Rad9/53BP1 promotes crossover recombination DNA repair by limiting the Sgs1 and Mph1 helicases

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Abstract

A DNA double strand break (DSB) is primed for homologous recombination (HR) repair through the nucleolytic processing (resection) of its ends, leading to the formation of a 3’ single-stranded DNA (ssDNA). Generation of the ssDNA is accompanied by the loading of several repair factors, including the ssDNA binding factor RPA and the recombinase Rad51. Then, depending upon the availability and location of a homologous sequence, different types of HR mechanisms can occur. Inefficient or slow HR repair results in the activation of the DNA damage checkpoint (DDC)¹. In budding yeast, the 53BP1 ortholog Rad9 acts as a scaffold, mediating signal from upstream kinases Mec1 and Tel1 (ATR and ATM in human) to downstream effectors kinases Rad53 and Chk1 (CHK2 and CHK1 in human). In addition to its role in DDC, Rad9 limits DSB resection². Remarkably, this function is conserved in 53BP1, also being implicated in cancer biology in human cells³,⁴.

Here we show that Rad9 limits the recruitment of the helicases Sgs1 and Mph1 on to a DSB, promoting Rad51-dependent recombination with long track DNA conversions, crossovers and break-induced replication (BIR). This regulation couples the DDC with the choice and effectiveness of HR sub-pathways, and might be critical to limit genome instability with implication for cancer research.
Main

Rad9 is the 53BP1 ortholog in *S. cerevisiae* (budding yeast). It plays fundamental role in DNA damage signalling, mediating phosphorylation from upstream kinases Mec1/ATR and Tel1/ATM. In addition to this function, we described a DDC-independent role of Rad9 in the regulation of DSB resection, limiting repair through single strand annealing (SSA). Indeed, the Rad9 binding to a DSB reduces the Exo1 and Dna2-Sgs1 recruitment, affecting the long-range resection. However, it is not understood how the formation of the 3’-end filament is coupled with the selection of appropriate HR sub-pathway, and whether Rad9 might have a role to determine that choice.

To investigate further the role of Rad9 in DSB repair, we performed a genetic assay with a diploid system, which allows to study the repair of DSB through gene conversion (GC) and break-induced replication (BIR), after the induction of a DSB by I-SceI in chromosome XV. Importantly, this system not only measures frequencies of crossover (CO) and noncrossover (NCO) in DSB repair, but also distinguishes between short and long tract gene conversion events (Fig. 1a). Upon I-SceI induction, in the absence of RAD9 we observed a striking increase in the percentage of short track GC (white colonies in the assay) (Fig. 1b). By screening colonies for specific markers, we also found that rad9Δ cells have reduced frequency of both CO and BIR events (Fig. 1c, and Extended data Table 1). Based on these results, we hypothesized that Rad9 might control strand invasion-mediated mechanisms to repair a DSB, in addition to its function in limiting SSA.

To study more precisely the role of Rad9 in the BIR process, we took advantage of an haploid genetic system engineered to test only this HR subpathway. Briefly, one DSB is induced in chromosome V by the endonuclease HO and it is repaired by BIR thanks to a donor sequence on chromosome XI (Fig. 1d). Interestingly, analysing the DSB repair by Southern blotting, in exponentially growing or nocodazole-arrested cells (Fig. 1e, f), we found that RAD9 deletion severely affected BIR, regardless of the cell cycle stage. Considering that DDC signalling to Pif1 helicase contributes to BIR, we speculated that this would be the reason of the rad9Δ cells defect in the assay. However, after plating the cells
in galactose to induce HO, we surprisingly observed higher cell lethality in \textit{rad9Δ} cells with respect to \textit{rad53-K227A chk1Δ} cells, that are defective in DDC signalling downstream from Rad9, and \textit{pif1Δ} cells (Fig. 1g). Therefore, cells lacking \textit{RAD9} would be defective in BIR for a reason other than deficient signalling to Pif1.

Then, we tested critical steps in Rad51-dependent HR in \textit{rad9Δ} cells. First, by chromatin immunoprecipitation (ChIP), we found that the recombination factors Rpa1, Rad52 and Rad51 were efficiently recruited at the cut site in \textit{rad9Δ} cells (Fig. 2a, b, c, and Extended data Fig. 1). However, their binding near to the break ends was significantly higher in the absence of \textit{RAD9}, especially at later time points. Since the amount of ssDNA close to the break site in wild type and \textit{rad9Δ} cells was similar (Extended data Fig. 2), the increased recruitment of Rpa1, Rad52 and Rad51 on the DSB was unlikely due to higher amount of available substrate in \textit{rad9Δ} cells. Instead, we hypothesized that the loading and oligomerization of Rad9 protein on the DSB might physically dampen the recruitment of the recombination factors, similarly to its role in limiting nuclease activity for DSB resection\textsuperscript{2,5,6}. To test this hypothesis, we expressed from a plasmid the wild type Rad9 or the two protein variants Rad9-2Ala and Rad9-7xA, both reducing the Rad9 binding and oligomerization on the DSB\textsuperscript{5,10}, in \textit{rad9Δ} cells. We found that both the Rad9 variants, contrary to the wild type form, did not completely rescue the lethality of \textit{rad9Δ} cells in the BIR assay (Extended data Fig. 3), supporting the idea that Rad9 might affect BIR though a physical role at the DSB site.

Despite the Rad51 binding at the DSB was increased (Fig. 2c), we found that it was not enriched at the donor site in \textit{rad9Δ}, after DSB induction in G2/M blocked cells (Fig. 2d). This result indicates that cells lacking \textit{RAD9} cannot form a stable Displacement-loop (D-loop) structure and synapsis between DSB and the donor template, providing molecular evidence of the failure in DSB repair by BIR. Consistent with these data, \textit{RAD9} deletion impaired the D-loop extension, measured through a PCR-based assay to a greater extent than \textit{rad53-K227A chk1Δ} and \textit{pif1Δ} mutations (Fig. 3a, b, c). These results suggest that Rad9 promotes strand invasion and D-loop extension in BIR, through a mechanism independent on the Chk1 and Rad53 signalling. Of note, previous works have
shown that the annealing between the two DNA strands in D-loop formation can either be promoted by a Rad51-dependent process or rejected by the Sgs1-Top3-Rmi1 complex and Mph1\textsuperscript{11-15}. Strikingly, after deletion of the genes coding for the two helicases Sgs1 and Mph1 in rad\textsuperscript{9}\Delta cells, both viability and primer extension analyses showed a rescue to the level found in pif\textsuperscript{1}\Delta cells (Fig. 3d, e, f). Moreover, by ChIP analysis, we also found higher binding of Sgs1 and Mph1 on to a persistent DSB in rad\textsuperscript{9}\Delta cells (Fig. 3g, h). As Sgs1 interacts with Rpa1 and Rad51\textsuperscript{16,17}, while Mph1 interacts with Rpa1 and Rad52\textsuperscript{18,19}, we are tantalized to suggest that the hyper-loading of Sgs1 and Mph1 at the DSB may be related to the increased recruitment of Rad51, Rad52 and Rpa1 in rad\textsuperscript{9}\Delta cells (Fig. 2a, b, c, and Extended Fig. 1). In particular, it has been recently shown that Sgs1 interacts with Rad51 and the sgs\textsuperscript{1}-F1192D (sgs\textsuperscript{1}-FD) mutation abolishes this interaction\textsuperscript{17}. We found that the sgs\textsuperscript{1}-FD partially rescued the viability of cells lacking RAD9 in the BIR assay (rad\textsuperscript{9}\Delta = 1.29 % ± 0.32; sgs\textsuperscript{1}-FD rad\textsuperscript{9}\Delta = 2.67 % ± 0.03; mph\textsuperscript{1}\Delta sgs\textsuperscript{1}-FD rad\textsuperscript{9}\Delta = 8.25 % ± 0.11) (Fig. 3d, i), similarly to the deletion of SGS1 (sgs\textsuperscript{1}\Delta rad\textsuperscript{9}\Delta = 5.73 % ± 0.83; mph\textsuperscript{1}\Delta sgs\textsuperscript{1}\Delta rad\textsuperscript{9}\Delta = 11.87 % ± 1.78) (Fig. 3d). This result suggests that lowering the interaction between Sgs1 and Rad51 might reduce strand rejection, favouring BIR in rad\textsuperscript{9}\Delta cells. As because the interplay between Rad9 and Sgs1 regulates also DSB resection\textsuperscript{5,6}, we measured the kinetic of ssDNA formation at different positions from the break in the sgs\textsuperscript{1}-FD mutant. We found that this allele did not alter resection of an irreparable DSB, which still remained faster in the sgs\textsuperscript{1}-FD rad\textsuperscript{9}\Delta double mutant cells (Extended data Fig. 2). These results, together with the viability of the sgs\textsuperscript{1}-FD rad\textsuperscript{9}\Delta cells in the BIR assay (Fig. 3i), suggest that the kinetic of the DSB resection is not responsible per se of the severe BIR defect of the rad\textsuperscript{9}\Delta cells.

Because of the reduced levels of CO and long track gene conversion in rad\textsuperscript{9}\Delta cells (Fig 1a, b, c), we investigated the role of Rad9 in DSB repair by using an ectopic gene conversion (eGC) system\textsuperscript{11}, which is coupled to DDC activation\textsuperscript{20}. This assay allows us to detect both the CO and NCO repair products by Southern blotting (Fig 4a). Of note, deletion of SGS1 and MPH1 greatly increase CO\textsuperscript{11,12}. Significantly, we found that rad\textsuperscript{9}\Delta cells had reduced repair product through both NCO and CO, with a major effect on the latter, when the DSB was induced in cells blocked in G2/M phase (Fig. 4b, c). Once again, by deleting SGS1 and MPH1 we
observed a partial rescue of the DSB repair defect in rad9Δ cells, specifically for the less frequent class of repair associated with CO (Fig. 4b, c).

Overall, this work provides experimental evidence of an unprecedented role of Rad9 in DSB repair, controlling the fate of 3′ Rad51 filament in HR. Interestingly, limiting the Sgs1 helicase, Rad9 antagonizes the formation of extended 3′ ssDNA for recombination while, limiting both Sgs1 and Mph1, reduces the disassembling of D-loop structure once repair DNA synthesis is commenced (Fig. 4d). This reduces SSA and favours DSB repair through HR sub-pathways that require stable D-loop, such as long tract GC, eGC and BIR, also increasing the frequency of CO outcome. Accordingly, RAD9 deletion limits sister chromatid exchanges and promotes Rad1/XPF-dependent translocations, likely through SSA 21,22. Moreover, in line with our model, eliminating the SGS1 gene in rad9Δ cells causes dramatic levels of translocations between homeologous sequences and complex chromosomal rearrangements 23,24.

In conclusion, we suggest that the Rad9 recruitment at a DSB guarantees that the DNA break is engaged into complex and slow HR sub-pathways only when cell division is restrained by the DDC activation. This reduces the risk of premature segregation of chromosomes with DNA linkages caused by unresolved HR intermediates, which leads to the formation of anaphase bridges and deleterious chromosome rearrangements25,26. As such, this regulation might be critical to limit genetic instability, a hallmark of cancer cells. Indeed, 53BP1-depleted cancer cells accumulate ultrafine anaphase bridges, suggesting that this mechanism might be evolutionary conserved also in higher eukaryotes27.

**Methods**

**Yeast strains, Media and Growth conditions.** All the strains listed in Extended data Table 2 are derivative of JKM179, JRL92, tGI354 and W303. To construct strains standard genetic procedures were followed28. Deletions and tag fusions were generated by the one-step PCR system. The sgs1-F1192D was obtained using a Cas9 mediated gene targeting system29. For the indicated experiments, cells were grown in YEP medium enriched with 2% glucose (YEP+glu), 3%
raffinose (YEP+raf) or 3% raffinose and 2% galactose (YEP+raf+gal). Unless specified all the experiments were performed at 28 °C. To block cells in G2/M, 20 μg/ml nocodazole was added to the cell culture.

**Cell viability assay.** JRL92 derivative strains were inoculated in YEP+raf, grown O/N at 28°C. The following day, cells were normalized and plated on YEP+glu and YEP+gal. Plates were incubated at 28 °C for three days. Viability results were obtained from the ratio between number of colonies on YEP+gal and YEP+glu. Standard error of the mean (SEM) was calculated on two independent experiments.

**Southern blot analysis.** Purified genomic DNA was digested with the appropriate restriction enzyme/s, probed with a specific 32P-labeled probe and scanned with a Typhoon Imager (GE healthcare). In JRL92 a CAN1 fragment was used as a probe, the % of BIR repair has been calculated using the donor band as a loading control. Repair in tGI354 background was analyzed as described previously20. Densitometric quantification of the bands intensity was performed using the ImageJ software. The SEM was calculated on three independent experiments.

**ChIP analysis.** ChIP analysis was performed as described previously5. Input and immunoprecipitated DNA were analysed by quantitative PCR, using a Bio-Rad CFX connect, or droplet digital PCR (ddPCR), using a Bio-Rad QX200 droplet reader. The oligonucleotides used are listed in Extended data Table 3. Data are presented as fold enrichment at the HO cut site (at the indicated distance from the DSB) over that at the KCC4 locus on the left arm of chromosome III or CAN1
locus on chromosome V. Then normalized to the corresponding input sample. SEM was calculated on two independent experiments.

**D-loop extension analysis.** To measure the DNA synthesis after D-loop formation during BIR we adopted a strategy that was described previously\(^8\). In our experiments, the genomic DNA at 0h and 24h after DSB formation was amplified by PCR with oligonucleotides on the *CAN1* locus and, as a control, on *TLC1* locus (Expanded data Table 3). See a scheme in Fig. 2b.

**Quantitative analysis of DSB end resection by Real time PCR.** Quantitative PCR (qPCR) analysis of DSB resection was performed as already described\(^30\). The oligonucleotides used are listed in Expanded data Table 2. The DNA was digested with the *Rsa*I restriction enzyme (NEB) that cuts inside the amplicons at several distances from the DSB, but not in the *KCC4* control region on chromosome III. qPCR was performed on both digested and undigested templates using StoS Quantitative Master Mix 2X SYBR Green (Genespin) with the Bio-Rad CFX connect qPCR system. The ssDNA percentage over total DNA was calculated using the following formula: % ssDNA = \(100/[(1 + 2^{\Delta Ct})/2]/f\), in which \(\Delta Ct\) values are the difference in average cycles between digested template and undigested template of a given time point and \(f\) is the HO cut efficiency measured by qPCR.

**DSB-induced recombination assay.** Homozygous diploids of the wild type and *rad9Δ* strains derived from LSY2205-11C and LSY2543 (W303 background) were grown in YEP+raf overnight at 28°C. Cells were plated on YEP+glu and YEP+gal plates to calculate plating efficiency and ensure *ADE2* auxotrophy before I-SceI induction. Separately, 2% galactose was added to the cell culture to induce I-SceI. After 1.5h of induction, cells were plated on YEP+glu plates and
incubated for 2-3 days at 28°C. Percentage of Red (the class of two long track conversions), White (the class of two short track conversions) and Red/White (the class of one long and one short track conversion) was determined only for recombinant colonies. All the plates were replicated on YEP+glu plates containing Hygromycin, Nourseothricin and Synthetic Complete (SC) medium-ura, SC-met, SC-ade, SC-ade+raf, SC-ade+gal plates to determine the percentage of recombinant colonies and distinguish the NCO, CO and BIR events in each sub-class of the colonies7.

**Statistical analysis.** Statistical analysis was performed using Microsoft Excel or Prism softwares. p-values were determined by an unpaired two-tailed t-test. No statistical methods or criteria were used to estimate sample size or to include or exclude samples.

1. Symington, L. S. & Gautier, J. Double-strand break end resection and repair pathway choice. *Annu Rev Genet* **45**, 247–271 (2011).
2. Lazzaro, F. *et al.* Histone methyltransferase Dot1 and Rad9 inhibit single-stranded DNA accumulation at DSBs and uncapped telomeres. *EMBO J.* **27**, 1502–12 (2008).
3. Bouwman, P. *et al.* 53BP1 loss rescues BRCA1 deficiency and is associated with triple-negative and BRCA-mutated breast cancers. *Nat Struct Mol Biol* **17**, 688–695 (2010).
4. Bunting, S. F. *et al.* 53BP1 inhibits homologous recombination in Brca1-deficient cells by blocking resection of DNA breaks. *Cell* **141**, 243–254 (2010).
5. Ferrari, M. *et al.* Functional Interplay between the 53BP1-Ortholog Rad9 and the Mre11 Complex Regulates Resection, End-Tethering and Repair of a Double-Strand Break. *PLoS Genet.* **11**, e1004928 (2015).
6. Bonetti, D. *et al.* Escape of Sgs1 from Rad9 inhibition reduces the
requirement for Sae2 and functional MRX in DNA end resection. *EMBO Rep.* **16**, 351–361 (2015).

7. Ho, C. K., Mazón, G., Lam, A. F. & Symington, L. S. Mus81 and Yen1 Promote Reciprocal Exchange during Mitotic Recombination to Maintain Genome Integrity in Budding Yeast. *Mol. Cell* **40**, 988–1000 (2010).

8. Lydeard, J. R., Jain, S., Yamaguchi, M. & Haber, J. E. Break-induced replication and telomerase-independent telomere maintenance require Pol32. *Nature* **448**, 820–823 (2007).

9. Vasianovich, Y., Harrington, L. A. & Makovets, S. Break-Induced Replication Requires DNA Damage-Induced Phosphorylation of Pif1 and Leads to Telomere Lengthening. *PLoS Genet.* **10**, e1004679 (2014).

10. Naiki, T., Wakayama, T., Nakada, D., Matsumoto, K. & Sugimoto, K. Association of Rad9 with double-strand breaks through a Mec1-dependent mechanism. *Mol Cell Biol* **24**, 3277–3285 (2004).

11. Ira, G., Malkova, A., Liberi, G., Foiani, M. & Haber, J. E. Srs2 and Sgs1-Top3 suppress crossovers during double-strand break repair in yeast. *Cell* **115**, 401–411 (2003).

12. Prakash, R. *et al.* Yeast Mphl helicase dissociates Rad51-made D-loops: Implications for crossover control in mitotic recombination. *Genes Dev.* **23**, 67–79 (2009).

13. Fasching, C. L., Cejka, P., Kowalczykowski, S. C. & Heyer, W.-D. Top3-rmi1 dissolve rad51-mediated d loops by a topoisomerase-based mechanism. *Mol. Cell* **57**, 595–606 (2015).

14. Jain, S., Sugawara, N., Mehta, A., Ryu, T. & Haber, J. E. Sgs1 and Mph1 Helicases Enforce the Recombination Execution Checkpoint During DNA Double-Strand Break Repair in Saccharomyces cerevisiae. *Genetics* **203**, 667–75 (2016).

15. Piazza, A. *et al.* Dynamic Processing of Displacement Loops during Recombinational DNA Repair. *Mol. Cell* **1**, 1–31 (2019).

16. Hegnauer, A. M. *et al.* An N-terminal acidic region of Sgs1 interacts with Rpa70 and recruits Rad53 kinase to stalled forks. *EMBO J* (2012). doi:10.1038/emboj.2012.195

17. Campos-Doerfler, L., Syed, S. & Schmidt, K. H. Sgs1 Binding to Rad51
Stimulates Homology-Directed DNA Repair in Saccharomyces cerevisiae. *Genetics* 208, 125–138 (2018).

18. Banerjee, S. *et al.* Mph1p promotes gross chromosomal rearrangement through partial inhibition of homologous recombination. *J. Cell Biol.* 181, 1083–1093 (2008).

19. Silva, S. *et al.* Mte1 interacts with Mph1 and promotes crossover recombination and telomere maintenance. *Genes Dev.* 30, 700–717 (2016).

20. Dibitetto, D. *et al.* Slx4 and Rtt107 control checkpoint signalling and DNA resection at double-strand breaks. *Nucleic Acids Res.* 44, 669–682 (2016).

21. Conde, F. *et al.* The Dot1 histone methyltransferase and the Rad9 checkpoint adaptor contribute to cohesin-dependent double-strand break repair by sister chromatid recombination in Saccharomyces cerevisiae. *Genetics* 182, 437–446 (2009).

22. Fasullo, M., Bennett, T., AhChing, P. & Koudelik, J. The Saccharomyces cerevisiae RAD9 checkpoint reduces the DNA damage-associated stimulation of directed translocations. *Mol. Cell. Biol.* 18, 1190–1200 (1998).

23. Vasan, S., Deem, A., Ramakrishnan, S., Argueso, J. L. & Malkova, A. Cascades of Genetic Instability Resulting from Compromised Break-Induced Replication. *PLoS Genet.* 10, e1004119 (2014).

24. Schmidt, K. H., Wu, J. & Kolodner, R. D. Control of Translocations between Highly Diverged Genes by Sgs1, the Saccharomyces cerevisiae Homolog of the Bloom’s Syndrome Protein. *Mol. Cell. Biol.* 26, 5406–5420 (2006).

25. Germann, S. M. *et al.* TopBP1/Dpb11 binds DNA anaphase bridges to prevent genome instability. *J. Cell Biol.* 204, 45–59 (2014).

26. Chan, Y. W., Fugger, K. & West, S. C. Unresolved recombination intermediates lead to ultra-fine anaphase bridges, chromosome breaks and aberrations. *Nat. Cell Biol.* 20, 92–103 (2018).

27. Tiwari, A., Addis Jones, O. & Chan, K.-L. 53BP1 can limit sister-chromatid rupture and rearrangements driven by a distinct ultrafine DNA bridging-breakage process. *Nat. Commun.* 9, 677 (2018).

28. Longtine, M. S. *et al.* Additional modules for versatile and economical PCR-based gene deletion and modification in Saccharomyces cerevisiae. *Yeast*
14, 953–961 (1998).

29. Anand, R., Memisoglu, G. & Haber, J. Cas9-mediated gene editing in Saccharomyces cerevisiae. Protoc. Exch. (2017). doi:10.1038/protex.2017.021a

30. Ferrari, M., Twayana, S., Marini, F. & Pellicoli, A. A qPCR-Based Protocol to Quantify DSB Resection. in Methods in Molecular Biology 1672, 119–129 (2018).

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Author contributions

M.F. performed experiments in Fig. 1, 2, 3, 4 and Expanded data Fig. 1, 2, 3.

C.C.R. performed experiments in Fig 1b, c, and contributed to Fig. 2 and Fig. 3.

S.L. contributed to the ChIP analyses in Fig 2c and Fig. 3g, h, and Expanded data Fig. 1a.

A.P. supervised and coordinated all aspects of the work.

M.F., C.C.R. and A.P. wrote the manuscript.
**Competing interests**

The authors declare no competing interests.

**Figure legends**

**Fig. 1: Rad9 promotes crossover recombination DSB repair.**

a, scheme of the genetic system to test recombination events in LSY2205-11C/LSY2543 background.

b, Distribution of the recombinant colony types in the indicated LSY2205-11C and LSY2543 derivative diploid strains, with SEM of three experiments. Red (two long track conversions), White (two short track conversions) and Red/White (one long and one short track conversion) were determined from recombinant colonies after I-SceI induction.

c, Distribution of CO, NCO and BIR among all the colony types of the experiments in (b), with SEM of two independent diploids.

d, scheme of the genetic system to test BIR in YRL92 background.

e, Southern blot of *AvpI*-digested DNA to monitor DSB repair through BIR in the indicated YRL92 strains.

f, Densitometric analysis of the BIR band of three experiments as in (e), with SEM.

g, BIR efficiency measured by cell viability in the indicated YRL92 strains, with SEM of three experiments.

**Fig. 2: Rad9 limits Rpa1, Rad52 and Rad51 hyper-loading at the DSB site, and favours Rad51 binding at the donor site.**
ChIP analysis at the indicated positions near the cut site, after 3 hours since DSB induction, for the binding of Rpa1-Myc (a), Rad52-HA (b) and Rad51 (c), with SEM of three experiments. The indicated JKM strains were blocked in G2/M with nocodazole. The same experiment as in Extended data Fig. 2.

d, Rad51 enrichment on donor site at Chromosome XI, 3h and 6h after DSB on Chromosome V in the indicated JRL92 strains, with SEM of three experiments. Cells were blocked in G2/M with nocodazole.

**Fig. 3: Rad9 promotes D-loop extension in BIR by limiting Sgs1 and Mph1.**

a, scheme of the PCR-based assay to test D-loop extension in JRL92 background.
b, PCR-based primer extension assay to test D-loop in the indicated JRL92 strains.
c, Quantification of the D-loop extension in three experiments as in (c), with SEM.
d, BIR efficiency measured by cell viability in the indicated JRL92 strains, with SEM of three experiments.
e, as in (b), with the indicated JRL92 strains.
f, Quantification of the D-loop extension as in (e), with SEM of three experiments.
g, Sgs1 binding at the DSB site in the indicated JKM strains blocked in G2/M with nocodazole, with SEM of three experiments.
h, Mph1 binding at the DSB site in the indicated JKM strains blocked in G2/M with nocodazole, with SEM of three experiments.
i, as in (d).

**Fig. 4: Rad9 promotes recombination events that require stable D-loop.**
a, scheme of the genetic system to test ectopic gene conversion (eGC) in tGI354 background.

b, Southern blot of EcoRI-digested DNA to monitor DSB repair through eGC in the indicated tGI354 strains. CO, Crossover; NCO, Noncrossover.

c, Densitometric analysis of the CO and NCO bands of three experiments as in (b), with SEM.

d, working model: Rad9 controls HR, coordinating resection and strand rejection steps. After a DSB, Rad9 binding at the lesion limits: i) the Sgs1-Dna2 axis for the long-range resection and SSA; ii) the Sgs1-Top3-Rmi1 and Mph1 axes for the strand-rejection, favouring long-lived D loop and complex HR sub-pathways.

Extended data Fig. 1: Rad9 limits Rpa1, Rad52 and Rad51 hyper-loading at the DSB site.

ChIP analysis at the indicated positions near the cut site, 4.5 hours after DSB induction, for the binding of Rad51 (a), Rad52-HA (b) and Rpa1-Myc (c), with SEM of three experiments. The indicated JKM strains were blocked in G2/M with nocodazole. The same experiment as in Fig 2.

Extended data Fig. 2: rad9Δ and sgs1-FD mutations do not alter DSB resection closed to the DSB.

DSB resection analysis at 0.15 kb (a), 1.4 kb (b) and 10 Kb (c) from the DSB at the indicated times after HO induction, with SEM of three experiments. The indicated JKM strains were blocked in G2/M with nocodazole.
Extended data Fig. 3: *rad9*-2Ala and *rad9*-7xA alleles partially rescue BIR of *rad9Δ* cells.

BIR efficiency measured by cell viability in the indicated YRL92 strains, with SEM of two experiments.

Extended data Table 1: Distribution of colonies related to Fig. 1c.

Extended data Table 2: List of the yeast strains used in this study.

Extended data Table 3: List of the oligonucleotides used in this study.
all the colonies were screened to classify recombination events
**Fig. 2**

(a) **Rpa1 ChIP at 3 hours from DSB**

(b) **Rad52 ChIP at 3 hours from DSB**

(c) **Rad51 ChIP at 3 hours from DSB**

(d) **Rad51 ChIP on the donor site**

*p*=0.019

*p*=0.011
Fig. 4

(a) Recombination products

(b) Loading control

Parental

CO

NCO

(b) Hours after DSB + nocodazole

0 3 6 9 12 24 0 3 6 9 12 24 0 3 6 9 12 24 0 3 6 9 12 24

Wild type

rad9Δ

mph1Δ sgs1Δ

rad9Δ

(c) % of DSB repair

0 10 20 30 40 50 60 70 80 90

wild type

rad9Δ

mph1Δ sgs1Δ

mph1Δ sgs1Δ

rad9Δ

(d) Complex HR w/CO

(BIR, long-track GC)

long-lived D-loop

Rad9

RPA

Rad51

DNA synthesis

Rad9 reduction/depletion

(Sgs1-Top3-Rmi1)

(Mph1)

strand rejection

Rad9 reduction/depletion

(Sgs1-Dna2)

long-range resection

SSA

long-range resection

SSA

(d) Complex HR w/CO

(BIR, long-track GC)

long-lived D-loop

Rad9

RPA

Rad51

DNA synthesis

Rad9 reduction/depletion

(Sgs1-Top3-Rmi1)

(Mph1)

strand rejection

Rad9 reduction/depletion

(Sgs1-Dna2)

long-range resection

SSA
Extended data Fig. 1

(a) Rad51 ChIP at 4.5 hours from DSB

(b) Rad52 ChIP at 4.5 hours from DSB

(c) Rpa1 ChIP at 4.5 hours from DSB
% of viability after DSB formation

wild type  rad9+TxA  rad9-2Axa  rad9Δ

p=0.001
p=0.042
p=0.003

p=0.003
Extended data Fig. 1

(a) Rad51 ChIP at 4.5 hours from DSB

(b) Rad52 ChIP at 4.5 hours from DSB

(c) Rpa1 ChIP at 4.5 hours from DSB
Extended data Fig. 2

0.15 Kb from DSB site

% of ssDNA vs. hours after DSB

1.4 Kb from DSB site

% of ssDNA vs. hours after DSB

10 Kb from DSB site

% of ssDNA vs. hours after DSB

Legend:
- wild type
- rad9Δ
- sgs1Δ
- sgs1-FD
- sgs1Δ rad9Δ
- sgs1-FD rad9Δ
Extended data Fig. 3

% of viability after DSB formation

- wild type
- rad9-7xA
- rad9-2AxA
- rad9Δ

$p=0.001$
$p=0.042$
$p=0.003$
### Extended data Table 1: Distribution of colonies related to Fig. 1b:

|                  | Diploid 1   | Diploid 1   | Diploid 2   | Diploid 2   |
|------------------|-------------|-------------|-------------|-------------|
|                  | WT          | *rad9Δ*     | WT          | *rad9Δ*     |
| White/Red sectors| BIR         | 4           | 3           | 5           | 4           |
|                  | BIR-like    | 1           | 0           | 4           | 0           |
|                  | CO          | 6           | 2           | 6           | 4           |
|                  | Unclassified LOH | 0     | 0           | 0           | 0           |
|                  | NCO         | 40          | 18          | 68          | 13          |
| Red colonies (Recombinants) | BIR         | 0           | 0           | 3           | 1           |
|                  | BIR-like    | 5           | 0           | 0           | 0           |
|                  | CO          | 7           | 2           | 6           | 2           |
|                  | Unclassified LOH | 1     | 13          | 3           | 8           |
|                  | NCO         | 105         | 35          | 64          | 31          |
| White colonies | BIR         | 0           | 0           | 0           | 0           |
|                  | BIR-like    | 0           | 1           | 0           | 0           |
|                  | CO          | 3           | 1           | 5           | 0           |
|                  | Unclassified LOH | 13   | 8           | 10          | 11          |
|                  | NCO         | 91          | 171         | 72          | 150         |

Red (Non recombinants)  | 12           | 5           | 7           | 5           |

|                  | Total colonies | 288         | 259         | 253         | 229         |
|                  | Total recombinants | 276 | 254         | 246         | 224         |
Extended data Table 2. List of yeast strains described in this work.

| Strain name | Parental strain | Genotype | Source |
|-------------|-----------------|----------|--------|
| Y1600       | JKM17           | MATa ho hml::ADE1 hmr::ADE1 ade1-100 leu2-3, 112 lys5, trpl::hisG ura3-52 lys3::ade3::GAL10::HO | Lee et al., 1998 |
| Y603        | tGI354          | ho hml::ADE1 MATa-inc hmr::ADE1 ade1 leu2-3,112 lys5 trpl::hisG ura3-52 ade3::GAL10::HO (arg5,6::MATa::HPH) | Ira et al., 2003 |
| Y3172       | JRL092          | ho mat::hisG hml::hisG HMRa-stk ura3A851 trplA63 leu2A::KAN ade3::GAL10::HO can1,1-1446::HOC::HPH ykl215c::LEU2::can1A289 | Lydeard et al., 2007 |
| Y3480       | JRL092          | ho mat::hisG hml::hisG HMRa-stk ura3A851 trplA63 leu2A::NAT ade3::GAL10::HO can1,1-1446::HOC::HPH ykl215c::LEU2::can1A289 | This study |
| Y4197       | Y3480           | rad53::K227A::KANMX6@RAD53 chkl::TRP1 | This study |
| Y3513       | Y3480           | rad53::K227A::KANMX6@RAD53 chkl::TRP1 | This study |
| Y3308       | Y3172           | pif1::TRP1 | This study |
| Y3387       | Y3172           | sgs1::NAT | This study |
| Y3475       | Y3172           | mph1::NAT | This study |
| Y3981       | Y3172           | mph1::NAT sgs1::URA3 | This study |
| Y3454       | Y3172           | sgs1::NAT rad9::TRP1 | This study |
| Y3488       | Y3172           | mph1::NAT rad9::TRP1 | This study |
| Y3982       | Y3172           | sgs1::URA3 mph1::NAT rad9::TRP1 | This study |
| Y4377       | Y3480           | sgs1::F1192D rad9::TRP1 mph1::KANMX6 | This study |
| Y4351       | Y3480           | sgs1::F1192D rad9::TRP1 | This study |
| Y4314       | Y3480           | sgs1::F1192D | This study |
| Y4302       | Y3172           | rad9::NAT rad9-3xHA::TRP1 | This study |
| Y4312       | Y3172           | rad9::NAT rad9-2Ala-3xHA::TRP1 | This study |
| Y4304       | Y3172           | rad9::NAT rad9-7xA-3xHA::TRP1 | This study |
| Y4279       | Y1600           | MATa RAD9::HPH SGS1-13xMYC::KANMX6 | This study |
| Y4046       | Y1600           | MATa SGS1::13xMYC::KANMX6 | This study |
| Y4283       | Y1600           | MATa RPA1::13xMYC::KANMX6 | This study |
| Y4289       | Y1600           | MATa RPA1::13xMYC::KANMX6 RAD9::HPH | This study |
| Y3892       | Y1600           | MATa RAD52::3xHA::KANMX6 | This study |
| Y3890       | Y1600           | MATa RAD52::3xHA::KANMX6 rad9::HPH | This study |
| Y4332       | Y1600           | MATa sgs1::F1192D | This study |
| Y4488       | Y1600           | MATa sgs1::F1192D rad9::HPH | This study |
| Y2995       | Y603            | rad9::NAT | This study |
| Y4375       | Y603            | sgs1::KANMX6 mph1::TRP1 | This study |
| Y4429       | Y603            | rad9::NAT sgs1::KANMX6 mph1::TRP1 | This study |
| Y3412       | LSY2205-11C      | MAT alpha, ade2-I lys2::GAL-I-SCEI his3::HPHMX4 | Ho et al. 2010 |
| Y3414   | LSY2543 | \textit{MAT a, met22::KTRP1 ade2-n his3::NATMX4} | Ho et al. 2010 |
|---------|---------|-------------------------------------------------|----------------|
| Y3831   | Y3412   | \textit{rad9::KANMX6}                            | This study     |
| Y3849   | Y3414   | \textit{rad9::KANMX6}                            | This study     |
**Extended data Table 3.**

**List of the oligonucleotides used for ChIP and DSB resection analyses.**

| Name  | Sequence (5’-3’)                             | Distance from DSB | Source                  |
|-------|----------------------------------------------|-------------------|-------------------------|
| QMAT1F| CCTGGTTTTTGGTTTTGTAGAGTGG                    | 0.15 kb (Chr III) | Kim et al. 2007         |
| QMAT1R| GAGCAAGACGATGGGAGTTTC                       | 0.15 kb (Chr III) | Kim et al. 2007         |
| QMAT1.4F|                                    | 1.4 kb (Chr III) | This study              |
| QMAT1.4R|                                   | 1.4 kb (Chr III) | This study              |
| QMAT2.F| ATTGCGACAAGGCTTCACCC                      | 4.8 kb (Chr III)  | Kim et al. 2007         |
| QMAT2.R| CCCATCACAGGTTTTATGGTTCC                    | 4.8 kb (Chr III)  | Kim et al. 2007         |
| QMAT3.1.F| GGGTTCAACACTGCCACCTTTG                     | 10 kb (Chr III)   | Dibitetto et al. 2016   |
| QMAT3.1.R| GGTGATGGAGATGGAGTAGGAACG                    | 10 kb (Chr III)   | Dibitetto et al. 2016   |
| QMAT4F| GCTCTTCTAGCGAACAACACG                      | 15 kb (Chr III)   | Kim et al. 2007         |
| QMAT4R| GCAATAACCCACGGAACACTG                      | 15 kb (Chr III)   | Kim et al. 2007         |
| QKCC4F| TCGTATCGGTGCTGCTATGAA                      | Chr III            | This study              |
| QKCC4R| CTCTCGAAAATTTGGCTGTCATTG                   | Chr III            | This study              |
| QCan1F| CAATGGTGTTAGCTGGCTGCTGCC                   | Chr XI (JRL92 strain) | This study            |
| QCan1R| TGGCTCTACTACAGTACAACCTCG                   | Chr XI (JRL92 strain) | This study            |

**List of the oligonucleotides used for the primer extension assay in the JRL092 strains.**

| Name | Sequence (5’-3’) | Position | Source                  |
|------|------------------|----------|-------------------------|
| P1   | GAGGATACGTTCCTATGGAG | CAN1 on Chr V | This study              |
| P2   | GTCTTTGCTATGATCTTCCC | CAN1 on Chr XI | This study              |
| P3   | CTACCTCTCCTAAAGCCTTCG | TLC1 on Chr II | This study              |
| P4   | TGGTCTATCAGTAACACCTCG | TLC1 on Chr II | This study              |