The early interaction of the outer membrane protein PhoE with the periplasmic chaperone Skp occurs at the cytoplasmic membrane.

Harms, N.; Koningstein, G.; Dontje, W.; Muller, M.; Oudega, B.; Luirink, S.; Cock, H.

published in
Journal of Biological Chemistry
2001

DOI (link to publisher)
10.1074/jbc.M011194200

document version
Publisher's PDF, also known as Version of record

Link to publication in VU Research Portal

citation for published version (APA)
Harms, N., Koningstein, G., Dontje, W., Muller, M., Oudega, B., Luirink, S., & Cock, H. (2001). The early interaction of the outer membrane protein PhoE with the periplasmic chaperone Skp occurs at the cytoplasmic membrane. Journal of Biological Chemistry, 276, 18804-18811. https://doi.org/10.1074/jbc.M011194200

General rights
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
• You may not further distribute the material or use it for any profit-making activity or commercial gain
• You may freely distribute the URL identifying the publication in the public portal

Take down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

E-mail address:
vuresearchportal.ub@vu.nl

Download date: 12. Jun. 2022
The Early Interaction of the Outer Membrane Protein PhoE with the Periplasmic Chaperone Skp Occurs at the Cytoplasmic Membrane*

Nellie Harms§, Gregory Koningstein‡, Wendy Dontje‡, Matthias Muller‡, Bauke Oudega‡, Joen Luirink‡, and Hans de Cock¶

From the §Department of Molecular Microbiology, Institute of Molecular Biological Sciences, Biocentrum Amsterdam, De Boelelaan 1087, Amsterdam 1081 HV, The Netherlands, ¶Molecular Microbiology and Institute of Biomembranes, Utrecht University, Padualaan 8, Utrecht 3584 CH, The Netherlands, and the Institut für Biochemie und Molekularbiologie, Universität Freiburg, Herman-Herder-Strasse 7, D-79104 Freiburg, Germany

Spheroplasts were used to study the early interactions of newly synthesized outer membrane protein PhoE with periplasmic proteins employing a protein cross-linking approach. Newly translocated PhoE protein could be cross-linked to the periplasmic chaperone Skp at the periplasmic side of the inner membrane. To study the timing of this interaction, a PhoE-dihydrofolate reductase hybrid protein was constructed that formed translocation intermediates, which had the PhoE moiety present in the periplasm and the dihydrofolate reductase moiety tightly folded in the cytoplasm. The hybrid protein was found to cross-link to Skp, indicating that PhoE closely interacts with the chaperone when the protein is still in a transmembrane orientation in the translocase. Removal of N-terminal parts of PhoE protein affected Skp binding in a cumulative manner, consistent with the presence of two Skp-binding sites in that region. In contrast, deletion of C-terminal parts resulted in variable interactions with Skp, suggesting that interaction of Skp with the N-terminal region is influenced by parts of the C terminus of PhoE protein. Both the soluble as well as the membrane-associated Skp protein were found to interact with PhoE. The latter form is proposed to be involved in the initial interaction with the N-terminal regions of the outer membrane protein.

The majority of the proteins destined for the extracytoplasmic environment in *Escherichia coli* are synthesized as precursors in the cytoplasm that are translocated across the inner IM in a non-native state. To reach their final conformation and destiny they need to be prevented from premature folding and degradation in the cytoplasm. It is well established that molecular chaperones and folding catalysts are required to channel proteins into productive export, folding, and assembly pathways. In the cytoplasm one of the first events that takes place when a nascent PhoE emerges from the ribosome is the interaction with trigger factor, a cytoplasmic chaperone. Using a cross-linking approach, the latter protein was found to interact with nascent PhoE chains as short as 57 amino acids (1–3). Subsequently, at a late co-translational or early post-translational stage the precursor protein interacts with SecB (4–6). This SecB-precursor complex is then targeted to SecA, the peripheral subunit of the membrane-embedded SecYEG complex (7). PhoE precursors are then translocated across the IM in a non-native state. When appearing at the periplasmic side of the IM, these polypeptides undergo complex folding processes and have to be prevented from misfolding, aggregating, and degrading. Proteins destined for the outer membrane or the extracellular environment require correct targeting to these locations. Periplasmic chaperones and folding catalysts have been implicated to be involved in the folding and assembly pathway of OMPs. It was demonstrated previously that OMPs are exposed to the periplasmic chaperones during the assembly process (8). A periplasmic system consisting of molecular chaperones and folding catalysts has been described (9, 10). Among them, the peptidyl-prolyl isomerases (FkpA, SurA, RotA, and PpiD) carry out the isomerization around the Xaa-Pro peptide bond. In addition to the isomerase activity the FkpA and SurA proteins were found to possess chaperone activity (11, 12). The second type of folding catalysts are the protein disulfide isomerase (Dsb proteins) carrying out disulfide bond formation and thiol-disulfide exchanges (13).

In addition, Skp functions as a molecular chaperone in the periplasm of Gram-negative bacteria, possibly specifically required for the biogenesis of OMPs. Skp is a basic soluble protein in the periplasm, but it was also found in IM fractions (14, 15). The protein exists in two different states as was demonstrated by their different sensitivity toward proteases (16). The conversion between these states can be modulated in *vitro* by phospholipids, lipopolysaccharides, and bivalent cations (16). The function of these two different forms of Skp in *vivo* is not yet known. Previously, it was demonstrated that Skp binds selectively to OmpA and proteins of the bacterial porin family (16, 17). In addition, non-OMPs seem to act as a substrate for binding (18). Skp interacts with OmpA in close vicinity of the IM, and it was found to be involved in the release of the OMP from this membrane (15). In addition, cells defective in Skp and the periplasmic protease DegP accumulate protein aggregates in the periplasm (15). These results suggest that Skp is a molecular chaperone involved in generating and maintaining solubility of early folding intermediates of OMPs in the periplasm.

The molecular mechanism of the PhoE assembly pathway

---

* The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1794 solely to indicate this fact.

§ To whom correspondence should be addressed. Tel.: 31-20-444-7176; Fax: 31-20-444-7136; E-mail: harms@bio.vu.nl.

† The abbreviations used are: IM, inner or cytoplasmic membrane; OMP, outer membrane protein; IPTG, isopropyl-1-thio-D-galactopyranoside; BS3, bis(sulfosuccinimidyl) suberate; RNC, ribosome nascent chain complex; DSS, disuccinimidyl suberate; DHFR, dihydrofolate reductase.
Skp Interacts with a PhoE Translocation Intermediate

has been investigated extensively in vivo and in vitro. However, little is known about which periplasmic chaperones and folding catalysts are involved in the late stages of translocation over the IM, in the release from the IM, and in the early stages of folding in the periplasm. Furthermore, the order of interactions with accessory proteins and the timing of the various reactions are largely unknown. In the studies described in this report, we use an unbiased approach to study these processes. We present evidence that the initial interaction of the non-native porin protein PhoE at the periplasmic side of the IM is with Skp. This interaction occurs already early during translocation of PhoE, i.e. when PhoE is still in a transmembrane orientation in the translocase. At the N terminus of PhoE, two Skp-binding sites could be identified. In contrast to OmpA, release of PhoE from the IM appeared not to be dependent on Skp.

**EXPERIMENTAL PROCEDURES**

**Bacterial Strains and Growth Conditions**—The E. coli K12 strain MC4100 (F′ lacU169 araD138 rpsL thi relA) was used for the spheroplast labeling experiments and for the isolation of lysate and inner membrane vesicles (19). MC4100 and RC354c, containing a deletion in the skp gene (17), were used for the isolation of periplasmic extracts (20). BG87 (21) was used for the isolation of trigger factor-depleted membrane vesicles (19). MC4100 and RC354c, containing a deletion in the phoE-dhfr gene (a gift from T. Langer, Munich, Germany). Subsequently, the phoE-dhfr fusion gene was cloned downstream of the arabinose promoter on plasmid pMPK4 (26). Plasmid pGAH317, carrying the wild-type skp gene, has been described (27).

**Spheroplast Labeling, Cross-linking, and Release**—For pulse labeling of spheroplast cells were grown in GB1 medium with casamino acids and 35S-methionine. During the chase 50 μg/ml chloramphenicol (30 μg/ml) were added when indicated, and in the case of a cross-link experiment also 1 mM bis(sulfosuccinimidyl) suberate (BS3) dissolved in labeling medium (Pierce). After the 5-min incubation the spheroplasts were washed twice with quench buffer (0.1 mM glycine, 10 mM NaHCO3, 250 mM sucrose, pH 8.5). The spheroplasts were subsequently collected by centrifugation for 2 min at 10,000 × g and resuspended in 0.5 ml of cold H2O.

**Flotation Gradient Centrifugation**—To purify membranes and membrane-associated protein complexes, crude membranes were subjected to flotation gradient analysis as described previously (28).

**Pulse-Chase Labeling**—PhoE was grown either as described above and used for pulse labeling with 35S-methionine (30 μCi per ml of cells) followed by a chase with cold methionine (final concentration, 2 mM). Samples were taken at several time intervals and precipitated with trichloroacetic acid.

**RESULTS**

Skp Interacts with PhoE at the Periplasmic Side of the Cytoplasmic Membrane—PhoE is translocated over the IM in a non-native state. Upon entering the periplasm, the protein has to be released from this membrane to assemble into the outer membrane in its native trimeric structure. We wanted to determine which chaperones and folding catalysts assist in these processes and to study the timing of the interaction with these accessory proteins. To that end, we used an unbiased approach to analyze the molecular environment of the newly translocated PhoE protein or PhoE translocation intermediates at the periplasmic side of the IM. MC4100 and MC4100 Δskp containing inducible phoE gene were collected into spheroplasts, incubated for 5 min with 35S-methionine, and chased with cold methionine. Subsequently, cells were treated with the membrane-impermeable cross-linker BS3 either in the presence or absence of added periplasmic extract. After immunoprecipitating the PhoE proteins in the pellet or...
Supernatant fractions with anti-sera against PhoE, proteins that interact with this OMP were identified. As shown in Fig. 1, lanes 13 and 14, a cross-linking product with a molecular mass of ~52 kDa was found in the supernatant fraction. The 52-kDa product corresponds in size to the combination of one molecule of PhoE (36 kDa) and one Skp (16 kDa). The band was absent when periplasmic proteins were not added (lane 11) or when a periplasmic extract of a skp mutant was used (lane 12). These data indicate that Skp closely interacts with PhoE during or after the process of secretion into the spheroplast medium, as was shown previously (16).

In the spheroplast pellet fraction cross-linking products of ~71, 57, and 52 kDa were detected (lanes 5–8), suggesting that these products are still associated to the IM. The intensity of the 71- and 57-kDa bands was independent of the addition of periplasmic proteins. This indicates that the cross-linking adducts were not periplasmic, but membrane-associated proteins. The cross-linking adducts were not further identified, but the size of the 71-kDa product suggests that this represents a PhoE dimer. The band of 52 kDa could be immunoprecipitated with a specific antiserum raised against Skp (lane 9). A faint reaction was also obtained without the addition of a periplasmic extract containing Skp (lanes 5 and 6), indicating that the residual amount of Skp associated with the spheroplasts interacted with PhoE. These results showed that Skp interacts with PhoE at the level of the IM. It is, however, also conceivable that the Skp-PhoE cross-linking product in the pellet fraction represents an interaction of Skp with PhoE aggregates. To distinguish between these possibilities, spheroplasts were lysed and IMs were purified by flotation gradient centrifugation (Fig. 2).

The vast majority of PhoE protein was associated with the membranes present in fraction 2 (see lanes 3, 4, and 6). In this fraction the IM protein YidC (32) was detected as a control protein (results not shown). Furthermore, the typical three cross-linking products of 75, 57, and 52 kDa were also detected in this membrane fraction. The 52-kDa band could be immunoprecipitated with either αPhoE (lane 6) or αSkp (lane 8). The three cross-linking bands could also be obtained when the cross-linking was carried out after the flotation (lane 4). These results showed that the Skp-PhoE interaction occurs in close proximity of the membrane, resulting in a membrane-associated PhoE-Skp complex.

The interaction of Skp with PhoE at the membrane was also investigated in vitro. The precursor of PhoE was synthesized in an E. coli-derived translation system in the presence of inverted IM vesicles. The precursors could be imported into these vesicles with an efficiency of ~25% as was monitored by their resistance against externally added proteinase K (Fig. 3, lanes 1, 4, and 7). This efficiency of targeting and insertion was also found in other studies with in vitro synthesized prePhoE (33). Cross-linking with the membrane-permeable cross-linker DSS gave rise to a cross-linking product of 52 kDa that could be immunoprecipitated with anti-serum directed against PhoE (lanes 2, 5) and Skp (lanes 3 and 6). This cross-linking product must have been formed, at least for a large part, inside the vesicles (periplasmic side), because it was resistant to externally added proteinase K (lanes 5 and 6). Importantly, this proteinase K-resistant cross-linking product could not be obtained when vesicles were used isolated from a skp mutant (lanes 13 and 14). The 52-kDa cross-linking product that could be immunoprecipitated with anti-Skp (lane 11) was degraded by the protease (lane 14), indicating that the cross-linking adduct originates from the lysate, which was isolated from the wild type.

**Skp Interacts with a Translocation Intermediate of PhoE**—We subsequently wanted to determine whether Skp interacts with PhoE during translocation across the IM. Previously, it was demonstrated that a PhoE-LacZ hybrid protein spans the IM, with the PhoE moiety in the periplasm and the β-galactosidase moiety in the cytoplasm. In addition, such LacZ...
Skp Interacts with a PhoE Translocation Intermediate

**Fig. 2. The PhoE-Skp complex is membrane-associated.** E. coli cells of wild type MC4100 containing plasmid pNN100 were converted to spheroplasts, induced with IPTG, and pulse-labeled. During the chase periplasmic extract was added from an Skp-overproducing strain to spheroplasts, induced with IPTG, and pulse-labeled. Membranes were isolated and loaded on a flotation gradient. The gradient was fractionated into four fractions, and the upper two fractions are shown. The fractions were used in immunoprecipitation experiments with either αPhoE (lanes 5, 6) or αSkp (lanes 7, 8). Lanes 1–4 contain the proteins that are present in membrane fractions that were cross-linked after the flotation. An asterisk indicates the cross-link product of PhoE with Skp.

Fusion proteins were shown to block translocation of other proteins across the IM, due to jamming of the Sec pathway (34). In other studies dihydrofolate reductase (DHFR) was used to create OmpA-DHFR translocation intermediates that have similar effects as LacZ fusions (35) (36). We made use of this strategy to study the environment of PhoE translocation intermediates. We constructed a hybrid protein containing the N-terminal 300 amino acid residues of PhoE and monomeric and oligomeric forms of Skp. The cross-linking products were not detected when the isolated RNC precipitated with control buffer (lane 5). Similar results were obtained with a shorter version of the PhoE-DHFR hybrid protein caused jamming of the Sec translocase. Proteinase K treatment of spheroplasts that were induced for synthesis of the hybrid protein and subsequently pulse labeled, showed that the PhoE moiety of the mature hybrid protein was degraded by the protease, whereas the DHFR moiety was not (results not shown). These data demonstrate that the PhoE-DHFR hybrid protein is in a transmembrane orientation, because it is accessible for protease added from the outside and it blocks the translocation apparatus. The vast majority of the translocons must be occupied with this fusion protein, because an efficient block in protein export is observed.

To analyze the environment of these translocation intermediates, spheroplasts were induced with arabinose, pulse-labeled in the presence of additional periplasmic proteins, and subsequently treated with the soluble cross-linker BS3. In the pellet fractions several cross-linked bands could be identified after immunoprecipitation with antisera raised against DHFR (Fig. 5, lane 3). The strong cross-linking product of ~97 kDa possibly represented a dimer of the hybrid protein. The very weak band of ~65 kDa could also be immunoprecipitated with antisera raised against Skp (lane 2). In addition, by using sequential immunoprecipitation we found that this cross-linking product could be immunoprecipitated first by anti-Skp and subsequently by anti-PhoE (lane 1). The 65-kDa cross-linking product was membrane-associated, because it was found in the same two fractions of a flotation gradient as the inner membrane protein YidC (32) (results not shown). These results demonstrated that the PhoE-DHFR hybrid protein interacted with Skp and that the interaction of Skp with PhoE occurred during translocation, i.e., when PhoE is in a transmembrane orientation. The amount of PhoE-DHFR-Skp complex found in this experimental setup is much less compared with the amount of PhoE-Skp found in the experiments described above and shown in Fig. 1. It is possible that cross-linking targets are not properly exposed in the hybrid protein, but it is also conceivable that a C-terminal PhoE-truncate binds less efficiently to Skp compared with the full-length protein.

**The Skp Binding Site(s) in PhoE—**The results obtained so far indicate an early interaction between Skp and the first 300 amino acids of PhoE protein. To determine which regions of the PhoE protein are important for Skp binding, we investigated whether Skp is able to bind to shorter, nascent chains of PhoE. PhoE ribosome nascent chain complexes (RNCs) were synthesized in vitro, isolated by centrifugation over a sucrose cushion, incubated with a periplasmic extract, and subsequently treated with BS3. In the past we found that nascent chains as long as ~55 amino acids could interact with trigger factor (2). Because trigger factor will block binding sites on the nascent chain in these experiments, it is required to use a trigger factor-depleted lysate. In addition we used constructs that lack a signal sequence to mimic the situation at the periplasmic side of the membrane as good as possible. As shown in Fig. 6a, an RNC containing 137 amino acids of PhoE gave rise to cross-linking products of ~32 and ~48 kDa (lane 6). Both cross-linking products could be immunoprecipitated with αSkp (lane 7) and represent most probably complexes between the 137-mer of PhoE and monomeric or oligomeric forms of Skp. The cross-linking products were not detected when the isolated RNC complexes were incubated with control buffer (lane 2–4). Similar results were obtained with a shorter version of the PhoE protein. RNCs of a 100-mer of PhoE resulted in cross-linking products of 29 and 45 kDa that could be immunoprecipitated with αSkp (Fig. 6b, lanes 5, 6). These products were found when a wild type periplasm was used but were absent when an extract was used of a skp mutant strain (lanes 2, 3). The 100-mer of PhoE exposed ~65 amino acid residues out of the
ribosome, indicating that a binding site for Skp is present at the extreme N terminus of PhoE.

Interaction of Skp with Mutant PhoE Proteins Synthesized in Vitro—De Cock et al. (16) demonstrated that Skp interacts with PhoE proteins that were synthesized in vitro, because these PhoE proteins were specifically precipitated with antibodies directed against Skp. The absence or presence of the signal sequence did not interfere with complex formation. We used this approach to study the Skp-binding site of PhoE in more detail by using a set of PhoE mutants with internal deletions in the mature part of the protein. Wild type and mutant PhoE proteins were synthesized in an E. coli derived in vitro system (Fig. 7, TL). After synthesis, purified Skp was added followed by the addition of puromycin to release all nascent chains. PhoE proteins associated to Skp were immunoprecipitated with antibodies directed against Skp. In all cases PhoE-Skp complexes could be precipitated (Fig. 7, αSkp). However, the relative amount of PhoE that could be co-precipitated varied between the different mutants (Table I). Deletion of amino acids 3–11 in the mature portion of PhoE protein reduced Skp binding efficiency only moderately by ~35% as compared with full-length PhoE protein. However, larger deletions in the N-terminal part of PhoE protein reduced the binding efficiencies much further, i.e. >90%. A deletion of amino acids 48–146 reduced the efficiency of binding another 20-fold to >99%. This can be explained by assuming that two independent Skp-binding sites exist in the N-terminal part of PhoE protein, i.e. between 3–91 and 110–174 PhoE. Deletion of amino acids 48–146 would delete a major part of both sites resulting in an almost complete reduction of binding to Skp.

A mutant PhoE protein containing the N-terminal 204 amino acids, but lacking the C-terminal part (Fig. 7, lane 9) leads to a reduction of the binding efficiency with ~65%, suggesting that indeed the N-terminal part of PhoE protein contains the most important Skp-binding sites. Interestingly, however, a smaller deletion in the C-terminal part of PhoE, amino acids 249–298 (Fig. 7, lane 7), reduced the efficiency much further (>90%). In conclusion, the N-terminal region seems to be important for Skp binding whereas different parts of the C terminus of PhoE seem to influence the binding to these N-terminal sites. However, it cannot be excluded that differences in folding characteristics of the various deletion mutants affect Skp binding due to alterations in exposure of the Skp-binding sites.

Skp Is Not Responsible for the Release of PhoE from the Inner
Skp Interacts with a PhoE Translocation Intermediate

**FIG. 5.** The 300PhoE-DHFR translocation intermediate interacts with Skp. MC4100 containing pAra300PhoE-DHFR was converted to spheroplasts, induced with arabinose, and pulse-labeled. During the chase, periplasm from an Skp-overproducing strain and the cross-linker BS³ was added (lanes 1–3). Pellet fractions were used in immunoprecipitation experiments with αDHFR (lane 3), αSkp (lane 2), or αSkp followed by αPhoE (lane 1). The cross-link product of 300PhoE-DHFR with Skp is indicated with an arrow.

**FIG. 6.** Skp interacts with ribosome nascent chain complexes of PhoE. a, ribosome nascent chain complexes of 137 amino acids of PhoE were synthesized, purified over a sucrose cushion, incubated with buffer (A) (lanes 1–4) or a periplasmic extract of an Skp-overproducing strain (D) (lanes 5–8), and subsequently cross-linked with BS³, b, a 100-amino acid nascent chain was used and either incubated with a periplasmic extract of an skp mutant (B) (lanes 1–3) or a wild type strain (C) (lanes 4–6). Cross-linked products were identified by immunoprecipitation with either αPhoE or αSkp. The cross-link product of PhoE with Skp is indicated with an arrow.

**Membrane—**Recently, it was proposed that Skp is required for the release of OmpA protein from spheroplasts. Furthermore, Skp was proposed to be involved in the acquirement of a soluble periplasmic form of newly translocated OMPs (15). From the data presented in Fig. 1 (lanes 1 and 15) we observed that PhoE remained associated to the membrane to a large extent. Addition of a periplasmic extract isolated from a Skp-overproducing strain (Fig. 1, lanes 4 and 18) did not significantly influence the amount of released PhoE. To investigate the release of PhoE from the spheroplasts we carried out release experiments and found that 55% of PhoE remained associated to the membrane in a wild type strain. It has been reported that the amount of Skp in spheroplasts is lower in the presence of bivalent cations (14). When we leave out magnesium ions from our spheroplast preparations, the amount of PhoE that remained associated to the membranes was slightly increased (63%). In these spheroplast preparations the release of OmpA into the supernatant was 90%. On the contrary, in the strain lacking Skp protein, PhoE and OmpA were both for a large part detected in the pellet fraction (65 and 61%, respectively). The data indicate that Skp is required for the release and formation of a soluble form of OmpA protein, whereas the release of PhoE seems to be independent of Skp.

**DISCUSSION**

Proteins destined for the extracytoplasmic environment pass the IM in a non-native conformation. Upon entering the periplasm, these proteins have to be protected immediately against degradation, aggregation, and misfolding. To that end, periplasmic chaperones are supposed to be involved in directing these proteins into a productive folding and/or assembly pathway. Little is known which periplasmic chaperones and folding catalysts are involved in the late stages of IM translocation and the early stages of folding. We used an unbiased approach to study the early interactions in the periplasm of accessory proteins with the OMP PhoE, and we found an early interaction with the periplasmic chaperone Skp. It has been suggested that Skp plays an important role in these early events as a specific chaperone for OMPs (15, 16), but the timing and the location of this interaction was not clear. The data presented in this paper showed that the initial interaction of PhoE with Skp occurs at the periplasmic side of the IM. A hybrid protein consisting of 300 N-terminal amino acids of PhoE protein fused to mouse DHFR formed a transmembrane translocation intermediate. This hybrid protein interacted with Skp as monitored by protein cross-linking, indicating that PhoE interacts with the chaperone already during translocation. In addition, the data implicate the presence of one or more Skp-binding sites in the N-terminal region of PhoE. Indeed, we found that Skp was able to closely interact with an N-terminal PhoE fragment of 65 amino acid residues in *in vitro* cross-linking experiments. Furthermore, deletions in the N-terminal as well as the C-terminal part of the PhoE protein resulted in reduction of precipitation of PhoE with αSkp. The data can be explained by assuming the presence of two Skp-binding regions in the N terminus of PhoE. Absence of certain parts of the C terminus in PhoE mutant proteins might influence the conformation of this protein, and, as a consequence, the exposure of binding sites is different. The immunoprecipitation experiments were performed in the absence of membranes, which might influence the manner and extend to which Skp interacts with PhoE protein. However, it should be emphasized that detergents are present during immunoprecipitations, which can be regarded as a membrane-mimicking environment. Taken together, the results demonstrated that Skp interacts with PhoE protein once the extreme N-terminal part, containing these binding sites, appears in the periplasm. We therefore suggested that early translocation intermediates of OMPs are already recognized by this molecular chaperone. It is unlikely that the specific interaction of Skp with OMPs is due to recog-
FIG. 7. Immunoprecipitations of in vitro formed complexes between Skp and wild type or mutant PhoE proteins with Skp antiserum. Wild type and mutant PhoE proteins were synthesized as described under “Experimental Procedures.” Part of translations mixtures (TL) were analyzed directly (5 μl) demonstrating the position of all translation products (indicated by a small arrow) and chloramphenicol acetyltransferase (Cat). The other part (45 μl) was analyzed after immunoprecipitations with Skp antiserum (α-Skp). Lanes: 1, mPhoE; 2, Δ3–11; 3, Δ3–15; 4, Δ3–42; 5, Δ8–91; 6, Δ110–174; 7, Δ249–298; 8, Δ56–93; 9, Δ204–330; 10, Δ56–93; 11, Δ48–146.

Table 1

Relative efficiency of immunoprecipitations of PhoE proteins with Skp

| Protein         | Relative efficiency |
|-----------------|---------------------|
| WT (mPhoE)     | 100                 |
| Δ3–11          | 65                  |
| Δ3–18          | 7                   |
| Δ3–42          | 8                   |
| Δ8–91          | 8                   |
| Δ110–174       | 4                   |
| Δ249–298       | 5                   |
| Δ63–287        | 3                   |
| Δ204–330       | 33                  |
| Δ56–93         | 5                   |
| Δ48–146        | 0.2                 |

nition of a linear amino acid sequence, because the extreme N terminus of PhoE protein does not reveal a conserved sequence. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.

Previously, it was demonstrated that Skp exists in two different states characterized by their different sensitivities to proteases in vivo (16). In addition, Skp was localized as a soluble periplasmic as well as a membrane-associated protein. Both forms have apparently the ability to interact with OMPs. In co-immunoprecipitation and cross-linking experiments to ribosome-nascent chains, the soluble, protease-sensitive form was able to form a soluble complex with OMPs. Here, we demonstrated that full-length PhoE protein, imported into IM vesicles, interacts with Skp that is co-purified with IM vesicles, thereby decreasing the possibility for detection. It is therefore more likely that the basis for the selective interaction of Skp with OMPs is at the level of recognizing certain structural elements, possibly β-strands or β-sheets.
21. Guthrie, B., and Wickner, W. (1990) J. Bacteriol. 172, 5555–5562
22. Den Blaauwen, T., Buddelmeijer, N., Aarsman, M. E. G., Hameete, C. M., and Nanninga, N. (1999) J. Bacteriol. 181, 5167–5175
23. Nouwen, N., Tommassen, J., and De Kruijff, B. (1994) J. Biol. Chem. 269, 16929–16933
24. Van Gelder, P., De Cock, H., and Tommassen, J. (1994) Eur. J. Biochem. 236, 783–787
25. de Cock, H., Hekstra, R. E. W., and Tommassen, J. (1990) Biochemistry 29, 177–182
26. Mayer, M. P. (1995) Gene 163, 41–46
27. Holck, A., and Kleppe, K. (1988) Gene 77, 117–124
28. Valent, Q. A., Scotti, P. A., High, S., de Gier, J.-W. L., von Heijne, G., Lentzen, G., Wintermeyer, W., Oudega, B., and Luirink, J. (1998) EMBO J. 17, 2504–2512
29. High, S., Flint, N., and Dobberstein, B. (1991) J. Cell Biol. 113, 25–34
30. Bosch, D., Leunissen, J., Verhakel, J., de Jong, M., van Erp, H., and Tommassen, J. (1986) J. Mol. Biol. 189, 449–455
31. Römsch, K., Webb, J., Linglebach, K., Gausepohl, H., and Dobberstein, B. (1990) J. Cell Biol. 111, 1793–1802
32. Bonnefoy, N., Chalvet, F., Hamel, P., Slonimski, P., and Dujardin, G. (1994) J. Mol. Biol. 239, 201–212
33. Houben, E. N. G., Scotti, P. A., Valent, Q. A., Brunner, J., de Gier, J.-W., Oudega, B., and Luirink, J. (2000) FEBS Lett. 238, 1–5
34. Oliver, D. B., and Beckwith, J. (1981) Cell 25, 765–772
35. Arkowitz, R. A., Joly, J. C., and Wickner, W. (1993) EMBO J. 12, 243–253
36. Joly, J. C., and Wickner, W. (1993) EMBO J. 12, 255–263
37. Bocquet-Pages, C., Lazdunski, C., and Lazdunski, A. (1981) Eur. J. Biochem. 118, 105–111
38. Ried, G., Hindennach, I., and Henning, U. (1990) J. Bacteriol. 172, 6048–6053
39. Pages, C., Lazdunski, C., and Lazdunski, A. (1982) Eur. J. Biochem. 122, 381–386
40. Bolla, J.-M., Lazdunski, C., and Pages, J.-M. (1988) EMBO J. 7, 3595–3599