6-IncRNA Assessment Model for Monitoring and Prognosis of HER2-Positive Breast Cancer: Based on Transcriptome Data

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Background: In view of the high malignancy and poor prognosis of human epidermal growth factor receptor 2 (HER2)-positive breast cancer, we analyzed the RNA expression profiles of HER2-positive breast cancer samples to identify the new prognostic biomarkers.

Methods: The linear fitting method was used to identify the differentially expressed RNAs from the HER2-positive breast cancer RNA expression profiles in the Cancer Genome Atlas (TCGA). Then, a series of methods including univariate Cox, Kaplan-Meier, and random forests, were used to identify the core long non-coding RNAs (lncRNAs) with stable prognostic value for HER2-positive breast cancer. A clinical feature analysis was performed, and a competing endogenous RNA network was constructed to explore the role of these core lncRNAs in HER2-positive breast cancer. In addition, a functional analysis of differentially expressed messenger RNAs in HER-2 positive breast cancer also provided us with some enlightening insights.

Results: The high expression of four core lncRNAs (AC010595.1, AC046168.1, AC069277.1, and AP000904.1) was associated with worse overall survival, while the low expression of LINC00528 and MIR762HG was associated with worse overall survival. The 6-lncRNA model has an especially good predictive power for overall survival (p < 0.0001) and 3-year survival (the area under the curve = 0.980) in HER2-positive breast cancer patients.

Conclusion: This study provides a new efficient prognostic model and biomarkers of HER2-positive breast cancer. Meanwhile, it also provides a new perspective for elucidating the molecular mechanisms underlying HER2-positive breast cancer.

Keywords: human epidermal growth factor receptor 2-positive breast cancer, random forests, prognosis, long non-coding RNA, competing endogenous RNA

Abbreviations: HER2, human epidermal growth factor receptor 2; TCGA, the Cancer Genome Atlas; lncRNA, long non-coding RNA; ceRNA, competing endogenous RNA; mRNA, messenger RNA; ROC, receiver operating characteristic; AUC, area under the curve; miRNA, microRNA; GEPIA, Gene Expression Profiling Interactive Analysis; GO, gene ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; STRING, search tool for the retrieval of interacting genes; PPI, protein-protein interaction; WGCNA, weighted gene co-expression network analysis; AMPK, Adenosine 5′-monophosphate (AMP)-activated protein kinase
INTRODUCTION

About 15–20% of breast cancers are HER2 positive [1]. HER2 positivity, namely HER2 protein overexpression, is an important indicator of strong tumor aggressiveness and poor prognosis [2]. Due to the strong heterogeneity of breast cancer, the prognosis and treatment of HER2-positive breast cancer are significantly different from other breast cancer subgroups [3, 4]. Although new treatments have improved HER2-positive breast cancer prognosis in recent years, its persistently high mortality indicates that the situation is still alarming. Therefore, the identification of new HER2-positive breast cancer specific biomarkers is particularly urgent.

Recent comprehensive transcriptome analysis revealed that there is a specific IncRNA expression pattern in different subtypes of breast cancer [5]. The discovery of IncRNAs, ranging in length from 200 nucleotides to 100 kilobases, has helped scientists break free from their conventional view that gene regulation in biology mainly involves protein-coding genes [6, 7]. IncRNAs can regulate gene expression at the transcriptional, post-transcriptional and epigenetic levels, which in turn affects the clinical course of the disease [8]. Compared with mRNAs and microRNAs (miRNAs), IncRNAs may be a direct indicator of tumor status [9, 10]. Studies have shown that the function of IncRNAs is associated with disease progression and patient survival of cancers, such as non-small cell lung cancer, prostate cancer and gastric cancer [11–13]. Previous studies have established prognostic models for breast cancer through RNA and alternative splicing [14, 15], but the establishment of a IncRNA system to predict HER2 positive breast cancer patients’ prognosis is hardly seen.

In this study, we applied a series of reliable bioinformatics methods to analyze the transcriptome data of HER2-positive breast cancer. Six core IncRNAs associated with stable prognosis value were identified using the randomForestSRC package and a good prognostic model was constructed. To explore the role of these core IncRNAs in HER2-positive breast cancer, we constructed a corresponding ceRNA network and performed a clinical feature analysis. In addition, we performed functional analysis of differentially expressed mRNAs and assessed the prognostic value of the core IncRNAs in various tumors in Gene Expression Profiling Interactive Analysis (GEPIA) for more comprehensive information. Features recognized in this study also helped us gain further understanding of the biological behavior of HER2-positive breast cancer at the molecular level.

MATERIALS AND METHODS

Downloading of Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer RNA Expression Data and the Corresponding Clinical Data

Highly developed microarrays and high-throughput sequencing technologies have lent powerful support for deciphering cancer genetic code [16]. TCGA provides unprecedented large-scale data for cancer researches, which provide great opportunities for revealing genomic variation in a variety of tumors [17]. RNA expression profile data of HER2-positive breast cancer samples including IncRNA, miRNA, and mRNA sequence information, were downloaded from TCGA (https://portal.gdc.cancer.gov/). These RNA data were derived from 113 HER2-positive breast cancer tissues and 105 tumor-adjacent normal breast tissues. The corresponding clinical data of the 113 HER2-positive breast cancer patients were also downloaded (Supplementary Table S1). This study has strictly followed the TCGA publication guidelines (https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga/using-tcga/citing-tcga) and all data are available without restrictions on their use in publications.

Identification of Differentially Expressed RNAs in Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer and the Functional Analysis of Differentially Expressed Messenger RNAs

We used the limma package [18] to identify differentially expressed RNAs (including IncRNAs, miRNAs, and mRNAs) between the 113 HER2-positive breast cancer tissues and 105 tumor-adjacent normal breast tissues. Then we selected the differentially expressed mRNAs for functional enrichment analysis including gene ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis in the R language environment [19].

Construction and Analysis of the Protein-Protein Interaction Network

The Search Tool for the Retrieval of Interacting Genes (STRING) database (https://string-db.org) was used to construct a protein-protein interaction (PPI) network [20]. We input the names of differentially expressed mRNAs into the multiple proteins option input box of STRING database in the form of a list and selected “homo” for “species.” The interaction score of nodes was greater than 0.4. The PPI network was downloaded and visualized in Cytoscape. Furthermore, Cytoscape-MCODE was used to perform module analysis on the PPI network [21, 22]. Modules with score >4 and nodes ≥5 were selected for the functional enrichment analysis in the STRING database. Cytoscape-Centiscape was used to analyze the internal connectivity of the PPI network [23]. The statistical criterion for functional enrichment analysis was FDR <0.05.

Identification of Long Non-Coding RNAs With Prognostic Value for Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer

First, we selected IncRNAs with prognostic value for HER2-positive breast cancer patients by applying two methods, including univariate Cox regression analysis and Kaplan-Meier survival analysis (statistical criterion: p < 0.05). To achieve that, the survival package was used [24]. Then the overlap of the prognostic IncRNAs resulted from the above two methods was obtained. Next, the randomForestSRC package [25] was used to perform the high-dimensional variable selection for survival data on the overlapped IncRNAs, and IncRNAs with stable prognostic value for
HER2-positive breast cancer were selected as core lncRNAs for subsequent analysis. Random forests is one of the classical methods for dimensionality reduction, and its results are highly reliable.

**Construction of the Prognostic Model for Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer**

The multiCox model was constructed using the core lncRNAs. This model was used to predict the survival risk of HER2-positive patients. The 113 HER2-positive breast cancer samples from TCGA were classified into high-risk and low-risk groups, with the median signature risk score as the cut-off value. Furthermore, we used the receiver operating characteristic (ROC) curve to evaluate the predictability of the 6-lncRNA model for the 3-year survival rate for HER2-positive breast cancer. To achieve that, the survivalROC package was used [26]. The results of the grouping, according to the classification, were compared with the actual 3-year survival status of HER2-positive breast cancer patients to obtain a false positive rate and a true positive rate. These rates were respectively represented on the abscissa (X axis) and the ordinate (Y axis) to plot the ROC curves. AUC, the area under the curve, was calculated as a measure of prognostic ability. The greater the AUC, the better the predictive ability. The risk score is the sum of the Cox regression coefficient multiplied by the expression value of each core lncRNA. The calculation formula is as follows:

\[
\text{Risk score} = \beta_1 \times \text{expression level of lncRNA1} + \beta_2 \times \text{expression level of lncRNA2} + \cdots + \beta_n \times \text{expression level of lncRNA}_n
\]

\(\beta\) is the Cox regression coefficients.

**Results**

Differentially Expressed RNAs in Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer

By using the limma package, we obtained data concerning the difference in RNA expression levels in HER2-positive breast cancer. When FDR < 0.05 and \(|\log_{2}\text{FC}| > 1\) were set as the thresholds for the significance of the gene expression difference, we identified 350 differentially expressed lncRNAs, 163 differentially expressed miRNAs, and 1910 differentially expressed mRNAs in those 113 HER2-positive breast cancer samples (Figure 1).

Results of the Functional Analysis of Differentially Expressed Messenger RNAs in Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer Patients and the Analysis of the Protein-Protein Interaction Network

The number of mRNAs selected according to the statistical criteria of FDR < 0.05 and \(|\log_{2}\text{FC}| > 1\) reached 1910. In order to improve the accuracy of this study, from that pool we further selected 411 differentially expressed mRNAs based on two criteria (\(|\log_{2}\text{FC}| > 2\) and FDR < 0.05) for a separate functional enrichment analysis. Results with the lowest FDR values are shown in Figure 2. The PPI network downloaded from the STRING database showed that there were significant enrichment regions (Supplementary Figure S1). Cytoscape-MCODE module analysis further showed that there were six significant modules (score >4 and nodes ≥5) in the PPI network (Supplementary Figure S2), among which the following three modules were most significant: Module 1 (score = 48.857, nodes = 50), Module 2 (score = 14, nodes = 14), and Module 3 (score = 9.357, nodes = 29). Supplementary Table S2 shows the data of the module analysis downloaded from Cytoscape.

Module 1 was primarily associated with mitosis. Module 2 was closely associated with immune functions. Module 3 was primarily involved in lipid metabolism. Module 4 was primarily involved in cell differentiation, stratum corneum formation, and signaling by Receptor Tyrosine Kinases. Module 5 was involved in post-translational protein phosphorylation. Module 6 participated in digestion and absorption, and degradation of the extracellular matrix. The results of the modules' functional analysis are included in Supplementary Table S3. The connectivity analysis
by Cytoscape-Centiscape of the PPI network showed that 88 differentially expressed mRNAs had a degree greater than 20, and they mainly concentrated in Module 1 (50), Module 2 (5), Module 3 (15), Module 4 (10), and Module 6 (5). The main results of the connectivity analysis are included in Supplementary Table S4.

The Core Long Non-Coding RNAs With Stable Prognostic Value for Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer

There were 504 lncRNAs with significant prognostic value for HER2-positive breast cancer by the univariate Cox regression analysis. Results from 383 lncRNAs indicated significant prognostic value for HER2-positive breast cancer by Kaplan-Meier survival analysis. The number of the overlap of the lncRNAs identified by the two methods was 126. Figure 3 is plotted with the Venn online software (https://bioinfogp.cnb.csic.es/tools/venny/index.html). After a high-dimensional selection of the survival random variables was performed on the 126 lncRNAs using the randomForestSRC package, and the following six stable survival-related lncRNAs were obtained: AC010595.1, AC046168.1, AC069277.1, AP000904.1, LINC00528, and MIR762HG. In consideration of the differential expression analysis, we found that the expression levels of five core lncRNAs (AC010595.1, AC046168.1,
AC069277.1, AP000904.1 and MIR762HG) in HER2-positive breast cancer tissues were higher than those in adjacent normal breast tissues (|t| > 2 and FDR < 0.05), while there was no significant difference in the expression of LINC00528. Taking into account the results of clinical feature analysis, this may be because the expression level of LINC00528 in early HER2-positive breast cancer (T1/2) was higher than advanced HER2-positive breast cancer (T3/4) (|t| = 1.930803441, p = 0.060616568). The details of these six core lncRNAs are shown in Tables 1 and 2.

### The Prognostic Model of Human Epidermal Growth Factor Receptor 2-Positive Breast Cancer

Each of the six core lncRNAs had a good single-factor prognosis for overall survival and 3-year survival in HER2-positive breast cancer patients (Figure 4: p < 0.05; Figure 5: AUC > 0.7). The multCox model of the six core lncRNAs (AC010595.1, AC046168.1, AC069277.1, AP000904.1, LINC00528, and MIR762HG) had a high-level capability of risk assessment for HER2-positive breast cancer patients. The formula for determining the risk score is given below:

$$\text{Risk score} = (4.39 \times \text{expression level of AC010595.1}) + (5.56 \times \text{expression level of AC046168.1}) + (20.61 \times \text{expression level of AC069277.1}) + (13.89 \times \text{expression level of AP000904.1}) + (-18.45 \times \text{expression level of LINC00528}) + (-4.09 \times \text{expression level of MIR762HG}).$$

According to the median signature risk score, the 113 HER2-positive breast cancer samples from TCGA were divided into high-risk and low-risk groups with significant overall survival differences (p < 0.0001), and the ROC curve showed that the 3-year survival rates were predicted with high accuracy (AUC = 0.980) (Figure 6).

### The Competing Endogenous RNA Network of Core Long Non-Coding RNAs and Results of the Functional Enrichment Analysis

Sixteen miRNAs and 129 mRNAs were identified for the construction of the ceRNA network of core lncRNAs. Figure 7 shows the ceRNA network. Three core lncRNAs (AC010595.1, AP000904.1 and MIR762HG) were closely associated in the ceRNA network, and the functional analysis results of their target mRNAs were similar: they were involved in biological processes including muscle organ development, regulation of vasculature development, and angiogenesis. They were enriched in KEGG pathways of proteoglycans in cancer, cGMP-PKG signaling pathway. They were also closely associated with focal adhesion. Notably, the KEGG analysis showed that target mRNAs of AP000904.1 were also enriched in the PI3K-Akt signaling pathway and Ras signaling pathway. Moreover, FGF2 (|log2FC| = 2.60345034144707, FDR = 7.58730025761752E-75), the famous cancer-related gene, was found to be a target mRNA of three core lncRNAs (AC010595.1, AP000904.1 and MIR762HG) in the ceRNA network. Based on the above analysis, we also discovered that FGF2 had a high connectivity in the PPI network and was subordinate to Module 3 (degree = 58).

The GO analysis showed that the target mRNAs of LINC00528 were involved in biology processes activated by immune cells such as T cells, lymphocytes and leukocytes, and were enriched in positive regulation of leukocyte cell-cell adhesion. The KEGG analysis showed that these target mRNAs were also enriched in...
hematopoietic cell lineage, besides being involved in primary immunodeficiency, Th1 and Th2 cell differentiation, and T cell receptor signaling pathway.

The functional analysis showed that the target mRNAs of AC046168.1 participated in the biology processes of epidermal cell differentiation, and epidermis development. The target mRNAs of AC069277.1 were associated with epigenetics, and were involved in the biology process of transcription corepressor activity and KEGG pathways of transcriptional mis-regulation in cancer, and necroptosis. Table 3 lists the primary results of the functional enrichment analysis of target mRNAs of each core lncRNA. A Q-value less than 0.05 is considered statistically significant. Supplementary Figures S3–S8 show the bubble diagrams.

Results of the Clinical Feature Analysis of Core Long Non-Coding RNAs and Their Prognostic Value in Various Tumors Based on Gene Expression Profiling Interactive Analysis

The results from the Student’s t-test showed that the six core lncRNAs were associated with some clinical parameters of HER2-positive breast cancer. It is worth noting that the expression level of LINC00528, to a certain extent, can distinguish between patients with early HER2-positive breast cancer and patients with advanced HER2-positive breast cancer: the expression level of LINC00528 in early HER2-positive breast cancer patients (T1/2) was higher than that in advanced HER2-positive breast cancer patients (T3/4) (t = 1.93083441, p = 0.060616568). Compared with non-targeted molecular therapy group, the expression level of LINC00528 was significantly higher in the targeted molecular therapy group (|t| > 2). These results are consistent with the better prognosis in the high expression group of LINC00528. The details are shown in Supplementary Table S5.

At present in GEPIA (http://gepia.cancer-pku.cn/), only relevant information of LINC00528 was available while the data of the other five core lncRNAs were not. We analyzed the prognostic value of LINC00528 for all 31 diseases included by GEPIA and found that the high expression of LINC00528 was associated with better overall survival in breast invasive carcinoma, cervical squamous cell carcinoma and endocervical adenocarcinoma, head and neck squamous cell carcinoma, lung adenocarcinoma and skin cutaneous melanoma, while the high

### Table 2 | The expression of core lncRNAs in HER2-positive breast cancer.

| Symbol | Log2FC | t       | p value    | FDR        |
|--------|--------|---------|------------|------------|
| AC010595.1 | -0.076688216 | -4.56493508 | 8.69E-06 | 3.85E-05 |
| AC046168.1 | -0.034387366 | -2.338474983 | 0.020433454 | 0.045385091 |
| AC069277.1 | -0.007222874 | -2.40156115 | 0.017169908 | 0.038882814 |
| AP000904.1 | -0.024568155 | -4.349568207 | 1.48E-05 | 6.23E-06 |
| LINC00528 | 0.009887881 | 0.948387176 | 0.343761433 | 0.458696976 |
| MIR762HG | -0.154237904 | -3.930359618 | 0.000114125 | 0.000419305 |

FC, fold change; FDR, false discovery rate.
expression of LINC00528 was associated with worse overall survival in acute myeloid leukemia, stomach adenocarcinoma and uveal melanoma (logrank \( p < 0.05 \)). These results were downloaded from GEPIA. Figure 8 shows the details. The references for analysis platforms and packages were provided in Table 4.

**DISCUSSION**

The continuous advancement of molecular oncology has made it possible to identify the genetic characteristics of tumor subtypes. These genetic characteristics are often closely associated with the activity and therapeutic opportunities of oncogenic pathways, hence becoming important biomarkers and drug targets for tumors [31]. In combination with the high heterogeneity of breast cancer genomes, the identification of subtype-specific related gene markers can provide important support for clinical strategies to improve breast cancer subtypes. Currently, advanced bioinformatics tools based on sophisticated algorithms are constantly available, allowing us to identify potential driving events in diseases from massive genomic data [32–34]. Based on transcriptome data, this study identified key biomarkers closely related to the prognosis of HER2-positive breast cancer and explored the molecular pathogenesis of HER2-positive breast cancer.
The potential of lncRNA as a prognostic factor for tumors has been recognized. As a complement to mRNAs and miRNAs, lncRNA-rich transcripts provide us with important materials for obtaining tumor prognostic factors. Through reliable analysis methods such as univariate Cox, Kaplan-Meier and random forests, we eventually identified the following six stable survival-related lncRNAs as core lncRNAs in HER2-positive breast cancer: AC010595.1, AC046168.1, AC069277.1, AP000904.1, LINC00528, and MIR762HG. We associated the high expression of four core lncRNAs (AC010595.1, AC046168.1, AC069277.1, and AP000904.1) with worse overall survival, from which we inferred that they promoted the development of cancer. The low expression of LINC00528 and MIR762HG was associated with worse overall survival, suggesting that they may have been tumor suppressor genes. Each of the core lncRNAs had good predictive power for 3-year survival, with AP000904.1 (AUC = 0.914) and LINC00528 (AUC = 0.929) being optimal.

The multi-factor regression model composed of these six core lncRNAs has a non-negligible advantage. It has a very good risk assessment ability of HER2-positive breast cancer patients (p < 0.05), and a powerful prognostic ability for 3-year survival (AUC = 0.980). This provides an effective classification tool for the clinical prognosis assessment of HER2-positive breast cancer. We also intended to test the prognostic model using an independent data set in Gene Expression Omnibus. Unfortunately, we were not able to find a fully applicable data set. This is partly because there are few related studies on these six core lncRNAs. The lack of large clinical cohorts to validate the model is one of the limitations of this study. However, considering that the six core lncRNAs in this study have shown stable prognostic value for HER2-positive breast cancer in a variety of prognostic analyses, the 6-lncRNA prognostic model is relatively reliable and worthy of further study.

Currently, non-coding regions are not fully annotated and we could hardly find any reports on these six prognostic lncRNAs. Therefore, our understanding of these core lncRNAs is not sufficiently comprehensive. More researches and tests are needed for clinical transformation. Non-coding genes usually function in complex regulatory networks [35]. To explore the functional relevance of core lncRNAs in HER2-positive breast cancer, we constructed a corresponding ceRNA network. Three core lncRNAs (AC010595.1, AP000904.1, and MIR762HG) were found closely related in the ceRNA network. Their target mRNAs were significantly enriched in the biological processes of muscle organ development, regulation of vasculature development, and angiogenesis, and closely associated with focal adhesion. In consideration of the importance of angiogenesis and cell adhesion in the pathogenesis of tumors, it could be inferred that AC010595.1, AP000904.1 and MIR762HG have jointly affected the growth and metastasis of HER2-positive breast cancer through ceRNA mechanisms. Furthermore, the famous cancer-related gene FGF2 was found to be a target mRNA of the three core lncRNAs in the ceRNA network. FGF2, an important member of the tyrosine kinase receptor family, plays a key role in the development of tumors. By reviewing the other analysis results in this study, we also identified that FGF2 was a low expressed mRNA in HER2-positive breast cancer (|log2FC| = 2.60345034144707, FDR = 7.58730025761752E-75) and occupied a core position in the PPI network (degree = 58). These newly discovered relationships between FGF2 and the core lncRNAs are valuable.

In addition, the KEGG analysis also showed AP000904.1 target mRNAs directly participated in the PI3K-Akt signaling...
**TABLE 3 |** The main functional analysis results of the target mRNAs of each core lncRNA.

| ID          | Description                               | Q value   | Count |
|-------------|-------------------------------------------|-----------|-------|
| AC010595.1  | **Biology process**                       |           |       |
| GO:0007517  | Muscle organ development                   | 0.003517703 | 8     |
| GO:1901342  | Regulation of vasculature development      | 0.003517703 | 7     |
| GO:0001525  | Angiogenesis                               | 0.003517703 | 8     |
| GO:0061337  | Cardiac conduction                         | 0.003736857 | 5     |
| **Cellular component** |                           |           |       |
| GO:0005901  | Caveola                                    | 0.002572495 | 4     |
| GO:0044853  | Plasma membrane raft                       | 0.002572495 | 4     |
| GO:0045121  | Membrane raft                              | 0.002572495 | 6     |
| GO:0098857  | Membrane microdomain                       | 0.002572495 | 6     |
| GO:0098589  | Membrane region                            | 0.005744063 | 6     |
| GO:0001653  | Peptide receptor activity                  | 0.014043265 | 4     |
| **KEGG pathway** |                                    |           |       |
| hsa05205    | Proteoglycans in cancer                    | 0.021546616 | 5     |
| hsa04022    | cGMP-PKG signaling pathway                 | 0.05294302  | 4     |
| hsa04510    | Focal adhesion                             | 0.068542716 | 4     |
| AC046168.1  | **Biology process**                       |           |       |
| GO:0009913  | Epidermal cell differentiation             | 0.006405587 | 3     |
| GO:0008544  | Epidermis development                      | 0.006980753 | 3     |
| AP000904.1  | **Biology process**                       |           |       |
| GO:0007517  | Muscle organ development                   | 0.004405768 | 8     |
| GO:1901342  | Regulation of vasculature development      | 0.004405768 | 7     |
| GO:0001525  | Angiogenesis                               | 0.004405768 | 8     |
| **Cellular component** |                     |           |       |
| GO:0005901  | Caveola                                    | 0.003856935 | 4     |
| GO:0044853  | Plasma membrane raft                       | 0.003856935 | 4     |
| GO:0045121  | Membrane raft                              | 0.003856935 | 6     |
| GO:0098857  | Membrane microdomain                       | 0.003856935 | 6     |
| GO:0098589  | Membrane region                            | 0.00852471  | 6     |
| GO:0043235  | Receptor complex                           | 0.019729483 | 5     |
| GO:0001653  | Peptide receptor activity                  | 0.017088444 | 4     |
| **KEGG pathway** |                                    |           |       |
| hsa05205    | Proteoglycans in cancer                    | 0.031738163 | 5     |
| hsa04022    | cGMP-PKG signaling pathway                 | 0.071853414 | 4     |
| hsa04510    | Focal adhesion                             | 0.081765703 | 4     |
| hsa04151    | PI3K-Akt signaling pathway                 | 0.081765703 | 5     |
| hsa04014    | Ras signaling pathway                      | 0.081765703 | 4     |
| LINC00528   | **Biology process**                       |           |       |
| GO:0042110  | T cell activation                          | 7.85E-14   | 14    |
| GO:0002696  | Regulation of lymphocyte activation        | 4.69E-11   | 12    |
| GO:0050867  | Positive regulation of leukocyte activation| 5.43E-11   | 11    |
| GO:0050856  | Positive regulation of cell activation     | 5.90E-11   | 11    |
| GO:0051251  | Positive regulation of lymphocyte activation| 4.14E-10  | 10    |
| GO:0050870  | Positive regulation of T cell activation   | 4.14E-10   | 9     |
| GO:0050863  | Regulation of T cell activation            | 4.14E-10   | 10    |
| GO:0030098  | Lymphocyte differentiation                 | 4.35E-10   | 10    |
| GO:1903039  | Positive regulation of leukocyte cell-cell adhesion | 4.70E-10  | 9     |
| GO:1903037  | Regulation of leukocyte cell-cell adhesion  | 5.41E-10   | 10    |
| **Cellular component** |                           |           |       |
| GO:0008552  | Side of membrane                           | 2.59E-10   | 11    |
| GO:0042101  | T cell receptor complex                     | 4.08E-10   | 5     |
| GO:0009897  | External side of plasma membrane           | 3.02E-07   | 7     |
| GO:0008688  | Cytoplasmic side of plasma membrane        | 1.37E-05   | 5     |
| GO:0088002  | Plasma membrane receptor complex            | 1.39E-05   | 5     |
| GO:0098652  | Cytoplasmic side of membrane               | 1.64E-05   | 5     |
| GO:0043235  | Receptor complex                           | 0.000173207 | 5     |
| **KEGG pathway** |                                    |           |       |
| hsa05162    | Measles                                    | 2.59E-06   | 6     |
| hsa046340   | Primary immunodeficiency                   | 4.06E-06   | 4     |
| hsa04658    | Th1 and Th2 cell differentiation           | 4.06E-06   | 5     |

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TABLE 3  (Continued) The main functional analysis results of the target mRNAs of each core lncRNA.

| ID         | Description                          | Q value     | Count |
|------------|--------------------------------------|-------------|-------|
| hsa04640   | Hematopoietic cell lineage           | 4.06E-06    | 5     |
| hsa04660   | T cell receptor signaling pathway    | 4.06E-06    | 5     |
| MIR762HG   | Biology process                      |             |       |
| GO:0007517 | Muscle organ development             |             |       |
| GO:1901342 | Regulation of vasculature development| 0.004744758 | 10    |
| GO:0001667 | Ameboidal-type cell migration        |             |       |
| GO:0045765 | Regulation of angiogenesis           |             |       |
| GO:0001525 | Angiogenesis                         | 0.008004132 | 10    |
| GO:0061337 | Cardiac conduction                   |             |       |
| GO:0005925 | Focal adhesion                       | 0.010229707 | 8     |
| GO:0005924 | Cell-substrate adherens junction     |             |       |
| GO:0005901 | Caveola                              |             |       |
| GO:0000655 | Cell-substrate junction              |             |       |
| GO:0044853 | Plasma membrane raft                 |             |       |
| GO:0045121 | Membrane raft                        |             |       |
| GO:0098857 | Membrane microdomain                 |             |       |
| GO:0005911 | Cell-cell junction                   |             |       |
| GO:0043255 | Ion channel binding                  |             |       |
| GO:0005539 | Glycosaminoglycan binding            |             |       |
| GO:0001653 | Peptide receptor activity            |             |       |
| GO:0016247 | Channel regulator activity           |             |       |
| KEGG pathway | Proteoglycans in cancer            |             |       |
| hsa005205  | cGMP-PKG signaling pathway           |             |       |

KEGG, Kyoto Encyclopedia of Genes and Genomes.

FIGURE 8  Kaplan-Meier curves of the core lncRNA LINC00528 in various cancers based on GEPIA. The Kaplan-Meier curves show the prognostic significance of LINC00528 in a variety of cancers according to the survival analysis based on GEPIA. The red lines represent patients with high gene expression, and blue lines represent patients with a low gene expression. (A) BRCA: breast invasive carcinoma; (B) CECS: cervical squamous cell carcinoma and endocervical adenocarcinoma; (C) HNSC: head and neck squamous cell carcinoma; (D) AML: acute myeloid leukemia; (E) LUAD: lung adenocarcinoma; (F) SKCM: skin cutaneous melanoma; (G) STAD: stomach adenocarcinoma; (H) UVM: uveal melanoma. Abbreviations: GEPIA, Gene Expression Profiling Interactive Analysis.
pathway as well as the Ras signaling pathway. It is well known that these two pathways are the classical signal transduction pathways activated mainly by HER-2 and other members of the HER/ErBb receptor family after they form heterodimers. They can form cross-talk with other signaling pathways, and participate in biological functions including cell proliferation, migration, infiltration, anti-apoptosis and promotion of angiogenesis [36]. AP000904.1 is likely to be a key regulator for HER2 overexpressing tumors that deserves more attention.

The functional enrichment analysis showed that target mRNAs of LINC00528 were closely associated with immune activity and they participated in biology processes activated by various immune cells such as T cells, lymphocytes and leukocytes. LINC00528 was especially enriched in the biology processes of positive regulation of leukocyte cell-cell adhesion and regulation of leukocyte cell-cell adhesion (including multiple molecules of the CD family). This suggests that LINC00528 is closely associated with tumor cell metastasis. Based on the results of prognostic analysis and clinical characterization, we hypothesized that the low expression level of LINC00528 at the late stage is likely to be an important factor for the late metastasis of HER2-positive breast cancer. Survival analysis based on GEPIA also showed that LINC00528 had a good prognostic value in various tumors (p < 0.05), which suggested that LINC00528 could play an important role in the mechanism of cancer metastasis.

Functional analysis of differentially expressed mRNAs has also provided us with some enlightening insights. Several differentially expressed mRNAs in HER2-positive breast cancer (including CXCL2, CXCL12, CXCL10, CXCL11, CXCL9 and CXCL14) were enriched in the biology processes of chemokine receptor binding and chemokine activity. The result provides a molecular basis for elucidating the metastatic pattern of HER2-positive breast cancer. Interestingly, the differentially expressed mRNAs were significantly enriched in the malaria KEGG pathway (FDR = 0.0149807601977498). Previous studies have shown that anti-malarial drugs can cause cell cycle arrest and apoptosis in HER2-enriched breast cancer cells [37]. Furthermore, the PPI network showed modules which were significant for the exploration of the biological behavior of HER2-positive breast cancer. Module 1 was the most significant and primarily involved in mitosis, which is consistent with the characteristics of HER2 overexpression. Module 2 showed the immune regulation in HER2-positive breast cancer. Notably, Module 3 participated in the Adenosine 5′-monophosphate-activated protein kinase (AMPK) signaling pathway. AMPK as a regulator of cellular energy metabolism is an ideal target for tumor therapy [38, 39].

CONCLUSION

This study provides a new efficient prognostic model of HER2-positive breast cancer. The 6-lncRNA prognostic model is relatively reliable and worthy of further study. The six core lncRNAs are also potential drug targets. Currently there are almost no reports regarding the six core lncRNAs. This study has thus laid a foundation for a further understanding of the roles of these core lncRNAs and for obtaining the key pathways. Meanwhile, it provides a new perspective for elucidating the molecular mechanisms underlying HER2-positive breast cancer.

DATA AVAILABILITY STATEMENT

Publicly available datasets were analyzed in this study. This data can be found here: https://portal.gdc.cancer.gov/. Code availability: We conducted data analysis in R (3.4.0) language environment. The R packages used are available on the website: https://mirrors.tuna.tsinghua.edu.cn/CRAN/. We also applied GEPIA (http://gepia.cancer-pku.cn/) for further analysis, which has been explained appropriately in the paper.

AUTHOR CONTRIBUTIONS

XZ and JW were involved in the concept and design of the study. XZ drafted the manuscript. All authors were involved in acquisition, analysis, and interpretation of the data, revised the manuscript, and approved the final version.

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CONFLICT OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.por-journal.com/articles/10.3389/pore.2021.609083/full#supplementary-material.
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