Approaching the Secrets of N-Glycosylation in *Aspergillus fumigatus*: Characterization of the AfOch1 Protein

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Abstract

The mannosyltransferase Och1 is the key enzyme for synthesis of elaborated protein N-glycans in yeast. In filamentous fungi genes implicated in outer chain formation are present, but their function is unclear. In this study we have analyzed the Och1 protein of *Aspergillus fumigatus*. We provide first evidence that poly-mannosylated N-glycans exist in *A. fumigatus* and that their synthesis requires AFoch1 activity. This implies that AFoch1 plays a similar role as *S. cerevisiae* ScOch1 in the initiation of an N-glycan outer chain. A ΔAfOch1 mutant showed normal growth under standard and various stress conditions including elevated temperature, cell wall and oxidative stress. However, sporulation of this mutant was dramatically reduced in the presence of high calcium concentrations, suggesting that certain proteins engaged in sporulation require N-glycan outer chains to be fully functional. A characteristic feature of AFoch1 and Och1 homologues from other filamentous fungi is a signal peptide that clearly distinguishes them from their yeast counterparts. However, this difference does not appear to have consequences for its localization in the Golgi. Replacing the signal peptide of AFoch1 by a membrane anchor had no impact on its ability to complement the sporulation defect of the ΔAfOch1 strain. The mutant triggered a normal cytokine response in infected murine macrophages, arguing against a role of outer chains as relevant *Aspergillus* pathogen associated molecular patterns. Infection experiments provided no evidence for attenuation in virulence; in fact, according to our data the ΔAfOch1 mutant may even be slightly more virulent than the control strains.

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Introduction

Eukaryotic proteins that enter the secretory pathway in the endoplasmatic reticulum (ER) are glycosylated by distinct sets of enzymes that catalyze either the formation of N- or O-linked glycans. Glycosylation can influence the folding of proteins, their biological activity and half-life. So far, fungal N-glycans have mainly been analyzed in yeasts, whereas a remarkable diversity exists in the further processing, branching and elongation of this core structure. In yeasts N-glycans are characterized by their distinct and complex high-mannose structures [1].

The Och1 proteins of *S. cerevisiae* and *C. albicans* are α1,6-mannosyltransferases that initiate a distinct branch in the N-glycan core thereby providing the platform for the subsequent formation of a large poly-mannosylated outer chain [2,3]. The Δoch1 mutants of *S. cerevisiae* and *C. albicans* are sensitive to elevated temperatures [2,3]. The *C. albicans* mutant showed furthermore an attenuated virulence in a murine model of infection [3] and triggered a reduced cytokine response in infected murine macrophages, a phenotype that was linked to recognition of outer chains by the mannose receptor [4].

The N-glycosylation pathway has been studied in detail in *S. cerevisiae* [5]. The early steps take place in the cytoplasm and the ER and are essential and highly conserved between yeasts and filamentous fungi. The so-called high mannose pathway is a cascade of enzymes engaged in the synthesis of elaborated N-glycans. The corresponding genes encoding a set of enzymes reaching from Och1 to Mnn1 in *S. cerevisiae* have been identified in filamentous fungi based on sequence homology [6]. However, data on enzyme activity and N-glycan structures in filamentous fungi are still very limited and provided so far no evidence for the existence of elaborated outer chains.

*Aspergillus fumigatus* is a pathogenic mold that can cause severe, life-threatening infections in immunocompromised patients [7]. Recognition of this pathogen by the innate immune system is crucial for its successful abatement. Glycostructures are excellent targets for the germ-line encoded pattern recognition receptors (PRRs) of innate immune cells. We have recently characterized two *A. fumigatus* mutants in the mannosyltransferase genes *mnt1* and *mnt4*. The Mnt1 protein catalyzes an early and essential step in the
formation of O-linked glycans, whereas MitA is required for the synthesis of GDP-mannose:inositol-phosphorylceramide (MIPC)-derived glycosphingolipids. We found no differences in the cytokine response induced in murine macrophages infected with these mutants and their corresponding control strains. This argues against a role of O-linked glycans and MIPC-derived glycosphingolipids as major pathogen-associated molecular patterns (PAMPs) of *A. fumigatus* [8,9]. In this study we generated a mutant in the *afochl* gene, encoding the orthologue of the yeast Och1 proteins, and characterized its phenotype. We provide evidence for a mannosyltransferase activity of this protein and its requirement for sporulation under distinct growth conditions.

**Results**

The *A. fumigatus* gene AFUA_5G08580

The ochl genes of *S. cerevisiae* and *C. albicans* encode α1,6-mannosyltransferases that are essential for the outer chain elongation of N-linked glycans [2,3]. A BlastP search in the *A. fumigatus* genome data base using the ScOch1 sequence revealed the highest homology for AFUA_5G08580. This ORF is predicted to contain one intron. PCR amplification from *A. fumigatus* chromosomal and cDNA using primers ochl-5' and ochl-3' resulted in two amplicons of different size. Sequencing of the PCR product derived from cDNA revealed an ORF that differed from the one predicted in the genome entry AFUA_5G08580. (The product derived from cDNA revealed an ORF that differed from the one predicted in the genome entry AFUA_5G08580. (The sequence of the full length mRNA has been submitted to the EMBL data base: Accession number FR666740.) The fact that the mRNA was amplified using a primer localized 173 bp downstream of the STOP codon indicates the presence of a longer 3'-region, similar to the one predicted for the homologous *A. nidulans* gene AN4716.4-T.

Using the corrected protein sequence AoCh1 shows 33.7% identity (43.4% similarity) and 43.2% identity (56.3% similarity) to the Ochl protein of *S. cerevisiae* and *C. albicans*, respectively. These values are similar to the homology of the two yeast Ochl proteins (37.3% identity; 51.3% similarity). All three sequences share two stretches of high homology: one around the putative glycosyltransferase sugar-binding region (Pfam domain PF04488), comprising a characteristic DXD motif, and a second in the C-terminal region (Supplementary Figure S1).

**Generation and characterization of a Δafoch1 mutant**

A deletion construct was generated that comprises 1 kb regions up- and down-stream of the *afochl* gene flanking a hygromycin resistance cassette (Figure 1A). This linear construct was used for transformation of protoplasts of strain AF293 [10]. PCR analysis of a hygromycin-resistant clone that was used throughout this study is shown in Figure 1B. Single integration of the deletion construct was verified by Southern blot (data not shown). For complementation the mutant was transformed with a construct that comprises the *afochl* gene (under the control of the *A. nidulans* gpdA promoter) and a pyrithiamine resistance cassette. Several pyrithiamine resistant clones were obtained and analyzed by PCR. The results obtained for the clone used in subsequent studies are shown in Figure 1B.

The *Afochl* mutant and the control strains (the parental strain AF293 and the complemented mutant) showed comparable growth and sporulation on AMM, YG and Sabouraud plates at 37°C (Figure 2A–A’, 3A–A’ and data not shown). The *S. cerevisiae ochl* mutant is impaired in growth at elevated temperatures [11] and its *C. albicans* counterpart is moreover sensitive to different cell wall stressors, e.g., calcifluor white and SDS [3]. Growth of the *Afochl* mutant was analyzed under these and additional stress conditions.

We observed no phenotype at temperatures up to 48°C, on YG plates containing different stressors (calcifluor white (200 µg/mL), Congo red (100 µg/mL), SDS (0.05%), sodium deoxycholate (0.1%)), in a disk diffusion assay using 30% H2O2, and in E-tests with voriconazol, caspofungin and amphotericin B (data not shown).

**Impaired sporulation of the Δafochl mutant under high calcium conditions**

We recently described that the *A. fumigatus* mannosyltransferase MitA is essential for synthesis of complex mannos:inositol-phosphorylceramide-derived glycolipids and that the corresponding mutant is impaired in growth in the presence of high calcium concentrations [8]. Under similar conditions (YG agar plates containing 350 mM calcium) the colony size of the Δafochl mutant was only slightly reduced compared to the control strains (Figure 2B-B’), but the mutant colonies showed a striking broad white rim of non-sporulating mycelium (Figure 2B’). No defect in sporulation or growth was detectable on plates containing 550 mM MgCl2 (data not shown), demonstrating that this effect is not due to an increased concentration of divalent or chloride ions in the medium.

Further experiments revealed no sporulation phenotype of the mutant on Aspergillus minimal medium (AMM) plates (containing 1% or 4% glucose) supplemented with up to 300 mM calcium (Figure 2C-C’ and data not shown). This indicates that calcium per se is not sufficient to trigger the sporulation defect. On Sabouraud plates the sporulation phenotype was already evident at 100 mM calcium (Figure 3B-B’) and became even more apparent after 240 h incubation, when colonies of the control strains showed complete area-wide sporulation (Figure 3F-F’). Sabouraud medium comprises 4% glucose, pancreatic digest of casein and peptone (3 g/L each). To determine whether the complex components (that are not present in AMM) are required, we analyzed growth on plates with 100 mM calcium, 4% glucose and either 10 g/L casein digest or peptone. The sporulation defect was only apparent on plates containing casein (Figure 3C-C’), demonstrating a requirement for a so far undefined component present in casein, but not in peptone. A characteristic feature of Sabouraud medium is the acidic pH of 5.6. Adjusting the pH to 7.0 abolished the sporulation phenotype (Figure 3E-E’). Growth of the mutant on AMM plates adjusted to pH values of 5.0, 6.0 and 7.0 revealed no defect in sporulation, demonstrating that a low pH is required but not sufficient to induce the sporulation phenotype of the Δafochl mutant (data not shown).

**Aspergillus fumigatus Och1 is involved in N-glycan biosynthesis**

The Ochl proteins of *S. cerevisiae* and *C. albicans* are α1,6-mannosyltransferases acting on the Man9GlcNAc2 core of N-glycans. In the *C. albicans* Δochl mutant a reduced N-glycosylation of an N-acetyl-glucosaminidase was described [3]. Using sequence homology we identified the putative orthologous *A. fumigatus* N-acetyl-glucosaminidase (AFUA_8G05020) and performed a similar experiment with protein extracts obtained after growth in AMM broth with glucosamine as the sole carbon source. EndoH digestion reduced the apparent molecular weight indicating the presence of N-glycans, but no difference was observed between the Δafochl mutant and the control strains (data not shown).

Continuing along these lines, N-glycans of secreted proteins from *A. fumigatus* wild type, Δafochl and Δafoch1+afoch1 were released by PNGase F treatment and analysed by capillary electrophoresis after fluorescent labelling. As seen in Figure 4 and...
Figure 1. Construction of the Δafoch1 mutant and the complemented strain. (A) Structure of the genomic afoch1 gene and the deleted afoch1::hph/tk locus. Approximately 1 kb of the 5' and 3' regions of afoch1 (gray boxed areas) were used for construction of the deletion cassette. The positions of the primers used for PCR amplifications and the resulting PCR products (PCR 1–4) are indicated. (B) Equal amounts of genomic DNA of AF35, Δafoch1 and Δafoch1 + afoch1 were used as template for PCR amplification of the regions indicated in panel A (PCR 1–4).

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Figure 2. Impaired sporulation of the Δafoch1 mutant in the presence of high calcium concentrations. Colonies of the parental strain AF35 (A, B, C), the Δafoch1 mutant (A', B', C') and the complemented strain (A'', B'', C'') were grown at 37°C on YG medium (A to A''), YG medium supplemented with 350 mM CaCl$_2$ (B to B'') and AMM medium supplemented with 500 mM CaCl$_2$ (C to C '').

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in previous reports [12,13], *A. fumigatus* secreted proteins mainly carry N-glycans composed of the Man$_5$GlcNAc$_2$ core predominantly substituted with a galactofuranose residue. Additional N-glycan structures labelled with an asterisk in Figure 4 are observed when *A. fumigatus* is grown in Sabouraud broth (first electropherogram, panel A) but not in minimal media (first spectrum, panel B). Importantly, these peaks are absent from the Δαfoch1 strain (second electropherogram, panel A) and increased in the complemented mutant (third electropherogram, panel A). In minimal media, these additional N-glycans are only observed if Δαfoch1 is expressed from the strong gdh1 promoter of the complementation construct. Moreover, N-glycans of higher molecular weight are also present when *A. fumigatus* is grown in Sabouraud but not in minimal media broth (first and third electropherogram of the Supplementary Figure S2). The majority of these structures (labelled with an asterisk) are dependent on Δαfoch1 expression. These data clearly indicate the involvement of Δαfoch1 in the biosynthesis of N-glycans. They suggest that synthesis of polymannosylated N-glycans occurs in *A. fumigatus* and might be triggered in response to specific environmental conditions. These results would be in agreement with Δαfoch1 playing a similar role as *S. cerevisiae* ScOch1 in the initiation of an outer chain.

**Infection experiments**

The och1 mutant of *C. albicans* was shown to induce a reduced cytokine response in murine macrophages in comparison to the corresponding wild type strain [4]. Infection of murine, peritoneal macrophages with the Δαfoch1 and the control strains revealed no significant difference in the levels of secreted TNFα, IL-6 or IL-10 (Figure 5A and data not shown). To analyze whether the Δαfoch1 mutant is attenuated in virulence, as reported for its *C. albicans* counterpart [3], we infected mice using a systemic model of infection with immunocompetent mice and an intranasal infection model with immunocompromised mice. We observed no attenuation for the Δαfoch1 mutant in either model of infection (Figure 5B and C). Unexpectedly, infections with the Δαfoch1 led to a significantly faster killing in the systemic model of infection (*p* = 0.002, Figure 5B) and indications of a slightly higher virulence were also observed in the intranasal infection model (Figure 5C).

**Och1-like proteins of yeast and filamentous fungi differ in their N-termini**

Yeasts and filamentous fungi usually harbor several members of the Och1-family. The Hoc1 proteins are homologous to Och1 and build a second, phylogenetically distinct subfamily of putative mannosyltransferases. ScHoc1 resides in the Golgi, but the gene is unable to complement an *S. cerevisiae* Δαoch1 mutant [14]. Another member of the Och1-family, ScOcr1 (Och1-related protein), was shown to contribute to the elongation of N-glycan outer chains and to O-glycosylation of proteins [15]. A phylogenetic tree derived from the protein sequences of Och1, Hoc1 and Ocr1 proteins from *S. cerevisiae*, *F. angusta*, *C. albicans* and *A. fumigatus* is shown in the Supplementary Figure S3. As expected, families of Och1 and Hoc1 proteins are apparent, but apart from Δαfoch1 none of the Och proteins of *A. fumigatus* were allocated to one of these subfamilies. In fact, Δαfoch3 appears to be more closely related to the GDP-mannose:inositol-phosphorylceramide–mannosyltransferase AmMitA than to the other putative or proven 1,6-mannosyltransferases (Supplementary Figure S3).

A closer inspection of the Δαfoch1 polypeptide sequence revealed a remarkable difference to its yeast orthologs. All yeast Och1 proteins are predicted to contain N-terminal membrane anchors, whereas a cleavable signal peptide is predicted for Δαfoch1. We extended this analysis to other ScOch1 homologues. The results are summarized in Figure 6 (more detailed information is given in the Supplementary Table S1). N-terminal membrane anchors are predicted for the Och1-like proteins of *S. cerevisiae*, *C. albicans*, Schizosaccharomyces pombe, *Pichia pastoris*, *Pichia angusta* (formerly: *Hansenula polymorpha*) and *Cryptococcus neoformans*, whereas N-terminal signal sequences are predicted for the Och1 proteins of *A. fumigatus*, *A. nidulans*, *A. niger*, *Neurospora crassa*, Magnaporthe grisea,
Histioplasma capsulatum and Ashbya gossypii. Strikingly, N-terminal membrane anchors are found in yeast, whereas signal peptides are predicted for filamentous fungi. The Och1 protein of Ashbya gossypii is of particular interest. Ashbya gossypii belongs to the Saccharomycetaeae and this relationship is also evident from the phylogenetic tree shown in Figure 6. However, A. gossypii grows only in the filamentous form and AgOch1 harbors an N-terminal signal sequence. In conclusion, these findings suggest a link between filamentous growth and Och1 proteins having a signal peptide.

The AfOch4 protein

In the A. fumigatus Och family signal peptides are only predicted for AfOch1 and AfOch4 (Supplementary Table S2). This prompted us to analyze the putative mannosyltransferase AfOch4 in more detail. Information on the generation and complementation of a Δafoch1 mutant is given in Supplementary Figure S4. The mutant grows normally and showed no phenotype under all conditions tested for the afoch1 mutant including high calcium concentrations (data not shown). Transformation of the Δafoch1 mutant with the afoch4 gene (under the control of the constitutive gpdA promoter) did not rescue the sporulation phenotype on calcium plates (Supplementary Figure S4, panel C) suggesting that AfOch1 and AfOch4 have different biological functions.

Localization of the AfOch1 protein

The ScOch1 protein comprises an N-terminal membrane anchor and is localized in the cis-Golgi (Nakayama et al., 1992). After cleavage of its N-terminal signal peptide, AfOch1 is released into the lumen of the ER and can thereby enter the secretory pathway. To investigate its localization we fused the afoch1 gene in frame to the 5’-end of the rfp gene (encoding the red fluorescent protein). This construct was able to rescue the sporulation defect of the Δafoch1 mutant (data not shown). Microscopic analysis revealed a strong RFP fluorescence that was focused in distinct intracellular compartments (Figure 7A and B). These large spot-like structures disappeared after treatment with Brefeldin A (Figure 7C and D), which is known to disrupt the Golgi apparatus [16]. This demonstrates that AfOch1 is retained in the Golgi although it lacks an N-terminal membrane anchor.

The functional importance of the AfOch1 N-terminus

To determine whether the signal peptide of AfOch1 is required for it to function properly, we exchanged nine amino acids flanking the predicted cleavage site by mutagenesis of the complementation plasmid (Figure 8A). The resulting AfOch1* is predicted to contain a non-cleavable N-terminal anchor sequence (Signal peptide probability: 0.117; Signal anchor probability: 0.883). Transformation of the Δafoch1 mutant with the afoch1* gene rescued the sporulation defect on calcium plates (Figure 8B–D), indicating that the N-terminal signal sequence is not essential for the function of AfOch1.

Discussion

The N-glycosylation of eukaryotic proteins proceeds in several steps: synthesis of the core glycan on the lipid carrier dolichylypyr-
demonstrate that poly-mannosylated N-glycans are formed by \textit{A. fumigatus} under certain circumstances. Evidence for outer chains was obtained for the wild type strain, but not for the mutant, indicating that \textit{AfOch1} is essentially required. Evidence for poly-mannosylated N-glycans was detectable for the wild type after growth in Sabouraud, but not in AMM broth, whereas the complemented strain synthesized these glycostructures in both media. Expression of \textit{och1} in the complementation construct is driven by the constitutive \textit{gpd}A promoter and is therefore not completely restoring the wild type situation. However, this constitutive expression led to the formation of outer chains in AMM broth and thereby provided a valuable hint that expression of \textit{afoch1} from its native promoter is strongly influenced by the growth medium. \textit{AfOch4} a homolog of \textit{AfOch1} is not able to complement the \textit{afoch1} mutant. Whether \textit{AfOch2} or \textit{AfOch3} play a role in the initiation or elongation of N-glycan outer chains remains to be determined.

\textit{A. fumigatus} \textit{afoch1} mutant is impaired in virulence and triggers lower levels of proinflammatory cytokines during infection of murine macrophages [3,4]. Similar infection experiments revealed no difference between the \textit{Afoch1} mutant and the control strains, suggesting that N-glycan outer chains of \textit{A. fumigatus} are no relevant target for murine pattern recognition receptors. Our findings may reflect a fundamental difference between the cell wall of yeast and \textit{A. fumigatus}. A thick layer of mannansylated proteins on the core cell wall is characteristic for the former, but seems to be absent in \textit{Aspergillus}. Since \textit{afoch1} expression is dependent on the growth conditions, \textit{AfOch1} may not be expressed during in vitro infection of macrophages. However, these experiments do not necessarily reflect the situation in the infected host. Analysis of the virulence of the \textit{Afoch1} mutant in two murine models of infection revealed no attenuation. Surprisingly, infection with the mutant led to a faster killing in the systemic model of infection. Whether this truly reflects a slight increase in virulence remains to be determined. For the time being our data at least suggest that \textit{AfOch1} is expressed during infection. To prove this, specific antibodies for the outer chains of \textit{A. fumigatus} N-glycans would be valuable tools.

In contrast to the \textit{Och1} protein of \textit{S. cerevisiae} \textit{AfOch1} harbors a signal sequence that is most likely cleaved off during transport over the ER membrane. This difference prompted us to analyze other members of the \textit{Och1}-family. Strikingly, the \textit{Och1} proteins of the filamentous fungi \textit{A. fumigatus}, \textit{A. nidulans}, \textit{A. niger}, \textit{N. crassa} and \textit{M. grisea} harbor signal sequences, whereas the \textit{Och1} protein from the yeasts \textit{S. cerevisiae}, \textit{S. pombe}, \textit{P. angusta}, \textit{P. pastoris} and \textit{C. albicans} have N-terminal anchor sequences. \textit{Ashbya gossypii} is closely related evolutionarily to \textit{S. cerevisiae}, and both \textit{Och1} proteins are highly homologous, but \textit{AgOch1} harbors a signal peptide. Interestingly, \textit{A. gossypii} grows in hyphal filaments and has been described as “filamentous yeast” [26]. \textit{Candida albicans}, \textit{S. cerevisiae} and \textit{H. capsulata} can switch between a hyphal and a yeast form. The two former have anchor sequences, while the latter is strongly predicted to harbor a signal peptide. \textit{Histoplasma capsulata} commonly grows in filamentous hyphae, but switches to a pathogenic yeast morphotype at 37°C. Hence, it is tempting to speculate that the presence of an N-terminal signal peptide is somehow linked to a hyphal morphology under conventional growth conditions. It is interesting to note that the elaborated outer chains of \textit{S. cerevisiae} and \textit{C. albicans} were all isolated from the yeast morphotype [3,11]. Whether outer chains are abundant in the pathogenic hyphal morphotype of \textit{C. albicans} remains to be determined.

After cleavage of its N-terminal signal sequence, \textit{AfOch1} is released into the lumen of the ER and may travel to the cell
surface. However, enzymes that are required for N-glycan synthesis conventionally reside in the Golgi. They are generally supposed to be membrane-anchored and to interact with luminal target proteins [27]. We have localized an RFP-fusion of AfOch1 in distinct intracellular compartments. The sensitivity of these structures to Brefeldin A strongly suggests that they represent the Aspergillus Golgi apparatus. Proper localization of the fusion protein was verified by its ability to complement the sporulation defect of the \(A.\) fumigatus \(D_{afoch1}\) mutant. In yeast, the localization of ScOch1 in the Golgi involves retrograde transport of this membrane anchored protein [28]. However, similar mechanisms operate with other Golgi mannosyltransferases, e.g., ScHoc1, which harbors a signal peptide [14]. Thus, so far unknown proteins may assist ScHoc1 and AfOch1 to remain at the membrane of the Golgi. A mutated AfOch1* protein that lacks the cleavage site and is predicted to harbour an N-terminal membrane anchor instead is able to rescue the sporulation defects of the \(D_{afoch1}\) mutant. These data indicate redundant sorting mechanisms that assure that Golgi proteins remain in this organelle, which is also suggested by the finding that a mutated form of ScMnn1 lacking its N-terminal membrane anchor is retained in the Golgi [29]. It is possible that the conserved stretch of amino acids in the C-terminal part of AfOch1 is involved in protein-protein interactions that mediate this sorting mechanism.

In conclusion, we provide evidence that AfOch1 is expressed in \(A.\) fumigatus and that this expression depends on the growth medium. Under permissive conditions outer chains are detectable in the N-glycans of \(A.\) fumigatus. This strongly suggests that AfOch1, like its yeast counterparts, is an \(a_1,6\)-mannosyltransferase that initiates the branching of the outer chain. A mutant lacking this enzyme showed no phenotype in plate assays under standard and stress conditions. However, it is strongly impaired in sporulation in the presence of high calcium concentrations. The precise mechanism underlying this phenotype is unknown, but it
provides a convenient mean to analyze AfOch1 activity. In AfOch1 a signal sequence replaces the N-terminal membrane anchor that is characteristic for Och1 proteins in yeast. This seems to be generally the case in filamentous fungi, suggesting a link between the growth in filamentous hyphae and the presence of an N-terminal signal sequence in Och1 proteins. Why Och1 proteins differ in their N-termini and how this correlates to a different abundance of elaborated outer chains in yeast and filamentous fungi is still an open question, but our results provide a base to analyze this in more detail.

Materials and Methods

Strains and culture conditions

The *A. fumigatus* ΔakaA strain AfS35, the procedure for isolation of conidia, and the composition of the yeast glucose medium (YG) and the Aspergillus Minimal Medium (AMM) have been described previously [10,30]. Sabouraud medium, casein and pepton were purchased from Becton-Dickinson.

Sequence analysis and database searches

Database searches were performed using BlastP and BlastN and the following databases: GeneBank/EMBL/DDJB, the Central Aspergillus Data Repository (CADRE) (http://www.cadre-genomes.org.uk/Aspergillus_fumigatus/), the Saccharomyces Genome Database (http://www.yeastgenome.org/), and the Candida Genome Database (http://www.candidagenome.org/). Predictions of signal sequences were performed using SignalP 3.0 (http://www.cbs.dtu.dk/services/SignalP/). N-glycosylation sites were predicted using NetNGlyc 1.0 (http://www.cbs.dtu.dk/services/NetNGlyc/). Predictions for O-β-GlcNAc attachment sites were performed using YinOYang (http://www.cbs.dtu.dk/services/YinOYang/). Sequence homologies were analyzed using the EMBoss Pairwise Alignment Algorithms (http://www.ebi.ac.uk/Tools/emboss/align/). Prediction of protein domains was performed using InterProScan Search (http://www.ebi.ac.uk/Tools/InterProScan/). Phylogenetic trees were obtained using CLUSTAL (http://www.ebi.ac.uk/Tools/clustalw2/).

Genetic nomenclature

At a meeting in Copenhagen in 2004 the *Aspergillus fumigatus* genome sequencing group proposed a nomenclature based on the Demerec bacterial system that should be used for all species (i.e. *abdD* for genes). In our manuscript we followed their recommendations.

Construction of the Δafoch1 and Δafoch4 mutants and the complemented strains

All oligonucleotides used in this study are summarized in Supplementary Table S3. The *afoch1*/AFUA_5G08580 gene was amplified from cDNA using oligonucleotides och1-5′ and och1-3′. The PCR product was cloned, sequenced and submitted to the database.

To construct a suitable replacement cassette a 3.5 kb hygromycin resistance cassette was excised from pSK346 using the SfiI restriction enzyme. The flanking regions of the *afoch1* gene (approx. 1.0 kb each) were amplified by PCR from chromosomal DNA using the oligonucleotide pairs och1-upstream-5′/och1-upstream-3′ and och1-downstream-5′/och1-downstream-3′. These oligonucleotides harbored incompatible SfiI sites. After digestion with SfiI a ligation of the three fragments (resistance cassette and flanking regions) yielded a 5.4 kb deletion cassette that was purified using the PrepEase Gel Extraction Kit (USB, Cleveland, USA). The fragment was cloned into the pBluescriptKS vector (Stratagene, La Jolla, USA) using oligonucleotide derived NotI sites. Purified NotI fragments from the resulting plasmid were used for transformation.

For complementation the *afoch1* gene was amplified using oligonucleotides och1-5′ and och1-3′. The PCR product was purified and cloned into pSK379 to drive expression from the *gpdA*
promoter. The resulting plasmid was isolated using the Pure Yield Plasmid MidiPrep System (Promega, Mannheim, Germany). A. fumigatus protoplasts were generated and transformation was performed essentially as described previously [9]. The resulting protoplasts were transferred to AMM plates containing 1.2 M sorbitol and either 200 μg/ml hygromycin (Roche, Applied Science, Mannheim, Germany) or 0.1 μg/ml pyrithiamine (Sigma-Aldrich, Deisenhofen, Germany).

The generation of the AfOch1 mutant was performed accordingly using oligonucleotides summarized in Supplementary Table S3.

Genomic DNA analysis
A. fumigatus clones which showed the expected resistance on selective plates were further analyzed by PCR. In the first PCR, one oligonucleotide (och1-cast-5’) that hybridized immediately upstream of the afoch1 gene was combined with a second primer (och1-3’) localized at the 3’ end of the afoch1 gene (compare Fig. 1). This reaction (PCR1) was used to detect the afoch1 gene in its wild type context. The core afoch1 gene was amplified using oligonucleotides och1-5’ and och1-3’ (PCR2). The correct integration of the deletion cassette was analyzed at the 5’ end using oligonucleotides och1-cast-5’ and hph-3-SmaI (PCR3) and at the 3’ end using oligonucleotides trpCt-fwd and och1-3’UTR-rev (PCR4) (Fig. 1).

Mutation of the 5’-end of the afoch1 gene corresponding to the cleavage site of the signal peptide was performed using the above described complementation plasmid. An inverse long range PCR using the oligonucleotides och1-TMD-5’ and och1-TMD-3’ and the Long Amp Taq Polymerase (New England Biolabs) was performed according to the instructions of the vendor. The resulting 9554 bp fragment was subsequently purified and blunt-ended using T4 DNA Polymerase (Fermentas). After phosphorylation the fragment was ligated given rise to a mutated complementation vector. After sequencing this vector was introduced into the Δafoch1 mutant.

Phenotypic testing on plates
Isolated conidia were counted using a Neubauer chamber. For drop dilution assays, series of tenfold dilutions derived from a starting solution of 1×10⁷ conidia per ml were spotted in aliquots of 3 μl onto plates. These plates were supplemented with the indicated agents and incubated at the indicated temperatures. For quantification of the radial growth, 3 μl containing 1×10⁶ conidia were spotted in the center of a 9 cm Petri dish. The diameter of the colonies was determined over time.

E-test strips of voriconazole, amphotericin B and caspofungin were obtained from Inverness Medical (Cologne, Germany) and used as described previously [9].

N-glycan analysis
N-glycan preparation and separation was carried out essentially as described previously [31]. Briefly, glycoproteins from 1 mL supernatant of A. fumigatus cultures grown for 3 days at 37°C were transferred to Immobilon P Multiwell plates (Millipore). After N-glycan release with peptide:N-glycanase (PNase F, New England Biolabs) in 50 mM NH₄HCO₃ pH 8.4 and labelling with 8-amino-1,3,6-pyrenetrisulfonic acid, N-glycans were separated on a capillary electrophoresis DNA Sequencer (ABI PRISM® 9100-Avant Genetic Analyzer, Applied Biosystems, Foster City, CA, USA) using an injection time of 30 to 90 s. Reference glycans were purchased from Dextra Laboratories (Reading, UK).

Construction and analysis of strains expressing an AfOch1-RFP fusion proteins
The afoch1 gene from nucleotide 1 to 1053 was amplified using oligonucleotides och1-5’ and och1-3’ b. The resulting 1053 bp fragment was cloned into pJW101 using the PmeI restriction site. This derivative of pSK379 comprises the gene for the monomeric red fluorescence protein 1 (mRFP1) [9]. The resulting plasmid pJW102 was transformed into A. fumigatus protoplasts as described above. To test whether the AfOch1-RFP-containing organelles are sensitive to Brefeldin A, hyphae grown in minimal medium (AMM) at 37°C were incubated for 4 h in the presence of 20 μg/ml BrefeldinA (Sigma, Deisenhofen, Germany). Microscopic analysis was performed using a SP-5 confocal laser scanning microscope (Leica Microsystems, Heidelberg, Germany).

Infection of isolated murine macrophages
Primary resting murine macrophages were isolated by peritoneal lavage of C57Bl/6 mice using standard procedures. The subsequent cultivation was performed using RPMI1640 medium supplemented with 5% fetal calf serum. Cells were seeded in 96 well plates at a density of 2.5×10⁵ cells per well. On the next day, the cultures were infected with 2.5×10⁵ conidia per well. After 15 h, the supernatants were harvested and concentrations of TNFα, IL-10 and IL-6 were determined using the Cytometric Bead Array (Becton Dickinson, Heidelberg, Germany) according to the instructions of the vendor.

Mouse infection experiments
All animal experiments were performed in accordance to the national regulations in Germany. Analysis of the mutant in a systemic model of infection was essentially performed as described previously (Wagener et al., 2006). Briefly, 2×10⁶ conidia of the Δafoch1 mutant, the complemented mutant or the parental strain As35 in a final volume of 300 μl PBS containing 0.02% Tween 20 were injected retroorbitally into male CD-1 mice. Survival of infected animals was monitored once a day.

Alternatively, we used an intranasal infection model with immunocompromized female outbred CD-1 mice. Briefly, mice were immunosuppressed by intraperitoneal injection of cortisone acetate (25 mg/mouse; Sigma-Aldrich) on days −3 and 0. On day 0 the mice were anesthetized with fentanyl (0.06 mg/kg, Janssen-Cilag, Germany), midazolam (1.2 mg/kg, Roche, Germany) and medetomidin (0.5 mg/kg, Pfizer, Germany) and infected intranasally with 1×10⁶ conidia in 20 μl PBS. Controls received PBS only. Survival was monitored for 14 days. During this period, mice were examined clinically at least twice daily and weighted individually every day.

Kaplan-Meier survival curves were compared using the log rank test (SPSS 15.0 software). P values <0.05 were considered statistically significant.

Supporting Information
Figure S1 Sequence alignment of the Och1 proteins of Saccharomyces cerevisiae (ScOch1; YGL038C), Candida albicans (CaOch1; orf19.7391) and A. fumigatus (AfOch1; AFUA_5G08580).

Figure S2 The presence of larger N-glycans in AfOch1 mutant.

Figure S3 Characterization of the AfOch1 Protein.

Figure S4 The presence of larger N-glycans in A. fumigatus requires Och1 expression and depends on culture conditions.

Electropherograms of fluorescently labelled N-glycans enzymatically released from secreted glycoproteins of A.
Aspergillus fumigatus parental strain AfS35, Δafoch1 and Δafoch1 + afoch4 strains grown either in Sabouraud broth or Aspergillus minimal media. The grey bar indicates the migration of the Man5GlcNAc2 core N-glycan used as standard (M9). Asterisks indicate ΔOch1 dependent N-glycans. (TIF)

**Figure S3** Probability of N-terminal signal sequences and membrane anchors of selected proteins of the Och1 family from *A. fumigatus*, *S. cerevisiae*, *C. albicans* and *P. angusta*. The sequences were analyzed using the SignalP 3.0 algorithm. The accession numbers of probabilities for a signal sequence or a membrane anchor is given in Panel A as the homologues to the reference ScOch1 protein sequence are given. (DOC)

**Table S1** Probability of N-terminal signal sequences and membrane anchors of Och1 proteins. The Och1 sequences of the indicated fungal species were analyzed using the SignalP 3.0 algorithm. The accession numbers of the analyzed sequences and their homology to the reference ScOch1 protein sequence are given. (DOC)

**Table S2** Probability of N-terminal signal sequences and membrane anchors for members of the Och1 family of *A. fumigatus*, *S. cerevisiae*, *C. albicans*, and *P. angusta*. The Och1 sequences of the indicated fungal species were analyzed using the SignalP 3.0 algorithm. The probabilities of N-terminal signal peptides and membrane anchors, as well as the homologies to the reference ScOch1 protein sequence are given. (DOC)

**Table S3** Oligonucleotides used in this study. (DOC)

**Author Contributions**

Conceived and designed the experiments: FE, FHR. Performed the experiments: AK, JW, JE, BJ. Analyzed the data: FE, FHR, JH. Wrote the paper: FE. 

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