Alternative Splicing Signature in human Crohn’s Disease

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Research Article

Keywords: Alternative Splicing, Crohn's disease, post-transcriptional regulation

DOI: https://doi.org/10.21203/rs.3.rs-113422/v3

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Abstract

Background: Although hundreds of risk loci for Crohn’s disease (CD) have been identified, the underlying pathogenesis of CD remains unclear. Recently, evidence has shown that aberrant gene expression in CD patients’ colon tissues was associated with progression of CD. We reasoned that post-transcriptional regulation, especially alternative splicing (AS), may also play important roles in the pathogenesis of CD.

Results: Herein we re-analyzed a public mRNA-seq data from NCBI GEO dataset (GSE66207) and identified approximately 3,000 unique AS events in CD patients compared to the healthy controls. “Lysine degradation” and “Sphingolipid metabolism” are the top two AS events enriched terms in CD patients. In a validation study, we also sequenced 8 subjects and demonstrated that key genes which were previously linked to CDs, such as IRF1, STAT3, also had significant AS events in CDs.

Conclusion: In conclusion, our study provided a landscape of AS events in CD, especially as the first study focused on a Chinese cohort. Our data suggested that dysregulation of AS may be a new mechanism that contribute to the pathogenesis of CD.

Introduction

Crohn’s disease (CD), as an inflammatory bowel disease (IBD), was recognized to be possibly caused by dysregulated mucosal immune responses to the gut flora, in these genetically susceptible patients [1]. To date, more than 200 risk loci for IBD have been identified, which are also enriched in multigenic regulatory modules [2, 3]. Although genome-wide association studies as well as subsequent meta-analysis could explain the underlying pathophysiology of CD to some extent, the pathogenesis of CD remains not fully understood. Recently, studies showed that dysregulated gene expression in the gut mucosa of CD patients may determine the initiation and progression of CD [4]. A following study also showed there exists a big difference of gene expression in inflamed and noninflamed intestinal mucosa of CD patients when compared with the healthy mucosa of controls [5]. Moreover, a study that focused on integrated analysis of micro-RNAs (miRNA) and mRNA expression demonstrated that miRNAs also play critical roles in regulating CD patients’ gene expressions [6]. Some of these circulating miRNAs were also thought to be potential prognosis markers for CD patients [7]. These data suggested that transcriptional and post-transcriptional regulation may play important roles in determining the etiology of CD.

Protein diversity determines the complexity of eukaryotic cellular processes. Alternative splicing (AS) of the precursor mRNA is one of the essential mechanisms to increase the protein diversity and regulate the intricate protein-RNA network [8, 9]. Nearly 95% of the total human multi-exon genes involve AS events [10]. Evidences indicates that AS plays crucial roles in not only oncogenic processes including proliferation, apoptosis, hypoxia, angiogenesis, immune escape and metastasis [11] [12], but also in basic developmental processes and tissue identity [13]. A pioneering study, which focused on alternative splicing of pre-mRNA in IBDs, has identified 47 splicing factors and 33 intron retention events that were
dysregulated in mucosal tissue of patients [14]. However, due to the lack of next generation sequencing technology, only 149 splicing factors and 145 intron retention events were screened in that pioneer study.

We reasoned that dysregulated AS could contribute to the pathogenesis of CD. Thus, we obtained public mRNA-seq data from NCBI GEO dataset (GSE66207) [6], which includes 33 colon tissue mRNA-seq data (20 are CD and 13 are healthy controls). Using Miso and related AS event analysis software [15], we identified approximately 3,000 significant AS events in CD patients compared to the controls. 117 biological pathways are significantly enriched for these AS events, of which some are highly related to inflammation related responses. IRF1, STAT3, and MAOA were identified as the key relevant genes involved in these AS events. In a validation study, we also identified 64% genes of significant AS events overlapped with the results from the public dataset, indicating a comprehensive role of AS in CD. We believe our results may shed a light on understanding the mechanism of post-transcriptional regulation in CD progression.

**Methods**

**Sample Collection**

A total of four patients with CD from Shengjing Hospital of China Medical University were collected between June 2020 and September 2020, and served as the experimental group. All eligible patients had an established diagnosis of CD according to endoscopic and histologic assessments. Colonic biopsy specimens were taken from the rectum, ulcer margin of sigmoid colon and inflamed portions. Four patients with normal distal colon confirmed by surgical pathology served as the control group. The study was approved by the institutional review board of Shengjing Hospital of China Medical University, and informed consent was obtained from each patient.

**Data preparation**

RNA-seq data was obtained from NCBI GEO dataset (GSE66207) [6]. The sequencing was performed via ILLUMINA (Illumina HiSeq 2500) mean run: 26.98M spots. The mean size for the sequencing file includes 2.58G bases according to the SRX886282 project detail. This dataset included 20 Crohn's disease patients' and 13 normal colon tissues' raw sequence data and mRNA expression data (Patient info were included in the Table 1). Fastq files were aligned on human Hg19 genome by STAR-2.7.1a [16] (Read length = 150, Average read depth = 50). The indexed .bam file were generated by Samtools (1.10) [17] for next step analysis. Raw files in validation the experiment were prepared in same method.

**Identification of significant alternative splicing Events**

Alternative splicing events were identified and quantified using MISO version 0.5.4 [18]for 33 samples. A read length of 48 was applied and other parameters were in default in MISO. The level of alternative splicing events was defined as Percentage Spliced In (Psi). To get differentially expressed alternative splicing events between Crohn's disease patients and normal samples, we applied Wilcoxon test on all
alternative splicing events with at least 3 patients and 1 normal sample. Significant events were defined as ones with p-values < 0.05. Same method was applied in transferability study part.

Pathway Analysis

Pathway analysis of the splicing and data was performed using Enrichr [19] [20] with Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway library. Only the pathways with p-value < 0.05 were considered as related pathways. WebGestalt[21], SUMER [22] and Cytoscape (3.8.0) [23] were applied in pathway analysis of transferability study part. Over-Representation Analysis (ORA) and geneontology biological process were applied as enrichment method and functional database respectively in WebGestalt. The enriched category with the gene sizes less than 5-kb and false discovery rate (FDR) above 0.05 was removed in WebGestalt results. The results of different alternative splicing types were input into SUMER to get pathway network. Finally, Cytoscape (3.8.0) was applied to modify color and text size of the network from SUMER.

Visualization

Sankey diagram was generated by Plotly in python. UpSet plots were generated by ComplexHeatmap [24] in R. Sashimi plots were generated by IGV (version 2.8.3) [25]. In order to avoid random error, we randomly selected and merged RNA-seq alignment data from three Crohn's disease patients and three normal samples respectively. Other graphs were plotted using ggplot in R.

RNA-seq expression analysis

Pair-ended RNA-seq libraries for the transferability study were prepared using the NEBNext Ultra RNA with Poly-A selection kit and was sequenced on an Illumina Hi-Seq 4000 (Genergy, Shanghai). Kallisto software was then used to quantify RNA-seq raw counts based on a pseudoalignment algorithm [26] (Read Length = 200, default; Mean read depth = 50). Differential gene expression was determined with log2foldchange > 1.5 and P < 0.05 genes with > 1 count per million. Any gene with a P-value greater than False Discovery Rate (FDR), after Benjamini−Hochberg correction for multi-testing, was deemed significantly differentially expressed under the test conditions as compared to the controls.

qRT-PCR analysis

Colon tissues from 4 control and 4 CD patients used for quantitative real-time PCR (qPCR) assays. Total RNA was extracted from cells using TRIpure reagents (Bioteke Beijing) and then reverse-transcribed using a first-strand cDNA synthesis kit (TSK302S, RT6 cDNA Synthesis Kit Ver 2). Reverse-transcribed products were used as templates for qPCR using the 2×T5 Fast qPCR Mix (SYBR Green I). The primers used for different target genes were as follows: STAT3 (exon1)-F: GGCGAGGATTGGCTGAAGGG; STAT3 (exon1)-R: CAGGCCGAAGGGCCTCTC; STAT3 (Lengthened 3'UTR)-F: tttctggaagttaaagtagatacagca; STAT3 (Lengthened 3'UTR)-R: ggccactgcattcaaattcc; IRF1(exon1)- F: GGCAGAGCTCGCCACTCCTTAGTC; IRF1(exon1)-R: AGGCAGAGGTTGCCGGGTT; IRF1 (Lengthened 3'UTR)-F: acatgtggctagtgccagtg; IRF1 (Lengthened 3'UTR)-R: cacagatgctgctccaaaaa.
Results

Splicing Types

In the study of identifying alternative splicing events of the data from GSE66207, we found eight types of alternative splicing (AS) events including alternative 3'splice site (A3SS), alternative 5'splice site (A5SS), alternative first exon (AFE), alternative last exon (ALE), mutually exclusive exons (MXE), retrained introns (RI), skipped exon (SE), and Tandem 3' UTR. 2,980 significant alternative splicing events were discovered in 20 Crohn's disease patients (Fig. 1A) (Table S1). Among these types, SE has the most significant alternative splicing events, but the percentage over total number of SE events is rather low. 4.1% of AFE events and 4% of Tandem 3'UTR events were recognized as significant events, indicating a higher level compared to the others (Fig. 1B). The distribution and intersection of gene symbols among 8 events types for 2980 significantly different AS events were shown in Fig. 1C.

Pathway Analysis

The AS events which were related to Crohn's disease contribute to 117 different biological pathways (KEGG pathway) (Fig. 2). Among these significantly enriched pathways, “Lysine degradation”, “Sphingolipid metabolism”, and “Proteoglycans in cancer”, are the most enriched terms, which have the p-value $3 \times 10^{-5}$, $3.6 \times 10^{-4}$ and $3.6 \times 10^{-4}$, respectively (Table S2). Most of the pathways were associated with only one splicing type. However, “Human cytomegalovirus infection” and “Lysine degradation” pathways were affected by four splicing types. We then combined related AS events pathway analysis results with the RNA-seq expression results. We found 8 inflammation related genes were regulated by CD related AS events. Among these 8 genes, Interferon Regulatory Factor 1 (IRF1), Signal Transducer And Activator Of Transcription 3 (STAT3) and Monoamine Oxidase A (MAOA), are identified to have the differentially expressed AS events with the lowest p-Value of Wilcoxon test and most significant differentially expression level, between the Crohn's disease and normal group (Figure. 3A-C). Three genes were all observed with SE splicing events. STAT3 was also observed with AFE splicing events. According to the pathway analysis, IRF1 and STAT3 are involved in prolactin signaling pathways which have a multitude of effects relating to immunoregulation and protection [27]. Moreover, MAOA is related to tryptophan metabolism, which was also highly associated with inflammation, stress response, and microbiome homeostasis [28]. All of three genes in the different sample groups show significantly different RNA expression levels.

PCA analysis

We summarized 2,980 related AS events which occurred across all 33 samples with Percent Spliced In (PSI) values generated by MISO. To characterize the AS events between disease and normal sample, we performed a principal component analysis (PCA) on the MISO results of 2,980 related AS events. PC1-PC3 accounted for 33% of the variance (Fig. 4). The biological differences between CD patients and normal samples were captured by the first, the second and third principal component (PC).

Splicing types in transferability study
To validate AS events that we identified in the GSE66207 datasets, we collected samples and performed RNA-seq on four colon tissues from patients with CD and four controls from Shengjing Hospital of China Medical University. Between June 2020 and September 2020, a total of 4 patients diagnosed on the basis of the standard clinical, endoscopic, and histological criteria of CD at the Gastroenterology Departments of the Shengjing Hospital of China Medical University were included and served as the validation group. The biopsies were obtained during endoscopy. Patients who had infectious colitis, indeterminate colitis, Behcet’s disease, intestinal tuberculosis, and colorectal cancer were excluded. Colon biopsies that were obtained from healthy volunteers during endoscopy served as the control group. The clinical disease activity was assessed by the measurement of the Crohn’s disease activity index for CD by a physician who was blinded to the colonoscopy appearance of the patient. The study was approved by the institutional review board of Shengjing Hospital of China Medical University, and informed consent was obtained from each patient (Table 1).

Table 1
Clinical Characteristics of the Patients (A total of 4 patients with CD were included between June 2020 and September 2020. The clinical characteristics of all patients are shown in Table 1. There were no significant differences with regard to age and sex between the experimental and control groups (P > 0.05).)

| Patient | Age (years) | Gender | Course of disease (years) | Age at diagnosis | Disease phenotype | Disease location | CDAI | Medication |
|---------|-------------|--------|---------------------------|-----------------|------------------|-----------------|------|------------|
| patient1 | 16          | Male   | 1.5                       | 15              | B3               | L3              | 160  | Anti-TNF-a therapy |
| patient2 | 22          | Female | 0.5                       | 22              | B1               | L3              | 170  | 5-ASA and Anti-TNF-a therapy |
| patient3 | 30          | Female | 4                         | 29              | B1p              | L3              | 410  | Anti-TNF-a therapy |
| patient4 | 23          | Male   | 3                         | 21              | B2p              | L2              | 240  | Anti-TNF-a therapy and Corticosteroids |

Locations: L1 = terminal ileum; L2 = colon; L3 = ileocolon. Behaviors: B1 = nonstricturing; B2 = stricturing, B3 = penetrating; P = perianal disease.

ASA, aminosalicylic acid.

Using the same analysis pipeline, we identified 1,715 significant AS events (Table S3) in this validation analysis. Interestingly, SE and AFE are still the most two type events and MXE is the least one, which is
similar to the result from the public dataset (Fig. 5A). But the percentage over the total number of all events types are relatively lower than the previous study (Fig. 5B). In 1,715 significant AS events, AFE and ALE were also identified across multiple genes, which had the most intersections (Fig. 5C).

**Biological Process Pathway analysis in transferability study**

We performed biological process pathway analysis in transferability study based on gene lists of 1,731 significant AS events. Since there are only less than 200 related genes in MXE, RI and Tandem 3’ UTR events types, we didn't identify significant biological processes of these three types of events. In the other five AS type events, we found several biological process networks were highly enriched using the WebGestalt analysis. Among these significantly enriched networks, “defense response”, “cell killing”, “response to cytokine” and their related modules were the most enriched clusters (Fig. 6). These results suggested potential dysregulation of AS may play important roles in host-microbiome interactions in these patients with CD.

**Comparison between the data from public dataset and our transferability study**

Finally, we compared the results from the analysis of the public dataset and the results from the analysis of our own cohort. In terms of the genes that had significant AS events, we identified 64% genes that were identified to include significant AS events in transferability study were overlapped with the results we found in the public dataset (Fig. 7A). Interestingly, IRF1 and STAT3 are still identified to have significant Tandem 3’UTR events and showed the significantly altered expression at mRNA level in the patients (Fig. 7B). In order to validate these findings, we performed qRT-PCR assays on the 8 samples (4 CD vs 4 control). We initially validated the upregulation of STAT3 and IRF1 expression in the CD samples (Fig. 7C), which is consistent with the previous findings [29, 30]. We also did the qRT-PCR assays specially targeting the different usage of 3’UTR regions of these two genes. Interestingly, the expression level of the lengthened 3’UTR are both significantly increased in the CD samples, compared to the control. (Fig. 7D-7E). Even after normalized to the overall mRNA expression (Exon 1), the lengthened 3’UTR expression are still profoundly upregulated in the CD samples (Fig. 7F). These results further validated our findings of significant AS in the CD related crucial genes, indicating the potential roles of AS in regulating these CD-related gene expression, thus contributing to the etiology of the disease. Finally, we collected 404 splicing factors from a large splicing factor database for expression analysis [31]. We excluded 34 factors that were barely expressed in the RNA-seq data (FPKM (Fragments Per Kilobase of transcript per Million mapped reads) < 1). As a result, we identified five splicing factors genes were significantly differentially expressed (p-adj-value < 0.05) in both dataset (Table 2). We also noticed that these splicing factors all had the same trend in terms of the expression level alteration. We speculated these splicing factors may be important in regulating the AS events in CD. In conclusion, these results suggested a strong similarity between our transferability study and the public dataset. We also speculated the difference from these analyses may be due to the different genetic background of these samples, as the first cohort was from the University of North Carolina (assumably a cohort of mixed ethnicity) the validation cohort consists of all Han Chinese. Besides, the average age of the first cohort was 45 years old, the average age of our
cohort was 21 years old, indicating a significant difference on age between the two population. However, the sex ratio between the two cohort was similar (1:1 vs 0.7:1). These factors could also potentially affect the AS results.

### Table 2
Transferability study RNA-seq splicing factor raw count fold change of 8 samples

| Gene symbol | Description                                      | GSE66207-Log2Foldchange CD vs Control | Validation experiment-Log2Foldchange CD vs Control |
|-------------|--------------------------------------------------|----------------------------------------|---------------------------------------------------|
| HSPA1A      | Heat Shock Protein Family A (Hsp70) Member 1A     | 2.4950086                              | 1.73285                                           |
| RNF213      | Ring Finger Protein 213                           | 1.5825733                              | 3.760895                                          |
| MOV10       | Mov10 RISC Complex RNA Helicase                   | 1.5125286                              | 1.304929                                          |
| HSPA5       | Heat Shock Protein Family A (Hsp70) Member 5      | 1.3313641                              | 3.431851                                          |
| LSM10       | LSM10, U7 Small Nuclear RNA Associated             | 1.2935081                              | 1.631783                                          |

### Discussions

Dysregulation of mRNA splicing can cause human disease, and more and more studies showed that targeting alternative splicing could lead to the development of novel therapeutics [32]. To date, accumulated data have already highlighted the crucial roles of AS in many human diseases, including cancer, neurodegenerative disorders, immune and infectious diseases [33–35]. Herein, we presented a comprehensive analysis of AS events in CD using a well-established public dataset and a validation dataset from sequencing a Chinese cohort. In our study, we found SE is the most significantly enriched AS event in CD. This suggested that the CD patients’ transcriptome is much more complex and chaotic compared to the healthy controls, due to a more sophisticated mRNA isoform system. The PCA plot also demonstrated that even if only using different AS events as the principal components, we were still able to differentiate the CD group and control group, suggesting that the mRNA AS event itself, other than the expression level, could be a solid comprehensive signature for CD patients. All these results proved our hypothesis that dysregulated AS events may contribute to the etiology of CD.

Among the AS event related significantly enriched pathways, “Lysine degradation” pathway is the most enriched pathway. EZH1 and EZH2 are two genes involved in this pathway, which have been previously linked to the progression of IBD [36, 37]. They both have dysregulated SE events in CD patients.

“Sphingolipids metabolism” is the most significantly enriched pathways in A3SS events. Interestingly, sphingolipids have recently recognized as mediators of inflammation and potential therapeutic targets for IBDs [38]. Our results suggested that metabolism of sphingolipids in CD patients may differ from the healthy people due to AS, resulting in a dysregulated sphingolipid pool. Moreover, the three most
dysregulated genes in inflammatory response related pathways were all previously linked to IBD. IRF1, as an interferon regulatory factor, has been correlated with CD activity index and CD endoscopic index of severity [39]. In a cohort study, IRF1 has a 72% increase in gene expression among CD patients compared to the controls [40]. STAT3, as a central component in immune response signal transduction, if has gain-of-function mutation, will cause multi-organ early auto-immune diseases [41]. Notably, STAT3 has also been indicated as one of the crucial targets for treating IBDs, although activation of STAT3 is likely to occur in both innate and acquired cell types [29]. MAOA, as a monoamine oxidase, though is not directly associated with IBDs, is highly related with inducing oxidative stress in obese people who are suffering chronic inflammation [42]. Recently, dysregulation of MAOA has also been linked to specific microbe alteration [43]. In conclusion, both top AS related significantly enriched pathways and significantly dysregulated genes are highly related with IBD and CD. Our results provided new insights that post-transcriptional regulation could also contribute to IBD via regulating these key pathways and genes.

Although our study provided a comprehensive AS analysis on colon tissues from CD patients, and also have identified nearly 3,000 unique AS events in CD patients, our studies still have certain limitations. In the first place, although we validated the AS events of STAT3 and IRF1 genes, the experimental validation of discovered AS events in many other genes is still lacking. Moreover, whether these AS events affect protein diversity also requires experimental validation. The validation studies should include examining the protein level of these AS regulated genes, and the detecting the downstream effect on certain signaling transduction pathways. Finally, even if we have detected several splicing factors were dysregulated in CD patients, a RIP-seq or CLIP-seq study should be performed to further detect the responsible splicing factors, RNA binding proteins for CD related AS events. Nonetheless, our study demonstrated a new potential pathophysiology of CDs which is regulated by AS. Our study also presented the first landscape of AS in CD patients. Our data also suggested that drugs that target AS related genes and splicing factors should be considered for further screening for CD patients.

**Conclusion**

Our study presented a landscape of alternative splicing (AS) in CD, and provided the first AS related transcriptional analysis in Chinese cohort. Integrated analysis of two datasets revealed that AS may play a crucial role in determining the pathogenesis of CD.

**Abbreviations**

CD: Crohn's disease; AS: alternative splicing; IBD: inflammatory bowel disease; miRNAs: Micro-RNA; KEGG: Kyoto Encyclopedia of Genes and Genomes; FDR: False Discovery Rate; 3’ UTR: three prime untranslated region.

**Declarations**

- **Ethics approval and consent to participate**
The study was approved by the institutional review board of Shengjing Hospital of China Medical University, and informed consent was obtained from each patient (2020Yc002). All the experiment protocol for involving human data was in accordance with the guidelines of national/international/institutional or Declaration of Helsinki in the manuscript.

Consent for publication

Not applicable.

Availability of data and material

RNA-seq data for 20 CD AS analysis was obtained from NCBI GEO dataset (GSE66207). Data for 4 CD validation sequencing are available from the corresponding author on request. Please contact author for data requests.

Competing interests

The authors declare no competing financial interests.

Funding

This work was supported partly by the Science and Technology Program of Liaoning Province (No. 2019-BS-140).

Authors' contributions

Y. Tan designed research and collected the patients' samples; D. Li, Y. Liang and J. Lu analyzed data; D. Li and Y. Tan wrote the paper.

Acknowledgements

We appreciate the assistance from Intanx Life (Shanghai) Co. Ltd. in data processing and consulting.

References

1. Simmons A: Genes, viruses and microbes. Nature 2010, 466(7307):699–700.
2. Momozawa Y, Dmitrieva J, Théâtre E, Deffontaine V, Rahmouni S, Charloteaux B, Crins F, Docampo E, Elansary M, Gori A-S et al: IBD risk loci are enriched in multigenic regulatory modules encompassing
putative causative genes. *Nature Communications* 2018, 9(1):2427.

3. Hong SN, Park C, Park SJ, Lee CK, Ye BD, Kim YS, Lee S, Chae J, Kim J-I, Kim Y-H: Deep resequencing of 131 Crohn's disease associated genes in pooled DNA confirmed three reported variants and identified eight novel variants. *Gut* 2016, 65(5):788–796.

4. van Beelen Granlund A, Flatberg A, Østvik AE, Drozdov I, Gustafsson BI, Kidd M, Beisvag V, Torp SH, Waldum HL, Martinsen TC: Whole genome gene expression meta-analysis of inflammatory bowel disease colon mucosa demonstrates lack of major differences between Crohn's disease and ulcerative colitis. *PloS one* 2013, 8(2).

5. Hong SN, Joung J-G, Bae JS, Lee CS, Koo JS, Park SJ, Im JP, Kim YS, Kim JW, Park WY et al: RNA-seq Reveals Transcriptomic Differences in Inflamed and Noninflamed Intestinal Mucosa of Crohn's Disease Patients Compared with Normal Mucosa of Healthy Controls. *Inflammatory Bowel Diseases* 2017, 23(7):1098–1108.

6. Peck BCE, Weiser M, Lee SE, Gipson GR, Iyer VB, Sartor RB, Herfarth HH, Long MD, Hansen JJ, Isaacs KL et al: MicroRNAs Classify Different Disease Behavior Phenotypes of Crohn's Disease and May Have Prognostic Utility. *Inflammatory Bowel Diseases* 2015, 21(9):2178–2187.

7. Jensen MD, Andersen RF, Christensen H, Nathan T, Kjeldsen J, Madsen JS: Circulating microRNAs as biomarkers of adult Crohn's disease. *Eur J Gastroenterol Hepatol* 2015, 27(9):1038–1044.

8. Wang Y, Liu J, Huang B, Xu YM, Li J, Huang LF, Lin J, Zhang J, Min QH, Yang WM: Mechanism of alternative splicing and its regulation. *Biomedical reports* 2015, 3(2):152–158.

9. Chen M, Manley JL: Mechanisms of alternative splicing regulation: insights from molecular and genomics approaches. *Nature reviews Molecular cell biology* 2009, 10(11):741–754.

10. Black DL: Mechanisms of alternative pre-messenger RNA splicing. *Annual review of biochemistry* 2003, 72(1):291–336.

11. David CJ, Manley JL: Alternative pre-mRNA splicing regulation in cancer: pathways and programs unhinged. *Genes Dev* 2010, 24(21):2343–2364.

12. Oltean S, Bates DO: Hallmarks of alternative splicing in cancer. *Oncogene* 2014, 33(46):5311–5318.

13. Baralle FE, Giudice J: Alternative splicing as a regulator of development and tissue identity. *Nature Reviews Molecular Cell Biology* 2017, 18(7):437.

14. Hässler R, Kerick M, Mah N, Hultschig C, Richter G, Bretz F, Sina C, Lehrach H, Nietfeld W, Schreiber S et al: Alterations of pre-mRNA splicing in human inflammatory bowel disease. *Eur J Cell Biol* 2011, 90(6–7):603–611.

15. Katz Y, Wang ET, Airoldi EM, Burge CB: Analysis and design of RNA sequencing experiments for identifying isoform regulation. *Nature methods* 2010, 7(12):1009–1015.

16. Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S, Batut P, Chaisson M, Gingeras TR: STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 2013, 29(1):15–21.

17. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R: The sequence alignment/map format and SAMtools. *Bioinformatics* 2009, 25(16):2078–2079.
Katz Y, Wang ET, Airoldi EM, Burge CB: Analysis and design of RNA sequencing experiments for identifying isoform regulation. Nat Methods 2010, 7(12):1009–1015.

Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, Clark NR, Ma’ayan A: Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. BMC bioinformatics 2013, 14(1):128.

Kuleshov MV, Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, Koplev S, Jenkins SL, Jagodnik KM, Lachmann A: Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. Nucleic acids research 2016, 44(W1):W90-W97.

Liao Y, Wang J, Jaehnig EJ, Shi Z, Zhang B: WebGestalt 2019: gene set analysis toolkit with revamped UIs and APIs. Nucleic acids research 2019, 47(W1):W199-W205.

Savage SR, Shi Z, Liao Y, Zhang B: Graph algorithms for condensing and consolidating gene set analysis results. Molecular & Cellular Proteomics 2019, 18(8 suppl 1):S141-S152.

Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T: Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome research 2003, 13(11):2498–2504.

Gu Z, Eils R, Schlesner M: Complex heatmaps reveal patterns and correlations in multidimensional genomic data. Bioinformatics 2016, 32(18):2847–2849.

Thorvaldsdóttir H, Robinson JT, Mesirov JP: Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. Briefings in bioinformatics 2013, 14(2):178–192.

Bray NL, Pimentel H, Melsted P, Pachter L: Near-optimal probabilistic RNA-seq quantification. Nature biotechnology 2016, 34(5):525–527.

Ben-Jonathan N, Hugo ER, Brandebourg TD, LaPensee CR: Focus on prolactin as a metabolic hormone. Trends Endocrinol Metab 2006, 17(3):110–116.

Gostner JM, Geisler S, Stonig M, Mair L, Sperner-Unterweger B, Fuchs D: Tryptophan Metabolism and Related Pathways in Psychoneuroimmunology: The Impact of Nutrition and Lifestyle. Neuropsychobiology 2020, 79(1):89–99.

Sugimoto K: Role of STAT3 in inflammatory bowel disease. World journal of gastroenterology: WJG 2008, 14(33):5110.

Clavell M, Correa-Gracian H, Liu Z, Craver R, Brown R, Schmidt-Sommerfeld E, Udall J, Jr., Delgado A, Mannick E: Detection of interferon regulatory factor-1 in lamina propria mononuclear cells in Crohn's disease. J Pediatr Gastroenterol Nutr 2000, 30(1):43–47.

Seiler M, Peng S, Agrawal AA, Palacino J, Teng T, Zhu P, Smith PG, Buonamici S, Yu L: Somatic Mutational Landscape of Splicing Factor Genes and Their Functional Consequences across 33 Cancer Types. Cell Rep 2018, 23(1):282–296.e284.

Zhao S: Alternative splicing, RNA-seq and drug discovery. Drug Discovery Today 2019, 24(6):1258–1267.

Wang G-S, Cooper TA: Splicing in disease: disruption of the splicing code and the decoding machinery. Nature Reviews Genetics 2007, 8(10):749–761.
34. Scotti MM, Swanson MS: RNA mis-splicing in disease. *Nature Reviews Genetics* 2016, 17(1):19.
35. Kim HK, Pham MHC, Ko KS, Rhee BD, Han J: Alternative splicing isoforms in health and disease. *Pflügers Archiv-European Journal of Physiology* 2018, 470(7):995–1016.
36. Fernandez-Becker NQ, Moss AC: In silico analysis of T-bet activity in peripheral blood mononuclear cells in patients with inflammatory bowel disease (IBD). *In Silico Biol* 2009, 9(5–6):355–363.
37. Lou X, Zhu H, Ning L, Li C, Li S, Du H, Zhou X, Xu G: EZH2 Regulates Intestinal Inflammation and Necroptosis Through the JNK Signaling Pathway in Intestinal Epithelial Cells. *Dig Dis Sci* 2019, 64(12):3518–3527.
38. Sukocheva OA, Lukina E, McGowan E, Bishayee A: Sphingolipids as mediators of inflammation and novel therapeutic target in inflammatory bowel disease. *Adv Protein Chem Struct Biol* 2020, 120:123–158.
39. Tang R, Yang G, Zhang S, Wu C, Chen M: Opposite effects of interferon regulatory factor 1 and osteopontin on the apoptosis of epithelial cells induced by TNF-α in inflammatory bowel disease. *Inflamm Bowel Dis* 2014, 20(11):1950–1961.
40. Huff CD, Witherspoon DJ, Zhang Y, Gatenbee C, Denson LA, Kugathasan S, Hakonarson H, Whiting A, Davis CT, Wu W et al: Crohn's disease and genetic hitchhiking at IBD5. *Mol Biol Evol* 2012, 29(1):101–111.
41. Milner JD, Vogel TP, Forbes L, Ma CA, Stray-Pedersen A, Niemela JE, Lyons JJ, Engelhardt KR, Zhang Y, Topcagic N: Early-onset lymphoproliferation and autoimmunity caused by germline STAT3 gain-of-function mutations. *Blood* 2015, 125(4):591–599.
42. Sturza A, Olariu S, Ionică M, Duicu OM, Vâduva AO, Boia E, Muntean DM, Popoiu CM: Monoamine oxidase is a source of oxidative stress in obese patients with chronic inflammation (1). *Can J Physiol Pharmacol* 2019, 97(9):844–849.
43. Xie Y, Wang C, Zhao D, Wang C, Li C: Dietary Proteins Regulate Serotonin Biosynthesis and Catabolism by Specific Gut Microbes. *Journal of Agricultural and Food Chemistry* 2020.

**Figures**
Figure 1

Alternative splicing identification in CD patients and normal colon samples. A. Significant AS event counts in different AS types between Crohn's disease patients and normal colon samples. B. The percentage of significant AS events over all AS events in different AS types. C. Distribution of AS types and intersection between gene symbols of 2,980 related AS events.
Figure 2
Pathway Analysis. The biological pathways associated with the related AS events in different splice type.
Figure 3

IGV Sashimi Plot of three related genes (IRF1, MAOA and STAT3) expression and potential splicing patterns (circle = junction coverage, line = potential splicing event).
Figure 4

PCA plot. Scatterplot pairs of first three principal components for Percent Spliced In (PSI) values across 33 samples. The red triangles represent CD patients and the blue circle represent normal samples.
Figure 5

Alternative splicing identification in CD patients and normal samples in transferability study. A. Significant AS event counts in different AS types between CD patients and normal samples. B. The percentage of significant AS events over all AS events in different AS types. C. Distribution of AS types and intersection between gene symbols of 1,715 related AS events.
Figure 6

Enriched biological process networks which were identified in our validation study.
Figure 7

Linkage between the results from public dataset and the results from our transferability study. A. Venn diagram showed the 1781 genes that were identified to include significant AS events in transferability study were overlapped with the results from GSE66207. B. IGV Sashimi plot showed IRF1 and STAT3 also had significant tandem 3'UTR events as well as significantly altered expression in patients with CD compared to the controls. C. qRT-PCR analysis showed the exon1 mRNA expression of the STAT3 and IRF1 were both significantly upregulated in the CD group. ***: Student's t-test p-value < 0.001; bar = SEM. D. Scheme shows how the primers are designed to target the lengthened 3'UTR of the STAT3 and IRF1 genes in the CD patients. E. qRT-PCR analysis showed the expression of the lengthened 3'UTR region of STAT3 and IRF1 are significantly upregulated in the CD samples, matching the results of Figure 7B. ***: Student's t-test p-value < 0.001; bar = SEM. F. Normalized results of lengthened 3'UTR / exon 1 expression of STAT3 and IRF1 gene. ***: Student's t-test p-value < 0.001; bar = SEM.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- TableS1.xlsx
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