Mitis Group Streptococci Express Variable Pilus Islet 2 Pili

Dorothea Zähner1,4*, Ashish R. Gandhi1, Hong Yi3, David S. Stephens1,2,4

1 Division of Infectious Diseases, Department of Medicine, Atlanta, Georgia, United States of America, 2 Department of Microbiology and Immunology, Emory University School of Medicine, Atlanta, Georgia, United States of America, 3 Robert P. Apakarian Integrated Electron Microscopy Core, Emory University, Atlanta, Georgia, United States of America, 4 Department of Veterans Affairs Medical Center (Atlanta), Decatur, Georgia, United States of America

Abstract

**Background:** Streptococcus oralis, Streptococcus mitis, and Streptococcus sanguinis are members of the Mitis group of streptococci and agents of oral biofilm, dental plaque and infective endocarditis, disease processes that involve bacteria-host interactions. Their close relative, the human pathogen S. pneumoniae uses pilus-islet 2 (PI-2)-encoded pili to facilitate adhesion to eukaryotic cells.

**Methodology/Principal Findings:** PI-2 pilus-encoding genetic islets were identified in S. oralis, S. mitis, and S. sanguinis, but were absent from other isolates of these species. The PI-2 islets resembled the genetic organization of the PI-2 islet of S. pneumoniae, but differed in the genes encoding the structural pilus proteins PitA and PitB. Two and three variants of pitA (a pseudogene in S. pneumoniae) and pitB, respectively, were identified that showed ∼20% difference in nucleotide as well as corresponding protein sequence. Species-independent combinations of pitA and pitB variants indicated prior intra- and interspecies horizontal gene transfer events. Polyclonal antisera developed against PitA and PitB of S. oralis type strain ATCC35037 revealed that PI-2 pili in oral streptococci were composed of PitA and PitB. Electronmicrographs showed pilus structures radiating >700 nm from the bacterial surface in the wild type strain, but not in an isogenic PI-2 deletion mutant. Anti-PitB-antiserum only reacted with pili containing the same PitB variant, whereas anti-PitA antisera was cross-reactive with the other PitA variant. Electronic multilocus sequence analysis revealed that all PI-2-encoding oral streptococci were closely-related and cluster with non-PI-2-encoding S. oralis strains.

**Conclusions/Significance:** This is the first identification of PI-2 pili in Mitis group oral streptococci. The findings provide a striking example of intra- and interspecies horizontal gene transfer. The PI-2 pilus diversity provides a possible key to link strain-specific bacterial interactions and/or tissue tropisms with pathogenic traits in the Mitis group streptococci.

Citation: Zähner D, Gandhi AR, Yi H, Stephens DS (2011) Mitis Group Streptococci Express Variable Pilus Islet 2 Pili. PLoS ONE 6(9): e25124. doi:10.1371/journal.pone.0025124

Editor: Stefan Bereswill, Charité-University Medicine Berlin, Germany

Received August 2, 2011; Accepted August 25, 2011; Published September 22, 2011

Copyright: © 2011 Zähner, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by National Institutes of Health grant #R01AI070829 (www.nih.gov) and Department of Veterans Affairs VA Merit Award (www.va.gov). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: dzahner@emory.edu

Introduction

Oral streptococci of the Mitis group streptococci [1,2], such as Streptococcus mitis, Streptococcus oralis, and Streptococcus sanguinis are close relatives of the important human pathogen Streptococcus pneumoniae. They are members of the normal human oral flora, but in contrast to S. pneumoniae only occasionally cause acute or chronic disease as opportunistic human pathogens. But as early colonizers in the development of oral biofilms (for review see [3]), they are associated with gingivitis and caries and occasionally cause subacute infective endocarditis [4]. They share with S. pneumoniae the ability to develop natural competence for genetic transformation [5]; and extensive horizontal gene transfer, including virulence genes, has been documented between these species [1,6,7,8]. Hence the genetic diversity within these species results in strain-specific capabilities to form intra- and inter-species biofilms, to interact with salivary components or to bind platelets, all of which are factors important in host colonization and disease development.

Pili, filmbial extensions on the surface of bacteria have recently been identified in many Gram-positive species, including streptococci [9,10,11,12,13], enterococci [14,15], bacilli [16], lactobacilli [17], and actinomyces [18,19], and they have diverse, and often unresolved roles in environmental interactions or pathogenesis [20]. These pili are typically encoded by gene clusters, sometimes called pilus islets [21], that contain in addition the genes required for pilus biosynthesis [22]. Typically, these gene clusters encode a pilus backbone protein and one or two accessory pilus proteins, which in some pili are either located at the tip of the pilus [23,24], and may serve as adhesin [24], or at the base of the pilus, acting as a linker between the pilus backbone polymer and the cell wall [24,25,26,27]. All Gram-positive pilus proteins share a C-terminal LPXTG-motif or a variant thereof, as part of a cell wall sorting signal (CWSS). This CWSS is a typical feature of Gram-positive surface proteins that are attached to the cell wall [28,29]. In addition, at least one sortase enzyme is encoded in the pilus gene cluster. This sortase polymerizes pilus proteins to form the pilus structure [30]. Attachment of the pilus structure to the peptidoglycan precursor of the cell wall is usually performed by the housekeeping sortase, mostly referred to as sortase A [31], which in addition attaches other LPXTG-motif containing surface proteins to the cell.
surface and is encoded outside the pilus gene cluster. The surface exposure of Gram-positive pili has made them attractive vaccine candidates [32].

In Mitis group streptococci, pili have been identified in S. sanguinis and S. pneumoniae [9,13]. S. pneumoniae can harbor two different types of pilus islets, the pilA pilus islet, also called pilus islet 1 (PI-1) [9] and the type two pilus islet, named pilus islet 2 (PI-2) [21]. The pilus gene cluster in S. sanguinis resembles the PI-1 pilus islet in S. pneumoniae [13]. The two pilus islets in S. pneumoniae encode two antigenically different pili [21]. The pneumococcal PI-2 islet has been shown to be present in about 10 to 21% of pneumococcal isolates [21,33]. The pneumococcal PI-2 islet is unusual since it encodes a pilus formed solely by a pilus backbone protein, PitB [21]. A second gene, *pitC*, encodes signature motifs of a Gram-positive surface protein, i.e. an N-terminal signal sequence and a C-terminal CWSS, but it is a pseudogene due to a stop-codon causing its premature termination. The PitB-based pilus has been shown to be present as a single structure on the bacterial surface, in contrast to the typical multitude of pili structures per bacterial cells observed for other Gram-positive bacteria [21,22]. An additional unusual feature of the pneumococcal PI-2 islet is that of the two encoded sortase genes, *srtG1* and *srtG2*, the latter is a pseudogene in most of the analyzed PI-2 gene clusters [21]. The pneumococcal PI-2 islet shows an unusual high sequence conservation of >99.9% identity over the entire 6.8 kb region among all PI-2 containing pneumococcal isolates, and is inserted in all analyzed strains in the same insertion site, located between the *papT* and *hemH* genes [21,33]. A PI-2-piliated pneumococcal strain has been shown to mediate host cell adhesion in a pilus-dependent manner [21].

We report the first identification of PI-2 pili in *S. oralis* and *S. mitis*, with several strains expressing pili that, in addition to the pilus backbone protein, have a putative adhesin protein attached to the pilus backbone structure. Genetic variability between genes encoding the PI-2 pilus proteins showed that the PI-2 islet in oral streptococci was subject to horizontal gene transfer between these species. Further, by immunogold electron microscopy (EM), pili were found to share the morphology of the pneumococcal PI-2 pilus. The sequence variation in PitB was shown to result in antigenic variation. The variability of the pilus proteins within and beyond the species suggests a role for PI-2 pili in the strain-specific tissue tropism of the Mitis group of streptococci.

### Results

**PI-2 pilus islets in oral Mitis group streptococci**

In a search for pilus-encoding islets that resemble PI-2 of *S. pneumoniae*, the publicly available genomic sequences of oral Mitis group streptococci and selected additional strains (Table 1), isolates that have frequently been used as reference strains in studies of oral streptococci, were examined. PI-2 pilus islets were identified in genomic sequencing projects of the *S. oralis* type strain ATCC35037 [34], *S. mitis* ATCC6249, *S. sanguinis* ATCC49296, and *S. sp. C300* (all part of the “The NIH human microbiome project” [http://nihroadmap.nih.gov/hmp]), and *S. oralis* Uo5 [35] (Figure 1). In addition, PI-2 pilus islets were identified by PCR in *S. oralis* strain ATCC10557 and strain 34 (both formerly classified as *S. sanguinis* [34,36] (Figure 1). ATCC10557 is an isolate from a patient with endocarditis often used as a *S. oralis* reference strain in adhesion and disease-associated studies [37], and strain 34 is most commonly used in studies of experimental biofilm formation involving *S. oralis* [19]. After initial identification of the PI-2 islet in ATCC10557 and 34 by PCR, the entire DNA sequence of the PI-2 genetic islets was determined. In all seven strains (Figure 1) the PI-2 islet encompassed 6.8 kb and was inserted between the *papT* and *hemH* genes, consistent with the insertion site of the PI-2 islet in *S. pneumoniae* [21]. In all seven isolates the PI-2 islet encoded the genes for the same five proteins: for two putative pilus proteins, named according to the pneumococcal PI-2 islet, *pitA* and *pitB*, a signal peptidase like-protein, *spb3* [21,38]; and two putative sortases, *srtG1* and *srtG2*, respectively (Figure 1). The overall similarity between the PI-2 nucleotide sequences of the seven strains ranged from 87% to 94% identity, and their GC-content was 37.8 to 38.4%, which is lower than the average GC-content of the available genomic sequences of the hosting *S. oralis*, *S. mitis*, *S. sanguinis* strains, and *S. sp. C300*, respectively (41%), but closer to *S. pneumoniae* strains (39%), suggesting that the PI-2 islet may have evolved in a different species, with slightly lower GC-content.

The *pitC* gene, which is a pseudogene in the *S. pneumoniae* PI-2 pilus islet, was the first gene of the oral streptococci PI-2 islet (Figure 1) and was predicted to encode in all seven streptococcal strains a functional surface protein, with a typical N-terminal signal peptide and a LPXTG-like motif (VPETG) in the C-terminal CWSS. In addition, PitA in all five strains was predicted to contain a von Willebrand factor type A (vWA) domain [39], characteristic of many eukaryotic extracellular-matrix binding

### Table 1. Streptococcal strains used in this study.

| Strain | Relevant genotypic features | Source/alternative name/reference |
|--------|-----------------------------|----------------------------------|
| *S. oralis* ATCC35037 | PI-2 | ATCC\(^a\); NTCT11427, SK23; type strain |
| ATCC350373P12 | ATCC 35037-derivative, but PI2 ::km | This study |
| *S. oralis* ATCC10557 | PI-2 | ATCC; SK2 |
| ATCC105573P12 | ATCC 10557-derivative, but PI2 ::km | This study |
| *S. oralis* 34 | PI-2 | [36] |
| *S. mitis* ATCC6249 | PI-2 | ATCC |
| *S. sanguinis* ATCC49296 | PI-2 | ATCC |
| *S. pneumoniae* GA47901 | PI-2 | [33] |
| *S. pseudopneumoniae* BAA-960 | PI-2 | ATCC; CDC-SS-1757; type strain |
| *S. gordonii* ATCC10558 | PI-2 | ATCC; type strain |

\(^a\)American Type Culture Collection.

doi:10.1371/journal.pone.0025124.t001
proteins, such as α2β1 integrin, an important human receptor for collagen [40,41]. The vWA domain resided in the N-terminal region of PitA (amino acid residues 77 to 355 in the PitA sequence of ATCC35037), and contained the characteristic MIDAS motif (Asp65-X-Ser67-X-Ser69 and Asp249) required for metal-conjugation [41,42]. vWA domains have been identified in several Gram-positive pilus proteins that act as adhesins [17,27,43], suggesting a similar role for PitA.

The pitB genes in six of the seven strains (exception S. oralis 34) encoded a protein with a non-canonical LPXTG-like motif (VTPTG) in the CWSS. The same motif is also present in PitB of S. pneumoniae [21]. In addition, amino acid residues that have been shown for pneumococcal PitB to form two intramolecular isopeptide bonds, each formed between a Lys and Asn residue [44] were conserved in all four alleles (Figure S1). Additional motifs described for some Gram-positive pilus backbone proteins, i.e. the pilin motif and the E-box-motif [30], were missing in PitB of the oral streptococci, consistent with their absence in pneumococcal PitB [21]. In strain S. oralis 34, the start codon was missing and alternative start codons were not apparent, suggesting that pitB was not transcribed in this strain (Figure 1).

The three additional proteins encoded in PI-2, SipA, which has actually occurred at the border or within the highly conserved sipA gene and the flanking pepT and sortase genes (Figure 2). The DNA sequence variation between the different pitA regions in Mitis group streptococci.

doi:10.1371/journal.pone.0025124.g001

Figure 1. Schematic representation of PI-2 pilus-encoding regions in Mitis group streptococci. Type strains (T) and strains from genomic sequencing projects (G) are indicated. GenBank accession numbers: S. oralis ATCC35037: AEDW01000020; S. oralis Uo5: FR720602; S. oralis SK255: AFNM00000000; S. mitis ATCC6249: AEEI01000012; S. pneumoniae GA47901: AFGR00000000; S. pneumoniae TIGR4: AE005672; S. sanguinis ATCC49296: AEPO01000013; S. sanguinis ATCC10556: AFNM20000000; S. sp. C300: ACJR00000000. Shaded areas indicate pseudogenes (one asterisk indicates a mutation in the start codon and two asterisks indicate the position of a stop-codon).

doi:10.1371/journal.pone.0025124.g002

Figure 2. Conservation of genes and proteins in the PI-2 pilus-encoding regions of Mitis group streptococci. Sequence identity within the same variants and between different variants is indicated. GenBank accession numbers are as provided for figure 1. An asterisk indicates a mutation in the start codon; two asterisks the position of a stop-codon.

doi:10.1371/journal.pone.0025124.g003
PI-2 pili on the surface of oral streptococci

To determine whether PI-2 pilus islet-containing strains expose pili on the cell surface, cell wall extracts of strains representing the five different pilus variants, i.e. ATCC35037, ATCC10557, 34, ATCC6249, and ATCC49296 (which encodes pili identical to Us5 and C300), were prepared and analyzed for the presence of pilus protein-containing polymers by Western blots. Gram positive pilus polymers produce a characteristic high molecular weight (HMW) banding pattern in Western blots developed with an antibody against a pilus protein [10,11]. Anti-PitB and anti-PitA polyclonal antiseras were developed against the ATCC35037 proteins, and PI-2 deletion mutants were constructed in ATCC35037 and ATCC10557 to verify the specificity of the anti-PitB and anti-PitA polyclonal antiseras for PI-2-encoded proteins. A Western blot using anti-PitB antiserum revealed HMW bands in cell wall extracts of the ATCC35037 wild type strain (Figure 3A). Corresponding bands were not detected in cell wall extracts of the isogenic PI-2 deletion mutant (Figure 3A), which confirmed the HMW bands detected in the wild type strain were encoded in the PI-2 islet, and that the anti-PitB antiserum was specific for a PI-2 encoded protein. A similar HMW banding pattern was detected in cell wall extracts of S. mitis ATCC6249 (Figure 3A), the strain that encoded the same PitB variant as ATCC35037. However, no HMW banding pattern or band corresponding to the molecular weight of the PitB monomer (37 kDa) was detected in cell wall extracts of ATCC10557, its PI-2 deletion mutant, S. oralis 34 or S. sanguinis ATCC49296. This indicated that either these strains did not express the PitB protein or the anti-PitB antiserum was not cross-reactive with the PitB variants expressed in these strains (Figure 3A). In addition, PitB antiserum was not cross-reactive with PitB of S. pneumoniae, and polyclonal antiserum developed against PitB of S. pneumoniae [33] was not reactive with any of the identified PitB variants in oral streptococci (data not shown).

To determine whether PitA was present in pilus structures, the cell wall extracts of the five strains and the two PI-2 deletion mutants were examined for HMW bands with the polyclonal anti-PitA antiserum (Figure 3B). HMW banding patterns were observed in cell wall extracts of S. oralis ATCC35037, ATCC10557, and S. sanguinis ATCC49296 (Figure 3B). The HMW patterns were absent from both isogenic PI-2 deletion mutants, which confirmed the specificity of the anti-PitA antiserum for a PI-2 encoded protein in both strains. In addition, the anti-PitA antiserum reacted with two bands corresponding to the approximate molecular weight of the PitA monomer (88 kDa) in the ATCC10557 wild type strain that were absent in the isogenic deletion mutant (Figure 3B). The two bands suggested the presence of two differently processed species of PitA monomers or a partial instability of the PitA monomer. The HMW banding pattern in ATCC10557 was faint compared to ATCC35037 and ATCC49296, and faint relative to the two bands of the PitA monomer in this strain, suggesting that pilus polymerization occurred in this strain, but was less efficient than in the other two strains (Figure 3B). Detection of HMW bands in ATCC10557 and ATCC49296 with anti-PitA but not with anti-PitB antiserum strongly suggested that a PitB pili backbone was formed in these strains, but that the PitB variants encoded in these strains eluded detection by the anti-PitB antiserum. HMW bands were not detected in S. oralis 34 or S. mitis ATCC6249. However, in S. oralis 34 two bands that corresponded to the molecular weight of the PitA monomer (88 kDa) were detected, demonstrating that PitA was expressed, but not polymerized (Figure 3B). Since pitB is a pseudogene in S. oralis 34, this finding was consistent with PitB being the supposed pili backbone protein of PI-2 pili, as has been shown for the S. pneumoniae PI-2 pili, and PitA an ancillary pilus protein. However, detection of PitA in S. oralis 34 also
demonstrated that anti-PitA antiserum was able to react with this PitA variant, which is about 20% different from PitA of ATCC35037. Since PitA of S. oralis 34 and S. mitis ATCC6249 are very similar (>94%), the lack of detection of HMW bands or PitA monomer in the latter strain suggests that PitA may not be expressed in ATCC6249. Taken together, these findings demonstrated the formation of pilus polymers in S. oralis ATCC35037, S. oralis ATCC10557, S. mitis 6249 and S. sanguinis ATCC49296. In addition, the results confirmed that PitA, at least in ATCC35037, ATCC10557 and ATCC49296, was part of PI-2 pilus polymers. The PitB variants were antigenically different from each other, whereas the PitA variants showed some cross-reactivity with the used polyclonal antiserum.

To confirm and evaluate the morphology of pilus structures on the surface of the streptococcal isolates that encode PI-2 islets, bacteria were analyzed by EM. Immunogold-labeled anti-PitB antiserum and anti-PitA-antiserum were used to study the wild type strains and deletion mutants. Anti-PitB-antiserum reacted with pili on the surface of S. oralis strain ATCC35037 and S. mitis ATCC6249 (Figure 3C). The label was distributed evenly along the pilus structures, which appeared mostly as single pili per bacterium. The pilus length ranged between 0.7–2.5 μm. Consistent with the results obtained in Western blots, anti-PitB antiserum failed to identify pili on the surface of S. oralis ATCC10557 and 34, as well as S. sanguinis ATCC49296 (data not shown). Anti-PitA antiserum revealed no pilus structures on the surface of any of the tested strains (data not shown), suggesting that PitA was not present along the pilus backbone. Pili structures were not visible by negative staining alone and other defined surface structures were not discernible (not shown).

Taken together, pilus structures were observed on the surface of S. oralis strain ATCC35037 and S. mitis ATCC6249. Strong evidence for the presence of PI-2 pili was obtained for S. oralis ATCC10557 and S. sanguinis strain ATCC49296 in Western blots using anti-PitA antiserum. The anti-PitB antiserum lacked cross-reactivity with pilus protein variants different from ATCC35037.

Phylogenetic relationship of PI-2-containing isolates and conservation of the PI-2 insertion site in Mitis group streptococci

PI-2 pilus encoding islets have been identified in strains of four different species: S. pneumoniae, S. oralis, S. mitis and S. sanguinis, and also in the unassigned streptococcal strain C300, and the islet was always inserted between the pepT-hemH genes (Figure 1; [21,33]). To determine the phylogenetic relationship between the PI-2-containing strains identified in this study and the extent of conservation of the pepT-hemH region throughout the Mitis group streptococci electronic Multilocus Sequence Analysis (eMLSA, [46]) was performed. eMLSA is based on seven independent genetic loci, selected to reflect the phylogenetic relationship among viridans streptococci [46]. The analysis included all strains of species of Mitis group streptococci with currently available genomic sequences (deposited as complete or partial genomic sequences at the National Center for Biotechnology [NCBI]) and the additional strains used in this study. Figure 4 shows that PI-2-containing oral streptococci all belonged to a cluster of strains that included the S. oralis type strain ATCC35037. The identification of S. mitis strain ATCC6249 within this cluster of mostly S. mitis strains is consistent with earlier reports describing S. mitis strains that were more closely related to S. oralis strains than to other S. mitis strains, disclosing S. mitis as a very heterogeneous species among streptococci [6]. Surprisingly, also S. sanguinis ATCC49296 was more closely related to strains of S. oralis than to S. sanguinis (Figure 4). This finding suggests that S. sanguinis ATCC49296 has been misclassified as S. sanguinis. Hence, all PI-2-containing oral streptococcal strains are closely related to S. oralis strains.

The pepT-hemH region was conserved throughout most of the species of Mitis group streptococci, including the important human pathogen S. pneumoniae and even the more distantly related Streptococcus gordonii (Figure 4). The sequence identity of the approximately 150-bp intergenic region between pepT and hemH ranged from >52% for S. gordonii to 67–96% for all other species, and between 76–92% and 72–90% for the flanking genes pepT and hemH, respectively, relative to S. oralis SK255. The exceptions were Streptococcus parasanguinis (formerly S. parasanguis), the close relative of S. parasanginis, Streptococcus orta [47], and Streptococcus cristatus (formerly S. cristus) and an isolate of Streptococcus infantis (Figure 4). In these species or strains, respectively, unrelated putative proteins are encoded adjacent to pepT, without conservation of the intergenic region, and hemH homologs, if present, were encoded in a distant region of the genome, likely excluding these strains from acquisition of PI-2 at this site. In all S. sanguinis strains (except the above discussed ATCC49296, supporting its misclassification) a 4.1 kb region, which is unrelated to PI-2, was inserted between pepT and hemH (Figure 1). This finding demonstrates that the pepT-hemH insertion site is a hotspot of recombination and not specific for PI-2.

Bagnoli et al. [21] described a 7-bp sequence (TCGTTTT) putative insertion site for PI-2, which is duplicated in some of the pneumococcal PI2-encoding strains. However, this duplicated sequence was absent from the borders of the PI-2 islets in the oral streptococci, suggesting acquisition of PI-2 by homologous recombination within the 150-bp intergenic region between pepT and hemH or the flanking genes. The conservation of the pepT-hemH region among most of the Mitis group streptococci provides the possibility for PI-2 acquisition by other strains of the Mitis group streptococci. PI-2 was not identified in any other chromosomal location.

Taken together, PI-2 pilus encoding islets were identified in strains of S. oralis and S. mitis and the insertion site between pepT and hemH was conserved among most species of Mitis group streptococci. The expressed PI-2 pili were composed of a strain-specific combination of PitA and PitB variants, which results from intra- and interspecies horizontal gene transfer.

Discussion

This is the first report of the identification of PI-2 pili in oral streptococci of the Mitis group. PI-2 pili were identified in strains of S. oralis and S. mitis, species that can be associated with a broad range of pathogenesis traits. These traits lead to early biofilm formation in dental plaque development, adhesion to heart valves resulting in bacterial endocarditis; and involve intra- and interspecies interactions, as well as interactions with host structures and tissues, such as salivary proteins or platelets in the genesis of endocarditis. Adhesion and colonization, have been described for strains of the Mitis group streptococci [48,49], but the molecular basis for these events is not fully understood and in other Gram positive bacteria, such as actinomyces or enterococci [19,50], these events are conferred by pili. Thus, PI-2 pili of Mitis group streptococci are candidates for contributing to adhesive interactions.

PI-2 pili have formerly been identified in S. pneumoniae. However, pneumococcal PI-2 pili are composed solely of the pilus backbone protein PitB [21]. In contrast, in oral Mitis group streptococci, PI-2 pili were composed of PitB and PitA (the latter is encoded as a pseudogene in S. pneumoniae). PitA is a putative adhesin, which is suggested by the presence of a VWA domain.
The vWA domain is present in both PitA variants, such as in ATCC35037 and ATCC6249 despite their 20% sequence variation. Further, the second sortase gene encoded in the PI-2 pilus islet, srtG2, also a pseudogene in most pneumococci [21], is predicted to be functional in all the analyzed oral streptococcal PI-2 islets. Thus, PI-2 pili in oral Mitis group streptococci retained features that have been lost in the pneumococcal PI-2 pilus islet.

Analysis of the PI-2 islets provided evidence for horizontal gene transfer (HGT) events, which were not limited to the acquisition of the entire PI-2 islet but also involved recombination events within the islet, as evident by the presence of combinations of particular PitA or PitB variants found within and beyond species (Figure 2). The observed sequence variation between the pitA and pitB variants, respectively, is consistent with the variation in previously
identified antibiotic resistance-conferring mosaic genes in penicillin binding proteins (PBP) of *S. pneumoniae* that originated from PBPs of *S. mitis* or *S. oralis* [51,52,53] and mosaic blocks in the virulence factor NanA [54], with a typical difference between 20%–30% [51,52]. Western blot and immunogold-labeling EM revealed that the PilB variant of ATCC35037/ATCC6249 had different antigenic properties than ATCC10557 and ATCC49296/U65/C5300, respectively. Hence, the 20%–25% sequence variation between the PilB variants was sufficient to abolish polyclonal antibody cross-reactivity. PI-2 pilus sequence variation may contribute to host evasion or specific pathogenic events and has implications for pilus proteins as vaccine candidates.

The modular structure of the pilus proteins, with highly conserved signal sequences and motifs in the cell wall sorting signal, in combination with conserved biosynthetic genes, provides a framework to incorporate variable domains that may result in pili with different functions. The PilB variants as well as the PilB variants may facilitate adhesion. The PilB-based PI-2 pilus in *S. pneumoniae* mediates adhesion to eukaryotic cells, although with far less efficiency than observed for other Gram-positive pili [21] that employ the accessory pilus adhesins for this interaction. The putative adhesive vWA domain in PilA may mediate binding to various substrates. Konto-Ghiorghi et al. have shown that the vWA domain in the pilus protein Gbs1474 (PilC) of *S. agalactiae* is critical for adherence to epithelial cells [27]. In addition, Gbs1474 has a vWA-domain-independent function in biofilm formation [27]. Dissection of the functional role of streptococcal PI-2 pili will need to consider the role of both pilus proteins and also address the broad spectrum of bacteria-bacteria as well as bacteria-host adhesive interactions associated with the Mitis group streptococci.

A striking feature of the *S. pneumoniae* PI-2 islet is the inactivation of pilA and srtG2. In general, gene inactivation was not observed in the streptococcal PI-2 gene clusters. But the pilB gene of *S. oralis* strain 34 lacked a start codon, and pilus structures were not seen. However, PilA monomer was identified in cell wall extracts of this strain and may exert an adhesive function on the bacterial surface, even though not as part of a pilus structure. Also, the absence of PilA in strain ATCC6249 requires further evaluation. Further studies will have to determine whether selective gene inactivation but retention of parts of the biosynthetic machinery of the PI-2 pili may provide an additional molecular mechanism to enhance variation in structure and function of PI-2 pili, and whether this may modulate the tissue tropism of certain strains.

Identification of the PI-2 islet in species of *S. mitis*, *S. oralis* and the more distantly related *S. sanguinis* suggested a distribution of PI-2 throughout the entire streptococcus Mitis group. However, eMLSA revealed that all PI-2-containing strains were closely related to each other and to PI-2 lacking strains of *S. oralis*. This included *S. sanguinis* ATCC49296, which was in fact identified as *S. oralis* based on biochemical characterization in an earlier study [55], and based on eMLSA should be considered misclassified as *S. sanguinis*. Although HGT and the ability to develop competence for genetic transformation [56,57,58] has been observed throughout the Mitis group of streptococci, the increasing number of available genomic sequences has revealed that HGT is a particularly common theme among *S. pneumoniae*, *S. mitis* and *S. oralis* [7]. The identified distribution of PI-2 islets suggests that they are part of a common gene pool shared by *S. pneumoniae*, *S. mitis*, and *S. oralis*.

Filamentous surface structures have long been observed on the surface of oral streptococci, including strains of *S. oralis*, *S. mitis* and *S. sanguinis* (most of them then classified as *S. sanguis*) by electron microscopy [39,60,61,62,63]. They were visualized by negative staining and described as tufts, fibrils or fimbriae, and ranged from a 30 nm to ≤700 nm that either coexisted on the bacterial cell or seemed to be mutually exclusive [60,63]. *S. oralis* strains ATCC35037, ATCC10557, and 34 were described to express fibrils, but no fimbriae [64]. The molecular nature of these structures remained elusive until cross-reactive antibodies against the Csh-like protein, first identified in *S. gordoni* [25], detected fibrils of 50–80 nm in several *S. oralis* and *S. sanguinis* strains [64]. CshA homology are encoded in *S. oralis* and *S. mitis*, but due to major sequence variation in the binding domain, their function remains to be determined [64]. However, the PI-2 pili described in this study were considerably longer (>700 nm), and therefore inconsistent with these fibrilar surface structures. The failure to identify pili in earlier studies is consistent with our inability to detect pilus by negative staining alone. In fact, the lack of sufficient contrast of Gram-positive pilus by negative staining has contributed to the delay in identification of Gram-positive pili [10]. Several previous studies [60,62,63] have tried to correlate the observed surface structures on Mitis group streptococci with pathogenesis relevant traits, such as coaggregation and binding to salivary proteins [60,62,63], but the results have been inconclusive.

Dissection of the role of PI-2 pili may help to elucidate some of these events.

Taken together, genetically variable PI-2 pili were identified and characterized in multiple strains of *S. oralis* and *S. mitis*. The observed intra- and interspecies variation in the genes encoding these pili provide evidence for horizontal gene transfer events between and within PI-2 islets of these species. Further work will have to determine whether the pilus protein variants confer different adhesive properties to the respective strain. The discovery of PI-2 pili in oral Mitis group streptococci may provide a key to dissect the heterogeneity of interspecies and host interactions associated with these bacteria.

**Materials and Methods**

**Bacterial strains and growth conditions**

All streptococcal strains used in this study are listed in Table 1. Streptococcal strains were routinely grown in Todd-Hewitt broth (BD) supplemented with 0.5% yeast extract (THY) and antibiotics, if appropriate. Antibiotics were used in the following concentrations: spectinomycin 110 µg/ml; kanamycin 50 µg/ml for *Escherichia coli* and 800 µg/ml for streptococcal strains.

**PCR-based screen for PI-2**

To identify PI-2 islets primers sipA_so_up and sipA_so_dn (Table S1), designed against the PI-2 specific gene sipA [21] were used. To determine the absence of PI-2 from the described insertion site primers pepT_so_F and hemH_so_R (Table S1) were designed against the flanking genes pepT and hemH. All primers were designed based on the sequence of *S. oralis* strain ATCC35037.

**Electronic Multilocus Sequence Analysis (eMLSA)**

The phylogenetic relationship between strains used in this study was determined by multilocus sequence analysis [46] following the instructions provided online for the viridans streptococci project (www.viridians.emlsa.net). Multilocus sequence information of strains from genomic sequencing projects was retrieved from the according loci of the genomic sequences, in case the data was not already included in the eMLSA database, and from strains with not yet available sequence information by amplification and sequencing of the corresponding loci. Primers and conditions were used according to the online instructions. Alignment of concatenated sequences was performed with MEGA version 5.0 [65].
Construction of PI-2 deletion mutants

3′- and 5′-flanking regions of PI-2 of *S. oralis* strain ATCC35037 were amplified using primer pairs pepT_so_F/PitA_so_3 and SrtG2_so/hemH_so_R, respectively (Table S1). Both PCR products were used in a PCR based overlap-extension reaction [66], introducing a Mfd restriction site between the two fused fragments. The PCR product was cloned into the pCR-Blunt II-TOPO vector (Invitrogen), resulting in pCR8-ΔPI-2. The kanamycin-cassette of pSP151 [67] was amplified using primers KM_F and KM_R (Table S1), cloned into the TOPO vector pCR2.1 (Invitrogen), recovered pSF151 [67] was amplified using primers KM_F and KM_R (Table S1) and cloned into the E. coli expression vector pET-32a+ (Stratagene). The thioredoxin/6xHis-fusion proteins were purified using the B-PER 6xHis Fusion Purification Kit (Pierce). The N-terminal thioredoxin/6xHis-fusion proteins were purified using the B-PER 6xHis Fusion Purification Kit (Pierce). The N-terminal thioredoxin/6xHis-tag was removed by cleavage with recombinant enterokinase (Novagen) according to the manufacturer’s recommendations. Purified PitA and PitB proteins were used to produce polyclonal antisera in rabbits (Covance Research Products Inc.).

Production of recombinant PitA and PitB

The regions of *pitA* and *pitB*, corresponding to the mature proteins (bp 106 to bp 2484 and bp 91 to bp 1128, respectively) were amplified from chromosomal DNA of *S. oralis* ATCC35037 using primer pairs PitA_so_SmaI_F/PitA_so_NotI_R, and PitB_so_BamHI_F/PitB_so_Xho_R, respectively (Table S1) and cloned into the E. coli expression vector pET-32a+ (Stratagene). The thioredoxin/6xHis-fusion proteins were purified using the B-PER 6xHis Fusion Purification Kit (Pierce). The N-terminal thioredoxin/6xHis-tag was removed by cleavage with recombinant enterokinase (Novagen) according to the manufacturer’s recommendations. Purified PitA and PitB proteins were used to produce polyclonal antisera in rabbits (Covance Research Products Inc.).

Preparation of streptococcal cell wall extracts

Cell wall extracts of streptococcal strains were prepared from bacteria grown on trypticase soy agar (TSA) plates containing 5% sheep blood (BD) overnight at 37°C. Bacteria were collected from plates by adding 750 μl 10 mM Tris, pH 8 buffer to the plate surface, followed by resuspension and recovery of the bacteria. Samples were centrifuged for 5 min at 13000 x g and bacterial pellets were used for further processing. Bacterial pellets (approximately 40 mg per strain) were resuspended in 250 μl 10 mM Tris, pH 8 buffer to the plate surface, followed by resuspension and recovery of the bacteria. Samples were centrifuged for 5 min at 13000 x g and bacterial pellets were used for further processing. Bacterial pellets (approximately 40 mg per strain) were resuspended in 250 μl 10 mM Tris, pH 8.0 supplemented with 400 U/ml mutanolysin (Sigma-Aldrich, St. Louis, MO, USA) and 30% raffinose. After 2 h of incubation at 37°C, cell wall extracts were collected by centrifugation at 12,000 × g for 15 min. 25 μl cell wall extract were mixed with sodium dodecyl sulfate–polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer and boiled for 10 min immediately before separation on a 3%–8% Tris-Acetate polyacrylamide gel (NuPAGE; Invitrogen). For immunostaining, proteins were transferred to a nitrocellulose membrane and detected with polyclonal anti-PitA (1:50,000 diluted) or anti-PitB antiserum (1:20,000).

Negative staining and immunogold electron microscopy

Strains were grown on TSA-blood agar plates (5%-sheep blood, BD) overnight at 37°C and 5% CO₂. Bacteria were resuspended in PBS before being applied to carbon-coated copper grids (300 mesh, EMS), and stained with 1% methylamine tungstane for 30 seconds. Alternatively, strains were applied to carbon-coated nickel grids (300 mesh, EMS) and immunostained according to established protocols [21,70]. Anti-PitA and anti-PitB antiserum were used at a 1:10,000 and a 1:4,000 dilution, respectively, and a 10 nm-gold-conjugated goat anti-mouse IgG was used as secondary antibody at 1:20 dilution. Following the immunogold staining, the grids were negative stained as described above and analyzed using a Hitachi H-7500 transmission electron microscope.

Nucleotide sequence accession numbers

The following GenBank accession numbers were assigned to PI-2 of *S. oralis* ATCC10557: JF496566 and S. oralis 34: JF496567.

Supporting Information

Figure S1 Alignment of PitB proteins of oral Mitis group streptococci and *S. pneumoniae*. PitBs of *S. oralis* Uo5 and *S. sp.* C300 are not shown (identical to PitB of *S. sanguinis* ATCC49296). Only residues different from the consensus sequence are indicated. Sequence positions based on the *S. oralis* ATCC35037 sequence are indicated above the alignment. The Signal peptides (SP) and cell wall sorting signals (CWSS) are underlined and the LPXTG-like motif (VTPTG) is shown in bold. Signal peptides were predicted by the SignalP program. Conserved amino acids involved in intramolecular isopeptide bonds in *S. pneumoniae* PitB are indicated by an asterisk followed by the number for bond 1 and 2, respectively. GenBank accession numbers are as in Fig. 1. Alignment was performed with ClustalW software.

(PPT)

Figure S2 Evidence for past recombination events in PI-2 pilus islets of Mitis group streptococci. A. Schematic overview of PI-2 pilus islets. Sequences of *S. oralis* Uo5 and *S. sp.* C300 are not shown (almost identical to *S. sanguinis* ATCC49296). Gene regions encoding conserved motifs are indicated by a black bar below the gene. SP: Signal peptide, vWA: von Willebrand A domain, CWSS: cell wall sorting signal. Red squares indicate the regions of sudden, major changes in sequence polymorphisms aligned in panels B-E. In panels B-E. Regions of major sequence polymorphisms are indicated in grey. GenBank accession numbers: *S. oralis* strains ATCC35037: AEDW01000020; ATCC10557: JF496566; and 34: JF496567; *S. mitis* ATCC6249: AEEN01000012; *S. sanguinis* ATCC49296: AEPO01000013; and *S. pneumoniae* GA47884: GU256423. Alignment was performed with ClustalW software.

(PPT)

Table S1 Primers used in this study.

(DOC)

Acknowledgments

We thank Dr. Robert Palmer (National Institute of Dental and Craniofacial Research) for providing *S. oralis* strain 34.

Author Contributions

Conceived and designed the experiments: DZ. Performed the experiments: DZ ARG HY DSS. Wrote the paper: DZ DSS.

References

1. Whatmore AM, Efratiou A, Pikerill AP, Broughton K, Woodward G, et al. (2000) Genetic relationships between clinical isolates of *Streptococcus pneumoniae*, *Streptococcus oralis*, and *Streptococcus mitis*: characterization of “Atypical” pneumococci and organisms allied to *S. mitis* harboring *S. pneumoniae* virulence factor-encoding genes. Infect Immun 68: 1374–1382.

2. Kilian M (2005). *Streptococcus and Lactobacillus*. In: Topley and Wilson’s microbiology and microbial infections (eds P Borriello, P R Murray, and G Funke).

3. Kolenbrander PE (2000) Oral microbial communities: biofilms, interactions, and genetic systems. Annu Rev Microbiol 54: 413–437.

Type 2 Pili of Mitis Group Streptococci
27. Konto-Ghiorghi Y, Mairey E, Mallet A, Dumenil G, Caliot E, et al. (2009) Dual
26. Dramsi S, Caliot E, Bonne I, Guadagnini S, Prevost MC, et al. (2006) Assembly
22. Telford JL, Barocchi MA, Margarit I, Rappuoli R, Grandi G (2006) Pili in
20. Mandlik A, Swierczynski A, Das A, Ton-That H (2008) Pili in Gram-positive
16. Budzik JM, Marraffini LA, Schneewind O (2007) Assembly of pili on the surface
31. Swaminathan A, Mandlik A, Swierczynski A, Gaspar A, Das A, et al. (2007)
30. Ton-That H, Schneewind O (2004) Assembly of pili in Gram-positive bacteria.
13. Okahashi N, Nakata M, Sakurai A, Terao Y, Hoshino T, et al. (2010) Pili of oral
12. Yamaguchi T, Matsumoto M, Sugimoto Y, Soutome S, Oho T (2009) Gene
cloning and characterization of S. pneumoniae fimM involve in saliva-mediated
aggregation. Res Microbiol 160: 609–616.
11. Okahashi N, Nakata M, Sakurai A, Terao Y, Hoshino T, et al. (2010) Pili of oral
28. Schneewind O, Model P, Fischetti VA (1992) Sorting of protein A to the
17. Johnston C, Hinds J, Smith A, van der Linden M, Van Eldere J, et al. (2010)
16. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
15. Ronda C, Garcia JL, Lopez R (1988) Characterization of genetic transformation
14. Machi J, Hinds J, Smith A, van der Linden M, Van Eldere J, et al. (2010)
13. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
12. Ronda C, Garcia JL, Lopez R (1988) Characterization of genetic transformation
11. Ronda C, Garcia JL, Lopez R (1988) Characterization of genetic transformation
10. Lauer P, Rinaudo CD, Soriani M, Margarit I, Maione D, et al. (2005) Genome
9. Barocchi MA, Ries J, Zogaj X, Hemsley C, Albiger B, et al. (2006) A
8. Johnston C, Hinds J, Smith A, van der Linden M, Van Eldere J, et al. (2010)
7. Donati C, Hiller NL, Tettelin H, Muzzi A, Croucher N, et al. (2010) Structure
6. Kilian M, Poulsen K, Blomqvist T, Ha ˚varstein LS, Bek-Thomsen M, et al.
5. Ronda C, Garcia JL, Lopez R (1988) Characterization of genetic transformation
4. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
3. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
2. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
1. Doern CD, Burnham CA (2010) It's Not Easy Being Green: the viridans group
62. Jameson MW, Jenkinson HF, Parnell K, Handley PS (1995) Polypeptides associated with tufts of cell-surface fibrils in an oral *Streptococcus*. Microbiology 141: 2729–2738.

63. Willcox MD, Drucker DB (1989) Surface structures, co-aggregation and adherence phenomena of *Streptococcus oralis* and related species. Microbios 59: 19–29.

64. Elliott D, Harrison E, Handley PS, Ford SK, Jaffray E, et al. (2003) Prevalence of Coli-like fibrillar surface proteins among mitis group oral streptococci. Oral Microbiol Immunol 18: 114–120.

65. Tamura K, Peterson D, Peterson N, Sutcher G, Nei M, et al. (2011) MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. Mol Biol Evol. [Epub ahead of print]; doi: 10.1093/molbev/msr121.

66. Ho SN, Hunt HD, Horton RM, Pullen JK, Pease LR (1989) Site-directed mutagenesis by overlap extension using the polymerase chain reaction. Gene 77: 51–59.

67. Tao L, LeBlanc DJ, Ferretti JJ (1992) Novel streptococcal-integration shuttle vectors for gene cloning and inactivation. Gene 120: 105–110.

68. Alloing G, Martin B, Granadel C, Claverys JP (1998) Development of competence in *Streptococcus pneumoniae*: pheromone autoinduction and control of quorum sensing by the oligopeptide permease. Mol Microbiol 29: 73–83.

69. Yoshida Y, Yang J, Peaker PE, Kato H, Bush CA, et al. (2008) Molecular and antigenic characterization of a *Streptococcus oralis* coaggregation receptor polysaccharide by carbohydrate engineering in *Streptococcus gordonii*. J Biol Chem 283: 12654–12664.

70. Yi H, Leunissen JLM, Shi GM, Gutekunst CA, Hersch SM (2001) A novel procedure for pre-embedding double immunogold–silver labeling at the ultrastructural level. Journal of Histochemistry & Cytochemistry 49: 279–283.