RNA interference is mediated by 21- and 22-nucleotide RNAs

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Double-stranded RNA (dsRNA) induces sequence-specific posttranscriptional gene silencing in many organisms by a process known as RNA interference (RNAi). Using a Drosophila in vitro system, we demonstrate that 21- and 22-nt RNA fragments are the sequence-specific mediators of RNAi. The short interfering RNAs (siRNAs) are generated by an RNase III–like processing reaction from long dsRNA. Chemically synthesized siRNA duplexes with overhanging 3′ ends mediate efficient target RNA cleavage in the lysate, and the cleavage site is located near the center of the region spanned by the guiding siRNA. Furthermore, we provide evidence that the direction of dsRNA processing determines whether sense or antisense target RNA can be cleaved by the siRNA–protein complex.

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target RNA can be cleaved by the siRNP endonuclease complex.

**Length requirements for processing of dsRNA to 21- and 22-nt RNA fragments**

Lysate prepared from *D. melanogaster* syncytial embryos recapitulates RNAi in vitro, providing a tool for biochemical analysis of the mechanism of RNAi [Tuschl et al. 1999; Zamore et al. 2000]. In vitro and in vivo analysis of the length requirements of dsRNA for RNAi has revealed that short dsRNA (<150 bp) are less effective than longer dsRNAs in degrading target mRNA [Ngo et al. 1998; Tuschl et al. 1999; Caplen et al. 2000; Hammond et al. 2000]. The reasons for reduction in mRNA degrading efficiency are not understood. We therefore examined the precise length requirement of dsRNA for target RNA degradation under optimized conditions in the *Drosophila* lysate. Three series of dsRNAs were synthesized and directed against firefly luciferase (Pp-luc) reporter RNA. The dual luciferase assay was used to monitor specific suppression of target RNA expression [Tuschl et al. 1999; Fig. 1A,B]. Specific inhibition of target RNA expression was detected for dsRNAs as short as 38 bp, but dsRNAs of 29–36 bp were not effective in this process. The effect was independent of the target position and the degree of inhibition of Pp-luc mRNA expression correlated with the length of the dsRNA; that is, long dsRNAs were more effective than short dsRNAs.

It has been suggested that the 21–23-nt RNA fragments generated by processing of dsRNAs are the mediators of RNA interference and cosuppression [Hamilton and Baulcombe 1999; Hammond et al. 2000; Zamore et al. 2000]. We therefore analyzed the rate of 21–23-nt fragment formation for a subset of dsRNAs ranging in size from 501 to 29 bp. Formation of 21–23-nt fragments in *Drosophila* lysate (Fig. 2) was readily detectable for 39–501 bp dsRNAs but was significantly delayed for the 29-bp dsRNA. This observation is consistent with a role of 21–23-nt fragments in guiding mRNA cleavage and provides an explanation for the lack of RNAi by 30-bp dsRNAs. The length dependence of 21–23 mer formation is likely to reflect a mechanism to prevent the undesired activation of RNAi by short intramolecular base-paired structures of cellular RNAs.

**Mapping of the cleavage sites on sense and antisense target RNAs**

Addition of dsRNA and 5′-capped target RNA to the *Drosophila* lysate results in sequence-specific degradation of the target RNA [Tuschl et al. 1999]. The target mRNA is only cleaved within the region of identity with the dsRNA, and many of the target cleavage sites are separated by 21–23 nt [Zamore et al. 2000]. Thus, the number of cleavage sites for a given dsRNA was expected to roughly correspond to the length of the dsRNA divided by 21. We mapped the target cleavage sites on a sense and an antisense target RNA that was 5′ radiolabeled at the cap [Zamore et al. 2000; Fig. 3A,B]. Stable 5′ cleavage products were separated on a sequencing gel, and the position of cleavage was determined by comparison with a partial RNase T1 and an alkaline hydrolysis ladder from the target RNA.

Consistent with the previous observation [Zamore et al. 2000], all target RNA cleavage sites were located within the region of identity to the dsRNA. The 39-bp dsRNA produced a strong and a weak (often hardly de-
was always located 7–10 nt downstream from the 5′ boundary of the region covered by the dsRNA (data not shown). This suggests that the point-of-target RNA cleavage can be determined by the end of the dsRNA and could imply that processing to 21–23mers starts from the ends of the duplex.

Cleavage sites on sense and antisense targets for the longer 111-bp dsRNA were much more frequent than anticipated, and most of them appear in clusters separated by 20–23 nt (Fig. 3A,B). As for the shorter dsRNAs, the first cleavage site on the sense target is 10 nt from the 5′ end of the region spanned by the dsRNA, and the first cleavage site on the antisense target is located 9 nt from the 5′ end of region covered by the dsRNA. It is unclear what causes this disordered cleavage, but one possibility could be that longer dsRNAs may not only get processed from the ends but also internally, or there are some specificity determinants for dsRNA processing that we do not yet understand. Some irregularities to the 21–23 nt spacing were also noted previously [Zamore et al. 2000].

dsRNA is processed to 21- and 22-nt RNAs by an RNase III–like mechanism

To understand better the molecular basis of dsRNA processing and target RNA recognition, we decided to analyze the sequences of the 21–23-nt fragments generated by processing of 39-, 52-, and 111-bp dsRNAs in the Drosophila lysate. We first examined the 5′ and 3′ termini of the RNA fragments. Periodate oxidation of gel-purified 21–23-nt RNAs followed by β-elimination indicated the presence of a terminal 2′ and 3′ hydroxyl [data not shown]. The 21–23mers were also responsive to alkaline phosphatase treatment, implying the presence of a 5′ terminal phosphate [data not shown]. The presence of 5′ phosphate and 3′ hydroxyl termini suggests that the dsRNA could be processed by an enzymatic activity similar to Escherichia coli RNase III [for reviews, see Dunn 1982; Nicholson 1999; Robertson 1982, 1990].

To directionally clone the 21–23-nt RNA fragments, 3′ and 5′ adapter oligonucleotides were ligated to the purified 21–23mers using T4 RNA ligase. The ligation products were reverse transcribed, PCR-amplified, concatemerized, cloned, and sequenced. Over 220 short RNAs were sequenced from dsRNA processing reactions of the 39-, 52-, and 111-bp dsRNAs [Fig. 4A]. We found the following length distribution: 1% 18 nt, 5% 19 nt, 12% 20 nt, 45% 21 nt, 28% 22 nt, 6% 23 nt, and 2% 24 nt. Sequence analysis of the 5′ terminal nucleotide of the processed fragments indicated that oligonucleotides with a 5′ guanosine were underrepresented. This bias was most likely introduced by T4 RNA ligase, which discriminates against 5′ phosphorylated guanosine as donor oligonucleotide (Romanii et al. 1982), no significant sequence bias was seen at the 3′ end. Many of the -21 nt fragments originating from the 3′ ends of the sense or antisense strand of the duplexes include 3′ nucleotides that are derived from untemplated addition of nucleotides during RNA synthesis using T7 RNA
Figure 3. Mapping of sense and antisense target RNA cleavage sites. (A) Denaturing gel electrophoresis of the stable 5' cleavage products produced by 1 h incubation of 10 nM sense or antisense RNA 32P-labeled at the cap with 10 nM dsRNAs of the p133 series in Drosophila lysate. Length markers were generated by partial nuclease T1 digestion and partial alkaline hydrolysis (OH) of the cap-labeled target RNA. The regions targeted by the dsRNAs are indicated as black bars on both sides. The 20-23-nt spacing between the predominant cleavage sites for the 111-bp dsRNA is shown. The horizontal arrow indicates unspecific cleavage not caused by RNAi. (B) Position of the cleavage sites on sense and antisense target RNAs. The sequences of the capped 177-nt sense and 180-nt antisense target RNAs are represented in antiparallel orientation such that complementary sequence are opposing each other. The region targeted by the different dsRNAs are indicated by differently colored bars positioned between sense and antisense target sequences. Cleavage sites are indicated by circles (large circle for strong cleavage, small circle for weak cleavage). The 32P-radiolabeled phosphate group is marked by an asterisk.
polymerase. Interestingly, a significant number of endogenous Drosophila ∼21-nt RNAs were also cloned, some of them from LTR and non-LTR retrotransposons (data not shown). This is consistent with a possible role for RNAi in transposon silencing (Ketting et al. 1999; Tabara et al. 1999).

The ∼21-nt RNAs appear in clustered groups (Fig. 4A) that cover the entire dsRNA sequences. For the 39-bp dsRNA, two clusters of ∼21-nt RNAs were found from each dsRNA-constituting strand [including overhanging 3’ ends]. Only one of the clusters from each strand can be correlated with a strong cleavage hot spot on the target sense or antisense RNA (Fig. 3A,B), indicating that dsRNA processing produced primarily two functional small RNAs originating from the 3’ ends of the duplex. Perhaps the ∼21-nt RNAs are present in double-stranded form in the endonuclease complex, but only one of the strands can be used for target RNA recognition and cleavage.

The ∼21-mer clusters for the 52- and 111-bp dsRNA...
are less well defined when compared to the 39-bp dsRNA. The clusters are spread over regions of 25–30 nt most likely representing several distinct subpopulations of 21-nt duplexes and, therefore, guiding target cleavage at several nearby sites. These cleavage regions are still predominantly separated by 20–23-nt intervals. The rules determining how dsRNA can be processed to ∼30 nt fragments are not yet understood, but it was observed previously that the ∼21–23-nt spacing of cleavage sites could be altered by a run of uridines (Zamore et al. 2000). The specificity of dsRNA cleavage by E. coli RNase III appears to be mainly controlled by antideterminants, that is, excluding some specific base pairs at given positions relative to the cleavage site (Zhang and Nicholson 1997). The sequence dependence of dsRNA processing and target RNA cleavage in RNAi needs to be examined further.

To test whether sugar-, base-, or cap-modification were present in processed 21-nt RNA fragments, we incubated radiolabeled 505-bp Pp-luc dsRNA in lysate for 1 h, isolated the ∼21-nt products, and digested it with P1 or T2 nuclease to mononucleotides. The nucleotide mixture was then analyzed by two-dimensional thin-layer chromatography (Fig. 4B). None of the four ribonucleotides were modified, as indicated by P1 or T2 digestion. We have previously analyzed adenosine to inosine conversion in the ∼21-nt fragments (after a 2 h incubation) and detected a small extent (∼0.7%) deamination (Zamore et al. 2000). Shorter incubation in lysate (1 h) reduced this inosine fraction to barely detectable levels. RNase T2, which cleaves 3’ of the phosphodiester linkage, produced nucleoside 3’-phosphate and nucleoside 3’,5’-diphosphate, thereby indicating the presence of a 5’-terminal monophosphate. All four nucleoside 3’,5’-diphosphates were detected and indicate that the inter-nucleotidic linkage was cleaved with little or no sequence specificity for the residue 3’ to the cleavage site, and according to the sequence analysis of the cloned ∼21-nt fragments, no significant sequence bias was observed for the residue 5’ of the cleavage site. In summary, the ∼21-nt fragments are unmodified and were generated from dsRNA such that 5’-monophosphates and 3’-hydroxyls were present at the 5’-ends. Analysis of the products of dsRNA processing indicated that the ∼21-nt fragments are generated by a reaction with all the characteristics of an RNase III cleavage reaction (Dunn 1982; Robertson 1982, 1990; Nicholson 1999).

**Synthetic 21- and 22-nt RNAs mediate target RNA cleavage**

We chemically synthesized 21- and 22-nt RNAs, identical in sequence to some of the cloned ∼21-nt fragments, and tested them for their ability to mediate target RNA degradation [Fig. 5A–C]. The 21- and 22-nt RNA duplexes were incubated at 100 nM concentrations in the lysate, a 10- to 20-fold higher concentration than the 52-bp control dsRNA. Under these conditions, target RNA cleavage was readily detectable. Tenfold reduced concentrations of 21- and 22-nt duplexes (10 nM) still caused target RNA cleavage but to a smaller extent (data not shown). Increasing the duplex concentration from 100 to 1000 nM, however, did not further increase target degradation (data not shown), perhaps because of a limiting protein factor within the lysate. Single-stranded sense or antisense 21- and 22-nt RNAs at 100 nM concentration did not affect target RNA expression, most likely because single-stranded RNAs are not stable in the lysate and degraded to mononucleotides within minutes (data not shown). We also found that preannealing of the short antisense RNAs to the target mRNA before the addition of lysate had no effect on target RNA expression (data not shown).

RNase III makes two staggered cuts in both strands of the dsRNA, leaving a 3’ overhang of 2 nt. The 21- and 22-nt RNA duplexes with 2- or 3-nt overhanging 3’ ends (duplexes 1, 4, 6) were more efficient in reducing the target RNA expression than the corresponding blunt-ended dsRNAs [duplexes 2, 5, 7] or the dsRNA with 4 nt overhang [duplex 3]. Duplexes 6 and 7 are generally more effective for RNAi than duplexes 1–5, probably as a consequence of target RNA accessibility (because of RNA self-structure or RNA-coating proteins) or because of sequence-specific effects in the reconstitution of the RNA-degrading complexes. The interference effects determined in the translation-based assay [Fig. 5B] correlate well with the intensity of the cleavage bands observed by targeting 5’ radiolabeled model substrates with the 21- and 22-nt RNA duplexes [Fig. 5C]. Together, these data suggest that 2–3 nt of overhanging 3’ ends are beneficial for reconstitution of the RNAi nuclease complex and may be required for high-affinity binding of the short RNA duplex to the protein components. A 5’ terminal phosphate, although present after dsRNA processing, was not required to mediate target RNA cleavage and was absent from the short synthetic RNAs.

The synthetic 21- and 22-nt duplexes guided cleavage of sense as well as antisense targets within the region covered by the short duplex. This is interesting, considering that the presumably base-paired clusters of ∼21-nt fragments derived from the 39-bp dsRNA (Fig. 2) can only be correlated to a predominant cleavage site on either the sense or the antisense target but not both. We interpret this result by suggesting that only one of two strands present in the ∼21-nt duplex is able to guide target RNA cleavage and that the orientation of the ∼21-nt duplex in the nuclease complex is determined by the initial direction of dsRNA processing. It also implies that the processed short RNAs are present in a tight ribonucleoprotein complex and do not dissociate and rebind during the time scale of the experiment. The presentation of an already perfectly processed ∼21-nt duplex to the in vitro system, however, does allow formation of the active sequence-specific nuclease complex with two possible orientations of the symmetric RNA duplex. This results in cleavage of sense as well as antisense targets within the region of identity with the ∼21-nt RNA duplex.

The target cleavage site is located near center of the region covered by the 21- or 22-nt RNAs, 11 or 12 nt

RNA interference is mediated by 21- and 22-nt RNAs...
Synthetic 21- and 22-nt RNAs mediate target RNA cleavage. (A) Graphic representation of control 52-bp dsRNA and synthetic 21- and 22-nt dsRNAs. The sense strand of 21- and 22-nt short interfering RNAs (siRNAs) is shown in blue, the antisense strand in red. The sequences of the siRNAs were derived from the cloned fragments of 52- and 111-bp dsRNAs (Fig. 4A), except for the 22-nt antisense strand of duplex 5. The siRNAs in duplexes 6 and 7 were unique to the 111-bp dsRNA-processing reaction. The two 3’ overhanging nucleotides indicated in green are present in the sequence of the synthetic antisense strand of duplexes 1 and 3. Both strands of the control 52-bp dsRNA were prepared by in vitro transcription, and a fraction of transcripts may contain untemplated 3’ nucleotide addition. The target RNA cleavage sites directed by the siRNA duplexes are indicated as orange circles (see legend to Fig. 4A) and were determined as shown in Figure 5B. (B) RNA interference assay. To evaluate the efficiency of target RNA degradation, control 52-bp dsRNA (5 nM) or 21- and 22-nt RNA duplexes 1–7 (100 nM) targeting full-length Pp-luc mRNA were tested in the translation-based RNAi assay as described in Figure 1B. The relative luminescence of target to control luciferase normalized to a buffer control (buf) is plotted; error bars indicate standard deviations calculated from at least two independent experiments. (C) Position of the cleavage sites on sense and antisense target RNAs. The target RNA sequences are as described in Figure 3B. Control 52-bp dsRNA (10 nM) or 21- and 22-nt RNA duplexes 1–7 (100 nM) were incubated with target RNA at 25°C for 2.5 h in Drosophila lysate. The stable 5’ cleavage products were resolved on the gel. The cleavage sites are indicated in Figure 5A. The region targeted by the 52-bp dsRNA or the sense [s] or antisense [as] strands are indicated by the black bars to the side of the gel. The cleavage sites are all located within the region of identity of the dsRNAs. For precise determination of the cleavage sites of the antisense strand, a lower percentage gel was used (data not shown).
RNA interference is mediated by 21- and 22-nt RNAs

downstream of the first nucleotide that is complementary to the 21- or 22-nt guide sequence [Fig. 4A,B]. Displacing the sense strand of a 22-nt duplex by two nucleotides (cf. duplexes 1 and 3 in Fig. 5A) displaced the cleavage site of only the antisense target by two nucleotides. Displacing both sense and antisense strands by two nucleotides shifted both cleavage sites by two nucleotides (cf. duplexes 1 and 4). We predict that it will be possible to design a pair of 21- or 22-nt RNAs to cleave a target RNA at almost any given position.

The specificity of target RNA cleavage guided by 21- and 22-nt RNAs appears exquisite, as no cleavage sites are detected outside of the region of complementarity to the 21- and 22-nt RNAs [Fig. 5C]. It should, however, be noted that the nucleotides present in the 3' extension of the 21- and 22-nt RNA duplex may contribute less to substrate recognition than the nucleotides near the cleavage site. This is based on the observation that the 3'-most nucleotide of the antisense strand of active duplexes 1 or 3 [Fig. 5A] is not complementary to the target. A detailed analysis of the specificity of RNAi can now be readily undertaken using synthetic 21- and 22-nt RNAs.

On the basis of the evidence that synthetic 21- and 22-nt RNAs with overhanging 3' ends mediate RNA interference, we propose to name the ~21-nt RNAs short interfering RNAs, or siRNAs, and the respective RNA-protein complex a small interfering ribonucleoprotein particle, or siRNP.

3' overhangs of ~20 nt on short dsRNAs inhibit RNAi

We have also analyzed dsRNAs with 17–20 nt overhanging 3' ends that were less potent than blunt-ended dsRNAs [data not shown]. The inhibitory effect of long 3' ends was particularly pronounced for dsRNAs <100 bp. The effect was not caused by imperfect dsRNA formation, based on native gel analysis [data not shown]. We tested if the inhibitory effect of long overhanging 3' ends could be used as a tool to initiate dsRNA processing at only one of the two ends of a short RNA duplex.

We synthesized four combinations of the 52-bp model dsRNA, blunt-ended, 3' extension only on the sense strand, the 3' extension only on the antisense strand, and the double 3' extension on both strands and mapped the target RNA cleavage sites and monitored ~21-nt formation after incubation in lysate [Fig. 6A–C]. The first and predominant cleavage site of the sense target was lost when the 3' end of the antisense strand of the duplex was extended, and the strong cleavage site of the antisense target was lost when the 3' end of sense strand of the duplex was extended. Extending the 3' ends on both strands rendered the 52-bp dsRNA virtually inactive. These observations correlate with the formation of ~21-nt fragments from blunt-ended dsRNAs or dsRNAs with only one 3' extension and the absence of ~21-nt fragments when both 3' ends of the duplex are extended [Fig. 6C]. One explanation for the dsRNA inactivation by ~20-nt 3' extensions could be the association of single-stranded RNA-binding proteins that could interfere with the association of one of the dsRNA-processing factors at this end. This is supported by the significantly longer persistence of the double 3' extended dsRNA in the lysate [Fig. 6C]. Together, these results are consistent with our model where only one of the strands of the siRNA duplex in the assembled siRNP is able to guide target RNA cleavage. The orientation of the strand that guides RNA cleavage is defined by the direction of the dsRNA processing reaction. A block at the 3' end of the sense strand will only permit dsRNA processing from the opposing 3' end of the antisense strand. This, in turn, generates siRNPs in which only the antisense strand of the siRNA duplex is able to guide sense target RNA cleavage. The same is true for the reciprocal situation. The less pronounced inhibitory effect of long 3' extensions in the case of longer dsRNAs (≥500 bp, data not shown) suggests that long dsRNAs may also contain internal dsRNA-processing signals or may get processed cooperatively because of the association of multiple cleavage factors.

A model for dsRNA-directed mRNA cleavage

The new biochemical data update the model for how dsRNA targets mRNA for destruction [Fig. 7]. Based on the 21–23-nt length of the processed RNA fragments, it had already been speculated that an RNase III–like activity may be involved in RNAi [Bass 2000]. Double-stranded RNA is first processed to short RNA duplexes of predominantly 21 and 22 nt in length and with staggered 3' ends similar to an RNase III–like reaction [Dunn 1982; Robertson 1982; Nicholson 1999]. This hypothesis is further supported by the presence of 5' phosphates and 3' hydroxyls at the termini of the siRNAs [Fig. 4B] as observed in RNase III reaction products [Dunn 1982, Nicholson 1999].

Bacterial RNase III and the eukaryotic homologs Rnt1p in Saccharomyces cerevisiae and Pac1p in Schizosaccharomyces pombe have been shown to function in processing of ribosomal RNA as well as snRNA and snoRNAs [see, for example, Chanfreau et al. 2000]. Little is known about the biochemistry of RNase III homologs from plants, animals, or human. Two families of RNase III enzymes have been identified predominantly by database-guided sequence analysis or cloning of cDNAs. The first RNase III family is represented by the 1327-amino-acid D. melanogaster protein drosha [accession no. AF116572]. The carboxyl terminus is composed of two RNase III domains and one dsRNA-binding domain, and the amino terminus is of unknown function. Close homologs are also found in C. elegans [accession no. AF160248] and human [accession no. AF189011; Filipov et al. 2000, Wu et al. 2000]. The drosha-like human RNase III was recently cloned and characterized [Wu et al. 2000]. The gene is ubiquitously expressed in human tissues and cell lines, and the protein is localized in the nucleus and the nucleolus of the cell. Based on results inferred from antisense inhibition studies, a role of this protein for rRNA processing was suggested. The second class is represented by the C. elegans gene K12H4.8 [accession no. S44849] coding for an 1822-amino-acid pro-
Figure 6. Long 3’ overhangs on short dsRNAs inhibit RNAi. (A) Graphic representation of 52-bp dsRNA constructs. The 3’ extensions of sense and antisense strand are indicated in blue and red, respectively. The observed cleavage sites on the target RNAs are represented as orange circles analogous to Figure 4A and were determined as shown in B. (B) Position of the cleavage sites on sense and antisense target RNAs. The target RNA sequences are as described in Figure 3B. DsRNA (10 nM) was incubated with target RNA at 25°C for 2.5 h in Drosophila lysate. The stable 5’ cleavage products were resolved on the gel. The major cleavage sites are indicated with a horizontal arrow and are also represented in A. The region targeted by the 52-bp dsRNA is represented as a black bar at both sides of the gel. (C) Processing of 52-bp dsRNAs with different 3’ extensions. Internally 32P-labeled dsRNAs (5 nM) were incubated in Drosophila lysate, and reaction aliquots were analyzed at the indicated time points. An RNA size marker [M] has been loaded in the left lane, and the fragment sizes are indicated. Double bands at time zero are caused by incompletely denatured dsRNA.
tein. This protein has an amino-terminal RNA helicase motif, which is followed by two RNase III catalytic domains and a dsRNA-binding motif, similar to the *drosha* RNase III family. There are close homologs in *S. pombe* (accession no. Q09884), *Arabidopsis thaliana* (accession no. AF187317), and *D. melanogaster* (accession no. AE003740), and human (accession no. AB028449) (Jacobsen et al. 1999; Filippov et al. 2000; Matsuda et al. 2000). It is tempting to speculate that the K12H4.8 RNase III/helicase is the likely candidate to be involved in RNAi.

Genetic screens in *C. elegans* identified *rde-1* and *rde-4* as essential for activation of RNAi, without an effect on transposon mobilization or cosuppression (Tabara et al. 1999; Dernburg et al. 2000, Grishok et al. 2000). It is tempting to speculate that the K12H4.8 RNase III/helicase is the likely candidate to be involved in RNAi.

Processing of dsRNA to siRNA duplexes appears to start from the ends of both blunt-ended dsRNAs or dsRNAs with short (1–5 nt) 3’ overhangs and proceeds in ~21–23-nt steps. Long (~20 nt) 3’ staggered ends on short dsRNAs suppress RNAi, possibly through interaction with single-stranded RNA-binding proteins. The suppression of RNAi by single-stranded regions flanking short dsRNA and the reduced rate of siRNA formation from short 30-bp dsRNAs may explain why structured regions within mRNAs do not lead to activation of RNAi. In *C. elegans*, it was observed recently that injection of a 26-bp dsRNA could trigger RNAi of the *unc-22* gene; however, a >250-fold higher concentration of 26-bp dsRNA was necessary compared to an 81-bp dsRNA control (Parrish et al. 2000). It is conceivable that siRNA production from the 26-bp dsRNA was rate limiting and was compensated for by increasing the concentration of 26-bp dsRNA.

In our model, we presume that the dsRNA-processing proteins or a subset of them remain associated with the siRNA duplex and preserve its orientation, as determined by the direction of the dsRNA processing reaction. Only the siRNA sequence associated with the blue protein is able to guide target RNA cleavage. The endonuclease complex is referred to as small interfering ribonucleoprotein complex or siRNP. It is presumed here that the endonuclease that cleaves the dsRNA may also cleave the target RNA, probably by temporarily displacing the passive siRNA strand not used for target recognition. The target RNA is then cleaved in the center of the region recognized by the sequence-complementary guide siRNA. Because the cleavage site is displaced by 10–12 nt relative to the dsRNA processing site, a conformational rearrangement or a change in the composition of an siRNP must occur before target RNA cleavage.

RNA interference is mediated by 21- and 22-nt RNAs
Elbashir et al.

vation in *C. elegans* that certain chemical modifications (e.g., 2′-aminouridine, 2′-deoxythymidine, or 5-iodouridine) incorporated into dsRNA are well tolerated at the sense, but not the cleavage-guiding antisense, strand [Parrish et al. 2000].

The finding that synthetic 21- and 22-nt siRNA duplexes can be used for efficient mRNA degradation demonstrates that the targeting step can be uncoupled from the dsRNA-processing step. This raises the prospects of using siRNA duplexes as new tools for sequence-specific regulation of gene expression in functional genomics as well as biomedical studies. The siRNAs may be effective in mammalian systems, where long dsRNAs cannot be used because they activate the dsRNA-dependent protein kinase [PKR] response [Clemens 1997]. As such, the siRNA duplexes may represent a new alternative to antisense or ribozyme therapeutics.

Materials and methods

In vitro RNAi

In vitro RNAi and lysate preparations were performed as described previously [Tuschl et al. 1999; Zamore et al. 2000] using a final concentration of 0.03 mg/mL creatine kinase in the RNAi reaction. It is critical to use freshly dissolved creatine at a final concentration of 0.03 mg/mL creatine kinase in the optimal ATP regeneration. The RNAi translation assays [Fig. 1] were performed with dsRNA concentrations of 5 nM and an extended preincubation period at 25°C for 15 min before the addition of in vitro transcribed, capped, and polyadenylated *Pp*-luc and *Rr*-luc reporter mRNAs. The incubation was continued for 1 h, and the relative amount of *Pp*-luc and *Rr*-luc protein was analyzed using the dual luciferase assay [Promega] and a Monolight 3010C luminometer [PharMingen].

RNA synthesis

Standard procedures were used for in vitro transcription of RNA from PCR templates carrying T7 or SP6 promoter sequences (see, for example, Tuschl et al. 1999). Synthetic RNA was prepared using Expedite RNA phosphoramidites [Proligo]. The 3′-labeled *Pp*-luc plasmid (pGEM-luc sequence; Tuschl et al. 1999) or the respective ratio for radiolabeled nucleoside triphosphates other than UTP. Labeling of the cap of the target RNAs was performed as described previously [Zamore et al. 2000]. The target RNAs were gel purified after cap labeling.

Cleavage site mapping

Standard RNAi reactions were performed by preincubating 10 nM dsRNA for 15 min followed by addition of 10 nM capped target RNA. The reaction was stopped after a further 2-h incubation [Fig. 3A] or 2.5-h incubation [Figs. 5C, 6B] by proteinase K treatment [Tuschl et al. 1999]. The samples were then analyzed on 8% or 10% sequencing gels. The 21- and 22-nt synthetic RNA duplexes were used at 100 nM final concentration [Fig. 5B,C].

Cloning of ~21-nt RNAs

The 21-nt RNAs were produced by incubation of radiolabeled dsRNA in *Drosophila* lysate in absence of target RNA [200 µL reaction, 1 h incubation, 50 nM dsP111, or 100 nM dsP52 or dsP93]. The reaction mixture was subsequently treated with proteinase K [Tuschl et al. 1999], and the dsRNA-processing products were separated on a denaturing 15% polyacrylamide gel. A band, including a size range of at least 18–24 nt, was excised and then eluted into 0.3 M NaCl overnight at 4°C in siliconized tubes. The RNA was recovered by ethanol precipitation and then dephosphorylated (30 µL reaction, 30°C, 30 min, 10 U alkaline phosphatase, Roche). The reaction was stopped by phenol/chloroform extraction, and the RNA was ethanol precipitated. The 3′-labeled oligonucleotide [pUUUaaccgc cactctctc: uppercase, RNA2; lowercase, DNA; p, phosphate; x, 4-hydroxymethylbenzyl] was then ligated to the dephosphorylated ~21-nt RNA [20 µL reaction, 37°C, 30 min, 5 µL 3′-adapter, 50 mM Tris-HCl at pH 7.6, 10 mM MgCl2, 0.2 mM ATP, 0.1 mg/mL acetylated BSA, 15% DMSO, 25 U T4 RNA ligase; Amersham-Pharmacia] (Pan and Uhlenbeck 1992). The ligation reaction was stopped by the addition of an equal volume of 8 M urea/50 mM EDTA stopmix and directly loaded on a 15% gel. The ligation products were separated by denaturing gel electrophoresis in 1× TAE-buffer. The 52-bp dsRNAs with the 17-nt and 20-nt 3′ overhangs [Fig. 6] were annealed by incubating at 95°C for 1 min and then rapidly cooled to 70°C and followed by slow cooling to room temperature over a 3-h period [50 µL annealing reaction, 1 µM strand concentration, 300 mM NaCl, 10 mM Tris-HCl at pH 7.5]. The dsRNAs were then phenol/chloroform extracted, ethanol precipitated, and dissolved in lysate buffer.

Transcription of internally 32P-radiolabeled RNA used for dsRNA preparation [Figs. 2, 4] was performed using 1 mM ATP, CTP, GTP, 0.1 or 0.2 mM UTP, and 0.2–0.3 µM [α-32P]UTP (3000 Ci/mmol) or the respective ratio for radiolabeled nucleoside triphosphates other than UTP. Labeling of the cap of the target RNAs was performed as described previously [Zamore et al. 2000]. The target RNAs were gel purified after cap labeling.

Two different PCR products that only contained a single promoter sequence.

DsRNA annealing was carried out using a phenol/chloroform extraction. Equimolar concentration of sense and antisense RNA [50 nM to 10 µM, depending on the length and amount available] in 0.3 M NaOAc [pH 6] were incubated at 90°C for 30 sec and then extracted at room temperature with an equal volume of phenol/chloroform and followed by a chloroform extraction to remove residual phenol. The resulting dsRNA was precipitated by addition of 2.5–3 volumes of ethanol. The pellet was dissolved in lysis buffer [100 mM KCl, 30 mM HEPES-KOH at pH 7.4, 2 mM MgOAc2], and the quality of the dsRNA was verified by standard agarose gel electrophoresis in 1× TAE-buffer. The 52-bp dsRNAs with the 17-nt and 20-nt 3′ overhangs were annealed by incubating at 95°C for 1 min and then rapidly cooled to 70°C and followed by slow cooling to room temperature over a 3-h period [50 µL annealing reaction, 1 µM strand concentration, 300 mM NaCl, 10 mM Tris-HCl at pH 7.5]. The dsRNAs were then phenol/chloroform extracted, ethanol precipitated, and dissolved in lysis buffer.

Transcription of internally 32P-radiolabeled RNA used for dsRNA preparation [Figs. 2, 4] was performed using 1 mM ATP, CTP, GTP, 0.1 or 0.2 mM UTP, and 0.2–0.3 µM [α-32P]UTP (3000 Ci/mmol) or the respective ratio for radiolabeled nucleoside triphosphates other than UTP. Labeling of the cap of the target RNAs was performed as described previously [Zamore et al. 2000]. The target RNAs were gel purified after cap labeling.

Cloning of ~21-nt RNAs

The 21-nt RNAs were produced by incubation of radiolabeled dsRNA in *Drosophila* lysate in absence of target RNA [200 µL reaction, 1 h incubation, 50 nM dsP111, or 100 nM dsP52 or dsP93]. The reaction mixture was subsequently treated with proteinase K [Tuschl et al. 1999], and the dsRNA-processing products were separated on a denaturing 15% polyacrylamide gel. A band, including a size range of at least 18–24 nt, was excised and then eluted into 0.3 M NaCl overnight at 4°C in siliconized tubes. The RNA was recovered by ethanol precipitation and then dephosphorylated (30 µL reaction, 30°C, 30 min, 10 U alkaline phosphatase, Roche). The reaction was stopped by phenol/chloroform extraction, and the RNA was ethanol precipitated. The 3′-labeled oligonucleotide [pUUUaaccgc cactctctc: uppercase, RNA2; lowercase, DNA; p, phosphate; x, 4-hydroxymethylbenzyl] was then ligated to the dephosphorylated ~21-nt RNA [20 µL reaction, 37°C, 30 min, 5 µL 3′-adapter, 50 mM Tris-HCl at pH 7.6, 10 mM MgCl2, 0.2 mM ATP, 0.1 mg/mL acetylated BSA, 15% DMSO, 25 µM U T4 RNA ligase; Amersham-Pharmacia] (Pan and Uhlenbeck 1992). The ligation reaction was stopped by the addition of an equal volume of 8 M urea/50 mM EDTA stopmix and directly loaded on a 15% gel. The ligation products were separated by denaturing gel electrophoresis in 1× TAE-buffer. The 52-bp dsRNAs with the 17-nt and 20-nt 3′ overhangs were annealed by incubating at 95°C for 1 min and then rapidly cooled to 70°C and followed by slow cooling to room temperature over a 3-h period [50 µL annealing reaction, 1 µM strand concentration, 300 mM NaCl, 10 mM Tris-HCl at pH 7.5]. The dsRNAs were then phenol/chloroform extracted, ethanol precipitated, and dissolved in lysis buffer.

The phosphorylation reaction was stopped by phenol/chloroform extraction, and the RNA was recovered by ethanol precipita-
tion. Next, the 5' adapter (tactatacgaacctAAA: uppercase, RNA; lowercase, DNA) was ligated to the phosphorylated ligat

tion product as described above. The new ligation product was

gel purified and eluted from the gel slice in the presence of

reverse transcription primer (GACTAGCTGGAAATTCAGGATGCCGCTTAAA: bold, EcoRI site), used as carrier. Reverse

transcription (15 µL reaction, 42°C, 30 min, 150 U Superscript II

reverse transcriptase; Life Technologies) was followed by PCR

using a 5' primer CAGCCAACCGAAATTCATAGCAGCTC

TAAA (bold, EcoRI site) and the 3' RT primer. The PCR product

was purified by phenol/chloroform extraction and ethanol

precipitated. The PCR product was then digested with

EcoRI (NEB) and concatamerized using T4 DNA ligase (high concentration, NEB). Concatamers of a size range of 200–800 bp were separated on a low-melt agarose gel, recovered from the gel by a standard melting and

melon extraction procedure, and ethanol precipitated. The unpaired ends were filled in by incubation with Taq

polymerase under standard conditions at 72°C for 15 min, and

the DNA product was directly ligated into the pCR2.1-TOPO

vector using the TOPO TA cloning kit (Invitrogen). Colonies

were screened using PCR with M13-20 and M13 Reverse se-

quencing primers. PCR products were directly submitted for

custom sequencing (Sequence Laboratories Go

quencing primers. PCR products were directly submitted for

custom sequencing [Sequence Laboratories Götingen]. On av-

erage, four to five ~21-mer sequences were obtained per clone.

2D-TLC analysis

Nuclease P1 digestion of radiolabeled, gel-purified siRNAs and

2D-TLC was carried out as described [Zamore et al. 2000].

Nuclease T2 digestion was performed in 10 µL reactions at 50°C

for 3 h in 10 mM ammonium acetate (pH 4.5) using 2 ng/µL
carrier tRNA and 30 U ribonuclease T2 (Life Technologies). The

migration of nonradioactive standards was determined by UV

detection. The identity of nucleoside-3'-monophosphates using

[γ-32P]ATP and T4 polynucleotide kinase [data not shown].

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mer sequences were obtained per clone.

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