Simultaneous Determination of 17 Pesticide Residues in Rice by GC/MS using a Direct Sample Introduction Procedure and Spiked Calibration Curves

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Abstract

A reliable, rapid and accurate method based on spiked calibration curves and direct sample introduction was developed for determination of 17 pesticide residues in rice by gas chromatography-mass spectrometry single quadrupole selected ion monitoring GC/MS-SQ-SIM. Sample preparation is based on extraction with acetonitrile without clean up. The use of spiked calibration curves for constructing the calibration curve substantially reduced adverse matrix-related effects. The average recovery of pesticides at 6 concentration levels was in range of 97.5-102.1%. The method was proved to be repeatable with RSDr in range of 0.7%-19.8% for all of the concentration levels. The limits of detection and limit of quantifications for all the pesticides were < 10 ng/g and < 25 ng/g, respectively. The developed method was applied for simultaneous determination of the selected pesticides in 23 rice samples collected from Tehran retail market in March 2009.

Although many studies have been conducted regarding the determination of pesticides by using GC-MS, this is the first attempt in Iran using GC-MS-SIM technique that successfully can determine 17 pesticides with difference in physicochemical properties in rice.

Keywords: Pesticides; Spiked calibration curve; GC/MS; Rice.
in March 2009, were purchased and analyzed according to the validated method.

**Experimental**

**Materials and Methods**

**Chemicals**

All pesticide standards were purchased from Dr. Ehrenstorfer Co. (Augsburg, Germany). All organic solvents, intended for extraction, were at least of LC grade and purchased from Merck (Darmstadt, Germany). Bulk quantities of anhydrous Na$_2$SO$_4$ and NaCl were obtained from Merck (Darmstadt, Germany).

**GC–SQ/MS**

An Agilent Technologies 6890N Network GC System chromatographs (Wilmington, USA) with a SQ detector and equipped with an Agilent 7683B autosampler (Agilent technologies, USA) was used. A HP-5 capillary column (30 m × 0.25 mm I.D., 0.25 μm film thicknesses) was used for separation.

**Calibration standards**

Individual stock standard solutions (1 mg/mL) were prepared in ethyl acetate and stored in dark at -20°C. They were kept for 1 h at ambient temperature prior to their use. A mixed stock standard solution of pesticides was prepared in ethyl acetate at concentrations specified in Table 2. Spiked calibration standards at half maximum residue levels of 0.5 MRLs, 1 MRLs, 1.5 MRLs, 2 MRLs, 2.5 MRLs and 5 MRLs were prepared by addition of 250 μL, 500 μL, 750 μL, 1000 μL, 1500 μL and 2500 μL of mixed standard stock solution respectively to 30 g of blank rice samples in each case. In those pesticides for which no MRL has been set, mixed standard stock solutions were prepared according to Table 2.

The list of selected pesticides along with some of their physicochemical properties and their MRLs are presented in Table 1.

In order to overcome the adverse matrix-related effects, it was decided to make the calibration standards by spiking blank rice samples with certain amounts of pesticides and constructing the calibration curve using these spiked calibration standards.

Sample preparation

For sample preparation, an aliquot of 20
μL of internal standard solution (1000 mg/L) was added to 30 g of rice sample in a waring blender and after being left for 1.5 h at ambient temperature in dark, 120 mL acetonitrile was added. The mixture was blended at high speed for 1 min. Six grams of NaCl was added to the mixture and blending was continued for an additional 60 sec. The slurry was transferred to a proper centrifuge tube and the residue in blender was rinsed with 50 mL acetonitrile and added to the centrifuge tube. After centrifugation for 7 min at 3000 rpm in - 5°C, the supernatant was filtered through 15 g of Na$_2$SO$_4$ and the filtrate evaporated to dryness. The residue was reconstituted in 1 mL toluene and 1 μL of the solution was injected into gas chromatograph.

Recovery studies
For recovery determination, spiked rice samples at concentration levels of 0.5, 1, 1.5, 2, 2.5 and 5 MRLs were prepared in triplicates and then treated according to the procedure described previously. The recoveries were calculated using the spiked calibration curves.

GC-SQ-MS analysis
The GC-SQ-MS was employed with helium as the carrier gas at a constant flow of 1 mL/min. The oven temperature started at 75°C and remained at this temperature for 3 min, then increased to 180°C at 25°C/min ramp rate and finally, increased to 300°C at 5°C/min ramp, holding at 300°C for 10 min. Injection port was adjusted at 250°C and splitless mode was used.

After acquisition of the total ion chromatogram for the mixed stock standard solutions in scan mode, peaks were identified by their retention time and mass spectra. The identification was confirmed by comparing the relative abundances for three ions (one quantifier and two qualifiers) of the experimental standards to known relative abundances of the Pest Library reference spectra. The most abundant ion that showed no evidence of chromatographic interference and had the highest signal-to-noise ratio was taken for quantification purposes. A GC–SQ–MS chromatogram of 17 pesticides and internal standard (TPM) is shown in Figure 1.

Quantitation
The concentration of pesticides was determined by interpolation of the relative peak areas for each pesticide to internal standard peak area in the sample on the spiked calibration

| No. | Compound             | Structural group | M.F.          | M.W.  | MRL (ppm) |
|-----|----------------------|------------------|---------------|-------|-----------|
| 1   | Dichlorvos           | Organophosphorus | C$_3$H$_7$Cl$_2$O$_4$P | 220.98 | -----     |
| 2   | Diazinon             | Organophosphorus | C$_3$H$_7$N$_2$O$_3$PS | 304.35 | 0.20      |
| 3   | Carbaryl             | Carbamate        | C$_6$H$_9$NO  | 201.22 | 1.00      |
| 4   | Fenitrothion         | Organophosphorus | C$_3$H$_7$NO$_3$PS | 277.2  | 1.00      |
| 5   | Fipronil             | Phenylprazole    | C$_3$H$_7$O$_3$F$_2$NOS | 437.2  | 0.01      |
| 6   | Malathion            | Organophosphorus | C$_3$H$_7$O$_3$PS | 330.36 | -----     |
| 7   | Oxadiazon            | Oxadiazole       | C$_7$H$_9$Cl$_2$N$_2$O$_3$ | 345.22 | 0.02      |
| 8   | Primicarb            | Carbamate        | C$_7$H$_9$N$_2$O$_2$ | 238.00 | -----     |
| 9   | *Chlorpyrifos        | Organophosphorus | C$_7$H$_7$Cl$_3$NO$_3$PS | 350.59 | 0.50      |
| 10  | *Chlorpyrifos-methyl | Organophosphorus | C$_7$H$_7$Cl$_3$NO$_3$PS | 322.5  | 0.10      |
| 11  | Fenthion             | Organophosphorus | C$_7$H$_7$O$_3$PS | 278.33 | 0.05      |
| 12  | Propiconazole        | Triazole         | C$_7$H$_7$Cl$_3$N$_2$O$_3$ | 342.00 | 0.05      |
| 13  | Propargite           | Sulfite ester    | C$_7$H$_7$O$_3$S | 350.00 | -----     |
| 14  | Edifenphos           | Organophosphorus | C$_7$H$_7$O$_3$PS | 310.4  | 0.10      |
| 15  | Fenvalerate          | Pyrethroid       | C$_7$H$_7$CNO$_3$ | 419.92 | -----     |
| 16  | Pirimiphos-methyl    | Organophosphorus | C$_7$H$_7$N$_2$O$_3$PS | 305.00 | 1.00      |
| 17  | Iprodione            | Dicarboximide    | C$_7$H$_7$Cl$_3$N$_2$O$_3$ | 330.17 | 10.00    |

Table 1. Physicochemical properties and MRLs of the selected pesticides.
curve. In order to compensate for losses during sample processing and instrumental analysis, internal standard (TPM) was used.

Application to real samples
Twenty-three rice samples were purchased from local markets of Tehran in March 2009. In order to avoid any possible thermal decomposition of pesticide residues, 200 g rice sample was milled with Romer mill (Stylemaster Drive, USA) with 100 g dry ice. A 30 g portion of the powder was subjected to the process of sample preparation described previously.

Results

Gas chromatographic determination
Analysis was performed in the SIM mode based on the use of one target and two qualifier ions. Pesticides were identified according to their retention times, target and qualifier ions. The quantitation was based on the peak area ratio of the targets to that of internal standard. Table 3 summarizes pesticides studied with their target and qualifier ions used in SIM mode in this study.

Method validation
Linearity of the calibration curves
Calibration curves were constructed for each compound using six different concentration levels. TPM was used as internal standard. For identification of pesticides, the retention time and three ions (one for quantitation and two for identification) were used.

The 17 pesticides showed linearity in SIM mode. Linear spiked calibration curves for all the pesticides were obtained with correlation factors more than 0.997. Table 4 shows calibration data (equation and regression coefficient) of interest pesticides in spiked rice calibration curves. The spiked calibration curve of chlorpyrifos in rice

| Concentrations (μg/mL) |
|------------------------|
| 0.6 | 1.2 | 3 | 6 | 12 | 30 | 60 |
| Fipronil | Oxadiazon | Fenthion | Propiconazole |
| Dichlorvos | Primicarb | Malathion | Edifenphos |
| Fenvaerate | Propargite | Chlorpyrifos | Chlorpyrifos-methyl |
| Diazinon | Chlorpyrifos |
| Carbaryl | Iprodione | Fenitrothion |
| Primiphos-methyl |

Figure 1. Representative chromatograms of 17 pesticides and internal standard.
Simultaneous Determination of 17 Pesticide Residues in Rice by GC/MS

Limits of detection and limits of quantification

Limits of quantification (LOQs) of the proposed method were calculated by considering a value 10 times that of background noise in spiked rice samples. The LOQs for all the pesticides in this method were calculated < 25 ppb.

Recovery

Table 5 presents the recovery and repeatability of 6 concentration levels. The recovery of pesticides at 6 concentration levels was in range of 97.48 to 102.15%. The method was proved to be repeatable with RSDr in range of 0.7% to 19.8% at all spiking levels. The recoveries and repeatabilities are in accordance to the criteria set by SANCO Guideline (25).

Real samples

For application to real samples, twenty-three rice samples were purchased from local markets of Tehran in March 2009 and analyzed according to the method described above. For evaluation of analysis, one QC sample at 1 MRL level was carried out in each working round. One of the 23 samples showed contamination with diazinon, pirimiphos-methyl and chlorpyrifos at concentrations of 0.10, 0.02, 0.09 ppm, respectively, which were below the MRLs of these pesticides in Iran. Figure 3 shows the overlaid chromatogram of a spiked rice sample at 1 MRL levels (a) and contaminated rice sample (b) in SIM mode.

Discussion

Matrix-induced response enhancement is seemingly the most discussed matrix effect negatively impacting quantitation accuracy of certain analytes in GC (12). When a real sample is injected, the blocked of active sites (especially free silanol groups) in the GC inlet and column by the matrix components reduces losses of susceptible analytes caused by adsorption or degradation on these active sites. This phenomenon results in larger analyte signals in matrix-containing solutions in comparison to the matrix containing free solutions, which makes the convenient use of calibration standards in solvent only impractical. This would lead to overestimations of the calculated concentrations.

| No. | Compound       | Retention time | Diagnostic ions | Quantification ions |
|-----|----------------|----------------|-----------------|--------------------|
| 1   | Dichlorvos     | 7.66           | 185., 109, 220  | 109                |
| 2   | Diazinon       | 11.87          | 304, 276, 179   | 304                |
| 3   | Primicarb      | 12.45          | 238.2, 166.1    | 166.1              |
| 4   | Chlorpyrifos-methyl | 13.15      | 286, 125, 323   | 286                |
| 5   | Carbaryl       | 13.29          | 144, 125.9, 115.1 | 144               |
| 6   | Fenitrothion   | 13.87          | 277, 260, 214   | 277                |
| 7   | Pirimiphos-methyl | 13.88         | 305, 290, 276.1 | 305                |
| 8   | Malathion      | 14.03          | 285, 173, 158   | 173                |
| 9   | Fenthion       | 14.320         | 278, 169, 262.9 | 278                |
| 10  | Chlorpyrifos   | 14.478         | 314, 197, 257.8 | 314                |
| 11  | TPM (Istd)     | 14.699         | 244, 165        | 244, 165           |
| 12  | Fipronil       | 15.89          | 367, 420, 351   | 367                |
| 13  | Oxadiazon      | 17.548         | 344.1, 302, 258 | 258                |
| 14  | Iprodione      | 19.57          | 244.1, 187, 161 | 187                |
| 15  | Edifenphos     | 19.784         | 310, 200.9, 173 | 310                |
| 16  | Propiconazole  | 19.89, 20.28   | 259, 190.9, 172.9 | 259           |
| 17  | Propargite     | 20.433         | 350.2, 335.2, 201.1 | 350.2          |
| 18  | Fenvalerate    | 28.13, 28.42   | 419.2, 225.1, 167 | 225.1          |
Another potential problem associated with matrix injections involves gradual accumulation of nonvolatile matrix components in the GC system, resulting in formation of new active sites and gradual decrease in analyte responses. This effect, sometimes called matrix-induced diminishment (26) negatively impacts ruggedness (i.e., long-term repeatability of analyte peak intensities, shapes, and retention times), which is a highly important factor in routine GC analysis (17).

Theoretically, elimination of matrix components or active sites would surmount the matrix-induced enhancement effect; but, complete and permanent GC system deactivation or comprehensive sample cleanup is practically impossible (26, 27, 28).

Since an efficient elimination of the sources of the matrix is not possible in practice, the analysts are required to compensate for the effect using alternative calibration methods. The current compensation approaches include the use of the followings: (A) matrix-matched standards, (B) standard addition method, (C) isotopically labeled internal standards (not feasible in multiresidue pesticide analysis due to their unavailability or high price) and (D) usage of analyte protectants (18).

In the present study, we used spiked calibration standard approach to overcome the problems caused by the matrix. In this approach, calibration standards are prepared by the addition of standard solution to blank rice samples that are subjected to the same sample preparation procedure which is intended to be used for unknown samples. In this way, the standard sample matrices will have the same composition as the unknown samples.
and therefore the effect of matrix is reflected in both standards and unknown samples. The calibration curve is constructed using these spiked calibration standards and it is easily used to calculate the concentration of analyte(s) in unknown sample without being concerned about the matrix effects.

**Conclusion**

A simple and rapid method was developed to determine 17 pesticide residues in rice; a main food in Iranian food basket.

The proposed method not only allowed the simultaneous determination and confirmation of 17 pesticides with good recoveries and low detection limits, but also showed to be useful in routine analysis due to its fast and easy procedure.

The developed method has the advantage of using spiked calibration curves that minimize the matrix interferences leading to higher accuracy for pesticides analyses.

**Acknowledgment**

The authors are very grateful to the Food and Drug Control Labs (FDCL) for their financial supports.

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**Table 5.** Average recoveries (%) and range of relative standard deviations (%) obtained by GC-MS analysis of rice at 6 spiking levels (n=3) in rice samples.

| Compound            | 0.5MRL | 1MRL | 1.5MRL | 2MRL | 2.5MRL | 5MRL | Total Recovery (%)(n=18) | Range of RSD (%) |
|---------------------|--------|------|--------|------|--------|------|--------------------------|------------------|
| Dichlorvos          | 95.6   | 108.5| 99.5   | 104.3| 103.3  | 99.5 | 101.79                  | 2.4 - 16.7       |
| Diazinon            | 102.4  | 101.3| 94.9   | 100.0| 102.4  | 99.7 | 100.16                  | 3.9 - 10.8       |
| Primicarb           | 87.2   | 99.7 | 100.0  | 97.5 | 107.1  | 98.7 | 98.40                   | 1.7 - 19.3       |
| Chlorpyrifos-methyl | 103.6  | 98.8 | 95.9   | 98.6 | 104.1  | 99.5 | 100.12                  | 3.4 - 13.7       |
| Carbaryl            | 85.7   | 97.3 | 102.6  | 99.8 | 104.8  | 98.8 | 98.19                   | 4.6 - 10.7       |
| Fenitrothion        | 117.8  | 104.4| 93.4   | 97.4 | 98.8   | 100.9| 102.13                  | 2.6 - 7.5        |
| Pirimiphos-methyl   | 116.3  | 101.2| 94.3   | 95.0 | 102.9  | 100.4| 101.69                  | 0.8 - 9.1        |
| Malathion           | 104.5  | 105.4| 85.8   | 101.7| 105.1  | 99.4 | 100.35                  | 2.5 - 19.1       |
| Fenithion           | 80.2   | 104.8| 106.7  | 101.7| 96.7   | 99.9 | 98.36                   | 3.3 - 15.9       |
| Chlorpyrifos        | 103.0  | 103.9| 98.2   | 98.4 | 99.9   | 100.3| 100.62                  | 2.1 - 9.3        |
| Iprodione           | 88.9   | 113.2| 100.6  | 99.7 | 99.2   | 99.8 | 100.25                  | 2.7 - 10.7       |
| Fipronil            | 91.7   | 109.5| 99.1   | 92.7 | 104.6  | 99.8 | 99.58                   | 4.9 - 16.1       |
| Oxadiazon           | 121.8  | 100.0| 92.7   | 93.2 | 104.7  | 100.3| 102.15                  | 3.5 - 15.5       |
| Edifenphos          | 107.5  | 105.4| 95.3   | 94.2 | 103.3  | 100.2| 100.00                  | 3.1 - 14.9       |
| Propiconazole       | 117.8  | 94.8 | 93.7   | 98.4 | 103.7  | 99.9 | 101.40                  | 3.6 - 11.8       |
| Propargite          | 77.9   | 96.7 | 107.6  | 102.1| 101.6  | 98.9 | 97.48                   | 1.4 - 19.8       |
| Fenvalerate         | 110.6  | 104.7| 105.9  | 89.8 | 95.7   | 101.9| 101.45                  | 0.7 - 5.5        |
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