The presence of disseminated tumour cells in the bone marrow is inversely related to circulating free DNA in plasma in breast cancer dormancy

Patients with primary breast cancer frequently have evidence of minimal residual disease (MRD) in the absence of any clinical or radiological evidence of metastases (Slade et al, 2009). The detection of circulating tumour cells (CTCs) in the blood and disseminated tumour cells (DTCs) in the bone marrow of disease-free patients with breast cancer has been well documented. Their presence identifies those patients with a worse prognosis, although many remain disease free for many years or decades (Meng et al, 2004; Braun et al, 2005; Slade et al, 2009). In addition, circulating free DNA (cfDNA) has been found in higher concentrations in breast cancer patients than healthy controls and has been proven to share similar genetic features to the primary tumour (Leon et al, 1977; Stroun et al, 1989). Larger fragment sizes of cfDNA, greater than the apoptotic limit, have also been detected in the blood of cancer patients; believed to be derived from tumour cell necrosis and lysis (Jahr et al, 2001; Wang et al, 2003; Umetani et al, 2006). The relationship of these measures of MRD to long-term breast cancer dormancy has not been fully established.

Many post-operative studies have principally been carried out by sampling patients’ bone marrow or blood shortly after surgery or adjuvant chemotherapy during the period of highest risk of relapse; however, very few studies have focused on the later ‘low risk’ period, that is, later than 4 years after surgery. Patients in this period with MRD present are likely to have cells in a dormant state although the precise mechanisms are unclear. We decided to concentrate on this period for several reasons. First, a proportion of patients relapse during this period: it is generally agreed that there is a constant rate of relapse of around 1–2% annually (van der Sangen et al, 2011); many patients develop evidence of widespread metastatic disease in intervals between clinic visits, and this is a considerable cause of morbidity. Second, we reasoned that if a test, or combination of tests, could be shown to indicate recurrent disease during this time, this may provide a reason to instigate systemic therapy at an earlier stage, with the aim of eradicating the disease. Finally, it was possible that we would find a subset of patients who had no evidence of disease at any time point using any test; these patients should not need to be followed up in clinic, thus reducing patient anxiety and cost.

The phenomenon of ‘dormancy’ in relation to cancer has been extensively described in the literature (Meng et al, 2004; Almog et al, 2009; Willis et al, 2010). The exact mechanisms of dormancy are still not clearly understood; however, evidence of a relationship...
between tumour dormancy and increased apoptosis exists. In a mouse model under angiogenic suppression, tumour cell proliferation was balanced by a reciprocal amount of cell death (Holmgren et al, 1995). We hypothesised that by combining tests indicating cell death (cfdNA) with other measures of viability (DTC and CTC detection), we could determine the relationship between the two processes.

The comparison of cfdNA with other markers of MRD has not been fully assessed, although evidence for a positive association of cfdNA with viable CTCs detected by an epithelial immunospot assay has been described (Schwarzenbach et al, 2009a). Conversely, no correlation of cfdNA with DTCs measured by immunocytochemistry (ICC) was found (Schwarzenbach et al, 2009b). In this study, we aimed to find a combination of tests to characterise dormancy mechanisms in breast cancer patients, which would enable possible routes of therapeutic intervention. For this, we compared DTCs measured by two methods (quantitative RT-PCR measurement of cytokeratin 19 (CK19) and ICC, using a pan-cytokeratin antibody) and CTCs by CellSearch with two measures of cfdNA. The tests consisted of the measurement of two overlapping amplicons: a 96-bp amplicon, representing total cfdNA and a larger 291-bp amplicon that was above the apoptotic limit (185–200-bp) and believed to be mostly cancer associated. These measurements of MRD were compared with each other and with prognostic factors in patients with breast cancer on long-term follow-up.

MATERIALS AND METHODS

Primary breast cancer patients and control groups

Three control groups were recruited to measure CTCs and cfdNA in the blood; these consisted of 34 healthy female controls (29 of which had CTCs measured) and 51 patients with benign breast disease (BBB; 28 of which had CTCs measured). Thirty-one patients with metastatic breast cancer were used as a control group to confirm raised levels of cfdNA and a higher number of CTCs (eight of these patients had CTCs measured) as compared with healthy controls (Allard et al, 2004; Van der Auwera et al, 2009). All patient groups including the primary breast cancer patients signed a written consent form; both form and protocol had been approved by the local ethics committee (RREC numbers 0975). Disseminated tumour cells in the bone marrow were not measured in the control groups for ethical reasons, but our test for bone marrow DTCs has been standardised and extensively validated in previous studies (Borgen et al, 1999; Slade et al, 1999, 2005, 2009). The amount of blood taken for CTC measurement was 22.5 ml in the healthy controls and patients with BBB and 7.5 ml in the patients with metastatic breast cancer. Twenty millilitres of blood was taken for all cfdNA analyses.

In this study, we recruited 64 patients who had been treated for primary breast cancer. Twenty-two patients had previously been recruited to our MRD studies (Slade et al, 2005, 2009). Twenty-nine patients were recruited from a trial of adjuvant high-dose chemotherapy (Coombes et al, 2005) and were selected because they had a high risk of relapse. The remaining 13 patients were selected as additional patients for statistical purposes. The inclusion criteria included a histologically confirmed breast cancer and consent to the follow-up procedure (see below). Exclusion criteria included evidence of recurrent disease on conventional staging with isotopic bone scan and computed tomography of the chest and liver/abdomen, before study entry. All patients agreed to repeat bone marrow sampling at the outset of the study followed by repeat blood sampling for further tests to measure MRD. In all, 43 (67%) patients received adjuvant chemotherapy and 58 (91%) patients received adjuvant endocrine therapy (Table 1).

### Table 1 Primary breast cancer patient characteristics

| Lymph node-positive patients (%) | Lymph node-negative patients (%) | Total (%) |
|----------------------------------|----------------------------------|-----------|
| **Menopausal status**            |                                  |           |
| Pre-menopausal                   | 11 26                            | 7 32      | 18 28     |
| Post-menopausal                  | 31 74                            | 15 68     | 46 72     |
| **Histology**                    |                                  |           |
| Invasive ductal                  | 31 74                            | 16 73     | 47 73     |
| Invasive lobular                 | 7 17                             | 2 9       | 9 14      |
| Other/mixed invasive             | 4 9                              | 4 18      | 8 13      |
| **Tumour size**                  |                                  |           |
| T1                               | 15 36                            | 20 91     | 35 55     |
| T2                               | 16 38                            | 2 9       | 18 28     |
| T3                               | 7 17                             | 0 0       | 7 11      |
| T4                               | 0 0                              | 0 0       | 0 0       |
| Unknown                          | 4 9                              | 0 0       | 4 6       |
| **Tumour grade**                 |                                  |           |
| I 1                              | 2 5                              | 23 6      |
| II 20                            | 48 9                             | 4 11      | 29 45     |
| III 20                           | 48 8                             | 36 28     | 44        |
| Unknown                          | 1 2                              | 0 0       | 1 2       |
| **Hormone receptor status**      |                                  |           |
| ER positive                       | 29 69                            | 17 77     | 46 72     |
| ER negative                      | 10 24                            | 5 9       | 15 23     |
| Unknown ER status                | 3 7                              | 0 0       | 3 5       |
| PgR positive                     | 17 40                            | 13 59     | 30 47     |
| PgR negative                     | 19 45                            | 7 32      | 26 41     |
| Unknown PgR status               | 6 14                             | 2 9       | 8 12      |
| **HER2 status**                  |                                  |           |
| Positive                         | 13 31                            | 3 14      | 16 25     |
| Negative                         | 26 62                            | 17 77     | 43 67     |
| Unknown                          | 3 7                              | 2 9       | 5 8       |
| **Endocrine therapy**            |                                  |           |
| Yes                              | 41 98                            | 17 77     | 58 91     |
| No                               | 1 2                              | 5 23      | 6 9       |
| **Chemotherapy**                 |                                  |           |
| Yes                              | 35 83                            | 8 36      | 43 67     |
| No                               | 7 17                             | 14 64     | 21 33     |
| **Total number of patients**     | 42 66                            | 22 34     | 64 100    |

Abbreviations: ER = oestrogen receptor; PgR = progesterone receptor.

Detection of CTCs and DTCs from blood and bone marrow

The skin was incised before the bone marrow aspirates were taken to minimise the risk of epithelial contamination. Between 2 and 5 ml of bone marrow was aspirated into syringes primed with preservative-free heparin (Leo Labs, Risborough, UK). Cells were cytocentrifuged at a concentration of 5 × 10^5 per cytopsin; air-dried and stored at −20 °C before use. Slides were then stained for a common epitope of cytokeratin as previously described (Pantel et al, 1994; Slade et al, 1999; Smith et al, 2000). Samples that were isotype positive were deemed uninterpretable and therefore excluded from the results. The MCF-7 cell line was used as a positive control. Real-time qRT-PCR for CK19 and ABL mRNA was performed as described previously (Slade et al, 2005). The standard used for quantification was the artificial construct in the range 10^1–10^4 for CK19 and 10^2–10^6 for ABL per 2.5 µl (Slade et al, 1999). All assays were performed with duplicate standards, a non-cDNA control and a positive cDNA control were extracted from the MCF-7 cell line. A CK19:ABL ratio greater or equal to...
0.1% (one CK19/1000 ABL transcripts) was regarded as positive. All the samples were anonymised to the person performing the assays. For the detection of CTCs, 7.5 ml of blood from the metastatic patients and 3 x 7.5 ml from the control and primary breast cancer patients, were collected in CellSave preservative tubes (Veridex, LLC, Warren, NJ, USA) from patients in London, anonymised and transported at room temperature to the Department of Surgery and Cancer, Imperial College London for processing within 72 h of collection as recommended by the manufacturer. The CellSearch System (Veridex, LLC, Warren, NJ, USA) was used for the isolation and enumeration of CTCs from each 7.5 ml of blood separately using specific morphological criteria to identify CTCs (Allard et al, 2004).

**CfDNA detection and analysis**

Blood samples were collected in EDTA tubes and processed within 3 h of venesection. Plasma was separated by centrifugation at 1200 g for 10 min, and plasma was taken from the upper phase and decanted into fresh polypropylene tubes. Tubes were then spun again to remove any contaminating leucocytes (Chiu et al, 2001), and the resulting supernatant was aliquoted into sterile tubes and stored at –80 °C. Plasma samples were allowed to thaw to room temperature and re-spun in a bench-top centrifuge to remove any remaining cell debris as recommended (Page et al, 2001), and cfDNA was isolated as described previously (Page et al, 2011). To estimate cfDNA quantity, the concentration of a 96-bp GAPDH amplicon served as a measure of total cDNA quantity, whereas a larger overlapping 291-bp amplicon measured total cfDNA above the limit of apoptosis. The integrity of cfDNA was calculated as a ratio of the 96-bp and 291-bp amplicons (291-bp/(291-bp + 96-bp)). Primers and conditions are as described previously (Page et al, 2011).

**Study design**

The primary objective of this prospective, longitudinal study was to investigate the natural history of DTCs in bone marrow aspirates from 64 primary breast cancer patients. This study also had secondary objectives to investigate the presence and quantity of CTCs and cfDNA in blood. Few patients relapsed with recurrent

**Table 2** CTC measurements from blood and cell-free DNA quantity in plasma are lower in healthy controls and benign breast disease patients than in metastatic breast cancer.

|                     | Healthy controls | Benign breast disease | Metastatic breast cancer |
|---------------------|------------------|-----------------------|--------------------------|
| Cell-free DNA 96-bp (ng/ml⁻¹) |                  |                       |                          |
| No. of patients     | 34               | 51                    | 31                       |
| Mean                | 0.24             | 0.54                  | 3.5                      |
| 95% Confidence interval | 0.09 – 0.63     | 0.27 – 1.07           | 1.48 – 8.29              |
| Median              | 0.86             | 1.27                  | 2.45                     |
| Range               | 0 – 4.52         | 0 – 8.36              | 0.03 – 1044              |
| Cell-free DNA 291-bp (ng/ml⁻¹) |                  |                       |                          |
| No. of patients     | 34               | 51                    | 31                       |
| Mean                | 0.24             | 0.44                  | 3.5                      |
| 95% Confidence interval | 0 – 0.20        | 0.1 – 0.40            | 0 – 1.5                  |
| Median              | 0                | 0.22                  | 0.44                     |
| Range               | 0 – 1.37         | 0 – 1.94              | 0 – 602                  |
| Cell-free DNA integrity |                  |                       |                          |
| No. of patients     | 34               | 51                    | 31                       |
| Mean                | 0.10             | 0.28                  | 0.21                     |
| 95% Confidence interval | 0 – 0.13        | 0.11 – 0.25           | 0 – 0.34                 |
| Median              | 0                | 0.2                   | 0.07                     |
| Range               | 0 – 0.39         | 0 – 0.49              | 0 – 0.83                 |
| Circulating tumour cells |                  |                       |                          |
| No. of patients     | 29               | 28                    | 8                        |
| Volume of blood measured | 22.5 ml         | 22.5 ml               | 7.5 ml                   |
| Mean                | 0.03             | 0.29                  | 24                       |
| 95% Confidence interval | 0.04 – 0.11     | 0.09 – 0.66           | 7.29 – 55.29             |
| Median              | 0                | 0                     | 9                        |
| Range               | 0 – 1            | 0 – 5                 | 0 – 109                  |
| No. of patients with CTCs | 1 (3%)          | 4 (14%)               | 6 (75%)                  |

**Table 3** Minimal residual disease marker values in lymph node-positive and lymph node-negative primary breast cancer patients in samples taken over time.

|                      | Lymph node-positive patients at surgery (n = 42) | Lymph node-negative patients at surgery (n = 22) | P-value |
|----------------------|--------------------------------------------------|--------------------------------------------------|---------|
| Cell-free DNA 96-bp (ng/ml⁻¹) | Mean 19.19                                       | 14.19                                            | <0.0001* |
| 95% CI               | 0.61 – 18.93                                     | 0.24 – 7.52                                      |         |
| Median               | 1.06                                            | 3.01                                             |         |
| Range                | 0 – 2626                                        | 0 – 440                                          |         |
| No. of samples       | 323                                             | 152                                              |         |
| Cell-free DNA 291-bp (ng/ml⁻¹) | Mean 4.35                                       | 2.66                                             | 0.0248* |
| 95% CI               | 0.16 – 4.90                                     | 0.05 – 1.43                                      |         |
| Median               | 0.17                                            | 0.38                                             |         |
| Range                | 0 – 785                                         | 0 – 78                                           |         |
| No. of samples       | 322                                             | 149                                              |         |
| Cell-free DNA integrity | Mean 0.22                                       | 0.18                                             | 0.479   |
| 95% CI               | 0.001 – 0.029                                    | 0.001 – 0.037                                    |         |
| Median               | 0.17                                            | 0.13                                             |         |
| Range                | 0 – 1                                           | 0 – 1                                            |         |
| No. of samples       | 257                                             | 136                                              |         |
| Disseminated tumour cells (qRT-PCR) | Mean 0.09                                       | 0.07                                             | 0.0154* |
| 95% CI               | 0 – 0.012                                       | 0.001 – 0.023                                    |         |
| Median               | 0.053                                           | 0.04                                             |         |
| Range                | 0 – 1.25                                        | 0 – 1.12                                         |         |
| No. of samples       | 475                                             | 133                                              |         |
| Disseminated tumour cells (ICC) | Mean 0.94                                       | 0.79                                             | 0.063   |
| 95% CI               | 0.005 – 0.15                                     | 0.009 – 0.28                                     |         |
| Median               | 0                                               | 0                                                |         |
| Range                | 0 – 15                                          | 0 – 12                                           |         |
| No. of samples       | 463                                             | 139                                              |         |
| Circulating tumour cells | Mean 0.55                                       | 0.15                                             | 0.982   |
| 95% CI               | 0.012 – 0.38                                     | 0.003 – 0.099                                    |         |
| Median               | 0                                               | 0                                                |         |
| Range                | 0 – 23                                          | 0 – 2                                            |         |
| No. of samples       | 153                                             | 72                                               |         |

Abbreviation: CTC = circulating tumour cell. aCalculated as the ratio of the 96-bp and 291-bp amplicons (291-bp/(291-bp+96-bp)). bDetected with the CellSearch system. cSamples were considered positive for CTCs when at least one cell was detected per total sample volume.

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Molecular Diagnostics

and Disseminated tumour cells (qRT-PCR)

Circulating tumour cells

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disease therefore MRD values were compared with each other and with prognostic factors in a univariate analysis. For ethical and technical reasons, samples measuring CTCs and cfDNA were taken at fewer time points as detailed below.

Statistical methods

The main continuous variables of 96- and 291-bp amplicons in cfDNA, ICC and qRT-PCR assays for DTC, and CTC assay were positively skewed. The Spearman rank correlation (corrected for ties) was used to assess monotonic relationships between continuous variables. The Mann–Whitney test was used to compare continuous scores between two groups. One-way analysis of variance was used to compare more than two groups. A binary comparison of minimal residual disease markers in primary breast cancer patients with hormone receptor and HER2 receptor status of the tumours measuring 8.3 years). In terms of their primary staging, 20 patients had cancer were followed up for 1–11.5 years after surgery (a median of 8.3 years). In terms of their primary staging, 20 patients had tumours measuring <20 mm with no involved lymph nodes detected (T1N0), 4 patients had 1–3 involved lymph nodes detected and 2 patients had tumours measuring >20 mm with no involved lymph nodes detected and 38 high-risk patients had >3 involved lymph nodes at surgery. For the purposes of this study, patients were grouped into node-positive (n = 42) or node-negative (n = 22) groups (Table 1). Table 1 also shows patient characteristics and adjuvant systemic therapy details. The node-positive patients had bone marrow aspirates taken every 6 months, and node-negative patients every 12 months for ethical reasons. Blood samples were obtained 3–6 monthly post-operatively over a follow-up period of up to 11.5 years. For cfDNA, 2–13 (a mean of 7.4) plasma samples were analysed per patient and 0–7 (a mean of 3.5) samples for CTC analysis using the CellSearch system. For DTC analysis, 0–21 (a mean of 9.5) bone marrow aspirates were taken from each patient (both ICC and qRT-PCR if the volume was sufficient). A total of 168 bone marrow aspirates were taken at the same time points as the blood sampling and used in the direct comparison of DTCs, CTCs and cfDNA. All samples for CTC analysis also had corresponding samples for cfDNA analysis. Total measurements of DTCs, CTCs and cfDNA were used when comparing values from lymph node positive with lymph node-negative breast cancer patients.

Minimal residual disease measurements in control groups

Only 1 of 29 (3%) healthy female controls had a single CTC detected and 4 of 28 (14%) patients with BBD had CTCs present in

Table 4  Comparison of minimal residual disease markers in primary breast cancer patients with hormone receptor and HER2 receptor status of the primary tumour

|                         | ER status |       | HER2 status |       | PR status |       | Triple negative status |       |
|-------------------------|-----------|-------|-------------|-------|-----------|-------|------------------------|-------|
|                         | Positive  | Negative | Positive  | Negative | Positive  | Negative | Yes | No |
| Total circulating free DNA 96-bp (ng/ml) | No. of samples 341 | 105 | 128 | 308 | 220 | 187 | 81 | 364 |
| Mean 21.67 | 4.82 | 21.2 | 15.14 | 9.50 | 30.23 | 124 | 20.43 |
| 95% CI 3.42 to 39.92 | 3.14 to 6.51 | 4.81 to 7.42 | 1.81 to 32.10 | 4.11 to 14.9 | 2.54 to 63.0 | 1.70 | 1.35 |
| Median 1.2 | 2.29 | 0.89 | 1.91 | 0 | 0 | 0 | 0 |
| Range 0–2626 | 0–53.71 | 0–1581 | 0–2626 | 0–440 | 0–2626 | 0–53.71 | 0–2626 |
| P-value 0.0044 | 0.0130 | 0.71 | 0.0444 |

Circulating free DNA 291-bp (ng/ml)

|                         | No. of samples 336 | 106 | 129 | 305 | 215 | 188 | 82 | 360 |
| Mean 4.46 | 4.46 | 8.08 | 1.79 | 2.08 | 1.68 | 1.68 | 1.68 |
| 95% CI –29.2 to 9.20 | –0.79 to 2.87 | –4.11 to 20.28 | 1.01 to 257 | 0.68 to 3.47 | 1.53 to 15.22 | 0.84 to 4.83 | 0.22 to 6.63 |
| Median 0.23 | 0.19 | 0.1 | 0.28 | 0.38 | 0.16 | 0.24 | 0.23 |
| Range 0–785.4 | 0–56 | 0–785.4 | 0–78.03 | 0–124 | 0–785 | 0–56 | 0–785.4 |
| P-value 0.792 | 0.188 | 0.23 | 0.50 |

Disseminated tumour cells (qRT-PCR) ¹

|                         | No. of samples 440 | 135 | 171 | 399 | 273 | 261 | 106 | 476 |
| Mean 0.087 | 0.096 | 0.87 | 0.09 | 0.07 | 0.10 | 0.09 | 0.09 |
| 95% CI 0.07–0.10 | 0.07–0.11 | 0.07–0.11 | 0.08–0.10 | 0.06–0.09 | 0.08–0.12 | 0.07–0.11 | 0.08–0.10 |
| Median 0.05 | 0.07 | 0.05 | 0.048 | 0.05 | 0.05 | 0.08 | 0.05 |
| Range 0–1.25 | 0–0.79 | 0–0.79 | 0–1.25 | 0–1.12 | 0–1.25 | 0–0.65 | 0–1.25 |
| P-value 0.11 | 0.39 | 0.022 | 0.06 |

Disseminated tumour cells (ICC) ²

|                         | No. of samples 136 | 435 | 175 | 389 | 272 | 260 | 102 | 474 |
| Mean 0.93 | 1.02 | 0.81 | 0.82 | 0.82 | 0.79 | 0.91 | 0.82 |
| 95% CI 0.67–1.36 | 0.72–1.00 | 0.60–1.03 | 0.75–1.11 | 1.47 | 1.82 | 1.70–1.60 | 0.74–1.02 |
| Median 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Range 0–15 | 0–12 | 0–9 | 0–15 | 0–12 | 0–15 | 0–15 | 0–12 |
| P-value 0.34 | 0.45 | 0.23 | 0.16 |

Circulating tumour cells ³

|                         | No. of samples 166 | 48 | 74 | 135 | 125 | 72 | 34 | 178 |
| Mean 0.36 | 0.67 | 0.54 | 0.38 | 0.36 | 0.40 | 0.51 | 0.34 |
| 95% CI 0.08 to 0.64 | –0.15 to 1.49 | –0.09 to 1.17 | 0.08 to 0.67 | 0.03 to 0.77 | –0.03 to 1.06 | –0.25 to 2.08 | 0.08 to 0.61 |
| Median 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Range 0–23 | 0–19 | 0–23 | 0–19 | 0–23 | 0–19 | 0–19 | 0–23 |
| P-value 0.38 | 0.60 | 0.73 | 0.15 |

Abbreviations: CI = confidence interval; ICC = immunocytochemistry; qRT-PCR = quantitative reverse transcription-PCR. ¹Statistically significant difference of median values by Mann–Whitney test or mean values by t-test. ²Calculated as the percentage ratio of qRT-PCR CK1/ABL (≥0.1 is considered positive). ³Detected by pan-CK antibody. Samples were considered positive for circulating tumour cells when at least one cell was detected in 22.5 ml of blood by the CellSearch system.
Comparisons of markers of MRD with established prognostic factors in 64 breast cancer patients on long-term follow-up

Samples from multiple time points taken from each of the 64 primary breast cancer patients on long-term follow-up were included in this pooled analysis. In all, 8 of the 64 patients relapsed. There were no significant differences in cfDNA concentration, cfDNA integrity, DTC ICC or CTCs in the eight patients who had relapsed compared with the 56 who had not. However, there was a significant difference in the level of DTC detection in BM by qRT-PCR (P = 0.025, Mann–Whitney). Next, we compared measures of MRD in all 64 patients, comparing lymph node-negative vs lymph node-positive patients. The mean quantities of both the 96- and 291-bp amplicons in cfDNA were correlated with the number of lymph nodes positive at surgery (P < 0.0001 and P = 0.0248, respectively; Table 3). However, there was extensive overlap in the value ranges between node-positive and node-negative patients (Supplementary Figure 1). Disseminated tumour cells (qRT-PCR) in the BM were positively correlated with node-positive patients (P = 0.0154). There was no association between DTCs (ICC) or CTCs and node status. The measures of MRD (DTCs, CTCs and cfDNA) from individual blood or bone marrow samples were also compared with conventional prognostic factors in the 64 patients. In cfDNA, the 96-bp amplicon was significantly higher in patients with ER-negative (P = 0.004), HER2-negative (P = 0.013) and triple-negative status (P = 0.004) compared with those whose tumours express ER or HER2. The DTC (qRT-PCR) median values were also higher in PR-negative patients (P = 0.02) and triple-negative patients but this did not reach statistical significance (P = 0.06; Table 4).

When considering tumour size, grade and menopausal status, there were three significant associations: the 96-bp cfDNA amplicon was significantly higher in patients with high-grade breast cancer also showed a higher mean number of CTCs per ml of blood by the CellSearch system.

### Table 5
Comparison of minimal residual disease markers in primary breast cancer patients with menopausal status and with tumour size and grade of the primary tumour

|                | Post  | Pre  | T1    | T2    | T3    | I    | 2    | 3    |
|----------------|-------|------|-------|-------|-------|------|------|------|
| Total circulating free DNA 96-bp (ng/ml⁻¹) |       |      |       |       |       |      |      |      |
| Mean 95% CI   | 32.61 | 4.31 | 8.3   | 40.99 | 3.73  | 13.84| 7.26 | 27.73|
| Median        | 3.32  | 6.19 | 2.6   | 6.02  | 1.31  | 1.37 | 1.29 | 1.33 |
| Range         | 0–2626| 1–1976| 0–4396| 0–2626| 0–4541| 0–3721| 0–1160| 0–5636|
| No. of samples| 212   | 262  | 254   | 139   | 53    | 38   | 217  | 2    |
| P-value       | 0.120 |      | 0.026 |      | 0.032 |      |      |      |

| Circulating free DNA 291-bp (ng/ml⁻¹) |       |      |       |       |       |      |      |      |
| Mean 95% CI   | 6.77  | 1.35 | 2.38  | 7.74  | 0.74  | 1.47 | 1.78 | 5.99 |
| Median        | 0.78  | 1.32 | 1.11  | 3.66  | 0.2  | 0.33 | 0.39 | 1.33 |
| Range         | 0–785.4| 0–56 | 0–1242| 0–785.4| 0–1368| 0–1604| 0–785.4| 0–1368|
| No. of samples| 211   | 260  | 251   | 139   | 52    | 36   | 215  | 215  |
| P-value       | 0.033 |      | 0.102 |      | 0.305 |      |      |      |

| Disseminated tumour cells (qRT-PCR) |       |      |       |       |       |      |      |      |
| Mean 95% CI   | 0.087 | 0.088| 0.08  | 0.09  | 0.104 | 0.08 | 0.08 | 0.1   |
| Median        | 0.07–1 | 0.07–1 | 0.07–1 | 0.07–1 | 0.07–1 | 0.07–1 | 0.07–1 | 0.07–1 |
| Range         | 0–1.25 | 0–1.124 | 0–1.124 | 0–1.124 | 0–1.124 | 0–1.124 | 0–1.124 | 0–1.124 |
| No. of samples| 258   | 358  | 256   | 192   | 113   | 49   | 257  | 290  |
| P-value       | 0.846 |      | 0.846 |      | 0.519 |      |      |      |

| Disseminated tumour cells (ICC) |       |      |       |       |       |      |      |      |
| Mean 95% CI   | 0.8   | 0.97 | 0.88  | 0.73  | 1.12  | 0.92 | 0.88 | 0.93 |
| Median        | 0.62–0.97 | 0.78–0.12 | 0.67–0.18 | 0.56–0.89 | 0.66–1.57 | 0.31–1.52 | 0.69–0.07 | 0.72–1.13 |
| Range         | 0–11  | 0–15 | 0–12  | 0–7   | 0–15  | 0–12 | 0–9  | 0–15 |
| No. of samples| 259   | 351  | 259   | 215   | 87    | 47   | 262  | 284  |
| P-value       | 0.307 |      | 0.378 |      | 0.809 |      |      |      |

| Circulating tumour cells |       |      |       |       |       |      |      |      |
| Mean 95% CI   | 0.37  | 0.46 | 0.17  | 0.09  | 1.03  | 0.35 | 0.15 | 0.68 |
| Median        | 0.09–0.83 | 0.14–0.79 | 0.09–0.25 | 0–0.6–1.25 | 0–0.34 to 2.4 | 0.01–0.68 | 0.07–0.23 | 0.12–2.12 |
| Range         | 0–23  | 0–19 | 0–2   | 0–3   | 0–23  | 0–3  | 0–3  | 0–23 |
| No. of samples| 100   | 125  | 120   | 71    | 29    | 23   | 95   | 107  |
| P-value       | 0.026 |      | 0.197 |      | 0.201 |      |      |      |

Abbreviations: CI = confidence interval; ICC = immunocytochemistry; qRT-PCR = quantitative reverse transcription-PCR. *Statistically significant by one-way analysis of variance, Kruskal–Wallis or Mann–Whitney U-test. +Calculated as the percentage ratio of qRT-PCR CK19/ABL (0.1 or more is considered positive). **Detected by pan-CK antibody. #Samples were considered positive for circulating tumour cells when at least one cell was detected in 225 ml of blood by the CellSearch system.
tumours ($P = 0.032$), and the 291-bp cfDNA amplicon ($P = 0.033$) and the number of CTCs detected ($P = 0.026$) were both higher in post-menopausal women compared with pre-menopausal women (Table 5). However, there was extensive overlap in the value ranges between groups (Supplementary Table 1).

Relationships between different measures of MRD

In all, 41 out of the 64 patients on follow-up for primary breast cancer had both DTCs and CTC values measured at the same time. Circulating tumour cells were positively correlated with DTCs measured by ICC only ($P = 0.01$). This confirms our previous findings: 16 out of 19 (84%) high-risk primary breast cancer patients with DTCs present by ICC also had CTCs present (Slade et al., 2009). However, there was no statistically significant correlation between CTCs and DTCs measured by qRT-PCR (Supplementary Table 1).

Next, we investigated the relationship between cfDNA with DTCs and CTCs by analysing all the matched patient samples with bone marrow aspirates and blood samples performed at the same clinic visit or within a month of either sample being taken. A total of 168 matched bone marrow and blood samples had both DTC analysis (ICC and qRT-PCR) and qPCR measurement of 96- and 291-bp amplicons in cfDNA. In all, 8 out of 64 patients had no matched samples; the rest had both a mean and median of 3 per patient. Up to 179 matched samples were analysed for both CTCs and cfDNA measuring both 96-bp and 291-bp amplicons. Interestingly, an inverse correlation was found between the presence of DTCs as measured by CK19 qRT-PCR and the quantity of the 291-bp amplicon in cfDNA ($P = 0.0469$) (Figure 1B). The box plot demonstrates that those patients with DTCs (qRT-PCR) had lower median and inter-quartile ranges of the 291-bp cfDNA amplicon than the group with no DTCs present. No other correlations were evident.

DISCUSSION

Patients with a high risk of relapse based on tumour stage are more likely to have bone marrow DTCs at presentation (Braun et al., 2006), and these patients are more likely to demonstrate CTCs during follow-up (Slade et al., 2009). In addition, we and others have shown that cfDNA is higher in breast cancer patients than healthy controls (Gal et al., 2004; Huang et al., 2006). It is therefore important to establish the relationship between the presence of DTCs, CTCs and cfDNA levels during long-term follow-up.

Our results show that many patients with breast cancer continue to have evidence of MRD long after the completion of their adjuvant treatment despite the fact that they have no clinically evident recurrent disease. Measures of MRD designed to detect both living and non-viable cells such as CTC/DTG (ICC) and DTC (qRT-PCR) are intermittently present; their presence does not appear to indicate inevitable relapse. Higher levels of all MRD measures were more likely in patients who had an ER-negative, HER2-negative or triple-negative tumour and who had a higher grade of tumour although only values for the 96-bp cfDNA amplicon were significantly higher. Mean quantities of both amplicons (96 and 291 bp) in cfDNA were correlated with the number of lymph nodes positive at surgery ($P<0.0001$ and $P = 0.0248$, respectively). However, there was extensive overlap in the value ranges between groups for both cfDNA and the other measures of MRD suggesting that individual markers have limited prognostic value in the long-term follow-up of primary breast cancer patients (Supplementary Figures 1 and 2).

Of interest, cfDNA levels of the larger 291-bp amplicon appear inversely related to bone marrow dissemination as measured by qRT-PCR. At first glance, these results appear counter-intuitive. However, as an increase in larger sized fragments in cfDNA may result from dying micrometastases, evidence of viable DTCs as disclosed by an increase in bone marrow CK19 mRNA by qRT-PCR may be less likely to be found in patients in whom this is happening. Immunocytochemistry is not a good measure of viable micrometastases as the method detects both viable and non-viable cells. For example, a proportion of CTCs detected by the CellSearch system have expression of a marker of apoptosis indicating they are not all viable (Rossi et al., 2010). Cells that are viable are also more likely to have intact mRNA and so may explain why DTCs and CTCs measured by CK19 mRNA are a better predictive indicator of prognosis (Smith et al., 2000; Benoy et al., 2006; Farmen et al., 2008).

Although cfDNA is more informative, in that all blood samples contain measurable cfDNA, these results suggest that simple quantification of cfDNA alone is not a useful prognostic marker during the follow-up period and a panel of MRD markers may be required. For example, evidence that cfDNA is tumour derived using genomic technologies may provide more information about genomic alterations of clinical significance (Shaw et al., 2011).

Figure 1 The largely cancer specific 291-bp cfDNA amplicon in plasma is inversely related to the presence of DTCs (qRT-PCR CK1/9ABL ≥ 0.1%) in the bone marrow of primary breast cancer patients on follow-up. Comparison of (A) 96-bp cfDNA and (B) 291-bp cfDNA in patient samples grouped by the presence or absence of DTCs (ICC and qRT-PCR) and CTCs (CellSearch) in paired bone marrow aspirate and blood samples from 64 primary breast cancer patients on follow-up. The box plots show the median and inter-quartile ranges and the whiskers indicate 1.5 x the inter-quartile range (*statistically significant, Mann–Whitney non-parametric t-test).
The fluctuations over time may be an important determinant, along with combined investigations with DTCs and CTCs to provide further information on the viability and metastatic potential of these micrometastases.

Our data raise important questions regarding the issue of dormancy in breast cancer. We and others have shown that rare DTCs and CTCs can persist for many years after the end of breast cancer treatment (Meng et al, 2004; Slade et al, 2005, 2009). As the half-life of CTCs in the blood is estimated at 1–2 h (Meng et al, 2004), their presence may represent a dynamic balance between proliferation from micrometastatic niches and cell death. This turnover of residual cells is possibly one of the reasons why cancer-derived cfDNA (above the apoptotic limit) persists in blood for so long after diagnosis and treatment. A mathematical model of breast cancer dormancy has recently been developed and demonstrates that long-term breast cancer dormancy can be maintained by a small subset of growth-restricted, dangerous micrometastases (Willis et al, 2010). This theoretical model agrees with the observations described herein of the persistence of a low but steady number of CTCs and DTCs observed from growth-restricted and not necessarily relapse-inducing micrometastases. The additional measurement and molecular characterisation of cfDNA derived from dying tumour cells may help to distinguish these phases of dormancy.

In conclusion, the inverse correlation of DTCs (by qRT-PCR) in the bone marrow with the larger 291-bp amplicon in cfDNA indicates that the dormancy period of breast cancer is potentially characterised by two distinct phases: one in which measures of viable DTCs are absent, during which measures of cell death manifesting as cfDNA in the blood are evident, and an alternate phase, characterised by evidence of viable DTCs, but lower levels of cfDNA.

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Conflict of interest

The authors declare no conflict of interest.

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