Three new species of eriophyoid mites
(Acari, Eriophyoidea) from Xinjiang
Uygur Autonomous Region, China

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Abstract
Three new species of eriophyoid mites from Xinjiang Uygur Autonomous Region, China, are described and illustrated. They are Paracolomerus gonglius sp. n. and Phyllocoptruta beggerianae sp. n. collected on Rosa beggeriana Schrenk ex Fisch. & C. A. Mey. (Rosaceae), and Rhyncaphytoptus fuyuniensis sp. n. collected on Cotoneaster ignavus E. L. Wolf (Rosaceae). All eriophyoid mites described here are vagrants on the undersurface of leaves and any apparent damage was not observed.

Keywords
Taxonomy, Colomerini, Phyllocoptrini, Rhyncaphytoptinae, Rosaceae

Introduction
Eriophyoid mites (Acari: Eriophyoidea) have been recognized as important pests in agriculture and forestry all over the world (Lindquist et al. 1996). Their stylets are involved in piercing plant cells, injecting saliva into them and sucking cell sap (de Lillo et al. 2002). The saliva causes cytological, biochemical and physiological changes in the pierced plants (de Lillo and Monfreda 2004, Petanović and Kielkiewicz 2010a).
Eriophyoid mites induce plant malformations as galls, complex symptoms or vector pathogens disturbing the normal growth of plants (Petanović and Kielkiewicz 2010b). This is the case of Colomerus vitis (Pagenstecher), Aceria pallida Keifer and Tegolophus zizyphagus (Keifer) which induce erinea, galls or leaf edge curls and cause economic losses to grape, matrimony vine and jujube, respectively, in Xinjiang (Lu and Mao 1990, Zang 1998, Yang et al. 2012). However, about half eriophyoid mite species are vagrants on the surface of leaves and do not cause any apparent damage (Huang 2008, Skoracka et al. 2010, Petanović and Kielkiewicz 2010b). These mites, occurring in a large amount, may cause non-distortive changes and affect the normal growth of the plants (Oldfield 1996). Usually eriophyoids are tiny in size and hard to see with unaided eyes. Sometimes their symptoms can be confused with those due to viruses, nutrient deficiency and physiological disorders (Van Leeuwen et al. 2010). Therefore, it is necessary to study the systematic account of Eriophyoidea for having a further contribute in better understanding their significance in Agriculture.

Kuang (1995) first explored and reported the eriophyoid mite fauna in Xinjiang. After that, a number of field surveys were conducted in the same area and further 31 species were reported so far. Out of 31 species, 1 species belongs to the family Phytoptidae, 2 species belong to the family Diptilomiopidae and 28 species belong to the family Eriophyidae (Table 1). The fact that more than 1000 species have been recognized from China (personal data of X.-F. Xue) suggests that many more areas need to be explored more carefully. For this purpose eriophyoid mites were collected by Ji-Wei Li from Tianshan Mountains, Altai Mountains, Farmlands and Gurbantunggut Desert of Xinjinag in 2013 and 2014.

In the present study, we describe 2 new species of the genera Paracolomerus and Phyllocoptruta collected on Rosa beggeriana (Fig. 1) and one new species of the genus Rhyncaphytoptus collected on Cotoneaster ignavus (Fig. 1), all from Xinjiang. Also, this is the first description of the genus Paracolomerus from Rosaceae.
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Table 1. List of eriophyoid mites from Xinjiang Uygur Autonomous Region.

| Family         | Subfamily | Tribe         | Species                                      | Reference                     |
|----------------|-----------|---------------|----------------------------------------------|-------------------------------|
| Phytoptidae    | Nalepinae | Nalepellini   | *Setoptus koraiensis* Kuang & Hong, 1995     | Xue et al. 2012a: 12.         |
| Eriophyidae    | Cecidophyinae | Colomerini | *Colomerus vitis* (Pagenstecher, 1857)       | Kuang 1995: 42–43.            |
|                |           |               | *Paracolomerus gongliu* sp. n.               | This study                    |
|                | Eriophyinae | Eriophyini    | *Eriophyes catacardiae* Keifer, 1962          | Kuang 1995: 47.               |
|                |           | Acerini       | *Aceria bromi* Kuang & Zhang, 1992           | Kuang 1995: 53–54.            |
|                |           |               | *Aceria changiensis* Kuang & Pang, 1997      | Kuang and Pang 1997: 230–231. |
|                |           |               | *Aceria dispars* (Nalepa, 1891)              | Song et al. 2008: 13.         |
|                |           |               | *Aceria haloxylonis* Xue, Zhang, Li & Hong, 2012 | Xue et al. 2012b: 203–208.   |
|                |           |               | *Aceria nimit* Hall, 1967                    | Kuang et al. 2005: 29.        |
|                |           |               | *Aceria pallida* Keifer, 1964                 | Hong et al. 2006: 230.        |
|                |           |               | *Aceria tamaricis* (Trotter, 1901)           | Kuang et al. 2005: 33–34.     |
|                |           |               | *Aceria tosichella* Keifer, 1969              | Kuang 1995: 64–65.            |
| Phylloxoptinae | Phylloxoptini |          | *Castaneirinus alchemillae* (Liro, 1940)     | Kuang et al. 2005: 58–59.     |
|                |           |               | *Epireptes sabiniae* Xue & Hong, 2005        | Xue et al. 2012a: 31.         |
|                |           |               | *Phylloxoptes pyrivagnus* Kadono, 1985        | Kuang et al. 2005: 68–69.     |
|                |           |               | *Phylloxoptera beggeriana* sp. n.            | This study                    |
|                |           |               | *Proctus tabulaeformis* Xue, Song, Amrine & Hong, 2007 | Xue et al. 2012a: 39.         |
| Anthocoptini   |           |               | *Aculodes shibezeiensis* Kuang, Lu & Zhang, 2005 | Kuang et al. 2005: 81–82.     |
|                |           |               | *Aculops alopecuroides* Kuang, 1998           | Kuang 1998: 410–411.          |
|                |           |               | *Aculops sativis* Xue, Song & Hong, 2007     | Xue et al. 2012a: 41–42.      |
|                |           |               | *Aculus amygdali* Xue & Hong, 2005           | Xue et al. 2012a: 43.         |
|                |           |               | *Aculus schlechtendi* (Nalepa, 1890)         | Kuang 1995: 120–121.          |
|                |           |               | *Aculus tenuothrix* (Nalepa, 1889)           | Kuang 1995: 131–132.          |
|                |           |               | *Tegolophus zizyphagoides* (Keifer, 1939)    | Kuang 1995: 146–147.          |
|                |           |               | *Tetra cuira* Xue, Song & Hong, 2006         | Xue et al. 2012a: 54.         |
|                |           |               | *Tetra nitratea* Li, Li, Zhang & Xue, 2014   | Li et al. 2014: 348–351.      |
|                |           |               | *Tetra sativa* Li, Li, Zhang & Xue, 2014     | Li et al. 2014: 339–343.      |
| Family          | Subfamily        | Tribe                      | Species                  | Reference                        |
|-----------------|------------------|----------------------------|--------------------------|----------------------------------|
| Diptilomiopidae | Rhyncaphytoptinae|                            | *Rhyncaphytoptus fuyuniensis* sp. n. | This study                      |
|                 |                  |                            | *Rhyncaphytoptus yilisis* Song, Xue & Hong, 2007 | Song et al. 2007: 63–65.         |
|                 |                  |                            | *Rhyncaphytoptus ziziphi* Kuang, 2005 | Kuang et al. 2005: 157–158.       |
| Tetra shiheziensis | Wang & Lu, 2004 |                            |                          | Wang and Lu 2004: 266–267.       |
| Tetra tianchiensis | Li, Li, Zhang & Xue, 2014 |                            |                          | Li et al. 2014: 335–339.         |
| Tetra tianschanicae | Li, Li, Zhang & Xue, 2014 |                            |                          | Li et al. 2014: 330–334.         |
| Tetra viciae     | Li, Li, Zhang & Xue, 2014 |                            |                          | Li et al. 2014: 343–348.         |
Materials and methods

Specimens of mites were collected from Xinjiang Uygur Autonomous Region, China. The morphological terminology used here follows Lindquist (1996). The generic classification of the eriophyoid mites is made according to Amrine et al. (2003), together with the comparison of genera erected after 2003. Specimens were cleared in Keifer’s booster and slides were mounted using modified Berlese medium (Amrine and Manson 1996). The number of measured specimens (n) is given within parentheses in the description. All specimens were examined, measured, taken photos and drawn with the aid of an Olympus Bx61 microscope using phase contrast. The measurements and drawings were based on the methods outlined by de Lillo et al. (2010) and abbreviations follow Amrine et al. (2003). For each species, the holotype female measurement precedes the corresponding range for paratypes (given in parentheses). For males, only ranges are given. All measurements are given in micrometres and are lengths unless specified. Type specimens are deposited at the Department of Plant Protection, College of Agriculture, Shihezi University, Xinjiang Uygur Autonomous Region, China.

Results

Family Eriophyidae Nalepa, 1898
Subfamily Cecidophyinae Keifer, 1966
Tribe Colomerini Newkirk & Keifer, 1975
Genus Paracolomerus Keifer, 1975

Paracolomerus gonglius sp. n.
http://zoobank.org/36D41CF4-E10C-47F1-8635-3E2CC695A2CB
Fig. 2

Description. FEMALE (n=6). Body vermiform, 187 (175–217, excluding gnathosoma), 50 (42–51) wide, 48 (40–47) thick; light yellow. Gnathosoma 24 (23–26), projecting obliquely down, pedipalp coxal setae (ep) 2 (2–3), dorsal pedipalp genual setae (d) 7 (6–8), unbranched, cheliceral stylets 21 (20–23). Prodorsal shield 30 (29–32), 37 (33–37) wide; median line almost complete, interrupted in the middle with short sloping lines on either side at the posterior end, admedian lines complete, submedian lines broken, with several short lines and granules on the lateral side; frontal shield lobe absent. Scapular tubercles near rear shield margin, 24 (23–24) apart, scapular setae (sc) 15 (14–15), projecting posterior. Coxigenital region with 5 (5–6) microtuberculated semiannuli. Coxal plates with several short lines, anterolateral setae on coxisternum I (1b) 7 (7–8), 11 (10–11) apart, proximal setae on coxisternum I (1a) 25 (25–28), 13 (12–13) apart, proximal setae on coxisternum II (2a) 44 (39–44), 24 (23–24) apart, tubercles 1b and 1a apart 5 (5–6), tubercles 1a and 2a 8 (7–8) apart. Internal coxisternal apodeme absent. Legs with usual series of setae. Leg I 27 (26–28), femur 8 (7–8), basiventral femoral setae (bv) 12
Figure 2. Schematic drawings of *Paracolomerus gonglius* sp. n.: **AL** Lateral view of anterior body region **CG** Female coxigenital region **D** Dorsal view **em** Empodium **IG** Internal female genitalia **LO** Lateral view of annuli **L1** Leg I **PM** Lateral view of posterior opisthosoma. Scale bar: 15 μm (**D**); 10 μm (**AL, CG, IG, PM**); 7.5 μm (**LO**); 5 μm (**L1**); 2.5 μm (**em**).
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Leg II 25 (24–26), femur 7 (7–8), basiventral femoral setae (bv) 13 (13–14); genu 5 (4–5), antaxial genual setae (l) 7 (7–8), located in the middle; tarsus 7 (6–7), setae ft’ 15 (14–15), setae ft” 22 (20–22), seta u’ 4 (4–5); tarsal empodium (em) 6 (5–6), simple, 5-rayed, tarsal solenidion (ω) 7 (7–8), rod-like. Leg II 25 (24–26), femur 7 (7–8), basiventral femoral setae (bv) 13 (13–14); genu 5 (4–5), antaxial genual setae (l”) 8 (8–10); tibia 5 (4–5); tarsus 7 (6–7), setae ft’ 6 (6–7), setae ft” 23 (23–25), seta u’ 4 (4–5); tarsal empodium (em) 6 (5–6), simple, 5-rayed, tarsal solenidion (ω) 9 (9–10), rod-like. Genital coverflap 11 (11–12), 22 (21–22) wide, coverflap with two rows of ridges, the basal one with 12 (11–13) longitudinal ridges, the other one with 9 (8–10) longitudinal ridges, proximal setae on coxisternum III (3a) 17 (17–20), 17 (17–18) apart. MALE. Unknown. Type host plant. Rosa beggeriana Schrenk ex Fisch. & C. A. Mey. (Rosaceae). Relation to the host plant. Vagrant on leaves; no apparent damage was observed. Type locality. Gongliu county, Xinjiang Uygur Autonomous Region, China (43°09’37"N, 81°36’34"E), elevation 1,396 m; 3 August 2014, coll. Ji-Wei Li. Type material. Holotype: female (slide number SHZU Paracolomerus 1.1, marked Holotype). Paratypes: 6 females mounted on 6 separate microscope slides. Etymology. The specific designation gonglius comes from the name of location, Gongliu, where the new species was collected. Differential diagnosis. All traits are in accordance with the type species Paracolomerus casimiroae Keifer, 1975 of the genus Paracolomerus (opisthosomal annuli subequal, legs with usual series of setae, scapular tubercles on rear shield margin, scapular setae projected posteriorly) except for ventral surface ornamentation of coxa I (lines do not circle around tubercles 1a and meet at sternum in P. gonglius sp. n.; lines originate at setae 1b, circle distally around tubercles 1a and meet at sternum, enclose most of the coxal surface in P. casimiroae). This species is similar to P. fopingacer Xue, Song & Hong, 2011, from Acer sp. L. (Aceraceae), but can be differentiated from the latter by median line almost complete, with 5–6 short lines on the lateral sides of prodorsal shield (median line present for half, without short lines on the lateral sides in P. fopingacer), frontal shield lobe absent (frontal shield lobe acuminate in P. fopingacer) and 5-rayed empodium (6-rayed empodium in P. fopingacer). Remarks. To date, only three species were reported from the genus Paracolomerus, P. casimiroae Keifer, 1975, P. davidiae Kuang & Hong, 1995 (in Kuang 1995) and P. fopingacer.
Subfamily Phyllocoptinae Nalepa, 1892
Tribe Phyllocoptini Nalepa, 1892
Genus *Phyllocoptruta* Keifer, 1938

*Phyllocoptruta beggerianae* sp. n.
http://zoobank.org/D33691AE-25DD-4A17-A854-E7C9925C96FF

Fig. 3

**Description.** FEMALE (n=9). Body fusiform, 207 (182–207, excluding gnathosoma), 49 (46–51) wide, 43 (40–46) thick; white. **Gnathosoma** 28 (27–30), projecting obliquely down, pedipalp coxal setae (*ep*) 3 (2–3), dorsal pedipalp genual setae (*d*) 9 (8–9), unbranched, cheliceral stylets 26 (25–27). **Prodorsal shield** 42 (40–43), 43 (41–44) wide, median line formed by lined short lines, admedian lines complete and connected posteriorly, submedian lines present at the posterior half, with several short lines and granules; frontal shield lobe rounded, broad-based, 5 (4–5). Scapular tubercles ahead of rear shield margin, 23 (22–25) apart, scapular setae (*sc*) 16 (16–18), projecting forward and convergent. **Coxigenital region** with 9 (7–9) microtuberculated semiannuli. Coxal plates with several short lines and granules, anterolateral setae on coxisternum I (*1b*) 11 (10–11), 12 (11–12) apart, proximal setae on coxisternum I (*1a*) 27 (26–31), 10 (9–10) apart, proximal setae on coxisternum II (*2a*) 46 (42–46), 25 (23–25) apart, tubercles *1b* and *1a* apart 7 (6–7), tubercles *1a* and *2a* 9 (8–9) apart. Internal coxisternal apodeme 3 (3–4). Legs with usual series of setae. **Leg I** 36 (35–37), femur 10 (9–10), basiventral femoral setae (*bv*) 14 (13–15); genu 5 (4–5), antaxial genual setae (*l'* 22 (22–25); tibia 8 (8–9), paraxial tibial setae (*l*') 12 (10–12), located at 1/3 from dorsal base; tarsus 9 (8–9), setae *ft'* 19 (19–21), setae *ft''* 22 (22–25), seta *u'* 10 (9–10); tarsal empodium (*em*) 8 (7–8), simple, 8-rayed, tarsal solenidion (*ω*) 10 (9–10), rod-like. **Leg II** 29 (28–30), femur 8 (8–9), basiventral femoral setae (*bv*) 16 (15–16); genu 4 (4–5), antaxial genual setae (*l'*') 8 (6–8); tibia 5 (5–6); tarsus 8 (7–8), setae *ft''* 9 (8–10), setae *ft''* 23 (23–25), seta *u'* 9 (8–9); tarsal empodium (*em*) 8 (7–8), simple, 8-rayed, tarsal solenidion (*ω*) 10 (9–10), rod-like. **Opisthosoma** dorsally with a furrow in the middle; 35 (33–38) dorsal annuli, elliptical microtubercles on the rear margin; 74 (72–77) ventral annuli, microtubercles on the rear margin, circled on the anterior ventral annuli, and linear and spiny on the last posterior ventral annuli. Setae *c2* 28 (25–28) on ventral annulus 14 (12–14), 47 (45–48) apart; setae *d* 52 (48–52) on ventral annulus 28 (27–29), 35 (32–35) apart; setae *e* 33 (33–36) on ventral annulus 49 (47–51), 13 (12–14) apart; setae *f* 28 (26–29) on 6th ventral annulus from rear, 16 (16–17) apart. Setae *h1* 4 (3–4), *h2* 77 (75–83). **Genital coverflap** 13 (11–13), 19 (18–20) wide, coverflap with 3 transverse lines basally, 11 (11–13) longitudinal ridges distally, proximal setae on coxisternum III (*3a*) 43 (41–44), 15 (14–15) apart.

**MALE** (n=2). Similar in shape and prodorsal shield arrangement to female, 155–169. Prodorsal shield 32–35, 28–31 wide; scapular setae *sc* 16–17, 21–23 apart. Opisthosoma dorsally with a furrow, 32–37 annuli, ventrally with 74–81 annuli, dor-
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Figure 3. Schematic drawings of *Phyllocoptruta beggerianae* sp. n.: AL Lateral view of anterior body region AD Dorsal view of anterior body region CG Female coxigenital region D Dorsal view em Empodium GM Male genital region IG Internal female genitalia LO Lateral view of annuli PM Lateral view of posterior opisthosoma. Scale bar: 15 μm (D); 10 μm (AD, AL, CG, IG, GM, PM); 7.5 μm (LO); 2.5 μm (em).
sal and ventral microtubercles are similar to females. Male genitalia 17–18 wide, setae 3a 18–20, 14–15 apart.

**Type host plant.** *Rosa beggeriana* Schrenk ex Fisch. & C. A. Mey. (Rosaceae).

**Relation to the host plant.** Vagrant on leaves; no apparent damage was observed.

**Type locality.** Xinyuan county, Xinjiang Uygur Autonomous Region, China (43°36′29″N, 82°17′56″E), elevation 758 m; 29 July 2014, coll. Ji-Wei Li.

**Type material.** Holotype: female (slide number SHZU Phyllocoptruta 1.1, marked Holotype). Paratypes: 16 females and 2 males mounted on 18 separate microscope slides.

**Etymology.** The specific designation *beggerianae* comes from the epithet of the host plant, *beggeriana*.

**Differential diagnosis.** This species is similar to *Phyllocoptruta clematoclethra* Xue, Song & Hong, 2010, from *Clematoclethra* sp. Maxim. (Actinidiaceae), but can be differentiated from the latter by admedian lines connected posteriorly (admedian lines separate in *P. clematoclethra*), scapular tubercles ahead of rear shield margin, scapular setae 16–18 (scapular tubercles on rear shield margin, scapular setae 3–4 in *P. clematoclethra*), female genital coverflap with 3 transverse basal lines (coverflap without transverse lines in *P. clematoclethra*) and 8-rayed empodium (5-rayed empodium in *P. clematoclethra*).

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**Family Diptilomiopidae Keifer, 1944**

**Subfamily Rhyncaphytoptinae Roivainen, 1953**

**Genus Rhyncaphytoptus Keifer, 1939**

*Rhyncaphytoptus fuyuniensis* sp. n.

http://zoobank.org/CE644BB3-52C6-43E6-AAC7-06BC248177F7

Figs 4–5

**Description.** FEMALE (n=8). Body fusiform, 256 (216–267, excluding gnathosoma), 60 (55–64) wide, 58 (54–62) thick; light yellow. Gnathosoma 61 (55–64), projecting downwards, pedipalp coxal setae (ep) 3 (2–3), dorsal pedipalp genual setae (d) 7 (6–7), unbranched, cheliceral stylets 83 (76–88). **Prodorsal shield** 29 (28–30) excluding the thin anterior process length from frontal lobe, 47 (46–49) wide, sub-triangular in anterior shape; long and flexible frontal lobe ending with a thin anterior process, the process extends for 14 (13–15). Median line very short, on 1/5 anterior part of prodorsal shield; admedian lines complete and connected at base with transverse lines, forming a vase-shaped outline; semicircled line between the scapular tubercles. Scapular tubercles ahead of rear shield margin, 30 (28–31) apart, scapular setae (sc) 47 (46–50), projecting forward. **Coxigenital region** with 15 (14–16) microtuberculated semiannuli. Coxal plates with 1–3 short lines, anterolateral setae on coxisternum I (1b) 10 (10–12), 10 (10–11) apart, proximal setae on coxisternum I (1a) 32 (29–33), 10 (10–11) apart, proximal setae on coxisternum II (2a) 45 (42–47), 29 (28–30) apart, tubercles 1b and 1a apart 7 (6–7), tubercles 1a and 2a 10 (9–11) apart. Internal coxisternal apodeme 7 (6–7). Legs with
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usual series of setae. **Leg I** 42 (41–43), femur 13 (12–14), basiventral femoral setae (bv) 13 (13–15); genu 7 (6–7), antaxial genual setae (l’) 23 (23–25); tibia 10 (10–11), paraxial tibial setae (l) 7 (7–8), located at 1/3 from dorsal base; tarsus 8 (7–8), setae ft’ 20 (18–20), setae ft’’ 26 (23–26), seta u’ 5 (4–5); tarsal empodium (em) 8 (7–8), simple, 10-rayed, tarsal solenidion (ω) 8 (8–9), rod-like. **Leg II** 39 (38–40), femur 13 (12–13), basiventral femoral setae (bv) 14 (14–16); genu 6 (5–6), antaxial genual setae (l’) 9 (9–11); tibia 8 (7–9); tarsus 8 (8–9), setae ft’ 10 (9–11), setae ft’’ 30 (26–30), seta u’ 5 (4–5); tarsal empodium (em) 8 (7–8), simple, 10-rayed, tarsal solenidion (ω) 10 (10–11), rod-like. **Opisthosoma** dorsally arched, 25 (20–25) dorsal annuli, 92 (90–104) microtuberculated ventral annuli; the anterior dorsal annuli smooth (for about 5/6 of them), the anterior ventral annuli with circled microtubercles (for about 2/3 of them), the posterior part of dorsal and ventral annuli with linear and spiny microtubercles. Setae c2 13 (12–14) on ventral annulus 19 (17–21), 59 (53–61) apart; setae d 51 (46–51) on ventral annulus 37 (36–42), 45 (41–45) apart; setae e 26 (26–29) on ventral annulus 55 (53–64), 25 (24–26) apart; setae f30 (27–30) on 7th ventral annulus from rear, 21 (20–22) apart. Setae h1 3 (3–4), h2 75 (70–79). **Genital coverflap** 15 (14–16), 30 (28–30) wide, coverflap with many granules basally, proximal setae on coxisternum III (3a) 14 (13–14), 20 (19–21) apart.

**MALE** (n=5). Similar in shape and prodorsal shield arrangement to female, 202–243. Prodorsal shield 22–25 without the frontal lobe length, 45–48 wide; scapular setae sc 40–46, 27–30 apart. Opisthosoma dorsally with 19–21 annuli; ventrally with 78–85 annuli, dorsal and ventral microtubercles are similar to females. Male genitalia 21–22 wide, setae 3a 12–13, 18–20 apart.
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**Type host plant.** *Cotoneaster ignavus* E. L. Wolf (Rosaceae).

**Relation to the host plant.** Vagrant on leaves; no apparent damage was observed.

**Type locality.** Fuyun county, Xinjiang Uygur Autonomous Region, China (47°17’39”N, 89°58’26”E), elevation 1,359 m; 15 August 2014, coll. Ji-Wei Li.

**Type material.** Holotype: female (slide number SHZU Rhyncaphytoptus 7.1, marked Holotype). Paratypes: 12 females and 15 males mounted on 27 separate microscope slides.

**Etymology.** The specific designation *fuyuniensis* comes from the name of location, Fuyun, where the new species was collected.

**Differential diagnosis.** This species is similar to *Rhyncaphytoptus buxifoliae* Song, Xue & Hong, 2009, from *Cotoneaster buxifolius* Lindl. (Rosaceae), but can be differentiated from the latter by median line very short, on 1/5 anterior part of prodorsal shield (prodorsal shield with incomplete median line on posterior 1/2 in *R. buxifoliae*), scapular tubercles small (scapular tubercles 5–13 long in *R. buxifoliae*) and with a long and flexible frontal lobe (lack a distinct, long frontal lobe in *R. buxifoliae*).

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