Glioblastoma Immune Landscape and the Potential of New Immunotherapies

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Glioblastoma (GBM) are the most common tumors of the central nervous system and among the deadliest cancers in adults. GBM overall survival has not improved over the last decade despite optimization of therapeutic standard-of-care. While immune checkpoint inhibitors (ICI) have revolutionized cancer care, they unfortunately have little therapeutic success in GBM. Here, we elaborate on normal brain and GBM-associated immune landscapes. We describe the role of microglia and tumor-associated macrophages (TAMs) in immune suppression and highlight the impact of energy metabolism in immune evasion. We also describe the challenges and opportunities of immunotherapies in GBM and discuss new avenues based on harnessing the anti-tumor activity of myeloid cells, vaccines, chimeric antigen receptors (CAR)-T and -NK cells, oncolytic viruses, nanocarriers, and combination therapies.

Keywords: glioblastoma, immune response, macrophage, immunotherapy, CART-T cell

PREFACE

The adult human brain is a tissue of vast complexity, composed of multiple cell types defined by their location, function, or molecular characteristics. Five main classes of cerebral cells have been described: neurons, astrocytes, oligodendrocytes, endothelial cells, and microglia. Interactions among these cell types orchestrate the structure and function of the brain in electrical signaling, axonal ensheathing, regulation of blood flow, metabolic coupling and immune surveillance. For instance, astrocytes which are key effectors of the brain’s energy metabolism, convert glucose into lactate, which is delivered to neurons and retro-converted into pyruvate to fuel the Krebs cycle (1). The neurovascular unit (NVU), which encompasses the blood-brain barrier (BBB), is a functional physiological unit that regulates the blood/cerebral parenchyma interface. It is composed of endothelial cells, smooth muscle cells, pericytes, astrocytes, microglia and neurons. The NVU governs brain homeostasis, controlling cerebral perfusion and protecting from potential pathogens or toxins present in the blood. The NVU is significantly altered in CNS malignancy, especially in glioblastoma (GBM), which are grade IV malignant glioma that are highly vascularized with dense tortuous and leaky blood vessels, permitting massive immune cell infiltration in the tumor core. GBMs are mainly derived from neural stem cells, differentiating into astrocytic or neuronal lineages. This cancer is one of the deadliest types in humans, with an average survival time of <15 months upon diagnosis.
Even with the standard-of-care treatment, consisting of surgical resection when possible, followed by radiation and chemotherapy with the drug Temozolomide (TMZ), the estimated recurrence rate is more than 90%. Recurrence is mostly caused by the regrowth of highly invasive cells that spread out of the tumor core, partially due to its hypoxic and acidic environment (2), and are therefore not removed by surgical resection. The long-standing assumption that GBM tumors were clonal masses with identical molecular characteristics have recently been challenged. Indeed, tumor single cell transcriptomics have identified several GBM cellular states with notable plasticity modulated by the tumor microenvironment (3, 4).

IMMUNE MECHANISMS OF THE HEALTHY CENTRAL NERVOUS SYSTEM (CNS)

Prior to delving into the immune landscape and immunosuppressive mechanisms of GBM, we briefly overview the architecture of the CNS immune system under physiological conditions, highlighting its unique lymphatic drainage system, immune cell populations and leukocyte trafficking (Figure 1A).

Anatomically, the brain parenchyma is surrounded by the meninges, a series of three membranes under the skull, namely the dura mater, the arachnoid membrane and the pia mater (Figure 1B). The brain bathes in cerebrospinal fluid (CSF), generated at the blood-CSF barrier, by epithelial cells of the choroid plexus, through diffusion, pinocytosis and active transport from arterial blood in fenestrated capillaries (Figure 1C). The CSF flows around the brain four ventricles into the subarachnoid space (SAS) in a unidirectional flux through the action of cilia on the choroid plexus and ependymal cells that line the ventricles. It enters the brain parenchyma through aquaporin 4, water channels on the end-feet of astrocytes surrounding the vasculature, and communicates with the interstitial fluid (ISF) through the glymphatic system, a network of perivascular channels formed by astroglia for waste elimination (5). The CSF is reabsorbed by the venous blood in venous sinuses at arachnoid villi. Such turnover occurs three to twelve times daily suggesting that the CSF is an immunologically active fluid. Indeed, the CSF drains trafficking leukocytes to the deep cervical lymph nodes (DCLNs) via the newly discovered meningeal lymphatic vessels in the dura mater (6, 7), or by channeling along cranial nerves through the cribriform plate to the nasal

**FIGURE 1 | Architecture of the CNS immune system.** (A) Schematic illustration of the human brain anatomy namely the brain parenchyma, choroid plexus, ventricles, cerebrospinal fluid (CSF), meninges, dural and nasal lymphatics and the deep cervical lymph nodes (DCLNs). (B) The meninges. These are three membranes that envelop the brain, namely the dura mater, the arachnoid membrane and the pia mater. (C) The neurovascular unit (NVU), blood brain barrier (BBB) and perivascular space. The glia limitans formed by astrocytes end feet ensheath the capillary basement membrane and its pericytes. The perivascular space contains microglia-like perivascular macrophages also dubbed border-associated macrophages (BAMs) and antigen-presenting cells (APC). Microglia are found in the brain parenchyma. (D) The glymphatic system. The CSF enters the brain parenchyma through aquaporin 4, water channels on the end-feet of astrocytes surrounding the vasculature, and communicates with the interstitial fluid (ISF) carrying solutes and small antigens through the glymphatic system, a network of perivascular channels formed by astroglia for waste elimination.
mucosa where it accesses its afferent lymphatics. The ISF, which carries parenchymal solutes and small soluble antigens but not parenchymal immune cells, reaches the DCLNs by channeling along the tight space of the basement membrane lining the walls of cerebral capillaries and arteries. The blood supply of the brain enters through capillaries and post-capillary venules, that push the pia mater in the SAS to form perivascular spaces (Virchow-Robin spaces). The brain vasculature is ensheathed by the BBB (Figure 1D) formed by endothelial cells connected by complex tight junctions and pericytes in the capillary basement membrane, and surrounded by the pia mater, the subpial space and the glia limitans, a thin membrane barrier at the parenchymal basement membrane formed by astrocyte foot processes.

The CNS has long been considered as a site of immune privilege. This was based on earlier findings that transplanted tissue grafts in the brain parenchyma elicit slow adaptive immune responses and are not readily rejected (8), and on the presumed lack of lymphatic vessels. Further, a paucity of innate immune responses to pathogen- or danger-associated molecular patterns (PAMPs and DAMPs) has been reported (9, 10). However, mounting evidence challenge this notion and demonstrate active immunosurveillance in the healthy CNS (11). Together with the discovery of a dural meningeal lymphatic system (6, 7), several studies have shown that unlike the brain parenchyma, the cerebral ventricles elicit immune responses leading to graft rejection (12, 13). Thus, the CNS exhibits compartment-specific immunity regulated by leukocyte entry across endothelial, epithelial and glial cell layers of the blood-brain and blood-CSF barriers. These barriers segregate the parenchyma from the peripheral immune system at steady state while permitting immune communications in the CSF-filled SAS and ventricular space. Such compartmentalization is also reflected by spatially and functionally diverse resident immune cell subsets.

The recent use of high-dimensional single cell approaches [e.g., mass cytometry and single cell RNA sequencing (scRNAseq)] in mice (14) and humans (15), along with intravascular leukocyte tracking and fate mapping systems in reporter mice, has uncovered diverse resident immune cells in the healthy CNS and mapped their localization to different CNS compartments. Microglia, which are derived from a yolk sac progenitor, are found exclusively in the brain parenchyma. A distinct subset of embryonically-derived microglia-like macrophages line the meninges, the choroid plexus and the perivascular spaces, and are dubbed border-associated macrophages (BAMs). Microglia and BAMs make up the bulk of the healthy CNS immune cells accounting for ~80% and ~10% of all CNS steady state leukocytes, respectively. Blood-derived monocytes (Ly6C<sup>hi</sup> and Ly6C<sup>lo</sup>), monocyte-derived cells (MdCs), dendritic cells (DCs) and neutrophils are also present in the healthy CNS, albeit at lower frequencies (<3%) (14). T and B cells, innate lymphocytes (ILCs), natural killer (NK), NKT, eosinophils and mast cells are rare (<1%) but also found at steady state. While microglia and BAMs share several surface markers (CD45<sup>lo</sup> CD11b<sup>hi</sup> F4/80<sup>+</sup> CD64<sup>+</sup> MTeK<sup>+</sup> Cx3CR1<sup>+</sup>), they differ in the expression of SIGLEC-H, which is typically found on microglia but not on BAMs. In contrast, the latter express CD206, CD38 and CD88. Both subsets potentially act as antigen-presenting cells (APCs), as they can upregulate, in a context-dependent fashion, the expression of CD11c, MHCII and co-stimulatory molecules. For instance, microglia of the white matter express higher levels of MHCII, CD68 and HLA-DR compared to gray matter microglia, and upregulate pro-inflammatory cytokines such as SPPI (osteopontin) with age (15). There is little evidence that microglia and BAMs migrate to the periphery to prime T cells. Instead they are thought to maintain tissue homeostasis and to locally re-stimulate T cells.

On the other hand, brain DCs traffic to the DCLNs using one of two routes: a specific route involving the rostral migratory stream (16), olfactory bulb, cribiform plate, and nasal mucosal lymphatics or via the dural lymphatics (Figure 1A). At steady state, DC trafficking contributes to CNS immune tolerance by inducing regulatory T cells (T<sub>reg</sub>). Endothelial cells of the meningeal lymphatic vessels are also presumed to maintain brain antigens-reactive T cells in an anergic state (7). Efferent T cells reach the CNS through the choroid plexus or subarachnoid veins and extravasate into the CSF-filled ventricular space and SAS. In the absence of antigen encounter, T cells are eliminated from the CNS by apoptosis or CSF drainage. Cognate antigen recognition on perivascular or leptomeningeal APCs is required for activated T cells to cross the glia limitans into the parenchyma. T cell activation in the brain is often detrimental leading to neuroinflammation and tissue damage. However, this is not always the case, as T cells can mediate neuroprotective effects in response to CNS injury (17).

**Glioblastoma (GBM) Subtypes and Their Associated Immune Landscapes**

In 2007, the WHO graded CNS tumors based on histological criteria (grade II-IV) (18). In 2010, Verhaak et al. used an unsupervised gene expression analysis of 200 GBM and two normal brain samples to identify four GBM subtypes based on molecular signatures (Table 1). These were referred to as neutral (NE), proneural (PN), classical (CL) and mesenchymal (MES) (19). The NE subtype, in which the normal brain samples clustered, was characterized by the expression of neuronal gene markers, and was later shown by the same team to be non-tumor specific (20). The PN subtype, associated with the best median patient survival, had two genomic features, *PDGFRA* alterations and point mutations in *IDH1*, and was characterized by elevated expression of oligodendrocytic and pro-neural development genes. The CL subtype had high rates of *EGFR* gene amplification co-occurring with aberrations in the RB pathway. It exhibited high expression of neural precursors and stem cell markers, and elevated expression of effectors of the Notch and sonic hedgehog pathways. The MES subtype, linked to the least favorable outcome, had predominant *NF1* gene aberrations and *PTEN* mutations. As its name implies, it included an epithelial-to-mesenchymal signature indicative of de-differentiated/trans-differentiated tumors. It also had the highest inflammatory signature with a notable upregulation of...
TABLE 1 | GBM molecular classification and associated immune phenotypes.

| Classifier | Neural | Proneural | Classical | Mesenchymal |
|------------|--------|-----------|-----------|-------------|
| Genetics<sup>a</sup> | Expression of neuron markers such as NEFL, GABRA1, SYT1 and SLC12A5 | PDGFRα mutations, especially in the Ig-domain | Chromosome 7 amplification paired with chromosome 10 loss | Focal hemizygous deletion of a region at 17q11.2 |
| Association with GO categories linked to the neuron projection and axon and synaptic transmission | Point mutation in IDH1 associated with higher CpG island methylation | High level of EGFR amplification | Low expression of NF1 | Co-mutations of NF1 and PTEN |
| | Focal amplification of the locus at 4q12 harboring PDGFRα | High level of EGFR alterations | Expression of mesenchymal markers (CHI3L1, CD44, MERK1, YKL40 and MET) | High expression of genes implicated in the NFKB and tumor necrosis factor superfamily pathways (TRADD, RELB, TNFRSF11A) |
| | High level of PDGFRα expression | Lack of TP53 mutations | High expression of microglial markers such as CD68 and PTPRC |
| | TP53 mutation | Focal hp21.3 homozygous deletion, targeting CDKN2A | | |
| | Loss of heterozygosity | High expression of neural precursors and stem cell markers | | |
| | Chromosome 7 amplification paired to loss of chromosome 10 only in 50% of the cases | | | |
| | High expression of oligodendrocytic development genes | Expression of proneural development genes | | |

| Immune cell infiltrates<sup>b</sup> | Tumor core | | | |
| | Macrophages (CD163) | Macrophages (CD163) | Macrophages (CD163) + | Macrophages (CD163) + + + |
| | | | | |
| | Microglia (CD68) ++ | Microglia (CD68) | Microglia (CD68) + | Microglia (CD68) + + + |
| | Perivascular area | CD4 T cells ++ | CD4 T cells | CD4 T cells + + + |
| | | CD8 T cells | CD8 T cells | CD8 T cells |
| | | | | |
| | Immune markers<sup>c,d</sup> | PD-1 | IL-12, PD-1 | Galec tin 3, IL-10, IL-23, TGFβ, PD-L1, CD163, CCR2, CCL-22, CD47, CSF-1, Micia-1, IL-6, CTLA-4, Arginase, CD204, IL-1, IL-15, IL-7, CD278, IDO |
| | | | | |
| | Re-classification<sup>e</sup> | <i>; Healthy brain</i> | Combination of OPC-and NPC-like | AC-like | MES-like |
| | | | | | |
| | Associated gene mutation with the re-classification<sup>f</sup> | PDGFRα and CDK4 mutations, respectively | EGFR mutation | NF1 mutation | |

<sup>a</sup>Verhaak RG et al. (19), Cancer Cell 17: 98-110.  
<sup>b</sup>Martinez-Lage M et al. (20), Acta Neuropathol Commun 7: 203.  
<sup>c</sup>Doucette T et al. (21), Cancer Immunol Res 1: 112-122.  
<sup>d</sup>Wang Q et al. (22), Cancer Cell 32: 42-56.  
<sup>e</sup>Neftel C et al. (4), Cell 178:835-849.

Genes in the TNF and NF-κB pathways. Several studies from the Cancer Genome Atlas (TCGA) project subsequently defined a core of recurrent driver genomic alterations in GBM, involving TP53, RB1, NF1, PDGFRA, EGFR, PTEN, and CTNNBD2 (21–24). Genetic alterations in IDHI or IDH2, TERT, and co-deletion of chromosome arms 1p and 19q (1p/19q codel) were rather found in low grade gliomas (LGG; grades II-III) (23, 25). In 2016, the WHO reclassified CNS tumors to integrate molecular information to the diagnosis criteria (26). This classification divided adult gliomas into three groups: (1) oligodendrogliomas, which harbor IDH mutations and 1p/19q codel, (2) astrocytomas, which are IDH mutant but without the 1p/19q codel, and (3) GBM, which are mostly IDH wild-type (WT) (Figure 2). It also introduced histone 3 K27M mutation as a molecular feature of pediatric diffuse midline glioma (27). More recent integration of results from scRNAseq, in vivo single cell lineage tracing and genomic and transcriptomic analyses from TCGA refined the GBM subtypes by identifying four plastic GBM cellular states. These were characterized by six transcriptomic meta-modules and genetic alterations in EGFR, PDGFRα, CDK4, and NF1 (4). Two meta-modules enriched in mesenchymal genes, including hypoxia and glycolysis genes, were referred to as MES1 and MES2, and corresponded to the TCGA-MES subtype in Verhaak et al. (19). An astrocytes-like (AC) module was consistent with the TCGA-CL, and three additional modules referred to as oligodendrocyte progenitor cells-like (OPC) and neural progenitor cells-like (NPC)1 and NPC2, corresponded to the TCGA-PN sub-type (Table 1). Neftel et al. showed, using patient-derived xenografts (PDX) in mice, that tumor cells were able to transit from one cellular state to another, indicative of a plasticity that was modulated by the tumor microenvironment (4). The immune landscape of the GBM subtypes was initially explored by transcriptomics (19, 20, 29). These studies confirmed that the MES subtype exhibited elevated expression of pro-inflammatory mediators together with immunosuppressive factors and immune checkpoints (Table 1). CIBERSORT
FIGURE 2 | Molecular classification of gliomas. Adapted from the 2016 WHO classification of brain tumors by DeWitt JC et al. (26).

Analysis (30) revealed more TAMs, neutrophils and CD4\(^+\) T cells expression signatures in MES, whereas an activated DCs signature was found in CL (20). Analysis of a separate glioma classification system based on IDH1 mutation status and DNA methylation (31) similarly revealed elevated TAMs and neutrophils signatures in one subgroup of IDH1 wild-type (WT) tumors, that was of the MES profile (20). To reassess these findings at the protein level, Martinez-Lage et al. used an automated immunohistochemistry-based analysis of tissue microarray (TMA) from a cohort of 98 patients to define the immune cell counts in each GBM subtype. Microglia and blood-derived TAMs were the most prevalent cells in all four GBM subtypes, but were highest (>80% of all leukocytes) in the MES subtype. Whereas, CD8\(^+\) T cell frequencies were similar in all groups, the MES subtype had slightly more CD4\(^+\) T cells (~1%) (28).

Alternative stratification of GBM based on consensus immunome clusters (CIC) identified two immunologically active GBM clusters (32). These clusters expressed genes associated with cytotoxic T lymphocyte (CTLs) and NK cell activation, such as granzyme B (GZMB) and interferon gamma (IFNG), and genes linked to feedback inhibitory mechanisms including FOXP3, immune checkpoint inhibitors (CTLA-4, PD-1, TIM3, VISTA) and their ligands e.g., PD-L1 and galectin-9 (32). Nevertheless, these CICs did not discriminate patients with respect to survival outcome, potentially due to the low frequencies of CTLs and NKs and the strong immunosuppressive environment mediated by the myeloid compartment. Indeed, GBM tumor-infiltrating lymphocytes (TILs) display an exhausted phenotype (33), and GBM-infiltrating NK cells express reduced levels of activating receptors e.g., NKp30, NKG2D, and DNAX accessory molecule-1 (DNAM-1) (32).
GBM-ASSOCIATED MYELOID CELLS
DIVERSITY, ONTOGENY AND TUMOR GEOGRAPHY

Myeloid cells are key determinants of tumor progression and patient outcome in several cancers (34), and are being actively pursued as targets of new immunotherapies (35, 36). The predominance and diversity of myeloid cells in GBM has warranted extensive analysis of their phenotypes and functions in this cancer. This is critical for discriminate therapy, as general targeting of macrophages with inhibition of colony stimulating factor 1 receptor (CSF1R) failed to enhance overall survival in recurrent GBM (37). The use of lineage tracing systems in glioma mouse models revealed distinct GBM-associated myeloid cell ontogeny, i.e., TAMs derived from microglia (MG-TAMs) or from hematopoietic stem cells in the bone marrow (BM-TAMs) (38). RNAseq analysis of these subsets highlighted the impact of ontogeny-imposed chromatin states and tumor cues on their functions in tumor growth and response to therapy. For instance, differential resistance to the anti-angiogenesis therapy bevacizumab was reported to be mediated by BM-TAMs (39). ATAC-seq and transcription factor (TF) landscape analysis identified TFs linked to microglia identity [e.g., MEF2 (40)] in MG-TAMs, whereas BM-TAMs were enriched in TFs involved in monocyte to macrophage differentiation, i.e. RUNX, CEBP, PU.1, IRF4 and STAT3. Notably, a RUNX-induced gene, integrin subunit alpha 4 (Itgα4, also known as Cd49d) was identified as a distinguishing cell surface marker between the two TAM subsets in both mice and humans. It is expressed on BM-TAMs but epigenetically suppressed in microglia and MG-TAMs. Further analysis, using three different scRNAseq platforms, uncovered 66 core genes that distinguish the two TAM lineages (41). CX3CR1, which is commonly used to isolate microglia in mice, is not specific to microglia, since monocytes upregulate its expression as they differentiate in tissues. Instead, the purinergic receptor P2RY12 has recently emerged as a new microglia marker. MG-TAMs are therefore CD11b+ CX3CR1+ P2RY12− CD49D− whereas BM-TAMs are CD11b+ CX3CR1+ P2RY12− CD49D+ (41) (Figure 3). Both TAM subsets display a “non-canonical” state, expressing both M1 and M2 markers. However, BM-TAMs exhibit higher expression of immunosuppressive cytokines and effectors of oxidative metabolism, characteristic of the M2 phenotype (41). Collectively, while several studies confirm a critical role of BM-TAMs in GBM, MG-TAMs are not mere bystanders. A recent report, exploring the efficacy and targets of the phagocytosis checkpoint inhibitor anti-CD47, demonstrated that MG-TAMs are important effectors of glioma cell phagocytosis contributing to overall survival of glioma-bearing mice (42).

RNAseq analysis of distinct anatomically defined tumor regions (e.g., leading edge, infiltrating region, necrotic zone, blood vessels etc.) and in situ hybridization for BIN1 (an MG-TAM marker) or TGFβ1 (a BM-TAM marker), revealed tumor geographic variation in TAM composition. BM-TAMs were enriched near the blood vessels whereas MG-TAMs were found in infiltrated white matter (41). This was confirmed in a glioma model using the Cx3cr1GFP,Ccr2RFP reporter mouse, which showed that BM-TAMs, which constituted 85% of the total TAM population, localized in the perivascular areas of the tumor core, whereas MG-TAMs accounting for 15% of all TAMs, were restricted to the peritumoral area (43) (Figure 3). Besides differential gene expression profiles, these two TAM subsets have different morphological and migratory characteristics, as shown by 2-photon microscopy. MG-TAMs are stationary, larger in size and more branched than BMDM-TAMs that are highly mobile and smaller (44). Clinically, BMDM-TAM infiltration correlates with poor patient survival (28, 41).

TAM RECRUITMENT AND IMMUNOSUPPRESSIVE MECHANISMS IN GBM

Interleukin (IL)-6, produced by vascular endothelial cells and TAMs, has been implicated in several pro-tumoral processes in GBM: (1) it contributes to the disruption of the BBB by downregulating intercellular tight junction proteins on endothelial cells (45). Concordantly, endothelial cell-specific deletion of IL-6 prevented glioma growth and improved mouse survival (46); (2) it reinforces GBM metabolic dependence on aerobic glycolysis (47), as discussed below; and (3) it promotes the recruitment of macrophages through the induction of CCL5/CXCL5 and favors their alternative activation through PPARγ/HIF-2α signaling (46). The CCL2-CCR2 pathway is equally important for BM-TAM recruitment. Glioma cells instruct this pathway through indoleamine 2,3-dioxygenase (IDO)-dependent production of kynurenine (KYN), a metabolite that triggers CCR2 upregulation through aryl hydrocarbon receptor (AHR). Myeloid-specific deletion of AHR in mice blunted BM-TAMs glioma infiltration. In humans, the KYN-AHR pathway is upregulated in GBM and is associated with an unfavorable outcome (48). A direct correlate has been established between loss of PTEN and BM-TAM recruitment via lysyl oxidase (LOX), a macrophage chemoattractant that signals through the β1 integrin (ITGβ1)-PYK2 pathway. Concordantly, YAP1, LOX and β1 integrin are elevated in GBM, and are associated with reduced overall survival. LOX-elicted TAMs infiltrate the tumor microenvironment and support glioma growth via SPP1 (osteonectin), which inhibits glioma cell apoptosis, promotes angiogenesis and sustains the TAM tolerogenic phenotype by signaling through the Integrin αvβ5 (49, 50).

GBM and other brain tumors are notorious for eliciting local and systemic immunosuppression, mediated in great part by TAMs. TAM-derived TGFβ was initially considered as a key inducer of systemic immune tolerance (51). However, targeting this immunosuppressive cytokine alone did not impact the survival of mice bearing brain tumors (52), implicating additional mechanisms. The expression of PD-L1 on circulating monocytes and BM-TAMs might similarly trigger systemic immunosuppression, through a feed forward mechanism involving IL-10 (53). Beyond soluble immunosuppressive cytokines, direct cell-cell contacts, e.g., through PD-L1 (54), tolerogenic HLA molecules (55) and the apoptosis-inducing...
| Identifier     | Study title                                                                 | Interventions                                                                 | Number expected to be enrolled | Primary completion |
|----------------|------------------------------------------------------------------------------|-------------------------------------------------------------------------------|---------------------------------|-------------------|
|                | **Phase III clinical trials**                                                |                                                                               |                                 |                   |
| NCT04277221    | ADCTA for adjuvant immunotherapy in standard treatment of recurrent glioblastoma multiforme (GBM) | Biological: Autologous dendritic cell/tumor antigen, ADCTA                     | 118                             | December 31, 2022 |
| NCT03548571    | Dendritic cell immunotherapy against cancer stem cells in glioblastoma patients receiving standard therapy | Biological: Dendritic cell immunization Drug: Adjuvant temozolomide            | 60                              | May 1, 2021       |
| NCT02667587    | An investigational immuno-therapy study of temozolomide plus radiation therapy with nivolumab or placebo, for newly diagnosed patients with glioblastoma (GBM, a malignant brain cancer) | Drug: Nivolumab Drug: temozolomide Radiation: Radiotherapy Other: Nivolumab Placebo | 693                             | February 11, 2022 |
| NCT022617589   | An investigational immuno-therapy study of nivolumab compared to temozolomide each given with radiation therapy, for newly-diagnosed patients with glioblastoma (GBM, a malignant brain cancer) | Drug: Nivolumab Drug: Temozolomide Radiation: Radiotherapy                      | 550                             | January 17, 2019  |
|                | **Phase II clinical trials**                                                 |                                                                               |                                 |                   |
| NCT04145115    | A study testing the effect of immunotherapy (ipilimumab and nivolumab) in patients with recurrent glioblastoma with elevated mutational burden | Biological: Ipilimumab Biological: Nivolumab                                   | 37                              | May 31, 2023      |
| NCT02649582    | Adjuvant dendritic cell-immunotherapy plus temozolomide in glioblastoma patients | Biological: Dendritic cell vaccine plus temozolomide chemotherapy               | 20                              | December 2020     |
| NCT03927222    | Immunotherapy targeted against cytomegalovirus in patients with newly-diagnosed WHO grade IV unmethylated glioma | Biological: Human CMV pp65-LAMP mRNA-pulsed autologous DCs containing GM CSF Drug: Temozolomide Biological: Tetanus-Diphtheria Toxoid (Td) (and 2 more...) | 48                             | December 2023     |
| NCT03916757    | V-Boost Immunotherapy in glioblastoma multiforme brain cancer                | Biological: V-Boost                                                           | 20                              | April 15, 2020    |
| NCT03650257    | A large-scale research for immunotherapy of glioblastoma with autologous heat shock protein gp96 | Biological: gp96 Drug: Temozolomide radiation: Radiotherapy                     | 150                             | August 20, 2021   |
| NCT03548571    | Dendritic cell immunotherapy against cancer stem cells in glioblastoma patients receiving standard therapy | Biological: Dendritic cell immunization Drug: Adjuvant temozolomide            | 60                              | May 1, 2021       |
| NCT04013672    | Study of pembrolizumab plus SurVaxM for glioblastoma at first recurrence     | Drug: Pembrolizumab Drug: SurVaxM Drug: Sargramostim Drug: Montanide ISA 51    | 51                              | December 31, 2020 |
| NCT01567202    | Study of DC vaccination against glioblastoma                                 | Procedure: Surgery Drug: Chemotherapy Radiation: Radiotherapy (and 2 more...) | 100                             | December 1, 2019  |
| NCT02799238    | Autologous lymphoid effector cells specific against tumor (ALECSAT) as add on to standard of care in patients with glioblastoma | Biological: ALECSAT Radiation: Radiotherapy Drug: Temozolomide                  | 62                              | June 2020         |
| NCT02799238    | Cediranib maleate and olaparib compared to bevacizumab in treating patients with recurrent glioblastoma | Biological: Bevacizumab Drug: Cediranib Drug: Cediranib maleate Drug: Olaparib | 70                              | May 31, 2020      |
| NCT02337686    | Pembrolizumab in treating patients with recurrent glioblastoma              | Other: Laboratory Biomarker Analysis Biological: Pembrolizumab Other: Pharmacological study Procedure: Therapeutic Conventional Surgery | 20                             | December 31, 2020 |
| NCT01174121    | Immunotherapy using tumor infiltrating lymphocytes for patients with metastatic cancer | Biological: Young TIL Drug: Aldesleukin Drug: Cyclophosphamide (and 2 more...) | 332                             | December 29, 2023 |

(Continued)
| Identifier | Study title | Interventions | Number expected to be enrolled | Primary completion |
|------------|-------------|---------------|-------------------------------|-------------------|
| NCT04225039 | Anti-GITR/Anti-PD1/Stereotactic radiosurgery, in recurrent glioblastoma | Drug: INCMGA00012  
Drug: INCAGN01876  
Drug: SRS  
Procedure: Brain surgery | 32 | February 2025 |
| NCT04049669 | Pediatric trial of indoximod with chemotherapy and radiation for relapsed brain tumors or newly diagnosed DIPG | Drug: Indoximod  
Radiation: Partial Radiation  
Radiation: Full-dose Radiation (and 4 more...) | 140 | October 2, 2024 |
| NCT03491683 | INO-5401 and INO-9012 delivered by electroporation (EP) in COMBINATION WITH cemiplimab (REGN2810) in newly-diagnosed glioblastoma (GBM) | Biological: INO-5401  
Biological: INO-9012  
Biological: Cemiplimab (and 2 more...) | 52 | January 18, 2021 |
| NCT03047473 | Avelumab in patients with newly diagnosed glioblastoma multiforme | Biological: Avelumab | 30 | September 2022 |
| NCT03174197 | Atezolizumab in combination with temozolomide and radiation therapy in treating patients with newly diagnosed glioblastoma | Drug: Atezolizumab  
Radiation: Radiation therapy | 60 | June 30, 2020 |
| NCT03395587 | Efficiency of vaccination with lysate-loaded dendritic cells in patients with newly diagnosed glioblastoma | Biological: Autologous, tumor lysate-loaded, mature dendritic cells (DC)  
Drug: Standard therapy | 136 | September 6, 2022 |
| NCT03158389 | NCT neuro master match–N²M² (NOA-20) | Drug: APG101  
Drug: Alectinib  
Drug: Idasanutlin (and 4 more...) | 350 | September 30, 2023 |
| NCT02968940 | A phase I/IIa study evaluating temferon in patients with glioblastoma & unmethylated MGMT | Drug: Temferon | 21 | December 2022 |
| NCT03899857 | Pembrolizumab for newly diagnosed glioblastoma | Drug: Pembrolizumab | 56 | December 2022 |
| NCT01204684 | Dendritic cell vaccine for patients with brain tumors | Biological: Autologous tumor lysate-pulsed DC vaccination  
Biological: Tumor lysate-pulsed DC vaccination +0.2% resiquimod  
Biological: Tumor-lysat pulsed DC vaccination +adjuvant polyICLC | 60 | January 31, 2021 |
| NCT02236165 | Phase 2 Study of Durvalumab (MEDI4736) in Patients With Glioblastoma | Drug: Durvalumab  
Radiation: Standard radiotherapy  
Biological: Bevacizumab | 159 | November 2018 |
| NCT023102436 | Non-viral TCR gene therapy | Drug: Fludarabine  
Drug: Cyclophosphamide  
Drug: Aldesleukin  
Biological: Sleeping Beauty Transposed PBL | 210 | December 31, 2028 |
| NCT03412877 | Administration of autologous T-cells genetically engineered to express T-cell receptors reactive against mutated neoantigens in people with metastatic cancer | Drug: Cyclophosphamide  
Drug: Fludarabine  
Drug: Aldesleukin (and 2 more...) | 270 | March 23, 2027 |
| NCT02794883 | Tremelimumab and durvalumab in combination or alone in treating patients with recurrent malignant glioma | Biological: Durvalumab  
Other: Laboratory Biomarker Analysis  
Procedure: Surgical Procedure  
Biological: Tremelimumab | 36 | December 2019 |
| Identifier     | Study title                                                                                                                                                                                                 | Interventions                                                                                                                                                                                                 | Number expected to be enrolled | Primary completion |
|----------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------|-------------------|
| NCT02382977    | Study to evaluate safety, tolerability, and optimal dose of candidate GBM vaccine VBI-1901 in recurrent GBM subjects                                                                                           | Biological: VBI-1901                                                                                                                                                                                         | 38                              | October 2020      |
| NCT02382977    | Study to evaluate safety, tolerability, and optimal dose of candidate GBM vaccine VBI-1901 in recurrent GBM subjects                                                                                           | Biological: DNX-2401 Biological: Pembrolizumab                                                                                                     | 49                              | December 2020     |
|                | **Phase I clinical trials**                                                                                                                                                                                |                                                                                                                                                                                                             |                                 |                   |
| NCT02649582    | Adjuvant dendritic cell-immunotherapy plus temozolomide in glioblastoma patients                                                                                                                          | Biological: Dendritic cell vaccine plus temozolomide chemotherapy                                                                               | 20                              | December 2020     |
| NCT04165941    | Novel gamma-delta γδ T cell therapy for treatment of patients with newly diagnosed glioblastoma                                                                                                             | Biological: DRI cell therapy                                                                                                                        | 12                              | January 2022      |
| NCT03961971    | Trial of anti-tim-3 in combination with anti-PD-1 and SRS in recurrent GBM                                                                                                                                   | Drug: MBG453                                                                                                                                                                                                  | 15                              | February 2022     |
| NCT03426891    | Pembrolizumab and vorinostat combined with temozolomide for newly diagnosed glioblastoma                                                                                                                   | Drug: Pembrolizumab Drug: Vorinostat Drug: Temozolomide Radiation: Radiotherapy                                                                     | 32                              | April 2021        |
| NCT02208362    | Genetically modified T-cells in treating patients with recurrent or refractory malignant glioma                                                                                                             | Biological: IL13Rα2-specific, hinge-optimized, 41BB-costimulatory CAR/truncated CD19-expressing Autologous T lymphocytes Other: Laboratory biomarker analysis Other: Quality-of-life assessment (and 5 more...) | 92                              | May 2020          |
| NCT04323046    | Immunotherapy (nivolumab and ipilimumab) before and after surgery for the treatment of recurrent or progressive high grade glioma in children and young adults                                                  | Biological: Ipilimumab Biological: Nivolumab Drug: Placebo Administration (and 2 more...)                                                           | 45                              | March 1, 2022     |
| NCT0407706     | Nivolumab, BMS-986205, and radiation therapy with or without temozolomide in treating patients with newly diagnosed glioblastoma                                                                             | Biological: IDO1 inhibitor BMS-986205 Biological: nivolumab Radiation: Radiation Therapy Drug: Temozolomide                                                 | 30                              | June 9, 2022      |
| NCT04201873    | Pembrolizumab and a vaccine (ATL-DC) for the treatment of surgically accessible recurrent glioblastoma                                                                                                       | Biological: Dendritic cell tumor cell lysate vaccine Biological: Pembrolizumab Other: Placebo Administration Drug: Poly ICCL                                                                            | 40                              | August 1, 2024    |
| NCT04003649    | IL13Ralpha2-targeted chimeric antigen receptor (CAR) T cells with or without nivolumab and ipilimumab in treating patients with recurrent or refractory glioblastoma                                              | Biological: IL13Ralpha2-specific Hinge-optimized 4-1BB-co-stimulatory CAR/Truncated CD19-expressing autologous TN/MEM cells Biological: Ipilimumab Biological: Nivolumab (and 2 more...) | 60                              | January 22, 2022  |
| NCT03714334    | DNX-2440 oncolytic adenovirus for recurrent glioblastoma                                                                                                                                                   | Drug: DNX-2440 injection                                                                                                                                                                                    | 24                              | April 16, 2022    |
| NCT02852655    | A pilot surgical trial to evaluate early immunologic pharmacodynamic parameters for The PD-1 checkpoint inhibitor, pembrolizumab (MK-3475), in patients with surgically accessible recurrent/progressive glioblastoma | Drug: MK-3475                                                                                                                                                                                                | 35                              | March 28, 2018    |
| NCT04207481    | NKG2D-based CAR T-cells immunotherapy for patient with r/r NKG2D+ solid tumors                                                                                                                             | Biological: NKG2D-based CAR T-cells                                                                                                                                                                          | 10                              | December 1, 2022  |
| NCT03491688    | INO-5401 and INO-9012 delivered by electroporation (EP) in combination with cemiplimab (REGN2810) in newly-diagnosed glioblastoma (GBM)                                                                     | Biological: INO-5401 Biological: INO-9012 Biological: Cemiplimab (and 2 more...)                                                                     | 52                              | January 18, 2021  |
| NCT03174197    | Atezolizumab in Combination with temozolomide and radiation therapy in treating patients with newly diagnosed glioblastoma                                                                                 | Drug: Atezolizumab Radiation: Radiation Therapy Drug: Temozolomide                                                                                                                   | 60                              | June 30, 2020     |

(Continued)
| Identifier  | Study title                                                                 | Interventions                                                                 | Number expected to be enrolled | Primary completion   |
|------------|------------------------------------------------------------------------------|--------------------------------------------------------------------------------|-------------------------------|----------------------|
| NCT03389230 | Memory-enriched T cells in treating patients with recurrent or refractory grade III-IV glioma | Biological: CD19CAR-CD28-CD3zeta-EGFRt-expressing Tm-enriched T-lymphocytes Biological: CD19CAR-CD28-CD3zeta-EGFRt-expressing Tn/mem-enriched T-lymphocytes Other: Laboratory Biomarker Analysis Procedure: Leukapheresis | 42                            | June 14, 2021         |
| NCT03344250 | Phase I EGFR BATs in newly diagnosed glioblastoma                           | Drug: EGFR BATs with TMZ following SOC RT/TMZ Drug: Weekly EGFR BATs following SOC RT/TMZ | 18                            | October 1, 2020      |
| NCT03158389 | NCT neuro master match- N² M² (NOA-20)                                       | Drug: APG101 Drug: Alectinib Drug: Idasanutlin (and 4 more...)                    | 350                           | September 30, 2023  |
| NCT03868109 | A phase I/IIa study evaluating temferon in patients with glioblastoma & unmethylated MGMT | Drug: temFeron                                                                 | 21                            | December 2022       |
| NCT0392545  | Combination of immunization and radiotherapy for malignant gliomas (InSituVac1) | Combination product: Combined immune adjuvants and radiation                     | 30                            | April 1, 2020       |
| NCT03341806 | Avelumab with laser interstitial therapy for recurrent glioblastoma          | Drug: Avelumab Combination Product: MRI-guided LITT therapy                      | 30                            | September 2020      |
| NCT02062827 | Genetically engineered HSV-1 phase 1 study for the treatment of recurrent malignant glioma | Biological: M032 (NSC 733972)                                                    | 36                            | September 2020      |
| NCT02223103 | Safety and immunogenicity of personalized genomic vaccine and tumor treating fields (TTFIELDS) to treat glioblastoma | Drug: Poly-ICLC Device: Tumor Treating Fields Biological: Peptides                | 20                            | May 22, 2020        |
| NCT02766699 | A study to evaluate the safety, tolerability and immunogenicity of EGFR(V)-EDV-dox in subjects with recurrent glioblastoma multiforme (GBM) | Drug: EGFR(V)-EDV-Dox                                                            | 20                            | December 2019       |
| NCT02619239 | Dose-escalation study to evaluate the safety and tolerability of GX-17 in patients with glioblastoma | Drug: GX-17                                                                     | 15                            | January 31, 2021    |
| NCT02010606 | Phase I study of a dendritic cell vaccine for patients with either newly diagnosed or recurrent glioblastoma | Biological: Dendritic cell vaccination, in addition to standard temozolomide chemotherapy and involved field radiation therapy Biological: Dendritic cell vaccination, with optional bevacizumab treatment for patients previously treated with bevacizumab | 39                            | April 2020          |
| NCT02502708 | Study of the IDO Pathway inhibitor, indoximod, and temozolomide for pediatric patients with progressive primary malignant brain tumors | Drug: Indoximod Drug: Temozolomide Radiation: Conformal radiation (and 2 more...) | 81                            | December 12, 2019   |
| NCT02382977 | Study to evaluate safety, tolerability, and optimal dose of candidate GBM vaccine VBI-1901 in recurrent GBM subjects | Biological: VBI-1901                                                             | 38                            | October 2020        |
| NCT03043391 | Phase 1b study PVSRIP0 for recurrent malignant glioma in children            | Biological: Polio/Rhinovirus Recombinant (PVSRIP0)                              | 12                            | July 1, 2020        |
| NCT03576612 | GMCI, rivulomab, and radiation therapy in treating patients with newly diagnosed high-grade gliomas | Biological: AdV-tk Drug: Valacyclovir Radiation: Radiation (and 3 more...)       | 36                            | February 28, 2021   |
| NCT03657576 | Trial of C134 in patients with recurrent GBM                                 | Biological: C134                                                                 | 24                            | September 2022      |
| NCT03152318 | A study of the treatment of recurrent malignant glioma with rQNestin34.5v2   | Drug: rQNestin Drug: Cyclophosphamide Procedure: Stereotactic biopsy             | 106                           | July 2021           |
TABLE 2 | Continued

| Identifier       | Study title                                                                 | Interventions                      | Number expected to be enrolled | Primary completion |
|------------------|-----------------------------------------------------------------------------|------------------------------------|-------------------------------|--------------------|
| NCT03911388      | HSV G207 in children with recurrent or refractory cerebellar brain tumors  | Biological: G207                   | 15                            | September 1, 2022  |
| NCT02457845      | HSV G207 alone or with a single radiation dose in children with progressive or recurrent supratentorial brain tumors | Biological: G207                   | 18                            | October 2020       |
| NCT00634231      | A phase I study of AdV-tk + prodrug therapy in combination with radiation therapy for pediatric brain tumors | Biological: AdV-tk Drug: Valacyclovir Radiation: Radiation | 12                            | December 2015      |

receptor Fas (56) contribute to immune escape. A recent study reported a role of tumor-associated glycosylation in local and systemic immunosuppression (57). This was mediated through a direct interaction between O-linked glycans on glioma cells with their receptor, Macrophage Galactose-type Lectin (MGL), on TAMs leading to immunosuppression signaling. Of note, the current GBM standard of care often prescribes dexamethasone to alleviate cerebral edema. This immunosuppressive corticosteroid further contributes to the GBM immunosuppressive environment, interfering with anti-tumor immunity and presenting a challenge for the future of immunotherapies in this cancer.

METABOLIC REMODELING OF THE GBM TUMOR MICROENVIRONMENT

Hypoxia and necrosis are well-known features of GBM. HIF-1α, stabilized by the inhibition of prolyl hydroxylase (PHD) activity in hypoxia, is a transcription factor that modifies the expression of thousands of genes, notably effectors of glycolysis and lactic fermentation. The expression of glucose transporters (GLUT1), glycolytic enzymes (PDK1, Hexokinase or PKM2), and lactate dehydrogenase A (LDHA) help in replenishing NAD⁺ to support the glycolytic process. Monocarboxylate transporter (MCT)4 expression is also increased following stabilization of HIF-1α, leading to passive release of lactate out of the cells (58). Production of H⁺ happens during glycolysis, lactic fermentation, but also during respiration when CO2 is hydrated into HCO₃⁻ and H⁺ ions by carbonic anhydrases (CAs). H⁺ ions eflux from the cytoplasm via H⁺ ATPases and Na⁺/H⁺ exchangers (NHEs) leads to a decrease in the extracellular pH. Tumor acidosis promotes cancer cell invasion through cytoskeletal remodeling, but also by modulating the activity of immune cells in the tumor microenvironment. For instance, LDHA-mediated production of lactic acid was shown to blunt the cytotoxic activity of CTLs and NK cells in melanoma through inhibition of NFAT expression (59). This supports previous findings demonstrating that lactate accumulation in T cells, due to decreased efflux via MCT1 (which controls lactate shuttling in a gradient dependent manner), blunted CTL activity (60). TAMs reinforce GBM metabolic shift to aerobic glycolysis through IL-6 that enhances the activity of phosphoglycerate kinase 1 (PGK1) by promoting its phosphorylation (47). Glioma cells also display a high dependence on amino acid metabolism accompanied by an elevated uptake of branched chain amino acids (BCAA). Through the overexpression of branched chain amino acid transaminase 1 (BCAT1), glioma tumors excrete elevated levels of branched-chain ketoacids (BCKA) through MCT1. Which influx into TAMs and blunt their phagocytic activity (61). GBM TAMs were also shown to drive T cell dysfunction through elevated expression of the ectonucleosidase CD39 that, together with CD73, induces the production of the immunosuppressive metabolite adenosine (48).

THE FUTURE OF IMMUNOTHERAPIES IN GBM

Immune Checkpoint Inhibitors (ICI)
Immune checkpoint inhibitors (ICI) targeting the PD-1 or CTLA-4 pathways have revolutionized cancer therapy in the last decade. However, they have had little clinical benefit in GBM, at the least in the adjuvant setting. The recently published results of the open-label, randomized, phase 3 trial CheckMate-143, which evaluated nivolumab vs. bevacizumab in patients with recurrent GBM were disappointing, as there was no significant difference in median overall survival (mOS) between the two arms (62). The two ongoing phase 3 trials CheckMate-498 and CheckMate-548 evaluating the use of nivolumab in patients with newly-diagnosed GBM, either methylguanine methyltransferase (MGMT)- unmethylated or MGMT-methylated, also failed to meet their primary endpoints, according to an update by Bristol-Myers Squibb. In the neoadjuvant setting, the results are controversial. The anti-PD-1 nivolumab, administered as a neoadjuvant, did not impact patient survival in resectable GBM in a phase 2 clinical trial (63). In contrast, another study reported a survival benefit of the anti-PD-1 pembrolizumab in 35 patients with recurrent and resectable GBM (64). Collectively, the dismal results of ICI in GBM may be due to the poor immunogenicity of GBM tumors. In 2017, the FDA approved the use of the anti-PD-1 pembrolizumab in solid tumors with microsatellite instability high (MSI-H) or mismatch repair deficiency (dMMR) tumors. This year, it further approved the use of pembrolizumab for the treatment of adult and pediatric patients with non-resectable or metastatic tumor mutation burden-high (TMB-H) solid tumors. dMMR gliomas are rare (65), but earlier results from two case
reports showed a response to pembrolizumab in one pediatric (66) and one adult (67) patients. Despite these promising results, a recent study reported that PD-1 blockade did not impact mOS in hypermutated gliomas, consistent with an observed lack of TILs in these cancers (68). However, another study reported significant clinical and radiological responses of nivolumab in two young siblings with biallelic mismatch repair deficiency (66), suggesting that ICI therapy might benefit pediatric GBM with high mutational burden [e.g., with MSH6 mutations (69)]. It is plausible that treatments that increase mutational burden might synergize with ICI, as has been shown in other cancers (70). Nanoscale immunoconjugates (NICs), which deliver ICIs, covalently attached on a natural biopolymer scaffold, across the BBB using transferrin receptor (TIR)-mediated transcytosis, or via angiopep-2 (AP-2)-LDLR-related protein 1 (LRP1), were shown to outperform free ICIs in increasing TILs and improving survival in a murine glioma model (71). However, this remains to be tested in patients. Alternative immunotherapies for GBM are being explored. These are primarily focused on vaccines, chimeric antigen receptors (CAR)-T cells, oncolytic viruses and strategies that harness the anti-tumor activity of myeloid cells or the use of adipose stem/stromal cells (ASC) and stromal vascular fraction (SVF) injected in the surgical cavity [reviewed in Bateman et al. (72)].

Vaccines

In the vaccine arena, three phase 3 clinical trials have been completed with different outcomes. ACT IV, a phase III trial evaluating Rindopepimut (also known as CDX-110), a 13-amino acid peptide vaccine targeting EGFRvIII, a constitutively active mutant form of EGFR expressed in ~30% of GBM patients, in combination with TMZ was terminated for futility, as no significant difference in mOS was observed in patients with newly-diagnosed GBM (73). The failure of this approach might be due to heterogenous expression of EGFRvIII within the tumor or loss of its expression leading to clonal outgrowth of resistant cells. A second phase III trial that evaluated an autologous tumor lysate-pulsed DC vaccine (DCVaxL™) in combination with TMZ showed some clinical benefit, reporting longer progression free survival (PFS) and mOS in patients with recurrent GBM (74). However, this is a logistically complicated approach as it requires personalization, apheresis, and DC expansion prior to administration back into patients. A third phase III trial conducted in Japan using personalized peptide vaccination for HLA-24+ recurrent GBM did not meet the primary nor the secondary endpoints (75). More recently, two phase I/ib trials reported beneficial effects of personalized peptide vaccines. The first, the Glioma Actively Personalized Vaccine Consortium (GAPVAC), employed two sets of personalized peptide vaccines designed according to patients tumor mutations, transcriptomic and immuno-pediotimal profiles, and showed that these vaccines were able to elicit sustained CD8+ T cell and CD4+ Th1 responses against neoantigens (76). The second, which employed a pool of synthetic long peptides mimicking neoantigens, also reported the generation of poly-functional neoantigen-specific CD4+ T cells and CD8+ T cells in the periphery and enhanced infiltration of TILs (77). Together, these trials indicate that vaccine approaches are feasible as they elicit anti-tumor immune responses but whether this will translate into clinical benefit, as a monotherapy, requires additional testing.

**CAR-T and CAR-NK Cells**

CAR-T cells are patients-derived T cells engineered to express a CAR, which consists of the antigen-recognition region of an antibody fused in tandem with the cytoplasmic domains of the T cell receptor chain CD3ζ and costimulatory receptors (e.g., CD28 and/or 4-1BB). Currently, approved CAR-T cells target CD19 in B cell malignancies. The challenges of this therapy include the identification of tumor-specific or tumor-associated antigens, especially important in solid tumors, circumventing antigen loss, and countering the exhaustion of transferred CAR-T cells, among others. Several trials and pre-clinical studies have been conducted using CAR-T cells in GBM. The first was a case report that used an IL13Rα2-CAR-T cells in one patient. The CAR-T was delivered through repeated infusions in the resected tumor cavity followed by infusions in the ventricular system. This regimen led to the regression of all cranial and spinal tumors accompanied by a notable immune activity in the CSF (78). A first-in-human study including 10 patients with recurrent GBM followed. This study evaluated EGFRvIII-CAR-T cells injected intravenously. While the CAR-T cells expanded in the blood and trafficked to the tumor, the antigen was lost in 5 out 7 patients and the tumor microenvironment exhibited elevated expression of inhibitory molecules and a high frequency of Treg cells (79). Improvement of CAR-T therapy requires the identification of a tumor-associated antigen expressed stably throughout tumor growth and with limited heterogeneity. Chondroitin sulfate proteoglycan 4 (CSPG4) was found to fit this criterion. It is highly expressed in 67% of GBM cells and is sustained by TNF derived from microglia. Intracranial delivery of CSPG4-CAR-T cells was effective in vivo in nude mice transplanted with CSPG4-expressing glioma cells or neurospheres (80). Transgenic expression of cytokines, such as IL-15, was also demonstrated as a mean to improve anti-glioma activity of CAR-T cells, as shown with IL13Rα2-CAR-T cells (81). Since the final CAR-T cell product is a mix of CD4+ CAR-T and CD8+ CAR-T cells, another mean to refine this approach is to characterize the T cell subset that mediates anti-tumor activity. Using orthotopic GBM mouse models and IL13Rα2-CAR-T cells, the CD4+ CAR-T cell subset was found to be more effective than the CD8+ CAR-T cells, which were rapidly exhausted (82). Co-expression of the IL-8 receptor, CXCR1/CXCR2, was found to enhance CAR-T cells trafficking and persistence in the tumor in a glioma mouse model (83). Engineering EGFRvIII-CAR-T cells to co-express a bispecific T-cell engager (BiTE) against wild-type EGFR was demonstrated to ameliorate this therapy by countering the heterogeneity of EGFRvIII expression (84). A CAR-engineered NK cell targeting both WT EGFR and EGFRvIII mutant, NK-92-EGFR-CAR, was similarly efficient in targeting and killing GBM cells in mice engrafted with patients’ mesenchymal GBM stem cells (85). Additional CAR target antigens in GBM include B7-H3 (86, 87), HER2 (88-90) and EphA2 (91), as demonstrated in preclinical studies, and in a phase I dose escalation clinical trial using a HER2-CAR (92). Interestingly, generation of a
Daubon et al. Glioblastoma Immune Landscape and Immunotherapies

FIGURE 4 | Immunotherapies for the treatment of GBM. Classical immune checkpoint inhibitors (ICIs) i.e., anti-PD-1/PDL-1 and anti-CTLA4 were ineffective in GBM. Current approaches include modulating TAMs (anti-CD47 to boost phagocytosis, nano-immunoconjugates to modulate TAM phenotype, aptamers to inhibit TAM recruitment), personalized peptide vaccines, chimeric antigen receptor (CAR)-T and CAR-NK cell approaches and oncolytic viruses. BiTEs, Bi-specific T-cell engagers.

tricistronic transgene encoding three CAR molecules against HER2, EphA2 and IL13Rα2, dubbed universal CAR-T (UCAR), was shown to overcome interpatient heterogeneity and target 100% of tumor cells (93). Another approach to overcome problems of tumor heterogeneity and antigen escape, is a new CAR design employing a toxin as the targeting entity was developed and tested in a murine model of glioma. This is based on GBM cells’ affinity to bind chlorotoxin (CLTX) by matrix metalloproteinase-2. (CLTX)-CAR-T cells efficiently limited tumor growth in the absence of off-target effects (94).

Oncolytic Viruses

Oncolytic viruses (OV) constitute an interesting therapeutic approach in GBM, as besides their lytic activity, they might overcome GBM immunosuppression by stimulating innate immunity. Several types of OVs have been tested including replication-competent viruses such as polio and measles viruses, Herpes simplex viruses (HSV), adenoviruses and retroviruses. Notably, recombinant non-pathogenic polio-rhinovirus chimera (PVSR1PO), which binds the poliovirus receptor CD155 on cancer cells, was evaluated in 61 GBM patients via intratumoral injection and was effective in 21% patients who survived past 36 months (95). Replication-deficient adenoviruses, e.g., aglatimagene besadenovec, have also been used as vectors to deliver tumoricidal genes such as the HSV thymidine kinase that converts ganciclovir into a toxic nucleotide analog that poisons infected dividing cells. Two phase II clinical trials evaluated this Adv-tk viro-immunotherapy in GBM and reported improved PFS and OS (96, 97). An oncolytic HSV expressing E-cadherin, a ligand for the inhibitory NK receptor KLRG1, resulted in a better outcome in a glioma mouse model, by inhibiting NK cells and permitting viral spread (98). More recently, a Zika OV was shown to specifically target GBM stem cells (GSCs) rather than neural precursor cells, through a SOX2-Integrin αvβ5 Axis (99), suggesting a potentially superior anti-tumoral activity for brain tumor therapy. A triple combination of anti-CTLA-4, anti-PD-1 and a recombinant oncolytic HSV expressing mouse IL-12 (G47Δ-mIL12) cured most mice in two glioma models. CD4+ T cells, CD8+ T cells and M1 macrophages mediated this response, highlighting the need for combinatorial approaches in future trials (100).

Macrophage-Based Immunotherapies

Additional promising strategies for GBM immunotherapy include harnessing the anti-tumor activity of myeloid and NK cells. Targeting the phagocytosis checkpoint CD47 using a humanized anti-CD47 antibody, HuSF9-G4, has shown promise in a glioma PDX mouse model of five aggressive pediatric brain cancers (101). Furthermore, anti-CD47 in combination with TMZ was shown to enhance phagocytosis and promote cytotoxic CD8+ T cell priming by stimulating antigen cross-presentation through cGAS-STING activation (102). Members of
the Let-7 micro-RNA family have also been used as a therapeutic tool in a mouse glioma model; they showed microglial anti-tumor activity by stimulating TLR7 (103). Alternatively, blocking TAM recruitment or polarization has also shown some efficacy in preclinical models. A 4-1BB-osteopontin (OPN) bi-specific aptamer for instance increased median survival by neutralizing macrophage infiltration while co-stimulating effector T cell activity (50). Di-mannose nanocarriers that bind the mannose receptor CD206 on M2 macrophages, used to deliver in vitro-transcribed mRNA encoding M1-polarizing transcription factors, were shown to reprogram TAMs and improve survival in different cancer models (ovarian, lung metastasis) including GBM (104).

**Perspectives**

There is a significant need to develop novel GBM immunotherapies. To date, more than 70 clinical trials with the terms GBM and immunotherapy are found in the clinicaltrials.gov webpage, of which 7 are phase III, 31 phase II and 37 phase I trials (Table 2). These trials explore the various strategies described above notably personalized vaccines, adoptive cell transfer therapy and combinations. It is our hope that this endeavor will soon impact patients’ lives (Figure 4).

**AUTHOR CONTRIBUTIONS**

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

**FUNDING**

MS was funded by the ARC Foundation, IDEX Bordeaux and the New Aquitaine Region. TD was funded by ARC Foundation, Ligue Contre le Cancer, SIRIC BRIO.

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.