Adaptive evolution and demographic history contribute to the divergent population genetic structure of *Potato virus Y* between China and Japan

Fangluan Gao | Wenchao Zou | Lianhui Xie | Jiasui Zhan

Fujian Key Laboratory of Plant Virology, Institute of Plant Virology, Fujian Agriculture and Forestry University, Fuzhou, China

Correspondence
Jiasui Zhan, Fujian Key Laboratory of Plant Virology, Institute of Plant Virology, Fujian Agriculture and Forestry University, Fuzhou, China.
Email: Jiasui.zhan@fafu.edu.cn

Funding information
China Agriculture Research System, Grant/Award Number: CARS-10-P11.

Abstract

*Potato virus Y* (PVY) is an important plant pathogen causing considerable economic loss to potato production. Knowledge of the population genetic structure and evolutionary biology of the pathogen, particularly at a transnational scale, is limited but vital in developing sustainable management schemes. In this study, the population genetic structure and molecular evolution of PVY were studied using 127 first protein (P1) and 137 coat protein (CP) sequences generated from isolates collected from potato in China and Japan. High genetic differentiation was found between the populations from the two countries, with higher nucleotide diversity in Japan than China in both genes and a *K*\textsubscript{ST} value of .216 in the concatenated sequences of the two genes. Sequences from the two countries clustered together according to their geographic origin. Further analyses showed that spatial genetic structure in the PVY populations was likely caused by demographic dynamics of the pathogen and natural selection generated by habitat heterogeneity. Purifying selection was detected at the majority of polymorphic sites although some clade-specific codons were under positive selection. In past decades, PVY has undergone a population expansion in China, whereas in Japan, the population size of the pathogen has remained relatively constant.

**KEYWORDS**

Bayesian skyline plots, demographic history, natural selection, phylogenetic analysis, *Potato virus Y*

1 | INTRODUCTION

Genetic drift, gene flow, and natural selection are three main evolutionary forces shaping the spatial population genetic structure of species (Zhan & McDonald, 2004). Under constrained gene flow, stochastic changes in allele frequencies among geographic populations can result in random fixation of neutral alleles, leading to nonadaptive differentiation (Wright, 1938). On the other hand, divergent selection for various ecological or physiological characters among genetically isolated populations may lead to adaptive population subdivision (Koskella & Vos, 2015; Yang et al., 2016). In plant pathology, understanding how populations are spatially structured is important to project the evolutionary trajectories of pathogens and formulate approaches for sustainable plant disease management (Zhan, Thrall, & Burdon, 2014). For example, many soil-borne plant pathogens are highly spatially structured (Gilbert, 2002) and regional R gene deployment may be an appropriate method for an effective and durable control of plant disease caused by the pathogens. On the other hand, recombination can facilitate the reshufflings of genomes and R gene pyramids may be less efficient in managing sexual pathogens usually
characterized by higher genetic variation distributed at a fine scale (McDonald & Linde, 2002).

Plant pathogens can vary greatly in spatial population genetic structure (Barrett, Thrall, Burdon, & Linde, 2008; Zhan, Thrall, Papaix, Xie, & Burdon, 2015), depending on the biotic and abiotic factors they associated with such host genetics, pathogen biology, physical environments, and the ways of human intervention during and postagricultural production. These factors can affect individually and interactively on the extent of genetic drift, gene flow, and selection, therefore influencing the generation and maintenance of spatial population structure (Bergholz, Noar, & Buckley, 2011; Burdon & Thrall, 2008; Zhan & McDonald, 2013). Natural selection is expected to play a central role in the spatial population genetic structure of plant pathogens in agricultural ecosystems (Thrall et al., 2011), primarily due to variation in host genetics, fungicide applications, climatic conditions, and agricultural practices among regions. Directional selection for virulence, fungicide resistance, and other ecological characters related to particular biogeographic environments can drive the rapid accumulation of adaptive genetic differentiation in plant pathogen populations (Achtman & Wagner, 2008). At the same time though, variation in these same factors among regions can also strongly influence the demographic dynamics of plant pathogens, generating nonadaptive genetic differentiation in the pathogen populations. However, despite recognition of its importance, an in-depth understanding of how patterns of spatial population genetic structure are generated and maintained and the main evolutionary mechanisms responsible for these patterns are still limited for many plant pathogens, but critical for sustainable plant disease management (He, Zhan, Cheng, & Xie, 2016; Zhan et al., 2015).

Potato virus Y (PVY) is a member of the genus Potyvirus in the family Potyviridae. Its genome has a single-stranded positive-sense RNA of ~9.7 kb, encoding a polyprotein that is cleaved into 10 mature functional proteins (King, Lefkowitz, Adams, & Carstens, 2011). Additionally, a short polypeptide (PIPO) is expressed within the P3 cistron by frame shifting (Chung, Miller, Atkins, & Firth, 2008). Among the 11 functional proteins encoded, the first protein (P1) and the coat protein (CP) are thought to play an important role in the adaptation of potyviruses to host species (Valli, López-Moya, & García, 2007). Furthermore, the CP gene has been frequently used in strain identification, species classification, and phylogenetic analysis of potyviruses (Cuevas, Delaunay, Rupar, Jacquot, & Elena, 2012).

PVY is one of the most destructive pathogens affecting potato (Solanum tuberosum L.), the third largest food crop in the world (Birch et al., 2012) and widely grown in many Asian countries including China and Japan. It can cause 40%–70% yield reduction in potato production (Nolte, Whitworth, Thornton, & McIntosh, 2004) and significantly reduce the quality of seed tubers. As the largest producer in the world, China accounts for 26.3% and 22.2% of the global potato acreages and yields, respectively (Wang et al., 2011), and potato production in the country is expected to substantially increase in coming decades due to government support and dietary shifts (Kearney, 2010). PVY is one of the main factors constraining further development of Chinese potato industry (Wang et al., 2011).

Knowledge of the population genetics and evolutionary biology of PVY, particularly at a transnational scale, is relatively limited compared to other important plant pathogens such as Magnaporthe (Tredway, Stevenson, & Burpee, 2005), Pyrenophora (Gurung, Short, & Adhikari, 2013), Verticillium (Short et al., 2015), and Phytophthora (Tian et al., 2016). Many factors such as technology and resource availability may partially contribute to this shortage. In addition, importation of living pathogens even for scientific reasons is strictly forbidden in many countries including China due to quarantine regulations. As a consequence, analysis of biotrophic pathogens with no or few morphological characters used to be very challenging for many researchers. With the advance of PCR-based sequencing technology and sharing of sequence data in many public domains, empirical analysis of the population genetic structure and evolutionary biology of biotrophic plant pathogens such as PVY at an international scale has become possible. This approach has been used by several laboratories to infer the evolution of PVY (Cuevas et al., 2012; Ogawa, Tomitaka, Nakagawa, & Ohshima, 2008), usually involving bona fide geographical populations. Ogawa and colleagues (Ogawa, Nakagawa, Hataya, & Ohshima, 2012; Ogawa et al., 2008) compared the population structure of PVY in Japan, Europe, and North America and found some unique evolutionary patterns associated with the pathogen in Japan. For example, there were only three nonrecombinant subpopulations in Japan (PVYO, PVYN, and PVYNNTN), whereas six subpopulations existed worldwide. Furthermore, unlike results from other regions where the subpopulations all have the same age, it is believed that the Japanese PVYO subpopulation is older than the PVYN and PVYNNTN subpopulations.

The objectives of this study were to (i) compare the population genetic structure of PVY in China and Japan and (ii) determine the main evolutionary and demographic mechanisms responsible for the observed population genetic structure.

2 | MATERIALS AND METHODS

2.1 | Viral sequences

Eighty-five PVY isolates, confirmed by enzyme-linked immunosorbent assay (ELISA) with a broad-spectrum PVY antibody (Agdia, Elkhart, USA), were randomly collected from potato (Solanum tuberosum) across a range of geographical locations in China including Fujian, Hunan, and Hebei provinces in 2011 and 2012 (Figure 1). Total RNAs were extracted using an RNAsimple Kit according to the manufacturer’s instructions (TianGen, Beijing, China). Full-length cDNAs were synthesized by RT-PCR using Oligo(dT)18 and TransScript® First-Strand cDNA Synthesis SuperMix (TransGen, Beijing, China) and amplified with two pairs of degenerate primers as described previously (Gao et al., 2014). PCR amplifications of cDNAs were performed in a total volume of 50 μl composed of 5.0 μl of TransTaq™ 10× HiFi Buffer II, 4.0 μl of dNTPs (2.5 mM), 2.0 μl of forward primer (10 μmol/l), 2.0 μl of reverse primer (10 μmol/l), 34.5 μl of ddH2O, 0.5 μl of TransTaq™ HiFi Polymerase (5 U/μl), and 2.0 μl of template cDNA. The PCR
The program was initially denatured at 94°C for 5 min; followed by 30 cycles of 94°C for 30 s, 53°C (P1 gene), or 55°C (CP gene) for 30 s and 72°C for 1 min; ended with an extension at 72°C for 10 min. PCR products were separated on 1% agarose gels by electrophoresis, visualized using a UV transilluminator and cleaned using a TIANgel Maxi Purification Kit (TianGen).

Purified PCR fragments were ligated into T-tailed pEASY-T5 Zero vector (TransGen) and transformed into competent *Escherichia coli* strain Trans1-T1 (TransGen). Recombinant plasmids were extracted and sequenced in both directions by Nanjing GenScript Biological Technology Co., Ltd. (GenScript, Nanjing, China). Due to high mutation rate in RNA viruses, at least four cDNA clones derived from two separate PCR reactions were sequenced for each PVY isolate to ensure consensus. Only the sequence identical at least three clones was used for further analyses to eliminate potential heterogeneity introduced by Taq polymerase. All sequences generated in this study were deposited in GenBank databases. In addition, 24 sequences of P1 gene and 26 sequences of CP gene from other parts of China (including Guizhou, Shandong, Heilongjiang, Liaoning, and Shaanxi provinces) and 34 sequences each of CP and P1 genes from Japan were retrieved from GenBank (Table S1, Figure 1). Consequently, the combined sequences from China were generated from PVY samples collected between 2005 and 2012 and the Japanese sequences were generated from samples collected between 1995 and 2012. As host-driven adaptation could affect the diversification of viral isolates, only PVY sequences derived from potato were retrieved and included in the analysis of population genetic spatial structure.

2.2 Genetic diversity and population genetic differentiation

Nucleotide sequences of the P1 and CP genes were aligned using the Muscle algorithm (Edgar, 2004) implemented in MEGAS5 (Tamura et al., 2011). A nucleotide identity matrix was generated using BioEdit software (Hall, 1999) after all gaps were removed. Haplotype diversity ($H_T$) and nucleotide diversity ($\pi$) were estimated using DnaSP 5.0 (Librado & Rozas, 2009). Pairwise $F_{ST}$, a measure of genetic differentiation among populations, was computed in Arlequin 3.5 (Excoffier & Lischer, 2010). Genetic differentiation among populations was also evaluated by $K_{ST}$ and $S_{nr}$ using DnaSP 5.10 (Hudson, 2000; Hudson, Boos, & Kaplan, 1992; Librado & Rozas, 2009). The hypothesis of deviation from null population differentiation was tested by 1000 permutations of the original data.
2.3 | Recombination and phylogeographic analyses

Putative recombination joints (RJ) and parental sequences were identified using seven methods (RDP, GENECOV, BOOTSCAN, MaxChi, CHIMAERA, SISCAN, and 3SEQ) implemented in the RDP4 suites (Martin et al., 2010). The probability of a putative recombination event was corrected by a Bonferroni procedure with a cutoff of \( p < .01 \). To avoid false identification, only events supported by at least four of the seven methods were considered to be recombinants. Recombinants were removed from the subsequent reconstruction of phylogenetic trees.

Phylogenetic trees were reconstructed by the Bayesian inference (BI) implemented in MrBayes 3.2.5 (Ronquist et al., 2012) and maximum likelihood (ML) implemented in MEGA5 (Tamura et al., 2011) using the GTR + G + I nucleotide substitution model determined by MrModeltest (Nylander, 2008). BI was run in 2,000,000 generations of Markov chains that were sampled every 100 generations to establish convergence of all parameters. The effective sample size (ESS) of parameters was checked by Tracer 1.6 to ensure values above 200 as advised by the programmers with the first 25% of sampled trees burn-in. Topology robustness of ML trees was assessed by 1,000 bootstraps. ML-BPs and BI-PPs were plotted on Bayesian 50% majority-rule consensus trees using FigTree 1.4.2 and Illustrator CS5 (Adobe).

The effect of geographic origin on PVY populations was evaluated by phylogeny–trait association analysis, and BaTS 2.0 (Parker, Rambaut, & Pybus, 2008) was used to compute an association index (AI), parsimony score (PS), and maximum monophyletic clade (MC). Low AI and PS scores and high MC scores suggest a strong PVY–geographic association. The topology of reconstructed trees was tested using BaTS by comparing it with the randomized trees generated from 10,000 reshufflings of tip characters. Topology robustness was determined in BaTS by comparing it with the null distribution of trees obtained from 10,000 bootstraps of tip characters.

2.4 | Natural selection

HyPhy 2.10b (Kosakovsky Pond, Frost, & Muse, 2005) and PAML 4.7 (Yang, 2007) were used to identify nucleotide sites in P1 and CP cistrons that were likely to be involved in PVY adaptation. Three codon-based approaches, that is, IFEL (internal branches fixed-effects likelihood), REL (random-effects likelihood), and MEME (mixed effects model of evolution) (Kosakovsky Pond et al., 2006, 2011; Murrell et al., 2012), were included in the HyPhy package. Only sites simultaneously identified by IFEL and MEME with \( p < .05 \) and \( > .95 \) posterior probability identified by REL were considered to be under selection. In addition, the ratio of nonsynonymous (dN) to synonymous (dS) substitution (\( \omega = dN/dS \)) was calculated for each gene using CODEML algorithm implemented in PAML 4.7 (Yang, 2007). Three different nested models (M3 vs. M0, M2a vs. M1a, and M8 vs. M7) were compared and likelihood-ratio tests (LRTs) were applied to select the one that best fitted the data. When the LRT was significant (\( \text{p-value} < .01 \)), the Bayes empirical Bayes (BEB) method (Yang, Wong, & Nielsen, 2005) was used to identify amino acid residues that are likely to have evolved under positive selection based on a posterior probability threshold of .95.

2.5 | Population historic dynamics

Two different approaches were used to explore the demographic history of PVY populations in the two countries. First, Tajima’s D and Fu’s \( F_{S} \) statistics implemented in Arlequin 3.5 (Excoffier & Lischer, 2010) were used to determine the neutrality of the P1 and CP genes by 1,000 permutations of the original sequences. Tajima’s D test identifies evolutionary events such as population expansion, bottlenecks, and selection by comparing the estimated number of segregating sites with the mean pairwise difference among sequences (Tajima, 1989). Fu’s \( F_{S} \) is sensitive to population demographic expansion and usually displays a negative value (Fu, 1997) in expanding populations. Following these calculations, Bayesian skyline plots (BSP) was generated to explore demographic history using BEAST 1.8.2 (Drummond, Suchard, Xie, & Rambaut, 2012). Sampling times of the sequences were used to calibrate the molecular clock. Date-randomization tests (DRTs) were performed in R 3.3.1 using the Tip Dating Beast package (Rieux & Khatchikian, 2016) to determine the temporal signal in data sets. A data set was considered to have an adequate spread in sampling time if its average rate did not fall within the 95% confidence intervals (CIs) generated from 20 replicates of randomized dates (Ramsden, Holmes, & Charleston, 2009; Duchêne, Duchêne, Holmes, & Ho, 2015). A Bayes factors test indicated that the relaxed uncorrelated exponential model was a better fit to the sequence data than the relaxed uncorrelated lognormal model and was chosen to estimate the molecular clock of P1 and CP genes. The MCMC was run for \( 5 \times 10^6 \) generations to ensure convergence of all parameters. Convergence and ESS (\( > 200 \)) of the parameters were checked using Tracer 1.6.

3 | RESULTS

3.1 | Sequence variation in P1 and CP genes

Seventy-three P1 and 78 CP genes were sequenced in this study and were deposited in GenBank (Table S1). Two P1 sequences had a nucleotide T insertion at position 33 (accession number: KF722821) and a nucleotide (A) insertion at position 814 (accession number: KF771018), respectively. Moreover, two P1 sequences (accession numbers: KX451346 and KX451344) contained premature terminators at position 208–210 and 619–621. A single CP sequence (accession number: KC296822) had an 11-nucleotide deletion at the beginning of the gene. These nonfunctional sequences were removed from further analyses of the population genetic structure. In addition, 34 complete P1 and CP sequences originated from Japan were retrieved from GenBank (Table S1). Consequently, a total of 127 complete P1 and 137 complete CP sequences were included in the population genetic analysis of the virus. The average nucleotide identities in the P1 and CP genes among the viral isolates from China were 89.9% (72.3%–100%) and 96.3%
TABLE 1  Sample sizes and genetic variation of the P1 and CP genes in the Potato virus Y populations sampled from China and Japan

| Gene | Country | Sample size | Haplotypes | Haplotype diversity | Nucleotide diversity |
|------|---------|-------------|------------|---------------------|---------------------|
| P1   | China   | 93          | 90         | .999                | .100                |
|      | Japan   | 34          | 27         | .980                | .109                |
|      | All     | 127         | 117        | .998                | .119                |
| CP   | China   | 103         | 99         | .999                | .036                |
|      | Japan   | 34          | 22         | .914                | .043                |
|      | All     | 137         | 121        | .994                | .053                |

TABLE 2  Statistical tests for population differentiation between P1 and CP genes in the Potato virus Y populations sampled from China and Japan

| Gene | $K_{ST}$ | $p$   | $S_{lm}$ | $p$   | $F_{ST}$ | $p$   |
|------|----------|-------|----------|-------|----------|-------|
| P1   | .141     | .000*** | .972     | .000*** | .294     | .000*** |
| CP   | .289     | .000*** | .949     | .000*** | .520     | .000*** |
| Concatenated | .216 | .000*** | .977 | .000*** | .390 | .000*** |

(.01 < $p$ < .05).

**.001 < $p$ < .01.

***$p$ < .001.

(87.0%–100%), while those from Japan were 89.1% (71.5%–100%) and 95.7% (88.7%–100%), respectively. All typical motifs of potyviruses were detected in the deduced amino acid sequences. However, the conserved N25 of CP protein in the reference sequences (O-139, N-605, C1-SON41, Adgen, and Chile3) was replaced by S (isolates NTNHIR3 and T13) or T (isolate ONGOB6) in the Japanese isolates and the E68 residue was replaced by G (isolates XQ03) or K (isolate Laiwu3) in Chinese isolates (Figure S1).

One hundred and seventeen haplotypes were identified in the 127 complete sequences of P1 gene with an overall haplotype diversity of .998 and nucleotide diversity of .119 when the sequences from China and Japan were considered together (Table 1). The most common haplotype was detected four times in P1 sequences and 10 times in CP sequences. No identical haplotypes were detected in the two countries in both genes. When the sequences were considered according to individual geographic location, higher nucleotide but lower haplotype diversity was found in Japan than in China. In the CP genes, 121 haplotypes were identified in the 137 complete sequences, with an overall haplotype diversity of .984 and nucleotide diversity of .053 when sequences from the two countries were pooled. Similar to the P1 gene, higher nucleotide diversity but lower haplotype diversity was found in the CP gene from Japan than from China when they were considered separately.

3.3  | Recombination analyses

Forty-two P1 sequences, all originating from China, were identified with a high level of confidence as recombinants by all seven methods implemented in the RDP package ($p$-values ≤1.27 × 10^{-2}, Table S2). All of these sequences were derived from three recombination events. Similarly, three recombination breakpoints were detected in eight CP sequences from China and four CP sequences from Japan by at least six of the seven approaches ($p$-values ≤1.69 × 10^{-2}, Table S2).

3.4  | Phylogenetic analysis

After removal of putative recombinants, 85 complete P1 sequences and 125 complete CP sequences were included in the phylogenetic analysis. Both P1 and CP sequences fell into two distinct clades (N clade and O clade) with high posterior probabilities (PPs ≥ .99) and maximum likelihood (ML) bootstrap (BPs ≥ 70%) supports (Figure 2). In the N clade of P1 tree, all 41 Chinese isolates were clustered into subclade 1 and all but three (NTNHIR3, NTNKGAM2, and NTNTK1) of the 27 Japanese sequences were grouped into subclade 2. Similarly, all 10 Chinese isolates were clustered into subclade 3 and all seven Japanese isolates were grouped into subclade 4. In the N clade of the CP tree, all nine Chinese sequences were clustered into subclade 1, whereas all 23 Japanese sequences were grouped into subclade 2. In contrast, O clade was composed of a mixture of Chinese and Japanese sequences. A similar pattern was found when only the P1 and CP sequences from GenBank (i.e., excluding the sequences generated in this study) were used to construct a phylogenetic tree (data not shown). Distinct differences in the population genetic structure of PVY between China and Japan were also found by phylogeny–geography association analysis. Significant MC ($p$ < .01), AI ($p$ < .001), and
PS (p < .001) were detected in both P1 and CP genes when sequences from the two countries were compared (Table 3). However, in general, a phylogeny–geography association was not found within countries (MC: p > .05; Table 3) with the exception of PVY isolates from Fujian and Heibei in China and Kyushu and Honshu in Japan.

3.5 | Detecting natural selection

Purifying selection was detected in the majority of polymorphic sites by PAML packages (Figure 3). Evidence of positive selection was detected in the 1st codon of CP sequences in the O clade using the PAML approach (PP > .99, Table 4). No positive selection was detected in the N clade. In P1 sequences, no positive selection was detected in either the N or O clade by PAML (Table 4). However, positive selection in the P1 gene was detected in the 3rd and 5th codons of the N clade and the 3rd, 78th, and 241st codons of the O clade by the REL approach implemented in the HyPhy package.

The REL approach also detected positive selection in the 1st, 9th, 11th, 15th, and 58th codons of CP sequences in the O clade and in the 1st codon of the N clade. Similarly, positive selection was found in the 1st codon both in the O and N clades of CP sequences by the MEME approach (p < .05; Table 4). Positive selection was also found in the 1st and 138th codons of CP sequences in the O clade by IFEL (<.05; Table 4). However, only the 1st codon in the N-terminal region of the CP protein, one of the cleavage sites in the PVY genome, was found to be under positive selection by both PAML and three approaches implemented in the HyPhy package. Further analysis showed that Glu (87.50%) dominated at the Nib/CP cleavage site in the N clade of the CP protein, whereas Ala (40.86%) and Glu (56.99%) were found to have a high frequency at the cleavage site in the O clade of the CP protein.

3.6 | Demographic history

Significantly negative Fu’s F3 was detected in both P1 and CP sequences from China (Table 5); Tajima’s D was positive for the P1 sequences and negative for the CP sequences from China, but none of them were significant. All Tajima’s D and Fu’s F3 in the sequences from Japan were positive but not significant (Table 5). All data included in the demographic analysis passed the DRTs that showed no overlaps between the original estimate of evolutionary rate and 95% CIs generated from 20 replicates of date randomization (Figure S2). Coalescence-based BSP revealed an explicit demographic history for the PVY populations of China and Japan (Figure 4)—showing that the PVY from China experienced a population expansion prior to a period of stability, whereas the population from Japan remained relatively constant throughout the past decades.

FIGURE 2  Bayesian phylogenetic trees of Potato virus Y (PVY) isolates based on P1 (a) and CP (b) coding regions. For three key nodes in a tree, the Bayesian posterior probabilities and maximum likelihood bootstrap percentage are indicated above the branches (Bayesian posterior/bootstrap). PVY isolates from this study are indicated by black dots. The distance unit is substitutions/site.

4 | DISCUSSION

Consistent with previous results (Ogawa et al., 2008, 2012; Tian et al., 2011), high genetic diversities (Table 1) were found in both P1 and CP genes in the current study. Each of these two genes encodes a protein with ecological functions that are important to the survival and adaptation of PVY in local biotic and abiotic environments (Quenouille, Vassilakos, & Moury, 2013). In addition to the high mutation rate in RNA viruses (Domingo, Sheldon, & Perales, 2012), high genetic variation found in both current and previous studies is possibly attributed to the high recombination rate in PVY populations (Gibbs & Ohshima, 2010). In recent decades, recombinant PVYs were found to be prevalent in the global population of the pathogen sampled from potato production areas (Quenouille et al., 2013). In this study, 52.69% and 11.76% of concatenated sequences in the Chinese and Japanese populations show a mixed genomic structure of PVYN and PVYD (Table S2). These results re-enforce widespread concern related to the propensity for rapid adaptation of PVY to changing biotic and abiotic environments. Such adaptation may cause problems for the deployment of major gene-mediated host resistance in potatoes, tomato, and other crops (Karasev & Gray, 2013).

Our study also reveals a significant difference in the molecular population genetic structure between the PVY sequences originating from potato hosts in China and Japan—a pattern that is supported by comparative analyses of genetic variation (Table 1), population differentiation (Table 2), and phylogeny–geography association (Table 3, Figure 2) in the two genes (each represented by ~130 sequences). It has been documented that measurements of the diversity of genetic variation are highly sensitive to sample sizes (Bashalkhanov, Pandey, & Rajora, 2009) and populations with larger sample sizes tend to have higher observed genetic diversity. Despite smaller sample sizes included in the Japanese than Chinese populations, both P1 and CP sequences originating from Japan show higher nucleotide diversity than those from China, suggesting that differences in nucleotide diversity between the two viral populations are unlikely to be the result of sampling error. However, the higher haplotype diversity in Chinese sequences may be associated with the somewhat larger sample size as compared to Japanese sequences (93 vs. 34 isolates). Indeed, when we used a bootstrap-strapping approach described previously (Zhan, Pettway, & McDonald, 2003) to standardize sample sizes with the smaller population (Japan in this case) and recalculated haplotype diversity, the difference in genetic variation between the two populations disappeared (Table S3). The bootstrap was conducted using the Resampling Stats add-in package for Excel (Blank, Seiter, & Bruce, 2001) with 100 replicates. For each bootstrap replication, haplotype diversity was recorded from a random sample of 34 sequences (the actual size of Japanese population). The mean and variance of the diversity were calculated and used for a t-test.
Table 3: Tests of geography–phylogeny association for P1 and CP genes in PVY isolates originated from China and Japan

| Gene | Country | Statistic | Isolates | Observed mean (95% HPD) | Null mean (95% HPD) | Significance |
|------|---------|-----------|----------|-------------------------|---------------------|--------------|
| P1   |         | AI        |          | 0.06 (0.02, 0.19)        | 4.61 (3.72, 5.35)   | <.001***     |
|      |         | PS        |          | 3.15 (3.00, 4.00)        | 26.79 (23.77, 29.41)| <.001***     |
|      | China   | MC (Guizhou) | 2        | 1.44 (1.00, 2.00)        | 1.01 (1.00, 1.05)  | .01**        |
|      |         | MC (Heilongjiang) | 2   | 2.00 (2.00, 2.00)        | 1.01 (1.00, 1.03)  | .02*         |
|      |         | MC (Henan)  | 1        | n/a                     | n/a                 | n/a          |
|      |         | MC (Hunan) | 20       | 6.00 (6.00, 6.00)        | 1.98 (1.46, 2.54)  | .02*         |
|      |         | MC (Fujian) | 17       | 5.80 (4.00, 6.00)        | 1.78 (1.33, 2.27)  | .01**        |
|      |         | MC (Hebei) | 7        | 3.91 (2.00, 4.00)        | 1.16 (1.00, 1.75)  | .01**        |
|      |         | MC (Shaanxi) | 2   | 1.00 (1.00, 1.00)        | 1.00 (1.00, 1.03)  | .01**        |
|      | Japan   | MC (Kyushu) | 34       | 23.42 (17.00, 24.00)     | 2.82 (2.22, 3.82)  | .01**        |
|      |         | MC (Okinawa) | 3       | 3.58 (2.00, 6.00)        | 1.88 (1.32, 2.84)  | .05*         |
|      |         | MC (Hokkaido) | 7    | 2.61 (1.00, 4.00)        | 1.18 (1.00, 2.00)  | .07**        |
|      |         | MC (Honshu) | 6        | 2.00 (2.00, 6.00)        | 1.09 (1.00, 1.30)  | .02*         |
|      |         | MC (Shanxi) | 24       | 2.48 (2.00, 4.00)        | 1.98 (1.55, 2.41)  | .43**        |
|      |         | MC (Fujian) | 45       | 7.23 (7.00, 9.00)        | 2.87 (2.21, 3.66)  | .01**        |
|      |         | MC (Hebei) | 12       | 2.19 (2.00, 3.00)        | 1.27 (1.01, 1.99)  | .05*         |
|      |         | MC (Hunan) | 3        | 1.40 (1.00, 2.00)        | 1.01 (1.00, 1.04)  | .10**        |
|      | Japan   | MC (Kyushu) | 17       | 3.11 (2.00, 5.00)        | 1.58 (1.14, 2.13)  | .02*         |
|      |         | MC (Okinawa) | 3    | 1.12 (1.00, 2.00)        | 1.01 (1.00, 1.04)  | .10**        |
|      |         | MC (Hokkaido) | 6   | 2.26 (2.00, 3.00)        | 1.05 (1.00, 1.14)  | .10**        |
|      |         | MC (Honshu) | 4        | 2.00 (2.00, 2.00)        | 1.02 (1.00, 1.12)  | .02*         |

AI, association index; PS, parsimony score; MC, maximum monophyletic clade; HPD, highest probability density interval; n/a: no data available because of insufficient sample size (n < 2).

Significance thresholds: *.01 < p < .05; **.001 < p < .01; ***p < .001.

Three indices were used to test population differentiation between Chinese and Japanese PVY sequences. \( K_{ST} \) and \( F_{ST} \) measure the relative proportion of total genetic diversity attributable to among-population differences. They range from .00 to 1.00. A value of one for \( K_{ST} \) or \( F_{ST} \) indicates that populations investigated are completely isolated, while a value of zero indicates the populations studied are identical (Hudson et al., 1992). \( K_{ST} \) or \( F_{ST} \) values between .15 and .25 indicate high population differentiation, and values greater than .25 indicate very high genetic differentiation among populations (Balloux & Lugon-Moulin, 2002). \( S_{mn} \) measures how often the nearest sequences are found in the same populations. Its value would be close to one when populations are highly structured and near .5 when populations are identical (Hudson, 2000). Therefore, \( K_{ST} \) and \( F_{ST} \) are approximately equal to \( S_{mn}^{-.5} \). All three indices are higher than .15 in the current study (Table 2), suggesting a high differentiation of PVY sequences between the two populations. \( K_{ST} \) is the most powerful index for detecting spatial population structure for recombination and high mutation populations (Hudson et al., 1992), which is the case in PVY (Blanchard, Rolland, Lacroix, Kerlan, & Jacquot, 2008).

Phylogenetic analyses provide additional support for differentiation between the Chinese and Japanese populations of PVY (Figure 2). Sequences tend to cluster according to their geographic origin, consistent with previous results derived from more and wider geographic locations (Cuevas et al., 2012). Although some clades such as the O clade of the CP tree are composed of sequences from both locations, the hypothesis of random association between PVY sequences...
from China and Japan and their geographic origins is rejected by all three statistics (AS, MC and PS) (Table 3). Phylogenetic trees are reconstructed under the assumption of constant evolutionary rates and many evolutionary processes may affect the correct reconstruction of phylogenetic topology (Revell, Harmon, & Collar, 2008). The polyphyletic nature of these different clades could be generated by some sequences experiencing a complex evolutionary history such as recombination. For example, in the P1 tree, three Japanese sequences (NTNHIR3, NTTNK1, and NTNKGM2) were clustered together with Chinese sequences in a subclade (Figure 2). It is likely they are recombinants that were not detected by the techniques used here. Our analyses suggest that the CP sequences of these three isolates are also most likely derived from recombination (Table S2).

Both stochastic and deterministic events could lead to the observed pattern of spatial structure (Caruso et al., 2011). Stochastic events caused by genetic drift may lead to nonadaptive population differentiation between the two countries. In nature, the extent of population differentiation generated by random genetic drift is expected to increase over geographic distance. Although geographic distance between some locations (ex. 2,771.10 km between Guizhou and Heilongjiang) within China is greater than that between China and Japan (ex. 819.41 km between Fujian in China and Okinawa in Japan), PVY sequences were clustered according to country origin (Figure 2, Table 3). Because our data (non-neutrality) are not appropriate for testing the stochastic event, the contribution of genetic drift could not be ruled out. However, we believe that deterministic events are the main factor responsible for the observed pattern of spatial population genetic structure in the pathogen.

The hypothesized contribution of deterministic events to the observed pattern of spatial population genetic structure is supported by neutrality tests in the deduced amino acid sequences and in the

![FIGURE 3](image-url)

Sliding window plot of dN/dS ratios for P1 (a) and CP (b) genes. Sites under neutral (dN/dS = 1) are indicated in red dotted line. The window size is 20 codons, and the offset between windows is one codon.

TABLE 4 Putative codons in the P1 and CP sequences detected under positive selection using the PAML package and three approaches implemented in the HyPhy Software

| Gene | Clade  | PAML Model M8 | HyPhy IFEL | REL | MEME |
|------|--------|---------------|------------|-----|------|
|      | Codon  | Codon p Value | Codon pp   | Codon p Value | Codon p Value |
| P1   | N Clade| 3 .999*       |            |     |      |
|      |        | 5 1.000**     |            |     |      |
|      | O Clade| 3 .970*       | 78 .984*   |     |      |
|      |        | 241 .972*     | 138 .981*  |     |      |
| CP   | N Clade| 1 .994**      | 1 .023*    | 1   | .029*|
|      |        | 1 .991**      | 9 .981*    | 1   | .007**|
|      | O Clade| 11 .959*      | 15 .977*   | 58  | .982*|

*aFor each PVY clade, model M8 performed better than model M7 in an LRT. Model M2a also performed better than models M0, M1a, or M3 in LRTs and provided results similar to M8.

*bPP, Posterior probability that individual codon positions belong to the positively selected category using the Bayes Empirical Bayes (BEB) method implemented in PAML and the random-effects likelihood (REL) approach implemented in the HyPhy, respectively.

*Posterior probability >95% or p < .05.

**Posterior probability >99% or .001 < p < .01.
sequences were included in the analysis (data not shown). Positive selection in cleavage sites has also been found in other viruses such as the human immunodeficiency virus (HIV, Banke, Lillemark, Gerstoft, Obel, & Jørgensen, 2009). Successful cleavages to form functional cis-elements are crucial for survival and reproduction of PVY (Tena Fernandez et al., 2013). This process is catalyzed by proteases that are under constant change by mutation (Yu, Benton, Bovee, Sessions, & Lloyd, 1995). Positive selection in the CP cleavage site could serve as a reversal mechanism compensatory to mutations in the protease in PVY and other viruses (Banke et al., 2009).

Differences in spatial structure between the Chinese and Japanese sequences may also result from differences in the demographic dynamics of the PVY populations. Sudden increases or decreases in population size associated with demographic events can affect the generation, maintenance, and distribution of genetic variation, not only directly through genetic drift and mutation, but also indirectly through impacts on the efficiency of natural selection to remove or amplify mutations as well as on migration and recombination (Wang & Whitlock, 2003). Indeed, when we performed demographic analyses, we found that PVY populations in China were small but had undergone recent expansion, possibly associated with increasing potato production in the current years (Wang et al., 2011), while in Japan, they were large but stably maintained (Figure 4). This result is consistent with the potato cultivation of the two countries in the past decades. Potato acreage in Japan maintained stable over the last several decades (Kawakami et al., 2015) while increased in China from ~2.3 million hectares in 1980s to ~5.4 million hectares recently (http://faostat.fao.org). The temporal scale of samples can impact the estimate of demographic dynamics (Drummond, Pybus, & Rambaut, 2003). In our study, the temporal scale (1995–2012) in the PVY sequences from Japan was 10 years longer than that (2005–2012) from China. However, we do not believe that this difference would have affected our conclusions because all data passed the DRTs (Ramsden, Holmes & Charleston, 2009; Duchêne et al., 2015) with a high level of confidence.

In summary, our study represents one of a few attempts to understand patterns and causes of spatial population genetic structure across political borders in PVY, a destructive pathogen of potato and many other Solanaceous crops. The finding of two subpopulations indicates distinct gene pools exist in PVY from China and Japan. This suggests that strict quarantine regulation is needed to prevent the movement of novel alleles or allelic combination of PVY between the two countries when trading plant materials that are hosts for this pathogen (e.g., potato and tobacco). However, sequences included in this analysis were relatively limited both in sample sizes and sites. In particular, the temporal scales might be relatively short for analysis of demographic events. Further study with larger, multiple-location, and longer temporal scale samples may be required to confirm the results and generalize the findings.

**ACKNOWLEDGEMENTS**

This work was supported by the China Agriculture Research System (Grant No. CARS-10-P11), P. R. China, granted to Jiasui Zhan. We
thank Drs. Jeremy J. Burdon and Peter Thrall in CSIRO, Australia, for proof-reading the manuscript and Dr. Zhenguo Du in FAFU and Dr. Lin Zhang in NNU for comments and suggestions to the manuscript.

DATA ARCHIVING STATEMENT

Sequence data obtained in this study are available on GenBank under the accession numbers KF722798–KF722799, KF722801, KF722804–KF722805, KF722807–KF722808, KF722810, KF722812–KF722813, KF722815–KF722818, KF722820–KF722825, KF722827–KF722830, KF722832–KF722834–KF722841, KF771008–KF771016, KF771018–KF771020, KX451323–KX451351, and KC296789–KC296790 (CP gene).

REFERENCES

Achtman, M., & Wagner, M. (2008). Microbial diversity and the genetic nature of microbial species. Nature Reviews Microbiology, 6, 431–440.

Balloux, F., & Lugon-Moulin, N. (2002). The estimation of population differentiation with microsatellite markers. Molecular Ecology, 11, 155–165.

Banke, S., Lillemark, M. R., Gerstoft, J., Obel, N., & Jørgensen, L. B. (2009). Positive selection pressure introduces secondary mutations at Gag cleavage sites in Human immunodeficiency virus Type 1 harboring major protease resistance mutations. Journal of Virology, 83, 8916–8924.

Barrett, L. G., Thrall, P. H., Burdon, J. J., & Linde, C. C. (2008). Life history determines genetic structure and evolutionary potential of host–parasite interactions. Trends in Ecology & Evolution, 23, 678–685.

Bashalkhanov, S., Pandey, M., & Rajora, O. P. (2009). A simple method for estimating genetic diversity in large populations from finite sample sizes. BMC Genetics, 10, 1–10.

Bergholz, P. W., Noar, J. D., & Buckley, D. H. (2011). Environmental patterns are imposed on the population structure of Escherichia coli after fecal deposition. Applied and Environmental Microbiology, 77, 211–219.

Birch, P. R. J., Bryan, G., Fenton, B., Gilroy, E. M., Hein, I., Jones, J. T., ... Toth, I. K. (2012). Crops that feed the world: Potato: are the trends of increased global production sustainable? Food Security, 4, 477–508.

Blanchard, A., Rolland, M., Lacroix, C., Kerlan, C., & Jacquot, E. (2008). Potato virus Y: A century of evolution. Current Topics in Virology, 7, 21–32.

Blank, S., Seiter, C., & Bruce, P. (2001). Resampling stats in excel version 4. Arlington, VA.

Burdon, J. J., & Thrall, P. H. (2008). Pathogen evolution across the agro-ecological interface: Implications for disease management. Evolutionary Applications, 1, 57–65.

Caruso, T., Chan, Y., Lacap, D. C., Lau, M. C., McKay, C. P., & Pointing, S. B. (2011). Stochastic and deterministic processes interact in the assembly of desert microbial communities on a global scale. The ISME Journal, 5, 1406–1413.

Chung, B. Y., Miller, W. A., Atkins, J. F., & Firth, A. E. (2008). An overlapping essential gene in the Potyviridae. Proceedings of the National Academy of Sciences of the United States of America, 105, 5897–5902.

Cuevas, J., Delaunay, A., Rupar, M., Jacquot, E., & Elena, S. F. (2012). Molecular evolution and phylogeography of Potato virus Y based on the CP gene. Journal of General Virology, 93, 2496–2501.

Domingo, E., Sheldon, J., & Perales, C. (2012). Viral quasispecies evolution. Microbiology and Molecular Biology Reviews, 76, 159–216.

Drummond, A. J., Pybus, O. G., & Rambaut, A. (2003). Inference of viral evolutionary rates from molecular sequences. Advances in Parasitology, 54, 331–488.

Drummond, A. J., Suchard, M. A., Xie, D., & Rambaut, A. (2012). Bayesian phylogenetics with BEAUti and the BEAST 1.7. Molecular Biology and Evolution, 29, 1969–1973.

Duchêne, S., Duchêne, D., Holmes, E. C., & Ho, S. Y. W. (2015). The performance of the site-randomization test in phylogenetic analyses of time-structured virus data. Molecular Biology and Evolution, 32, 1895–1906.

Edgar, R. C. (2004). MUSCLE: Multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Research, 32, 1792–1797.

Excoffier, L., & Lischer, H. E. (2010). Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. Molecular Ecology Resources, 10, 564–567.

Fu, Y. X. (1997). Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. Genetics, 147, 915–925.

Gao, F., Chang, F., Shen, J., Shi, F., Xie, L., & Zhan, J. (2014). Complete genome analysis of a novel recombinant isolate of potato virus Y from China. Archives of Virology, 159, 3439–3442.

Gibbs, A., & Ohshima, K. (2010). Potyviruses and the digital revolution. Annual Review of Phytopathology, 48, 205–223.

Gilbert, G. S. (2002). Evolutionary ecology of plant diseases in natural ecosystems. Annual Review of Phytopathology, 40, 13–43.

Gurung, S., Short, D. P., & Adhikari, T. B. (2013). Global population structure and migration patterns suggest significant population differentiation among isolates of Pyrenophora tritici-repentis. Fungal Genetics and Biology, 52, 32–41.

Hall, T. A. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series, 41, 95–98.

He, D. C., Zhan, J., Cheng, Z. B., & Xie, L. (2016). Viruliferous rate of small brown plant hopper is a good indicator of rice stripe disease epidemics. Scientific Reports, 6, 21376.

Hudson, R. R. (2000). A new statistic for detecting genetic differentiation. Genetics, 155, 2011–2014.

Hudson, R. R., Boos, D. D., & Kaplan, N. L. (1992). A statistical test for detecting geographic subdivision. Molecular Biology and Evolution, 9, 138–151.

Jansky, S. H., Jin, L. P., Xie, K. Y., Xie, C. H., & Spooner, D. M. (2009). Potato production and breeding in China. Potato Research, 52, 57–65.

Kasave, A. V., & Gray, S. M. (2013). Continuous and emerging challenges of potato virus Y in potato. Annual Review of Phytopathology, 51, 571–586.

Kawakami, T., Oochori, H., & Tajima, K. (2015). Seed potato production system in Japan, starting from foundation seed of potato. Breeding Science, 65, 17–25.

Kearney, J. (2010). Food consumption trends and drivers. Philosophical Transactions of the Royal Society B: Biological Sciences, 365, 2793–2807.

King, A. M. Q., Lefkowitz, E., Adams, M. J., & Carstens, E. B. (2011). Virus Taxonomy, Ninth Report of the International Committee on Taxonomy of Viruses. Amsterdam: Elsevier Academic Press.

Kosakovsky Pond, S. L., Frost, S. D. W., Grossman, Z., Gravenor, M. B., Richman, D. D., & Leigh Brown, A. J. (2006). Adaptation to different human populations by HIV-1 revealed by codon-based analyses. PLoS Computational Biology, 2, e62.

Kosakovsky Pond, S. L., Frost, S. D. W., & Muse, S. V. (2005). HyPhy: Hypothesis testing using phylogenies. Bioinformatics, 21, 676–679.

Kosakovsky Pond, S. L., Murrell, B., Fourment, M., Frost, S. D. W., Delpont, W., & Scheffler, K. (2011). A random effects branch-site model for detecting episodic diversifying selection. Molecular Biology and Evolution, 28, 3033–3043.

Koskella, B., & Vos, M. (2015). Adaptation in natural microbial populations. Annual Review of Ecology, Evolution, and Systematics, 46, 503–522.
Librado, P., & Rozas, J. (2009). DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. Bioinformatics, 25, 1451–1452.

Martin, D. P., Lemey, P., Lott, M., Moulton, V., Posada, D., & Lefebvre, P. (2010). RDP3: A flexible and fast computer program for analyzing recombination. Bioinformatics, 26, 2462–2463.

McDonald, B. A., & Linde, C. (2002). Pathogen population genetics, evolutionary potential, and durable resistance. Annual Review of Phytopathology, 40, 349–379.

Murrell, B., Wertheim, J. O., Moola, S., Weighill, T., Scheffler, K., & Kosakovsky Pond, S. L. (2012). Detecting individual sites subject to episodic diversifying selection. PLoS Genetics, 8, e1002764.

Nolte, P., Whitworth, J. L., Thornton, M. K., & McIntosh, C. S. (2004). Effect of seedborne Potato virus Y on performance of Russet Burbank, Russet Norkotah, and Shepody potato. Plant Disease, 88, 248–252.

Nylander, J. A. A. (2008). MrModeltest v2.3. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.

Ogawa, T., Nakagawa, A., Hataya, T., & Ohshima, K. (2012). The genetic structure of populations of Potato virus Y in Japan; based on the analysis of 20 full genomic sequences. Journal of Phytopathology, 160, 661–673.

Ogawa, T., Tomitaka, Y., Nakagawa, A., & Ohshima, K. (2008). Genetic structure of a population of Potato virus Y inducing potato tuber necrotic ringspot disease in Japan; comparison with North American and European populations. Virus Research, 131, 199–212.

Park, J., Rambaut, A., & Pybus, O. G. (2008). Correlating viral phenotypes with phylogeny: Accounting for phylogenetic uncertainty. Infection, Genetics and Evolution: Journal of Molecular Epidemiology and Evolutionary Genetics in Infectious Diseases, 8, 239–246.

Quenouille, J., Vassilakos, N., & Moury, B. (2013). Potato virus Y: A major crop pathogen that has provided major insights into the evolution of viral pathogenicity. Molecular Plant Pathology, 14, 439–452.

Ramsden, C. H., Holmes, E. C., & Charleston, M. A. (2009). Hantavirus evolution in relation to its rodent and insectivore hosts: No evidence for codivergence. Molecular Biology and Evolution, 26, 143–153.

Revell, L. J., Harmon, L. J., & Collar, D. C. (2008). Phylogenetic signal, evolutionary process, and rate. Systematic Biology, 57, 591–601.

Rieux, A., & Khatchikian, C. E. (2016). TipDatingBeast: An R package to assist the implementation of phylogenetic tip-dating tests using BEAST. Molecular Ecology Resources, doi:10.1111/1755-0998.12603 [Epub ahead of print].

Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D. L., Darling, A., Hohna, S., ... Huelsenbeck, J. P. (2012). MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology, 61, 539–542.

Short, D. P., Gurung, S., Gladieux, P., Inderbitzin, P., Atallah, Z. K., Nigro, F., ... Subbarao, K. V. (2015). Globally invading populations of the fungal plant pathogen Verticillium dahliae are dominated by multiple divergent lineages. Environmental Microbiology, 17, 2824–2840.

Tajima, F. (1989). Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics, 123, 585–595.

Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., & Kumar, S. (2011). MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Molecular Biology and Evolution, 28, 2731–2739.

Tena Fernandez, F., Gonzalez, I., Doblas, P., Rodriguez, C., Sahana, N., Kaur, H., ... Canto, T. (2013). The influence of cis-acting P1 protein and translational elements on the expression of Potato virus Y helper-component protease (HCPro) in heterologous systems and its suppression of silencing activity. Molecular Plant Pathology, 14, 530–541.

Thrall, P. H., Oakeshott, J. G., Fitz, G., Southerton, S., Burdon, J. J., Sheppard, A., ... Denison, R. F. (2011). Evolution in agriculture: The application of evolutionary approaches to the management of biotic interactions in agro-ecosystems. Evolutionary Applications, 4, 200–215.

Tian, Y. P., Liu, J. L., Zhang, C. L., Liu, Y. Y., Wang, B., Li, X. D., ... Valkonen, J. P. (2011). Genetic diversity of Potato virus Y infecting tobacco crops in China. Phytopathology, 101, 377–387.

Tian, Y. E., Yin, J. L., Sun, J. P., Ma, Y. F., Wang, Q. H., Quan, J. L., & Shan, W. X. (2016). Population genetic analysis of Phytophthora infestans in northwestern China. Plant Pathology, 65, 17–25.

Tredway, L. P., Stevenson, K. L., & Burpee, L. L. (2005). Genetic structure of Magnaporthe grisea populations associated with St. Augustinegrass and tall fescue in Georgia. Phytopathology, 95, 463–471.

Vall, A., Lopez-Moya, J. J., & Garcia, J. A. (2007). Recombination and gene duplication in the evolutionary diversification of P1 proteins in the family Potyviridae. Journal of General Virology, 88, 1016–1028.

Wang, B., Ma, Y., Zhang, Z., Wu, Z., Wu, Y., Wang, Q., & Li, M. (2011). Potato viruses in China. Crop Protection, 30, 1117–1123.

Wang, J., & Whitlock, M. C. (2003). Estimating effective population size and migration rates from genetic samples over space and time. Genetics, 163, 429–446.

Wright, S. (1938). Size of population and breeding structure in relation to evolution. Science, 87, 425–431.

Yang, Z. (2007). PAML 4: Phylogenetic analysis by maximum likelihood. Molecular Biology and Evolution, 24, 1586–1591.

Yang, Z., Wong, W. S., & Nielsen, R. (2005). Bayes empirical Bayes inference of amino acid sites under positive selection. Molecular Biology and Evolution, 22, 1107–1118.

Yang, L. N., Zhu, W., Wu, E. J., Yang, C., Thrall, P. H., Burdon, J. J., ... Zhan, J. (2016). Trade-offs and evolution of thermal adaptation in the Irish potato famine pathogen Phytophthora infestans. Molecular Ecology, 25, 4047–4058.

Yu, S. F., Benton, P., Bovee, M., Sessions, J., & Lloyd, R. E. (1995). Defective RNA replication by poliovirus mutants deficient in 2A protease cleavage activity. Journal of Virology, 69, 247–252.

Zhan, J., & McDonald, B. A. (2004). The interaction among evolutionary forces in the pathogenic fungus Mycosphaerella graminicola. Fungal Genetics and Biology, 41, 590–599.

Zhan, J. S., & McDonald, B. A. (2013). Experimental measures of pathogen competition and relative fitness. Annual Review of Phytopathology, 51, 131–153.

Zhan, J., Pettway, R. E., & McDonald, B. A. (2003). The global genetic structure of the wheat pathogen Mycosphaerella graminicola is characterized by high nuclear diversity, low mitochondrial diversity, regular recombination, and gene flow. Fungal Genetics and Biology, 38, 286–297.

Zhan, J. S., Thrall, P. H., & Burdon, J. J. (2014). Achieving sustainable plant disease management through evolutionary principles. Trends in Plant Science, 19, 570–575.

Zhan, J. S., Thrall, P. H., Papaix, J., Xie, L., & Burdon, J. J. (2015). Playing on a pathogen’s weakness: Using evolution to guide sustainable plant disease control strategies. Annual Review of Phytopathology, 53, 19–43.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Gao F, Zou W, Xie L, Zhan J-S. Adaptive evolution and demographic history contribute to the divergent population genetic structure of Potato virus Y between China and Japan. Evol Appl. 2017;10:379–390. https://doi.org/10.1111/eva.12459