Complete Genome Sequence of *Shewanella chilikensis* Strain DC57, Isolated from Corroded Seal Rings at a Floating Oil Production System in Australia

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**ABSTRACT** Here, we describe the genome of *Shewanella chilikensis* strain DC57, a facultatively anaerobic bacterium isolated from corroded seal rings at a floating oil production system in Australia. The genome of strain DC57 has a size of 4.91 Mbp and harbors 4,178 predicted protein-encoding genes.

*Shewanella chilikensis* is a facultatively anaerobic, Gram-negative, and rod-shaped bacterium (1). Members of the genus *Shewanella* have been reported to be associated with microbiologically influenced corrosion (2–6). *Shewanella* spp. have the ability to use a variety of electron acceptors, including nitrate, thiosulphate, and iron oxides (7), which indicates that these microorganisms can cause corrosion by different mechanisms.

*S. chilikensis* strain DC57 was isolated from corroded seal rings at a floating oil production system located in waters on the North West Shelf of Western Australia. Corrosion products were collected and inoculated in anaerobic phenol red broth medium (8). After positive growth in tubes incubated at 40°C, the culture was plated onto phenol red agar and incubated in anaerobic jars with AnaeroGen sachets (Oxoid). DC57 was purified using the streaking method until an axenic culture was obtained, as determined by microscopy. Single colonies were transferred to phenol red broth medium, and DNA was extracted using the DNeasy PowerSoil kit (Qiagen). Extracted DNA was sequenced with a combination of sequencing platforms. For Illumina sequencing, the library was prepared with the Nextera XT DNA sample preparation kit, and paired-end reads were generated on the MiSeq platform using the MiSeq reagent kit v3-600, as recommended by the manufacturer (Illumina, San Diego, CA, USA). For Nanopore sequencing, genomic DNA was prepared using the ligation sequencing kit 1D (SQK-LSK109) without any size selection. Sequencing was performed with the MinION Mk18 device and a SpotON flow cell R9.4, as recommended by the manufacturer (Oxford Nanopore Technologies, Oxford, UK). Base calling was performed using Albacore v2.3.1. Quality filtering of the reads was performed with fastp v0.19.4 (9), which resulted in 3,370,098 short reads (Illumina) with an average length of 245 bp, and 654,567 long reads (Nanopore) with an average length of 1,813 bp. A hybrid assembly strategy using Unicycler v0.4.7 (10) was applied to perform a de novo genome reconstruction, with overlap removal, circularization, and rotation. The assembly was validated with Bandage v0.8.1 (11). Default parameters were used for all software unless otherwise specified.

The complete genome of DC57 comprises a single circular chromosome (4,910,425 bp) with an overall GC content of 52.35% and 162-fold coverage. Annotation was performed with the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.10...
(12), which predicted 4,434 genes, including 104 tRNA genes, 25 rRNA genes, 4 noncoding RNA genes, 4,178 genes encoding proteins with predicted functions, and 123 genes encoding hypothetical proteins. Classification was performed by calculating the average nucleotide identity (ANI) with the Python module for ANI analyses (pyANI) v0.2.7 (13). This analysis revealed that DC57 is closely related to *S. chilikensis* strain JCS (GenBank accession number NZ_NIJM00000000.1) with an ANI value of 98.86%.

The genome analysis revealed the presence of the metal reduction pathway (MTR), two pathways for nitrate reduction (NAP and NAR), and genes for thiosulfate reduction (*phsA* and *glpE*), which could be related to the corrosive potential of the strain.

**Data availability.** The genome sequence of *Shewanella chilikensis* strain DC57 was submitted to GenBank under accession number CP045857. The raw reads were deposited in the NCBI SRA database under accession numbers SRR11492373 and SRR11492374.

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**REFERENCES**

1. Sucharita K, Sasikala C, Park SC, Baik KS, Seong CN, Ramana CV. 2009. *Shewanella chilikensis* sp. nov., a moderately alkaliphilic gammaproteobacterium isolated from a lagoon. Int J Syst Evol Microbiol 59: 3111–3115. [https://doi.org/10.1099/ijs.0.010918-0](https://doi.org/10.1099/ijs.0.010918-0).
2. Miller RB, Lawson K, Sadek A, Monty CN, Senko JM. 2018. Uniform and pitting corrosion of carbon steel by *Shewanella oneidensis* MR-1 under nitrate-reducing conditions. Appl Environ Microbiol 84:e00790-18. [https://doi.org/10.1128/AEM.00790-18](https://doi.org/10.1128/AEM.00790-18).
3. Miller RB, II, Sadek A, Rodriguez A, Iannuzzi M, Giai C, Senko JM, Monty CN. 2016. Use of an electrochemical split cell technique to evaluate the influence of *Shewanella oneidensis* activities on corrosion of carbon steel. PLoS ONE 11:e0147899. [https://doi.org/10.1371/journal.pone.0147899](https://doi.org/10.1371/journal.pone.0147899).
4. Philips J, Van Den Driessche N, De Paepe K, Prévotveau A, Granick JA, Arends JBA, Rabaey K. 2018. A novel *Shewanella* isolate enhances corrosion by using metallic iron as the electron donor with fumarate as the electron acceptor. Appl Environ Microbiol 84:e01154-18. [https://doi.org/10.1128/AEM.01154-18](https://doi.org/10.1128/AEM.01154-18).
5. Dawood Z, Brözel VS. 1998. Corrosion-enhancing potential of *Shewanella putrefaciens* isolated from industrial cooling waters. J Appl Microbiol 84:929–936. [https://doi.org/10.1046/j.1365-2672.1998.00414.x](https://doi.org/10.1046/j.1365-2672.1998.00414.x).
6. Schutz MK, Schlegel ML, Libert M, Bildstein O. 2015. Impact of iron-reducing bacteria on the corrosion rate of carbon steel under simulated geological disposal conditions. Environ Sci Technol 49:7483–7490. [https://doi.org/10.1021/acs.est.5b00693](https://doi.org/10.1021/acs.est.5b00693).
7. Fredrickson JK, Romine MF, Beliaev AS, Auchtung JM, Driscoll ME, Gardner TS, Nealon KH, Osterman AL, Pinchuk G, Reed JL, Rodionov DA, Rodrigues JLM, Saffarini DA, Serres MH, Spormann AM, Zhulin IB, Tiedje JM. 2008. Towards environmental systems biology of *Shewanella*. Nat Rev Microbiol 6:592–602. [https://doi.org/10.1038/nrmicro1947](https://doi.org/10.1038/nrmicro1947).
8. NACE International. 2014. Field monitoring of bacterial growth in oil and gas systems. TM0194. NACE International, Houston, TX.
9. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. [https://doi.org/10.1093/bioinformatics/bty560](https://doi.org/10.1093/bioinformatics/bty560).
10. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. [https://doi.org/10.1371/journal.pcbi.1005595](https://doi.org/10.1371/journal.pcbi.1005595).
11. Wick RR, Schultz MB, Zobel J, Holt KE. 2015. Bandage: interactive visualization of de novo genome assemblies. Bioinformatics 31:3350–3352. [https://doi.org/10.1093/bioinformatics/btv383](https://doi.org/10.1093/bioinformatics/btv383).
12. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. [https://doi.org/10.1093/nar/gkw569](https://doi.org/10.1093/nar/gkw569).
13. Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. Anal Methods 8:12–24. [https://doi.org/10.1039/C5AY02550H](https://doi.org/10.1039/C5AY02550H).