Intravoxel incoherent motion (IVIM) modeling of diffusion MRI during chemoradiation predicts therapeutic response in IDH wildtype glioblastoma

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Abstract

Background: Prediction of early progression in glioblastoma may provide an opportunity to personalize treatment. Simplified intravoxel incoherent motion (IVIM) MRI offers quantitative...
estimates of diffusion and perfusion metrics. We investigated whether these metrics, during chemoradiation, could predict treatment outcome.

**Methods:** 38 patients with newly diagnosed IDH-wildtype glioblastoma undergoing 6-week/30-fraction chemoradiation had standardized post-operative MRIs at baseline (radiation planning), and at the 10th and 20th fractions. Non-overlapping T1-enhancing (T1C) and non-enhancing T2-FLAIR hyperintense regions were independently segmented. Apparent diffusion coefficient ($\text{ADC}_{\text{T1C}}$, $\text{ADC}_{\text{T2-FLAIR}}$) and perfusion fraction ($f_{\text{T1C}}$, $f_{\text{T2-FLAIR}}$) maps were generated with simplified IVIM modelling. Parameters associated with progression before or after 6.9 months (early vs late progression, respectively), overall survival (OS) and progression-free survival (PFS) were investigated.

**Results:** Higher $\text{ADC}_{\text{T2-FLAIR}}$ at baseline [Odds Ratio (OR) = 1.06, 95% CI 1.01–1.15, $p = 0.025$], lower $f_{\text{T2-FLAIR}}$ at fraction 10 (OR = 2.11, 95% CI 1.04–4.27, $p = 0.018$), and lack of increase in $\text{ADC}_{\text{T2-FLAIR}}$ at fraction 20 compared to baseline (OR = 1.12, 95% CI 1.02–1.22, $p = 0.02$) were associated with early progression. Combining $\text{ADC}_{\text{T2-FLAIR}}$ at baseline, $f_{\text{T2-FLAIR}}$ at fraction 10, ECOG and MGMT promoter methylation status significantly improved AUC to 90.3% compared to a model with only ECOG and MGMT promoter methylation status ($p = 0.001$). Using multivariable analysis, neither IVIM metrics were associated with OS but higher $f_{\text{T2-FLAIR}}$ at fraction 10 (HR = 0.72, 95% CI 0.56–0.95, $p = 0.018$) was associated with longer PFS.

**Conclusion:** $\text{ADC}_{\text{T2-FLAIR}}$ at baseline, its lack of increase from baseline to fraction 20, or $f_{\text{T2-FLAIR}}$ at fraction 10 significantly predicted early progression. $f_{\text{T2-FLAIR}}$ at fraction 10 was associated with PFS.

**Keywords**

Glioblastoma; Survival; Progression-free Survival; Diffusion MRI; Intravoxel incoherent motion imaging; Radiotherapy

For patients with glioblastoma, median survival has not significantly changed from 6.9 months since the landmark Stupp trial in 2005 [1]. The current standard of care is maximal safe surgical resection followed by chemoradiation. The radiation planning MRI is the last imaging timepoint before the start of chemoradiation. Subsequently, the first MRI is obtained 6–8 weeks after completing chemoradiation, translating to approximately 3 months where no imaging is performed.

The challenge with early imaging response determination is threefold. First, we do not routinely image during chemoradiation; therefore, there is no information beyond clinical assessments until the first post-chemoradiation MRI. Second, the interpretation of the first post-chemoradiation MRI is confounded by changes associated with radiation, such as pseudoprogression [2,3]. Lastly, we remain reliant on crude changes in 2-dimensional measurements from standard MRI to determine response without advanced quantitative imaging. Should we have the ability to know during or soon after chemoradiation which patients would not benefit from standard of care using imaging biomarkers, we could better individualize patient care pathways early on.
Intravoxel incoherent motion (IVIM) imaging is based on diffusion-weighted imaging (DWI) MRI and can provide separate estimates for quantitative metrics of diffusion, representative of structural changes [4], and the perfusion fraction [5] which is a measure related to microcirculation. Furthermore, it has the benefit of not requiring the use of gadolinium-based contrast agents, and short acquisition times. Few studies have investigated diffusion and/or perfusion MRI parameters in glioblastoma during chemoradiation [6–8], and they have been limited with respect to: sample size; assessment of only enhancing tumor as opposed to non-enhancing T2-FLAIR hyperintense regions; mixed sample populations including patients with recurrent glioblastoma on Bevacizumab or other clinical trial drugs; and/or, typically the intra-treatment MRI was acquired at only one timepoint over the course of chemoradiation. A summary of these differences is provided in Supplementary Table 1 [6–18].

We propose using a previously validated simplified IVIM model [19,20] to investigate the prognostic potential of diffusion and perfusion metrics at multiple timepoints over the course of chemoradiation in patients with glioblastoma, with the enhancing tumor and surrounding non-enhancing T2-FLAIR hyperintense regions interrogated separately. For the simplified IVIM model, 3 b-values were be used (0, 500, and 1000 s/mm$^2$), as this follows consensus guidelines for brain tumor imaging in clinical trials for DWI [21] and confers the additional benefit of not requiring specialized hardware or pulse sequence design.

**Materials & methods**

**Patient population**

This prospective study was approved by the local institutional Research Ethics Board (#430–2015). Written informed consent was obtained from all patients. A total of 50 consecutive patients with a potential new high-grade glioma diagnosis on neuroimaging were considered for recruitment. Twelve patients were excluded: four with WHO grade II or III gliomas on pathology; three withdrew; three discontinued chemoradiation; one progressed with treatment interruptions; and one had an IDH mutation, which is a distinctly different molecular and clinical profile compared to IDH wildtype (IDHwt) glioblastoma [22]. Therefore, a total of 38 patients with IDHwt glioblastoma who completed standard radiation (60 Gy in 30 fractions) concurrent with temozolomide were included. Prognostic factors such as age at diagnosis, Eastern Cooperative Oncology Group (ECOG) performance status, extent of resection, IDH status and MGMT promoter methylation status (MGMT$_{PMS}$) were extracted from electronic medical records.

**MR imaging**

All patients were scanned on a single 1.5 T Philips Ingenia system (Philips Medical Systems, Best, The Netherlands). Acquired sequences included 3D T2-FLAIR, pre- and post-contrast 3D T1-weighted (T1) and echo planar DWI with three b-values (0, 500, and 1000 s/mm$^2$) [21]. DWI was trace-weighted with an average of 3 orthogonal directions, and the addition of the 500 s/mm$^2$ b-value requires less than a minute of additional scan time compared to scanning with only b-values of 0 and 1000 s/mm$^2$. No signal averaging was
performed at \( b = 0 \) s/mm\(^2\). Detailed imaging parameters are provided in Supplementary Table 2. Patients were scanned at radiation planning, and at fractions 10 and 20.

T2-FLAIR images were coregistered to T1 post-contrast (T1C) images using Elastix registration software [23]. Volumes of interest (VOIs) were then manually delineated using the semi-automatic thresholding software Amira (version 2019.2, Thermo Fischer Scientific, Berlin, Germany). VOIs consisted of non-overlapping T1 enhancing (Fig. 1A, 1F) and surrounding nonenhancing T2-FLAIR hyperintense regions (Fig. 1B, 1G). Areas of intrinsic T1 hyperintensity representing hemorrhagic material were not included in T1C contour delineations. Necrotic/cystic regions, surgical cavity and large vessels were excluded from all VOIs. VOIs were then coregistered to \( b = 0 \) of DWI and re-sampled with the resolution of the DWI sequence (Fig. 1C–E). VOIs were also drawn on the contralateral normal appearing white matter (cNAWM) and grey matter (cNAGM). No eddy current correction was performed.

MRI quantification

A previously validated simplified IVIM model [19,20] was used to calculate the diffusion coefficient (\( D \), Fig. 1D) and perfusion fraction (\( f \), Fig. 1E) maps. The technique assumes the DWI signal loss due to blood flow in the microvasculature has negligible contribution to DWI images acquired at high \( b \)-values. As in prior studies [19,24], voxels with values of \( f < 0\% \) or \( f > 30\% \) were considered non-physiological and excluded.

In order to provide a diffusion metric that is widely used clinically and to enable comparison with literature, the apparent diffusion coefficient (ADC) was also calculated using the slope of the natural logarithm of the \( b = 1000 \) over \( b = 0 \) images (Fig. 1C).

Statistics

Parametric maps were calculated voxelwise, and expressed as median and interquartile range (IQR) over the VOIs. All outcome definitions were calculated using the date of the baseline (post-surgical radiation planning) MRI as reference.

The primary endpoint of the study was to determine whether IVIM metrics could be used to predict progression before or after 6.9 months (209 days), the median time to progression reported by Stupp et al. [1]. The secondary endpoint of the study was to see if the same metrics were associated with progression-free survival (PFS) and overall survival (OS). PFS was defined as the time until disease progression in accordance with RANO criteria [25], and the date of death or last follow-up was used to calculate OS, both in months. PFS and OS were calculated using the Kaplan-Meier product-limit method and differences were assessed using the log-rank test. Cox regression was used for multivariable survival analyses.

Significant findings on univariate analyses were tested in multivariable analyses. Logistic regression analysis was used to assess the association of ADC, \( D \), and \( f \) with early vs. late progression, and the performance was assessed using the area under the receiver operating characteristic curve (AUC). All \( p \)-values were two-sided, and a \( p < 0.05 \) was considered
statistically significant. Statistical analysis was performed using SAS (version 9.4 for Windows; SAS Institute, Inc., Cary, NC, USA).

Results

Patient details are summarized in Table 1. Age at diagnosis ($p = 0.74$), extent of resection ($p = 0.68$), MGMT$_{PMS}$ ($p = 0.18$) and ECOG ($p = 0.06$) were not significantly associated with early or late progression in univariate analyses. Due to limited sample size, we assessed the impact of the age [26], extent of resection [27], MGMT$_{PMS}$ [28], and ECOG [29] on PFS and OS via univariate analyses to determine what covariates should be included in multivariable analyses; only MGMT$_{PMS}$ and ECOG were significant (Supplementary Fig. 1) and included in further analyses.

Five patients (13%) demonstrated pseudoprogression within the follow-up period. With retrospective follow-up, all five cases were confirmed to have true progression after 6.9 months, and therefore were considered late progressors.

Median ADC$_{cNAWM}$ was $0.76 \times 10^{-3}$ mm$^2$/s (IQR, 0.72–0.78 $\times 10^{-3}$ mm$^2$/s) and median ADC$_{cNAGM}$ was $0.88 \times 10^{-3}$ mm$^2$/s (IQR, 0.82–0.92 $\times 10^{-3}$ mm$^2$/s). Median ADC values stratified by the timepoint of acquisition, VOI, and progression status are summarized in Table 2 and Fig. 2A–B. Higher ADC$_{T2-FLAIR}$ at baseline was significantly associated with early progression after adjusting for ECOG and MGMT$_{PMS}$ (adjusted OR (aOR) = 1.06, 95% CI 1.00–1.01, $p = 0.025$). Using the median as the cut-off ($1.07 \times 10^{-3}$ mm$^2$/s) within ADC$_{T2-FLAIR}$ to predict early vs late progression, yielded a sensitivity and specificity of 76.5% and 66.7%, respectively. No significant associations with early progression were observed at any other timepoint for ADC$_{T1C}$ nor ADC$_{T2-FLAIR}$.

When assessing changes in the ADC relative to baseline, ADC$_{T1C}$ continually increased although these changes were not significantly associated with outcome (fraction 10, $p = 0.17$; fraction 20, $p = 0.22$). Comparatively, ADC$_{T2-FLAIR}$ of early progressors did not increase from baseline to fraction 20 whereas ADC of late progressors increased between these timepoints. Lack of increase in ADC$_{T2-FLAIR}$ from baseline to fraction 20 was associated with early progression (aOR = 1.12, 95% CI 1.02–1.22, $p = 0.02$).

With regards to $D$, as $D$ is known to approximate ADC when the effect of perfusion fractions are small as in the case for brain [30], their correlation was assessed for every timepoint of acquisition and VOI. At every timepoint and VOI, a correlation above 94.5% and $p < 0.0001$ were observed. To prevent multicollinearity, and since ADC is more commonly used in the clinical setting, all subsequent analyses was performed using ADC rather than $D$. Further details on $D$ are provided in the Supplementary Table 3 and Supplementary Fig. 2.

With regards to $f$, median $f_{cNAWM}$ was 10.4% (IQR, 9.7–11.6%) and median $f_{cNAGM}$ was 12.5% (IQR, 11.1–12.9%). Median $f$, stratified by timepoint of acquisition, VOI, and progression status are shown in Table 2 and Fig. 2C–D. Early progressors demonstrated lower $f$ at all timepoints, although this was only significant at fraction 10 where lower $f$ was associated with early progression in both T1C (aOR = 1.87, 95% CI 1.11–3.15, $p = 0.018$) and T2-FLAIR (aOR = 2.11, 95% CI 1.04–4.27, $p = 0.039$). Using the median as the cut-off
(12%) for \( f_{T2-FLAIR} \) at fraction 10 to predict early vs late progression, yielded a sensitivity and specificity of 70.6% and 66.7%, respectively. Similarly, using the median as the cut-off (15%) for \( f_{T1C} \) at fraction 10 yielded a sensitivity and specificity of 64.7% and 61.9%, respectively.

When assessing changes in \( f \) relative to baseline, no significant changes for \( f_{T1C} \) (fraction 10, \( p = 0.30 \); fraction 20, \( p = 0.24 \)) or \( f_{T2-FLAIR} \) (fraction 10, \( p = 0.75 \); fraction 20, \( p = 0.98 \)) were observed, and none were associated with early progression.

When assessing our primary endpoint, median ADC\(_{T2-FLAIR}\) at baseline, \( f_{T1C} \) and \( f_{T2-FLAIR} \) at fraction 10 were combined into a single model and adjusted for ECOG and MGMT\(_{PMS}\) to assess whether these variables could better predict progression status (early vs late). \( f_{T1C} \) at fraction 10 was not significant in the presence of median of ADC\(_{T2-FLAIR}\) at baseline and \( f_{T2-FLAIR} \) at fraction 10 (\( p = 0.15 \)), and therefore was not included in subsequent multivariable analyses.

To assess the contribution of ADC\(_{T2-FLAIR}\) and \( f_{T2-FLAIR} \) in predicting outcome, we compared the performance between the combinations of ECOG and MGMT\(_{PMS}\) vs ADC\(_{T2-FLAIR}\) and \( f_{T2-FLAIR} \) vs all 4 parameters together. The model with ECOG and MGMT\(_{PMS}\) had an AUC of 73.4%, whereas IVIM parameters had an AUC of 83.9%. The combination of all 4 parameters in a model was significantly improved with an AUC of 90.3% (\( p = 0.001 \)). In the multivariable model, all parameters except for ECOG (aOR = 0.15, 95% CI 0.02–1.14, \( p = 0.066 \)) were significant (ADC\(_{T2-FLAIR}\), aOR = 1.08, 95% CI 1.01–1.15, \( p = 0.028 \); \( f_{T2-FLAIR} \), aOR = 2.32, 95% CI 1.07–5.05, \( p = 0.034 \); MGMT\(_{PMS}\), aOR = 0.08, 95% CI 0.01–0.86, \( p = 0.039 \)).

When assessing our secondary endpoint using Kaplan Meier, the median ADC\(_{T2-FLAIR}\) at baseline (1.07 × 10\(^{-3}\) mm\(^2\)/s) was significantly associated with OS (\( p = 0.037 \); Fig. 3A) but not PFS (\( p = 0.48 \); Fig. 3C), with higher ADC\(_{T2-FLAIR}\) significantly associated with a shorter OS. Conversely, the median \( f_{T2-FLAIR} \) at fraction 10 (12%) was significantly associated with PFS (\( p = 0.016 \); Fig. 3D) but not OS (\( p = 0.48 \), Fig. 3B), with lower \( f_{T2-FLAIR} \) significantly associated with shorter PFS. Combining these parameters, patients with both high ADC\(_{T2-FLAIR}\) and low \( f_{T2-FLAIR} \) had significantly worse OS (\( p = 0.026 \)) and PFS (\( p = 0.002 \)).

Results from Cox regression are summarized in Table 3. While an ECOG of 0 and methylated MGMT\(_{PMS}\) demonstrated significant associations with longer OS (HR = 0.19, 95% CI 0.063–0.59, \( p = 0.004 \); and HR = 0.28, 95% CI 0.1–0.8, \( p = 0.017 \), respectively), neither IVIM parameters were significant. Higher \( f_{T2-FLAIR} \) (HR = 0.72, 95% CI 0.56–0.95, \( p = 0.018 \)) and methylated MGMT\(_{PMS}\) (HR = 0.25, 95% CI 0.1–0.63, \( p = 0.003 \)) were associated with longer PFS.

**Discussion**

ADC and \( f \) of the entire T1C, and the surrounding nonenhancing T2-FLAIR volumes, were evaluated before and during chemoradiation in patients with newly diagnosed, IDHwt glioblastoma. While there are advantages to more sophisticated IVIM models (e.g., ability to
calculate the pseudo-diffusion coefficient, $D^*$ [12]), we chose the simplified model to increase the generalizability and clinical utility of the results. We demonstrated that various quantitative metrics from the T2-FLAIR at baseline, and at fractions 10 and 20, can act as prognostic biomarkers. Metrics from the T2-FLAIR volume exhibited stronger associations with outcome compared to those from the T1C volume, with $f_{T2-FLAIR}$ significantly predictive of PFS.

We report that the use of the simplified IVIM model here generated similar values for ADC within the cNAWM and cNAGM as those reported in the literature [31–33], and similarly for $f_{cNAWM}$ [12,14,19,24,34–36] and $f_{cNAGM}$ [36,43], lending strength to our results.

We further report that at baseline, a greater ADC$_{T2-FLAIR}$ value is significantly associated early progression, and trended towards worse OS. While previous, single time-point studies found that lower ADC was associated with poorer outcomes [9,12] (Supplementary Table 1), significant methodological differences may be the cause of this discordance. In these earlier studies, circular regions of interest were drawn over areas of highest and lowest ADC on the pre-operative MRI of patients with grade 2 to 4 gliomas. Comparatively, we reported on the entirety of the T1C and T2-FLAIR regions on post-operative MRI, where all or most of the enhancing tumor has already been resected, as most (81.6%) of our cases underwent gross-or sub-total resection. In addition, on radiation planning MRI the T1C may include tumor but also subacute ischemic changes and granulation tissue compared to the pre-operative MRI where the T1C is only tumor. Moreover, our patient dataset was limited but homogenous, comprising of only patients with wildtype IDH glioblastomas. We hypothesize that greater ADC$_{T2-FLAIR}$ after post-surgical resection reflects a more biologically aggressive tumor, which is the cause of earlier progression ($p = 0.029$, Fig. 2B). This effect was also observed in Li et al. [6], who reported higher normalized ADC values in the T2-FLAIR of patients that died within the year compared to those that survived longer, although it was not significantly related to OS or PFS. We surmise that more aggressive tumors cause more disruption of blood brain barrier and increased interstitial edema leading to increased ADC, whereas less aggressive tumors cause less disruption of blood brain barrier.

During chemoradiation, the relative change in the ADC may be more reflective of a direct relationship between cellularity, tumor cell kill, and interstitial edema. We hypothesize that the lack of increase in ADC$_{T2-FLAIR}$ by fraction 20 compared to baseline ($p = 0.02$) of early progressors reflects persistent tumour burden or even cell proliferation and hence, represents a surrogate of treatment resistance. This hypothesis is supported by a recent study that utilized functional diffusion maps in patients with Grade 3 and 4 gliomas before and midway through radiotherapy, and found that patients with a high percentage of unchanging ADC compared to baseline had worse prognosis [8]. Similar results were also observed in another study evaluating ADC in patients with brain metastases during a course of 30 Gy in 10 fractions of whole brain radiation therapy, where it was reported that ADC decreased during treatment in non-responding tumors [37], and likewise in Li et al. [6] and Wen et al. [7], who studied post-operative patients with newly diagnosed glioblastoma. However, differences in methodology and the inclusion of patients undergoing mixed therapies limit direct comparison with our results, and none of the studies reported the status of IDH mutation or MGMT$_{PMS}$. 

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With regards to $f$, our reported values fall within the large range of values reported in the literature [12,14,19,34,35,38]. This variation is likely due to differences in acquisition protocols and patient populations. We observed $f$ to be higher at all timepoints in late progressors within both T2-FLAIR and T1C volumes (Fig. 2C–D). $f$ is related to relative cerebral blood volume (rCBV) [5,12] which is highly associated with relative cerebral blood flow (rCBF) in brain tumors [10,39]. Within this context, higher rCBF has been shown to be associated with better prognosis in glioblastoma [40–42]. Therefore, our results are consistent with better overall perfusion within the tumor capillaries and subsequently, better responsiveness during chemoradiation. Furthermore, our results may signify the importance of early vascular normalization processes that enhance perfusion. However, it is also possible that the increase observed is due to a free water effect due to increased interstitial water and edema. Moreover, $f$ may be affected by permeability effects, particularly when there is a compromised blood brain barrier. Further studies are required to validate this hypothesis.

To the authors’ knowledge, there are a limited number of studies that have investigated $f$ in patients with glioblastoma. Puig et al. [10] investigated the association of IVIM metrics with outcomes in patients with newly diagnosed glioblastoma prior to surgery and found that patients with higher $f_{T1C}$ had shorter OS, with no association between $f_{T2-FLAIR}$ and outcome. Federau et al. [9] also observed a higher $f$ within high-grade gliomas, however, no association was observed between the $f_{T1C}$ and OS, and T2-FLAIR was not investigated. Both studies utilized pre-operative MRIs and $f$ was calculated using the full IVIM model; VOIs were then manually placed over the highest $f$ for analyses. These methodological differences may be the cause of the discrepancies of our results.

When analyzing the combined effect of the median ADC$_{T2-FLAIR}$ at baseline and median $f_{T2-FLAIR}$ at fraction 10, adjusted for MGMT$_{PMS}$ and ECOG, ADC$_{T2-FLAIR}$ trended towards significance for OS whereas $f_{T2-FLAIR}$ maintained its significance for PFS (Table 3). As ADC$_{T2-FLAIR}$ was significant on univariate analysis ($p = 0.04$), and a significant increase in AUC was observed when ECOG, MGMT$_{PMS}$, and IVIM parameters were combined ($p = 0.001$), we suspect that the loss of significance for ADC$_{T2-FLAIR}$ is primarily due to power. We hypothesize that understanding both metrics is important to determine tumour responsiveness.

This study has limitations. The data may be biased due to the single-centre nature of the data; furthermore, all scans were from a single vendor. However, these may also represent a strength as the consistency allows for less variability in the acquisitions. Furthermore, inclusion of the entire T2-FLAIR hyperintensity and T1C as opposed to subjective placement of regions of interest of a defined size is a strength, though validation in a larger, multi-institutional cohort with different scanners is still required. Compared to conventional IVIM models, using only 3 b-values of diffusion images without eddy currents correction affects the accuracy of IVIM parameters leading to the potential for increased variation. However, our suggested model improves feasibility and facilitates translation and implementation into daily clinical practice.
In conclusion, when using simplified IVIM metrics to differentiate early versus late progression, ADC\textsubscript{T2-FLAIR} at baseline, as well as \( f_{T2-FLAIR} \) and \( f_{T1C} \) at fraction 10 were significant, though \( f_{T1C} \) at fraction 10 did not maintain its significance in the presence of the other two parameters. A model combining ADC\textsubscript{T2-FLAIR} at baseline, \( f_{T2-FLAIR} \) at fraction 10, MGMT\textsubscript{PMS} and performance status demonstrated higher AUC compared to when variables were assessed separately. Lack of increase in ADC\textsubscript{T2-FLAIR} over the course of chemoradiation was associated with early progression, and can potentially act as an imaging biomarker of response to treatment in differentiating progression. In time-dependent analyses, higher ADC\textsubscript{T2-FLAIR} at baseline was significantly associated with shorter OS, and a lower \( f_{T2-FLAIR} \) at fraction 10 was associated with shorter PFS. In a multivariable model with ECOG and MGMT\textsubscript{PMS}, no IVIM metrics were significantly associated with OS, however \( f_{T2-FLAIR} \) at fraction 10 retained its significance with PFS.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Abbreviations:

- DWI: diffusion-weighted imaging
- T2-FLAIR: T2-weighted fluid-level attenuated inversion recovery
- IVIM: intravoxel incoherent motion
- IDH: isocitrate dehydrogenase 1
- IDHwt: wildtype IDH
- ECOG: Eastern Cooperative Oncology Group
- MGMT: \( O^6 \)-methylguanine-DNA methyltransferase
- VOI: volume of interest
- cNAWM: contralateral normal appearing white matter
- cNAGM: contralateral normal appearing grey matter
- ADC: apparent diffusion coefficient
- RANO: response assessment in neuro-oncology
- AUC: area under the receiver operating characteristic curve
- aOR: adjusted odds ratio

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Fig. 1.
Images of a patient included in the study. Contrast-enhanced T1 image (A), T2-FLAIR image (B), ADC map (C), $D$ map (D) and $f$ map (E). Region of interest (green) of the enhancing parts of the mass (F), region of interest (magenta) of non-enhancing T2-FLAIR hyperintensity (G), with both regions of interest overlaid on ADC map (H), $D$ map (I) and $f$ map (J). ADC and $D$ color bar scale is $\times 10^{-3}$ mm$^2$/s, while $f$ color bar is shown in percent.
Fig. 2.
Changes in median ADC (A, B) and f (C, D) values during chemoradiation in T1 contrast-enhancing areas (A, C) and nonenhancing FLAIR hyperintense areas (B, D) stratified by early (red) vs. late (blue) progressors. Vertical lines represent one standard deviation, and stars (*) indicate where significant differences were observed.
Fig. 3.
Kaplan-Meier analyses demonstrating the association with ADC<sub>T2-FLAIR</sub> (A, C) and f<sub>T2-FLAIR</sub> (B, D) on OS (A–B) and PFS (C–D). Grey vertical line on the plots indicates the 6.9 month timepoint which reflects the threshold for early vs. late progression used for this analyses.
### Table 1

Patient demographics.

| Patients                             | All patients ($n = 38$) | Early progressors ($n = 17$) | Late Progressors ($n = 21$)* |
|--------------------------------------|-------------------------|-----------------------------|------------------------------|
| Male ($n, \%$)                       | 25 (65.8%)              | 11 (64.7%)                  | 14 (66.6%)                   |
| Median age at diagnosis (range)      | 57.50 (20–68)           | 57 (20–68)                  | 58 (41–67)                   |
| MGMT promoter methylation            |                         |                             |                              |
| Methylated                           | 19 (50.0%)              | 7 (41.2%)                   | 12 (57.1%)                   |
| Unmethylated                         | 15 (39.5%)              | 9 (52.9%)                   | 6 (28.6%)                    |
| Unknown                              | 4 (10.5%)               | 1 (5.9%)                    | 3 (14.3%)                    |
| ECOG at diagnosis                    |                         |                             |                              |
| 0                                    | 24 (63.2%)              | 8 (47.1%)                   | 16 (76.2%)                   |
| 1                                    | 13 (34.2%)              | 8 (47.1%)                   | 5 (23.8%)                    |
| 2                                    | 1 (2.6%)                | 1 (5.9%)                    | 0 (0%)                       |
| Extent of Resection                  |                         |                             |                              |
| Gross total resection ($n, \%$)      | 9 (23.7%)               | 3 (17.6%)                   | 6 (28.6%)                    |
| Sub-total resection ($n, \%$)        | 22 (57.9%)              | 10 (58.9%)                  | 12 (57.1%)                   |
| Biopsy Only ($n, \%$)                | 7 (18.4%)               | 4 (23.5%)                   | 3 (14.3%)                    |
| Median number of days between the baseline MRI and initiation of chemoradiation (range) | 7 (0–14) | 6 (0–14) | 7 (7–12) |
| MRIs Completed                       |                         |                             |                              |
| Fraction 0 ($n, \%$)                 | 38 (100%)               | 17 (100%)                   | 21 (100%)                    |
| Fraction 10 ($n, \%$)                | 38 (100%)               | 17 (100%)                   | 21 (100%)                    |
| Fraction 20 ($n, \%$)                | 36 (94.7%)              | 16 (94.1%)                  | 20 (95.2%)                   |
| Median time to progression (range) in months | 7.2 (1.8–21.9) | 4.3 (1.8–6.9)*** | 11.1 (7.1–21.9)*** |
| Median follow-up (range) in months **| 14.5 (1.8–27.9) | 8.9 (1.8–18.2) | 18.4 (8.6–27.9) |

* At the date of last inquiry, 5 patients (13.2%) had not experienced progression of disease. As this date was > 6.9 months from baseline MRI, we considered these patients as late progressors.

** At the date of last inquiry, 18 patients (4 of the early progressors and 14 of the late progressors) were still alive and were therefore censored.

*** 2 patients progressed, one of whom passed away, prior to their standard of care 6–8 week post-treatment scan. Their date of death was used as their date of progression.
Median and interquartile range (IQR) for ADC and $f$, stratified by outcome. Significant values as determined by univariate analysis are shown in bold, and show the association between the parameter with early progression.

| Map            | Baseline MRI | Fraction 10 MRI | Fraction 20 MRI |
|----------------|--------------|-----------------|-----------------|
|                | T1C          | T2-FLAIR        | T1C             | T2-FLAIR        | T1C                  | T2-FLAIR              |
| ADC ($10^{-3}$ mm$^2$/s) |              |                 |                 |                 |                      |                      |
| Early          | 1.05 (0.96–1.17) | **1.20 (1.07–1.27)** | 1.02 (0.90–1.17) | 1.11 (1.01–1.29) | 1.08 (0.95–1.20) | 1.14 (0.99–1.25) |
| Late           | 1.06 (0.97–1.12) | **1.01 (0.91–1.14)** | 1.06 (0.99–1.29) | 1.07 (0.94–1.30) | 1.14 (1.01–1.30) | 1.11 (1.01–1.29) |
| $p$-value      | 0.77         | **0.029**       | 0.14            | 0.53            | 0.16               | 0.97               |
| $f$ (%)        |              |                 |                 |                 |                      |                      |
| Early          | 14.1 (13.2–15.0) | **13.9 (12.6–15.5)** | 11.7 (10.7–11.9) | **11.3 (10.7–12.5)** | 14.3 (13.5–15.4) | 11.6 (10.9–12.4) |
| Late           | 14.6 (13.6–15.8) | **15.3 (14.1–16.4)** | 12.0 (10.7–14.0) | **12.5 (11.7–13.8)** | 15.1 (14.1–15.8) | 12.2 (11.5–13.5) |
| $p$-value      | 0.21         | 0.045           | **0.028**       | 0.35            | 0.06               |                    |

*After adjusting for ECOG and MGMT promoter methylation status, the significance for these variables were: $\text{ADC}_{T2-FLAIR}: p = 0.025$; $\text{DT}_{T2-FLAIR}: p = 0.011$; $\text{f}_{T1C}: p = 0.018$; $\text{f}_{T2-FLAIR}: p = 0.039$. 
Table 3

Cox regression analysis using median ADC_{T2-FLAIR} from baseline, median f_{T2-FLAIR} from fraction 10, adjusted for ECOG and MGMT promoter methylation status.

| Outcome | Parameter                                      | HR    | 95% CI     | p-value |
|---------|-----------------------------------------------|-------|------------|---------|
| OS      | Baseline median ADC_{T2-FLAIR} (1.07 × 10^{-3} mm^2/s) | 1.03  | 0.1–1.06   | 0.1     |
|         | Median f_{T2-FLAIR} from fraction 10 (12%)    | 0.84  | 0.61–1.14  | 0.25    |
|         | ECOG 0 vs 1–2                                 | 0.19  | 0.063–0.59 | 0.004   |
|         | MGMT promoter methylation status (methylated vs unmethylated) | 0.28  | 0.096–0.8  | 0.017   |
| PFS     | Baseline median ADC_{T2-FLAIR} (1.07 × 10^{-3} mm^2/s) | 1.01  | 0.99–1.04  | 0.43    |
|         | Median f_{T2-FLAIR} from fraction 10 (12%)    | 0.72  | 0.56–0.95  | 0.018   |
|         | ECOG 0 vs 1–2                                 | 0.41  | 0.17–1.01  | 0.052   |
|         | MGMT promoter methylation status (methylated vs unmethylated) | 0.25  | 0.097–0.63 | 0.0033  |

Significant values in bold.