Abstract. Background/Aim: Dual-specificity protein phosphatase 4 (DUSP4) negatively regulates MAPK signaling and is involved in various cellular processes. We herein evaluated the relationship between DUSP4 expression and clinicopathological characteristics in a large series of gastric cancer samples. Materials and Methods: DUSP4 expression was examined by immunohistochemistry in 508 gastric cancer samples. Cases were classified according to the TCGA molecular classification and HER2 amplification. Kaplan-Meier plots were used to predict the relationship between mRNA expression of DUSP4 and survival. Results: Low expression of DUSP4 was significantly correlated with larger tumor size, higher pT category, positive nodal status, higher stage, lymphovascular invasion, perineural invasion, worse overall survival, and worse recurrence-free survival. No correlation was observed between DUSP4 expression and molecular characteristics. Bioinformatics analysis showed that low mRNA expression was associated with a poor prognosis. Conclusion: Low expression of DUSP4 is associated with aggressive phenotypes of gastric cancer and a poor prognosis.

Gastric cancer is a common epithelial malignant neoplasm of the digestive system. According to the GLOBOCAN 2018 database, gastric cancer shows the fifth highest incidence and the third highest mortality rate worldwide (1). It is the most common cancer in East Asia, including Korea. Although the age-standardized rate of incidence and death is declining, it still has a high mortality rate (2). Therefore, it is important to investigate novel treatment and prognostic biomarkers for gastric cancer.

Surgical resection is the only method for curative treatment of gastric cancer, but the prognosis is poor for advanced or metastatic gastric cancer. Therefore, adjuvant chemotherapy has been performed to improve survival, and recent advances in molecular biology have led to discovery of various targeted therapies. For example, the use of trastuzumab, an anti-human epidermal growth factor receptor-2 (HER2) drug, was approved with cytotoxic drugs for the treatment of metastatic HER2-overexpressing gastric cancer. Additionally, clinical trials for other molecular targets (EGFR, CLDN18.2, VEGFR) are ongoing (3).

Dual-specificity protein phosphatase 4 (DUSP4), also known as mitogen-activated protein kinase phosphatase (MKP)-2, negatively regulates mitogen-activated protein kinase (MAPK) signaling and is involved in various cellular processes such as cell proliferation and immune response (4). There have been many studies on the role of DUSP4 in cancer. In vitro studies of EGFR-mutated lung cancer (5), papillary thyroid carcinoma (6, 7), and pancreatic cancer (8) revealed increased DUSP4 expression. Low expression of DUSP4 was associated with a poor prognosis in a study of diffuse large B-cell lymphoma (9), whereas high expression of DUSP4 was associated with lymph node metastasis and extrathyroidal extension in a study of papillary thyroid carcinoma (7). The relationship between DUSP4 expression and aggressive features showed contradictory results in studies of colorectal cancer (10, 11). Low DUSP4 expression was associated with a poor prognosis in a study of triple-negative breast cancer (12); however, high DUSP4 expression was associated with a poor prognosis in early T-stage breast cancer (13). In vitro studies of gastric cancer cell lines revealed that up-regulated DUSP4 expression was associated with drug resistance, epithelial-mesenchymal transition,
migration, and invasion (14, 15). Immunohistochemical analysis of gastric cancer tissue confirmed that low DUSP4 expression was associated with sex, tumor size, depth of invasion, and distant metastasis (16).

In this study, we evaluated the relationship between DUSP4 expression and the clinicopathological characteristics of gastric cancer using semi-quantitative immunohistochemical (IHC) staining in a large series of resected gastric cancer samples. Additionally, the association between DUSP4 expression and specific molecular subtypes was evaluated.

**Materials and Methods**

**Clinical data and pathological evaluation.** Gastric cancer cases were retrospectively enrolled as patients who underwent surgical resection between February 2005 and July 2010 at the Hanyang University Hospital. A total of 520 cases were collected, and 9 cases were excluded from the study because they had distant metastasis at the time of diagnosis or neoadjuvant therapy. In addition, two cases had a positive surgical margin, and one case was excluded because the patient died immediately after surgery. Consequently, 508 cases were included in this study. To compare the expression of DUSP4, 27, 54, and 63 samples of the normal gastric mucosa (≥1 cm from the tumor), adenoma, and lymph node metastasis were additionally obtained, respectively.

Medical records were reviewed to confirm the patient’s clinical information including age, sex, adjuvant chemotherapy, death, and recurrence date. A review of hematoxylin and eosin (H&E) slides and pathologic reports was performed by two pathologists (SSB and SSP). Tumor location, and size, gross type, histology, lymphovascular invasion and perineural invasion, and tumor-node-metastasis (TNM) stage were evaluated. Tumor histology was classified based on the Lauren classification and 2019 WHO classification, and the TNM stage was classified according to the 8th edition of the American Joint Committee on Cancer (AJCC). The clinicopathological characteristics of gastric cancer patients are summarized in Table I.
Tissue microarray construction and immunohistochemistry. Tissue microarray systems (Tissue Microarray Set, Labro, Seoul, Republic of Korea) and formalin-fixed paraffin-embedded (FFPE) tissue were used for tissue microarray (TMA) construction. The blocks of gastrectomy or endoscopic submucosal dissection (ESD) specimens were used for normal gastric mucosa, tubular adenoma, and gastric cancer tissue. For LN tissue, the block with the largest metastatic tumor in the LN dissection specimen was used. After determining the representative area on the H&E slides by light microscopy, tissue was collected through a 3.0 mm punch.

Immunohistochemistry was performed to evaluate the expression of DUSP4. Deparaffinization and rehydration were performed on 4 μm sections by immersing the sections in xylene and graded series of ethanol. Heat-induced epitope retrieval (100°C for 20 min in sodium citrate buffer) and blocking of endogenous peroxidase using the peroxidase-blocking solution (S2023, Dako, Glostrup, Denmark) for 15 min were performed. Sections were incubated at 4°C overnight with the diluted DUSP4 antibody (1:200, Rabbit DUSP4 polyclonal antibody, ab72593, Abcam, Cambridge, MA, USA).

Labeled polymer and 3,3’-diaminobenzidine tetrahydrochloride chromogen (K5007, EnVision™ Detection System, Dako, Glostrup, Denmark) were sequentially used for detection according to the manufacturer’s instructions.

Table II. DUSP4 expression in various gastric tissues.

| Samples           | Cases | Low expression (%) | High expression (%) | p-Value |
|-------------------|-------|--------------------|---------------------|---------|
| Normal mucosa     | 27    | 4 (14.8%)          | 23 (85.2%)          | <0.001  |
| Tubular adenoma   | 54    | 7 (13.0%)          | 47 (87.0%)          |         |
| Gastric cancer    | 508   | 233 (45.9%)        | 275 (54.1%)         |         |
| LN metastasis     | 63    | 34 (54.0%)         | 29 (46.0%)          |         |

DUSP4, Dual-specificity protein phosphatase 4; LN, lymph node.

Figure 1. Immunohistochemical staining of dual-specificity protein phosphatase 4 (DUSP4) in gastric cancer. (a): Negative; (b): weakly positive expression; (c): intermediate expression; (d): strong positive expression, ×200.
Interpretation of IHC staining. DUSP4 expression was evaluated by two pathologists without access to clinical data (SSB and SSP). For the semi-quantitative assessment, histoscore (H-score) was calculated based on the intensity and percentage of nuclear DUSP4 expression. The intensity of staining was classified as 0 to 3 (0: negative, 1: weak, 2: intermediate, 3: strong) and the positive rate was scored from 0 to 100. Following this, the optimal cutoff value of DUSP4 expression was determined using the receiver operating characteristics (ROC) curve, and cases were divided into high or low expression.

Microsatellite instability and Epstein-Barr virus status. The TCGA network classified gastric cancer into four molecular subtypes using comprehensive molecular evaluation: Epstein-Barr virus (EBV), microsatellite instability (MSI), genomic stability (GS), and chromosomal instability (CIN) (17). In this study, IHC staining of mismatch repair proteins was performed to identify the MSI subtype and EBV-encoded RNA in situ hybridization (EBER-ISH) was performed to confirm the EBV subtype. IHC staining was performed on the entire tumor section for MLH1 (G168-728, Cell Marque, Rocklin, CA, USA), PMS2 (MRQ-28, Cell Marque, CA, USA), MSH2 (G219-1129, Cell Marque, CA, USA), and MSH6 (PU29, Leica Biosystems, Nussloch, Germany) and classified as MSI subtype when one or more results were negative. Cases with diffuse positivity when stained using the INFORM EBER Probe (Roche, Basel, Switzerland) on TMA slides were classified as EBV subtype.

HER2 status. IHC and silver DNA in situ hybridization (SISH) tests were performed on all sections of the TMA to evaluate HER2 amplification. Automatic staining was performed according to the manufacturer’s instructions with a rabbit monoclonal anti-HER-2 antibody (4B5, Roche) and INFORM HER2 Dual ISH DNA probe cocktail (Roche). HER2 amplified cases were defined as an IHC score 3 or a HER2/ chromosome 17 (CEP17) ratio of ≥2.0 (18).

Bioinformatics. Kaplan-Meier plotter (KM plotter, http://kmplot.com/analysis/) is an online analysis tool that provides the gene expression data for 1,065 gastric cancer samples (19). It is used to predict the association between the mRNA expression of biomarker candidates and the survival of gastric cancer patients. The effect of DUSP4 expression on the prognosis of gastric cancer patients was estimated using this tool in this study.

Statistical analyses. The Kruskal-Wallis test was performed to compare DUSP4 expression in normal gastric mucosa, tubular adenoma, gastric cancer, and lymph node metastasis. Pearson’s chi-square ($\chi^2$) and Fisher’s exact tests were performed to evaluate the correlation between DUSP4 expression and clinicopathological factors. The Kaplan-Meier method with log-rank test was performed to evaluate the influence of DUSP4 expression on overall survival (OS) and recurrence-free survival (RFS), and univariate and multivariate Cox regression analyses were performed to identify the significant prognostic factors. SPSS software version 25.0 (IBM, Armonk, USA) was used for all statistical analyses, and $p$-value <0.05 was considered statistically significant.

Results

The pattern of DUSP4 expression. Nuclear DUSP4 expression was variable, and representative images are presented in Figure 1. Cases were divided into high-expression group (H-score ≥200) and low-expression group (H-score <200) using the ROC curve. High DUSP4 expression was observed in 23 cases (85.2%) of normal gastric mucosa, 47 cases (87.0%) of tubular adenoma, 275 cases (54.1%) of gastric cancer, and 29 cases (46.0%) of LN tissue with gastric cancer metastasis (Table II).
The correlation between DUSP4 expression and clinicopathological features. The correlation between DUSP4 expression and clinicopathological features was investigated and is summarized in Table III. Low expression of DUSP4 was significantly correlated with larger tumor size \((p<0.001)\), presence of lymphovascular invasion \((p<0.001)\), presence of perineural invasion \((p<0.001)\), higher pT category \((p<0.001)\), nodal metastasis \((p<0.001)\) and higher stage \((p<0.001)\). No significant correlation was observed between DUSP4 expression and other features including age, gender, tumor location, Lauren classification, and histological type.

The correlation between DUSP4 expression and molecular characteristics. According to the TCGA molecular classification, 34 cases (6.7%) were classified as EBV subtype and 40 cases (7.9%) were classified as MSI subtype. The remaining 434 cases (85.4%) were classified as GS or CIN subtype. In this study, these two subtypes were not distinguished and classified as EBV-negative or microsatellite stable (MSS). There were 24 HER2-amplified cases (4.7%) and these cases were EBV-negative or MSS. There was no correlation between DUSP4 expression and molecular characteristics (Table III).

Prognostic significance of various factors including DUSP4 expression. In univariate analyses, various factors showed a significant association with OS or RFS. Low DUSP4 expression \((p=0.028)\), older age \((p<0.001)\), larger tumor size \((p<0.001)\), diffuse and mixed type of Lauren classification \((p=0.016)\), higher pT category \((p<0.001)\), nodal metastasis \((p<0.001)\), higher TNM stage \((p<0.001)\), lymphovascular invasion \((p<0.001)\), and perineural invasion \((p<0.001)\) were associated with short OS. Low DUSP4 expression \((p=0.004)\), larger tumor size \((p<0.001)\), undifferentiated and other types of WHO histological classification \((p<0.001)\), diffuse and mixed type of Lauren classification \((p<0.001)\), higher pT category \((p<0.001)\), nodal metastasis \((p<0.001)\), higher TNM stage \((p<0.001)\), lymphovascular invasion \((p<0.001)\), and perineural invasion \((p<0.001)\) were associated with short RFS. In multivariate analyses, older age \((p<0.001)\), higher TNM stage \((p<0.001)\), and perineural invasion \((p=0.041)\) were associated with short OS, and higher TNM stage \((p<0.001)\) was associated with short RFS. However, low DUSP4 expression was not statistically significant (Table IV). Kaplan-Meier survival analyses revealed that low DUSP4 expression was significantly associated with short OS and RFS (log-rank test, \(p=0.027\) and \(p=0.003\), respectively; Figure 3). Similar to our results, the

### Table III. Correlation between DUSP4 expression and clinicopathological features and molecular characteristics \((n=508)\).

| Variables                        | Low expression (%) | High expression (%) | \(p\)-Value |
|----------------------------------|--------------------|---------------------|-------------|
| Age                              |                    |                     | 0.984       |
| <65 years                        | 140 (45.9%)        | 165 (54.1%)         |             |
| ≥65 years                        | 93 (45.8%)         | 110 (54.2%)         |             |
| Gender                           |                    |                     | 0.284       |
| Female                           | 66 (42.3%)         | 90 (57.7%)          |             |
| Male                             | 167 (47.4%)        | 185 (52.6%)         |             |
| Tumor size                       |                    |                     | <0.001      |
| <4.0 cm                          | 98 (35.0%)         | 182 (65.0%)         |             |
| ≥4.0 cm                          | 135 (59.2%)        | 93 (40.8%)          |             |
| Location                         |                    |                     | 0.575       |
| Proximal                         | 80 (44.2%)         | 101 (55.8%)         |             |
| Distal                           | 153 (46.8%)        | 174 (53.2%)         |             |
| Histologic type*                 |                    |                     | 0.252       |
| Differentiated and others        | 85 (42.7%)         | 114 (57.3%)         |             |
| Lauren                           | 148 (47.9%)        | 161 (52.1%)         |             |
| Intestinal                       | 87 (41.0%)         | 125 (59.0%)         | 0.065       |
| Diffuse and mixed                | 146 (49.3%)        | 150 (50.7%)         |             |
| pT category                      |                    |                     | <0.001      |
| pT1 and pT2                      | 108 (34.3%)        | 207 (65.7%)         |             |
| pT3 and pT4                      | 125 (64.8%)        | 68 (35.2%)          |             |
| Nodal status                     |                    |                     | <0.001      |
| Negative                         | 115 (38.5%)        | 184 (61.5%)         |             |
| Positive                         | 118 (56.5%)        | 91 (43.5%)          |             |
| Stage                            |                    |                     | <0.001      |
| I                                | 93 (33.1%)         | 188 (66.9%)         |             |
| II and III                       | 140 (61.7%)        | 87 (38.3%)          |             |
| Lymphovascular invasion          |                    |                     | <0.001      |
| Absent                           | 94 (37.0%)         | 160 (63.0%)         |             |
| Present                          | 139 (54.7%)        | 115 (45.3%)         |             |
| Perineural invasion              |                    |                     | <0.001      |
| Absent                           | 110 (34.9%)        | 205 (65.1%)         |             |
| Present                          | 123 (63.7%)        | 70 (36.3%)          |             |
| EBV status                       |                    |                     |             |
| Negative                         | 213 (44.9%)        | 261 (55.1%)         | 0.116       |
| Positive                         | 20 (58.8%)         | 14 (41.2%)          |             |
| MSI status                       |                    |                     |             |
| MSS                              | 219 (53.2%)        | 249 (46.8%)         | 0.151       |
| MSI                              | 14 (35.0%)         | 26 (65.0%)          |             |
| HER2 status                      |                    |                     |             |
| No amplification                 | 220 (45.5%)        | 264 (54.5%)         | 0.403       |
| Amplification                    | 13 (54.2%)         | 11 (45.8%)          |             |

*Differentiated: Well-differentiated tubular adenocarcinoma, moderately differentiated tubular adenocarcinoma, papillary adenocarcinoma; Undifferentiated: poorly differentiated tubular adenocarcinoma, poorly cohesive carcinoma; Others: mucinous adenocarcinoma, other histologic subtypes; Stage, AJCC 8th edition. EBV, Epstein-Barr virus; MSS, microsatellite stable; MSI, microsatellite instability; HER2, human epidermal growth factor receptor-2.
analysis using the KM plotter demonstrated that low mRNA expression of DUSP4 was associated with unfavorable survival outcomes (log-rank test, \(p<0.001\); Figure 4).

**Discussion**

In the present study, immunohistochemical staining for DUSP4 was performed in normal gastric mucosa, tubular adenoma, gastric cancer, and LN tissue with gastric cancer metastasis. Our results demonstrated that the expression of DUSP4 was significantly lower in gastric cancer and metastatic tumors than that in normal gastric mucosa and tubular adenoma. Low expression of DUSP4 was significantly associated with larger tumor size, higher \(pT\) stage, nodal metastasis, lymphovascular invasion, and perineural invasion, which suggests that DUSP4 down-regulation is associated with gastric cancer progression. Similar to the KM plotter analysis results, Kaplan-Meier survival analyses revealed that low DUSP4 expression was significantly associated with overall survival and recurrence-free survival.
DUSPs are a subclass of protein tyrosine phosphatases that dephosphorylate threonine and tyrosine residues on MAPKs and consequently act as negative regulators of MAPK signaling. DUSP4 is included in the first subfamily together with DUSP1, DUSP2, and DUSP5. It is mainly located in the nucleus and is known to show substrate specificity for MAP kinases p38, c-Jun N-terminal kinase (JNK), and extracellular-signal-regulated kinase (ERK) (20). Promoter hypermethylation and copy loss are representative molecular alterations affecting DUSP4 expression in cancer. Waha et al. demonstrated de novo methylation and transcriptional silencing of DUSP4/MKP-2 in primary glioma tissues and glioma cell lines (21) and Schmid et al. reported that DUSP4 promoter hypermethylation repressed DUSP4 expression in diffuse large B-cell lymphoma (9). Chitale et al. and Mazumdar et al. revealed that copy loss of DUSP4 at chromosome 8p was found in EGFR-mutant lung cancers and estrogen receptor (ER)-negative breast cancers (22, 23). Several studies have shown that the function, whether a tumor suppressor or tumor promoter, varies depending on the type of cancer, and the exact impact of DUSP4 expression on cancer progression and prognosis is still unclear.

Recently, Baglia et al. and Gaggianesi et al. reported that low expression of DUSP4 is generally considered to be associated with a poor prognosis or aggressive features in breast cancer (12, 24). Mazumdar et al. demonstrated that the overexpression of DUSP4 suppressed the MAPK, NF-kB, and Rb signaling pathways and inhibited cell growth (23). Balko et al. showed that low DUSP4 expression was correlated with high ERK activity and basal-like subtype. DUSP4 promoter methylation is most common in basal-like subtype among the molecular subtypes of breast cancer (25). Ichimanda et al. reported that colorectal cancer tissue showed a higher expression of DUSP4 than that in normal tissue. However, compared to the superficial region, DUSP4 expression decreased in the deep region of the tumor, and the activity of ERKs increased (26). Saigusa et al. demonstrated that decreased DUSP4 expression in colorectal cancer was significantly correlated with tumor progression and distant metastasis (10). Chitale et al. showed that 8p loss was observed more frequently in EGFR mutant adenocarcinoma of the lung, and re-expression of DUSP4 reduced growth and knockdown of DUSP4 leads to enhanced growth (22). They suggested that DUSP4 might act as a tumor suppressor in human malignancy.

Some authors have reported that DUSP4 in tumor carcinogenesis may play a role as a tumor promoter. Briston et al. reported that ERK1/2 activity was increased when DUSP4 protein was highly expressed and that depletion of DUSP4 decreased ERK activity in EGFR-mutant lung cancer cells (5). Lee et al. demonstrated that DUSP4 expression was increased in thyroid cancer cell lines (TPC1, WRO82-1, and XTC) (6). Ma et al. showed that high DUSP4 expression was associated with aggressive features such as lymph node metastasis and extrathyroidal extension in papillary thyroid carcinoma (7). Kim et al. reported that high DUSP4 expression was associated with poor prognosis in early T-
stage breast cancer (13). Gröschl et al. showed that DUSP4 is associated with microsatellite instability (MSI) in colorectal cancer and DUSP4 overexpression increased cell proliferation in colorectal cancer cell lines (27). They suggested that DUSP4 might act as a tumor promoter in human malignancy.

In gastric cancer, there is no conclusive report regarding the exact role of DUSP4 expression in tumor progression and prognosis. Kang et al. reported that DUSP4 overexpression increased doxorubicin resistance in gastric cancer cells and the knockdown of DUSP4 increased the cytotoxicity of gastric cancer cells to doxorubicin. Up-regulation of DUSP4 promotes the epithelial-mesenchymal transition (14). Xu et al. revealed that the knockdown of DUSP4 reduced the mobility and invasiveness of gastric cancer cells, and DUSP4 mRNA expression was increased in gastric carcinoma compared to that in the adjacent tissues. DUSP4 expression showed a significant increase in 30 gastric cancer tissues detected by immunohistochemistry and western blotting (15). Zhang et al. demonstrated that low DUSP4 expression was significantly associated with various factors related to cancer progression, such as larger tumor size, deep invasion of the tumor, and distant metastasis in a study of 89 patients with gastric cancer (16). In our study, low DUSP4 expression was significantly correlated with aggressive phenotypes and poor prognosis in a large series of 508 patients with gastric cancer. The association between DUSP4 expression and specific molecular subtypes was investigated, however, there was no significant correlation between DUSP4 expression and specific molecular subtypes.

As a strength of this study, it was possible to evaluate the clinicopathological significance of DUSP4 expression in gastric cancer through larger number of cases than previous studies, and similar results were confirmed using bioinformatics analysis. Also, this is the first study to investigate the association between the expression of DUSP4 and a specific molecular subtype of gastric cancer. However, there is a limitation in that we only evaluated the expression of DUSP4 through immunohistochemistry. The molecular mechanism underlying the role of DUSP4 expression in gastric cancer was not studied, and further studies such as DUSP4 promoter hypermethylation are needed.

Figure 4. Analysis using the Kaplan-Meier (KM) plotter demonstrated that low mRNA expression of DUSP4 was associated with unfavorable survival outcome (n=875; Log-rank test, p<0.001).
In conclusion, our results showed that DUSP4 expression in gastric cancer and lymph node metastasis was reduced compared to that in normal tissue and tubular adenoma. Low DUSP4 expression is associated with cancer progression and poor prognosis in patients with gastric cancer. Our results suggest that DUSP4 may act as a tumor suppressor in gastric cancer. The exact role of DUSP4 in gastric cancer and its potential as a novel therapeutic target for gastric cancer should be investigated in further studies.

Compliance with Ethical Standards

The Institutional Review Board of Hanyang University Hospital (HYUH 2019-04-032-001) approved the study and waived the requirement for informed consent.

Conflicts of Interest

The Authors declare that no potential conflicts of interest exist relevant to this article.

Authors’ Contributions

Seongsik Bang: tissue microarray construction; assessment of the immunohistochemical slides, review of histological material, data analysis, drafting of the manuscript, preparation of the illustrations, preparation of the final text. Seungyun Jee: review of histological material; Hyunsung Kim: review of histological material; Kiseok Jang: revision of the manuscript; Hosub Park: revision of the manuscript; Jaekyung Myung: revision of the manuscript; Dongho Choi: provided clinical data and antibody; Su-Jin Shin: review of immunohistochemical slides, review of histological material, data analysis, drafting of the manuscript, preparation of the final text. Seung Sam Paik: elaboration of the research concept, general supervision of the study, revision of the manuscript and preparation of the final text.

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