SIRPγ-CD47 Interaction Positively Regulates the Activation of Human T Cells in Situation of Chronic Stimulation

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A numerous number of positive and negative signals via various molecules modulate T-cell activation. Within the various transmembrane proteins, SIRPγ is of interest since it is not expressed in rodents. SIRPγ interaction with CD47 is reevaluated in this study. Indeed, we show that the anti-SIRPγ mAb clone LSB2.20 previously used by others has not been appropriately characterized. We reveal that the anti-SIRPα clone KWAR23 is a Pan anti-SIRP mAb which efficiently blocks SIRPα and SIRPγ interactions with CD47. We show that SIRPγ expression on T cells varies with their differentiation and while being expressed on Tregs, is not implicated in their suppressive functions. SIRPγ spatial reorganization at the immune synapse is independent of its interaction with CD47. In vitro SIRPα-γ/CD47 blockade with KWAR23 impairs IFN-γ secretion by chronically activated T cells. In vivo in a xeno-GvHD model in NSG mice, the SIRPγ/CD47 blockade with the KWAR23 significantly delays the onset of the xeno-GvHD and deeply impairs human chimerism. In conclusion, we have shown that T-cell interaction with CD47 is of importance notably in chronic stimulation.

Keywords: graft-versus-host disease, SIRPA, chronic stimulation, T cell, CD47, SIRPG

INTRODUCTION

T cells are adaptive immune cells that are educated in the thymus to defend the organism and eliminate pathogens efficiently by recognizing the MHC-antigen complex via their TCR. Mature T cells express a variety of receptors which engagement will stabilize, enhance, or decrease their activation upon antigen recognition. In this study, we focused on SIRPγ, a T-cell restricted surface molecule from the signal-regulatory protein (SIRP) family. SIRP family comprise transmembrane glycoproteins expressed on immune cells and central nervous system (1–3). Three main members have been described: SIRPα, SIRPβ, and SIRPγ with homologous extracellular immunoglobulin (Ig)-like domains but distinct transmembrane and intracytoplasmic domains (4–8). Briefly, SIRPα (CD172a) is expressed on hematopoietic progenitors, myeloid cells, dendritic cells, NK cells, and...
neurons (4, 9). SIRPα cytoplasmic tail contains immunoreceptor tyrosine-based inhibition motifs (ITIMs) (10) resulting in inhibitory signals. SIRPα ligates CD47, an ubiquitously expressed molecule, and this interaction is responsible of the “don’t eat me signal” which leads to the inhibition of phagocytosis by myeloid cells (11). SIRPβ (CD172b) associates with DAP12 (12, 13) which contains an immunoreceptor tyrosine-based activation motif (ITAM). Yet, ligand of SIRPβ is still unknown. Finally, SIRPγ (CD172g) has been described recently (14) probably because the gene of SIRPγ is absent in rodents and found only in humans and primates (15). SIRPγ has a very short intracytoplasmic tail of 4-amino acids (aa) incapable of transducing signals on its own. SIRPγ also interacts with CD47 as SIRPα (14–16), but the SIRPα/CD47 affinity (Kd 2 µM) is 10 times stronger than the one of SIRPγ/CD47 (Kd 23 µM) (17).

It has been shown that the interaction between SIRPγ and CD47 plays a key role in T-cell transendothelial migration under shear flow conditions (17) in cell-cell adhesion and promotes antigen-specific T-cell proliferation and costimulation (16). However, these observations rely on the use of the anti-SIRPγ clone LS62.20 which we report in this study as not being able to efficiently block the SIRPγ/CD47 interaction at the concentration tested. Thus, we reevaluate the impact of SIRPγ in human T-cell biology using a different anti-SIRP monoclonal antibody (clone KWAR23), which we characterize here as capable to efficiently block SIRPγ and SIRPα interactions with CD47. We show that SIRPγ expression varies on CD4 and CD8 T-cell subpopulations and, while being expressed by regulatory T cells (Tregs), we demonstrate that SIRPγ/CD47 interaction is not implicated in their suppressive function. Upon antigen presenting cell (APC) recognition, SIRPγ clusters at the immune synapse independently of its ligation to CD47 on the APC. IFN-γ secretion by chronically activated T cells is impaired by SIRPγ/CD47 interaction blockade in vitro. Finally, when used in vivo in a T-cell-dependent xeno-GvHD model mediated by the injection of human PBMC in NSG-reipient mice, the anti-SIRPα-γ mAb KWAR23 impairs human cell engraftment, leading to a delayed GvHD onset. Importantly, all CD4 and CD8 T-cell compartments were compromised in KWAR23-treated mice. In conclusion, we have shown that T-cell interaction with CD47 is of importance notably in chronic stimulation.

MATERIALS AND METHODS

Reagents

All the reagents used in the study are listed in the Table 1.

Cells

Human peripheral blood mononuclear cells (PBMC) were isolated from fresh cytopheresis ring from healthy donors obtained from the Etablissement Français du Sang (Nantes, France). Written informed consent was provided according to institutional guidelines. Human T cells were isolated by negative selection with the Miltenyi’s Pan T-cell isolation kit according to the manufacturer’s protocol. Jurkat T cells (ATCC, clone E6-1) were cultured at 10⁵ cells/ml in complete RPMI medium (10% GE Healthcare HyClone™ Fetal Bovine Serum (Chicago, IL, USA), 1% penicillin-streptomycin Gibco® (Waltham, MA, USA), 1% glutamine, 1% sodium pyruvate Gibco®, 1% Hepes Gibco®, and 1% MEM NEAA Gibco®) at 37°C, 5% CO₂.

Development of KO and KI Jurkat

The GeneArt CRISPR Nuclease Vector kit (Life Technologies, Carlsbad, CA, USA) was used. Guide RNAs for SIRPγ and CD47 were selected thanks to the CRISPOR website. The following were the sRNA selected: sgRNA anti-SIRPγ #1 TCCCCGTG GAGCGTGTCCTG TGG, sgRNA anti-SIRPγ #2 TGTAT GTGCCGCCACATCTGC TGG, sgRNA anti-CD47 #1 CTTGTTTAGAGCTCCATCAA AGG, and sgRNA anti-CD47 #2 TCCATGCTTTGTTACTAATA TGG; PAM sequence is indicated in italic. Briefly, Jurkat cells suspended in Opti-MEM (Gibco) medium at 10⁶ cells/ml were electroporated with a NEPA21 (Nepalgene) and placed immediately in warm complete RPMI medium in six-well plates and incubated at 37°C, 5% CO₂. After amplification, cells were stained using anti-SIRPγ mAb and/or anti-CD47 mAb for FACS sort with the ARIA II (BD Biosiences, Franklin Lakes, NJ, USA) depending on the nonexpression of the corresponding epitopes: SIRPγ, CD47. Sorted cells were amplified for ulterior use. Genomic DNAs were extracted, and PCR amplified with the following primers SIRPγ F1 5’-GCCTCACTGT CCCTCAATTGT-3’; SIRPγ R1 5’-GGGATGAGGGAGGT CCATGTT-3’; SIRPγ F2 5’-TGGTGACCCAGTCATCTGAAATA-3’; SIRPγ R2 5’-GGGTTGACAAACAGTTTGA-3’; CD47 F1 5’- CTCTAAAGCCTTTCAAAGCAGCA-3’; CD47 R1 5’-AGGAGTACAGGTTGCACCA-3’; CD47 F2 5’-ACTACACC TGCATGTTCCA-3’; and CD47 R2 5’-CAGGTTGCAC AGCAAAAT-3’.

Jurkat SIRPγ-KO cells were then transduced with lentivirus encoding OST-SIRPγ. SIRPγ expression on transduced cells was higher than in their WT counterpart. SIRPγ-high cells were FACS sorted and amplified for ulterior use.

ELISA-Binding Assay

For human SIRPα-binding assay, recombinant human SIRPα was immobilized on plastic at 0.5 µg/ml (SIRPα) in carbonate buffer (pH 9.2). After saturation, purified antibodies were added in range (from initial concentration at 10 µg/ml) to measure binding. After incubation and washing, peroxidase-labeled donkey antihuman IgG was added and revealed by conventional methods.

For human SIRPγ-binding assay, purified antibodies were captured in range (from initial concentration at 5 µg/ml) with a coated donkey antihuman IgG (H+L) immobilized at 2 µg/ml in borate buffer. After washing, biotinylated SIRPγ (#11828-H08H, SinoBiological, Beijing, China, biotinylation performed by OSE Immunotherapeutics, Nantes, France) was added at 1 µg/ml and detected by streptavidin peroxidase, then revealed by conventional methods.

Human SIRPγ-Binding Assay on Human Lymphocyte and Monocyte by Flow Cytometry

PBMC, isolated by Ficoll from blood sample, were incubated with human Fc receptor-binding inhibitor diluted at 1/50.
### TABLE 1 | Key resource table.

| Reagent type       | Designation | Source                          | Reference                          | Host and subclass | Clone name |
|--------------------|-------------|---------------------------------|------------------------------------|-------------------|------------|
| Antibody           | Antihuman CD3 | BD Biosciences                  | 557943, 555332, 583725             | Mouse IgG1, κ     | UCHT1      |
| Antibody           | Antihuman CD4 | BD Biosciences                  | 562424, 560158, 55546, 560788      | Mouse IgG1, κ     | RPA-T4     |
| Antibody           | Antihuman CD4 | BD Biosciences                  | 557822                             | Mouse IgG1, κ     | SK3        |
| Antibody           | Antihuman CD8 | BD Biosciences                  | 564116                             | Mouse IgG1, κ     | SK1        |
| Antibody           | Antihuman CD14 | BD Biosciences                  | 558121                             | Mouse IgG2b, κ    | MSE2       |
| Antibody           | Antihuman CD45RA | BD Biosciences                  | 550855, 563870, 550994              | Mouse IgG2b, κ    | HI100      |
| Antibody           | Antihuman CD45 | BD Biosciences                  | 563792                             | Mouse IgG1, κ     | HI30       |
| Antibody           | Antihuman CD27 | BD Biosciences                  | 564301                             | Mouse IgG1        | L128       |
| Antibody           | Antihuman CD279 (PD1) | BioLegend, San Diego, CA, USA  | 329918                             | Mouse IgG1, κ     | EH12.2H7   |
| Antibody           | Antihuman CCR7 | BD Biosciences                  | 557648                             | Mouse IgG1, κ     | 3D12       |
| Antibody           | Antihuman CD95 | BD Biosciences                  | 559773                             | Mouse IgG1, κ     | DX2        |
| Antibody           | Antihuman CD28 | BD Biosciences                  | 562296                             | Mouse IgG1, κ     | CD28.2     |
| Antibody           | Antihuman CD47 | BD Biosciences                  | 560371                             | Mouse IgG1, κ     | B6H12      |
| Antibody           | Antihuman CD25 | BD Biosciences                  | 555431, 563701                      | Mouse IgG1, κ     | M-A251     |
| Antibody           | Antihuman CD127 | BD Biosciences                  | 557938, 562662                      | Mouse IgG1, κ     | HIL-7R-M21 |
| Antibody           | Antihuman CD127 | BioLegend                       | 351318                             | Mouse IgG1, κ     | A019D5     |
| Antibody           | Antihuman SIRP | BioLegend                       | 336606                             | Mouse IgG1, κ     | LSB2.20    |
| Antibody           | Antihuman CD47 | R&D Systems, Minneapolis, MN, USA | FAB4670G                          | Mouse IgG1, κ     | 472603     |
| Antibody           | Antihuman Foxp3 | eBioscience, San Diego, CA, USA | 45-4776-42                         | Rat IgG2a, κ      | PCH101     |
| Antibody           | Antimouse CD45 | BD Biosciences                  | 550994                             | Rat IgG2a, κ      | 30-F11     |
| Antibody           | Antimouse SIRPα | BD Biosciences                  | 740285                             | Rat IgG1, κ       | P84        |
| Antibody           | Antimouse CD11b | BD Biosciences                  | 552850                             | Rat IgG2b, κ      | M1/70      |
| Antibody           | Antimouse Cε | BioLegend                       | 410708, 409004                      | Rat IgG2a, κ      | M1310G05   |
| Antibody           | Antimouse CD3 | In house                        | Hybridoma                          | Mouse IgG2a, κ    | OKT3       |
| Antibody           | Pan antihuman SIRP | Produced by OSE Immunotherapeutics | WO2015138600A2                     | Chimeric human-IgG4 | Kwar23    |
| Antibody           | Antihuman SIRPα | OSE Immunotherapeutics         |                                   | Mouse IgG         | 18D5       |
| Antibody           | Antihuman CD47 | BioLegend                       |                                   | Mouse IgG1, κ     | B6H12      |
| Antibody           | Antimouse IgG (H+L) | Jackson Immunoresearch, West Grove, PA, USA | 709-005-098                     | Donkey            |
| Antibody           | Antimouse IgG | Jackson Immunoresearch          | 709-035-149                        | Donkey            |
| Antibody           | Antimouse IgG Fc | BioLegend                       | 409303                             |                 |
| Recombinant protein | sSIRPα | SinoBiological                  | 11612-H08H                         | Human              |
| Recombinant protein | SIRPα | SinoBiological                  | 11828-H08H                         | Human              |
| Recombinant protein | CD47 | SinoBiological                  | 12283-H02H                         | Human              |
| Recombinant protein | CD47 | AcroBiosystems, Newark, DE, USA | CD7-H82F6                         | Human              |
| Commercial kit      | Pan T-cell isolation kit SEE | Jackson Immunoresearch, West Grove, PA, USA | 130-096-535                     |                 |
| Toxin               | Human Fc receptor-binding inhibitor | BD Pharmingen, San Diego, CA, USA | 564220                            |                   |
| Toxin               | Human IFNγ ELISA set | BD Biosciences                  | 555142                             |                   |
| Toxin               | Alamur Blue | Invitrogen, Waltham, MA, USA    | DAL1100                            |                   |
| Toxin               | Streptavidin peroxidase | Jackson Immunoresearch          | 016-030-084                        |                   |
| Toxin               | Cell Proliferation Dye eFluor™ 450 | eBioscience™                   | 65-0842-85                        |                   |
| Toxin               | Cell Proliferation Dye eFluor™ 670 | eBioscience™                   | 65-0840-85                        |                   |
| Cell tracker        | Cell Tracker™ Green CMFDA Dye | ThermoFisher, Waltham, MA, USA   | C7025                              |                   |
| Reagent             | Poly-L-lysine | Sigma                          | P4832                              |                   |
| Microslides         | iBIDI 8 wells | iBIDI                          | 80626                              |                   |
Then, antibodies were incubated for 30 min at 4°C and washed before being stained 30 min at 4°C with PE-labeled anti-human IgG Fc and with a mix of 50-fold diluted antibodies to determine cell subpopulations (CD3+ for T lymphocyte and CD14+ for monocyte). Samples were analyzed on CytoFlex cytfluorometer (Beckman Coulter France, Villepinte, France) after gating on different subpopulations.

**Human SIRPγ and Human SIRPα Antagonist Activity Measured by ELISA**

For antagonist activity, enzyme-linked immunosorbent assay (ELISA) assay, recombinant human SIRPγ, or recombinant human SIRPα were immobilized on plastic at respectively 2 µg/ml (SIRPγ) and 0.5 µg/ml (SIRPα) in carbonate buffer (pH 9.2). During saturation, biotinylated human CD47 were preincubated at a unique concentration (final concentration at 3 µg/ml) with purified antibodies in range at room temperature for 15 min. Preincubated (CD47-biot/purified antibodies) mixes were then incubated on immobilized SIRPγ (O/N at room temperature) or on immobilized SIRPα (for 2 h at 37°C). After incubation and washing, streptavidin-peroxidase was added and revealed by conventional methods.

**Flow Cytometry Assays**

PBMC (10⁵ per well) were suspended in 200 µl of FACS buffer (phosphate-buffered saline 1x, 1% bovine serum albumin, Sigma-Aldrich, 0.1% Azide Sigma-Aldrich, St. Louis, MO, USA) and plated in a V-bottom 96-well plate prior to being centrifuged at 2,500 rpm for 1 min. For the extracellular staining, cells were labeled with a mix of antibodies for 30 min on ice. For the intracellular labeling, BD Cytotoxic/Cytoperm kit was used according to the manufacturer’s protocol with recommended volume of intracellular antibodies. Cells were washed twice before analysis on the LSRII (BD Biosciences). After the acquisition, data were analyzed using FlowJo software.

**Coculture of Tregs and Effector T Cells**

Purified T cells were stained with anti-CD4, anti-CD25, and anti-CD127 for 30 min at 4°C in the dark, cells were washed, and DAPI was used to discriminate dead cells. Cells were then sorted using the ARIA II (BD Biosciences). Tregs and effector T cells were further stained with two different cell proliferation dyes (CPD) for 10 min at 37°C and cultured either alone (control conditions) or together, in U-96-well costar plates previously coated or not with anti-CD3 (clone OKT3) at 2 µg/ml. When indicated, soluble anti-SIRPα/γ (KWAR23) antibody was added at 10 µg. After 5 days in culture, T-cell proliferation was assessed by flow cytometry using the LSRII (BD Biosciences).

**T-Cell Activation**

T cells were stained with CPD prior to be activated in plates coated with 0.5 µg/ml of anti-CD3 (OKT3) and 2–3 µg/ml of soluble anti-CD28.2. Cells were incubated 3 or 5 days at 37°C, 5% CO₂, then assessed by flow cytometry. In some experiments, T cells were stimulated with 0.5–1 µg/ml of coated OKT3 with or without coated CD47-Ig (10 µg/ml).

**Chronically Activated T Cells**

PBMC at 7.5 x 10⁶ were stimulated in complete medium three times in six-well plates coated with 3 µg/ml of anti-CD3 (OKT3) and 3 µg/ml of soluble anti-CD28.2. Each stimulation is consist of a 48–72-h culture in six-well plates coated with 3 µg/ml of anti-CD3 (OKT3) and 3 µg/ml of soluble anti-CD28.2. Each time, cells were washed and counted prior to be restimulated. Chronically stimulated PBMC were then cultured for 48 h in plates coated with 0.5 µg/ml of anti-CD3 (OKT3) and 10 µg/ml of human CD47-Fc recombinant protein in the presence or not of mAbs targeting the SIRP/CD47 interactions. Supernatants were collected after 48 h of stimulation with OKT3 and CD47-Fc to measure IFN-γ secretion, and cell fitness was analyzed by Alamar Blue detection at 530 nm after 5 days stimulation.

**Confocal Microscopy**

Jurkat, resting, or activated T cells were stained for 30 min at 4°C in the dark with an anti-SIRPγ (LSB2.20) and an anti-CD47 mAb. Cells were fixed with PFA at 4% for 10 min at room temperature (RT), permeabilized with PBS + 0.1% Triton for 5 min at RT, and further stained with DAPI for 5 min at RT in the dark. ProLong was added to the cells and the solution carefully placed between a glass slide and a 12-µM round coverslip and left for at least 48 h polymerization before being analyzed with confocal microscopy (Nikon Super Resolution SIM Confocal A1) using the NIS software. The images obtained were analyzed using ImageJ software.

**Time-Lapse Microscopy**

IBIDI (Gräfelfing, Germany) 8-well slides were coated with poly-1-lysine at 0.001%. Raji cells were primed with 10 ng/ml of SEE for 30 min at 37°C, and WT or SIRPγ<sup>high</sup> Jurkat cells were stained with a cell tracker green for 15 min at 37°C, washed thoroughly then stained with an anti-SIRPγ mAb (clone LSB2.20) for 30 min at 4°C. Cells were then washed, and Raji and Jurkat cells were deposited in the IBIDI slide. Once the cells are stabilized, the time lapse video starts. To assess the role of SIRPγ in the synapse formation, SIRPγ/CD47 interaction was blocked on SIRPγ<sup>high</sup> cells for 15 min at 37°C with the KWAR23 mAb at 10 µg/ml, then mixed with Raji for 15 min prior to be plated in IBIDI wells. SIRPγ polarization into the synapse was analyzed using ImageJ software. Briefly, a ROI was drawn on the whole cell surface and analyzed with colo2 plugin.

**Animals**

We used a xenot-GvHD model of immunodeficient mice to assess the role of SIRPγ in GvHD occurrence. NSG mice were
pur chased from the Jackson Laboratory (Bar Harbor, ME) and bred by the LabEx IGO humanized rodent platform. Mice (males and females) were used between 8 and 12 weeks old. All animals were housed under specific pathogen-free conditions, according to institutional guidelines. NSG mice were irradiated at 1.5 Gy on Day 0 prior to an i.v. injection of 10^7 human PBMC from healthy volunteers on Day 0. Mice were treated 2 times/week with PBS or anti-SIRPα-γ (KWAR23 mAb) at 5 mg/kg from D1 to D42. Human cell engraftment and weight loss were monitored during the experiment. If losing more than 20% of its maximum body weight, the mice were sacrificed. Human chimerism analyzed by flow cytometry is calculated as follows: (% of hCD45)/(% of hCD45 + % of mCD45) * 100. The relative number of CD4/CD8 T cells in the spleen is calculated as follows: (% human CD4/CD8 in spleen) * (% human CD4/CD8 in spleen). This study was carried out according to authorization from the French Ministry of Research, APAFIS # 4678. Intrapitoneal residual macrophages were collected as previously described in [Ray:2010cp]. Fc block was added to the cells following manufacture recommendation prior the incubation with the antibodies.

RESULTS
The KWAR23 Is a Pan Anti-SIRP mAb, Which Efficiently Inhibits SIRPγ-CD47 Interaction
KWAR23 mAb is a recombinant antibody which has been selected after immunization with a SIRPα recombinant molecule by Ring and collaborators (18). Given the homology of 79% when comparing the extracellular portions of the SIRPα, SIRPγ, and SIRPβ (15), we hypothesized that KWAR23 could also recognize other SIRP protein family members. We directly addressed this question by labelling human cells which are known to either uniquely express SIRPα, i.e., monocytes or described to express SIRPγ, i.e., T lymphocytes. KWAR23 efficiently binds 100% of monocytes when the concentration used is sufficient to do the anti-SIRPα-specific mAb (Figures 1A, B, left panels); similar results were obtained when U937 (SIRPα− cell line) were stained (data not shown). Interestingly, increased concentration of KWAR23 resulted in T-cell staining which reached a plateau when 60% of human T cells were stained. No binding was seen with the anti-SIRPα-specific mAb (clone 18D5), suggesting that KWAR23 recognizes a molecule other than SIRPα, presumably SIRPα on T cells (Figures 1A, B, right panels). Furthermore, Jurkat WT and Jurkat SIRPγ cells were stained with KWAR23 and LSB2.20, a specific anti-SIRPγ developed by Piccio and colleagues (16). While both Abs similarly stained Jurkat WT cells, no staining was observed on Jurkat SIRPγKO cells with either Ab, identifying SIRPγ as the sole KWAR23 epitope on Jurkat cells (Figure 1C). To confirm these results, we setup two ELISA in which recombinant SIRP molecules were used. The anti-SIRPα-specific mAb recognized SIRPα but not SIRPγ-recombinant protein, while the KWAR23 recognized SIRPα, SIRPγ, and even SIRPβ (Figure 1D and data not shown for SIRPβ). Thus, KWAR23 appears as a pan anti-SIRP antibody.

KWAR23 has been selected by Ring and collaborators for its ability to specifically block the SIRPα/CD47 interaction as does the anti-SIRPα Ab (19) (Figure 1E, left panel). LSB2.20 inhibits the SIRPγ/CD47 (16). Thus, we evaluated LSB2.20, KWAR23, and the anti-SIRPα for their ability to interfere with SIRPγ/CD47 binding. Surprisingly, the LSB2.20 did not efficiently block the SIRPγ/CD47 interaction at the concentration tested, neither did the anti-SIRPα. On the contrary, KWAR23 was found to efficiently abrogate SIRPγ/CD47 interaction (Figure 1E, right panel).

Thus, we redefined the KWAR23 as a Pan anti-SIRP antibody able to specifically block SIRPα-γ/CD47 interactions. SIRPγ Expression Varies on T-Cell Populations and While Being Highly Expressed on Treg, Is Not Implicated in Their Suppressive Functions
SIRPγ is expressed on T cells (16), and this expression can be altered by the presence of an eQTL (rs2281808) (20). So far, SIRPγ expression on different T-cell subsets has not been described. Thus, we analyzed SIRPγ expression on PBMC by FACS analysis and showed that naïve (CD3+CD45RA-CD28+) and central memory (CM, CD3+CD45RA-CD28+) T cells highly express SIRPγ whereas effector memory (EM, CD3+CD45RA-CD28-) T cells express to a lesser extent SIRPγ on their surface. Interestingly, TEMRA CD8 T cells (TEMRA, CD3+CD8+CD45RA-CD28- ) express SIRPγ at a lower level than EM T cells (Figures 2A, B). Since SIRPγ ligates CD47, a ubiquitously expressed molecule, we investigated CD47 expression on T-cell subsets. We show that it follows the same expression pattern as SIRPγ (Figures 2A, C). We then evaluated SIRPγ expression on T cells upon polyclonal stimulation. Proliferating T cells rapidly and significantly upregulates SIRPγ on their surface. Interestingly, upregulation of SIRPγ is increasing from Days 3 to 5 of activation. Conversely, the expression of CD47 on proliferating T cells decreases upon T-cell divisions with a higher impact at Day 5 as compared with Day 3 (Figures 2D, F).

We then investigated the expression of SIRPγ and CD47 on Tregs by gating on CD3+CD4+CD25+CD127low cells and demonstrated that both molecules are highly expressed on naïve (CD45RA+) and memory/activated (CD45RA-) Tregs while CD47 is significantly less expressed by memory/activated Tregs (Figures 2G–I). Prior to questioning the role of SIRPγ on Treg functions, we evaluated the consequences of SIRPγ blockade on T-cell proliferation upon CD3 and CD28 stimulations. As shown in Figure 2K, the specific blockade of SIRPγ by KWAR23 slightly reduces T-cell proliferation. We have previously shown that CD28 stimulation can inhibit Treg function (21), thus in an experimental set-up where T effector cells were only stimulated with coated OKT3, we analyzed the impact of SIRPγ blockade on Tregs suppressive function. After 5 days of culture in anti-CD3-coated wells, effector T cells (Teff) proliferated, while Tregs cultured in the same conditions did not (Figure 2J). When cocultured at a 1:1 Teff: Treg ratio, Tregs were able to inhibit Teff proliferation regardless of the presence or not of KWAR23 (Figure 2L), suggesting that SIRPγ is not involved in Tregs
suppressive function, although the direct effect of KWAR23 on OKT3-stimulated Teff and T reg cell would fully comfort our observations.

We further investigated the expression of SIRPγ on other cell types and detected it on MAIT cells, NKT cells (Supplementary Figures S1A–C), and in more than 60% of ILC1 cells (Supplementary Figures S1D, E). SIRPγ was not expressed on B cells neither on NK cells (data not shown) as already reported by Piccio (16).

Altogether, our results show that SIRPγ expression varies upon T-cell differentiation and T-cell stimulation, suggesting that this molecule could impact T-cell functions.
Figure 2 | SIRPγ expression varies on T-cell subpopulations, upon T-cell activation and is not implicated in Tregs function. (A) T-cell subpopulations gating strategy and their SIRPγ and CD47 expressions. (B) Geometric mean of SIRPγ and (C) CD47 on T-cell subpopulations; each symbol represents independent HV. Statistical analysis was performed by a Friedman test followed by a Dunn’s multiple comparison test. (D–F) SIRPγ and CD47 expressions were analyzed after T-cell stimulation with coated anti-CD3 and anti-CD28 mAbs. (D) Overlay allowing the comparison of SIRPγ (left) and CD47 (right) expression on unstimulated purified T cells (i.e., Day 0 in blue) vs. proliferating CPDlow T cells (i.e., Day 5 in red). (E) Geometric mean (± SEM) of SIRPγ or (F) CD47 on T cells as a function of their number of divisions, n = 10 HV analyzed in two independent experiments. Kruskal-Wallis test was used. (G) Treg gating strategy based on CD25 and CD127 expression is represented as dot plots; representative histograms of SIRPγ and CD47 staining on Treg subpopulations are shown. (H) Geometric mean of SIRPγ or (I) CD47 on Tregs are plotted. Each symbol represents independent HV. Statistical analysis was performed by a Friedman test followed by a Dunn’s multiple comparisons test. (J) A representative histogram of T effector (Teff) and T regulatory (Treg) cell proliferation under control conditions is shown at Day 5. (K) The effect of SIRPγ blockade with KWAR23 (10 µg/ml) was evaluated on T-cell proliferation upon OKT3 and anti-CD28 stimulation (both at 1 µg/ml), n = 2 independent experiments. (L) The consequences of SIRPγ blockade on Treg suppressive function was assessed by coculturing CPDlow/CPD-labeled Teff with CPDhigh/CPD-labeled Treg cells in the presence or not of stimulatory molecule (OKT3) with or without SIRPγ blockade (KWAR23). Percentage of CPDlow effector T cells (± SEM) is presented regarding culture conditions. n = 4 independent experiments. *p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001; ****p ≤ 0.0001; ns, not significant.
SIRPγ and CD47 Do Not Colocalize on Jurkat, Neither on Resting nor on Proliferating T Cells

Since SIRPγ and CD47 are both expressed on T cells with a similar expression pattern on the subpopulations, we hypothesized that CD47 may sequester SIRPγ at the cell surface by cis ligation on resting T cells. We first addressed this question using confocal microscopy by analyzing SIRPγ and CD47 expression patterns on wild-type (WT) Jurkat cells and on CRISPR-CAS9-modified Jurkat cells, i.e., Jurkat SIRPγ KO and Jurkat CD47 KO. On WT Jurkat cells, SIRPγ clusters in big punctuate patterns, whereas CD47 is expressed as smaller ones with a broad localization (Figure 3A). SIRPγ and CD47 therefore do not seem to colocalize on Jurkat cells. In the absence of SIRPγ, i.e., on SIRPγ KO Jurkat cells, CD47 expression is not altered and seems equivalent to that observed on WT Jurkat cells (Figure 3A). Surprisingly, the knockout of CD47 modifies the patterns of SIRPγ. SIRPγ is expressed on the whole cell surface instead of being in a big punctuate pattern at a specific localization of the cell surface as in WT Jurkat cells (Figure 3A). This might suggest that CD47 would impact SIRPγ distribution on the plasma membrane but not reciprocally. We also evaluated the expression of SIRPγ and CD47 on freshly isolated T cells; we identified a similar expression as on WT without any colocalization (Figures 3B, C). Given that SIRPγ and CD47 expression varies upon T-cell stimulation (Figures 2D–F), we analyzed their distribution on resting and on activated T cells. Upon T-cell stimulation, SIRPγ is upregulated and expressed on the whole T-cell surface as compared with resting T cells (i.e., freshly isolated T cells), in which SIRPγ is localized in big punctuate patterns (Figure 3B). However, CD47 distribution does not change upon T-cell activation (Figure 3B). The Pearson value clearly showed that there is no colocalization of SIRPγ and CD47 neither at steady state nor upon T-cell activation since the value obtained were not close to 1 (Figure 3C).

SIRPγ Polarizes at the Immune Synapse

SIRPγ/CD47 interaction has been shown to enhance superantigen-mediated T-cell costimulation (16); we therefore hypothesized that SIRPγ might be localized at the immune synapse. Interestingly, we observed a polarization of SIRPγ by time-lapse microscopy when Jurkat and SEE-primed Raji cells are in close contact. Jurkat cells emit pseudopods when they are closed to Raji cells and SIRPγ clearly polarized at the cell-cell contact (Figures 4A, B; Supplementary Video S1). To ensure that this cell-cell contact was a synapse, in one experiment, cells were fixed, permeabilized, and stained with phalloidin. The staining showed a polarization of actine at the cell-cell contact indicating the synapse formation (Figure 4C). Interestingly, CD47 (stained only on Raji cells) was localized at the immune synapse (Figure 4C). We obtained similar results when using T cells (stained with an anti-SIRPγ Ab) from healthy volunteer (HV) rather than Jurkat (Figure 4D). We further quantified SIRPγ expression intensity at the synapse and on the opposite side of the cell as explained in Figure 4E in all acquired time-lapse microscopy videos and showed a clear SIRPγ accumulation at the immune synapse (Figure 4F). Interestingly, blocking the SIRPγ-CD47 interactions with KWAR23 mAb did not affect SIRPγ polarization at the synapse (Figure 4F), suggesting that SIRPγ spatial reorganization of the synapse does not depend on its interaction with CD47 on the Raji cells.

Blocking SIRPγ/CD47 Interaction Impairs IFN-γ Secretion in Chronically Activated T Cells

To further decipher the importance of the SIRPγ/CD47 interaction on T-cell function, we stimulated T cells with low doses of OKT3 (0.5 or 1 µg/ml) and CD47-Fc, both coated, to provide a costimulatory signal as shown by Piccio and colleagues (16). With an experiment setup different than the one used by Piccio, we showed that the addition of coated CD47-Ig during T-cell stimulation did not increase their proliferation (Figure 5A);
however, coated CD47-Ig increases T-cell metabolism independently of its interaction with SIRPγ since the addition of KWAR23 has no impact (Figure 5B). Importantly, coated CD47-Fc significantly enhances IFN-γ secretion of OKT3-stimulated T cells; once again the addition of KWAR23 in the culture has no effect (Figure 5C). Our results show that T cells freshly stimulated with coated OKT3 and coated CD47-Fc benefited from CD47 in a SIRPγ-independent pathway.

CD47 is upregulated by different cells upon chronic infection, notably viral infection (22), and tumor cells. We therefore evaluated the impact of CD47-Fc stimulation on T cells chronically stimulated with three rounds of anti-CD3 anti-CD28 stimulations. Chronically activated T cells further stimulated with CD47-Fc kept their fitness as shown in the Figure 5D. However, their culture on a OKT3-CD47 coating increased their IFN-γ secretion (Figure 5E). To evaluate the role of the SIRPγ/CD47 interaction in this observation, the blocking anti-SIRPγ antibody KWAR23 or the blocking anti-CD47 antibody B6H12 were respectively added. As shown in Figure 5F, the addition of KWAR23 did not impair cell fitness, while B6H12 seriously impaired T-cell viability when used at 10 µg/ml (Figure 5F). This mAb has already been shown to induce T-cell apoptosis (23). KWAR23 significantly decreased IFN-γ secretion while a tendency was observed with the blocking anti-CD47 antibody B6H12 (Figure 5G). This observation suggests a role of the SIRPγ/CD47 interaction in chronically activated T cells.

**Blocking SIRPα-γ/CD47 Interactions In Vivo Impairs Human T-Cell Chimerism and Delays the Xeno-GvHD Onset**

Since SIRPγ is absent in rodents, we took advantage of a graft-versus-host disease (GvHD) model, in which human PBMC injected in NSG-recipient mice would be chronically stimulated by the xenogeneic environment. The role of SIRPγ in this experimental condition was evaluated by treating or not the irradiated recipient mice with 5 mg/kg of KWAR23 twice a week. In this situation, the xenogenic GvHD is human T cells
Inhibition of SIRPγ/CD47 interaction reduces IFN-γ secretion by chronically activated T cells.

(A–C) T cells were stimulated once with coated OKT3 (0.5–1 µg/ml) with or without coated CD47-Fc (10 µg/ml) in the presence or not of KWAR23. (A) T-cell proliferation after 5 days of stimulation was analyzed by CPD dilution assay (n = 3 independent experiments with a total of four different HV). (B) T-cell fitness was evaluated with Alamar Blue at Day 5; n = 4. (C) IFN-γ dosage in the supernatant of T cells after 2 days of stimulation in depicted stimulation conditions (n = 12 within seven independent experiments). (A–C) Statistical analyses were performed with a one-way ANOVA test followed by Kruskal-Wallis; *p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001, ns, not significant. (D, E) T cell subjected to three rounds of stimulation with coated OKT3 and anti-CD28 were further stimulated by a coated OKT3 stimulation with or without coated CD47-Fc (10 µg/ml). (D) T-cell fitness was evaluated with Alamar Blue at Day 5. (E) Supernatant were collected 48 h after the last stimulation and IFN-γ secretion was evaluated by ELISA. (D, E) Statistical analyses were performed by a Wilcoxon test. ns, not significant. (F, G) T cells subjected to three rounds of stimulation as previously mentioned were further stimulated during 48 h by a coated OKT3 stimulation with or without coated CD47-Fc (10 µg/ml) with various concentrations of KWAR23, B6H12, or LSB2.20. T-cell fitness was evaluated with Alamar Blue, n = 2 (mean ± SEM). (G) Supernatants collected at 48 h were analyzed by ELISA. Statistical analyses were performed by a Kruskal-Wallis test, *p ≤ 0.05 between KWAR23 and ISO and KWAR23 and LSB2.20 when Ab were used at 1.1 µg/ml; *p ≤ 0.05 between KWAR23 and ISO when Ab were used at 1.1 µg/ml; n = 3–5 (mean ± SEM).
driven due to their cross-reactivity with host MHCs (24). We first observed that mice survival was significantly extended when treated with KWAR23 (Figure 6A). At Day 6, mice treated with the KWAR23 showed a tendency of a lower human chimerism, although the frequency of human cells within the blood was quite low 1-week posthumanization. This observation was clearly significant at Day 14 (Figure 6B), suggesting that SIRPα blockade impairs T-cell survival, activation, proliferation, or localization within the host. This chimerism mostly comprised human CD3+ cells in both groups as shown by blood phenotypic analysis at Day 14 (Figure 6C). Human CD8 T cells played a primary role in xeno-GvHD pathogenicity (24); however, the CD4/CD8 ratio analyzed at D14 did not show an accumulation of CD8 T cells in the blood (Figure 6D). At Day 14, lower absolute numbers of human CD4 and CD8 circulating T cells were detected in the KWAR23-treated group (Figure 6E) which could explain the delay of the observed xeno-GvHD onset. Mice with a NOD background such as NSG mice, are superior to other strains for human immune cell engraftment thanks to their SIRPα whose affinity for human CD47 is even higher than the one for their natural ligand (mCD47), ensuring a functional “don’t eat me” signal (25). NSG mice are known to have functional residual macrophages. Given the weak chimerism observed in the KWAR23-treated group and the specificity of the KWAR23 to block human SIRPα-γ/hCD47 interactions, we wondered whether the KWAR23 cross-reacted with mouse SIRPα leading to the disruption of mSIRPα/hCD47 don’t eat me signal and eventually the phagocytosis of human cells. Intraportal macrophages collected from NSG mouse are all CD11b+ SIRPα+ and are not stained by the KWAR23 (Supplementary Figure S2A), ruling out our previously depicted hypothesis.

We evaluated the consequences of the KWAR23 treatment on T-cell activation. We analyzed PD1 expression on human circulating cells. All human T cells were PD1+ regardless of the treatment (Figure 6F). Therefore, the delay of the GvHD onset in the KWAR23-treated group was not due to a defect of human T-cell activation. In vitro, we have shown that chronically activated T cells secreted less IFN-γ when SIRPγ/CD47 interaction was blocked (Figure 5G). Thus, we measured IFN-γ concentration in mice sera collected at Day14. As expected, KWAR23-treated mice secreted significantly less IFN-γ than control mice (Figure 6G). This can be explained either by the lower amount of human T cells within the KWAR23-treated group, or by a direct effect of the treatment on chronically activated T cells (as shown in vitro in Figure 5G). Because human T cells are fully activated in the xenogeneic environment, we wondered if the KWAR23 treatment would impact the CD47’s downmodulation observed in vitro following T-cell stimulation (Figure 2F). Indeed, the downmodulation of human CD47 might reduce the “don’t eat me” signal and increase their susceptibility to residual mouse macrophages. KWAR23 did not stress CD47 down-regulation (Supplementary Figure S2B) indicating that the treatment would not increase T-cell susceptibility to mouse macrophages.

We then evaluated SIRPγ on human T cells in both groups by using the anti-SIRPγ mAb clone LSB2.20. We first validated that the KWAR23 did not impair the staining using LSB2.20 (Supplementary Figure S2C). The frequency of human CD4 and CD8 SIRPγ+ T cells in the KWAR23-treated group was not different to that in the PBS-treated mice (Figures 6H, I). However, SIRPγ geometric mean was significantly lower in the KWAR23-treated group (Figures 6H–J).

Thus, blocking SIRPγ/CD47 interaction in vivo impaired human T-cell chimerism by specifically affecting human T-cell numbers leading to the delay of xeno-GvHD.

Blocking SIRPα-γ/CD47 Interactions

In Vivo Impair Human T-Cell Differentiation

Because SIRPγ expression varies upon T-cell differentiation in the blood from HV (Figures 2A–C), we evaluated the impact of the treatment on T-cell differentiation in vivo. Three out of four HV whose PBMC were i.v. injected were phenotyped prior to their injection in NSG mice; naïve CD4, central memory CD4, and central memory CD8 T cells were the most abundant subpopulations (Supplementary Figures S3A, B). After i.v. hPBMC injection in untreated recipient mice, naïve T-cell compartment shrank in favor to central memory and effector memory cells regardless of the treatment. Whether or not this observation reflects naïve T-cell differentiation into central memory and effector memory cells, together with a memory T-cell compartment activation is difficult to evaluate. However, the inhibition of SIRPα-γ/CD47 interactions in vivo did not bias neither CD4 nor CD8 T-cell subpopulation frequencies (Figures 7A, B) while it clearly affected the absolute number of CD4 and CD8 T in all subpopulations except the TERMA CD8 T cells (Figures 7C, D). To further document the effect of KWAR23-treatment, some mice were sacked and their splenocytes were analyzed at Day 14. We confirmed that human cell engraftment was impaired in the spleen also upon KWAR23 treatment (Figure 8A). The CD4/CD8 ratio was significantly lower in the KWAR23-treated group (Figure 8B). A relative number of CD4 was significantly lower in the KWAR23-treated group (Figure 8C) without affecting the frequency of CD4 or CD8 T-cell subpopulations (Figures 8D, E) nor the relative number of those subpopulations (Figures 8F, G).

Altogether, our results suggest that KWAR23 treatment affects survival of all human T-cell subpopulations independently of mice macrophage implication.

DISCUSSION

SIRPγ protein is only expressed in humans and nonhuman primates (26) and binds CD47, an ubiquitous protein with an affinity 10-fold lower than that of SIRPα for CD47 (15). So far, reports on the potential role of the SIRPγ were performed with an anti-SIRPγ antibody (clone LSB2.20) which has not been appropriately characterized limiting the interpretation of previous reports on the SIRPγ biology. Thus, we decided to reevaluate the impact of SIRPγ in human T-cell biology using a different anti-SIRP monoclonal antibody (clone KWAR23) which we characterized here as capable to efficiently block
SIRPγ and SIRPα interactions with CD47. Addressing this question by blocking at the CD47 protein as previously reported (16) is challenging since CD47 is a multifaceted target engaging with various ligands in cis and in trans-interaction (27, 28), and some anti-CD47 mAb known to block SIRP/CD47 interaction (clone B6H12) can also trigger T-cell apoptosis (23).

Here, we report that SIRPγ is expressed by all naïve and central memory T cells and is upregulated after activation as previously reported (15), confirming that T cells might benefit
from this expression. While a previous study (16) reported that anti-SIRPγ (LSB2.20 clone) or anti-CD47 (clone B6H12) negatively impact human T-cell activation, we were first surprised that the specific blockade of SIRPγ/CD47 interaction using appropriate tools did not seem to directly impact T-cell activation upon one round of stimulation at least on T-cell functions addressed in our experiments, i.e., proliferation, metabolism, and IFN-γ secretion which are one of the many aspects of T-cell activation. The benefits of CD47-Ig coating on OKT3-stimulated T cells might therefore be dependent on a trans CD47/CD47 interaction as mentioned by others (27, 28).

SIRPγ extracellular portion consists of three domains (D1, D2, and D2), and isoforms containing D1/D2/D3, D1/D2, or D1 only, have been described (14, 16). SIRPγ/CD47 interaction was suggested to occur the SIRPγ D1 portion (29, 30). The LSB2.20 was raised against a SIRPγ-D1/D2-Ig fusion protein. Thus, one could consider that LSB2.20 epitope is not on the same extracellular domain than CD47-binding site. This would explain that targeting SIRPγ with either CD47-Ig or LSB2.20 on T cells would differently act on their effector functions as presented here and in a former study by Piccio for CD47-Ig or LSB2.20 stimulation, respectively (16). Recently, a patent by Ruozhen and colleagues in which epitopes of different anti-SIRPγ are mentioned, indicates that LSB2.20-binding domain is on SIRPγ D1 (31), so on the same domain than the CD47-binding site. Furthermore, they also showed LSB2.20 inefficient to increase T-cell proliferation upon CD3 stimulation (31).

Given the broad distribution of CD47, one could consider that its implication in T-cell biology through SIRPγ ligation should be finely tuned. Thus, we hypothesized that an upregulation of CD47 could impact SIRPγ ligation and lead to a biological effect. Indeed, CD47 is upregulated in a number of cancer cells, notably by MYC oncogene, a transcription factor, which binds promoter of cd47 and Pdl-1 genes (32). CD47 upregulation has been very recently shown in infected cells (22, 33). Here, we describe that SIRPγ clusters at the immune synapse and CD47 on the target cells also clusters at the cell/cell contact in a juxtacrine fashion. These observations confirm that SIRPγ expression and reorganization should also benefit human T cells. Unlike CD28 (34) on T cells or SIRPα on macrophages (35), SIRPγ localization at the immune synapse seem independent of its ligation to its known ligand CD47. This may reflect that SIRPγ/CD47 interaction needs to exceed a threshold for its effector functions, explaining SIRPγ localization to the immune synapse independently of CD47 engagement as a sensor searching for sufficient CD47 expression on the target cells.

It is worth noting that we could not reveal CD47 on WT Jurkat cells when using SIRPγ-Fc by FACS, while the SIRPα-Fc could. This may be due to the 10-fold lower affinity of SIRPγ to the level of CD47 expressed by Jurkat cells or to the conformation of either CD47 or SIRPγ. pH may influence protein/protein interactions as acidic pH has just unrevealed VISTA as a new ligand of PSGL1 (36), given that CD47 is
upregulated by tumor cells in a usually acidic environment, pH variation might also modify CD47/SIRPγ affinity.

In this study, all our experimental readouts focused on T-cell biology considering SIRPγ as a ligand. However, the 4 aa long SIRPγ intracellular tail is unexpected to transduce a signal by its own. Unlike SIRPβ, SIRPγ is not associated with DAP12 (4, 15, 16). Recently, the polymorphism of SIRPα (i.e., SIRPα-V1 and SIRPα-V2) has been shown to stimulate CD47 when donor differed from the recipient in one or both Sirpa alleles resulting in innate immune responses (37). Although these experiments were addressed in mice and not yet confirmed in humans, one can consider that upregulation of SIRPγ could contribute in a CD47 signaling.

Interestingly, all poxviruses that infect vertebrates (except four) contain predicted vCD47s (38). Even though their amino acid sequence similarity with their host parent gene is comprised between 18% and 29%, all of them conserved the structure with a N-terminal IgV domain, five transmembrane helices, and a short cytoplasmic tail (38). Although smallpox was recently eradicated by vaccination, the virus was present for thousands of years; SIRPγ may have interacted with vCD47. Recently, Myers and colleagues have shown that mice chronically infected with Friend retrovirus or LCMV Clone 13 accumulate some SIRPα+CD8+ T cells (39). They also evidenced this population in patients with chronic HCV infections by using a CyTOF approach. The expression of SIRPα was lower in these CD8 T cells than in

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**FIGURE 8** | Inhibition of SIRPαγ/CD47 interactions alters memory cells in the spleen of humanized mice. At Day 14, splenocytes were analyzed by FACS. (A) Human chimerism is shown; data were analyzed with a Mann-Whitney test. (B) CD4/CD8 ratio is shown regarding the treatment; data were analyzed with a Mann-Whitney test. (C) Relative number of human CD4 and CD8 in the splenocytes of recipient mice at Day 14. Data were analyzed with a one-way ANOVA test followed by Kruskal-Wallis. (D) Percentage of human CD4 and CD8 subpopulations is presented based on FACS analysis with the following patterns: CD45RA+CD27+ effector memory cells (EM), CD45RA−CD27+ central memory cells (CM), CD45RA−CD27− naive cells, and CD4+ CD45RA−CD27− for TEMRA CD8 cells. (E) Relative number of each subpopulation of CD4+ and (F) CD8+ T cells are presented. n = 1 experiment with PBMC from one HV. Data were analyzed with a one-way ANOVA test followed by Kruskal-Wallis. *p ≤ 0.05; **p ≤ 0.01; ns, not significant.
monocytes. In mice, the SIRPα expression on T cells was associated with effector functions; in humans, SIRPα/CD8+ T cells show higher activation status as compared with their SIRPα−CD8− counter parts (39). KWAR23, blocking anti-SIRPγ antibody, treatment was mostly effective in chronically activated T cells in vitro (Figure 5) and in vivo (Figure 6A). We previously reported that SIRPγ/CD47 blockade inhibits IFN-γ release in human tumor-antigen T-cell cross-priming assays using a CD8+ T-cell clone isolated and expanded from a melanoma patient (40, 41). Altogether, this suggests that SIRPγ/CD47 interaction might be more important in the activation biology of memory or chronically activated human T cells, than in naïve T-cell co-stimulation as previously hypothesized. Moreover, SIRPγ/CD47 interaction could play other roles, such as positively control the human T-cell transendothelial migration under shear flow conditions (17) or tissue migration (40). Since KWAR23 antibody blocks both SIRPα and SIRPγ interactions with CD47, we might consider that the observed effects were not only due to the sole SIRPγ blockade but also to SIRPα/CD47 blockade. However, on one hand in vivo, this antibody displays no cross-reactivity for mouse SIRPα and on the other hand in vitro in our chronically activated human T-cell models, only T cells remained after several polyclonal stimulation rounds.

Finally, we were surprised that the SIRPγ spatial reorganization at the synapse did not depend on its interaction with CD47. As previously mentioned, SIRPγ affinity for CD47 is relatively low and CD47 homophilic interactions have already been shown (27) which could explain CD47 clustering at the Raji membrane, although CD47KO target cells (here Raji) would be needed to fully confirm our observation. SIRPγ might interact with a yet unknown other receptor (beside CD47) which interaction might not be blocked by the KWAR23 antibody.

In the study, using a blocking anti-SIRPα-γ mAb, we report that T-cell interaction with CD47 is of importance not only for T-cell migration but also for their activation particularly in chronic stimulation situations. Whether these observations only involve SIRPγ or SIRPγ and SIRPα proteins remain to be clarified.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

Blood from healthy individuals was obtained at the Etablissement Français du Sang (Nantes, France). Written informed consent was provided according to institutional guidelines. The patients/participants provided their written informed consent to participate in this study. This study was carried out according to authorization from the French Ministry of Research, APAFIS # 4678.

AUTHOR CONTRIBUTIONS

SD, VN-D, MN, NE-D, J-MH, LB, CM, VT, and KB performed the experiments. SD, VN-D, NE-D, LB, and CM analyzed the experiments. NP, GB, and FH designed the experiment and overviewed the study. GB contributed to the writing of the paper. NP and GB obtained funding for the project. SD, NP, and FH wrote the paper. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2021.732530/full#supplementary-material

Supplementary Figure S1 | Expression of SIRPγ on blood cells other than T cells. SIRPγ expression has been analyzed on blood immune cells other than T cells. (A) Dot plot of SIRPγ expression on MAIT (CD45+CD3+TCRαβ+CD161+NKp14+) and NKT (CD45+CD3+CD56+) cells. (B) Geometric mean of SIRPγ on MAIT and NKT cells. Each symbol represents a HV. (C) Percentage of MAIT and NKT cell-expressing SIRPγ. (D) SIRPγ on ILCs (CD45+Lin−CD127+) is presented as dot plots; NCR marker, CRTH2, and CD161 were used to analyze the subsets of ILC. (E) Percentage of ILC-expressing SIRPγ, n = 3 independent HV.

Supplementary Figure S2 | The KWAR23 treatment does not affect the analysis of human cells in vivo. (A) Intraperitoneal NSG cells were collected and stained for mouse SIRPα and mouse CD11b. The cross reactivity of the KWAR23 (antihuman SIRPα-γ against mSIRPα) was evaluated. CD11b and SIRPα+ expression on intraperitoneal macrophages is presented as dot plot and the cross reactivity of the KWAR23 (revealed with the antihuman IgG PE) is presented in the red histogram while the control staining with the antihuman IgG PE only is presented in the black histogram (upper panel). In the same experiment, human PBMC were used as control and were either stained with KWAR23 and revealed with the antihuman IgG PE (red histogram) or were only stained with the antihuman IgG PE (black histogram). (B) Geometric mean (± SEM) of CD47 expression on CD8+ stained T cells is presented as a function of their division numbers, in the presence (red) or not (black) of KWAR23 during an anti-CD3+ anti-CD282 stimulation. n = 4 HV analyzed in one experiment. (C) SIRPγ staining with LSB2.20 on human CD3+ PBMC (left dot
Supplementary Figure S3 | T-cell phenotype of human PBMC prior to their i.v. injection in NSG. PBMC from three HV were analyzed by FACS prior their i.v. injection in NSG-irradiated recipient mice. Percentage of CD4 (A) and CD8 (B) subpopulations are presented based on FACS analysis with the following patterns.

Supplementary Video S1 | SIRPα polarizes at the Jurkat-Raji immune synapse. Time-lapse microscopy of the polarization of SIRPα at the Raji-Jurkat cell contact. Jurkat cells were stained with a cell tracker (green) and the anti-SIRPα (red); Raji cells (not stained) were primed with SEE.
33. Cham LB, Torrez Dulgeroff LB, Tal MC, Adomati T, Li F, Bhat H, et al. Immunotherapeutic Blockade of CD47 Inhibitory Signaling Enhances Innate and Adaptive Immune Responses to Viral Infection. *Cell Rep* (2020) 31:107494. doi: 10.1016/j.celrep.2020.03.058

34. Sanchez-Lockhart M, Graf B, Miller J. Signals and Sequences That Control CD28 Localization to the Central Region of the Immunological Synapse. *J Immunol* (2008) 181:7639–48. doi: 10.4049/jimmunol.181.11.7639

35. Morrissey MA, Kern N, Vale RD. CD47 Ligation Repositions the Inhibitory Receptor SIRPα to Suppress Integrin Activation and Phagocytosis. *Immunity* (2020) 53:290–302.e6. doi: 10.1016/j.immuni.2020.07.008

36. Johnston RJ, Su LJ, Pinckney J, Critton D, Boyer E, Krishnakumar A, et al. VISTA Is an Acidic pH-Selective Ligand for PSGL-1. *Nature* (2019) 574:565–70. doi: 10.1038/s41586-019-1674-5

37. Dai H, Friday AJ, Abou-Daya KL, Williams AL, Mortin-Toth S, Nicotra ML, et al. Donor SIRPα Polymorphism Modulates the Innate Immune Response to Allogeneic Grafts. *Sci Immunol* (2017) 2(12). doi: 10.1126/sciimmunol.aam6202

38. Farré D, Martínez-Vicente P, Engel P, Angulo A. Immunoglobulin Superfamily Members Encoded by Viruses and Their Multiple Roles in Immune Evasion. *Eur J Immunol* (2017) 47:780–96. doi: 10.1002/eji.201746984

39. Myers LM, Tal MC, Torrez Dulgeroff LB, Carmody AB, Messer RJ, Gulati G, et al. A Functional Subset of CD8+ T Cells During Chronic Exhaustion Is Defined by SIRPα Expression. *Nat Commun* (2019) 10:794–15. doi: 10.1038/s41467-019-08637-9

40. Gauttier V, Pergam S, Durand J, Biteau K, Mary C, Morello A, et al. Selective SIRPα Blockade Reverses Tumor T Cell Exclusion and Overcomes Cancer Immunotherapy Resistance. *J Clin Invest* (2020) 130:6109–23. doi: 10.1172/JCI135528

41. Vignard V, Lemercier B, Lim A, Pandolfini M-C, Guilloux Y, Khammari A, et al. Adoptive Transfer of Tumor-Reactive Melan-A-Specific CTL Clones in Melanoma Patients Is Followed by Increased Frequencies of Additional Melan-A-Specific T Cells. *J Immunol* (2005) 175:4797–805. doi: 10.4049/jimmunol.175.7.4797

**Conflict of Interest:** SD, LB, CM, VT, KB, and NP are employees and shareholders of OSE Immunotherapeutics, a company developing SIRPα antagonists. CM, VT, and NP are authors of patents related to SIRPα antagonists. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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