Cripto-1 vaccination elicits protective immunity against metastatic melanoma

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ABSTRACT
Metastatic melanoma is a fatal disease that responds poorly to classical treatments but can be targeted by T cell-based immunotherapy. Cancer vaccines have the potential to generate long-lasting cytotoxic CD8+ T cell responses able to eradicate established and disseminated tumors. Vaccination against antigens expressed by tumor cells with enhanced metastatic potential represents a highly attractive strategy to efficiently target deadly metastatic disease. Cripto-1 is frequently over-expressed in human carcinomas and melanomas, but is expressed only at low levels on normal differentiated tissues. Cripto-1 is particularly upregulated in cancer-initiating cells and is involved in cellular processes such as cell migration, invasion and epithelial–mesenchymal transition, which are hallmarks of aggressive cancer cells able to initiate metastatic disease. Here, we explored the potential of Cripto-1 vaccination to target metastatic melanoma in a preclinical model. Cripto-1 was overexpressed in highly metastatic B16F10 cells as compared to poorly metastatic B16F1 cells. Moreover, B16F10 cells grown in sphere conditions to enrich for cancer stem cells (CSC) progressively upregulated cripto-1 expression. Vaccination of C57Bl/6 mice with a DNA vaccine encoding mouse Cripto-1 elicited a readily detectable/strong cytotoxic CD8+ T cell response specific for a H-2 Kb-restricted epitope identified based on its ability to bind H-2Kb molecules. Remarkably, Cripto-1 vaccination elicited a protective response against lung metastasis and subcutaneous challenges with highly metastatic B16F10 melanoma cells. Our data indicate that vaccination against Cripto-1 represents a novel strategy to be tested in the clinic.

Introduction
The largest hurdle in the treatment of melanoma patients is to ensure that the disease does not disseminate in the patient, leading to metastasis and eventually death. T cell-based immunotherapy has emerged at the forefront for treating metastatic disease and success has been achieved with the treatment of metastatic melanoma and other tumor types using immune-checkpoint blockade and autologous antitumor T cells. The former releases the spontaneous antitumor T cell immunity through removal of inhibitory signaling mediated by the PD1 or CTLA4 molecules. In the latter case, adoptive transfer of ex vivo expanded and activated tumor infiltrating T cells has successfully cured a proportion of metastatic melanoma patients. The efficacy of these strategies relies entirely on the presence of a pre-established antitumor immunity in the patients, which may frequently not be the case.

This limitation can be overcome by educating the immune system through vaccination, which has a long successful history in the prevention of pathogen infections. However, less success has been achieved by vaccines targeting tumor-associated antigens (such as MAGE-A, GP100, NY-ESO-1, Tyrosinase and Her2) to treat cancer, showing limited therapeutic efficacy in clinical trials. Vaccines have been shown to elicit both humoral and cellular responses in a substantial proportion of the patients, though these responses are typically not potent enough to remove a large tumor burden. In consequence, removing bulky tumors through vaccination-induced antigen-specific T cell immunity remains a challenge that may be overcome by targeting a minor subpopulation of melanoma cells with enhanced potential to establish disseminated tumors, such as CSC.

Cripto-1 is a glycoprotein that plays a critical role during embryogenesis and is overexpressed in more than 50% of human carcinomas, as well as melanomas, but is expressed only at low levels on normal differentiated tissues. Cripto-1 is involved in cellular processes such as cell migration, invasion and epithelial–mesenchymal transition, which are hallmarks of metastatic cancer cells. Cripto-1 is particularly upregulated in a small subpopulation of cancer-initiating cells, usually referred to as CSC, which has enhanced potential to metastasize and establish new tumor masses at distant sites in melanoma and other types of cancer. Specific targeting of cancer-initiating cells has shown to efficiently eradicate established and disseminated melanoma lesions and, therefore, it represents a highly attractive strategy to target deadly metastatic disease. Here, we describe that DNA vaccination against the tumor-associated antigen Cripto-1 that elicits specific CD8+ T cell immune responses, leading to decreased tumor burden and reduced metastatic spread in an aggressive syngeneic melanoma model.
Results

Expression of cripto-1 in melanoma cell lines

We first analyzed the presence of Cripto-1 in different murine tumor models. To this end, tumor cell lines from C57BL/6 (B6) background were screened for Cripto-1 expression by western blot. The Balb/c D2F2 cell line and mouse Cripto-1-transfectant D2F2mCR were used as negative and positive controls for Cripto-1 expression, respectively. Significant expression was detected in B16F10 and RetV melanoma cell lines, the latter derived from the metastatic melanoma ret transgenic mouse model. B16F1, a less metastatic B16 melanoma sub-line, and the MCA205 sarcoma cell line had weaker expression of Cripto-1 (Fig. 1A). Then, B16F10 cells were grown in a sphere culture system (Fig. S1) to expand the proportion of cancer-initiating cells with enhanced metastatic potential. Interestingly, Cripto-1 expression was progressively increased after each round of sphere culture (Fig. 1B). To evaluate Cripto-1 expression in healthy tissues we screened the healthy mouse tissue gene expression data set obtained by Su et al. (GSE1133: GLP1073) (Fig. 1C). As expected, within the data set we found that Cripto-1 is expressed during early embryogenesis, and expression is downregulated beyond day 8.5 into adulthood (Fig. S2). These results indicate that Cripto-1 has the potential to be used as a melanoma-associated antigen in a vaccination context.

Generation of specific CD8^+ T cell responses in pmCR immunized mice

To evaluate the potential of Cripto-1 as a melanoma-associated antigen, we first defined the epitopes able to be recognized by CD8^+ T cells. A library consisting of 33 long overlapping peptides (15-mers) covering the full-length mouse Cripto-1 amino acid sequence was screened for the ability to bind H-2 molecules using the RMA-S MHC class I stabilization assay. Three of the peptides tested were able to stabilize H-2 K^b at the cell surface of RMA-S cells (mCR1-15, mCR16-30 and mCR46-60), and as expected, the positive control H2 K^b epitopes, Trp2180-188 and OVA257-264, strongly stabilized H-2 K^b (Fig. 2A). We then identified the presence of MHC class I-restricted epitopes within these 15-mer peptides using an in silico prediction analysis (http://www.cbs.dtu.dk/services/NetMHCpan/) (Fig. 2B). We next evaluated the immunogenicity of the predicted 9-mer epitopes by testing their ability to be processed, presented and recognized by CD8^+ T cells in mice vaccinated with plasmid DNA encoding full-length mouse cripto-1 (pmCR). One of these predicted peptides, mCR16-25 (Fig. 3A left panel), elicited readily detectable CD8^+ T cells able to produce IFN-g and TNF-a after ex vivo peptide stimulation as acquired by flow cytometry in an ex vivo intracellular cytokine staining (Fig. 3A right panel). None or very low responses specific for mCR1-9 and mCR46-55 were detected (Fig. S3A). We further demonstrated that vaccination-induced mCR16-25-specific CD8^+ T cells mediate in vivo cytotoxic killing. Vaccinated mice received spleen cells pulsed with either mCR16-25 or OVA control peptide and stained with high (CFSEhi) or low (CFSElo) concentrations of CFSE, respectively. One day later, specific killing of mCR16-25-pulsed CFSEhi target cells relative to OVA-pulsed CFSElo internal control splenocytes was analyzed by flow cytometry. Killing of mCR16-25 pulsed cells was observed in Cripto-1 vaccinated mice (45% ± 1%) (Fig. 3B). Then, we tested the ability of vaccination-induced mouse Cripto-1-specific CD8^+ T cells to recognize mouse melanoma B16F10 melanoma cells in vitro. CD8^+ T cells isolated from spleen of mouse Cripto-1-vaccinated animals secreted significant amounts of IFN-g in response to stimulation with B16F10 melanoma cells and mCR16-25 peptide but not after control stimulation with...
OVA peptide or left unstimulated (Fig. 3C). These results indicate that cripto-1 is an immunogenic melanoma-associated antigen and that vaccination-induced cripto-1-specific cytotoxic CD8\(^+\) T cells have the potential to recognize and eliminate highly aggressive metastatic melanoma cells.

As a result of these findings, we next investigated the potential of vaccination against mouse Cripto-1 to reduce tumor burden and lung metastasis in the B16F10 melanoma model.

We vaccinated mice twice with DNA plasmids encoding mouse Cripto-1 and 2 weeks later challenged them with B16F10 melanoma cells. Mice were vaccinated twice and 2 weeks later received a s.c. challenge with B16F10 melanoma cells and tumor growth was monitored. A significant delay in tumor growth was observed in pmCR-vaccinated mice, as compared to the control empty vector (pVAX) immunized mice (Fig. 4A). The delayed tumor growth led to significantly extended survival in pmCR-vaccinated mice (Fig. 4B). We then evaluated the ability of pmCR vaccination to protect against metastatic lung colonization of i.v. injected B16F10 cells. A significant decrease in the number of metastatic lung foci (Fig. 4C, D) was observed in mice vaccinated with mouse Cripto-1. These results indicate that vaccines encoding Cripto-1 induce immunity with the potential to target highly metastatic melanoma cells. Altogether, these results led us to conclude that pmCR vaccination efficiently induces mouse Cripto-1-specific CD8\(^+\) T cell responses able to target tumor burden and metastatic spreading of melanoma.

**Discussion**

Our current understanding of the complex nature of tumors has recently established that a small population of cancer cells within the heterogeneous tumor mass is particularly efficient in initiating the formation of disseminated cancerous lesions. These cells, referred to as cancer initiating cells or CSCs, have unlimited self-renewal potential and are resistant to classical therapies.\(^{25}\) The development of therapies selectively targeting this highly aggressive population to prevent metastatic disease therefore is of considerable importance. Cripto-1 has been shown to be expressed on many different tumors,\(^{11}\) including uveal and cutaneous melanomas.\(^{26,27}\) Interestingly, CR is particularly upregulated in CSC populations found in human melanoma\(^{13}\) and contributes to a more tumorigenic phenotype,\(^{28}\) resembling many of the pathways upregulated in metastatic malignant melanoma. Here, we identified that Cripto-1 is expressed in mouse metastatic melanoma models, including highly metastatic B16F10 cell line (Fig. 1A). Moreover, B16F10 cells grown in sphere culture conditions to enrich for CSC progressively upregulated cripto1 expression, indicating a direct correlation between CR expression and aggressiveness. We further demonstrate that CR can be used to target highly metastatic melanoma in the setting of a cancer vaccine. In mice that were vaccinated with DNA plasmids encoding mouse Cripto-1, we found that growth of the highly aggressive B16F10 cell line was significantly inhibited (Fig. 4A, B). Of particular interest, vaccination against mouse Cripto-1 led to a marked reduction in the metastatic spread of B16F10 tumor cells to lungs (Fig. 4C, D).

Vaccination has been a success story in protecting us from a plethora of pathogens, but unfortunately the success seen in this context has not been translated well to therapeutic vaccination against cancer. Cancer vaccines strongly rely on the generation of cytotoxic CD8\(^+\) T lymphocytes (CTLs) to mediate effective antitumor immune response.\(^{29}\) Perhaps it should not be expected that vaccine-induced antitumor CTL responses would be capable of removing bulky tumors, particularly when they consist of a heterogeneous cancer cell population that mediate a strong immnosuppression.\(^{30}\) Instead, vaccines should enable the immune system to reach distant sites of...
disease that evade the capability of traditional therapies. It is therefore essential that anticancer vaccines focus on the elimination of highly metastatic populations, such as CSC. It has been shown that adoptive transfer of CTL redirected against CSC via a CD20-specific chimeric antigen receptor efficiently eliminates melanoma tumors. In this work, elimination of less than 2% of the tumor cells was able to eradicate established melanoma lesions in mice. The same principle was successfully translated into humans, where rituximab (an anti-CD20 monoclonal antibody) produced the regression of chemotherapy-refractory metastatic melanoma.

This study shows the potential of targeting the tumor-associated antigen cripto-1 with a plasmid DNA-based vaccination approach capable of inhibiting aggressive tumor growth and metastatic spread. The key to the potential of DNA vaccines lies in their ability to elicit CTL responses specific for the plasmid-encoded antigens. Herein, we are the first to describe a CTL response to mouse cripto-1 (Fig. 2). Cripto-1, as well as being a GPI anchored extracellular protein, interacts with Notch1 in the ER/golgi apparatus. This lends itself to MHC class I immunoproteasome processing. By identifying strong stabilizing partners for H-2 Kb with the help of the TAP-1 deficient RMA-S cell line we were able to identify potential CTL epitopes and confirm them in silico (Fig 2B). CD8+ T cells from vaccinated mice were able to generate IFNγ and TNF-α in response to one of the predicted epitopes (Fig. 3A). We further validated that mCR16-25 is an epitope that can be processed and presented to CD8+ T cells during vaccination. CD8+ T cells from mouse Cripto-1-vaccinated mice could indeed recognize mCR16-25 and display epitope specific killing in vivo.

In summary, our findings show that targeting cripto-1 using DNA vaccination elicits CD8+ T cell immunity capable of significantly reducing aggressive metastatic melanoma tumor growth. We identify, for the first time, CTL epitopes specific to mouse cripto-1, and have in ongoing work also several human CTL epitopes defined. The possibility of targeting cripto-1 expressed on the small CSC population is attractive, particularly in an adjuvant setting to avoid metastatic spread.

**Material and methods**

**Mice and cell lines**

C57BL/6 mice were bred, maintained at the Microbiology and Tumor Biology Center (Karolinska Institutet, Stockholm, Sweden) and were handled by strict adherence to the European guidelines and University Ethical Committee. Animal studies performed were reviewed and approved by the Regional Animal ethics committees; Stockholms Norra Djurförsöksstina Nämnd Avdelning 2, Sweden with ethical permit number N426/11. RetV (generously donated by Prof. V. Umansky, DKFZ Heidelberg), MCA205, B16F10, B16F1, D2F2 and RMA-s were maintained with GlutaMAX-RPMI supplemented with 10% heat-inactivated FCS, 50 IU/mL penicillin, and 50 μg/mL streptomycin (Life Technologies). Cell lines were maintained at 37°C with 5% CO2 at 95% humidity and were split as was
necessary using 0.05% Trypsine/EDTA (Life Technologies). To generate Cripto-1 overexpressing cell lines, mouse Cripto-1 lentiviral particles were acquired (Amsbio, Abindon, UK) and were used to transduce D2F2 with mouse Cripto-1. Followed by cell sorting for Cripto-1 positive cells using FACS.

**Western blots**

Cell lysates were prepared with 1M RIPA buffer (50nM Tris-HCl, pH 7.4, 1% Triton-X, 0.5% Na-deoxycholate, 0.1% SDS, 150 mM NaCl, 2 mM EDTA, 50 mM NaF) with 1x protease inhibitor (Roche, Cat. No. 04693159001) at 1×10⁶ cells/mL directly after collection from cell culture. Prior to loading on the gel protein concentrations were determined with BCA protein assay (Thermo Scientific, Rockford, IL) according to the manufacturers protocol. Twenty µg protein per sample was loaded on 10% NuPAGE Bis-Tris acrylamide gels (Invitrogen) and run at 200 V for 45 min with MOPS SDS running buffer (Invitrogen) followed by transfer onto PVDF membrane (Immobilon-P; Millipore, Bedford, MA) for 3 h at 40V. Blocking of the membrane was done using TBS-0.5% Tween 20 (Sigma-Aldrich), 2.5% milk powder or 2.5% BSA, followed by wash in TBS-0.5% Tween 20 and incubation with primary antibodies: rabbit α-human Cripto antibody, cross reactive to mouse 1:1,000 (Rockland, cat.no. 600-401-997) and mouse α-b-Actin antibody 1:25,000 (Sigma-Aldrich) overnight at 4°C. Secondary staining was done using α-rabbit IgG, HRP-linked (Cell Signaling Technology) and α-mouse IgG, HRP-linked (Cell Signaling Technology) for 1 h at room temperature. Development was done using Amersham ECL Prime Western Blotting Detection Reagent (GE Healthcare). Using a LAS-1000 CCD camera system (Fujifilm, Tokyo, Japan) luminescence was detected.

**Vaccination and plasmids**

Mice were treated on week 8 and week 10 either by intradermal injection of 40 µg of plasmid injected in PBS followed by electroporation protocol as described previously using IGEA plate electrodes. Cripto-1 encoding plasmids were generously provided by Bianco C et al. and cloned into the pVAX vector (Invitrogen, Carlsbad, CA, USA). Plasmid based vaccines were produced by transformation of E. coli (TOP10, Invitrogen) with pVAX plasmid and grown in Luria-Bertani medium containing Kanamycin (50 µg/mL). To generate endotoxin-free vaccine, plasmids were purified using GigaPrep Endofree Kit (Qiagen GMBH, Hilden, Germany).

**Tumor models**

B16F10 tumor cells were transplanted into the C57BL/6 mice to model melanoma. B16F10 was injected s.c (50,000) or i.v. (200,000) in 100 µL PBS after being harvested when in vitro growth was logarithmic and at 80% confluence. Tumor size was monitored by palpation with calipers and mice were sacrificed when they became moribund or when the tumor reached a volume of 1,000 mm³. Mice that were injected i.v. were sacrificed at day 14 and lungs were excised and B16F10 foci were enumerated.

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**Figure 4.** Vaccination against mouse Cripto-1 increases survival and reduces metastatic lung burden in B6 mice challenged with mouse Cripto-1 expressing B16F10. (A) Tumor growth was compared in B6 mice (n = 10 per group) vaccinated prophylactically with either pmCR or pVAX with two doses of 20 µg plasmid DNA delivered i.d. with electroporation prior to s.c. challenge with 5×10⁶ B16F10 cells with 2-way ANOVA test; **p < 0.01. (B) Survival of B6 mice vaccinated with pmCR or pVAX was compared after challenge with s.c. B16F10 with Mantel–Cox test; **p < 0.01. The results shown here are representative of two experiments. (C) Representative photos of B16F10 metastatic lung burden in vaccinated mice harvested 14 d post i.v. challenge of 2×10⁵ tumor cells. D, reduction in lung metastasis was evaluated by enumerating foci of tumors in lungs of B16F10 i.v. challenged mice and compared with non-parametric t-test; *p < 0.05. The results shown here are representative of three experiments.
Immunological assays

Overlapping 15 amino acid Cripto-1 peptides were generated to cover the whole protein. RMA-s cells were washed with RPMI medium without FCS and kept at room temperature for 2 h. 2 × 10^5 cells were seeded into 96-well plates containing complete medium as well as peptides at a concentration of 100 μg/mL for 6 h. Cells were washed and stained with α-mouse-H2-K^b-FITC (BioLegend, 116505) to detect cell surface MHC class I molecules by flow cytometry. Mouse derived peptides, mCR16-25 (SAFEFGPVA), mCR46-55 (RSFQFVPSV), mCR1-9 (MGYFSSSVVL), Surv20-28 (ATFKNWPVEL), TRP2140-148 (SVYDFVVWVL), OVA257-264 (SIINFEKL) were acquired from China Peptides (ChinaPeptides Co. Ltd. Shanghai, China) at >95% purity. Peptides were used to stimulate mouse lymphocytes in peripheral blood harvested from immunized mice. Cells were seeded into 96-well plates and stimulated with 10 μg/mL of MHC class I-restricted peptide. After 2 h, Golgi-Plug (Becton, Dickson and Company) was added for the last 6 h of stimulation. Cells were stained with α-mouse-CD8^+ α-mouse-IFNγ, α-mouse-TNF-α using the Cytofix/Cytoperm Fixation/Permeabilization Solution (Becton, Dickson and Company) according to manufacturer’s instruction prior to acquisition of cells on LSRII FACS (Becton, Dickson and Company). Data were analyzed using FlowJo (Tree Star). CD8^+ cells were isolated from immunized mice splenocytes using MACS Beads CD8^+ positive selection (Miltenyi Biotec). 10^5 lymphocytes were seeded into 96-well plates and either co-cultured with 5 × 10^4 B16F10 cells or peptides (10 μg/mL) or non-stimulated overnight and supernatants were harvested after 18 h. IFNγ was evaluated using Mouse IFNγ ELISA development kit (MabTech, Nacka Strand, Sweden) following manufactures instruction. In vivo cytotoxicity was evaluated as previously described. Splenocytes were harvested from naïve C57BL/6 mice and labeled with 0.2, lo, or 2 μM, hi, of CFSE in PBS for 5 min. Ten percent FCS containing PBS was used to halt labeling of the cells. Hi naïve target splenocytes were pulsed with target peptide (10 μg/mL) and lo naïve splenocytes were pulsed with control peptide (10 μg/mL). Labeled splenocytes were mixed and injected i.v. into naïve, antigen immunized (AI) or control immunized (CI) mice. After 20 h spleens were harvested from the mice and acquired by flow cytometry. In vivo cytotoxicity was calculated as follows: 100 — ((percentage of hi mCR16-25 peptide-pulsed targets in AI or CI mice/percentage of lo control targets in AI or CI mice)/(percentage of hi mCR16-25 peptide-pulsed targets in naïve mice/lo control targets in naïve mice) × 100). Samples were acquired on BD LSRII and analyzed on FlowJo.

Cancer cell spheroid culture

B16F10 were used in the generation of spheroid cultured tumor cells. B16F10 cells were seeded into Ultra-Low Cluster Plate (Costar) with 50,000 cells in 3 mL of melanoma spheroid culture medium. B16F10 melanoma spheroid culture medium consisted of MBM-4 (Lonza) containing the following: CaCl_2, bovine pituitary extract (BPE), recombinant human Fibroblast Growth Factor (rhFGF), recombinant human Insulin, hydrocortisone, PMA, GA-1000 and 10% FBS (Lonza, CC-3249). Cells were monitored daily and split, using enzymatic and mechanical dissociation, every third day for melanoma or depending on sphere aggregate cluster size. Cells were collected prior to sphere culturing and at every consecutive passage, which were denoted as P1, P2 and P3 as their passage number indicated.

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

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References

1. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. Nature 2011; 480:480-9; PMID:22193102; http://dx.doi.org/10.1038/nature10673
2. Shin DS, Ribas A. The evolution of checkpoint blockade as a cancer therapy: what's here, what's next? Curr Opin Immunol 2015; 33:23-35; PMID:25621841; http://dx.doi.org/10.1016/j.coi.2015.01.006
3. Pardoll DM. Immunology beats cancer: a blueprint for successful translation. Nat Immunol 2012; 13:1129-32; PMID:23160205; http://dx.doi.org/10.1038/ni.2392
4. Rosenberg SA, Rosenberg SA, Dudley ME. Adoptive cell therapy for the treatment of patients with metastatic melanoma. Curr Opin Immunol 2009; 21:233-40; PMID:19304471; http://dx.doi.org/10.1016/j.coi.2009.03.002
5. Snyder A, Makarov V, Merghouth T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS et al. Genetic Basis for Clinical Response to CTLA-4 Blockade in Melanoma. N Engl J Med 2014; 371:2189-99; PMID:25459260; http://dx.doi.org/10.1056/NEJMoai1406498
6. Rizvi NA, Hellmann MD, Snyder A, Kristporg M, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science 2015; 348:124-8; PMID:25765070; http://dx.doi.org/10.1126/science.aaa1348
7. Down GA, Dranoff G. Immunofluorescence therapy for cancer. Annu Rev Immunol 2009; 27:83-117; PMID:19007331; http://dx.doi.org/10.1146/annurev.immunol.021908.132544
8. Finn OJ. Cancer vaccines: between the idea and the reality. Nat Rev Immunol 2009; 27:83-117; PMID:19007331; http://dx.doi.org/10.1146/annurev.immunol.021908.132544
9. Schmidt P, Abken H. The beating heart of melanomas: a minor subset of cancer cells sustains tumor growth. Oncoarget 2011; 2:313-20; PMID:21487158; http://dx.doi.org/10.1016/j. oncarget.2011.04.029
10. Bianco C, Strizzi L, Normanno N, Khan N, Salomon DS. Cripto-1: An oncofetal gene with many faces. Curr Top Dev Biol 2005; 67:85-133; PMID:15949532; http://dx.doi.org/10.1016/S0070-2153(05)67003-2
11. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144:646-74; PMID:21376230; http://dx.doi.org/10.1016/j.coi.2009.03.002
12. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144:646-74; PMID:21376230; http://dx.doi.org/10.1016/j.coi.2011.02.013
13. Strizzi L, Margaryan NV, GIfgar A, Hardy KM, Normanno N, Salomon DS, Hendrix MJ. The significance of a Cripto-1 positive
subpopulation of human melanoma cells exhibiting stem cell-like characteristics. Cell Cycle 2013; 12:1450-6; PMID:23574716; http://dx.doi.org/10.4161/cc.24601

14. Schlaak M, Schmidt P, Bangard C, Kurschat P, Mauch C, Abken H. Regression of metastatic melanoma in a patient by antibody targeting of cancer stem cells. OncoTarget 2012; 3:322-30; PMID:22289880; http://dx.doi.org/10.18632/oncotarget.437

15. Llader A, Ljungberg K, Tufvesson H, Tazzari M, Roos A, Roos A-K, Quest AFG, Kiessling R. Intradermal DNA electroporation induces survival-specific CTLs, suppresses angiogenesis and confers protection against mouse melanoma. Cancer Immunol Immunother 2009; 58:91-92; PMID:19526360; http://dx.doi.org/10.1006/excr.2001.5195

16. Wechselberger C, Wechselberger C, Ebert A, Ebert AD, Bianco C, Bianco C, Khan NI, Khan NI, Sun Y, Sun Y et al. Cripto-1 Enhances Migration and Branching Morphogenesis of Mouse Mammary Epithelial Cells. Exp Cell Res 2001; 266:95-105; PMID:11339828; http://dx.doi.org/10.1006/excr.2001.5195

17. Ligtenberg MA, Rojas-Colonelli N, Kiessling R, Lladser A. NF-kappa B activation during intradermal DNA vaccination is essential for eliciting tumor protective antigen-specific CTL responses. Hum Vaccin Immunother 2014; 9:2189-95; PMID:23884215; http://dx.doi.org/10.1006/excr.2001.5195

18. Kato M, Kato M, Takahashi M, Takahashi M, Akhand AA, Akhand AA, Liu W, Liu W, Dai Y, Dai Y et al. Transgenic mouse model for skin malignant melanoma. Oncogene 1998; 17:1885-8; PMID:9778055; http://dx.doi.org/10.1038/sj.ong.1202077

19. Nakamura K, Yoshikawa N, Yamaguchi Y, Kagota S, Shinozuka K, Kunitomo M. Characterization of mouse melanoma cell lines by their immunodiagnostic factor that should not be ignored. Oncogene 2009; 29:1093-102; PMID:19948478; http://dx.doi.org/10.1016/j.annorel.2009.01.001

20. Kato M, Takahashi M, Takahashi M, Takahashi M, Akhand AA, Akhand AA, Liu W, Liu W, Dai Y, Dai Y et al. Transgenic mouse model for skin malignant melanoma. Oncogene 1998; 17:1885-8; PMID:9778055; http://dx.doi.org/10.1038/sj.ong.1202077

21. Su AI, Su AI, Wiltshire T, Wiltshire T, Batalov S, Batalov S, Lapp H, Lapp H, Ching KA, Ching KA et al. A gene atlas of the mouse and human protein-encoding transcriptomes. Proc Natl Acad Sci USA 2004; 101:6062-7; PMID:15075390; http://dx.doi.org/10.1073/pnas.0400782101

22. Bloom MB, Perry-Lalley D, Robbins PF, Li Y, El-Gamil M, Rosenberg SA, Yang JC. Identification of tyrosinase-related protein 2 as a tumor rejection antigen for the B16 melanoma. J Exp Med 1997; 185:453-9; PMID:9053445; http://dx.doi.org/10.1088/jem.185.3.453

23. Su AI, Su AI, Wiltshire T, Wiltshire T, Batalov S, Batalov S, Lapp H, Lapp H, Ching KA, Ching KA et al. A gene atlas of the mouse and human protein-encoding transcriptomes. Proc Natl Acad Sci USA 2004; 101:6062-7; PMID:15075390; http://dx.doi.org/10.1073/pnas.0400782101

24. De Luca A, Lamura L, Strizzi L, Roma C, D’Antonio A, Margaryan N, Pirozzi G, Hsu M-Y, Botti G, Mari E et al. Expression and functional role of CRIPTO-1 in cutaneous melanoma. Br J Cancer 2011; 105:1030-8; PMID:21863025; http://dx.doi.org/10.1038/bjc.2011.324

25. Mallikarjuna K, Vajjayanthi P, Krishnakumar S. Cripto-1 expression in uveal melanoma: An immunohistochemical study. Exp Eye Res 2007; 84:1060-6; PMID:17412323; http://dx.doi.org/10.1016/j.exer.2007.01.019

26. Bianco C, Rangel MC, Castro NP, Nagaoka T, Rollman K, Gonzales M, Salomon DS. Role of Cripto-1 in stem cell maintenance and malignant progression. Am J Pathol 2010; 177:532-40; PMID:20616345; http://dx.doi.org/10.1038/ajpath.2010.100102

27. Gross S, Geldmacher A, Sharav T, Losch F, Walden P. Immunosuppressive mechanisms in cancer: Consequences for the development of therapeutic vaccines. Vaccine 2009; 27:3398-400; PMID:19200836; http://dx.doi.org/10.1016/j.vaccine.2009.01.001

28. Schmidt P, Kopecky C, Hombach A, Zgrino P, Mauch C, Abken H. Eradication of melanomas by targeted elimination of a minor subset of tumor cells. Proc Natl Acad Sci USA 2011; 108:2474-9; PMID:21282657; http://dx.doi.org/10.1073/pnas.1009069108

29. Ljunggren HG, Ljunggren H-G, Stam NJ, Stam NJ, Ohlén C, Ohlén C, Neeffes JJ, Neeffes JJ, Höglund P, Höglund P et al. Empty MHC class I molecules come out in the cold. Nature 1990; 346:476-80; PMID:2198471; http://dx.doi.org/10.1038/346476a0

30. Wechselberger C, Wechselberger C, Ebert A, Ebert AD, Bianco C, Bianco C, Khan NI, Khan NI, Sun Y, Sun Y et al. Cripto-1 Enhances Migration and Branching Morphogenesis of Mouse Mammary Epithelial Cells. Exp Cell Res 2001; 266:95-105; PMID:11339828; http://dx.doi.org/10.1158/0008-5472.CAN-05-3153