LETTER TO THE EDITOR

Clinical response to everolimus in a patient with Hodgkin’s lymphoma harboring a TSC2 mutation

Hodgkin’s lymphoma (HL) is a lymphoid malignancy that has a high cure rate with modern chemoradiation regimens. However, about 10–20% of patients with advanced stage HL develop relapsed or progressive disease, with 5% of patients presenting with primary progressive HL that has a 5-year overall survival of only 26%. It is of paramount importance to develop novel treatments for this population of patients. Recently, several novel drugs were found to be active in refractory and/or relapsed HL, including the anti-CD30 monoclonal antibody brentuximab vedotin, the anti-PD1 monoclonal antibody nivolumab and the mTOR inhibitor everolimus. In a phase II clinical trial, everolimus was shown to induce responses in 47% of patients with relapsed HL, including one patient who achieved clinical complete remission (CR). However, the lack of biomarkers for response are one of the many challenges that preclude a more widespread use of this drug in the therapy of HL. The advent of next-generation sequencing may help to determine molecular biomarkers associated with response to a specific drug, particularly when patients with an exceptional response are evaluated.

We herein report the case of a female patient with primary refractory HL that achieved a near CR with single-agent everolimus. Genomic analysis of the tumor at the time of progression to the last therapy demonstrated damaging mutations of the TSC2 gene that may lead to increased mTOR pathway activation.

A 31-year-old female patient was diagnosed with stage IVB nodular sclerosis HL in July 2012. She had primary refractory
disease and did not respond to ABVD and several different salvage chemotherapeutic regimes, including ICE, GNP, DHAP, BEACOPP, IVAC and bendamustine. She never underwent autologous peripheral blood stem cell transplantation due to lack of chemosensitive disease. She was treated with brentuximab vedotin, but had disease progression after six cycles, presenting with lung and pleural involvement and compression of spinal roots (Figure 1a). Patient was then started on therapy with off-label everolimus, based on the results of the aforementioned clinical trial, as she was considered to be refractory to chemotherapy. After 2 months of therapy with single-agent everolimus (10 mg orally once daily), a new positron emission tomography-computed tomography scan showed that the patient had achieved a near CR (Figure 1b) with disappearance of most sites of disease and > 90% reduction in the remaining sites. As she had no HLA-matched donors, she underwent haploidentical allogeneic stem cell transplantation 1 month after achieving a response to everolimus. Unfortunately, she passed away due to transplant-related complications (cytomegalovirus pneumonia) on day +68.

Because of this patient-significant response to everolimus, an inhibitor of the PI3K-Akt-mTOR pathway, we hypothesized that this patient’s tumor might harbor mutations associated with increased activity of this pathway. We sent the formalin-fixed paraffin-embedded block of a lymph node biopsy done at the time of disease progression on brentuximab to be analyzed by means of a comprehensive targeted next-generation-sequencing-based genomic profiling (FoundationOne Heme, Cambridge, MA, USA) using DNA and RNA in a CLIA certified laboratory (Foundation Medicine). This method consists of the analysis of the entire coding sequence of 405 cancer-related genes, 31 selected introns frequently involved in rearrangements and RNA sequencing of 265 genes commonly fused in cancer, to a median coverage of > 500x and > 20 000 000 reads. No microdissection of Reed–Sternberg cells was performed. No germline DNA was available for sequencing. The results revealed 21 genomic abnormalities, including a missense mutation on gene TSC2. The V1711M mutation on TSC2 is a conservative missense mutation localized on the Rap-GAP domain of the tuberin protein. The sorts intolerant from tolerant score (in silico tool that predicts whether changes in amino acids in protein sequences are deleterious) was 0.02, suggesting that this mutation is damaging. Furthermore, this mutation has been previously reported in a patient with squamous cell skin carcinoma of the face. Besides the TSC2 mutation, several other oncogenic variants were detected (Table 1).

Inactivating mutations on TSC2 and TSC1 have been described in tuberous sclerosis, and the TSC1–TSC2 complex is a critical negative regulator of mTORC1 (mTORC1). Within the TSC1–TSC2 complex, TSC1 stabilizes TSC2, whereas TSC2 acts as a GTPase-activating protein (GAP) for the small GTPase Rheb (Ras homolog enriched in brain). When active, the TSC1–TSC2 complex inhibits mTORC1 by stimulating the conversion of Rheb–GTP to Rheb–GDP. Therefore, the ability of a variety of upstream pathways to affect mTORC1 activity is dependent on modifications that functionally inhibit or activate the TSC1–TSC2 complex. The GAP domain, where the V1711M mutation is located, is crucial for TSC2 function, and loss of TSC2 activity may lead to mTORC1 pathway activation. It has been suggested that TSC1/TSC2 inactivation predicts for responses to mTOR inhibitors. Indeed, TSC1 mutations have been found in 8% of bladder malignant tumors, and are correlated with sensitivity to everolimus. In another report, a nonsense mutation in TSC2 (Q1178*) was described in a patient with anaplastic thyroid cancer who had an impressive response to everolimus. Activation of the mTOR pathway is found in the majority of cases of HL and HL cell lines, but the mechanisms that lead to this increased activity are currently unknown.

We herein describe the first case of TSC2 mutation in a patient with advanced HL who had an impressive clinical response to everolimus. One limitation of this report is that no functional studies were done to confirm the oncogenic role of this particular TSC2 mutation in HL. However, we do believe that the response to everolimus seen in this case suggests that mutations of TSC2 and other genes of the mTOR pathway should be further investigated in patients with HL.

### Table 1. Oncogenic genomic alterations found

| Gene      | AA change | VAF (%) or fold amplification |
|-----------|-----------|------------------------------|
| TSC2      | p.V1711M  | 50                           |
| DNMT3A    | p.R986L   | 11                           |
| PTCH1     | p.G288D   | 12                           |
| ATR       | p.W2104*  | 7                            |
| BRCAl     | p.R2842C  | 42                           |
| B2M       | p.M11     | 13                           |
| B2M       | p.M1K     | 26                           |
| MSH6      | p.T1219I  | 9                            |
| TP53      | p.K291N   | 19                           |
| TP53      | p.R248Q   | 21                           |
| AXL1      | p.Q1517*  | 9                            |
| XPO1      | p.E571K   | 32                           |
| AXIN1     | p.P245    | 9                            |
| FBXO11    | p.R640fs*5| 9                            |
| CD36      | p.Y325*   | 51                           |
| CDC73     | Splice site (c.370+1G>A) | 10 |
| JAK2      | Amplification | 11x                        |
| CD274     | Amplification | 11x                        |
| PDCD1LG2  | Amplification | 11x                        |
| CCND3     | Amplification | 22x                        |
| KDM4C     | Amplification | 11x                        |

Abbreviations: AA, amino acid; VAF, variant allele fraction.

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