Research Paper

Anti-Gastrins Antiserum Combined with Lowered Dosage Cytotoxic Drugs to Inhibit the Growth of Human Gastric Cancer SGC7901 Cells in Nude Mice

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Abstract

The objective of this study was to determine the effect of anti-gastrin antiserum in combination with varied dosages of cytotoxic drugs (5-Fluorouracil (5FU) + Cisplatin (CDDP)) in vivo growth of the human gastric cancer cell-line, SGC-7901, which expressed cholecystokininB/gastrin receptors and secreted gastrin. The anti-gastrin antiserum was obtained by immunizing rabbits using a novel immunogen vaccine, which was composed of the common amino-terminal portion of human carboxy-amidated gastrin-17 (G17) and glycine-extended gastrin-17 (gly-G17) and the common carboxy-terminal portion of progastrin (in a 50:50 mixture) all covalently linked to tetanus toxoid (TT) by specific peptide spacers. The antiserum neutralized both G17 and gly-G17 by enzyme-linked immunosorbent assay (ELISA), and a synthetic progastrin peptide, as well, using an E. coli expressed his-tagged progastrin.

The tumor was implanted subcutaneously into the backside of BALB/c nude mice, and the combination antibody-drug treatment using low dose combination chemotherapy had significantly reduced median tumor volumes (62% reduction; p =0.0018) and tumor weights (53% reduction; p =0.0062) when compared to the conventional high dose chemotherapy treated control mice that had a corresponding similar reductive effect, using just the two standard cytotoxic drugs alone; namely by reducing the tumor volumes (65%; p =0.0016) and tumor weights (59% reduction; p=0.0033). Importantly, the immunological treatment had little of the toxicities and side-effects of the full chemotherapy doses alone, which was effected by using a significant decrease in the dosage of chemotherapeutic drugs, while maintaining the same level of efficacy at reduction of tumor growth.

Key words: Gastrins; Progastrin; Cancer; Cytotoxic Drugs; Antibody.

Introduction

Gastrins are peptide hormones secreted by G cells in antrum of stomach and mucosal membrane. Gastrin’s post-translational processing is from a gastrin precursor, preprogastrin, which gives rise to a variety of products including progastrin (pro-G), glycine-extended gastrin-34 (gly-G34), amidated gastrin-34 (G34), glycine-extended gastrin-17 (gly-G17) and amidated gastrin-17 (G17), each with a distinctive spectrum of biological activity. Progastrin itself stimulates colonic epithelial proliferation, and their bio-processed intermediates (Gly-extended progastrins and gastrins) stimulate colonic epithelial proliferation.
and gastric epithelial differentiation, as well as does the C-terminally amidated gastrins (collectively, “the gastrins”); all of which are able to stimulate colonic proliferation, gastric epithelial proliferation and differentiation, and acid secretion [1].

Recently, numerous researchers have shown that the gastrins, specially including gly-G17 [2] and G17 [3], and progastrin all could promote tumorigenesis (e.g. gastric [4], colon [5], pancreatic [6] and non-small cell lung cancers [7]) in an autocrine, paracrine or endocrine manner [8-10], which has also warranted the gastrins as a preferred anti-tumor target in cancers expressing these stimulatory factors.

Gastric cancer is one of the most common malignant tumors in the world and second most frequent cause of cancer deaths, however more than 75% of patients present with gastric carcinomas that belong to advanced stages, and lose the opportunity for curative surgery; leaving primarily chemotherapy as a major and important role in its treatment. At present, 5-Fluorouracil (5-FU) combined with cisplatin (CDDP) is a standard combined treatment in gastric carcinoma chemotherapy, however at this stage of disease its’ efficiency and median survival time (MST) are just about 20%, and 7 months respectively [11,12]. One of main reasons leading to a low efficiency is that the conventional drug combinations are highly cytotoxic, which often result in off-targeting-toxic effects (neutropenia, anemia, etc..) and may cause serious adverse reactions and unwanted side-effects (e.g. high grade emesis, emaciation and hypoinnunity). This is especially true for patients that have their tumors becoming increasingly resistant to the drugs and by simply increasing the dosage of chemotherapeutic drugs their adverse symptoms and quality of life, become increasingly more severe. Thus the need for novel combination treatments in antineoplastic therapy that would promote efficacious tumor reductions or even increased curative effects, and at the same time, also reduce or avoid serious adverse reactions, has got the attention of researchers and clinicians treating advanced gastric cancers.

With reference to the proliferative mechanism of the gastrins in promoting gastrin-stimulated tumors, this study elucidates the deployment of a novel vaccine combination, that is composed of the common COOH-terminal amino acid sequences of progastrin, and the NH2-terminal amino acids of G17 and gly-G17 all covalently linked to Tetanus toxoid (TT) via peptide spacers, which induce the production of a spectrum of progastrin and gastrin neutralizing (anti-gastrins) antibodies, both the amidated and glycine extended forms, and thus reduces the availability of these growth factors to stimulated tumor proliferation. In addition we have also developed a combination treatment, which has an additive action by using the anti-gastrins antiserum and a reduced amount of chemical drugs (5-FU + CDDP) in an anti-tumor protocol. Our research contents thus includes: a) the use of anti-gastrins vaccine that raises antibodies neutralizing G17, gly-G17, and progastrin, all growth proliferative species and b) the anti-gastrins antiserum combined with reduced cytotoxic drug combinations, (5-FU + CDDP) to inhibit the human gastric cancer cells, SGC7901 xenografted into nude mice.

Materials and Methods

Cell line

SGC7901 is a human gastrin carcinoma cell-line. The cell-line was maintained in vitro in DMEM (GIBCO, US) supplemented with 5% heat-inactivated fetal bovine serum (FBS) (GIBCO, US), 1% penicillin-streptomycin and grown at 37°C in humidified conditions gassed with 5% CO2.

Immunogen

The immunogen is composed of an equal mixture of the common carboxy-terminal progastrin amino acids and NH2-terminal amino acids of G17 and gly-G17 all covalently linked to Tetanus Toxoid (TT) by peptide spacers.

Immunization procedure

The vaccine (0.25 mg/kg) was injected i.m. into left or right leg of rabbits (n = 6). Rabbits were initially immunized using three injection at 2-week intervals. Control animals were normal rabbits that were used as a source for IgG sera.

Anti-human G17 gastrins antibody levels of vaccine-immunized rabbits [13, 14]

Rabbits were ear bl ed at time points throughout the experiment and at termination by cardiac puncture euthanization under terminal anaesthesia. Anti-human G17 antibody levels were determined by ELISA. 110 µl per well of a 1 µg/ml solution of human G17-bovine serum albumin (BSA) conjugate (AoKe Corporation) in a coating buffer (1.5 mg/ml Na2CO3, 3 mg/ml NaHCO3, pH = 9.6) was coated into 96-well Immunulon U plates (Corning, USA) by an overnight incubation at 4°C. The positive, negative and tested sera at 3.16-fold serial dilutions, starting at a dilution of 1:100, were prepared in antibody dilution buffer [phosphate buffer saline tween-20 (PBST), 1% BSA]. Subsequent steps used the PBST (8 mg/ml NaCl, 3 mg/ml Na2HPO4·12H2O, 2.5 mg/ml KCl, 0.2 mg/ml KH2PO4, 0.05% tween-20) without BSA was used for washings. The 96-well plates were washed 4X to free of non-bound conjugates, then the sera were added (100 µl/well). After 1.5 hour incubation at room
temperature (RT), the plates were washed four times and a goat anti-rabbit IgG (H + L) alkaline phosphatase conjugate was added (1:1000 dilution in antibody dilution buffer, 100 µl/well). After 1.5 hour incubation at RT, the plates were washed four times to remove nonbound reagent, and 100 µl/well of pNPP substrate solution (1 mg/ml) was added in substrate buffer (0.01 mg/ml MgCl₂·H₂O, 10% diethanolamine). After 5 min incubation in dark at RT, 100 µl/well of stop buffer (1.0 M NaOH) was added and absorbance was measured on a microplate reader 405nm (reading wave)/490nm (reference wave). The values of 490 nm were subtracted from that of 405nm, and the antibody titer was calculated by using ED₅₀ (50% effective dose) module of SCANLT software.

**Specificity of antibodies raised in rabbits against anti-gastrin vaccine [13,15]**

A competitive ELISA was used to assess the specificity of the affinity of antibodies for G17 peptide. A fixed concentration of antiserum (1:50 dilution) was combined with the same volume of various inhibitors at 10-fold serial dilutions and incubated for 1 hour at room temperature. Then taking the mixture as primary antibody, other procedures were the same as the ELISA in detecting antihuman G17 antibody levels of vaccine-immunized rabbits. The inhibitors included *E. coli* expressed his-tagged Progastrin, G17, gly-G17, G34, vasoactive intestinal peptide (VIP), TT, and buffer (no inhibitor). Samples were run in quadruplicate, and means were calculated for each concentration. The % Inhibition relative to no inhibitor added (antiserum + buffer) was calculated for each inhibitor tested: %Inhibition = 100%(A uninhibited - A inhibited) / A uninhibited, where A = Absorbance.

**Establishment of carcinoma models**

Semi-confluent cell monolayers were harvested with 0.25% trypsin-EDTA. Confluent cell monolayers were washed twice and suspened in sterile PBS at a concentration of 1.0×10⁷ cells/ml in an ice-bath. A 0.45 ml volume was then injected into female nude mice (6-8 weeks old, weighting 17-20 g) subcutaneously on the back. The tumor pieces (3-5 mm³ in saline) were implanted into animals subcutaneously on the back. The mice were maintained in sterile isolation, fed and watered *ad libitum*. When tumors grew to 100 mm³, the mice were grouped to distribute and equalize tumor volumes.

**Treatments**

After being distributed into seven groups: a 5-FU at 50 mg/kg was combined with CDDP at 5 mg/kg as standard high-dose chemotherapeutic combination group (H.C.) that was administered i.p. every 10 days, and a normal rabbit (gastrin-negative IgG containing) serum at 200 mg/kg as a negative control (N.C.), which was administered i.p. every 2 days. Other groups consisted of: the anti-gastrins antiserum at doses of 200mg/kg, 150mg/kg, and 100 mg/kg (indicated as combined high dose (H); combined medium dose (M), and combined low dose (L) groups respectively). Each of the groups were administered respective sera, i.p. every 2 days that was all combined with 5-FU at the 40% lower chemotherapeutic dosages of 20 mg/kg and CDDP at 2 mg/kg administered i.p. every 7 days. The anti-gastrins antiserum alone was at 200 mg/kg given as a singular antiserum group that was administered i.p. every 2 days, while chemotherapy group of 5-FU at 20 mg/kg combined with CDDP at 2 mg/kg was given as a singular chemical combination for this group, and was administered i.p. every 7 days. Each experimental group was composed of 8 animals.

Animals were inspected daily and weighed every 2 days. Tumor volumes were measured twice weekly in a blind manner by an independent observer. The measurement was performed 3 times daily; measurements were taken and the mean of the 3 measurements at each time point was recorded. The mice were treated for a period of 26 days, sacrificed and the tumors removed for study. The weight, volume of tumors and weight of body were also carried out by an independent observer who was blind to the treatment groups. Means and standard deviation of the means were calculated for each group (means ± SD).

**Histological evaluation of the SGC-7901 tumors**

Tumor specimens were 4% formalin-fixed. Sections were then embedded in paraffin, 4-5 µm sections were cut by a microtome and stained with haematoxylin and eosin (HE). Assessment of tumour necrosis area was made by a histopathologist in a blind manner by image analysis.

**Statistical analysis**

All experiments were repeated three times. The data were analyzed between groups using Student’s t test. p values < 0.05 were considered statistically significant.

**Results**

**Human Anti-Gastrin17 Antibody Levels of Vaccine-immunized Rabbits**

Six rabbits were immunized with the immunogen vaccine and their antibody titers followed for a period of 8 weeks following three immunizations. Fig. 1 shows the antibody levels against G17 as assessed by the described ELISA technique. Following three
immunizations, antibodies were detectable by week 4 in all rabbits. The initial surge of antibodies by week 4 was followed by a second surge by week 6 (p < 0.05 vs week 4), after which the antibody titers declined and approached the level of week 4 by week 8 (p > 0.05 vs week 4) (Fig. 1).

Figure 1. Time scale of antibody titers of rabbits immunized by vaccine (0.25mg/kg) at the time points shown. Each point represents mean antibody titers (n = 6). Antibody titers were measured by an ELISA assay using a 1:100 dilution of antisera. Immunizations are indicated by arrows. *p < 0.05 vs 4 weeks, **p > 0.05 vs 4 weeks.

Normal rabbits were selected as negative control in the study, of which the protein concentration was equal to that of antiserum, but the G17 antibody titers was undetectable.

During the period of immunization, there was no local or systemic reactions induced (e.g. swelling, or festering) in the injection site of rabbits. The weight, which significantly increased (p < 0.05, data not shown) at termination, their appetites and other clinical features of the rabbits also showed no abnormalities. Besides using the dose of 0.25 mg/kg, another two doses including 0.125 mg/kg and 0.5 mg/kg immunogen have been studied. However the antibody titers for the 0.125 mg/kg dose was deemed too low, whereas the dose of 0.5 mg/kg had some serious local tissue reactive responses, and was therefore not used, even though there were high levels of antibody (data not shown).

The antiserum used to inhibit human gastric cancer SGC7901 cell growth in nude mice was prepared from the blood collected at week 6 by the optimal immunization program adopted in this study. Animals received three immunization injections initially and, a booster immunization would be administered if anti-gastrin antisera levels declined by more than 60% over the experimental times.

Gastrin specificity of antiserum raised by vaccine immunization of rabbits

Fig. 2 presents the percent inhibition of antibodies binding to G17-BSA as a function of inhibitor concentration. As can be seen in the figure, G17 and gly-G17 significantly inhibited antibodies binding to G17-BSA with their concentrations increasing (p < 0.05). Approximately 80% inhibition was attained with the G17 and gly-G17, however no inhibition was obtained by using TT, G34 and VIP. The NH2-terminals of G17 and gly-G17 was the same with the shared peptides epitope of immunogen vaccine, and so the results suggest that the affinity of antibody in antiserum was specific for binding the G17 and gly-extended G17 peptides.

Inhibition of antibodies binding to progastrin C-terminal peptide-bound to BSA was also effected and was a function of inhibitor concentration, using progastrin-specific peptides and trypsin digests of his-tagged (Ni-NTA column purified) expressed progastrin (data not shown).

TT was the carrier protein of vaccine, which could be recognized by antigen presenting cells and presented to CD4+ Th cells. Th cells then provided the second signal needed in B cells activation, which could exert immunological enhancement and promote production of a high levels of neutralizing antibodies. The results indicated that TT also could did not interfere in the role of binding of gastrins to immunogen-induced antibodies.

G34 and VIP were two types of gastrointestinal hormones, which were neuropeptides distributed at low levels, but widely distributed throughout the gastrointestinal tract, and central and peripheral nervous systems, having various biological activities, and were important in maintaining the normal physiological status and function of body. G34 could promote acid secretion and gastrointestinal mucous membrane proliferation, VIP could regulate blood pressure, boost myocardial contractility and dialate vessels. Anycross reactivity between those hormones and the immunogen induced antibodies could pre-
dictively produce untoward effects of the immunogen induced antibodies; however the results indicate that there were no non-specific interferences occurring between them.

**In vivo analysis of the therapeutic effect of anti-gastrins antiserum in combination with low dosage chemical drugs (5-FU + CDDP) on human SGC-7901 gastric tumors**

Fig. 3A, 3B and 3C show the effect of anti-gastrins antiserum in combination with cytotoxic drugs on the growth curve, final volume and weight of SGC-7901 tumors respectively. As shown in Fig. 3A, the tumor growth trend of the standard high-dose chemotherapeutic combination (H.C.) corresponded to that of combined high dose (H) antibody group with the lower levels of 5FU + CDDP. Similarly median volumes (Fig. 3B) of tumors from standard high-dose chemotherapeutic combination (H.C.) and the combined high dose antibody and lower-dose chemo (H) groups, had no significant difference (p = 0.8089) with one another; each were reduced by 65% (p = 0.0016), and 62% (p = 0.0017) respectively when compared to antibody negative controls (N.C.). The volumes of combined medium dose (M), combined low dose (L), single antibody (S.A.) and low dosage single chemical (S.C.) groups were not significantly different from that of negative controls (N.C.) group (p = 0.1391, 0.1008, 0.3010, 0.0894, respectively).

Median weights (Fig. 3C) of tumors from standard high-dose chemotherapeutic combination treated (H.C.) and combined high dose antibody (H) groups, showed no significant differences (p = 0.6859) between each other, and were reduced by 59% (p = 0.0033), and 53% (p = 0.0062) respectively when compared to negative controls (N.C.). The weights of the combined medium (M), combined low dose (L), single antibody (S.A.) and sole chemotherapies (S.C.) groups were also not significantly different from that of negative control (N.C.) group (p = 0.2324, 0.2642, 0.3114, 0.1270, respectively).

Table 1 shows the %RTP of H.C. and combined H groups are 22% (p = 0.0001), 28% (p = 0.0018) respectively, the %TWI of H.C. and combined H groups are 59% (p = 0.0033), 53% (p = 0.0062) respectively. The results indicated that treatment with a combination of anti-gastrins antiserum and lowered doses of cytotoxic drugs could achieve the optimal therapeutic effect as that of using a high dose of cytotoxic drugs with respect to inhibiting the growth of the human SGC-7901 gastric cancer tumors.

The antiserum and chemical drugs used in combined H group were separated to form the S.A. and S.C. groups respectively to study the interrelation between them after the combined using. As shown in RTV, S.C., whose dose was equal to the 40% dose of H.C., had weak anti-tumor effect, S.A. had no anti-tumor effect, however combining application of them to form the combined H group could produce synergy anti-tumor effect. The results suggested that
the anti-gastrins sera could increase the sensitivity of the tumors to chemotherapeutic drugs. We inferred that besides promoting tumor proliferation, invasion, metastasis and anti-apoptosis, the gastrins as growth factors might also enhance the multi-drug resistance (MDR) of tumor, but whose mechanisms needed to be studied further in detail.

Table 1. The relative tumor proliferation and tumor weight inhibition of treatment groups

| Groups          | Relative tumor volumes (Mean ± SD) | Relative tumor proliferation (%) | Tumor weight inhibition (%) |
|-----------------|------------------------------------|---------------------------------|-----------------------------|
| N.C.            | 22.29 ± 10.85                      | /                               | /                           |
| H.C.            | 4.30 ± 3.49**                      | < 40                            | > 30                        |
| Combined H      | 6.16 ± 2.65**                      | > 40                            | > 30                        |
| Combined M      | 14.73 ± 5.25*                      | 66                              | 16                          |
| Combined L      | 13.24 ± 4.49*                      | 59                              | 13                          |
| S.A.            | 15.78 ± 8.79                       | 71                              | 14                          |
| S.C.            | 12.72 ± 4.89*                      | 57                              | 23                          |

The %Relative tumor proliferation (RTP) relative to negative control was calculated for each treatment: %RTP = 100% × (TreatmentRTV / Negative ControlRTV), which should be less than 40% (p < 0.05, Student’s t test) to define the therapeutic effect. RTV (relative tumor volume) = tumor volumes at termination / tumor volumes in group. The %Tumor weight inhibition (TWI) relative to negative control was calculated for each treatment: %TWI = 100% × (1 - Median weightT / Median weightN.C.), which should be greater than 30% (p < 0.05, Student’s t test) to define the therapeutic effect. T = treatment. *p < 0.05 vs N.C., **p < 0.01 vs N.C.

Fig. 4A and Fig. 4B show the body weight changes and the final median body weight of all groups during the experiment, respectively. Over the duration of the experimental period, the body weights of standard high-dose chemotherapeutic combination (H.C.) had a significant decline (p < 0.05) compared to that of the other groups. The standard high-dose chemotherapeutic combination group animals had body weights significantly reduced by 20% (p = 0.0006) compared to those in other groups; moreover, animals in the standard high-dose chemotherapeutic combination group (H.C.) also exhibited diarrhea after each high dose administration. Further the weights of other groups showed no significant differences among themselves, unlike the high dosage positive controls (p > 0.05). Additionally all animals given low dosage chemotherapeutics, or just antiserum alone, showed no abnormal clinical reactions, unlike standard high-dose chemotherapeutic combination given the higher dosages of chemotherapeutic drugs.

The weights of standard high-dose chemotherapeutic combination treated animals (H.C.) were reduced by 17% (p = 0.0035) compared to the weights of the negative controls (N.C.) at termination. The other groups had no significant difference when compared to the negative controls (p > 0.05 vs N.C.). These findings suggest that the standard (higher) doses of the chemotherapeutic drugs used in standard high-dose chemotherapeutic combination (H.C.) have significantly stronger toxicities and side effects; whereas the antiserum had no such effects.

Histological evaluation of SGC7901 tumors

Table 2 shows the mean scores of N.C., H.C. and combined H groups graded for necrosis; samples were 0.875, 2.625, 2.25 respectively, which indicate the extent of pathology and degree of necrosis. The degree of standard high-dose chemotherapeutic combination (H.C.) necrosis corresponded to that of the combined high dose (H) group and also revealed that the tumors from animals treated with high chemotherapeutics (H.C. and combined H) groups sustained a greater degree of necrosis (Fig. 5B, Fig. 5C) when compared to tumors from negative controls (N.C.) (Fig. 5A).

Table 2. Pathology necrosis graded scores stained with HE

| Groups          | Mean scores |
|-----------------|-------------|
| Numbers with different levels of necrosis | 0 points (< 25%) | 1 points (25%–50%) | 2 points (50%–75%) | 3 points (> 75%) |
| N.C.            | /           | /                 | 2                | 5               | 1               | /                | 0.875             |
| H.C.            | /           | /                 | 3                | 5               |                 | 2.625             |
| Combined H      | /           | 2                 | 2                | 4               |                 | 2.25              |
Figure 5. Histology of SGC-7901 tumor from SCID mice. A, A section from tumor-cells of animals treated with negative control sera (N.C.) ×40. B, A section from tumor-cells of animals treated with standard high-dose chemotherapeutic combination protocols (H.C.) ×40. C, A section from tumor-cells of animals treated with combined-high dosage (H) group protocols ×40. All tumors were formalin fixed, paraffin embedded, and stained with hematoxylin and eosin. All assessments were made "blind" by an independent pathologist.

Discussion

The action mechanism of gastrins conforms to the signal-receptor theory that gastrins as ligands combine with their cognate receptors to stimulate relevant signaling pathways and thus exert pro-proliferative biological effects. The gastrins mainly stimulate cholecystokinin (CCK) family receptors (e.g. CCK-A, CCK-B or “classical” receptors) [16]; however there are also a number of potential “non-classical” receptors that have been reported to bind gastrins, especially the pro-gastrin species [17]. Many signal pathways (e.g. PKC [18], ERK/MAPK [19], PI3K/Ak [20], P125-FAK [21], JAK/STAT [22] signals) can be activated through the “classical” CCK receptors by the gastrins, which involve many of the known signal transduction pathways and share extensive signaling cross relationships [23]. Thus there is a strong body of information showing that the gastrins are able to produce many pro-malignant biological effects including promoting tumor proliferation [24], invasion [25], metastasis [26] and anti-apoptosis [27].

Currently there are mainly three kinds of methods aimed at targeting, primarily, gastrin 17 to effect an anti-tumor response, by inhibiting the secretion of this gastrin hormone (e.g. somatostatin [28], iRNA [29]), receptor antagonists (e.g. L-364718, L-365260, YM-022) [30] and by immunization to neutralize gastrin 17 (e.g. monoclonal antibody, vaccines [31]). A number of high affinity CCK-A and CCK-B receptor antagonists [32,33] have been exploited to inhibit the proliferative effects of gastrin-17, however they lacked specificity as they also blocked the actions of all the potential ligands of these receptors, such as G34 and CCK, meanwhile the receptors in tumors also were seen to undergo mutations and thus became insensitive to this receptor approach and therefore they could not block the potential proliferative activity of the hormone and its precursors that were being bound by non-classical receptor proteins that were also involved in tumor growth [30]. The use of somatostatin also lacked specificity and its use might upset the normal role of other hormones in the body. Monoclonal antibodies against receptors might have had positive effects, but not on mutated receptors, and long-term usage introduces immunogenicity and infusion-related reactions and is extremely expensive to be widely used in so frequent a cancer as gastric cancer is in Asia. The immunogen-vaccine designed to neutralize gastrins would overcome many of these shortcomings both in theory and in practice, which has become the research focus of this study.

This novel immunogen-vaccine in the presented study, a conjugated Gastrins-TT, consists of three parts, and includes the common amino-terminal por-
The proliferation of GI cancers; and so by inhibiting the S-phase of the cell cycle, but could not demonstrate the precise roles of various gastrins.

Significant inhibition of *in vitro* growth was not shown with the anti-gastrin antisera alone (data not shown), in these cells, which might be possibly due to the proliferation effect of the nutritional content that is associated with the crude antisera, and would necessitate the use of affinity purified anti-sera to perhaps demonstrate the inhibiting effect of higher amounts of anti-human gastrin antibodies.

In conclusion, this work indicated that the beneficial therapeutic effect of combined low dosage cytotoxic chemotherapy with anti-human gastrin antisera, which was as effective at inhibiting the gastric cancer xenograft tumor proliferation of higher dosage chemotherapy, but with lessened toxic side-effects. Such studies would warrant further combination studies with lower dose chemotherapeutics, along with anti-gastrin sera on gastrin responsive GI tumors, and might signify new clinically important protocols for their combined use in the future treatments of gastric and other GI cancers.

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**Competing Interests**

The authors have declared that no competing interest exists.

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