INTRODUCTION

Parkinson’s disease (PD) is the second-most common neurodegenerative disorder and is characterized by the progressive degeneration of dopaminergic (DA) neurons and a decrease in striatal dopamine. PD is associated with clinical movement disorders, including a tremor at rest, rigidity of the limbs, bradykinesia (slowness and paucity of voluntary movement), and postural instability (a tendency to fall even in the absence of...
weakeness or cerebellar balance disturbance) [1-3]. Although we do not fully understand the etiology of PD, accumulating evidence suggests that microglia, which are the resident immune cells of the brain, are crucial mediators of the brain inflammatory processes that lead to neurotoxicity, and that excessive microglial activation contributes to the initiation and progression of PD [3-5]. However, it is largely unknown what endogenous biomolecules initiate and stimulate microglial activation, even though the control of microglial activators, which stimulate neurotoxic inflammation, may be a useful strategy for the prevention of the degeneration of the nigrostriatal DA projection in the adult brain.

Toll-like receptors (TLRs) are pattern recognition receptors that recognize specific pathogen-associated molecular signatures and subsequently initiate inflammatory and immune responses [3, 6]. TLR4 recognizes various ligands, such as lipopolysaccharide (LPS), envelope proteins, heat-shock proteins, fibrinogen, and hyaluronan [3, 6]. The activation of TLR4 in immune cells induces increases in the levels of inflammatory cytokines [3, 7]. Although the pattern of TLR expression in the brain is controversial, there are many reports suggesting that microglia are important cells for TLR4-mediated immune responses, which may be involved in neurodegenerative diseases such as Alzheimer's disease (AD) and PD [8-10]. Moreover, increases in TLR4 expression have been observed in α-synuclein-overexpressing transgenic mice and in patients with multiple system atrophy [11], although alterations in TLR4 expression in patients with PD are still unclear. These results suggest that an increase in microglial TLR4 may be crucial for the pathogenesis of PD, and that the discovery of endogenous molecules involved in the induction of microglial TLR4 may be useful in guiding the development of knowledge-based targeted therapeutics for PD.

We previously reported that prothrombin kringle-2 (pKr-2), which is a domain of prothrombin that is generated by active thrombin, is able to induce the death of DA neurons in the rat SN through microglial activation, even though pKr-2 itself was not directly toxic to neurons [5]. Moreover, we recently found that patients with PD have increased pKr-2 expression in the SN, and that nigrostriatal DA projections may degenerate due to neurotoxic inflammation following pKr-2 upregulation-induced production of microglial TLR4 in the SN of adult murine brain [3]. These results suggest that pKr-2 might be a potential pathogenic factor in PD, and that limiting pKr-2-induced microglial activation may be an effective therapeutic strategy for protecting DA neurons in the adult brain.

**MICROGLIA AND NEUROINFLAMMATION**

Microglia are the resident immune cells in the central nervous...
system (CNS). In the resting state, microglia have small cell bodies and numerous processes and can support neuronal function and survival [22]. In pathological conditions, microglia stimulated by various activators undergo phagocytic morphological changes, which are characterized by enlarged cell bodies and short processes, and these altered microglia exert beneficial effects that repair tissue by releasing anti-inflammatory cytokines and neurotrophic factors [22, 23]. However, the major cause of neurotoxic inflammation in the CNS is the response of microglia to a variety of stimuli, such as infection, trauma, and toxins [19], and activated microglia produce neurotoxic inflammatory cytokines, such as interleukin (IL)-1β, tumor necrosis factor (TNF)-α, and IL-6 [22, 24]. In addition, activated microglia can produce reactive oxygen species (ROS), such as O$_2^-$ and O$_3^-$ derived oxidants, via the activation of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, which is also involved in neuroinflammatory processes [25]. Moreover, under proinflammatory conditions, microglia can release more ROS than neurons [26]. Excessive ROS production is implicated in neurodegenerative diseases [27]. Accordingly, many activated microglia are observed in post-mortem tissue from patients with PD [3]. In addition, highly increased levels of proinflammatory cytokines, such as TNF-α, IL-1β and interferon-γ, are also found in the SN of PD patients [3, 19, 20]. Similar to the increases in proinflammatory cytokines, the levels of NADPH oxidase are upregulated in microglia in the post-mortem midbrain tissue of PD patients [28].

Consistent with the observation of increases in NADPH oxidase and neurotoxic cytokines in the brains of patients with PD [3, 19, 20, 28], there are many reports indicating the significance of these factors involved in the degeneration of the nigrostriatal DA system in animal models of PD. 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) was discovered to be a neurotoxic compound in drug addicts who showed parkinsonian symptoms [29] and has been widely used in the production of PD animal models [30]. In the SN of adult mice, MPTP induces microglial activation, which is involved in neurotoxicity [31]. The blockade of microglial activation following treatment with minocycline, a broad-spectrum tetracycline antibiotic, inhibits the production of microglial-derived deleterious factors, and exerts neuroprotective effects in the MPTP-lesioned SN [30]. These suggest that microglial activation is involved in the PD pathogenesis related to DA neuronal damage. In addition, the increased expression of gp91phox, which is an NADPH oxidase subunit, is observed in the activated microglia of MPTP-injected mice [28]. In fact, the upregulation of gp91phox within activated microglia is markedly reduced by minocycline treatment, which leads to the protection of DA neurons in vivo [28].

6-hydroxydopamine (6-OHDA) is also often used to produce PD models for the testing symptomatic therapies and the study of the mechanisms of DA neuronal death [1]. Since the structure of 6-OHDA is similar to that of dopamine, it is taken up into DA neurons by the dopamine transporter, where it acts as a neuronal toxin by generating ROS [30]. In addition to direct neuronal toxicity, 6-OHDA-induced neurotoxicity can promote microglial activation in nigrostriatal DA projection areas [32, 33], resulting in the production of neurotoxic proinflammatory cytokines [34, 35]. In addition, microglial activation induced by 6-OHDA treatment parallels the activation of NADPH oxidase, including the p47phox and gp91phox subunits, in nigral microglia [36]. Similar to the effects of MPTP, the 6-OHDA-induced toxic effects in DA neurons are attenuated by minocycline treatment [37]. Taken together, these results suggest that microglial-derived neuroinflammation, which includes the production of neurotoxic cytokines and activation of NADPH oxidase within microglia, may play a crucial role in DA neuronal degeneration in animal models of PD. In addition to MPTP and 6-OHDA, studies using rotenone, which is an odorless, colorless, crystalline isoflavone used as a broad-spectrum insecticide, piscicide, and pesticide, have also indicated that the production of neurotoxic cytokines and activation of NADPH oxidase in microglia play important roles in PD pathogenesis [38, 39].

The above observations indicate that excessive increases in proinflammatory mediators and ROS production following microglial activation lead to severe neurotoxicity, resulting in the deterioration of the DA system in the adult brain. While numerous studies have examined the development of therapeutic and preventive agents against PD, there are currently no cures that stop or slow down the progressive degeneration of DA neurons and PD symptoms. Therefore, efforts to develop therapeutic and preventive agents for PD need to aim at inhibiting microglial-derived neuroinflammation, which would suppress microglial activation and its pathogenic mechanisms.

**CONTROL OF NEUROINFLAMMATION BY MICROGLIAL TLR4**

TLR4 initiates the activation of the innate immune response by recognizing pathogens as a pattern-recognition receptor. LPS is a representative ligand for TLR4 [40]. The TLR4-mediated activation of NF-κB and mitogen-activated protein kinases that occurs via myeloid differentiation factor 88-dependent and -independent pathways induces the expression of proinflammatory cytokines and chemokines [6, 41, 42]. TLR4, which is highly expressed in the brain, is stimulated by LPS in microglia [43, 44]. Neuroinflammation following microglial TLR4

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activation has been implicated in neurodegenerative disease. The knockout of TLR4 protects against traumatic brain injury [45], focal cerebral ischemic injury [46], and pertussis toxin-induced experimental autoimmune encephalomyelitis [47]. In AD, beta-amyloid (Aβ) deposition promotes inflammatory reactions, and TLR4 is upregulated in the glial cells surrounding the Aβ plaques in the brains of patients with AD [48]. The destructive mutation of TLR4 inhibits microglial activation by Aβ deposition, which results in a significant decrease in the levels of proinflammatory cytokines and chemokines [9, 48]. These findings suggest that TLR4 is essential for the Aβ-induced release of proinflammatory cytokines and chemokines, which is a hallmark of microglial activation and neurotoxicity [8, 9, 48, 49]. Microglial TLR4 is also associated with neuroinflammation in PD. Recent reports indicate that increased expression of microglial TLR4 is found in the post-mortem tissue of patients with PD [3]. In addition, TLR4 deficiency attenuates MPTP-induced neurotoxicity and is correlated with the inhibition of microglial activation in the SN [50]. Alpha-synuclein contributes to microglial activation during PD progression. The generation of ROS and proinflammatory cytokines is reduced in TLR4-deficient microglia after treatment with α-synuclein [10]. Moreover, LPS-induced ROS generation can be mediated by direct interaction between microglial TLR4 and NADPH oxidase, suggesting that microglial TLR4 may be a beneficial target in inhibiting NADPH oxidase activity [51]. Therefore, the control of microglial TLR4 expression may be a potential and important target for therapeutic intervention in neurodegenerative disease.

PRODUCTION OF pKr-2 AND ITS GENERAL ROLES in vivo

Prothrombin, which is known to be synthesized mainly in the liver and then secreted into the bloodstream [52], is cleaved to produce fragment 1-2 (kringle regions) and thrombin during activation [53, 54]. Activated thrombin, a serine protease which converts soluble fibrinogen into insoluble fibrin for blood coagulation, can cleave pKr-1 and -2 (Fig. 1) [55]. The functions of pKr-2 are well-established in the field of angiogenesis. For instance, pKr-2, which is purified from LPS-treated rabbit serum, acts as an angiogenic inhibitor during bovine capillary endothelial cell proliferation [56]. Treatment with pKr-2 induces the suppression of basic fibroblast growth factor-triggered endothelial cell growth and angiogenesis in the chorioallantoic membrane of chick embryos [57]. Moreover, treatment with pKr-2 inhibits endothelial cell proliferation and angiogenesis by inactivating the cyclin D1/cyclin-dependent kinase 4 (CDK4) complex through the induction of ROS production and upregulation of nuclear CDK inhibitors [58]. It also inhibits fibrin formation and platelet aggregation by binding to thrombin and inducing conformational changes at its active site, resulting in a reduction in the clotting activity of thrombin [59]. In addition, recombinant human pKr-2 reduces the immunoreactivity of matrix metalloproteinases 2 and 9 and the expression of vascular endothelial growth factor, which results in the inhibition of B16F10 melanoma cell metastasis [60].

Although many studies have investigated the functions of pKr-2 [56-60] and prothrombin is expressed in brain tissues [52], few reports have examined the roles of pKr-2 in the CNS. The accumulation of prothrombin and thrombin, which might be due to blood-brain barrier leakage, has been shown in the brains of patients with PD and AD [61-63], which suggests a possible increase in pKr-2 expression. Moreover, we previously reported that the upregulation of pKr-2 could contribute to microglial activation, resulting in neurodegeneration in the SN of murine brains [3]. Therefore, these results suggest that pKr-2 expression is increased in the lesioned brain and that its upregulation is involved in neurotoxic effects in the adult brain.

pKr-2 AS AN ENDOGENOUS PATHOGEN IN THE NIGROSTRIATAL DA SYSTEM

pKr-2 can trigger microglial activation, resulting in the production of neurotoxic cytokines such as TNF-α and IL-1β, and inflammatory mediators such as inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 [5, 64, 65]. In addition, pKr-2-activated microglia produce O2− and O− deriving oxidants through the activation of NADPH oxidase, which is also involved in neurotoxic events in the murine cortex [64]. Moreover, we recently
suggested that pKr-2 upregulation is involved in the pathogenesis of PD, and that the control of microglial pKr-2 expression and pKr-2-induced microglial TLR4 overexpression might be important for protecting the nigrostriatal DA system against PD [3]. These results help the understanding of the relationships among pKr-2, microglia, and TLR4 (Fig. 2). To ascertain whether pKr-2 is involved in PD via the activation of microglial and microglial TLR4-mediated neuroinflammatory responses, we quantitatively analyzed pKr-2 levels in the SN of patients with PD and found that TLR4 was involved in pKr-2-induced microglial activation (Fig. 2). The expression levels of pKr-1-2, which is a precursor of active thrombin-cleaved pKr-2, and pKr-2 were significantly increased in the post-mortem brain tissue of patients with PD \[p=0.018 \text{ and } p=0.008 \text{ vs. age-matched controls (CON), respectively; Fig. 2A and 2B}\]. As previously reported [3], 41% of the pKr-2 expression was localized within ionized calcium binding adaptor molecule 1 (Iba1)-positive microglia in the SN of patients with PD (Fig. 2C; data not shown). Moreover, the levels of Iba1 expression were significantly increased in the SN of pKr-2-treated wild-type (WT) mice \[p<0.001 \text{ vs. phosphate buffered saline (PBS)-treated WT mice; Fig. 2D}\]. However, this increase was significantly reduced by TLR4 deficiency \[p=0.011 \text{ vs. pKr-2-treated WT mice; Fig. 2D}\]. Taken together, these results suggest that an increase in pKr-2 expression is involved in microglial activation via TLR4 upregulation in the SN of adult brain (Fig. 2), even though it is unclear whether pKr-2-specific receptors exist in microglia and how pKr-2 induces an increase in microglial TLR4.

**CONTROL OF pKr-2-INDUCED NEUROTOXICITY**

Since inhibiting inflammation is an important strategy for the prevention of neuronal damage in neurodegenerative disease, therapies for the control of the activation of microglia have been proposed. Although the exact mechanisms are unknown, our

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**Fig. 2.** The expression of pKr-2 in the substantia nigra (SN) of patients with Parkinson’s disease (PD); (A, B) Western blot analysis for pKr-1-2 and pKr-2, respectively, in the human SN. Patients with PD had increases in the levels of pKr-1-2 (A) and pKr-2 (B) in the SN compared to age-matched controls (CON). \[p=0.018 \text{ vs. CON (t-test; n=4, each group)}; \]

\[p=0.008 \text{ vs. CON (t-test; n=3, each group)}\]. (C) The immunofluorescence staining for pKr-2 (red) and Iba1 (green) indicates that the expression of pKr-2 is co-localized within Iba1-positive cells in the SN of patients with PD. Scale bar, 10 μm. (D) Western blot analysis of Iba1 expression in the pKr-2-treated SN. Three days after treatment with pKr-2 (24 μg/2 μl) or phosphate-buffered saline (PBS, vehicle control), the levels of pKr-2 increased Iba1 expression were significantly decreased in TLR4 knockout (TLR4 \[−\]) mice compared to wild type (WT) mice. \[p<0.001 \text{ vs. PBS-treated mice (CON); } \]

\[p=0.011 \text{ vs. pKr-2-treated WT mice (one-way ANOVA and Tukey’s post hoc analysis; n=4, each group)}\].

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recent results strongly suggest that TLR4 and pKr-2 are closely associated with neuroinflammation in PD [3]. Similar to our results involved in neurodegeneration in the DA system, previous studies have shown that TLR4 expression is upregulated in the MPTP-treated animal model of PD [66] and that microglial TLR4 is directly activated by α-synuclein treatment [10]. Taken together, these results suggest that the modulation of microglial TLR4 by controlling pKr-2 production may provide us with important clues regarding the mechanisms responsible for inflammation-associated neurodegeneration in PD and open innovative therapeutic perspectives for the treatment of PD. For instance, treatment with minocycline has neuroprotective effects in preclinical studies of neurodegenerative diseases [2, 30, 67-71]. In particular, treatment with minocycline protects DA neurons against pKr-2-induced neurotoxicity through the inhibition of inflammatory responses in the brains of adult mice [2]. Moreover, its treatment reduces the expression of proinflammatory cytokines and iNOS, which is significantly increased by activated microglia following pKr-2 upregulation. These results suggest that the development of efficient anti-inflammatory drugs against pKr-2 may be useful for protecting DA neurons in the SN of lesioned adult brain.

CONCLUSION

The discovery of endogenous biomolecules that initiate and stimulate microglial activation and result in neurodegeneration via neurotoxic inflammatory events in the adult brain is very important, as the control of microglial activators may be a useful strategy in preventing the degeneration of the nigrostriatal DA projection in the adult brain [3, 5]. In addition, the discovery of endogenous molecules involved in the induction of microglial TLR4, which may be crucial for the pathogenesis of PD, may also be useful in guiding the development of knowledge-based targeted therapeutics for PD. pKr-2 upregulation may lead to the disruption of the nigrostriatal DA projection by microglial activation, resulting in the production of neurotoxic inflammatory biomolecules, such as ROS following NADPH oxidase activation, and proinflammatory cytokines, such as TNF-α and IL-1β [3, 5, 64]. Moreover, pKr-2 expression is significantly increased and co-localized in activated microglia in the SN of patients with PD, and the upregulation of microglial TLR4 might be a key mechanism for pKr-2-induced neurotoxic inflammation in the nigrostriatal DA system of murine brain [3]. Although it is still unclear how pKr-2 is translocated within microglia and further studies are needed to clarify the relationship between pKr-2 and the transcription factors involved in the induction of microglial TLR4 (Fig. 3), our results suggest that pKr-2 upregulation in the SN may be a potential pathogenic mechanism in PD, and that the induction of microglial TLR4 following pKr-2 overexpression may be an important target mechanism in the development of therapeutics against pKr-2-induced neurodegeneration in the nigrostriatal DA system of the adult brain.

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**Fig. 3.** Schematic representation of pKr-2-mediated neurotoxicity in the SN. Breakdown of the blood-brain barrier may be involved in an influx of prothrombin, which consequently resulting in the production of thrombin and pKr-2 in the SN. The increased pKr-2 may be translocated within microglia, and microglial activation may be mediated by the induction of microglial TLR4 following pKr-2 translocation. Activated microglia can produce neurotoxic cytokines and reactive oxygen species (ROS). These neurotoxic factors induce dopaminergic (DA) neuronal death.
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