A Beginners Guide to the Metabolism, Functions and Pharmacological Potential of Sphingolipids

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Abstract

This brief review is designed to acquaint the reader with a basic understanding of the functions of sphingolipids in the newly emerging field of lipidomics. This includes the derived so-called complex sphingolipids found in membranes. Sphingolipids have been implicated as important messenger molecules in many areas of cell function including cell growth, apoptosis, inflammation and migration. Complex sphingolipids have been shown to be involved in vital membrane functions such as receptor clustering and vesicle trafficking. Obviously these functions are significant factors in the progression of many disease states such as the development of cancerous cells and tumorigenesis. Accordingly these lipid messengers and membrane components have attracted much pharmaceutical interest and useful drugs are now starting to be approved and prescribed for patients in the U.S. This review introduces sphingolipid metabolism and function and briefly discusses their exploitation for drug development.

Keywords: Apoptosis; Cancer; Drug targets; Sphingolipids

Introduction

Sphingolipids are a very important and large family of interrelated lipid molecules at the forefront of the newly emerging field of lipidomics [1]. Since the discovery that Diacylglycerol (DAG), a lipid derived from membrane components via the action of phospholipase C, acts as a so-called secondary messenger to activate protein kinase C and the liberation of arachidonic acid by phospholipase A, as a precursor for eicosanoid production [2,3], the idea that lipids were just for energy storage or passive membrane components was redundant. It is now widely accepted that many lipids, especially sphingolipids, are vital in signaling and regulating very important cellular processes including cell growth, apoptosis, inflammation and migration [1]. However, unlike the relatively simple and linear signaling pathway of the ubiquitous secondary messenger cyclic adenosine mono phosphate, there are significant additional layers of complexity that must be considered with sphingolipids. For simplicity in this review the term sphingolipid is used for all of the molecules illustrated in figure 1 (excluding the de novo starting molecules utilized by the transferase and the degradation products produced by the lyase) regardless of whether they have been found to have signaling functions. As mentioned these are a large family of interrelated signaling molecules (Figure 1), some of which can exhibit opposite effects on the same cellular process such as apoptosis. Being interrelated means disruption of one part of the pathway can produce a “ripple effect,” affecting the levels of several of these molecules in turn [1]. Physically, some sphingolipids are very hydrophobic so limiting them to a particular organelle membrane, often where synthesized, or even a specific membrane face. Other sphingolipids are more hydrophilic and can enter the cytoplasm. There can also be differences in the alkyl chain lengths, position and number of double bonds and additional chemical groups (typically hydroxy). There are currently 6 known isoforms of the enzyme Ceramide Synthase (CerS; also known as sphingosine N-acetyltransferase) which is central in sphingolipid metabolism (Figure 1) and these iso forms exhibit preferences for acyl CoAs of different chain lengths (16 vs.18 vs.24 carbons) [4,5]. Sphingolipids are also converted into so-called “complex” sphingolipids by the addition of various “head groups” (such as phosphocholine, glucose or galactose) to become more amphipathic membrane components. In addition ceramide is used in the important group of membrane components known as gangliosides which are sialylated glycol sphingolipids found in cell membranes generally but in particular in the grey matter of the CNS [6,7]. Complex sphingolipids have been implicated in such important functions as endocytosis and receptor clustering and the formation of functional microdomains or so-called “lipid rafts” in cell membranes. Additionally gangliosides are involved in brain development and thought to be involved in synaptic transmission and the modifications implicated in such vital processes as learning and memory [8]. The sphingolipid ceramide itself can be a membrane component and is found in higher than normal levels in these microdomains involved in certain signaling pathways both in plasma and mitochondrial membranes [9]. Complex sphingolipids are themselves recycled and can add to and affect the pool of simpler sphingolipids (Figure 1). Virtually all the published data on sphingolipid function was obtained from research utilizing laboratory animals or cultured cells. A brain sphingolipid ("sphingosin") was actually first described in the 19th century [10] but it was not until 1947 that the collective term sphingolipid was applied [11] and by 1983 many individual sphingolipids had been described [12]. Sphingolipids are defined by a sphingoid base backbone (1,3-di-hydroxy, 2-aminooalkane) typified by sphingosine (Figure 2). Sphingosine is specifically D-erythro-1,3-dihydroxy, 2-aminooctadec-4-ene (also known as trans-4-sphingenine). Sphingolipids can vary in chain length, position and number of double bonds and other functional groups such as a hydroxyl group at position 4. Most mammalian sphingolipids studied so far contain an 18 carbon alkyl chain and therefore the following discussion focuses on these variants but the obvious potential for a vast super family of these molecules varying in
chain length, etc. should not be overlooked. For an extremely detailed and comprehensive review of the structure of sphingolipids the reader is recommended to access the Merrill laboratory web site [13; www.sphingomap.org].

Sphingolipid Metabolism

A detailed review of sphingolipid metabolism is beyond the scope of this review but is covered in-depth in the book “Sphingolipids as Signaling and Regulatory Molecules” [14]. To summarize, de novo sphingolipid synthesis starts with the condensation of the amino acid serine with palmitoyl-CoA to form 3-keto-dihydrosphingosine (also known as 3-ketosphinganine) catalyzed by serine palmitoyl transferase in the endoplasmic reticulum [15] (Figure 1). It has been shown in yeast and mouse fibroblast LM cells that a serine influx can drive de novo synthesis [16,17]. An abnormal subunit in this enzyme is responsible for hereditary sensory Neuropathy Type 1 which was the first human genetic disease associated with sphingolipid metabolism [18,19]. The 3-keto-dihydrosphingosine is then reduced to form dihydro sphingosine (also known as sphingamine) by an NADPH-dependent reductase [20]. This reaction is reversed by addition of an acyl chain of variable length by amide formation via the action of CerS [4,21]. This reaction can be reversed by one of several ceramidases which exhibit organelle-specific expression and, like the sphingomyelinasises, various pH optiums (acid, neutral and alkaline) [22,23]. The resulting molecule is known as dihydroceramide and is normally modified by the addition of a double bond in the 4-5 position by dihydroceramidase desaturase [24] to form ceramide (Figure 2). Ceramide can be considered a common central point or “hub” in sphingolipid metabolism [1] (Figure 1). It was thought initially that dihydroceramide might have specific biological activity but research, especially concerning the activation of protein phosphatase 2A (PP2A), has shown that ceramide itself has far more activity for all targets discovered so far [25]. Ceramide can then be phosphorylated via ceramide kinase after transporation to the高尔gi apparatus [26] to form ceramide-1-phosphate and this reaction can also be reversed by ceramide-1-phosphate phosphatase [27]. Ceramide in particular has important messenger functions that will be discussed later. Ceramide is also the main entry point to complex sphingolipid generation which occurs principally in the golgi apparatus. However, ceramide is very hydrophobic and needs to be transported to the golgi apparatus either via vesicular transport or by binding to the Ceramide Transfer Protein [28] (CERT; figure 3). Once in the Golgi various head groups such as glucose, galactose or phosphocholine can be added to the C1 position via the action of glucosyl ceramide synthase, ceramide galactosyltransferase or sphingomyelin synthase respectively [29,30]. These complex sphingolipids can be recycled via various hydrolases to release ceramide back into the sphingolipid pool [31,32]. Sphingomyelin is broken down first to sphingomyelin synthase respectively [29,30]. These complex sphingolipids

Figure 1: Summary of the metabolic pathways of the most important sphingolipids showing their interrelationships. De novo sphingolipid synthesis starts with serine and palmitoyl CoA while the recycling of “complex” sphingolipids will also affect the levels of these compounds. The two most currently researched intermediates, ceramide and Sphingosine-1-Phosphate (S-1-P), are highlighted with their principal and often antagonistic actions; the so-called sphingolipid “h生物质” can be visualized as the metabolic linking of “complex” sphingolipids (far left) to S-1-P (far right) via ceramide and sphingosine. Single arrows denote an irreversible step while dual arrows indicate a reversible step; C-1-P is ceramide-1-phosphate.

Figure 2: Cartoons comparing the structures of the most important sphingolipids plus the drug fingolimod and its active phosphorylated metabolite fingolimod phosphate.

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Movement of some of the most important sphingolipids within the cell can be understood by the concept of a "rheostat" affecting, in particular, the crucially important cell process of apoptosis; an increase in ceramide with a decrease in S-1-P favoring apoptotic pathways (a "left shift" in figure 1) whereas a decrease in ceramide with an increase in S-1-P favoring anti-apoptotic pathways (a "right shift" in figure 1) [49].

**Sphingolipid Function: Ceramide**

Although many sphingolipids appear to have various and important messenger functions, we will limit our principal discussion here to those best characterized so far and potentially the most important member's ceramide and S-1-P especially as they relate to apoptosis. Apoptosis, also known as programmed cell death, is a normal cellular process whereby old, unnecessary or unhealthy cells can be safely eliminated. It is of particular importance in controlling and preventing the development of cancer cells and tumorigenesis. To briefly summarize, apoptosis can be stimulated via 2 general pathways; the extrinsic and intrinsic pathways. As their names suggest, the extrinsic results from external ligands binding to so-called "death receptors" (FasL or TNF-α) while the intrinsic is triggered either by nuclear or mitochondrial dysfunction. Regardless of the route the final result is the activation of various caspasases and apoptosis. It is thought that ceramide, released from sphingomyelin in lysosomes by the action of acid sphingomyelinase [1,34], can form channels in the outer mitochondrial membrane. This allows the release of pro-apoptotic proteins such as cytochrome c, procaspases, apoptosis inducing factor, heat shock proteins, Smac/Diablo and endonuclease G which are crucial for apoptosis [50]. Acid sphingomyelinase release of ceramide is stimulated by TNFa, CD95 and certain chemotherapeutic agents [51]. Ceramide activates cathepsin D to induce apoptosis via activating Bid [34]. Ceramide's other principal targets include PP1 and PP2A which are involved in regulating apoptosis and cell growth arrest [52,53]. Ceramide also targets the kinase suppressor Rap which helps regulate TNFa-mediated activation of ERK1 and 2 in intestinal epithelial cells [54,55], activation of c-Raf and the Mitogen-Activated Protein Kinase (MAPK) pathway in kidney glomerular mesangial cells [56], and many members of the PKC family [57-59]. Ceramide can bind to and activate Stress-Activated Protein Kinase (SAPK/JNK) and cause kidney glomerular epithelial cells to undergo apoptosis distinct from the previously mentioned effect in mesangial cells via MAPK [60]. Ceramide is also implicated in metabolic syndrome and the development of type 2 diabetes. Excess saturated fatty acid levels and inflammatory cytokines such as TNF-α cause an increase in ceramide which, via PP2A, activates PKB/Akt and lowers insulin sensitivity [61]. Finally it has been shown that both ceramide and S-1-P can cause autophagy. Autophagy is the cellular recycling of organelles and macromolecules principally for energy homeostasis; it is also implicated in certain immune responses, stress, neurodegeneration, aging and cancer [62]. Normally Akt inhibits autophagy via mTOR kinase; if growth conditions become unfavorable this inhibition is removed [63,64]. Tamoxifen, which can induce autophagy, is known to act via sphingolipids [65,66]. Ceramide, a synthetic Ser/Thr protein phosphatase activator, can cause autophagy via beclin-1 upregulation in MCF-1 cells similarly to tamoxifen [66]. Sphingosine-1-phosphate appears to activate autophagy by inhibiting mTOR but independently of AKT [67].

**Sphingosine-1-Phosphate**

Sphingosine-1-phosphate is typically found at very low concentrations within cells, but higher in plasma, and is thought to act principally via extracellular GPCRs [44]. Sphingosine-1-phosphate has been shown to be essential for immune cell migration from lymphoid
Glucosylceramide is transported via the lipid transport protein FAPP2 [97]. Unlike galactosylceramide, glucosylceramide is essential to life in mammals; mice lacking this synthase are embryonic lethal but the condition can be rescued by addition of exogenous glucosylceramide. It is thought that these sphingolipids are essential for correct cell-cell recognition [98]. Mouse knockouts of glucosylceramide synthase confined to the skin or nervous system lead to either a breakdown of the hydrophobic barrier of the skin (excessive water loss followed by death) or death 11-24 days after birth, apparently due to a lack of proper brain development respectively [99,100]. Galactosylceramide is produced by ceramide galactosyltransferase from ceramide and UDP-galactose in the endoplasmic reticulum [101]. Although this enzyme appears to have limited tissue expression (kidneys, testes and intestines) it is found in myelinating glial cells both in the CNS and peripheral nervous system (oligodendrocytes and schwann cells respectively). In the CNS, myelin has a significant galactosylceramide content and knockout mice for ceramide galactosyltransferase display severe motor weakness among other problems such as male infertility [102,103]. Sphingomyelin is the commonest complex sphingolipid found in mammalian cells and is essential for survival [14,104]. There are 3 sphingomyelin synthases found primarily in the golgi apparatus but also in the cell membrane where they may be involved in maintaining sphingolipid content [30,105]. The synthesis of sphingomyelin from ceramide and phosphatidylcholine also produces DAG which has opposite effects to ceramide on proliferation and cell survival [106]. Vitamin D stimulates sphingomyelinase activity and results in an increase in ceramide levels due to sphingomyelin breakdown [107]. Apart from the generation of signaling molecules, complex sphingolipids are necessary for maintaining membrane structure and certain functional microdomains [108,109]. They are also binding sites for various proteins as well as certain bacterial and viral pathogens and toxins [110,111]. Some sphingolipids are actually covalently attached to membrane proteins [112]. Importantly it has been shown that depletion of complex sphingolipids from plasma membranes due to fumonisin-mediated disruption of sphingolipid metabolism (a fungal toxin that inhibits CerS) affects endocytosis of folate via human folate receptor α. Folate is required for correct development of the CNS and a deficiency in animals and humans causes an increase in Neural Tube Defects, (NTDs). Fumonisins have been shown to cause this in animal models and are strongly implicated in increased incidences of NTDs in humans where fumonisin-contaminated foods have been consumed [113].

Pharmacological Targeting

Sphingolipids are involved or implicated in many vital cell processes and pathologies. The simple rheostat model suggests that a shift in the intracellular sphingolipid profile towards ceramide versus S-1-P will favor apoptosis (Figure 1) and therefore the possible removal of cancer cells. Inhibition of SphKs should decrease the generation of S-1-P and were therefore an obvious early target of research but even though decreases in S-1-P were often seen, sometimes with a corresponding increase in ceramide, the results were mixed [114-118]. The specific SphK2 inhibitor ABC294640 did inhibit several cancerous cell types in culture and decrease S-1-P levels in mice but also has demonstrated anti-estrogenic effects [118,119]. Neutralizing of S-1-P via monoclonal antibodies looked promising with murine models showing a decrease in lung cancer metastasis but a human version has just recently failed a phase III trial for the treatment of renal cancer (Sonepcizumab) [120,121]; http://adisinsight.springer.com/drugs/800024045). However this drug is still being considered for the
treatment of wet age-related macular degeneration such as the prevalence of sphingolipid signaling. Research is also underway to develop drugs that activate S-1-P lyase as a means to decrease S-1-P [122]. Significant features of many cancers are their resistance to death receptor ligands and mitochondrial-induced apoptosis. Cancer cells typically have disrupted apoptotic pathways with an imbalance of pro-versus anti-apoptotic proteins [5]. Low levels of ceramide are observed in certain breast, colon and kidney cancers [123-126]. Elevation of ceramide levels should favor receptor clustering and therefore apoptosis [127]. Additionally, ceramide-enriched “platforms” can allow cell membrane invaginations sufficient to interact with mitochondria and trigger apoptosis in a blurring of the extrinsic and intrinsic pathways (the so-called “kiss of death”) [128,129]. Ceramide has the additional problem of being severely hydrophobic so drug delivery is a factor that needs to be considered if trying to directly increase ceramide levels by supplementation [130]. In the light of often contradictory results it is now becoming fairly obvious that a more refined rheostat model needs to be developed which could take into account such co-founders as subcellular location and differing alkyl chain lengths, etc. in ceramide and its derivatives (see CerS previously) [131]. It may be that a pharmacogenetic profile will need to be taken of the patient’s sphingolipid-metabolizing enzymes to be fitted to a model to predict the desired, patient-specific manipulation, possibly requiring multiple pharmacological agents. Finally the rheostat model can be by-passed by targeting the actual receptors involved. In the case of S-1-P, these are the S1PRs activated by extracellular S-1-P, especially S1PR1 whose activation is involved in several cancers. This particular avenue of research is now producing second generation drugs. One of the most promising has been fingolimod which is a sphingosine analog that is phosphorylated into an S-1-P analog by SphK2 and acts as an agonist on all the S1PR’s except S1PR2 [132,133] (Figure 2). Although an agonist it actually reduces the effect of S1PR1 activation by triggering receptor internalization [132,134], a form of cellular adaptation. Additionally fingolimod itself can induce apoptosis via the dephosphorylation of oncogenic proteins by PP2A [135-138]. However, fingolimod was found not to be particularly useful in combatting cancer but in treating Multiple Sclerosis (MS) where its inhibition of immune cell movement into the CNS was efficacious [69]. This drug has been approved by the FDA for the treatment of MS for several years now and is reviewed in far greater depth in the 2013 Nature Reviews of Drug Discovery (volume 12, number 9) which lists many sphingomimetics reviewed in far greater depth in the 2013 Nature Reviews of Drug Discovery (volume 12, number 9) which lists many sphingomimetics.

Conflict of Interest
The author has no conflicts of interest nor has published or has being considered for publication any part of this manuscript elsewhere.

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