Research Article

Dexmedetomidine Ameliorates Lung Injury Induced by Intestinal Ischemia/Reperfusion by Upregulating Cannabinoid Receptor 2, Followed by the Activation of the Phosphatidylinositol 3-Kinase/Akt Pathway

Meng Chen,1 Xue-Tao Yan,2 Li Ye,1 Jun-Jiao Tang,1 Zong-Ze Zhang,1 and Xiang-Hu He1

1Department of Anesthesiology, Zhongnan Hospital of Wuhan University, Wuhan, Hubei 430071, China
2Department of Anesthesiology, Shenzhen Bao’an Maternity and Child Health hospital, Shenzhen 518133, China

Correspondence should be addressed to Xiang-Hu He; hexh1220@aliyun.com

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Intestinal ischemia/reperfusion (I/R) is a clinical emergency, which often causes lung injury with high morbidity and mortality. Although dexmedetomidine has been identified to have a protective effect on lung injury caused by intestinal I/R, its specific mechanism is still elucidated. In recent years, the cannabinoid (CB2) receptor pathway has been found to be involved in I/R injury of some organs. In the current study, we investigated whether the CB2 receptor pathway contributes to the protective effect of dexmedetomidine on the intestinal I/R-induced lung injury in rats. Dexmedetomidine treatment upregulated the expression of CB2 receptor and suppressed the I/R-induced increases in lung injury scores, inflammatory cell infiltration, lung wet/dry ratio, MPO activity, MDA level, inflammatory cytokines, and caspase-3 expression while augmenting SOD activity and Bcl-2 expression, indicating attenuation of lung injury. Dexmedetomidine treatment also increased the expression of Akt. The protective effects of dexmedetomidine treatment were reversed by the CB2 receptor antagonist AM630 or the PI3K inhibitor wortmannin. And the CB2 receptor antagonist AM630 also downregulated the expression of Akt. Thus, our findings suggest that treatment with dexmedetomidine provides a protective role against lung injury caused by intestinal I/R in rats, possibly due to the upregulation of the CB2 receptor, followed by the activation of the PI3K/Akt pathway.

1. Introduction

Intestinal ischemia/reperfusion (I/R) is a clinical emergency frequently occurring in multiple clinical conditions, including acute mesenteric ischemia, abdominal aortic aneurysm surgery, small bowel transplantation, shock, and cardiopulmonary bypass [1]. In addition to causing local intestinal damage, I/R is often followed by distant organ injury, especially lung injury, associated with high morbidity and mortality [2]. Although the exact mechanism is poorly understood, some factors, such as oxidative stress, activated neutrophils, complement components, released inflammatory cytokines, and cell apoptosis, are believed to be involved in the development of I/R-induced lung injury [3–5].

Dexmedetomidine (DEX), a highly selective alpha-2 adrenoceptor (α2AR) agonist, exhibits anesthetic-sparing, analgesia, and sympatholytic properties and is widely used as a sedative agent in clinical anesthesia and ICU management [6]. Previous studies have identified DEX as having protective effects in some models of I/R injury, such as in the liver [7, 8], lung [9], heart [10], kidney [11], and intestine [12] because of its antioxidant, anti-inflammatory, and anti-apoptotic properties. Further research has shown that some molecular mechanisms, such as α2AR/phosphatidylinositol 3-kinase (PI3K)/Akt [13], HMGB1-TLR4-MyD88-NF-κB [14], and JAK/STAT [15], are involved in the process of protection of DEX against some organ I/R injuries. It has also been demonstrated that treatment with DEX mitigated lung...
injury caused by various factors, including intestinal I/R [16]. Although the protective effect of DEX treatment on intestinal I/R-induced lung injury has been investigated, the potential molecular mechanism by which DEX ameliorates lung injury remains unclear.

Recently, the role of cannabinoid (CB) receptors, especially the CB₂ receptor, has attracted much attention in health and disease. Although the CB₂ receptor was first identified in the peripheral immune system, it is now expressed in most organ systems, including in the cells of the brain, heart, liver, and cardiovascular and gastrointestinal (GI) systems [17]. Early studies focused mainly on the role of the CB₂ receptor in fundamental physiological processes. However, accumulating evidence supports that the CB₂ receptor has a critical role in regulating many disease processes, including inflammation [18], cancer [19], pain [20], and I/R injury [21], with it shown that the CB₂ agonist, HU-910, protected against hepatic I/R injury by attenuating oxidative stress, inflammatory response, and apoptosis [22]. Using the selective CB₂ agonist, JWH-133, and CB₂ gene-deficient mice, Defer et al. [23] also revealed that activating the CB₂ receptor promoted the survival of cardiac myocytes and protected against I/R-induced myocardial infarction through the direct inhibition of myocyte and fibroblast death and the prevention of myofibroblast activation. The role of the CB₂ receptor in the GI tract has been investigated, including in regulating abnormal motility, modulating intestinal inflammation, and limiting visceral sensitivity and pain [18, 24]. However, it remains unclear whether the CB₂ receptor is involved in the intestinal I/R-induced lung injury.

Thus, we hypothesized that DEX might improve intestinal I/R-induced lung injury by activating the CB₂ receptor pathway. In this study, we established a rat model of lung injury induced by intestinal I/R and investigated the effect of DEX on intestinal I/R-induced lung injury and its potential mechanisms.

2. Materials and Methods

2.1. Animals. Male Sprague-Dawley rats weighing 220-270 g were obtained from the Department of Laboratory Animal Center of the Wuhan University and housed under standardized conditions of food, water, light, and temperature. All animals were subjected to fasting for 12 h before experiments, but they were allowed free access to tap water throughout the experimental procedure. All procedures were approved by the Research Committee on Ethics of the Zhongnan Hospital of Wuhan University.

2.2. Drugs. DEX was acquired from Jiangsu Nhwa Pharmaceutical Corporation Ltd. (Jiangsu, China). The CB₂ antagonist AM630 was obtained from Tocris (Baldwin, MO, USA). The PI3K inhibitor wortmannin was purchased from Sigma-Aldrich (St. Louis, MO, USA). AM630 and wortmannin were dissolved in DMSO. DMSO alone was used in control experiments.

2.3. Experimental Protocol. Fifty-six male Sprague-Dawley rats were randomly assigned to the following seven groups (n = 8): sham operation (Sham) group, intestinal I/R (I/R) group, dexametomidine-treated I/R (DEX) group, dexametomidine- and AM630-treated I/R (DA) group, AM630-treated I/R (AI) group, dexametomidine- and wortmannin-treated I/R (DW) group, and wortmannin-treated I/R (WI) group. Animals in the DEX, DA, and DW groups were infused continuously with dexametomidine (i.v.) at 5 μg·kg⁻¹·h⁻¹ for 1 h before ischemia. This dose was chosen based on the results from a previous study [12]. Animals in the DA, AI, DW, and WI groups were given the CB₂ receptor antagonist, AM630 (3 mg/kg, i.p.), or the PI3K inhibitor, wortmannin (15 μg/kg, i.v.), before the injection of dexametomidine. Animals in the Sham or I/R groups were given an equal volume of the vehicle at the same time point.

2.4. Surgical Procedure. The model of intestinal I/R injury was established as we previously described [25]. Each animal was anesthetized with an intraperitoneal injection of sodium pentobarbital (50 mg/kg). An upper midline laparotomy was performed to expose the abdomen and the superior mesenteric artery (SMA). An atrumatic microvascular clamp was applied to the SMA for 1 h of ischemia induction, after which the occluding clamp was removed for 2 h of reperfusion. Sham-operated animals underwent the same surgical procedure as the experimental rats, except for the induction of I/R injury. The animals were sacrificed with pentobarbital overdose, and lung samples were harvested after reperfusion and used to assess biochemical parameters or fixed in buffered formalin for histopathological evaluation.

2.5. Histological Assessment. Lung samples were immersed in 10% formalin and then embedded in paraffin. Tissue blocks were cut into 5 μm sections and stained with hematoxylin and eosin (HE). The morphologic examination of lung tissues was carried out under a light microscope by two pathologists blinded to the experiment. The histological injury was graded using our previously described method [26]: grade 0, no diagnostic change; grade 1, mild neutrophil infiltration and mild to moderate interstitial congestion; grade 2, moderate neutrophil infiltration, perivascular edema formation, and partial destruction of the pulmonary architecture; and grade 3, dense neutrophil infiltration and the complete destruction of the pulmonary architecture.

2.6. Assessment of Pulmonary Edema. The left lower lung lobe was removed at the end of reperfusion, weighed, and then dried in an oven at 80°C for 24 h. The wet weight to dry weight (W/D) ratio of the lung tissue was calculated and used as an indicator for estimating pulmonary edema.

2.7. Assay of Myeloperoxidase Activity. Frozen lung tissues were homogenized on ice using a homogenizer. The myeloperoxidase (MPO) activity was measured according to the manufacturer’s instructions (Jiancheng Biologic Project Company, Nanjing, China). One MPO activity unit was defined as the amount of lung tissue that converted 1 μmol of hydrogen peroxide to water per minute at 37°C. The MPO activity was expressed in units per gram (U/g) of wet weight.
2.8. Assay of Lipid Peroxidation and Superoxide Dismutase Activity. Frozen lung tissues were homogenized on ice using a homogenizer and centrifuged at 4000 g for 10 min at 4°C. Malondialdehyde (MDA) level and superoxide dismutase (SOD) activity in the supernatant were determined according to the manufacturer’s instructions using commercial kits (Jiancheng Bioengineering Institute, Nanjing, China). The SOD activity was expressed in U/mg protein, and the MDA level was expressed in nmol/mg protein.

2.9. Assay of Lung Tissue Cytokines. Frozen lung tissues were homogenized on ice using a homogenizer and centrifuged at 4000 g for 10 min at 4°C. Levels of tumor necrosis factor alpha (TNF-α) and interleukin 6 (IL-6) in the supernatant were assayed according to the manufacturer’s instructions using ELISA kits (R&D Systems, Minneapolis, MN, USA). The concentrations of TNF-α and IL-6 were expressed in pg/mg protein.

2.10. Immunohistochemistry. Immunohistochemistry was conducted on a StepOne Real-Time PCR instrument (Life Technologies, Grand Island, NY). The primers used for RT-PCR were as follows: caspase-3 forward 5′-actactgcggatctgact-3′; reverse 5′-taacgcggcctgagata-3′; Bax forward 5′-gaacgctatggcctgaca-3′; reverse 5′-gttgaagcgtgacagac-3′; Bcl-2 forward 5′-cctcttcgctgcacctg-3′; reverse 5′-gggggtcacctcctgttg-3′; Akt forward 5′-ggagaactctggctgacag-3′; reverse 5′-gggggttacctcctgttg-3′; GAPDH forward 5′-tgtggtgctgacacgag-3′; reverse 5′-gggttgctgacacgag-3′. PCR conditions were as follows: 95°C for 5 min; 35 cycles at 95°C for 20s, 60°C for 20s, and 72°C for 20s. The PCR products were separated on 2% agarose gel electrophoresis, then transferred to polyvinylidene fluoride membranes (Millipore, Bedford, MA, USA) that were incubated with primary antibodies against cleaved caspase-3, Bax, Bcl-2 (Santa Cruz Biototechnology, USA), and p-Akt (Cell Signaling Technology, USA) overnight at 4°C. The membranes were washed with PBS-0.05% Tween 20 the next day and incubated for 1 h in horseradish peroxidase-conjugated secondary antibody (Jackson ImmunoResearch). Another wash of the membranes with PBS-0.05% Tween 20 followed the 1 h incubation period, at the end of which protein bands were visualized using an enhanced chemiluminescence kit (Amersham, Piscataway, NJ, USA). All band densities were quantified by densitometry using the Quantity One software (BioRad, Hercules, CA, USA).

2.11. Real-Time Quantitative Reverse Transcription-PCR. The total RNA of rat lung tissues was extracted using TRlipol reagent according to the manufacturer’s instructions (Takara, Japan). A real-time quantitative reverse transcription-PCR (RT-PCR) analysis was performed using a SYBR Premix Ex Taq™ Kit (Takara, Japan), and the reactions were conducted on a StepOne™ Real-Time PCR instrument (Life Technologies, Grand Island, NY). The primers used for PCR were as follows: caspase-3 forward 5′-actactgcggatctgact-3′; reverse 5′-taacgcggcctgagata-3′; Bax forward 5′-gaacgctatggcctgaca-3′; reverse 5′-gttgaagcgtgacagac-3′; Bcl-2 forward 5′-cctcttcgctgcacctg-3′; reverse 5′-gggggtcacctcctgttg-3′; Akt forward 5′-ggagaactctggctgacag-3′; reverse 5′-gggggttacctcctgttg-3′; GAPDH forward 5′-tgtggtgctgacacgag-3′; reverse 5′-gggttgctgacacgag-3′. PCR conditions were as follows: 95°C for 5 min; 35 cycles at 95°C for 20s, 60°C for 20s, and 72°C for 45 s. GAPDH was selected as an internal control, and the target gene expression was normalized to GAPDH expression and calculated using the 2ΔΔCt method.

2.12. Western Blot Analysis. Frozen lung samples were homogenized and centrifuged at 12,000 g for 10 min at 4°C. The protein concentrations in the supernatant were measured using a BCA protein assay kit (Beyotime Institute of Biotechnology, Shanghai, China). Equal amounts of lysate were subjected to 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis, then transferred to polyvinylidene fluoride membranes (Millipore, Bedford, MA, USA) that were incubated with primary antibodies against cleaved caspase-3, Bax, Bcl-2 (Santa Cruz Biototechnology, USA), and p-Akt (Cell Signaling Technology, USA) overnight at 4°C. The membranes were washed with PBS-0.05% Tween 20 the next day and incubated for 1 h in horseradish peroxidase-conjugated secondary antibody (Jackson ImmunoResearch). Another wash of the membranes with PBS-0.05% Tween 20 followed the 1 h incubation period, at the end of which protein bands were visualized using an enhanced chemiluminescence kit (Amersham, Piscataway, NJ, USA). All band densities were quantified by densitometry using the Quantity One software (BioRad, Hercules, CA, USA).

2.13. Statistical Analysis. All data are expressed as mean values ± SEM. Differences among the groups were analyzed using the one-way analysis of variance (ANOVA), followed by the Student-Newman-Keul (SNK) test for multiple comparisons. Differences were considered significant at P < 0.05.

3. Results

3.1. The Effects of Dexmedetomidine Treatment on the Histological Injury. Lung histological injury was evaluated and scored by two pathologists to investigate the role of DEX treatment against lung injury caused by intestinal I/R; representative morphological changes are presented in Figure 1. No significant morphological changes were observed in the Sham group (Figure 1(a)). I/R induced obvious lung tissue injury, manifested by the apparent destruction of the pulmonary architecture, interstitial edema, hemorrhage, and massive neutrophil infiltration (Figure 1(b)). The use of DEX attenuated lung injury significantly, with mild neutrophil infiltration and interstitial congestion (Figure 1(c)). Significant morphological changes were observed in the other four groups (Figures 1(d)–1(g)). The average scores were used for analysis (Figure 1(h)). These data showed that the lung injury scores in the I/R group were higher than those in the Sham group (P < 0.05) and were attenuated by DEX treatment in the DEX group (P < 0.05) but not in the other four groups (P > 0.05).

3.2. The Effects of Dexmedetomidine Treatment on Lung Tissue W/D Weight Ratio. The W/D weight ratio is used as an important indicator of lung tissue damage. As presented in Figure 2(a), the lung tissue W/D weight ratio was higher in all groups that underwent intestinal I/R than in the Sham group (P < 0.05). DEX treatment reduced the lung tissue W/D weight ratio compared to that of the I/R rats (P < 0.05). However, the protective role of DEX treatment against lung injury caused by intestinal I/R was reversed by the administration of the CB2 receptor antagonist, AM630, or the P13K inhibitor, wortmannin (P < 0.05). There was no significant difference in the lung tissue W/D weight ratio in the animals treated with AM630 alone or wortmannin alone compared with animals in the I/R group (P > 0.05).
Figure 1: Continued.
3.3. The Effects of Dexmedetomidine Treatment on MPO Activity. MPO activity is an indicator of neutrophil infiltration. We assessed the MPO activity to investigate the role of neutrophil infiltration in intestinal I/R-induced lung injury. As shown in Figure 2(a), I/R increased the MPO activity in lung tissues compared to that of the sham-operating rats (P < 0.05). Treatment with DEX decreased the MPO activity in lung tissues compared to that of the I/R rats (P < 0.05). In contrast, treatment with both AM630 and wortmannin reversed the decrease in MPO activity compared to that of the DEX rats (P < 0.05). Treatment with AM630 alone or wortmannin alone had no significant effect on MPO activity compared to that of the I/R rats (P > 0.05).

3.4. The Effects of Dexmedetomidine Treatment on SOD Activity and MDA Level. Oxidative stress is one of the main causes of lung injury induced by intestinal I/R. We assessed the level of MDA, an indicator of lipid peroxidation damage, and the activity of SOD, an antioxidant enzyme, to examine...
the role of oxidative stress in lung injury. As shown in Figure 3, I/R increased the MDA level and decreased the SOD activity compared to those of the sham-operating rats ($P < 0.05$). DEX treatment led to a decrease in MDA level and an increase in SOD activity ($P < 0.05$). However, the protective effect of DEX treatment on lung injury was reversed by the application of AM630 or wortmannin ($P < 0.05$). Treatment with AM630 alone or wortmannin alone had no significant impact on MDA level and SOD activity compared to those of the I/R rats ($P > 0.05$).

3.5. The Effects of Dexmedetomidine Treatment on the Levels of TNF-$\alpha$ and IL-6. We assessed the levels of TNF-$\alpha$ and IL-6 in the lung to evaluate the inflammatory cytokines in the lung injury caused by intestinal I/R. As shown in Figure 4, I/R led to an increase in the levels of TNF-$\alpha$ and IL-6 compared to those of the sham-operating rats ($P < 0.05$). Treatment with DEX decreased the levels of TNF-$\alpha$ and IL-6 compared to those of the I/R rats ($P < 0.05$). In contrast, treatment with AM630 or wortmannin reversed the protective role of DEX by increasing the levels of TNF-$\alpha$ and IL-6 ($P < 0.05$). AM630 alone or wortmannin alone had no significant impact on the levels of TNF-$\alpha$ and IL-6 compared to those of the I/R rats ($P > 0.05$). These results suggest that DEX treatment has a protective role against I/R-induced lung injury by reducing cytokine production.
3.6. The Effects of Dexmedetomidine Treatment on Macrophage Infiltration and p-Akt Expression in Lung Tissue by Immunohistochemical Assay. To assess macrophage infiltration in the lungs, we detected F4/80 expression by immunohistochemistry. Dexmedetomidine treatment reduced macrophage infiltration in the lung tissues following I/R. (a) Sham group. (b) Intestinal ischemia/reperfusion (I/R) group. (c) Dexmedetomidine-treated I/R (DEX) group. (d) Dexmedetomidine- and AM630-treated I/R (DA) group. (e) AM630-treated I/R (AI) group. (f) Dexmedetomidine- and wortmannin-treated I/R (DW) group. (g) Wortmannin-treated I/R (WI) group (original magnification, ×400). (h) Integrated optical density (IOD) of F4/80 expression in all groups. Data are expressed as mean ± SEM, n = 6 rats per group. *P < 0.05 versus the Sham group, #P < 0.05 versus the I/R group, &P < 0.05 versus the DEX group.

Figure 5: Immunohistochemical analysis of macrophage infiltration in the lung tissues of all groups. Macrophage infiltration was investigated by detecting F4/80 expression with immunohistochemistry. Dexmedetomidine treatment reduced macrophage infiltration in the lung tissues following I/R. (a) Sham group. (b) Intestinal ischemia/reperfusion (I/R) group. (c) Dexmedetomidine-treated I/R (DEX) group. (d) Dexmedetomidine- and AM630-treated I/R (DA) group. (e) AM630-treated I/R (AI) group. (f) Dexmedetomidine- and wortmannin-treated I/R (DW) group. (g) Wortmannin-treated I/R (WI) group (original magnification, ×400). (h) Integrated optical density (IOD) of F4/80 expression in all groups. Data are expressed as mean ± SEM, n = 6 rats per group. *P < 0.05 versus the Sham group, #P < 0.05 versus the I/R group, &P < 0.05 versus the DEX group.

3.6. The Effects of Dexmedetomidine Treatment on Macrophage Infiltration and p-Akt Expression in Lung Tissue by Immunohistochemical Assay. To assess macrophage infiltration in the lungs, we detected F4/80 expression by immunohistochemistry. As shown in Figure 5. In the Sham group, there were only a few positive staining in lung sections. Remarkable strong staining was observed in the rats subjected to I/R, and the expression of F4/80 in the I/R group, DEX group, DA group, AI group, DW group, and WI group was higher than that in the Sham group (P < 0.05). In contrast, treatment with DEX decreased the expression of F4/80 compared to that of the I/R rats (P < 0.05). Treatment with AM630 or wortmannin increased the expressions of F4/80 compared to that of the DEX-treated rats (P < 0.05). AM630 alone or wortmannin alone had no significant impact on the expressions of F4/80 compared to that of the I/R rats (P > 0.05). This suggests that treatment with DEX may inhibit macrophage infiltration.

The lung tissues were obtained to measure the expression of p-Akt by immunohistochemistry, as shown in Figure 6. The results of immunohistochemistry showed that the level of p-Akt expression was low in the Sham group. I/R increased p-Akt expression, but the differences in the values between the Sham and I/R groups were not statistically significant
Compared with the I/R group, the expression of p-Akt was enhanced in the DEX group ($P < 0.05$). The increase of p-Akt expression caused by DEX treatment was reversed by treatment with AM630 or wortmannin ($P < 0.05$). Compared with the I/R group, AM630 alone or wortmannin alone had no significant impact on p-Akt expression ($P > 0.05$).

### 3.7. The Effects of Dexmedetomidine Treatment on the Expressions of I/R Injury-Induced Apoptosis in Lung Tissues

Cell apoptosis is one of the main causes of lung injury caused by intestinal I/R. To investigate the role of cell apoptosis in lung injury caused by intestinal I/R, we evaluated the expressions of proapoptotic and antiapoptotic mRNA and proteins in lung tissues using RT-PCR and Western blot analysis. As shown in Figures 7(a)–7(c) and 8(a)–8(e), I/R upregulated the expressions of caspase-3 and Bax mRNA and cleaved caspase-3 and Bax proteins but reduced the expressions of Bcl-2 mRNA and protein ($P < 0.05$). The Bcl-2/Bax ratio was decreased significantly in the I/R rats ($P < 0.05$). Compared with the I/R group, the expressions of caspase-3 and Bax mRNA and cleaved caspase-3 and Bax proteins were downregulated significantly, the expressions of Bcl-2 mRNA and protein were increased significantly, the expressions of Bcl-2 mRNA and protein were

![Figure 6: Immunohistochemical analysis of the expression of p-Akt in all groups.](image)
decreased, and the Bcl-2/Bax ratio was reduced in the rats given AM630 or wortmannin ($P < 0.05$). AM630 alone or wortmannin alone had no significant impact on the expressions of caspase-3 mRNA and cleaved caspase-3 protein, Bcl-2, and Bax mRNA and proteins compared to those of the I/R rats ($P > 0.05$).

3.8. The Effects of Dexmedetomidine Treatment on the Expressions of the PI3K/Akt Pathway. To investigate the protective mechanism of DEX on lung injury caused by intestinal I/R, we evaluated the expressions of Akt mRNA and proteins in lung tissues by RT-PCR and Western blot analysis. As shown in Figures 7(d), 8(a), and 8(f), I/R increased only the expressions of Akt mRNA and p-Akt protein, but there was no statistical difference, compared to those of the sham-operating rats ($P > 0.05$). Treatment with DEX increased the expressions of Akt mRNA and p-Akt protein compared to those of the I/R rats ($P < 0.05$). Treatment with AM630 or wortmannin downregulated the expressions of Akt mRNA and p-Akt protein compared to those of the DEX-treated rats ($P < 0.05$). AM630 alone or wortmannin alone had no significant impact on the expressions of Akt mRNA and p-Akt protein compared to those of the I/R rats ($P > 0.05$). These results suggest that the PI3K/Akt pathway may be involved in the protective role of CB2 receptor-mediated DEX treatment of I/R-induced lung injury. Collectively, our findings suggest that dexmedetomidine ameliorates lung injury induced by intestinal ischemia/reperfusion by upregulating the CB2 receptor expression or activating the PI3K/Akt pathway (Figure 9).

4. Discussion

Intestinal I/R may not only cause local intestinal injury but also induce remote lung injury. In this study, we recognized...
Figure 8: The effects of dexmedetomidine treatment on the protein expressions of cleaved caspase-3, Bax, Bcl-2, and p-Akt in all groups. (a) The expressions of cleaved caspase-3, Bax, Bcl-2, and p-Akt in lung tissues were detected using Western blot analysis. (b) Cleaved caspase-3 expression. (c) Bax expression. (d) Bcl-2 expression. (e) Bcl-2/Bax ratio. (f) p-Akt expression. Data are expressed as mean ± SEM, n = 6 rats per group. *P < 0.05 versus the Sham group, *P < 0.05 versus the I/R group, &P < 0.05 versus the DEX group. I/R: intestinal ischemia/reperfusion; DEX: dexmedetomidine-treated I/R; DA: dexmedetomidine- and AM630-treated I/R; AI: AM630-treated I/R; DW: dexmedetomidine- and wortmannin-treated I/R; WI: wortmannin-treated I/R.
that intestinal I/R led to lung injury, manifested by an increase in lung injury scores and an increased lung wet/dry ratio. At the same time, our results showed that dexmedetomidine attenuated lung injury by upregulating the CB2 receptor expression. However, the protective effects of dexmedetomidine treatment on lung injury induced by intestinal I/R were reversed by the CB2 receptor antagonist AM630. We also found that the CB2 receptor-mediated protective role of dexmedetomidine treatment of lung injury was achieved by activating the PI3K/Akt pathway.

Studies have suggested that oxidative stress is one of the major factors contributing to I/R injury [27]. In this study, we investigated and found that intestinal I/R resulted in an increase in MDA level and a decrease in SOD activity in the lung, aggravating lung tissue injury manifested by the increase in lung injury scores and lung wet/dry ratio. MDA, an end product of lipid peroxidation, is used to assess tissue peroxidative injury, while SOD, an antioxidant enzyme, is used to evaluate the antioxidant level of tissues. Several studies have indicated that I/R often increases the level of MDA and decreases the activity of SOD, leading to the aggravation of tissue damage [12, 25, 26, 28]. Our findings are consistent with those of these studies. Also, some therapeutic methods could reduce tissue damage by decreasing MDA levels and increasing SOD activity, such as antioxidant enzymes [25, 29, 30], and preconditioning [12].

Dexmedetomidine reportedly has an antioxidant effect, providing a protective role against damage in some tissues [7–12]. Here, we found that dexmedetomidine treatment alleviated lung injury caused by intestinal I/R via the inhibition of increased MDA levels and decreased SOD activity.

Our findings are consistent with previous studies suggesting that the inhibition of lipid peroxidation and enhancement of antioxidant enzyme activity in the lungs might, at least, be partially involved in the protective mechanisms of dexmedetomidine in response to lung injury caused by intestinal I/R. Inflammation and cell apoptosis play significant roles in the progression of lung injury caused by various factors, including I/R and shock. Some studies have indicated that pretreatment with some substances, such as cyclic arginine-glycine-aspartate peptide (cRGD) [31] and anti-osteopontin Ab [32], protects lung injury caused by intestinal I/R, lowers neutrophil infiltration, suppresses inflammation, and inhibits lung apoptosis after intestinal I/R. Ben et al. [33] established that lungs from TLR4 mutant mice undergoing intestinal I/R exhibited a marked reduction in epithelial apoptosis, pulmonary MPO activity, and TNF-α and IL-6 levels and manifested a significantly smaller histological injury. In this present study, intestinal I/R increased neutrophil and macrophage infiltration, pulmonary MPO activity, the levels of TNF-α and IL-6, and cell apoptosis, manifested by the increase in the expressions of caspase-3 mRNA and cleaved caspase-3 protein and decrease in the Bcl-2/Bax ratio that aggravated intestinal I/R-induced lung injury. In contrast, dexmedetomidine treatment reversed these changes, attenuating lung injury scores and lung wet/dry ratio and alleviating lung injury caused by intestinal I/R. Our results are, hence, consistent with these previous findings.

Although dexmedetomidine treatment has a protective role against intestinal I/R-induced lung injury, the exact mechanism is not entirely clear. Recent research has revealed a critical role for the CB2 receptor in regulating many disease
processes, including in the immune system, cardiovascular and respiratory systems, bone, gastrointestinal tract and liver, and reproductive system [17, 34]. The CB2 receptor has also been verified to represent a very promising therapeutic target in gastrointestinal inflammatory conditions [18, 35]. Although the activation of the CB2 receptor may have positive symptomatic effects on inflammatory bowel disease through its anti-inflammatory effects [36], it is not clear whether the CB2 receptor is involved in the process of intestinal I/R-induced lung injury. In this study, our investigation of the role of the CB2 receptor in the lung injury caused by intestinal I/R showed that treatment with dexmedetomidine increased the expression of the CB2 receptor and attenuated lung injury, manifested by the decrease in lung injury scores and wet/dry ratio. Furthermore, the protective effect of dexmedetomidine treatment on lung injury caused by intestinal I/R was reversed by the CB2 receptor antagonist, AM630. Our results suggest that dexmedetomidine treatment provided the protective role against intestinal I/R-lung injury by activating the CB2 receptor. The role of the CB2 receptor in the I/R injuries of some other organs has also been demonstrated, including in the myocardium [37, 38], liver [22, 39], and brain [40]. Our findings are, therefore, consistent with these studies.

The PI3K/Akt is known as an intracellular signaling pathway and has been confirmed to provide a protective role against I/R injury by regulating cell proliferation, apoptosis, and oxidative stress [41]. PI3K/Akt activation is reportedly involved in the protective effect of DEX against the I/R injuries in some organs, including the lung [42, 43], heart [44], and kidney [45]. In our study, we also found that DEX treatment increased the expressions of Akt mRNA, p-Akt protein, and Bcl-2 mRNA and protein, reduced the expressions of caspase-3 mRNA, cleaved caspase-3 protein, and levels of TNF-α, IL-6, and MDA, and attenuated lung injury scores. However, when given the PI3K inhibitor wortmannin, the above protective effects of DEX were reversed, suggesting that DEX reduced intestinal I/R-induced lung injury by activating the PI3K/Akt pathway, consistent with earlier studies.

Also, some studies have revealed that the activation of the PI3K/Akt pathway plays a part in the protective mechanism of cannabinoid receptors. Both CB1 and CB2 receptors stimulate the proliferation of neural stem/precursor cells through the PI3K/Akt pathway [46]. Previous research showed that the CB2 receptor agonist, JWH133, protected the heart against I/R-induced apoptosis through the involvement of the PI3K/Akt pathway [47]. However, it is not clear whether the PI3K/Akt pathway mediates the protective effect of DEX activating the CB2 receptor. Here, DEX upregulated the expressions of Akt mRNA and p-Akt protein, attenuated lung injury scores, and reduced intestinal I/R-induced lung injury. In contrast, the use of both the CB2 receptor antagonist, AM630, and the PI3K inhibitor, wortmannin, reversed the protective roles of DEX against lung injury. Additionally, the CB2 receptor antagonist AM630 downregulated the expression of Akt mRNA and p-Akt protein and increased lung injury scores. Our results indicate that DEX protected against lung injury caused by intestinal I/R via the CB2-mediated PI3K/Akt pathway.

The present study has some limitations. Although we investigated that the protective roles of DEX against lung injury caused by intestinal I/R were reversed by both the CB2 receptor antagonist, AM630, and the PI3K inhibitor, wortmannin, there was insufficient evidence to draw the conclusion that DEX treatment provides a protective role against intestinal I/R-induced lung injury through the activation of the CB2-mediated PI3K/Akt pathway. Further study using deficient animals is needed.

5. Conclusions

Our data show that DEX treatment provided a protective effect on lung injury caused by intestinal I/R, with the protective effects attributed mainly to the upregulation of the CB2 receptor and subsequent activation of the PI3K/Akt pathway. The current findings provide a basis for the development of novel strategies for the treatment of lung injury.

Data Availability

The authors have authorised me to promise that the data in our manuscript can be provided to anyone who needs it. If anyone needs these data, please contact me by the email hexh1220@aliyun.com.

Conflicts of Interest

The authors declare that there is no conflict of interests.

Authors’ Contributions

Meng Chen and Xue-Tao Yan contributed equally to this work and should be considered co-first authors.

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