3,4-Dicaffeoylquinic Acid, a Major Constituent of Brazilian Propolis, Increases TRAIL Expression and Extends the Lifetimes of Mice Infected with the Influenza A Virus

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Brazilian green propolis water extract (PWE) and its chemical components, caffeoylquinic acids, such as 3,4-dicaffeoylquinic acid (3,4-diCQA), act against the influenza A virus (IAV) without influencing the viral components. Here, we evaluated the anti-IAV activities of these compounds in vivo. PWE or PEE (Brazilian green propolis ethanol extract) at a dose of 200 mg/kg was orally administered to Balb/c mice that had been inoculated with IAV strain A/WSN/33. The lifetimes of the PWE-treated mice were significantly extended compared to the untreated mice. Moreover, oral administration of 3,4-diCQA, a constituent of PWE, at a dose of 50 mg/kg had a stronger effect than PWE itself. We found that the amount of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) mRNA in the mice that were administered 3,4-diCQA was significantly increased compared to the control group, while H1N1 hemagglutinin (HA) mRNA was slightly decreased. These data indicate that PWE, PEE or 3,4-diCQA possesses a novel and unique mechanism of anti-influenza viral activity, that is, enhancing viral clearance by increasing TRAIL.

1. Introduction

Influenza is a common infectious disease, and various anti-influenza drugs are already on the market. For example, M2 ion channel inhibitors (amantadine and rimantadine) and neuraminidase inhibitors (zanamivir, oseltamivir, etc.) have been used to treat influenza viral infections in many areas, including the United States, Europe, and Asia [1–3]. Although oseltamivir is one of the most popular anti-influenza drugs, many resistant strains have already been discovered [4, 5]. Fortunately, the resistant strains that have been found to date are not yet pandemic. However, we need to carefully monitor the emergence and prevalence of new strains with pandemic characteristics, such as the H1N1 pandemic with drug resistance, which may occur because of the high incidence of mutations [6] and genomic rearrangements [7]. To avoid the worst, the discovery of novel anti-influenza agents that do not carry the risk of creating resistant strains would be urgently required.

Propolis, a resinous substance collected by honeybees from various plant sources, is used by the bees to protect the nest entrance against intruders and bacteria. Since ancient times, it has been used as a traditional folk medicine in many countries. Recent studies have demonstrated that propolis has a wide range of pharmacological properties, including antibacterial [8–10], antioxidant [11], anti-inflammatory...
The antiviral activity of propolis has been reported and includes anti-BBMV [16], anti-HSV [17–21], antipoliiovirus [22], anti-IBDV [10], anti-reovirus [10], antipotato viruses [23], and anti-HIV [24] activities [25, 26]. The anti-influenza activity of propolis has been also described in several reports [27–30], but the details of its activity were different. The reason for this may be differences in the plant origin of the propolis [14].

Recently, we have demonstrated that Brazilian green propolis water extract (PWE) and its components, particularly 3,4-dicafeoylquinic acid (3,4-diCQA), have anti-IAV activity [31]. Moreover, we found that a central mechanism of these compounds is the enhancement of cellular viability via unknown cellular processes rather than activity against the viral components [31].

Here, we examined the in vivo effects of propolis extracts on the lifetimes of mice infected with the IAV. To obtain insights into the mechanism of the antiviral effects of PWE and PEE, we examined tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) and IAV hemagglutinin (HA) gene expression in the lungs, because it has been reported that the ethanol extract of Polish propolis has anti-tumor activity via TRAIL enhancement [32].

2. Methods

2.1. Reagents and Compounds. Oseltamivir phosphate (Tamiflu) was purchased from the Chugai Pharmaceutical Co., Ltd. (Tokyo, Japan). Chlorogenic acid was purchased from the Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). All media and reagents for cell culture were purchased from Invitrogen (Carlsbad, CA, USA), Sigma (St. Louis, MO, USA), and Wako Pure Chemicals (Osaka, Japan). Water extracts of Brazilian green propolis (Minas Gerais State, Brazil) originating from Baccharis dracunculifolia [33] were obtained from the API Co., Ltd. (Gifu, Japan). 3,4-diCQA (99% or 82% purity) was purified from 25% ethanol-obtained from the API Co., Ltd. (Gifu, Japan). Chlorogenic acid was purchased from Japan SLC, Inc. (Hamamatsu, Japan) and housed at 4°C until use.

2.2. Mice. Female BALB/c mice (5 weeks old) were obtained from Japan SLC, Inc. (Hamamatsu, Japan) and housed at room temperature (maintained at 23 ± 3°C with a relative humidity range of 32–64% and a regular 12 hr light/dark cycle. The mice were fed a CE-2 rodent diet from CLEA Japan Inc. (Tokyo, Japan) and allowed free access to water.

2.3. Viruses. The influenza virus wild-type strain A/WSN/33 (H1N1), generated from cloned cDNAs using plasmid-based reverse genetics [34], was kindly supplied by Dr. Yoshihiro Kawaoka (Division of Virology, Department of Microbiology and Immunology, Institute of Medical Science University of Tokyo, Japan). The viruses were stored at –80°C until use.

2.4. Cells. Madin-Darby canine kidney (MDCK) cells were a kind gift from Professor Hideto Fukushi (United Graduate School of Veterinary Sciences, Gifu University) and were maintained in α-minimal essential medium (MEM) containing 10% fetal bovine serum (FBS) and penicillin-streptomycin.

2.5. Virus Preparation in Cell Culture and Determination of the TCID50 Value. MDCK cells were cultured in T-75 culture flasks. Confluent cells were washed with phosphate-buffered saline (PBS) and then incubated with the virus (0.1 multiplicity of infection (MOI)) in 2mL of viral growth medium (D-MEM (Dulbecco’s Modified Eagle Medium) containing 0.625 μg/mL of trypsin and 0.1% bovine serum albumin (BSA)) for 1 hr at 37°C. Then, 10mL of viral growth medium was added, and the cells were incubated for 24 hrs at 37°C. The culture supernatant was harvested by centrifugation (3000 rpm at 4°C) and stored at –80°C until use. TCID50 (50% tissue culture infective dose) values were determined by infecting the MDCK cells with serial dilutions of the viral suspension in 96-well microtiter plates, and the TCID50 values were calculated using the method of Reed and Muench [35] after 6 days.

2.6. Mouse Adaptation and Preparation of Virus. Viruses that had been passaged 2 or 3 times in MDCK cells were further serially passaged in 6-week-old female BALB/c mice. The first passaged virus lot was obtained by intranasally inoculating a 20-μL suspension of the starting virus (5.1 × 10⁸ TCID₅₀ per mL) into each animal, which had been lightly anesthetized using diethyl ether. At 48 hrs postinoculation (hpi), the mice were sacrificed. Their lungs were removed, placed into PBS (2 mL for the lungs of two mice) supplemented with 0.1% BSA and penicillin-streptomycin, and homogenized using a PRO200 homogenizer (PRO Scientific Inc.; Oxford, CT, USA). The suspension was further centrifuged at 3000 rpm at 4°C, stored at –80°C, and termed MP1 (mouse passage 1). The subsequent passages were similarly performed by inoculating 40 μL of the viral solution of the previous passage into each mouse. The second- and higher-passage virus lots were termed MP2, MP3, and so forth.

2.7. Anti-Influenza Therapeutic Efficacy in Mice

2.7.1. Assessment of Survival Time. Six-week-old female BALB/c mice were lightly anesthetized with diethyl ether and inoculated with 40 μL of a mixture of MP6 to MP12 (i.e., mouse-adapted viruses 8.3–53 × 10⁸ TCID₅₀/mouse, resp.) once per day for 4 days via intranasal instillation. The mice were categorized into 6 groups: (1) control (vehicle); (2) oseltamivir phosphate (0.5 mg/kg of body weight); (3) PWE (100 mg/kg); (4) PEE (100 mg/kg); (5) 3,4-diCQA (99% purity; 50 mg/kg); (6) chlorogenic acid (50 mg/kg). The compounds (in a 5% Arabic gum solution) were orally administered to the mice twice per day at intervals of more than 6 hrs for 6 days (from 0 to 5 dpi (days postinfection)). Administration began 4 hrs before viral inoculation. Mouse survival was observed once per day for 12 days after viral inoculation.

2.7.2. Assessment of mRNA Expression in the Lungs. Six-week-old female BALB/c mice were lightly anesthetized with
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diethyl ether and inoculated with 20 μL of MP14, a mouse-adapted virus (4.9 × 10⁷ TCID₅₀/mouse) once per day for 2 days via intranasal instillation. The mice were categorized into three groups: (1) control (vehicle); (2) 3,4-diCQA (82% purity; 50 mg/kg); (3) oseltamivir phosphate (0.5 mg/kg). Each compound (in a 5% Arabic gum solution) was orally administered to the mice twice per day at 6-hour intervals for 8 days (from 0 to 7 dpi). Administration began 4 hrs before viral inoculation. In the mRNA expression study (n = 13–14 at 0 dpi), the 6 mice with the lowest body weight in each group and the remaining surviving mice in each group were killed at 4 and 7 dpi, respectively, and the mRNA expression in the lungs was determined using quantitative real-time PCR (qPCR), as described below.

Total RNA was extracted from the lung homogenate using the TriPure Isolation Reagent (Roche Diagnostics, Mannheim, Germany), according to the manufacturer’s protocol. Briefly, the mice were sacrificed, and their lungs were removed, placed into a solution (1.5 mL for the lungs of one mouse) of TriPure, and homogenized using a PRO200 homogenizer. After centrifugation, the water phase of the lysate was recovered, and RNA was precipitated with the ethanol and rinsed. The purified RNA was dissolved in 50–100 μL of nuclease-free water. The amount of RNA was estimated using a NanoDrop (Thermo Scientific, Wilmington, DE, USA). The cDNA from a 500-ng aliquot was synthesized using a PrimeScript RT reagent kit (Takara Bio Inc., Shiga, Japan) and oligo dT primers according to the manufacturer’s protocol. Quantitative real-time PCR for the IAV or mouse gene was then performed. Briefly, the aliquot of the RT reaction solution was amplified using SYBR Premix Ex Taq II (Takara Bio Inc., Shiga, Japan) and a Real-Time PCR Thermal Cycler (“Thermal Cycler Dice Real Time,” Takara Bio Inc., Shiga, Japan), according to the manufacturer’s protocol. H1N1-specific primers were selected based on HA (hemagglutinin) mRNA using Primer Express software (PE Applied Biosystems, Waltham, MA, USA). The sequences of the HA primer sets were purchased from Takara Bio Inc. (Shiga, Japan). The sequences of the mouse TRAIL primer sets have been described elsewhere [36]. The mouse HPRT (hypoxanthine guanine phosphoribosyl transferase) primer sets were purchased from Takara Bio Inc. (Shiga, Japan). The relative quantities of H1N1 HA mRNA and mouse TRAIL mRNA were normalized to the mRNA expression of mouse HPRT (a housekeeping gene).

2.8. Statistics. The survival times were statistically compared using the Kaplan-Meier log-rank test and pairwise multiple comparison procedures (the Holm-Sidak method) using StatView 3.5 (SAS Institute, Cary, NC, USA). The body weights were compared using one-way analysis of variance (ANOVA). When any significance was detected by ANOVA, Dennett’s nonparametric test was then applied using EKUSERU-Tokei 2006 (Social Survey Research Information Co., Ltd., Tokyo Japan). The relative mRNA expression of the target gene was compared between treatment groups using a paired t-test or Student’s t-test. A P value of less than 0.01 (P < 0.01) was considered to be statistically significant.

3. Results

3.1. PWE, PEE, and 3,4-diCQA Increase the Survival Times of Mice Infected with the Influenza Virus. In preliminary experiments, MDCK culture-derived WSN/33 (H1N1, IAV) was only moderately lethal to mice, so we adapted the parent WSN/33 to mice using serial passages to enhance its virulence [37–39], as described in the Section 2. Mouse-adapted WSN/33 was able to kill more than 80% of mice within 10 days when the mice inoculated daily for 3 to 4 days. The anti-influenza activities of PWE, PEE, 3,4-diCQA, chlorogenic acid, and oseltamivir phosphate were evaluated using the mouse-adapted WSN/33 strains.

The vehicle treatment (control) group exhibited a significant decrease in body weight from 0 to 7 dpi. Treatment with PWE, PEE, or chlorogenic acid did not affect this phenotype compared to the control. However, 3,4-CQA (at 5 dpi alone, P < 0.01) and oseltamivir (at 5 dpi, P < 0.05; at 6-7 dpi, P < 0.01) slightly and strongly, respectively, counteracted the body weight loss caused by the viral infection (Figures 1(a) and 1(b)).

In the same experiment, 88.9% of mice in the control group died by 12 dpi, while all of the oseltamivir-treated mice survived. The survival time of the oseltamivir group was significantly longer than that of the control group (Figure 2(a), Table 1). Moreover, although chlorogenic acid had no effect, PWE, PEE, or 3,4-CQA treatment increased the survival rate (Figures 2(a) and 2(b), Table 1).

3.2. 3,4-diCQA Increases TRAIL Expression but Decreases HA mRNA Expression in Mouse Lungs Infected with IAV. We examined TRAIL and HA mRNA expression in the lungs of mice infected with IAV. Treatment with 3,4-CQA or oseltamivir had no effect on TRAIL mRNA expression at 4 dpi (Figure 3). At 7 dpi, the 3,4-CQA treatment group exhibited an increase in TRAIL mRNA expression (P < 0.05), while the oseltamivir treatment group exhibited no change in the expression of TRAIL mRNA (Figure 3).

In the same experiment, the oseltamivir treatment group exhibited less HA mRNA expression than the control at 4 and 7 dpi (P < 0.05; Figure 4). The 3,4-diCQA treatment group exhibited similar HA mRNA expression at 4 dpi and slightly less expression at 7 dpi than the control group (Figure 4).

Overall, oseltamivir treatment decreased IAV HA mRNA expression rapidly after infection and had no effect on TRAIL mRNA expression, whereas 3,4-diCQA treatment moderately decreased IAV HA mRNA expression and increased TRAIL mRNA expression with a delay.

4. Discussion

First, we must point out that there are some controversies regarding the anti-influenza effects of various propolis-related substances, especially in ex vivo versus in vivo experiments. We have shown that PWE and PEE can increase
Figure 1: Effects of orally administrated oseltamivir phosphate, PWE, PEE, 3,4-diCQA, and chlorogenic acid on the body weight of BALB/c mice infected with the IAV. (a) control (●), oseltamivir (0.5 mg/kg; ■), 3,4-diCQA (50 mg/kg; ♦), (b) PWE (100 mg/kg; ▲), PEE (100 mg/kg; ▼), or chlorogenic acid (50 mg/kg; ○) was administrated twice daily starting at 4 hours before the first viral infection until 5 days postinfection (5 dpi). The data represent the mean ± SE for the mice that survived. ∗P < 0.05 and ∗∗P < 0.01 versus vehicle control.

Figure 2: Survival curves of BALB/c mice infected with the IAV to assess the in vivo effects of orally administrated (a) control, oseltamivir (0.5 mg/kg), 3,4-diCQA (50 mg/kg), (b) PWE (100 mg/kg), PEE (100 mg/kg), or chlorogenic acid (50 mg/kg). The colors correspond to those used in Figures 1(a) and 1(b). The infected mice were observed for 12 days. The statistical results are shown in Table 1.

the lifetimes of mice with IAV infections. In a previous report, anti-influenza activity was found for PWE [31] but not PEE (personal communication, Urushisaki T.) when using MDCK cells. Here, we speculate that, in ex vivo experiments, some substances (as yet unidentified) in PEE are cytotoxic to MDCK cells and may mask its anti-influenza activity. We have also shown that 3,4-diCQA can increase the lifetimes of mice with IAV infections (Table 1). 3,4-diCQA, a natural product, is an ester of two polyphenolic caffeic acids and one cyclitol (−)-quinic acid. While chlorogenic acid (a natural product, an ester of one caffeic acid and one cyclitol (−)-quinic acid) does not have anti-influenza activity in vivo, these two compounds have anti-influenza activity ex vivo [31]. This may occur due to differences in the pharmacokinetic properties of these compounds in vivo.

The major chemical ingredients in PWE and PEE are listed in Table 2 [40, 41], and these show that the amount of 3,4-diCQA in PWE and PEE is 3.3–6.1% and 1.9–3.5%, respectively. It must be noted that the anti-influenza activities of PWE and PEE can only be partially explained by 3,4-diCQA. Thus, it is very likely that unknown compounds that efficiently act against the IAV are also contained in PWE and PEE.

We have also shown that 3,4-diCQA increases the mRNA expression of TRAIL in the lungs of IAV-infected mice (Figure 4). Although TRAIL is an apoptosis-inducing factor in tumor cells [42], it also induces the apoptosis of influenza virus-infected cells in infected animals via the TRAIL receptor (DR5), whose expression is induced by the virus. Ishikawa et al. [36] have shown that TRAIL mRNA expression is induced by an influenza virus infection in mouse lungs, and an anti-TRAIL monoclonal antibody (mAb) delays influenza virus clearance in mice. Moreover, Brincks et al. [43] have shown that morbidity and the influenza virus
Table 1: Survival of mice infected with IAV after the oral administration of various substances.

| Treatment             | n  | Dose                  | Duration (dpi) | Survival time (dpi, mean ± SE) | Statistical significance |
|-----------------------|----|-----------------------|----------------|--------------------------------|-------------------------|
| 5% Arabic gum         | 9  | 0–5                   | 7.4 ± 0.7      |                                |                         |
| oseltamivir           | 8  | 0.5 mg twice/kg/day   | 0–5            | >12.0*                         | **                      |
| PWE                   | 9  | 100 mg twice/kg/day   | 0–5            | 10.0 ± 0.9                     | *                       |
| PEE                   | 9  | 100 mg twice/kg/day   | 0–5            | 10.1 ± 0.9                     | *                       |
| 3,4-diCQA             | 7  | 50 mg twice/kg/day    | 0–5            | 10.3 ± 1.5                     | *                       |
| Chlorogenic acid      | 9  | 50 mg twice/kg/day    | 0–5            | 8.1 ± 0.8                      | N.S.                    |

*standard error not determined; * P < 0.05; ** P < 0.01; N.S. not significant.

Figure 3: TRAIL expression in the lungs of BALB/c mice infected with the IAV. Vehicle, 3,4-diCQA (50 mg/kg), or oseltamivir (0.5 mg/kg) was administrated twice daily starting at 4 hrs before the first viral infection until 7 days post-infection (7 dpi). Total RNA was recovered at 4 dpi or 7 dpi, and relative mRNA expression was measured using real-time PCR. The quantity of mouse TRAIL mRNA expression was normalized to the mRNA expression of mouse HPRT. The quantities are shown as mean ± SE. ** P < 0.01.

Figure 4: HA expression in lungs. IAV HA mRNA expression was normalized to the mRNA expression of mouse HPRT. The quantities are shown as mean ± SE. ** P < 0.01 (Student’s t-test).

We confirmed that Brazilian green propolis extracts (PWE and PEE) have anti-influenza activities. Here, we also hypothesized that their mode of action at least partially includes two mechanisms: an unknown cytoprotective mechanism [31] and the enhancement of viral clearance via TRAIL overexpression. We hypothesize that both are induced by 3,4-diCQA and/or unknown active constituents of Brazilian green propolis.

An important point regarding our hypothesis concerning the anti-influenza effects of propolis or its constituent, 3,4-diCQA, is that it might trigger or enhance the self-defense machineries of the host. Although viruses can easily become resistant to anti-influenza drugs, such as oseltamivir, that directly target viral proteins [4, 5], it would be more difficult for viruses to become resistant to antiviral agents whose target molecules are host molecules or machineries. Therefore, health supplements such as propolis may be useful as an alternative strategy for protection against influenza virus infections.

Many traditional Chinese medicinal herbs appear to have potential anti-influenza effects [46], but their mechanisms...
are not well understood. Our findings show that PWE and PEE and their constituent, 3,4-diCQA, may be useful as a potent lead compound for anti-influenza medicine. This may promote research into anti-influenza medicine developed from traditional substances.

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### Table 2: Concentrations and molecular weights of the constituents of Brazilian green propolis [40, 41].

| Components                      | Content (w/w%) in PWE | Content (w/w%) in PEE | Molecular weight (g/mol) |
|--------------------------------|-----------------------|-----------------------|--------------------------|
| Chlorogenic acid                | 2.7–3.6               | 0.6–0.8               | 354.3                    |
| Caffeic acid                    | 0.2                   | 0.1–0.6               | 180.2                    |
| 3,5-dicaffeoylquinic acid       | 4.3–4.9               | 2.4–2.7               | 516.5                    |
| 3,4-dicaffeoylquinic acid       | 3.3–6.1               | 1.9–3.5               | 516.5                    |
| 4,5-dicaffeoylquinic acid       | —                     | —                     | 516.5                    |
| 3,4,5-tricaffeoylquinic acid    | 0.2                   | 0.6                   | 678.6                    |
| Artepillin C                    | 0.2–0.6               | 11.4–14.0             | 300.4                    |
| Baccharin                       | 0.0                   | 6.8                   | 562.6                    |
| Drupanin                        | 0.1                   | 1.8                   | 232.3                    |
| Isosakuranetin                  | —                     | —                     | 286.3                    |
| p-coumaric acid                | 3.7                   | 2.3–2.5               | 164.2                    |
| Ferulic acid                   | 0.1                   | 0.0–0.1               | 194.2                    |
| Quinic acid                     | —                     | —                     | 192.2                    |

*Data not available.*
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