Do Disruptions in the Circadian Timing System Contribute to Autonomic Dysfunction in Huntington’s Disease?

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Huntington’s disease (HD) patients suffer from a progressive neurodegenerative disorder that inflicts both motor and non-motor symptoms. HD is caused by a CAG repeat expansion within the first exon of the 
\textit{huntingtin} (HTT) gene that produces a polyglutamine repeat that leads to protein misfolding, soluble aggregates, and inclusion bodies detected throughout the body. Both clinical and preclinical research indicate that cardiovascular dysfunction should be considered a core symptom in at least a subset of HD patients. There is strong evidence for dysautonomia (dysfunctional autonomic nervous system, ANS) in HD patients that can be detected early in the disease progression. The temporal patterning of ANS function is controlled by the circadian timing system based in the anterior hypothalamus. Patients with neurodegenerative diseases including HD exhibit disrupted sleep/wake cycle and, in preclinical models, there is compelling evidence that the circadian timing system is compromised early in the disease process. Here we review data from preclinical models of HD that explore the intersection between disruption of circadian rhythms and dysautonomia. This work will lead to new therapeutic strategies and standards of care for HD and other neurodegenerative diseases.

*Abbreviations: ATII, Angiotensin II; AVP, arginine vasopressin; ANS, autonomic nervous system; BP, blood pressure; BMAL1, Brain and muscle aryl-hydrocarbon receptor nuclear translocator-like 1; BDNF, brain-derived neurotrophic factor; CCM, cardiomyocyte-specific Clock mutant; CV, cardiovascular; CNS, central nervous system; CLOCK, Circadian Locomotor Output Cycles Kaput; Cry, cryptochrome; DD, constant darkness; DMV, dorsal motor nucleus of the vagus; HR, heart rate; HF, high-frequency; HD, Huntington’s disease; \textit{Htt}, huntingtin; IML, intermediolateral; BK, large-conductance calcium- and voltage-activated potassium currents; KLF15, kruppel-like factor 15; LD, light/dark; LF, low-frequency; NP, nitroprusside; NTS, nucleus tractus solitarius; PVN, paraventricular nucleus; Per, period; PPARα, peroxisome proliferator activated receptor alpha; \textit{pdk4}, pyruvate dehydrogenase kinase 4; ROR, retinoic acid-related orphan receptor; SCN, suprachiasmatic nucleus; \textit{ucp3}, uncoupling protein 3; VIP, vasoactive intestinal peptide.

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INTRODUCTION

Huntington’s Disease is a Genetically-determined Neurodegenerative Disease

Huntington’s disease (HD) patients suffer from a progressive neurodegenerative disorder that inflicts cognitive, psychiatric, and motor symptoms [1,2]. HD is caused by a CAG repeat expansion within the first exon of the huntingtin (HTT) gene which produces a polyglutamine repeat that leads to protein misfolding, soluble aggregates, and inclusion bodies detected [3,4]. The normal function of the protein (Huntingtin) is unknown; however, the mutated form leads to dysfunction of a broad range of cellular processes including cytoskeletal organization, metabolism, and transcriptional activities. Based on the broad distribution of the HTT, the mutation would be expected to produce symptoms throughout the body. Indeed, recent work suggests HD is a systemic illness affecting the entire body and data from animal studies suggests that core features of the disease can be modified by treatments that target tissues outside of the nervous system [5]. To further preclinical research, a large number of animal models of HD have been developed, each with strengths and weaknesses (see [6]). The work covered in this review was mostly conducted on the Q175, BACHD, R6/2, and R6/1 mouse lines.

Cardiovascular Dysfunction May Be Common in Huntington’s Disease

Cardiovascular (CV) events are a major cause of early death in the HD population and occur at a higher rate compared to the rest of the population [7,8]. Possible cardiomyopathies have not been well explored in HD patients. One recent study examined the electrocardiograms of over 500 early symptomatic HD patients and found evidence for low heart rate (bradycardia) and conduction abnormalities in a significant fraction of the patients [9]. Preclinical models of HD have found clear evidence for reduced contractility and cardiac output [10-15]. For example, a recent study of the cardiomyocytes of BACHD mice found evidence for increased calcium/calmodulin-dependent protein kinase II activity as well as structural abnormalities in the mitochondria [16]. Also, metabolic analysis of heart tissue from patients and mouse models found evidence for energy equilibrium imbalances in cardiac cells [17]. Finally, the cardiac-specific expression of polyQ repeats leads to CV dysfunction, suggesting that CV disease may be the result of local abnormalities [18,19].

In addition, there is strong evidence for dysautonomia (dysfunctional autonomic nervous system, ANS) in HD patients that can be detected early in the disease progression. The sympathetic nervous system appears to be most impacted during the very early stages of HD [20-22]. As the disease advances, the parasympathetic activity progressively decreases as well [22-24]. Therefore, both the clinical and preclinical research indicate that CV dysfunction should be considered a core symptom of at least a subset of HD patients [25].

Sleep and Circadian Dysfunction are an Integral Component of Huntington’s Disease Pathophysiology

Sleep disorders are prevalent in HD patients and have detrimental effects on the daily functioning and quality of life of patients and their caregivers [26,27]. The most common symptoms found in these studies include a delay in sleep onset, fragmented sleep during the night, and daytime sleepiness. Importantly, these disruptions in the sleep/wake cycle occur early in the disease progression and so could serve as a biomarker for HD as well as a target for interventions. The sleep/wake cycle is conceptualized as being driven by two, anatomically-distinct processes: a homeostatic sleep mechanism (process S) as well as by the circadian timing system (process C). To date, there is little evidence that HD impacts sleep homeostasis, but there is growing evidence for HD-driven disruption in circadian timing. Still, it is difficult to determine whether the disease alters the circadian timing system in humans, and animal models provide critical insights. Mouse models of HD also exhibit a progressive and rapid breakdown of the circadian rest/activity cycle that closely mimics the condition observed in human patients, typified by loss of consolidated sleep, increased activity during the rest phase, and more sleep during the active phase [11,28-30]. Importantly, the disruptions are seen under both light/dark (LD) cycle as well as the mice are held in constant darkness (DD). This latter step is critical to establish that the circadian system is compromised. Disorganized circadian timing leads to undesirable effects throughout the body [31], altering the function of key organ systems including heart, pancreas, liver, lungs, as well as the brain. Collectively this prior research supports the hypothesis that circadian dysfunction is an integral component of HD pathophysiology. We have been testing the hypothesis that dysfunction in the circadian system contributes to the CV disease in HD (Figure 1) and, in this review, we will summarize our progress.

TOPICS

Baroreceptor Reflex is Blunted in Huntington’s Disease

One of the classic physiological tests of the ANS function is the measurement of the baroreceptor reflex in which changes in blood pressure (BP) evoke alterations
in the heart rate (HR). Baroreceptors are mechanoreceptors located in the carotid sinus and aortic arch that sense the pressure changes. Although they are sensitive to both increases and decreases in arterial pressure, their primary role is to respond to a sudden fall in arterial pressure. This sudden decrease is detected by baroreceptors and signals are sent to the medulla to elevate the sympathetic activity and reduce the parasympathetic activity. These changes lead to vasoconstriction and increased HR to bring back arterial pressure into the normal range. Recordings from the BACHD [32] as well as the Q175 [15] lines of HD mice indicate that the baroreceptor reflex is dramatically altered (Figure 2). The administration of Angiotensin II (ATII) or Nitroprusside (NP) leads to episodes of hypertension or hypotension, respectively. Changes in BP as a result of drug administration should elicit a compensatory response of HR via the baroreceptor reflex in order to normalize BP levels. The Q175 mutants showed a dramatically blunted response in HR to the transient hypotension induced by NP suggesting that the sympathetic branch has impaired. The BACHD mutants showed a blunted response to both ATII and NP indicating both sympathetic and parasympathetic arms were impacted. In both cases, we confirmed that there were no genotypic differences in the change in BP evoked by the injected drug as well as performed pharmacological controls to showing that appropriate receptor blockers prevented the change in HR.

HD patients complain of dizziness and light-headedness upon standing, all symptoms of baroreceptor dysregulation resulting in orthostatic hypotension [20,33]. While not well documented in HD patients, orthostatic hypotension has been extensively examined in Parkinson’s disease (PD) [34]. In this neurodegenerative disease, the neurogenic hypotension impacts about 30 percent of the patients with dizziness or lightheadedness, fatigue when standing, and difficulty walking to be the most common symptoms [35,36]. In a significant number of patients, the hypotension is responsible for the falls which one of the main complications of PD that has a major impact on the quality of life. Patients with neurogenic orthostatic hypotension exhibit an enhanced reduction in systolic BP (40 vs. 20 mmHg) in response to a stressor and thus are vulnerable for falls due to failure to maintain BP [37]. Like the PD patients, we expect that a subset of HD patients will be vulnerable to neurogenic orthostatic hypotension.

Heart Rate Variability is Reduced in Huntington’s Disease

Heart rate variability (HRV) is a measure of variation in the beat-to-beat interval that reflects the dynamic balance of sympathetic and parasympathetic control of heart function. Traditionally, HRV is generally considered an indication of CV health and low HRV proposed as a predictor for CV disease and mortality [38,39]. More recently, HRV has become a popular index of cardiac autonomic control in the biobehavioral sciences due to its
The ANS (Figure 3). Both the LF and HF domains of the HRV exhibited robust daily rhythms. In the Q175 line [15], the HRV was low in young mutants, and this reduction was largest during the rest phase. The power in the LF domain was significantly reduced in the young mutants with the most prominent effects during the night. A similar reduction in HRV was previously observed in the BACHD line [11,14] as well as in the R6/1 model [44]. In HD patients, a similar decrease in HRV has also been reported during the presymptomatic and early stages of HD progression [20-23]. Prior studies reported HRV deficits during the Valsalva maneuver, hand-grip test, and the head up tilt test in HD patients [22,24,44-46]. As far as we know, the spectral power analysis has not been carried out on data from patients. In human subjects, HRV relationship with stress disorders and other illnesses [40]. In humans, it is typically measured for short intervals and is increasingly available in health records as a non-invasive measure of CV function. One of the most commonly used methods for HRV evaluation is power spectral density analysis in which high-frequency (HF) and low-frequency (LF) bands are extracted from the HRV signal, and the spectral power is calculated (e.g., [41]). Traditionally, the LF power is viewed as a measure of regulation by the sympathetic branch of the ANS, although it is more likely a general index of ANS function [42,43]. In our work, we measured HRV in WT and HD mutant mice over several days in both LD and DD conditions. In WT mice, HRV displayed a robust diurnal and circadian rhythm consistent with circadian regulation of the ANS (Figure 3). Both the LF and HF domains of the HRV exhibited robust daily rhythms. In the Q175 line [15], the HRV was low in young mutants, and this reduction was largest during the rest phase. The power in the LF domain was significantly reduced in the young mutants with the most prominent effects during the night. A similar reduction in HRV was previously observed in the BACHD line [11,14] as well as in the R6/1 model [44].

In HD patients, a similar decrease in HRV has also been reported during the presymptomatic and early stages of HD progression [20-23]. Prior studies reported HRV deficits during the Valsalva maneuver, hand-grip test, and the head up tilt test in HD patients [22,24,44-46]. As far as we know, the spectral power analysis has not been carried out on data from patients. In human subjects, HRV

Figure 2. Baroreceptor reflex is disrupted in mouse models of HD. In the baroreceptor reflex, changes in BP are detected by the baroreceptors and evoke compensatory changes in ANS. The Q175 and BACHD mutants showed a dramatically blunted response in HR to the transient hypotension induced by nitroprusside (NP) suggesting that the sympathetic branch has impaired. The effect of NP was blocked by a beta-adrenergic receptor blocker (β-blocker) propranolol. The Q175 mutants did not show a significantly altered response to transient hypertension induced by angiotensin II. HD patients complain of dizziness and light-headedness upon standing, all symptoms of baroreceptor dysregulation resulting in orthostatic hypotension. Data from [15,32].
Interestingly, these natural rhythms may increase the risk for vulnerable individuals and daily rhythms in the symptoms of CV disease peak in the morning hours (e.g., [48]). In nocturnal mice, diurnal and circadian rhythms in HR and LP power in WT mice. As shown in the panels, HRV increased during sleep while LF power (sympathetic outflow) was higher when the mice were active. In the HD mutants (Q175), the HRV was low in young mutants and this reduction was largest during the rest phase. The power in the LF domain was significantly reduced in the young mutants with the biggest effects during the night. A similar reduction in HRV was previously observed in the BACHD line [11,14] as well as in the R6/1 model [44]. In HD patients, a similar decrease in HRV has also been reported during the presymptomatic and early stages of HD progression.

Abnormal Diurnal and Circadian Rhythms in Heart Rate Observed in Huntington’s Disease

In humans and other animals, HR varies dramatically with acute physical demands, i.e., higher HR when we are physically active while low HR when sedentary. In addition to these acute changes, there are also robust 24-hr circadian rhythms in the heart and vasculature to prepare the CV system for higher output during the day in humans. Interestingly, these natural rhythms may increase the risk for vulnerable individuals and daily rhythms in the symptoms of CV disease peak in the morning hours (e.g., [48]).

In nocturnal mice, diurnal and circadian rhythms in CV output peak in the night. While influenced by activity levels, the rhythms occur even when measured in windows of time when the mice were inactive. As measured by telemetry, diurnal and circadian rhythms in HR are disrupted in the BACHD, Q175, R6/1 lines of mice [11,14,15,44]. For example, the Q175 exhibited highly pronounced tachycardia during their normal sleep time, with high HR and reduced amplitude in the HR rhythm (Figure 4). Besides, the normal strong correlation between activity and HR,
Possible Mechanisms Underlying the Dysautonomia

The circuits involved in the generation of rhythms in HR and autonomic function are relatively well defined. The central circadian clock in the suprachiasmatic nucleus (SCN) orchestrates the peripheral clocks via ANS [52-56]. Complete separation between pre-sympathetic and pre-parasympathetic neurons starts from the SCN (Figure 5) [56,57]. These separate pre-sympathetic and pre-parasympathetic neurons project to the paraventricular nucleus (PVN) and its separated pre-autonomic neurons have projections to either the preganglionic sympathetic neurons in the intermediolateral (IML) column of the spinal cord or the preganglionic neurons of the dorsal motor nucleus of the vagus (DMV). There are axon collaterals of the pre-sympathetic PVN neurons in the nucleus tractus solitarius (NTS). Particularly, the SCN utilizes complete separation of sympathetic and parasympathetic neurons to convey the circadian information to the periphery including the heart, liver, and adrenal gland. Various neuroendocrine and autonomic neurons are located in the PVN where appropriate hormonal and autonomic responses are coordinated [56,57].

HD is a neurodegenerative disease so the most obvious cause of dysautonomia is the loss of cell populations in the hypothalamus or brain stem. We examined possible anatomical changes within the SCN of the BACHD mouse at 3 months of age just as the motor symptoms
can first be measured. We found that the male, but not the female, SCN was smaller than WT controls. There were no differences in peptide expression (arginine vasopressin, AVP; vasoactive intestinal peptide, VIP) within the SCN with genotype or sex. In the more severely impacted R6/2 mice, the SCN shows decreased expression of VIP and its receptor, VPAC2 [58]. VIP plays a key role in synchronizing cell populations within the SCN and its reduction would be expected to disrupt the population rhythms in neural activity from this structure. The SCN sends projections out to regulate the temporal patterns of activity in several major arousal centers including the orexin expressing cell population in the lateral hypothalamus. The expression levels of orexin are reduced in HD models (e.g., [59,60]). Changes in the expression of brain-derived neurotrophic factor (BDNF) have also been observed. Expression of BDNF is normally high in the brainstem where this neuromodulator plays a crucial for regulating HR [61-63]. The levels of BDNF are known to be reduced in the brainstem of mutant mice and resorting BDNF level can ameliorate HD pathology and prolong the lifespan in HD mice [64]. Importantly, restoring BDNF levels also returned the HR of the HD mice to the control levels [65]. Therefore, in the mouse models, there is evidence for the degeneration of neurons in the SCN as well as the brain stem. There is also evidence for the strong expression of p62 immunopositive protein aggregates in axons of brainstem fiber tracts including the vagal nerve and the NTS [70].

In HD patients, there is evidence that the disease causes similar changes in expression as well as degeneration in the brain regions involved in the central nervous system (CNS) regulation of CV function. In the hypothalamus, there is evidence for the reduction in the expression of orexin and AVP in the brains of the HD patients [66,67]. In addition, the levels of BDNF and its receptors (TrkB) are reduced in HD patients [68], and the HD aggregates directly interfere with the transcription of BDNF [69]. Finally, there is evidence for degeneration in the

Figure 5. The circuits involved in the generation of rhythms in HR and autonomic function are relatively well defined. The central circadian clock in the suprachiasmatic nucleus (SCN) orchestrates the peripheral clocks via ANS. It appears that separate populations of SCN neurons project to the paraventricular nucleus (PVN) and innervate pre-sympathetic and pre-parasympathetic cell populations. These separate pre-sympathetic and pre-parasympathetic neurons project to either the preganglionic sympathetic neurons in the intermediolateral (IML) column of the spinal cord or the preganglionic neurons of the dorsal motor nucleus of the vagus (DMV). There are axon collaterals of the pre-sympathetic PVN neurons in the nucleus tractus solitarius (NTS). Both pathways innervate the heart to regulate HR via release of acetylcholine (ACh) and norepinephrine (NE). Also, the SCN regulates the HPA axis and the secretion of cortisol. The glucocorticoid receptors are also potent regulators of CV function. In HD patients, there is evidence for the degeneration of neurons in the SCN as well as the brain stem. There is also evidence for the strong expression of p62 immunopositive protein aggregates in axons of brainstem fiber tracts including the vagal nerve and the NTS [70].
brainstem nuclei in HD patients including loss of neurons in the substantia nigra, pontine nuclei, reticulotegmental nucleus of the pons, superior and inferior olives [70]. This study also reported the strong expression of p62 immunopositive protein aggregates in axons of brainstem fiber tracts including the vagal nerve and the nucleus of the solitary tract. These structures are centrally involved in the CNS regulation of the CV system. Thus, by the end of life, HD certainly impacts the brain regions responsible for the CNS regulation of CV function and establishes a possible structural cause for this dysfunction.

**Altered Suprachiasmatic Nucleus-driven Physiology Output in Huntington’s Disease**

The circadian system is composed of cell-autonomous clock gene expression rhythms that are synchronized and adaptively phase aligned in tissues throughout the body by a rhythmic output from the SCN [71]. Individual SCN neurons express rhythms in spontaneous firing rate, with higher firing rates observed during the day and low rates at night [72,73]. The BACHD and Q175 mouse models of HD exhibit decreased electrical activity in the SCN during the day [11,74]. This decrease in daytime firing in the SCN was not seen in the R6/2 model [75] although firing rate deficits were seen in an SCN-driven output in the orexin neurons [60]. Using electrophysiological techniques, we found that SCN neural activity rhythms were lost early in the disease progression and were accompanied by loss of the normal daily variation in resting membrane potential in the mutant SCN neurons [74]. The low neural activity could be transiently reversed by direct current injection thus demonstrating that the neurons have the capacity to discharge at WT levels. Exploring the potassium currents known to regulate the electrical activity of SCN neurons, our most striking finding was that these cells in the mutants exhibited an enhancement in the large-conductance calcium- and voltage-activated potassium (BK) currents. The expression of the pore-forming subunit (*Kcnma1*) of the BK channel was higher in the mutant SCN. These findings demonstrate that SCN neurons of both BACHD and Q175 HD models exhibit early pathophysiology and that dysregulation of BK current may be responsible.

**Circadian Molecular Feedback Loop May Be Impacted by Huntington’s Disease**

The rhythms in neural activity in the SCN are driven by cell autonomous molecular feedback loops. At a molecular level, circadian rhythms are generated by the intracellular transcriptional/translational feedback loop, driving daily oscillations with a period of approximately 24-hrs in the expression of core clock proteins. CLOCK (Circadian Locomotor Output Cycles Kaput) and BMAL1 (Brain and muscle aryl-hydrocarbon receptor nuclear translocator-like 1) the positive components of the clock bind to E-box sequences and drive the expression of the negative elements period (*Per*) and cryptochrome (*Cry*) which can inhibit their own transcription by repressing the CLOCK/BMAL1 heterodimer. Once the levels of PER and CRY are decreased, the new cycle of transcription/translation is started by CLOCK-BMAL1. This feedback loop constitutes a 24-hr period of the internal circadian rhythms. The circadian nuclear receptor, Rev-erba and a retinoic acid-related orphan receptor (ROR) regulate the *Bmal1* expression via activation and repression, respectively. Post-translational modifications are crucial for regulating the clock. Casein kinase 1 (CK1) phosphorylate PER and CRY which is vital for the circadian cycle length. The expression of other clock-controlled genes and output genes is modulated by the CLOCK/BMAL1 heterodimer, which involves many physiological functions [76]. We found that the circadian rhythms in PER2-driven bioluminescence were not altered in the SCN in the BACHD [77]; however, deficits in gene expression rhythms were found in the SCN of the more severely impacted R6/2 model [28]. Thus, the evidence that the molecular circadian clockwork is disrupted is so far mixed, and more work is required to identify at what stage in the disease progression these disruptions occur.

These molecular rhythms are not just expressed in the SCN and work done in the R6/2 model clearly demonstrates that circadian rhythms of clock-driven genes that are critical metabolic outputs in the liver are abolished in vivo [78]. This deficiency is accompanied by arrhythmic expression of the clock genes *Cry1* and *Dbp*, and a phase-advanced *Per2* cycle. There is overwhelming evidence that circadian clock genes including *Bmal1*, *Clock*, *Rev-erba*, *Per 1*, *Per 2*, *Cry1*, and *Cry2* are rhythmically expressed in cardiomyocytes [79,80]. Functionally this timing system has a major impact on the expression of the genes involved in cardiac metabolism and electrical activity display circadian expression. For example, the metabolic genes, pyruvate dehydrogenase kinase 4 (*pdk4*) and uncoupling protein 3 (*ucp3*), are known to be regulated by peroxisome proliferator-activated receptor alpha (PPARα). PPARα displays a robust rhythmic expression in the heart and is also shown to modulate *Bmal1* by directly binding to the *Bmal1* promoter [81]. Daily oscillations of the expression in *Pdk4* and *ucp3* are associated with oscillations in the clock gene expression with the peak during the active phase. Glucose levels are known to exhibit diurnal rhythms. Glucose transporters 1 and 4 (Glut1,4) have a peak in the expression level at the same time, suggesting increased glucose transport and utilization at the same time with diurnal variations. Potassium channels, Kv1.5 and Kv4.2, as well as calcium transients
show rhythmicity in protein levels [82].

There is still much work to do to understand how the molecular clock controlling transcription interacts with the physiology of CV function. The general assumption is that the temporal pattern of transcription favors ATP production and energy utilization during the active phase while allowing remodeling and repair to dominate during rest. One particularly interesting case study involves the rhythmically expressed kruppel-like factor 15 (KLF15). Depletion of KLF15 in cardiomyocytes leads to a disorganized circadian behavior despite an intact core clock [83]. KLF15 transcriptionally controls the rhythmic expression of Kv channel-interacting protein 2, a critical subunit required for generating a transient potassium current. Deficiency or excess of KLF15 causes loss of rhythmic QT variation, abnormal repolarization and enhanced susceptibility to ventricular arrhythmias [84]. We do not know whether the HD mutation impacts KLF15 or other critical rhythmic outputs. We consider this lack of knowledge a major hole in the literature as HD-driven alterations in circadian output could be an important clinical symptom of the disease.

**Genetic or Environmental Disruption of the Circadian Timing System Leads to Cardiovascular Symptoms**

Regardless of the specific cause, prior work has shown that the genetic or environmental disruption in circadian rhythms can result in CV symptoms. For example, mice held in a 20-hr (LD10:10) cycle exhibit worse cardiac damage in response to high BP compared to controls held on a normal 24-hr cycle (LD 12:12) [85]. These lighting conditions alter rhythms in clock gene expression in both the heart and brain. Furthermore, “tau” mutant hamsters have a mutation in CK1 which results in a short endogenous cycle length of approximately 22-hrs [86] and, when they are held under a 24-hr LD cycle, these mutant hamsters die early with cardiomyopathy including fibrosis [86]. However, when the mutant hamsters are housed in a 22-hr LD cycle or are made arrhythmic by the destruction of their circadian clock (SCN), the hamsters are protected from developing cardiac dysfunction [86]. These results suggest that discord between the internal circadian clock (22 hrs) and external environment (24 hrs) can trigger cardiac disease. To provide a final example, Bmal1-knock out (KO) mice exhibit arrhythmic circadian behavior, age-related dilated cardiomyopathy, and shortened life span [87-90]. These symptoms are also seen in the cardiomyocyte-specific Bmal1 KO mice [91]. To understand the role of the circadian clock within the cardiomyocyte on myocardial biology, a cardiomyocyte-specific Clock mutant (CCM) mouse model was assessed [92,93]. CCM mice show decreased diurnal rhythms in HR, reduced cardiac efficiency, and bradycardia [88]. The cardiac hypertrophic markers are elevated in CCM mice [93]. These data suggest an essential role of cardiomyocyte circadian clock in CV function and highlight the possibility that at least some of the CV symptoms seen in HD could be the consequence of the well-established circadian dysfunction.

Finally, sleep disruption and work schedules that disrupt the circadian system are also risk factors for CV disease in humans [48]. Epidemiological studies show that approximately 40 percent of the population in North America do not get the recommended amount of sleep, and the reduced sleep duration is associated with a high risk of cardiac disease [94]. In the laboratory, placing healthy adults in an inverted sleep/wake and meal cycle for three days is sufficient to increase their BP and levels of the inflammatory markers [95]. A short 2 hr disruption of the sleep/wake can increase resting HR and cortisol levels in healthy subjects [96]. The sleep and rhythms disruption due to work schedule is associated with the adverse endocrine and CV profiles in subjects, putting them at increased CV risk [96]. In a study performed on healthy male shift workers, 24-hr electrocardiogram recordings are used to investigate the effect of the work schedule on the cardiac autonomic control [97]. Results show a reduction in the cardiac sympathetic modulation based on the HRV analysis, suggesting an increased risk for CV events [97]. Therefore, the data from humans is consistent with our assertion that the sleep and circadian dysfunction seen in HD could contribute to the CV symptoms seen in the patients.

**SUMMARY AND OUTLOOK**

There are several lines of evidence for dysautonomia in HD. First, HD patients complain of dizziness and light-headedness upon standing, all symptoms of baroreceptor dysregulation resulting in orthostatic hypotension [21,33]. Work in animal models has confirmed dysfunction in the baroreceptor reflex [15,32]. Second, HD patients [20-23] as well as mouse models [11,14,15,44] show reduced HRV. This variation in heart rate is a well-established marker of cardiac autonomic control and a reduction in HRV is associated with poor CV outcomes. Third, pathological analysis of HD patients has shown degeneration and intranuclear neuronal inclusions throughout the brainstem including regions known to be centrally involved in the autonomic regulation of the CV system [70]. We do not yet know whether these regions are also impacted in animal models of HD. There is evidence to suggest that the postganglionic sympathetic and intrinsic neurons in the heart may exhibit reduced functionality in mouse models of HD [49]. The disruption of the autonomic system is a well-established consequence of PD [34] and, collectively, this data argues that dysau-
Dysautonomia should also be considered in HD as well.

The body of evidence presented in this review is consistent with the hypothesis that the disruption of the SCN circuit contributes to the autonomic dysfunction seen in HD. If correct, this hypothesis has a number of predictions. First, the normal diurnal and circadian rhythms in HR and BP driven by the ANS will be disrupted in HD. To date, the disrupted rhythms in HR, BP, and baroreceptor reflex from data in HD animal models and patients support this hypothesis. Second, removing the mutant HTT from the heart may not be sufficient to rescue cardiac function. There is evidence of direct effects of mutant HTT in the heart (reviewed in [25]) but we emphasize that the dysautonomia is likely to be the result of dysfunction in the CNS circuit controlling autonomic function. This prediction is clinically important as clinical trials with mutant HTT lowering agents are ongoing and understanding the therapeutic targets are important for the success of these trials. Third, treatment with drugs focusing on dysautonomia should be targeted at specific times of the daily cycle. For example, beta-adrenergic receptor blockers may produce the most substantial benefits when taken before bed. Fourth, cardiac function shows a robust diurnal variation including rhythms in HR and BP. This body of preclinical data described in this review suggests that monitoring of the CV system in HD patients should start at an early age so therapeutic interventions can be employed to slow the progression of these pathological processes and prevent early death. Most importantly, our data suggest that this early screening must include observations during the usual sleep hours of the day, as early anomalies may go undetected at the times of day that patients would usually interact with clinicians. Finally, we predict that treatments or lifestyle changes that improve the circadian timing system will reduce the autonomic dysfunction in HD.

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