YAP1 Promotes Tumor Invasion and Metastasis in Nasopharyngeal Carcinoma with Hepatitis B Virus Infection

Introduction: Nasopharyngeal carcinoma (NPC) patients with HBsAg (+) commonly present with high frequencies of distant metastasis and poor survival rate; however, the mechanism has not been elucidated.

Materials and Methods: We analyzed the yes-associated protein 1 (YAP1) expression between HBsAg (+) and HBsAg (-) of NPC patients, then analyzed the relationship of YAP1 with survival. We further explored the anti-tumor role in NPC cell lines using YAP1 siRNA technique, and checked whether YAP1 regulates epithelial–mesenchymal transition (EMT). The relationship between HBV X protein (HBx) and YAP1 was also tested using Dual-Luciferase reporter assay. Finally, we explored anti-YAP1 to inhibit tumor metastasis using the xenograft mice model.

Results: In the current study, we found that YAP1 expression was higher in HBsAg (+) samples than in the HBsAg (-) samples, as a clinical signature, suggesting that YAP1 could be used as a prognostic factor for NPC. Our results showed that the HBx could regulate YAP1, further promoting cellular invasiveness through EMT. Anti-YAP1 can also decrease metastasis in vivo.

Conclusion: Our findings suggest that YAP1 is a promising prognostic factor in NPC and could be used as a potential treatment target for NPC with HBV infection.

Keywords: YAP1, hepatitis B, HBx, nasopharyngeal carcinoma, metastasis
NPC patients with HBV infection exhibit a poor prognosis, and presently published data are not very clear.

The Hippo pathway has been shown to be critical in tumorigenesis, and recent data from The Cancer Genome Atlas (TCGA) strongly support that this pathway affects overall cancer survival.10 Yes-associated protein 1 (YAP1), a novel and significant protein in the Hippo pathway, is critical in tumor cell proliferation and invasion.11–13 Increasing evidence support that epithelial–mesenchymal transition (EMT) was required for epithelial cells to acquire malignant ability. Shao et al’s findings showed YAP1 can trigger transcriptional regulation of EMT as a significant component of oncogenic RAS signaling.11 A study from Omori et al showed that co-overexpression of YAP1, SOX2 and CD44v9 was closely associated with poor prognosis for head and neck cancer, and these molecular might induce cancer stem cell, which was related to EMT and angiogenesis.12 In another study, YAP1 was proven to drive the metaplastic transformation of squamous cell carcinoma to spindle cell carcinoma and its mechanism was that YAP1 could induce ZEB1 and EMT.13 Recently, our analysis of the signature of key distant metastasis proteins, using a proteomic approach with the reverse phase protein array (RPPA) platform, showed that YAP1 protein was highly expressed in metastatic patients.14 Furthermore, we analyzed and compared differential expression in NPC, with or without HBsAg, and found that YAP1 was highly expressed in HBsAg (+) patients. Therefore, we hypothesize that YAP1, as an oncogene, acts as a distant metastasis promoter in HBsAg (+) NPC. Understanding how YAP1 promotes metastasis could give a hint for new therapeutic approaches in NPC treatment.

In the current study, we explore the regulation mechanisms of YAP1 in NPC with HBV infection. Hepatitis B virus X protein (HBx) encoded by HBV is essential in primary hepatocellular carcinoma.15,16 This is the first study to demonstrate that HBx can regulate YAP1 in NPC patients with HBV infection, and mechanically, YAP1 promotes NPC cell invasion through EMT. Our findings show that YAP1 could serve as a promising prognostic and treatment target for NPC with HBV infection.

Materials and Methods

Cell Culture

Human NPC cell lines (CNE1 and SUNE1) used in this study were obtained from the Cancer Center of Sun Yat-Sen University (Guangzhou, China). The use of the cell lines was approved by the Ethics Committee of the First People’s Hospital of Foshan (Guangdong Province, China). Cells were cultured using RPMI-1640 medium with 10% fetal bovine serum, 0.5% penicillin–streptomycin sulfate in 5% carbon dioxide humid incubator as in previous studies.17,18

Ethics Statement

Animal experiment was approved from the Institutional Animal Care and Use Ethics Committee of the First People’s Hospital of Foshan, following guidelines for the welfare of the laboratory animals. All human samples were collected by following the protocols of the Ethics Committee of the First People’s Hospital of Foshan (Guangdong Province, China). And all patients provided written informed consent, in accordance with the Declaration of Helsinki. According to ethical and legal standards, all patient tissue samples and clinical data were anonymously processing.

RPPA Data and ONCOMINE Database Analysis

We compared the difference in YAP1 expression between HBsAg (+) and (-) groups based on proteomics analysis using an RPPA platform, which has been described previously.14 The ONCOMINE database (www.oncomine.com/nasopharyngeal cancer) was used to analyze yap gene expression with clinical TNM staging.

Clinical Data and Tissue Samples

We collected two independent databases for the study of YAP1, including 6 normal nasopharyngeal mucosa samples and 25 NPC diagnosed patients with HBsAg (+). To further confirm YAP1 as a prognostic factor for NPC survival, 70 formalin-fixed paraffin-embedded (FFPE) patient specimens (35 metastatic cases and 35 non-metastatic) were analyzed for YAP1 expression. All clinical data of patients are summarized in Table 1. Clinical staging was conducted according to the staging system of the American Joint Committee on Cancer (AJCC, 7th edition).

Plasmid and siRNA Transfection

Cells (1×10^5 cells/well) were seeded into 6-well plates 24 h before transfection. Then, 120 nM–160nM of YAP1 siRNA or control siRNA oligonucleotides (with/without 1–4 μg of HBx plasmid), or 6 μg of Myc-YAP1-pcDNA plasmid were transfected using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) protocol, transfected cells were culture in RPMI-1640 with 10% FBS and no
penicillin–streptomycin for 48 h. The siControl and YAP1 siRNA (YAP1-homo-1355) were purchased from Shanghai GenePharma Co., Ltd.

Establishment of shRNA-Stable Cells
YAP1 shRNA oligonucleotides were cloned into a retrovirus system (Clontech, Mountain view, CA, USA) to establish YAP1 knockdown stable cell line. In short, co-transfected with a YAP1 shRNA-vector DNA with vectors pLP and pVSVG into the 293T-packaging cell. Forty-eight hours after transfection, the supernatant was collected, then further to infect the target NPC cells. Stable clones were selected with 14 days of 0.6 μg/mL puromycin treatment, and further confirmed the positive clone by Western blot. At last, positive clones were maintained in 0.2 μg/mL puromycin culture condition.

Cell Viability Assay and Colony Formation
We evaluate relative cell viability using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Briefly, NPC cells were plated into 96-well plates overnight, 48 h after si-YAP1 with or without HBx plasmid transfection. The spectrophotometric absorbance of the samples was read after the MTT labeling reagent was added. The IC50 values were evaluated by using GraphPad Prism 4 (GraphPad Software, La Jolla, CA, USA).

To analysis whether YAP1 or HBx affect growth ability, SUNE1 and CNE1 cells (400 cells per well) were treated with siYAP1 with or without HBx plasmid transfection 12 h after cells seeded into 6-well plates. Cells were fixed, stained after 10 days of culturing. And colonies (50 or more cells) were counted with an inverted microscope.

Wound-Healing Assay
siYAP1 and HBx DNA plasmid transfection were performed 12 h after SUNE1 and CNE1 cells were plated into 6-well plates. The next day, a scratch (across the center of the cell plate) was made by using a sterile 200-μL-pipette tip, and then incubated for 24 h or 48 h. Images of the cells were acquired and the migration distance was measured using phase-contrast optics. The scratch wound widths were defined as the percentage of cells that could migrate into the scratch area.

Cell Invasion Assays
We evaluated the tumor cell invasion ability by using a Transwell Chamber (Corning Incorporated, Life Sciences, USA) with 8-μm pores coated with Matrigel (BD, Biosciences, USA). Cells were treated with siYAP with or without HBx infection 24h before plated into the upper chamber, and a higher percentage of fetal bovine serum (10%) was placed in the lower chamber. The plates were incubated for 24 h, then fixed with 10% methanol and 0.1% crystal violet for 15 min at room temperature, the images were acquired for further analysis.

Western Blot
Total cell lysates were extracted and then added on 10% SDS-PAGE to separate the targeting protein, next, transferred to nitrocellulose polyvinylidene difluoride membranes, then incubated with primary polyclonal antibodies overnight (anti-YAP1 #8418,1:1000 dilution, anti-Snail, #3879,1:800 dilution, anti-E-cadherin #14472,1:1000 dilution, anti-N-cadherin #13116,1:1000 dilution, anti-LATS1 #3477,1:700 dilution, anti-MST1 #14946,1:700 dilution; Cell Signaling Technology, USA). The next day, it was probed with secondary antibodies before detecting for the blots, the later one was using the chemiluminescence system (Bio-Rad, CA, USA).

Table 1: Patients Characteristics in the Current Study

| Characteristics | YAP1 Expression | P |
|-----------------|-----------------|---|
|                 | Low (N=40)      | High (N=30)   |
| **Sex**         |                 |               |
| Male            | 36 (90.0)       | 23 (76.7)     | 0.118 |
| Female          | 4 (10.0)        | 7 (23.3)      |     |
| **Age (years)**|                 |               |
| <50             | 22 (55.0)       | 15 (50.0)     | 0.431|
| ≥50             | 18 (45.0)       | 15 (50.0)     |     |
| **T category**  |                 |               |
| T1              | 1 (2.5)         | 1 (3.3)       | 0.895|
| T2              | 11 (27.5)       | 6 (20.0)      |     |
| T3              | 10 (25.0)       | 9 (30.0)      |     |
| T4              | 18 (45.0)       | 14 (46.7)     |     |
| **N category**  |                 |               |
| N0              | 7 (17.5)        | 2 (6.7)       | 0.291|
| N1              | 8 (20.0)        | 10 (33.3)     |     |
| N2              | 20 (50.0)       | 12 (40.0)     |     |
| N3              | 5 (12.5)        | 6 (20.0)      |     |
| **TNM staging**|                 |               |
| I               | 0 (0.0)         | 0 (0.0)       | 0.301|
| II              | 5 (12.5)        | 1 (3.3)       |     |
| III             | 14 (35.0)       | 9 (30.0)      |     |
| IVA-B           | 21 (52.5)       | 20 (66.7)     |     |
RNA Extraction and Quantitative RT-PCR (qRT-PCR)

Total RNA was extracted using RecoverAll™ Total Nucleic Acid Isolation Kit (Thermo Fisher Scientific, CA, USA), and collected using TRIzol (Ambion, Invitrogen, Carlsbad, CA, USA), by following the manufacturer's protocol. qRT-PCR was performed using Power SYBR-Green PCR master mix (Applied Biosystems Inc.) to detect mRNA levels of YAP1 and HBx; glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used to normalize the YAP1 and HBx values. The cycling instruction was as follows: 2 min of 50°C and 95°C for 10 min followed by 40 cycles at 95°C for 15 sec and 1 min of annealing/extension at 60°C. U6 snRNA or glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used to normalize the YAP1 and HBx values, respectively. Data were analyzed by the 2^−ΔΔCq method to determine relative changes in gene expression. The primers for PCR amplification were as follows: YAP forward, 5'-GGGTGTTCATCCATTCTC-3'; YAP reverse, 5'-CTCTGTTGTATCATCC-3'; HBx forward, 5'-GTGGGATGAGCACGAC-3', and HBx reverse, 5'-TACGAC CAGAGGCATAACAGG-3'.

Dual-Luciferase Reporter Assay

SUNE1 cells were transfected using pGL3-Basic (0.2 ng/well) or reporter constructs of the YAP gene, as indicated (pGL3-348, 0.2 ng/well). Relative luciferase activity results were obtained. SUNE1 cells were co-transfected with pGL3-348 (0.2 ng/well) and HBx expression plasmid (pCMV-HBx, 0.2 or 0.3 ng/well) or control pcDNA (0.2 ng/well), respectively. The promoter activities of YAP were measured. All experiments were conducted in triple independent experiments. Primers for YAP promoter (5'-3'):

pGL3-348 forward CGGGGTATCCAGGCGACGGTCACGT

Immunohistochemical Analysis

A total of 101 FFPE human specimens (6 normal nasopharyngeal mucosa samples, 25 NPC patients diagnosed with HBsAg (+), 70 NPC cases with intact clinical data and median 5-year follow-up) were analyzed with each specimen being de-paraffinized and sectioned. Brieﬂy, the sections were incubated with primary antibodies against YAP1 (#8418,1:200 dilution, Cell Signaling Technology, USA) and HBx (#ab39716, 1:50 dilution, Abcam, MA, USA) at 4°C overnight, then incubated with the biotinylated secondary antibody, at last subjected to the Liquid DAB+ Substrate Chromogen System (Dako, Carpinteria, CA). At least 500 tumor cells (NPC) or normal cells (nasopharyngeal inflammation) were counted for YAP1 and HBx expression evaluation in five representatives’ high-power microscopy fields. Nuclear or cytoplasmic yellow or brown staining was considered positive expression. The proportion score represented the positive staining cells as follows: 0 = less than 5%; 1 = 5% to 25%; 2 = 25% to 50%; 3 = greater than 50%. The intensity score represented the estimated average staining as follows: 0 = no staining; 1 = weak staining; 2 = intermediate staining; 3 = strong staining. The overall protein expression defined as the average of the proportion and intensity scores (ranges = 0, 1, 2–4 and 6 to 9, respectively). We determined negative (−), weakly positive (+), positive (++) and highly positive (+++) if the scores were 0, 1, 2–4, and 6–9, respectively.

Xenograft Tumors in Nude Mice

Animal experiments were performed following the guidelines of the Institutional Animal Care and Use Ethics Committee of the First People’s Hospital of Foshan, Guangdong Province, China. Eight-week-old female BALB/c nude mice were purchased from the Medical Experimental Animal Center of Guangdong Province (Guangzhou, China). To create a mice metastasis model, SUNE1 cells (200 μL PBS containing 1×10⁶ SUNE-1 cells) stably expressing shYAP1 or control shRNA (with or without HBx DNA plasmid, respectively) was injected into mice tail vein (n=6 per group). Six weeks later, mice were euthanized, and then their lungs were collected. All lung samples were then paraffin-embedded, sectioned and stained for morphology, and also anti-Snail (#sc271977, 1:100 dilution, Santa Cruz Biotechnology, USA), anti-E-Cadherin (#14472,1:200 dilution, Cell Signaling Technology, USA), anti-Vimentin (#5741,1:200 dilution, Cell Signaling Technology, USA), anti-TWIST (#ab49254, 1:80 dilution, Abcam, MA, USA) and anti-HBx (#sc373890, 1:50 dilution, Santa Cruz Biotechnology, USA) immunohistochemistry staining was performed in these samples.

Statistical Analysis

For only two groups, Student’s t-test was used for statistical analysis of the results. The characteristic of the high YAP1 and low YAP1 expression were compared using the chi-square test. Kaplan–Meier method was used to analyze
actuarial survival rates and the Log rank test was used to compare differences in survival. Pearson’s correlation coefficient was used to analyze the association between YAP and HBx expression. Two-tailed P values <0.05 were considered statistically significant. Statistical analysis was performed using SPSS software (version 16.0, IBM Corporation, Armonk, NY).

**Results**

**YAP1 Expression in NPC Tissue with HBV Infection Correlated with Poor Prognosis**

We assessed YAP1 expression using IHC analysis, which turned out that YAP1 expression was much higher in NPC tissue (n=25) than normal nasopharyngeal mucous epithelium (n=6, p=0.035, Figure 1A and C), which was consistent with results from the RPPA data reported previously. Due to the sample limitation, YAP1 expressions in HBsAg (+) NPC (n=5) show no statistic significant compare to HBsAg (-) NPC (n=65) (p=0.557, Supplementary Figure 1A); however, we could see a tendency that a higher YAP1 expression in HBsAg (+) NPC. Furthermore, we found that HBx expression positively correlated with YAP1 expression (Pearson correlation=0.396; p=0.027, Figure 1C), which gave us a clue that YAP1 expression is positively related to HBsAg (+) NPC. What is more, survival analysis showed that high expression of YAP1 was likely associated with poor overall survival (39.7% vs.74.3%, p=0.043, Figure 1D) and also show right survival tendency in progression-free survival and metastasis-free survival (Supplementary Figure 1B and C). The significance of positive finding suggests that YAP1 might also serve as an effective predictive biomarker for NPC.

**YAP1 Inhibition Reduced Tumor Proliferation, Migration and Invasion**

Next, bio-function of YAP1 in NPC cells was explored. We analyzed whether RNAi-mediated YAP1 inhibition could replicate the tumor-suppressive effect of SUNE1 and CNE1 cells. Our results showed that cell growth and colony formation ability in NPC cells were significantly inhibited when YAP1 knockdown (Figure 2A and B, *p<0.05). And Wound-healing and invasion assays showed that SUNE1 and CNE1 cells migrated at a significantly slower rate when YAP1 levels were decreased through RNA interference comparing to the control groups (Figure 2C, *p<0.05, **p<0.01). SUNE1 and CNE1 cells had fewer invasions in the lower chamber after transfected with YAP siRNA, and the invasion rate of each cell was 50% lower compared to its

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**Figure 1** High YAP1 expression in NPC tissue with HBV infection correlated with poor prognosis.

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Figure 2 YAP1 inhibition reduced NPC cell proliferation, migration and invasion.
control group (Figure 2D, *p<0.05, **p<0.01). These results proved that YAP1 inhibition could reduce NPC cell proliferation, migration, and invasion, which indicate that targeting YAP1 could suppress NPC growth and invasion.

YAP1 promotes tumor metastasis through regulating the EMT pathway

To further explore the pathway regarding YAP1 mediated NPC metastasis, we evaluated EMT related markers by performing PCR and Western blot analyses. The expression levels of EMT key regulators (LATS1, MST1, Snail, E-cadherin, and N-cadherin) were detected after NPC cells were treated with siYAP1 or YAP1 DNA plasmid. With the depletion of YAP1, MST1 and E-cadherin increased, while LATS1 decreased in both CNE-1 and SUNE1 cell lines, and N-cadherin decreased were detected only in CNE1 cell, E-cadherin and Snail increased only in SUNE1 cell line (Figure 3A and B, *p<0.05, **p<0.01). Meanwhile, as YAP1 exogenously increased (Figure 3C and D), MST1 and E-cadherin expressions were decreased in CNE1 cell; however, LATS1 still decreased in both CNE1 and SUNE1 cell lines, and Snail expression showed no marked difference between the two treatments. Nevertheless, differential expression of the EMT markers, after alteration of YAP1, indicates that the regulatory role of YAP1 in NPC metastasis may be through EMT.

HBx, correlated with YAP1, promotes tumor proliferation, migration and invasion

In our previous study, we had found that HBsAg (+) patients account for 10.3% of the NPC group and had a poor survival rate if the HBsAg (+) patient showed a high level of pretreated EBV DNA; therefore, we explored the role of HBx in NPC cell lines. We found that HBx plasmid transfecion could promote cell growth and colony formation in SUNE1 and CNE1 cells, even after treatment with YAP1 siRNA (Figure 4A and B, *p<0.05, **p<0.01). Interestingly, this procedure could restore NPC cell’s YAP1 expression. By using an in vitro invasion and migration assay, we also found a significant increase in the rate of NPC cell invasion and migration under these conditions (Figure 4C, *p<0.05, **p<0.01). Additionally, with an increase of HBx, YAP1 was highly expressed (Figure 5A), while transfected HBx with si-YAP1 together, LATS1 decreased, and MST1 increased in both cell lines, but Snail increased only in SUNE1; however, E-cadherin and N-cadherin expression showed no difference in CNE1 and SUNE1 (Figure 5B and C, *p<0.05, **p<0.01). Notably, HBx promoted NPC cell invasion through EMT, which correlated with YAP1. The effects of HBx and YAP1 modulation suggest that targeting of HBx and/or YAP1 could regulate NPC growth, invasion and migration.

Anti-YAP1 inhibits tumor cell metastasis in vivo, and HBx directly up-regulates YAP1 expression through binding to its promoter region

We established metastasis in a mouse model by injecting sh-YAP1 SUNE1 cells or control shRNA (with or without HBx DNA plasmid, for each group, respectively) into nude mice’s tail veins, and found lung metastasis colonization was remarkably suppressed in both the shYAP1 and shYAP1 with HBx groups, only one in six mice was found, respectively, and in the vehicle group, three in six mice were found with lung metastasis colonization, while treatment with HBx found four in six mice with lung colonization sites (Figure 6A and B). We performed IHC analysis for vimentin and E-cadherin expression of the lung metastasis colonization site of mice, vimentin and E-cadherin expression were decreased in ShYAP1 group compared to the vehicle group, while with HBx exogenously increase could enhance vimentin and E-cadherin expression significantly; however, in ShYAP1 with HBx exogenously increase group, vimentin and E-cadherin expression showed no such increasing level compared to the vehicle with HBx group (Figure 6C). Thus, we concluded that HBx could induce metastasis, while anti-YAP1 could inhibit tumor cell metastasis in vivo.

To further delineate the interaction of HBx with YAP1, we performed a Dual-Luciferase reporter assay. Literature indicated that the YAP promoter was located at nt 2232/1115. Thus, we attempted to confirm this in our model. Cotransfection of HBx expression plasmid (pCMV-X, 0.2 ng/well), pGL3-YAP1 or pGL3-Basic control (0.2 ng/well) was performed in SUNE1 cells. Relative luciferase activity of pGL3-YAP1 (pGL3-348) increased by 5- or 10-fold on average by HBx in the HBx and YAP1 gene cotransfection group. However, the pGL3-Basic control construct was not activated by HBx, let alone the pGL3-Basic control or pGL3-YAP1 (pGL3-348) group (Figure 6D). Consistent with the study before, we proved that the regulation site of HBx is located −348 of YAP1 in NPC.
Figure 3 YAP1 promotes NPC metastasis by regulating the EMT pathway.
Figure 4 HBx promotes NPC cell proliferation, migration and invasion, correlated to YAP1.
Figure 5 HBx regulate YAP1 expression and EMT.
Collectively, we concluded that YAP1 is a driving factor in NPC cell metastasis, and HBx plays a promoting role in this process, but the interaction between these two markers needs further investigation.

**Discussion**

Human viral infection has increasingly been established as playing a significant role in the hallmarks of cancer oncogenesis. Within the endemic area of NPC, EBV associated non-keratinizing squamous cell carcinoma has accounted for approximately 95%, and interestingly, 10.3% of patients were infected with chronic HBV. However, its molecular mechanism remains unknown. In the current study, we first revealed a higher YAP1 expression in NPC with HBV infection, and showed that HBx could regulate YAP1, which further increasing NPC cell invasion through the EMT process. Our findings demonstrate that YAP1 could be used as a promising prognostic and treatment target for NPC with HBV infection.

**Figure 6** Anti-YAP1 inhibits tumor cell metastasis in vivo, and HBX directly up-regulates YAP1 expression through binding to its promoter region.
YAP1, a key downstream effector of the Hippo pathway, functions as a transcriptional co-activator. YAP1 is highly expressed in various types of cancers, which could promote cell proliferation and invasion. Evidence indicates that YAP1 has great clinical significance. According to TCGA data analyses, its expression was correlated with survival in gastric, lung, colorectal and hepatocellular cancers. YAP1 could drive cell proliferation in oral squamous cell carcinoma and promote glycolysis in the progression of NPC. ONCOMINE database analysis confirmed that gene expression of YAP1 was increasingly associated with T advanced and N advanced staged NPC tissue, as Supplementary Figure 2 data shown. High expression of YAP1 was closely associated with poor survival, based on our previous RPPA proteomic profiling, which we confirmed in the current study by using IHC to determine YAP1 and HBx differential protein expression and clinical prognosis within the patient sample groups. As expected, cell proliferation, migration and invasion were decreased when YAP1 expression inhibited. These results suggest that YAP1 is a major driver protein in NPC metastasis. However, the mechanism underlying the promotion of NPC metastasis by YAP1 has not yet been elucidated, which leads us for further investigation.

The Hippo pathway is crucial for organ development and homeostasis maintenance. The key members of the mammalian Hippo pathway include a kinase cascade of mammalian sterile 20-like kinase 1/2 (MST1/2) and large tumor suppressor kinase 1/2 (LATS1/2). MST1/2 interact with and phosphorylate LATS1/2 kinase. Active LATS1/2 kinase inactivates YAP1 and PDZ-binding motif (TAZ) to affect their localization. Considering YAP1 is a key component of Hippo pathway, and its important function in homeostasis maintenance, as shown in our results, YAP1 expression could influence MST1/2 and LATS1/2, which lead us to wonder how YAP1 function in homeostasis maintenance. It is reported that YAP1 promotes tumor proliferation by targeting important genes including Ki-67, SOX4, H19 and c-MYC, the latter one was positively related to EMT. Our RPPA data also indicated that YAP1 was strongly correlated with c-MYC expression (Supplementary Figure 1D), which is consistent with those studies. Indeed, we showed that anti-YAP1 inhibits NPC cell migration and invasion by regulating EMT. To further investigate the underlying mechanisms, we attempted to identify potential targets of YAP1 in regulating NPC metastasis. HBx reportedly plays an important role in hepatocellular carcinoma and can up-regulate YAP1 expression. Interestingly, a similar phenomenon of HBx regulating YAP1 has also been observed in NPC cell lines. We confirmed that YAP1 was a direct target of HBx by using luciferase reporter assay, which could prove our hypothesis. Thus, we can further conclude that YAP1 induces cell invasion and migration through the EMT axis, which could be mediated by HBx.

During tumor progression, EMT could trigger aggressive features including cell migration, invasion, and treatment resistance. EMT is a factor in cancer metastasis. Hallmarks of EMT are including loss of E-cadherin, increase of mesenchymal and extracellular matrix biomarkers N-cadherin, and vimentin. Varieties of signaling pathways are involved with EMT regulation, including TGF-β/Snail, GSK3β, EGFR, and ERK/MAPK. It is reported that YAP/TAZ signaling plays an important role in EMT regulation. YAP/TAZ activation could promote cell proliferation and EMT. YAP1 could activate Slug and vimentin-mediated EMT with the transcription factor FOS. Weiss et al found that Snail/Slug-YAP/TAZ complexes could modulate self-renewal and cell differentiation. Crosstalk between YAP/TAZ and EMT transcription factors like Snail/Slug and Twist could promote cancer cell EMT process, which further acquires malignant features.

In our study, we showed that anti-YAP1 inhibits cell migration and invasion, and with E-cadherin highly re-expressed, while LATS1 and MST1 showed a little decrease. However, when YAP1 expression exogenously increased, we found that E-cadherin expression loss, LATS1 and MST1 decreased, together with Snail expression level increasing. All those evidence lead us to conclude that YAP1 promotes metastasis by regulating EMT in NPC cells. Further, we had identified HBx is the potential target of YAP1 in regulating NPC metastasis. HBx has been reported to take an important role in hepatocellular carcinoma, and it could up-regulate YAP1 expression. HBx could dysfunction intercellular adhesion, and induce EMT and metastasis in HCC. Research showed that HBx could induce HMGA2 by regulating EMT, subsequently leading to HCC cell invasion. These evidence suggest that HBx might regulate YAP1 in the EMT process. Indeed, we proved that HBx regulated promoter region of YAP was located at nt –348 through luciferase reporter gene assays. And with HBx increasing expression, NPC cell invasion and migration ability enhanced. Our study firstly reported that YAP1 induces cell invasion and migration through EMT axis, and could be mediated by HBx.
activity in NPC, which could provide a new strategy for NPC metastasis clinical treatment thinking.

In summary, the present study demonstrated that YAP1 promotes metastasis through the regulation of the EMT axis which can be induced by HBx. Our findings offer new therapeutic approaches for NPC treatment and serve as a promising prognosis and treatment target for NPC patients with HBV infection.

Acknowledgment
Zeli Huang, Bojin Su, and Fang Liu are co-first authors in this study.

Funding
This work was financially supported by grants from the National Natural Science Foundation of China (No.81402244 [to Ying Huang] and 81602392 [to Tao Xu]), a Special Fund Project for 2016 University of Texas MD Anderson cancer center sister hospital network fund (SINF).

Disclosure
The authors report no conflicts of interest in this work.

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