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Studies of Laboulbeniales on Myrmica ants (IV): host-related diversity and thallus distribution patterns of Rickia wasmannii

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Abstract – Fungal species identities are often based on morphological features, but current molecular phylogenetic and other approaches almost always lead to the discovery of multiple species in single morpho-species. According to the morphological species concept, the ant-parasitic fungus Rickia wasmannii (Ascomycota, Laboulbeniales) is a single species with pan-European distribution and a wide host range. Since its description, it has been reported from ten species of Myrmica (Hymenoptera, Formicidae), of which two belong to the rubra-group and the other eight to the phylogenetically distinct scabrinodis-group. We found evidence for R. wasmannii being a single phylogenetic species using sequence data from two loci. Apparently, the original morphological description (dating back to 1899) represents a single phylogenetic species. Furthermore, the biology and host-parasite interactions of R. wasmannii are not likely to be affected by genetic divergence among different populations of the fungus, implying comparability among studies conducted on members of different ant populations. We found no differences in total thallus number on workers between Myrmica species, but we did observe differences in the pattern of thallus distribution over the body. The locus of infection is the frontal side of the head in Myrmica rubra and M. sabuleti whereas in M. scabrinodis the locus of infection differs between worker ants from Hungary (gaster tergites) and the Netherlands (frontal head). Possible explanations for these observations are differences among host species and among populations of the same species in (i) how ant workers come into contact with the fungus, (ii) grooming efficacy, and (iii) cuticle surface characteristics.

Key words: Ant-associated fungi, Laboulbeniomycetes, Molecular evolution, Ribosomal DNA, Thallus density.
Introduction

Ants harbour a vast diversity of microbial parasites and pathogens. Fungal species of ants are usually pathogenic, but some species, notably members of Laboulbeniales (Ascomycota), are ectoparasitic and do not cause the death of the hosts. Laboulbeniales are developmentally unique in that they do not produce mycelia; instead, they produce multicellular units, thalli, which attach externally to the integument of the host. Rickia wasmannii Cavara, 1899 [9] (Fig. 1) is a species of Laboulbeniales that infects diverse ants in the genus Myrmica Latreille, 1804 (Hymenoptera, Formicidae) in Europe. Knowledge on the biology of R. wasmannii is accumulating and this species has quickly become one of the most thoroughly researched species of Laboulbeniales [2–5, 11–13, 15, 29, 32, 52, 53, 70].

Non-random positional patterns on the integument [48], variation in host usage across geographical regions [29], and habitat specificity [53] have recently been explored for R. wasmannii. On the other hand, the phylogenetic diversity of R. wasmannii from different host species remains unknown. This question deserves to be explored, as it was shown recently for a few Laboulbeniales examples that there is phylogenetic structuring within presumed species. For species in the genera Gloeandromyces Thaxt. and Hesperomyces Thaxt., phylogenetic segregation by host species has been observed. For example, in both G. pageanus Haelew. and G. streblae Thaxt., two phylogenetic clades can be found: one clade for isolates removed from Trichobius dugesiioides Wenzel, 1966 (Diptera, Streblidae) and another clade for isolates from T. joblingi Wenzel, 1966 [33, 34]. Similarly, Hesperomyces virens Thaxt. consists of multiple clades, each clade corresponding to a species with strict host specificity [30].

Even though the main hosts of R. wasmannii all belong to a single genus of ants (for a discussion of alternative hosts, see [53]), the different host species are placed in two clades that are phylogenetically not closely related (referred to as species groups in [39, 56]). Myrmica rubra (Linnaeus, 1758) and M. ruginodis Nylander, 1846 belong to the rubra-group, whereas the other known hosts belong to the scabrinodis-group. These are M. gallienii Bondroit, 1920; M. hellenica Finzi, 1926; M. sabuleti Meinert, 1861; M. scabrinodis Nylander, 1846; M. slovaca Sadil, 1952; M. specioides Bondroit, 1918; M. spinosior Santschi, 1931; and M. vandeli Bondroit, 1920 [4, 29]. Assessing whether R. wasmannii shows phylogenetic segregation by host species or host species group is important to better understand its interactions with different ant hosts. Studies using infected and non-infected Myrmica ants have been done to assess the parasite’s effects on ant behaviour and physiology. Interpretation of these results is complicated when the taxonomic status of different fungal populations is uncertain. Comparing interactions between a fungal parasite and its different hosts is only reliable when the fungal populations represent a single phylogenetic species.

Building on the hypothesis that R. wasmannii is a complex of species, potentially segregated by host species (or species group), it is logical to assume that thallus distribution patterns may be different on various ant hosts. If we were to find variable patterns of thallus distribution, these would have to be (partly) attributed to the fungal partner, the ant partner, environmental factors, or a combination of these. To try to shed light on this complex interaction of factors, we took an integrative approach (sensu [30]) and generated independent sets of data, that is, barcode sequences of R. wasmannii isolates and thallus density counts by body part.

During this study, we sampled infected ants from different regions in Europe and sequenced two loci to assess intraspecific phylogenetic diversity in R. wasmannii. Collected host ants represent three Myrmica species belonging to the rubra- and scabrinodis-groups [56]. After having accumulated many collections of R. wasmannii-infected ants, we assessed thallus densities per body part from different host species (M. rubra, M. sabuleti, and M. scabrinodis) and from different populations of the same host species (M. scabrinodis).

Material and methods

Collection of ants

Ants were collected directly from nests in seven locations in four countries (Fig. 2): Austria (Vienna), Belgium (Moelingen), Hungary (Bükkszentkereszt, Rakaca, Üjléta), and the Netherlands (Savelsbos, Wijhe-Eys). Long-term preservation was in 80–96% ethanol. Identification of ants was based on Seifert [64] and Radchenko and Elmes [56]. Voucher specimens are deposited at the Naturalis Biodiversity Center (Leiden, The Netherlands) and the Hungarian Natural History Museum (Budapest, Hungary). Identification of mounted thalli was done under light microscope, based on Thaxter [70] and De Kesel et al. [15].

DNA extraction, PCR amplification, and sequencing

DNA was isolated from 3 to 100s of thalli using extraction protocols described in [31] or a modified REPLI-g Single Cell Kit (Qiagen, Valencia, California) [30]. The internal transcribed spacer (ITS) region (ITS1–5.8S–ITS2) and the 5′ end of the nuclear ribosomal large subunit (28S) were amplified, for ITS using primer pairs ITS1f [26] & ITS4 [77] and ITS9f/mun [19] & ITS4, for partial ITS–28S using the newly designed Rickia-specific primer RickITS2 (5′–CTAGTTGTAATTGCA-TATTTTAGTG–3′) & LR3 [74], and for 28S-only using
Polymerase chain reactions (PCR) used 13.3 µL of RedExtract Taq polymerase (Sigma–Aldrich, St. Louis, Missouri), 2.5 µL of each 10 µM primer, 5.7 µL of H₂O, and 1.0 µL of template DNA. In some cases, 0.25 µL of dimethyl sulfoxide (DMSO) was added as a PCR enhancer (and 5.45 µL of H₂O). All amplifications were done in an Applied Biosystems 2720 thermal cycler (Foster City, California) with initial denaturation at 94 °C for 3:00 min; followed by 35 cycles of denaturation at 94 °C for 1:00 min, annealing at 50 °C for 0:45 min, and extension at 72 °C for 1:30 min; and final extension at 72 °C for 10:00 min.

PCR products were loaded onto TAE 1% agarose gels for electrophoresis at 100 V for 25 min and UV transillumination was used to check the product size. Products showing strong bands on gel were purified with Qiaquick PCR Purification Kit (Qiagen) or DF100 PCR cleaning kit (Geneaid, New Taipei City, Taiwan) and sequenced using the same primers and 1 µL of purified PCR product per 10 µL sequencing reaction. Sequencing reactions were performed using the Big Dye® Terminator v3.1 Cycle Sequencing Kit (Life Technologies, Carlsbad, California). For molecular work performed in Hungary, sequencing was outsourced to Microsynth AG (Wolfurt-Bahnhof, Austria). Sequence fragments were assembled, trimmed, and manually edited at ambiguous sites in Sequencher 4.10.1 (Gene Codes Corporation, Ann Arbor, Michigan). The identities of our consensus sequences were

Figure 1. The head of a Myrmica sabuleti worker, heavily infected with Rickia wasmannii.

Figure 2. Field sites where ants for this project have been collected. Field sites are located in Europe (Austria, Belgium, Hungary, The Netherlands).
confirmed by performing BLAST searches at http://ncbi.nlm.nih.gov/blast/Blast.cgi. Edited sequences are deposited in NCBI GenBank (accession numbers in Table 1).

Datasets and phylogenetic analyses

Individual datasets for ITS and 28S were constructed in order to assess intraspecific phylogenetic diversity in Rickia wasmannii. Alignments were done using MUSCLE v3.7 [18] on the Cipres Science Gateway v3.3 [50] and checked in BioEdit v7.2.6 [36]. Ambiguously aligned regions and uniformative positions were removed using trimAl v1.3 [8] with 60% gap threshold and minimal coverage of 50%. We also constructed a combined ITS + 28S dataset. The aligned sequence data for each region were concatenated in MEGA7 [43] to create a matrix of 804 bp with phylogenetic data for 16 isolates.

Maximum parsimony (MP) analyses were run using PAUP on XSEDE [69]. MP was estimated with heuristic searches consisting of 500 stepwise-addition trees obtained using random sequence addition replicates followed by tree bisection-reconnection (TBR) branch swapping (MultiTrees in effect) and saving all equally most-parsimonious trees. Robustness of branches was estimated by maximum parsimony bootstrap proportions using 500 replicates, with heuristic searches consisting of 10 stepwise-addition trees obtained using random sequence addition replicates followed by TBR branch swapping, with MaxTrees set at 100. Maximum likelihood (ML) analyses were run using IQ-TREE [10, 51] from the command line. Nucleotide substitution models were selected under Akaike information criteria for small sample size (AICc) with the help of jModelTest 2 [14] in Cipres [50]. For the ITS dataset, the TPM1 + G model was selected (−lnL = 712.0173); for 28S, the TrN + G model (−lnL = 1371.6022). ML was inferred for each individual dataset under the appropriate model, and for the concatenated dataset under partitioned models. Ultrafast bootstrap analysis was implemented with 1000 replicates [37]. Phylogenetic reconstructions with bootstrap values (BS) were visualised in FigTree v1.4.3 (http://tree.bio.ed.ac.uk/software/figtree/).

Species delimitation

We used three species delimitation methods to validate species limits of or within Rickia wasmannii (fide [30, 34]): Automatic Barcode Gap Discovery method [55], General Mixed Yule Coalescent method [54], and a Poisson tree processes model approach [79]. All analyses were done with both the ITS and 28S datasets; the ITS region has been proposed as the universal barcode for all fungi [63] whereas the 28S locus was recently put forward as potential barcode for Laboulbeniomycetes because it is easy to amplify and has high discriminative power [30, 75]. We used the following parameters in the online version of ABGD (https://wwwabi.snv.jussieu.fr/public/abgd/abgdweb.html): \(P_{\text{min}} = 0.001, P_{\text{max}} = 0.01, \text{steps} = 10, N_0 \text{ bins} = 20\). We evaluated results for both the Jukes-Cantor (JC69) and Kimura two-parameter (K80) distance metrics [40, 41] and for four gap width values (X); 0.1, 0.5, 1.0, and 1.5. We used the online version of bPTP (http://species.h-its.org) with default values for all parameters (number of MCMC generations, thinning, burn-in, seed). Finally, we conducted GMYC in R (R Core Team 2013) using the packages mcl [49] and splits [25]. The MCC tree from Bayesian inference (BI) served as input for both the bPTP and GMYC analyses.

Bayesian analyses were run for individual datasets with a Markov Chain Monte Carlo (MCMC) coalescent approach implemented in BEAST v1.8.4 [17], under a strict molecular clock assuming a constant rate of evolution across the tree. We selected the Birth-Death Incomplete Sampling speciation model [66] as tree prior and the nucleotide substitution model selected by jModelTest 2 [14] under AICc. Four independent runs were performed from a random starting tree for 10 million generations with a sampling frequency of 1000. Using the same settings failed to converge for the ITS dataset, and we thus

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**Table 1.** Overview of Rickia sequences used in this study. All isolates for which sequences were generated are listed, with GenBank accession numbers as well as host species, country, and year of collection.

| Isolate             | Species               | ITS         | 28S         | Host          | Country     | Year |
|---------------------|-----------------------|-------------|-------------|---------------|-------------|------|
| ADK6272a            | R. wasmannii          | MK500050    | MK500050    | Myrmica sabuleti | Belgium     | 2015 |
| ADK6274c            | R. wasmannii          | MK500051    | –           | Myrmica sabuleti | Belgium     | 2015 |
| DE_Rak4             | R. wasmannii          | KT800050    | KT800021    | Myrmica scabrinodis | Hungary     | 2014 |
| Bükkzszenkereszti2016 | R. wasmannii         | MK500052    | –           | Myrmica scabrinodis | Hungary     | 2016 |
| Ujjeta2014          | R. wasmannii          | MK500053    | MK490857    | Myrmica scabrinodis | Hungary     | 2014 |
| Ujjeta2015-4        | R. wasmannii          | MK500054    | –           | Myrmica scabrinodis | Hungary     | 2015 |
| Wien2015-1          | R. wasmannii          | MK500055    | MK490858    | Myrmica rubra  | Austria     | 2015 |
| D. Haelew. 1234a     | R. wasmannii          | MH040595    | MH040595    | Myrmica sabuleti | Netherlands | 2013 |
| Wien2016-1          | R. wasmannii          | MK500056    | –           | Myrmica rubra  | Austria     | 2016 |
| Wiensabuleti2016-1  | R. wasmannii          | MK500057    | –           | Myrmica sabuleti | Austria     | 2016 |
| SR1s                | R. pachyiulus         | MH040593    | MH040593    | Pachyiulus hungaricus | Serbia     | 2015 |
| SR8s                | R. pachyiulus         | MK500058    | MK500058    | Pachyiulus hungaricus | Serbia     | 2015 |
| SR13s               | R. pachyiulus         | MK500059    | MK500059    | Pachyiulus hungaricus | Serbia     | 2015 |
| SR4s                | R. laboulbenioides    | MH040592    | MH040592    | Cylindroiulus punctatus | Denmark    | 2015 |
| SR5s                | R. laboulbenioides    | MK500060    | MK500060    | Cylindroiulus punctatus | Denmark    | 2015 |
| SR12s               | R. uncigeri           | MK500061    | MK500061    | Unciger foetidus | Denmark     | 2015 |
optimised settings, selecting the GMRF Bayesian Skyride coalescent tree prior and increasing the number of generations to 80 million (with sampling frequency of 8000). Settings of priors were entered in BEAUti [17] to generate an XML file, which was run using BEAST on XSEDE in Cipres (two runs) and locally from the command line (two runs). The resulting log files were entered in Tracer [57] to check trace plots for convergence and to adjust burn-in. Burn-in values were changed for each log file to achieve net Effective Sample Sizes of ≥200 for sampled parameters. While removing a portion of each run as burn-in, log files and trees files were combined in LogCombiner. TreeAnnotator was used to generate consensus trees (0% burn-in) and to infer the Maximum Clade Credibility (MCC) tree, with the highest product of individual clade posterior probabilities.

**Thallus density counts**

Thallus density was determined on 354 *Myrma*ica workers. Workers originated from Austria (Vienna), Hungary (Bükkzséntkereszt, Rakaca, Újléta), and the Netherlands (Savelbos, Wijlre-Eys). Thalli of workers were counted under a stereomicroscope at 40×. Thalli were counted on workers of *M. rubra* (34 workers from Vienna), *M. sabuleti* (three workers from Savelbos, 47 from Wijlre-Eys), and *M. scabrinodis* (50 workers from Bükkzséntkereszt, 100 workers from Rakaca, 100 from Újléta, 20 from Wijlre-Eys). Counts were done on recently sampled workers. Counting took place with the workers submerged in H2O, which increased visibility of thalli.

**Statistical analyses**

We used both absolute and relative values of counted thallus numbers for each body part in statistical analyses. The former is simply the number of thalli counted on a given body part, whereas the latter is calculated as the absolute number of thalli on a given body part divided by total number of thalli on the worker body. We used *R* for all presented statistical data analyses (R Core Team 2018).

**Absolute and relative thallus numbers**

To test for significant differences in total number of thalli between *Myrmica* species, we used a quasi-Poisson generalized linear regression model, in which the number of counted thalli was the response variable, and ant species was the predictor. Quasi-Poisson was preferred over a classical Poisson model, because the count-data showed considerable over-dispersion. Model summaries for models containing factor variables in *R* generally present parameter estimates contrasting them to an arbitrarily selected factor level, so with factor variables with more than two levels, some contrasts are not shown. To acquire factor level comparisons not shown in the summary, the package *lsmeans* was used [45].

Next, we compared absolute thallus number of given body parts between species. To do so, we used multiple Conover-Iman tests of the package *conover.test* [16]. In each test, we tested the difference between *Myrmica* species in the counted thallus number on a given body part. Following the tests (resulting in 48 comparisons), we applied Bonferroni’s *P*-value adjustment to avoid Type I error results. In the results, we only considered tests as significant if Bonferroni-adjusted *P*-values were below 0.05. We compared relative thallus number on given body parts as well, also using Conover-Iman tests. Similarly, we used Bonferroni’s adjustment on the *P*-values from the Conover-Iman test results.

To visualise species differences in the pattern of infection over the body of ants we used the Barnes-Hut implementation of t-distributed stochastic neighbour embedding (t-SNE [72, 73]) with the package *Rtsne* [42]. We chose this method over implementing a Principle Component Analysis approach, because in t-SNE we could explicitly specify the number of dimensions onto which to reduce the original data. Therefore, we were able to plot infection patterns (both of absolute and relative thallus number) on a 2D scatterplot. For t-SNE we used the square root-transformed values for both absolute and relative thallus numbers.

**Potential origin of infection on the body**

It has been suggested that infection with *R. wasmannii* starts from the ant head [15, 32, 48]. If so, one would expect to see that, in the early stages of infection, only (or mostly) the head is parasitised. Therefore, the relative number of thalli should be high during the first stages of infection (= when total number of thalli is small). Consequently, if the infection spreads from the head to other body parts, we should see a decrease in the relative thallus number on the head simultaneously with the increase of total number of thalli on the whole body.

First, we checked the range (minimum and maximum values) of the relative thallus number on each body part of infected ants, separately for the three ant species. We selected those body parts to be of interest in which one) the minimum value of relative thallus number was larger than zero and two) the maximum value was the largest in comparison to other body parts. In *M. rubra*, the frontal (or dorsal) side of the head and the gaster tergites satisfied our criteria. In *M. sabuleti*, only the frontal side of the head had a minimum relative thallus number value larger than zero, and it had the largest maximum value among all body parts. In *M. scabrinodis*, there was no body part on which the minimum value of relative thallus number was larger than zero, and so we selected the body part with the largest maximum value, which was the gaster tergites. Notably, on the gaster tergites of *M. scabrinodis*, we observed the lowest incidence of zero values in relative thallus number as well. As a result, we decided to use this body part as a starting point to test our hypothesis about the infection’s spread.

To test whether there is indeed a significant negative association between total thallus number and relative thallus number on selected body parts (frontal side of the head, gaster tergites), we used quasi-binomial generalised linear regression models. Using these, we were able to reliably fit models on a numeric scale ranging from zero to one (i.e. on the scale of the data) and to control for over-dispersion in the data. Two models were fitted. We specified the response variables to be the relative thallus number on the frontal side of the head and on the gaster tergites in the first and second model, respectively. In both models, predictor variables were total number of thalli on the
Table 2. Summary of results of MP, ML, BI, and species delimitation analyses (ABGD, bPTP, GMYC). Explanation of symbols and values used: \( pp \) = posterior probability; + under ABGD represents supported clades; 4 under ABGD means that the analysis found support for 4 species hypotheses \( (fule [55]) \) within \( R. wasmannii \) under prior maximum distance \( (P) = 0.001, 0.001292, \) and 0.001668; numbers under bPTP and GMYC are Bayesian support values for delimited species hypotheses.

| Putative species | MP BS | ML BS | \( pp \) | ABGD \( P = 0.001 \) | ABGD \( P = 0.002783 \) | ABGD \( P = 0.01 \) | bPTP | GMYC |
|------------------|-------|-------|---------|-----------------|-------------------|-----------------|------|------|
| \( R. wasmannii \) | 100   | 100   | 1.0     | +               | +                 | +               | 0.869| 0.64 |
| \( R. laboulbenoides \) | 99.3  | 97    | 1.0     | +               | +                 | +               | 0.996| 0.96 |
| \( R. uncigeri \) | 100   | 98    | 1.0     | +               | +                 | +               | 1.000| 0.00 |
| \( R. pachyiuli \) | 99.8  | 98    | 1.0     | +               | +                 | +               | 0.962| 0.80 |
| \( R. wasmannii \) | 100   | 100   | 1.0     | 4               | +                 | +               | 0.974| 0.38 |
| \( R. laboulbenoides \) | 99.9  | 97    | 1.0     | +               | +                 | +               | 0.994| 0.76 |
| \( R. uncigeri \) | 99.8  | 98    | 1.0     | +               | 1.000             | 0.00            | 0.993| 0.63 |
| \( R. pachyiuli \) | 100   | 98    | 1.0     | +               | +                 | +               | 0.993| 0.63 |

Figure 3. Phylogenetic reconstruction of \( Rickia \) species using a combined ITS + 28S dataset. The topology is the result of maximum likelihood inference. For each node, ML and MP bootstraps are presented above and below the branch leading to that node. For each \( R. wasmannii \) isolate, isolate name, \( Myrmica \) host epithet, and country code (AT, Austria; BE, Belgium; HU, Hungary; NL, The Netherlands) are presented. To the right of the phylogeny, results of species delimitation methods are summarised, from left to right: ABGD of the aligned ITS & 28S datasets under most parameters combinations; ABGD of the 28S dataset for \( P = 0.001, 0.001292, \) and 0.001668; numbers under bPTP and GMYC are Bayesian support values for delimited species hypotheses.
whole body, a factor variable generated by specifying species name and country of origin, and the interaction term between these two variables. The factor variable had four levels: (1) *M. rubra* from Austria, (2) *M. sabuleti* from the Netherlands, (3) *M. scabrinodis* from Hungary, and (4) *M. scabrinodis* from the Netherlands.

In both models, we used square root-transformed response and predictor variables. Also, to be able to infer on mean species-level differences using the intercept estimates of the models, we centred the square root-transformed predictor variable at zero by subtracting the mean of the variable from each of its values. Furthermore, in the Results section we report actual regression coefficients for the slopes from each factor level, using the package *jtools* [47].

### Results

#### Phylogenetic analyses, and species delimitation

The ITS dataset comprised 258 characters, of which 182 were constant and 70 were parsimony-informative. A total of 16 isolates were included (Table 1): *Rickia wasmannii* (five isolates as ingroup), *R. laboulbenioides* De Kesel (two isolates), *R. pachyiuli* M. Bechet & I. Bechet (three isolates), and *R. uncigeri* Scheloske (one isolate). *Rickia wasmannii* was retrieved as a monophyletic clade with maximum support from MP, ML, and BI (not shown). In this clade were included *R. wasmannii* isolates of thalli removed from *M. rubra* (two isolates), *M. sabuleti* (four isolates), and *M. scabrinodis* (four isolates).

The 28S dataset comprised 547 characters, of which 423 were constant and 113 were parsimony-informative. A total of 11 isolates were included (Table 1): *Rickia wasmannii* (five isolates as ingroup), *R. laboulbenioides* De Kesel (two isolates), *R. pachyiuli* (three isolates), and *R. uncigeri* (one isolate). *Rickia wasmannii* was retrieved as a monophyletic clade with maximum support from MP, ML, and BI (not shown), including isolates from

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**Table 4.** Contrasts acquired from the model estimating between-species differences in total thalli number. 

| Contrasts of Myrmica spp. | Estimate | SE  | z-ratio | P     |
|---------------------------|----------|-----|---------|-------|
| *M. rubra* – *M. sabuleti* | -0.070   | 0.155 | -0.45   | 0.689 |
| *M. rubra* – *M. scabrinodis* | -0.046   | 0.128 | -0.36   | 0.712 |
| *M. sabuleti* – *M. scabrinodis* | 0.024    | 0.105 | 0.23    | 0.817 |

**Table 3.** Results of the Automatic Barcode Gap Discovery (ABGD) analyses. X, relative gap width; JC69, Jukes-Cantor substitution model; K80, Kimura 2-parameter substitution model.

| Distance | JC69 | K80 |
|----------|------|-----|
| 0.001    | 3    | 3   |
| 0.001292 | 3    | 3   |
| 0.001668 | 3    | 3   |
| 0.002154 | 3    | 3   |
| 0.002783 | 3    | 3   |
| 0.003594 | 3    | 3   |
| 0.004642 | 3    | 3   |
| 0.005995 | 3    | 3   |
| 0.007743 | 3    | 3   |

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**Figure 4.** Total number of thalli on worker bodies of the three examined *Myrmica* species (n = 34 for *M. rubra*, n = 50 for *M. sabuleti*, n = 270 for *M. scabrinodis*).
Table 5. Results of Conover-Iman tests on the species differences of body parts in absolute thallus density. The column “Largest thallus density” shows which host species had highest number of thalli on a given body part; species names in parentheses indicate that the difference between the specified species and one of the other two species is not significant; “none” means that the three species did not differ significantly from one another.

| Body part            | Comparisons of *Myrmica* spp. | t      | P (adjusted) | Largest thallus density |
|----------------------|-------------------------------|--------|--------------|-------------------------|
| Antennae             | *M. rubra* – *M. sabuleti*     | −5.28  | <0.001       | *M. sabuleti*           |
|                      | *M. rubra* – *M. scabrinodis*  | −1.56  | 1.000        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 5.78  | <0.001       |                         |
| Head (frontal)       | *M. rubra* – *M. sabuleti*     | −3.15  | 0.029        | (M. sabuleti)           |
|                      | *M. rubra* – *M. scabrinodis*  | −3.09  | 0.034        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 0.90  | 1.000        |                         |
| Head (ventral)       | *M. rubra* – *M. sabuleti*     | −0.52  | 1.000        | (M. sabuleti)           |
|                      | *M. rubra* – *M. scabrinodis*  | 2.26   | 0.254        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 3.43  | 0.012        |                         |
| Pronotum             | *M. rubra* – *M. sabuleti*     | −4.54  | <0.001       | *M. sabuleti*           |
|                      | *M. rubra* – *M. scabrinodis*  | −2.96  | 0.048        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 3.06  | 0.036        |                         |
| Mesonotum            | *M. rubra* – *M. sabuleti*     | −4.64  | <0.001       | (M. sabuleti)           |
|                      | *M. rubra* – *M. scabrinodis*  | −3.48  | 0.011        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 2.59  | 0.127        |                         |
| Propodeum            | *M. rubra* – *M. sabuleti*     | −2.65  | 0.110        | None                    |
|                      | *M. rubra* – *M. scabrinodis*  | −2.80  | 0.076        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 0.52  | 1.000        |                         |
| Petiole              | *M. rubra* – *M. sabuleti*     | −3.29  | 0.018        | (M. scabrinodis)        |
|                      | *M. rubra* – *M. scabrinodis*  | −4.05  | 0.001        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | −0.03  | 1.000        |                         |
| Postpetiole          | *M. rubra* – *M. sabuleti*     | −2.46  | 0.173        | None                    |
|                      | *M. rubra* – *M. scabrinodis*  | −2.30  | 0.242        |                         |
| Gaster tergites      | *M. rubra* – *M. sabuleti*     | 0.83   | 1.000        |                         |
|                      | *M. rubra* – *M. scabrinodis*  | −2.35  | 0.224        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 4.17  | 0.001        |                         |
| Gaster sternites     | *M. rubra* – *M. sabuleti*     | 3.48   | 0.011        | (M. rubra)              |
|                      | *M. rubra* – *M. scabrinodis*  | 0.07   | 1.000        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | −4.95  | <0.001       |                         |
| Procoxa              | *M. rubra* – *M. sabuleti*     | 3.39   | 0.014        | *M. rubra*              |
|                      | *M. rubra* – *M. scabrinodis*  | 4.74   | <0.001       |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 0.70  | 1.000        |                         |
| Profemur             | *M. rubra* – *M. sabuleti*     | 1.22   | 1.000        | None                    |
|                      | *M. rubra* – *M. scabrinodis*  | 0.60   | 1.000        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | −1.06 | 1.000        |                         |
| Mesofemur            | *M. rubra* – *M. sabuleti*     | 2.67   | 0.107        | (M. rubra)              |
|                      | *M. rubra* – *M. scabrinodis*  | 3.80   | 0.003        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 0.64  | 1.000        |                         |
| Mesofemur            | *M. rubra* – *M. sabuleti*     | 0.25   | 1.000        | None                    |
|                      | *M. rubra* – *M. scabrinodis*  | 1.53   | 1.000        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | 1.44  | 1.000        |                         |
| Metacoxa             | *M. rubra* – *M. sabuleti*     | 3.30   | 0.018        | *M. rubra*              |
|                      | *M. rubra* – *M. scabrinodis*  | 3.79   | 0.003        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | −0.29 | 1.000        |                         |
| Metafemur            | *M. rubra* – *M. sabuleti*     | 1.48   | 1.000        | None                    |
|                      | *M. rubra* – *M. scabrinodis*  | 1.28   | 1.000        |                         |
|                      | *M. sabuleti* – *M. scabrinodis* | −0.63 | 1.000        |                         |

*M. rubra* (one isolate), *M. sabuleti* (two isolates), and *M. scabrinodis* (two isolates). The concatenated ITS + 28S dataset comprised 804 characters, of which 604 were constant and 183 were parsimony-informative. A total of 16 isolates were included. Once again, *R. wasmannii* was retrieved as a monophyletic clade with maximum support from MP and ML (Fig. 3).

Results of the species delimitation methods are summarised in Tables 2 and 3 and Figure 3. The number of putative species of *Rickia* was three in the ITS dataset with ABGD: *Rickia laboulbenioides* + *uncigeri*, *R. pachyiuli*, and *R. wasmannii*. In the 28S dataset, this number varied from three to seven, depending on the prior intraspecific divergence parameter, whereas other parameters (relative gap width, distance metrics employed) had no influence on the results (Table 3). The bPTP analysis of both ITS and 28S topologies resulted in four highly supported species: *Rickia laboulbenioides*, *R. pachyiuli*, *R. wasmannii*, and *Rickia uncigeri*. Additionally, the resolution of the relationships among these species was high in the 28S dataset.
Figure 5. Visualisation of infection patterns based on absolute thallus number on 16 body parts, using t-SNE to reduce the number of dimensions of the data set (n = 354).

R. uncigeri, and R. wasmannii. The GMYC analysis of the ITS resulted in the recognition of four species, all with moderate to high support except for R. uncigeri (of which only a single isolate was included). The GMYC analysis of the 28S led to comparable results, but in this analysis also the R. wasmannii clade received low support (pp = 0.38).

Absolute and relative thallus numbers

There were no significant differences between Myrmica species in total thallus number (Table 4, Fig. 4). In the Conover-Iman tests comparing absolute thallus number of each body part between species, we found 20 significant differences after Bonferroni’s P-value adjustment (Table 5). Overall, M. sabuleti specimens were more heavily infected on their antennae, head, pronotum, and mesonotum compared to workers of M. rubra and M. scabrinodis, whereas M. scabrinodis ants appeared to have the highest thallus density on their petiole and gaster tergites in comparison to the other host species. Myrmica rubra workers showed highest thallus density on their gaster sternites, procoxa, mesocoxa, and metacoxa (Fig. 5).

In the Conover-Iman tests comparing relative thallus number of each body part between species, we found 29 significant differences after Bonferroni’s P-value adjustment (Table 6). In comparison to M. rubra and M. scabrinodis, values of relative thallus number were highest in M. sabuleti on the antennae, head, pronotum, mesonotum, propodeum, and petiole. On the gaster tergites, M. scabrinodis had larger proportions of thalli compared to the other host species. Furthermore, M. rubra workers had highest proportions of thalli on their gaster sternites, procoxa, mesocoxa, mesofemur, metacoxa, and metafemur, compared to the other host species (Fig. 6).

Potential origin of infection on the body

The total number of thalli was significantly negatively associated with relative thallus number on the frontal side of the head in M. rubra (Estimate = −0.054, SE = 0.014, t = −3.81, P < 0.001) and in M. sabuleti (Estimate = −0.046, SE = 0.011, t = −3.97, P < 0.001). In the case of M. scabrinodis, the regression coefficients differed between ants from the Netherlands and Hungary: in the Netherlands, the association between total number of thalli and relative thallus number on the frontal side of the head was negative and relatively strong (Estimate = −0.076, SE = 0.029, t = −2.57, P = 0.011), whereas in Hungary, the regression coefficient was positive and weaker than in the other groups (Estimate = 0.013, SE = 0.006, t = 2.27, P = 0.024). These associations are shown in Figure 7.

In the model in which we fitted the relative thallus number of gaster tergites on total thallus number, the regression coefficient was significantly negative for M. scabrinodis ants collected in Hungary (Estimate = −0.042, SE = 0.005, t = −9.02, P < 0.001), but it was not significant in workers from the Netherlands (Estimate = 0.039, SE = 0.026, t = 1.52, P = 0.129). The association was not significant in M. rubra either (Estimate = −0.006, SE = 0.010, t = −2.57, P = 0.011). However, we found a significantly positive effect in M. sabuleti (Estimate = 0.024, SE = 0.011, t = 2.16, P = 0.031). These associations are shown in Figure 8.

Discussion

Rickia wasmannii was described in the 19th century, based on morphological characters only [9]. Myrmica hosts of R. wasmannii belong in two phylogenetically distinct species groups (rubra-group and scabrinodis-group). The genus Myrmica quickly diversified around the Eocene–Oligocene transition. The scabrinodis-group is among the oldest species groups (21.46 ± 4.00 Mya), whereas the estimated crown age for the rubra-group is 10.88 ± 2.12 Mya, in the Late Miocene [39].

Our results demonstrate that R. wasmannii does not encompass divergent genetic lineages segregated by host. In all molecular phylogenetic reconstructions, R. wasmannii isolates formed a monophyletic clade with maximum support. Infected workers of Myrmica spp. were collected in Austria, Belgium, Hungary, and the Netherlands. Even so, there is no geographic signal. The ITS sequences of R. wasmannii are all identical, whereas there are 0, 1, or 2 nucleotide differences among LSU sequences. We conclude that, contrary to species of Gloeomyces and Hesperomyces, in R. wasmannii neither geography nor host species are drivers of divergent evolution. The absence of host specificity in R. wasmannii is quite different from what has been observed in a Myrmica-associated group of endosymbiotic bacteria; Spiroplasma species co-diverged with their hosts over evolutionary time [1].

Most species of Laboulbeniales have been described based on morphological characters. In fact, only four species and four formae have been described based on combined morphological and molecular data [27, 34, 35]. In addition, for only a handful of species the taxonomic status has been assessed using molecular phylogenetic data following description. For example, Corethromyces bicolor Thaxt., after having been transferred to another genus, was re-installed in the genus Corethromyces Thaxt. based on DNA studies [76], and using sequence data from three loci, distinct clades within...
Table 6. Results of Conover-Iman tests on the species differences of body parts in relative thallus number. The column “Largest proportion of thalli” shows which host species had highest proportion of thalli on a given body part; species names in parentheses indicate that the difference between the specified species and one of the other two species is not significant; “none” means that the three species did not differ significantly from one another.

| Body part          | Comparisons of Myrmica spp. | t    | P (adjusted) | Largest proportion of thalli |
|--------------------|------------------------------|------|--------------|------------------------------|
| Antennae           | M. rubra – M. sabuleti      | −6.77| <0.001       | M. sabuleti                 |
|                    | M. rubra – M. scabrinodis   | −1.77| 0.542        |                              |
|                    | M. sabuleti – M. scabrinodis| 7.68 | <0.001       |                              |
| Head (frontal)     | M. rubra – M. sabuleti      | −3.59| 0.005        | M. sabuleti                 |
|                    | M. rubra – M. scabrinodis   | −1.68| 0.616        |                              |
|                    | M. sabuleti – M. scabrinodis| 3.20 | 0.017        |                              |
| Head (ventral)     | M. rubra – M. sabuleti      | −0.15| 1.000        | (M. sabuleti)               |
|                    | M. rubra – M. scabrinodis   | 4.49 | <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 5.53 | <0.001       |                              |
| Pronotum           | M. rubra – M. sabuleti      | −7.40| <0.001       | M. sabuleti                 |
|                    | M. rubra – M. scabrinodis   | −4.04| 0.001        |                              |
|                    | M. sabuleti – M. scabrinodis| 5.91 | <0.001       |                              |
| Mesonotum          | M. rubra – M. sabuleti      | −7.33| <0.001       | M. sabuleti                 |
|                    | M. rubra – M. scabrinodis   | −4.36| <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 5.43 | <0.001       |                              |
| Propodeum          | M. rubra – M. sabuleti      | −4.23| <0.001       | (M. sabuleti)               |
|                    | M. rubra – M. scabrinodis   | −3.29| 0.013        |                              |
|                    | M. sabuleti – M. scabrinodis| 2.22 | 0.231        |                              |
| Petiole            | M. rubra – M. sabuleti      | −4.03| 0.001        | M. sabuleti                 |
|                    | M. rubra – M. scabrinodis   | −4.94| <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 0.03 | 1.000        |                              |
| Postpetiole        | M. rubra – M. sabuleti      | −2.00| 0.368        | None                        |
|                    | M. rubra – M. scabrinodis   | −1.32| 0.934        |                              |
| Gaster tergites    | M. rubra – M. sabuleti      | 3.73 | 0.003        | (M. scabrinodis)            |
|                    | M. rubra – M. scabrinodis   | −2.38| 0.160        |                              |
|                    | M. sabuleti – M. scabrinodis| −8.20| <0.001       |                              |
| Gaster sternites   | M. rubra – M. sabuleti      | 5.26 | <0.001       | (M. rubra)                  |
|                    | M. rubra – M. scabrinodis   | 0.58 | 1.000        |                              |
|                    | M. sabuleti – M. scabrinodis| −6.90| <0.001       |                              |
| Procoxa            | M. rubra – M. sabuleti      | 5.11 | <0.001       | M. rubra                    |
|                    | M. rubra – M. scabrinodis   | 7.41 | <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 1.38 | 0.934        |                              |
| Profemur           | M. rubra – M. sabuleti      | 1.85 | 0.489        | None                        |
|                    | M. rubra – M. scabrinodis   | 1.64 | 0.616        |                              |
|                    | M. sabuleti – M. scabrinodis| −0.73| 1.000        |                              |
| Mesofemur          | M. rubra – M. sabuleti      | 3.49 | 0.007        | M. rubra                    |
|                    | M. rubra – M. scabrinodis   | 5.00 | <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 0.88 | 1.000        |                              |
| Metacoxa           | M. rubra – M. sabuleti      | 4.18 | 0.001        | M. rubra                    |
|                    | M. rubra – M. scabrinodis   | 5.34 | <0.001       |                              |
|                    | M. sabuleti – M. scabrinodis| 0.27 | 1.000        |                              |
| Metacoxa           | M. rubra – M. sabuleti      | −4.07| 0.025        | M. rubra                    |
|                    | M. rubra – M. scabrinodis   | 3.17 | 0.018        |                              |
|                    | M. sabuleti – M. scabrinodis| −0.68| 1.000        |                              |

*Hesperomyces virescens* were found, each corresponding to host-specific species [30].

Species in the genus *Coreomyces* Thaxt. do not show host specificity – similar to *R. wasmannii. Coreomyces corixae* (green clade in [68]), for example, occurs on water boatmen (*Heteroptera, Corixidae*) in the genera *Callicorixa* White 1873, *Hesperocorixa* Kirkaldy 1908, and *Sigara* Fabricius, 1775 [67]. As more examples of Laboulbeniales fungi are explored, we can start linking speciation patterns to presence and absence of fungal traits. One candidate trait that may have an influence in host-dependent speciation in Laboulbeniales is the presence of a haustorium. Haustoria are rhizoidal structures that can be simple
or branched and penetrate the host’s integument to provide additional holdfast and to increase surface area, presumably for nutrient uptake. Benjamin [6] believed that all Laboulbeniales produce haustoria. This is contrary to Tragust et al. [7] who, based on light and electron microscopy techniques, found no evidence for penetration in four species of Laboulbeniales: Laboulbenia camponoti S.W.T. Batra, L. formicarum Thaxt., Rickia lenoirii Santam., and R. wasmanni.

A note about millipede-associated Laboulbeniales

The sequences of R. laboulbenoides, R. pachyiuli, and R. uncigeri were generated for this study and are the first published ones for millipede-associated Laboulbeniales. Laboulbeniales on millipedes occur in five genera: Diplopodomyces W. Rossi & Balazuc, Rickia, the recently described Thaxterimyces Santam., Reboleira & Enghoff, Triainomyces
Similar to our findings with bat fly-associated Laboulbeniales fungi [33], we expect that parasitism of millipedes by Laboulbeniales arose several times independently. Some species of Rickia on millipedes are known to parasitise several millipede hosts. For example, *R. candelabriformis* Santam., Enghoff &
Reboleira, R. gigas Santam., Enghoff & Reboleira, and R. lophophora Santam., Enghoff & Reboleira [60] are potential next targets to study intraspecific diversity, to assess whether our current observations for R. wasmannii hold for the entire genus.

**Species delimitation analyses**

The ABGD analysis of the 28S dataset found different numbers of putative species depending on the prior intraspecific divergence (P), which is in line with previous work. Puillandre et al. [55] put forward to use $P = 0.01$, because under this setting, ABGD results in the same number of putative species found using different approaches. In our analyses of both the ITS and 28S datasets, ABGD found three species under this setting: R. laboulbenioides + uncigeri, R. pachyiuli, and R. wasmannii. Checking the distance matrices, we found that the lowest number of inter-species nucleotide differences was observed between R. laboulbenioides and R. uncigeri (Tables 7 and 8). For example, in the ITS dataset, R. laboulbenioides differed in six nucleotides from R. uncigeri, whereas it differed in 16 nucleotides from R. pachyiuli and in 24 nucleotides from R. wasmannii (details in Table 7). Apparently, ABGD was not able to identify the divergence among R. laboulbenioides and R. uncigeri isolates as a “barcode gap” (fide [55]), which will likely be resolved once we generate and include more sequences of R. uncigeri. The GMYC results are congruent with the results from the other species delimitation methods. One clade lacks support, the singleton clade R. uncigeri, and this is no surprise because GMYC looks at intraspecific branching versus interspecific branching.

The lack of phylogenetic structuring among R. wasmannii populations may be attributed to two different but not mutually exclusive scenarios: (1) intermittent gene flow homogenising populations and (2) recent spread of the fungus starting from a small founder population. The first scenario is possible because co-occurring arthropods may share Laboulbeniales parasites. Interspecific ascospore transmission in sympatric species has been observed for R. wasmannii parasitising M. scabrinodis, mites, and a Microdon myrmicae larva (Diptera, Syrphidae) in ant nests [53]. The second scenario can best be illustrated with the following example. Laboulbenia formicarum Thaxt. is thought to have spread from North America to Europe on an unknown ant host [23], followed by host shifts to European-native and invasive ant host species during its rapid spread in recent years [24, 28]. It was shown for M. rubra that it survived the last glacial period in multiple refugia and expanded its distribution along different routes [46]. It might be possible that R. wasmannii has undergone postglacial spread with its host, followed by multiple host shifts to other Myrmica species. Microsatellite studies are required to assess population-wide genetic differences, e.g., to answer the question whether incipient sympatric speciation is taking place.

**Habitat specificity and host spectrum**

Rickia wasmannii is a single species, clearly shared by a number of Myrmica hosts and with a vast distribution area. The species is non-penetrating [71] and compared to taxa with a haustorium such as H. virescens, it has several hosts but only if these occupy a similar habitat (Myrmica nests). This habitat specificity – preference for Myrmica nests and habitat choices – can explain the wide distribution on multiple Myrmica species and ant nest inquilines [53]. Moreover, the fact that there is overlap and even contact between Myrmica populations of different species [78] implies that regular or at least sufficient interspecific transfer of R. wasmannii occurs between host taxa. It also means that the different host taxa, their specific habitat choices, and the nature of their nests, allow the development of the fungus population. Considering the high thallus densities observed, we doubt there is enough reason to consider one Myrmica species as a main host (fide [62]) over other species. In this context, we propose that R. wasmannii is a true eurytopic species with a wide ecological amplitude. It is expected that other species of Myrmica may also carry this parasite. However, absence of R. wasmannii on a given Myrmica species does not necessarily mean that this ant species, its nests, and/or its habitat selection are unsuitable for this fungus. Indeed, in areas where several infected nests of M. scabrinodis occur, some adjacent nests can be entirely free of R. wasmannii [15]. This has also been observed for M. sabuleti, where infection frequency of workers can vary from 0 to 100% among nests that are only a few meters apart (P. Boer, unpublished data).

**Distribution of thalli on worker bodies**

The original morphological description [8] holds to the phylogenetic species concept. This implies that differences in thallus numbers of different body parts between ant species

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### Table 8. Distance matrix of the aligned 28S tRNA sequences.

| Isolate | Species | GenBank acc. no. | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 |
|---------|---------|------------------|---|---|---|---|---|---|---|---|---|---|---|
| 1 SR4s  | Rickia laboulbenioides | MH040592     | 0 |   |   |   |   |   |   |   |   |   |   |
| 2 SR5s  | Rickia laboulbenioides | MK500060     | 39| 39|   |   |   |   |   |   |   |   |   |
| 3 SR8s  | Rickia laboulbenioides | MK500058     | 39| 39| 39| 39|   |   |   |   |   |   |   |
| 4 SR1s  | Rickia pachyiuli       | MH040593     | 39| 39| 39| 39| 39| 39|   |   |   |   |   |
| 5 SR13s | Rickia pachyiuli       | MK500059     | 39| 39| 39| 39| 39| 39| 39| 39|   |   |   |
| 6 SR12s | Rickia uncigeri        | MK500061     | 39| 39| 39| 39| 39| 39| 39| 39| 39| 39| 39|
| 7 Ujle2014 | Rickia wasmannii      | MK490857     | 79| 79| 79| 79| 79| 79| 79| 79| 79| 79| 79|
| 8 DE_Rak4 | Rickia wasmannii      | KT800021     | 79| 79| 79| 79| 79| 79| 79| 79| 79| 79| 79|
| 9 Wien2015-1 | Rickia wasmannii | MK490858     | 80| 80| 80| 80| 80| 80| 80| 80| 80| 80| 80|
| 10 D. Haelw. 1234a | Rickia wasmannii | MH040595     | 79| 79| 79| 79| 79| 79| 79| 79| 79| 79| 79|
| 11 ADK5272a | Rickia wasmannii | MK500050     | 79| 79| 79| 79| 79| 79| 79| 79| 79| 79| 79|
and populations must be explained by behaviour, cuticular chemical profiles, and/or environmental stresses \cite{7, 20, 21, 65}. In our dataset, there was no evidence for differences between host species in total number of thalli on worker bodies. If the ant species in the study area are of the same body size, this might suggest that the overall number of thalli on a worker’s body is simply a factor of the worker’s age, irrespective of host species; older workers show heavier infection by \textit{R. wasmannii} \cite{3}.

We did observe differences between \textit{Myrmica} species in the pattern of infection over the body. Tests on both absolute and relative thallus number indicate that \textit{M. sabuleti} workers are more heavily infected on the first few body segments compared to other hosts (Tables 5 and 6). \textit{Myrmica rubra} workers show highest thallus densities on the coxa and femur, whereas in \textit{M. scabrinodis} highest thallus densities were found on the gaster tergites. These results might indicate differences among host species in how ants come into contact with the fungus, or even differences in grooming efficacy. Which body parts are workers able to groom (and thus stop ascospores from adhering and developing) more effectively? Another possibility involves differences in the cuticle itself \cite{20, 65}; surface characteristics may have a fundamental impact on the success of an ascospore to adhere to the cuticle and develop to a mature thallus.

Based on our statistical analyses, it is likely that in \textit{M. rubra} and \textit{M. sabuleti} the locus of infection (the area where the infection originates) is the frontal side of the head. For \textit{M. scabrinodis} workers from the Netherlands, the locus of infection also appears to be the frontal side of the head. However, for \textit{M. scabrinodis} specimens from Hungary, infection likely starts from the gaster tergites. These results indicate differences among populations of the same species in a wide geographical range. Different \textit{Myrmica} species display divergent foraging, allo-grooming, and secretion emission activities \cite{7}. This likely leads to differences in how workers enter into contact with ascospores, which should be investigated with behavioural studies of their hosts.

**Conflict of interest**

The authors declare that they have no conflicts of interest in relation to this article.

**Authors’ contributions**

D.H. coordinated the study, performed molecular phylogenetic and species delimitation analyses, and wrote the manuscript with input from all co-authors; P.B., F.B., A.T., and A.D.K. collected ants; P.B. and F.B. counted thalli on worker bodies; Z.R. performed statistical analyses; D.H., A.S.P.S.R., and W.P.P. extracted DNA and generated sequence data; A.D.K. provided the photograph for Figure 1 and the drawing included in Figure 3; A.D.K. and O.N. provided expertise at all stages of research.

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