Supplementary Materials and Methods

**Figure S1.** The growth curve of SM10λπ, PAO1 and EC600 under indoles. Cells (1 × 10^7 CFU/ml) were cultured in the presence of indicated concentrations of indoles and isometric solvent ethyl alcohol at 37°C for 6 h. Values are means ± SEMs from at least three independent experiments.

**Figure S2.** Validation of constructed strains. Indicated strains in the logarithmic phase were collected for extracellular indoles, cell optical density measurement and real-time PCR analysis of tnaA expression. The indoles expression level was indicated in terms of extracellular indoles production (µM) divided by cell optical density (OD600). Values are means ± SEMs from at least three independent experiments. ***, P < 0.001.

**Table S1.** Minimal inhibitory concentration (MIC) of SM10λπ against ciprofloxacin.

| No. | 1    | 2    | 3    | 4    | 5    | 6    | 7    | 8    | 9    | 10   | 11   | 12   |
|-----|------|------|------|------|------|------|------|------|------|------|------|------|
|     |      |      |      |      |      |      |      |      |      |      |      |      |
| First| -    | -    | -    | -    | -    | -    | -    | +    | +    | +    | +    | +    |
| Second| -    | -    | -    | -    | -    | -    | -    | +    | +    | +    | +    | +    |
| Third | -    | -    | -    | -    | -    | -    | -    | +    | +    | +    | +    | +    |
| CIP (µg/ml) | 2^0 | 2^-1 | 2^-2 | 2^-3 | 2^-4 | 2^-5 | 2^-6 | 2^-7 | 2^-8 | 2^-9 | 2^-10 | 2^-11 |

**Table S2.** The information of strains and plasmids used in this work.
| Strains/plasmids | Genotype or characteristics | Source |
|------------------|-----------------------------|--------|
| **Escherichia coli** |                             |        |
| SM10λπ           | thi thr leu tonA lacY supE recA::RP4-2-Tc::Mu Km λpir | (1)    |
| SM10λπ ΔtnaA     | Mutants of *E. coli* SM10λπ deficient in *tnaA* gene | This work |
| ΔtnaA-vector     | SM10λπ ΔtnaA with pSTV28 vector introduced | This work |
| ΔtnaA-TnaA       | SM10λπ ΔtnaA with pSTV28-TnaA introduced | This work |
| SM10λπ-TnaA      | *E. coli* SM10λπ with pSTV28-TnaA introduced | This work |
| EC600            | LacZ-, Gm^S^, Rif^R^    | Our lab |

| **Pseudomonas aeruginosa** |                             |        |
| PAO1             | Wild-type strain, Gm^S^, Amp^R^ | (2)    |

| **Plasmids** |                             |        |
| pSTV28         | Control plasmid, containing P<sub>lac</sub> promoter, Cm^R^ | Our lab |
| pSTV28-tnaA    | pSTV28 derivative, *Escherichia coli* SM10λπ <i>tnaA</i> overexpression plasmid, controlled by the constitutive P<sub>lac</sub> promoter, Cm^R^ | This work |
| pUCP24T        | 370 bp oriT fragment from pCVD442 cloned into pUCP24, ori1600, Gm^R^ | Our lab |
| pKD3           | oriR6K, FRT::cat::FRT template plasmid Cm^R^, Amp^R^ | (3)    |
| pKD46          | oriR101 repA101ts P-araB-gam-bet-exo Amp^R^ | (3)    |
| pCP20          | pSC101 temperature-sensitive replicons, Flp (λ Rp), cI857, Cm^R^, Amp^R^ | (3)    |

Gm^R^, Rif^R^, Amp^R^ and Cm^R^ stand for gentamycin, rifampin, ampicillin and chloramphenicol resistance, respectively.

**Table S3.** Sequences of RNA and DNA oligonucleotides.

| Name | Sense primer (5′-3′) | Antisense primer (5′-3′) | Source |
|------|----------------------|--------------------------|--------|
|      |                      |                          |        |
| Promers for qPCR |                             |                          |        |
| <i>tnaA</i> | TCACCCGCGAAACCTACAAA | GTCTTTTCATGCACAGCAGGC | Our lab |
| <i>korA</i> | GCTTACCGAAAGCCAGTTCCAG | GCAAGTTCCTTGCTTCGGAACGC | (4)    |
| <i>korB</i> | AAGGAAAAGGCCGCGAAGGAG | TCGATGAGCGCGACCAGTTTC | (4)    |
| <i>trbA</i> | TGGAACCTCCCCTACCTCTTT | CCACACTGATGCGTTCGTAT | (5)    |
| <i>trbB</i> | CGCGGTGCGCATCTTCACG | TGCCCGAGCCAGTACCAGCAATG | (6)    |
| <i>rtaA</i> | GAAGCCCATCGCGTGCCCTGTAG | GCCGACGATGACGAAGGACGG | (5)    |
| <i>rtaI</i> | ATCACGAAAGGAACCATCCTT | TTGAACCTGTGGTGCGGTTGAC | (7)    |
| <i>rtaJ</i> | CGAACGAAAGAGCGATGAGG | TCGTGGTGAGCCAGAAGTTT | This work |
| <i>rpoD</i> | TATCTGCTGGAACAGTACGATGTG | TGTTGTACATCGCGGCTCG | (8)    |

Primers for <i>tnaA</i> promoter cloning

| Name | Sense primer (5′-3′) | Source |
|------|----------------------|--------|
| <i>tnaA</i> | TATGACCATGATTACGAATTTCGGT, ATAGCAGATG | This work |

**Plasmids construction**
For construction of TnaA-expressing plasmid, TnaA coding region was amplified by PCR from *E. coli* SM10λπ chromosomal DNA. Amplified fragments were purified by gel extraction kit (ShengGong, Shanghai, China), then digested with *Hind III* and *BamH I* (Takara, Dalian, Liaoning, China), and cloned into the corresponding sites of pET32a (+).

**Construction of *E. coli* SM10λπ tnaA deficient mutants**

Construction of *E. coli* SM10λπ tnaA mutant followed the same steps described by Datsenko *et al* (3) except 100 mM L-arabinose was used to induce *P*araB promoter of pKD46. Briefly, the insert fragments were amplified with primer D-*tnaA*-F (TGTAATTTACAGGGATCACTGTAATTAAAATAAATGAAGGATTATGTATG TGGCTGGAGCTGCTTCG) and D-*tnaA*-R (TGTAGGGTAAAGAGTGGCTAACATCCTTATAGCCACTCTGTAGTATAAAT GGAATTAGCCATGCTTC). Afterwards, 100 μL cells were transfected with 1000ng PCR products using electroporation according to the manufacturer's instructions. Recombinants were selected on LB agar (Cm, 15μg/mL) and then transformed with pCP20 to eliminate FRT-flanked *cat* gene. Both mutants were verified by PCR (primers C-*tnaA*-F: TTTGCCCTTCTGTAGCCATC and C-*tnaA*-R: ACCATAACACCCCCAAATGC) and DNA sequencing.

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