The MITF(E318K) variant confers moderate risk for cutaneous melanoma. While there are small studies suggesting that this risk is associated with other malignancies (e.g. renal cell carcinoma), little is known about the role of this variant in specifying risk for other cancers. In this study, we perform a systematic review and meta-analysis of the published data as a backdrop to a whole-exome sequence (WES)-based characterization of MITF(E318K) risk for various cancers in sporadic samples from the TCGA and several genetically-enriched patient cohorts. We found minimal evidence of MITF(E318K)'s contribution to non-melanoma cancer risk among individuals with low inherited risks of melanoma (OR 1.168; 95% CI 0.78–1.74; p = 0.454), suggesting that earlier reports of an association between this variant and other malignancies may be related to shared environmental or polygenic risk factors rather than MITF(E318K). Interestingly, an association was observed with uterine carcinosarcoma, (OR 9.24; 95% CI 2.08–37.17; p = 0.024), which was not previously described. While more research needs to be completed, this study will help update cancer screening recommendations for patients with the MITF(E318K) variant.

The microphthalmia-associated transcription factor (MITF) is a known master regulator of melanocyte development with functions ranging from pigment production to differentiation, survival, and cell-cycle production of melanocytes. A rare functional variant of MITF(E318K) has been shown to disrupt a conserved SUMOylation site and to confer a two- to fourfold risk for cutaneous melanoma. Moreover, there have been small studies suggesting that this variant is also associated with other cancers such as renal cell carcinoma (RCC). However, since epidemiologic observations have documented an association between cutaneous melanoma and many other malignancies, including RCC, it is possible that MITF(E318K) represents a germline “passenger” mutation in these other cancers without directly impacting the risk of these other cancers such as RCC. In this study we perform a systematic review and meta-analysis of the published data as a backdrop to a whole-exome sequence (WES)-based characterization of MITF(E318K) risk for various cancers in sporadic populations from the TCGA database and genetically-enriched patient populations.

Results
Meta-analysis of the data compiled from nine published studies (n = 331) demonstrated that the variant was significantly correlated with melanoma (Fig. 1; odds ratio (OR) 2.37; 95% confidence interval (CI) 1.89–2.97; p < 1E−5; I2=19%). Studies were conducted on a mixture of hereditary and sporadic melanoma populations. A few studies identified a relationship of this variant with renal cell carcinoma (RCC), pancreatic cancer, and pheochromocytoma/paraganglioma. Bertolotto et al. identified over a fivefold increased risk for carriers to develop RCC, melanoma, or both cancers in 829 patients (OR 5.55; 95% CI 2.59–12.91; p = 1.2E−6) and identified a fivefold risk of developing RCC only in 164 “genetically-enriched” patients who were wild-type for RCC-predisposing genes (OR 5.19; 95% CI 1.37–16.87; p = 0.008). In contrast, other studies did not find an association with sporadic RCC. Ghiorzo et al. identified a 31-fold increased risk of melanoma in carriers with a personal or family history of pancreatic cancer (OR 30.85; 95% CI 6.85–138.9; p = 0.0005), but did not find significant enrichment of the variant in sporadic pancreatic cancer patients (0/210 patients had the variant).
Castro-Vega et al. screened 555 unrelated pheochromocytoma/paraganglioma patients, and found that the variant was enriched compared to controls (OR 3.19; 95% CI 1.34–7.59; p = 0.005)11.

To better understand the risks of this rare functional polymorphism on other cancers, we then systematically evaluated prevalence of the MITF(E318K) variant in multiple cancer cohorts using germline whole-exome sequence data from the TCGA panel and from several genetically-enriched cohorts to validate these associations (Table 1). The TCGA cohort consists of patients mostly with late-onset cancer and unascertained family history, and therefore their risk of comorbid cancers is theoretically low. This set up enables evaluation of primary cancer risk conferred by MITF(E318K). Since the variant is mainly found in the European population (minor allele frequency (MAF) in gnomAD14 = 0.00245 for European; MAF = 6.01E−04 for African; MAF = 0 for Asian), we performed the analysis using only European patients identified through principal component analysis (PCA).

Table 1. Relationship of MITF(E318K) with different cancer types in TCGA and genetically-enriched patient cohorts. Cancers whose lower boundary of 95% CI is greater than 1.0 are highlighted in Italics.

| Cohort                      | # MITF(E318K) carriers | N total samples | MAF       | Fisher OR (95% CI) | p-value |
|-----------------------------|------------------------|-----------------|-----------|--------------------|---------|
| Control                     | 63                     | 11,106          | 0.00284   | Ref                | Ref     |
| TCGA thyroid carcinoma      | 1                      | 363             | 0.00318   | 0.48 (0.05–2.57)   | 0.724   |
| TCGA glioblastoma           | 1                      | 326             | 0.00153   | 0.54 (0.05–2.86)   | 1.000   |
| TCGA breast carcinoma       | 3                      | 776             | 0.00193   | 0.68 (0.22–1.96)   | 0.800   |
| TCGA lung adenocarcinoma    | 2                      | 501             | 0.00200   | 0.70 (0.17–2.63)   | 1.000   |
| TCGA low grade glioma       | 2                      | 447             | 0.00224   | 0.79 (0.19–2.96)   | 1.000   |
| TCGA bladder carcinoma      | 1                      | 209             | 0.00239   | 0.84 (0.08–4.50)   | 1.000   |
| TCGA lung SCC               | 2                      | 391             | 0.00256   | 0.90 (0.22–3.39)   | 1.000   |
| TCGA/enriched breast cancer | 6                      | 976             | 0.00307   | 1.08 (0.50–2.39)   | 0.823   |
| TCGA prostate adenocarcinoma| 1                      | 226             | 0.00221   | 1.13 (0.11–6.55)   | 0.597   |
| TCGA pheochromocytoma/paraganglioma | 1      | 145             | 0.00345   | 1.22 (0.12–6.55)   | 0.565   |
| TCGA liver carcinoma        | 1                      | 125             | 0.00400   | 1.41 (0.14–7.64)   | 0.512   |
| TCGA kidney/papillary clear cell carcinoma | 1 | 123             | 0.00407   | 1.44 (0.14–7.77)   | 0.507   |
| TCGA cervical SCC/endocervical adenocarcinoma | 1 | 114             | 0.00439   | 1.51 (0.14–8.24)   | 0.496   |
| TCGA pancreatic adenocarcinoma | 1              | 116             | 0.00431   | 1.52 (0.15–8.26)   | 0.487   |
| TCGA rectum adenocarcinoma | 1                      | 113             | 0.00442   | 1.57 (0.15–8.49)   | 0.478   |
| TCGA head/neck SCC          | 4                      | 436             | 0.00459   | 1.62 (0.63–4.23)   | 0.322   |
| TCGA sarcoma                | 1                      | 102             | 0.00490   | 1.74 (0.17–9.44)   | 0.444   |
| TCGA colon adenocarcinoma   | 3                      | 300             | 0.00500   | 1.77 (0.58–5.15)   | 0.251   |
| TCGA uterine corpus endometrial carcinoma | 4 | 385             | 0.00519   | 1.79 (0.66–4.83)   | 0.295   |
| MGH genetically-enriched cutaneous melanoma (GECM) | 3 | 250             | 0.00600   | 2.13 (0.69–6.22)   | 0.178   |
| TCGA cutaneous melanoma     | 5                      | 409             | 0.00617   | 2.17 (0.93–5.29)   | 0.094   |
| TCGA cutaneous melanoma + MGH GECM | 8    | 659             | 0.00607   | 2.15 (1.03–4.37)   | 0.061   |
| MGH genetically-enriched breast cancer | 3 | 200             | 0.00750   | 2.67 (0.87–8.33)   | 0.111   |
| TCGA ocular melanoma        | 1                      | 47              | 0.01064   | 3.81 (0.37–21.69)  | 0.237   |
| TCGA uterine carcinoma sarcoma | 2 | 39              | 0.02564   | 9.24 (2.08–37.17)  | 0.024   |

To better understand the risks of this rare functional polymorphism on other cancers, we then systematically evaluated prevalence of the MITF(E318K) variant in multiple cancer cohorts using germline whole-exome sequence data from the TCGA panel and from several genetically-enriched cohorts to validate these associations (Table 1). The TCGA cohort consists of patients mostly with late-onset cancer and unascertained family history, and therefore their risk of comorbid cancers is theoretically low. This set up enables evaluation of primary cancer risk conferred by MITF(E318K). Since the variant is mainly found in the European population (minor allele frequency (MAF) in gnomAD14 = 0.00245 for European; MAF = 6.01E–04 for African; MAF = 0 for Asian), we performed the analysis using only European patients identified through principal component analysis (PCA). We
used a set of common (MAF > 0.01) LD-pruned variants to perform PCA and used k-means to identify European cluster of samples. The final dataset contained European 6143 cancer cases and 11,106 European non-cancer controls. Comparison of MITF(E318K) frequency in the controls sample (MAF = 0.28%; Table 1) matched well with the frequency expectation for Europeans (MAF in gnomAD Europeans = 0.25%), suggesting that our control cohort provides a reference point for MITF free from ancestral bias.

The risk of developing any cancer in aggregate with the MITF(E318K) mutation was slightly increased, but this did not reach significance (OR 1.294; 95% CI 0.88–1.88; p = 0.19). Similarly, the risk for all non-melanoma cancers was also increased but not significantly (OR 1.168; 95% CI 0.78–1.74; p = 0.454). Among the 25 cancers tested, uterine carcinomas (OR 9.24; 95% CI 2.08–37.17; p = 0.024) and melanomas (OR 2.15; 95% CI 1.03–4.37; p = 0.061) exhibited the strongest associations with the variant. We did not find any significant association of MITF(E318K) with renal cell carcinoma (RCC) (p = 0.5068), pancreatic cancer (p = 0.487), or pheochromocytoma/paraganglioma (p = 0.565) in the TCGA cohort.

Further, we tested whether differences in ascertainment between cancer cohorts could explain the lack of the previously reported association with RCC. Specifically, it is expected that the burden of germline risk variants should be lower in cohorts with unascertained family history (TCGA) than in genetically-enriched samples. We compared allele frequencies of MITF (E318K) for the familial (MAF = 0.6%) and sporadic (MAF = 0.6%) melanoma cohorts and did not observe any difference, suggesting that the sporadic cohort should be powerful enough to detect primary cancer risks associated with MITF(E318K).

Therefore, in our analysis of RCC, pancreatic cancer, pheochromocytoma/paraganglioma cohorts, there does not appear to be a strong risk of primary cancer conferred by MITF(E318K). Importantly, TCGA non-melanoma cohorts are unlikely to have substantial inherited risk of cutaneous melanoma due to the relatively high age of participants and no ascertainment, resulting in the lack of an association signal for MITF(E318K). Previous studies reporting the non-melanoma cancer association of MITF(E318K) have not yet assessed inherited melanoma risks for the RCC and other cancer cohorts.

**Discussion**

This study is an important addition to the published literature as it represents the largest MITF(E318K) variant analysis to date with more samples collectively between cases and controls than found in previously published studies. In addition, with WES, we utilize principal component analysis (PCA) to match for European ancestry which allows for a much cleaner calculation of risk compared to most other studies that relied solely on MITF(E318K) counts without regard for population mix. Although we corroborated the association of the variant with melanoma, we found minimal evidence of MITF(E318K)’s contribution to the risk of non-melanoma cancers among individuals with low inherited risks of melanoma.

These results suggest that earlier reports of an association between this variant and other malignancies may be related to shared environmental or polygenic risk factors rather than this specific MITF polymorphism. For example, multiple studies have shown an association between RCC and melanoma. In a 2018 analysis of the Surveillance, Epidemiology, and End Results (SEER) database, the Standardized Incidence Ratio (SIR) for developing secondary primary melanoma among RCC patients was 2.31, and the SIR for developing secondary primary RCC among melanoma patients was 2.87. In previous MITF(E318K) studies, inclusion of high melanoma risk patients with a primary manifestation of RCC or other cancers could have driven increased frequency of MITF(E318K) in observed non-melanoma cohorts. Common putative risk genes for RCC and melanoma – BAPI, MITF, CDKN2B – suggest that similar pathways are disrupted in both disorders; however, individual variant risks should be evaluated when shared genetic background is taken into account. For example, Christensen et al. investigated 48 families with early onset RCC, a family history of RCC, a family history of RCC and melanoma, or both RCC and melanoma diagnosis in the same individual. MITF(E318K) was found only in a RCC-affected member of a family with multiple melanomas. Consequently, authors concluded that BAPI, MITF or CDKN2B are not frequent causes of hereditary renal cancer.

This common polygenic background and shared environmental factors likely contributed, at least in part, to Bertolotto et al.’s observation of an enrichment of the variant in RCC and melanoma cases. While this study did report an association with RCC-only, the cohort was young, had family history of RCC, and had rare histological subtypes, unlike the TCGA patients. Thus, MITF(E318K) may play a role in driving cancer formation in a subset of “genetically-enriched” RCC patients by a still undisclosed biological mechanism. Of note, several other researchers have also failed to identify an association of sporadic RCC with MITF(E318K). This may also explain Ghiorzo et al.’s findings, as their association was only noted in melanoma patients with a personal or family of pancreatic cancer, and not in patients with sporadic pancreatic cancer. The underpinnings of variance regarding pheochromocytoma/paraganglioma is unclear, but may be due to the difference in age between our and Castro-Vega et al.’s cohorts.

Beyond cutaneous melanoma, our study also found a putative risk association between the germline MITF(E318K) and uterine carcinomas (malignant mixed Müllerian tumors) although the low frequency of the variant and, expectedly, low numbers of variant carriers, preclude us from rigorously adjusting for multiple hypotheses. Interestingly, uterine carcinomas (mixed Müllerian tumors) have been reported to exhibit melanocytic differentiation, and increased MITF expression. While more research needs to be completed, this study will help update cancer screening recommendations for patients with the MITF(E318K) variant.

A limitation of this study is the relatively small sample size of some of the TCGA cohorts. This, along with the rarity of the MITF variant, contributes to the limited power of analysis. Nevertheless, it may be worthwhile to better characterize the relationship of MITF(E318K) with different cancer types in a larger study with explicit evaluation of inherited polygenic predisposition to melanoma, and to look for biology of how MITF(E318K) mediates risks for co-occurrence of other cancers with melanoma.
Methods

Literature review and meta-analysis. We searched MEDLINE and Harvard HOLLIS from database inception to February 25, 2020 by using various combinations of the terms "melanoma", "MITF", "E318K", and "cancer". Studies without non-cancer controls were excluded. No pertinent non-English articles were found. Meta-analysis was conducted using the Review Manager 5.3 software, published by the Cochrane community. All variables were considered dichotomous, and analyzed using the Mantel–Haenszel statistical method and fixed effect analytic method. Heterogeneity was assessed using the Chi² test and I² statistic. See Supplementary File S1 for full methods and excluded studies.

TCGA and enriched cohorts. Datasets of germline cancer (TCGA sporadic and MGH genetically-enriched cases) and control exome sequences are described in previous publications from our lab.14-16. Details of enriched patient cohorts can be found in Supplementary Methods S1. All datasets can be accessed through dbGAP using the following accession numbers: phs000178.v1.p1, phs000823.v1, phs000822.v1.p1, phs000806.v1.p1, and phs000814.v1.p1.

Aggregated set of samples was used for joint variant calling using Picard/GATK pipeline. Principal component analysis (PCA) using common LD-pruned autosomal variants was used to identify a cluster of European samples (k-means). Individual genotypes for the MITF(E318K) variant were filtered. Only genotypes with depth of coverage more than 10X and genotype quality more than 20× were used for analysis and all cohorts had the target variant called in more than 95% of the samples. Fisher exact test (two-sided) was used to test association of coverage more than 10X and genotype quality more than 20× were used for analysis and all cohorts had the target variant called in more than 95% of the samples. Fisher exact test (two-sided) was used to test association of coverage more than 10X and genotype quality more than 20× were used for analysis and all cohorts had the target variant called in more than 95% of the samples. Fisher exact test (two-sided) was used to test association with a specific cancer type. Odds ratio and 95% confidence interval was calculated using Graph Prism using the Baptista-Pike method. MAF was calculated by MITF(E318K) # Minor Alleles/# Total Samples/2.

Data availability

The TCGA datasets were generated using the data publicly available at https://portal.gdc.cancer.gov. The genetically-enriched datasets can be accessed through dbGAP using the following accession numbers: phs000178.v1.p1, phs000823.v1, phs000822.v1.p1, phs000806.v1.p1, and phs000814.v1.p1.

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References

1. Yokoyama, S. et al. A novel recurrent mutation in MITF predisposes to familial and sporadic melanoma. Nature 480, 99–103. https://doi.org/10.1038/nature10630 (2011).
2. Bertolotto, C. et al. SUMOylation-defective MITF germline mutation predisposes to melanoma and renal carcinoma. Nature 480, 94–98. https://doi.org/10.1038/nature10539 (2011).
3. Berwick, M. et al. MITF E318K’s effect on melanoma risk independent of, but modified by, other risk factors. Pigment Cell Melanoma Res. 27, 485–488. https://doi.org/10.1111/jpcr.12215 (2014).
4. Ghiorzo, P. et al. Prevalence of the E318K MITF germline mutation in Italian melanoma patients: associations with histological subtypes and family cancer history. Pigment Cell Melanoma Res. 26, 259–262. https://doi.org/10.1111/jpcr.12047 (2013).
5. Gromowski, T. et al. Prevalence of the E318K and V320I MITF germline mutations in Polish cancer patients and multiorgan cancer risk: a population-based study. Cancer Genet. 207, 128–132. https://doi.org/10.1016/j.cancergen.2014.03.003 (2014).
6. Mangas, C. et al. Genetic susceptibility to cutaneous melanoma in southern Switzerland: role of CDKN2A, MC1R and MITF. Br. J. Dermatol. 175, 1030–1037. https://doi.org/10.1111/bjd.14097 (2016).
7. McMeniman, E. K. et al. The interplay of sun damage and genetic risk in Australian multiple and single primary melanoma cases and controls. Br. J. Dermatol. https://doi.org/10.1111/bjd.18777 (2019).
8. Potrony, M. et al. Prevalence of MITF p. E318K in patients with melanoma independent of the presence of CDKN2A causative mutations. JAMA Dermatol. 152, 405–412. https://doi.org/10.1001/jamadermatol.2015.4556 (2016).
9. Wadt, K. A. et al. Molecular characterization of melanoma cases in Denmark suspected of genetic predisposition. PLoS ONE 10, e0122662. https://doi.org/10.1371/journal.pone.0122662 (2015).
10. Ozola, A. & Pjanova, D. The lack of E318K MITF germline mutation in Latvian melanoma patients. Cancer Genet. 208, 355–356. https://doi.org/10.1016/j.cancergen.2015.03.011 (2015).
11. Castro-Vega, L. J. et al. The MITF p. E318K variant, as a risk factor for pheochromocytoma and paraganglioma. J. Clin. Endocrinol. Metab. 101, 4764–4768. https://doi.org/10.1210/jc.2016-2103 (2016).
12. Christensen, M. B. et al. Exploring the hereditary background of renal cancer in Denmark. PLoS ONE 14, e0215725. https://doi.org/10.1371/journal.pone.0215725 (2019).
13. Stoehr, C. G. et al. The microphthalmia-associated transcription factor p. E318K mutation does not play a major role in sporadic renal cell tumors from caucasian patients. Pathobiology 83, 165–169. https://doi.org/10.1159/000435311 (2016).
14. Karczewski KJ, Tao G, Cummings BR, Alfeldt J, Wang Q, Collins RL, Laricchia K, Ganna A, Birnbaum DP, Gauthier LD, Brand S, Ferriera S, Gabriel S, Gentry J, Gupta N, Jeandet T, Kaplan D, Llanwarne C, Munshi R, Novod S, Petrillo N, Roazen D, Ruano-Rulio V, Saltzman A, Schleicher M, Soto J, Tibbetts K, Tolonen C, Wade G, Talkowski ME, The Genome Aggregation Database Consortium, Neale BM, Daly MJ. Variation across 141,456 human exomes and genomes reveals the spectrum of loss-of-function intolerance across human protein-coding genes. bioRxiv. 1, 1. https://doi.org/10.1101/531210 (2019).
15. Kim, K. et al. Association between melanoma and renal-cell carcinoma for sequential diagnoses: a single-center retrospective study. Cancer Epidemiol. 57, 80–84. https://doi.org/10.1016/j.canep.2018.08.003 (2018).
16. Granter, S. R., Wellbrock, K. N., Quigley, C. & Fisher, D. E. Role for microphthalmia transcription factor in the diagnosis of metastatic malignant melanoma. Appl. Immunohistochem. Mol. Morphol. 10, 47–51. https://doi.org/10.1097/00129039-20020300-00008 (2002).
17. Roma, A. A., Malpica, A. & Deavers, M. T. Malignant melanoma arising in an ovarian carcinosarcoma: case report and review of the literature. Int. J. Gynaecol. Pathol. 30, 158–162. https://doi.org/10.1097/PGP.0b013e31819eb85 (2011).
18. Artomov, M. et al. Case-control analysis identifies shared properties of rare germline variation in cancer predisposing genes. Eur. J. Hum. Genet. 27, 824–828. https://doi.org/10.1038/s41431-019-0346-0 (2019).
19. Artomov, M. et al. Rare variant, gene-based association study of hereditary melanoma using whole-exome sequencing. *J. Natl. Cancer Inst.* **109**, 1. https://doi.org/10.1093/jnci/djx083 (2017).

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**Author contributions**

S.G. and H.T. conducted the systematic review and meta-analysis. M.A., S.G., and H.T. conducted the TCGA analysis. A.S., K.S., L.E., M.A., S.M., C.N., S.G., and H.T. prepared and analyzed the genetically-enriched data. All authors prepared and edited the manuscript. S.G. submitted the manuscript. S.G. and H.T. serve as guarantors.

**Competing interests**

The authors declare no competing interests.

**Additional information**

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