Chronic Oral Infection with *Porphyromonas gingivalis* Accelerates Atheroma Formation by Shifting the Lipid Profile

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Abstract

**Background:** Recent studies have suggested that periodontal disease increases the risk of atherothrombotic disease. Atherosclerosis has been characterized as a chronic inflammatory response to cholesterol deposition in the arteries. Although several studies have suggested that certain periodontopathic bacteria accelerate atherogenesis in apolipoprotein E-deficient mice, the mechanistic link between cholesterol accumulation and periodontal infection-induced inflammation is largely unknown.

**Methodology/Principal Findings:** We orally infected C57BL/6 and C57BL/6.KOR-Apoeshl (B6.Apoeshl) mice with *Porphyromonas gingivalis*, which is a representative periodontopathic bacterium, and evaluated atherogenesis, gene expression in the aorta and liver and systemic inflammatory and lipid profiles in the blood. Furthermore, the effect of lipopolysaccharide (LPS) from *P. gingivalis* on cholesterol transport and the related gene expression was examined in peritoneal macrophages. Alveolar bone resorption and elevation of systemic inflammatory responses were induced in both strains. Despite early changes in the expression of key genes involved in cholesterol turnover, such as liver X receptor and ATP-binding cassette A1, serum lipid profiles did not change with short-term infection. Long-term infection was associated with a reduction in serum high-density lipoprotein (HDL) cholesterol but not with the development of atherosclerotic lesions in wild-type mice. In B6.Apoeshl mice, long-term infection resulted in the elevation of very low-density lipoprotein (VLDL), LDL and total cholesterol in addition to the reduction of HDL cholesterol. This shift in the lipid profile was concomitant with a significant increase in atherosclerotic lesions. Stimulation with *P. gingivalis* LPS induced the change of cholesterol transport via targeting the expression of LDL-receptor-related genes and resulted in the disturbance of regulatory mechanisms of the cholesterol level in macrophages.

**Conclusions/Significance:** Periodontal infection itself does not cause atherosclerosis, but it accelerates it by inducing systemic inflammation and deteriorating lipid metabolism, particularly when underlying hyperlipidemia or susceptibility to hyperlipidemias exists, and it may contribute to the development of coronary heart disease.

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Introduction

Coronary heart disease (CHD) is the leading cause of death in Japan and other developed countries. The major pathway underlying CHD pathology is atherosclerosis. Several risk factors for atherosclerosis have been identified, including smoking, hypertension, hyperglycemia, hypercholesterolemia and genetic factors. However, atherosclerosis can develop in the absence of these classic risk factors [1]. Recent epidemiological studies have suggested a link between atherosclerosis and infection/inflammation. Associations have been reported with *Chlamydia pneumoniae*, *Helicobacter pylori*, and cytomegalovirus [2,3], as well as with dental infections, particularly those associated with periodontitis [4,5].

Periodontitis is a chronic infectious disease initiated by a group of periodontopathic bacteria, such as *Porphyromonas gingivalis*. Periodontitis destroys the tooth-supporting tissues and leads to tooth loss if not adequately treated. Of the several hypotheses proposed to account for the effect of chronic infections on the development of atherosclerosis [6], one—the induction of low-grade systemic inflammation by periodontal infection—is particularly important. In fact, case-control and intervention studies [7,8] have clearly demonstrated that high-sensitivity C-reactive
atheromatous plaques, and the serum lipid profile became more lipid profile. Additionally, infection accelerated the development of in the aorta and liver, irrespective of abnormalities in the serum models.

of short-term infection models rather than chronic infection atherogenesis have not been elucidated, primarily because of the which periodontal infection affects tissues and organs related to affect and modify existing risk factors. However, the ways in investigate the mechanism by which periodontal infection may (HDL) cholesterol levels were decreased compared to those cholesterol levels were increased and high-density lipoprotein (LDL) periodontitis, total cholesterol and low-density lipoprotein (LDL) cholesterol levels were increased and high-density lipoprotein (HDL) cholesterol levels were decreased compared to those in healthy subjects [13,14,15]. Therefore, it is reasonable to investigate the mechanism by which periodontal infection may affect and modify existing risk factors. However, the ways in which periodontal infection affects tissues and organs related to atherogenesis have not been elucidated, primarily because of the limitations of the experimental design described above and the use of short-term infection models rather than chronic infection models.

Here, we compared short-term and long-term infections in normocholesterolemic and hyperlipidemic mice. We found that oral inoculation of P. gingivalis affected the gene expression profiles in the aorta and liver, irrespective of abnormalities in the serum lipid profile. Additionally, infection accelerated the development of atheromatous plaques, and the serum lipid profile became more proatherogenic in the presence of preexisting hyperlipidemia.

Results

Alveolar bone loss due to oral Infection with P. gingivalis

Gross visual inspection of P. gingivalis-infected and sham-infected wild-type mice under a stereoscopic microscope (Fig. 1A–D) revealed the alveolar bone pathology. The distance between the cement-enamel junction (CEJ) and the alveolar bone crest (ABC) in the mandibular molars (measured by micro-CT) was significantly different between the infected mice and the control mice. P. gingivalis infection significantly increased the distance between the CEJ and the ABC in the interdental region ($P<0.005$ compared to sham-infected mice; Fig. 1E). The effect of P. gingivalis infection tended to be greater in B6.Apoeshl mice compared with wild-type mice ($P<0.01$).

Systemic response to oral Infection with P. gingivalis

Oral infection with P. gingivalis induced a significant elevation in serum IL-6 levels during short-term infection, regardless of whether hyperlipidemia was present (Fig. 2A). During short-term infection, serum amyloid A (SAA) was increased in infected mice compared with those subjected to sham infection, but this difference did not reach a statistically significant level. Further elevation of IL-6 and SAA was observed during long-term infection, and significant differences were observed in both wild-type and B6.Apoeshl mice (Fig. 2B). The serum levels of P. gingivalis W83-specific IgG were significantly greater ($P<0.01$) in P. gingivalis-infected mice compared with sham-infected mice after short-term infection. In sham-infected mice, no production of antibodies against P. gingivalis was observed during either short-term or long-term infection periods. However, the infection did induce significantly greater antibody production during short-term infection. Further elevation of antibody levels was observed in response to long-term infection (Fig. 2C). There was no difference in antibody production observed between wild-type and B6.Apoeshl mice.

Deterioration of the serum lipid profile id associated with the infection period

In both mouse strains, oral infection with P. gingivalis had no significant effect on the serum lipid profile during short-term infection. In wild-type mice, only HDL cholesterol levels were decreased by long-term infection. In contrast, all cholesterol levels shifted toward atherogenic levels during long-term P. gingivalis infection in B6.Apoeshl mice. Total and LDL cholesterol levels were elevated and HDL cholesterol levels were decreased in P. gingivalis-infected mice compared with sham-infected mice (Table 1).

An increase in atherosclerosis is associated with P. gingivalis infection in B6.Apoeshl mice but not in wild-type mice

Analysis of aortic atherosclerosis demonstrated a significant increase in lesion area during the experimental period in the absence of infection in B6.Apoeshl mice, but not wild-type mice (short-term infection vs. long-term infection in sham-infected mice). P. gingivalis infection further increased the lesion size during each infection period, and atherosclerotic plaques comprised >40% of the total vessel area (Fig. 3B). A similar effect on the response to infection was observed in the aortic sinus lesion volume (Fig. 3C). Conversely, no apparent lesions were observed at any point during the experimental period in wild-type mice.

Changes in atherogenicity-related gene expression profiles in aortic tissue are associated with the infection period

Oral infection with P. gingivalis significantly increased the expression of Toll-like receptor (TLR) 2 (Fig. 4A) and TLR4 (Fig. 4B) in infected mice compared with sham-infected mice, regardless of whether hyperlipidemia was present. In wild-type mice, a longer bacterial burden induced a further elevation in gene expression, whereas the difference between sham-infected and P. gingivalis-infected mice was smaller in long-term infected B6.Apoeshl mice.

Chemokine (C-C motif) ligand 2 (Ccl2) and early growth response-1 (Egr1), both of which are vascular inflammation-related genes, were also significantly up-regulated in the aortic tissues of P. gingivalis-infected mice relative to sham-infected mice (Fig. 4C and 4D). There was no difference in the Egr1 expression pattern either between short-term infection and long-term infection or between wild-type and B6.Apoeshl mice. The duration of the infection period did not affect Ccl2 expression in sham-infected mice, and P. gingivalis infection consistently induced the up-regulation of gene expression.
Figure 1. Determination of alveolar bone loss (A–D). Three-dimensional (A and B) and two-dimensional (C and D) microcomputed tomography images of representative samples from sham-infected mice (A and C) and infected mice (B and D) are presented. The infected group (A) showed an increase in the distance from the cement-enamel junction (CEJ) to the alveolar bone crest compared with the sham-infected group (B). Two-dimensional micro-CT sections for the infected group (C) but not the sham-infected group (D) showed alveolar bone loss at the bifurcation.
Numerical analysis of alveolar bone loss between the control group and the infected group (E; N = 10 in each group) was performed. The distance between the cement-enamel junction and alveolar bone crest in each tooth root was determined from three-dimensional micro-CT images using image visualization software. Box plots present medians and the 25th and 75th percentiles as boxes and the 10th and 90th percentiles as whiskers.

Because of the importance of lipid metabolism in atherogenesis and the involvement of infection and inflammation in lipid metabolism, the gene expression levels of several cholesterol transport molecules liver X receptors (LXR) a, b and ATP-binding cassette A1 (ABCAC1) were examined. Both Lxrα and Lxrβ were down-regulated during short-term infection, and further down-regulation was observed during long-term infection in wild-type mice (Figs. 4E and 4F). In B6.Apoeshl mice, Lxrα showed changes similar to those in wild-type mice, whereas Lxrβ did not show any difference following infection. Aheal was down-regulated in P. gingivalis-infected mice, with a greater reduction being observed during long-term infection. There was no difference in Aheal expression between wild-type and B6.Apoeshl mice (Fig. 4G).

Changes in atherogenicity-related gene expression profiles in the liver are associated with the infection period

The expression levels of the inflammation-related markers Saa1/2 and Il6 were significantly up-regulated during P. gingivalis infection (Figs. 5A and 5B). The expression pattern showed differences between wild-type and B6.Apoeshl mice. Long-term infection tended to produce enhanced up-regulation of Saa1 in wild-type mice, whereas the difference in gene expression was smaller between sham-infected and P. gingivalis-infected B6.Apoeshl mice during long-term infection. No such difference was observed for Il6 expression. The lipid metabolism-related genes Lxrα and Lxrfwere down-regulated during short-term infection and further down-regulated under long-term infection, regardless of whether hyperlipidemia was present (Figs. 5C and 5D). The gene expression profile of Aheal showed a trend similar to that of the Lxrs, but a significant reduction was only observed during long-term infection (Fig. 5E). In addition, LDL receptor (Ldlr) and inducible degrader of the LDL receptor (Idol), another LXR target gene, were analyzed. The expression of Ldlr tended to be lower in P. gingivalis-infected mice compared with sham-infected mice (Fig. 5F). Consistent with Lxrs expressions, Idol expression was also decreased in P. gingivalis-infected mice compared with sham-infected mice. The change was greater during long-term infection in both wild-type and B6.Apoeshl mice (Fig. 5G).

In contrast to the Lxrs and Aheal, a marked difference in hydroxymethylglutaryl-coenzyme A reductase (Hmgcr) expression was observed between wild-type and B6.Apoeshl mice. Infection with P. gingivalis significantly increased the expression of Hmgcr as the bacterial burden in wild-type mice increased, while there was no change in the expression levels of this gene in B6.Apoeshl mice (Fig. 5H). A significant down-regulation was also observed in Adipor2, which is another antiatherogenic molecule (Fig. 5I).

Because triglyceride levels were significantly down-regulated during P. gingivalis infection, particularly during long-term infection, the expression levels of genes critical for the regulation of triglyceride synthesis were analyzed. Both fatty acid synthase (Fas) and sterol regulatory element-binding protein 1c (Srebf1) in mice expression levels were significantly down-regulated during long-term infection but not during short-term infection in wild-type mice (Figs. 3J and 5K). In B6.Apoeshl mice, Fas expression was significantly lower in infected mice compared to sham-infected mice, regardless of the length of the infection period. Srebf1 expression levels were significantly down-regulated only during long-term infection in this strain of mice.

TLR ligands induce phosphorylation of IRF3 in macrophages

As shown in Fig. 6, the TLR4 ligand E. coli LPS and TLR2 ligand Pam3CSK4 clearly induced the phosphorylation of IRF3 in macrophages. P. gingivalis LPS, which is known to act as an agonist for both TLR2 and TLR4, also induced IRF3 phosphorylation, although the activity of P. gingivalis LPS was much less potent than that of E. coli LPS.

Effects of P. gingivalis and E. coli LPS on cholesterol uptake and efflux from macrophages

Oral infection with P. gingivalis down-regulated the expression of the Lxrs and Aheal in both strains of mice. However, in contrast to the progression of the atherosclerotic lesions in B6.Apoeshl mice, no development of such lesions was observed in wild-type mice. These results suggested that either cholesterol efflux may be much more impaired by the infection in B6.Apoeshl mice than wild-type mice. Alternatively cholesterol efflux may be similar between wild-type and B6.Apoeshl mice, but cholesterol uptake may be higher in B6.Apoeshl mice. This would result in increased cholesterol accumulation and lesion progression. To distinguish between these two hypotheses, the effect of LPS treatment on cholesterol efflux by peritoneal macrophages was quantified in both strains of mice. We found that the apoA1-dependent cholesterol efflux induced by GW3965, a synthetic LXR agonist, was significantly inhibited by LPS treatment. The effect of GW3965 and inhibitory effect of LPS on GW3965-induced apoA1-dependent cholesterol efflux were similar between wild-type mice and B6.Apoeshl mice (Fig. 7A). Basal cholesterol uptake of B6.Apoeshl macrophages was much higher compared to that of wild-type macrophages. LPS treatment in the absence of GW3965 significantly increased cholesterol uptake in both wild-type macrophage and B6.Apoeshl macrophage (Fig. 7B). These results suggested that increased atherosclerotic lesions may be due to the increased uptake of cholesterol into B6.Aoeshl macrophages compared with wild-type macrophages, rather than differences in cholesterol efflux. The differences of these cellular characteristics were further manifested by the LPS stimulation.

Effects of P. gingivalis and E. coli LPS on LDLR expression in macrophages

The gene expression of LDLR in B6.Apoeshl macrophages was lower than that in wild-type macrophages and stimulation with P. gingivalis LPS and E. coli LPS increased the gene expression in both strains of mice. Although GW3965 suppressed the Ldls gene expression, either LPS abrogated the effect of GW3965. In contrast to the Ldls, the gene expression of Idol was higher in B6.Apoeshl macrophages compared with wild-type macrophages. While GW3965 increased the Idol expression in macrophages of both strains of mice, LPS suppressed the effect of GW3965 (Fig. 8A).

Culture in the sterol-depletion medium strongly induced LDLR protein expression in the macrophage and the addition of GW3965 potently decreased LDLR protein levels in macrophages. P. gingivalis LPS stimulation not only markedly attenuated the suppressive activity of GW3965 on the LDLR expression but also up-regulated the LDLR protein expression in B6.Apoeshl mice.
Both LPS enhanced LDLR expression in the absence of GW3965 (Fig. 8B).

Discussion

In both wild-type and B6.Apoeshl mice, oral infection with \textit{P. gingivalis} induced alveolar bone resorption, which is a characteristic feature of chronic periodontitis. Both strains of mice also showed elevated levels of serum antibodies against this bacterium, indicating a systemic immune response. Because periodontal tissue destruction is stimulated by inflammatory mediators, rather than by the bacteria themselves [16], the alterations in gene expression levels, the serum lipid profile and serum inflammatory mediators observed in this study could be a consequence of oral infection with \textit{P. gingivalis}. Despite the various systemic effects of oral infection in wild-type mice, the infection did

Figure 2. Effects of oral infection with \textit{P. gingivalis} on serum levels of interleukin (IL)-6 (A), serum amyloid A (SAA; B), and anti-\textit{P. gingivalis} antibody (C; \textit{N = 5 in each group}). All experiments were performed in triplicate wells for each condition and repeated at least twice. Representative data are shown. Box plots present medians and the 25th and 75th percentiles as boxes and the 10th and 90th percentiles as whiskers. Significant differences were observed between the infected group and the sham-infected group (* \(P<0.05\); ** \(P<0.01\), Mann-Whitney U-test). doi:10.1371/journal.pone.0020240.g002
oral infection was attenuated during long-term infection. *P. gingivalis* significantly up-regulated. However, the stimulatory effect of the expression of Egr1 was further elevated after long-term infection. In this context, previous studies have examined the role of periodontal infection using ApoE-deficient mice [9,10]. These studies demonstrated that either oral or intravenous infection using ApoE-deficient mice has the same effects as *C. pneumonia* infection has the same effects as *P. gingivalis* infection, aortic expression of both Tlr2 and Tlr4 was significantly enhanced in B6.Apoeshl mice but this effect was less pronounced than in wild-type mice. Because the expression of both Egr1 and Ccl2 is positively regulated by IL-6 in endothelial cells [18], increased serum IL-6 levels in B6.Apoeshl mice compared with wild-type mice would be expected to enhance the transcription of these molecules. Contrary to this assumption, IL-6 did not up-regulate these molecules. These results again suggest that abnormal lipid metabolism may have affected inflammatory responses in endothelial cells. The lipid metabolism–inflammatory response relationship was further highlighted by the observation of alveolar bone resorption. The effect of *P. gingivalis* infection tended to be greater in B6.Apoeshl mice than in wild-type mice. Further studies should be directed at understanding the role of lipid metabolism during the inflammatory response in blood vessels.

Additionally, infection and subsequent inflammation affected lipid metabolism-related gene expression. While the observed changes in Lxrα and Abca1 expression were similar between wild-type and B6.Apoeshl mice with both short-term and long-term infections, Lefβ gene expression was not affected by oral infection with *P. gingivalis* in B6.Apoeshl mice.

LXRs have been identified as central regulators of lipid homeostasis. LXRα is expressed primarily in the liver, intestine, adipose tissue, and macrophages, whereas LXRβ is expressed in many cell types. Endothelial cells also express functional LXRs, and treatment with LXR agonists can up-regulate ABC transporters. This result suggests that LXRs have an antiatherogenic effect in endothelial cells [19,20]. A number of studies have demonstrated that ABCA1 is an efficient exporter of cholesterol from macrophages and other cells, a major determinant of plasma HDL cholesterol levels and a potent cardioprotective factor [21]. The concomitant reduction of Lxrα and Abca1 seen in the livers of infected mice suggests that LXRs are functionally affected by oral infection and Ccl2 was significantly enhanced in B6.Apoeshl mice but this effect was less pronounced than in wild-type mice. Because the expression of both Egr1 and Ccl2 is positively regulated by IL-6 in endothelial cells [18], increased serum IL-6 levels in B6.Apoeshl mice compared with wild-type mice would be expected to enhance the transcription of these molecules. Contrary to this assumption, IL-6 did not up-regulate these molecules. These results again suggest that abnormal lipid metabolism may have affected inflammatory responses in endothelial cells. The lipid metabolism–inflammatory response relationship was further highlighted by the observation of alveolar bone resorption. The effect of *P. gingivalis* infection tended to be greater in B6.Apoeshl mice than in wild-type mice. Further studies should be directed at understanding the role of lipid metabolism during the inflammatory response in blood vessels.

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### Table 1. Serum lipid profile associated with infection period in wild-type and B6.Apoeshl mice.

|        | Wild-type |         |        | Long-term |         |
|--------|-----------|---------|--------|-----------|---------|
|        | Short-term | Long-term |        | Short-term | Long-term |
|        |          |          | P-value |          |          |
|        |          |          |          |          |          |
| Total  | 102.05±4.73 | 103.01±4.70 | 0.841   | 100.31±6.99 | 99.06±3.72 | 0.841 |
| CM     | 0.58±0.26 | 0.31±0.17 | 0.151   | 0.56±0.23 | 0.55±0.24 | 0.917 |
| VLDL   | 8.09±2.82 | 6.67±1.08 | 0.421   | 8.94±2.12 | 9.83±3.37 | 1     |
| LDL    | 12.72±1.51 | 11.88±0.97 | 0.309   | 11.54±2.37 | 11.88±2.17 | 0.548 |
| HDL    | 80.66±6.12 | 86.53±5.66 | 0.222   | 89.81±1.71 | 84.88±2.53 | 0.008 |
| TG     | 76.79±32.59 | 47.37±21.04 | 0.151   | 20.43±5.21 | 12.41±3.05 | 0.016 |
|        | B6.Apoeshl |         |        |          |          |
|        | Short-term | Long-term |        | Short-term | Long-term |
|        |          |          | P-value |          |          |
|        |          |          |          |          |          |
| Total  | 750.26±157.93 | 775.51±165.26 | 1       | 614.29±47.19 | 756.18±33.93 | 0.008 |
| CM     | 43.33±5.11 | 35.31±2.33 | 0.056   | 41.21±3.91 | 35.67±2.67 | 0.056 |
| VLDL   | 458.71±111.32 | 479.66±143.16 | 1       | 398.89±53.60 | 493.64±45.21 | 0.032 |
| LDL    | 249.38±46.71 | 252.22±34.07 | 1       | 179.09±29.93 | 255.51±27.64 | 0.016 |
| HDL    | 34.83±7.73 | 34.29±8.24 | 0.841   | 38.31±4.34 | 23.35±1.81 | 0.008 |
| TG     | 45.21±41.60 | 59.04±26.80 | 0.548   | 67.81±8.30 | 38.21±10.62 | 0.008 |

Values are indicated as mg/dL. CM indicates chylomicron; VLDL, very low density lipoprotein; LDL, low density lipoprotein; HDL, high density lipoprotein. N=5 in each group.
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Figure 3. Effects of oral infection with *P. gingivalis* on aortic atherosclerosis in wild-type and B6.Apoeshl mice. (A) Representative aortas from wild-type and B6.Apoeshl mice are depicted. (B) Aortic atherosclerosis expressed as a percentage of the total area (N=8 in each group). Box plots present medians and the 25th and 75th percentiles as boxes and the 10th and 90th percentiles as whiskers. Significant differences were observed between the infected group and the sham-infected group (*P*<0.01, Mann-Whitney U-test). (C) Representative aortic sinus cross sections from wild-type and B6.Apoeshl mice. Original magnifications, 40 x. doi:10.1371/journal.pone.0020240.g003
infection with *P. gingivalis*, possibly via crosstalk between LXR and TLR signaling. Castillo et al. [22] demonstrated that activation of TLR3 and TLR4 by microbial ligands blocks the induction of LXR target genes, including *Abca1*, in cultured macrophages and aortic tissues *in vivo*. Crosstalk between LXR and TLR signaling is mediated by interferon regulatory factor 3 (IRF3), a specific effector of TLR3/4 that inhibits the transcriptional activity of LXR. *P. gingivalis* contains diverse TLR ligands, including LPS, lipoproteins, and CpG DNA, which are recognized by TLR4, TLR2, and TLR9, respectively. Although the agonistic activity of *P. gingivalis* LPS is weaker than *E. coli* LPS [23], we found that *P. gingivalis* LPS could also induce phosphorylation of IRF3 in macrophages, albeit to a lesser extent than *E. coli* LPS (Fig. 6). Therefore, TLR signaling-mediated IRF3 activation may be a mechanism for the down-regulation of Lxr and *Abca1*.

The functional effect of the change in Lxr and *Abca1* expression levels on the shift in serum lipoproteins (particularly HDL cholesterol) was confirmed by the inhibition of cholesterol efflux by *P. gingivalis* LPS and *E. coli* LPS in peritoneal macrophages. GW3965-induced apoA1-dependent cholesterol efflux was completely abolished by stimulation with LPS. However, the rate of basal and GW3965-induced cholesterol efflux was lower in B6.Apoeshl mice compared with wild-type mice. On the other hand, cholesterol uptake was higher in B6.Apoeshl mice compared to wild-type mice and the uptake was enhanced by the LPS stimulation in both strains of mice.

Plasma cholesterol level is regulated by the LDLR and the expression of the LDLR is regulated at transcriptional and post translational levels. Recently, Zelcer et al. [24] found that LDLR expression is controlled by the LXR-Idol axis where LXR induces Idol, an E3 ubiquitin ligase that triggers LDLR degradation. Given that Lxr are down-regulated in the *P. gingivalis*-infected mice, it is probable that Idol expression is also suppressed in the infected mice compared to sham-infected mice and this change may relate to up-regulation of LDLR. In fact, the expression of the *Idol* gene was significantly lower in the liver of the infected mice and the change was more prominent during the long-term infection in B6.Apoeshl mice. Although the *ldlr* expression tended to be lower in *P. gingivalis*-infected mice compared to sham-infected mice, the difference was not statistically significant. These changes may have negative effect on Idol-mediated LDLR degradation resulting in the enhanced LDLR expression and subsequent decrease of plasma cholesterol. However, total and LDL cholesterol showed no change in wild-type mice but rather elevation in B6.Apoeshl mice during the long-term infection. *In vitro* stimulation of macrophages with LPS attenuated the GW3965-mediated suppression of the LDLR expression and the effect of *P. gingivalis* LPS was more potent than *E. coli* LPS. The reason for this paradox is unclear. However, Li et al. [25] showed that the acute inflammatory response decrease LDLR receptor expression in the liver. In contrast, the same inflammatory stimuli increased the LDLR expression in macrophages. Our results are consistent with these findings. In addition to Idol, the propionate convertase subtilisin-like kexin type 9 (PCSK9) is also implicated in the regulation of LDLR expression [26]. Although we have not determined whether PCSK9 contributes significantly to LDLR degradation and how its activity is affected by LPS, Feingold et al. [27] showed that inflammation stimulates PCSK9 expression leading to increased LDLR degradation but the underlying mechanisms remain elusive. Nevertheless, oral infection of *P. gingivalis* does affect the expression of the genes that regulate LDLR expression at protein level.

In wild-type mice, genes involved in lipid metabolism, but not the lipid profile itself, were found to be altered by short-term infection with *P. gingivalis*. By contrast, long-term infection reduced HDL cholesterol levels. The exact reasons for the discrepancy between gene expression levels and the lipid profile are not known. However, because the reduction in the expression of the *Abca1* gene, a rate-limiting factor in HDL biogenesis [28], was more pronounced during long-term infection than during short-term infection, it is possible that the suppression of *Abca1* during long-term infection was sufficiently robust to induce a decrease in HDL cholesterol. Few effects of short-term infection with *P. gingivalis* on total cholesterol and the size of the lipoproteins in plasma have been reported [10]. Therefore, it appears that longer-term and chronic infections are necessary to shift the plasma lipid profile toward an atherogenic state, despite the observation of proatherogenic changes in gene expression during short-term infection.

In addition to its effects on HDL cholesterol, *P. gingivalis* infection induced a significant down-regulation of serum triglycerides during long-term infection. *Fas* and *Sreb1*, which are genes involved in the regulation of triglyceride synthesis, were also down-regulated. Enzymes involved in triglyceride synthesis are transcriptionally regulated by SREBP-1c [29] and LxRs [30] in the liver. LxRs are also important regulators of FAS [31]. The conversion of glucose into triglycerides is nutritionally regulated, and both insulin and glucose signaling pathways are involved. Given that LxRs are key regulators of insulin and glucose signaling pathways that were down-regulated in infected mice of both strains, the resulting down-regulation of *Fas* and *Sreb1* and subsequent decrease in serum triglycerides could be due to *P. gingivalis* infection.

Severe periodontitis has been associated with a modest decrease in HDL and LDL cholesterol levels and a more robust increase in plasma triglycerides [32,33]. Although the effects of periodontal infection on the lipid profile of long-term infected B6.Apoeshl mice are slightly different from those seen in human periodontitis, particularly with regard to triglycerides, it is clear that infection does affect lipid metabolism. However, further studies are necessary to clarify the relationship between chronic infection and lipid metabolism. Nevertheless, the systemic inflammation induced by *P. gingivalis* oral infection is associated with a shift to a proatherogenic serum lipid profile that leads to elevated LDL cholesterol and decreased HDL cholesterol levels [34].

In summary, *P. gingivalis* is a representative periodontopathic bacterium that does not have the ability to induce atherosclerosis but can accelerate the progression of atherosclerosis by affecting blood vessel and liver function when hyperlipidemia is present. The results of the present study provide a potential mechanism for this bacterium’s contribution to atherogenesis and shed light on the underlying mechanism by which periodontitis affects atherosclerosis and subsequent CHD in humans.

### Materials and Methods

**Reagents**

GW3965 hydrochloride, SANDOZ 58-035, Apolipoprotein A-1, simvastatin sodium salt, mevalonic acid and lipoprotein-deficient fetal bovine serum (LPDS) were purchased from Sigma-Aldrich.
Figure 5. Comparison of the relative gene expression levels in the liver between the control group and the infected groups or between the short-term and long-term infected groups (N = 5 in each group). The relative quantity of mRNA was normalized to the relative quantity of GAPDH mRNA. The box plots present medians and the 25th and 75th percentiles as boxes and the 10th and 90th percentiles as whiskers. (* P < 0.05; ** P < 0.01, Mann-Whitney U-test) doi:10.1371/journal.pone.0020240.g005
analyzed by western blotting. Blot is a representative of three of Pam3CSK4 for 12 hrs. Phosphorylation of IRF3 cellular extracts was presence of an AnaeroPackTM (Mitsubishi Gas Chemical Co. Inc., Tokyo, Japan) for 48 hours at 37°C. Bacterial suspensions were prepared in phosphate-buffered saline (PBS) without Mg2+/Ca2+ using established growth curves and spectrophotometric analysis.

Figure 6. Induction of IRF3 phosphorylation in RAW 264.7 macrophages by TLR2 and TLR4 ligands. Cells were stimulated with 1.0 µg/ml of \textit{P. gingivalis} LPS, 0.1 µg/ml of \textit{E. coli} LPS or 0.1 µg/ml of Pam3CSK4 for 12 hrs. Phosphorylation of IRF3 cellular extracts was analyzed by western blotting. Blot is a representative of three independent experiments.

The number of CFUs was standardized by measuring optical density (600 nm).

**Oral Infection**

The murine experimental periodontitis model was developed according to Baker et al. [37] with slight modifications. The animals received sulfamethoxazole and trimethoprim at final concentrations of 700 µg/ml and 400 µg/ml, respectively, in water bottles \textit{ad libitum} for 10 days. This treatment was followed by 3 days without antibiotics. The experimental group was then infected. A total of 10^7 CFU of live \textit{P. gingivalis} suspended in 100 µl of PBS with 2% carboxymethyl cellulose (Sigma-Aldrich) was given to each mouse via a feeding needle. This suspension was given either 10 times (short-term infection) or 42 times (long-term infection) at 3-day intervals. The control group received the same pretreatment and was sham infected without the \textit{P. gingivalis}. Two days after the final treatment, the mice were fasted for 16 hours, and oral swabs were obtained and tested for the presence of \textit{P. gingivalis} as previously described [38]. Mice were then euthanized with CO2, and their tissues were removed.

Quantification of Alveolar Bone Loss

For quantitative three-dimensional (3-D) analysis of alveolar bone loss, mandibles were scanned by micro-computed tomography (micro-CT) (SkyScan 1174; SkyScan, Kontich, Belgium). The micro-CT settings were as follows: pixel size, 1024×1024; slice thickness, 6 µm; magnification, 6×; voltage, 50 kV; and electrical current, 0.08 mA. The 3-D views were constructed with the imaging software program CTAn ver.1.5.0 (SkyScan), and alveolar bone loss was analyzed with the software program CTvol ver.1.9.4 (SkyScan). The sagittal plane of the specimens was set parallel to the x-ray beam axis. The distance from the distal CEJ of the first molar in the middle of the lingual-distal root to the interdental ABC between the first and second molars was measured in the sagittal plane parallel to the lingual-distal root. The measurements of alveolar bone loss in mice were performed by blinded examiners (SM and DY).

Serum Lipoprotein, SAA, IL-6 and IgG Antibody Levels

Sera were obtained prior to euthanasia, and serum cholesterol and triglyceride profiles were analyzed at Skylight Biotech Inc. (Akita, Japan). Serum IL-6 and serum amyloid A (SAA) were measured using commercial ELISA kits (Thermo Scientific and Kamiya Biomedical, Seattle, WA, respectively). \textit{P. gingivalis}-specific IgG antibodies levels were measured by ELISA according to a previously described method [33].

Real-time PCR

Total RNA isolated from aorta, liver tissues and macrophages was extracted using an RNaseasy Mini kit and treated with DNase I (Qiagen, Germantown, MD) according to the manufacturer’s instructions. Aliquots of RNA were then reverse-transcribed to cDNA using random primers (Takara Bio Inc., Shiga, Japan) and M-MLV reverse transcriptase (Invitrogen, Carlsbad, CA). Primers and probes specific for real-time PCR were purchased from Applied Biosystems (Foster City, CA). Reactions were carried out in a 25-µl mixture in the ABI PRISM 7900HT Sequence Detection System (Applied Biosystems) using TaqMan Gene Expression Assays (Applied Biosystems) containing 900 nM primer and 250 nM probe. The reactions consisted of a 10-minute incubation at 95°C, followed by 40 cycles of a two-step amplification procedure consisting of annealing/extension at
60°C for 1 minute and denaturation for 15 seconds at 95°C. ABI PRISM SDS 2.0 software (Applied Biosystems) was used to analyze the standards and carry out the quantifications. The relative quantity of each mRNA was normalized to the relative quantity of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA.

Assessment of the Atherosclerotic Lesion Area

The extent of atherosclerotic lesions in the aortic tree was determined by en face quantification. The aorta was harvested, opened longitudinally, fixed with 4% PFA/PBS and stained with oil red O (Sigma-Aldrich). The stained area was analyzed using ImageJ 1.42 q (Wayne Rasband, National Institutes of Health).

Figure 7. Lipopolysaccharide from *P. gingivalis* and *E. coli* inhibit cholesterol efflux and enhance cholesterol uptake. (A) Peritoneal macrophages from wild-type and B6.Apoeshl mice were loaded with BODIPY-cholesterol and treated with vehicle, 1 μg/ml *P. gingivalis* LPS or 0.1 μg/ml *E. coli* LPS in the presence or absence of 1 μM GW3965. Data are presented as apoA1-dependent cholesterol efflux. (B) Cholesterol uptake was estimated as the total of cellular and effluxed cholesterol. The results are shown as the mean ± S.D. of three independent experiments. There were significant differences found between the wild-type and B6.Apoeshl macrophages (unpaired t-test; *P* < 0.05) and between control and LPS-treated macrophages in the absence (paired t-test; *P* < 0.05) or presence of GW3965 (paired t-test; †*P* < 0.05).
Figure 8. Lipopolysaccharide from \textit{P. gingivalis} and \textit{E. coli} alter the LDLR expression in macrophages. Peritoneal macrophages from wild-type and B6.Apoeshl mice were cultured in the sterol depletion medium for 8 hours and the effect of LPS was evaluated in the presence or absence of GW3965 following 18 hours of incubation. (A) Total RNA was extracted from the cells and gene expression of LDLR and Idol was analyzed by...
and expressed as a proportion of the lesion within the total vessel area. In addition, frozen sections of the proximal aorta were prepared [39]. Briefly, five sections (10 µm thick) per mouse, each separated by 100 µm, were stained with oil red O.

Western blots

For IRF-3 analysis, RAW 264.7 macrophages were seeded in a 24-well culture plate (TPP, Trasadingen, Switzerland) at a concentration of 1 x 10⁶ cells/ml in D-MEM supplemented with 1% penicillin/streptomycin. After 12 hours of incubation, the cells were stimulated with 50 ng/ml of Pam3CSK4 in medium.

The protein expression of LDLR was evaluated in peritoneal macrophages obtained from wild-type and B6.Apoeshl mice. The macrophages were collected 4 days after intraperitoneal injection of aged sterile 3% thio-glycollate and cultured in a D-MEM plate (TPP, Trasadingen, Switzerland) at a concentration of 1 x 10⁶/well in D-MEM supplemented with 10% LPDS, 5 µM simvastatin, and 100 µM mevalonic acid (sterol-depletion medium) for 8 hours and then treated with LPS from P. gingivalis (1 µg/ml) and E. coli LPS (0.1 µg/ml) in the presence or absence of GW3965 (1 µM) for 18 hours.

The cultured cells were washed twice with ice-cold PBS, and the protein was extracted using M-PER Mammalian Protein Extraction Reagent (Pierce Biotechnology) supplemented with a Halt Protease Inhibitor Cocktail Kit (Thermo Scientific) and Halt Phosphatase Inhibitor Cocktail (Thermo Scientific) according to the manufacturer’s instructions. Cell debris was pelleted by centrifugation at 12000 x g for 10 minutes at 4°C. The protein concentration in the supernatant was determined using a Pierce BCA Protein Assay kit (Thermo Scientific) according to the manufacturer’s instructions.

Twelve micrograms of each sample was solubilized using SDS sample buffer, separated by SDS-PAGE, transferred to a polyvinylidene difluoride membrane (Immobilon-P; Millipore Co., Bedford, MA), subjected to western blotting with each antibody, and developed with ECL using Lumi Vision PRO 400EX (Aisin, Aichi, Japan). For reprobing of GPDH, the membranes were washed 3 times with wash buffer (Tris-buffered saline containing 0.1% Tween-20 and 0.5% BSA) and subjected to western blotting with anti-GAPDH antibody, as described above.

Cholesterol influx and efflux in macrophages

Peritoneal macrophages from wild-type and B6.Apoeshl mice were collected 4 days after intraperitoneal injection of aged sterile 3% thio-glycollate and plated in a 12-well culture plate (TPP, Trasadingen, Switzerland) at a concentration of 1 x 10⁶/well in D-MEM containing 5% FBS and 1% penicillin/streptomycin, hereafter referred to as medium. Efflux assays were slightly modified from those described by Venkateswaran et al. [40]. Adherent cells were incubated with medium supplemented with ACAT inhibitor (58-035; 2 µg/ml) and BODIPY-cholesterol (10 µg/ml) for 24 hrs. The labeled cells were then treated with P. gingivalis LPS (1.0 µg/ml) or E. coli LPS (0.1 µg/ml) and/or GW3965 for 4 hrs. After cells were washed with HBSS, ApoA1 (10 µg/ml) was added to the culture and the cells were incubated for an additional 12 hrs. Following incubation, the supernatants were removed and the cells were dissolved using M-PER Mammalian Protein Extraction Reagent. The labeled cholesterol in the supernatants and cell fractions was determined using a Versa Fluor Fluorometer (Bio-Rad, Hercules, CA) equipped with the appropriate filters. Uptake of cholesterol in the cells was estimated as the total of released and cellular labeled cholesterol, whereas the ApoA1-dependent efflux of labeled cholesterol from the cells into the medium was determined by calculating the percentage of the total labeled cholesterol in the cells for each condition.

Statistical Analysis

Nonparametric data were evaluated using the Mann-Whitney U test with GraphPad Prism (GraphPad Software, Inc., La Jolla, CA) for two-group comparisons. A probability value (P) <0.05 was considered statistically significant.

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Author Contributions

Conceived and designed the experiments: KT TN KY. Performed the experiments: TM NT YA H. Miyashita H. Miyazawa SM KT. Analyzed the data: KT TN KY. Contributed reagents/materials/analysis tools: NT YA H. Miyashita SM KT. Wrote the paper: KY.

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