Effect of oral administration of GABA on thermoregulation in athletes during exercise in cold environments: A preliminary study

Hongli Wang, Lin Cheng and Yanbai Han*

College of Physical Education and Health, Guangxi Normal University, Guilin, China

**Background:** γ-aminobutyric acid (GABA), a common ingredient in sports supplements and other health products, regulates body temperature in the preoptic area and anterior hypothalamus (PO/AH). To date, no study has examined the effect of GABA on thermoregulation during exercise in humans in a cold temperature environment (11 ± 0.3°C, 45% ± 2% relative humidity).

**Methods:** We performed a randomized, double-blind study. Ten trained male athletes consumed either a drink (3 ml/kg weight) containing GABA (1,000 mg, trial G) or an equivalent amount of placebo drink (trial C) before exercise. They rested for 20 min and then cycled at 60% of maximum output power for 40 min, pedaling at 60 rpm, and recovered for 20 min. Core temperature ($T_c$), skin temperature (upper arm, chest, thigh, calf), and heart rate (HR) were monitored at rest ($T_0$), exercise begins ($T_{20}$), 20 min of exercise ($T_{40}$), the exercise ends ($T_{60}$), and at recovery ($T_{90}$).

**Results:** Compared to $T_0$, $T_c$ decreased significantly at $T_{20}$ and increased significantly at $T_{40}$, $T_{60}$ and $T_{90}$ ($p < 0.01$). From 35–80 min, the $T_c$ was higher in trial G (peaked at 37.96 ± 0.25°C) than in trial C (37.89 ± 0.37°C), but it failed to reach significant difference ($p > 0.05$); $T_{sk}$ continued to increase during exercise and was significantly higher than $T_0$ at $T_{40}$ ($p < 0.05$), $T_{60}$ and $T_{90}$ ($p < 0.01$). There was no significant difference in $T_{sk}$ between the two trials ($p > 0.05$).

**Conclusion:** Our findings provide initial evidence that oral administration of GABA does not affect thermoregulation and has no adverse effects on the body as an ergogenic exercise supplement during exercise in cold environments.

**KEYWORDS**
γ-aminobutyric acid, nutritional supplements, thermoregulation, exercise, cold environment

**Introduction**

Major sporting events such as the Winter Olympic Games and Winter Paralympic Games are held during the cold winter season, with athletes inevitably training and competing in cold conditions. For many athletes, participation in sport (e.g., skating, skiing, and running) in cold environments is part of daily life. Studies have shown that ambient temperatures of 10–16°C can induce physiological reactions such as...
peripheral vascular contraction and peripheral blood flow reduction to maintain the body heat balance (1). In a cold environment, the body’s energy expenditure increases, resulting in excitation of the sympathetic nervous system and higher catecholamine concentrations. These changes cause vasoconstriction of the terminal blood and skin vessels, increased heart rate (HR) and cardiac output, decreased skin and limb sensitivity, reduced skeletal muscle contractility and coordination, and reduced joint mobility. Prolonged exercise (1–4 h) in cold weather may result in excessive heat loss from the body, affecting thermoregulation and resulting in hypothermia. Individuals may be at risk for thermoregulatory fatigue or even loss of life (1–4).

Human body temperature is comprised of core and shell temperatures. Core temperature \( T_c \) is the temperature of the body’s internal organs (e.g., heart, lungs, brain, and abdominal viscera), usually measured during exercise by rectal, esophageal, and gastrointestinal temperatures. Shell temperature is defined as the temperature of the body’s peripheral tissues (skin, subcutaneous tissues, and muscles). Skin temperature, in particular, is influenced by the thickness of the skin and subcutaneous adipose tissue. It is also affected by the skin blood flow and ambient temperature and humidity (5). The preoptic area and anterior hypothalamus (PO/AH) is the thermoregulatory center of the body. Its role is to maintain a normal and relatively stable body temperature, ensuring a dynamic balance between heat production and loss (6).

\( \gamma \)-aminobutyric acid (GABA), with the molecular formula \( \text{C}_4\text{H}_7\text{NO}_2 \), is a predominant inhibitory neurotransmitter in the hypothalamus. It was first identified in the brain in 1950, with GABAergic synapses noted as more abundant in the PO/AH than other brain regions (7–9). GABA is also widely found in plants, animals, and microorganisms. In addition to natural sources such as germinated brown rice, soybeans, fruit, and lactic acid bacteria, GABA can also be produced by chemical synthesis, plant enrichment, and microbial fermentation, allowing for better application of GABA in plants and animals. There has been widespread interest in the functional and health effects of GABA because it is a small molecular weight non-protein amino acid proved safe to eat and used in many foods and beverages (including novelty foods) in recent years. Studies have shown that oral administration of GABA reduces neuronal activity, prevents overheating of nerve cells, and has physiological effects such as preventing arteriosclerosis, reducing stress, improving sleep, enhancing exercise-induced muscle hypertrophy, regulating cardiac arrhythmias, lowering blood pressure and blood lipids and enhancing liver function. Moreover, GABA has positive effects in the treatment of psychiatric disorders, epilepsy, convulsions, Huntington’s and Parkinson’s disease (10–15).

Evidence from animal studies suggests a positive relationship between GABA and thermoregulation, and an understanding of the underlying mechanism is gradually emerging. Thermoregulation of freely-moving rats during pharmacological stimulation of GABA in normal (23°C), cold (5°C), and hot (35°C) environments indicated that GABA has a crucial thermoregulatory role in the PO/AH. Body temperature decreased in hot ambient environments through the central administration of GABA in animals, while it rose in cold environments (16). In humans, GABA’s role in decreasing core body temperature at rest and during exercise in hot environments is well established (17). However, no studies have examined the use of GABA as a food supplement in a functional beverage that increases core body temperature during exercise in cold environments, provides effective measures to counteract cold conditions, prevents cold injury, and improves athletic performance during training and competition. Cycling in cold environments (15°C) poses a serious challenge to the human conditioning system compared with temperate environments (18–25°C), and longer exercise duration may result in a greater risk of injury (18–20). When athletes cannot increase heat production to compensate for environment-related heat loss, their endurance and performance will decrease, and the risk of injury will increase. This study investigated whether oral administration of GABA increases core body temperature in athletes in a cold environment. To clarify the effects and underlying mechanisms of GABA, we hypothesized that the systemic administration of GABA to trained athletes during exercise would induce higher core body temperature in a cold environment. To test this, we compared thermoregulatory responses in athletes after oral administration of a GABA-containing sports drink or a placebo during exercise in a cold environment over two trials. If oral administration of GABA leads to elevated human body temperature during exercise in cold environments, evidence-based practical strategies could then be used to resist the negative effects of cold environments. This would have important practical applications for ensuring safe training and improved performance in sports carried out in the cold.

Methods

Participants

Ten healthy male college athletes (mean ± standard deviation, age: 21.4 ± 1.0 years, height: 181.4 ± 5.2 cm, weight: 72.5 ± 5.4 kg, \( \text{VO}_{2\text{max}} \): 63.0 ± 5.7 ml/kg\(^{-1}\)min\(^{-1}\)) volunteered to participate in the study. The participants were middle-long distance runners who had taken part in regular sports training for a minimum of 3 years, training at least 6 h per week. The sample size calculation (effect size = 0.25, alpha level = 0.05, power = 0.80) using G*Power (version 3.1.9.7; Dusseldorf, Germany) indicated that 10 participants were required for this study (21, 22). The investigators enrolled participants after assessing them and based on the following criteria: (1) had no
chronic diseases and cold acclimatization; (2) physical activity for at least 30 min per day, three times per week; (3) no cardiovascular, metabolic, or respiratory disease history; (4) no fever; and (5) no use of any supplements or medications that affect thermoregulation. One week before the study, participants were informed about the procedures and dangers involved and signed informed consent forms. All research protocols followed the principles of the latest declaration of Helsinki and received approval from the ethical committee of Guangxi Normal University.

Research design

The study was conducted using a randomized, double-blind, cross-over design. Each participant underwent a familiarization session and two experimental trials in a randomized order. Participants completed two trials during the randomized second and third visits: a controlled trial (trial C) and a GABA trial (trial G). To ensure drug washout, each condition was separated by at least 4 days (23). Both trials were performed at the same time of day to minimize the influence of circadian rhythms on body temperature and other biological variables. Participants were instructed to follow the same diet on testing days to minimize the effects of nutrition. All participants were asked to abstain from alcohol, caffeine, tea (>48 h), strenuous exercise (>24 h), and eating a standard meal (>2 h) before beginning each experiment. Both trials were conducted in winter (December-January) with a room temperature of 11°C and temperature and humidity controlled by air conditioning and humidification equipment set at 11 ± 0.3°C and 45 ± 2% relative humidity.

Preliminary testing and familiarity

All participants completed body composition (BOD POD Body Composition Tracking System, USA) and height (SYHW-80D stadiometer, China) measurements on arrival at the laboratory for the first time. Before assessing VO_{2max} in the preliminary experiment, participants performed a 5-min warm-up. Maximum oxygen consumption was measured using a graded exercise test on a friction braked bicycle ergometer (Monark889E, Sweden). The test began at 0 kp and then increased load by 0.25 kp every 15 s while maintaining a pedal frequency of 60 rpm until volitional exhaustion. Exhaustion was characterized as the condition when participants rated perceived exertion at 19-20 on the Borg scale, pedal frequency was <60 rpm for more than 5 s despite verbal encouragement, and HR reached a maximum value (220-age). A week before the experiments, participants were familiarized with the equipment and procedures involved in the study. The seat heights, bar heights, and positions on the cycle simulators were adjusted independently for each participant prior to the test, and the settings were recorded so they could be reproduced in each subsequent experimental trial.

Drug treatment

The participants were given GABA (1000 mg) dissolved in 3 ml/kg body weight of sports drink (GABA drink) in trial G or an equivalent amount of sports drink (placebo) in trial C. The GABA (500 mg/tablet, Swanson) doses followed previous research (23–25). Participants were randomly assigned to receive either a placebo or GABA drink on the first visit. The other drink was given at a second visit. The temperature of both placebo and GABA drinks was 8°C at the time of drinking. The sports drinks (MI zone beverage, Danone, China) contained 88 KJ of energy, 4.8 g of carbohydrates, and 0 g of fat and protein.

Exercise protocol

The participants performed at 60% of the maximum load for 40 min at a speed of 60 rpm (23). The mean load of the participants was 2.0 ± 0.3 kp.

Procedures

At 13:00 on the day of the experiment, participants arrived at the laboratory in a fed state (at least 2 h after their last meal). On arrival, their post-void nude body mass was recorded. An HR telemetry band (Polar Beat, Kempele, Finland) was fitted to the participant’s chest. Temperature sensors were attached to four places on the skin surface (chest, upper arm, thigh, and leg) on the left side of the body to calculate the weighted mean skin temperature. Afterward, participants inserted a rectal thermistor 10 cm from the anal sphincter to measure T_r. Then, at 13:55, after the participants were quietly seated on the ergometer, temperature sensors were connected to the data logger. Participants were then given equal volumes of either a placebo (trial C) or a GABA beverage (trial G). At 14:00, the experiment started, during which the temperature measurement software was used to continuously monitor the rectal temperature, skin temperature, and HR. It has been reported that plasma GABA concentrations decrease after oral administration of GABA, peak 20-40 min after ingestion, and remain significantly higher than baseline for at least an hour after ingestion (17). After a 20-min rest, at 14:20, participants performed continuous exercises at 60% of maximum output with a pedaling frequency of 60 rpm. At 14:40, participants were offered a sports drink to prevent dehydration (3 ml/kg body weight) while they remained seated on the ergometer. At 15:00, the exercise is completed, followed by a 20-min recovery period. The final body mass measurement was taken immediately after the exercise. The total time included 20 min rest plus 40 min exercise for the measurement plus 20 min resting time, giving a total of 80 min for the measurements. Each participant spent a
total time of 140 min in the lab. The experimental protocol for this study is shown in Figure 1.

Measurements

The main outcome measures of thermoregulation were core body temperature ($T_c$), chest skin temperature ($T_{chest}$), upper arm skin temperature ($T_{arm}$), thigh skin temperature ($T_{thigh}$), calf skin temperature ($T_{calf}$), and HR. $T_c$ (YSI401AC, YSI Precision, USA) and skin temperature (YSI409AC, YSI Precision, USA) probes were connected to a data logging device, and data were recorded every minute (Squirrel Meter Logger, Grant Instruments, Cambridge, UK). Skin thermistors were attached to four sites on the left side of the body using transparent Tegaderm patches surgical tape (3M, UK), shown in Figure 2. The same investigator applied probes to the skin to ensure consistent pressure was applied to each spot and each person. Pen markings provided reference points for each location. The mean skin temperature ($T_{sk}$) was calculated using Ramanathan’s proposed formula: $T_{sk} = 0.3 (T_{chest} + T_{arm}) + 0.2 (T_{thigh} + T_{calf})$ (26). In addition, HR was measured in minute intervals throughout the experiment using polar telemetry (Polar Electro Oy, Kempele, Finland), and environmental conditions were recorded with a thermometer (JXBS-3001-TH-5-U, China). Estimated total sweating volume was derived by measuring the change in dry body mass (BM), measured immediately before and after the trial as $\text{BM before} - \text{BM after} + 3 \text{ ml/kg (weight of sports drink)}$. Dehydration was defined as a body mass loss of 2% or more (27).

Statistical analysis

The data were analyzed using SPSS version 23.0 (Chicago, IL, USA), and the values were expressed as mean ± standard deviation. First, each outcome variable was examined for normality and sphericity. The one-sample Kolmogorov-Smirnov test for normal distribution was used, and log transformation was used for statistical analysis when the data were not normally distributed. When the assumption of sphericity was violated, the Greenhouse-Geisser correction was applied to reduce the probability of a type I error. We used a 2 [GABA-containing drink and placebo sport drink conditions] × 5 (rest ($T_0$), exercise onset ($T_{20}$), exercise ($T_{40}$) 20 min, the exercise ends ($T_{60}$), and recovery ($T_{80}$)] analysis of variance (ANOVA) with repeated measures to assess the changes in mean HR, $T_c$, and $T_{sk}$ values. If a significant time × condition interaction was revealed, post hoc analysis was conducted using paired t-tests with the Bonferroni stepwise correction method to identify the pairwise differences. Body mass and sweat loss were analyzed using a paired t-test to determine differences in pre-exercise and post-exercise. Effect sizes were reported as Cohen for t-tests and partial eta squared ($\eta^2$) for ANOVA. Statistical significance was established as $p < 0.05$.

Results

Effect of GABA on $T_c$ and $T_{sk}$

As shown in Figure 3A, no significant interaction was noted $[F (1.357, 24.434) = 1.181, p = 0.191, P\eta^2 = 0.091]$. There was
Effect of GABA on skin temperature in different areas

Figure 4A shows the changes in $T_{\text{arm}}$ for both groups in the experiment. There was no significant effect of GABA on $T_{\text{arm}}$ [F (1, 18) = 0.002, $p = 0.996$, $\eta^2_p = 0.000$] and no condition $\times$ time interaction [F (1, 118.368) = 0.572, $p = 0.558$, $\eta^2_p = 0.031$]. However, there was a significant main effect of time [F (2, 35.68) = 10.081, $p < 0.01$, $\eta^2_p = 0.359$]. At T0, $T_{\text{arm}}$ was comparable between trial G (33.13 $\pm$ 0.92°C) and trial C (33.21 $\pm$ 0.97°C) ($p > 0.05$). Furthermore, the increase in $T_{\text{arm}}$ over time was greater in trial G than trial C during exercise but the difference was not statistically significant ($p > 0.05$).

As shown in Figure 4B, there was no significant effect of GABA on $T_{\text{chest}}$ [F (1, 17) = 0.039, $p = 0.846$, $\eta^2_p = 0.002$], and no condition $\times$ time interaction [F (1, 174.64) = 0.17, $p = 0.815$, $\eta^2_p = 0.010$]. However, there was a significant main effect of time F (1, 29.604) = 24.737, $p < 0.01$, $\eta^2_p = 0.593$). At T0, $T_{\text{chest}}$ values of trial G and trial C were 33.17 $\pm$ 0.59°C, 33.26 $\pm$ 0.60°C respectively. During exercise, $T_{\text{chest}}$ values increased and were significantly higher than T0 at T40 and T60 ($p < 0.01$), $T_{\text{chest}}$ values in trial G were lower than those in trial C ($p > 0.05$).

As shown in Figure 4C, there was no significant effect of GABA on $T_{\text{thigh}}$ [F (1, 18) = 0.478, $p = 0.498$, $\eta^2_p = 0.026$], and no condition $\times$ time interaction [F (2, 238.40) = 0.897, $p = 0.426$, $\eta^2_p = 0.047$] was noted. However, there was, again, a significant main effect of time [F (2, 238.40) = 64.016, $p < 0.01$, $\eta^2_p = 0.781$]. Compared with T0, the $T_{\text{thigh}}$ values of both groups increased in T20<T40. T40-T60 entered the platform stage during exercise ($p < 0.01$), and decreased after 10 min of recovery ($p < 0.01$); at each time point, there was no significant difference between the two trials.

As shown in Figure 4D, there was no significant effect of GABA on $T_{\text{calf}}$ [F (1, 18) = 0.101, $p = 0.754$, $\eta^2_p = 0.006$] and no condition $\times$ time interaction [F (1, 665.29) = 0.23, $p = 0.755$, $\eta^2_p = 0.013$]. Again, a significant main effect of time [F (1, 665.29) = 4.352, $p < 0.05$, $\eta^2_p = 0.195$] was found. $T_{\text{calf}}$ values decreased at T20 ($p < 0.01$); it rose after 10 minutes of exercise, was significantly higher than T0 at T40 ($p < 0.01$), and decreased during the recovery period. There was no significant difference between the two trials.

Effect of GABA on HR

There was no significant effect of GABA on HR [F (1, 18) = 0.006, $p = 0.938$, $\eta^2_p = 0.000$] and no condition $\times$ time interaction [F (1, 419.253) = 0.023, $p = 0.589$, $\eta^2_p = 0.023$]. There was a significant main effect of time [F (1, 419.253) = 257.104, $p < 0.01$, $\eta^2_p = 0.935$]. During exercise, HR increased over time ($p < 0.01$) and peaked at 141.80 $\pm$ 15.63 b/min with GABA drink and 139.80 $\pm$ 17.35 b/min with the placebo drink ($p > 0.05$) (Figure 5).

The dehydration of the participants in the experiment

Compared to the pre-exercise weight, there was no significant difference in both groups. There was no difference between the two groups in sweating volume and dehydration rate ($p > 0.05$). The dehydration rate was <1%, indicating that no mild dehydration occurred in the participants of the study (Table 1).
Discussion

To the best of our knowledge, no study has previously explored the effects of oral administration of GABA on exercise thermoregulation responses of individuals in a cold environment. The main finding of this study was that oral administration of GABA did not influence the thermoregulation responses of individuals who exercise at cold temperatures. The thermoregulation parameters of $T_c$, $T_{sk}$, HR, and sweat loss did not differ between experimental and control groups.

The effect of GABA on human thermoregulation during rest

At rest, the main challenge for human thermoregulation comes from cold ambient temperature conditions. The goal of body thermoregulation is to maintain core temperature by both feedback and feedforward mechanisms impinging on the integrative circuitry of the PO/AH (28). Maintaining a stable core temperature relies on balancing the heat generated and lost from conduction, convection, radiation, and evaporation (29). The cool sensation that elicits the cold defense response involves the activation of the cation channel conductance of the transient receptor potential 8 (TRPM8) in the skin’s cold sensation thermoreceptors. The integrating center of the PO/AH receives signals from thermoreceptors in the skin that generate a typical physiological response to cold environments, including muscle shivering and cutaneous vasoconstriction-preventing heat loss and generating heat (29–31).

In our results, the values of $T_c$ at $T_0$ in trial C and trial G were $37.45 \pm 0.17^\circ$C, and $37.33 \pm 0.25^\circ$C, respectively. The changes in the $T_{sk}$, $T_{chest}$, and $T_{thigh}$ were relatively small, while $T_{arm}$ and $T_{calf}$ both decreased consistently with time (Figures 3, 4). This finding concurs with Kruk et al. (32), who concluded that pre-exercise cold exposure resulted in a significant decrease in rectal temperature. With the increased temperature gradient between the body and the environment, the skin acts as a heat radiator for the body, and when the skin temperature is higher than the air temperature, heat is transferred from the body to the environment. As heat dissipation increases, the decrease in core body temperature causes the activation of mechanisms that preserve or generate heat. Skin temperature is controlled by blood flow to the skin surface, and when reacting to a cold environment the blood flow to the skin surface decreases so that less heat is transferred from the body core to the skin surface. Thus, the vasoconstrictive response to exposure to cold causes a decrease in heat dissipation and guarding of core temperature, resulting in a decrease in shell temperature. Repetitive rhythmic muscle contractions produce shivering to release heat, usually starting with contractile activity or redistribution of blood from the trunk and skeletal muscles, followed by blood flow to the distal part of the extremities (33). There were no significant differences in the values of $T_c$ and $T_{sk}$ between the groups in our studies. This may have been because oral administration of GABA was not yet effective within 20 min, or chemical stimulation did not affect the body.
FIGURE 3
Change in upper arm (A), chest (B), thigh (C), calf (D) temperature in trial G (black) and trial C (white). Values are means ± SD. Bars represent standard deviations. * P < 0.05, ** P < 0.01, significant difference compared to T₀.

The effect of GABA on human thermoregulation during exercise

The hypothalamus is the center of thermoregulation with the PO/AH integrating temperature information from all parts of the body and regulating body temperature by suppressing heat production in other parts (16). GABA is widely distributed in the assorted hypothalamic nuclei, especially in the PO/AH, and has a regulatory role in the body’s temperature (36). There are two types of temperature-sensitive neurons in the PO/AH – heat-sensitive neurons and cold-sensitive neurons controlling heat loss and heat generation, respectively (37). Animal studies show that GABA and GABA-A receptors in the PO/AH receive cold information from the skin and induce a thermogenic response (38).

Exercise increases heat output. However, a small portion of the heat generated becomes mechanical energy and is mostly dissipated as heat. The study results showed that the Tc of trial G and trial C continued to increase by 0.63 ± 0.31, 0.56 ± 0.35 respectively, until the end of the exercise. The Tc values in trial G tended to be higher than those of trial C from 35 to 60 min (Figure 3A). This suggests that both groups’ gradual increase in core temperature was because of increased heat production by skeletal muscle and the liver. Heat thermoreceptors transmit signals to the PO/AH, which increases heat dissipation by stimulating vasodilation and sweat secretion in the skin. The skin is the body’s main radiator and circulating blood transfers heat from the core to the skin surface to dissipate heat.

In our study, both groups’ Tsk, Tarm, and Tcalf increased during exercise (Figures 3B, 4A,D). In trial G, the value of Tarm and Tcalf was higher than trial C. This may have been because the skin vasodilates and more blood flows to the skin surfaces, distributing blood to the shell in exercise. It was reported that forearm blood flow increases rapidly with core temperature in humans (39). Another study showed that people who exercise regularly did not have reduced leg blood flow during submaximal performance cycling (40). In a study by Galloway (41), rectal temperature reached 39.2°C and mean skin temperature decreased from 25°C to 22.5°C when individuals cycled at 70% VO₂max at an ambient
temperature of 11°C. Participants performing sustained exercise at moderate intensities may be less likely to have decreased rectal temperatures in an environment of 3–11°C (42). Another study demonstrated that with temperatures between −10°C and 20°C, there was no difference in rectal temperature when individuals exercised at 64% VO2max, suggesting an elevated core temperature separate from ambient temperature (43). These studies indicate that exercise intensity is sufficient to prevent a decrease in core temperatures and that cold-sensitive receptors in the PO/AH are not activated, resulting in the inability of GABA to function.

Hyperthermia, with a core body temperature above 38°C, is thought to be an important factor contributing to central fatigue and limiting endurance exercise capacity (44). However, there were no significant differences in core and mean skin temperatures between the groups during exercise. GABA consumption did not significantly increase core body temperature, and thus the risk and negative effects of hyperthermia during exercise were reduced. It is also possible that oral GABA raises core temperature under cold conditions and then lowers core temperature because of increased heat production from exercise. The two effects of GABA would then be largely neutralized, maintaining core temperature in the normal range rather than further raising it. These findings suggest that GABA is a safe food supplement ingredient even when used in cold conditions during exercise.

Some animal studies suggest that GABA may not be able to cross the blood–brain barrier (BBB) (45, 46), which may explain the lack of modulatory effect of GABA administration in the present experiment. Nevertheless, additional evidence suggests that small concentrations of GABA can reach the BBB (47, 48). This should be clarified in future studies using nuclear magnetic resonance spectroscopy (MRS) in humans to study the effect of GABA administration on GABA concentration. While we recognize the limitations of our work, animal experiments have used an ambient temperature nearly 5°C colder than the present experiment. The ambient temperature in our study may have been insufficiently cold to stimulate cold-sensitive neurons in the preoptic-thalamic anterior, yielding inconsistent results. A possible explanation for GABA's limited effect on the core body and mean skin temperature could be the relatively moderate ambient conditions in which the exercise tests were performed. Exercise at an ambient temperature of 10°C suggests that the exercise stimulus was sufficient to balance heat generation and heat dissipation for both groups. The cooling mechanism of increasing heat storage was sufficient for the participants to overcome core temperature increases during exercise, like increased skin heat dissipation during exercise in the cold.

The effect of GABA on human thermoregulation during recovery

Both groups showed a linear decrease in core body temperature and mean skin temperature after exercise during the recovery period, with values higher than baseline values, implying that participants did not recover sufficiently. However, core body temperature and mean skin temperature were higher in trial G than in trial C and some parts of the skin temperature decreased. However, Tc and Tsk did not differ between the groups. It may be that at the end of the exercise, the core body temperature deviated from the value of the turning point. The thermoregulatory system then started to work, and the heat-sensitive neurons received a signal from the PO/AH leading to stimulation of the heat signal and more impulses being sent. This promoted sweat secretion through neurotransmission, increasing blood flow to the skin and promoting heat dissipation, resulting in lower body temperature. The preoptic areas of the hypothalamus react to heat and the medial dorsal area to cold, with a degree of antagonism between these two brain regions (49). When heat production increases, activation of neurons in the preoptic area inhibits the response to cold during exercise. Therefore, there was no increase in core body temperature for exercising participants administered GABA orally.
The effect of GABA on human HR

Heart rate is also an essential indicator in the process of thermoregulation. During exercise, the core temperature rises due to the movement of skeletal muscles, and the body needs to dissipate heat through the skin, so the blood flow to the skin is enhanced, leading to an increase in heart rate. It has been reported that within a certain range, for every 2°C increase in body temperature, cardiac output increases by ~10%, and peripheral resistance decreases by 20%. During the transition from rest to exercise, the rapid increase in heart rate (HR) predominantly mediated by cardiac vagal withdrawal, supported by increased cardiac sympathetic activity after a few seconds (50, 51). This experiment showed that after exposure to 11°C for 20 min, both groups of the heart rate compared to 0 min increased. During exercise, the heart rate significantly increased in trials G and C, peaking at 141.80 ± 15.63 b/min, 139.80 ± 17.35 b/min, respectively. When the athletes are exercising in the cold, blood is redistributed from the central part of the body to the peripheral tissues to increase heat dissipation from the skin, cardiovascular demands may dramatically increase due to the dual demand for blood supply by the working muscles and the skin, which in turn increases the heart rate. After exercise, the heart rate maintains stability after a rapid descent to pre-exercise level. However, there were no differences among groups in heart rate. Upon entering the recovery period, increased venous reflux caused baroreceptor reflex. HR decreased rapidly in our study unlike previous research that showed GABA activation in conscious rats induces press or response and tachycardia (52, 53). However, the extent to which these findings can be extrapolated to humans remains unclear. Previous studies have indicated that central GABAergic mechanisms are involved in the HR responses at the onset of exercise. Teixeira (54) studied participants randomly performing three bouts of 5× passive and active cycling under placebo and oral administration of diazepam (10 mg). The GABAergic mechanisms contributed significantly to the muscle mechanome reflex-mediated HR responses at the onset of exercise. Possible explanations for this may be that diazepam crosses the BBB (55). Contrary to Teixeira’s (54) findings, we did not find that GABA affects HR. The results of studies on whether GABA can cross the BBB are inconsistent, and the findings in our study are likely because GABA cannot cross the BBB.

Dehydration of participants in the experiment

Sweat production and the subsequent evaporation are the principal modes of heat loss in humans during exercise. Exercise elevates core and skin temperatures, contributing to the increased sweat rate. When the body core temperature rises above the critical level of 37°C (98.6°F), sweating increases the rate of heat loss. This experiment was conducted before and during exercise with 3 ml/kg body weight each time. By measuring body weight before and after exercise, we concluded that the rate of sweat loss between the experimental and control groups did not reach the level of dehydration, and there was no significant difference between the two groups, indicating that the exercise capacity of the participants was not affected by dehydration in the two exercise groups.

Conclusion

There are several limitations to this study. First, although we estimated an appropriate sample size using the G-power software, the number of participants only met the minimum sample size requirement. Second, Only 1,000 mg of GABA was validated for its effect on thermoregulation in cold environments. In future studies a larger sample size should be used to study the effect of different doses of GABA intake for prolonged exercise on thermoregulation and reveal the dose-effect relationship. Finally, it has been reported that serum GABA concentrations peak at 20–40 min and can be maintained at higher than pre-dose levels for at least 1 h. The present experiment involved a rest period of 20 min, but it is possible that plasma GABA concentrations vary in each individual. In the present study, we did not measure plasma GABA concentrations. Therefore, future research should measure plasma GABA levels and clarify the mechanisms involved in conjunction with animal experiments.

Oral administration of GABA was not shown to be effective in increasing core body temperature of individuals when cycling for 40 min in a cold environment, but it is a safe supplemental ingredient for exercise and other healthy diets. Core temperature can be increased by exercise for a short period of time in cold environments. However, it is important to wipe sweat and keep warm after exercise to avoid upper respiratory tract infections. Whether different doses of GABA have any potential effect on thermoregulation during exercise of longer duration in cold environments requires further investigation.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary material.

Ethics statement

The studies involving human participants were reviewed and approved by Guangxi Normal University.
The patients/participants provided their written informed consent to participate in this study.

Author contributions

HW conceived and supervised the study. YH and HW designed the experiments and carried out the experiments. LC analyzed the data and wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding

This study was supported by the Innovation Project of Guangxi Graduate Education (No.YCBZ2022069).

Acknowledgments

The authors would like to thank the participants in this study for their collaboration and the native English-speaking scientists at Charlesworth Group for editing our manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2022.883571/full#supplementary-material

References

1. Wakabayashi H, Oku J, Tipton MJ. Exercise performance in acute and chronic cold exposure. J Phys Fit Sports Med. (2015) 4:177–85. doi: 10.7600/jpfsm.4.177
2. Castellani JW, Sawka MN, DeGroot DW, Young AJ. Cold thermoregulatory responses following exertional fatigue. Front Biosci (Schol Ed). (2010) 2:654–65. doi: 10.2741/s106
3. Castellani JW, Tipton MJ. Cold stress effects on exposure tolerance and exercise performance. Compr Physiol. (2015) 6:643–69. doi: 10.1002/cphy.c140081
4. Ros GS. Fatal events related to running competitions in the mountains. Wilderness Environ Med. (2021) 32:176–80. doi: 10.1016/j.wem.2020.12.001
5. Lim CL, Byrne C, Lee JK. Human thermoregulation and measurement of body temperature in exercise and clinical settings. Ann Acad Med Singap. (2008) 37:347–53. doi: 10.1097/MAJ.0b013e318145a721
6. Rothhaas R, Chung S. Role of the preoptic area in sleep and thermoregulation. Front Neurosci. (2021) 15:664781. doi: 10.3389/fnins.2021.664781
7. Boudaba C, Szabo K, Tasker JG. Physiological mapping of local inhibitory inputs to the hypothalamic paraventricular nucleus. J Neurosci. (1996) 16:7151–60. doi: 10.1523/JNEUROSCI.16-22-0151.1996
8. Decavel C, Van den Poel AN. GABA: a dominant neurotransmitter in the hypothalamus. J Comp Neurol. (1990) 302:1019–37. doi: 10.1002/cne.9030209423
9. Yasumatsu M, Yazawa T, Otokawa M, Kawasawa K, Hasegawa H, Aihara Y. Monoamines, amino acids and acetylcholine in the preoptic area and anterior hypothalamus of rats: measurements of tissue extracts and in vivo microdialyses. Comp Biochem Physiol A Mol Integr Physiol. (1998) 121:11-23. doi: 10.1016/S0195-6433(98)10096-X
10. Hepsomali P, Groeger JA, Nishihira J, Scholey A. Effects of oral gamma-aminobutyric acid (GABA) administration on stress and sleep in humans: an automated psychophysiological assessment. Front Neurosci. (2020) 14:923. doi: 10.3389/fnins.2020.00923
11. Hinton T, Jelinek HF, Vienckou V, Johnston GA, Matthews S. Effect of GABA-fortified oolong tea on reducing stress in a university student cohort. Front Nutr. (2019) 6:27. doi: 10.3389/fnut.2019.00027
12. Nakamura H, Takishima T, Kometai T, Yokogoshi H. Psychological stress-reducing effect of chocolate enriched with gamma-aminobutyric acid (GABA) in humans: assessment of stress using heart rate variability and salivary chromogranin A. Int J Food Sci Nutr. (2009) 60 Suppl 5:106–13. doi: 10.1080/09637480802558508
13. Ngo DH, Vo TS. An updated review on pharmaceutical properties of gamma-aminobutyric acid. Molecules. (2019) 24:2678. doi: 10.3390/molecules24152678
14. Oketch-Rabah HA, Madden EE, Roe AL, Bett JM. United states pharmacopeia (USP) safety review of gamma-aminobutyric acid (GABA). Nutrients. (2021) 13:2742. doi: 10.3390/nu13082742
15. Sakashita M, Nakamura U, Horie N, Yokoyama Y, Kim M, Fujita S. Oral supplementation of gamma-aminobutyric acid and whey protein improves whole body fat-free mass in men after resistance training. J Clin Med Res. (2019) 11:428–34. doi: 10.14740/jcmebr3817
16. Ishiwata T, Saito T, Hasegawa H, Yazawa T, Kotani Y, Otokawa M, et al. Changes of body temperature and thermoregulatory responses of freely moving rats during GABAergic pharmacological stimulation to the preoptic area and anterior hypothalamus in several ambient temperatures. Brain Res. (2005) 1048:32–40. doi: 10.1016/j.brainres.2005.04.027
17. Miyazawa T, Kawabata T, Suzuki T, Imai D, Hamamoto T, Yoshikawa T, et al. Effect of oral administration of GABA on temperature regulation in humans during rest and exercise at high ambient temperature. Osaka City Med J. (2009) 55:99–108.
18. Nybo L, Rasmussen P, Sawka MN. Performance in the heat-physiological factors of importance for hyperthermia-induced fatigue. Compr Physiol. (2014) 4:657–89. doi: 10.1002/cphy.c130012
19. Tatterson AJ, Hahn AG, Martin DT, Febbraio MA. Effects of heat stress on physiological responses and exercise performance in elite cyclists. J Sci Med Sport. (2000) 3:118–30. doi: 10.1016/S1440-2440(00)80080-8
20. Davies MJ, Clark B, Welvaert M, Skorski S, Garvicani-Lewis LA, Saunders P, et al. Effect of environmental and feedback interventions on pacing profiles in cycling: a meta-analysis. Front Physiol. (2016) 7:591. doi: 10.3389/fphys.2016.00591
21. Faul F, Erdfelder E, Buchner A, Lang AG. Statistical power analyses using G’Power 3: tests for correlation and regression analyses. Behav Res Methods. (2009) 41:1149–60. doi: 10.3758/BRM.41.4.1149
22. Paul F, Erdfelder E, Lang AG, Buchner A. G'-Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. Behav Res Methods. (2007) 39:175–91. doi: 10.3758/BF03193416

23. Miyawara T, Kawabata T, Okazaki K, Suzuki T, Imai D, Hamamoto T, et al. Oral administration of γ-aminobutyric acid affects heat production in a hot environment in resting humans. J Physiol Anthropol. (2012) 31:3. doi: 10.1186/1880-6805-31-3

24. Mandle T, King J, Collacciott E, Jones DA. Drink temperature influences fluid intake and endurance capacity in men during exercise in a hot, dry environment. Exp Physiol. (2006) 91:925–33. doi: 10.1113/expphysiol.2006.034223

25. Yaspeleas BB 3rd, Ivy JL. Effect of carbohydrate supplements and water on exercise metabolism in the heat. J Appl Physiol. (1991) 71:680–7. doi: 10.1152/jappl.1991.71.2.680

26. Ramanathan NL. A new weighting system for mean surface temperature of the human body. J Appl Physiol. (1964) 19:531–3. doi: 10.1152/jappl.1964.19.3.531

27. Sawka MN, Noakes TD. Does dehydration impair exercise performance? Med Sci Sports Exerc. (2007) 39:1209–17. doi: 10.1249/mss.0b013e3181244664

28. Morrison SF, Nakamura K. Central mechanisms for thermoregulation. Annu Rev Physiol. (2010) 81:285–308. doi: 10.1146/annurev-physiol-020510-141546

29. Tan CL, Knight ZA. Regulation of body temperature by the nervous system. Neuron. (2018) 93:31–48. doi: 10.1016/j.neuron.2018.02.022

30. Tansey EA, Johnson CD. Recent advances in thermoregulation. Adv Physiol Educ. (2015) 39:134–48. doi: 10.1152/advan.00126.2014

31. Madden CJ, Morrison SF. Central nervous system circuits that control body temperature. Neurousci Lett. (2019) 696:225–32. doi: 10.1016/j.neulet.2018.11.027

32. Erik B, Pekkarinen H, Harri M, Manninen K, Hanninen O. Thermoregulatory responses to exercise at low ambient temperature performed after precooling or preheating procedures. Eur J Appl Physiol Occup Physiol. (1990) 59:416–20. doi: 10.1007/BF02388622

33. Bell DG, Tikuisis P, Jacobs I. Relative intensity of muscular contraction during shivering. J Appl Physiol. (1992) 72:2336–42. doi: 10.1152/jappl.1992.72.6.2336

34. Song K, Wang H, Kamm GB, Pohle W, Reis FC, Heppenstall P, et al. The TRPM2 channel is a hypothalamic heat sensor that limits fever and can drive temperature-dependent body weight homeostasis. J Neurosci. (2016) 36:5034–46. doi: 10.1523/JNEUROSCI.0213-16.2016

35. Kimura H, Kuriyama K. Distribution of γ-aminobutyric acid (GABA) in the rat hypothalamus: functional correlates of GABA with activities of appetite controlling mechanisms. J Neurochem. (1975) 24:903–7. doi: 10.1111/j.1471-4159.1975.tb03634.x

36. Hori T. An update on thermosensitive neurons in the brain: from cellular biology to thermal and non-thermal homeostatic functions. Ipn J Physiol. (1991) 41:1–22. doi: 10.1107/jpophysiology.41.1

37. Osaka T. Cold-induced thermogenesis mediated by GABA in the preoptic area of anesthetized rats. Am J Physiol Regul Integr Physiol. (2004) 287:R306–313. doi: 10.1152/ajpregu.00003.2004

38. Waterhouse J, Aizawa S, Nevill A, Edwards B, Weinert D, Atkinson G, et al. Rectal temperature, sweat rate, distal sweat flow, and forearm blood flow following mild exercise at two phases of the circadian cycle. Chronobiol Int. (2007) 24:63–85. doi: 10.1080/07420580601142551

39. Proctor DN, Newcomer SC, Koch DW, Le KU, MacLean DA, Leuenberger UA. Leg blood flow during submaximal cycle ergometry is not reduced in healthy older normally weightwomen. J Appl Physiol. (2003) 94:1859–69. doi: 10.1152/japplphysiology.00898.2002

40. Galloway SD, Maughan RJ. Effects of ambient temperature on the capacity to perform prolonged cycle exercise in man. Med Sci Sports Exerc. (1997) 29:1240–9. doi: 10.1097/00005768-199709000-00018

41. Nimmo M. Exercise in the cold. J Sports Sci. (2004) 22:898–915. discussion-6. doi: 10.1080/02640410400005883

42. Layden JD, Patterson MJ, Nimmo MA. Effects of reduced ambient temperature on fat utilization during submaximal exercise. Med Sci Sports Exerc. (2002) 34:774–9. doi: 10.1097/01.MSS.0000057682.002500-00008

43. Nybo L. CNS fatigue provoked by prolonged exercise in the Heat. Front Biosci (Elite Ed). (2010) 2:779–92. doi: 10.2741/e138

44. Knudsen GM, Poulsen HE, Poulsen OB. Blood-brain barrier permeability in galactosamine-induced hepatic encephalopathy. No evidence for increased GABA-transport. J Hepatol. (1988) 6:187–92. doi: 10.1016/S0168-8278(88)80330-8

45. Kuriyama K, Ste PY. Blood-brain barrier to H3-gamma-aminobutyric acid in normal and amino oxoacetic acid-treated animals. Neuropharmacology. (1971) 10:103–4. doi: 10.1016/0028-3908(71)90013-X

46. Al-Sarraf H. Transport of 14C-gamma-aminobutyric acid into brain, cerebrospinal fluid and choroid plexus in neonatal and adult rats. Brain Res Dev Brain Res. (2002) 139:121–9. doi: 10.1016/S0165-3806(02)00537-0

47. Shyamaladervi N, Jayakumar AR, Sujatha R, Paul V, Subramanian EH. Evidence that nitric oxide production increases gamma-aminobutyric acid permeability of blood-brain barrier. Brain Res Bull. (2002) 57:231–6. doi: 10.1016/S0361-9230(01)00755-9

48. Zhao ZD, Yang WZ, Gao C, Fu X, Zhang W, Zhou Q, et al. A hypothalamic circuit that controls body temperature. Proc Natl Acad Sci U S A. (2017) 114:2042–7. doi: 10.1073/pnas.1616251114

49. Smith SA, Mitchell JH, Garry MG. The mammalian exercise pressor reflex in health and disease. Exp Physiol. (2006) 91:89–102. doi: 10.1113/expphysiology.2005.032667

50. Williamson JW, Nóbrega AC, Winchester PK, Zim S, Mitchell JH. Instantaneous heart rate increase with dynamic exercise: central command and muscle-heart reflex contributions. J Appl Physiol. (1995) 78:1273–9. doi: 10.1152/jappl.1995.78.4.1273

51. Catelli JM, Giakas WJ, Sved AF. GABAergic mechanisms in nucleus tractus solitarius alter blood pressure and vasopressin release. Brain Res. (1987) 403:279–89. doi: 10.1016/0006-8993(87)90065-5

52. Landulpho CD, Dias AC, Colombari E. Cardiovascular mechanisms activated by microinjection of baclofen into NTS of conscious rats. Am J Physiol Heart Circ Physiol. (2005) 288:H716–h23. doi: 10.1152/ajpheart.2004.102422

53. Teixeira AL, Ramos PS, Samora M, Sabino-Carvalho JL, Ricardo DR, Colombari E, et al. GABAergic contribution to the muscle mechanoreflex-mediated heart rate responses at the onset of exercise in humans. Am J Physiol Heart Circ Physiol. (2018) 314:H716–h23. doi: 10.1152/ajpheart.00857.2017

54. Tedeschi G, Smith AT, Dhillon S, Richards A. Rate of entrance of benzodiazepines into the brain determined by eye movement recording. Br J Clin Pharmacol. (1983) 15:103–7. doi: 10.1111/j.1365-2125.1983.tb01471.x