Alterations in Retinal Oxygen Delivery, Metabolism, and Extraction Fraction During Bilateral Common Carotid Artery Occlusion in Rats

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Ocular ischemic syndrome (OIS) is a condition that is caused by occlusion or severe narrowing of the vasculature that supplies the eye, usually the internal and/or common carotid arteries.1 It is characterized by reduced ocular blood flow, loss of visual acuity, orbital pain, and abnormalities in the anterior and posterior segments of the eye.1 Most patients with OIS are also at risk for, or suffer from, cardiovascular and/or cerebrovascular diseases.2–4 Since OIS may appear in the eye before signs and symptoms from the cerebrovascular system,1 patients with OIS should be investigated for the presence of potential cerebral disease.

Experimental permanent bilateral common carotid artery occlusion (BCCAO) is a procedure that causes ischemia by reducing perfusion pressure to tissues that are supplied by the common carotid arteries. It was first used to study cerebral ischemia in rats5 and was later adopted as an experimental model for OIS, showing changes in retinal vessels, capillary network and arterial filling time.6 With prolonged BCCAO of 3 to 300 days duration, retinal anatomical and functional abnormalities, including progressive retinal degeneration and reduced retinal electrophysiological function have been demonstrated.2–13 Additionally, retinal electrophysiologic,13–15 and biochemical14–17 changes have been reported within 24 hours of BCCAO.

It is well-established that autoregulatory mechanisms maintain or augment total retinal blood flow (TRBF) during reduced perfusion pressure and light flicker stimulation.18,19 Accordingly, the initial compensatory response to BCCAO is expected to be retinal vasodilation, though not previously reported to our knowledge. Once the vasodilatory capacity is expended, TRBF is reduced thereby affecting the rate of inner retinal oxygen delivery (DO2) which is equal to the product of TRBF and retinal arterial oxygen content. To maintain inner retinal oxygen metabolism (MO2) under a reduced DO2 condition, the tissue must extract more of the supplied oxygen, consequently increasing the oxygen extraction fraction (OEF), defined as the ratio of MO2 to DO2.20 However, there is limited knowledge about changes in TRBF, DO2, MO2, and OEF immediately following BCCAO.

Methods for measurements of retinal MO2 in rats have been reported by an oxygen-sensitive microelectrode technique, visible light optical coherence tomography (OCT) and by combining photoacoustic ophthalmoscopy with spectral domain OCT.21–25 We have previously used phosphorescence lifetime and blood flow imaging for measurements of DO2 and MO2 in rats.26–28 Recently, we reported alterations in DO2, MO2, and OEF in a rat model of graded retinal ischemia induced by ligating the contralateral carotid artery and compressing the ipsilateral carotid artery at graded levels.30 However, in this study, sequential measurements under graded retinal ischemia in the same rat were obtained, thus precluding independent data points that are needed for comparative analysis and
mathematical modeling of DO\textsubscript{2}, MO\textsubscript{2}, and OEF. The current study was conducted to test two hypotheses: 1) DO\textsubscript{2} and MO\textsubscript{2} decrease and OEF increases immediately after BCCAO and 2) MO\textsubscript{2} is maintained by elevation of OEF during moderate reductions in DO\textsubscript{2}.

**Material and Methods**

**Animals**

All procedures were approved by the University of Southern California Institutional Animal Care and Use Committee and adhered to the articles of the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. The study was performed in 14 adult (age: 12–18 weeks) male Long-Evans rats (weight: 300–500 g; Charles River, Wilmington, MA, USA). Rats were acclimated for 3 days before being subjected to random grouping of cohorts. They were kept under environmentally controlled conditions with a 12-hour/12-hour light/dark cycle at 20 to 22°C, were fed a standard rat diet, and had free access to food and water. Prior to performing the experimental procedures, the rats were anesthetized with xylazine (5 mg/kg) and ketamine (90 mg/kg) by intraperitoneal injections. Additional xylazine and ketamine were administered during the procedures to maintain anesthesia. The rats were placed on a heated pad for performing the surgical procedure, and the eyes were kept hydrated by administering saline drops every 10 minutes. Phenylephrine 2.5% (Paragon, Portland, OR, USA) and Tropicamide 1% (Bausch and Lomb, Tampa, FL, USA) were instilled into both eyes for pupillary dilation prior to imaging. For BCCAO, the common carotid arteries (CAs) were accessed via an anterior midline prelaryngeal incision and were cleanly dissected from the vagus and sympathetic nerves. In 5 rats, 5-0 silk sutures were used to ligate both CAs (ligation group) and in another five rats, atraumatic vascular clamps (jaw dimension of 6 × 1 mm; Fine Science Tools, Braintree, MA, USA) were applied to the CAs (clamp group). Four rats were subjected to exactly the same surgical procedure without occlusions (sham group). The femoral artery was cannulated for administration of 2-μm polystyrene fluorescent microspheres (Life Technologies, Eugene, OR, USA) at a concentration of 10\textsuperscript{4} particles/mL and Pd-Porphine (Frontier Scientific, Logan, UT, USA) at a dosage of 20 mg/kg. For imaging, the rats were placed on an animal holder equipped with a copper tubing water heater. A glass coverslip with 2.5% hypromellose ophthalmic demulcent solution (HUB Pharmaceuticals, Plymouth, MI, USA) was applied to the cornea to maintain hydration and eliminate its refractive power. Both eyes of each rat were imaged within 30 minutes after BCCAO or sham procedures. Personnel who conducted the experiments were knowledgeable of group allocation during the imaging sessions.

**Blood Flow Imaging**

A previously described imaging system was used to capture red-free and fluorescent image sequences and measure vessel diameter (D) and venous blood velocity (V), respectively.\cite{26} For D measurements, the light illumination of a slit lamp biomicroscope coupled with a green filter (540 ± 5 nm) was used to capture retinal images. Registered mean images were analyzed to determine the vessel boundaries based on the full width at half maximum of intensity profiles perpendicular to the vessel centerline at several consecutive locations along each vessel.\cite{26} Individual retinal vessels were averaged to obtain mean arterial and venous D (D\textsubscript{A}, D\textsubscript{V}) per eye. For V measurements, a 488-nm diode excitation laser and an emission filter (560 ± 60 nm) were used to acquire 520 fluorescence images at 108 Hz. Multiple image sequences were analyzed to determine the displacement of microspheres along each vein segment over time. In each vein, V was calculated by averaging 15 to 30 measurements. Additionally, measurements in individual veins were averaged to calculate a mean venous V (V\textsubscript{V}) per eye. Blood velocity was measured in veins which have a smaller variance during the cardiac cycle compared to arteries. Blood flow was calculated as V × π × D\textsuperscript{2} / 4 for each vein and summed over all the major veins to calculate TRBF.

**Oxygen Tension Imaging**

Retinal vascular oxygen tension (PO\textsubscript{2}) imaging was performed using our previously established optical section phosphorescence lifetime imaging system.\cite{27,28} A vertical laser line (532 nm) was projected on the retina at an angle and an infrared filter with a cutoff wavelength of 650 nm was placed in the imaging path. The phosphorescence emission from the oxygen-sensitive molecular probe, Pd-Porphine, within all the major retinal arteries and veins was imaged. The phosphorescence lifetime was determined using a frequency-domain approach and converted to PO\textsubscript{2} using the Stern-Volmer expression.\cite{29,30} Three PO\textsubscript{2} measurements were averaged for each retinal major vein and artery.

**Oxygen Delivery, Metabolism and Extraction Fraction**

The O\textsubscript{2} content of the retinal blood vessels was determined as the sum of oxygen bound to hemoglobin and dissolved in blood\cite{31}: O\textsubscript{2} content = SO\textsubscript{2} × HgB × C + k × PO\textsubscript{2}, where SO\textsubscript{2} is the oxygen saturation calculated from the rat hemoglobin dissociation curve using measured PO\textsubscript{2} and blood pH values from literature, HgB is the rat hemoglobin concentration value (13.8 g/dL),\cite{32} C is the maximum oxygen-carrying capacity of hemoglobin (1.39 mL O\textsubscript{2}/gHb),\cite{33} and k is the solubility of oxygen in blood (0.0032 mL O\textsubscript{2}/dL mm Hg).\cite{34} Measurements of O\textsubscript{2} content in individual vessels were averaged to obtain mean arterial and venous O\textsubscript{2} contents (O\textsubscript{2A}, O\textsubscript{2V}) per eye. O\textsubscript{2A} was computed as the difference between O\textsubscript{2A} and O\textsubscript{2V}, O\textsubscript{2A}, and O\textsubscript{2V} were calculated as: TRBF × O\textsubscript{2A}, TRBF × O\textsubscript{2A}, and O\textsubscript{2A}/DO\textsubscript{2}, respectively.

**Statistical Analysis**

Statistical analyses were performed using statistical software (SSPS Statistics, version 24; IBM Armonk, NY, USA). Linear mixed model analysis was performed on oxygen metrics (O\textsubscript{2A}, O\textsubscript{2V}, TRBF, DO\textsubscript{2}, O\textsubscript{2}, OEF) with group (sham, clamp, ligation) and eye (right, left) as fixed factors and animal as a random factor. Estimates (β) for the effect of group were determined. The relationships of O\textsubscript{2} and OEF with DO\textsubscript{2} were estimated using the nonlinear least squares curve-fitting tool in MATLAB. Significance was accepted at P ≤ 0.05.

**Results**

A schematic diagram of the methodology for measurements of retinal vascular PO\textsubscript{2} is shown in Figure 1. Retinal arterial and venous PO\textsubscript{2} in the rat from the sham group was higher than the rat from the ligation group (Figs. 1A, 1C). Retinal vessel boundaries, as automatically detected, are shown outlined on red-free retinal images from rats in the sham and ligation groups (Figs. 1B, 1D). The position of one intravenous microsphere at two time points is displayed in a yellow box overlay on the red-free retinal images. Reduced blood velocity
can be visualized in the rat from the BCCAO group (Fig. 1D) compared to the rat from the sham group (Fig. 1B) by the shorter distance the microsphere traveled during the same time interval.

The Table lists mean and standard deviation of retinal vascular O$_2$ contents, Vv, and TRBF in the sham, clamp and ligation groups. Compared to the sham group, O$_2$A was lower in both clamp (β = -2.9 mL O$_2$/dL) and ligation (β = -3.8 mL O$_2$/dL) groups (P < 0.01). Similarly, O$_2$V was lower in both clamp (β = -2.7 mL O$_2$/dL) and ligation (β = -5.3 mL O$_2$/dL) groups (P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04). O$_2$AV was also lower in the ligation group compared to the clamp group (β = -2.6 mL O$_2$/dL; P < 0.04).

There were no statistically significant differences in Dv and DA (P > 0.2). However, Vv was lower in the ligation group compared to the sham (β = -4.2 mm/minute) and clamp (β = -4.9 mm/minute) groups (P ≤ 0.006). Vv was also lower in the clamp group compared to the sham group (β = -4.9 mm/minute; P = 0.003). Likewise, TRBF was lower in the ligation group compared to the sham group (β = -6.2 L/minute) and clamp (β = -3.8 L/minute) groups (P ≤ 0.02). Occlusion of carotid arteries had a general effect of reducing retinal O$_2$A, O$_2$V, Vv, and TRBF, while Dv and DA were not affected significantly. Additionally, O$_2$AV was not significantly changed with BCCAO due to comparable reductions in O$_2$A and O$_2$V.

Figure 2 displays mean and standard deviation of DO$_2$ and MO$_2$ in the sham, clamp and ligation groups. DO$_2$ was 948 ± 255 nLO$_2$/minute, 532 ± 356 nLO$_2$/minute, and 175 ± 137 nLO$_2$/minute in the sham, clamp, and ligation groups, respectively. DO$_2$ in the ligation group was lower compared to that in the sham (β = -772 nLO$_2$/minute) and clamp (β =
DO₂ in the clamp group was lower compared to that in the sham group (β = -415 nL O₂/minute; P = 0.03). MO₂ was 466 ± 103 nL O₂/minute, 306 ± 135 nL O₂/minute, and 171 ± 135 nL O₂/minute in the sham, clamp, and ligation groups, respectively. MO₂ in the ligation group was lower compared to that in the sham (β = -295 nL O₂/minute) and clamp (β = -135 nL O₂/minute) groups (P ≤ 0.05). MO₂ in the clamp group was lower compared to that in the sham group (β = -160 nL O₂/minute; P = 0.03). Immediately after ligation of carotid arteries, DO₂ and MO₂ were decreased to 18% and 37% of the mean values in the sham group, respectively.

Figure 3 shows the mean and standard deviation of OEF in the sham, clamp, and ligation groups. OEF was 0.51 ± 0.11, 0.71 ± 0.29, and 0.98 ± 0.02 in the sham, clamp, and ligation groups, respectively. OEF was higher in the ligation group compared to that in the sham (β = 0.47) and clamp (β = 0.20) groups (P ≤ 0.05). However, OEF in the clamp group was not significantly different than that in the sham group (P = 0.1). The increase in OEF during carotid artery ligation indicates that up to 98% of the delivered oxygen was metabolized by the retinal tissue.

Figure 4 depicts the relationship between MO₂ and DO₂ based on compiled data from all 3 groups. The data were described by an exponential function: MO₂ = 520*(1 - e^{-0.002*DO₂}; R² = 0.81). According to the mathematical model, MO₂ reached an asymptotic maximum value of 520 nL O₂/minute at high levels of DO₂. As DO₂ decreased, MO₂ initially decreased minimally, but eventually decreased more severely at low levels of DO₂. Using the equation derived from the mathematical model, at 50% of the mean MO₂ of the sham group (233 nL O₂/minute), DO₂ was calculated to be 297 nL O₂/minute. Based on this relationship, it may be possible to estimate MO₂ based on measurements of DO₂ and derive threshold values for maintaining MO₂.

Based on compiled data from all three groups, the relationship between OEF and DO₂ is depicted in Figure 5. The data were described by an exponential function: OEF = 0.998*(1 - e^{-649/DO₂}); (R² = 0.77). According to the mathematical model, OEF initially increased as DO₂ decreased, but eventually reached an asymptotic maximum value of 1 at low levels of DO₂. Using the DO₂ value of 297 nL O₂/minute, the value at which MO₂ was reduced to 50% of the mean in the sham group, OEF was calculated to be 0.89, reflecting a 75% increase with respect to the mean OEF value of 0.51 in the sham group. From this relationship, OEF threshold values for maintaining MO₂ may be estimated.

**DISCUSSION**

Reduced oxygen supply to the retina due to carotid artery occlusion in patients with OIS often leads to vision loss. Therefore, it is important to assess alterations in DO₂, MO₂ and OEF under the experimental BCCAO condition to gain knowledge that may be relevant to carotid occlusive disease and other retinopathies also characterized by reduced oxygen supply. The current study confirmed the first hypothesis by demonstrating reductions in DO₂ and MO₂ and an increase in OEF under BCCAO. Furthermore, we were able to confirm the second hypothesis that the retina can accommodate moderate reductions in DO₂ and relatively maintain MO₂, though there is a decrease in MO₂ mediated by severe reductions in DO₂.

During BCCAO, TRBF was considerably reduced to 27% of normal blood flow, presumably from the vertebral arteries via
the circle of Willis. This observation is consistent with other studies that showed presence of some retinal blood flow during BCCAO. Moreover, other studies have shown collateral circulation in the brain and severe bilateral hemispheric ischemia only by occluding the vertebral arteries in addition to the CAs.

The reason for the observed reduction in retinal $O_{2a}$ during BCCAO is not entirely clear. However, other studies have shown under conditions of reduced retinal blood flow with BCCAO or reperfusion following occlusion of ophthalmic vessels, retinal $O_{2a}$ was similarly reduced. The main vascular compensatory response to BCCAO has been shown to be vasodilation and increased blood flow in the vertebral and basilar arteries, but this does not normalize blood flow to the circle of Willis and brain. Along this relatively long path of slow-moving column of blood from the circle of Willis to the eye, there was likely more time for oxygen to be diffused and consumed by the arterial walls, as well as by the optic nerve and peripapillary retinal tissues central to the measurement site. This may account for the measured decrease in $O_{2a}$.

The finding of decreased $MO_2$ mediated by a severe reduction in $DO_2$ within 30 minutes after BCCAO is consistent with previous studies that showed the electroretinogram (ERG) b-wave amplitude was reduced within 20 and 45 minutes of BCCAO. In the current study, $MO_2$ was reduced by 65%, comparable to a previously reported reduction of 46% in the ERG b-wave amplitude. Since generation of the ERG b-wave requires energy, its reduction is expected in the presence of reduced $DO_2$. Additionally, Barnett et al. did not report any retinal histologic damage after 45 minutes of BCCAO, which suggests that $DO_2$ and $MO_2$ alterations after 30 minutes of BCCAO do not cause immediately observable retinal structural damage. Future studies can be directed at whether immediate or sustained alterations in $DO_2$ and $MO_2$ can predict retinal morphological and functional abnormalities observed with prolonged BCCAO.

Initially, the principal compensatory mechanism of the retina to respond to reduced perfusion pressure is to undergo vasodilation, ensuring TRBF and $DO_2$ are maintained and consequently $MO_2$ remains unchanged. In the current study, retinal vasodilation was not observed at the time of the measurements, suggesting that the vasodilatory response for maintaining TRBF was either already expended or not yet activated. Since based on Poiseuille formula, both blood flow and velocity depend on vessel diameter, the finding of reductions in both TRBF and $V_v$ suggests inadequate vasodilation of retinal arteries and of downstream microvasculature for reducing vascular resistance immediately following BCCAO. Similar to our finding, Riva and coworkers found as perfusion pressure was decreased by increasing intraocular pressure in humans that retinal venous diameters near the optic nerve remained unchanged while blood velocity and flow decreased. They also found a simultaneous decrease in vascular resistance confirming the presence of vasodilation in the microvasculature downstream from their measurement site, though not adequate to maintain blood flow and velocity. It is probable that a similar phenomenon was active under BCCAO. Moreover, several studies have reported reductions in blood flow when autoregulatory mechanisms are expended under severe decreased perfusion pressure conditions in humans, monkeys and cats. Nevertheless, with less severe reductions in perfusion pressure, regulation by both TRBF and $V_v$ is expected to minimize changes in $MO_2$.

In the current study, the use of clamp incompletely occluded the CAs in some cases, thus providing data under varying levels of reduced TRBF. This allowed for mathematical modeling of the relationships of $MO_2$ and OEF with $DO_2$. Under moderate reductions in $DO_2$, a minimum change in $MO_2$ was demonstrated. The relative maintenance of $MO_2$ was mediated by an increase in OEF as long as adequate oxygen was available to be extracted. However, with further reductions in $DO_2$, the venous $O_2$ content became negligible and thereby OEF reached a maximum value of unity, indicating that all of the delivered oxygen was metabolized. Under severe $DO_2$ reduction, OEF was maximized and $MO_2$ became limited by $DO_2$, in agreement with our previous report.

One advantage of the current study was quantitative measurements of TRBF and $DO_2$. Thus, findings on the effects of ischemia on $MO_2$ and OEF are not limited to the BCCAO model of retinal ischemia and may be applicable to other methods of reducing retinal blood flow to similar levels. Of particular interest is the information we obtained on $MO_2$. 

![Graph showing the relationship between retinal OEF and oxygen delivery (DO2) based on compiled data obtained within 30 minutes of sham procedure or bilateral common carotid artery occlusion in rats. The data were described by an exponential function: OEF = 0.998* (1 - e^{-0.185*x2}), r² = 0.77; n = 28. Initially, OEF increased with decreasing DO2, but with more severe reductions in DO2, it increased more steeply and approached the theoretical maximum value of one, indicating oxygen was completely extracted from the retinal vasculature.](image-url)
Since energy is required for most retinal functions and MO₂ is an indicator of retinal energy production, it should closely reflect the injurious impact of retinal ischemia.

Recently methods have become available for measurements of MO₂, DO₂, and OEF in human subjects. Thus, evaluation of these parameters may potentially lead to improvements in diagnosis and therapy of human retinal ischemic diseases. For example, thresholds for MO₂, DO₂, and OEF may be derived and applied to predict prognosis for disease progression or response to treatment for individual patients. Moreover, MO₂ may prove to be a valuable quantitative outcome in therapeutic trials that evaluate delivery of oxygen or neuroprotective drugs. Finally, now that it is possible to measure MO₂, research can focus on its role in retinal ischemia and other diseases, thereby leading to future advances in new metabolically-based treatments.

The current study had some limitations. First, the study was limited to retinal physiologic measurements and did not investigate corresponding biochemical abnormalities, electrophysiologic dysfunction, or histopathologic changes under BCCAO, which have been previously described in literature. Nevertheless, the finding of reduced MO₂ was consistent with previously reduced electrophysiologic responses immediately after BCCAO. Second, the systemic physiologic condition of the rats was not controlled or monitored during imaging, though we expect conditions were similar in sham and BCCAO groups. Third, the findings of the current study may not be generalizable beyond the animal species/strain under investigation. Fourth, the vascular clamp produced variable levels of CA occlusions apparently due to the displacement of the clamp during positioning of the animal for imaging. This, in turn, provided a limited number of data points under each graded level of decreased DO₂. Additional data are needed for more robust mathematical modeling of the relationships. Finally, future studies are needed to determine the effect of prolonged TRBF reduction or TRBF recovery on MO₂ and OEF.

In conclusion, this study demonstrated decreases in MO₂ and DO₂ and an increase in OEF immediately after permanent BCCAO. Furthermore, OEF was maximized as MO₂ and DO₂ were matched during severe ischemia. Additionally, nonlinear mathematical models were proposed for predicting MO₂ and OEF based on DO₂. The findings improve knowledge of mechanisms that can maintain MO₂ and may clarify the pathophysiology of retinal ischemic injury.

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