Septic shock secondary to an acute necrotizing community-acquired pneumonia with bacteremia due to Pseudomonas aeruginosa

P. Riviere*, D. Patin, E. Delaporte, H. Mahfoudi, S. Lecaitel, F. Poher, P. Villette, J. Ducaux, P. Jouault, G. Brunin

Hôpital Duchenne, Rue Monod, Service de réanimation, 62200 Boulogne-sur-Mer, France

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A B S T R A C T
* Pseudomonas aeruginosa is an uncommon cause of necrotizing acute community-acquired pneumonia (CAP). Only thirteen cases have been previously reported in the literature. In this article, we describe a case of previously healthy 80-year-old male patient, who presented in septic shock caused by necrotizing CAP. Despite inadequate empiric antimicrobial treatment, the patient survived and was able to return to his home after three weeks of hospitalization. To the best of our knowledge, this is the second case of septic shock secondary to P. aeruginosa necrotizing CAP and bacteremia, with optimal clinical outcome. We highlight the evolution of this pathology remains unpredictable, despite the factors related to the host and the bacterium.

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Introduction

Pseudomonas aeruginosa is a cause of nosocomial pneumonia in mechanically ventilated patients or with risk factors. Rarely, it can be responsible for acute community-acquired pneumonia (CAP). The risk factors identified for CAP are structural lung abnormalities, tobacco, alcohol, and contact with P. aeruginosa contaminated water. In this article, we describe a case of an 80-year-old male patient admitted to intensive care unit (ICU), for septic shock due to P. aeruginosa necrotizing CAP and bacteremia.

Case presentation

An 80-year-old male presented to the emergency department at the end of January for three day history of severe right sided chest pain associated with dysnea and fever. He denied any use of alcohol or tobacco. Past medical history consisted of hypertension, hypothyroidism, and calcified pleural plaques of unknown cause: he was a baker and had no past history of asbestos exposure. Six weeks prior to the presentation, he was diagnosed with autoimmune hemolytic anemia (AIHA) during a short hospitalization and was getting treated with oral prednisone 70 mg daily. Recent laboratory work up consisted of negative HIV, hepatitis B and hepatitis C serologies. Antinuclear antibodies were positive with anti double-stranded DNA titer 1:360 but did not meet the criteria for lupus. Plasma protein electrophoresis was normal. A Positron emission tomography (PET) scan and a chest-abdomen-pelvis computed tomography (CT) were normal.

At arrival to the emergency department, the patient was dyspneic with respiratory rate of 27 breaths/min, heart rate of 93 beats/min and fever of 39°C. His lung auscultation was negative for crackles. The patient was hypotensive, with blood pressure of 80/60 mmHg, which did not respond to the initial intravenous fluids, thus required vasopressor support with norepinephrine. He was admitted to the ICU with the diagnosis of sepsis shock. The laboratory tests showed leukocytosis with white blood cell count of 27,100/mm3, hemoglobin level of 14.5 g/dL and platelet count of 187,000/mm3. The arterial blood gas collected on two liters of oxygen showed pH of 7.48, PaO2 of 62 mmHg, and PaCO2 of 35 mmHg. C-reactive protein (CRP) was elevated at 130 mg/L. The patient had acute kidney injury with creatinine at 1.386 mg/dl (baseline: 0.795 mg/dl). Lactic acid was elevated to 31.53 mg/dl. Procalcitonin (PCT) was increased to 14.6 mg/mL. Blood cultures were obtained. Pneumococcal and Legionella urine antigens were negative. The anteroposterior (AP) chest x-ray (Fig. 1A) showed pulmonary consolidation in the right upper lobe (RUL) with a cavity. Chest, abdomen and pelvic CT (Fig. 1B) showed RUL consolidation associated with septated cavitation. Empiric antimicrobial therapy was started with intravenous (IV) cefotaxime 2 g x 3 daily and IV levofloxacin 500 mg twice daily. On the second day of hospitalization, blood cultures became positive for pan-sensitive P. aeruginosa. The antimicrobials were changed to IV cefepime 2 g over eight hours twice daily for three weeks.

* Corresponding author.
E-mail address: pierre.rivierepr@gmail.com (P. Riviere).

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The patient underwent bronchoscopy with bronchoalveolar lavage (BAL) with positive culture of *P. aeruginosa* at the concentration of $10^5$/mL. The respiratory strain susceptibility pattern was identical to the blood culture. The acid-fast bacilli stain was negative. The patient's clinical condition improved on IV cefepime. The vasopressor was rapidly weaned on the second day and oxygen therapy was stopped on the fourth day of hospitalization. He was transferred out of the ICU to the pulmonaryology floor and was later discharged from the hospital after 3 weeks of IV cefepime. Repeat chest x-ray at 2 weeks (Fig. 1C) showed dramatic decrease of the pulmonary consolidation with residual 10 cm cavity.

Discussion

Necrotizing CAP due to *P. aeruginosa* is rare. In the literature, we found 13 cases of necrotizing CAP due to *P. aeruginosa* associated with cavitation. Table 1 summarizes the 11 most recent cases [1–10]. The main risk factors for necrotizing CAP due to *P. aeruginosa* are: lung structural abnormalities (including cystic fibrosis, emphysema, bronchial dilatation, chronic obstructive pulmonary disease), immunocompromised condition (including neoplasia, neutropenia, prolonged antibacterial therapy, immunosuppression), and exposure to contaminated liquids (including hot tub, spa, humidifier) [1,2]. This patient had none of these risk factors prior to the hospitalization during he was diagnosed AIHA, however he had been treated with prednisone 70 mg daily for six weeks prior to presentation with necrotizing CAP. The PET scan and the chest-abdomen-pelvis CT during his hospitalization on hematology floor did not reflect abnormality except pleural plaques.

The literature review did not show any specific radiologic symptomatology of necrotizing CAP due to *P. aeruginosa*. However Hatchette et al. [11] described two-thirds of this pneumonia occurs in the right upper lobe, which was the case for our patient. Fetzer et al. reviewed a series of 7 autopsies and described two presentations of *P. aeruginosa* CAP: the presence of diffuse subpleural hemorrhagic nodular lesions and occasional necrotic parenchyma; or the formation of a 5 cm necrotic lung lesion within the lobular boundaries that included multiple necrotic nodules surrounded by hemorrhagic parenchyma [12].

Regarding antimicrobial management for CAP, the literature is summarized in Table 1 [1–10]. It is generally not appropriate to administer empiric anti-Pseudomonas therapy. However, based on the severity of the clinical presentation, the need for care in the ICU, along with the presence of *P. aeruginosa* infection risk factors, broader antimicrobial therapy can be considered. In our case, the risk factors were old age and the use of corticosteroids. Our patient was immunocompromised after receiving a cumulative dose of prednisone greater than 600 mg which is associated with a risk of opportunistic infections. Furthermore, *P. aeruginosa* is more likely to colonize recently hospitalized patients, as it was the case for this patient. These additional risk factors may discuss an initial broader antimicrobial therapy. There are only few studies comparing anti-pseudomonal monotherapy versus combination of antibiotics. The current assumption is that two anti-pseudomonal antibiotics are required in case of bacteremia or severe pneumonia due to *Pseudomonas*. Indeed, one report found that the therapeutic...
association was associated with a higher mortality rate than monotherapy for older patients [13]. While in some cases the evolution of the CAP due to P. aeruginosa can be devastating, in the present case, the outcome was favorable despite inadequate initial antimicrobial therapy and IV cefepime monotherapy during 3 weeks. This case shows that anti-pseudomonal monotherapy may be a better option for older patients. Indeed, although the most commonly used antimicrobial is ceftazidime, no study has shown a difference in survival between cefepime and ceftazidime in severe bacterial infections. In addition, it seems that cefepime has a better diffusion into the lung. This is the only the second case of septic shock secondary to P. aeruginosa necrotizing CAP with bacteremia in a patient with favorable outcome [14].

For our patient, host risk factors were old age and the use of corticosteroids. Pathogen related virulence factors in similar cases have not been described in the literature. For our patient pathogen related virulence factors have been investigated: it is a wild P. aeruginosa that does not have virulence factors. The strain does not produce the exotoxin ExoU of the type III secretion system (major factor of virulence in the lung). Detection of Colistin resistance was negative. These elements do not explain the speed of installation or the severity of the infection. We propose that a future research of P. aeruginosa virulence factors leading to cavitary pneumonia could explain the severity and rapid progression of the disease.

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