Short report

**Insulin-degrading enzyme is exported via an unconventional protein secretion pathway**

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**Abstract**

Insulin-degrading enzyme (IDE) is a ubiquitously expressed zinc-metalloprotease that degrades several pathophysiologically significant extracellular substrates, including insulin and the amyloid β-protein (Aβ), and accumulating evidence suggests that IDE dysfunction may be operative in both type 2 diabetes mellitus and Alzheimer disease (AD). Although IDE is well known to be secreted by a variety of cell types, the underlying trafficking pathway(s) remain poorly understood. To address this topic, we investigated the effects of known inhibitors or stimulators of protein secretion on the secretion of IDE from murine hepatocytes and HeLa cells. IDE secretion was found to be unaffected by the classical secretion inhibitors brefeldin A (BFA), monensin, or nocodazole, treatments that readily inhibited the secretion of α1-antitrypsin (AAT) overexpressed in the same cells. Using a novel cell-based Aβ-degradation assay, we show further that IDE secretion was similarly unaffected by multiple stimulators of protein secretion, including glyburide and 3'-O-(4-benzoyl)benzoyl-ATP (Bz-ATP). The calcium ionophore, A23187, increased extracellular IDE activity, but only under conditions that also elicited cytotoxicity. Our results provide the first biochemical evidence that IDE export is not dependent upon the classical secretion pathway, thereby identifying IDE as a novel member of the select class of unconventionally secreted proteins. Further elucidation of the mechanisms underlying IDE secretion, which would be facilitated by the assays described herein, promises to uncover processes that might be defective in disease or manipulated for therapeutic benefit.

**Results**

Accumulating evidence from cell and animal modeling studies and human molecular genetics implicates impaired function of IDE in the pathogenesis of type 2 diabetes mellitus and Alzheimer disease (AD) [1-3]. IDE is the prototypical member of an evolutionarily distinct superfamily of zinc-metalloproteases possessing several features that distinguish it from conventional metalloproteases, including an "inverted" zinc-binding motif (HxxEH) [4] and an unusual tertiary structure [5-7]. Another distinguishing feature of IDE is its subcellular localization: the vast majority of IDE is present in the cytosol, with smaller amounts present in mitochondria, peroxisomes, and endosomes [8]. A small fraction of IDE—estimated to be 3% to 10% of the total—is also trafficked to the extracellular space, and it is this pool which interacts with known substrates of IDE, such as insulin and Aβ [9]. Despite a large number of studies demonstra-
ing that IDE is secreted and/or associated with the cell surface (e.g., [10-13]), little else is known about the underlying export pathway(s).

In previous work, we showed that alternative translation initiation of IDE mRNA at an initiation codon upstream of the canonical one leads to the incorporation of a 41-amino acid N-terminal sequence [14]. Bioinformatic analysis showed that this sequence was highly conserved (e.g., ~80% identity between humans and pufferfish), implying that it serves some important function, and some prediction programs predicted it to be a \textit{bona fide} signal sequence [14]. Subsequent experimental analysis, however, showed unequivocally that this sequence instead encodes a mitochondrial presequence, as demonstrated by the localization of green fluorescent protein tagged with this sequence, by electron microscopy, and by other methods [14,15]. Moreover, overexpression of IDE isoforms encoding this sequence had no effect on the degradation of extracellular substrates, in clear contradistinction to isoforms translated beginning at the canonical initiation codon [14,15].

Despite IDE’s apparent lack of a classical signal peptide, it is possible that the protein may nevertheless be exported through the classical secretory pathway, whether mediated by undefined posttranslational modifications (e.g., proteolysis to reveal a cryptic signal sequence), by association with heterologous classically secreted proteins, or by other means dependent upon a functional classical secretion pathway. To address this possibility, we co-transfected immortalized murine hepatocytes with full-length, unmodified cDNAs encoding human IDE and human α1-antitrypsin (AAT), a widely studied classically secreted protein [16]. Three days after transfection, cells were washed then treated with each of 3 different inhibitors of classical secretion: BFA (a blocker of ER-to-Golgi transport), monensin (an ionophore of monovalent cations that disrupts Golgi-dependent transport), and nocodazole (an inhibitor of microtubule polymerization that interferes with multiple forms of vesicular transport), or 0.1% EtOH as a vehicle control. Cell lysates and conditioned medium were collected, and the latter was concentrated 100-fold, then both were probed by western blotting for IDE and AAT. As expected, each of the treatments led to a significant decrease in the secretion of mature, ~62-kDa AAT and a concomitant increase in intracellular AAT or immature forms thereof (Fig. 1A, B). In striking contrast, however, the levels of secreted IDE were unaffected by any of these inhibitors of classical secretion (Fig. 1A, B). We confirmed that these compounds produced no significant cytotoxic effects at the concentrations tested, as judged by quantification of lactate dehydrogenase (LDH) release (data not shown).

To study effects on secreted IDE proteolytic activity \textit{per se}, we adapted our well-characterized fluorescence polarization-based Aβ degradation assay [17] for use in cells. To that end, HeLa cells transfected with IDE cDNA or empty vector were incubated for varying lengths of time with fluorescein-Aβ\textsubscript{1-40}\textsubscript{[Lys-LC-biotin]} (FAβB) [17]. A time-dependent increase in FAβB hydrolysis was observed that was strongly increased in the IDE-overexpressing cells (Fig. 2). In agreement with prior findings [12], FAβB hydrolysis was inhibited nearly completely by insulin (20 μM), a competitive inhibitor of IDE. Collectively, these results suggest that nearly all the Aβ-degrading activity produced by these cells is attributable to IDE, making FAβB degradation a reliable surrogate marker of extracellular IDE activity.

We then used this experimental paradigm to evaluate the effects of several inhibitors and activators of protein secretion on endogenously secreted IDE activity. Consistent with the results obtained above (Fig. 1), BFA, monensin and nocodazole at several concentrations produced no
significant effects on the secretion of IDE from HeLa cells (Fig. 3A). We then tested several activators of conventional and unconventional protein secretion, including the ATP-binding cassette (ABC) transporter inhibitor glyburide, which is known to stimulate insulin secretion from pancreatic beta cells; Bz-ATP, a known stimulator of P2X7 receptors, which have been proposed to mediate the unconventional secretion of IL-1β [18]; and the calcium ionophore, A23187. No significant effect on IDE activity was elicited by glyburide or Bz-ATP, but significant enhancement was observed by treatment with A23187 for 2 h. Quantification of the effects of the different treatments on LDH release in parallel experiments, however, revealed that A23187 was significantly cytotoxic under the conditions tested (Fig. 3B). Subsequent analysis showed that treatment with A23187 for shorter durations (e.g. ≤ 30 min) failed to produce significant effects on extracellular IDE activity or LDH release (not shown).

The primary goal of the present study was to establish whether the secretion of IDE proceeds–directly or indirectly–via the classical secretion pathway. Using two independent methods, we show that the secretion of catalytically active IDE is not affected by multiple inhibitors of classical secretion. This result places IDE in the emerging category of unconventionally secreted proteins, which are characterized by the lack of a classical signal peptide, the lack of posttranslational modifications specific for the ER and Golgi compartments, and resistance to BFA and other inhibitors of Golgi-dependent export [19]. The list of proteins documented to be secreted by unconventional means is relatively small (i.e., approximately 20; see Ref. [19]), but it includes a number of medically relevant proteins, such as fibroblast growth factor-1 and -2 (FGF-2), interleukin-1α and IL-1β, thioredoxin, galectins, and certain heat-shock and cytoskeletal proteins [19,20]. Proteins associated specifically with neurodegenerative diseases have also been shown to be secreted by unconventional means, including α-synuclein [21] and prion protein [22].

The mechanisms underlying unconventional protein secretion are largely mysterious, and only beginning to be elucidated for just a subset of proteins. Nevertheless, unconventional secretion can be usefully subdivided into vesicular and nonvesicular pathways, with the former category applying predominantly to integral-membrane pro-
A23187 (Sigma) were prepared and stored according to manufacturer’s recommendations. Cells were grown under normal cell culture conditions (37°C, 5% CO₂ in a humidified chamber).

**Methods**

**Chemicals**

BFA, monensin, nocodazole, glyburide, Bz-ATP, and A23187 (Sigma) were prepared and stored according to manufacturer’s recommendations.

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**Cell Culture and Transfections**

SV40-transformed wild-type murine hepatocytes were plated in 24-well plates (2 × 10⁵ cells/well), transfected with appropriate cDNAs using TransIT LT1 according to manufacturer’s recommendations (Mirus) and maintained in Hepatocyte Medium (3H Biomedical AB) supplemented with 20% fetal bovine serum. For experiments with HeLa cells a “reverse transfection” procedure was carried out in 384-well format (Mirus). Briefly, cDNA-TransIT LT1 complexes were prepared in Opti-MEM I low-serum medium (Invitrogen), added to 384-well plates (20 μL/well), then HeLa cells suspended in DMEM/20% FBS were added to the wells (20 μL/well; 10,000 cells/well). Cells were grown under normal cell culture conditions (37°C, 5% CO₂ in a humidified chamber).

**Collection of Secreted and Intracellular IDE and AAT for Biochemical Detection**

Three days after transfection, immortalized murine hepatocytes were washed 2 times in DMEM/0.1% bovine serum albumin (BSA), then incubated with the latter medium supplemented with appropriate chemicals or 0.1% EtOH as a vehicle control. Cells were pre-treated with compounds for 30 min, washed, then cells were allowed to condition the chemical-containing media for 2 h. Conditioned medium (250 μL/well, 6 wells/condition for each experiment) was collected, cleared of cellular debris by centrifugation, then concentrated 100-fold at 4°C using 2-mL YM-30 Microcon spin columns (Amicon) according to manufacturer’s recommendations. Cells were washed twice with DMEM/0.1%BSA then lysed in PBS supplemented with 0.1% Triton X100 and 1× Complete Mini protease inhibitor cocktail (Roche) (30 μL/well, 6 wells/condition). After incubation on ice for 30 min, the lysates were centrifuged at 10,000 g at 4 °C for 10 min to pellet cell debris and unlysed cells.

**Western Blot Analysis**

Cell lysates and concentrated conditioned media were subjected to SDS-PAGE under reducing conditions using 10% polyacrylamide tris-glycine mini gels (Invitrogen) and transferred to nitrocellulose membranes as described [25]. Membranes were blocked in 5% non-fat milk in tris-buffered saline supplemented with 0.2% Tween-20 (TBS-T), incubated for 1 h with antibodies against IDE (IDE-1; 1:1000; Ref. [13]; generous gift of Dennis Selkoe, Harvard Medical School) or AAT (GALT-80A; 1:1000; Immunology Consultants Laboratory), washed with TBS-T, probed with peroxidase-conjugated anti-rabbit IgG and anti-goat IgG secondary antibodies, respectively (1:5000; Vector Labs), and detected by enhanced chemiluminescence using SuperSignal West Pico Substrate (Pierce). Western blots were quantified using NIH Image 1.62 (NIH).
Cell-based FA/B Degradation Assay

HeLa cells plated in 384-well format (10,000 cells/well) were washed twice in DMEM/0.1% BSA, then incubated at 37 °C with the latter medium supplemented with FA/B (0.5 μM; 40 μL/well). Reactions were terminated by addition of DMEM/0.1% BSA supplemented with avidin (2.5 μM) and 1.10 phenanthroline (5 mM; 10 μL/well). Fluorescence polarization ($r_{ex} = 485$, $r_{em} = 535$) was determined using a SpectraMax M5 microplate reader (Molecular Devices). Controls included wells lacking cells (0% hydrolysis), wells supplemented with 10 nM recombinant IDE (100% hydrolysis) or excess insulin (20 μM) to establish IDE-dependent vs. IDE-independent hydrolysis. For experiments investigating the effects of chemicals on FA/B degradation, in vitro tests were conducted to guard against effects on the fluorescence polarization read-out or on the activity of recombinant IDE.

Cytotoxicity assay

LDH release was quantified using the Cytotox 96 cytotoxicity assay (Promega) according to manufacturer’s recommendations.

Abbreviations

AAT: α1-antitrypsin; Aβ: amyloid β-protein; AD: Alzheimer disease; BFA: brefeldin A; Bz-ATP: 3′-O-(4-benzoyl)benzoyl-adenosine triphosphate; FA/B: fluorescein-Aβ1-40[Lys-LC-biotin]; FGF-2: fibroblast growth factor-2; IDE: insulin-degrading enzyme; IL-1β: interleukin-1β; LDH: lactate dehydrogenase; P2X7 receptor: P2X purinergic receptor 7.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

IZ participated in the design of the study and executed the cell biological and enzymatic assays. LL generated the immortalized murine hepatocyte cell line, generated recombinant IDE and assisted IZ in the execution of all experiments. ML designed the experiments, analyzed data and wrote the manuscript.

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