Screening of the DNA mismatch repair genes MLH1, MSH2 and MSH6 in a Greek cohort of Lynch syndrome suspected families

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Abstract

Background: Germline mutations in the DNA mismatch repair genes predispose to Lynch syndrome, thus conferring a high relative risk of colorectal and endometrial cancer. The MLH1, MSH2 and MSH6 mutational spectrum reported so far involves minor alterations scattered throughout their coding regions as well as large genomic rearrangements. Therefore, a combination of complete sequencing and a specialized technique for the detection of genomic rearrangements should be conducted during a proper DNA-testing procedure. Our main goal was to successfully identify Lynch syndrome families and determine the spectrum of MLH1, MSH2 and MSH6 mutations in Greek Lynch families in order to develop an efficient screening protocol for the Greek colorectal cancer patients’ cohort.

Methods: Forty-two samples from twenty-four families, out of which twenty two of Greek, one of Cypriot and one of Serbian origin, were screened for the presence of germline mutations in the major mismatch repair genes through direct sequencing and MLPA. Families were selected upon Amsterdam criteria or revised Bethesda guidelines.

Results: Ten deleterious alterations were detected in twelve out of the twenty-four families subjected to genetic testing, thus our detection rate is 50%. Four of the pathogenic point mutations, namely two nonsense, one missense and one splice site change, are novel, whereas the detected genomic deletion encompassing exon 6 of the MLH1 gene has been described repeatedly in the LOVD database. The average age of onset for the development of both colorectal and endometrial cancer among mutation positive families is 43.2 years.

Conclusion: The mutational spectrum of the MMR genes investigated as it has been shaped by our analysis is quite heterogeneous without any strong indication for the presence of a founder effect.

Background

Colorectal cancer (CRC) is the second most common cause of cancer-related deaths in the industrialized countries [1]. Approximately 5 to 10% of all colorectal cancer cases are due to highly penetrant alleles, which are inherited mostly in an autosomal dominant fashion [2]. Lynch syndrome, also referred to as Hereditary Non Polyposis Colorectal Cancer (HNPCC), is the most common hereditary colon cancer syndrome [1]. However, its actual tumour spectrum is quite heterogeneous. Particularly, it confers high susceptibility to the development of cancer in the female reproductive tract [3], while the risk for developing cancer at other organs such as ovaries, stomach, small bowel, brain and urinary tract ranges from 2% to 13% [4]. It is attributed to germline mutations in either of the mismatch repair (MMR) genes: MLH1, MSH2, MSH6 and PMS2 [5]. The main function of the MMR gene products is to identify and correct...
mismatches as well as short insertion or deletion loops, which occur during replication and recombination [6]. In case of disabled MMR machinery, errors resulting to the contraction/ expansion of tandemly repeated sequences, known as microsatellites, accumulate. This condition is termed as microsatellite instability (MSI). Detection of MSI has been a useful prescreening laboratory tool for the recognition of suspected Lynch syndrome cases [2,7].

In clinical practice, the diagnosis of Lynch syndrome is mainly based on the Amsterdam criteria (AMS) [8]. However, families of small size or with atypical features, such as later age of onset for CRC, might be missed if only the AMS criteria are taken into consideration. Therefore, the Bethesda guidelines, which include all clinical conditions, have emerged as criteria of suspicion [2,8].

The selection of the actual “high-risk” patients is a matter of significance, as a proper genetic testing should combine sequencing of the MMR gene coding regions with techniques for the detection of large genomic rearrangements. This is so, because no hotspots have been reported in the MMR genes while deletions/duplications account for a substantial part of mutations associated with Lynch syndrome [9-11]. Nevertheless, the presence of founder mutations have been well documented in some populations, thus facilitating the procedure of genetic testing [1].

The aim of this study was to successfully identify Lynch syndrome families and to report the MLH1, MSH2 and MSH6 mutational spectrum within Greek colorectal cancer families. More specifically, the nature and frequency of the pathogenic alterations has been elucidated in order to develop an efficient DNA-based screening protocol for the Greek colorectal cancer patients’ cohort.

Methods

Patients

The Amsterdam criteria and the revised Bethesda guidelines were used as selection criteria. Ultimately, twenty-four families were subjected for genetic testing, i.e. monitoring of point mutations or large genomic rearrangements in MLH1, MSH2 and MSH6 genes. Seventeen of the selected families fulfilled the Amsterdam criteria while seven families satisfied at least one of the revised Bethesda guidelines. All families were referred to Molecular Diagnostics Laboratory - NCSR “DEMOKRITOS”. All patients gave us written informed consent for participation in our study. The study was approved by the Bioethics Committee of the National Centre for Scientific Research “Demokritos” (Reference Number 240/EHΔ/10.8) in agreement with the 1975 Helsinki statement, revised in 1983.

An additional 951 sporadic colorectal cancer cases, collected from Hellenic Community of Oncology (HECOG), were screened for the genomic rearrangement identified, resulting in the deletion of exon 6 of MLH1 gene.

DNA extraction

Total genomic DNA was isolated from peripheral blood lymphocytes following the salt extraction procedure [12]. The quantity and quality of the DNA samples was determined with “NanoDrop” (Thermo Fisher Scientific, MA, USA).

DNA sequencing

All exons of the MLH1, MSH2 and MSH6 genes, including splicing junctions, were amplified by polymerase chain reaction (PCR). The PCR products were purified with “Nucleofast 96 PCR Plates” (Macherey-Nagel, Germany) and then subjected to automated cycle sequencing with the Big Dye Terminator Cycle v3.1 sequencing kit (Applied Biosystems, Foster City, CA) and electrophoresis on an ABI Prism 310 Sequencer (PerkinElmer, Applied Biosystems, Foster City, CA). Primer pairs were designed with “Primer 3” software and are available upon request.

RNA extraction and RT-PCR

Total RNA was extracted from peripheral blood lymphocytes using Trizol (Invitrogen, Paisley, UK) following standard protocol. 1000 ng of total RNA was reverse transcribed using 60 μM of random hexamers and 10 units of MMLV reverse transcriptase (Roche Diagnostics, Mannheim, Germany) at 50°C for 30 min, followed by enzyme inactivation at 85°C for 5 min. Subsequently, cDNA was amplified on a new PCR reaction using the proper primer set.

Multiplex Ligation-Dependent Probe Amplification (MLPA)

MLPA was performed with P003 kit (MRC-Holland, Netherlands) according to the manufacturer’s instructions. Fragment analysis was conducted on ABI Prism 310 Genetic Analyzer using GeneMarker software (Applied Biosystems, Foster City, CA). 35 peaks corresponding to each exon of the MLH1 and MSH2 genes, as well as 7 peaks corresponding to DNA sequences outside these genes, were identified. Their migration was calculated according to the GeneScan ROX-500 size standard (Applied Biosystems, Foster City, CA). Decrease of 30 to 50% in the peak area(s) was considered as deletion of the corresponding exon(s), while increase of 30 to 50% as duplication of the corresponding exon(s).

Long range polymerase chain reaction and breakpoint analysis

150 ng of genomic DNA was amplified in a 50 μl reaction volume using 1,5 mM Mg²⁺, 300 μM of each
Table 1 Features of families carrying a germline mutation in either of the MMR genes

| Patient ID | Clinical manifestations (age of onset) | Family history | Criteria used for selection |
|------------|----------------------------------------|----------------|-----------------------------|
| F33        | Endometrial Ca (40)                     | mother hysterectomy at 40y-CRC at 70y, 1st brother CRC at 50y, 2nd brother CRC at 32y | AMS II |
| F39        | CRC (50)                               | father stomach Ca at 64y, sister ovarian Ca at 56y, one 2nd degree relative (P) brain Ca at 47y, one 2nd degree relative (P) pancreatic Ca at 58y, 2 2nd degree relatives (P) CRC at 52y & 80y respectively | AMS II |
| F41        | Endometrial Ca (~45)-CRC (77)          | 1 daughter endometrial Ca at 45y-CRC at 58y, 1 grandson died from CRC at 29y, 1 granddaughter <5 polyps at 33y, two brothers CRC >50y, 1 sister endometrial Ca at 26y, 1 niece endometrial Ca at 40y | AMS II |
| F74        | CRC (42)-endometrial Ca (64)           | mother endometrial Ca-CRC, brother CRC at 26y, grandmother (M) CRC, uncle (M) CRC at 37y-CRC at 34y, cousin (M) CRC at 40y | AMS II |
| F84        | Endometrial Ca (40)                    | mother died from pulmonary embolism, grandmother (M) stomach Ca at 60y, 2 2nd degree relatives CRC, 1 2nd degree relative CRC-metachronous endometrial Ca | AMS II |
| F263       | CRC (42)-metachronous CRC (47)         | mother endometrial Ca at 50y, grandfather (M) stomach Ca, 4 2nd degree relatives CRC, 1 2nd degree relative endometrial Ca at 34y | AMS II |
| F111       | CRC (42)                               | father renal Ca-prostate Ca, grandfather (P) CRC, uncle (P) CRC | Revised Bethesda |
| F150       | CRC (40)                               | father CRC at 48y, uncle (P) CRC at 50y | AMS I |
| F656       | CRC (59)                               | father CRC at 41y, 1st sister CRC at 41y, 2nd sister CRC at 65y, grandmother (P) gynecological Ca at 7y | AMS II |
| F1278      | CRC (33)                               | father CRC at 56y, grandmother (P) gynecological Ca at 7y-CRC at ~43y | AMS II |
| F68        | CRC (28)-metachronous CRC (44)         | father prostate Ca, brother 4 polyps at 36y, 1 2nd degree relative CRC at 45y-metachronous CRC at 80y, 1 2nd degree relative duodenum adenocCa at 42y | Revised Bethesda |
| F1376      | Ca cecum (30)                          | mother CRC at 50y-polyp in the female reproductive tract at ~45y, grandfather (M) CRC at ~80y, 5 2nd degree relatives CRC <60y, 1 2nd degree relative endometrial Ca at ~50y-metachronous CRC at 65y | AMSII |
| 15RB (Serbian origin) | CRC (49)                               | father CRC at 60y, 2 brothers CRC at 49 & 59y respectively, 1 sister endometrial Ca at 48y-metachronous CRC at 57y, 1 sister pancreatic Ca at 63y, 1 niece CRC-synchronous stomach Ca at 25y, 2 2nd degree relatives (P) CRC at 50 & 60y respectively, 1 2nd degree relative (P) bladder Ca at 60y | AMSII |

*Family members written in bold italic carry the particular mutation running in the family **(P): Paternal, (M): Maternal, CRC: ColoRectal Cancer, Ca: Cancer.
Three of the pathogenic nucleotide substitutions located in exons 5, 7 and 13 of the MSH2 gene resulted in a premature stop codon. The nonsense mutations MSH2, p.Gln314X (exon 5) and p.Gln413X (exon 7) are novel, to the best of our knowledge. The MSH2, p.Arg711X (exon 13) has been reported several times in the LOVD database, as well as in an older study conducted in Greek colorectal cancer patients [15].

Two splice site changes were detected in four separate families. RT-PCR analysis of the MLH1, c.116+5G > C mutation, which is located in intron 1, results in a 227-base pair insertion taking place between exons 1 and 2 (data not shown). The specific mutation has been reported twice in the literature, while both bioinformatic prediction tools and in vitro techniques (conversion/cDNA analysis) demonstrated that it activates a cryptic splice site [16,17]. On the other hand, the MLH1, c.790+1G > A mutation, which is located in intron 9, induces skipping of exons 9 and 10 [18]. This mutation was identified in two distinct families, one of Greek and one of Cypriot origin. Thirteen different cancer cases have been reported in the Cypriot family, all potentially related to Lynch syndrome. However, one of the proband’s daughters, who developed breast cancer at the age of 42 years, does not carry the particular mutation. The detailed family pedigree is shown on figure 1.

Moreover, direct sequencing of DNA from samples F1278 and 1SRB revealed the presence of two discrete missense changes located in exon 13 of the MSH2 gene. Exons 7 to 13 of the MSH2 gene span the highly conserved ABC-ATPase domain, thus amino acid substitutions in this domain may have a severe impact on the protein’s function [19]. The MSH2, p.Cys759Arg is characterized as pathogenic in the “MMR Gene Unclassified Variants Database” http://www.mmrmissense.info. The MSH2, p.Arg711Pro mutation, which was identified in a Serbian family, is reported for the first time and its pathogenicity was assessed through bioinformatics tools (Table 3). All the protein web tools used (SIFT, PolyPhen, MAPP-MMR [20]) conclude that it is a deleterious change. In addition, it may influence the splicing process through the abrogation/creation of enhancer/silencer motifs as prediction algorithms for the presence of such sequences have demonstrated (Table 3).

The MSH2, c.1276G > C involves a novel G to C transversion located at the last nucleotide of the exon 7 of the MSH2 gene. According to the in silico analysis performed, the wild type splice donor is abolished (Table 4). Furthermore, the majority of the algorithms used for the prediction of enhancer/silencer motifs by the Human Splice Finder (v.2.4) web tool http://www.umd.be/HSF/ [21] indicate that the c.1276G > C induces the abrogation or creation of such binding sites.

The MSH6, c.3202 C > T mutation, located on exon 5 of the gene, has been identified in an individual diagnosed with colorectal cancer at the age of 40. The specific mutation, although has been previously reported, is of interest since it is the first MSH6 mutation reported in a Greek family.

Detection of benign polymorphisms

Eleven intronic variants, three missense and four synonymous changes have been identified in the MLH1, MSH2 and MSH6 genes (Table 5). Two of the reported polymorphisms (MSH2, c.646-46delC and MSH6, c.3678 A > G) are novel.
The MSH2, c.646-46delC, located in intron 3 of the gene, has not been previously described. As it was detected in one of our MLH1/MSH2 mutation-negative patients, we tried to further characterize it through in silico analysis (Table 4). The Human Splice Finder (v.2.4) software demonstrated a significant variation in the branch site between the “wild type” and the “mutant” sequence, while both the NNSplice (0.9) http://www.fruitfly.org/seq_tools/splice.html and NetGene2 http://www.cbs.dtu.dk/services/NetGene2/ software packages presented no significant variation in either of the major splice sites. Subsequently, RNA sample was collected from the index patient in order to examine any possible alteration of the above variant on the RNA level. cDNA prepared was amplified with primers spanning exons 2 to 5 of the MSH2 gene and revealed the presence of one discernible band corresponding to the wild type transcript (data not shown). Therefore, it is obvious that the MSH2, c.646-46delC is a non-pathogenic variation.

**Gene dosage alterations**

The eleven families tested negative for point mutations were screened for the presence of large genomic rearrangements using MLPA. The MLPA analysis indicated the presence of genomic rearrangements in two out of twelve families. Particularly, one deletion encompassing exon 6 of the MLH1 gene (c.454-?_545+?del) and one duplication involving exon 12 of the MSH2 gene (c.1760-?_2005+?dup) have been demonstrated.

The MLH1, c.454-?_545+?del was confirmed through an individual second MLPA experiment which involved the proband of the family (263), as well as one affected and one unaffected first-degree relative (Figure 2). Subsequently, a long range PCR was performed using a sense primer located in exon 5 and an antisense primer.
located in intron 7, which preferentially amplified a smaller fragment of about 2.5 kb, instead of the expected wild type fragment of 4937 bp (figure 3). The 2.5 kb fragment was subjected to sequencing in order to uncover the flanking regions of the deleted segment. Then, the 2.5 kb fragment was further amplified using various combinations of intronic primers. Sequencing of the smaller segments produced revealed the boundaries of the deletion, starting between nucleotides 157967-158005 in intron 5 and ends between nucleotides 160376-160414 in intron 6 (GenBank Accession number AC011816). The breakpoints belong to AluSx_C repeats according to the results of RepeatMasker http://www.repeatmasker.org/cgi-bin/WEBRepeatMasker (Figure 3). The exact breakpoints could not be determined due to the presence of two identical 33 bp sequences located within the AluSx_C elements which flank the deletion.

In order to assess the frequency of the MLH1, c.454-?_545+?del mutation in the Greek population, 951 unselected sporadic CRC cases were screened using diagnostic primers. The specific deletion was not detected among these samples, indicating that it is not a recurrent mutation in the context of the Greek population. In the case of the duplication of exon 12 of the MSH2 gene identified by MLPA, the results have been confirmed by a second individual MLPA experiment, but it was not possible to be confirmed by another method.

Discussion and Conclusions
Lynch syndrome is a heterogeneous disorder in respects to its molecular basis, as well as its phenotypic expression. Therefore, the selection of the putative mutation carriers as well as the detection of the causative germ-line alterations is a challenging task. A variety of point mutations, such as substitutions, small insertions/deletions and splice site alterations, as well as large genomic rearrangements have been reported in the international InSiGHT (LOVD) database [10]. Particularly, genomic deletions account for approximately 10% of MLH1 and MSH2 mutations, while genomic duplications have been observed in approximately 1% of the Lynch syndrome cases [11,22]. Furthermore, the fact that there are multiple susceptible genes that predispose to Lynch syndrome, immunohistochemistry and/or MSI should be used, where possible, as a pre-screening method to successfully identify the high risk families. In our series of

| Methods of in silico analysis | Result | Comments |
|-----------------------------|--------|----------|
| SIFT score                  | 0.00→pathogenic | If SIFT score < .05 then the aa substitution is predicted to affect protein function |
| PolyPhen                    | Probably damaging (3.071)→pathogenic | If PolyPhen score > 2 then the aa substitution is predicted to affect protein function |
| MAPP-MMR                    | 40.700→pathogenic | If MAPP-MMR score > 4.55 then the aa substitution is predicted to affect protein function |
| NNSplice (0.9)              | SD: 1.00/1.00 No |
|                             | SA: 0.95/0.95 Change |
| NetGene2 Server             | SD: 0.58/0.62 No |
|                             | SA: 0.71/0.71 Change |
| Human Splice Finder v2.4    | SD: 74.29/74.4 |
| (HSF)                       | SA (c.2123):76.67/86.97 |
|                             | SA (c.2126):75.87/79.36 |
|                             | BP:81.68/86.61 |
| Rescue ESE                  | ESE Finder: c.2129 SF2/ASF (IgM-BRCA1)→new site |
| PESE octamers No difference | EIEs (Zhang et al): c.2127→new site |
| ESS (Wang et al)            | ESE from HSF: c.2132 (9G8)→broken site |
| Fas-Ess hexamers            | Silencer motifs (Sironi et al): c.2127→new site |
| PESS octamers               | Other splicing motifs (Goren et al): c.2130→new site |

*SD: splice donor, SA: splice acceptor, BP: branch point aa: aminoacid. **These algorithms are included in the HSF analysis.
experiments, tumor samples from the patients studied were not available and therefore, direct sequencing analysis of the MMR genes was performed.

Nine pathogenic point mutations and one genomic rearrangement have been detected in twelve out of seventeen (70.5%) AMS+ families studied. A 29.5% of AMS+ families that included at least two colorectal or other Lynch syndrome-associated cancer cases remained unresolved. Additionally, no deleterious mutations were detected in patients with young age of onset for CRC, i.e. < 40 years, but with no family history. Our results coincide with observations of previous studies which have demonstrated that a proportion as high as 50% of AMS+ families harbour no pathogenic mutations in either of the major MMR genes, namely MSH1 and MSH2 genes [9]. Nevertheless, the detection rate among AMS+ families was much higher than in families selected upon Bethesda guidelines, indicating the already reported higher specificity of the modified Amsterdam criteria in clinical practice.

| Gene (exon/intron) | Nucleotide change | Method | Result | Comments |
|--------------------|------------------|--------|--------|----------|
| **MSH2 (exon 7)**  | c.1276G > A      | Human Splice Finder (v.2.4) (HSF) (*) | "wt" donor site broken (84.7/77.68) | Scores predicted for the wt seq/ score predicted for the mut seq |
|                    |                  | ESE Finder: c.1274 (SRp55)→new site |
|                    |                  | Rescue ESE: c.1271/c.1272→site broken |
|                    |                  | ESE from HSF: c.1273/c.1276 (9G8)→site broken |
|                    |                  | Silencer motifs (Sironi et al): c.1271/c.1272→site broken |
|                    |                  | Fas-Ess hexamers: c.1275→site broken |
|                    |                  | NNSplice (0.9) | "wt"donor site broken (0.91/-) |
|                    |                  | NetGene2 Server | "wt" donor site broken (0.83/-) |
| **MSH2 (intron 3)**| c.646/46delC    | Human Splice Finder (v.2.4) (HSF) (*) | Variation in one of the potential branch points (c.646-48) (79.39/23.83) | Scores predicted for the wt seq/ score predicted for the mut seq |
|                    |                  | ESE Finder | Rescue ESE |
|                    |                  | PESE octamers | EIEs (Zhang et al) No |
|                    |                  | ESE from HSF change | |
|                    |                  | Silencer motifs/no | |
|                    |                  | hnRNP motifs | |
|                    |                  | Other splicing motifs | |
|                    |                  | PESS octamers: c.646-49 (46.39/87.56)/c.646-46→new site |
|                    |                  | NNSplice (0.9) | SD: 0.90/0.91 No |
|                    |                  | NetGene2 Server | SD: 0.76/0.76 No |

(*) these algorithms are included in the HSF analysis.

Table 4 *In silico* analysis of novel sequence variations.

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Since the particular study includes a relatively small sample size, which can be a limitation when trying to provide a complete mutational spectrum, all available information on Greek putative Lynch syndrome families have been combined [15]. Thirty-three families have been screened for the presence of germline mutations in the *MLH1* and *MSH2* genes, while thirteen of them have been screened for the presence of germline mutations in the *MSH6* gene. Sixteen mutations have been detected in eighteen of these families (Figure 4). To the best of our knowledge, eight of the pathogenic mutations recorded have never been described before.

Genomic rearrangements in the *MLH1* and *MSH2* genes represent 6% of the mutations identified in Greece, a percentage that falls within the range reported previously [9,22-25]. More specifically, in a German patients’ cohort genomic deletions account for 10.6% of colorectal cancer families, while in the Dutch population genomic aberrations are encountered in 6.5% of AMS+ families [23,24]. If only AMS+ families are taken into account, this proportion rises up to 8.3%, which is in accordance with the observation made by Martínez-Bouzas et al [26]. Nonsense mutations and splice site changes seem to prevail among point alterations identified in the Greek patients’ cohort. A quite interesting finding of this study is the high rate of novel mutations, which is calculated to be 53.3%, highlighting the distinct heterogeneous nature of the Greek mutational spectrum of the *MLH1* and *MSH2* genes.

Despite the aforementioned heterogeneity, two of the mutations described, the *MLH1*, c.790+1G > A transition and the *MLH1*, c.116+5G > C transversion were detected in two distinct families during this study. The *MLH1*, c.790+1G > A has been reported several times in different populations and further analysis is required in order to investigate the possible founder effect of this mutation within the Greek or Cypriot population. The extensive clustering of cancer cases within members of family 41 is of particular interest, since thirteen family members spread across three successive generations were diagnosed with five different cancer types. Interestingly, two of the family members were diagnosed with breast and thyroid cancer, respectively. These two types of cancer are not typical phenotypic features of the Lynch syndrome. Both patients were genotyped and found not to harbour the particular pathogenic mutation, indicating that they were probably sporadic cancer cases. On the contrary, other reports have been able to identify breast cancer in MMR mutation carriers, enhancing the theory that breast cancer can be a rare phenotypic feature of Lynch syndrome [27-31].

On the other hand, the *MLH1*, c.116+5G > C has been reported only once before [16]. There is a possibility that the two families (F656, F1376) harbouring the *MLH1*, c.116+5G > C mutation may be related, but it was not feasible to track down all the family relatives, since most of them live abroad. Further investigations are required in order to elucidate whether the particular alteration has a founder effect within the Greek population.

The aforementioned data underscore the heterogeneity of the *MLH1* and *MSH2* mutational spectrum in the Greek population.

| Gene   | Location | Nucleotide change | Protein effect | Families with the variant |
|--------|----------|-------------------|----------------|---------------------------|
| *MLH1* | Intron 6  | c.545+72T > A     | N.A            | 2                         |
|        | Exon 8   | c.655A > G        | p.Ile219Val    | 12*                       |
|        | Intron 14| c.1668-19A > G    | N.A            | 6*                        |
| *MSH2* | Intron 1  | c.211+8G > C      | N.A            | 2                         |
|        | Intron 1  | c.211+9C > G      | N.A            | 4                         |
|        | *Intron 3*| c.646-46delC      | N.A            | 1                         |
|        | Exon 6   | c.965G > A        | p.Gly322Asp    | 6                         |
|        | Intron 10| c.1661+12G > A    | N.A            | 14*                       |
|        | Intron 12| c.2006-6T > C     | N.A            | 9*                        |
| *MSH6* | Intron 1  | c.261-36 A > G    | N.A            | 2                         |
|        | Exon 1   | c.116 G > A       | p.Gly39Glu     | 2                         |
|        | Exon 1   | c.186 C > A       | p.Arg62Arg     | 2*                        |
|        | Exon 2   | c.276 A > G       | p.Pro92Pro     | 4*                        |
|        | Exon 3   | c.540 T > C       | p.Asp180Asp    | 4*                        |
|        | Intron 4  | c.3173-101 C > G  | N.A            | 1                         |
|        | Exon 5   | c.3438+14 A > T   | N.A            | 8*                        |
|        | *Intron 7*| c.3646+29_3646+32delCTAT | N.A. | 3*                       |
|        | *Intron 8*| c.3802-40 C > G  | N.A          | 8*                        |
|        | *Exon 8* | c.3678 A > G      | p.Ala1226Ala   | 1                         |

Table 5 Polymorphisms in either of the major MMR genes

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Greek population, since each Greek family harbours a distinct deleterious germline mutation, which can be of any type. Furthermore, the first germline pathogenic mutation in *MSH6* gene in a Greek family is reported here. Therefore, a combination of techniques that will be able to detect both small size alterations within the three genes and large deletions/insertions is required for routine genetic testing of putative Lynch syndrome patients, which is in accordance with other studies.

Moreover, we have achieved to determine the breakpoints of the *MLH1*, c.454–?_545+?del and subsequently to develop diagnostic primers for its rapid detection. This deletion was described initially by Viel and her colleagues [32] and has been reported several times since then. The possible founder effect of the specific mutation within the Greek population has been excluded when 951 unselected colorectal cancer cases were screened and no carriers of the specific mutation have been identified.

It is of great significance to obtain systematically data about the exact nature and frequency of the pathogenic mutations encountered in a particular patients’ cohort, in order to customize the genetic testing to the population’s needs, which will eventually result in the reduction of the high cost of genetic testing along with the optimization of the detection rate.

Our results indicate the distinct, heterogeneous nature of Lynch syndrome’s associated mutations in a Greek colorectal cancer patients’ cohort, as well as the significant contribution of Amsterdam criteria in the...
Figure 3 Breakpoint analysis of the MLH1, c.454-?_545+?del. The products after long range PCR are shown in the upper part of the figure. In the first two lanes corresponding to DNA samples of the 263CRC family members we obtained a discernible band of ~2.5 kb, as well as a fainted band with the expected size of ~5 kb, while in the third lane with control DNA sample we got only the band of ~5 kb which corresponds to the expected “wt” allele. Lane 4: 100 bp ladder, lane 5: negative control, lane 6: 1 kB ladder. The PCR product obtained by the combination of F3 and R1 primers was sequenced with R1 and revealed the deletion breakpoints, which are indicated by purple arrows upon the chromatogram. The size and the boundaries of the deletion are depicted schematically in the lower part of the figure.
identification of putative Lynch syndrome families. Consequently, the compilation of an accurate and detailed family history by the physician is a critical step for the diagnosis of Lynch syndrome.

Abbreviations
CRC: Colorectal cancer; HNPCC: Hereditary non polyposis colorectal cancer; MMR: Mismatch repair genes; MSI: Microsatellite instability; AMS: Amsterdam; MLPA: Multiplex ligation-dependent probe amplification; PCR: Polymerase chain reaction; InSiGHT: International society for gastrointestinal hereditary tumours.

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Authors’ contributions
GT checked the family histories for their compliance with AMS clinical criteria for the diagnosis of Lynch syndrome, carried out the molecular genetic testing for MLH1 and MSH2, i.e sequencing and MLPA, as well as the in silico analysis and wrote the paper. FF checked the family histories, carried out a part of the sequencing for MLH1 and MSH2, carried out all the sequencing for MS56 and revised the paper, RS designed the study and revised the paper, GN referred families to our laboratory after collecting a family history, MM referred families to our laboratory after collecting a family history, MBM referred a putative Lynch syndrome family of Serbian origin to our laboratory for final diagnosis through genetic testing, GF referred families to our laboratory after collecting a family history, CP referred families to our laboratory after collecting a family history, MF referred families to our laboratory after collecting a family history, MB referred a putative Lynch syndrome family of Serbian origin to our laboratory for final diagnosis through genetic testing, GF referred families to our laboratory after collecting a family history as well as blood samples from sporadic colorectal cancer patients collected by HECOG, DY designed the study and revised the paper. All authors have read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

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