Molecular docking analysis of penta galloyl glucose with the bcl-2 family of anti-apoptotic targets

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Abstract:
Apoptosis requires cellular proteins from the B-cell lymphoma 2 (BCL-2) family linked to breast cancer. Therefore, it is of interest to document the Molecular docking analysis data of penta galloyl glucose with the bcl-2 family of anti-apoptotic targets (Bcl-2, BCL-XL, Caspase 3, and Caspase 9). Data shows that Pentagalloyl glucose have optimal binding features with Bcl-2, BCL-XL, Caspase 3, and Caspase 9 proteins with binding energy of -8.6,-7,-7.5 and 4.4 kcal/mol respectively for further consideration in this context.

Keywords: Breast cancer, apoptosis, penta galloyl glucose, molecular docking

Background:
Breast cancer is the most frequent cancer among women around the world. Approximately 30% of women diagnosed with early-stage breast cancer go on to develop metastatic breast cancer, which necessitates therapy with anti-breast cancer therapeutic drugs. Although many modern anti-breast cancer medicines can slow
tumor development, the effect is usually short-lived, and patients frequently acquire resistance to the anticancer drugs utilized, resulting in about half of all treated patients relapsing [1]. This fact showed that new medicines that are more effective, safe, and have the ability to expand the lifespan of breast cancer patients are required. In multicellular organisms, the entire process of cell division and cell death is strictly regulated. When either of these biological processes is disrupted, it has an impact on normal development and homeostasis, which can lead to cancer. Cell proliferation pathways are enhanced during cancer progression, while programmed cell death or apoptotic pathways are decreased; thus, cancer has a repertoire of these failing processes [2]. Apoptosis has been researched in a variety of processes, including normal cell turnover, embryonic development, morphogenesis, aging, cell population regulation, and tumor progression, since its commencement [3]. Although apoptosis is a frequent process for removing undesired, damaged, or hazardous cells, other mechanisms such as anoikis, necroptosis, entosis, netosis, pyroptosis, or ferroptosis are also involved in programmed cell death [4]. Extrinsic stimuli such as viral infections, xenobiotics, and pollutants can trigger apoptosis, as can intrinsic factors such as DNA damage, reactive oxygen species (ROS), and reactive nitrogen species (RNS) [5]. Apoptosis is also involved in the clearance of immune cells that have been damaged by illness or noxious substances during immunological responses [6, 7].

The B-cell lymphoma-2 (Bcl-2) family, which includes both pro- and anti-apoptotic proteins, is important in the mitochondrial pathway of apoptosis because it promotes the release of cytochrome c and Smac (Second mitochondrial-derived activator of caspases) into the cytosol, resulting in caspase-dependent cell death [8,9]. Bak and Bax, multi-domain proteins, and Bim, Bid, Puma, and Noxa, BH3 domain only proteins, are members of the Bcl-2 family of pro-apoptotic proteins [10]. Bcl-xL, Bcl-2, and Mcl-1 are the most well-known anti-apoptotic proteins. The BH3-only proteins induce apoptosis via interacting with Bak and Bax or binding to anti-apoptotic proteins, causing Bak and Bax to be released [11]. As a result, the ratio of anti- and pro-apoptotic proteins in the Bcl-2 family determines whether a cell lives or dies. Anti-apoptotic Bcl-2 proteins are regularly expressed in most tumor tissues and cancer cell lines [12]. Prostate cancer, breast cancer, B-cell lymphomas, and colorectal cancer, for example, all have high levels of Bcl-2. Overexpression of Bcl-2 family anti-apoptotic proteins promotes cell proliferation and survival, as well as therapeutic resistance due to apoptosis evasion. As a result, they are well-established pharmacological targets for cancer therapy. The development of anti-apoptotic protein Bcl-2 inhibitors has a lot of clinical potential.

Molecular docking
AutoDock in PyRx PyRx (0.8) GUI was used to check the binding capability of the selected antibiotics with each selected target. Docking provides valuable insight into the interaction between the proteins with ligand. The structures of the Pentagalloylglucose were prepared before docking experiment by using make ligand command present in PyRx. Make macromolecule option was used to prepare the protein structures. AutoDock program along with the Lamarckian genetic algorithms (LGA) was used to perform docking experiments. The Lamarckian GA parameters used in the analysis consist of 30 independent runs, 150 size of population, a 25,000,000 energy evaluations, 27,000 number of generation, mutation rate of 0.02, and 0.8 crossover rate. Docking was carried out with in the pre-defined grid size for protein. For each ligand protein complex, individual docking procedures have been performed. The results were ranked in the order of increasing docking energies. The lowest binding energy of each cluster was assumed to be representative [15].

Results and Discussion
Docking experiments on Pentagalloylglucose compounds and anti-apoptotic proteins were conducted. The interactions between anti-apoptotic proteins and Pentagalloylglucose, as well as the binding energies of the complexes, were used to examine the AutoDock results. The accuracy of the AutoDock data was confirmed by looking at the anti-apoptotic proteins' lowest binding free energy and hydrogen bonds with penta galloyl glucose. Table 1 shows the results of the docking analysis.

Table 1: Molecular docking results of penta galloyl glucose with selected target proteins

| S. No | Protein | Binding energy (Kcal/mol) | H-bond interaction |
|-------|---------|---------------------------|--------------------|
| 1     | BCL-2   | -8.6                      | PRO-120, PHE-121, ARG-124, ASN-169, TRP-173, GLU-176 |
| 2     | BCL-XL  | -7                        | ARG-100, ASN-136  |
Docking with BCL-2
Bcl-2 was first discovered as an oncogene implicated in human B cell follicular lymphoma [16]. Vaux et al. found this protein’s anti-apoptotic action in a system where apoptosis was caused by IL-3 deficiency in 1988 [17]. Bcl-2 was soon found to prevent apoptosis generated by a variety of stressors, including serum deprivation, heat shock, and chemotherapeutic chemicals, implying that Bcl-2 can prevent apoptosis through a common mechanism. Certain types of necrotic cell death are also inhibited by Bcl-2. Bcl-2 was found to prevent apoptosis through a common mechanism.

Docking with BCL-XL
JNK signaling targets Bcl-xL, a major antiapoptotic molecule. In response to microtubule inhibitors and other apoptotic stimuli like ionizing radiation or chemotherapy, Bcl-xL is phosphorylated [19]. BCL-XL was chosen as one of the probable targets for molecular docking with Pentagalloylglucose in this investigation. According to the docking data, BCL-XL has a binding affinity for Pentagalloylglucose by forming three hydrogen bonds with binding energies of 7 kcal/mol. It shows strong binding with bcl-xl, and the interaction of this docking is depicted in Figure 1.

Docking with Caspase-3
Caspases play an important role in programmed cell death (apoptosis). Caspase-3 is a commonly activated death protease that catalyzes the selective cleavage of a variety of important cellular proteins. Figure 1 and Table 1 demonstrate the molecular docking results of caspase 3 with Pentagalloylglucose. Pentagalloylglucose has a high affinity for caspase 3, according to the docking data. The docking results revealed that in the creation of three hydrogen bonds, interacted with binding energies of -7.5 kcal/mol.

Docking with Caspase-9
Caspases-9, the most studied initiator caspase, is a crucial component in the intrinsic or mitochondrial pathway, which is activated by a variety of stimuli including as chemotherapies, stress agents, and radiation. Caspase-9 is hypothesized to involve homodimerization monomeric zymogens and is triggered on the apoptosome complex to maintain catalytic state. Failure to activate caspase-9 has far-reaching physiological and pathophysiological consequences, including degenerative and developmental problems, as well as cancer. Caspase-9 docking studies with Pentagalloylglucose revealed an interaction with a binding energy of -4.4 kcal/mol and the formation of five hydrogen bonds. All these interaction were shown in the figure 1 and the details of interaction were shown in Table 1. Docking simulation research revealed that all of the identified targets implicated in apoptotic proteins had a robust interaction behavior with Pentagalloylglucose in this investigation. It revealed that Pentagalloylglucose could be a promising anticancer candidate worthy of future investigation.

Conclusions:
Our findings show that Pentagalloylglucose have optimal binding features with Bcl-2, BCL-XL, Caspase 3, and Caspase 9 proteins with binding energy of -8.6,-7,-7.5 and 4.4 kcal/mol respectively for further consideration in this context.

Conflict of Interest:
There is no conflict of interest from any of the authors.

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