Original Research Article

Infections with yeast and yeast like fungi in cancer patients with special emphasis on non-neoformans cryptococcal infections: Retrospective study

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Candida spp. and Cryptococcus spp. are important fungal pathogens in cancer patients. Non-albicans Candida are of special concern, since some are highly virulent and show reduced susceptibility to antifungal agents. Non-neoformans Cryptococcal infections have also shown an incremental rise over the past four decades. The aim of this retrospective study was to evaluate the prevalence, distribution and susceptibility pattern of various species of Candida and Cryptococcus causing infections in cancer patients. This Retrospective study was conducted from Jan 2011 to Aug 2016. Yeasts/yeast like fungi isolated in various specimens received for fungal and also bacterial culture were included. Fully automated VITEK 2 compact was used for final species identification and susceptibility testing of isolated yeasts. The prevalence of infections was 1.36% during the study period with prevalence being 0.54% in patients with haematological cancer and 2.45 in patients with solid tumours. Among various solid tumours maximum infection rate was seen in patients with head and neck cancers i.e. 4.01%. A total of 9.09% growths were responsible for Blood Stream Infections (BSI). Most common Candida sp. isolated was C.tropicalis (37.06%) followed by C.albicans (36.87%). Out of total 35 Cryptococcus sp. isolated 30 were Cryptococcus laurentii and only 5 were Cryptococcus neoformans. Low level of resistance was shown by C.albicans to all the antifungal agents. C.tropicalis also showed low resistance with only 2.01% resistance to Fluconazole and Amphotericin B. Higher resistance rate was observed in C.krusei with 7.69% isolates resistant to Fluconazole and 15.38% resistant to Amphotericin B. Rate of resistance shown by C.glabrata to Fluconazole was also quite high i.e. 9.09%. Most of the Candida sp. showed good sensitivity to both Caspofungin and Micafungin except C.krusei with 15.38% resistance to both candins. Out of all the antifungal agents tested for Voriconazole was the most effective for all the yeasts isolated with highest resistance rate being 7.69% shown by C. krusei. All Cryptococcus neoformans isolated were sensitive to the antifungal agents tested for. i.e. Amphotericin B, Flucytosine, Fluconazole and Voriconazole. Out our study emphasizes the need to make new prophylaxis policies for Candida infections. Also further studies should be conducted to determine the antifungal susceptibility pattern of Cryptococcus laurentii.

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1. Introduction

Patients with cancer are considered a population at high risk for developing invasive fungal infections. Candida spp. and Cryptococcus spp. are the yeasts most frequently isolated in clinical practice. They are important nosocomial pathogens in cancer patients and are associated with substantial morbidity and mortality, prolonged hospitalization and increased healthcare expenditures.¹,² Several reasons have been proposed for the increase in invasive fungal infections in cancer patients including extended survival of cancer patients as well as advances in supportive care by the use of antineoplastic and immunosuppressive agents, improved control of bacterial infections by using broad-spectrum antibiotics, hematopoietic stem cell transplantation, prosthetic devices and grafts and more aggressive surgery.³–⁵

Candida is a normal commensal of the skin, gastrointestinal and genitourinary tracts.⁶ Candida sp. continue to be
the most common fungal pathogens in patients with cancer. They account for 75% of total fungal infections. Although *Candida albicans* remains the most prevalent species, there has been a clear shift towards non-*albicans* *Candida* species namely *Candida tropicalis*, *Candida parapsilosis*, *Candida krusei* particularly found in the neutropenic patients and *Candida glabrata* found especially in patients with solid tumour. Non-*albicans Candida* are of special concern, since some are highly virulent and are associated with treatment failure due to reduced susceptibility to antifungal agents. Reported increase in non-*albicans Candida* might have been mediated by one or more confounding risk factors in addition to selection for species that were less susceptible to azoles.

The genus *Cryptococcus* comprises several species which are able to cause infections in human s and animals. Infections caused by them are frequently related to the exposure to avian droppings especially pigeons which are reservoirs for *Cryptococcus* species. *Cryptococcus neoformans* and *Cryptococcus gattii* are the major pathogens within the genus. Other *Cryptococcal* species have traditionally been considered non-pathogenic, however, there has been an incremental rise in non-*neoformans* *Cryptococcal* infections namely *Cryptococcus albidus*, *Cryptococcus laurentii*, *Cryptococcus luteolus*, *Cryptococcus uniguttatus*, *Cryptococcus curvatus* and *Cryptococcus humicola* over the past four decades. This increase may be due to enhanced awareness of such infections, improved laboratory detection of non-*neoformans* *Cryptococcal* species, wide use of antifungals favouring the appearance of rare and more resistant species and a rise in the number of at-risk patients.

There are very few studies from India on the pattern of yeast infections in cancer patients. The Gujarat Cancer & Research Institute (GCRI), Ahmedabad, India is the largest cancer hospital of the country and provides state-of-the-art diagnostic and therapeutic services to the patients of all types of origin and financial background suffering from cancer. GCRI caters to a large number of patients mainly from the states of Gujarat, Rajasthan, Madhya Pradesh, Chattisgarh and also many patients from Maharashtra, Uttar Pradesh, Bihar and Jharkhand. It is presumed that the findings of this study would faithfully reflect the pattern of infections due to yeast and yeast like fungi in cancer patients from western India and to an extent from that of developing nations like ours.

2. Aims and objectives

The Aim of this Retrospective study was

1. To identify the different species of *Candida* and *Cryptococcus* causing infections in cancer patients,
2. To study their prevalence in various type of cancers and in different samples and
3. To find their susceptibility pattern to antifungal agents

3. Materials and Methods

The study was conducted retrospectively. A total of 572 samples were studied during the period of Jan 2011 to Aug 2016. Patients of both sexes and all age groups ranging from 1 to 86 years were included in the study.

The various specimens included were those routinely submitted for diagnosing the infectious agent in the Microbiology Laboratory at Gujarat Cancer & Research Institute (GCRI), Ahmedabad from patients diagnosed with cancer. Yeasts/ yeast like fungi isolated in samples received for fungal culture and also for bacterial culture were included. The various specimens from which yeasts/ yeast like fungi were isolated included Blood, Urine, Sputum, samples from Surgical Site Infections (SSI), Stool, Ascitic fluid, Bronchoalveolar Lavage (BAL), Endotracheal secretions, Pleural fluid, various Tips like central line catheter tip, Hickman catheter tip, Endotracheal tube etc. and other samples which include swabs, pus and tissue samples from sites other than SSI. Cerebrospinal fluid (CSF) samples did not show any growth of *Candida* sp. or *Cryptococcus* sp. during the study period and thus CSF has not been mentioned in this study.

Patients clinically suffering with fever, cough, expectoration and radiological findings of chest, burning micturition and symptoms of septicemia were considered to have infection. Patients on chemotherapy, not getting treated by administration of antibiotics and still having persisting symptoms of infection were an indication for reporting yeasts. In stool, urine and sputum samples colony count of less than 1000 CFU/ ml of yeasts were considered as colonization and these isolates were not included in the study. Microscopically sputum samples having gram positive oval/ round budding yeast cells with pseudohyphal elements and patients having clinical symptoms were considered for reporting yeasts. In case of urine samples microscopic findings of gram positive oval/ round budding yeast cells along with inflammatory cells and again patients showing clinical symptoms of urinary tract infection were considered to be infectious. Isolation of yeast in urine was considered significant when there was reproducibility of growth in two separately collected urine samples from the same patient. For stool samples the clinical history of diarrhea, nausea, Absolute Neutrophil Count (ANC) of patient and again microscopic findings were taken into consideration before reporting yeasts/ yeast like fungi. Also yeasts were reported when there was no growth of any pathogenic bacteria in these samples (Stool, urine and sputum).

Multiple episodes in the same patient were counted as separate infections unless they were caused by the same fungal agent.
Yeast/ yeast like fungi were identified on the basis of colony morphology of growth and gram stain findings. Fully automated VITEK 2 compact was used for final species identification and susceptibility testing of isolated yeasts.

Data were collected using WHONET software version 5.6 and were compiled in Microsoft excel.

3.1. Ethics statement

No informed consent was obtained from the individual patients whose data were analyzed in this non interventional study. It is not necessary to obtain approval from a medical ethics committee for this type of observational study since it contains no directly identifiable data.

3.2. Statistical analysis

The prevalence of fungal infections in various types of cancers was analyzed. The association between fungal infections in Haematological cancers and Solid tumours was studied using Chi square test. P-value less than 0.05 was considered significant.

4. Results

During the study period, total 1.36% (572/ 42013) samples showed infection with Candida or Cryptococcus sp. These 572 samples were studied further.

The gender ratio was 1:1 (50% each) showing no male or female preponderance.

Figure 1 shows percentage of infections due to yeast/ yeast like fungi in different age groups. Patients above the age of 40 years constituted 65.91%(377/572) of the total fungal infections, maximum i.e. 23.78% in age group 51 to 60 years followed by 21.50% in patients above 60 yrs and 20.63% in 41 to 50 years of age. Minimum infection rate (4.72%) was seen in 11 to 20 years of age group patients.

Table 1 showed prevalence of yeast and yeast like fungal infection in patients with haematological cancer to be 0.54%(129/23927) which is lesser than in patients with solid tumours i.e. 2.45%(443/18086). The result was statistically significant with a p value of less than 0.00001.

Among solid tumours the infection rate was maximum i.e. 4.01% (97/2416) in patients with head and neck cancers followed by 3.70%(117/ 3166) in GIT cancer patients, 2.43%(40/1649) in patients with Respiratory cancers and 1.95%(97/ 4976) in patients with Gynaecological cancers as shown in Table 2.

Figure 2 A total of 9.62% of Candida and Cryptococcal growths were responsible for Surgical Site Infections (SSI) while 9.09% were responsible for Blood Stream Infections (BSI). Ascitic fluid, ET secretions, Pleural fluid, BAL, Tip and other samples constituted remaining 16.43% of the growths.

Table 3 shows that out of total 572 yeasts and yeast like fungi isolated 93.88%(537/572) were Candida sp. while 6.12%(35/572) were Cryptococcus sp. Among Candida grown more than half i.e. 63.13%(339/537) were non albicans Candida sp. and 36.87% (198/537) were Candida albicans. Overall most common Candida sp. isolated was C.tropicalis i.e. 37.06% (199/ 537). Other species of Candida isolated commonly were C.glabrata (9.62%), C.parapsilosis (6.67%), C.famata (2.27%), C. krusei (2.27%) and C.guilliermondii (1.22%). Only 2 species of Cryptococcus were isolated. Cryptococcus laurentii constituted 85.71%(30/35) while Cryptococcus neoformans constituted only 14.29%(5/35) of the total Cryptococcal growths indicating rise in non neoformans Cryptococcal infections.

Calbicans and C.tropicalis were the two most frequent species isolated in all types of samples as listed in Table 4. C.tropicalis was seen more commonly in Ascitic fluid (100% - 1 out of 1 isolate), Blood (38.46%), Endotracheal (ET) secretions (100% - 1 out of 1 isolate), Pleural fluid (44.44%) and Urine (47.89%) while C.albicans was more frequent in Bronchoalveolar lavage (BAL) (70%), Sputum (47.62%), Stool (39.52%), Tip (38.46%) and other samples (45%). Samples from Surgical Site infections (SSI) showed equal growth of both C.tropicalis and C.albicans (34.55% each). Blood samples showed growth of C.tropicalis (38.46%) most commonly followed by C.parapsilosis (34.62%), C.albicans (5.77%) and C.guilliermondii (5.77%). Ascitic fluid, BAL and ET secretions did not show any Cryptococcal growth. All the remaining samples showed growth of Cryptococcus laurentii more commonly than Cryptococcus neoformans. Out of total 30 Cryptococcus laurentii isolated 26 isolates (86.67%) were from Stool, Urine, Sputum and Blood samples.

Low level of resistance was shown by Calbicans to all the antifungal agent s tested for as shown in Table 5. C.tropicalis also showed low resistance with only 2.01% (4 out of 199 isolates) resistant to Fluconazole and Amphotericin B. C.parapsilosis showed no resistance to Amphotericin B and only 2.7% resistance (1 out of 37 isolates) to Fluconazole. Higher resistance rate was observed in C.krusei with 7.69% isolates resistant to Fluconazole and 15.38% resistant to Amphotericin B. Rate of resistance shown by C.glabrata to Fluconazole was also quite high i.e. 9.09%. Maximum resistance to Fluconazole was shown by C.haemulonii i.e. 66.67% (2 out of 3 isolates). Most of the Candida sp. showed good sensitivity to both Caspofungin and Micafungin except C.krusei (15.38% isolates were resistant to both the candins). Out of all the antifungal agents tested for Voriconazole was the most effective for all the yeasts isolated with highest resistance rate being 7.69% shown by C.krusei. All Cryptococcus neoformans isolated were sensitive to the antifungal agents.
tested for i.e. Amphotericin B, Flu cytosine, Fluconazole and Voriconazole. Susceptibility pattern of Candida famata, Candida spheraica and Cryptococcus laurentii could not be obtained due to limitations of VITEK 2 Compact instrument.

5. Discussion

Fungal infections are an important cause of morbidity and mortality in cancer patients who are vulnerable to these infections. Despite the limitations related to its retrospective nature, our study provides important information.

Microscopic examination gives early indication of the presence of yeasts. The observation of yeasts in normally sterile tissue or fluids is significant, provided the specimens have been collected aseptically. The possibility of yeast as a pathogen must be considered when the budding yeast cells are present and hyphae are abundant, long and thin. The prevalence of yeast/yeast like fungi during the study period was 1.36% in our study which was comparable with a study by Paswan et al which showed incidence of yeast infections to be 1.6% in patients with haematological malignancies. Another prospective study by El-Mahlawy et al on fungal infection in children with cancer showed higher yeast infection rate i.e. 2.9%.

Candida and Cryptococcal infections were seen equally in males and females. Patients with age more than 40 years showed 65.91% of the total yeast and yeast like fungi infections in our study probably due to more immunocompromised status in older patients. The findings were similar to a study by Hajjeh et al in which 72% candedemia cases occurred among persons > 45 years old.

Higher infection rate was seen in patients with Head and neck cancers, Gastrointestinal tract cancers, Respiratory cancers and Gynaecological cancers in our study as yeasts occur as normal flora in oral cavity, skin, lower genitourinary tract and gastrointestinal tract. When the immunity becomes low these commensals may cause infection. Infection rate among patients with haematologic malignancies was only 0.54% in our study. A study by L. Pagano et al showed higher rate of infection (1.6%) due to yeasts in patients with haematologic malignancies. The main risk for fungal infections in patients with haematologic malignancies is neutropenia. This neutropenia results from intensive cytotoxic chemotherapy and radiotherapy to totally ablate malignant bone marrow stem cells and haematopoetic stem cell transplantation resulting in graft vs host reaction.

Urine, stool and sputum samples were together responsible for 64.86% of total growths in this study which is again because of yeasts occurring as commensals at these sites. A study by Timothy et al gave similar results with 75% of yeasts recovered from Respiratory and urine specimens. Candidiasis is the fourth common cause of nosocomial Blood Stream infections worldwide, accounting for 9% of all such infections in the United States. In this study blood samples were responsible for 9.09% of total growth of yeast/yeast like fungi.

Candida sp continue to be the most common fungal pathogen in patients with cancer. In our study 93.88% yeasts recovered were Candida sp. Overall most common species of Candida isolated was Candida tropicalis (37.06%) followed by Calbicans (36.87%). This is of concern as Candida tropicalis shows a higher invasive capacity and 50 to 60% of the colonized patients develop disseminated Candidiasis. In a study by Paswan et al predominance of Candida tropicalis was more as compared to Candida albicans in cancer patients. Non-albicans Candida (NAC) constituted 63.13% of the total growths of Candida in this study. Other studies have also shown increasing trend towards non-albicans Candida. Prophylactic use of antifungal agents like Fluconazole are responsible for rise in NAC infections. NACs are of special concern, since some are highly virulent and are associated with treatment failure due to reduced susceptibility to antifungal agents.

Our study showed out of total 572 isolates 35 (6.12%) were Cryptococcus sp. A study by L. Pagano et al on epidemiology of fungal infections in patients with haematologic malignancies showed that 8 out of 192 yeasts (4.17%) isolated were due to Cryptococcus sp. The prevalence of Cryptococcal infection increased during the Acquired Immunodeficiency Syndrome (AIDS) pandemic. Out of total Cryptococcal growths 85.71% were Cryptococcus laurentii while only 14.29% were Cryptococcus neoformans in our study. A study on neoformans Cryptococcal infections: a Systematic Review stated that 80% of non-neoformans Cryptococcal infections are due to Cryptococcus laurentii and Cryptococcus albidus. Non-neoformans cryptococcal infections have been reported rarely in humans but reports of cases of infections due to Cryptococcus laurentii have been increasing during the past decade. This increase may be due to enhanced awareness of such infections and improved laboratory detection of these infections. In addition, although the wide use of antifungals has efficiently reduced the incidence of the most prevalent pathogenic fungi, it has also favoured the appearance of niches for rare and may be more resistant species. The major risk factors for non-neoformans Cryptococcal infections include impaired cell mediated immunity due to haematologic malignancy, corticosteroid therapy, organ transplantation. A study by D. Averbuch et al showed that most of the Cryptococcus laurentii isolated were in cancer patients.

Calbicans continues to be the single most common species causing candidemia in USA, however Blood Stream Infections (BSI) due to non-albicans Candida species
are increasing. A programme of epidemiology and fungal susceptibility performed in the USA, Canada and South America, called SENTRY demonstrated Candida glabrata to be the second most frequent species (first being C. albicans) causing Candidaemia followed by C. parapsilosis. A study from Barcelona, Spain showed C. parapsilosis to be the most common species responsible for BSI after C. albicans. In our study C. tropicalis (38.46%) was the most common species isolated from blood followed by Candida parapsilosis (34.62%) while Candida albicans was responsible for only 5.77% of the total BSI. Another study from India by Paswan et al. also showed predominant isolation of C. tropicalis (49%) from blood. C. tropicalis was also the species most commonly isolated from other sterile body fluids like Urine, Pleural fluid and Ascitic fluid in our study. Other samples showed C. albicans to be the most common yeast isolated. A study by Keihn et al. showed that C. albicans was the most frequent isolate from all the samples including Blood, urine, Pleural fluid etc. followed by C. tropicalis and C. glabrata. In the early 1990s, increasing use of fluconazole to treat HIV-infected patients with recurrent oropharyngeal candidiasis resulted in the selection of Candida species intrinsically less susceptible to azoles and is responsible for emergence of less susceptible non- albicans Candida species.

In this study growth of Cryptococcus laurentii was seen more commonly than Cryptococcus neoformans in all the samples showing Cryptococcal growths. Stool, Urine, Sputum and Blood samples together showed 86.87% of the total Cryptococcus laurentii growths. The major reservoir of Cryptococcus sp. is droppings of pigeons and other birds. The natural habitat of Cryptococcus laurentii is unknown. However, a study by Mattson et al. showed feral pigeons to be carriers of medically significant fungi like Cryptococcus laurentii and Cryptococcus uniguttulatus. The ubiquitous presence of pigeons in our hospital may be a possible source of infection by Cryptococcus laurentii in our study. Infection is usually acquired by inhalation. After the initial pulmonary infection, Cryptococcus laurentii may spread to other organ systems, particularly in immunosuppressed patients even if pulmonary infection is asymptomatic. Also the presence of invasive devices have been shown to be a significant risk factor associated with Cryptococcus laurentii infection. Thus Blood Stream Infections due to Cryptococcus laurentii may be acquired via the intravenous catheters in our study. Another route of transmission may be nosocomial spread of infection. Cerebrospinal fluid (CSF) have been considered to be an important sample for isolation of fungal infections especially Cryptococcus sp. But in our study CSF did not show any growth of yeast or yeast like fungi during the study period.

Low rates of resistance among C. albicans to various antifungal agents including Fluconazole were seen in our study. These findings have important implications for the management of C. albicans infections as Fluconazole is commonly used for treatment of uncomplicated Candida infections. A low level of Fluconazole resistance was found among C. tropicalis and C. parapsilosis isolates, and a high level of resistance was detected among C. krusei isolates in our study which was consistent with other studies. Another finding in our study was C. glabrata showing 9.09% resistance to Fluconazole which was even higher than that shown by C. krusei (7.69%). C. krusei is considered resistant to Fluconazole but resistance pattern of C. glabrata to Fluconazole is variable. High resistance to Amphotericin B was shown by C. krusei and C. guillermondii in our study. C. krusei showed high resistance to other drugs also including Caspofungin, Micafungin and Voriconazole but showed 100% sensitivity to Flucytosine. Also Voriconazole showed least in vitro resistance to most of the Candida species in our study. Voriconazole has a broad spectrum of activity and is available both orally and parenterally, and may be suitable as second-line therapy in selected patients resistant to first line agents like Fluconazole.

The 5 isolates Cryptococcus neoformans were 100% sensitive to all the antifungal agents tested for i.e. Amphotericin B, Flucytosine, Fluconazole and Voriconazole in this study. Candida e. Caspofungin and Micafungin are inactive against Cryptococcus species due to greater proportion of (1, 3)-α-D glucan linkages present in cell wall polymers of Cryptococcus sp. against which the candins act. Therefore susceptibility tests of Cryptococcus neoformans was not performed to Candins. Susceptibility pattern of Cryptococcus laurentii could not be obtained due to limitations of our instrument – VITEK 2 Compact. There are a few studies stating that there is a favourable response of Cryptococcus laurentii to appropriate antifungal therapy but different degrees of susceptibilities to antifungal agents are seen in vitro.

![Fig. 1: Fungal infections in different age groups](image-url)
Table 1: Clinical diagnosis and prevalence of fungal infections

| Diagnosis         | No. of samples Infected | No. of samples Not Infected | Percentage |
|-------------------|-------------------------|-----------------------------|------------|
| Hematological Cancers | 129                     | 23798                       | 0.54       |
| Solid Tumors      | 443                     | 17643                       | 2.45       |
| Total             | 572                     | 41441                       | 1.36       |

p value is <0.00001.

Table 2: Rate of infection in various types of Solid tumours

| Diagnosis                  | No. of samples received | No. of infected samples | Percentage |
|----------------------------|-------------------------|-------------------------|------------|
| i. CNS Tumour              | 1719                    | 27                      | 1.57       |
| ii. GIT cancer             | 3166                    | 117                     | 3.70       |
| iii. Gynaecological cancers| 4976                    | 97                      | 1.95       |
| iv. Head and neck cancers  | 2416                    | 97                      | 4.01       |
| v. Respiratory cancers     | 1649                    | 40                      | 2.43       |
| vi. Other solid tumours *  | 4160                    | 65                      | 1.56       |
| Total                      | 18086                   | 443                     | 2.45       |

*Includes tumours like Breast cancer, Prostate cancer, Renal cancer, Urinary bladder cancer, Bone cancers etc.

Table 3: Species of Candida and Cryptococcus isolated

| Species                  | No. of samples | Percentage |
|--------------------------|----------------|------------|
| Candida sp.              | 537            | 93.88      |
| i. C. albicans           | 198            | 36.87      |
| ii. C. famata            | 13             | 2.42       |
| iii. C. glabrata         | 55             | 10.24      |
| iv. C. guilliermondii    | 7              | 1.30       |
| v. C. haemulonii         | 3              | 0.56       |
| vi. C. kefyr             | 4              | 0.74       |
| vii. C. krusei           | 13             | 2.42       |
| viii. C. lipolytica      | 2              | 0.37       |
| ix. C. lusitaniae        | 1              | 0.19       |
| x. C. parapsilosis       | 37             | 6.89       |
| xi. C. pelliculosa       | 1              | 0.19       |
| xii. C. rugosa           | 2              | 0.37       |
| xiii. C. spherica        | 1              | 0.19       |
| xiv. C. tropicalis       | 199            | 37.06      |
| xv. C. utilis            | 1              | 0.19       |
| Cryptococcus sp.         | 35             | 6.12       |
| i. Crypto. laurentii     | 30             | 85.71      |
| ii. Crypto. neoformans   | 5              | 14.29      |

6. Conclusion

Patients with cancer are at particular risk for infections with yeast and yeast like fungi. With the increase in immunocompromised patients and widespread use of immunosuppressive agents non- albicans Candida and non-neoformans Cryptococcus are emerging human pathogens. The low incidence of Fluconazole resistance among isolates of Calbicans is reassuring. But the role of azole chemoprophylaxis in development of drug resistance in less susceptible Candida sp. needs to be examined and new prophylaxis policies need to be made. Also non-neoformans Cryptococcus are easy to miss, so a high degree of clinical suspicion, improved culture and identification techniques are required. Our findings emphasize that further studies need to be conducted to determine the antifungal susceptibility pattern of Cryptococcus laurentii.

6.1. Author’s contribution

All the authors were responsible for study conception, design, analysis and interpretation of data. The authors participated in drafting and revising the article and gave final approval for the version to be submitted.

7. Conflicts of Interest

All authors declare that they have no conflicts of interest with respect to the research, authorship and/or publication of this article.
Table 4: Distribution of species of *Candida* and *Cryptococcus* isolated in different specimens

| Species          | Ascitic Fluid | BAL | Blood | ET Secretions | Pleural fluid | Sputum | SSI | Stool | Tip | Urine | Others |
|------------------|---------------|-----|-------|---------------|---------------|--------|-----|-------|-----|-------|--------|
| *C. albicans*    | 0             | 7   | (70)  | 3             | 0             | 2      | 50  | 19    | 49  | 5     | 36     |
| *C. famata*      | 0             | 0   | 0     | 0             | 0             | 2      | 1 (1.9) | 1 (1.82) | 4 | 1     | 5       |
| *C. glabrata*    | 0             | 1   | (10)  | 0             | 0             | 1      | 8   | 6     | 16  | 1     | 17     |
| *C. guilliermondii* | 0          | 0   | 3     | 0             | 0             | 0      | 0   | 0     | 2   | 0     | 0       |
| *C. haemulonii*  | 0             | 0   | 1     | 0             | 0             | 0      | 1   | (1.82) | 2   | 0     | 1 (0.7) |
| *C. kefyr*       | 0             | 0   | 0     | 0             | 0             | 0      | 0   | 0     | 5   | 0     | 0       |
| *C. krusei*      | 0             | 0   | 2     | 0             | 0             | 3      | (2.86) | 1     | 1   | (0.81) | 1 (0.7) |
| *C. lipolytica*  | 0             | 0   | 0     | 0             | 0             | 0      | 0   | 0     | 0   | 0     | 1 (0.7) |
| *C. lusitaniae*  | 0             | 0   | 0     | 0             | 0             | 0      | 0   | 0     | 0   | 1     | 1 (0.7) |
| *C. parapsilosis*| 0             | 0   | 18    | 0             | 1             | 3      | 6   | 0     | 1   | 3     | 5       |
| *C. pelliculosa* | 0             | 0   | 1     | 0             | 0             | 0      | 0   | 0     | 0   | 0     | 0       |
| *C. rugosa*      | 0             | 0   | 0     | 0             | 0             | 1      | (0.95) | 0     | 0   | 1     | 0       |
| *C. sphaerica*   | 0             | 0   | 0     | 0             | 0             | 0      | 0   | 0     | 1   | (0.81) | 0       |
| *C. tropicalis*  | 1 (100)       | 2   | (20)  | 20            | 1 (100)       | 4      | (44.44) | 31   | (29.52) | 32   | (25.81) |
| *C. utilis*      | 0             | 0   | 0     | 0             | 0             | 1      | (0.81) | 0     | 0   | 0     | 0       |
| *Crypto. laurentii* | 0         | 0   | 4     | 0             | 1             | (11.11) | 5   | (4.76) | 1   | (1.82) | 10     |
| *Crypto. neoformans* | 0      | 0   | 0     | 0             | 0             | 1      | (0.81) | 0     | 0   | 1     | 0       |
| **Total**        | 1 (100)       | 10  | (100) | 52            | (100)        | 1 (100) | (100) | 9 (100)| (100) | 105   | (100) |

Table 5: In vitro susceptibilities of *Candida* sp. and *Cryptococcus neoformans* to various antifungal agents

| Species (Total No. of isolates) | Antifungal agent | No. of resistant isolates (%) |
|---------------------------------|------------------|------------------------------|
| *C. albicans* (198)             | Amphotericin B   | 2 (1.01)                     |
|                                 | Caspofungin       | 5 (2.53)                     |
|                                 | Fluconazole       | 2 (1.01)                     |
|                                 | Micafungin        | 5 (2.53)                     |
|                                 | Voriconazole      | 3 (1.52)                     |
| *C. glabrata* (55)              | Amphotericin B   | 2 (3.64)                     |
|                                 | Caspofungin       | 2 (3.64)                     |
|                                 | Fluconazole       | 2 (3.64)                     |
|                                 | Micafungin        | 7 (12.73)                    |
|                                 | Voriconazole      | 2 (3.64)                     |
| *C. guilliermondii* (7)         | Amphotericin B   | 2 (28.57)                    |
|                                 | Caspofungin       | 1 (14.29)                    |
|                                 | Fluconazole       | 1 (14.29)                    |
|                                 | Micafungin        | 1 (14.29)                    |
|                                 | Voriconazole      | 1 (14.29)                    |
| *C. haemulonii* (3)             | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. kefyr* (4)                  | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. krusei* (13)                | Amphotericin B   | 2 (15.38)                    |
|                                 | Caspofungin       | 2 (15.38)                    |
|                                 | Fluconazole       | 2 (15.38)                    |
|                                 | Micafungin        | 2 (15.38)                    |
|                                 | Voriconazole      | 2 (15.38)                    |
| *C. lipolytica* (2)             | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. lusitaniae* (1)             | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. parapsilosis* (37)          | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. pelliculosa* (1)            | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. rugosa* (2)                 | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *C. tropicalis* (199)           | Amphotericin B   | 4 (2.01)                     |
|                                 | Caspofungin       | 2 (1.01)                     |
|                                 | Fluconazole       | 4 (2.01)                     |
|                                 | Micafungin        | 2 (1.01)                     |
|                                 | Voriconazole      | 3 (1.51)                     |
| *C. utilis* (1)                 | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
| *Cryptococcus neoformans* (5)   | Amphotericin B   | 0                            |
|                                 | Caspofungin       | 0                            |
|                                 | Fluconazole       | 0                            |
|                                 | Micafungin        | 0                            |
|                                 | Voriconazole      | 0                            |
Fig. 2: Isolation of yeast/ yeast like fungi from different samples
*Includes Central line catheter tip, Hickman catheter tip and Endotracheal tube (ET) tip etc.
**Includes swabs, pus and tissue samples from sites other than surgical site infection.
Note: None of the CSF samples showed growth of Candida sp. or Cryptococcus sp. during the study period and so CSF sample has not been mentioned.

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