Research Article

ASSESSMENT OF DISEASE RESISTANCE AND HIGH YIELDING TRAITS OF COMMON BUCKWHEAT GENOTYPES IN SUBTROPICAL CLIMATE OF NEPAL

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ABSTRACT

Twelve common buckwheat (Fagopyrum esculentum Moench) genotypes received from Hill Crops Research Program, Kabre, Dolakha were evaluated for resistance against major foliar fungal diseases and high yielding traits at the research field of National Maize Research Program, Rampur, Chitwan during winter seasons of two consecutive years 2017 and 2018. The design of the experiment was Randomized Complete Block having three replications. The unit plot size was 3m x 2m with 25cm row to row spacing and continues plant to plant spacing was maintained and net harvested plot was 6 sq meters. The experiment was planted at last week of October in both years. The recommended dose of fertilizer was 30:30:0 for N:P:K kg ha⁻¹ respectively and seed rate 50 kg ha⁻¹. Disease data were recorded for downy mildew (Perenospora fagopyri), powdery mildew (Erysiphe polygoni) and botrytis leaf blight (Botrytis cinerea) severity on 20 randomly tagged plants/plot. The yield and yield attributing traits were recorded. Buckwheat genotypes differed significantly (p<0.05) on disease severity, grain yield and yield attributing traits. Genotypes CBBP-01, KIF-72-22-520, GF5283, ACC#2234 and ACC#2213 were resistant to powdery mildew, downy mildew and botrytis leaf blight and also resulted in higher grain yield. Grain yield had found strong negative correlation with the fungal disease severity of buckwheat. These genotypes might be useful for the development of major foliar fungal disease resistant high yielding common buckwheat variety in inner terai region of Nepal.

Keywords: buckwheat, Disease resistance, Genotypes, Yielding traits

INTRODUCTION

Common or sweet buckwheat (Fagopyrum esculentum Moench) and bitter buckwheat (Fagopyrum tataricum (L.) Gaertn) both belongs to family Polygonaceae.

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Received: 30.03.2020
Accepted: 24.05.2020
Globally 18 species are known to be the members of genus *Fagopyrum* including two cultivated species *Fagopyrum esculentum* and *F. tataricum* (Oshaki et al., 2001). It is a sixth staple food crop next to rice, wheat, maize, finger millet, and barley in term of cultivation area (10296 ha), production (11472 mt) and yield (1114 kg ha\(^{-1}\)) in Nepal (MoAD, 2019). It is considered as a pseudo-cereal, poor man’s crop, and underexploited or neglected crops, occupying major place for Nepalese agriculture system and contributing greatly in food and nutrition security of remote areas especially in high hilly regions (Baniya, 1995). It is the best crop in higher altitude in terms of adaptation to different climatic variables and easily fitted to different cropping patterns due to short duration (Joshi, 2008). It is cultivated on marginal land in 61 out of 75 districts of Nepal from about 60 to 4500 masl, especially hilly and mountain districts. Sweet or common buckwheat varieties are generally grown in mid hill, terai and inner terai region but Tartary or bitter buckwheat varieties are grown in higher altitude (Baral et al., 2014). There are altogether 19 local landraces of sweet or common buckwheat and 37 for Tartary or bitter buckwheat listed from Nepal (Joshi, 2008). Although the scope and demand of buckwheat is increasing due to its medicinal and nutritional values but the yield potential is reducing due to several biotic and abiotic constraints. The reason behind the low productivity of most of the crops in Nepal is due to the attack of many plant diseases at different stages of crop. Disease is one of the major biotic constraints to reduce buckwheat yield and also deteriorate the quality of product that ultimately reduce the market price. Buckwheat plants are affected by wide range of pathogens with fungal diseases being the most important. Fungal diseases decrease in productivity through infection and damage to leaves, stems, roots and pods as well as reduce market value due to discolor seeds (Subedi et al., 2015). Among biotic constraints, downy mildew (*Perenospora fagopyri*), powdery mildew (*Erysiphe polygoni*), botrytis leaf blight (*Botrytis cinerea*), rust (*Puccinia fagopyri*), damping-off and root rot (*Rhizoctonia niasholani*, *Pythium* spp, *Fusarium* spp) were the major fungal diseases contributes the wide yield gap in buckwheat genotypes (Manandhar et al., 2016). Resistant varieties provide a more effective and more consistent method of disease control without causing any hazards to environment. Transfer of available resistance traits in a cultivated background is especially important for farmers lacking resources to control the disease (Subedi, 2015). Therefore, the present study was undertaken in order to find out the resistant/tolerant buckwheat genotypes against major foliar diseases and the direct and indirect contributions of these traits toward yield.

**MATERIALS AND METHODS**

Twelve buckwheat genotypes received from Hill Crops Research Program (HCRP), Kavre, Dolakha were evaluated for sources of disease resistance and yield related traits in National Maize Research Program (NMRP), Rampur, Chitwan during winter seasons of two consecutive years 2017 and 2018. The geographical location of the research area was 27°37’ N latitude and 84°25’ E longitude at an altitude of 256
ABMSL and has sub-tropical climate. The design of the experiment was randomized complete block having three replications. The unit plot size was 3 m x 2 m with 25 cm row to row spacing and continues plant to plant spacing was maintained and net harvested plot was 6 square meters. The experiment was planted at last week of October in both years. The recommended dose of fertilizer was 30:30:0 N:P:K kg ha\(^{-1}\) and seed rate 50 kg ha\(^{-1}\). Other agronomic practices were followed as recommended (HCRP, 2017). Disease data were recorded for downy mildew (\textit{Perenospora fagopyri}) and powdery mildew (\textit{Erysiphe polygoni}) severity in 0-9 scale (Manandhar et al., 2016) whereas botrytis leaf blight (\textit{Botrytis cinerea}) severity in 1-9 scoring scale on 20 randomly tagged plants/plot (Morrall and Mckenzie, 1974). Yield and yield attributing traits like days to flowering, days to maturity, plant height (cm), grain yield (kg ha\(^{-1}\)) and hundred seed weight (g) were recorded. Percent Disease Index (PDI) is computed on the basis of recorded data according to the formula (Wheeler, 1969).

\[
PDI (\%) = \frac{\text{Sum of all numerical values}}{\text{No of plants observed}} \times \frac{100}{\text{Maximum diseases rating}}
\]

All data were analyzed statistically using Microsoft Excel and MSTAT-C computer package program. Treatment means were compared using duncans multiple range test (DMRT) at 5% level of significance (P≤0.05). The relationship between disease severity and yield were also calculated.

**RESULTS AND DISCUSSION**

**Experiment during 2017**

Data revealed that statistically significant differences were observed in the traits flowering days, days to maturity, plant height (cm), grain yield (kg ha\(^{-1}\)), hundred seed weight (g), powdery mildew, downy mildew and botrytis leaf blight severity among buckwheat genotypes during winter season of 2017 (Table 1). The flowering days ranged from (19-25 days), maturity days (57-68 days), plant height (94-106 cm) and hundred seed weight (3.30 - 4.20 g). Buckwheat genotypes KIF-72-22-520 (1958.56 kg ha\(^{-1}\)), CBBP-01 (1950 kg ha\(^{-1}\)), GF 5283 (1868.44 kg ha\(^{-1}\)), ACC#2234 (1857.22 kg ha\(^{-1}\)) and PL 15 (1850.44 kg ha\(^{-1}\)) were the high yielding compare to standard check Mithe Phapar-1 (1830 kg ha\(^{-1}\)).

Out of 12 buckwheat genotypes, CBBP-01, KIF-72-22-520, ACC#2234, GF 5283, ACC#2213 and PL-15 were resistant having percent disease index (PDI) of 26.35, 27.08, 30.05, 31.45, 31.65 and 32.25% respectively whereas Chitwan local (local check) was susceptible having PDI of 56.88% to powdery mildew disease. The downy mildew severity was found lower in genotypes KIF-72-22-520 (32.42%), CBBP-01 (34.25%), GF 5283 (35.58%) and ACC#2234 (38.45%) compared to...
standard check Mithe Phapar-1 (41.45%). The higher downy mildew severity was recorded in genotype ACC#6529 (65.92%) followed by Chitwan local (61.72%). Similarly, botrytis leaf blight severity was found lower in genotypes CBBP-01 (37.45%), KIF-72-22-520 (38.88%), GF 5283 (41.35%), ACC#2234 (43.95%) and ACC#2213 (45.48%) compared to standard check Mithe Phapar-1 (48.58%). The higher botrytis leaf blight severity was recorded in genotype ACC#6529 (73.35%) followed by Chitwan local (66.58%) (Table 1).

Table 1. Response of buckwheat genotypes to yield parameters and major foliar fungal diseases at NMRP, Rampur, Chitwan, Nepal during winter season of 2017

| Genotypes          | Fl_days | Mat_days | PHT (cm) | GY (kg ha⁻¹) | HSWt (g) | Percent disease index (%) | PM | DM | BLB |
|--------------------|---------|----------|----------|--------------|----------|---------------------------|----|----|-----|
| ACC#5670           | †23b    | 66abc    | 96f      | 1677.11d    | 3.30f    | 47.68c                     | 56.42d | 62.28e |
| ACC#2213           | 22c     | 64cde    | 99j      | 1755.11c    | 3.90bcd  | 31.65d                     | 42.75g | 45.48f |
| ACC#6529           | 22c     | 60f      | 102cd    | 1438.33e    | 4.20a    | 64.14d                     | 65.92a | 73.35c |
| ACC#2234           | 23bc    | 63de     | 101de    | 1857.22b    | 4.10ab   | 30.05d                     | 38.45g | 43.95f |
| PC-30              | 23bc    | 65bcd    | 101de    | 1660.56d    | 3.80de   | 50.02e                     | 55.45f | 62.58c |
| GF 5283            | 24abc   | 68ab     | 101de    | 1868.44b    | 3.50f    | 31.45d                     | 35.58ef| 41.35g |
| CBBP-01            | 19f     | 61f      | 103bc    | 1950.00a    | 4.10ab   | 26.35c                     | 34.25f | 37.45h |
| KIF-72-22-520      | 25a     | 68a      | 105ab    | 1958.56a    | 3.40f    | 27.08e                     | 32.42f | 38.88h |
| PL 15              | 25a     | 67ab     | 106b     | 1850.44b    | 4.10ab   | 32.25d                     | 42.62d | 55.42d |
| ACC#493            | 24abc   | 66abc    | 101de    | 1463.33e    | 3.80cde  | 55.32b                     | 58.22g | 64.38bc |
| Mithe Phapar-1 (SC)| 22c     | 63de     | 102cd    | 1830.00b    | 3.60ef   | 32.52d                     | 41.45g | 48.58h |
| Chitwan Local (LC) | 19f     | 57f      | 97e      | 1473.56b    | 3.30f    | 56.88b                     | 61.72b | 66.58b |

Grand mean         22.58  64.06  101.08  1731.89  3.80  40.45  47.10  53.36
F test             **  **  **  **  **  **  **  **
LSD (0.05)         2.06  2.26  2.18  38.60  0.03  2.43  2.89  2.33
CV, %              5.40  2.10  1.30  1.30  4.00  3.50  3.60  2.60

Note: † Means of 3 replications, Means in column with same superscript is not significantly differed by DMRT (P<0.05). Fl_days – flowering days, Mat_days – maturity days, PHT- Plant height, GY- grain yield, HSWt- hundred seed weight, PM- Powdery mildew, DM- Downy mildew, BLB- Botrytis leaf blight, %- percentage, cm-centimeter, kg ha⁻¹ - kilogram per hectare, g- gram, SC- standard check, LC- local check, **- highly significant

Experiment during 2018

During 2018 also, trends of both disease development, yield and yield attributing traits were similar. The yield (kg ha⁻¹) and yield attributing traits like flowering days,
days to maturity, plant height (cm) and hundred seed weight (g) were significantly varied with buckwheat genotypes (Table 2). The flowering days ranged from (22-29 days), maturity days (61-71 days), plant height (100-108 cm) and hundred seed weight (3.60 - 4.50 g). Buckwheat genotypes CBBP-01 (2050 kg ha$^{-1}$), KIF-72-22-520 (2048.89 kg ha$^{-1}$), GF 5283 (1984.44 kg ha$^{-1}$), ACC#2234 (1953.89 kg ha$^{-1}$) and ACC#2213 (1952.78 kg ha$^{-1}$) were the high yielding compare to standard check Mithe Phapar-1 (1950 kg ha$^{-1}$) (Table 2).

Table 2. Performance of buckwheat genotypes to yield parameters and major foliar fungal diseases at NMRP, Rampur, Chitwan, Nepal during winter season of 2018

| Genotypes         | Fl_ Days | Mat_ days | PHT (cm) | GY (kg ha$^{-1}$) | HSWt (g) | Percent disease index (%) |
|-------------------|----------|-----------|----------|-------------------|----------|--------------------------|
|                   |          |           |          |                   |          | PM | DM | BLB |
| ACC#5670          | †25d     | 68bc      | 100a     | 1784.44c          | 3.60d    | 52.05g | 63.55b | 68.25c |
| ACC#2213          | 26cd     | 67d       | 103cd    | 1952.78b          | 4.20b    | 36.68d | 46.35e | 54.28f |
| ACC#6529          | 25d      | 63c       | 105bc    | 1526.67d          | 4.60d    | 67.48e | 69.38a | 75.72c |
| ACC#2234          | 27bc     | 67d       | 103c     | 1953.89b          | 4.50c    | 36.72df | 43.78f | 51.45g |
| PC-30             | 28ab     | 70b       | 102cd    | 1785.56c          | 4.00bc   | 51.38d | 61.48c | 65.28d |
| GF 5283           | 26cd     | 67d       | 101bc    | 1984.44ab         | 4.00bc   | 35.58f | 40.72g | 49.55f |
| CBBP-01           | 23c      | 63bc      | 106ab    | 2050.00a          | 4.20b    | 30.05b | 35.62b | 40.58c |
| KIF-72-22-520     | 29a      | 71a       | 108a     | 2048.89a          | 3.80cd   | 33.45f | 39.08g | 46.22b |
| PL 15             | 28ab     | 69ab      | 108a     | 1949.11b          | 4.20b    | 37.38c | 49.42d | 56.38e |
| ACC#493           | 27bc     | 69bc      | 105b     | 1550.00d          | 4.10b    | 65.45b | 69.18a | 72.42b |
| Mithe Phapar-1 (SC) | 27bc   | 66d       | 103b     | 1950.00b          | 4.20b    | 36.55df | 47.55c | 55.62ed |
| Chitwan Local (LC)| 22e      | 61e       | 100f     | 1586.22d          | 3.80cd   | 62.48gd | 64.72b | 71.42b |
| Grand mean        | 26.08    | 66.72     | 103.72   | 1843.50           | 4.10     | 45.44  | 52.57  | 58.93 |
| F test            | **       | **        | **       | **                | **       | **     | **    | **   |
| LSD (0.05)        | 1.38     | 1.74      | 1.61     | 64.60             | 0.03     | 1.50   | 1.86   | 1.93 |
| CV, %             | 3.10     | 1.50      | 0.90     | 2.10              | 3.70     | 1.90   | 2.10   | 1.90 |

Note: † Means of 3 replications, Means in column with same superscript is not significantly differed by DMRT (P≤0.05). Fl days – flowering days, Mat_days – maturity days, PHT- Plant height, GY- grain yield, HSWt- hundred seed weight, PM- Powdery mildew, DM- Downy mildew, BLB- Botrytis leaf blight, %- percentage, cm-centimeter, kg ha$^{-1}$ – kilogram per hectare, g- gram, SC- standard check, LC-local check, **- highly significant.

Genotypes, CBBP-01, KIF-72-22-520, GF 5283, ACC#2213 and ACC#2234 were resistant having percent disease index (PDI) of 30.05, 33.45, 35.58, 36.68 and 36.72% respectively compared to standard check Mithe Phapar-1 (36.55%) whereas ACC#6529, ACC#493 and Chitwan local (local check) was susceptible having PDI of 67.48, 65.45 and 62.48% respectively to powdery mildew disease. The downy
mildew severity was found lower in genotypes CBBP-01 (35.62%), KIF-72-22-520 (39.08%), GF 5283 (40.72%) and ACC#2234 (43.78%) and ACC#2213 (46.35%) compared to standard check Mithe Phapar-1 (47.55%) (Table 2). The higher downy mildew severity was recorded in genotype ACC#6529 (69.38%) followed by ACC#493 (69.18%) and Chitwan local (64.72%). Similarly, botrytis leaf blight severity was found lower in genotypes CBBP-01 (40.58%), KIF-72-22-520 (46.22%), GF 5283 (49.55%), ACC#2234 (51.45%) and ACC#2213 (54.28%) compared to standard check Mithe Phapar-1 (55.62%). The higher botrytis leaf blight severity was recorded in genotype ACC#6529 (75.72%) followed by ACC# 493 (72.42%) and Chitwan local (71.42%) (Table 2).

Relationship between grain yield (kg ha\(^{-1}\)) and powdery mildew severity (%)

During 2017-2018 cropping season of buckwheat, yield was found to had highly significant negative correlation \((r = -0.99)\) with the percent disease index of powdery mildew disease. The predicted linear regression line was also displayed downward slope i.e. \(y = -0.07x + 167.46\), with regression coefficient \(R^2 = 0.97\), where ‘y’ denoted predicted crop yield of buckwheat and ‘x’ stood for PDI of powdery mildew of buckwheat (Fig. 1). The estimated regression line indicated that the unit rise in the PDI of powdery mildew disease (within 0-9 scale), there existed possibilities of yield reduction by 167.39 kg ha\(^{-1}\).

![Figure 1](image_url)

**Figure 1.** Relationship between crop yield (kg ha\(^{-1}\)) and PDI of buckwheat powdery mildew at Rampur, Chitwan, Nepal during 2017-2018

**Relationship between grain yield (kg ha\(^{-1}\)) and Downy mildew severity (%)**

A significant linear negative correlation \((r = -0.96)\) between yield and downy mildew severity was observed representing the best fit having \(R^2 = 92\%\) (Fig. 2). Obviously
the yield decreased with the increase in Percent Disease Index (PDI). The predicted linear regression line was also displayed downward slope i.e. $y = -0.06x+158$, with regression coefficient $R^2 = 0.93$, where ‘y’ denoted predicted crop yield of buckwheat genotypes and ‘x’ stood for PDI of downy mildew of buckwheat (Fig. 2). The estimated regression line indicated that the unit rise in the PDI of downy mildew disease (within 0-9 scale), there existed possibilities of yield reduction by 157.94 kg ha$^{-1}$.

![Figure 2](image.png)

**Figure 2.** Relationship between crop yield (kg ha$^{-1}$) and PDI of buckwheat downy mildew at Rampur, Chitwan, Nepal during 2017-2018

**Relationship between grain yield (kg ha$^{-1}$) and botrytis leaf blight severity (%)**

A linear negative correlation ($r = -0.95$) between yield and botrytis leaf blight severity was observed representing the best fit having $R^2 = 89\%$ (Figure 3). Obviously the yield decreased with the increase in Percent Disease Index (PDI). The predicted linear regression line was also displayed downward slope i.e. $y = -0.06x+160.73$, with regression coefficient $R^2 = 0.90$, where ‘y’ denoted predicted crop yield of buckwheat genotypes and ‘x’ stood for PDI of botrytis leaf blight of buckwheat (Fig. 3). The estimated regression line indicated that the unit rise in the PDI of botrytis leaf blight disease (within 1-9 scale), there existed possibilities of yield reduction by 160.67 kg ha$^{-1}$.
The genetic improvement of buckwheat is an important area to determine its potential for production and rapid distribution. Disease resistance has been found quite useful for the improvement of desired traits and addressing the productivity and yield related problems in buckwheat (Luitel et al., 2017). Even within a species of plant that is susceptible to a particular species of pathogen however there is considerable variation in both the susceptibility of the various plant cultivars towards the pathogen and the virulence of the various pathogen races toward the plant variety. The genetics of such host-pathogen interaction are of considerable biological interest and of the utmost importance in developing disease control strategies through breeding for resistance (Agrios, 2005). Although buckwheat is normally a cold tolerant and is not attacked by many diseases or pests but a number of diseases and pests have been reported on this crop (Joshi and Paroda, 1991). The major diseases are powdery mildew (Erysiphe polygoni), downy mildew (Peronospora ducumeti), stem rot (Botrytis cinerea), chlorotic leaf spot (Alternaria alternata), leaf spot (Septoria polygonica), smut (Sphacelothecaj agopyri), root and stem rot (Phytophthoraagopyri), brown leaf spot (Ascochytaitalica), rust (Puccinia agopyri), root and collar rot (Sclerotinia libertianai) and root rot (Fusarium spp.). Attacks of several viruses also cause reduction in plant height and losses in grain yield. From this experiment, buckwheat genotypes CBBP-01, KIF-72-22-520, GF5283, ACC#2234 and ACC# 2213 showed higher yield with lower foliar fungal disease severity. The findings of this experiment is in agreement with the Baniya et al. (1995) who...
reported that out of 309 common buckwheat landraces, days to 50% flowering varied from 26 to 45 while days to 95% maturity varied from 67 to 98. Plant height ranged from 25 to 116.4 cm. There was a large variation noted in thousand grain weight from 10.2 to 31.8 g. The ratings for disease were from no disease to 100% infection for powdery mildew and from no disease to 60% infection for downy mildew showing a wide variation that could be utilized in crop improvement programmes. The experiment conducted by Baral et al. (2014) showed that buckwheat genotypes T-Vaskar (1282 kg ha\(^{-1}\)), GF 5289 (1205 kg ha\(^{-1}\)) and IR-13 (1212 kg ha\(^{-1}\)) as promising lines in the year 2012. Baniya et al. (1995) also reported that two main buckwheat diseases powdery mildew and downy mildew show negative correlation with yield and yield attributing traits indicating low incidence of disease is associated with high values of each respective trait. The resistance in buckwheat genotypes to different fungal diseases could be the presence of phenolic and flavonoids compounds like flavonols, anthocyanins and C-glucosyl-flavones (Mikulajova et al., 2016).

CONCLUSION
Grain yield had strong negative correlation with the major foliar fungal disease severity of buckwheat. Genotypes CBBP-01, KIF-72-22-520, GF5283, ACC#2234 and ACC# 2213 were resistant to powdery mildew, downy mildew and botrytis leaf blight that resulted in higher grain yield. The study can be useful for selecting suitable genotypes for the development of major foliar fungal disease resistant high yielding common buckwheat variety in subtropical climate of Nepal.

ACKNOWLEDGEMENTS
The fund for this study was achieved from Nepal Agricultural Research Council (NARC). Acknowledgements go to both NMRP and HCRP team and other technical staffs of NARC for their valuable suggestions, continuous support and facilities during experimentation period.

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