Prenatal is Still Neglected in Resource-Limited Areas despite the Use of Tests That Optimize the Diagnosis of Infectious Diseases in Brazil

Galileu Barbosa Costa1**, Raquel Gois Bastos2†, Ney Cristian Amaral Boa Sorte34, Antônio Conceição da Purificação3, Beneli Miranda dos Santos3, Bernardo Galvão-Castro4, Sara Nunez Vaz5, Lauro Juliano Marín2 and Sandra Rocha Gadelha2**

1Núcleo de Epidemiologia e Bioestatística, Centro de Pesquisas Gonçalo Moniz, Fiocruz, Salvador, Brazil
2Laboratório de Farmacogenômica e Epidemiologia Molecular, Departamento de Ciências Biológicas, Universidade Estadual de Santa Cruz, Ilhéus, Bahia, Brazil
3Associação de Pais e Amigos dos Excepcionais (APAE), Salvador, Bahia, Brazil
4Escola Baiana de Medicina e Saúde Pública, Fundação Bahiana para Desenvolvimento das Ciências, Salvador, Bahia, Brazil
5Centro de Pesquisa Gonçalo Moniz, Fundação Oswaldo Cruz (FIOCRUZ), Salvador, Bahia, Brazil

*Corresponding author: Galileu Barbosa Costa, Núcleo de Epidemiologia e Bioestatística, Centro de Pesquisas Gonçalo Moniz, Fiocruz, Rua Waldemar Falcão, 121, Candeal, Salvador, Bahia, Brazil, 40296-710, Tel: +5573988112802; Sandra Rocha Gadelha, Universidade Estadual de Santa Cruz, Departamento de Ciências Biológicas, Campus Soane Nazaré de Andrade, Rodovia Jorge Amado, km 16, Salobrinho, Ilhéus, Bahia, Brazil, 45662-900, Tel: +557388384075

Abstract
Vertically transmitted infections are caused by a diversity of pathogenic microorganisms, and pregnant women are routinely screened to evaluate the risks and reduce the burden of general disorders in their unborn. We assessed the seroprevalence of syphilis, toxoplasmosis, hepatitis B and C, HIV and Human T lymphotropic virus (HTLV) in pregnant women using public healthcare services in two main cities in South region of Bahia State, Brazil. Blood samples were collected using filter paper cards from 943 pregnant women aged between 12 and 45 years (average age of 24.3 yr). The average of pregnancies was 2.15, and most women reported they had never experienced abortion. ELISA assays were applied to detect antibodies against HBV, HCV, HIV, HTLV, syphilis, and toxoplasmosis. The prevalence rates of IgG antibodies found were 0.2% for HBV, 0.1% for HCV, 0.5% for HIV, 1.4% for HTLV, 2.8% for T. pallidum, and 1.5% for T. gondii. Only 47.5% of pregnant women returned to confirmatory tests. The rate of return for confirmatory test was low, even using tests that promote the diagnostic system. Our findings highlight how pre-natal care is still neglected, reinforcing the lack of awareness and the need for more incisive health education policies in the region.

Keywords
Pregnant women, Infectious diseases, Prenatal, Resource-limited areas, Public health

Introduction
Pregnant women, unborn fetuses, and neonates represent populations of high-risk individuals with increased susceptibility to infectious microbes that can pose significant health risks during pregnancy, depending on the pathogenesis and disease outcome [1-3]. The occurrence of infectious diseases during pregnancy is associated with spontaneous abortion, stillbirth, preterm delivery, and low birth weight [4,5], as well as persistent and long-lasting infections, even if not expressed at birth [6,7].

Infections transmitted to the fetus result in severe birth defects such as microcephaly or even fetal death, and is usually inversely related to gestational age at ac-
Material and Methods

Study population

A cross-sectional study was carried out during November 2011-October 2012 with pregnant women attended at Basic Healthcare Units from two main cities in the South State of Bahia, Northeast region of Brazil. Itabuna (14° 48’ 09.7” S 39° 16’ 38.8” W) and Ilhéus (14° 49’ 33.7” S 39° 02’ 03.7” W). Data on age, number of pregnancies, number of abortions, and place of residence were collected and converted to variables to be tested for correlation with the screened pathogens.

Samples collection and laboratory processing

Drops of blood were collected by using sterile lancet in digital region in the first or second chirodactyl (Figures 1A and 1B), after local asepsis with 70% ethanol [19,20]. The blood was soaked with three to four rounds of filter paper and dried at room temperature, protected from sunlight and from possible contaminants. After completely dried, the filter papers were bagged and kept at room temperature. The filter papers were sent to the Diagnostic and Research Center (CEDIP), APAE, Salvador, where they were processed and tested. Blood samples without anticoagulants were also collected, centrifuged and serum harvested and stored at -20 °C until tested.

Detection of Hepatitis B and C, HIV, HTLV, Syphilis, and toxoplasmosis

The samples were processed to detect anti-HIV 1+2, anti-HTLV-1 and 2, anti-VHC antibodies, anti T. pallidum (IgM and IgG), anti- T. gondii IgM antibodies and HBsAg on filter paper and plasma using the enzyme linked immunosorbent assay (ELISA). The immunological markers (antibodies) were obtained by an elution process, using a buffered solution containing protein, detergent, stabilizers and 0.1% sodium azide, and then subjected to ELISA, as recommended by the manufacturer. The following commercial kits on the material obtained on filter paper were used: IMUNOSCREEN HIV 1+2 - SS®, IMUN-
OSCREEN HTLV I and II - SS®, IMUNOSCREEN ANTI-HCV-SS®, IMUNOSCREEN TOXO IGM - SS®, IMUNOSCREEN SIFILIS - SS® and IMUNOSCREEN HBsAG - SS®, produced by MBIOLOG Diagnósticos LTD, Contagem, MG, Brazil.

**Confirmatory tests**

The active search system, in order to collect new samples to confirm the result obtained by the filter, was coordinated primarily by the APAE Salvador, through the specific sector, informing the Health Department of each city the pregnant woman results (positive screened cases). Since these women tend not to seek reference services frequently, when altered results were detected in the screening, it became necessary to seek these pregnant women in each Basic Health Unit, interfering with the service’s natural flow. In Ilhéus, a collection point for weekly collections of these samples was set up. In Itabuna, some of these samples were collected in the existing laboratory at the reference center, and others, in their own health units. Reference services for the monitoring of pregnant women are: Women’s Health Reference Center (CMAE) in Ilhéus and Reference Center Dr. Julio Brito (CRJB) in Itabuna. Of the pregnant women who answered the recall, two tubes of 4 ml venous blood were collected, with one dry tube, and the other containing the anticoagulant Ethylenediamine tetraacetic acid (EDTA). After blood samples were collected, they were taken to the Pharmacogenomics Laboratory of Molecular Epidemiology of the UESC (LAFEM), where they were centrifuged at 2500 revolutions per minute (RPM) for five minutes to separate the serum and the cellular portion. The serum obtained from the dried tube was aliquoted into two wells of 2.0 ml, with one stored and the other sent to the APAE Salvador, where the confirmatory test was performed. The tube containing EDTA was also aliquoted into two microcentrifuge tubes for subsequent DNA extraction.

**Sequencing and subtyping of HIV and HTLV positive samples**

Genomic DNA was extracted by using the QIAGEN QIAamp® DNA Blood Kit, according to the manufacturer’s instructions. All DNA samples were quantified using the GeneQuant pro Amersham Biosystem. For the HTLV-1 subtyping, we used a nested-PCR technique, specific for the LTR 5’ region of the HTLV-1, using on the first round, the following primers: ATLTR1 (5’-TGACACTGACCTACACTGAGCCGAGCCCCAAAT-3’) and ATLTR2 (5’-TCGTATCCTACTGTACCTGACCTAGCCGAGCCCCAAAT-3’); and for the second round, the primers ATLTR11 (5’-ACTAAGGCTCTGACGTCTCCCCC-3’) and ATLTR12 (5’-CGGTACTTGGCCGCTGACGCAACGGCCGAGCCCCAAT-3’). For the HIV-1 subtyping, we amplified the envelope (env) and group antigen (gag) regions by nested-PCR. For the env region, during the first round, we used the primers ED5 (5’-ATGGGATCAAGCCTAAAGCCATGTG-3’) and MM1 (5’-GGTGAATATCCTGCCCTAAT-3’); and in the second round, the primers ED31 (5’-CTCAAGGCATTACCAAGGCCCTGTCCAAAG-3’) and MM4 (5’-CCTCCTACTATGTTTGAGCCGACCATAAGCTATGAA-3’). For the gag region, we used during the first round, the primers GAG1 (5’-GGGAGACGTCTAGTATTAAGC-3’) and H1P202 (5’-CTAATACCTGTATCACTCCTGCTGT-3’); and in the second round, the primers GAG2 (5’-GGGAAAATCCGTGTAAAGCCGACGACGACCCCAACGACGGC-3’) and G17 (5’-TCCACATTTCACAGCCCTCTTTTTT-3’).

The sequencing was performed using the automated sequencer ABI3100, with the Taq FS Dye Terminator Cycle Sequencing Kit (Applied Biosystems), with the HTLV-1 primers: ATLTR11 and ATLTR12; and the HIV-1 primers: ED31, MM4, ES7 (5’-CTGTTAATGGCCAGCTACG-3’) and ED14 (5’-TCTTGCCTGAGCTTTTGATGCCTACGACGACGACCCCAACGACGGC-3’) for env; and GAG2, G17, H1G77 (5’-TCACCTAGAATTTTTTGAATGATCGTCGGG-3’) and M214 (5’-GAAACCAGCTTACATATCTCCT-3’) for gag. To determine subtype, we used the LASP HTLV-1 Automated Subtyping (Version 1.0) tool, available at http://www.bioafrica.net/rega-genotype/html/subtypinghtlv.html. To determine the HIV subtypes, we used the REGA HIV-1 & 2 Automated Subtyping Tool (Version 2.0), available at http://www.bioafrica.net/rega-genotype/html/subtypinghiv.html, which also has tools for the detection of recombinant forms using the Simplot algorithm.

**Ethical considerations**

This study was approved by the Research Ethics Committee of Escola Bahiana de Medicina e Saúde Publica under registration number 110/2010. Informed consent was obtained from all participants, authorizing the use of their samples for further studies. In case of minors, consent was signed by parents or guardians.

**Results**

A total of 943 pregnant women were enrolled in this study, in which 336 were from Ilhéus and 647 were from Itabuna. Participant’s age ranged from 12 to 47 years (average of 24.3 years). The number of pregnancies per woman ranged from 1 to 12 (with an average of 2.2), and most women (87.2%) reported they never have had history of miscarriage.

Of the total surveyed, 61 pregnant women had a positive serology using filter paper as collecting model, representing an overall prevalence of 6.47% (61/943), being 8.0% (27/336) in the city of Ilhéus, and 5.6% (34/607) in Itabuna. No cases of coinfection were observed. The specific prevalence for each pathogen was as follows: 0.1% (1/943) for HCV, 0.2% (2/943) for HBV, 0.5% (5/943) for HIV, 1.4% (13/943) for HTLV, 1.5% (14/943) IgM positive for T. gondii, and 2.8% (26/943) VDRL positive (T. pallidum).

Of the 61 positive pregnant women in the screening, only 29 (47.5%) returned to collect a venous sample and perform the confirmatory test. One of these had a positive serology for HCV (100% return - 1/1), two for HIV (20% - 2/10), nine for HTLV (69% - 9/13), nine for T. pallidum (35% - 9/26) and eight for T. gondii (57% -
8/14). Venous blood was collected upon a return visit. Serum from the collection was used to perform an Electrochemiluminescence (ECL) and PCR in case of HIV; ELISA and Western Blot for HTLV; VDRL and FTA-Abs for T. pallidum; IgM search for T. gondii, and anti-HCV search for HCV. After the new tests were performed, we confirmed: the HCV, two samples for HIV and all tested for T. pallidum. There were no false-positives for these three agents. However, of the nine positive samples for HTLV, only six were confirmed (33.3% were false-positives) and of the eight returns with positive results for T. gondii, only one was confirmed (Table 1).

Upon molecular characterization of retroviruses, the detected HTLV strains in 6 samples clustered within the Cosmopolitan subtype, Transcontinental subgroup (HTLV-1aA) (Table 2). On the other hand, the detected HIV strains in 2 samples clustered with subtypes B and F (F1) (Table 3).

**Discussion**

In Brazil, infectious diseases during pregnancy are relatively common, especially affecting disadvantaged populations, and creating challenges to the Public Health system, regarding the establishment of serological screening strategies. On the other hand, studies have revealed that the difficulty encountered by pregnant women to apply for laboratory tests is one of the most significant deficiencies in the quality of prenatal care [21]. It is also important to emphasize that women with multiple pregnancies could have a previous unknown serological status, with a higher possibility of vertical transmission to their children, and underestimating the prevalence of vertical infections.

A study conducted during 2000-2001 in Salvador (capital of Bahia) revealed that 30% of the 486 surveyed pregnant women did not undergo prenatal care, and most of them attended prenatal care after the second trimester, which reveal a failure in the service [17]. Additionally, the lack of a better effective and integrated network service among healthcare centers, laboratories, and hospitals to provide the results of vertically transmitted infections was another important concern raised in that study [15]. Another study found that only 35.4% of pregnant women enrolled in the Program for the Humanization of Prenatal and Birth (PHPN) in Brazil has been submitted to HIV testing [16]. According to Cavalcante and collaborators, one of the main barriers to reduce the rates of mother-to-child transmission of HIV, especially in low-income regions, is the complexity of operating a more standardized prenatal care, including the sampling methods and the performance of laboratory tests [18].

In this study, the average number of days to obtain the results of pregnant women was around 20 days. However, the healthcare professionals at the units reported that results for laboratory tests performed by traditional methods took up to 70 days to be available, making difficult any intervention strategies related to positive cases. Although the application of filter paper has already demonstrated to be an efficient method compared to traditional plasma and serum samples collection (including a reduction for the results of routine laboratory tests), we found that the lack of awareness and education of these pregnant women on the importance of prenatal care.

The filter paper methodology has showed a good level of accuracy compared to venipuncture, helping to reduce the waiting time for test results [15]. The sample collection on filter paper also represents a low-cost tool, with a possibility of higher operational capacity, and low-risks during sample transportation and storage, being a feasible method for public services in areas far from major centers and low-resources [14-19]. It
can be vertically transmitted in pregnant women from South region of Bahia. We also observed that health-care professionals and patients in the SUS prenatal service from that region face many difficulties, being the lack of knowledge of vertical transmission pointed as one of the main concerns, reinforcing the need for more incisive health education policies. Although we used tests that promote a fast diagnostic, the rate of return for confirmatory tests was still low, highlighting how the awareness of pre-natal care remains a challenge for the population. Finally, given the severity of the disorders associated to the screened pathogens in this study, checking for the presence or susceptibility to some of these infections needs to be reinforced, mainly for women living in low-resource areas with limited access to healthcare system.

Competing Interests
The authors declare no conflict of interests in this study.

Funding
Coordenação de Aperfeicoamento de Pessoal de Nível Superior (CAPES) and Fundação de Amparo ao Pesquisador do Estado da Bahia (FAPESB).

Acknowledgements
We would like to thank all the professionals from the Primary Care of SUS healthcare facilities in Ilhéus and

| Name                  | Length | Report | Assignment                          | Support | Genome                        |
|-----------------------|--------|--------|-------------------------------------|---------|-------------------------------|
| LTR10_consensus_sequenceNEW | 605bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 100.0   | HTLV-I Consolation HFK1       |
| LTR12_consensus_sequenceNEW | 605bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 100.0   | HTLV-I Consolation HFK1       |
| LTR14_consensus_sequenceNEW | 607bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 98.0    | HTLV-I Consolation HFK1       |
| LTR17_consensus_sequenceNEW | 606bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 96.0    | HTLV-I Consolation HFK1       |
| LTR20_consensus_sequenceNEW | 605bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 100.0   | HTLV-I Consolation HFK1       |
| LTR_24_consensus_sequenceNEW | 605bp  | Report | subtype_a(subgroup_A) (Transcontinental) | 94.0    | HTLV-I Consolation HFK1       |

*Only bootstrap values above 70% were considered significant and described.*
Itabuna cities, Bahia State. We also thank Filipe Almeida Rego for the support with molecular analysis.

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