COMPARATIVE STUDIES ON THE PHYSIOCHEMICAL PROPERTIES, PHENOLIC COMPOUNDS AND ANTIOXIDANT ACTIVITIES IN 13 JAPANESE PLUM CULTIVARS GROWN IN THE SUBTROPICAL REGION OF CHINA

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Abstract. The physiochemical parameters and antioxidant properties of plum cultivars, grown in subtropical regions of China are little known. However, in this comparative study, these properties of 6 landraces and 7 introduced cultivars showed evaluated. The major nonvolatile constituents were a significant difference among cultivars. Color parameter values were strongly influenced by cyanidin 3-O-glucoside and cyanidin 3-O-rutinoside. Remarkably high antioxidant activity, high total phenolic, and ascorbic acid contents were found in black or purple flesh genotypes. Catechinic acid, the most important phenolic acid in Japanese plum, together with vanillic acid, caffeic acid and syringic acid, accounted for > 96% of the total phenolic content. The total phenolics and ascorbic acid concentration revealed significant contributions to antioxidant capacity detected by α, α-diphenyl-β-picrylhydrazyl (DPPH), \( r = 0.78^{**}, 0.73^{**} \) and ferric reducing antioxidant power (FRAP) \( r = 0.74^{**}, 0.57^* \). Moreover, there was a clear correlation between the total phenolic compounds and the ascorbic acid content \( r = 0.76^{**} \). Comparison of physicochemical characteristics and antioxidant profiles revealed that both introduced cultivars and landraces had good adaptability in the subtropical region.

Keywords: Prunus salicina Lindl, chemical compounds, nutrient content, subtropical cultivation, correlation analysis

Introduction

People consume fruits not only for their taste, but also their vital nutritional content (Liu, 2013), strong antioxidants and health promotion capabilities (Dai and Mumper, 2010; Goodarzi et al., 2018). Bioactive compounds including phenolics, anthocyanins and phytochemicals as well as vitamins E and C are considered to be beneficial properties of fruits (Mirmiran et al., 2009; Wang et al., 2018). Hence, a great deal of research has been carried out, in order to improve the quantity and quality of bioactive compounds in commonly consumed fruits.
Plum (*Prunus salicina* Lindl) commonly known as Japanese plum, is an important stone fruit crop commercially grown in China, Spain and USA. China ranks first in plum production (Li et al., 2015; Vlaic et al., 2018). Likewise, other fruits, various kinds of polyphenolic compounds such as phenolic acids, flavonols, anthocyanins (Turturică et al., 2018) and antioxidant activities have been identified in plum (Gil et al., 2002). Plum are good source of antioxidant that protects from biomolecular damage including aging caused by free radicals (Cefali et al., 2018). However, European plums (*Prunus domestica* L.) and Japanese plums (*Prunus salicina* Lindl.) have several different concentrations of phytochemicals, nutritional properties and antioxidant activities (Arion et al., 2014; Fanning et al., 2014; Jaiswal et al., 2013). Different cultivars have a greater variation of phenolic composition and concentration under various environmental conditions (Bochi et al., 2015; Wang et al., 2018). Several reports have been found on Japanese plum cultivars focusing on large size and homogeneous color (red, purple or yellow) (Lozano et al., 2009). Recently, more attentions have been focused on the functional properties of plum, their reasonable source of dietary-fiber, ascorbic acid, phenolic compounds, anthocyanins and other compounds with antioxidant properties (Fanning et al., 2014; Jaiswal et al., 2013; Kim et al., 2003).

In the past 20 years, many plum cultivars such as Blackamber, Wickson, Fiar, and Methely have been introduced, in order to enhance the plum production scale. Most of the plums are adapted to subtropical regions of Southeastern China. However, little information is available about the phytochemical properties of Chinese genotypes growing in subtropical regions of China (Byrne et al., 2000; Liu et al., 2007). Assuming that physicochemical characteristic, phenolic compounds and antioxidant properties of different Japanese Plums cultivars are influenced in subtropical regions. The main objectives of this study were to analyze the physicochemical characteristics, phenolic compounds and antioxidant properties of 13 Japanese plum cultivars grown in southeastern China.

**Materials and methods**

**Plant materials**

Fruit samples of 13 plum cultivars were harvested at commercial maturity stage during early June to end of August from Fujian Agriculture and Forestry Experiment Station Orchard at the hillock reservoir of Gutian county, Fujian, located in Southeastern China (N26°38'56.67", E118°49'1.02", Elevation: 323 m). The daily effective accumulated temperature is > 10 °C, and the annual average rainfall is about 5000 mm. Among 13 cultivars, 7 were introduced (Friar, Gariota, Blackamber, Santa Rosa, Elodrao, Methely, and Akihime) and 6 were landraces (Furongli, Hongnai, Yanzhili, and Xiguali as local cultivars of Fujian, Crown and Cuipingwannai were from the college of Horticulture, Fujian Agriculture and Forestry University. The dates of taking the samples are displayed in Table A1 in the Appendix. After harvesting, fruits were immediately transported to the laboratory and sorted in a refrigerator up to 4 °C, according to the uniformity of their firmness. Plums without stones were cut into several pieces and then stored at -80 °C until extraction. To ensure the uniformity of frozen material before further experiments, the fruits were ground to a fine powder by using liquid nitrogen.
Chemical standards

Standards for sucrose, glucose, fructose and sorbitol, as well as malic, citric, shikimic and fumaric acids were obtained from Fluka Chemical (New York, USA). Catechin, vanillic, gallic, caffeic, syringic, ferulic and chlorogenic acids, quercetin, cyanidin 3-O-glucoside and cyanidin 3-O-glucoside were purchased from Sigma-Aldrich (Oslo, Norway).

Gas chromatography-mass spectrometer (GC-MS) determination of sugar compounds and organic acids

Organic acids and sugars were extracted according to the protocol described by Liu et al. (2007) and Glew et al. (2003) The extraction of derivatives was carried out by using the method of Jiao et al. (2010). 1 μl of the derivative sample was injected into GC-MS using the split mode 50:1 and injector temperature was 230 °C. Ultra-pure helium served as carrier gas at the constant flow of 1.0 mL·min⁻¹. The oven was programmed at the following profile: initial temperature of the column was 70 °C (5 min hold), followed by an increasing the rate of 5 °C·min⁻¹ to 310 °C and then hold for 1 min. The mass operating parameters were: interface temperature was 250 °C, ion trap temperature was 200 °C, ion source was EI mode, electron energy was 70 eV, the solvent delay was 8 min. All data were obtained from the full-scan mass spectra within the range of 50 - 600 amu. The organic acids and sugars were identified by comparing the retention times and mass spectral data with the corresponding standards. Concentrations were determined according to the calibration of external standard solution.

High performance liquid chromatography (HPLC) analysis of phenolic compounds and anthocyanins

Plum extraction was carried out by the method of Gil et al. (2002). Samples were filtered through a 0.45 µm filter and then used for the analysis of phenolic compounds by HPLC (Kelebek et al., 2015). Anthocyanins were analyzed at 530 nm by HPLC (Usenik et al., 2008).

Antioxidant activity evaluation

2 g sample was extracted with 15 ml solution of 80% methanol (v/v) at 30 °C, then by an ultrasonic extraction with the power of 300 W for 40 min. Furthermore, it was centrifuged at the acceleration of 10000×g for 10 min (4 °C). For the analysis of the antioxidant activities of the plum fruits, combined supernatants of two replications were filtered by using a 0.45 µm filter. Three methods, i.e., the DPPH(α, α-diphenyl-β-picrylhydrazyl, DDPH) (Arion et al., 2014), ABTS(2, 2’-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), ABTS) (Maria do Socorro et al., 2010) and FRAP (ferric reducing antioxidant power) (Benzie and Strain, 1996) were used in order to test the antioxidant activity of the plum fruits.

Statistical analysis

Three replications were used for each treatment (n = 3) and statistical analysis were conducted by using SPSS software (version 21.0). Results were expressed as mean values ± standard deviation (SD). To determine whether the bioactive compounds would contribute to the antioxidant capacity, Pearson’s correlation coefficients were calculated at p < 0.05 and p < 0.01 confident levels for all variables. An analysis of
variance was performed followed by Student’s t-test (two-tailed distribution, unequal variance). Relationships between the physicochemical variables and either cultivar of P. salisida or antioxidants capacities of the known DPPH, FRAP and ABTS were analyzed using CCA (Canoco 4.5).

Results

Phenolic compounds and anthocyanins

Eight phenolic compound markers were identified in plums fruit at 280 nm. The acidic phenolics like caffeic acid, chlorogenic acid, ferulic acid and quercetin had more intense absorptance at 320 nm (Fig. 1). The concentration of the phenolic compounds of the 13 Prunus cultivars was shown in Table 1. The catechinic acid was the most abundant, especially in purple- or red-fresh cultivars Furongli, Xiguali and Yanzhili, which was two times higher than that 10 other cultivars. Additionally, abundant polyphenolic compounds were vanillic acid. Interestingly, the cultivars Xiguali, Gariota, Hongnai and Santa Rosa had outstanding high values of vanillic acid (Fig. 2). The hydroxycinnamic acid derivatives, caffeic acid, ferulic acid and chlorogenic in plums showed significant variation in 13 cultivars tested, in which caffeic acid predominated with the mean value of 7.68 µg·g⁻¹ fresh weight (FW). The acidic phenolics such as chlorogenic acid, ferulic acid and quercetin had low levels, and the neutral phenolic gallic acid tended to have low values as well. It should be noted that catechinic and syringic acids made 84.8% and 11.9% of the phenolic materials, respectively in Furongli.

Table 1. Correlation matrix between anthocyanins with chromatic parameters of plum cultivars

|                  | L'         | a'         | b'         | Chroma (C)  | Hue angle (H) |
|------------------|------------|------------|------------|-------------|---------------|
| Cyanidin 3-O-glucoside | -0.689**  | 0.707**    | -0.738**   | -0.632**    | 0.438**       |
| Cyanidin 3-O-rutinoside | -0.682**  | 0.708**    | -0.736**   | -0.648**    | 0.428**       |
| Total anthocyanins     | -0.697**  | 0.717**    | -0.748**   | -0.646**    | 0.441**       |

**P < 0.01 by Pearson’s test
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Figure 2. Contents of polyphenols in 13 plum cultivars (µg/g FW). The bars represent the mean ± standard deviation

Concerning the anthocyanins content, we observed that cyanidin 3-O-glucoside quantities were two times higher than that of cyanidin 3-O-rutinoside in plums (Figs. 3 and 4). Cyanidin 3-O-glucoside was the predominant compound among anthocyanins and detected in Xiguali, Methely and Blackamber, which corresponds to a mean content of 335.0 mg·100 g⁻¹ FW among the 10 cultivars tested. The cyanidin 3-O-rutinoside was also detected in abundance and its mean value was 161.5 mg·100 g⁻¹ FW in 10 cultivars. In all cases, the levels of cyanidin 3-O-glucoside, cyanidin 3-O-rutinoside and total anthocyanins correlated negatively or positively (P < 0.01) with each of the color parameters L*, a*, b*, C and H, respectively (Table 1). It seems that the highest values of a* and H corresponded positively to the samples with the highest anthocyanins content. The lowest values of b*, L* and C were negatively correlated with the anthocyanin levels. The darker the fruits, the more negative was the correlation of the color parameters of L*, b* and C to the amount of total anthocyanins, cyanidin 3-O-glucoside and cyanidin 3-O-rutinoside in plums. These results showed that chromatic parameters have a significantly correlate with the evolution of fruit color and anthocyanins levels and indicated that color parameters could be used to monitor pigment evolution and anthocyanins content of plum cultivars.

Figure 3. Chromatogram of anthocyanins of plum cultivars. (1) Cyanidin 3-glucoside, (2) cyanidin 3-rutinosid, (3) unknown
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Total phenolics, ascorbic acid and antioxidant capacity

Thirteen cultivars were analyzed with respect to their content of total phenolics, ascorbic acid and antioxidant capacities (Table 2). Total phenolic contents were classified into three categories: low (150-200 mg of gallic acid equivalent (GAE) per 100 g FW), medium (200-250 mg of GAE per 100 g FW) and high (> 250 mg of GAE per 100 g FW). The highest contents of total phenolics were observed among the red- or purple-fresh fruits of Xiguali, Furongli and Yanzhili. The yellow-fresh fruits of Crown and Cuipingwannai had the lowest total phenolics except for Blackamber.

Table 2. Antioxidant capacity, total phenolic content and ascorbic acid content of 13 plum cultivars

| Cultivars    | Total phenolic contents (mg·100 g⁻¹ FW) | Ascorbic acid (mg·100 g⁻¹ FW) | DPPH (%)     | FRAP (µmol·g⁻¹ FW) | ABTS (%) |
|--------------|----------------------------------------|--------------------------------|--------------|--------------------|----------|
| Gariota      | 192.68±15.62cd                         | 20.04±1.78ef                   | 93.95±0.31c  | 10.51±1.53b        | 99.84±0.00a |
| Santa Rosa   | 229.38±9.29bc                          | 29.31±1.03c                   | 93.30±0.10de | 11.49±1.18ab       | 99.63±0.08a |
| Cuipingwannai| 174.82±16.03d                          | 7.92±0.46i                    | 93.46±0.11cd | 8.17±1.64c         | 95.78±3.02b |
| Blackamber   | 154.38±18.61e                          | 18.83±0.49f                   | 93.18±0.10e  | 7.46±1.51c         | 84.69±4.01c |
| Hongnai      | 216.74±29.49e                          | 21.71±1.10e                   | 93.75±0.06cd | 10.78±1.79b        | 99.68±0.23a |
| Akihime      | 208.24±12.09c                          | 5.74±0.63j                    | 93.79±0.26cd | 10.73±1.74b        | 99.20±0.57a |
| Friar        | 187.04±11.40cd                         | 10.63±0.85h                   | 92.49±0.41f  | 4.79±0.82d         | 55.10±4.35d |
| Crown        | 170.73±4.77d                           | 12.16±0.24h                   | 93.50±0.15cde| 7.77±1.46d         | 85.22±3.83c |
| Yanzhili     | 257.53±16.57b                          | 33.38±1.40b                   | 94.52±0.00b  | 10.16±1.11b        | 99.44±0.33a |
| Eldorado     | 170.91±17.61d                          | 16.85±0.63g                   | 93.67±0.26cd | 7.35±1.55c         | 96.06±4.23a |
| Furongli     | 281.05±9.59a                          | 21.78±1.32e                   | 94.48±0.11b  | 13.12±0.82a        | 99.84±0.00a |
| Methely      | 206.94±6.91c                           | 26.29±1.33d                   | 93.71±0.21cd | 11.74±0.64b        | 99.68±0.13a |
| Xiguali      | 291.53±8.73a                           | 44.00±1.82a                   | 96.14±0.25a  | 12.12±1.12ab       | 99.79±0.08a |

Values represent mean ± standard deviation (n = 3). Different letters denote significant differences in the level of P < 0.05 by Duncan’s test.
In this study, 13 different cultivars were investigated for their total antioxidant capacity, using the DPPH, FRAP and ABTS methods as shown in Table 2. In general, the antioxidant activities of the black or red landraces Furongli and Xiguali were higher than those of the other cultivars. The results showed statistically significant correlation in antioxidant capacity on DPPH within 13 cultivars. The highest DPPH radical scavenging activities for the 13 cultivars fruit extracts was measured in Xiguali, followed by Furongli and Yanzhili, while the lowest values were observed in Friar. The reduction power towards ABTS radical cation was more than 95% in almost all plum cultivars except for Blackamber and Friar. The values obtained by FRAP were less than 10 μmol·g⁻¹ FW for yellow-flesh cultivars like Crown, Cuipingwannai, Blackamber, Eldorada, and Friar. The FRAP values of red-flesh or red-skin cultivars were higher than those of yellow-flesh cultivars, except Akihime. In general, the measurement from the three methods (DPPH, ABTS and FRAP) demonstrated that Furongli and Xiguali had the highest levels of antioxidants but the lowest was in the Friar cultivar.

Relationships between organic acids and sugars

Correlation analysis of the typical biological characteristics on the components of sugar and organic acid in 13 plums cultivars (Table 3), found that, in terms of biological characteristics, succinic played a leading role, showed a extremely significantly positive correlation with Citric \((r = 0.98^{**})\) and sorbitol \((r = 0.54^*)\). In addition, Shikimic showed a significantly positive correlated with Cis-aconitic \((r = 0.59^*)\) and a significantly negatively correlated with Ascorbic \((r = -0.57^*)\). In the relationship in soluble sugars, sucrose, fructose and glucose were the main three sugars in plums, and there was a significant correlation among them. In addition, sorbitol and glucose were also significantly correlated \((r = 0.67^{**})\). It can be seen that the interdependent among soluble sugars is significantly stronger than that among organic acids, which may be due to a circulation of the metabolic pathways of these three sugars while not exist in organic acids. It is worth noting, the total phenolics and ascorbic indicated a significant positive correlation \((r = 0.76^{**})\).

### Table 3. Correlation matrix between organic acids and sugars in 13 plum cultivars

|                  | Malic | Citric | Shikimic | Succinic | Fumaric | Cis-aconitic | Sucrose | Glucose | Fructose | Sorbitol | Ascorbic |
|------------------|-------|--------|----------|----------|---------|--------------|---------|---------|----------|----------|----------|
| Citric           | -0.06 |        |          |          |         |               |         |         |          |          |          |
| Shikimic         | 0.24  | 0.27   |          |          |         |               |         |         |          |          |          |
| Succinic         | -0.06 | 0.98** | 0.20     |          |         |               |         |         |          |          |          |
| Fumaric          | 0.41  | 0.34   | 0.40     | 0.37     |         |               |         |         |          |          |          |
| Cis-aconitic     | 0.00  | -0.12  | 0.59*    | -0.15    | 0.36    |               |         |         |          |          |          |
| Sucrose          | 0.10  | 0.18   | -0.18    | 0.27     | -0.01   | -0.11         |         |         |          |          |          |
| Glucose          | 0.25  | 0.09   | 0.35     | 0.14     | 0.05    | 0.00          | 0.75**  |         |          |          |          |
| Fructose         | 0.29  | 0.14   | 0.46     | 0.17     | 0.14    | 0.29          | 0.70**  | 0.97**  |          |          |          |
| Sorbitol         | 0.28  | 0.51   | 0.37     | 0.54*    | 0.17    | -0.35         | 0.52    | 0.67**  | 0.65     |          |          |
| Ascorbic         | -0.21 | -0.53  | -0.57*   | -0.44    | -0.47   | -0.21         | 0.55*   | 0.34    | 0.22     | -0.16    |          |
| Total phenolics  | -0.13 | -0.47  | -0.45    | -0.42    | -0.45   | 0.22          | 0.37    | 0.04    | 0.03     | -0.28    | 0.76**   |

* and ** indicate correlated significantly at the level of P < 0.05 and P < 0.01 respectively

Relationships between bioactive phytochemicals and antioxidant capacity

The amount of total phenolics, ascorbic acid, individual phenolic compounds and anthocyanins showed a good linear relationship with each other (Table 4). The total phenolics and ascorbic acid concentration of the 13 plum cultivars correlated
significantly with the antioxidant capacity detected by DPPH ($r = 0.78^{**}, 0.73^{**}$) or FRAP ($r = 0.74^{**}, 0.57^{*}$), but not so significantly by ABTS ($r = 0.41, 0.41$). Moreover, there was a significant correlation between the total phenolic compounds and the ascorbic acid ($r = 0.76^{**}$). We found that catechinic acid had a stronger contribution to the antioxidant constituents than DPPH ($r = 0.63^{*}$), FRAP ($r = 0.55^{*}$) and the total phenolics ($r = 0.77^{**}$). The high significant correlations between caffeic acid and DPPH ($r = 0.64^{*}$), ascorbic acid ($r = 0.64^{*}$) and vanillic acid ($r = 0.63^{*}$) indicated that catechinic- and caffeic acids contributed more than ascorbic acid to the total phenolics and antioxidant activity. Cyanidin 3-O-glucoside also shows a significantly correlated ($r = 0.94^{**}$) with cyanidin 3-O-rutinoside in the flesh of plum cultivars (Fig. 5; Table 5). In addition, a positive significant correlation between the ascorbic acid, cyanidin 3-O-glucoside ($r = 0.63^{*}$), and cyanidin 3-O-rutinoside ($r = 0.60^{*}$) was identified. Anthocyanins were important bioactive compounds contributing to ascorbic acid. The clear trend between the antioxidant capacity and the Prunus genotypes was analyzed by Canonical correspondence analysis (CCA) (Fig. 5). The antioxidant capacity of the purple- or red-flesh cultivars Furongli, Xiguali and Yanzhili showed a bigger variation than that of the other cultivars and correlated with the contents of total phenolics (Fig. 5). In general, black and red Prunus cultivars had a higher antioxidant capacity compared with yellow-flesh genotypes such as Methely, Santa Rosa, Akihime, Crown, Hongnai, and Cuipingwannai (Table 4).

**Discussion**

The presence of polyphenolics correlated with previous investigations in which they showed that plums were rich in phenolic compounds and there are great differences among the phenolic contents in Prunus. Salicina and P. domestica (Arion et al., 2014; Mubarak et al., 2012; Slimestad et al., 2009). The occurrence of catechinic acid, vanillic acid and syringic acid has already been reported in plums (Jaiswal et al., 2013). However, some reports also showed smaller amounts of chlorogenic, caffeic acid and quercetin (Kim et al., 2003; Mubarak et al., 2012).
Table 4. Correlation matrix between phenolic compounds, anthocyanins, total phenolics content, ascorbic acid and antioxidant capacities using DPPH, FRAP and ABTS

|                  | Gallic acid | Catechinic acid | Caffeic acid | Vanillic acid | Chlorogenic | Syringic acid | Ferulic acid | Quercetin | Cyanidin 3-O-glucoside | Cyanidin 3-O-rutinoside | Total phenolics | Ascorbic acid | DPPH | FRAP |
|------------------|-------------|-----------------|-------------|--------------|-------------|--------------|--------------|-----------|-------------------------|--------------------------|------------------|---------------|------|------|
| Catechinic acid  | -0.26       |                 |             |              |             |              |              |           |                         |                          |                  |               |      |      |
| Caffeic acid     | 0.5         | 0.6             |             |              |              |              |              |           |                         |                          |                  |               |      |      |
| Vanillic acid    | 0.29        | 0.1             | 0.63*       |              |             |              |              |           |                         |                          |                  |               |      |      |
| Chlorogenic      | -0.08       | 0.22            | 0.26        | 0.06         |             |              |              |           |                         |                          |                  |               |      |      |
| Syringic acid    | -0.26       | 0.85**          | -0.29       | -0.21        | -0.04       |              |              |           |                         |                          |                  |               |      |      |
| Ferulic acid     | 0.14        | 0.29            | 0.43        | 0.72*        | -0.17       | 0.1          |              |           |                         |                          |                  |               |      |      |
| Quercetin        | -0.13       | -0.1            | 0.25        | 0.49         | -0.17       | -0.19        | 0.78**       |           |                         |                          |                  |               |      |      |
| Cyanidin 3-O-glucoside | 0.02     | 0.05            | 0.52        | 0.17         | 0.22        | -0.21        | 0.05         | -0.21     |                         |                          |                  |               |      |      |
| Cyanidin 3-O-rutinoside | -0.01   | 0.17            | 0.37        | 0.15         | 0.17        | -0.04        | 0.05         | -0.23     | 0.94**                  |                          |                  |               |      |      |
| Total phenolics  | 0.2         | 0.77**          | 0.48        | 0.36         | 0.23        | 0.48         | 0.32         | -0.25     | 0.33                     | 0.39                     |                  |               |      |      |
| Ascorbic acid    | 0.06        | 0.39            | 0.65*       | 0.53         | 0.1         | -0.01        | 0.27         | -0.13     | 0.63*                   | 0.6*                     | 0.76**            |               |      |      |
| DPPH             | -0.02       | 0.63*           | 0.64*       | 0.46         | 0.23        | 0.33         | -0.09        | 0.46      | 0.39                     | 0.78**                   | 0.73**            |               |      |      |
| FRAP             | 0.09        | 0.55*           | 0.44        | 0.45         | 0.12        | 0.39         | 0.32         | -0.05     | 0.13                     | 0.19                     | 0.74**            | 0.57**        | 0.66** |      |
| ABTS             | 0.13        | 0.28            | 0.36        | 0.34         | 0.17        | 0.15         | 0.07         | -0.02     | -0.01                    | -0.02                    | 0.41              | 0.41          | 0.59  | 0.81**|

* and ** indicate correlated significantly at the level of P < 0.05 and P < 0.01 respectively

Table 5. Correlation matrix between anthocyanins with chromatic parameters of plum cultivars

|                  | L'   | a'   | b'   | Chroma (C) | Hue angle (H) |
|------------------|------|------|------|------------|---------------|
| Cyanidin 3-O-glucoside | -0.689** | 0.707** | -0.738** | -0.632** | 0.438**       |
| Cyanidin 3-O-rutinoside | -0.682** | 0.708** | -0.736** | -0.648** | 0.428**       |
| Total anthocyanins   | -0.697** | 0.717** | -0.748** | -0.646** | 0.441**       |

* and ** indicate correlated significantly at the level of P < 0.05 and P < 0.01 respectively
The plums contain mainly cyanidin 3-O-glucoside and cyaniding 3-O-rutinoside (Kim et al., 2003). Slimestad et al. (2009) reported that cyanidin 3-O-rutinoside represents more than 60% of the total anthocyanin content. It should be noted that cyanidin 3-O-glucoside and 3-O-rutinoside in fruit extracts of the yellow-flesh cultivars Crown, Akihime and Cuipingwannai were not detected due to their low contents, which is consistent to previous findings (Kim et al., 2003; Lozano et al., 2009).

The ascorbic acid content showed significant differences \((p < 0.05)\) in the 13 selected cultivars. The ascorbic acid detected in our experiments was higher rather than previous reports. Most of the fruits contained considerable amounts of ascorbic acid (Asami et al., 2003; Fang et al., 2017) which were higher than 10 mg·100 g\(^{-1}\), and apparently higher than that in Spain (Gil et al., 2002). These results demonstrated that plum cultivars grown in subtropic areas have higher level of ascorbic acid than those grown in other regions.

Many assays were used for the determination of antioxidant activity in fruits and vegetables (Bustos et al., 2018; Moo-Huchin et al., 2015; Nayak et al., 2015). These observations indicated that purple- or red-flesh Prunus cultivars had higher contents of the total phenolics (except cultivar Gariota) compared with other cultivars. However, Other researchers have also reported that plum is rich in total phenolic contents (Gil et al., 2002; Granato et al., 2018). Unfortunately, the range of total phenolics in plum differs among different reports due to their cultivar differences, agricultural practices, growth conditions and seasons (Arion et al., 2014; Kim et al., 2003).

These results indicated that the total phenolic compounds and ascorbic acid have a strong contribution to the antioxidant activity as also described in previous reports (Kim et al., 2003; Lozano et al., 2009; Mubarak et al., 2012). In some other reports, no correlation between antioxidant capacity and vitamin C was found (Hassimotto et al., 2005), even a negative influence of vitamin C was detected (Kalt et al., 1999). These similar results showed that cyanidin 3-O-rutinoside and cyaniding 3-O-glucoside were responsible for the pigmentation of the fruits (Kim et al., 2003; Raynal et al., 1989; Tomás-Barberán et al., 2001). These results were supported by previous reports which have shown that the main phytochemicals responsible for the antioxidant capacity in plums were total phenolics, ascorbic acid, and phenolic compounds (Gil et al., 2002; Vlaic et al., 2018), although other compounds may also be involved (Kim et al., 2003).
task to choose the most appropriate method to evaluate high-quality cultivars. Comparison of physicochemical characteristics and antioxidant profiles revealed that both introduced cultivars and landraces had good adaptability in the subtropical region. Thus, in order to achieve the maximum health-beneficial effects of plum consumption and their optimal organoleptic and nutritive properties, it would be advisable to get the cultivars with a high content of ascorbic acid, catechinic acid, caffeic acid, and syringic acid. The next work, we will conduct a deeply study on the main polyphenols and antioxidant activities in plums, and explore the best hybrid breeding model of plums to improve the functional components and nutrient content of fruits.

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**APPENDIX**

**Table A1. The dates of collecting samples in this work**

| Cultivar     | Time     |
|--------------|----------|
| Xiguali      | 6/20/2018|
| Methely      | 7/25/2018|
| Furongli     | 8/5/2018 |
| Eldorado     | 7/25/2018|
| Yianzhili    | 6/8/2018 |
| Crown        | 8/5/2018 |
| Friar        | 7/25/2018|
| Akihime      | 8/10/2018|
| Hongnai      | 8/10/2018|
| Blackamber   | 8/10/2018|
| Cuipingwannai| 8/20/2018|
| Santa Rosa   | 7/25/2018|
| Gariota      | 7/20/2018|