Video Validation of Tri-Axial Accelerometer for Monitoring Zoo-Housed *Tamandua tetradactyla* Activity Patterns in Response to Changes in Husbandry Conditions

Sofía Pavese 1, Carlos Centeno 2, Lorenzo Von Fersen 3, Gabina V. Eguizábal 1,4, Luis Donet 2, Camila J. Asencio 1,4, Daniel P. Villarreal 5 and Juan Manuel Busso 1,4,*

Simple Summary: In recent years, the use of technologies to remotely monitor animal behaviour has increased. However, when they are used for the first time in a new species, validation is needed. Therefore, a tri-axial accelerometer was used to monitor lesser anteater (*Tamandua tetradactyla*) behaviour. First, the influence of using a vest on the animals’ behaviour was evaluated. Second, correlations between video recordings and accelerometer data were made, allowing the researchers to obtain summary measures for each behaviour and a threshold to discriminate activity/inactivity events. Sensitivity and precision variables revealed the robustness of accelerometer monitoring. Third, animals responded to a reduction in enclosure complexity by increasing their activity. The relevance of the accelerometer was determined comparing data (activity level and activity cycle) with video recordings. The results indicate that an accelerometer attached to a vest is a reliable technology for monitoring the activity pattern of *T. tetradactyla* under semi-controlled environmental conditions.

Abstract: Accelerometers are a technology that is increasingly used in the evaluation of animal behaviour. A tri-axial accelerometer attached to a vest was used on *Tamandua tetradactyla* individuals (*n* = 10) at Biodiversity Park. First, the influence of using a vest on the animals’ behaviour was evaluated (ABA-type: A1 and A2, without a vest; B, with a vest; each stage lasted 24 h), and no changes were detected. Second, their behaviour was monitored using videos and the accelerometer simultaneously (experimental room, 20 min). The observed behaviours were correlated with the accelerometer data, and summary measures (X, Y and Z axes) were obtained. Additionally, the overall dynamic body acceleration was calculated, determining a threshold to discriminate activity/inactivity events (variance = 0.0055). Then, based on a 24 h complementary test (video sampling every 5 min), the sensitivity (85.91%) and precision (100%) of the accelerometer were assessed. Animals were exposed to an ABA-type experimental design: A1 and A2: complex enclosure; B, decreased complexity (each stage lasted 24 h). An increase in total activity (%) was revealed using the accelerometer (26.15 ± 1.50, 29.29 ± 2.25, and 35.36 ± 3.15, respectively). Similar activity levels were detected using video analysis. The results demonstrate that the use of the accelerometer is reliable to determine the activity. Considering that the zoo-housed lesser anteaters exhibit a cathemeral activity pattern, this study contributes to easily monitoring their activities and responses to different management procedures supporting welfare programs, as well as ex situ conservation.
Keywords: acceleration logging; activity; animal behaviour; ethogram; remote observation; telemetry; Pilosa; Xenarthra

1. Introduction

In recent years, the application of technologies to remotely monitor animal activity (e.g., accelerometers) have made it possible to increase sampling capacity, with minor sampling effort and observer bias [1–5]. Tri-axial accelerometers are sensors that simultaneously measure the acceleration of an object/organism in space along three dimensions, X, Y and Z [6]. These devices offer information that can be processed to identify animals’ activity states [7]. In brief, information is reflected by two main components: static acceleration, related to the force of gravity and the orientation of the accelerometer in space, and dynamic acceleration, associated with the movement of an animal [8]. Normally, accelerometers are mounted to the animal’s body with a device that is designed for such use (e.g., a collar or vest), but this does not affect its behaviour. The effect of mounting the accelerometer varies considerably according to the taxon and the technique used [9], and it has been studied in several groups [10–12].

When researchers use an accelerometer to determine activity and behavioural patterns in a species for the first time, validation is a key concern [13,14]. If accelerometers are used without previous validation, the robustness of the results and replicability of experiments are compromised. Brown et al. [13] indicate that the validation process can be carried out by synchronising behavioural observations (i.e., video recordings) with simultaneously obtained data from the accelerometer. From this point, it is possible to classify accelerometer data variability more precisely, using manual methods, statistical algorithms or even specific software [13].

Tamandua tetradactyla (Mammalia: Pilosa: Myrmecophagidae) is a medium-sized anteater species endemic to South America. The Myrmecophagidae family includes Myrmecophaga trygactyla (giant anteater) and the Tamandua species: T. tetradactyla (lesser or southern anteater) and T. mexicana (northern anteater) [15–17]. The lesser anteater conservation status varies according to geographical scale. The animals are threatened in Córdoba (Argentina) [18], near threatened in Argentina [19] and of little concern globally [20].

Many aspects of Tamandua spp. biology remain to be elucidated [21,22], and their activity and behavioural pattern are no exceptions [17,23]. In the wild, T. tetradactyla individuals have been observed to be active during both light and dark phases. They show mainly arboreal habits, although they also rest, feed and move in the ground [17,24,25]. In captivity, similar observations were made in previous studies using infrared cameras [26,27]. Considering that behavioural patterns are adaptive responses determined by individual intrinsic characteristics (e.g., physiology and behaviour), as well as external factors, validating the accelerometer will allow us to expand our knowledge on T. tetradactyla behaviour. The present study was developed in light of the general hypothesis that the use of a tri-axial accelerometer would enable the characterisation of the activity pattern of zoo-housed T. tetradactyla without affecting it. Therefore, the purposes of this study were to: (1) evaluate the effect of a vest-mounted accelerometer on the general activity pattern; (2) correlate accelerometer and video recording information, evaluating fluctuations of the X, Y and Z data series (accelerometer) in relation to changes in the orientation of the animals’ movements (video); and (3) characterise the activity pattern during husbandry changes using an accelerometer.

2. Materials and Methods

2.1. Studied Animals and Housing Conditions

Ten adult Tamandua tetradactyla (henceforth referred to as ‘lesser anteaters’; n = 5 females and 5 males; average weight: 6.5 ± 0.8 and 8.5 ± 0.6 kg, respectively) were studied at Biodiversity Park (former Córdoba Zoo, 31°12.32′ S; 64°16.84′ W), Córdoba, Argentina.
Lesser anteaters were housed in individual contiguous enclosures, maintained under natural conditions of photoperiod, temperature and humidity. A diet was administered following the Manual of Nutrition and Diet for Wild Animals in Captivity [28]. Additionally, considering that food-based environmental enrichment has proven useful in favouring the expression of natural behaviours in this species [29], other food items were offered (e.g., honey, ants, and fruits).

Housing conditions were characterized by enclosures designed following recommendations for the species [21,30]. All enclosures were similar in physical characteristics and size, and were located next to each other in the same area of the institution; more details can be found in the work of Eguizábal et al. [26]. In brief, three walls were made of concrete, and the front wall was made of glass (allowing public view), while the roof was made of wire mesh (10 × 10 cm) and the floor of soil and wooden substrate (Figure 1). To achieve the possible beneficial effects of providing new exploration opportunities, animals were rotated between enclosures every 2 weeks.

**Figure 1.** Housing of adult lesser anteaters (*Tamandua tetradactyla*) at Biodiversity Park. Enclosures were under natural conditions of photoperiod, temperature and humidity. Image shows the view from the rear of enclosure 2.

**2.2. Video Cameras and Tri-Axial Accelerometer**

Behaviour was continuously monitored using infrared video cameras (HIKVISION Turbo HD- IR Turret Camera- DS 2CE56C2T IRM) and recorded using a digital video recorder (HIKVISION Turbo HD DVR- DS 7200 Series). All observations were made by the same researcher (S.P.).

The accelerometer was a Techno SmArt tri-axial-AXY-3 device [31]. It weighed 0.7 g, and its dimensions were 9 × 15 × 4 mm. The equipment was configured for continuous sampling at a frequency of 10 Hz (10 data/second/axis) and with a dynamic range of 8 g. The accelerometer was placed in the pocket of the vest, positioning the X axis in correspondence with the anterior–posterior body axis, the Y axis with the right/left and the Z axis with up/down (Figure 2, Panel A).
Animals 2022, xx, x 4 of 21

The accelerometer was placed in the pocket of the vest, positioning the X axis in correspondence with up/down (Figure 2, Panel A). The activity pattern of lesser anteaters was assessed through video analysis. The activity pattern of lesser anteaters to expose the machine in the test and due to its low weight (0.7 g: 0.009% of the animals’ average body weight).

The accelerometer was not placed in the vest pockets, since it was considered unnecessary to expose the machine in the test and due to its low weight (0.7 g: 0.009% of the animals’ average body weight).

To evaluate the effect of the vest on which the accelerometer was mounted, the activity pattern was assessed through video analysis. The activity pattern of lesser anteaters was studied using an ABA-type experimental design over 3 consecutive days in May 2019. This experimental design was chosen due to the low number of lesser anteaters available (N = 6) at that moment in the institution. In this design, each individual acted as its own control when subjected to an experimental treatment [32]. Sampling began on day one in order to obtain behavioural baseline data without the vest (stage A1); on day two, with the vest on...
Animals 2022, 12, 2516

(stage B); and ended the day after, without the vest (stage A2). It should be noted that the study started at 14:00 h, when lesser anteaters received their daily food ration and were usually active. Thus, each experimental day started at 14:00 h and ended at 13:59 h of the following calendar day.

The activity pattern of each lesser anteater was continuously monitored using a video camera located in the enclosure. The animals’ activity was sampled using the focal animal sampling method [33], registering activity events (records/day), total activity (hours/day) and activity per hour (percentage/hour). Each activity event started when an active behaviour (e.g., feeding, exploration, and locomotion) was registered, and ended when rest behaviour (the only inactive behaviour) was detected [26]. The resolution time was 1 s, and any event that lasted less than 1 s was considered irrelevant.

For the statistical analysis, activity events were analysed using a mixed general linear model (MGLM), and a Poisson error distribution was assumed. ABA stages (A1, B, and A2) were included as a fixed term, and sex and individual were random factors. The same factorial structure was used for total activity statistical analysis, but with a mixed linear model (MLM). Activity per hour was not subjected to statistical analysis due to its descriptive nature. When data did not meet a normal distribution, a natural logarithm transformation was applied.

2.4. Study 2—Video Analysis Validation of the Accelerometer

In order to evaluate fluctuations in the X, Y and Z data series (accelerometer) in relation to changes in the orientation of the animals’ movements (video), each lesser anteater (N = 10) was subjected to a 20 min test in an experimental room during August–September 2019. The experimental room (surface: 3 × 3 m; height: 2.4) was located in the laboratory next to the animal enclosures; more details can be found in the work of Zárate et al. [34,35]. The room had 4 homogeneous walls, a cement floor and a single entrance through a door, and was under controlled lighting and temperature conditions. In order to stimulate the expression of different behaviours, the experimental room had a climbing structure made of wood and wire mesh (vertical movement and climbing), and one feeder located on the floor and one feeder located above the climbing structure (feeding) (Figure 2, Panel B). These elements in the experimental room were provided to motivate a high number of different behaviours. It was expected that the behaviours registered in the videos were associated with different values of acceleration.

Prior to starting the test, the vest was mounted on each lesser anteater housed in the enclosure, and then, in the experimental room, the tri-axial accelerometer was placed in the pocket of the vest (Figure 2, Panel C). The test began after a 5 min acclimation period per animal, and a food ration was administered by two feeders after 10 min. Animal behaviour was simultaneously monitored by two means: accelerometer and video recordings. Both devices were temporally synchronized using their software. Video recordings were obtained via two cameras located in the ceiling of the experimental room, one in a central position and one in a corner.

The animals’ behaviour was sampled using a focal sampling method and employing a continuous recording rule [33]. First, through video analysis, we registered behavioural events using a previously developed ethogram [26]; inactive behaviour: resting; and active behaviours: motionless, feeding, locomotion, descent locomotion (or climbing down), self-grooming, ascendent locomotion (or climbing up), inverted locomotion (inverted climbing), exploration, alert, repetitive locomotion, and others. Then, we classified the same behavioural events in the serial data of the accelerometer (X, Y, and Z values), considering the beginning and the end of each behavioural event registered in the video. The X, Y and Z variables (acceleration values for each respective axis) and the overall dynamic body acceleration (ODBA) were considered according to Ladds et al. [36]. In order to obtain the value of the dynamic acceleration (m/s²) of each axis, static acceleration was calculated as a 3 s average and then subtracted from each of the axes [37].
Table S1 in the Supplementary Material, reports the summary measures obtained for the variables X, Y, Z and ODBA for each behaviour. These measures were used in the exploratory analysis to characterise the resulting pattern of fluctuations associated with the behaviours. Following the simplest approach for acceleration waveform statistics utilized frequently by others researchers [13], such as Mean, Minimum, Variance, among others, we were not able to establish criteria to discriminate active behaviours (e.g., locomotion vs. repetitive locomotion). Otherwise, ODBA variance was more useful than X, Y, Z variances to discriminate behavioural events classified as active and inactive. In order to explore this parameter (ODBA variance), a cluster analysis using the average Euclidean distance (non-hierarchical K-means method) was used [13]. The threshold was established at 0.0055, since it was observed that the active behaviours had greater or equal values.

To test ODBA variance in real context, a 24 h pilot test was performed during October 2019. The accelerometer was mounted on one lesser anteater in its enclosure, and the animal’s behaviour was monitored for 24 h using two cameras (one positioned in the enclosure and one in the shelter). Active and inactive behavioural events were recorded continuously; as an example, one active event may include the expression of different behaviours. Each activity event started when an active behaviour (e.g., feeding, exploration, or locomotion) was registered, and ended when rest behaviour (the only inactive behaviour) was detected [26]. Each event observed in the video recordings was associated with the corresponding accelerometer data series. Using these data, a matrix was obtained comparing the number of observed events in the video with those predicted using the accelerometer (according to the threshold value 0.0055). The precision and sensitivity percentages were calculated based on this matrix: precision = true positive/(true positive + false positive); sensitivity = true positive/(true positive + false negative). True positive: coincidence in observed event using video recordings and classification of event using the accelerometer; false positive: observed inactive event using video recordings and classified using the accelerometer as an active event; false negative: observed active event using video recordings and classified using the accelerometer as an inactive event [38,39].

2.5. Study 3—Assessment of Biological Relevance of Using Accelerometer to Monitor Activity Pattern

The activity pattern and activity level (the proportion of time that animals spend active) are behavioural information that can provide an indicator of how animals respond to changes in their environment. To evaluate the biological relevance of using an accelerometer to measure the activity level of lesser anteaters (N = 6), animals were studied in an ABA-like experimental design [32] for three consecutive days in October 2019. Lesser anteaters were studied on: day one in a routine housing condition (stage A1: baseline), day two in an enclosure with reduced complexity (stage B: treatment), and day three with their housing condition restored (stage A2). During stages A1 and A2, the complex enclosure contained a wooden shelter, several climbing structures (e.g., logs, stairs, and wire mesh ceiling), plants, and soil and wood substrate (Figure 3, Panel A). During stage B, lesser anteaters were rotated (<15 s of handling transportation) to the contiguous enclosure, in which most climbing structures were eliminated (Figure 3, Panel B).

Activity pattern was simultaneously monitored by two means: an accelerometer and video recordings. The active and inactive behaviours (records/day) of each lesser anteater were continuously monitored using two video cameras located at the enclosure, via the ethogram employed in Studies 1 and 2. First, the animals’ activity was recorded on a video camera at 5 min intervals using an instantaneous sampling method [40]. Based on previous studies [33,40,41], activity was measured for 9 s at each sampling point (1 record/sample point). A total of 288 frequency records were obtained per individual each day. Second, the activity was then associated with the obtained accelerometer data, calculating the ODBA for each sampling point (5 min intervals). After this, ODBA variance was obtained, which allowed us to classify the values of 288 sampling points per stage as active or inactive events depending on whether they exceeded the ODBA threshold.
Figure 3. Diagram representing the enclosures used during Study 3 to evaluate behavioural response of adult lesser anteaters (*Tamandua tetradactyla*) to enclosure complexity. An ABA-type experimental design was carried out. Panel A: complex enclosure with climbing structures (stages A1 and A2). Panel B: enclosure with reduced complexity (stage B).

In addition, the software ActogramJ [42] was used to build actograms based on active/inactive records from both the videos and accelerometer data. Actograms represent the way that the animals’ activity is distributed throughout the day, allowing us to ascertain their activity patterns. Using the individual actograms, smoothed mean actograms were then constructed for each of the ABA stages. It was also possible to calculate the acrophase (time of the day of peak activity) and start/end of activity (time) using both technologies.

For statistical analysis, based particularly on Study 2, it should be clarified here that the objective of the study was to measure activity/inactivity, and not detailed behaviours. First, the effect of treatment on activity patterns was analysed, considering the information obtained using the video recordings and accelerometer analysed. A mixed general linear model (MGLM) was used, and a Poisson error distribution was assumed. ABA stages...
(A1, B, and A2) were included as a fixed term, and sex and individual characteristics were random factors. Then, a T-test was employed to compare the results of the activity level per stage obtained using the camera and accelerometer. Additionally, a T-test was also employed to compare the values of the acrophase, start and end of activity obtained using both previous methodologies.

Again, a matrix for percentages of precision and sensitivity was obtained by comparing the number of active events observed in the video recordings with those predicted using the accelerometer.

2.6. Statistical Analysis

For all studies, normality was verified using the modified Shapiro–Wilks test, and variance homogeneity was verified using the F-test of equality of variances. For a posteriori tests, Fisher’s test was applied when the statistical analysis showed a \( p \)-value \( \geq 0.05 \). All analyses were performed using InfoStat [43]. The significance level was 5% for all tests. Values are reported here as the mean \( \pm \) SEM unless otherwise noted.

3. Results

3.1. Study 1—Effects of Accelerometer Mounting Using a Vest

No changes during ABA were detected for activity events (A1 = 3.67 ± 0.88; B = 2.50 ± 0.22; A2 = 2.50 ± 0.43 records/day) or total activity (A1 = 5.68 ± 0.79; B = 7.34 ± 1.93; A2 = 6.15 ± 0.98 h/day). Figure 4 shows the activity per hour (percentage/hour) according to ABA stages for both sexes.

3.2. Study 2—Correlations between Data from Accelerometer and Video Recordings

The cluster analysis for ODBA variance values is shown in Figure 5. Behaviours with lower acceleration values (rest, feeding, motionless, and others) have a common origin in the same node, in contrast to the rest. Moreover, these behaviours have an ODBA variance value close to 0, while the others have higher values (Figure 6).

During the complementary pilot test, video analysis allowed the determination of 71 activity and inactivity events on one animal housed in the enclosure. Fluctuations in the positional axes and corresponding ODBA values detected over a 24 h period using the accelerometer are illustrated in Figure 7 (Panels A and B, respectively).

During the complementary pilot test, additional information was obtained to support validation. By associating the data obtained from the video recordings and accelerometer, it was possible to compare the number of observed events in the videos with those predicted using the accelerometer: 26 active events (true positive), 35 inactive events (true positive), 10 active events classified as inactive (false negative) and 0 inactive events classified as active (false positive). These results correspond to a 100.00% precision and an 85.91% sensitivity rate. Examples of true positives and a false negative are shown in Figure 8 (time scale is not associated with the length of each event).

3.3. Study 3—Characterisation of Behavioural Response to Changes in Enclosure Complexity Using an Accelerometer

When analysing the effect of enclosure complexity on total activity, variations were detected for video recording data (B > A2 > A1 records/day; \( F_{2,46651} = 161.4; p < 0.0001 \)). The percentages of activity for these stages were 35.36 ± 3.15; 29.29 ± 2.25, and 26.15 ± 1.50, respectively. The same pattern was also observed in the study of ODBA variance data obtained from the accelerometer (B > A2 > A1 records/day; \( F_{2,5181} = 26.1; p < 0.0001 \); Figure 9; an example of an individual profile).

By linking the data from the video recordings and the accelerometer, it was possible to compare the number of events observed in the video recordings with those predicted using the accelerometer: 1121 active events (true positive), 3613 inactive events (true positive), 450 active events classified as inactive (false negative) and 0 inactive events classified as active (false positive). These results correspond to a 100.00% precision and a 91.30% sensitivity rate.
Finally, comparing total activity between the two technologies, significant differences were found in two of the three stages: the A1 and A2 stages (A1: $T = -5.22, p = 0.0004$; A2: $T = -2.24, p = 0.0486$). In both cases, the averages obtained using the accelerometer were higher than those of the video analyses. In addition to this, no statistical differences were detected between technologies when the acrophase ($T = 0.84, p = 0.4334$), start ($T = 0.49, p = 0.6332$) and end of activity ($T = -0.15, p = 0.8833$) were compared. For instance, the acrophase assessed using video recordings was $6.17 \pm 1.14$ h, and using the accelerometer, it was $8.91 \pm 3.05$ h.

Figure 4. Percentage of activity per hour of adult *Tamandua tetradactyla* (n = 3 females, Panel A; 3 males, Panel B) exposed to semi-controlled conditions during ABA stages. Behaviour was recorded continuously for 3 consecutive days (A1: baseline, without vest; B: treatment, vest on; A2: posterior, without vest). The beginning of the X axis corresponds to feeding time.
Figure 5. Cluster analysis for the ODBA variance values associated with the behaviours of adult Tamandua tetradactyla (n = 5 females and 5 males) exposed to test in the experimental room. The hierarchical sequence of cluster formation was produced taking into account the average comparison. Abbreviations: O: others; R: resting; M: motionless; F: feeding; L: locomotion; DL: descent locomotion; SG: self-grooming; AL: ascendent locomotion; E: exploration; and A: alert.

Figure 6. Overall dynamic body acceleration (ODBA) variance values of behaviours of adult Tamandua tetradactyla (n = 5 females and 5 males) exposed to test in the experimental room. Behaviour was monitored continuously by means of a tri-axial accelerometer. Box plots show median values (horizontal line), average (dark square dot), 25th and 75th percentiles (bottom and top lines), SD ± 1 (whiskers) and values outside the range (black circles). Abbreviations: O: others; R: resting; M: motionless; F: feeding; L: locomotion; DL: descent locomotion; SG: self-grooming; AL: ascendent locomotion; E: exploration; and A: alert.
Figure 7. Profiles of the accelerometer data series (Panel A) and ODBA dot plot (Panel B) obtained during the 24 h pilot test carried out on an adult Tamandua tetradactyla individual exposed to semi-controlled conditions in its enclosure. Panels A–C: X, Y and Z axes. Panel D: vertical lines indicate 60 min periods throughout the test. Light and dark periods associated with light/dark photoperiodic phases are shown.
During the complementary pilot test, additional information was obtained to support validation. By associating the data obtained from the video recordings and accelerometer, it was possible to compare the number of observed events in the videos with those predicted using the accelerometer: 26 active events (true positive), 35 inactive events (true positive), 10 active events classified as inactive (false negative) and 0 inactive events classified as active (false positive). These results correspond to a 100.00% accuracy and an 85.91% sensitivity rate. Examples of true positives and a false negative are shown in Figure 8 (time scale is not associated with the length of each event).

Figure 8. Comparison of a fraction of behavioural events observed using video recordings and predicted using an accelerometer. Data are from the 24 h pilot test carried out on an adult Tamandua tetradactyla. In the upper part, images show a frame of 1 min of video recordings. Images a and c show a lesser anteater that was active (locomotion and others), and image b shows an inactive state of the same animal (rest). In the middle part, X, Y, and Z profiles are shown for the same fractions of the video. In the lower part, detected ODBA variance values obtained from accelerometer monitoring for each behavioural event are illustrated. According to activity threshold value (0.0055), a was classified as active (true positive) and b and c as inactive (true positive and false negative, respectively).
Figure 9. Activity pattern of individual *Tamandua tetradactyla* (as an example) exposed to semi-controlled conditions in response to enclosure complexity. Experimental stages: during A1 and A2, lesser anteaters were in the complex environment, and during B, they were in the enclosure with reduced complexity. ODBA variance was calculated for each sampling point (90 records/sampling; 12 samplings/hour). The horizontal dotted line indicates the threshold value to discriminate active behaviours from inactive ones.
4. Discussion

The use of a tri-axial accelerometer was validated to assess activity patterns in adult *Tamandua tetradactyla* under human care. In Study 1, the effect of mounting the accelerometer using a vest on the animals’ activity was discarded as a source of variation in the activity pattern. This result enabled its use to monitor activity in Studies 2 and 3. During Study 2, it was possible to correlate the fluctuations in axis activity obtained using the accelerometer with the behaviours observed in video recordings. Based on Study 2, after various statistics and their values in relation to the detected behavioural activities were explored, the ODBA variance was determined as a threshold parameter that allowed for discrimination between activity and inactivity events; further research is necessary to categorise different behaviours. Finally, during Study 3, findings revealed that a tri-axial accelerometer was useful to monitor the responses of lesser anteaters to changes in environmental complexity by increasing their activity in the less complex enclosure.

Given the great potential that can be attributed to accelerometers, a first step in applying them to a new species is to find a suitable mounting device that minimally affects the behaviour of the animals [44]. With this in mind, a vest was designed specifically for use on *T. tetradactyla* individuals to allow the proper and secure positioning of the accelerometer on the animals’ bodies. The vest allowed us to keep the X axis aligned with the longitudinal body axis, with no apparent rotations or changes in position. No obvious short-term changes were observed in the animals’ activity throughout the day when using the vest (i.e., activity cycle). In fact, in the authors’ observations, aversive or disturbed behaviours were detected. However, more studies are necessary to assess the long-term effects of using vests on animals, such as thermal discomfort and/or skin damage. In addition, one problem with the design of the vest was material durability, as it was observed that some individuals damaged the vest with their claws during self-grooming, so it might be interesting to explore new textile materials that fit their bodies just as well but are more durable. Nevertheless, in relation to the other mounting devices that have been used for studying tamanduas and giant anteaters’ behaviour (*Myrmecophaga tridactyla*), the vest could be a more appropriate mounting accessory, avoiding anaesthesia and the use of rigid elements that could harm individuals [45,46]. Thus, the vest could also be a tool for attaching other small devices, such as GPS tracking systems, magnetometers, further broadening the range of behavioural studies in zoos or animal facilities [4,13,47].

During Study 2, detailed information regarding the movements and positions of lesser anteaters’ bodies for most behaviours was obtained from accelerometer monitoring. Only the behaviours of repetitive locomotion and inverted climbing were not observed in the experimental room, and had to be further characterised in the 24 h pilot test. From cluster analysis, a first approach was made, and it was determined that behaviours with low dynamic acceleration (ODBA variance close to 0; rest, motionless and feeding) and minor variations in the X, Y and Z axes had a common node. Furthermore, the associations between acceleration profiles and behavioural activities observed in video recordings revealed that the behaviours that resulted in least variations in the values of the axes involved fewer body movements. The same results in terms of rest behaviour have been found in studies of other species using accelerometers [1,39]. The lesser anteaters studied ingested artificial food with minimal body movements, helping with their tongue and/or claws but without large displacements, for which there were no marked fluctuations in acceleration values. It is worth noting that artificial food (i.e., a semi-liquid balanced diet) is very different from what the species can find in nature (i.e., social insects). However, when offering other feeding resources, which are frequently administered in food-based environmental enrichment at zoos, the fluctuations in acceleration values could be greater. It would be interesting to explore the relationship between preys and *T. tetradactyla* individuals using an accelerometer, considering that the ODBA variance parameter is useful to assess energy expenditure [13,48].

After various statistics and their values in relation to the detected behavioural activities were explored, the ODBA variance was determined as a threshold parameter that allowed
for discrimination between activity and inactivity events. This threshold proved to be robust, with high percentages of precision and sensitivity during both Studies 2 and 3. Further video exploration revealed that the false negatives were events in which an animal interrupted its resting behaviour by opening its eyes or adjusting its position to continue resting. Therefore, acceleration was very low, and behaviour was misclassified as rest according to the accelerometer records. When active behaviours were taken into account, a wide range of ODBA variance values were detected, especially for the different types of locomotion, so they could not be distinguished from each other using this parameter (or with the other statistical parameters analysed). Perhaps this is because the ODBA variance is affected by variables such as the mean, the sampling frequency, and the resolution of the accelerometer [37]. Considering that ODBA is an important parameter to assess energy expenditure, and that its combination with other measures can contribute to the integration of biomechanics, behaviour and ecology [13,48,49], further studies should focus on its application to distinguish active behaviours in T. tetradactyla. It is also important for further studies in T. tetradactyla to take into account that the ODBA parameter was useful to discriminate between active behaviours when analysis was performed using automatic machine learning [39,50,51]. This was demonstrated in other studies employing other parameters (such as static lateral acceleration) and processing accelerometer data through the automatic classification of active behaviours [52,53].

During Study 3, the use of the accelerometer proved its biological relevance in characterising the activity pattern of T. tetradactyla individuals in enclosures with different complexities of climbing elements. The analysis of both video and accelerometer data indicated that, in the environment with reduced complexity, lesser anteaters were more active. As it was not possible to define a threshold to discriminate between behaviours, future studies are necessary to detect if animals perceive husbandry changes as positive (e.g., increased exploration) or negative (e.g., increased repetitive locomotion). Furthermore, when animals returned to the complex enclosure (A2 stage), they were more active than in the basal stage (A1), perhaps indicating that the treatment effect lasted. There is limited information on this subject, but a case study showed that a greater expression of active behaviours was generated when two T. tetradactyla individuals were subjected to different types of enrichment [54]. Similarly, in the present study, the reduced environmental complexity could have acted as a novel environment, fostering the increased activity and perhaps the ability to deal with a new situation. This interpretation is supported by the fact that no increases in the expression of abnormal behaviours or stereotypies were observed (in the videos).

In the wild, it has been found that 47% of the mammals that inhabit the forests of Brazil (one of the current distribution sites of T. tetradactyla) have changed their period of activity in response to different human disturbances [55]. For example, Dasypus novemcinctus (a xenarthra-related species) has been found to increase its diurnal activity with habitat fragmentation [56]. The authors of previous studies estimate that this could be due to factors such as stress or difficulties in acquiring food, and hypothesise that the flexibility in activity patterns contributes to the ability of this species to adapt to changes induced by human activities. Although no evident changes in the activity cycle (the acrophase, start and end of activity) were found for T. tetradactyla individuals, the increased activity in the least complex environment could be understood as a positive response to environmental changes. In the context of zoological institutions, we consider a beneficial reaction of individuals, since frequently the lack of behavioural response is linked to a chronic state of stress.

When comparing total activity values obtained using both methodologies, more records per day were observed in the videos (during A1 and A2). This difference could be explained by the sensitivity of the accelerometer monitoring, which, despite being high (91.3%), indicated that not all active events were detected as such. For example, on several occasions when animals were feeding, they were almost immobile, and only extended and retracted their tongues, causing this behaviour to be misclassified as rest. Moreover, as mentioned above, some false negative events arose when animals briefly interrupted their resting
behaviour (opening their eyes, changing their position, etc.), which indicates an underestimation of their activity, albeit a biologically irrelevant one. Other studies have shown that accelerometers are useful for clearly discriminating between resting behaviour and active behaviours. However, the reduced performance of accelerometers is evident based on accuracy and precision parameters when they were applied to monitor behaviours with low acceleration, such as feeding and other movements [53,57,58]. Nevertheless, when researchers want to analyse results, it is also important to consider the position of the accelerometer on the animal body (e.g., ear or neck) and/or the species-specific feeding strategy; for example, results on feeding behaviour collected using accelerometers mounted on the necks of cattle will be different for grazing animals in a field vs. animals in a feedlot [38,59].

There are some practical aspects to consider when using the accelerometer to monitor the behaviour of the lesser anteater: (a) donning the vest requires three or four trained individuals and can be completed in a short time (<5 min); (b) in contrast, the accelerometer can be easily attached and removed by a single person; and (c) taking into account that data were manually processed, future researchers may benefit from applying a software that processes information automatically, which may facilitate the understanding of the associations between records obtained from both technologies.

Overall, the present study proved the usefulness of the tri-axial accelerometer in monitoring T. tetradactyla activity patterns, especially in evaluating the effect of changes in environmental complexity. In addition, the importance of the validation and calibration of the accelerometer in captivity was proven, similarly to other studies, before applying this methodology in other environments, such as the wild [36,60,61]. Hopefully, further studies on zoo-housed lesser anteaters should assess the long-term effect of vests on behavioural activity and skin health, before results may serve as a basis for researchers studying the behaviour of free-ranging lesser anteaters. Finally, our multidisciplinary approach could be improved by developing semi-automatic or automatic data processing that maximizes the potential of the accelerometer.

5. Conclusions

In the present study, by evaluating the behaviour of adult Tamandua tetradactyla individuals using a tri-axial accelerometer, it was found that:

- The use of a vest for mounting the accelerometer did not influence the total activity or activity pattern.
- The pattern of fluctuations in the accelerometer validated by video recordings made it possible to clearly discriminate resting behaviour from active behaviours.
- The use of the accelerometer was biologically relevant to the characterization of the response to changes in environmental complexity, employing ODBA variance as a parameter.

In this study, the use of the accelerometer placed in a vest was demonstrated to be a reliable method to determine the activity patterns of lesser anteaters under semi-controlled conditions. Considering that zoo-housed lesser anteaters exhibit diurnal and nocturnal activity patterns, this study contributes to finding a strategy to easily monitor their activities and responses to different management procedures supporting welfare programs, as well as ex situ conservation.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/ani12192516/s1, Table S1: Descriptive statistics obtained for the X, Y, Z axes and the ODBA according to the behaviours observed in Tamandua tetradactyla individuals during 20-minute test at the experimental room and 24 h complementary test at the enclosure.

Author Contributions: Conceptualisation, S.P. and J.M.B.; methodology, S.P., C.C., L.V.F., L.D., G.V.E., C.J.A., D.P.V. and J.M.B.; investigation, S.P. and J.M.B.; resources, J.M.B., L.V.F., C.C. and D.P.V.; writing—original draft preparation, S.P., G.V.E. and J.M.B.; writing—review and editing, S.P., G.V.E., J.M.B., C.J.A., C.C. and L.V.F.; supervision, J.M.B. and C.C.; project administration, J.M.B.;
funding acquisition, J.M.B. and L.V.F. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Argentinian Ministry of Science, Technology and Productive Innovation, grant number BID.PICT.2014.2642.

**Institutional Review Board Statement:** The study was conducted according to the Argentinean National Law for Animal Protection (Nº 14346) and the CONICET Ethics Committee (Resolution 1047 Annex II, 2005), and approved by the Bioethics and Laboratory Animal Commission on 9 April 2015, following the Annex to the Regulation of ‘Care and Use of Laboratory Animals’ (IBVY/CONICET- FCEFyN-UNC). Furthermore, the *Guidelines for the ethical use of animals in applied animal behaviour research* provided by the International Society for Applied Ethology were followed (Sherwin et al., 2003).

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** The data presented in this study are available upon reasonable request from the corresponding authors.

**Acknowledgments:** The authors would like to thank the staff members at the Parque de la Biodiversidad (the former Córdoba Zoo, renamed by the local government in December 2020) for their assistance and collaboration during the studies and Nuremberg Zoo for providing the accelerometer. The vest was made by Florencia Rivero. The vectorized image of lesser anteater used in Figures 2 and 3 was designed by graphic designer Antonela Munizaga. Help with statistical analysis was provided by Julio A. Di Rienzo. The study corresponds to the final thesis of Sofia Pavese for her degree in biology, and the research was revised by professors at the Universidad Nacional de Córdoba. We are grateful to Danielle Brown for her comments on an earlier version of this study, which enabled us to significantly improve the manuscript. The manuscript was revised by the service offered at https://www.proofreadingservices.com/all-services.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Clemente, C.J.; Cooper, C.E.; Withers, P.C.; Freakley, C.; Singh, S.; Terrill, P. The private life of echidnas: Using accelerometry and GPS to examine field biomechanics and assess the ecological impact of a widespread, semi-fossorial monotreme. *J. Exp. Biol.* 2016, 219, 3271–3283. [CrossRef] [PubMed]
2. Williams, C.T.; Barnes, B.M.; Buck, C.L. Integrating physiology, behavior, and energetics: Biologging in a free-living arctic hibernator. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* 2016, 202, 53–62. [CrossRef] [PubMed]
3. Pagano, A.M.; Rode, K.D.; Cutting, A.; Owen, M.A.; Jensen, S.; Ware, J.V.; Robbins, C.T.; Durner, G.M.; Atwood, T.C.; Oubbard, M.E.; et al. Using tri-axial accelerometers to identify wild polar bear behaviors. *Endanger. Species Res.* 2017, 32, 19–33. [CrossRef]
4. Hughey, L.F.; Hein, A.M.; Strandburg-Peshkin, A.; Jensen, E.H. Challenges and solutions for studying collective animal behaviour in the wild. *Philos. Trans. R. Soc. B Biol. Sci.* 2018, 373, 20170005. [CrossRef] [PubMed]
5. Reinhardt, K.D.; Vyazovskiy, V.V.; Hernandez-Aguilar, R.A.; Imron, M.A.; Nekaris, K.A.I. Environment shapes sleep patterns in a wild nocturnal primate. *Sci. Rep.* 2019, 9, 1–13. [CrossRef] [PubMed]
6. Gleiss, A.C.; Wilson, R.P.; Shepard, E.L. Making overall dynamic body acceleration work: On the theory of acceleration as a proxy for energy expenditure. *Methods Ecol. Evol.* 2011, 2, 23–33. [CrossRef]
7. Gervasi, V.; Brunberg, S.; Swenson, J.E. An individual-based method to measure animal activity levels: A test on brown bears. *Wildl. Soc. Bull.* 2006, 34, 1314–1319. [CrossRef]
8. Shepard, E.L.; Wilson, R.P.; Quintana, F.; Laich, A.G.; Liebsch, N.; Albareda, D.A.; Halsey, L.G.; Gleiss, A.; Morgan, D.T.; Myers, A.E.; et al. Identification of animal movement patterns using tri-axial accelerometry. *Endanger. Species Res.* 2008, 10, 47–60. [CrossRef]
9. Withey, J.C.; Bloxton, T.D.; Marzluff, J.M. Effects of Tagging and Location Error in Wildlife Radiotelemetry Studies. In *Radio tracking and Animal Populations*; Academic Press: Cambridge, MA, USA, 2001; pp. 43–75. [CrossRef]
10. Herbst, L.; Redford, K. Home-range size and social spacing among female common long-nosed armadillos (*Dasypus novemcinctus*). *Natl. Geogr. Res. Explor.* 1991, 7, 236–237.
11. Golabek, K.A.; Jordan, N.R.; Clutton-Brock, T.H. Radiocollars do not affect the survival or foraging behaviour of wild meerkats. *J. Zool.* 2008, 274, 248–253. [CrossRef]
12. Swenson, J.E.; Wallin, K.; Ericsson, G.; Cederlund, G.; Sandegren, F. Effects of ear-tagging with radiotransmitters on survival of moose calves. *J. Wildl. Manag.* 1999, 63, 354–358. [CrossRef]
13. Brown, D.D.; Kay, R.; Wikelski, M.; Wilson, R.; Klimley, A.P. Observing the unwatchable through acceleration logging of animal behavior. *Anim. Biotelemetry* 2013, 1, 20. [CrossRef]
14. Kröschel, M.; Reineking, B.; Werwie, F.; Wildi, F.; Storch, I. Remote monitoring of vigilance behavior in large herbivores using acceleration data. *Anim. Biotelemetry* 2017, 5, 10. [CrossRef]

15. Redford, K.H.; Eisenberg, J.F. *Order Xenarthra (Edentata). Mammals of the Neotropics: The Southern Cone*; The University of Chicago Press: Chicago, IL, USA, 1992; pp. 46–68.

16. Torres, R.; Monguilot, J.; Bruno, G.; Michelutti, P.; Ponce, A. Ampliación del límite austral de la distribución del oso melero (*Tamandua tetradactyla*) en la Argentina. *Nótilas Faunísticas* 2009, 39, 1–5.

17. Hayssen, V. *Tamandua tetradactyla* (Pilosa: Myrmecophagidae). *Mamm. Species* 2011, 43, 64–74. [CrossRef]

18. Tamburini, D. Orden Cingulata. In *Mamíferos de Córdoba y su Estado de Conservación*; Torres, R., Tamburini, D., Eds.; Editorial de la Universidad Nacional de Córdoba: Córdoba, Argentina, 2018; pp. 87–111.

19. Varela, D.; Cirignoli, S.; Torres, R.M.; Superina, M. *Tamandua Tetradactyla*. en *Categorización 2019 de los Mamíferos de Argentina Según su Riesgo de Extinción*; SayDS–SAREM, Ed.; Secretaría de Gobierno de Ambiente y Desarrollo Sostenible: Buenos Aires, Argentina, 2019. [CrossRef]

20. Miranda, F.; Fallabrino, A.; Arteaga, M.; Tirira, D.G.; Meritt, D.A.; Superina, M. Maintenance of Xenarthra in Captivity. In *The Biology of the Xenarthra*; Vizzacino, S.F., Loughry, W.J., Eds.; University Press of Florida: Gainesville, FL, USA, 2008; pp. 232–243.

21. Superina, M.; Miranda, F.; Plese, T. Monitoring of Xenarthra in captivity. In *The Biology of the Xenarthra*; Vizzacino, S.F., Loughry, W.J., Eds.; University Press of Florida: Gainesville, FL, USA, 2008; pp. 232–243.

22. Montgomery, G.G. Movements, foraging and food habits of the four extant species of Neotropical vermilinguas (*Mammalia: Myrmecophagidae*). In *The Evolution and Ecology of Armadillos, Sloths, and Vermilinguas*; Montgomery, G.G., Ed.; Smithsonian Institution Press: Washington, DC, USA, 1985; pp. 365–377.

23. Fernandes, T.N.; Young, R.J. Fluctuations in the tympanic membrane temperatures of non-restrained captive giant anteaters and southern tamanduas. *J. Zool.* 2008, 274, 94–98. [CrossRef]

24. Eguizábal, G.V.; Palme, R.; Superina, M.; Asencio, C.J.; García Capocasa, M.C.; Busso, J.M. Characterization and correlations of behavioral and adrenocortical activities of zoo-housed lesser anteaters (*Tamandua tetradactyla*). *Zoo Biol.* 2019, 38, 334–342. [CrossRef] [PubMed]

25. Eguizábal, G.V.; Palme, R.; Superina, M.; Asencio, C.J.; Villarreal, D.P.; Borrelli, L.; Busso, J.M. Non-Invasive Assessment of the Seasonal Stress Response to Veterinary Procedures and Transportation of Zoo-Housed Lesser Anteater (*Tamandua tetradactyla*). *Animals* 2022, 12, 75. [CrossRef] [PubMed]

26. Dierenfeld, E.S.; Graffam, W.S. *Manual de Nutrición y Dietas para Animales Silvestres en Cautiverio* (Ejemplos para Animales de América Latina); Zoo Conservation Outreach Group: New Orleans, LA, USA, 1996.

27. Eguizábal, G.V.; Palme, R.; Villarreal, D.; Del Borgo, C.; Di Rienzo, J.A.; Busso, J.M. Assessment of adrenocortical activity and behaviour of the collared anteater (*Tamandua tetradactyla*) in response to food-based environmental enrichment. *Zoo Biol.* 2013, 32, 632–640. [CrossRef]

28. Rojano-Bolaño, C.; Miranda-Cortés, L.; Ávila-Avilán, R.; Álvarez-Otero, G. Parámetros hematológicos de Hormigueros gigantes (*Myrmecophaga tridactyla*) Linnaeus, 1758) de vida libre en Pore, Colombia. *Vet. Zootec.* 2014, 8, 85–99. [CrossRef]

29. TecnosM’Art Europe. GPS Tracking Systems for Animals. Available online: https://www.technosmart.eu/ (accessed on 5 September 2019).

30. Ladds, M.A.; Thompson, A.P.; Kadar, J.P.; Slip, D.J.; Hocking, D.P.; Harcourt, R.G. Super machine learning: Improving accuracy and reducing variance of behaviour classification from accelerometer. *Anim. Biotelemetry* 2017, 5, 1–9. [CrossRef]

31. Shepard, E.L.; Miller, G.G.; Montgomery, G.G. *Mammals of the Neotropics* (Pilosa: Myrmecophagidae). *Mamm. Species* 2011, 43, 64–74. [CrossRef]

32. Vázquez Diosdado, J.A.; Barker, Z.E.; Hodges, H.R.; Amory, J.R.; Croft, D.P.; Bell, N.J.; Codling, E.A. Classification of behaviour in housed dairy cows using an accelerometer-based activity monitoring system. *Anim. Biotelemetry* 2015, 3, 15. [CrossRef]

33. Shepherd, E.L.; Wilson, R.P.; Halsey, L.G.; Quintana, F.; Laich, A.G.; O’Connor, C.E.; Penheric, C.J. Guidelines for the ethical use of animals in applied animal behaviour research. *Appl. Anim. Behav. Sci.* 2003, 81, 291–305, (updated 2017, International Society for Applied Ethology). [CrossRef]

34. Shephard, E.L.; Wilson, R.P.; Halsey, L.G.; Quintana, F.; Laich, A.G.; O’Connor, C.E.; Penheric, C.J. Guidelines for the ethical use of animals in applied animal behaviour research. *Appl. Anim. Behav. Sci.* 2003, 81, 291–305, (updated 2017, International Society for Applied Ethology). [CrossRef]

35.科技SmArt Europe. GPS Tracking Systems for Animals. Available online: https://www.technosmart.eu/ (accessed on 5 September 2019).

36. Scheppe, E.L.; Wilson, R.P.; Halsey, L.G.; Quintana, F.; Laich, A.G.; O’Connor, C.E.; Penheric, C.J. Guidelines for the ethical use of animals in applied animal behaviour research. *Appl. Anim. Behav. Sci.* 2003, 81, 291–305, (updated 2017, International Society for Applied Ethology). [CrossRef]
42. Schmid, B.; Helfrich-Förster, C.; Yoshii, T. A new ImageJ plug-in “ActogramJ” for chronobiological analyses. J. Biol. Rhythm. 2011, 26, 464–467. [CrossRef] [PubMed]

43. Di Rienzo, J.A.; Casanova, F.; Balzarini, M.G.; Gonzalez, L.; Tablada, M.; Robledo, C.W. InfoStat versión 2019. Grupo InfoStat, Faculty of Agricultural Sciences, National University of Córdoba, Argentina, 2019. Available online: http://www.infostat.com.ar (accessed on 16 March 2022).

44. Gleiss, A.C.; Dale, J.J.; Holland, K.N.; Wilson, R.P. Accelerating estimates of activity-specific metabolic rate in fishes: Testing the applicability of acceleration data-loggers. J. Exp. Mar. Biol. Ecol. 2010, 385, 85–91. [CrossRef]

45. Brown, D.D. Activity Patterns and Space Use of Northern Tamandua Anteaters (Tamandua mexicana) on Barro Colorado Island, Panama; University of California: Davis, CA, USA, 2011.

46. Di Blanco, Y.E.; Pérez, J.I.; Díaz, P.; Sporring, Y.K. Cinco años de radiomarcaje de osos hormigueros (Myrmecophaga tridactyla): Mejoras implementadas y lecciones aprendidas. Edentata 2012, 13, 49–55. [CrossRef]

47. Cooke, S.J.; Hinch, S.G.; Wikelski, M.; Andrews, R.D.; Kuchel, L.J.; Wolcott, T.G.; Butler, P.J. Biotelemetry: A mechanistic approach to ecology. Trends Ecol. Evol. 2004, 19, 334–343. [CrossRef] [PubMed]

48. Nathan, R.; Spiegel, O.; Fortmann-Roe, S.; Harel, R.; Wikelski, M.; Getz, W.M. Using tri-axial acceleration data to identify behavioral modes of free-ranging animals: General concepts and tools illustrated for griffon vultures. J. Exp. Biol. 2012, 215, 986–996. [CrossRef]

49. Halsey, L.G.; Shepard, E.L.; Wilson, R.P. Assessing the development and application of the accelerometry technique for estimating energy expenditure. Comp. Biochem. Physiol. Part A Mol. Integr. Physiol. 2011, 158, 305–314. [CrossRef]

50. McClune, D.W.; Marks, N.J.; Wilson, R.P.; Houghton, J.D.R.; Montgomery, I.W.; McGowan NE; Gormley, E.; Scantlebury, M. Tri-axial accelerometers quantify behaviour in the Eurasian badger (Meles meles): Towards an automated interpretation of field data. Anim. Biotelemetry 2014, 2, 5. [CrossRef]

51. Kumpulainen, P.; Valdeoirola Cardó, A.; Sompi, S.; Törnqvist, H.; Vääätäjä, H.; Majaranta, P.; Gizatdinova, Y.; Hoog Antink, C.; Surakka, V.; Kujala, M.V.; et al. Dog behaviour classification with movement sensors placed on the harness and the collar. Appl. Anim. Behav. Sci. 2021, 241, 105393. [CrossRef]

52. Barwick, J.; Lamb, D.W.; Dobos, R.; Welch, M.; Trotter, M. Categorising sheep activity using a tri-axial accelerometer. Comput. Electron. Agric. 2018, 145, 289–297. [CrossRef]

53. Nekaris, K.A.I.; Campera, M.; Chimienti, M.; Murray, C.; Balestri, M.; Showell, Z. Training in the Dark: Using Target Training for Non-Invasive Application and Validation of Accelerometer Devices for an Endangered Primate (Nycticebus bengalensis). Animals 2022, 12, 411. [CrossRef]

54. De Godoy Peixoto, F.B.; Ambrózio, M.T.G.; Colbachini, H.; Padilha, F.L.A.; Costa, F.R. Enriquecimento ambiental aplicado a tamanduás-mirins (Tamandua tetradactyla) no Aquário de São Paulo: Estudo de caso. Pesqui. Ensino Ciências Exatas Nat. 2019, 3, 119–124. [CrossRef]

55. Mendes, C.P.; Carreira, D.; Pedrosa, F.; Beca, G.; Lautenschlager, L.; Akkawi, P.; Berce, W.; Ferraz, K.M.M.B.; Galetti, M. Landscape of human fear in Neotropical rainforest mammals. Biol. Conserv. 2020, 241, 108257. [CrossRef]

56. Norris, D.; Michalski, F.; Peres, C.A. Habitat patch size modulates terrestrial mammal activity patterns in Amazonian forest fragments. J. Mammal. 2010, 91, 551–560. [CrossRef]

57. Wang, Y.; Nickel, B.; Rutishauser, M.; Bryce, C.M.; Williams, T.M.; Elkaim, G.; Wilmers, C.C. Movement, resting, and attack behaviors of wild pumas are revealed by tri-axial accelerometer measurements. Mov. Ecol. 2015, 3, 2. [CrossRef] [PubMed]

58. Nogoy, K.M.C.; Chon S-i; Padilha, F.L.A.; Lee, D.-H.; Choi, S.H. High Precision Classification of Resting and Eating Behaviors of Cattle by Using a Collar-Fitted Triaxial Accelerometer Sensor. Sensors 2022, 22, 5961. [CrossRef]

59. Sprinkle, J.E.; Sagers, J.K.; Hall, J.B.; Ellison, M.J.; Yelich, J.V.; Brennan, J.R.; Taylor, J.B.; Lamb, J.B. Predicting Cattle Grazing Behavior on Rangeland using Accelerometers. Rangel. Ecol. Manag. 2021, 76, 157–170. [CrossRef]

60. Hammond, T.T.; Springthorpe, D.; Walsh, R.E.; Berg-Kirpatrick, T. Using accelerometers to remotely and automatically characterize behavior in small animals. J. Exp. Biol. 2016, 219, 1618–1624. [CrossRef]

61. Chen, J.; Brown, G.; Fudickar, A. Simulation-based validation of activity logger data for animal behavior studies. Anim. Biotelemetry 2021, 9, 31. [CrossRef]