Abstract. The aim of this study was to determine the myocardial area at risk (AAR), infarction-core size (IS) and the salvaged myocardial zone (SMZ), and to evaluate the imaging and histological characteristics of intramyocardial hemorrhage (IMH) after myocardial infarction using non-contrast T2 mapping on 7T magnetic resonance imaging (MRI). Twenty Sprague Dawley (SD) rats were randomly divided into the sham and model groups (n=10 in each). In the model group, myocardial infarction models were established by left anterior descending branch ligation. After 24 h, all animals were imaged on a 7.0 Tesla system with cine spiral imaging, T2 mapping with late gadolinium enhancement (LGE). The rats were then sacrificed for measurement of the IS and AAR using 2,3,5-triphenylterazolium chloride (TTC) and hematoxylin and eosin (H&E) staining. T2 mapping revealed that the AAR in the model group was significantly higher than that in the sham group. No remarkable T2 value was noted in the entire heart of the sham group. LGE and TTC staining demonstrated similar IS. T2 mapping and H&E staining revealed a similar AAR as well. T2 mapping characterized the IMH as a phenomenon resulting from the area of hypointensity in the hyperintensity involving the infarct-core zone and corresponding T2 value 928.6±1.52 msec with IMH vs. 35.8±2.61 msec without IMH; n=3 with 18 slices; P=0.032). In conclusion, non-contrast T2 mapping was a reliable approach to quantitatively evaluate the SMZ and IMH.

Introduction

Myocardial area at risk (AAR) is defined as the extent of ischemic myocardium following coronary artery occlusion, which comprises the nonviable infarction-core size (IS) and the viable salvaged myocardial zone (SMZ). Thus, measurement of AAR and IS allows for determination of the SMZ, which is accepted as a stronger predictor of therapeutic efficacy after myocardial infarction (1). Moreover, intramyocardial hemorrhage (IMH) constantly emerges during the development of myocardial infarction, which subsequently induces corresponding alterations in T2 value on T2 maps, inducing subsequent injuries and elevating the morbidity and mortality (2-4). However, the imaging characteristics on T2 maps need to be illustrated.

During the past decades, multiple indirect strategies, such as nuclear imaging and echocardiography, have been introduced to evaluate myocardial viability. Unlike those approaches with the limitations of acoustic windows and radiation, cardiovascular magnetic resonance imaging (CMR) does not employ ionizing radiation and offers higher spatial and temporal resolution. Furthermore, CMR may quantitatively assess cardiac function, and its application for assessing viability is widely accepted (5-8). Conventional application of T2-weighted magnetic resonance imaging (MRI) to delineate the AAR, a dark blood turbo spin echo (TSE) technique has been employed to quantitatively measure the AAR and SMZ post-reperfusion (9), and T2-weighted MRI has been utilized to identify the incidence of myocardial edema. However, these techniques have certain limitations with regard to adaptation of image variations associated with respiratory or cardiac motion, the signal associated with surface coil intensity inhomogeneity and the effect of blood flow (10). In 2007, T2 mapping was introduced to assess the AAR at the early stage (11-13) and
was subsequently performed in patients with myocardial infarction (14). Late gadolinium enhancement imaging (LGE) may be utilized to identify myocardial infarction characterized as increased signal intensity due to cellular membrane rupture and gadolinium-diethylenetriamine pentaacetic acid (Gd-DTPA) diffusion into intracellular spaces. Furthermore, a previous study by our group suggested that if myocardial edema were identified by novel faster T2 mapping, the AAR could be determined in spite of rats having an increased heart rate (15). Combined use of T2 mapping and LGE delineates the SMZ in ischemic myocardium and defines IMH as the area of hypointensity in the hyperintensity of the infarct-core zone. The present study was designed to establish a stable and reliable platform using a non-invasive technique to assess the SMZ and IMH in a rat model despite a high heart rate.

Materials and methods

Study design. A total of 20 4-week-old male Sprague Dawley (SD) rats with a mean weight of (250±20 g) provided by the Experimental Animal Center of Guiyang Medical University (Guiyang, China) were included in the present study and randomly assigned to the model and sham groups (n=10). The rats were accustomed at 20˚C, 70% humidity and 12-h light/dark cycle with free access to food and water. In the model group, the animal models with myocardial infarction were established by left anterior descending branch (LAD) ligation as previously described (16). The animals were anesthetized by intraperitoneal injection of sodium pentobarbital (Sichuan Kelun Pharmaceutical Co., Ltd., Sichuan, China) at a dosage of 50 mg/kg. Subsequently, the rats underwent thoracotomy and mechanical ventilation was monitored by real-time electrocardiogram (ECG). The LAD ligation was closed using a 7-0 silk suture. In the sham group, the animals received thoracotomy without LAD occlusion. The present study was approved by the Institutional Animal Research Committee of the First People's Hospital of Guiyang and was performed according to the recommendations of the Guide for the Care and Use of Laboratory Animals of the First People's Hospital of Guiyang (Guiyang, China).

MRI protocols. General. All MRI was performed on a 7T MRI (BioSpec 70/30; Bruker, Karlsruhe, Germany) 24 h after the corresponding treatments. First, cardiac function was assessed by cine spiral imaging of the entire left ventricle (LV), followed by T2 mapping imaging and LGE imaging. In each group, three rats were sacrificed and the heart was collected for 2,3,5-triphenyltetrazolium chloride (TTC) staining. Subsequently, the heart tissue was prepared for histological examination. Upon CMR, the rats were anesthetized with isoflurane (5 vol%; Sichuan Kelun Pharmaceutical Co., Ltd.) in a 2:1 mixture of air (0.3 l/min) and oxygen (0.1 l/min), and then maintained under anesthesia (1.5-2.5 vol% isoflurane) in a custom-built cradle to maintain a stable cardiac and respiratory rate. Each rat was placed in the prone position on a surface coil. ECG signal, respiratory and heart rates were obtained from three subcutaneous copper needles loaded in the bilateral hind limbs. Novel faster T2 mapping. After maintaining the respiratory rate between 30 and 50 cycles per minute, novel faster T2 mapping imaging was performed by changing the concentration of isoflurane. Five single-sliced simplified T2-mapping images were obtained on the short-axis slices during the mid-diastolic and end-inspiratory phases using ECG and respiratory gating systems. Imaging parameters were as follows: Repetition ime/echo time (TR/TE), 1,500 msec/10, 20, 30 msec; data acquisition time, 44 msec; matrix size, 192x192; field of view (FOV), 50x50 mm; and slice thickness, 1.5 mm. The last TE of 30 msec was selected based on the approximate T2 value of myocardial tissue at 7T MR. Analysis of variance (ANOVA) test was used for determination of AAR by novel faster T2 mapping.

LGE imaging. A standard multi-slice short axis late gadolinium fast low-angle shot-cine sequence was performed and cine MR images were acquired from the base to the apex. The imaging sequence was as follows: TR/TE, 8 msec/3.1 msec; flip angle, 10; matrix size, 256x256; FOV, 50x50 mm; slice thickness, 1.5 mm; and 25 frames for each slice. Contrast enhancement was examined 10 min after intravenous administration of Gd-DTPA via the hind limbs (0.2 mmol/kg; Magnevist, Bayer Health Care Pharma AG, Berlin, Germany) and anesthetics (1.5-2.5 vol% isoflurane) in a custom-built cradle were used. The imaging parameters were TR/TE, 5.2 msec/1.8 msec; flip angle, 25; matrix size, 256x256; FOV, 50x50 mm; slice thickness, 1.5 mm, and 25 frames for each slice. All CMR parameters are illustrated in Table I.

CMR imaging. Cardiac function at the end-diastolic and end-systolic phases was evaluated using cine spiral imaging, and the LV borders were manually delineated and calculated by multiplying the size involving slice thickness and gap. T2 mapping imaging data were analyzed using specific software in Matlab 7.1 (Mathwork Inc., Natick, MA, USA). The T2 value was obtained by manually drawing the region of interest (ROI), ~0.06 cm² (88 pixels) on T2 maps from the base to apex with a total of 6 slices. The AAR was then defined as the area of ischemic myocardium with a T2 value standard deviation (SD) of >2 from the remote myocardium on T2 mapping, and the infarction area was defined with a signal intensity SD of >5 above the mean of remote myocardium on LGE images, both of which were calculated by manual delineation of the square centimeter. Thereafter, the SMZ was obtained by subtracting the infarction size by LGE from the AAR. The IMH was identified as the area of hypointensity within the hyperintensity infarction area on T2 maps. The AAR, IS and SMZ were all expressed as the percentage of LV after parameter standardization. All manual data were independently measured by two experienced radiologists who were blinded to the study.

Histopathological analysis. After CMR imaging, the heart samples were prepared as paraffin-embedded slices (5-µm thick sections), and each slice was incubated with 1% 2,3,5-triphenyltetrazolium chloride (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) for 15 min at 37˚C and then observed under a microscope (magnification, x200). The infarction area was stained white, whereas the viable myocardium was stained red. LGE and TTC outcomes were visually matched by comparing in vivo images and ex vivo slices by two
experienced radiologists blinded to the study. Stratified analysis was performed according to whether IMH was present, which was assessed by measuring the area of hypointensity within the hyperintensity infarction area in T2 maps. Histological sections were subjected to hematoxylin and eosin (H&E) staining.

Statistical analysis. Values are expressed as the mean ± standard error of the mean. Comparison between pairs of groups was performed using an independent-samples t-test. The correlation between LGE and TTC staining, T2 mapping and H&E staining, and LGE and H&E staining was analyzed by linear regression analysis using the Pearson coefficient r. ANOVA was used for determination of AAR by novel faster T2 mapping. The detection of AAR and IS between two groups was statistically compared using the paired t-test. P<0.05 was considered to indicate a statistically significant difference.

Results

Animal grouping. The variations in the respiratory gating and ECG throughout the process of CMR affected the increase in time to acquire images. Sixteen rats were included in the final analysis. Two rats were excluded from the sham group due to low-quality images caused by motion artifacts. In the model group, one rat that died during anesthesia was excluded, and the other rat was excluded due to low-quality images due to technical reasons. In addition, three rats in the model group yielded hemorrhage according to the T2 maps.

CMR and histological assessment of myocardial infarction models. As illustrated in Table I and Fig. 1, the average end-diastolic volume (EDV) in the sham group was 125.6±5.27 µl, which was significantly higher compared with 82.4±5.03 µl in the model group (P=0.004; n=16) (Fig. 2). As illustrated in Fig. 3, the LGE images indicated a clear IS as a hypointensity region involving cardiac muscle by drawing a region (yellow dotted line; Fig. 3B), which was perfectly matched by TTC staining (blue dotted line; Fig. 3C). The IS was stained red, distinguishable from viable myocardium stained white. T2-mapping images (Fig. 3D) with three ETs of 10, 20 and 30 sec revealed that the T2 value was in pseudo color (Fig. 3E). All T2 maps were scaled between 0 and 80 msec. The AAR is shown in the simplified T2 maps (red dotted line; Fig. 3F), which corresponded to the T2 maps (red dotted line; Fig. 3G). In an overlay of Fig. 3E and F, the SMZ was calculated by subtracting the IS from the AAR and is presented between the red and yellow dotted lines (Fig. 3G).

Determination of AAR by novel faster T2 mapping. ANOVA revealed that the AAR in the model group defined by novel faster T2 mapping was 38.6±1.52 msec, which was significantly higher than 26.8±1.48 msec in the sham group (P=0.007); however, no significant difference was observed between the base and apex T2 values in the sham group (P=0.056). In the model group, the mean AAR, IS and SMZ were 38.6±1.52 msec (P=0.039), 42.4±1.67 msec (P=0.002) and 35.8±1.31 msec (P=0.016) compared with that in the sham group, as illustrated in Fig. 4A.

As shown in Fig. 4B, the proportion of AAR determined by T2 mapping had a good correlation with that determined by H&E staining. A statistical significance was noted between AAR determined by T2 mapping (61.98±10.34%) and H&E staining (60.28±9.53%; n=3 with 18 slices; P=0.0062; R²=0.96).

Agreement of IS determined by LGE and TTC. As shown in Fig. 5, the proportion of IS by LGE and TTC staining was in good correlation, indicating a significant difference

| Method         | TR (msec) | TE (msec) | FA  | Slice thickness (mm) | Reconstructed matrix | FOV  | NEX | TA (sec) | Acquisition time (sec) |
|----------------|-----------|-----------|-----|----------------------|----------------------|------|-----|----------|------------------------|
| LG-FLASH-cine  | 8         | 3.1       | 10  | 1.5                  | 256x256              | 5x5  | 1   | 51       | 51                     |
| Faster T2 mapping | 1500     | 10/20/30  | -   | 1.5                  | 192x192              | 5x5  | 1   | 216      | 300                    |
| FISP-cine      | 5.2       | 2         | 2.5 | 1.5                  | -                    | 5x5  | 1   | 67       | 180                    |

FISP, fast imaging with steady state precession; LG-FLASH, late gadolinium fast low angle shot; TE, echo time; TR, repetition time; FA, fractional anisotropy; NEX, number of excitations; FOV, field of view; TA, time to appearance.
in terms of IS between LGE and TTC staining (47.99±10.56 vs. 46.26±9.94; n=3 with 18 slices; P=0.0064; R²=0.96).

**Figure 3.** Magnetic resonance images obtained by combination of faster T2-mapping and LGE vs. histological examination. IS was clearly denoted as an extensive hyperintense region on (A) LGE, obtained by (B) drawing a region (yellow dotted line), which perfectly matched with (C) the TTC image (blue dotted line). (D) T2-mapping images with three echo times of 10, 20 and 30 sec, yielding (E) T2 value in pseudo color; all T2 maps were scaled between 0 and 80 msec. (F) The AAR was shown on the simplified T2 maps (red dotted line), corresponding to (G) the T2 map (red dotted line) in an overlay of (E) and (F). SMZ was presented between the zones within the red and yellow dotted lines. LGE, late gadolinium enhancement imaging; TTC, 2,3,5-triphenylterazolium chloride staining; AAR, area at risk; IS, infarction size.

**Table 1.** Comparison of IS between LGE and TTC staining.

| Parameter | LGE | TTC |
|-----------|-----|-----|
| IS (mm²)  | 47.99±10.56 | 46.26±9.94 |

**Determination of SMZ by novel faster T2 mapping.** The SMZ size was expressed as the percentage of LV for parameter
standardization and determined using three methods. The SMZ was 13.48±3.64% of the LV (range, 7.72‑20.6%) according to quantification by T2 mapping and 13.04±3.68% of the LV (range, 7.81‑20.5%) as determined by AAR minus IS, which was consistent with 13.68±9.94% of the LV (range, 7.85‑21.5%; n=3 with 18 slices; P=0.0068 and P=0.0067, R²=0.95 and 0.94) as assessed by H&E staining (Fig. 6).

In vivo and ex vivo quantification of IMH. Intramyocardial hemorrhage was observed in three rats from the model group at 24 h. IMH occurred after infarction corresponding to the T2 maps, which were surrounded by IS (green area; Fig. 7A). These characteristics referred to the ‘patchy’ hypointensity in the hyperintensity zone (white dotted line), as illustrated in Fig. 7A. Rough examination of the heart also confirmed the presence of IMH (red dotted line) in Fig. 7B, which was consistent with the findings of H&E staining (blue dotted line) in Fig. 7C, histopathological findings in Fig. 7D (blue dotted line) and E. On T2-mapping images, IMH was characterized as the area of hypointensity in the hyperintensity infarction zone (Fig. 7A), accompanied by the following changes in the T2 value: 35.8±1.52 msec with IMH and 32.2±1.92 msec without IMH (n=3; P=0.035).

Discussion

Myocardial AAR is defined as the incidence of ischemic myocardium after myocardial infarction, which consists of non-viable IS and viable SMZ. Consequently, measurement of AAR and IS allows for the determination of SMZ, which is recognized as a convincing predictor of viable myocardium and therapeutic efficacy after myocardial infarction (8-18).

The evidence from the present study indicated that combined application of in vivo non-contrast T2 mapping by MRI and LGE allows for accurate quantification of multiple ischemic components, including the AAR, IS and SMZ in a rat model. The SMZ detected by novel faster T2 mapping was independently correlated with histological outcomes. In addition, a ‘patchy’ area was found due to the hypointensity attenuation in the hyperintensity infarction-core zone on T2 maps, which was subsequently confirmed by H&E staining.

Ischemic heart disease has been proven to be associated with poor function of LV, and cine MRI provides essential data and pivotal insights into LV function (19). Conventional techniques, such as Doppler or ultrasound and echocardiography in canine models or humans, have been used to assess myocardial function. In the present study, cine MRI provided EDV data of the LV to facilitate the real-time evaluation of the regional function, although the rats exhibited a relatively high heart rate.

The T2 value in the model group was significantly higher compared with that in the sham group. The present as well as a previous study by our group demonstrated no significant difference in the sham group from the base to apex on the T2 maps (15). However, these results were not consistent with the findings by other studies (20), probably due to differences in experimental animals and recruitment of human subjects. In the rat models with myocardial infarction, evaluation of T2 mapping resulted in AAR, IS and SMZ of 38.6±1.52, 42.4±1.67 and 35.8±1.31 msec, corresponding to different T2 values.

The AAR refers to the signs of edema induced by myocardial injury, which includes SMZ and IS. The SMZ is a dynamic parameter and depends upon the delineation of AAR and IS. SMA also serves as a vital predictor of ensuing myocardial infarction (21-23). Conventionally, T2-weighted MRI has been
employed to delineate and assess the AAR after myocardial infarction as a dark blood TSE technique. However, all of these approaches possess disadvantages, such as difficulty in adapting images to respiratory or cardiac motion and the signal associated with surface coil intensity inhomogeneity (24). T2 mapping has been first utilized to assess myocardial edema without the above limitations in 2007 (25,26) and subsequently applied in patients diagnosed with myocardial infarction (27). In the present study, AAR was evaluated by a novel faster T2 mapping with three ETs of 10, 20 and 30 sec, and a T2 value in pseudo color was documented. These results represented an apparent linear correlation between AAR determined by faster T2 mapping and H&E staining.

IS is another pivotal parameter for the evaluation of SMZ following myocardial infarction. LGE has been confirmed to possess a relatively high sensitivity of up to 99% for the evaluation of myocardial infarction, myocardial fibrosis and scars (28). In the present study, IS assessed by LGE was correlated with TTC staining. However, LGE was slightly overestimated in the actual infarction zone, which may explain for the peri-infarction zone being surrounded by edema after acute myocardial infarction, resulting in increased signal intensity. Of note, LGE has several limitations due to gadolinium dispersion, which is dependent on the time of image acquisition and dose of contrast agent (29,30), which was observed in the process throughout the present

Figure 6. SMZ assessed by T2 mapping imaging compared with H&E staining. SMZ, salvaged myocardium control; Remote infarction; H&E, hematoxylin and eosin staining; AAR, area at risk; IS, infarction size; LV, left ventricle.

Figure 7. Assessment of IMH by T2 mapping vs. histological examination. (A) T2 mapping demonstrated that the IMH (white dotted line) was surrounded by IS (green area), corresponding to the ‘patchy’ hypointensity in the hyperintensity zone. (B) Rough examination of the heart tissue confirmed the incidence of IMH (red dotted line), perfectly matching with (C) the H&E staining (blue dotted line), and (D and E) the histological analysis [blue dotted line; magnification, x100 in (D) and x200 in (E)]. H&E, hematoxylin and eosin; IMH, intramyocardial hemorrhage; IS, infarction size.
study. Thus, in the present investigation, the experimental procedures were performed at 10 min after injection of 0.2 mmol/kg Gd-DTPA.

In the present study, three approaches were utilized to determine different parameters of the myocardial size at risk. Previous studies have stated that the SMZ is a time-dependent parameter, considered a vital predictor of poor prognosis induced by ventricular arrhythmia (31,32). According to a mathematical equation, SMZ was calculated by subtracting IS from AAR. The SMZ calculated from the AAR minus the IS determined by LGE was slightly smaller than that assessed by histological examination, probably resulting from the above-mentioned limitation of LGE. However, the SMZ assessed by faster T2 mapping perfectly matched with the findings of H&E staining, suggesting that T2 mapping is a feasible technique to determine SMZ.

IMH is considered a sign of severe injury induced by vascular cell damage (33). By establishing in vivo rat models with cardiac infarction, three out of eight rats presented with IMH on the T2 maps in current study. Faster T2 mapping is capable identifying the imaging characteristics as the ‘striped’ hypointensity in the hyperintensity infarction-core zone. This phenomenon may explain why the T2 value obtained by IMH was significantly lower than that in the infarction-core area and was equivalent to the T2 data in the sham group. Previous investigation has demonstrated that T2-mapping imaging is able to quantitatively measure the T2 value, which can distinguish various ischemic components (34). In the present study, T2 value in the infarction zone with IMH was significantly lower compared with that in the infarction area without IMH, which was subsequently validated by H&E staining outcomes. Consequently, faster T2-mapping imaging is a feasible technique to quantitatively measure IMH in a rat model with myocardial infarction. However, IMH is merely one component of varying characteristics after myocardial infarction (20), and alternative components, such as inflammation and myocardial vascular obstruction (35,36), should be urgently explored by more investigations.

The present study had several limitations that should be acknowledged. First, the myocardial infarction model was assessed at only one time-point. In addition, the sample size was relatively small. Furthermore, rats with a high heart rate were examined under mechanically ventilated conditions and extrapolation of the results to human subjects should be interpreted with caution. Third, all ROIs for different segments of T2 values and myocardial infarction components were determined by two radiologists, which may have induced manual bias. It was difficult to visualize the color-coded T2 maps. Combination with other methods of CMR should therefore be recommended for detecting tissue changes after myocardial infarction.

Taken together, faster T2-mapping imaging is suitable for visualizing and quantifying myocardial AAR, IS and SMZ, and for evaluating the severity of IMH in a rat model on 7T MRI. This technique serves as a reliable non-invasive method to quantitatively assess the SMZ and IMH, contributing to monitor the viable myocardium during gene or stem cell therapy after myocardial infarction. Moreover, the mechanism of IMH will be further investigated in our subsequent research.

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