High Doses of Vitamin C and Leukaemia: In Vitro Update

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Summary
Vitamin C (ascorbic acid) is an essential nutrient with a number of beneficial effects on the human body. Although the majority of mammals can synthesize their own Vitamin C, humans and a few other species, do not produce it and depend on dietary sources for their Vitamin C supply. Among its many effects on cell function and metabolism, Vitamin C has shown, in vitro, a powerful anticancer effect against a number of human tumour cell lines, including myeloid leukaemia. There are many different mechanistic explanations for the anticancer/anti leukemic effects of Vitamin C and the aim of the present review is to illustrate these mechanisms, showing the results of some preliminary in vitro investigations, and outlining their potential clinical relevance.

Introduction
Vitamin C is an essential nutrient with a number of beneficial functions, for the organism [1], such as
1. Helping the metabolism of tyrosine, folic acid and tryptophan,
2. Increasing the elimination of cholesterol,
3. Contributing to the synthesis of catecholamine’s,
4. Helping the body to absorb and breakdown histamine,
5. Enhancing the absorption of non-heme iron,
6. Promoting the synthesis of collagen (its most widely known physiological function),
7. Neutralizing free radicals (it is a reducing agent, “scavenger” of free radicals, and a founder among the natural antioxidants),
8. Protecting DNA from damage due to free radicals and mutagens,
9. Reducing the risk of premature death,
10. Fighting off widespread environmental pollutants,
11. Preventing the development of nitrosamines.

Though ubiquitous, ascorbate is not produced by humans, guinea pigs, some primates, a particular type of fruit eating bat, the majority of fishes and birds [2], who depend on diet for the assumption and use of this fundamental nutrient.

Vitamin C and Leukaemia: Historical Background

The first mention of the therapeutic potentialities of Vitamin C in leukaemia, can be found in the book “The Healing Factor: Vitamin C against disease”, written by the biochemist Irwin Stone, in 1974 [3]. In his book, Stone refers to a study, performed in 1936 by Stephen and Hawley [4], demonstrating, for the first time, that when the blood is separated into plasma, red blood cells, and white blood cells, there is a 20- to 30-fold concentration of Vitamin C in the white blood cells, as compared to plasma.

Following this report, Barkhan and Howard, by studying a few cases of chronic myelogenous and lymphatic

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leukaemia, added the evidence that leukemic patients have substantially lower than normal plasma levels of Vitamin C [5].

As noted by Stone, although this knowledge could suggest the use of Vitamin C as a therapeutic agent, in leukaemia, the first clinical trials showed contrasting results, due to the inappropriately low doses administered.

Later on, Vogt, in a literature review [6], confirmed that there are high deficits of Vitamin C in leukemic patients, as also confirmed by the reports of Kyhos and Coll [7] in 1945, and Waldo and Zipf, in 1955 [8].

Since Vitamin C is a natural component of human nutrition, endowed with innumerable beneficial effects on human body [9-11], it is, to say the least, astonishing, to realize how little scientists have done, in eight decades, to verify the role and potentialities of this nutrient, in the prevention and treatment of human leukaemia.

According to Stone, leukaemia reduces the body stores of Vitamin C to very low levels, and any residual Vitamin C circulating in the blood is scavenged and locked in the excessive numbers of leukocytes usually contained in the blood of these patients. As a direct consequence, the plasma levels of Vitamin C are reduced to zero or close thereto, and tissues are no longer supplied with this most important metabolite, since it is accumulated in leukocytes.

Stone defined "biochemical scurvy" the condition of insufficient Vitamin C supply to body tissues, and proposed that its correction required the administration of Vitamin C at a rate of 25 grams or more per day.

In 2012, seventy-six years after the first observations on the distribution of Vitamin C in leukocytes, an investigation on 131 patients affected by different types of leukaemia, definitively confirmed that leukemic patients have significant lower plasma Vitamin C than normal controls. The reduction of plasma Vitamin C levels in leukaemia, as predicted by Stone, is due to an increased uptake and utilization by the active proliferating leukocytes, leading to tissue biochemical scurvy and consequent increased tendency to bleeding and infections, which are the hallmark of this pathological condition [12,13].

With the above data at hand, it is clear that leukaemia can be viewed as a condition of functional Vitamin C deficiency, associated with biochemical scurvy, and therefore, all leukemic patients are suitable candidates for the treatment with this nutrient.

How Much Vitamin C to Treat Leukaemia? The Concept of "Mega-Doses"

In 1949, Frederik Klenner first reported the successful treatment of bulbar poliomyelitis, with high doses of Vitamin C administered by intramuscular, intravenous and oral route [14].

The same Author had established clinical protocols using massive doses of Vitamin C, to treat a number of different viral conditions [15,16], but it was Stone who formerly defined the concept and the rationale for the use of "mega-doses" (also known as "pharmacologic" doses) of Vitamin C in leukaemia and other cancers.

In particular, Stone observed that man and only a few other species do not produce their own Vitamin C, while the great majority of mammals do, according to their physiologic requirements [17].

This observation led Stone to hypothesize that, due to either insufficient intake or increased consumption of the nutrient, or both, man could easily undergo a condition that he defined "hypoascorbemia". Hypoascorbemia is a reduced amount of circulating Vitamin C (also called "ascorbic acid"), due to the lack of the enzyme L-Gulonolactone Oxidase (GLO), as a consequence of an "inborn error of carbohydrate metabolism" [18-20].

This defect, now very well acknowledged and characterized [21], led Stone to hypothesize that to be in good health, man needs mega-doses (several grams a day), rather than doses in the order of milligrams, as stated by the Recommended Daily Allowances (RDAs) [22-24].

The rationale behind the use of mega-doses of Vitamin C was further refined by the chemist and twofold Nobel Prize, Linus Pauling, who soon became an enthusiastic supporter of the use of this nutrient, in high doses, not only to prevent disease [25-31], but also to treat a number of pathologic conditions, ranging from common cold [32-37], to cancer [38-53], and AIDS [54].

Intravenous Vitamin C and Cancer

Studies on dose-concentration relationship in humans, performed by Levine and co-workers [55], revealed that at oral doses exceeding 250 mg/day, the plasma levels of Vitamin C reach a plateau, and any further increase in the amount administered by mouth, does not determine significant increase in plasma concentration. This is due to multiple "control" mechanisms, including, among others, intestinal absorption, tissue accumulation, renal reabsorption and excretion, and utilization. On the contrary, the intravenous administration of high doses of Vitamin C, bypassing the above control mechanisms, allows plasma concentrations that are 100-fold or higher than maximally tolerated oral doses, and the peak could last for hours within the millimolar (mM) range [56,57].

More importantly, at plasma concentrations easily achievable by intravenous administration (5-10 mM
for 1-2 hours), Vitamin C induced death in 75% of 48 cancer cell lines tested in vitro [58], but had no toxic effect on human peripheral white blood cells, fibroblasts, or epithelial cells. This selective cytotoxic effect would be achieved since at high doses, parenteral ascorbate is a peroxide delivery system for the generation of sustainable ascorbate radical and H$_2$O$_2$ in the extracellular space, with consequent oxidative damage to cancer cells [59,60].

Therefore, high doses of Vitamin C would be cytotoxic to cancer cells because of their pro-oxidant, rather than anti-oxidant effects [61,62], even though, some Authors remark that the pro-oxidant activity of Vitamin C, may not be relevant, in vivo [63-65].

It has been suggested that the selective cytotoxic activity of high doses of Vitamin C against cancer cells, as compared to the normal ones, is due to their reduced levels of antioxidant enzymes, catalase, superoxide dismutase, and glutathione peroxidase. The reduced level of antioxidant enzymes leads to cellular damage by accumulation of hydrogen peroxide [66-69], with subsequent intracellular redox imbalance and oxidative damage to different cellular structures [70-72]. However, the proposed mechanistic explanation of the selective cytotoxic action of Vitamin C on cancer cells has always remained elusive and generic.

More recently, Yun and co-workers [73], by investigating the effects of high doses of Vitamin C on KRAS and BRAF mutants cells derived from Colorectal Cancer (CRC), have further refined the mechanist explanation of the anticancer properties of Vitamin C. In particular, according to the Authors, the death of KRAS and BRAF cell mutants of CRC is not caused by the Vitamin C itself, but rather, by its oxidized form, Dehydroascorbic Acid (DHA). While Vitamin C enters cells though specific receptors, called Sodium-Vitamin C Co-Transporters (SVCTs) [74], DHA competes with glucose, for intracellular uptake by Glucose Transporters (GLUT), mainly 1 and 4 subtype receptors [75,76].

Interestingly, both KRAS and BRAF activating mutations are responsible of the upregulation of GLUT1 expression in different types of cancer, including CRC [77,78]. However, as reported by Yun and coll [73], the upregulation of GLUT-1 expression, is not always associated with increased sensitivity of tumour cell lines to the cytotoxic effects of DHA.

Further investigation into the metabolic makeup of KRAS and BRAF mutations CRC-derived cell lines, showed an accumulation of glycolytic intermediates upstream Glyceraldehyde-3-Phosphate-Dehydrogenase (GAPDH) and a contemporary depletion of the metabolites downstream GAPDH, indicating an inhibition or severe reduction of its enzymatic activity, which appears to be the key of the cytotoxic effect of DHA.

In summary, the data reported by Yun and coll. on the effect of DHA on CRC cell lines, indicate that in glycolysis-addicted KRAS and BRAF mutated cell lines, high amounts of DHA are transported into the cancer cells, through the overexpressed GLUT-1 receptors, and then reduced again to Vitamin C inside the cells. The reduction of DHA to Vitamin C scavenges Glutathione (GSH), thus inducing redox imbalance and oxidative stress. Oxidative stress, in turn, leads to inactivation of GAPDH, inhibition of glycolysis, and energetic crisis, which leads to cell death [79].

More precisely, beyond being inactivated directly by ROS (including H$_2$O$_2$), GAPDH function is also hindered by the depletion of Nicotinamide Adenine Dinucleotide (NAD+), caused by the activation of the DNA repairing enzyme, Poly ADP-Ribose Polymerase (PARP), induced by the DNA. In fact, the increased production of ROS, in cancer cells, due to the high doses of Vitamin C, produces increased DNA damage and consequent activation of PARP. PARP, in turn, consumes NAD+ and NAD+ depletion (together with the consequent ATP depletion) determines the energetic crisis leading cancer cells to death [80].

**Vitamin C and H$_2$O$_2$**

The view that Vitamin C in high concentrations, administered by intravenous infusion, acts as a pro-oxidant, leading to the formation of H$_2$O$_2$, thus inducing oxidative damage to cancer cells, is not new. In 1969, when man first walked on the moon, Benade and co-workers had already demonstrated that Vitamin C could selectively kill cancer cells, without harming normal cells. The Authors suggested that the cytotoxic effect of ascorbate could be due (“in major part”) to the intracellular (not extracellular!) generation of toxic hydrogen peroxide produced upon oxidation of the ascorbate, by the cells (and not by the conversion of DHA back to Vitamin C!). This view, was corroborated by the fact that the toxicity of ascorbate was greatly enhanced by the concomitant administration of 3-Amino-1, 2, 4-Triazole (ATA) that inhibit the enzyme catalase, “thus decreasing or destroying the ability of the cancer cells to detoxify H$_2$O$_2$ effectively” [81].

On the other hand, a number of scientific reports demonstrate that human cancer cells show low levels of antioxidant enzymes (including, among others, catalase and glutathione peroxidase), and therefore cannot detoxify hydrogen peroxide [82-84].

Moreover, accumulating evidence suggests that cancer cells produce high amounts of hydrogen peroxide...
Two years later (1976), Cameron and Pauling showed

In 1974, Irwin Stone had already reported on the
toxic effects of hydrogen peroxide on cancer cells
In no cases, the antiviral properties of Vitamin C have
been ascribed to its presumed pro-oxidant activity, when administered in high concentrations, for the treatment of viral infections;
The toxic effects of hydrogen peroxide on cancer cells
were known since 1957, when Reginald Holman published a paper on “Nature”, showing that rat implanted with Walker 256 adenocarcinoma, and treated by simply replacing their drinking water with 0.45% hydrogen peroxide, showed a rate of cure of 50-60%. The time taken for complete disappearance of the tumour was 15-50 day depending on the tumour size when the treatment was started [94]. Holman’s work was based on the assumption (later confirmed by studies on Vitamin C) that malignant cells are deficient in catalase, and, as such, sensitive to oxidizing agents;
In 1974, Irwin Stone had already reported on the
treatment of leukaemia with high doses of Vitamin C, given by mouth [22], and in the same year, Cameron and Campbell published the results of a clinical trial, showing that, in “untreatable” cancer patients, ascorbate in high doses can bring about some significant improvement in morbidity and mortality [95];

Two years later (1976), Cameron and Pauling showed that the survival of untreatable cancer patients increased by a factor of about 3 in the majority of cases, and about 20 in 10% of them, under treatment with 10 grams of ascorbate per day, starting with the intravenous route, followed by oral administration [48];
Later on, the same authors confirmed, in another article, that 10 or more grams of ascorbate per day, significantly prolong the survival and improve the quality of life of untreated cancer patients [51];
The ten grams proposed as a standard treatment, in these cases, can certainly be considered “high” doses, if compared with the RDAs, but are still much lower than the doses used in the most recent anticancer clinical trials with intravenous ascorbate [96];
The results of the two clinical trials published by Cameron & Pauling, were confirmed by Murata and co-workers, in 1982 [97], and therefore, they cannot be considered isolated reports;
The clinical studies performed at the Mayo Clinic to repeat the experience reported by Cameron & Pauling [98,99], did not follow the method suggested by the two scientists, and therefore failed in its (presumed) intent, even though the results of this work were taken as the “definitive proof” that Vitamin C had no anticancer properties;
The Mayo Clinic work showed, among others, that Vitamin C is ineffective in Colorectal Cancer (CRC), a conclusion definitively debunked by recent evidences reported by Yun and co-workers [72,78]. The Mayo Clinic, on the other hand, always refused to provide Pauling and other Authors with their data;
Abram Hoffer, who worked with Linus Pauling, and treated a number of patients with high doses of oral Vitamin C, obtaining essentially the same results as Pauling and Cameron, wrote “Dr. Pauling risked his enormous scientific credibility by his views on using large doses of vitamins. His work on Vitamin C and the common cold and the flu, and later on the role it plays in controlling the ravages of cancer are well known and, in the opinion of Orthomolecular scientists, are correct. We have come to the same conclusions by observing what Vitamin C has done for tens of thousands of patients. It is the most remarkable anti-stress antioxidant and is finding its place in the treatment of more and more difficult chronic diseases including AIDS, cancer, infections, toxemias, schizophrenia and more” [100,101];
Art Robinson, who, in 1994, published an important article showing that mice with cancer that were given high dose vitamin C in the diet, or fed a diet of raw vegetables, lived up to 20 times longer than controls [102], and all this, happened a long time before the NIH mouse experiment reported by Chen [58-60];
Hugh Riordan and his research team repeated and
The pharmacokinetic studies of Levine and Padayatty, on Vitamin C, indicate that after oral administration of 200 mg of the nutrient, the maximum plasma concentrations obtained, are not superior to 70-80 µM. This is due to a “tight control”, operated by several different mechanisms, including, among others: bioavailability, intestinal absorption, tissue accumulation, renal reabsorption and excretion, and utilization rate as a function of homeostasis. On the contrary, when Vitamin C is administered intravenously, “tight control” is bypassed, until renal excretion restores equilibrium, depending on the dose administered [55-57].

Therefore, according to these data, the intravenous administration of Vitamin C is the only way to achieve plasma concentrations in the order of millimoles, necessary to kill cancer cells.

However, this view is in disagreement with the following evidences:

a) The results obtained by intravenous administration of Vitamin C, do not show the same large effects reported by Robinson, feeding squamous cell carcinoma implanted mice, with large doses of the nutrient [102];

b) Abram Hoffer [100] used oral high doses of Vitamin C in cancer patients and obtained essentially the same significant results as Cameron & Pauling [48-51], Cameron & Campbell [95], and Murata [97];

c) Although Mark Levine maintains that “… only injected ascorbate (Vitamin C) might deliver the concentrations needed to see an anti-tumour effect” [108], neither the legendary scientist Linus Pauling, nor the consultant surgeon, Ewan Cameron, seemed to know the difference between oral and intravenous administration. In fact, in their clinical trial, the protocol started with a few days of 10 grams of intravenous Vitamin C, followed by 10 grams of oral Vitamin C for the whole life [48,51]. Cameron & Campbell, who had already reported on the successful treatment of cancer with oral Vitamin C, two years before Cameron & Pauling [48,51], regarding the administration route, had already observed “… with increasing experience, we tend now to believe that the intravenous regime is probably unnecessary as a routine measure, and need only to be employed in situations where vomiting, anorexia, or other complications of malignancy, preclude oral administration” [95];

d) Plasma concentrations above the 400 µM have been reported, after the administration of a single dose of oral liposomal Vitamin C [109];

e) At times of stress or illnesses (including cancer), the body may absorb extra Vitamin C, as demonstrated by the principle of “bowel tolerance” to the nutrient administered by mouth. According to this principle, when the body is saturated with Vitamin C, a slight diarrhoea may appear, due to intestinal elimination of the nutrient; however, during stress or disease, the amount of oral Vitamin C a patient can tolerate, before the appearance of diarrhoea, increases in proportion with the severity of the condition [110];

This means that the “tight control” hypothesized by Levine and Padayatty, over the plasma concentration of Vitamin C, is either inexistent or relative to disease conditions or stress. To achieve the maximum plasma levels, a typical person may need 20 grams of oral Vitamin C spread throughout the day (3-4 grams every four hours); but cancer patients may require far more [111], and such massive intake may result in plasma concentrations that the tumour may absorb, generation hydrogen peroxide that kills cancer cells.

f) More recently, the paradigm according to which antioxidants inhibit tumorigenesis predominantly by decreasing ROS-mediated DNA damage and mutations [112,113], has been challenged by experimental data, showing that antioxidants such as N-Acetylcyesteine (NAC) and Vitamin C exerts their anti-tumorigenic activity by downregulating HIF-1α [114].

Interestingly, these data were obtained not by injecting, but simply feeding mice with large amounts of NAC and Vitamin C, thus once more validating the role of oral administration of Vitamin C (and other antioxidants), in fighting cancer.

**Vitamin C and Leukaemia: An In Vitro Update**

As we have previously demonstrated, high (“pharmacologic”) concentrations of Vitamin C (in the form of the sodium salt of ascorbic acid) are capable of eliciting a clear-cut pro-apoptotic/cytotoxic effect on human Pro-myelocytic Leukaemia derived cell lines (HL60), *in vitro* [115] (Figure 1 and Figure 2).

This effect is already evident at concentrations of Vitamin C of 1 mM in the culture medium, and it is proportional to the amount of Vitamin C.

Since clinical investigations using high doses of Vitamin C to treat cancer, have reported plasma levels of more than 30 [116], and up to 49 mM [117], it seems reasonable to conclude that protocols using high amounts of Vitamin C, administered by intravenous injection, are not strictly necessary to kill cancer cells in APL.
It is of interest to consider that in our procedure, the leukemic cells were exposed to increasing concentrations of Vitamin C for no more than two hours, then accurately “washed”, re-suspended in fresh culture medium, which did not contain Vitamin C, and further incubated.

Further investigations, performed by our research group, showed that a plasma concentration of 3 mM of Vitamin C in the culture medium, is sufficient to kill more than a half of the cells in culture (LC$_{50}$), in a number of different human myeloid leukaemia cell lines, including HL60, K562, U937, NB4, Nb4-R1, and NB4/As

![Figure 1:](image1)

Figure 1: The microphotographs refer to the cyto-morphologic modifications of HL60 cell lines (Human Acute Promyelocytic Leukemia - APL) exposed for two hours to increasing concentrations of Vitamin C. It is evident that increasing the concentration of Vitamin C in the medium (from 1 to 5 mM), APL cells show an increasing degree of morphologic alterations indicating progressive cell death (apoptosis, autophagy, autorschizis). With the Hoechst/PI fluorescent staining, vital cells are colored in blue, while dead/apoptotic cells are stained in red. M.G.G. = May Grunwald Giemsa cell staining; Hoechst33342/Propidium Iodide (PI) = Vital Staining; C = Control (untreated) sample; 1 mM, 3 mM, 5 mM = Vitamin C at 1, 3, and 5 millimoles in the culture medium; Original magnification: 400X.

![Figure 2:](image2)

Figure 2: Viability profile of (Human) Acute Promyelocytic Leukemia (APL) cell line (HL60) exposed for two hours to increasing concentrations of Vitamin C. Cytofluorimetric analysis. It is evident that increasing the concentration of Vitamin C in the medium (from 1 to 5 mM), the number of dead cells increases proportionally; C = Control (untreated) sample; 1 mM, 3 mM, 5 mM = Vitamin C at 1, 3, and 5 millimoles in the culture medium.

It is of interest to consider that in our procedure, the leukemic cells were exposed to increasing concentrations of Vitamin C for no more than two hours, then accurately “washed”, re-suspended in fresh culture medium, which did not contain Vitamin C, and further incubated...
Cytotoxic effect of Vitamin C on human myeloid leukemia cell lines

**Figure 3:** The diagram shows the almost uniform decrease in the number of vital leukemic cells in the culture medium, after exposing them to increasing concentrations of Vitamin C, for two hours. The figure illustrates Table 1.

Vitamin C LC$_{50}$

**Figure 4:** Highlighted with colored circles, the LC$_{50}$ for each human myeloid leukemia cell line tested. The figure illustrates Table 1.

for additional 18-24 hours, before the evaluation of cell survival and apoptosis. Given the results obtained, it is reasonable to conclude that the Vitamin C added to the culture medium (in the form of sodium ascorbate), is
Table 1: The table shows the number of vital cells after two hours of exposure to increasing concentrations of Vitamin C in the culture medium. The cell lines used in this experiment are variants of human myeloid leukemia cells, and include: HL60, NB4, K562, U937, NB4-R1, NB4/As. It is evident that the total number of cells in culture decreases by increasing the concentration of Vitamin C in the culture medium.

| Exp. 1   |   |   |   |   |   |   |
|----------|---|---|---|---|---|---|
|          | HL60 (2 h) | NB4 (2 h) | K562 (2 h) | U937 (2 h) | NB4-R1 (2 h) | NB4/As (2 h) |
| Contr.   | 471 | 912 | 663 | 1189 | 337 | 819 |
| VC 0.5   | 296 | 680 | 669 | 476  | 82.3 | 42.7 |
| VC 1     | 108 | 496 | 628 | 245  | 39.7 | 31.9 |
| VC 3     | 22.6| 163 | 226 | 56.4 | 47   | 32.2 |
| VC 5     | 15.1| 143 | 82  | 30.2 | 35   | 48.4 |
| VC 7     | 6.15| 85.4| 32.2| 10.6 | 24.6 | 17.6 |
| Contr.   | 869 | 1,020| 694 | 829  | 936  | 958  |
| VC 0.5   | 423 | 956 | 399 | 704  | 624  | 823  |
| VC 1     | 349 | 887 | 445 | 585  | 560  | 674  |
| VC 3     | 217 | 337 | 200 | 232  | 335  | 581  |
| VC 5     | 143 | 74.6| 111 | 118  | 344  | 402  |
| VC 7     | 89.5| 147 | 35.2| 93.7 | 255  | 329  |

| Exp. 2   |   |   |   |   |   |   |
|----------|---|---|---|---|---|---|
|          |   |   |   |   |   |   |
| Contr.   | 843 | 1,130| 1,020| 969  | 967  | 859  |
| VC 0.5   | 545 | 924 | 660 | 716  | 678  | 722  |
| VC 1     | 438 | 835 | 507 | 689  | 668  | 649  |
| VC 3     | 343 | 395 | 113 | 480  | 551  | 499  |
| VC 5     | 218 | 346 | 35.2| 411  | 443  | 453  |
| VC 7     | 181 | 241 | 17.6| 320  | 380  | 414  |

| Exp. 3   |   |   |   |   |   |   |
|----------|---|---|---|---|---|---|
|          |   |   |   |   |   |   |

C = control (untreated) sample; VC = Vitamin C; VC 0.5 mM, VC 1 mM, VC 3 mM, VC 5 mM = Vitamin C at 0.5, 1, 3, and 5 millimoles in the culture medium.

rapidly internalized by the leukemic cells, and its “toxic” effects continue for hours (days), even when the nutrient has been removed from the culture medium. This is in agreement with the notion that both normal and leukemic white blood cells tend to accumulate/concentrate Vitamin C [119-122] to levels that are 10-100 fold higher than plasma [123,124], and it is also in disagreement with the view that hydrogen peroxide forms outside the tumour cells [58-60].

Neutrophils, in particular, accumulate vitamin C via the Sodium-Dependent Vitamin C Co-Transporter 2 (SVCT2) [125], and have intracellular levels of 1-2 mM [126]. Therefore, while there is agreement on the fact that, in solid tumours, Vitamin C, initially oxidized to Dehydroascorbic Acid (DHA), is internalized by the cell, via GLUT 1 and 4, and finally reduced again to ascorbic acid, with consumption of glutathione, this does not seem to apply to the case of acute myeloid leukaemia.

More importantly, the parallel exposure of normal hematopoietic precursors (CD34+), isolated from cord blood, to Vitamin C, at the concentrations that are cytotoxic for leukemic cells did not affect their survival, or impair their capacity to proliferate and differentiate in response to myeloid growth factors. These data confirm that Vitamin C is harmless for normal hematopoietic precursors.

Hypoxia Inducible Factor (HIF): The Forgotten Pathway

Hypoxia and induction of Hypoxia-Inducible Factors (HIF) is a hallmark of many tumours [127,128]. HIF-1 is a heterodimeric transcription factor discovered in 1991 [129], composed of two subunits, α and β. The HIF-1α subunit is oxygen sensitive and it is induced by hypoxic conditions, which are very common in cancer. Direct transcriptional targets of HIF-1 include genes regulating, among others, growth and apoptosis, cell migration, energy metabolism, angiogenesis, vasomotor regulation, matrix and barrier functions, and transport of metal ions and glucose [130].

In normoxic conditions, the HIF-1α unit is down-regulated by Vitamin C dependent hydroxylases, while in hypoxic conditions (such as those existing in many different types of cancer) HIF-1α hydroxylation is repressed, with consequent increase in HIF-dependent gene transcription, neo-angiogenesis and tumour growth and progression [131].

More importantly, since Vitamin C stimulates HIF-1α prolyl hydroxylases, low levels of Vitamin C promote tumour growth and progression, by reducing HIF-1α hydroxylation [132], thereby stabilizing HIF1-α, while high levels of HIF renders cancer cells more sensitive to Vitamin C-induced toxicity [75]. To confirm this view, Kuiper and coll [133] have recently found an inverse relationship between intra-tumour levels of Vitamin C and HIF activity in both endometrial cancer [134], and Colorectal Carcinoma (CRC) [135].

In 1925, Otto Warburg observed that cancer cells manifest increased rates of lactate production under aer-
obic conditions ("Warburg Effect") or, in other words, they preferentially utilize glycolysis, instead of oxidative phosphorylation, for metabolism even in the presence of oxygen [136,137].

"Hypoxia" (low oxygen concentration) is a hallmark of solid tumours, usually occurring at the centre of the tumour mass, where blood vessels are abnormal or insufficient to supply adequate amounts of oxygen [138]. In response to the reduced oxygen tension, the HIF is activated to mediate the primary transcriptional adaptation to hypoxic stress in cancer cells [139,140].

As previously mentioned, HIFs regulate angiogenesis, cell survival, proliferation, apoptosis, adhesion, and metabolism by transcriptionally activating downstream targets such as vascular endothelial growth factor and erythropoietin. Therefore, HIF (HIF1, in particular) plays a major role in tumour growth, and clinical data suggest that the upregulation of HIF, as determined by the low oxygen tension, is usually associated with increased mortality in a number of different cancers [141-143], and may represent a relevant target for new therapeutic approaches to the disease [144-146].

The HIF Pathway in Leukaemia

The role of HIF-1α in leukaemia, and in particular in Acute Myeloid Leukaemia (AML), has only recently emerged and it is still somewhat controversial. One possible explanation for this delayed interest in the role of hypoxia in leukemia could be the fact that leukemia is not considered a "solid" tumour, and therefore, the role of oxygen, in its pathogenesis, has been considered inconsequential for long time.

This erroneous view, has been recently reviewed, as data have emerged, demonstrating that leukemic cells are sensitive to the oxygen tension, and hypoxia influences leukemic cell proliferation, differentiation, and resistance to chemotherapy [147].

Migliavacca & coll ha recently demonstrated an oncogenic function of HIF-1α, in the M5 Fab subtype of AML [148]. In particular, the authors have demonstrated that, in M5 AML, HIF-1α mediates the ability of leukemic cells to migrate and invade extramedullary sites.

The same group, has demonstrated that PML-RARα and other fusion proteins involved in the pathogenesis of Acute Promyelocytic Leukemia (APL) behave as transcriptional coactivators of HIFs, and both HIFs and PML-RARα, enhances the progression of APL, by promoting cell migration, homing to bone marrow, and bone marrow neo-angiogenesis [149,150].

Further investigations [151] have demonstrated that HIF-1α plays critical and pleiotropic roles in in the pathogenesis of Chronic Lymphocytic Leukemia (CLL). Globally, elevated levels of HIF-1a have been reported in AML [152-155], APL [150], Acute Lymphoblastic Leukemia (ALL) [153,156], and Chronic Myelogenous Leukemia (CML) [157,158]. Furthermore, HIF-1α overexpression conditions disease severity and outcome in both AML and Myelodysplastic Syndrome (MDS) [153,159-161].

Overall, the available data show that hypoxia and HIF-mediated signalling, play a crucial role in leukaemia, and targeting HIF with specific drugs or natural inhibitors, such as Vitamin C, represents a potentially useful approach to its treatment [154,162].

What to Do next?

The anticancer properties of Vitamin C are known, since at least six decades, even though its use in clinical practice, has only recently re-emerged, after the demonstration that, in relatively high concentrations, it can selectively kill a number of different human tumour cells, both in vitro and in vivo [58-61,73,102,114,116].

The proof of the anticancer efficacy of Vitamin C in high doses, administered by mouth, has been reported four decades ago, by the legendary scientist, Linus Pauling [48,51], and further confirmed, very recently, by experimental, in vitro and in vivo data [73,79,102,114].

Vitamin C is a natural compound, an antioxidant and a life-saving nutrient with multiple beneficial effects on the human body, that man, some primates and a few other mammals do no longer produce. Beyond being a natural and essential nutrient, Vitamin C shows, in vitro, an outstanding efficacy in killing a number of different cancer cells, with an efficiency that no other anticancer drug, presently available on the market, has ever shown.

Vitamin C is extremely selective in its anticancer effect, being able to kill only cancer cells, by sparing, at the same time, all the other cells of the organism, thus being totally harmless for normal cells, and, as such, very well tolerated and without significant side effects. In fact, the only (relative) contraindication to its use, is the lack of the enzyme Glucose-6-Phosphate-Dehydrogenase (G6PDH), a rare genetic condition also known as "fuvism". More importantly, within an expensive and often artificially inflated market, such as that of the anticancer drugs [163,164], Vitamin C, with its low cost, represents an outstanding opportunity, for both the patients and the healthcare system.

Unfortunately, in spite of all the above characteristics (or just simply "because" of them!), Vitamin C, has never been easily or favourably accepted as an anticancer drug, by the western, business-oriented Medicine. This also explains why, although the data on its anticancer efficacy are outstanding and straightforward, many scien-
As we have seen, the idea that the oral administration of Vitamin C, in high doses, is not effective against cancer is a conceptual artefact born from questionable interpretations of pharmacokinetics data, after oral and/or intravenous administration.

On the other hand, the idea that Vitamin C, administered in high doses by intravenous infusion, behaves as a pro-drug of H₂O₂, thus inducing pro-oxidant damage to cancer cells, beyond being experimentally questionable, has not led to clinically significant results or outcomes [165-169], even though, encouraging results (to say the least!) emerge from unbiased interpretation of the available data [170]. In particular, as it has been shown up to 110 g/m²/day are very well tolerated by the majority of patients [117], and even in the absence of any significant clinical remission, intravenous Vitamin C is almost invariably associate with a clear-cut improvement in patient’s quality of life [96].

As a result, history repeat itself! … and just as Vitamin C was dismissed as ineffective, against cancer, more than thirty years ago, on the ground of flawed clinical trials [98,99], nowadays it runs again the risk of being definitively buried, in spite of the large amount of scientific evidence, demonstrating its extraordinary efficacy in fighting cancer!

It is clear that much remains to be understood about the cytotoxic effects of Vitamin C against cancer, and much more can (and must!) be done, to both improve the intravenous therapy, and further investigate the oral administration route of the high doses of the nutrient.

Improving the intravenous treatment, can (should!) be achieved, by considering:

a) The type of pharmaceutical preparation, the sodium salt of the ascorbic acid to be preferred, when administered by the intravenous route [171];

b) The time and schedule of administration (slow infusion to be preferred) [172,173];

c) The level of tissue oxygenation (cell cultures are better oxygenated than tumour tissues, and this may explain the differences in the outcomes between in vitro and in vivo treatment of cancer) [174]. In clinical settings, an improved tumour tissue oxygenation could be obtained with either ozone or hyperbaric oxygen;

d) The level of blood glucose (glucose may interfere with the uptake of ascorbate by cancer cells) [175,176], and the possibility of associating an adequate dietetic regimen to the treatment with high doses of oral or intravenous Vitamin C.

On the other hand, regarding the oral administration of high doses of Vitamin C to treat cancer, it will be important to consider:

a) The successful results of the oral administration of Vitamin C in different experimental settings [102,114];

b) The results of the clinical studies performed by Cameron & Campbell [95], Cameron & Pauling [48,51], and Murata [97];

c) The rationale behind the use of Vitamin C as an antioxidant (rather than a pro-oxidant) and inhibitor of the HIF [114];

d) The possibility of using “alternative” formulations (such as liposomes), with increased absorption and improved bioavailability [109,173].

Concluding Remarks

The rationale behind the use of high doses of Vitamin C in the treatment of acute leukaemia is strong and very well grounded. In summary:

I. Leukemic patients, almost invariably show a severe deficiency of this nutrient;

II. While it is currently supposed to kill cancer cells by inducing pro-oxidant damage, Vitamin C, is also very effective as an antioxidant, by inhibiting the Hypoxia Inducible Factor (HIF);

III. The mechanistic explanation of the pathogenesis of myeloid leukaemia, includes the possibility that a Vitamin deficiency may induce the neoplastic transformation of myeloid precursors, through an upregulation of the HIF, and the consequent cascade of HIF-dependent cancer genes;

IV. Although administered by intravenous infusion, in the majority of clinical trials performed so far, Vitamin C appears to be effective, in fighting cancer, even when administered by mouth;

V. Vitamin C is very well tolerated, and has no side or undesired effects;

VI. Experimental in vitro data, unequivocally show the cytotoxic effect of Vitamin C against leukaemia as shown in our study on leukemic and normal cell lines, Vitamin C can kill almost every type of Acute and Chronic Myeloid Leukaemia derived cell, without doing any harm to their normal counterpart (CD34+) cells;

VII. Vitamin C is a natural compound, and it is very cheap;

VIII. Do we need more information or evidence, to start clinical trials on Vitamin C, in the treatment of acute and chronic myeloid leukaemia?
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Conflict of Interests

None to declare.

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