miR-155 induction in microglial cells suppresses Japanese encephalitis virus replication and negatively modulates innate immune responses

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Abstract

Background: Microglial cells, which are resident macrophages of the central nervous system, play important roles in immune responses and pathogenesis. Japanese encephalitis virus (JEV) is a neurotropic virus that infects microglial cells in brain. Several microRNAs including miR-155 and miR-146a play an important role in defining the microglia inflammatory profile. In this study, we have investigated the effect of miR-155 and miR-146a modulation on JEV infection as well as innate immune responses in human microglial cells.

Methods: In vitro studies were performed in JEV-infected human microglial CHME3 cells. miR-155 or miR-146a were overexpressed and total RNA and protein were extracted following JEV-infection. Expression of genes involved in innate immune responses was studied by PCR array, quantitative real-time PCR (qPCR), western blot and Fluorescence activated cell sorter (FACS). JEV replication was monitored by studying the viral RNA by qPCR, protein by western blot, and titres by plaque assay.

Results: Overexpression of miR-155 in CHME3 cells resulted in significantly reduced JEV replication whereas miR-146a overexpression had an insignificant effect. Additionally, interferon regulatory factor 8 (IRF8) and complement factor H (CFH) were induced during JEV infection; however, this induction was attenuated in miR-155 overexpressing cells following JEV infection. Further, JEV-induced NF-κB regulated downstream gene expression was attenuated. Interestingly, an increased level of CD45, a negative regulator of microglia activation and a reduced phosphorylated-Signal Transducers and Activators of Transcription (p-STAT1) expression was observed in miR-155 overexpressing cells upon JEV infection.

Conclusion: Induction of miR-155 in human microglial cells may negatively modulate JEV-induced innate immune gene expression and may have a beneficial role in limiting JEV replication in human microglial cells.

Keywords: NeurimmiRs, JEV, CD45, Microglia activation

Background

Japanese encephalitis (JE) is an acute central nervous system (CNS) inflammatory disease caused by infection with Japanese encephalitis virus (JEV); a small, enveloped, plus-strand RNA virus belonging to the Flaviviridae family. It is the leading cause of viral encephalitis in south-east Asia, India, and China where three billion people are at risk of contracting the disease, yet its pathogenesis remains poorly understood. While neurons are believed to be the primary target of JEV in the brain, a recent report has suggested that microglial cells can be directly infected with JEV [1,2]. Microglial cells are the resident immune cells of the CNS and have a critical role in host defense against invading pathogens. However, substantial evidence suggests that infection of microglia by JEV may actually lead to neuronal cell death through uncontrolled production of pro-inflammatory cytokines. Therefore, downregulation of cytokine production can serve to dampen the inflammatory response and can contribute to better virus clearance and increased protection against JEV [3]. Indeed, inhibition of chronic neuroinflammation, particularly due to microglial activation, has been
suggested to be a practical strategy in the treatment of neurodegenerative diseases [4,5].

Recently, a new class of regulatory RNAs, called microRNAs (miRNAs) have emerged that modulate immune response and play key regulatory roles in virus-host interactions. These miRNAs serve as universal regulators of differentiation, activation, and polarization of mammalian cells including microglia and macrophages in normal and diseased CNS [6]. Thus, modulation of cellular miRNA expression during viral infection may be an important determinant of disease outcome [7]. Recent reports suggested that a subset of miRNAs, termed as NeurimmiRs, co-exist in the brain and peripheral organs [8,9]. These miRNAs can affect both neuronal and immune functions and thus constitute important therapeutic targets for those diseases that affect both the immune system and brain functions. Among them, miR-155 and miR-146a are multifunctional and widely reported to modulate different stages of innate immune response during inflammation and infection [10-12]. Thus, miR-155 and miR-146a were shown to increase in the microglial cells in response to stimulation with Lipopolysaccharides (LPS) and Polyinosinic-polycytidylic acid (polyIC), respectively, and they seem to play a fundamental role in the microglial inflammatory profile [13,14]. These miRNAs are also associated with interferon (IFN) signaling pathways [15,16]. Moreover, miR-155 and miR-146a not only modulate Toll-like receptors (TLRs)-mediated innate immune response, but also target complement regulatory proteins and facilitate complement activation [17-19]. This phenomenon is very important to eliminate the virus from infected cells. Furthermore, both miR-146a and miR-155 have been shown to play an important role in viral infection. For example, Wu et al. [20] reported an increased dengue virus 2 (DENV2) replication in miR-146a overexpressing cells, whereas overexpression of miR-155 significantly suppressed human immunodeficiency virus (HIV) infection in activated macrophages [21].

Since JEV is a neurotropic virus it is likely that NeurimmiRs play an important role in virus replication and immunopathology. Using a global miRNA array we have identified differentially expressed NeurimmiRs in human microglial cells during the course of JEV infection. Of these, we have focused our study on miR-155 and miR-146a and have investigated their effect on JEV replication and their role in the modulation of microglia-mediated innate immune response during JEV infection. For this purpose, in vitro studies were performed in JEV-infected human microglial CHME3 cells. Our results indicate that miR-155 induction might have a beneficial role for the host by limiting JEV replication through modulation of microglia-mediated innate immune responses.

Materials and methods

Cells, antibodies, miRNA mimics, and inhibitors

Human microglial cells (CHME3) were provided by the National Brain Research Centre, Manesar, India. Porcine stable kidney (PS) cell line was procured from National Centre for Cell Science, Pune, India. CHME3 cells were grown in Dulbecco’s Modified Eagle’s Medium (DMEM, Invitrogen, Carlsbad, CA, USA) and PS cells in Eagle’s Minimal Essential Medium (MEM, Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (FBS), 2 mM L-glutamine, and 100 μg/ml penicillin-streptomycin (Invitrogen, Carlsbad, CA, USA). Primary antibodies against MyD88, Ikke, IRF8, p-STAT1, STAT1 and HRP-conjugated secondary antibodies were purchased from Cell Signaling Technology, Beverly, MA, USA. CD-45 antibodies were from BD Biosciences, San Jose, CA, USA. Rabbit antibody against JEV NS1 protein was produced in-house. MicroRNA mimics and inhibitors were from Invitrogen, Carlsbad, CA, USA (Assay ID: MC12601, mimic for hsa-miR-155-5p; MH12601, inhibitor for hsa-miR-146a-5p; MC10722, mimic for hsa-miR-155-5p; MH10722, inhibitor for hsa-miR-146a-5p, mirVana® miRNA Mimic Control #1). NF-kB inhibitor ammonium pyrrolidine dithiocarbamate, and PI3K inhibitor LY294002 were from Sigma-Aldrich, Saint Louis, MO, USA.

Transfection of cells and virus infection

The P20778 strain of JEV was propagated in PS cells and titrated by plaque assay [22]. Microglial cells were seeded in 6-well tissue culture plates at a density of 0.5 × 10⁶ cells/well and transfected 24 hours later using Lipofectamine 2000® reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer’s protocol. For over-expression or inhibition studies, 25 pmol of a miRNA mimic or inhibitor, respectively, was transfected. Cells were washed with 1 × PBS after 24 hours transfection and infected with JEV at multiplicity of infection 5 (MOI = 5). Culture supernatant was collected for virus titration as plaque forming unit (PFU)/ml and cells lysate was used for protein and RNA studies at different times post-infection (pi).

RT² Profiler PCR array

The Human NFκB Signaling Targets PCR array (#PAHS-0225Z) (SA Biosciences/Qiagen, Hilden, Germany) was used to determine the profile of genes associated with the human innate and adaptive immune responses. Total RNA was extracted from JEV-infected and uninfected or specific mimic- and inhibitor-transfected CHME3 cells using RNeasy mini kit (Qiagen, Hilden, Germany) with inclusion of a DNase I treatment step. cDNA was prepared from 1 μg total RNA using a RT² PCR array first strand kit (Qiagen, Hilden, Germany). Quantitative real-
time PCR (qPCR) was performed with an ABI PRISM 7500 (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's instructions.

**Western blot analysis**

Control and treated microglial cells were washed with 1 × PBS and lysed in cell lysis buffer (Sigma-Aldrich, Saint Louis, MO, USA) in the presence of protease inhibitor cocktail (Roche Diagnostics, Basel, Switzerland). The concentration of the protein lysate was determined using the Bradford method. The protein sample (50 μg) was electrophoresed on 10% sodium dodecyl sulfate-polyacrylamide gel (SDS-PAGE) and transferred onto a nitrocellulose membrane. The membrane was then blocked using 5% non-fat dry milk in PBST (PBS containing 0.05% Tween-20) for one hour at room temperature on a shaker. After blocking, the membrane was incubated with rabbit anti-human primary antibody overnight at 4°C with gentle shaking. After three washes of the membrane for 10 minutes each with PBST, the western blots were developed using chemiluminescence reagents (Santa Cruz Biotechnology, Dallas, Texas, USA). The following thermal cycling profile was used for the PCR analysis: 95°C for 15 minutes, 40 cycles at 94°C for 15 seconds, and 55°C for 30 seconds. Each qPCR reaction was performed in triplicate and the mean Ct value for each sample was used for data analysis. Expression levels of miRNAs were normalized to that of U6 snRNA.

**Flow cytometry assay**

Expression of CD-45 isoforms on CHME3 cells was examined by fluorescence activated cell sorter (FACS) analysis. Cells were fixed using 2% para-formaldehyde and stained with CD-45 antibody conjugated with Alexa-488 dye (BD Biosciences, San Jose, CA, USA) as per the manufacturer’s instructions. Samples were analyzed on FACS Calibur cell analyser (BD Biosciences, San Jose, CA, USA). Data were analyzed using FlowJo flow cytometry analysis software (Tree Star Inc, Ashland, OR, USA).

Quantitation of miRNA expression

Cells were lysed in TRIzol reagent (Invitrogen, Carlsbad, CA, USA) for RNA isolation and the total RNA was isolated using RNAspray kit (Qiagen, Hilden, Germany). RNA concentration was quantified using a NanoDrop 2000 spectrophotometer (Thermo Fisher, Waltham, MA, USA). RNA (1 μg) was reverse transcribed using prime script first-strand cDNA Synthesis kit (Takara Bio Inc. Otsu, Shiga, Japan) in a 20 μl reaction according to the manufacturer’s protocol. The expression of cellular genes was studied by qPCR with the fluorescent DNA-binding dye SYBR green (Power SYBR Green PCR master kit; Applied Biosystems, Foster City, CA, USA) by the real-time fluorescence detection method. JEV RNA was quantified as described earlier [23]. Each quantitative PCR reaction was performed in triplicate and the mean threshold cycle (Ct) value for each sample was used for data analysis. The RNA transcript levels were normalized to that of GAPDH.

**Results**

**JEV infection modulates expression of NeurimmIRs in microglial cells**

In order to understand how JEV infection modulates immune pathogenesis-related miRNAs, we carried out a global human miRNA array study to identify differentially expressed miRNAs in human microglial CHME3 cells in response to JEV infection. Of the several miRNAs modulated during the course of infection, we focused here on a subset of seven miRNAs that have previously been defined as NeurimmIRs [8,9]. Following the bioinformatics and statistical analysis, we found that six of these NeurimmIRs were differentially expressed in CHME3 cells during JEV infection (Figure 1A). We observed that miR-125b and miR-132 remained down-regulated throughout the course of infection. While miR-146a and miR-326 were up-regulated at an early time point (6 hours pi), these were down-regulated as
the infection progressed. On the other hand, miR-124 and miR-155 were suppressed at early time points, but were up-regulated at later time points. No change was seen in miR-212 levels during JEV infection.

As this study focused on miR146a and miR-155 we validated their expression in human microglial cells by qPCR using the TaqMan assay. The qPCR data generally conformed to the microarray data. Thus, miR-146a expression levels showed a small increase at 6 hours pi followed by a reduction at 24 hours pi. For miR-155 however, the expression levels showed a small decrease at 6 hours pi that was followed by a moderate (approximately 1.8-fold) increase at 48 hours pi (Figure 1B).

miR-155 suppresses JEV replication in microglial cells

In order to understand the effect of miR-155 and miR-146a modulation on JEV replication, we overexpressed these miRNAs in microglial cells by transfecting specific mimics and infected the cells with JEV. At 24 hours pi, levels of the JEV NS1 protein and JEV RNA were significantly reduced in miR-155 overexpressing microglial cells, but these were largely unaffected in miR-146a overexpressing cells (Figure 2A and B). A significant suppression of JEV titre (approximately 1 log2) was observed in miR-155 overexpressing cells while miR-146a overexpression had no effect on JEV titre (Figure 2C). These data clearly show that miR-155 had an inhibitory effect on JEV replication in microglial cells.

miR-155 modulates JEV-induced IFN-β expression in microglial cells

The innate immune response involving the induction of type I IFN constitutes the first line of antiviral defenses and could contribute to reduced virus replication. Since miR-155 and miR-146a are known to modulate IFN-β expression in peripheral organs [15,16], we focused on
how these miRNAs affect IFN-β mRNA expression in JEV-infected microglial cells. For this, we overexpressed these miRNAs in CHME3 cells by transfecting specific mimic (mimic-155 or mimic-146a) 24 hours prior to JEV infection. Our data showed that JEV-induced IFN-β mRNA expression in CHME3 cells and the levels of this JEV-induced IFN-β mRNA were significantly reduced in miR-155 overexpressing cells, while only a marginal effect was observed in miR-146a overexpressing cells (Figure 3A). In fact, the suppressive effect of miR-155 on IFN-β mRNA expression was reversed when the miR-155 inhibitor was used. Thus cells transfected with the miR-155 inhibitor prior to JEV infection showed an enhanced IFN-β mRNA expression when compared to control mimic-transfected cells infected with JEV (Figure 3A).

Infection of mammalian cells by viruses leads to expression of interferon-stimulated genes (ISG) [24]. We have studied the effect of miR-155 and miR-146a overexpression on ISG expression in JEV-infected microglial cells. Figure 3B shows that JEV-induced ISG expression was significantly repressed in miR-155 overexpressing cells, whereas it was only marginally affected in miR-146a overexpressing cells. The suppressive effect of miR-155 overexpression on IRF7 expression could be decreased by the miR-155 inhibitor (Figure 3C). These data indicate that JEV infection of miR-155 overexpressing microglial cells resulted in a reduced expression of IFN-β and ISG.

miR-155 attenuates JEV-induced IRF8 expression in microglial cells

The interferon regulatory factor (IRF) family of transcription factors regulate the entire type I IFN system. IRF8, a member of IRF family, is a myeloid lineage transcriptional regulator that plays an essential role in the microglial activation and is involved in inducing innate immune response. IRF8 and IRF3 can cooperatively induce rapid IFN-β production in monocytes [25]. It is also reported that TLR receptor signaling can activate NF-κB through IRF8 in myeloid lineage cells [26]. We therefore, studied IRF3 and IRF8 expression in human microglial cells during JEV infection. Our results suggested that IRF8 was induced during JEV infection and significantly up-regulated mRNA levels were observed at 48 hours pi. However, there was relatively little increase in IRF3 expression during JEV infection (Figure 4A, left panel). Western blot results corroborated this finding where enhanced IRF8 protein levels were seen in JEV-infected cells at 48 hours pi (Figure 4A, right panel).

Interestingly, JEV-induced IRF8 expression was suppressed in miR-155 overexpressing cells and this could be reversed using the miR-155 inhibitor (Figure 4B, left panel). Importantly, expression of IRF8 was not affected in miR146a overexpressing cells (Figure 4B, right panel). These data demonstrate that miR-155 can suppress JEV-induced IRF8 expression and this might result in the reduced IFN-β levels seen above.

miR-155 attenuates JEV-induced NF-κB pathway gene expression in microglial cells

NF-κB-mediated pro-inflammatory gene expression plays an important role in innate immune response against viral infection. Figure 5A shows that NF-κB activation is required for IFN-β induction in JEV-infected cells as inhibition of NF-κB activity led to reduced IFN-β mRNA expression in human microglial cells. Since JEV-induced IFN-β expression was suppressed in miR-155 overexpressing cells (Figure 3A), we hypothesized that miR-155 might attenuate JEV-induced innate immune gene expression. In order to identify the innate immune genes that are

Figure 3 JEV-induced IFN-β mRNA expression in human microglial cells. CHME3 cells were transfected with CM, M-155, M-146a, or I-155 and mock-infected or infected with JEV 24 hours later. Total RNA was extracted 24 hours pi, and levels of various transcripts were determined by qPCR normalized against those of GAPDH. (A) Relative IFNβ mRNA expression in JEV-infected cells compared to mock-infected cells. (B) Relative abundance of various ISG transcripts in JEV-infected cells compared to mock-infected cells. (C) Relative expression of IFIT1 in M-155 and I-155 transfected cells compared to CM transfected cells following JEV infection. CM, control mimic; GAPDH, Glyceraldehyde 3-phosphate dehydrogenase; I-155, h, hours; inhibitor 155; IFN, interferon; ISG, interferon-stimulated genes; JEV, Japanese encephalitis virus; M-146a, mimic 146a; M-155, mimic 155; pi, post-infection. **, P < 0.005.
modulated by miR-155 during JEV infection, a PCR array was performed covering 84 genes directly or indirectly regulated via the \(\text{NF-}\kappa\text{B}\) pathway. The heat map shown in Figure 5B, depicts dysregulated genes in CHME3 cells during JEV infection. Cells transfected with the control mimic followed by JEV infection had 26 genes up-regulated (over 2-fold). On the other hand, 32 genes were up-regulated in miR-155 overexpressing cells infected with JEV. Interestingly, 19 of these up-regulated genes were common in both sets. However, 16 out of these 19 genes exhibited attenuation of their expression in miR-155 overexpressing JEV-infected cells (Figure 5C). We studied the expression of several of these mRNAs by qPCR (Figure 5D).

\text{IL-12B} and \text{IL-4} are anti-inflammatory genes and the expression of these was up-regulated whereas \text{PTGS2} (\text{COX2}), an inflammatory mediator, was significantly suppressed in miR-155 overexpressing cells. \text{CCR-5}, which has an important role in JEV infection, \cite{27} was up-regulated in these cells. These qPCR data conformed to the PCR array data.

Interestingly, miR-155 and miR-146a are reported to be involved in the reduction of \text{NF-}\kappa\text{B} activity by targeting specific genes \((\text{Ikk}\epsilon, \text{MyD88}, \text{TAB2}, \text{TRAF6}, \text{IRAK1})\) in the TLR-mediated \text{NF-}\kappa\text{B} activating pathways \cite{15,28}. PCR array data showed that JEV-induced MyD88 mRNA expression was suppressed in miR-155 overexpressing cells. The reduced MyD88 protein level was confirmed by western blot. However, \text{Ikk}\epsilon protein levels were not affected (Figure 5E). As observed in the PCR array, \text{IL-6} mRNA expression was markedly reduced in M-155 overexpressing cells following JEV infection. Transcription factor \text{C/EBP}\beta, which activates \text{IL-6} gene expression, is among the potential direct mRNA targets of miR-155. \text{TGF-\beta activated kinase 1/MAP3K7 binding protein 2 (TAB2)} is another important multifunctional molecule that activates \text{NF-}\kappa\text{B}. Our results showed that both \text{C/EBP}\beta and \text{TAB2} mRNA expression were significantly attenuated in miR-155 overexpressing cells following JEV infection (Figure 5F). These results indicate that miR-155 can differentially target multiple genes in the \text{NF-}\kappa\text{B} signaling pathway, resulting in significant modulation of the expression of innate immune genes in JEV-infected human microglial cells.

**miR-155 enhances JEV-induced CD45 expression on microglial cells**

Following an infection or an injury, resting microglial cells could polarize into two major activated subtypes...
categorized as M1 and M2. The M1 subtype overproduces pro-inflammatory cytokines and promotes cell-mediated immunity, whereas M2 microglia tends to dampen inflammation and clear cellular debris [29]. In this context, CD45, a hematopoietic cell-specific protein tyrosine phosphatase, acts as a negative regulator of cytokine receptor signaling leading to altered microglial activation [29-31]. We observed that JEV infection in miR-155 overexpressing human microglial cells upregulated CD45 expression on cell surface (Figure 6A). As a downstream effect of this modulation, we have checked p-STAT1 expression (which is important in pro-inflammatory cytokine production) as well as the expression of ISG. We observed that JEV infection in the control mimic or miR-146a overexpressing cells led to enhanced p-STAT1 expression. On the other hand,
reduced p-STAT1 expression was observed in miR-155 overexpressing cells infected with JEV (Figure 6B). We have also examined the mRNA expression of other cytokine genes by qPCR. Thus, IL-1β, TNF-α, and IL-10 expression was significantly reduced in miR-155 over-expressing JEV-infected cells, but IL-4 levels were not affected (Figure 6C). Together, these data show that overexpression of miR-155 during JEV infection enhances CD45 expression on the microglial cell surface and negatively modulates cytokine expression.

miR-155 suppresses JEV-induced complement factor H expression in microglial cells

Complement factor H (CFH) is a central regulator of the complement system and viruses could up-regulate CFH expression to inhibit excessive complement activation against them. CFH expression can be regulated by miR-155 or miR-146 as both of them have matched seed sequences on the 3’-untranslated region (UTR) of the CFH gene [17,18]. In this study, we examined the expression of CFH and other complement pathway regulators in microglial cells following JEV infection. We found that CFH mRNA expression was significantly enhanced at 48 hours pi (Figure 7A, left panel). Expression of other complement regulators (CD46, CD55, or CD59) showed a marginal increase during the early phase of JEV infection (6 hours pi) but at the later time points it was same as in uninfected control cells (Figure 7A, right panel). Moreover, JEV-induced CFH expression was NF-κB dependent as NF-κB inhibition prior to JEV infection did not enhance CFH expression (Figure 7B). Further, overexpression of miR-155, but not miR-146a, attenuated the CFH expression in microglial cells during JEV infection (Figure 7C).

**Discussion**

Microglia plays key roles in both innate and adaptive immune response in the CNS. In the mammalian immune system miRNAs control differentiation as well as innate and adaptive immune responses. A subset of these miRNAs (designated NeurimmiRs) notably affects both immune and neuronal functions. In this study, we have
focused on two NeurimmirRs, miR-155 and miR-146a, and have studied their role in JEV replication and microglial activation during the infection. These two miRNAs are widely reported to have an immense effect on innate immune response in the context of various external stimuli (such as TLR ligands, virus, bacteria, and other microorganisms). However, very little is known about their role in innate immune modulation during neurotropic flavivirus infection. Here, we have used cultured human microglial cells to understand the effect of JEV infection on these two miRNAs. We have also studied how overexpression of these two miRNAs in microglial cells could affect JEV replication and how these miRNAs could modulate innate immune responses as well as microglial activation. We observed that the expression of miR-155 was enhanced in microglial cells during JEV infection. This may be due to the fact that miR-155 production is transiently under the control of NF-κB which is activated during JEV infection and inflammation [28,32]. Since both these miRNAs are known to have a role in antiviral immunity, we hypothesized that upregulation of these miRNAs might play a role in controlling JEV infection. Interestingly, we found that JEV replication was significantly restricted in miR-155 overexpressing cells.

To gain an insight into the mechanism of how miR-155 overexpression suppressed JEV replication, we analyzed JEV genomic RNA in silico with miRNA target prediction algorithms (PITA and RNA22) but found no potential target sites for miR-155. Hence, miR-155 is unlikely to target JEV RNA directly. We next examined IFN-β in JEV-infected miRNAs overexpressing cells as inducible miR-155 in response to the vesicular stomatitis virus (an RNA virus) infection in macrophage was
shown to induce type I IFN signaling and inhibit viral replication [16]. However, to our surprise we found significant reduction in JEV-induced IFN-β as well as downstream ISG mRNA expression in cells where miR-155 was overexpressed. This suggests that miR-155 uses different mechanisms to exert an antiviral effect against JEV in microglial cells. In fact, Swaminathan et al. [21] have shown that miR-155 exerts an anti-HIV-1 effect by targeting several HIV-1 dependency factors involved in post-entry, pre-integration events, leading to severely diminished HIV-1 infection. Thus, miR-155 can use different mechanisms for its antiviral effect depending upon the virus and the environmental stimuli generated during the particular virus infection of a certain cell type.

In order to understand reduced IFN-β expression in JEV-infected miR-155 overexpressing cells, we checked the expression of IRFs. Among the several IRFs reported, we focused our study on IRF8 as it plays a major role in IFN signaling, response to infection, and maturation of myeloid lineages [33]. IRF8 is also involved in the rapid induction of IFN-β in human monocytes [25]. Moreover, IRF8 may activate a program of gene expression that transforms microglia into a reactive phenotype [34]. In this study, we observed that JEV infection in microglia can induce IRF8 expression, but the same expression was attenuated in miR-155 overexpressing cells and may be a possible reason for reduced IFN-β production in miR-155 overexpressing cells. Interestingly, bioinformatics analysis predicted IRF8 as one of the potential targets for miR-155. We cloned 3’-UTR of IRF8 in the luciferase reporter system for miRNA target validation but found no change in the luciferase readout in miR-155 overexpressing cells (data not shown). We have shown that IRF8 is induced during JEV infection. Since JEV replication and titers are reduced in miR-155 overexpressing cells, it could result in reduced IRF8 induction.

IRF8 is also involved in TLR-mediated NF-κB activation [26]. Since JEV-induced IRF8 expression in miR-155 overexpressing cells was attenuated, we sought to understand its effect on NF-κB pathway. For this we examined 84 genes by RT-PCR array whose expression and function, either directly or indirectly, depend on NF-κB activation. We found several genes, including IFN-β, MyD88, STAT1, PTGS2, and IL-12B, which are induced by JEV and show attenuated expression in miR-155 overexpressing cells. Recently, CCR5 receptor expression in a mouse model of JE was reported to play an important role in recovery as well as promote host survival against JEV [27]. Interestingly, increased CCR5 expression was also observed in our cell culture based study. Thus, modulation of NF-κB mediated signaling pathway genes by JEV-induced miR-155 expression might play a role in reduced JEV replication in microglial cells.

Specific induction of active SHP2 phosphatase dephosphorylates IRF8, which in turn becomes an active repressor and down-regulates TLR-mediated gene expression [35]. In hematopoietic cell lineage CD45 (a Src-homology 2 domain (SH2)-containing protein tyrosine phosphatase) plays an important role in regulating cytokine receptor mediated signal [36]. Analysis of microglia ex vivo revealed that IRF8-deficient microglia had significantly increased levels of CD45 [37]. Thus attenuated IRF8 expression in miR-155 overexpressing cells may result in enhanced CD45 expression on microglial cells.

Following an infection, resting microglial cells can get activated into M1 or M2 phenotypes. The M1 phenotype is a pro-inflammatory state and induces neuropathology, whereas M2 is anti-inflammatory state that could have a neuroprotective role [36]. It is reported that CD45 can negatively regulate CD40L-CD40-induced microglial M1 activation; an effect leading to the promotion of the M2 phenotype. Moreover, this CD45-mediated activation state appears to decrease harmful cytokine production [36]. In this study, we observed an increased CD45 expression in JEV-infected miR-155 overexpressing cells and relatively reduced p-STAT1 expression. Further, mRNA expression for pro-inflammatory cytokines IL-1β and TNF-α was reduced in miR-155 overexpressing cells. CD45 has also been shown to down-regulate NF-κB, an important mediator of pro-inflammatory cytokines [38]. Thus, increased CD45 expression in JEV-infected miR-155 expressing cells may explain reduced expression of NF-κB pathway genes in our study. Taken together, these data show that miR-155 induction can modulate the JEV-induced microglial activation to a state that may be beneficial to the host. Further in vivo study is needed to clarify this issue.

Complement activation is considered to be an important component of innate immune response against invading pathogens. CFH is one of the regulators which negatively regulate the complement activation. Viruses can utilize CFH to evade the innate immune response. The non-structural protein NS1 of the West Nile virus, a flavivirus, inhibits complement activation by binding to CFH [39]. CFH can be induced by NF-κB activation and can be regulated through miRNAs [17]. Interestingly, both miR-155 and miR-146a have the same target sequence in the 3’-UTR of CFH and this has been experimentally proven for both the miRNAs [17]. We have shown that JEV infection can induce CFH expression in human and mouse microglial cells and this induction is attenuated in miR-155 overexpressing cells. Therefore, it is possible that miR-155 expression in JEV-infected microglial cells inhibits CFH expression, which in turn may benefit the host by facilitating complement activation against JEV.

In a study published earlier this month, Thounaojam et al. [40] showed an up-regulation of miR-155 in mouse...
microglial cells (BV-2) and in the mouse and human brain during JEV infection. They suggested that miR-155 had a pro-inflammatory role as its inhibition decreased TBK-1, IRF3/7, and NF-κB phosphorylation both in BV-2 cells as well as in the mouse brain. These results point to a role for miR-155 that is opposite to what we have observed in our study. A major difference between these two studies is that while results reported by Thounaojam et al. [40] are derived from mouse BV-2 cells and the mouse brain where various different kinds of cells may get infected with JEV, our results are derived entirely from in vitro cultured human microglial cells CHME3. Additionally, the GP78 strain of JEV used by Thounaojam et al. [40] is a slow growing virus, both in cultured cells and the mouse brain, with lesions in the virus-cell fusion process [22]. Another reason for these differences may be related to the smaller increase (approximately 4-fold) of miR-155 during JEV infection in BV2 cells [40], compared with the super maximal concentration in CHME3 cells (approximately 100-fold) that occurred during miR-155 overexpression in the present study. Several reports have suggested that, depending on the expression level, miR-155 can modulate cellular functions by targeting genes in different pathways. Ceppi et al. [41] showed that low level miR-155 expression enables the activation of p38 MAPK pathway, favoring IL-1β expression, which induces inflammation in an autocrine manner. However, the same pathway was inhibited when the miR-155 expression level went up significantly, ultimately reflecting an altered immune profile. Similar observations were made by Xiao et al. [42] who reported a 3-fold induction of miR-155 and pro-inflammatory cytokine responses during Helicobacter pylori infection, whereas the overexpression of miR-155 negatively regulated the pro-inflammatory responses.

Intracellular signaling pathways that are concomitantly activated by the same stimulus often interact with one another through a cross regulatory feedback mechanism. The miR-155 is a multifunctional microRNA and it can modulate inflammatory responses in both a positive and negative way [40-48]. Besides its positive role in NF-κB activation and subsequent pro-inflammatory response, accumulating evidence has demonstrated that it can constitute a negative feedback loop in the NF-κB signaling pathway by targeting multiple key proteins, which ultimately leads to repression of, or at least the limitation of NF-κB activation in response to viral or microbial stimuli [41,42,45-48]. Therefore, miR-155 could modulate inflammation depending on various factors including its expression level, the cell type, and the environmental stimuli.

Overall our study suggests that miR-155 modulation can act on multiple levels to control JEV infection of microglial cells and induce innate immune responses that may be beneficial to the host. It can enhance CD45 expression, reduce pro-inflammatory cytokines and CFH expression by targeting several key genes, and suppress JEV replication in microglial cells. These data point to miR-155 playing an important role in modulating JEV-induced microglial activation that may be beneficial in limiting JEV infection in the host. Additional studies, both in vitro and in vivo, are needed to further understand the role of miR-155 during JEV infection in switching microglial activation towards the neuroprotective state.

Abbreviations
ANNOVA: Analysis Of Variance; CCR5: C-C Chemokine Receptor type 5; CEBPB: CCAAT/Enhancer Binding Protein beta; CFH: Complement Factor H; CNS: Central Nervous System; COX-2: Cyclooxygenase-2; DENV 2: Dengue Virus 2; DMEM: Dulbecco’s Modified Eagle’s Medium; DMSO: Di Methyl Sulfoxide; FACS: Fluorescence Activated Cell Sorter; FBS: Fetal Bovine Serum; GAPDH: Glyceraldehyde 3-Phosphate Dehydrogenase; HIV: Human Immunodeficiency Virus; HRP: Horseradish Peroxidase; IRS: Interferon Regulatory Factor; ISG: Interferon Stimulated Genes; JEV: Japanese Encephalitis Virus; LPS: Lipopolysaccharide; MEM: Minimal Essential Medium; MOL: Multiplicity Of Infection; MyD88: Myeloid Differentiation primary response gene (88); NF-κB: Nuclear Factor kappa light chain enhancer of activated B cells; NF-κB: Non fluorescent Quencher-Minor Groove Binder; PCR: Polymerase Chain Reaction; PFU: Plaque Forming Unit; PI3K: Phosphatidylinositol-4,5-bisphosphate 3-kinase; Poly (IC): Polyinosinic-polycytidylic acid; PS: Porcine Stable kidney cell line; p-STAT1: Phosphorylated-Signal Transducers and Activators of Transcription; PTGS-2: Prostaglandin-endoperoxide Synthase 2; qPCR: Quantitative real-time PCR; SDS-PAGE: Sodium Dodecyl Sulfate-Polyacrylamide Gel; STAT1: Signal Transducers and Activators of Transcription; TBK1: TANK Binding Kinase 1; TLR: Toll Like Receptor; TNF-α: Tumor Necrosis Factor-alpha.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
AB was responsible for experimental design, data analysis, and drafting the manuscript. SP, BK, and PJ performed the RNA extraction, PCR array, RT-PCR, and transfection and virus infection experiments. SV contributed to the idea, supervised the experiments, and participated in editing the manuscript. All authors have read and approved the final manuscript.

Acknowledgements
This work was supported by the Department of Biotechnology (DBT), Government of India grants number BT/MB/01/VDR/C08/08 and BT/PR6714/ MED/29/617/2012. We are thankful to Dr Manpreet Kaur for her excellent technical advice. We are thankful to Dr Arinbas Basu (National Brain Research Centre, Manesar, India) for providing the human microglial cells (CHME3).

Received: 24 March 2014 Accepted: 13 May 2014 Published: 29 May 2014

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doi:10.1186/1742-2094-11-97

Cite this article as: Pareek et al.: miR-155 induction in microglial cells suppresses Japanese encephalitis virus replication and negatively modulates innate immune responses. Journal of Neuroinflammation 2014 11:97.