After 7-valent pneumococcal conjugate vaccine (PCV) for children was introduced in Japan in November 2010, we examined changes in Streptococcus pneumoniae serotypes and in genetic antimicrobial drug resistance of isolates from adults with invasive pneumococcal diseases. During April 2010–March 2013, a total of 715 isolates were collected from adults with invasive pneumococcal diseases. Seven-valent PCV serotypes in adults decreased from 43.3% to 23.8%, most noticeably for serotype 6B. Concomitantly, 23-valent pneumococcal polysaccharide vaccine (PPSV23) serotypes decreased from 82.2% to 72.2%; non-PPSV23 serotypes increased from 13.8% to 25.1%. Parallel with serotype changes, genotypic penicillin-resistant S. pneumoniae decreased from 32.4% to 21.1%, and 6 non-PPSV23 serotypes emerged (6D, 15A, 15C, 16F, 23A, and 35B). Respective vaccine coverage rates for 13-valent PCV and PPSV23 differed by disease: 73.9% and 84.3% for patients with pneumonia, 56.4% and 69.2% for patients with bacteremia and sepsis, and 45.7% and 69.3% for patients with meningitis.

Streptococcus pneumoniae can cause invasive pneumococcal diseases (IPDs), which can be treated with generally effective antimicrobial drugs. In adults, well-known risk factors for IPDs include underlying conditions such as splenectomy, functional asplenia, immunodeficiency, and HIV infection. Risk is also increased with age >65 years, diabetes, renal dialysis, and chronic hepatic dysfunction (1,2).

In 2010, the US Advisory Committee on Immunization Practices (ACIP) issued recommendations that all persons >65 years of age should be vaccinated with 23-valent pneumococcal polysaccharide vaccine (PPSV23) to prevent pneumococcal diseases (3). In 2012, the ACIP also recommended that 13-valent pneumococcal conjugate vaccine (PCV13), which the US Food and Drug Administration approved in 2011, be given to adults ≥19 years of age if they have immunocompromising conditions (4). Recently, the ACIP recommended routine administration of both PCV13 and PPSV23 to all adults ≥65 years of age (5).

In the United States, 7-valent pneumococcal conjugate vaccine (PCV7) has been used to vaccinate children since 2000, resulting in individual and herd immunity and a decline in pneumococcal infection in children (6–9). After PCV7 was introduced, serotype 19A pneumococcal strains with penicillin resistance increased (8–10); the change in 2012 to PCV13 covers serotype 19A (11).

Presently, pneumococcal conjugate vaccines (PCVs) are incorporated into pediatric vaccination schedules in >120 countries, and PCV13 has been approved for adults in >100 countries. Pneumococcal infections in adults have decreased as an indirect effect of widespread use of PCVs for children (2,6,8,9,12). However, in countries where PCV7 or PCV13 was introduced, overall coverage of serotypes by the vaccine gradually decreased because of pneumococcal serotype replacement from vaccine type to nonvaccine type. In particular, increases of nonvaccine serotypes, such as 6C, 15A, 23A, and 35B, have been reported in the United States (13,14) and other countries (15–18).

In Japan, PCV7 vaccination was introduced for children <5 years of age in November 2010 by the Provisional Fund for the Urgent Promotion of Vaccination. PCV7 was incorporated into routine vaccination schedules in April 2013 and replaced by PCV13 in November 2013. PCV7 rapidly decreased IPD infections in children. However, pneumococcal infections caused by non-PCV7 serotypes (19A, 15A, 15B, 15C, and 24) increased in children during 2012 (19).

Meanwhile, Japan became the first “super-aging” society in the world in 2013, when 25.1% of the nation’s
population was ≥65 years of age (Statistics Bureau, Ministry of Internal Affairs and Communications, http://www.stat.go.jp/english/data/handbook/c0117.htm). Lifestyle-related and other diseases that affect response to infection have increased in older adults, and pneumonia has again become the third leading cause of death in Japan. Nevertheless, the voluntary PPSV23 immunization rate among older persons long remained <18%. In November 2014, the Japanese Ministry of Health, Labour and Welfare began promoting routine vaccination with PPSV23 for adults ≥65 years of age.

Because of these developments, we sought to clarify changes in serotypes and in genetic antimicrobial resistance in isolates from adults with IPDs after PCV7 for children was introduced in Japan. We describe capsular serotypes, drug-resistance genotypes, and multilocus sequence typing in isolates from adults with IPDs, and we examine the relationships of these factors to specific IPDs.

**Methods**

**Patients and Pneumococcal Strains**

Our study included adult patients (≥19 years of age) with IPD. Pneumococcal isolates from sterile clinical samples such as blood, cerebrospinal fluid, pleural effusion, and joint fluid were collected from clinical laboratories at 341 hospitals participating in this IPD surveillance study. Each hospital had a microbiology laboratory, and the hospitals were distributed nearly uniformly throughout Japan. The hospitals participated in the surveillance project after written permission was obtained from laboratory or hospital directors.

From April 2010 through March 2013, a total of 715 pneumococcal isolates were collected for the study: 275 during April 2010–March 2011, the first surveillance period; 213 during April 2011–March 2012, the second period; and 227 during April 2012–March 2013, the third period. This large-scale IPD surveillance for adults was performed in conjunction with IPD surveillance for children (19). During the first surveillance period, the rate of voluntary PCV7 vaccination for children was ≈10%. The second period occurred simultaneously with the Urgent Promotion of Vaccination Incentive for children, and the immunization rate was ≈50%–60%. The third period occurred just before the transition from PCV7 to PCV13 for children, and the PCV7 immunization rate was ≈80%–90%. However, the rate of voluntary vaccination with PPSV23 in adults remained <18% throughout the surveillance periods.

The collected pneumococcal isolates were sent promptly from each clinical laboratory to our laboratory (Molecular Epidemiology for Infectious Agents, Kitasato Institute for Life Sciences, Kitasato University, Tokyo, Japan), accompanied by survey forms completed by attending physicians but not identifying patients in compliance with ethical guidelines for epidemiology in Japan. The surveys collected information on patient’s age at illness onset, sex, clinical manifestations, blood test results at hospitalization, and outcome. Clinical manifestations and diagnoses were verified in all patients, according to diagnostic criteria and guidelines for sepsis and related conditions (20).

**Serotypes and Antimicrobial Drug–Resistant Genotypes**

Serotypes were determined by the capsular quellung reaction by using antiserum purchased from the Statens Serum Institute (Copenhagen, Denmark). By using real-time PCR methods described previously (21), alterations were identified in 3 penicillin-binding protein (PBP) genes mediating β-lactam resistance in *S. pneumoniae*: *pbp1a* (encoding enzyme PBP1A), *pbp2x* (encoding PBP2X), and *pbp2b* (encoding PBP2B). Amino acid substitutions within or near each PBP’s conserved amino acid motifs, such as serine-threonine (Thr)–methionine-lysine (STMK), were detected by using real-time PCR. The *mef* (A) and *erm* (B) genes mediating macrolide resistance were also identified by real-time PCR (21). Quinolone resistance was analyzed by sequencing the quinolone-resistance–determining region in the genes *gyrA*, *gyrB*, *parC*, and *parE* in strains with MICs of levofloxacin of >4 μg/mL (22).

Gene analysis identified genotypes (g) on the basis of their responses to β-lactam antimicrobial drugs: penicillin-susceptible *S. pneumoniae* (gPSSP) possessing 3 normal *pbp* genes; penicillin-intermediate *S. pneumoniae* (gPISP), also classified as gPISP (pbp2x), gPISP (pbp2b), gPISP (pbp1a+pbp2x), or gPISP (pbp2x+pbp2b); and penicillin-resistant *S. pneumoniae* (gPRSP), which possessed all 3 abnormal *pbp* genes (19,21). Macrolide-resistant genotypes were also represented variously: macrolide-susceptible *S. pneumoniae* that possessed no resistance genes; macrolide resistance mediated by the *mef*(A) gene, MLR-*mef*(A); macrolide resistance mediated by the *erm*(B) gene, MLR-*erm*(B); and macrolide resistance involving both genes, MLR-*mef*(A)+*erm*(B).

Identification of capsular type by the quelling reaction and resistance genotyping by real-time PCR was performed within 1 day of sample collection for each strain. Results were reported immediately to the medical staff at the referring hospital.

**Antimicrobial Susceptibility Testing**

MICs of 7 intravenous antimicrobial agents (penicillin, ampicillin, cefotaxime, ceftriaxone, meropenem, panipenem, and vancomycin) were determined for all pneumococcal isolates by agar-dilution methods (23). We obtained each agent from the respective manufacturer.
Multilocus Sequence Typing
Multilocus sequence typing (MLST; http://www.MLST.net) was performed for nonvaccine serotype strains, except for PPSV23 serotypes, according to previously described methods with minor modifications (24). Primers used were based on sequences listed by the US Centers for Disease Control and Prevention (http://www.cdc.gov/ncidod/biotech/strep/alt-MLST-primers.htm). Analyses by using MLST and eBURST (Department of Infectious Disease Epidemiology, Imperial College of London, UK) were performed, as described by the MLST website (http://spneumoniae.mlst.net).

Statistical Analysis
We assessed differences in serotypes by age groups, clinical symptoms and signs, resistance types, and surveillance periods. To determine whether differences were statistically significant, we performed χ² tests or the Fisher exact test by using Ekuseru-Toukei 2012 software for statistics (Social Survey Research Information, Tokyo, Japan).

Results
Yearly Changes in Pneumococcal Serotypes
During each annual surveillance period (April–March) during 2010–2013, pneumococcal serotypes from IPD isolates varied (Table 1). All strains were isolated from sterile samples, such as blood, cerebrospinal fluid, pleural effusion, or joint fluid.

During all surveillance periods, the estimated voluntary inoculation rate for PPSV23 in adults was <18% in Japan. In February 2010, the vaccination incentive for PCV7 in children began on a voluntary basis. In November 2010, the Special Fund for the Urgent Promotion of Vaccination was initiated, and PCV7 vaccination of children <5 years of age became an official priority throughout

**Table 1. Pneumococcal serotypes in isolates from adult patients with invasive pneumococcal diseases, Japan, April 2010–March 2013***

| Vaccine serotype | 2010–2011, no. (%) | 2011–2012, no. (%) | 2012–2013, no. (%) | p value† |
|------------------|-------------------|-------------------|-------------------|---------|
| PCV7             |                   |                   |                   |         |
| 4                | 14 (5.1)          | 17 (8.0)          | 7 (3.1)           | 0.075   |
| 6B               | 41 (14.9)         | 24 (11.3)         | 12 (5.3)          | 0.003   |
| 9V               | 7 (2.5)           | 5 (2.3)           | 1 (0.4)           | 0.168   |
| 14               | 21 (7.6)          | 16 (7.5)          | 17 (7.5)          | 0.998   |
| 18C              | 1 (0.4)           | 1 (0.5)           | 1 (0.4)           | ND      |
| 19F              | 14 (5.1)          | 8 (3.8)           | 7 (3.1)           | 0.511   |
| 23F              | 21 (7.6)          | 13 (6.1)          | 9 (4.0)           | 0.232   |
| Total            | 119 (43.3)        | 84 (39.4)         | 54 (23.8)         | <0.001  |
| PCV13            |                   |                   |                   |         |
| 1                | 1 (0.4)           | 1 (0.5)           | 0                 | ND      |
| 3                | 45 (16.4)         | 27 (12.7)         | 42 (18.5)         | 0.226   |
| 5                | 0                 | 0                 | 1 (0.4)           | ND      |
| 6A               | 11 (4.0)          | 4 (1.9)           | 6 (2.6)           | 0.367   |
| 7F               | 9 (3.3)           | 2 (0.9)           | 3 (1.3)           | 0.128   |
| 19A              | 18 (6.5)          | 11 (5.2)          | 17 (7.5)          | 0.593   |
| Total            | 203 (73.8)        | 129 (60.6)        | 123 (54.2)        | <0.001  |
| PPSV23           |                   |                   |                   |         |
| 10A              | 10 (3.6)          | 8 (3.8)           | 11 (4.8)          | 0.754   |
| 11A              | 3 (1.1)           | 8 (3.8)           | 5 (2.2)           | 0.142   |
| 15B              | 3 (1.1)           | 4 (1.9)           | 5 (2.2)           | 0.605   |
| 22F              | 10 (3.6)          | 19 (8.9)          | 18 (7.9)          | 0.040   |
| Other‡           | 8 (2.9)           | 1 (0.5)           | 8 (3.5)           | 0.082   |
| Total            | 226 (82.2)        | 165 (77.5)        | 164 (72.2)        | 0.036   |
| Nonvaccine serotype |                 |                   |                   |         |
| 6C               | 13 (4.7)          | 12 (5.6)          | 17 (7.5)          | 0.407   |
| 15A              | 6 (2.2)           | 10 (4.7)          | 8 (3.5)           | 0.310   |
| 15C              | 0                 | 4 (1.9)           | 4 (1.8)           | ND      |
| 23A              | 2 (0.7)           | 8 (3.8)           | 8 (3.5)           | 0.053   |
| 35B              | 7 (2.5)           | 6 (2.8)           | 9 (4.0)           | 0.626   |
| 37               | 3 (1.1)           | 0                 | 1 (0.4)           | ND      |
| 38               | 3 (1.1)           | 2 (0.9)           | 3 (1.3)           | 0.928   |
| Other§           | 3 (1.1)           | 2 (0.9)           | 5 (2.2)           | 0.454   |
| Total            | 37 (13.5)         | 44 (20.7)         | 55 (24.2)         | 0.007   |
| Nontypeable¶     | 1 (0.4)           | 0                 | 2 (0.9)           |         |

*Years run from April 1 to March 31 of the following year. ND, not determined because of a low number of strains.
†p values characterize comparisons of 2010–2011, 2011–2012, and 2012–2013.
‡Serotypes 12F, 20, and 33F.
§Serotypes 6D, 7C, 16F, 18B, 24F, and 34.
¶These serotypes were excluded from the statistical analysis because they were unclassifiable.
Japan. Estimates of the PCV7 immunization rate in children was <10% in 2010, 50%–60% in 2011, and 80%–90% in 2012 (19).

PPSV23 coverage of serotypes found in IPD isolates decreased significantly each year, from 82.2% in 2010 to 77.5% in 2011 to 72.2% in 2012 (p = 0.036). Respective coverages with PCV7 and PCV13 also decreased significantly, from 43.3% and 73.8% in 2010 to 39.4% and 60.6% in 2011 to 23.8% and 54.2% in 2012, respectively (p<0.001). Although serotype 22F increased significantly in IPD isolates, yearly decreases of serotype 6B contributed markedly to decreases in PPSV23 coverage. Nonvaccine serotypes (i.e., non-PPSV23) increased significantly during the surveillance periods from 13.5% in 2010 to 20.7% in 2011 to 24.2% in 2012 (p = 0.007).

Introduction of PCV7 in children indirectly affected adults with IPD, shown by changes in each serotype during 2010–2012 (online Technical Appendix Figure 1, http://wwwnc.cdc.gov/EID/article/21/11/14-2029-Techapp.pdf). Decreases occurred in PCV7 serotypes and serotype 6A, the latter of which exhibits cross-immunity with 6B, but almost all other serotypes except those in PCV13 increased, especially serotypes 22F, 6C, 23A, 15C, 15A, and 35B.

**Yearly Changes in Genotypes Resistant to Penicillin, Macrolides, and Quinolones**

Genotypes resistant to penicillin, macrolides, and quinolones varied during the surveillance periods (Table 2). Among genotypes resistant to penicillin, 40% were gPISP (pbp2x), 26% were gPRSP, 10.2% were gPISP (pbp1a+pbp2x), and 5.9% were gPISP (pbp2x+pbp2b). Only 16.1% were gPSSP strains without any abnormal pbp genes. Other genotypes were gPISP (pbp2b; 1.3%) and gPISP (pbp1a+2b; 0.6%). In contrast to a gradual increase in gPISP (pbp2x+pbp2b; p = 0.025), gPRSP decreased significantly over the surveillance periods (p = 0.008). In addition, 89.7% of all pneumococcal strains possessed mef(A) or erm(B) genes that mediated macrolide resistance: 23.6% carried the mef(A) gene mediating intermediate macrolide resistance; 56.9% carried the erm(B) gene mediating high macrolide resistance; and 9.1% possessed both genes. Macrolide-susceptible strains accounted for only 10.3%. Furthermore, only 0.7% of strains were highly resistant to levofloxacin (MIC >16 μg/mL) and possessed mutations in both gyrA and parC genes. Intermediate-resistance strains (MIC 4 μg/mL) that possessed the parC mutation accounted for 1.3% of strains.

**Yearly Changes in Resistance Genotypes and Serotypes**

Yearly changes in genotypes and serotypes affecting β-lactam resistance also occurred during the 3 periods (Figure). Decreases closely related to a reduction of serotypes 6B, 19F, and 23F, but not of serotype 14, were found for a number of gPRSP. Emergence of gPRSP was evident among 6 non-PPSV23 serotypes (6D, 15A, 15C, 16F, 23A, and 35B), although such resistance strains were few. Almost all non-PPSV23 serotype strains showed macrolide resistance (online Technical Appendix Figure 2), which did not change significantly from year to year (Table 2).

**Invasive Pneumococcal Diseases and Patient Age**

To examine relationships between IPDs and patient age at illness onset, we classified IPDs into 4 categories (Table 3): pneumonia (n = 375), including cases of empyema (n = 17) and pleuritis (n = 16); bacteremia and sepsis with no obvious focus (n = 172), including sepsis (n = 51), severe

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**Table 2. Isolates positive for invasive pneumococcal diseases and resistant to penicillin, macrolide, and quinolone, by genotype, Japan, April 2010–March 2013**

| Genotype        | Total, no. (%) | 2010–2011, no. (%) | 2011–2012, no. (%) | 2012–2013, no. (%) | p value†  |
|-----------------|----------------|-------------------|-------------------|-------------------|-----------|
| Penicillin resistance |                |                   |                   |                   |           |
| gPISP           | 115 (16.1)     | 42 (15.3)         | 40 (18.8)         | 33 (14.5)         | 0.431     |
| gPISP (pbp2x)   | 286 (40.0)     | 96 (34.9)         | 89 (41.8)         | 101 (44.5)        | 0.076     |
| gPISP (pbp2b)   | 9 (1.3)        | 5 (1.8)           | 0                 | 4 (1.6)           | ND        |
| gPISP (pbp1a+2x)| 73 (10.2)      | 30 (10.9)         | 23 (10.8)         | 20 (8.8)          | 0.701     |
| gPISP (pbp1a+2b)| 4 (0.6)        | 3 (1.1)           | 1 (0.5)           | 0                 | ND        |
| gPISP (pbp2x+2b)| 42 (5.9)       | 10 (3.6)          | 11 (5.2)          | 21 (9.3)          | 0.025     |
| gPRSP (pbp1a+2x+2b)| 186 (26.0)    | 89 (32.4)         | 49 (23.0)         | 48 (21.1)         | 0.008     |
| Macrolide resistance |              |                   |                   |                   |           |
| Resistance gene (-) |              |                   |                   |                   |           |
| mef(A) gene     | 74 (10.3)      | 34 (12.4)         | 16 (7.5)          | 24 (10.6)         | 0.216     |
| erm(B) gene     | 169 (23.6)     | 63 (22.9)         | 56 (26.3)         | 50 (22.0)         | 0.538     |
| mef(A) and erm(B) | 407 (56.9)    | 152 (55.3)        | 123 (57.7)        | 132 (58.1)        | 0.777     |
| Quinolone resistance‡ |             |                   |                   |                   |           |
| gyrA+parC       | 5 (0.7)        | 2 (0.7)           | 3 (1.4)           | 0                 | ND        |
| parC            | 9 (1.3)        | 5 (1.8)           | 1 (0.5)           | 3 (1.3)           | ND        |

†p values characterize comparison of the 3 surveillance periods.
‡Quinolone resistance, Ser81Phe substitution in GyrA and Ser79Tyr in ParC, respectively.
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sepsis (n = 57), and septic shock (n = 24); meningitis (n = 127); and others (n = 41), including cellulitis (n = 7), arthritis (n = 6), spondylitis (n = 4), and peritonitis (n = 3). Pneumonia (52.4%) was the most common category of IPDs, followed by bacteremia and sepsis (24.1%) and meningitis (17.8%). Mean age of patients with meningitis was 65 years, younger than those with pneumonia or bacteremia and sepsis (mean age 74 and 70 years, respectively). Meningitis occurred significantly more often among patients <64 years of age, in contrast to pneumonia, which occurred most often in persons ≥75 years of age (p<0.001).

Data for deaths and survival were available for 656 (91.7%) of the 715 patients. Proportions of deaths occurring within 30 days of hospitalization increased significantly with age: 6.0% at 19–49 years, 17.8% at 50–64 years, 17.9% at 65–74 years, 25.4% at 75–84 years, and 26.3% at ≥85 years (p = 0.002). Overall rate of death occurring within 30 days after hospitalization was 21.3%; overall rate for serious sequelae was 10%.

Serotype Distribution by Disease

A comparison of proportions of serotypes included in PCV7, PCV13, PPSV23, and non-PPSV23 by disease category (i.e., pneumonia, bacteremia and sepsis, meningitis, and others) showed that frequency of serotypes differed considerably by type of IPD (Table 4). Prevalence of serotypes 3, 14, and 19A was higher in isolates of patients with pneumonia than in those from patients with other illnesses (p value range <0.001–0.076). In contrast, serotypes 10A, 15A, 23A, and 23F were significantly more prevalent in isolates of patients with meningitis than in those from patients with other illnesses (p value range <0.001–0.004).

Coverages for vaccines among all isolates collected and examined during the study duration were 35.5% for PCV7, 63.6% for PCV13, and 77.6% for PPSV23. Respective coverages of PCV13 and PPSV23 differed significantly for each IPD group: 73.9% and 84.3% for pneumonia, 56.4% and 69.2% for bacteremia and sepsis, 45.7% and 69.3% for meningitis, and 56.1% and 78.0% for others (p<0.001 for

Figure. Yearly changes in number of serotypes and in penicillin resistance in genotypes found in isolates from adults with invasive pneumococcal diseases, Japan, April 2010–March 2013. Serotypes are shown for each of the 3 yearly surveillance periods: April 2010–March 2011, April 2011–March 2012, and April 2012–March 2013. Short tic marks on horizontal axis represent yearly number of isolates for specific serotypes; longer tic marks represent the 3-year surveillance period for each serotype. gPSSP, genotypic penicillin-susceptible Streptococcus pneumoniae; gPISP, genotypic penicillin-intermediate resistant S. pneumoniae; gPRSP, genotypic penicillin-resistant S. pneumoniae. Parentheses enclose abnormal pbp genes that mediate penicillin resistance.
The most prevalent serotype was serotype 3 (15.9%), followed by serotypes 6B (10.8%), 14 (7.6%), 22F (6.6%), 19A (6.4%), and 23F (6.0%). Death rates did not differ significantly by pneumococcal serotype.

Antimicrobial Drug Susceptibility by Genotype

The online Technical Appendix Table shows susceptibilities (50% MIC, 90% MIC, and MIC range) of 6 parenteral β-lactam agents for *S. pneumoniae* strains (n = 710). PBP gene alterations affect MICs of penicillin, ampicillin, cefotaxime, and meropenem (online Technical Appendix Figure 3). Most strains in this study were gPRSP with *pbp2x* gene alterations that reduced susceptibility to cephalosporin agents rather than to penicillin. MICs of cefotaxime and ceftriaxone were affected more by *pbp2x* alterations than by *pbp2a* alterations. In contrast, MICs of penicillin, ampicillin, and meropenem were affected more by *pbp2b* alterations than by *pbp2x* alterations. Strains showing MICs ≥4μg/mL to cefotaxime had *pbp2a* gene alteration and also had *pbp1a*, *pbp2x*, and *pbp2b* gene alterations (data not shown).

MLST in Non-PCV13 Serotype Strains

MLST results for 92 strains (non-PCV13 serotypes) collected during 2012 and results for 2 gPRSPs (serotypes 15B collected during 2010 and 23A collected during 2011) indicate that sequence types (STs), 8 of which were new, and clonal complexes (CCs) varied (Table 5). We identified 3 new STs of gPRSPs possessing macrolide-resistant genes: ST6138 (CC81) in serotype 15C, ST8351 (CC3117) in serotype 16F, and ST9619 (CC156) in serotype 23A. Also, we identified gPRSPs of ST282 (CC81) in serotype 6D, reported from South Korea in 2008 and derived from Vietnam serotype 23F in 1996; ST63 (CC63) in serotype 15A, derived from Sweden\(^{13,14,16–18,31}\) in 1992; ST83 (CC81) in serotype 15B, reported from South Korea in 2007 and derived from Taiwan serotype 23F in 1997; and ST558 (CC558) in serotype 35B, reported from the United States in 1999 (Table 5). We confirmed that all gPRSPs examined in our study had abnormal PBPs by DNA sequencing: substitutions of Thr371Serine or Thr371Alanine (Ala) within conserved amino acid motif STMK in PBP1A; Thr338Ala within STMK in PBP2X; and Thr445Ala adjacent to the serine-serine-asparagine amino acid motif in PBP2B, respectively.

**Table 3.** Isolates positive for invasive pneumococcal diseases in adults, by age group, Japan, April 2010–March 2013

| Disease                | No. (%) | Mean age, y (±SD) | p value |
|------------------------|---------|------------------|---------|
| Pneumonia*             | 375 (52.4) | 33 (39.8) | 66 (42.0) | 103 (49.8) | 105 (60.7) | 68 (71.6) | 74 (14.9) | <0.001 |
| Bacteremia and sepsis† | 172 (24.1) | 20 (24.1) | 42 (28.8) | 55 (26.6) | 34 (19.7) | 12 (20.7) | 20 (6.3) | 65 (14.4) | <0.001 |
| Meningitis             | 127 (17.8) | 24 (28.9) | 38 (24.2) | 37 (17.9) | 22 (12.7) | 6 (3.9) | 0 | 67 (14.5) | 0.133 |
| Other‡                 | 41 (5.7) | 6 (7.2) | 11 (7.0) | 12 (5.8) | 12 (6.9) | 0 | 67 (14.5) | 0.133 |
| Total                  | 715 (49) | 83 (15.7) | 207 (22.1) | 173 (26.6) | 95 (14.9) | 70 (14.9) | 70 (14.9) | 70 (14.9) | 70 (14.9) |

*Includes empyema (n = 17) and pleuritis (n = 16).
†Bacteremia (n = 39), sepsis (n = 51), severe sepsis (n = 58), and septic shock (n = 24).
‡Cellulitis (n = 7), arthritis (n = 6), spondylitis (n = 4), peritonitis (n = 3), and other conditions.
vaccine implementation (19), but the non-PCV7 serotypes 19A, 15A, 15B, 15C, and 24 increased each year. These results reflect simultaneous replacement of PCV7 serotypes with non-PCV7 serotypes among pneumococci that colonize the nasopharynx of children (K. Ubukata et al., unpublished data). Consequently, frequency of transmission of PCV7 serotypes from children to adults would be expected to decrease.

In this study, we found epidemiologic changes among pneumococcal strains isolated from adult patients with IPD; vaccine-type strains decreased significantly (6–9). However, the prominent decrease in serotype 6B detected by our surveillance differed from findings in the United States, where serotype 14 decreased most prominently after PCV7 introduction (14). IPD cases in Japan resulting from serotype 14 decreased significantly for adults during our ongoing surveillance in 2014. Given Japan’s high population density, effects of herd immunity were likely present in children and adults. However, non-PCV7 types, especially serotype 22F, increased.

A decrease in penicillin-nonsusceptible S. pneumoniae strains has been associated with serotype changes occurring after vaccine introduction (29,30,32). In our previous genetic studies, the gPRSP with 3 PBP alterations encoded by the \( pbp1a \), \( pbp2x \), and \( pbp2b \) genes also decreased significantly (19). However, our current study found increases in the gPISP with \( pbp2x \) gene alterations that reduce susceptibilities to cephalosporin agents (e.g., cefotaxime and ceftriaxone), rather than reducing susceptibility to penicillin. This trend likely reflects preference among medical practices in Japan for cephalosporin and macrolide agents.

We also identified some gPRSPs not previously reported in Japan, such as ST63 (CC63) of serotype 35B, identified in the United States (33); ST63 (CC63) of serotype 15A derived from Sweden (14, 35); ST282 (CC81) of serotype 6D in South Korea (34, 35); and ST83 (CC81)
of serotype 15B, originating in Taiwan serotype 23F. The
remaining 3 STs identified in gPRSP, serotypes 15C, 16F,
and 23A, appear to be novel STs arising in Japan. ST6138
of serotype 15C and ST9619 of serotype 23A were esti-
ated to be extended from ST83 and ST338 (Pneumococcal
Molecular Epidemiology Network clone Colombia23F-26),
respectively (36).

Recent pneumococcal genomic analysis (37,38) indicated
occurrence of capsule switching by genetic recom-
bination among strains of different serotypes. Recombina-
tion of regions consisting of the capsule cps gene locus
and 2 adjacent pbp1a and pbp2x genes results in capsular
switching and penicillin resistance. A pneumococcal strain
arising from such an event will show high penicillin re-
sistance from mutations in pbp genes because of selection
pressure from antimicrobial drugs. MLST and genotypic
results of gPRSPs identified in non-PCV13 strains in this
study support occurrence of such genetic events.

In relation to serotypes and IPDs in adults, PPSV23
coverage has been reported to differ by disease (45.7% and
69.3% for patients with meningitis. This find-
ing supports recommendations by the ACIP (5) for routine
vaccinations with both PCV13 and PPSV23 in all adults
≥65 years of age, especially those with serious underlying

| Serotype (total no. isolates) | No. isolates, N = 92 | Penicillin resistance genotype | Macrolide resistance gene | Clonal complex | ST | Earliest report of same ST† Year | Country (city) |
|-------------------------------|----------------|--------------------------------|---------------------------|---------------|---|-------------------|---------------|
| 6C (17)                       |                | gPISP(pbp2x+2b)                | erm(B)                    |               | 315 | 386               | 1996          | Portugal (Lisbon)             |
|                               |                | gPISP(pbp2x)                   | erm(B)                    |               | 2924 | 2924              | 2003          | Japan (Hyogo)                  |
|                               |                | gPISP(pbp2x)                   | erm(B)                    |               | 2924 | 6183              | 2008          | Japan (Kanagawa)               |
|                               |                | gPISP(pbp2x)                   | erm(B)                    |               | 2924 | 2017              | 2008          | Japan (Fukuda)                 |
|                               |                | gPISP(pbp2x+2b)                | erm(B)                    |               | 156  | 5241              | 2008          | Japan (Kumamoto)               |
|                               |                | gPISP(pbp2x+2b)                | mef(A) or erm(B)          |               | 5832 | 5025‡             | 2012          | Japan (Kanagawa)               |
|                               |                | gPISP(pbp2x+2b)                | mef(A) or erm(B)          |               | 5832 | 5025‡             | 2009          | Japan (China)                  |
|                               |                | gPISP(pbp2x+2b)                | mef(A) or erm(B)          |               | 5832 | 5025‡             | 2009          | Japan (China)                  |
| 6D (1)                        |                | gPRSP                          | mef(A)                    |               | 81   | 282               | 1996;         | Vietnam [23F]; South Korea [6D] |
| 10A (11)                      |                | gPISP(pbp2x)                   | Non                       |               | 156  | 1263              | 1998          | USA (Arizona/New Mexico)       |
|                               |                | gPISP(pbp2x)                   | Non                       |               | 156  | 3395‡             | 2012          | Japan (Toyama)                 |
|                               |                | gPISP(pbp2x)                   | erm(B)                    |               | 113  | 5236              | 2007          | Japan (Kanagawa)               |
|                               |                | gPISP(pbp1a+2x)                | erm(B)                    |               | 113  | 3078‡             | 2012          | Japan (Yamaguchi)              |
| 11A (5)                       |                | gPISP(pbp1a+2x)                | erm(B)                    |               | 63   | 63                | 1992          | [Sweden15A-25]                 |
|                               |                | gPRSP                          | erm(B)                    |               | 63   | 63                | 1992          | [Sweden15A-25]                 |
| 15B (6)                       |                | gPISP(pbp2x)                   | mef(A)                    |               | 199  | 199               | 1987          | [Netherlands15A-37]            |
|                               |                | gPISP(pbp2x)                   | mef(A)                    |               | 199  | 199               | 1987          | [Netherlands15A-37]            |
|                               |                | gPISP(pbp2x)                   | mef(A)                    |               | 199  | 5609‡             | 2012          | Japan (Tokyo)                  |
|                               |                | gPRSP                          | mef(A)                    |               | 81   | 83                | 1997          | Taiwan [23F]                   |
|                               |                | gPRSP                          | mef(A)                    |               | 81   | 6138‡             | 2013          | Japan (Tokyo)                  |
| 15C (4)                       |                | gPISP(pbp2x)                   | mef(A)                    |               | 199  | 199               | 1987          | [Netherlands15A-37]            |
|                               |                | gPISP(pbp1a+2x)                | mef(A)                    |               | 199  | 199               | 1987          | [Netherlands15A-37]            |
|                               |                | gPRSP                          | mef(A)                    |               | 81   | 6138‡             | 2013          | Japan (Tokyo)                  |
| 16F (1)                       |                | gPRSP                          | mef(A)                    |               | 3117 | 8351‡             | 2011          | Japan (Mie)                    |
| 20 (3)                        |                | gPISP(pbp2x)                   | erm(B)                    |               | 4745 | 4745              | 2005          | China (Wuhan, Hubel)           |
|                               |                | gPISP(pbp1a+2x)                | erm(B)                    |               | 433  | 433               | 1997          | Poland (center)                |
|                               |                | gPRSP                          | mef(A) or erm(B)          |               | 433  | 433               | 1997          | Poland (center)                |
|                               |                | gPRSP                          | mef(A)                    |               | 113  | 5236              | 2007          | Japan (Kanagawa)               |
|                               |                | gPRSP                          | mef(A)                    |               | 113  | 7158              | 2007          | Japan (Aichi)                  |
| 23A (9)                       |                | gPISP(pbp2x+2b)                | erm(B)                    |               | 156  | 338               | 1995          | Japan (Kumamoto)               |
|                               |                | gPISP(pbp2x+2b)                | erm(B)                    |               | 156  | 5242              | 2009          | Japan (Kumamoto)               |
|                               |                | gPISP(pbp2x+2b)                | erm(B)                    |               | 156  | 5246              | 2008          | Japan (Niigata)                |
|                               |                | gPRSP                          | erm(B)                    |               | 156  | 9619‡             | 2011          | Japan (Toyama)                 |
| 35B (9)                       |                | gPISP(pbp2x)                   | erm(B)                    |               | 1816 | 2755              | 2004          | China (Shanghai)               |
|                               |                | gPISP(pbp2x)                   | mef(A) or erm(B)          |               | 1816 | 2755              | 2004          | China (Shanghai)               |
|                               |                | gPRSP                          | Non                       |               | 558  | 558               | 1999          | USA (New York) [35B]           |
|                               |                | gPRSP                          | mef(A) or erm(B)          |               | 558  | 558               | 1999          | USA (New York) [35B]           |

*PCV13, 13-valent pneumococcal conjugate vaccine; ST, sequence type. Brackets in final column represent clones reported in the Pneumococcal Molecular Epidemiology Network Clone Collection (http://web1.sph.emory.edu/PMEN/index.html).
†Earliest year and first location are shown for the applicable ST registered in the Multilocus Sequence Typing website (http://spneumoniae.mlst.net).
‡Identified as a new ST in this study.
diseases. For pneumococci with a large number of capsular types, development of a novel vaccine against components other than the capsule is expected (40). Overuse of antimicrobial agents should be avoided worldwide to avoid selection of novel resistant strains capable of undermining vaccine efficacy.

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References
1. Alanee SR, McGee L, Jackson D, Chiou CC, Feldman C, Morris AJ, et al. Association of serotypes of Streptococcus pneumoniae with disease severity and outcome in adults: an international study. Clin Infect Dis. 2007;45:46–51. http://dx.doi.org/10.1086/518538
2. Muhammad RD, Oza-Frank R, Zell E, Link-Gelles R, Butler JC, et al. Antimicrobial susceptibility and serotype distribution of Streptococcus pneumoniae in New York City. Antimicrob Agents Chemother. 1996;40:2321–6.
3. Centers for Disease Control and Prevention. Geographic variation in invasive pneumococcal disease in the United States. Clin Infect Dis. 2011;53:137–43. http://dx.doi.org/10.1093/cid/cir326
11. Centers for Disease Control and Prevention. Licensure of a 13-valent pneumococcal conjugate vaccine (PCV13) and recommendations for use among children—Advisory Committee on Immunization Practices (ACIP). MMWR Morb Mortal Wkly Rep. 2010;59:258–61.
12. Lexau CA, Lynfield R, Danila R, Plishivlii T, Facklam R, Farley MM, et al. Changing epidemiology of invasive pneumococcal disease among older adults in the era of pediatric pneumococcal conjugate vaccine. JAMA. 2005;294:2043–51. http://dx.doi.org/10.1001/jama.294.16.2043
13. Gertz RE Jr, Li Z, Pimenta FC, Jackson D, Juni BA, Lynfield R, et al. Increased penicillin nonsusceptibility of nonvaccine-serotype invasive pneumococci other than serotypes 19A and 6A in post–7-valent conjugate vaccine era. J Infect Dis. 2010;201:770–5. http://dx.doi.org/10.1086/650496
14. Richter SS, Heilmann KP, Dohrn CL, Riahi F, Diekema DJ, Doern GV. Pneumococcal serotypes before and after introduction of conjugate vaccines, United States, 1999–2011. Emerg Infect Dis. 2013;19:1074–83. http://dx.doi.org/10.3201/eid1907.121830
15. Miller E, Andrews NJ, Waight PA, Slack MP, George RC. Herd immunity and serotype replacement 4 years after seven-valent pneumococcal conjugate vaccine introduction in England and Wales: an observational cohort study. Lancet Infect Dis. 2011;11:760–8. http://dx.doi.org/10.1016/S1473-3099(11)70090-1
16. Steens A, Bergsaker MA, Aaberge IS, Rønning K, Vestrheim DF. Prompt effect of replacing the 7-valent pneumococcal conjugate vaccine with the 13-valent vaccine on the epidemiology of invasive pneumococcal disease in Norway. Vaccine. 2013;31:6232–8. http://dx.doi.org/10.1016/j.vaccine.2013.10.032
17. Moore CE, Paul J, Foster D, Mahar SA, Griffiths D, Knox K, et al. Reduction of invasive pneumococcal disease 3 years after the introduction of the 13-valent conjugate vaccine in the Oxfordshire region of England. J Infect Dis. 2014;210:1001–11. http://dx.doi.org/10.1093/infdis/jiu213
18. Harboe ZB, Dalby T, Weinberger DM, Benfield T, Melbak K, Slotved HC, et al. Impact of 13-valent pneumococcal conjugate vaccination in invasive pneumococcal disease incidence and mortality. Clin Infect Dis. 2014;59:1066–73. http://dx.doi.org/10.1093/cid/ciu524
19. Chiba N, Morozumi M, Wajima T, Shoju M, Iwata S, Ubukata K, et al. Changes in capsule and drug resistance of pneumococci after introduction of PCV7, Japan, 2010–2013. Emerg Infect Dis. 2014;20:1132–9. http://dx.doi.org/10.3201/eid2007.131485
20. Levy MM, Fink MP, Marshall JC, Abraham E, Angus D, Cook D, et al. 2001 SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference. Crit Care Med. 2003;31:1250–6. http://dx.doi.org/10.1097/01.CCM.0000050454.01978.3B
21. Chiba N, Morozumi M, Ubukata K. Application of the real-time PCR method for genotypic identification of beta-lactam resistance isolates from invasive pneumococcal diseases. Microb Drug Resist. 2012;18:149–56. http://dx.doi.org/10.1089/mdr.2011.0102
22. Pan XS, Ambler J, Meltiar S, Fisher LM. Involvement of topoisomerase IV and DNA gyrase as ciprofloxacin targets in Streptococcus pneumoniae. Antimicrob Agents Chemother. 1996;40:2321–6.
23. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; 21st informational supplement. Vol. 31. Document M100-S21. Wayne (PA): The Institute; 2011.
24. Sakai F, Chiba N, Ono A, Yamagata Murayama S, Ubukata K, Sunakawa K, et al. Molecular epidemiologic characteristics of Streptococcus pneumoniae isolates from children with
meningitis in Japan from 2007 through 2009. J Infect Chemother. 2011;17:334–40. http://dx.doi.org/10.1007/s10156-010-0180-3

25. Griffin MR, Zhu Y, Moore MR, Cynthia G, Whitney CG, Grijalva CG. U.S. hospitalizations for pneumonia after a decade of pneumococcal vaccination. N Engl J Med. 2013;369:155–63. http://dx.doi.org/10.1056/NEJMoa1209165

26. Angoulvant F, Levy C, Grimpel E, Varon E, Lorrot M, Biscardi S, et al. Early impact of 13-valent pneumococcal conjugate vaccine on community-acquired pneumonia in children. Clin Infect Dis. 2014;58:918–24. http://dx.doi.org/10.1093/cid/ciu006

27. Shea KM, Weycker D, Stevenson AE, Strutton DR, Pelton SI. Modeling the decline in pneumococcal acute otitis media following the introduction of pneumococcal conjugate vaccines in the US. Vaccine. 2011;29:8042–8. http://dx.doi.org/10.1016/j.vaccine.2011.08.057

28. Taylor S, Marchisio P, Vergison A, Harriague J, Hausdorff WP, Haggard M. Impact of pneumococcal conjugate vaccination on otitis media: a systematic review. Clin Infect Dis. 2012;54:1765–73. http://dx.doi.org/10.1093/cid/cis292

29. Kyaw MH, Lynfeld R, Schaffner W, Craig AS, Hadler J, Reingold A, et al. Effect of introduction of the pneumococcal conjugate vaccine on drug-resistant Streptococcus pneumoniae. N Engl J Med. 2006;354:1455–63. http://dx.doi.org/10.1056/NEJMoa051642

30. Hsu HE, Shutt KA, Moore MR, Beall BW, Bennett NM, Craig AS, et al. Effect of pneumococcal conjugate vaccine on pneumococcal meningitis. N Engl J Med. 2009;360:244–56 http://dx.doi.org/10.1056/NEJMoa080836

31. Richter SS, Diekema DJ, Heilmann KP, Dohrn CL, Riahi F, Doern GV. Changes in pneumococcal serotypes and antimicrobial resistance after introduction of the 13-valent conjugate vaccine in the United States. Antimicrob Agents Chemother. 2014;58:6484–9. http://dx.doi.org/10.1128/AAC.03344-14

32. Hampton LM, Pan XS, Ambler J, Mehtar S, Farley MM, Schaffner W, et al. Prevention of antibiotic-nonsusceptible Streptococcus pneumoniae with conjugate vaccines. J Infect Dis. 2012;205:401–11. http://dx.doi.org/10.1093/infdis/jir755

33. Beall B, McEllistrem MC, Gertz RE Jr, Boxrud DJ, Besser JM, Harrison LH, et al. Emergence of a novel penicillin-nonsusceptible, invasive serotype 35B clone of Streptococcus pneumoniae within the United States. J Infect Dis. 2002;186:118–22. http://dx.doi.org/10.1086/341072

34. Song JH, Baek YJ, Ko KS. Comparison of capsular genes of Streptococcus pneumoniae serotype 6A, 6B, 6C, and 6D isolates. J Clin Microbiol. 2011;49:1758–64. http://dx.doi.org/10.1128/JCM.02628-10

35. Ko KS, Baek YJ, Song JH. Multidrug–resistant Streptococcus pneumoniae serotype 6D clones in South Korea. J Clin Microbiol. 2012;50:818–22. http://dx.doi.org/10.1128/JCM.05895-11

36. Pai R, Gertz RE, Whitney CG, Beall B; Active Bacterial Core Surveillance Team. Clonal association between Streptococcus pneumoniae serotype 23A, circulating within the United States, and an internationally dispersed clone of serotype 23F. J Clin Microbiol. 2005;43:5440–4. http://dx.doi.org/10.1128/JCM.43.11.5440-5444.2005

37. Brueggemann AB, Pai R, Crook DW, Beall B. Vaccine escape recombinants emerge after pneumococcal vaccination in the United States. PLoS Pathog. 2007;3:e168. http://dx.doi.org/10.1371/journal.ppat.0030168

38. Ko KS, Baek YJ, Song JH. Multidrug–resistant Streptococcus pneumoniae serotype 6D clones in South Korea. J Clin Microbiol. 2012;50:818–22. http://dx.doi.org/10.1128/JCM.05895-11

39. Grabenstein JD, Weber DJ. Pneumococcal serotype diversity among adults in various countries, influenced by pediatric pneumococcal vaccination uptake. Clin Infect Dis. 2014;58:854–64. http://dx.doi.org/10.1093/cid/cit600

40. Lee LH, Gu XX, Nahm M. Towards new broader spectrum pneumococcal vaccines: the future of pneumococcal disease prevention. Vaccines. 2014;2:112–28 http://dx.doi.org/10.3390/vaccines2010112.

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Serotype Changes and Drug Resistance in Invasive Pneumococcal Diseases in Adults after Vaccinations in Children, Japan, 2010–2013

Technical Appendix

**Technical Appendix Table.** Susceptibility to 6 parenteral β-lactam antimicrobial agents by genotype class in isolates from adults with invasive pneumococcal diseases, Japan, April 2010–March 2013.

| Antimicrobial agent and genotype class* | MIC<sub>50</sub> (μg/mL) | MIC<sub>90</sub> (μg/mL) | MIC range (μg/mL) |
|----------------------------------------|--------------------------|--------------------------|-------------------|
| Penicillin                            |                          |                          |                   |
| gPSSP (n = 115)                        | 0.016                    | 0.031                    | 0.004–0.031       |
| gPISP (pbp2x) (n = 285)                | 0.063                    | 0.063                    | 0.016–0.125       |
| gPISP (pbp1a+2x) (n = 73)              | 0.125                    | 0.25                     | 0.063–1           |
| gPISP (pbp2x+2b) (n = 41)              | 0.125                    | 0.5                      | 0.031–0.5         |
| gPRSP (pbp1a+2x+2b) (n = 184)          | 2                        |                          | 0.25–4           |
| Ampicillin                            |                          |                          |                   |
| gPSSP                                 | 0.031                    | 0.031                    | 0.004–0.031       |
| gPISP (pbp2x)                         | 0.063                    | 0.125                    | 0.008–0.125       |
| gPISP (pbp1a+2x)                      | 0.125                    | 0.5                      | 0.031–1           |
| gPISP (pbp2x+2b)                      | 0.25                     | 0.5                      | 0.063–1           |
| gPRSP (pbp1a+2x+2b)                   | 1                        | 2                        | 0.125–4           |
| Cefotaxime                            |                          |                          |                   |
| gPSSP                                 | 0.016                    | 0.031                    | 0.008–0.25        |
| gPISP (pbp2x)                         | 0.25                     | 0.5                      | 0.008–0.5         |
| gPISP (pbp1a+2x)                      | 1                        | 1                        | 0.125–4           |
| gPISP (pbp2x+2b)                      | 0.25                     | 0.5                      | 0.031–0.5         |
| gPRSP (pbp1a+2x+2b)                   | 1                        | 2                        | 0.125–8           |
| Ceftriaxone                            |                          |                          |                   |
| gPSSP                                 | 0.016                    | 0.063                    | 0.008–0.125       |
| gPISP (pbp2x)                         | 0.25                     | 0.25                     | 0.016–0.5         |
| gPISP (pbp1a+2x)                      | 0.5                      | 1                        | 0.25–1            |
| gPISP (pbp2x+2b)                      | 0.25                     | 0.25                     | 0.063–0.5         |
| gPRSP (pbp1a+2x+2b)                   | 0.5                      | 1                        | 0.25–2            |
| Meropenem                             |                          |                          |                   |
| gPSSP                                 | 0.016                    | 0.031                    | 0.008–0.063       |
| gPISP (pbp2x)                         | 0.016                    | 0.063                    | 0.008–0.125       |
| gPISP (pbp1a+2x)                      | 0.063                    | 0.125                    | 0.016–0.25        |
| gPISP (pbp2x+2b)                      | 0.063                    | 0.5                      | 0.016–0.5         |
| gPRSP (pbp1a+2x+2b)                   | 0.5                      | 1                        | 0.063–4           |
| Panipenem                             |                          |                          |                   |
| gPSSP                                 | 0.004                    | 0.004                    | 0.002–0.008       |
| gPISP (pbp2x)                         | 0.004                    | 0.008                    | 0.002–0.016       |
| gPISP (pbp1a+2x)                      | 0.016                    | 0.016                    | 0.004–0.063       |
| gPISP (pbp2x+2b)                      | 0.016                    | 0.031                    | 0.004–0.031       |
| gPRSP (pbp1a+2x+2b)                   | 0.063                    | 0.125                    | 0.008–0.25        |

*Results of gPISP (pbp2b) (n = 9) and gPISP(pbpTa+2b) (n = 3) were omitted because of a low number of strains.
Technical Appendix Figure 1. Comparison of proportional increases and decreases in serotypes in isolates from adults with invasive pneumococcal diseases, Japan, from first surveillance period (April 2010–March 2011) to third surveillance period (April 2012–March 2013). Blue indicates serotypes included in PCV7; green indicates serotypes included in PCV13; orange indicates serotypes included in PPSV23; and purple indicates serotypes not included in PPSV23.
Technical Appendix Figure 2. Changes in number of serotypes and in macrolide (ML) resistance by genotype in isolates from adults with invasive pneumococcal diseases for 3 yearly surveillance periods, Japan, April 2010–March 2013. ML susceptible indicates not possessing any ML-resistance gene; ML resistance [mef(A)] indicates possessing mef(A) gene mediating ML-intermediate resistance to 14-membered MLs and azalide; ML resistance [erm(B)] indicates possessing erm(B) gene mediating ML-high resistance; and ML resistance [mef(A)+erm(B)] indicates possessing both resistance genes.
Technical Appendix Figure 3. Distribution of susceptibilities to 4 parenteral β-lactam antimicrobial agents (penicillin, ampicillin, cefotaxime, and meropenem) by genotype in isolates from adults with invasive pneumococcal diseases, Japan, April 2010–March 2013. gPSSP indicates genotypic penicillin-susceptible Streptococcus pneumoniae; gPISP indicates genotypic penicillin-intermediate–resistant S. pneumoniae; and gPRSP indicates genotypic penicillin-resistant S. pneumoniae. Parentheses enclose abnormal pbp genes mediating penicillin resistance.