Proapoptotic BIM Impacts B Lymphoid Homeostasis by Limiting the Survival of Mature B Cells in a Cell-Autonomous Manner

Rui Liu¹, Ashleigh King¹,², Philippe Bouillet³,⁴, David M. Tarlinton⁵, Andreas Strasser³,⁴ and Jörg Heierhorst¹,²*

¹Molecular Genetics Unit, St. Vincent’s Institute of Medical Research, Fitzroy, VIC, Australia, ²Department of Medicine, St.Vincent’s Health, The University of Melbourne, Fitzroy, VIC, Australia, ³Molecular Genetics of Cancer Division, Walter and Eliza Hall Institute of Medical Research, Parkville, VIC, Australia, ⁴Department of Medical Biology, The University of Melbourne, Parkville, VIC, Australia, ⁵Department of Immunology and Pathology, Monash University, Melbourne, VIC, Australia

The proapoptotic BH3-only protein BIM (Bcl2l11) plays key roles in the maintenance of multiple hematopoietic cell types. In mice, germline knockout or conditional pan-hematopoietic deletion of Bim results in marked splenomegaly and significantly increased numbers of B cells. However, it has remained unclear whether these abnormalities reflect the loss of cell-intrinsic functions of BIM within the B lymphoid lineage and, if so, which stages in the lifecycle of B cells are most impacted by the loss of BIM. Here, we show that B lymphoid-specific conditional deletion of Bim during early development (i.e., in pro-B cells using Mb1-Cre) or during the final differentiation steps (i.e., in transitional B cells using Cd23-Cre) led to a similar >2-fold expansion of the mature follicular B cell pool. Notably, while the expansion of mature B cells was quantitatively similar in conditional and germline Bim-deficient mice, the splenomegaly was significantly attenuated after B lymphoid-specific compared to global Bim deletion. In vitro, conditional loss of Bim substantially increased the survival of mature B cells that were refractory to activation by lipopolysaccharide. Finally, we also found that conditional deletion of just one Bim allele by Mb1-Cre dramatically accelerated the development of Myc-driven B cell lymphoma, in a manner that was comparable to the effect of germline Bim heterozygosity. These data indicate that, under physiological conditions, BIM regulates B cell homeostasis predominantly by limiting the life span of non-activated mature B cells, and that it can have additional effects on developing B cells under pathological conditions.

Keywords: apoptosis, cell death, BCL-2 family, BIM, BH3-only protein, B lymphocytes, B cell lymphoma

INTRODUCTION

Members of the BCL-2 family of proteins play crucial roles in the regulation of the mitochondrial apoptotic cell death pathway (1). The BH3-only protein BIM, a proapoptotic member of this family, is particularly critical for regulating cell death/survival in the hematopoietic system. Germline knockout (KO) of Bim causes an increase in the number of cells in several hematopoietic subsets, including B cells, T cells, monocytes, and granulocytes, with a marked splenomegaly (2). In these mice, mature B cells are approximately doubled in number compared to wild-type (WT) controls. Upon in vivo antigen stimulation, B cells can differentiate into antibody-secreting plasma cells, which are also
greatly increased in number in *Bim* KO mice (2). This increase in plasma cells combined with defects in negative selection of autoreactive B cells (3) is thought to lead to the development of a severe auto-antibody-driven systemic lupus erythematosus-like autoimmune pathology with immune-complex glomerulonephritis on a mixed 129Sv/JxC57BL/6 genetic background (2). These symptoms of autoimmunity, however, are significantly moderated on a C57BL/6 background (4). *Bim* KO mice on an inbred C57BL/6 background show an abnormally increased proportion of low-affinity B cell receptor (BCR)/surface-IgM expressing B cells in the germinal center, and they accumulate low-affinity memory B cells (5). Both of these cell populations would normally be eliminated by apoptotic cell death during selection for B cells with improved affinity for antigen arising from somatic mutation of their *IgV* genes (5). Conversely, loss of *BIM* specifically increases the survival of autoreactive immature B cells in the bone marrow, which demonstrated that BIM plays a key role in apoptosis activation by autoreactive BCRs during this developmental stage (3).

*BIM* expression levels increase progressively during B cell development (pre-pro-B < pro-B/pre-B < immature B < mature B) (6, 7), which may explain why loss of *Bim* has such profound effects on immature and mature B cell populations. However, loss of *Bim* can also increase cell numbers at earlier stages of B cell development under pathological conditions, for example, by supporting the survival (but not proliferation and differentiation) of developing B cells in the absence of IL-7 or the IL-7 receptor in *vivo* and *in vitro* (6, 8). In addition to *BIM*, other BH3-only proteins such as BMF and PUMA are expressed in B lymphoid cells, and their loss can also lead to increased B cell numbers (7) or synergistically increase B cell numbers in combination with the *Bim* KO (9), highlighting functional redundancies among the proapoptotic proteins.

The B lymphoid expansion resulting from the germline *Bim* KO is transplantable and affects both the follicular and marginal zone compartment (8). In addition, a floxed *Bim* allele has recently been generated, and its conditional deletion throughout the hematopoietic system using *Vav-Cre* recapitulates key features of the germline *Bim* KO phenotype, including increased white blood cell numbers and splenomegaly (10). Collectively, these findings indicate that the B cell-related features of the *Bim* KO phenotype emanate from an impact that is intrinsic to the hematopoietic cell lineage. However, whether these effects on B cell homeostasis are solely due to the loss of a function of *BIM* specifically within the B lymphoid cell lineage, or whether they may be in part due to an indirect, reactive consequence of losing *BIM*-dependent apoptosis in another hematopoietic cell type remains unresolved. In addition, if the alterations observed are due to the loss of B cell-intrinsic functions of *BIM*, it remains to be resolved to what extent they are caused by increased B cell production during their development in the bone marrow, or prolonged survival of mature B cells in the periphery. The relevance of these issues has recently been highlighted by the finding that conditional deletion of *Bim* in myeloid cells (using *LysM-Cre*) can in fact lead to increased splenic B cell numbers and immune-complex glomerulonephritis similar to that observed in germline *Bim* KO mice (11).

Thus, to investigate whether *BIM* regulates B cell homeostasis in a cell-intrinsic manner and to resolve the stage(s) of B cell development at which *BIM* may exert its most critical functions, we have here employed two different B lymphoid-specific CRE recombinase mouse strains for the conditional deletion of *Bim*: *Mb1-Cre* for deletion during the early developmental pro-B cell stage in the bone marrow (12), and *Cd23-Cre* for deletion at the nearly fully matured transitional B cell stages in peripheral lymphoid tissues (13).

### MATERIALS AND METHODS

**Mice**

Animal experiments were performed according to the Australian Code for the Care and Use of Animals for Scientific Purposes, 8th Edition (2013), and approved by the St. Vincent’s Hospital Melbourne Animal Ethics Committee, approval numbers 019/13 and 002/17.

*Mb1-Cre* (12), *Cd23-Cre* (13), *Eqy-Myc* (14), and *Bim-floxed* (10) mice had been generated, or backcrossed for at least 10 generations, on a C57BL6 background, and were housed in specific pathogen-free micro-isolators. All mice for bone marrow and splenic cell analyses were used at 8 weeks ± 3 days of age, and not selected on gender. Ethical endpoints for tumor-prone mice in survival analyses were determined by trained animal technicians who were blinded to the genotypes of individual mice.

**Immunoblot and Genotyping**

Western blotting was performed as described (15) using antibodies against BIM (Cell Signaling Technology, C34C5) and β-actin (EMD Millipore/Merck, MAB1501) as a loading control. DNA from toe biopsies was genotyped using primers 5'-AAGAATCTAGGTTGACTCTAG-3' and 5'-AACCAACTGTACCTTGCTATA-3' resulting in PCR products of ~1 kbp for the *Bim* floxed, ~0.8 kbp for the WT, and ~0.3 kbp for the *Bim*-deleted alleles. In addition, biopsies were genotyped with *Bim* KO primers (5'-AAGAATCTAGGTTGACTCTAG-3', 5'-CATTTGACTGAGATAGTGGTTGA-3', and 5'-CCCGTGTGACCACTAGTGA-3'; WT ~0.5 kbp, KO ~0.6 kbp) to exclude mice containing germline *Bim* null alleles as a result of ectopic CRE recombinase activity.

**Cell Preparations**

Hind limbs were dissected and skin, muscle, and soft tissues were removed carefully. Both ends of the femur and tibia were cut and the bone marrow was flushed out with 1 mL of ice-cold MACS buffer (1× PBS, pH 7.2, 0.5% BSA, and 2 mM EDTA) using a 21 G needle. Bone marrow cell suspensions from femur and tibia were combined and mixed thoroughly.

Freshly dissected mouse spleens were weighed and gently homogenized in 2 mL cold MACS buffer on ice. Cell suspensions were passed sequentially through 70 and 40-μm cell strainers to remove debris, followed by washes of tubes and strainers with 4 mL MACS buffer to recover remaining cells.

*B-1a* cells were isolated by peritoneal lavage using 26 and 23 G needles with PBS containing 2% fetal bovine serum (FBS).
Flow Cytometry
The following reagents were used for cell staining: B220-APC (eBioscience™, 17-0452-83), B220-FITC (Biolegend, 103206), BP1-PE (BD, 553735), CD11b-Pacific blue (eBioscience™, 48-0112-82), CD8-APC (eBioscience™, 17-0081-82), CD4-PE (eBioscience™, 12-0041-83), CD5-FITC (BD, 553021), CD11b-APC-Cy7® (BD, 557657), CD19-APC eFluor780 (eBioscience™, 47-0193-82), CD19-PerCP-Cy5.5® (eBioscience™, 45-0193-82), CD21-PE (Biolegend, 123410), CD23-biotin (BD, 553317), CD24-FITC (BD, 561777), CD43-biotin (BD, 553269), CD43-PE (BD, 553271), GR1-biotin (eBioscience, 13-5931-85), IgD-eFluor450 (eBioscience™, 48-5993-82), IgM-PE-Cy7® (BD, 552867), Brilliant Violet 605™ Streptavidin (Biolegend, 405229), and propidium iodide (Sigma, P4864-10ML). Our gating strategies for staining of bone marrow B lymphoid fractions according to Hardy et al. (16), total and mature splenic B cell numbers, and peritoneal cavity B-1a cell analyses were recently described (15, 17). Hardy fractions were gated as follows: A, B220+ CD43+ CD24high BP1+; B, B220+ CD43+ CD24high BP1+; C, B220+ CD43+ CD24high BP1+; D, B220+ CD43+ IgM- IgD-; E (immature B cells), B220+, CD43-, IgM-, IgD-; F (recirculating B cells), B220+, CD43+, IgD+. T1 transitional B cells were gated as B220+, IgMhigh IgD--; T2 transitional B cells as B220+, IgMhigh IgDhigh; and mature B cells as B220+, IgM+, IgD+. Hardy fractions A–D correspond to the Basel classification (18–20) as follows: Hardy A is equivalent to pre-pro-B cells (B220+ CD43+ CD19+); Hardy B/C represent pre-B/pro-B I cells (B220+ CD43+ CD19+ c-kit+ CD25– SL–); Hardy C corresponds to large pre-B II cells (B220+ CD43+ CD19+ c-kit+ CD25+ SL+); and Hardy D corresponds to small pre-B II cells (B220+ CD43+ CD19+ c-kit+ CD25+ SL+). For follicular and marginal zone B cell proportions, splenic cells were first gated on live B220+ CD19+ lymphocytes, followed by gating on CD21/35 and CD23 (follicular B cells: CD21/35+CD23−; marginal zone B cells: CD21/35−CD23+). FACS data were analyzed using FlowJo 10.3 software (Tree Star).

Statistical Analysis
Data from independent experimental replicates were analyzed using GraphPad Prism software. The numbers of independent samples are indicated in the figures and tables. Error bars indicate the mean ± SEM. p Values were calculated using the two-tailed unpaired Student’s t-test. The Mantel–Cox test was used for survival analyses.

RESULTS
Conditional Loss of BIM During Early Stages of B Lymphopoiesis Leads to Increased B Cell Numbers
To analyze cell-intrinsic effects of the loss of BIM on the B cell lineage, we crossed mice containing floxed Bim alleles (10) with mice containing the Mb1-Cre knock-in allele (12). Within the B lymphoid lineage, Mb1-Cre is active from the early pro-B cell stage (12), coinciding with the initiation of Igh gene locus rearrangement by V(D)J recombination. However, Mb1-Cre can have spurious ectopic activity in the germline (21), especially when transmitted via male breeders. During genotyping, we noticed unusually high frequencies (compared to our experience with other floxed loci, such as Asclz, Dynlin1, tp53) of ectopic recombination of the floxed Bim allele, even in the female germline. Germline heterozygosity for Bim can result in some haplo-insufficiency phenotypes, such as reduced life span (2), pronounced acceleration of B cell lymphoma development in Eμ-Myc mice (22), and protection from polycystic kidney disease in the absence of Bcl2 (23). We therefore only used bona fide Mb1-CreBimfl/fl mice for the analyses described here and excluded mice containing inadvertently rearranged germline Bim−/− mice to avoid any indirect effects from Bim heterozygosity in other cell types on the B lymphoid lineage.

Mb1-Cre Bim−/− deleted mice contained ~2-fold more total B cells in the spleen than matched Mb1-Cre Bim+/+ control mice (p < 0.0001), and this was particularly pronounced among mature B cells, with a much smaller effect on transitional B cells (Figure 1A). In contrast, the numbers of innate-like B-1a cells in the peritoneal cavity [which emanate from distinctive fetal stem cells, as opposed to ongoing hematopoiesis for conventional B cells (24)] were not increased in Mb1-Cre Bim−/− deleted mice (Figure 1B). Efficient B cell-specific Bim deletion and loss of all the three BIM protein isoforms [which result from alternative splicing of the Bim pre-mRNA (25)] was confirmed by PCR genotyping and immunoblot analyses (Figures 1C,D).

This expansion of the mature B cell pool was also reflected in an ~4-fold increase in recirculating B cells in the bone marrow of Mb1-Cre Bim−/− deleted mice (Hardy fraction F; Figures 1E,F).
Conditional *Bim* deletion also led to other significant changes in cellularity during distinct stages of B cell development in the bone marrow. This included a near-doubling of cells in the pro-B cell stage (when *Mb1-Cre* first becomes active; Hardy fraction B) that was carried over into the subsequent early pre-B cell stage (fraction C, when *Igh* V(D)J recombination is completed), and a <2-fold expansion of small pre-B cells (Hardy fraction D, during which *Igl* loci undergo VJ recombination) (Figure 1E). However, *Mb1-Cre Bim*-deleted mice contained normal numbers of cycling pre-B cells (Hardy fraction C'), the stage when cells expressing a successfully rearranged pre-BCR undergo ~5 cycles of clonal expansion. Surprisingly, *Mb1-Cre Bim*-deleted mice also contained normal numbers of immature B cells (Hardy fraction E), the stage when cells expressing self-reactive BCRs are eliminated by a mechanism that involves BIM-dependent apoptosis (3).

Collectively, these data indicate that loss of BIM can affect several stages of B cell development in a lineage-intrinsic manner, and that this is balanced by compensatory changes at other developmental stages.
Conditional Bim Deletion During the Transitional Stages of B Cell Development Leads to Cell-Intrinsic Expansion of the Mature Follicular B Cell Pool

The finding that the numbers of immature B cells (Hardy fraction E), which represent the final stage of B lymphoid development in the bone marrow, were not abnormally increased in Mb1-Cre Bim-deleted mice (Figure 1B) suggested that the expansion of the peripheral B cell pools in these mice may be due to an increased life span of mature B cells, rather than increased cell production during development. To test this hypothesis, we intercrossed mice containing floxed Bim alleles with mice containing a Cd23-Cre transgene. Cd23-Cre becomes active in transitional B cells after they have left the bone marrow, and recombination of floxed genes is only complete at the mature B cell stage in peripheral lymphoid organs (13). Cd23-Cre did not display ectopic germline activity toward the floxed Bim allele.

Consistent with our hypothesis, Cd23-Cre-mediated deletion of Bim led to a significant expansion of total and mature B cell numbers in the spleen and in the circulation compared to Cd23-Cre Bim<sup>+/+</sup> control mice (Figure 2A; Table 1). The increase in the numbers of mature B cells in Cd23-Cre Bim-deleted mice was comparable to the splenic B cell expansion in Mb1-Cre Bim-deleted mice (Figure 2A; note that the differences between Mb1-Cre Bim<sup>+/+</sup> and Cd23 Bim<sup>+/+</sup> mice were statistically not significant). Further analyses indicated that the expansion of the mature splenic B cell pool was restricted to the CD23-positive follicular B cell compartment, with no increase in the marginal zone B cells (Figure 2C, middle panel). To determine if this specificity for follicular cells was simply due to lower Cd23-Cre activity in the marginal zone B cells (which express only very low levels of Cd23), we repeated similar analyses using Mb1-Cre Bim-deleted mice. Interestingly, loss of Bim during early B cell development, before the divergence of follicular and marginal zone cells, again led to a specific expansion of the follicular B cell compartment but not of marginal zone B cells (Figure 2C, right panel). These results indicate that marginal zone B cells are largely indifferent to the loss of Bim, which is consistent with the notion that they have a much longer life span than follicular B cells (26).

We also enumerated B lymphoid cells in the bone marrow of Cd23-Cre Bim-deleted mice. In line with the expansion of the mature B cell pool in the spleen (Figure 2A) and circulation (Table 1), the numbers of recirculating mature B cells (Hardy fraction F) were significantly increased in Cd23-Cre Bim<sup>+/+</sup> mice compared to control animals (Figure 2B). As Cd23-Cre is not active at this early developmental stage, this finding indicates that this pro-B cell expansion may be a secondary consequence of the increased mature B cell pool, possibly due to altered cytokine profiles. This indirect expansion of pro-B cells in Cd23-Cre Bim-deleted mice, therefore, implies that the quantitatively similar increase of pro-B cells in Mb1-Cre Bim-deleted mice may also be due to cell-extrinsic effects. In addition to the effects on the B lymphoid compartment, Cd23 Bim-deleted mice also exhibited a modest ~50% increase of splenic CD8<sup>+</sup> T cells, but other cell types in the circulation and spleen were not significantly changed compared to controls (Tables 1 and 2).
Collectively, these results from the analysis of the Cd23-Cre-mediated Bim deletion mice indicate that loss of Bim within mature B cells is sufficient to increase peripheral B cell numbers in a cell-intrinsic manner, and that this can cause a secondary increase in pro-B cells and CD8⁺ T cells.

| Blood                        | Control (cells x 10⁶/ml) | Cd23-Cre Bim⁺⁻ (cells x 10⁶/ml) | Significance |
|------------------------------|--------------------------|---------------------------------|--------------|
| White blood cells            | 12.53 ± 0.93             | 20.43 ± 3.05                    | p = 0.0477   |
| B cells                      | 5.270 ± 0.32             | 12.28 ± 2.51                    | p = 0.0324   |
| CD4⁺CD8⁻ T cells            | 1.628 ± 0.403            | 2.336 ± 0.306                   |              |
| CD8⁺CD4⁻ T cells            | 2.592 ± 0.292            | 2.049 ± 0.181                   |              |
| CD4⁺CD8⁺ T cells            | 0.069 ± 0.042            | 0.166 ± 0.147                   |              |
| Granulocytes                 | 1.868 ± 0.505            | 1.414 ± 0.537                   |              |
| Monocytes                    | 1.425 ± 0.227            | 1.496 ± 0.183                   |              |
| Red blood cells              | 8.400 ± 120              | 8.977 ± 305                     |              |
| Platelets                    | 950 ± 193                | 1,157 ± 79                      |              |

n = 4.

| Spleen                      | Control (cells x 10⁶) | Cd23-Cre Bim⁺⁻ (cells x 10⁶) | Significance |
|------------------------------|-----------------------|-------------------------------|--------------|
| B cells                      | 82.21 ± 3.3546        | 117.17 ± 14.28                | p = 0.0327   |
| CD4⁺CD8⁻ T cells            | 31.17 ± 2.20          | 35.93 ± 3.34                  |              |
| CD8⁺CD4⁻ T cells            | 16.50 ± 1.06          | 23.45 ± 2.01                  | p = 0.0141   |
| CD4⁺CD8⁺ T cells            | 0.655 ± 0.207         | 1.206 ± 0.205                 |              |
| Granulocytes                 | 6.865 ± 0.846         | 4.625 ± 1.075                 |              |
| Monocytes                    | 6.473 ± 0.42          | 5.140 ± 0.514                 |              |

Controls are Cd23-Cre⁺⁺ Bim⁺⁺ mice; n = 4.

Loss of Bim Prevents the Elimination of Activation-Resistant Mature B Cells In Vitro

To assess how loss of Bim may lead to increased numbers of mature B cells, we performed in vitro B cell activation experiments using LPS and measured proliferation by dilution of CellTrace Violet (CTV). LPS induces a bi-phasic response in mature resting B cells. During the first phase, small B lymphocytes are activated to escape from their G₀ state by increasing their cell mass, and in the second phase these much larger lymphoblasts enter multiple rounds of cell division (27). It has previously been proposed that LPS activation of B cells leads to the inactivation of BIM as a result of multisite phosphorylation by ERK and subsequent ubiquitin-dependent proteasomal degradation (28). Interestingly, LPS-treated Cd23-Cre Bim⁻⁻ deleted B cells produced a similar number of dividing cells and a similar CTV dilution profile compared to Cd23-Cre control B cells (Figure 3A). However, whereas the small, non-blasting and undivided lymphocytes were almost completely lost from the cultures of control B cells after 4 days of LPS treatment, a substantial number of these resting cells survived in the cultures of Cd23-Cre Bim⁻⁻ deleted B cells (Figure 3B). Analysis of the CTV dilution assays, using the proliferation platform of the FlowJo software package, indicated that relative to the numbers of remaining undivided cells, there was a ~4-fold reduction in the average number of divisions per cell in the cultures of Bim⁻⁻ deleted B cells compared to the Cd23-Cre control B cells (Division index, Figure 3C). This difference was reduced to ~0.2-fold fewer divisions when only the activated cells that had divided at least once were taken into consideration (Proliferation index, Figure 3D). Qualitatively
similar results were also obtained in response to treatment with CD40L (Figures 3E,F). Taken together, these results indicate that loss of BIM primarily increases the survival of activation-resistant resting B lymphocytes that do not exit the G0 state, with a relatively minor effect on the survival of actively proliferating B lymphoblasts.

**Heterozygous Conditional Bim Deletion in Developing B Cells Accelerates MYC-Driven B Cell Lymphomagenesis**

While the above experiments indicate that under normal physiological conditions B cell-intrinsic loss of Bim exerts its main effect on the mature B cell pool, there are several instances where germline loss of Bim can have profound effects on murine B cell development under pathological conditions (6, 8, 29). For example, in the context of MYC-driven B cell lymphomagenesis, it has been shown that the loss of even a single copy of Bim leads to dramatically accelerated lymphoma development emanating from the pre-B and immature B cell stages in the bone marrow of Eμ-Myc transgenic mice (22). We therefore also compared the effects of heterozygous germline versus heterozygous B lymphoid-intrinsic conditional Bim deletion on the time course of lymphoma development in Eμ-Myc mice.

Compared to Mb1-Cre Eμ-Myc control mice (that also contained an Mb1-Cre knock-in allele; median survival 113 days), conditional Mb1-Cre Bim+/− Eμ-Myc mice (median survival 72 days) exhibited significantly accelerated lymphoma development (Figure 4A). The survival curve for these B lymphoid-specific conditional Bim heterozygous mice was not significantly different from the survival curve for otherwise isogenic germline Bim heterozygous mice [that also contained an Mb1-Cre knock-in allele (15)] (Figure 4A). Among the tumors that formed in the Mb1-Cre Bim-deleted Eμ-Myc mice, there were higher proportions of surface IgM-negative precursor B cell lymphomas compared to the Mb1-Cre Eμ-Myc control mice (Figure 4B), although this difference was statistically not significant. It should be noted that heterozygous conditional Bim deletion did not affect normal B cell development compared to Mb1-Cre control mice (Figure 4C). Overall, these data indicate that the effect of Bim heterozygosity on the lymphoma development of Eμ-Myc transgenic mice is largely B lymphoid-intrinsic, without quantifiably significant compounding effects of Bim deficiency in other tissues, including their developmental niche in the bone marrow.
DISCUSSION

One of the most prominent phenotypes of germline Bim KO mice is a profound increase in the number of peripheral B cells, and an associated splenomegaly, but it was unclear to what extent this was due to B lymphoid-specific functions of BIM or influenced by the absence of BIM in other tissues and hematopoietic cell types. Notably, the numbers of splenic B cells in the conditional Mbl-Cre Bim%-deleted mice reported here are strikingly similar to those of germline Bim KO mice (on the same genetic background, at the same age and under the same housing conditions, albeit at different times) that also contained a Mbl-Cre knock-in allele (29) (Figure 5). In contrast, the total spleen cell numbers were significantly more increased in the germline Bim%-null mice compared to the B lymphoid-specific Bim%-deleted mice (Figure 5). These data therefore indicate that the B cell expansion in the Bim KO is largely due to cell-intrinsic changes within the B-lymphoid cell lineage, but the loss of Bim in other cell types is also an important contributor to the severity of the splenomegaly.

The pool of mature B cells is constantly regenerated in mice in unison with the elimination of non-reactive or dormant B cells that are not encountering antigens specific to their unique BCRs (30, 31). This raises the question whether loss of BIM affects B cell numbers by regulating the generation of new B cells or by regulating the survival of existing B cells. In addition to having increased numbers of mature B cells, the conditional Mbl-Cre Bim%-deleted mice also contained more pro-B and pre-B cells than control animals (fractions B, C, and D; Figure 1E). However, this expansion of B lymphoid cells from earlier developmental stages was reversed to normal cellularity at the immature B cell stage (fraction F; Figure 1E), which represents the final stage before developing B cells emigrate from the bone marrow to peripheral lymphoid tissues including the spleen. This may be due to BIM-deleted cells undergoing fewer divisions at the large pre-B cell stage, perhaps due to limited availability of growth factors that drive this cell proliferation, or the elimination of excess cells by BH3-only proteins other than BIM. For example, the Bnf KO leads to more pronounced elevation of pre-B cells and immature-transitional B cells in the bone marrow than the Bim germline KO (7). Regardless, the observation that Mbl-Cre Bim%-deleted mice have increased numbers of mature B cells but normal numbers of immature B cells suggests that the expansion of mature B cells caused by the absence of BIM may largely be due to their increased life span rather than increased survival of cells at earlier stages of B lymphopoiesis. This notion is supported by our analysis of conditional Cd23-Cre Bim mice, which delete Bim only during the final differentiation stages after B lymphoid cells have left the bone marrow, yet exhibit a quantitatively comparable peripheral mature B cell expansion to the Mbl-Cre Bim mice.

The conclusion of our in vitro analyses, that BIM regulates B cell homeostasis primarily through the stochastic elimination of superfluous mature B cells, is also corroborated by our in vitro data, which show that loss of BIM had a much more pronounced effect on apoptotic cell death of untreated resting cells than on the survival of LPS-stimulated cells after 24–48 h in culture. The latter suggests that other proapoptotic BH3-only proteins, possibly PUMA (5), or other cell death mechanisms, such as death receptor-mediated apoptosis (32), may be more important than BIM in the killing of activated B lymphoblasts.

While the considerations above relate to how BIM regulates B cell homeostasis under physiological conditions in otherwise normal mice, it has previously been shown that germline loss of Bim can also impact B cell development, for example, in the context of other mutations or under pathological conditions. A prominent example of this is the accelerated development of B cell lymphomas emanating from pre-B cells or immature B cells in Ep-Myc transgenic mice (22). Our data indicate that the conditional and germline loss of Bim have quantitatively comparable effects in this context. Other examples of the effect of BIM on B cell development include the partial or complete rescue of B cell development defects of IL-7-deficient or IL-7 receptor-deficient mice (6, 8), dicer-deficient mice (33), or Asciz-deficient mice (29) by germline Bim KO. Indeed, we recently found that conditional Bim deletion can fully rescue the B cell developmental defects of Mbl-Cre Dynn11%-deleted mice (17), which are mechanistically and quantitatively similar to those observed in the Mbl-Cre Asciz%-deleted mice. This indicates that BIM exerts its effects on B cell development under pathological conditions, at least during B lymphomagenesis, also in a cell-intrinsic manner in the B lymphoid lineage.

Finally, while our data indicate that the expansion of mature B cells in Bim KO mice is largely driven in a cell-intrinsic manner, it has been reported that substantial B cell expansions can also be triggered by the conditional deletion of Bim within only the myeloid cell compartment (11). However, it should be noted that these analyses of LysM-Cre Bim%-deleted mice were performed in much older mice (8 months versus 8 weeks of age for our mice) and after they had developed a multi-organ autoimmune syndrome. Nevertheless, these findings highlight that it would be worthwhile to monitor the Cd23-Cre Bim%-deleted mice for longer periods to determine if B cell-specific loss of BIM eventually leads to age-dependent deposition of immune complexes in susceptible tissues, such as the glomeruli in the kidney, and the development of autoimmune syndrome-like pathologies, although these would be expected to be relatively mild on a C57BL/6 background (see above).

ETHICS STATEMENT

Animal experiments were performed according to the Australian Code for the Care and Use of Animals for Scientific Purposes, 8th Edition (2013), and approved by the St. Vincent’s Hospital Melbourne Animal Ethics Committee, approval numbers 019/13 and 002/17.
AUTHOR CONTRIBUTIONS

RL was involved in design, performing and analysis of experiments, and contributed to drafting of the manuscript. AK was involved in design, performing, and analysis of experiments. PB, DT, and AS provided resources and were involved in discussing experiments and editing the manuscript. JH was involved in the conception of the study, the design, analysis, and supervision of experiments, and drafting and editing of the manuscript.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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