The relationships between neuroglial alterations and neuronal changes in Alzheimer’s disease, and the related controversies I: Gliopathogenesis and glioprotection

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Abstract

Since Alois Alzheimer described the pathology of Alzheimer’s disease in 1907, an increasing number of studies have attempted to discover its causes and possible ways to treat it. For decades, research has focused on neuronal degeneration and the disruption to the neural circuits that occurs during disease progression, undervaluing in some extent the alterations to glial cells even though these alterations were described in the very first studies of this disease. In recent years, it has been recognized that different families of neuroglia are not merely support cells for neurons but rather key and active elements in the physiology and pathology of the nervous system. Alterations to different types of neuroglia (especially astroglia and microglia but also mature oligodendroglia and oligodendroglial progenitors) have been identified in the initial neuropathological changes that lead to dementia, suggesting that they may represent therapeutic targets to prevent neurodegeneration. In this review, based on our own studies and on the relevant scientific literature, we argue that a careful and in-depth study of glial cells will be fundamental to understanding the origin and progression of Alzheimer’s disease. In addition, we analyze the main issues regarding the neuroprotective and neurotoxic role of neuroglial changes, reactions and/or involutions in both humans with Alzheimer’s disease and in experimental models of this condition.

Keywords: Alzheimer’s disease, gliopathogenesis, glioprotection, astroglia, oligodendroglia, microglia, astrogliosis, microgliosis

Introduction

Alzheimer’s disease (AD) is a neurodegenerative syndrome that leads to dementia and was first described more than a century ago by Alois Alzheimer.1 A few years ago, AD was described as an “epidemic of the 21st century”, both due to the large number of cases expected (more than 20 million people by 2020) and the enormous economic and social burden it places on countries around the globe.2 For many decades, researchers from different disciplines around the world have been attempting to find the causes of this neurodegenerative process and to find a treatment to prevent or treat it once dementia has set in.

Since the original description of the disease, research into AD has mainly focused on assessing the alterations to the neurons and circuits that might provoke the cognitive decline associated with dementia (the “neuron-centric” theory of AD).1-7 Over the years, different techniques (neuropathological, biochemical, biomolecular, genetic, etc.) have been used to study neurons in normal and pathological situations, both in “healthy” and “Alzheimer” individuals, as well as in experimental models of AD.8-18 In most human studies and in many pathogenic models of AD, was considered that glial cells also “suffer concomitant alterations”, and these alterations are thought to aggravate neurodegeneration,19-32 representing key changes in the pathogenic cascades that provoke neurodegeneration/dementia. However, the specific study of glial cells in AD begins many decades after (except for exceptions that are later mentioned). It was necessary to develop new technologies to study the types and subtypes of glial cells and their functions, as well as to understand and accept that these cells were intimately involved in the functions of neurons and neuronal circuits, to focus on them the investigation of neurodegenerative disorders.22-31 As such, in recent years, astrogliosis and microgliosis, the main drivers of neuroinflammation,24,25,27-31,33-39 as well as changes in oligodendroglial cells, have been integrated into all the pathogenic theories of AD.33-36,40 In fact, we can even consider that “neuroglial” theories of AD now exist.

The importance of glia in AD was recognized long ago by the Spanish neuroscientist Nicolas Achucarro, a collaborator of both Alois Alzheimer and Santiago Ramón y Cajal. In Achucarro’s studies (1910-1925), he pointed out how astroglia and microglial cells undergo important changes during human neurodegeneration, indicating that they should not be underestimated.3,19,21,26,41 However, there was a lack of technologies capable of differentiating subtypes of neuroglial cells and their functions to analyze their real involvement in the disease. In more recent decades, new research supported by new technologies (histochemical, biochemical, genic, and so on), significantly increased our understanding of glial cells (23-36, 40). Accordingly, glial cells are no longer merely considered...
support cells for neurons and neuronal circuits; instead, they are thought to function very actively in maintaining the morphology and correct functioning of the nervous system as well as in the adaptive responses on which cognitive and behavioral processes are based.\textsuperscript{32-31} Therefore, alterations to glia underlie pathological processes, and thus, it is logical that therapeutic interventions aimed at maintaining or improving their activity will help prevent or treat neurodegeneration.

There is already much support for the claim that studying glial alterations in AD will provide a basis to understand and treat this disease.\textsuperscript{23,48,49,52-57} However, unfortunately, we remain far from understanding the precise role of the different glial cells in each phase of every neuropathological process.\textsuperscript{3,26,58,59} Furthermore, many of the observations made in the human brain, as well as results obtained in animal models of AD, have generated some controversy about the true role (neuroprotective or neurotoxic) of the changes experienced by the different types of glial cells. Nevertheless, we do have enough information to establish that understanding the alterations to glia will shed some light on the mechanisms underlying different neuropathological processes; this information will certainly help to reveal therapeutic targets to fight these diseases. In this first part of this review, we consider the general characteristics of neuroglial cells in AD to understand their potential role in the pathogenesis, diagnosis and treatment (preventive or palliative) of this condition. We consider these issues in the light of differentiated and/or concurrent pathogenic and neuroprotective/neuroreparative processes. In the second part of this monograph (“The relationships between neuroglial alterations and neuronal changes in Alzheimer’s Disease, and the existing controversies. II Gliotherapy and multimodal AD therapy”) we will study the therapeutic possibilities that are being analyzed to try to prevent or palliate the onset or progression of this disease, based on treatments aimed at maintaining the normal functions of neuroglial cells or normalizing the alterations that occur in the progression of AD.

**Some considerations about neuroglial cells**

From Santiago Ramón y Cajal’s pioneering studies and those of his disciples and peers,\textsuperscript{59-22,27,41,60-69} we know that different types of glial cells accompany neurons in the central nervous system (CNS). These glial cells can be grouped into three main classes, initially characterized by their shape and their relationship with neurons: astroglia,\textsuperscript{3,19-26,61-66} oligodendroglia\textsuperscript{70,71} and microglia.\textsuperscript{27,28,58,67,69} However, as years have passed, we can now further differentiate these cells according to their functions, especially in terms of maintaining or not the activity of neurons and neural circuits or the production of different types of neuroactive substances (neurotoxic or neuroprotective astrocytes\textsuperscript{43,59} or microglial cells\textsuperscript{44-46} (see later).

**Astroglia**, also known as astrocytes, are cells of a neuroepithelial origin that adopt a series of morphologies in different CNS regions: protoplasmic astrocytes, predominantly in the gray matter; fibrillar astrocytes, predominantly in the white matter; neuroepithelial cells in the cerebellum; subpial astrocytes, etc.\textsuperscript{60-66}

Over the years, it has been shown that these cells are not only mere support cells for neurons but also that they contribute in a transcendental way in neuronal functions. As such, they not only maintain the homeostasis of the environment in which neuronal circuits develop but also participate in the development and maintenance of synapses, as well as in neurotransmission throughout the life of the individual.\textsuperscript{35,40,42,43,47,48,50-53,72}

Astroglia display a high degree of plasticity (glioplasticy), adapting to different situations in conjunction with their accompanying neurons to achieve the optimal degree of neuron functionality and the best response to normal and/or abnormal changes in the CNS.\textsuperscript{3,26} A significant advance in the study of astroglia has been understanding that some cell subtypes are able to produce active substances that protect neurons and contribute to neurotransmission (neurotrophic factors, neuroprotective substances, gliotransmitters, etc.). However, it was also found that reactive astrocytes can produce neurotoxic substances that induce neurodegeneration (cytokines, chemokines, free radicals, nitric oxide -NO-, prostaglandins, etc.)\textsuperscript{23-26,35,42,43,47,53}

In recent years, the study of astroglia has intensified with the application of new biochemical and molecular biology techniques\textsuperscript{50,52,53,73-87} and has revealed new characteristics and functions. As a result, in addition to the morphologically defined subtypes, new subtypes have been characterized through the expression of different genes in resting situations and as part of adaptive, neuroreparative, neuroprotective or neurotoxic responses.\textsuperscript{78-86} Single-cell transcriptomic and epigenomic analysis is rapidly becoming the method of choice to identify candidate regulators of cell identity and to distinguish the heterogeneity in neuroglial subsets. Such approaches enable developmental processes and disease responses to be studied more accurately.\textsuperscript{79-85}

The most widely used marker of astroglia is GFAP. Its use has shown that there are many cells that clearly express this marker, and there are a number of cells that express it weakly and many that do not. Also, there are cells that contain different GFAP isoforms in both normal and pathological situations that are associated with hypertrophy and/or hyperplasia.\textsuperscript{86-88} Indeed, similar behavior has been observed with vimentin and other astroglia markers.\textsuperscript{80,81} Accordingly, CD44 is an extracellular marker of astrocytes, and CD44\textsuperscript{*} and CD44\textsuperscript{+} cells have been identified in different subtypes of protoplasmic astrocytes, both with short or long extensions, and have been located in different layers of the cerebral cortex, hippocampus, and internal regions of the brain.\textsuperscript{83} Likewise, other astroglia markers related to the metabolism of the neurotransmitter glutamate, such as gluta-mine synthetase or glutamate transporter-1\textsuperscript{74,76,89} are present in different subtypes of normal and reactive astrocytes. These markers can be found in different regions of the CNS, both in relation to normal neuronal circuits and in areas of neurodegeneration in human brains, as well as in AD models. Perhaps the most important revelation in recent years has been the
demonstration of the heterogeneity of astrocytes in relation to their location in the different cortical and hippocampal layers. Superficial, mid-, and deep astrocyte identities were recently identified in a pattern that reflected a gradient across layers that was distinct from that of neurons. Features of these astrocyte layers were established in the early postnatal cortex, and these layers persist in the cortex of adult mice and humans. The presence of astrocyte layers in the adult cortex was confirmed through single-cell RNA sequencing and spatial reconstruction analysis, again establishing that these layers do not correspond to known neuronal layers. Both in rodents and in humans, these glial cells express genes whose functions have yet to be clarified, both in normal situations and during degeneration.

From the outset of studies on astroglia, different hypertrophic changes in some elements (together with cellular hyperplasia in some instances) were evident in response to aggressive agents (lesions, toxins, neurodegenerative processes or during aging), as well as in involutive processes. In a study on gene transcriptome analyses of reactive astrocytes, it was found that 57 genes (among others, C3, GBP2 - guanine nucleotide binding protein 2-, Serping1), were preferentially expressed in LPS-induced A1 reactive astrocytes, and 150 genes (S100a10 - calcium-binding protein A10-, PTX3 - pentraxin-3-, S1Pr3, and others), were preferentially expressed in induced A2 astrocytes. C3 is, in many studies, the most commonly used specific marker for A1 astrocytes, while S100a10 and PTX3 are the most commonly used specific markers for A2 astrocytes.

A1 reactive astrocytes seem to be mainly induced by reactive microglia (through TNF-α, L-1 and complement C1 subtypes). Different direct and indirect studies support this opinion: a) in triple knockout mice (IL-1β −/−, TNFα −/− and C1qa −/−), it has been observed that A1 reactivity decreased after systemic LPS injection, a model for neuroinflammatory research; b) this subtype of reactive astroglia is not generated in mice devoid of microglia (CSFR knockout mice). Another inflammatory cytokine, IL-18, effector cytokine processed by NLRP3 inflammasomes, is a novel described inducer of A1 astrocytes, as well NO and oxidative products. These A1 inducers could downregulate the expression of A2 astrocytes.

A1 astrocytes can be identified in vivo by C3d complement upregulation. Other suggested markers are H2-T23, Fkbp5 and lipp. These neurotoxic astrocytes can enhance local neurodegeneration or, conversely, they may help remove unrecoverable neurons in an attempt to repair neuronal circuits. A1 astrocytes also produce different neurotoxins not well identified until now, D-serine, NO and proinflammatory cytokines (TN-α and so on). These substances, produced in different quantities in the different subtypes of A1 astrocytes, induce neuronal apoptosis and different alterations in oligodendrocytes (including pro-oligodendrocytes) and microglial cells. Different local physiological or pathophysiological processes can reprogram the neuroinflammatory process that has started or that is ongoing, or, on the contrary, to initiate an anti-inflammatory response depending on normality of the CNS (although, unfortunately, it is the rarest possibility). TGF-β and FGFs have been reported to reduce neuroinflammation caused by activated microglia and astrocytes. Further found that TGF-β1 downregulated several genes related to the A1 phenotype. A1 astrocytes can enhance local neurodegeneration or, conversely, they may help remove unrecoverable neurons in an attempt to repair neuronal circuits.

A2 astrocytes are mainly induced by cytokines IL-1β and IL10 (markers of anti-inflammatory M2 phenotype microglia). It is also notable the communication between M2 microglia and A1 astrocytes in antiinflammatory processes because a reduction of A1 toxic molecules was observed. It was recently reported that TGF-β1 and BMP4 of the TGF-β superfamily signaling system modulated the plasticity of reactive astrocytes towards the A2 phenotype. However, the main inducer of A2 astrocytes seems to be a chemokine. Prokineticin 2 (PK2), being its major source, PK2 treatment or overexpression in primary astrocytes or the mouse brain can induce the reactivity of A2 astrocytes. A2 astrocytes increase the production of neurotrophic factors, anti-inflammatory cytokines IL6 and IL10, and thrombospondins. A2 astrocytes are considered key elements in neuroprotection and the induction of synaptogenesis and neurorepair, as well as the main defense against neuroinflammation originated in microglial cells.

Many results of studies on the induction of A1 and A2 astrocytes, as well as the production of neurotropic or neuroprotective substances derived from astrocyte subtypes, give rise to controversies that are difficult to solve. Therefore, it is necessary to intensify research in various fields and models (molecular, cell and animal models) to reach valid conclusions. In a recent review on the different types/subtypes of reactive astrocytes, a consensus (subscribed by 78 researchers) is proposed to define subtypes based on different markers (morphology, location, genetics, RNA expression, neuronal and glial effects) in order to clarify the subtypes of astrocytes and their specific role in glioprotection and gliopathology.

Oligodendroglia, also known as oligodendrocytes or oligodendroglial cells, are cells of neuroepithelial origin for which there is a wide range of morphological subtypes, including those that are related to the formation of myelin and other cell types that accompany neurons in the gray matter (OLs). A recently described subclass of oligodendroglia (or a new class of neuroglia?) was demonstrated to be made up of cells considered to be oligodendroglial progenitor cells (OPCs), pro-oligodendrocytes,
polydendrocytes or NG2+ cells (neuron glia protein 2 positive cells, which is the main marker of these cells) that not only give rise to mature oligodendrocytes and other cells in the CNS, such as astrocytes, but they can also regulate neural, glial, and vascular systems and receive neurotransmitter signals before the final myelinating state. OPCs in the subependymal zone do not receive any synapses but in the grey matter can have glutamatergic and GABAergic synaptic receptors but cells in the white matter seem only to have glutamatergic synapses.

Different subtypes of non-motile myelinating oligodendrocytes (OL) with a distinct stellate morphology, with fine processes in parallel or connected to myelinated internodes, have been described. Other classifications have been based on the number of their enveloping processes of neuronal axons and the characteristics of these axons. Oligodendrocytes undergo many maturation states before reaching their destination and becoming a mature myelinating OL.

OPCs in the brain are produced in three distinct waves (from embryonic to postnatal ages) and from distinct regions (the ventral ventricular germinal zone and the dorsal aspect of the spinal cord), but only OPCs and their progeny from the last two waves persist until adulthood. OPCs from different regional origins have diverse properties (i.e., the dorsally derived OPCs have a higher remyelination capacity). The oligodendrocyte-specific G protein-coupled receptor GPR17 is a cell-intrinsic timer of myelination. The study of OPC properties have revealed that these cells are indeed not a uniform population with equal behaviors or functions. OPCs in different regions show different responsiveness to growth factors or mitogens (OPCs in the white matter, but not in the grey matter, respond to the platelet derived growth factor-PDGF and vary in their capacity to differentiate when transplanted into other CNS areas). However, it remains unclear whether the reported diversity of OPC properties represent subtypes of OPCs with distinct functions, or if they reflect different states of cells with the same function as they progress along their lineage. Very different phenotypes have been described in different studies, but no clear subtypes related with CNS regions, age or pathological states. This cell subtype appears to proliferate and enter damaged areas of the CNS, accumulating to the greatest extent in areas with neurodegenerative lesions. In a recent study, senescent pro-oligodendrocytes were found to associate in large numbers with plaques in both AD models and human brains.

Microglia, also known as microglial cells, include various cell subtypes of mesodermal origin, such as “resident”, “resting” or “quiescent” microglia that originate from macrophages that invade the CNS during brain development and “invasive” microglial cells that invade the developed CNS in response to pathogenic changes in the brain. Macrophages can enter the brain parenchyma through the Virchow–Robin spaces to generate new microglial cells. For many years it has been impossible to differentiate invading myeloid cells from resident microglia. In recent years, the true existence of invasive microglia in neurodegenerative diseases (mainly neurotoxic considered) has been widely questioned. Although it has been shown in stroke, multiple sclerosis and epilepsy, it seems that this does not occur in Alzheimer’s disease. It has been shown that microglial cells associated with amyloid plaques and involuted neurons, although they show reactive monocyte phenotypes, are cells derived from resident microglia and not from new monocytes entering the new AD neuroinflammatory scenario.

There are various morpho-functional subtypes of microglia, with a round morphology (predominantly phagocytic) or with extensions, contributing to the main line of immune defense in the nervous system. Microglia exhibit regional and age-dependent phenotypes, and their coupled proliferation and apoptosis maintain the rapid turnover of microglia in the adult brain, mainly in changes of the normal nervous tissue status. Reu et al, 2017, report that microglia, unlike most other hematopoietic lineages, renew slowly at a median rate of 28% per year, and some microglia last for more than two decades. Microglia population in the human brain is sustained by continuous slow turnover throughout adult life. Microglia are highly reactive cells (many types of “activated”, “reactive” or “hypertrophic” microglial cells have been described). Activated microglia present an increased number of amoeboid forms and decreased arborization in cells with prolongations, and typically are distinguished by overexpression of human leukocyte antigen-antigen D related (HLADR) as well as a group of cluster of differentiation (CD) molecules such as CD40, CD45, and CD68. For many years, reactive microglial cells were considered to exhibit two opposite states of proinflammatory/neurotoxic substance production (proinflammatory cytokines, such as IL-1α, IL-1, IL-6, IL-12, IL-15, IL-17, monocyte chemoattractant protein-1, and TNF-α; reactive oxygen -ROS- and nitrogen -NOS- species) or anti-inflammatory/neuroprotective (anti-inflammatory cytokines, including IL-10, IL-4, transforming growth factor-beta -TGF-growth factors). Thus, two phenotypes were, respectively, distinguished: M1 and M2 were termed. These substances are close related to the induction of diverse reactive astroglial cells (see astroglia section). Two main classes of reactive microglia (M1 and M2) do not develop in clearly specific brain regions or disease phases. Indeed, markers for both populations can be either up- or downregulated in many different areas in the brain in association with a variety of neurodegenerative diseases. New technologies (including two-photon imaging, whole-genome transcriptomic and epigenomic analysis with complementary bioinformatics, unbiased proteomics, cytometry by time of flight (CyTOF; Fluidigm) cytometry) need to be used to define different subtypes of microglia reactive cells. A comprehensive gene expression encyclopedia of glia cells exists to help researchers.
Ransohoff (2016)\textsuperscript{151} raises an important question \textit{(`A polarizing question: do M1 and M2 microglia exist?)}, considering that this microglial polarization has not been established by research findings, being adopted this nomenclature in an attempt to simplify data interpretation at a time when the ontogeny and functional significance of microglia had not yet been characterized.

Microglial cells can communicate with other cell types by releasing soluble factors as well as exchanging active molecules an RNA through secreted extracellular vesicles (EV)/exosomes, releasing soluble factors as well as exchanging active molecules characterized.

Microglial cell reactivity is critical in neuroinflammation and is closely related to most of the neuropathological cascades that lead to dementia.\textsuperscript{138-140,158-160} However, microglial dystrophy has been correlated with the presence of neurofilibrillary degeneration in situ, suggesting that neurodegeneration is secondary to age-related microglial deterioration.\textsuperscript{161} Microglial dystrophy has been detected in the human brain,\textsuperscript{129,162} and the term “dystrophy” is used to refer to different morphological abnormalities that affect cytoplasmic microglial extensions, such as spheroid swellings, demarcations, beaded or tortuous processes, and—most conspicuously—fragmented processes. This latter phenomenon probably represents the most advanced stage of cytoplasmic deterioration that affects microglia, and the term “cytorrhexis” has been proposed to describe specific microglial cytoplasmic fragmentation.\textsuperscript{129}

**Neuroglial Responses in AD**

Astrogliosis and microgliosis are key aspects in most theories regarding the pathogenic “cascades” leading to AD,\textsuperscript{3,24-28,31,33-35,138-143,150-153,158-161} although other changes in astroglial, oligodendroglial and microglial cells have also been described\textsuperscript{181,191,192,123,126,159,163-165} but have not been thoughtfully considered in the neuropathological theories on AD.

Most neuroglial studies have been carried out using only a limited number of specific markers for certain types or subtypes of neuroglial cells (normal and/or reactive). The results of these studies have been considered useful for a sufficient definition of the neuroglial typology of the region/area studied in the brain. However, many astroglial cells are not GFAP- or vimentin-immunopositive, and many microglial cells are not IBA-1 or LN-3-immunopositive. As an example, Figure 1 shows a high density of neuroglial nuclei (Congo red in this case) to visualize cells. If the density of different types and subtypes of neuroglial cells (astrocytes, oligodendroglial cells, including NG2+ cells, and microglia) is studied in parallel sections with the most commonly used conventional markers in research, the sum of the partial results obtained from each neuroglial type/subtype is always lower (17-32% in our studies, pending of publication) than the density of neuroglial cell nuclei. An important number of neuroglial cells of specific phenotypes will go unnoticed.

It is true that there are different brain areas where important conventional astrogliosis and/or microgliosis processes can be observed (see later). However, there are also areas where there is a reduction in astrogliosis or microgliosis (Figure 2) or an abnormal manifestation of astrogial cells presenting unusual markers (such as amyloid in astrocytes; (Figure 3) that are not commonly considered. New techniques are indispensable for studying well-characterized glial cells in different regions/areas of the brain.\textsuperscript{79-85}

The main relevance of neuroinflammation in AD pathogenesis is indisputably accepted. In this sense, the differential involvement of neuroinflammatory molecules, mainly released by microglial cells during the development of the disease, may contribute to the modulation of characteristics and the severity of the neuropathological changes, driving, in part, AD phenotypic diversity.\textsuperscript{58} Amyloid production and deposition are closely related to neuroinflammatory reactions of astroglial and microglial cells. Moreover, astrocytes and microglial cells (in different transitional states) seem to have an important involvement in tau production, degradation, processing and propagation.\textsuperscript{166-168} However, we still do not have a clear idea of the different toxic or neuroprotective mechanisms that actually occur. The ability to interpret the effects of all these neuroglial changes is hindered by the fact that the exact role of these glial cells in AD is not fully understood. Indeed, most neuroglial alterations, such as gliosis, are also considered processes inherent to normal CNS aging, and they have been associated with other neurodegenerative diseases. Therefore, many mysteries remain regarding the influence of glial cells on the pathogenesis of AD.\textsuperscript{3,23-26,153,158} Indeed, it is still unclear

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**Figure 1.** Brodmann’s area 46 of a brain from an AD case, Braak and Braak V. Layer IV-V. Zone of high density of glial cells scattered in the cortical parenchyma and not related to amyloid plaques. Congo red stain / confocal observation with green filter. A large number of neuroglial nuclei (from astrocytes, oligodendrocytes, and microglial cells) is observed. The technique does not allow to differentiate the different types or subtypes of neuroglial cells, but it demonstrates the high density of neuroglial cells that are present in this region of the CNS altered by AD pathology. Bar = 200 microns
whether glial modifications or their reactive states fulfill a primary or secondary role in the “cascades” or events that drive neurodegeneration.

Importantly, there is no single relationship between the different types of glial cells and neurons affected by accumulations of aberrant proteins (phospho-tau, synuclein) (Figures 4A, 4B) or the amyloid plaques (Figures 5, 6 and 7). In fact, different types of amyloid plaques are not associated with specific patterns of glial accompaniment. The number of astroglial cells closed involved varies between 2 and 15 (Toledano and colleagues, unpublished data) (Figures 5, 6 and 7), with neighboring similar plaques also displaying considerable variability (this was observed in all regions of the brain, regardless of the number of astrocytes in each studied area). Many plaques are devoid of astroglial accompaniment, in special in areas of low astroglial density (astroglial involution?). Microglia seem to be more consistent in their relationship with amyloid plaques, as microglia are evident with more than 75% of all plaque types, both in their mass and in the periphery.
Glial alterations must always be considered when attempting to understand the pathogenesis of neurodegenerative diseases, especially AD, and when attempting to develop successful therapeutic strategies. In fact, glial cells are currently considered to represent a promising target to establish effective therapies for AD, mainly because other therapies targeting neurons have failed to produce promising results.

**The special features of neuroglial cells in human AD**

**Astrogial responses.** There is some debate as to whether astrocytes display any special features in AD. Unusual expression of GFAP isoforms has been described, and specific patterns of astroglial reactivity have been proposed to be closely associated with certain pathological AD lesions or specific areas of amyloidogenesis, but this is a debatable matter (Figures 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, and 11).

“Dramatic” and “generalized” astrogliosis has been described in AD, and a wide number of hypertrophic/GFAP-hyperimmunoreactive astrocytes have been described in many regions of the brains of AD subjects. Moreover, special patterns of astrogliosis have been observed in some specific CNS regions, but do not always occur. It has been proposed that cortical...
Astrogliosis in AD follows an obvious laminar pattern, with a heavy band evident in layers I-III and another band present in layer V (Figures 2, 3, and 4), even though this pattern has not been reported universally. Reactive astrocytes have been reported close to amyloid (simple or complex) plaques in the hippocampus (Figure 2). Normal aging is sometimes associated with considerable cortical gliosis but does not follow any specific pattern. Studies of astroglial markers show a complex age-dependent remodeling of these cells in different brain regions, although in most AD cases, the hypertrophy of immunoreactive astrocytes in the gray and white matter is heterogeneous (in some cases, hypertrophy is more pronounced at the interface between the two types of matter). Subcortical gray matter astrogliosis is commonly observed in both normal aging and AD, although “variable” and “disorganized astrogliosis” is more often observed in AD, even in areas where astrogial cells are lost to a large extent (Figure 6). Such areas have been considered zones in which senescence, atrophy or dystrophy of these cells occurs, provoking neuronal pathologies; the maintenance of normal astrocytes is necessary for the normal activity of neurons and their adaptive changes.

Several studies suggest that the number of astroglial cells in many regions of the CNS remains (more or less) constant throughout life, irrespective of the evolution of pathological processes such as AD. Phenotypic changes but not proliferation have been proposed as glial responses to AD. We recently found that astrogliosis (defined by the distribution of GFAP) in the brains of 65- and 85-year-old AD patients (with a disease course of 10-15 years) was less pronounced than that observed in normal 85-year-old individuals. Astrogliosis in AD patients seems to be quite variable in different brain regions and reflects the state of involution in each brain. Astroglial GFAP-immunopositive cells and immunonegative hyperplasia (documented by the increase in astroglial nuclei and the absence of microglia) are the sum of both general and local reactions to neural changes; these changes reflect the status of the environment. In neurodegenerative processes, neuronal and astrocyte death, as well as some hypertrophic surviving astrocytes, seem to compensate for the loss of astroglial subtypes, as demonstrated in the cerebellum or in the molecular layer of the hippocampus.
Despite these considerations, astrogliosis is thought to be associated with neuropathological alterations in AD, and changes in the number of astrocytes are associated with amyloid plaques in several studies. The significance of reactive astrogliosis around these pathological structures is unclear (noxious or protective), and different plaque-astroglial cell relationships have been observed: plaques with a scar-like crown of astroglia, plaques with isolated peripheral astrocytes, plaques infiltrated by astroglial processes, plaques not associated with astroglia, etc. A generally weaker astrocyte response to β-amyloid (Ab) plaques has been associated with cognitive impairment (perhaps a loss of defense responses), and reactive astrogliosis in the neuropil of affected areas is considered to be harmful. In a recent study, the highest neurotoxic plaques were related to reduced contact with astrocytes. In terms of neurofibrillary tangles, an association between neurons with tangles and glial cells was observed. Indeed, there appears to be a clear relationship between such neurons and reactive astrocytes, although different types of glial-neuron relationships have been described.

We present a series of images showing different reactive astrocytes (GFAP immunopositive) related to neuropathological alterations (or possible foci of alterations) in AD subjects which are not generally considered in current studies. In Figure 4A, a zone of high density of immunopositive GFAP astrocytes is shown, mainly associated with dystrophic neurons and not associated with amyloid plaques. In some areas of this brain region, diffuse involutive hypertrophic astrocytes are evident that undergo a process of astrogliosis (degeneration of the cell body and its processes) (Figure 4B). This process could be a transition stage towards astrocyte involution. In some other areas where there is no marked AD neuropathology, foci of astroglial hypertrophy (Figures 8 and 9) similar to those that appear in advanced AD brains (included areas of astroglial involution – Figure 6) can be seen and may be indicative of a zone of degenerative onset.

Astrocytes that accumulate amyloid or aberrant tau protein deposits have often been found in human AD patients (Figure 3), in experimental AD models and in animals that present AD-like amyloidosis (monkeys and simians). Moreover, astroglial cells may generate amyloids, raising the possibility that astrocytes participate in the generation of amyloid that leads to AD. Extracellular vesicles from 3xTgAD mouse and AD patient astrocytes have been shown to be transporting elements of substances that cause damage to neurons, glial cells and endothelial cells, aggravating AD. Hypertrophic astrocytes with abnormal gene expression, such as calretinin, have also been described. Indeed, there is an increase in the GFAP 1 isoform, one of the nine splice variants of GFAP described in astrocytes from different species, as AD progresses. Complex and region-specific changes to other astroglial markers (glutamine synthetase, S100β) have also been detected in both the aging brain and in AD. Thus, the diverse astrogliial responses in the brains of AD patients could reflect the multifactorial nature of this disease (a systemic disease that alters glial responses in different ways), although these responses may also reflect the particular features of the distinct CNS regions.

**Oligodendroglial responses.** Alterations to oligodendrocytes have rarely been studied in AD, although demyelination is a secondary characteristic feature of this disease. Demyelination assessment using new brain imaging techniques is widely used for in vivo diagnosis, although demyelination is mainly considered a secondary pathological event in AD. As such, more studies are needed to assess whether demyelination is a primary pathological process in AD. Metabolic changes in AD oligodendrocytes have been observed, such as alterations in glycolytic and ketolytic gene expression.

In recent years, much interest has been generated by NG2+ cells. These cells seem to be modified in AD subjects, even though the significance of these alterations (neuroprotective or neurodegenerative) is not completely clear.

**Microglial responses.** There is an intimate relationship between microglia and neurons at the synaptic level. Microglia modulate activity-dependent functional and structural plasticity indispensable to normal synaptic function and cognition. Alterations in microglia-synapse interactions are key for AD presentation and progression.

Different reactive microglia have been described in association with neuropathological AD lesions, whereas less prominent differences in these cells are associated with normal senility or with other neurodegenerative diseases. Microglial cells with abundant extensions, scattered though the parenchyma, have been observed in different regions of brains affected of AD (Figures 12 and 13). Different morphologies have also been described in other areas of these brains. The forms with extensions also invade some subtypes of amyloid plaques and the round forms are close the plaques as well diffusely distributed in the brain parenchyma (Figures 12, 13, and 14). Different microglial phenotypes have been described, but no clear microglial responses have been associated to specific alterations in specific areas of the brain or in specific phases of the progression of AD.

A decrease in ramified (considered healthy) microglia was recently described in subjects with advanced AD, with imaging analysis demonstrating a reduction in the arborized area and skeletal complexity. It was concluded that activated microglia were not associated with AD but that they were increased in nondemented controls with a stronger AD-type pathology. Moreover, the authors considered that microglial clusters were only occasionally associated with Ab- and tau-positive plaques but that these elements represented less than 2% of the total microglial population. We have shown that the number of microglial cells in the cerebellum and hippocampus is more closely related to the age of the individuals than to their AD pathology.
Neuroinflammation is currently thought to be one of the main mechanisms driving the pathogenesis of AD, although this process can also explain degenerative changes in aging and in other neurodegenerative processes. Microgliosis could protect against neuronal degenerative changes (via phagocytosis of damaged neurons or amyloid plaques or via the production of neuroprotective agents, although a secondary effect might provoke alterations through an excess of neurotoxins in the neural microenvironment (e.g., chemokines and cytokines or inducers of oxidative stress). A large increase in both resident and reactive microglial cells is often observed in the parenchyma, as well as around the vessels in AD brains (Fig 12). Microglial activation in AD appears to be Aβ-dependent, with Aβ binding to receptors such as RAGE, scavenger receptors and toll-like receptors (TLR2, TLR4 and TLR6) representing an important cause of microglial activation in mouse models of AD. However, clear patterns of microgliosis are not generally evident in AD, although a higher density of microglial cells in subpial zones, as well as at the transition between the neuronal and molecular layers of the hippocampus and cerebellum, has often been seen. Many authors consider that microglial cells accumulate in areas more strongly associated with amyloid deposits, even infiltrating amyloid plaques, although they also appear in less severely affected areas. In some models of AD, microgliosis is observed before the onset of AD pathology, which is why inflammation/microgliosis has been considered the origin of AD in some theories. Significantly, Aβ immunotherapy seems to downregulate microglial activation and reduce the inflammation-mediated component of AD.

Despite microglial reactions, the genotype of microglial cells in different individuals is closely related to AD progression. Microglial TREM2 (Triggerin Receptor Expressed on Myeloid cells 2) facilitates adaptive regulation of amyloid plaque formation; as amyloid fibrils become compacted into plaques, adaptive regulation reduces the local induction of neurodegeneration due to the TREM2 activity of healthy microglial cells. However, other TREM2 variants, such as the R47H variant, are associated not only with a higher risk of AD but also with earlier symptom onset and accelerated dementia. Other single-nucleotide polymorphisms in genes that are exclusively or largely expressed in microglia, including CD33, CR1, ABCA7 and SHIP1, are associated with an increased AD risk. Human and mouse single-nucleus transcriptomics revealed TREMP2-dependent and TREMP2-independent...
cellular responses in AD,\textsuperscript{189} and TREMP2 haploinsufficiency impairs microglial barrier function, decreasing amyloid compaction.\textsuperscript{207} In this sense, colony-stimulating factor 1 receptor signaling seems to be necessary for microglial viability.\textsuperscript{207}

Microglial dystrophy has been demonstrated to be correlated with the presence of neurofibrillary degeneration, suggesting that neurodegeneration is secondary to aging-related microglial deterioration.\textsuperscript{164,165} Dystrophic microglia are also associated with different neurodegenerative diseases.\textsuperscript{208} Moreover, “aged microglia” seem to affect synaptic function.\textsuperscript{209} The inability of microglia to remove amyloid deposits has been considered a cause of “microglial exhaustion”, which in turn promotes neurofibrillar neurodegeneration, brain failure and dementia.\textsuperscript{161}

It has been suggested that the inability of microglia to remove aggregated amyloid causes microglial exhaustion and thus exacerbates already ongoing age-dependent microglial deterioration.\textsuperscript{209,210} The eventual total loss of functional microglia in advanced AD could promote widespread NFT’s, dementia and brain failure. Microglial dystrophy is probably caused by oxidative stress, and in this sense, it can be considered pivotal in this disease.\textsuperscript{138,161,162}

In summary, there is structural evidence of microglial heterogeneity in human AD from light microscopy and transmission and scanning electron microscopy in AD models in association with amyloid and tau pathology,\textsuperscript{130,198} as well as molecular evidence of the production of a large variety of neurotoxic substances. The sequence of appearance of the microglial reaction and that of the disappearance of normal microglia in human AD has been discussed in various studies.\textsuperscript{130,162} Certain studies show microglial alterations in nonsymptomatic or early AD phases and others show microglial alterations in advanced phases. The loss of healthy microglia has also been described only in severely affected regions of AD brains.\textsuperscript{130}

Chronically activated microglia secrete proinflammatory cytokines related to the induction and/or progression of AD, but the state transition from a resting state to an activated state and the exact meaning of each phase (depending on the panel of cytokines/chemokines secreted) remain unclear.\textsuperscript{210} Natural beneficial effects can occur, or therapeutically induced effects can be produced in this state or during this period of transition, but careful research is necessary.

\textit{Induced glial responses in AD models}

In some experimental models of AD, such as those involving mechanical, anoxic or toxic damage to the cortical regions involved in cognitive functions (e.g., the entorhinal cortex),\textsuperscript{199} significant changes in resident neuroglial cells are induced (astroglia and microglia), both in local lesion areas and in areas innervated by injured neurons (e.g., the hippocampus).\textsuperscript{211,212} Similarly, damage to cholinergic cells in the \textit{nucleus basalis magnocellularis (nbm)}\textsuperscript{211} of 4-month-old rats causes transient changes in “proximal” areas, e.g., nondamaged structures neighboring the nbm that are not innervated by this nucleus but that maintain a vascular relationship with it; this damage also causes substantial and permanent changes in the ipsilateral cortex to which it is directly connected synaptically (layers I-V of the motor and somatosensory cortical regions). Moreover, the indirectly connected contralateral cortex displays long-term reactive astrogliosis, a cortical alteration that persists for relatively long periods (13-20 months). In contrast, the proximal response lasts from 1 day to 13 months, and tends to disappear thereafter. Tightly interwoven subsets of astrocytes with distinct GFAP immunoreactivity have been observed, while nbm lesions in 20-month-old animals produce similar but weaker patterns of glial reactivity, in addition to glial reactivity related to old age. The maintenance of reactive astrocytes for many months after the occurrence of a lesion suggests an influence of factors other than those produced by nbm neurons that were initially damaged. It is possible that similar reactive astrocytes in humans could promote AD-related neurodegeneration and that nbm cholinergic involution might provoke cortical involution by inducing reactive astrocytosis and/or microgliosis.

Reactive processes of different sets of neuroglial cells have been described in different transgenic mouse models of AD. In some cases, such glial alterations have been described early in life and prior to the appearance of any neuropathological hallmarks of AD, while others develop after the appearance of aberrant deposits of amyloid or tau protein\textsuperscript{162,163} (Figure 15). Enrichment of the neurodegenerative signature in microglia has been observed in AD models.\textsuperscript{213}

\begin{center}
\textbf{Figure 15.} Electron microscopy image of an amyloid plaque of the frontoparietal cortex (layer V) in a transgenic model of AD (APP + PS1, to which two human genes have been inserted - Amyloid Precursor Protein and Pre-Seniline 1- that induce AD of family type). In the center of the image, an amyloid plaque is observed, with a dense amyloid “core” and less electrodense radial amyloid extensions. The boundaries of a hypertrophic astroglial cell are marked in red, and the extensions of a microglial cell associated to the amyloid plaque are marked in blue. In black, hypertrophic neurites filled with vesicular forms indicative of degeneration of dendrites and axons of affected neurons are delimited. The almost “normal” appearance neuropil shows small alterations compared to control mice (increase in diameter and alterations of subcellular structures in dendrites and axons; synaptic alterations; varicosities in neuroglia extensions). Bar = 25 microns
\end{center}
How can neuroglial alterations be interpreted in human AD tissue and AD models?

Morphological and functional changes to neuroglia are always evident in the brains of humans suffering from AD, as well as in the brains of animal models of AD, although these changes are quite variable in nature (Figure 15). These neuroglial variations can be interpreted in different manners. It may be that the main controversy, when one wants to interpret the role of astroglia and microglia associated with the neuropathological manifestations of AD, is whether the associated glial reactions tend to eliminate aberrant protein accumulations and/or to separate them from the rest of the tissue or, on the contrary, if these cellular elements promote further development of neuropathology. In all the studies on this subject, and in all the reviews on it, this controversy is always considered by all authors, and an undisputed conclusion is never reached. Neuroprotective and neurotoxic glial reactions seem to coexist. We still do not have markers (or a set of markers) that can be defined, with absolute certainty, regarding the role of each of the neuroglial cells present in certain regions of the CNS in each phase of a disease. Perhaps the most important justifications of the involvement and the simultaneous or exclusive neurotoxic and/or neurodegenerative effects of neuroglial cells in AD are indicated below.

1) Differences in the response of different neuroglial families or subtypes.

The plasticity of glial cells, particularly in terms of reactive gliosis, includes modifications to the structure of glial processes, changes in cell motility and—more importantly—modifications in the production of diverse neuroprotective (neurotrophic factors, gliotransmitters, etc.) and neurotoxic substances (cytokines, chemokines, free radicals, prostaglandins, NO or other neurotoxins). The expression of different GFAP isoforms in astrocytes, as well as the different reactive forms of these normal neuroglial cells (or subsets of astroglia) must also be kept in mind. Chronically activated microglia secrete proinflammatory cytokines related to the induction and/or progression of AD, but the state transition from a resting state to an activated state and the exact meaning of each phase (depending on the panel of cytokines/chemokines secreted) remain unclear. These changes alter the concentrations of local factors, which could in turn induce neuropathological changes in small areas that may hinder the primary neuroprotective response driven by neuroglia. Neurotoxic effects are commonly described, but natural beneficial effects could occur. Moreover, therapeutically induced effects could be produced in these state transitions. Careful research is necessary both to correctly interpret all neuroglial changes and to develop protective or corrective glial therapies.

2) Differences in pathological neuronal degenerative changes in AD.

An important number of studies on neurodegeneration in AD focus only on neuronal tau-dependent neurofibrillary tangles and dense core of amyloid plaques. However, other well-documented pathological alterations, such as the existence of different types of dystrophic neurites, other aberrant protein aggregates in neurons and different types of amyloid deposits (diffuse or focused amyloids, which forms various types of plaques), are overlooked. All these alterations are related to neuroglial changes of varying intensity and make different contributions to the development of the disease. The alterations occur differently in each region/area of the brain parenchyma depending on the characteristics of the initial cellular or molecular changes.

AD is a multifactorial syndrome, both in its genesis and in the consequences of its different pathological modifications. In this line of thought, Fiala (2007) pointed out several hypotheses that try to explain the local production of amyloids and the pathogenesis of the plaques: 1) a vascular (extracerebral) origin of amyloids leading to perivascular deposits and synaptic/dendritic amyloid lesions; 2) a glial origin, inducing neurite deposition and synaptic/dendritic amyloid lesions; 3) neuronal secretion, glial activation and glial neurotoxicity; 4) neuronal secretion, glia-induced amyloid "fibrillation" in the neuropil, and synaptic/dendritic amyloid lesion; 5) Abeta release by neuronal lysis, deposit production and activation of microglia, and glial toxicity; and 6) dystrophic axon lysis (including amyloid spread), extracellular amyloidosis, glial reaction, neuronal and glial toxicity. Likewise, there are various hypotheses that assume that the hyperphosphorylated tau protein and other aberrant proteins give rise to neurite dystrophy and neuronal dysfunction or death. Astrocytes and microglial cells (in different transitional states) seem to have an important involvement in tau production, degradation, processing and propagation, but we still do not have a clear idea of the different toxic or neuroprotective mechanisms that actually occur.

Expression and secretion of ApoE isoforms in astrocytes and microglial cells during neuroinflammation processes are of special importance in neurodegeneration. ApoE, the main member of the apolipoprotein family in CNS, is a protein involved in a wide variety of functions, including lipid transport, neuromodulation, neuronal plasticity, neuronal repair, neurite outgrowth and regulation of Aβ formation and clearance. ApoE also modulates the inflammatory response of microglia and astrocytes, the cells that secrete the greatest amounts of this apolipoprotein. Three major isoforms, apoE2, apoE3, and apoE4, encoded by the ε2, ε3, and ε4 alleles, exits in humans. These isoforms have different abilities to carry out the assigned functions in the CNS. ApoE4 not only increases an individual’s risk for AD, but also for the outcome from neurological injuries with dramatic brain inflammation. Conversely, ApoE2 and 3 seem to be protective factors, increasing anti-inflammatory actions. ApoE4 genotype was associated with lower levels of secreted apoE from astrocytes and microglia, as well as higher levels of the larger apoE species that
remained inside microglia. In neurons, apo 2 and 3, and apoE4 have different intracellular trafficking profiles: apoE4 is retained in the endoplasmic reticulum (ER) and Golgi apparatus, causing functional disturbances. In astrocytes, apoE4 causes ER stress. In studies carried out on cell cultures of astrocytic microglial cells that express different apo2, 3 or 4 isoforms, it has been observed that APOE2 astrocytes and microglia secreted three to five times more apoE than APOE4 cells, with APOE3 cells intermediate, supporting the hypothesis that APOE4 predisposes to greater inflammation in primary cells. The presence of apoE4 is associated with overactive proinflammatory phenotypes such as elevated NO, TNFα, and IL-6. However, expression of astrocitic apoE3 decreased the levels of IL-6 and IL-1β. The studies of Liu et al., 2017 conclude that expression of apoE4 during the initial seeding stage of AD is sufficient to drive amyloid pathology and plaque-associated neuritic dystrophy, while the presence of apoE4 after the initial seeding stage has minimal impact of amyloid pathology, highlighting the importance of early alterations of apoE in AD.

The above mentioned pathological alterations (tau and amyloid related alterations, apo 4 dysfunctions) may be the origin of the varying neuronal changes and development of AD and should be considered when trying to interpret the role of neuroglia in specific regions/areas in the study of each brain. This complex neurodegenerative process considered here involves the accumulation of products in the parenchyma, the astroglial and neuronal secretion of amyloids, amyloid fibrillation by microglia, neuronal lysis and of dystrophic axons, etc. Different compounds can be demonstrated in the plaques. Reactive subtypes of microglia and astroglia are involved in all these pathological processes, although their roles seem to differ in terms of the formation and development of the distinct types of plaque. Different subtypes of neuroglial cells are expected to be observed. Plaques do not grow indiscriminately because neuroglial cells regulate plaque growth through the phagocytosis of amyloid deposits. “Burn plaques” are often thought to reflect the plaque lysis produced by glial cells. Reactive astrocytes located in close proximity to either diffuse or compact plaques may exert a neuroprotective role in the aging brain, although the astroglial response to Aβ plaques is associated with cognitive impairment by many authors.

3) Differences in the involvment of the functional glioneurovascular units.

In a focal area or in a larger area spanning different regions of the brain, involutive processes can affect cellular elements in close morphofunctional relationships (neurons, glial cells and vascular structures) that configure the basic trophic and functional units to maintain the elements of the neural circuits. Vascular risk factors can result in dysregulation of the neurovascular units producing hypoxia and altered transport to the blood (including clearance of amyloid) as well as neuronal degeneration and/or neuroglial toxic responses inducing parenchymal and vascular accumulation of Aβ. The set of toxic factors produced by the elements of the neuroglivascular units leads to an acceleration of the neurodegenerative process. For this reason, it is possible to observe areas with very different alterations in cellular elements. Neurovascular unit dysfunction is a main inducer of AD.

4) Similarities and differences in the distinct neuroglial responses in aging, AD and other neurodegenerative diseases.

Enhanced astrogliosis and microgliosis in association with aging, AD and non-AD neurodegenerative diseases have been described. In conjunction with astrocyte dysfunction in “senescence” and dystrophy, impaired microglial functionality has been demonstrated in “senescence”, asthenia, and dystrophy, affecting microglial motility, proteostasis, phagocytosis and cell signaling. The number of astrocytes has often been inversely correlated with synaptic density but not in all disorders. In AD, this inverse relationship seems to occur in all cortical brain regions, yet it was only evident in the frontal pole in association with frontal lobe degeneration. The number of astrocytes appears to be maintained throughout life, even in patients suffering from AD. In contrast, microglial density appears to increase with neurodegeneration, although the increase in microglial density associated with physiological aging is accentuated in healthy individuals over 75 years of age, exceeding that observed in individuals with AD. Moreover, microglial density seems to depend on the years of disease evolution and not on the patient’s age. Indeed, it was proposed that phenotypic changes underlie most glial responses but not glial proliferation.

Agonal events may be responsible for atrophic/senescent subtypes of neuroglial cells, mainly due to changes in pH. This possibility should be investigated in studies of human brains.

Conclusions regarding glial alterations in AD and future therapeutic perspectives

In summary, there are several aspects of neuroglia that may be particularly important to understand the pathology of AD and to develop preventive therapies.

As research progresses on the characteristics and functions of the different types of neuroglial cells, which include wide diversities of morphofunctional and gene reactions as well as involvements of various subsets of these cells (mainly focused in astrogial and microglial cells), greater new possibilities for the involvement of neuroglial cells are found both in the maintenance of brain functions in adulthood, aging and in neurodegenerative diseases, as well as in the triggering and progress of neurodegenerative processes. New mechanisms of neuroprotection/neuroreparation, as well as of neurodegeneration are continually being described in the scientific literature, with therapeutic possibilities.
(developed in the second part of this monography). As conclusions we want to highlight:

1) In the brains of individuals who suffer from AD, alterations of neuroglial cells are consistently observed in different regions of the brain, but the types of alterations observed are highly variable, both in the morphological and functional aspects, both in their neuroprotective/neuroreparative and neurotoxic nature, both in their presence in large areas and in small regions or as cells of the same subtype but with different phenotype closely intermingled. As above mentioned, it is true that there are different brain areas where important conventional astroglial and/or microglial processes can be observed (aspect that is considered in a large number of publications as a specific or characteristic "marker" of the underlying neurodegeneration in AD). However, there are also areas where there is a reduction in astrogliosis or microgliosis. In addition, as it has been shown in this monograph, there are significant differences in the relationships of neuroglial cells with amyloid plaques and altered neurons. On the other hand, insufficient research has been done on A2 and M2 anti-inflammatory cells in human AD (which may shed light on the neuropathological pathways of AD and/or define new therapeutic targets). All types of glial cells seem to be affected: astrocytes can present both astrogliosis (hypertrophy/hyperactivity and/or hyperplasia) (of A1 or A2 phenotypes) and involuion (morphological and functional); microglial cells exhibit proinflammatory changes to subsets of resident and newly (?) incorporated cells as well as anti-inflammatory changes; and oligodendrocytes demonstrate a loss of cellular elements, demyelination, and a decrease in the number and function of NG2+ cells (although these cells could proliferate and try to recover the damaged neurons). All these changes can drive the manifestation of symptoms and the variable progression of AD, while many of these changes may appear in physiological senility, they are much more marked in AD.

2) Complex relationships between the various morpho-functional forms (normal, reactive, dystrophic, "senescent" – these currently valued for their expression of specific macromolecules\(^ {240-242} \)) of the main neuroglial types accompanying neurons (both normal and dystrophic) are working during all life, both in normal or abnormal circumstances (Figure 16). Neuroglia cells maintain close interrelationships ("crosstalk of glial cells")\(^ {209,243} \) offering a modulate response of the entire neuronal group close in contact with neurons, both normal and dystrophic, in specific areas. Neuroactive neuroglia glial substances (cytokines, chemokines, prostaglandins, NO, free radicals, …), can finally produce (directly or in-directly) neurotoxicity/neuronal involuion or stimulate neuronal recovery. In the first case, the first action could tend to eliminate neurotoxic neurons to improve homeostasis, but in the case of AD can spread neurodegeneration in degenerating areas.

3) The production of amyloids, amyloid deposition in the parenchyma of nervous tissue and in the wall of blood vessels, and the accumulation of phosphorylated tau and other proteins in the soma (tangles) as well as in the axons and dendrites of neurons (dystrophic neurites) produces large and complex reactive responses in all neuroglial cells. In contrast, alterations to reactive neuroglial cells induce the formation of amyloid and aberrant intraneuronal deposits, consequently inducing morphological and functional degeneration of neurons. The neuronal or neuroglial process affected can lead to the establishment of a neurodegenerative disease; together, alterations to the cells and the affected processes contribute to disease progression.

4) The reactivity of astrogia, oligodendroglia and microglia to some extent represents mechanisms that initially seem to counteract the damaging changes in the neural milieu. These changes apparently attempt to correct neural dysfunction and neuronal circuits or eliminate the neurons that induce toxicity or that are undergoing involuion/death.

5) The development of AD can be studied in experimental models from very early stages of life, especially in transgenic mice that express Ab and phosphorylated tau (which are deposited in the nerve parenchyma—amyloid—and neurons—tau protein—in the human brain). However, these studies have not provided much information regarding the pathogenesis of AD in humans, although they have enabled phenotypic variations in neurons and glial cells to be defined during the course of the disease and in aging. Many of these studies highlighted the neuropathological alterations and the differences in cognitive performance of these animals, such that the extrapolation of these results must be analyzed carefully.\(^ {36,160,161} \)

6) Novel techniques that combine cellular and molecular approaches\(^ {79-84} \) will reveal new insights to improve the present understanding of the genetic changes to cells that accompany the changes in neurons affected by neurodegenerative processes. Thus, it should be possible to determine what causes neurodegenerative involuion and disease progression and to identify new therapeutic targets.

As a conclusion, it should be noted that neuroglial cells are a) fully involved in the neuropathology of AD and b) that the
different types and subtypes of these cells can be both neuroprotective/neuroreparative or neurotoxic, either simultaneously or consecutively depending on the subset of cells. Future research should aim to elucidate the true role of each cell subtype and the possible transitions from their normal phenotype to a reactive state. This research should focus on each phase in the evolution of AD and develop specific strategies for each situation. Some of the specific objectives should be to determine the specific characteristics of astroglial, oligodendrogial and microglial cells from each specific region/area of normal, aged and AD brains in each phase of the disease; the neurotoxic or neuroprotective roles that these cells play; the role of each subtype in the formation of amyloid deposits (diffuse amyloid or plaques) or, conversely, in the clearance of amyloid; the effect that these cells have on the blood-brain barrier; the effects that these cells have on neuronal function; the factors that induce responses from each of these cell types; and the intracellular communication pathways that drive the phenotypic changes characteristic of each cell subtype. These issues remain largely unresolved but should be clarified through extensive research into the behavior of neuroglia.

As above mentioned in the Introduction, in the second part of this monograph (“The relationships between neuroglial alterations and neuronal changes in Alzheimer’s Disease, and the existing controversies. II Gliotherapy and multimodal AD therapy”) we will study the therapeutic possibilities that are being analyzed to try to prevent or palliate the onset or progression of this disease, based on treatments aimed at maintaining the normal functions of neuroglial cells or normalizing the alterations that occur in the progression of AD.

Author contributions
All the authors have been carried out jointly the research on AD and have written this monography.

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REFERENCE
1. Alzheimer A. Über eine eigenartige Erkrankung der Hirnrinde, Allg. Zschr Psychiat. 1907;10:136-148.
2. WHO. World report on aging and health. Geneva: WHO; 2015. Available at https://www.who.int/ageing/publications/world-report-2015/en/2015
3. Toledano A, Merino JJ, Rodríguez JJ. Neuroglia in Alzheimer’s Disease: From cohort to contestant in the disease progression and its therapy. Cur Alzheimer Res. 2016;13:318-320.
4. Armstrong RA. What causes Alzheimer’s disease? Folia Neuropathol. 2013;5:169-188.
5. He Y, Chen Z, Gong G, Evans A. Neuronal networks in Alzheimer’s disease. Neuroscientist. 2009;15:333-350.
6. Schmitt HP. Neuro-modulation, aminergic neuro-disinhibition and neuro-degeneration. Draft of a comprehensive theory for Alzheimer disease. Med Hypotheses. 2005;65:1106-1119.
7. Robertson JM. The Gliocentric Brain. Int J Mol Sci. 2018;19:3033. doi:10.3390/ijms19103033.
8. Perry G, Rizzuto N, Autilio-Gambetti L, Gambetti P. Paired helical filaments from Alzheimer disease patients contain cytoskeletal components. Proc Natl Acad Sci U S A. 1985;82:3916-3920.
9. Chan-Palay V, Lang W, Haesler U, Köhler C, Yasargil G. Distribution of altered hippocampal neurons and axons immunoreactive with antisera against neuropeptide Y in Alzheimer’s-type dementia. J Comp Neurol. 1986;248:376-394.
10. Miranda AM, Ashok A, Chan RB, et al. Effects of APOE4 allelic dosage on lipidomic signatures in the entorhinal cortex of aged mice. *Transl Psychiatry*. 2022;12:129. doi:10.1038/s41398-022-01881-6.

11. Filipponi A, Esposito E, Mannino D, et al. The contribution of altered neuronal autophagy to neurodegeneration. *Pharmacol Ther*. 2022;268:101878. doi:10.1016/j.pharmthera.2022.101878.

12. Rey CC, Robert V, Bouisset G, et al. Altered inhibitory function in hippocampal plasticity with a preferential role of the flavonoid hesperidin. *J Mol Histol*. 2021;52:1043-1065.

13. Cheng YJ, Lin CH, Lane HY. Involvement of Cholinergic, Adrenergic, and Glutamatergic Network Modulation with Cognitive Dysfunction in Alzheimer’s disease. *Int J Mol Sci*. 2021;22:22591. doi:10.3390/ijms22052591.

14. Shao F, Wang M, Guo Q, et al. Characterization of Alzheimer’s Disease-Associated Excitatory Neurons via Single-Cell RNA Sequencing Analysis. *Front Aging Neurol*. 2021;13:742176. doi:10.3389/ftagn.2021.742176.

15. Ramón-Cajal y. Contribution a la connaissance de la neurone cerebrale et cerebelleuse dans la parade generale progressive. *Trab Lab Infect Biol Univ Madrid*. 1925;23:157-216.

16. Achacoso N, Gayarre M. Contribución al estudio de la neurologia en la corteza de la demencia senil y su participación en la alteración celular de Alzheimer. *Trab Lab Infect Biol Univ Madrid*. 1914;12:1-38.

17. Achacoso N, Gayarre M. La corteza cerebral en la demencia paralítica con el nuevo método del oro y sublimado de Caja. *Trab Lab Infect Biol Univ Madrid*. 1914;12:68-93.

18. Penfield W. Neurology: normal and pathological. *In: Penfield W*, ed. *Cytology and cellular pathology of the nervous system*, 1932;2:423-479.

19. Papa M, De Luca C, Pette F, et al. Astrocyte neuron interplay in maladaptive plasticity. *Neurobiol Dis*. 2014;72:15-28.

20. Verkhratsky A, Rodriguez JJ, Parpura V. Astroglia in neurological diseases. *Future Neurosci*. 2018;3:149-162.

21. Verkhratsky A, Parpura V, Montana V, et al. Glial cells in (patho)physiology. *J Neurochem*. 2012;124:1-47.

22. Azizi G, Mirshafiey A. The potential role of proinflammatory and anti-inflammatory cytokines in Alzheimer disease pathogenesis. *Immunopharmacol Immunomunopathol*. 2012;4:81-98.

23. Perez-Alvarez A, Navarrete M, Covelo A, et al. Structural and functional plasticity of astrocyte processes and dendritic spine interactions. *J Neurosci*. 2020;58:312-325.

24. Prinz M, Priller J, Sisodia SS, et al. Heterogeneity of CNS myeloid cells and their complex roles in Alzheimer disease: implications for therapy. *Acta Neuropathol*. 2013;126:479-497.

25. Rajkovic A, Hocking A, Griffee J, et al. Microglial regulation of cerebrovascular functions. *Glia*. 2021;70:60-72.

26. McGeer PL, Itagaki S, Boyes BE, McGeer EG. Reactive microglia are positive for HLA DR in the substantia nigra of Parkinson’s disease brains. *Neurology*. 1998;18:1285-1291.

27. McGeer PL, Rogers J, McGeer EG. Inflammation, anti-inflammatory agents and Alzheimer disease: The last 12 years. *J Alzheimers Dis*. 2006;9:271-276.
74. Veldhanski A, Nedergaard M, Hertz L. Why are astrocytes important? Neuro chem. Res. 2015;40:389–401.

75. Smlklick A. Dynamic learning and memory, synaptic plasticity and neurogenesis: an update. Front Behav. Neurosci. 2014;8:106.

76. Kuljevicew-Nawrot M, Sykova E, Chvatal A, et al. Astrocytes and glutamate homeostasis in Alzheimer’s disease: a decrease in glutamate synthase, but not in glutamate transporter-1, in the prefrontal cortex. ASN Neuro. 2013;5:273-282.

77. Liddelow SA, Barres BA. Reactive astrocytes: production, function, and therapeutic potential. Immunity. 2017;46:957-967.

78. Liddelow SA, Guttenplan KA, Clarke LE, et al. Neurotrophic reactive astrocytes are induced by axotomy in mice. Nature. 2015;514:481-485.

79. Bayraktar OA, Bartels T, Holmquist S, et al. Astrocyte layers in the mammalian cerebral cortex revealed by a single-cell in situ transcriptomic map. Nat. Neurosci. 2020;23:500-509.

80. Chal H, Díaz-Castro R, Shigetomi E, et al. Neuronal circuit-specialized astrocytes: transcriptomic, proteomic, morphological, and functional evidence. Neurev. 2017;54:396-397.

81. Lin CCJ, Yu K, Hatcher A, et al. Identification of diverse astrocyte populations and their malignant analogs. Nat. Neurosci. 2017;20:396-405.

82. Ivov MM, Eriksen GA, Shkolikhev MM, et al. The aging astrocyte transcriptome from multiple regions of the mouse brain. Cell Rep. 2018;22:269-285.

83. Souanov AA, Wu X, Tsankova NN, et al. Phenotypic heterogeneity and plasticity of isocortical and hippocampal astrocytes in the human brain. J. Neurosci. 2014;34:2285-2298.

84. Cahoy JD, Emery B, Kaulahl A, et al. A transcriptome database for astrocytes, neurons, and oligodendrocytes: A new resource for understanding brain development and function. J. Neurosci. 2008;28:264-278.

85. Batiuk MY, Martirosyan A, Wahis J, et al. Identification of diverse astrocyte populations and their malignant analogs. Nat. Neurosci. 2020;11:1220.

86. Alvaraz MI, Rivas L, Laczun C, et al. Astroglial cell subtypes in the cerebella of Alzheimer disease brains suggest possible roles for both A1 and A2 astrocytes in Alzheimer disease. Aging Cell. 2015;40:389-401.

87. Li T, Liu T, Chen X, et al. Microglia induce the transformation of A1/A2 reactive astrocytes via the CXCR7/PI3K/Akt pathway in chronic post-surgical pain. Neuroinflammation. 2020;23:500-509.

88.divides G, Stavropoulos A, Manioudaki M, et al. Activation of both transforming growth factor-β and bone morphogenetic protein signalling pathways upon traumatic brain injury restrains pro-inflammatory and boosts tissue reparatory responses of reactive astrocytes and microglia. Brain Commun. 2019;1:e0028. doi:10.1093/braincomms/fcz028.

89. Gordon R, Neal M, Luo J, et al. Prokineticin-2 upregulation during neuronal injury mediates a compensatory protective response against dopaminergic neuronal degeneration. Nat. Commun. 2016;7:12932.

90. Alvarez MI, Rivas L, Lacruz C, et al. Astroglial cell subtypes in the cerebella of Alzheimer disease mice via the downregulation of TLR4/MyD88 signaling. Glia. 2015;66:1429-1451.

91. Ziskin JL, Nishiyama A, Rubio M, et al. Vascular release of glutamate from unmyelinated axons in white matter. J. Neurosci. 2007;30:321-330.

92. Verkharrtsky A, Nedergaard M, Hertz L. Why are astrocytes important? Neurochem. Res. 2015;40:389-401.

93. Liu LR, Liu JC, Bao JS, et al. Interaction of Microglia and Astrocytes in the Neurovascular Unit. Front. Immunol. 2020;11:1024. doi:10.3389/fimmu.2020.01024.

94. Crawford AH, Tripathi BB, Richardson WD, Franklin RJ. Developmental origin of oligodendrocyte lineage cells determines response to demyelination and susceptibility to age-associated functional decline. Cell Rep. 2016;15:761-773.

95. Crawford AH, Tripathi BB, Richardson WD, Franklin RJ. Developmental origin of oligodendrocyte lineage cells determines response to demyelination and susceptibility to age-associated functional decline. Cell Rep. 2016;15:761-773.

96. Hill RA, Patel KD, Medved J, et al. NG2 cells in white matter but not gray matter proliferate in response to PDGF. J. Neurochem. 2013;133:14558-14566.

97. Marques S, Gil J, Collod-Broche S, et al. Oligodendrocyte heterogeneity in the aging juvenile and adult central nervous system. Science. 2016;352:1326-1329.

98. Foerster S, Hill MF, Franklin RJM. Diversity in the oligodendrocyte lineage: Plasticity or heterogeneity? Glia. 2019;25:2411.

99. Viganó F, Dimou L. The heterogenic nature of NG2-oligodendrocyte Brain Res. 2015;1638:129-137. doi:10.1016/j.brainres.2015.09.012.

100. Kang SH, Fukaya M, Yang JK, et al. NG2-CNS glial progenitors remain committed to the oligodendrocyte lineage in postnatal life and following neurodegeneration. Neuro. 2010;68:668-671.

101. Clermente D, Ortega MC, Meleno-Jerez C, et al. The effect of glia-glia interactions on oligodendrocyte precursor cell biology during development and in demyelinat- ing diseases. Front Cell Neurosci. 2017;12:268.

102. Mensis AR. Carroll WM Oligodendrocyte lineage cells in chronic demyelination of multiple sclerosis optic nerve. Brain. 2015;25:517-530.

103. Zhang P, Ichimoto Y, Grammatikakis I, et al. Senolytic therapy alleviates Aβ-associated oligodendrocyte progenitor cell senescence and cognitive deficits in an Alzheimer’s disease model. Nat. Neurosci. 2019;22:719-728.

104. Nicholls MR, St-Pierre MK, Wendeln AC, et al. Inflammatory mechanisms in neurodegeneration. J. Neurochem. 2019;149:562-581.

105. Streit WJ, Khoshbouei H, Bechmann I. Dystrophic microglia in late-onset Alzheimer disease neuropathology. Brain Res. 2019;143:179-224.
132. Villacampa N, Henecka MT. Microglia in Alzheimer’s disease: Local heroes. J Exp Med. 2020;217:e20192311.

133. Planas 2018Gállosiá M, Méndez-Muñiz F, Ortxoa-de-Amezaga A, et al. dendritic Cells and Microglia Have Non-redundant Functions in the Inflamed Brain with Protective Effects of Type 1 cDCs. Cell Rep. 2020;33:108291. doi:10.1016/

134. Ajami B, Samanuk M, Wiegbohner P, et al. Single-cell mass cytometry reveals distinct populations of brain microglial cells in mouse neuroinflammation and neurodegeneration models. Nat Neurosci. 2018;21:541–551.

135. Varvel NH, Neher JJ, Bosch A, et al. infiltrating monocytes promote brain in-
flammation and exacerbate neural damage after status epilepticus. Proc Natl Acad Sci U S A. 2016;113:E5665–E5674. doi:10.1073/pnas.1604263111.

136. Reed-Geaghen EG, Croxford AL, Becher B, Landergh GE. Plaque-associated microglial cells derive from resident microglia in an Alzheimer disease model. J Exp Med. 2020;217:e2019374. doi:10.1084/jem.2019374.

137. Nummerjean A, Kirchhoff F, Helmschen F. Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo. Science. 2005;308:1314–1318.

138. Colton C, Wilcock DM. Assessing activation states in microglia. CNS Neurol Drug Targets. 2010;9:174–191.

139. Colton CA. Heterogeneity of Microglial Activation in the Innate Immune Re-
sponse in the Brain. J Neuroimmunol. 2009;9:349–198.

140. Choy V, Le Charpentier T, Lebon S, et al. Characterization of phenotype markers and neuronontatic potential of polarized primary microglia in vitro. Brain Behav Immun. 2013;32:70–85.

141. Grabert K, Michoel T, Karavolos MH, et al. microglial brain region-dependent diversity and selective regional sensitivities to aging. Nat Neurosci. 2016;19: 504–516.

142. De Biase LM, Schoebel KE, Fusfeld ZH, et al. Local cues establish and maintain region-specific phenotypes of basal ganglia microglia. Neuron. 2017;95:341–356.

143. Olah M, Patrić V, Villanacci AC, et al. A transcriptomic atlas of aged human microglia. Nat Commun. 2018;9:1–8.

144. Reu P, Khosravi A, Bernard S, et al. The lifespan and turnover of microglia in the mouse brain. Immunity. 2013;39:81–94.

145. Togo T, Akiyama H, Kondo H, et al. Expression of CD40 in the brain of Alzheimer’s disease: A comparative species review. Cell Rep. 2021;10:1138.

146. McPherson JL, Itagaki S, Togo H, McGeer EG. Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. Neurobiol Lett. 1987;79:195–200.

147. Varvel NH, Neher JJ, Bosch A, et al. Expression of CD40 in the brain of Alzheimer’s disease and other neurological diseases. Brain Res. 2000;885:117–121.

148. Hendrickx DA, van Eden CG, Schuurman KG, et al. Staining of HLA-DR, Iba1 and CD68 in human microglia reveals partially overlapping expression depending on cellular morphology and pathology. J Neuroinflammation. 2017;14:9:2–22.

149. Guttenplan KA, Liddelow SA. Astrocytes and microglia: Models and tools. J Exp Med. 2019;216:71–83.

150. Murray PJ, Allen JE, Biswas SK, et al. Macrophage activation and polarization: Nomenclature and experimental guidelines. J Leukoc Biol. 2013;95:817–825.

151. Varvel NH, Neher JJ, Bosch A, et al. Does Alzheimer’s disease alters oligodendrocytic and neurodegeneration. J Alzheimers Dis. 2017;54:67–77.

152. Serrano-Pozo A, Mielke ML, Gomez-Isla T, et al. Reactive glia not only associates with plaques but also parallels tangles in Alzheimer’s disease. Am J Pathol. 2011;179:1373–1384.

153. Toledano A, Alvarez MI, Lopez-Rodriguez AB, et al. Does Alzheimer’s disease exist in all primates? Alzheimer pathology in non-human primates and its pathophysiological implications (II). Neurology. 2014;29:42–55.

154. Togo T, Akiyama H, Kondo H, et al. Expression of CD40 in the brain of Alzheimer’s disease. Brain Res. 2000;885:117–121.

155. Toledano A, Alvarez MI, Lopez-Rodriguez AB, et al. Does Alzheimer’s disease exist in all primates? Alzheimer pathology in non-human primates and its pathophysiological implications (II). Neurology. 2012;2:354–369.

156. Ferrer J, Garcia MA, Gonzalez IL, et al. Aging-related tau astroglopathy (ARTAG): not only tau phosphorylation in astrocytes. Brain Pathol. 2018;28:965–985.

157. Beckman D, Chakrabarty P, Ott S, et al. A novel tau-based rhesus monkey model of Alzheimer’s disease: A comprehensive gene expression encyclopedia of glia cells in health and disease. Glia. 2015;63:1495–1506.

158. Munoz-Antolín JJ, Faisal AS, et al. Targeting Microglia-Synapse Interactions in Alzheimer’s Disease? J Alzheimers Dis. 2017;58:597–612.

159. Arai J, Schubert D, Sapp E, et al. Microglia generation in chronic amyloidosis in mice and men. J Leukoc Biol. 2013;95:817–825.
217. Manelli AM, Stine WB, Van Eldah IJ, LaDu MJ. ApoE and Abeta-42 interaction: Effects of isoform and conformation on structure and function. J Mol Neurosci. 2004;24:235-246.

218. Banger SW, Harmon AD. Microglial activation by Alzheimer amyloid precursor protein and modulation by apolipoprotein E. Nature. 1997;388:787-881.

219. Laaskowitz DT, Horbuth K, roses AD. Apolipoprotein E and the CNS response to injury. J Cerebr Blood Flow Metab. 1998;18:665-671.

220. Di Battiata AM, Heinsinger NM, Rebeck GW. Alzheimer’s disease genetic risk factor APOE-epsilon4 also affects normal brain function. Curr Alzheimer Res. 2016;13:1200-1207.

221. Fernandez CG, Hamby M, McReynes ML, Roy WJ. The role of APOE4 in disrupting the homeostatic functions of astrocytes and microglia in aging and Alzheimer’s Disease. Front Aging Neurosci. 2019;11:14.

222. Hyman BT, Gomez-Isla T, West H, et al. Clinical and neuropathological correlates of apolipoprotein E genotype in Alzheimer’s disease. Ann NY Acad Sci. 1996;777:158-165.

223. Kockx M, Traini M, Korthardes L. Cell-specific production, secretion, and function of apolipoprotein E. J Mol Med (Berlin). 2018;96:361-371.

224. Brown CM, Whight E, Conlon CA, et al. Apolipoprotein E isoform mediated regulation of nitric oxide release. Free Rad Biol Med. 2002;32:1071-1075.

225. Brodbek J, McGuire J, Liu Z, et al. Structure-dependent impairment of intracellular apolipoprotein E4 trafficking and its detrimental effects are rescued by small-molecule structure correctors. J Biol Chem. 2011;286:17217-17226.

226. Zhong N, Ramaswamy G, Weigburgh KH. Apolipoprotein E4 domain interaction induces endoplasmic reticulum stress and impairs astrocyte function. J Neurosci. 2009;29:2773-2780.

227. Rieker C, Miglavska E, Vaucher A, et al. Apolipoprotein E4 expression causes gain of toxic function in isogenic human induced pluripotent stem cell-derived endothelial cells. Arterioscler Thromb Vasc Biol. 2019;39:e195-e207.

228. Zhang Kj, Zhang Hf, Zhang Xm, et al. Apolipoprotein E isoform-specific effects on cytokine and nitric oxide production from mouse Schwann cells after inflammatory stimulation. Neurosci Lett. 2011;499:175-180.

229. Liu CC, Zhao N, Fu Y, et al. ApoE4 accelerates early seeding of amyloid pathology. Neuron. 2017;96:1024-1032.

230. Soto-Rojas LO, Pacheco-Herrero M, Martinez-Gomez PA, et al. The neurovascular unit dysfunction in Alzheimer’s Disease. Int J Mol Sci. 2021;22:2022.

231. Kirschin GW, Krey R, Ge S. The Hippocampal Neuro-Glia-Vascular Network: Metabolic Vulnerability and Potential Neurogenetic Regulation in Disease. Brain Pathol. 2018;13:129-144.

232. Colonna M, Brioschi S. Neuroinflammation and neurodegeneration in human brain at single-cell resolution. Nat Rev Immunol. 2020;20:81-92.

233. Gahande-Rodriguez E, Keane L, Capasso M. Microglial phagocytosis in aging and Alzheimer’s disease. J Neurosci. 2020;40:284-298.

234. Stuijf P, Schlachetzki JCM. Microglia in Alzheimer’s Disease. Curr Alzheimer Res. 2019;17:29-43.

235. Vogels T, Murgoci AN, Hromadka T. Intersection of pathological tau and mi-croglia at the synapse. Acta Neuropathol Commun. 2019;7:109.

236. Sterkova A, Lu A, Manuso R, et al. Novel Alzheimer risk genes determine the microglia response to amyloid-ß but not to TAU pathology. EMBO Mol Med. 2020;12:e10606.

237. Madore C, Yin Z, Leibowitz J, et al. Microglia, Lifestyle Stress, and Neurodegeneration. Immunity. 2020;52:222-240.

238. Yao K, Zu HB. Microglial polarization: Novel therapeutic mechanism against Alzheimer’s disease. Inflammopharmacology. 2020;28:95-110.

239. Van der Kant R, Goldstein LS, Ossenkoppel R. Amyloid-ß-independent regulators of tau pathology in Alzheimer disease. Nat Rev Neurosci. 2020;21:21-35.

240. Dumitri GP, Lee X, Basile G, et al. A biomarker that identifies senescent human cells in culture and in aging skin in vivo. Proc Natl Acad Sci USA. 1995;92:936-9367.

241. Kunz DJ, Decary S, Hong Y, et al. Senescence-associated (beta)-galactosidase reflects an increase in lysosomal mass during replicative aging of human endothelial cells. J Cell Sci. 2000;113:3631-3622.

242. Dehacq-Chainaux F, Erasumilsky JD, Campisi J, et al. Protocols to detect senescence-associated beta-galactosidase (SA-beta-gal) activity, a biomarker of senescent cells in culture and in vivo. Nat Protoc. 2009;4:1799-1806.

243. Xie L, Zhang N, Zhang Q, et al. Inflammatory factors and amyloid-ß-induced microglial polarization promote inflammatory crosstalk with astrocytes. Aging (Albany NY). 2020;12:22538-22549.