Research Article

Bathing Effects of Various Seawaters on Allergic (Atopic) Dermatitis-Like Skin Lesions Induced by 2,4-Dinitrochlorobenzene in Hairless Mice

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We evaluated the preventive effects of four types of seawater collected in Republic of Korea on hairless mice with 2,4-dinitrochlorobenzene- (DNCB-) induced allergic/atopic dermatitis (AD). The anti-inflammatory effects were evaluated by measuring tumor necrosis factor- (TNF-) α and interleukins (ILs). Glutathione (GSH), malondialdehyde (MDA), superoxide anion, and inducible nitric oxide synthase (iNOS) were measured to evaluate the antioxidant effects. Caspase-3 and poly (ADP-ribose) polymerase (PARP) were observed to measure the antiapoptotic effects; matrix metalloproteinase- (MMP-) 9 levels were also evaluated. Mice with AD had markedly higher clinical skin severity scores and scratching behaviors; higher TNF-α and ILs (1β, 10, 4, 5, and 13) levels; higher MDA, superoxide anion, caspase-3, PARP, and MMP-9 levels; and greater iNOS activity. However, the severity of AD was significantly decreased by bathing in seawaters, but it did not influence the dermal collagen depositions and skin tissue antioxidant defense systems. These results suggest that bathing in all four seawaters has protective effects against DNCB-induced AD through their favorable systemic and local immunomodulatory effects, active cytoprotective antiapoptotic effects, inhibitory effects of MMP activity and anti-inflammatory and antioxidative effects.

1. Introduction

Allergic/atopic dermatitis (AD), a chronic inflammatory skin disease associated with cutaneous hyperreactivity, affects approximately 3% of infants, 10–20% of children, and 1–3% of adults worldwide [1]. Patients with AD develop extremely itchy skin followed by severe scratching behavior that induces the production of proinflammatory cytokines [2]. This in turn activates immune cells and initiates an inflammatory cycle of AD accompanied by erythema, keratosis, and scaling [3]. All of these symptoms are the consequence of an imbalanced immune response to various allergens [4]. Another defining characteristic of the allergic immune system is the capacity to generate elevated immunoglobulin E (IgE) antibodies and type 2 helper T cells (Th2), which are critical for IgE synthesis [5]. The IgE level is associated with the severity of AD, and the
abnormal skin barrier in patients with AD, a key feature of the disease, contributes to the increased severity [6]. The functions of IgE in allergic inflammation suggest that IgE and IgE-mediated activation of mast cells and eosinophils contribute to AD. IgE can sensitize mast cells in the skin, culminating in the production of inflammatory mediators such as cytokines (interleukin- (IL-) 4, IL-5, IL-13, and tumor necrosis factor- (TNF)-α) when cell-bound IgE is cross-linked by allergens. The cytokines IL-4 and IL-13, which are released by mast cells, contribute to the Th2 response. TNF-α, which is produced by macrophages, also plays an important role in the acute phase of AD [7]. Moreover, oxidative stress is involved in the pathogenesis of AD [4].

Adrenocorticosteroids and antihistamine agents have shown favorable ameliorating effects for the treatment of AD, but they have also shown serious side effects [8]. Accordingly, researchers have explored alternative therapies and natural products in a vigorous attempt to ameliorate AD [4, 6]. Hydrotherapy is among these alternative approaches [9] and may be used as a complementary therapy. Indeed, several studies have tested different types of water baths and reported beneficial effects on dermatological disorders [10–13]. Even when the composition of the water baths differs, each has somewhat unique characteristics [14]. Hydrotherapy can reportedly modulate lymphocyte proliferation and cytokine production [14], and some compositions of mineral waters have shown favorable antioxidant effects [15–18]. Moreover, various types of seawaters have shown favorable effects on different types of dermatitis [19–21].

In the present study, we evaluated the anti-AD effects of various types of seawaters collected in Republic of Korea, namely, west surface seawater (WSSW) collected from Wepo-ri, Ganghwa-do; west saline groundwater (WSGW) collected from Yonggongoncheon, Seokmo-do; east surface seawater (ESSW) collected from Nagok-ri, Uljin; and east saline groundwater (ESGW) collected from Hoojeong-ri, Uljin, (Table 1). The anti-AD effects of these seawaters were assessed using a hairless mouse model of 2,4-dinitrochlorobenzene- (DNCB-) induced AD [22].

2. Methods

2.1. Animals and Husbandry. Total one-hundred twenty-six 6-week female SKH-1 hairless mice (OrientBio, Seongnam, Republic of Korea) were prepared, and seven groups of eight mice each were selected based on the body weights at 5 weeks after DNCB sensitization based on the body weights, clinical skin severity scores, and scratching behaviors. Animals were allocated four per polycarbonate cage in a temperature (20–25°C) and humidity (50–55%) controlled room. Light: dark cycle was 12 hr:12 hr, and standard rodent chow (Samyang, Seoul, Republic of Korea) and water were supplied free to access. All laboratory animals were treated according to the national regulations of the usage and welfare of laboratory animals and approved by the Institutional Animal Care and Use Committee in Daegu Haany University (Gyeongsan, Gyeongbuk, Republic of Korea).

2.2. Preparation of Seawaters and DEXA. Colorless clear solutions of WSSW, WSGW, and ESSW and light yellowish solution of ESWG were collected around Wepo-ri (Ganghwa-do, Republic of Korea), Yonggongoncheon (Seokmo-do, Republic of Korea), Nagok-ri (Uljin, Republic of Korea), and Hoojeong-ri (Uljin, Republic of Korea), respectively, and used after filtration with pore size 1.2 μm GF/C Grass Microfiber Filter (Korea Filter Paper Co., Ltd., Seoul, Republic of Korea) except for WSGW, which was not filtered before being used in this experiment. Salinity and mineral compositions of individual seawaters were listed in Table 1. White powders of water-soluble DEXA (Sigma-Aldrich, St. Louis, MO, USA) was obtained and used in the present study as a potent reference agent. All test materials tested in this experiment were stored at 4°C in a refrigerator to protect from light and humidity until being used. Seawaters were warmed around 37°C, at least 30 min before bathing.

2.3. Inducement of AD. AD-like dermatitis was induced by sensitization of 1% DNCB (dissolved in a 3:1 mixture of acetone and olive oil) once a day for 1 week and boosted by 0.5% DNCB, three times a week for 28 days according to established previous methods [4, 6] with some modifications. DNCB solutions were topically applied on the dorsal back skins in a volume of 200 μL/mouse. In intact control mice, only vehicle (3:1 mixtures of acetone and olive oil) was topically applied, instead of DNCB solutions in this experiment (Figure 1).

| Seawaters | Temperature (°C) | Depth (m) | Salinity (%) | Ca | Mg | K | Na |
|-----------|-----------------|-----------|--------------|----|----|---|----|
| GUGW      | 30              |           | 0.40         | 81.00 | 121.00 | 38.00 | 569.00 |
| WSSW      | 22              | 5         | 26.20        | 234.61 | 923.67 | 481.34 | 7081.18 |
| WSGW      | 70              | 700       | 22.40        | 3243.08 | 207.79 | 325.91 | 3850.67 |
| ESSW      | 21.9            | 5         | 34.00        | 417.50 | 1264.00 | 383.00 | 10672.00 |
| ESWG      | 21.7            | 689       | 26.00        | 1856.00 | 1012.00 | 38.00 | 3178.00 |

Seawaters were filtered with pore size 1.2 μm GF/C Grass Microfiber Filter (Korea Filter Paper Co., Ltd., Seoul, Republic of Korea) except for WSGW, which was not filtered before being used in this experiment. GUGW = general underground water; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggongoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESWG = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea).

Table 1: Salinity and mineral compositions of individual seawaters used in this study.
2.4. Bathing and Topical Application of DEXA. Each of eight mice per group was freely bathing on the mouse polycarbonate cages (200 × 260 × 130 mm; DJ-101, Daejong Instrument Ind. Co., Seoul, Republic of Korea) containing about 1,900 mL of warm seawaters around 37°C and fasting plates being 4 cm deep for 20 min/day. Water-soluble DEXA was dissolved in distilled water as 1% solution and topically applied on the dorsal back skins once a day for 6 weeks from 5 weeks after DNCB sensitization. Intact and DNCB control mice were bathing on the distilled water instead of seawaters to provide the same swimming stresses. Six weeks of bathing periods in this study were selected according to previous bathing effects of mineral-rich water [23] (Table 1, Figure 1).

2.5. Changes in Body Weights. Changes of body weight were measured at once a week from 1 day before initial DNCB sensitization to end of 6 weeks of continuous bathing in the 4 different types of seawaters or topical application of 1% DEXA using an automatic electronic balance (Precisa Instrument, Dietikon, Switzerland). To reduce the individual differences, the total body weight gains during 11 weeks of the whole experimental periods and body weight gains during 6 weeks of bathing or topical application of 1% DEXA were calculated as follows, respectively:

\[
\text{Total body weight gains during 11 weeks of the whole experimental periods} = \text{Body weight at sacrifice} - \text{Body weight at initial DNCB sensitization (g/head)},
\]

(1)

\[
\text{Body weight gains during 6 weeks of bathing or topical application of 1% DEXA} = \text{Body weight at sacrifice} - \text{Body weight at initial bathing or topical application of 1% DEXA (g/head)}.
\]

(2)

2.6. Evaluation of Clinical Skin Severity Scores. Five signs of skin lesions, (1) pruritus/itching, (2) erythema/hemorrhage, (3) edema, (4) excoriation/erosion, and (5) scaling/dryness, were graded as follows: 0 (no symptoms), 1 (mild), 2 (moderate), and 3 (severe), and totalized scores (max = 15) were regarded as clinical skin severity score based on the previous report [4] with some modifications. Scoring was conducted at 35, 38, 42, 49, 56, 63, 70, and 77 days after first DNCB sensitization, respectively.

2.7. Evaluation of Scratching Behavior. Each mouse was placed individually in a routine polycarbonate mouse cage and their behavior was monitored for 30 min, at 35, 38, 42, 49, 56, 63, 70, and 77 days after first DNCB sensitization, respectively. Scratching of the rostral back and biting of the caudal back were observed; scratching movements by the hind paw were defined as a scratching bout that ended when the mice either licked their hind paw or placed their hind paw back on the floor, and a series of one or more biting movements were counted as one episode that ended when the mouse returned to the straight-forward position [4].

2.8. Serum Total IgE Level Measurement. At sacrifice, about 1 mL of venous blood was collected from vena cava under anesthesia with 2 to 3% isoflurane (Hana Pharm. Co., Hwaseong, Republic of Korea) in the mixture of 70% N₂O and 28.5% O₂, and serum was separated by centrifuging at 15,000 rpm for 10 min under 4°C, using clotting activated serum tube. Total IgE levels in serum were determined by sandwich ELISA using the mouse IgE ELISA set (BD Biosciences, San Diego, CA, USA) according to previous methods [4, 6]. Briefly, plates were coated with capture antibody in
ELISA coating buffer and incubated overnight at 4°C. Plates were washed with PBS-Tween 20 (0.05%) and subsequently blocked (10% FBS in PBS) for 1 hr at 20°C. Serial dilutions of standard antigen or sample in dilution buffer (10% FBS in PBS) were added to the plates and plates were incubated for 2 hrs at 20°C. After washing, biotin-conjugated anti-mouse IgE and streptavidin-horseradish peroxidase conjugate were added to the plates and plates were incubated for 1 hr at 20°C. Finally, tetramethylbenzidine substrate solution was added to the plates and after 15 min of incubation in the dark, a 2NH2SO4 solution was added to stop the reaction. Optical densities were measured at 450 nm on an automated ELISA reader (Tecan; Männedorf, Switzerland).

2.9. Lymphatic Organ Weight Measurements. At sacrifice, 24 hrs after end of last 42th bathing, the spleen and left submandibular LN in each mouse were collected after eliminations of the surrounding connective tissues, muscles, and any debris. Individual weights of lymphatic organs were measured at g levels regarding absolute wet weights. To reduce the individual body weight differences, the relative weight (% as body weights) was calculated using body weight at sacrifice and absolute organ weights as follows according to our previously established methods [24]:

\[
\text{Relative organ weight (\% versus body weights)} = \frac{\text{Absolute organ weight}}{\text{Body weight at sacrifice}} \times 100.
\] (3)

2.10. Splenic Cytokine Content Measurements. Splenic concentrations of TNF-α, IL-1β, and IL-10 were measured by ELISA using commercially available kits, mouse TNF-α ELISA kit (BD Biosciences/Pharmingen, San Jose, CA, USA), mouse IL-1β ELISA kit (Genzyme, Westborough, MA, USA), and mouse IL-10 ELISA kit (Genzyme, Westborough, MA, USA), respectively, as previously described [25]. Approximately 10–15 mg of tissue samples were homogenized in a tissue grinder containing 1 mL of lysis buffer (PBS containing 2 mM PMSF and 1 mg/mL of aprotinin, leupeptin, and pepstatin A) as described by Clark et al. [26]. Analysis was performed with 100 mL of standard (diluted in lysis buffer) or 10, 50, or 100 mL of tissue homogenate. Each sample was run in duplicate, and a portion of the sample was analyzed for protein. Data are expressed as pg/mg of protein. For each assay a standard curve was generated and, based on replicates of the measured absorbance, demonstrated an average coefficient of variance of <10%.

2.11. Quantitative RT-PCR. Total RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA, USA), according to the method described in previous studies [6, 27]. The RNA concentrations and quality were determined by CFX96 Real-Time System (Bio-Rad, Hercules, CA, USA). To remove contaminating DNA, samples were treated with recombinant DNase I (DNA-free; Ambion, Austin, TX, USA). RNA was reverse transcribed using the reagent High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA) according to the manufacturer’s instructions. The expression of GAPDH mRNA was used as a control for tissue integrity in all samples.

2.12. GSH Assay. Cutaneous GSH levels were determined using a fluorescence assay as previously described [28]. Firstly, the skin (1:3, w/w dilution) was homogenized in 100 mM NaH2PO4 (pH 8.0; Sigma-Aldrich, St. Louis, MO, USA) containing 5 mM EDTA (buffer I). After that, homogenates were treated with 30% trichloroacetic acid (Sigma-Aldrich, St. Louis, MO, USA) and centrifuged twice (at 1,940 ×g for 6 min and at 485 ×g for 10 min) and the fluorescence of the resulting supernatant was measured in a fluorescence spectrophotometer (RF-5301PC; Shimadzu Corp., Tokyo, Japan). Briefly, 100 μL of the supernatant was mixed with 1 mL of buffer I and 100 μL of o-phenalddehyde (1 mg/mL in methanol; Sigma-Aldrich, St. Louis, MO, USA). The fluorescence was determined after 15 min (kem = 350 nm; kem = 420 nm). The standard curve was prepared with different concentrations of GSH (0.0-75.0 μM). Protein levels in the skin homogenates were measured using the method of Lowry et al. [29]. Results are presented as μM of GSH/mg of protein.

2.13. Lipid Peroxidation. Firstly, the protein content of homogenate (10 mg/mL in 1.15% KCl) was measured using the Lowry et al. method [29]. Thiobarbituric acid reactive substances (TBARS) measurement was used to evaluate lipid peroxidation as previously described [30]. For this assay, trichloroacetic acid (10%; Sigma-Aldrich, St. Louis, MO, USA) was added to the homogenate to precipitate proteins. This mixture was then centrifuged (3 min, 1,000 ×g). The protein-free sample was extracted and thiobarbituric acid (0.67%) was added. The mixture was kept in water bath at 100°C for 15 min. MDA, an intermediate product of lipoperoxidation, was determined by difference between absorbances at 535 and 572 nm on a microplate spectrophotometer reader (Tecan; Männedorf, Switzerland) and the results are reported as nM/mg of protein [31].

2.14. Superoxide Anion Production. The quantitation of superoxide anion production in tissue homogenates (10 mg/mL in 1.15% KCl) was performed using the nitro blue tetrazolium (NBT) assay [32]. Briefly, 50 μL of homogenate was incubated with 100 μL of NBT (1 mg/mL; Sigma-Aldrich, St. Louis, MO, USA) in 96-well plates at 37°C for 1 hr. The supernatant was then carefully removed and the reduced formazan solubilized by adding 120 μL of 2 M KOH and 140 μL of DMSO. The NBT reduction was measured at 600 nm using a microplate spectrophotometer reader (Tecan; Männedorf, Switzerland). The protein content was used for data normalization.

2.15. Histopathology. Samples from dorsal back skins, spleen, and left submandibular LN were separated and fixed in 10% neutral buffered formalin and then embedded in paraffin, sectioned (3–4 μm), and stained with hematoxylin and eosin (H&E) for general histopathology, Masson’s trichrome (MT) for collagen fiber, or toluidine blue for mast cells according to
our established methods [27], and after that the histopathological profiles of each sample were observed under light microscope (Nikon, Tokyo, Japan). To detail changes more, mean epithelial thicknesses in the epidermis (μm), mean numbers of inflammatory and mast cells infiltrated in the dermis (cells/mm² of dermis), total splenic thicknesses (mm/central regions), mean numbers of white pulp (white pulps/mm² of splenic parenchyma) and red pulp cells (×10⁷ cells/mm² of splenic parenchyma), total submandibular LN thicknesses (mm/central regions), cortex lymphoid follicle numbers (follicles/mm² of cortex), and mean thicknesses (μm/LN) of submandibular LN were calculated for general histomorphometric analysis using a computer-assisted image analysis program (iSolution FL, ver. 9.1, IMT i-solution Inc., Quebec, Canada) under H&E stain with collagen fiber occupied regions in the dermis (%/mm² of dermis) under MT stain, according to our previously established methods [25, 27], respectively. The histopathologist was blinded to group distribution when this analysis was made.

2.16. Immunohistochemistry. After deparaffinization of prepared skin, submandibular LN, or spleen histological paraffin sections, citrate buffer antigen (epitope) retrieval pretreatment was conducted as previously [27]. Briefly, preheat water bath with staining dish containing 10 mM citrate buffers (pH 6.0) until temperature reaches 95–100°C. Immerse slides in the staining dish and place the lid loosely on the staining dish. Allow incubation for 20 minutes and turn off the water bath. Place the staining dish at room temperature and allow the slides to cool for 20 minutes. After epitope retrievals, sections were immunostained using avidin-biotin complex (ABC) methods for caspase-3, PARP, NT, 4-HNE, MMP-9, IFN-γ, iNOS, IL-1β, IL-2, and TNF-α according to our previous studies [27, 33, 34]. Briefly, endogenous peroxidase activity was blocked by incubation in methanol and 0.3% H₂O₂ for 30 minutes, and nonspecific binding of immunoglobulin was blocked with normal horse serum blocking solution (Vector Lab., Burlingame, CA, USA, dilution 1:100) for 1 hr in humidity chamber. Primary antiserum was treated for overnight at 30 minutes, and nonspecific binding of immunoglobulin was blocked with normal horse serum blocking solution (Vector Lab., Burlingame, CA, USA, dilution 1:100) for 1 hr in humidity chamber. Primary antiserum was treated for overnight at 4°C in humidity chamber and then incubated with biotinylated universal secondary antibody (Vector Lab., Burlingame, CA, USA, dilution 1:50) and ABC reagents ( Vectastain Elite ABC Kit, Vector Lab., Burlingame, CA, USA, dilution 1:50) for 1 hr at room temperature in humidity chamber. Finally, they were incubated in peroxidase substrate reagents (Vector Lab., Burlingame, CA, USA) for 3 min at room temperature. All sections were rinsed in 0.01M PBS for 3 times, between steps. The cells or fibers occupied by over 30% of immunoreactivities, the density, of each antiserum, for caspase-3, PARP, NT, 4-HNE, MMP-9, IFN-γ, iNOS, IL-1β, IL-2, and TNF-α as compared with intact dermal keratinocytes or dermal fibers, were regarded as positive, and the mean numbers of caspase-3, PARP, NT, and 4-HNE immunoreactive cells in the epidermis (cells/100 epithelial cells), and mean IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cell numbers in the dermis (cells/mm² of dermis), spleen (cells/mm² of spleen), and submandibular LN (cells/mm² of LN) were also counted using an automated image analysis process as our established methods [27, 33, 34] with some of our modifications, respectively. In addition, the occupied percentages by MMP-9 immunoreactive fibers were also calculated in the dermis (%/mm² of dermis), as MMP-9 immunoreactivities, in this experiment. The histopathologist was blinded to the group distribution when performing the analysis.

2.17. Statistical Analyses. All data were expressed as mean ± standard deviation (SD) of eight hairless mice. Multiple comparison tests for different dose groups were conducted. Variance homogeneity was examined using the Levene test. If the Levene test indicated no significant deviations from variance homogeneity, the obtained data were analyzed by one-way ANOVA test followed by least-significant differences multicomparison (LSD) test to determine which pairs of group comparison were significantly different. In case of significant deviations from variance homogeneity was observed at Levene test, a nonparametric comparison test; Kruskal-Wallis H test was conducted. When a significant difference is observed in the Kruskal-Wallis H test, the Mann-Whitney U (MW) test was conducted to determine the specific pairs of group comparison, which are significantly different. Statistical analyses were conducted using SPSS for Windows (Release 14.0K, IBM SPSS Inc., Armonk, NY, USA). In addition, the percent changes between intact vehicle and DNCB control were calculated to observe the severities of AD-like lesions induced by DNCB in this study, and the percent changes as compared with DNCB control and hairless mice bathing in seawaters or 1% DEXA topically applied mice were also calculated to help in the understanding of the efficacy, as follows according to our previous report [35], respectively:

\[
\text{Percentage changes as compared with intact vehicle control (\%)} = \frac{\text{Data of DNCB control} - \text{Data of intact vehicle control mice}}{\text{Data of intact vehicle control mice}} \times 100, \tag{4}
\]

\[
\text{Percentage changes as compared with DNCB control (\%)} = \frac{\text{Data of test material treated mice} - \text{Data of DNCB control mice}}{\text{Data of DNCB control mice}} \times 100. \tag{5}
\]

3. Result

3.1. Changes on the Body Weight and Gains. Significantly lower body weights were demonstrated in DNCB control mice at 6 and 7 days after initial DNCB sensitization than in intact vehicle control mice (p < 0.01), but significantly higher body weights were transiently noticed at 3 weeks after the first DNCB boosting in DNCB control mice than in intact vehicle control mice (p < 0.05). Therefore, there were no significant changes in body weight gain during the total 11-week experimental period or during the 6-week bathing periods between DNCB control mice and intact vehicle control mice. In addition, neither topical application of 1% dexamethasone (DEXA) nor bathing with all four seawaters influenced the body weight or body weight gain compared to
the DNBC control mice throughout the entire experimental period (Figure 2).

3.2. Clinical Skin Severity Score Changes. Among all of the animals, AD-induced mice were selected 1 day before the initial treatment of DEXA or bathing according to the 10 clinical skin severity scores in the categories of pruritus/itching, erythema/hemorrhage, edema, excoriation/erosion, and scaling/dryness. The clinical skin severity scores were significantly higher in the DNBC control mice than in the intact vehicle control mice from 24 h before initial bathing or topical application of 1% DEXA to the end of the experimental period. However, these increases in clinical skin severity scores were significantly lower in the intact vehicle control mice than in the DNBC control mice from 1 week after the initial topical application of 1% DEXA or bathing in WSGW ($p < 0.01$ and $p < 0.05$, resp.). In addition, significantly lower clinical skin severity scores were detected in the intact vehicle control mice than in the DNBC control mice from 2 weeks after initial bathing with ESGW and from 3 weeks after initial bathing with WSSW and ESSW ($p < 0.01$ and $p < 0.05$, resp.) (Table 2).

3.3. Changes on the Scratching Behaviors. As noted above, AD-induced mice were selected 1 day before the initial treatment of DEXA or bathing; 4 weeks after the initial DNBC boosting, those with >300 episodes of head-scratching behavior per 30 min along with increases in the 10 clinical skin severity scores were selected. DNBC control mice showed significantly greater increases in scratching behavior than intact vehicle control mice from 24 h before initial bathing or topical application of 1% DEXA until the end of the experimental period ($p < 0.01$). However, these increases in scratching behavior were significantly less marked in intact vehicle control mice than in DNBC control mice from 1 week after the initial topical application of 1% DEXA until the end of the experimental period ($p < 0.01$ or $p < 0.05$). In addition, significantly less scratching behavior was detected in intact vehicle control mice than in DNBC control mice from 2 weeks after the initial bathing in WSGW and ESSW and from 4 weeks after the initial bathing in WSSW and ESSW until the end of the experimental period ($p < 0.01$ and $p < 0.05$, resp.) (Table 3).

3.4. Effects on the Serum Total IgE Levels. Significantly higher serum total IgE levels were detected in DNBC control mice than in intact vehicle control hairless mice ($p < 0.01$). However, significantly lower serum total IgE levels were detected in intact vehicle control mice than in DNBC control mice after topical treatment with 1% DEXA and bathing in ESGW, WSGW, ESSW, and WSSW, in that order ($p < 0.01$) (Figure 3). The serum total IgE levels in DNBC control mice were 249.23% higher than those in the intact vehicle control mice, and these levels decreased by 57.63%, 20.50%, 32.30%, 29.46%, and 41.55% after topical application of 1% DEXA and bathing in WSSW, WSGW, ESSW, and ESGW, respectively.

3.5. Changes on the Submandibular LN and Spleen Weights. Significantly higher submandibular lymph node (LN) and absolute and relative spleen weights were detected in DNBC control mice than in intact vehicle control hairless mice ($p < 0.01$). However, significantly lower absolute and relative submandibular LN and spleen weights were detected in intact vehicle control hairless mice than in DNBC control mice after topical treatment with 1% DEXA and bathing in ESGW, WSGW, ESSW, and WSSW, in that order ($p < 0.01$) (Table 4).

3.6. Effects on the Splenic Cytokine Contents. Significantly higher splenic tissue levels of TNF-α, IL-1β, and IL-10 were detected in DNBC control mice than in intact vehicle control hairless mice ($p < 0.01$). However, significantly lower splenic tissue levels of TNF-α, IL-1β, and IL-10 were detected in intact vehicle control hairless mice than in DNBC control mice after topical
Table 2: Changes on the clinical skin severity scores during 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Index | Treatments | Controls | DEXA | Bathing on |
|-------|------------|----------|------|------------|
|       | Intact     | DNCB     | WSSW | WSGW | ESSW | ESGW |
| Days after initial DNCB sensitization |            |          |      |      |      |      |
| 35    | 1.00 ± 0.76 | 12.13 ± 1.55a | 12.25 ± 1.39a | 12.25 ± 1.83a | 12.00 ± 1.51a | 12.13 ± 1.25a |
| 38    | 1.13 ± 0.99 | 11.75 ± 1.28a | 11.00 ± 1.51a | 11.38 ± 1.41a | 11.13 ± 0.99a | 11.00 ± 1.20a |
| 42    | 0.75 ± 0.71 | 11.50 ± 1.41a | 8.75 ± 1.28ab | 10.75 ± 1.04a | 10.25 ± 1.16ac | 10.50 ± 1.20a |
| 49    | 1.00 ± 0.53 | 10.63 ± 1.41a | 7.00 ± 1.20ab | 9.88 ± 0.83a  | 8.88 ± 1.13ab | 9.75 ± 1.16a  |
| 56    | 1.00 ± 0.53 | 10.13 ± 1.46a | 6.13 ± 1.13ab | 9.00 ± 1.20ac | 8.13 ± 0.99ab | 9.00 ± 1.20ac |
| 63    | 1.25 ± 0.71 | 9.75 ± 1.49a | 4.75 ± 1.49ab | 7.75 ± 1.04ab | 6.88 ± 1.25ab | 7.63 ± 0.92ab |
| 70    | 1.13 ± 0.64 | 9.13 ± 1.46a | 4.00 ± 1.31ab | 7.25 ± 0.71ab | 6.25 ± 1.04ab | 6.88 ± 0.64ab |
| 77    | 2.00 ± 0.76 | 8.38 ± 1.30a | 3.50 ± 1.07ab | 6.25 ± 0.71ab | 5.25 ± 1.16ab | 5.88 ± 0.64ab |

Values are expressed as mean ± SD of eight hairless mice, scores (max = 15). AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea). *p < 0.01 as compared with intact control by LSD test; †p < 0.01 and ‡p < 0.05 as compared with DNCB control by LSD test.

3.7. Changes on the Skin Tissue Cytokine mRNA Expressions (RT-PCR Analysis). Significantly higher TNF-α, IL-4, IL-5, and IL-13 mRNA expression levels in the dorsal back skin were detected in DNCB control mice compared to intact control hairless mice, as determined by RT-PCR analysis (p < 0.01). However, significantly lower TNF-α, IL-4, IL-5, and IL-13 mRNA expression levels in the dorsal back skin were detected in intact control hairless mice than in DNCB control mice after topical treatment with 1% DEXA and bathing in ESGW, WSGW, ESSW, and WSSW, in that order (p < 0.01 and p < 0.05, resp.) (Table 5).

3.8. Effects on the Skin Tissue Antioxidant Defense Systems. Significantly lower glutathione (GSH) levels and significantly higher lipid peroxidation and superoxide anion production levels in the dorsal back skin were detected in DNCB control mice than in intact control hairless mice (p < 0.01). However, significantly higher GSH levels and lower lipid peroxidation and superoxide anion production levels in the dorsal back skin were detected in intact control hairless mice than in DNCB control mice after bathing in ESGW, WSGW, ESSW, and WSSW, in that order (p < 0.01 or p < 0.05). Topical application of 1% DEXA did not influence the skin tissue expression.
Values are expressed as mean ± SD of eight hairless mice, frequencies/30 min. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea). a as compared with DNCB control by LSD test; b p < 0.05 as compared with intact control by LSD test; c p < 0.01 as compared with DNCB control by LSD test.

3.9. Histopathological Changes on the Dorsal Back Skin Tissues. Histopathological signs of AD-related hypersensitivity were significantly lower in intact control hairless mice than in DNCB control mice after bathing in ESGW, WSGW, ESSW, and WSSW, in that order (p < 0.01 or p < 0.05). Topical application of 1% DEXA also resulted in significantly lower increases in the mean epithelial thicknesses, numbers of mast and inflammatory cells infiltrating the dermis, caspase-3, PARP, NT, and 4-HNE immunoreactive epidermal cells, dermal MMP-9 immunoreactivity, dermal IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cells (p < 0.01) but did not influence the percentages of collagen fibers occupied dermal regions (Tables 8 and 9, Figures 4–6).

3.10. Histopathological Changes on the Splenic Tissues. Significantly higher total splenic thickness, number of red pulp lymphoid cells, and numbers of IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cells were observed in DNCB control mice than in intact vehicle control hairless mice (p < 0.01). However, these hypersensitivity-related signs of splen hypertrophy were significantly lower in intact vehicle control hairless mice than in DNCB control mice after topical treatment of 1% DEXA and bathing in ESGW, WSGW, ESSW, and WSSW, in that order (p < 0.01 and p < 0.05, resp.). No meaningful changes in the number of white pulp cells were demonstrated after treatment with DNCB, topical application of 1% DEXA, or bathing in any of the four types of seawater (Table 10, Figures 7 and 8).

3.11. Histopathological Changes on the Submandibular LN Tissues. Significantly higher total submandibular LN thickness, number of cortex lymphoid follicles, cortex thickness, and numbers of IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cells in the submandibular LN tissues were observed in DNCB control mice than in intact vehicle control hairless mice (p < 0.01). However, these hypersensitivity-related signs of submandibular LN hypertrophy were significantly inhibited by topical treatment with 1% DEXA and bathing in
Table 5: Changes on the splenic cytokine contents after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Groups       | Tumor necrosis factor-α | Splenic cytokine contents (pg/mg protein) | Interleukin-10 |
|--------------|-------------------------|------------------------------------------|----------------|
|              |                         | Splenic cytokine contents (pg/mg protein)|                |
|              |                         | Tumor necrosis factor-α                  |                |
| Controls     |                         | 164.92 ± 23.50                          | 216.95 ± 54.25 |
|              |                         | 530.62 ± 113.24                          | 688.86 ± 131.07 |
| Reference    |                         | 198.70 ± 19.33                           |                |
| Bathing on   |                         | 373.00 ± 39.06                           |                |
| WSSW         |                         | 322.99 ± 67.87                           |                |
| WSGW         |                         | 352.92 ± 64.20                           |                |
| ESSW         |                         | 296.31 ± 65.51                           |                |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungancheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); LSD = least-significant differences multiple-comparison. a p < 0.01 as compared with intact control by LSD test; b p < 0.01 as compared with DNCB control by LSD test; c p < 0.01 and d p < 0.05 as compared with intact control by MW test; e p < 0.01 and f p < 0.05 as compared with DNCB control by MW test.

Table 6: Changes on the skin mRNA expressions after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Groups       | Skin mRNA expressions (relative expressions/GAPDH mRNA) | Spleen                        | Submandibular LN |
|--------------|---------------------------------------------------------|-------------------------------|------------------|
|              |                                                         |                               |                  |
| Controls     |                                                         | 1.03 ± 0.08                   | 1.01 ± 0.05      |
|              |                                                         | 5.62 ± 1.22                   | 4.94 ± 0.74      |
| Reference    |                                                         | 2.74 ± 1.20                   | 1.88 ± 0.56      |
| Bathing on   |                                                         | 4.18 ± 0.52                   | 3.87 ± 0.60      |
| WSSW         |                                                         | 3.46 ± 0.77                   | 3.06 ± 0.50      |
| WSGW         |                                                         | 3.94 ± 0.62                   | 3.59 ± 0.66      |
| ESSW         |                                                         | 2.82 ± 1.22                   | 2.55 ± 0.79      |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungancheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); GAPDH = glyceraldehyde 3-phosphate dehydrogenase. a p < 0.01 as compared with intact control by MW test; b p < 0.01 and c p < 0.05 as compared with DNCB control by MW test.

ESGW, WSGW, ESSW, and WSSW, in that order (p < 0.01) (Table 11, Figures 9 and 10).

4. Discussion

In this study, we examined the anti-AD effects of various types of seawater collected from different regions in Republic of Korea in a hairless mouse model of DNCB-induced AD after 6 weeks of bathing for 20 min once a day for 42 days. The results were compared to those after topical application of 1% DEXA.

Generally, marked decreases in body weight were noted during DNCB sensitization, but the body weights increased despite the development of hypersensitivity after chronic and repeated DNCB exposure. Therefore, DNCB sensitization and boosting did not critically influence the total body weight gains of mice [6, 36]. In the present study, significantly lower body weights were also demonstrated in DNCB control mice at 6 and 7 days after initial DNCB sensitization than in intact vehicle control mice, but significantly higher body weights were transiently noticed at 3 weeks after the first DNCB boosting in DNCB control mice than in intact vehicle control mice. Therefore, no significant changes in body weight gains were noted between the DNCB control mice and intact vehicle control mice during the total 11-week experimental period or 6-week bathing period; these findings are quite similar to those of previous studies [6, 36]. In addition, topical application of 1% DEXA and bathing in all four different...
Figure 4: Continued.
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Figure 4: Representative histological images of dorsal back skin tissues, taken from unexposed intact or DNCB-induced AD mice bathing on seawaters or topical application of DEXA. Marked increases of mean epithelial thicknesses due to hyperplasia/hypertrophy of epidermal keratinocytes were detected on the dorsal back skin tissues in DNCB control mice with noticeable increases of the numbers of mast and inflammatory cells infiltrated into dermis and abnormal collagen depostions, respectively. However, these histopathological hypersensitivities related AD signs were significantly inhibited by bathing on the ESGW, WSGW, ESSW, and WSSW as compared with DNCB control mice, in that order, respectively. Topical application of 1% DEXA also significantly reduced the increases of mean epithelial thicknesses, numbers of dermal infiltrated mast, and inflammatory cells induced by DNCB treatment but did not influence the percentages of collagen fiber occupied dermal regions as compared with DNCB control mice, in this experiment. (a) Intact vehicle control mice bathing on the distilled water; (b) DNCB control mice bathing on the distilled water; (c) AD mice bathing on the WSSW; (d) AD mice bathing on the WSGW; (e) AD mice bathing on the ESSW; (f) AD mice bathing on the ESGW; (g) 1% DEXA topically applied AD mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); EP = epidermis; DE = dermis; MT = Masson's trichrome. Scale bars = 40 μm.

Table 7: Changes on the skin antioxidant defense systems after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Groups       | Glutathione (μM/mg of protein) | Lipid peroxidation-malondialdehyde (nM/mg of protein) | Superoxide anion production (NBT reduction/OD at 600 nm) |
|--------------|--------------------------------|-------------------------------------------------------|--------------------------------------------------------|
| Controls     |                                |                                                       |                                                        |
| Intact       | 1.49 ± 0.27                    | 0.38 ± 0.13                                           | 0.41 ± 0.12                                            |
| DNCB         | 0.44 ± 0.17                    | 2.66 ± 0.70d                                          | 1.68 ± 0.27d                                           |
| Reference    |                                |                                                       |                                                        |
| DEXA         | 0.43 ± 0.13                   | 2.58 ± 0.87d                                          | 1.57 ± 0.62d                                           |
| Bathing on   |                                |                                                       |                                                        |
| WSSW         | 0.67 ± 0.10c                   | 1.83 ± 0.35df                                         | 1.28 ± 0.26df                                          |
| WSGW         | 0.83 ± 0.22ab                  | 1.38 ± 0.22de                                         | 1.03 ± 0.22de                                          |
| ESSW         | 0.76 ± 0.18ab                  | 1.72 ± 0.20df                                         | 1.14 ± 0.17de                                          |
| ESGW         | 1.01 ± 0.27ab                  | 1.03 ± 0.31de                                         | 0.73 ± 0.25de                                          |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); OD = optical density; NBT = nitro blue tetrazolium; LSD = least-significant differences multicomparison. *p < 0.01 as compared with intact control by LSD test; †p < 0.01 and ‡p < 0.05 as compared with DNCB control by LSD test; ‡p < 0.01 as compared with intact control by MW test; §p < 0.01 and ¶p < 0.05 as compared with DNCB control by MW test.

seawaters did not influence the body weight or body weight gains compared to DNCB control mice throughout the entire experimental period.

The skin lesions in patients with AD are generally characterized by infiltration of various inflammatory cells such as mast cells, basophils, eosinophils, and T cells [1, 37]. AD is also associated with several common symptoms including itching, erythema, eczematous skin lesions, chronic relapse, and pruritus [2]. Application of DNCB onto the skin also causes inflammation and dermal sclerosis along with these common symptoms [38]. Therefore, clinical skin severity scores based on the five main skin lesions and symptoms (pruritus/itching, erythema/hemorrhage, edema, excoriation/erosion, and scaling/dryness) have been used as valuable predictor of progression of AD with scratching behavior [4]. Mast cells mediate inflammatory responses such as
Figure 5: Continued.
hypersensitivity and allergic reactions, and the allergen cross-linking of surface IgE-dependent mast cell activation stimulates degranulation and release of histamine, leukotrienes, proteases, prostaglandins, and cytokines [6]. Activated mast cells release a variety of inflammatory mediators following cross-linking of IgE-receptor complexes at the high-affinity IgE receptor I. Of these mediators, histamine is generally considered to be a marker of mast cell degranulation in immediate allergic reactions and is a potent inducer of itching. Histamine is a characteristic major mediator in mast cell storage granules and directly triggers type I allergic responses [6, 39]. In the present study, IgE-mediated hypersensitivity, dermal sclerosis, and inflammatory and mast cell infiltration induced by DNBC treatment were significantly decreased by bathing in ESWG, WSWG, ESSW, and WSSW, in that order, and by topical treatment of 1% DEXA. These findings suggest that bathing in these four different types of seawaters around Republic of Korea can inhibit the symptoms of DNCB-induced dermatitis.

The pathogenesis of AD is complex, involving genetic, environmental, and immunological factors. In particular, IL-4, IL-5, and IL-13, which are produced by Th2 cells, may have especially key roles in the onset and development of AD [40]. Although the etiology and pathology of AD are not fully understood, a recent study reported that typical symptoms of AD involve increased levels of Th2-mediated cytokines [41]. Th2 cells are dominant during the acute phase of AD, whereas Th1 cells are dominant and contribute to pathogenesis during the chronic phase [42]. An elevated IgE level is a hallmark of AD, and the expression of IL-4 contributes to this elevation. IL-4 stimulates IgE production in B cells. IgE released from B cells binds...
Figure 6: Continued.
to mast cells, which then degranulate and release various biological mediators in patients with IgE-mediated AD [43]. AD is dependent upon the secretion of the cytokines IL-4, IL-5, and IL-13 by Th2 cells that are generated from precursors. Most patients with AD have increased eosinophils and IgE levels due to elevated IL-4, IL-5, and IL-13 produced by Th2 cells [44]. TNF-α is a well-known proinflammatory cytokine [45] and is markedly increased in DNCB-associated dermatitis [46]. IL-2 is normally produced by T cells during an immune response [47] and has a well-documented role in the induction of pruritus in AD [48]. IFN-γ is a 20 kDa to 25 kDa glycoprotein produced by CD8+ T cells and natural killer (NK) cells in response to IL-2. IFN-γ has complex effects on B- and T-cell functions and enhances NK cell and macrophage activities [49]. Increases in IFN-γ activities have also been observed in DNCB-induced dermatitis [50]. Increases in iNOS activities related to the proinflammatory agents endotoxin, IL-1β, TNF-α, and IFN-γ can induce shock.

Table 9: Immunohistochemical skin tissues histomorphometry after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Antiserum | Controls | DEXA | Bathing on | DEXA |
|-----------|----------|------|------------|------|
| Caspase-3 | 3.13 ± 1.96 | 85.00 ± 7.33 | 37.00 ± 10.62 | 54.25 ± 10.62 |
| PARP      | 6.50 ± 2.20 | 85.00 ± 10.01 | 21.38 ± 5.40 | 48.00 ± 14.58 |
| Nitrotyrosine | 16.38 ± 3.74 | 87.63 ± 6.55 | 16.63 ± 3.50 | 42.50 ± 10.63 |
| 4-HNE     | 3.63 ± 1.41 | 85.38 ± 12.78 | 24.25 ± 7.03 | 64.13 ± 11.47 |
| MMP-9     | 10.37 ± 5.62 | 55.34 ± 10.24 | 38.74 ± 10.80 | 34.69 ± 7.06 |
| IFN-γ     | 7.42 ± 2.04 | 138.52 ± 25.95 | 39.46 ± 15.48 | 83.71 ± 13.62 |
| iNOS      | 6.68 ± 2.14 | 237.05 ± 28.70 | 62.15 ± 20.37 | 133.28 ± 39.82 |
| IL-1β     | 4.83 ± 2.37 | 185.70 ± 26.59 | 41.73 ± 14.26 | 129.77 ± 31.57 |
| IL-2      | 3.52 ± 1.75 | 86.46 ± 20.19 | 17.15 ± 6.12 | 30.47 ± 11.94 |
| TNF-α     | 5.22 ± 1.29 | 65.15 ± 14.79 | 16.42 ± 5.19 | 24.26 ± 3.42 |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); IFN = interferon; IL = interleukin; iNOS = inducible nitric oxide synthase (2); TNF = tumor necrosis factor; ABC = avidin-biotin complex. All being ABC immunostain. Scale bars = 40 μm.

FIGURE 6: Representative immunohistochemical images of dermal IFN-γ, iNOS, IL-1β, IL-2, and TNF-α in the dorsal back skin tissues, taken from unexposed intact or DNCB-induced AD mice bathing on seawaters or topical application of DEXA. Dramatic infiltrations of dermal IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cells were observed in DNCB control mice as compared with intact vehicle control hairless mice, respectively. However, these increases of immunoreactivities of IFN-γ, iNOS, IL-1β, IL-2, and TNF-α in the dermis were significantly inhibited by bathing on the ESGW, WSWG, ESSW, and WSSW as compared with DNCB control mice, in that order, respectively. Topical application of 1% DEXA also significantly reduced the increases of dermal IFN-γ, iNOS, IL-1β, IL-2, and TNF-α immunolabeled cells induced by NDNCB treatment as compared with DNCB control mice, in this experiment. (a) Intact vehicle control mice bathing on the distilled water; (b) DNCB control mice bathing on the distilled water; (c) AD mice bathing on the WSSW; (d) AD mice bathing on the WSGW; (e) AD mice bathing on the ESSW; (f) AD mice bathing on the ESGW; (g) 1% DEXA topically applied AD mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); IFN = interferon; IL = interleukin; iNOS = inducible nitric oxide synthase (2); TNF = tumor necrosis factor; ABC = avidin-biotin complex. All being ABC immunostain. Scale bars = 40 μm.
Figure 7: Continued.
and inflammatory responses in the body [51], and overexpression of iNOS is also involved in the pathogenesis of AD [52]. Therefore, downregulation of iNOS, IL-2, TNF-α, and IFN-γ expression has been used to predict the favorable effects of test materials in patients with various allergic diseases. In addition, AD induces systemic hypersensitivity and marked proliferation of central and peripheral lymphocytes, especially T cells [53]. In the present study, these systemic and local hypersensitivities induced by DNCB treatment were significantly inhibited by bathing in ESGW, WSGW, ESSW, and WSSW, in that order, as well as by topical treatment of 1% DEXA. These findings are considered to be direct evidence that bathing in these four different types of seawater in Republic of Korea inhibits the symptoms of DNCB-induced dermatitis through their potent systemic and local immunomodulatory effects. A previous study found that hydrotherapy can modulate lymphocyte and cytokine production [14].

AD leads to an imbalance between ROS and endogenous antioxidants, causing depletion of endogenous antioxidants such as GSH [18, 54]. GSH is a representative endogenous antioxidant that prevents tissue damage by maintaining low

![Image](image_url)

**Figure 7:** Representative histological images of splenic tissues, taken from unexposed intact or DNCB-induced AD mice bathing on seawaters or topical application of DEXA. Marked hypertrophic changes due to hyperplasia of red pulp lymphoid cells were detected on the splenic tissues in DNCB control mice as compared with intact vehicle control hairless mice, respectively. However, these hypersensitivities related splenic hypertrophic signs were significantly inhibited by topical treatment of 1% DEXA and bathing on the ESGW, WSGW, ESSW, and WSSW as compared with DNCB control mice, in that order, respectively. No meaningful changes on the numbers of white pulps were demonstrated by treatment of DNCB as compared with intact vehicle control and also by topical application of 1% DEXA or bathing on the all four types of seawaters as compared with DNCB control mice, in this experiment. (a) Intact vehicle control mice bathing on the distilled water; (b) DNCB control mice bathing on the distilled water; (c) AD mice bathing on the WSSW; (d) AD mice bathing on the WSGW; (e) AD mice bathing on the ESSW; (f) AD mice bathing on the ESGW; (g) 1% DEXA topically applied AD mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggunoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hojojeong-ri (Uljin, Republic of Korea); WP = white pulp; RP = red pulp. All being hematoxylin-eosin stain. Scale bars = 400 μm.

**Table 10:** Splenic tissues histomorphometry after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Index                  | Controls | DEXA | Bathing on |
|------------------------|----------|------|------------|
|                       |          |      | WSSW       |
|                       |          |      | WSGW       |
|                       |          |      | ESSW       |
|                       |          |      | ESGW       |
| Total Th               | 1.55 ± 0.28 | 3.42 ± 0.59 | 1.67 ± 0.29 | 2.34 ± 0.39 | 1.92 ± 0.25 | 2.08 ± 0.25 | 1.73 ± 0.12 |
| White pulp            | 12.75 ± 1.49 | 14.00 ± 1.69 | 12.88 ± 2.03 | 14.38 ± 2.13 | 13.00 ± 1.51 | 13.25 ± 1.67 | 13.63 ± 2.72 |
| Red pulp cells        | 2.91 ± 0.87 | 29.40 ± 7.33 | 5.09 ± 2.65 | 21.83 ± 3.20 | 15.54 ± 2.15 | 18.37 ± 2.19 | 11.21 ± 4.27 |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggunoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hojojeong-ri (Uljin, Republic of Korea); Th = thickness; IFN = interferon; IL = interleukin; iNOS = inducible nitric oxide synthase (2); TNF = tumor necrosis factor. *p < 0.01 and **p < 0.05 as compared with intact control by MW test; ***p < 0.01 and ****p < 0.05 as compared with DNCB control by MW test.
Figure 8: Continued.
levels of ROS and acts as a protective antioxidant factor in tissues [55]. GSH also acts as a cofactor for glutathione peroxidase and glutathione reductase, which reduce hydrogen peroxide and lipid hydroperoxides [56]. 4-HNE is an \(\alpha,\beta\)-unsaturated hydroxyalkenyl produced by lipid peroxidation in cells and has been used as a valuable tissue lipid peroxidation marker. It is currently being considered a possible causal agent of numerous diseases [57]. NT is a product of tyrosine nitration mediated by reactive nitrogen species such as peroxynitrite anion and nitrogen dioxide. It is detected in many pathological conditions and is a marker of iNOS-dependent, reactive nitrogen species-induced nitrative stress [58, 59]. In the present study, depletion of endogenous antioxidants and increases in oxidative stress were significantly inhibited by bathing in ESGW, WSGW, ESSW, and WSSW, in that order. Hydrotherapy has potent antioxidant effects in various types of dermatitis [15–18]. In further support of the antioxidant effects of bathing in WSSW, WSGW, ESSW, and ESGW, all four types of seawater inhibited lipid peroxidation as determined by the MDA concentration of the TBARS assay.

**Table 11:** Submandibular LN tissues histomorphometry after 6 weeks of continuous bathing on seawaters or topical application of DEXA in DNCB-induced AD mice.

| Index                  | Controls | DEXA    | Bathing on |
|------------------------|----------|---------|------------|
|                        | Intact   | DNCB    | WSSW       |
|                        |          |         | WSGW       |
|                        |          |         | ESSW       |
|                        |          |         | ESGW       |
| Total Th               | 0.73±0.17| 1.70±0.39| 0.98±0.20  |
|                        |          | 1.34±0.14| 1.09±0.17  |
|                        |          | 1.28±0.22| 0.97±0.12  |
| Follicles Th           | 11.38±2.77| 35.00±5.81| 14.50±2.73 |
|                        |          | 28.25±5.60| 22.88±3.60 |
|                        |          | 25.25±5.75| 14.63±2.07 |
| Cortex Th              | 353.27±104.03| 981.34±123.88| 399.76±98.96 |
|                        |          | 822.82±100.84| 753.92±123.13 |
|                        |          | 807.70±119.32| 528.73±128.20 |
| Immune reactive cells  |          |         |            |
| TNF-\(\gamma\)         | 7.52±2.03| 131.36±24.32| 20.13±11.80 |
|                        |          | 94.60±12.60| 65.21±23.35 |
|                        |          | 91.63±14.44| 38.52±11.13 |
| IFN-\(\gamma\)         | 7.80±5.27| 206.72±15.82| 18.97±5.99  |
|                        |          | 157.95±25.35| 79.41±15.14 |
|                        |          | 112.41±14.85| 46.65±27.73 |
| iNOS                   | 8.65±4.11| 153.55±24.80| 19.38±7.03  |
|                        |          | 113.57±21.64| 64.26±12.58 |
|                        |          | 82.10±26.17| 37.52±15.17 |
| IL-1\(\beta\)          | 15.07±6.75| 123.60±20.31| 43.47±12.23 |
|                        |          | 63.23±95.66| 78.81±18.19 |
|                        |          | 42.50±14.74|            |
| IL-2                   | 2.85±1.54| 84.57±17.53| 15.30±5.39  |
|                        |          | 45.56±13.55| 29.20±7.20  |
|                        |          | 31.96±8.98| 21.36±7.02  |

Values are expressed as mean ± SD of eight hairless mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); IFN = interferon; IL = interleukin; iNOS = inducible nitric oxide synthase (2); TNF = tumor necrosis factor; ABC = avidin-biotin complex. All being ABC immunostain. Scale bars = 40 \(\mu\)m.
Figure 9: Continued.
In the present study, these increases in apoptotic effect by modulating the activities of caspase-3 and PARP. The results obtained in this study suggest that bathing on the all four different types of seawaters collected around Republic of Korea has strong inhibitory activity against DNCB-induced skin sclerosis by modulating the activities of MMP-9.

5. Conclusion

The results obtained in this study suggest that bathing on the all four different types of seawaters collected around Republic of Korea has favorable protective effects against DNCB-induced AD through their favorable systemic and local immunomodulatory effects, active cytoprotective anti-apoptotic effects, inhibitory effects of MMP activity, and anti-inflammatory and antioxidative effects, in the order of ESGW, WSGW, ESSW, and WSSW, at least in the condition of this experiment. These different efficacies along different collecting regions may be directly related to the different salinity and mineral compositions. Therefore, it is expected that bathing on the seawaters modulated their salinity and mineral compositions.

Figure 9: Representative histological images of submandibular LN tissues, taken from unexposed intact or DNCB-induced AD mice bathing on seawaters or topical application of DEXA. Noticeable hypertrophic changes due to hyperplasia of cortex lymphoid cells were detected on the submandibular LN tissues in DNCB control mice as compared with intact vehicle control hairless mice, respectively. However, these submandibular LN hypersensitivities related hypertrophic signs were significantly inhibited by topical treatment of 1% DEXA and bathing on the ESGW, WSGW, ESSW, and WSSW as compared with DNCB control mice, in that order, respectively. (a) Intact vehicle control mice bathing on the distilled water; (b) DNCB control mice bathing on the distilled water; (c) AD mice bathing on the WSSW; (d) AD mice bathing on the WSGW; (e) AD mice bathing on the ESSW; (f) AD mice bathing on the ESGW; (g) 1% DEXA topically applied AD mice. AD = allergic/atopic-like dermatitis; DNCB = 2,4-dinitrochlorobenzene; DEXA = dexamethasone-water soluble; WSSW = west surface seawater collected around Wepo-ri (Ganghwa-do, Republic of Korea); WSGW = west saline groundwater collected at Yonggungoncheon (Seokmo-do, Republic of Korea); ESSW = east surface seawater collected around Nagok-ri (Uljin, Republic of Korea); ESGW = east saline groundwater collected around Hoojeong-ri (Uljin, Republic of Korea); LN = lymph node; CO = cortex; FO = lymphoid follicle; ME = medulla. All being hematoxylin-eosin stain. Scale bars = 400 μm.
Figure 10: Continued.
mineral compositions maybe serve as a predictable alternative therapy in AD patients in future.

Conflict of Interests
The authors declare that they have no conflict of interests.

Authors’ Contribution
Choong Gon Kim and Meehye Kang contributed equally to this work.

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