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Modulation of epithelial immunity by mucosal fluid

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Mucosal epithelial cells, including those at the ocular surface, resist infection by most microbes in vivo but can be susceptible to microbial virulence in vitro. While fluids bathing mucosal surfaces (e.g. tears) contain antimicrobials, potentially pathogenic microbes often thrive in these fluids, suggesting that additional mechanisms mediate epithelial resistance in vivo. Here, tear fluid acted directly upon epithelial cells to enhance their resistance to bacterial invasion and cytotoxicity. Resistance correlated with tear fluid-magnified activation of NFκB and AP-1 transcription factors in epithelial cells in response to bacterial antigens, suggesting priming of innate defense pathways. Further analysis revealed differential regulation of potential epithelial cell defense genes by tears. siRNA knockdown confirmed involvement of at least two factors, RNase7 and ST-2, for which tears increased mRNA levels, in protection against bacterial invasion. Thus, the role of mucosal fluids in defense can include modulation of epithelial immunity, in addition to direct effects on microbes.

Pseudomonas aeruginosa is a Gram-negative opportunistic bacterium that does not normally infect healthy epithelial-lined tissue surfaces in vivo. However, epithelial cells derived from these tissues become exquisitely sensitive to a variety of virulence mechanisms of P. aeruginosa and other opportunistic pathogens if grown in cell culture media in vitro. One difference between the in vivo environment and standard in vitro cell culture conditions for mucosal epithelia is the presence versus absence of mucosal fluid. Indeed, it is commonly assumed that bacteriostatic or bactericidal activities of molecular factors in mucosal fluids protect against infection in vivo. Supporting a role for mucosal fluids in defense, we have previously shown that human tear fluid can protect corneal epithelial cells against P. aeruginosa invasion, and against cytotoxicity towards, corneal epithelial cells in vitro, and against virulence in vivo, if added with the bacterial inoculum. However, in the same study we reported that human tear fluid removed from the eye does not consistently inhibit the growth of P. aeruginosa strains that it protects cells against. This result suggested mechanisms independent of direct antimicrobial activity were involved in the epithelial protective effects of tear fluid. The same is likely to be true of other mucosal fluids, many of which are known to support survival of a rich and often complex microbiota.

Here, we used P. aeruginosa, tear fluid and corneal epithelial cells to test the hypothesis that mucosal fluid can act directly upon epithelial cells to enhance their resistance to bacterial-induced invasion and cytotoxicity mechanisms of bacteria.

**Results**

**Exposure to cells did not enhance bacteriostatic activity of tear fluid.** Epithelial cell induced activation of tear fluid bacteriostatic activity would provide a different explanation for why incubation of cells with tear fluid protects the cells from P. aeruginosa. To explore this alternate hypothesis, bacteriostatic activity of human tears pre-exposed to cultured corneal epithelial cells overnight was compared to tear fluid that had not been pre-exposed to cultured cells in vitro. Results showed that three of the four P. aeruginosa strains tested could grow in human tear fluid whether or not the tears were pre-exposed to human corneal epithelial cells in vitro, with no significant increase in bacteriostatic activity noted after the tears had been exposed to cells (Fig. 1a).

**Pre-exposing epithelial cells to human tear fluid increased cellular resistance to bacterial-induced invasion or cytotoxicity.** We next explored the impact of human tear fluid on cellular susceptibility to P. aeruginosa invasion or cytotoxic virulence mechanisms. Cells pre-exposed to human tear fluid for 16 h were compared to cells pre-exposed only to media (controls) for their susceptibility to P. aeruginosa invasion and cytotoxic activity, respectively. Before adding the bacterial inoculum, tear fluid was removed by washing cells with phosphate buffered saline. After washing, cells were challenged with 10⁴ cfu P. aeruginosa for 3 h. Cells pre-exposed to
Tear fluid had significantly reduced susceptibility to invasion by all of three bacterial strains tested, with 2.5-, 5- and 3.5-fold reductions compared to controls observed for \( P. \) aeruginosa strains 19660, 6294 and PAO1, respectively (Fig. 1b). Tear pre-exposure also increased cellular resistance to killing by the cytotoxic \( P. \) aeruginosa strain 6206 (3 fold) (Fig. 1c, d).

**Tear fluid magnified bacterial-induced activation of NFkB and AP-1 transcription factors.** The above data suggested that the mechanism(s) by which tear fluid protects against \( P. \) aeruginosa invasion and cytotoxicity involves modulation of corneal epithelial cell defenses. Thus, we used a 10-pathway Reporter Assay, which utilizes a transcription factor-responsive firefly luciferase construct and a constitutively-expressing \( Renilla \) luciferase construct, to determine the impact of tear fluid on transcription factor activation in corneal epithelial cells in response to bacterial challenge. Corneal epithelial cells were incubated with either tears or media for 16 h, washed, then incubated for 3 h with \( P. \) aeruginosa supernatant containing surface antigens (LPS, flagellin and pilin). Results showed that tear fluid pre-exposure magnified bacterial antigen-induced increases in luciferase expression controlled by NFkB and AP-1 transcription factors by ∼5- and 20-fold, respectively (Fig. 2a).

**Impact of tear fluid on expression of specific known epithelial-expressed antimicrobial factors.** Increased resistance to bacterial invasion and cytotoxicity accompanied by priming of two known innate defense signaling pathways, suggested that inducible antimicrobial factors in corneal epithelial cells might be upregulated by tear fluid. We began by exploring such factors already known to be expressed by these cells that have been shown to be inducible\(^9\)–\(^12\). Thus, mRNA for hBD-2, hBD-3 and LL-37 was examined after exposure to either media or tear fluid (Fig. 2b), with or without subsequent inoculation with \( P. \) aeruginosa antigens (Fig. 2c). Only hBD-3 mRNA expression was increased by tear fluid, and by only ∼1.5 fold (Fig. 2b).

Since our unpublished results showed cell lysate fractions containing factors < 3 kDa had antimicrobial activity against \( P. \) aeruginosa (data not shown), we next focused on molecules of that size known to be expressed by other epithelial cell types to explore if tear fluid induced their expression. Histatin 3, a precursor to histatin 5 and P113, was found expressed in significant quantities by the corneal epithelial cells, but only after tear pre-exposure (Fig. 2b). Bacterial exposure further enhanced the expression of histatin 3 in cells that had been pre-exposed to tear fluid. Histatin 5, derived by proteolytic cleavage of histatin 3, has potent antimicrobial activity and is produced by parotid and submandibular salivary glands\(^13\)–\(^14\). P113

![Figure 1](https://www.nature.com/scientificreports/)

**Figure 1 | Pre-exposing Epithelial Cells to Human Tear Fluid Protects against \( P. \) aeruginosa Invasion and Cytotoxicity, without affecting Tear Bacteriostatic Activity.** a, \( P. \) aeruginosa (\( \sim10^4 \) cfu) were suspended in cell culture media, tears, or tears pre-exposed to corneal epithelial cells for 3 h at 37 °C. Tear bacteriostatic activity was unaffected by prior exposure to epithelia. b, c, d, Pre-exposure to tear fluid protected corneal epithelial cells against \( P. \) aeruginosa invasion (b) and cytotoxicity (strain 6206) measured by trypan blue staining of dead/dying cells (c, d). Epithelial cells were exposed to tear fluid or cell culture media for 16 h then inoculated with \( P. \) aeruginosa (\( \sim10^4 \) cfu in cell culture media, 3 h at 37 °C, see methods).
(<1.5 kDa), a cleaved product of its parent molecule, histatin 5, also has potent antibacterial activity against *P. aeruginosa*.

**Microarray analysis of tear fluid effects.** To determine if there might be other critical defense genes differentially regulated by tear fluid, we performed full human genome microarray analysis of cells pre-exposed to tear fluid, with and without subsequent exposure to bacterial antigens. With exposure to tear fluid only, 192 and 29 probe sets were up- and down-regulated, respectively by at least 4 fold or more (Fig. 3a). Pre-exposure to tear fluid for cells that were then exposed to *P. aeruginosa* antigens (compared to *P. aeruginosa* antigen challenge without tear pre-exposure) up- and down-regulated 151 and 160 probe sets, respectively by at least 4 fold or more (Fig. 3b). Two genes, RNase7 and ST2 were highly upregulated by multiple probe sets in both sets of experiments; tear fluid upregulated RNase7 by 28–40 fold and ST2 by 18–198 fold, and cells exposed to *P. aeruginosa* antigens with pre-exposure to tears upregulated RNase7 by 24–47 fold and ST2 by 33–171 fold. Semi-quantitative RT-PCR confirmed upregulation of these factors (Fig. 4a, b, c).

**siRNA knockdown studies.** Since RNase7 and ST2 have both been shown to play protective roles in immunity, and mRNA levels of each were significantly increased by tear fluid pre-exposure, which also protected cells against *P. aeruginosa*, the impact of siRNA knockdown of these molecules on epithelial cell resistance to *P. aeruginosa* was explored. Knockdown of either RNase7 or ST2, was found to significantly enhance susceptibility of the cells to *P. aeruginosa* invasion (Fig. 4d). These results suggest that each of these two factors, for which mRNA levels were increased by tear fluid pre-exposure, are directly involved in cellular resistance to *P. aeruginosa* invasion.

**Discussion**

The data presented in this article show that tear fluid can induce protective immunity in corneal epithelial cells against *P. aeruginosa* invasion and cytotoxic virulence mechanisms. Tear fluid exposure was found to change the expression of many genes, with the most significant increased mRNA levels observed for RNase7 and ST2. Accordingly, knockdown of either RNase7 or ST2 increased cellular susceptibility to *P. aeruginosa* invasion, showing both factors play a role in tear fluid-induced epithelial immunity to this pathogen.

RNase7 is a known antimicrobial (~14.5 kDa) with potent ribonuclease activity first identified in human skin, found to play a major role in determining the skin antimicrobial barrier. It has potent antimicrobial activity against various pathogens including *E. coli*, *Pseudomonas aeruginosa*, *Candida albicans*, *Staphylococcus...*
also explain its capacity to reduce pathology during infection; i.e., modulation of epithelial cell susceptibility to bacterial virulence factors. Given the complexity of already known effects of ST2, particularly on TLR signaling, further studies will be needed to elucidate exactly how ST2 protects cells against bacteria. However, possibilities include that it impacts production of epithelial-expressed antimicrobials via its effects on TLR-mediating signaling or Th2 responses, which each regulate expression of antimicrobial peptides. Alternatively ST2 could impact, via effects on microbe recognition, bacterial trafficking to intraepithelial perinuclear vacuoles that are capable of killing intracellular bacteria. Tear fluid was found to prime cells for NFκB and AP-1 transcription factor activation, supporting that downstream innate immune responses are impacted.

Recent studies have shown that the normal microbiota at some mucosal surfaces can induce immunity in epithelial cells lining the mucosa, involving TLRs which detect microbe-associated molecular patterns (MAMPs). DAMPs (damage associated molecular patterns) can also trigger TLR receptor signaling and therefore downstream effects. Since the ocular surface lacks significant microbiota, other mechanisms must be involved in maintaining the normal immune tone of its surface epithelial cells during health. Data presented in this report showing that tear fluid can modulate epithelial immunity suggests the presence of TAMPs (tear associated molecular patterns). Whether these act on the same receptors/signaling pathways as other factors that modulate epithelial immunity, and whether components of tears responsible are similar in structure to MAMPs or PAMPs, remains to be determined.

While we have found changes to gene expression induced by tear fluid, some involved in epithelial immunity to bacteria, it remains possible that tear fluid influences cell immunity in other ways, for example if tear components directly impact the surface biochemistry of the epithelial cells, e.g. by binding directly to cells or by modifying cellular factors directly. Further studies will be needed to address that possibility.

The significance of these findings extends to the design of in vitro cell culture studies. Standard methods for studying epithelial cell biology, including cellular microbiology, involve growth of cells in tissue culture media. The results of this study suggest that growth media should be formulated to mimic the appropriate mucosal fluid when studying mucosal epithelial cells in vitro.

Factors that stagnate mucosal fluids often cause susceptibility to infection, such as cystic fibrosis, intubation, and contact lens wear. While reduced clearance of microbes and/or biofilm formation very likely contributes to susceptibility in these instances, the impact of prolonged stagnation on mucosal fluid biochemistry and its subsequent impact on epithelial immune tone, could be another topic worthy of investigation.

In summary, the results of this study show that the role of mucosal fluids in defense against infection stretches beyond effects on microbe viability or virulence to actually enhancing cellular resistance to microbes. These findings add to our understanding of how mucosal surfaces resist infection during health, and suggest new avenues for developing biocompatible agents to prevent, rather than simply treat, pathology in susceptible patients.

**Methods**

**Cell culture.** Human telomerase-immortalized corneal epithelial cells (hTCEpi) were maintained in 10 cm tissue culture treated Petri dishes (Becton Dickinson, Franklin Lake, NJ) in serum-free KGM-2 medium (Lonza, Walkersville, MD) until confluent as previously described. Cells were then seeded onto 96-well tissue culture plate (Becton Dickinson, Franklin Lake, NJ) and grown to 80–90% confluence. 2 days prior to experiment, the cells were incubated in high calcium KGM-2 medium containing 1.15 mM Ca²⁺ for 16 h, then washed with sterile phosphate-buffered saline (PBS; Sigma, St. Louis, MO) before being incubated in either 40 µl of fresh human tear fluid or high calcium KGM-2 without antibiotics either for 6 h or 16 h depending on the experiment. All cells were incubated at 37˚C with 5% CO₂ while culturing and during experiments.

**Figure 3 | Microarray Analysis of Epithelial Cells Exposed to Tear and/or P. aeruginosa antigens.** a, b Heat-maps of differentially regulated probe sets in epithelial cells incubated with human tear fluid for 6 h (a) or cells pre-exposed to tear fluid for 16 h followed by 3 h incubation with P. aeruginosa antigens (b). Red and green boxes indicate up- and down-regulation, respectively, of probe sets with tear fluid.
Figure 4 | RT-PCR Confirmation of Tear Induced Expression of RNase7 and ST2 and Their Protective Effect against P. aeruginosa Invasion. a, b, c, d, siRNA knockdown of RNase7 or ST2 in corneal epithelial cells increased P. aeruginosa invasion.

Preparation of bacteria. All bacteria were grown on trypticase soy agar plates overnight at 37°C. Bacteria were suspended in basal KBM-2 media (without antibiotics) to a spectrophotometer optical density of 0.1 at OD_{600} which is equivalent to \( \sim 1 \times 10^6 \text{ cfu/ml} \) for invasion and cytotoxicity experiments. P. aeruginosa clinical isolates 6294 and 6206 were used for invasion and cytotoxicity assays, respectively. Some experiments also involved the use of two laboratory isolates of P. aeruginosa, strains PAO1 and PAO1-K-960.

P. aeruginosa supernatant. P. aeruginosa strain PAO1 was grown as single colonies on trypticase soy agar at 37°C overnight. A single colony was then inoculated into 5 ml of trypticase soy broth and grown with aeration to late log phase (37°C). The culture broth was then centrifuged at 14,000 rpm (16,000 \( \times \) g) for 30 min and the supernatant collected in a syringe, sterilized with a 0.22 \( \mu \)m polymer filter (Corning Star Corporation, Cambridge, MA), aliquoted and stored at \(-80^\circ\text{C}\) until use in experiments. Bacterial supernatant diluted 1:5 with basal KBM-2 media (without antibiotics), or an equivalent amount of basal cell culture medium, was used to stimulate corneal epithelial cells for experiments.

Invasion and cytotoxicity assays. Corneal epithelial cells were either pre-exposed to high Ca\(^{2+}\) media or human tear fluid for 16 h (for tear fluid pre-exposure experiments), washed with 100 \( \mu \)l of pre-warmed PBS, then inoculated with 40 \( \mu \)l of P. aeruginosa strain 6294 or strain 6206 diluted in basal KBM-2 media to a concentration of \( \sim 1 \times 10^6 \text{ CFU/ml} \). Triplicate wells were used for each sample. Epithelial cells were incubated with bacteria for 3 h (37°C, 5.0% CO\(_2\)). The cells were then washed with PBS and incubated with 100 \( \mu \)l of gentamicin solution (0.4% [vol/vol]; 200 \( \mu \)g/ml; BioWhittaker, Walkerville, MD) in cell culture media. After a 1 h incubation at 37°C, 100 \( \mu \)l of Trypan blue solution (10% [vol/vol]) in basal KBM-2 media was added, and incubated for 15 min. The Trypan blue solution was then replaced with 100 \( \mu \)l of basal KBM-2 media. Phase-contrast images were taken on an inverted microscope (Olympus IX-70, Olympus America Inc., Center Valley, PA) attached to a video camera (Optronics, Goleta, CA) at 200 \( \times \) magnification. Epithelial cells were considered vulnerable to bacterial cytotoxicity (i.e. dead/dying) if they exhibited Trypan blue staining. Blue stained cells were also quantified for statistical analysis. To quantify intracellular bacteria, epithelial cells were lysed in Triton X-100 (0.25% [vol/vol]; LabChem Inc., Pittsburgh, PA) in PBS for 15 minutes. The cells were then scraped for complete lysis and undiluted and \( 10^{-1} \) dilutions plated onto MacConkey agar plates to enumerate surviving (intracellular) bacteria by viable count.

Bacterial growth assay. Bacterial growth and viability were tested by adding 50 \( \mu \)l of bacteria suspended in either cell culture media, or tears with or without exposure to corneal cells, to empty Eppendorf tubes. These suspensions were incubated at 37°C for 3 h and quantified by viable count. Initial suspensions were also quantified to study bacteriostatic or bactericidal effects of tear fluid. These were done in triplicates.

Transfection and luciferase array assay. Human corneal epithelial cells were reverse transfected using SureFECT (SABiosciences, Frederick, MD) onto Cancer 10-Pathway Reporter Array (SABiosciences, Frederick, MD). Each reporter consists of a mixture of an inducible transcription factor responsive firefly luciferase reporter and a constitutively expressing Renilla luciferase construct (20:1). Firefly and Renilla luciferase activities of cells incubated either in media or tear fluid for 16 h followed by 3 h incubation with P. aeruginosa supernatant were measured. The change in the activity of each signaling pathway was calculated by comparing the normalized luciferase activities of the reporter of each group.

RNA purification. Total RNA of corneal epithelial cells incubated in either cell culture media or human tear fluid for 6 h and cells either pre-exposed to media or tear fluid for 16 h followed by 3 h incubation with P. aeruginosa supernatant were...
extracted with the RNeasy Kit (Qiagen, Valencia, CA) using Qiagenexedar columns for cell lysis and inserting Qiagen on-column DNase steps to remove any contaminating genomic DNA.

**RNA target preparation/GeneChip microarray analysis.** RNA quality was assessed by using the Agilent Model 2100 Bioanalyzer (Agilent Technologies, Palo Alto, CA). Ten micrograms of total RNA was processed on the microarray by using the Affymetrix GeneChip one-cycle target labeling kit (Affymetrix, Santa Clara, CA) according to the manufacturer's recommended protocols. The resultant biotinylated cRNA was fragmented and then hybridized to the GeneChip Human Genome U133 Plus 2.0 Arrays (Affymetrix). The arrays were washed, stained, and scanned using the Affymetrix Model 450 Fluidics Station and Affymetrix Model 3000 scanner, following manufacturer's recommended protocols, at the University of California Functional Genomics Laboratory. Expression values were generated by using Microarray Suite (MAS) v5.0 software (Affymetrix). Each sample and hybridization underwent a quality control evaluation, including cRNA amplification of >4-fold, percentage of probe sets reliably detecting between 40% and 60% present call, and 3'-5' ratio of GAPDH gene.

**Evaluation and normalization of affymetrix GeneChip data.** The hybridizations were normalized by using the RNA (robust multichip averaging) method from Biocductor to obtain summary expression values for each probe set. Gene expression levels were analyzed on a logarithmic scale. The given background-corrected and normalized log2 values were anti-logged and fold expression levels were analyzed on a logarithmic scale. The given background-corrected and normalized log2 values were anti-logged and fold expression levels were analyzed on a logarithmic scale. The given background-corrected and normalized log2 values were anti-logged and fold expression levels were analyzed on a logarithmic scale. The given background-corrected and normalized log2 values were anti-logged and fold expression levels were analyzed on a logarithmic scale.

**Statistical analysis.** Data were expressed as mean ± s.d. (n = 3 or 4 samples per group) unless otherwise stated. Student's t-test was used to evaluate the statistical significance of differences between groups with P values <0.05 considered significant. All experiments were repeated at least three times.
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**Author contributions**

JM and CT performed all experimental work. All authors contributed to the experimental design, interpretation of data, and writing the manuscript. SMJF and DJE supervised the project.

**Additional information**

Competing financial interests: The authors declare no competing financial interests.

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