How to rule out non-neoplastic hypercortisolemia (previously known as pseudo-cushing)

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The term non-neoplastic hypercortisolism (NNH) is used frequently nowadays, interchangeable with the pseudo-Cushing (pCS) state, in order to differentiate the functional non-neoplastic hypercortisolism that mirrors the neoplastic activation of the HPA axis, termed endogenous Cushing’s syndrome (CS). Diagnosis of neoplastic CS is often complex, especially when modest cortisol excess is present. Indeed, functional activation of the hypothalamic-pituitary-adrenal (HPA) axis can be caused by a variety of illnesses/conditions (e.g., psychiatric disorders, alcoholism, obesity, or polycystic ovary syndrome) [1]. Although some clinical features (such as bruisability, facial plethora, proximal myopathy, and large purple cutaneous striae) are highly specific for CS, they are not sensitive. Furthermore, NNH/pCS-related conditions may have clinical characteristics that overlap with CS, making the differential diagnosis very challenging [2].

Before ruling out NNH/pCS with second-line testing the possibility of a false-positive result from first-line testing for hypercortisolism should be considered. The low specificity of immunoassays may cause false-positive results from interfering circulating steroids (mainly cortisone) in late-night salivary cortisol and urinary-free cortisol. The use of estrogen-progesterone contraceptives or CYP3A4 inducing drugs may cause false-positive results at low-dose dexamethasone suppression test (LDDST). Special attention to the pre-analytical phase (clear and written instructions for sample collection, withdrawal of interfering drugs), use of mass spectrometry assays, and dexamethasone (dex) measurement may be useful tools to avoid false-positive results. In addition, physicians should also select the most appropriate test for the considered patient: avoid measuring circadian rhythm in shift workers, urinary cortisol in case of impaired renal function, and so on [3].

Several second-line tests have been proposed to distinguish NNH/pCS from neoplastic CS (Table 1), but there is still no agreement on the gold standard one among them. The combined dex-Corticotropin Releasing Hormone (dex-CRH) test assumes that only CS patients will sustain a cortisol response to CRH stimulation following dex suppression, allowing NNH/pCS and CS to be distinguished by specified thresholds. Proposed by Yanovski et al. in 1993, the combination of LDDST and CRH test for in-patients (the cohort of pCS subjects mainly presenting with affective disorders) with a cut-off for cortisol level at 15’ after CRH stimulation over 38 nmol/l, reported 100% sensitivity and specificity [4]. Despite stricter cortisol cut-offs and consideration of stimulated-ACTH levels, other studies did not corroborate this high diagnostic accuracy. Different study protocols (number and time of dex doses, ovine vs human CRH at fixed/weight-adjusted doses), different accuracy of cortisol assays for low levels, concomitant medications, even without measurement of serum dex, and differences in the enrolled population (number of cases, severity of hypercortisolism, subtypes of CS and NNH/pCS) can all contribute to different results [5]. Other tests, such as LDDST [6], midnight serum cortisol [7], and the desmopressin (DDAVP) test [8] indicated greater NNH/pCS detection accuracy. CRH test (without dex suppression) is useful in the differential diagnosis of...
ACTH-dependent CS; nonetheless, it performed poorly in Yanovski’s study in terms of discrimination between NNH/ pCS [4]. Arnaldi et al gave it new life by using a bimodal cut-off and generating promising results [9, 10] that would allow physicians to confirm CS and learn more about the cause of ACTH excess with a single test. Further studies are however needed to confirm these findings.

ACTH-secreting adenomas may aberrantly express vasopressin receptor 2 (VR-2). As a result, the synthetic analog DDAVP could only elicit ACTH response in Cush- ing’s Disease (CD) patients, because the normal pituitary expresses VR-3, which has a low DDAVP affinity [11]. A rise in ACTH levels above 6 pmol/L has been proposed to differentiate CD from NNH/pCS, achieving the best result in patients with mild hypercortisolism [12]. An alternative bimodal cut-off has been proposed to boost its accuracy [13]; both approaches demonstrated acceptable results in a larger retrospective cohort [14]. This test is simpler and less expensive than dex-CRH, and it is available in the US and other countries. In regions where CRH is not available, Araya et al. recommended a dex-DDAVP test as an alternative, employing cortisol as a less expensive readout [15]. In a direct comparison of dex-CRH and DDAVP, Pecori Giraldi et al. found that the latter performed better in confirming CD. As a result, a step-by-step approach involving a

Table 1  Second line tests in the differential diagnosis of CS vs. pCS. Δ = Delta, dex = dexamethasone, CRH = Corticotropin Realising Hormone, DDAVP = Desmopressin, CS = Cushing’s Syndrome, CD = Cushing’s Disease, pCS = pseudo-Cushing’s syndrome, ACS = Adrenal Cushing’s Syndrome, EAS = Ectopic ACTH Secretion, Se = Sensitivity, Sp = Specificity. *Paediatric patients

| Study               | Test     | CS           | pCS / controls | Cut-off                                      | Se (%) | Sp (%) |
|---------------------|----------|--------------|----------------|----------------------------------------------|--------|--------|
| Yanovski JA, 1993   | Dex-CRH  | CD 35, ACS 2, EAS 2 | pCS 19         | Cortisol at 15′: 38 nmol/L (1.4 µg/dL)        | 100    | 100    |
| Martin NM, 2006     | Dex-CRH  | CD 8, ACS 4  | pCS 3, controls 16 | Cortisol at 15′: 50 nmol/L (1.8 µg/dL)        | 100    | 88     |
| Gatta B, 2007       | Dex-CRH  | CD 17 (mild) | pCS 14         | Cortisol at 15′: 110 nmol/L (4 µg/dL)         | 100    | 86     |
| Erickson D, 2007    | Dex-CRH  | CD 21        | pCS 30         | Cortisol at 15′: 70 nmol/L (2.5 µg/dL)        | 90     | 90     |
| Pecori-Giraldi F, 2007 | Dex-CRH  | CD 29, ACS 3 | pCS 23         | Cortisol at 15′: 75 nmol/L (2.7 µg/dL)        | 100    | 82     |
| Reimondo G, 2008    | Dex-CRH  | CD 13, ACS 3 | pCS 15         | Cortisol at 15′: 44 nmol/L (1.6 µg/dL)        | 93.7   | 93.3   |
| Valassi E, 2009     | Dex-CRH  | CD 60        | pCS 41         | Cortisol at 15′: 38 nmol/L (1.4 µg/dL)        | 86.3   | 84.7   |
| Alwani RA, 2014     | Dex-CRH  | CD 53        | pCS 20         | Cortisol at 15′: 87 nmol/L (3.2 µg/dL)        | 94     | 100    |
| Batista D, 2008*    | Dex-CRH  | CD 11        | pCS 11 (Obese) | Cortisol at 15′: 88 nmol/L (3.2 µg/dL)        | 91     | 95     |
| Yanovski JA, 1993   | CRH      | CD 35, ACS 2, EAS 2 | pCS 19         | Sum of post-CRH cortisol levels > 3450 nmol/L (125 µg/dL) | 64     | 100    |
| Arnaldi G, 2009     | CRH      | CD 51, EAS 7 | pCS 26, controls 31 | Basal serum cortisol > 331 nmol/L (12 µg/dL) and ACTH peak > 12 pmol/L (54 pg/mL) | 91.3   | 98.2   |
| Tirabassi G, 2011   | CRH      | CD 30        | pCS 18, controls 12 | Basal serum cortisol > 331 nmol/L (12 µg/dL) and ACTH peak > 12 pmol/L (54 pg/mL) | 96.6   | 100    |
| Malerbi DA, 1996    | DDAVP    | CD 14        | pCS 11         | Δ-Cortisol ≥ 4 times intra-assay variation coefficient | 100    | 64     |
| Moro M, 2000        | DDAVP    | CD 76        | pCS 30, controls 67 | Δ-ACTH ≥ 6 pmol/L 0′ – 30′ (27.2 pg/mL) | 86.8   | 90.7   |
| Pecori-Giraldi F, 2007 | DDAVP   | CD 27        | pCS 21         | Δ-ACTH ≥ 6 pmol/L 0′ – 30′ (27.2 pg/mL) | 90     | 96.7   |
| Tirabassi G, 2010   | DDAVP    | CD 52        | pCS 28, controls 31 | Basal serum cortisol > 331 nmol/L (12 µg/dL) and Δ-ACTH > 4 pmol/L (18 pg/mL) | 75     | 89.8   |
| Tirabassi G, 2011   | DDAVP    | CD 30        | pCS 18, controls 12 | Basal serum cortisol > 331 nmol/L (12 µg/dL) and Δ-ACTH > 4 pmol/L (18 pg/mL) | 90.3   | 91.5   |
| Rollin G, 2015      | DDAVP    | CD 68        | pCS 56         | Peak ACTH of 15.8 pmol/L (36.8 pg/mL) | 90.8   | 94.6   |
| Araya V, 2017       | Dex-DDAVP | CD 36        | pCS 9, controls 7 | Δ-Cortisol ≥ 166 nmol/L (6 µg/dL) | 96.9   | 93.7   |
high level of diagnostic agreement, resulting in no CS misdiagnosis [10].

No single test among those proposed seems to guarantee a perfect distinction between CS and NNH/pCS patients. Furthermore, the wide variation in the population recruited, the NNH/pCS definition criteria, and the cut-offs used in different studies make it difficult to define a gold standard in this intricate differential diagnosis. To summarize, ruling out NNH/pCS-related disorders is required in cases of moderate hypercortisolism, equivocal clinical presentation, or suspected non-neoplastic hypercortisolism. According to the recent Pituitary Society consensus report, the patient’s clinical history and the duration of symptoms are of utmost importance in the differential diagnosis of NNH/pCS [17]. The specific treatment of the underlying condition that led to non-neoplastic hypercortisolism can achieve the recovery of HPA-axis function: clinical assessments and first-line testing should be repeated 3–6 months after baseline [1, 3, 17]. Second-line dynamic tests can be performed in referral centers with expertise in the differential diagnosis of CS and knowledge of their cut-offs. To rule out a CD diagnosis in a doubtful circumstance, a skilled utilization of dynamic diagnostics along with frequent clinical and hormonal surveillance is required [2, 17].

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Declarations

Conflict of interest All authors declare that they have no conflicts of interest for this topic that might be perceived as influencing the impartiality of the reported research.

Research involving human participants and patient consent Informed consent was obtained in individual studies, but not performed for this study.

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