Recent developments in the chiroptical properties of chiral plasmonic gold nanostructures: bioanalytical applications

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Abstract
The presence of excess L-amino acid in the Murchison meteorite, circular polarization effect in the genesis of stars and existence of chirality in interstellar molecules contribute to the origin of life on earth. Chiral-sensitive techniques have been employed to untangle the secret of the symmetries of the universe, designing of effective secure drugs and investigation of chiral biomolecules. The relationship between light and chiral molecules was employed to probe and explore such molecules using spectroscopy techniques. The mutual interaction between electromagnetic spectrum and chirality of matter give rise to distinct optical response, which advances vital information contents in chiroptical spectroscopy. Chiral plasmonic gold nanoparticle exhibits distinctive circular dichroism peaks in broad wavelength range thereby crossing the limits of its characterization. The emergence of strong optical activity of gold nanosystem is related to its high polarizability, resulting in plasmonic and excitonic effects on incident photons. Inspired by the development of advanced chiral plasmonic nanomaterials and exploring its properties, this review gives an overview of various chiral gold nanostructures and the mechanism behind its chiroptical properties. Finally, we highlight the application of different chiral gold nanomaterials in the field of catalysis and medical applications with special emphasis to biosensing and biodetection.

Keywords Gold nanostructures · Chirality · Circular dichroism · Plasmonic nanoparticle · Surface plasmon resonance · Chiroptical properties

Introduction
Chirality is a fascinating property of certain objects in nature [1]. Chirality means handedness, is observed in snail shells, gourd tendrils, human hands and even spiral galaxies [2, 3]. An object is said to be chiral if it is not superimposable on its mirror image [4]. Chirality plays a major role in the origin of life. The building blocks of life such as amino acids, carbohydrates and nucleic acids are chiral [5, 6]. Deoxyribonucleic acid/ribonucleic acid (DNA/RNA) consists of residues with the same handedness owing to act as template in replication of cell division process. In prebiotic chemical world, these macromolecules synthesized from small molecules are prone to have existed as racemic mixtures [5]. Life on earth manifests a priority for L-amino acids and D-sugars, in spite of a thermodynamic stability equilibrium of the L- and D-enantiomers of these molecules. The bioactivities of a chiral molecule depend on its handedness; for example, one enantiomer might act as a drug while the other might be toxic [7]. Accordingly, chirality of molecules has a vital role in functionality and interaction with other molecules.

Chirality is an intrinsic property of certain molecules or nanostructures, which on interaction with light, generate a significant contribution to both absorption and scattering [1, 8]. The interaction of chiral molecules with circularly polarized photons, circular dichroism (CD) effect occurs relating the variation in molecular absorption of clockwise and counter-clockwise circularly polarized lights [9]. CD spectroscopy is a potential technique to unravel molecular chirality and conformational analysis in a chemical reaction [10]. The chiroptical response of a chiral ensemble...
can be tuned and enhanced by the integration of plasmonic nanoparticles [11]. Chirality existence in nanosystem taking place via (i) intrinsic chirality depends on the geometry of the particle (ii) incorporation of chiral templates like DNA and peptides and (iii) plasmon-induced chirality [12]. Nanoparticles with chiral plasmonic properties generate chirooptical activity owing to its ability to create broadband circular polarization states of photons [13]. The origin of chirality in nanoparticle is well investigated using plasmonic chiral gold nanoparticles (Au NPs). The interaction of electronic states of ligands with the surface gold atoms give rise to distinct chirality effects in chiral Au NPs [14].

In this review, we present the origin of chirality of gold nanostructures, different types of chiral gold nanostructures, enhancement of chirality in Au NPs and its applications in physical, chemical and biological fields. The distinct electromagnetic properties, shift in wavelengths, energy changes associated with Au NPs and hybrid plasmonic Au NPs are also illustrated. We concentrate our attention on the phenomena of electronic interactions between ligands and surface gold atoms, variation in optical properties such as plasmonic resonance and CD. The mechanism of innate chirality and external factors contributing for chirality of gold nanostructures were also studied. Finally, we provide an outlook for numerous bioanalytical applications based on plasmonic chiral gold nanostructures.

**Chiral gold nanostructures**

A chiral dimer of gold nanorods (Au NRs) could be synthesized by using a chiral-binding agent between the adjacent nanorods in a solution. The size parameter and aspect ratio of the Au NRs affect its optical activity. Exciton coupling theory highlights the dipole–dipole interaction between dimer of Au NRs arranged as prolate ellipsoids in a medium of refractive index \( n = 1.5 \). The energy level of the localized surface plasmon resonance (LSPR) consists of hybrid modes such as symmetric (\( \omega_+ \)) and antisymmetric (\( \omega_- \)). The scaling of the system increased the extinction efficiency by a factor of 6, and overall extinction efficiency of the longitudinal modes can be attributed to the side-by-side assembly of the dimer, large refractive index of the adsorbed polystyrene and the interaction of plasmon resonances to changes in the refractive index. The absence of detectable CD signal before and after dimerization of Au NRs, owing to the existence of racemic mixture of chiral dimers in solution (Fig. 1a).

Transmission electron microscopy (TEM) images of the Au NDs depict that the particles have smaller aspect ratio (Fig. 1b) and remain in different configurations (Fig. 1c). The UV–visible extinction spectroscopy of the Au NDs gives a red shift after the formation of dimers (Fig. 1d and e). The red shift in the transverse mode as well as in the longitudinal mode can be attributed to the side-by-side assembly of the dimer, large refractive index of the adsorbed polystyrene and the interaction of plasmon resonances to changes in the refractive index. The absence of detectable CD signal before and after dimerization of Au NDs, owing to the existence of racemic mixture of chiral dimers in solution (Fig. 1f and g) [17].

The DNA-driven nanoparticle chiral pyramids comprising of Au NPs, CdSe@ZnS quantum dots and silver nanoparticles (Ag NPs) give CD signals in a wide range from 350- to 550-nm region. Chiral Au NPs synthesized using D-glutamic acid and L-glutamic acid inhibit the formation of human serum albumin fibrils. The surface chirality of Au NPs strongly influences the fibrillation process, which is more for D-glutamic acid–based Au NPs [18]. The Au114(SR)60 with chiral icosahedral shell (1-Au60) have unique optical, electronic, vibrational and structural properties [19].

The chiral handedness of Au NPs obtained from the helical glutathione (GSH) oligomers result in the formation of end-to-end crossed (EEX) Au NR junctions [20]. Peptide conjugates and alkyne-terminated aliphatic substrates \( \text{C}_{18}-(\text{PEP}_{\text{Au}}^{M-\text{ox}})_{2} \) direct the formation of plasmonic chiroptically active single helical Au NPs, where peptide conjugate \( \text{PEP}_{\text{Au}}^{M-\text{ox}} = \text{AYSSGAPPMP}^{\text{ox}}\text{PPF} \), \( \text{C}_{18} \) 18 carbon chain and ox = oxidation [21]. The chirality of Au NRs depends on the aspect ratio. The chirality
Fig. 1 Structural and spectral characteristics of Au NDs and Au ND dimers. a Cartoon depiction of the dimer formation scheme through polymer capture, resulting in parallel (top), twisted (middle) and twisted with planar displacement (bottom) dimers. b, c TEM images of the Au NDs before and after dimerization, respectively. Scale bars are 100 nm. d, e Ensemble UV–Vis extinction spectra of the Au ND and dimer samples, respectively. The Au NDs show maxima at 521 and 812 nm. The maxima for Au ND dimers are at 534 and 816 nm. f, g Ensemble CD spectra of the Au ND and dimer samples, respectively. No peaks are observed in either case. “Reprinted with permission from Ref. 17. Copyright (2016) American Chemical Society”
increases with increase in aspect ratio. It depends upon the concentration of AgNO₃ in growth solution. The length of Au NRs increases, whereas diameter decreases with increase in concentration of AgNO₃. The origin of intrinsic chirality in nanocrystals is due to the point defects, dislocation and chiral defects in the nanosystem, which is greater in Au NRs with high aspect ratio. Here Au NR–based colorimetric assay was used for the detection of L-valine in the presence of D-valine with a detection limit of 0.1 mM [22]. The human serum albumin and porcine serum albumin–guided chiral assembly of Au NRs exhibit left-handed CD responses, whereas equine serum albumin, sheep serum albumin and bovine serum albumin executed on the converse (Fig. 2a). The interaction of albumin with Au NRs is electrostatic. The cryogenic TEM image of human serum albumin–guided and bovine serum albumin–guided Au NRs manifested left-handed and right-handed twisted side-by-side geometry respectively (Fig. 2c and d). These spatial asymmetry geometries indicated the template-dependent chirality of serum albumins. The handedness of the chiroptical properties depends on the surface charge distributions of the serum albumins and the ambient pH [23].

Chiral plasmonic gold nanoparticle

A multi-chirality evolution step method for the synthesis of chiral plasmonic Au NPs with increase in particle uniformity has been utilized for the enhancement of chiroptic response. The Au NPs having 432 point-group symmetry (432 helicoid 111) shows well-defined gaps between the nanoparticle, increased dis-symmetry factor of g = 0.31 and distinct cubic boundaries [24]. Chiral Au NPs are produced by using peptides with random coil and α-helix. These Au NPs show CD signals at the surface plasmon resonance (SPR) wavelength (≈ 520 nm) and in the visible region of spectrum get red-shifted on aggregation [25]. A core-satellite nanostructure of Au NRs (core) and Au NPs (satellites) were created using DNA as linkers. CD spectrum displayed plasmonic CD (PCD) corresponding to Au NRs and Au NPs at SPR wavelength region (Fig. 3). The chirality depends upon the number of Au NPs around Au NRs, structure of core-satellites nanosystem, ligand induction, dipole–dipole interactions and chiral arrangement of surface atoms or molecules [26].

Chiral cholesterol-capped Au NPs were constructed and synthesized uniformly dispersed mixtures in different nematic liquid crystal hosts. These nanoassemblies prepared in the presence of disulphide chiral bias are strong chiral inducers as compared to that in the absence of a chiral bias [27]. The chiral discrimination of tyrosine and phenylalanine
enantiomers with cysteine-modified Au NPs was analysed with Raman spectra. It was found that the left-handed enantioomer combination of tyrosine or phenylalanine with same handedness cysteine-based Au NPs gives enhanced Raman signals. Similar observation was obtained for D-enantiomer as well. This type of interaction occurs due to (i) the presence of three interaction sites in the vicinity of D-selector and D-enantiomer of L-selector and L-enantiomer and (ii) possibility of more hydrogen bonds between different amino acids of same handedness as that of different handedness [28]. The blending of the results obtained from CD spectra and Raman spectra can be used for the discrimination of enantiomers of amino acids. Chiral liquid crystalline Au NPs (LC-Au NPs) were assembled into a fluid lamellar structure showing chiroptical activity. The arylamine in liquid crystal ligands reduce Au(III) to Au(0) and bind to the Au NPs core. The LC-Au NPs exhibit temperature-dependent bisignate CD signals. The thick coating of Au NPs by mesogens produces a negative refractive index. This physical quantity can be utilized for the investigation of metamaterials [29].

Chiral plasmonic gold nanoantenna

The combination of Au NRs with a 3D chiral structure known as nanoantenna delivers powerful chirality probes for functionalization with proteins, DNA and other metal nanoparticles. The ordering of gold nanorods on a fibre backbone gives a three-dimensional chiral structure known as plasmonic gold nanoantenna. The fluid dispersion of chiral fibres is obtained by adding water in anthraquinone-based oxalamide-1 present in DMF/ethanol solution. This plasmonic gold nanoantenna generates strong surface plasmon–mediated circular dichroism (SP-CD) with high anisotropy factors (g-factor) across visible and near-infrared wavelengths (600–900 nm). These Au NRs in fluid suspension manifest a g-factor of 0.022, which is comparable to proteins complexes (0.06) and polyaromatic compounds (0.05). The SP-CD band arises as a result of 3D chiral arrangement of the Au NRs, and the intensity of SP-CD decreases with an increase in disorder of the particle. Here, each particle is considered as an electric dipole where the SP-CD depends on the geometry and composition of the structure [30].

Plasmonic helical nanoantenna on thin gold layer converts subwavelength longitudinal near-fields into circularly polarized wave and vice versa. The nanoantenna consists of L-handed four-turn Au-coated carbon helix coated on cylindrical pedestal. The excitation of an L-handed helix with circular polarization generates a longitudinally oriented dipole containing the surface plasmons. This effect occurs in a broad wavelength range owing to the waveguide mode of surface plasmons propagating along the nanoantenna [31]. The stability of such nanoantenna is excellent. The nature of nanoantenna with right-handedness upon excitation could also contribute to its usage in practical applications.

Hybrid chiral gold nanostructures

The chiral gold-upconversion nanoparticle pyramids exhibit strong plasmonic circular dichroism at 512 nm and luminescence in the range 500–600 nm. The resulting nanopyramids enumerate the following process: (i) bathochromic shift in UV–Visible spectrum, (ii) strong CD signal and (iii) luminescence resonance energy transfer (LRET) from
upconversion nanoparticle to Au NPs. The merit of this nanoassembly is that multiple optical on–off changes take place on interaction with active analytes [32]. The chiral gold nanoparticle–chromophore hybrid complexes possess tuneable strong chiroptical activity. The chiroptical activity is tuned by optimizing the spectral overlap between plasmon resonances and chiral collective molecular excitations. The chiral Au NP–chromophore hybrid assembly with the aid of DNA-template establishes a spectral overlap between plasmons and collectively coupled chiral excitons. This leads to an enhancement in molecular CD along with a triggered plasmonic CD. This nanosystem can act as a bisignate CD probe for nanosensing strategies [33].

Core-satellite/shell chiral gold nanostructures

The chiral DNA-modified shell core-satellite nanostructures based on Au NPs have been self-assembled for the detection of mycotoxin. The shell core structure consists of Au–Ag alloyed hollow sphere as core with a thin layer of gold as outer shell. The core-satellite structure was developed with the dipole–dipole interaction between DNA-modified shell core structure and Au NPs, which produce a strong chiral signal [34]. The gold/silver core shell nanorods functionalized with DNA has enhanced plasmonic properties and exhibit increased circular dichroism responses. The Ag shell thickness can be used as a parameter for tuning the LSPR. The DNA origami platform holds great promise for synthesizing bimetallic nanoparticles [35]. The chiral core-satellite silver nanorod (Ag NR)-gold nanoparticle nanostructure has been fabricated by using L-cysteine and D-cysteine as bridging molecules. The L- and D-cysteine-based core-satellite structure gives mirrored CD signals. Increase in concentration of both enantiomers produces a red shift in CD spectra. Here, the origin of optical activity is explained as (i) electronic interaction between chiral cysteine molecules and achiral metal electrons and (ii) vicinal effects developed from the dissymmetric fields via the formation of chemical bond between dissymmetric centre and the metal core [36].

The chiral cysteine molecules intercalated between Au NR core and Au shell, which is modified with cetyltrimethylammonium bromide and cetylpyridinium chloride (Fig. 4a). The overall morphology of the core–shell structure consists of an additional arrow head–like end cap as that of Au NR core (Fig. 4b). The modification of the core–shell structure with CTAB imparts positive charge on its surface. The trisodium citrate molecules link the modified core–shell assembly side-by-side (Fig. 4c and d). The surfactants transfer and amplify the chirality induced by the adsorbed chiral molecule. The chiral force developed, owing to the formation of hot spots via dipole–dipole interaction between chiral molecule and achiral plasmonic gold nanostructures [37]. The surfactants and chiral thiols could induce strong chiroptical effects in gold-nanoparticle based core–shell structures.

DNA-based chiral gold nanostructures

Three-dimensional (3D) plasmonic chiral gold nanostructures were developed through programmable transformation of Au NPs using DNA origami as template. A two-dimensional (2D) DNA origami template organized the Au NPs into a 3D helix with engineerable optical chirality [38]. The plasmonic coupling between Au NRs in a pattern is arranged in the presence and absence of a gold nanosphere using DNA origami. Here, the gold nanosphere acts as a transmitter. The two Au NRs having 54-nm length and 23-nm width are separated by a surface-to-surface distance of 62 nm and tilted by 90° to each other. These nanosystems were assembled in thiol-functionalized DNA (Fig. 5b and d). The presence of gold nanosphere (Fig. 5a and c) in between Au NRs creates hotspots via near-field mechanism and produced enhanced CD response [39]. The interaction of these nanosystems with biomolecules for utilizing it as a biosensor is in chaos. The Au NPs were assembled into 3D tetrahedrons with DNA origami template. The CD spectra of the 3D tetrahedrons exhibit bisignate signatures. The CD signal depends on the size of the Au NPs as well as the interparticle distance (Fig. 5). For same-sized Au NPs (20 nm), the intensity of CD signal continually increases with decrease in interparticle distance and decreases with decrease in size of Au NPs (13 nm) [40].

The arrangement of four identical Au NPs into 3D asymmetry tetramer using DNA origami template exhibiting 3D chirality [41]. Modular assembly of spherical Au NPs, Au NRs with definite numbers and heterogeneous nanostructures were fabricated with sample DNA origami template [42]. The DNA barrel scaffold induces the assembly of left- or right-handed tethers for the building-up of the chiral plasmonic Au NPs. The scalability of the DNA barrel structure into nanowire, followed by the deposition of Au NPs, could be initiated as chiroplasmonic wave-guides [43]. A dynamic Au NP plasmonic chiral nanostructures assembled using the DNA origami method in which the DNA alignment process gives several-fold increased CD signal [44].

The Au NR–based chiral plasmonic-excitonic systems were assembled on DNA template. The nanosystem shows Rabi splitting and anticrossing behaviour in the CD spectra. Rabi splitting occurs whenever the plasmon and exciton resonance energies are the same. The energy difference between symmetric mode and excitons is known as Rabi splitting. The splitting increases with an increase in the extent of coupling between excitonic molecule and chiral plasmonic nanostructures. The plasmon-like branch and exciton-like branch show transformations until the CD
resonances of the two branches become synchronized, thus exhibiting anticrossing property [45].

**Polymer-based chiral gold nanostructures**

The chiral self-assembled Au NRs with strong CD signal in the visible-near IR region has been successfully constructed using poly (2-hydroxyethyl methacrylate) (PIPEMA) and poly (methacrylate hydroethyl-3-indolepropionate) (PHEMA). The pristine Au NRs do not induce any CD response. The intensity of bisignate longitudinal peak at 591 nm (positive peak) and 730 nm (negative peak) is stronger than the transverse peak at 528 nm (Fig. 6a). Here, the intensity of CD signal depends on the molecular weight of the polymers used as well as the pH values of the solution (Fig. 6a and d). The TEM image and tilt angle (0–60°) TEM image show an L-handed side-by-side arrangement of Au NRs (Fig. 6b and c) [46].

The chiral arrangement of Au NPs using D- and L-helical pores in block copolymer films templates was fabricated. The arrangement of Au NPs in a porous poly(butadiene)-b-poly(ethylene oxide) film prepared using D-tartaric acid and L-tartaric acid with projection perpendicular and parallel to the cylinder axis are shown in Fig. 7. The TEM image consists of Au NPs with a pitch of average 40 nm (Fig. 7a and d) and 3D tomography further confirms the different helical chiral arrangements (Fig. 7c) [47].

Fig. 4  a Schematic diagram of embedding sergeants in the interface region of Au NR core and Au shell (upper panel). A chiral SS dimer is used to demonstrate the formation of chiral plasmonic assemblies (lower panel). The surfactant layer (soldiers) on the nanorod surface is omitted for clarity. b Representative TEM images of Au NR cores and AuNR@Cys2@Au0.05 core–shell nanostructures. c SEM and TEM images of AuNR@Cys@Au SS oligomers. d A HRTEM image showing rod side facet link mode in a rod trimer. AA/Au3+ molar ratio = 1.6. “ Reproduced from Ref. 37 with permission from the Royal Society of Chemistry”
Origin of chirality

The plasmon-coupled circular dichroism (PCCD) emerges due to the dipole–dipole interaction between plasmonic nanoparticle and chiral adsorbate molecules. The PCCD was achieved in Au NRs-BSA aggregates. The CD arises in these aggregates by: (i) intrinsic structural chirality with enantiomeric excess of right-handed configuration, (ii) hot spots created between closely spaced Au NRs. The high-angle annular dark-field scanning transmission electron microscopy (HAADF-STEM) imaging indicates that non-parallel dimer of Au NRs was circular differential scattering (CDS) active at 720 nm (Fig. 8A and B) whereas parallel dimer of Au NRs was CDS active at 618 nm (Fig. 8E and F). The tomographic reconstruction and simulations indicate that nonparallel dimer is chiral (Fig. 8C and D) and parallel dimer is achiral (Fig. 8G and H) [48].

The gold nanostructures lock the chirality of circularly polarized light through the plasmonic field-driven asymmetric displacement of nanoparticles in dynamic ensemble followed by particle-to-particle growth [49]. Chiral gold nanostructures were synthesized using L-cysteine and D-cysteine as capping agents [50]. The L-cysteine-encoded gold nanoparticle absorbs right-handed/clockwise circularly polarized light (CPL) at 534 nm and left-handed/anticlockwise CPL at 638 nm. In the CD spectrum of D-cysteine-encoded Au NPs, an inverted response at the same peak position is obtained (Fig. 9a). The 150-nm-sized rhombic dodecahedron Au NPs have 60-nm-sized edge with bent at an angle of \(+\phi\), while for D-cysteine the same sized nanoparticle has edge with bent at an angle of \(-\phi\) (Fig. 9b and c) [51]. The chiro-optical activity of amino acid and peptide directed synthesized gold nanoparticle depends on edge length, gap width, gap angle, growth rate, curvature and nature of additives [50]. The origin of chirality in \(\text{Ag}_{24}\text{Au}_1(R/S-\text{BINAS})_x(\text{DMBT})_{18-2x}\) where \(x\) ranges from 1 to 7, \((\text{DMBT}=2,4\text{-dimethylbenzenethiolate, BINAS}=R/S-1,1',2',2''\text{-dithiophene})\) is due to the transitions within \(\text{Ag}_{24}\text{Au}_1\text{S}_{18}\) framework and conjugation effects within the aromatic system of the ligand[52].
**Fig. 6**  
(a) CD spectra of the original GNRs and the assemblies induced by PIPEMA192 under different pH values. The numbers in the label correspond to the pH value in the solution.  
(b) Representative TEM image of the assembled nanostructures at pH 13.4.  
(c) Top: tilt angle TEM images of a representative assembled structure, bottom: the corresponding 3D model structures. The numbers in the label correspond to the title angle.  
(d) CD spectra of the GNR assemblies induced by PIPEMA with different molecular weights. “Reprinted with permission from Ref. 46. Copyright (2019) American Chemical Society”

**Fig. 7**  
TEM image of chiral arrangement of Au NPs in a porous PBdEO film prepared using D-TA with projection (a) perpendicular to the cylinder axis and (b) parallel to the cylinder axis. The doping amount of TA in the parent BCP film is 16 wt %.  
(c) 3D tomography of chiral arrangement of Au NPs in a porous PBdEO film using D-TA.  
(d) TEM image of chiral arrangement of Au NPs in a porous PBdEO film prepared using L-TA with projection perpendicular to the cylinder axis.  
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Fig. 8  CDS-active Au NR dimer-BSA complexes with chiral and achiral configurations. A and E HAADF-STEM tilt-series images of a chiral dimer (A) and an achiral dimer (E) and the corresponding geometric models extracted from the tomographic reconstructions. B and F Experimental single-particle CDS spectra of the chiral dimer (B) from (A) and the achiral dimer (F) from (E). The experimental spectra are shown with a pink envelope that represents the experimental error. C and G Simulated scattering spectra of the chiral dimer (C) and achiral dimer (G) for incident LCP and RCP light and the corresponding CDS spectra. The insets in (C) show the charge plots calculated at 720 and 805 nm. The insets in (G) show the charge plots calculated at 618 and 670 nm. The charge plots in C and G share the same scale bar. The dashed lines in C and D refer to the plasmon modes at 720 (green) and 805 nm (orange). The dashed lines in F and G refer to the plasmon modes at 618 (green) and 670 nm (orange). D and H Cross-sectional views of calculated near-field enhancements (|E/E₀|²) for the chiral dimer (D) at 720 and 805 nm and the achiral dimer (H) at 618 and 670 nm. These results demonstrate that although the dimer in (E) to (H) is not chiral on the basis of its geometry, CDS is observed and must originate from chiral BSA molecules located in inter-NR hotspots. a.u., arbitrary units; E, electric field; k, incident wave vector. “ Reproduced with permission from Ref. 48. Copy right 2019, The American Association for the Advancement of Science”
The three main spectroscopic techniques for the characterization of chirality in materials include optical rotatory dispersion (ORD), CD and Raman optical activity (ROA). CD is related to the imaginary part of the chirality parameter “k.” It depends on ellipticity, which is a function of the proportionate difference in transmission. The ORD is related to the real part of chirality parameter. It is expressed as the unequal rotation for plane polarized light of different wavelengths [53]. Optical activity is the ability of a molecule to rotate the plane polarized light [54].

**Intrinsic plasmonic chirality**

In 3D chiral plasmonic nanomaterials, the circular dichroism effect occurs due to the interaction of (i) electric dipole and magnetic dipole and (ii) electric and magnetic dipoles with dipole-quadruple responses of the system. A 3D chiral configuration resulted from the wave vector, electric dipole and magnetic dipole. The interplay of electric and magnetic dipole results in an imaginary part of mixed electric and magnetic polarizability. This mixed polarizability is a dot product of both these dipoles. The helical arrangement of Au NPs bring about coupled plasmon waves disseminating along a helical path associated with absorption of these waves with respect to the handedness of the plasmonic nanohelices [55].

**Extrinsic plasmonic chirality**

Extrinsic plasmonic chirality resulted from the mutual alignment of the 2-D metamaterial and the direction of wave propagation [54]. Extrinsic chirality of 3D-chiral metamaterials occurs by the oblique incidence onto metasurfaces, if it lacks line of mirror symmetry in the direction of plane of incidence [56, 57]. The extrinsic chirality via asymmetric transmission due to oblique incidence can occur in square arrays of plasmonic nanospheres, semiconductor quantum dots, planar metamaterials and lossy double-periodic gratings [58]. The extrinsic chirality through photoluminescence emission takes place by breaking the mirror symmetry in the presence of asymmetric Au coating [59]. The oblique excitation/illumination of a 2D achiral plasmonic nanostructure at a particular incident angle exhibits a non-zero plasmonic CD response through extrinsic chirality [60]. Such extrinsic chirality happens by the strong interplay of electric and magnetic dipoles leading the way to mixed polarizability. Under oblique incidence, symmetry breaking and the projection of the oscillating magnetic moment on the plane perpendicular to the wave propagation induced by CPL are nonzero [61].

The huge extrinsic CD of a plasmonic splitting rectangle ring originates due to the generation of circular current by acting as a magneton is verified using the FEM simulation. In analytical model, the CD of meta-molecule from left and right circularly polarized light occurs due to mixed electric and magnetic polarizability [62]. The Au–Ag heterodimer behaves like a plasmonic magneton. It possesses multipolar Fano-resonances and powerful CD effect. Both Fano-resonance profile and CD signal depends on size, gap and shape of nanorice heterodimer; the CD spectra are sensitive to ambient medium in visible and near IR regions. In addition to it, the CD effect varies constructively with Fano-asymmetry factor [63].

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**Fig. 9** Opposite handedness of chiral nanoparticles depending on the cysteine enantiomer. a Circular dichroism (CD) spectrum of nanoparticles synthesized using L-cysteine (L-Cys, black) and D-cysteine (D-Cys, red). SEM images of resultant chiral nanoparticles obtained using L-Cys (b) and D-Cys (c). Schematic models and magnified SEM images are shown in the inset. Auxiliary lines indicating oppositely bent edges at an angle of +ϕ for L-Cys and −ϕ for D-Cys are illustrated in the SEM images. Scale bar, 100 nm. Reproduced with permission from Ref. 51. Copyright © 2020, Springer Nature.
Chiral fields in dipoles under external excitation

The generation of strong chiral fields in the vicinity of a dipole dimer is illustrated in Fig. 10. An enhanced and uniform chiral field was formed in the gap between the two dipoles placed in such a way that two lobes with the same chirality get overlapped. In the corners of Au block dimers, strong field enhancement occurs, resulting in drastic chiral fields. The chiral metal nanostructure and non-chiral structure with CPL illumination, the chiral field is non-uniform and the enhancement changes between positive and negative at different wavelength. In the case of helical structures excited with linearly polarized light, the chiral field is uniform. Here, the enhancement factors are either negative or positive [64].

Discrete dipole approximation

In multipoles, thousands of dipoles are considered as individual particles. The plasmonic CD response of such multipoles is taken by averaging the variance in extinction cross sections, \( Q \), of left circularly polarized from right circularly polarized plane waves by considering different orientation of the helical superstructures \( CD = \langle Q_{IL} - Q_{IR} \rangle \Omega \), where \( Q_{IL} \) and \( Q_{IR} \) indicates extinction cross section of left circularly polarized plane waves, and right circularly polarized plane waves, respectively, \( \Omega \) represents the set of different orientations.

**Fig. 10** Formation schematic of enhanced chiral near-fields with uniform optical chirality in the gap of a coupled point dipole dimer (a) and Au spherical nanoparticle (10-nm diameter) dimer (b). a Analytically calculated chiral near-field distributions of (i) one dipole, (ii) two dipoles with a large gap \( d \) of 0.06λ, and (iii–iv) two dipoles with a small gap \( d \) of 0.04λ. Black arrows show the dipole momentum. Signs of “+” and “−” in the scale bar indicate the field is left- and right-handed, respectively, which applies to all figures in the following. b Numerically calculated chiral near-field distributions of (i) one sphere, (ii) two spheres with a large gap \( d \) of 10 nm and (iii–iv) two dipoles with a small gap \( d \) of 2 nm. c Corresponding electric field distribution of the case (b)-iii. The right coordinates give incident polarization and direction. d Schematic of the directions of incident fields and scattered fields by a dipole dimer. Reproduced with permission from Ref. 64. Copyright © 2015, Springer Nature
of orientations of the helical superstructure with respect to the incident wave-vector [65]. The total CD signal in a system comprising of a nanocrystal and a chiral molecule is \( \text{CD}_{\text{NP,molecule}} = \text{CD}_{\text{NP}} + \text{CD}_{\text{molecule}} \). Where \( \text{CD}_{\text{NP}} = \langle Q_{\text{NP,}+} - Q_{\text{NP,-}} \rangle \Omega \), where \( Q_{\pm} \) are the rate of absorption corresponding to two incident electromagnetic waves. The molecule produces a strong Fano effect in the CD spectra. This CD band of molecule appears at the plasmon frequency. The plasmonic nanoparticles like gold and silver nanoparticle exhibit an absorption enhancement with small separation between molecules and nanoparticle [66].

### Optical coupling between gold nanorods and chiral surfactants

The Au NRs coated with chiral mesoporous silica shells (CMS) manifest strong CD signals in the visible light and NIR region (Fig. 11a). The CD activity is due to the optical coupling between Au NRs and the chiral cysteine molecules docked in the CMS shells. The small CD signals at 520 nm correspond to the feeble transverse LSPR while strong CD signal ranging from 500 to 900 nm are attributed to the intense longitudinal LSPR (Fig. 11b). In the absence of chiral mesoporous silica, the mixing of Au NRs with pure chiral molecules lacks CD signal in the visible region, while that in the UV region remains. With an increase in aspect ratio of Au NRs, the plasmonic peak around 700 nm gets red-shifted. It indicates that the origin of plasmon-induced chiral property is due to the strong near-field coupling between chiral molecules within the mesoporous shell and Au NRs [67].

### Chirality induced by preferred handedness of the polymer chain

Using the chain transfer polymerization of PIPEMA and PHEMA leads to the self-assembly of Au NRs, with CD response at Vis–NIR region. The CD spectra of the nanoassembly give a maximum intensity peak at pH of 13.4. The origin of chirality of Au NRs is controlled by the preferred handedness of the chiral polymers owing to the syndiotacticity and steric hindrance of the side groups of the polymer chain. Moreover, the intensity of the CD signals is controlled by the molecular weight of the polymers [46].

### Born-Kuhn model

The plasmonic form of the Born-Kuhn model can be well illustrated as two identical corner-stacked, vertically aligned nanorods having an angle of 90° between them. These orthogonal nanorods are equivalent to chiral plasmonic dimer. In coupled oscillator model the excitation of a mode depending on symmetry of light corresponds to antisymmetric case. Hybridization model gives idea about sign and spectral position of the resulting optical modes. The plasmonic Born-Kuhn model demonstrates the tuning of vertical inter-rod distance, which can open on to selective excitation of bonding and antibonding plasmonic chiral modes [68]. The Born-Kuhn-type plasmonic analog produced with 3D gold chiral metasurfaces was used for the analysis of third-harmonic generation circular dichroism (THG-CD). The THG-CD spectra consist of broader peak and dip than that of linear CD spectra. In addition to this, the nonlinear CD peaks is almost zero at the peak position of linear CD spectra [69].

### Tuning of chirality of Au NPs

The chiroptical activity of chiral Au NPs can be tuned by the peptide sequence. The sequence engineering of A3 gold-binding peptide (AYSSGAPPMPFF) can be used for optimizing chiroptical properties of chiral helical Au NPs. The similar CD spectra show that all peptides have polyproline II secondary structure (Fig. 12). The position of methionine
(M) or methionine sulfoxide (M-ox) within the peptide sequence controls the aspect ratio of Au NPs. The nature of the peptides determines the formation of spherical, oblong or rod-shaped Au NPs. The shift in the M-ox position of peptides gives large-sized oblong Au NPs with high intensity of CD peaks [70]. A series of Au NRs capped with axially chiral binaphthyl thiols and its dispersion in nematic liquid crystals 5CB have been synthesized. The enhancement of chirality by a related series of polyhedral Au NRs can be employed to identify chirality transfer. The nematic liquid crystal phase reflects the variation in chiral induction strength and facilitated cooperative effects with helically arranged chiral objects [71]. In a polar protic solvent, chiral gold nanoclusters (Au NCs) undergo self-aggregation into supraparticles, while in polar aprotic solvent, self-aggregation does not occur. The degree of self-aggregation increases with an increase in polarity of polar protic solvent, which induces amplification and chirality of supraparticles [72].

The Au NPs functionalized with a highly sensitive and conformational responsive enantiomeric pairs of axially chiral binaphthyl ligands were employed to induce the chirality transfer from nanoscale solid particle to a bulk achiral liquid crystal phase. The Au NP–conjugated binaphthyl ligands were used for analysing the dependent factors of chirality transfer such as size of Au NPs, chiral correlation length, the local number, dihedral angle and density of chiral binaphthyl ligands [73]. A coordinating thermally sensitive organogelator template controls the chirality in the in situ synthesis of hybrid Au NPs. The sol form of the template induces coordination between Au(III) and azobenzene moiety results in chirality inversion, whereas the gel form of the template cannot coordinate with HAuCl₄ so that original chirality is maintained [74]. The chiral poly(phenylene-alt-benzothiadiazole) doped with helically shaped Au NPs via dipole–dipole interaction leads to plasmonic enhancement of optical chirality parameter (Fig. 12) [75]. The circular differential Mie scattering (CDMS) of Au NPs is linearly proportional to the resonance strength and chirality parameter (k) of the molecules [76]. The three chiral Au NR-chromophore hybrid structures were constructed by self-assembling achiral cyanine dye K21J-type aggregates, cyanine dye (cy3) and cyanine dye (cy5) with Au NRs with the aid of DNA templates. The chirality of the hybrid complexes depends on the spectral overlap between the SPR and chiral collective molecular excitations [33].

**Mechanism of CD effect in chiral plasmonic gold nanostructures**

The mechanism of CD effect in chiral plasmonic gold nanostructure is based on coulomb interaction, plasmon-induced mechanism and dipole–dipole interaction. The coulomb interaction between molecule and nanoparticle creates an angle separation between electric and magnetic dipole of the molecule (Scheme 1a and b). The interaction of nanoparticle creates a CD effect via lowering the symmetry of the molecule. The dipole of the molecule induces a chiral current inside the nanoparticle. It results in a Fano effect in the CD spectra. The CD spectra due to the interaction of a molecule and nanoparticle in the presence of an electric field comprises of (i) incident field, (ii) field induced by nanoparticle and (iii) field created by the molecule. The asymmetric shapes of CD bands of a nanoassembly are called Fano-type effect. In Scheme 1a, the electric and magnetic dipole $\mathbf{\mu}_{12}$ and $\mathbf{m}_{21}$, respectively, are not perpendicular to each other, thereby resulting in CD signal. Here, R is centre to centre separation and $\Delta$ is the distance between the centre of the molecule and nanoparticle surface [66].

The chiral adsorbate on Au NPs interacts with the electronic states of the nanoparticle and generates a CD band at the plasmon wavelength (Scheme 1c and d) [77]. This type of mechanism is applied to the nanosystem in which the absorption peak of the chiral molecule is far away from plasmonic band of the nanoparticle. Here, the CD effect is induced due to the modification of the surface state of the nanoparticle via the adsorption of the chiral molecule [66].
In hybrid materials such as azo-compounds and Au NPs in polymer films, the two components of dipole vectors can freely rotate in a direction perpendicular to the direction of propagation of light. The CPL can induce supramolecular chirality with one vector on the surface of Au NPs and the second dipole vector is fixed within the Au NPs (Scheme 1e and f). Here, the initial weak adsorption of chiral species on the surface of Au NPs limited the motion of the resulting complex (Scheme 1e) [78].

Applications of chiral plasmonic gold nanostructures

Chiral plasmonic gold nanostructures are established as a probe for a wide variety of applications including catalysis, biosensing and biodetection. Chiral, multipetal gold nanoflowers (Au NFs) were prepared by the reduction of HAuCl₄ with the aid of core-satellite silver nanoparticle seeds. The Hg²⁺/Au NF combination catalyses the transformation of rhodamine B to rhodamine 110 and nitroarenes to aminoarene. The bright-field TEM images of Au NF look like mermaid flower (Fig. 13a and b). The HRTEM images consist of lattice planes with spacing 2.33 Å corresponding to (111) plane of fcc Au NFs (Fig. 13c). The STEM-HAADF (scanning transmission electron microscopy/high-angle annular dark field) (Fig. 13d), energy-dispersive x-ray (EDX) (Fig. 13e), energy-filtered TEM (EFTEM) image (Fig. 13g), CD spectra and wide-angle XRD profile of Au NF prove the characteristic properties of gold. Relative thickness map of individual Au NF (Fig. 13f) is shown in Fig. 13h [79].

Biosensing

Chiral plasmonic gold nanostructures exhibiting CD were used for biosensing because they are sensitive to asymmetry in sample structure, quick easy method in obtaining measurements, shorter time scale, less amount of sample required.
and recovery of sample is possible [80]. CD exploits the differential absorption of right- and left-handed plane polarized light sensitive to asymmetric in structure of molecules [81]. Recently, chiral plasmonic gold nanostructures offer new platform for highly sensitive detection strategies. Because of their biocompatibility and plasmonic properties, gold nanostructures can make sensible effect in the detection process.

Chiral plasmonic gold nanostructures can find the way towards various biosensing applications, including label-free single-molecule/virus detection. The antibody-conjugated self-assembled superstructure of chiroplasmonic gold nanoparticles (CAu NPs) and CdTe NPs quantum dots (QDs) nanohybrids could detect picomolar concentrations of avian influenza A (H5N1) virus. The CdTe NPs exhibit a particle size of 6.5 nm with strong emission wavelength at 710 nm and possibly could overlap with plasmonic states in the nanohybrids. Here, prolate, flower and urchin-shaped Au NPs having plasmonic peaks at 548 nm, 565 nm and 590 nm, respectively, were chosen to make nanohybrids. The urchin-shaped Au NPs outperform all other Au NPs because of their broadened plasmonic peak and maximum extent of overlap with excitonic wavelength of CdTe NPs. The sensitivity of the nanohybrid-based bioassay could detect up to 1 pgmL⁻¹ of H₅N₁ virus, which was 10 times more than that of bioassay based on bare urchin-like Au NPs. Moreover, chiroptical response is more sensitive as compared to visual colour response–based commercial kit, which could detect up to 1 ngmL⁻¹ (Table 1). The sensing performance was also extended to detect avian influenza A (H₄N₆) virus, coronavirus and fowl adenovirus. The concept of optical rotation activity and energy coupling characteristics due to chiral plasmon-exciton interactions change the chirality enhancing the sensitivity of chiral plasmonic nano sensors [82].

A chiral assembly of 16-mercaptohexadecanoic acid–functionalized Au NPs immobilized with tropomyosin (TROP) and anti-tropomyosin monoclonal (mAb) antibodies as trimer was developed for the sensing and quantification of shell fish allergen tropomyosin. In the nanosensor, Au NPs-TROP comparatively binds with Au NPs-mAb and free antigen. The AuNPs-TROP and Au

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**Fig. 13** a and b BF-TEM images of Au NFs and a single Au NF, respectively, and c HRTEM image of a part of Au NF. d STEM-HAADF image. e EDX line profile. f Unfiltered image of a single Au NF. g EFTEM image of that Au NF. h Relative thickness map. i Line profile over the red rectangular box indicated in h. j CD spectra of chiral Au NFs. k Wide-angle XRD profile of Au NFs. “Reprinted with permission from Ref. 79. Copyright (2015) American Chemical Society”
NPs-mAb could form aggregates of different oligomers and contributes to the increase in CD intensity. The biosensor was designed in such a way that a decrease in CD intensity takes place whenever the concentration of free antigen increases, resulting in a decrease in the concentration of aggregates. In comparing CD absorption curves of the probe in the presence of non-allergic proteins, the latter does not affect the strong CD intensity of the probe, indicating the specificity of the biosensor [83].

The deposition of gold or shells around gold heterodimers gives tuneable CD signals, which depends on the composition, shell thickness and sequence of metallic layer (Fig. 14). The chiroptical assemblies of nanoparticles produce amplified chiroptical signal, were used for DNA detection at zeptomolar level. The chiroplasmonic band of Au heterodimers (HDs) at 525 nm exhibited a red shift pink to purple by an Au shell (@Au) deposition on it. The strong peak at 581 nm of HD@Au gets amplified with a g-factor of $1.21 \times 10^{-2}$. The formation of shell on HDs decreased the gap between two nanoparticles thereby enhanced the CD bands. The narrow CD spectra with the highest g-factor is obtained in HD@Au with g-factor $1.03 \times 10^{-2}$ and double-shelled Ag shell followed by Au shell on HD (HD@Ag@Au) with g-factor $0.44 \times 10^{-2}$ or Au shell followed by Ag shell on HD (HD@Au@Ag) with g-factor $0.28 \times 10^{-2}$. The strong CD bands of HD@Au were utilized for DNA detection. The DNA concentration quantitatively affects the HD@Au and optical activity of the resulting dispersions. The LOD of the HD@Au-based biosensor was low (Table 1) as compared to fluorometric QD assay (LOD = 1 nM) [84] and colorimetric biobar-code assay (LOD = 500 zM) [85]. Here, the enhanced CD signal in combination with polymeric chain reaction is attributed to the highly sensitive detection of DNA [86].

The Ag NP-Au NP heterodimers linked with antigen–antibody bridges exhibit chiroplasmonic properties. These dimers with scissor-like conformation were utilized for the bioanalysis of a cancer biomarker prostate-specific antigen and environmental pollutant microcystin-LR (Fig. 15). Here, the Au NPs and Ag NPs were modified with anti-microcystin-LR (MCLR) corresponding antibodies and BSA-anti-microcystin-LR, respectively, mixed together so that bridged heterodimers were formed. These heterodimers produce two prominent CD signals. The existence of plasmonic nanoparticles enhanced the chirality via coupling of angled conformation with circularly polarized light. The advantage of this study over previously studied cases is the explanation of the origin of chiral signals in heterodimers. The occurrence of chirality in bridged dimers depends on dihedral angle, $\theta$. The toxin MCLR results in the formation of dimers, which exhibit bisignate CD signal between 526 nm ad 399 nm. The LOD (Table 1) for MCLR is 250 times lower as compared to ELISA assay. In addition to this, the chiroplasmonic method was extended to the detection of prostate-specific antigen. The intensity of CD signal increases with concentration of PSA because of the assembly of heterodimers [87].

Plasmonic Au NPs were self-assembled with lanthanide-doped upconversion nanoparticles in the proximity of suitable DNA frame. The assembled pyramids exhibit

| Target                  | Nanomaterial                                      | Real sample       | Linear range                  | Detection limit | Ref     |
|-------------------------|--------------------------------------------------|-------------------|--------------------------------|-----------------|---------|
| Avian influenza A H4N6 virus | Antibody conjugated self-assembled gold nano structure and CdTe NPs | Chicken blood     | 100–0.01 HAU/50µL             | 0.0315 HAU/50 µL | [82]    |
| Fowl adenoviruses-9 (FAdVs-9) | Antibody conjugated self-assembled gold nano structure and CdTe NPs | Chicken blood     | 3.64 PFU mL$^{-1}$            |                 | [82]    |
| Bronchitis virus (IBV)   | Antibody conjugated self-assembled gold nano structure and CdTe NPs | Chicken blood     | $10^2$–$10^4$ EID/50µL        | 47.91 EID/50 µL | [82]    |
| Tropomyosin DNA          | Chiral Au NP trimer                               | Shell fish        | 0.1–15 ngmL$^{-1}$            | 21 pgmL-1 (S/N = 3) | [83]    |
| Prostate-specific antigen| Au heterodimers                                   | N/A               | 160 zM–1.6 pM                 | 17 zM           | [86]    |
| MCLR                    | Functionalized Ag NP-Au NP heterodimers           | Water from lake   | 0.002 to 0.5 ng/mL            | 0.0008 ng/mL    | [87]    |
| miRNA                   | Gold-upconversion nanoparticle                   | HeLa cells        | 0.073 to 43.65 fmol/10 $\text{pg}_{\text{RNA}}$ | 0.03 fmol/10 $\mu$g$_{\text{RNA}}$ | [32]    |
| ATP                     | Gold nanoparticle heterodimers                   | Live cell         | 1.5 to 4.2 mM                 | 0.2 mM          | [89]    |
| Zn$^{2+}$               | Au@AuAg yolk-shell nanorods                      | Human SH-SY5Y cells | $76.6 \times 10^{-6}$ M/10$^6$ cells to $211.5 \times 10^{-6}$ M/10$^6$ cells | $38.7 \pm 0.3 \times 10^{-6}$ M/10$^6$ cells | [90]    |

Table 1 Summary of chiral plasmonic gold nanostructure-based assays for various detection targets
chiroptical properties in the visible region, whereas the luminescence of upconversion nanoparticles get quenched by the LRET [88]. In the presence of miRNA, the DNA frame undergoes complete dissociation resulting in the separation of Au NPs and upconversion nanoparticle. The chiroplasmonic DNA-driven gold-upconversion nanopyramids exhibit dual signals of strong CD at 521 nm and remarkable luminescence in 500–600 nm. The nanoprobe was applied for the real-time detection of intracellular micro-RNA in living cells (Table 1). The intensity of CD spectra gets decreased, whereas that of fluorescence gets increased as the concentration of miR-21 gets increased in the cells. The sensitivity of CD detection is 4 times stronger than that of fluorescence detection. The CD signals are more prominent than luminescent signals, owing to the plasmonic enhancement via intrinsic chirality of DNA molecule and spin angular momentum of photon of the nanopyramids [32].

A chiral-aptasensor based on Au NP heterodimer could be used to quantify intracellular ATP levels. The chiral nanoassembly exhibit special CD signal with high background intensity and avoid optical interference with autofluorescence of cellular organelles. The decay of CD peak...
of heterodimers at 523 nm gets enhanced in the presence of ATP, owing to the chiroplasmic detection of ATP. This type of chiral Au NP sensors can be explored to monitor and investigate biological activity in living organisms [89].

The chiral Au@AuAg yolk-shell nanorods were synthesized using chiral D- and L-penicillamines. The circular dichroism of plasmonic core-satellite superstructures depend on concentration of chiral molecules, hot spots in the nanogaps of Au NRs, aspect ratio of Au NRs and coulombic interaction between Au NRs and D- or L-penicillamines. This superstructure with increased g-factor (0.021) and strong plasmonic CD intensity was employed as the photothermal probes for the quantitative detection of zinc ions in Parkinson’s cells (human SH-SY5Y cells). Here, the CD response of the nanoassembly originates from the coulombic interaction between Au NR and D/L-penicillamines. The plasmonic CD also gets enhanced due to the formation of hot spots created between nanogaps in the nanoassembly. The biosensor is designed in such a way that the CD response of the nanoassembly at 530 nm and 740 nm decreases with increase in concentration of Zn^{2+} ion in cells, suggesting the disassembly of the nanostructure via the dissociation of DNAzyme by Zn^{2+} ions. The nanoassembly shows good selectivity over other metal ions, negligible cytotoxicity and its CD response has no interference with proteins or peptides [90].

The chiroplasmic active optical fibre probe was assembled with gold and silver layers followed by immobilization of helicene enantiomers. The nanoassembly was able to produce chiroplasmonic waves via chirality transfer for helicene to plasmonic bands. The helicene used were rac-9-(3-pyridylethynyl)[7]helicene, rac-9-(4-pyridylethynyl)[7]helicene and racemic 2-(3-thienyl)[6]helicene. The complete information of a molecule in the solution manifests the dependence of surface chirality of glutathione onto the dissociation of DNAzyme by Zn^{2+} ions. The chiral glutathione-stabilized Au NPs were employed as the chiral selector for the enantioselective recognition of S-naproxen and R-naproxen [93].

The 3.3-nm chiral L-glutathione and D-glutathione-capped Au NPs named as L3.3 and D3.3, respectively, were used for the chiral recognition of amylloid beta peptide and inhibition of Aβ42 fibrillation occurs in Alzheimer’s diseases. The inhibition activity of Aβ42 fibrillation was greatly enhanced by glutathione stabilized Au NPs as compared to pristine glutathione molecules. The conformational changes associated with the interaction of AuNPs with Aβ42 were analysed using CD spectroscopy. The positive CD band at 195 nm and negative CD peak at 215 nm in the presence of D3.3 give a distinguished effect as compared to L3.3. It manifests the dependence of surface chirality of glutathione-coated Au NPs. Here, the Au NP prevents the structural change of Aβ42 from its bare conformation to β-sheet conformation in solution. The chiral glutathione-stabilized Au NPs have opened a new era for bioanalysis toward the fatal Alzheimer’s diseases [94]. The L- and D-glutathione can act as a reducing agent for the synthesis of gold nanoclusters (Au NCs.) The positive surface charge of Au NCs is attained by using the cationic ligand, 4,6-diamino-2-pyrimidinethiol. The spectra of both L- and D-glutathione-based Au NCs consist of mirrored CD peaks. The D-glutathione-protected Au NCs are less toxic to cells as well as organs. Thus, the application of it in vivo is safer than its L-glutathione-based counterparts. The study was extended into mice and identified the superiority of D-form over L-form [95]. Several stages of studies and modifications are necessary for the application of such nanoassembly in daily life.

The Au NPs were grafted with 2-mercaptoacetyl-L(D)-valine (L(D)MAV) and poly(acryloyl-L(D)-valine (L(D)-PAV) chiral molecules. The PAV-Au NPs give strong CD signals as compared to MAV-Au NPs because of the presence of a greater number of carboxyl groups in PAV-Au NPs. The mirror image CD spectra of L-PAV-Au NPs and D-PAV-Au NPs appear between 190 and 300 nm region. The internalization of chiral D-PAV-Au NPs in A549 and HepG2 cells were significantly greater than MAV-Au NPs and L-PAV-Au NPs. This chiral selective interaction arises as a result of preferable interaction between D-PAV molecules on Au NPs and L-phospholipid-based cell membrane [96].

Biodetection

A chiral assembly of sugammadex-Au NPs was used as a colorimetric chiral sensor for detecting lysine and asparagine enantiomers. In addition to this, it exhibits excellent catalytic activity towards the reduction of 4-nitrophenol with sodium borohydride [92]. The two-dimensional chiral arrangements of gold nanorods give strong second harmonic generation chiroptical effects, where chirality originates through inter-particle plasmonic interactions and mutual arrangement. These “meta-molecules” could aid as building blocks for a novel “watermark”, which could be read-out encoded pattern by means of a sophisticated nonlinear microscopy technique. The chiral N-acetyl-L-cysteine-modified Au NPs were employed as a chiral selector for the enantioselective recognition of S-naproxen and R-naproxen [93].
The plasmonic hot spots chiroptical activity of DNA-bridged chiral Au NPs/Au NRs dimers concede one to monitor the internalization of it in mammalian cells. The CD peaks in the visible region get reversed from negative to positive owing to the variation in the conditions of the dimers (Fig. 16a) inside and outside the cervical cancer HeLa cells, which is in good agreement with simulated CD spectra. The chirality reversal arises due to the variation in the elastic, electrostatic and van der Waals interactions resulting in the conformational changes of DNA-bridged Au NR dimers. The image from biological transmission electron microscopy (bio-TEM) proved that the dimers are dispersed in the cytosol of HeLa cells (Fig. 16b). Cryo-TEM tomography and its statistical analysis showed that the chirality of the dimers transformed from right-handed to left-handed conformation: the average dihedral angles are $-12.5 \pm 2^\circ$ (inside) and $+9.6 \pm 1^\circ$ (outside) of the cells (Fig. 16c and d). The chiroplasmonic gold nanostructures can be used to extend its contribution in photodynamic therapy and quantitative estimation of nanoscale interactions [97].

The chiral Au NPs with high polarizability can interact with circularly polarized light (CPL), which may induce stringent cellular response to chiral photons. The Au NPs stabilized with D/L-cysteine and functionalized with single-stranded DNA (ssDNA). The DNA-bridged chiral Au NPs were established for the distinction of neural stem cells (NCS) into neurons with the aid of CPL. The nanoassemblies enter the stem cells and trigger conformational changes, entangled with actin nanofibres. The nanoparticle denoted as $C_{30}(D)S_{5}-C_{20}(L)$ or $C_{30}(L)S_{5}-C_{20}(D)$ is fabricated by connecting 5-nm dangling nanoparticle with DNA-functionalized D- or L-cysteine-capped Au NPs having 30 nm or 20 nm (L- and D- stands for handedness of cysteine). The $C_{30}(D)S_{5}-C_{20}(L)$ nanoassemblies with a negative CD peak at 524 nm reconfigure to as $C_{30}(D)-C_{20}(L)S_{5}$ in the presence of Fox3mRNA with a positive CD. The CD of the nanoassemblies originates from the mirror asymmetry

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**Fig. 16** Chiral geometry of NP dimers. **a** TEM image of NP dimers in cell culture media; scale bar, 100 nm. **b** Bio-TEM images of NP dimers in the HeLa cells; scale bars, 100 nm. **c** TEM tomography images (bottom) of NP dimers both outside and inside cells with schematics of dimers’ geometry (top). **d** Statistical analysis of the dihedral angles $\theta$ for NP dimers inside and outside the cell as determined from cryo-TEM tomography images. The error bars correspond to the standard error of the mean ($n = 3$). The sign of the dihedral angle in these nanoscale structures was chosen in accord with the IUPAC convention. **e** Simulated CD spectra of NP dimers intra- and extracellular localization of NP dimers based on geometries from d. “ Reproduced with permission from Ref. 97. Copyright © 2017, Springer Nature”
of the nanoassembly [98]. The CPL impose polarization-reliant constrain on the nanofibres via nanoassemblies, owing to the differentiation of NCS. The biological effects of CPL based on Au NP–based nanoassemblies can be used for further exploration in bio detection applications [99].

The gold Janus nanoparticles capped with hydrophobic hexanethiolate ligands on one face and hydrophilic 3-mercapto-1,2-propanediol on the other face of the core. This Janus nanoparticle self-assembled into hollow vesicle-like nanostructures in both aqueous and organic phases. It exhibits circular dichroism absorption in the visible region, where the intensity of CD increases with increase in concentration of Au NPs. The gold Janus nanoparticle on interaction with L-alanine gives no change in plasmonic CD, whereas D-alanine shows negative plasmonic CD. The simulation study shows that a vertex formed on the Janus particle, which gives unique nature for it. The chiral vesicle-like Au NPs possess high enantioselectivity towards D-alanine compared to L-alanine. These types of Janus nanosystem could be extended for the identification, separation and biosensing of various biomolecules [100].

A chiral carbon nanotube electropolymerized with dopamine and loaded with Au NPs on it was utilized for the quantitative and qualitative electrochemical recognition of tryptophan and tyrosine isomers. Here, the dopamine directs the uniform dispersion of Au NPs on carbon nanotubes. The conductivity of Au NPs blended with chirality of carbon nanotube established as a novel chiral electrochemical biosensing system for amino acids [101]. The chiral plasmonic Au NR–based biosensors were employed for the hydrogen bonding (Hb) and zwitterionic electrostatic interactions (Zw-Es) between amine and carboxylic acid group in aqueous media. Such nanosensors are sensitive to conformations and selective to chirality at the nanoscale interfaces in Hb and Zw-Es interactions. The SERS technique could be coupled to chiral plasmonic sensors for developing multifunctional biosensors to investigate the chirality in biological systems [102].

**Conclusion**

Plasmonic nanoparticles utilizing CD spectroscopy have vital role in bioanalytical applications, and therefore, the development of effective biosensors and biodetection probes is of great importance. Bioanalytical tools exhibit importance in various economic background. The progress in this field is forging ahead to offer effective contribution in bioanalytical fields. The requirements for these include the following: (i) High selectivity. For many real samples such as pesticides containing vegetables, enantiomeric drugs, amino acids, peptides, proteins, genetic materials and cells, either broad-spectrum bioanalytical tools capable of unique identification or excellent target-specific probes are necessary. (ii) High sensitivity. The concentration levels of samples are generally below the detection limits of the bioanalytical tools, at which stage the biomolecules are usually undetectable using current platforms. The sensitive bioanalytical sensors allow us to investigate low concentration level of biomolecules. (iii) Low cost. The effectiveness of a bioanalytical probe depends on the extensive use of it among a large population, which is possible only if the probe is cost effective.

Importantly, the development of Au NP–based nanoprobe attains great improvements in bioanalytical field. Among the properties of Au NPs, chiroptical properties have attained much interest due to its high sensitivity and unique features. The interaction of chiral plasmonic Au NPs with electromagnetic radiation generates unique optical signatures. The CD responses of chiral plasmonic Au NPs depend on the interaction with different biomolecules and are distinctive like a finger print. Different types of chiral gold nanostructure produce a wide range of SPR wavelength. These optically tuneable nanosystems were functionalized with several biomolecules or synthetic molecules in various applications.
The combination of plasmonic properties and CD response of such nanoassembly could be utilized for fabricating bio-analytical tools.

Although a great progress in Au NP–based chiriptical responsive system has been achieved, many confronting challenges remains. For example, in the case of hot spot formation–based CD response, the practicability of maintaining distance between plasmonic nanoparticles using linker molecules in a macroscopic system is questionable. Secondly, the detection of large biomolecule-like proteins with the CD spectroscopy using plasmonic nanoparticle is difficult due to the possibility of decay of intensity of electric field between the nanoparticles. In addition to this, the mechanism of CD response is still not clear. Thirdly, the reproducibility of the detection systems is a challenging factor too. Moreover, the application of CD-responsive-based Au NPs in a point of care stage still demands interdisciplinary approach–based research by experts from various fields.

Declarations

Conflicts of interest The authors declare no competing interests.

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