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Ontogenetic and Among-Individual Variation in Foraging Strategies of Northeast Pacific White Sharks Based on Stable Isotope Analysis

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Abstract

There is growing evidence for individuality in dietary preferences and foraging behaviors within populations of various species. This is especially important for apex predators, since they can potentially have wide dietary niches and a large impact on trophic dynamics within ecosystems. We evaluate the diet of an apex predator, the white shark (Carcharodon carcharias), by measuring the stable carbon and nitrogen isotope composition of vertebral growth bands to create lifetime records for 15 individuals from California. Isotopic variations in white shark diets can reflect within-region differences among prey (most importantly related to trophic level), as well as differences in baseline values among the regions in which sharks forage, and both prey and habitat preferences may shift with age. The magnitude of isotopic variation among sharks in our study (>5% for both elements) is too great to be explained solely by geographic differences, and so must reflect differences in prey choice that may vary with sex, size, age and location. Ontogenetic patterns in δ¹³C values vary considerably among individuals, and one third of the population fit each of these descriptions: 1) δ¹³C values increased throughout life, 2) δ¹³C values increased to a plateau at ~5 years of age, and 3) δ¹³C values remained roughly constant values throughout life. Isotopic data for the population span more than one trophic level, and we offer a qualitative evaluation of diet using shark-specific collagen discrimination factors estimated from a 3+ year captive feeding experiment (Δ¹³Cshark-diet and Δ¹⁵Nshark-diet equal 4.2% and 2.5%, respectively). We assess the degree of individuality with a proportional similarity index that distinguishes specialists and generalists. The isotopic variance is partitioned among differences between-individual (48%), within-individuals (40%), and by calendar year of sub-adulthood (12%). Our data reveal substantial ontogenetic and individual dietary variation within a white shark population.

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Introduction

Although diet is often treated as a species-level trait, variation in diet composition and foraging behavior occurs within most species. This variation can be attributed to at least three factors–habitat-specific variation in prey availability; differences in the cost-benefit ratios of potential prey among the sexes, or age- or size-classes of consumers; and phenotypic variation among what often appear to be otherwise similar individuals [1–5]. Collectively, this dietary variation influences the fitness of consumers and their ecological and evolutionary impacts on prey species, communities, and ecosystems [1–5].

White sharks (Carcharodon carcharias) are apex predators that can have cascading effects on marine ecosystems [6,7], but our understanding of their foraging ecology is fragmentary and often biased by spectacular accounts, especially attacks on humans and other large mammals. In the northeastern Pacific Ocean, white sharks were once considered a nearshore species that preyed primarily on pinnipeds, a perception arising from many studies focused on coastal sites near pinniped colonies where shark foraging behavior was easy to observe [8–13]. This view has been challenged by recent satellite tagging data from white sharks off the coast of California and Baja California, Mexico, which revealed migration between the North American continental shelf and two offshore areas [14 to 26° N and 125 to 140° W] [14–17]. Isotopic data from tagged individuals corroborated offshore foraging on lower trophic level prey and indicated similar dietary preferences within this population [18]. Although observations of white shark predation on non-pinniped prey are rare, stomach contents include remains from invertebrates, turtles, fish, and sharks [19]. Here, we assess population-level diet variation, potential ontogenetic shifts in prey preferences, and individual diet specialization through analysis of carbon and nitrogen isotope variation.

The stable isotope composition of a tissue reflects a temporal integration of dietary and environmental inputs (albeit mediated by animal physiology), and can thus be used as a natural tracer for foraging variation. The most commonly used stable isotope ratios
in dietary studies are $^{13}$C/$^{12}$C and $^{15}$N/$^{14}$N. Physiological processes lead to a subtle increase in $^{13}$C- and $^{15}$N-concentration with trophic level in consumers (~1% and ~3% per trophic step, respectively) [20]. In addition, there are $^{13}$C/$^{12}$C and $^{15}$N/$^{14}$N gradients throughout the eastern basin of the Pacific Ocean controlled by a variety of factors related to primary production at the base of the food web [21–23]. For carbon, upwelling brings $^{13}$C-depleted deep water to the surface and nutrient-driven phytoplankton blooms off the California coast increase the baseline carbon isotope values ~2–3% relative to less productive offshore areas [21–23]. The spatial nitrogen isotope gradient has a similar range, but is more complex as the source of nutrients (i.e., nitrate, ammonium, $N_2$) and processes of biological incorporation or geochemical cycling (i.e., $N_2$ fixation, nitrate or ammonium assimilation, or denitrification) vary with productivity regimes [23–27]. These baseline trends are conserved in consumer tissues; for example, pinnipeds foraging in productive $^{13}$C-enriched nearshore habitats on the Pacific Rim have carbon isotope values ~2% greater than those foraging offshore [20].

Assessments of consumer diet with stable isotope analysis must address these spatial and trophic variations among potential prey. This technique has been used to chronicle patterns of diet and habitat use in a variety of marine predators, including marine mammals (see review by [29]), marine turtles [30–32], and sharks [33]. Two issues that are important to disentangling the effects of spatial versus trophic variations on consumer isotope values are biological parameters (i.e., discrimination factors and turnover rates) and distinct prey isotope values. In complex systems, however, these sources of variation are difficult to tease apart. The few isotopic studies featuring sharks often calculate trophic position and report isotopic differences among tissues [33–39], but other ecological insights, such as dietary preferences and habitat use, are limited.

The concept of ecological niche [40] was adopted in isotopic ecology because stable isotope values vary with aspects of a consumer’s diet and habitat through time [41]. Although ecological and isotopic niches cannot be directly translated, isotopic variation within and among individual consumers of a particular species or population reflects the niche width [41–43]. Data from multiple individuals can yield an estimate of population-level isotopic niche width, but to capture within-individual dietary variation, multiple measurements of an individual’s diet and habitat preferences over time are required. Serially-sampled accretionary structures (i.e., feathers, baleen, vibrissae, turtle scutes, shark vertebrae, etc.) produce ontogenetic time series that can be used to establish within-individual isotopic niche width [32,36,43–46]. These patterns can be compared among individuals to identify generalists with overlapping isotopic values or specialists that occupy just part of the population’s isotopic range [43]. Although bony fish are featured in studies of individuality, we are aware of only one other study featuring elasmobranchs (shark, skates, and rays) despite their considerable diversity, wide distribution, and functional importance in marine and estuarine systems [2,33].

We investigated isotopic time series from the vertebrae of white sharks collected from the northeast Pacific Ocean between 1957 and 2000. Because white shark vertebrae grow by accretion, isotopic values from growth bands record lifetime variations in an individual’s diet. We hypothesized $^{15}$N values to increase with age to reflect a shift to high trophic-level prey. Furthermore, a quantitative analysis of carbon and nitrogen isotopic data will reveal the degree of dietary variation within and among individuals in this white shark population.

### Methods

(a) Collection and Preparation of Vertebræ for Isotopic Analysis

The vertebrae of 15 adult white sharks were sampled from existing collections (Table 1). Fourteen specimens were caught off the California coast (locations are listed in Table 1, if available). One specimen (42094–2) was caught offshore from Baja California, Mexico, and may be part of the population that aggregates near Guadalupe Island [17]. Because the isotopic pattern from this individual was similar to that for some of the sharks caught off the California coast, we included data from this shark in our analyses.

All of the vertebrae were stored frozen or dry, with two exceptions (27015 and 27601), which were preserved in alcohol until sampling (Table 1). While some prior studies have found greater isotopic variability and a shift to lower C:N atomic values for ethanol-preserved muscle [47,48], we found no differences in isotopic or C:N atomic variability between frozen and ethanol preserved vertebrae, and so we included data from ethanol preserved specimens in our analyses.

White shark vertebrae grow by accretion in concentric rings and have annual growth bands that are used to age individuals [49]. We followed sampling and preparation techniques in Kim and Koch [40] for these vertebrae. Briefly, a 1-mm thick section was cut from the parasagittal plane with a diamond saw. Then, sections were polished and annual rings, defined as one opaque and one translucent growth band pair, were independently counted and marked by SLK and a researcher who conducts elasmobranch age and growth studies at California State University, Moss Landing Marine Lab (MLML). Individual growth rings were sampled from the corpus calcærum using a New Wave MicroMill (MLML and the Marine Analytical Lab, University of California, Santa Cruz [UCSC]) from a maximum depth of 1.2 mm (File S1). Samples were decalcified using 0.5 M EDTA (pH 8), rinsed 10 times with milliQ water to isolate collagen, and freeze-dried for stable isotope analysis (method modified from [40]). Samples were weighed to 300–400 µg into tin boats (3×5 mm, Costech) and analyzed at the Stable Isotope Laboratory at UCSC on an elemental analyzer coupled to an isotope ratio-monitoring mass spectrometer (Thermo-Scientific Delta-XP IR-MS). Isotope ratios are expressed in δ values, where:

$$\delta X = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1000$$

X is the element of interest, h is the high mass number, and R is the high mass-to-low mass ratio. Units are parts per thousand (per mil, ‰) deviations from a standard. The δ$^{13}$C and δ$^{15}$N values were referenced to V-PDB and AIR, respectively. Replicates of a gelatin standard within each analysis session allowed for mass and drift corrections. Comparisons of this standard within and between runs yielded SD of <0.2% and <0.3% for δ$^{13}$C and δ$^{15}$N values (n = 77), respectively.

Because of the temporal span of our data, we corrected for anthropogenic changes in the δ$^{13}$C values of Earth surface carbon reservoirs that occurred over the past 70 years [50]. As δ$^{15}$C values did not change constantly through this period, two atmosphere-derived rates (0.05% per year for 1937–1960 and 0.022% per year for 1961–2000) were used to correct samples to 2000, the year of death for the most recent individuals [51].
or mass based on von Bertalanffy growth functions (VBGF, [54]): use annual vertebral growth bands [49] and relate them to length approximately 300–350 cm long [16,52]. Additionally, white rookeries. The smaller white sharks observed at An˜o Neuvo Island, from observation and tagging studies of white sharks near pinniped variance (ANOVA). These age classes were based on length data values varied significantly with age using a one-way analysis of variation. For each shark, we fit three alternative functions to the time series data: i) a constant increase in $L(t) = L_\infty - (L_\infty - L_0)e^{-kt}$ (2)

where $t$ is time, $L_\infty$ is the maximum (or asymptotic) length, $L_0$ is length at birth ($t=0$), and $k$ is an empirically derived growth constant. According to VBGF, 300 cm corresponds to white sharks that are 5 years old [55,56]. Based on tooth morphology and white shark presence at pinniped rookeries, we classified isotopic data from growth bands corresponding to birth through the end of the 5th year as “young” and to ≥6 years old as “sub-adult to adult.”

We next evaluated individual differences in ontogenetic dietary variation. For each shark, we fit three alternative functions to the time series data: i) a constant $\delta^{15}N$ value (assuming random variation but no prevailing trend with age); ii) a continuous increase in $\delta^{15}N$ values with age, fit using a first-order polynomial; and iii) a non-linear, asymptotic increase in $\delta^{15}N$ values with age, fit using VBGF (Equation 2, substituting $\delta^{15}N$ values for length). We removed statistically significant outlier data (studentized residuals that fall outside 95% prediction interval for new data points) prior to final fitting. We tested the statistical significance of each function at $\alpha = 0.05$, and used degrees-of-freedom-adjusted R$^2$ values to evaluate goodness of fit for each model. We then assigned individuals to one of three classes based on which of these models (no ontogenetic shift, linear increase or asymptotic increase) was significant and provided the best fit. For individuals exhibiting an asymptotic increase, we designated the age of transition from young to sub-adult through adult diets as the point at which the VBGF function equaled 90% of the asymptotic value. For individuals exhibiting no significant ontogenetic shift and having high $\delta^{15}N$ values in early growth bands, we sampled and analyzed portions of the central vertebra, which form from maternal resources before parturition, to assess the potential of metabolic turnover within the vertebra.

### Table 1. Summary of biological and collection data and proportional similarity index ($w_j$) for white sharks.

| ID     | Year of death | Sex | # Growth bands | Location caught         | Collection of acquisition | Mean adult | $\delta^{13}C$ value, % (SD) | $\delta^{15}N$ value, % (SD) | $w_j$ | Method of storage |
|--------|---------------|-----|---------------|--------------------------|---------------------------|------------|------------------------------|------------------------------|------|-------------------|
| 26245  | 1957          | F   | 20            | Monterey Bay, CA         | CAS                       | −13.3 (0.6) | 20.2 (0.3)                   | 0.17                         | Dry  |                   |
| 26678  | 1959          | ?   | 11            | Selva Beach, CA          | CAS                       | −11.4 (0.2) | 18.8 (1.0)                   | 0.23                         | Dry  |                   |
| 27015  | 1960          | ?   | 9             | Stinson Beach, CA        | CAS                       | −12.0 (0.2) | 18.1 (0.4)                   | 0.25                         | Ethanol |                 |
| 26781  | 1960          | ?   | 9             | Tomales Bay, CA          | CAS                       | −11.6 (0.3) | 18.9 (0.5)                   | 0.24                         | Ethanol |                 |
| 42994-2| 1965          | ?   | 7             | Eastern Pacific, Baja Mexico | LACM                    | −14.1 (0.6) | 16.7 (0.3)                   | 0.33                         | Dry  |                   |
| WS CM  | 1976          | ?   | 10            | Point Reyes, CA          | GC                        | −13.6 (0.6) | 17.6 (0.7)                   | 0.50                         | Dry  |                   |
| WH 17  | 1980          | ?   | 14            | California               | MLML                      | −13.8 (0.8) | 19.0 (0.6)                   | 0.73                         | Frozen |                 |
| WS 21  | 1983          | M   | 16            | Anacapa Island, CA       | MLML                      | −14.9 (0.2) | 16.0 (0.4)                   | 0.26                         | Frozen |                 |
| WS 100 | 1985          | ?   | 19            | Santa Barbara, CA        | MLML                      | −12.6 (0.6) | 18.1 (0.8)                   | 0.90                         | Frozen |                 |
| WS KG  | 1986          | ?   | 13            | SE Farallones, CA        | KG                        | −13.0 (0.5) | 19.6 (0.2)                   | 0.59                         | Dry  |                   |
| WS 101 | 1991          | ?   | 11            | CA                       | MLML                      | −13.4 (0.2) | 16.6 (0.4)                   | 0.74                         | Frozen |                 |
| WS 128 | 1992          | ?   | 17            | CA                       | MLML                      | −13.3 (0.8) | 18.4 (0.9)                   | 0.42                         | Frozen |                 |
| 42898 PR| 1998          | ?   | 15            | Point Reyes, CA          | SA                        | −12.4 (0.5) | 18.5 (0.3)                   | 0.08                         | Dry  |                   |
| 56731-1| 2000          | ?   | 17            | Catalina Island, CA      | LACM                      | −12.3 (0.5) | 19.0 (0.8)                   | 0.73                         | Dry  |                   |
| CC3    | 2000          | F   | 14            | Morro Bay, CA            | LML                       | −12.0 (0.2) | 19.8 (0.5)                   | 0.33                         | Frozen |                 |

Abbreviations are as follows: California Academy of Sciences (CAS), Natural History Museum of Los Angeles County (LACM), G. Chan (GC), Moss Landing Marine Lab (MLML), K. Goldman (KG), S. Anderson (SA), and Long Marine Lab (LML).

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### (b) Quantitative Analysis of Ontogeny

To test for the existence of an ontogenetic shift at the population level in a generalized linear model, we established 2 age classes and had individuals as a source of variation to determine if $\delta^{15}N$ values varied significantly with age using a one-way analysis of variance (ANOVA). These age classes were based on length data from observation and tagging studies of white sharks near pinniped rookeries. The smaller white sharks observed at An˜o Neuvo Island, Southeast Farallon Islands, Tomales Bay, and Point Reyes are approximately 300–350 cm long [16,32]. Additionally, white shark teeth develop finer serrations beginning at 300 cm, indicating a functional shift [53]. Age and growth studies often use annual vertebral growth bands [49] and relate them to length or mass based on von Bertalanffy growth functions (VBGF, [34]):

(c) Potential Prey for Qualitative Assessment of White Shark Diet

We compiled isotopic data for potential prey gleaned from studies in northeastern Pacific ecosystems to provide a dietary context for our sub-adult to adult isotopic data from white sharks. We limited our assessment to qualitative patterns, rather than estimating specific proportions of dietary inputs, because the large number of potential prey and their variance would likely produce indeterminate results [57]. The potential prey we included were as follows: California sea lion [28,40,58]; harbor seal [28,41]; northern elephant seal [28,41–43]; harbor porpoise [32,36,43–45,59]; dolphins (Berman and Newsome, unpublished data); various tuna species [32,36,43–46,60]; neritic fish [43,61]; blue and hammerhead sharks [62]; and cephalopod species [63,64]. The localities and isotopic values for all prey are compiled in File S2. All prey isotopic values were “corrected” (and error propagated) to resemble collagen for comparison.

While our shark data span from 1950–2000, the prey data are relatively recent and thus may not reflect secular shifts in baseline
isotope values due to climate change or changes in productivity. This source of variation is difficult to constrain, but there is little evidence for major secular shifts (i.e., >1 or 2‰), and variation of this magnitude will not affect our qualitative assessment of diet.

(d) Assessing the Degree of Individuality

To evaluate niche overlap, we used Pianka's measure [65] under multivariate normality [66], which is as follows:

$$\bar{w}_y = \frac{g(s)g(x)dx}{\sqrt{g^2(s)dx} \sqrt{g^2(x)dx}} \cdot (3)$$

Equation 3 allows for the calculation of proportional overlap (in 2-dimensional niche space) between two individuals or populations (i and j), accounting for multivariate covariance and density distributions [g(s)]. We measured the degree of isotopic niche overlap, or "proportional similarity," between each individual's sub-adult/adult isotopic range and the population-level isotopic range (i.e., all sub-adult/adult growth bands for 15 individuals).

Averaged across individuals, this proportional similarity index [66], Equation 3; [66]) allowed us to examine the degree of individual specialization; individual specialists would be expected to have a low degree of overlap (\(w_y << 1\)), while generalists would have extensive overlap (\(w_y \approx 1\)).

We also assessed the effects of individual differences on sub-adult/adult isotopic values within a generalized linear model. This analysis allowed us to assess the relative amount of variance explained by differences among individuals vs. variation within individuals. We included a temporal category, "calendar year of adulthood," defined as the calendar year a shark reached 6 years of age. The three categories for calendar year of sub-adulthood were based on pinniped populations: before the passage of the Marine Mammal Protection Act (pre-1972), during the period when pinniped populations were increasing (1972–1986), and after pinniped populations doubled from pre-1972 counts (post-1986) [67,68]. Other biological details (i.e., sex, location caught, etc.) were not included in our analysis because the information was not available for all specimens (Table 1). The best-fit models for \(\delta^{13}C\) and \(\delta^{15}N\) values (weighted equally and independently) were selected based on minimal Akaike Information Criterion (AIC) values [69]. All statistical analysis was performed in MatLab (version 8.0).

(e) Discrimination Factors

There are offsets between prey and consumer \(\delta^{13}C\) and \(\delta^{15}N\) values, known as trophic discrimination factors, which reflect preferential sorting during metabolism and incorporation into tissues [20,70,71]. To compare potential prey and consumer isotopic values, discrimination factors defined as:

$$\Delta^\delta X = \delta^\delta X_{consumer} - \delta^\delta X_{prey} \cdot (4)$$

must be applied to account for trophic enrichment of \(^{13}\)C and \(^{15}\)N. The average carbon and nitrogen discrimination factors that are widely used are 0.4‰ (SD = 1.3‰) and 3.4‰ (SD = 1.0‰), respectively [72], but actual values vary with diet, physiology, and tissue [22,73,74].

We conducted a controlled feeding study with captive leopard sharks (Triakis semifasciata) fed squid over 1250 days [62]. The care and protocol for euthanizing the leopard sharks were approved by the UCSC Chancellor’s Animal Research Committee (permit code: Koch 0901) and were in accordance with Institutional Animal Care and Use Committee (IACUC) standards. Briefly, the leopard sharks (n = 3) were caught in the San Francisco Bay from August 2003 to January 2006 and maintained at Long Marine Lab, UCSC in polyethylene tanks (2.3 m diameter, 1.2 m water depth) with a continuous flow of filtered seawater from the Monterey Bay (temperature range: 13°–17°C; salinity range: 30–34) until July 2009. The sharks were sacrificed using a lethal dose of tricaine methanesulfonate (MS-222) and vertebrae were extracted and frozen at −20°C. A pair of adjacent vertebrae from the anterior column were cleaned and selected for analysis. For each pair of vertebrae, one vertebra was thin-sectioned to measure growth bands (following sectioning methods of [75] and adapted by [76]) and growth bands in the other vertebra were drilled and collagen prepared for stable isotope analysis. Growth bands were measured 3 times from each shark’s vertebrae non-consecutively using a microscope and transmitted light. The outermost bands without statistically different isotopic values were averaged as the \(\delta^\delta X_{consumer}\) value in Equation (4).

Results

(a) White Sharks

A comparison of young and subadult to adult white sharks, blocked across individuals, demonstrated a significant ontogenetic shift (F1, 206 = 23.19, p < 0.0001, r^2 = 0.69), confirming that there is an ontogenetic shift in dietary preferences or habitat use in the northeastern Pacific white shark population. Five individuals showed a non-linear, asymptotic increase in \(\delta^{15}N\) values, with the transition to the sub-adult to adult diet occurring at approximately 4 years of age (Figure 1A). Five individuals exhibited a linear relationship between \(\delta^{15}N\) and age, with a mean increase of 0.127 yr^-1 ± 0.073 (p < 0.0001; Figure 1B). The remaining 5 individuals showed no significant relationship between age and \(\delta^{15}N\) values (Figure 1C). For individuals with high \(\delta^{15}N\) values (>17.0‰) before age 6 and linearly increasing or no ontogenetic shift (i.e., 26678, 56731–1, CC3, WS 100, WS CM), the average \(\delta^{15}N\) values in the central vertebra, which are formed prior to parturition, were 0.6–3.4‰ less than the growth bands for ages 1–5 (Figure 1). A comparison of individual age vs. \(\delta^{13}C\) values did not reveal significant patterns (File S3). These three patterns of individual variation in \(\delta^{15}N\) values within the population are robust and point to substantial differences in the ontogeny of foraging behavior among individuals.

A bivariate plot of isotopic data from sub-adult/adult growth bands illustrates the dietary diversity within the northeastern Pacific white shark population (Figure 2). The population-level \(\delta^{13}C\) and \(\delta^{15}N\) values range from −14.5 to −11.7‰ and 17 to 21‰, respectively. Within this range, certain individuals (e.g., WS CM, WS 100, WS 128) exhibit isotopic ratios consistent with a diet rich in lower trophic level prey, whereas other individuals (e.g., 26245, WS KG, CC3) appear to consume primarily high trophic level prey. One individual, WS 21, was an outlier from the population pattern, with low \(\delta^{13}C\) and \(\delta^{15}N\) values.

Our data reveal that northeast Pacific white sharks occupy a wide isotopic niche, as expected for a generalist population. However, closer inspection reveals a range of strategies among individuals, which may relate to sex, location, size, or individuality, as illustrated by the size and placement of individual bivariate confidence ellipses relative to the population (Figure 3A). The sharks in the sample population varied considerably in terms of isotopic overlap, with most sharks having low degrees of overlap (\(w_y < 0.5\)), but a few having high values (\(w_y < 0.8\)) suggesting more generalized diets (Figure 3B).
Figure 1. $\delta^{15}N$ values versus growth increment number (age) for 15 white sharks. A) Individuals modeled with a VBGF curve. B) Individuals showing a significant linearly increasing trend. C) Individuals showing no significant pattern. Average pre-parturition $\delta^{15}N$ values ($n = 3$) are indicated as grey filled circles for individuals with relatively high juvenile $\delta^{15}N$ values (>17‰).

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The modal range of the \( w_y \) value was 0.23–0.33 for this population (Table 1). The generalized linear model of \( d_{13}C \) and \( d_{15}N \) data showed significant effects of individual variation (\( F_{12,192} = 22.76, p < 0.0001 \)). The combined variance in \( d_{13}C \) and \( d_{15}N \) values was explained largely by differences among individuals (48%) and within-individual effects (40%). The calendar year of sub-adulthood accounted for 12% of the variance in a model with both isotope values weighted equally. Post hoc comparisons using the Tukey HSD test indicated the significance of all pairwise differences between calendar year of sub-adulthood categories for both isotopes (all \( p \)-values < 0.0001 except between pre-1972 and post-1986 \( d_{13}C \) means \( p = 0.018 \)).

(b) Discrimination Factors for Vertebrae

Because dietary carbon and nitrogen incorporate relatively slowly into shark tissues [77], discrimination factors were based on the average isotopic values from the last 1–1.5 years of the experimental sharks (outermost 12 mm). The average width for the last 6 bands (representing the final 3 years for the shark) differed among individuals, but the total thickness for the final 3 years ranged from 15.3 to 18.6 mm (Table 2). Average \( d_{13}C \) and \( d_{15}N \) values from the outermost 12 mm did not differ significantly among sharks (Kruskal-Wallis Test, \( d_{13}C \) values: \( H = 3.51, 2 \) d.f., \( p = 0.17 \) and \( d_{15}N \) values: \( H = 1.32, 2 \) d.f., \( p = 0.52 \); Table 2). The average \( d_{13}C \) and \( d_{15}N \) values (\( n = 6, SD \)) near the birthmark, where the corpus calcerum changes angle, was \(-15.4\%\) (0.3) and \(18.5\%\) (0.5), respectively, which is significantly different from the outermost bands that represent body tissues in steady state with the captive squid diet (Kruskal-Wallis Test, \( H = 12.5, 1 \) d.f., \( p = 0.0004 \)). Based on the average isotopic value of the sharks’ diet (Table 2; [62]), the vertebral collagen \( d_{13}C \) and \( d_{15}N \) values (SD) are \(4.2\%\) (0.7) and \(2.5\%\) (1.1), respectively.

Discussion

Isotopic analysis of white shark dietary patterns reveals ontogenetic and among-individual variation. This finding contrasts with previous dietary assessments based on coastal observations and stable isotope data, which suggest that nearshore pinnipeds were the preferred prey for this population [18,19,35,46].

(a) Ontogenetic Patterns

Previous studies [16,36,52,53] have suggested a shift from low to high trophic level prey with age in white sharks, and the overall increase in \( d_{15}N \) values with age reported here is consistent with this scenario. We expected the time series from all sharks to exhibit a trend of early increase in \( d_{15}N \) values, followed by a plateau once individuals had switched to a high-trophic level adult diet. While this pattern was evident for some individuals, it was not the dominant trend in our sample (Figure 1). The variation in ontogenetic patterns cannot be explained by long-term environmental changes, as sharks that exhibited the asymptotic increase in \( d_{15}N \) values and those that did not spanned the temporal range of our study.

Figure 2. Carbon and nitrogen isotope values from sub-adult to adult growth bands (\( \geq \)6 years old). The colored symbols are from white sharks; open symbols represent years \( \leq 1986 \) and closed symbols represent years \( >1986 \). Isotopic values for potential prey data are the grey boxes and are as follows: 1) northern elephant seal, 2) California sea lion, 3) harbor seal, 4) dolphin, 5) harbor porpoise, 6) tuna, 7) neritic fish, 8) offshore cephalopod, 9) nearshore cephalopod, 10) blue shark, 11) hammerhead shark. The mean prey isotope values were corrected for trophic enrichment (\( \Delta^{13}C = 4.2\% \) and \( \Delta^{15}N = 2.5\% \)) and collagen-to-muscle (\( \Delta^{13}C = 2.0\% \) and \( \Delta^{15}N = 0\% \)), if necessary (prey data and citations are listed in File S2). doi:10.1371/journal.pone.0045068.g002
The lack of a rise in $\delta^{15}$N values in some individuals is due to high values in years 1–5. Below, we discuss three possible explanations for these high values.

1. If young sharks scavenged carcasses of pinnipeds or large squid [78], they would have high $\delta^{15}$N values. Because feeding observations of juvenile white sharks are rare, this hypothesis is untested.

2. A residual signal from maternally-derived nutrients may label these early growth increments because of long incorporation rates [77]. We consider this unlikely, as rapid juvenile growth [56,79] likely erases the isotopic signal from maternal resources beyond the first growth increment.

3. A small but significant amount of metabolic turnover within vertebral centra could label growth bands 1–5 with material that reflects the high trophic level diets of adults. This explanation would require near complete turnover of collagen in grown increments 1–5, which seem unlikely given the densely mineralized acellular cartilage in shark vertebrae, but remodeling could occur during sustained swimming, as evidenced in bony fish [80,81].

(b) Vertebrae Discrimination Factors

The trophic enrichment used for white shark prey comparisons was based on the leopard shark discrimination factors (Δ$^{13}$C and Δ$^{15}$N values equal 4.2‰ and 2.5‰, respectively). These shark-specific collagen discrimination factors are greater than other tissues (i.e., blood or muscle) [22], but similar to other collagen discrimination factors [82,83]. Because collagen has a high glycine content, which is relatively $^{13}$C-enriched compared to other amino acids [94], its $\delta^{13}$C values are greater than muscle. In contrast to our study, previously published shark vertebrae discrimination factors were obtained from relatively short growth periods (<200 days) and collagen was not isolated within the calcified vertebral tissue [85,86]. Because organic components within a tissue can have different isotopic values and vary between individuals and species, it is important to isolate and compare similar substrates, when possible. Furthermore, the prey isotope values of these previous studies are confounding factors, as Hussey et al. [85] estimated prey values from weight-based feeding logs and Malpica-Cruz et al. [86] fed sharks a pellet diet that was low in protein relative to natural diets, two important factors in determining discrimination factors [74,87,88].

(c) Assessment of Sub-adult to Adult White Shark Diet

Isotopic results from white shark vertebrae indicate a diverse diet and support their classification as a generalist population (Figure 2). Representing the entire dietary range of white shark prey species and localities is not feasible. Therefore we present data on a subset of common potential prey identified from stomach content studies that span a diverse range of trophic levels and habitats (Figure 2). One potential prey group we omitted was large whales because white sharks selectively consume their blubber [11] and collagen is primarily routed from dietary protein [89,90].

Although most white sharks exhibit a range of intermediate isotopic values, which is consistent with previous isotopic evidence for both nearshore and offshore foraging [18], strategies may vary among individuals. In combination, isotopic values, ontogenetic patterns and $x_0$ values can indicate the extent of pinniped consumption. For example, individuals with intermediate isotope values, ontogenetic shift to higher trophic level, and a high degree of specialization (27015, 26781, and WS KG) are likely foraging on pinnipeds when near shore. However, intermediate isotopic values suggest that there are some offshore inputs (with lower $\delta^{13}$C and $\delta^{15}$N values). Other individuals (i.e., WH 17, WS 100, WS 128, 56731–1) that also have intermediate isotope values, but with little to no ontogenetic shift and a low degree of specialization, are likely opportunistic, non-specialized foragers. One distinct outlier among the sharks in our population is WS 21, which had low $\delta^{13}$C and $\delta^{15}$N values throughout its lifetime (Figure 2). These isotopic values suggest this individual did not consume marine mammals and its foraging ecology likely diverged from the well-studied California and Baja populations [14–18].

These isotopic results demonstrate the broad dietary range of white sharks, but caution should be taken when attempting to determine prey more specifically. The prey isotope values (after trophic enrichment correction) mostly encompass the white shark data, but overlap of a consumer’s $\delta^{13}$C and $\delta^{15}$N values with a prey could also result from integration across several outlying prey taxa. For example, the isotopic values for 26781 overlap with dolphin and harbor porpoise (preys 4 and 5, respectively, in Figure 2) whereas CC3’s values overlap with California sea lion, dolphin, and harbor porpoise (preys 2, 4, and 5, respectively, in Figure 2). However, it is likely that the isotopic mixing space for these sharks also included the following outlying prey: northern elephant seals, harbor seals, tuna, off-, and nearshore cephalopods (preys 1, 2, 6, 8, and 9 respectively, in Figure 2). Overall, the isotopic data for 26781 and CC3 suggest marine mammals were

| Individual | Average width of last 6 growth bands ± SD (n), mm | Average $\delta^{13}$C value ± SD (n), ‰ | Average $\delta^{15}$N value ± SD (n), ‰ |
|------------|----------------------------------|----------------------------------|----------------------------------|
| Diet       | 3.10±0.38 (18)                   | $-18.5\pm0.3$ (43)               | $13.3\pm0.7$ (43)               |
| CS         | 2.54±0.51 (18)                   | $-14.4\pm0.5$ (8)                | $15.9\pm0.8$ (8)                |
| FS         | 2.99±0.41 (24)                   | $-14.6\pm0.3$ (4)                | $15.3\pm0.5$ (4)                |

Table 2. Average width of last 6 growth bands and average isotopic values from outer-12 mm of vertebrae from leopard sharks fed a constant diet of squid over 1250 days.
the dominant prey but cephalopods and tuna were also likely consumed when individuals were offshore, similar to results from satellite tagged white sharks [10].

(d) Individuality
The isotopic data in aggregate suggest that northeast Pacific white sharks are generalists at the population level, but further analysis reveals a high degree of individual specialization within the population. Individual differences within the white shark population can be attributed to changes in prey preference and foraging location with ontogeny. A core constraint on our analysis of niche occupancy and breadth is that if isotopic values differ between two specimens, then either prey type or foraging location (or both) must differ between the specimens. The converse is not true, however. Isotopic similarity between two specimens could result from consumption of the same prey in the same location, but also from fortuitous combinations of different prey types in different locations. Because we do not attempt to specify the particular prey, our assessment of individuality is a conservative measure and reflects minimal niche differences.

The proportional similarity index \( \bar{w}_{ij} \) quantified and compared isotopic variation within each individual to the population’s isotopic distribution. The \( \bar{w}_{ij} \) values for the sampled population indicated 8 specialists \( \bar{w}_{ij} = 0.73–0.90 \), 4 generalists \( \bar{w}_{ij} = 0.73–0.90 \), and 3 animals with intermediate values \( \bar{w}_{ij} = 0.42–0.59 \); Table 1). We note the distinction between \( \bar{w}_{ij} \) (which describes the proportional overlap between an individual’s isotopic values and the population’s average) and isotopic niche breadth (the absolute range of isotopic variation in an individual), as these two metrics are not necessarily correlated. For example, isotopic niche breadth was similar for specimens 42094–2 and 42898 PR (represented by \( \bar{w}_{ij} \) values for the sampled population indicated 8 specialists \( \bar{w}_{ij} = 0.33 \), 4 generalists \( \bar{w}_{ij} = 0.73–0.90 \), and 3 animals with intermediate values \( \bar{w}_{ij} = 0.42–0.59 \); Table 1). The higher \( \bar{w}_{ij} \) value for 42898 PR reflects the fact that its isotopic niche, while narrow, overlaps with a greater number of conspecifics.

Conclusions
Stable isotope analysis of white shark vertebrae provided lifetime records of diet and revealed a variety of feeding patterns. For example, we found significant variability in the degree and timing of shifts in \( ^{15} \text{N} \) values with age among individuals. Although there was a significant difference between young and adult diets, not all individuals displayed a shift to a higher trophic level prey. The isotopic data suggest that as a population, subadult to adult white sharks are generalist predators and consume a diverse array of high and low trophic level prey from nearshore and offshore habitats. However, a comparison between individual and population isotopic niche overlap revealed a high degree of dietary individuality.

Although confounding factors, such as the lack of biological metadata and unknown effects of physiological characteristics, prevent specific dietary assessments, this study provides the first evidence of ontogenetic and individual dietary variation in white sharks. One potential explanation for this pattern may be a mechanism to increase foraging success in species experiencing high intraspecific competition (i.e., the niche variation hypothesis [91]). Instead of individuals using all resources (i.e., potential prey or foraging locations) equally, individual foraging patterns diverge to create a spectrum of specialists that do not differ in survival or reproductive success [2,4,43,92,93]. This isotopic study, in conjunction with satellite tagging studies, is expanding our understanding of the foraging ecology of white sharks.

Supporting Information

File S1 Photo showing annual growth bands before (left) and after drilling (right) on specimen WS101. Ages are also noted next to the growth bands. (TIF)

File S2 The compiled prey isotope values from the literature and unpublished data used in Figure 2. (DOC)

File S3 The ontogeny of \( ^{15} \text{N} \) values for the 15 white sharks analyzed for this study. Individuals are in the same A, B, and C groups as Figure 1. (TIF)

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Author Contributions
Conceived and designed the experiments: SLK PLK. Performed the experiments: SLK MTT JAE PLK. Analyzed the data: SLK MTT JAE PLK. Contributed reagents/materials/analysis tools: SLK MTT JAE PLK. Wrote the paper: SLK MTT JAE PLK.

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Author Contributions

Conceived and designed the experiments: SLK PLK. Performed the experiments: SLK MTT JAE PLK. Analyzed the data: SLK MTT JAE PLK. Contributed reagents/materials/analysis tools: SLK MTT JAE PLK. Wrote the paper: SLK MTT JAE PLK.
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