A Narrative Review on LI-RADS Algorithm in Liver Tumors: Prospects and Pitfalls

Federica De Muzio 1, Francesca Grassi 2, Federica Dell’Aversana 2, Roberta Fusco 3,*, Ginevra Danti 4,5, Federica Flammia 4,5, Giuditta Chiti 4,5, Tommaso Valeri 6,7, Andrea Agostini 6,7, Pierpaolo Palumbo 5,8, Federico Bruno 5,9, Carmen Cutolo 10, Roberta Grassi 2,5, Igino Simonetti 11, Andrea Giovagnoni 6,7, Vittorio Miele 4,5,*, Antonio Barile 9, and Vincenza Granata 11

Department of Medicine and Health Sciences V. Tiberio, University of Molise, 86100 Campobasso, Italy; demuziofederica@gmail.com
Division of Radiology, Università degli Studi della Campania Luigi Vanvitelli, 81100 Naples, Italy; francesca.grassi1@studenti.unicampania.it (F.G.); federica.dellaversana@studenti.unicampania.it (F.D.); roberta.fusco@polincianapoli.it (R.G.)
Medical Oncology Division, Igea SpA, 80013 Naples, Italy
Division of Radiology, Azienda Ospedaliera Universitaria Careggi, 50134 Florence, Italy; ginevra.danti@gmail.com (G.D.; federicalflammia91@gmail.com (F.F.); giudittachiti@gmail.com (G.C.); vmiele@sirm.org (V.M.)
Italian Society of Medical and Interventional Radiology (SIRM), SIRM Foundation, 20122 Milan, Italy; palumbopierpaolo89@gmail.com (P.P.); federico.bruno.1988@gmail.com (F.B.)
Department of Clinical Special and Dental Sciences, University Politecnica delle Marche, 60126 Ancona, Italy; t.valeri@univpm.it (T.V.); a.agostini@staff.univpm.it (A.A.); a.giovagnoni@univpm.it (A.G.)
Department of Radiological Sciences, University Hospital Ospedali Riuniti, Via Tronto 10/a, 60126 Torrette, Italy
Area of Cardiovascular and Interventional Imaging, Department of Diagnostic Imaging, Abruzzo Health Unit 1, 67100 L’Aquila, Italy
Emergency Radiology, San Salvatore Hospital, Via Lorenzo Natali 1, 67100 L’Aquila, Italy; abarile63@gmail.com
Department of Medicine, Surgery and Dentistry, University of Salerno, 84084 Fisciano, Italy; carmencutolo@hotmail.it
Radiology Division, Istituto Nazionale Tumori-IRCCS-Fondazione G. Pascale, Via Marniano Semmola, 80131 Naples, Italy; igino.simonetti@istitutotumori.na.it (I.S.); v.granata@istitutotumori.na.it (V.G.)
* Correspondence: r.fusco@igeamedical.com

Abstract: Liver cancer is the sixth most detected tumor and the third leading cause of tumor death worldwide. Hepatocellular carcinoma (HCC) is the most common primary liver malignancy with specific risk factors and a targeted population. Imaging plays a major role in the management of HCC from screening to post-therapy follow-up. In order to optimize the diagnostic-therapeutic management and using a universal report, which allows more effective communication among the multidisciplinary team, several classification systems have been proposed over time, and LI-RADS is the most utilized. Currently, LI-RADS comprises four algorithms addressing screening and surveillance, diagnosis on computed tomography (CT)/magnetic resonance imaging (MRI), diagnosis on contrast-enhanced ultrasound (CEUS) and treatment response on CT/MRI. The algorithm allows guiding the radiologist through a stepwise process of assigning a category to a liver observation, recognizing both major and ancillary features. This process allows for characterizing liver lesions and assessing treatment. In this review, we highlighted both major and ancillary features that could define HCC. The distinctive dynamic vascular pattern of arterial hyperenhancement followed by washout in the portal-venous phase is the key hallmark of HCC, with a specificity value close to 100%. However, the sensitivity value of these combined criteria is inadequate. Recent evidence has proven that liver-specific contrast could be an important tool not only in increasing sensitivity but also in diagnosis as a major criterion. Although LI-RADS emerges as an essential instrument to support the management of liver tumors, still many improvements are needed to overcome the current limitations. In particular, features that may clearly distinguish HCC from cholangiocarcinoma (CCA)
and combined HCC-CCA lesions and the assessment after locoregional radiation-based therapy are still fields of research.

Keywords: liver; diagnosis; LI-RADS

1. Introduction

Liver cancer is the sixth most detected tumor and the third leading cause of tumor death worldwide [1–7]. Hepatocellular carcinoma (HCC) is the most common primary liver malignancy (more than 80%), with a higher prevalence in male patients (3:1 M/F) and in developing countries (72% in Asia compared to 9.8% and 5% in Europe and the USA, respectively) [2]. At present, chronic liver disease is the main cause of HCC development, with viral cirrhosis remaining the primary cause of carcinogenesis [8]. Indeed, HBV and HCV chronic infection are still responsible for 56 and 20% of deaths from liver cancer, respectively [1,2]. However, in the last few decades, due to an increase in obesity and type 2 diabetes, also nonalcoholic fatty liver disease (NAFLD) is becoming an emerging cause of HCC (annual incidence of approximately 2.4–12.8%), especially in the Western world [9–14]. Other risk factors include environmental exposure to aflatoxins, anabolic steroids, oral contraceptives and tobacco and alcohol abuse [1]. Finally, albeit in a small percentage of cases, some benign lesions such as adenomas (5% of all cases) may undergo neoplastic degeneration [15–17].

Imaging plays a major role in the management of HCC from screening to post-therapy follow-up [18–39]. In particular, ultrasonography (US) is the method of choice for tumor screening, while the role of computed tomography (CT) and magnetic resonance imaging (MRI) for HCC diagnosis and post-treatment assessment is largely consolidated in clinical practice [3,40–61].

In order to optimize diagnostic accuracy and using a universal report that allows more effective communication among the multidisciplinary team caring for the patient, several classification systems have been proposed over time. Of these, the most valid and comprehensive is the Liver Imaging Reporting and Data System (LI-RADS), which provides guidance on all aspects of HCC imaging, from techniques for imaging acquisition to assessing treatment response and directing management. The latest version of ACR LI-RADS includes four algorithms: 
(a) US-LI-RADS for HCC screening and surveillance; 
(b) contrast-material-enhanced US for HCC diagnosis (CEUS-LI-RADS); 
(c) CT/MRI for HCC diagnosis and radiologic T stage (CT/MRI LI-RADS); and 
(d) CT/MRI for HCC treatment response assessment (TR LI-RADS) [62–65]. This system was conceived to emphasize positive predictive value and specificity in the diagnosis of HCC. On the other hand, unlike other diagnostic algorithms, LI-RADS includes an LR-M category that aims to preserve specificity and increase sensitivity in the definition of malignant lesions other than HCC [12,16]. Non-HCC malignancies include a broad spectrum of pathologies and some of them, such as intrahepatic cholangiocarcinoma (iCCA) or combined hepatocellular and cholangiocarcinoma (cHCC-CCA), are being recognized in patients with the same risk factors of HCC (chronic hepatitis or cirrhosis) [16,23,39]. Thereby, the correct differential diagnosis is essential for appropriately targeting treatment.

The aim of this narrative review was to evaluate the main current applications of LI-RADS in liver cancer, primarily in HCC and to assess the main radiological features that can guide differential diagnosis.

2. LI-RADS

LI-RADS is a system supported by the American College of Radiology (ACR), resulting from the work of radiologists experienced in liver pathology and integrated with the latest guidelines of the American Association for the Study of Liver Diseases (AASLD) [66]. The first version, proposed in 2011, aimed to standardize imaging and reporting on CT and MRI
examinations in patients at risk of HCC. Based on scientific evidence, this version has been updated introducing other algorithms, which cover the entire diagnostic and therapeutic management of HCC patients [3].

Currently, LI-RADS comprises four algorithms addressing screening and surveillance, diagnosis on CT/MRI, diagnosis on contrast-enhanced ultrasound (CEUS) and treatment response on CT/MRI. Each algorithm has a core document that includes relevant indications to be applied in clinical practice [64]. A specific manual (LI-RADS manual) details the complex aspects of liver diseases, including imaging parameters, reporting instructions, templates and management recommendations. In order to adopt a universal language both in daily clinical practice and research, LI-RADS has published even a standardized lexicon, simplifying the communication between the different experts involved in HCC management [64,67]. The main focus of the LI-RADS system is to achieve high specificity and positive predictive value (PPV) in HCC detection and diagnosis, avoiding false-positive examinations. To ensure a high PPV, LI-RADS should be employed only in populations with a high pre-test probability of disease [3]. At present, LI-RADS and the American Association for the Study of Liver Diseases (AASLD) are in agreement in defining the target population at risk of HCC as those adult patients (≥18 years old) with cirrhosis, in the absence of current or prior HCC (incidence of HCC exceeds 1.5%/year) and subsets of patients with chronic HBV infection in whom the incidence of HCC exceeds 0.2% per year. In addition to the criteria for defining the high-risk screening population, patients with evidence of HCC or previous HCC are included in the diagnostic algorithm [67].

Both US-LI-RADS and diagnostic LI-RADS do not address patients with vascular causes of liver cirrhosis (e.g., cardiac hepatopathy, Budd–Chiari syndrome) because these conditions are associated with hypervascular benign liver lesions, which increase the risk of false positives and reduce the PPV for the diagnosis of HCC [67–81]. Although with some regional differences, patients with hepatitis C in the absence of cirrhosis and adults with nonalcoholic steatohepatitis are not included in the screening. Furthermore, patients in Child–Pugh class C who are not candidates for transplantation are excluded, considering their poor life expectancy [2].

An important point of the LI-RADS philosophy is the choice to adopt the term “observation” when referring to identified liver formations during the screening or diagnostic phase [67]. The generic term “observation” embraces the whole spectrum of possible abnormalities, ranging from simple cysts to clear neoplastic lesions [82–94]. In addiction, LI-RADS guidelines suggest the imaging techniques that could be used, according to the different diagnostic phases and the availability. At present, CT and MRI are the most widely used imaging modalities in the Western world, while the Asian-Pacific, Japanese and Chinese guidelines recommend CEUS for the diagnosis of HCC [95].

2.1. US-LI-RADS

The main purpose in the management of patients with HCC is to detect the disease at an early stage, when it is potentially treatable by resection or liver transplantation. Indeed, the average 5-year survival of advanced and untreated cases is only 15% [96]. US is proposed as a tool for cancer screening, and it has already been widely demonstrated in other oncological settings [97–100]. With regard to HCC, several studies showed the benefits in terms of reduced mortality and median survival of using US surveillance, particularly every six months in patients at risk of HCC with or without alpha-fetoprotein assay [101]. US shows a sensitivity in detecting HCC at an early stage of 63% and at any stage of 78–94% [102] and a specificity of 89% [103].

The ACR working group developed a standardized algorithm for US screening and surveillance in high-risk patients (US-LI-RADS) [67,104]. US-LI-RADS provides the classification of liver observations into three categories according to the probability of HCC, including an evaluation of the overall quality of the examination to define its reliability [67,104].
2.1.1. Technique

In order to achieve high diagnostic accuracy, the ACR US-LI-RADS working group illustrates some guidance on the technical execution of the US examination [104–108]. The examination should be performed using primarily a convex probe (Figures 1 and 2), possibly combined with a linear transducer to explore the liver margins. Technical parameters have to be adjusted considering the single patient habitus to allow adequate penetration of the ultrasound beam and optimize the acoustic window [108]. Power and color Doppler are helpful in assessing vascular structures and any signs of vascularization of intraparenchymal presumed alterations [108].

![Figure 1. HCC on VIII seg. B mode assessment (A,B). The lesion shows inhomogeneous structure (arrow).](image1)

![Figure 2. HCC on VII seg. US (A) and CEUS assessment. The lesion (arrow) shows APHE during arterial phase (B) with washout during portal (C) and late (D) phase of contrast study.](image2)

Each observation should be assessed in three dimensions in the longitudinal and transverse plane, reporting their precise location and anatomical relationships. Optional cine clips may facilitate the comparison in follow-up exams [108].

2.1.2. Ultrasound Category and Visualization Score

The US-LI-RADS algorithm includes both an evaluation score of the observations and an analysis of the quality of the images. The three possible categories (US-1, US-2 and US-3) should be applied to the entire examination, and each category corresponds to a subsequent clinical management [104–108].
The US-1 category includes the detection of clear benign formations (simple cysts, focal areas of fat sparing or hemangiomas), leading the patient to routine follow-up in 6 months [109].

The US-2 category is assigned when not definitely benign <10 mm formations are detected. Close US follow-up at 3–6 months is recommended to intercept an overthreshold-old growth (>1 cm), which requires further diagnostic characterization. The stability or subthreshold size in two years, redeploy the patient in the US surveillance program every six months [109].

US-3 corresponds to a positive examination for lesions >10 mm that are probably HCC. In these circumstances, a contrast-enhanced CT, MRI or US is mandatory [109].

During the examination, the radiologist should also pay attention to identifying a new thrombus in a vein, which could be a sign of tumor invasion. The second component of US-LI-RADS is the visualization score, which is an assessment of three categories (A-B-C) of the overall quality and/or perceived sensitivity of the exam [104,109]. Although the visual score should be used to indicate the expected level of sensitivity of a screening and surveillance US examination, it does not directly influence patient management and remains open territory for research [109].

2.2. CT/MRI LI-RADS

Accurate diagnosis and staging of HCC can be achieved by CT (Figure 3) or MRI (Figure 4) in the absence of invasive methods, when precise and stringent criteria are applied [110–130]. Although MRI proves to be more sensitive in the characterization of liver lesions, particularly with hepatospecific contrast agents, there is no unambiguous indication from the pool of experts [131–148].

![Figure 3. HCC on VIII seg. CT assessment. The lesion (arrow) shows APHE during arterial phase (A) of contrast study with washout in portal phase (B) and capsule appearance (C) in late phase.](image)

The choice to use CT rather than MRI remains at the discretion of individual institutions.

2.2.1. CT/MRI Technique

The LI-RADS guidelines provide information on minimum technical requirements for CT and MRI examinations to be considered reliable. CT examinations should be performed on multidetector devices (≥8 detectors), while a magnetic field of at least 1.5 T is indispensable for MRI [67]. Intravenous contrast medium administration and a multiphasic acquisition are fundamental. Three phases of acquisition—arterial, portal and late phases—are necessary to comply with the LI-RADS requirements [67]. In particular, the arterial phase could be well timed, sufficiently late, to detect one of the hallmarks of HCC, namely the arterial-phase hyperenhancement (APHE). The late phase is acquired at 2–5 min, with some differences when a hepatospecific contrast agent is employed. When using gadoxetate disodium, no conventional late phase is obtained; instead, images are acquired in the transitional and hepatobiliary phases at 2–5 min and 15–20 min after injection, respectively [67]. If gadobenate dimeglumine is used, a conventional delayed
phase is usually acquired at 2–5 min with optional imaging of the hepatobiliary phase at 1–3 h after injection [67].

![Figure 4](image1.png)

**Figure 4.** HCC on II seg. MRI assessment. The lesion (arrow) shows hyperintense signal on T2-W sequence (A), hypo-iso signal on T1 sequences ((B): in phase and (C): out phase) with APHE during arterial phase (D) of contrast study, capsule appearance during transitional phase (E) and hypointense signal in EOB phase (F).

In-phase and out-of-phase T1-weighted basal sequences and T2-weighted sequences (with or without fat suppression) prior to contrast injection are necessarily integrated into the MRI study, while diffusion-weighted imaging (DWI), subtraction sequences and hepatospecific phase images are optionally conducted [67].

Although there is no guidance on the assessment of the severity of artifacts, the radiologist should assign an LR-NC category (LI-RADS not categorizable), when an observation cannot be fully assessed due to missing or degraded images [67].

2.2.2. CT/MRI Categories

LI-RADS includes eight diagnostic categories (-LR), reflecting the probability that an observation is benign, HCC, a neoplasm other than HCC or a tumor in a vein (TIV) [67].

The algorithm is built to guide the radiologist through a stepwise process, recognizing both major and ancillary features.

Currently, LI-RADS major imaging criteria include APHE (Figure 5), washout, capsule appearance, threshold growth and size [67].

![Figure 5](image2.png)

**Figure 5.** HCC on VI seg. MRI assessment. The lesion (arrow) shows APHE during arterial phase (A) with washout and capsule appearance (B) during portal phase of contrast study.
While the major criteria should always be satisfied, ancillary features, such as restricted diffusion (Figure 6), could be used at the discretion of the radiologist to increase or decrease the category, except to upgrade an observation from LR-4 to LR-5 [149–151].

The categories LR-1 (definitely benign) and LR-2 (probably benign) include simple cysts or nodules, <20 mm in size, that do not show features of malignancy. LR-3 (intermediate likelihood) refers to some perfusions or true nodules with one or two major malignant features. Specifically, observations <20 mm could be categorized as LR-3 if they have APHE as the sole major feature. Observations lacking APHE are categorized as LR-3 if they have a size <20 mm and ≤1 additional major feature or size ≥20 mm and no additional major features. In these circumstances, it should be indicated to repeat enhanced diagnostic imaging in 3 to 6 months [67]. The LR-4 observations are probable HCCs. The LR-4 category includes observations <10 mm with nonrim APHE and ≥1 additional major feature; 10 to 19 mm observations with nonrim APHE and “capsule” as the only major feature; observations ≥20 mm with nonrim APHE and no additional major features. The LR-4 category also includes observations without rim APHE: observations <20 mm and ≥2 additional major features and observations ≥20 mm and with ≥1 additional major feature. According to LI-RADS guidelines, LR-4 patients are referred for biopsy, presumptive treatment or follow-up, depending on the multidisciplinary team’s decision [67].

Malignancy categories include LR-5 (definitely HCC), LR-M (malignant lesion other than HCC) and LR-TIV (malignancy with tumor in vein) [67].

APHE and size ≥10 mm are required for LR-5 categorization. The LR-5 category includes observations >20 mm with ≥1 additional major feature and 10 to 19 mm observations with ≥2 additional major features. Observations measuring 10 to 19 mm can also be categorized as LR-5 if they have only one additional major feature: either non-parenchymal “washout” or threshold growth. In the absence of diagnostic doubt, pathologically confirmed lesions and benign lesions of non-hepatocellular origin do not require LI-RADS classification.

CT and MRI also contribute to the staging of liver disease, particularly to the evaluation of possible macrovascular invasion, which falls into the LR-TIV category (Figure 7) and contraindicates the curative surgical approach [13,152].
2.2.3. HCC Diagnosis

On imaging, HCC reveals a distinctive dynamic vascular pattern of APHE followed by washout in portal-venous phase. Due to its high specificity, this temporal enhancement hallmark is incorporated in all current diagnostic systems. As emerged from a recent meta-analysis, the APHE and washout are the strongest independent imaging features associated with HCC [155]. APHE is defined as the presence of nonrimlike enhancement of an observation that is unequivocally of higher intensity/attenuation than background liver [155]. This appearance is referable to the high arterial flow in advanced HCC due to angiogenesis and formation of nontriadal or unpaired neoarteries [156].

Among the LI-RADS major features, APHE shows the highest sensitivity (85%) for progressed HCC, but it is characterized by a low specificity (57%) [157]. Indeed, this feature may also be seen in benign entities such as hemangiomas or perfusion alterations, premalignant lesions such as dysplastic nodules or small non-HCC tumors [158]. Holland et al., in 46 patients with cirrhosis who underwent MRI before liver transplantation, noticed that most of the hypervascular lesions in the arterial phase (93%) were actually benign formations [159].

These findings were confirmed by Granata et al. in 17 hyperplastic nodules that resulted hypervascular during MRI examinations in a retrospective evaluation of 70 patients with HCC [23].

A meaningful number of observations with APHE could potentially be incorrectly classified as HCC; therefore, this criterion alone is considered inadequately accurate [23]. The false-positive rate could be lowered by combining APHE with the “washout” feature [23,160]. The washout of an observation refers to the progressively hypointense/hypodense appearance compared to the surrounding liver parenchyma during the portal-venous phase and the delayed phase [67,161].
This contrast-enhancement behavior corresponds to a reduced portal supply, increased cellularity and reduced extracellular volume of the tumor.

APHE and washout appearance are not a feature exclusive to HCC and could be observed in cirrhotic or dysplastic nodules, raising the issue of specificity when applied as a sole criterion on liver observations [162–172].

Combining detection of APHE and washout could significantly increase specificity, reaching values close to 100% when detected in high-risk patients with nodules $\geq 20$ mm [173].

This advantage is still associated with poor sensitivity, especially in smaller-sized lesions, showing inconspicuous washout [174].

The capsule appearance is the other major criterion that may improve the performance of the LI-RADS algorithm. The “capsule” presence is a distinctive sign of advanced HCC, with the highest specificity value among all major features (90–96%) [23,175]. However, this tumor appearance often coincides with the APHE/washout pattern, limiting its added value in terms of sensitivity [176].

Favoring specificity is positively accepted in Western countries, where the main purpose is to avoid false positives in order to select patients for liver transplantation. On the other hand, in Asian countries, locoregional therapies are often employed in the first instance, and therefore HCC should be intercepted at an early stage. Since LI-RADS is proposed as a universal guide in HCC management, the problem of sub-optimal sensitivity should be solved [177].

2.2.4. Ancillary Features

Ancillary features (AFs) could play a decisive role. Although these imaging criteria alone preclude definitive diagnosis of HCC, they provide additional data for tumor characterization, improving the sensitivity rate of major features. The importance of AFs applied especially to MRI emerges from the literature. Between all AFs, the hypointensity in the hepatospecific phase (HPH) appears independently associated with the diagnosis of HCC [178].

HPH has shown to increase sensitivity in liver observation assessment, even the small ones (10–19 mm). This last clinical scenario is particularly challenging, reaching a failure rate of 87% when only major criteria are identified in small liver lesions [179].

Vernuccio et al. recorded a sensitivity of 84% applying HPH to LR-3 small observations with APHE upgraded to the LR-4 category [180]. This sensitivity value resulted considerably higher compared to those of LI-RADS major criteria (from 0% to 35%) [179].

The signal of an observation in the hepatobiliary phase depends on the expression of the OATP8 transporter, which provides the contrast agent uptake. The OATP8 expression declines during hepatocarcinogenesis, and therefore the assessment of signal intensity in the hepatobiliary phase may guide the detection and characterization of hepatocellular nodules in the cirrhotic liver. The HPH could also be seen in non-hepatocellular benign lesions such as hemangiomas or non-HCC malignancies due to the physiological absence of the OATP8 expression. Therefore, HPH is not sufficiently specific to be used as the sole criterion, but as shown in several studies, it could be associated with more AFs in order to overcome this limitation [180–188]. Cannella et al. observed an increase in specificity (from 86.1% to 96.2%) for HCC detection, combining HPH with three or more AFs favoring malignancy in 155 patients in LR-3 and LR-4 LI-RADS categories [189].

Additionally, Lee et al. observed that the application of AFs both of malignancy in general (mild–moderate T2 hyperintensity and HPH) and favoring HCC in particular (nonenhancing “capsule” and mosaic architecture) to the LR-4 category assigned on the basis of the major criteria significantly increases the sensitivity in the diagnosis of HCC (69.4–76.9% compared with the standard LR-5 66.2%), without impairing specificity (95.3–96.5%) [190].

Several studies have focused on the application of gadobenate dimeglumine (Gd-BOPTA) in the diagnosis of HCC and its possible advantages as a major feature in tumor assessment. Although the diagnostic performance and liver parenchymal enhancement observed during the hepatobiliary phase seem similar for both gadoxetic Gd-EOB and Gd-BOPTA [191,192], some data suggest that Gd-BOPTA exhibits a higher maximum
enhancement value of the hepatic artery, portal and hepatic veins during the respective vascular phases [193].

Therefore, these peculiarities could be useful when applying the LI-RADS algorithm. Cortis et al. demonstrated increases in sensitivity from 53.2% to 75.8% for lesions of ≥20 mm and from 53.5% to 62.1% for lesions of 10–19 mm when HPH on Gd-BOPTA was included as an additional major feature to the typical APHE/washout pattern in cirrhotic patients with hypervascular HCC [194]. According to this evidence, Zhang et al. recorded an increased sensitivity in HPH on Gd-BOPTA inclusion as a major feature, without impact on specificity. Furthermore, the authors surprisingly proved that HPH on Gd-BOPTA could even replace the enhancing capsule as a major criterion in the LI-RADS algorithm, with sensitivity improvement, especially for lesions 10–19 mm in size [195]. Further studies are needed, but HBP images on Gd-BOPTA MRI could be considered an important tool for the HCC characterization of any size.

2.2.5. LR-M Category

Intrahepatic cholangiocarcinoma (iCCA) (Figure 8) and combined hepatocellular-cholangiocarcinoma (cHCC-CCA) (Figure 9) could develop in a cirrhotic liver displaying HCC-like characteristics [12,16].

Figure 8. CCA on VII seg. The lesion (arrow) shows targetoid appearance in T2-W sequence (A) with nonrim APHE (B) and progressive contrast enhancement during late (C) phase of contrast study.

Figure 9. cHCC-CCA on VII seg. The lesions (arrow) show APHE during arterial phase of contrast study (A) without washout in portal phase (B) and progressive contrast enhancement during late phase (C) of contrast study.

In LI-RADS, the LR-M category collects all those observations with features suggestive of malignancy but missing major criteria specific for HCC (classic APHE and washout/capsule appearance). The LR-M features include: rimlike APHE, peripheral washout, liver surface retraction, adjacent biliary obstruction, mixed pattern and infiltrative margin [67].

In several studies, the rimlike APHE emerges as the most common sign suggesting malignancy other than HCC, being present in 50–84% of examined lesions. The second most frequently encountered feature resulted a delayed or progressive central enhancement during PVP or DP after extracellular agent injection, observed in 42–96% of liver
observations [196]. Regarding the washout appearance, the literature is unclear, given the lack of distinction between peripheral lesion washout and HPH. However, these limitations are of minor importance since “washout” is described in a small percentage of non-HCC cases (4–6%) [197–199].

Another ancillary feature associated with the LR-M category is represented by a mild–moderate T2 hyperintensity. Cannella et al. found that a peculiar signal in the T2 sequences, the “targetoid” appearance associated with tumor vascular involvement, could be a key sign of non-HCC primary liver carcinomas (PLCs) [200]. Researchers retrospectively analyzed 165 iCCAs and 74 chHCC-CCAs, with 136 HCCs as controls. All lesions were pathologically proven and prior imaged on contrast-enhanced CT or MRI. Combining targetoid appearance on T2-weighted images and tumor vascular involvement showed high specificity (92–100%) for the diagnosis of non-HCC PLCs, with a good inter-reader agreement [200]. These results are in line with previous studies, which stressed the role of T2-weighted appearance in iCCA detection. In particular, Sheng et al. proved that dynamic enhancement patterns and T2-weighted images signal were the most important MRI discriminants between ICC and atypical small HCC (≤3 cm) in high-risk patients [201].

LR-M features defined by LI-RADS are most closely associated with the imaging appearance of ICC as the second most frequent neoplasm in cirrhotic liver [202]. Histologically, ICC presents a central fibrotic core of imbibed stroma and a predominantly peripheral cellularization/vascularization. This inner stratification reflects in a so-called “targetoid” enhancement pattern on imaging, with a peripheral arterial hyperenhancement in a rimlike aspect, followed by a peripheral washout with a progressive and delayed enhancement of the central core [203,204]. The same targetoid pattern could be appreciated in diffusion-weighted imaging (DWI), in which the outer layer shows more restriction than the central zone. The targetoid organization is also documented during the hepatobiliary phase on MRI: a central retention of contrast, less intense of background liver, associated with a peripheral irregular rim of hypointensity [204]. However, as demonstrated by Kim et al., the contrast enhancement patterns of ICC in cirrhotic liver depend on tumor size. Small ICCs (<3 cm) may not exhibit the typical targetoid vascular pattern, with a difficult differential diagnosis from HCC [205]. Furthermore, the targetoid appearance is not the exclusive prerogative of ICCs. Lee et al. observed that the targetoid pattern showed the highest sensitivity (75.8%) to correctly identifying chHCC-CCA as LR-M in a group of 99 patients with pathologically proven chHCC-CCA or HCC [206]. This evidence was confirmed by Choi et al. Researchers observed in 194 patients with cirrhosis and surgically proven single primary liver cancer that although the targetoid mass features on Gd-EOB were more common in ICCs (39–59%), it could be detected even in chHCC-CCs (9–21%) and HCCs (1–3%) [207]. Therefore, a mass displaying any of the features of dynamic contrast enhancement, hepatobiliary phase imaging or DWI should be generally categorized as LR-M.

In the literature, the targetoid appearance on HPH, the presence of rimlike APHE and peripheral “washout” would seem sufficiently distinctive of ICCs [208,209]. However, it should be emphasized that these are not definitive data, considering that most studies focused on the differential diagnosis between ICC and HCC have compared the characteristics of ICC with atypical HCC in a non-targeted population.

To date, only a few publications have described the imaging appearance of chHCC-CCA and the features that can be distinguished only in these lesions compared to HCC or ICC [12,16]. Several studies have evaluated the possibility of differentiating chHCC-CCA from ICC through the different contrast enhancement patterns. Park et al. divided 82 surgically confirmed chHCC-CCAs lesions into subgroups according to vascular enhancement pattern on Gd-EOB-MRI [210]. Forty-eight lesions showing nonrim APHE were assigned to the hypervascular group, while thirty-four lesions demonstrating rim, peripheral or isoenhancement were assigned to the nonhypervascular group. On pathological specimens, the hypervascular group encountered a major HCC component, poor CC elements of pathological analysis and was associated with increased overall survival after curative surgery than the other group [210]. In a subsequent study, Kim et al. identified in
cHCC-CCAs an intermediate-form APHE, characterized by an irregular distribution with some areas thicker than others. This type of APHE seemed able to distinguish cHCC-CCAs from ICCs [211].

Regarding other liver tumors, Granata et al. [16] assessed the diagnostic performance of LI-RADS in not-at-risk-of-HCC populations, showing that high diagnostic accuracy was obtained by LI-RADS classification between malignant and benign lesions. The presence of AFs could help the radiologist toward a correct diagnosis [16]. However, LI-RADS has a low capability to identify borderline lesions as BT-IPNB and hepatic adenomatosis as benign lesions due to oxaliplatin [16].

2.3. CEUS-LI-RADS

CEUS is a US examination widely accepted in many clinical contexts, performed by injecting intravenously microbubble-based contrast agents (USCAs). USCAs are small (from 1 to 10 µm) biodegradable particles of low-solubility gases (e.g., perfluoropropane, perfluorocarbon or sulfur hexafluoride) covered by a flexible lipid shell [212–217]. This type of contrast agent is confined within the intravascular space (blood pool agents) and almost completely exhaled after 20 min from the injection. Through the application of a specific US imaging mode, the microbubbles produce an echo pulse that is evidenced by tissue signal removal [218].

ACR incorporated CEUS into the LI-RADS system in 2016, driven by working groups of hepatologists and radiologists, who have demonstrated the diagnostic accuracy of CEUS in the characterization of focal liver lesions [67]. Albeit with some substantial differences from the other diagnostic algorithm, the CEUS-LI-RADS model allows a dynamic classification of hepatic focalities in different categories. CEUS provides a real-time image of the vascular behavior of intercepted lesions, eliminating the concept of observation applied in the other fields of the LI-RADS system.

Otherwise, CEUS is a focused exam, being inadequate for staging and response to therapy, lacking the panoramic and standardized visualization of CT/MRI. Considering these premises, CEUS-LI-RADS introduces different diagnostic features, requiring its own algorithm with distinctive unique properties [219].

2.3.1. Technique

US equipment should comprise contrast-enhanced imaging setting, which generally includes a dual-screen and on-screen chronometer.

Prior to enhanced imaging, baseline scans are performed to assess the target nodule, choosing the optimal position for the patient in order to reduce out-of-plane motion from breathing. Using the B-mode image as a guide, the probe is positioned over the lesion on both screens simultaneously to facilitate enhancement characterization [217]. Then, a contrast agent is injected into an antecubital vein using a ≥ 20 G catheter, followed by a saline flush. Currently, there are two commercially available US contrast agents for CEUS-LI-RADS: Definity® (in USA, Canada)/Luminity® (outside USA, Canada) and Lumason® (in USA)/Sono-Vue® (outside USA).

Arterial phase begins within 20 s after injection of the contrast agent and persists until 30–45 s, depending on the cardiocirculatory situation. PVP begins 30–45 s after injection and ends at 120 s, while the late phase starts 2 min after injection, lasting until the clearance of microbubbles from liver tissue [217].

It is advisable to perform the examination continuously from contrast medium injection to the peak of arterial phase enhancement, in order to intercept APHE. Conversely, PVP and LP examinations are executed intermittently (every 30 sec) to minimize the destruction of microbubbles, allowing one to observe the late washout [217].

Finally, a survey recording is recommended from the contrast vascular input until the peak of APHE.
2.3.2. CEUS-LI-RADS Categories

As in the CT/MRI model, eight diagnostic categories with related imaging work-up suggestions cover the spectrum of lesions possibly discernible at CEUS.

Nodules are classified based on major criteria: size (<10, ≥10 <20 mm, ≥20 mm), the APHE appearance and the presence and type of washout.

CEUS LR-1 and 2 categories correspond to definitely and probably benign observations, respectively. LR-1 comprises formations with unequivocally benign contrastographic behavior, whereas LR-2 is assigned to isoenhancing lesions (solid nodule <10 mm/nonmasslike formations of any size) and LR-3 nodules stable in size for ≥2 years [219,220].

Patients with CEUS LR-1 and LR-2 can resume routine 6-month US surveillance, without the need for further diagnostic investigations [219].

LR-3 (intermediate probability of malignancy) includes solid nodules ≥10 mm with isoenhancement in all phases, nodules <20 mm showing late washout onset (≥60 s) and a mild degree of washout. CEUS LR-3 also includes nodules <10 mm, which show only APHE as a vascular criterion. Guidelines suggest repeating the examination with contrast administration within 3–6 months, but in selected cases, the multidisciplinary team could require a biopsy [219].

The LR-4 categories (probable HCC) include nodules ≥10 mm with APHE and no appreciable washout and those with APHE, late washout onset (≥60 s) and a mild degree of washout but measuring <10 mm. Finally, also foci ≥20 mm with no APHE but late washout onset (≥60 s) and a mild degree of washout may also be characterized as CEUS LR-4 [219].

All these conditions should undergo biopsy assessment or short-term (<3 months) imaging follow-up, relying on the multidisciplinary discussion decision [219].

The LR-5 nodules fulfill all major criteria and are treated as definitely HCC without biopsy confirmation [219].

CEUS algorithm also provides an LR-M class that includes not HCC malignant lesions, which usually need biopsy confirmation. The imaging features for LR-M are rim APHE, early (<60 s) washout or marked washout resulting in a “punched-out” appearance within 2 min after contrast injection. If the image quality is poor and does not allow lesion definition, the radiologist is called upon to assign the LR-NC category [219,220].

As CT/MRI, LR-TIV refers to tumor tissue in the portal or hepatic veins.

Ancillary features favoring malignancy, without specificity for HCC, include definite interval size increase, whereas definite interval size reduction or stability ≥2 years favor benignity. Features favoring HCC, in particular, include a mosaic appearance of the nodule and a nodule-in-nodule appearance in the arterial phase [219]. These features are applied less often in the CEUS-LI-RADS algorithm, with the same limitation of the CT/MRI model in upgrading lesions from LR-4 to LR-5 [219].

2.3.3. CEUS-LI-RADS vs. CT/MRI LI-RADS

CEUS enables the possibility of real-time assessment of APHE, which might be missed or misinterpreted in other imaging studies in case of inappropriate sequences/acquisition timing. This property is particularly useful in definitive LR-2 categorization of nodules, presenting dubiously HCC (LR-3-LR4), for a suspected arterial phase appearance on the CT/MRI algorithm. Among all benign focal liver formations, the assessment of hemangiomas and arteriportal shunts could result particularly challenging [221,222]. Indeed, rapidly filling hemangiomas may present as homogeneous APHE nodules on CT/MRI, mimicking HCCs. Similarly, arteriportal shunts could be perceived as nodules or could hide malignant fociulations, lowering the diagnostic accuracy of CT/MRI categorization [221].

The peculiarity of CEUS in intercepting APHE is reflected positively not only in the downgrade of LR-3 or LR-4 formations but also in the definitive diagnosis of HCC, assigning the category LR-5 to probable malignant lesions [219]. Indeed, several studies have shown that CEUS could intercept APHE in nodules categorized as LR-5, formerly misdiagnosed as LR3 or LR-4 at CT/MRI examinations.
In particular, Maruyama et al. recorded that 7/27 focal liver lesions (<20 mm) characterized as non-hypervascular (APNHE) on prior contrast-enhanced CT then showed APHE at CEUS and resulted as HCCs on histological specimens [223].

As highlighted, APHE is a necessary major feature to achieve HCC diagnosis. According to the CEUS algorithm, any nodule ≥ 10 mm with APHE is classified as LR-4 or LR-5, depending on the presence or absence of the washout criterion, respectively. This assumption is based on the “B-mode nodules visibility” as a prerequisite for CEUS evaluation. As a corollary, the LR-3 category of CEUS includes more cases of HCCs compared to the CT/MRI algorithm, since the examination is based on actual nodules, whereas LR-3 formations on CT/MRI include observations even <10 mm that are not necessarily well defined [224].

Albeit less frequently, mass-forming ICCs are encountered during US surveillance. On CEUS, ICC often shows rim-APHE, which is an uncommon finding in HCC. Rim-APHE was a key feature in Huang et al.’s study on 228 nodules (99 ICCs and 129 LR-M HCCs) showing sensitivity and specificity for differential diagnosis with HCCs of 70.4% and 68.8%, respectively. In addition, combing the rim-APHE with elevated CA 19-9 and normal alpha-fetoprotein (AFP) values increased sensitivity to 100% [225]. Rim-APHE should not be confused with the CT/MRI capsule appearance major criterion, which could not be appreciated on CEUS due to its fibrotic nature with large interstitial spaces and low blood volume. ICC also shows an early (<60 s) and prominent washout compared to HCC, which conversely proves a mild and late washout (≥60 s) after contrast injection [219].

Therefore, the LR-M category on CEUS-LI-RADS could be confidentially assigned in the presence of rim-APHE, early washout (<60 s), marked washout visible within the first 2 min after contrast injection or any combination of these three features [219,225].

2.4. LI-RADS Treatment Response Algorithm

HCC treatment was traditionally based on surgical or locoregional ablation technique [226–237]. The LI-RADS treatment response algorithm (TRA) is proposed as a comprehensive approach to standardize the disease assessment after locoregional therapies for HCC, using contrast-enhanced CT or MRI [226,227]. Compared to other criteria, such as modified Response Evaluation Criteria in Solid Tumours (mRECIST) [238], which evaluate the patient-level response, TRA provides the viability after treatment of the individual liver lesions [227].

Considering that the management of HCC often requires multiple therapeutic interventions, this approach could guide the multidisciplinary team caring for the patient in closely monitoring the evolution of the disease or in choosing the appropriate therapy.

Similarly to the LI-RADS diagnostic algorithm, the TRA classifies treated HCC into three different categories: LR-TR viable, LR-TR equivocal or LR-TR nonviable (Figure 10), according to the post-treatment enhancement pattern [227]. LR-TR viable is assigned to liver observations demonstrating persistent arterial phase hyperenhancement or washout or a pattern enhancement similar to pre-treatment imaging characteristics [227]. Generally, residual neoplastic cells could be distributed in a nodular, mass-like, or in thick irregular tissue within or along the treated lesion. If the observation shows no enhancement or if only a typical post-treatment enhancement is observed, LR-TR nonviable is assigned. In challenging cases that are difficult to define, an intermediate category (LR-TR equivocal) could be assigned. The role of the intermediate category is still questioned [227].

Chaudhry et al. [239] evaluated the application of the TRA in 36 patients with 53 lesions after ablative therapy prior to liver transplantation, comparing the category assigned on imaging with the pathological finding. Assessing equivocal lesions as viable, the researchers observed an increase in sensitivity in predicting complete necrosis to 81–87%, compared to 40–70% when treating the same lesions as nonviable. Furthermore, the majority of lesions classified as LR-TR equivocal (five out of six (83%)) exhibited incomplete necrosis in histopathology specimens [239]. Similarly, Shropshire et al. found incomplete necrosis in 12/17 (71%) of the lesions categorized as equivocal after transarterial embolization [240].
Hence, the application of the LR-TR *equivocal* category would seem to increase the sensitivity in residual disease detection, intercepting some cases that might escape from a treatment program for the residual presence of HCC [240].

![Figure 10](image-url). Treated HCC on II seg. Nonviable lesion. The lesion (arrow) shows necrotic appearance during arterial (A) and portal (B) phase of contrast study.

On the other hand, what also emerges from these studies is the unsatisfactory diagnostic accuracy of the nonviable LR-TR category in predicting a complete response to therapy. In both Chaudry’s and Shropshire’s investigations, only a low percentage of lesions assigned as nonviable were actually completely necrotic at histopathology (49% and 38–46%, respectively) [239–242].

Currently, insufficient data are available in evaluating the impact of nonviable LR-TR, in cases of minimal residual disease at pathological relief on patients’ overall survival or progression-free survival. Given the possibility of residual microfoci of disease that are difficult to distinguish on imaging after local-regional therapy, the interval for imaging follow-up should remain close, regardless of LR-TR category [67].

Although the application of TRA seems plausible in clinical practice, there are still important gaps. For instance, an area that has not yet been sufficiently investigated concerns the evaluation of TRA performance in assessing treatment response after locoregional radiation-based therapy (transarterial radioembolization or external beam radiotherapy). Further research should be promoted to improve the algorithm, decrease the rate of LR-TR *equivocal* categorizations and provide clinical assessment of residual disease after all types of local-regional therapies.

3. Conclusions

LI-RADS is the most comprehensive system for the diagnosis and management of HCC. Four LI-RADS algorithms cover three individual clinical contexts: screening and surveillance, diagnosis/staging and assessment of treatment response.

CEUS-LI-RADS and CT/MRI LI-RADS algorithms could achieve confident HCC diagnosis, providing tailored management recommendations for *observations* found on US surveillance in patients considered at risk. In particular, CEUS-LI-RADS can afford a contrast-enhanced examination in absence of radiation exposure. This could be an important advantage, especially in young patients.

Currently, LI-RADS major imaging criteria should always be satisfied, while ancillary features could be used at the discretion of the radiologist to increase or decrease the category. However, the sensitivity value of these combined criteria is inadequate. Recent evidence proved that HPH on Gd-EOB-DTPA could be an important tool not only in increasing sensitivity but also in diagnosis as a major criterion. Although LI-RADS emerges as an
essential instrument to support the management of liver tumors, still many improvements are needed to overcome the current limitations. In particular, features that may clearly distinguish HCC from ICC and cHCC-CC lesions and the assessment after locoregional radiation-based therapy are still fields of research. In fact, although the application of TRA seems plausible in clinical practice, there are still important gaps. An area that has not yet been sufficiently investigated concerns the evaluation of TRA performance in assessing treatment response after transarterial radioembolization or external beam radiotherapy. Further research should be promoted to improve the algorithm, decrease the rate of LR-TR equivocal categorizations and provide clinical assessment of residual disease after all types of local-regional therapies.

Author Contributions: Each authors contributed equally at the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors are grateful to Alessandra Trocino, librarian at the National Cancer Institute of Naples, Italy.

Conflicts of Interest: The authors have no conflict of interest to be disclosed. The authors confirm that the article is not under consideration for publication elsewhere. Each author has participated sufficiently to take public responsibility for the content of the manuscript.

References
1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 2021, 71, 209–249. [CrossRef] [PubMed]
2. World Health Organization (WHO). Global Health Estimates 2020: Deaths by Cause, Age, Sex, by Country and by Region, 2000–2019; WHO: Geneva, Switzerland, 2020.
3. Granata, V.; Grassi, R.; Fusco, R.; Belli, A.; Cutolo, C.; Pradella, S.; Grazzini, G.; La Porta, M.; Brunese, M.C.; De Muzio, F.; et al. Diagnostic evaluation and ablation treatments assessment in hepatocellular carcinoma. Infect. Agent Cancer 2021, 16, 53. [CrossRef] [PubMed]
4. Shin, N.; Choi, J.A.; Choi, J.M.; Cho, E.S.; Kim, J.H.; Chung, J.J.; Yu, J.S. Sclerotic changes of cavernous hemangioma in the cirrhotic liver: Long-term follow-up using dynamic contrast-enhanced computed tomography. Radiol. Med. 2020, 125, 1225–1232. [CrossRef] [PubMed]
5. Bottari, A.; Silipigni, S.; Carerj, M.L.; Cattaf, A.; Maimone, S.; Marino, M.A.; Mazziotti, S.; Pitrone, A.; Squadrito, G.; Ascenti, G. Dual-source dual energy CT in the evaluation of hepatic fractional extracellular space in cirrhosis. Radiol. Med. 2020, 125, 7–14. [CrossRef] [PubMed]
6. Bracci, S.; Dolciami, M.; Trobìani, C.; Izzo, A.; Pernazza, A.; D’Amati, G.; Manganaro, L.; Ricci, P. Quantitative CT texture analysis in predicting PD-L1 expression in locally advanced or metastatic NSCLC patients. Radiol. Med. 2021, 126, 1425–1433.
7. Granata, V.; Petrillo, M.; Fusco, R.; Setola, S.V.; de Lutio di Castelguidone, E.; Catalano, O.; Piccirillo, M.; Albino, V.; Izzo, F.; Petrillo, A. Diagnostic evaluation and ablation treatments assessment in hepatocellular carcinoma. Gastroenterol. Res. Pract. 2013, 2013, 469097. [CrossRef] [PubMed]
8. Dimitroulis, D.; Damaskos, C.; Valsami, S.; Davakis, S.; Garmpis, N.; Spartalis, E.; Athanasiou, A.; Moris, D.; Sakellariou, S.; Kykalos, S.; et al. From diagnosis to treatment of hepatocellular carcinoma: An epidemiologic problem for both developed and developing world. World J. Gastroenterol. 2017, 23, 5282–5294. [CrossRef]
9. White, D.L.; Kanwal, F.; El-Serag, H.B. Association between nonalcoholic fatty liver disease and risk for hepatocellular cancer, based on systematic review. Clin. Gastroenterol. Hepatol. 2012, 10, 1342–1359. [CrossRef]
10. Mokrane, F.Z.; Lu, L.; Vavasseur, A.; Otal, P.; Peron, J.M.; Luk, L.; Yang, H.; Ammari, S.; Saenger, Y.; Rousseau, H.; et al. Radiomics machine-learning signature for diagnosis of hepatocellular carcinoma in cirrhotic patients with indeterminate liver nodules. Eur. Radiol. 2020, 30, 558–570. [CrossRef]
11. Karmazanovsky, G.; Gruzdev, I.; Tikhonova, V.; Kondrattyev, E.; Revishvili, A. Computed tomography-based radiomics approach in pancreatic tumors characterization. Radiol. Med. 2021.
12. Granata, V.; Fusco, R.; Venanzio Setola, S.; Sandomenico, F.; Luisa Barretta, M.; Belli, A.; Palaia, R.; Tatangelo, F.; Grassi, R.; Izzo, F.; et al. Major and ancillary features according to LI-RADS in the assessment of combined hepatocellular-cholangiocarcinoma. Radiol. Oncol. 2020, 54, 149–158. [CrossRef] [PubMed]

13. Granata, V.; Fusco, R.; Setola, S.V.; Picone, C.; Vallone, P.; Belli, A.; Incollongo, P.; Albino, V.; Tatangelo, F.; Izzo, F.; et al. Microvascular invasion and grading in hepatocellular carcinoma: Correlation with major and ancillary features according to LI-RADS. Abdom. Radiol. 2019, 44, 2788–2800. [CrossRef]

14. Ganne-Carrié, N.; Chaffaut, C.; Bourcie, V.; Archambeaud, I.; Perarnau, J.M.; Oberti, F.; Roulot, D.; Moreno, C.; Louvet, A.; Dao, T.; et al. Estimate of hepatocellular carcinoma incidence in patients with alcoholic cirrhosis. J. Hepatol. 2018, 69, 1274–1283. [CrossRef] [PubMed]

15. Sempoux, C.; Balabaud, C.; Bioulac-Sage, P. Malignant transformation of hepatocellular adenoma. Hepatol. Oncol. 2014, 1, 421–431. [CrossRef] [PubMed]

16. Granata, V.; Fusco, R.; Venanzio Setola, S.; Barretta, M.L.; Iasevoli, D.M.A.; Palaia, R.; Belli, A.; Patrone, R.; Tatangelo, F.; Grassi, R.; et al. Diagnostic performance of LI-RADS in adult patients with rare hepatic tumors. Eur. Rev. Med. Pharmacol. Sci. 2022, 26, 399–414. [CrossRef]

17. Granata, V.; Fusco, R.M.; Catalano, O.; Filice, S.; Avallone, A.; Piccirillo, M.; Longi, M.; Palaia, R.; Grassi, R.; Izzo, F.; et al. Uncommon neoplasms of the biliary tract. Radiological findings. Br. J. Radiol. 2017, 90, 20160561. [CrossRef] [PubMed]

18. Granata, V.; Fusco, R.; de Lutio di Castelguidone, E.; Avallone, A.; Palaia, R.; Delrio, P.; Tatangelo, F.; Botti, G.; Grassi, R.; Izzo, F.; et al. Diagnostic performance of gadoxetic acid-enhanced liver MRI versus multidetector CT in the assessment of colorectal liver metastases compared to hepatic resection. BMC Gastroenterol. 2019, 19, 129. [CrossRef] [PubMed]

19. Petralia, G.; Summers, P.E.; Agostini, A.; Ambrosini, R.; Cianci, R.; Cristel, G.; Calistri, L.; Colagrande, S. Dynamic contrast-enhanced MRI in oncology: How we do it. Radiol. Med. 2020, 125, 1288–1300. [CrossRef]

20. Ria, F.; Samei, E. Is regulatory compliance enough to ensure excellence in medicine? Radiol. Med. 2020, 125, 904–905. [CrossRef]

21. Zhang, A.; Song, J.; Ma, Z.; Chen, T. Combined dynamic contrast-enhanced magnetic resonance imaging and diffusion-weighted imaging to predict neoadjuvant chemotherapy effect in FIGO stage IB2-IIA2 cervical cancers. Radiol. Med. 2020, 125, 1233–1242. [CrossRef]

22. Crimi, F.; Capelli, G.; Spolverato, G.; Bao, Q.Y.; Florio, A.; Milite Rossi, S.; Cecchin, D.; Albertoni, L.; Campi, C.; Pucciarelli, S.; et al. MRI T2-weighted sequences based texture analysis (TA) as a predictor of response to neoadjuvant chemo-radiotherapy (nCRT) in patients with locally advanced rectal cancer (LARC). Radiol. Med. 2020, 125, 1216–1224. [CrossRef] [PubMed]

23. Granata, V.; Fusco, R.; Avallone, A.; Filice, F.; Tatangelo, F.; Piccirillo, M.; Grassi, R.; Izzo, F.; Petrollo, A. Critical analysis of the major and ancillary imaging features of LI-RADS on 127 proven HCCs evaluated with functional and morphological MRI: Lights and shadows. Oncotarget 2017, 8, 51224–51237. [CrossRef] [PubMed]

24. Kirienko, M.; Ninatti, G.; Cozzi, L.; Voulaz, E.; Gennaro, N.; Barajon, I.; Ricci, F.; Carlo-Stella, C.; Zucali, P.; Sollini, M.; et al. Computed tomography (CT)-derived radiomic features differentiate prevascular mediastinal masses as thymic neoplasms versus lymphomas. Radiol. Med. 2020, 125, 951–960. [CrossRef] [PubMed]

25. Zhang, L.; Kang, L.; Li, G.; Zhang, X.; Ren, J.; Shi, Z.; Li, J.; Yu, S. Computed tomography-based radiomics model for discriminating the risk stratification of gastrointestinal stromal tumors. Radiol. Med. 2020, 125, 465–473. [CrossRef]

26. Fusco, R.; Sansone, M.; Granata, V.; Setola, S.V.; Petrollo, A. A systematic review on multiparametric MR imaging to risk stratification of gastrointestinal stromal tumors. J. Clin. Med. 2021, 10, 1259. [CrossRef]

27. De Muzio, F.; Cutolo, C.; Dell’Aversana, F.; Grassi, F.; Ravo, L.; Ferrante, M.; Danti, G.; Flamia, F.; Simonetti, I.; Palumbo, P.; et al. Complications after Thermal Ablation of Hepatocellular Carcinoma and Liver Metastases: Imaging Findings. Diagnostics 2022, 12, 1151. [CrossRef]

28. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.V.; Simonetti, I.; Dell’Aversana, F.; Grassi, F.; Bruno, F.; Belli, A.; et al. Complications Risk Assessment and Imaging Findings of Thermal Ablation Treatment in Liver Cancers: What the Radiologist Should Expect. J. Clin. Med. 2022, 11, 2766. [CrossRef]

29. De Filippo, M.; Puglisi, S.; D’Amuri, F.; Gentili, F.; Paladini, I.; Carrafielo, G.; Maestroni, U.; Del Rio, P.; Ziglioli, F.; Pagnini, F. CT-guided percutaneous drainage of abdominopelvic collections: A pictorial essay. Radiol. Med. 2021, 126, 1561–1570. [CrossRef]

30. Fusco, R.; Granata, V.; Sansone, M.; Rega, D.; Delrio, P.; Tatangelo, F.; Romano, C.; Avallone, A.; Pupo, D.; Giordano, M.; et al. Validation of the standardized index of shape tool to analyze DCE-MRI data in the assessment of neo-adjuvant therapy in locally advanced rectal cancer. Radiol. Med. 2021, 126, 1044–1054. [CrossRef]

31. Barabino, M.; Gurgitano, M.; Angileri, S.A.; Franceschelli, G.; Santambrogio, R.; Mariani, N.M.; Opocher, E.; Carrafielo, G. LI-RADS to categorize liver nodules in patients at risk of HCC: Tool or a gadget in daily practice? Radiol. Med. 2021, 126, 5–13. [CrossRef]

32. Cholangiocarcinoma Working Group. Italian Clinical Practice Guidelines on Cholangiocarcinoma—Part I: Classification, diagnosis and staging. Dig. Liver Dis. 2020, 52, 1282–1293. [CrossRef] [PubMed]

33. Cholangiocarcinoma Working Group. Italian Clinical Practice Guidelines on Cholangiocarcinoma—Part II: Treatment. Dig. Liver Dis. 2020, 52, 1430–1442. [CrossRef] [PubMed]

34. Gabelloni, M.; Di Nasso, M.; Morganti, R.; Faggioni, L.; Masi, G.; Falcone, A.; Neri, E. Application of the ESR iGuide clinical decision support system to the imaging pathway of patients with hepatocellular carcinoma and cholangiocarcinoma: Preliminary findings. Radiol. Med. 2020, 125, 531–537. [CrossRef] [PubMed]
35. Granata, V.; Bicchierai, G.; Fusco, R.; Cozzi, D.; Grazzini, G.; Danti, G.; De Muzio, F.; Maggialetti, N.; Smorchkova, O.; D’Elia, M.; et al. Diagnostic protocols in oncology: Workup and treatment planning. Part 2: Abbreviated MR protocol. Eur. Rev. Med. Pharmacol. Sci. 2021, 25, 6499–6528. [PubMed]

36. Laurelli, G.; Falcone, F.; Gallo, M.S.; Scala, F.; Losito, S.; Granata, V.; Cascella, M.; Greggi, S. Long-Term Oncologic and Reproductive Outcomes in Young Women With Early Endometrial Cancer Conservatively Treated: A Prospective Study and Literature Update. Int. J. Gynecol. Cancer 2016, 26, 1650–1657. [CrossRef]

37. Gatti, M.; Calandrini, M.; Bergamasco, L.; Darvizeh, F.; Grazioli, L.; Inchingolo, R.; Ippolito, D.; Roussé, S.; Velti, A.; Fonio, P.; et al. Characterization of the arterial enhancement pattern of focal liver lesions by multiple arterial phase magnetic resonance imaging: Comparison between hepatocellular carcinoma and focal nodular hyperplasia. Radiol. Med. 2020, 125, 348–355. [CrossRef]

38. Orlacchio, A.; Chegai, F.; Roma, S.; Merolla, S.; Bosa, A.; Franciosi, S. Degradable starch microspheres transarterial chemoembolization (DSMs-TACE) in patients with unresectable hepatocellular carcinoma (HCC): Long-term results from a single-center 137-patient cohort prospective study. Radiol. Med. 2020, 125, 98–106. [CrossRef]

39. Cutolo, C.; Dell’Aversana, F.; Fusco, R.; Grazzini, G.; Chiti, G.; Simonetti, I.; Bruno, F.; Palumbo, P.; Pierpaoli, L.; Valeri, T.; et al. Combined Hepatocellular-Cholangiocarcinoma: What the Multidisciplinary Team Should Know. Diagnostics (Basel) 2022, 12, 890. [CrossRef]

40. Fiorentino, A.; Gregucci, F.; Bonaparte, I.; Vitulano, N.; Surgo, A.; Mazzola, R.; Di Monaco, A.; Carbonara, R.; Alongi, F.; Langialonga, T.; et al. Stereotactic Ablative radiation therapy (SABR) for cardiac arrhythmia: A new therapeutic option? Radiol. Med. 2021, 126, 155–162, Epub 13 May 2020. [CrossRef]

41. Nakamura, Y.; Higaki, T.; Honda, Y.; Tatsugami, F.; Tani, C.; Fukumoto, W.; Narita, K.; Kondo, S.; Akagi, M.; Awai, K. Advanced CT techniques for assessing hepatocellular carcinoma. Radiol. Med. 2021, 126, 925–935. [CrossRef]

42. Granata, V.; Fusco, R.; Belli, A.; Borzillo, V.; Palumbo, P.; Bruno, F.; Grassi, R.; Ottaiano, A.; Nasti, G.; Pilone, V.; et al. Conventional, functional and radiomics assessment for intrahepatic cholangiocarcinoma. Infect. Agent Cancer 2022, 17, 13. [CrossRef] [PubMed]

43. Nakamura, V.; Fusco, R.; Setola, S.V.; Simonetti, I.; Cozzi, D.; Grazzini, G.; Grassi, F.; Belli, A.; Miele, V.; Izzo, F.; et al. An update on radiomics techniques in primary liver cancers. Infect. Agent Cancer 2022, 17, 6. [CrossRef]

44. Barile, A.; Conti, L.; Lanni, G.; Calvisi, V.; Masciocchi, C. Evaluation of medial meniscus tears and meniscal stability: Weight-bearing MRI vs arthroscopy. Eur. J. Radiol. 2013, 82, 633–639. [CrossRef] [PubMed]

45. Caruso, D.; Polici, M.; Zerunian, M.; Nacci, I.; Marasco, M.; Magi, L.; Tarallo, M.; Gargiulo, S.; Iannicelli, E.; et al. CT-based radiomics for prediction of therapeutic response to Everolimus in metastatic neuroendocrine tumors. Radiol. Med. 2022, Epub ahead of print. [CrossRef] [PubMed]

46. Agazzi, G.M.; Ravanelli, M.; Rocca, E.; Medicina, D.; Balzarini, P.; Pessina, C.; Vermi, W.; Berruti, A.; Maroldi, R.; Farina, D. CT texture analysis for prediction of EGFR mutational status and ALK rearrangement in patients with non-small cell lung cancer. Radiol. Med. 2021, 126, 786–794. [CrossRef]

47. Iacobellis, F.; Di Serafino, M.; Brillantino, A.; Mottola, A.; Del Giudice, S.; Stavolo, C.; Festa, P.; Patlas, M.N.; Scaglione, M.; Romano, L. Role of MRI in early follow-up of patients with solid organ injuries: How and why we do it? Radiol. Med. 2021, 126, 1328–1334. [CrossRef]

48. Liu, J.; Wang, C.; Guo, W.; Zeng, P.; Liu, Y.; Lang, N.; Yuan, H. A preliminary study using spinal MRI-based radiomics to predict high-risk cytogenetic abnormalities in multiple myeloma. Radio. Med. 2021, 126, 1226–1235.

49. Qin, H.; Que, Q.; Lin, P.; Li, X.; Wang, X.R.; He, Y.; Chen, J.Q.; Yang, H. Magnetic resonance imaging (MRI) radiomics of papillary thyroid cancer (PTC): A comparison of predictive potential of multiple classifiers modeling to identify cervical lymph node metastases before surgery. Radiol. Med. 2021, 126, 1312–1327.

50. Gannon, C.J.; Izzo, F.; Aloia, T.A.; Pignatta, S.; Nasti, G.; Vallone, P.; Orlando, R.; Scordino, F.; Curley, S.A. Can hepatocellular cancer screening increase the proportion of long-term survivors? Hepatogastroenterology 2009, 56, 1152–1156. [PubMed]

51. Danti, G.; Berti, V.; Abenavoli, E.; Briganti, V.; Linguanti, F.; Mungai, F.; Pradella, S.; Miele, V. Diagnostic imaging of typical lung carcinoids: Relationship between MDCT, 111In-Octreoscan and 18F-FDG-PET imaging features with Ki-67 index. Radiol. Med. 2020, 125, 715–729. [CrossRef]

52. Scapicchio, C.; Gabelloni, M.; Barucci, A.; Cioni, D.; Saba, L.; Neri, E. A deep look into radiomics. Radiol. Med. 2021, 126, 1296–1311. [CrossRef] [PubMed]

53. Salvati, F.; Rossi, F.; Limbucci, N.; Pistoia, M.L.; Barile, A.; Masciocchi, C. Mucoid metaplastic-degeneration of anterior cruciate ligament. J. Sports Med. Phys. Fit. 2008, 48, 483–487.

54. Benedetti, G.; Mori, M.; Panzeri, M.M.; Barbera, M.; Palumbo, D.; Sini, C.; Mufatti, F.; Andreasi, V.; Steidler, S.; Doglioni, C.; et al. CT-derived radiomic features to discriminate histologic characteristics of pancreatic neuroendocrine tumors. Radiol. Med. 2021, 126, 745–760. [CrossRef] [PubMed]

55. Barile, A.; Bruno, F.; Arrigoni, F.; Splendiani, A.; Di Cesare, E.; Zappia, M.; Guglielmi, G.; Masciocchi, C. Emergency and Trauma of the Ankle. In Seminars in Musculoskeletal Radiology; Thieme Medical Publishers: New York, NY, USA, 2017; Volume 21, pp. 282–289. [CrossRef]

56. Nardone, V.; Reginelli, A.; Grassi, R.; Boldrini, L.; Vacca, G.; D’Ippolito, E.; Annunziata, S.; Farchione, A.; Belfiore, M.P.; Desideri, I.; et al. Delta radiomics: A systematic review. Radiol. Med. 2021, 126, 1571–1583. [CrossRef] [PubMed]

57. Brunese, L.; Brunese, M.C.; Carbone, M.; Ciccone, V.; Mercalfo, F.; Santone, A. Automatic PI-RADS assignment by means of formal methods. Radiol. Med. 2021, 127, 83–89. [CrossRef]
58. van der Lubbe, M.F.; Vaidyanathan, A.; de Wit, M.; van den Burg, E.L.; Postma, A.A.; Bruinjes, T.D.; Bilderbeek-Beckers, M.A.L.; Dammeijer, P.F.M.; Bossche, S.V.; Van Rompaey, V.; et al. A non-invasive, automated diagnosis of Menière’s disease using radiomics and machine learning on conventional magnetic resonance imaging: A multicentric, case-controlled feasibility study. Radiol. Med. 2021, 127, 72–82. [CrossRef]

59. Granata, V.; Fusco, R.; Avallone, A.; Cassata, A.; Palaia, R.; Delrio, P.; Grassi, R.; Tatangelo, F.; Grazzini, G.; Izzo, F.; et al. Abbreviated MRI protocol for colorectal liver metastases: How the radiologist could work in pre surgical setting. PLoS ONE 2020, 15, e0241431. [CrossRef]

60. Granata, V.; Fusco, R.; Venanzio Setola, S.; Mattace Raso, M.; Avallone, A.; De Stefano, A.; Nasti, G.; Palaia, R.; Delrio, P.; Petrillo, A.; et al. Liver radiologic findings of chemotherapy-induced toxicity in liver colorectal metastases patients. Eur. Rev. Med. Pharmacol. Sci. 2019, 23, 9697–9706. [CrossRef]

61. Granata, V.; Fusco, R.; Maio, F.; Avallone, A.; Nasti, G.; Palaia, R.; Albino, V.; Grassi, R.; Izzo, F.; Petrillo, A. Qualitative assessment of EOB-GD-DTPA and Gd-BT-DOTA MR contrast studies in HCC patients and colorectal liver metastases. Infect. Agent Cancer 2019, 14, 40. [CrossRef]

62. Chernyak, V.; Tang, A.; Do, R.K.G.; Kamaya, A.; Kono, Y.; Santillan, C.S.; Fowler, K.J.; Bashir, M.R.; Cunha, G.M.; Fetzer, D.T.; et al. Liver imaging: It is time to adopt standardized terminology. Radiol. Med. 2022. Epub ahead of print. [CrossRef]

63. Terzi, E.; Giamperoli, A.; Iavarone, M.; Leoni, S.; Granito, A.; Forgione, A.; Tovoli, F.; Piscaglia, F. Prognosis of Single Early-Stage Hepatocellular Carcinoma (HCC) with CEUS Inconclusive Imaging (LI-RADS LR-3 and LR-4) Is No Better than Typical HCC (LR-5). Cancers 2022, 14, 336. [CrossRef]

64. Masciocchi, C.; Arrigoni, F.; La Marra, A.; Mariani, S.; Pizzagalli, M.; Barile, A. Treatment of focal benign lesions of the bone: MRgFUS and RFA. Br. J. Radiol. 2016, 89, 20150356. [CrossRef]

65. Moura Cunha, G.; Chernyak, V.; Fowler, K.J.; Sirlin, C.B. Up-to-Date Role of CT/MRI LI-RADS in Hepatocellular Carcinoma. J. Hepatol. Carcinoma 2021, 8, 513–527. [CrossRef] [PubMed]

66. Marrero, J.A.; Kulik, L.M.; Sirlin, C.B.; Finn, R.S.; Abecassis, M.M.; Roberts, L.R.; Heimbach, J.K. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guideline by the American Association for the Study of Liver Diseases. Hepatology 2018, 68, 723–750. [CrossRef] [PubMed]

67. American College of Radiology. CT/MRI Liver Imaging Reporting and Data System v2018 Core. Available online: https://www.acr.org/ClinicalResources/Reporting-and-Data-Systems/LI-RADS/CT-MRI-LIRADS-v (accessed on 15 April 2020).

68. Granata, V.; Fusco, R.; Avallone, A.; Catalano, O.; Filice, F.; Leongito, M.; Palaia, R.; Izzo, F.; Petrillo, A. Major and ancillary magnetic resonance features of LI-RADS to assess HCC: An overview and update. Infect. Agent Cancer 2017, 12, 23. [CrossRef] [PubMed]

69. Granata, V.; Fusco, R.; Belli, A.; Danti, G.; Bicci, E.; Cusano, R.; Petrillo, A.; Izzo, F. Diffusion weighted imaging and diffusion kurtosis imaging in abdominal oncological setting: Why and when. Radiol. Med. 2022, 17, 25.

70. Ierardi, A.M.; Gaibazzi, N.; Tuttolomondo, D.; Fusco, S.; La Mura, V.; Peviani, F.; Aceto, S.; Blasi, F.; Cozzi, D.; Carrafiello, G.; et al. Deep vein thrombosis in COVID-19 patients in general wards: Prevalence and association with clinical and laboratory variables. Radiol. Med. 2021, 126, 774–778. [CrossRef] [PubMed]

71. Argalia, G.; Tarantino, G.; Ventura, C.; Campioni, D.; Tagliati, C.; Guardati, P.; Kostandini, A.; Marzioni, M.; Giuseppetti, G.M.; Giovagnoni, G. Shear wave elastography and transient elastography in HCV patients after direct-acting antivirals. Radiol. Med. 2021, 126, 894–899. [CrossRef] [PubMed]

72. Zener, R.; Orepoulos, G.; Beecroft, R.; Rajan, D.K.; Jaskolka, J.; Tan, K.T. Transabdominal Direct Sac Puncture Embolization of Type II Endoleaks after Endovascular Abdominal Aortic Aneurysm Repair. J. Vasc. Interv. Radiol. 2018, 29, 1167–1173. [CrossRef] [PubMed]
80. Xian, M.F.; Huang, Y.; Xie, W.X.; Pan, K.M.; Zeng, D.; Huang, H.; Li, M.D.; Xie, X.Y.; Kuang, M.; Lu, M.D.; et al. LR-M observations on contrast-enhanced ultrasound: Detection of hepatocellular carcinoma using additional features in comparison with current LI-RADS criteria. AJR Am. J. Roentgenol. 2021, 219, 76–85. [CrossRef]

81. Guo, H.L.; Zheng, X.; Cheng, M.Q.; Zeng, D.; Huang, H.; Xie, X.Y.; Lu, M.D.; Kuang, M.; Wang, W.; Xian, M.F.; et al. Contrast-Enhanced Ultrasound for Differentiation Between Poorly Differenitated Hepatocellular Carcinoma and Intrahepatic Cholangiocarcinoma. J. Ultrasound Med. 2021, 41, 1213–1225. [CrossRef]

82. Barile, A.; Sabatini, M.; Iannessi, F.; Di Cesare, E.; Splendiani, A.; Calvisi, V.; Masicocchi, C. Pigmented villonodular synovitis (PVNS) of the knee joint: Magnetic resonance imaging (MRI) using standard and dynamic paramagnetic contrast media. Report of 52 cases surgically and histologically controlled. Radiol. Med. 2004, 107, 356–366.

83. Kim, Y.Y.; Yeom, S.K.; Shin, H.; Choi, S.H.; Rhee, H.; Park, J.H.; Cho, E.S.; Park, S.; Lee, S.S.; Park, M.S. Clinical staging of mass-forming intrahepatic cholangiocarcinoma: Computed tomography versus magnetic resonance imaging. Hepatol. Commun. 2015, 5, 2009–2018. [CrossRef]

84. Ichikawa, S.; Yamamoto, H.; Morita, T. Comparison of a Bayesian estimation algorithm and singular value decomposition algorithms for 80-detector row CT perfusion in patients with acute ischemic stroke. Radiol. Med. 2021, 126, 795–803. [CrossRef] [PubMed]

85. Rampado, O.; Depaoli, A.; Marchisio, F.; Gatti, M.; Racine, D.; Ruggeri, V.; Ruggirello, I.; Darvizeh, F.; Fonio, P.; Ropolo, R. Effects of different levels of CT iterative reconstruction on low-contrast detectability and radiation dose in patients of different sizes: An anthropomorphic phantom study. Radiol. Med. 2021, 126, 55–62. [CrossRef] [PubMed]

86. Cicero, G.; Mazzotti, S.; Silipigni, S.; Blandino, A.; Cantisani, V.; Pergolizzi, S.; D’Angelo, T.; Stagno, A.; Maimone, S.; et al. Dual-energy CT quantification of fractional extracellular space in cirrhotic patients: Comparison between early and delayed equilibrium phases and correlation with oesophageal varices. Radiol. Med. 2021, 126, 761–767. [CrossRef]

87. Ponnoprat, D.; Inkeaw, P.; Chaijaruwanich, J.; Traisathit, P.; Sripaporn, P.; Inmutto, N.; Na Chiangmai, W.; Pongnikorn, D.; Chitapanarux, I. Classification of hepatocellular carcinoma and intrahepatic cholangiocarcinoma based on multi-phase CT scans. Med. Biol. Eng. Comput. 2020, 58, 2497–2515. [CrossRef]

88. Tsunematsu, S.; Chuma, M.; Kaniyama, T.; Miyamoto, N.; Yabusaki, S.; Hatanaka, K.; Hatamushi, T.; Kamachi, H.; Yokoo, H.; Kakisaka, T.; et al. Intratumoral artery on contrast-enhanced computed tomography imaging: Differentiating intrahepatic cholangiocarcinoma from poorly differentiated hepatocellular carcinoma. Abdom. Imaging 2015, 40, 1492–1499. [CrossRef]

89. Zhao, Y.J.; Chen, W.X.; Wu, D.S.; Zhang, W.Y.; Zheng, L.R. Differentiation of mass-forming intrahepatic cholangiocarcinoma from poorly differentiated hepatocellular carcinoma: Based on the multivariate analysis of contrast-enhanced computed tomography findings. Abdom. Radiol. 2016, 41, 978–989. [CrossRef]

90. Ruys, A.T.; van Beem, B.E.; Engelbrecht, M.R.; Bipat, S.; Stoker, J.; Van Gulik, T.M. Radiological staging in patients with hilar cholangiocarcinoma: A systematic review and meta-analysis. Br. J. Radiol. 2012, 85, 1255–1262. [CrossRef] [PubMed]

91. Ichikawa, S.; Isoda, H.; Shimizu, T.; Tamada, D.; Taura, K.; Todai, K.; Onishi, H.; Motosugi, U. Distinguishing intrahepatic mass-forming biliary carcinomas from hepatocellular carcinoma by computed tomography and magnetic resonance imaging using the Bayesian method: A bi-center study. Eur. Radiol. 2020, 30, 5992–6002. [CrossRef]

92. Chu, H.; Liu, Z.; Liang, W.; Zhou, Q.; Zhang, Y.; Lei, K.; Tang, M.; Cao, Y.; Chen, S.; Peng, S.; et al. Radiomics using CT images for preoperative prediction of futile resection in intrahepatic cholangiocarcinoma. Eur. Radiol. 2021, 31, 2368–2376. [CrossRef]

93. Megibow, A.J. Clinical abdominal dual-energy CT: 15 years later. Abdom. Radiol. 2020, 45, 1198–1201. [CrossRef]

94. Schicchi, N.; Fogante, M.; Palumbo, P.; Agliata, G.; Esposto Pirani, P.; Di Cesare, E.; Giovagnoni, A. The sub-millisievert era in imaging techniques for the diagnosis of hepatocellular carcinoma: LI-RADS. Chin. Clin. Oncol. 2021, 10, 3. [CrossRef] [PubMed]

95. El-Serag, H.B. Hepatocellular carcinoma. N. Engl. J. Med. 2011, 365, 1118–1127. [CrossRef]

96. Brizi, M.G.; Perillo, F.; Cannone, F.; Tuzza, L.; Manfredi, R. The role of imaging in acute pancreatitis. Radiol. Med. 2021, 126, 1017–1029. [CrossRef]

97. Assadsangabi, R.; Babaei, R.; Songco, C.; Ivanovic, V.; Bobinski, M.; Chen, Y.J.; Nabavizadeh, S.A. Multimodality oncologic evaluation of superficial neck and facial lymph nodes. Radiol. Med. 2021, 126, 1074–1084. [CrossRef]

98. Granata, V.; Grassi, R.; Fusco, R.; Galdiero, R.; Setola, S.V.; Palaia, R.; Belli, A.; Silvestro, L.; Cozzi, D.; Brunese, L.; et al. Pancreatic cancer detection and characterization: State of the art and radiomics. Eur. Rev. Med. Pharmacol. Sci. 2021, 25, 3684–3699. [CrossRef]

99. Granata, V.; Fusco, R.; Catalano, O.; Setola, S.V.; de Lutio di Castelguidone, E.; Piccirillo, M.; Palaia, R.; Grassi, R.; Granata, F.; Izzo, F.; et al. Multidetector computer tomography in the pancreatic adenocarcinoma assessment: An update. Infect. Agent Cancer 2016, 11, 57. [CrossRef]

100. Zhang, B.H.; Yang, B.H.; Tang, Z.Y. Randomized controlled trial of screening for hepatocellular carcinoma. J. Cancer Res. Clin. Oncol. 2004, 130, 417–422. [CrossRef] [PubMed]

101. Singal, A.; Volk, M.L.; Waljee, A.; Salgia, R.; Higgins, P.; Rogers, M.A.; Marrero, J.A. Meta-analysis: Surveillance with ultrasound for early-stage hepatocellular carcinoma in patients with cirrhosis. Aliment. Pharmacol. Ther. 2009, 30, 37–47. [CrossRef] [PubMed]

102. Chou, R.; Cuevas, C.; Fu, R.; Devine, B.; Wasson, N.; Ginsburg, A.; Zakhia, B.; Pappas, M.; Graham, E.; Sullivan, S.D. Imaging Techniques for the Diagnosis of Hepatocellular Carcinoma: A Systematic Review and Meta-analysis. Ann. Intern. Med. 2015, 162, 697–711. [CrossRef]
147. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.V.; Dell’Aversana, F.; Otaiano, A.; Nasti, G.; Grassi, R.; Pilone, V.; et al. EOB-MR Based Radiomics Analysis to Assess Clinical Outcomes following Liver Resection in Colorectal Liver Metastases. *Cancers* **2022**, *14*, 1239. [CrossRef] [PubMed]

148. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.V.; Dell’ Aversana, F.; Otaiano, A.; Avallone, A.; Nasti, G.; Grassi, F.; et al. Contrast MR-Based Radiomics and Machine Learning Analysis to Assess Clinical Outcomes following Liver Resection in Colorectal Liver Metastases: A Preliminary Study. *Cancers* **2022**, *14*, 1110. [CrossRef] [PubMed]

149. Boatright, C.; Peterson, J.; Williams, V.L.; Best, S.; Ash, R. LI-RADS v2018: Utilizing ancillary features on gadoxetate-enhanced MRI to modify final LI-RADS category. *Abdom. Radiol.* **2020**, *45*, 3136–3143. [CrossRef] [PubMed]

150. Palumbo, P.; Masedu, F.; De Cataldo, C.; Cannizzaro, E.; Bruno, F.; Pradella, S.; Arrigoni, F.; Valentì, M.; Splendiani, A.; Barile, A.; et al. Real-world clinical validity of cardiac magnetic resonance tissue tracking in primitive hypertrophic cardiomyopathy. *Radiol. Med.* **2021**, *126*, 1532–1543. [CrossRef]

151. Barile, A. Correction to: Some thoughts and greetings from the new Editor-in-Chief. *Radiol. Med.* **2021**, *126*, 1377. [CrossRef] [PubMed]

152. Cutolo, C.; De Muzio, F.; Fusco, R.; Simonetti, I.; Bellì, A.; Patrone, R.; Grassi, F.; Dell’Aversana, F.; Pilone, V.; Petriolo, A.; et al. Imaging Features of Post Main Hepatectomy Complications: The Radiologist Challenging. *Diagnoses* **2022**, *12*, 1323. [CrossRef]

153. Liu, Y.I.; Shin, L.K.; Jeffrey, R.B.; Kamaya, A. Quantitatively defining washout in hepatocellular carcinoma. *AJR Am. J. Roentgenol.* **2013**, *200*, 84–89. [CrossRef]

154. Mähringer-Kunz, A.; Steinle, V.; Dübèr, C.; Weinmann, A.; Koch, S.; Schmidtmann, I.; Schotten, S.; Hinrichs, J.B.; Graafen, D.; dos Santos, D.P.; et al. Extent of portal vein tumour thrombosis in patients with hepatocellular carcinoma: The more, the worse? *Liver Int.* **2019**, *39*, 324–331. [CrossRef] [PubMed]

155. van der Pol, C.B.; McInnes, M.D.F.; Salameh, J.P.; Levis, B.; Cherryak, V.; Sirlin, C.B.; Bashir, M.R.; Allen, B.C.; Burke, L.M.B.; Choi, J.Y.; et al. CT/MRI and CEUS LI-RADS Major Features Association with Hepatocellular Carcinoma: Individual Patient Data Meta-Analysis. *Radiology* **2022**, *302*, 326–335. [CrossRef]

156. Motosugi, U.; Ichikawa, T.; Sou, H.; Sano, K.; Tominaga, L.; Muhi, A.; Araki, T. Distinguishing hypervascular pseudolesions of the liver from hypervascular hepatocellular carcinomas with gadoxetic acid-enhanced MR imaging. *Radiology* **2010**, *256*, 151–158. [CrossRef]

157. Shin, J.; Lee, S.; Yoon, J.K.; Chung, Y.E.; Choi, J.Y.; Park, M.S. LI-RADS Major Features on MRI for Diagnosing Hepatocellular Carcinoma: A Systematic Review and Meta-Analysis. *J. Magn. Reason. Imaging* **2021**, *54*, 518–525. [CrossRef]

158. Tang, A.; Singal, A.G.; Mitchell, D.G.; Hecht, E.M.; Fowler, K.J.; Kulik, L.; Parikh, N.D.; Kono, Y.; Sirlin, C.B. Introduction to the Liver Imaging Reporting and Data System for Hepatocellular Carcinoma. *Clin. Gastroenterol. Hepatol.* **2019**, *17*, 1228–1238. [PubMed]

159. Holland, A.E.; Hecht, E.M.; Hahn, W.Y.; Kim, D.C.; Babb, J.S.; Lee, V.S.; West, A.B.; Krinsky, G.A. Importance of small (<or = 20-mm) enhancing lesions seen only during the hepatic arterial phase at MR imaging of the cirrhotic liver: Evaluation and comparison with whole explanted liver. *Radiology* **2005**, *237*, 934–944.

160. Jang, H.J.; Kim, T.K.; Khalili, K.; Yazdi, L.; Menezes, R.; Park, S.H.; Sherman, M. Characterization of 1-to 2-cm liver nodules detected on hcc surveillance ultrasound according to the criteria of the American Association for the Study of Liver Disease: Is quadriphasic CT necessary? *AJR Am. J. Roentgenol.* **2013**, *201*, 314–321. [CrossRef] [PubMed]

161. Luca, A.; Caruso, S.; Milazzo, M.; Mamone, G.; Marrone, G.; Miraglia, R.; Maruzzelli, L.; Carollo, V.; Minervini, M.I.; Vizzini, G.; et al. Multidetector-row computed tomography (MDCT) for the diagnosis of hepatocellular carcinoma in cirrhotic candidates for liver transplantation: Prevalence of radiological vascular patterns and histological correlation with liver explants. *Eur. Radiol.* **2010**, *20*, 898–907. [CrossRef] [PubMed]

162. Chernyak, V.; Fowler, K.J.; Kamaya, A.; Kielar, A.Z.; Elsayes, K.M.; Bashir, M.R.; Kono, Y.; Do, R.K.; Mitchell, D.G.; Singal, A.G.; et al. Liver Imaging reporting and data system (LI-RADS) version 2018: Imaging of hepatocellular carcinoma in at-risk patients. *Radiology* **2018**, *289*, 816–830. [CrossRef]

163. Kamath, A.; Roudenko, A.; Hecht, E.; Sirlin, C.; Chernyak, V.; Fowler, K.; Mitchell, D.G. CT/MR LI-RADS 2018: Clinical implications and management recommendations. *Abdominal. Radiol.* **2019**, *44*, 1306–1322. [CrossRef]

164. Paoletti, M.; Muzic, S.I.; Marchetti, F.; Farina, L.M.; Bastianello, S.; Pichiecchio, A. Diferential imaging of atypical demyelinating lesions of the central nervous system. *Radiol. Med.* **2021**. [CrossRef] [PubMed]

165. Nazari, M.; Shiri, I.; Hajianfar, G.; Oveisì, N.; Abdollahi, H.; Deebband, M.R.; Oveisì, M.; Zaidì, H. Noninvasive Fuhrman grading of clear cell renal cell carcinoma using computed tomography radiomic features and machine learning. *Radiol. Med.* **2020**, *125*, 754–762. [CrossRef] [PubMed]

166. Fusco, R.; Granata, V.; Mazzei, M.A.; Meglio, N.D.; Roscio, D.D.; Moroni, C.; Montì, R.; Cappabianca, C.; Picone, C.; Neri, E.; et al. Quantitative imaging decision support (QIDS™) tool consistency evaluation and radiomic analysis by means of 594 metrics in lung carcinoma on chest CT scan. *Cancer Control* **2021**, *28*, 1073274820985786. [CrossRef] [PubMed]

167. Cellina, M.; Pirovano, M.; Ciocca, M.; Gibelli, D.; Floridi, C.; Oliva, G. Radiomic analysis of the optic nerve at the first episode of acute optic neuritis: An indicator of optic nerve pathology and a predictor of visual recovery? *Radiol. Med.* **2021**, *126*, 698–706. [CrossRef] [PubMed]
168. Dietrich, C.F.; Nolsoe, C.P.; Barr, R.G.; Berzigotti, A.; Burns, P.N.; Cantisani, V.; Chammas, M.C.; Chaubal, N.; Choi, B.I.; Clevert, D.-V.; et al. Guidelines and Good Clinical Practice Recommendations for Contrast Enhanced Ultrasound (CEUS) in the liver-update 2020 WFUMB in cooperation with EFSUMB, AFSUMB, AIUM, and FLAUS. Ultrasound Med. Biol. 2020, 46, 2579–2604. [CrossRef]

169. Jo, P.C.; Jang, H.J.; Burns, P.N.; Burak, K.W.; Kim, T.K.; Wilson, S.R. Integration of contrast-enhanced US into a multimodality approach to imaging of nodules in a cirrhotic liver: How i do it. Radiology 2017, 282, 317–331. [CrossRef]

170. Kielar, A.Z.; Chernyak, V.; Bashir, M.R.; Do, R.K.; Fowler, K.J.; Mitchell, D.G.; Cerny, M.; Elsaies, K.M.; Santillan, C.; Kamaya, A.; et al. LI-RADS 2017: An update. J. Magn. Reson. Imaging 2018, 47, 1449–1474. [CrossRef]

171. An, C.; Lee, C.H.; Byun, J.H.; Lee, M.H.; Jeong, W.K.; Choi, S.H.; Lim, Y.-S.; Kim, Y.S.; Kim, J.H.; et al. Intraindividual comparison between gadoxetate-enhanced magnetic resonance imaging and dynamic computed tomography for characterizing focal hepatic lesions: A multicentre, multireader study Korean. J. Radiol. 2019, 20, 1616–1626. [CrossRef]

172. Kim, Y.Y.; Kim, M.J.; Kim, E.H.; Roh, Y.H.; An, C. Hepatocellular carcinoma versus other hepatic malignancy in cirrhosis: Performance of LI-RADS version 2018. Radiology 2019, 291, 72–80. [CrossRef]

173. Cho, J.Y.; Lee, J.M.; Sirln, C.B. CT and MR imaging diagnosis staging of hepatocellular carcinoma: Part II Extracellular agents, hepatobiliary agents, and ancillary imaging features. Radiology 2014, 273, 30–50. [CrossRef]

174. Schima, W.; Heiken, J. LI-RADS v2017 for liver nodules: How we read and report. Cancer Imaging 2018, 18, 14. [CrossRef] [PubMed]

175. Ishigami, K.; Yoshimitsu, K.; Nishihara, Y.; Irie, H.; Asayama, Y.; Tajima, T.; Nishie, A.; Hirakawa, M.; Ushijima, Y.; Okamoto, D.; et al. Hepatocellular carcinoma with a pseudocapsule on gadolinium-enhanced MR images: Correlation with histopathologic findings. Radiology 2009, 250, 435–443. [CrossRef] [PubMed]

176. De Gaetano, A.M.; Catalano, M.; Pompli, M.; Marini, M.G.; Rodríguez Carrero, P.; Gulli, C.; Infante, A.; Iezzi, R.; Ponziani, F.R.; Cerrito, L.; et al. Critical analysis of major and ancillary features of LI-RADS v2018 in the differentiation of small (≤ 2 cm) hepatocellular carcinoma from dysplastic nodules with gadobenate dimeglumine-enhanced magnetic resonance imaging. Eur. Rev. Med. Pharmacol. Sci. 2019, 18, 7766–7801. [CrossRef]

177. Tang, A.; Cruite, I.; Mitchell, D.G.; Sirln, C.B. Hepatocellular carcinoma imaging systems: Why they exist, how they have evolved, and how they differ. Abdom. Radiol. 2018, 43, 3–12. [CrossRef]

178. Xie, S.; Zhang, Y.; Chen, J.; Jiang, T.; Liu, W.; Rong, D.; Sun, L.; Zhang, L.; He, B.; Wang, J. Can modified LI-RADS increase the sensitivity of LI-RADS v2018 for the diagnosis of 10–19 mm hepatocellular carcinoma on gadoxetic acid-enhanced MRI? Abdom. Radiol. 2022, 47, 596–607. [CrossRef]

179. Vernuccio, F.; Porrello, G.; Cannella, R.; Vernuccio, L.; Midiri, M.; Giannitrapani, L.; Soresi, M.; Brancatelli, G. Benign and
199. Rimola, J.; Forner, A.; Reig, M.; Vilana, R.; de Lope, C.R.; Ayuso, C.; Bruix, J. Cholangiocarcinoma in cirrhosis: Absence of contrast washout in delayed phases by magnetic resonance imaging avoids misdiagnosis of hepatocellular carcinoma. Hepatology 2020, 71, 3770–3781. [CrossRef]

200. Kang, Y.; Lee, J.M.; Kim, S.H.; Han, J.K.; Choi, B.I. Intrahepatic mass-forming cholangiocarcinoma: Enhancement patterns on gadoxetate dimeglumine-enhanced MRI. Radiology 2018, 288, 118–128. [CrossRef]

201. Sheng, R.F.; Zeng, M.S.; Rao, S.X.; Ji, Y.; Chen, L.L. MRI of small intrahepatic mass-forming cholangiocarcinoma and atypical small hepatocellular carcinoma (≤3 cm) with cirrhosis and chronic viral hepatitis: A comparative study. Clin. Imaging 2014, 38, 265–272. [CrossRef]

202. Joo, I.; Lee, J.M.; Yoon, J.H. Imaging Diagnosis of Intrahepatic and Perihilar Cholangiocarcinoma: Recent Advances and Challenges. Radiology 2018, 288, 7–13. [CrossRef]

203. Granata, V.; Grassi, R.; Fusco, R.; Setola, S.V.; Belli, A.; Ottaviano, A.; Nasti, G.; La Porta, M.; Danti, G.; Cappabianca, S.; et al. Intrahepatic cholangiocarcinoma and its differential diagnosis at MRI: How radiologist should assess MR features. Radiol. Med. 2021, 126, 1584–1600. [CrossRef]

204. Kim, S.J.; Lee, J.M.; Han, J.K.; Kim, K.H.; Lee, J.Y.; Choi, B.I. Peripheral mass-forming cholangiocarcinoma in cirrhotic liver. AJR Am. J. Roentgenol. 2007, 189, 1428–1434. [CrossRef]

205. Lee, H.S.; Kim, M.J.; An, C. How to utilize LR-M features of the LI-RADS to improve the diagnosis of combined hepatocellular-cholangiocarcinoma on gadoxetate-enhanced MRI? Eur. Radiol. 2019, 29, 2408–2416. [CrossRef] [PubMed]

206. Choi, S.H.; Lee, S.S.; Park, S.H.; Kim, K.M.; Yu, E.; Park, Y.; Shin, Y.M.; Lee, M.G. LI-RADS Classification and Prognosis of Primary Liver Cancers at Gadoxetic Acid-enhanced MRI. Radiology 2019, 290, 388–397. [CrossRef] [PubMed]

207. Tang, A.; Bashir, M.R.; Corwin, M.T.; Cruite, I.; Dietrich, C.F.; Do, R.; Ehmam, E.C.; Fowler, K.J.; Hussain, H.K.; Jha, R.C.; et al. Evidence Supporting LI-RADS Major Features for CT- and MR Imaging-based Diagnosis of Hepatocellular Carcinoma: A Systematic Review. Radiology 2018, 286, 29–48. [CrossRef] [PubMed]

208. Fowler, K.J.; Potetzke, T.A.; Hope, T.A.; Costa, E.A.; Wilson, S.R. LI-RADS M (LR-M): Definite or probable malignancy, not specific for hepatocellular carcinoma. Abdom. Radiol. 2018, 43, 149–157. [CrossRef] [PubMed]

209. Zou, X.; Luo, Y.; Morelli, J.N.; Hu, X.; Shen, Y.; Hu, D. Differentiation of hepatocellular carcinoma from intrahepatic cholangiocarcinoma and combined hepatocellular-cholangiocarcinoma in high-risk patients matched to MR field strength: Diagnostic performance of LI-RADS version 2018. Abdom. Radiol. 2021, 46, 3168–3178. [CrossRef] [PubMed]

210. Park, S.H.; Lee, S.S.; Yu, E.; Kang, H.J.; Park, Y.; Kim, S.Y.; Lee, S.J.; Shin, Y.M.; Lee, M.G. Combined hepatocellular-cholangiocarcinoma: Gadoxetic acid-enhanced MRI findings correlated with pathologic features and prognosis. J. Magn. Reson. Imaging 2017, 46, 267–280. [CrossRef] [PubMed]
211. Kim, S.S.; Lee, S.; Choi, J.Y.; Lim, J.S.; Park, M.S.; Kim, M.J. Diagnostic performance of the LR-M criteria and spectrum of LI-RADS imaging features among primary hepatic carcinomas. *Abdom. Radiol.* 2020, 45, 3743–3754. [CrossRef]

212. Fanelli, F.; Cannavale, A.; Chisci, E.; Citone, M.; Falcone, G.M.; Michelagnoli, S.; Miele, V. Direct percutaneous embolization of aneurysm sac: A safe and effective procedure to treat post-EVAR type II endoleaks. *Radiol. Med.* 2021, 126, 258–263. [CrossRef]

213. Bartolotta, T.V.; Randazzo, A.; Bruno, E.; Taibbi, A. Focal liver lesions in cirrhosis: Role of contrast-enhanced ultrasonography. *World J. Radiol.* 2022, 14, 70–81. [CrossRef]

214. Sugimoto, K.; Saito, K.; Shirata, N.; Kamiyama, N.; Sakamaki, K.; Takahashi, H.; Wada, T.; Kakegawa, T.; Tomita, Y.; Abe, M.; et al. Comparison of modified CEUS LI-RADS with sonazoid and CT/MRI LI-RADS for diagnosis of hepatocellular carcinoma. *Hepatol. Res.* 2022. Epub ahead of print. [CrossRef]

215. Catalano, O.; Sandomenico, F.; Vallone, P.; Setola, S.V.; Granata, V.; Fusco, R.; Lastoria, S.; Mansi, L.; Petrillo, A. Contrast-Enhanced Ultrasound in the Assessment of Patients with Indeterminate Abdominal Findings at Positron Emission Tomography Imaging. *Ultrasound Med. Biol.* 2016, 42, 2717–2723. [CrossRef]

216. Avallone, A.; Pecori, B.; Bianco, F.; Aloj, L.; Tatangelo, F.; Romano, C.; Granata, V.; Marone, P.; Leone, A.; Botti, G.; et al. Critical role of bevacizumab scheduling in combination with pre-surgical chemo-radiotherapy in MRI-defined high-risk locally advanced rectal cancer: Results of the BRANCH trial. *Oncotarget* 2015, 6, 30394–30407, PMCID:PMC4745808. [CrossRef] [PubMed]

217. Granata, V.; Fusco, R.; Catalano, O.; Avallone, A.; Palaia, R.; Botti, G.; Tatangelo, F.; Granata, F.; Cascella, M.; Izzo, F.; et al. Diagnostic accuracy of magnetic resonance, computed tomography and contrast-enhanced ultrasound in radiological multimodality assessment of peribiliary liver metastases. *Plos ONE* 2017, 12, e0179951. [CrossRef]

218. Schellhaas, B.; Hammon, M.; Strobel, D.; Pfeifer, L.; Kielisch, C.; Göertz, R.S.; Cavallaro, A.; Janka, R.; Neurath, M.F.; Uder, M.; et al. Interobserver and intermodality agreement of standardized algorithms for non-invasive diagnosis of hepatocellular carcinoma in high-risk patients: CEUS-LI-RADS versus MRI-LI-RADS. *Eur. Radiol.* 2018, 28, 4254–4264, Epub 19 April 2018. [CrossRef] [PubMed]

219. Bartolotta, T.V.; Terranova, M.C.; Gagliardo, C.; Taibbi, A. CEUS LI-RADS: A pictorial review. *Radiographics* 2020, 40, 1253–1266. [CrossRef] [PubMed]

220. Wilson, S.R.; Lyshchik, A.; Piscaglia, F.; Cosgrove, D.; Jang, H.J.; Sirlin, C.; Dietrich, C.F.; Kim, T.K.; Willmann, J.K.; Kono, Y. CEUS LI-RADS: Algorithm, implementation, and key differences from CT/MRI. *Abdom. Radiol.* 2018, 43, 127–142. [CrossRef] [PubMed]

221. Kim, T.K.; Jang, H.J.; Wilson, S.R. Benign liver masses: Imaging with microbubble contrast agents. *Ultrasound Q.* 2006, 22, 31–39. [PubMed]

222. Vilgrain, V.; Boulos, L.; Vullierme, M.P.; Denys, A.; Terris, B.; Menu, Y. Imaging of atypical hemangiomas of the liver with pathologic correlation. *Radiographics* 2000, 20, 379–397. [CrossRef]

223. Maruyama, H.; Takahashi, M.; Ishibashi, H.; Yoshikawa, M.; Yokosuka, O. Contrast-enhanced ultrasound for characterisation of hepatic lesions appearing non-hypervascular on CT in chronic liver diseases. *Br. J. Radiol.* 2012, 85, 351–357. [CrossRef]

224. Kim, T.K.; Noh, S.Y.; Wilson, S.R.; Kono, Y.; Piscaglia, F.; Jang, H.J.; Lyshchik, A.; Dietrich, C.F.; Willmann, J.K.; Vezeridis, A.; et al. Contrast-enhanced ultrasound (CEUS) liver imaging reporting and data system (LI-RADS) 2017—a review of important differences compared to the CT/MRI system. *Clin. Mol. Hepatol.* 2017, 23, 280–289. [CrossRef] [PubMed]

225. Huang, J.Y.; Li, J.W.; Ling, W.W.; Li, T.; Luo, Y.; Liu, J.B.; Lu, Q. Can contrast enhanced ultrasound differentiate intrahepatic cholangiocarcinoma from hepatocellular carcinoma? *World J. Gastroenterol.* 2020, 26, 3938–3951. [CrossRef] [PubMed]

226. Kieler, A.; Fowler, K.J.; Lewis, S.; Yaghmai, V.; Miller, F.H.; Yarmohammadi, H.; Kim, C.; Chernyak, V.; Yokoo, T.; Meyer, J.; et al. Locoregional therapies for hepatocellular carcinoma and the new LI-RADS treatment response algorithm. *Abdom. Radiol.* 2018, 43, 218–230. [CrossRef] [PubMed]

227. Granata, V.; Grassi, R.; Fusco, R.; Setola, S.V.; Belli, A.; Piccirillo, M.; Pradella, S.; Giordano, M.; Cappabianca, S.; Brunese, L.; et al. Abbreviated MRI Protocol for the Assessment of Ablated Area in HCC Patients. *Hepatol. Res.* 2016, 46, 1655–1662. [CrossRef] [PubMed]

228. Capone, F.; Costantini, S.; Guerriero, E.; Calemma, R.; Napolitano, M.; Scala, S.; Izzo, F.; Castello, G. Serum cytokine levels in patients with hepatocellular carcinoma. *Eur. Cytokine Netw.* 2010, 21, 99–104. [CrossRef]
234. Perrone, F.; Gallo, C.; Daniele, B.; Gaeta, G.B.; Izzo, F.; Capuano, G.; Adinolfi, L.E.; Mazzanti, R.; Farinati, F.; Elba, S.; et al. Cancer of Liver Italian Program (CLIP) Investigators. Tamoxifen in the treatment of hepatocellular carcinoma: 5-year results of the CLIP-1 multicentre randomised controlled trial. *Curr. Pharm. Des.* 2002, 8, 1013–1019. [CrossRef] [PubMed]

235. Bimonte, S.; Barbieri, A.; Rea, D.; Palma, G.; Luciano, A.; Cuomo, A.; Arra, C.; Izzo, F. Morphine Promotes Tumor Angiogenesis and Increases Breast Cancer Progression. *Biomed. Res. Int.* 2015, 2015, 161508. [CrossRef] [PubMed]

236. Polesel, J.; Talamini, R.; Montella, M.; Maso, L.D.; Crovatto, M.; Parpinel, M.; Izzo, F.; Tommasi, L.G.; Serraino, D.; La Vecchia, C.; et al. Nutrients intake and the risk of hepatocellular carcinoma in Italy. *Eur. J. Cancer* 2007, 43, 2381–2387. [CrossRef] [PubMed]

237. Pignata, S.; Gallo, C.; Daniele, B.; Elba, S.; Giorgio, A.; Capuano, G.; Adinolfi, L.E.; De Sio, I.; Izzo, F.; Farinati, F.; et al. CLIP Investigators. Characteristics at presentation and outcome of hepatocellular carcinoma (HCC) in the elderly. A study of the Cancer of the Liver Italian Program (CLIP). *Crit. Rev. Oncol. Hematol.* 2006, 59, 243–249. [CrossRef] [PubMed]

238. Chaudhry, M.; McGinty, K.A.; Mervak, B.; Lerebours, R.; Li, C.; Shropshire, E.; Ronald, J.; Commander, L.; Hertel, J.; Luo, S.; et al. The LI-RADS version 2018 MRI treatment response algorithm: Evaluation of ablated hepatocellular carcinoma. *Radiology* 2020, 294, 320–326. [CrossRef]

240. Shropshire, E.L.; Chaudhry, M.; Miller, C.M.; Allen, B.C.; Bozdogan, E.; Cardona, D.M.; King, L.Y.; Janas, G.L.; Do, R.K.; Kim, C.Y.; et al. LI-RADS Treatment Response Algorithm: Performance and Diagnostic Accuracy. *Radiology* 2019, 292, 226–234. [CrossRef]

241. Agostini, A.; Borgheresi, A.; Mari, A.; Floridi, C.; Bruno, F.; Carotti, M.; Schicchi, N.; Barile, A.; Maggi, S.; Giovagnoni, A. Dual-energy CT: Theoretical principles and clinical applications. *Radiol. Med.* 2019, 124, 1281–1295. [CrossRef] [PubMed]

242. Masciocchi, C.; Conti, L.; D’Orazio, F.; Conchiglia, A.; Lanni, G.; Barile, A. Errors in Musculoskeletal MRI. In *Errors in Radiology*; Romano, L., Pinto, A., Eds.; Springer: Milano, Italy, 2012. [CrossRef]