**A B S T R A C T**

Fungi are microorganisms considered decomposers of the organic matter, by the action of several enzymes that can present biotechnological potential, emphasizing the holocelulases (endoglucanases, exoglucanases, β-glucosidases, endoxilanases, β-xylosidases, among others). Among holocellulolytic fungi described in literature, *Penicillium*, *Aspergillus* and *Trichoderma* species are the most important. This study aimed to evaluate the efficiency in production of holocellulolytic accessories enzymes (β-glucosidase, β-xylosidase and α-L-arabinofuranosidase) by *Trichoderma atroviride* 102C1, a mutant strain obtained in our laboratory, using different lignocelluloses biomass as substrates. The mutant strain 102C1 was cultivated at 28°C in salt mineral solution supplemented with corn steep liquor as nitrogen source (1.26% w/v), and wheat bran, sorghum bagasse, sugarcane straw (*in natura* and pretreated with steam explosion) or sugarcane bagasse (*in natura* and pre-treated with steam explosion), as carbon source (2.5% w/v), 200 rpm, for 5 days. The best results for (β-glucosidase, β-xylosidase and α-L-arabinofuranosidase activities were observed in sorghum bagasse (13.09 U.ml⁻¹), sugarcane straw pretreated (5.24 U.ml⁻¹) and wheat bran (192.6 U.ml⁻¹), respectively, between 3 and 5 days fermentation. These results suggest the use of different agro-industrial by-products to obtain holocellulolytic accessories enzymes by the mutant fungi *T. atroviride* 102C1 and the possibility for biotechnology process.
Introduction

Holocellulases are one of the largest industrial enzymes worldwide, by dollar volume, because of their use in cotton processing, paper recycling, as detergent enzymes, in juice extraction, and as animal feed additives. However, holocellulases will become the largest volume industrial enzyme, if ethanol, butanol, or some other fermentation product of sugars, produced from biomass by enzymes, becomes a major transportation fuel (Wilson, 2009). The mainly holocellulases producers in nature are the filamentous fungi, like Aspergillus, Penicillium and Trichoderma. The holocellulase complex from Trichoderma sp., one of the most important microorganisms used in industry is consisted mainly of cellobiohydrolases (EC 3.2.1.91), endoglucanases (EC 3.2.1.4), β-glucosidases (EC 3.2.1.21), endoxylanases (EC 3.2.1.8), β-xylosidases (EC 3.2.1.37) and α-arabinofuranosidases (EC 3.2.1.55). These enzymes act synergistically to the hydrolysis of lignocellulose biomass in sugars (pentoses and hexoses) and could be produced by bacteria and filamentous fungi in nature, which a potential for biotechnology purposes, through the use of agro-industrial by-products within the biorefinery concept (Sanchez, 2009).

The Brazilian sugarcane system of agroenergy is considered as the most efficient system (Santos et al., 2012). In 2015, Brazil produced about 593 million tons of sugarcane, producing about 150 million tons of sugarcane bagasse and 150 million tons of sugarcane straw. Therefore, in order to meet wider needs, a significant increase in the production of ethanol would be possible only if the basic knowledge necessary for the development of technologies that will be capable to obtain energy from lignocellulosic materials present in sugarcane is developed (Soccol et al., 2010). The conversion of lignocellulose biomass requires a mixture of enzymatic complex, including cellulases (endo-1,4-glucanase, exo-1,4-celllobiohydrolase and β-1,4-glucosidase) as well as the hemicellulases (endoxylanase, β-1,4-xylosidase, α-L-arabinofuranosidase). Some of these enzymes are considered accessories in bioconversion of lignocellulose biomass (Hansen et al., 2015). There are many studies using β-glucosidase and β-xylosidase enzymes and few reports using α-arabinofuranosidase.

The β-1,4-glucosidases are essential for complete hydrolysis of cellulose fiber. These enzymes cleave cellobiose and celloooligosaccharides liberating molecules of glucose as the end product (Gottschalk et al., 2010). β-1,4-Xylosidases are essential enzymes of the microbial xylanolytic system, and contribute to decrease the inhibition of xylanases by the end-product of xylan hydrolysis. This enzyme is cell-associated in most bacteria and yeast, but it is freely found in the culture media of some fungi (Michelin et al., 2012). α-L-Arabinofuranosidases are accessory enzymes that cleave 4-L-arabinofuranosidicα linkages and act synergistically with other hemicellulases and pectic enzymes to promote the complete hydrolysis of hemicelluloses and pectins (Grigorevski-Lima et al., 2013).

In the present work, the production of three enzymes with important accessory role in lignocellulose biodegradation, i.e. β-1,4-glucosidases, β-1,4-xylosidase and α-L-arabinofuranosidase, were studied using different agro-industrial by-products, by the mutant strain Trichoderma atroviride 102C1.

As agro-industrial by-products the authors used sugarcane bagasse (SCB) and straw (SCS) in natura, sugarcane bagasse (SCSE) and straw (SSSE) pre-treated by steam explosion, sorghum bagasse (SB) and wheat bran (WB).
Materials and Methods

Microorganism

Initially the strain *Trichoderma atroviride* 676 was originally isolated from Amazon forest soil and identified by Dr Maria Ines Sarquis, at Fundação Oswaldo Cruz (Rio de Janeiro, Brazil). After nitrosoguanidine and U.V. radiation exposition, the mutant *T. atroviride* 102C1 was selected as holocellulolytic promising strain (Grigorevski-Lima *et al.*, 2013; Oliveira *et al.*, 2014; Oliveira *et al.*, 2016).

Spore suspensions of the mutant strain were prepared according to Hopwood *et al.*, (1985) after cultivation (28ºC/15 days) in yeast extract-malt extract-agar medium (1966) and maintained as stock cultures in 20% (v/v) glycerol at -20ºC. Spore concentration was determined using Neubauer counting chamber.

Production of Holocellulolytic Enzymes

*T. atroviride* 102C1 cells were cultured in a growth medium containing either of the six different agro-industrial by-products as carbon source (2.5% w/v): (i) SCB, (ii) SCS; (iii) SBSE; (iv) SSSE; (v) SB; (vi) WB and corn steep liquor (CSL) as main nitrogen source (1.26% w/v).

Media were always supplemented with modified Mandel solution (Mandels and Weber, 1969) containing (g.l⁻¹): urea, 0.3; (NH₄)₂SO₄, 1.4; KH₂PO₄, 2.0; CaCl₂, 0.3; MgSO₄.7H₂O, 0.3; FeSO₄.7H₂O, 0.005; CoCl₂.6H₂O, 0.02; MnSO₄.4H₂O, 0.016; ZnSO₄.7H₂O, 0.014. Cultivation was performed in submerged fermentation using *T. atroviride* 102C1, in 125 ml Erlenmeyer flasks filled 1/5 of its volume with a culture medium based (initial pH 5.5), which was inoculated with a spore suspension to a final concentration of 10⁶ spores.ml⁻¹. Cells were incubated at 28ºC, in an orbital shaker (200 rev.min⁻¹), for up to 5 days. At each day, the whole content of a shake flask was filtered through a glass microfiber filter (Whatman GF/A), in triplicate, and the culture supernatants obtained were used in enzymatic assays.

The chemical characterization of six agro-industrial by-products was carried out by Laboratório de Controle Bromatológico e Microscópico (LabC Brom / UFRJ), based on methodology of Mendez *et al.*, (1985) and Van Soest (1963).

Enzyme assays

The β-1,4-glucosidase, β-1,4-xylosidase and α-L-arabinofuranosidase activities were determined by release of *p*-nitrophenol obtained by hydrolysis of the substrates *p*-nitrophenyl-β-D-glucopyranoside (Sigma-Aldrich®), *p*-nitrophenyl-β-D-xilopyranoside (Sigma-Aldrich®) and *p*-nitrophenyl-α-L-arabinofuranoside (Sigma-Aldrich®), respectively, to 10 mM.

Enzyme assays were prepared with 200 µl of 0.5M sodium acetate buffer pH 5.0, 650 µl for β-1,4-glucosidase and β-1,4-xylosidase or 600 µl for α-L-arabinofuranosidase of distilled water and 50 µl for β- glucosidase and β-xylosidase or 100 µl for α-L-arabinofuranosidase of the enzyme extract. Reaction happened for 10 minutes at 50ºC with the addition of 100 µl of the substrate corresponding to each enzyme. After this period, the enzyme reaction was stopped with the addition of 500 µl of 1 M Na₂CO₃ pH 10.0. The reading of the amount of *p*-nitrophenol released during the enzyme assays was performed in spectrophotometer at 420nm (Da Silva *et al.*, 2010).

A unit β-1,4-glucosidase, β-1,4-xylosidase and α-L-arabinofuranosidase (U) was defined as...
the amount of enzyme that released 1 µmol of \(p\)-nitrophenol at 50°C in 1 minute. All assays were performed in triplicates, and results were expressed as average values. Variations in the multiple assays were < 10%.

**Results and Discussion**

\(\beta\)-Glucosidase (BGL), \(\beta\)-xylosidase (BXL) and \(\alpha\)-L-arabino-furanosidases (ARF), holocellulolytic enzymes, are very important enzymes in the process of complete biodegradation of lignocellulosic biomass releasing fermentable sugars (glucose, xylose and arabinose).

In this research, the mutant strain *Trichoderma atroviride* 102C1 was studied aiming at the production of holocellulolytic accessories enzymes using different agro-industrial by-products as main carbon source. *T. atroviride* 102C1 is already known as endoglucanase and endoxylanase producer when grown in lignocellulose biomass (Kóvacz et al., 2008; Kóvacz et al., 2009; Oliveira et al., 2014; Oliveira et al., 2016).

For accessories enzymes production from *T. atroviride* 102C1, six different carbon sources (SCB, SCS, SBSE, SSSE, WB and SB) were used as an inducer. Individual evaluation of the carbon sources revealed that use of SB resulted in the highest production of \(\beta\)-glucosidase (13.09 U.ml\(^{-1}\)), after 4 days, compared with other sources used (Figure 1).

Annually, several researches on \(\beta\)-glucosidase production by microorganisms have been carried out, confirming its relevance in cellulose fiber biodegradation. The production of \(\beta\)-glucosidase (BGL) by *Trichoderma* species using lignocellulosic residues are reported in literature. Grigorevski-Lima *et al.*, (2013) produced a maximal \(\beta\)-glucosidase activity (0.17 U.ml\(^{-1}\)) in the parental strain of *T. atroviride* 102C1 after 4 fermentation-days using sugarcane bagasse (*in natura*). Rana *et al.*, (2014) have reported \(\beta\)-glucosidase production with *T. reesei* RUT-C30 (ATCC 56765) using corn straw pretreated by alkali explosion as 4.77 U.ml\(^{-1}\), after 7 fermentation-days in STR bioreactor. Kóvacs, Szakacs and Zacchi [18] have showed enzyme production using *T. reesei* RUT-C30, *T. atroviride* TUB F-1505 and *T. atroviride* TUB F-1505 (mutant of wild strain TUB F-1505) with \(\beta\)-glucosidase activity of <0.2, 4.9 and 7.1 U.ml\(^{-1}\), respectively, using steam pretreated spruce as feedstock.

Delabona *et al.*, (2012) produced a maximal \(\beta\)-glucosidase activity (9.18 U.ml\(^{-1}\)) with *T. harzianum* P49P11 using sugarcane bagasse pretreated, after 4 fermentation-days. The authors observed that the BGL production by the mutant strain *T. atroviride* 102C1 was very promising using agro industrial by-products, especially SB and WB at 5 days of fermentation.

When \(\beta\)-xylosidase (BXL) production was studied, the maximal activity (5.24 U.ml\(^{-1}\)) was detected using SSSE after 4 fermentation-days (Figure 2). Jiang *et al.*, (2011) produced a maximal \(\beta\)-xylosidase activity (0.25 U.ml\(^{-1}\)) with *T. reesei* RUT-C30 (ATCC 56765) using lactose 1% (w/v), after 7 fermentation-days. Menezes *et al.*, (2010) observed a maximal \(\beta\)-xylosidase activity (0.09 U.ml\(^{-1}\)) with *Pleurotus* sp. BCCB068 after 40 fermentation-days, using xylan as substrate.

Guerfali, Maalej-Achouri e Belghith (2013) have showed enzyme production using *Talaromyces thermophilus* with \(\beta\)-xylosidase activity of 1.4 U.ml\(^{-1}\), in the presence of wheat bran 2% (w/v), after 7 fermentation-days. There are few reports in literature concerning to BXL production by filamentous fungi. Once again, the production of BXL by mutant *T. atroviride* 102C1 was higher than reported in literature (Table 1).
Table 1 The dry matter-based composition of the biomass components (%)

| Source | Cellulose  | Hemicellulose | Lignin    |
|--------|------------|---------------|-----------|
| SCB    | 32.78 + 0.72 | 42.35 + 1.69  | 10.74 + 0.19 |
| SBSE   | 45.40 + 0.64 | 10.53 + 0.64  | 11.91 + 0.55 |
| SCS    | 25.63 + 2.66 | 37.76 + 2.21  | 14.31 + 0.65 |
| SSSE   | 15.72 + 3.81 | 17.34 + 0.69  | 19.79 + 1.06 |
| WB     | 7.17 + 0.11  | 26.96 + 0.45  | 2.78 + 0.19  |
| SB     | 44.91 + 0.16 | 26.22 + 1.34  | 9.00 + 0.25  |

Fig. 1 Fermentation time-course for β-1,4-glucosidase (BGU), production by *T. atroviride* 102C1, in submerged fermentation, upon the use of different substrates: (▲) SCB; (●) SBSE; (▲) SCS; (△) SSSE; (■) WB and (●) SB
Fig. 2 Fermentation time-course for β-1,4-xylosidase (BXU), production by *T. atroviride* 102C1, in submerged fermentation, upon the use of different substrates:

(♦) SCB; (-○-) SBSE; (◆) SCS; (-▲-) SSSE; (■) WB and (●) SB

Fig. 3 Fermentation time-course for α-L-arabino-furanosidase (ARA), production by *T. atroviride* 102C1, in submerged fermentation, upon the use of different substrates:

(♦) SCB; (-○-) SBSE; (▲) SCS; (-▲-) SSSE; (■) WB and (●) SB
Individual evaluation of the carbon sources revealed that use of WB resulted in the highest production of $\alpha$-L-arabinofuranosidase (ARF), 192.60 U.ml$^{-1}$, after 3 days of submerged fermentation (Figure 3). Almeida et al., (2011) produced 0.045 U.ml$^{-1}$ of $\alpha$-L-arabinofuranosidase with Acremonium zeae EA0802 using oat spelts xylan, after 18 fermentation-days. Temer, Terrasan and Carmona (2014) have showed enzyme production using Penicillium janczewskii with $\alpha$-L-arabinofuranosidase activity of 0.7 U.ml$^{-1}$, using orange waste mixed brewer’s spent grain as feedstock, after 10 days of fermentation. Ioannes et al., (2000) observed $\alpha$-L-arabinofuranosidase activity (0.7, 0.85 and 1.00 U.ml$^{-1}$) with P. purpurogenum using oat spelts xylan, beet pulp and L-arabitol, respectively, after 4 days of fermentation. Visser et al., (2013) have showed enzyme production using Penicillium pinophilum and Chrysoporthe cubensis with $\alpha$-L-arabinofuranosidase activity of 0.27 and 0.52 U.ml$^{-1}$, using knife-milled elephant grass 3% (w/v) and wheat bran, respectively, after 7 fermentation-days. Guerfali, Maalej-Achouri and Belghith (2013) produced a maximal $\alpha$-L-arabinofuranosidase (1.05 U.ml$^{-1}$) with Talaromyces thermophilus using wheat bran 2% (w/v), after 8 days. At the current days, there are few reports concerning ARF production. The mutant strain 102C1 was capable to produce a high ARF activity in the presence of WB (192.60 U.ml$^{-1}$), in a short period time (3 days). This enzyme activity value was higher than those reported in the literature.

The use of accessories enzymes in biodegradation of hemicellulose fraction from lignocellulosic biomass could be very important. When combine $\beta$-xylosidase and $\alpha$-L-arabinofuranosidase in an enzymatic blend, almost doubled xylose release from water extractable wheat arabinoxylan compared to $\beta$-xylosidase treatment alone, for example. The removal of 1-3 linked arabinose from singly substituted xylopyranosyls near non-reducing ends provided access for $\beta$-xylosidase (Rasmussen et al., 2012). The combination of endo-1,4- $\beta$-xylanase and $\beta$-xylosidase to hydrolyse wheat arabinoxylan, could be increased 2.5-fold when $\alpha$-L-arabinofuranosidase was added (Rasmussen and Meyer, 2010; Rasmussen et al., 2012; McCleary et al., 2015).

Lignocellulosic biomass is the most abundant source of renewable sugars that can be fermented into biofuels. However, when focus in their hydrolysis, many efforts still need to be coordinated. The study of accessories enzymes such as $\beta$-1,4-glucosidase, $\beta$-1,4-xylosidase and $\alpha$-L-arabinofuranosidase, can be of great significance to assist in the complete hydrolysis of plant biomass, allowing greater accessibility of the main enzymes (endoglucanase, cellulbiohydrolase and endoxylanase) of the cellulose and hemicellulose fibers. In this work, we could observe different effects of the carbon sources (SCB, SBSE, SCS, SSSE, WB and SB) on enzymes production. The present authors are convinced that our results of the fermentation study of accessories enzymes using different substrates as main carbon source prove that our mutant 102C1 might be suitable strain for practical applications, allowing its use in biotechnological applications, particularly in the hydrolysis of agro-industrial by-products, such as wheat bran (WB) and sorghum bagasse (SB).

**Conflict of Interest**

The authors declare that there are not conflicts of interest in this study.

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References

Almeida, M.N., Guimarães, V.M., Bischoff, K.M., et al., 2011. Cellulases and hemicellulases from endophytic Acremonium species and its application on sugarcane bagasse hydrolysis. Appl Biochem Biotechnol 165: 594-610.

Da Silva, A.S., Inoue, H., Endo, T., Yano, S, Bon, E.P.S. 2010. Milling pretreatment of sugarcane bagasse and straw for enzymatic hydrolysis and ethanol fermentation. Bioresource Technol. 101: 7402-7409.

Damasio, A.R.L., Pessela, B.C., Segato, F., Prade, R.A., Guisan, J.M., and Polizeli, M.L.T.M. 2012. Improvement of fungal arabinofuranosidase thermal stability by reversible immobilization. Proc Biochem 47: 2411-2417.

Delabona, P.S., Farinas, C.S., da Silva, M.R., Azzoni, S.F., and Pradella, J.G.C. 2012. Use of a new Trichoderma harzianum strain isolated from the Amazon rainforest with pretreated sugar cane bagasse for on-site cellulase production. Bioresource Technol 107: 517-521.

Gottschalk, L.M.F., Oliveira, R.A., and Bon, E.P.S. 2010. Cellulases, xylanases, β-glucosidases and ferulic acid esterase produced by Trichoderma and Aspergillus act synergistically in the hydrolysis of sugarcane bagasse. Biochem Eng J 51: 72-78.

Grigorevski-Lima, A.L., Oliveira, M.M.Q., Nascimento, R.P., Bon, E.P.S., and Coelho, R.R.R. 2013. Production and partial characterization of cellulases and xylanases from Trichoderma atroviride 676 using lignocellulosic residual biomass. Appl Biochem Biotech 169: 1373-1385.

Guerfali, M., Maalej-Achouri, I., and Belghith, H. 2013. Hydrolytic potential of Talaromyces thermophilus β-xylosidase and its use for continuous xylose production. Food Technol Biotechnol 51: 479-487.

Hansen, G.H., Lubeck, M., Frisvad, J.C., Lubeck, P.S., and Andersen, B. 2015. Production of cellulolytic enzymes from ascomycetes: comparison of solid state and submerged fermentation. Proc Biochem 50: 1327-1341.

Hopwood, D.A., Bibb, M.J., Chater, K.F., et al., 1985. Genetic manipulation of Streptomyces, a Laboratory Manual. The John Innes Institute, Norwich, United Kingdom.

Ioannes, P., Peirano, A., Steiner, J., Eyzaguirre, J. 2000. An α-L-arabinofuranosidase from Penicillium purpurogenum: Production, purification and properties. J Biotechnol 76: 253-258.

Jiang, X., Geng, A., He, N., and Li, Q. 2011. New isolate of Trichoderma viride strain for enhanced cellulolytic enzyme complex production. J Biosci Bioeng 111: 121-127.

Kovácz, K., Megyeri, L., Szakacs, G., Kubicek, C.P., Galbe, M., and Zacchi, G. 2008. Trichoderma atroviride mutants with enhanced production of cellulase and beta-glucosidase on pretreated willow. Enz Microb Technol 43: 48-55.

Kovácz, K., Szakacs, G., and Zacchi, G. 2009. Comparative enzymatic hydrolysis of pretreated spruce by supernatants whole fermentation broths and washed mycelia of Trichoderma reesei and Trichoderma atroviride. Biore Technol 100: 1350-1357.

Mandels, M., and Weber, J. 1969. The production of cellulases. In: Edited by Gould RF. Cellulases and their applications. Washington, DC: American Chemical Society. Advances in Chemistry Series, 95; pp. 391-414.

McCleary, B.V., McKe, V.A., Draga, A., Rooney, E., Mangan, D., and Larkin, J. 2015. Hydrolysis of wheat flour arabinoxylan, acid-debranched wheat flour arabinoxylan and arabino-xyloligosaccharides by α-xylanase, α-L-arabinofuransidase and β-xylosidase. Carbohydrate Research 407: 79-96.
Mendez, M.H.M., Derivi, S.C.N., Rodrigues, M.C.R., Fernandes, M.L., Machado, R.L.D. 1985. Método de fibra detergente neutro modificado para amostras ricas em amido. Ciência & Tecnologia de Alimentos 5: 123-131.

Menezes, C.R., Silva, I.S., Pavarina, E.C., Faria, A.F., Franciscon, E., and Durrant, L.R. 2010. Production of xylolithiosaccharides from enzymatic hydrolysis of xylan by white-rot fungi Pleurotus. Acta Sci Technol 2: 37-42.

Michelin, M., Peixoto-Nogueira, S.C., Silva, T.M., et al., 2012. A novel xylan degrading b-D-xylosidase: purification and biochemical characterization. W J Microbiol Biotechnol 28(11): 3179-3186.

Oliveira, M.M.Q., Grigorevski-Lima, A.L., Bon, E.P.S., Coelho, R, R, R, and Nascimento, R.R. 2014. Trichoderma atroviride 102C1 mutant: A high endoxylanase producer for assisting lignocellulosic material degradation. J Microb Biochem Technol 6: 236-241.

Rana, V., Eckard, A.D., Teller, P., and Ahring, B.K. 2014. On-site enzymes produced from Trichoderma reesei RUT-C30 and Aspergillus saccharolyticus for hydrolysis of wet exploded corn stover and loblolly pine. Bioresource Technol 154: 282-289.

Rasmussen, L.E., and Meyer, A.S. 2010. Endogeneous β-D-xylosidase and α-L-arabinofuranosidase activity in flax seed mucilage. Biotechnol Letts 32: 1883-1891.

Van Soest, P.J. 1963. Use of Detergent in the Analysis of Fibrous Feed I, Preparation of Fiber Residues of Low Nitrogen. J AOAC Int 46: 925-929.

Wilson, D.B. 2009. Cellulases and biofuels. Curr Opin Biotechnol 20: 295-299.

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