Structural and functional characteristics of cGMP-dependent methionine oxidation in Arabidopsis thaliana proteins

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Abstract

Background: Increasing structural and biochemical evidence suggests that post-translational methionine oxidation of proteins is not just a result of cellular damage but may provide the cell with information on the cellular oxidative status. In addition, oxidation of methionine residues in key regulatory proteins, such as calmodulin, does influence cellular homeostasis. Previous findings also indicate that oxidation of methionine residues in signaling molecules may have a role in stress responses since these specific structural modifications can in turn change biological activities of proteins.

Findings: Here we use tandem mass spectrometry-based proteomics to show that treatment of Arabidopsis thaliana cells with a non-oxidative signaling molecule, the cell-permeant second messenger analogue, 8-bromo-3,5-cyclic guanosine monophosphate (8-Br-cGMP), results in a time-dependent increase in the content of oxidised methionine residues. Interestingly, the group of proteins affected by cGMP-dependent methionine oxidation is functionally enriched for stress response proteins. Furthermore, we also noted distinct signatures in the frequency of amino acids flanking oxidised and un-oxidised methionine residues on both the C- and N-terminus.

Conclusions: Given both a structural and functional bias in methionine oxidation events in response to a signaling molecule, we propose that these are indicative of a specific role of such post-translational modifications in the direct or indirect regulation of cellular responses. The mechanisms that determine the specificity of the modifications remain to be elucidated.

Keywords: Methionine oxidation, Reactive oxygen species, 3,5-cyclic guanosine monophosphate, Tandem mass spectrometry-based proteomics, Arabidopsis thaliana

Findings

The debate of whether methionine (Met) oxidation of proteins is a purely chemical consequence of cellular oxidative damage or a protective mechanism against oxidative damage, or indeed a post-translational modification that can act as a specific cellular signal and/or response, is ongoing [1-3]. To shed light on this question we treated Arabidopsis suspension culture cells with the cell permeable second messenger analogue 8-bromo 3,5-cyclic guanosine monophosphate (8-Br-cGMP). Cyclic GMP has a signaling role in many plant responses, including responses to light [4], hormones [5-8], signaling peptides [9], salt and drought stress [10,11], ozone, and defence responses [12-14]. Given that cGMP is not an oxidising agent and does not induce protein Met oxidation in vitro (Additional file 1), we tested if cGMP causes protein oxidation in vivo. To this end we used an OxiSelect™ Intracellular ROS Assay Kit (Cell Biolabs, Inc.) and show that cGMP can cause protein oxidation (Figure 1). We therefore conclude that any protein Met oxidation event resulting from cGMP treatment is most likely the result of direct or indirect cellular processes. To further characterise cGMP-dependent Met oxidation in vivo, a proteomic analysis was performed on A. thaliana (ecotype Col-0) cell
In our proteomic analysis we considered a peptide as containing oxidised Met residue when it was identified with high confidence (≥ 95%) in at least two biological replicates. A total of 385 cGMP-dependent methionine oxidised proteins were identified (Additional file 2, tab “AF1”). Assigned spectral counts (Additional file 2, tab “AF2”) were used to estimate the relative ratio of peptides containing oxidized Met residue(s) as compared to total number of peptides identified in the sample.

An example of a tandem mass spectrometry result demonstrating oxidative modification of TiO₂-enriched peptides extracted from 8-Br-cGMP-treated cells is shown in Figure 2. Peptides containing single oxidised Met residue show an increase in mass to charge (m/z) ratio of 15.9994 that corresponds to the average mass of an oxygen atom. For example, the peptide fragment (DHDKPIQQVIAEMTDGGVDR) of the alcohol dehydrogenase 1 (AT1G77120) in non-oxidised form has the m/z ratio of 850.3723 (Figure 2A), while after oxidation of Met residue, the m/z ratio shifts to 866.3673 (Figure 2B).

Further, we identified peptides with oxidised Met that occurred in all three biological replicates at different time points. We noted an increase in the total number of peptides containing residues of oxidised Met after cGMP treatment from 221 to 633 and then 1451 at 0, 30 and 60 minutes, respectively (Figure 3A and Additional file 2, tab “AF2”). These numbers represent 1.4%, 19.4% and 13%, respectively, of the total number of peptides identified at each time point. Thus, the percentage of Met oxidised peptides identified is the highest at 30 minutes. In addition, the numbers of oxidised Met peptides detected at each time-point suggest that the total number of oxidised Met residues increased nearly 3-fold during the first 30 minutes of treatment and 7-fold after 60 minutes of treatment (Additional file 2, tab “AF2”). Of these redundant peptide fragments containing oxidised Met, 14 at 0 minutes, 113 at 30 minutes and 288 at 60 minutes were unique for each time-point. The total Met oxidised peptides correspond to 34 (at 0'), 136 (at 30') and 281 (at 60') Arabidopsis proteins, from which 10, 94, and 224 identified oxidized Met proteins are unique for each time-point, respectively (Figure 3B). This finding implies either that cGMP-dependent Met oxidized Met residues have undergone proteolysis.

Gene ontology (GO) analysis (Fatigo+; http://babelomics3.bioinfo.cipf.es/) [18] of the unique proteins containing oxidised Met residues was undertaken. The result shows a significant enrichment of GO terms (adjusted p-value<1.00e⁻⁰⁵) in categories including ‘response to stress’, ‘response to abiotic stimuli’, ‘response to oxidative stress’, and ‘response to oxygen and ROS metabolic process’ (see Additional file 2, tab “AF3”). Moreover, the gene number in these GO
categories also increased over time (Figure 4). It therefore appears that the application of a membrane-permeable analogue of the second messenger cGMP induces Met oxidation in a set of proteins that are over-represented in specific functional groups.

Proteins enriched in the GO categories: ‘response to oxidative stress’, ‘response to ROS’, ‘response to oxygen’, and ‘ROS metabolic process’ showed four main patterns.

1. Loss of peptides or proteins containing oxidised Met residue(s) and reduction in the number of peptide copies after treatment, e.g. glyceraldehyde-3-phosphate dehydrogenase (AT1G13340; AT3G04120) and 60S acidic ribosomal protein (AT2G27720). Loss of Met oxidised peptides can occur due to degradation of copies of proteins with oxidised Met residues or the reduction of oxidised Met residues.

2. Increase in the number of oxidised peptide fragments after treatment, e.g. peroxidase (AT2G22420; AT4G08770). This may imply that cGMP either indirectly induces over-expression of specific proteins with peptide fragments susceptible to oxidation or preferentially induces oxidation of Met residues in specific proteins without necessarily inducing transcription and/or translation.

3. New peptides detected with Met residues oxidised after treatment, e.g. ATP synthase (AT5G08670),

![Figure 2](image.png)

**Figure 2** MS/MS spectra of ADH1 containing non-oxidised (A) and oxidised (B) methionine residues. Three biological replicates of 10 μM 8-Br-cGMP-treated cells and H2O mock treated controls were collected at 0, 30 and 60 min. Proteins were precipitated using 10% (w/v) trichloroacetic acid in acetone, re-solubilised in 7 M urea, 2 M thiourea and 4% (w/v) CHAPS, reduced, alkylated and trypsin digested. Peptides were fractionated by cation exchange chromatography. Methionine oxidised peptides were enriched using TiO2 beads and re-suspended in 5% (v/v) acetonitrile and 0.1% (v/v) formic acid prior to identification and quantitation by LTQ Orbitrap coupled with a nanoelectrospray ion source. Peptides (5 μL) were injected onto a 50 mm × 0.3 mm Magic C18AQ column. The top 10 precursor ions were selected with a resolution of 60,000 for fragmentation using normalized collision-induced dissociation set at 35.0. Spectra were searched against TAIR using MASCOT, with a precursor mass tolerance of 10 ppm, a fragment ion mass tolerance of 0.3 Da, one missed cleavage, carbamidomethyl cysteine residues as fixed modification and oxidation and dioxidation of methionine residues as variable modifications. Proteins with a score > 95% were considered positively identified (corresponding score ≤ 31). Spectra were further processed with the Scaffold™ software using the ‘Trans-Proteomic Pipeline’ algorithm (threshold 95%). Oxidised Met residues showed an increase in mass/charge ratio (m/z) of 15.9994. Arrows show Met residues at position 13 in the fragment DHDKPIQQVIAEWTDDGVDR of AT1G77120 before oxidation (m/z ratio 850.3723) (A) and after oxidation (m/z ratio 866.3673) (B).

**Table 1** Enrichment of methionine oxidized peptides (oxMet) using TiO2 with and without DHB

|                      | Without enrichment | TiO2 enrichment without DHB | TiO2 enrichment with DHB |
|----------------------|--------------------|-----------------------------|--------------------------|
|                      | R1     | R2     | R3     | R4     | R1     | R2     | R3     | R4     | R1     | R2     | R3     | R4     |
| Assigned spectra     | 123109 | 67688  | 42897  | 89362  | 38403  | 56545  | 14450  | 4288   | 28092  | 11880  | 8567   | 3812   |
| Assigned spectra oxMet pep. | 3073   | 288    | 49     | 3930   | 15803  | 4273   | 6303   | 424    | 26913  | 4370   | 6586   | 1318   |
| % oxMet              | 2.5    | 0.4    | 0.1    | 4.4    | 4.12   | 7.6    | 43.6   | 9.9    | 95.8   | 36.8   | 76.9   | 34.6   |

*The cell suspension culture was treated with 10 μM H2O2 for 30 minutes. The experimental procedures are as described in the legend to Figure 2.*
and (4) multiple Met residues becoming oxidised after treatment, e.g. in the heat shock protein 70 (AT3G12580) (Additional File 2, tab “AF1”).

A total of 299 proteins (78% of all unique proteins containing oxidised Met residues) comprised only one oxidised Met residue, while 86 proteins (22% of all unique proteins containing oxidised Met residues) had ≥ 2 oxidised Met residues (Additional file 2, tab “AF4”). Of those multi-oxidised 86 proteins 10 proteins had ≥ 4 Met residues oxidised (Additional file 2, tab “AF5”). The ‘low expression of osmotically responsive gene 1’ (AT1G56070), ‘tubulin 9’ (AT4G20890), ‘heat shock protein 70’ (AT5G02500) and ‘male gametophyte defective 1’ (AT2G21870) contain the largest number of oxidised residues, representing 16%, 23%, 29% and 50%, respectively, of their total Met content (Table 2 and Additional file 2, tab “AF5”). It is of interest that heat shock proteins AT3G02500, AT3G12580 and AT5G09590 each had at least four oxidised Met residues.

The protein with the highest number of Met oxidised peptide fragments in response to cGMP is the methylesterase PCR A (AT1G11580) with 235 oxidised fragments representing ≈ 30% of the identified peptides (Table 3 and Additional file 2, tab “AF5”). Among the proteins with peptides show the greatest extend of Met oxidation, peroxidase superfamily protein (AT2G22420) showed the highest ratio of Met oxidised peptides to all identified peptide fragments of the protein (114 out of 160), representing ≈ 70% of total peptides detected and assigned to this protein. Localised in the cytosol, this family of proteins is mainly involved in oxidation-reduction processes and responses to oxidative stress. In addition to ROS being generated by activation of a plasma membrane NADPH oxidase [19], an extracellular cell wall peroxidase is also involved in the biosynthesis of H₂O₂ [20] and plays an important role in plant resistance to pathogens [21]. A previous study using antisense expression of a French bean peroxidase cDNA in Arabidopsis showed a reduction in mRNA level of peroxidase (AT3G49120), which in turn led to a reduction in the oxidative burst and eventually a reduced resistance to fungal and bacterial pathogens [22]. The authors hypothesized that peroxidases have a role in sustaining and/or initiating ROS that signal early defence responses in plants. Given that peroxidases themselves are highly susceptible to Met oxidation, and that this modification limits their activity, it suggests a negative feedback mechanism and links Met oxidation to the control of cellular redox balance. It has also been reported that O₂ and NO, both oxidising agents, induce transcriptional activation of scavenger-encoding genes, like alternative oxidase and glutathione peroxidase [12]. Furthermore, oxidation of Met residues has been shown to target both specific functional domains and consequently modify functional characteristics, e.g. in cytochrome c [23] and peroxidases [24], as well as regions outside functional domains, like in the nascent polypeptide-associated complex where modifications do not seem to affect functionality.

Given that Met oxidation can profoundly alter cellular responses, the question is if there is evidence for site selectivity and if so, what determines it. In order to address this question, we have subjected all 385 proteins with one or several oxidised Met to further analysis and noted the following. Firstly, the average Met frequency in the set of proteins containing oxidised Met is 0.027 as compared to the other amino acids (AA) in the complete proteome where it is 0.025 [25], and hence is nearly the same; we also noted that none of the 50 Arabidopsis proteins with the highest frequency of Met occurrence
(0.09) contained any oxidised Met residues. This indicates that Met frequencies in proteins per se are not a factor that determines increased oxidation. Secondly, of the 385 proteins containing at least one modified Met residue, only eight residues were on the N-terminus, and of the 150 double Met (-MM-) residues, five proteins had both residues oxidised and eight proteins - only one. Thirdly, of the 575 oxidised Met residues detected, 75 had a glutamic acid (Glu) and 68 had an aspartic acid (Asp) as an immediate C-terminal neighbour and this bias is likely, at least in part, due to preferential enrichment of these AAs by TiO2 [26], however we also find the uncharged alanine (Ala: 67) enriched on the C-terminus. The most frequent N-terminal neighbours are glutamic acid (Glu: 67) and again the uncharged alanine (Ala: 65). The least frequent C- and N-terminal neighbours are tryptophan (Trp: 2; 0,

### Table 2 cGMP-dependent proteins with multiple methionine residues oxidised and their enriched GO terms

| Accession No. | Description | *OxMet/total Met residues | Annotations | Enriched GO term(s) |
|---------------|-------------|---------------------------|-------------|---------------------|
| AT1G56070     | Low expression of osmotically responsive gene 1 | 5/38 | RC, translational elongation | RTS |
| AT2G21870     | Male gametophyte defective 1 | 5/10 | Co2+, Cu2+ and Zn2+ binding | |
| AT4G20890     | Tubulin 9 | 5/22 | GTP binding, GTP catabolic process, RCA | |
| AT5G02500     | Heat shock protein 70 | 5/17 | Responses to heat, bacterium, H2O2 and HLI, RCa, RTS, RV | RTS, RV |
| AT3G12580     | Heat shock protein 70 | 4/16 | Responses to heat, bacteria, H2O2, HLI, RCa, RTS, RV | RTS, RV |
| AT3G27240     | Cytochrome C1 family | 4/8 | Heme binding, Fe2+ binding | |
| AT4G02930     | GTP binding elongation factor Tu family protein | 4/14 | ATP, Co2+ and Zn2+ binding, RCa | |
| AT1G4310      | Unknown protein | 4/12 | | |
| AT4G27390     | Translocase outer membrane 20 | 4/8 | Metal ion binding | |
| AT4G10480     | Nascent polypeptide-associated complex | 4/10 | Transcription regulation and mitochondrial translocation | |

HLI- high light intensity, RC- response to cold, RCA- response to cadmium ion, RTS- response to temperature stimulus (GO:0009266), RV- response to virus (GO:0009615), adjusted p < 1.78e-2. *Ox- oxidised.
respectively) and cysteine (Cys: 1; 2, respectively). We further compared the observed frequency of amino acids flanking Met positions to their theoretically expected values implied from the analysis of overall AA frequencies in the entire Arabidopsis proteome [25]. This process involved counting all of the occurrences of AAAs in the proteome and establishing their relative frequency in the proteome, and the analysis was done in Matlab (Version R2010b). We note that, while under-represented (i.e. the observed frequency in positions flanking Met is lower than their average in the entire proteome), leucine and serine are the most frequent flanking AAAs. In turn, Met flanking a Met is 35% over-represented on the N-terminal side and 25% over-represented on the C-terminal side. In contrast, oxidised Mets have much reduced relative preference for Met (-14% on N-terminus and -44% on C-terminus), albeit based on a very limited sample. Other under-represented flanking AAAs are cysteine (Cys) and proline (Pro).

The rapidly evolving field of redox proteomics provides new evidence supporting the notion that oxidation of Met residues may have a great impact on protein activity, regulation of biochemical pathways and cellular function in response to changing environmental conditions. This is consistent with the observed Met oxidation accumulation in plants under low temperature conditions and the fact that plant methionine sulfoxide reductase (MSR) confers increased tolerance to freezing [27]. It is also conceivable that the differential oxidation footprint of Met is a result of different susceptibility depending e.g. on the conformation of the protein or on differential access for repair of the MSR to different proteins or protein domains. Furthermore, our results are an indication that many of the Arabidopsis proteins involved in modulating the level of reactive oxygen species (ROS), including ROS-scavenging and ROS-producing proteins [19], may - at least in part - be regulated by oxidation of their Met residues.

### Additional files

**Additional file 1:** The file contains the list of methionine oxidised peptides and their assigned spectral counts following *in vitro* cGMP treatment experiment.

**Additional file 2:** The file contains the lists of all methionine oxidised proteins and fragments and the result of the gene ontology analysis.

### Competing interests

The authors declare that they have no competing interests.

### Authors’ contributions

CG conceived the project. CM, IT, BP, LT and KSL designed and performed the experiments, BJ performed the bio-informatics analyses, and CG wrote the manuscript with critical input from all authors. All authors read and approved the final manuscript.

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