Combination Therapy With Casirivimab/Imdevimab and Remdesivir for Protracted SARS-CoV-2 Infection in B-cell-Depleted Patients

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Profoundly B-cell-depleted patients can have prolonged severe acute respiratory syndrome coronavirus 2 infections with evidence of active viral replication, due to inability to mount an adequate humoral response to clear the virus. We present 3 B-cell-depleted patients with prolonged coronavirus disease 2019 infection who were successfully treated with a combination of casirivimab/imdevimab and remdesivir.

Keywords. B-cell-depleted; monoclonal antibodies; protracted SARS-CoV-2.

Immunocompromised hosts remain at risk for severe coronavirus disease 2019 (COVID-19) infection, particularly those with B-cell depletion, as impaired antibody response could lead to ineffective viral clearance [1]. These patients can have protracted infections with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), with low cycle threshold (Ct) values with nucleic acid amplification testing (NAAT) and recovery of infectious virus in cell culture [2–5]. These patients are at risk not only for recurrent hospitalizations, but for emergence of escape mutants that can spread to communities.

The antiviral remdesivir has been shown to shorten time to recovery [6]. The combination of monoclonal antibodies (mAb) casirivimab/imdevimab reduces risk of hospitalization and death and reduces viral load (VL) in upper respiratory tract specimens without emergence of escape mutants [7, 8]. We hypothesized that combination of mAb and remdesivir would be safe and effective in achieving viral clearance and symptom resolution without risking emergence of escape mutants. We present 3 B-cell-depleted patients with prolonged COVID-19 who were treated successfully with combination casirivimab/imdevimab under an emergency Investigational New Drug (E-IND) application with off-label remdesivir. Treatment was a single dose of 1200 mg intravenously of each of casirivimab and imdevimab and a 5-day course of remdesivir (200-mg loading dose intravenously, then 100 mg daily thereafter).

PATIENT CONSENT

Written consent was obtained from each patient.

The design of the work was approved by the Johns Hopkins Institutional Review Board (IRB) and conforms to standards currently applied in the United States. In addition, E-IND approval was obtained from Regeneron Pharmaceuticals and the US Food and Drug Administration (FDA).

Patient #1

A 45-year-old man with multiply treated relapsed/refractory follicular lymphoma, history of nonmyeloablative haploidentical bone marrow transplant (BMT) complicated by graft-vs-host disease (GVHD) on prednisone and ibrutinib, and hypogammaglobulinemia was admitted 1 year after BMT with cough, headache, and loss of taste and smell, but not hypoxemia. He was diagnosed with COVID-19 via NAAT and received convalescent plasma. Two months later, he had persistent cough without fevers or hypoxemia, and a chest computed tomography (CT) scan showed bilateral ground-glass opacities. SARS-CoV-2 via NAAT on nasopharyngeal (NP) swab was persistently positive with low Ct (Table 1). Extensive evaluation was negative for other etiologies.

He received casirivimab/imdevimab on day 70 after diagnosis and off-label remdesivir on days 75–79; SARS-CoV-2 NAAT became negative on day 84 (Table 1), with resolution of cough by 2 weeks and normalization of chest CT by 4 weeks. His preceding leukopenia and neutropenia (absolute neutrophil count 0.79 K/cu mm) normalized by 2 weeks post-treatment. Ten months later, he is clinically well except for baseline GVHD.

Patient #2

A 52-year-old woman with follicular lymphoma (in remission after R-CHOP) and common variable immunodeficiency (CVID) on weekly subcutaneous immunoglobulin G replacement was hospitalized with fever, cough, and headache. She...
tested positive for SARS-CoV-2 NAAT on NP swab with chest CT showing bilateral ground-glass opacities and received a 10-day course of dexamethasone. Attempts to wean dexamethasone were unsuccessful and led to extensive evaluation revealing only persistent SARS-CoV-2 NAAT with low Ct values (Table 1). She received casirivimab/imdevimab on day 111 and remdesivir on days 115–119. Her Ct values on SARS-CoV-2 NAAT increased until NAAT became negative on day 146. She is clinically well 10 months after treatment.

Patient #3
A 62-year-old woman with refractory diffuse large B-cell lymphoma received chimeric antigen receptor (CAR) T-cell therapy (after fludarabine and cyclophosphamide), complicated by cytokine release syndrome. Three weeks after CAR T cells, she was diagnosed with mild COVID-19 via NAAT, with cough and fatigue, and received bamlanivimab under Emergency Use Authorization. She continued with intermittent fevers and developed hypoxemia with persistently positive SARS-CoV-2 NAAT with low Ct. She received casirivimab/imdevimab on day 47 and remdesivir on days 46–50; repeat SARS-CoV-2 NAAT Ct slowly increased until NAAT was negative on day 64. She is clinically well 8 months after treatment.

SAFETY
After baseline physical exam, weekly symptom assessments, CBC with differential (CBC), complete metabolic panel (CMP), and NP swab for SARS-CoV-2 NAAT were obtained on all

| Patient Number* | Day From Diagnosis | Platform | Ct Values | Developed AA Substitutions |
|-----------------|--------------------|----------|-----------|---------------------------|
| Patient #1      | 0                  | NeuMoDx  | N 13.69, Nsp2 14.37 |                           |
|                 | 1                  | Convalesenct plasma infusion |                           |                           |
|                 | 19                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 14.7 |                           |
|                 | 57                 | GenMark ePlex | N/A*      | NSP8:I156L NS6:E13K       |
|                 | 61                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 19.1 |                           |
|                 | 70                 | REGN-COV infusion |                           |                           |
|                 | 77                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 30.1 |                           |
|                 | 84                 | Roche 6800 | Negative |                           |
|                 | 91                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | Negative |                           |
|                 | 216                | NeuMoDx  | Negative |                           |
| Patient #2      | 0                  | NeuMoDx  | N 18.82, Nsp2 19.71 |                           |
|                 | 20                 | GenMark ePlex | N/A*      |                           |
|                 | 26                 | NeuMoDx  | N 24.10, Nsp2 25.74 |                           |
|                 | 33                 | GenMark ePlex | N/A*      |                           |
|                 | 38                 | GenMark ePlex | N/A*      |                           |
|                 | 103                | GenMark ePlex | N 19.27, Nsp2 19.94 | NSP2:K34S |
|                 | 111                | REGN-COV infusion | –         | –                        |
|                 | 117                | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 26.2 | NSP13:D36G S:N334K S:S939F |
|                 | 126                | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 31.6 |                           |
|                 | 132                | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 37.7 |                           |
|                 | 139                | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 37.2 |                           |
|                 | 147                | NeuMoDx  | Negative |                           |
| Patient #3      | 0                  | GenMark ePlex | N/A*      |                           |
|                 | 1                  | Bamlanivimab infusion | –         | –                        |
|                 | 23                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 23.3 | S:E484K                  |
|                 | 38                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 21.9 |                           |
|                 | 39                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 22.2 |                           |
|                 | 45                 | NeuMoDx  | N 21.11, Nsp2 22.43 |                           |
|                 | 47                 | REGN-COV infusion | –         | –                        |
|                 | 50                 | NeuMoDx  | N 28.69, Nsp2 30.01 | NSP6:L37F             |
|                 | 57                 | Cepheid (Xpert Xpress SARS-CoV-2/Flu/RSV) | N2/E 29.1 |                           |
|                 | 64                 | Roche 6800 | Negative |                           |

Abbreviations: AA, amino acid; Ct, cycle threshold.

*Clade/lineages: patient 1, 20A/B.1.409; patient 2, 20B/B.1.1.434; patient 3, 20G/B.1.2.

N/A represents positive results; Ct values are not reported by this assay.
BRIEF REPORT

IMMUNOLOGIC PARAMETERS

B Cells
Patient #1 did not have lymphocyte subsets performed before casirivimab/imdevimab, but had an absolute lymphocyte count (ALC) of 0.16 K/cu mm. One month later, he had a CD19+ lymphocyte count and percentage of 0, which was considered to have been baseline. Patient #2 had lymphocyte subsets 3 months before casirivimab/imdevimab, with CD19+ count and percentage of 0, and ALC of 0.12 K/cu mm. Patient #3 did not have lymphocyte subsets, but had an ALC of 0.19 K/cu mm before casirivimab/imdevimab. Normal ranges are ALC 1.1–4.8 K/cu mm, CD19+ lymphocyte total number 12–645/μL, and CD19+ lymphocyte percentage 6%–19%.

Immunoglobulins
Patient #1 had a total immunoglobulin G (IgG) level of 179 mg/dL (normal, 610–1616 mg/dL), immunoglobulin A (IgA) 16 mg/dL (normal, 61–348 mg/dL), and immunoglobulin M (IgM) <10 mg/dL (normal, 35–242 mg/dL) before casirivimab/imdevimab. Patient #2 was receiving subcutaneous immunoglobulin replacement (6 g weekly), with an IgG level of 417 mg/dL, IgA 27 mg/dL, and IgM <10 mg/dL. Patient #3 had an IgG level of 525 mg/dL, IgA 50 mg/dL, and IgM <10 mg/dL. Thus, all 3 had low baseline Ig levels, including all with IgM below the limit of quantitation.

Anti-SARS-CoV-2 Antibodies
Before casirivimab/imdevimab, Patient #1 had an anti-S1-subunit IgG of 0.03 arbitrary units (arb’U) on the Euroimmun assay (positive >1.24 arb’U) (Figure 1). One week after casirivimab/imdevimab, this rose to 171.60 arb’U. Patient #2 had an anti-SARS-CoV-2 S1 spike subunit IgG of 0.09 arb’U before casirivimab/imdevimab. Patient #3 had a positive anti-S1-subunit IgG antibody of 8.50 arb’U 5 weeks after bamlanivimab but before casirivimab/imdevimab.

DISCUSSION

These 3 cases highlight protracted COVID-19 courses in B-cell-depleted immunocompromised patients. These patients have absent adaptive humoral responses, which are crucial to controlling viral infection, as shown by the inverse correlation between NAAT positivity and antibody seroconversion [9, 10]. This is further supported by reductions in VL seen after treatment with casirivimab/imdevimab [8]. It is now known that persistent NAAT positivity can represent replication-competent virus in viral cultures, and prolonged shedding could result in emergence of variants [2, 3].

Antibody-mediated neutralization is virus-specific, as demonstrated by lack of cross-reactivity between neutralizing

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**Figure 1.** Subunit IgG values over time for all cases (Euroimmun SARS-CoV-2 S1 subunit ELISA kits assay, semiquantitative values). Abbreviations: ELISA, enzyme-linked immunosorbent assay; IgG, immunoglobulin G; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.
antibodies against the SARS-CoV-2 receptor binding domain (RBD) and those for SARS-CoV-1 and MERS-CoV [9]. Patients lacking circulating B-cells and/or with profound hypogammaglobulinemia are unable to mount neutralizing antibody responses, can have persistent symptoms [5] but still retain effective virus-specific T-cell responses [5].

When choosing therapy for these patients, a significant concern is preventing emergence of resistance. Remdesivir shortens duration of symptoms, likely by inhibiting viral proliferation [6]. Monoclonal antibody therapy with casirivimab/imdevimab could enhance clearance in patients without endogenous immune response or with high VL, without emergence of escape mutants [7]. We hypothesized that combination mAb would prevent escape mutants by binding to 2 distinct and nonoverlapping regions of the receptor binding domain [11], while addition of adjunctive remdesivir would help reduce viral burden [7].

Sequencing was done on the majority of samples from each patient. In this limited set, 12 amino acid substitutions were noted (Table 1); the most common mutations were in the spike protein. Patients #1 and #3 had received CP and bamlanivimab, respectively, before development of mutations, but the role of these therapies in selecting for mutations is unclear. Patient #3 received bamlanivimab before the development of S:E484K in lineage B.1.2, a lineage with a relative paucity of spike protein substitutions. This substitution has been associated with mAb escape when using monotherapy and with infections in vaccinated individuals [12]. Although some novel mutations were noted during or immediately after treatment in Patient #2 (Table 1), this did not translate into treatment and/or clinical failure. Of note, Patients #1 and #2 originally presented before the availability of monoclonal antibodies for early treatment.

Poor outcomes from COVID-19 have been reported in patients treated with anti-CD20 antibody therapy [1]. Predicting which patients might have severe complications is important; for example, SARS-CoV-2 RNAemia is being investigated as a marker of severity [1]. Of note, leukopenia/neutropenia in our first patient was attributed to viral suppression from persistent SARS-CoV-2, given its resolution shortly after casirivimab/imdevimab; SARS-CoV-2 has been found in bone marrow biopsies by real-time reverse transcription polymerase chain reaction, suggesting a potential role [5].

Emerging data show low antibody responses to COVID-19 mRNA vaccines in patients with CLL (39.5%) and those receiving anti-CD20 therapies within 12 months before vaccination [13]. This population will remain highly vulnerable, and more research is needed to define the best preventive and therapeutic approaches [14–16]. This case series, as well as other emerging reports of the utility of monoclonal antibody therapy in prolonged COVID-19 infection [17], highlights a potential role for passive immunotherapy, with or without an antiviral agent.

In conclusion, a combination of monoclonal antibodies and remdesivir was effective and well tolerated in 3 B-cell-depleted patients with protracted COVID-19 infection. It is possible that this regimen may have prevented emergence of escape mutants, although further study is needed to confirm this. Additional studies are needed to evaluate this treatment option in a larger population.

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