Late starting treadmill exercise improves spatial leaning ability through suppressing CREP/BDNF/TrkB signaling pathway following traumatic brain injury in rats

Il-Gyu Ko1,2, Sung-Eun Kim1,2, Lakkyong Hwang1,2, Jun-Jang Jin1,2, Chang-Ju Kim1,2, Bo-Kyun Kim3, Hong Kim4,*

1Department of Physiology, College of Medicine, Kyung Hee University, Seoul, Korea
2Kohwang Medical Research Institute, Kyung Hee University, Seoul, Korea
3Department of Emergency Technology, College of Health Science, Gachon University, Incheon, Korea
4Department of Oriental Sports Medicine, College of Biomedical Science, Daegu Haany University, Gyeongsan, Korea

INTRODUCTION

Traumatic brain injury (TBI) is known to results in deficits in spatial learning and memory function (Kim et al., 2010a; Shen et al., 2013). In generally, TBI causes cell death and neurological dysfunction through direct physical disruptions of tissue or pathways (primary injury) as well as through delayed biochemical changes induced by trauma (secondary injury). TBI triggers a complex cascade of apoptotic events which can contribute to delayed secondary injury processes.

Apoptosis, or programmed cell death, is a physiological form of cell death that is important for normal embryologic development and cell turnover in adult organisms. Apoptosis plays a crucial role in delayed neuronal loss after both acute and chronic brain injury. In the TBI-induced apoptosis, Bcl-2 family plays as the key executioner of apoptosis and caspase family acts as the final pathway of apoptosis (Kim et al., 2010a; Park et al., 2014; Zhang et al., 2005).

Brain-derived neurotrophic factor (BDNF) and its receptor, tyrosine kinase B (TrkB), are broadly expressed in the mammalian brain. BDNF/TrkB signaling is critical for neuronal survival, morphogenesis, and plasticity (Numakawa et al., 2010). Activ...
tion of BDNF/TrkB signaling protects against TBI through cAMP-response element binding protein (CREB) cascade (Wu et al., 2014). CREB is a transcription factor in the brain and it is implicated in neuronal survival and spatial learning ability (Griesbach et al., 2009; Shen et al., 2013).

Physical activity ameliorates neurological dysfunction after TBI through inhibition of apoptosis, attenuation of neuro-inflammation and oxidative stress, and improving neuro-restoration (Archer et al., 2012; Piao et al., 2013). However, the effect of exercise on TBI is controversial, depending on the time of the start of the exercise.

Farmer et al. (2004) defined that appropriate time window for voluntary exercise after brain injury is dependent on injury severity in which an increase in injury severity prolongs the period of nonresponsiveness to voluntary exercise. Voluntary exercise endogenously upregulates BDNF and enhances recovery when it was delayed after TBI, however, when exercise was rapidly administered to TBI, the molecular response to exercise is disrupted and recovery might be delayed (Griesbach et al., 2004). Early started exercise may be deleterious by exacerbating posttraumatic symptomology and disrupting restorative processes (Griesbach, 2011).

In the present study, we investigated the effects of late starting treadmill exercise on spatial learning ability in relations with CREB- BDNF signaling pathway using TBI rats. For this study, radial 8-arm maze test, TUNEL (terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling) staining, caspase-3 immunohistochemistry, and western blot for Bax, Bcl-2, BDNF, TrkB, CREB, and p-CREB were performed.

MATERIALS AND METHODS

Animals and treatments

Male Sprague-Dawley rats, weighing 90±5 g (4 weeks old), were used for the experiments. The experimental procedures were performed in accordance with the animal care guidelines of the National Institutes of Health and the Korean Academy of Medical Sciences. This study was approved by the Kyung Hee University Institutional Animal Care and Use Committee (Seoul, Korea, KHUASP[SE]-17-089). The animals were randomly divided into the following four groups (n = 10 in each group): sham-operation group, sham-operation and treadmill exercise group, TBI-induced group, and TBI-induced and treadmill exercise group.

Induction of TBI

TBI was made based on the previous study (Park et al., 2014). For the induction of TBI, the rats were anesthetized with Zoletil 50 (10 mg/kg, intraperitoneally; Vibac Laboratories, Carros, France). A circular craniotomy (5.0 mm) was performed using a Dremel motor tool and a specially designed drill bit that prevented damage to the meninges and cortex (2.4 mm lateral to the midline, and 4.2 mm posterior to the coronal suture). The contusion injury was created with an electromagnetic contusion device (Impact One, Stereotaxic Impactor; MyNeurolab, St. Louis, MO, USA) using a sterile stainless steel impactor tip (3.0-mm diameter) that was activated at a velocity of 3.00 m/sec. The impactor tip was positioned above the cortex, and resulted in a 2.5-mm compression to the cortex.

Exercise protocol

The rats in the exercise groups were scheduled to run on a treadmill for 30 min once a day for 8 weeks (56 days) starting 3 weeks (21 days) after TBI induction. The rats in the exercise groups were forced to run on a motorized treadmill (Columbus Instruments, Columbus, OH, USA). The exercise load consisted of running at speeds of 2 m/min for the first 5 min, 5 m/min for the next 5 min, and then 8 m/min for the last 20 min, at an inclination of 0°.

Radial 8-arm maze test

Spatial learning ability was determined using a radial 8-arm maze apparatus, according to the previously described method (Ji et al., 2015; Kim et al., 2010b). The rats were deprived of water for 24 hr and allowed to explore the water for 5 min. The test was conducted on the 76 days after TBI-induced operation (55 days after starting the treadmill exercise). The time spent in seeking water at the end of the arms was counted. The test was terminated when a rat found water in all 8 arms or when over 8 min elapsed. Re-entry into the previously visited arms was counted as an error. The number of correct choice before the first error was also counted.

Tissue preparation

The animals were sacrificed after last treadmill running. The rats were anesthetized using Zoletil 50 (10 mg/kg, intraperitoneally; Vibac Laboratories), transcardially perfused with 50 mM phosphate-buffered saline, and fixed with a freshly prepared solution consisting of 4% paraformaldehyde in 100 mM phosphate buffer (pH, 7.4). Brains were dissected, and storage overnight same fixative, then it was transferred to 30% sucrose for cryoprotection. For the immunohistochemistry, the slices were coronal sectioned at 40 µm thick using a cryostat (Leica, Nussloch, Germany).
Immunohistochemistry for caspase-3

Immunohistochemistry was conducted to evaluate the caspase-3 expression in the hippocampal dentate gyrus, according to the previously described method (Kim et al., 2014; Ko et al., 2009). Free-floating tissue sections were incubated overnight with mouse anti-caspase-3 antibody (1:500; Santa Cruz Biotechnology, Santa Cruz, CA, USA) at a dilution of 1:1,000, and the sections were then incubated for 1 hr with biotinylated anti-mouse antibody for caspase-3 (1:200; Vector Laboratories, Burlingame, CA, USA). Next, the sections were incubated with avidin-biotin-peroxidase complex (Vector Laboratories) for 1 hr at room temperature. The sections were finally mounted onto gelatin-coated slides. The slides were air dried overnight at room temperature, and coverslips were mounted using Permount (Fisher Scientific, New Jersey, NJ, USA).

Western blotting

Western blot for the expressions of Bax, Bcl-2, BDNF, TrkB, CREB, and p-CREB were performed, according to the previously described method (Kim et al., 2014; Ko et al., 2009). The hippocampal tissues were homogenized on ice, and lysed in a lysis buffer containing 50 mM HEPES (pH, 7.5), 150 mM NaCl, 10% glycerol, 1% Triton X-100, 1 mM phenylmethylsulfonyl fluoride, 1 mM EGTA, 1.5 mM MgCl₂·6H₂O, 1 mM sodium orthovanadate, and 100 mM sodium fluoride. Protein content was measured using a Bio-Rad colorimetric protein assay kit (Hercules, CA, USA). Protein samples (30 μg) were separated on sodium dodecyl sulfate-polyacrylamide gel and transferred onto a nitrocellulose membrane. The membranes were incubated with 5% skim milk in Tris-buffered saline containing 0.1% Tween-20 and then incubated overnight at 4°C with the following primary antibodies: mouse anti-β-actin antibody, mouse anti-Bcl-2 antibody, mouse anti-Bax antibody, rabbit anti-BDNF antibody, rabbit anti-TrkB antibody, rabbit anti-CREB antibody, rabbit anti-p-CREB antibody (1:1,000; Santa Cruz Biotechnology). Subsequently, membranes were incubated for 1 hr with attempt secondary antibodies (1:2,000; Vector Laboratories), and band detection was performed using the enhanced chemiluminescence detection kit (Santa Cruz Biotechnology).

Data analysis

The detected bands of Bax, Bcl-2, BDNF, TrkB, CREB, and p-CREB were calculated densitometrically using Molecular Analyst ver. 1.4.1 (Bio-Rad). The area of the granular layer of the hippocampal dentate gyrus from each slice was measured by Image-Pro Plus image analysis system (Media Cybernetics Inc., Silver Spring, MD, USA). The number of caspase-3-positive cells in the hippocampal dentate gyrus was counted hemilaterally under a light microscope (Olympus, Tokyo, Japan), and expressed as the number of cells per square millimeter of granular area in the hippocampal dentate gyrus.

Differences among the groups were evaluated using IBM SPSS Statistics ver. 21.0 (IBM Co., Armonk, NY, USA). For the comparison among the groups, one-way analysis of variance was performed followed by Duncan post hoc test. All values are expressed as the mean±standard error of the mean. Statistically significant differences were established at P < 0.05.

RESULTS

Effect of treadmill exercise on the spatial learning ability in the radial 8-arm maze test

The time of complete performance, the number of correct, and the number of error in the radial 8-arm maze test are presented in Fig. 1.
Fig. 1. These results showed that induction of TBI increased completed time, decreased number of correct, and increased number of error. In contrast, treadmill exercise for 8 weeks decreased completed time, increased correct number, and reduced error number in the TBI-induced rats.

**Effect of treadmill exercise on caspase-3 expression in the hippocampal dentate gyrus**

Photomicrographs of caspase-3-positive cells in the hippocampal dentate gyrus are presented in Fig. 2. These results showed that induction of TBI enhanced caspase-3 expression in the hippocampal dentate gyrus. In contrast, treadmill exercise for 8 weeks suppressed caspase-3 expression in the TBI-induced rats.

**Effect of treadmill exercise on Bax and Bcl-2 expressions in the hippocampus**

The relative expressions of Bax and Bcl-2 in the hippocampus are presented in Fig. 3. These results showed that induction of TBI enhanced the expression Bax and enfeebled the expression of Bcl-2. Therefore, induction of TBI enhanced the ratio of Bax to Bcl-2, represents facilitation of apoptosis in the hippocampus. However, treadmill exercise during 8 weeks suppressed the expression of Bax and enhanced the expression of Bcl-2, resulting in suppression of Bax to Bcl-2 ratio in the TBI-induced rats.

**Effect of treadmill exercise on BDNF and TrkB expressions in the hippocampus**

The relative expressions of BDNF and TrkB in the hippocampus are presented in Fig. 4. These results showed that induction of TBI enhanced BDNF and TrkB expressions in the hippocampus. In contrast, treadmill exercise during 8 weeks suppressed BDNF and TrkB expressions in the TBI-induced rats.

**Effect of treadmill exercise on the ratio of p-CREB to CREB in the hippocampus**

The relative expressions of CREB and p-CREB are presented in Fig. 5. These results showed that induction of TBI enhanced the ratio of p-CREB to CREB in the hippocampus. In contrast, treadmill exercise during 8 weeks suppressed the ratio of p-CREB to CREB in the TBI-induced rats.

**DISCUSSION**

There are many reports that exercise counteracts the cognitive deficits associated with the brain injury (Griesbach et al., 2009; Kim et al., 2010a; Ko et al., 2009). In the present study, longer complete time, less correct choice, and more error choice were observed in the TBI-induced rats, representing that TBI deteriorated spatial learning ability. Treadmill exercise shorted complete time, increased correct number, and decreased error number in the TBI-induced rats, showing that treadmill exercise alleviated TBI-induced impairment of spatial learning ability. Such a memory improving effect of physical exercise has been reported in many other studies. Griesbach et al. (2004) reported that physical exercise activated neuroplasticity mechanisms within the hippocampus and counteracted cognitive deficit induced by experimental TBI. Treadmill exercise alleviated TBI-induced short-term
Late starting treadmill exercise improves spatial learning ability in rats memory impairment by suppressing TBI-induced apoptotic neuronal cell death (Kim et al., 2010a; Park et al., 2014).

During the process of secondary injury of TBI, apoptosis-regulatory factors are implicated in the susceptibility of neurons to cell death (Zhao et al., 2015). Bax and Bcl-2 are two distinct members in the regulation of apoptosis. Bcl-2 is functionally characterized...
BDNF is altered by TBI, and this increment of BDNF might be considered as one of the mechanisms promoting neuroprotection and neuronal repair after damage (Guan et al., 2003). Similarly, up-regulation of cell proliferation and/or neurogenesis during pathological situations has been suggested as a compensatory response to TBI-induced apoptotic cell death (Kernie and Parent, 2010).

In the present study, the expressions of BDNF and TrkB in the hippocampus were increased at 11 weeks after TBI induction. The expressions of BDNF and TrkB in neurons are linked with the phosphorylation of CREB (Griesbach et al., 2009; Wu et al., 2014). BDNF/TrkB signaling is critical for promoting neuronal survival following brain damage. Activation of BDNF/TrkB signaling protects against TBI via the phosphatidylinositide 3-kinase (PI3K)/Akt, but not extracellular regulated kinase pathway, and this protective effect may be amplified via the PI3K/Akt/CREB cascade (Wu et al., 2014). Such being the cases, the present study showed that long-term after TBI induction, the ratio of p-CREB/CREB in the hippocampus was enhanced. In the present results, phosphorylation of CREB was facilitated through activation of BDNF/TrkB pathway. These mechanisms contribute for acceleration of neuronal repair following TBI. In the previous studies, neurotrophic factors were up-regulated in traumatic insult at few hours after the brain lesion and this up-regulation lasted for 4 weeks (Feliciano et al., 2014; Madathil et al., 2010).

It is still unclear whether excessive expression of neurotrophic factors including BDNF promotes or delays functional recovery from TBI. Physical exercise suppressed overexpression of neurotrophic factors following nerve injury (Byun et al., 2005; Griesbach, 2011). In the present study, treadmill exercise ameliorated TBI-induced overexpression of BDNF and TrkB, which suppressed phosphorylation of CREB in the hippocampus. Our study maintained recovery period for 3 weeks after TBI, and treadmill exercise lasted during 8 weeks, it was progressed total 11 weeks after induction of TBI. There is still controversy on the starting time of exercise after brain injury. If brain is energetically compromised, exercise-induced increment in metabolic demand may negatively impact neurological recovery (Griesbach et al., 2012).
Initiation of physical exercise in the first few days after TBI caused suboptimal induction of neuroplasticity/neuroprotective pathways such as BDNF/TrkB pathway, and consequently resulted in poor functional or neuropathological outcomes (Zhao et al., 2015).

Here in this study, we demonstrated that treadmill exercise for 11 weeks starting 3 weeks after TBI also improved spatial learning ability by suppressing apoptotic neuronal cell death in the hippocampus. These effects of treadmill exercise appeared through suppressing TBI-induced activation of CREB/BDNF/TrkB signaling pathway. Based on the present results, late starting treadmill exercise is also a potential useful therapeutic strategy for the treatment of TBI.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

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