Synthesis of Plasmepsin II Inhibitors – Potential Antimalarial Agents

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Abstract: A new series of plasmepsin II (PM II) inhibitors has been prepared based on 4-aminopiperidine-tert-butyl-carbamate (I). These compounds might be useful as antimalarial drugs acting via a new mechanism, and therefore be less susceptible to parasite resistance now often observed with current antimalarial therapies. Some of the final compounds prepared exhibited encouraging inhibitory activity towards PM II.

Keywords: Malaria, Plasmepsin II, Reductive Amination, Suzuki Coupling.

Introduction

The World Health Organization (WHO) estimates that more than 280 million people are infected with malaria. This situation worsens because drug resistant strains of the malaria parasite are spreading quickly. Among four different strains, *Plasmodium falciparum* causes the largest number of lethal infections and often shows substantial resistance against Chloroquin, still the most important drug used to treat malaria infections.

Plasmodia are transmitted by *Anopheles gambia* mosquitoes, which are developing greater pesticide resistance. Climate changes, changing agricultural habits and increased mobility are assumed to be responsible for the spread of the mosquito and the disease into formerly unaffected regions. Political
conflicts in regions affected by malaria have made disease control difficult. Thus, possible new drug targets are urgently needed which allow for the development of new medicines acting via different mechanisms in order to avoid fast development of resistance [1].

Among the various targets, the hemoglobin degradation pathway - involving several parasite specific enzymes like the plasmepsins [6] and the falcipains [2] - is of major interest. One possibility to strike against the parasite is to inhibit an enzyme within this pathway [2], interrupting the nutrition source and therefore eliminating the parasite by starvation.

Scheme 1.

Scheme 1 presents a simplified overview of processes in the food vacuole and shows many interesting targets for drug development [3-5]. PM II has been chosen because its inhibition follows a completely new and promising pathway of fighting against the malaria parasite having a good potential of being devoid of fast emerging resistance. This target fits well within our core competencies in the field of aspartic protease inhibition as a therapeutic strategy [6], which has been proven especially valid in the field of HIV-protease inhibition to treat AIDS. The lead compounds depicted below (Scheme 2) emerged from a high throughput screening (HTS) of a commercial library containing 50,000 compounds for PM II inhibitory activity.

One of the preferred lead structures found in the HTS effort considered as being suitable for optimization as PM II inhibitors is depicted in Scheme 2 exhibiting an IC\textsubscript{50} on PM II of 3.6 \(\mu\)M. By variation of different parameters like ring size, functional groups of substituents or chain length of
alkyl groups, the preferred key intermediate identified for further optimization cycles was 4-amino-
piperidine-tert-butylcarbamate (1) [7].

Scheme 2.

Part of the work presented here, was undertaken to elaborate the Structure-Activity-Relationships
(SAR) with respect to variations of R^II and R^IV (Scheme 2, R^I=H; for R^III either the trans-4-pentyl-
cyclohexanoyl- or the 4-pentyl[2.2.2]bicyclooctanoyl-unit was introduced as a replacement of the 4-
pentyl-benzoyl-group for which results are published in [6]). Therefore the synthesis of biaryl
aldehydes 4 by Suzuki coupling reactions [8] (Scheme 3) was investigated and optimized. Aldehydes 4
were then coupled in a reductive amination reaction onto N-Boc-4-aminopiperidine (1) [7,9]. The
subsequent acylation of amine 5 with suitable carboxylic acids or carboxylic acid chlorides (e.g. 6)
under standard reaction conditions resulted in the amides 7. Cleavage of the Boc-protecting group with
4 M HCl in dioxane gave the precursors 8. Final reductive amination with a diverse set of aldehydes
(e.g. 9) following standard literature procedures in a parallel chemistry set-up resulted in the target
inhibitors 10 which were tested for inhibition of plasmspin II in a FRET-assay [10]. In order to assess
the specificity and the selectivity of the PM II inhibitors (e.g. 10), the compounds were tested in
FRET-assays for the inhibition of other aspartic proteases like plasmepsin IV, cathepsin D or cathepsin
E. For the whole set of final compounds depicted in Tables 1 and 2, substantially lower or no
inhibition of these aspartic proteases was detected.

The main goal of this investigation was to find out what types of biaryl units in combination with
either the trans-4-pentylcyclohexanoyl- or the 4-pentyl[2.2.2]bicyclooctanoyl-unit in position R^III led
to the most active PM II inhibitors (e.g. 10; Scheme 4). In order to confirm the superior influence of a
biaryl moiety on the inhibitory activity, a limited number of compounds bearing a 4-alkyl- or 4-
alkoxy-substituted benzyl group were also prepared (see Tables 1 and 2).
Scheme 3.

Suzuki couplings:

\[
\begin{align*}
\text{HCO} & \quad + \quad \text{Br}_{\text{N}} \quad \text{O} \\
\text{2} & \quad \text{3 mol\% Pd(PPh3)_4} \\
& \quad \text{K}_2\text{CO}_3 \text{ (aq.)} \\
& \quad \text{Toluene, i-PrOH} \\
\text{3} & \quad \text{4} \\
\end{align*}
\]

Scheme 4. Representative synthetic pathways for the preparation of 4-amino-piperidine based PMII inhibitors

1

\[
\begin{align*}
a) & \quad 4, \text{MeOH} \\
b) & \quad \text{NaBH}_4 \\
\end{align*}
\]

5

\[
\begin{align*}
\text{Cl} & \quad \text{NEt}_3, \text{CH}_2\text{Cl}_2 \\
\end{align*}
\]

6

7

\[
\begin{align*}
\text{4 M HCl / dioxane} \\
\end{align*}
\]

8

9

\[
\begin{align*}
\text{NaBH(OAc)}_3 & \quad \text{CH}_2\text{Cl}_2 \\
\end{align*}
\]
Results and Discussion

Tables 1 and 2 summarize the results for the two series of PM II inhibitors prepared according to the pathway depicted in Scheme 4. Table 1 shows compounds bearing the trans-4-pentyl-cyclohexanoyl unit as R\textsuperscript{III}. With respect to R\textsuperscript{1} the differences in activity between the compounds containing a 4-pyridyl moiety and the compounds containing a 3-pyridyl moiety are in the range of a factor of 2, with no clear preference for either pyridyl moiety. Replacement of the pyridyl units by an ethoxy group results in a 6- to 10-fold loss of activity (with the exception of the compounds given in row 6, Table 1, with R\textsuperscript{2} = 2-imidazolyl, where already the pyridyl substituted compound exhibits low activity). With respect to R\textsuperscript{2}, the small lipophilic 2-methyl-propyl unit leads to more active compounds than the introduction of larger, heteroatom containing (e.g. hydroxy-phenyls (rows 2, 3, Table 1) or pyridyls (rows 4, 5, Table 1) or highly hydrophilic units like 2-imidazolyl, leading to an almost complete loss of activity in this series.

Table 1:

| Row | R\textsuperscript{1} | R\textsuperscript{2} | IC\textsubscript{50} (PM II) in nM |
|-----|-------------------|--------------------|-------------------------------|
| 1   | 32                | 50                 | 189                           |
| 2   | 117               | 64                 | 651                           |
| 3   | 151               |                    |                               |
| 4   | 189               |                    |                               |
| 5   | 137               | 61                 | 937                           |
| 6   | 2255              |                    | 2736                          |

N\textsubscript{N}N
N\textsubscript{O}
Table 2 gives the results for the PM II inhibitors bearing the 4-pentyl[2.2.2]bicyclooctanoyl moiety in position R\textsuperscript{III}. The compounds possessing a 4-pyridyl group in position R\textsuperscript{1} are generally more active than the compounds having an isopropyl unit in this position. This reflects the same SAR tendencies already elaborated for the first series of inhibitors given in Table 1. In the second group of inhibitors (Table 2) the introduction of a 2-imidazolyl unit in position R\textsuperscript{2} is much less detrimental for the inhibitory activity (col. 1, row 4, Table 2). But again the small lipophilic 2-methyl-propyl group leads to the most active derivatives. The introduction of the 4-hydroxy-phenyl (row 2, Table 2) or the 4-pyridyl (row 3, Table 2) unit results in derivatives exhibiting comparable activities as analogous compounds from Table 1. The influence on changes in activity by moving from trans-4-pentyl-cyclohexanoyl as R\textsuperscript{III} to 4-pentyl[2.2.2]bicyclooctanoyl as R\textsuperscript{III} seems to be weak. Both substituents lead to the same SAR within the respective series of PM II inhibitors.

**Table 2:**

| Row | R\textsuperscript{1} | IC\textsubscript{50} (PM II) in nM |
|-----|----------------------|-------------------------------|
| 1   |                      | 37                            | 313                           |
| 2   | -O-H                | 92                            | 808                           |
| 3   | -N                  | 107                           | 4799                          |
| 4   | -N                  | 108                           | 697                           |
| 5   | -S                  |                                | 1247                          |
| 6   |                      |                                | 651                           |
The most active compounds prepared within these optimization efforts exhibit potencies which are a 100-fold higher than the activity of the initial lead compound depicted in Scheme 2. It has been shown that 4-aminopiperidine might well serve as a template to develop non-peptidic inhibitors of PM II allowing for the use of a parallel chemistry setup for the fast preparation of small focused libraries. In future work the potency of the PM II inhibitors series described needs to be further improved. In addition the physicochemical properties of the inhibitors (e.g. clogP, polar surface area or molecular weight) have to be optimized towards more drug-like values. Investigations are ongoing and results will be published in due course. The rather poor physicochemical properties of the final compounds depicted in Tables 1 and 2 could raise questions about a non-specific interaction of the inhibitors with the enzyme. According to preliminary results obtained from X-ray analysis there is strong evidence for a specific interaction of the inhibitors with the active site of the enzyme. This fact confirms the results obtained in the selectivity screening discussed above.

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Experimental

General

$^1$H-NMR spectra were recorded at 300 MHz using a Varian Oxford NMR 300 and chemical shifts are reported in $\delta$ values (ppm) using DMSO-d$_6$ as internal standard (unless otherwise specified). Purities of all compounds were over 95% and were determined by either $^1$H-NMR or LC-MS (Waters Micromass; ZMD-platform with ESI-probe with Alliance 2790 HT; Colum: 2x30mm, Gromsil ODS4, 3µm, 120A; Gradient: 0 – 100% acetonitrile in water, 6 min, with 0.05% formic acid, flow: 0.45mL/min). All solvents were stored over molecular sieves, except wet solvents used in Suzuki reactions. All reagents were used as received from commercial sources without further purification.

Selected Procedures [11]

4-Pyridin-4-yl-benzaldehyde (4)

To a solution of 3-bromopyridine hydrochloride (3) (2.92 g, 15 mmol, 1 eq) in toluene (30 mL) was added iso-propanol (30 mL), 4-formylphenylboronic acid (2) (2.48 g, 16.5 mmol, 1.1 eq) and 2 M aqueous K$_2$CO$_3$ (40 mL, 80 mmol, 5.3 eq). The solution was degassed with N$_2$ for 15 minutes. After adding Pd(PPh$_3$)$_4$ (0.519 g, 0.45 mmol, 0.03 eq), the solution was refluxed for 7 h under argon. The reaction mixture was diluted with water and extracted three times with ethyl acetate. The combined organic layers were washed twice with brine, dried over MgSO$_4$ and the solvent was evaporated.
Recrystallization from ethyl acetate gave 2.13 g (77%) of 4-pyridin-4-yl-benzaldehyde (4) as yellow needles. LC: \( t_r \) 2.41'; MS: 184 [M+H]+; \(^1^H\)-NMR: 10.10 (s, 1H), 8.71 (d, 2H), 8.06 (s, 4H), 7.77 (d, 2H).

**4-(4-Pyridin-4-yl-benzylamino)-piperidine-1-tert-butyl carbamate (5)**

A solution of 4 (1.07 g, 5.33 mmol, 1 eq) and 1 (976 mg, 5.33 mmol, 1 eq) in methanol (30 mL) was refluxed for 4 h. After cooling to r.t., NaBH\(_4\) (222 mg, 5.87 mmol, 1.1 eq) was added carefully over 30 minutes. After 2 h of additional refluxing, the solution was carefully diluted with water and extracted three times with ethyl acetate. The combined organic layers were washed twice with brine, dried over MgSO\(_4\) and the solvent was evaporated to give 1.90 g (96%) of compound 5 as a yellow solid which could be used in further transformations without purification [7]. LC: \( t_r \) 2.68'; MS: 368 [M+H]+; \(^1^H\)-NMR: 8.63 (d, 2H), 7.82-7.41 (m, 6H), 3.89-3.74 (m, 3H), 3.79 (s, 1H), 3.08-1.73 (m, 5H) 1.40 (br s, 9H) 1.28-1.09 (m, 2H).

**4-[(4-trans-Pentyl-cyclohexanecarbonyl)-(4-pyridin-4-yl-benzyl)-amino]-piperidine-1-tert-butyl carbamate (7)**

To a solution of trans-4-pentylcyclohexanecarboxylic acid (485 mg, 2.45 mmol, 1 eq) in CH\(_2\)Cl\(_2\) (20 mL) at 0°C was added oxalic acid chloride (0.35 mL, 4.14 mmol, 1.7 eq) and 3 drops of DMF. The mixture was stirred in an N\(_2\) atmosphere for 1 h and carefully evaporated (40°C, 200 mbar, argon at inlet) [12]. The obtained yellowish viscous acid chloride 6 was dissolved in an Ar atmosphere in CH\(_2\)Cl\(_2\) (10 mL), followed by the addition of NEt\(_3\) (1.2 mL, 8.61 mmol, 3.5 eq) and 5 (900 mg, 2.45 mmol, 1 eq) dissolved in CH\(_2\)Cl\(_2\) (10 mL). Stirring was continued at r.t. for 12 h. The mixture was diluted with ethyl acetate, washed once with saturated Na\(_2\)CO\(_3\) solution and with brine, dried over MgSO\(_4\) and the solvent was evaporated. The crude material was purified by column chromatography (silica gel 60, DCM/Hex/MeOH 10:4:1) to give 623 mg (46%) of the amide 7 as white filaments [7]. LC: \( t_r \) 5.85'; MS: 548 [M+H]+; \(^1^H\)-NMR in CDCl\(_3\) (shows both amide rotamers with a ratio of ~1:1): 8.63-8.52 (m, 2H), 7.65-7.14 (m, 6H), 4.73-4.60 (m, 1H), 4.31, 4.24 (s, 2H, benzylic methylene of both rotamers), 4.19-3.92 (m, 2H), 2.77-0.56 (m, 36H).

**4-trans-Pentyl-cyclohexanecarboxylic acid piperidin-4-yl-(4-pyridin-4-yl-benzyl)-amide hydrochloride (8)**

A solution of compound 7 (617 mg, 1.13 mmol, 1 eq) in 4 M HCl / dioxane (10 mL, 40 mmol, 35.4 eq) was stirred for 4 h at r.t. under argon. The solvent was evaporated to give 8 as a white solid in quantitative yield [7]. LC: \( t_r \) 3.67' (purity of >95%), MS: 448 [M+H]+.
4-trans-Pentyl-cyclohexanecarboxylic acid [(1-(4-hydroxy-benzyl)-piperidin-4-yl)-(4-pyridin-4-yl-benzyl)-amide (10)

To a solution of compound 8 (63 mg, 0.131 mmol, 1 eq) in CH$_2$Cl$_2$ (10 mL) was added 4-hydroxy-benzaldehyde (64 mg, 0.524 mmol, 4 eq) and NaBH(OAc)$_3$ (138 mg) and stirring was continued at r.t. under argon for 12 h. The reaction mixture was washed once with saturated NaHCO$_3$ solution, dried over MgSO$_4$ followed by chromatographic purification to give 42.3 mg (58%) of the final compound 10 as a white solid. LC: $t_r$ 4.33'; MS: 555 [M+H]$^+$; $^1$H-NMR: 8.68-8.47 (m, 2H), 7.66-6.61 (m, 10H), 5.27 (s, 1H), 4.59, 4.56 (s, 2H, benzylic methylene of both rotamers) 3.86 (m, 1H), 3.19 (s, 2H), 3.07-2.79 (m, 2H), 2.62-2.41 (m, 1H), 2.31-0.59 (m, 26H).

References

1. (a) Bell, A. *I. Drugs* 2000, 3, 310-317; (b) Bell, A. *Curr. Opin. Anti-infect. Invest. Drugs* 2000, 2, 63-70.
2. Rosenthal, P. J.; Sijwali, P.S.; Singh, A.; Shenai, B. R. *Curr. Pharm. Des.* 2002, 8, 99-110.
3. http://sites.huji.ac.il/malaria/hemoglobindigest.html
4. Rosenthal, P. J. *Antimalarial Chemotherapy: Mechanism of Action, Resistance, and New Directions in Drug Discovery*; Humana Press: New Jersey, 2001.
5. Berry, C. *Curr. Opin. Drug Disc. Dev.* 2000, 3, 624-629.
6. Boss, C.; Richard-Bildstein, S.; Weller, T.; Fischli, W.; Meyer, S.; Binkert, C. *Curr. Med. Chem.* 2003, 10, 883-907.
7. Actelion Pharmaceuticals Ltd (Boss, C.; Fischli, W.; Meyer, S.; Richard-Bildstein, S.; Weller, T.), 2002, WO 02/24649.
8. (a) Miyaura, N.; Suzuki, A. *Chem. Rev.* 1995, 95, 2457-2483; (b) Watanabe, T.; Miyaura, N.; Suzuki; A. *Synlett* 1992, 207-210; (c) Gong, Y.; Pauls, H. W. *Synlett* 2000, 829-831.
9. Abdel-Magid, A. F.; Carson, K. G.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. *J. Org. Chem.* 1996, 61, 3849-3862.
10. Matayoshi, E. D.; Wang, G. T.; Krafft, G. A; Erickson, J. *Science* 1990, 247, 954-958.
11. Mueller, R. *Diploma Thesis* 2002, University of Applied Sciences Basel, Muttenz.
12. Taber, D. F.; Deker, P. B.; Gaul, M. D. *J. Am. Chem. Soc.* 1987, 109, 7488-7494.

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