Association Study of Serine Racemase Gene with Methamphetamine Psychosis

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Abstract: Experimental studies have demonstrated that not only dopaminergic signaling but also glutamatergic/NMDA receptor signaling play indispensable roles in the development of methamphetamine psychosis. Our recent genetic studies provided evidence that genetic variants of glutamate-related genes such as DTNBP1, GLYT1, and G72, which are involved in glutamate release and regulation of co-agonists for NMDA receptors, conferred susceptibility to methamphetamine psychosis. Serine racemase converts l-serine to d-serine, which is an endogenous co-agonist for NMDA receptors. Three single nucleotide polymorphisms (SNPs) in the promoter region of the serine racemase gene (SRR), rs224770, rs3760229, and rs408067, were proven to affect the transcription activity of SRR. Therefore, we examined these SNPs in 225 patients with methamphetamine psychosis and 291 age- and sex-matched controls. There was no significant association between methamphetamine psychosis and any SNP examined or between the disorder and haplotypes comprising the three SNPs. However, rs408067 was significantly associated with the prognosis for methamphetamine psychosis and multi-substance abuse status. The patients with C-positive genotypes (CC or CG) of rs408067 showed better prognosis of psychosis after therapy and less abuse of multiple substances than the patients with GG genotypes. Because the C allele of rs408067 reduces the expression of SRR, a lower d-serine level or reduced NMDA receptor activation may affect the prognosis of methamphetamine psychosis and multiple substance abuse. Our sample size is, however, not large enough to eliminate the possibility of a type I error, our findings must be confirmed by replicate studies with larger samples.

Keywords: Methamphetamine psychosis, serine racemase, glutamate, NMDA receptors, SNP.

INTRODUCTION

Methamphetamine is widely abused around the world [1, 2]. In Japan, it has been the most popular illegal abused substance since World War II, and its use produces serious social problems [3, 4]. Development of novel or innovative medicines that are more efficient for treatment of methamphetamine use disorders is necessary. Methamphetamine has a strong psychostimulant effect on the CNS, and elucidation of the neuronal mechanisms underlying methamphetamine-induced psychological dependence and psychosis is considered a top priority. Many experimental studies using a behavioral sensitization model, an animal model of methamphetamine dependence and psychosis, have revealed that the dopamine system in the brain plays central and indispensable roles in the induction and expression of methamphetamine use disorders [5]. These findings seem reasonable because methamphetamine is an indirect dopamine agonist. In addition, animal studies have also revealed that glutamate and its N-methyl-D-aspartate (NMDA) receptors play important roles in methamphetamine use disorders. Thus, methamphetamine administration enhanced glutamate release in the rat accumbens and ventral tegmentum area (VTA) [6, 7]. Repeated amphetamine administration results in enhanced neuronal responsiveness to locally applied glutamate in the VTA [8, 9] and frontal cortex [10]. Repeated methamphetamine administration induces behavioral sensitization, which was blocked by co-administration of MK-801 (dizocilpine) or CPP (D, L-3 [(+/−)-2-carboxypiperazin-4-yl]-propyl-1-phosphonic acid), an NMDA-receptor antagonist by systemic administration or microinjection into the VTA [11-14]; further, infusion of iGluR antagonists into the accumbens reduced the motor stimulant effects of amphetamine [15]. Reductions in glutamate transmission by administration of the glutamate-release inhibitor riluzole [16], the glutamate transporter activator MS-153 [17], infusion of an AMPA (α-amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid)/KA (kainite) antagonist into the accumbens [18], and virally mediated overexpression of EAAT2 (a glial high affinity glutamate transporter, SLC1A2) in the accumbens [19] all have been shown to attenuate the development of amphetamine conditioned place preference. Mice deficient in GluRepsilon 1, which is a human ortholog of the NR2A subunit of the
NMDA receptor, showed attenuation of acute methamphetamine-induced hyperlocomotion and low dose methamphetamine-induced sensitization [20]. These experimental findings indicate that enhanced glutamate transmission via NMDA and/or AMPA receptors due to methamphetamine administration is essential for induction of sensitization and conditioning to the drug [5]. Therefore, it is possible that genomic variants of genes encoding molecules involved in glutamate signaling may affect individual susceptibility to methamphetamine dependence and psychosis. Based on this hypothesis, we previously analyzed the genetic association of several genes related to glutamate signaling, such as DTNBPI, GLYT1, and G72, and found that all these genes were significantly associated with methamphetamine use disorders [21-23]. Serine racemase is an enzyme that converts l-serine to d-serine and increases the level of d-serine, which is a co-agonist for the NMDA receptor and regulates activity of NMDA receptors in the brain [24, 25]. Therefore, we investigated a possible association between the serine racemase gene (SRR) and methamphetamine psychosis.

MATERIALS AND METHODS

Subjects

The subjects comprised 225 patients with methamphetamine psychosis (181 male and 44 female; mean age ± SD, 37.5 ± 11.9) and 274 age-, gender-, and geographical-origin matched healthy controls (217 male and 57 female; mean age ± SD, 37.6 ± 13.1). All subjects were unrelated Japanese born and living in relatively restricted areas of Japan. All patients were out- or inpatients in psychiatric hospitals of the Japanese Genetics Initiative for Drug Abuse (JGIDA). Consensus diagnoses of the patients were made by two trained psychiatrists according to ICD-10 criteria on the basis of unstructured interviews and medical records. The patients suffered from F15.5 (methamphetamine psychosis) and also F15.2 (methamphetamine dependence). The controls had no individual or family history of drug dependence or major psychotic disorders such as schizophrenia or bipolar disorder. This study was approved by the ethics committee of each JGIDA institution. After a complete description of the study to the subjects, written informed consent was obtained.

Clinical Phenotypes

The patients with methamphetamine psychosis were divided into subgroups according to the following clinical characteristics: age at first consumption of methamphetamine (younger or older than 20 years), latency to the onset of psychotic symptoms after the first consumption (less than or more than 3 years), prognosis of methamphetamine psychosis after therapy (transient type: psychotic symptoms disappeared within 1 month after discontinuance of methamphetamine use and treatment with an antipsychotic, or prolonged type: psychotic symptoms lasted more than 1 month even after discontinuance of methamphetamine use and treatment with an antipsychotic), spontaneous relapse to a psychotic state, and the presence or absence of multiple-substance abuse. Detailed information on the clinical subgroups of methamphetamine dependence and psychosis was reported elsewhere [22, 26].

Genotyping

Peripheral blood was obtained from the subjects, and genomic DNA was extracted from peripheral leukocytes using the standard method. Based on our previous study of schizophrenia, three SNPs in the promoter region, rs224770, rs3760229, and rs408067, were analyzed. The first two SNPs were genotyped using TaqMan®-based techniques (Applied Biosystems 7300/7500 Real Time PCR System, Applied Biosystems Japan Ltd., Tokyo, Japan) with TaqMan probes (assay ID C_1027003_10 for rs224770 and C_1625595_10 for rs3760229). The polymerase chain reaction was carried out in a total volume of 7 μL containing 3.2 μL TaqMan® Universal PCR Master Mix, 0.32 μL TaqMan® probe, and 20 ng of DNA. The amplification protocol was denaturation at 95°C for 10 min, followed by 40 cycles of denaturation at 92°C for 15 sec, and annealing and extension at 60°C for 1 min. Rs408067 was genotyped by the restriction enzyme fragment length polymorphism method. PCR was carried out in a total volume of 15 μl with 10% dimethyl sulfoxide and 0.75 units of Taq DNA polymerase in the reaction mixture with mismatch probes (forward: 5′ CCG GCC GCA CCT CTT GCC AGC CGC GC 3′, reverse: 5′ CCC CCT GCC GCC CTC CCT TTC CGA AGC 3′). Initial denaturation was performed for 5 min at 95°C, then 35 cycles of 30 sec of denaturing at 95°C, 30 sec of annealing at 68°C, and 30 sec of extension at 72°C were performed, followed by a final extension at 72°C for 5 min. The PCR products were then analyzed on 4.5% agarose gels after digestion with EheI.

Statistical Analysis

Deviation of the genotypes from Hardy-Weinberg equilibrium was tested using a chi-square goodness-of-fit test. The statistical significance of the difference was assessed by a chi-square test (genotype comparison) or log likelihood ratio test (allele comparison) at a significance level of 0.05. The pairwise linkage disequilibrium (LD) and haplotype frequencies were estimated by the EH algorithm using the SNPalyze program (Dynacom Co., Mobara, Chiba, Japan).

RESULTS

Genotype distributions and allele frequencies of the three SNPs of the SRR gene of the patients with methamphetamine psychosis and control subjects are shown in Table 1. The genotype distributions of the control subjects did not deviate from Hardy-Weinberg equilibrium at any of the three SNPs (rs224770, χ²=0.86, P=0.35; rs3760229, χ²=0.11, P=0.74; rs408067, χ²=0.48, P=0.49). In the single marker analysis, no SNP showed a significant difference between patients and control subjects in the frequency distribution of the genotype (rs224770, χ²=1.23, P=0.54; rs3760229, χ²=2.51, P=0.29; rs408067, χ²=1.21, P=0.55) or allele (rs224770, χ²=0.73, P=0.39; rs3760229, χ²=0.026, P=0.87; rs408067, χ²=0.022, P=0.88).

Table 2 shows the results of the pairwise linkage disequilibrium (LD) among the three SNPs of the SRR gene using the D’ and r² values as an index. The D’ and r² between each pair of SNPs were above 0.7 and 0.3, respectively, which indicated strong LD between the SNPs. Accordingly, the three SNPs are located on the same LD block. The LD structure found in the present study is consistent with our
previous study of schizophrenia [27]. Then, we analyzed three marker haplotypes consisting of rs224770, rs3760229, and rs408067 (Table 3). Five haplotypes were found, and the G-T-G haplotype was the major one. There was no significant difference in estimated frequencies of haplotypes between the patients with methamphetamine psychosis and controls (global permutation $p$ value = 0.068).

Effects of the SNPs on several clinical phenotypes, age at first abuse of methamphetamine, latency of psychosis from the first abuse, prognosis of psychotic symptoms after therapy, complication of spontaneous relapse of psychosis and multi-substance abuse status, were analyzed. Allelic and genotypic distributions of rs224770 and rs3760229 were not associated with these clinical phenotypes (Table 4A and 4B);

### Table 1. Association between the SRR Gene and Methamphetamine Psychosis

| Rs 2224770 | Group | N | Genotype | Allele | p value |
|------------|-------|---|----------|--------|---------|
|            |       |   | G/G (%)  | G/A (%)| A/A (%) |        |
| Control    | 291   |   | 169 (58.1)| 101 (34.7)| 21 (7.2)| 0.54 |
| METH       | 221   |   | 118 (53.4)| 87 (39.4)| 16 (7.2)|        |

| Rs 3760229 | Group | N | Genotype | Allele | p value |
|------------|-------|---|----------|--------|---------|
|            |       |   | T/T (%)  | T/C (%)| C/C (%) |        |
| Control    | 292   |   | 94 (32.2)| 140 (47.9)| 58 (19.9)| 0.29 |
| METH       | 225   |   | 66 (29.3)| 123 (54.7)| 36 (16.0)|        |

| Rs 408067 | Group | N | Genotype | Allele | p value |
|-----------|-------|---|----------|--------|---------|
|            |       |   | G/G (%)  | G/C (%)| C/C (%) |        |
| Control    | 291   |   | 169 (58.1)| 102 (35.1)| 20 (6.9)| 0.55 |
| METH       | 225   |   | 128 (56.9)| 86 (38.2)| 11 (4.9)|        |

### Table 2. Linkage Disequilibrium Among the Three SNPs of the SRR Gene

| Rs2224770 | Rs3760229 | Rs408067 |
|-----------|-----------|----------|
| rs2224770 | 0.4142    | 0.9082   |
| rs3760229 | 1         | 0.3631   |
| rs408067  | 0.953     | 0.9374   |

### Table 3. Association of Haplotypes of the SRR Gene with Methamphetamine Psychosis

| Haplotype | Control | Patients |
|-----------|---------|----------|
| rs2224770 | rs3760229 | rs408067 |
| N=580     | N=440    |          |
| G  | T  | G  | 0.552 | 0.554 |
| A  | C  | C  | 0.237 | 0.233 |
| G  | C  | G  | 0.194 | 0.167 |
| A  | C  | G  | 0.009 | 0.032 |
| G  | T  | C  | 0.007 | 0.009 |

Global permutation $p$ value = 0.068.
### Table 4A-C. Association of Subgroups with Methamphetamine Psychosis According to Clinical Phenotype

#### 4A

| Subgroup                  | N  | G/G (%) | G/A (%) | A/A (%) | p value  | G (%)  | A (%)  | p value |
|---------------------------|----|---------|---------|---------|-----------|--------|--------|---------|
| **Age at First Use**      |    |         |         |         |           |        |        |         |
| 20y<                      | 118| 62(52.5)| 47(39.8)| 9(7.6)  | 0.98      | 171(72.5)| 65(27.5)|          |
| 20y>=                     | 100| 54(54.0)| 39(39.0)| 7(7.0)  | 0.82      | 147(73.5)| 53(26.5)|          |
| **Latency of Psychosis**  |    |         |         |         |           |        |        |         |
| 3y<                       | 101| 52(51.5)| 40(39.6)| 9(8.9)  | 0.40      | 144(71.3)| 58(28.7)|          |
| 3y>=                      | 92 | 53(57.6)| 35(38.0)| 4(4.3)  | 0.23      | 141(76.6)| 43(23.4)|          |
| **Prognosis of Psychosis**|    |         |         |         |           |        |        |         |
| Transient                 | 112| 53(47.3)| 50(44.6)| 9(8.0)  | 0.14      | 156(69.6)| 68(30.3)|          |
| Prolonged                 | 90 | 55(61.1)| 30(33.3)| 5(5.6)  | 0.066     | 140(30.4)| 40(22.2)|          |
| **Spontaneous Relapse**   |    |         |         |         |           |        |        |         |
| No                        | 116| 68(58.6)| 41(35.3)| 7(6.0)  | 0.88      | 177(76.3)| 55(23.7)|          |
| Yes                       | 81 | 50(61.7)| 26(32.1)| 5(6.2)  | 0.81      | 111(58.4)| 36(22.2)|          |
| **Multi-substance abuse** |    |         |         |         |           |        |        |         |
| None or mild              | 137| 67(50.8)| 52(39.4)| 13(9.8) | 0.15      | 186(70.5)| 78(29.5)|          |
| Heavy                     | 82 | 47(57.3)| 32(39.0)| 3(3.7)  |           | 126(76.8)| 38(23.2)|          |

#### 4B

| Subgroup                  | N  | T/T (%) | T/C (%) | C/C (%) | p value  | T (%)  | C (%)  | p value |
|---------------------------|----|---------|---------|---------|-----------|--------|--------|---------|
| **Age at First Use**      |    |         |         |         |           |        |        |         |
| 20y<                      | 119| 34(28.6)| 67(56.3)| 18(15.1)| 0.88      | 135(56.7)| 103(43.2)|          |
| 20y>=                     | 103| 31(30.1)| 54(52.4)| 18(17.5)| 0.88      | 116(56.3)| 90(43.7)|          |
| **Latency of Psychosis**  |    |         |         |         |           |        |        |         |
| 3y<                       | 102| 32(31.4)| 53(52.0)| 17(16.7)| 0.83      | 117(57.4)| 87(42.6)|          |
| 3y>=                      | 95 | 28(29.5)| 55(57.9)| 12(12.6)| 0.36      | 111(58.4)| 79(41.6)|          |
| **Prognosis of Psychosis**|    |         |         |         |           |        |        |         |
| Transient                 | 114| 29(25.4)| 68(59.6)| 17(14.9)| 0.36      | 126(55.3)| 102(44.7)|          |
| Prolonged                 | 92 | 32(34.8)| 46(50.0)| 14(15.2)| 0.36      | 110(59.8)| 74(40.2)| 0.36    |
| **Spontaneous Relapse**   |    |         |         |         |           |        |        |         |
| No                        | 120| 30(25.0)| 69(57.5)| 21(17.5)| 0.20      | 129(53.8)| 111(46.2)|          |
| Yes                       | 94 | 31(33.0)| 51(54.3)| 12(12.8)| 0.20      | 113(60.1)| 75(39.9)| 0.20    |
| **Multi-substance abuse** |    |         |         |         |           |        |        |         |
| None or mild              | 135| 37(27.4)| 71(52.6)| 27(20.0)| 0.14      | 145(53.7)| 125(46.3)|          |
| Heavy                     | 83 | 26(31.1)| 49(59.0)| 8(9.6)  |           | 101(60.8)| 65(39.2)| 0.14    |
however, rs408067 was significantly associated with a prognosis for methamphetamine psychosis and multi-substance abuse status (Table 4C). The patients with C-positive genotypes (CC or CG) of rs408067 showed the transient type of psychosis significantly more frequently after standard therapy with antipsychotics than the patients with the GG genotype (p=0.039). This indicated that C-positive genotypes of rs408067 may have a better response to antipsychotic treatment and better prognosis of methamphetamine psychosis. The patients with the C allele or C-positive genotypes of rs408067 also showed significantly less heavy multi-substance abuse than those with the G allele or GG genotype (p=0.036 and 0.029, respectively). They abuse fewer illegal substances other than methamphetamine, such as morphine, cocaine, or cannabis.

**DISCUSSION**

The present study revealed that the SRR gene did not affect susceptibility to methamphetamine use disorders, but it was associated with several clinical phenotypes of methamphetamine dependence and psychosis. Carriers of the C allele of rs408067 showed the less prolonged type of methamphetamine psychosis compared with non-carriers, indicating a better prognosis after therapy with antipsychotics and less multi-substance abuse of illegal drugs other than methamphetamine. Our previous study [27] found that rs408067, which is located in the promoter region of the SRR gene, may be functional because an *in vitro* luciferase assay showed that the C allele had 60% lower promoter activity than the G allele did. Therefore, having the C allele of rs408067 may produce a decrease in the SRR level and d-serine conversion, and a decreased level of d-serine, a co-agonist for NMDA receptors, may result in reduced activity of NMDA receptors. Therefore, reduced NMDA receptor signaling due to possession of the C allele of rs408067 of the SRR gene may be a protective factor against a worse prognosis of methamphetamine psychosis and heavy multi-substance abuse behaviors.

A set of molecules are involved in the regulation of glutamate and NMDA receptor signaling. One of those is dysbindin, which is encoded by the DTNBP1 gene. Dysbindin was shown to be involved in glutamate release from synaptic terminals [28]. Recently, we found that the polymorphisms of P1635 (rs3213207) and SNPA (rs2619538) and the three-locus haplotype of P1655 (rs2619539)-P1635-SNPA of the DTNBP1 gene showed significant associations with methamphetamine psychosis [22]. The P1635 polymorphism of DTNBP1 was also associated with a worse prognosis for methamphetamine psychosis. We also reported that two genes involved in regulation of d-serine were associated with methamphetamine psychosis [21, 23]. They were the GLYT1 and G72 genes. GLYT1 encodes glycine transporter type 1, which locates in glia close to NMDA receptors and reuptakes glycine and d-serine; both are allosteric co-agonists of

### Table 4. contd….

| Subgroup | N  | G/G (%) | G/C (%) | C/C (%) | p value | G (%) | C (%) | p value |
|----------|----|---------|---------|---------|---------|-------|-------|---------|
| Age at First Use | | | | | | | | |
| 20y< | 119 | 67(56.3) | 45(37.8) | 7(5.9) | 179(75.2) | 59(24.8) | | |
| 20y>= | 103 | 59(57.3) | 40(38.8) | 4(3.9) | 0.95 | 158(76.7) | 48(23.3) | 0.85 |
| Latency of Psychosis | | | | | | | | |
| 3y< | 103 | 58(56.3) | 40(38.8) | 5(4.9) | 156(75.7) | 50(24.3) | | |
| 3y>= | 94 | 56(59.6) | 36(38.3) | 2(2.1) | 0.57 | 148(78.7) | 40(21.3) | 0.48 |
| Prognosis of Psychosis | | | | | | | | |
| Transient | 114 | 58(50.9) | 52(45.6) | 4(3.5) | 168(73.7) | 60(26.3) | | |
| Prolonged | 92 | 60(65.2) | 27(29.3) | 5(5.4) | 0.056 | 147(79.9) | 37(20.1) | 0.14 |
| Spontaneous Relapse | | | | | | | | (0.039) |
| No | 120 | 65(54.2) | 47(39.2) | 8(6.7) | 177(73.8) | 63(26.2) | | |
| Yes | 94 | 56(59.6) | 36(38.3) | 2(2.1) | 0.27 | 148(78.7) | 40(21.3) | 0.23 |
| Multi-substance abuse | | | | | | | | |
| None or mild | 135 | 71(52.6) | 54(40.0) | 10(7.4) | 196(72.6) | 74(27.4) | | (0.029) |
| Heavy | 83 | 53(63.9) | 29(34.9) | 1(1.2) | 0.067 | 135(81.3) | 31(18.7) | 0.038 |

P values in parentheses indicate those between GG vs C-positive genotypes (GC and CC).
NMDA receptors. GLYT1 was significantly associated with methamphetamine use disorders. G72 encodes an activator of d-amino acid oxidase [29], which metabolizes d-serine by oxidation. Two haplotypes of the G72 gene in the 5’ and 3’ regions were significantly associated with methamphetamine psychosis [21]. Thus, our previous and present findings consistently indicate that genetic variants that may affect neural signaling of glutamate and NMDA receptors affect susceptibility to methamphetamine-use disorders including psychosis and associated clinical characteristics.

The sample size in the present study was not too small, but it was not large enough to exclude the possibility of a type I error. Therefore, our findings must be replicated in an independent larger sample and other populations.

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REFERENCES

[1] Maxwell, J.C.; Rutkowski, B.A. The prevalence of methamphetamine and amphetamine abuse in North America: a review of the indicators, 1992-2007. *Drug Alcohol Rev.*, 2008, 27, 229-235.

[2] Griffiths, P.; Mravcik, V.; Lopez, D.; Klemova, D. Quite a lot of smoke but very limited fire—the use of methamphetamine in Europe. *Drug Alcohol Rev.*, 2008, 27, 236-242.

[3] Tatemoto, S. Methamphetamine psychosis. *Folia Psychiatr. Neurol. Jpn. Suppl.*, 1963, 7, 377-380.

[4] Ujike, H.; Sato, M. Clinical features of sensitization to methamphetamine observed in patients with methamphetamine dependence and psychosis. *Ann. N. Y. Acad. Sci.*, 2004, 1025, 279-287.

[5] Ujike, H. Stimulant-induced psychosis and schizophrenia: the role of sensitization. *Curr. Psychiatry Rep.*, 2002, 4, 177-184.

[6] Xue, C.J.; Ng, J.P.; Li, Y.; Wolf, M.E. Acute and repeated systemic amphetamine administration: effects on extracellular glutamate, aspartate, and serine levels in rat ventral tegmental area and nucleus accumbens. *J. Neurochem.*, 1996, 67, 352-363.

[7] Wolf, M.E.; Xue, C.J. Amphetamine-induced glutamate efflux in the rat ventral tegmental area is prevented by MK-801, SCH 23930, and ibotenic acid lesions of the prefrontal cortex. *J. Neurochem.*, 1999, 73, 1529-1538.

[8] White, F.J.; Hu, X.T.; Zhang, X.F.; Wolf, M.E. Repeated administration of cocaine or amphetamine alters neuronal responses to glutamate in the mesocambellus dopamine system. *J. Pharmacol. Exp. Ther.*, 1995, 273, 445-454.

[9] Zhang, X.F.; Hu, X.T.; White, F.J.; Wolf, M.E. Increased responsiveness of ventral tegmental area dopamine neurons to glutamate after repeated administration of cocaine or amphetamine is transient and selectively involves AMPA receptors. *J. Pharmacol. Exp. Ther.*, 1997, 281, 699-706.

[10] Peterson, J.D.; Wolf, M.E.; White, F.J. Altered responsiveness of medial prefrontal cortex neurons to glutamate and dopamine after withdrawal from repeated amphetamine treatment. *Synapse*, 2000, 36, 342-344.

[11] Karlet, R.; Calder, L.D.; Chaudhry, I.A.; Turkcanis, S.A. Blockade of "reverse tolerance" to cocaine and amphetamine by MK-801. *Life Sci.*, 1989, 45, 599-606.

[12] Battisti, J.J.; Uretsky, N.J.; Wallace, L.J. NMDA glutamate receptor role in the development of context-dependent and independent sensitization of the induction of stereotypy by amphetamine or apomorphine. *Behav. Brain Res.*, 2000, 114, 167-174.

[13] Kalivas, P.W.; Aleskader, J.E. Involvement of N-methyl-D-aspartate receptor stimulation in the ventral tegmental area and amygdala in behavioral sensitization to cocaine. *J. Pharmacol. Exp. Ther.*, 1993, 267, 486-495.

[14] Cador, M.; Bijou, Y.; Callihol, S.; Stinus, L. D-amphetamine-induced behavioral sensitization: implication of a glutamatergic medial prefrontal cortex-ventral tegmental area innervation. *Neuroscience*, 1999, 94, 705-721.

[15] Pulvirenti, L.; Swedlow, N.R.; Koob, G.F. Microinjection of a glutamate antagonist into the nucleus accumbens reduces psychostimulant locomotion in rats. *Neurosci. Lett.*, 1989, 103, 213-218.

[16] Tschentke, T.M.; Schmidt, W.I. Blockade of morphine- and amphetamine-induced conditioned place preference in the rat by riluzole. *Neurosci. Lett.*, 1998, 242, 114-116.

[17] Nakagawa, T.; Fujio, M.; Ozawa, T.; Minami, M.; Satoh, M. Effect of MS-153, a glutamate transporter activator, on the conditioned rewarding effects of morphine, methamphetamine and cocaine in mice. *Behav. Brain Res.*, 2005, 156, 233-239.

[18] Layer, R.T.; Uretsky, N.J.; Wallace, L.J. Effects of the AMPA/kainate receptor antagonist DNBQX in the nucleus accumbens on drug-induced conditioned place preference. *Brain Res.*, 1993, 617, 267-273.

[19] Fujio, M.; Nakagawa, T.; Sekiya, Y.; Ozawa, T.; Suzuki, Y.; Minami, M.; Satoh, M.; Kaneko, S. Gene transfer of GLT-1, a glutamate transporter, into the nucleus accumbens shell attenuates methamphetamine- and morphine-induced conditioned place preference in rats. *Eur. J. Neurosci.*, 2005, 22, 2744-2754.

[20] Miyamoto, Y.; Yamada, K.; Nagai, T.; Mori, H.; Mishina, M.; Furukawa, N.; Noda, Y.; Nabeshima, T. Behavioural adaptations to addictive drugs in mice lacking the NMDA receptor epsilon1 subunit. *Eur. J. Neurosci.*, 2004, 19, 151-158.

[21] Kotaka, T.; Ujike, H.; Okahisa, Y.; Takaki, M.; Nakata, K.; Kodama, M.; Inada, T.; Yamada, M.; Uchimura, N.; Iwata, N.; Sora, I.; Iyo, M.; Ozaki, N.; Kuroda, S. G72 gene is associated with susceptibility to methamphetamine psychosis. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, 2009, 33, 1046-1049.

[22] Kishimoto, M.; Ujike, H.; Motohashi, Y.; Tanaka, Y.; Okahisa, Y.; Kotaka, T.; Harano, M.; Inada, T.; Yamada, M.; Komiyama, T.; Hori, T.; Sekine, Y.; Iwata, N.; Sora, I.; Iyo, M.; Ozaki, N.; Kuroda, S. The dysbindin gene (DTNBP1) is associated with methamphetamine psychosis. *Biol. Psychiatry*, 2008, 63, 191-196.

[23] Morita, Y.; Ujike, H.; Tanaka, Y.; Kishimoto, M.; Okahisa, Y.; Kotaka, T.; Harano, M.; Inada, T.; Komiyama, T.; Hori, T.; Yamada, M.; Sekine, Y.; Iwata, N.; Iyo, M.; Sora, I.; Ozaki, N.; Kuroda, S. The glycine transporter 1 gene (GLYT1) is associated with methamphetamine-use disorder. *Am. J. Med. Genet. B, Neuropsychiatr. Genet.*, 2008, 147B, 54-58.

[24] Wolosker, H.; Sheh, K.N.; Takahashi, M.; Mothet, J.P.; Brady, R.O., Jr.; Ferris, C.D.; Snyder, S.H. Purification of serine racemase: biosynthesis of the neurmodulator D-serine. *Proc. Natl. Acad. Sci. USA*, 1999, 96, 721-725.

[25] Xia, M.; Liu, Y.; Figueroa, D.J.; Chiu, C.S.; Wei, N.; Lawlor, A.M.; Lu, P.; Sur, C.; Koblan, K.S.; Connolly, T.M. Characterization and localization of a human serine racemase. *Brain Res. Mol. Brain Res.*, 2004, 125, 96-104.

[26] Ujike, H.; Harano, M.; Inada, T.; Yamada, M.; Komiyama, T.; Sekine, Y.; Sora, I.; Matsu, K.; Nomura, A.; Nakata, K.; Ozaki, N. Nine- or fewer repeat alleles in VNTR polymorphism of the dopamine transporter gene is a strong risk factor for prolonged methamphetamine psychosis. *Pharmacogenom. J.*, 2003, 3, 242-247.

[27] Morita, Y.; Ujike, H.; Tanaka, Y.; Otani, K.; Kishimoto, M.; Morio, A.; Kotaka, T.; Okahisa, Y.; Matsuhisa, M.; Morikawa, A.; Hamase, K.; Zaitus, K.; Kuroda, S. A genetic variant of the serine racemase gene is associated with schizophrenia. *Biol. Psychiatry*, 2007, 61, 1200-1203.

[28] Numakawa, T.; Yagasaki, Y.; Ishimoto, T.; Okada, T.; Suzuki, T.; Iwata, N.; Ozaki, N.; Taguchi, T.; Tatsumi, M.; Kamijima, K.; Straub, R.E.; Weinberger, D.R.; Kunugi, H.; Hashimoto, R. Evidence of novel neuronal functions of dysbindin, a susceptibility gene for schizophrenia. *Hum. Mol. Genet.*, 2004, 13, 2699-2708.

[29] Chumakov, I.; Blumenfeld, M.; Guerassimenko, O.; Cavarec, L.; Palicio, M.; Abderrahim, H.; Bouguerbier, L.; Barry, C.; Tanaka, H.; La Rosa, P.; Puch, A.; Tahri, N.; Cohen-Akine, A.; Delabrousse, S.; Lissarrague, S.; Picard, F.P.; Maurice, K.; Essioux, Y. Koyabayashi et al. Current Neuropharmacology, 2011, Vol. 9, No. 1
L.; Millasseau, P.; Grel, P.; Debailleul, V.; Simon, A.M.; Caterina, D.; Dufaure, I.; Malekzadeh, K.; Belova, M.; Luan, J.J.; Bouillot, M.; Sambucy, J.L.; Primas, G.; Saumier, M.; Boukkiri, N.; Martin-Saumier, S.; Nasroune, M.; Peixoto, H.; Delaye, A.; Pinchot, V.; Bastucci, M.; Guillou, S.; Chevillon, M.; Sainz-Fuertes, R.; Meguenni, S.; Aurich-Costa, J.; Cherif, D.; Gimalac, A.; Van Duijn, C.; Gauvreau, D.; Ouellette, G.; Fortier, I.; Raelson, J.; Sherbatich, T.; Riazanskaia, N.; Rogaev, E.; Raeymaekers, P.; Aerssens, J.; Konings, F.; Luyten, W.; Macciardi, F.; Sham, P.C.; Straub, R.E.; Weinberger, D.R.; Cohen, N.; Cohen, D. Genetic and physiological data implicating the new human gene G72 and the gene for D-amino acid oxidase in schizophrenia. Proc. Natl. Acad. Sci. USA, 2002, 99, 13675-13680.