De-novo transcriptome assembly for discovery of putative microsatellite markers and transcription factors in black pepper (Piper nigrum)

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ABSTRACT

Black pepper (Piper nigrum L.) (2n= 52; Piperaceae), is a perennial, trailing woody flowering vine belonging to Piperaceae family. Genus piper has more than 1000 species (Jaramillo and Manos 2001). It is categorized into different types based on the degree of maturation and type of processing method used. It is mainly a self-pollinated plant and cultivated commercially by orthotropic stem cutting method (Krishnamoorthy and Va 2011). It is known as “King of spices” due to its global trade, widespread dietary, medicinal, preservative and insecticidal uses (Quijano-Abril et al. 2006). It has great nutritional and agricultural significance with antioxidant, anti-inflammatory and anticancerous properties (Gulcin I. 2005).

Black pepper is known to have originated in tropical evergreen forests of Western Ghats of India. It is one of the highly traded spices of the world and cultivated as a major cash crop in more than 30 tropical countries of the world (Ahmad et al. 2010, Tian et al. 2006). Vietnam is the world’s leading and largest producer and exporter of pepper, producing about 35% of the world’s P. nigrum crop. Globally, India contributes second highest area of black pepper (132000 ha, 2017) after Indonesia (181978 ha, 2017) (FAO 2017).

The productivity of the crop is affected by both biotic as well as abiotic stresses. Since, the crop is one of the costliest spice germplasms, these stresses lead to major economic losses. Currently, a few studies on this crop transcriptome/ RNA-Seq has been done. Also, the whole genome sequencing of black pepper is yet unavailable.

The continuous global rise in temperature and loss in the crop production due to abiotic stresses warrants the study on pathways and its mechanisms involved in abiotic stress tolerance. This will be useful for further investigation to elucidate stress improvement, deciphering pathways as well as mining of genic region SSR markers along with the primer generation which will further be useful in QTL mapping population and breeding programmes for germplasm improvement.

Key words: Black pepper, de novo assembly, Markers, RNA-sequence, Transcription factors

Black pepper (Piper nigrum L.) (2n = 52), a trailing woody flowering vine belongs to Piperaceae family. Genus piper has more than 1000 species (Jaramillo and Manos 2001). It is categorized into different types based on the degree of maturation and type of processing method used. It is mainly a self-pollinated plant and cultivated commercially by orthotropic stem cutting method (Krishnamoorthy and Va 2011). It is known as “King of spices” due to its global trade, widespread dietary, medicinal, preservative and insecticidal uses (Quijano-Abril et al. 2006). It has great nutritional and agricultural significance with antioxidant, anti-inflammatory and anticancerous properties (Gulcin I. 2005).

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MATERIALS AND METHODS

Data Set: Single end and paired end transcriptomic SRA data of *P. nigrum* were obtained from different genotypes available at National Centre for Biotechnology Information (http://www.ncbi.nlm.nih.gov). SRAToolkit was used to convert SRA data into Fastq format, resulting in separate files for forward and reverse data for each sample.

Pre-processing and de novo assembly: FastQC tool (http://www.bioinformatics.babraham.ac.uk/projects/fastqc/) (Andrews 2010) was used for the visualization of reads quality, before and after the pre-processing. Quality of the data was based on various parameters such as basic statistics, per sequence quality scores, per base sequence content, adapter content, per sequence GC content, sequence length distribution, per base N content, sequence duplication levels, over-represented sequences, per tile sequence quality and K-mer content. Trimmomatic tool (version 0.36) was used for removal of low quality reads (Bolger et al. 2014). Trimming of bases were done from 3’ and 5’ end, keeping headcrop as 10-12 and phred-score of 33. These high quality pre-processed reads were used for de novo transcriptome assembly using SOAPdenovo-Trans (version 0.99) assembler (Xie et al. 2014) that provides higher contiguity, faster execution and lower redundancy, which not only removes sequencing errors but also shortens ambiguous contigs (default ≤100 bp) caused by repeats. This was followed by CAP3 assembler for removal of redundant sequences (Huang and Madan 1999).

Homology Search, Annotation and Functional Characterization: Homology search of transcripts from *P. nigrum* transcriptome assembly were performed against NCBI non-redundant database (ftp://ftp.ncbi.nlm.nih.gov/blast/db/) using Blastx algorithm as standalone local ncbi-blast-2.2.31+ with threshold E-value 1e-3 (Altschul et al., 1990). For further research, Blast2GO tool (https://www.blast2go.com/) (Conesa et al., 2005) was used for their mapping and annotation. The functional classification was done using the Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) databases for broader overview of the crop species and the pathways involved.

Prediction of transcription factors involved in stress tolerance: Transcriptional factors from the generated assembly were predicted using PlantTFDB 4.0 (http://planttfdb.cbi.pku.edu.cn/download.php) (Jin et al. 2016). Blastx algorithm as standalone local ncbi-blast-2.2.31+ with threshold E-value 1e-6 (Altschul et al. 1990) were employed.

Mining of genic region putative molecular markers and primer designing: Mining of genic region putative SSR markers was done from de novo transcriptome assembly of *P. nigrum* using perl script of MISA (Micro SAtellite identification tool) (http://pgrc.ipk-gatersleben.de/misa/) (Thiel et al. 2003). The SSR loci containing repeat units of 1–6 nucleotides only were considered. For identification of genic region SSR markers, default parameters like ten repeating units for mononucleotides, six repeating units for dinucleotides and five repeating units for trinucleotides, tetranucleotides, pentanucleotides and hexanucleotides were considered. Maximum difference between two SSRs was kept as 100 bp. SSR specific primers were designed using Primer3 V0.4.0 (Untergasser et al. 2012).

RESULTS AND DISCUSSION

Pre-processing and de novo assembly: Entire public domain available genomic resource of species *P. nigrum* were mined successfully to discover transcription factors, molecular markers and to perform functional annotation. We retrieved 5 sets of single end and 4 sets of paired end transcriptomic SRA data of *P. nigrum* L. from NCBI (Table 1). A total of 516057410 single end and 157856524 paired end reads of black pepper genotype samples were retrieved with reads length 101 bp. After data pre-processing using Trimmomatics and visualization by FastQC, a total of 52545628 and 967085 low quality reads from single end and paired end samples, respectively, were removed. The remaining high quality reads were then used for the downstream analysis. The de novo transcriptome assembly by SOAP de novo-trans generated a total of 53690 transcripts with percent GC as 43.82% and GC count 14993012 bp. The N50 value of the contig was 688 bp.

Homology Search: Homology search of transcripts were performed using blastx to identify the similar genes present in the database. Out of 53690 transcripts, we found that 34996 transcripts showed similarity with other genes present in the database, while 15894 transcripts were novel as they didn’t showed any similarity or were without hits. A total of 639 transcripts were involved in mapping. Top hit species distribution revealed that maximum hits were found with *Nelumbo nucifera* i.e. 4938 transcripts, followed by *Macleaya cordata* and *Elaeis guineensis* in 3098 and 1597 transcripts, respectively. Total transcripts were found to be involved in 111 pathways. Maximum number of transcripts were found to be involved in biosynthesis of antibiotics i.e. 46, followed by 15 and 13 transcripts in purine metabolism and Glycolysis / Gluconeogenesis, respectively (Fig1). Blast2GO annotation results revealed that these transcripts gets categorized into three sub categories such as biological process, molecular functions and cellular components. In biological process, molecular function and cellular

Table 1. List of transcriptome data retrieved for analysis (PE: Paired End; SE: Single End)

| S.No. | Accession IDs | Description |
|-------|--------------|-------------|
| 1     | SRR1776865   | Vegetative phase plantlets (PE) |
| 2     | SRR1777719   | Root transcriptome (PE) |
| 3     | SRR1781514   | Root transcriptome (PE) |
| 4     | SRR408047    | Leaf transcriptome (PE) |
| 5     | SRR1164727   | SE |
| 6     | SRR1818148   | Fruit transcriptome (SE) |
| 7     | SRR1583631   | Root transcriptome (SE) |
| 8     | SRR3341858   | Root transcriptome (SE) |
| 9     | SRR3341859   | Root transcriptome (SE) |
DE-NOVO TRANSCRIPTOME ASSEMBLY IN BLACK PEPPER

stress tolerance: Since transcription factors are major players in development and adoptive responses in abiotic stress, thus they also play major role in putative candidate genes involved in transcriptional regulation of abiotic stress mediation. Transcription factors (TF) were identified by performing blast (Blastx tool) against the plant transcription factor database (PlantTFDB 4.0) using blastx tool. A total of 14005 transcription factors among 53690 transcripts and 39685 transcripts were novel as they were without hits with expected e-value ≤ 6. Most abundant transcription factors were bHLH, MYB, NAC, ERF, C2H2 and WRKY represented by 1470, 1418, 1089, 783, 672 and 620 transcripts, respectively. Top fifteen most

Fig 1. Top 16 KEGG pathways of transcripts of P. nigrum.

component, a total of 10230, 2316 and 6096 transcripts were found, respectively (Fig 2).

Prediction of transcription factors involved in

Fig 2. Gene ontology of transcripts. (Green, blue and yellow colour represent the biological process, molecular functions and cellular components, respectively).
abundant transcription factors identified from transcripts are represented in Fig 3.

Among the discovered TFs, basic helix-loop-helix (bHLHs) is well known regulator of abiotic defence mechanism. It activates the different types of genes which are involved in sensing of environmental signals by plant like hormone signalling. Myeloblastosis related proteins (MYB) is a huge and diverse family and is found mostly in all eukaryotes and are known to be highly expressed in drought. They are found to be involved in different processes, such as the control of cellular and organ morphogenesis, circadian rhythm, secondary metabolism as well as regulation of stomatal movements as a response to drought stress (Shin et al. 2011). C2H2 zinc-finger (C2H2-ZF) proteins are a large gene family in plants that participate in biotic and abiotic stress responses as well as various aspects of normal plant growth and development. This domain has also shown to mediate RNA, protein interactions and is known to be involved in overlapping responses to a variety of stress conditions including environmental stress regulation (Liu et al. 2015). TFs from other families such as HD-Zip (homeodomain-leucine zipper), GRAS (GAI – Gibberellin-acid insensitive), RGA – Repressor of GA1, SCR - Scarecrow), HSF (heat shock factor) and NF-Y (nuclear factor Y) which are found in our study, have been known to cope not only with abiotic stresses like salt, temperature and drought stresses but also with other stresses such as heat shock, oxygen deficiency, high light and nutrient deficiency ( Lan et al. 2017). Members of WRKY domain protein family have also been found in our study which are known to contain at least one conserved DNA-binding region including highly conserved a zinc finger motif \((CX_{4-7}CX_{22-23}HXH/C)\) and \(\text{WRKY}GQK\) peptide sequence (Pandey and Somssich 2009).

**Mining of genic region putative molecular markers and primer designing:** A total of 4770 SSRs were mined from the *de novo* transcriptome assembly of *P. nigrum*, Out of these 355 transcripts were with more than one SSR loci. There was less abundance (135) of compound SSR. Tri-nucleotide repeats were most abundant (1407) followed by mono-type (2573) and di nucleotide (754). Being coding region, these transcripts are expected to have higher abundance of tri-nucleotide repeats (Huang et al. 2016) (Table 2). In order to use the discovered SSR loci, primers were computed using PRIMER3 tool and 382 primer pairs were obtained successfully. These are ready to use primers for genotyping which requires wet-lab validation.

Mining of SSR markers from transcriptomic data can cater the need of crisis of molecular markers having advantage in terms of time and cost effectiveness. SSR serves as the most versatile molecular markers. In the present study, we discovered SSR loci from genic regions using transcriptomic data which offers several advantages in terms of stability and transferability. These markers can be used in linkage mapping, studies related to genetic variability and functional diversity (Kujur et al. 2013). Applications of such markers are evident in crops like tomato and pepper, sugarcane, basil, sesame, African oil palm and tea (Taxak et al. 2017). These SSR primers could represent a valuable and useful genomic resource of *P. nigrum* which will facilitate further advancements in genetic and molecular studies in the endeavour of better productivity of *P. nigrum* germplasm, especially in the era of rising abiotic stress.
Table 2  Summary statistics of mined genic region putative SSRs and primers from *P. nigrum*

| Information regarding SSRs | De-novo Assembly |
|-----------------------------|------------------|
| Sequences examined          | 53690            |
| Identified SSRs             | 4770             |
| SSR containing sequences    | 4377             |
| Sequences containing >1 SSR | 355              |
| SSRs present in compound Formation | 135             |
| Mono- nucleotide            | 2573             |
| Di- nucleotide              | 754              |
| Tri- nucleotide             | 1407             |
| Tetra- nucleotide           | 32               |
| Penta- nucleotide           | 3                |
| Hexa- nucleotide            | 1                |
| Total no. of primers        | 382              |

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