Modulation of SIRT1-Foxo1 Signaling axis by Resveratrol: Implications in Skeletal Muscle Aging and Insulin Resistance

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Abstract
Aging individuals and diabetic patients often exhibit concomitant reductions of skeletal muscle mass/strength and insulin sensitivity, suggesting an intimate link between muscle aging and insulin resistance. Foxo1, a member of the FOXO transcription factor family, is an important player in insulin signaling due to its inhibitory role in glucose uptake and utilization in skeletal muscle. Phosphorylation of Foxo1 is thought to mitigate the transactivation of pyruvate dehydrogenase lipoamide kinase 4 (PDK4), which is a negative regulator of the glycolytic enzyme pyruvate dehydrogenase (PDH). In contrast, how aging would regulate acetylation/deacetylation machineries and glucose utilization in skeletal muscle through the Foxo1/PDH axis remains largely undetermined. Accumulating body of evidence have shown that resveratrol, a natural polyphenol in grapes and red wine, activates the longevity-related protein sirtuin 1 (SIRT1) and augments insulin sensitivity in addition to its well-documented effects on mitochondrial energetics. The present review summarizes the role of Foxo1/SIRT1 in insulin signaling in skeletal muscle and proposes the insight that activation of SIRT1 deacetylase activity to deacetylate and suppress the Foxo1-induced transactivation of PDK4 may represent an anti-hyperglycemic mechanism of resveratrol in aging skeletal muscle.
Introduction

Skeletal muscle is an important target for anti-aging/anti-diabetic interventions provided its role in glucose clearance upon insulin stimulation and reductions of muscle mass and insulin sensitivity are not uncommon in older individuals and diabetic patients. In addition to the induction of gene products related to hepatic gluconeogenesis, aberrant changes in expression and/or post-modification of the transcription factor Foxo1 has also been implicated in the disturbance to insulin signaling and glucose uptake and utilization in skeletal muscle. The role of Foxo1 in energy homeostasis is further complicated by its plausible effects on mitochondrial metabolism. Resveratrol is a natural polyphenol found in grapes and red wine known to activate the deacetylase activity of longevity-related protein SIRT1 to enhance mitochondrial metabolism. Although emerging evidence leans support to the notion that resveratrol may orchestrate insulin and metabolic signaling through the SIRT1/Foxo1 axis, whether SIRT1 may also deacetylate and repress the hyperglycemic effects of Foxo1 in the aged skeletal muscle would need to be further investigated. This review summarizes the role of Foxo1-SIRT1 axis in insulin signaling in skeletal muscle and proposes the insight that activation of SIRT1 deacetylase activity to deacetylate and suppress the Foxo1-induced transactivation of PDK4 may represent an anti-hyperglycemic mechanism of resveratrol in the aging skeletal muscle.

Intimate link between skeletal muscle aging and insulin resistance

Muscle aging and diabetes are not mutually exclusive. Skeletal muscle is known as an effector to mediate approximately 65% of glucose clearance in response to insulin stimulation. An earlier study has demonstrated that aging elevated circulating levels of insulin and triacylglycerol and attenuated glucose uptake and utilization in the soleus muscle of obese rats [1], suggesting a relationship that exists between aging and reduced glucose clearance in skeletal muscle. Functional studies in humans have revealed that the handgrip strength was reduced with concomitant increase in homeostasis model assessment (HOMA) insulin resistance index in older, non-diabetic adults [2], thus implying that the aging skeletal muscle may have a pathogenic role in insulin resistance. The HOMA index is first described as a mathematical model predictive of pancreatic β-cell function (i.e., site of insulin production) calculated from basal plasma insulin and glucose concentrations [3]. Importantly, experimental data collected from diabetic patients unraveled that insulin resistance, rather than insulin deficiency, has a predominant role in the development of diabetes [3]. These data hence raise the possibility that skeletal muscle aging may promote the pathogenesis of diabetes whereas conditions of impaired insulin sensitivity may aggravate the functional deteriorations of the aging skeletal muscle. There appears to be an intimate link between skeletal muscle aging and insulin resistance.

The need of effective anti-aging/anti-hyperglycemic regimens in the skeletal muscle

A large amount of resources are dedicated to medical care associated with muscle aging and insulin resistance. The facts that the prevalence of diabetes increases with age and younger adults account largely for the “pre-diabetic” population exhibiting impaired glucose tolerance are worrisome [4]. According to the Centers for Disease Control and Prevention, it is estimated that older individuals would represent approximately 19.6% of the population in the United States by the year 2030 whereas an economic burden of US$218 million was imposed by medical care and loss of productivity related to diabetes in 2007, thereby translating into an annual cost of $700 per person for medical expenses related to diabetes [5]. Provided that diabetic patients manifest reduced muscle mass
and strength measured as appendicular arm lean mass and grip strength, respectively [6]
whereas a 30-day supplementation trial with the natural polyphenol resveratrol improves
the HOMA resistance index and efficiency of energy production machinery in vastus lateralis
muscle of obese men [7], it is conceivable to speculate that regimens which augment skeletal
muscle mass and function in the senior population would be an attractive candidate in the
prevention and treatment of diabetes.

Resistance exercise has been shown to augment muscle mass and strength in aged
women [8] and increase glucose disposal in diabetic patients [9]. These beneficial outcomes,
unfortunately, are often limited by the lack of motivation to exercise in the elderly population
[10]. It is also noteworthy that physical inactivity is an important factor contributing to
reduced muscle mass and increased prevalence of disability in older adults [11]. Nevertheless,
over 50% of diabetic patients regardless of presentation of obesity have sedentary lifestyles
and the likelihood to exercise regularly is reduced with increasing body mass index in non-
diabetic individuals [12]. Caloric restriction, a dietary regimen involving ~30% to 40%
reduction of daily energy intake, has been shown to improve insulin sensitivity in both aged
rats [13] and overweight, non-diabetic adults [14] but the issue of compliance in humans
remains to be studied. Taken together, anti-aging/anti-diabetic interventions which gain high
compliance (i.e., dietary supplementation) while mimicking the anti-hyperglycemic effects
of physical exercise/caloric restriction (e.g., resveratrol) [15, 16] may help pronouncedly the
problems of muscle aging and insulin resistance.

While the molecular mechanisms of insulin signaling cascade and their perturbations
leading to insulin resistant state have been reviewed extensively elsewhere [17, 18], the
present review attempts to discuss the perspective that modulation of the SIRT1/Foxo1-
related insulin signaling axis in the aging skeletal muscle might represent a novel anti-
hyperglycemic mechanism of resveratrol.

Complexity of Foxo1 signaling: crosstalk between insulin and mitochondrial
signaling

Foxo1 is a member of the “Forkhead box” (FOXO) transcription factor family which
has multiple regulatory roles in signaling pathways including, but not limited to oxidative
stress, apoptotic cell death, catabolism and insulin sensitivity. The physiological effects
of Foxo1 originate from the binding of insulin, a hormonal peptide which promotes the
reduction of circulating glucose, to its native insulin receptor (Fig. 1). This binding triggers
the phosphorylation and activation of insulin receptor. The intrinsic tyrosine kinase activity
of insulin receptor then phosphorylates insulin receptor substrate (IRS) to induce the
canonical PI3K-Akt phosphorylation cascade (reviewed by [18-21]). This pathway involves
the activation of the 2 isoforms of 3-phosphoinositide dependent protein kinase (PDK),
namely PDK1 and PDK2 to phosphorylate and activate protein kinase B (Akt) at threonine
308 and serine 473, respectively. In addition, it is worth-noting that Ser473 of Akt can also
be phosphorylated by mammalian target of rapamycin complex 2 (mTORC2) [22]. Recent
efforts have exhibited that phosphorylation of mTORC2 at serine 2481 is essential to the
induction of phospho-Akt(Ser473) by insulin, an effect which was abolished by inhibition of PI3K
[23]. These studies thereby suggest that Akt can be activated through multiple pathways.
Activation of Akt then abrogates the gene transactivation capacity of Foxo1 through the
induction of inhibitory phosphorylation at threonine 24, serine 256 or serine 319, leading to
the retention and degradation of Foxo1 in the cytoplasm (Fig. 1).

There are studies proposing that activation of Foxo1 may have protective effects under
hyperglycemic conditions. Targeted disruption of Foxo1 in liver was found to aggravate
lipid abnormalities measured as triglyceride and cholesterol and up-regulate genes related
to cholesterol synthesis in streptozotocin-induced diabetic mice [24]. Although a 4-hour
transient exposure of hyperglycemia/hyperinsulinemia reduced the transcript levels of both
isoforms of IRS, these reductions were associated with up-regulation of Foxo1 and CD36,
which is a gene involved in fatty acid oxidation in vastus lateralis muscle of healthy, non-obese humans [25]. These studies appear to suggest that Foxo1 may modulate mitochondrial signaling to maintain lipid homeostasis during conditions of insulin resistance. In contrast, an earlier study has demonstrated that prolonged glucose infusion for 5 days blunted the expression and membranous translocation of glucose transporter 4 (GLUT4) in the hind limb muscle of rats [26], hence raises the concern that whether insulin resistance affects pathways of mitochondrial metabolism or mitigates glucose uptake in the skeletal muscle can be largely determined and complicated by the duration of glucose infusion.

Alterations of Foxo1 have been implicated in mitochondrial adaptation associated with aging, insulin resistance and physical exercise. Previous studies have demonstrated that diabetic patients had reduced citrate synthase activity and smaller mitochondria in the vastus lateralis muscle [27] whereas senior, non-diabetic subjects exhibited poor performance in oral glucose tolerance test and reduced whole-body mitochondrial oxidative capacity [28]. Moreover, it has been reported that the mitochondrial function measured as oxygen consumption and adenosine triphosphate (ATP) production was reduced with concomitant mitigations in the protein abundances of Foxo1, PEPCK and G6pase in hepatocytes treated with berberine [29] whereas Foxo1 knockdown attenuated the reduction of mitochondrial DNA copy number induced by various muscle wasting stimuli including serum starvation, dexamethasone and myostatin in cultured C2C12 myotubes [30]. Recent findings have suggested that the phosphorylation of Foxo1 at serine 256 was reduced with up-regulation of peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1α), which is the central metabolism regulator and G6pase, which is an enzyme involved in gluconeogenesis in the liver in a mouse model of hepatitis C infection [31]. It has also been observed in the skeletal muscle that the mRNA levels of Foxo1, pyruvate dehydrogenase lipoamide kinase (PDK4), PGC-1α and aminolevulinate delta synthase 1 (ALAS1, an important protein for proper functioning of the electron transport chain) were elevated in response to acute exercise [32]. However, it remains largely unknown whether the manipulation of Foxo1 by post-translational modification machineries would affect insulin sensitivity and mitochondrial function in the aging skeletal muscle. Transactivation of Foxo1 targets in liver and skeletal muscle are illustrated in Fig. 2.
Foxo1 as a negative regulator of glucose utilization in the skeletal muscle

Nevertheless, an accumulating body of evidence suggests that Foxo1 is an important player during the development of insulin resistance. The concept that Foxo1 has pro-hyperglycemic properties is established based on a series of works that involve manipulations of the endogenous level and activity of Foxo1. Genetic ablation of Foxo1 in hypothalamus is known to enhance glucose uptake in gastrocnemius muscle [33]. Recent efforts have demonstrated that reducing the endogenous level of Foxo1 by knockdown approach abolished the elevation of MAPK phosphatase-3 (MKP-3), a factor known to induce transcriptional activation of gluconeogenic genes including PEPCK and G6pase in the liver, thereby suggesting that hepatic gluconeogenesis may involve orchestration by Foxo1 and PGC-1α. The role of Foxo1 in insulin signaling in skeletal muscle may largely be owing to transactivation of PDK4, an inhibitor of the glycolytic enzyme PDH. More importantly, the activities of Foxo1 and PGC-1α are in part governed by SIRT1-mediated deacetylation (i.e., acetylation activates Foxo1 whereas deacetylation activates PGC-1α). This raises the possibility that SIRT1 may regulate concomitantly pathways associated with insulin resistance and mitochondrial metabolism. Furthermore, the modulating effects of SIRT1 on energy homeostasis in skeletal muscle may require an upstream activating signal mediated through the energy sensor AMPK in response to resveratrol stimulation.
in conditions of impaired insulin sensitivity. One of such effects are mediated by pyruvate dehydrogenase lipoamide kinase isozyme 4 (PDK4), an inhibitor of the glycolytic enzyme pyruvate dehydrogenase (PDH) (Fig. 3). It has been reported that insulin stimulation increased the phosphorylations of Akt at Ser473/Thr308 and Foxo1 at Thr24 and reduced the mRNA content of PDK4 in the gastrocnemius muscle [38]. It is noteworthy that the elevation of Foxo1 phosphorylation was accompanied by increased whole-body glucose uptake and attenuated hepatic glucose production [38]. In contrast, experimental simulation of insulin-resistant states by infusion of intralipid/lactate and insulin in rats reduced the protein contents of phosphorylated Akt$^{\text{Ser307}}$ and Foxo1$^{\text{Ser319}}$ and induced transcriptional activation of PDK4 in the gastrocnemius muscle [39]. Consistent findings have shown that acute administration of the PDH activator dichloroacetate in humans mitigated the elevations of Foxo1 and PDK4 transcripts induced by an occasional high-fat diet in quadriceps muscle and reduced the level of circulating lactate during exercise [40]. These existing data thus lean support to the notion that interventions repressing the expression/activity of Foxo1 may improve glucose utilization in the skeletal muscle and thus, may have preventive/therapeutic values against insulin resistance.

**Activation of SIRT1 by resveratrol: more than enhancing mitochondrial metabolism**

Resveratrol (3,5,4'-hydroxystilbene) is a natural polyphenolic compound commonly found in grapes, berries and red wine and has pleiotropic effects mediated by multiple signaling pathways. The striking finding that resveratrol extends remarkably the lifespan of yeast by 70% through the activation of silent information regulator 2 (sir2) [41] has initiated an array of studies to unravel the beneficial effects of resveratrol on general health. Sirtuin 1 (SIRT1) is known as the mammalian ortholog of sir2 and is one of the most-studied members among the seven isoforms of sirtuins. It has been shown by microarray analysis that resveratrol up-regulated genes related to mitochondrial metabolism in C2C12 myocytes and skeletal muscle of mice challenged with high-fat diet [15]. However, these transcriptional changes were abrogated by treatment with SIRT1-specific shRNA, hence suggesting an indispensable role of SIRT1 [15]. It is thought that the transcriptional effects of resveratrol require the deacetylation mechanism of SIRT1. This postulation is supported by the findings demonstrating that 1) SIRT1 deacetylated and activated the central metabolism regulator PGC-1α and 2) the effects of SIRT1 activation were recapitulated by infection with acetylation-defective PGC-1α mutants regardless of resveratrol treatment [15].

Although the SIRT1/PGC-1α axis is generally accepted as the central signaling dogma of resveratrol-induced augmentation of mitochondrial oxidative capacity [15, 42], a growing body of studies has reported inconsistent findings. It has been previously shown by our group and others that resveratrol improved cardiac function but without affecting the expression of PGC-1α [16, 43]. These observations are in agreement with the findings showing that an 8-week resveratrol intervention had no effects on the protein abundances of PGC-1α and complex IV subunits in triceps muscle [44]. Intriguingly, the protein contents of PGC-1α and citrate synthase activity in gastrocnemius muscle were also reported to be unchanged following 10 months of resveratrol intervention [45]. Nonetheless, the exact explanations for the unaltered mitochondrial metabolic markers are not known. There are also contradictory findings indicating that forced expression of defective SIRT1 indeed enhanced acetylation of PGC-1α and expression of mitochondrial proteins in both C2C12 myotubes and triceps muscle [44]. Additional research is warranted to further understand the mechanisms of these inconsistent changes in response to resveratrol treatment. In addition, these disparities bring forward the possibility that the beneficial effects of SIRT1 activation induced by resveratrol may not be necessarily confined to the induction of mitochondrial metabolism.
Resveratrol may confer anti-hyperglycemic effects through modulation of SIRT1-Foxo1

It is well-documented that resveratrol enhances glucose transport/uptake in skeletal muscle. While resveratrol increases basal glucose uptake in L6 myotubes in a dose-dependent manner [46], the skeletal muscle of diabetic patients exhibits increased expression of SIRT1 and GLUT4 in response to a 12-week daily supplementation with resveratrol [47]. It has also been reported that a 16-month supplementation with resveratrol increased glucose uptake in insulin-stimulated muscles [16] and membrane localization of GLUT4 in the diabetic heart was improved in response to 5 days of resveratrol feeding [48]. However, it remains to be elucidated whether activation of SIRT1 by resveratrol may enhance the expression/membrane localization of GLUT4, resulting in improved glucose clearance in response to insulin-stimulation.

SIRT1 is known to modulate upstream signaling of Foxo1. Reduced contractile function of skeletal muscle has been demonstrated to accompany the down-regulation of SIRT1 and IRS1 [49]. Although further studies are required to dissect the physiological effects of SIRT1-mediated deacetylation of IRS1, phosphorylation of IRS1 at serine-307 is considered as a molecular event that precedes and associates with the development of glucose intolerance [50, 51]. The significance of Akt phosphorylation in insulin sensitivity was further confirmed in studies demonstrating that a 4-week resveratrol supplementation trial reduced postprandial glucose and increased phosphorylation of Akt at serine 473 in the platelet of diabetic patients [52] and muscle biopsies of diabetic patients manifested reduced level of...
phospho-AktSer473, an effect which was mimicked by knocking down of SIRT1 in L6 myoblasts [53]. These data bring forward the idea that SIRT1 would increase phosphorylation of Akt, which is a marker indicative of intact insulin signaling. It appears that this postulation is well-supported by the observation that activation of SIRT1 deacetylase activity by caloric restriction potentiates the phosphorylation of AktSer473/Thr308 induced by insulin in isolated soleus muscles [54]. Taken into consideration that the molecular mechanism by which resveratrol increases phosphorylation of Akt may at least in part, be ascribed to the exposure of Akt phosphorylation sites in response to activating deacetylation by SIRT1 [55], it is conceivable to postulate that the anti-hyperglycemic effects of resveratrol in aging muscle may require the deacetylation mechanism of SIRT1.

Most of our understandings in the role of Foxo1 in skeletal muscle physiology are acquired from experimental models of muscle disuse atrophy. These studies have shown that the capacity of Foxo1 to induce transcriptional activation of atrophic genes can be governed by acetylation/deacetylation. The histone acetyltransferase p300 has been reported to acetylate and increase nuclear localization and transactivating activity of Foxo1 in the skeletal muscle [56]. In contrast, resveratrol was observed to increase the SIRT1 deacetylase activity and alleviate the elevations of protein contents of acetylated Foxo1, atrogin1 and MuRF-1 in dexamethasone-treated L6 myotubes [57]. Although it is well-established that the activity of PDH, a glycolytic enzyme in skeletal muscle would be blunted in response to phosphorylation and inactivation of Foxo1 [38-40], how acetylation/deacetylation mechanisms would modulate the activity of Foxo1, expression of PDK4 and the subsequent glucose utilization in the aging skeletal muscle are poorly understood. It has been reported that the protein content of SIRT1 was increased with concomitant reduction of acetylation status of Foxo1 in the skeletal muscle of diabetic patients following a 3-month intake of epicatechin-containing drink [58]. Recent efforts have also unraveled that resveratrol elevated the protein contents of phospho-Foxo1 and GLUT4 in the adipose tissues of high-fat diet-fed mice [59]. However, it merits further investigation whether these findings could be extrapolated in conjunction with potential involvements of acetylation/deacetylation of Foxo1 in the aging skeletal muscle. Collectively, it is plausible to speculate that activation of PDH and increased membrane trafficking of GLUT4 secondary to SIRT1-mediated deacetylation of Foxo1 may represent an anti-hyperglycemic mechanism of resveratrol (Fig. 3), although the validity of this speculation remains to be confirmed by future research.

**The SIRT1/Foxo1 axis: a pathway plausibly regulated by AMPK**

AMP-activated protein kinase (AMPK) is an energy sensor which activates in response to elevated level of adenosine monophosphate, a marker indicative of energy insufficiency. Previous works have exhibited that AMPK was required to mediate metabolic adaptation. Genetic inactivation of AMPK was observed to prevent the reduction of Foxo1 acetylation and up-regulation of GLUT4 and CPT1b in skeletal muscle of mice after 20 hours of fasting [37]. These findings are in agreement with the observation that acute bouts of swimming exercise failed to diminish the acetylation status of PGC-1α in the skeletal muscle of AMPK-deficient mice [37]. It is also worth-noting that the attenuated acetylations of Foxo1 and PGC-1α induced by fasting/exercise were accompanied by an increase in nicotinamide adenine dinucleotide (NAD), an effect which was abrogated in the absence of AMPK [37]. Given that NAD is a cofactor of SIRT1 deacetylase activity, the modulating effects of resveratrol/SIRT1 on energy homeostasis in skeletal muscle may require activating signal from AMPK. This speculation is supported by a recent study showing that neither resveratrol nor over-expression of SIRT1 enhanced the contents of cellular ATP and mitochondrial DNA in C2C12 cultures treated with AMPK-shRNA, an observation mimicked by pharmacological blockade of SIRT1 with EX527, implying that AMPK may act upstream of SIRT1 [60]. While the mechanisms underlying the finding that a 33-month resveratrol supplementation reduces the HOMA insulin resistance index without affecting body mass remains unclear [61], activation of AMPK by resveratrol
may increase the enzymatic activity of SIRT1 to deacetylate and repress the hyperglycemic effects of Foxo1 and enhance the PGC-1α-related transcription cascade leading to improved mitochondrial metabolism (Fig. 2 & 3). However, it has also been reported that acute exposure to higher concentrations of resveratrol mitigated phosphorylations of AMPK and Akt and glucose uptake in insulin-stimulated human myotubes [62]. Collectively, it merits further research to identify the molecular mechanisms accounting for these dose-related discrepancies and plausible cross-talk between PGC-1α and Foxo1 in the context of orchestrating insulin signalling and mitochondrial metabolism in the skeletal muscle.

Conclusions

Increasing studies have demonstrated that suppression of Foxo1 at both expression and activity levels enhances insulin sensitivity, an effect largely attributable to increased glucose uptake and utilization in the skeletal muscle and reduced gluconeogenesis in the liver. In addition to the enhancement of mitochondrial metabolism through the SIRT1-PGC-1α axis, it is worth-noting that resveratrol, a natural polyphenol in grapes and red wine, also has anti-hyperglycemic effects including stimulation of glucose uptake and prevention of adverse alterations in insulin signaling in skeletal muscle. Despite the fact that more intense research is required before the clinical application of resveratrol as an anti-diabetic intervention, repression of the Foxo1-induced reduction in glucose uptake and utilization by activation of SIRT1 deacetylase may represent a novel, yet promising anti-hyperglycemic mechanism of resveratrol.

Abbreviations

Akt (Protein kinase B); ALAS1 (Aminolevulinate delta synthase); AMPK (AMP-activated protein kinase); ATP (Adenosine triphosphate); CD36 (Cluster of differentiation 36); CPT1b (Carnitine palmitoyltransferase 1b); Foxo1 (Forkhead transcription factor 1); G6pase (Glucose-6-phosphatase complex); GLUT4 (Glucose transporter 4); HOMA (Homeostasis model assessment); MAPK (Mitogen-activated protein kinase); MEK (Mitogen-activated protein kinase kinase); MKP-3 (MAPK phosphatase–3); mTORC2 (Mammalian target of rapamycin complex 2 (mTORC2)); MURF-1 (Muscle ring finger-1); NAD (Nicotinamide adenine dinucleotide); PDH (Pyruvate dehydrogenase); PDK1/2 (3-phosphoinositide dependent protein kinase 1/2 (PDK)); PDK4 (Pyruvate dehydrogenase lipoamide kinase isozyme 4); PEPCK (Phosphoenolpyruvate carboxykinase); PGC-1α (Peroxisome proliferator-activated receptor gamma coactivator 1-alpha); sir2 (Silent information regulator 2); SIRT1 (Sirtuin 1).

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