Thymic hyperplasia after chemotherapy in adults with mature B cell lymphoma and its influence on thymic output and CD4$^+$ T cells repopulation

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ABSTRACT
To investigate the thymic regenerative potential in adults accepting chemotherapy for lymphoma. The dynamics of thymic activity in 54 adults from baseline to 12 mo post-chemotherapy was analyzed by assessing thymic structural changes with serial computed tomography (CT) scans, and correlating these with measurements of thymic output by concurrent analysis of single-joint (sj) T-cell receptor excision circles (sjTREC) and CD31$^+$ recent thymic emigrants (RTE) in peripheral blood. Furthermore, the consequence of thymic renewal on peripheral CD4$^+$ T cell recovery after chemotherapy was evaluated. Time-dependent changes of thymic size and thymic output assessed by both sjTREC levels and CD31$^+$ RTE counts in peripheral blood were observed during and after chemotherapy. Enlargement of thymus over baseline following chemotherapy regarded as rebound thymic hyperplasia (TH) was identified in 20 patients aged 18—53 y (median 33 y). By general linear models repeated measure analysis, it was found that, patients with TH (n = 20) had a faster recovery of sjTREC levels and CD31$^+$ RTE counts after chemotherapy than patients with comparable age, gender, diagnosis, disease stage, thymic volume and output function at baseline but without TH (n = 18) (p = 0.035, 0.047); besides, patients with TH had a faster repopulation of both naïve CD4$^+$ T cell and natural regulatory CD4$^+$ T cell subsets than those without TH (p = 0.042, 0.038). These data suggested that adult thymus retains the capacity of regeneration after chemotherapy, especially in young adults. The presence of TH could contribute to the renewal of thymopoiesis and the replenishment of peripheral CD4$^+$ T cell pool following chemotherapy in adults.

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Introduction
A variety of factors other than age such as stress and toxicity could result in an alteration in the structure and cellular composition of the thymus. However, the thymus tissue is plastic and has the capacity for renewal on recovery.1 Thymus atrophy has been observed following chemotherapy for malignancies, with a reduction in cortical lymphocytes, a shrinkage of the thymic lobules, and a decrease in the production of naïve T cells. After the cessation of therapy, the thymus regrows, and restores the output of naïve T cells.2 Excessive re-growth of the thymus may sometimes occur during recovery, and is termed rebound TH.2 TH may be related with a robust thymic regeneration, and is characterized by an increase in thymic size and density without changing its normal shape and histological appearance.3 This phenomenon is common in children and adolescents, and can occasionally be observed in young adults, but is rare in older patients,2,4-7 possible due to a decreased regenerative capacity of thymus with age. However, the small number of adults studied to date has provided only limited knowledge on the renewal capacity of adult thymus.

The assessment of thymic activity after chemotherapy is mainly based on studies of the regeneration of T lymphocytes with naïve phenotype (CD45RA$^+$CD4$^+$) and thymic imaging.2,6-10 The main limitation of these approaches is that they do not actually measure RTE but rather the more mature naïve T cells, which can proliferate in the peripheral blood or be converted from memory T cells.11 Moreover, thymic function is not always consistent with its mass. Therefore, more reliable approaches are needed for deep understanding of the dynamics of thymic activity. Over the past decade, sjTREC formed during T cell receptor rearrangement and CD31$^+$ naïve CD4$^+$ T cells were regarded as more reliable markers of thymic output, and have been widely used in the studies of haematopoietic stem cell transplantation (HSCT), human immuno-deficiency virus (HIV) infection and congenital immunodeficiency disorders, but less in chemotherapy.10-14

Full immune recovery after chemotherapy depends on thymic output of new RTE to replenish the peripheral naïve T cells pool. In the young, TH predicts a renewal of thymic activity, and contributes to the reconstitution of naïve T cells with a wide T cell receptor (TCR) repertoire.9 In contrast, homeostatic
proliferation is thought to be the major contributor to immune reconstitution in adults, leading to a delayed recovery of naive T cell populations after chemotherapy. No correlation has been found between thymic size and early recovery of peripheral naive CD4+ counts after chemotherapy in adults. However, aged thymus still has the renewal capacity to rebuild a T cell repertoire. TH after chemotherapy observed in younger adults was found to be correlated with a faster and more complete recovery of naive T cells. Patients’ age-related thymic output and long-term consequences for their immune system remains to be fully understood.

The present study was designed to investigate the thymic regenerative potential in adults with lymphoma after chemotherapy, by assessing thymic structural change with serial CT scans and correlated these with measurements of thymic output by concurrent analysis of sjTREC and CD31+ RTE. Furthermore, the consequence of thymic renewal on peripheral CD4+ T cells recovery after chemotherapy was evaluated.

**Results**

**Dynamics of thymic volume during and after chemotherapy**

Time-dependent changes of thymic size were observed in 54 patients studied: thymic volume showed an inverse correlation with the age (Spearman test r = −0.707, p < 0.001); during treatment, the thymic soft tissue mass reduced in 16/54 (30%) and remained stable in 37/54 (69%) patients, yet increased in one patient; after the end of treatment, thymic enlargement was obvious in 24/54 (44%) patients, minimal change of thymus was detected in the remaining 30/54 (56%) cases.

TH was identified in 20/54 (37%) patients aged 18–53 y (median 33 y), whose clinical characteristics are shown in Table 1. The change in their thymic size was particularly remarkable (Fig. 1): prior to therapy, 17/20 (85%) of them had a TI of 0–2 with no or minimal soft tissue in the thymus; during therapy, all but one patient (95%) attained a TI of 0–2; yet, at the onset of TH, 11/20 (55%) of them achieved a TI of at least three with moderate to mass-like thymic soft tissue. Thus, the thymic size recovered from chemotherapy in these patients not merely returned to the pretreatment status but to a higher level.

**Dynamics of thymic output during and after chemotherapy**

To determine the change of thymic output during and after chemotherapy, CD31+ RTE counts and sjTREC levels in peripheral blood were measured serially from baseline to 12 mo post-

| characteristics                        | patients with TH (n = 20) | patients without TH (n = 18) | p     |
|----------------------------------------|--------------------------|-----------------------------|-------|
| Median age (year)                      | 33(18–53)                | 37(28–49)                   | 0.082 |
| Gender (%)                             |                          |                             |       |
| Male                                   | 9/20 (45)                | 11/18 (61)                  | 0.341 |
| Female                                 | 11/20 (55)               | 7/18 (39)                   |       |
| Disease type (%)                       |                          |                             |       |
| DLBCL                                  | 9/20 (45)                | 11/18 (61)                  | 0.341 |
| HL                                     | 5/20 (25)                | 2/18 (11)                   |       |
| Other types                            | 6/20 (30)                | 5/18 (28)                   |       |
| Disease stage (%)                      |                          |                             |       |
| I–II                                   | 7/20 (35)                | 7/18 (39)                   | 1.000 |
| III–IV                                 | 13/20 (65)               | 11/18 (61)                  |       |
| CD4+T cells counts (× 10⁹/L)           |                          |                             |       |
| At baseline                            | 676 ± 382                | 572 ± 358                   | 0.382 |
| At the nadir                           | 351 ± 215                | 329 ± 189                   | 0.597 |
| Thymic index at baseline (%)           |                          |                             |       |
| 0                                      | 2/20 (10)                | 5/18 (28)                   | 0.410 |
| 1                                      | 6/20 (30)                | 3/18 (17)                   |       |
| 2                                      | 9/20 (45)                | 9/18 (50)                   |       |
| 3                                      | 2/20 (10)                | 1/18 (5)                    |       |
| 4                                      | 1/20 (5)                 | 0/18 (0)                    |       |
| Thymic output at baseline              |                          |                             |       |
| CD31+RTE (× 10⁹/L)                     | 155 ± 127                | 136 ± 124                   | 0.742 |
| sjTREC (copies/10⁶PBMCs)                | 9873 ± 9621              | 8526 ± 8625                 | 0.793 |

Abbreviations: DLBCL, diffuse large B cell lymphoma; HL, Hodgkin lymphoma; RTE, recent thymic emigrants; sjTREC, single-joint T-cell receptor excision circles.
chemotherapy (Fig. 2). Both the numbers of CD31$^+$ RTE and sjTREC levels started to reduce with the administration of cytotoxic drugs, and got down to the nadir at the end of treatment ($p = 0.003, 0.015$, respectively), then remained low during the first half year of follow-up; thereafter, CD31$^+$ RTE counts and sjTREC levels rose significantly at 9 and 6 mo after the end of chemotherapy respectively ($p < 0.001$), and were not significantly different from the pretreatment levels ($p > 0.05$).

**The impact of thymic hyperplasia on thymic output**

It should be noted that, the level of thymic output will vary according to a number of background factors, including age, gender, disease status and thymic activity at baseline. To assist the evaluation of the influence of TH on thymic output after chemotherapy, we compared quantitative changes of CD31$^+$ RTE and sjTREC in peripheral blood prior, during and after chemotherapy in patients with ($n = 20$) and without ($n=18$) TH (Table 1). Patients in two groups were not significantly different in age, gender, diagnosis, disease stage, thymic volume and output function (CD31$^+$ RTE and sjTREC) at baseline, and CD4$^+$ cell numbers at baseline and the nadir ($p > 0.05$). By general linear models repeated measure analysis, impact of the presence of TH was found on the recovery of both CD31$^+$ RTE counts and sjTREC levels between two groups ($p = 0.035, 0.047$, respectively). The numbers of CD31$^+$ RTE and sjTREC levels in subjects with TH were higher than those in subjects without TH at each time point beyond 6 mo of follow-up ($p < 0.05$), indicating a fast recovery of thymic output in patients with TH (Fig. 3A and B).

**The impact of thymic hyperplasia on CD4$^+$ T cells repopulation**

Next, we assessed whether the presence of TH could promote the reconstitution of naïve CD4$^+$ T cells after chemotherapy. Since the replenishment of both naïve CD4$^+$ T cells and natural regulatory CD4$^+$ T cells (nTregs) in the peripheral blood following lymphopenia relies on thymic de novo production,20,21 we performed a serial analysis of peripheral naïve CD4$^+$ T cells and nTregs counts from baseline to 12 mo post-chemotherapy in patients with ($n = 20$) and without TH ($n = 18$) (Table 1). By repeated-measure analyses adjusted for the presence of thymic rebound using general linear models, it was found that, subjects with TH could reconstitute the peripheral naïve CD4$^+$ T cell pool significantly faster than subjects without TH during the 12 mo follow-up ($p = 0.042$). The numbers of peripheral naïve CD4$^+$ T cells in both groups got down to the nadir at the end of chemotherapy ($p = 0.005, 0.019$, respectively), and rose significantly at 6 and 9 mo after chemotherapy discontinuation in cases with and without TH, respectively ($p = 0.028, 0.036$) (Fig. 4A). Similarly, the repopulation of nTreg was found to be
faster in subjects with TH ($p = 0.038$), in whom the numbers of nTreg rose significantly at 6 mo after the completion of treatment ($p = 0.032$) before declining to the nadir at the end of treatment ($p = 0.007$). Yet, no significant increase in nTreg counts was observed in those without TH within one year of follow-up ($p > 0.05$) (Fig. 4B).

**Discussion**

During the involutional process, the densely cortical and medullary tissues of the thymus reduce and the thymopoietic productivity decline. In adults over 40 y, the thymus is usually mostly fatty in composition, but remains active and generates new T cells at a lower rate. Since thymic activity is dependent upon age, the renewal ability of the thymus maybe also influenced by host age. Information on the thymic renewal capacity following chemotherapy has been obtained from different age groups. Thymic atrophy was reported to be observed in over 90% of young patients (2–35, mean 17 y) receiving chemotherapy for malignancies; withdrawal of chemotherapy resulted in variable enlargement of the thymus; reactive TH or rebound, defined as a diffuse enlargement of thymus (re-growth 50% or greater than baseline volume) on CT scans was observed in 25% of them, suggesting a high regenerative capacity of the thymus in children, adolescents and young adults. In a group of older adults (26–63, mean 46 y), however, this thymic renewal capacity is limited. Only a low frequency (22%) of thymic atrophy followed by a low incidence (11%) and minimal degree of hyperplasia was observed after high-dose chemotherapy and autologous stem cell transplantation. In this study, the thymus appeared to atrophy during chemotherapy in 30% of adults aged 18–59 y (median 35 y), and regrow on recovery in nearly half of them. Moreover, TH was found to be a relatively common phenomenon occurring in 37% of those adults, of whom the eldest was 53 y old. No TH was found in those above 60 y. It is concluded that adult thymus retains the capacity of regeneration, especially in young adults. In agreement with our findings, Sfikakis et al. demonstrated an enlargement of the previously atrophic thymus in 63% of younger adults (18–49 y, median 30 y), but in none of the elderly.

In consistent with the changes in thymic size, time-dependent changes of thymic output could be observed during and after chemotherapy. Thymic output monitored by determining the numbers of CD31+ RTE and indirectly by measuring the levels of sjTREC in peripheral blood decreased to the nadir at the end of chemotherapy, but recovered to baseline levels within one year after completion of chemotherapy. It can be speculated that, cytotoxic drugs and glucocorticoid hormones used in chemotherapy could effectively extinguish thymic output, followed by a restoration of thymic output after the removal of drug toxicity and the elimination of stress imposed by the tumor with successful therapy. Furthermore, it is confirmed that, TH observed in young adults post-chemotherapy could influence the regeneration rate of both CD31+ RTE and sjTREC in the peripheral blood. The faster recovery of thymic output observed in subjects with TH following chemotherapy suggested that thymic structural re-growth served as a basis for the renewed thymopoiesis. Thymic enlargement concurrent with the restoration of thymic output further supported the hypothesis that TH after chemotherapy predicts a robust renewal of thymopoiesis. Multiple mechanisms including the numeric and functional recovery of cortical thymic epithelial cells that determine the overall lymphopoietic capacity of the thymus might play important roles in the thymic regeneration and contribute to the process of TH following chemotherapy. In support of this point, HSCT recipients with TH was reported to display a robust renewal of thymopoiesis during recovery from lymphopenia, as evidenced by increases in sjTREC bearing cells with broad TCR diversity in the peripheral blood.

The potential role of the adult thymus in T cell immune recovery following chemotherapy-induced lymphopenia remains the subject of debate. CD4+ T cell repopulation subsequent to lymphopenia relies on two pathways: production of T cells de novo and CD4+ T cells. The thymic pathway is commonly compromised in adults, and CD4+ T cells rely more upon expansion to restore peripheral T cell pool, which could reduce the TCR diversities and increase the risk for infections and cancer. In the study of Vedel et al., no association was found between thymic size and early recovery of peripheral naïve CD4+ counts after chemotherapy, suggesting a limited role of adult thymus in CD4+ T cells repopulation. However, other two studies revealed that thymic output was still an important pathway for CD4+ T cell reconstitution in adults, and TH correlated significantly with a faster and more complete recovery of naïve CD4+ T cells after chemotherapy. As reported, TH after
chemotherapy, observed in this cohort of adults we studied, could promote the repopulation of naïve CD4+ T cells within one year of follow-up, although no impact of TH has been found on the numeric recovery of total CD4+ T cells in our previous study.26 Hence, it is concluded that, TH after chemotherapy in adults, as in children and adolescents, predicts a renewal of thymic activity, and contributes to the reconstitution of peripheral T cell populations by generating naïve T cells de novo. Besides, thymic enlargement occurring in adults under HSCT and anti-viral therapy for HIV, was reported to be correlated with higher TREC frequency and greater numbers of naïve and total CD4+ T cells as well, consistent with the idea that the adult thymus remains active late in life and retains the capacity to orchestrate normal T lymphopoiesis.10,27

Furthermore, this data explored the role of adult thymus in maintaining peripheral Tregs pool after chemotherapy. Tregs is a heterogeneous population that consists of thymic derived natural (n)Treg and peripheral converted induced (i)Treg cells.21 It has been shown that, most recovering Tregs after chemotherapy were peripherally derived, possibly due to the excessive proliferation of iTregs under immune activation and the impaired thymic output of nTregs.28 This study revealed that, the numbers of peripheral nTregs declined significantly with cytoreductive therapy; during recovery, the repopulation of nTregs was faster in subjects with TH, indicating an increased thymic production of nTregs correlated with thymic renewal. This data provided the first evidence of an important role of renewed thymopoiesis in the Tregs repopulation following chemotherapy. Likewise, the replenishment of nTregs subsequent to lymphopenia induced by HSCT, HIV infection or autoimmune disease, was found to rely on de novo thymic production.29,31

Admittedly, there are some limitations to this study. sjTRECs are not replicated during mitosis and thus, become diluted with each successive iteration of cell division. Therefore, levels of sjTRECs in rapidly dividing cells can underestimate the ability of thymic regeneration.32 On the other hand, besides the sole thymic T cell neogenesis, CD31+ naïve CD4+ T cells can proliferate and maintain their CD31 expression in peripheral blood. Thus, the CD31+ RTE data potentially underestimate thymic derivation as well.33 Thymic activity evaluation by sjTRECs in naïve CD4+ T cells could be more sensitive and informative.

In conclusion, this study confirmed adult thymus retains the capacity of regeneration after chemotherapy, especially in young adults, manifesting as an increase in thymic volume and output function. TH was related to a faster recovery of CD4+ naïve T cells and nTreg after chemotherapy, indicating that the thymic output pathway may play a role in the reconstitution of CD4+ T cells in young adults. Adult thymus appears to be pivotal for reconstitution of T lymphoid immunity following chemotherapy, and could be the target of therapeutic improvements.

Materials and methods

Patients

Eighty-four lymphoma patients with mature B cell lymphoma admitted at the Department of Hematology, the First Affiliated Hospital of Nanjing Medical University between January 2013 and January 2015 were studied. All of them accepted chemotherapy at lymphoma diagnosis. Patients with diffuse large B cell lymphoma (DLBCL) were treated with R-CHOP (rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone) or R-DA-EPOCH (rituximab, etoposide, prednisone, vincristine, cyclophosphamide, and doxorubicin) regimen. For patients with Hodgkin lymphoma (HL), ABVD (adriamycin, bleomycin, vinblastin and dacarbazine) regimen was administered. Patients with follicular lymphoma (FL) or marginal zone lymphoma (MZL) accepted R-CHOP regimen mentioned above or R-COP (rituximab, cyclophosphamide, vincristine and prednisone) regimen. Eight patients who had no response to therapy or had disease progression within one year after the end of treatment were excluded. Sixteen patients who had response to chemotherapy and underwent HSCT as first-line consolidation were excluded. In six patients, it was not possible to determine the size of the thymus because of the lymphoma growth. The remaining 54 patients (32 DLBCL, 12 HL, 6 FL, 4 MZL), aged 18–59 y (median 35 y), were included. CT examinations were performed prior (baseline), during (after three cycles of chemotherapy) and after (0, 3, 6, 9, 12 mo post-chemotherapy) treatment. Simultaneously, blood samples were collected for flow cytometric analyses and the obtainment of peripheral blood mononuclear cells (PBMC) by density gradient centrifugation. This single-center retrospective study was approved by hospital ethical committee and admitted by the patients.

Imaging of thymus

Serial analysis of thymus structural changes was performed by reviewing CT images. The thymic size was scored using thymic index (TI) on a scale from 0 to 5 as described elsewhere:19 0, no soft tissue; 1, minimal soft tissue, barely recognizable; 2, minimal soft tissue, more obvious; 3, moderate soft tissue; 4, moderate soft tissue of greater extent, almost mass-like; and 5, mass-like appearance, suggesting hyperplasia or thymoma. Thymic enlargement, or subsequent regression, was defined as a change of at least 1 on the 0–5 scale. Enlargement of the thymus over baseline in the absent of any clinical, laboratory or radiological sign of disease progression was interpreted as TH.

Detection of CD31+ RTE by flow cytometry

Repeated analyses of peripheral blood lymphocyte subsets were performed in parallel with the CT examination. Peripheral blood samples were collected and stained immediately using the whole blood lysis technique. Phenotypic analyses were made with combinations of FITC, PE, APC and PerCP-conjugated mAbs (Becton Dickinson, USA) against CD3, CD4+, CD45RA, CCR7, CD31, CD25, CD127 and isotype control, using FACS Calibur (Becton Dickinson, USA) as described.34,35 The number of CD4+ cells expressing naïve phenotype (CD3+CD4+CD45RA–CCR7+), CD31+ naïve CD4+T cells (CD3+CD4+CD45RA+CD31+) and CD4+ regulatory T cells (Tregs) with naïve phenotype (CD4+CD25+CD127lowCD45RA+) was determined. The fluorescence of 40,000 cells was measured. Whole blood lymphocyte counts were performed with an automated analyzer, and the absolute numbers of cell subsets were determined by multiplying the total lymphocyte
count by the fraction of lymphocytes bearing the specific phenotypic markers.

**Detection of sjTREC by RQ-PCR**

Serial quantification of sjTREC in DNA of PBMCs was done by TaqMan real-time quantitative PCR assay using a StepOnePlus (Applied Biosystems, USA) according to the method of Tang et al. DNA was extracted from PBMCs using the QIAamp Blood DNA Mini Kit (QIAGEN, German) according to instructions, and the DNA concentration was determined by spectrophotometry (Eppendorf, Germany) prior to further analysis. The sequences of the forward and reverse primers and probe for the sjTREC were 5′ AACAGCCTTTGGGACACTATCG 3′, 5′ GCTGAACITATTG CA ACTGTGAG 3′, and 5′ 6-FAM-CCACATCCCTTGAACCATGCTGACACCTC-TAMRA 3′. As an internal control, the expression of the RAG2 gene was simultaneously analyzed using the following forward and reverse primers and probe: forward primer, 5′ GCAACTGGGAAATGGACTG 3′; reverse primer, 5′ GGTTG CAAAATTCATCATCACCAC 3′, and the probe 5′ 6-FAM-CCCCTTGCTTCTTG TTGATGTGGACGTGTGTGA-TAMRA 3′. Each PCR mixture contained around 10,000 copies of genomic DNA, 0.8 μL TREC or RAG2 probe, 0.4 μL each sjTREC or RAG2 primer, 0.4 μL ROX Reference Dye (50 ×) and 10 μL Premix Ex Taq (2 ×) (TaKaRa, China). The thermal cycling conditions were 2s at 95°C, and 50 cycles of 1s at 94°C and 20s at 60°C. A standard curve based on a plasmid preparation containing the sjTREC target sequence was plotted, and sjTREC values for samples were calculated using the StepOne software (Applied Biosystems, USA). Samples were analyzed in triplicates, and the median was calculated. The percentage of the copies of the sjTREC gene to that of the RAG2 gene was defined as the sjTREC level.

**Statistical analysis**

Continuous variables are expressed as mean ± standard deviation (SD) and categorical ones as number of cases (percentage). K–S test was used for normality test. Pearson or Spearman correlation analysis was applied for correlation analysis of two factors. Differences in numerical data were compared by the independent t test or Mann–Whitney U test. Differences in categorical data were compared using the chi-square test or Fisher exact test. The changes of each individual’s thymic output and lymphocyte subsets at different time points were assessed by general linear models repeated-measure analysis using the statistical tests within subject contrasts. The impact of TH on the recovery of thymic output and lymphocyte subsets within individual patients was evaluated by general linear models repeated-measure analysis using between-subject contrasts adjusted for the presence of TH. Data analysis was performed using SPSS21 statistical software. A value of p < 0.05 was considered to be significant.

**Disclosure of potential conflicts of interest**

No potential conflicts of interest were disclosed.

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