Research Article

Study of the Antimicrobial Activity of the Chinese Dong Ethnic Minority Medicine, Madeng’ai

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The overuse of antibiotics has contributed to the emergence of multidrug-resistant bacteria, which poses a challenging task for clinical therapy. Thus, new agents with antibiotic efficacy against multidrug-resistant infections are needed. The traditional Dong ethnic minority medicines have emerged as a new source for prodrug selection. Among them, Madeng’ai (Potentilla-freyniana Bornm) is widely used by the folk for anti-infection and wound healing, although the mechanisms remain unclear. In this study, the antimicrobial activities of Dong medicine Madeng’ai were evaluated both in vitro and in vivo. S. aureus, E. coli, E. facetalis, P. aeruginosa, K. pneumoniae, and A. baumannii were cultured in LB media, different concentrations of Madeng’ai powder solution were added to the LB agar plates to evaluate minimal inhibitory concentration. An animal study was performed on a mouse excisional wound model combined with bacterial solution injection in the wound area. After Madeng’ai or PBS treatment, hematoxylin and eosin analysis were used for pathological analysis of skin tissues from the infected area. Madeng’ai powder solution over 2 mg/mL concentration completely inhibited E. coli growth. At 4.0 mg/mL, Madeng’ai significantly inhibited the growth of E. facetalis, Pseudomonas aeruginosa (PAE), Klebsiella pneumoniae, and Acinetobacter baumannii. The mouse model revealed that Madeng’ai could suppress the growth of MRSA and PAE and accelerate healing of cutaneous wounds. Madeng’ai, a newly discovered Dong ethnic minority medicine possesses considerable antimicrobial activity against both human normal pathogenic bacteria and multiresistance bacteria such as Pseudomonas aeruginosa, S. aureus, and Acinetobacter baumannii. Therefore, Madeng’ai has great potential for further study and clinical application.

1. Introduction

Although a wide range of antibiotics have been developed, their overuse and misuse has led to the emergence of antimicrobial resistance (AMR), which is a threat to international public health [1, 2]. It is estimated that in Norway, Iceland, and the European Union, 25,000 AMR-associated deaths have occurred annually since 2009, while in the USA, this number is estimated to be 99,000 [3, 4]. The threat of AMR in the USA is well documented in the CDC’s report titled “Antibiotics Resistance Threats in the United States, 2013.” The UK’s National Risk Assessment (NRA) reported that the number of AMR infections would continue to increase significantly. In China, the National Health and Family Planning Commission reported the “National Action Plan for Containing Antibacterial Resistance (2016–2020)” in 2016 [5].

AMR increases the risk of uncontrolled infection, sepsis, septic shock, microbial toxic syndrome, and even mortality. It is estimated that globally, there were 48.9 million cases of
sepsis and 11 million sepsis deaths in 2017 [6]. In Japan, a prospective nationwide cohort study of sepsis carried out from 2016-2017 found that *E. coli* was the most common cause of sepsis, *S. aureus* was the most common Gram-positive bacteria, methicillin-resistant *S. aureus* (MRSA) was the most fatal bacteria, and that Gram-positive bacteria were slightly more common in sepsis than Gram-negative bacteria [7]. Goldstein et al. reported that the rates of sepsis-associated hospitalization and mortality have increased markedly in the last 2 decades. *E. coli* is the main cause of septicemia-associated hospitalization in patients aged >50 years and the main cause of mortality in adults (18–84 years old) [8]. A Chinese study by Chen et al. found that Gram-positive bacteria are as prevalent as Gram-negative bacteria in adults with sepsis and that in neonates, sepsis from Gram-positive bacteria was more common than sepsis from Gram-negative bacteria [9]. Thus, it is crucial to monitor the development of antimicrobial resistance. Novel antimicrobial agents with efficacy against AMR strains are urgently needed [2, 10].

Natural products have served as a major source of antibacterial agents. For instance, honey has been used for millennia as a topical treatment for wounds and burns. Manuka honey is reported to inhibit a wide range of microorganisms [10]. *Artemisia argyi* (Chinese mugwort) has been used as a traditional Chinese herbal medicine for millennia. Xiao et al. reported that *Artemisia argyi* Lev. et Vant essential oil extracted through simultaneous distillation has high antimicrobial activity against *L. monocytogenes*, *E. coli*, *P. Vulgaris*, *S. enteritidis*, and *A. Niger* [11]. It is reported that leaf extracts from *Potentilla fruticosa*, *Potentilla glabra*, and *Potentilla parvifolia*, traditional Chinese herbs of the *Rosaceae* family, possess significant microbial inhibition effects [12]. *Potentilla fruticosa* was the most potent inhibitor of Gram-positive bacteria and *P. aeruginosa*, while *Potentilla parvifolia* exhibited broad-spectrum antifungal and antibacterial effects [12]. These reports indicate that some herbs and natural products have great potential for antimicrobial use. *Potentilla freyniana Bornm*, a member of the genus *Potentilla* and family *Rosaceae*, is widely distributed in southern China provinces, including in Hunan, Guizhou, and Guangxi. Madeng’ai (Figure 1) is a variant of *Potentilla freyniana Bornm*, and has long been medicinally used by the Chinese Dong ethnic minority for the treatment of infection, wound healing, and empyrosis. This phenomenon inspires us that Madeng’ai may have beneficial effects on human pathogenic bacteria infection, although no relevant study has been published. Due to the widespread drug resistance, new chemicals or herbal medicines are urgently needed for the treatment of antimicrobial resistant (AMR) infections. For instance, methylene blue has now been proven to be highly effective in inhibiting the growth of *Mycobacterium tuberculosis* [13]. In this study, using *in vitro* and *in vivo* studies, we showed that Madeng’aihas antimicrobial activity on human normal pathogenic and multiresistant bacteria, and this Dong medicine has great potential for further study and application.

2. Materials and Methods

2.1. Plant Materials. Madeng’ai samples were collected from a grassland 800 meters above sea level, in Tongdao County, Huaihua City, Hunan Province, and identified by the Hunan Provincial Key Laboratory of Dong Medicine, Hunan University of Medicine according to its growing area, botany morphology of leaf, root, stem, and flower color, and other comprehensive factors. The plant materials were powdered and stored at 4°C until use.

2.2. Microorganisms. *Staphylococcus aureus* (*S. aureus*, ATCC25923), *Escherichia coli* (*E. coli*, ATCC259220), *Enterococcus faecalis* (*E. faecalis*, ATCC29212), *Pseudomonas aeruginosa* (*P. aeruginosa*, ATCC2785), *Klebsiella pneumoniae* (*K. pneumoniae*) and *Acinetobacter baumannii* (*A. baumannii*) were obtained from the First Affiliated Hospital of Chongqing Medical University, China. The latter two bacteria were isolated from patients’ sputum and belong to nonmultiple drug-resistant bacteria based on drug susceptibility tests (using the minimum inhibitory concentration method).

2.3. Minimum Inhibitory Concentration (MIC) Evaluation. MIC refers to the lowest concentration of extracts capable of inhibiting visible bacterial growth. According our preliminary study, Madeng’ai powder (0, 100, 200, 300, 400, and 500 mg) was added into flasks containing 100 mL of LB broth media and autoclaved to generate 0, 1, 2, 3, 4, and 5 mg/mL Madeng’ai extract suspensions. Next, 100 μL aliquots of MRSA or E.coli cultures with a density of 5 x 10^5 CFU/mL (0.5 McFarland method) were added to the 100 μL LB broth containing different Madeng’ai extracts and cultured overnight, 37°C, with shaking at 220 rpm for 2-3 hours until an OD600 (Optical Density) of 0.5 was reached in low concentration of Madeng’ai extract groups. Finally, 100 μL aliquots from the 6 cultures of MRSA or E.coli were spread on...
normal LB agar plates and cultured overnight. The inhibitory role of Madeng’ai extracts on bacteria was judged by the naked eye, ignoring single colony within the area of the inoculated spot.

2.4. Antibacterial Activity Evaluation in Animal Experiments. Mouse experiments adhered to the animal use guidelines of the National Institutes of Health and were approved by the Ethics Committee for Animal Experiments of Chongqing Medical University. All mice were housed in laminar flow cabinets under specific pathogen-free conditions at room temperature with a 12 h light/dark cycle and with ad libitum access to food and water. Twelve female BABL/c mice (5–6 weeks old) were randomly divided into the MRSA-medicine, MRSA-PBS, PAE-medicine, and PAE-PBS groups. Next, the backs of the mice were shaved and a 0.5 cm diameter wound was made using a sterile surgical scissor. Methicillin-resistant Staphylococcus aureus and P. aeruginosa were cultured in LB broth at 37°C with shaking at 220rpm for 2–3 hours to reach an OD600 nm value of 0.5. Microorganisms were counted using the plate counting method and adjusted to a density equivalent to 10^6 CFU/mL. A 100 μL suspension of MRSA or PAE was then subcutaneously injected into the wound area within 30 min to avoid changes in inoculum density. For animal studies, powdered Madeng’ai (10 g) root was added to 50 mL of autoclaved Milli-Q water and autoclaved to generate a pasty suspension (concentration: 0.2g/mL). After 48 h, the Madeng’ai extract was applied twice daily on the wound surfaces of the MRSA/PAE-medicine group using a cotton swab. PBS was similarly applied to the wounds of the MRSA/PAE-PBS group.

2.5. Wound Recovery and Changes in Mice Weight. The weight of the mice and the rate of recovery were monitored daily for 10 days after wound treatment with the Madeng’ai extracts and PBS control.

2.6. Rate of Bacteria Growth and Histological Analysis after Treatment. After wound treatment with Madeng’ai or PBS for 10 days, wound regions were disinfected with 75% ethanol. The wound surfaces were then cut down with sterile surgical scissors and swabbed on LB agar plates for bacteria culture. Wound surfaces were then sectioned at 4 μm and stained with H&E for histological analysis.

2.7. Statistical Analysis. Data are presented as the mean ± standard deviation (mean ± SD), and analyzed by SPSS software for Windows (version 23.0). A comparison of means between groups were performed by the Student’s t-test. Differences among the experimental groups were analyzed by one-way analyses of variance followed by LSD or Dunnett’s T post hoc test. p < 0.05 was considered significant.

3. Results

3.1. Minimum Inhibitor Concentration (MIC). The antimicrobial activities (MIC values) of Medeng’ai against common Gram-positive and Gram-negative bacteria are shown on Figure 2. For S. aureus, Madeng’ai exhibited macroscopic inhibitory activity at 2 mg/mL as almost no macroscopic S. aureus colonies were observed at this concentration. At 1 mg/mL, Madeng’ai suppressed the growth of S. aureus LB agar plates as fewer S. aureus colonies were observed relative to untreated cultures. Madeng’ai extracts also inhibited the growth of E. coli on LB agar plates with the strongest inhibition observed at 4 mg/mL with no observable macroscopic E. coli colonies. Although 1 mg/mL of the extract suppressed E. coli, 2 and 3 mg/mL almost completely inhibited E. coli growth on LB agar plates when compared to untreated controls.

To determine its antimicrobial spectrum, its inhibitory effect was examined against various bacteria on an LB agar plate (Figure 3). This analysis showed that 4.0 mg/mL Madeng’ai had macroscopic inhibitory activity against E. faecalis (Gram-positive) and P. aeruginosa (Gram-negative) as well as against K. pneumoniae and A. baumannii (both Gram-negative), isolated from patients’ sputum.

3.2. In vivo Antimicrobial Activity. The wound healing rates in the MRSA-medicine and MRSA-PBS groups are shown in Figure 4. On Day 0 (before treatment), infected wound areas were obviously visible in both the MRSA-medicine group and the MRSA-PBS group. However, on Day 6 (after treatment), the wound areas in the MRSA-medicine group were markedly smaller when compared to Day 2. Moreover, wound margins were more distinct and regular on Day 6 when compared to Day 2 and wound secretions were reduced relative to Day 2. In contrast, on Day 6, mice in the MRSA-PBS group (MRSA untreated) had larger infected wound areas than on Day 2. Moreover, wound margins were less well-defined on Day 6 when compared to Day 2 and wound secretions were increased relative to Day 2. On Day 10, the wounds on MRSA-medicine mice were markedly healed and had no redness, swelling, or macroscopic wound area when compared to MRSA-PBS mice, whose wounds were still infected, open, red in color, and swollen.

The wound healing status of the PAE-medicine and PAE-PBS groups are shown on Figure 5(a). The results revealed that the infected wound recovered better in the PAE-medicine group when compared to the PAE-PBS group. On Day 0 (before treatment), infected wound areas were obviously visible in both the PAE-medicine group and PAE-PBS groups. However, on treatment Day 6, the infected wound area in the PAE-medicine group was smaller than on Day 2. Moreover, the infected wound margin was more defined and regular, and the wound secretions were reduced when compared to Day 2. On Day 10, the wounds on PAE-medicine mice had clearly healed and did not have redness, swelling, or macroscopic wound area when compared to PAE-PBS mice whose wounds were still infected, red, swollen, and open.
3.3. Bacterial Growth Condition and Histological Analysis after Treatment. The growth rates of infected bacteria from wound secretions after MRSA and PAE treatment surfaces (recovered or unrecovered) are shown in Figures 4(b) and 5(b), respectively. Very few bacterial colonies were observed upon culturing bacteria from MRSA-infected wounds treated with Madeng’ai extracts on LB agar plate. However, numerous MRSA colonies were observed upon culturing bacteria from the wounds of PBS-treated mice. Similar observations were made upon culturing bacteria from PAE-infected wounds when compared to bacteria cultured from the wounds of PBS-treated mice.

H&E analyses of wound surfaces (recovered or unrecovered) after treatment revealed vast numbers of inflammatory cells (bluish-violet cells marked with red circles) were observed on the upper pictures of both (Figures 4(c) and 5(c)), indicating that MRSA/PAE wound infection persisted after PBS treatment. However, the number of inflammatory cells was markedly reduced upon treatment of MRSA- and PAE-infected wounds with Madeng’ai extract.

Figure 2: (a) MIC results of Madeng’ai against S. aureus and E. coli. (b) The number of colonies on plates of different concentrations of Madeng’ai against S. aureus and E. coli. *** p < 0.01 versus blank control group.

Figure 3: (a) The inhibitory effect of Madeng’ai against E. faecalis, P. aeruginosa (PAE), K. pneumoniae, and A. baumannii. (b) The number of colonies on plates of different concentrations of Madeng’ai against S. aureus and E. coli. *** p < 0.01 versus blank control group.
Moreover, the epidermis was more intact in treated animals than in PBS controls, indicating that Madeng’ai promoted the healing of infected wounds (Figures 4(c) and 5(c)). No obvious changes in mouse body weight were noticed during the treatment period (Figures 4(d) and 5(d)).

4. Discussion

In this study, we show that on LB agar plates, 2 mg/mL of Madeng’ai markedly inhibits the growth of *S. aureus* and *E. coli*, respectively. Moreover, our data show that the extract has broad spectrum activity at a MIC of 4 mg/mL against Gram-positive and Gram-negative bacteria. Further analysis revealed that at 4 mg/mL, the extract could also inhibit the growth of other common infectious bacteria, including *E. faecalis*, PAE, *K. pneumoniae*, and *A. baumannii*. It should be noted that the 0, 1, 2, 3, 4, and 5 mg/mL of Madeng’ai extracts we used in this study is the concentration of crude herb powder rather than herb extracts or pure compounds. Since the Madeng’ai powder or boiled soup was traditionally used by the Dong and Miao ethnic minorities of China as analgesic and antibacterial agents, we believe the active ingredients could be extracted during the autoclaving process (20 mins). Although some active compounds may be inactive during this process, and the extract efficiency may not be high due to the short

**Figure 4**: The effects of Madeng’ai on bacterial growth and histological and body change of methicillin-resistant *S. aureus*-infected mice. (a) Recovery of methicillin-resistant *S. aureus*-infected wounds on BALB/c mice after Madeng’ai or PBS treatment. (b) LB agar of methicillin-resistant *S. aureus* growth from infected wound secretions after treatment with Madeng’ai or PBS. (c) H&E analysis of skin tissues from wounds infected with methicillin-resistant *S. aureus* after treatment with Madeng’ai or PBS. (d) Changes in mouse body weight during treatment.
extraction time, the data reported in this work is promising. The antibacterial activities of water and alcohol extracts of Madeng’ai under various conditions and the active compounds responsible for the antibacterial activities should be investigated in further study. Crucially, mouse experiments revealed that the Madeng’ai extract has strong antimicrobial activity against methicillin-resistant S. aureus and PAE and promoted wound healing in mice compared to untreated controls. Moreover, bacterial culture results revealed that negligible levels of MRSA and PAE existed in the secretions of Madeng’ai treated mice. H&E analysis revealed that treatment with Madeng’ai reduced inflammatory infiltration and promoted skin regeneration in the wound area.

Large numbers of drugs with antimicrobial activity have been discovered and deployed clinically. Anne H. Norris and colleagues recommended that the choice of antimicrobial agents be based more on the outpatient parenteral antimicrobial therapy model than on the drug’s pharmacokinetics [14]. Antibiotics like vancomycin, cefazolin, or aminoglycosides may be limited to patients who receive parenteral antimicrobials during dialysis sessions [15]. Li et al. showed that decisions on antimicrobial prophylaxis should be based on contamination in the surgical field. For instance, antibiotics should be used to prevent postoperative surgical site infection, which is mainly caused by S. aureus. Besides, before colon and rectal surgery, antibiotics were also applied to combat the infections may be caused by E. coli.
Empirical therapy for bacterial infections is widely accepted because it is effective. For instance, vancomycin and antistaphylococcal penicillin are used empirically to inhibit *S. aureus* bacteremia and vancomycin is the first choice treatment for MRSA [17, 18]. Fosfomycin-trometamol and nitrofurantoin have been recommended as first-line antibiotics for uncomplicated cystitis. Fluoroquinolones are first-line oral treatments for uncomplicated pyelonephritis as well as for *E. coli*, the leading cause of uncomplicated urinary tract infection [19–21].

However, the development of antibiotics has been accompanied by antimicrobial resistance, and mounting effort has gone into understanding the mechanisms of antimicrobial resistance. Antimicrobial resistance may happen when the target cell’s wall is poorly permeable or permeability is specifically reduced for a given antibiotic [22]. Resistance may also result from loss of chemical affinity between an antimicrobial agent and its target, as happens in streptococci [23]. Additionally, antimicrobial activity may be suppressed by the bacterium, as happens in β-lactamase production [24]. Other mechanisms of antimicrobial resistance include the extrusion of antimicrobial agents from bacterial cells by efflux systems [25] and the production of extracellular polymer substances in biofilms that may impair the diffusion of antimicrobial agents through electrostatic and steric interactions [26]. Thus, novel, effective antimicrobials are urgently needed.

Although we have not determined the antimicrobial mechanism of Madeng’ai, numerous studies have indicated the antimicrobial role of *Potentilla* species attributed to their phenolic compounds [12, 27]. Thus, future studies should investigate the underlying mechanisms of the antimicrobial activity of Madeng’ai. Furthermore, because of their anti-inflammatory and vasoconstrictive effects, *Potentilla* species are used to treat topical inflammatory skin disorders [28–30] and diabetes mellitus [31, 32]. Therefore, the therapeutic potential of Madeng’ai against microbial infections and other diseases warrants further investigation.

5. Conclusion

Taken together, Dong ethnic minority medicine Madeng’ai possesses antimicrobial activity against both human normal pathogenic bacteria but also multiresistance bacteria such as *Pseudomonas aeruginosa*, *S. aureus*, and *Acinetobacter baumannii*. Therefore, Madeng’ai has great potential for further study and application.

**Abbreviations**

| Abbreviation | Description                                      |
|--------------|--------------------------------------------------|
| AMR          | Antimicrobial resistance                         |
| MIC          | Minimum inhibitory concentration                 |
| MRSA         | Methicillin-resistant *Staphylococcus aureus*     |
| *E. coli*    | *Escherichia coli*                               |
| PAE          | *Pseudomonas aeruginosa*                         |
| *K. pneumoniae* | *Klebsiella pneumoniae*                        |
| *A. baumannii* | *Acinetobacter baumannii*                      |
| *E. faecalis* | *Enterococcus faecalis*                         |
| *S. aureus*  | *Staphylococcus aureus*                         |

**Data Availability**

All data used to support the findings of this study are included within the article.

**Ethical Approval**

All animal experiments were performed in line with the guidelines for animal use of the National Institutes of Health. All procedures for mice experiments were approved by the Ethics Committee of Animal Experiments of Chongqing Medical University.

**Conflicts of Interest**

The authors declare that there are no conflicts of interest.

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**References**

[1] S. A. McEwen and P. J. Collignon, “Antimicrobial resistance: a one health perspective,” *Microbiology Spectrum*, vol. 6, no. 2, pp. 6–2, 2018.

[2] L. Brinkac, A. Voorhies, A. Gomez, and K. E. Nelson, “The threat of antimicrobial resistance on the human microbiome,” *Microbial Ecology*, vol. 74, no. 4, pp. 1001–1008, 2017.

[3] M. Ferri, E. Ranucci, P. Romagnoli, and V. Giaccone, “Antimicrobial resistance: a global emerging threat to public health systems,” *Critical Reviews in Food Science and Nutrition*, vol. 57, no. 13, pp. 2857–2876, 2017.

[4] B. Spellberg, M. Blaser, R. J. Guidos, and D. N. Gilbert, “Policy Paper. Combating antimicrobial resistance: policy recommendations to save lives,” *Clinical Infectious Diseases An Official Publication of the Infectious Diseases Society of America*, vol. 52, no. Suppl 5, p. S397, 2011.

[5] Y. Xiao and L. Li, “A national action plan to contain antimicrobial resistance in China: contents, actions and expectations,” *AMR Control*, vol. 15, no. 3, pp. 17–20, 2017.

[6] K. E. Rudd, S. C. Johnson, K. M. Agesa et al., “Global, regional, and national sepsis incidence and mortality, 1990–2017: analysis for the Global Burden of Disease Study,” *The Lancet*, vol. 395, no. 10219, pp. 200–211, 2020.

[7] Y. Umemura, H. Ogura, K. Takuma et al., “Current spectrum of causative pathogens in sepsis: a prospective nationwide cohort study in Japan,” *International Journal of Infectious Diseases*, vol. 103, pp. 343–351, 2021.

[8] E. Goldstein, D. R. MacFadden, Z. Karaca, C. A. Steiner, C. Viboud, and M. Lipsitch, “Antimicrobial resistance prevalence, rates of hospitalization with septicemia and rates of mortality with sepsis in adults in different US states,” *International Journal of Antimicrobial Agents*, vol. 54, no. 1, pp. 23–34, 2019.

[9] X. C. Chen, Y. F. Yang, R. Wang, H. F. Gou, and X. Z. Chen, “Epidemiology and microbiology of sepsis in mainland China
in the first decade of the 21st century,” *International Journal of Infectious Diseases*, vol. 31, pp. 9–14, 2015.

[10] V. C. Nolan, J. Harrison, J. E. E. Wright, and J. A. G. Cox, “Clinical significance of manuka and medical-grade honey for antibiotic-resistant infections: a systematic review,” *Antibiotics*, vol. 9, no. 11, p. 766, 2020.

[11] X. Guan, D. Ge, S. Li, K. Huang, J. Liu, and F. Li, “Chemical composition and antimicrobial activities of Artemisia argyi L. essential oils extracted by simultaneous distillation-extraction, subcritical extraction and hydro-distillation,” *Molecules*, vol. 24, no. 3, p. 483, 2019.

[12] S. S. Wang, D. M. Wang, W. J. Pu, and D. W. Li, “Phytochemical profiles, antioxidant and antimicrobial activities of three Potentilla species,” *BMC Complementary and Alternative Medicine*, vol. 13, no. 1, pp. 321–411, 2013.

[13] D. Gazel and Ö. Zanapaloçu Gazel, “In vitro activity of methylene blue on *Mycobacterium tuberculosis* complex isolates,” *Tuberk Toraks*, vol. 69, no. 2, pp. 279–284, 2021.

[14] A. H. Norris, N. K. Shrestha, G. M. Allison et al., “2018 Infectious diseases society of America clinical practice guideline for the management of outpatient parenteral antimicrobial therapy,” *Clinical Infectious Diseases*, vol. 68, no. 1, pp. e1–e35, 2019.

[15] C. B. Cunha and E. M. D’Agata, “Implementing an antimicrobial stewardship program in out-patient dialysis units,” *Current Opinion in Nephrology and Hypertension*, vol. 25, no. 6, pp. 551–555, 2016.

[16] S. Li, Z. J. Han, J. He et al., “Society for Translational Medicine expert consensus on the use of antibiotic drugs in thoracic surgery,” *Journal of Thoracic Disease*, vol. 10, no. 11, pp. 6536–6574, 2018.

[17] J. S. Davis, S. van Hal, and S. Tong, “Combination antibiotic treatment of serious methicillin-resistant *Staphylococcus aureus* infections,” *Seminars in Respiratory and Critical Care Medicine*, vol. 36, no. 01, pp. 003–016, 2017.

[18] R. Diaz, V. Afreixo, E. Ramalheira, C. Rodrigues, and B. Gago, “Evaluation of vancomycin MIC creep in methicillin-resistant *Staphylococcus aureus* infections—a systematic review and meta-analysis,” *Clinical Microbiology and Infections*, vol. 24, no. 2, pp. 97–104, 2018.

[19] F. M. E. Wagenlehner, G. Schmiemann, U. Hoyme et al., “National S3 guideline on uncomplicated urinary tract infection: recommendations for treatment and management of uncomplicated community-acquired bacterial urinary tract infections in adult patients,” *Urologe A*, vol. 50, no. 2, pp. 153–169, 2011.

[20] C. C. Butler, K. O’Brien, M. Wootton et al., “Empiric antibiotic treatment for urinary tract infection in preschool children: susceptibilities of urine sample isolates,” *Family Practice*, vol. 33, no. 2, pp. 127–132, 2016.

[21] B. Regasa Dadi, T. Abebe, L. Zhang, A. Mihret, W. Abebe, and W. Amogne, “Drug resistance and plasmid profile of uropathogenic *Escherichia coli* among urinary tract infection patients in Addis Ababa,” *The Journal of Infection in Developing Countries*, vol. 12, no. 08, pp. 608–615, 2018.

[22] S. Karakonstantis and I. Saridakis, “Colistin heteroresistance in Acinetobacter spp.: systematic review and meta-analysis of the prevalence and discussion of the mechanisms and potential therapeutic implications,” *International Journal of Antimicrobial Agents*, vol. 56, no. 2, Article ID 106065, 2020.

[23] J. M. Sierra, E. Fusté, F. Rabanal, T. Vinuesa, and M. Viñas, “An overview of antimicrobial peptides and the latest advances in their development,” *Expert Opinion on Biological Therapy*, vol. 17, no. 6, pp. 663–676, 2017.

[24] J. Y. Madec, M. Haenni, P. Nordmann, and L. Poirel, “Extended-spectrum β-lactamase/AmpC-and carbapenemase-producing Enterobacteriaceae in animals: a threat for humans?” *Clinical Microbiology and Infections*, vol. 23, no. 11, pp. 826–833, 2017.

[25] R. Kabra, N. Chauhan, A. Kumar, P. Ingale, and S. Singh, “Efflux pumps and antimicrobial resistance: paradoxical components in systems genomics,” *Progress in Biophysics and Molecular Biology*, vol. 141, pp. 15–24, 2019.

[26] R. A. Harper, G. H. Carpenter, G. B. Proctor et al., “Diminishing biofilm resistance to antimicrobial nanomaterials through electrolyte screening of electrostatic interactions,” *Colloids and Surfaces B: Biointerfaces*, vol. 173, pp. 392–399, 2019.

[27] D. Augustynowicz, K. P. Latté, and M. Tomczyk, “Recent phytochemical and pharmacological advances in the genus Potentilla L. sensu lato—An update covering the period from 2009 to 2020,” *Journal of Ethnopharmacology*, vol. 266, Article ID 113412, 2021.

[28] U. Wöllle, J. Hoffmann, B. Haarhaus, V. Rao Mittapalli, and C. M. Schenpp, “Anti-inflammatory and vasoconstrictive properties of *Potentilla erecta*—A traditional medicinal plant from the northern hemisphere,” *Journal of Ethnopharmacology*, vol. 204, pp. 86–94, 2017.

[29] J. Hoffmann, F. Casetti, U. Bullerkotte et al., “Anti-inflammatory effects of agrimoniin-enriched fractions of *Potentilla erecta*,” *Molecules*, vol. 21, no. 6, p. 792, 2016.

[30] M. T. Tomovic, S. M. Cupara, M. T. Popovic-Milenkovic, B. T. Ljujic, M. J. Kostic, and S. M. Jankovic, “Antioxidant and anti-inflammatory activity of *Potentilla reptans* L.,” *Acta Polonica Pharmaceutica*, vol. 72, no. 1, pp. 137–145, 2015.

[31] L. Han, T. Li, M. Du, R. Chang, B. Zhan, and X. Mao, “Beneficial effects of *potentilla discolor* bunge water extract on inflammatory cytokines release and gut microbiota in high-fat diet and streptozotocin-induced type 2 diabetic mice,” *Nutrients*, vol. 11, no. 3, p. 670, 2019.

[32] N. Wang, F. Zhu, M. Shen et al., “Network pharmacology-based analysis on bioactive anti-diabetic compounds in *Potentilla discolor* bunge,” *Journal of Ethnopharmacology*, vol. 241, Article ID 111905, 2019.

[33] Z. Tang, Y. Zhao, Z. Zhang et al., *Antimicrobial Activity Study of Chinese Medicine Madeng’ai*, Research Square, Durham, North Carolina, USA, 2021.