A Cryptic Species of the *Tylonycteris pachypus* Complex (Chiroptera: Vespertilionidae) and Its Population Genetic Structure in Southern China and nearby Regions

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Abstract

Three distinct bamboo bat species (*Tylonycteris*) are known to inhabit tropical and subtropical areas of Asia, i.e., *T. pachypus*, *T. robustula*, and *T. pygmaeus*. This study performed karyotypic examinations of 4 specimens from southern Chinese *T. p. fulvidus* populations and one specimen from Thai *T. p. fulvidus* population, which detected distinct karyotypes (2n=30) compared with previous karyotypic descriptions of *T. p. pachypus* (2n=46) and *T. robustula* (2n=32) from Malaysia. This finding suggested a cryptic *Tylonycteris* species within *T. pachypus* complex in China and Thailand. Morphometric studies indicated the difficulty in distinguishing the cryptic species and *T. p. pachypus* from Indonesia apart from the external measurements, which might be the reason for their historical misidentification. Based on 623 bp mtDNA COI segments, a phylogeographic examination including *T. pachypus* individuals from China and nearby regions, i.e., Vietnam, Laos, and Cambodia, was conducted to examine the population genetic structure. Genealogical and phylogeographical results indicated that at least two diverged lineages existed in these regions (average 3.4 % of Kimura 2-parameter distances) and their population structure did not match the geographic pattern. These results suggested that at least two historical colonizations have occurred by the cryptic species. Furthermore, through integration of traditional and geometric morphological results, morphological differences on zygomatic arches, toothrows and bullae were detected between two lineages in China. Given the similarity of vegetation and climate of Guangdong and Guangxi regions, we suggested that such differences might be derived from their historical adaptation or distinct evolutionary history rather than the differences of habitats they occurred currently.

Key words: cryptic species, karyotypic examination, morphometric studies, population genetic structure, *Tylonycteris*.

Introduction

Bamboo bats (genus *Tylonycteris*, Chiroptera, Vespertilionidae) are a group of small bats with a remarkably flattened braincase, small size, light weight, and unique well-developed thumbpads at the bases of the first fingers and flat footpads on hindfeet. They have the peculiar habit of roosting inside hollow
bamboo stems. Their dental formula is \( \frac{2113}{32} \times 2 = 32 \) [1-3]. Three species are recognized at present, which are found in tropical and subtropical Asia and the Amboi Islands, i.e., \( T. pachypus \) (Temminck, 1840) [1], \( T. robustula \) Thomas, 1915 [2], and \( T. pygmaeus \) Feng et al. 2008 [3]. Among them, greater bamboo bat (\( T. robustula \)) is the largest species with dark and more gray pelage, and \( T. pygmaeus \), a recently recognized species, is the smallest one in \( Tylonycteris \), while the lesser bamboo bat (\( T. pachypus \)) is a medium-sized species with a distinct golden brown pelage [3, 4]. Although several forms of \( T. pachypus \) have been attributed historically, e.g., \( T. fulvidus \), \( T. meyeri \), and \( T. aurex \) [4], after examinations of their external and skull features Tate assigned them to a single \( T. pachypus \) complex [5]. Subsequently, in consideration of their smaller size relative to \( T. robustula \) and their distinct supraorbital tubercles, all members of this complex were condensed into one species, namely \( T. pachypus \) [4, 6-11]. Nowadays, five subspecies of \( T. pachypus \) are proposed [3, 4, 12, 13]: \( T. p. pachypus \) (Temminck, 1840) originally described from Java [1]; \( T. p. fulvidus \) (Blyth, 1859) from Burma [14]; \( T. p. meyeri \) Peters, 1872 from Philippines [15]; \( T. p. aurex \) Thomas, 1915 from Bombay, India [2]; \( T. p. bhaktii \) Oei, 1960 from Lombok [16]. Individuals from southern China, Vietnam, Laos, Thailand, and Burma are often referred to as \( T. p. fulvidus \), which is a large-sized form within the \( T. pachypus \) that is similar to \( T. p. aurex \) from southern India, although it has a more drab brown appearance and a relatively shorter forearm [3, 4, 12, 13]. However, many puzzles are still unsolved, including the validity of this subspecies, karyotype of \( T. p. fulvidus \), morphological differences between \( T. p. pachypus \) and \( T. p. fulvidus \) from China, and their population genetic structure.

In this study, a karyotypic examination of \( T. p. fulvidus \) individuals from two Chinese populations (Guangdong and Guangxi provinces) and one Thai population was conducted, and a cryptic species was identified by its unique chromosomal characteristics when compared with previous study that referred to the karyotype of other \( Tylonycteris \) [3]. Secondly, a phylogeographic analysis was performed for the examination of population genetic structure of \( T. p. fulvidus \) from China and nearby regions including Vietnam, Laos, and Cambodia. Thirdly, morphometric comparisons using individuals of \( T. p. fulvidus \) from China and \( T. p. pachypus \) from Malaysia as well as the \( T. p. fulvidus \) individuals from two distinct lineages were conducted to investigate their morphological differences.

**Material and methods**

**Sampling**

Twenty-four specimens of \( T. p. fulvidus \) were collected from Guangdong and Guangxi provinces in China between 2000 and 2012 (Table 1). All of the voucher specimens were adult stage according to the stage of epiphyseal–diaphyseal fusion, and were further preserved in 70 - 100% ethanol and deposited at the College of Life Sciences, Guangzhou University, Guangdong, for further morphometric and phylogeographic analyses. To analyze the karyotype, \( T. p. fulvidus \) were collected in Guangdong province (2 males, 1 female) and in Guangxi province (1 male), and \( T. robustula \) was collected in Guangxi province (2 males, 5 females). In addition, one \( T. p. fulvidus \) was collected from Chiangmai, Thailand on 1982 (no. 11276).

**Table 1.** Haplotype diversity, nucleotide diversity, and haplotypes for different populations of \( Tylonycteris pachypus fulvidus \).

| Population (China) | N | Vouch number of specimen | Number of haplotypes (±SD) | Haplotype diversity (±SD) | Nucleotide diversity (±SD) | Haplotype (label of related individuals) |
|-------------------|---|--------------------------|---------------------------|--------------------------|---------------------------|----------------------------------------|
| Guangdong         | 16| 2000156, 04353 1/2, 06219, 09339 1/2, 09340, 09343, 01001 1/2, 02001, 0437 1/2, 0439, 0440, 2000-09, 10199 1/2, 12173, 12174 1/2, 12175 | 4 | 0.81±0.13 | 0.003±0.002 | TP-H5, TP-H6, TP-H7, TP-H17 |
| Guangxi           | 8 | 10221, 10222, 10223, 10224, 10225, 10226 1/2, 10227 1/2, 10230 | 5 | 0.93±0.12 | 0.007±0.005 | TP-H2, TP-H8, TP-H9, TP-H14, TP-H16 |
| Vietnam           | - | - | 6 | - | 0.017±0.010 | TP-H1, TP-H2, TP-H3, TP-H4, TP-H9, TP-H10 |
| Laos              | - | - | 4 | - | 0.016±0.011 | TP-H9, TP-H11, TP-H13, TP-H15 |
| Cambodia          | - | - | 1 | - | - | TP-H12 |

*Specimens that were amplified successfully are labeled in bold.
Karyotypic analyses

Chromosomal preparations were made from tail and lung tissue cultures following Harada and Yosida [17]. Differential staining using the G-band and C-band techniques was applied following Seabright [18] and Sumner [19], respectively. The nomenclature of chromosomes followed Levan et al. [20]. The diploid number (2n) and the total number of autosomal arms (FN) were determined by observing 30 metaphase cells in each specimen.

Phylogeographic analyses

All of the specimens were used in the phylogeographic analyses to infer the population genetic structure of T. p. fulvidus from China, although only 7/15 Guangdong specimens and 6/8 Guangxi specimens were amplified and sequenced successfully (Table 1). In addition, three T. robustula specimens from Guangxi were also included in a genealogic analysis in this study (no. 08004, 07345 and 07346). Genomic DNA was isolated from approximately 20 mg of muscle tissue using a Universal Genomic DNA Extraction Kit (TAKARA). A partial segment of mtDNA cytochrome oxidase subunit I gene (COI), an acknowledged barcoding region used for species and subspecies identification, was amplified by polymerase chain reaction (PCR) using the primers that deposited in NBCI-nt: F (5’- TGT AAA ACG ACG GCC AGT TCT CAA CCA ACC ACA AAG ACA TTG G -3’) and R (5’- CAG GAA ACA GCT ATG ACT AGA CTT CTG GGT GCC CAA AGA ATC A -3’). PCRs were performed using a final volume of 50 μl, which contained approximately 20 mg of muscle tissue using a Universal Genomic DNA Extraction Kit (TAKARA). A partial segment of mtDNA cytochrome oxidase subunit I gene (COI), an acknowledged barcoding region used for species and subspecies identification, was amplified by polymerase chain reaction (PCR) using the primers that deposited in NBCI-nt: F (5’- TGT AAA ACG ACG GCC AGT TCT CAA CCA ACC ACA AAG ACA TTG G -3’) and R (5’- CAG GAA ACA GCT ATG ACT AGA CTT CTG GGT GCC CAA AGA ATC A -3’). PCRs were performed using a final volume of 50 μl, which contained approximately 5.0–50 ng DNA, 0.2 mM of each dNTP, 0.4 mM of each primer, 1.5 mM MgCl2, and 2.0 U Taq polymerase (TAKARA), using the manufacturer’s buffer. Amplification was performed using a MyCycler Thermal Cycler (BioRad) as follows: 94C for 4 min; 37 cycles at 94C for 30 s, 50C for 30 s, and 72C for 1 min; and 72C for 5 min. DNA sequencing was performed using an ABI PRISM 3700 DNA Analyzer (Applied Biosystems). The chromatograms were checked and edited using GENEIOUS [21] and aligned with MUSCLE [22]. All sequences of T. pachyppus and T. robustula were blasted against the NCBI-nt database with GENEIOUS [21] and only matching sequences with a max score value of >1,000 were included in subsequent phylogeographic and genealogic analysis.

To verify the monophyly of Tylonycteris species and T. p. fulvidus Chinese population, all matching sequences from nearby regions were included, i.e., sequences from Vietnam, Laos, and Cambodia (GenBank Accession Numbers: GU684765, GU684774, GU684806, HM541981-HM541989, HM914916, HM914920, HM914921, GU684753, GU684758, GU684766, GU684779, GU684781, HM541990-HM542004, HM914929, and HM914947). The genealogical relationships were reconstructed among all uniquely identified sequences, using the COI segments from Myotis yumanensis and Pipistrellus subflavus (GenBank accession numbers GU723138 and GU723142) as outgroups. A maximum likelihood (ML) phylogeny was reconstructed with RAxML V7.2.7 [23] using 500 bootstrap replicates via the CIPRES Science Gateway V3.1 [24]. A neighbor-joining (NJ) tree was constructed with MEGA 4 [25] using the approximate model selected by Modeltest 3.06 [26]. A Bayesian analysis was conducted with MrBayes 3.1.2 [27]. Convergence was indicated when the standard deviation (SD) of the split frequencies was < 0.01 [27]. Chains were sampled every 2,000 generations. The starting trees were generated randomly and the prior probability indicated that all trees were equally likely. Divergence times were estimated with COI sequences using Bayesian MCMC as implemented in BEAST [28]. Because no direct estimates of COI mutation rate are available for bats and no appropriate fossil record that could be used for divergence time estimation, we estimated minimum and maximum divergence times using two substitution rates, 2% and 5% per million years [29]. Similar estimates of 2.6% for phyllostomid bats [30], 2.3–5% in Carolinia [31], and 4% from fossil calibrations [32] have been suggested. The nucleotide substitution model was the same as that used for phylogenetic analysis. Two MCMC chains were run for 10,000,000 generations with a burnin of 2,500,000. The means and 95% confidence interval (CI) of the divergence times were estimated from two samplings using Tracer v.1.4.1 [33], and the trees were summarized using TreeAnnotator v.1.4.8 [28].

The haplotype number, haplotype diversity (h), and nucleotide diversity (π) were calculated using ARLEQUIN 3.01 [34]. A hierarchical analysis of molecular variation (AMOVA) [34] was performed in ARLEQUIN using 1,000 permutations to estimate the partitioning of genetic variation among different populations of T. p. fulvidus. To overcome the pitfalls of traditional phylogenetic reconstruction in intra-species phylogenetic studies and to explore the phylogeographic history of T. p. fulvidus, a haplotype network was also constructed using the statistical parsimony method [35] in TCS [36].

Morphometric analyses

The following external and cranial measurements were taken using electronic vernier calipers (Guogen, Serial number: 00000315) during the morphometric analyses: length of the hind foot (HF), ear length (E), forearm length (FA), tibia length (TB),...
length of third digit metacarpal (III'), length of fourth digit metacarpal (IV'), length of fifth digit metacarpal (V'), greatest length of skull (GLS), condylobasal length (CBL), condylocanine length (CCL), height of braincase (HBC), breadth of braincase (BBC), height of occipital (HOC), occipital condyle width (OW), zygomatic width (ZW), interorbital breadth (IOB), palatal bridge length (PBL), upper tooth row (C-M3), width of the crowns of the upper canines (C1-C1), width of the crowns of the upper molars (M3-M3), lower tooth row (C-M3), and the mandibular length (MDL). A principal component analysis (PCA) was performed for each type of traits (external and cranial measurements) of 20 T. p. fulvidus specimens from the Guangdong and Guangxi populations, as well as 3 T. p. pachypus specimens from Komodo and Java, Indonesia, to compare the overall similarities in their external and cranial characteristics. The variation between different variables from two Chinese populations (lineages) was analyzed using Student’s t-test. All statistical analyses were performed using SPSS Statistics 17.0 (SPSS Inc. 2000).

Through geometric morphometric analysis, size and shape variations were used to discriminate between two distinct lineages of T. p. fulvidus from China. A Nikon D80 camera with AF MICRO NIKKOR 60mm 1:2.8 D lens, mounted on a tripod at a fixed distance of 10 cm from the skull (which was always mounted on graph paper), was used to take dorsal, lateral and mandibular images for skulls from our samples. Landmark placement and further analysis were performed using the thin plate spline (TPS) series of programs [37-39]. The program tpsDig version 2.1 [40] was used to capture landmarks in two dimensions for dorsal (10 landmarks) ventral (34 landmarks) and mandibular views (29 landmarks) (Fig. 1A-C, for a more detailed landmark description see Appendix I). In order to superimpose the data, landmarks were subjected to Generalized Procrustes Analysis (GPA) which removes variation in digitizing location, orientation, and scale, and superimposes the objects in a common coordinate system [41] using the Morphologika2 software program (version 2.5) [42]. The size difference between two lineages was tested using t-test basing on the centroid size (CS) [43] of each individual that obtained using Geometric Morphometrics Tools Package (GMTP) version 2.1 [44]. To elucidate the shape differences between two lineages, a Principal Components Analysis (PCA) was employed in Morphologika2 (version 2.5) [42] to calculate principal axes of variation. Discriminant functions were generated from the samples from two different populations on the basis of PC scores from PCA. A Discriminant Analysis with cross-validations was carried out to assess the power of the discriminant function in SPSS Statistics 17.0 (SPSS Inc. 2000). Thin plate splines were also produced for a visual representation of the morphological differences in the skull between lineages using Morphologika2 (version 2.5) [42].
Results

Distinct karyotypes of *T. p. fulvidus* from China and Thailand

Chromosome numbers of *T. p. fulvidus* from Guangdong and Guangxi of China were 2n=30, FN=56 (Table 2; Fig. 2A-D), which was identical with the karyotypic features from Thai sample. There were eight large metacentric or submetacentric pairs, two pairs of small submetacentrics, four pairs of medium to small subtelocentrics in the autosomes, with a medium-sized subtelocentric X and a small, submetacentric Y chromosome (Table 2; Fig. 2A-D). One autosome pairs of subtelocentric chromosome (no. 12) had secondary constrictions adjacent to the centromere (Fig. 2A-D). The chromosome number of *T. robustula* from Guangxi province was 2n=32, FN=52 (Table 2; Fig. 2E). There were eight metacentric or submetacentric pairs, one pair of small submetacentrics, two pairs of subtelocentrics and four pairs of acrocentrics in the autosomes, with a medium-sized acrocentric X chromosome (Table 2; Fig. 2E). The diploid number, the placement of centromeres, and the size of the biarmed elements of *T. p. fulvidus* from China and Thailand differed from the chromosomes of *T. p. pachypus* (2n=46) and *T. robustula* (2n=32) from the Malayan Peninsula [45] and Guangxi province (Table 2).

Table 2. Karyotype descriptions for *Tylonycteris pachypus pachypus* (Temminck, 1840), *T. p. fulvidus* (Blyth, 1859) and *T. robustula* Thomas, 1915.

| Species/subspecies                          | 2n | FN | Pairs of autosomes | Sex chromosomes | Reference            |
|---------------------------------------------|----|----|--------------------|-----------------|----------------------|
| *Tylonycteris pachypus* (Malayan peninsula) | 46 | 56 | 4                  | 2 A             | M Yong et al. (1971) |
| *Tylonycteris robustula* (Malayan peninsula) | 32 | 56 | 11                 | 2 A             | M Yong et al. (1971) |
| *Tylonycteris robustula* (Guangxi, China)   | 32 | 52 | 9                  | 2 A             | M present study      |
| *Tylonycteris pachypus fulvidus* (Guangdong, China) | 30 | 56 | 10                 | 4 ST           | M present study      |
| *Tylonycteris pachypus fulvidus* (Guangxi, China) | 30 | 56 | 10                 | 4 ST           | M present study      |
| *Tylonycteris pachypus fulvidus* (Chiangmai, Thailand) | 30 | 56 | 10                 | 4 ST           | M present study      |

* M, meta/submeta-centric; ST, subtelocentrics; A, acrocentric.

Figure 2. The karyotypes of *Tylonycteris pachypus fulvidus* from Guangdong and Guangxi, and *T. robustula* from Guangxi analyzed in this study. Conventional (A, no. 2000156), G-banded (B, no. 2000156), and C-banded (C, no. 2000156) karyotypes of *T. p. fulvidus* from Guangzhou, Guangdong province, China; conventional (D, no. 10230) karyotype of *T. p. fulvidus* from Chongzuo, Guangxi province, China; conventional (E, no. 10229) karyotype of *T. robustula* from Chongzuo, Guangxi province, China.
Phylogeny and population genetic structure

The monophyly of *T. pachypus* and *T. robustula* was well supported using the most commonly used phylogenetic methods (NJ, ML, and Bayes) (Fig. 3A), and the splitting event should have occurred during Pliocene or Pleistocene period (1.35-4.99 Myr) (Fig. 3A-B). The blast results obtained from the NCBI-nt database showed that the COI sequences of the *T. pachypus* specimens from nearby regions were very similar to the haplotypes from the Chinese populations. Furthermore, some were actually identical (Table 1). Two divergent lineages of *T. p. fulvidus* were emerged (Figs. 3A and 4), and their splitting event was estimated to be around 0.24 - 1.21 Myr ago according to our divergence time estimations (Fig. 3A-B). The intertwined relationships among haplotypes and their similar external features (see subsequent sections for full details) suggested that individuals from the regions should be the same as the cryptic species found in China. Note that one *T. robustula* haplotype (GenBank accession numbers: HM914921) clustered into the *T. pachypus* lineage (Fig. 3A). Such phenomenon may be derived from the contamination of samples in molecular experiment, the misidentification of the specimens, or the incomplete lineage sorting. However, due to difficulty in checking the specimens and discriminating the potential causes, this haplotype was excluded in our sequent phylogeographic analysis.

Based on 623 base pairs from the partial mitochondrial cytochrome oxidase subunit 1 (COI) gene, 17 unique haplotypes were identified from the Chinese and nearby populations of *T. pachypus* (Figs. 3A and 4; Table 1). Thirty-nine polymorphic sites were detected, but no insertions or deletions were found among the haplotypes. Haplotypes TF-H5, TF-H6, TF-H7, and TF-H17 were found only in the Guangdong population, while haplotype TF-H9 were found within the Guangxi, Laos, and Vietnam populations (Figs. 3A and 4; Table 1). Using a traditional regional group setting (Chinese group vs. Southeast Asia group), the variances among groups within populations and among populations within groups were -14.60% and 54.87% (*P*<0.01), respectively (Table 3). Using a no group setting, the variances among populations (52.09%, *P*<0.01) and within populations (47.91%) were roughly the same as the regional group setting (Table 3). However, when the groups were arranged according to the genealogical result that detected two diverged lineages in these regions, the variance among groups (lineages) increased to 64.30% (*P*<0.05) (Figs. 3A and 4; Table 3).

![Figure 3](http://www.ijbs.com)

**Figure 3.** Genealogical reconstruction of *Tylonycteris pachypus fulvidus* from China and nearby regions and the divergence time of related nodes. (A) Biogeographical distribution of haplotypes identified in this study and their phylogenetic relationships based on Bayesian, maximum likelihood (ML), and neighbor-joining (NJ) methods. The values on the nodes represented the posterior probabilities from BEAST, MrBayes, the ML bootstrap values, and the NJ bootstrap values, respectively. Bayesian estimates of divergence time used 2 fixed substitution rates of 2% per million years and 5% per million years. Geographical distributions of major groups were mapped onto the phylogenetic trees and the haplotypes of specimens used in the karyotypic analysis are highlighted in the light gray and gray bars. The *T. robustula* haplotype (GenBank accession numbers: HM914921) that clustered into the *T. pachypus* lineage is labeled in bold. (B) Summarized table of Bayesian estimates of divergence time of related nodes using two fixed substitution rates.
The statistical parsimony haplotype network of COI also produced two divergent cladograms (where over 14 mutation steps were required for connection), which resembled the relationships detected by phylogenetic reconstruction (Figs. 3A and 4). Sequence divergence between the two lineages ranged from 3.0% to 4.2% of Kimura 2-parameter model distances (average 3.4%) for COI segment. These results were consistent with the AMOVA results, which indicated that most of the genetic variance was attributable to variance among lineages. Our haplotype network and phylogenetic topologies indicated the monophyly of Guangdong population, whereas an intertwined relationship of haplotypes was found in the Guangxi and southern Vietnam and Laos populations, i.e., Lao Cai, Tuyen Quang, Ha Noi, Nam Khan, and Khammouan (Figs. 3A and 4).

| Structure                      | Source of variation       | Variation (%) | Fixation indices | P     |
|-------------------------------|---------------------------|---------------|------------------|-------|
| China group vs. Southeast Asia group | Among regions             | -14.60        | -0.18            | 0.7   |
|                               | Among populations/within regions | 54.87        | 0.57             | <0.01* |
|                               | Within populations        | 59.87         | 0.49             | <0.01* |
| Two diverged lineages         | Among lineages            | 64.3          | 0.64             | <0.05* |
|                               | Among populations/within lineages | 22.22        | 0.62             | <0.01** |
|                               | Within populations        | 13.48         | 0.87             | <0.01** |
| No group                      | Among populations        | 43.2          | 0.52             | <0.01** |
|                               | Within populations        | 56.8          |                  |       |

*Statistically significant (P < 0.05), **statistically significant (P < 0.01).
NS, nonsignificant.

**External and cranial size differences**

The external and cranial measurements collected in this study are provided in Table 4. The PCA analysis of the external characteristics showed that the eigenvalues of the first two principal components were 3.64 and 1.47, respectively, which explained 73.01% of the total variance. Plots of principal components 1 and 2 indicated that the *T. p. pachypus* could not be distinguished from the *T. p. fulvidus* (Fig. 5A). The eigenvalues of the first three principal components for the cranial measurements were 6.39, 3.87, and 2.61, respectively, which explained 85.79% of the total variance (Table 5). The number of specimens from Indonesia was limited in this study, but the 3D plots of principal components 1, 2, and 3 detected differences between *T. p. fulvidus* from China and *T. p. pachypus*. 

Figure 4. TCS network of COI haplotypes for the *Tylonycteris pachypus fulvidus*. The size of each circle is proportional to the frequency of the specific haplotype used in this study.
from Indonesia. The first principal component was strongly correlated with CBL, CCL, BBC, OW, ZW, and IOB, which might reflect a skull size effect (Table 5). The second principal component was strongly correlated with tooth measurements, such as C-M3, M3-M3, and C-M3, while the third principal component was strongly correlated with PBL and HOC (Table 5). The plots derived from the Guangdong specimens were difficult to distinguish from the Guangxi specimens (Table 5), but the pairwise comparison (t-test) of the Guangdong and Guangxi specimens showed that the BBC, C-M3, M3-M3, and C-M3 measurements were significantly different ($P < 0.05$), and the specimens of Guangdong are slightly larger than the Guangxi specimens (Table 4).

![Figure 5](http://www.ijbs.com) **Figure 5.** Principal components plots based on the external and cranial of *Tylonycteris pachypus* measurements. (A) Principal components plots of the external measurements showing components 1 and 2. (B) principal components 3D plots of cranial measurements for *T. p. fulvidus* showing components 1, 2, and 3. The contribution of each axis for total variation is indicated in parenthesis.

**Table 4.** External and cranial measurements (mm) of *Tylonycteris pachypus* used in this study.

| T. p. fulvidus (Guangdong) | n   | T. p. fulvidus (Guangxi) | n   | *P* | T. p. pachyapus (Indonesia) | n   |
|----------------------------|-----|--------------------------|-----|-----|-----------------------------|-----|
| FA                        | 26.07±0.89 (24.86–27.86) | 12  | 25.96±0.83 (24.56–27.00) | 8   | 0.98 | 26.01±0.94 (25.09–26.97) | 3   |
| E                         | 6.80±0.93 (4.90–8.34)    | 9   | 6.79±0.52 (5.88–7.50)    | 8   | 0.79 | 5.68±0.20 (5.15–5.90)    | 3   |
| II0                       | 24.33±0.66 (23.58–25.83) | 12  | 24.50±0.80 (23.41–25.38) | 8   | 0.19 | 24.66±1.27 (23.23–25.66) | 3   |
| IV0                       | 24.07±0.60 (23.30–25.20) | 12  | 24.20±0.69 (23.34–24.93) | 8   | 0.34 | 24.22±0.87 (23.22–24.80) | 3   |
| V0                        | 23.66±0.60 (22.82–24.96) | 12  | 23.86±0.67 (22.79–24.66) | 8   | 0.36 | 24.23±1.05 (23.03–24.93) | 3   |
| HF                        | 6.15±0.48 (4.22–5.72)    | 11  | 4.67±0.45 (4.00–5.52)    | 8   | 0.11 | 5.32±0.14 (5.19–5.46)    | 3   |
| TB                        | 11.25±0.45 (10.33–11.81) | 12  | 11.17±0.67 (9.94–11.93)  | 8   | 0.72 | 10.79±1.11 (9.91–12.04)  | 3   |
| GLS                       | 11.29±0.30 (10.65–11.65) | 12  | 11.06±0.22 (10.72–11.44) | 8   | 0.06 | (10.49, 11.01)           | 2   |
| CBL                       | 11.08±0.27 (10.58–11.45) | 12  | 10.84±0.20 (10.55–11.09) | 8   | 0.61 | (8.86, 9.08)             | 2   |
| CCL                       | 10.68±0.25 (10.12–10.99) | 12  | 10.45±0.21 (10.05–10.69) | 8   | 0.11 | 10.84±1.18 (9.63–12.20)  | 4   |
| HBC                       | 3.61±0.22 (3.26–3.97)    | 12  | 3.49±0.10 (3.31–3.61)    | 8   | 0.28 | (3.56, 3.74)             | 2   |
| BBC                       | 6.67±0.17 (6.39–6.98)    | 12  | 6.56±0.10 (6.40–6.70)    | 8   | 0.06 | 6.55±0.17 (6.40–6.80)    | 4   |
| HOC                       | 1.14±0.04 (1.06–1.21)    | 12  | 1.07±0.10 (0.93–1.20)    | 8   | 0.06 | (3.17, 3.18)             | 2   |
| OW                        | 4.33±0.10 (4.19–4.52)    | 12  | 4.26±0.16 (3.97–4.44)    | 8   | 0.29 | (3.79, 3.93)             | 2   |
| ZW                        | 7.82±0.27 (7.41–8.25)    | 12  | 7.94±0.10 (7.76–8.05)    | 8   | 0.57 | 8.15±1.21 (7.19–9.50)    | 3   |
| IOB                       | 3.42±0.09 (3.23–3.55)    | 12  | 3.45±0.08 (3.36–3.55)    | 8   | 0.88 | (3.17, 3.23)             | 2   |
| PBL                       | 3.82±0.28 (3.48–4.27)    | 12  | 3.97±0.20 (3.61–4.16)    | 8   | 0.29 | (4.77, 4.99)             | 2   |
| C-M3                      | 3.88±0.14 (3.36–3.87)    | 12  | 3.41±0.06 (3.32–3.50)    | 8   | <0.01| 3.71±0.35 (3.31–4.00)    | 4   |
| C1-C2                     | 3.71±0.19 (3.45–4.08)    | 12  | 3.57±0.10 (3.44–3.68)    | 8   | 0.06 | (3.08, 3.35)             | 2   |
| M3-M3                     | 5.10±0.16 (4.75–5.50)    | 12  | 4.86±0.13 (4.70–5.04)    | 8   | <0.01| (4.61, 5.07)             | 2   |
| C-M4                      | 4.00±0.12 (3.73–4.20)    | 12  | 3.73±0.10 (3.60–3.84)    | 8   | <0.01| (3.88, 4.01)             | 2   |
| MDL                       | 8.13±0.17 (7.86–8.34)    | 12  | 7.99±0.17 (7.73–8.27)    | 8   | >0.07| (7.48, 7.69)             | 2   |

*P-value for t-test comparisons between Guangdong and Guangxi specimens.*
Table 5. Factor loadings, eigenvalues, and the variance explained by each principal component based on the cranial measurements of *Tylonycteris pachypus fulvidus* examined in this study.

| Variable | PCA 1  | PCA 2  | PCA 3  |
|----------|--------|--------|--------|
| GLS      | 0.27   | 0.77   | -0.23  |
| CBL      | 0.84   | 0.22   | -0.48  |
| CCL      | 0.89   | 0.24   | -0.07  |
| HBC      | 0.57   | 0.16   | 0.69   |
| BBC      | 0.88   | 0.31   | 0.28   |
| HOC      | -0.51  | -0.09  | 0.83   |
| OW       | 0.89   | 0.17   | -0.18  |
| ZW       | 0.85   | 0.01   | 0.05   |
| IOB      | 0.93   | -0.02  | -0.09  |
| PBL      | -0.01  | -0.21  | 0.93   |
| C-M³     | 0.19   | 0.88   | -0.01  |
| C¹-C¹    | 0.43   | 0.70   | -0.34  |
| M³-M³    | 0.05   | 0.91   | -0.14  |
| C-M³     | -0.03  | 0.90   | 0.20   |
| MDL      | 0.93   | 0.26   | -0.06  |
| Eigenvalues | 6.39   | 3.87   | 2.61   |
| % of variance explained | 42.60 | 25.78 | 17.41 |

Cranial differences between two lineages

Geometric morphometric analysis confirmed earlier traditional morphometric results and indicated the *T. p. fulvidus* from Guangdong were significantly larger than Guangxi specimens (dorsum: $\ln CS_{Guangzhou}=7.16$, $\ln CS_{Guangxi}=7.13$, *t*-test $P=0.02$; ventrum: $\ln CS_{Guangzhou}=7.55$, $\ln CS_{Guangxi}=7.53$, *t*-test $P=0.05$; mandible: $\ln CS_{Guangzhou}=7.08$, $\ln CS_{Guangxi}=7.03$, *t*-test $P<0.01$). Both the Mahalanobis and Procrustes distances of the dorsum, ventrum and mandible between the lineages mean were significant ($D=9.95$, $P=0.02$ and $d=0.02$, $P<0.01$ for the dorsum; $D=2.72$, $P=0.01$ and $d=0.03$, $P<0.01$ for the ventrum; $D=3.75$, $P<0.01$ and $d=0.03$, $P<0.01$ for the mandible). The number of the axes required to explain 99% of the overall shape variance was 13 principal components (PC) for dorsum, 16 for the ventrum and 16 for the mandible. The first two PC accounted for 29.10% and 17.13% of the overall shape variance of dorsum, 24.18% and 13.30% for the ventrum, 24.28% and 18.50% for the mandible. From the PC plots of the dorsum, ventrum and mandible, two lineages could be mostly separated (Fig. 6A-C). At the cross-validation of the discriminant function, 7 out of 16 specimens, 4 out of 18 specimens and 3 out of 19 specimens were misclassified at the dorsum, ventrum and mandible, respectively. As regards the shape changes, Guangzhou population has an elongated zygomatic arches, and wider toothrows of the upper jaw, elongated bullae, and larger and longer toothrows of the mandible (Fig. 6A-C).

Figure 6. Plots of principal components factors 1 and 2 for the dorsal (A) and ventral (B) sides of the cranium, and the labial side of the mandible (C), as well as the thin plate splines (TPS) of *Tylonycteris pachypus fulvidus* from Guangdong and Guangxi, China.
Discussion

Karyotypic information is one of the most valuable data used in systematic and genetic studies of bats [46]. It is important for identifying cryptic species, clarifying debates in taxonomic settings, assessing the relationships between taxa, and studying the processes of speciation and evolution [46]. The chromosomes of bats have been studied and reviewed extensively, and taxon samplings have increased continuously, but the karyotypic data available for many species and subspecies are still limited. The present chromosomal study of T. p. fulvidus from China and Thailand detected a cryptic species within the T. pachypus complex, which was identified based on its diploid number, the placement of centromeres, and the size of biarmed elements compared with the karyotypes of T. p. pachypus and T. robustula from Malay Peninsula (Table 3). The chromosomal rearrangements among them supported the existence of distinct Tylonycteris cryptic species and reproductive isolation among recognized subspecies. Because, if a species/subspecies becomes subdivided into two geographic forms, one of them is characterized by a series of chromosomal rearrangements and the hybrids between these geographic forms are expected to experience reduced fertility due to the meiotic difficulties caused by the heterozygosity of the rearrangements [47-49].

An accepted hypothesis for the karyotypic evolution of vesperilionids bats is conservatism in the chromosome arms and Robertsonian translocation due to fusion of the whole long arms of two acrocentric chromosomes, which resulted in a decrease in the chromosomal diploid number [50-52]. If so, T. p. pachypus should be a primitive form within Tylonycteris because it contains higher chromosomal diploids number than any other reported Tylonycteris species (Table 2), whereas cryptic species from Southeast Asia may be a more derived form that emerged following Robertsonian fusion events involving subacrocentric or/and acrocentric autosomes. However, this hypothesis needs to be validated using other chromosome staining methods, such as fluorescence in situ hybridization (FISH) [53-55]. The assumed evolutionary scenario within Tylonycteris also raises another puzzling question about their phylogenetic relationship and phenotypes because T. p. pachypus from Southeast Asia is more similar to the cryptic species found in this study in terms of its external features than T. robustula [3]. In a broader sense, the external and life habit similarities of Tylonycteris species suggest that they are excellent subjects for studies of speciation based on karyotypic organization and genetic variations, as well as the dependence of genetic mechanisms on phenotypic constraints.

High genetic similarities of the related COI segments of T. pachypus specimens from Laos, Cambodia, and Vietnam, as well as individuals from China, the intertwined phylogenetic relationships, and their similar external features suggest that the specimens from Laos, Cambodia, and Vietnam should be attributed as the same cryptic species from China and Thailand. The similar external features and pelage of T. p. fulvidus from China and T. p. pachypus from Indonesia [3, 12] could be a major cause of the historical failure to distinguish them (Figs. 1A-E and 5A; Table 4). This phenomenon might have occurred because the long-term convergent evolution of Tylonycteris species constrained their external differentiation due to their similar and specialized habitats and ecotypes [3, 5, 10, 12]. The results of the multivariate analyses of the cranial measurements showed that although the skull and tooth size characteristics of the 2 species overlapped, cryptic species from China could be distinguished from T. p. pachypus based on HOC and PBL, where PC 3 was strongly correlated (Fig. 5B). These measurements may be critical for the identification of T. pachypus subspecies in future.

The contrasting patterns of the haplotype relationships, distribution, and branching provide insights into the evolutionary processes that have shaped the population genetic structure of T. p. fulvidus in China and nearby regions. The AMOVA analysis and phylogeographic reconstruction both detected a population genetic structure that did not match the geographic distribution, and this common pattern is often observed in small mammals characterized by ecomorphological traits that limit dispersal [56, 57]. The coexistence of two divergent lineages (average 3.4% of Kimura 2-parameter model distances for COI segment) in these regions suggests a complex evolutionary history for this cryptic species with multiple historical colonization events (at least twice) in China. All of the haplotypes from Guangdong population, which formed a single clade, were grouped with haplotypes from Cambodia and populations from the far south of Vietnam (e.g., Attapu, Binh Phuoc, Lam Dong, Kaoh Kong, and Ho Chi Minh city), rather than nearby populations (Figs. 3A and 4). This pattern might be explained by a historical long-distance colonization from southern Asia to Guangdong and the extinction of transitional populations, or biogeographical rearrangements due to the environmental changes of Pleistocene [58]. The intertwined haplotypes of the clade inhabiting Guangxi, northern Vietnam, and Laos might be attributed to historical colonization by a single ancestral population containing diverse lineages or frequent gene flow among...
populations. Given their ecomorphological traits implying limited dispersal capability [56, 57], however, it is suggested that a lineage derived from a diverse ancestral population may be more likely. Both the traditional and geometric morphological analyses detected size and shape differences between two divergent lineages. Considering the fact that most of variations are related to zygomatic arch, upper jaw, mandible and bullae (Figs. 5B and 6A-C), these differences may be resulted from the ecological feature regarding foraging, echolocation, varied diets, and distinct evolutionary history [59-67]. It is worthwhile to note that since the similar latitude and close distance between Guangdong and Guangxi, they share similar sub-tropic vegetation and climate [68]. In consideration of the distinct evolutionary scenarios of two lineages in present study, we suggested that such difference might be derived from their historical adaptation or evolutionary history rather than the differentiation due to the habitat they occurred currently. However, to confirm this hypothesis and clarify the causes of cranial differences, a comparative study on diet and echolocation of two lineages is required in the future.

This study integrated karyotypic, morphometric, and phylogeographic data to recognize a distinct cryptic species within T. pachypus. Its primary phylogeographic framework in China and nearby regions, and morphometric differences between two lineages were also studied. The results indicated that all T. p. fulvidus specimens from China and nearby regions belong to a newfound cryptic species. Similarities in their external features may be a major cause of historical misidentifications. This study also addressed the question of the source of the species diversity found within the Tylonycteris complex, which appears to have undergone long-term convergent evolution. Cryptic species may be a significant problem that affects many taxonomic groups in bats [69-73]. It is required to clarify the species boundary and distribution range of T. pachypus complex, especially in the continental Asia attributed to T. p. fulvidus, T. p. pachypus, and T. p. aurex.

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**Competing Interests**

The authors have declared that no competing interest exists.

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