Role of RNA secondary structure in emergence of compartment specific hepatitis B virus immune escape variants

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Abstract

AIM
To investigate the role of subgenotype specific RNA secondary structure in the compartment specific selection of hepatitis B virus (HBV) immune escape mutations.

METHODS
This study was based on the analysis of the specific observation of HBV subgenotype A1 in the serum/plasma, while subgenotype A2 with G145R mutation in the peripheral blood leukocytes (PBLs). Genetic variability found among the two subgenotypes was used for prediction and comparison of the full length pregenomic RNA (pgRNA) secondary structure and base pairings. RNA secondary structures were predicted for 37 °C using the Vienna RNA fold server, with default parameters. Visualization and detailed analysis was done using RNA shapes program.

RESULTS
In this analysis, using similar algorithm and conditions, entirely different pgRNA secondary structures for subgenotype A1 and subgenotype A2 were predicted, suggesting different base pairing patterns within the
two subgenotypes of genotype A, specifically, in the HBV genetic region encoding the major hydrophilic loop. We observed that for subgenotype A1 specific pgRNA, nucleotide 358\(^{\text{C}}\) base paired with 1738\(^{\text{T}}\) and nucleotide 587\(^{\text{C}}\) base paired with 607\(^{\text{T}}\). However in sharp contrast, in subgenotype A2 specific pgRNA, nucleotide 358\(^{\text{T}}\) was opposite to nucleotide 588\(^{\text{C}}\), while 587\(^{\text{C}}\) was opposite to 359\(^{\text{T}}\), hence precluding correct base pairing and thereby lesser stability of the stem structure. When the nucleotides at 358\(^{\text{C}}\) and 587\(^{\text{C}}\) respectively (as observed specifically in the PBL associated A2 sequences), these nucleotides base paired correctly with 588\(^{\text{C}}\) and 359\(^{\text{T}}\), respectively.

**CONCLUSION**

The results of this study show that compartment specific mutations are associated with HBV subgenotype specific alterations in base pairing of the pgRNA, leading to compartment specific selection and preponderance of specific HBV subgenotype with unique mutational pattern.

**Key words:** Hepatitis B; Compartmentalization; Peripheral blood leukocytes; pgRNA; RNA secondary structure; G145R

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Core tip: We have previously shown that, in our study population, distribution of hepatitis B virus (HBV) subgenotypes A1 and A2 is highly biased in the serum/plasma and peripheral blood leukocyte (PBL) compartments respectively. Analysing the predicted base pairing patterns of pregenomic RNAs (pgRNAs), specific for HBV subgenotype A1 and A2, we demonstrate that the potent immune escape mutation G145R evolves specifically in the context of HBV subgenotype A2. The PBL compartment is exposed to strong anti-HBs immunity, and thus G145R is highly advantageous for the virus to persist. This explains the exclusive preponderance of subgenotype A2 in the PBL compartment, sharply contrasting the prevalence of subgenotype A1 in the serum/plasma.

INTRODUCTION

Viral compartmentalization signify infection, persistence and replication of viruses in off-target cells/tissues or anatomical compartments of the host, and this phenomenon is now believed to be a crucial event in many important viral infections, including hepatitis B virus (HBV), human immunodeficiency virus, hepatitis C virus, etc.[1-5]. Recent molecular evolutionary studies have demonstrated that viruses evolve independently under the influence of unique immunological milieu in a given compartment, leading to the selection and emergence of specific viral variants, which endow the virus with an advantage to survive and persist in that particular compartment[2,6,7]. Such compartment specific viral evolution have extremely important implications in emergence and re-emergence of immune-escape mutants, antiviral resistant mutants, their long term persistence and transmission through different non-conventional routes[1-7].

HBV is the prototype member of the Hepadnaviridae family of enveloped viruses with a very unique partially double-stranded DNA genome[8]. Despite having a DNA genome, HBV exclusively uses an RNA intermediate (the pregenomic RNA or the pgRNA) and a virus encoded reverse transcriptase to replicate its genome through a complex mechanism of primer shifting[8]. Even though HBV is classically considered to be a hepatotropic virus, HBV related nucleic acids and proteins have long been detected in different tissues, suggesting that it replicates and propagates in various non-hepatic tissues[9]. Interestingly, some of these extrahepatic sites have been shown to act as reservoirs and also the source of reinfection after surgical and therapeutic interventions[9,10]. Recently, ours and other research groups have provided convincing evidences that the HBV strains and their mutational signature pattern present in different extra-hepatic compartments, are often characteristically distinct from the HBV strains circulating in the serum/plasma/hepatic compartments and that immune escape/drug resistance mutations are significantly more frequent in different extrahepatic compartments in HBV carriers[11,12,14].

In our previous studies, we have recognized the subgenotype A1 (Afro-Asian subgenotype) as the predominant subgenotype of HBV genotype A circulating in the sera/plasma of our study population and that the occurrence of G145R mutation therein was sporadic[12-14]. In sharp contrast, we documented the confined and exclusive existence of HBV subgenotype A2 with the potent “immune escape” mutation G145R within the peripheral blood leukocytes (PBL), across the study population, irrespective of the HBV genotype/subgenotype circulating in the serum/plasma of the respective individual[2]. G145R is the mutation signifying Glycine to Arginine substitution at amino acid residue 145 in the major hydrophilic loop (MHL), a B-cell epitope of the hepatitis B surface antigen (HBsAg), which provides a strong immune escape property. These observations strongly signify that viral mutants with G145R do have an explicit replicative advantage within the PBLs, that are exposed to strong anti-HBs immunity and that this mutation emerges specifically in the perspective of subgenotype A2, but not in subgenotype A1. Moreover, all the subgenotype specific nucleotide substitutions in the MHL encoding region of A1 (505\(^{\text{T}}\), 514\(^{\text{C}}\), 616\(^{\text{T}}\) and 619\(^{\text{C}}\)) and A2 (505\(^{\text{T}}\), 514\(^{\text{C}}\), 616\(^{\text{T}}\) and 619\(^{\text{C}}\)) are
subgenotype A2, respectively, following the method number DQ315784 (India) for subgenotype A1 and full length sequences, namely-GenBank accession generated separately by editing two well defined base-pairing and folding of the pgRNA.

These nucleotide differences were consequently used (summarised in the Table 1) were determined earlier in this analysis. For subgenotype A1 and A2 specific pgRNA second­ary structure predictions, we demonstrate that the selection and emergence of G145R based pairing of the A2 specific pgRNA in a way, which favours the emergence of G145R.

In the present work, we compared the changes in the base pairing of the pgRNA due to subgenotype A1 and A2 specific substitutions in the MHL encoding region. Based on the RNA secondary structure predictions, we demonstrate that the selection and emergence of G145R within HBV subgenotype A2 sequences in the PBL compartment occurs due to the differential base pairing characteristics in the subgenotype A2 specific pgRNA.

MATERIALS AND METHODS

Sequences for analysis

HBV surface gene sequences, corresponding to the nucleotide 341 to 660 of the HBV genome (nucleotide position counted from the unique EcoRI site in the HBV genome; GenBank accession number DQ315784 (India) for subgenotype A1 and GenBank accession number AJ309370 (France) for subgenotype A2, respectively, following the method described previously for generation of full length pgRNA sequences. The pgRNA templates so generated were unpolyadenylated and included the terminal redundancy. These two sequences served as the base sequences for prediction of secondary structures, to which nucleotide substitutions observed in the MHL encoding and flanking regions of serum associated A1 and PBL associated A2 (as mentioned in the previous section) were substituted respectively at appropriate nucleotide positions. Finally, these two template sequences (approximately 3.3 kb) were subjected to RNA secondary structure prediction and comparison.

Prediction of RNA structure

For prediction of the secondary structures, the pgRNA sequences generated as stated above were submitted to the Vienna RNA secondary structure server. The server predicts the minimum free energy (mfe) secondary structures for single RNA sequences using an algorithm proposed by Zuker and Stiegler, and also calculates the equilibrium base-pairing probabilities by means of partition function (pf) algorithm proposed by McCaskill. Apart from the mfe and pf, the server also provides a centroid structure, which indicates the reliability of the predictions, while the dot-plot which provides information on base-pairing probabilities of all the possible predicted structures. All the secondary structure predictions were performed for a temperature of 37 °C, keeping all the other parameters to default. Visualization, annotation and analysis of the mfe structures were performed using the RNAshapes program. As the present study was focused on the genetic variability of the HBV genome encoding the MHL region of the surface gene, we restricted our detailed analysis of base pairing pattern to the secondary structure of the part of pgRNA, corresponding to the MHL encoding sequence.

RESULTS

Changes in the nucleotide base pairing of the predicted secondary structure of pgRNA

The gross structural features of the subgenotype A1 and A2 specific pgRNA were found to be entirely distinct (Figure 1). The difference in the pgRNA structure was also evident from the mountain plots showing the mfe, pf, centroid, entropy and the dot plot of the two subgenotypes. The difference in other features are summarised in the Table 2. Detailed scrutiny of the pgRNA secondary structures corresponding to the MHL encoding genetic regions, revealed entirely distinct pgRNA secondary structures with discrete intra­molecular base pairing patterns due to the subgenotype specific and variations between A1 and A2 sequences (Figures 2 and 3). Interestingly, when we focussed on the base pairing of the nucleotides encoding the MHL region, we noted that in subgenotype A1 specific pgRNA, nucleotide 358 base paired with 1738 and nucleotide 587 base paired with 607.

| Nucleotide Position of the HBV genome | Base present in reference GenBank sequences | Base in genotype A1 sequences isolated from serum/plasma | Base in genotype A2 sequences isolated from PBL |
|--------------------------------------|---------------------------------------------|-------------------------------------------------------|-----------------------------------------------|
| 505                                  | c                                           | t                                                     | t                                             |
| 514                                  | c                                           | c                                                     | a                                             |
| 616                                  | a                                           | g                                                     | a                                             |
| 619                                  | t                                           | t                                                     | c                                             |

1Nucleotide positions indicate distance from the unique EcoRI site in the HBV genome; 2Sequences isolated from our study population. HBV: Hepatitis B virus; PBL: Peripheral blood leukocyte.
However in sharp contrast, in subgenotype A2 specific pgRNA, nucleotide 358 was opposite to nucleotide 588, while 587 was opposite to 359, hence precluding correct base pairing and thereby less stability of the stem structure. When the nucleotides at 358 and 587 were replaced with 358 and 587 respectively (as observed specifically in the PBL associated A2 sequences), these nucleotides base paired correctly with 588 and 359, respectively (Figure 3), forming a correctly paired stem-loop structure, hence stabilizing the local conformation. Nevertheless, the effects of other substitutions were not as influential as these two changes. The exclusive detection of subgenotype A2 sequences with the abovementioned substitutions in the PBL clearly suggest the selective advantage of the pgRNA with 358 and 587, and in turn the importance of G145R immune escape mutation in the PBL compartment.

**DISCUSSION**

In this study, we present interesting observations about the possible mechanism of compartment specific selection of immune escape HBV mutants. Based on our previous studies done on serum/plasma isolated HBV genotypes, we have documented the predominance of at least three distinct HBV genotypes in our study population, namely genotype D (most abundant) followed by genotypes C and A1. However, when we investigated the paired HBV sequences isolated from

| Table 2 Comparison of the thermodynamic characteristics of the minimum free energy secondary structure predictions for subgenotypes A1 and A2 pgRNA |
|---------------------------------|---------------|---------------|
| Features                        | Subgenotype A1 | Subgenotype A2 |
| Minimum free energy of the optimal secondary structure | -1052.10 kcal/mol | -1049.50 kcal/mol |
| Free energy of the thermodynamic ensemble | -1099.56 kcal/mol | -1098.93 kcal/mol |
| Minimum free energy of the centroid secondary structure | -722.20 kcal/mol | -679.21 kcal/mol |
| Ensemble diversity              | 863.25         | 954.99        |

Figure 1  Comparative diagram showing the differences between different aspects of the predicted secondary structures of the pgRNA, specific for HBV subgenotype A1 and subgenotype A2. A: Predicted minimum free energy (mfe) structures, coloured by base-pairing probabilities (according to the rainbow scale shown in the middle, denoting base pair probabilities from 0 to 1). Colour of the unpaired regions denotes the probability of being unpaired; B: Mountain plot representing the mfe structure (red line), the thermodynamic ensemble of RNA structures (green line), and the centroid structure (blue line). Positional entropy for each position is presented below the mountain plot; C: Dot-plot showing the base-pairing probabilities of the two predictions.
serum/plasma and the PBLs, we surprisingly observed the exclusive preponderance of the genotype A in the PBL, irrespective of the HBV genotypes circulating in the serum/plasma of any given individual\(^2\). More interestingly, the genotype A sequences isolated from the PBL was found to be markedly distinct from that of genotype A sequences isolated from serum/plasma, in terms of subgenotype and specific nucleotide sub-

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**Figure 2** Diagram showing the genetic variability and the part of the predicted mfe structure, corresponding to the genetic region encoding the major hydrophilic loop of subgenotype A1. A: Consensus nucleotide sequence (corresponding to positions 341 to 660 of the HBV genome) and predicted amino acid sequence (corresponding to residues 63 to 168 of the HBsAg). Subgenotype A1 specific nucleotides (505\(^C\), 514\(^C\), 616\(^A\) and 619\(^T\)) and two variable nucleotides (358\(^T\) and 587\(^G\)) are indicated by arrowheads; B: Detailed base pairing pattern of the mfe pgRNA structure, specific for subgenotype A1. Aforementioned variable sites are encircled by pink circles and indicated by arrows and numbers correspond to their nucleotide position in the HBV genome. HBV: Hepatitis B virus; HBsAg: Hepatitis B surface antigen.
institution patterns. More precisely, subgenotype A1 of HBV genotype A was prevalent in the serum/plasma while in sharp contrast; subgenotype A2 was solely isolated from the PBL\cite{2}.

In the present study, we sought to examine the selective advantage of subgenotype A2 in the PBL compartment with the help of advance computational prediction and analysis programs. We focussed our

Figure 3  Diagram showing the genetic variability and the part of the predicted mfe structure, corresponding to the genetic region encoding the major hydrophilic loop of subgenotype A2. A: Consensus nucleotide sequence (corresponding to positions 341 to 660 of the HBV genome) and predicted amino acid sequence (corresponding to residues 63 to 168 of the HBsAg). Subgenotype A2 specific nucleotides (505\textsuperscript{T}, 514\textsuperscript{A}, 616\textsuperscript{G} and 619\textsuperscript{C}) and two co-evolving nucleotides (358\textsuperscript{C} and 587\textsuperscript{A}) are indicated by arrowheads; B: Detailed base pairing pattern of the mfe pgRNA structure, specific for subgenotype A2. Aforementioned variable sites are encircled by pink circles and indicated by arrows and numbers correspond to their nucleotide position in the HBV genome. Part of the mfe structure is amplified in the inset for better visualization of the base pairing. HBV: Hepatitis B virus; HBsAg: Hepatitis B surface antigen.
analysis on the examination of nucleotide sequences encoding the dominant B-cell epitope (MHL) of the HBV surface antigen, since in a number of other viruses, analogous genetic regions (epitope regions of the envelope protein) have been shown to undergo faster evolution to facilitate the emergence of compartment specific immune-escape variants\[^2\]. Interestingly, when we compared serum/plasma circulating subgenotype A1 sequences with PBL confined subgenotype A2 sequences, we found that only four subgenotype specific nucleotide substitutions differentiate both sequences\[^2\]. However, all these four subgenotype specific nucleotide substitutions in the MHL encoding region were found to be synonymous in nature (i.e., the sequence of amino acids in the MHL remains same between subgenotypes A1 and A2), suggestive of the fact that MHL epitope diversity might not be directly relevant to the selection of subgenotype A2 over subgenotype A1 in PBL. On the other hand, in addition to these four subgenotype specific nucleotide substitutions, two additional nucleotide substitutions (358\[^{\text{t}}\] and 587\[^{\text{s}}\]) were evident with PBL associated A2 sequences, across the study population, which we have earlier shown to be co-evolving in the PBL\[^2\]. Interestingly, we further noted that nucleotide substitution 358\[^{\text{t}}\] was also synonymous, while substitution 587\[^{\text{s}}\] was non-synonymous and translated into the potent immune escape G145R mutation of HBsAg. Earlier studies have demonstrated that by virtue of its definite advantages, G145R mutation helps HBV to dynamically evade anti-HBs specific immune response, thereby ensuring viral persistence in anatomical compartments, which are exposed to strong anti-HBs immunity\[^2\]. The association of these five synonymous nucleotide substitutions and a potent immune escape mutation with PBL associated Ae/A2 sequences led us to hypothesize that the advantageous 587\[^{\text{s}}\] (G145R) might be selected at the pgRNA base pairing level and to verify this hypothesis, this comparative study was undertaken.

On comparison of the pgRNA secondary structures, we observed that the invariant association of subgenotype A2 with the selection of nucleotide 587\[^{\text{s}}\] (causing G145R) most possibly occur in the context of genotype A2 specific altered pgRNA base pairing patterns. Fascinatingly, we observed that substitution of a uracil (U, corresponding to Thymidine, T in DNA sequence) to cytosine (C) at position 358 altogether changed the local base pairing pattern of the pgRNA (358\[^{\text{t}}\] paired with nucleotide 588\[^{\text{t}}\] instead of the normal pairing with 1738\[^{\text{t}}\] in subgenotype A1). In the context of this altered base pairing, a single nucleotide change (U to C) at 359 was found to stabilize the stem structures by pairing with the wild type 587\[^{\text{t}}\], just opposite to it. However, the nucleotide at position 359 encodes a Cysteine residue at amino acid position 69 of HBsAg, which is extremely essential for the generation of subviral 20 nm HBsAg particles, and thus, any non-synonymous substitution at this position is most likely to be detrimental for the virus persistence\[^24\]. Therefore, based on the predicted secondary structures, we hypothesized that, instead of selecting an altered nucleotide at this exceptionally essential position (359), a compensatory alteration of a single nucleotide (G to A) at position 587 is expected to serve dual purpose, firstly it may help stabilize the stem structure (by pairing with the highly conserved 359\[^{\text{t}}\]) and secondly it results in the emergence of a potent immune escape G145R mutation, both of which appears to be highly advantageous for the virus.

The HBV polymerase lacking proof reading function has been implicated in the generation of random mutations and generation of “quasi-species”, of which the genomes (viral DNA) or pregenomes (pgRNA) having mutations useful for escaping the immune response of the host are gradually selected and subsequently become the prevalent viral population\[^28\]. Apart from virus polymerase induced random mutations, host PBL associated APOBEC (apolipoprotein B mRNA editing enzyme, catalytic polypeptide) family of cellular cytidine deaminases have also been shown to induce hyper-mutation (G to A mutations) in the HBV genome, which has been suggested to cause genetic diversification and consequently selective evolution among the divergent genomes\[^26-28\]. Nevertheless, the finding of selective predominance of the point mutation leading to G145R in the present study is highly significant in the context of PBL, since PBLs are exposed to strong anti-HBs humoral immune response and HBV variants with G145R are capable of strongly neutralizing this immune response, without any compromise in the replicative competence, thereby ensuring viral perseverance\[^26,30\]. Whatever is the source of genetic diversification, in the present work we describe a probable mechanism of RNA folding, through which divergent viral genomes/ pregenomes having favourable mutations are selected for propagation.

We acknowledge that the RNA folding predictions are based on statistical/mathematical algorithms and the biological relevance of these predictions are based on their corroboration with the biological data. Interestingly, the results of the present RNA folding predictions beautifully elucidate the observed co-evolution of the mutations at positions 358 and 587, which supports the biological relevance of the observed predictions. The results of the present study further implies that, certain HBV mutations are selected at the subgenomic RNA level (as they are synonymous at the protein level), which may significantly alter the base pairing of the pgRNA, which in turn may hasten the selection of mutations at other sites. Interestingly, the mechanism suggested in this work is very much similar to the mechanism described for HBV genotype specific selection of the most widely studied HBV precore mutation (1896\[^{\text{t}}\]), which emerges to stabilize the stem-loop structure of the epsilon “ε” signal of pgRNA\[^{\text{24,25}}\]. Altogether the present study, support the findings of Kidd-Ljunggren et al\[^{\text{30}}\], that demonstrate the implications of genotype specific differences in the pgRNA secondary structures in the emergence of genotype specific variations in the HBV genome.

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In conclusion, our results based on the predicted RNA secondary structures suggest the role of HBV genotype/subgenotype specific base pairing patterns of the pgRNA in selection/emergence of advantageous mutations. Furthermore, the observed association of a potent immune escape mutation with a particular HBV subgenotype, confined in a specific anatomical compartment indicate the possible mechanism of genotype/subgenotype specific compartmentalization of HBV, which may have important implications in extrahaepatic maintenance and transmission of HBV through hitherto unknown routes.

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