Serum Concentrations of Selected Heavy Metals in Patients with Alcoholic Liver Cirrhosis from the Lublin Region in Eastern Poland

Andrzej Prystupa 1, Anna Błażewicz 2, Paweł Kiciński 3,†, Jarosław J. Sak 4,5,†,*
Jarosław Niedziałek 5,† and Wojciech Żaluska 5

1 Department of Internal Medicine, Medical University of Lublin, Staszica 16, 20-081 Lublin, Poland; aprystupa@wp.pl
2 Department of Analytical Chemistry, Medical University of Lublin, Chodźki 4a (Collegium Pharmaceuticum), 20-093 Lublin, Poland; anna.blazewicz@umlub.pl
3 Department of Family Medicine, Medical University of Lublin, Staszica 11, 20-081 Lublin, Poland; pawelkici@wp.pl
4 Department of Ethics and Human Philosophy, Medical University of Lublin, Staszica 4/6 (Collegium Maximum), 20-059 Lublin, Poland
5 Department of Nephrology, Medical University of Lublin, Jaczewskiego 8, 20-954 Lublin, Poland; wtzaluska2@poczta.onet.pl
6 Individual Medical Practice, Lublin, Ludwika Hirsztelda 5/11, 20-092 Lublin, Poland; niedzialseauke@poczta.fm
* Correspondence: jareksak@tlen.pl; Tel.: +48-814-486-850
† These authors contributed equally to this work.

Abstract: According to the WHO report, alcohol is the third most significant health risk factor for the global population. There are contrary reports about heavy metals concentrations in patients with alcoholic liver cirrhosis. The aim of this study was to investigate serum concentrations of selected heavy metals in patients with alcoholic liver cirrhosis living in the eastern part of Poland according to cirrhosis stage. The participants came from various hospitals of the Lublin region were enrolled. The study group included 46 male and 16 female patients. The control group consisted of 18 healthy individuals without liver disease. High Performance Ion Chromatography was used to determine the concentrations of metal ions (Cd, Zn, Cu, Ni, Co, Mn, and Pb) in serum samples. The concentrations of copper, zinc, nickel, and cobalt were found to be significantly lower in patients with alcoholic liver cirrhosis compared to the control group. The serum concentration of cadmium was significantly higher in patients with advanced alcoholic liver cirrhosis compared to the control group. We hypothesize that disorders of metabolism of heavy metals seem to be the outcome of impaired digestion and absorption, which are common in cirrhosis, improper diet, environmental and occupational exposure.

Keywords: liver cirrhosis; alcohol; heavy metals; microelements

1. Introduction

According to the World Health Organization’s report, alcohol is the third most significant health risk factor for the global population [1]. Chronic alcohol intake is associated with an increased risk of malnutrition, chronic pancreatitis, alcoholic liver disease (steatosis, hepatitis, cirrhosis), and cancer [2]. Liver cirrhosis is characterized by diffuse fibrosis of the hepatic connective tissue, which causes deterioration of its structure, degeneration and destruction of hepatocytes.
Exposure to inappropriate doses of heavy metals results in several adverse health effects [3]. The role of heavy metals in the pathogenesis of liver cirrhosis and its complications is not fully understood. Heavy metals may exacerbate liver damage (e.g., lead, cadmium) but some of them are essential for the organism (microelements, e.g., cobalt, zinc). There are contrary reports about heavy metals concentrations in patients with alcoholic liver cirrhosis.

The aim of this study was to investigate serum concentrations of selected heavy metals in patients with alcoholic liver cirrhosis living in the eastern part of Poland according to cirrhosis stage and to compare them with concentrations in controls.

2. Experimental Section

2.1. Patients

Patients with alcoholic liver cirrhosis treated in various hospitals of the Lublin region were randomly enrolled. The study group included 46 male and 16 female patients, aged 54.9 ± 10.4 years. All patients presented a history of heavy alcohol consumption (mainly vodka and cheap wine) in the absence of positivity for serological markers of viral, autoimmune, and metabolic diseases. The diagnosis of liver cirrhosis was based on clinical features, laboratory tests and abdominal ultrasound. The Child-Turcotte-Pugh criteria (Child-Pugh score) as P-Ch A, P-Ch B, P-Ch C, were used to assess the severity of liver cirrhosis. The control group consisted of 18 (12 male and six female) healthy individuals, aged 43.7 ± 14.6 years, without liver disease and without a history of alcohol abuse. The exclusion criteria were tobacco smoking and vitamin and mineral supplements taken over the last three months, acute kidney damage, stage III-V chronic kidney disease, and acute infections. Characteristics of the study population are given in Tables 1 and 2.

The study design was approved by the Bioethics Committee of the Medical University of Lublin (agreement number KE-0254/190/2011). All subjects gave their written informed consent.

Table 1. Characteristics of the control and study group.

| Variables                  | Control Group (n = 18) | Study Group (n = 62) |
|----------------------------|------------------------|----------------------|
| Age (years)                | 43.7 ± 14.6            | 54.9 ± 10.4          |
| Percentage of males (%)    | 67%                    | 74%                  |
| Alcohol abuse (years)      | -                      | 14.05 ± 5.17         |
| Body Weight (kg)           | 67.5 ± 8.8             | 68.7 ± 13.7          |
| Total bilirubin (mg/dL)    | 0.39 ± 0.29            | 5.33 ± 7.07          |
| Albumin (g/dL)             | 5.24 ± 0.55            | 2.63 ± 0.66          |
| INR                        | 1.24 ± 0.15            | 1.52 ± 0.35          |
| Total protein (g/dL)       | 6.2 ± 0.31             | 5.6 ± 0.84           |
| ALT (U/L)                  | 17.9 ± 5.9             | 64.3 ± 123.8         |
| AST (U/L)                  | 18.2 ± 7.1             | 115.5 ± 166.5        |
| PLT (G/L)                  | 231.3 ± 29.8           | 133.7 ± 73.9         |
| MCV (fL)                   | 84.7 ± 3.4             | 93.7 ± 9.3           |
| Urea (mg/dL)               | 24.5 ± 10.1            | 32.4 ± 26.1          |
| Sodium (mmol/L)            | 140 ± 3.2              | 134.3 ± 5.4          |
| Potassium (mmol/L)         | 4.3 ± 0.39             | 3.8 ± 0.65           |
| CRP (mg/L)                 | 2.51 ± 2.29            | 21.07 ± 10.96        |

| Complications              |                        |                      |
|----------------------------|------------------------|----------------------|
| Oesophageal varices (%)    | -                      | 53%                  |
| Ascitis (%)                | -                      | 42%                  |
| Encephalopathy (%)         | -                      | 64%                  |

Reference ranges provided by the laboratory that performed determinations: total bilirubin (0–1.2 mg/dL), albumin (3.5–5.2 g/dL), total protein (6–8 g/dL), ALT—alanine aminotransferase (5–40 U/L), AST—aspartate aminotransferase (5–40 IU/L), urea (21–43 mg/dL), PLT—blood platelets (120–400 K/µL), INR—International Normalized Ratio (0.86–1.30), MCV—mean cell volume (80–94 fL), Na—sodium (136–145 mmol/L), K—potassium (3.5–5.1 mmol/L), CRP—C-reactive protein (<5 mg/L).
Table 2. Concentrations of heavy metals in the control group and patients with various stages of liver cirrhosis according to Child-Pugh classification.

| Heavy Metals | Controls | P-Ch A (n = 19) | P-Ch B (n = 20) | P-Ch C (n = 23) | p *  |
|--------------|----------|----------------|----------------|----------------|------|
| Cadmium (Cd) (mg/L) | 0.0054 ± 0.0007 | 0.0061 ± 0.0028 | 0.0076 ± 0.0045 | 0.0078 ± 0.0044 | <0.05 |
| Cobalt (Co) (mg/L) | 0.0042 ± 0.0020 | 0.0028 ± 0.0015 | 0.0024 ± 0.0014 | 0.0024 ± 0.0014 | <0.01 |
| Copper (Cu) | 1.2663 ± 0.0771 | 1.1178 ± 0.2240 | 1.1080 ± 0.2024 | 1.1529 ± 0.1599 | <0.001 |
| Manganese (Mg) (mg/L) | 0.0089 ± 0.0006 | 0.0126 ± 0.0214 | 0.0144 ± 0.0214 | 0.0154 ± 0.0158 | NS   |
| Zinc (Zn) (mg/L) | 1.3607 ± 0.1123 | 1.0371 ± 0.3373 | 0.9579 ± 0.2441 | 1.0095 ± 0.3164 | <0.001 |
| Nickel (Ni) (mg/L) | 0.0019 ± 0.0001 | 0.0007 ± 0.0001 | 0.0005 ± 0.0001 | 0.0003 ± 0.0011 | NS   |
| Lead (Pb) (mg/L) | 0.0384 ± 0.0076 | 0.0447 ± 0.0135 | 0.0439 ± 0.0127 | 0.0478 ± 0.0176 | NS   |
| Iron (Fe) (mg/L) | 1.3682 ± 0.1452 | 1.2288 ± 0.2272 | 1.3540 ± 0.3264 | 1.3170 ± 0.2687 | NS   |

NS—not significant (p > 0.05). * Results of tests of intergroup multiple comparisons are presented in figures.

2.2. Blood Samples

The material for the study was the peripheral blood obtained from the ulnar vein. Blood samples were collected after an 8–12-h overnight fast between 8.00 and 10.00 a.m. into clot tubes, 7 mL in volume. Serum was separated by centrifugation at 1000 rpm for 10 min. Serum samples taken from healthy subjects and alcoholic liver cirrhosis patients were pre-treated and analyzed in the same way. They were transported and stored frozen at −20 °C in polypropylene containers. 1 mL of the sample was divided into two parts (each 0.5 mL) in order to have two independent solutions prior to the mineralization procedure. The mineralization procedure was carried out in the NovaWave Microwave Tunnel Digestion System (Scp Science, Montreal, QC, Canada) using Teflon® vessels.

Each time an acidic digestion with 99% nitric acid water solution was applied (1 mL of HNO₃: 9 mL H₂O). The conditions of the mineralization procedure had been previously optimized [4]. The obtained solutions were poured into volumetric flasks (PTFE).

The detailed procedure of preparation of standard solutions, operating HPIC conditions, and validation of the methods applied were described in the previous papers [4,5]. The limits of detection (LOD) values (in µg/mL) were as follows: 0.022 (Cd), 0.026 (Co), 0.048 (Cu), 0.006 (Mn), 0.056 (Zn), 0.006 (Ni), 0.009 (Fe), and 0.0008 (Pb). The method accuracy was verified by the certified reference material (human serum): Seronorm™ Trace Elements Serum L-2 (Billingstad, Norway).

2.3. Instrumentation and Reagents

High Performance Ion Chromatography (HPIC) was used to determine concentrations of metal ions in serum samples.

A Dionex DX-500 ion chromatograph (Sunnyvale, CA, USA) composed of an IP25 isocratic pump, a pre-column IonPac CG5A, an IonPac CS5A column, and a AD20 UV-VIS detector was used for measurements. A Chromeleon (Dionex, Sunnyvale, CA, USA) chromatography workstation was used for instrument control and data acquisition. Post column reagent (PCR) was applied for the spectrophotometric detection. One liter of PCR was prepared with 0.5 mmole of 4-(2-pyridylazo)resorcinol (PAR), 1.0 mole of 2-dimethylamino-ethanol, 0.3 mole of sodium bicarbonate, and 0.50 mole of ammonium hydroxide dissolved in deionized water (18 MΩ·cm). All reagents were of analytical grade. The eluent was filtered and degassed before use. The pyridine-2,6-dicarboxylic acid (PDCA) eluent enables the separation of copper, nickel, zinc, cobalt, cadmium, and manganese. The composition of 1 L of the PDCA eluent concentrate (simplifying eluent preparation and improves reproducibility), which was five times diluted with deionized water and used as a mobile phase during chromatographic analysis, was as follows: 7.0 mmole of PDCA, 66 mmole of potassium hydroxide, 5.6 mmole of potassium sulfate, and 74 mmole of formic acid. The oxalic acid eluent concentrate (10 times diluted with deionized water) was used as a mobile phase during determinations of lead.
ions. The composition of the oxalic eluent was as follows: 8.0 mM oxalic acid, 50 mM potassium hydroxide, and 100 mM tetramethylammonium hydroxide.

All mobile phase components were obtained from Sigma-Aldrich, Darmstadt, Germany. Aqueous solutions of metal salts were prepared by dilution of Titrisol standard metal salt solutions (Merck Millipore, Darmstadt, Germany). PAR was obtained from Dionex, Sunnyvale, CA, USA.

All analyses were carried out using pre-washed polypropylene flasks and vials.

2.4. Statistical Analysis

STATISTICA 10 PL (StatSoft, Inc., Tulsa, OK, USA) was used for statistical analysis. Continuous variables were expressed as mean ± standard deviation (SD). Before analysis, variables were checked for normality using the Shapiro-Wilk test; the Brown-Forsythe test was applied to test equality of variances. Variables were compared using the t-student test or ANOVA test when normally distributed or Mann-Whitney test or Kruskall-Wallis test, when non-normally distributed. Analysis of covariance (ANCOVA) was performed to examine differences between groups representing the controls and the patients with the consecutive stages of cirrhosis on a heavy metal concentration after controlling for variation of age (considered to be a nuisance variable). Correlations between two variables were analyzed with the Pearson correlation coefficient or Spearman’s rank correlation coefficient when appropriate. For all tests, \( p < 0.05 \) was considered as statistically significant.

3. Results

3.1. Concentrations of Heavy Metals in Patients with Alcoholic Liver Cirrhosis Compared to the Control Group

Serum concentrations of heavy metals in controls and patients with various stages of alcoholic liver cirrhosis were presented in Table 2 and in Figures 1–5.

Significant differences in cadmium concentrations were observed in the study population (Figure 5); the concentrations in question were significantly lower in the control group (0.0054 ± 0.0007 mg/L) than in patients with advanced cirrhosis, i.e., in Child-Pugh C patients (0.0078 ± 0.0044 mg/L). Moreover, copper concentrations were significantly higher in the control group (1.2663 ± 0.0771 mg/L) compared to Child–Pugh A (1.1178 ± 0.2240 mg/L), B (1.1080 ± 0.2024 mg/L), and C (1.1529 ± 0.1559) patients. Otherwise, there were no significant differences in its concentrations among patients with various stages of cirrhosis (Figure 2). Concentrations of zinc were significantly lower in Child–Pugh A (1.0371 ± 0.3373 mg/L), B (0.9579 ± 0.2441 mg/L), and C (1.0095 ± 0.3164) patients compared to controls (1.3607 ± 0.1123 mg/L) (Figure 3). Similarly to copper, there were no significant differences in its concentrations among subgroups with different stages of cirrhosis. Furthermore, concentrations of nickel were significantly higher in the control group (0.0019 ± 0.001 mg/L) compared to Child-Pugh B (0.0005 ± 0.001 mg/L) and C (0.0003 ± 0.0011 mg/L) patients (Figure 4). Serum concentrations of cobalt were significantly lower in Child–Pugh B (0.0026 ± 0.0015 mg/L) and C (0.0024 ± 0.0014) patients compared to controls (0.0042 ± 0.0020 mg/L) (Figure 1).

An increasing trend in concentrations of manganese and lead was observed in patients with cirrhosis, yet the elevations were not statistically significant (Table 2). There were no significant inter-subgroup differences in iron concentrations (Table 2).

Additionally, analysis of covariance (ANCOVA) confirmed that there was a significant effect of the independent variable (i.e., “subgroup”, which divides the population into four categories: healthy, and alcoholics with classes A, B, and C of liver cirrhosis,) on serum concentration of cadmium, copper, zinc, nickel, and cobalt after controlling for the effect of age (considered to be a confounding factor).
Figure 1. Concentrations of cobalt in individual subgroups.

Figure 2. Concentrations of copper (Cu) in individual subgroups.
Figure 3. Concentrations of zinc (Zn) in individual subgroups.

Figure 4. Concentrations of nickel (Ni) in individual subgroups.
were negatively correlated with MCV (\( r = -0.33, p < 0.01 \)), copper (\( r = -0.29, p = 0.01 \)), zinc (\( r = -0.3, p = 0.01 \)), and nickel (\( r = -0.47, p < 0.0001 \)). A significant, albeit weak, correlation was found between age versus the concentration of cadmium (\( r = 0.23, p < 0.05 \)) and lead (\( r = 0.24, p < 0.05 \)). No significant correlations were observed between concentrations of the heavy metals studied and gender.

### 3.3. Correlations between Concentrations of Heavy Metals and Laboratory Results

Moderate positive correlations were demonstrated between the platelet count versus concentrations of zinc (\( r = 0.34, p < 0.01 \)) and nickel (\( r = 0.48, p < 0.0001 \)), and a negative correlation between the platelet count and concentration of cadmium (\( r = -0.28; p < 0.05 \)). Otherwise, concentrations of cobalt were negatively correlated with MCV (\( r = -0.36, p = 0.001 \)). Moreover, the AST activity was negatively correlated with concentrations of cobalt (\( r = -0.35, p < 0.01 \)), zinc (\( r = -0.29; p < 0.05 \)), and nickel (\( r = -0.24, p < 0.05 \)) while the concentration of total bilirubin was negatively correlated with the concentration of nickel (\( r = -0.24; p < 0.05 \)). The International Normalized Ratio (INR) value was negatively correlated with the concentration of nickel (\( r = -0.47; p < 0.001 \)). Furthermore, a negative correlation was found between the concentration of C-reactive protein (CRP) and concentrations of cobalt (\( r = -0.26; p < 0.05 \)) and copper (\( r = -0.27; p < 0.05 \)).

### 3.4. Concentrations of Heavy Metals according to the Presence of Ascites, Encephalopathy, and Esophageal Varices

The concentration of nickel was demonstrated to be significantly lower in patients with ascites as compared to cirrhotic patients without ascites (0.0003 ± 0.001 vs. 0.0007 ± 0.001 mg/L; \( p < 0.05 \)). Concentrations of zinc and nickel were lower in patients with esophageal varices as compared to cirrhotic patients without varices (0.0003 ± 0.001 vs. 0.0007 ± 0.001 mg/L; \( p < 0.05 \)).
cirrhotic patients without esophageal varices (0.0003 ± 0.008 vs. 0.0007 ± 0.001 mg/L; \( p < 0.05 \) and 0.958 ± 0.304 vs. 1.048 ± 0.294; \( p < 0.05 \), respectively).

3.5. Correlations among Heavy Metal Concentrations

A significant correlation was demonstrated between concentrations of cadmium and manganese \((r = 0.48; p < 0.0001)\), cobalt and nickel \((r = 0.29; p < 0.05)\), copper and zinc \((r = 0.3; p < 0.05)\), as well as zinc and nickel \((r = 0.39; p < 0.001)\).

4. Discussion

“Heavy metals” is an imprecise term that involves a variously-defined collection of metals and semimetals characterized by high density. Our study presents analysis of serum concentrations of the selected heavy metals in patients with alcoholic liver cirrhosis. Some of the metals studied play important biological roles as microelements necessary for proper activities of numerous enzymes and for other processes (Cu, Co, Zn, Fe, Mn, Ni). However, their excess can cause toxic effects. Moreover, Cd and Pb are toxic elements whose presence in the human body is unnecessary. The patients included to the study were from Lublin and its region (Southeastern Poland). The Lublin province is an agricultural, poorly-industrialized region, and our subjects were predominantly farmers and unemployed individuals; thus, the potential effect of environmental and occupational exposure was minimized.

Our study findings demonstrated a significant increase in serum concentrations of cadmium in P-Ch C patients compared to controls. The major sources of this element in the body are environmental (including cigarette smoke) and occupational exposures. Cadmium is predominantly absorbed via the respiratory and digestive route, as well as the skin, although to a markedly lesser degree, and accumulates mainly in the kidneys and liver, where it exerts its toxic effects \([6,7]\). Chronic exposure to cadmium has been demonstrated to lead to hepatocyte damage \([8]\). Cadmium is an element of high toxicity and long half-life, which has negative effects on reproductive capacities and the cardiovascular system \([9]\). In humans, cadmium exerts carcinogenic effects affecting numerous cellular and molecular mechanisms (including redox imbalance, DNA repair, and inhibition of apoptosis) \([10]\). Kazi et al. have reported elevated concentrations of cadmium in patients with liver cirrhosis. Moreover, they have demonstrated significantly higher concentrations of cadmium in patients with HCC. However, their study regarded patients with liver cirrhosis induced by viral hepatitis and not alcoholic cirrhosis, as in our study; moreover, they did not analyze the effects of cirrhosis stage on concentrations of cadmium \([11,12]\). Further studies are required to assess the cause-effect relationship between elevated concentrations of cadmium and the development of HCC. Noteworthy, our findings revealed a correlation between the concentration of cadmium and reduced platelet count, which is likely to be of clinical importance.

Concentrations of cobalt were significantly lower in patients with alcoholic liver cirrhosis as compared to controls. Cobalt gets into the body in a few ways: firstly with food; secondly by the respiratory system; thirdly, by the skin. Cobalt is necessary for maintaining normal physiological functions of the human body. The organic form of cobalt is a component of vitamin B12 and is essential for formation of amino acids, some proteins in the nervous cells, and neurotransmitters \([13]\). Hunt et al. have demonstrated significantly reduced concentrations of cobalt in the livers of subjects with cirrhosis \([14]\). Low serum concentrations of cobalt in patients with liver cirrhosis can result from malnutrition and impaired absorption.

Copper is an essential nutrient trace metal involved in a multitude of cellular processes. Copper is used as a cofactor in numerous enzymatic mechanisms \([15]\). Sawa et al. have showed that Cu levels in serum and liver tissues were significantly higher in patients with liver cirrhosis compared to the control group. The main pathway of Cu excretion is via the biliary tract. In patients with long-term obstructive jaundice or primary biliary cirrhosis, serum and hepatic Cu levels are increased \([16]\). Rahelic et al. have found that serum concentrations of copper were significantly higher in patients with alcoholic liver
cirrhosis in comparison to controls [17]. In our study, serum concentrations of copper were significantly lower in patients with liver cirrhosis compared to controls. Low concentrations of copper in the study group might have resulted from impaired absorption and malnutrition. Our results are consistent with the study by Fields et al. who demonstrated in animal model that alcohol consumption may aggravate copper deficiency [18].

Nickel results in increased levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT) and gamma-glutamyl-transpeptidase (SGGT) in the liver and serum of animals and humans exposed to nickel salts [19]. Volini et al. have reported significantly increased hepatic concentrations of nickel in the early and advanced stages of hepatic cirrhosis [20]. In our study, serum concentrations of nickel were significantly lower in patients with liver cirrhosis compared to controls. Similar results were reported by McNeely et al. Serum concentrations of nickel were found to be significantly lower in 18 patients with liver cirrhosis compared to controls. According to the authors, decreased levels of serum nickel in liver cirrhosis may reflect diminished concentrations of serum nickeloplasmin and albumin [21]. Absorbed nickel is eliminated in the urine [22]. Patients with ascites in the course of liver cirrhosis are usually treated with diuretics. This may lead to increased excretion of nickel and its decreased serum level.

Liver plays an important role in maintaining systemic zinc homeostasis. In our study, the concentration of zinc was significantly lower in P-Ch A-C patients compared to controls. Several studies have concluded that zinc levels are decreased in serum of individuals with liver cirrhosis. Goode et al. have demonstrated that plasma zinc concentrations were reduced in patients with liver cirrhosis compared to controls and correlated strongly with plasma albumin concentrations [23]. Albumin is the major zinc carrier in the plasma and binds approximately 80% of plasma zinc [24]. Inadequate dietary intake, as well as the impaired absorption or increased clearance of zinc, may be responsible for the decrease of zinc content in patients with liver diseases [25]. Yoshida et al. measured serum Zn levels before and 60, 120, 180 min after administration of Zn sulfate. Lower increases in serum zinc levels 120 and 180 min after ingestion in cirrhotic patients than in controls indicated Zn malabsorption, which might be caused by portal hypertensive gastrocolonomopathy [26]. Diuretic therapy in patients with cirrhosis and ascites results in increased renal zinc excretion [27]. Reding et al. have showed that short-term oral zinc supplementation improves hepatic encephalopathy by correcting zinc deficiency that compromises conversion of ammonia to urea [28].

In our study, serum levels of manganese and lead were higher in cirrhotic patients than those in controls but the differences were not statistically significant. Rahelic et al. have revealed that serum concentrations of manganese were significantly higher in patients with liver cirrhosis compared to the control group [17]. Moscarello has not found any significant difference in concentrations of manganese between cirrhotic patients and controls [29]. Manganese is secreted in bile, thus, its concentration increases in cholestatic liver disease [30]. According to Jurczyk et al., the mean value of serum iron did not differ significantly among patients with chronic viral hepatitis, alcoholic liver cirrhosis and alcoholic hepatitis [31].

5. Conclusions

Disorders of heavy metal metabolism seem to be the outcome of impaired digestion and absorption, which are common in cirrhosis, improper diet, environmental and occupational exposure. Malnutrition is one of the essential liver cirrhosis-related problems [32]. Low serum concentrations of zinc, copper, cobalt, and nickel are most likely caused by inadequate dietary intake and malabsorption. Moreover, substantial effects of environmental exposure connected with work and place of residence cannot be excluded, which could, at least partially, explain the diverse results reported by various authors.

Further studies are needed to determine the prognostic value of impaired metabolism of heavy metals and effects of their supplementation on the clinical course of alcoholic liver cirrhosis.
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