Probiotic Supplements Improve Blood Glucose and Insulin Resistance/Sensitivity among Healthy and GDM Pregnant Women: A Systematic Review and Meta-Analysis of Randomized Controlled Trials

Yu-Qing Pan,1,2 Qing-Xiang Zheng1,2, Xiu-Min Jiang2,3, Xiao-Qian Chen2, Xiao-Yun Zhang4, and Jing-Ling Wu4

1School of Nursing, Fujian Medical University, No. 1 Xuefu North Road, University Town, Shangjie Zhen, Minhou County, Fuzhou City, Fujian Province, China
2Fujian Maternity and Child Health Hospital Affiliated to Fujian Medical University, No. 18 Daoshan Street, Gulou District, Fuzhou City, Fujian Province, China
3Fujian Obstetrics and Gynecology Hospital Affiliated to Fujian Medical University, No. 18 Daoshan Street, Gulou District, Fuzhou City, Fujian Province, China
4School of Nursing, Fujian University of Traditional Chinese Medicine, Fuzhou, China

Correspondence should be addressed to Xiu-Min Jiang; jzc0427@163.com

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Background. Probiotic supplements may be seen as a promising way to improve glucose metabolism. This study aimed to evaluate the effects of probiotic supplements on blood glucose, insulin resistance/sensitivity, and prevention of gestational diabetes mellitus (GDM) among pregnant women. Methods. Eleven electronic databases were searched from inception to May 2020. Two authors independently identified randomized controlled trials (RCTs), assessed the eligibility and quality of the included studies, and then extracted data. The primary outcomes were fasting plasma glucose (FPG), 1 h and 2 h plasma glucose after 75 g oral glucose tolerance test (OGTT), HbA1c, fasting plasma insulin, insulin resistance, and insulin sensitivity. Fixed and random effect models were used to pool the results. Results. A total of 20 RCTs involving 2972 participants were included according to the inclusion and exclusion criteria. The pooled results of this research showed that probiotic supplements could reduce the level of FPG (mean difference (MD) = −0.11; 95% CI = −0.15 to −0.04; P = 0.0007), serum insulin (MD = −1.68; 95% CI = −2.44 to −0.92; P < 0.00001), insulin resistance (MD = −0.36; 95% CI = −0.53 to −0.20; P < 0.00001), and insulin sensitivity (MD = −21.80; 95% CI = −31.92 to −11.67; P < 0.00001). Regarding the subgroup analysis of different pregnant women, the effects of probiotics on FPG, insulin, and insulin resistance were more obvious among GDM and healthy women than among overweight/obese women. Furthermore, the differences were not significant in HbA1c (MD = −0.05; 95% CI = −0.12 to 0.03; P = 0.23), 1 h OGTT (MD = −0.07; 95% CI = −0.25 to 0.10; P = 0.42), and 2 h OGTT (MD = −0.03; 95% CI = −0.17 to 0.12; P = 0.72). Conclusion. This review found that probiotic supplements had certain functions to reduce the level of FPG and improve insulin, insulin resistance, and insulin sensitivity, especially for GDM and healthy pregnant women.

1. Introduction

Gestational diabetes mellitus (GDM) is the most common pregnancy complication, and its prevalence is continually rising worldwide due to increased obesity and the average age of pregnant women [1], from 4.5% to 20.3% in the Western Pacific [2] and 14.8% in China [3]. Overweight and obesity can contribute to half of GDM’s prevalence [4]. Meanwhile, obesity occurs in up to 30% of women [5], and obese women have a higher risk for GDM in comparison
with normal-weight women [6]. Both GDM and obesity induce metabolic traits, including hyperglycemia, hyperinsulinemia, and insulin resistance [7, 8], as well as imposing a huge economic burden [1, 9, 10], especially in developing countries. GDM and obesity also cause ongoing maternal and neonatal health problems [2, 7, 8, 11–13]. Women with GDM are more likely to develop diabetes at rates of 20%–60% in five to ten years after pregnancy, and the incidence of metabolic diseases in their offspring also significantly increases [13]. Therefore, the prevention of obesity may be directly associated with a lower risk of GDM, and the prevention and treatment of hyperglycemia and insulin resistance among pregnant women have become a global concern.

Lifestyle intervention is the major method to control maternal hyperglycemia and insulin resistance, including medical nutrition therapy, exercise intervention, and self-monitoring of blood glucose [14]. Women with GDM whose blood sugar cannot be controlled at an ideal level by diet or exercise should be accepted for pharmacological therapy [14]. Although insulin therapy is the most common and safest pharmacological therapy, it is very labor-intensive and time-consuming for nurses and creates a financial burden on women with GDM [9]. Oral medications, such as metformin and sulfonylureas, are associated with higher risks for adverse pregnancy outcomes, such as large-for-gestational age, neonatal hypoglycemia, and birth injury. Pregnant women may also face many barriers during clinical implementation [15–17]. Owing to the poor management of lifestyle intervention and the limitations of pharmacotherapy, seeking a better way to improve hyperglycemia and insulin resistance is essential.

Probiotic is defined as beneficial live microorganisms in the host when they reach an adequate dose [18], and it plays an important role in improving the intestinal microenvironment, modulating the immune system, and preventing systemic disease and inflammation [19]. Previous studies have demonstrated that gut microbiota promotes the digestion of complex polysaccharides to produce monosaccharides and short-chain fatty acids (SCFAs) [20], which have positive relations with metabolism [21]. The composition and function of obese-diabetic microbiota change in comparison with healthy gut microbiota, which leads to metabolic disorders such as overweight/obesity, elevated blood glucose, insulin resistance, and inflammation [22, 23]. Accordingly, probiotics may be seen as a potential, economic, and practical approach to improve blood glucose and insulin resistance/sensitivity among pregnant women.

Even though previous studies indicated the effects of probiotics in preventing and treating GDM, some conflicting conclusions were still drawn from other studies. Zhang et al. (2019) [24] showed that probiotics could reduce the blood glucose level of women with GDM. On the contrary, several system reviews indicated that probiotic supplements did not reduce the blood glucose of women with GDM in comparison with the placebo group [25–27]. Moreover, in another two system reviews [28, 29], such probiotic supplements had a positive effect on women without a GDM diagnosis. Less is also known about the effects of probiotics on different pregnancy status as well as the duration, dosages, and type of probiotic interventions.

This study aimed to synthesize more high-quality randomized controlled trials (RCTs) to provide evidence of the effects of probiotic supplements on glycemic control, insulin resistance/sensitivity, and prevention of GDM among pregnant women.

2. Method

This review completely followed the PRISMA guidelines [30].

2.1. Search Strategy. We searched 11 electronic databases (Embase, Web of Science, Medline, PubMed, Cochrane, CINAHL, Clinicaltrials.gov, China National Knowledge Infrastructure, WanFang Data, Chinese Scientific Journal Database, and SinoMed) from April 2020 to May 2020. The search strategy combined MeSH terms and free words that were listed as follows: “pregnant,” “obstetric,” “obesity,” “overweight,” “gestational diabetes,” “gestational diabetes mellitus,” “probiotic,” “symbiotic,” “lactobacilli,” “streptococcus,” “bifidobacter,” “saccharomy,” “yeast,” “yogurt,” and “bacteria.” The retrieval was adjusted to different features of each database. To expand the retrieval area and advance the recall of the search engine, the reference list of included studies and relevant reviews were further tracked through the snowballing method.

2.2. Selection Criteria. The inclusion criteria of this study were as follows: (1) RCTs; (2) studies published in Chinese or English; (3) pregnant women with or without overweight or obesity (body mass index (BMI) 25.0–29.9 kg/m2 or ≥30.0 kg/m2 at the first antenatal visit, resp.), GDM (diagnosed in the second or third trimester of pregnancy, without identified diabetes before gestation), and over 16 years old; and (4) probiotic supplement was used as an intervention method.

Studies were excluded if they constituted duplicated publication, the information of glucose index (e.g., fasting glucose, insulin, and HOMA-IR) was not reported, or the relevant data could not be extracted.

2.3. Data Extraction. Two review authors (Pan and Zheng) independently selected literature according to the inclusion and exclusion criteria after screening the title, abstract, and full text and then extracted data. The information we extracted from the eligible studies included (1) basic information of the study: first author, year of publication, country, and sample size; (2) characteristics of participants: age and BMI at baseline; (3) intervention details: probiotic species, dose, frequency, and duration; (4) primary outcomes, including fasting plasma glucose (FPG), 1 h and 2 h plasma glucose post 75 g oral glucose tolerance test (OGTT), and glycated Hb (HbA1c); and (5) secondary outcomes, including fasting plasma insulin (FPI), homeostasis model assessment insulin resistance (HOMA-IR), homeostasis
model of assessment-estimated β cell function (HOMA-B), quantitative insulin sensitivity check index (QUICKI), and incidence of GDM. We resolved any disagreement through discussion or turned to a third reviewer (Jiang) when consensus was not reached.

2.4. Quality Assessment. Two review authors (Pan and Zheng) used the Cochrane risk of bias tool, Review Manager 5.3, to evaluate independently the methodological quality of each RCT. The classification of risk bias included (1) random sequence generation, (2) allocation concealment, (3) blinding of participants and personnel, (4) blinding of outcomes assessment, (5) incomplete outcome data, (6) selective reporting, and (7) other biases. Each classification was rated as “high risk,” “low risk,” or “unclear.” Once more than one entry was assessed as “high risk,” the quality of study would be regarded as high risk for bias. Any disagreement was resolved through discussion or by consulting a third reviewer (Jiang).

2.5. Data Synthesis. Review Manager software (version 5.3) was used for meta-analysis following the Cochrane handbook [31]. Two review authors (Pan and Zheng) cross-checked the entered data to ensure that they were strictly correct. Continuous variables were calculated with the mean difference (MD). Statistical heterogeneity was assessed with the Chi² and I². If P < 0.01 for the Chi² test and I² < 50% indicated that heterogeneity was not significant, then the fixed-effects model was utilized to merge the results. Otherwise, the meta-analysis used the random-effects model (P > 0.01 for the Chi² test and I² > 50%). Moreover, to explore the potential clinical heterogeneity, a subgroup analysis was performed for each group meta-analysis according to different participants, types, dosages, and duration of probiotic intervention. If I² was still greater than 50%, then the sensitivity analysis was conducted to guarantee the stability of results and remove the dubious studies. The descriptive analysis method was adopted if substantial heterogeneity still existed. The 95% confidence intervals (95% CI) were calculated in each statistical analysis, and P < 0.05 was regarded as significant for analysis. A funnel plot test was utilized to assess potential publication bias if more than 10 studies were included.

3. Results

A total of 4,644 articles were identified by searching 11 electronic databases. Fifty-two studies were kept after irrelevant articles (n = 4474), duplication (n = 98), and reviews (n = 20) were removed through screening titles and abstracts. Of the remaining studies retrieved, ten studies were protocols, two studies were reviews, and eight studies were duplicated publications. Among the rest of the studies, two studies did not utilize probiotics to intervene, eight studies reported irrelevant outcomes, and one study was a case report. The lone Chinese literature just reported the incident of GDM without any other relevant index. Hence, we finally included 20 RCTs based on a careful checking of the full text according to the selection criteria (Figure 1).

3.1. Characteristics of Included Studies. Twenty studies [32–51] with a total of 2,972 participants were included (Table 1). Each study had an average of 148 participants, ranging from 50 to 433, from New Zealand, Australia, Iran, Ireland, Finland, and Thailand. The participants were divided into three subgroups: overweight or obese pregnant women, women with GDM, and healthy pregnant women. The intervention types included probiotic capsules (16 studies [32–34, 36–45, 48, 51]), food (one study [47]), and probiotic yogurt (three studies [35, 46, 50]). Different species and combinations of probiotics were used in each study. The frequency of intervention in most studies was once per day. The duration of intervention in included studies was four weeks, six weeks, eight weeks, nine weeks, and from enrolment until delivery. According to the measurement time of each outcome indicator, this review divided the duration into two subgroups: short term (≤12 weeks) and long term (>12 weeks). The most common species of probiotics included Lactobacillus rhamnosus, Bifidobacterium lactis, Lactobacillus acidophilus, Lactobacillus casei, and Lactobacillus fermentum. The doses of probiotics ranged from 10⁶ colony-forming units (CFUs) to 10¹¹ CFUs, and the most common dose was 10⁹ CFUs. The subgroups were set to small dose (<10⁹ CFU) [33–35, 39, 46, 47, 50] and large dose (≥10⁹ CFU) [32, 36–38, 40–45, 48, 49, 51]. Table 1 shows the characteristics of all included studies.

3.2. Quality Assessments of the Literature. Figure 2 shows the methodological quality and risk for the bias of all included studies. Random sequence generation and allocation concealment were conducted in 18 included studies. Blinding of participants and personnel was reported in 19 studies, of which 5 studies reported triple blinding. A total of 7 studies used intention-to-treat analysis, 8 studies were considered “low risk for bias,” and only one study was evaluated as “high risk” for selective reporting. There was only one study (5%) with a small sample size, which was less than or equal to 50 participants.

3.3. Effect of Probiotic Supplements on Blood Sugar

3.3.1. Assessment of Efficiency on FPG. In 20 studies with 2,555 participants, the FPG level was reduced in the intervention group (MD = −0.11; 95% CI = −0.15 to −0.04; P = 0.0007) compared with that in the control group (Table 2). For the effects of probiotic supplementation on different participants, the pooled results indicated that the intervention group significantly differed from the placebo group among women with GDM (MD = −0.12; 95% CI = −0.24 to −0.01; P = 0.04) and healthy pregnant women (MD = −0.07; 95% CI = −0.11 to −0.02; P = 0.004). However, there were no significant differences in overweight/obese women for the FPG level. After sensitivity analyses, more concentrated results were obtained in the overweight/obese
subgroup, changing from (MD = −0.09; 95% CI = −0.20 to 0.02; \( P = 0.12 \)) to (MD = −0.13; 95% CI = −0.21 to −0.06; \( P = 0.0006 \)), and the 95% CI (−0.24 to −0.01) was shortened (−0.32 to −0.13) in the subgroup of GDM. When the different duration of probiotic intervention was considered, all the results illustrated that short-term (MD = −0.13; 95% CI = −0.22 to −0.04; \( P = 0.006 \)) intervention benefited pregnant women to improve the FPG level, whereas long-term intervention had no benefits. After sensitivity analysis, both short-term (MD = −0.12; 95% CI = −0.20 to −0.03; \( P = 0.006 \)) and long-term (MD = −0.11; 95% CI = −0.18 to −0.05; \( P = 0.0005 \)) interventions were effective and were only slightly different. In terms of the intervention types, probiotic capsule (MD = −0.008; 95% CI = −0.17 to −0.03; \( P = 0.008 \)) improved the FPG level more than probiotic yogurt did (MD = −0.09; 95% CI = −0.18 to 0.00; \( P = 0.05 \)), which was merely at the junction with zero boundaries. Only one study [47] reported probiotics in the form of diet. Significant heterogeneity (\( I^2 = 77\% \)) among the probiotic capsule subgroup was decreased after three skeptical studies were removed, and the result of the meta-analysis also improved precision. A large dose of probiotic supplement was statistically significant (MD = −0.10; 95% CI = −0.17 to −0.03; \( P = 0.004 \)).

3.3.2. Assessment of Efficiency on 1h and 2h OGTT and HbA1C. Six included studies reported the OGTT, including 1,434 participants of 1h OGTT and 1,491
| Article, year (country) | Participants characteristics | Mean age, BMI (intervention/control) | Intervention/control (sample size) | Probiotic species and dose | Frequency and duration | Outcomes |
|-------------------------|-----------------------------|--------------------------------------|-----------------------------------|---------------------------|------------------------|----------|
| Oksene-Gafa et al., 2019 (New Zealand) [32] | Pregnant women with obesity | 28.9 ± 5.7/28.6 ± 5.7, 38.9 ± 6.5/38.2 ± 5.7 | Probiotic/placebo (115/111) | Lactobacillus rhamnosus LGG and Bifidobacterium lactis Bb12 (6.5 × 10⁹ CFU) Lactobacillus rhamnosus LGG and Bifidobacterium animalis subspecies lactis Bb12 (>1 × 10⁹ CFU) | 1 capsule daily from 12¹⁰⁻¹⁷⁺⁶ weeks of gestation until delivery | OGTT, HbA1c, and incident of GDM |
| Callaway et al., 2019 (Australia) [33] | Overweight or obese pregnant women | 31.3 ± 4.7/31.7 ± 4.8, 31.9 ± 7.5/31.6 ± 7.2 | Probiotic/placebo (219/214) | | 1 capsule daily from enrolment (15.9 ± 1.9) until delivery | OGTT and incident of GDM |
| Lindsay et al., 2014 (Ireland) [34] | Pregnant women with obesity | 31.4 ± 5.0/31.0 ± 5.2, 32.9 ± 2.4/34.1 ± 2.7 | Probiotic/placebo (64/76) | Lactobacillus salivarius UCC118 (10⁹ CFU) | 1 capsule daily from 24 weeks to 28 weeks of gestation | FBG, insulin, HOMA-IR, and incident of GDM |
| Asgharian et al., 2020 (Iran) [35] | Pregnant women with obesity | 29.5 ± 6.2/29.4 ± 5.5, 29.2 ± 3.3/30.3 ± 4.1 | Probiotic yogurt/conventional yogurt (83/82) | Lactobacillus acidophilus La5 and Bifidobacterium lactis Bb12 (10⁸ CFU) | 100g/day from 24 weeks of gestation until delivery | FPG, OGTT, and incident of GDM |
| Pellonperä et al., 2019 (Finland) [36] | Overweight or obese pregnant women | 30.4 ± 4.8/30.8 ± 4.8/30.8 ± 4.6/30.4 ± 4.1, 30.0 ± 4.2/29.9 ± 4.7/29.3 ± 3.9/29.7 ± 4.2 | Fish oil + placebo/fish oil + probiotics + placebo/fish oil + probiotics/placebo + placebo(110/109/109/110) | Lactobacillus rhamnosus HN001 and Bifidobacterium animalis ssp. lactis 420 (10⁹ CFU) | Daily consumption from randomization (12.5 ± 3.1) until delivery | OGTT, FPG, insulin, HOMA-IR, and incident of GDM |
| Jafarnejad et al., 2016 (Iran) [37] | Pregnant women with GDM | 32.4 ± 3.1/31.9 ± 4.0, 26.8 ± 2.7/27.4 ± 3.1 | Probiotic/placebo (41/41) | | Twice per day from 16 weeks of gestation for 8 weeks | FPG, HbA1c, insulin, and HOMA-IR |
| Karamali, 2016 (Iran) [38] | Pregnant women with GDM | 31.8 ± 6.0/29.7 ± 4.0, 28.6 ± 4.2/28.5 ± 3.4 | Probiotic/placebo(30/30) | Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum (6 × 10⁶ CFU/g) Lactobacillus delbrueckii (112.5 × 10⁶ CFU) Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum (6 × 10⁶ CFU/g) | A daily capsule from diagnosis for 6 weeks | FPG, insulin, HOMA-IR, HOMA-B, and QUICKI |
| Lindsay et al., 2015 (Ireland) [39] | Pregnant women with GDM | 33.5 ± 5.0/32.6 ± 4.5, 29.06 ± 6.70/28.94 ± 5.79 | Probiotic/placebo (74/75) | Lactobacillus salivarius UCC118 (10⁹ CFU) | 1 capsule daily from diagnosis until delivery | FPG, insulin, HOMA-IR, and HOMA-IR |
| Article, year (country) | Participants characteristics | Mean age, BMI (intervention/control) | Intervention/control (sample size) | Probiotic species and dose | Frequency and duration | Outcomes |
|------------------------|-----------------------------|--------------------------------------|-----------------------------------|---------------------------|-----------------------|----------|
| Ahmadi et al., 2016 (Iran) [40] | Pregnant women with GDM | 28.5 ± 5.8/28.7 ± 3.4, 28.7 ± 4.5/28.4 ± 2.7 | Symbiotic/placebo (35/35) | Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum (6 x 10⁶ CFU/g), L. acidophilus (5 x 10⁶ CFU/g), L. plantarum (1.5 x 10⁶ CFU/g), and L. fermentum (7 x 10⁶ CFU/g), and L. gasseri (2 x 10⁶ CFU/g) | A daily capsule from diagnosis for 6 weeks | FPG, insulin, HOMA-IR, HOMA-B, and QUICKI |
| Nabhani et al., 2018 (Iran) [41] | Pregnant women with GDM | 29.4 ± 5.8/30.3 ± 5.6, 26.4 ± 4.1/28.2 ± 4.7 | Symbiotic/placebo (47/48) | L. acidophilus (5 x 10¹⁰ CFU/g), L. plantarum (1.5 x 10¹⁰ CFU/g), L. fermentum (7 x 10⁹ CFU/g), and L. gasseri (2 x 10⁹ CFU/g) | A daily capsule from diagnosis for 6 weeks | FPG, insulin, HOMA-IR, and QUICKI |
| Kijmanawat et al., 2019 (Thailand) [42] | Pregnant women with GDM | 32.50 ± 5.02/30.72 ± 5.05,- | Probiotic/placebo (30/30) | Streptococcus thermophilus (STY-31), and Lactobacillus delbrueckii bulgaricus (LBY-27) (>4 x 10⁸ CFU) | 1 capsule daily from diagnosis for 8 weeks | FPG, insulin, HOMA-IR, and QUICKI |
| Dolatkhah et al., 2015 (Iran) [43] | Pregnant women with GDM | 28.14 ± 6.24/26.48 ± 5.23, 31.41 ± 3.92/29.86 ± 3.39 | Probiotic/placebo (32/32) | Lactobacillus acidophilus, Bifidobacterium bifidum, and Lactobacillus fermentum (8 x 10⁶ CFU/g) | 1 capsule daily from diagnosis for 6 weeks | FPG, insulin, HOMA-IR, and QUICKI |
| Babadi et al., 2019 (Iran) [44] | Pregnant women with GDM | 28.8 ± 4.3/29.0 ± 4.2/26.5 ± 2.7 | Probiotic/placebo (25/25) | Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum, and Lactobacillus fermentum (8 x 10⁶ CFU/g) | Daily capsule for 6 weeks | FPG, insulin, HOMA-IR, and QUICKI |
| Jamilian et al., 2018 (Iran) [45] | Pregnant women with GDM | 31.2 ± 5.9/29.9 ± 3.7, 26.4 ± 4.2/27.5 ± 3.3 | Probiotic/placebo (30/30) | Lactobacillus acidophilus, Bifidobacterium bifidum, L. reuteri, and Lactobacillus fermentum (8 x 10⁶ CFU/g) | Daily capsule for 6 weeks | FPG, insulin, HOMA-IR, and QUICKI |
| Sahhaf et al., 2019 (Iran) [46] | Pregnant women with GDM | 31.64 ± 5.97/31.67 ± 5.49, 31.67 ± 5.44/29.67 ± 3.03 | Probiotic yogurt/conventional yogurt (42/42) | Lactobacillus acidophilus and Bifidobacterium lactis (10⁶ CFU) | 300 mg/day for 8 weeks | FPG and HbA1c |
| Taghizadeh et al., 2014 (Iran) [47] | Healthy pregnant women | 26.4 ± 6.3/29.0 ± 4.6, 27.9 ± 5.1/28.2 ± 4.1 | Symbiotic food/control food (28/28) | Lactobacillus sporogenes (1 x 10⁶ CFU/g) | Twice times per day with 18 g for 9 weeks | FPG, insulin, HOMA-IR, HOMA-B, and QUICKI |
participants of 2 h OGTT (Table 2). The results of the meta-analysis indicated that 1 h OGTT (MD = -0.07; 95% CI = -0.25 to 0.10; \( P = 0.42 \)) and 2 h OGTT (MD = -0.03; 95% CI = -0.17 to 0.12; \( P = 0.72 \)) did not have a significant difference between the intervention and control groups. In the subgroup analyses stratified by pregnancy status and intervention form, the results of 1 h OGTT (MD = -0.04; 95% CI = -0.24 to 0.16; \( P = 0.71 \)) and 2 h OGTT (MD = 0.01; 95% CI = -0.15 to 0.18; \( P = 0.89 \)) among overweight/obese pregnant women did not differ from those among the control group; probiotic capsule did not have a significant reduction in 1 h OGTT (MD = -0.23; 95% CI = -0.24 to 0.70; \( P = 0.75 \)) and 2 h OGTT (MD = 0.03; 95% CI = -0.11 to 0.18; \( P = 0.65 \)). Only one study reported OGTT in the subgroup of healthy women and probiotic yogurt. Neither of the above two subgroups drew statistical differences between the intervention and control groups. In the outcomes of 1 h and 2 h OGTT, a subgroup analysis of dosage could not be performed because of all included RCTs that adopted a large dose of probiotic bacteria. Meanwhile, HbA1c was reported in a total of 389 pregnant women in three studies. The results showed no significant difference (MD = -0.05; 95% CI = -0.12 to 0.03; \( P = 0.23 \)) between the intervention and control groups. No heterogeneity existed (\( I^2 = 0 \% \)), and there were no available studies to conduct a subgroup analysis.

### 3.4. Effect of Probiotics on Insulin

#### 3.4.1. Assessment of Efficiency on Fasting Plasma Insulin

Information on the FPI level was measured in 16 studies involving 1,446 participants (Table 2). A significant reduction was observed in comparison with the intervention group and the control group (MD = -1.68; 95% CI = -2.44 to -0.92; \( P < 0.00001 \)). Compared with the pregnancy status of overweight/obese, the level of FPI among women with GDM (MD = -2.40; 95% CI = -3.70 to -1.09; \( P = 0.0003 \)) and healthy women (MD = -1.77; 95% CI = -2.40 to -1.14; \( P < 0.00001 \)) was significantly decreased. After sensitivity analysis, the results were more precise and \( I^2 \) values were reduced to zero. For different probiotic intervention durations, short-term intervention could promote insulin control (MD = -1.55; 95% CI = -2.17 to -0.93; \( P < 0.00001 \)) instead of long-term intervention (MD = 0.42; 95% CI = -2.38 to 3.18; \( P = 0.77 \)). After sensitivity analysis, the results of the short-term intervention were stable, but long-term...
3.4.2. Assessment of Efficiency on HOMA-IR and HOMA-B. There were 16 studies involving 1,383 participants, where the measured HOMA-IR was significantly improved (MD = −0.36; 95% CI = −0.53 to −0.20; P < 0.00001) in comparison with the control group (Table 2). For different pregnancy status, the results illustrated that probiotic supplements in women with GDM (MD = −0.59; 95% CI = −0.88 to −0.30; P < 0.00001) and healthy women (MD = −0.34; 95% CI = −0.48 to −0.20; P < 0.00001) had a positive effect on HOMA-IR. However, positive results could not be obtained in overweight/obese women regardless of whether a sensitivity analysis was conducted or not. With regard to the different duration of interventions, the results of subgroup analyses showed that short-term intervention (MD = −0.47; 95% CI = −0.64 to −0.31; P < 0.00001) reaped benefits; long-term intervention had no positive result (MD = −0.15; 95% CI = −0.61 to 0.32; P = 0.53), but the result of such intervention changed after sensitivity analysis (MD = −0.27; 95% CI = −0.47 to −0.08; P = 0.006). Moreover, four studies with 242 participants measured HOMA-B (Table 2). Similarly, the results of the meta-analysis indicated that probiotics had positive effects on reducing the HOMA-B (MD = −21.80; 95% CI = −31.92 to −11.67; P < 0.00001). When it came to different pregnancy status, the pooled results showed that probiotics were also beneficial to women with GDM (MD = −25.25; 95% CI = −39.04 to −11.47; P = 0.0003) and healthy women (MD = −17.74; 95% CI = −32.66 to −2.82; P = 0.02). For the probiotic capsule subgroup, the pooled effectiveness on HOMA-IR (MD = −0.40; 95% CI = −0.63 to −0.18; P = 0.0004) and HOMA-B (MD = −22.76; 95% CI = −32.32 to −12.19; P < 0.00001) was shown to have significant differences. Still, the dosage of probiotics did not affect the effect on HOMA-IR and HOMA-B.

3.4.3. Assessment of Efficiency on QUICKI. Nine studies enrolling 582 participants measured QUICKI (Table 2). QUICKI was improved in the probiotic group in comparison with the control group (MD = 0.01; 95% CI = 0.00 to 0.03; P = 0.001). For different pregnancy status, the results of this meta-analysis indicated that QUICKI in women with GDM (MD = 0.00; 95% CI = 0.00 to 0.01; P = 0.03) and healthy women (MD = 0.02; 95% CI = 0.01 to 0.03; P = 0.001) had a slight difference compared with women in the control group. Regarding the different types of probiotics, significant differences were shown in the probiotic capsule (MD = 0.01; 95% CI = 0.00 to 0.01; P = 0.002). Given the number of studies, meta-analysis was conducted only for the large-dose group, and a statistical difference was observed between intervention groups and placebo groups.

3.5. Effect of Probiotics on the Incidence of GDM. The incidence of GDM was measured in seven studies with a total of 1,645 participants. There was no statistical difference between the intervention and control groups (RR = 1.03; 95% CI = 0.94 to 1.18; P = 0.67). For different pregnant women, the same result was obtained (RR = 1.09; 95% CI = 0.95 to 1.25; P = 0.24). Regarding the different duration
Table 2: Meta-analysis of the effects of probiotics in different subgroups.

| Outcome or subgroup | Studies | N   | Statistical method                       | Effect estimate       | P         | I² (%) |
|---------------------|---------|-----|------------------------------------------|-----------------------|-----------|--------|
| FPG                 | 20      | 2555| Mean difference (IV, random, 95% CI)     | −0.11 [−0.17, −0.04]  | 0.0007    | 72     |
| Different pregnancy status for FPG | 6 | 1233 | Mean difference (IV, random, 95% CI) | −0.09 [−0.20, 0.02] | 0.12  | 78     |
| GDM                 | 10      | 740 | Mean difference (IV, random, 95% CI)     | −0.12 [−0.24, −0.01]  | 0.04  | 76     |
| Healthy             | 4       | 579 | Mean difference (IV, random, 95% CI)     | −0.07 [−0.11, −0.02]  | 0.004    | 0      |
| Different intervention durations for FPG | 12 | 827 | Mean difference (IV, random, 95% CI) | −0.13 [−0.22, −0.04] | 0.006 | 74     |
| Long term           | 8       | 1725| Mean difference (IV, random, 95% CI)     | −0.08 [−0.17, 0.00]  | 0.06  | 72     |
| Different intervention forms for FPG | 3 | 282 | Mean difference (IV, random, 95% CI) | −0.09 [−0.25, 0.04] | 0.15  | 29     |
| Food                | 1       | 52  | NA                                       | NA                    | NA  | NA     |
| Capsule             | 16      | 2221| Mean difference (IV, random, 95% CI)     | −0.10 [−0.17, −0.03]  | 0.008  | 77     |
| Different probiotic bacteria doses for FPG | 3 | 206 | Mean difference (IV, random, 95% CI) | −0.11 [−0.25, 0.04] | 0.15  | 29     |
| 1 h OGTT            | 6       | 1434| Mean difference (IV, fixed, 95% CI)      | −0.07 [−0.25, 0.10]  | 0.42  | 0      |
| Different pregnancy status for 1 h OGTT | 5 | 1060 | Mean difference (IV, fixed, 95% CI) | −0.04 [−0.24, 0.16] | 0.71  | 11     |
| Healthy             | 1       | 374 | NA                                       | NA                    | NA  | NA     |
| Different intervention forms for 1 h OGTT | 1 | 125 | NA                                       | NA                    | NA  | NA     |
| Yogurt              | 5       | 1309| Mean difference (IV, fixed, 95% CI)      | 0.23 [−0.24, 0.70]   | 0.75  | 0      |
| 2 h OGTT            | 6       | 1491| Mean difference (IV, fixed, 95% CI)      | −0.03 [−0.17, 0.12]  | 0.72  | 45     |
| Different pregnancy status for 2 h OGTT | 5 | 1097 | Mean difference (IV, fixed, 95% CI) | 0.01 [−0.15, 0.18] | 0.89  | 52     |
| Healthy             | 1       | 394 | NA                                       | NA                    | NA  | NA     |
| Different intervention forms for 2 h OGTT | 5 | 1373 | Mean difference (IV, fixed, 95% CI) | 0.03 [−0.11, 0.18] | 0.65  | 0      |
| Yogurt              | 1       | 118 | NA                                       | NA                    | NA  | NA     |
| Capsule             | 5       | 1373| Mean difference (IV, fixed, 95% CI)      | 0.03 [−0.11, 0.18]   | 0.65  | 0      |
| HbA1c               | 3       | 389 | Mean difference (IV, fixed, 95% CI)      | −0.05 [−0.12, 0.03]  | 0.23  | 0      |
| Insulin level       | 16      | 1446| Mean difference (IV, random, 95% CI)     | −1.68 [−2.44, −0.92]  | <0.00001 | 76     |
| Different pregnancy status for insulin | 3 | 502 | Mean difference (IV, random, 95% CI) | 0.94 [−3.36, 5.24] | 0.67  | 90     |
| GDM                 | 9       | 605 | Mean difference (IV, random, 95% CI)     | −2.40 [−3.70, −1.09]  | 0.0003  | 70     |
| Healthy             | 4       | 339 | Mean difference (IV, random, 95% CI)     | −1.77 [−2.40, −1.14]  | <0.00001 | 24     |
| Different intervention durations for insulin | 12 | 825 | Mean difference (IV, random, 95% CI) | −2.27 [−3.13, −1.41] | <0.00001 | 72     |
| Long term           | 4       | 621 | Mean difference (IV, random, 95% CI)     | 0.42 [−2.34, 3.18]   | 0.77  | 87     |
| Different intervention forms for insulin | 1 | 70  | NA                                       | NA                    | NA  | NA     |
| Food                | 1       | 52  | NA                                       | NA                    | NA  | NA     |
| Capsule             | 14      | 1324| Mean difference (IV, random, 95% CI)     | −1.66 [−2.59, −0.73]  | 0.0005  | 76     |
| Different probiotic bacteria doses for insulin | 2 | 122 | Mean difference (IV, fixed, 95% CI) | −1.92 [−2.48, −1.36] | <0.00001 | 0      |
| Large dose          | 14      | 1423| Mean difference (IV, fixed, 95% CI)      | −1.04 [−1.22, −0.85]  | <0.00001 | 76     |
| HOMA-IR level       | 16      | 1383| Mean difference (IV, random, 95% CI)     | −0.36 [−0.53, −0.20]  | <0.00001 | 74     |
| Different pregnancy status for HOMA-IR | 3 | 502 | Mean difference (IV, random, 95% CI) | 0.07 [−0.55, 0.70] | 0.81  | 89     |
| GDM                 | 9       | 610 | Mean difference (IV, random, 95% CI)     | −0.39 [−0.88, −0.30]  | <0.00001 | 64     |
| Healthy             | 4       | 271 | Mean difference (IV, random, 95% CI)     | −0.34 [−0.48, −0.20]  | <0.00001 | 7      |
| Different intervention durations for HOMA-IR | 12 | 830 | Mean difference (IV, random, 95% CI) | −0.47 [−0.64, −0.31] | <0.00001 | 57     |
| Long term           | 4       | 553 | Mean difference (IV, random, 95% CI)     | −0.02 [−0.31, 0.47]   | 0.94  | 87     |
| Different intervention forms for HOMA-IR | 12 | 830 | Mean difference (IV, random, 95% CI) | −0.47 [−0.64, −0.31] | <0.00001 | 57     |
and intervention types of probiotics, long-term intervention (RR = 1.03; 95% CI = 0.90 to 1.18; P = 0.69) and probiotic capsule (RR = 1.05; 95% CI = 0.92 to 1.21; P = 0.44) did not decrease the incidence of GDM. For this outcome, a large dose of probiotic bacteria was used without exception.

3.6. Sensitivity Analysis and Publication Risk Bias. After the subgroup analysis, a sensitivity analysis was performed to remove several articles (Callaway et al. [33]; Pellonperä et al. [36]; Jafarnejad et al. [37]; Dolatkhah et al. [43]) that mainly contributed to substantial heterogeneity. This review largely produced similar results but better precision with less heterogeneity, and most pooled results showed relative stability (Table 3). Except for the sensitivity analyses of the FPG in the overweight/obese women subgroup and long-term intervention subgroup, the FPG and HOMA-IR in long-term intervention subgroup, all of them, had opposite results. The funnel plots of the FPG level, insulin, and HOMA-IR were also visually symmetric, which showed no significant publication risk bias. Most of the included studies focused on the top of the funnel, which further demonstrated that the sample size was large and this review is credible (Figure 3).

4. Discussion

The pooled analyses indicated that probiotic supplements had positive effects on improving FPG level, insulin level, insulin resistance, and insulin sensitivity, especially in GDM and healthy pregnant women. After

Table 2: Continued.

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Yogurt              | 1       | 70| NA                 | NA              | NA      | NA     |
| Food                | 1       | 52| NA                 | NA              | NA      | NA     |
| Capsule             | 14      | 1261| Mean difference (IV, random, 95% CI) | −0.40 [−0.63, −0.18] | 0.0004 | 77     |

Different probiotic bacteria doses for HOMA-IR

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Small dose          | 2       | 251| Mean difference (IV, random, 95% CI) | −0.29 [−0.41, −0.17] | <0.00001 | 94     |
| Large dose          | 14      | 1261| Mean difference (IV, random, 95% CI) | −0.29 [−0.34, −0.24] | <0.00001 | 77     |

HOMA-B

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| GDM                 | 2       | 130| Mean difference (IV, fixed, 95% CI) | −25.25 [−39.04, −11.47] | 0.0003  | 0      |
| Healthy             | 2       | 112| Mean difference (IV, fixed, 95% CI) | −17.74 [−32.66, −2.82] | 0.02    | 0      |

Different intervention forms for HOMA-B

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Food                | 1       | 52| NA                 | NA              | NA      | NA     |
| Capsule             | 3       | 190| Mean difference (IV, fixed, 95% CI) | −22.76 [−33.32, −12.19] | <0.00001 | 0      |

Different probiotic bacteria doses for HOMA-B

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Small dose          | 1       | 52| NA                 | NA              | NA      | NA     |
| Large dose          | 3       | 190| Mean difference (IV, fixed, 95% CI) | −22.76 [−33.32, −12.19] | <0.00001 | 0      |

QUICKI

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| GDM                 | 6       | 381| Mean difference (IV, fixed, 95% CI) | 0.00 [0.00, 0.01] | 0.03    | 49     |
| Healthy             | 3       | 201| Mean difference (IV, fixed, 95% CI) | 0.02 [0.01, 0.03] | 0.001   | 0      |

Different intervention durations for QUICKI

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Short term          | 8       | 493| Mean difference (IV, fixed, 95% CI) | 0.00 [0.00, 0.01] | 0.02    | 28     |
| Long term           | 1       | 89| NA                 | NA              | NA      | NA     |

Different intervention forms for QUICKI

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Food                | 1       | 52| NA                 | NA              | NA      | NA     |
| Capsule             | 8       | 530| Mean difference (IV, fixed, 95% CI) | 0.01 [0.00, 0.01] | 0.002   | 55     |

Different probiotic bacteria doses for QUICKI

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Small dose          | 1       | 52| NA                 | NA              | NA      | NA     |
| Large dose          | 8       | 530| Mean difference (IV, fixed, 95% CI) | 0.01 [0.00, 0.01] | 0.002   | 55     |

Incidence of GDM

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| The incident rate of GDM | 7 | 1645| Risk ratio (M-H, fixed, 95% CI) | 1.03 [0.94, 1.18] | 0.67    | 37     |

Different pregnancy status for the incident of GDM

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Overweight/obesity  | 6       | 1272| Risk ratio (M-H, fixed, 95% CI) | 1.09 [0.95, 1.25] | 0.24    | 4      |
| Healthy             | 1       | 373| NA                 | NA              | NA      | NA     |

Different intervention durations for the incident of GDM

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Short term          | 1       | 136| NA                 | NA              | NA      | NA     |
| Long term           | 6       | 1508| Risk ratio (M-H, fixed, 95% CI) | 1.03 [0.90, 1.18] | 0.69    | 48     |

Different intervention forms for the incident of GDM

| Outcome or subgroup | Studies | N | Statistical method | Effect estimate | P       | I² (%) |
|---------------------|---------|---|--------------------|-----------------|---------|--------|
| Yogurt              | 1       | 128| NA                 | NA              | NA      | NA     |
| Capsule             | 6       | 1516| Risk ratio (M-H, fixed, 95% CI) | 1.05 [0.92, 1.21] | 0.44    | 31     |

NA, not available.
Table 3: Sensitivity analysis on effects of probiotic supplement on glucose metabolism.

| Outcome | Before sensitivity analysis | After sensitivity analysis |
|---------|----------------------------|---------------------------|
|         | Effect estimate | P   | I² (%) | Remove study | Effect estimate | P   | I² (%) |
| FPG     | -0.11 [-0.17, -0.04] | 0.0007 | 72 | Callaway et al., 2019; Dolatkhah et al., 2015 | -0.10 [-0.15, -0.05] | <0.00001 | 46 |
|         | -0.09 [-0.20, 0.02] | 0.12 | 78 | Callaway et al., 2019 | -0.13 [-0.21, -0.06] | 0.0006 | 34 |
| GDM     | -0.12 [-0.24, -0.01] | 0.04 | 76 | Jafarnejad et al., 2016; Lindsay et al., 2015; Nabhani et al., 2018 | -0.22 [-0.32, -0.13] | <0.00001 | 35 |
| Short term | -0.13 [-0.22, -0.04] | 0.006 | 74 | Dolatkhah et al., 2015; Jafarnejad et al., 2016 | -0.12 [-0.20, -0.03] | 0.007 | 48 |
| Long term | -0.08 [-0.17, 0.00] | 0.06 | 72 | Callaway et al., 2019 | -0.11 [-0.18, -0.05] | 0.0005 | 35 |
| Capsule | -0.10 [-0.17, -0.03] | 0.008 | 77 | Callaway et al., 2019; Dolatkhah et al., 2015; Pellonperä et al., 2019B | -0.08 [-0.13, -0.02] | 0.008 | 38 |
| Large dose | -0.10 [-0.17, -0.03] | 0.004 | 75 | Callaway et al., 2019; Dolatkhah et al., 2015; Jafarnejad et al., 2016 | -0.12 [-0.17, -0.06] | <0.00001 | 41 |
| 2 h OGTT | 0.01 [-0.15, 0.18] | 0.89 | 52 | Asgharian et al., 2020 | 0.10 [-0.07, 0.27] | 0.26 | 0 |
| Insulin | -1.68 [-2.44, -0.92] | <0.00001 | 76 | Jafarnejad et al., 2016; Karamali et al., 2016; Pellonperä et al., 2019B | -1.52 [-2.03, -1.02] | <0.00001 | 44 |
| GDM     | -2.40 [-3.70, -1.09] | 0.0003 | 70 | Ahmadi et al., 2016; Jafarnejad et al., 2016; Karamali, 2016 | -0.99 [-1.18, -0.80] | <0.00001 | 0 |
| Short term | -2.27 [-3.13, -1.41] | <0.00001 | 72 | Ahmadi et al., 2016; Jafarnejad et al., 2016; Karamali, 2016 | -1.55 [-2.17, -0.93] | <0.00001 | 49 |
| Long term | 0.42 [-2.34, 3.18] | 0.77 | 87 | Pellonperä et al., 2019B | -1.32 [-2.02, -0.63] | 0.0002 | 0 |
| Capsule | -1.66 [-2.59, -0.73] | 0.0005 | 76 | Jafarnejad et al., 2016; Pellonperä et al., 2019B | -1.52 [-2.14, -0.91] | <0.00001 | 38 |
| Large dose | -1.04 [-1.22, -0.85] | <0.00001 | 76 | Jafarnejad et al., 2016; Pellonperä et al., 2019B | -1.04 [-1.23, -0.86] | <0.00001 | 38 |
| HOMA-IR | -0.36 [-0.53, -0.20] | <0.00001 | 74 | Pellonperä et al., 2019B; Taghizadeh et al., 2014 | -0.39 [-0.52, -0.27] | <0.00001 | 49 |
| GDM     | -0.59 [-0.88, -0.30] | <0.00001 | 64 | Dolatkhah et al., 2015; Jafarnejad et al., 2016 | -0.59 [-0.93, -0.25] | 0.0007 | 44 |
| Short term | -0.45 [-0.62, -0.29] | <0.00001 | 57 | Karamali et al., 2016 | -0.42 [-0.57, -0.28] | <0.00001 | 49 |
| Long term | -0.15 [-0.61, 0.32] | 0.53 | 85 | Pellonperä et al., 2019B | -0.27 [-0.47, -0.08] | 0.006 | 0 |
| Capsule | -0.40 [-0.63, -0.18] | 0.0004 | 77 | Jafarnejad et al., 2016; Pellonperä et al., 2019B | -0.39 [-0.54, -0.24] | <0.00001 | 37 |
subgroup analysis, the effects of the probiotic capsule were better than those of probiotic yogurt, short-term intervention (≤12 weeks) seemed to be more effective in glucose metabolism, and a large dose of probiotics (≥10⁹ CFU) played a role in decreasing FPG. However, probiotic supplements neither improved the level of HbA1c and 1 h and 2 h OGTT nor reduced the incidence of GDM.

Compared with previous systematic reviews, this present review included more RCTs and conducted several subgroups to control clinical heterogeneity, including different pregnancy status, duration of probiotic intervention, intervention types, and dosages, except for when grouping studies by probiotic species, because each combination of probiotic chains was only utilized in one or two studies. Sensitivity analyses were also performed to confirm the

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**Table 3: Continued.**

| Outcome | Before sensitivity analysis | After sensitivity analysis |
|---------|----------------------------|---------------------------|
|         | Effect estimate | P   | I² (%) | Effect estimate | P   | I² (%) |
| Different probiotic bacteria doses for HOMA-IR | | | | | | |
| Large dose | −0.29 [−0.34, −0.24] | <0.00001 | 77 | Karamali, 2016; Pellonperä et al., 2019B | −0.30 [−0.35, −0.26] | <0.00001 | 43 |
| QUIICKI | | | | | | |
| Capsule | 0.01 [0.00, 0.01] | 0.002 | 55 | Babadi et al., 2019 | 0.01 [0.01, 0.02] | <0.00001 | 0 |
| Different intervention durations for the incident of GDM | | | | | | |
| Long term | 1.05 [0.81, 1.36] | 0.69 | 52 | Wickens et al., 2017 | 1.18 [0.89, 1.57] | 0.24 | 36 |

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**Figure 3: Funnel plot.** (a) FPG. (b) HOMA-IR. (c) Insulin.
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improved credibility of the results. Furthermore, this review carefully evaluated the characteristics and qualities of each included study and concluded cautiously.

Probiotic supplements had a certain function in improving the level of FPG in this review, though it was discrepant in different pregnant women. Based on this outcome, there was no meta-analysis discussing the effects of probiotics on the improvement of blood glucose in obese pregnant women. The meta-analysis results of this study illustrated that probiotics seem to only affect blood glucose in women with GDM or healthy women. This was consistent with Barengolts et al. [52], who pointed out no evidence to prove that probiotics could help control glucose in obese participants. The probable reason for this finding could be the varied composition of the gut microbiome according to the states of gestation [53, 54]. Mokkala et al. pointed out that the inflexibility of gut microbiota may influence the probiotics to regulate glucose metabolism [55]. The gut microbiota in early pregnancy is comparable to that in nonpregnancy women. Women with GDM have lower biodiversity of the intestinal microbiota in the first trimester. The diversity of gut microbiota among prepregnancy obese pregnant women is the lowest in the early and third trimesters of pregnancy [53]. However, the result of the overweight/obese women subgroup changed obviously after sensitivity analysis. This might be contributed by one dubious study [33] that had contrary findings. Therefore, it is still unclear whether probiotics can improve blood sugar in overweight/obese pregnant women. Of particular interest is that pregnancy decreased insulin sensitivity and increased insulin, insulin resistance, and HbA1c compared with those in nonpregnancy women. Thus, the effect of probiotics could be limited among pregnant women. Additionally, the subgroup analysis of different duration of probiotic intervention found that short-term (≤12 weeks) probiotic had more positive effects on improving FPG than did long-term intervention (>12 weeks). Similarly, short-term intervention could affect FPI and HOMA-IR, which is opposite to the meta-analysis results of Han et al. [56]. With the advance of gestation, the gut microbiota associated with inflammatory states is increased substantially in healthy pregnant women. Therefore, probiotic supplements might be more helpful to control blood glucose in the short term. The available studies only performed subgroup analyses of different intervention forms in FPG. Probiotic capsules were a better choice than probiotic yogurt in the level of FPG because only a certain dose of probiotics can confer a benefit, and probiotic capsules have a more accurate dose compared with probiotic yogurt. The traditional storage method of adding probiotics to dairy products or food limits the survival of probiotics [57]. The advanced technology of making capsules ensures probiotic survival, including drying technology, microencapsulation of probiotic bacteria, improved encapsulating material, capsule size, and structure [57–59]. Intestinal flora plays an important role in energy metabolism (including diabetes and obesity) [22, 23]. There is no evidence indicating that the best dosage of probiotics can regulate blood glucose, although it is universally accepted that an adequate dose of probiotics could make a difference. The results of this meta-analysis suggested that probiotic bacteria equal to and more than 10^9 CFU per day is significantly more effective than a lower dosage. While the results in this study did not show a remarkable effect on controlling glucose, the positive performance of probiotics cannot be denied. Thus, probiotics are expected to be an assistant treatment strategy for diabetes.

The specific mechanism of probiotics remains elusive. An increasing number of researches demonstrate that microbes and metabolites work together with metabolic function and human health. Probiotics can improve metabolism by regulating the gut microbiota that will produce numerous organic compound metabolites, such as SCFAs [49] and bile acids [60]. They are the key point in the development of metabolic diseases and the improvement of insulin sensitivity, energy metabolism, and appetite suppression [21]. The imbalance of the intestinal flora may contribute to the abnormal absorption of lipopolysaccharides and increase aberrant circulating levels of SCFAs and bile acids [21, 61]. Therefore, dysbacteriosis is a vital factor of metabolic diseases like obesity, GDM, and insulin resistance [13]. Probiotics can work through the following three mechanisms: firstly, probiotics interact with gut flora and consequently produce metabolites like SCFAs; secondly, probiotics improve the intestinal epithelial barrier; and thirdly, probiotics regulate the secretion of proinflammatory mediators like tumor necrosis factor-α, interleukin-6, and intestinal glucagon-like peptide 1. The reduction of them can improve glycemic control and insulin sensitivity [19, 56, 62]. Therefore, probiotic supplements are expected to be a promising approach for glucose metabolism.

Probiotic supplements can improve insulin level, β-cell function, insulin resistance, and insulin sensitivity, especially in healthy pregnant women or those with GDM in this study. These results resembled those of several previous systematic reviews [24–29]. In particular, we found that probiotic supplements had no positive effects on insulin and HOMA-IR among overweight/obese women. Therefore, it is necessary to conduct more research to confirm this result. Meanwhile, this review did not conduct a meta-analysis of HOMA-B and QUICKI for overweight/obese women owing to a lack of related data. In addition, the probiotic capsule, which was the most applied probiotic form in this review, had effective improvement on insulin, HOMA-IR, HOMA-B, and QUICKI. Correspondingly, both small dose and large dose had positive effects. Moreover, a short-term intervention (≤12 weeks) could make more of a difference instead of a long-term intervention.

Supplementing probiotics did not have positive effects on reducing the incidence of GDM in this study, no matter the status of the pregnant women and how much probiotics they took. In the previous conclusion, the effects of improving blood glucose were mild in healthy women and barely reduced FPG among overweight/obese pregnant women. Hence, probiotics may not be an ideal preventive strategy.

This review has several limitations. Firstly, even though both English and Chinese literature were searched, only English literature was finally included. Secondly, most of the
included studies came from Iran, but some full texts from Iran could not be accessed, which might contribute to risk bias in the results and affect the generality of this study. Thirdly, there was high-level heterogeneity in the meta-analysis process which came from participants in different pregnancy status as well as the difference in duration, forms, species, and doses of the probiotic interventions. Meanwhile, given the limitations of the original article, even though participants were divided into overweight/obese women, women with GDM, and healthy pregnant women, there were still women who were concurrent with overweight/obese and GDM, which contributed to some heterogeneity for results. Moreover, the beginning and termination of intervention of the included studies differed, which might, to a certain extent, affect the results. Fourthly, outcome measurements only included glucose metabolism, insulin sensitivity, and the incidence of GDM; other maternal and adverse outcomes were not included in this review, which might lead to a limitation. Finally, while this current meta-analysis was not registered and may have small deviations, we still strictly followed the process of the systematic review.

In conclusion, probiotics could modulate blood sugar and improve insulin level within a certain range in healthy and GDM women instead of overweight/obese pregnant women. There were no related meta-analyses discussing the effects of probiotics among obese/overweight pregnant women. The specific mechanism that causes this difference among pregnant women in normal and pathological pregnancies remains unknown. One of the possibilities may be that the richness, diversity, and sensitivity of intestinal flora interfere with the role of probiotics, thereby calling for more high-quality trials with a larger number of participants to explore the effects of probiotics in obese/overweight pregnant women. A significant reduction was likewise observed for metabolism parameters in taking short-term probiotics (≤12 weeks). A dosage of $10^9$ CFU per day or more could be considered an effective dose to modulate FPG. It is suggested that the regulatory effects of probiotics are short-lived. Furthermore, probiotic capsules might be more effective than probiotic yogurt in terms of FPG level. However, probiotic supplements neither improve the levels of HbA1c and 1 h and 2 h OGTT nor reduced the incidence of GDM. The optimal probiotic dose in this study is also still unclear. Further high-quality, multicentre, and large-scale RCTs are needed to ensure the safety of probiotics, probe into the positive effects on pregnant women and infants, and confirm the dosage and duration of probiotic supplements. Additional research can be conducted to further determine the effects of probiotics on insulin resistance and insulin sensitivity among overweight/obese women.

### Abbreviations

- **GDM**: Gestational diabetes mellitus
- **RCT**: Randomized controlled trial
- **OGTT**: 75 g oral glucose tolerance test
- **CI**: Confidence interval
- **MD**: Mean difference

### Data Availability

All the data of the review are available from public electronic databases.

### Disclosure

The funder had no role in study design, data collection, and analysis, decision to publish, or preparation of the manuscript. Yu-Qing Pan and Qing-Xiang Zheng are the co-first authors.

### Conflicts of Interest

The authors declare no conflicts of interest.

### Authors’ Contributions

Yu-Qing Pan was responsible for manuscript drafting and revision, data collection, data extraction, and data analysis; Qing-Xiang Zheng contributed to study design, manuscript drafting and revision, data analysis, and data interpretation; Xiu-Min Jiang was responsible for study design, data interpretation, and manuscript revision; Xiao-Qian Chen, Jing-Ling Wu, and Xiao-Yun Zhang contributed to data analysis and data interpretation.

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### Supplementary Materials

Supplementary file 1: search strategy for each database used in this review. Supplementary file 2: PRISMA 2009 checklist. (Supplementary Materials)

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