Clinical influence of switching companion diagnostic tests for EGFR-TKIs from Therascreen to Cobas v2

Ken Uchibori1, Natsuki Takano1,2, Ryo Manabe1, Ryosuke Tsugitomi1, Shinsuke Ogusu1, Takehiro Tozuka1, Hiroaki Sakamoto1, Hiroshi Yoshida1, Yoshiaki Amino1, Ryo Ariyasu1, Satoru Kitazono1, Noriko Yanagitani1 & Makoto Nishio1

1 Department of Thoracic Medical Oncology, The Cancer Institute Hospital of Japanese Foundation for Cancer Research, Tokyo, Japan
2 Department of Pulmonary Medicine and Oncology, Graduate School of Medicine, Nippon Medical School, Tokyo, Japan

Keywords
Companion diagnostic (CDx); EGFR mutation; EGFR-TKI; next-generation sequencing (NGS); polymerase chain reaction (PCR).

Correspondence
Makoto Nishio, Department of Thoracic Medical Oncology, The Cancer Institute Hospital of Japanese Foundation for Cancer Research, 3-8-31 Ariake, Koto-ku, Tokyo 135-8550, Japan.
Tel: +81 3 3520 0111
Fax: +81 3 3520 0141
Email: mnishio@jfcr.or.jp

Received: 11 October 2020;
Accepted: 7 December 2020.
doi: 10.1111/1759-7714.13797

Thoracic Cancer 12 (2021) 906–913

Abstract

Background: Several companion diagnostic (CDx) tests for epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs) have been approved. In our institute, the CDx test for EGFR-TKIs was changed from the Therascreen test (Therascreen) to the Cobas EGFR v2 test (Cobas) because only Cobas was approved for the use of osimertinib in patients with EGFR-mutated non-small cell lung cancer (NSCLC) with T790M mutations. The clinical influence of switching the CDx test has not yet been examined comprehensively.

Methods: All serial patients with lung cancer tested for EGFR mutations with CDx tests between February 2014 and February 2016 at the Cancer Institute Hospital of the Japanese Foundation for Cancer Research (JFCR) were enrolled in this analysis.

Results: Therascreen was used as a CDx test for EGFR-TKI therapy in 607 patients between February 2014 and January 2015, and Cobas was used in 621 patients between February 2015 and February 2016. EGFR mutations were detected in 218 patients (35.9%) and 244 patients (39.3%) tested with Therascreen and Cobas, respectively. At the initial diagnosis, 400 and 459 patients were tested with Therascreen and Cobas, respectively. EGFR mutation subtypes, including del19, L858R, and others, were detected in 13.0%, 17.0%, and 2.5% of patients using Therascreen and 17.4%, 14.4%, and 1.5% of patients using Cobas, respectively.

Conclusions: No significant impact of switching from Therascreen to Cobas as the CDx test for EGFR mutations in clinical practice was observed. However, the detection pattern of the EGFR mutation subtypes between the two CDx tests was slightly different.

Key points
Significant findings of the study: We examined the influence of changing the EGFR test in 1228 patients in total. The detection rate of EGFR mutations was similar. However, the detection pattern for EGFR subtype mutations was slightly different between the two tests.
What this study adds: Switching CDx tests from target polymerase chain reaction (PCR)- to next-generation sequencing (NGS)-based methods may lead to obvious changes in clinical practice. When the CDx test is required to change, the investigation of this influence is warranted in future studies.
Introduction

Epidermal growth factor receptor (EGFR) gene-activating mutations are the primary oncogenic drivers in lung adenocarcinoma. The overall survival of patients with EGFR-mutated lung cancer has improved remarkably from one year to approximately three to four years after the introduction of EGFR tyrosine kinase inhibitors (EGFR-TKIs). The detection of EGFR-activating mutations using companion diagnostic (CDx) tests is mandatory to initiate treatment with EGFR-TKIs. Variable EGFR tests are used in clinical practice in Japan. Therascreen, Cobas EGFR v2, and Oncomine CDx are the CDx tests used for EGFR therapy that were approved in 2011, 2016, and 2019, respectively. These CDx tests each have a unique profile in terms of the detection method and degree of specificity for EGFR gene mutations (Table S1), especially EGFR exon 19 deletions (del19), which have been shown to have many patterns in DNA sequences, leading to the identification of ~30 types.

Therascreen (Therascreen) can detect EGFR gene mutations using a polymerase chain reaction (PCR)-based Scorpion-ARMS method that covers three types of G719X, 19 types of del19, three types of exon 20 ins, S768I, T790M, L858R, and L861Q, with a detection sensitivity of 1%–10%. Cobas EGFR v2 (Cobas) can detect EGFR gene mutations using a PCR-based Cobas method that covers three types of G719X (exon 18), 29 types of del19, five types of exon 20 ins, S768I, T790M, two types of L858R, and L861Q, with a detection sensitivity of 3%–5%. Importantly, Cobas is the only CDx test for T790M mutations to prescribe osimertinib. Oncomine CDx (Oncomine) was the first approved CDx test based on next-generation sequencing (NGS) in Japan, and it targets multiple oncogenes including EGFR, ALK, ROS1, and BRAF. The profile of EGFR mutations detected by Oncomine includes four types of G719X (exon 18), 21 types of del19, three types of exon 20 ins, S768I, T790M, two types of L858R, L861Q, and L861R, with a detection sensitivity of 6%–8%.

The results of testing EGFR mutations might depend on which CDx test is used, and it is essential to determine the influence of switching the CDx test in clinical practice. In our institute, we were evaluating EGFR mutations with Therascreen, which was approved as a CDx test for EGFR-TKIs in January 2013. In February 2015, we switched from Therascreen to EGFR-Cobas for EGFR mutation testing when osimertinib was approved for the treatment of patients with T790M-positive lung cancer who relapsed on prior EGFR-TKI therapy because only Cobas was approved as a CDx test for the use of osimertinib in patients with T790M mutations. The details of detectable del19 mutations are different between Therascreen and Cobas. Therascreen and Cobas do not cover all types of del19 mutations, and there are some differences in the mutations covered by these tests, as shown in the package inserts. Detectable uncommon mutations, such as G719X and ex 20 insertions, are more relevant in testing with Cobas than with Therascreen. The main objective of this study was to compare the frequency of EGFR mutations and the distribution of mutation subtypes after the replacement of Therascreen with Cobas.

Methods

We analyzed all patients who were tested for EGFR mutations between February 2014 and February 2016 at the Cancer Institute Hospital of the Japanese Foundation for Cancer Research (JFCR). Therascreen was used to detect EGFR mutations from February 2014 to January 2015, and Cobas was used to detect EGFR mutations from February 2015 to February 2016. Cytology samples or tissue samples were obtained by transbronchial biopsy, computed tomography (CT)-guided biopsy, and surgery. All samples were analyzed at the CLIA certified commercial laboratories SRL and LSI using Therascreen and Cobas, respectively.

We evaluated the frequency of each EGFR mutation type, including activating mutations (exon 19 deletion [del19] and exon 21 point mutations [L858R]), uncommon mutations (G719X, S768I, L861Q/R, and exon 20 ins), and T790M, according to the detection methods Therascreen and Cobas using a subgroup analysis involving all participants, overall patients at the initial diagnosis, adeno/non-adeno subtypes at the initial diagnosis, and rebiopsy at failure on EGFR-TKIs. The difference in background characteristics and the detection frequencies of subtypes between the two groups was calculated statistically using the chi-squared test. Differences in the median age were analyzed with the Mann-Whitney U test. The GraphPad prism 7 (GraphPad Software, San Diego, CA) was used for statistical analysis. Differences were considered statistically significant at $P < 0.05$.

The study protocol was reviewed and approved by the Ethics Committee of the Cancer Institute Hospital, JFCR (IRB number: 2020–1139).

Results

Patient characteristics

Of 1287 patients who underwent EGFR mutation tests, 607 patients were examined with Therascreen and 621 patients were examined with Cobas. The remaining 59 patients were excluded because they had malignancies other than lung cancer (Fig 1). The patient characteristics of all tested cases are shown in Table 1. A few cases were
doubly counted because the plural time of tests was examined for these cases in the study period. The patients' age and the timing of testing were statistically different between the two groups, whereas other factors, including sex, smoking history, pathology, stage, PS, and tested specimens, were similar.

The detection rate of common EGFR mutations (del19 and L858R) was 32.8% in the Therascreen group and

---

**Figure 1** Patient flow diagram. Adeno, adenocarcinoma; Initial test, EGFR test at the initial attempt; Nonadeno, histology other than adenocarcinoma; ≥2 times, EGFR test at the second attempt or over.

**Table 1** Patient characteristics

|                | Therascreen (N = 627) | Cobas (N = 621) | P (χ²) |
|----------------|-----------------------|-----------------|--------|
| Age            | Median (range)        |                 |        |
| Sex            | Male/female           |                 |        |
| Smoking history| Never/ex or current/unknown |             |        |
| Pathology      | Adeno/nonadeno        |                 |        |
| Stage          | I–III/IV or recurrence|                 |        |
| PS (ECOG)      | 0, 1/≥2               |                 |        |
| Timing of test | Initial/2nd or more   |                 |        |
| Tested specimens| Cytology/histology    |                 |        |
| EGFR status    | Wild/positive/not detected |             |        |
|                | Wild                  |                 |        |
|                | Ex19 del              |                 |        |
|                | Ex21 L858R            |                 |        |
|                | T790M (de novo/acquired) |             |        |
|                | Other                 |                 |        |
|                | Not detected          |                 |        |

Adeno, adenocarcinoma; Ex19 del, EGFR exon 19 deletion; Ex20 ins, EGFR exon 20 insertion; Ex21 L858R, EGFR exon 21 L858R point mutation; ex or current, ex or current smoker; initial, initial attempt of EGFR test; Never, never smoked; nonadeno, pathology other than adenocarcinoma; not detected: the result was not detected with EGFR test; positive, EGFR mutation-positive; wild, wild-type; second or more, EGFR test at second (or more) times.
36.7% in the Cobas group, whereas that of uncommon mutations, such as G719X and exon20 ins, was 3.1% and 2.6%, respectively. T790M mutations were detected in 3.6% and 3.2% of all tested patients in the Therascreen and Cobas groups, respectively. Only one case was a de novo T790M mutation occurring at the initial diagnosis. T790M cases were not added to the EGFR-positive cases because they were already counted as del19 or L858R-positive cases. The prevalence of EGFR mutations between the two groups did not differ significantly.

**Patient characteristics at the initial test**

Focusing on the patients whose EGFR status was evaluated at the initial test, 400 patients were examined by Therascreen and 459 patients were analyzed by Cobas (Table 2). The characteristics were similar between the two groups, with slight differences in age, pathology, and tested specimens. The proportion of adenocarcinoma was dominant in each group (75.8% in Therascreen and 79.5% in Cobas).

The proportion of del19, L858R, ex20 ins, and other minor mutations was 13.0%, 17.0%, 1.0%, and 1.5% in the Therascreen group and 17.4%, 14.4%, 0.7%, and 0.9% in the Cobas group, respectively. A de novo T790M mutation was found in only one case (0.25%) in the Therascreen group. The frequencies of del19 and L858R among EGFR mutations were 40% and 52.3% in the Therascreen group and 52.3% and 43.1% in the Cobas group, respectively.

In contrast to our expectations, del19 mutations were observed numerically but not significantly more frequently ($P = 0.073$) in the Cobas group compared with the Therascreen group, whereas the prevalence of L858R was slightly higher in the Therascreen group than in the Cobas group but without statistical significance ($P = 0.291$). The distribution of uncommon mutations was not different between the two groups ($P = 0.306$).

**Patients with adenocarcinoma at the initial test**

From the perspective of detection performance regarding common EGFR mutations in lung adenocarcinoma at the initial EGFR testing, our results showed that the detection rates of overall mutations and common mutations (del19 and L858R) were similar (42.2% vs. 41.9% and 39.3% vs. 40.0% for Therascreen and Cobas, respectively) (Table 3). The frequencies of del19 and L858R among EGFR mutations were 39.8% and 53.1% in the Therascreen group and 52.3% and 43.1% in the Cobas group, respectively. Similar to the analysis of all participants, del19 mutations in the Cobas group and L858R mutations in the Therascreen group were more prevalent numerically but not statistically significant than those in each other’s group ($P = 0.099$ and 0.161, respectively). The distribution of uncommon mutations was not different between the two groups ($P = 0.378$).

In the nonadenocarcinoma group, there were no differences in the background characteristics of patients included for each

---

**Table 2** Characteristics of all patients at the initial EGFR test

|               | Therascreen ($N = 400$) | Cobas ($N = 459$) | P-value ($\chi^2$) |
|---------------|-------------------------|-------------------|-------------------|
| Age           | Median (range)          | 67 (29–90)        | 69 (38–92)        | 0.216 |
| Sex           | Male/female             | 232/168           | 261/198           | 0.737 |
| Smoking history | Never/ex or current/unknown | 138/262/0     | 179/279/1         | 0.247 |
| Pathology     | Adeno/nonadeno          | 303/97            | 365/94            | 0.185 |
| Stage         | I–III/V or recurrence   | 291/109           | 332/127           | 0.891 |
| PS (ECOG)     | 0,1/2                   | 386/14            | 437/22            | 0.346 |
| Tested specimens | Cytology/histology     | 187/213           | 214/245           | 0.970 |
| EGFR status   | Wild/positive/not detected | 265/130/5    | 300/153/6         | 0.963 |
|               | Wild                    | 265               | 300               | 0.784 |
|               | Ex19 del                | 52 (+T790M 1)     | 80                | 0.073 |
|               | Ex21 L858R              | 68                | 66                | 0.291 |
|               | T790M                   | 1                 | 0                 | 0.795 |
|               | Other                   | G719X-1           | G719X-1           | 0.306 |
|               |                         | S768I-2           | G719X+S768I-1     |         |
|               |                         | Ex20 ins-4        | Ex20 ins-3        |         |
|               |                         | L861Q-2           | L861Q-0           |         |
|               |                         | L858R+S768I-1     | L858R+S768I-2     |         |
|               | Not detected            | 5                 | 6                 | 0.957 |

Adeno, adenocarcinoma; Ex19 del, EGFR exon 19 deletion; Ex20 ins, EGFR exon 20 insertion; Ex21 L858R, EGFR exon 21 L858R point mutation; ex or current, ex or current smoker; Never, never smoked; nonadeno, pathology other than adenocarcinoma; not detected, the result was not detected with EGFR test; positive, EGFR mutation-positive; wild, wild-type.
detection method. Only one del19 and one S768I EGFR mutation in the Therascreen group were confirmed in this cohort.

Details of rebiopsies at relapse on EGFR-TKIs

A rebiopsy is usually performed at failure on first- or second-generation EGFR-TKI therapy to evaluate the development of T790M secondary mutations. In this study, 55 and 66 cases were assessed by Therascreen and Cobas, respectively. The characteristics of patients who underwent a rebiopsy were generally similar in the clinic, although seven cases with histology results other than adenocarcinoma or squamous cell carcinoma and predominance in cytology specimens among the evaluated samples were observed in the Cobas group (Table 4).

The comparison of the detection sensitivity for T790M mutations revealed no significant difference between Therascreen (34.5% [19/55]) and Cobas (30.8% [20/65]) (P = 0.762). The proportions of T790M-negative, wild-type, and undetected tumors were similar between the groups (45.5% vs. 44.6%, 5.5% vs. 15.4%, and 7.3% vs. 3.1% for Therascreen and Cobas, respectively). The informative rebiopsy rates in this study estimated by excluding the wild-type and undetected tumors were 87.7% (48/55) and 81.5% (53/65) for Therascreen and Cobas, respectively. The detection power of T790M in informative biopsies was 39.6% for Therascreen and 37.7% for Cobas. These results suggest that the performance of the two methods is not significantly different.

Discussion

We evaluated the EGFR mutation status for the treatment of lung cancer according to the results of approved CDx tests to prescribe EGFR-TKIs. However, we usually use only one test in each institution. To the best of our knowledge, there is no report other than the current study that evaluates the influence of replacing the CDx test in clinical practice, but some reports have compared Scorpion and Cobas using identical specimens.16–20

Table 3 Patient characteristics at the initial EGFR test divided into pathological types

|                     | Adeno     | Nonadenocarcinoma | P-value (χ²) | Adeno     | Nonadenocarcinoma | P-value (χ²) |
|---------------------|-----------|-------------------|--------------|-----------|-------------------|--------------|
| Age                 | Median (range) | 66 (29–90) | 0.07 | 71 (43–87) | 0.679 |
| Sex                 | Male/female | 160/143 | 0.585 | 72/25 | 0.273 |
| Smoking history     | Never/ex or current/unknown | 128/175/0 | 0.285 | 10/87 | 0.487 |
| Stage               | I–III/IV or recurrence | 221/82 | 0.643 | 70/27 | 0.217 |
| PS (ECOG)           | 0,1/2    | 295/8 | 0.228 | 91/6 | 0.955 |
| Tested specimens    | Cytology/histology | 125/178 | 0.919 | 62/35 | 0.768 |
| EGFR status         | Wild/positive/not detected | 174/128/3 | 0.899 | 93/2/2 | 0.319 |
| Ex19 del            | Wild | 174 | 0.853 | 93 | 0.853 |
|                     | Ex19 del (+T790M) | 51 | 0.999 | 1 | 0.326 |
|                     | Ex21 L858R | 68 | 0.161 | 0 | — |
|                     | T790M     | 1 | 0.272 | 0 | — |
| Other               | G719X-1   | G719X-1 | 0.378 | G719X-0 | 0.323 |
|                     | S768I-1   | S768I-1 | — | S768I-0 | — |
|                     | Ex20 ins-4 | + S768I-1 | — | Ex20 ins-0 | — |
|                     | Ex20 ins-3 | + S768I-1 | — | Ex20 ins-3 | — |
|                     | Ex20 ins-0 | + S768I-2 | — | Ex20 ins-0 | — |
| Not detected        | 3 | 0.653 | 2 | 1 | 0.579 |

Adeno, adenocarcinoma; Ex19 del, EGFR exon 19 deletion; Ex20 ins, EGFR exon 20 insertion; Ex21 L858R, EGFR exon 21 L858R point mutation; ex or current, ex or current smoker; Never, never smoked; nonadenocarcinoma, pathology other than adenocarcinoma; not detected, the result was not detected with EGFR test; positive, EGFR mutation-positive; wild, wild-type.
As a result, no significant difference between the characteristics of patients tested with the two CDx tests was observed, except for age. The frequency of $EGFR$ mutations and the distribution of mutation subtypes did not differ significantly before and after the replacement of Therascreen with Cobas. Our results show that the frequency of $EGFR$ mutations at the initial testing among all patients and those with adenocarcinoma was 32.5% versus 33.3% and 42.2% versus 41.9% for Therascreen versus Cobas, respectively. The frequencies of del19 and L858R among $EGFR$ mutations in adenocarcinoma were 39.8% and 53.1% in the Therascreen group and 52.3% and 43.1% in the Cobas group, respectively. These results were similar to previous reports showing that $EGFR$ mutations account for 30%–40% of non-small cell lung cancer (NSCLC) and 40%–55% of lung adenocarcinoma and that del19 mutations account for 40%–49% and 46%–59%, respectively, whereas L858R mutations account for 39%–47% and 25%–38% of $EGFR$ mutations among Asians and Caucasians, respectively.21–24 The frequency of uncommon mutations among $EGFR$ mutation-positive adenocarcinoma cases (7.0% [9/128] in Therascreen and 4.6% [7/153] in Cobas) was not significantly different. It was difficult to compare the details because of the small number of cases. Only two $EGFR$-positive cases were confirmed among the nonadenocarcinoma cohort, supporting the established knowledge that $EGFR$ mutations are rarely found in nonadenocarcinoma histology.25,26

At the initial diagnosis test, we anticipated that more del19 cases might be found in the Cobas group than in the Therascreen group based on the wider coverage of Cobas for detectable del19 types. As we expected, the number of del19 mutations in the current study was larger in the Cobas group than in the Therascreen group, although this difference was not statistically significant. On the other hand, the distribution of L858R among $EGFR$ mutation-positive cases was reduced to 42.3% for Cobas from 53.1% in the Therascreen group, with no statistical significance. These disparities between the two groups in the detection rate of $EGFR$-activating mutations and the distribution of mutation subtypes stayed within the range mentioned above, suggesting that the replacement of Therascreen with Cobas has minimal influence in clinical practice. One of the reasons that switching the CDx test did not influence the detection rate in clinical practice was that both methods are mutation-specific target PCR-based CDx tests.

As cytological specimens are acceptable for PCR-based $EGFR$ testing,27 about half of the samples assessed were cytological specimens (46.8% for Therascreen and 46.6% for Cobas). The ratio of cytological samples was similar between the two groups. The combined use of cytological and histological samples did not affect the detection rate of $EGFR$ mutations for either Therascreen or Cobas. Regarding the perspective of testing at failure on $EGFR$-TKIs, we should note that cytological samples were more prevalent in the current study than those in previous studies.28,29

| Table 4 | Patient characteristics who underwent rebiopsy at relapse on EGFR-TKIs |
|---------|---------------------------------------------------------------|
|         | Therascreen ($N = 55$)                                        | Cobas ($N = 65$) | $P$-value ($\chi^2$) |
| Age     | Median (range)                                               |                |                      |
|         | 65 (34–83)                                                   | 67 (40–86)      | 0.130               |
| Sex     | Male/female                                                  | 15/40           | 24/41               | 0.261               |
| Smoking history | Never/ex or current/unknown            | 31/24/0         | 29/36/0             | 0.200               |
| Pathology | Adeno/nonadeneno                                           | 55/0            | 59/6                | 0.021               |
| P(ESOG) | 0, 1/c/2                                                   | 48/7            | 51/4                | 0.340               |
| Tested specimens | Cytology/histology                                        | 39/16           | 58/7                | 0.011               |
| EGFR status | Wild/positive/not detected                                  | 3/48/4          | 10/53/2             | 0.144               |
|         | Wild                                                       | 3               | 10                  | 0.081               |
|         | Ex19 del                                                  | 10              | 12                  | 0.969               |
|         | Ex21 L858R                                               | 15              | 17                  | 0.890               |
| T790M   | G179X + T790M                                             | 0               | G179X + T790M       | 2                   |
|         | De19 + T790ML8                                            | 15              | De19 + T790M        | 11                  |
|         | 58R + T790M                                              | 4               | L858R + T970M       | 7                   |
| Other   | G719X                                                    | 1               | G719X               | 1                   |
|         | L858R + Ex20ins                                           | 1               | Ex20ins             | 3                   |
|         | S768I                                                     | 0               | S768I               | 0                   |
|         | L861Q                                                     | 1               | L861Q               | 0                   |
|         | L858R + del19                                             | 1               |                     |                     |
| Not detected |                                                | 4               | 2                   | 0.293               |

Adeno, adenocarcinoma; ex or current: ex or current smoker; Ex19 del, $EGFR$ exon 19 deletion; Ex21 L858R, $EGFR$ exon 21 L858R point mutation; Ex20 ins, $EGFR$ exon 20 insertion; Never, never smoked; nonaden, pathology other than adenocarcinoma; not detected, the result was not detected with $EGFR$ test; positive, $EGFR$ mutation-positive; wild, wild-type.
In this study, the detection power of T790M mutations in informative biopsies was 39.6% for Therascreen and 37.7% for Cobas, which are slightly lower than those reported previously. However, the performance of the two tests was similar.

Recently, NGS-based CDx tests, such as the Oncomine CDx System and FoundationOne, have been approved for EGFR-TKIs and other molecular agents. The method used to evaluate driver oncogenes in lung cancer has been switched gradually from single-plex mutation-specific target PCR CDx tests, such as Cobas or Therascreen, to multi-plex CDx tests based on NGS. Clinical properties between PCR- and NGS-based tests may be quite different. Therefore, we should pay attention to the influence of switching from target PCR- to NGS-based CDx tests, which might lead to obvious changes in clinical practice.

Some limitations of this study should be addressed. First, this is a retrospective study from a single institution comparing two distinct patient groups. Second, the EGFR detection profiles of Therascreen and Cobas were not compared directly by assessing identical specimens. The results of this study might include selection bias. The concordance rate between Therascreen and Cobas was reported to be 98.0% when they were evaluated using identical specimens. However, these points may be permitted because the primary focus of this study was to investigate the influence of switching the EGFR test in clinical practice. We considered it preferable to compare sequential patients in a year at a single institution rather than at multiple institutions. The consistency in sample collection and handling procedures might provide advantages for this study to evaluate the clinical properties of CDx tests. Moreover, the results of this study might reflect the quality of CDx tests in the real world because variable sample sizes and samples obtained in practice were assessed, whereas a usual performance test is conducted using samples of ideal size and quality. Taken together, the results of this study provide useful information to examine the influence of switching the CDx test in clinical practice.

In conclusion, the switching of CDx tests from Therascreen to Cobas showed minimal influence in clinical practice, but the distribution pattern of EGFR mutation subtypes might differ. Switching CDx tests from target PCR- to NGS-based methods may lead to obvious changes in clinical practice. The investigation of this influence is warranted in future studies.

Acknowledgments

The authors assume full responsibility for the analyses and interpretation of these data. The authors would like to thank Enago (www.enago.jp) for the English language review.

Disclosure

K. Uchibori reports the employment of a family member at Daiichi Sankyo. M. Nishio reports honoraria from Ono Pharmaceutical, Bristol-Myers Squibb, Pfizer, Chugai Pharmaceutical, Eli Lilly, Taiho Pharmaceutical, AstraZeneca, Boehringer-Ingelheim, MSD, and Novartis, research funding from Novartis, Daiichi Sankyo, Taiho Pharmaceutical, Bristol-Myers Squibb, Boehringer-Ingelheim, Ono Pharmaceutical, Eli Lilly, Chugai Pharmaceutical, AstraZeneca, Merck Serono, MSD, and Pfizer. N. Yanagitani reports honoraria from Chugai Pharmaceutical. All the other authors have stated that they have no conflicts of interest.

References

1. Kohno T, Tsuta K, Tsuchihara K, Nakaoku T, Yoh K, Goto K. RET fusion gene: Translation to personalized lung cancer therapy. Cancer Sci 2013; 104 (11): 1396–400.
2. Giacone G. Epidermal growth factor receptor inhibitors in the treatment of non-small-cell lung cancer. J Clin Oncol 2005; 23 (14): 3235–42.
3. Takano T, Fukui T, Ohe Y et al. EGFR mutations predict survival benefit from gefitinib in patients with advanced lung adenocarcinoma: A historical comparison of patients treated before and after gefitinib approval in Japan. J Clin Oncol 2008; 26 (34): 5589–95.
4. Inoue A, Kobayashi K, Maemondo M et al. Updated overall survival results from a randomized phase III trial comparing gefitinib with carboplatin-paclitaxel for chemo-naive non-small cell lung cancer with sensitive EGFR gene mutations (NEJ002). Ann Oncol 2013; 24 (1): 54–9.
5. Yang JC, Wu YL, Schuler M et al. Afatinib versus cisplatin-based chemotherapy for EGFR mutation-positive lung adenocarcinoma (LUX-Lung 3 and LUX-Lung 6): Analysis of overall survival data from two randomized, phase 3 trials. Lancet Oncol 2015; 16 (2): 141–51.
6. Yoshioka H, Shimokawa M, Seto T et al. Final overall survival results of WJTOG3405, a randomized phase III trial comparing gefitinib versus cisplatin with docetaxel as the first-line treatment for patients with stage IIIB/IV or postoperative recurrent EGFR mutation-positive non-small-cell lung cancer. Ann Oncol 2019; 30 (12): 1978–84.
7. Mok TS, Cheng Y, Zhou X et al. Improvement in overall survival in a randomized study that compared dacomitinib with gefitinib in patients with advanced non-small-cell lung cancer and EGFR-activating mutations. J Clin Oncol 2018; 36 (22): 2244–50.
8. therascreen EGFR RGQ PCR Kit [Package insert]. QIAGEN GmbH, Hilden, Germany 2019.
9. Hsiue EH, Lee JH, Lin CC, Yang JC. Profile of the therascreen(R) EGFR RGQ PCR Kit as a companion diagnostic for gefitinib in non-small cell lung cancer. Expert Rev Mol Diagn 2016; 16 (12): 1251–7.
Valle A, Le Loupp AG, Denis MG. Efficiency of the Therascreen(R) RQQ PCR kit for the detection of EGFR mutations in non-small cell lung carcinomas. Clin Chim Acta 2014; 429: 8 –11.

11 cobas® EGFR Mutation Test v2 [Package insert]. Roche Molecular Systems, Inc., Indianapolis, IN 2016.

12 Malapelle U, Sirera R, Jantsus-Lewinrette E et al. Profile of the Roche cobas(R) EGFR mutation test v2 for non-small cell lung cancer. Expert Rev Mol Diagn 2017; 17 (3): 209–15.

13 OncomineTM Dx Target Test [Package insert]. Thermo Fisher Scientific Inc., Carlsbad, CA, 2017.

14 OncomineTM Dx Target Test [Summary of Safety and Effectiveness Data (SSED)]. Thermo Fisher Scientific Inc., Carlsbad, CA, 2017.

15 Chung KP, Wu SG, Wu JY et al. Clinical outcomes in non-small cell lung cancers harboring different exon 19 deletions in EGFR. Clin Cancer Res 2012; 18 (12): 3470–7.

16 Feng Q, Yang ZY, Zhang JT, Tang JL. Comparison of direct sequencing and amplification refractory mutation system for detecting epidermal growth factor receptor mutation in non-small-cell lung cancer patients: A systematic review and meta-analysis. Oncotarget 2017; 8 (35): 59552–62.

17 Young EC, Owens MM, Adeyiyi I et al. A comparison of methods for EGFR mutation testing in non-small cell lung cancer. Diagn Mol Pathol 2013; 22 (4): 190–5.

18 Lopez-Rios F, Angulo B, Gomez B et al. Comparison of molecular testing methods for the detection of EGFR mutations in formalin-fixed paraffin-embedded tissue specimens of non-small cell lung cancer. J Clin Pathol 2013; 66 (5): 381–5.

19 Ho HL, Chang FP, Ma HH et al. Molecular diagnostic algorithm for epidermal growth factor receptor mutation detection in Asian lung adenocarcinomas: Comprehensive analyses of 445 Taiwanese patients with immunohistochemistry, PCR-direct sequencing and scorpion/ARMS methods. Respiratory 2013; 18 (8): 1261–70.

20 Wu M, Pan X, Xu Y, Wu S, Wu X, Chen B. Methodological comparison of the allele refractory mutation system and direct sequencing for detecting EGFR mutations in NSCLC, and the association of EGFR mutations with patient characteristics. Oncol Lett 2018; 16 (1): 1087–94.

21 Arcila ME, Nafa K, Chaft JE et al. EGFR exon 20 insertion mutations in lung adenocarcinomas: Prevalence, molecular heterogeneity, and clinicopathologic characteristics. Mol Cancer Ther 2013; 12 (2): 220–9.

22 Inoue A, Yoshida K, Morita S et al. Characteristics and overall survival of EGFR mutation-positive non-small cell lung cancer treated with EGFR tyrosine kinase inhibitors: A retrospective analysis for 1660 Japanese patients. Jpn J Clin Oncol 2016; 46 (5): 462–7.

23 Leduc C, Merlio JP, Besse B et al. Clinical and molecular characteristics of non-small-cell lung cancer (NSCLC) harboring EGFR mutation: Results of the nationwide French cooperative thoracic intergroup (IFCT) program. Ann Oncol 2017; 28 (11): 2715–24.

24 Li S, Li L, Zhu Y et al. Coexistence of EGFR with KRAS, or BRAF, or PIK3CA somatic mutations in lung cancer: A comprehensive mutation profiling from 5125 Chinese cohorts. Br J Cancer 2014; 110 (11): 2812–20.

25 Shukuya T, Takahashi T, Kai R et al. Efficacy of gefitinib for non-adenocarcinoma non-small-cell lung cancer patients harboring epidermal growth factor receptor mutations: A pooled analysis of published reports. Cancer Sci 2011; 102 (5): 1032–7.

26 Kobayashi K, Soejima K, Fukunaga K et al. Key prognostic factors for EGFR-mutated non-adenocarcinoma lung cancer patients in the Japanese Joint Committee of Lung Cancer Registry Database. Lung Cancer 2020; 146: 236–43.

27 Goto K, Satouchi M, Ishii G et al. An evaluation study of EGFR mutation tests utilized for non-small-cell lung cancer in the diagnostic setting. Ann Oncol 2012; 23 (11): 2914–9.

28 Chouaid C, Dujon C, Do P et al. Feasibility and clinical impact of re-biopsy in advanced non small-cell lung cancer: A prospective multicenter study in a real-world setting (GFPC study 12-01). Lung Cancer 2014; 86 (2): 170–3.

29 Nosaki K, Satouchi M, Kurata T et al. Re-biopsy status among non-small cell lung cancer patients in Japan: A retrospective study. Lung Cancer 2016; 101: 1–8.

30 Kimura H, Ohira T, Uchida O et al. Analytical performance of the cobas EGFR mutation assay for Japanese non-small-cell lung cancer. Lung Cancer 2014; 83 (3): 329–33.

Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher’s website:

Table S1 The profile of detectable EGFR mutation by Cobas V2, Therascreen and Oncomine.