A novel FLOURY ENDOSPERM2 (FLO2)-interacting protein, is involved in maintaining fertility and seed quality in rice

Rintaro Suzuki¹, Tomohiro Imamura¹,a, Yoko Nonaga¹, Hiroaki Kusano¹,b, Hiroshi Teramura¹, Ken-Taro Sekine², Tetsuro Yamashita³, Hiroaki Shimada¹,*

¹Department of Biological Science and Technology, Tokyo University of Science, 6-3-1 Niijuku, Katsushika, Tokyo 125-8585, Japan; ²Faculty of Agriculture, University of the Ryukyus, 1 Senbaru, Nishihara, Okinawa 903-0213, Japan; ³Faculty of Agriculture, Iwate University, 3-18-8 Ueda, Morioka, Iwate 020-8550, Japan

*E-mail: shimadah@rs.noda.tus.ac.jp Tel: +81-3-5876-1469 Fax: +81-3-5876-1614

Received November 15, 2019; accepted December 12, 2019 (Edited by M. Yamaguchi)

Abstract  Crop plants accumulate a large amount of storage starch and storage proteins in the endosperm. Genes involved in the biosynthesis of these substances work in concert during development of the rice endosperm. The rice flo2 mutant produces aberrant seeds with reduced grain quality. FLOURY ENDOSPERM 2 (FLO2), the causative gene of the flo2 mutant, is considered to be a regulatory protein that controls the biosynthesis of seed storage substances. FLO2 contains tetratricopeptide repeat (TPR) motifs that may mediate protein–protein interactions. In this study, we identified the protein that interacts with the TPR motif of FLO2. We generated a transformant that produced the FLAG-tagged fusion FLO2 protein in the flo2 mutant and used this in the shotgun proteomic analysis. A protein, which we named FLOC1, interacted with FLO2. In vitro pull-down assays indicated that the TPR motif was involved in this interaction. A knock-down transformant of FLOC1 showed significantly reduced fertility and generation of seeds with abnormal features. These findings suggest that FLOC1 is involved not only in seed fertility but also in seed quality. These phenotypes were also observed on the RNAi transformants of the flo2 mutant although the effect of the flo2 mutation remained. These findings imply that there is a difference in the functions of FLO2 and FLOC1 although both of appear to be involved in the control of seed quality during seed formation.

Key words: developing seed, fertility, protein–protein interaction, rice, seed formation.

Introduction

Rice is an important crop that accounts for more than 20% of the world’s grain production. The rice endosperm forms within a few weeks after flowering, and accumulates a large amount of storage substances, such as starch and storage proteins (Zhou et al. 2013). The quality of the rice endosperm affects the appearance, taste, and yield of crop production, both of which are one of the goals of rice breeding. Elucidating the mechanism that determines endosperm quality provides important knowledge for creating crops with high-quality, high-yield grains (Zhang 2007).

In rice, several key enzymes are involved in starch biosynthesis, such as AGPase, granule-bound starch synthase, soluble starch synthase, and starch branching enzyme (Caballero et al. 2008; Hirose and Terao 2004; Mizuno et al. 1993; Müller-Röber et al. 1992). Storage proteins, such as glutelin, globulin, and prolamin, are also produced during seed development and accumulate in the endosperm (Krishnan and White 1995; Tanaka et al. 1980). These proteins work in concert during a short period during development stage. The rice floury mutants, flo1 to flo5, exhibit the aberrant features in the endosperm. The causative genes for flo3, flo4, and flo5 have been identified as the 16-kDa globulin gene, OsPPDKB and OsSSSIIIa, respectively (Kang et al. 2005; Nishio and Iida 1993; Ryoo et al. 2007). Other floury mutants have been reported recently. The causative gene for flo6 encodes a CBM-containing protein, which may influence the starch content and its physicochemical features (Peng et al. 2014).

The rice flo2 mutant produces the aberrant endosperm that shows cloudy, dwarf, and pulverized features, and results in a significantly reduced grain quality (Kawasaki et al. 1996). In this mutant, the expression levels of many genes involved in starch biosynthesis and storage protein biosynthesis are significantly decreased. FLOURY
ENDOSPERM 2 (FLO2), product of the causative gene of the flo2 mutant, the FLO2 gene, is considered to be a regulatory protein that controls the biosynthesis of seed substance storage (She et al. 2010). However, it is unclear how FLO2 regulates the genes involved in the biosynthesis of the storage substances in developing seeds.

FLO2 is a large protein comprising 1720 amino acid residues. The middle region of this protein contains three repeats of the tetratricopeptide repeat (TPR) motif. In other part, no homology to any other protein with known function has been found (She et al. 2010). The TPR motif is a degenerate 34-amino acid tandem repeat that forms scaffolds that mediate protein–protein interactions and can assemble multiple protein complexes (Chadli et al. 2008). We have found a LEA protein and bHLH, a potential transcription factor, as candidate proteins that interact with FLO2 (She et al. 2010). We have predicted that FLO2 builds a complex of multiple proteins. In this study, we sought to identify another regulatory factor that interacts with FLO2 and which may be involved in the regulatory mechanism controlling the quality of rice seed development.

Materials and methods

Plant materials and growth conditions

Japonica rice (Oryza sativa L. cv. Nipponbare) was used as the wild-type plant. The flo2 mutant, EM37, which was generated by MNU treatment, was used (Satoh and Omura 1981). Seeds were germinated at 30°C in a dark chamber and were grown in a greenhouse. After flowering, the rice plants were cultivated in the growth chamber under 12-h-light (28°C) and 12-h-dark (25°C) conditions. The gene for the fusion FLO2 protein was created as follows. We chemically synthesized a 1.5-kb DNA fragment that encodes the C-terminal portion of the FLO2 protein followed by the 3xFLAG tag sequence, and replaced it with the BanIII–XhoI region of the DNA fragment containing the full-length FLO2 gene, which was inserted in the pGWB1 vector (She et al. 2010). The resultant gene was introduced into the flo2 mutant. The transformants were generated and grown in the greenhouse.

Generation of transformants containing the RNAi construct for the Os03g0663800 gene

For the RNAi construct, two parts of the Os03g0663800 gene were PCR amplified from its cDNA using KOD-plus-NEO (Toyobo, Osaka, Japan) and cloned using pENTER/D-TOPO kits (Invitrogen, Waltham, MA, USA). These fragments corresponded to the positions 1 to 305 and 1564 to 1689 of the cDNA, and were connected and introduced into the pANDA vector, which was driven by the CaMV 35S promoter (Miki and Shimamoto 2004). The resultant plasmid was used for the transformation of the wild-type and flo2 mutant using the Agrobacterium-mediated method (Hiei et al. 1994). The transformants were grown on Murashige–Skoog plates (Murashige and Skoog 1962) supplemented with 50 µg·l⁻¹ hygromycin B (Fuji-Film Wako Pure Chemical Industries, Ltd. Corp., Tokyo, Japan) to screen for hygromycin-resistant lines before their cultivation in soil.

The plants were grown in a growth cabinet or greenhouse. The gene introduced into the transformants was confirmed by PCR using the primer set corresponding to the GUS linker region in the pANDA vector, 5′-CAT GAA GAT GCG GAC TTA CG and 5′-ATC CAG TTC TTC TTC GCC CC and 5′-ATC CAC GCC GTA TTC CG.GG for the GUS linker. PCR was performed with an initial denaturation at 94°C for 2 min followed by 35 cycles at 94°C for 30 s, 57°C for 30 s, and 72°C for 30 s using a GeneAmp PCR System 2720 (Applied Biosystems, Foster City, CA, USA).

RNA isolation and reverse transcription (RT)-PCR

Total RNA was isolated from rice using the RNeasy Mini Kit (Qiagen, Venlo, Netherlands). The first-strand cDNA was synthesized from 1 µg of total RNA using a ReverTra Ace cDNA synthesis kit (Toyobo) with an oligo-dT(20) primer. RT-PCR was performed using the GeneAmp PCR System 2720 mentioned above. To detect the transcript for the RNAi construct, the region of the GUS linker lying in the regions of the RNAi transcript was used. The procedure for amplification comprised initial denaturation at 94°C for 2 min followed by 35 cycles at 94°C for 30 s, 57°C for 30 s, and 72°C for 30 s. The Actin I transcript (acc no. AK100267) was used as the control and was amplified by an initial denaturation at 94°C for 2 min followed by 30 cycles at 94°C for 30 s, 57°C for 30 s, and 72°C for 30 s using the primer set of 5′-CCC TCC GTA AAG GAA GTA CAG TGT and 5′-GTC GA AGA ATT AGA AGC ATT TCC.

RNA blot analysis

For the RNA-blot analysis, 20 µg of total RNA was applied in each lane of a 1% agarose gel containing 1xMops running buffer (20 mM Mops, pH 7.0, 8 mM acetate, and 1 mM EDTA) supplemented with 38% formaldehyde and electrophoresed, and the bands were then transferred onto Hybond-N’ nylon membranes (GE Healthcare, Buckinghamshire, UK). The membranes were subjected to detection of the transcript. Hybridization was performed according to Ausubel et al. (1987) using the DIG-labeled probe corresponding to the 3′ region of the Os03g0663800 gene, which was PCR-amplified using the primers, 5′-ATC CAG TTC TTC TTC GCC CC and 5′-CCC CCT GTG CTG CTG CTT CCC, in which the region between positions of 685 and 819 in the cDNA for the Os03g0663800 gene (acc no. AK105347) was amplified. DIG-labeled probe was synthesized using PCR DIG Labeling Mix (Roche, Basel, Switzerland). Fragments hybridized to the probe were detected by the interaction with anti-digoxigenin-AP Fab fragments (Roche) using an ImageQuant LAS 4000 imager (GE Healthcare).

Fractionation, digestion, and identification of proteins interacting with FLO2

Developing rice seeds (15 days after flowering) were ground...
in TBS buffer (200 mM Tris-HCl, pH 7.5, 10% glycerol, 5 M NaCl, and 0.01% Protease Inhibitor Cocktail (Sigma-Aldrich, St Louis, MO, USA)). The cell lysate was centrifuged at 1000 g to remove the cell debris and insoluble materials (P1 fraction). The resulting supernatant (S1) was fractionated by centrifugation at 11,400 g into the fractions comprising precipitates (P2) and supernatant (S2) of the cell lysates. The P2 fraction was suspended in the same amount of TBS buffer. The P1, P2, and S2 fractions were used for further analysis. A 10-ml aliquot of the S2 fraction was incubated with the agarose beads with 5 µl of monoclonal anti-FLAG antibody (Sigma-Aldrich) at 4°C for 2 h. After incubation, the reaction mixture was applied to a Micro Bio-Spin chromatography column (Bio-Rad, Hercules, CA, USA), and the agarose beads were collected by centrifugation. After washing with TBS buffer, the proteins bound to the agarose beads were extracted with 150 µl of extraction buffer (60 mM Tris-HCl, pH 6.8, 2% SDS, 10% glycerol, and 1.5% DTT) with heating.

The fractions obtained were analyzed by SDS-polyacrylamide gel electrophoresis (PAGE) and protein blot analysis using the anti-FLAG antibody. SDS-PAGE was performed according to Ausubel et al. (1987). In parallel, the protein samples were loaded onto an SDS-polyacrylamide gel and electrophoresed for a short time to remove the impurities in the fraction. The gel containing the protein fraction was excised and subjected to protein digestion by trypsin using an In-Gel Tryptic Digestion Kit (Thermo Scientific, Vantaa, Finland). Digested peptides were applied onto a Magic C18 AQ nano-column (Michrom Bioresources, Auburn, CA, USA) in an Advance UHPLC system (Michrom Bioresources) equilibrated with 0.1% formic acid in acetonitrile, and the peptides were eluted using a linear gradient from 5 to 45% acetonitrile at a flow rate of 500 nl min⁻¹. The digested peptides were analyzed using an LTQ Orbitrap XL (Thermo Scientific) mass spectrometer operated with Xcalibur software (version 2.0.7, Thermo Scientific). Peptides were identified using an in-house Mascot server (MS/MS ion search, Mascot version 2.5, Matrix Science Inc.).

**Phylogenetic analysis**

Alignment of the proteins was performed using Clustal X program from the Clustal website (http://www.clustal.org/). The phylogenetic tree was constructed using the Phylip neighbor-joining method with bootstrap values from 1,000 neighbor-joining bootstrap replicates (Felsenstein 2005). The tree was visualized using the TreeView program (Page 1996).

**In vitro pull-down assay for detecting the protein–protein interactions**

3xFLAG-tagged proteins, corresponding to the entire and the portions of FLO2 proteins, and the 6xHis-tagged target protein were used for the in vitro pull-down experiment. For the 3xFLAG-tagged FLO2 proteins, the fragments encoding the entire and part of FLO2, corresponding to the positions 1–1720, 568–1720, and 933–1050 in the amino acid sequence, were prepared by PCR amplification from FLO2 cDNA, and connected with the fragment for 3xFLAG tag at their 3’ ends. The 6xHis-tagged target proteins were constructed as follows. A fragment encoding the protein encoded by the Os03g0663800 gene was chemically synthesized. The nucleotide sequence for this protein was modified without any alteration of its amino acid sequence for efficient in vitro translation (Supplementary Figure S1). Using this artificial gene, the fragments for the truncated proteins, corresponding to the positions 1–314 and 265–562 of the amino acid sequence, were PCR amplified. They were connected with the fragment for the 6xHis tag and then inserted into the vector, pEU-dMac3, for cell-free protein synthesis. pEU-dMac3 was constructed by replacing the E01 sequence with the dMac3 translational enhancer in pEU-E01. dMac3 is a portion of the 5’ untranslabeled region of OsMac3 mRNA, comprising 161 nucleotides, which exhibits sufficient activity as a translational enhancer (Aoki et al. 2014).

Using the resultant plasmids, mRNAs were synthesized using an in vitro transcription system. They were used in the preparation of the corresponding proteins using a wheat germ protein synthesis kit (CellFree Science, Yokohama, Japan). The general procedure followed the manufacturer’s protocol. The generated proteins were evaluated by SDS-PAGE and protein blot analysis. The 3xFLAG-tagged FLO2 protein was applied to ANTI-FLAG M2 Affinity Gel (Sigma-Aldrich), and the 6xHis-tagged target proteins were added to the resin. The resin was washed with a buffer (25 mM Tris-HCl, pH 7.5, 150 mM NaCl, 10 mM EDTA, 10 mM MgCl₂, 1% Triton X-100, and 2% glycerol), and the proteins interacting with the resin were eluted with boiling buffer (60 mM Tris-HCl, pH 6.8, 2% SDS, 10% glycerol, and 1.5% DTT). The eluted fraction was subjected to SDS-PAGE, and the His-tagged proteins were detected using protein blot analysis with the anti-6xHis tag antibody (Clontech-takara, Kusatsu, Japan). As a control, reactions were performed in the absence of FLO2 protein.

**Results**

**Generation of a rice transformant containing the fusion gene encoding FLO2 fused with the FLAG tag**

FLO2 is strongly expressed in developing seeds, coincident with the expression of other genes that encode storage starch and storage proteins (She et al. 2010). To examine the behavior of FLO2 protein, we created a transformant containing the fusion gene encoding the FLO2 fused to the FLAG tag, which was driven by the native FLO2 promoter (Figure 1A).

This fusion gene was introduced into the rice flo2 mutant, and three independent transformants were obtained. These transformants, designated as FLO2-MF, produced normal-shaped seeds whose features were similar with those of the wild-type plants. It has been reported that the flo2 mutant produces seeds with floury endosperm and that their size and weight are
FLOC1 is involved in seed formation

Reduced (Kawasaki et al. 1996). Our result indicated that introduction of the fusion FLO2 gene resulted in restoration of the phenotype of the flo2 mutant (Figure 1B). This observation suggests that the fusion FLO2 gene was expressed sufficiently to induce the expression of functional FLO2 in the developing seeds of FLO2-MF.

Next, we attempted to detect the fusion FLO2 protein in the developing seeds. A crude extract was prepared from the developing seeds of FLO2-MF, and was subjected to protein blot analysis using an anti-FLAG antibody. A band corresponding to the 250 kDa protein was detected in the developing seeds of FLO2-MF (Figure 1C). This size was considered to represent the full-length protein transcribed from the fusion FLO2 gene.

**Search for the factor that interacts with FLO2**

Using the developing seeds of FLO2-MF, we sought to identify the protein that interacts with FLO2. A cell lysate was prepared from the developing seeds harvested at 15 days after flowering (Figure 2A). The resultant solution was combined with an anti-FLAG antibody (FLAG-IP), and the immunoprecipitated fraction, which was expected to contain the fusion FLO2 protein, was collected (Figure 2B). In parallel, to obtain a control fraction, a similar procedure was performed using the developing seeds of the wild-type plants. All of these fractions were then subjected to shotgun proteomic analysis.

The shotgun proteomic analysis produced many peptide fragments, which were candidate proteins that interacted with FLO2 (Supplementary Tables S1 to S3). We screened for the unique protein in both FLO2-MF and control by first removing the proteins bound nonspecifically to the antibody. We obtained two proteins that we considered to be unique in the FLO2-MF transformant. One was a novel protein whose function is unclear, and the other was FLO2.

This protein composed 562 amino acid residues and was encoded by the Os03g46100 gene. The corresponding cDNA to this protein was registered as AK105347. This gene had a nucleotide sequence with extremely high GC content (Supplementary Figure S1). The deduced amino acid sequence contained multiple repeats of some amino acid residues, such as glutamine and glutamic acid (Figure 3A). This protein was predicted to be a member of the cupin super family that had two cupin domains with a molecular weight of 63 kDa (acc. no. Q75GX9) (Figure 3B). However, this protein has not been reported and its function is unclear. We named this protein...
protein FLOC1 (FLO2-interacting cupin domain protein 1). The RiceXPro database (http://ricexpro.dna.affrc.go.jp/) showed that this gene was expressed specifically in immature seeds (Supplementary Figure S2A).

FLOC1 had two homologue genes in the rice genome, designated as FLOC2 and FLOC3, whose cDNAs were registered as AK102651 and AK105307. The predicted FLOC2 and FLOC3 proteins had 28% and 35% similarity to FLOC1, respectively. FLOC2 and FLOC3 were found to contain a single cupin domain, whereas FLOC1 had two cupin domains (Figure 3B). These genes were also specifically expressed in the developing seeds (Supplementary Figures S2B and S2C). Both FLOC2 and FLOC3 were not included in the list of the proteins interacted with FLO2.

The orthologues of FLOC1 were widely distributed in various plants, such as wheat, sorghum (Sorghum bicolor), grape (Vitis vinifera), and Arabidopsis thaliana (Figure 3C). However, no orthologues with FLOC1 were found in yeast and animals. These results suggest that the FLOC1, FLOC2, and FLOC3 proteins are unique and are conserved in higher plants.
Detection of the interaction between FLO2 and FLOC1

To examine the interactions between FLO2 and FLOC1, we performed an in vitro pull-down assay to identify the protein–protein interactions. As the target, the genes encoding a truncated FLOC1 protein containing cupin domain 1 (N-region FLOC1), a truncated FLOC1 protein containing cupin domain 2 (C-region FLOC1), and the entire FLOC1, were constructed (Figure 4A). Similarly, the genes for a series of FLAG-tagged FLO2 proteins, the entire FLO2, a truncated FLO2 protein containing the C-terminal region, and a small protein containing the region for TPR motif, were constructed (Figure 4B). These proteins were produced by the wheat germ cell-free protein production system using the assembled vectors to the translational enhancer dMac3. Sufficient amounts of the proteins were obtained (Figure 4C), and the proteins were subjected to the following pull-down assay.

The pull-down assay detected the interaction between FLO2 and FLOC1. All of the FLOC1 proteins, the entire FLOC1, the N-region FLOC1, and the C-region FLOC1 interacted with the proteins corresponding to the entire FLO2, truncated FLO2, and the region containing the TPR motif (Figure 4C). No protein was detected on any reaction in absence of FLO2 protein. These results suggested that both the cupin domains are involved in interactions with the TPR motif of FLO2.

Analysis of the phenotype of the transformant containing the RNAi for FLOC1

To examine the physiological roles of FLOC1, we examined the phenotype caused by the lack of FLOC1. Performing this analysis in the presence or absence of FLO2 would indicate whether FLOC1 and FLO2 have independent functions, complement those functions, or a synergistic function. An RNAi construct for FLOC1 was constructed using the sequences in the 5′ and 3′ regions of FLOC1 (Figure 5A). These were introduced into wild-type and the flo2 mutant to generate transformants (Figure 5B), in which the expression of the FLOC1 gene was suppressed (Figure 5C). These transformants...
R. Suzuki et al.

53

Copyright © 2020 The Japanese Society for Plant Cell and Molecular Biology

grew normally during the vegetative growth phase and headed similarly as did the wild-type plants (Figure 5D). However, the RNAi transformants showed a significantly reduced fertility rate (Figure 5F). In addition, seeds produced by these transformants showed morphological abnormalities, such as small size and severely wrinkled features (Figure 5H). These results suggest that knockdown of FLOC1 affected both the seed quality and fertility. Phenotypes of low fertility and abnormal seed features were also observed in the RNAi transformants derived from the flo2 plants, although these formed floury seeds (Figure 5E, G, I). These findings suggest that, as well as FLO2, FLOC1 plays important roles in seed development, but they have distinct functions.

Discussion

The rice endosperm accumulates a large amount of storage substances. Enzymes involved in the production of these substances are the key factors that determine the yield and grain quality. Rice FLO2 plays a crucial role in seed development, for example, by regulating the biosynthesis of storage starch and proteins in the endosperm (She et al. 2010). We produced transformants harboring the fusion FLO2 protein gene in the flo2 mutant. Considering the observation that these
transformants produced normal-shaped seeds, we suggest that the fusion FLO2 protein worked efficiently as FLO2 in developing seeds and compensated for the flo2 mutation (Figure 1). In this study, we performed shotgun proteomic analysis to identify a novel protein that interacts with the fusion FLO2, which we expected to be involved in endosperm formation during endosperm development.

FLO2 is predicted to be a protein that interacts with other proteins to exhibit its function. We have found that a LEA protein and a bHLH transcription factor can interact with FLO2 (She et al. 2010). However, we had no data to indicate that they interact directly with the TPR motif of FLO2, and in this study, we attempted to identify other proteins that interact with the TPR motif of FLO2. Given that the fusion FLO2 protein was detected in the developing seeds by the anti-FLAG antibody, in this study, we tried to identify the proteins that can interact with FLO2 and found the novel protein, FLOC1, as a candidate protein that can interact with FLO2.

FLOC1 had two cupin domains, whose functions were unknown. FLOC1 was wildly conserved in the plant genomes. Rice FLOC1 had two homologues, FLOC2 and FLOC3 (Figure 3). FLOC1, FLOC2, and FLOC3 were expressed specifically in the developing seeds. This expression pattern coincided with that of FLO2. FLOC2 and FLOC3 had a single cupin domain, whereas FLOC1 had two cupin domains. In addition, FLOC2 and FLOC3 were not included in the list of the candidates that can interact with FLO2, which suggests that FLOC2 and FLOC3 may be functionally divergent from FLOC1.

The pull-down assay identified the interaction between FLOC1 and FLO2 and that the TPR motif may be involved in this interaction (Figure 4). The TPR motif comprises tandem repeats of 34 amino acid residues, and adopts a right-handed helical loop–helix structure with an amphipathic channel; such channels are involved in many protein–protein interactions (Allan and Ratajczak 2011). Proteins containing cupin domain are widely distributed. It has been reported that germin and germin-like proteins (GLPs) play an important role in plant development and defense (Lane et al. 1993). However, the function of the cupin domain remains unclear.

FLO2 is also expressed in leaves, although the function of FLO2 in leaves is unknown (She et al. 2010). It has been reported that A. thaliana FLO2 is involved in the regulation of translocation and transport of assimilates and in quality control of substance supply or transfer (Kihira et al. 2017). The TPR motif has been shown to exhibit great flexibility and variability in terms of its substrate specificity (Nojima et al. 2017). FLOC1 was absent in leaves, and, therefore, it was expected that FLO2 would interact with other factors.

The knock-down transformants harboring the RNAi construct for FLOC1 exhibited significantly reduced fertility, and produced aberrant seeds with small and severely wrinkled features (Figure 5). This suggests that FLOC1 is involved in both seed fertility and seed quality. It has been reported that the flo2 mutation results in the formation of grains with white and floury endosperm that contains loosely packed, small, and spherical starch granules with large air spaces (She et al. 2010). In the RNAi transformants derived from the wild-type plant, the seeds became small and formed abnormal features. This suggests that a lack of FLOC1 caused insufficient grain filling and resulted in aberrant seed formation. In the RNAi transformants derived from the flo2 mutant, these phenotypes were also observed along with the formation of floury seeds (Figure 5). This observation suggests that the effect of the flo2 mutation remained in these transformants, which implies difference in the functions of FLO2 and FLOC1 even though both are involved in the control of seed quality during seed formation.

Acknowledgements
We thank Miho Kihira, Hiromi Mutsuro-Aoki, Mariko Ohnuma, and Daiki Miyano for their assistance in the experimental work.

References
Allan RK, Ratajczak T (2011) Versatile TPR domains accommodate different modes of target protein recognition and function. Cell Stress Chaperones 16: 353–367
Aoki H, Teramura H, Schepetinikov M, Ryabova LA, Kusano H, Shimada H (2014) Enhanced translation of the downstream ORF attributed to a long 5' untranslated region in the OsMac1 gene family members, OsMac2 and OsMac3. Plant Biotechnol 31: 221–228
Ausubel FM, Brent R, Kingston RE, Moore DD, Seidman JG, Smith JA, Struhl K (1987) Current Protocols in Molecular Biology. Wiley, New York, sections 4.9 and 10.6
Caballero L, Bancel E, Debton C, Branlard G (2008) Granule-bound starch synthase (GBSS) diversity of ancient wheat and related species. Plant Breed 127: 548–553
Chadli A, Bruinsma ES, Stengard B, Toft D (2008) Analysis of Hsp90 cochaperone interactions reveals a novel mechanism for TPR protein recognition. Biochem 47: 2850–2857
D'Andrea LD, Regan L (2003) TPR proteins: The versatile helix. Trends Biochem Sci 28: 655–662
Felsenstein J (2005) PHYLIP (Phylogeny Inference Package) Version 3.69. http://evolution.genetics.washington.edu/phylip.html
Hiei Y, Ohta S, Komari T, Kamishiro T (1994) Efficient transformation of rice (Oryza sativa L.) mediated by Agrobacterium and sequence analysis of the boundaries of the T-DNA. Plant J 6: 271–282
Hirose T, Terao T (2004) A comprehensive expression analysis of the starch synthase gene family in rice (Oryza sativa L.). Plant Cell Physiol 47: 2850–2857
Kang HG, Park S, Matsuoka M, An G (2005) White-core endosperm floury endosperm-4 in rice is generated by knockout
mutations in the C4-type pyruvate orthophosphate dikinase gene (OsPPDKB). *Plant J* 42: 901–911
Kawasaki T, Mizuno K, Shimada H, Satoh H, Kishimoto N, Okumura S, Ichikawa N, Baba T (1996) Coordinated regulation of the genes partitioning in starch biosynthesis by the rice Flurry-2 locus. *Plant Physiol* 110: 89–96
Kihira M, Taniguchi K, Kaneko C, Ishii Y, Aoki H, Koyanagi A, Kusano H, Suzuki N, Yin Y-G, Kawachi N, et al. (2017) *Arabidopsis thaliana* FLO2 is involved in efficiency of photoassimilate translocation, which is associated with leaf growth and aging, yield of seeds and seed quality. *Plant Cell Physiol* 58: 440–450
Kishimoto N, Okumura S, Ichikawa N, Baba T (1996) Coordinated regulation of the genes partitioning in starch biosynthesis by the rice Flurry-2 locus. *Plant Physiol* 110: 89–96
Kawasaki T, Mizuno K, Shimada H, Satoh H, Kishimoto N, Okumura S, Ichikawa N, Kawachi N, et al. (1996) Coordinated regulation of the genes partitioning in starch biosynthesis by the rice Flurry-2 locus. *Plant Physiol* 110: 89–96
Kihira M, Taniguchi K, Kaneko C, Ishii Y, Aoki H, Koyanagi A, Kusano H, Suzuki N, Yin Y-G, Kawachi N, et al. (2017) *Arabidopsis thaliana* FLO2 is involved in efficiency of photoassimilate translocation, which is associated with leaf growth and aging, yield of seeds and seed quality. *Plant Cell Physiol* 58: 440–450

Krishnan HB, White IA (1995) Morphometric analysis of rice seed protein bodies (Implication for a significant contribution of prolamine to the total protein content of rice endosperm). *Plant Physiol* 105: 1991–1995
Lane BG, Dunwell JM, Ray JA, Schmitt MR, Cuming AC (1993) Germin, a protein marker of early plant development, is an oxalate oxidase. *J Biol Chem* 268: 12239–12242
Miki D, Shimamoto K (2004) Simple RNAi vectors for stable and transient suppression of gene function in rice. *Plant Cell Physiol* 45: 490–495
Mizuno K, Kawasaki T, Shimada H, Satoh H, Kobayashi E, Okumura S, Arai Y, Baba T (1993) Alteration of the structural properties of starch components by the lack of an isoform of starch branching enzyme in rice seeds. *J Biol Chem* 268: 19084–19091
Müller-Röber B, Sonnewald U, Willmitzer L (1992) Inhibition of the ADP-glucose pyrophosphorylase in transgenic potatoes leads to sugar-storing tubers and influences tuber formation and expression of tuber storage protein genes. *EMBO J* 11: 1229–1238
Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiol Plant* 15: 473–497
Nishio T, Iida S (1993) Mutants having a low content of 16-kDa allergenic protein in rice (*Oryza sativa* L.). *Theor Appl Genet* 86: 317–321
Nojima S, Fujishima A, Kato K, Ohuchi K, Shimizu N, Yonezawa K, Tajima K, Yao M (2017) Crystal structure of the flexible tandem repeat domain of bacterial cellulose synthesis subunit C. *Sci Rep* 7: 13018
Page RDM (1996) TreeView: An application to display phylogenetic trees on personal computers. *Comput Appl Biosci* 12: 357–358
Peng C, Wang Y, Liu F, Ren Y, Zhou K, Lv J, Zheng M, Zhao S, Zhang L, Wang C, et al. (2014) *FLOURY ENDOSPERM6* encodes a CBM48 domain-containing protein involved in compound granule formation and starch synthesis in rice endosperm. *Plant J* 77: 917–930
Ryoo N, Yu C, Park CS, Baik MY, Park IM, Cho MH, Bhoo SH, An G, Hahn TR, Jeon JS (2007) Knockout of a starch synthase gene OsSSIIIa/Flo5 causes white-core floury endosperm in rice (*Oryza sativa* L.). *Plant Cell Rep* 26: 1083–1095
Satoh H, Omura T (1981) New endosperm mutations induced by chemical mutagens in rice, *Oryza sativa* L. *Jpn J Breed* 31: 316–326
She K-C, Kusano H, Koizumi K, Yamakawa H, Hakata M, Imamura T, Fukuda M, Naito N, Tsurumaki Y, Yaeshima M, et al. (2010) A novel factor *FLOURY ENDOSPERM2* is involved in regulation of rice grain size and starch quality. *Plant Cell* 22: 3280–3294
Tanaka K, Sugimoto T, Ogawa M, Kasai Z (1980) Isolation and characterization of two types of protein bodies in the rice endosperm. *Agric Biol Chem* 44: 1633–1639
Zhang Q (2007) Strategies for developing green super rice. *Proc Natl Acad Sci USA* 104: 16402–16409
Zhou S-R, Yin L-L, Xue H-W (2013) Functional genomics based understanding of rice endosperm development. *Curr Opin Plant Biol* 16: 236–246