Evaluation of Approach to a Conspecific and Blood Biochemical Parameters in TAAR1 Knockout Mice

Ilya S. Zhukov 1,2,*, Maria A. Ptukha 1, Ekaterina A. Zolotoverkhaja 3, Ekaterina L. Sinitca 2, Ilya Y. Tissen 2, Inessa V. Karpova 2, Anna B. Volnova 1 and Raul R. Gainetdinov 1,4,*

1 Institute of Translational Biomedicine, Saint Petersburg State University, Universitetskaya Nab., 7–9, 199034 Saint Petersburg, Russia; ptukhamaria@yandex.ru (M.A.P.); a.volnova@spbu.ru (A.B.V.)
2 S. Anichkov Department of Neuropharmacology, Institute of Experimental Medicine, Federal State Budgetary Scientific Institution, Academica Pavlova Str. 12, 197376 Saint Petersburg, Russia; sin.e@list.ru (E.L.S.); iljatis@mail.ru (I.Y.T.); inessa.karpova@gmail.com (I.V.K.)
3 Laboratory of Biochemical Toxicology and Pharmacology, Golikov Research Center of Toxicology, Federal Medical-Biological Agency, Bekhtereva Str. 1, 192019 Saint Petersburg, Russia; e.zolotoverkhaja@yandex.ru
4 Saint Petersburg State University Hospital, Saint Petersburg State University, Universitetskaya Nab., 7–9, 199034 Saint Petersburg, Russia
* Correspondence: ilya.zhukov@skolkovotech.ru (I.S.Z.); gainetdinov.raul@gmail.com (R.R.G.)

Abstract: It is known that the trace amine-associated receptor 1 (TAAR1) receptor is involved in limbic brain functions by regulating dopamine transmission and putative reward circuitry. Moreover, other TAARs are expressed in the olfactory system of all studied vertebrate species, sensing innate socially-relevant odors, including pheromones. Therefore, one can assume that TAARs may play a role in rodent social and sexual behavior. A comparative behavioral and biochemical analysis of TAAR1 knockout (TAAR1-KO) and wild-type mice is also important for the preliminary evaluation of the potential side effects of future TAAR1-based therapies. In our studies, we adapted a sexual incentive motivation test for mice to evaluate the sexual behavior of TAAR1-KO and wild-type mice. Furthermore, we measured testosterone and other biochemical parameters in the blood. As a result, we found only minimal alterations in all of the studied parameters. Thus, the lack of TAAR1 does not significantly affect sexual motivation and routine lipid and metabolic blood biochemical parameters, suggesting that future TAAR1-based therapies should have a favorable safety profile.

Keywords: trace amines; TAAR; mice sexual motivation; biochemistry; testosterone; TAAR1; safety profile

1. Introduction

In 2001, two independent groups discovered a family of monoamine-related G protein-coupled receptors (GPCRs) named Trace Amine-Associated Receptors (TAARs) [1–3]. In mammals, nine subfamilies of trace amine-associated receptors (TAAR1-9) genes are known. Three of them are pseudogenes in humans [4,5]. The classic examples of trace amines are β-phenylethylamine (PEA), p-tyramine (TYR), tryptamine (TRP), and p-octopamine (OCT). Trace amines are structurally and functionally close to classical monoamine neurotransmitters such as dopamine, serotonin, and norepinephrine, but their tissue concentrations are more than 100 times lower [6]. Nowadays, TAAR1 is one of the most investigated trace amine-associated receptors. Its expression was found in the limbic brain areas and certain peripheral tissues. Significant alterations in the brain dopamine, serotonin and glutamate function were found in TAAR1 knockout (TAAR1-KO) mice [4,5]. Based on elevated dopamine transmission, supersensitivity of D2 dopamine receptors and enhanced responsiveness to amphetamine, TAAR1-KO mice were proposed as a model of schizophrenia [4,5]. In fact, the first drug based on TAAR1 agonism has successfully passed Phase II of clinical trials for the treatment of schizophrenia [7].
Previous studies demonstrated that alterations in mesolimbic and mesocortical dopaminergic neurons could affect several aspects of rodent sexual behavior [8–16]. Recent studies in dopamine transporter (DAT) knockout rats confirmed a key role of dopamine in sexual behavior. They provided evidence that the permanently elevated dopamine levels triggered by DAT gene silencing can significantly affect male sexual motivation [17]. TAAR1 can modulate the dopamine system via the formation of the TAAR1/D2R heteromer complex [5]. Several lines of evidence indicate that the TAAR1 is involved in limbic networks and can be involved in putative reward functions [4,5]. At the same time, TAAR2-TAAR9 are expressed in the olfactory epithelium of all studied vertebrate species, functioning as sensors of socially-relevant innate odors, including pheromones [18–20]. Moreover, recent studies indicate that TAAR5 and likely other “olfactory” TAARs are also expressed in the limbic brain areas and modulate adult neurogenesis [21–23]. These observations suggest that such an impact of TAARs on the central nervous system (CNS) functions can potentially affect social and sexual functions. In the present study, we used a non-contact sexual incentive motivation test (SIMT) to evaluate sexual motivation. This male sexual behavior assessment method was used mostly in rats, but we adapted this method for mice. Testosterone (TSTO) is a pivotal hormone involved in regulating male sexual function, acting both at the central and peripheral levels [24]. The analysis of sexual behaviors and TSTO blood levels in mice lacking TAAR1 allows for the evaluation of potential risks related to future TAAR1-based therapies.

Most investigations in the TAAR1 field were focused on brain neurotransmission functions [25,26]. However, TAAR1 is known to also be widely expressed outside of the nervous system. Activated platelets can release PEA and TYR [27]. These compounds chemoattract neutrophils via the TAAR1 and TAAR2 heterodimer complex [28]. TAARs may be generally involved in the process of leukocyte recruitment to the injury sites [5]. At the same time, the lack of TAAR1 and TAAR5 does not lead to significant changes in platelets and other hematological parameters, even in older mice [29,30]. On the other hand, a recent study demonstrated that increased TAAR1 expression in monocytes mediated anti-inflammatory effects in multiple sclerosis [31]. As a result, one can expect a significant role of TAAR1 in immune regulation.

TAAR1 can also be involved in thyroid regulation. Primary cilia (PC) are microtubule-based sensory organelles with various receptors and channels involved in thyroid regulation [32]. The expression of TAAR1 was localized at the PC of thyroid epithelial cells in in vitro and in situ experiments [33]. Further studies demonstrated that the deletion of the TAAR1 gene led to phenotypic changes in thyroid morphology and its functional activity [34]. Furthermore, the visualization of the trafficking of mouse TAAR1 to the cilia of thyroid epithelial cells was performed with a green fluorescent protein [35]. In addition, it was proposed that high TAAR1 expression can be a positive prognosticator for overall survival in ovarian cancer patients [36]. Interestingly, ovarian cancer is regulated by thyroid hormones and their derivatives [37,38]. However, the role of TAARs in the non-canonical regulation of the thyroid system is still unclear, and further studies are needed.

TAAR1 receptor expression was also found in pancreatic β-cells, the stomach, and the intestines [39]. A significant role of TAAR1 and probably other TAARs in type 2 diabetes and obesity was indicated [40]. Thus, such an impact of TAAR1 on the biological system may lead to metabolic and lipid exchange imbalances. Recently, we found significantly decreased low-density lipoprotein cholesterol (LDL-cholesterol) changes in cholesterol levels in TAAR9 knockout rats [41]. Thus, the analysis of routine biochemical parameters in the blood of TAAR1 knockout (TAAR1-KO) mice is of interest. In a previous study, we investigated TAAR1-KO mice to evaluate the safety profile for TAAR1 potential treatments from the perspective of clinical hematology, basic behavioral tests, and thyroid regulation [29]. The current study focused on the sexual motivation behavior of TAAR1-KO mice, fulfilled with additional hormone parameters and biochemical screening.
2. Materials and Methods

2.1. Animals

All animal studies were carried out according to the Ministry of Health of Russian Federation guidelines and the principles adopted by the FELASA and RusLASA organizations’ welfare of laboratory animal use. All experiments were approved by the Saint Petersburg State University Ethical Committee for Animal Research (No. 131-03-1 of 16 July 2020). Wild-type (WT) and TAAR1-KO mice were derived by crossing (over 20 generations) heterozygous TAAR1 C57BL6/129SvJ animals. Experimental male mice (30 weeks old) and WT female mice (14 weeks old) were housed 3–5/cage, maintained under standard lab conditions (room temperature and humidity were 21 ± 5 °C and 40–70%, respectively), and provided with food and water ad libitum. All experiments were conducted during the light phase. The mice were habituated to the experimental room for at least 1 h before the behavioral experiments.

2.2. Sample Collection and Storage

To prepare serum for biochemical screening and automated ELISA, mice were decapitated, and blood was collected into VACUETTE blood collection tubes for serum (Greiner Bio-One, Austria, Kremsmünster), incubated in a vertical position for 15 min, and then kept at +4 °C until centrifugation. Samples with coagulated blood were centrifuged at 1500 rpm for 15 min at +4 °C. Serum was transferred into dry clean tubes and stored until analysis at −20 °C for no more than 3 days.

2.3. Measurement of Biochemical Parameters

TAAR1 biochemical screening was performed on automatic analyzer Random Access A-25 (Biosystems S.A., Spain, Barcelona), which was used utilizing the spectrophotometer principle. Serum samples were stored at −20 °C before analysis. The following biochemical parameters were analyzed: alanine aminotransferase (ALT), aspartate aminotransferase (AST), total protein, urea, triglycerides (TG), lactate dehydrogenase (LDH), creatine kinase, alkaline phosphatase (ALP), total cholesterol (TC), low-density lipoprotein cholesterol (LDLC), high-density lipoprotein cholesterol (HDLC), albumin, total bilirubin (TB), creatinine. The full data, number of samples and dilution factors are presented in the Supplementary Materials (Table S1).

2.4. Measurement of Testosterone

We used an automatic analyzer based on the ELISA principle, Advia Centaur XP (Siemens Healthineers, Germany, Erlangen), to measure testosterone. Serum samples were diluted with sterile pyrogen-free 0.9% sodium chloride solution in a ratio of 1:3.

2.5. Interassay Repeatability

Before analyzing the serum and blood samples, the equipment was decontaminated, calibrated, and checked by a laboratory quality control. Interassay repeatability was evaluated by calculating the coefficient of variation (CV) of 10 consecutive internal quality control material measurements in three different controls: high, normal, and low. CVs were calculated as standard deviation (SD)/mean × 100.

2.6. Sexual Incentive Motivation Test Protocol

1. Hormone-induced estrous. WT female mice in estrous were used as a sexual incentive. To induce estrous, adult female mice were given 10 µg estrogen benzoate and 500 µg progesterone intraperitoneally 48 and 2 h before the experiment, respectively [42]. The stage of the cycle was checked 1 h before the experiment via an assessment of vaginal smears [43].

2. Experimental setup and analysis. A modified sexual incentive motivation test (SIMT) was used to evaluate the sexual behavior of TAAR1-KO and WT male mice [44]. The setup consisted of 4 experimental chambers (15 × 30 cm²), each with an adjacent
incentive cage separated by a permeable wall (Figure 1). A female mouse in estrous
was placed in the incentive cage 20 min before the experiment. A male mouse was then
placed into the experimental chamber for 20 min, while the behavior was recorded
and then processed using Noldus EthoVision XT (Version 11.5; Noldus Information
Technology, Wageningen, The Netherlands). To speed up the data collection and
analysis, a set-up of 4 such independent cages was used simultaneously.

3. To assess the recognition of sexually relevant stimuli, two zones were differentiated
for the analysis: a 10 × 10 cm² zone adjacent to the incentive cage called “female”,
and a 4 × 3 cm² zone closest to the cage called “nose”. The following parameters
potentially descriptive of sexual behavior were analyzed: percentage of time spent in
the “female” zone by the center-point of the animal’s body, number of visits to the
“female” zone by the center-point, percentage of time spent in the “nose” zone by the
nose-point, number of visits to the “nose” zone by the nose-point.

4. Experimental design. Firstly, to assess the validity of the suggested method, 15 WT
male mice were tested in two conditions: in the presence of a sexual incentive and
with empty incentive cages. The parameters described in the previous paragraph
were compared to assess the validity of individual parameters and the method itself
for the evaluation of sexual behavior. 16 TAAR1-KO and 15 WT male mice were tested
in SIMT.

![Figure 1. Modified sexual incentive motivation test setup. Setup includes experimental chamber (15 × 30 × 50 cm³) and incentive cage separated by a wire-screen (dashed line) from each chamber. White circle with female mouse—incitement cage; blue square—“female” zone, 10 × 10 cm²; purple rectangle—“nose” zone, 4 × 3 cm².](image)

2.7. Statistical Analysis

In the SIMT, a two-way analysis of variance (ANOVA) with repeated measures was
used to compare all data, which was preliminarily tested for Gaussian distribution with the
D’Agostino-Pearson normality test. Hormonal and biochemical parameters data between
two groups were analyzed using a non-parametric Mann–Whitney test. Analyses were
performed using GraphPad Prizm 8 (GraphPad Software, San Diego, CA, USA).

3. Results

3.1. Lack of TAAR1 Does Not Affect Sexual Motivation

Figure 2a,b demonstrate the most critical parameters of the SIMT test. The control
group (CTRL) without female mice in chambers shows that in the absence of sexual
incentive, male mice have no place preference within the experimental area (Figure 2c).
There was a statistically significant effect of the presence of females on the number of visits
to the female zone (Figure 2c) and the percent of the time in the nose zone (Figure 2b) of
WT mice (F (1, 107) = 20.93, p < 0.0001 and F (2, 167) = 26.81, p < 0.0001, respectively).
Figure 2a,b demonstrate the most critical parameters of the SIMT test. The control group (CTRL) without female mice in chambers shows that in the absence of sexual incentive, male mice have no place preference within the experimental area (Figure 2c). There was a statistically significant effect of the presence of females on the number of visits to the female zone (Figure 2c) and the percent of the time in the nose zone (Figure 2b) of WT mice ($F(1, 107) = 20.93, p < 0.0001$ and $F(2, 167) = 26.81, p < 0.0001$, respectively).

In the presence of sexual incentive, there are no significant differences in the number of visits to the female zone (Figure 2a) and the percent of the time in the nose zone (Figure 2b), between WT and TAAR1-KO mice ($F(1, 115) = 1.246, p = 0.2666$ and $F(1, 115) = 1.254, p = 0.2651$, respectively). All other sexual motivation behavioral parameters also revealed minimal alterations. The complete results of the SIMT tests are presented in the Supplementary Materials (Figure S1). In addition, the blood testosterone analysis (Figure 3a) did not reveal significant differences between WT and TAAR1-KO mice.
Figure 3. Comparative analysis of basic biochemical and hormonal parameters in the blood of TAAR1-KO and WT mice. (a) Testosterone, (b) alanine aminotransferase (ALT), (c) aspartate aminotransferase (AST), (d) total protein, (e) urea, (f) de Ritis ratio (AST/ALT), (g) lactate dehydrogenase (LDH), (h) creatine kinase, (i) alkaline phosphatase (ALP), (j) albumin, (k) total bilirubin (TB), (l) creatinine. The biochemical screening did not reveal significant differences in any of the demonstrated parameters. Only the creatine kinase comparative analysis shows minimal alterations (WT = 741.7 ± 75.02; KO = 463.5 ± 78.47). Data are mean ± SEM. * p < 0.05.

3.2. TAAR1 Gene Knockout Does Not Significantly Affect Biochemical Parameters

The comparative analysis of TAAR1 and WT did not reveal significant differences in major biochemical parameters, such as alanine aminotransferase (ALT), aspartate aminotransferase (AST), total protein, urea, triglycerides (TG), lactate dehydrogenase (LDH), alkaline phosphatase (ALP), total cholesterol (TC), low-density lipoprotein cholesterol (LDLC), high-density lipoprotein cholesterol (HDLC), albumin, total bilirubin (TB), creatinine (Figures 3b and 4). Only creatinine kinase levels were significantly decreased in mutant mice (Figure 3h).
we assessed hematological parameters with a routine biochemistry screening panel. TAAR1-
TAAR9) are also similarly involved in the transmission of innate odors into limbic brain
parameters. Data are mean ± SEM.

Figure 4. Comparative analysis of main lipid exchange biochemical parameters and ratios in the
blood of TAAR1-KO and WT mice. (a) Total cholesterol (TC), (b) high-density lipoprotein cholesterol
(HDLC), (c) low-density lipoprotein cholesterol (LDLC), (d) total cholesterol/high-density lipoprotein
ratio (TC/HDLC), (e) ratio of low-density lipoprotein cholesterol and high-density lipoprotein
cholesterol (LDLC/HDLC), (f) triglycerides (TG). There are only minimal alterations in all presented
parameters. Data are mean ± SEM.

4. Discussion

In the present study, we adapted a SIMT behavioral test for mice and evaluated the
effect of TAAR1 gene deletion on sexual motivation and testosterone levels. Furthermore,
we assessed hematological parameters with a routine biochemistry screening panel. TAAR1-
based therapies have a strong potential in the treatment of several human disorders such
as schizophrenia, addiction, depression, diabetes, and obesity [4,5]. TAAR1 agonists have
already entered phase III clinical trials to treat schizophrenia [7]. Therefore, it is essential to
consider the potential side effects of TAAR1-based therapies and preliminarily evaluate the
safety profile in the periphery.

There are no commonly accepted protocols concerning non-contact sexual motivation
and social recognition that have been reliably established for mice. Therefore, we had to
integrate several features from the existing rat test paradigm [45]. Classic tests often use
additional males as social recognition validation objects. However, the social hierarchies
of rodents are regulated by odor and sniffing interactions [46]. Furthermore, rodents are
sensitive to other male odor stimuli, and the existence of alpha males in the experimental
arena may affect sexual motivation and lead to additional social stress [47]. Thus, only
male and female mice socially interacted through the cage in our experiment. It should be
noted, however, that the lack of male conspecifics does not exclude the possibility that the
female was approached not only due to sexual motivation, but also due to social interaction
or a combination of these. Moreover, the arena parameters were changed to accommodate
the smaller size of mice. Finally, we performed a fast screening neurobiological test, which
allows one to evaluate sexual motivation in mice quickly. The adapted SIMT method can
be used in future pharmacological experiments and as a preliminary step in copulatory
sexual tests.

While TAAR1-KO mice demonstrated minimal alterations in sexual motivation, further
studies of this kind in the TAAR field are warranted. Recent studies revealed TAAR5
localization in the glomerular and deeper olfactory bulb layers projecting to the limbic brain
olfactory circuitry [21]. It is well established that sexual behavior is regulated in mice via the
olfactory system [48], and the TAAR5 agonist trimethylamine is considered a pheromone
in mice [49]. Moreover, TAAR5 knockout mice show increased adult neurogenesis and a
higher number of dopamine neurons [22]. It is likely that other “olfactory” TAARs (TAAR2-
TAAR9) are also similarly involved in the transmission of innate odors into limbic brain
areas regulating emotions and adult neurogenesis [23]. Thus, the evaluation of the role of TAARs in sexual behaviors may become a prospective direction for future studies.

The analysis of testosterone and other routine blood biochemical parameters in TAAR1-KO and WT mice also demonstrated minimal alterations. Only creatine kinase showed a significant decrease in mutant mice. Such changes may be related to an increased locomotor activity in TAAR1-KO mice [50]. These observations indicate that despite the known expression of TAAR1 in pancreatic β-cells, the stomach, the intestines, the thyroid gland, and leucocytes, the lack of TAAR1 minimally affects lipid and metabolic processes in normal conditions. Potentially, TAAR1-mediated non-canonical mechanisms in the periphery could be revealed under pharmacological or specialized diet challenge conditions.

In conclusion, even such a drastic manipulation as the elimination of TAAR1 did not cause significant alterations in sexual or social motivation, testosterone levels, and blood biochemical parameters. These observations suggest that future TAAR1-based therapies will likely have a good safety profile.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/brainsci12050614/s1, Figure S1: Comparative SIMT analysis of TAAR1-KO and WT mice; Figure S2: Additional parameters of SIMT test; Table S1: Comparative biochemical and hormonal analysis of TAAR1-KO and WT mice.

Author Contributions: Conceptualization, I.S.Z., A.B.V., I.V.K. and R.R.G.; SIMT methodology, I.S.Z., M.A.P., I.Y.T. and A.B.V.; formal analysis, I.S.Z., A.B.V. and M.A.P.; investigation, I.S.Z., E.A.Z., I.Y.T. and I.V.K.; E.L.S. (performed the experiments with biochemical and testosterone parameters); resources, R.R.G.; data curation, I.S.Z., M.A.P., A.B.V. and R.R.G.; writing—original draft preparation, I.S.Z. and R.R.G.; writing—review and editing, I.S.Z., M.A.P., E.A.Z., I.V.K., A.B.V. and R.R.G.; visualization, I.S.Z., M.A.P., A.B.V. and R.R.G.; supervision, R.R.G.; project administration, R.R.G.; funding acquisition, R.R.G. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by the project ID: 93018770 of the St. Petersburg State University, St. Petersburg, Russia.

Institutional Review Board Statement: The experiments described in this paper were approved by the Saint Petersburg State University Ethical Committee for Animal Research (protocol No. 131-03-1 of 16 July 2020) according to the RusLASA and FELASA organizations’ welfare of laboratory animal use.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data will be available upon request from the corresponding author.

Acknowledgments: Breeding and support of TAAR1-KO mice was performed by the Resource Center Vivarium of the Institute of Translational Biomedicine, St. Petersburg State University, St. Petersburg, Russia.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References
1. Bunzow, J.R.; Sonders, M.S.; Arttamangkul, S.; Harrison, L.M.; Zhang, G.; Quigley, D.I.; Darland, T.; Suchland, K.L.; Pasumamula, S.; Kennedy, J.L.; et al. Amphetamine, 3,4-methylenedioxymethamphetamine, lysergic acid diethylamide, and metabolites of the catecholamine neurotransmitters are agonists of a rat trace amine receptor. Mol. Pharmacol. 2001, 60, 1181–1188. [CrossRef] [PubMed]
2. Borowsky, B.; Adham, N.; Jones, K.A.; Raddatz, R.; Artymyshyn, R.; Ogozalek, K.L.; Durkin, M.M.; Lakhmani, P.P.; Bonini, J.A.; Pathirana, S.; et al. Trace amines: Identification of a family of mammalian G protein-coupled receptors. Proc. Natl. Acad. Sci. USA 2001, 98, 8966–8971. [CrossRef] [PubMed]
3. Premont, R.T.; Gainetdinov, R.R.; Caron, M.G. Following the trace of elusive amines. Proc. Natl. Acad. Sci. USA 2001, 98, 9474. [CrossRef] [PubMed]
4. Berry, M.D.; Gainetdinov, R.R.; Hoemer, M.C.; Shahid, M. Pharmacology of human trace amine-associated receptors: Therapeutic opportunities and challenges. Pharmacol. Ther. 2017, 180, 161–180. [CrossRef]
5. Gainetdinov, R.R.; Hoener, M.C.; Berry, M.D. Trace Amines and Their Receptors. *Pharmacol. Rev.* 2018, 70, 549–620. [CrossRef]

6. Boulton, A.A. Letter: Amines and theories in psychiatry. *Lancet* 1974, 2, 52–53. [CrossRef]

7. Koblan, K.S.; Kent, J.; Hopkins, S.C.; Krystal, J.H.; Cheng, H.; Goldman, R.; Loebel, A. A Non–D2-Receptor-Binding Drug for the Treatment of Schizophrenia. *N. Engl. J. Med.* 2020, 382, 1497–1506. [CrossRef]

8. Everitt, B.J. Sexual motivation: A neural and behavioural analysis of the mechanisms underlying appetitive and copulatory responses of male rats. *Neurosci. Biobehav. Rev.* 1990, 14, 217–232. [CrossRef]

9. Pfaus, J.G.; Phillips, A.G. Role of Dopamine in Anticipatory and Consummatory Aspects of Sexual Behavior in the Male Rat. *Behav. Neurosci.* 1991, 105, 727–743. [CrossRef]

10. Hull, E.M.; Du, J.; Lorrain, D.S.; Matuszewich, L. Extracellular dopamine in the medial preoptic area: Implications for sexual motivation and hormonal control of copulation. *J. Neurosc.* 1995, 15, 7465–7471. [CrossRef]

11. Argiolas, A.; Melis, M.R. Central control of penile erection: Role of the paraventricular nucleus of the hypothalamus. *Prog. Neurobiol.* 2005, 76, 1–21. [CrossRef] [PubMed]

12. Melis, M.R.; Argiolas, A. Central control of penile erection: A re-visititation of the role of oxytocin and its interaction with dopamine and glutamic acid in male rats. *Neurosci. Biobehav. Rev.* 2011, 35, 939–955. [CrossRef] [PubMed]

13. Argiolas, A.; Melis, M.R. Neuropeptides and central control of sexual behaviour from the past to the present: A review. *Prog. Neurobiol.* 2013, 108, 80–107. [CrossRef] [PubMed]

14. Pfaus, J.G. Dopamine: Helping Males Copulate for at Least 200 Million Years: Theoretical Comment on Kleitz-Nelson et al. (2010). *Behav. Neurosci.* 2010, 124, 877–880. [CrossRef] [PubMed]

15. Sanna, F.; Bratzu, J.; Piludu, M.A.; Corda, M.G.; Melis, M.R.; Giorgi, O.; Argiolas, A. Dopamine, noradrenaline and differences in sexual behavior between Roman high and low avoidance male rats: A microdialysis study in the medial prefrontal cortex. *Front. Behav. Neurosci.* 2017, 11, 108. [CrossRef]

16. Hull, E.M.; Dominguez, J.M. Male Sexual Behavior. In *Knobil and Neill’s Physiology of Reproduction*; Academic Press: Cambridge, MA, USA, 2015; pp. 2211–2285. [CrossRef]

17. Sanna, F.; Bratzu, J.; Serra, M.P.; Leo, D.; Quartu, M.; Boi, M.; Espinoza, S.; Gainetdinov, R.R.; Melis, M.R.; Argiolas, A. Altered Sexual Behavior in Dopamine Transporter (DAT) Knockout Male Rats: A Behavioral, Neurochemical and Intracerebral Microdialysis Study. *Front. Behav. Neurosci.* 2020, 14, 58. [CrossRef]

18. Horowitz, L.F.; Saraiva, L.R.; Kuang, D.; Yoon, K.H.; Buck, L.B.; Horowitz, L.F.; Saraiva, L.R.; Yoon, K.H. Olfactory receptor patterning in a higher primate. *J. Neurosci.* 2014, 34, 12241–12252. [CrossRef]

19. Liberles, S.D.; Buck, L.B. A second class of chemosensory receptors in the olfactory epithelium. *Nature* 2006, 442, 645–650. [CrossRef]

20. Syed, A.S.; Sansone, A.; Röner, S.; Bozorg Nia, S.; Manzini, I.; Korsching, S.I. Different expression domains for two closely related amphibian TAARs generate a bimodal distribution similar to neuronal responses to amine odors. *Sci. Rep.* 2015, 5, 13935. [CrossRef]

21. Espinoza, S.; Sukhanov, I.; Efimova, E.V.; Kozlova, A.; Antonova, K.A.; Illiano, P.; Leo, D.; Merkulyeva, N.; Kalinina, D.; Musienko, P.; et al. Trace Amine-Associated Receptor 5 Provides Olfactory Input Into Limbic Brain Areas and Modulates Emotional Behaviors and Serotonin Transmission in mice. *Front. Mol. Neurosci.* 2020, 13, 18. [CrossRef]

22. Efimova, E.V.; Kozlova, A.A.; Razenko, V.; Katolikova, N.V.; Antonova, K.A.; Sotnikova, T.D.; Merkulyeva, N.S.; Veshchitskii, A.S.; Kalinina, D.S.; Korzhevskii, D.E.; et al. Increased dopamine transmission and adult neurogenesis in trace amine-associated receptor 5 (TAAR5) knockout mice. *Neuropsychopharmacology* 2021, 182, 108373. [CrossRef] [PubMed]

23. Efimova, E.V.; Katolikova, N.V.; Kanov, E.V.; Gainetdinov, R.R. Trace amine-associated receptors at the cross-road between innate olfaction of amines, emotions, and adult neurogenesis. *Neural Regen. Res.* 2022, 17, 1257–1258. [CrossRef] [PubMed]

24. Vignozzi, L.; Corona, G.; Petrone, L.; Filippi, S.; Morelli, A.M.; Forti, G.; Maggi, M. Testosterone and sexual activity. *J. Endocrinol. Invest.* 2005, 28, 39–44. [PubMed]

25. Sukhanov, I.; Espinoza, S.; Yakovlev, D.S.; Hoener, M.C.; Sotnikova, T.D.; Gainetdinov, R.R. TAAR1-dependent effects of apomorphine in mice. *Int. J. Neuropsychopharmacol.* 2014, 17, 1683–1693. [CrossRef] [PubMed]

26. Espinoza, S.; Lignani, G.; Caffino, L.; Maggi, S.; Sukhanov, I.; Leo, D.; Mus, L.; Emanuelue, M.; Ronzitti, G.; Harmeier, A.; et al. TAAR1 Modulates Cortical Glutamate NMDA Receptor Function. *Neuropsychopharmacology* 2015, 40, 2217–2227. [CrossRef]

27. D’Andrea, G.; Terrazzino, S.; Fortin, D.; Farruggio, A.; Rinaldi, L.; Leon, A. HPLC electrochemical detection of trace amines in human plasma and platelets and expression of mRNA transcripts of trace amine receptors in circulating leukocytes. *Neurosci. Lett.* 2003, 346, 89–92. [CrossRef]

28. Babusyte, A.; Kotthoff, M.; Fiedler, J.; Krautwurst, D. Biogenic amines activate blood leukocytes via trace amine-associated receptors TAAR1 and TAAR2. *J. Leukoc. Biol.* 2013, 93, 387–394. [CrossRef]

29. Zhukov, I.S.; Kubarskaya, L.G.; Tissen, I.Y.; Kozlova, A.A.; Dagayev, S.G.; Kashuro, V.A.; Vlasova, O.L.; Sinitsa, E.L.; Karpova, I.V.; Gainetdinov, R.R. Minimal Age-Related Alterations in Behavioral and Hematical Parameters in Trace Amine-Associated Receptor 1 (TAAR1) Knockout Mice. *Cell. Mol. Neurobiol.* 2020, 40, 273–282. [CrossRef]

30. Zhukov, I.S.; Kubarskaya, L.G.; Karpova, I.V.; Vaganova, A.N.; Karpenko, M.N.; Gainetdinov, R.R. Minor changes in erythrocyte osmotic fragility in trace amine-associated receptor 5 (Taar5) knockout mice. *Int. J. Mol. Sci.* 2021, 22, 7307. [CrossRef]

31. Barnes, D.A.; Galloway, D.A.; Hoener, M.C.; Berry, M.D.; Moore, C.S. Taar1 expression in human macrophages and brain tissue: A potential novel facet of ms neuroinflammation. *Int. J. Mol. Sci.* 2021, 22, 11576. [CrossRef]
32. Lee, J.; Sul, H.J.; Kim, K.H.; Chang, J.Y.; Shong, M. Primary Cilia Mediate TSH-Regulated Thyroglobulin Endocytic Pathways. *Front. Endocrinol.* 2021, 12, 1075. [CrossRef] [PubMed]

33. Szumskas, J.; Qatato, M.; Rehders, M.; Führer, D.; Biebermann, H.; Grandy, D.K.; Köhrle, J.; Brix, K. Trace Amine-Associated Receptor 1 Localization at the Apical Plasma Membrane Domain of Fisher Rat Thyroid Epithelial Cells Is Confined to Cilia. *Eur. Thyroid J.* 2015, 4, 30–41. [CrossRef] [PubMed]

34. Qatato, M.; Szumskas, J.; Skripnik, V.; Rijnkjes, E.; Köhrle, J.; Brix, K. Canonical TSH Regulation of Cathepsin-Mediated Thyroglobulin Processing in the Thyroid Gland of Male Mice Requires Taar1 Expression. *Front. Pharmacol.* 2018, 9, 221. [CrossRef] [PubMed]

35. Qatato, M.; Venugopalan, V.; Al-Hashimi, A.; Rehders, M.; Valentine, A.D.; Hein, Z.; Dallito, U.; Springer, S.; Brix, K. Trace Amine-Associated Receptor 1 Trafficking to Cilia of Thyroid Epithelial Cells. *Cells* 2021, 10, 1518. [CrossRef] [PubMed]

36. Vogelsang, T.I.R.; Vattai, A.; Schmoeckel, E.; Kaltofen, T.; Chelariu-Raicu, A.; Zheng, M.; Mahner, S.; Mayr, D.; Jeschke, U.; Trillisch, F. Trace amine-associated receptor 1 (Taar1) is a positive prognosticator for epithelial ovarian cancer. *Int. J. Mol. Sci.* 2021, 22, 8479. [CrossRef] [PubMed]

37. Shinderman-Maman, E.; Cohen, K.; Moskovich, D.; Hercbergs, A.; Werner, H.; Davis, P.J.; Ellis, M.; Ashur-Fabian, O. Thyroid hormones derivatives reduce proliferation and induce cell death and DNA damage in ovarian cancer. *Sci. Rep.* 2017, 7, 16475. [CrossRef] [PubMed]

38. Mousa, S.A.; Lin, H.Y.; Tang, H.Y.; Hercbergs, A.; Luidens, M.K.; Davis, P.J. Modulation of angiogenesis by thyroid hormone and hormone analogues: Implications for cancer management. *Angiogenesis* 2014, 17, 463–469. [CrossRef]

39. Revel, F.G.; Moreau, J.-L.; Gainetdinov, R.R.; Ferragud, A.; Velázquez-Sánchez, C.; Sotnikova, T.D.; Moraity, S.R.; Harmeier, A.; Groebeke Zbinden, K.; Norcross, R.D.; et al. Trace amine-associated receptor 1 partial agonism reveals novel paradigm for neuropsychiatric therapeutics. *Biol. Psychiatry* 2012, 72, 934–942. [CrossRef]

40. Raab, S.; Wang, H.; Uhles, S.; Cole, N.; Alvarez-Sánchez, R.; Künecke, B.; Ullmer, C.; Matile, H.; Bedoucha, M.; Norcross, R.D.; et al. Incretin-like effects of small molecule trace amine-associated receptor 1 agonists. *Mol. Metab.* 2016, 5, 47–56. [CrossRef]

41. Murtazina, R.Z.; Zhukov, I.S.; Korenkova, O.M.; Popova, E.A.; Kuvarzin, S.R.; Efimova, E.V.; Kubarskaya, L.G.; Batotsyrenova, E.G.; Zolotoverkhaya, E.A.; Vaganova, A.N.; et al. Genetic deletion of trace-amine associated receptor 9 (TAAR9) in rats leads to decreased blood cholesterol levels. *Int. J. Mol. Sci.* 2021, 22, 2942. [CrossRef]

42. Liu, Y.; Wu, Y. Progesterone Intramuscularly or Vaginally Administration May Not Change Live Birth Rate or Neonatal Outcomes in Artificial Frozen-Thawed Embryo Transfer Cycles. *Front. Endocrinol.* 2020, 11, 539427. [CrossRef] [PubMed]

43. Cora, M.C.; Kooistra, L.; Travlos, G. Vaginal Cytology of the Laboratory Rat and Mouse: Review and Criteria for the Staging of the Estrous Cycle Using Stained Vaginal Smears. *Toxicol. Pathol.* 2015, 43, 776–793. [CrossRef] [PubMed]

44. Bai, Y.; Li, Y.; Lv, Y.; Liu, Z.; Zheng, X. Complex motivated behaviors for natural rewards following a binge-like regimen of morphine administration: Mixed phenotypes of anhedonia and craving after short-term withdrawal. *Front. Behav. Neurosci.* 2014, 8, 23. [CrossRef] [PubMed]

45. Ågmo, A. Unconditioned Sexual Incentive Motivation in the Male Norway Rat (*Rattus norvegicus*). *J. Comp. Psychol.* 2003, 117, 3–14. [CrossRef] [PubMed]

46. Wesson, D.W. Sniffing Behavior Communicates Social Hierarchy. *Curr. Biol.* 2013, 23, 575–580. [CrossRef]

47. Caramaschi, D.; de Boer, S.F.; de Vries, H.; Koolhaas, J.M. Development of violence in mice through repeated victory along with changes in prefrontal cortex neurochemistry. *Behav. Brain Res.* 2008, 189, 263–272. [CrossRef]

48. Ishii, K.K.; Touhara, K. Neural circuits regulating sexual behaviors via the olfactory system in mice. *Brain Sci.* 2022, 12, 614. [CrossRef]

49. Wallraabenstein, I.; Kuklan, J.; Weber, L.; Zborala, S.; Werner, M.; Altmüller, J.; Becker, C.; Schmidt, A.; Hatt, H.; Hummel, T.; et al. Human trace amine-associated receptor TAAR5 can be activated by trimethylamine. *PLoS ONE* 2013, 8, e54950. [CrossRef]

50. Sukhanov, I.; Dorofeikova, M.; Dolgorukova, A.; Dorotenko, A.; Gainetdinov, R.R. Trace amine-associated receptor 1 modulates the locomotor and sensitization effects of nicotine. *Front. Pharmacol.* 2018, 9, 329. [CrossRef]