Research article

The ameliorative activity of Chrysobalanus orbicularis in streptozotocin-induced type II diabetes mellitus rat model

Lisa Ilobekemen Ekakitie a, Babatunji Emmanuel Oyinloye a, b, Basiru Olaitan Ajiboye a, *, 1

a Phytomedicine, Biochemical Toxicology and Biotechnology Research Laboratories, Department of Biochemistry, Afe Babalola University, Ado-Ekiti, Ekiti State, Nigeria
b Biotechnology and Structural Biology (BSB) Group, Department of Biochemistry and Microbiology, University of Zululand, Kwa Dlangezwa 3886, South Africa

1. Introduction

Diabetes mellitus is a significant issue on the earth’s planet today and characterized by clinical abnormalities (Piero et al., 2015). This condition happens because of glucose homeostasis impairment, characterized by supreme or relative insufficiency of the hormone insulin or inefficacy of its fringe activity (Amira et al., 2016). This is showed with constant hyperglycemia a noteworthy side effect of diabetes mellitus, influencing carbohydrate, fat, and protein digestion (Sorour et al., 2018).

Among the various kinds of diabetes mellitus, type II is the most common, both at international and in sub-Sahara Africa, with Nigeria positioned at fifth (Ajiboye et al., 2020). Hindrance of insulin signalling in skeletal muscle, liver and fat tissues because of the advanced insulin resistance is the primary driver of type II diabetes mellitus. This disorder advances and prompts organ dysfunction and unexpected death (Joudaki and Setorki, 2019). Exogenous variables like ecological factors just as hereditary qualities, diet and weight contribute extraordinarily to the improvement of this ailment (Piero et al., 2015). Lately, characteristic spices have been utilized broadly for the administration of type II diabetes mellitus and its difficulties and it has been accounted for that it is more secure with next to zero side effects (Ibitoye et al., 2017) consequently the inclination of elective medications over manufactured drugs (Kooti et al., 2016).

In Africa there are a few restorative spices used to oversee diabetes mellitus and its intricacy, this organic assorted variety converts into an auxiliary decent variety (Pires et al., 2016) empowering the disclosure of new types of bioactive substances with antidiabetic potential documented by Paracampo (2011). In Nigeria, there are various plants under this classification and a model is Chrysobalanus orbicularis, which belongs to the family Chrysobalanaceae, known for its crucial therapeutic qualities locally in the Southern part of Niger Delta in Nigeria just as they are utilized for the mainstream Niger Delta pepper soup. The plant is generally called “Omillo”, logical data on the impacts of this generally expended leaf has not been settled. Showing the scientific efficacy of the

Chrysobalanus orbicularis is a medicinal plant present in West Africa in the Itsekiri speaking part of Nigeria. It is used conventionally in diabetes mellitus management. This research investigates the ameliorative activity of the aqueous leaf extract of C. orbicularis in a streptozotocin-induced diabetic rat model. Freshly prepared streptozotocin (40 mg/kg body weight [BW]) was administered intraperitoneally to induce diabetes. Three diabetic groups were placed on aqueous leaf extract of C. orbicularis at 11.076, 22.134, and 44.268 mg/kg BW respectively; a group was placed on metformin (44.28 mg/kg BW), and the other two groups were the diabetic control and normal control. The experiment lasted for 28 days, thereafter, fasting blood glucose levels and body weight variations were recorded. Also, glycogen level, antioxidant enzyme, hexokinase and glucose-6-phosphatase activities, malonaldehyde (MDA) as well as glucose transporters 2 and 4 levels were analyzed using standard procedures. Diabetic rats administered aqueous extract of C. orbicularis leaf significantly (p < 0.05) decreased the fasting blood glucose and MDA levels, and glucose-6-phosphatase activity. In addition, administration of aqueous extract of C. orbicularis leaf to diabetic rats demonstrated a momentous increase in liver glycogen level, superoxide dismutase, catalase, glutathione peroxidase, reduced glutathione, glutathione transferase, and hexokinase activities as well as GLUT-2 and GLUT-4 levels. The data from this study suggest that the aqueous extract of C. orbicularis leaf may be beneficial in the management of diabetic mellitus and its secondary effects.

### Keywords:
- Diabetes mellitus
- Streptozotocin
- Chrysobalanus orbicularis
- Body weight
- Antioxidant activity

---

* Corresponding author.
E-mail address: bash1428@yahoo.co.uk (B.O. Ajiboye).

1 Current Address: Phytomedicine and Molecular Toxicology Research Laboratory, Department of Biochemistry, Federal University-Oye-Ekiti, Ekiti State, Nigeria.

https://doi.org/10.1016/j.heliyon.2021.e06596
Received 19 December 2020; Received in revised form 21 February 2021; Accepted 23 March 2021
2405-8440/© 2021 Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
plant reinforcement as of late an investigation was done on the antioxid-
and inhibitory activities of enzymes linked to type II diabetes the
role of the novel *Chrysobalanus orbicularis* by Ekakitie et al. (2020)
indicating the scientific efficacy of the plant. In light of an ethno-
botanical study nonetheless, this existing study assesses the ameliorative action of
aqueous leave extract of *Chrysobalanus orbicularis* in streptozotocin-prompted type II diabetes mellitus rats model.

2. Materials and methods

2.1. Chemicals and reagents

Streptozotocin and other chemicals used in this study were purchased
from Sigma Chemical Co., St Louis, MO, USA. Although, the enzyme
assy kits were secured from Randox Laboratories Ltd., Antrim, UK. It is
important to note that analytical grade was taken into consideration for
all the chemicals and reagents.

2.2. Plant materials and authentication

*Chrysobalanus orbicularis* leaf was secured in March 2019 at exactly 9
a.m in Itsekiri layout, Sapele, Delta State, Nigeria. The leaf was subse-
quent authenticated at Forestry Research Institute of Nigeria (FRIN)
Ibadan, Nigeria, and FHI:112232 was assigned as its herbarium number.

2.3. Preparation of plant materials

Firstly, the obtained leaves were washed under running water and
dried at 25 °C for two weeks and then pulverized into fine particles (using
a food processor). Then, two hundred grams of the powdered sample was
extracted in two thousand millilitres of distilled water by maceration,
according to the method of Nwozo and Oyinloye (2011) for 72 h, filtered
with cheesecloth, and freeze-dried to obtain the dried extract. Based on
the ethnobotanical survey, a cup of the filtrate was also dried and 0.2523
g was obtained as the yield. This is corresponding to the dose con-
sumption of a 70 kg man. This was further extrapolated to get 22.134
mg/kg. The obtained yield was kept at 4 °C using a universal bottle for
further studies.

2.4. Experimental animal

Sixty male albino rats, weighing between 130 to 140 g were secured
from the Animal Holding unit of Afe Babalola University, Ado-Ekiti,
Nigeria. The animals were acclimatized for seven days in a laboratory
environment of 12 h light/dark phase and had free access to food and
water before the commencement of the experiment at a temperature of
25 °C. This research study was approved by the Animal Ethical Com-
mittee of Afe Babalola University, Ado-Ekiti, Ekiti State, Nigeria
(ABUAD/PG/SCI/022).

2.5. Induction of diabetes mellitus

Streptozotocin (STZ) was liquefied in citrate buffer at a concentration of
15 mg/mL. The animals fasted overnight and the initial fasting blood
glucose (FBG) level for each of the animals was checked before inducing
diabetes by injecting 40 mg/kg of STZ intraperitoneally. Forty-eight
hours after STZ administration, FBG levels of all the experimental ani-
mals were determined using the ACCU check glucometer. As documented
previously by Ajiboye et al. (2018), animals with FBG levels greater than
or equal to 250 mg/dL were employed in this study.

2.6. Animal groupings

This was divided randomly into six groups of ten animals in each
group, which was housed in a pathogen-free environment.

2.7. Treatment of animal with plant extract and metformin

The extract of *C. orbicularis* leaf and metformin were orally adminis-
tered to the corresponding animals using a needle and intubator. This
administration route was preferred due to its safety, ease of ingestion as
well as preventing pain. Notably, the administration was carried out
between 9 and 10 am during the experimental period.

2.8. Tissues processing

The animals were fasted overnight (12 h) and euthanized with
halothane anesthesia on the 28th day of the experiment. The whole blood
of each rat was collected through cardiac puncturing, permitted to clot,
centrifuged at 3000 rpm for 900 s to achieve a clear solution called
serum, and employed for different analyses.

2.9. Biochemical analysis

FGB level was carried out according to the procedure of Ahmad et al.
(2007) via ACCU check glucometer. The obtained blood from the tail end
of each rat was dropped into the testing area of the glucometer. Then the
actual FGB level was revealed in the screen of the glucometer (mg/dL).
The FGB was recorded before STZ induction, at 48 h induction of STZ, on
the 14th day of the experiment, and finally on the 28th day of the
experiment before the sacrifice of the animals.

Liver glycogen was evaluated via a method of Murat and Serfaty
(1975). Superoxide dismutase (SOD), catalase (CAT), glutathione
peroxidase (GPx), glutathione reductase (GSH), glutathione transferase
(GST), GLUT 2, GLUT 4 and lipid peroxidation [measured as malon-
dialdehyde (MDA)] were determined via commercial kits. Also, hexoki-
nase and glucose-6-phosphatase activities were respectively carried out
via Akinosoye et al. (1987) and Baginsky et al. (1974).

2.10. Data analysis

Data were accepted as a mean ± SD of ten replicates. For the statisti-
cal analysis of all the results obtained, graph pad prism 5 (GPP) soft-
ware was used. A one-way ANOVA was used to conduct Tukey’s post hoc
examination. The meaningful difference was set at p < 0.055.

3. Results

3.1. Body weight of diabetic rats administered aqueous extract of
*C. orbicularis* leaf

The body weights of the rats were not substantially different from
each other at the beginning of the experiment (Figure 1). However, there
was a substantial decline in the body weight of diabetic control rats at
days 14 and 28 compared with other classes. Whereas there was a sub-
stantial increase in body weight of diabetic rats, administered at different
doses of *C. orbicularis* leaf.
3.2. Fasting blood glucose level of diabetic rats administered aqueous extract of C. orbicularis leaf

The result (Figure 2) indicates that before the injection of STZ, fasting blood glucose levels were not substantially different from each other. Important changes in fasting blood glucose levels occurred at 48 h of STZ induction in all except in the normal control group. Three doses of the aqueous extract of _C. orbicularis_ leaf showed (at 14th and 28th days) a substantial decrease with non-significant (p > 0.05) difference in the fasting blood glucose levels when compared with the control group.

3.3. Liver glycogen of diabetic rats administered aqueous extract of C. orbicularis leaf

In contrast to other groups, there was a substantial reduction in liver glycogen in diabetic control rats. However, a substantial increase in liver glycogen levels in diabetic rats administered three separate doses of aqueous extract of _C. orbicularis_ leaf was observed at the end of the experiment (Figure 3).

3.4. Oxidative stress biomarkers of diabetic rats administered aqueous extract of C. orbicularis leaf

STZ administration resulted in substantial depletion in superoxide dismutase, catalase, glutathione peroxidase, glutathione (reduced) and glutathione transferase concentrations, as well as augmented levels of MDA relative to other groups. However, the activities of superoxide dismutase, catalase, glutathione peroxidase, glutathione (reduced) and glutathione transferase increased significantly (p < 0.05) as well as a decrease in MDA levels in diabetic rats (Figure 4).

3.5. Hexokinase and glucose-6-phosphatase specific activities of diabetic rats administered aqueous extract of C. orbicularis leaf

The result revealed a substantial reduction in diabetic control of hexokinase-specific activity as well as a momentous upsurge in glucose-6-phosphatase-specific activity of diabetic control rats linked to other groups. On the other hand, a substantial increase in hexokinase activity was observed with a significant (p < 0.05) decrease in glucose-6-phosphatase activity in the liver of diabetic rats given three separate doses of aqueous extract of _C. orbicularis_ leaf (Figure 5).

3.6. Glucose transporters (GLUT 2 and GLUT 4) of diabetic rats administered aqueous extract of C. orbicularis leaf

Figure 6 displayed a substantial decline in diabetic control levels of GLUT 2 and GLUT 4 when compared with other groups. On the 28th day...
of this study, there was a substantial increase in the GLUT 2 and GLUT 4 levels of diabetic animals given three separate doses of aqueous extract of *C. orbicularis* leaf, with no distinct significance (*p* > 0.05) concerning the non-diabetic control group.

4. Discussion

Oxidative stress has been associated with the aetiology of numerous illnesses including diabetes mellitus. Oxidative stress causes membrane harm driving at long last to membrane rupture in various cell types (Kirubananthan et al., 2019). Cells have a proficient antioxidant defence agent protection framework against ruinous prompted by oxidative stress. A rise in blood glucose incites oxidative stress bringing about an expanded creation of oxygenated free radicals and diminished antioxidant enzyme activities (Liu et al., 2008). This may bring about intracellular structure adjustment and eventually in influence the normal cellular functions, prompting pathogenesis and the development of diabetic complexities. The current study portrays the ameliorative effect of the ethnobotanical doses of aqueous leaf extract of *C. orbicularis* in the

---

**Figure 4.** Oxidative stress biomarkers level in streptozotocin-induced diabetic rats after administration of aqueous extract of *C. orbicularis* leaf. Values are expressed as mean ± standard deviation (SD) of ten replicates. Bar with the same * are not significantly different at *p* > 0.05. Bar with different * are significantly different at *p* < 0.05. Normal control: non-diabetic control; met: metformin; diab: diabetic; AACO: *Chrysobalanus orbicularis* aqueous leaf extract; GPx: Glutathione peroxidase; GSH: Glutathione reductase; GST: Glutathione transferase SOD: superoxide dismutase; CAT: catalase; MDA: Malondialdehyde.

**Figure 5.** Hexokinase and Glucose-6-phosphatase specific activity in streptozotocin-induced diabetic rat after administration of aqueous extract of *C. orbicularis* leaf. Values are expressed as mean ± standard deviation (SD) of ten replicates. Bar with the same * are not significantly different at *p* > 0.05. Bar with different * are significantly different at *p* < 0.05. Normal control: non-diabetic control; DC: diabetic control; Diab: diabetic; met: metformin; AACO: *Chrysobalanus orbicularis* leaf.
therapeutic management of diabetes mellitus, a significant piece of information in drug advancement.

Currently, an aqueous extract of *C. orbicularis* leaf had the potential to increase the weight of induced diabetic rats. Khaleel et al. (2015) reported among numerous scientists that streptozotocin-induced diabetic rats are known by body weight loss, probably due to protein breakdown and the incompetence to provide gluconeoegenesis amino acid linked to insulin deficiency, resulting in muscle wasting and tissue breakdown in diabetic rats (Anand et al., 2008). Whereas, administration of various doses of aqueous extract of *C. orbicularis* leaf to the diabetic rat was able to restore weight loss probably due to insulin secretion enhancement.

Streptozotocin has been employed as an inducing agent to cause diabetes mellitus by causing selective damage to insulin generating pancreatic β-cells (Zhang et al., 2015). Pancreatic β-cell damage affects insulin secretion and eventually leads to a disorder known as hyperglycaemia (Sicree et al., 2006). The ability of the *C. orbicularis* leaf aqueous extract to reduce hyperglycaemia to normoglycaemia may be due to its high antioxidant content, which may regenerate the damaged beta cells (of the pancreas) that were impaired (Ekakitie et al., 2020). Administration of aqueous extract of *C. orbicularis* leaf to induced diabetic rats exhibited brilliant performance probably by inhibiting the absorption of glucose from the intestine, thereby helps to release glucose from the liver, or it could increase the number of insulin receptors, which in turn increases the sensitivity to insulin levels of the target tissues.

Liver glycogen is an essential biomarker for the assessment of the normoglycaemic effect of plant extracts or drugs (Huang et al., 2000). The rise in liver glycogen levels of the aqueous extracts of *C. orbicularis* leaf administered to diabetic rats can be due to its action in inhibiting glycogen phosphorylase. This is a central enzyme in glycogenolysis that, by inhibiting glycogen phosphorylase, causes glucagon and epinephrine to release glucose into the blood. This activity inhibits the entry of glucose into the blood, thereby reducing the amount of glucose in large quantities (Ajiboye et al., 2018). Besides, the aqueous extract of *C. orbicularis* leaf administered to diabetic rats trigger an enhancement in insulin secretion. Though the reduction in these glucose transporters could be associated with declining glucose uptake in diabetic mellitus states (Taderera et al., 2019). Thus this could be one of the mechanisms of antidiabetic potential of *C. orbicularis*.

5. Conclusions

The results of this research show that an aqueous extract of *C. orbicularis* leaf has antidiabetic activities, as it ameliorates the streptozotocin-induced type II rats model. Therefore, using ethnobotanical dosage, the plant extract may be useful for treating diabetic mellitus and its complications. To the best of our understanding, this is the first study reporting the ethnobotanical value of this plant in diabetes mellitus management.

Declarations

Author contribution statement

Basiru Ajiboye, Lisa Ilobekemen Ekakitie, Babatunji Emmanuel Oyinloye: Conceived and designed the experiments; Performed the...
experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

**Funding statement**

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

**Data availability statement**

Data will be made available on request.

**Declaration of interests statement**

The authors declare no conflict of interest.

**Additional information**

No additional information is available for this paper.

**References**

Ahmad, M.S., Pchetscheckriedeler, M., Ahmed, N., 2007. Aged garlic extract and Salicyl cysteine prevent formation of advanced glycation end products. Eur. J. Pharmacol. 561, 32–38.

Ajiboye, B.O., Ojo, O.A., Adeyoun, O., Imiere, O., Oyinloye, B.E., Ogumodode, O., 2018. Ameliorative activity of Ethanol extract of Artocarpus heterophyllus stem bark on Alloxan-induced diabetic rats. Adv. Pharmacuet. Bull. 8 (1), 141–147.

Ajiboye, B.O., Ojo, O.A., Oyinloye, B.E., Akuboh, O., Okosola, M.A., Idowu, O., 2020. In vitro antioxidant and inhibitory activities of polyphenolic-rich extracts of Syzygium cuminini (Linn) Skeels leaf on two important enzymes relevant to type II diabetes mellitus. Pak. J. Pharm. Sci. 33 (2), 523–529.

Akinyosoye, F., Fawole, M., Akinyahju, J., 1987. Studies on some enzymes of carbohydrate metabolism in Geotrichum candidum. Niger J Microbiol 7, 154–161.

Amira, R.E.B., Samy, A.H., Abeer, A.A., Yehia, A.H., Tarek, M.M., 2016. Saponins and their potential role in diabetes mellitus. Diabetes Manag. 7 (1), 148–158.

Anand, P., Murali, Y.K., Tandon, V., 2008. Insulinotropic effect of Brassica nigra improves glucose homeostasis in streptozotocin-induced diabetic rats. Exp. Clin. Endocrinol. Diabetes 117 (6), 251–256.

Baginski, S.E., Foli, P.P., Zak, B., 1974. Glucose-6-phosphatase. Methoden der enzymatischen Analyse 1, 835–842.

Ekakitie, L.I., Ojo, O.A., Oyunloye, B.E., Ajiboye, B.O., 2020. Antioxidant and inhibitory activities of enzymes linked type II diabetes mellitus: the novel role of C. orbicularis leaf extract. Iran. J. Toxicol. 3 (14).

Huang, X., Vaag, A., Hansson, M., 2000. Impaired insulin-stimulated expression of the glycogen synthase gene in skeletal muscle of type 2 diabetic patients is acquired rather than inherited. J. Clin. Endocrinol. Metabol. 85, 1584–1590, 2000.

Bhitoye, G.B., Ghali, U.M., Adekunle, J.B., Uwazie, J.N., Ajiboye, T.O., 2017. Antidiylpidemic, anti-inflammatory, and antioxidant activities of aqueous leaf extract of Dioscoreophllumcuministii (Stapf) Diels in high-fat diet-fed rats. Evid Based Complement Alternat Med 8128125, 2017.

Joudaki, R., Setorki, M., 2019. The protective effect of Saturejaehoriactierrrhizohydracolchic extract on streptozotocin-induced diabetes through modulating glucose transporter 2 and 4 expression and inhibiting oxidative stress. Pharmaceut. Biol. 57 (1), 318–327.

Khaleel, N., Saif, A., Anuha, S., Shaik, H.S., 2015. Effect of streptozotocin on glucose levels inAlbino wistar rats. J. Pharmaceut. Sci. Res. 7 (2), 67–69.

Kiranbhananathan, G., Vijayan, S.G., Thangaraj, A., Sundaram, R., 2019. Antioxidant potential of theoflavin ameliorates the activities of keyenzymes of glucose metabolism in high fat diet and streptozotocin – induced diabetic rats. Redox Rep. 24 (1), 41–50.

Koot, W., Farshchipour, M., Asadazadeh, Z., Ashthary-Larky, D., Asadi-Samani, M., 2016. The role of medicinal plants in the treatment of diseases: a systematic review of Calendula officinalis. Electron. Physician 8 (14), 92–95.

Liu, W., Hei, Z., Nie, H., 2008. Berberine ameliorates renal injury in streptozotocin-induced diabetic rats by suppression of both oxidative stress and aldose reductase. Chin. Med. J. 121, 706–712.

Mahmoud, M.R., El-Abbar, H.S., Saleh, S., 2002. The effect of Nigella sativa oil against the liver damage induced by Schistosomamansoni infection in mice. J. Ethnopharmacol. 79, 1–11.

Murat, J.C., Serfaty, A., 1975. Simple enzymatic determination of polysaccharid(glycogen) content of animal tissues. Clin. Chem. 20, 1576–1577.

Murray, R.K., Grammer, D.K., Mayesa, P.A., 2000. Harper’s Biochemistry, 25th ed. Appleton and Lange, Stamford, CT.

Nwozo, S.O., Oyinloye, B.E., 2011. Hepatoprotective effect of aqueous extract of Aframomummelegueta on ethanol-induced toxicity in rats. Acta Biochim. Pol. 58 (3), 355–358.

Paracanmo, N.E.N.P., 2011. Comumeroporrotetivar. anguifoliusadflad. (Connaraceae): Tradicionalmenteutilizadacomobarbatimang No Pará. Embapa Amazonia Oriental. InfotecadeEmbrapa: Belém, PA.

Piero, M.N., Nzzo, G.M., Njagi, J.M., 2015. Diabetes mellitus – a devastating metabol is dialogue. Asian J. Biomed. Pharmaceut. Sci 4 (40), 1–7.

Pires, F.B., Dolivitch, C.B., Prá, V.D., Móngelo, D.L., Schneider, V.M., Loose, R.F., Schmidt, M.E.P., Bressan, L.P., Mazzuti, M.A., Rosa, M.B., 2016. AnOverview about the chemical composition and Biological Activity of Medicinal species found in the Brazilian Amazon. J. App PharmSci 6 (12), 233–238.

Sircre, R., Shaw, J., Zimm, P., 2006. The global burden. Diabetes and impaired glucose tolerance. Prevalence and projections. In: Gan, D. (Ed.), Diabetes Atlas, third ed. International Diabetes Federation, Brussels, pp. 18–103.

Sorour, H.A., Selim, M.M., El-Moselhy, L.E., Ahmed, S.G., 2018. Ameliorative effect of watermelon rind ingestion on the pancreas of diabetic female albino rat (histological, immunohistochemical and morphometric study). Egypt. J. Histol. 42 (1), 10–22.

Taderera, T., Chagonda, L.S., Gomo, E., Katereke, D., Shai, L.L., 2019. Ananasstrepynphy fungi aqueous extract stimulate glucose uptake in established C2C12 muscle cell lines. Afr. Health Sc i. 19 (2), 2219–2229.

Zhang, J.G., Liu, Q., Liu, Z.L., Li, Y., Li, T.T., 2015. Antihyperglycemic activity of Anoectochilusrhorborgi polyacchariose in diabetic mice induced by high-fat diet and streptozotocin. J. Ethnopharmacol. 164, 180–185.