First record of *Chilomima clarkei* (Amsel) (Lepidoptera: Crambidae) in *Manihot esculenta* Crantz (Euphorbiaceae) in Amapá state, Brazil

Ricardo Adaime1, Rodrigo Souza Santos2,3, Adilson Lopes Lima4, Jhulie Emille Veloso dos Santos2 & Alexandre Specht2

1. Embrapa Macapá, Laboratório de Proteção de Plantas, Brazil. 2. Embrapa Acre, Brazil. 3. Faculdade de Macapá (FAMA), Brazil. 4. Embrapa Cerrados, Brazil.

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**Corresponding author:**
Rodrigo Souza Santos
rodrigo.s.santos@embrapa.br

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**Abstract:** The present study reports *Chilomima clarkei* (Amsel) (Lepidoptera: Crambidae) infesting stems of *Manihot esculenta* Crantz (Euphorbiaceae) in Amapá state, Brazil, for the first time. Considering the importance of this species as a pest, it is necessary to publicize its occurrence and provide information about the main symptoms of the attacked plants and morphological characters of development stages to identify it quickly in crops. As there are no registered products for this pest’s control, producers are advised to pay attention to any sign of infestation. To avoid it from spreading, they are also recommended to remove and burn all the infested parts detected in the plantations, in addition to selecting healthy manivas (stem cuttings) at the time of replanting, preventing the infestation of new crops.

**Keywords:** Amazon; Stem borer; Infestation; Stem borer caterpillar; Moth

**Cassava,** *Manihot esculenta* Crantz (Euphorbiaceae), also known in Brazil as *aipim* or *macaxeira* (NASSAR et al. 2008), is a semi-perennial, bushy plant with tuberous root (CÉBALLOS & LA CRUZ 2012). It has the physiological characteristic of storing starch in its roots (FUKUDA et al. 2006).

In the north region of Brazil, cassava and its varieties are widely cultivated and consumed by the population, in addition to being an important economic activity for producers, especially the family-based ones (RODRIGUES et al. 2007; FAZOLIN & ESTRELA 2016). In 2018, the area devoted to cassava cultivation in Brazil was 1,222,019 ha, 427,993 ha in Amapá state, Brazil (IBGE 2020). In 2018, the area devoted to cassava cultivation in Brazil was 1,222,019 ha, 427,993 ha in Amapá state, Brazil (IBGE 2020).

Among the insects that attack cassava in the field are a wide variety of lepidopterans, known as butterflies and moths, associated with cultivation (SILVA et al. 1968). Among the moths, representatives of the following families are reported: 1. Saturniidae [Rothschildia spp., Bunaea alcinoe (Stoll), Imbrasia dione Fabricius, Nudaurelia petiveri (Guérin-Méneville), Rothschildia aurota (Walker), Rothschildia hesperus (Linnaeus), Rothschildia jacobae (Walker) and Samia Cynthia (Drury)], 2. Sphingidae [Cocytius duponchel (Poez), Erinnyis alope (Drury), Erinnyis crameri (Schaus), Erinnyis ello Linnaeus, Manduca albiplaga (Walker), Manduca sexta (Linnaeus)], 3. Arctiidae [Alpenus nigropunctatus (Bethune-Baker), Disparctica vittata (Bruce), Hypercompe hambletoni (Schaus) and Phoenixprocta sanguinea (Walker)], 4. Geometridae [Coloeca simula (Warren) and Hyposidra talaca (Walker)], 5. Hepialidae [Endocitta sericea (Swinhoe)], 6. Lasiocampidae [Pachypasa papiy Tams], 7. Limacodidae [Prototoia sjoestedt (Aurivillius)], 8. Lymantriidae [Callithea horsfieldii (Saunders)], 9. Erebidae [Hypercompe hambletoni (Schaus)], 10. Noctuidae [Agrotis ipsilon (Hufnagel), Spodoptera eridania (Cramer), Spodoptera littoralis (Boisdau), Spodoptera litura Fabricius, Chrysodeixis includens (Walker), Tiracola plagiai (Walker) and Felita straburra (Fabricius)], 11. Nymphalidae [Anarta jatrophae (Linnaeus), Colobura derce (Linnaeus) and Charaxes affinis Butler], 12. Psychidae [Euxeta variegata (Snellen)], 13. Pyralidae [Asciodes gordialis (Guenée), Cadra figuliella (Gregson), Chilo belifalpis (Hamphson), Chilozele trapeziana (Sepp), Condylorrhiza vestigialis (Gréigson), Omiodes indicata (Fabricius), Pyphocera atfiusella Hampson and Glaphyria sp.], 14. Crambidae [Chiloimima clarkei (Amsel), Chilozele trapeziana (Sepp) and Condylorrhiza vestigialis (Guenée)], 15. Uranidae [Urania fulgens (Walker)] and 16. Lycaenidae [Eumaeus atola (Poez) (MOSCHEL 1878; MÜLLER 1886; BONNINGHAUSEN 1899; SAUER 1943; CURSEIL 1955; EHRLICH & RAVEN 1964; SILVA et al. 1968; SILVA et al. 1981; COTO et al. 1995; BELLOTTI et al. 2002; MÜLLER-HILLER 2008; VANDER HEYDEN 2009; MONTIEL et al. 2011; 2013; 2014; 2016; 2018; SPECHT et al. 2015; BRITO et al. 2019; NHM 2020).

Of all these lepidopterans, the representatives of Sphingidae, whose caterpillars voraciously attack the leave, must be...
highlighted as pests. As well as the representatives of Crambidae, due to the boring habit, whose larvae and pupae stay inside the stems (branches), protected from the weather, natural enemies, and even chemical control agents (Bellotti 2008). In the north region of the country, the cassava hornworm, Erinnyis ello (Linnaeus) (Lepidoptera: Sphingidae) is notably the key pest of the crop, due to the voracity with which the caterpillars consume the leaves and the economic losses resulting from its infestations in the region (Rodrigues et al. 2007; Fazolin & Estrela 2016).

The record presented here refers to a technical visit made to a rural property (00°40'14.1 "N; 50°46'48.6 "W), located in the municipality of Itaubal do Piririm, state of Amapá, in November 2017. There were reports of an unknown phytosanitary problem in cassava cultivation. During an inspection in the plantation, holes in the stems and sawdust-like material were verified in the plants (Figure 1A).

Nearly 30% of the stems were infested, showing sawdust-like frass. During the inspection, stems with signs of infestation were collected. Then, with a pocket knife, they were cut lengthwise, and galleries were checked. These galleries were formed by the consumption of soft tissue (medulla), and caterpillars and pupae with characteristics of lepidopterans were found inside them (Figures 1B and 1C).

Other stems were collected and taken to the Embrapa Amapá Plant Protection Laboratory, in Macapá, to be maintained in laboratory conditions until adults emergence for further identification. After approximately 40 days, 15 moths were obtained and identified as Chilomima clarkei (Amsel) (Lepidoptera: Crambidae) (Figure 1D). Voucher specimens from the research were deposited at the Embrapa Amapá Plant Protection Laboratory.

The adult of C. clarkei is 25 to 35 mm long, with the forewings characterized by dark bronze bands. The moths are nocturnal, resting on plant stems during the day, in a head-down position. Females lay up to 200 eggs on the stems near the axillary buds. After four to six days, they hatch into larvae, which feed on the superficial layer of the stem and weave a type of capsule where they are protected. In the fifth instar, the larvae penetrate the stem to feed, completing their development and turning into pupae, which remain protected inside the stems. Depending on environmental conditions, the larval stage lasts 32 to 64 days, and the pupal stage 12 to 17 days (Löhr 1983; Bellotti 2008).

In South America, there are records of C. clarkei in some countries such as Colombia, Venezuela, and Argentina (Vides et al. 1996; Almonacid et al. 2016). In Brazil, it is considered a secondary pest of cassava cultivation, which sporadically causes economic damage in some localities (Rodriguez et al. 2009). In the 1990s, there was a significant infestation of this species on the Atlantic Coast of Colombia, generating a very expressive social problem (Bellotti et al. 2002).

The pest limits cassava production in Colombia, both for obtaining roots and genetic material for propagation (Vides et al. 1996). When the infestation is very high, there is a decrease in the quantity and quality of propagation material. Studies of economic damage with artificial infestations (8 to 12 larvae/plant and 16 to 20 larvae/plant) showed a 45 to 62% reduction in root production (Löhr 1983). In Brazil, the economic impact of this species on cassava crops is still unknown.

The identification of C. clarkei is essential because, along with it, some other lepidopterans and coleopterans form a borer complex, such as species of Sternocoelus (= Coelosternus) (Coleoptera: Curculionidae) that occur in Brazil, whose signs and damage in plants are similar (Gallo et al. 2002; Bellotti 2008).

**Figure 1.** Chilomima clarkei (Lepidoptera: Crambidae). A. stem of Manihot esculenta with symptoms of infestation by larvae; B. larva at the end of development, removed from the inside of the cassava stem; C. pupa inside the stem; D. adult moth in dorsal view. Photos: Adilson Lopes Lima.
So far, there are no products registered with the Ministry of Agriculture, Livestock and Food Supply (MAPA) to control *C. clarkei* in cassava cultivation (AGROFIT 2020). In this regard, we reinforce the importance of registering the species with potential damage to cassava crop in the regions cultivated in Brazil, so that further studies aimed at their management may be demanded. However, there are records of the parasitoids *Braccon* sp., *Apanteles* sp., *Brachymyzeria* sp., *Agathis* sp. (Hymenoptera: Braconidae), *Tetrastichus howardii* (Olliff) (Hymenoptera: Eulophidae) and *Trichogramma* sp. (Hymenoptera: Trichogrammatidae) in the literature, parasitizing eggs and caterpillars of *C. clarkei* and the entomopathogens *Metarhizium anisopliae* (Metchnikoff) Sorokin and *Beauveria bassiana* (Bals.) Vuill. (Loehr 1981; Bellotti et al. 2008), acting as natural enemies of this pest.

For main management methods, producers are recommended to select healthy *manivas* (stem cuttings for planting) at the time of replanting, to avoid the contamination of new crops. In plantations where the pest has been detected, producers should remove and burn all the infested parts, avoiding dissemination during regrowth. In infested areas, besides the destruction of the attacked parts, Bertorelli et al. (2006) recommend using light traps with ultraviolet fluorescent lamps (model Luiz de Queiroz) for monitoring and reducing the moth population in the area.

Considering the social and economic relevance of the cassava cultivation for the Amapá state and the Brazilian Amazon, it is imperative to direct efforts towards publicizing the occurrence of *C. clarkei*, describing morphological characters of development stages and characterizing the main symptoms of the attacked plants. Thus, it will be possible to identify this pest in the crops quickly, so that decisions are made before its population levels cause economic losses. Studies should also be carried out to quantify regional damages and, specifically, develop or adapt management methods to avoid significant losses to production.

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