Regulatory Action of all trans Cancer Acid on Metastasis Induced Lung Cell Metabolic Changes during Implantation of B16F10 Cancer Cells in C57BL6 Mice

V.M. Berlin Grace1*, D. David Wilson2, S. Saranya1 and Rohit Peardon1

1Department of Biotechnology, Karunya Institute of Technology and Sciences, Coimbatore, India. 2School of Science, Arts and Management, Karunya Institute of Technology and Sciences, Coimbatore, India.

Abstract

The changes that occur during metastasis lodging is under intense research now to develop preventive new drugs to fight against the deadly metastasis. The molecular drug, all trans Retinoic Acid (ATRA) has regulatory effects on signal mediated metabolism. In this study, we have analyzed the metastasis facilitating metabolic changes in mice lung when a highly metastatic melanoma cell line (B16F10) having potency to lodge in lung was implanted via tail vein injection into C57BL/6 mice (1×10^6 cells/ml in PBS). One group of implanted mice were treated with 0.60 mg of ATRA per Kg body weight daily for 21 days. The alteration of protein, enzymatic and non-enzymatic antioxidants (SOD, Catalase, GPX, GSH) levels and the lipid profile with cholesterol level were evaluated in the lung tissues. The ATRA treatment caused 62.16% inhibition on metastatic nodule formation. Compared to normal mice, the cancer control mice showed an increased (p < 0.01**) total protein, LPO and NO and a decreased antioxidant. In ATRA treated group, all these levels were reverted to near normal levels with a high significance (p < 0.01**) difference from untreated cancer mice. The lipid profile and cholesterol level also were altered in cancer and were normalized in ATRA treated group with high significance (p < 0.01**). All these results implies that the metabolic changes induced in the lung tissue during metastatic lodging of melanoma cells were prevented and regularized by the ATRA treatment in vivo which give a scope of anti-metastatic therapy using ATRA.

Keywords: Lung metastasis, ATRA, C57BL/6 mice, melanoma cell line, Metabolism change

*Correspondence: berlinbiochem@gmail.com; Tel: +91-9894051175

(Received: November 18, 2020; accepted: May 12, 2021)
INTRODUCTION
Cancer is a multi-factorial disorder, originating due to infection, inflammation, pollution-smoke, radiation, oxidative stress, diet, life style and genetics (Anand et al.). Most of the lung cancer patients when diagnosed are at metastasis stage as it metastasizes very fast which may be due to rich blood flow. Hence, now the cancer researchers are concentrating more on management of cancer metastasis, fast progression as well as the associated changes in the body. The Vitamin A active metabolite, ATRA is a molecular therapeutic agent which has control over several genes involved in the molecular pathogenesis of cancer progression and metastasis (Cui et al.) by binding to its receptor RARs and regulating target genes (Altucci et al.). ATRA has shown vital regulatory role in embryonic development of several tissues and organs (Romero et al.). As an example, the functional retinoic acid response elements (RAREs) was found within flanking sequences of some of the most 3’ Hox genes (Marshall et al.) which are the resperoner to ATRA during early embryonic development stages in control anterior/posterior patterning (Holland et al.). ATRA alone or in combination is now used as a potent therapeutic agent for Acute Promyelocytic Leukaemia (APL) complete remission rate as high as 85% to 95% (Osman et al; Zhang et al and Chou et al) and was found to induce differentiation (Zuccari et al.) and apoptosis (Altucci et al.). ATRA has been proven to treat vascular disorders and abnormal angiogenesis (Maiti et al.).

The oxidative stress and inflammatory mediators generated during cancer development aggravate the lung cancer progression and enhance the metastasis (Valavanidis et al.). The oxidative stress markers and inflammatory mediators set up a favourable tumour microenvironment at cancer initiation itself and enhance progression, promote angiogenesis, invasion, tumour cell migration, leading to metastasis. (Vaidya et al.). Some studies have shown a relationship between retinoic acid and the generation of nitric oxide (NO) (Rafa et al., Grosjean et al., Hirokawa et al., Seguin et al.), prostaglandins (Mamidi et al.; Devaux et al., Hill et al.) and the expression of cyclooxygenase 1 (COX-1) (Habib et al, Nusing et al) and COX-2 (Kanekura et al.). Retinoid have various effects on cytokines by decreasing IL-12 and IFN as well as increasing IL-4 (Villarroya et al.). Cell line implantation and metastasis is reported to generate several reactive species and radicals and the cancer condition itself reduces the antioxidant enzyme level in our body. However, it is unknown whether oxidative stress is the cause or the consequence of disease, but Reactive Oxygen Species (ROS) promotes tumor progression and make up the tumor microenvironement (Liou and Storz, 2010; Weinberg et al) which promote immune cell infiltration (Kotsafti et al) and apoptosis (Altucci et al.) which are the responder to ATRA. RoS activate cell survival signalling, leading to cancer cell migration and metastasis (Aggarwal et al.). However, reduction of these stress parameters and inflammation mediators help to manage the metastatic complications in lung cancer. Vitamin A and retinoid family have been traditionally recognized as chain breaking anti-oxidant or redox-active molecules (Khaper et al., Mantymaa et al.). ATRA at very low level has enhanced the levels of MnSOD2 and Cu.ZnSOD1 to protect from oxidative damage in a neuronal cell line study (Ahlemeyer et al.). Lung metastasis involve oxidative stress and inflammation (Li et al.).

Lipids as the key membrane composition, source of energy and regulator of signal transduction pathways, plays vital role in cancer progression and undergo rapid change both at molecule and gene levels in cancer (Watkins et al). Study in mice has shown that the treatment with ATRA reduced body weight and adiposity independent of changes in food intake (Puigserver et al.) and has improved the glucose tolerance level (Felipe et al.). ATRA has been proven to induce body fat loss which very well correlated with an activation of brown adipose tissue (BAT) (Bonet et al. and Puigserver et al.) and it was also reported to reduce adipogenic/lipogenic capabilities (Ribot et al.), thereby enhancing the oxidative metabolism and thermo genesis in white adipose tissue (WAT) depots, leading to fatty acid mobilization as well as healthy oxidation in other tissues (Mercader et al., 2006). Thus retinoic acid was found to regulate the adipose tissue development as well as deposition of lipid at different tissues (Hiroshi et al.).

This study was therefore designed to analyze the vital changes happening in the lung of C57BL6 mice during cancer cell line implantation
and the regulatory role of ATRA in this condition by considering all these findings regarding metastasis associated metabolic and physiologic changes.

**MATERIAL AND METHODS**

**Animals**

Four to six weeks old mice (C57BL/6) were procured from the National Institute of Nutrition, Hyderabad, and subjected in this study. The mice were maintained in animal house at suitable condition as per the guidelines of CPCSEA during the experimental period by providing normal feed with water ad libitum.

**Chemicals**

ATRA was purchased from Sigma Chem. Co. (St. Louis, MO) and the other chemicals of analytical grade used were purchased from Himedia, India.

**Cancer Cells**

The melanoma cell line (B16F10) used for developing lung metastatic cancer in mice was obtained from the National Centre for Cell Science, Pune, India and sub-cultured in buffered medium (RPMI1640), supplemented with antibiotic, penicillin and streptomycin. They were recovered by harvesting with trypsin and EDTA and then (PBS, pH 7.4) wash was given.

**Study Design and Groups**

Four experimental groups (6 C57BL/6 mice per group) were included in this study. The group 1 mice was maintained as normal group and the mice of groups 2-4 were injected via tail vein with 0.1 ml of B16F10 melanoma cells (1×10⁶ cells/ml in PBS) to develop metastatic mice model. The group 3 and 4 mice were simultaneously treated with i.p injection of 0.1 ml Olive oil and ATRA (0.60 mg per Kg body weight) in olive oil respectively for 21 consecutive days as per the literature (Suzuki et al.). Furthermore no mortality or behavioral changes were shown by the mice until 50 days of observation in acute toxicity study carried out earlier for the dose up to 1mg/Kg body weight. The protocol for these experiment was followed in line with the guidelines of CPCSEA and the approval for this study was obtained from the Institutional Animal Ethical Committee of Karunya University [IAEC/KU/BT/12/019]. On the 22nd day of metastasis induction and treatment, all the mice were sacrificed. The lung tissues were excised out, washed and observed visually through petri dish glass for metastatic nodules to count. From the number of nodules, we have calculated the percent of inhibition of nodule formation:

\[
\text{Inhibition Percent (%) = } \frac{\text{No. of nodules in metastasis control - No. of nodules in treated}}{\text{No. of nodules in metastasis control}} \times 100
\]

Then a small portion of it was cut, placed in a buffer formalin (10%), and embedded in paraffin for histologic studies. The remaining portion of lung tissues from each mouse were subjected to other analyses as detailed below.

Group 1: Normal mice

Group 2: B16F10 melanoma cell line implanted metastasis control mice

Group 3: B16F10 melanoma cell line implanted mice + Olive oil treatment

Group 4: B16F10 melanoma cell line implanted mice + ATRA treatment

**Estimation of ATRA level in the tissue of lung**

Lung tissues were homogenized with 10% ascorbic acid and ethanol. After vortexing, it was subjected to lipid extraction with n-hexane (2 ml), then vortexed. The hexane layer was collected after centrifugation for 10 minutes (2000 rpm) and air dried. The ATRA concentration in the residue after re-dissolving it in methanol (250µl) was then analysed using reversed phase HPLC system. The ATRA was detected at a wavelength of 310 nm with a mixture of acetonitrile in 4.5 to 6.5 ratio, run in the Phenomenex Luna column (250 mm×4.6 mm) at 1ml/minute flow rate. The peak area was calculated from the standard ATRA (400 to 2000ng/ml).

**Total protein assay in lung tissues**

A portion of lung tissue was homogeneized with buffer and the supernatant was separated by centrifugation. Then the total protein level was estimated against the standard protein bovine serum albumin (BSA) by the widely used standard method (Lowry et al.) and the optical density was measured at 680 nm. From the standard graph obtained from the increasing concentrations of standard BSA, the level of protein present in the lung tissue extract was calculated and expressed in µg/g tissue.

**Evaluation of oxidative stress levels in lung tissue**

The lung tissue was homogenised and the supernatant after centrifugation was subjected to analysis of various biochemical parameters which are related to oxidative stress induced in lung
during implantation of metastatic cell line. The assays for the following enzymatic antioxidants were performed using standard colorimetric methods described by the researchers in very early years as follows: The method described by Kakkar et al., was used for the assay of Super Oxide Dismutase (SOD) (Kakkar et al.). The Catalase assay was performed as describe previously (Sinha et al.). The Glutathione Peroxidase (GPX) assay was carried out by following the relevant literature (Rotruck et al.).

Similarly, the non-enzymatic antioxidant Glutathione (GSH) estimation was done in the supernatant of TCA (10%) precipitated homogenate by following the standard colorimetric method (Ellman et al.) and from the plotted standard graph and from which the GSH level in sample was calculated.

The extent of lipid peroxidation in lung tissue due to metastasis was assessed by measuring the production of thiobarbituric acid reactive substances (TBARS) by following the standard method (Ohkawa et al.) using MDA as standard. The assay for the generated nitric oxide (NO) was carried out in 250 µl of lung tissue homogenate by incubating it with Griess reagent for 15 minutes and reading at 546nm (John et al.)

Lipid profiling
The blood serum of experimental mice were subjected to lipid profiling along with estimation of total cholesterol level by following the standard Zak’s method. Lipid profiling was done with the use of “Erba lipid profile kit. Briefly, the reagent was mixed with serum and the absorbance was read using colorimeter at 505 nm after 5 minutes incubation (37°C).

RESULTS
Metastatic nodules formed in lung
The number of nodules formed due to metastatic lodging in lung of all mice was visually counted and the mean number of nodules in each group of mice is given in Table 1. The effect of ATRA treatment on prevention was then calculated by comparing with the number of nodules found in the lung of metastasis control mice as follows and expressed as percent inhibition in Table 1.

The mean values given are from 6 mice and the p value is considered Non significant if it is > 0.05; * means significant (p > 0.05), and ** means a highly significant (p > 0.01)

ATRA and total protein in lung tissue
The ATRA peak was obtained at RT value of 4.5 and from the standard calibration curve.

Table 1. Number of nodules found on lung and the percent inhibition by ATRA

| Groups                | Mean number of nodules | Inhibition (%) |
|----------------------|------------------------|----------------|
| 1 - Normal           | Nil                     | NA             |
| 2 - Metastasis control | 59.52 ± 01             | NA             |
| 3 – Metastasis + vehicle treated | 56.61 ± 11 \textsuperscript{ns} | 4.88          |
| 4 – Metastasis + ATRA treated | 22.52 ± 07\textsuperscript{**} | 62.16         |

Table 2. The total protein level in lung tissue

| Groups                | Mean protein level (µg/g) | ATRA level (µg/ml homogenate) |
|----------------------|---------------------------|-------------------------------|
| 1 - Normal           | 24.23 ± 12                | 0.223 ± 0.05                  |
| 2 - Metastasis control | 47.32 ± 10\textsuperscript{**} | 0                             |
| 3 – Metastasis + vehicle treated | 46.21 ± 17\textsuperscript{ab ns} | 0                             |
| 4 – Metastasis + ATRA treated | 31.22 ± 06\textsuperscript{ab**} | 0.476 ± 0.08 \textsuperscript{**} |

NA – Not applicable
Normally the level of ATRA will be maintained in a lower quantity in serum and in lung. Hence in our study also the quantity of ATRA present in the lung of normal mice was found to be very low (0.223 ± 0.05) whereas, in the metastasis control group and the olive oil treated group the ATRA level was completely reduced and was undetectable. But the treatment with ATRA has shown an elevation (p > 0.01**) in the level of ATRA than the normal level in group 4 animal as shown in Table 2.

The increase in total protein level when compared to normal mice was found to be highly significant (p > 0.01**) in the metastasis control group as well as in olive oil treated group whereas the increase in ATRA treated group was a minimum (p> 0.05*). ATRA treated group shown a greater difference from control group while a non-significance was observed in olive oil treated group as shown in Table 2. This may be resulted from the cell physiologic/functional changes due to increase in cell surface receptor, glycoproteins, lytic enzymes such as collagenase, cathepsin, plasminogen activator (cleave peptide bond) which favour mobility and dissemination of metastatic cells.

The values given are the mean ± SD of 6 mice and the p value is considered Non significant if it is > 0.05ns; significant if it is > 0.05*, and highly significant if it is > 0.01**. aGroup 1 Normal Vs Groups 2-4; bGroup 2- Control Vs Groups 3 and 4

Profile of Oxidative Stress Markers in the Lung Tissue

In this study we have observed a significant level of changes in lung antioxidant levels in the experimental groups during metastasis (Table 3). A highly significant (p > 0.01**) reduction in antioxidant enzymes was observed in metastasis control groups. Comparatively, the ATRA treated group has shown a highly significant increase in all these enzymes while, in the vehicle treated group, no significant reduction was observed in olive oil treated group (Table 3).

The mean values of 6 mice are given with SD. ns means not significant (p > 0.05); * means

### Table 3. Levels of antioxidants in the lung tissue

| Groups                        | SOD (U/Mg protein) | Enzymatic Catalase (µM/Mg protein) | GPX (µg/Mg protein) | Non-enzymatic GSH (µg/Mg protein) |
|-------------------------------|--------------------|-----------------------------------|--------------------|----------------------------------|
| 1 - Normal                    | 4.35± 0.02         | 0.091 ± 0.18                      | 43.17 ± 0.14       | 1.93 ± 0.05                      |
| 2 - Metastasis control        | 1.87± 0.03         | 0.031 ± 0.02                      | 19.09 ± 0.08       | 0.52 ± 0.16                      |
| 3 – Metastasis + vehicle treated | 1.75± 0.06 | 0.037 ± 0.04                      | 19.21 ± 0.13       | 0.59 ± 0.13                      |
| 4 – Metastasis + ATRA treated | 3.65± 0.11         | 0.072 ± 0.05                      | 36.05 ± 0.07       | 1.22 ± 0.06                      |

### Table 4. Extent of lipid peroxidation and Nitric Oxide levels in the lung tissue

| Groups                        | LPO activity (nM/g) | NO level (µM/g) |
|-------------------------------|--------------------|-----------------|
| 1 - Normal                    | 12.21 ± 0.07       | 1.39 ± 0.07     |
| 2 - Metastasis control        | 40.33± 0.04         | 3.98± 0.12      |
| 3 – Metastasis + vehicle treated | 40.52± 0.02 | 3.65± 0.11      |
| 4 – Metastasis + ATRA treated | 20.14± 0.09         | 2.04± 0.08      |

### Table 5. Lipid profile and cholesterol levels in the serum (mg/dl)

| Groups                        | Cholesterol | TGL | HDL | LDL | VLDL |
|-------------------------------|-------------|-----|-----|-----|------|
| 1 - Normal                    | 110 ± 2     | 97 ± 4 | 42 ± 3 | 67 ± 1 | 21 ± 2 |
| 2 - Metastasis control        | 225 ± 1     | 32 ± 2 | 18 ± 2 | 107 ± 2 | 41 ± 1.5 |
| 3 – Metastasis + vehicle treated | 223 ± 2   | 31 ± 3 | 18 ± 3 | 105 ± 1  | 40 ± 3 |
| 4 – Metastasis + ATRA treated | 130 ± 1     | 85 ± 1 | 36 ± 1 | 78 ± 3  | 29 ± 2 |


significant (p > 0.05) ** means highly significant (p > 0.01). Also, a Group 1 Normal Vs Groups 2 Control; b Group 2 Control Vs Groups 3 and 4 treatments. We have also observed a significant levels of changes in the extent of cell membrane lipid peroxidation as well as the reactive NO radical in the lung tissues among the groups of experimental mice during metastasis as shown in Table 4. The extent of lipid peroxidation and the NO level have increased in the metastasis control group in a highly significant (p > 0.01**) level while ATRA treated group has shown a highly significant (p > 0.01**) reduction than control. However, the olive oil treated group showed only non-significant difference from the control group.

The mean values from 6 mice are given with SD. a means not significant (p > 0.05); b means significant (p< 0.05) ** means highly significant (p > 0.01). Here, a Group 1 Normal Vs Groups 2 Control; b Group 2 Control Vs Groups 3 and 4 treatments

**Lipid Profile and Level of Cholesterol in the Lung Tissue**

The results of lipid profile and cholesterol are given in Table 5. A highly significant (p > 0.01**) increase was shown in cholesterol and HDL levels in the metastasis control. On contrary to it, a highly significant (p > 0.01**) decrease was noticed for TGL in the control group. All these changes were brought to near normal level in the ATRA treated group while the olive oil treatment which showed similar alterations like control group could not make any reversion in the levels.

The data is given as mean values from 6 mice with SD. a means not significant (p > 0.05); b means significant (p< 0.05) ** means highly significant (p > 0.01). Here, a Group 1 Normal Vs Groups 2 Control; b Group 2 Control Vs Groups 3 and 4 treatments

**DISCUSSION**

It is understood from the literature that most of the physiologic regulatory functions of Vitamin A mainly metabolism, cell growth, apoptosis and reproduction are exhibited through its one of the active metabolite, ATRA along with other retinoid (Mangelsdorf et al.). The chemotherapeutic role of ATRA as a cell differentiating molecular agent has become well known due to its efficient action on treating Acute Promyelocytic Leukemia (APL) and also by its key role in treating many other deadly diseases (Farooqui et al.). In addition to its therapeutic role ATRA has crucial role as chemo preventive agent against the development of cancer, angiogenesis and inflammation. In the past work which has been done to study the anti-inflammatory effect of ATRA in inflammatory arthritis has shown that the incidence of arthritis were lower in mice treated with ATRA than a normal mice (Yuji et al.). Some of the molecular actions of ATRA treatment on cancer were found to be arresting cell cycle at G1 as well as prolonging cell division rather killing tumor cells which is an identity for molecular therapy and also it could reduce DNA synthesis as well as colony formation. Being a differentiating agent it could also induce morphological changes in tumour cells (Wu et al.).

The cellular ATRA level is tightly regulated by two types of binding proteins such as CRABP I and II which direct it to nucleus for interaction with its high affinity receptors RARs (α, β, and γ) for exhibiting molecular action on target cells along with its isomer, 9-cis-retinoic acid which can also bind to RXRs (α, β, and γ) (Chen et al.; Mangelsdorf et al.; Heyman et al.). According to the variation in receptor types availability in each organ ATRA exhibits regulation of various cellular functions. Presently intense research is focused towards the therapeutic efficiency of ATRA for many solid cancers in addition to APL and vascular disorders (Zuccari et al.; Maiti et al.).

Recent studies have highlighted that the hypoxia condition in cancer cell infiltration induces ROS production and inflammatory responses which facilitate the tissue damage mediated migration of cancer cells, resulting in lung metastasis (Li et al.). A recent mice model study also has demonstrated that the hypoxia induced oxidative stress and inflammatory response promoted melanoma lung metastasis (Li et al.). In our study ATRA has shown an impressive antioxidant activity to reduce the free radicals present in the body by the different antioxidant enzymes. The previous reports states that the free radicals can damage the DNA and can cause a tumour cell proliferation. When ATRA was given to the cancer induced mice it has shown an impressive enzymatic antioxidant activity. Our study was done on SOD, Catalase, NO, GPx, GSH, LPO and the results states that ATRA has the...
When these antioxidant levels are reduced in living systems, the excess free radicals generated during cancer metastasis and progression via activated LPO action as we have demonstrated in our study will facilitate further progression and establishment of cancer in metastasized site. It was reported by other studies also that the oxidative stress created by the free radicals have implication in the pathogenesis of many clinical disorders, including cancer (Narendirakannan et al.). They also have highlighted that there should be enough antioxidants or natural agents that are capable of augmenting the activity antioxidant enzymes to prevent implications of free radicals including cancer progression. In addition, the membrane lipids also plays key role in generating such free radicals as well as in signal transduction leading to alteration in cell growth and apoptosis (Watkins et al.).

On the basis of the previous reviews, our idea of doing the lipid profiling was to identify that when ATRA is given to the normal mice and the cancer cell line implanted mice what will be the effect on the lipid content of the body. There is no adverse change in the lipid content of the body for ATRA treatment however, there is a significant decrease in the cholesterol level in mice and increase in the LDL level. The result shows that ATRA has no adverse effect on the lipid content of the body and there were no significant side effect seen in ATRA treated mice.

On the basis of the result which came out of different parameter done to analyse the physiological effect of ATRA on lung cancer in mice has shown a potent effect of ATRA.

Since ATRA is already been use as a chemo-preventive drug for the APL, it can be used for the treatment of lung cancer also to prevent the metastasis and progression. Though ATRA is a lipid, by lipid profiling it is seen that it has not shown any adverse effect on the lipid content in mice body. The conclusion of all these parameters is that ATRA has potent anti-cancer drug properties against lung metastasis.

**Acknowledgments**

The authors acknowledge the financial support given by the Management of Karunya Institute of Technology and Sciences via Karunya Short Term Research. The shared facilities from the Cancer Nano-therapeutic Research Centre established with the funding of the Department of Science and Technology – Science and Engineering Research Board and the Department of Biotechnology New Delhi.

**Conflict of Interest**

The authors declare that there is no conflict of interest.

**Authors’ Contribution**

All authors designed the experiments. SS & RP performed the experiments. VMBG analyzed the data and DDW wrote the manuscript along with BG. All authors read and approved the manuscript.

**Funding**

Management of Karunya Institute of Technology and Sciences via Karunya Short Term Research Grant (KSTRG:2008-09). Funding of the Department of Science and Technology – Science and Engineering Research Board (DST-SERB-Ref: No.SB/YS/LS-252/2013, dated 15 May, 2014) and the Department of Biotechnology (Ref. No.: BT/ PR14632/NNT/28/824/2015, dated 02-09-2016) New Delhi.

**Data Availability**

All datasets generated or analyzed during this study are included in the manuscript.

**Ethics Statement**

This study was carried out in accordance with the recommendations of NIH guidelines for animal use, Animal Care and Use Committee (NIH). The protocol for these experiment was followed in line with the guidelines of CPCSEA and the approval for this study was obtained from the Institutional Animal Ethical Committee of Karunya University [IAEC/KU/BT/12/019].

**References**

1. A Vaishali, HS Tuli, A Varol , et al. Role of Reactive Oxygen Species in Cancer Progression: Molecular Mechanisms and Recent Advancements. Biomolecules. 2019; 9:735. doi: 10.3390/biom9110735
2. Ahlemeyer B, Bauerbach E, Plath M, et al. Retinoic acid reduces apoptosis and oxidative stress by preservation of SOD protein level. Free Radic Biol
1. Habib A, Hamade E, Mahfouz R, Nasrallah MS, de Thé H, Bazarbachi A. Arsenic trioxide inhibits ATRA-induced prostaglandin E2 and cyclooxygenase-1 in NB4 cells, a model of acute promyelocytic leukemia. Leukemia. 2008;22(6):1125-30. PMID: 18354491. doi: 10.1038/leu.2008.59

2. Heyman RA, Mangelsdorf DJ, Dyck JA et al. 9-cis retinoic acid is a high affinity ligand for the retinoid X receptor. Cell. 1992;68:397-406. doi: 10.1016/0022-8674(92)90479-V

3. Hirokawa K, Masanori Utsuyama, Yi-Xin Zeng, ChieriKurashima, Kasai Michiyuki. Immunological alterations with aging-laying a stress on recent progress in Japan, Archives of Gerontology and Geriatrics. 1994;19(2):171-183. doi: 10.1016/0167-4943(94)90038-8

4. Ronkinson IB, Agonist and antagonist of retinoic acid receptors cause similar changes in gene expression. Physiol Rev. 2001;81:1329-1340. doi: 10.1152/physrev.2001.81.3.1329

5. Balakrishnan N, Panda A, Raj N, Shrivastava A, Prathani R. The evaluation of nitric oxide scavenging activity of Acalypha indica Linn root. Asian J Res Chem. 2009;2(2):148-150.

6. Cui J, Gong M, He Y, Li G, He T, Bi Y. All-trans retinoic acid inhibits proliferation, migration, invasion and induces differentiation of hepa-1 cells through reversing EMT in vitro. International Journal of Oncology. 2016;48(1):349-357. doi: 10.3892/ijo.2015.3235

7. Devaux Y, Zangrando J, Schreno B, et al. Long noncoding RNAs in cardiac development and ageing. Nat Rev Cardiol. 2015;12(7):415-25. doi: 10.1038/nrcardio.2015.55

8. Ellman GL, Courtney KD, Andresjiv V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem Pharmacol. 1961;7:88-97. doi: 10.1016/0006-2952(61)90145-9

9. Farooqui AA, Yang HC, Horrocks L. Involvement of phospholipase A2 in neurodegeneration. Neurochem Int. 1997;30:517-522. [PubMed: 9152992] doi: 10.1016/S0744-1327(96)00122-2

10. Flores F, Bonet ML, Ribot J, Palau A. Modulation of resistin expression by retinoic acid and vitamin A status. Diabetes. 2004;53:882-889. doi: 10.2337/diabetes.53.4.882

11. Grosjean F, Cerneau P, Bourdillon A, Bastianelli D, Peyronnet C, Duc G. Feeding value, for pig, of near isogenic faba beans containing or not tannins and with high or low levels of vicine or convicine. J. Rech. Porc. 2001;33:205-210.
31. Mantymaa P, Guttorm T, Sitonen T, et al. Cellular redox state and its relationship to the inhibition of clonal cell growth and the induction of apoptosis during all-trans retinoic acid exposure in acute myeloblasticleukemia cells. Haematologica. 2000; 85:238-245.

32. Marshall H, Morrison A, Studer M, Pöpperl H, Krumlauf R. Retinoids and Hox genes. The FASEB Journal. 1996;10: 969-978. doi: 10.1096/fasebj.10.8.8801179

33. Mercader J, Ribot J, Murano I et al. Remodeling of FLR1 transporter requires phospholipase C and is repressed by Mediator. J Biol Chem. 2006;281(31):22152-6. doi: 10.1074/jbc.M506728200

34. Ribot J, Felipe F, Bonet ML, Palou A. Changes of adiposity in response to vitamin A status correlate with changes of PPAR gamma 2 expression. Endocrinology. 2006;147:5325-5332. doi: 10.1210/en.2006-0760

35. Nozaki Y, Yamagata T, Sugiyama M, Ikoma S, Kinoshita K, Funuchi M. Anti-inflammatory activity of all-trans-retinoic acid in inflammatory arthritis. Clinical immunology. 2006;119:272-9. doi: 10.1016/j.clim.2005.11.012

36. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides of blood lipids. Anal Biochem. 1979;95:318-27. doi: 10.1016/0003-9861(79)90099-9.

37. Osman AEG, Jennifer A, Jane EC, et al. Treatment of all-trans-retinoic acid in inflammatory arthritis. Mediators of Inflamm. 2018;2018:8971943. doi: 10.1155/2018/8971943

38. Palace VP, Khaper N, Qin Q, Singal PK. Antioxidant and anti-inflammatory effects of retinoic acid in the colon. Inflamm. Res. 2004;53:421-6. doi: 10.1007/s00011-004-2455-5

39. Puigserver P, Vázquez R, Giralt M. Retinoids and retinoid receptors in the control of energy balance: a synthesis of theory and empirical evidence. Int J Environ Res Public Health. 2013;10(9):3886-3907. doi: 10.3390/ijerph10093886

40. Rafa H, Benkhelifa S, Younes SA, et al. All-Trans Retinoic Acid Modulates TLR4/NF-κB Signaling Pathway Targeting TNF-α and Nitric Oxide Synthase 2 Expression in Colonic Mucosa during Ulcerative Colitis and Colitis Associated Cancer. Mediators of Inflamm. 2017;2017:7353252. doi: 10.1155/2017/7353252

41. Ribeiro Neto E, Vazquez F, Bonet ML, Palou A. Changes of adiposity in response to vitamin A status correlate with changes of PPAR gamma 2 expression. Obes Res. 2001;9:500-509. doi: 10.1038/oby.2001.65

42. Romero C, Desai P, DeLillo N, Vancura A. Expression of FLR1 transporter requires phospholipase C and is repressed by Mediator. J Biol Chem. 2006;281(9):5677-85. doi: 10.1074/jbc.M506728200

43. Rottruck JF, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. Science. 1973;179(4073):588-90. PMID: 4686466. doi: 10.1126/science.179.4073.588

44. Sano H, Kawahito Y, Wilder RL, et al. Expression of cyclooxygenase-1 and -2 in human colorectal cancer. Cancer Res. 1995;55(17):3785-9. PMID: 7641194

45. Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972;47(2):389-94. doi: 10.1016/0003-2697(72)90132-7

46. Suzuki T, Harai H, Nakano M, et al. Site-specific labeling of cytoplasmic peptide-N-glycanase by N,N’-diacetylchitobiose-related compounds. J Biol Chem. 2006;281(31):22152-60. doi: 10.1074/jbc.M603236200

47. Vaidya FJ, Chhipa AS, Sagar N, Pathak C. Oxidative Stress and Inflammation Can Fuel Cancer. In: Maurya P, Dua K. (eds) Role of Oxidative Stress in Pathophysiology of Diseases. Springer, Singapore. 2020. doi: 10.1007/978-981-15-1568-2_14

48. Valavanidis A, Vlachogianni T, Fiotakis K, Loridas S. Pulmonary oxidant stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. Int J Environ Res Public Health. 2013;10(9):3886-3907. doi: 10.3390/ijerph10093886

49. Villarroya F, Iglesias R, Giralt M. Retinoids and retinoid receptors in the control of energy balance: novel pharmacological strategies in obesity and diabetes. Curr Med Chem. 2004;11:795-805. doi: 10.2174/0929867043455747

50. Watkins AL, Hilleston W, Morecroft SE. Audit quality: A Synthesis of theory and empirical evidence. Journal of Accounting Literature. 2004;23: 153-193.

51. Weinberg F, Ramnath N, Nagarath R. Reactive Oxygen Species in the Tumor Microenvironment: An Overview. Cancers. 2019;11(8):1191. doi: 10.3390/cancers11081191

52. Wu S, Donigan A, Platsoucas CD, Jung W, Sopranzaki M, Sopranzaki KJ. All-trans retinoic acid blocks cell cycle progression of human ovarian adenocarcinoma cells at late G-1. J Exp Cell Res. 1997;232: 277-286. doi: 10.1006/excr.1997.3495

53. Zhang X, Liu L, Yao Y, et al. Treatment of non-high-risk prostate cancer with realgar-indigo (ATRA): study protocol for a randomized controlled trial. Trials. 2020;21:7. doi: 10.1186/s13063-019-3983-2

54. Zuccari G, Carosio R, Fini A, Montaldo PG, Orienti I. Modified polyvinylalcohol for encapsulation of all-transretinoic acid in polymeric micelles. J Control Rel. 2005;103: 369-380. doi: 10.1016/j.jconrel.2004.12.016