Identification of partial trisomy 13q in two unrelated patients using single-nucleotide polymorphism array and literature overview

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Abstract

Background: Partial trisomy 13q is a less common chromosomal abnormality with a great clinical variability, among them, isolated partial trisomy 13q is extremely rare. Here, we report two new unrelated cases of partial trisomy 13q in Chinese families aiming to emphasize the genotype–phenotype correlation in partial trisomy 13q.

Methods: Enrolled in this study were two unrelated cases of partial 13q trisomy from two families in Quanzhou region South China. Karyotype and single-nucleotide polymorphism (SNP) array analysis were employed to identify chromosome abnormalities and copy number variants in the families.

Results: A 72.9-Mb duplication in 13q14.11q34 region was identified using SNP array analysis in Patient 1 with an intellectual disability, developmental delay, seizures, gastric perforation, and other congenital malformations from a family with paternal inv(13)(p12q14.1). SNP array detection in Patient 2 revealed a 92.4-Mb duplication in 13q12.11q34 region combined with an 8.4-Mb deletion in Xq27.3q28 region with intellectual disability, developmental delay, cleft palate, and duplication of the cervix and the vagina. No chromosomal abnormality was elicited from the parents of Patient 2.

Conclusions: In this study, we presented two new unrelated cases of partial trisomy 13q with variable features in Chinese population, which may enrich the spectrum of the phenotypes partial trisomy 13q and further confirm the genotype–phenotype correlation.

Keywords: Partial trisomy 13q, Karyotype analysis, SNP array, Seizures, Developmental delay

Introduction

Trisomy 13, a common chromosomal disorder characterized by severe intellectual disability and life-threatening physical abnormalities, can be detected effectively during the prenatal period using non-invasive prenatal testing [1]. Partial trisomy 13q is a rare chromosomal abnormality with variable clinical phenotypes and distinctive clinical features similar with those in trisomy 13. Partial trisomy 13q most commonly results from parental balanced translocations or inversions, and rarely from de novo [2–5]. A distinct phenotype of partial proximal and partial distal trisomy of chromosome 13q has...
been defined [6]. Proximal trisomy 13q usually shows strabismus, low and flat nasal bridge, persistent foetal hemoglobin (Hb), and the increased number of nuclear projects on neutrophils and other congenital malformations. On the other hand, the distal trisomy 13q often presents features of polydactyl, hemangioma, bushy eyebrows, long eyelashes, long philtrum, thin upper lip, high arched palate, ocular aberrations, and other facial malformations [7, 8].

Deletion of Xq27.3q28 region is a rare chromosomal disorder typically characterized as developmental delay, intellectual disability, and characteristic facial features [9]. Male patients show more severe clinical features than females, which resemble as Hunter syndrome and Fragile X syndrome [10, 11]. However, female patients harboring Xq27.3q28 deletion demonstrate variable clinical manifestations [9, 12, 13].

In this study, we present two new unrelated cases of partial trisomy 13q with variable features in two Chinese families. The first case involved a patient who harbored an isolated duplication in 13q14.11-qter region. The other case was a girl who harbored a large duplication of 13q12.11-qter combined with Xq27.3q28 deletion.

Material and methods
Subjects
Two new unrelated cases of partial trisomy 13q with variable features in two Chinese families were enrolled in this study from Fujian province, South China. Both of their parents denied consanguineous marriage and any family history of inherited disease. Both families received pretest consultation and signed the written information, subsequently, karyotype and chromosomal microarray analysis were performed in the patients. Ethics Committee approval was obtained from the Institutional Ethics Committee of Quanzhou Women’s and Children’s Hospital to the commencement of the study (2020No.31).

Karyotype analysis
Approximately 2~3 ml parental peripheral blood was collected from the patients and their parents for karyotype analysis. Around 20 ml amniotic fluid was obtained by amniocentesis for fetal chromosome karyotype analysis. The cultured amniotic fluid cells and peripheral blood lymphocytes were harvested using a SinochromeChromprepII automatic chromosome harvesting system according to the standard protocol (Shanghai Lechen Biotechnology Co., Ltd.) we described before [14]. Twenty metaphases were analyzed for peripheral blood karyotype and 30 metaphases were analyzed for fetal karyotype. Nomenclature of chromosomal karyotype was conducted according to ISCN 2020 [15].

DNA extraction
Approximately 3~5 ml peripheral bloods were collected from the patients and their parents for chromosomal microarray analysis. Approximately 10 ml amniotic fluid was obtained by amniocentesis for fetal chromosome microarray analysis. Genomics DNA were extracted from peripheral blood using QIAamp DNA Blood Kit (QIAGEN, Germany) referred to the manufacturer’s protocol (www.qiagen.com).

SNP array detection
The single-nucleotide polymorphism array analysis was carried out using Affymetrix Cytoscan 750 K chip (Life Technologies, American) referred to the protocol described before [16]. The single-nucleotide polymorphism and copy number variants (CNVs) were analyzed using the Genotyping Console and Chromosome Analysis Suite software. The CNVs pathogenicity interpretation was conducted according to the American College of Medical Genetics (ACMG) standards and guidelines [17]. The Database of Genomic Variants (DGV) (http://dgv.tcag.ca/dgv), Online Mendelian Inheritance in Man (OMIM) (https://omim.org/), DECIPHER (https://deciphergenomics.org/) and PubMed (https://www.ncbi.nlm.nih.gov/pubmed/), as well as other databases, were used as reference resources.

Results
Patient 1
The proband in Family 1 was a girl who was the first child in this family. The patient was born full-term delivery with 50 cm (+0.18SD) in height and 2.45 kg (−2.3SD) in weight. The baby was found to have feeding difficulties and jaundice after birth. Her developmental milestone was dramatically delayed. She was unable to walk and speak until 4 years of age. A diagnosis of motor and language delay and intellectual disability was made. Gastric perforation occurred at the age of 5, for which surgical treatment was conducted. Then, seizures were observed at the age of 6, which could be controlled with antiepileptic drugs. Ultrasonography showed that the heart and kidneys were normal, but no brain MR image was available in this patient. At present, she is 8 years old, 118 cm (−2.0SD) tall and weighs 28 kg (+0.6SD), with no obvious facial dysmorphic features, except that her left ear is slightly smaller than the right one. She is now receiving compulsory education in the first grade of a primary school, which is one year behind the average children. She can read and write a little, with learning disability in arithmetic logic. In the 2nd pregnancy of Family 1, miscarriage occurred at the gestational age of 12+ weeks. At
present, amniocentesis is carried out for prenatal diagnosis in the 3rd pregnancy of this family.

**Patient 2**
The proband 2 was also a girl born as the first child of Family 2. She was delivered at the gestational age of 37 weeks, with 50 cm (+0.18SD) in height and 3.0 kg (−0.6SD) in weight. The newborn had cleft palate and duplication of the cervix and vagina. Her developmental milestone was obviously delayed. At one year of age, she was able to sit independently, but presented vertical neck instability and could not turn over at the age of 1 year. In addition, no deciduous teeth were observed. The child’s psychological assessment result showed that the total development quotient was 38. At the age of 1 year and one month, she was 73.8 cm (−0.9SD) tall, 7.6 kg (−2.3SD) in weight, and her head circumference was 44.2 cm (−0.7SD) without obvious facial deformity. She could not walk and speak at the age of 2 years. A diagnosis of motor and language developmental delay and intellectual disability was made. No brain MRI or ultrasound images of different organs were available. At present, amniocentesis is recommended and conducted for prenatal diagnosis in the 2nd pregnancy of Family 2.

**Karyotype analysis results**
Karyotype analysis in Patient 1 demonstrated a derivation of chromosome 13, which was described as 46,XX,rec(13) dup(13q)inv(13)(p12;q14.1)dp (Additional file 1). Subsequent parental karyotype analysis showed that the mother’s karyotype was 46,XX and that of the father was 46,XY,inv(13)(p12;q14.1) (Fig. 1). In the 3rd pregnancy of family 1, karyotype analysis showed that the fetal karyotype was 46, X?,inv(13)(p12;q14.1).

The karyotype analysis in Patient 2 elicited an additional chromosomal material present on the long arm of chromosome X, and described as 46,XX,add(X)(q27.3) (Additional file 1). Both parents had a normal karyotype, indicating that the chromosomal abnormality in Patient 2 was a de novo structural variation.

**Fig. 1** Karyotype analysis result of the proband’s father in Family 1. The arrows indicate chromosomal aberration breakpoints. The karyotype of the proband’s father was 46,XY,inv(13)(p12;q14.1)
Subsequent prenatal diagnosis was carried out in the 2nd pregnancy of Family 2, showing a normal karyotype and no obvious ultrasound anomalies.

**SNP array analysis results**

SNP array analysis results showed a large 72.9-Mb fragment duplication (arr[GRCh37]13q14.11q34(42,195,553_115,107,733)×3) (Fig. 2A) in 13q14.11q34 region in Patient 1, which contained 178 OMIM genes. According to the ACMG guidelines, partial trisomy 13q was interpreted as pathogenic. In addition, no copy number variant was observed in the parents or in the fetus of the 3rd pregnancy of the parents.

SNP array analysis in Patient 2 showed a 92.4-Mb duplication in 13q12.11q34 region (arr[GRCh37]13q12.11q34(22,618,244_115,107,733)×3) (Fig. 2B) associated with a 8.4-Mb deletion in Xq27.3q28 region (arr[GRCh37]Xq27.3q28(146,773,695_155,233,098)×1) (Fig. 2C). Both variants were interpreted as pathogenic CNVs referred to ACMG guidelines. In addition, no copy number variant was observed in the fetus of the 2nd pregnancy of the parents.

**Discussion**

Partial trisomy 13q is a rare chromosomal abnormality with variable clinical phenotypes, with few cases of isolated partial trisomy 13q are available in the literature. In this study, we report two new unrelated cases of partial trisomy 13q in two Chinese families. In addition, we reviewed the clinical findings of isolated partial trisomy 13q in partial reported cases (Table 1), and found that most variants were inherited from the parental chromosome 13 inversions, and few of them were from direct parental inheritance or from de novo.

As present, the genotype–phenotype correlation in partial trisomy 13q has not been fully understood. Partial trisomy 13q4qter is known with variable clinical features similar to trisomy 13. Previous studies have identified isolated trisomy 13q13qter in patients with intellectual disability and facial dysmorphism inherited from their mothers who carried a pericentric inversion of chromosome 13 [20, 26]. Some studies suggested that partial trisomy 13q11q13 may not contribute to most of the features of trisomy 13; in contrast, other studies suggested the region of 13q32qter as the main region causing the clinical features of trisomy 13 [27], which was further
confirmed by the study that elicited a 13q32qter duplication in a patient with several abnormalities [4]. While, a mild clinical feature was observed in an 8-year-old male harboring 13q32qter duplication [28]. In addition, another study conducted by Krygier et al. [6] demonstrated a pure 13q31.1qter duplication in a patient presenting a relatively mild phenotype, suggesting that the critical region of trisomy 13 may be placed close to somewhere in the proximal region. In addition, a patient who carried the 13q14qter duplication exhibited the clinical features including cleft lip/palate, low set ears, depressed nasal bridge, hypertelorism, and epicanthal fold [29]. In the present study, we report two unrelated cases with 13q14q34 duplication, presenting developmental delay, intellectual disability, and other congenital malformations, which are consistent with the previous studies.

As for 13q31.3q32.3 duplication, the GPC5 and GPC6 are indicated as candidate genes for polydactyl [30], while some other studies containing the genes showed the absence of polydactyl [18], suggesting incomplete penetrance in these variants. Similarly, both cases in our study covered the 13q31.3q32.3 region did not have polydactyl. Several studies have shown that patients with pure partial trisomy 13q had the clinical features of cleft lip and palate [23, 29], as well as the Patient 2 in our study shared the smallest region of 13q22q31, indicating that this region may be the critical region for cleft lip and palate phenotype. The duplication of 13q31.1qter was identified in patients with seizures in previous studies [6, 22], and presented in our study as well. Therefore, it is suggested that the distal 13q region may be responsible for seizures. In addition, haemangiomas has been reported to be associated with trisomy of 13q32qter [31], no feature of haemangiomas was observed in our case at present, while, we can not rule out the occurrence of haemangiomas in the further. At present, there are few case reports describing patients who carried 13q duplication and exhibited gastrointestinal abnormalities. However, a previous study elicited internal malrotation in a patient with tetrasomy 13q31qter [32], as well as the case presented in the DECIPHER database (ID:395,925) who harbored a 15.1-Mb duplication in 13q13.2q21 region. In this study, we observed an additional feature of gastric perforation in Patient 1, but whether this is ascribed to partial trisomy 13q needs more investigation.

### Table 1 Clinical findings in partial reported cases with isolated partial trisomy 13q

| References            | Age/Sex | Duplication | Inheritance | Clinical features                                                                 |
|-----------------------|---------|-------------|-------------|----------------------------------------------------------------------------------|
| Krygier et al. [6]    | 12/F    | 13q31.1qter | /           | Learning difficulties, poor speech, facial dysmorphism, seizures, slightly development delay |
| Atack et al. [18]     | 12/M    | 13q31.1q32.3 | Paternal   | Developmental delay, learning disability, facial dysmorphic                        |
| Mathijssen et al. [19]| 2.5/M   | 13q21.3q31.1 | Maternal   | Intellectual disability, developmental delay, dental abnormality, feeding problem, seizures, strabismus, behaviour problem |
| Mehra et al. [20]     | 7/M     | 13q13qter   | Maternal inv(13)(p12q31) | Intellectual disability, developmental delay, learning disability, facial dysmorphic |
| Fraccaro et al. [21]  | 4/F     | 13q21q33.3  | De novo    | Trigonocephaly, low hair implantation, facial dysmorphic, bilateral clinodactyl of the fifth toes, psychomotor retardation |
| Habedank et al. [22]  | 10/M    | 13q22qter   | Maternal inv(13)(p11q21) | Psychomotor retardation, spastic diplegia of the legs, and myoclonic and akinetic seizures, facial dysmorphic, abnormal fingers and toes |
| Williamson et al. [23]| 34/F    | 13q22qter   | Presumptive paternal inv(13)(p11q22) | Short stature, mentally retarded with ptosis, cleft soft palate, and polydactyly |
| Machado et al. [24]   | Newborn/F | 13q14qter  | De novo    | Cyanosis, hydropsy, hypotony, akinesia, and abdomen distension, short neck, and facial dysmorphism |
| Chen et al. [25]      | Fetus   | 13q14.1qter | Paternal inv(13)(p12q14.1) | Intrauterine growth restriction and oligohydramnios, craniofacial dysmorphism, and camptodactyly of the right hand, bilateral subependymal cysts, left renal hydronephrosis, dilated coronary sinus with a persistent left superior vena cava, mild prominence of the left temporal horn |
| This study (Patient 1)| 8/F     | 13q14.11qter| Paternal inv(13)(p12q14.1) | Intellectual disability, motor and speech developmental delay, feeding problem, gastric perforation, seizures |

F Female; M Male
mild clinical features who had der(X)t(X;13)(p21;q32), which may ascribe to inactivation of der(X) chromosome [34]. However, another study revealed a female with a karyotype of 46,X,der(X)t(X;4)(q22;q24) and showed that a high proportion (30%) of tested autosomal genes escaped inactivation, indicating that autosomal material lacking X chromosome specific features is associated with the spreading and/or maintenance of inactivation [35]. In this study, we can not rule out the effect of trisomy 13q that contribute to the clinical features of intellectual disability and developmental delay in Patient 2. In addition, previous studies presented urogenital anomalies including hypospadias, cryptorchidism and duplicated ureter in patients with partial trisomy 13q and partial tetrasomy [28, 32], and the feature of cleft palate as well [23, 29], but no report was available in the literature to clarify the relationship between Xq27.3q28 deletion and the clinical anomalies mentioned above. Thus, we believe the urogenital anomaly of the cervix and the vagina duplication, and cleft palate observed in Patient 2 may be ascribed to partial trisomy 13q. However, more work is needed to determine the genotype–phenotype relationship as shown in Patient 2.

In conclusion, partial trisomy 13q is a rare chromosomal abnormality, especially isolated partial trisomy 13q. In this study, we presented two new unrelated cases of partial trisomy 13q with variable features, exhibiting developmental delay, intellectual disability, and other congenital malformations, which were consistent with the previous studies. In comparison with other studies, we indicated that 13q22q31 region may be critical for cleft lip and palate phenotype and the distal region of 13q may be responsible for seizures. In addition, our study enriched the phenotype spectrum of partial trisomy 13q and further confirmed the genotype–phenotype correlations.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s13039-022-00608-y.

Additional file 1: Karyotype results of Patient 1 and Patient 2.

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Author contributions
JZ designed the study; JZ and CC wrote the article; HZ, YJ, WF and YL performed routine chromosome analysis and data analysis; GW, YX, and XW modified and proofread the paper. All authors approved the final article.

References
1. Ürel Demir G, Doğan OA, Şimşek Kiper PO, et al. Coexistence of trisomy 13 and SRY (-) XX ovotesticular disorder of sex development. Fetal Pediatr Pathol. 2017;36(6):445–51.
2. Martin-de Saro M, Compean Z, Aguilar K, et al. Partial trisomy 13q/Monosomy 3p resulting from a paternal reciprocal 3p;13q translocation in a boy with facial dysmorphism and hypertrophic cardiomyopathy. Mol Syndromol. 2021;12(5):305–11.
3. Du J, Tan YQ, Li LY, Lu GX. Molecular cytogenetic detection of partial chromosome 13q trisomy and its relation with the clinical features of torticolis. Zhonghua Yi Xue Yi Chuan Xue Za Zhi. 2003;20(3):189–92.
4. Rao VV, Carpenter NJ, Gucsavas M, Coldwell J, Say B. Partial trisomy 13q1q identified by sequential fluorescence in situ hybridization. Am J Med Genet. 1995;58(1):50–3.
5. Patil SJ, Phadke SR. Pericentric inversion causing duplication and deletion of chromosome region 13q22 → 13qter in the offspring. Am J Med Genet A. 2007;143A(1):82–4.
6. Krygier M, Lipska-Ziętkiewicz BS, Koczkowska M, Wierzbza J, Limon J. Mild phenotype of a large partial 13q trisomy. Clin Dysmorphol. 2014;23(4):155–7.
7. Xiao W, Gao Z, Meng Q, Zhang M. Cytogenetic and molecular genetic diagnosis of a neonate with partial 13q trisomy and partial 5p monosomy. Zhonghua Yi Xue Yi Chuan Xue Za Zhi. 2014;31(6):747–9.
8. Rogers JF. Clinical delineation of proximal and distal partial 13q trisomy. Clin Genet. 1984;25(3):211–9.
9. Katoh K, Aiba K, Fukushima D, et al. Clinical and molecular genetic characterization of two female patients harboring the Xq27.3q28 deletion

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Availability of data and materials
The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent for participate
Ethics Committee approval was obtained from the Institutional Ethics Committee of Quanzhou Women’s and Children’s Hospital to the commencement of the study (2020No.31).

Consent for publication
We confirmed that all subjects participating in this study signed written informed consent for publication of their own and their children's genetic data and relevant information.

Competing interests
The authors declare that they have no conflict of interests.
with different ratios of X chromosome inactivation. Hum Mutat. 2020;41(8):1447–60.

10. Gercz J, Geideon AK, Sutherland GR, Mulley JC. Identification of the gene FMR2, associated with FRAXE mental retardation. Nat Genet. 1996;13(1):105–8.

11. Birot AM, Delobel B, Gronnier P, Bonnet V, Maire L, Bozon D. A S-megabase familial deletion removes the IDS and FMR-1 genes in a male Hunter patient. Hum Mutat. 1996;7(3):266–8.

12. Clarke JT, Wilson PJ, Morris CP, et al. Characterization of a deletion at Xq27-q28 associated with unbalanced inactivation of the nonmutant X chromosome. Am J Hum Genet. 1996;51(2):316–22.

13. Marshall LS, Simon J, Wood T, et al. Deletion Xq27.3q28 in female patient with global developmental delays and skewed X-inactivation. BMC Med Genet. 2013. https://doi.org/10.1186/1471-2350-14-49.

14. Zhuang J, Wang Y, Zeng S, Lv C, Lin Y, Jiang Y. A prenatal diagnosis and genetics study of five pedigrees in the Chinese population with Xp22.31 microduplication. Mol Cytogenet. 2019. https://doi.org/10.1186/s13039-019-0461-1.

15. Wang H. Introduction and interpretation of the updated contents of the international system for human cytogenomic nomenclature. Zhonghua Yi Xue Yi Chuan Xue Za Zhi. 2021;38(12):1165–70.

16. Zhuang J, Zhang N, Fu W, et al. Cytogenetic and molecular analysis of distal 4q duplication with distinctive phenotype using single-nucleotide polymorphism array. Mol Cytogenet. 2021;14(1):46.

17. Kearney HM, Thorland EC, Brown KK, et al. American college of medical genetics standards and guidelines for interpretation and reporting of postnatal constitutional copy number variants. Genet Med. 2011;13(7):680–5.

18. Atack E, Fairtlough H, Smith K, Balasubramanian M. A novel paternal du- tion into autosomal material of an x;autosome translocation: evidence for a difference between autosomal and X-chromosomal DNA. Am J Hum Genet. 2011;3(7):680–5.

19. Mathijsen IB, Hoovers JM, Mul AN, Man HY, Ket JL, Hennekam RC. Array comparative genomic hybridization analysis of a familial duplication of chromosome 13q: a recognizable syndrome. Am J Med Genet A. 2005;136(1):76–80.

20. Mehra S, Christ L, Jung L, Zinn AB, Schwartz S. Characterization of a familial balanced rec(13) in a child with mild MR and his half-sibling with two structurally rearranged chromosomes 13. Am J Med Genet A. 2005;137(2):217–21.

21. Fraccaro M. De novo duplication 13q (46, XX, dup(13)(q12–q33)). Hum Genet. 1981;68(4):444.

22. Habedank M. Familial pericentric inversion of chromosome 13 resulting in duplication 13q22 to qter. J Med Genet. 1982,19(3):227–9.

23. Williamson EM, Miller JF, Seabright M. Pericentric inversion (13) with two different recombinants in the same family. J Med Genet. 1980;17(4):309–12.

24. Machado IN, Heinrich JK, Campanhol C, Rodrigues-Penes RM, Oliveira FM, Banini R. Prenatal diagnosis of a partial trisomy 13q (q14–qter): phenotype, cytogenetics and molecular characterization by spectral karyotyping and array comparative genomic hybridization. Genet Mol Res. 2010;9(1):441–8.

25. Chen CP, Lin SP, Lee CC, Chen WL, Chen LF, Wang W. Perinatal findings of partial trisomy 13q (13q14.1–qter) resulting from paternal pericentric inversion of chromosome 13. Prenat Diagn. 2005,25(6):527–8.

26. Lucas J, Le Mee F, Picard F, Le Marec B, Junien C. Trisomy 13q13 leads to qter pure par recombinacion aneuquisome d’une inversion pericentrique maternelle [Pure trisomy 13q13–qter caused by aneuquisome recombinacion of a maternal pericentric inversion]. Ann Genet. 1983;26(3):187–90.

27. Warburton PE, Dolled M, Mahmood R, et al. Molecular cytogenetic analysis of eight inversion duplications of human chromosome 13q that each contain a neocentromere. Am J Hum Genet. 2000;66(6):1794–806.

28. Helali N, Iafolla AK, Kahler SG, Qumsiyeh MB. A case of duplication of 13q32–qter and deletion of 18p11.32–qter with mild phenotype: patau syndrome and duplications of 13q revisited. J Med Genet. 1996;33(7):600–2.

29. Ou CY, Hsu TY, Chang JC, Chang SY. Partial trisomy 13 (46, XY, dup(13) (q14–31)): a case report. Changgeng Yi Xue Za Zhi. 1998;21(1):82–5.

30. van der Zwaag PA, Dijkhuizen T, Gerssen-Schoorl KB, et al. An interstitial duplication of chromosome 13q31 3q32.1 further delineates the critical region for postaxial polydactyly type A2. Eur J Med Genet. 2010;53(1):45–9.

31. Gordon PL, Dalton JD, Martens PR, Tharapel AT, Wilroy RS. Elucidation of the centromere involvement in an inversion (13) by fluorescent in situ hybridisation. J Med Genet. 1993;30(5):414–6.

32. Haddad V, Aboura A, Tosca L, et al. Tetrasomy 13q31.1qter due to an inverted duplicated neocentric marker chromosome in a fetus with multiple malformations. Am J Med Genet A. 2011;158A(4):894–900.

33. Mattei MG, Matter JF, Ayme S, Giraud F. X-autosome translocations: cytogenetic characteristics and their consequences. Hum Genet. 1982;64(1):295–309.

34. Blennoe V, Sahlin S. Rescue from the effects of trisomy 13q32–qter owing to skewed X inactivation in a der(X)(X;13)(p21;q32) carrier. J Med Genet. 1999;36(9):718–9.

35. White WM, Willard HF, Van Dyke DL, Wolff DJ. The spreading of X inactiva- tion into autosomal material of an xautosome translocation: evidence for a difference between autosomal and X-chromosomal DNA. Am J Hum Genet. 1998;63(1):20–8.

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