Role of sodium-dependent Pi transporter/Npt2c on Pi homeostasis in klotho knockout mice different properties between juvenile and adult stages

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Abstract
SLC34A3/NPT2c/NaPi-2c/Npt2c is a growth-related NaPi cotransporter that mediates the uptake of renal sodium-dependent phosphate (Pi). Mutation of human NPT2c causes hereditary hypophosphatemic rickets with hypercalciuria. Mice with Npt2c knockout, however, exhibit normal Pi metabolism. To investigate the role of Npt2c in Pi homeostasis, we generated α-klotho−/−/Npt2c−/− (KL2cDKO) mice and analyzed Pi homeostasis. α-Klotho−/− (KLKO) mice exhibit hyperphosphatemia and markedly increased kidney Npt2c protein levels. Genetic disruption of Npt2c extended the lifespan of KLKO mice similar to that of α-Klotho−/−/Npt2a−/− mice. Adult KL2cDKO mice had hyperphosphatemia, but analysis of Pi metabolism revealed significantly decreased intestinal and renal Pi (re)absorption compared with KLKO mice. The 1,25-dihydroxy vitamin D3 concentration was not reduced in KL2cDKO mice compared with that in KLKO mice. The KL2cDKO mice had less severe soft tissue and vascular calcification compared with KLKO mice. Juvenile KL2cDKO mice had significantly reduced plasma Pi levels, but Pi metabolism was not changed. In Npt2cKO mice, plasma Pi levels began to decrease around the age of 15 days and significant hypophosphatemia developed within 21 days. The findings of the present study suggest that Npt2c contributes to regulating plasma Pi levels in the juvenile stage and affects Pi retention in the soft and vascular tissues in KLKO mice.

KEYWORDS
FGF23/klotho, growth, intestine, kidney, phosphate transporter

1 | INTRODUCTION
The serum inorganic phosphate (Pi) concentration is controlled by the intestines and kidneys, which regulate Pi absorption/excretion; the bones, liver, and muscles, which store Pi; and regulatory factors (Kaneko, Tatsumi, Segawa, & Miyamoto, 2017). Sodium-dependent phosphate transporters (the solute carrier 34 [SLC34] family) have an important role in regulating serum Pi levels. SLC34A1/NPT2A/NaPi-2a and SLC34A3/NPT2C/NaPi-2c are...
predominantly expressed in the proximal tubules of the kidney (Kaneko et al., 2011; Lederer & Miyamoto, 2012; Levi et al., 2019). The phenotypic differences resulting from NPT2A and NPT2C gene mutations may be due to differential dominance of the transporters involved in Pi homeostasis. In humans, NPT2C mutations cause hereditary hypophosphatemic rickets with hypercalciuria (HHRH), a primary renal Pi wasting disorder that results in increased serum 1,25-dihydroxy vitamin D3 \([1,25(\text{OH})_2\text{D}_3]\) concentrations with associated intestinal \(\text{Ca}^{2+}\) hyperabsorption, hypercalciuria, and rickets/osteomalacia (Bergwitz & Miyamoto, 2019; Bergwitz et al., 2006; Lorenz-Depiereux et al., 2006; Yamamoto et al., 2007). In mice, disruption of Npt2c (Npt2 designate a mouse NaPi-2) causes hypercalciuria and rickets/osteomalacia (Miyamoto, 2019; Bergwitz et al., 2008). The Npt2c transcript, however, was detected in several tissues, suggesting that extrarenal Npt2c has an important role (Segawa, Onitsuka, Kuwahata, et al., 2009). Furthermore, only Npt2a/Npt2c double knockout (KO) mice exhibit a physiology similar to that of patients with HHRH (Segawa, Onitsuka, Furutani, et al., 2009). In rodents, Npt2a is a major functional NaPi cotransporter in the proximal tubules (Kaneko et al., 2017; Lederer & Miyamoto, 2012; Levi et al., 2019). Npt2c is important for Pi reabsorption in weanling animals but mediates a very small percentage of Pi reabsorption in adult animals (Segawa et al., 2002). A recent study demonstrated that mice with kidney-specific Npt2c deletion show no prominent phenotype (Myakala et al., 2014). The Npt2c transcript, however, was detected in several tissues, suggesting that extrarenal Npt2c has an important role (Kuwahara et al., 2012; Nishimura & Naito, 2008).

Fibroblast growth factor (FGF)23 is a hormone that promotes renal Pi excretion by decreasing Pi reabsorption in the proximal tubules while concurrently reducing the plasma \(1,25(\text{OH})_2\text{D}_3\) concentration by both decreasing its biosynthesis and increasing its metabolism (Hu, Shi, & Moe, 2019). FGF23 requires an additional cofactor, \(\alpha\)-klotho, to bind with high affinity and signal efficiently through its cognate FGF receptors. Klotho and FGF receptors form a heterodimeric receptor for FGF23 (Urakawa et al., 2006). \(\alpha\)-Klotho-mutant (kl/kl) mice exhibit blunted FGF23 signaling and increased Pi reabsorption, hyperphosphatemia, and ectopic calcification (Kuro-o et al., 1997; Segawa et al., 2007). Feeding kl/kl mice a low Pi diet can rescue the kl/kl mouse phenotype because renal klotho protein is upregulated in klotho-deficient animals (Morishita et al., 2001; Segawa et al., 2007). These findings suggest that kl/kl mice are not suitable for examining the effects of dietary Pi.

\(\alpha\)-Klotho knockout (KLKO) mice have a phenotype similar to that of kl/kl mice (Alexander et al., 2009; Hu et al., 2019, 2011; Kuro-o et al., 1997; Morishita et al., 2001; Ohnishi, Nakatani, Lanske, & Razzaque, 2009a, 2009b; Segawa et al., 2007; Tsujikawa, Kurotaki, Fujimori, Fukuda, & Nabeshima, 2003; Yoshida, Fujimori, & Nabeshima, 2002). A detailed analysis of Pi metabolism in KLKO mice has not yet been reported. Ohnishi et al reported that serum Pi levels are an important determinant of calcification in \(\alpha\)-klotho\(^{-/-}\)/NaPi2a\(^{-/-}\) mice, and lowering serum Pi levels can reduce or eliminate soft-tissue and vascular calcification, even in the presence of extremely high serum calcium (Ca) and active vitamin D levels (Ohnishi, Nakatani, Lanske, & Razzaque, 2009a).

In the present study, we investigated the role of Npt2c in KLKO mice. Our findings revealed that although Npt2c does not affect the plasma Pi concentration, it has the same effects as Npt2a to suppress calcification and increase the lifespan in KLKO mice.

## 2 MATERIAL AND METHODS

### 2.1 Experimental animals

KLKO mice and Npt2aKO mice were purchased from CLEA Japan Inc. (Tokyo, Japan) and Jackson Laboratories, respectively. In the present study, we used KLKO mice produced by Tsujikawa et al. (2003). The generation of Npt2cKO mice was described previously (Segawa, Onitsuka, Kuwahata, et al., 2009). KLKO heterozygous mice and Npt2a or Npt2c heterozygous mice were crossed to generate KLKO (\(\alpha\)-klotho\(^{-/-}\), Npt2a\(^{-/+}\), Npt2c\(^{-/+}\)), \(\alpha\)-klotho/Npt2a double KO (KL2aDKO; \(\alpha\)-klotho\(^{-/-}\), Npt2a\(^{-/-}\), Npt2c\(^{+/-}\)), and \(\alpha\)-klotho/Npt2c double KO (KL2cDKO; \(\alpha\)-klotho\(^{-/-}\), Npt2a\(^{+/-}\), Npt2c\(^{-/-}\)) mice. Genomic DNA was extracted from tail clippings and amplified by polymerase chain reaction (PCR) using the specific primers listed in Table 1. The mice were weaned at 3 weeks of age and were provided unlimited access to water and standard mouse chow (MF; Oriental). The mice were maintained under pathogen-free conditions and handled in accordance with the Guidelines for Animal Experimentation of Tokushima University School of Medicine (T29-3).

### TABLE 1 Primers for genotyping

| Application          | Primer name | Primer sequence (5’-3’) |
|----------------------|-------------|------------------------|
| Klotho genotyping    | Klotho 1    | GATGGAGGCCACAGGATTGT   |
|                      | Klotho 2    | TGTGCGGTAAGCTTGTGT    |
|                      | Klotho 3    | CGACGTTCAGACTGTG     |
| Npt2a genotyping     | Npt2a 1     | ATGGCTACCACCTAGCTCA   |
|                      | Npt2a 2     | CAAGTGGAATGTGCTTTT    |
|                      | Npt2a 3     | TGCTACTTCATTTTGTACGC |
| Npt2c genotyping     | Npt2c 1     | CTTCCATACATGGCAG      |
|                      | Npt2c 2     | CTGCAATTACCAGACTTCC   |
|                      | Npt2c 3     | CGATATCGCCGCTCCGATC  |
2.2 Metabolic cages to collect urine and fecal samples

The mice were individually placed in metabolic cages at 10:00 a.m. for quantitative urine and fecal collection for 24 hr with free access to food and water. Fecal samples were ashed according to a modified protocol (Ikuta et al., 2019; Ikuta et al., 2018). The fecal samples were collected and placed in beakers and allowed to dry at 110°C for no more than 24 hr. The samples were then ashed at 250°C for 3 hr and at 550°C for 24 hr in a muffle furnace. The samples were cooled, weighed, and digested in HCl with heat, and the sample volume was standardized to 5 ml.

2.3 Biochemical measurements

Plasma, urinary, and fecal Pi, and Ca were determined using commercial kits (Wako), respectively. Blood Ca\textsuperscript{2+} was analyzed using a Siemens Rapid Lab 348 Ca/pH analyzer. Concentrations of plasma FGF23, parathyroid hormone (PTH), and 1,25(OH)\textsubscript{2}D\textsubscript{3} were determined using the intact FGF23 ELISA kit (KAINOS Laboratories), intact PTH ELISA kit (Immunotopics Inc.), and 1,25-(OH)\textsubscript{2} Vitamin D ELISA Kit (Immundiagnostik), respectively.

2.4 RNA extraction and cDNA synthesis

Total RNA was extracted from mouse tissues using ISOGEN (Wako) according to the manufacturer’s instructions. After treatment with DNase (Invitrogen), cDNA was synthesized with or without the Moloney murine leukemia virus, reverse transcriptase (Invitrogen), and oligo(dT)\textsubscript{12–18} primer. Specific primers for NaPi transporters, Cyp27A1, Cyp24A1, IL-6, IL-1β, and GAPDH were used for the PCR reactions. The PCR primer sequences are shown in Table 2. To check for genomic DNA contamination, a reverse transcriptase negative control experiment was performed (data not shown).

2.5 Protein sample purification

Brush border membrane vesicles (BBMVs) were prepared from kidney and intestine using the Ca\textsuperscript{2+} precipitation method, and used for immunoblotting and Pi transport analyses, as described previously (Furutani et al., 2013; Ikuta et al., 2018; Schlingmann et al., 2016).

2.6 Transport assay

Transport of \textsuperscript{32}P into BBMVs was measured by the rapid filtration technique as described previously (Ikuta et al., 2018; Segawa et al., 2004). The transport rate of Pi into the kidney BBMVs was determined at 30, 60 and 120 s at 25°C with an inward gradient of 100 mM NaCl or 100 mM KCl and 0.1 mM KH\textsubscript{2}PO\textsubscript{4} (pH 7.5). The Na\textsuperscript{+}-dependent Pi transport activity (activity rate in the presence of Na\textsuperscript{+} in the absence of Na\textsuperscript{+}) is shown. All measurements were performed in triplicate.

Intestinal absorption was assessed on the basis of the \textsuperscript{32}P blood level after gavage of a test solution using a previously described protocol with modifications (Matsuo et al., 2005; Van Cromphaut et al., 2001). The test solution (pH7.4) contained 128 mM NaCl, 4.7 mM KCl, 2.5 mM CaCl\textsubscript{2}, 1.2 mM MgSO\textsubscript{4}, and 4 mM KH\textsubscript{2}PO\textsubscript{4} (80 μCi/ml). For the study, 5 μl of the test solution per gram body weight was administered by gavage. Blood samples were obtained at the indicated time-points and analyzed by liquid scintillation counting.

| Name                 | Sense primer sequences (5’-3’) | Anti sense primer sequence (5’-3’) |
|---------------------|-------------------------------|----------------------------------|
| 1aOHase (qPCR)      | GAGCAAAACTCCAGGAAGCACAG      | TGAGGAATGACAGGAGAAGG            |
| 240Hase (qPCR)      | TGGGAAGATGTAGGTGGTACCC       | TCGATGCAGGGCTTGACTG             |
| Npt2a (qPCR)        | AGTCTCATTTGGAATTTTGTGCA      | GCCAGTGGCCTCTACCCCT             |
| Npt22 (qPCR)        | TAATCTTCTCAGTCCACCTGTGCT     | CAGTGGAAATTGGCAGTCCTCAA        |
| Npt2c (semi-quantitative PCR) | AGGTCCCCAACCCTACTCTG       | TGCCTAGTAGCTGGAAAGCA           |
| Npt2b (qPCR)        | CCTGGGACCTGGCTGAACCT        | AATGCAGAGGGCTCTCCCTT           |
| PiT1 (qPCR)         | CCCATTGCATGGAAGGAGGA         | GCCACCTGGTGATGTAGTCTG           |
| PiT2 (qPCR)         | CTCAGAAAGGAGCACTGCAAGAG    | AACGTTGACCCGTATCTCCT            |
| IL-6 (qPCR)         | CACAGTGCCAGAGGAGGAGAC       | TTGCCCAGTGACACACCTT             |
| IL-1β (qPCR)        | TGCACACCTTTTTGAGCATG     | ATGTGGTCTCGAGGATTTG             |
| GAPDH (qPCR)        | CTGCACACCAACACTGCTTAGC     | CATCCACAGACCCTCTTGG             |
| GAPDH (semi-quantitative PCR) | i CTGCACACCAACACTGCTTAGC | GGCTGCTTCACACCTTCTT             |
2.7 | Immunoblotting

Protein samples were heated at 95°C for 5 min in sample buffer in the presence of 2-mercaptoethanol and subjected to 8% sodium dodecyl sulfate-polyacrylamide gel electrophoresis. The separated proteins were transferred by electrophoresis to Immobilon-P polyvinylidene difluoride (Millipore) and treated with diluted antibodies. Signals were detected using Immobilon Western (Millipore).

2.8 | Immunofluorescence staining

Mouse kidneys were fixed with the 4% paraformaldehyde solution (pH 7.2) overnight at 4°C, washed with phosphate buffered saline (PBS), cryoprotected with 10% and 20% sucrose at 4°C, embedded in OCT compound (Miles), and frozen in hexane at −80°C. Frozen sections (5 μm thick) were collected onto MAS-coated slides (Matsunami Glass IND, Ltd.) and air-dried (Ohkido, Segawa, Yanagida, Nakamura, & Miyamoto, 2003; Segawa et al., 2005). Serial sections were incubated with primary antibodies overnight at 4°C. Alexa Fluor 488 anti-rabbit (Molecular Probes) and Alexa Fluor 568 anti-mouse (Molecular Probes) were used as secondary antibodies for 60 min at room temperature. Thereafter, the sections were mounted with Dapi Fluoromount G™ (Southern Biotech).

2.9 | Antibodies

Rabbit anti-Npt2a and Npt2c polyclonal antibodies were generated as described previously and used for immunoblotting and immunohistochemistry (Ohkido et al., 2003; Segawa et al., 2005). Mouse anti-actin monoclonal antibody (Millipore) was used as an internal control. Horseradish peroxidase (HRP)-conjugated anti-rabbit or anti-mouse IgG was utilized as the secondary antibody (Jackson Immuno Research Laboratories, Inc, West Grove, PA, USA). The diluted antibodies for immunoblotting were as follows: anti-Npt2a (1:15,000), -Npt2c (1:1,500), and -actin (1:10,000). The diluted antibodies for immunofluorescence staining were as follows: anti-Npt2a (1:500), -Npt2c (1:1,000), and -villin (Millipore) (1:500).

2.10 | Histologic analysis

Mouse tissues were fixed with 4% paraformaldehyde overnight at 4°C and embedded in paraffin. Serial sections (5 μm thick) of several tissues were mounted on MAS-coated slides (Matsunami Glass IND, Ltd.). The sections were treated for hematoxylin and von Kossa staining prior to light microscopic observations. The von Kossa staining for mineral deposits was performed by applying 5% silver nitrate to the sections and exposing them to bright light for 30 min (Schlingmann et al., 2016). The sections were slightly counterstained with hematoxylin.

2.11 | Bone analysis

The femurs of all groups were fixed with 4% paraformaldehyde overnight at 4°C, decalcified for 4 weeks with 10% EDTA, and then, embedded into paraffin for immunohistochemical examinations. For von Kossa staining, tibiae were immersed in a mixture containing 2% paraformaldehyde and 2.5% glutaraldehyde diluted in a 0.067 M cacodylate buffer (pH 7.4) and post-fixed with 1% osmium tetraoxide in a 0.067 M cacodylate buffer for 4 hr at 4°C. After post-fixation, the tibiae were embedded in epoxy resin (Epon 812, Taab, Berkshire, UK) and sliced at a 500 nm thickness using an ultramicrotome.

Immunohistochemical analyses of mouse bone sections were performed as described previously. Briefly, the sections were immersed into 0.3% H₂O₂ in methanol for 30 min to block endogenous peroxidase. To reduce nonspecific binding, 1% bovine serum albumin (BSA; Serologicals Proteins Inc.) in PBS (1% BSA-PBS) was applied to the sections for 20 min. The sections were then incubated with rabbit polyclonal antisera against tissue-nonspecific alkaline phosphatase (ALP) (Oda et al., 1999). We rapidly purified soluble forms of glycosylphosphatidylinositol-anchored proteins using human tissue-nonspecific ALP, or rabbit polyclonal anti-dentin matrix protein-1 (DMP-1; Takara Bio) with 1% BSA-PBS at room temperature for 2 hr as previously described, with some modifications (Oda et al., 1999). The thus treated sections were incubated with HRP-conjugated anti-rabbit

FIGURE 1 | Physiological analysis, renal phosphate transport, and transporter expression in KLKO mice. (a) Serum fibroblast growth factor (FGF)23, (b) plasma 1,25(OH)₂D₃, (c) plasma parathyroid hormone (PTH), (d) blood ionized Ca, (e) plasma Pi, (f) urinary Ca, and (g) urinary Pi excretion. Male mice at 8-weeks of age (n = 5–20) were used. Values are mean ± SE. *p < .05, **p < .01. Metabolic cages were used for 24-hr collection of urine from mice. (h) Renal Na⁺-dependent and (i) -independent Pi transport activity in renal BBMV isolated from the kidneys of 8-week-old WT and KLKO mice. Values are mean ± SE. *p < .05. (j) Western blot analysis of renal BBMV isolated from the kidneys of 8-week-old WT and KLKO mice (n = 3–5). Each lane was loaded with 20 μg of BBMVs. Actin was used as an internal control. Relative intensity of Npt2a and Npt2c expression in WT mice was defined as 1.0. Values are mean relative intensity ± SE. ***p < .01 versus WT mice. (k) Immunofluorescence staining of DAPI (blue), villin (red), and Npt2a or Npt2c (green) in kidney sections of 8-week-old WT and KLKO mice. Sections were prepared from kidneys embedded in OCT compound and frozen. Scale bar; 100 μm
antibody (Chemicon International, Temecula) for 1 hr, and thereafter, the immunoreactivities were visualized by using diaminobenzidine tetrahydrochloride as a chromogen.

For double detection of ALP and tartrate-resistant acid phosphatase (TRAP), the sections immunostained for ALP were incubated with a mixture of 8 mg of naphthol AS-BI
phosphate (MilliporeSigma), 70 mg of red violet LB salt (MilliporeSigma), and 50 mM L(+)-tartaric acid (0.76 g; Nacalai Tesque) diluted in 60 ml of a 0.1 M sodium acetate buffer (pH 5.0) for 20 min at room temperature. Methyl green was used to counterstain in all sections.

2.12 Statistical analysis

Data are expressed as means ± SE. Differences among multiple groups were analyzed by ANOVA. The significance of differences between the two experimental groups was established by ANOVA followed by Student's t-test. A p value of less than .05 was considered significant.

3 RESULTS

3.1 Physiologic analysis, renal phosphate transport, and transporter expression in KLKO mice

The phenotypes of the KLKO mice were confirmed at 8 weeks of age (Figure 1). Compared with Klotho +/+ (WT) mice, KLKO mice exhibited high plasma concentrations of FGF23 and 1,25(OH)2D3, hyperphosphatemia, and hypercalciuria, but not ionized Ca or PTH, as described previously (Figure 1a–f) (Ohnishi et al., 2009a). Urinary Pi excretion was significantly lower in KLKO mice than in WT mice (Figure 1g).

Renal Na+-dependent Pi transport activities were significantly higher in KLKO mice than in WT mice (Figure 1h). Na+-independent Pi transport activities did not differ between WT and KLKO mice (Figure 1i). Npt2a protein expression levels did not differ significantly between WT and KLKO mice (Figure 1j). In contrast, Npt2c protein expression levels were significantly higher in KLKO mice than in WT mice (Figure 1j). Npt2a and Npt2c protein expression was confirmed by immunofluorescence staining (Figure 1k). Npt2a and Npt2c mRNA levels were not significantly different between WT and KLKO mice (Figure S1).

3.2 Biochemical analysis of KL2cDKO mice

Feeding of a low Pi diet can rescue the kl/kl mouse phenotype (Morishita et al., 2001;Segawa et al., 2007). In the present study, we confirmed the effects of a low Pi diet on KLKO mice (Figure S2). KLKO mice were fed a low Pi diet from 8 weeks of age. A low Pi diet increased the body weight and extended the lifespan of KLKO mice, similar to that of kl/kl mice (Figure S2a). After 8 days, a low Pi diet significantly decreased plasma Pi levels in KLKO mice compared with baseline levels before starting the test diet (Figure S2b).

To clarify the role of Npt2c in hyperphosphatemia in KLKO mice, we generated α-klotho −/−Npt2c−/− (KL2cDKO) mice for comparison with the α-klotho −/−Npt2a−/− (KL2aDKO) mice (Figure 2). Disruption of Npt2c in KLKO mice extended the lifespan of the KLKO mice (Figure 2a). KLKO mice had a significantly lower body weight than WT mice (p < .01 versus 4–14-week-old WT mice). KL2aDKO and KL2cDKO mice also had significantly lower body weights than WT mice (KL2aDKO; p < .01 versus 3–20-week-old WT mice, KL2cDKO; p < .05 versus 4–20-week-old WT mice). KL2aDKO and KL2cDKO mice had significantly greater body weights, however, than KLKO mice (KL2aDKO or KL2cDKO; p < .05 versus 12-week-old KLKO mice Figure 2b). The body weight gain was similar between KL2aDKO and KL2cDKO mice (Figure 2b).

Plasma FGF23, 1,25(OH)2D3, and Pi levels, and urinary Pi excretion levels of the mice at 8 weeks of age are shown in Figure 2c–f. The phenotypes of the KL2aDKO mice were consistent with the previous description (Ohnishi et al., 2009a). Plasma FGF23 levels were also high in KL2cDKO mice, similar to KLKO and KL2aDKO mice (Figure 2c). KLKO, KL2aDKO, Npt2aKO, and KL2cDKO mice had higher levels of 1,25(OH)2D3 than WT mice (Figure 2d). Real-time PCR of renal 1αOHase and 24OHase mRNA was performed (Figure S3). The 1αOHase mRNA levels were significantly higher in KLKO, KL2aDKO, Npt2aKO, and KL2cDKO mice than in WT mice (Figure S3). In contrast, 24OHase mRNA levels were lower in KLKO, KL2aDKO, Npt2aKO, KL2cDKO, and Npt2cKO mice than in WT mice (Figure S3b). KL2aDKO mice had significantly lower plasma Pi levels than KLKO mice, as described previously (Figure 2e) (Ohnishi, Kato, Akiyoshi, Atfi, & Razzaque, 2011). The plasma Pi levels remained high in KL2cDKO mice, similar to that in KLKO mice (Figure 2f). Ohnishi et al. (2009a) did not measure urinary Pi excretion levels. In the present study, we measured urinary Pi excretion using metabolic cages. KL2aDKO mice had much higher urinary Pi excretion levels compared with the other groups of mice (Figure 2f). Urinary Pi excretion levels were significantly higher in KL2cDKO mice than in KLKO mice (Figure 2f). Hypercalcemia and hypercalciuria were also more severe.
in KL2aDKO mice compared with KLKO mice and the other mouse groups (Figure S3c and d). KL2cDKO mice did not exhibit hypercalcemia, but had higher levels of urinary Ca excretion compared with WT mice (Figure S3c and d). Urinary Ca excretion levels were not significantly different between KLKO and KL2cDKO mice (Figure S3c and d).

### 3.3 Disruption of Npt2c in KLKO mice recovered renal and vascular calcification

Renal and vascular calcification was observed in KLKO mice at 8 weeks of age (Figure 3b, f and j), but not in WT (Figure 3a, e and i) and Npt2cKO (Figure 3d, h and l) mice on the basis of von Kossa staining, as described previously (Segawa, Onitsuka, Kuwahata, et al., 2009). Disruption of Npt2c in KLKO mice at 8 weeks of age abolished the renal and vascular calcification (Figure 3c, g and k). The kidneys and arteries of elderly mice were evaluated with von Kossa staining (Figure 3m–r). Plasma Pi levels of 40-week-old KL2cDKO mice also showed hyperphosphatemia (Plasma Pi concentration WT 7.36 ± 0.48 mg/dl, KL2cDKO 12.4 ± 0.75, Npt2c 7.49 ± 0.52, n = 4–8). No renal and vascular calcification, however, was observed in 40-week-old KL2cDKO mice (Figure 3n and q).

### 3.4 Npt2c plays an important role in KLKO mice during the juvenile period

Because Npt2c is highly expressed during the juvenile period, we previously reported that Npt2c is a growth-related Pi transporter (Ohkido et al., 2003; Segawa et al., 2002). KLKO mice...
at 5 weeks of age showed hyperphosphatemia and hypercalciumia, but not hyperphosphaturia or hypercalcemia (Figure 4a and b, and Figure S4a and b). The Npt2a protein expression levels were significantly lower in 5-week-old KLKO mice compared with WT mice (Figure 4c). In contrast, the Npt2c protein expression levels were significantly higher in 5-week-old KLKO
mice compared with WT mice (Figure 4c). Furthermore, Npt2a and Npt2c protein expression in 5-week-old KLKO mice was confirmed by immunofluorescence staining (Figure 4d). Npt2a mRNA levels were significantly lower in KLKO mice than in WT mice at 5 weeks of age (Figure S4c). Npt2c mRNA levels, however, were not different between WT and KLKO mice at 5 weeks of age (Figure S4e).

Disruption of Npt2c in KLKO mice at 5 weeks of age significantly decreased the plasma Pi levels compared with KLKO mice, but plasma Pi levels were significantly higher in KL2cDKO mice than in WT and Npt2cKO mice (Figure 4e). Urinary Pi excretion did not differ significantly among groups (Figure 4f). Furthermore, KL2aDKO and Npt2aKO mice exhibited hypercalcemia and hypercalciuria compared with the other mouse groups (Figure S4d and e). KL2cDKO mice had slight, but significant hypercalcemia compared with WT and KLKO mice (Figure S4d). Furthermore, KL2cDKO mice had hypercalciuria compared with WT mice, but were not different from KLKO mice (Figure S4e). KL2aDKO and KL2cDKO mice at 5 weeks of age had higher 1,25(OH)₂D₃ levels than WT mice (Figure 4g). Furthermore, the 1,25(OH)₂D₃ levels were significantly suppressed in KL2cDKO mice, but not in KL2aDKO mice compared with KLKO mice (Figure 4g).

Real-time PCR of renal 1αOHase and 24OHase mRNA was performed (Figure S4f and g). The 1αOHase mRNA levels tended to be higher in KLKO, KL2aDKO, Npt2aKO, and KL2cDKO mice than in WT mice (Figure S4f). In contrast, 24OHase mRNA levels tended to be lower in KLKO, KL2aDKO, Npt2aKO, KL2cDKO, and Npt2cKO mice than in WT mice (Figure S4f). Real-time PCR on inflammation markers, interleukin (IL)-6 and IL-1β, are shown in Figure 4h and i. Disruption of Npt2c in KLKO mice significantly decreased the IL-6 and IL-1β mRNA levels to those in KLKO mice (Figure 4f and g).

3.5 Bone histochemical analysis

Bone histochemical analysis was performed in 5-week-old mice (Figure 5). The bone volume of the femoral metaphyses in Npt2aKO and KL2aDKO mice seemed to be markedly reduced, while those in KLKO, Npt2cKO, and KL2cDKO mice appeared to be similar or slightly decreased compared with those in WT mice (Figure 5A–L).

Von Kossa staining to evaluate the metaphyseal mineralization in Npt2aKO and KL2aDKO mice revealed markedly reduced metaphyseal trabeculae in both groups (Figure 5O, P, C, d). Npt2aKO mice exhibited the broad areas of unmineralized bone matrix in the metaphyses, while KL2aDKO mice showed only slightly unmineralized bone matrix in the corresponding areas, indicating that bone mineralization of KL2aDKO mice seemed to have recovered to some extent by Npt2a deficiency (Figure 5O, P, c, d). Unlike Npt2aKO mice, mineralization in metaphyseal trabeculae was similar between Npt2cKO mice and WT mice (Figure 5M, R, a, f). Despite a similar metaphyseal bone volume as in WT, KLKO and KL2cDKO mice exhibited very large of unmineralized bone matrix compared with the WT mice (Figure 5M, N, Q, a, b, e).

Histologically assessment by double immunostaining for ALP/TRAP revealed similar numbers of ALP-positive osteoblasts and TRAP-reactive osteoclasts in all the groups (Figure 5g–l). Interestingly, an intense accumulation of DMP-1 immunoreactivity was observed in the osteocyte lacunae of KLKO and KL2cDKO mice (Figure 5n and q). In contrast, Npt2aKO, KL2aDKO, and Npt2cKO mice showed a distribution pattern of DMP-1 immunoreactivity in the osteocytic lacunar-canalicular system that was similar to that of WT mice (Figure 5m, o, p, r).

3.6 Intestinal Pi absorption in KL2cDKO mice

Intestinal Pi absorption in KLKO mice has not been studied in detail. KLKO mice may have increased intestinal Pi absorption due to the high plasma vitamin D levels. The high vitamin D levels may also contribute to hyperphosphatemia in KLKO mice. Fecal Pi excretion was measured in juvenile (5-week-old) and adult KLKO (8-week-old) mice using metabolic cages (Figure 6a–d). In the juvenile period, KLKO, KL2cDKO, and Npt2cKO mice had higher food intake (mg/g body weight) than WT mice, but there was no difference in fecal Pi excretion (Figure 6a and b). In adults, food intake (mg/g body weight) did not differ between groups, but fecal Pi excretion was significantly lower in KLKO mice compared with WT mice (Figure 6c and d). Disruption of Npt2c in 8-week-old KLKO mice, however, recovered the fecal Pi excretion level compared with 8-week-old KLKO mice (Figure 6d). Furthermore, to confirm the intestinal Pi absorption, ³²P was administered orally and the Pi absorption rate was measured at 8 weeks of age (Figure 6e). The Pi absorption rates (³²P transfer from intestine to the blood) were significantly increased in KLKO mice compared with WT mice (Figure 6e). Disruption of Npt2c in 8-week-old KLKO mice, however, recovered the fecal Pi excretion level compared with 8-week-old KLKO mice (Figure 6d). Furthermore, to confirm the intestinal Pi absorption, ³²P was administered orally and the Pi absorption rate was measured at 8 weeks of age (Figure 6e).

Next, we investigated the reasons for the suppression of fecal Pi excretion in KL2cDKO mice compared with KLKO mice. Npt2c is mainly expressed in the kidney (Ohkido et al., 2003; Segawa et al., 2002). In the present study, we evaluated Npt2c in the mouse intestine. Npt2c mRNA expression was detected in the proximal and distal mouse intestine (Figure 6f). Furthermore, intestinal Npt2c mRNA expression levels were significantly increased in 8-week-old KLKO mice, but not in 5-week-old KLKO mice, compared with WT mice (Figure 6g).
Intestinal Pi transporter mRNA expression is shown in Figure 7. Intestinal PiT1 and PiT2 mRNA levels were significantly increased in KLKO mice compared with WT mice (Figure 7a and b). Intestinal Npt2b mRNA levels were not different in KLKO mice compared with WT mice (Figure 7c). Disruption of Npt2c in KLKO mice suppressed the induction of PiT1 and PiT2 mRNA (Figure 7a and b).

3.7 | Role of Npt2c in controlling blood Pi levels in juvenile mice

Finally, we confirmed renal Npt2a and Npt2c protein expression around the juvenile period (Figure 5a and b). As described previously, Npt2a protein expression gradually increased with growth (Figure 5a and b). In contrast, Npt2c protein expression rapidly increased at 28 days of age (28D; Figure 5a and b). PTH and FGF23 levels were measured in mice at 15, 21, 28, and 60 days (Figure 5c and d). Plasma PTH levels in mice at 21, 28, and 60 days were significantly lower than those at 15 days (Figure 5c). In contrast, FGF23 levels in mice were lower at 21 and 28 days than at 15 and 60 days (Figure 5d). Next, we measured plasma Pi levels in juvenile Npt2cKO mice (Figure 8). In juvenile Npt2cKO mice, plasma Pi levels began to decrease around 15 days of age and showed significant hypophosphatemia at 21 days (Figure 8).

4 | DISCUSSION

In the present study, to investigate the roles of Npt2c on FGF23/klotho signals in Pi homeostasis, we evaluated the KL2cDKO mouse phenotype. High plasma 1,25(OH)₂D₃ levels in KLKO mice may contribute to hyperphosphatemia by enhancing intestinal Pi absorption and renal Pi reabsorption. KLKO mice exhibited increased renal Npt2c protein levels. These phenotypes were also observed in kl/kl mice, but both Npt2a and Npt2c were increased in the kl/kl mice. The KLKO mouse phenotypes were similar to those described in several previous studies (Alexander et al., 2009; Hu et al., 2019, 2011; Kuro-o et al., 1997; Morishita et al., 2001; Ohnishi et al., 2009a; Ohnishi, Nakatani, Lanske, & Razzaque, 2009b; Segawa et al., 2007; Tsujikawa et al., 2003; Yoshida et al., 2002).

The lifespan of both KL2aDKO and KL2cDKO mice was significantly extended compared with that of KLKO mice. In KL2aDKO mice, despite the increased plasma 1,25(OH)₂D₃ levels, we predicted that the increased renal
Pi loss would reduce Pi toxicity and extend the lifespan, as well as improve soft-tissue and vascular calcification compared with KLKO mice. In the present study, a low Pi-diet prolonged the lifespan of KLKO mice. These observations are consistent with the reported phenotypes of KL2aDKO mice (Ohnishi et al., 2009a). In KL2aDKO mice, serum Pi levels are an important determinant of calcification and lowering serum Pi levels can reduce or eliminate...
soft-tissue and vascular calcification, even in the presence of extremely high serum Ca and 1,25(OH)_{2}D_{3} levels compared with KLKO mice. Survival curves indicated that the lifespan of KL2cDKO mice was extended similarly to that of KL2aDKO mice compared with KLKO mice. The plasma Pi levels in adult KL2cDKO mice (8-week-old) did not decrease, however, and plasma 1,25(OH)_{2}D_{3} levels also remained high. Renal Pi excretion was significantly increased in KL2cDKO mice compared with KLKO mice. In addition, adult (8-week-old) KLKO mice had significantly decreased fecal Pi excretion, and increased the levels of intestinal PiT1, PiT2, and Npt2c mRNA compared with WT mice. In vivo {^{32}P} oral administration, to KL2cDKO mice significantly suppressed intestinal Pi absorption compared with that in the KLKO mice. Thus, in KL2cDKO mice, although there was no change in the plasma Pi concentration, the body Pi load was reduced by increased intestinal and renal Pi excretion compared with that in KLKO mice. Such changes are expected to result in the suppression of calcification (kidney and aorta).

On the other hand, plasma Pi levels were reduced in 5-week-old KL2cDKO mice compared with KLKO mice. Renal Npt2c is specifically increased during the juvenile

FIGURE 8 Npt2cKO mice had hypophosphatemia in the juvenile period. Plasma Pi concentration in WT and Npt2cKO mice at 1, 5, 10, 15, 21, and 28 days of age (n = 10–30). Values are mean ± SE, *p < .05 versus WT mice
period, suggesting that Npt2c function is involved in renal Pi reabsorption. Urinary Pi excretion levels, however, were not altered in 5-week-old KL2cDKO mice compared with KLKO mice. Furthermore, previous and current data revealed no change in Pi excretion in 5-week-old Npt2cKO mice. We predicted that organs other than the kidney that express Npt2c contribute to the decreased plasma Pi concentration.

Deletion of Npt2c in KLKO mice did not have a clear recovery effect on bone phenotype. In addition, bone abnormalities in KLKO mice may have been caused by factors such as increased FGF23 due to klotho deficiency rather than maintenance of the plasma Pi concentration, suggesting that it is involved in the retention of Pi in the body. On the other hand, human NaPi-2c, like mouse NaPi-2a, is expected to be an important molecule for maintaining plasma Pi levels. Although patients with HHRH typically present in childhood with rickets and/or nephrolithiasis, patients occasionally present as adults with low bone density (Dasgupta et al., 2014; Dhir, Li, Hakonarson, & Levine, 2017). NaPi-2c may have different roles in the body depending on age.

Finally, Npt2c may contribute to maintaining plasma Pi levels for growth during the juvenile period and may also be important for Pi retention in adults by controlling intestinal and renal Pi absorption. Figure 9 summarizes the KLKO, KL2aDKO, and KL2cDKO mouse phenotypes. Analysis of mouse NaPi-2c may be useful toward understanding the role of NaPi-2c in phenotypically different HHRH patients.

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CONFLICT OF INTEREST

None declared.

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REFERENCES

Alexander, R. T., Woudenberg-Vrenken, T. E., Buurman, J., Dijkman, H., van der Eerden, B. C., van Leeuwen, J. P., … Hoenderop, J. G. (2009). Klotho prevents renal calcium loss. Journal of the American Society of Nephrology, 20, 2371–2379. https://doi.org/10.1681/ASN.2008121273

Bergwitz, C., & Miyamoto, K. I. (2019). Hereditary hypophosphatemic rickets with hypercalciuria: Pathophysiology, clinical presentation, diagnosis and therapy. Pflugers Archiv: European Journal of Physiology, 471, 149–163.

Bergwitz, C., Roslin, N. M., Tieder, M., Loredo-Osti, J. C., Bastepe, M., Abu-Zahra, H., … Juppner, H. (2006). SLC34A3 mutations in patients with hereditary hypophosphatemic rickets with hypercalciuria predict a key role for the sodium-phosphate cotransporter NaPi-IIc.
in maintaining phosphate homeostasis. The American Journal of Human Genetics, 78, 179–192. https://doi.org/10.1086/499409

Dasgupta, D., Wee, M. J., Reyes, M., Li, Y., Simm, P. J., Sharma, A., … Bergwitz, C. (2014). Mutations in SLC34A3/NPT2c are associated with kidney stones and nephrocalcinosis. Journal of the American Society of Nephrology, 25, 2366–2375.

Dhir, G., Li, D., Hakonarson, H., & Levine, M. A. (2017). Late-onset hereditary hypophosphatemic rickets with hypercalcuria (HHRH) due to mutation of SLC34A3/NPT2c. Bone, 97, 15–19. https://doi.org/10.1016/j.bone.2016.12.001

Furutani, J., Segawa, H., Aranami, F., Kuwahara, S., Sugano, M., Bannai, K., … Miyamoto, K. (2013). Dietary inorganic phosphorus regulates the intestinal peptide transporter PepT1. Journal of Renal Nutrition, 23, e11–e20. https://doi.org/10.1053/j.jrn.2012.02.006

Hikone, K., Hasegawa, T., Tsuchiya, E., Hongo, H., Sasaki, M., Yamamoto, T., … Amizuka, N. (2017). Histochemical examination on periodontal tissues of klotho-deficient mice fed with phosphate-insufficient diet. Journal of Histochemistry and Cytochemistry, 65, 207–221.

Hu, M. C., Shi, M., & Moe, O. W. (2019). Role of alphaKlotho and FGFR23 in regulation of type II Na-dependent phosphate co-transporters. Pflügers Archiv - European Journal of Physiology, 471, 99–108.

Ikuta, K., Segawa, H., Hanazaki, A., Fujii, T., Kaneko, I., Shiozaki, Y., … Miyamoto, K.-I. (2019). Systemic network for dietary inorganic phosphate adaptation among three organs. Pflügers Archiv - European Journal of Physiology, 471(1), 123–136. https://doi.org/10.1007/s00424-018-2242-9

Ikuta, K., Segawa, H., Sasaki, S., Hanazaki, A., Fujii, T., Kushi, A., … Miyamoto, K. I. (2018). Effect of Npt2b deletion on intestinal and renal inorganic phosphate (Pi) handling. Clinical and Experimental Nephrology, 22, 517–528. https://doi.org/10.1007/s10157-017-1497-3

Kaneko, I., Segawa, H., Furutani, J., Kuwahara, S., Aranami, F., Hanabusa, E., … Miyamoto, K. (2011). Hypophosphatemia in vitamin D receptor null mice: Effect of rescue diet on the developmental changes in renal Na+–dependent phosphate co-transporters. Pflügers Archiv : European Journal of Physiology, 461, 77–90.

Kaneko, I., Tatsumi, S., Segawa, H., & Miyamoto, K. I. (2017). Control of phosphate balance by the kidney and intestine. Clinical and Experimental Nephrology, 21, 21–26. https://doi.org/10.1007/s10157-016-1359-4

Kuro-o, M., Matsumura, Y., Aizawa, H., Kawaguchi, H., Suga, T., Utsugi, T., … Nabeshima, Y. I. (1997). Mutation of the mouse klotho gene leads to a syndrome resembling ageing. Nature, 390, 45–51. https://doi.org/10.1038/36285

Kuwahara, S., Aranami, F., Segawa, H., Onitsuka, A., Honda, N., Tominaga, R., … Miyamoto, K. (2012). Identification and functional analysis of a splice variant of mouse sodium-dependent phosphate transporter Npt2c. The Journal of Medical Investigation, 59, 116–126. https://doi.org/10.2152/jmi.59.116

Lederer, E., & Miyamoto, K. (2012). Clinical consequences of mutations in sodium phosphate co-transporters. Clinical Journal of the American Society of Nephrology, 7, 1179–1187. https://doi.org/10.2215/CJN.09090911

Levi, M., Gratton, E., Forster, I. C., Hernando, N., Wagner, C. A., Biber, J., … Murer, H. (2019). Mechanisms of phosphate transport. Nature Reviews Nephrology, 15, 482–500. https://doi.org/10.1038/s41581-019-0159-y

Lorenz-Depiereux, B., Benet-Pages, A., Eckstein, G., Tenenbaum-Rakover, Y., Wagenstaller, J., Tiosano, D., … Strom, T. M. (2006). Hereditary hypophosphatemic rickets with hypercalcuria is caused by mutations in the sodium-phosphate cotransporter gene SLC34A3. American Journal of Human Genetics, 78, 193–201. https://doi.org/10.1086/499410

Matsu, A., Negoro, T., Seo, T., Kitao, Y., Shindo, M., Segawa, H., & Miyamoto, K. (2005). Inhibitory effect of JTP-59557, a new triazol derivative, on intestinal phosphate transport in vitro and in vivo. European Journal of Pharmacology, 517, 111–119. https://doi.org/10.1016/j.ejphar.2005.05.003

Morishita, K., Shirai, A., Kubota, M., Katakura, Y., Nabeshima, Y., Takeshige, K., & Kamiya, T. (2001). The progression of aging in klotho mutant mice can be modified by dietary phosphorus and zinc. The Journal of Nutrition, 131, 3182–3188. https://doi.org/10.1093/jn/131.12.3182

Myakala, K., Motta, S., Murer, H., Wagner, C. A., Koesters, R., Biber, J., & Hernandez, N. (2014). Renal-specific and inducible depletion of NaPi-IIc/Scl34a3, the cotransporter mutated in HHRH, does not affect phosphate or calcium homeostasis in mice. American Journal of Physiology Renal Physiology, 306, F833–F843. https://doi.org/10.1152/ajprenal.00133.2013

Nishimura, M., & Naito, S. (2008). Tissue-specific mRNA expression profiles of human solute carrier transporter superfamilies. Drug Metabolism and Pharmacokinetics, 23, 22–44. https://doi.org/10.2133/dmpk.23.22

Oda, K., Amaya, Y., Fukushi-Irie, M., Kinameri, Y., Ohhuye, K., Kubota, I., … Kobayashi, J. (1999). A general method for rapid purification of soluble versions of glycosylphosphatidylinositol-anchored proteins expressed in insect cells: An application for human tissue-nonspecific alkaline phosphatase. Journal of Biochemistry, 126, 694–699.

Ohkido, I., Segawa, H., Yanagida, R., Nakamura, M., & Miyamoto, K. (2003). Cloning, gene structure and dietary regulation of the type-IIc NaPi cotransporter in the mouse kidney. Pflügers Archiv - European Journal of Physiology, 446, 106–115. https://doi.org/10.1007/s00424-003-1010-6

Ohnishi, M., Kato, S., Akiyoshi, J., Atfi, A., & RAZZEAQUE, M. S. (2011). Dietary and genetic evidence for enhancing glucose metabolism and reducing obesity by inhibiting klotho functions. The FASEB Journal, 25, 2031–2039. https://doi.org/10.1096/fj.10-167056

Ohnishi, M., Nakatani, T., Lanske, B., & RAZZEAQUE, M. S. (2009a). In vivo genetic evidence for suppressing vascular and soft-tissue calcification through the reduction of serum phosphate levels, even in the presence of high serum calcium and 1,25-dihydroxyvitamin D levels. Circulation: Cardiovascular Genetics, 2, 583–590. https://doi.org/10.1161/CIRCGENETICS.108.847814

Ohnishi, M., Nakatani, T., Lanske, B., & Razzaque, M. S. (2009b). Reversal of mineral ion homeostasis and soft-tissue calcification of klotho knockout mice by deletion of vitamin D 1alpha-hydroxylase. Kidney International, 75, 1166–1172.

Schlingmann, K. P., Ruminska, J., Kaufmann, M., Dursun, I., Patti, M., Kranz, B., … Konrad, M. (2016). Autosomal-recessive mutations in SLC34A1 Encoding sodium-phosphate cotransporter 2A Cause Idiopathic Infantile Hypercalciemia. Journal of the American Society of Nephrology, 27, 604–614

Segawa, H., Kaneko, I., Takahashi, A., Kuwahata, M., Ito, M., Ohkido, I., … Miyamoto, K. (2002). Growth-related renal type II
Na/Pi cotransporter. *The Journal of Biological Chemistry*, 277, 19665–19672.

Segawa, H., Kaneko, I., Yamanaka, S., Ito, M., Kuwahata, M., Inoue, Y., ... Miyamoto, K. (2004). Intestinal Na-P(i) cotransporter adaptation to dietary P(i) content in vitamin D receptor null mice. *American Journal of Physiology Renal Physiology*, 287, F39–F47.

Segawa, H., Onitsuka, A., Furutani, J., Kaneko, I., Aranami, F., Matsumoto, N., ... Miyamoto, K. (2009). Npt2a and Npt2c in mice play distinct and synergistic roles in inorganic phosphate metabolism and skeletal development. *American Journal of Physiology-Renal Physiology*, 297, F671–F678. https://doi.org/10.1152/ajprenal.00156.2009

Segawa, H., Onitsuka, A., Kuwahata, M., Hanabus, E., Furutani, J., Kaneko, I., ... Miyamoto, K. (2009). Type IIC sodium-dependent phosphate transporter regulates calcium metabolism. *Journal of the American Society of Nephrology*, 20, 104–113. https://doi.org/10.1681/ASN.2008020177

Segawa, H., Yamanaka, S., Ito, M., Kuwahata, M., Shono, M., Yamamoto, T., & Miyamoto, K. (2005). Internalization of renal type IIC Na-Pi cotransporter in response to a high-phosphate diet. *American Journal of Physiology Renal Physiology*, 288, F587–F596.

Segawa, H., Yamanaka, S., Ohno, Y., Onitsuka, A., Shiozawa, K., Aranami, F., ... Miyamoto, K. (2007). Correlation between hyperphosphatemia and type II Na-Pi cotransporter activity in klotho mice. *American Journal of Physiology Renal Physiology*, 292, F769–F779.

Tsujikawa, H., Kurotaki, Y., Fujimori, T., Fukuda, K., & Nabeshima, Y. (2003). Klotho, a gene related to a syndrome resembling human premature aging, functions in a negative regulatory circuit of vitamin D endocrine system. *Molecular Endocrinology*, 17, 2393–2403. https://doi.org/10.1210/me.2003-0048

Urakawa, I., Yamazaki, Y., Shimada, T., Iijima, K., Hasegawa, H., Okawa, K., ... Yamashita, T. (2006). Klotho converts canonical FGF receptor into a specific receptor for FGF23. *Nature*, 444, 770–774. https://doi.org/10.1038/nature05315

Van Cromphaut, S. J., Dewerchin, M., Hoenderop, J. G., Stockmans, I., Van Herck, E., Kato, S., ... Carmeliet, G. (2001). Duodenal calcium absorption in vitamin D receptor-knockout mice: Functional and molecular aspects. *Proceedings of the National Academy of Sciences of the United States of America*, 98, 13324–13329.

Yamamoto, T., Michigami, T., Aranami, F., Segawa, H., Yoh, K., Nakajima, S., ... Ozono, K. (2007). Hereditary hypophosphatemic rickets with hypercalciuria: A study for the phosphate transporter gene type IIC and osteoblastic function. *Journal of Bone and Mineral Metabolism*, 25, 407–413.

Yoshida, T., Fujimori, T., & Nabeshima, Y. (2002). Mediation of unusually high concentrations of 1,25-dihydroxyvitamin D in homozygous klotho mutant mice by increased expression of renal 1alpha-hydroxylase gene. *Endocrinology*, 143, 683–689.

**SUPPORTING INFORMATION**

Additional supporting information may be found online in the Supporting Information section.

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