Novel exostosin-2 mutation identified in a Chinese family with hereditary multiple osteochondroma

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Abstract. Hereditary multiple osteochondroma (HMO) is an autosomal dominant genetic disorder characterized by multiple outgrowing bony tumors capped by cartilage, generally affecting the metaphyses. The disease is known as hereditary multiple exostoses, familial exostosis, multiple cartilaginous exostoses or hereditary malformation of cartilage. The prevalence of HMO in Europe and the United States is ~1:100,000, although it has not been reported in China. The disease is often accompanied by pain, asymmetry and skeletal malformations, including forearm and leg bending deformities, limb length discrepancies, and knee internal and external rotation abnormalities. Mutations to exostosin-1 (EXT1) and EXT2 mutations cause insufficient heparan sulfate biosynthesis, leading to chondrocyte proliferation, abnormal bone growth in neighboring regions, multiple exostoses, and ultimately malignant transformation. The risk of malignant degeneration to osteochondrosarcoma increases with age, despite the low lifetime risk (~1%). The present study selected three generations of a single family with HMO and explored the potential underlying mechanism.

Introduction

Hereditary multiple osteochondroma (HMO), also known as hereditary multiple exostoses, is characterized by multiple exogenous exostoses in the skeletal system (1,2). The incidence of HMO in Europe is ~1:100,000 (3), and in the United States is ~1:50,000 (4); however, to the best of our knowledge, its incidence in China is presently unknown. It occurs more frequently in males than in females (~1.5:1). Symptoms in females are not serious and the disease is often not diagnosed (5,6). HMO is autosomal dominant (7) and has an penetrance of 100% (3,4). Approximately 70% of patients with HMO have a family history of the disorder (5,6). The disease affects regions of the humerus (10-50%), forearm (39-60%), knee (33%), ankle (25%), and other long bones; the scapula and ribs are affected, and the majority of osteochondroma is distributed symmetrically (4,10-12).

Studies concerning the etiology and molecular genetic background of HMO have indicated that the majority cases are associated with mutations in the exostin (EXT) tumor inhibition genes (13); specifically, EXT1, EXT2, and EXT3, which are located at 11p11-13, 8c23-24.1, and on chromosome 19p, respectively, are associated with HMO (14,15). EXT encodes a type II transmembrane protein that is closely associated with the synthesis of heparan sulfate (HS), a key component of cartilage (16). Mutations in the EXT gene affect glycosyltransferase activities, resulting in an HS synthesis barrier. Nonsense mutations, frameshift mutations and splice site mutations account for 80% of mutations in patients with HMO.

In the present study, the clinical data of members from three generations of a single family with HMO were collected and statistically analyzed. Using two known genes of HMO, EXT1 and EXT2, the genomic DNA of the family members was directly sequenced using a polymerase chain reaction (PCR), in order to identify the pathogenesis of the family with HMO and explore the potential underlying mechanism. The results of the present study may be useful for the clinical genetic counseling of similar diseases in the future.

Materials and methods

Patients and clinical data. A three-generation HMO family, JX-JJ001 (Jiangxi, China) were the subject of the present study (Fig. 1). Diagnostic criteria: X-ray examination revealed that the long bone metaphysis had at least two or more osteochondromas. According to the above criteria, it was confirmed that five patients (4 male, 1 female) in the family exhibited generation inheritance of HMO. All participants provided informed consent for clinical genetic counseling of similar diseases in the future.
Molecular analysis. The peripheral blood from the pedigree patients and the non-patient members was drawn and added to the EDTA anticoagulant tube. Next, the DNA was extracted from the blood using the AxyPrep Nucleic Acid Purification kit (Axygen Scientific, Inc., Union City, CA, USA) in accordance with the manufacturer's protocol. Following this, a Nanodrop2000 spectrophotometer (Nanodrop Technologies, Wilmington, DE, USA) was used to quantify DNA. Primer 5.0 (Premier Biosoft International, Palo Alto, CA, USA) was used to design primers and primer sequences (Table II) for all exons of EXT1 and EXT2 (including the 100 bp base of exon and intron junctions). PCR (KAPA2G Fast Multiplex Mix; Kapa Biosystems, Inc., Wilmington, MA, USA) was used for the expansion of genomic DNA, with thermocycling conditions as follows: 94°C for 5 min, followed by 94°C for 30 sec and 60°C for 30 sec, for 30 cycles each, followed by 72°C for 1 min, 72°C for 5 min and 12°C for 10 min. Amplified DNA product was subjected to Sanger sequencing analysis using the BigDye Terminator v3.1 Cycle Sequencing kit (Applied Biosystems; Thermo Fisher Scientific, Inc., Waltham, MA, USA) in accordance with the manufacturer's protocol. Following this, a Nanodrop2000 spectrophotometer (Nanodrop Technologies; Thermo Fisher Scientific, Inc., Wilmington, MA, USA) was used to quantify DNA. Primer 5.0 (Premier Biosoft International, Palo Alto, CA, USA) was used to design primers and primer sequences (Table II) for all exons of EXT1 and EXT2 (including the 100 bp base of exon and intron junctions). PCR (KAPA2G Fast Multiplex Mix; Kapa Biosystems, Inc., Wilmington, MA, USA) was used for the expansion of genomic DNA, with thermocycling conditions as follows: 94°C for 5 min, followed by 94°C for 30 sec and 60°C for 30 sec, for 30 cycles each, followed by 72°C for 1 min, 72°C for 5 min and 12°C for 10 min. Amplified DNA product was subjected to Sanger sequencing analysis using the BigDye Terminator v3.1 Cycle Sequencing kit (Applied Biosystems; Thermo Fisher Scientific, Inc., Waltham, MA, USA) and an ABI 3730XL automatic sequencer (Applied Biosystems; Thermo Fisher Scientific, Inc.). Samples from the patients and healthy controls were bi-directionally sequenced and the sequencing results were analyzed using Sequencer Demo 3.0 (Gene Codes Corporation, Ann Arbor, MI, USA) and Mutation Surveyor Demo V4.0 software (SoftGenetics, LLC, State College, PA, USA), with National Centre for Biotechnology Information (EXT1, NM_000127.2; EXT2, NM_001178083.1; https://www.ncbi.nlm.nih.gov/) sequences used as reference genes.

Results

DNA sequencing identified a novel mutation to the EXT2 gene in all patients with HMO. The tissue structure (Fig. 2) of patients with HMO was characterized by a cartilage cap that was covered by a fibrous perichondrium and merged into an underlying spongy bone. Each exon of the EXT1 and EXT2 genes was sequenced in 5 patients (I1, I2, I3, I11, and I12), as shown in Fig. 1) and 3 normal members (I2, I1 and I44, as shown in Fig. 1) of the HMO family JX-JJ001. Following the data filtering procedures, all of the HMO patients were found to have mutations in the EXT2 gene. A novel nonsense mutation was identified on the EXT2 gene in the five HMO patients: c.676C>T (amino acid R23*), (Fig. 3), it is worth noting that this novel nonsense 23R>X mutation occurred only in the five HMO patients and was not found in other family members. Therefore, we hypothesize that this novel EXT2 mutation site may be a pathogenic mutation site. According to this finding, the DNA of 300 healthy volunteers who were not associated with the family members was sequenced, and it was found that none of the EXT2 genes in the 300 healthy volunteers had a 23R>X mutation. Thus, this novel nonsense mutation 23R>X in the EXT2 gene is leading to pathogenicity of the HMO family JX-JJ001.

23R>X mutation had a notable effect on the function of the EXT2 gene. To understand the effect of 23R>X on the function of the EXT2 gene, further computer simulation analysis was performed. The protein encoded by the EXT2
The gene was composed of 728 amino acids and had two notable domains: the exostosin domain (between the 101 and 380 amino acids) and the glycosyltransferase domain (between the 466 and 711 amino acids) (Fig. 3B). Amino acid 23 (Arg) was present before the exostosin and glycosyltransferase domains. Furthermore, evolutionary conservation analysis revealed that the amino acids were highly conservative in multiple species (Fig. 4). This result indicates that this amino acid serves a notable role in the function of the EXT2 gene and remains in the process of biological evolution. This nonsense mutation 23R>X causes the EXT2 transcription process to be terminated prematurely and produces a truncated EXT2 protein, lacking the C-terminal 705 amino acids (Fig. 3B). Since the glycosyltransferase domain serves a notable role in HS biosynthesis, the mutation 23R>X is likely to have a significant effect on the function of the EXT2 gene, particularly affecting the biosynthesis of HS. The same result was also observed with the online prediction tool Mutation Taster, which revealed that using stop codon to replace 23 arginine affects the function of the EXT2 protein and leads to the occurrence of the disease (probability score, 6.0) (Fig. 5).

Discussion

Although HMO is observed in Europe and the United States at an incidence of 1:50,000-100,000, to the best of our knowledge its incidence in China is not known (3,4). Clinical characteristics often vary among individuals: Patients typically have symmetrical bony protrusions with cartilage caps in long bone growth plates, as well as short, flat and/or irregular bones; certain patients have disproportionate limbs and short stature (20). The onset age for half of all patients is <3 years old. During growth and development, exostoses grow gradually; the growth plates eventually disappear and exostoses stop growing until puberty (20). The clinical symptoms and complications of HMO are associated with the location and size of exostoses; these symptoms include pain, short stature, skeletal deformities, oppression around the blood vessels and nerves.

Table I. Clinical data for five hereditary multiple osteochondroma patients.

| Patient | Sex | Height, cm | Age of onset, years | Age at time of study, years | Localization |
|---------|-----|------------|---------------------|-----------------------------|--------------|
| 1       | Male | 172.5      | 75                  | 75                          | Femurs, tibia, fibula and knee joints |
| 2       | Female | 157.4      | 37                  | 45                          | Femurs, tibia, fibula and knee joints |
| 3       | Male | 159.2      | 38                  | 42                          | Femurs, tibia, fibula and knee joints |
| 4       | Male | 160.5      | 7                   | 15                          | Femurs, tibia, fibula and knee joints |
| 5       | Male | 129.4      | 6                   | 10                          | Femurs, tibia, fibula and knee joints |

Table II. Primer sequences used to sequence the EXT2 genes.

| Primers | Forward | Reverse |
|---------|---------|---------|
| EXT2-E1 | 5'-ATTGCCCTCCAGGAAATGTTA-3' | 5'-GCAGGAGTGGAAATCGGAG-3' |
| EXT2-E2 | 5'-GGCGTGTGTTGCCACAGTTAC-3' | 5'-ACCACTCAAGAGCAGAAAGCA-3' |
| EXT2-E3 | 5'-CTGTGTGGAGTTCCAGGAGTTT-3' | 5'-TGCCAGGACATAAGCCCTAACT-3' |
| EXT2-E4 | 5'-TGTACCGAGAGGATAAGCAG-3' | 5'-AAACGAGGAGAAGCAGAACTC-3' |
| EXT2-E5 | 5'-GGCGGCTAGACGTGTAAGG-3' | 5'-CACAAGACACCAGACACATCCAAG-3' |
| EXT2-E6 | 5'-GGCGTCAACCTCCCTGAAAC-3' | 5'-CCTTGGTTTGTGAACCTGCTCT-3' |
| EXT2-E7 | 5'-GAAGGAGGTTTGGGATGTTTGT-3' | 5'-AAAGTACCCCCACCTAGGACATT-3' |
| EXT2-E8 | 5'-AAAGTGTGCCTGTTGAGTG-3' | 5'-ACTGCTGAAACCCTCTGTTG-3' |
| EXT2-E9 | 5'-TAAAGGAAATTAGCCTACCTGGAG-3' | 5'-CCCAAGTTAAAGACACTCTC-3' |
| EXT2-E10 | 5'-GTAAGAAGCCACCAAGCCTGC-3' | 5'-GTATGGCCAGGCTTGAGATT-3' |
| EXT2-E11 | 5'-CTTTGGAATTGATGAGACCG-3' | 5'-CCCACACTCCACAGCACCCT-3' |
| EXT2-E12 | 5'-TCTTACAGAATCCCCATTAGACTCT-3' | 5'-ATGAGTATCTCGAAAGTGGAGA-3' |
| EXT2-E13 | 5'-GCCCTCTTTTATCCTTCTATT-3' | 5'-GACCGCATCATACTAGAACCCT-3' |
| EXT2-E14 | 5'-CTTTGAGTATCTGCGGTG-3' | 5'-GCCCTTCCAGGACATTACAA-3' |
| EXT2-E14-CX | 5'-ATGGAGGTGACTATGGCTAC-3' | 5'-AGGAATTGCTGTATTGCTTCTCTC-3' |
| EXT2-E15-1 | 5'-TTTGCTGTATCTCCTCAACCTCT-3' | 5'-AGGAATTGCTGTATTGCTTCTC-3' |
| EXT2-E15-2 | 5'-TACATCAATGAGGTCTTCTTTCAGGGA-3' | 5'-ACGCTGACTGGCACAACAACTA-3' |
| EXT2-E15-CX | 5'-AATCTTGCTGTAATCTTACCTAATA-3' | 5'-AATCTTGCTGTAATCTTACCTAATA-3' |

EXT2-E1, exostosin-2 exon 1.
spontaneous pneumothorax and changes in joint mobility (4). The most serious complication is malignant chondrosarcoma or osteosarcoma, and the malignant transformation rate is 0.5-5% (12).

Human HMO is associated with at least two genes: \(\text{EXT1}\) on 8q24.11-q24.13 and \(\text{EXT2}\) on 11p12-11 (21,22). These two tumor suppressor genes are widely expressed; \(\text{EXT1}\) contains 11 exons and 10 introns and encodes 746 amino acids, whereas \(\text{EXT2}\) contains 15 exons and 15 introns, with exons 2-14 form an open reading frame encoding 728 amino acids. The two genes are highly similar at the amino acid level, particularly in the C-terminal region. According to Human Mutation Database statistics, 595 HMO-associated mutations have been detected to date, including 401 mutations in \(\text{EXT1}\) and 194 mutations in \(\text{EXT2}\); these include missense/nonsense mutations, small insertion/deletion mutations, splice site changes and large deletions (23). Although, to the best of our knowledge, the precise functions of these two genes is unknown, the proteins are localized in the type II transmembrane glycoproteins of the endoplasmic reticulum, which is associated with the synthesis of HS proteoglycan (HSPG) (17).

HSPG is composed of a core protein and an attached glycosaminoglycan chain (24). The proteins encoded by \(\text{EXT1}\) and \(\text{EXT2}\) form a heterologous oligomer in the Golgi apparatus of the majority of human cells (25). The heterologous oligomer, a glycosyltransferase complex, is involved in the polymerization of HS chains (26). The oligomer can add glucuronic acid and N-acetylglucosamine residues to HS chains alternatingly, forming HSPGs (27). The activity of the glycosyltransferase complex formed by the \(\text{EXT1}/\text{EXT2}\) oligomer is higher compared with that of \(\text{EXT1}\) or \(\text{EXT2}\) alone (28,29).

HS is widely expressed in cell membranes and the extracellular
matrix serves a notable role in receptor ligand binding, signal transduction and other physiological processes (30). HS proteoglycans are essential in several signaling pathways, including those involving fibroblast growth factors (FGFs), vascular endothelial growth factor and transforming growth factor-β, and may affect the concentration gradient of various morphogens on the cell surface, including bone morphogenetic proteins and hedgehog (31). HS proteoglycans can bind hedgehog ligands to control the spread of the hedgehog ligand in the extracellular matrix (32).

Hedgehog is a secretory protein that hydrolyzes itself, and the hedgehog signaling pathway serves a notable role in the regulation of growth plate chondrocytes (33-35). Hypertrophic chondrocytes of the growth plate may secrete Indian hedgehog (Ihh) (36,37). Ihh functions with parathyroid hormone-related protein (PTHrP) and regulates endochondral bone formation.
via a negative feedback loop (38). Ihh can induce periarticular hypertrophic chondrocytes to increase the secretion of PTHrP, which can suppress the proliferation of chondrocytes into hypertrophic chondrocytes (18). Ihh can also individually control chondrocyte differentiation and induce periosteal osteogenesis (39). In the regulation of endochondral bone growth via Ihh-PTHrP, multiple factors serve notable roles, including bone morphogenetic proteins (BMPs), which promote Ihh expression in cartilage cells (40).

Mutations in EXT1 and EXT2 can lead to the expression of truncated proteins that are associated with decreased activity of the glycosyltransferase complex (17). Synthesis of the HS chain is then reduced, which leads to the formation of incomplete HS proteoglycans (26,27). The incomplete HS proteoglycans cause ligand-receptor binding disorders in a number of signaling pathways, including Ihh, BMPs, FGF, and Wnt, which affects the concentration gradient of ligands on the cell surface or in the extracellular matrix (30,31). Disorders in the Ihh signaling pathway cause Ihh secretion and increase secretion of PTHrP by periarticular hypertrophic chondrocytes (18). This influences the proliferative potential of chondrocytes into hypertrophic chondrocytes and causes chondrocytes to undergo a long proliferative period, and ultimately HMO (41-44). Additionally, abnormal HS proteoglycans may change the morphology and differentiation potential of perichondrocytes, resulting in perichondrial cells that function as growth-plate-like cells and give rise to chondrocytes that clonally expand and develop into an exostosis, leading to HMO (25).

A lower disease burden is observed in individuals with EXT2 mutations than in those with EXT1 mutations (15). In a study examining 172 individuals from 78 families, Porter et al (12) verified a higher severe disease frequency in persons with mutations in EXT1 than in those with EXT2 mutations, as evidenced by shortened stature, skeletal deformities (shortened forearm or bowing and knee deformities), and function (decreased range of motion in the elbow, forearm and/or knee) (13). Individuals with EXT2 mutations have fewer exostoses, a lower incidence of limb misalignment with longer limb segments and height, and less frequent pelvic and flat bone involvement than those with EXT1 mutations (45).

The present study reveals a novel heterozygous nonsense mutation (c.67C>T) in EXT2, using whole-exome sequencing. The mutation was validated by Sanger sequencing and its negative influence on the expression of EXT2 was examined via in silico analysis. The results of the current study provide information regarding the association between the EXT gene family and HMO. These findings may facilitate early diagnosis and prenatal genetic screening of HMO.

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Competing interests

The authors declare that they have no competing interests.

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