Management strategies of enterovirus D68 outbreaks: current perspectives

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Abstract: Following its discovery in California in 1962, enterovirus D68 (EV-D68) was reported only sporadically around the world. In August 2014, a marked increase of EV-D68 cases in young children with severe respiratory infections was reported in the USA and Canada and later in Europe and Asia. Some of these cases were also found to be associated with acute flaccid paralysis, which exacerbated public health concern, and has since triggered international efforts to strengthen both EV-D68 and acute flaccid paralysis surveillance systems. This review summarizes the current knowledge on EV-D68, offering an overview of EV-D68 epidemiology, clinical presentations, diagnostic methodologies, and treatment strategies, as well as surveillance and outbreak management.

Keywords: enterovirus D68, AFP, diagnostics, treatment, surveillance, outbreak

Introduction

Human enteroviruses (EVs) are responsible for a wide variety of human infections, ranging from mild, nonspecific illness to severe diseases, such as hand, foot, and mouth disease, respiratory infections, herpangina, and encephalitis, among others. Enterovirus D68 (EV-D68), an acid-labile EV belonging to EV group D, is mainly associated with respiratory disease and was identified for the first time in hospitalized children presenting with pneumonia and bronchiolitis in California in 1962.1 In the following years, this virus was sporadically reported until 2008, when cases of EV-D68 were detected in the Philippines, Japan, the Netherlands, and the USA.2 The majority of the detections were among young children under 4 years of age, presenting with respiratory complications and requiring hospitalization.3,4 Starting from August 2014, an increase in the number of EV-D68 cases was initially reported in the USA, followed by Canada and Europe. This marked increase in EV-D68 cases, many of which were associated with neurological complications, has since triggered international public health concern and set in motion recommendations for worldwide increased surveillance for EV-D68.2,5

This review summarizes the current knowledge on EV-D68, including an overview of EV-D68 surveillance and outbreak management strategies. With this purpose, a PubMed database search, limited to publications from the last 10 years, was performed using EV-D68, EV-D68 AND drug therapy, EV-D68 AND drug effects, EV-D68 AND pathogenicity, and EV-D68 AND vaccine as keywords. This search resulted in 238 records, for which the abstracts were screened based on a set of inclusion and exclusion criteria. These included EV-D68 epidemiology, clinical presentation, surveillance, and outbreak management and excluded case reports, modeling, and environmental
studies. As a result of this screening, 78 publications were systematically reviewed, of which 59 were included for the current review.

Epidemiology

The increase in EV-D68 cases reported in young children in the summer of 2014 triggered an alarm in the international public health community. It was soon after revealed that an unprecedented high number of EV-D68 cases also presented with acute flaccid paralysis (AFP), namely 120 cases in the USA and another three in Europe, two of which were in Norway and one in France.6–8 In view of these detections and the increasing evidence of a potential association of EV-D68 with AFP, the European Centre for Disease Prevention and Control advised European Union (EU)/European Economic Area (EEA) countries to strengthen respiratory sample screening for EVs and EV typing and to strengthen AFP surveillance as well as EV surveillance, with a particular focus on cases with affected central nervous system function.5 These measures envisaged enhancing polio surveillance and allowed for the detection of unusual clusters of neurological disease or trends among nonpolio AFP cases. For this purpose, new diagnostic methods were developed specifically for detection of EV-D68, and several retrospective studies ensued, showing an increase in EV-D68 cases in preceding years, many of which were considered at the time of detection as rhinovirus (RV) infections.9,10 Retrospective studies in the Netherlands showed an increased circulation of the virus in the country from 2010 to 2014.3,11

In the following year, 2015, few or no EV-D68 cases were reported, after which an upsurge of cases re-occurred in 2016, both in Europe and in the USA.12,13 During the summer of 2016, an increase in cases was reported in the Netherlands, the majority of which affected children younger than 5 years old.14 In Sweden, 74 EV-D68 confirmed cases, among which there was one fatal case and another 10 presenting with severe respiratory or neurological symptoms, were reported during August–September 2016. The majority of cases were, once again, young children under 5 years of age.15 Cases have been found in all ages according to surveillance reports, and the most affected age group has yet to be defined. An increase in EV-D68 was also observed in other European countries, such as France, Italy, the UK, and Portugal, with reports of some cases being associated with AFP.16 A recent review has shown a causal relationship between EV-D68 and acute flaccid myelitis (AFM), supported by the application of Bradford Hill criteria.17

EV-D68 classification and pathogenesis

EV-D68 is a small, nonenveloped virus with a single stranded, positive-sense RNA genome of about 7.5 kilobases, which includes a single open reading frame which encodes four structural proteins (VP1 to VP4) and seven nonstructural proteins (2A to 2C and 3A to 3D), a 5′ untranslated region (UTR) with a hairpin-loop secondary structure, and a 3′UTR with a poly(A) tract18 (Figure 1). Based on the VP1 nucleotide sequence, EV-D68 strains have been classified into three distinct genetic clades; A, B, and C.18 These are further divided into subclades A1 and A2, B1 and B2, and, since the 2014 outbreaks, two more subclades were described, B3 and D.14,15,19–21 In 2014, the predominant circulating EV-D68 subclade was B1,6,20 and in 2016, the new subclade B3.14,15 EV-D68 belongs to the family Picornaviridae and genus EV, which comprises 12 species designated EV A to H and RV A to C.22 EV-D68 belongs to the EV D species, along with EV-D70, associated with acute hemorrhagic conjunctivitis; EV-D94, causative agent of AFP; EV-D111 and EV-D120, identified in nonhuman primates.18 EV-D68 is biologically similar to RV, in that it is mainly associated with respiratory disease, and was previously named RV87. However, it was not until recently that severe cases of respiratory disease due to EV-D68 were reported.3,9,10,23

In general, EV infect via the fecal–oral route and are normally transmitted by inhalation of aerosolized virus-containing material. However, manual transmission to the airway from environmental surfaces also occurs. After an incubation period of 1–2 weeks, symptoms may develop. During the first week of illness, the virus can generally be found in the nasopharynx, where it attaches to sialic acid
(SA) receptors in the epithelial cells of the upper respiratory tract (URT) mucosa. EV-D68 spreads locally and can enter the bloodstream to gain access to organs such as meninges and other parts of the central nervous system. EV-D68 has been shown to have a higher affinity for α-2-6-linked SA than α-2-3-linked SA receptors as is the case of other respiratory viruses. The ability of the virus to bind to α-2-3-linked SA receptors is an observation that supports that the virus can cause infections in the lower respiratory tract (LRT). Animal models have demonstrated that the virus can spread from URT to LRT after nasal infection and cause symptoms in the lungs.24

Clinical presentation
EV-D68 was first detected in cases presenting with respiratory symptoms like rhinorrhea, sneezing, coughing, or sore throat combined with fever and malaise. EV-D68 can cause both URT and LRT infection and has been associated with bronchitis, bronchiolitis, wheezing illness, and exacerbation of asthma in children.18,25-27 Fatal respiratory infections with EV-D68 have been reported both in children and in adults, particularly in patients with comorbidities or immunosuppression.18,28 In addition, EV-D68 has been found in cases suffering from various neurological diseases. A fatal outcome has also been described in a meningomyeloencephalitis patient with EV-D68 positive cerebrospinal fluid (CSF).29

Outbreak investigations of children with polio-like disease presenting with AFP or AFM have shown that EV-D68 can be found in respiratory specimens from a considerable number of cases, particularly those with severe infection.6,10-15 Many cases commonly show prodromal symptoms, such as fever from URT infections, and usually improve before development of paralysis, which, if it occurs, is generally observed within a week’s time. Other symptoms, such as headache, stiff neck, and pain in the back and affected limbs, are commonly observed as URT symptoms decrease. A common feature among these cases is that the virus is not detected in CSF samples or stool specimens at onset of paralysis, possibly due to late or insufficient sampling. A recent review suggests that EV-D68 may act as a trigger to AFM and describes an average time period of around 4 days until maximum muscle impairment is reached, involving decreased or absent reflexes.31 In that study, an asymmetric distribution of paralysis was observed, with mostly upper limbs affected, and general motor function was still impaired a year later upon follow-up consultations. Other clinical manifestations included bowel or bladder dysfunction. Asthma was one of the most common comorbidities among the cases.31

EV-D68 diagnostics
EV-D68 sample collection should be performed according to clinical manifestations and preferably within the first days following symptom onset. Specimens include CSF, stool, respiratory specimens, eg, nasopharyngeal aspirates, nasopharyngeal swabs, bronchoalveolar lavage, and blood, in severe cases.36 Serum collection for serological testing of EV-D68 is not a common practice in routine diagnostics. The need for sampling is associated with severity of the clinical presentation, and collecting multiple samples from different sites is important for increasing the chances of successful detection of EV-D68. Stool sample collection from patients with AFP is fundamental to exclude poliovirus. Respiratory specimens should also be included for analysis, as this virus is rarely detected in CSF and stool, and a negative result in these types of specimens does not rule out EV-D68 infection. Blood samples should also be considered, as EV-D68 viremia may, in some cases, be higher in blood than in the CSF.

Diagnostic testing for EV relies on molecular methods such as PCR targeting the 5’ UTR of the genome. This region presents high similarity between EV and RV and as such may in some cases result in false negative or false RV positives. Furthermore, due to high genetic similarity among the different EV types, this methodology does not allow for a reliable differentiation between EV-D68, RV, and other EV, such as coxsackie virus and echovirus, and as such should not be used as the preferred method for EV type discrimination. The general strategy for EV-D68 confirmation has been to perform pan-EV PCR on the UTR and/or the VP1 or VP2 followed by sequencing of the PCR products.37,38 However, this procedure is time-consuming, costly, and unfeasible for many diagnostic laboratories.

Food and Drug Administration-cleared molecular assays are currently available for detection of EV and RV in clinical samples through respiratory panel multiplex assays. An EV-D68-specific RT-PCR method was quickly developed by the CDC following the outbreak in 2014 and was approved by the Food and Drug Administration under emergency use authorization only.39-41 Other RT-PCR methods, with different degrees of specificity and sensitivity, have also been developed42-44 as well as a duplex-assay for simulations screening and differentiation between non-D68 EV positive and D68 positive virus.45

Complete viral genomes of EV-D68 have been generated through high-throughput sequencing of overlapping amplicons from primary specimens, allowing for identification of new clades as well as by other whole-genome sequencing strategies.19,46-49 In addition, a “Fernon” reference strain has been
provided by the French National Enterovirus Reference Laboratory to allow laboratories to test the accuracy of EV and EV/RV assays. EV-D68 viruses have also been included in Quality Control for Molecular Diagnostics External Quality Assessment panels.

An evaluation of the EV diagnostic analysis performed in Europe in 2014 has shown that many commercial available diagnostic assays do not discriminate adequately between RV and EV and/or do not discriminate EV-D68 from other EV. In both the Europe and the USA, few clinical diagnostic laboratories employ systematic EV detection followed by genotyping in the routine screening strategy of respiratory materials. As EV-D68 is known to cross-react with RV and EV targets, it is recommended to do EV-D68 confirmation by sequencing or to perform at least two assays with different target regions for detection. The VP1 region has been the preferred region for EV-D68 genotyping.

**Antiviral treatment and vaccines**

Currently, there is no specific antiviral therapy or vaccine available for EV-D68 infections. Mild disease cases might benefit from supportive therapies but are otherwise self-limiting. In the case of severe LRT disease, supplemental oxygen, bronchodilators, systemic corticosteroids, and ventilator support are the main interventions used. Treatment for AFM is empiric and often includes plasmapheresis and high-dose corticosteroids, aimed at reducing immune responses and inflammation or human intravenous immunoglobulin (hIVG) with potential immunomodulatory or antiviral effects. The rare and sporadic occurrence of the disease makes randomized control trials for investigating treatment effects difficult. Lack of suitable animal models has further hampered a more comprehensive understanding of the EV-D68 pathogenesis, therapy, and vaccine development. Recent studies in experimental mouse models indicate that immunomodulatory strategies, such as use of human intravenous immunoglobulin containing high titers of anti-EV-D68 or vaccination, might be efficient for prevention and treatment of EV-D68-mediated neurological disease, while corticosteroids should be used with caution. Moreover, studies of three potential therapies against EV-D68 in experimental mouse models suggest that the primary target of treatment should be the virus itself, rather than reducing the immune response.

Several direct-acting EV inhibitors have been developed, including capsid binders and inhibitors of viral enzymes required for genome replication such as protease inhibitors, 3Dpol inhibitors, and 2C ATPase inhibitors. So far, none have been approved for clinical use. The most widely studied compounds are capsid binders, of which pleconaril, vapendavir, and pocavir are undergoing clinical trials. However, a disadvantage of capsid binders is the emergence of resistant EV-D68 variants, which complicates their clinical use. Protease inhibitors may be more promising as direct-acting antiviral drugs, particularly since these often have broad-spectrum anti-enteroviral effects. Rupintrivir is the most studied among the protease inhibitors tested against EV and has shown broad-acting in vitro effects. However, clinical development was halted due to poor oral availability and lack of effect in naturally infected patients in clinical studies. Nevertheless, several rupintrivir derivatives are under development as well as preclinical studies of nonpeptidomimetic small molecule inhibitors with a better bioavailability than rupintrivir. 3Dpol inhibitors such as ribavirin have shown promising results for some viruses; however, few have yet been developed against EV. While several 2CATPase inhibitors have been identified, their effect against EV remains to be tested in clinical trials. Drug repurposing studies of 2CATPase inhibitors identified fluoxetine (ie, Prozac®) as a potential drug for EV, and it was shown to reduce EV-D68 growth in vitro. However, adding to issues related to dosage and side effects, a recent study found no effect on EV-D68 viral loads or symptoms in an experimental mouse model. Finally, inhibitors of host factors involved in EV infection might be used in treatment. These may have broad-spectrum antiviral activity but are also associated with drawbacks such as adverse effects and toxicity in the patient.

The challenges of developing specific antiviral therapies against EV-D68 emphasize the need for vaccine development. Recent establishment of a suitable neonatal mouse model for EV-D68 disease, and identification of the cotton rat as a potential powerful animal model for natural EV-D68 infection, hopefully might accelerate future vaccine development.

**EV-D68 surveillance and outbreak management**

As a result of the EV-D68 outbreaks in 2014, many countries implemented measures to strengthen their national EV surveillance systems, such as enhanced screening of severe respiratory infections, according to well-defined case definitions, specific EV-D68 diagnostic testing, dissemination of information among healthcare providers and the general public, and enhanced screening of AFP cases. Ongoing surveillance is key for timely detection and effective response to outbreaks and to ensure supportive
care. This surveillance, added to the mounting evidence of the AFP and EV-D68 association, as well as the cyclical EV-D68 pattern of 2–4 years during summer–autumn seasons, makes clinicians and laboratory staff evermore prepared to recognize and respond to potential EV-D68 outbreaks. An effective response to an outbreak requires coordination and clear communication among responders in multidisciplinary teams, including clinicians, nurses, infection prevention control officers, laboratory staff, epidemiologists, and public health officials, among others. A systematic and flexible outbreak investigation is essential to ensure the ultimate welfare of patients, by attending to all steps in the outbreak response process. These steps span from the early recognition of EV-D68 infection symptoms by the clinician who triggers the alert or by laboratory personnel detecting an increase in EV-D68 positive results, followed by the subsequent steps of an outbreak investigation, i.e., developing hypotheses, searching and describing further cases, implementing control measures, which should be done as early into the investigation as possible, to communication of final results. Given that, in cases of severe EV-D68 infection, the timeframe between disease onset and development of critical neurological symptoms is extremely short, usually spanning only a couple of days, the initial recognition and treatment stages are vital. Continued surveillance is essential to monitor EV-D68 incidence, circulating molecular strains as well as spectrum of illness and potential changes in patterns of illness severity.

**Concluding remarks**

Since its discovery in 1962, EV-D68 has made sporadic appearances until a series of outbreaks in 2014 caught worldwide public health attention. As a result, many countries strengthened their EV surveillance systems, to include also specific EV-D68 screening, with a particular focus on AFP and AFM cases. The implementation of VP1 and VP2 sequencing strategies, an extended EV surveillance including molecular epidemiology of both circulating and outbreak strains and a comprehensive viral characterization, would be an invaluable public health addition for EV reference laboratories. The underlying mechanisms for the apparent association between AFP and EV-D68 are still unclear, and merit further investigation as the number of reports suggesting this association increases. In addition, further studies are needed addressing the underlying host and viral factors, associated with severity of disease, as well as development of vaccines, in order to circumvent the current therapeutic challenges.

**Disclosure**

The authors report no conflicts of interest in this work.

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