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Amplification Dynamics of Platy-1 Retrotransposons in the Cebidae Platyrrhine Lineage

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Abstract

Platy-1 elements are Platyrrhine-specific, short interspersed elements originally discovered in the Callithrix jacchus (common marmoset) genome. To date, only the marmoset genome has been analyzed for Platy-1 repeat content. Here, we report full-length Platy-1 insertions in other New World monkey (NWM) genomes (Saimiri boliviensis, squirrel monkey; Cebus imitator, capuchin monkey; and Aotus nancymaae, owl monkey) and analyze the amplification dynamics of lineage-specific Platy-1 insertions. A relatively small number of full-length and lineage-specific Platy-1 elements were found in the squirrel, capuchin, and owl monkey genomes compared with the marmoset genome. In addition, only a few older Platy-1 subfamilies were recovered in this study, with no Platy-1 subfamilies younger than Platy-1-6. By contrast, 62 Platy-1 subfamilies were discovered in the marmoset genome. All of the lineage-specific insertions found in the squirrel and capuchin monkeys were fixed present. However, 15% of the lineage-specific Platy-1 loci in Aotus were polymorphic for insertion presence/absence. In addition, two new Platy-1 subfamilies were identified in the owl monkey genome with low nucleotide divergences compared with their respective consensus sequences, suggesting minimal ongoing retrotransposition in the Aotus genus and no current activity in the Saimiri, Cebus, and Sapajus genera. These comparative analyses highlight the finding that the high number of Platy-1 elements discovered in the marmoset genome is an exception among NWM analyzed thus far, rather than the rule. Future studies are needed to expand upon our knowledge of Platy-1 amplification in other NWM genomes.

Key words: insertion, polymorphism, evolution, subfamilies.

Introduction

Transposable elements (TEs) are discrete pieces of DNA that are able to move from one genomic location to another. These elements can be broadly categorized based on their movement mechanism, either “cut-and-paste” or “copy-and-paste.” The former category includes DNA transposable elements that mobilize via a DNA intermediate. (Hellen and Brookfield 2013). The latter category includes retrotransposable elements that move via an RNA intermediate (Batzer and Deininger 2002; Konkel et al. 2010). In primates, retrotransposable nonautonomous short interspersed elements (SINEs) such as Alu elements (Houck et al. 1979) and autonomous long interspersed elements (LINEs) called L1s make up roughly 10% and 17% of the genome by mass, respectively (Lander et al. 2001; Batzer and Deininger 2002; Richardson et al. 2015). The high copy number Alu elements are ~300 base pairs (bp) long with a dimeric structure, separated by a middle A-rich region. These elements are derived from the 7SL RNA, a component of the signal recognition particle (Batzer and Deininger 2002; Konkel et al. 2010). These elements are mobilized via a retrotransposition mechanism called target-primed reverse transcription (TPRT) (Luan et al. 1993;
et al. 2005; Osterholz et al. 2009). This is in part due to poor of NWM has been under debate (Ray and Batzer 2005; Ray 2015). Since the first study of NWM cladistics, the phylogeny of monkeys), Atelidae (large monkeys with prehensile tails), or Platyrrhini (formerly subsumed into one genus, Cebus) (Alfaro et al. 2012), and Callitrichinae which includes marmosets (genera Callitrichis, Callimico, Cebuella, and Mico) and tamarins (genera Leontopithecus and Saguinus) (Garbino and Martins-Junior 2018). While reported divergence times and radiation of these three clades have varied, there is a general consensus that the rapid radiation occurred over a short time of 1–2 Myr. Estimates of when this divergence occurred range from 19.25 Ma (Perelman et al. 2011) to 23.2 Ma (Schneider 2000).

Recently, a new retrotransposable element was discovered in the common marmoset genome and subsequently found to be specific to the Platyrrhini parvorder. Deemed, “Platy-1,” these ~100 bp elements have the hallmarks of movement via TPRT (Konkel et al. 2016). In addition, these elements share some structure and sequence similarity with Alu elements, a primate-specific SINE, suggesting that Platy-1 likely originated from an Alu element and is 7SL RNA derived (Konkel et al. 2016). Roughly 2,200 Platy-1 elements were found in the common marmoset genome [calJac3], prompting a closer look at other NWMs with whole genome sequence data available.

Although once thought to be “junk” DNA (Kazazian 2011), TEs have had an unexpected influence on primate biology in terms of disease, phenotypes, and evolution. TEs can cause genomic instability via double-stranded breaks (Belgnaoui et al. 2006; Gasior et al. 2006) and nonhomologous recombination (Han et al. 2005a; Startek et al. 2015), potentially influencing meiosis, depending upon the location of the insertion as well as resulting in the contraction or expansion of genome size. In addition, based on their insertion location, TEs can affect transcriptional control via influencing alternative splicing if inserted into the coding region of a gene, disrupting the formation of a gene product, or influencing the methylation status of the TE’s surrounding environment (Cordaux and Batzer 2009). It is therefore informative to understand the amplification dynamics of mobile elements in order to understand how genomes have evolved, particularly because of parallel evolution in which many mobile elements may be active in multiple lineages simultaneously. Due to parallel evolution, each NWM lineage will have its own unique distribution of not only TE families but also distinctive subfamilies within each family. For example, the discovery of 46 Saimiri lineage-specific Alu subfamilies was recently reported (Baker et al. 2017), most of which derived from the larger group of established AluTa subfamilies. AluTa subfamilies are specific to NWM as the result of a unique fusion event between two anthropoid AluS subfamilies (Ray and Batzer 2005) and have been used to study phylogenetic relationships.

The purpose of this study was to characterize the Platy-1 elements found in the current genome assemblies of other Platyrrhine primates in order to determine the quantity of Platy-1 elements as well as the amplification dynamics in comparison to the common marmoset genome (Konkel et al. 2016).

Materials and Methods

Platy-1 Lineage Specificity

The capuchin monkey (Cebus imitator), owl monkey (Aotus nancymaeae), and squirrel monkey (Saimiri boliviensis) genomes were obtained from NCBI (cebus-Cebus_imitator-1.0; owl-Anan_1.0) and the University of California Santa Cruz (UCSC) genome browser (squirrel-saiBo1). Assembly statistics for each genome are available in Table 1 and representative photographs for each species are shown in fig. 1). These genomes were analyzed for Platy-1 elements using RepeatMasker (RepeatMasker-Open-4.0) utilizing a custom library of the 62 Platy-1 subfamilies previously defined (Konkel et al. 2016) and all current Alu subfamily consensus sequences obtained from RepBase (Jurka et al. 2005). Full-length Platy-1 elements were defined as possessing a start position no less
than 4 bp and an end position not shorter than two nucleotides prior to the A-tail within the consensus sequence (Konkel et al. 2016). Full-length elements were extracted from the RepeatMasker output using a custom python script. These elements, along with 600 bp of flanking sequence on both the 5’ and the 3’ ends of the Platy-1 insertion, were compared with the common marmoset (Callithrix jacchus/calJac3) and the remaining NWM genomes using a locally installed version of BLAT (Kent 2002) to determine lineage specificity. Specificity was determined by visualizing the BLAT alignments using pslPretty and observing a ~100 bp gap. For each locus an alignment file was generated in BioEdit (Hall 1999) to be used for the design of oligonucleotide primers.

Platy-1 Shared Elements

To analyze shared elements among NWM, the whole-genome aligner mugsy (Angiuoli and Salzberg 2011) was utilized. All Platy-1 elements with flanking sequence for each lineage (squirrel, capuchin, and owl monkeys as well as marmoset) were put into one FASTA file. The resulting four FASTA files were then aligned as if they were whole genomes using the whole genome function in mugsy. The output .maf file was visualized using GMAJ (globin.bx.psu.edu/dist/gmaj/; last accessed March 26, 2019) and manually assessed for alignment precision. This analysis proved fruitful for obtaining elements that were shared among all four of the genomes analyzed. However, elements that were computationally predicted to be shared between only two or three of the four genomes, typically had gaps in the sequence assembly of the genome(s) in which the insertion was absent, thus obscuring any potential phylogenetically informative data. To overcome this limitation, for the pool of elements not shared among all four genomes, we used BLAT followed by a custom python script to obtain orthologous sequences from each genome and then aligned all four sequences for each locus using BioEdit (Hall 1999).

Oligonucleotide Primer Design

The loci determined to contain Platy-1 elements unique to each NWM were put into individual files containing the orthologous sequences from marmoset, squirrel monkey, owl monkey, and capuchin monkey genomes. These sequences were aligned using CLUSTALW (Thompson et al. 1994) and/or MUSCLE (Edgar 2004). Forward and reverse oligonucleotide primers for polymerase chain reaction (PCR) were designed using Primer3 (v.0.4.0) and checked in BioEdit to ensure minimal mismatches to allow for the amplification of a PCR product in all genomes specified. In silico PCR was used to confirm the oligonucleotide primers would amplify only one product in multiple species. The same process was followed for the shared Platy-1 elements (supplementary file 1, Supplementary Material online).

DNA Samples

DNA samples are described in supplementary file 2, Supplementary Material online. Briefly, there were four panels utilized for this study: a NWM panel, a squirrel monkey panel, an owl monkey panel, and a capuchin monkey panel. The NWM panel contained three Old World monkeys (OWM) and sixteen NWM species representing the three NWM families. This DNA panel was used to screen elements for lineage-specificity. The squirrel monkey panel included DNA samples from 32 individuals of the genus Saimiri.
representing five species, the owl monkey panel included DNA samples from 23 individuals of the genus Aotus representing five species, and the capuchin monkey panel included DNA from 14 different capuchin monkeys, 8 representing five species, and the capuchin monkey panel in- apella Cebus imitator MgCl2, 10 indicating 25

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mutation rate of 0.006024 per base per million years (my) (Konkel sequence, a feature available in the RepeatMasker output. The 

The age of the Platy-1 elements was estimated by utilizing the 

PCR amplification was performed in 25 μl reactions containing 25 μg of template DNA, 200 nM of each primer, 1.5 mM MgCl2, 10× PCR buffer (1×: 50 mM KCl; 10 mM Tris–HCl, pH 8.4), 0.2 mM dNTPs, and 1 unit of Taq DNA polymerase. The

The squirrel monkey genome had a considerably higher number of lineage-specific insertions, with the majority of the elements belonging to the 4a subfamily (supplementary file 3, Supplementary Material online). Of the 119 loci analyzed by PCR, 88 were homozygous present, while 31 remained polymorphic for insertion presence/absence among 23 Aotus individuals analyzed (supplementary file 3, Supplementary Material online and fig. 3B). The owl monkey genome had a considerably higher number of lineage-specific insertions, with the majority of the elements being either 4 or 4a subfamily members (table 2 and fig. 3C). Of the 119 loci analyzed by PCR, 88 were homozygous present, while 31 remained polymorphic for insertion presence/absence among 23 Aotus individuals analyzed (supplementary file 3, Supplementary Material online; figs. 2C and 4). The Aotus genus was the only one of four genera in this study to show evidence of ongoing Platy-1 mobilization. Given the rapid radiation of these four genera as discussed in a review article by Schneider and Sampaio (2015), our data suggests the emergence of the 4a Platy-1 subfamily approximately between 19 and 20 Ma.

Table 1 Genome Assembly Statistics

| Genome       | Common Name       | Assembly Name                  | N50 (Contig) | N50 (Scaffold) | Coverage | Number of Gaps | Size (bp) |
|--------------|-------------------|--------------------------------|--------------|----------------|-----------|----------------|-----------|
| Cebus imitator | Capuchin monkey   | Cebus_imitator-1.0              | 41,196       | 5,274,112      | 81×       | 133,441        | 2.72×10⁶  |
| Sapajus apella | Owl monkey        | Anan_1.0                        | 28,503       | 8,280,397      | 113.4×    | 215,259        | 2.93×10⁵  |

Note.—The assembly statistics for the NWM genomes used in this study are shown above.

Results

Lineage-Specific Platy-1 Insertions in NWM

A total of 387, 605 and 335 Platy-1 loci were retrieved from the RepeatMasker analysis of the capuchin [Cebus_imitator-1.0], owl monkey [Anan_1.0], and squirrel monkey [saiBol1] genomes, respectively (table 2). Of these, 171, 378, and 158 were determined to be full-length insertions, as previously defined (see Materials and Methods; Konkel et al. 2016). These values are strikingly low as compared with the 2,268 full-length Platy-1 elements previously identified in marmoset [cAlJac3] (Konkel et al. 2016). In the capuchin genome, there were 22 predicted lineage-specific Platy-1 insertions, with 16 insertions conducive to locus-specific PCR (table 2, figs. 2A and 3A). The squirrel monkey genome had a similarly low number of lineage-specific insertions, 36, with 18 of these analyzed by PCR (table 2, figs. 2B and 3B). With 145 loci, the owl monkey genome had the largest number of lineage-specific insertions of the three NWM genomes investigated. Of these, 119 insertions were analyzed using locus-specific PCR (table 2, figs. 2C and 3C).

Of the capuchin monkey lineage-specific Platy-1 insertions, the majority belonged to the 4a subfamily (fig. 3A). All of the 16 loci subjected to PCR (see Materials and Methods) were homozygous for the presence of the insertion (supplementary file 3, Supplementary Material online). A similar trend was observed for the squirrel monkey lineage-specific insertions, as all 18 PCR-analyzed loci in this lineage were fixed present and the majority of these insertions also belonged to the 4a subfamily (supplementary file 3, Supplementary Material online and fig. 3B). The owl monkey genome had a considerably higher number of lineage-specific insertions, with the majority of the elements being either 4 or 4a subfamily members (table 2 and fig. 3C). Of the 119 loci analyzed by PCR, 88 were homozygous present, while 31 remained polymorphic for insertion presence/absence among 23 Aotus individuals analyzed (supplementary file 3, Supplementary Material online; figs. 2C and 4). The Aotus genus was the only one of four genera in this study to show evidence of ongoing Platy-1 mobilization. Given the rapid radiation of these four genera as discussed in a review article by Schneider and Sampaio (2015), our data suggests the emergence of the 4a Platy-1 subfamily approximately between 19 and 20 Ma.

Among the 31 polymorphic loci identified in the owl monkey genome, the allele frequency variation across 23 Aotus
FIG. 2.—Lineage-specific Platy-1 elements. (A) The presence of the Ceb_5 capuchin monkey specific Platy-1 element is indicated by the higher of the two bands present (254-bp band), while the absence is indicated by the lower of the two bands present (139-bp band). (B) The presence of the Ply4a-27 squirrel monkey specific Platy-1 element is indicated by the higher of the two bands present (355-bp band), while the absence is indicated by the lower of the two bands present (240-bp band). (C) The presence of the U_OM_89423_v3 owl monkey specific Platy-1 element is indicated by the higher of the two bands present (511-bp band), while the absence is indicated by the lower of the two bands present (377-bp band). Lanes: 1: 100-bp ladder; 2: TLE (negative control); 3: Human (HeLa); 4: Chimpanzee; 5: African green monkey; 6: Wooly monkey; 7: White-bellied spider monkey; 8: Black-handed howler monkey; 9: Bolivian red howler monkey; 10: Common marmoset; 11: Pygmy marmoset; 12: Geoffroy’s marmoset; 13: Red-cheeked mustached tamarin; 14: Geoffroy’s saddle-back tamarin; 15–17: Capuchin monkey; 18: Squirrel monkey; 19: Owl monkey; 20: Northern white-faced saki; 21: Bolivian gray titi; 22: 100-bp ladder. The bars above the gel electrophoresis image indicate the following: Blue-Old World Monkey; Gold-NWM; Green-Atelidae; Purple-Cebidae; Red-Callithrichinae; Gray-Pitheciidae. Scientific names of the primates are indicated below the gel images.
individuals revealed a distinct separation between the two recognized groups of owl monkey, red- and gray-necked (Menezes et al. 2010), for at least three loci. For these loci there was a clear separation of species with (homozygous present) and without (homozygous absent) a Platy-1 insertion (supplementary file 3, Supplementary Material online and fig. 4), reflecting the red-necked (A. nancymaae, A. azarae) and gray-necked (A. lemurinus, A. trivirgatus, A. vociferans) divergence seen in South America (Menezes et al. 2010). The majority of the lineage-specific Platy-1 elements discovered in this study were members of previously-defined Platy-1 subfamilies 4 or 4a based on the subfamily consensus sequences reported in Konkel et al. (2016). Evidence of recent mobilization within Aotus, and no observed mobilization activity among the Saimiri, Cebus, or Sapajus genera prompted us to construct a sequence alignment of all owl monkey lineage-specific Platy-1 elements (supplementary file 5 and fig. S1, Supplementary Material online).

After comparing owl monkey-specific loci to the Platy-1-4 consensus sequence reported in Konkel et al. (2016), at least two distinct diagnostic mutations occurred since Aotus diverged from the other genera. Among the loci present in the owl monkey genome, there were multiple shared diagnostic mutations at positions 19 (G to C transversion), 26 (T to A transversion), 64 (G to T transversion), 70 (C to G transversion), 79 (C to T transition), and 82 (A to G transition). This newly discovered subfamily was named Platy-1-4b_aotus (n = 58) and is aligned in (fig. 5). The nomenclature convention is as follows: this subfamily appears to have derived from Platy-1-4 but had different diagnostic substitutions than Platy-1-4a and was discovered in owl monkeys.

Another diagnostic change was identified among Aotus-specific loci, some of which were fixed present while others

### Table 2

|                      | Total | Full-Length | Lineage-Specific | PCR |
|----------------------|-------|-------------|------------------|-----|
| **Capuchin monkey**  | 387   | 171         | 22               | 16  |
| **Squirrel monkey**  | 335   | 158         | 36               | 18  |
| **Owl monkey**       | 605   | 378         | 145              | 119 |

Note.—The table shows the total Platy-1 elements extracted from the RepeatMasker output (Total), the full-length elements extracted from the RepeatMasker output (Full-length), the elements that were predicted to be lineage-specific, and full-length loci analyzed using locus specific PCR for each NWM.

![Graph](https://example.com/graph.png)

**Fig. 3**.—Predicted lineage-specific Platy-1 elements and PCR analyses. A comparison of the number of the predicted lineage-specific and PCR-analyzed loci is shown, as well as the number of elements belonging to each Platy-1 subfamily in the aforementioned categories. (A) Capuchin monkey, (B) squirrel monkey, and (C) owl monkey. Note the differences in scale between capuchin and squirrel versus owl monkeys.
were polymorphic. While sharing all diagnostic mutations that compose Platy-1-4b_aotus, there were three additional diagnostic mutations at positions 12 (G to A transition), 62 (A to G transition), and 100 (A to G transition). This new subfamily was termed Platy-1-4b3_aotus (n = 10). This follows standard nomenclature for naming repeats, as this subfamily has all the mutations of Platy-1-4b_aotus with 3 additional mutations (Batzer et al. 1996; fig. 5). Platy-1 mobilization in the Aotus lineage is consistent with the stealth model of SINE amplification dynamics (Han et al. 2005b) in which a few very old elements remain dormant for millions of years before slowly emerging with active daughter elements.

In the owl, squirrel, and capuchin monkey genomes analyzed in this study, no Platy-1 subfamilies younger than Platy-1-1 to Platy-1-6 were identified in the initial RepeatMasker analysis. This is in contrast to the marmoset genome where 62 subfamilies were discovered and all are present in [calJac3] (Konkel et al. 2016). As a part of that initial marmoset study, a subset of Platy-1 elements representing the majority (50 of 62) of subfamilies were analyzed by PCR to assess their distribution among NWM species. A graphic illustration of those results is shown in: (supplementary file 5 and fig. S2a, Supplementary Material online). Subfamilies shared among all NWM on this graph belonged to the oldest subfamilies (Platy-1-1 through Platy-1-3) and one insertion specific to all Cebidae belonged to subfamily Platy-1-4a. These data are in agreement with the RepeatMasker analysis performed in this study that identified the subfamily range between 4 and 4a as the source of lineage-specific elements reported here.
Divergence of Platy-1 Subfamilies in NWM

Among the Platy-1 subfamilies there is a wide range of nucleotide divergence values from the respective subfamily consensus sequences as identified by RepeatMasker (fig. 6). A higher percent divergence from the respective subfamily consensus sequence is considered generally indicative of the age of the insertion event, as older elements have more time to accumulate random mutations. Mobile elements, on average, accumulate mutations at a neutral rate. As a consequence, the divergence can be used as an estimate of the age of the insertion. Figure 6 illustrates that the vast majority of the full-length Platy-1 elements identified in this study are relatively old. This finding is consistent with the PCR results from the squirrel and capuchin monkey lineage-specific DNA panels in which all the lineage-specific insertions had reached fixation throughout the genus. The average age corresponding with the percent divergence of the predicted lineage-specific Platy-1 insertions reported for the capuchin monkey is 12.4 Ma with a range from 4.8 to 22 Ma, while the average age of the lineage-specific Platy-1 insertions reported for squirrel monkey is 13.2 Ma with a range from 3.2 to 27.7 Ma. Older Platy-1 subfamilies correspond with higher percent divergence and therefore higher average age (e.g., capuchin monkey and squirrel monkey Platy-1-1: 21.8 and 23.4 Ma, respectively) (supplementary file 4, Supplementary Material online).

The lower nucleotide divergence values of the Platy-1 insertions found in the owl monkey genome (fig. 6C) were consistent with more recent insertions and are in agreement with the polymorphic loci found via PCR. The average age of the predicted lineage-specific owl monkey Platy-1 insertions calculated from the percent divergence of the insertion sequence to its respective consensus sequence is 8.5 Ma, with a range from 0 to 25.4 Ma (supplementary file 4, Supplementary Material online).

It is of note that these age estimates are based on a retrotransposable element that is only ~100 bp in length and therefore could represent a fairly broad range. Even a one nucleotide change is equivalent to 1% divergence or ~1.66 Myr. However, these results are intended to emphasize that the NWM genomes studied here contain primarily older Platy-1 elements, as compared with the marmoset genome in which the relative divergence values and age estimates illustrate that the marmoset genome contains large quantities of younger elements.

Shared Platy-1 Elements

Over half (127/230; 55%) of the shared elements identified using a mugsy alignment (see Materials and Methods) were found within all four NWM genomes analyzed (marmoset, owl monkey, squirrel monkey, and capuchin monkey). The actual number is likely higher than this data set reflects due to lack of homology across multiple genome assemblies. These data are in agreement with the low number of lineage-specific insertions found in the NWM genomes analyzed in this study.

Five of the loci that were predicted to be lineage-specific in the squirrel monkey and capuchin monkey genomes were experimentally determined by PCR to be shared between the aforementioned genomes. These five
loci were fixed present in all individuals representing the Saimiri, Cebus, and Sapajus genera (supplementary files 1 and 3, Supplementary Material online). These data are consistent with the close established relationship between Saimiri, Cebus, and Sapajus. In addition, these shared elements all belonged to the 4a Platy-1 subfamily, indicating that the age of these elements could reflect the evolutionary divergence time of Saimiri, Cebus, and Sapajus from other Cebids.

**Discussion**

This study expanded upon the research reported by Konkel et al. (2016) by not only recovering Platy-1 insertions unique
to other NWM genomes but also analyzing the amplification dynamics of these insertions. It is striking to note that there are a considerably lower number of Platy-1 repeats in owl, capuchin, and squirrel monkeys compared with the expansion and proliferation of Platy-1 insertions seen in the marmoset genome (Konkel et al. 2016). However, when comparing the three NWM genomes included in this study, there is a larger number of total, full-length, and lineage-specific insertions found in the owl monkey genome than in the capuchin and squirrel monkey genomes (Table 2). Platy-1 mobilization in owl monkeys appears to have been relatively quiescent for millions of years, dating back to the 4a subfamily, and only recently resumed with modest retrotransposition activity leading to the origin of two new Aotus lineage-specific subfamilies. By contrast, Platy-1 retrotransposition in capuchin and squirrel monkeys remains quiescent. One possible explanation is polymorphic loci were subject to lineage sorting during speciation, potentially eliminating source drivers for Platy-1 mobilization. This explanation is consistent with the lower overall numbers of Platy-1 elements in the capuchin and squirrel monkey genomes and higher overall percent divergences of the elements from their consensus sequences. In addition, all of the lineage-specific loci ascertained from the capuchin and squirrel monkey genomes were determined to be fixed present. These data indicate negligible recent Platy-1 mobilization in these lineages. This slow propagation is likely not due to a lack of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in L1, the element that provides the necessary enzymes for TPRT, of available enzymatic machinery as it has been shown that L1, the element that provides the necessary enzymes for TPRT, has recently amplified in

These findings suggest that the extensive proliferation of Platy-1 elements in the common marmoset is the exception, rather than the norm in NWM genomes. Such disparities could be the result of differing effective population sizes after speciation, opposing environmental pressures, or genomic environment of the Platy-1 insertions in the different genera. There are also biological differences that might play a role. For example, marmosets have a unique aspect to their reproduction in that they mostly produce twins. The twinning of marmoset leads to genetic chimeras. This inherent genetic diversity in addition to the rapid reproduction of marmosets may have led to an environment favorable to retrotransposable element propagation. (Consortium 2014; Harris et al. 2014). The peak rate of Platy-1 propagation reportedly occurred with the rise of the marmoset ancestor (Konkel et al. 2016) when several Platy-1 subfamilies were active in parallel. In contrast, early Platy-1 evolution likely started with a low number of source elements resulting in very slow mobilization as illustrated by the NWM lineages analyzed in this study.

The polymorphic loci identified in this study that delineate between red- and gray-necked owl monkeys may be particularly useful for medical studies in which species identification is important. Owl monkeys have long been used as an animal model for malaria, with Aotus lemurinus griseimembra as the primary species used that is susceptible to the parasite responsible for causing this particular human malady (Herrera et al. 2002; Moreno-Pérez et al. 2017). While other owl monkey species have varying degrees of malaria susceptibility, the results with A. l. griseimembra have been particularly reproducible. Another owl monkey species, A. vociferans, is also susceptible to infection, but not as widely used as A. l. griseimembra. It is of note that both of these species are considered gray-necked (Herrera et al. 2002). Having reproducible genetic markers to distinguish between gray- and red-necked owl monkeys may be of value for biomedical studies. Although only a few polymorphic loci were identified in the owl monkey genome, with a small fraction of those showing a clear separation between gray- and red-necked owl monkeys, these markers provide a quick, simple, and unambiguous identification that is not currently available for this organism.

Platy-1 insertions observed in this study were often flanked by an Alu on either the 5’ or 3’ end of the Platy-1 element, and on rare occasions both sides. As reported by Konkel et al. (2016), Platy-1 elements with intact TSDs also possess endonuclease cleavage sites, indicating that Platy-1 elements have the same sequence and/or insertional preference as Alu elements. Although Platy-1 elements may occasionally highjack Alu movement, Platy-1 elements are present in substantially lower overall numbers than Alu insertions in NWM. For example, a recent study of the Saimiri genome determined that there were 739,636 full-length and 43,201 lineage-specific Alu elements (Baker et al. 2017), a much higher number than the full-length and lineage-specific Platy-1 elements analyzed in this study. In addition, the majority of the Platy-1 elements characterized in this study were shared among all four of the NWM genomes studied. This indicates that there are simply not enough phylogenetically informative Platy-1 elements to be able to resolve NWM relationships. Alu elements may provide the key to elucidating NWM relationships as they have previously resolved difficult primate connections (Ray et al. 2005; McLaren et al. 2012, 2013; Meyer et al. 2012; Walker et al. 2017; Jordan et al. 2018). A whole genome comparison of Alu insertion polymorphisms among the four NWM genomes described here may provide useful for elucidating some of the NWM relationships.

It is important to note that the repeats identified in this study were ascertained from the reference genome for all of the species studied. In addition, loci that were analyzed via PCR needed to be in conserved regions of all four genomes for confirmation of lineage-specificity. Sufficient time may have also passed that some of the insertions belonging to older subfamilies have experienced sufficient decay and were not recognized by RepeatMasker in the initial genome screening. It is therefore possible that the number of lineage-specific repeats and overall Platy-1 content in the NWM genomes analyzed is somewhat higher than reported.
Undoubtedly, all three genomes have a sharply lower number of Platy-1 elements compared with marmoset.

**Supplementary Material**

Supplementary data are available at Genome Biology and Evolution online.

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**Authors’ Contributions**

J.M.S. performed analyses, computational and benchtop experiments, and wrote the article. J.R.M., Y.S., and S.A.B. analyzed the Aotus and Saimiri, respectively, lineage-specific insertions by performing computational and benchtop analyses. B.T., L.C.R., C.P.S., and M.M.F. performed experiments. J.N.B. and M.K.K. performed computational analyses by ascertaining Platy-1 elements from the Aotus and Saimiri genomes, respectively. A.D.M., J.D.O., and K.A.P. provided samples and edited the article. M.A.B. and J.A.W. designed the research and edited the final article. All authors read and approved the final article.

**Literature Cited**

Alfarò JW, Silva JD, Jr, Rylands AB. 2012. How different are robust and gracile capuchin monkeys? An argument for the use of sapajus and cebus. Am J Primatol. 74(4):273–286.

Anguoli SV, Salzberg SL. 2011. Mugsy: fast multiple alignment of closely related whole genomes. Bioinformatics 27(3):334–342.

Baker JN, et al. 2017. Evolution of Alu subfamily structure in the saimiri lineage of New World monkeys. Genome Biol Evol. 9(9):2365–2376.

Batzer MA, Deininger PL. 2002. Alu repeats and human genomic diversity. Nat Rev Genet. 3(5):370–379.

Batzer MA, et al. 1996. Standardized nomenclature for Alu repeats. J Mol Evol. 42(1):3–6.

Belgnaoui SM, Gosden RG, Semmes OJ, Haoudi A. 2006. Human LINE-1 retrotransposon induces DNA damage and apoptosis in cancer cells. Cancer Cell Int. 6:13.

Boissinot S, Roos C, Furano AV. 2004. Different rates of LINE-1 (L1) retrotransposon amplification and evolution in New World monkeys. J Mol Evol. 58(1):122–130.

Caozzo O, Archidiacono N, Loruso N, Stanyon R, Rocchi M. 2016. The 14/15 association as a paradigmatic example of tracing karyotype evolution in New World monkeys. Chromosomes 125(4):747–756.

Cawthon Lang KA. 2005. Primate factsheets: owl monkey (Aotus) taxonomy, morphology and ecology [cited 2019 Mar 6]. Available from: http://pin.primate.wisc.edu/factsheets/entry/owl_monkey, last accessed March 26, 2019.
