is

$$\delta_{\text{anis}} = \{ \Delta \chi (3 \cos^2 \theta_1 - 1) + \Delta \chi (3 \cos^2 \theta_2 - 1) \} / 3R^3$$  \hspace{1cm} (4)

where $\theta_1$ is the angle between the radius vector and $x$-axis, $\theta_2$ is
the angle between the radius vector and $z$-axis. Effective
magnetic field experienced by the nucleus varies along three
orthogonal directions because of these anisotropic magnetic
susceptibilities.

Table 2: The CSA parameters at structurally and chemically distinct carbon sites of clocortolone pivalate

| Carbon from different chemical environment with isotropic chemical shift ($\delta_{\text{iso}}$) (ppm) | $\delta_{11}$ | $\delta_{22}$ | $\delta_{33}$ | Span (ppm) | Skew | Anisotropy | Asymmetry |
|---|---|---|---|---|---|---|---|
| 67.5(C1) | 87.9 | 72 | 42.6 | 45.3 | 0.3 | $-37.3$ | 0.6 |
| 52.3(C2) | 82.7 | 41.8 | 32.3 | 50.4 | $-0.6$ | $45.6$ | 0.3 |
| 42.9(C3) | 54.5 | 43.3 | 30.9 | 23.5 | 0.05 | $-17.9$ | 0.9 |
| 88(C4) | 113 | 87.1 | 64 | 49 | $-0.06$ | $37.5$ | 0.9 |
| 75.7(C5) | 106.7 | 65.3 | 55.2 | 51.5 | $-0.6$ | $46.4$ | 0.3 |
| 34.7(C6) | 46.6 | 34.5 | 22.9 | 23.7 | $-0.03$ | $17.9$ | 1 |
| 49(C7) | 71.6 | 46.7 | 28.8 | 42.8 | $-0.2$ | $33.9$ | 0.8 |
| 36.1(C8) | 46.3 | 38.1 | 24.1 | 22.2 | 0.3 | $-18.1$ | 0.7 |
| 86.8(C9) | 124.1 | 75.6 | 60.8 | 63.4 | $-0.5$ | $55.9$ | 0.4 |
| 70.1(C10) | 109.7 | 55.2 | 45.5 | 64.2 | $-0.7$ | $59.4$ | 0.2 |
| 39.9(C11) | 54.1 | 32.9 | 22.8 | 21.3 | $-1.0$ | $21.2$ | 0 |
| 17.7(C12) | 29.8 | 15.1 | 8.2 | 21.6 | $-0.4$ | $18.2$ | 0.6 |
| 87.7(C13) | 117.3 | 84 | 61.9 | 55.5 | $-0.2$ | $44.4$ | 0.7 |
| 165.7(C14) | 287.9 | 149.1 | 59.9 | 228.1 | $-0.2$ | $183.4$ | 0.7 |
| 201.0(C15) | 309.7 | 171.5 | 122.2 | 187.5 | $-0.5$ | $162.9$ | 0.5 |
| 20.6(C16) | 36.4 | 16.8 | 8 | 27.7 | $-0.4$ | $23.6$ | 0.5 |
| 28.2(C17) | 33.2 | 33.2 | 18.3 | 14.9 | 1 | $-14.9$ | 0 |
| 156.6(C18) | 269.8 | 150.6 | 49.3 | 220.5 | $-0.08$ | $169.9$ | 0.9 |
| 122(C19) | 203.2 | 117.8 | 45 | 158.2 | $-0.08$ | $121.8$ | 0.9 |
| 84.8(C20) | 118.7 | 78.3 | 57.3 | 61.4 | $-0.3$ | $50.9$ | 0.6 |
| 128.7(C21) | 209.5 | 133.9 | 42.8 | 166.8 | 0.1 | $-129$ | 0.9 |
| 187.9(C22) | 300.1 | 151.2 | 112.6 | 187.5 | $-0.6$ | $168.2$ | 0.3 |
| 178.6(C23) | 240.4 | 211 | 84.3 | 156.1 | 0.6 | $-141.4$ | 0.3 |
| 46.4(C24) | 59.2 | 53.2 | 26.8 | 32.5 | 0.6 | $-29.4$ | 0.3 |
| 32.3(C25) | 49.3 | 38.1 | 9.6 | 39.8 | 0.4 | $-34.1$ | 0.5 |
| 23.5(C26) | 42.5 | 14.1 | 14.1 | 28.4 | $-1$ | $28.4$ | 0 |
| 33.2(C27) | 51.1 | 36.7 | 11.9 | 39.2 | $0.3$ | $-32$ | 0.7 |

The existence of the non-bonded electron surrounding the
nucleus is responsible for the manifestation of magnetic
shielding and deshielding effect. When the non-bonded elec-
tron revolves around the nucleus in a clockwise direction,
a paramagnetic current is generated, and a magnetic field
is induced along the direction of the external magnetic field.
Naturally, the strength of the local magnetic field is increased.
This effect is called magnetic deshielding effect. On the other
hand, when a bonded electron revolves around the nucleus
along the counter clockwise direction, a magnetic field is
generated along the opposite direction of the external magnetic
field. As a consequence, the value of the local magnetic field is
decreased - magnetic shielding effect. These effects manifest
themselves as a large value of chemical shift anisotropy for
those carbon nuclei surrounded by nonspherical distribution
of electronic charge density.

3.3 Determination of correlation times associated with
molecular motions
The molecular motions associated with molecular rotation,
translational self-diffusion, and internal motion in non-rigid
molecular can be probed by nuclear spin relaxation. Several nuclear interactions like chemical shift anisotropy interaction, magnetic dipole-dipole interaction, the spin rotation interaction, the electric quadrupole interaction, and the scalar coupling of the first and second kind simultaneously contribute to the relaxation mechanism of a spin system. However, for $^{13}$C

Fig. 3  $^{13}$C 2D-PASS CP MAS NMR spectrum of clocortolone pivalate. The horizontal axis shows isotropic spectrum, and the vertical axis shows anisotropic spectrum. The spinning CSA sideband patterns of (a) C14, (b) C22, (c) C26, (d) C13, (e) C6, (f) C4, (g) C20, (h) C18, (i) C9 are shown in this figure.
nuclei, the relaxation mechanism is predominated by the chemical shift anisotropy interaction, and the hetero-nuclear dipole–dipole coupling. The magnitude of CSA tensor fluctuates with time because of the modulations caused by molecular rotations. The chemical shift anisotropy has a great influence in relaxation mechanism for nuclei exhibiting large chemical-shift ranges. The expression of spin–lattice relaxation time due to CSA interaction is

\[ T_1 = \frac{1}{\tau} \frac{1}{1 + \frac{1}{\tau} \sigma^2 \Delta \delta} \]

where \( \Delta \delta \) is the chemical shift anisotropy parameter. Fig. 4 (a) Bar-diagram of anisotropy parameter at chemically different carbon sites of clocortolone pivalate. (b) Bar-diagram of molecular correlation time at chemically different carbon sites. Molecular correlation time is high for those nuclei for which the value of chemical shift anisotropy parameter (\( \Delta \delta \)) is high and vice versa.
where $X$ represents $\text{H}$, $\text{Cl}$, and $\text{F}$. The bond distance $T_{\text{C2}} = 1.054$ Å/Å.

The contribution of dipole–dipole interaction on spin–lattice relaxation is given by:

$$\frac{1}{T_{1}^{\text{DPP}}} = \frac{1}{10} \left( \frac{\gamma_{C} \gamma_{X} h}{r_{\text{CX}}} \right)^{2} T_{2} \frac{3}{1 + \omega_{C}^{2} T_{2}^{2}} + \frac{1}{1 + \omega_{C}^{2} T_{2}^{2}}$$

where $X$ represents $\text{H}$, $\text{Cl}$, and $\text{F}$. The bond distance $r_{\text{CX}}$ is represented in Table 4. Larmor precession frequency $\omega = 2\pi f = 2 \times 3.14 \times 125.758$ MHz = 789.76024 MHz; $B = 11.74$ T; $\gamma_{C} = 10.7084$ MHz T$^{-1}$, $\gamma_{\text{H}} = 42.577$ MHz T$^{-1}$, $\gamma_{\text{N}} = 3.077$ MHz T$^{-1}$, $\hbar = 1.054 \times 10^{-34}$ J s.

The spin–lattice relaxation for $^{13}$C can be written as

$$\frac{1}{T_{1}^{\text{CSA}}} = \frac{1}{10} \left( \frac{\gamma_{C} \gamma_{X} h}{r_{\text{CX}}} \right)^{2} T_{2} \frac{3}{1 + \omega_{C}^{2} T_{2}^{2}}$$

At the presence of high magnetic field, chemical shift anisotropy (CSA) interaction dominate over hetero-nuclei dipole–dipole coupling for non-protonated carbon nuclei. Molecular correlation time is calculated by eqn (8).

Table 3 and Fig. 4(b) show molecular correlation time, which implies the time required for the rotation of the molecule through one radian, of clocortolone pivalate at differently carbon sites. The order of magnitude varies from $10^{-4}$ to $10^{-8}$ s. Molecular correlation time is of the order of $10^{-8}$ s for those carbon nuclei attached with neighbouring carbon or oxygen atoms by double bonds like C14, C15, C18, C19, C21, C22, and C23 (as shown in Fig. 1(a)). It implies the molecular correlation time is substantially high for those carbon nuclei associated with high values of chemical shift anisotropy parameter ($\Delta \delta$). On the contrary, molecular correlation time is of the order of $10^{-8}$ s for C12, C16, C17 carbon nuclei reside at the side portion of the molecule and the magnitude of chemical shift anisotropy parameter ($\Delta \delta$) is low for those carbon nuclei. Fig. 4 shows the bar diagram of chemical shift anisotropy parameter ($\Delta \delta$) and molecular correlation time at crystallographically and chemically different carbon nuclei sites.

The crystal structure of the molecule was first drawn by using Chem Draw software. The structure was optimized by density functional theory method with the help of GAUSSIAN 09W software. B3LYP-631G (d, p) basis set was used for the optimization process. The bond distances, shown in Table 4, were extracted from the final optimized structure of clocortolone pivalate.

The considerable variation of the spin–lattice relaxation time (Table 1), CSA parameters (Table 2) and molecular correlation time (Table 3) at numerous carbon nuclei sites of clocortolone pivalate, suggests that the nuclear spin dynamics hugely varied due to the presence of the different structural groups surrounding the carbon nuclei. How the dynamics of carbon nuclei spin of clocortolone pivalate is getting affected due to the presence of various chemical group surrounding the nuclei can be pictured vividly by this type of extensive studies. The drug, clocortolone pivalate has different substitution pattern for example methylation at C8, chlorine at C4 and fluorine at C13 and pivalate at C20 etc. These unique substitution patterns make it different from other steroids to be more potent, safe, efficacious and metabolically stable. Thus, dynamics at every chemically different carbon nuclei is important in this aspect.

**Table 3: Molecular correlation time at chemically different carbon sites of clocortolone pivalate**

| Carbon nuclei | Molecular correlation time (s) | Carbon nuclei | Molecular correlation time (s) |
|---------------|-------------------------------|---------------|-------------------------------|
| C1            | $2.6 \times 10^{-5}$           | C2            | $2.6 \times 10^{-5}$           |
| C3            | $7.4 \times 10^{-6}$           | C4            | $2.7 \times 10^{-5}$           |
| C5            | $6.6 \times 10^{-5}$           | C6            | $7.7 \times 10^{-6}$           |
| C7            | $2.7 \times 10^{-5}$           | C8            | $8.3 \times 10^{-6}$           |
| C9            | $6.9 \times 10^{-5}$           | C10           | $6.8 \times 10^{-5}$           |
| C11           | $2.9 \times 10^{-7}$           | C12           | $5.9 \times 10^{-8}$           |
| C13           | $6.4 \times 10^{-5}$           | C14           | $6.7 \times 10^{-4}$           |
| C15           | $3.7 \times 10^{-4}$           | C16           | $9.7 \times 10^{-8}$           |
| C17           | $3.5 \times 10^{-8}$           | C18           | $4.8 \times 10^{-4}$           |
| C19           | $2.1 \times 10^{-4}$           | C20           | $8.2 \times 10^{-5}$           |
| C21           | $2.5 \times 10^{-4}$           | C22           | $2.2 \times 10^{-4}$           |
| C23           | $4.6 \times 10^{-5}$           | C24           | $6.6 \times 10^{-6}$           |
| C25           | $4.2 \times 10^{-6}$           | C26           | $1.3 \times 10^{-7}$           |
| C27           | $1.5 \times 10^{-5}$           |               |                               |
4. Conclusion

The correlation between the structure and dynamics of clocortolone pivalate (extensively used for cutaneous disease states, such as psoriasis vulgaris, nummular eczema, contact dermatitis, atopic dermatitis, seborrheic dermatitis, and stasis dermatitis) are monitored by measuring CSA parameters, spin-lattice relaxation time, and molecular correlation time at twenty-seven crystallographically and chemically different carbon nuclei sites. The CSA parameters are substantially varied at chemically different carbon sites, indicate that the molecular orientation and electron distribution are different in numerous regions of this molecule. The asymmetry parameter is zero for C11, C17, C26, indicates that the spinning CSA sideband pattern for those nuclei are axially symmetric. And it is very small (≤0.3) for C2, C5, C10, C22, C23, C24, resonance line, suggests the spinning CSA side-band pattern for those resonance lines is nearly axially symmetric. On other hand, the spinning CSA sideband pattern is highly asymmetric for C3, C4, C6, C18, C19, C21 carbon resonance line. Anisotropy parameter is substantially high for double bonded C14, C15, C18, C19, C21, C22, and C23 nuclei. Spin–lattice relaxation time and molecular correlation time at chemically different carbon site of this molecule differ widely. Spin–lattice relaxation rate is slow for carbon nuclei reside at the carbon ring. On the contrary, it is very fast for C12, C17, C16, C26 carbon nuclei reside at the side portion of the molecule. Molecular correlation time is of the order of $10^{-4}$ s for those carbon nuclei attached with neighbouring carbon or oxygen atoms by double bonds like C14, C15, C18, C19, C21, C22, and C23. It implies that the molecular correlation time is very high for those carbon nuclei associated with high values of chemical shift anisotropy. On the contrary, molecular correlation time is of the order of $10^{-8}$ s for C12, C16, and C17 carbon nuclei present at the side portion of the molecule. The various portion of the molecule exhibit different degree of motions. The relationship between the structure and dynamics of the molecule is pictured by these types of investigation. This type of inclusive investigation will deliver a potential impact on the way of inventing advanced medicine in pharmaceutical industry.

Conflicts of interest

There are no conflicts to declare.

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References

1 J. M. Hanifin, K. D. Cooper, V. C. Ho, S. Kang, B. R. Krafchik, D. J. Margolis, L. A. Schachner, R. Sidbury, S. E. Whitmore, C. K. Sieck and A. S. Van Voorhees, Guidelines of care for atopic dermatitis, J. Am. Acad. Dermatol., 2004, 50, 391–404.
2 D. Y. Leung, R. A. Nicklas, J. T. Li, I. L. Bernstein, J. Blessing-Moore, M. Boguniewicz, J. A. Chapman, D. A. Khan, D. Lang, R. E. Lee and J. M. Portnoy, Disease management of atopic dermatitis: an updated practice parameter, Ann. Allergy, 2004, 93, S1–S21.
3 H. Saeki, M. Furue, F. Furukawa, M. Hide, M. Ohtsuki, I. Katayama, R. Sasaki, H. Suto and K. Takehara, Guidelines for management of atopic dermatitis, J. Dermatol., 2009, 36, 563–577.
4 J. Bourke, I. Coulson and J. English, Guidelines for the management of contact dermatitis: an update, Br. J. Dermatol., 2009, 160, 946–954.
5 A. Menter, N. J. Korman, C. A. Elmets, S. R. Feldman, J. M. Gelfand, K. B. Gordon, A. Gottlieb, J. Y. Koo, M. Lebwohl, H. W. Lim and A. S. Voorhees, Guidelines of care for the management of psoriasis and psoriatic
arthritus: section 3. Guidelines of care for the management and treatment of psoriasis with topical therapies, *J. Am. Acad. Dermatol.*, 2009, 60, 643–659.

6 B. Brazzini and N. Pimpinelli, New and established corticosteroids in dermatology, *Am. J. Clin. Dermatol.*, 2002, 3, 47–58.

7 I. C. Valencia and F. A. Kerdel, *Topical corticosteroids*. Fitzpatrick’s dermatology in general medicine, ed. Wolff K., Goldsmith L. A., Katz S. I., Gilchrest B. A., Paller A. S. and Leffell D. J., McGraw-Hill, New York, 7th edn, 2008, pp. 2102–2106.

8 K. L. Goa, Clinical pharmacology and pharmacokinetic properties of topically applied corticosteroids: a review, *Drugs*, 1988, 36, 51–61.

9 S. E. Jacob and T. Steele, Corticosteroid classes: a quick reference guide including patch test substances and cross-reactivity, *J. Am. Acad. Dermatol.*, 2006, 54, 723–727.

10 M. W. Greaves, *Pruritus*. *Rook's textbook of dermatology*, ed. Burns S., Breathnach, N. Cox and C. Griffiths, Wiley-Blackwell, 8th edn, 2010, pp. 21.1–21.18.

11 B. M. Magnusson, S. E. Cross, G. Winckle and M. S. Roberts, Percutaneous absorption of steroids: determination of in vitro permeability and tissue reservoir characteristics in human skin layers, *Skin Pharmacol. Physiol.*, 2006, 19, 336–342.

12 J. Bikowski, R. Pillai and B. Shroot, The position not the presence of the halogen in corticosteroids influences potency and side effects, *J Drugs Dermatol.*, 2006, 5, 125–130.

13 M. Katz and E. H. Gans, Topical corticosteroids, structure–activity and the glucocorticoid receptor: discovery and development process of “planned serendipity”, *J. Pharm. Sci.*, 2008, 97, 2936–2947.

14 P. Buchwald, Glucocorticoid receptor binding: a biphasic dependence on molecular size as revealed by the bilinear LinBiExp model, *Steroids*, 2008, 73, 193–208.

15 C. Holden, J. English, C. Hoare, A. Jordan, S. Kownacki, R. Turnbull and R. C. Staughton, Advised best practice forthe use of emollients in eczema and other dry skin conditions, *J. Dermatol. Treat.*, 2002, 13, 103–106.

16 M. R. Warner and C. Camisa, *Topological corticosteroids. Comprehensive Dermatologic Drug therapy*, Saunders, Philadelphia, 2nd edn, 2007, pp. 595–624.

17 J. Q. Del Rosso and L. H. Kircik, A comprehensive review of clocortolone pivalate 0.1% cream: structural development, formulation characteristics, and studies supporting treatment of corticosteroid-responsive dermatoses, *The Journal of Clinical and Aesthetic Dermatology*, 2012, 5, 20–24.

18 L. H. Kircik and J. Q. Del Rosso, The treatment of inflammatory facial dermatoses with corticosteroid: focus on clocortolone pivalate 0.1% cream, *J Drugs Dermatol.*, 2012, 5, 1194–1198.

19 J. Q. Del Rosso and L. H. Kircik, Transitioning from brand to generic with topical products and the importance of maintaining the formulation and therapeutic profiles of the original product: focus on clocortolone pivalate 0.1% cream, *J Drugs Dermatol.*, 2014, 13, s77–s83.

20 L. H. Kircik, A study to assess the occlusivity and moisturization potential of three topical corticosteroid productus using the skin trauma after razor shaving (STARS) bioassay, *J Drugs Dermatol.*, 2014, 13, 582–585.

21 J. Q. Del Rosso and L. H. Kircik, The role of a midpotency topical corticosteroid and the clinical relevance of formulation characteristics in the management of commonly encountered eczematous and inflammatory dermatoses in adults and children: focus on the pharmacologic properties of clocortolone pivalate 0.1% cream, *J Drugs Dermatol.*, 2013, 12, s5–s10.

22 L. H. Kircik, Clocortolone pivalate: a topical corticosteroid with a unique structure, *J Drugs Dermatol.*, 2013, 12, s3–s4.

23 S. Singh and B. K. Mann, Clinical utility of clocortolone pivalate for the treatment of corticosteroid-responsive skin disorders: a systematic review, *Clin. Cosmet. Investig. Dermatol.*, 2012, 5, 61–68.

24 W. Xu, X. Jia, W. Liu, X. Zhang, Y. Chen, L. Zhou and S. You, Identification and characterization of three impurities in clocortolone pivalate, *J. Pharm. Biomed. Anal.*, 2012, 62, 167–171.

25 D. A. Torchia, The measurement of proton-enhanced carbon-13 T1 values by method which suppresses artifacts, *J. Magn. Reson.*, 1978, 30, 613.

26 R. Tycko, G. Dabagh and P. A. Mirau, Determination of chemical shift anisotropy lineshapes in a two-dimensional magic angle spinning NMR experiment, *J. Magn. Reson.*, 1989, 85, 265–274.

27 S. F. Liu, J. D. Mao and K. Schmidt-Rohr, A robust technique for two-dimensional separation of undistorted chemical shift anisotropy powder patterns in magic angle spinning NMR, *J. Magn. Reson.*, 2002, 155, 15–28.

28 J. C. C. Chan and R. Tycko, Recoupling of chemical shift anisotropies in solid state NMR under high speed magic angle spinning and in uniformly 13C labelled systems, *J. Chem. Phys.*, 2003, 118, 8378–8389.

29 G. Hou, L. Byeon In-Ja, J. Ahn, A. M. Gronenborn and T. Polenova, Recoupling of chemical shift anisotropy by R-symmetry sequences in magic angle spinning NMR spectroscopy, *J. Chem. Phys.*, 2012, 137, 134201–134210.

30 A. Bax, N. M. Szeverenyi and G. E. Maciel, Correlation of isotropic shifts and chemical shift anisotropies by two-dimensional Fourier-transform magic angle hopping NMR spectroscopy, *J. Magn. Reson.*, 1983, 52, 147–152.

31 A. Bax, N. M. Szeverenyi and G. E. Maciel, Chemical shift anisotropy in powdered solids studied by 2D FT CP/MAS NMR, *J. Magn. Reson.*, 1983, 51, 400–408.

32 A. Bax, N. M. Szeverenyi and G. E. Maciel, Chemical shift anisotropy in powdered solids studied by 2D FT NMR with flipping of the spinning axis, *J. Magn. Reson.*, 1983, 55, 494–497.

33 Z. Gan, High-resolution chemical shift and chemical shift anisotropy correlation in solids using slow magic angle spinning, *J. Am. Chem. Soc.*, 1992, 114, 8307–8309.

34 W. T. Dixon, Spinning-sideband-free and spinning-sideband-only NMR spectra in spinning samples, *J. Chem. Phys.*, 1982, 77, 1800–1809.
35. O. N. Antzutkin, S. C. Shekar and M. H. Levitt, Two-dimensional sideband separation in magic angle spinning NMR, *J. Magn. Reson., Ser. A*, 1995, 115, 7–19.

36. B. J. Walder, K. K. Dey, D. C. Kaseman, J. H. Baltisberger and P. J. Grandinetti, Sideband separation experiments in NMR with phase incremented echo train acquisition, *J. Chem. Phys.*, 2013, 138, 174203.

37. M. Ghosh, S. Sadhukhan and K. K. Dey, Elucidating the internal structure and dynamics of α-chitin by 2DPASS-MAS-NMR and spin–lattice relaxation measurements, *Solid State Nucl. Magn. Reson.*, 2019, 97, 7–16.

38. M. Ghosh, B. P. Prajapati, N. Kango and K. K. Dey, A comprehensive and comparative study of the internal structure and dynamics of natural β-keratin and regenerated β-keratin by solid state NMR spectroscopy, *Solid State Nucl. Magn. Reson.*, 2019, 101, 1–11.

39. M. Ghosh, N. Kango and K. K. Dey, Investigation of the internal structure and dynamics of cellulose by 13C-NMR relaxometry and 2DPASS-MAS-NMR measurements, *J. Biomol. NMR*, 2019, 73, 601–616.

40. K. K. Dey and M. Ghosh, Understanding the effect of deacetylation on chitin by measuring chemical shift anisotropy tensor and spin lattice relaxation time, *Chem. Phys. Lett.*, 2020, 738, 136782.

41. K. K. Dey, S. Gayen and M. Ghosh, Investigation of the detailed internal structure and dynamics of itraconazole by solid state NMR measurements, *ACS Omega*, 2019, 4, 21627–21635.

42. J. Herzfeld and A. E. Berger, Sideband intensities in NMR spectra of samples spinning at the magic angle, *J. Chem. Phys.*, 1980, 73, 6021–6030.

43. N. F. Ramsey, Magnetic Shielding of Nuclei in Molecules, *Phys. Rev.*, 1950, 78, 699–703.

44. N. F. Ramsey, Chemical effects in nuclear magnetic resonance and in diamagnetic susceptibility, *Phys. Rev.*, 1952, 86, 243–246.

45. A. M. Orendt and J. C. Facelli, Solid state effects on NMR chemical shifts, *Annu. Rep. NMR Spectrosc.*, 2007, 62, 115–178.

46. M. P. Nicholas, E. Eryilmaz, F. Ferrage, D. Cowburn and R. Ghose, Nuclear spin relaxation in isotropic and anisotropic media, *Prog. Nucl. Magn. Reson. Spectrosc.*, 2010, 57, 111–158.

47. N. Tjandra, A. Szabo and A. Bax, Protein backbone dynamics and 13B chemical shift anisotropy from quantitative measurement of relaxation interference effects, *J. Am. Chem. Soc.*, 1996, 118(29), 6986–6991.

48. P. Dais and A. Spyros, 13C nuclear magnetic relaxation and local dynamics of synthetic polymers in dilute solution and in the bulk state, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1995, 27, 555–633.

49. F. A. L. Anet and D. J. O’Leary, The shielding tensor part II: understanding its strange effect on relaxation, *Concepts Magn. Reson.*, 1992, 4, 35–52.

50. D. Massiot, V. Montaillout, F. Fayon, P. Florian and C. Bessada, Order-resolved sideband separation in magic angle spinning NMR of half integer quadrupolar nuclei, *Chem. Phys. Lett.*, 1997, 272, 295–300.

51. F. Fayon, C. Bessada, A. Douy and D. Massiot, Chemical bonding of lead in glasses through isotropic vs. anisotropic correlation: PASS shifted Echo, *J. Magn. Reson.*, 1999, 137, 116–121.

52. M. S. Ironside, R. S. Stein and M. J. Duer, Using chemical shift anisotropy to resolve isotropic signals in solid-state NMR, *J. Magn. Reson.*, 2007, 188, 49–55.

53. R. Y. Dong, NMR relaxation rates, *Encyclopedia of Spectroscopy and Spectrometry*, 1999, pp. 1568–1575, DOI: 10.1006/rsps.2000.0264.

54. U. Haeberlen, *High Resolution NMR in Solids: Selective Averaging*, Academic Press, New York, 1976.

55. R. J. Abraham, M. Mobli and R. J. Smith, 1H chemical shifts in NMR: Part 19† carbonyl anisotropies and steric effects in aromatic aldehydes and ketones, *Magn. Reson. Chem.*, 2003, 41, 26–36.

56. H. M. McConnell, Theory of Nuclear Magnetic Shielding in Molecules: Long-Range Dipolar Shielding of protons, *J. Chem. Phys.*, 1957, 27, 226.

57. M. Randic, M. Novic and D. Plavsic, π-Electron currents in fixed π-sexet aromatic benzenoids, *J. Math. Chem.*, 2012, 50, 2755–2774.

58. L. D. Mendelsohn, ChemDraw 8 ultra, windows and macintosh versions, *J. Chem. Inf. Comput. Sci.*, 2004, 44(6), 2225–2226.

59. M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson and H. Nakatsuji, *Gaussian 09, Revision D. 01*, Gaussian. Inc., Wallingford, CT, 2009.