Sleep in Alzheimer's Disease–Beyond Amyloid

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A B S T R A C T

Sleep disorders are prevalent in Alzheimer's disease (AD) and a major cause of institutionalization. Like AD pathology, sleep abnormalities can appear years before cognitive decline and may be predictive of dementia. A bidirectional relationship between sleep and amyloid β (Aβ) has been well established with disturbed sleep and increased wakefulness leading to increased Aβ production and decreased Aβ clearance; whereas Aβ deposition is associated with increased wakefulness and sleep disturbances. Aβ fluctuates with the sleep-wake cycle and is higher during wakefulness and lower during sleep. This fluctuation is lost with Aβ deposition, likely due to its sequestration into amyloid plaques. As such, Aβ is believed to play a significant role in the development of sleep disturbances in the preclinical and clinical phases of AD. In addition to Aβ, the influence of tau AD pathology is likely important to the sleep disturbances observed in AD. Abnormal tau is the earliest observable AD-like pathology in the brain with abnormal tau phosphorylation in many sleep regulating regions such as the locus coeruleus, dorsal raphe, tuberomammillary nucleus, parabrachial nucleus, and basal forebrain prior to the appearance of amyloid or cortical tau pathology. Furthermore, human tau mouse models exhibit AD-like sleep disturbances and sleep changes are common in other tauopathies including frontotemporal dementia and progressive supranuclear palsy. Together these observations suggest that tau pathology can induce sleep disturbances and may play a large role in the sleep disruption seen in AD. To elucidate the relationship between sleep and AD it will be necessary to not only understand the role of amyloid but also tau and how these two pathologies, together with comorbid pathology such as alpha-synuclein, interact and affect sleep regulation in the brain.

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Abbreviations: Aβ, amyloid β; AD, Alzheimer's disease; APP, amyloid precursor protein; BF, basal forebrain; bvFTD, behavioral variant FTD; CBD, corticobasal degeneration; CSF, cerebrospinal fluid; DLB, dementia with Lewy bodies; DR, dorsal raphe; EDS, excessive daytime sleepiness; EEG, electroencephalography; FTD, Frontotemporal Dementia; htau, human tau; ISF, interstitial fluid; LC, locus coeruleus; LDT, laterodorsal tegmental nucleus; LH, lateral hypothalamus; mPFC, medial prefrontal cortex; NBM, nucleus basalis of Meynert; NFT, neurofibrillary tangle; NREM, non-rapid eye movement; PAG, periaqueductal gray matter; PLMD, periodic limb movement disorder; PPT, pedunculopontine tegmental nucleus; PSP, progressive supranuclear palsy; Pz, parafascial zone; RBD, REM sleep behavior disorder; REM, rapid eye movement; SCN, suprachiasmatic nucleus; SLD, sublaterodorsal area; SWS, slow wave sleep; TMN, tuberomammillary nucleus; VLO, ventrolateral preoptic nucleus; vPAG, ventral periaqueductal gray

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1. Introduction

For over 25 years, a high prevalence of sleep disorders has been identified in patients with Alzheimer’s disease (AD). Many studies have estimated that 25–66% of AD patients exhibit sleep disorders of some kind and slow sleep disturbances are one of the leading causes of AD patient institutionalization (Bianchetti et al., 1995; Guarnieri et al., 2012; Moran et al., 2005). Sleep is an important biological function and although sleep disturbances were once considered to be a bi-product of neurodegeneration, they are not limited to advanced disease states but occur even before cognitive decline and may be predictive of neurodegeneration (Hahn et al., 2013; Lim et al., 2013). AD pathology has been shown to begin long before the onset of the clinical cognitive impairment characteristic of AD. The preclinical stage of AD is characterized by the aggregation of amyloid-β (Aβ) peptide into amyloid plaques in the brain and a decrease of Aβ42 in the cerebrospinal fluid (CSF) as well as tau phosphorylation and aggregation into neurofibrillary tangles (NFT) and neuropil threads in neocortical regions (Holtzman et al., 2011). This molecular pathology, especially tau pathology, is then linked with neuronal death, synaptic loss, cognitive impairment, and AD diagnosis (Jack and Holtzman, 2013; Holtzman et al., 2011). The presence of sleep disturbances throughout preclinical and clinical disease progression underscores the important role sleep may play in disease pathology and progression.

Studies in humans and mouse models have begun to unravel the relationship between sleep, AD, and cognitive impairment, but much work remains. Due to the abundance of mouse models that develop amyloid deposition and biomarker tests available, much is known about the bidirectional interaction between sleep, Aβ, and its aggregation in the brain. However, to fully understand the interaction between AD and sleep, the contribution of tau and other pathologies cannot be ignored. Here, we will review the clinical relationship between AD and sleep and how amyloid pathology contributes to this interaction. We will further highlight how tau pathology in AD may interact with sleep disturbances including evidence from tauopathies such as frontotemporal dementia (FTD). Lastly, up to 60% of AD patients also have the presence of alpha-synuclein aggregation in the brain and we will highlight ways in which this pathology may contribute to sleep disturbances seen in AD as well as similarities between AD and dementia with Lewy bodies (DLB) (Hamilton, 2000).

2. Sleep and circadian disturbances in AD patients

AD patients exhibit a wide array of sleep, electroencephalography (EEG), and circadian dysfunction that is well reviewed in the literature (Musiek et al., 2015; Peter-Derex et al., 2015; Lim et al., 2014b). Excessive daytime sleepiness (EDS), sun-downing, and insomnia are among the most common reported disturbances in AD and stem from changes in sleep architecture and circadian rhythm. AD patients have been shown to have decreased sleep efficiency, non-rapid eye movement (NREM) sleep and slow wave sleep (SWS) as well as decreased rapid eye movement (REM) sleep and increased latency to REM sleep (Vitiello et al., 1990; Bliwise et al., 1989; Bonanni et al., 2005). Consequently, AD patients display an increase in wakefulness that is characterized by an increased number of awakenings at night (Moe et al., 1995; Vitiello et al., 1990; Bonanni et al., 2005). Taken together these changes result in an observed increase in sleep fragmentation in AD patients (Vitiello et al., 1990). Sleep disturbances are found to occur throughout clinical disease with most changes visible in mild disease stages and these disruptions worsen with disease severity (Vitiello et al., 1990; Bonanni et al., 2005; Liguori et al., 2014b). Increased REM sleep latency and wakefulness as well as decreased REM sleep and sleep efficiency are also found to correlate with impaired cognitive function (Moe et al., 1995; Liguori et al., 2014b). The abundance of sleep disturbances in AD patients even in early stages of cognitive impairment may be due to neuronal and synaptic loss that is already occurring at these stages of disease.

Prior to clinical diagnosis, sleep disturbances are also a risk factor for developing cognitive dysfunction and AD. A recent study in a preclinical population has shown using actigraphy that a decrease in sleep efficiency is visible even in cognitively normal people that are amyloid positive compared to those that are amyloid negative, suggesting a role for Aβ aggregation in preclinical sleep changes (Ju et al., 2013). Furthermore, clinical follow up studies have shown that cognitively normal older individuals with high sleep fragmentation had a 1.5-fold increased risk of developing AD and self-reported reduced sleep was associated with a 2-fold increased risk of AD development (Hahn et al., 2013; Lim et al., 2013). These early, preclinical changes in sleep suggest that poor sleep may be a risk factor and biomarker for AD and cognitive decline and demonstrates that pathological AD-type changes may be leading to sleep abnormalities prior to obvious cognitive symptoms. This supports the idea that sleep disruption may be prognostic for future cognitive decline.

Changes in sleep architecture are also accompanied by changes in EEG power. Quantitative EEG studies show that AD patients exhibit EEG slowing in both wake and REM sleep that increases with disease severity and suggest that REM sleep EEG slowing is a robust biomarker for AD severity (see (Petit et al., 2004) for in-depth review). Markers of circadian dysfunction have also been identified in AD patients and include increased nocturnal activity, decreased diurnal activity, core body temperature phase delay and amplitude decrease, clock gene phase changes across brain regions, and changes in sleep regulating hormones such as decreased nocturnal melatonin (sleep promoting) and increased hypocretin/orexin (wake promoting) (Voilcer et al., 2001; Harper et al., 2005; Liguori et al., 2014b; Wu et al., 2003; Cer makan et al., 2011). The relationship between AD progression and sleep modulating hormones is not straightforward however. Studies have shown that CSF orexin levels in moderate to severe AD are significantly increased compared to controls and high orexin levels were appropriately associated with increased nocturnal disturbance (Liguori et al., 2014b). However, post mortem analysis
increased activity rhythm fragmentation and increased nocturnal neurotensin positive neurons in the SCN is also correlated with matic AD (Jack and Holtzman, 2013; Holtzman et al., 2011). Simi-3. Amyloid and sleep thology and sleep degeneration.

observations demonstrate a direct relationship between AD pathology in the suprachiasmatic nucleus (SCN) has also been es-3.2. The bidirectional relationship between Aβ and sleep disturbance

changes observed in AD patients and provide a unique opportunity to explore the direct effect of APP/Aβ on sleep without the presence of other pathological characteristics of AD such as tau aggregation. Increases in wake and decreases in both NREM and REM sleep have been reported in most mouse models studied including APP/PS1, Sx FAD, PDAPP, TgCRND8, and Tg2576, similar to changes seen in AD patients (Roh et al., 2012; Schneider et al., 2014; Huitrón-Reséndiz et al., 2002; Colby-Milley et al., 2015; Zhang et al., 2005). The normalization of these deficits by Aβ immunization in APP/PS1 and Tg2576 mice demonstrates the ability of Aβ pathology itself (distinct from APP) to induce changes in sleep architecture (Wisor et al., 2005; Roh et al., 2012). Furthermore, APP transgenic mice have increased sleep fragmentation, increased latency to REM sleep, and decreased number of REM sleep bouts, changes that are reminiscent of those seen in clinical AD (Colby-Milley et al., 2015; Sethi et al., 2015; Zhang et al., 2005). Sleep changes in multiple models have also been shown to occur early in disease progression when plaque pathology is low and worsen with age, exemplifying the importance of Aβ aggregation on sleep disruption in early and advanced stages of AD (Colby-Milley et al., 2015; Huitrón-Reséndiz et al., 2002; Zhang et al., 2005). The effects of Aβ on sleep are not limited to mice and humans, but extend to Dro- sophila, where expressing Aβ results in decreased and fragmented sleep (Tabuchi et al., 2015). These studies across organisms demonstrate the importance and robust effect of amyloid pathology on sleep. EEG power also appears to be affected in most mouse models. Despite differences between models, there is a prevailing observation of decreased power with disease progression in the delta and/or theta frequencies (Colby-Milley et al., 2015; Zhang et al., 2005; Schneider et al., 2014). Furthermore, unlike humans with AD who display EEG showing, these mouse models show an increase in higher EEG frequencies, demonstrating that Aβ mouse models do not recapitulate all aspects of sleep changes seen in clinical AD (Colby-Milley et al., 2015; Zhang et al., 2005).

The ability of amyloid pathology to induce many changes similar to those seen in AD suggests that amyloid may be a driving force behind sleep changes in early AD and throughout disease progression. How Aβ induces sleep changes is still to be determined and is likely the result of many mechanisms. Changes could be due to the effect of Aβ and amyloid pathology on neuronal populations or brain regions crucial to sleep-wake systems. Although amyloid pathology in the subcortical and brainstem nuclei is shown to occur later in disease and cannot explain early changes in sleep (Serrano-Pozo et al., 2011), worsening of sleep deficits may be due to progression of pathology spreading from cortical to subcortical and later brainstem structures, as suggested in the CRND8 mouse model (Colby-Milley et al., 2015). In AD brain, diffuse plaques are found throughout the hypothalamus, which contains many sleep regulating regions including the VLPO, the main sleep promoting region of the brain where NFTs are infrequent, and amyloid pathology in this region is one possible mechanism for the sleep loss observed in AD (Standaert et al., 1991). Amyloid plaques are also the dominant clinical AD pathol-ogy observed in the periaqueductual grey matter (PAG), with 81% of AD cases having amyloid pathology in this dopaminergic wave-active area, although tau pathology is observed early in disease progression as well (Parvizi et al., 2000; Stratmann et al., 2016). Sleep changes could also be the result of neuronal circuit dys-function, such as Aβ induced breakdown of slow oscillations and coherence in the neocortex, thalamus and hippocampus and de-terioration of slow-wave activity (Busche et al., 2015). These are but a few of the possible mechanisms by which Aβ may induce changes in sleep.

3.1. Amyloid-precursor protein (APP)/Aβ overproduction induces sleep changes in animal models

APP/Aβ overproduction models recapitulate many of the sleep changes observed in AD patients and provide a unique opportunity to explore the direct effect of APP/Aβ on sleep without the presence of other pathological characteristics of AD such as tau aggregation. APP/PS1, Sx FAD, PDAPP, TgCRND8, and Tg2576, similar to changes seen in AD patients (Roh et al., 2012; Schneider et al., 2014; Huitrón-Reséndiz et al., 2002; Colby-Milley et al., 2015; Zhang et al., 2005). The normalization of these deficits by Aβ immunization in APP/PS1 and Tg2576 mice demonstrates the ability of Aβ pathology itself (distinct from APP) to induce changes in sleep architecture (Wisor et al., 2005; Roh et al., 2012). Furthermore, APP transgenic mice have increased sleep fragmentation, increased latency to REM sleep, and decreased number of REM sleep bouts, changes that are reminiscent of those seen in clinical AD (Colby-Milley et al., 2015; Sethi et al., 2015; Zhang et al., 2005). Sleep changes in multiple models have also been shown to occur early in disease progression when plaque pathology is low and worsen with age, exemplifying the importance of Aβ aggregation on sleep disruption in early and advanced stages of AD (Colby-Milley et al., 2015;
in the Tg2576 and APP/PS1 amyloid mouse models which both had a significant increase in amyloid plaque pathology following 6 weeks of sleep deprivation (Kang et al., 2009). Conversely, 8 weeks of increased sleep by treatment with an orexin receptor antagonist significantly decreased amyloid pathology in APP/PS1 mice, suggesting that sleep is a potent modulator of amyloid both acutely, of monomeric Aβ, and over time, of amyloid pathology progression (Kang et al., 2009). Drosophila that overexpress Aβ demonstrate the same increase in Aβ accumulation with sleep deprivation and decrease in accumulation with sleep increase, further supporting the strong effect of sleep on amyloid pathology (Tabuchi et al., 2015).

Increases in Aβ due to sleep disturbance could be the result of increased Aβ secretion, decreased clearance, or both mechanisms. In Drosophila, sleep deprivation induced Aβ increases are lost when neuronal hyperexcitability is inhibited, suggesting that Aβ increases with sleep deprivation are due to increased neuronal activity that accompanies wakefulness (Tabuchi et al., 2015). This observation is supported in mice where both sleep deprivation and neuronal activity increases Aβ secretion into the interstitial fluid (ISF) (Cirrito et al., 2005; Kang et al., 2009). Sleep has also been shown to facilitate clearance of Aβ from the ISF by means of the glymphatic system, and thus the decrease of clearance in this fashion may also contribute to the increases in Aβ when sleep is lost (Xie et al., 2013).

### 3.3. Amyloid-β and circadian rhythm

Circadian dysfunction is well documented in AD patients and studies have demonstrated a delicate interaction between AD, circadian function, and amyloid (see (Musiek et al., 2015) for in depth review). Aβ, like many proteins in the brain, exhibits a circadian fluctuation and this observation is seen in both humans and mice. Humans have increased CSF Aβ throughout the daytime that then decreases at night and these Aβ levels are inversely correlated with sleep (Huang et al., 2012). Mouse ISF Aβ exhibits the same circadian fluctuation in both wild type and APP/Aβ overexpression models (Roh et al., 2012; Kang et al., 2009). The ability of sleep deprivation to raise Aβ levels in the brain could be a direct result of this fluctuation. Lack of sleep may inhibit the reduction of Aβ that would normally occur during the restful period, which could affect amyloid pathology development in AD over a longer period of time. Furthermore, AD pathology affects this Aβ fluctuation. In humans, the amplitude of Aβ fluctuation decreases both with age as well as the presence of amyloid aggregation and amyloid positive patients with familial dominant AD have a loss of Aβ fluctuation (Roh et al., 2012; Huang et al., 2012). Lower CSF Aβ42 levels in humans is also associated with lower hypocretin levels and increased amyloid pathology, further suggesting the important relationship between abnormal circadian function and AD amyloid pathology (Slats et al., 2012). Aβ overexpressing APP/PS1 mice display a similar loss of circadian Aβ fluctuation in the ISF, likely due to sequestration of Aβ42 into amyloid plaques, a phenomenon also likely occurring in humans (Roh et al., 2012). The loss of fluctuation occurs with the onset of amyloid pathology and is rescued by Aβ42 immunization, demonstrating the causative role of amyloid pathology in the loss of Aβ fluctuation (Roh et al., 2012). In addition to Aβ fluctuation, amyloid overexpression mouse models also show changes in circadian processes such as decreased thermoregulation and increased period, suggesting that some form of Aβ is sufficient to induce circadian dysregulation similar to that seen in ageing and AD (Wisor et al., 2005; Huitrón-Reséndiz et al., 2002; Song et al., 2015). Mechanistically, APP/Aβ overexpression may affect circadian features through SCN degeneration, which is observed in 3xTg mice and humans with AD (Sterniczuk et al., 2010; Harper et al., 2008). Although rare, diffuse amyloid plaques are seen in the SCN of AD patients (Stopa et al., 1999). Aβ in culture and mouse models is also shown to induce changes in clock genes, such as decreases in PER2 and BMAL1, which could induce circadian dysregulation outside the SCN (Song et al., 2015). Taken together, these studies demonstrate an important interaction between Aβ and circadian rhythms that could contribute to and possibly accelerate disease progression.

### 4. Tau and sleep

The hyperphosphorylation and intercellular aggregation of tau protein is the second hallmark of AD and unlike Aβ, total and phosphorylated tau in the CSF has been shown to predict and correlate with cognitive decline in preclinical and clinical AD (Fagan et al., 2007; Mattsson et al., 2009). Furthermore, tau pathology has recently been shown to be the earliest observable AD-like change in human brain, with abnormal tau phosphorylation and aggregation in the LC beginning as early as young adulthood and extending to other connected regions even before amyloid is detected (Braak et al., 2011; Stratmann et al., 2016). Despite these observations, very little is known about the effect of tau pathology on sleep in preclinical or clinical AD. Currently, our understanding of the possible role of tau in AD related sleep disturbance is based off the presence of NFTs in sleep centers of the brain, limited studies in AD mouse models containing both APP and tau transgenes or a human tau transgene alone, and what is known about sleep in other tauopathies such as FTD. Together, these areas highlight the important role tau may play in sleep dysregulation in AD.

#### 4.1. Neurofibrillary tangles in sleep, wake, and circadian centers of the brain during AD pathogenesis

Sleep–wake cycles are tightly regulated in the brain by many brain regions, neurotransmitters, and flip-flop switches. Sleep regulating areas are found throughout the brain including the brainstem, midbrain, thalamus, hypothalamus, and BF (Fig. 1) (Saper et al., 2005). These many regions regulate arousal (ascending arousal pathway), promote and inhibit NREM and REM sleep, as well as regulate circadian rhythms and the biological clock (Fig. 1). Many of these regions, especially the wake-promoting arousal system, are shown to be affected by abnormal tau pathology in the human brain with many of the changes occurring during preretangle stages or stage 0 of tau pathology by Braak and Braak staging, prior to any cortical tau or amyloid pathology development (Fig. 1) (Braak et al., 2011; Stratmann et al., 2016; Stern and Naidoo, 2015; Braak and Del Tredici, 2011). Here we will discuss sleep, wake, and circadian regulating areas of the brain known to be affected by tau pathology to underscore the potential importance of tau in sleep dysregulation.

#### 4.1.1. Ascending arousal system

The ascending arousal system is composed of wake-active neuronal groups and includes many brain areas known to contain abnormal tau phosphorylation pathology early in life and with AD disease progression (Stern and Naidoo, 2015; Braak et al., 2011; Braak and Del Tredici, 2011). Tau pathology first appears in the brainstem LC and extends to the medial parabrachial nucleus (mPB), the dorsal raphe (DR), and the PAG (Braak et al., 2011; Stratmann et al., 2016; Braak and Del Tredici, 2011). Pretangle tau pathology is also found in the hypothalamic tuberomammillary nucleus (TMN), lateral hypothalamus (LH), and the BF (Braak et al., 2011; Stratmann et al., 2016; Braak and Del Tredici, 2011) (Fig. 1). All of these areas have abnormal tau phosphorylation prior to cortical tau or amyloid pathology and may play a role in tau...
induced sleep dysregulation during normal aging (Braak et al., 2011; Stratmann et al., 2016; Braak and Del Tredici, 2011). In AD, many regions of the ascending arousal system, including the pedunculopontine tegmental nucleus (PPT) and laterodorsal tegmental nucleus (LDT), have been shown to have robust NFT pathology, further supporting a role of tau in AD sleep disturbances (Parvizi et al., 2001; Stern and Naidoo, 2015). Braak et al. demonstrate that abnormal tau species are found in this area in a high percentage of young adults (70% at 20 years of age) prior to cortical tau or amyloid burden (Braak et al., 2011). Furthermore, NFTs in the LC are reproducibly found in AD patients as well as cognitively normal and mildly impaired adults without full blown dementia (Grudzien et al., 2007; Dugger et al., 2012). In AD, LC noradrenergic neurons are lost and LC neuron number is found to be correlated to cognitive decline in a non-demented cohort (Wilson et al., 2013; Brunnström et al., 2011). Increased NFTs in the LC is also related to more rapid cognitive decline (Wilson et al., 2013). NFTs in and loss of LC neurons may therefore be playing a role in sleep disturbances and EDS found in preclinical and early AD (Stern and Naidoo, 2015). In addition to AD pathology driving loss of LC neurons, these neurons are also susceptible to increased wakefulness (Zhang et al., 2014). Since increased wake is characteristic of AD, it follows that AD may be able to induce degeneration of this area, further contributing to disease and sleep dysfunction. Lastly, LC degeneration has been shown to promote Aβ pathology in APP23 mice and result in increased neuronal loss and memory deficits (Heneka et al., 2006). Tau pathology in the LC or wake induced neuronal loss could therefore affect amyloid pathology development and disease progression in AD. Taken together, tau pathology in the LC may have a profound effect on both sleep and cognition in AD pathogenesis and begin years before other pathologies are present, making it an important area for study of the interaction between sleep, tau, and AD. Furthermore, early pathology in the LC alone is not recapitulated, to our knowledge, in any AD or human tau (htau) mouse models. Such models would be invaluable in studying this interaction.

The serotonergic neurons of the DR are also wake active and in addition to pretangle stage detection of tau pathology in this region, NFT pathology in the DR was also identified in preclinical AD patients (stage 0, 1 and 2) as well as those with a clinical AD diagnosis (Saper et al., 2005; Stern and Naidoo, 2015; Chen et al., 2000; Stratmann et al., 2016; Rüb et al., 2000; Grinberg et al., 2011).
controls (Braak et al., 2011; Mesulam et al., 2004; Stratmann et al., 2016), and early AD, as well as in cognitively normal age matched in pre-tangle stages of AD development, mild cognitive impairment and tau pathology in this region (German et al., 1987; Parvizi et al., 2001; Stratmann et al., 2016).

Cholinergic neurons in the PPT and LTD are active during wake and REM sleep (Fig. 1) (Stern and Naidoo, 2015; Saper et al., 2005). Although studies of stage 0 AD tau pathology have not reported early tau pathology in these areas, NFTs are common in the LTD and PPT in clinical AD (Dugger et al., 2011; Dugger et al., 2012; Parvizi et al., 2001). One study showed that 69% of AD postmortem brains contained NFTs in the PPT without amyloid pathology present there, even in patients with short disease duration (Parvizi et al., 2001). Furthermore, 66% of patients had NFT pathology in the LTD and the presence of NFTs in this area correlated with disease progression (Parvizi et al., 2001). The wake-active, REM-inhibiting PAG of the midbrain has Stage 0 development of tau pathology (Stratmann et al., 2016). However, in the PAG one study reported that only 9% of clinical AD brains autopsied had NFT pathology, while 81% contained amyloid plaques (Parvizi et al., 2000). Therefore, the effect of NFTs in this area is unclear.

In the hypothalamus, abnormal tau pathology is present in the pretangle, precortical tau stage in the TMN and LH (Fig. 1) (Stratmann et al., 2016). Furthermore, in patients who died with dementia due to AD, hypothalamic NFTs are numerous and concentrated in the TMN and accompanied by wake-active histaminergic neuronal loss (Nakamura et al., 1993; Airaksinen et al., 1991a; Stern and Naidoo, 2015; Standaert et al., 1991). This focus of pathology suggests a role of tau in neuronal loss in the TMN and a possible contribution to sleep disruption, but the influence of amyloid cannot be ignored since diffuse amyloid plaques are also present throughout the hypothalamus in the AD brain (Standaert et al., 1991). Although most prominent in the TMN, NFTs can also be seen in the LH in clinical AD (Standaert et al., 1991). LH neurons are wake active and produce orexin, which acts to stabilize the sleep-wake switch, allowing for sustained arousal and sleep (Saper et al., 2005). Pretangle tau pathology has also been reported specifically in the prefrontal area which contains the orexinergic neurons (Stratmann et al., 2016). Although it is not known if tau pathology affects orexinergic neurons, orexin is an important marker of sleep-wake regulation that is shown to increase in the CSF of moderate to severe AD patients and is positively correlated to total tau and related to sleep impairment in AD (Ligouri et al., 2014b). Furthermore, orexin levels in cognitively normal elderly correlate with phosphorylated tau, suggesting that tau could be a driving factor in orexin and wakefulness changes in the brain (Osorio et al., 2016).

Tau pathology also reaches the magnocellular nuclei of the BF in pre-tangle stages of AD development, mild cognitive impairment, and early AD, as well as in cognitively normal age matched controls (Braak et al., 2011; Mesulam et al., 2004; Stratmann et al., 2016; Iraizoz et al., 1999). Cholinergic neurons in the basal forebrain (nucleus basalis of Meynert (NBM)) are active during wake and REM sleep and also play an important role in cognition with excitatory projection to the cortex (Fig. 1) (Saper et al., 2005; Stern and Naidoo, 2015). NBM neurons are lost in AD and cell number decreases with disease progression (Rogers et al., 1985; Iraizoz et al., 1999). NFTs may be to blame for this cell loss since remaining neurons in this area contain NFTs but very few, if any, amyloid plaques are present (Iraizoz et al., 1999; Rogers et al., 1985). Furthermore, NFTs in the NBM correlate with higher mental deterioration and decreased BF volume in AD correlates with increased cortical amyloid plaque burden, suggesting that tau pathology and loss of neurons in the BF could play a role in disease progression as well as cognitive decline and sleep disruption (Kerbler et al., 2015; Iraizoz et al., 1999).

Many regions of the ascending arousal system are affected by tau pathology early in life and prior to preclinical amyloid pathology development, supporting that tau may play a crucial role in sleep disturbance and sleep-wake system dysregulation not only in clinical AD but also in preclinical AD and normal aging. More work is needed to determine if tau pathology in these areas induces changes in sleep-wake regulation. It is known that lesions in some of the wake regulating areas discussed here do not induce changes in sleep-wake architecture (Fuller et al., 2011; Compf et al., 2010). However, tau pathology may induce dysregulation by affecting multiple areas, specific cell types, or in conjunction with pathology in sleep promoting regions.

4.1.2. Sleep promoting regions

AD pathology in and degeneration of sleep promoting regions of the brain could explain the loss of sleep seen in clinical AD. One of the main sleep promoting regions of the brain is the VLPO. The VLPO, located in the hypothalamus, projects to the ascending arousal system and inhibits this system during sleep (Fig. 1) (Saper et al., 2005). Surprisingly, the VLPO has not been shown to contain prominent NFT pathology (Standaert et al., 1991). However, there are other regions in the brain that function to generate NREM and REM sleep, such as the PFZ and the sublaterodorsal area (SLD), which may be affected by tau pathology (Fig. 1). The PFZ, a group of GABAergic neurons in the brainstem, is a SWS generating region recently identified in mice (Anaclet et al., 2014). Disruption of the PFZ could result in decreased SWS as seen in AD, but to our knowledge pathology in this region remains unstudied (Anaclet et al., 2014). The SLD and pre coeruleus is a REM generating region in the brain also elucidated in mice and rats (Krenzer et al., 2011; Lu et al., 2006). The presence of tau pathology in the SLD has not been specifically studied. However, the SLD REM producing region is located near the LC, the earliest location of tau pathology, so it is possible that tau pathology could accumulate in this region throughout aging and the progression of AD. Furthermore, the SLD receives input from the ventrolateral PAG which serves as a REM off region. The PAG has also been shown to have early presence of tau pathology, strengthening the possible effect of tau on REM sleep (Krenzer et al., 2011; Lu et al., 2006). Disruption to or cell loss in the SLD or disruption of the REM on/off balance could be a possible mechanism for loss of REM sleep and increased REM latency in AD. Other brain areas that are active in wake are also active during REM sleep and affected by tau pathology in pre-tangle stages, the BF, and in clinical AD, the LTD and PPT (Fig. 1). Degeneration in these regions could further decrease REM sleep pathway activation and sustenance.

4.1.3. Circadian regions

The SCN is the master pacemaker of the body and regulates circadian rhythm and inputs (Fig. 1). There is not significant tau pathology in this region with only scattered NFTs as well as rare, diffuse amyloid plaques observed in the SCN of AD brains (Stopa et al., 1999). Although the impact of this pathology on circadian rhythm is unknown it may contribute to the loss of vasopressin and neuropeptide hormones in the SCN and circadian disruption observed in AD patients (Musiek et al., 2015; Stopa et al., 1999). The TMN has also been shown to project to the SCN and contain tau pathology which could possibly affect input and regulation of the

2009). Beyond the presence of NFTs in the DR, neuron density is also decreased in AD patients (Chen et al., 2000). Although this loss of neurons or NFT number in the DR of AD patients was not correlated with cognitive decline, it is possible that loss of serotonergic neurons and NFT induced changes in this region contributes to sleep dysregulation in AD patients (Chen et al., 2000).

The mPB is also wake active and inhibited by the parafacial zone (PFZ) during NREM sleep (Anaclet et al., 2014). Abnormal tau pathology in the mPB was identified at stage 0 and NFTs are found in the mPB of up to 91% of postmortem clinical AD brains in the absence of amyloid pathology, supporting a link between clinical symptoms and tau pathology in this region (German et al., 1987; Parvizi et al., 2001; Stratmann et al., 2016).

Many regions of the ascending arousal system are affected by tau pathology early in life and prior to preclinical amyloid pathology development, supporting that tau may play a crucial role in sleep disturbance and sleep-wake system dysregulation not only in clinical AD but also in preclinical AD and normal aging. More work is needed to determine if tau pathology in these areas induces changes in sleep-wake regulation. It is known that lesions in some of the wake regulating areas discussed here do not induce changes in sleep-wake architecture (Fuller et al., 2011; Compf et al., 2010). However, tau pathology may induce dysregulation by affecting multiple areas, specific cell types, or in conjunction with pathology in sleep promoting regions.

4.1.3. Circadian regions

The SCN is the master pacemaker of the body and regulates circadian rhythm and inputs (Fig. 1). There is not significant tau pathology in this region with only scattered NFTs as well as rare, diffuse amyloid plaques observed in the SCN of AD brains (Stopa et al., 1999). Although the impact of this pathology on circadian rhythm is unknown it may contribute to the loss of vasopressin and neuropeptide hormones in the SCN and circadian disruption observed in AD patients (Musiek et al., 2015; Stopa et al., 1999). The TMN has also been shown to project to the SCN and contain tau pathology which could possibly affect input and regulation of the
SCN circadian clock (Stern and Naidoo, 2015; Airaksinen et al., 1991b; Braak et al., 2011; Standaert et al., 1991). More work is needed to identify if tau induces circadian changes or if tau itself is regulated in a circadian fashion to determine if there is a tau/circadian interaction.

4.2. Sleep changes in htau expressing mice

Mice expressing human tau transgenes with disease promoting mutations that cause FTD have been studied in recent years, but only a small number of sleep studies in these mice have been conducted to assess the ability of tau pathology to induce sleep disruption. In an effort to model AD, mice containing both amyloid pathology and human forms of tau have been studied. Circadian changes reminiscent of AD have been noted in the 3xTg mouse model, which has decreased vasopressin neurons in the SCN, increased daytime activity, and shorter circadian period (Sterniczuk et al., 2010). This demonstrates that tau and Aβ pathology, either alone or together, can induce circadian changes. A second amyloid and tau model, PLB1, has shown changes in sleep architecture similar to AD patients and Aβ-depositing mice including increased wake, decreased NREM and REM sleep, less frequent and shorter REM bouts, increased number of short NREM bouts, and overall sleep fragmentation that worsens with age (Jyoti et al., 2015; Platt et al., 2011). This model was further analyzed for the effect of the htau P301L transgene alone by extraction of the amyloid pathology producing portion of the transgene (Koss et al., 2016). Analysis of PLB1tau mice demonstrated for the first time that forebrain mutant htau expression alone alters sleep. Specifically, PLB1tau mice have increased wake and decreased NREM sleep, as well as more robust changes in EEG power than observed in PLB1 mice with both tau and amyloid (Koss et al., 2016). This work is the first evidence that tau pathology alone can induce sleep changes reminiscent of neurodegenerative disease. However, more work is needed in this model to determine what brain regions tau pathology accumulates in as well as other tau models to understand how tau may be inducing changes in sleep. Lastly, loss of tau in tau knock out mice induces changes in sleep including increased wake, decreased NREM sleep, and sleep fragmentation, demonstrating that endogenous tau function is important to sleep regulation and loss of tau function in tauopathies like AD, not just tau pathology, could contribute to the sleep changes observed (Cantero et al., 2010).

Tau may not only affect sleep and wake parameters but it is possible that sleep disruption could contribute to tau pathology progression, as seen with Aβ. One study in 3xTg mice showed an increase in insoluble tau and MC1 staining for conformational tau changes following 2 months of sleep deprivation by light cycle manipulation (Di Meco et al., 2014). This suggests a possible bidirectional relationship between tau pathology and sleep, similar to that seen with Aβ, which could accelerate disease progression. However, this model also includes APP/Aβ overexpression so the effect of amyloid in the observed changes cannot be ignored (Di Meco et al., 2014). Neuronal activity has also been shown to increase tau in the ISF, raising the possibility for extended wakefulness to impact tau pathology progression (Yamada et al., 2014). Although much work remains, these studies demonstrate that tau may be an important component of sleep-wake dysregulation in the development of AD and needs to be taken into consideration when dissecting and understanding the cause and effect of sleep changes in AD.

4.3. Sleep disturbances in tauopathies

The FTD spectrum includes multiple disorders, many of which contain tau pathology. Tau aggregation occurs in approximately half of the patients with behavioral variant FTD (bvFTD), which affects the frontal and temporal lobe, and 70% of nonfluent/agrammatic variant FTD cases, which affects the posterior frontal lobe and insula (Pressman and Miller, 2014; Pan and Chen, 2013). There are also 2 other pure tauopathies: progressive supranuclear palsy (PSP) and corticobasal degeneration (CBD) (Pan and Chen, 2013). PSP is characterized by globule NFTs that affect largely the brainstem and basal ganglia while CBD is characterized by ballooned neurites, globule corticobasal bodies, and fine filamentous inclusions in the cortex as well as the basal ganglia, subcortical nuclei, and brainstem (Dickson, 1999; Pan and Chen, 2013). In both of these disorders, there are also astrocytic tau inclusions (Dickson, 1999). Despite the variation in FTD, studies have shown that patients suffer from sleep disturbances as frequently as or possibly more than AD. One large clinical study showed that 65.7% of AD patients and 76% of FTD patients had sleep disturbances of some kind including insomnia, restless leg syndrome, sleep disordered breathing, and EDS (Guarnieri et al., 2012). This and other studies suggest that sleep is possibly better preserved in AD than FTD (Bonakis et al., 2014; Guarnieri et al., 2012; Pistacchi et al., 2014).

FTD patients experience many similar sleep disturbances as AD. Studies in bvFTD patients have shown decreased total sleep time and percent sleep, decreased sleep efficiency that is worse in MRI abnormal FTD, decreased REM sleep, and changes in NREM sleep (Anderson et al., 2009; Bonakis et al., 2014). When compared to AD patients, there are no striking differences in bvFTD patient sleep parameters. One study showed no differences in any sleep parameters between AD and bvFTD and while a second showed AD and bv-FTD to both display EDS and altered NREM sleep compared to controls, only the NREM/REM ratio was significantly decreased in FTD compared to AD, although REM sleep and total sleep were decreased in FTD only compared to controls (Bonakis et al., 2014; Kundermann et al., 2011). Finally, when analyzing orexin, no difference has been observed between FTD and controls but FTD orexin levels are decreased compared to AD (Çoban et al., 2013; Liguori et al., 2014a).

Studies in pure tauopathies have also shown these patients to exhibit sleep disturbances similar to those in AD. Patients with PSP have decreased total sleep time and sleep efficiency, decreased sleep spindles, increased sleep latency, and decreased REM sleep characterized by decreased number and duration of REM episodes (Gama et al., 2010; Montplaisir et al., 1997; Petit et al., 2004). The observed changes in REM sleep in PSP are suggestive of PPT degeneration (Montplaisir et al., 1997). As previously discussed, the PPT contains robust tau pathology in AD, and thus tau pathology in this brain region may contribute to the loss of REM sleep in AD. Furthermore, PSP exhibits slowing of wake EEG as well as EDS that correlates with disease progression, similar to changes observed in AD (Montplaisir et al., 1997; Gama et al., 2010; Petit et al., 2004). A patient with a TAU G389R mutation causing tau pathology and hereditary Pick’s disease also showed sleep fragmentation and decreased SWS (Gemignani et al., 2005). These observations demonstrate the ability of tau pathology to induce profound changes in sleep not unlike those seen in AD and supports a possible role for tau pathology in AD sleep disruption.

5. Alpha-synuclein aggregation, DLB, and sleep

Alpha-synuclein is a 140 amino acid protein predominantly expressed at pre-synaptic terminals of neurons and is thought to play a role in vesicle trafficking and release (Bendor et al., 2013). Intracytoplasmic aggregates of alpha-synuclein, present in Lewy bodies and Lewy neurites, are pathological hallmarks of a group of neurodegenerative diseases such as Parkinson disease, multiple system atrophy, and DLB. Up to 60% of patients with sporadic AD also have Lewy body pathology in the brain (Hamilton, 2000). AD
cases with Lewy bodies show pathology consistently in the amygdala but this can also be observed to spread through the neocortex with limited brainstem and substantia nigra involvement (Hamilton, 2000). Dominantly inherited AD patients also have a high prevalence of Lewy bodies in the brain that are concentrated in the amygdala but also found in limbic, neocortical, and brainstem regions (Lippa et al., 1998; Leverenz et al., 2006). Dominantly inherited AD patients with Lewy bodies outside of the amygdala also had a longer course of disease (Leverenz et al., 2006). Furthermore, there is some overlap in symptoms and pathology of AD and DLB which makes it worthwhile to examine the relationship between DLB and sleep here (McKeith, 2000; Lippa and Knopman, 2007; McKeith et al., 2005).

DLB is characterized by the onset of dementia like symptoms, cognitive dysfunction and visual hallucinations coincident with Parkinsonism. In addition to the brain stem and striatum, DLB features synuclein deposits in the insula, cingulate cortex, amygdala and hippocampus (Dickson et al., 1994; Tsuibo and Dickson, 2005; Harding and Halliday, 2001; Bertrand et al., 2004; Aarsland et al., 2004). DLB cases can further present with extensive Aβ deposits in the striatum (Serrano-Pozo et al., 2011). Parkinson's disease with dementia is similar to DLB except that Parkinsonism precedes cognitive impairment by at least one year and usually longer. A number of sleep disorders present along with DLB and some reports estimate that ~50% of DLB patients have sleep disorders and this can increase to over 70% in moderate to severe disease (Borroni et al., 2008; Farina et al., 2009). When compared to AD, it has also been suggested that DLB has an overall greater sleep disturbance with DLB patients exhibiting twice as many nocturnal sleep disturbances as AD (Grace et al., 2000; Bliwise et al., 2011). There is also some evidence to suggest measurable changes in sleep characteristics in DLB patients. Although research is limited, one study reported decreased stage 1 NREM sleep and increased stage 2 NREM sleep in DLB patients when compared with Parkinson disease patients (Terzaghi et al., 2013). Quantitative EEG studies in DLB have further reported EEG slowing as well as high variation and fluctuation of mean EEG power in wake (Petit et al., 2004; Walker et al., 2000). There is evidence of accumulation of synuclein pathology in the form of Lewy bodies in wake active neurons of various brain regions (LH, LC, TMN, NBM, DR) which could contribute to the sleep disorders and disturbances observed in DLB (Stern and Naidoo, 2015). Furthermore, like in AD and other neurodegenerative disorders, DLB patients show underlying circadian dysregulation. One study suggests that DLB patients have greater disruption in locomotor activity than AD patients and non-demented controls with significantly decreased diurnal activity (Harper et al., 2004). DLB also trended towards increased nocturnal activity compared to controls but remained less than AD (Harper et al., 2004). Temperature circadian phase occurred significantly later in both AD and DLB, although DLB patients showed lower core-body temperature amplitude than AD and was not significantly different from non-demented controls (Harper et al., 2004).

Outside of these generalized sleep and circadian disturbances, DLB is characterized by several specific sleep conditions that also aid in differentiating it from AD and other dementias including REM sleep behavior disorder (RBD), EDS, and periodic limb movement disorder (PLMD) (Borroni et al., 2008; Farina et al., 2009). RBD, a condition where patients physically ‘act out’ their vivid dreams during REM sleep, is now considered a part of the diagnostic criteria for DLB (Boeve et al., 1998; Irizaro et al., 2014; Ferman et al., 2011; McKeith et al., 2005). In fact, RBD can precede any dementia-like symptoms and can be considered to be the prodromal phase of DLB (Boeve et al., 2013; Donahgy and McKeith, 2014). Multiple longitudinal studies tracking frequency and emergence of neurodegenerative symptoms in patients diagnosed with RBD have found that a large portion of these patients go on to develop Parkinsonism or dementia. About 30–40% of patients were found to have Parkinsonism or dementia within 5 years of an RBD diagnosis and about 80% of RBD patients develop Parkinsonism or dementia within 12 years (Boeve et al., 2013; Schenck et al., 1996; Schenck et al., 2013; Postuma et al., 2009; Irizaro et al., 2006, 2013). Both LBD and RBD have been associated with loss of cholinergic neurons in the brain stem including the LC, PPT/LDT and substantia nigra (Dugger et al., 2012; Uchiyama et al., 1995). One study in dementia patients found that patients with DLB are much more likely to have RBD than patients with AD (Boeve et al., 2001). Although RBD is not a common feature of AD, it is interesting to note that patients with AD do show loss of neurons in LC and tau pathology is present in the PPT and LDT (Dugger et al., 2011; Brunström et al., 2011; Parvizi et al., 2001). PLMD is more commonly associated with restless leg syndrome in PD, but is also found separately in patients with DLB (Fantini et al., 2002). A study comparing patients with AD and DLB found a higher frequency of PLMD in DLB patients (Hibi et al., 2012). A lot remains unknown about PLMD, such as its etiology, true prevalence, and mechanism.

EDS is another sleep condition that although prevalent in AD, is shown to be more frequent in DLB (Roddy et al., 2007). As the name suggests, patients with EDS have difficulty maintaining wakefulness during the day. Like RBD, EDS is more prevalent in patients with DLB than AD as measured and confirmed by polysomnography and multiple sleep latency test (Roddy et al., 2007; Ferman et al., 2011). Although the mechanism(s) underlying EDS is not completely clear, there are some indications that orexinergic neurons in the lateral hypothalamus are lost in both DLB and AD, which might contribute to EDS in both diseases (Fronczek et al., 2007; Fronczek et al., 2012; Lessig et al., 2010). Furthermore, orexinergic neurons modulate the sleep-wake switch by virtue of their reciprocal actions on ascending arousal wakefulness neurons, many of which are known to be affected by AD and DLB pathology and could contribute to sleep disturbances in both diseases (Saper et al., 2010).

6. Conclusion

Sleep disturbances are prevalent in AD and may play a significant role in the advancement of pathology and disease progression. Although Aβ and amyloid pathology has been established to interact with sleep and circadian rhythms, tau pathology and also the co-morbidity of alpha-synuclein aggregation may be important contributors to sleep system dysfunction in AD. Tau pathology in the brainstem is the earliest AD-like pathological change that occurs during normal aging and abnormal tau pathology is present in many sleep centers of the brain even before the onset of amyloid pathology. Early studies in an htau expressing mouse model shows sleep disturbances with tau pathology alone. These observations combined with the high prevalence of sleep disorders in other tauopathies exemplifies the need to understand the interaction between tau and sleep in addition to and in conjunction with Aβ pathology.

Conflicts of interest

DMH co-founded and is on the scientific advisory board of C2N Diagnostics. DMH consults for Genentech, AbbVie, Eli Lilly, NeuroPhage, Biogen, and Denali. DMH receives research grants to Washington University from C2N Diagnostics, Eli Lilly, AbbVie, and Denali.
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