Association of Monocyte Chemoattractant Protein-1 (MCP-1) 2518A/G Polymorphism with Proliferative Diabetic Retinopathy in Korean Type 2 Diabetes

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Purpose: Monocyte chemoattractant protein-1 (MCP-1) is a chemokine that can increase adhesion molecule expression on monocytes and produce superoxide anions. Hyperglycemia induces MCP-1 production in vascular endothelial cells and retinal pigmented epithelial cells, and has been implicated as a causal factor in the facilitation of vascular complications in diabetes. In the present study, we evaluated the association of a single nucleotide polymorphism (SNP) in the MCP-1 gene with proliferative diabetic retinopathy (PDR) in a Korean population with type 2 diabetes.

Materials and Methods: We conducted a case-control study, which enrolled 590 subjects with type 2 diabetes, and SNP genotyping of c.2518A/G in the MCP-1 gene was performed using polymerase chain reaction followed by digestion with PvuII restriction enzyme.

Results: The prevalence of c.2518A/G polymorphism in diabetic patients was 13.2% (A/A), 47.1% (A/G) and 39.7% (G/G). In patients with diabetic retinopathy, the prevalence of PDR was significantly higher (\(p=0.009\)) in diabetic subjects with the c.2518A/A genotype (35.9%; n=78) compared to those with either the A/G or G/G genotype (22.3%, n=512). The prevalence of any other micro and macro-complications, including nephropathy and cerebrovascular events, were not different according to the c.2518A/G genotype. Conclusion: Our new genetic findings suggest that the c.2518A/A genotype in MCP-1 could be used as a susceptibility gene to predispose Koreans exhibiting type 2 diabetes for the development of PDR.

Key Words: MCP-1 polymorphism, type 2 DM, proliferative diabetic retinopathy

INTRODUCTION

Proliferative diabetic retinopathy (PDR) is a serious diabetic microvascular complication as a consequence of active angiogenesis in the retina.¹ It is a leading cause of visual loss with a substantial impact on the quality of life of diabetic patients. It has been observed that neovascularization plays a pivotal role in the development of PDR. This process involves the migration and proliferation of endothelial cells as well as the remodeling of the extracellular matrix.²⁻⁴ The pathogenic mechanism of PDR remains to be fully elucidated, but several cytokines and chemokines, includ-
The MCP-1 c.2518A/G polymorphism in the first 5' flanking region at position -2518 influences MCP-1 production and expression in response to an inflammatory stimulus. \(^{14}\) This genetic variability correlates with differences in monocyte MCP-1 production and may be responsible for clinical differences in disease severity.

Because of the importance of MCP-1 in the biology of PDR and the genetic single nucleotide polymorphism (SNP) evidence for role in the regulation of MCP-1 production, we postulated that there may be an association between MCP-1 c.2518A/G with the pathogenesis of PDR. This study was designed to evaluate a sub-population of Korean patients exhibiting type 2 diabetes with or without PDR.

**MATERIALS AND METHODS**

**Subjects**
A case-control study was designed in which 590 type 2 diabetic patients were enrolled with a defined ophthalmologic status. This study was approved by the Institutional Review Board of Chungbuk National University Hospital, and all patients gave informed consent prior to being included in the study. Inclusion criteria were age at diagnosis of diabetes ≥30 years and a known duration of diabetes of ≥5 years. Diabetes was diagnosed according to WHO criteria. \(^{15}\) All patients underwent biochemical tests and medical history. Diabetic retinopathy was assessed through dilated pupils by trained ophthalmologists. The patients were classified using the criteria based on the International Clinical Retinopathy Severity Scale: 1) no apparent diabetic retinopathy; 2) non-proliferative diabetic retinopathy (NPDR); or 3) PDR. Diabetic nephropathy was defined by the presence of microalbuminuria or overt albuminuria. Macrovascular complications, such as coronary artery disease or stroke, were assessed according to the available medical records.

**Determination of MCP-1 c.2518A/G genotype**
Peripheral leukocytes were isolated from EDTA-treated whole blood obtained from each patient, and genomic DNA was extracted for polymerase chain reaction (PCR) amplification of MCP-1. The following primers were used for amplification: forward 5'-CCGAGATGTTCCCCAGCAG' and reverse 5'-CTGCTTTGCTTGCTCTTT-3'. The DNA amplification was performed by cycling at 94°C for 1 minute, 55°C for 1 minute, and 72°C for 1 minute. After 40 cycles, the reaction was extended for an additional 10 minutes at 72°C. The amplified PCR product was subsequently restricted digest overnight at 37°C with PvuII (5U).\(^{14}\) Agarose gel electrophoresis of the digested products were analyzed to determine the presence of the various genotypes: 1) A/A genotype yields only a single 930 bp band; 2) G/G genotype results in two bands (222 bp and 708 bp); and 3) A/G genotype results in three bands (222 bp, 708 bp and 930 bp).

**Statistical analysis**
All statistical tests were performed with SPSS version 12.0 (SPSS Inc., Chicago, IL, USA). Genotype frequencies were calculated from the observed numbers. Hardy-Weinberg equilibrium was tested with the \(\chi^2\) test. The data was expressed as mean±standard deviation. Means and frequencies of variables were evaluated using Student’s t-test, and \(\chi^2\) test, respectively. All \(p\) values <0.05 were considered statistically significant.

**RESULTS**
In our subject population, 142 patients were diagnosed with proliferative diabetic retinopathy and the remaining 448 patients were categorized as having NPDR. The demographics of the study population according to the MCP-1 c.2518A/G genotype are summarized in Table 1. The clinical characteristics did not differ among genotype subgroups.

Genetic variation in the SNP analysis of c.2518A/G in the MCP-1 gene was analyzed with the following distribution: A/A (13.2%), A/G (47.1%) and G/G (39.7%). The genotypic distribution of the MCP-1 c.2518A/G was in Hardy-Weinberg equilibrium. In terms of severity of diabetic retinopathy (Table 2), the prevalence of PDR was signifi-
The pathogenesis of PDR remains poorly understood. Recent studies have implicated a number of cytokines and chemo-

Table 1. Clinical Characteristics of Patients with Type 2 Diabetes According to the MCP-1 c.2518A/G SNP

| Clinical characteristics | MCP-1 c.2518A/G SNP | p value |
|--------------------------|----------------------|---------|
|                          | AA (n=78)            | AG+GG (n=512) |         |
| Sex (F/M)                | 41:37                | 253:259    | 0.604   |
| Age (yrs)                | 59.73±9.71           | 59.67±10.96| 0.968   |
| BMI (kg/m²)              | 25.50±3.42           | 24.93±3.39 | 0.179   |
| Duration of DM (yrs)     | 12.89±7.21           | 11.84±7.54 | 0.246   |
| FBS (mg/dL)              | 143.92±56.02         | 133.33±39.77| 0.068   |
| PP2 (mg/dL)              | 215.57±69.68         | 209.85±72.46| 0.599   |
| HbA1c (%)                | 7.68±1.41            | 7.47±1.30  | 0.187   |
| C-peptide (ng/mL)        | 2.76±3.02            | 2.61±1.92  | 0.576   |
| Insulin (µIU/mL)         | 15.05±23.54          | 12.35±6.97 | 0.090   |
| Cholesterol (mg/dL)      | 170.91±32.67         | 171.70±34.49| 0.850   |
| Triglyceride (mg/dL)     | 171.74±114.89        | 158.03±92.29| 0.269   |
| HDL (mg/dL)              | 45.79±12.49          | 48.03±13.31| 0.193   |
| LDL (mg/dL)              | 98.39±28.27          | 101.23±29.47| 0.453   |
| BUN (mg/dL)              | 16.52±7.04           | 15.50±8.15 | 0.296   |
| Cr (mg/dL)               | 1.21±0.67            | 1.12±0.60  | 0.218   |
| SGOT (IU/L)              | 25.99±11.01          | 26.78±14.48| 0.644   |
| SGPT (IU/L)              | 30.46±18.13          | 30.25±23.29| 0.938   |

MCP-1, monocyte chemoattractant protein-1; SNP, single nucleotide polymorphism.

Table 2. Prevalence of Diabetic Proliferative Retinopathy According to the MCP-1 c.2518A/G SNP

| Genotype | p value |
|----------|---------|
| AA       | 28 (35.9%) |
| AG+GG    | 114 (22.3%)  |
| 0.009    |          |

PDR, proliferative diabetic retinopathy; MCP-1, monocyte chemoattractant protein-1; SNP, single nucleotide polymorphism.

Table 3. Prevalence of Diabetic Nephropathy According to the MCP-1 c.2518A/G SNP

| Genotype | p value |
|----------|---------|
| AA       | 28 (35.9%) |
| AG+GG    | 163 (31.8%)  |
| 0.475    |          |

DN, diabetic nephropathy; MCP-1, monocyte chemoattractant protein-1; SNP, single nucleotide polymorphism.

Table 4. Prevalence of Diabetic Macrovascular Complications According to the MCP-1 c.2518A/G SNP

| Genotype | p value |
|----------|---------|
| AA       | 11 (14.1%) |
| AG+GG    | 67 (85.9%)  |
| 0.918    |          |

CAD, coronary artery disease; MCP-1, monocyte chemoattractant protein-1; SNP, single nucleotide polymorphism.

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significantly higher (p=0.009) in subjects with the c.2518A/A genotype (35.9%; 28/78 patients) compared to those carrying either the A/G or G/G genotype [22.3% (114/512)].

The prevalence of diabetic nephropathy (Table 3) and other macrovascular complications including coronary heart disease and stroke (Table 4) were evaluated for their association with the MCP-1 c.2518A/G SNP, but no significant association was observed for this specific polymorphism.

DISCUSSION

The pathogenesis of PDR remains poorly understood. Recent studies have implicated a number of cytokines and chemo-
In the activation of a complex network of pathways to promote cellular migration and proliferation. In conjunction with these factors, there appears to be a critical role for genetics in the regulation of PDR, since only a subgroup of diabetic patients develop PDR, even between PDR and NPDR patients with similar glycemic control and duration of diabetes.

Among the genetic factors being considered as a susceptibility gene for PDR, MCP-1 is known to be associated with diabetic microvascular or macrovascular complications. MCP-1 is a chemokine that exerts several effects on monocytes and macrophages including induction of superoxide anion, cytokine production, and adhesion molecule expression. Moreover, intraocular MCP-1 levels have been shown to be significantly increased in diabetic retinopathy (DR) and associated with clinical stage of DR.

In the present study, the prevalence of PDR was significantly higher in diabetic patients with the c.2518A/A genotype compared to those with either the A/G or G/G genotype. However, the relationship between SNP polymorphism c.2518A/G in the MCP-1 gene with diabetic retinopathy remains controversial. Katakami, et al. reported that the G allele in the c.2518A/G polymorphism is a susceptibility allele for diabetic retinopathy in a Japanese population of diabetic patients. In this study, however, the mean duration of diabetes was for only a length of 9 years, which was much shorter than in other published studies. In another Japanese study, Yoshioka, et al. showed that there was no association between c.2518A/G polymorphism of the MCP-1 gene with diabetic retinopathy in a group of Japanese patients with type 2 diabetes. This study did not match the duration of diabetes and glycemic control with the previous study by Katakami, et al. Towards this end, our study was designed to match both the duration of diabetes (>12 years) and glycemic control between the various genotypes, i.e., A/A and A/G+G/G. Using this approach, the number of PDR patients was significantly higher in the A/A genotype population compared to those in the other A/G+G/G genotype population. This data would suggest that the c.2518A/A genotype of the MCP-1 gene could be a potential risk factor for PDR in Korean type 2 diabetic patients. The difference in the nucleotide that was associated with PDR between the current study and the Katakami, et al. study, who included 112 hemodialyzed patients with kidney failure. This was not a selection criterion in our protocol and we did not have any patients in this category. The role of the c.2518A/G polymorphism may play an important diagnostic role in kidney disease detection. Kim, et al. reported that urinary MCP-1 concentration was much greater in A/A genotype patients compared to the other genotypes in Korean patients with lupus nephritis. These results would suggest that the A allele of MCP-1 c.2518A/G polymorphism may be associated with kidney diseases in Korean patients.

In cardiovascular disease (CVD) and MCP-1 polymorphism, Buraczynska, et al. showed that the G allele of the MCP-1 gene is associated with an increased risk of CVD in hemodialyzed renal failure patients. In our study, MCP-1 polymorphism was not associated with CVD. Different patient traits and ethnicity may explain the differences in the role of the SNP in the MCP-1 gene with various other phenotypes. Interestingly, the distribution of allele frequency in the MCP-1 gene was different between Caucasian and Asian patients. G allele carriers were more prevalent in Asian populations (G allele : A allele=60 : 40), while A allele carriers were more prevalent in Caucasian populations (G allele : A allele=40 : 60).

There are some limitations to our study that could impact our conclusions. First, this was a cross-sectional study, and as such, we did not measure plasma MCP-1 in this study. For more accurate results, prospective studies will be needed to investigate the role of MCP-1 polymorphism in PDR in type 2 diabetic patients, and a larger prospective genome-wide association study that includes family history will be required to fully clarify the pathogenesis of PDR in type 2 diabetic patients.

In conclusion, this study suggests that the c.2518A/G polymorphism in the MCP-1 gene can be used as a novel method to detect susceptibility to PDR manifestation in Koreans with type 2 diabetes.

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