p53 as a prognostic factor in stage I breast cancer

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Summary Accumulation of the tumour-suppressor protein p53 in breast cancer is associated with several prognostic factors that indicate an aggressive, rapidly proliferating tumour with an unstable genome. To assess p53 accumulation in stage I breast cancer and to evaluate the prognostic value of both nuclear and cytoplasmic p53, 205 patients with node-negative breast cancer and tumour size less than or equal to 20 mm were examined. Immunohistochemistry was performed on frozen sections with the monoclonal antibodies PAb 1801 and DO1. Cellular p53 accumulation, within either the nucleus or the cytoplasm or in both, showed the same association with different pathobiological variables as nuclear accumulation alone. Eleven per cent of the tumours showed strong nuclear accumulation and were significantly correlated to age under 50 years, negative oestrogen receptor status, DNA aneuploidy, high S-phase fraction, high pathological grade and poor prognosis. The distant recurrence rate ratio was 6.2 (P = 0.002). It is thus concluded that p53 accumulation is of prognostic value in early stage breast cancer.

Keywords: p53; breast cancer; immunohistochemistry

Breast cancer treatment is primarily based on the traditional prognostic factors tumour size and presence of lymph node metastasis. A tumour 20 mm or less in size without lymph node involvement is associated with a relatively good prognosis, but within 5 years 13% of these will recur and after 20 years this figure rises to 25% (Rosen et al., 1993). In order to unveil this group of tumours that recur much work has been done to find reliable prognostic factors that can predict which patients would benefit from adjuvant therapy (McGuire et al., 1990). A potential prognostic factor in breast cancer is the accumulation of the tumour-suppressor protein p53 (Isola et al., 1992; Thor et al., 1992; Allred et al., 1993; Silvestrini et al., 1993; Stenmark-Askmalm et al., 1994). In case of DNA damage, active p53 regulates the cell cycle, giving time for repair or, if this fails, induces apoptosis. If p53 is inactive the growth-controlling function is lost and damage to the genome is thought to accumulate (Kastan et al., 1991; Yonish-Rouach et al., 1991; Lane, 1992). Indeed, it has been shown that human cancers often lose normal p53 activity (Vogelstein, 1990). The normal (wild-type) form of p53 is usually rapidly metabolised and is therefore not detectable by immunohistochemical methods. On the other hand, inactive or mutated p53 has a longer half-life and accumulates within the cell, thus becoming detectable by immunohistochemistry (Lane and Benchimol, 1990; Yonish-Rouach et al., 1991). p53 accumulation has been shown to occur in breast cancer, and it has been associated with several other prognostic factors indicating an aggressive, rapidly proliferating tumour with an unstable genome. The prevalence of p53 overexpression in small, node-negative breast cancer has been reported to be low (Davidoff et al., 1991a; Isola et al., 1992).

The purpose of the present study was to investigate p53 accumulation in 205 stage I breast cancers and to evaluate the prognostic value of both nuclear and cytoplasmic p53 as assessed by immunohistochemistry.

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Materials and methods

Patients

The study comprises 205 patients with stage I breast cancer, according to the International Union Against Cancer staging system, i.e. tumours smaller or equal to 20 mm, with a negative lymph node status and without distant metastasis. The patients were diagnosed during 1985–88 and registered at the Oncological Centre, South-East Health Care Region of Sweden. General mammographic screening was introduced in the region in 1987. None of the patients received any preoperative treatment. Patients were treated with breast-conserving surgery plus 54 Gy breast irradiation or modified radical mastectomy. Approximately half of the patients received adjuvant tamoxifen treatment, mainly patients over the age of 50 with oestrogen (ER) or progesterone (PR) receptor-positive tumours. Distant recurrence was registered in 15 patients. The median follow-up period was 5 years.

Immunohistochemistry

The monoclonal antibodies PAb1801 and DO1 (Oncogene Science, Manhasset, NY, USA) were used. As a negative control we used IgG antibodies (Sigma, St Louis, MI, USA). The tumours were collected from fresh surgical resections and stored below –70°C before they were sectioned. The 6 µm frozen sections were air dried and stored at –20°C. The sections were then fixed in acetone (4°C) for 10 min and air dried. Endogenous peroxidase activity was quenched with 0.6% hydrogen peroxide in methanol for 5 min; thereafter the slides were rinsed with phosphate-buffered saline (PBS) with 0.1% bovine serum albumin and 0.5% Tween (PBSA + Tween). To block endogenous avidin-binding activity the tissue was first treated with avidin (0.001%) and then, after rinsing with PBSA + Tween, with biotin (0.01%) (Sigma). The sections were rinsed and placed in PBSA + Tween for 5 min. Normal goat serum (1:5) was used for 20 min in order to block non-specific immunostaining. The sections were incubated with the primary antibodies Abl1801 (1:50) and DO1 (1:300) for 30 min, and then with biotinylated goat antibody (1:500) for 30 min. After being rinsed with PBSA + Tween, they were incubated with streptABCComplex/horseradish peroxidase (1:500) for 30 min (StreptABCComplex/HRP Duet, Dako, Glostrup, Denmark). The sections were rinsed with PBSA + Tween before being stained with 3,3-diaminobenzidine tetrahydrochloride (DAB).
in PBS with 0.036% hydrogen peroxide for 8 min and then rinsed with distilled water, counterstained with haematoxylin, dehydrated in a series of ethanol, cleared in xylene, and mounted.

**DNA flow cytometry**

DNA flow cytometry was used to determine DNA content and S-phase fraction. A suspension of isolated nuclei was prepared as described by Vindelöv et al. (1983) using a detergent (NP40), trypsin and RNase. Finally, propidium iodide was used to stain DNA. The suspensions were analysed with a Leitz MPV FLOW flow cytometer (Leitz, Wetzlar, Germany) interfaced to a Monroe OC8888 personal computer system (Litton Business, Morris Plains, NJ, USA). The DNA content of the tumour stemlines were related to the internal reference cells, which were chicken and trout erythrocytes. Tumours with a single G1 peak in the near-diploid range were classified as DNA-diploid and others as DNA-aneuploid. Considering a rectangular S-phase distribution, the number of cells in S-phase was estimated by multiplying the number of channels between the G2M and G1-M peaks by the mean number of registrations per channel in an interval judged as representative for replicating cells.

**Hormone receptor analyses**

Hormone receptor analysis was performed on specimens collected from fresh surgical resections and stored below -70°C. Tumours diagnosed before 1988 were analysed for ER and PR according to Wrangle et al. (1978). The cytosol was incubated with radioligands and the receptors were isolated by isoelectric focusing in polyacrylamide gel. Tumours collected after 1988 were analysed by the Abbott enzyme immunoassay (EIA) method. The receptor concentration was expressed as fmol of receptor per µg of DNA. A cut-off value of 0.1 fmol µg⁻¹ DNA was used for receptor positivity.

**Histological grading**

Blocks available at the Department of Pathology in Linköping were reviewed by a pathologist. Seventy cases were histologically typed and graded. Sixty-three cases were classified as ductal carcinomas and seven as lobular. One tumour with a predominant in situ component and only a small invasive component, and one case of medullar carcinoma were not graded. The grading was done according to Bloom and Richardson (1957).

**Statistical methods**

Chi-square tests for contingency tables were used for testing the significance of differences in p53 accumulation in relation to pathobiological variables. Survival curves were computed with the product limit method presented by Kaplan and Meier (1958). We used Cox's proportional hazards model (1972) to estimate and test the relation of p53 accumulation and other prognostic variables to distant recurrence-free survival. P-values ≤ 0.05 were regarded as significant.

**Results**

**p53 accumulation**

The sections were independently examined and scored by two of the authors. Nuclear staining was scored as negative or, if only a small proportion of the nuclei were stained (1–20%), as weakly positive, or as strongly positive if over 20% of the nuclei were clearly stained. Cytoplasmic staining was scored as negative or positive. The negative controls were all negative.

The staining patterns for both antibodies were very similar, with a 95% concordance for the nuclear staining and 93% for the cytoplasmic staining. The results for 1801 and DO1 were combined and the final score was defined as the highest score between the two antibodies (Table 1). Positivity irrespective of nuclear or cytoplasmic location was defined as cellular accumulation.

**p53 accumulation and pathobiological variables**

When comparing the degree of p53 accumulation pattern against the various pathobiological variables, the p53-negative tumours and tumours with weak positivity were similar and were thus grouped together. Our further analyses were therefore based upon the groups of negative or weakly positive compared with the group of strongly positive. One of the strongly positive cases is shown in Figure 1. Nuclear and cellular p53 accumulation showed the same relation to the other variables. Table II shows how nuclear p53 accumulation was related to different variables. Patient age under 50, negative ER status, DNA aneuploidy, high S-phase fraction and poorly differentiated tumours were significantly correlated with strong nuclear p53 accumulation.

**p53 accumulation in relation to distant recurrence-free survival**

The estimated distance recurrence-free survival after 5 years was 73% for those patients with tumours having a high nuclear p53 accumulation compared with 95% for those lacking or having little p53 accumulation (Figure 2). The distribution of patients having distant recurrence is shown in Table I. According to Cox regression analysis the recurrence rate ratio was 6.2 (P = 0.002). In the univariate Cox analysis, both high nuclear and high cellular p53 accumulation were associated with a poor prognosis, as were negative ER status, DNA aneuploidy and a high S-phase fraction (Table III). Cytoplasmic accumulation did not have any additional value to that of nuclear staining (Figure 3). In the multivariate analysis, S-phase fraction and DNA ploidy independently predicted a higher risk of distant recurrence. p53 accumulation added prognostic information to that of S-phase fraction and DNA ploidy when analysed separately, but not when

**Table 1 Number of cases and intracellular staining pattern of accumulated p53 for the antibodies PAb1801 and DO1 combined**

|       | C- | C+ |
|-------|----|----|
| N-    | 145 (7) | 7 |
| N+    | 16 (1) | 14 (1) |
| N+    | 4 (1) | 19 (5) |
|       | 165 | 40 |
|       | 205 |     |

N-, N+, N++, nuclear negative, weakly positive and strongly positive respectively. C-, C+, cytoplasmic negative and positive respectively. Numbers of patients having distant recurrence are within parentheses.
analysed with both at the same time. When all variables were included in the multivariate Cox analysis, p53 accumulation failed to be an independent prognostic factor, as did ER status and size.

Discussion

Accumulated p53 in node-negative breast cancer has been shown to be associated with an aggressive course and a poor prognosis (Isola et al., 1992; Allred et al., 1993; Silvestrini et al., 1993). This study showed that the same can be applied to small breast cancers (T1N0) and that the occurrence of p53 accumulation also is of importance in small tumours. Accumulation of p53 is more often seen in advanced stages of breast cancer (Davidoff et al., 1991a), but when seen in earlier stages the p53 alteration is maintained during breast cancer progression (Davidoff et al., 1991b; Bartkova et al., 1993). Alteration of the p53 gene is thus suggested to take place early in cancer progression, rather than being involved in the development of metastases.

Cellular p53 accumulation, irrespective of nuclear or cytoplasmic location, has been shown to add prognostic information in stage II breast cancer (Stenmark-Askmalm et al., 1994). In this study no additional prognostic value for cytoplasmic overexpression was found. The role of cytoplasmic p53 accumulation is controversial. In this study we did observe cytoplasmic staining but, when present, usually accompanied nuclear staining. It is possible that cytoplasmic accumulation reflects a more advanced stage of tumour progression since an increased prevalence of cytoplasmic accumulation has been found with increasing tumour stage in colorectal cancer using the CM1 antibody (Sun et al., 1992; Bosari et al., 1994). Leakage from the nucleus to the cytoplasm due to poor fixation has also been suggested as an explanation for cytoplasmic staining (Fisher et al., 1994).

Table II  Strong nuclear p53 accumulation for PAb1801 and DOI combined related to different pathological variables

|                     | n   | n (%) | P-value |
|---------------------|-----|-------|---------|
| Age (years)         |     |       |         |
| < 50                | 60  | 11 (18)| 0.038   |
| ≥ 50                | 145 | 12 (8) |         |
| Tumour size (mm)    |     |       |         |
| < 10                | 48  | 4 (8)  | 0.47    |
| 11–20               | 157 | 19 (12)|         |
| Oestrogen receptor status |       |       |         |
| Negative            | 52  | 14 (27)| <0.0001 |
| Positive            | 153 | 9 (6)  |         |
| DNA ploidy type     |     |       |         |
| Diploid             | 108 | 3 (3)  | <0.0001 |
| Aneuploid           | 97  | 20 (21)|         |
| S-phase fraction (%)|     |       |         |
| < 10                | 144 | 7 (5)  | <0.0001 |
| ≥ 10                | 61  | 16 (26)|         |
| Histological grade  |     |       |         |
| Well differentiated  | 30  | 1 (3)  | <0.0001 |
| Moderately differentiated | 21 | 1 (5) |         |
| Poorly differentiated| 19  | 9 (47) |         |
*Seventy cases were graded. N++, nuclear strongly positive.

Table III  Univariate regression analysis (Cox model) of distant recurrence related to strong nuclear p53 accumulation and other prognostic variables

|                     | n  | Rate ratio | 95% CI  | P-value |
|---------------------|----|------------|---------|---------|
| Nuclear p53 status  |    |            |         |         |
| Nuclear negative weakly positive | 182 | 1 | 2.2–17.4 | 0.002 |
| Nuclear positive    | 23 | 6.2       |         |         |
| Tumour size (mm)    |    |            |         |         |
| < 10                | 48 | 1         |         |         |
| 11–20               | 157| 0.5       | 0.17–1.5| 0.22    |
| Oestrogen receptor status |     |       |         |         |
| Negative            | 52 | 1         |         |         |
| Positive            | 153| 0.35      | 0.12–0.96| 0.048  |
| DNA ploidy type     |    |            |         |         |
| Diploid             | 108| 1         |         |         |
| Aneuploid           | 97 | 7.9       | 1.8–35.1| 0.0008  |
| S-phase fraction (%)|    |            |         |         |
| < 10                | 144| 1         |         |         |
| ≥ 10                | 61 | 6.4       | 2.03–20.1| 0.0006  |

Figure 2  Distant recurrence-free survival related to nuclear p53 immunoreactivity for PAb1801 and DOI combined, regardless of cytoplasmic accumulation. N– N+, negative or weakly positive; N+++, strongly positive.

Figure 3  Distant recurrence-free survival related to cytoplasmic and or nuclear p53 immunoreactivity for PAb1801 and DOI combined. N– N+, nuclear negative or weakly positive; N++, nuclear strongly positive; C+, cytoplasmic positive and C–, cytoplasmic negative.
However tumours can exhibit a very heterogeneous staining pattern, with tumour cells having only nuclear accumulation, both strong nuclear and cytoplasmic staining and mainly cytoplasmic staining present within the same tissue section. Distinct staining of both nucleus and cytoplasm throughout the section or staining of nuclei only were other staining patterns observed. Thus, it is unlikely that cytoplasmic staining is satisfactorily explained by poor fixation only.

In line with the results of others (Isola et al., 1992) strong accumulation of p53, in contrast to weak accumulation, was related to other prognostic factors. There are difficulties in interpreting the staining patterns in immunohistochemistry. Fisher et al. (1994) have described the importance of optimal fixation in paraffin-embedded tissue. In another study the staining intensity did not reflect the proportion of tumour cells having a gene mutation, but it did show that positive staining to a great extent corresponded to gene mutations (Jacquemier et al., 1994).

Results with the two primary antibodies correlated strongly. If at least one of the two antibodies showed positive staining, the tumour was regarded as positive. In order to reduce variation in immunoreactivity when using different antibodies, and to obtain a higher sensitivity, the staining results for different antibodies have also been combined in other studies (Allred et al., 1993; Jacquemier et al., 1994).

Strong nuclear p53 accumulation was significantly correlated to several pathobiological variables, thus indicating an aggressive, genetically instable and rapidly proliferating tumour. In fact, poorly differentiated tumours showed strong overexpression of p53. This is similar to results of other studies which show that a high malignancy grade is correlated with p53 accumulation (Isola et al., 1992; Domagala et al., 1993; Lipponen et al., 1993). Breast cancer patients with tumours exhibiting a high S-phase fraction have been shown to respond to adjuvant cytotoxic treatment (Stål et al., 1994). It would be of interest to investigate whether tumours with p53 accumulation, which often exhibit high S-phase fraction, respond to such treatment. On the other hand, p53 plays an active role in apoptosis and cells lacking functional p53 protein have been shown to be resistant to both radiation and cytotoxic drugs in vitro experiments (Lowe et al., 1993).

Our results show that p53 accumulation can predict recurrence-free survival. In this study p53 failed to be an independent prognostic factor when several variables were included in a multivariate analysis. However, it should be stated that the number of relapses was low. p53 accumulation added prognostic information to that of S-phase fraction, which in a previous study of stage I breast cancer was shown to be an independent prognostic factor after adjustment for other variables (Stål et al., 1993). In accord with the study of Isola et al. (1992), less than half of the highly proliferative tumours showed strong overexpression of p53 protein. A larger study is needed to further investigate the role of p53 as an independent prognostic factor in stage I breast cancer.

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