Oxalate oxidase (OxO), a manganese dependent enzyme, is involved in the catalysis of oxalate oxidation to carbon dioxide with the formation of hydrogen peroxide. OxO is present in the cell wall of plants. It increases the resistivity against diseases and external stress. Oxalate is found as waste metabolites in mammals, excess accumulation of oxalate results in hyperoxaluria and urolithiasis in humans. These disorders can be diagnosed by the help of OxO in many analytical methods. The present study is aimed on isolation of OxO enzyme from Pseudomonas strain under high salt stress. The OxO enzyme was produced in bulk under selected salt stress. It was partially purified by ammonium sulphate precipitation method, dialysis and ion-exchange chromatography. The OxO enzyme on enzymatic analysis showed potent activity concerning oxalic acid substrate. The microtiter plate analysis confirmed the ability of OxO enzyme purified from Pseudomonas strain in the diagnosis of oxalate-related disorders and in the future could be used to prepare the diagnostic biosensors.

**Keywords:** Oxalate oxidase, Salt stress, Pseudomonas strain, Peroxidase

**INTRODUCTION**

The daily consumption of a large amount of oxalate causes Oxalosis or the formation of calcium oxalate deposits in vital tissues or organs of the body, which could be fatal to humans (Sanz & Reig, 1992). Oxalate is observed as a crucial part of stones found in kidney disorders. Thus, the determination of oxalate content in urine is essential to patients with kidney stone problems. The oxalate determination can be done through clinical analysis of blood and urine using the OxO enzyme. Thus, the OxO became an essential candidate for researchers for the diagnosis of kidney stones (Pundir et al., 1993). The OxO enzyme mechanism involves the oxidation of oxalate to carbon dioxide along with hydrogen peroxide. The OxO is dependent on manganese ions. Researchers isolated the OxO enzyme from fungi, bacteria, and plants (Carrillo et al., 2004; Zhou et al., 1998). This enzyme produces two moles of carbon dioxide and one mole of hydrogen peroxide from one mole of oxalate and oxygen. Oxalate oxidase from wheat and barley is known as germin-like oxalate oxidases found in the cell walls of embryos during germination (Graz et al., 2016; Lane, 1994). The Basidiomycete Abortiporus biennis strain produces oxalate oxidase in the mycelium (Koyama, 1988). Researchers proposed the purification of membrane-bound oxalate oxidase from Pseudomonas sp. OX-53. Microbes are known for their potential contribution as a useful source of enzymes (Vázquez et al., 2019). Microorganisms with high yield, easy culturing, and isolation strategies proves low-cost source in many industrial applications (Chiriboga, 1966; Datta & Meuose, 1955; Dumas et al., 1993; Dunwell, 1998; Hu et al., 2015). Researchers used the OxO enzyme isolated from these sources in many diagnostic and clinical treatments for Urolithiasis (Chauhan et al., 2012). Conventional analytical techniques such as Gas and liquid chromatography were studied for OxO analysis, but they have chances of sample degradation (Fiorito & Córdoba de Torres, 2004; Harris et al., 2004). In the era of rapid and robust screening of samples in clinical laboratories, sensitivity is most important. Thus, it is the need of the hour to prepare the biosensors based on OxO for reliable detection of oxalate associated disorders (Yiribelli & Posen, 1980). Pseudomonas sp. isolated from soil under high-stress conditions.

**MATERIAL AND METHODS**

Pseudomonas Agar for fluorescein production (PAF), Pseudomonas Agar (PMP) for the detection of pyocyanin production, Normal saline solution, NaCl, Potassium oxalate (PO), Citrimide agar, Simmons Citrate, Methyl Red medium, Voges-Proskauer medium, Glycerol, Yeast extract, Peptone, Dipotassium phosphate, Magnesium sulphate, Oxalic acid, Potassium oxalate, Ammonium sulphate, Tris-Cl buffer, Protein ladder, 3-methyl-2-benzothiazolinone hydrazone (MBTH), N, N-dimethylamine (DMA), Ethylenediaminetetraacetic acid (EDTA). All chemicals were of analytical grade and purchased from Himedia, India, and standard oxalate oxidase enzyme from BioVision.
Identification of Pseudomonas culture by VITEK 2 system (software version VT2-08.01)

Isolated bacterial culture was inoculated on a nutrient medium and incubated at 37°C for 24 hrs. Suspensions were prepared by emulsifying bacterial isolates in 0.45% saline to the equivalent of a 0.5 McFarland turbidity standard, 1.5 × 10^n CFU/ml. The same suspension was used for identification on the VITEK 2 system. The 0.5 McFarland bacterial suspension was diluted; cards were automatically filled, sealed, and loaded into the VITEK 2 instrument for incubation and reading (García-Garrote et al., 2000).

Screening of medium for salt stress oxalic acid and potassium oxalate for isolated Pseudomonas strain

The growth medium was prepared in 250 ml flask composed of 1% glycerol, 1% yeast extract, 1% peptone, 0.1% dipotassium phosphate, 0.05% magnesium sulphate with 1% oxalic acid, and 2% potassium oxide as a variant for the screening of suitable substrate for bacterial growth to produce oxalate enzyme under stress condition.

Process optimization for maximum bacterial biomass production by a various dose of potassium oxalate salt

The bacterial broth (PAF) was prepared in which content increase the concentration of potassium oxalate ranges from 1 to 12% were added, sterilized by autoclaving at 1.5 lbs pressure (121°C) for 15 minutes. The isolated culture from 1.0% PO (Potassium oxalate) medium was inoculated in each flask and kept for observation for 24 hr at 37°C. 200 µl sample was added in 96 well plates, and the absorbance was noted at 625 nm using Bioteck H1-SYNERGY ELISA spectrophotometer.

Bulk production in suspension medium and harvesting of cells

Pseudomonas aeruginosa was grown in PAF medium broth (500 ml) with 2% Potassium oxalate supplemented in 1000 ml flask for 3-4 days at 37°C.

Characterization of Oxalate oxidase (OxO) enzyme

Ammonium sulphate fractionation and dialysis

Ammonium sulphate fractionation was performed in two stages, at an initial concentration of 0-60% of ammonium sulphate was added slowly until it completely dissolved in cold condition at 4°C for 5 hrs and later it was kept on stirring overnight at 4°C. After the precipitation process was completed, it was centrifuged at 4000 rpm, and the supernatant was collected and used for the second stage of the 80% ammonium sulphate fractionation process. The 60% pellet was resuspended in buffer (Tris-Cl pH 8) subjected to dialysis using a dialysis sack mwco (110) Hi-Media dialysis membrane. The dialysis sack was attached to a flat at one end and immersed in 100 times the volume of the sample and dialyzed against appropriate buffer (PBS) for 24 hr at 4°C with continuous stirring, resuspended in buffer and stored at 4°C. The appropriate dialysis buffer was determined by the subsequent purification procedure to be undertaken. After completion of 60% and 80% dialysis samples were stored for further analysis and assay purposes (Hu & Guo, 2009).

Protein estimation and SDS gel electrophoresis after partial purification of oxalate oxidase from oxalate degrading bacteria

The unknown protein quantification was done by Biuret method, and SDS-PAGE was performed on 12% separating and 5% stacking polyacrylamide gels at 25°C using TARSON Mini Vertical Electrophoresis Assembly (7080). The gels loaded with the sample were run at 25/50/100 V till the dye front reached the end of separating gel. The protein bands were visualized by staining with Coomassie brilliant blue (CBB) G-250 (Laemmli, 1970).

Ion Exchange Column Chromatography (Fractionations) of Partial Purified Enzyme

Column chromatography with DEAE-cellulose chromatography Macro-Prep®/DEAE medium was used, which is an anion exchange medium. It was purchased from BioRad laboratories. The glass chromatography column was packed up to 15 cm height 50% slurry of the Macro-Prep®/DEAE medium. It was equilibrated with 20 mM tris CL buffer of pH 8 for the column chromatography process. Gradients of 50 ml were prepared to range between 200 to 800 mM NaCl in 20 mM tris-CL buffer of pH 8. Further, 50 ml of 20 mM tris CL buffer column was used to wash the column. Next, a 60% Ammonium sulphate precipitated sample (partially purified enzyme) was dialedyzed, and it was loaded on the column. The column was washed with the same buffer to remove the unbound proteins, and the enzyme was eluted by applying a linear gradient of NaCl from 200 to 800 mM, and fractions of 2 ml were collected. Active fractions were pooled and stored for further analysis (Kotsira & Clonis, 1997).

Bacterial oxalate oxidase enzyme activity

MBTH (3-methyl benzothiazoline hydrazine) used to detect release of hydrogen peroxide in the enzymatic reaction of Barley oxidase enzyme up to 1mM. In the assay of oxalate oxidase, hydrogen peroxide is produced which is coupled with HRP which catalyses the MBTH-DMA (Kanauchi et al., 2009; Goodwin et al., 2017).

Oxalate + O₂ → 2CO₂ + H₂O₂ (catalyzed by oxalate oxidase)

MBTH + H₂O₂ + DMA → indamine dye (purple color) + 2H₂O (catalyzed by added peroxidases)

Substrate concentrations were prepared as 10-50 µL of 200 mM oxalic acid in 20 mM tris CI buffer pH 8. About 200 µL of crude bacterial enzyme, 60%, and 80% fractionated dialysis protein sample, and 50 µL substrate was mixed in 24 well plate and kept in incubation for 37°C for 30 min. After completion of incubation time, 300 µL of MBTH solution (MBTH solution was prepared in 0.2 M sodium acetate buffer (pH 4.0) with 0.79 mM DMA and 0.11 mM MBTH) was added along with 100 µL of 100 mM EDTA solution and incubated for 5 min. After that, 20 µL of horseradish peroxidase enzyme solution (HRP) was added (1 mg/mL, Himedia, TC487). HRP enzyme solution (Immediately before use, prepare a solution containing 1 mg/ml of Peroxidase, in cold deionized water). The plate was incubated for 1hr in dark condition. The reading was noted on SYNERGY H1 microplate reader Bioteck at 600 nm (Kanauchi et al., 2009; Goodwin et al., 2017).

RESULTS AND DISCUSSION

Biochemical characterization

Biochemical characterization tests were performed for isolated bacterial samples for the identification of Pseudomonas species. The test results are shown in Table 1. Figure 1A. Showed that no fluorescence on plates as compared to Figure 1B and Figure 1C. Yellow pigment like colonies on PAF plates in visible light conditions was observed, and under UV transilluminator (Figure 1B), it showed greenish fluorescence (Figure 1C). The Oxidase test was found to be positive for the isolated sample, as shown in Figure 2B (bacterial sample without PO) and Figure 2C (bacterial sample with PO) compared to Figure 2A which was a negative control broth. Catalase test was also found to be positive, as a bacterial broth sample reacted to H₂O₂ to produce effervescence shown in Figure 3B with reference to negative control Figure 3A.

Table 1 Biochemical characterization of isolated bacterial samples

| Sr. no | Biochemical test | Observation | Inference |
|-------|-----------------|-------------|-----------|
| 1     | Citrimide agar test | Fluorescence in UV light | +ve |
| 2     | Citrate test | Green to blue colour change | +ve |
| 3     | Oxidase test | Dark blue-purple colour change within 10-30 sec. | +ve |
| 4     | Catalase test | Bubble formation | +ve |
| 5     | Pigment formation | Yellow-green colour | +ve |
| 6     | Methyl Red test | No colour change | -ve |
| 7     | Voges Proskauer test | No colour change | -ve |
Identification of oxidase producing Bacterial culture by VITEK 2 system (software version VT2-08.01)

The isolated colony was observed used for preparing a bacterial suspension of 0.5 McFarland. That sample was used for identification, and results were obtained after 6.78 hrs. The given Gram-negative bacteria *Pseudomonas aeruginosa* was identified using the VITEK 2 system.

Screening of medium for salt stress oxalic acid and potassium oxalate for isolated *Pseudomonas* strain

The increasing bacterial growth was observed in 3% Potassium oxalate medium compared to 2% oxalic acid medium. The Potassium oxalate supplemented medium showed increased growth shown in Table 2 and Figure 4.

### Table 2 Effect on bacterial growth supplemented with oxalic acid and potassium oxalate in the growth medium

| Sr. No | Type of sample | Mean Absorbance (O.D.) |
|--------|----------------|------------------------|
| 1      | Control        | 0.062                  |
| 2      | Oxalic acid 2% | 0.258                  |
| 3      | Potassium oxalate 3% | 2.200               |

**Figure 4** Comparative bacterial growth under oxalic acid and Potassium oxalate supplemented medium

Process Optimization for Maximum Bacterial Biomass Production by different Concentration of Potassium Oxalate Salt

The cultured bacteria showed growth in salt stress range between 1 to 12% PO containing medium. The flask also showed bubbles formation, which are the indicators for the presence of oxidase enzyme. The results are shown in Table 3 and Figure 5. It is clear that 3% of PO concentration have maximum bacterial growth. Hence 3% concentration of PO is selected in growth medium or is found to be more suitable for bulk production of oxalate enzyme.

### Table 3 Effect on the growth of bacteria supplemented with different concentration of potassium oxalate (salt stress)

| Concentration of Potassium oxalate (%) | Mean absorbance | Bubble formation |
|---------------------------------------|----------------|-----------------|
| Control                               | 0.062          | -               |
| 1                                     | 0.22           | -               |
| 2                                     | 0.464          | +++++           |
| 3                                     | 0.934          | ++++++++        |
| 4                                     | 0.492          | +++             |
| 5                                     | 0.298          | ++              |
| 6                                     | 0.251          | ++              |
| 7                                     | 0.248          | ++              |
| 8                                     | 0.131          | ++              |
| 9                                     | 0.128          | +               |
| 10                                    | 0.125          | +               |
| 11                                    | 0.125          | +               |
| 12                                    | 0.120          | +               |
Bacterial oxidase enzyme was subjected to total protein content evaluation. It showed that protein content of crude bacterial oxidase enzyme is 0.086 mg/ml, 60% fractionated dialysis sample was 0.216 mg/ml and 80% fractionated dialysis sample was found to be 0.005 mg/ml. It was deduced by the BSA calibration graph (Figure 6 and Table 4). The BSA gel electrophoresis was performed for the crude enzyme, 60% and 80% dialyzed sample. The 60% dialyzed sample showed more prominent bands than crude and 80% dialyzed sample (Figure 4). The enzyme oxalate oxidase from Pseudomonas aeruginosa is observed to be the molecular weight between 45 Da to 95 Da in all the three samples (Sambrook & Russell, 2001; Steinberg, 2009).

Protein estimation and SDS gel electrophoresis after partial purification of oxalate oxidase from oxalate degrading bacteria

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Bacterial oxidase enzyme activity

In the reaction, oxalate oxidase catalyzes the production of hydrogen peroxide from oxalate; the hydrogen peroxide further reacts with MBTH and DMA in the presence of peroxidase enzyme to form a purple color product (Indamine dye). The color of indamine dye was read at 600 nm using SYNERGY H1 microtiter plate reader from Biotek®. The results indicate the increase in absorbance as the indamine concentration increases, which can be correlated with the activity of the oxalate oxidase enzyme. The standard linear curve was prepared using standard hydrogen peroxide. The activity can be defined as the amount of oxalate unit required to produce the 1 μmol of hydrogen peroxide at 55°C per minute. The H₂O₂ detection was performed using MBTH indamines dye measured using Result layout colorimetric scheme of MBTH based standard H₂O₂ detection using HRP of reaction samples shown in Figure 8. The activity was shown in Figure 9, it shows that a 60% dialyzed sample has more significant H₂O₂ release in the reaction compared to the standard OxO enzyme, 80% dialyzed sample, and crude enzyme (Goodwin et al., 2017; Goyal et al., 1999; Zhang et al., 1996). Some of the difficulties faced during study include the detection of H₂O₂ becomes unstable at higher temperature, thus the study should be done at lower temperatures.
Chauhan, N., Narang, J., Shweta, & Pundir, C. S. (2012). Immobilization of barley oxalate oxidase onto gold–nanoparticle-porous CaCO3 microsphere hybrid for amperometric determination of oxalate in biological materials. Clinical Biochemistry, 45(3), 253–258. https://doi.org/10.1016/j.clinbiochem.2011.11.004
Chiriboga, J. (1966). Purification and properties of oxalic acid oxidase. Archives of Biochemistry and Biophysics, 116, 516–523. https://doi.org/10.1016/0003-9861(66)90057-9
Dashek, V. W. (2018). Methods in Plant Biochemistry and Molecular Biology. CRC Press. https://doi.org/10.1201/9781351074483
Datta, P. K., & Meeruse, B. J. (1985). Microbial oxalic acid oxidase—a flavoprotein. Biochimica et Biophysica Acta, 17, 602–603. https://doi.org/10.1016/0005-276X(85)90142-6
Dumas, B., Saillard, A., Chevet, J. P., Freyssinet, G., & Pallett, K. (1993). Identification of barley oxalate oxidase as a germin-like protein. Comptes Rendus de l’Academie Des Sciences. Serie III, Sciences de La Vie, 316(8), 793–798. PMID: 8044704
Dunwell, J. M. (1998). Cupins: A New Superfamily of Functionally Diverse Proteins that Include Germins and Plant Storage Proteins. Biotechnology and Genetic Engineering Reviews, 15(1), 1–32. https://doi.org/10.1007/978-1-4471-0914-8_1
Dorito, P. A., & Córdoba de Torres, S. I. (2004). Optimized multilayer oxalate biosensor. Talanta, 62(3), 649–654. https://doi.org/10.1016/talanta.2003.09.010
Garcia-Argote, F., Cercenado, E., & Bouza, E. (2000). Evaluation of a New System, VITEK 2, for Identification and Antimicrobial Susceptibility Testing of Enterococci. Journal of Clinical Microbiology, 38(6), 2108–2111. https://doi.org/10.1128/JCM.38.6.2108-2111.2000
Goodwin, J. M., Rana, H., Ndungu, J., Chakrabarti, G., & Moomaw, E. (2017). Hydrogen peroxide inhibition of bicupin oxalate oxidase. PLOS ONE, 12(5), e0171764. https://doi.org/10.1371/journal.pone.0171764
Goyal, L., Thakur, M., & Pundir, C. S. (1999). Purification and properties of a membrane bound oxalate oxidase from Amaranthus leaves. Plant Science, 142(1), 21–28. https://doi.org/10.1016/S0168-9452(98)00251-9
Graz, M., Rachwał, K., Zan, R., & Jarosz-Wilkolazka, A. (2016). Oxalic acid degradation by a novel fungal oxalate oxidase from Abietorpus biennius. Acta Biochimica Polonica, 63(1), 595–600. https://doi.org/10.18388/abp.2016_1282
Harris, A. H., Freel, R. W., & Hatch, M. (2004). Serum oxalate in human beings and rats as determined with the use of ion chromatography. The Journal of Laboratory and Clinical Medicine, 144(1), 45–52. https://doi.org/10.1016/j.lab.2004.04.008
Hu, Y., & Guo, Z. (2009). Purification and characterization of oxalate oxidase from Arthrobacter. Acta Physiologica Plantarum, 31(2), 229–235. https://doi.org/10.1007/s11738-008-0227-y
Hu, Y., Xiang, M., Jin, C., & Chen, Y. (2015). Characteristics and heterologous expressions of oxalate degrading enzymes “oxalate oxidases” and their applications on immobilization, oxalate detection, and medical usage potential. Journal of Biotech Research, 6, 63–75.
Islam, F., & Roy, N. (2018). Screening, purification and characterization of cellulase from cellulase producing bacteria in molasses. BMC Research Notes, 11(1), 445. https://doi.org/10.1186/s13104-018-3558-4
Sambo, J. F., & Russell, D. (2001). Molecular Cloning: A Laboratory Manual (3-Volume Set). In Cold Springs Harbour Press (Vol. 1).
Kanauchi, M., Milot, J., & Bamforth, W. (2009). Oxalate and Oxalate Oxidase in Malt. Journal of the Institute of Brewing, 115(3), 232–237. https://doi.org/10.1004/jib.2010.0541.2009.00374.x
King, E. O., Ward, M. K., & Raney, D. E. (1954). Two simple media for the demonstration of pyocyanin and fluorescein. The Journal of Laboratory and Clinical Medicine, 44(2), 301–307. PMID: 13184240
Kotsira, V. Ph., & Clonis, Y. D. (1997). Oxalate Oxidase from Barley Roots: Purification to Homogeneity and some Properties. Plant Science, 123, 629–638. https://doi.org/10.1016/S0168-9452(96)00529-0
Koyama, H. (1988). Purification and Characterization of Oxalate Oxidase from Pseudomonas sp. OX-53. Agricultural and Biological Chemistry, 52(3), 743–748. https://doi.org/10.1271/bbb1961.52.743
Laemmli, U. K. (1970). Cleavage of Structural Proteins during the Assembly of the Head of Bacteriophage T4. Nature, 227(5259), 680–685. https://doi.org/10.1038/227680a0
Lam, D. L. (1994). Oxalate oxidase. In R. G. Mitarai (Ed.), Immobilization technology, immobilized enzymes and cells (pp. 173–269). CRC Press.
Lane, B. G. (1994). Oxalate, germin, and the extracellular matrix of higher plants. The FASEB Journal, 8(3), 294–301. https://doi.org/10.1096/fasebj.8.3.8143935
MacGregor, A. W. (1977). ISOLATION, PURIFICATION AND ELECTROPHORETIC PROPERTIES OF AN α-AMYLASE FROM MALTED BARLEY. Journal of the Institute of Brewing, 83(2), 100–103. https://doi.org/10.1186/abbs.1997.9898
Neut, D., Hendriks, J. G. E., Horn, J. R. van, Mei, H. C. van der, & Busscher, H. J. (2005). Pseudomonas aeruginosa biofilm formation and slime excretion on antibiotic-loaded bone cement. Acta Orthopaedica, 76(1), 109–114. https://doi.org/10.1080/00016470510030427
Palanivelu, P. (2018). Analytical Biochemistry and Separation Techniques - A Laboratory Manual for B.S., M. Sc., & M. Phil. Students- VI Edition.
Pundir, C. S., Kuchhal, N. K., & Satyapal, null. (1993). Barley oxalate oxidase immobilized on zirconia-coated alkylamine glass using glutaraldehyde. *Indian Journal of Biochemistry & Biophysics, 30*(1), 54–57.

Sanz, P., & Reig, R. (1992). Clinical and pathological findings in fatal plant oxalosis. A review. *The American Journal of Forensic Medicine and Pathology, 13*(4), 342–345. [https://doi.org/10.1097/00000433-199212000-00016](https://doi.org/10.1097/00000433-199212000-00016)

Steinberg, T. H. (2009). Chapter 31 Protein Gel Staining Methods: An Introduction and Overview. In R. R. Burgess & M. P. Deutscher (Eds.), *Methods in Enzymology* (Vol. 463, pp. 541–563). Academic Press. [https://doi.org/10.1016/S0076-6879(09)63031-7](https://doi.org/10.1016/S0076-6879(09)63031-7)

Vázquez, M. A., Cabrera, E. C. V., Aceves, M. A., & Mallol, J. L. F. (2019). Cellulolytic and ligninolytic potential of new strains of fungi for the conversion of fibrous substrates. *Biotechnology Research and Innovation, 3*(1), 177–186. [https://doi.org/10.1016/j.biori.2018.11.001](https://doi.org/10.1016/j.biori.2018.11.001)

Yriberrí, J., & Posen, S. (1980). A semi-automatic enzymic method for estimating urinary oxalate. *Clinical Chemistry, 26*(7), 881–884. [https://doi.org/10.1093/clinchem/26.7.881](https://doi.org/10.1093/clinchem/26.7.881)

Zhang, Z., Yang, J., Collinge, D. B., & Thordal-Christensen, H. (1996). Ethanol increases sensitivity of oxalate oxidase assays and facilitates direct activity staining in SDS gels. *Plant Molecular Biology Reporter, 14*(3), 266–272. [https://doi.org/10.1007/BF02671662](https://doi.org/10.1007/BF02671662)

Zhou, F., Zhang, Z., Gregersen, P. L., Mikkelsen, J. D., Neergaard, E. de, Collinge, D. B., & Thordal-Christensen, H. (1998). Molecular Characterization of the Oxalate Oxidase Involved in the Response of Barley to the Powdery Mildew Fungus. *Plant Physiology, 117*(1), 33–41. [https://doi.org/10.1104/pp.117.1.33](https://doi.org/10.1104/pp.117.1.33)