Environmental factors controlling the distributions of *Botryococcus braunii* (A, B and L) biomarkers in a subtropical freshwater wetland

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Here we report the molecular biomarker co-occurrence of three different races of *Botryococcus braunii* (*B. braunii*) in the freshwater wetland ecosystem of the Florida Everglades, USA. The specific biomarkers include C32–C34 botryococenes for race B, C27–C32 n-alkadienes and n-alkatrienes for race A, and lycopadiene for race L. The n-alkadienes and n-alkatrienes were present up to 3.1 and 69.5 µg/g dry weight (dw), while lycopadiene was detected in lower amounts up to 3.0 and 1.5 µg/g dw in periphyton and floc samples, respectively. Nutrient concentrations (P and N) did not significantly correlate with the abundances of these compounds. In contrast, n-alkadienes and n-alkatrienes were present in wider diversity and higher abundance in the floc from slough (deeper water and longer hydroperiod) than ridge (shallow water and shorter hydroperiod) locations. n-Alkadienes, n-alkatrienes, and lycopadiene, showed lower δ13C values from −40.0 to −35.5‰, suggesting that the source organisms *B. braunii* at least partially utilize recycled CO2 (13C depleted) produced from OM respiration rather than atmospheric CO2 (13C enriched) as the major carbon sources.

The green alga *B. braunii* is widely distributed in aquatic ecosystems, especially lakes and ponds1. The Botryococci are known to contain high amounts of a remarkably diverse range of unusual hydrocarbons, such as botryococenes, n-alkadienes and n-alkatrienes, C40 monoaromatic compounds including lycopadienes and related oxygenated compounds that provide source diagnostic information2,3. While the C23–C31 n-alkadienes and n-alkatrienes, and squalenes (less specific) were reported as indicators of race A of *B. braunii*5–7, botryococcene (C30–C37) related lipids and methylated squalenes (C31–C34) were believed to be specific biomarkers biosynthesized by race B of *B. braunii*2,4,5. In contrast, race L contains isoprenoid structures related to lycopadiene8–10. These biomarker compounds, especially the saturated forms of botryococenes and lycopadienes well preserved in sediments and rocks, were thus used as biomarkers for paleoreconstructions5,9.

The Florida Everglades is the largest, subtropical freshwater wetland in the United States. Since the early 20th century it has been drained significantly because of structural modifications for flood control, urban and agricultural development, which severely reduced its size, and over 5,000 km2 (50%) of the original landscape has been converted to agricultural and urban use during the last half century. Drainage of the wetlands resulted in shifts in the composition and distribution of vegetation cover, changes of the water quality and hydroperiod11. Currently, the vegetation shifts along the Everglades landscape from sloughs (deeper water, longer hydroperiod) with emergent and submerged plants, to ridges (shallow water, shorter hydroperiod) with *Cladium* sp. dominated communities, and scattered tree islands dispersed throughout this landscape12. Within this diverse distribution of plant species, periphyton mats, composed of abundant calcareous mixtures of diatoms, cyanobacteria and green algae, are distributed widely throughout this ecosystem13,14. In the Everglades, periphyton occurs primarily as benthic or floating mats instead of free floating phytoplankton. Thus, the suspended particulate organic matter is

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mostly found as flocculent material (floc), which consists of a non-consolidated layer of microorganisms, organic (detritus and disaggregated periphyton remains) and inorganic particles15. Although

_B. braunii_ is distributed widely in aquatic ecosystems, especially in tropical oligotrophic freshwater to brackish lakes 16, the microfossils of _Botryococcus_ have only been reported in soil cores of tree islands in the Everglades17. A series of botryococcenes with carbon numbers from C32 to C34 were also detected in periphyton, floc, surface and deeper soils across the Everglades wetlands 18, suggesting the existence of race B of _B. braunii_.

Although individual races of _B. braunii_ are widely distributed, reports of the co-existence of the different chemical races are rare. To our best knowledge, there is only one prior report of the co-existence of three races in a freshwater crater lake1. Here, we report the molecular characterization of various tracers of three races of _B. braunii_ including botryococcenes, long chain _n_-alkadienes, _n_-alkatrienes, and lycopadiene in environmental samples of the Everglades ecosystem, examine their stable carbon isotopes, and discuss possible controlling factors including nutrients and hydroperiod on their distribution and abundances.

### Experimental Methods

#### Sampling locations.

The sampling sites for this study feature a gradient of nutrient and hydroperiod in the Everglades (Fig. 1; Table 1; http://fcelter.fiu.edu/data)19. Briefly, Water Conservation Area 3 (WCA3) is located...
## Compounds Kovats retention indexes* Periphyton Floc

| Compounds | Kovats retention indexes* | Periphyton | Floc |
|-----------|---------------------------|------------|------|
| C_{27:3}  | 2630                      | —          | —    |
| C_{27:2}  | 2643                      | —          | —    |
| C_{26:3}  | 2727                      | —          | —    |
| C_{26:2}  | 2744                      | —          | —    |
| C_{25:3}  | 2826                      | 7981 (2457)| —    |
| C_{21:2}  | 2843                      | 1957 (974) | —    |
| C_{20:3}  | 2923, 2927                | —          | —    |
| C_{20:2}  | 2941, 2965                | —          | —    |
| C_{19:3}  | 3022                      | —          | —    |
| C_{18:2}  | 3040                      | 4521 (721)| 201 |
| C_{18:1}  | 3070, 3110                | —          | —    |

### Sample collection and preparation.

Periphyton and floc (regardless of ridge and slough environments) were collected from various locations across the Florida Everglades (Table 2). Additional floc samples were sampled from both ridge and slough environments, and during different times of the year from 2012 to 2014 within WCA3. Both submerged and floating periphyton were placed into Ziploc bags. Floc, surface soils were sampled following the procedures as described previously. Both leaves and roots (when applicable) of the dominant plants such as Nymphaeaceae, Utricularia sp., Chara sp., Cladium sp., Eleocharis sp., Typha domingensis P., and Typha latifolia were randomly sampled from different locations within ENP and WCA3. All samples were kept cool on ice during transport to the laboratory. The samples were transferred into pre-combusted glass jars and stored at −20 °C until further analysis.

All samples were processed and analyzed as described previously. Briefly, they were freeze-dried at −50 °C, then shredded and sieved through a 32 mesh (500 µm) sieve to remove coarse material. Samples (1–3 g dry weight) were subjected to ultrasonic extraction three times (0.5 h each) with dichloromethane (DCM) (Optima, Fisher, USA) as solvent. Total extracts were concentrated and then fractionated by adsorption chromatography over silica gel. The aliphatic fraction and aromatic hydrocarbon fraction were eluted sequentially using n-hexane and hexane: toluene (3:1, v:v), respectively. Ziploc bags used for sampling were washed with n-hexane and the extracts were employed as control blanks and randomly analyzed to exclude external contamination.

### Bulk parameter analysis.

Total nitrogen (TN) was measured by the high-temperature dry combustion method using a Carlo-Erba NA-1500 CNS Analyzer. Total P was analyzed with a Technicon Auto Analyzer II System (Pulse Instruments Ltd.), according to the standard method for orthophosphate P (EPA method 365.1). Bulk δ^{13}C values were also determined for floc samples using standard elemental analyzer isotope ratio mass spectrometer (EA-IRMS) procedures, and reported with respect to the Vienna PeeDee Belemnite (VPDB) standard for carbon. Precision of the δ^{13}C measurements was ±0.10‰.

### Gas chromatography-mass spectrometry (GC-MS).

GC-MS analyses were performed on a Hewlett-Packard 6890 GC fitted with a Rtx-1 capillary column (30 m, 0.25 mm ID, Restek, USA) interfaced to a HP 5973 MSD. Compounds were quantified by squalane as the internal standard, assuming a similar response factor. Kovats retention indexes (RI) were calculated following the formula: RI = 100 × (R_r−R_p)/(R_p−R_0) + 100n, where x denotes the compound of interest, R denotes the GC retention time, and n and n + 1 denote the carbon number for the nearest n-alkane and (n + 1)-alkane eluting before and after x, respectively on the GC. The identification of individual compounds was based on the comparison with published mass spectra and interpretation of the mass spectra.

### Gas chromatography-isotope ratio mass spectrometry (GC-IRMS).

The δ^{13}C values of individual n-alkadienes, n-alkatrienes and lycopadiene were measured using a GC-IRMS system, which consists of a HP 6890 GC equipped with a Rtx-1 fused silica capillary column (30 m, 0.25 mm ID), a combustion interface (Finnigan GC combustion IV), and a Finnigan MAT delta Plus mass spectrometer. Between every three
samples, three standard mixes containing squalane and C_{17} n-alkane (different concentrations as 30 ng/µL, 100 ng/µL and 500 ng/µL, with known δ^{13}C values for each compound) were analyzed to check instrument performance and also for correction purposes. A known amount of squalane was used as an internal standard. The δ^{13}C values are given in the per mil (‰) notation relative to the Vienna PeeDee Belemnite (VPDB) standard. The reproducibility of the GC-IRMS system was <0.5‰ for both standards and repeat analyses of selected samples (n = 3). Due to the co-elution of a few n-alkadiene or n-alkatriene isomers, and the relative lower concentration for some specific non-dominant isomers, only compounds present in sufficient quantities (intensity above 1000 mVs) could be accurately determined for reliable δ^{13}C values. Average values were reported if more than one δ^{13}C value was measured for isomers with the same carbon atom numbers.

Data analysis. Environmental data across multiple locations was obtained from the Florida Coastal Everglades Long Term Ecological Research database (FCE-LTER; http://fcelter.fiu.edu/) and used for comparison with the abundance of the biomarker compounds (botryococcenes, n-alkadienes, n-alkatrienes, and lycopadiene). Statistical analyses were performed using SPSS version 13.0 for Windows. Outliers were tested using the two-sided Grubbs test (P < 0.05). Significant correlations (P < 0.05) between floc physicochemical parameters and the biomarker compounds were determined using Pearson correlation. Significant differences between means of different groups of data were compared using the unpaired t-test (two-tailed, unequal variance).

Results

Identification of n-alkadienes, n-alkatrienes and lycopadiene. GC-MS analysis of the n-hexane eluted fraction from various periphyton and floc sample extracts showed the presence of n-alkadienes, n-alkatrienes and one lycopadiene (Fig. 2; Table 2). A total of 11 C_{27} to C_{32} n-alkadiene and n-alkatriene isomers eluting between the C_{26} to C_{32} n-alkanes were tentatively identified and their Kovats indexes are also given (Figs 2a–f; 3a–k; Table 2). These compounds all exhibit a terminal double bond and one or two mid-chain unsaturations with both Z and E stereochemistry. The mid-chain double bond positions could be further identified based on dimethyl disulfide adducts experiments, but this is not pursued in this present study. Generally, no carbon number predominance was found for these n-alkadienes and n-alkatrienes. Similar no odd or even carbon chain predominance was observed in C_{27}–C_{43} mono-, di- and (to a lesser extent) tri-unsaturated n-alkenes reported in lacustrine sediments. Lycopadi-14,18-diene was identified based on its retention time and mass spectrum match with that in...
the literature (Fig. 3)\(^1\). Another lycopadiene isomer with lower abundance was also identified (Fig. 3). A series of botryococcenes with 32 to 35 carbon atoms are detected in most periphyton and floc samples and elute between \(C_{26}\) to \(C_{29}\) \(n\)-alkanes, in agreement with a previous report (Fig. 2)\(^18\).

Spatial distribution of \(n\)-alkadienes, \(n\)-alkatrienes and lycopadiene. A higher molecular diversity of \(n\)-alkadienes and \(n\)-alkatrienes was detected in floc compared to periphyton (Table 2). Specifically, only \(C_{29}\) and \(C_{31}\) \(n\)-alkadienes, and \(C_{30}\) \(n\)-alkatriene were found in periphyton samples, while \(C_{27}\)–\(C_{31}\) \(n\)-alkatrienes were present in floc samples. Lycopadiene occurred in most of the periphyton samples, but rarely in floc samples. Floc samples (\(n = 86\)) from both ridge and slough locales within the WCA3 area and floc samples (\(n = 12\)) from SRS2 and TSPh2 were analyzed. The \(N\) and \(P\) (nitrogen and phosphorus) concentration of these floc samples were 9.7–46.2 mg/g dw and 73–884 µg/g dw, respectively. The total concentration of \(n\)-alkadienes and \(n\)-alkatrienes of these floc samples ranged from 135 to 6953 ng/g dw. Surprisingly, the abundance of the \(C_{29}\) \(n\)-alkatriene could be up to 2 times above that of the \(C_{29}\) \(n\)-alkane in the same sample (Fig. 2), which is in contrast with previous reports that \(n\)-alkadienes and \(n\)-alkatrienes usually show much lower abundances than the odd numbered \(n\)-alkane homologues\(^1\). No significant correlations were observed (\(P > 0.05\)) between nutrient concentrations and the concentrations of each compound group or the total concentrations in floc (Fig. 4). In addition, no significant correlations were observed between surface water nitrogen and phosphorus concentrations, and abundances of \(n\)-alkadienes and \(n\)-alkatrienes among different locations across the freshwater wetland.

Floc samples from ridge (\(n = 19\)) and slough (\(n = 12\)) environments (within 5 m distance) were analyzed from multiple years (2012 to 2014) within the WCA3 area. The concentrations of \(n\)-alkadienes and \(n\)-alkatrienes in the slough floc samples ranged from 2.0 to 69.5 µg/g dw (average as 13.9 µg/g dw). In contrast, the concentrations of \(n\)-alkadienes and \(n\)-alkatrienes in the ridge floc samples ranged from 0 to 6.6 µg/g dw (average as 1.6 µg/g dw; Fig. 4). The concentrations of \(n\)-alkadienes and \(n\)-alkatrienes were significantly higher in the slough than the ridge floc (unpaired student t-test, two tailed, \(P < 0.01\)). In addition, 8 transects were analyzed from slough to the ridge environments (\(n = 32\)) and obvious concentration decrease trends were observed (Fig. 4).

Compound specific carbon isotope analysis was performed on the dominant \(n\)-alkadienes, \(n\)-alkatrienes and lycopadiene. Due to incomplete GC resolution of some \(n\)-alkadiene or \(n\)-alkatriene isomers with the same carbon number, the \(\delta^{13}C\) values are reported as averages for those compounds (mixtures of \(Z\) and \(E\) isomers). Significantly lower
δ¹³C values were observed for the n-alkadienes, n-alkatrienes and lycopadiene (Table 2) than those of n-alkanes (−32.7 ± 1.8‰) and bulk samples (−30.7 ± 1.4‰). No significant differences in the averaged δ¹³C values were observed between n-alkadienes and n-alkatrienes, whereas the averaged δ¹³C values of the n-alkadienes and n-alkatrienes were lower than those of lycopadiene (Table 3).

### Discussion

**Co-occurrence of B. braunii (A, B, L) indicated by n-alkadienes, n-alkatrienes, and lycopadiene in the Everglades.** Lycopadiene has been reported as a specific biomarker for race L of *B. braunii*, while botryococcenes have been suggested to derive from race B of *B. braunii* in the Everglades. n-Alkanes and n-alkatrienes were not detected in floc and surface soil at the mangrove-dominated estuarine locations, nor in the leaves or roots of dominant plant species across the Everglades ecosystem. n-Alkadienes and n-alkatrienes have been reported in insect wax lipids, but they usually cover higher carbon chain lengths up to C₉⁻.
Odd numbered carbon (poly) unsaturated n-alkenones in the range C_{23}–C_{31} have been characterized in the chlorophyte *Chlorella emersonii*, the diatom *Rhizosolenia setigera* and two marine eustigmatophytes. *C_{30}*, C_{31} and C_{33} alkenes with one to four double bonds are also produced by some haptophytes, such as *Emiliania huxleyi*, *Isochrysis galbana*, *Gephyrocapsa oceanica* and *Chrysothella lamellosa* However, the n-alkadienes and n-alkatrienes detected in this study are all from only the freshwater wetland locations, and thus should not be derived from haptophytes. No significant correlations (concentration based) were observed between n-alkadienes, n-alkatrienes, and the C_{29} HBI (highly branched isoprenoid) across the whole sample set (n = 98, P > 0.05), excluding cyanobacteria as the major source of n-alkadienes, n-alkatrienes detected. Therefore, we suggest that the n-alkadienes and n-alkatrienes detected in this study likely derive from the A race of *B. braunii*. Combining the botryococenes and lycopadiene produced by the B and L races of *B. braunii*, the co-occurrence of three races (A, B and L) of *B. braunii* seems possible.

No significant correlations exist among the abundances of biomarkers of different races of *B. braunii*, which could be caused by: (1) variations in the populations of each race of *B. braunii* across our study area, (2) differences in the hydrocarbon concentration in each race, and (3) different physiological states for each race. Similar results have also been observed in another study. Mixtures of cis n-alkadiene and triene(s) or cis/trans dienes covering the carbon chain range from C_{25} to C_{31} have been characterized in the A race of *B. braunii* across our study area, (2) differences in the hydrocarbon concentration in each race, and (3) different physiological states for each race. Similar results have also been observed in another study. Mixtures of cis n-alkadiene and triene(s) or cis/trans dienes covering the carbon chain range from C_{25} to C_{31} have been characterized in the A race of *B. braunii*.

Environmental controls of *B. braunii* biomarkers and their implications in the Everglades. Although *B. braunii* is known to be sensitive to environmental changes, and botryococenes have been suggested to be applied as a proxy for eutrophication, the lack of correlations between nutrients and n-alkadienes and n-alkatrienes suggest that they seem not to be indicators for eutrophic conditions in this freshwater wetland. Actually, *Botryococcus* was also not suggested to directly reflect nutrient status of waters in the Everglades.

In contrast to nutrients, hydroperiod seems to be one of the controlling factors for the distribution of n-alkadienes and n-alkatrienes. Significant higher abundances of n-alkadienes and n-alkatrienes were observed in the ridge than slough floc, which could be explained by the following reasons: (1) the A race of *B. braunii* has the ability to float due to its high lipid concentrations, which leads to its enrichment in the slough environment; (2) more diagenetic degradation of n-alkadienes and n-alkatrienes occurs in the ridge environment due to stronger oxidation. In this study, n-alkadienes and n-alkatrienes were only observed in the floc at locations SRS2 and WCA3, in agreement with higher concentrations of botryococenes reported at these two sites. This could be mainly attributed to longer hydroperiod (WCA3 and SRS2), and lower water flow velocities (WCA3), resulting in reduced floc transport (Table 1). The longer hydroperiod could induce sub-oxic or anoxic conditions in the floc layer, and thus decrease carbon mineralization rates. However, other factors may also contribute to the concentration difference among different sites, such as the composition of periphyton and these require further investigations (Table 1).

n-Alkadienes and n-alkatrienes were only detected in WCA3 slough soils/sediments (Fig. 3), but not in all SRS and TSPh sites, which could likely be due to: (1) a more complex microbial composition in periphyton and floc compared to soils, (2) limited incorporation of periphyton and floc into soils, or (3) early diagenetic reworking or microbial degradation of these compounds by heterotrophs such as bacteria and fungi. Several sulfur-containing compounds and two thiophenes both with 20 carbon atoms (3-methyl-2-(3',7',11'-trimethyldecyl)thiophene and 3-(4',8',12'-trimethyltridecyl)thiophene were detected in most of the floc samples (data not shown), suggesting anoxic or sub-oxic conditions. If early diagenetic reduction of the unsaturations is one of the factors accounting for the absence of n-alkadienes and n-alkatrienes in all deeper soils/sediments, part of the C_{29}–C_{33} n-alkanes detected in sediments of the Florida wetland could also be derived from the n-alkadienes and n-alkatrienes. However, further investigation is needed. Lycopadiene was not detected in surface and deeper soils, likely due to: (1) a much lower amount of lycopadiene produced, or (2) diagenetic transformation of lycopadiene into higher molecular weight compounds. However, lycopadiene was reported in a few studies including freshwater lake sediments from the Holocene and an oil shale from the Pliocene. In addition, a monoaromatic lycopane derivative was reported from the Messel oil shale, and kerogen fractions of samples from oil shale layer 4 in the Eocene Huidian Formation, NE China. Adam et al. proposed that this compound could be a specific biomarker for race L of *B. braunii* in sediments deposited under freshwater and/ or brackish conditions. Though analyzing a Holocene freshwater lake sediment core, Zhang et al. suggested that n-alkadienes, botryococenes and lycopadienes can survive in oxic sediments for several decades, and the down core variation in these lipids likely reflects changes in environmental conditions either favoring the bloom or near-extinction of *B. braunii*. However, this present study shows that botryococenes were widely detected, and
n-alkadienes and n-alkatrienes were rarely present, while lycopadiene was not detected in soils, suggesting a recent contribution of these compounds likely due to the blooming of *B. braunii*.

The abundance of these compounds does not correlate with both bulk N and P concentrations in floc samples or surface water, suggesting that nutrients may not be the controlling factors for the distributions of these compounds in this ecosystem. In contrast, slough floc contains significantly higher amounts of n-alkadienes and n-alkatrienes than ridge floc. Thus, hydroperiod could be one of the controlling factors for the abundances of n-alkadienes and n-alkatrienes within this freshwater wetland. However, further investigation is needed to refine the application of these biomarkers as indicators of community structure of *B. braunii* in the Everglades.

**References**

1. Zhang, Z. P. et al. Biomarker evidence for the co-occurrence of three races (A, B and L) of *Botryococcus braunii* in El Junco Lake, Galápagos. *Org. Geochem.* **38**, 1459–1478 (2007).

2. Maxwell, J. R. et al. The Botryococcus—hydrocarbons of novel structure from the alga *Botryococcus braunii*, Kützing. *Phytochem.* **7**, 2157–2171 (1968).

3. Metzger, P., Largeau, C. & Casadevall, E. Lipids and macromolecular lipids of the hydrocarbon-rich microalga *Botryococcus braunii*. Chemical structure and biosynthesis. Geochemical and biotechnological importance. In *Fortsschritte der Chemie organischer Naturstoffe/Progress in the Chemistry of Organic Natural Products* (pp. 1–70). Springer Vienna (1991).

4. Metzger, P. et al. *Botryococcus braunii*: a rich source for hydrocarbons and related ether lipids. *Appl. Microbiol. Biot.* **66**, 486–496 (2005).

5. Volkman, J. K. Acyclic iso-prenoid biomarkers and evolution of biosynthetic pathways in green microalgae of the genus *Botryococcus*. *Org. Geochem*. **75**, 36–47 (2014).

6. Metzger, P. et al. Alkadiene-and botryococcene-producing races of wild strains of *Botryococcus braunii*. *Phytochem.* **24**, 2305–2312 (1985).

7. Metzger, P. et al. An n-alkatriene and some n-alkadienes from the A race of the green alga *Botryococcus braunii*. *Phytochem.* **25**, 1869–1872 (1986).

8. Derenne, S. et al. Direct relationship between the resistant biopolymer and the tetraterpenic hydrocarbon in the lycopadiene race of *Botryococcus braunii*. *Phytochem.* **29**, 2187–2192 (1990).

9. Adam, P. et al. C30 mono- to octadecamethyloctane derivatives as indicators of the contribution of the alga *Botryococcus braunii* race L to the organic matter of sussel oil shale (Eocene, Germany). *Org. Geochem.* **37**, 584–596 (2006).

10. Salmon, E. et al. Thermal decomposition processes in algal *Botryococcus braunii* race L. Part 1: experimental data and structural evolution. *Org. Geochem.* **40**, 400–415 (2009).

11. Davis, S. M. et al. Landscape dimension, composition, and function in a changing Everglades ecosystem. In: S. M. Davis, J. C. Ogden (Eds), *The Everglades: North America’s subtropical wetland*. St. Lucie Press, Delray Beach, Florida, pp. 419–444 (1994).

12. Gaiser, E. E. et al. Landscape patterns of periphyton in the Florida Everglades. *Crit. Rev. Environ. Sci. Tec.* **41**, 92–120 (2011).

13. Hagerthley, S. E. et al. Everglades periphyton: a biogeochemical perspective. *Crit. Rev. Environ. Sci. Tec.* **41**, 309–343 (2011).

14. Gropp, I. G. et al. The freshwater floc: a functional relationship of water and organic and inorganic floc constituents affecting suspended sediment properties. *Water Air Soil Poll.* **99**, 43–54 (1997).

15. Gao, M. et al. Occurrence and distribution of novel botryococcene hydrocarbons in freshwater wetlands of the Florida Everglades. *Chemosphere* **70**, 224–236 (2007).

16. Saunders, C. J. et al. Environmental assessment of vegetation and hydrological conditions in Everglades freshwater marshes using multiple geochemical proxies. *Aquat. Sci.* **77**, 271–291 (2015).

17. Neto, R. R. et al. Organic biogeochemistry of detrital floculent material (floc) in a subtropical, coastal wetland. *Biogeochem.* **77**, 283–304 (2006).

18. He, D. et al. Occurrence and distribution of monomethylalkanes in the freshwater wetland ecosystem of the Florida Everglades. *Chemosphere* **119**, 258–266 (2015).

19. De Mesmay, R. et al. Novel monoo-, di- and tri-unsaturated very long chain (C37–C43) n-alkanes in alkene-free lacustrine sediments (Lake Masoko, Tanzania). *Org. Geochem.* **38**, 323–333 (2007).

20. He, D. et al. Assessing source contributions to particulate organic matter in a subtropical estuary: A biomarker approach. *Org. Geochem.* **75**, 129–139 (2014).

21. He, D. et al. Compositions and isotopic differences of iso- and anteiso-alkanes in black mangroves (*Avicennia germinans*) across a salinity gradient in a subtropical estuary. *Environ. Chem.* **13**, 623–630 (2015).

22. Lockey, K. H. Insect hydrocarbon classes: implications for chemotaxonomy. *Insect Biochem.* **21**, 91–97 (1991).
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Author Contributions
D. He and R. Jaffé designed this study and performed the experiments. D. He performed most of the laboratory work and wrote the initial draft of the manuscript. Both B.R.T. Simoneit and R. Jaffé contributed to streamlining of the manuscript and contributed to data interpretation. All authors worked on the manuscript revisions.

Additional Information

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