CDC6 is a prognostic biomarker and correlated with immune infiltrates in glioma

Feng Wang1,2†, Fen Zhao1,3†, Li Zhang1,2†, Lai Xiong1,1, Qing Mao1,1, Yanhui Liu1,1, Xiaoguang Qiu4,1,2, Xiang Wang1,2, Lin Shui1,1, Xi Chen1,1, Kexing Ren1,1,2, Pixian Shui1,1, Qiongwen Zhang1,2,1, Yifei Deng1,2, Weimin Li2,1, Xiaqi Xie3,4, Dengbin Wu9,1, Tao Li1,4, Jinyi Lang1,1, Lei Liu1,2, Huaying Chen1,2, Jianguo Xu1,1, Sen Bai1,1, Zhiping Li1,1, Qiang Yue1,1, Ni Chen1,1, Bingwen Zhou1,1, Cheng Yi4,1, Yuquan Wei5,1, Yuchuan Fu1,1, Yong Luo1,1, Qiheng Gou1,2,1, Lunxu Liu1,1, Yuanzhao Liu1,1, Jingbo Kang1,1, Junjie Wang1,1, Dongcun Jing1,1, Fuquan Zhang1,1, Xiaoyan Yang1,1, Xianfeng Li1,1, Tao Jiang4,1, Zongcun Zhang1,1, Yizhi Zhou1,1 and Junlin Yi1,1

Abstract

Background: Cell division cycle 6 (CDC6) has been proven to be associated with the initiation and progression of human multiple tumors. However, its role in glioma, which is ranked as one of the common primary malignant tumor in the central nervous system and is associated with high morbidity and mortality, is unclear.

Methods: In this study, we explored CDC6 gene expression level in pan-cancer. Furthermore, we focused on the relationships between CDC6 expression, its prognostic value, potential biological functions, and immune infiltrates in glioma patients. We also performed vitro experiments to assess the effect of CDC6 expression on proliferative, apoptotic, migratory, and invasive abilities of glioma cells.

Results: As a result, CDC6 expression was upregulated in multiple types of cancer, including glioma. Moreover, high expression of CDC6 was significantly associated with age, IDH status, 1p/19q codeletion status, WHO grade and histological type in glioma (all $p < 0.05$). Meanwhile, high CDC6 expression was associated with poor overall survival (OS) in glioma patients, especially in different clinical subgroups. Furthermore, a univariate Cox analysis showed that high CDC6 expression was correlated with poor OS in glioma patients. Functional enrichment analysis indicated that CDC6 was mainly involved in pathways related to DNA transcription and cytokine activity, and Gene Set Enrichment Analysis (GSEA) revealed that MAPK pathway, P53 pathway and NF-κB pathway in cancer were differentially enriched in glioma patients with high CDC6 expression. Single-sample gene set enrichment analysis (ssGSEA) showed CDC6 expression in glioma was positively correlated with Th2 cells, Macrophages and Eosinophils, and negative correlations...
with plasmacytoid dendritic cells, CD8 T cells and NK CD56bright cells, suggesting its role in regulating tumor immunity. Finally, CCK8 assay, flow cytometry and transwell assays showed that silencing CDC6 could significantly inhibit proliferation, migration, invasion, and promoted apoptosis of U87 cells and U251 cells \( (p<0.05) \).

**Conclusion:** In conclusion, high CDC6 expression may serve as a promising biomarker for prognosis and correlated with immune infiltrates, presenting to be a potential immune therapy target in glioma.

**Keywords:** CDC6, Glioma, Prognosis, Immune infiltrates, Biomarker

**Background**

Glioma, accounting for almost 80% of malignant brain tumors, is the most common primary malignant tumor in central nervous system (CNS) with high degree of mortality and malignancy [1]. The current standard regimens for glioma are mainly focused on maximizing safe surgical resection of tumor and assisting in adjuvant radiotherapy and chemotherapy, while high frequencies of recurrence and metastasis remain a huge challenge [2]. In recent years, although the application of immunotherapy, targeted therapy, photodynamic therapy and electric field therapy have improved the prognosis of glioma patients, the effects remain far from satisfactory [3]. For these reasons, the identification of key molecules involved in glioma is urgent and highly demanded for improving the clinical outcome.

Cell division cycle 6 (CDC6), mapped to chromosome 17q21.3, is an essential licensing factor for DNA replication during G1 phase and S phase in eukaryotic cells [4]. CDC6 also plays crucial roles in the development and maintenance of the S-M phase checkpoint mechanisms in the cell cycle [5]. Furthermore, many previous studies have shown that abnormal expression of CDC6 is involved in oncogenic activities in a variety of malignancies, and may be a potential diagnostic and prognostic marker for related tumors. For example, Kim et al. determined that elevated expression of CDC6 was highly correlated with poor prognosis of prostate cancer (PCa) [6]. Mahadevappa et al. reported that expression of CDC6 was significantly higher in breast cancer, especially in estrogen receptor (ER) negative breast cancer, suggesting that it may be a potential prognostic marker and therapeutic target in breast cancer patients [7]. While its role in glioma is unclear. Therefore, we aimed to demonstrate the relationship between CDC6 expression and glioma.

To clarify the role of CDC6 in glioma, we explored the relationship between CDC6 expression and glioma. Our results revealed that CDC6 was upregulated in glioma tissues and high CDC6 expression was correlated with poor prognosis and immune infiltrates in glioma patients. These findings suggest that CDC6 may be a potentially promising target by regulating its interaction with infiltrating immune cells in glioma patients.

**Results**

**Clinical characteristics of the glioma patients**

The microarray data of GSE104291 was downloaded from The Cancer Genome Atlas (TCGA) and Genotype-Tissue Expression (GTEx) database, including glioma (contained 689 tumor samples and 1157 normal samples). A total of 298 female and 398 male patients were involved in the present study, of which 20.5% \( (n=143) \) were over age 60. As for race, most of the cases 637 (93.3%) were white. The WHO grade included 224 (35.3%) G2, 243(38.3%) G3 and 168 (26.5%) G4. IDH (isocitrate dehydrogenase) status involved 246 (35.9%) with IDH-wide-type (IDH-WT) and 440 (64.1%) with IDH-mutant (IDH-MT). 1p/19q codeletion status included 171 (24.8%) codel and 518 (75.2%) non-codel. In terms of primary therapy outcome, 112 (24.2%) were progressive disease (PD), 147 (31.8%) patients were stable disease (SD), 64 (13.9%) patients were partial response (PR), and 139 (30.1%) patients were complete response (CR). Besides, as for histological type, 195(28%) of patients were astrocytoma, 134 (19.3%) of patients were oligoastrocytoma, 199 (28.6%) of patients were oligodendroglioma and 168 (24.1%) were glioblastoma.

From The Chinese Glioma Genome Atlas (CGGA), 325 tumors with both gene expression data and clinical futures were analyzed. The clinical characteristics of the glioma patients including age, gender, histology, grade, IDH status, 1p/19q codeletion status, chemo-status, radio-status, and MGMTp_methylation status and etc. Among the 325 participants, 203 (62%) were male and 122 (38%) were female, and the median age of all participants was 40.5 years.

**Abnormally high expression of CDC6 in glioma**

As shown in Fig. 1A, principal component analysis (PCA) plot showed that no batch effect was found in the GSE10429 dataset. After the R language limma package processed, a total of 3742 differentially expressed genes (DEGs) were identified between glioma tissues...
and normal tissues, including 2179 downregulated genes and 1563 upregulated genes. The volcano plots presented the expression of DEGs (Fig. 1B), and among them, the expression level of CDC6 was significantly upregulated in the GSE10429 dataset.

We analyzed the expression of CDC6 in 689 glioma samples of TCGA database and 1157 normal samples of TCGA database combined GTEx database, and found that CDC6 was significantly high expressed in glioma samples (Fig. 1C; \( p < 0.001 \)). We further expanded the number of samples to evaluate the mRNA expression level of CDC6 across pan-cancer in TCGA tumors with the data of the GTEx database as controls. As shown in Fig. 1D, compared with normal tissues, CDC6 was significantly upregulated in 28 of 33 cancer types, including bladder urothelial carcinoma (BLCA), breast invasive carcinoma (BRCA), glioblastoma multiforme (GBM), head and neck squamous cell carcinoma (HNSC), kidney renal clear cell carcinoma (KIRC), renal papillary cell carcinoma (KIRP), brain lower grade glioma (LGG), lung adenocarcinoma (LUAD), lung squamous cell carcinoma (LUSC), ovarian serous cystadenocarcinoma (OV), prostate adenocarcinoma (PRAD), stomach adenocarcinoma (STAD), oral squamous cell carcinoma (OSCC) and so on (all \( p < 0.05 \)), while it was downregulated in acute myeloid leukemia (LAML) (\( p < 0.05 \)), no difference was found in kidney chromophobe (KICH), pheochromocytoma and paraganglioma (PCPG), mesothelioma (MESO) and uveal melanoma (UVM). The results indicated that the mRNA expression of CDC6 was also highly expressed across multiple types of cancer.

**Relationship between expression of CDC6 and survival in glioma**

Kaplan–Meier analysis revealed that glioma patients from the CGGA dataset with high CDC6 expression was correlated with poor OS (Fig. 1E; \( p < 0.001 \)). Then, similar analysis in glioma patients from the CGGA dataset demonstrated that OS was significantly decreased in patients with high CDC6 mRNA expression compared with those with low CDC6 mRNA expression (Fig. 1F; \( p < 0.001 \)). Furthermore, we investigated the correlations between CDC6 expression and prognosis (OS) in different clinical subgroups of glioma from TCGA database. The results showed that the higher expression of CDC6 had a worse OS in following clinical subgroups, including subgroup of age \( \geq 60 \) years (Fig. 2A; \( p < 0.001 \)), subgroup of female (Fig. 2B; \( p < 0.001 \)), subgroup of primary therapy outcome (PD) (Fig. 2C; \( p < 0.001 \)), subgroup of IDH status (WT) (Fig. 2E; \( p < 0.01 \)) and subgroup of 1p/19q codeletion status (non-codel) (Fig. 2F; \( p < 0.001 \)). However, no difference in OS was observed upon histological type (Glioblastoma) (Fig. 2D; \( p = 0.167 \)).

**Association between CDC6 expression and clinicopathological features of glioma patients**

To evaluate the association between CDC6 expression and clinicopathological features of glioma patients, the clinical characteristics of 689 glioma patients including RNA sequencing data from TCGA database were analyzed. Based on the mean value of CDC6 expression, the patients were divided into high- and low-CDC6 expression groups, we performed Wilcoxon rank-sum test, Kruskal–Wallis test and logistic regression analysis. As shown in Fig. 2G-O, Our results showed that higher expression levels of CDC6 were observed in patients with age \( > 60 \) years (Fig. 2G; \( p < 0.001 \)), IDH status (WT) (Fig. 2J; \( p < 0.001 \)), 1p/19q codeletion status (non-codel) (Fig. 2K; \( p < 0.001 \)), WHO grade (G3/G4) (Fig. 2L; \( p < 0.001 \)), and worse histological type (Fig. 2M-O; \( p < 0.001 \)). However, no statistically significant correlation was found between the expression levels of CDC6 and other clinical pathological characteristics, such as gender (Fig. 2H; \( p > 0.05 \)) and race (Fig. 2I; \( p > 0.05 \)). These data indicate that high expression of CDC6 is significantly associated with age, IDH status, 1p/19q codeletion status, WHO grade and histological type, respectively.

Univariate analysis using logistic regression demonstrated that CDC6 expression was correlated with poor prognostic features in glioma patients. High CDC6 expression was significantly correlated with age \( > 60 \) vs. \( <= 60 \): \( \text{OR}=4.212, \text{95\%CI}=2.795–6.490, p < 0.001 \), WHO grade (G4&G3 vs. G2: \( \text{OR}=9.413, \text{95\%CI}=6.408–14.077, p < 0.001 \), IDH status (WT vs. MT: \( \text{OR}=15.032, \text{95\%CI}=10.032–23.106, p < 0.001 \), 1p/19q codeletion status (non-codel vs. codel: \( \text{OR}=5.413, \text{95\%CI}=3.638–8.230, p < 0.001 \).
Fig. 1 (See legend on previous page.)
p < 0.001), primary therapy outcome (PD vs. CR&PR&SD: OR = 2.656, 95%CI = 1.720–4.118, p < 0.001) and histological type (glioblastoma vs. astrocytoma&oligoastrocytoma&oligodendroglioma: OR = 31.688, 95%CI = 16.724–68.190, p < 0.001). However, there was no significant difference in overall survival (OS) upon gender and race (all p > 0.05). Taken together, these results indicate that the glioma patients with high CDC6 expression are associated with worse clinicopathological characteristics and may serve as a biomarker of poor prognosis.

Cox univariate and multivariate analysis of prognostic factors in glioma
Univariate and multivariate Cox proportional hazard regression analyses were carried out with glioma patients from TCGA database. The univariate analysis indicated that high CDC6 expression was associated with the worse OS (HR = 4.608; p < 0.001). Other clinical parameters, such as gender (HR = 1.262; p = 0.062), age (HR = 4.668; p < 0.001), WHO grade (HR = 18.615; p < 0.001), IDH status (HR = 4.608; p < 0.001), 1p/19q codeletion status (HR = 4.428; p < 0.001), primary therapy outcome (HR = 3.542; p < 0.001) and histological type (HR = 9.114; p < 0.001) were also correlated with the worse OS time. Following multivariate Cox analysis, the results showed that gender (HR = 1.945; p = 0.004), age (HR = 4.689; p < 0.001), WHO grade (HR = 4.879; p = 0.006), IDH status (HR = 0.544; p = 0.026) and primary therapy outcome (HR = 3.643; p < 0.001) were independent prognostic factors in OS of glioma patients. However, we could not exhibit statistical significance of CDC6 expression in OS by multivariate Cox analysis (HR = 1.470; p = 0.078).

Functional enrichment analysis of DEGs
To elucidate the biological functions of CDC6, we analyzed the DEGs between high- and low- CDC6 expression groups based on the median CDC6 expression level. A total of 1357 DEGs were acquired with the threshold values of adjusted p value (p.adj) < 0.05 and |log2 FC| > 2, including 1321 upregulated genes and 36 downregulated genes, that were presented in volcano plots. Then, we conducted Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses of DEGs, revealing that the primary biological process (BP) contained pattern specification process, embryonic organ development, regionalization, and anterior/posterior pattern specification. The cellular component (CC) was mainly enriched in collagen-containing extracellular matrix, kinetochore, condensed chromosome, centromeric region, and DNA packaging complex. The molecular function (MF) was primarily involved in receptor ligand activity, DNA-binding transcription activator activity, RNA polymerase II-specific, cytokine activity, and extracellular matrix structural constituent. Further functional enrichment analyses showed significantly enriched KEGG pathway in cytokine-cytokine receptor interaction, transcriptional misregulation in cancer, cell cycle, systemic lupus erythematosus and ECM-receptor interaction.

GSEA identifies CDC6-related signaling pathway
To identify CDC6-related signaling pathways in glioma, GSEA between high- and low- CDC6 expression datasets was conducted to reveal significant differences (p. adj < 0.05, false discovery rate (FDR) < 0.25) in enrichment of the Molecular Signatures Database (MSigDB) Collection (c5.bp.v7.2.symbols.gmt (Gene ontology) and c2.cp. v7.2.symbols.gmt (Curated)). In all, 3 GO items including single organism behavior, gated channel activity, cognition, transporter complex and ligand gated channel activity were showed significantly differential enrichment in CDC6 high expression phenotype. The results showed that the biological processes strongly associated with CDC6 were cell proliferation and immune-related pathways. 6 KEGG items that exhibited significantly differential enrichment in the CDC6-high expression phenotype were identified, including MAPK activation, DNA repair, P53 signaling pathway, focal adhesion, core matrisome and NF-kB activation. The results revealed that these pathways positively associated with CDC6 were responsible for regulation of cell proliferation/apoptosis and tumor invasion. Taking together, these findings suggest that these biological processes and signaling pathways of CDC6-high expression, which are critically important in development and metastasis of tumor, may be a potential target for the treatment of glioma.

The correlation between CDC6 expression and immune cell infiltration
We further explored the association between the expression level of CDC6 and immune cell infiltration level quantified by ssGSEA in glioma using Spearman correlation.
Fig. 2 (See legend on previous page.)
The results showed that CDC6 expression was positively correlated with infiltration levels of Th2 cells, Macrophages, Eosinophils, etc., and negatively correlated with that of plasmacytoid dendritic cells (pDCs), natural killer (NK) CD56 bright cells, etc.

**CDC6 promotes proliferation and inhibits apoptosis in glioma cells**

To explore the effect of CDC6 on glioma progression, U87 and U251 cells were transfected with sh-CDC6–1004, and the transfection efficiency was confirmed by western blot analysis. The results showed that the expression levels of protein in the sh-CDC6–1004 group was significantly lower than that in the sh-NC group. CCK-8 was used to assess role of CDC6 on glioma cell proliferation, and the results showed that the proliferative viability of U87 and U251 cells in the sh-CDC6–1004 group was markedly lower than that in the sh-NC group, particularly at 72h. Flow cytometry was further performed to analyze apoptosis of the transfected cells, the results indicated that, compared with the sh-NC group, the sh-CDC6–1004 group showed greater apoptosis of U87 and U251 cells. Our results suggest that CDC6 promotes proliferation and inhibits apoptosis of glioma cells.

**CDC6 facilitates migration and invasion of glioma cells**

To further clarify whether CDC6 affects the ability of migration and invasion of glioma cells, we performed the transwell assays. The results showed that the number of migrated and invaded U87 and U251 cells in the sh-CDC6–1004 group was markedly lower than that in the sh-NC group. These findings reversely suggest that CDC6 promotes migration and invasion of glioma cells.

**Discussion**

CDC6, DNA replication factor, which associates with DNA replication origins and is required for replication initiation; hence, it is closely associated with tumorigenesis and development. In recent years, many studies about association between CDC6 expression and prognosis of multiple tumors have emerged recently. For example, Zhang et al. found that CDC6 was aberrantly expressed in lung cancer tissues, and overexpression of CDC6 was associated with poor OS of lung cancer patients [8]. Mahadevappa et al. reported that high level of CDC6 expression was significantly associated with a poorer survival time in ER positive breast cancer [7]. Kim et al. determined that CDC6 mRNA expression was significantly higher in PCa tissues than that in controls and elevated expression of CDC6 was highly correlated with poor prognosis of patients with PCa [6]. Consistently, here we found that glioma patients with high CDC6 expression had a worse prognosis than those with low CDC6 expression, especially in certain clinical subgroups, such as age >60 years, female, primary therapy outcome (PD), IDH status (WT) and 1p/19q codeletion status (non-codeletion). In subgroup of Glioblastoma, our results showed that CDC6 was highly expressed in tumor tissues, while survival analysis showed no significant correlation between the expression level of CDC6 and prognosis of patients. Considering that there were only 168 glioblastomas in this subgroup, the sample size was small and may not have sufficient statistical efficacy, and the conclusions may be biased. Moreover, CDC6 expression level as a risk factor in OS of glioma through univariate Cox regression analyses, and by multivariate Cox analysis, we could not exhibit statistical significance of CDC6 expression in OS. Considering the current data derived from the TCGA public database, retrospective studies still have their own bias due to potential confounding factors. Therefore, a prospective study should be performed to verify the prognostic predictive role of CDC6 in glioma in the future. The results are further supported by the prior research involving prognostic significance of abnormally high expression of CDC6 in GBM [9], which identified high CDC6 expression as prognostic variables for the OS. These results together thus suggest that CDC6 may have value as a glioma prognostic biomarker.

Many studies about biological function and signaling pathways of glioma have emerged recently. Wang et al. and Li et al. reported that enhanced activity of NF-κB signaling pathway promoted growth of GBM in vivo and induced ferroptosis of glioma cell lines [10]. Krex et al. reported that the p53 pathway was highly deregulated in GBM, the mutational status of TP53 was related with GBM growth [11]. Through GSEA, our study revealed that NF-κB signaling pathway, MAPK pathway and P53 pathway were differentially enriched in glioma patients with high CDC6 expression phenotype, indicating that CDC6 may promote glioma cell growth and migration via these pathways. Those all suggest that CDC6 may serve as a potential therapeutic target in glioma.

Tumor-infiltrating immune cells (TIICs) are indispensable component of the tumor microenvironment (TME), which play an important role in the growth and progression of tumors. In recent years, a large number of studies about the possible role of CDC6 in human TIICs have emerged. Cong et al. reported that CDC6 was related to B cell, T cell infiltration, macrophage infiltration and dendritic cell (DC) infiltration, and may be a prognostic factor for Clear Cell Renal Cell Carcinoma patients [12]. However, the correlation between CDC6 expression and immune cell infiltration in glioma has not been investigated. In present study, the results revealed that CDC6 expression was positively correlated with infiltration abundances
of Th2 cells, Macrophages and Eosinophilis, and negatively correlated with that of pDCs, CD8 T cells and NK CD56bright cells. These correlations could be indicative of a potential mechanism by which CDC6 inhibits the function of pDCs, CD8 T cells and NK CD56bright cells, subsequently promotes the function of Th2 cells, Macrophages, and Eosinophilis, which are responsible for maintaining the immunosuppressive local microenvironment of tumor, and thus contribute to the development and progression of tumor. These findings suggest that CDC6 may be a potentially promising target by regulating its interaction with infiltrating immune cells in glioma patients. However, the detailed underlying mechanisms still need to be further explored.

Abnormal and unrestricted cell growth is a hallmark of cancer and is caused by the misregulation of various crucial protein expression, which leads to the occurrence, progression and recurrence of glioma patients [13]. CDC6, a chromosome replication licensor in the cell cycle, plays a major role in cell proliferation, migration, invasiveness and tumor metastasis in several cancers. Deng et al. demonstrated that CDC6 protein level was highly expressed in ovarian cancer tissues, silencing CDC6 decreased cell proliferation and colony formation in HO8910 cells [14]. Zhao et al. found that CDC6 was highly expressed in gastric cancer cells, down-regulation of CDC6 inhibited cell proliferation, invasion, and promoted apoptosis of BGC823 and SGC7901 cell lines [15]. The present study demonstrated that the CDC6 protein level in U87 and U251 cells was higher than that in normal cell, and silencing CDC6 inhibited proliferation, migration and invasion, and promoted apoptosis of glioma cells. The result suggests that CDC6 may act as a novel oncogenic in glioma and may be a potential therapeutic target.

Although the present study suggested some correlations between CDC6 expression and glioma, there are still some limitations in this study. Firstly, number of healthy samples used as controls differed significantly from that of tumor samples, hence additional studies were required to maintain a balance of sample size. Secondly, the current study was performed primarily based on bioinformatic analyses and in vitro experiments, further more clinical samples of glioma patients are required to verify the abnormally expression of CDC6. Meanwhile, it is necessary to further elucidate the biological functions of CDC6 in glioma and the underlying molecular mechanisms in subsequent experiments. Last but not most, retrospective studies still have their own bias caused by potential confounding factors because of data from public databases, especially non-uniform intervening measures and lacking of some detailed information; herefore, a prospective study should be performed in the future to avoid analysis bias.

Conclusion
Our results showed that CDC6 expression was highly expressed in glioma tissues, and high CDC6 expression was significantly correlated with poor survival and immune infiltration of glioma, which might promote tumor tumorigenesis through abnormal inflammation and immune response of glioma patients. In addition, CDC6 was mainly enriched signaling pathways such as NF-κB signaling pathway, MAPK pathway and P53 pathway, therefore CDC6 may be involved in the progression of glioma. We also found that CDC6 promotes the proliferative, migratory and invasive capabilities of glioma cells, and inhibits apoptosis of glioma cells. Collectively, our results suggest that CDC6 may serve as a promising biomarker for prognosis in glioma and correlated with immune infiltrates, presenting to be a potential immune therapy target in glioma.

Abbreviations
CDC6: Cell division cycle 6; CGGA: The Chinese Glioma Genome Atlas; CNS: central nervous system; CR: complete response; DEGs: differentially expressed genes; FDR: false discovery rate; GBM: glioblastoma; GO: Gene Ontology; GEO: The Gene Expression Omnibus; GSEA: Gene Set Enrichment Analysis; GTEx: Genotype-Tissue Expression; HR: hazard ratio; IDH: isocitrate dehydrogenase; KEGG: Kyoto Encyclopedia of Genes and Genomes; MSigDB: Molecular Signatures Database; NES: Normalized enrichment scores; NK: natural killer; OS: overall survival; p.adj: adjusted p value; PCA: principal component analysis; PD: progressive disease; pDCs: plasmacytoid dendritic cells; PR: partial response; SD: stable disease; ssGSEA: Single-sample gene set enrichment analysis; TCGA: The Cancer Genome Atlas; TME: tumor microenvironments; WB: Western blot.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s12943-022-01623-8.

Additional file 1.

Acknowledgements
We thank Professor Ying Huang for suggestions and comments on the manuscript. We also thank Lei Dai for technical support.

Authors’ contributions
FW conceived and conducted the experiments. FZ, LX and LZ analysed the results and wrote the manuscript. FZ, LZ and JWW provided computational and statistical insight. QY and XXQ prepared all figures. FW, DBW and CY supervised the study and extensively revised the manuscript. All authors reviewed and approved the final manuscript.

Funding
The present study was financially supported by the National Natural Science Foundation of China.
Availability of data and materials
The results within this publication are in part based upon data generated by the GEO database (http://www.ncbi.nlm.nih.gov/geo), TCGA and GTEx database by UCSC XENAGenome (https://xenabrowser.net/datapages/), and CGGA database (http://www.cgga.org.cn/). In addition, all data used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Consent of publication
Not applicable.

Ethics approval and consent to participate
Not applicable.

Competing interests
The authors declare no competing interests.

Author details
1 Department of Internal Medicine, West China Hospital Cancer Center Head And Neck, Sichuan University, Chengdu, China. 2 Department of Cancer Center Head and Neck, West China Hospital, Sichuan University, Chengdu, China.
3 Department of Oncology, Chengdu First People’s Hospital, Chengdu 610041, Sichuan Province, China. 4 Department of Radiotherapy, Beijing Tian Tan Hospital, Capital Medical University, Beijing 100050, China. 5 Department of Pharmacy, Affiliated Hospital of Southwest Medical University, Luzhou, China. 6 Department of Radiotherapy, Chengdu Seventh Hospital, Chengdu, China. 7 Center for Precision Medicine, West China Hospital, Sichuan University, Chengdu, China. 8 Department of Critical Care Medicine, West China Hospital, Sichuan University, Chengdu, China. 9 Cancer Hospital, An Steel Group General Hospital, Anshan, Liao Ning, People’s Republic of China. 10 Department of Radiotherapy, Sichuan Cancer Hospital & Institute, Sichuan Cancer Center, School of Medicine, University of Electronic Science and Technology of China, Chengdu 610041, Sichuan, China. 11 Department of Radiotherapy, Beijing Hospital, Beijing, People’s Republic of China. 12 Department of Radiotherapy, The sixth Medical Center of PLA General Hospital, Beijing, People’s Republic of China.
13 Department of Radiation Oncology, Peking University Third Hospital, No. 49, Beijing, China. 14 Kawagawa Hospital Tsukiji, Fukutai, Japan. 15 Department of Radiation Oncology, Beijing Union Medical College Hospital, Beijing, China. 16 Department of Radiotherapy, First Hospital of Shan Xi Medical University, Taiyuan West, Taiyuan, China. 17 Qing Dao Central Hospital, 127 Si Liu South Road, Shi Bei District, Qing Dao, Shan Dong Province, China. 18 Shanghai High-Tech United Bio-Technological R&D Co., Ltd, Shanghai, China. 19 Department of Radiation Oncology, National Cancer Center National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China.

Received: 24 June 2022 Accepted: 12 July 2022
Published online: 25 July 2022

References
1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin. 2020;70:7–30.
2. Li G, Zhai Y, Wang Z, Wang Z, Huang R, Jiang H, et al. Postoperative standard chemoradiotherapy benefits primary glioblastoma patients of all ages. Cancer Med. 2020;9:1935–45.
3. Wang H, Xu T, Huang Q, Jin W, Chen J. Immunotherapy for malignant Glioma: current status and future trends. Trends Pharmacol Sci. 2020;41:123–38.
4. Lau E, Zhu C, Abraham RT, Jiang W. The functional role of Cdc6 in S-G2/M in mammalian cells. EMBO Rep. 2006;7:425–30.
5. Borlado LR, Mendez J. Cdc6: from DNA replication to cell cycle checkpoints and oncogenesis. Carcinogenesis. 2008;29:237–43.
6. Kim YH, Byun YJ, Kim WT, Jeong P, Yan C, Kang HW, et al. Cdc6 mRNA expression is associated with the aggressiveness of prostate Cancer. J Korean Med Sci. 2018;33:e303.
7. Mahadevappa R, Neves H, Yuen SM, Bai Y, McCruden CM, Yuen HF, et al. The prognostic significance of Cdc6 and Cdt1 in breast cancer. Sci Rep. 2017;7:985.
8. Zhang X, Xiao D, Wang Z, Zou Y, Huang L, Lin W, et al. MicroRNA-26a/b regulate DNA replication licensing, tumorigenesis, and prognosis by targeting Cdc6 in lung cancer. Mol Cancer Res. 2014;12:1535–46.
9. Zhao H, Zhou X, Yuan G, Hou Z, Sun H, Zhai N, et al. Cdc6 is up-regulated and a poor prognostic signature in glioblastoma multiforme. Clin Transl Oncol. 2021;23:565–71.
10. Li S, He Y, Chen K, Sun J, Zhang L, He Y, et al. RSL3 drives Ferroptosis through NF-kappaB pathway activation and GPX4 depletion in Glioblastoma. Oxidative Med Cell Longev. 2021;2021:2915019.
11. Krex D, Mohr B, Appelt H, Schackert HK, Schackert G. Genetic analysis of a multifocal glioblastoma multiforme: a suitable tool to gain new aspects in glioma development. Neurosurgery. 2003;53:1377–84.
12. Yicong Y, Wang Y, Denglong W, Baoying H. Increased Cdc6 expression associates with poor prognosis in patients with clear cell renal cell carcinoma. Front Oncol. 2021;11:666418.
13. Zhu L, Zheng Y, Hu R, Hu C. CKA2PL as an independent risk factor, closely related to the prognosis of Glioma. Biomed Res Int. 2021;2021:5486131.
14. Deng Y, Jiang L, Wang Y, Xi Q, Zhong J, Liu J, et al. High expression of Cdc6 is associated with accelerated cell proliferation and poor prognosis of epithelial ovarian cancer. Pathol Res Pract. 2016;212:239–46.
15. Zhao B, Zhang J, Chen X, Xu H, Huang B. Mir-26b inhibits growth and resistance to paclitaxel chemotherapy by silencing the Cdc6 gene in gastric cancer. Arch Med Sci. 2019;15:498–503.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.