The Fundamental Role of Bicarbonate Transporters and Associated Carbonic Anhydrase Enzymes in Maintaining Ion and pH Homeostasis in Non-Secretory Organs

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Abstract: The bicarbonate ion has a fundamental role in vital systems. Impaired bicarbonate transport leads to various diseases, including immune disorders, cystic fibrosis, tumorigenesis, kidney diseases, brain dysfunction, tooth fracture, ischemic reperfusion injury, hypertension, impaired reproductive system, and systemic acidosis. Carbonic anhydrases are involved in the mechanism of bicarbonate movement and consist of complex of bicarbonate transport systems including bicarbonate transporters. This review focused on the convergent regulation of ion homeostasis through various ion transporters including bicarbonate transporters, their regulatory enzymes, such as carbonic anhydrases, pH regulatory role, and the expression pattern of ion transporters in non-secretory systems throughout the body. Understanding the correlation between these systems will be helpful in order to obtain new insights and design potential therapeutic strategies for the treatment of pH-related disorders. In this review, we have discussed the broad prospects and challenges that remain in elucidation of bicarbonate-transport-related biological and developmental systems.

Keywords: bicarbonate; ion transporters; carbonic anhydrase; intracellular pH; maturation

1. Convergent Regulation of Ion Homeostasis

Ion homeostasis is an important process involved in various organ functions, including modulation of sensitivity to blood pressure, immune cell differentiation, fluid secretion, and fertilization of reproductive cells such as sperm and eggs. Ions utilize several paths to enter the cytosolic milieu, by accompanying other cations and anions through the involvement of cotransporters or energy-consuming channels. In this section, we have focused on the convergent regulation of various ion transporters and channels in various physiological and pathophysiological milieus.

Na\(^+\) is an essential element for all living organisms and a crucial ion for the modulation of osmotic pressure. Regulation of Na\(^+\) concentration is modulated by various ion transporters, including Na\(^+\)-K\(^+\)-Cl\(^-\) cotransporter 1 (NKCC1), electrogenic-type Na\(^+\)-HCO\(_3\)^\(-\) cotransporter (NBCe), and sodium–potassium ATPase (Na\(^+\)-K\(^+\)-ATPase). Coregulation of these transporters and a pump revealed that cytosolic Na\(^+\) level is strongly dependent on the extracellular K\(^+\) and HCO\(_3\)^\(-\) levels in cortical astrocytes, which are involved in pH regulation and volume regulation [1]. The HCO\(_3\)^\(-\)-dependent channels and transporters include cystic fibrosis transmembrane conductance regulator channel (CFTR) [2,3], NBC [4,5], Cl\(^-\)/HCO\(_3\)^\(-\) exchangers (CBE) [6,7], and Ca\(^{2+}\)-activated Cl\(^-\) channel TMEM16A (also known as Anoctamin-1) [8,9]. The NBC and the epithelial sodium channel (ENaC) are essential in the modulation of CFTR-dependent membrane potential and the cAMP/protein kinase A (PKA) signaling pathway in sperm capacitation [10]. The CFTR exists in epithelial cells and also exhibits critical
functions in lung homeostasis related to maintain the ion gradients. The mutation of CFTR, ΔF508 CFTR, causes chronic inflammatory responses and, eventually, cystic fibrosis (CF) [11]. The function of CFTR reciprocally regulates ENaC activity. In CF patients, ΔF508 CFTR and highly activated ENaC coexist in lung epithelia. Overexpressed wild-type CFTR effectively suppresses ENaC activity, whereas ΔF508 CFTR elevated ENaC activity in a CF airway epithelial model, suggesting that CFTR is functionally involved in regulation of inflammatory responses in the lungs [11].

The regulation of intracellular pH (pH₅) is mediated by HCO₃⁻ transporters coupled with Na⁺, including NBCe, electroneutral-type NBC (NBCn), and Na⁺-driven CBE (NDCBE) [12]. A splice variant of NBCe, NBCe2, is associated with salt sensitivity in the renal proximal tubule system. Although NBCe2 modulates renal HCO₃⁻ transport independently of hypertension, NBCe2 activity is hyperresponsive to intracellular Na⁺ concentration [13]. The Na⁺-H⁺ exchanger (NHE) and CBE activity were also higher in homozygous variants compared to wild-type variants of NBCe2 in the renal proximal tubule cells [14]. Involvement of NHE in the enhanced NBC system can be implicated in the regulation of steady-state pH₅. Although this evidence was gathered in vascular smooth muscle cells, elevated steady-state pH₅ and enhanced pH recovery from acidosis are mediated by NBCn1. Association of NHE activity maintains a steady-state pH₅ [15]. NDCBE, such as that coded by gene SLC4A8, transports extracellular Na⁺ and HCO₃⁻ into cells in exchange for intracellular Cl⁻ in brain and testis [12]. In contrast, one of the CBEs, anion exchanger 4 (AE4), promotes Cl⁻ influx to exchange K⁺ (or Na⁺) for HCO₃⁻ in secretory cells [16]. A recent study demonstrated that Cl⁻/HCO₃⁻ exchange of AE4 under Cl⁻-free conditions mediates Na⁺-HCO₃⁻ cotransport without changes in membrane potential [16]. In addition, the transport of Na⁺, Cl⁻, and HCO₃⁻ occurs in the presence of K⁺, along with Cs⁺, Li⁺, and Rb⁺ [16]. AE3, the functions of which are still unknown, has been implicated in acid–base homeostasis in the kidney [17]. However, Kampik et al. showed that expression of receptor potential melastatin (TRPM) 6 channel increased in the kidney of AE3-deficient mice [17]. Another CBE, AE1, harbors a binding site of the Na⁺-K⁺-ATPase β1 subunit, especially in the human kidney [18]. Interaction between AE1 and the Na⁺-K⁺-ATPase (especially the β1 subunit) is involved in achievement and retention of basolateral membrane residency in kidney cells [18]. Recently, kinase-mediated transporter regulation was revealed in the renal system. Renal tubular transport and reabsorption of water and electrolytes are regulated by two kinases, serine/threonine kinase Akt and serum-/glucocorticoid-inducible kinase 1 (SGK1), increasing Na⁺ reabsorption [19]. Signaling pathways involving the phosphorylation of Akt enhance activities of NBCe1, ENaC, and Na⁺-Cl⁻ co-transporter (NCC), and those involving SGK1 promote NHE3 activation, encouraging reabsorption of Na⁺. SGK1, furthermore, increases K⁺ secretion via the renal outer medullary potassium channel (ROMK) [19]. Major modulation of electrolyte transport occurs in the renal and exocrine systems, including salivary glands and pancreatic glands, and has already been extensively discussed [5,20]. In this review, we have focused on current knowledge of the regulation and roles of ion transporters in maintaining pH and appropriate HCO₃⁻ concentration with respect to physiological and pathophysiological conditions of the various organs, except renal and exocrine systems.

2. Carbonic Anhydrases: Regulation of HCO₃⁻

In this section, we discuss the regulatory enzymes of HCO₃⁻ transporters. The concentration of HCO₃⁻ provides the driving force or favorable circumstances for transporters. As regulatory enzymes, various isoenzymes of carbonic anhydrase (CA) are differentially localized in the subcellular regions (cytosolic forms: CA I, CA II, CA III, CA VII and CA XIII; membrane-associated forms: CA IV, CA IX, CA XII, and CA XIV; mitochondria-associated forms: CA VA and CA VB; and secreted form: CA VI) and their catalytic activities vary. CAs participate in essential biological processes such as respiration, pH balance, and bicarbonate transport [21,22]. During the conversion of carbon dioxide to HCO₃⁻, CAs are involved in HCO₃⁻ movement, and participate in complexes of HCO₃⁻ transporting systems [20]. Hydration of CO₂ was considered an important source for HCO₃⁻ [23,24]. Inhibition of CA with acetazolamide reduced duodenal HCO₃⁻ secretion [23]. In addition, pH regulation
of CA was addressed in various epithelia [25]. The role of CA on HCO$_3^-$ secretion was studied with HCO$_3^-$ transporters NBCn1 and NBCe2 in the duodenum [26]. Since CA was revealed to form a HCO$_3^-$ transport metabolon to accelerate HCO$_3^-$ flux, specific isoenzymes of CA have been implicated as the regulatory factor of HCO$_3^-$ transporters. CA II interacted with AE and DRA [27].

Our previous report addressed the fact that CA XII is physically associated with HCO$_3^-$ transporters anion exchanger 2 (AE2) and NBC, and regulates their activities [28]. The regulatory roles of CAs on the HCO$_3^-$ transporters have been reported to mediate cancer cell survival, cell migration, pH regulation, ion transport, and enamel formation in several biological systems [29–36]. Briefly, CA IX is involved in cell migration to facilitate the activities of AE2 and NBCe1 [30,31]. The role of oncogenic CAs will be discussed in Section 4.2. Inhibition of CA II and CA II knockout reduced the activity of NBCe1 in cortical astrocytes, suggesting that CA II is associated with an acid-loading role of NBCe1 into cells, and stabilized extracellular pH in brain tissue [33]. However, the precise regulation of CA isoforms and HCO$_3^-$ transporters remains unclear.

3. Regulatory Factors of HCO$_3^-$ Transporters

3.1. Regulatory Molecules

The HCO$_3^-$ transporters are mainly involved in the production of pancreatic and salivary fluid for HCO$_3^-$ secretion. The HCO$_3^-$-transporting mechanism is supported by several regulatory molecules and ions. This section focuses on the regulatory molecules of HCO$_3^-$ transporters, mostly found in secretory tissues such as salivary glands and pancreas [37–40]. The regulatory molecules include inositol-1,4,5-triphosphate (IP$_3$) receptor binding protein released with IP$_3$ (IRBIT), with-no-lysine (WNK) kinase, sterile 20 (STE20)-related proline/alanine-rich kinase (SPAK), spinophilin (SPL), and phosphatidylinositol 4,5-bisphosphate (PIP$_2$) [20,41,42]. These factors mediate the supportive or inhibited function of HCO$_3^-$ transport via NBCe1-B, SLC26A6, and AE2 [20,41–44].

The IRBIT protein is abundantly expressed in various tissues. The IRBIT binds the NH$_2$-terminal domain of IP$_3$ receptors (IP$_3$Rs) in resting state to inhibit the activity of IP$_3$Rs; otherwise, the IRBIT is separated from IP$_3$R when it stands in the stimulating state through the activation of G protein-coupled receptor [43,45]. The signaling cascade of released IRBIT is so far unknown. Mikoshiba et al. demonstrated that the IRBIT-binding site is located in the cytosolic N-terminus domain of NBCe-1B and clarified its interaction using a pull-down assay [46]. We previously reported that the N-terminus of NBCe1-B possesses a cluster of positively and negatively charged residues, and identified the positively charged cluster of the N-terminus of NBCe1-B as an IRBIT-binding domain [47]. The protein phosphatase 1 (PP1), as a supplementary protein of IRBIT, also possesses the enhanced role of NBCe1-B activity by the maintaining the membrane stability of NBCe1-B [38]. HCO$_3^-$ transporters are also regulated by WNK and SPAK kinases to suppress transportation of HCO$_3^-$ by decreasing the surface expression and the activity of HCO$_3^-$ transporters such as NBCe1-B and SLC26A6 [37,38]. WNKs mostly act with SPAK to phosphorylate and activate SPAK, and the activated SPAK subsequently phosphorylates NBCe1-B [38] and SLC26A6 [40,48]. The phosphorylation of these transporters decreases the membrane stability. The scaffolding functions of WNKs are mediated by their first 119 residues [49,50], and especially T-loops, to phosphorylate SPAK kinase [51]. SPAK kinases contain a proline-and arginine-rich domain (P/ARD), and a kinase domain including a serine motif (S-motif) and a COOH terminal (CCT) domain [51–53]. The CCT domain binds with SPAK binding motif [R/K]FX[V/I] of WNKs [54]. The SPAK phosphorylation site of NBCe1-B is S65 and T49 in the first 85 N-terminus residues, which acts as an autoinhibitory domain (AID) of NBCe1-B [39]. More recently, Lee et al. and Jeong et al. suggested that scaffolding protein SPL, as an actin cytoskeletal modulator, binds to AE2 and induces the enhanced AE2 activity [41,42]. SPL consists of a binding motif for F-actin, receptor, and canonical PP1 (R-K-I-H-F motif); PDZ domain; and three coiled-coil (C–C) domains [55]. Actually, the SPL can enhance AE2 activity through the 1–480 amino acid residues containing F-actin binding motif, receptor binding motif, and PP1 binding motif [42]. In addition, SPL
activity is regulated by Ca$^{2+}$ signaling with kinases including SPAK and Ca$^{2+}$/calmodulin-dependent protein kinase II (CaMKII) [41]. PIP$_2$ is the prevailing signaling molecule acting as a precursor of IP$_3$ and stimulating NBCe1-A activity [56]. In addition, PIP$_2$ indirectly increases NBCe1-B and NBCe1-C through increases of IP$_3$ and Ca$^{2+}$ [57]. The regulatory role of PIP$_2$ on ion transporters can be considered a new therapeutic approach not only in NBCe1, but also in K$_{ATP}$ [58,59], NHE1 [60], transient receptor potential canonical (TRPC) [61], and ATP-gated P2X channels [62,63]. Moreover, the activities of transporters are modulated by various phosphatase or kinase pathways and their multiple phosphorylation sites provide fine-tuning of transporters [64].

3.2. Cl$^-$ as A Signaling Ion

The Concentration of Cl$^-$ is responsible for regulating cellular functions to maintain fluid and electrolyte homeostasis through multiple transporters [65] such as CLC transporter [66], CBE [67], and CFTR [20,40]. The concentration of extracellular Cl$^-$ controls the body fluid content of ions, cellular volume, and blood pressure [68]. Likewise, Cl$^-$ plays a regulatory role in regulating HCO$_3^-$ absorption and secretion [69] and activity of NBC [70]. NBCe1-B in its resting state is inhibited by high concentrations of intracellular Cl$^-$ ([Cl$^-$]$_i$) through the Cl$^-$-interacting motif, GXXXP [70]. In NBCe1-B, the two GXXXP motifs sense intracellular Cl$^-$, and one of the motifs is revealed by the autoinhibitory module of IRBIT which interacts with high affinity for Cl$^-$. In resting state, the [Cl$^-$]$_i$ is maintained between 5 mM and 60 mM [71], which decreases activity of NBCe1-B to 40% of its highest activity, in contrast with a 140 mM concentration of Cl$^-$, which inhibited 60% of NBCe1-B activity [70]. This complicated mechanism with differential affinity for Cl$^-$ addresses that sensing with different range of [Cl$^-$]$_i$ saves NBCe1-B energy [70]. The [Cl$^-$]$_i$-sensing ability of GXXXP suggests a powerful regulatory factor; thus, further studies have to be conducted to control HCO$_3^-$ transportation more effectively.

4. HCO$_3^-$ and pH Regulation and Coordinated Transporters

Acid–base homeostasis with HCO$_3^-$ secretion is critically regulated in the kidney, lung, and exocrine systems such as salivary and pancreatic glands through various transporters, which have been extensively focused on in various reviews [5,20,72–74]. This section only focuses on pH regulation and the associated ion transporters including HCO$_3^-$ transporters in non-secretory systems. We have defined in this review that the non-secretory system includes the immune, tumorigenesis, tooth, vascular smooth muscle, heart, intestine, and reproduction systems.

4.1. Immune System

HCO$_3^-$ transport is necessary for the immune defense system. For example, human lung cancer cells (Calu3) co-cultured with lymphocytes from CFTR-null mice showed abolished HCO$_3^-$ secretion in the host defense system due to bacterial infection [75]. Since NBC activity has been implicated in the pH regulation of lymphocytes, NBCn1 has recently been shown to be strongly induced upon macrophage differentiation to acidify phagosomes [76,77]. Loss of SLC4A7 resulted in the increased intracellular acidification during phagocytosis, suggesting that intracellular pH homeostasis is associated with the anti-microbial function of macrophages [76]. Neutrophils also possess the HCO$_3^-$ transport mechanism involving NBCe1 or NBCn1, but not CBE, modulated by chemotactic agents, such as N-formylmethionyl-leucyl-phenylalanine (fMLF)/cytochalasin B, which regulates the basal pH [78,79]. Apart from the role of pH regulation, CD8$^+$ T cells are dependent on AE2 as the CBE for modulation of cell proliferation and activation [80]. Primary biliary cirrhosis (PBC) is observed in progressive autoimmune-mediated cholangitis, and PBC patient reveal a reduced expression of AE2. The precise role of AE2 in PBC still remains unknown; however, AE2-deficient mice exhibit intrahepatic T-cell activation [81]. Currently, evidence of identified transporters and regulation of activity in the immune system is relatively rare compared to in other biological systems (Table 1 and Figure 1). The critical role of HCO$_3^-$ transport in the cellular network, including epithelial cells in immune cells, should be studied carefully in the future.
4.2. Tumor System

Modulation of pH is a primary process in cancer cells with high metabolic and proliferative features. The cancer cells undergo more acid-producing processes and produce acidic metabolites H⁺ and CO₂. Therefore, adaptation of fluctuated extracellular pH is an essential process in tumors. Recently, a large cohort study on breast cancer, breast cancer cell lines, and a mouse model revealed enhanced expression of NHE1, NBCn1, and monocarboxylate transporters (MCT) MCT-1 and MCT-4 (Table 1) [82,83]. In addition, NBCe1 contributes to HCO₃⁻ transport in the LS174 colon adenocarcinoma cell line and MDA-MB-231 breast cancer cells (Table 1) [31]. Disrupted NBCn1 expression delays murine breast cancer development and progression [83,84]. Tumorigenic signaling and induced tumor hypoxia in mal-perfused regions of tumors are main features of tumorigenesis. The expression of oncogenic CAs such as CA IX or CA XII is enhanced in hypoxic regions [85–91]. These enzymes help to maintain an acidic environment and produce HCO₃⁻ to serve HCO₃⁻ transporters. Several HCO₃⁻ transporters interact with these CAs to facilitate the consumption of HCO₃⁻ [28,35,92]. Although the role of CA in the tumorigenesis is beyond the scope of this section, HCO₃⁻ transporters are, at least, upregulated and play critical roles in pH regulation in tumors [93] and will be further discussed in the final section. Data-mining analyses of changes in acid–base transporter expression revealed upregulated NHE1/3/4, various HCO₃⁻ transporters (AE3 and DRA), H⁺ pumps, and MCT-4 in pancreatic ductal adenocarcinoma [94]. Following a different approach for hematological malignancies, treatment with HCO₃⁻ transporter inhibitor 4-acetamido-4-isothio cyanostilbene-2,2-disulfonate (SITS) for T-cell lymphoma, designated Dalton’s lymphoma, increased the extracellular pH and induction of apoptosis [95]. Thus, blocking HCO₃⁻ transport and disrupting pH homeostasis could be suggested as a promising anticancer therapy. Beyond the role of pH regulation, several transporters are involved in cancer biology, including cell survival and migration (Table 1). Overexpression of AE2 in colon cancer is correlated with expression of Ki67 protein, a nuclear proliferation marker [96]. Enhanced expression of AE2 through the involvement of the transcription factor early growth response 1 (EGR1) promoted proliferation of colon cancer cells [96]. DRA is a critical Cl⁻/HCO₃⁻ exchanger involved in absorption of Cl⁻ in the colon. The downregulation of a DRA gene (SLC26A3) is observed in adenomas and adenocarcinomas of the colon [97]. In addition to the proliferative colonic crypt zone, SLC26A3-null mice present high-Cl⁻-content diarrhea and more acidic lumen due to enhanced NHE3 expression and H⁺-ATPase...
as compensation of adaptive regulation [98]. The pH regulatory role of transporters in colon or intestinal units is discussed intensively in a later section. Moreover, NBCn1 expression is increased in human breast carcinoma tissue [99]. NHE1 and NBCn1 are involved in acid extrusion with different effects on the cathepsin release in breast cancer [100] and breast cancer cell motility [101]. They appear to serve as acid regulators (Figure 2). Although the role of ion transporters in cell migration is crucial in tissue homeostasis, this topic is not discussed in this review.

**Figure 2.** Upregulated bicarbonate transporters in cancer cells. The activation of NBCn1 and the expression of CBE (AE3 and DRA) is upregulated in breast cancer and pancreatic ductal adenocarcinoma. In addition, NBCe1 and AE2 activation increases colon cancer cell proliferation.

4.3. **Tooth Developmental System**

Enamel formation requires functional activity of ion transport for pH regulation. Ameloblasts in the maturation stage regulate extracellular pH to control the regulatory networks for enamel mineralization (Table 1) [102,103]. Modulation of pH fluctuation is mediated by the cyclic transformation of ruffle-ended ameloblasts and smooth-ended ameloblasts [105]. During amelogenesis, ameloblasts in maturation stage express the gene transcripts for NBCe1 isoforms B-E [104]. Mutation of NBCe1 is associated with defects of enamel development, and its expression in ameloblasts and papillary cells depends on the developmental process to modulate pH [104]. Matured ameloblasts secrete $\text{HCO}_3^-$ into the forming enamel through the involvement of AE2 [105]. In addition, ameloblasts express SLC26A3, SLC26A4 (CBE), and SLC26A6, and the gene expression profile of null mice showed that SLC26A isoforms compensate their functions [105]. Moreover, SLC26A1 (CBE) and SLC26A7 (Cl$^-$/base transporter) were also highly expressed in maturation-stage rodent ameloblasts [106] (Figure 3). SLC26A1- and SLC26A7-null mice did not exhibit abnormal enamel formation [106]. There were compensation processes of interaction units such as CFTR, CA II, CA VI, AE2, NBCe1, AE4, and SLC26A9 (epithelial Cl$^-$ channel), and thus no obvious dental phenotype was revealed [102,106]. Bronckers summarized the major types of transmembrane molecules of ameloblasts [103]. The experimental and functional evidence of transporters and pH regulation in ameloblasts or dental cells needs to be elucidated. Recently, HCO$_3^-$ secretion was observed in ameloblasts from a rat cell line, HAT-7 cells, in a 2D culture system. HAT-7 cells are recommended as a useful functional model for transporters of ameloblasts [107]. Moreover, the characteristics of pH modulation in human dental pulp stem cells by NBC, NHE, AE, and Cl$^-$/OH$^-$ exchanger (CHE) were recently addressed [108].
Regulation of pH in vascular smooth muscle cells (VSMCs) has been studied in various disease models. Vascular wall affected by an agonist modulates pH to induce contraction or relaxation of mesenteric arteries. The regulation of NBC activity during artery contraction is associated with pH control (Table 1). The calcineurin A signaling pathway interacts with NBCn1, but not NHE, and modulates the NBC activity in VSMCs [109]. NBCn1-null mice were observed to exhibit attenuated myogenic tone in the presence of N-nitro-L-arginine methyl ester (L-NAME), and reduction in rhythmic contractile pattern compared to the wild type [110]. Disruption of NBCn1 inhibited nitric-oxide-mediated Rho kinase signaling and is involved in perturbed regulation of blood pressure [111]. The effect of alcohol on aortic smooth muscle cells modulated resting pH via the regulation of NBC, NHE, CHE, and AE [112]. Axial pH gradients enhanced migration and promoted filopodia via the NBCn1 in VSMCs, suggesting that NBCn1 and its HCO$_3^-$ component are involved in arterial remodeling [113,114]. Stimuli of NHE and NBCn1 under acidic extracellular pH inhibited NHE1 and NBCn1 activities in VSMCs [115].

4.5. Cardiac System

Acid–base balance is a critical factor in the cardiac system and is involved in excitation–contraction coupling. Thus, acid–base imbalance participates in cardiac dysfunction and arrhythmias. NHE1 in the gap junction at intercalated discs and NBC (not identified in subtype) in t-tubules were observed in rat ventricular myocytes to be involved in junction communication and excitation–contraction coupling, respectively [116].

The pathogenesis of cardiac tissue injury such as ischemic reperfusion injury involves various ion transporters such as NHE. During ischemia, the accumulation of intracellular H$^+$ is mediated by NHE [117]. The Cl$^-$/OX$^{2-}$ exchanger/CBE SLC26A6 was detected and dominantly expressed in

**Figure 3.** Enamel development induced by bicarbonate transporters. CBE (AE2, SLC26A1, and SLC26A7) and NBCs (NBCe1 (B–E)) transporters induce formation and development of enamel.
mouse cardiac myocytes to regulate pH balance [118,119]. However, the potential functions of cardiac SLC26A6 remain unknown. Recently, it has been reported that angiotensin II is responsible for the impairment of the NBCe1 in cardiac hypertrophy, and enhanced NBCn1 activity compensated the reduced function of NBCe1 (Table 1) [120]. NBCe1 is involved in myocardial damage by mediating Na\(^+\) and Ca\(^{2+}\) loading. Treatment with a-L3, a selective NBC inhibitor, improved myocardial function during ischemic reperfusion injury [120]. HCO\(_3^-\) transporters functionally interact with carbonic anhydrases (CAs) [27,121–123]. The CA localized in the plasma membrane and cytosol and the CA-coupled NBC in plasma membrane modulated the intracellular pH to transport HCO\(_3^-\) effectively into cytosol. Peetz et al. reported that the involvement of CA mediates the convergent NBC activity to modulate pH [35]. In addition, MCT activity enhanced lactate influx as an energy substrate in cardiomyocytes [35]. Although the modulatory mechanisms of NBC remain unclear, more recently, agonist stimulation of mineralocorticoid receptor, aldosterone, enhanced NBC activity via the activation of G-protein-coupled receptor GPR30/PI3K-AKT pathway in rat cardiomyocytes [124].

4.6. Digestive System

Duodenal, intestinal, and colonic HCO\(_3^-\) regulation is crucial for epithelial defense against acid and for mucus secretion, as well as for pH regulation [125–127]. Several reports have shown that splice variants of NBC are differentially expressed in the intestinal system. NBCn1 is localized in duodenal villous enterocytes, colonic crypt cells, and goblet cells of the intestine [128]. Electrogenic splice variants of NBC, NBCe1-B and NBCe1-C, are predominantly expressed in proximal colon, and the electroneutral forms, NBCn1-C or NBCn1-D, are widely expressed in proximal and distal colon [129]. The parietal cells have been identified by AE2 in the basolateral membrane [130]. Recently, the Cl\(^-\)/HCO\(_3^-\) exchanger SLC26A9 has been reported in mouse and human gastrointestinal tract [131]. Reduced expression of SLC26A9 has also been related with impaired secretion of HCO\(_3^-\) in the proximal duodenal mucus layer and enhanced death rate of CFTR-null mice [131]. The membrane expression of these transporters maintains intracellular pH homeostasis, and a potential role of coordinated CAs has been proposed in the gastrointestinal system [132]. Though the transporters are involved in HCO\(_3^-\) dependent mucosal function for epithelial protection of digestive tract (Table 1), the precise role and the differential expression of transporters and associated CAs should be studied more extensively.

4.7. Reproduction System

HCO\(_3^-\) is also critical to pH maintenance as well as ionic homeostasis in the reproduction system, including sperm production or sperm quality, male reproductive tract, and uterine epithelial cells [133]. AE2, NBCe1, NBCn1, and NDCBE were identified in sertoli cells, targets of hormonal signaling, and are associated with spermatogenesis [134,135] (Table 1 and Figure 4). Estradiol level is involved in the modulation of ion transporter expression [136–140]. CFTR also plays a critical role in male fertility. Age-dependent CFTR regulation correlates with sperm quality, including sperm motility. Reduced CFTR expression in sperm was observed in men with advanced age [141]. The male reproductive duct needs to maintain adequate luminal pH by modulating HCO\(_3^-\) transport through DRA, SLC26A6, and CFTR, and/or H\(^+\) transport through NHE3 [133,142–144]. In addition, SLC26A4 and SLC26A6 were expressed in the endometrial cells and localization of SLC26A6 was dependent on the menstrual cycle [145]. The uterine fluid is affected by changes in sex steroids due to the fluctuation in pH and Na\(^+\) concentration. Estradiol-induced NBCe1-A expression was enhanced in luminal and granular epithelial cells of uteri [146]. Recently, NBCn1 present in the apical membrane of endometrial cells was also implicated in the balance between HCO\(_3^-\) absorption and secretion [147] (Table 1 and Figure 5). However, in both males and females, the functional relationship between each transporter in maintenance of reproductive process needs to be elucidated more extensively.
Figure 4. Male reproductive regulation with bicarbonate transporters. Spermatogenesis in sertoli cells induced by movement of bicarbonate ions through NBCe1, NBCn1, AE2, and NDCBE.

Figure 5. Female reproductive regulation with bicarbonate transporters. CBE (SLC26A6) and NBC (NBCn1 and NBCe1-A) transporters play an important role in maintenance of the pH of endometrial cells.

Table 1. Identified HCO$_3^-$ transporters and their function in various systems.

| Physiological/Pathological System | Transporters | Localization and Function | References |
|----------------------------------|--------------|---------------------------|------------|
| Immune                           | NBCn1        | Macrophage differentiation | [76,77]    |
|                                  | NBCe1, NBCn1 | Neutrophils, maintenance of intracellular pH | [78,79] |
|                                  | AE2          | CD8$^+$ T cells, controlling cell proliferation | [80]      |
| Tumorigenesis                    | NBCe1        | Colon/breast cancer, inducing cell proliferation | [31]      |
|                                  | NBCn1        | Development and motility of breast cancer | [83,84,101] |
|                                  | AE3, DRA     | Pancreatic ductal adenocarcinoma | [94]      |
|                                  | AE2          | Colon cancer, promotion of cell proliferation | [96]      |
Table 1. Cont.

| Physiological/Pathological System | Transporters | Localization and Function | References |
|-----------------------------------|--------------|---------------------------|------------|
| Tooth development                 | NBCe1B–E, AE2 | Ameloblasts in maturation stage, enamel development | [104,105] |
|                                  | SLC26A1, SLC26A7 | Maturation-stage rodent ameloblasts, enamel formation | [106] |
|                                  | NBC, AE       | Human dental pulp stem cells, pH modulation | [108] |
| Vascular smooth muscle            | NBCn1         | Vascular smooth muscle cells, myogenic tone, regulation of blood pressure, migration, arterial remodeling | [109], [110], [111], [112–114] |
| Cardiac                           | NBCe1, NBCn1  | Cardiac hypertrophy | [120] |
| Digestive                         | NBCe1-B, NBCe1-C | Proximal colon | [129] |
|                                  | NBCn1-C, NBCn1-D | Proximal and distal colon | [129] |
|                                  | AE2           | Parietal cells | [130] |
|                                  | SLC26A9       | GI tract | [131] |
| Reproduction                      | CFTR, AE2, NBCe1, NBCn1, NDCBE | Sertoli cells, spermatogenesis | [134,135] |
|                                  | CFTR, DRA, SLC26A6 | Male reproductive duct, maintenance of luminal pH | [133,142–144] |
|                                  | NBCn1, SLC26A4, SLC26A6 | Endometrial cells | [145,147] |
|                                  | NBCe1-A       | Luminal and granular epithelial cells of uteri | [146] |

5. Future Perspectives and Challenges

Various aspects of ion homeostasis and \( \text{HCO}_3^- \) transporters in pH regulation and multiple cellular functions have been addressed in physiological and pathological systems. Throughout the body, impaired \( \text{HCO}_3^- \) transport is associated with various diseases. However, the precise regulatory mechanisms are relatively unexplored and need to be identified for several systems. We summarized the expression and function of \( \text{HCO}_3^- \) transporters and their associated CAs in various systems. Although all vital systems exhibit differential roles in maintaining their homeostasis, fundamental principles of transporter mechanism have emerged. Various splice variants of \( \text{HCO}_3^- \) transporters exist. Moreover, the supportive role of CAs on \( \text{HCO}_3^- \) transporters needs to be identified precisely. The combined modulation of ion transporters and CAs also needs to be elucidated and specified. In this study, understanding the correlation between these systems could be helpful in obtaining new insights into molecular \( \text{HCO}_3^- \) signaling mechanisms and the potential therapeutic strategies.

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