Sodium glucose cotransporter 2 (SGLT2) inhibitors are a new class of antidiabetic drugs that improve glycemic control by inhibiting reabsorption of glucose filtered through the renal glomerulus. Use of drugs in this class has increased because of their effect of decreasing body weight and a low risk for hypoglycemia, in addition to a relatively strong glucose-lowering effect. SGLT2 inhibitors such as canagliflozin and sotagliflozin (a SGLT1/SGLT2 dual inhibitor) also have a mild or moderate intestinal and renal SGLT1 inhibitory effect because of their relatively weak selectivity for SGLT2 over SGLT1. Recent evidence shows that these SGLT2 inhibitors with low SGLT2/SGLT1 selectivity elevate the level of circulating glucagon like peptide-1 (GLP-1), an incretin hormone that promotes insulin secretion in pancreatic β cells. This effect probably occurs partly via inhibition of intestinal SGLT1, and the elevation of active GLP-1 levels is especially apparent when these drugs are co-administered with dipeptidyl peptidase 4 (DPP4) inhibitors. These findings suggest that a combination of canagliflozin or sotagliflozin and a DPP4 inhibitor can provide a beneficial effect associated with elevation of circulating active GLP-1 and may serve as a treatment for patients with type 2 diabetes.

Keywords: Canagliflozin; Sotagliflozin; SGLT1; Type 2 diabetes

Sodium glucose cotransporter 2 (SGLT2) inhibitors are a new class of antidiabetic drugs that improve glycemic control by inhibiting reabsorption of glucose filtered through the renal glomerulus [1, 2]. Use of drugs in this class has increased worldwide because of the definitive effect of a decrease of body weight and a low risk for hypoglycemia, in addition to a relatively strong glucose-lowering effect independent of insulin action [1, 2]. Notably, a recent prospective study (the EMPG-REG OUTCOME study) showed that empagliflozin (a SGLT2 inhibitor) reduced cardiovascular outcomes and death from any cause in patients with type 2 diabetes at high risk for cardiovascular events [3]. In contrast, a beneficial effect on cardiovascular events could not be proved using dipeptidyl peptidase 4 (DPP4) inhibitors [4-6], another widely used class of antidiabetic agents. DPP4 inhibitors mainly improve glycemic control by increasing circulating active glucagon-like peptide 1 (GLP-1), an incretin hormone that promotes insulin secretion in pancreatic β cells, by blocking degradation of GLP-1 by DPP4 [7].

SGLT2 is expressed in the S1 segment of the proximal tubules in the kidney, and inhibition of this molecule results in a marked increase in urinary glucose excretion (UGE) [8-10]. SGLT2 is a high-capacity and low-affinity glucose transporter that is responsible for approximately 90% of glucose absorption in kidney [9]. Interestingly, glucose reabsorption in renal tubules mediated by SGLT2 is promoted in patients with type 2 diabetes, compared with non-diabetic subjects [11, 12]. SGLT1 is expressed in the S3 segment of the proximal renal tubules, which is located more distally from the glomerulus compared with the location of S1. SGLT1 is a low-capacity and high-affinity transporter that accounts for absorption of the remaining 10% of the glucose [8]. The effect of SGLT1 on glucose absorption can be enhanced in the presence of an SGLT2 inhibitor due to a compensatory mechanism [8, 13-15]. Therefore, SGLT1 transport of glucose may weaken the glucose-lowering effect of SGLT2 inhibitors [8, 13-15].

SGLT1 is also abundantly expressed in the brush-border membrane surface of villi lining the lumen of the upper small intestine, where it contributes to absorption of glucose or galactose from the gastrointestinal tract [16, 17]. Glucose absorption in the small intestine via SGLT1 in patients with type 2 diabetes generally increases relative to that in non-diabetic subjects [18]. A genetic deficiency of SGLT1 can cause glucose-galactose malabsorption in newborn infants that results in life-threatening dehydration due to severe diarrhea if they do not receive sugar free diet [19]. However, it is likely that mild to moderate pharmacological inhibition of SGLT1 in the small intestine reduces postprandial excursion of glucose without causing severe diarrhea or malabsorption [20]. In this respect, an SGLT2 inhibitor with a mild or moderate inhibitory effect on SGLT1 may improve glycemic control more effectively than highly selective SGLT2 inhibitors in patients with type 2 diabetes by increased...
Orally administered canagliflozin is rapidly absorbed in the gastrointestinal tract in a dose-dependent manner at 50 - 300 mg/day and its oral bioavailability is approximately 65%, with a maximum effect 30 - 120 min after administration [33, 36]. Accumulation of canagliflozin in plasma increases to 36% following multiple doses ranging from 100 to 300 mg/day [33, 36]. The half-life (t1/2) is 11 - 13 h and this relatively long t1/2 permits a once-a-day regimen [37]. Plasma protein binding is very high (99%) and is independent of the plasma concentration of canagliflozin and of liver and renal damage [33, 36]. Canagliflozin is metabolized into two inactive O-glucuronide metabolites, named M5 and M7. In healthy subjects, canagliflozin is excreted in feces as 41.5% unchanged, 7% hydroxylated, and 3.2% O-glucurononidated forms, and in urine as < 1% unchanged form, 7-10% M5 and 21-32% M7 [33, 36].

The half-maximal inhibitory concentrations (IC50) of canagliflozin in humans are 663 ± 180 nM for SGLT1 and 4.2 ± 1.5 nM for SGLT2, giving an SGLT2 selectivity of 150- to 160-fold [21, 38]. The threshold for glucose excretion (RTG) in kidney is reduced from 240 to 70 - 90 mg/dL at a dose of 300 mg of canagliflozin, and this results in UGE of approximate 77 - 119 g/day, which is comparable to a calorie loss of 308 - 476 kcal/day [33]. This increase of 24-h UGE by canagliflozin is about 25% higher than that with dapagliflozin at its clinically approved maximal dose of 10 mg [39]. The maximum plasma concentration of canagliflozin is not altered by renal dysfunction [33, 36]. Canagliflozin (300 mg) given before a meal inhibits postprandial glucose excursion in patients with type 2 diabetes [26]. In contrast, dapagliflozin in healthy subjects [39] and empagliflozin in patients with type 2 diabetes [32] have no effect on postprandial excursion of glucose. This postprandial effect of canagliflozin seems to be explained by its relatively strong inhibition of SGLT1, compared with dapagliflozin and empagliflozin. Importantly, blood levels of canagliflozin are not sufficient to inhibit systemic SGLT1, but the maximum level in the lumen in the gastrointestinal tract is high enough to inhibit SGLT1 in the brush-border membrane lining of the lumen of the small intestine [25, 38].

**Clinical efficacy of canagliflozin for glycemic control**

Changes of HbA1c from baseline after canagliflozin monotherapy for 26 weeks were -0.77% at 100 mg/day (baseline HbA1c: 8.1%) and -1.03% at 300 mg/day (baseline HbA1c: 8.0%) [40], each with a significant difference vs. placebo (+0.14%; baseline HbA1c: 8.0%). In a comparison of the effects of canagliflozin and glimepiride (a sulfonylurea) for 52 weeks in patients with type 2 diabetes treated with metformin, the changes in HbA1c were -0.82% with canagliflozin 100 mg/day, -0.93% with canagliflozin 300 mg/day, and -0.81% with titrated glimepiride, with significant superiority of canagliflozin 300 mg/day vs. glimepiride [41]. In a similar comparison of canagliflozin and sitagliptin (a DPP4 inhibitor) for 52 weeks in patients with type 2 diabetes treated with metformin, the changes in HbA1c were -0.73% with canagliflozin 100 mg/day, -0.88% with canagliflozin 300 mg/day, and -0.73% with sitagliptin 100 mg/day. Again, there was significant superiority of canagliflozin 300 mg/day vs. sitagliptin 100 mg/day [42]. A recent systematic review and network meta-analysis showed

Elevation of Circulating GLP-1 by Canagliflozin and Sotagliflozin

**Canagliflozin**

Canagliflozin was developed in Japan and was approved by the US Food and Drug Administration (FDA) in March 2013, as the first clinically available SGLT2 inhibitor in the USA. Canagliflozin was approved in Europe in November 2013 and in Japan in October 2014. In the USA and Europe, the recommended starting dose is 100 mg once a day before the first meal. This can be increased to 300 mg a day depending on the degree of glycemic control if the patient tolerates 100 mg and has an estimated glomerular filtration rate (eGFR) ≥ 60 ml/min/1.73 m² [33-36]. However, only use of a dose of 100 mg once a day is currently approved in Japan.

**Pharmacokinetics and pharmacodynamics**

Orally administered canagliflozin is rapidly absorbed in the...
changes of HbA1c with canagliflozin of -0.86% at 300 mg/day and -0.76% at 100 mg/day, while those with the clinically approved maximum doses of dapagliflozin (10 mg/day) and empagliflozin (25 mg/day) were both -0.66%, suggesting a relatively strong glucose-lowering effect of canagliflozin [43].

**Effect of canagliflozin on circulating GLP-1: animal studies**

Oguma et al investigated the effects of canagliflozin, teneligliptin (a DPP4 inhibitor) and their combination therapy on plasma glucose, insulin, and active GLP-1 (aGLP-1) levels in Zucker diabetic fatty (ZDF) rats, a genetic animal model of obese type 2 diabetes [24]. After oral administration of glucose solution, plasma glucose significantly increased in vehicle-treated ZDF rats, and canagliflozin and teneligliptin monotherapy both significantly suppressed plasma glucose compared with that in vehicle-treated ZDF rats. As expected, combination therapy of these drugs further decreased plasma glucose. Plasma insulin levels after glucose administration were increased by teneligliptin, but not influenced by canagliflozin. A mild, transient and non-significant increase of plasma aGLP-1 occurred with canagliflozin (10 mg/kg) after glucose administration. Canagliflozin also increased the area under the curve (AUC0 - 2h) of aGLP-1 by about three- and fourfold at doses of 3 and 10 mg/kg, respectively, compared with vehicle, but these increases were not significant. Teneligliptin (0.3 mg/kg) resulted in an approximately eightfold greater AUC0 - 2h of aGLP compared with vehicle, and again the change was not significant. However, a significant increase in AUC of about 47-fold (compared to vehicle) occurred with co-administration of canagliflozin (10 mg/kg) and teneligliptin (0.3 mg/kg). In this study, teneligliptin inhibited plasma DPP4 activity by about 75% and this was not influenced by addition of canagliflozin [24].

The same authors investigated the effect of intestinal SGLT1 inhibition on plasma aGLP-1 levels in normal and diabetic rodents [25]. In male C57BL6J mice (non-diabetic control mice), canagliflozin (10 mg/kg) significantly increased plasma aGLP-1 at 15 and 30 min in an oral glucose tolerance test (OGTT) compared with vehicle, and a marked elevation was noted in combination with sitagliptin (10 mg/kg) and teneligliptin (0.3 mg/kg). In SD rats (an animal model of type 2 diabetes), canagliflozin in combination with sitagliptin (10 mg/kg) dose-dependently increased AUC0 - 2h of aGLP-1 during the OGTT. Canagliflozin + sitagliptin (10 mg/kg) also significantly increased the AUC0 - 2h of aGLP-1, but the effect was weaker than that of sitagliptin + 3-(4-cyclopropylphenylmethyl)-1-(beta-D-glucopyranosyl)-4-methylinole (CGMI), which is a more potent inhibitor of SGLT1 than canagliflozin; the IC50 values of CGMI for human SGLT1 and SGLT2 are 22.1 and 1.39 nM, respectively, with a 15.9-fold selectivity for SGLT2 over SGLT1. Taken together, these findings suggest that both the dose and SGLT1 selectivity are important for increased GLP-1 secretion and that the effect is enhanced by a combination with a DPP4 inhibitor. This study also showed that the potentially inhibitory effect of canagliflozin on carbohydrate absorption in the gastrointestinal tract is transient after sucrose loading. Canagliflozin increased carbohydrate content in the upper and middle intestine (but not in the lower intestine) at 1 h, but the effect vanished at 6 h [25].

**Effect of canagliflozin on circulating GLP-1: clinical studies**

Polidori et al investigated the effects of canagliflozin on intestinal glucose absorption in 20 healthy subjects using a dual tracer method in a study with a two-period, crossover design [26]. Canagliflozin decreased postprandial glucose and insulin, increased 0- to 6-h UGE, and delayed the rate of appearance of oral glucose (RaO) in plasma, as evaluated using oral 14C-glucose. Canagliflozin significantly decreased AUC0 - 1h of RaO by 31% and AUC0 - 2h by 20%. In contrast, canagliflozin treatment over 2 - 6 h increased RaO, such that AUC0 - 6h of RaO was only 6% lower compared with placebo. It was concluded that canagliflozin can reduce postprandial plasma glucose and insulin levels by increasing UGE mediated by SGLT2 inhibition in kidney and by delaying glucose absorption in the intestine, probably due to inhibition of SGLT1. Consistent with delayed glucose absorption, there were changes in the levels of GLP-1 and other gut peptides, including glucose-dependent insulino-mimetic peptide (GIP), another incretin hormone that promotes insulin secretion in pancreatic β cells, and peptide YY (PYY), an anorectic hormone secreted from L cells that reduces food intake directly or via a vagal afferent pathway [44, 45]. AUC0 - 2h for total GLP-1 (tGLP-1) was significantly increased by 35% with canagliflozin compared with placebo. The increase of tGLP-1 was large from 30 min to 2 h, but not from 0 to 30 min. A similar tendency was found for AUC0 - 2h of aGLP-1, but the increase was not significant. Canagliflozin decreased incremental postprandial GIP by 50% and increased incremental postprandial PYY by 60% compared with placebo.

We recently investigated the effect of canagliflozin on GLP-1 levels in patients with type 2 diabetes [46]. Canagliflozin (100 mg a day) (n = 15) for 3 days significantly increased AUC0 - 2h of plasma aGLP1 from baseline by about two times, while no significant change was noted for 3 days observation in a control group without canagliflozin (n = 15). Notably, the increase in plasma aGLP-1 with canagliflozin reached a maximum at 30 min and was maintained from 0 to 2 h. Addition of teneligliptin resulted in a further increase of plasma aGLP-1, with an approximately fourfold increase of AUC0 - 2h of aGLP-1 during the OGTT. Canagliflozin + sitagliptin (10 mg/kg) also significantly increased the AUC0 - 2h of aGLP-1, but the effect was weaker than that of sitagliptin + 3-(4-cyclopropylphenylmethyl)-1-(beta-D-glucopyranosyl)-4-methylinole (CGMI), which is a more potent inhibitor of SGLT1 than canagliflozin; the IC50 values of CGMI for human SGLT1 and SGLT2 are 22.1 and 1.39 nM, respectively, with a 15.9-fold selectivity for SGLT2 over SGLT1. Taken together, these findings suggest that both the dose and SGLT1 selectivity are important for increased GLP-1 secretion and that the effect is enhanced by a combination with a DPP4 inhibitor. This study also showed that the potentially inhibitory effect of canagliflozin on carbohydrate absorption in the gastrointestinal tract is transient after sucrose loading. Canagliflozin increased carbohydrate content in the upper and middle intestine (but not in the lower intestine) at 1 h, but the effect vanished at 6 h [25].

**Sotagliflozin**

Sotagliflozin is a dual SGLT1/SGLT2 inhibitor that is currently under development in a phase 3 study for type 1 diabetes. A similar phase 3 study for type 2 diabetes is expected to start by the end of 2016.

**Pharmacokinetics and pharmacodynamics**

In healthy subjects, the maximum circulating concentration of sotagliflozin was 165 ng/mL at day 7 after multiple-dose administration, and t1/2 was 29 h [23]. Approximately 70% of sotagliflozin is rapidly absorbed and reaches the circulation within 15 min. In patients with type 2 diabetes, a maximum
plasma concentration of 230 - 307 ng/mL was reached at 30 min to 2 h after administration of 300 mg of sitagliptin, with a t½ of 13.5 ± 5.3 h and a steady-state plasma concentration achieved between days 7 and 14 [31]. Sitagliptin is mostly eliminated in a glucoside form in the urine, and is partially eliminated unchanged in the feces in healthy subjects [20]. Similarly to canagliflozin, the blood level of sitagliptin is insufficient to inhibit systemic SGLT1, while the level in the lumen in the gastrointestinal tract is high enough to inhibit SGLT1 [31]. The IC50 values of sitagliptin are 36 nM for human SGLT1 and 1.8 nM for human SGLT2, with selectivity of 20-fold for SGLT2 over SGLT1 [31]. Sitagliptin has approximately similar potency for inhibition of SGLT2 compared with other SGLT2 inhibitors, such as dapagliflozin, empagliflozin and canagliflozin, but is about 10-fold more potent than canagliflozin for inhibition of SGLT1 [21, 31]. The maximum UGE was 44 g at 24 h after a single dose of sitagliptin at 300 mg, and the maximum UGE on day 7 after multiple dosing was 36 g [23, 30], reflecting the SGLT2 inhibitory effect of sitagliptin in the kidney. This UGE is somewhat lower than those reported for other SGLT2 inhibitors: dapagliflozin (62 g) [47], canagliflozin (70 g) [48] and empagliflozin (74 g) [49]. The relative weak effect of sitagliptin for UGE may be due to the improved glycemic control through mechanisms based on SGLT1 inhibition.

Clinical efficacy of sitagliptin for glycemic control

In a phase 2a clinical trial in type 2 diabetes not adequately controlled with metformin (n = 36), patients were divided into three groups: sitagliptin 150 mg a day (n = 12), 300 mg a day (n = 12), and placebo (n = 12) [31]. Sitagliptin treatment for 28 days decreased HbA1c by -1.15% at 150 mg and -1.25% at 300 mg from baseline (8.2% and 8.5%, respectively) vs. -0.49% with placebo (8.2%). In a phase 2b dose-ranging study of sitagliptin in patients with type 2 diabetes poorly controlled with metformin (n = 299), groups with HbA1c at baseline of 8.0%, 8.3%, 8.4%, 8.1% and 7.9% were treated with 75 mg once daily (n = 59), 200 mg once daily (n = 60), 200 mg twice daily (n = 60), 400 mg once daily (n = 60), and placebo (n = 60), respectively [50]. Sitagliptin significantly and dose-dependently decreased HbA1c from baseline by 0.42%, 0.52%, 0.80%, and 0.92% in 12 weeks treatment at these respective doses, while reduction of HbA1c with placebo was only 0.09%. The reduction of HbA1c at 400 mg once daily was similar to that achieved with the maximum dose of canagliflozin (300 mg once daily).

Effect of sitagliptin on circulating GLP-1: animal studies

Sitagliptin significantly increased circulating aGLP-1, tGLP-1 and PYY in mice from 30 min to 6 h after meal challenge [27]. The increases in GLP-1 and PYY were significantly associated with increases in total glucose in the small intestine, cecum and colon, compared with a vehicle group. The increases in glucose were maintained over 6 h, suggesting that inhibition of SGLT1 in the gastrointestinal tract by sitagliptin is sustained for a relatively long time. In addition, the increase of glucose in the gastrointestinal tract was associated with a decrease in pH in the cecum. Significant increases in circulating levels of aGLP-1, tGLP-1 and PYY, and of glucose in the small intestine, cecum and colon after a glucose-containing meal challenge also occurred in SGLT1-/- mice that were littermates of the SGLT1+/+ mice. The peak aGLP-1 level in SGLT1-/- mice occurred at 1 h after the meal test, corresponding to the peak in the glucose level in the small intestine [27]. Similarly, in obese male C57BL6J mice fed a high fat diet, single dose sitagliptin or sitaglitn was associated with a significant increase in AUC0-∞ h for aGLP-1 after meal challenge, and combination therapy with these drugs markedly elevated the AUC for aGLP-1. After 14 days of once-a-day administration, sitagliptin and sitaglitn monotherapy also increased plasma aGLP-1 between 0 and 6 h after meal challenge compared with the vehicle, and combination therapy further increased aGLP-1 [28]. In KKAY mice (a model of type 2 diabetes), a significant increase of tGLP-1 at 2 h after glucose challenge was found with sitagliptin compared with vehicle [29]. Interestingly, in this study, long-term sitagliptin treatment caused an increase in food consumption in KKAY mice, as well as in lean rats and dogs, but not in OP-CD rats (an obese rat model) [29].

Effect of sitagliptin on circulating GLP-1: clinical studies

In healthy subjects (n = 12; treatment 10, placebo 2), sitagliptin significantly increased aGLP-1, tGLP-1 and PYY and significantly reduced postprandial glucose relative to placebo after breakfast [30]. In a single-dose study of sitagliptin at 300 mg in 12 patients with type 2 diabetes, there was a significant increase in UGE throughout the day after dosing, significant increases in aGLP-1, tGLP-1, and PYY, and significant decreases in plasma glucose and insulin between 0 and 13 h [31]. In a single center, three-treatment, three-crossover, randomized, open-label study in patients with type 2 diabetes (n = 18), sitagliptin monotherapy and combination therapy with sitaglitn increased 24-h UGE to similar degrees, suggesting that sitaglitn did not affect UGE. Sitagliptin given once a day before a morning meal did not significantly increase aGLP-1 from baseline. Combination therapy of sitagliptin and sitaglitn once a day before a morning meal increased aGLP-1, tGLP-1 and PYY, and decreased total GIP, compared with sitaglitn monotherapy. The concentrations of aGLP-1 at 2 h after a morning meal and 2 h after lunch were approximate 10 and 15-20 pM, respectively, with combination therapy, compared to approximate 5 and 7 pM, respectively, at baseline (control) [28].

Mechanisms of Elevation of Circulating GLP-1 by SGLT2 Inhibitors

The amount of secreted GLP-1 is associated with the size of a meal and the rate of gastric emptying after the meal [51, 52]. Fasting plasma GLP-1 is usually maintained at a very low lev-
SGLT1 in healthy subjects [30] and in patients with type 1 in SGLT1-/- mice [27]. Similar findings were obtained for basically consistent with the postprandial elevation of GLP-1 with type 2 diabetes, as described above [46]. This finding is and significantly promoted this effect in our study in patients phase GLP-1 elevation after a meal in healthy subjects [26], inhibit SGLT1 in the upper intestine, which suggests that these fatty acids from meals can reach the lower intestine, and the increased glucose levels in the intestine lumen due to inhibition of SGLT1 in the upper intestine, resulting in acute promotion of GLP-1 secretion by probable L cells in the lower intestine via an effect mediated by the afferent vagal nerve; however, there is no evidence to support this hypothesis in animal models or clinical studies. Other mechanisms occurring via the afferent vagal nerve and associated with SGLT1 inhibition in the upper intestine, but independent of SGLT3 stimulation, may also be present. A mechanism independent of SGLT1 inhibition may also occur, because empagliflozin, a highly selective SGLT2 inhibitor with selectivity of 2,680-fold over SGLT1 [21], also promotes GLP-1 secretion after a meal in patients with type 2 diabetes [32]. A single dose of empagliflozin (25 mg) following chronic administration for 4 weeks increased plasma aGLP-1 after mix meal dosing, although the elevation appeared to be relatively mild, especially when empagliflozin was given chronically. Interestingly, empagliflozin increased plasma GLP-1 levels, as well as endogenous glucose production (EGP) and glucagon, even before a meal. These changes may be associated with increased UGE by SGLT2 inhibition, but the mechanisms are unknown. Taken together, these findings suggest that SGLT2 inhibition itself might contribute to plasma GLP-1 elevation, probably via indirect mechanisms, although the effect via this mechanism appears to be mild.

The effects of canagliflozin and sitagliptin on the second peak (late phase) of GLP-1 elevation after a meal may occur through different mechanisms for the two drugs. Due to SGLT1 inhibition by these drugs, most glucose derived from food reaches the lower intestine without absorption in the upper intestine [75]. SGLT1 inhibition of canagliflozin in the intestinal lumen is transient and vanishes in a few hours [25]. Therefore, the effect of canagliflozin has almost disappeared when glucose reaches the lower intestine, and the increased glucose level can directly stimulate GLP-1 secretion via SGLT1 in L cells in the lower small intestine. In contrast, sitagliptin is a long-acting SGLT1 inhibitor in the intestinal lumen, and the effect may be maintained for 1 day [27, 28]. Sitagliptin is partially eliminated unchanged in the feces [20], and thus it is unlikely that sitagliptin promotes GLP-1 secretion by glucose stimulation of SGLT1 in L cells. The most plausible explanation for late phase GLP-1 elevation after a meal by sitagliptin may be based on mechanisms mediated by short chain fatty acids (SCFAs) such as acetate, butyrate, and propionate [27, 74]. SCFAs produced by bacterial fermentation of glucose which reached the colon due to SGLT1 inhibition by sitagliptin in the upper small intestine probably can directly simulate GLP-1 secretion via GPR41 (FFAR3) and GPR43 (FFAR2) in L cells in the colon [28, 75]. The effect of GLP-1 elevation in the late phase by canagliflozin also may be partially based on this mechanism, but it is unclear which of the effect via SGLT1 by glucose stimulation or that via GPR41/43 by SCFA stimulation is predominant for this agent. Furthermore, GLP-1 elevation in the late phase, as well as in the early phase, may occur independently of increased glucose in the lower intestine or colon, probably via mechanisms indirectly associated with increased UGE by SGLT2 inhibition, because empagliflozin mildly increases plasma GLP-1 in both phases [32].
Taken together, the detailed mechanisms of plasma GLP-1 elevation in the early phase by canagliflozin or sotagliflozin remain largely unclear. However, a mechanism via afferent vagal nerve activation mediated by increased SGLT3 stimulation due to an increase of glucose in the upper intestine based on SGLT1 inhibition or other mechanisms, independently of SGLT3, might be involved. In addition, mechanisms independent of SGLT1 inhibition and indirect via SGLT2 inhibition may be involved. Late phase elevation of plasma GLP-1 may involve glucose stimulation of SGLT1 (canagliflozin) and SCFA stimulation of GPR41/43 (sotagliflozin and possibly canagliflozin) in L cells. Mechanisms independent of SGLT1 inhibition and associated with SGLT2 inhibition may also be involved.

Conclusion and Future Perspectives

The UKPDS study showed that earlier strict intervention for glycemic control can prevent progression of microvascular complications in patients with type 2 diabetes [76]. Additionally, longer observation over 10 years after the intervention in the UKPDS study showed that this earlier intervention for glycemic control after onset of type 2 diabetes suppressed cardiovascular events [77]. This suggests the importance of strict glycemic control in the early stage for inhibition of progression of both micro- and macrovascular complications.

On the other hand, a hypoglycemia attack caused by treatment for diabetes [78] or obesity [79] can be a risk factor for cardiovascular events. In this respect, SGLT2 inhibitors may have an advantage for treatment of type 2 diabetes from an earlier stage because of their effect on decreasing body weight, a low risk for hypoglycemia, and a glucose-lowering effect independent of insulin action [1, 2]. However, combination therapy may also be important to achieve stricter glycemic control if the effect of monotherapy is insufficient. Thus, addition of a DPP4 inhibitor to an SGLT2 inhibitor may be reasonable because DPP4 inhibitors generally do not increase body weight or hypoglycemia when used without sulfonylureas or insulin [80].

Combination therapy with SGLT2 inhibitors with low SGLT2/SGLT1 selectivity such as canagliflozin or sotagliflozin and a DPP4 inhibitor markedly enhances elevation of circulating active GLP-1 compared with DPP4 inhibitor monotherapy [24, 25, 28]. Notably, recent clinical trials have shown that DPP4 inhibitors do not reduce cardiovascular events and mortality [4, 5, 6], whereas these events were significantly suppressed by liraglutide, a GLP-1 receptor agonist [81]. It is unclear whether a difference in the degree of elevation of plasma active GLP-1 or liraglutide between treatments by DPP4 inhibitors and by liraglutide can explain these results. However, this finding also suggests that a combination of a canagliflozin or sotagliflozin and a DPP4 inhibitor might provide beneficial effects associated with enhanced elevation of circulating active GLP-1 levels.

Several issues remain to be clarified. First, it is unclear if significant elevation of circulating GLP-1 is specific for SGLT2 inhibitors with low SGLT2/SGLT1 selectivity such as canagliflozin and sotagliflozin, which has a potent SGLT1 inhibitory effect. The clinical significance of involvement of intestinal SGLT1 inhibition on elevation of circulating GLP-1 by SGLT2 inhibitors remains unclear and the mechanisms are probably complex. Second, it will be interesting to investigate whether there is a difference in elevation of circulating GLP-1 between SGLT2 inhibitors with a long acting and relatively strong intestinal SGLT1 inhibitory effect, such as sotagliflozin, and those with a short acting and mild intestinal SGLT1 inhibitory effect, such as canagliflozin. Finally, there is a need to clarify whether the extent of elevation of active GLP-1 by combination therapy with a DPP4 inhibitor and a canagliflozin or sotagliflozin is sufficient to provide a clinically significant effect on glycemic control and a potential protective effect of GLP-1 on cardiovascular events.

In conclusion, SGLT2 inhibitors with low SGLT2/SGLT1 selectivity such as canagliflozin and sotagliflozin can significantly elevate circulating active GLP-1 levels, especially when used in combination with DPP4 inhibitors. This combination therapy may serve as a new strategy for treatment of patients with type 2 diabetes in this aspect.

Conflicts of Interest

Authors have no interest to disclose.

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KT wrote this manuscript. TI reviewed the manuscript.

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