Review Article

Tailored Extracellular Vesicles: Novel Tool for Tissue Regeneration

Linli Li,1,2 Peipei Wu,1,2 Hui Qian,1,2 Wenrong Xu,1,2 Hui Shi,1,2 and Jiajia Jiang1,2

1Aoyang Institute of Cancer, Affiliated Aoyang Hospital of Jiangsu University, 279 Jingang Road, Suzhou, 215600 Jiangsu, China
2Zhenjiang Key Laboratory of High Technology Research on EVs Foundation and Transformation Application, School of Medicine, Jiangsu University, 301 Xuefu Road, Zhenjiang, Jiangsu, China

Correspondence should be addressed to Hui Shi; shihui@ujs.edu.cn and Jiajia Jiang; jiangjiajia_2001@163.com

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Extracellular vesicles (EVs) play an essential part in multiple pathophysiological processes including tissue injury and regeneration because of their inherent characteristics of small size, low immunogenicity and toxicity, and capability of carrying a variety of bioactive molecules and mediating intercellular communication. Nevertheless, accumulating studies have shown that the application of EVs faces many challenges such as insufficient therapeutic efficacy, a lack of targeting capability, low yield, and rapid clearance from the body. It is known that EVs can be engineered, modified, and designed to encapsulate therapeutic cargos like proteins, peptides, nucleic acids, and drugs to improve their therapeutic efficacy. Targeted peptides, antibodies, aptamers, magnetic nanoparticles, and proteins are introduced to modify various cell-derived EVs for increasing targeting ability. In addition, extracellular vesicle mimetics (EMs) and self-assembly EV-mimicking nanocomplex are applied to improve production and simplify EV purification process. The combination of EVs with biomaterials like hydrogel, and scaffolds dressing endows EVs with long-term therapeutic efficacy and synergistically enhanced regenerative outcome. Thus, we will summarize recent developments of EV modification strategies for more extraordinary regenerative effect in various tissue injury repair. Subsequently, opportunities and challenges of promoting the clinical application of engineered EVs will be discussed.

1. Introduction

Extracellular vesicles (EVs) are nanosized biogenic particles which can be isolated from multiple types of cells, tissues, and body fluids. They contain multiple bioactive molecules such as nucleic acids, proteins, lipids, and metabolites and can mediate intercellular communication in a short- and long-distance way. In addition, their characteristics including nanosize, low immunogenicity and toxicity, and easy crossing of physiological barrier are of great interest [1–3]. EVs derived from various cells play a vital role in tissue regeneration because of biologically active molecules inherited from parent cells and their inherent characteristics. For instance, accumulating studies have demonstrated that EVs derived from mesenchymal stem cells (MSCs) participate in accelerating wound healing, promoting cartilage tissue and bone tissue and neuronal regeneration, attenuating liver and renal damage, and strengthening myocardial regeneration [4–9], the mechanisms of which mainly include antiapoptosis, the promotion of cell proliferation and differentiation and migration, angiogenesis, and immune regulation.

However, the limited therapeutic efficacy, poor targeting ability, low yield for production of naive EVs, and rapid clearance from the body partly restrict EV clinical application. Growing evidences have suggested that EVs can be modified to overcome the obstacles mentioned above, and the modified strategies predominantly involve cargo loading,
targeting modification, the use of EMs which are synthetic extracellular vesicle-mimetic nanovesicles, self-assembly EV like nanocomplex, and combined application of EVs with biomaterials. The direct injection of therapeutic agents like nucleic acids, peptides, proteins, and drugs remains a challenge because these therapeutic agents may have drawbacks including short-life, rapidly being cleared from the body, complications to nontreatment tissue, and inefficient ability to cross physiological barriers like blood-brain barrier (BBB). Increasing studies have demonstrated that encapsulating therapeutic agents into EVs through indirectly incorporating cargos into donor cells or directly packaging cargos into EVs is a feasible strategy to strengthen therapeutic outcome owing to extraordinary inherent characteristics of EVs [10, 11]. And currently, the clinical assessments of EVs secreted from MSCs are undergoing for nanosized delivery in the field of regenerative medicine [12]. Besides, the biodistribution of EVs also affect their repairing effects. While EVs possess an inherent targeting ability compared with other synthetic cargos carrier like liposomes, the targeting ability of EVs are associated with the sources, the membrane components, administration mode of EVs, and pathophysiological condition of host [13]. Increasing studies have demonstrated that better therapeutic effects can be achieved through presenting surface ligands on EVs to enhance tissue and organ targeting ability. The ligands may include targeting peptides, antibodies, several proteins, and aptamers. Furthermore, EVs encapsulating magnetic nanoparticles like Fe₃O₄ and superparamagnetic iron oxide nanoparticles (SPION) and macrophage cell membrane-fused EMs nanovesicles (MF-NVs) are also used to enhance EV targeting ability. As for the production of EVs, Shao et al. have shown that only 1-4 μg of EV proteins are produced from 1×10⁶ MSC cells per day, indicating that the EVs secreted from MSC cells are limited [14]. How to enhance EV yield is necessary for future application in clinic. Some strategies like pH variations or low-oxygen conditions and chemical stimuli are employed to improve EV yield. However, the effect in a long-term brought by these approaches on the physiological properties of EVs need to be further determined [12]. EMs and self-assemble EV like nanocomplex are a feasible method to overcome the difficulties in poor yield of EVs. The retention time of EVs in the body is another consideration for better regenerative outcome. In recent years, EV-based tissue engineering has attracted increasing attention in regeneration medicine because the combination of EVs and biomaterials including hydrogel, scaffolds, and dressings are capable of achieving sustained delivery of EVs, recruit endogenous cells to proliferate, provide space for cell growth, and then result in synergistically enhanced tissue repair effects.

In this review, we highlight the recent advanced strategies of engineering EVs (loading therapeutic molecules and drugs into EVs, strengthening EV targeting ability, designing EV mimetics and EV-mimicking nanocomplex, combining EVs with biomaterials) in various tissue (neural tissue, eye, lung, heart, liver, intestine, pancreas islet, renal, bone, muscle, and skin) (Figure 1). In addition, we provide a perspective on the prospects of modified EVs. We look forward to helping better understand key challenges and opportunities in the application of modified EVs in regeneration medicine.

2. Extracellular Vesicles

Extracellular vesicles are endosome-derived vesicles which can be released by nearly all cells [15, 16]. According to their size, biogenesis, and contents, EVs are mainly classified into three subtypes: exosomes, microvesicles (MVs), and apoptotic bodies. Exosomes are ranged from 30 to 180 nm in size, and the components of exosomes mainly include structural proteins (such as HSP90, Alix, TSG101, CD9, CD63, TSPAN29, and flotillin), specific proteins (such as MCH-I, MCH-II, CD80, CD86, FasL, and TGF-β), microRNAs, mRNA, and other noncoding RNA [17, 18]. In brief, the exosome biogenesis involves multiple processes in which plasma membrane is internalized to form early endosome followed by transition to late endosomes which is also called multivesicular bodies (MVBs) and intraluminal vesicle (ILVs) formation within the endosome. Finally, MVBs fuse with plasma membrane and exosomes are released from the ILVs.

MVs (50 to 1000 nm) are formed by the budding of plasma membrane, and MV markers are characterized by integrins, selectins, and CD40 ligand, and their components include mRNA, miRNA, other noncoding RNAs, and cytoplasmic and membrane proteins [15, 19]. Apoptotic bodies (500 to 2000 nm) contain part of both nuclear and fragments and cell organelles released by cells under the process of programmed death [20].

3. Neurological Disorders

3.1. Spinal Cord Injury. Spinal cord injury (SCI), as the second major contributor of paralysis, leads to temporary or permanent loss of sensory and motor functions and results in massive cell death, inflammation, vascular injury, severe oxidative stress, and glial scar formation at the lesion area [21, 22]. EVs derived from MSCs, M2 macrophage, neural stem cells, and neurons have been reported to have potential to treat SCI via inhibiting neuronal apoptosis, degeneration, inflammatory response, and glial scar formation and promoting axonal regeneration and angiogenesis. Furthermore, modified EVs similarly are employed in the therapy of SCI. miRNAs such as miR-124-3p [23], miR-26a [24], lncRNAs like lncGm37494 [25], and plasmid cDNA such as GIT1 [26] and sonic hedgehog [27] are transfected to donor cells, and then modified EVs are isolated to treat SCI (Table 1). As a result, modified EVs loaded with therapeutic agents greatly enhanced SCI recovery compared with the control group. Although the method of transfecing donor cells to load cargos is simple and widely used, it may affect other molecules in donor cells, and the loading efficiency is limited. Besides gene agents, EVs can also be used as a drug delivery tool to overcome the shortcomings of SCI therapy-related drugs such as short half-life, inefficient capacity to cross BBB, and easy clearance. Berberine-loaded M2 macrophage-derived EVs can efficiently cross BBB to target the injured spinal cord due to macrophage-derived EVs with inherent
ability to target inflammation, and subsequently, this engineered EVs showed a decent anti-inflammatory and anti-apoptotic effect [10]. Moreover, Kim et al. constructed iron oxide nanoparticle- (IONP-) encapsulated EM nanovesicles (NV-IONP) from IONP-treated human MSCs (hMSCs). IONP with magnetic guidance not only endowed NVs with targeting ability but also increase the contents of therapeutic growth factors in NV [28]. Similarly, taking the advantages of EMs and macrophage membranes, macrophage membrane-fused EM nanovesicles (MF-NVs) were generated to improve targeting ability and therapeutic efficiency [29]. Apart from systematic administration, topical transplantation of MSC-derived EVs which were immobilized in a peptide-modified adhesive hydrogel is also employed to comprehensively mitigate SCI microenvironment [30]. Furthermore, multifunctional scaffold capable of retaining paclitaxel delivered EVs within scaffold is designed, and this scaffold has capability in recruiting endogenous neural stem cells to the injured site, enhancing neural regeneration and decreasing scar deposition [21].

3.2. Stroke. Stroke is one of the leading causes of death and disability worldwide [31]. Currently, there is no drugs available for the cure of stroke. Mounting evidences have demonstrated that EVs from MSCs remote ischemic preconditioning cells, endothelial progenitor cells, M2 microglia, and astrocytes are involved in the treatment of stroke mainly through enhancing angiogenesis and neurogenesis. Furthermore, modified EVs from cells transfected therapeutic miRNAs such as miR-126 [32], miR-223-3p [33], miR-17-92 [34], circular RNAs like circSHOC2 [35], NGF mRNA [36], and plasmid cDNA such as CCR2 [37] and CXCR4 [38] have showed enhanced therapeutic effects in contrast with natural EVs. Although loading bioactive molecules into EVs did improve therapy efficacy, increasing targeting ability of EVs to the injured brain is another consideration. Recently, c(RGDyK) peptide (Figure 2) is conjugated on the surface of EVs via bio-orthogonal copper-free click chemistry method, and then cholesterol modified miR-210 is directly delivered into EVs (RGD-EV-miR-210) via incubation. As a result, RGD-EV-miR-210 is demonstrated to be promising NVs which are capable of targeting to the injured brain and promoting microvascular formation, eventually alleviating ischemia brain symptoms. Meanwhile, the method of directly loading cargos via incubation with cholesterol-miR-210 seems to be better in keeping the integrity of EVs compared with several approaches like electroporation and sonication [39]. Furthermore, the click chemistry method is quicker and simpler in contrast with fusing targeted peptide with EV membrane protein [2]. Similarly, Yang et al. generated NGF-EV-RV by cotransfecting RVG-LAMP2B and pCI-neo-NGF plasmids into donor cells,
**Table 1: Modified EVs in the therapy of neurological disorders.**

| Diseases | EV Sources | Specific substrates | Modification methods | Modified EVs | Biological effects | Ref. |
|----------|------------|---------------------|----------------------|--------------|-------------------|------|
| SCI      | BMSC       | miR-26a             | Cell transfection    | miR-26a-EV   | Axonal regeneration and less glial scarring | [24] |
| SCI      | BMSC       | miR-124-3p          | Cell transfection    | miR-124-3p-EV| M2 polarization; antiapoptosis              | [23] |
| SCI      | ADSC       | IncGm37494          | Cell transfection    | IncGm37494-EV| M2 polarization                                | [25] |
| SCI      | BMSC       | GIT1                | Cell transfection    | GIT1-EV      | Less glial scar formation; anti-inflammation and apoptosis | [26] |
| SCI      | BMSC       | Sonic hedgehog      | Cell transfection    | Shh-EV       | Neuronal regeneration                           | [27] |
| SCI      | BMSC       | Fe3O4-treated BMSCs | Magnetic particles; extrusion | Mag-EMs      | EMs accumulation at injured site; more therapeutic cargos packaging | [28] |
| SCI      | UCB-MSC    | Macrophage membrane-fused MSCs | Extrusion | MF-EMs | Targeting ability; anti-inflammation | [29] |
| SCI      | hMSC       | Peptide-modified hydrogel | Biomaterials | pGel-EV | Efficient retention and sustained release of EVs; nerve recovery | [30] |
| SCI      | M2 macrophage | Berberine         | Ultrasonic          | Ber-EV  | Targeted delivery; anti-inflammatory and antiapoptotic effect | [10] |
| SCI      | HucMSC     | PTX; BSP; linearly ordered collagen scaffolds | Incubation; extrusion anchor peptide; biomaterials | LOCS-BSP-PTX-EMs (LBMP) | High retention of EMs-PTX within scaffolds; neuron formation | [21] |
| Stroke   | BMSC       | miR-17-92           | Cell transfection    | miR-17-92p-EV| Axon-myelin remodeling and electrophysiological recovery | [34] |
| Stroke   | IPAS       | circSHOC2           | Cell transfection    | circSHOC2-IPAS-EV | Antiapoptosis and less neuronal damage | [35] |
| Stroke   | HEK293     | NGF; RVG peptide   | Cell transfection; fuse targeted peptide with LAMP2B | NGF-RVG-EV | Targeted delivery; anti-inflammation; cell survival | [36] |
| Stroke   | BMSC       | c(RGDyK) peptide; cholesterol-modified miR-210 | Click chemistry; incubation | MiR-210-RGD-EV | Targeted delivery; angiogenesis | [39] |
| Stroke   | BMSC       | Fe3O4-harboring BMSCs | Magnetic nanoparticles; serial extrusion | Mag-EMs | Targeted delivery | [40] |
| Stroke   | Macrophage | Edaravone           | Incubation           | Edv-EV     | Improvement of Edv bioavailability and brain targeting; neuroprotection effect | [11] |
| PD       | Murine DC  | shRNA minicircles; RVG peptide | Electroporation; fuse RVG with LAMP2B | shRNA-MC-RVG-EV | Targeted delivery; less alpha-synuclein aggregation | [43] |
| PD       | HEK293T    | DNA aptamers; RVG peptide | Aptamer; fuse targeted peptide with LAMP2B | Apt-RVG-EV | Targeted delivery; less α-synuclein aggregates and motor impairments | [46] |
| PD       | imDC       | RVG peptide; curcumin; ANP; siSNCA | Self-assembly EV-like nanocomplex | C/ANP/S-REV | Targeted delivery; nanoscavenger for clearing α-synuclein and less immune activation | [50] |
| PD       | Serum      | Dopamine            | Incubation           | DA-EV      | More brain distribution of dopamine effects | [49] |
| PD       | Mononuclear phagocyte | Catalase           | Incubation; permeabilization; freeze-thaw cycles; sonication; extrusion | CAT-EV | Anti-inflammation | [48] |
| AD       | HEK-293T and BMSC | miR-29b         | Cell transfection    | miR-29b-EV | Less the pathological effects of amyloid-β (Aβ) peptide | [52] |
| AD       | Plasma     | Quercetin           | Ultrasound incubation | Que-EV  | Improvement of brain targeting and Que bioavailability | [55] |
| AD       | Macrophage | Curcumin           | Pretreated donor cells | Cur-EV | Improved bioavailability of cur; less phosphorylation of the tau protein | [51] |
and NGF-EV-RV not only showed efficient targeting ability to injured region but also decreased inflammation, promoted cell survival, and increased the population of doublecortin-positive cells [36]. Apart from targeting peptides, magnetic NVs (MNVs) are also fabricated via incorporating IONPs into MSC, and then MNVs are made by means of extrusion. IONP enables MNVs to possess excellent targeting ability, upregulate the expression of grow factors, significantly decrease infarction volume, and improve motor function of the brain [40]. Edaravone (Edv) is a type of clinical drugs primarily used for the treatment of brain infarction. It is reported that macrophage-derived EVs encapsulating Edv are capable of releasing drugs at ischemic region, thereby improving bioavailability and neuroprotective effect of Edv and decreasing ischemic cerebral infarction [11]. In addition, Tian et al. constructed a simple, quick, and efficient drug delivery system via conjugating c(RGDyK) peptide to the surface of EV (cRGD-EV), and curcumin was loaded onto the cRGD-EV (cRGD-EV-cur). cRGD-EV-cur significantly inhibited the inflammatory response and cellular apoptosis in the injury region [41]. In clinic, numerous drugs are being “trialed” to treat stroke, loading these drugs to EVs with targeting ability to injury brain hold great potential in augmenting drug therapeutic effect.

3.3. Parkinson’s Disease. Alpha-synuclein (α-syn) aggregates play a key role in the pathogenesis of Parkinson’s disease (PD) which is the second most prevalent neurodegenerative disorder worldwide [42, 43]. Thus, decreasing α-syn expression is an essential target for the treatment of PD. In recent years, the EV-based delivery system of gene to downregulate the α-syn expression or reduce the α-syn pathological aggregates has emerged as a crucial tool for the therapy of PD. Specifically, shRNA minicircles (shRNA-MCs), a type of double-stranded DNA vectors which are smaller than plasmid and have longer term gene silencing function compared with siRNA, were loaded into RVG modified EVs by electroporation. This engineered strategy achieved specific delivery of shRNA-MCs into the brain and decreased α-syn aggregation [43]. Notably, it is necessary to evaluate the EV integrity and the change of shRNA-MCs when it comes to the method of electroporation. In recent years, there emerges a novel agent, aptamer, which is also called “chemical antibody” with targeted therapy and diagnostic ability. More importantly, aptamer with high specificity and affinity are low toxic, low immunogenic, smaller, and more extraordinary in tissue penetration in contrast with antibody [44, 45]. DNA aptamers that specifically recognize α-syn and retard preformed fibril recruiting endogenous α-syn into pathologic aggregates were packaged into EVs which were modified with the neuron-specific RVG peptide as well, and then the aptamer-loaded RVG-EVs greatly decreased pathological α-syn aggregates and improved motor dysfunction [46]. Aptamer-loaded RVG-EVs maybe a hopeful candidate for PD treatment. Apart from the application of shRNA-MCs and aptamer, bone marrow MSC- (BMSCs-) derived EV-mediated delivery of antisense oligonucleotides (ASO) that selectively ameliorate the α-syn expression also provide an alternative for the treatment of PD [47]. Moreover, catalase-loaded EVs secreted by monocytes macrophages and dopamine-loaded blood derived EVs have neuroprotective effects on PD [48, 49]. Based on gene therapy, Liu et al. recently constructed a multifunctional core-shell delivery platform named “nanoscavenger” to achieve synergetic therapeutic effect of gene and chemical drugs, which is composed of immature dendritic cell- (imDC-) derived EVs modified with RVG peptide, curcumin, phenylboronic acid-poly (2-(dimethylamino) ethyl acrylate) nanoparticle, and small interfering RNA targeting SNCA (REV-C/ANP/S). Among these components, core part consists of C/ANP/S which loaded simultaneously siRNA targeting SNCA (siSNCA) with the capability in attenuating the α-syn protein expression and curcumin with the ability in reducing the existing α-syn aggregates. RVG modified imDC-EV’s served as the shell which not only have functions in crossing BBB, targeting lesion area, increasing the drugs bioavailability, and decreasing systematic toxicity but also exhibit ability in clearing immune activation because of the existence of imDC-EVs [50]. This platform not only successfully delivered gene-chem cargos to lesion region in a collaborative and targeted way but also increased loading efficiency compared with direct packaging of cargos into EVs.

3.4. Alzheimer’s Disease. Alzheimer’s disease (AD) is the most common type of dementia, and growing evidences suggest that AD is associated with deposition of the amyloid-beta protein (Aβ) and neuronal fiber tangles [51]. Modified EVs mainly involve gene and drug loading and targeting peptide modification in the treatment of AD. Specifically, EVs packaging miR-29 secreted from cells transfected with miR-29 have enhanced therapeutic effects on spatial learning and memory in animals [52]. Natural EVs passively target and accumulate in some specific organs such as the liver and spleen depending on its inherited property, therefore decreasing its targeting efficiency in other organs and weakening drug therapeutic efficacy in disease treatment, especially in central nervous disease therapy. And a variety of therapeutic EVs were modified by specific recognizable
ligands, aptamer, etc. to achieve drug targeted delivery. In recent years, plasma-derived EVs were found to possess the unique properties including the innate ability of crossing the BBB and immunologic inertia; more importantly, plasma-derived EVs not only improve the bioavailability of drugs but also can achieve drug brain targeted delivery across BBB because some peptides inherited by plasma-derived EVs can specifically bind to receptors in the brain, thereby increasing the accumulation of EVs in the brain [53–55]. Therefore, considering that quercetin (Que) is promising in strengthening cognitive ability and the capability of plasma derived EVs in achieving drug brain targeted delivery and improving drug bioavailability, Que is loaded into plasma EVs with inherent brain targeting capability (EV-Que), and then Que endowed EV-Que with neuroprotective effects via inhibiting CDK5-mediated Tau phosphorylation [55]. EVs derived from curcumin primed cells are also reported to be capable of relieving the symptoms of AD [51]. In light of the capability of MSC-derived EVs to treat AD, Cui et al. proposed to use RVG peptide to modify EVs to realize targeting to cortex and hippocampus region and then significantly improved learning and memory capabilities in mice [56]. Moreover, for alleviating symptoms of AD, RNA aptamers that selectively bind membrane Ap42 were also packaged into EVs to reduce reactive oxygen species production [57].

4. Eye Diseases

In proliferative diabetic retinopathy (PDR), endothelial-tomesenchymal transition (EndoMT) can lead to the occurrence of pathological fibrosis [58]. It has been demonstrated that EndoMT in DR can be suppressed by EVs collected from MSC overexpressing IncRNA SNHG7, thereby delaying DR progression [59]. Moreover, miR-486-3p modified EVs secreted from BMSCs also enhanced the treatment efficacy of DR through inhibiting cell oxidative stress, inflammation, and apoptosis and promoting proliferation via TLR4/NF-κB axis [60]. Subretinal fibrosis, the end stage of neovascular age-related degeneration which can lead to severe and irreversible vision loss and subretinal fibrosis, is associated with epithelial-mesenchymal transition (EMT). Recently, Li et al. found that human umbilical cord MSC- (HucMSCs-) derived EVs encapsulating miR-27b can efficiently suppress EMT via miR-27b/ HOXC6 axis [61]. Suppressing abnormal blood vessel growth is an important aspect for the treatment of some ocular diseases such as retinopathy of prematurity (ROP) and PDR. KV11, an 11-amino acid peptide, shows an anti-angiogenesis effect in vitro and in several animal models. Meanwhile, considering that EVs may preferentially home to the cells from which they are secreted, KV11 was linked to epithelial cell- (EC-) derived EVs via an anchoring peptide, CP05 which can specifically bind to EC membrane protein CD63 (Table 2). KV11-EVs showed more excellent therapeutic outcome. Specifically, KV11-EVs were more efficiently delivered to the blood vessels of the mouse retina, more effective in inhibiting neovascularization and vessel leakage, and showed stronger effect on inhibiting VEGF-downstream signaling than KV11 alone. And the therapeutic effect of KV11-EVs was comparable to the intravitreal injection of VEGF-trap. More importantly, the administration mode of retroorbital injection in this system is safer and less invasive compared with intravitreal injection with single KV11 [62]. KV11-EVs may also be used for the therapy of other diseases associated with pathological angiogenesis, and it would be possible to add several other antiangiogenesis peptides to modify EVs. Taking advantage of anti-inflammation property of EVs derived from regulatory T cells (Treg-EVs) and the ability of VEGF antibody in suppressing VEGF activity, Tian et al. engineered Treg-EVs via conjugating VEGF antibodies to the surface of Treg-EVs to strengthen the repair outcome in choroidal neovascularization (CNV)[63]. As mentioned above, aptamer has more advantages over antibody; thus, we believe that it would be interesting to replace antibody with aptamer notwithstanding the synthesis of aptamer is challenging.

5. Lung-Related Disease

5.1. Acute Lung Injury. Recently, it is reported that EVs from clinical-grade allogenic adipose-derived MSCs (ADSCs) are efficient in the therapy of acute respiratory distress syndrome and healthy volunteers who received the inhalation of EVs have no serious side effects [64]. It demonstrated that EVs may serve as a promising candidate for the therapy of lung-related diseases in clinic. Acute lung injury (ALI) represents a clinical disorder that may associate with inflammation response, oxidative stress, and fibrosis progression [65]. For more pronounced therapeutic effect, EVs were engineered by overexpressing MSCs with miR-30b-3p to relieve inflammation more effectively in ALI mice [66]. Dexamethasone (DEX) has been employed to decrease inflammation in clinic; however, the side effects brought by DEX cannot be ignored. Platelet-derived extracellular vesicles (PEVs) with similar surface glycoproteins and transmembrane proteins of platelets include CD40L, glycoproteins Iba, alIb, and VI, and P-selectin, which endows PEVs with the ability

Table 2: Modified EVs in the therapy eye-related diseases.

| Diseases                        | EV Sources | Specific substrates          | Modification methods                           | Modified EVs            | Biological effects                                    | Ref. |
|---------------------------------|------------|------------------------------|------------------------------------------------|-------------------------|------------------------------------------------------|------|
| Pathological retinal angiogenesis| EC         | Anchoring peptide CP05; KV11 peptide | Link therapeutic peptide to EV via anchoring peptide | KV11-CP05-EV            | Less neovascularization and vessel leakage          | [62] |
| CNV                             | Treg       | VEGF antibody                | Conjugate antibody to EV via a peptide linker (cl) | CL-aV-EV                | Inhibition of inflammation and VEGF activity         | [63] |

Diseases EV Pathological retinal angiogenesis CNV
to recognize and targeted to the inflamed tissue [67, 68]. Therefore, DEX was loaded into PEVs (DEX–PEVs) via incubation to achieve targeted DEX delivery, and this strategy demonstrated a better therapeutic outcome to reduce hyperinflammation in the affected lungs and recover blood oxygen saturation. Moreover, the same dose DEX–PEV treatment in pneumonia mice significantly reduced side effects including abnormal behavioral symptoms of anxiety and irritability in contrast with single use of DEX [68] (Table 3). In addition to using the property of natural EVs to achieve targeted delivery of therapeutic agents to injured lung, RAGE-binding peptide (RBP) with lung-targeting ability, and anti-inflammation effect is displayed on the surface of EVs via fusing with LAMP2B. And then curcumin with hydrophobic peculiarity and ability in decreasing reactive oxygen species is loaded into EVs as well. EVs simultaneously carrying RBP and curcumin succeed in significantly relieving ALI symptom [69].

5.2. Coronavirus Disease 2019. Coronavirus disease 2019 (COVID-19) patient in serious conditions is usually caused by SARS-CoV-2-mediated severe cytokine storm, which contributes to a tissue damage including apoptosis and necrosis and damage to alveolar epithelial cells and vascular ECs, and sustained lung invasion by continuous infiltration of immune cells [70]. Multiple clinical trials are evaluating the efficacy of and MSC-derived EVs in the remission of COVID-19 in critically ill patients. Furthermore, Jamalkhah et al. present an unprecedented strategy of modifying MSC-EVs with interfering RNAs which would hinder viral propagation, inflammation induction, and immune escape in already-infected cells and obstruct the viral particles’ entrance to the uninfected cells and lung tissue to further enhance the antiviral immune [71]. Natural EV-like ginger-derived lipid vehicles (GDLVs) that is isolated from edible ginger plant with low toxicity and can be produced in large scale are nanoparticles similar to EVs containing proteins, lipids, and RNAs, in particular, small-sized RNA and irritability in contrast with single use of DEX [68] (Table 3). In addition to using the property of natural EVs to achieve targeted delivery of therapeutic agents to injured lung, RAGE-binding peptide (RBP) with lung-targeting ability, and anti-inflammation effect is displayed on the surface of EVs via fusing with LAMP2B. And then curcumin with hydrophobic peculiarity and ability in decreasing reactive oxygen species is loaded into EVs as well. EVs simultaneously carrying RBP and curcumin succeed in significantly relieving ALI symptom [69].

| Diseases | EV Sources | Specific substrates | Modification methods | Modified EVs | Biological effects | Ref. |
|----------|-------------|---------------------|----------------------|--------------|-------------------|-----|
| ALI      | BMSC        | miR-30b-3p          | Cell transfection    | miR-30b-3p-EV| Anti-inflammation more effectively | [66] |
| ALI      | Platelet    | DEX                 | Incubation           | DEX-EV       | Targeted delivery and less side effects | [68] |
| ALI      | HEK293 cells| Curcumin; RBP       | Incubation; fuse targeted peptides (RBP) with LAMP2B | RBP-EV       | Targeted delivery and increased therapeutic effect | [69] |
| COVID-19 | HEK-293T cells | miR396a-5p or rlcv-miRl1-28-3p | Incubation | miRNA-EV | Remarkably suppressed inflammation and cytopathic effect | [73] |
| COVID-19 | HEK-293T cells | SiRNA-GFP; RBD | Electroporation; cell transfection with RBD-VSVG fusion vector | SiRNA-GFP-RBD-EV | Targeted delivery | [74] |
| COVID-19 | HEK-293T cells | PM targeting sequence | Fuse PM targeting sequence with ACE2 | PM-ACE2-EVs | Increase the accumulation of ACE2 on EV and block the interaction of virus with cells | [75] |
| IPA      | Fibroblast cell | CLD-loaded liposomes | Hybrid nanovesicles | CLD-hybrid nanovesicles | Targeted delivery of antibiotic drug with high efficiency | [82] |
| AA       | M2 macrophage | Dnmt3a3asmart silence; PLGA nanoparticles | Synthetic EV-like nanocomplex | Dnmt3a3asmart silence | Increase nanocomplex stability; targeted delivery of nucleic acid | [86] |
PM targeting sequence increased the accumulation of ACE2 in EVs, and most of ACE2 are on the surface of PM-ACE2-EVs. The viral load of authentic SARS-CoV-2 was in EVs, and most of ACE2 are on the surface of PM-ACE2-PM targeting sequence increased the accumulation of ACE2.

5.3. Idiopathic Lung Fibrosis. There are few options for idiopathic lung fibrosis (IPF) treatment. Nowadays, only pirfenidone and nintedanib (NIN) are approved by FDA for the treatment for IPF, however, which are palliative and merely delay disease progression [76, 77]. It has been revealed that EVs from diverse cells have therapeutic potential to treat IPF disease [78–81]. In addition, EVs are further engineered for the therapy of IPF. Anti-fibrotic drug incapable of being successfully delivered to fibroblasts is a hurdle in the pulmonary fibrosis therapy. Recently, hybrid NVs composed of liposome loading with clodronate (CLD) and fibroblast-derived EVs are applied to deliver NIN. Among these components, liposome can increase NIN drug encapsulation efficiency. CLD and fibroblast-derived EVs enabled the EV biodistribution in the lung because the former reduces liver uptake of EVs via inducing apoptosis of macrophage at the liver, and thus hybrid EVs preferentially accumulate in the fibrotic lung; the latter shows efficient homing properties to their parent cells [82].

5.4. Allergic Asthma. Allergic asthma is an airway inflammatory disease characterized by bronchial hyperresponsiveness, mucosal edema, and airflow restriction [83]. It is reported that MSC-EVs can promote the immune-suppressive effect of Tregs by upregulating IL-10 and TGF-β1 from peripheral blood mononuclear cells of asthmatic patient [84]. In addition to the interaction of MSC-EVs with monocytes, the administration of MSC-EV by intranasal delivery expands lung IL-10-producing interstitial macrophages, thus contributing to protection against allergic asthma [85]. IncRNA Dnmt3aos plays a key role in M2 macrophages polarization which plays an important part in the occurrence of allergic asthma (AA). Based on this, a nanocomplex composed of smart silencer of Dnmt3aos (Dnmt3aos\textsuperscript{smart silencer}) encapsulated poly lactic-co-glycolic acid (PLGA) core and EV membrane of M2 macrophages shell was exploited. M2 macrophages EV membrane endowed nanocomplex with the ability to achieve targeted delivery of Dnmt3aos\textsuperscript{smart silencer} to injured lung tissue when PLGA improved the stability of nanocomplex thereby effectively delayed AA progression [86].

6. Cardiovascular-Related Diseases

6.1. Myocardial Infarction. Myocardial infarction (MI), namely, myocardial injury due to myocardial ischemia, is a leading cause of morbidity and mortality worldwide [87, 88]. EVs from MSCs, induced pluripotent stem cells (iPSCs), and immune cells like activated CD4\textsuperscript{+} T cells and DC, cardiac progenitor cells, and ECs have been demonstrated to play a pivotal role in cardioprotection after MI mainly via promoting cell proliferation and angiogenesis, ameliorating pyroptosis, and inhibiting inflammatory response and apoptosis. Furthermore, modified EV-mediated regeneration of MI mainly includes loading cargos into EVs by manipulation of parent cells, targeting peptide modification, EMs and EV-mimicking nanocomplex, and the combined use of EVs with biomaterials. For instance, therapeutic cargos like lncRNA KLF3-AS1 [89], miR-210 [90], miR-185 [91], miRNA-181a [92], Akt [93], SDF1 [94], TIMP2 [95], SIRT1 [96], MIF [97], CXCR4 [98], GATA-4 [99], and HIF-1α [100] are used to modify EVs and further enhanced therapeutic efficacy. In order to enhance cardiac-targeting of EVs, and several targeting peptides such as cardiomyocyte specific peptide (CMP), cardiac-targeting peptide (CTP), and ischemic myocardium-targeting peptide were fused with EV membrane protein LAMP2B (Table 4). The targeted peptide modified EVs exhibit increased cardiac retention and strengthened therapeutic efficacy in contrast with non-targeted modified EVs [101–103]. It is well known that low yields along with intricate purification processes of EVs are major hurdles for EVs clinical application. Yao et al. recently generated a type of MSC membrane-camouflaged EV-mimicking nanocomplex. In this nanocomplex, MSC membrane is coated on miR-21 loaded mesoporous silica nanoparticle (MSN) surface. This strategy fully took the advantages of MSC membrane which possesses intrinsic feature of protecting nanoparticles from immune clearance and possessing the capability in specific recognition to targeted cells. Meanwhile, MSN enabled high miRNA loading and effectively protects miR-21 from degradation [104]. Furthermore, apart from the treatment for nervous system-related diseases, IONP-MSC-derived NVs were also used in the therapy of MI [105]. Intriguingly, Liu et al. recently fabricated a “vesicle shuttle” which consists of a magnetic Fe₃O₄ core and a silica shell decorated with PEG corona which conjugated two types of antibodies (one bonded to EV surface membrane protein CD63, the other targeted to myosin-light-chain surface markers on injured cardiomyocytes). Thus, the magnetic-guided “vesicle shuttle” enabled efficient collection, targeted transport and release of EVs, and subsequently improved heart function after MI [106]. Various studies have demonstrated that EVs may be a promising therapeutic tool in MI, but it is reported that the retention of EVs is no more than 3 hours postmyocardial injection [107]. Recently, pluripotent stem cell-induced cardiomyocyte-derived EVs encapsulated hydrogel patch allowed for sustainable and slow release of packaged EVs in a rat model of acute MI and promoted infarcted heart recovery [108].

6.2. Atherosclerosis. Atherosclerosis is associated with EC dysfunction and injury which might be caused by inflammation and reactive oxygen species accumulation [109]. There are emerging studies suggesting that modified EVs also can be used in prevention or therapy of AS. MSC-derived miR-145-rich EVs inhibit HUVECs migration in vitro and reduced atherosclerotic plaque in vivo [110]. Furthermore, EVs from MSCs overexpressing miR-512-3p significantly inhibit ox-LDL-mediated EC damage by regulating the Keap1/Nrf2 signaling pathway [109]. Anti-inflammation is considered as a promising strategy for atherosclerosis
Table 4: Modified EVs in the therapy of cardiovascular related diseases.

| Diseases | EV Sources | Specific substrates | Modification methods | Modified EVs | Biological effects | Ref. |
|----------|------------|---------------------|----------------------|--------------|--------------------|------|
| MI       | hMSC       | IncRNA KLF3-AS1     | Cell transfection    | KLF3-AS1-EV  | Antiapoptosis and pyroptosis | [89] |
| MI       | BMSC       | miR-210             | Cell transfection    | miR210-EV    | Antiapoptosis effects   | [90] |
| MI       | BMSC       | GATA-4              | Cell transfection    | GATA-4-EV    | Antiapoptosis and promotion of cardiac function recovery | [99] |
| MI       | CDC        | CMP                 | Fuse targeted peptide with LAMP2B | CMP-EV | Higher cardiac EV retention; less apoptosis | [101] |
| MI       | BMSC       | Fe3O4-treated BMSCs | Magnetic particles; Extrusion | Mag-EM | Retention of EMs within infarcted heart; cardiac function recovery | [105] |
| MI       | Rat serum  | GMNPtc with anti-CD63 and anti-MLC antibody | Magnetic nanoparticles; antibody | GMNPtc-EV | Recruitment, transport, and targeted delivery of EVs; angiogenesis | [106] |
| MI       | iCM        | Hydrogel patch      | Biomaterials         | Patch-EV     | Sustained delivery of EVs | [108] |
| AS       | M2         | HAL                 | Electroporation      | HAL-EV       | Excellent inflammation-tropism and anti-inflammatory effects | [111] |
| AS       | HucMSC     | miR-145             | Cell transfection    | miR-145-EV   | Inhibition of cell migration and atherosclerotic plaque | [110] |
| AS       | HucMSC     | HucMSC              | Improved extrusion methods | FT/EM        | Higher production yield of FT/EM; anti-inflammation | [112] |

7. Digestive Diseases

7.1 Liver Injury. Currently, there is still no very effective antifibrosis therapy in clinical treatment. The cell-free treatment strategy represented by engineered EVs has brought new hope for the treatment of liver fibrosis. MiRNA (e.g., miR-122 and miR-181-5p) modified EVs by overexpressing ADSCs have a potential in treating liver fibrosis [113, 114]. STAT3 is closely associated with the pathogenesis of liver fibrosis. siRNAs or ASO targeting STAT3 was packaged into clinical grade fibroblast-like MSC-derived EVs by electroporation significantly downregulated STAT3 levels as well as improved liver function in liver fibrosis mice [115]. Although EVs encapsulating these nucleotides did enhance the efficiency in prevention and treatment of liver fibrosis, the efficiency of strategies in loading cargos to EVs mentioned above is limited. Based on this, recent studies by Li et al. established a new engineering strategy for RNA cargos encapsulation by fusing CD9 with human antigen R (HuR) which is an RNA binding protein with capacity in interacting with AU rich elements (AREs) of RNAs. And fused CD9-HuR succeeded in enriching and packaging miR-155 inhibitor with AREs modification into EVs, which subsequently reduced miR-155 level in liver and significantly decreased CCL4 induced liver fibrosis. Moreover, CD9-HuR functioned EV can be utilized to load CRISPR/Cas9 which is difficult to be loaded into EVs by other approaches owing to its long length [116] (Table 5). Thus, CD9-HuR functioned EVs have great potential in RNA delivery of interest because of high loading efficiency. Knockdown of specific RNA is of great importance in the treatment of several diseases, to date, which is predominantly achieved by RNA interference (RNAi). Recently, EVs engineered with LAMP2B-HuR was developed to enrich specific RNA for lysosome degradation, which provide an alternative strategy for RNA degradation in cells particularly macrophage resistant to RNAi. That is to say, this system can be applied for macrophage-related disease therapy [117]. Here, the enhanced therapeutic efficacy has also been found from small molecule drug or cytokine preconditioning of MSC-derived EVs in acute liver injury models. Sun et al. found that melatonin preconditioned-ADSC-derived EVs exhibited better protection against hepatic ischemia-reperfusion...
7.2. Intestine Injury. Emerging studies suggest that gene or protein modified EVs played a vital role in promoting intestine injury repair. miR-326 directly packaged to MSC-EVs protein modification is more scalable and economical [72]. EVs derived from BMSCs transfected with heme oxygen-1 (HO-1) which expression of CD9 [72] and thus provided a new insulin-sensitizing agent for the treatment of metabolic diseases (Table 6). In addition to EV-based gene therapy, BAY55-9837 peptide capable of inducing glucose-dependent insulin secretion however with shortcomings of short half-life, lack of targeting ability, and poor blood GLC response was loaded into EVs which were decorated with SPIONs, namely, BAY-EV-SPION. BAY-EV-SPION overcame shortcomings of single use of BAY55-9837 peptide, thereby significantly augmented insulin secretion [127].

8. Diabetes and Its Related Complication

8.1. Diabetes. Diabetes mellitus (DM) is a metabolic disease, and type 1 and type 2 diabetes mellitus are common in clinic. The T1DM is mainly caused by the elimination of β-cells because of autoimmune destruction, while T2DM occurs because of decreased insulin sensitivity of peripheral tissues and a certain degree of pancreatic islet β-cells damage [123]. It has been demonstrated that MSC-EVs can delay T1DM progression via immune regulation, promotion of β-cell regeneration, and insulin secretion. Moreover, miR-26a in β-cells through circulating EVs reversed obesity-induced insulin resistance and hyperinsulinemia [124]. Meanwhile, there emerged several modified EVs in the treatment of diabetes. EVs transfected with an miR-133b mimic by the Exo-Fect [125] and M2 polarized bone marrow-derived macrophages derived EVs packaged with miR-690 [126] both enhanced glucose tolerance, insulin sensitivity, and thus provided a new insulin-sensitizing agent for the treatment of metabolic diseases (Table 6). In addition to EV-based gene therapy, BAY55-9837 peptide capable of inducing glucose-dependent insulin secretion however with shortcomings of short half-life, lack of targeting ability, and poor blood GLC response was loaded into EVs which were decorated with SPIONs, namely, BAY-EV-SPION. BAY-EV-SPION overcame shortcomings of single use of BAY55-9837 peptide, thereby significantly augmented insulin secretion [127].

8.2. Diabetic Nephropathy. Diabetic nephropathy (DN), one of diabetes complications, is considered as the most severe microvascular complication after DM [128]. Modified EVs in the therapy of DN mainly focused on isolating EVs from MSCs transfected with miRNAs. For instance, BMSC-EVs encapsulating miRNA-let-7a, ADSC-EVs mediated the delivery of miR-215-3p, human urine-derived MSC-EVs overexpressing miR-16-5p, and ADSC-EVs carrying miR-125a further protect against diabetic nephropathy by targeting USP22, ZEB2, VEGFA, and HDAC1 respectively [129–132].

8.3. Diabetic Peripheral Neuropathy. Engineered EVs were also employed in the field of diabetic peripheral neuropathy (DPN), one of the most common chronic complications of diabetes mellitus. Fan et al. developed engineered MSC-EVs via transfecting parent cells with miR-146a, and this modified EVs markedly inhibited the peripheral blood inflammatory monocytes and the activation of ECs via inhibiting Toll-like receptor (TLR)-4/NF-κB signaling pathway [133]. Considering that both biochemical and electrical cues are essential for nerve regeneration, polypyrrole nanoparticles (PpyNps) with electrically conducting are widely exploited both in vitro and vivo for nerve regeneration. Sing et al. fused BMSC-EVs with PpyNps containing liposomes by means of freeze-thaw method, and this hybrid NVs coupled with exogenous electrical stimulation synergistically
enhanced regenerative outcome, offering a new treatment approach in DPN [134].

### 9. Renal Injury

Acute kidney injury (AKI) is defined by many factors that contribute to an abrupt loss of kidney function including a rapid increase in serum creatinine, a decrease in urine production, or both. AKI is a common complication in hospitalized patients with high morbidity and mortality [135]. Moreover, AKI is associated with an increased risk of chronic kidney disease (CKD) and end-stage renal disease. However, there are no definitive therapeutic methods to cure established AKI or prevent it from progressing to CKD. Preclinical studies have demonstrated that multiple cell-derived such as MSCs [136, 137], endothelial colony forming cells [138], renal tubular epithelial cells [139], urinary-derived EVs [140] have good efficacy in the treatment of AKI. And several studies have shown that therapeutic cargos (including proteins, miRNAs, plasmids, and drugs) engineered EVs can enhance the therapeutic efficacy of AKI.

Klotho, a protein that has a protective effect in the kidney, has been reported to be able to treat AKI. Grange et al. loaded Klotho recombinant protein into urinary-derived EVs using the Exo-Fect transfection reagent. EVs loaded with Klotho proteins significantly improved the recovery of renal function in an acute tubular injury model [140] (Table 7). EVs derived from engineered MSCs overexpressing miR-let7c delivered exogenous miR-let7c alleviated renal fibrosis [141]. Furthermore, EVs encapsulating miR-29 administrated by intramuscular injection showed more accumulation in injured-renaled, reduced kidney fibrosis in the CKD model and attenuated muscle wasting which is one of the complications of CKD more evidently [142]. The macrophage-derived microvesicles with inflammation tropism and anti-inflammation effects were applied for targeted delivery of DEX into inflamed kidney to alleviate inflammation and fibrosis [143]. Similarly, IL-10 was loaded into EVs from macrophage for AKI treatment and prevention of CKD considering that IL-10 with inflammatory effects however with several limitations includes instability and tendency to activate leukocytes in the circulation [144]. Moreover, studies have shown that melatonin stimulated MSC-derived EVs enhanced the protective effect of kidney injury in the CKD disease model [145]. In conclusion, these findings demonstrate the effectiveness and rity of a novel therapeutic cargo delivery strategy with promising clinical applications. Besides, to increase EV stability and retention in the treatment of AKI, a class of RGD hydrogels recently was formulated based on that RGD peptide can bind to integrins which are presented on the surface of EVs. And RGD hydrogels showed extraordinary effects on promotion of proliferation, antiﬁbrosis, antiapoptosis, and proautophagy [146].

### 10. Musculoskeletal Diseases

#### 10.1. Osteoporosis

As one of the most common chronic, aged-related disease, osteoporosis, characterized by low bone mass and deterioration in bone microarchitecture, is related to the imbalance of osteoblasts synthesizing bone and osteoclasts breaking down bone [147]. Accumulating evidences demonstrate that EVs could serve as a novel therapeutic tool or biomarker of osteoporosis. For example, as a potential biomarker of osteoporosis, has-circ-0006859 is upregulated in serum of osteoporosis compared health people with high sensitivity and specificity [148] and as a promising therapeutic agent, it has been widely reported that EVs derived from BMSCs can be used to alleviate osteoporosis. Furthermore, as a potential gene delivery tool, EVs from BMSCs packaging lncRNA MALAT1 [149], circRNA RTN4 [150], miR-935 [151], miR-29a [152], and miR-150-3p [153] exhibit stronger therapeutic efficacy compared with naive EVs. The bone-targeting ability of EVs is another aspect, which is associated with the source of EVs. For example, Song et al. found that EVs derived from EC showed distinct superiority in bone targeting ability in contrast with EVs derived from BMSCs and MC3T3[154]. Nevertheless, the specific targeting and therapeutic ability of natural EVs are limited. For example, Luo et al. found that bone marrow stromal cell (ST-) derived EVs (ST-EVs) can strengthen osteoblastic differentiation of BMSCs in vitro. However, the ST-EVs failed to prevent osteoporosis in the osteoporotic mice model because a large number of ST-EVs accumulated in the lung and liver are not bone tissue, and this phenomenon evokes researchers to employ a type of aptamer which was displayed on the surface of EVs (apt-EVs) to target BMSCs of bone marrow and more importantly, this apt-EVs endowed EVs with some excellent therapeutic efficacy in vivo [155] (Table 8). It is critical to inhibit adipogenic differentiation and promote osteogenic differentiation from BMSCs for the therapy of bone loss-related disease. It has been reported
that miR-188 was involved in switch between osteogenesis and adipogenesis, and antagoniR-188 can play a role in promoting bone formation [156, 157]. Hu et al. constructed a type of hybrid NVs via fusing EVs which display C-X-C motif chemokine receptor 4 (CXC4R) on the surface with liposomes carrying antagoniR-188. CXC4R+ EVs could be recruited selectively to the bone marrow which is enriched in stromal cell-derived factor 1 (SDF1), a ligand of CXCR4 and predominantly expressed by BMSCs. Thus, CXCR4 endowed hybrid NVs with targeting ability and antagoniR-188 enabled hybrid NVs to promote bone formation. Moreover, this kind of hybrid greatly strengthened cargo loading efficiency compared with separately using of EVs and greatly enhanced targeting capability in contrast with only application of liposomes [158].

10.2. Osteoarthritis. Osteoarthritis (OA) is the most prevalent type of chronic degenerative joint disease that affects over 300 million people throughout the world [159, 160]. During the development of OA, the pathologic changes in joints may involve cartilage damage, the subchondral bone remodeling, inflammatory activation in the synovium, etc. [161]. The current treatment of OA mainly includes pain management and arthroplasty for end-stage disease [162]. However, these strategies fail to achieve satisfactory results in improving bone homeostasis and delaying OA progression. Recently, the role of EVs has increasingly attracted attention in the therapy of OA, and the studies mainly focus on exploring the diagnostic significance and biological effects of endogenous EVs during OA and therapeutic effects of EVs predominantly from MSCs such as BMSCs, ADSCs, synovial MSCs (SMSCs), and infrapatellar fat pad MSCs [161]. The function of EVs in diagnosis and therapy of OA are mainly ascribed to EV content miRNAs (including miR-6878-3p, miR-210-5p, miR-26a-5p, miR-146a-5p, miR-6821-5p, and miR-92a-3p), lncRNAs (including lncRNA PCGEM1, and lncRNAKLF3-AS1), and proteins like CD73/ecto-5'-nucleotidase [163–167]. In addition, in order to augment OA treatment efficacy, the modified EVs have also been proposed. The modification strategies mainly involve cargo loading, bone-targeting modification, and combined application with biomaterials. miR-210 modified BMSC-EVs exhibit superiority in anti-inflammation and antiapoptosis of chondrocytes compared with BMSC-EVs [168]. EVs derived from MSCs overexpressing miR-92a-3p are capable of strengthening cartilage proliferation and matrix gene expression [167]. It has been suggested that EVs derived from SMSCs played a crucial role in promoting chondrocyte proliferation and migration. Nevertheless, SMSC-EVs have the drawbacks in inhibiting the synthesis of extracellular matrix (ECM) protein including aggrecan and collagen II. To overcome these shortcomings, SMSC-140-5-EVs was fabricated via overexpressing miR-140-5 into SMSCs because miR-140-5 has a function in cartilage homeostasis and targeting RalA to enhance SOX9 and aggregan expression in SMSCs [169]. Apart from loading cargos into EVs by transfecting gene into donor cells, the direct loading of cargos has also been employed (Figure 3). For example, activating transcription factor4, capable of modulating chondrocyte proliferation and bone formation, was loaded into serum EVs derived from OA mice via electroporation [170]. Furthermore, to overcome insufficient targeted delivery of EVs to chondrocytes across the dense, nonvascular ECM of cartilage, Liang et al. loaded miR-140 to DC-derived EVs which were engineered by chondrocyte affinity peptide (CAP) via fusing with the LAMP2B protein on the surface of EVs to realize targeted delivery of miR-140 to chondrocytes [171]. Transplantation of synovial fluid-derived MSCs (SF-MSCs) is critical for OA treatment but is less effective as a cartilage substitute owing to their fibroblastic capability after transplantation [172, 173]. Kartogenin (KGN) is a recently discovered small molecule compound that can mediate SF-MSC-specific differentiation into chondrocytes. However, KGN is characterized by low water solubility, which makes it difficult for accurate administration, easy to form precipitates in the cell, and exhibit low effective concentration thereby limited its chondrogenesis-promoting activity. Xu et al. engineered EVs derived from DCs to achieve targeted delivery of KGN to SF-MSCs, even dispersion of KGN in the cytosol, effective concentration in the SF-MSCs, enhanced chondrogenesis of SF-MSCs by fusing E7 peptide capable of targeting SF-MSCs with EV protein LAMP2B (E7-EVs), and delivering KGN into EVs via electroporation [173]. In the treatment of OA, biomedical scaffolds have also been proposed. The combined application of HucMSC-EVs with acellular cartilage ECM scaffold exhibited better therapeutic effects compared with the single HucMSC-EV group. Specifically, in the EV-scaffold group, the deep layer cells were arranged in a typical vertical band which were similar to the cellular arrangement in natural cartilage, while the cells in the repaired tissue in the human MSC-EV group were

| Diseases          | EV Sources | Specific substrates | Modification methods | Modified EVs       | Biological effects                           | Ref.  |
|-------------------|------------|--------------------|----------------------|--------------------|---------------------------------------------|-------|
| AKI               | Fibroblast | Klotho RGD hydrogels | Exo-Fect agent Biomaterials | Klotho-EV RGD hydrogel-EV | Renoprotective effects Augment retention and stability of EVs and treatment efficacy | [140] |
| AKI               | PL-MSC     | miR-let7c          | Cell transfection    | miR-let7c-EV       | Antifibrosis                                | [141] |
| Renal fibrosis    | BMSC       | miR-29             | Cell transfection    | miR-29-EV          | Antifibrosis                                | [142] |
| CKD               | Satellite cell | miR-29            | Cell transfection    | miR-29-EV          | Antifibrosis                                | [142] |

Table 7: Modified EVs in the therapy of renal injury.
arranged in a disorderly manner, which was significantly different from the normal cartilage structure [174]. Meanwhile, a kind of 3D printed scaffold with radially oriented channels and composed of decellularized cartilage ECM, and gelatin methacrylate (GelMA) hydrogel was loaded with EVs and then significantly facilitated the cartilage regeneration in the OA rabbit model [175].

### 10.3. Fracture

Fracture is a common traumatic injury, the bone itself possesses a certain ability to repair and the regenerative process comprises inflammation, angiogenesis, stem cell differentiation, osteogenesis, and chondrogenesis. However, approximately 5%-10% of fractures are complicated by delayed healing or nonunion [176, 177]. A large number of studies demonstrate that EVs could promote fracture

| Diseases | EV Sources | Specific substrates | Modification methods | Modified EVs | Biological effects | Ref. |
|----------|------------|--------------------|----------------------|--------------|--------------------|------|
| Osteoporosis | BMSC | miR-935 | Cell transfection | miR-935-EV | Osteoblast proliferation and differentiation | [151] |
| Osteoporosis | BMSC | miR-29a | Cell transfection | miR-29a-EV | Robust ability in angiogenesis and osteogenesis | [152] |
| Osteoporosis | ST | DNA aptamers targeting bone | Aptamer | Antagomir-188-hybrid NV | Increase bone-targeting; alleviate bone loss | [158] |
| Osteoporosis | NIH-3T3 cell | circRNA Rtn4 | Cell transfection | Rtn4-EV | Reduced cytotoxicity and apoptosis | [150] |
| OA | BMSC | miR-210 | Cell transfection | miR-210-EV | Proliferation and antiapoptosis | [168] |
| OA | BMSC | miR-92a-3p | Cell transfection | miR-92a-3p-EV | Promote cartilage proliferation | [167] |
| OA | SMSC | miR-140-5p | Cell transfection | SMSC-140-EV | Enhance cell proliferation and migration without damaging ECM secretion | [169] |
| OA | Serum | ATF4 | Electroporation | ATF4-EV | Inhibit chondrocyte apoptosis | [170] |
| OA | DC | MiR-140; CAP peptide | Electroporation; fuse targeted peptide with LAMP2B | miR-140-CAP-EV | Deliver miR-140 to deep cartilage regions and promote bone regeneration | [171] |
| OA | DC | KGN; E7 peptide | Electroporation; fuse targeted peptide with LAMP2B | KGN-E7-EV | Bone-targeting capability and higher cartilage differentiation | [173] |
| OA | HucMSC | ACECM scaffolds | Biomaterials | ACECM-EV | Sustained release of EVs; osteochondral regeneration | [174] |
| Fracture | ATDC5 | VEGF; PCL-scaffolds; anchor peptide CP05 | Cell transfection; anchor peptide; biomaterials | PCL-CP05-VEGF-EV | High grafting efficiency of EVs; osteogenic differentiation; angiogenesis | [183] |
| Fracture | hMSC | 3D Ti-scaffolds | Biomaterials | Ti-scaffold-EV | Bone regeneration ability | [185] |
| Fracture | BMP2-treated macrophage | Titanium oxide nanotubes | Biomaterials | NT-BMP2-EV | Avoid ectopic bone formation; osteogenic differentiation | [186] |
| Fracture | BMSC | TA modified 3D porous SPEEK | Biomaterials | TA-SPEEK-EV | Osteoimmunomodulation effect; sustained release of EVs; osseointegration | [190] |
| Fracture | hMSC | Noggin-suppressed hMSCs; MeGC hydrogel | Cell transfection; extrusion; biomaterials | H-OMN-EM | High yields of EM; robust bone regeneration | [184] |
| DMD | BMSC | IL6ST decoy receptors | Fuse interleukin receptor with EV protein | IL6ST-EV | Block IL6 transsignaling pathway | [194] |
| DMD | NIH3T3 cell | Myostatin propeptide | Fuse therapeutic peptide with CD63 | Myostatin-EV | Increase delivery and propeptide stability | [192] |
recovery via mediating immunomodulation, osteogenesis, and angiogenesis. For example, BMSC-EVs can be encapsulated by ECs and osteoblast cell effectively and accelerate osteogenesis and angiogenesis via BMP-2/Smad1/RUNX2 and HIF-1α/VEGF signaling pathway respectively [177]. Furthermore, MSC-EVs under hypoxia were able to strengthen angiogenesis, proliferation, and migration to a greater extent compared with EVs cultured in normal condition via transferring miR-126 [178]. In recent years, “osteoimmunology” was introduced into bone regeneration, suggesting strong crosstalk between immunology and the skeletal system [179]. For example, M2 macrophage-derived EV miR-5106 can be transferred into BMSCs and induced BMSC osteoblastic differentiation via targeting SIK2 and SIK3 genes [180]. Large segmental bone defect repair based on EVs need to meet the requirements of promoting bone cell proliferation, the reconstruction of internal vasculature, and topical delivery and controllable release of functional EVs at the defect site. Thus, a type of EV-mediated bone scaffold system was constructed. In this system, EVs from ATDC5 overexpressing VEGF were loaded into a class of microscale porous PCL scaffold via a CD63-specific EV anchor peptide CP05 (PCL-Cp05-EV-VEGF). As a result, PCL-Cp05-EV-VEGF was capable of promoting the ingrowth of new tissues, provided a 3D space for vasculature remodeling and better promoted bone regeneration [181–183]. In combined application of modified EVs with biomaterials, EMs were constructed from the noggin-knockdown hMSCs cultured in conditioned osteogenic medium (EMs-OMN) via extrusion. The production of this EMs was more scalable compared with EVs derived from hMSCs, and EM-OMN exhibited robust bone regeneration because the suppressed expression of noggin enhanced osteogenic properties of EMs-OMN. More importantly, a further enhanced osteogenesis in vitro and in vivo was observed in the EM-OMN laden MeGC hydrogel [184]. Osteogenic EVs were loaded into 3D Ti-scaffolds with multiple advantageous properties like biocompatibility, nontoxicity, good mechanical strength, optimal porosity for cell migration and proliferation, and high surface areas for cell attachment. The EV-coated Ti-scaffolds showed more excellent bone regeneration in contrast with EV-free Ti-scaffold implants [185]. Similarly, the titanium nanotubes functionalized EVs from the BMP2-stimulated macrophages promoted osteogenic differentiation and can avoid ectopic bone formation and reduce adverse effects [186]. It is considered that a suitable bone biomaterial should possess ability both in mediating osteogenesis and manipulating the immune response,
thereby exerting a synergistic effect for achieving satisfactory osseointegration [187]. For instance, the implantation of synthetic biomaterials may contribute to an activated M1 phenotype that subsequently secretes multiple proinflammatory cytokines, and the long-term exposure to inflammatory cytokines may eventually lead to osseointegration failure [188, 189]. Conversely, the M2 phenotype macrophages possess capability in secreting anti-inflammatory cytokines, which is favorable for bone regeneration environment formation [188, 190]. BMSC-derived EVs with osteogenic differentiation and immunomodulatory advantages were incorporated on tannic acid (TA) modified sulfonated polyetheretherketone (SPEEK) which can ensure sustained release of EVs and was in favor of improving osseointegration. EV-loaded TA-SPEEK enabled macrophage M2 polarization (an anti-inflammatory phenotype) via the NF-κB pathway, which represented more favorable bone immune microenvironment that was beneficial for further BMSCs osteogenic differentiation [190].

10.4. DMD. Duchenne muscular dystrophy (DMD) is a life-threatening disorder that is caused by the absence of functional dystrophin protein, resulting cell membrane fragility, muscle damage, inflammation, fibrosis, and ultimate degeneration [191, 192]. EVs from cardiomyocyte-derived cells (CDCs) were reported to be a therapeutic candidate for DMD; notably, the obtaining of CDCs is time-consuming and highly technical for isolation and purification maintenance [193]. Systemic administration of EVs derived from hMSCs, murine serum, and myotubes can delay pathological progression via improving membrane integrity in mdx mice without detectable toxicity [193]. However, these approaches were unable to achieve cure effects for DMD. Recently, in order to further augment therapeutic efficiency of EVs for DMD, several engineered EVs have been proposed. Inflammation was considered a major target for DMD therapy, and corticosteroids with capability of decreasing inflammation are regularly used in clinic, but the use of corticosteroids is accompanied with a large number of side effects. IL-6, a key inflammatory cytokine, plays a vital role in skeletal muscle pathophysiology through two different mechanisms: the classical pathways participating in anti-inflammatory and transsignaling pathways conversely, mediating chronic inflammation. In light of this, EVs derived from BMSCs were designed to express IL6 signal transducer decoy receptors to selectively inhibit the IL6 transsignaling pathway and have no effects on classical signaling, thereby provided a potential for the treatment of DMD [194]. More importantly, the importance of the IL6 transsignaling pathway in muscle-related pathologies was first elucidated, and decoy receptor EV platform may combine multiple bioactive molecules or targeting ligands to further enhance various tissue therapeutic efficacy. Myostatin propeptide can play a role in inhibiting mature myostatin. However, the direct administration of myostatin propeptide was restricted for broad application because of safety problem, poor serum stability, and low delivery efficiency. Ran et al. constructed a delivery platform via fusing the inhibitory domain of myostatin propeptide with EV membrane protein CD63, which increased delivery and serum stability of propeptide and enhanced the inhibitory efficacy of myostatin propeptide. As a result, strengthened muscle mass and functional protection without detectable toxicity in mdx mice were achieved [192].

11. Skin Wound

Delayed wound healing and scar formation remains two main challenges in skin wound defects. It has been widely reported that EVs from stem cells such as HucMSCs and menstrual blood-derived MSCs can participate in skin wound healing [195, 196]. The modification of EVs in the therapy of skin wound mainly considers encapsulating cargos into EVs and the combined application of EVs with nanomaterials. As for therapeutic gene delivery, EVs secreted from BMSCs overexpressing TSG-6 significantly enhanced anti-inflammation and alleviated the formation of pathological scar injury [197]. ADSC-EVs greatly enhanced granulation tissue formation and angiogenesis and obviously promoted wound healing [198]. Moreover, direct engineering of EVs is also employed to repair wound defects. For example, miR-21-5p mimics, a novel therapeutic agent for diabetic wound recovery, was transferred into EVs derived from ADSCs by electroporation, and the engineered EVs exhibited excellent therapeutic outcomes in mediating proliferation and migration of keratinocytes through Wnt/β-catenin signaling pathway in vitro and promoted reepithelialization, collagen remodeling, and angiogenesis in diabetic wound models [199] (Table 9). Compared with transfecting cargos into donor cells, electroporation technique may possess the advantages of better encapsulation efficacy, and the process is quick. However, admittedly, it is limited to extensive application because this method can undermine the integrity of EVs thereby influenced the efficiency of wound recovery [200–202].

Emerging studies have been reported to combine EVs with nanomaterials such as hydrogel (Figure 4), metal nanoparticles, and other kinds of dressings to provide a better solution for synergistically enhancing skin tissue regeneration. It is believed that self-healing hydrogels are the most promising wound dressings because they possess the peculiarity of hemostatic ability, self-healing, controlled biodegradation, being injectable, tissue-adhesion, antibacterial activity, anti-ultraviolet, sequential bioactive molecule release, and excellent biocompatibility [203–206]. For example, Xu et al. fabricated a thermosensitive polysaccharide-based FEP hydrogel scaffold with antibacterial activity, fast hemostatic ability, good UV-shielding, and pH-responsive EV release performance. And eventually, this kind of hydrogel enhanced its high ability of promoting diabetic wound healing with less scar formation and skin appendage regeneration [207]. Recently, an extremely effective three-dimensional porous natural-based methyl-cellulose-chitosan hydrogel was proposed to load placental MSC-derived EVs to synergistically promote severe wound healing [204]. Xu et al. found that chitosan/silk hydrogel sponge loaded with platelet-rich plasma EVs was more successful in accelerating wound healing compared with single use of EVs or hydrogel [207]. It is also reported that HUVEC-
derived EVs combined with GelMA hydrogel might provide a potential prospect for accelerated cutaneous wound healing [208]. Apart from hydrogels, other kinds of biomaterials are also used to combine with EVs. An antioxidant wound dressing OxOBand composed of polyurethane was synthesized and supplemented with EVs secreted from ADSCs, which can effectively alleviate hypoxia and oxidative stress, induce angiogenesis, and exhibit faster wound closure [209]. Zhang et al. have suggested that marine sponge Haliclona sp, as a novel microneedle, could provide a safe and effective tool to deliver EVs to the deep layer of skin thus increased the skin absorption of EVs and eventually could produce significant therapeutic effects against skin photoaging in mice [210]. Chitosan-silk fibroin dressing loaded with silver nanoparticles with broad-spectrum antimicrobial ability and EVs from HucMSCs (CTS-SF/SA/Ag-EVs dressing) was able to effectively inhibit the growth of bacterial and enhance wound healing in an infected diabetic wound model [211]. Metal oxide nanoparticle like Fe$_3$O$_4$ with low toxicity, advanced targeting capability, biodegradability, high saturation magnetization, and good biocompatibility was employed to increase the accumulation of EVs at injury site [212, 213]. Recently, Li et al. have successfully constructed IONP-labeled EVs derived from MSCs and found that IONP can serve as a magnet-guided navigation tool, increased the EV accumulation at the cutaneous wound thereby augmented wound healing, reduced scar formation, and increased collagen expression [214].

### 12. Other Diseases

Besides the disease mentioned above, modified EVs are also involved in other diseases. The fetal inflammatory response is associated with neonatal mortality and morbidity, which often results in spontaneous preterm birth (PTB). Accumulating studies are testing to inhibit inflammation through suppressing the inflammatory transcription factor NF-κB pathway; however, they are limited to clinical application partly due to key pharmacological issues such as placental permeability and low efficiency of drug delivery. In light of these, Sheller-Miller et al. and Yim et al. engineered EVs that contained an inhibitor of NF-κB called superrepressor (srIκB) using an innovative tool named "EV for protein loading via optically reversible protein-protein interactions" (EXPLORs) which enabled efficient delivery of protein cargos into the cytosol of target cells through controllable, reversible detachment from the EVs, thus allowing for decreased NF-κB activation and the fetal inflammatory response.

| Diseases                  | EV Sources | Specific substrates | Modification methods | Modified EV | Biological effects                                                                 | Ref.  |
|---------------------------|------------|---------------------|----------------------|-------------|------------------------------------------------------------------------------------|-------|
| Pathological scar         | BMSC       | TSG-6               | Cell transfection    | TSG-6-EV    | Further ameliorated pathological scar                                              | [197] |
| DFU                       | ADSC       | Nrp2                | Cell transfection    | Nrp2-EV     | Enhance wound healing                                                              | [198] |
| DFU                       | ADSC       | miR-21-5p           | Electroporation      | miR-21-5p-EV| Accelerate diabetic wound healing                                                  | [199] |
| DFU                       | SMSC       | miR-126-3p; CS hydrogel | Cell transfection; biomaterials | CS-SMC-126-EV| Angiogenesis                                                                      | [215] |
| DFU                       | ADSC       | FHE hydrogel        | Biomaterials         | EV          | Sustained release of EV; facilitate wound healing                                   | [216] |
| DFU                       | ADSC       | OxOBand dressing    | Biomaterials         | PRP-ZWP/EV  | Accelerate wound healing                                                            | [207] |
| DFU                       | ADSC       | PUAO-CPO-EV         | Less oxidative stress; anti-infection | PU-AO-EV    | Less oxidative stress; anti-infection                                             | [209] |
| DFU                       | ADSC       | FEP hydrogel        | PH-responsive EV release; fasten wound healing | FEP-EV      | PH-responsive EV release; fasten wound healing                                     | [206] |
| Full-thickness skin defects | MEL-5 cell | PD-L1; PF-127 hydrogel | Cell transfection  | PF-127-PD-L1-EV| Fasten reepithelialization                                                        | [217] |
| Full-thickness skin defects | ADSC | Alginate-based hydrogel | Biomaterials      | Alg-EV      | Improve wound closure                                                              | [218] |
| Full-thickness skin defects | HUVEC | GelMA hydrogel      | Biomaterials         | GelMA-EV   | Accelerate wound healing                                                            | [208] |
| Full-thickness skin defects | HucMSC | Silver nanoparticle; CTS-SF/SA | Metal nanoparticle; biomaterials | CTS-SF/SA/Ag-EV| Broad-spectrum antimicrobial activity; accelerate wound healing    | [184] |
| Photoaging                 | HucMSC     | Marine sponge Haliclona sp | Biomaterials      | SHS-EV      | Increased the skin absorption of Exo; significant antiphotoaging effects      | [211] |
| Burn injury                | ADSC       | Fe$_3$O$_4$ nanoparticles | Incubate magnetic particles with parent cells | Fe$_3$O$_4$-EV | Enhanced wound healing in a magnetic targeting way              | [214] |
response and delayed LPS-induced PTB [219, 220]. Moreover, EXPLORs implemented for srlkB loading into EVs were also used to alleviate systemic inflammation in sepsis [221] (Table 10). To improve endometrial regeneration and fertility restoration, an injectable ADSC-EV laden Ag-S coordinated PEG hydrogel was generated, and EV-hydrogel exerted effects on sustained release of EVs, antibacterial activity, promotion of neovascularization, suppression of fibrosis, and increased endometrial receptivity [222]. Triiodothyronine (T3) at low concentration can mediate the oligodendrocyte progenitor cell differentiation and may promote myelin regeneration. However, systematically administered T3 at low concentration failed to reach the injured area and high dosage of T3 resulted in various side effects like peripheral immune reaction and cytotoxicity. Given the advantages of EVs on drug delivery, Xiao et al. loaded T3 into EVs from neural stem cells overexpressing ligand PDGF-A (PDGF-A-EV+ T3), which can target to the lesion of the spinal cord because PDGFR is significantly elevated in demyelinated areas. More importantly, targeted PDGF-A-EV loaded with low dosage of T3 remarkably enhanced the delivery of T3 and significantly delayed experimental autoimmune encephalomyelitis development [223].

13. Conclusions and Future Directions

EVs have shown great potential in multiple tissue and organ (e.g., lung, neuro, brain, skin, diabetes, and eye) regeneration, and clinical trials with allogenic and autologous stem cell derived EVs are underway. Furthermore, modified EVs greatly improved therapeutic outcome. In this review, we summarized various EV engineering strategies in diverse tissue and organs and described their specific characteristics. As cargo carrier, EVs show many advantages over several synthetic nanoparticles such as avoidance of phagocytosis by macrophages because of the existence of “do-not-eat-me” signal CD47 on EV surface, low toxicity, and immunogenicity. In addition, the biodistribution of EVs at injured sites tends to be the premise of successful repairs; thus, diverse approaches involving targeted peptide, several proteins, antibody, magnetic nanoparticles, and aptamer are used to strengthen tissue and organ

TABLE 10: Modified EVs in the therapy of other disease.

| Diseases       | EV Sources | Specific substrates | Modification methods | Modified EVs | Biological effects                                                        | Ref. |
|----------------|------------|---------------------|----------------------|--------------|--------------------------------------------------------------------------|------|
| PTB            | HEK293T    | SrlkB               | EXPLOR system        | SrlkB-EV     | High loading efficiency of cargos postpone infection-induced PTB         | [219]|
| Endometrial damage | ADSC       | PEG hydrogel        | Biomaterials         | Hydrogel-EV  | Angiogenesis anti-infective and anti-fibrotic effect                      | [222]|
| Sepsis         | /          | SrlkB               | EXPLOR system        | SrlkB-EV     | Efficient encapsulation of cargos; anti-inflammation                    |      |
targeting capacity. EMs produced by extrusion, self-assembly EV mimicking nanocomplex, and edible food derived vesicle-like nanoparticles with biocompatibility and stability are more scalable in large production and more efficient in encapsulating therapeutic agents. Considering the retention and therapeutic efficacy of EVs are transient, the bioactive materials like hydrogels, scaffolds, and dressings are promising strategies for precise and sustained release of EVs.

Despite the promising prospect of modified EVs for clinical application, there are still several major challenges needed to be addressed. For EV manufacturing, selecting parent cells and culture condition are two essential aspects which are associated with EV composition, bioactivity, homing property, and production. In addition, understanding the EV comprehensive physicochemical characterization is necessary for EV engineering and safety in clinical application; however, the characterization of EVs remains challenging because of the inherent heterogeneity. In addition, EV isolation methods may affect EV purity, quantity, specificity, and exosomal membrane integrity, which have effects on engineering EV. For cargo loading, the loading efficiency, drug property, and experimental installation, complexity of loading process, and the effects on EV properties are needed to be considered. The targeting modification of EVs may affect the EV membrane protein, cause immune response, and affect the property of recipient cells. Although aptamers are considered as a promising method to achieve targeting ability, high cost and insufficient studies of binding sites restrict further application. For the combined application of EVs biomaterials like hydrogel, dressing, and scaffolds, the biomaterials possess different properties, and the administration mode is also different; thus, it is significant to choose a suitable one for a specific disease, and it is essential to take the properties of biomaterials into consideration including the possibility in leading to immune response. Furthermore, keeping sequential release of EVs is one of the important function of biomaterials, and understanding the dynamic release of EVs in vivo and determining the quantity of EVs loaded into biomaterials may achieve better therapeutic efficacy. In addition, owing to the pathological characteristics of diseases and tissue are different, we believe that developing and choosing an appropriate EV engineering strategy for specific disease therapy is promising for precise medicine. Besides, it would be possible to extend a certain engineering strategy for more extraordinary therapeutic efficacy in multiple diseases, and the coordination of two or more engineering strategies may enable a construction of multifunctional platform which is more potent in tissue and organ regeneration. Notably, the synthesis of platform tends to be more intricated.

In conclusion, modified EVs can serve as a promising candidate for tissue and organ regeneration. Although there are key issues needed to be addressed for modified EV clinical transition, we believe that the difficulties will gradually be solved with the medical research development, and we anticipate that this review will provide new possibilities for better engineering EVs.

**Abbreviations**

| Abbreviation | Description |
|--------------|-------------|
| EVs:         | Extracellular vesicles |
| EMs:         | Extracellular vesicle mimetics |
| MSCs:        | Mesenchymal stem cells |
| MVs:         | Microvesicles |
| MVBs:        | Multivesicular bodies |
| ILVs:        | Intraluminal vesicles |
| BBB:         | Blood-brain barrier |
| SPION:       | Superparamagnetic iron oxide nanoparticles |
| NP:          | Nanoparticle |
| SCI:         | Spinal cord injury |
| IONP:        | Iron oxide nanoparticle |
| hMSCs:       | Human MSCs |
| NVs:         | Nanovesicles |
| Edv:         | Edaravone |
| α-syn:       | Alpha-synuclein |
| PD:          | Parkinson’s disease |
| shRNA-MCs:   | shRNA minicircles |
| BMSCs:       | Bone marrow MSCs |
| ASO:         | Antisense oligonucleotides |
| Shh:         | Sonic hedgehog |
| Mag:         | Magnetic particles |
| MF:          | Macrophage membrane-fused |
| pGel:        | Peptide-modified hydrogel |
| Ber:         | Berberine |
| PTX:         | Paclitaxel |
| BSP:         | Biospecificity peptide |
| LOCS:        | Linearly ordered collagen scaffolds |
| IPAS:        | Ischemic-preconditioned astrocyte |
| RVG:         | Rabies viral glycoprotein |
| RGD:         | Arg-Gly-Asp-D-Tyr-Lys |
| Apt:         | Aptamer |
| DEX:         | Dexamethasone |
| DA:          | Dopamine |
| CAT:         | Catalase |
| Que:         | Quercetin |
| Cur:         | Curcumin |
| AD:          | Alzheimer’s disease |
| DC:          | Dendritic cell |
| PDR:         | Proliferative diabetic retinopathy |
| EndoMT:      | Endothelial-to-mesenchymal transition |
| EMT:         | Epithelial-mesenchymal transition |
| HucMSCs:     | Human umbilical cord MSCs |
| Treg:        | Regulatory T cells |
| CNV:         | Choroidal neovascularization |
| ROP:         | Retinopathy of prematurity |
| ADSCs:       | Adipose-derived MSCs |
| ALI:         | Acute lung injury |
| RBP:         | RAGE-binding peptide |
| DEX:         | Dexamethasone |
| PEVs:        | Platelet-derived extracellular vesicles |
| RBD:         | Receptor-binding domain |
| PM:          | Plasma membrane |
| IPF:         | Idiopathic lung fibrosis |
| CLD:         | Clodronate |
| NIN:         | Nintedanib |
| IIPF:        | Idiopathic lung fibrosis |
| AA:          | Allergic asthma |
| PLGA:        | Polylactic-coglycolic acid |
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Conflicts of Interest

The authors declare no conflict of interest.

Authors’ Contributions

Linli Li and Peipei Wu contributed equally to this work.
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