Background

In recent years, there has been a greater emphasis placed on the use of small animal disease models during the early phases of novel drug and diagnostic biomarker development in molecular imaging with positron emission tomography (PET). However, an increasing body of evidence indicates that small animal models should be used with care [1]. The metabolic, pharmacokinetic, and toxicological data obtained in small animals should be extrapolated to other species or to humans rather carefully. Species differences may result in significant differences between the efficacy and, consequently, the safety of the same drug in the various species. These differences may stem from various factors, including species differences in drug metabolism, resulting from the genetic variations in drug-metabolizing enzymes, which in turn may alter pharmacokinetics, drug clearance, drug efficacy, and safety [2]. Another possible factor resulting in species differences

Abstract

Background: We have explored the possibility that the serotonin 1B receptor radioligand \([^{11}C]AZ10419369\) is a substrate for adenosine triphosphate (ATP)-binding cassette (ABC) transporters, such as P-glycoprotein (P-gp), Mrp4, and Bcrp, in rodents and whether there is a species difference regarding its blood-brain barrier (BBB) penetration.

Methods: In a series of preclinical positron emission tomography measurements, we have administered \([^{11}C]AZ10419369\) to mice, rats, and guinea pigs under baseline conditions and, on separate experimental days, after administration of the ABC transporter inhibitor, cyclosporin A (CsA).

Results: During baseline conditions, the brain uptake was low in mice and rats, but not in guinea pigs. After CsA pretreatment, the peak whole brain uptake values of \([^{11}C]AZ10419369\) increased by 207% in mice, 94% in rats, and 157% in guinea pigs. Binding potentials (BP\(_{ND}\) ) could not be estimated during baseline conditions in mice and rats. After CsA pretreatment, the highest BP\(_{ND}\) values were obtained in the striatum and thalamus (BP\(_{ND}\) ≈ 0.4) in mice, while in rats, the highest binding areas were the striatum, thalamus, hypothalamus, and periaqueductal gray (BP\(_{ND}\) ≈ 0.5). In guinea pigs, we did not find any significant changes in BP\(_{ND}\) between baseline and CsA pretreatment, except in the striatum.

Conclusions: The results indicate that BBB penetration of \([^{11}C]AZ10419369\) was hindered by ABC transporter activity in mouse, rat, and guinea pig. This study highlights the importance of ABC transporters in the design of preclinical positron emission tomography (PET) studies.

Keywords: Positron emission tomography (PET); \([^{11}C]AZ10419369\); Serotonin 1B (5-HT\(_{1B}\) ) receptor; Cyclosporin A (CsA); Blood-brain barrier (BBB); ATP-binding cassette transporters (ABC transporters); Mouse; Rat; Guinea pig; Species difference
is related to the differences in the transport of drug molecules (i) in the periphery in organs such as the liver, kidney, and gastrointestinal tract or (ii) in the central nervous system across the blood-brain barrier (BBB). Differences in BBB penetration can have a serious impact on the usefulness and predictive value of small animal models in the drug development process [3].

The transport of molecules across the BBB is partly passive (diffusion, based on concentration gradient differences) and is partly active (based on various transporter molecules and mechanisms) [4]. The most important transporters belong to the adenosine triphosphate (ATP)-binding cassette (ABC) superfamily. In the brain, their role is distinctive in the transport across the BBB. In the human genome, 48 genes encode the ABC transporters, which, based on sequence homologies, are divided into seven subfamilies (designated ABCA-ABCG) [5,6]. Several transporters from the ABC family are expressed in the capillaries of the rat brain. ABCB1, ABCC4, and ABCG2 are also present, but out of these, the ABCB1 subfamily encoding P-glycoproteins (P-gp; also called multidrug resistance 1 (Mdr1a)) is the most abundant. In rats, the expression of P-gp is approximately 4 times higher than that of Bcrp (ABCG2) and 12 times higher than that of Mrp4 (ABCC4) [7]. In humans, however, it has been found that Bcrp has a higher expression level than P-gp [8].

P-gp is a 170-kDa lipoprotein and is a well-characterized member of the ABC family. P-gp is widely expressed in plasma cell membranes across all vertebrate species and is upregulated in multidrug-resistant tumors [5,6]. In the BBB, P-gp contributes to the control of the pharmacokinetics of various compounds. It has been suggested that P-gp-mediated drug efflux may play a decisive role in the development and maintenance of various disorders, including, among others, multidrug-resistant cancer and drug-resistant epilepsies [9]. Differences in P-gp expression across different species have been reported for rat, guinea pig, and monkey [10].

[11C]AZ10419369 was an 11C-radiolabelled analogue of a serotonin 1B (5-HT1B) candidate drug, AZ10419369, and a novel radioligand developed for the selective visualization of the 5-HT1B receptors with PET [11,12]. Whereas the radioligand [11C]AZ10419369 has been demonstrated to be a good marker of drug-induced occupancy at central 5-HT1B receptors in the primate brain [13,14], initial measurements showed poor brain penetration in rats (unpublished data). In the present study, we set out to explore the possibility that [11C]AZ10419369 is a substrate for ABC transporters in rodents and, consequently, if there is a species difference regarding its BBB penetration.

The main objective of the present investigation was to explore the differences in BBB transport of [11C] AZ10419369, with special regard to the ABC transporter activities, in three rodent species widely used in preclinical drug development studies: mouse, rat, and guinea pig. PET measurements were performed after i.v. injection of [11C]AZ10419369 under baseline conditions as well as after pretreatment with cyclosporin A (CsA), an inhibitor of ABC transporters.

**Methods**

**Animals**

Eight male C57BL/6 mice, seven male Wistar rats, and seven male Dunkin-Hartley guinea pigs were used in the study. The animals were housed in groups in individually ventilated cages in a thermoregulated (approximately 22°C), humidity-controlled facility under a 12-h/12-h light/dark cycle with access to food and water ad libitum.

All animal experiments were conducted according to the guidelines of the Swedish National Board of Laboratory under a protocol approved by the Ethics Review Board of Northern Stockholm, Sweden (N210/10, N557/11).

**Radiochemistry and drugs**

[11C]AZ10419369 was prepared at Karolinska Institutet by N-methylation of the desmethyl precursor (8-(1-piperazinyl)-5-methylchrom-2-en-4-one-2-(4-morpholino-phenyl) carboxamide, AstraZeneca R&D, Wilmington, DE, USA), using [11C]methyl triflate, as described in detail earlier [11]. CsA (50 mg/mL, formulated in 96% ethanol) was purchased from Novartis Sverige AB (Täby, Sweden).

**In vivo imaging with PET**

On the experimental day, the animals were anesthetized by inhalation of isoflurane (4% to 5% isoflurane in 100% oxygen). After induction of anesthesia, the isoflurane concentration was lowered to 1.5% to 2% (50:50 air/oxygen) and a cannula was inserted in the tail vein in mouse and rat and in the tarsal vein in the case of guinea pig. The animals were positioned in the PET camera in transaxial position with their heads in the center of the field of view. [11C]AZ10419369 was intravenously administered simultaneously with the start of the PET acquisition and was followed by a 0.1-mL saline flush.

Eight male C57BL/6 mice (28.3 ± 0.8 g) were intravenously injected with [11C]AZ10419369 twice, 1 week apart. The average injected radioactivity was 10.8 ± 0.9 MBq (n = 16), and the average injected mass was 0.025 ± 0.024 μg (n = 16). Seven male Wistar rats (322 ± 46 g) were intravenously injected with [11C]AZ10419369 twice, 1 week apart. The mean injected radioactivity was 21.5 ± 2.7 MBq (n = 14), and the average injected mass is not available. Seven male guinea pigs (514.4 ± 146.9 g) were intravenously injected with [11C]AZ10419369 twice, 1 week apart. The average injected radioactivity was 16.8 ± 3.2 MBq (n = 14), and the average injected mass was 0.020 ± 0.022 μg (n = 14). Upon completion of the first imaging session, the animal returned to its cage. Before the start of
the second PET measurement, the animals were pre-
treated with 50 mg/kg CsA, administered intravenously
15 min before the injection of the radioligand through the
same cannula as used for radioligand injection.

Imaging system and reconstruction
The PET measurements were acquired using the Mediso
nanoScan® PET-MRI and the nanoScan® PET-CT small
animal imaging systems (Mediso Ltd., Budapest, Hungary)
[15,16]. Two animals were examined at the same time in
the identical PET modules of the two PET systems. The
acquired list mode data was reconstructed into 25 time
frames (63-min scan = 4 × 10 s, 4 × 20 s, 4 × 60 s, 7 × 180 s,
6 × 360 s) in the case of mice and rats and 35 time frames
(93-min scan = 4 × 10 s, 4 × 20 s, 4 × 60 s, 7 × 180 s, 11 ×
360 s) in the case of guinea pigs.
The image reconstruction was made with a fully three-
dimensional maximum-likelihood expectation maximization
(MLEM) algorithm with 20 iterations, without scatter and
attenuation correction.

Data analysis
The reconstructed dynamic mouse and rat PET images
were co-registered to the inbuilt mouse and rat MRI tem-
plates available in PMOD, which also incorporates volume
of interest (VOI) sets for mice and rats (version 3.3, PMOD
Technologies Ltd., Zurich, Switzerland). Co-registration
was done manually with the use of mean PET images.
For the guinea pig scans, a template MRI with an in-house de-
developed VOI set was used to co-register with the PET scans.
With the help of these VOI sets, decay-corrected time ac-
tivity curves (TAC) were generated. Whole brain and re-
gional brain uptake values were expressed as percent
standard uptake value (%SUV), which was calculated by
normalizing regional radioactivity for injected radioactivity
and body weight. Uptake values were then calculated as a
summation of uptake from 10 min to the end of the PET
acquisition (total 53 min in the case of rats and mice and
113 min in the case of guinea pigs).
As the %SUV is considered a semi-quantitative value,
quantitative image analysis was additionally performed
using PMOD. The non-displaceable binding potential
(BP_{ND}) was estimated using the simplified reference tissue
model (SRTM) [17] using the cerebellum as reference re-
gion (mice and rats) or cerebellar gray matter (guinea
pigs). This is a validated quantitative analysis method for
[^{11}C]AZ10419369 binding in humans [13] and has been
used in non-human primates [14]. Statistical analyses were
performed with Student’s two-tailed, paired t-test.

Results
At baseline conditions, the guinea pigs showed distin-
guishable brain uptake compared to the surrounding tis-
sue, whereas low brain uptake was observed in the rats
and mice (Figure 1). There was a statistically significant
increase in %SUV after CsA pretreatment compared to
baseline images in all regions, for all animal species
(Table 1). Peak whole brain uptake values increased by
207% in mice, 94% in rats, and 157% in guinea pigs after
CsA pretreatment.
A fast clearance of[^{11}C]AZ10419369 was demon-
strated in mice and rats, while there was a slow washout
in guinea pigs in both the baseline and after pretreat-
ment with CsA (Figure 1). During baseline conditions,
peak brain uptake was at 0.4 min in mice and rats and
at 84 min in guinea pigs. After CsA administration, the
peak brain uptake in mice was at 1.5 min and at 1.2 min
in rats. However, in guinea pigs,[^{11}C]AZ10419369 did not
reach equilibrium during the 2-h-long PET acquisi-
tion in all regions examined besides the cerebellum.
As the initial counts in brain were very low in mice
and rats at baseline condition, the binding potential of
[^{11}C]AZ10419369 could only be estimated reliably in
guinea pigs. After administration of CsA, the brain up-
take had increased sufficiently, and therefore, we quanti-
fied BP_{ND} in all three species (Table 1).
After CsA pretreatment, the highest BP_{ND} values were
found in the striatum and thalamus in mice (BP_{ND} = 0.4).
In rats, the highest BP_{ND} values were observed in the stri-
atum, thalamus, hypothalamus, and periaqueductal gray
(BP_{ND} = 0.5). In guinea pigs, the cerebellar gray matter
was selected as the reference region instead of the whole
cerebellum since there was a relative high concentration
of[^{11}C]AZ10419369 in the white matter. We found no
changes in BP_{ND} between baseline and CsA pretreat-
ment in guinea pigs except in the striatum where a
significant decrease in binding (BP_{ND} 3.1 to 2.4) was
observed. This is probably due to the fact that none of
the regions could reach equilibrium during our experi-
ments making the SRTM calculations unreliable despite
the 2-h scan.

Discussion
An increasing number of recent observations [10,18,19]
indicate that the brain concentration of a drug can differ
significantly among various species. These species differ-
ences can be traced back to various factors, including
differences in plasma binding, metabolic enzyme activ-
ities, or BBB transport. As several biologically effective
compounds may be substrates of efflux transporters, like
P-gp, one of the most important mechanisms behind
species differences in drug disposition in the brain can
be traced back to this phenomenon, i.e., that a CNS drug
is a substrate of P-gp in a given species.
Although CsA had been mostly used as a P-gp inhibi-
tor in relation to the brain, in the periphery, it has been
shown to inhibit transporters from the solute carrier
membrane transport protein group as well [20,21]. Since
brain exposure of any molecule is a function of peripheral distribution, brain penetration, and metabolism, this fact has to be taken into consideration while assessing the results. Metabolic differences under different pretreatment conditions could be a critical factor in brain uptake levels, since non-identical free fraction of \([11C]AZ10419369\) in the blood leads to different availability of radioligand for transport across the BBB. Arterial blood sampling is

**Figure 1** PET images and TAC curves of \([11C]AZ10419369\) uptake in mouse, rat, and guinea pig. Representative uptake images (left) and time activity curves of striatal and cerebellar uptake (right) of \([11C]AZ10419369\) in mouse (top), rat (middle), and guinea pig (bottom) in baseline and pretreatment conditions. The images are a summation from 10 min until the end of the scan (total 53 min in the case of rat and mice and 113 min in the case of guinea pig), while TAC curves are presented as mean values and SD as error bars in mice \((n = 8)\), in rats \((n = 7)\), and in guinea pigs \((n = 7)\).
generally used in PET experiments to measure changes in the free fraction over time. This is a rather difficult task in rodents, particularly in mice, in which the required blood volume would exceed the total blood volume of the animal. The effects of blood loss from arterial blood sampling could reduce the blood volume to such extent that it may alter metabolism, influence Vt, or even cause severe health issues which could lead to exsanguination and consequently to the death of the animal.

While the effects of CsA on the periphery could have been assessed with arterial blood measurement, we did not perform this for the aforementioned reasons. These effects can be avoided by using various transporter knockout animal models. In a complementary study using Sprague-Dawley rats and Mdr1a knockout animals, we have examined the difference in brain uptake and 5-HT1B receptor availability with [11C]AZ10419369 microPET. Increased uptake and [11C]AZ10419369 BPND were observed in KO rats compared with WT in some brain regions, suggesting that the brain uptake of [11C]AZ10419369 is influenced at least by Mdr1a function (see Additional file 1). Earlier PET studies have indicated that the brain uptake of various P-gp substrates may differ markedly between rodent and primate species [10,22-24]. It has also been demonstrated that there can be species difference in BBB transport between various rodent species [10,25]. This fact can be substantiated by the comparison of amino acid content in P-gp in rats, mice, and guinea pigs: the amino acid homology between mouse and rat is 93%, between mouse and guinea pig is 82%, and between rat and guinea pig is 80% [10].

The experiments with mice and rats showed that the rather low brain uptake of [11C]AZ10419369 during baseline condition could be increased significantly with the use of CsA. The brain uptake increased significantly in all species, and the binding potential became measurable after CsA pretreatment in mice and rats. In guinea pigs, the uptake of [11C]AZ10419369 was increased

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### Table 1 Comparison of regional uptake and binding potential in the different brain regions

| Region          | Mouse Baseline | CsA | P value | Rat Baseline | CsA | P value | Guinea pig Baseline | CsA | P value |
|-----------------|----------------|-----|---------|--------------|-----|---------|---------------------|-----|---------|
| %SUV            | 33.4 ± 2.4     | 98.6 ± 10.8 | 0.0000 | 37.4 ± 4.1   | 119.0 ± 29.3 | 0.0002 | 259.6 ± 34.6       | 643.1 ± 192.4 | 0.0020 |
| Cortex          | 30.9 ± 2.6     | 95.1 ± 10.6 | 0.0000 | 38.7 ± 4.2   | 113.0 ± 26.7 | 0.0002 | 213.3 ± 27.3       | 566.9 ± 198.1 | 0.0027 |
| Striatum        | 28.6 ± 2.4     | 110.4 ± 12.1 | 0.0000 | 37.0 ± 5.8   | 146.9 ± 43.5 | 0.0003 | 394.4 ± 58.7       | 768.3 ± 299.9 | 0.0205 |
| Thalamus        | 29.3 ± 2.2     | 114.4 ± 14.1 | 0.0000 | 36.2 ± 5.3   | 145.5 ± 41.8 | 0.0003 | 301.5 ± 44.9       | 792.1 ± 216.4 | 0.0015 |
| Hypothalamus    | 40.3 ± 2.3     | 107.8 ± 10.3 | 0.0000 | 41.2 ± 5.9   | 143.6 ± 39.7 | 0.0003 | 354.9 ± 60.9       | 794.3 ± 204.1 | 0.0019 |
| Hippocampus     | 28.0 ± 2.2     | 102.6 ± 11.0 | 0.0000 | 35.6 ± 4.6   | 137.3 ± 40.1 | 0.0003 | 228.3 ± 31.7       | 605.7 ± 171.1 | 0.0011 |
| Central canal-PAG | 27.1 ± 1.8  | 106.3 ± 13.4 | 0.0000 | 37.4 ± 5.3   | 147.4 ± 38.7 | 0.0002 | 446.2 ± 71.8       | 986.5 ± 248.0 | 0.0027 |
| Colliculus inferior | 28.0 ± 2.6  | 107.1 ± 12.9 | 0.0000 | 36.7 ± 4.1   | 138.7 ± 38.0 | 0.0003 | 192.7 ± 30.3       | 900.5 ± 264.7 | 0.0005 |
| Colliculus superior | 28.1 ± 2.9  | 94.1 ± 10.2 | 0.0000 | 34.4 ± 4.3   | 127.9 ± 32.2 | 0.0002 | 428.4 ± 69.7       | 1 072.5 ± 291.8 | 0.0018 |
| Midbrain        | 30.8 ± 2.4     | 111.5 ± 12.5 | 0.0000 | 37.4 ± 4.3   | 143.6 ± 36.7 | 0.0002 | 409.5 ± 67.6       | 970.1 ± 245.5 | 0.0018 |
| Cerebellum      | 33.7 ± 3.1     | 81.1 ± 8.6  | 0.0000 | 34.0 ± 3.3   | 96.4 ± 18.3  | 0.0001 | 162.7 ± 24.8       | 467.5 ± 135.8 | 0.0006 |
| Cerebellum GM   | -              | -           | -       | -            | -           | -       | 120.8 ± 17.4       | 372.6 ± 122.1 | 0.0009 |

BPND

| Region          | Mouse Baseline | CsA | P value | Rat Baseline | CsA | P value | Guinea pig Baseline | CsA | P value |
|-----------------|----------------|-----|---------|--------------|-----|---------|---------------------|-----|---------|
| Whole brain     | 0.19 ± 0.04    | n/a | -       | 0.21 ± 0.10  | n/a | -       | 1.56 ± 0.39         | 1.57 ± 0.44 | n.s. |
| Cortex          | 0.12 ± 0.04    | n/a | -       | 0.15 ± 0.10  | n/a | -       | 1.00 ± 0.23         | 1.23 ± 0.30 | n.s. |
| Striatum        | 0.39 ± 0.08    | n/a | -       | 0.48 ± 0.21  | n/a | -       | 3.13 ± 0.90         | 2.36 ± 0.52 | 0.0224 |
| Thalamus        | 0.39 ± 0.16    | n/a | -       | 0.48 ± 0.18  | n/a | -       | 2.05 ± 0.55         | 2.29 ± 0.72 | n.s. |
| Hypothalamus    | 0.30 ± 0.06    | n/a | -       | 0.45 ± 0.15  | n/a | -       | 2.76 ± 0.74         | 2.81 ± 1.04 | n.s. |
| Hippocampus     | 0.25 ± 0.11    | n/a | -       | 0.39 ± 0.18  | n/a | -       | 1.25 ± 0.29         | 1.38 ± 0.38 | n.s. |
| Central canal-PAG | 0.15 ± 0.17  | n/a | -       | 0.50 ± 0.11  | n/a | -       | 4.06 ± 1.36         | 5.00 ± 3.04 | n.s. |
| Colliculus inferior | 0.17 ± 0.19  | n/a | -       | 0.42 ± 0.12  | n/a | -       | 3.08 ± 0.86         | 4.00 ± 1.54 | n.s. |
| Colliculus superior | 0.13 ± 0.06  | n/a | -       | 0.27 ± 0.14  | n/a | -       | 3.50 ± 1.15         | 3.65 ± 1.11 | n.s. |
| Midbrain        | 0.26 ± 0.20    | n/a | -       | 0.46 ± 0.11  | n/a | -       | 3.34 ± 1.01         | 3.40 ± 1.17 | n.s. |
| Cerebellum      | -              | -   | -       | -            | -   | -       | 0.52 ± 0.16         | 0.49 ± 0.44 | n.s. |

Regional uptake and binding potential of [11C]AZ10419369 in the different brain regions in mice, rats, and guinea pigs during baseline and CsA pretreatment conditions (each value represents mean ± SD). n/a, not applicable; n.s., not significant.
significantly, about the same amount as in mice and rats, but the \( \text{BP}_{\text{ND}} \) showed no changes compared to baseline, and this difference between species could disappear with longer scanning times. However, as the guinea pigs had breathing difficulties under long-term anesthesia, increased scan time was not applied for ethical reasons.

The difference in \( \text{BP}_{\text{ND}} \) observed in guinea pigs, compared with mice and rats, is most likely due to the differences in homology of the 5-HT\(_{1B}\) receptor in the guinea pig versus that in the rat and mouse. A previous report indicated that amino acid residue 335 in the 5-HT\(_{1B}\) receptor is a major determinant of its reactivity with several ligands [26]. It has also been shown that \(^{3}\text{H}\) AZ10419369 has high affinity \((\text{Kd} = 0.4 \text{nM})\) for both human and guinea pig 5-HT\(_{1B}\) receptors, and AZ10419369 binds to human \((\text{Ki} = 0.8 \text{nM})\) and guinea pig 5-HT\(_{1B}\) receptors \((\text{Ki} = 0.01 \text{nM})\) with higher affinity than it does to rat 5-HT\(_{1B}\) receptors \((\text{Ki} = 2.3 \text{nM})\) [12]. Furthermore, our results show good agreement with the in vivo results of Maier et al. (2009) on guinea pig brain distribution of \(^{3}\text{H}\) AZ10419369 [27]. Thus, the current study seems to confirm the difference in \( \text{Kd} \) across species in vivo, with lower binding affinity to the mouse and rat 5-HT\(_{1B}\) receptor and with the highest affinity to the receptor in the guinea pig.

The results from the present study further support the importance of evaluating the outcome of small animal model experiments with caution especially when it comes to drug development studies. Since there can be species differences in transporter function, as shown in the present study, different promising radioligands targeted for the brain may be discarded on incorrect conclusions.

**Conclusions**

This study showed that \(^{11}\text{C}\) AZ10419369 is a substrate for ABC transporters in mouse, rat, and guinea pig. Our results suggest that the activity of ABC transporters, such as P-gp, as well as possible species differences in the receptor affinity of radioligands, are important factors to consider in the design of preclinical PET studies.

**Additional file**

Additional file 1: Supplementary material to the manuscript by Tóth et al. Additional PET experiment using P-gp knockout and wild-type rats to explore the contribution of the ABC transporter P-gp on the brain uptake of \(^{11}\text{C}\) AZ10419369.

**Competing interests**

The authors declare that they have no competing interests.

**Authors’ contributions**

MTóth participated in the design of the study, carried out the PET experiments, performed the statistical analysis, and drafted the manuscript. JH participated in the design of the study, carried out the PET experiments, and revised the manuscript draft critically. AV, BG, and CH participated in the design of the study and drafted the manuscript. S.J. and JD participated in the design of the study and revised the manuscript draft critically. M.Tokunaga participated in the design of the supplementary material, carried out the supplementary PET experiments, performed the statistical analysis on the supplementary material, and revised the manuscript draft critically. MH participated in the design of the supplementary material and revised the manuscript draft critically. All authors read and approved the final manuscript.

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**References**

1. Markou A, Chiamulea C, Geyer MA, Tricklebank M, Steckler T: Removing obstacles in neuroscience drug discovery: the future path for animal models. Neuropsychopharmacology 2009, 34(1):74–89.
2. Shen HW, Jiang XL, Gonzalez FJ, Yu AM: Humanized transgenic mouse models for drug metabolism and pharmacokinetic research. Curr Drug Metab 2011, 12:997–1006.
3. Tournier N, Valette H, Peyronneau MA, Saba W, Kuhnast B, Dollé F, Scherrmann JM, Cisternino S, Bottlaender M: Transport of selected PET radiotracers by human P-glycoprotein (ABCB1) and breast cancer resistance protein (ABCG2): an in vitro screening. J Nucl Med 2011, 52(3):415–423.
4. Kanani P, John C, Zoghbi SS, Haldin C, Gottesman MM, Innis RB, Hall MD: Imaging the function of P-glycoprotein with radiotracers: pharmacokinetics and in vivo applications. Clin Pharmacol Ther 2009, 86:368–377.
5. Gottesman MM: Mechanisms of cancer drug resistance. Annu Rev Med 2002, 53:615–627.
6. Szakács G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM: Targeting multidrug resistance in cancer. Nat Rev Drug Discov 2006, 5:219–234.
7. Hoshi Y, Uchida Y, Tachikawa M, Inoue T, Ohtsuki S, Terai K, Tariq S: Quantitative atlas of blood-brain barrier transporters, receptors, and tight junction proteins in rats and common marmoset. J Pharm Sci 2013, 102(9):3343–3355.
8. Shawahna R, Uchida Y, Declèves Y, Ohtsuki S, Yousif S, Dauchy S, Jacob A, Chassour F, Daumas-Dupont C, Couraud PO, Terai K, Scherrmann JM: Transcriptomic and quantitative proteomic analysis of transporters and drug metabolizing enzymes in freshly isolated human brain microvessels. Mol Pharm 2011, 8(4):1332–1341.
9. Lötscher U: Drug transporters in the epileptic brain. Epilepsia 2007, 48(Suppl 1):118–119.
10. Syvärinen S, Lindhe O, Palmer M, Korunn BR, Rahman O, Långström B, Kruusen GM, Hammarlund-Udenaes M: Species differences in blood-brain barrier transport of three positron emission tomography radioligands with emphasis on P-glycoprotein transport. Drug Metab Dispos 2009, 37(2):635–643.
11. Andersson JD, Persson ME, Finnera SJ, Gulyás B, Heys R, Elmore CS, Farde L, Haldin C: Development of a PET radioligand for the central 5-HT1B receptor: radiosynthesis and characterization in cynomolgus monkeys of eight radiolabeled compounds. Nucl Med Biol 2011, 38(2):261–272.
12. Persson ME, Andersson J, Nyberg S, McCarthy DJ, Finnera SJ, Varnäs K, Takano A, Karlsson P, Gulyás B, Media AM, Lee CM, Powell ME, Heys JR, Potts W,
