Elamipretide Improves Mitochondrial Function in the Failing Human Heart

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VISUAL ABSTRACT

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HIGHLIGHTS
- Mitochondrial function is impaired in explanted failing pediatric and adult human hearts.
- Elamipretide is a novel mitochondria-targeted drug that is targeted to cardiolipin on the inner mitochondrial membrane and improves coupling of the electron transport chain.
- Treatment of explanted human hearts with elamipretide improves human cardiac mitochondrial function.
- The study provides novel methods to evaluate the influence of compounds on mitochondria in the human heart and provides proof of principle for the use of elamipretide to improve mitochondrial energetics in failing myocardium due to multiple etiologies and irrespective of age.

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All authors attest they are in compliance with human studies committees and animal welfare regulations of the authors’ institutions and Food and Drug Administration guidelines, including patient consent where appropriate. For more information, visit the JACC: Basic to Translational Science author instructions page.

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SUMMARY

Negative alterations of mitochondria are known to occur in heart failure (HF). This study investigated the novel mitochondrial-targeted therapeutic agent elamipretide on mitochondrial and supercomplex function in failing human hearts ex vivo. Freshly explanted failing and nonfailing ventricular tissue from children and adults was treated with elamipretide. Mitochondrial oxygen flux, complex (C) I and CIV activities, and in-gel activity of supercomplex assembly were measured. Mitochondrial function was impaired in the failing human heart, and mitochondrial oxygen flux, CI and CIV activities, and supercomplex-associated CIV activity significantly improved in response to elamipretide treatment. Elamipretide significantly improved failing human mitochondrial function. (J Am Coll Cardiol Basic Trans Science 2019;4:147-57) © 2019 The Authors. Published by Elsevier on behalf of the American College of Cardiology Foundation. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

MITOCHONDRIAL DYSFUNCTION AND CARDIOLIPIN IN HEART FAILURE

A substantial body of work has implicated abnormal mitochondrial function and consequent impaired energy production via oxidative phosphorylation as a factor in the development of heart failure (HF), leading to heightened interest in mitochondrial function as a therapeutic target (1–3). Efficient mitochondrial function is considered especially essential in the heart due to the sustained energy requirements of the myocardium. Cardiolipin, a unique phospholipid with 4 fatty acid side chains, is critical for maintaining normal adenosine triphosphate generation by anchoring the proteins of the electron transport chain onto the inner mitochondrial membrane architecture (4). In the heart, a mitochondrial inner membrane enriched with tetralinoleoyl cardiolipin (containing 4 linoleic acid side chains) facilitates the stability of the physical interaction between oxidative phosphorylation complex protein multimers in what has been termed the mitochondrial supercomplex, composed of complex (C) I, CIII, and CIV (5,6). Abnormalities in cardiolipin composition lead to mitochondrial dysfunction in the failing heart through disruption of the supercomplex and can be improved with dietary interventions in rodent models (7–9). These abnormalities in mitochondrial function have led to heightened interest in mitochondria as a therapeutic target in HF (3).

Elamipretide, a mitochondrial-targeted peptide

Elamipretide (formerly referred to as Bendavia, MTP-131, and SS-31) is an aromatic-cationic, cell-permeable tetrapeptide in a new class of mitochondrial-targeted drugs and is the first in its class to enter clinical trials in patients with HF (10–12). The peptide is targeted to mitochondria via cardiolipin where it has been shown in animal models to improve energetics and decrease reactive oxygen species, possibly by stabilizing the mitochondrial membrane and cytochrome c (13,14). Elamipretide rapidly enters tissue (within minutes of treatment) and has cardioprotective effects in ischemia/reperfusion injury in animal models (15–18). To date, there are no data on the direct effects of elamipretide, or other members of this novel class, on mitochondrial function in the human heart.

ELAMIPRETIDE IN HUMAN HF

The purpose of the present study was to investigate the effects of elamipretide on human cardiac mitochondrial function. Because this study was ex vivo and performed over the course of a few hours (too rapid to induce cardiolipin remodeling), any effects of the drug would likely be independent of cardiolipin molecular species alterations. Using freshly explanted human hearts, the current study found that: 1) elamipretide improves impaired mitochondrial function in HF, with no effect on normal mitochondrial function in nonfailing hearts; 2) elamipretide improves mitochondrial supercomplex function (CI, CIII, and CIV) but does not alter CII or CV activity; and 3) the short-term action of elamipretide is independent of any changes in cardiolipin side chain composition.

METHODS

CHEMICALS AND REAGENTS. Elamipretide was provided by Stealth BioTherapeutics, Inc. (Newton, Massachusetts), resuspended in water at 10 mM, and stored at −80°C. The elamipretide ex vivo treatment...
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**TABLE 1** Characteristics of Cardiac Samples According to Experiment

|                      | High-Resolution Respirometry | Enzyme Activity Assays | BN-PAGE In-Gel Activity | Cardiolipin Mass Spectrometry |
|----------------------|-----------------------------|------------------------|-------------------------|-------------------------------|
|                      | Nonfailing | Failing | Nonfailing | Failing | Nonfailing | Failing | Nonfailing | Failing | Nonfailing | Failing | Nonfailing | Failing |
| No. of heart samples | 13        | 23      | 11         | 12       | 19         | 21       | 5          | 10      |           |         |           |         |
| Male                 | 6         | 15      | 4          | 7        | 8          | 10       | 3          | 7       |           |         |           |         |
| Female               | 7         | 8       | 7          | 5        | 11         | 11       | 2          | 3       |           |         |           |         |
| Age, yrs             | 25 ± 7    | 31 ± 5  | 20 ± 8     | 29 ± 8   | 26 ± 5     | 25 ± 5   | 28 ± 13    | 29 ± 9  |           |         |           |         |
| Ejection fraction, % | 64 ± 5    | 24 ± 2  | 64 ± 4     | 27 ± 3   | 61 ± 3     | 22 ± 6   | 58 ± 2     | 20 ± 3  |           |         |           |         |

Values are n or mean ± SEM.
BN-PAGE = blue native polyacrylamide gel electrophoresis.

digestion as previously described with the following modifications for human tissue (20,21). Briefly, freshly explanted cardiac tissue in pieces no larger than 5 mm was placed in ice-cold BIOPS solution. Tissue was trimmed of fat, placed into mitochondrial isolation buffer, and minced by using scissors. Low-speed spins were at 1,500g and high-speed spins at 4,200g. One milligram of trypsin was used per gram of tissue. Mitochondria were resuspended in potassium buffer and protein quantified (BCA, Thermo Fisher Scientific, Waltham, Massachusetts).

**ENZYMATIC ACTIVITIES.** Isolated mitochondria were diluted to a final protein concentration of 1 mg/ml and incubated with 100 μM elamipretide or an equal volume of vehicle (water) for 1 h on ice. Enzymatic activities of CI, CIV, and citrate synthase were performed at 30°C as previously described (22,23).

**BLUE NATIVE POLYACRYLAMIDE GEL ELECTROPHORESIS.** Mitochondrial supercomplexes were separated by BN-PAGE using TGX 4-15% gels (Bio-Rad, Hercules, California). In-gel activity assays for CI, CIII, CIV, and CV were performed with sepal tissue processed according to published methods (22,24). Samples were treated with 100 μM elamipretide for 1 h. Supercomplex and free protein bands were quantified by using ImageJ software (National Institutes of Health, Bethesda, Maryland). Total supercomplex activity was calculated as a sum of all supercomplex bands.

**HIGH-RESOLUTION RESPIROMETRY.** Respiration of permeabilized cardiac fibers was measured by high-resolution respirometry (Oxygraph, Oroboros Instruments, Innsbruck, Austria) using a stepwise protocol to evaluate various components of the electron transport system (19,25). Approximately 30 mg of ventricular tissue was placed in BIOPS immediately after explant. One-half of the tissue was incubated in BIOPS containing 100 μM elamipretide at 4°C for 1 h. After incubation, tissue was cut into approximately 2-mg pieces and teased using forceps to separate fibers. The tissue was then placed in a solution of BIOPS containing 30 μg/ml saponin for 30 min to permeabilize
the plasma membrane and to allow substrate delivery to the mitochondria. Fibers were washed for 10 min at 4°C in ice-cold mitochondrial respiration medium containing 25 μM blebbistatin ± 100 μM elamipretide (26). Samples were blotted on filter paper, weighed, and placed in the chambers of the Oroboros O2K apparatus at 37°C containing respiration medium and 25 μM blebbistatin ± 100 μM elamipretide. Standard protocols were followed for calibration of the chambers and followed by stepwise addition of 5 mM pyruvate, 1 mM malate, 4 mM adenosine diphosphate (ADP), 10 mM glutamate, 10 mM succinate, 10 μM cytochrome c, and either 2 μg/ml oligomycin or 0.5 μM steps of carbonyl cyanide p-trifluoromethoxyphenylhydrazone (FCCP) (until the maximum rate was reached) followed by 2 μM rotenone. Oxygen flux rates were normalized per milligram of tissue wet weight.

**CARDIOLIPIN QUANTITATION.** Cardiolipin was quantified in a subset of the total cohort by using previously published methods with liquid chromatography coupled to electrospray ionization mass spectrometry in an API 4000 Mass Spectrometer (SCIEX, Framingham, Massachusetts) (20). After BIOPS or BIOPS Plus 100 μM elamipretide treatment for 4 h, heart tissue was frozen without buffer at −80°C. Tissue pieces were homogenized by using a glass-on-glass homogenizer in phosphate-buffered saline and lipids extracted according to previously published methods with 1 mmol tetramyristoylcardiolipin as an internal standard (Avanti Polar Lipids, Alabaster, Alabama) (20,27). Cardiolipin species were quantified per milligram of protein.

**STATISTICAL ANALYSIS.** Statistical analyses were performed by using Prism software version 6.0.
Treatment effects were analyzed by using a ratio-paired Student’s t-test, with \( p < 0.05 \) reported as trending toward significance (28). For time-based effects were analyzed by using an unpaired Student’s t-test. Datasets were tested for Gaussian distribution with the D’Agostino and Pearson omnibus test or the Shapiro-Wilk normality test. Data that did not conform to a Gaussian distribution (glutamate and supercomplex coupling control factor data) were log-transformed before analysis.

Graphs with bars show SEM for unpaired Student’s t-test data, and graphs with paired data show means plus SDs (chosen for clarity) flanked on either side of the paired data. Linear regression analysis was performed to assess for an association between age and significant outcomes (no associations were demonstrated).

**RESULTS**

**ELAMIPRETIDE IMPROVES RESPIRATION OF INTACT MITOCHONDRIA FROM THE FAILING HUMAN HEART.** Quantitation of the results of high-resolution respirometry in all samples is shown in Figure 1 with sample traces shown in Supplemental Figure 2. There was significantly lower mitochondrial normalized oxygen flux in HF vs nonfailing samples using pyruvate malate + ADP, glutamate (CI), and succinate (CI + CII) as substrates (Figures 1A to 1C). There were also significantly lower oxygen fluxes in HF with inhibition of flux through CI + CII using oligomycin or the uncoupler FCCP \( (p < 0.01) \) but not combined with the CI inhibitor rotenone (data not shown).

Elamipretide treatment of nonfailing permeabilized fibers did not alter mitochondrial oxygen flux (Figures 1D to 1F). However, in HF, there was a significant increase in oxygen flux with elamipretide treatment using the substrates pyruvate malate + ADP (Figure 1G) and glutamate (CI) (Figure 1H). Notably, there was no significant difference with elamipretide treatment with the addition of succinate (CI + CII) in HF (Figure 1I). Addition of pyruvate/malate, FCCP, rotenone, or oligomycin did not significantly affect oxygen flux through CI + II (data not shown).

**ELAMIPRETIDE INCREASES THE SUPERCOMPLEX COUPLING CONTROL FACTOR AND RESPIRATORY CONTROL RATIO IN FAILING MITOCHONDRIA.** The supercomplex coupling control factor ratio (SC CCF) is
a mathematical construct to calculate the influence of the supercomplex on a given respirometry measurement and is directly proportional to supercomplex integrity (derivation is given in the Supplemental Methods). The SC CCF was significantly lower with HF (Figure 2A). Elamipretide treatment in nonfailing hearts did not alter SC CCF (Figure 2B). Conversely, in HF, elamipretide treatment significantly increased SC CCF (Figure 2C). The respiratory control ratio (RCR) indicates a trend toward decreasing in HF (Figure 2D). There was no change in RCR with elamipretide treatment in nonfailing hearts (Figure 2E), whereas in HF, RCR was significantly increased with elamipretide treatment (Figure 2F).

**Activities of Mitochondrial CI and CIV in Subsarcolemmal Mitochondria Are Increased with Elamipretide.** Freshly isolated subsarcolemmal and interfibrillar mitochondria treated with elamipretide were used to measure total activity of CI and CIV as well as citrate synthase, a soluble enzyme in the mitochondrial matrix (29). Total CI activity was significantly lower in HF compared with nonfailing samples (Figure 3A), whereas CIV activity in subsarcolemmal mitochondria showed a trend toward a decrease in HF (Figure 3B). No difference in citrate synthase activity was observed between HF and the nonfailing group (Figure 3C). In nonfailing subsarcolemmal mitochondria, activities of these 3 enzyme complexes were unaffected by elamipretide treatment (Figures 3D to 3F). However, in failing subsarcolemmal mitochondria, both CI and CIV activities exhibited a trend for increase by elamipretide treatment (Figures 3G and 3H), while citrate synthase activity was unchanged (Figure 3I). None of the enzyme activities was altered in the elamipretide-treated interfibrillar mitochondria (data not shown).

**Supercomplex-Associated CIV Activity Is Altered with Elamipretide.** BN PAGE in-gel
activity was used to separate supercomplex from free CIV. Figure 4A shows representative gels for CI, CII, CIV, and CV activities. CIV activity is present in a free state, within the supercomplex, and in intermediate molecular weight forms not analyzed in this study. CI is only present in its supercomplex form. When these bands were quantified, the quantity of CIV in the supercomplex was not altered in HF (Figure 4B), whereas the free CIV activity was significantly lower (Figure 4C). In nonfailing samples, elamipretide treatment had no effect on CIV regardless of whether it is bound or free (Figures 4D and 4E). By contrast, in HF, the activity of CIV in the supercomplex was significantly higher after elamipretide treatment, whereas activity of free CIV was unchanged (Figures 4F and 4G). There were no significant changes detected in the activities of CI, CII, or CV in HF or with elamipretide treatment according to this assay (data not shown).

**CARDIOLIPIN COMPOSITION IS NOT ALTERED WITH ELAMIPRETIDE TREATMENT.** Tetralinoleoyl cardiolipin, the predominant cardiac cardiolipin species, with 4 linoleic acids and a mass-to-charge ratio of 1448, is shown as a percentage of the total (11 major species were used for total) cardiolipin content. The percentage of tetralinoleoyl cardiolipin was significantly lower in HF, consistent with our previously reported findings (Figure 5A), but elamipretide treatment had no effect on tetralinoleoyl cardiolipin in nonfailing or HF samples (Figures 5B and 5C) (7,30). Cardiolipin absolute amounts, percent totals of 8 major cardiolipin species, monolysocardiolipin, or the sum of all cardiolipin species were unaltered after elamipretide treatment (data not shown).
DISCUSSION

OVERVIEW OF NOVEL FINDINGS. To the best of our knowledge, this study is the first to investigate the effect of elamipretide on mitochondrial respiration in explanted human heart tissue. Elamipretide treatment improves left ventricular volumes in adults with HF (12), suggesting a potentially new avenue for therapeutic agents in HF (31). We extended these clinical trial data by showing that impaired mitochondrial function in the failing pediatric and adult human heart can be improved with elamipretide treatment. Elamipretide treatment improved the function of components of the electron transport chain when they were associated with the supercomplex in human cardiac mitochondria from the failing ventricle. Finally, acute elamipretide treatment of human cardiac tissue improved mitochondrial and supercomplex function even though the treatment was too rapid to allow remodeling of cardiolipin. This comprehensive analysis in human heart tissue provides additional insight into the mechanism by which elamipretide treatment improves the function of the respiratory chain in human cardiac mitochondria. These findings support the use of supercomplex stabilizing compounds for the treatment of human disease (16,32).

ELAMIPRETIDE IN HUMAN DISEASE. Clinical trials have used elamipretide to address renal, cardiac, and skeletal muscle disease, but this study is the first to show reversal of mitochondrial dysfunction in HF. Elamipretide improved 6-min walk test distances in patients with mitochondrial myopathy (33) and improved left ventricular volumes in patients with HF and left ventricular dysfunction (12). Similarly, elamipretide was associated with improved kidney function after renal angioplasty (34). To our knowledge, there is only 1 other study exploring the influence of elamipretide directly on human mitochondria. Wijermars et al. (35) examined the influence of SS-31 on mitochondrial function in normal renal biopsy samples subjected to ischemic stress. An important difference between their protocol and ours was that the biopsy specimens were pretreated with the compound during the ischemic stress followed by a demonstration of attenuation of mitochondrial dysfunction in the ex vivo study. Similar to our findings in the heart, there was no effect of elamipretide on mitochondrial function in the control renal biopsy specimen. Importantly, our study is the first to report the ability to improve mitochondrial dysfunction in human disease with acute treatment with elamipretide.

ELAMIPRETIDE’S MECHANISM OF ACTION WORKS THROUGH THE ENZYMES OF THE MITOCHONDRIAL SUPERCOMPLEX. Results of high-resolution respirometry showed that elamipretide improves human cardiac mitochondrial function through improved coupling of the supercomplex-associated enzyme complexes CI, CIII, and CIV. Oxygen flux after the addition of supercomplex-specific substrates, pyruvate malate, ADP, and glutamate was impaired in the failing heart. Elamipretide improved oxygen flux in failing samples under these conditions, indicating direct activity on the supercomplex. Succinate, the next substrate added in our experimental protocol, forces the majority of electron flow through CII, a component of the respiratory chain not associated with the supercomplex (36,37). CII-mediated oxygen flux was not influenced by HF, nor was it altered by

FIGURE 5  Tetralinoleoyl Cardiolipin With Elamipretide Treatment

(A) The effect of HF on tetralinoleoyl cardiolipin (mass-to-charge ratio, 1,448) in untreated samples is expressed as a percentage of total cardiolipin. Elamipretide-treated (B) NF hearts or (C) HF hearts showed no changes in tetralinoleoyl cardiolipin. Untreated hearts in B and C are shown with closed circles and treated hearts with open circles. NF, n = 5; HF, n = 10. Abbreviations as in Figure 1.
Elamipretide treatment. Furthermore, addition of rotenone, which inhibits CI and blocks all supercomplex-mediated respiration, permitted CI-mediated oxygen flux that cannot be augmented by elamipretide treatment of tissues (Supplemental Figure 3). These data show that the effect of elamipretide treatment occurs via its effect on CI/CIII/CIV supercomplex activity.

In the complementary spectrophotometry studies, total mitochondrial enzymatic activities of complexes CI and CIV were increased with elamipretide in the failing heart, with no effect on citrate synthase, a soluble enzyme not in the membrane that serves as a marker of mitochondrial content and nonmembrane-associated mitochondrial activity. Enzyme activities of CI and CIV assayed by using BN-PAGE analysis revealed increased supercomplex-associated CIV activity but not CI activity with elamipretide treatment. This result may indicate a stronger effect of elamipretide on recruitment of CIV to the supercomplex. Alternatively, BN-PAGE may not have the sensitivity to detect improvement in CI activity recorded with the enzymatic assay. Taken together, these data suggest that elamipretide improves mitochondrial function by directly affecting respiratory complexes specifically associated with the mitochondrial supercomplex, perhaps by recruiting free CIV into the supercomplex.

**Improvement of Mitochondrial Function by Elamipretide is Independent of Cardiolipin Remodeling.** Elamipretide reportedly rapidly targets mitochondria through the attraction between the positively charged residues of the peptide with the negatively charged cardiolipin head group, which is believed to improve phospholipid-dependent bioenergetics (13). Indeed, long-term treatment with elamipretide in animal models has been associated with increased mitochondrial cardiolipin remodeling to its tetralinoleoyl form (16,38). To date, however, it is unclear if cardiolipin remodeling after chronic treatment is a cause or a consequence of elamipretide’s mechanism of action. We would not anticipate that elamipretide could acutely alter cardiolipin isoforms in an ex vivo system, and in fact we report here that drug treatment can improve mitochondrial function in the absence of any change in cardiolipin species.

We instead hypothesize that elamipretide may elicit its acute effects by improving the stabilization of cardiolipin-protein interactions, allowing the maintenance of the mitochondrial supercomplex.

**Improvement of Mitochondrial Function is Independent of Age and HF Etiology.** There is a growing body of literature characterizing the unique adaptations that exist between the failing pediatric heart and the adult heart (39-45). These studies, combined with the differential response of children to adult-based HF therapies and limited improvement in pediatric HF outcomes over the past decade, suggest that investigations of pediatric-specific HF treatments are needed (46-48). Importantly, the present study included both pediatric and adult failing hearts and showed that elamipretide improves mitochondrial function across a number of different pathologic phenotypes on current evidence-based therapy. This finding is unique and provides encouraging evidence that the mitochondria may represent one final common pathway in HF and that mitochondrial-targeted therapies may have more generalizable efficacy complementary to existing HF therapies.

**Study Limitations.** First, this study was unable to elucidate the mechanisms underlying chronic exposure to elamipretide. Although this study concluded that elamipretide does have a positive effect on mitochondrial function via the enzymes of the supercomplex, and this mechanism does not involve changes in quantity or quality of cardiolipin, the exposure time to elamipretide was limited to a few hours. Future studies are needed to determine the influence of chronic treatment on human mitochondrial function. Second, this study was performed with all hearts that were sequentially collected by our tissue bank over the course of several years. Additional research in larger populations will be necessary to determine if specific disease phenotypes derive greater benefit than others. Third, systemic administration of elamipretide could affect all other mitochondrial-containing tissues and could cause off-target effects not measured by these assays. Fourth, because the experiments were performed in heart tissue in isolation, concentrations of elamipretide in the ex vivo experiments cannot be extrapolated to in vivo treatment. Lastly, because elamipretide binds to the mitochondrial membrane, other membrane-dependent processes such as cross-talk with the endoplasmic reticulum and calcium signaling could be involved; however, this study was limited in scope to the effect of elamipretide on the supercomplex and electron transport system. Any additional effects of elamipretide on human mitochondrial calcium transport and endoplasmic reticulum-mitochondria interactions would need to be analyzed in future studies.

**Conclusions**

A new class of mitochondrial-targeted drugs, which includes elamipretide, represents a promising new strategy for the treatment of HF (49). A central
problem in HF is the inability of the heart to adequately meet its own metabolic demands due to altered mitochondrial function and metabolism. These drugs offer targeted benefits through improvement of mitochondrial function and energy production. The present study is the first to assess the impact of a member of this new class of compounds directly on human heart tissue. The findings of the current study are the first to report rapid improvement in mitochondrial function in the human heart, likely through improved coupling of the mitochondrial supercomplex. These data support the use of mitochondrial-targeted pharmaceutical agents to improve energetics and mitochondrial function in the failing heart.

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APPENDIX For supplemental tables and figures, please see the online version of this paper.