RESEARCH ARTICLE

Lateral orbitofrontal cortex partitions mechanisms for fear regulation and alcohol consumption

Madelyn H. Ray*, Emma Hanlon, Michael A. McDannald

Department of Psychology, Boston College, Chestnut Hill, Massachusetts, United States of America

* madelyn.ray@bc.edu

Abstract

Anxiety disorders and alcohol use disorder are highly comorbid, yet identifying neural dysfunction driving comorbidity has been challenging. Lateral orbitofrontal cortex (IOFC) dysfunction has been independently observed in each disorder. Here we tested the hypothesis that the IOFC is essential to partition mechanisms for fear regulation and alcohol consumption. Specifically, the capacity to regulate fear and the propensity to consume alcohol are unrelated when IOFC is intact, but become linked through IOFC dysfunction. Male Long Evans rats received bilateral, neurotoxic IOFC lesions or sham surgery. Fear regulation was determined by establishing discrimination to danger, uncertainty, and safety cues then shifting the shock probability of the uncertainty cue. Alcohol consumption was assessed through voluntary, intermittent access to 20% ethanol. The neurotoxic lesion approach ensured IOFC dysfunction spanned testing in fear regulation and alcohol consumption. IOFC-lesioned rats demonstrated maladaptive fear generalization during probability shifts, inverting normal prediction error assignment, and subsequently consumed more alcohol. Most novel, fear regulation and alcohol consumption were inextricably linked only in IOFC-lesioned rats: extreme fear regulation predicted excessive alcohol consumption. The results reveal the IOFC is essential to partition mechanisms for fear regulation and alcohol consumption and uncover a plausible source of neural dysfunction contributing to comorbid anxiety disorders and alcohol use disorder.

Introduction

Anxiety disorders and alcohol use disorder are highly comorbid [1–4]. Despite this clear clinical relationship, the neural mechanisms mediating comorbid anxiety and alcohol use are only beginning to be understood. Orbitofrontal cortex (OFC) function is altered or disrupted in anxiety disorders [5, 6], as well as alcohol-use disorder [7–9]. Specifically, OFC hypoactivity is seen in people with anxiety disorders, such as PTSD [6], in addition to people with alcohol use disorder [7, 10]. This commonality may mark OFC dysfunction, via hypoactivity, as a comorbid link between anxiety disorders and alcohol-use disorder.

Consistent with dysfunction in psychiatric disorders, previous studies have demonstrated independent roles for the OFC in fear regulation [11–16] and alcohol consumption [17–19].
OFC lesions lead to generalized fear increases to shock-free contexts during contextual fear discrimination [13, 20], while OFC inhibition via DREADDs results in persistent cued fear in the face of extinction [15]. OFC lesions or DREADD inhibition increases voluntary alcohol consumption in alcohol-dependent mice [17]. Thus, the OFC normally works to reduce fear and moderate alcohol consumption.

We propose that while individually contributing to each, an essential function of the OFC, particularly the lateral OFC (lOFC), is to partition neurobehavioral mechanisms for fear regulation and alcohol consumption. A salient prediction of our hypothesis is that lOFC dysfunction should result in failure of this partition, linking the capacity to regulate fear and propensity to consume alcohol. We induced IOFC dysfunction via neurotoxic lesion in one group of rats, and left IOFC intact via sham surgery in another group. The neurotoxic lesion guaranteed OFC dysfunction across fear and alcohol testing, and provided face validity for OFC hypoactivity observed in anxiety disorders and alcohol use disorder. We established fear discrimination to cues predicting unique foot shock probabilities: danger (1.00), uncertainty (0.25), and safety (0.00). The ability to upregulate or downregulate fear was determined by increasing (p = 0.25 to p = 0.50) then decreasing (p = 0.50 to p = 0.125) the shock probability associated with the uncertainty cue in four-session blocks. Changes in fear to uncertainty, as well as generalization to danger and safety, were monitored during probability shifts. All rats were then given voluntary, intermittent access to alcohol (20% ethanol) [21], followed by voluntary access to an isocaloric control solution. Assessing fear regulation and alcohol consumption in the same subjects allowed us to directly analyze their relationship, providing an explicit test our IOFC partitioning hypothesis.

Methods and materials

Subjects

Subjects were forty-three male Long Evans rats weighing 275–300 g upon arrival (Charles River Laboratories; RGD Cat# 2308852, RRID:RGD_2308852). Rats were individually housed and maintained on a 12-h dark light cycle (lights off at 6:00 PM) with water ad libitum. Procedures adhered to the NIH Guide for the Care and Use of Laboratory Animals and were approved by the Boston College Institutional Animal Care and Use Committee.

Surgery

Stereotaxic surgery was performed under isoflurane anesthesia (2–5%) using aseptic technique. Twenty-two rats received bilateral infusions of N-Methyl-D-aspartic acid (20 μg/μl in Dulbecco’s PBS) aimed at the dorsolateral subdivision of the orbitofrontal cortex (0.20 μl, +4.00 AP, ±3.60 ML, -5.10 DV; 0.25 μl, +4.00 AP, ±3.80 ML, -5.50 DV from skull) [22–24]. Infusions were delivered via 2 μl syringe (Hamilton, Neuros) controlled by a microsyringe pump (World Precision Instruments, UMP3-2). Infusion rate was ~0.11 μl/min. At the completion of each infusion, the syringe was raised 0.1 mm then left in place for five min to encourage delivery to the target site. The remaining twenty-one rats received identical surgical treatment without infusions. Rats received carprofen (5 mg/kg) for post-operative analgesia.

Behavioral apparatus

Eight sound-attenuated enclosures each housed a behavior chamber with aluminum front and back walls, clear acrylic sides and top, and a metal grid floor. Grid floors were electrically connected to a shock generator. A single external food cup and central nose poke opening equipped with infrared photocells were present on one wall. Auditory stimuli were presented...
through two speakers mounted on the ceiling of each behavior chamber. Consumption testing took place in the home cage. Solutions were in 50 mL tubes with rubber stoppers and ball bearing sipper tubes [25]. Rat chow and standard water bottles were always present on the home cage during consumption testing.

**Nose poke acquisition**

Following recovery from surgery, rats were food restricted to 85% of their initial free feeding weight, then fed (2–20 g/day) to increase their target body weight by 1 g/day for the remainder of testing. Rats were shaped to nose poke for pellet (BioServ F0021 –protein/fat/carbohydrate blend) delivery using a fixed ratio 1 schedule: one nose poke yielded one pellet. Shaping sessions lasted 30 min or approximately 50 nose pokes. Over the next 3, 60-min sessions, rats were placed on variable interval (VI) schedules in which nose pokes were reinforced on average every 30 s (session 1), or 60 s (sessions 2 and 3). For the remainder of testing, nose pokes were reinforced on a VI-60 schedule independent of all Pavlovian contingencies.

**Pre-exposure**

In two separate sessions, each rat was pre-exposed to the three cues to be used in Pavlovian fear discrimination. Cues were auditory stimuli, 10-s in duration and consisted of repeating motifs of a broadband click, phaser, or trumpet. Stimuli can be heard or downloaded at http://mcdannalldlab.org/resources/ardbark. Previous studies have found these stimuli to be equally salient, yet highly discriminable [26–28]. The 42-min pre-exposure sessions consisted of four presentations of each cue (12 total presentations) with a mean inter-trial interval (ITI) of 3.5 min. The order of trial type presentation was randomly determined by the behavioral program and differed for each rat during each session throughout behavioral testing.

For all sessions, fear to each auditory cue was measured using a suppression ratio based on nose poke rates during the 20-s baseline period immediately preceding the 10-s cue period [28–31]: suppression ratio = (baseline nose poke rate–cue nose poke rate) / (baseline nose poke rate + cue nose poke rate). A ratio of 1 indicated complete suppression of nose poking during the cue and a high level of fear; 0, no suppression and no fear. Intermediate suppression ratios reflected intermediate fear levels.

**Initial fear discrimination**

For the next sixteen sessions, each rat underwent Pavlovian fear discrimination. Each 54-min session began with a ~5-min warm-up period during which no cues or shock were presented. In discrimination, each cue was associated with a unique probability of foot shock (0.5 mA, 0.5-s): danger (1.00), uncertainty (0.25), and safety (0.00). Foot shock was administered 1-s following cue offset. A single session consisted of four danger, six uncertainty-no shock, two uncertainty-shock, and four safety trials. Mean inter-trial interval was 3.5 min.

**Increasing foot shock probability associated with uncertainty**

In order to assess rats’ ability to upregulate fear, the foot shock probability associated with uncertainty was increased from 0.25 to 0.50 starting on session 17, and continuing through session 20 (Fig 1A, middle). The total number of uncertainty trials was held constant, only now four of the eight uncertainty trials resulted in foot shock. For this and all subsequent blocks, the number and structure of the danger and safety trials were held constant, as were session length and ITI.
Decreasing foot shock probability associated with uncertainty

In order to assess rats’ ability to downregulate fear, the foot shock probability associated with uncertainty was decreased from 0.50 to 0.125 starting on session 21, and continuing through session 24 (Fig 1A, right). Again, the total number of uncertainty trials was held constant, only now a single uncertainty trial resulted in foot shock. Finally, rats were returned to the original uncertainty foot shock probability (0.25) for sessions 25–28. All aspects of these sessions were identical to initial discrimination. Rats then underwent a single extinction session identical to pre-exposure in which each cue was presented four times but no shocks were delivered. Immediately following extinction, rats were given *ad libitum* access to home cage food and water for the remainder of the experiment.

Voluntary alcohol consumption

Nine days following the extinction session, permitting sufficient time to overcome any food deprivation, rats received eight sessions of voluntary, intermittent, 24-h access to alcohol (20%
ethanol in di H\textsubscript{2}O v/v; [21]. Bottles were tested, weighed, then put on homes cages at 9:00 am on Sundays, Tuesdays, and Thursdays; and removed at 9:00 am on Mondays, Wednesdays, and Fridays. A leaky home cage water bottle was always present to ensure a constant water source, and that any alcohol consumption was voluntary. Consumption of an isocaloric, bitter, non-alcoholic solution was assessed in a ninth, 24-h session through voluntary access to quinine-adulterated DuoCal\textsuperscript{1} [QuAD; 0.1% quinine (w/v), 28% DuoCal\textsuperscript{1} (w/v), in di H\textsubscript{2}O].

**Histology**

Upon the conclusion of voluntary access, rats were anesthetized with an overdose of isoflurane and perfused intracardially with 0.9% biological saline. Brains were extracted and stored in 4% (v/v) formalin and 10% (w/v) sucrose. Forty-micrometer sections were collected on a sliding microtome and Nissl-stained to verify lesion placement.

**Statistical analysis**

Behavioral data were acquired using Med Associates Med-PC IV software (MED PC, RRID: SCR_012156). Raw data were processed in Matlab (MATLAB, RRID:SCR_001622) to extract nose-poke rates during two periods, baseline (20 s prior to cue onset) and cue (entire 10 s). These periods were used to calculate the suppression ratio. Suppression ratios were analyzed with repeated measures ANOVA in SPSS (RRID:SCR_002865) and Statistica (RRID: SCR_014213). Partial eta squared ($\eta^2$) and observed power are reported for ANOVA results for indicators of effect size.

For the four-session blocks in which shock probability was altered, we calculated a fear regulation score (FRS) for each cue/subject by taking the average suppression ratio for the last two sessions of each block and subtracting the first session. This was done in order to reduce the session-by-session change in suppression ratio to a single value, critical for later linear regression and also allowing for clearer visualization of the change for each cue/group.

Consumption is reported in grams (solution consumed) / kg (body weight) per 24 hours (total time given access): g/kg/24 h. Novelty-corrected alcohol consumption was determined by subtracting QuAD consumption level from the eight-session mean of alcohol consumption. Alcohol consumption was tested for normality using the Anderson-Darling test and analyzed using the nonparametric Mann-Whitney U test, which does not assume normality.

Linear regression was performed in which FRS values from probability shift blocks (increasing and decreasing) were used as regressors to predict eight-session mean alcohol consumption. Statistical output of regression was the significance of the model ($R^2$, $F$, $p$ and estimate of error variance) as well as beta coefficients ($\beta$) for each regressor, with larger absolute values indicating stronger predictive relationships and $\beta$ sign indicating direction of relationship. Regression models produced by sham and IOFC rats were statistically compared using the Fisher r-to-z transformation. For all analyses, $p < 0.05$ was considered significant.

**Results**

Neurotoxic damage (cell loss and gliosis) was quantified for the ventrolateral, lateral, and dorsolateral subregions of the OFC. Six IOFC rats had unilateral or non-neurotoxic damage and were excluded from subsequent analyses. Sixteen IOFC rats showed damage primarily in the lateral and dorsolateral subregions of the OFC, with little damage in the ventrolateral subregion and no damage in the medial OFC. Shams showed no evidence of neurotoxic damage. Representative IOFC (Fig 1B, right), and sham (Fig 1B, left) sections are shown. Each subject’s lesion was drawn, made transparent, and stacked (Fig 1C). Darker areas indicate regions of greater overlap and more consistent damage.
Sham and lOFC rats showed equivalent baseline nose poking rates throughout pre-exposure, discrimination, probability shift blocks, and extinction (Fig 2A). Mean ± SEM pellets earned per session: Sham, 52.25 ± 0.9 and lOFC, 51.98 ± 1.3; independent samples t test found no group difference (t\(_{35} = -0.57, p = 0.57\)). ANOVA for baseline nose poke rate with session (31) and group (sham vs. lOFC) as factors demonstrated a main effect of session \(F(30,1050) = 53.28, p = 1.69 \times 10^{-187}\), partial η\(^2\) = 0.60, observed power = 1.00. Critically, ANOVA found no main effect or interaction with group (Fs < 0.74, ps > 0.84). Equivalent performance ensures that differences in suppression ratios between groups cannot be attributed to differences in baseline nose poke rates.

**Initial Pavlovian discrimination**

Sham and lOFC rats showed equivalent and excellent fear discrimination over the initial 16 sessions (Fig 2B and 2C). Suppression ratios were low in pre-exposure and initially increased to all cues. As discrimination proceeded, the suppression ratio for each cue diverged: high to danger, intermediate to uncertainty, and low to safety. In support, ANOVA [within factors: session (16) and cue (3); between factor: group (sham vs. lOFC)] revealed a main effect of cue \(F(2,70) = 154.64, p = 2.07 \times 10^{-26}\), partial η\(^2\) = 0.82, observed power = 1.00], session \(F(15,525) = 52.48, p = 4.23 \times 10^{-94}\), partial η\(^2\) = 0.60, observed power = 1.00], and a cue x session interaction \(F(30,1050) = 13.75, p = 1.05 \times 10^{-56}\), partial η\(^2\) = 0.28, observed power = 1.00]. No main effect or interaction with group was observed.

**Increasing foot shock probability associated with uncertainty**

Starting session 17, and continuing until session 20, the foot shock probability associated with uncertainty was increased from 0.25 to 0.50. Discrimination was generally maintained in both groups over the four sessions. However, shams showed little change in fear to any cue (Fig 3A, left), whereas lOFC rats tended to increase fear to all cues (Fig 3A, right). In support, ANOVA for suppression ratio [between factor: group; within factors: session (4) and cue (3)] found a main effect of cue \(F(2,70) = 117.91, p = 3.86 \times 10^{-23}\), partial η\(^2\) = 0.77, observed power = 1.00] and trend toward a group x session interaction \(F(3,105) = 2.53, p = 0.062, \text{partial } \eta^2 = 0.67, \text{observed power} = 0.61\].

The change in fear to each cue was more apparent using the fear regulation score (FRS; Fig 3B). Shams showed FRS values around zero for uncertainty but a tendency toward negative
FRS values for danger and safety. By contrast, lOFC rats showed positive FRS values to all three cues. ANOVA for FRS [between factor: group; within factor: cue (3)] found a main effect of group \( F(1,35) = 5.53, p = 0.03, \text{partial } \eta^2 = 0.14, \text{observed power 0.63} \). Illustrative of the group main effect, when FRS was averaged for all three cues, a significant difference emerged with lOFC rats showing significantly higher FRS values than shams (Fig 3B, bottom right; two-tailed t-test, \( t_{35} = 2.34, p = 0.03 \)). The lOFC is not necessary to upregulate fear to uncertainty, per se, but may normally prevent generalized increases in fear to safety and danger.

Decreasing foot shock probability associated with uncertainty

Starting session 21, and continuing until session 24, the foot shock probability associated with uncertainty was decreased from 0.50 to 0.125. Discrimination was generally maintained. However, shams decreased fear to all cues (Fig 3C, left), while IOFC rats selectively reduced fear to uncertainty and safety (Fig 3C, right). ANOVA for suppression ratio [within factors: session (4) and cue (3); between factor: group (sham vs. IOFC)] found a main effect of cue \( F(2,70) = 81.08, p = 5.94 \times 10^{-19}, \text{partial } \eta^2 = 0.70, \text{observed power 1.00} \), session \( F(3,105) = 5.95, p = 0.001, \text{partial } \eta^2 = 0.15, \text{observed power 0.91} \), and a significant session x cue x group interaction \( F(6,210) = 2.50, p = 0.023, \text{partial } \eta^2 = 0.067, \text{observed power 0.83} \).
Visualizing the FRS for each group/cue made the pattern clear (Fig 3D). FRS was consistently negative for all three cues in shams. Negative FRS values were also observed to uncertainty and safety for IOFC rats but were positive for danger. ANOVA for FRS uncovered a trend toward a cue x group interaction \[F_{(2,70)} = 2.46, p = 0.093, \text{partial } \eta^2 = 0.066, \text{observed power 0.48}\]. Direct comparison of FRS to danger revealed significantly lower values in sham compared to IOFC rats (Fig 3D, top left; two-tailed t-test, \(t_{35} = 2.28, p = 0.03\)). The IOFC is not necessary to downregulate fear to uncertainty but may normally permit generalized decreases in fear to danger.

**Alcohol consumption**

Alcohol consumption was non-normally distributed in sham \([A^2 = 2.15, p = 5.00 \times 10^{-4}]\) and IOFC rats \([A^2 = 1.46, p = 5.29 \times 10^{-4}]\). While there was variability, consumption levels were higher in IOFC rats in each of the eight alcohol sessions (Fig 4A). Increased consumption by IOFC rats was restricted to alcohol, as low and similar levels of QuAD consumption were observed in both groups. Mann-Whitney U for 8-session mean alcohol consumption revealed a trend toward greater consumption by IOFC rats \([U = 112, p = 0.089]\) but found significance for novelty-corrected alcohol consumption \([U = 91, p = 0.018]\) (Fig 4B).

**Predictive relationship between fear regulation and alcohol consumption**

If the IOFC partitions fear regulation from alcohol consumption, then we should be able to use fear regulation to uncertainty—the only cue for which shock probability was altered—to predict alcohol consumption only in IOFC rats. The uncertainty FRSs from increasing shock probability (data from Fig 3B, top right) and decreasing shock probability (data from Fig 3D, top right) were used as regressors to predict novelty-corrected alcohol consumption (data from Fig 4B). Novelty-corrected alcohol consumption was z-score transformed (mean = 0, standard deviation = 1) in order to relate the change in FRS to standard deviation change in consumption.

We were unable to use uncertainty fear regulation to predict alcohol consumption in shams (regression model: \(R^2 = 0.02, F = 0.15, p = 0.87, \text{error} = 1.09\)). Indeed, there was near zero correlation between upshift FRS and novelty-corrected alcohol consumption (Fig 5A) and near zero correlation between downshift FRS and novelty-corrected alcohol consumption (Fig 5B). The resulting regression model had considerable error and did not fit the observed pattern of alcohol consumption (Fig 5C). Similar failures were observed when using danger FRSs.

---

![Fig 4. Voluntary alcohol consumption.](https://doi.org/10.1371/journal.pone.0198043.g004)
(regression model: \( R^2 = 0.05, F = 0.45, p = 0.64, \text{error} = 1.06 \)), safety FRSs (regression model: \( R^2 = 0.04, F = 0.38, p = 0.69, \text{error} = 1.06 \)), or a single model containing all six FRSs (regression model: \( R^2 = 0.16, F = 0.46, p = 0.83, \text{error} = 1.19 \)). Sham regression also failed if uncorrected alcohol consumption was predicted (regression model: \( R^2 = 0.03, F = 0.27, p = 0.77 \)).
error = 1.08). A power analysis revealed that 490 sham subjects would be necessary to observe a predictive relationship between uncertainty fear regulation and alcohol consumption (p < 0.05, power ≥ 0.80). Simple correlations revealed zero relationship between novelty-corrected alcohol consumption the upshift FRS (R = -0.04, p = 0.89; Fig 5A) and downshift FRS (R = -0.11, p = 0.65; Fig 5B).

By contrast, uncertainty fear regulation predicted alcohol consumption in IOFC rats (regression model: R² = 0.63, F = 10.84, p = 0.0017, error = 0.43, observed power 0.97). The beta coefficient for FRS during upshift (β = 3.78) and FRS during downshift (β = -3.75) were equivalently large, but in opposing directions. The resulting regression model had less error, and better fit the observed pattern of alcohol consumption (Fig 5F). The predictive relationship between FRS and alcohol consumption was restricted to uncertainty, as individual analyses for danger (regression model: R² = 0.10, F = 0.73, p = 0.50, error = 1.04) and safety (regression model: R² = 0.20, F = 3.30, p = 0.23, error = 0.92) did not reveal significant models. A virtually identical result was found for IOFC rats when using uncertainty FRSs to predict uncorrected, 8-session mean alcohol consumption (regression model: R² = 0.61, F = 10.12, p = 0.002, error = 0.45). The Fisher r-to-z transformation revealed a significant difference between r values in the IOFC and sham groups for novelty-corrected alcohol consumption (Z = 2.60, p = 0.0099) and uncorrected alcohol consumption (Z = 2.40, p = 0.016). The IOFC model produced a significantly stronger correlation coefficient and, therefore, greater fit than the sham model. When simple correlations were examined, the upshift FRS was positively, but not significantly, correlated with novelty-corrected consumption (R = 0.37, p = 0.16; Fig 5D), while the downshift FRS was significantly, negatively correlated with novelty-corrected consumption (R = -0.52, p = 0.04; Fig 5E).

**Discussion**

**LOFC contribution to fear regulation**

The IOFC was not required for mastery of initial fear discrimination. Somewhat surprisingly, shams showed little change in fear to uncertainty—or any cue—when shock probability was increased. Extensive training of the initial discrimination may have meant that more than four sessions would be required to increase fear to uncertainty. By contrast, IOFC rats increased fear to all three cues. Only in IOFC-damaged rats, when the uncertainty cue worsened, all cues worsened. Shams readily decreased fear to uncertainty when the foot shock probability was decreased, generalizing this decrease to safety and danger. LOFC rats decreased fear to uncertainty, generalized to safety, but failed to generalize to danger. Only in IOFC-intact rats, when the uncertainty cue improved, all cues improved. This overall pattern is consistent with findings from the reward literature, in which OFC lesions typically leave the acquisition of discrimination intact, but alter performance during reversal and the ability to track changes in cue-reward contingencies [32–39].

Of course, we cannot rule out the possibility that IOFC lesions altered cue encoding during initial fear discrimination, despite leaving behavior intact, and that probability shifts only revealed this deficit. For example, IOFC lesions could have increased sensitivity to uncertainty, making these rats more responsive to probability changes in either direction. Yet, fear to uncertainty—the only cue experimentally altered—never significantly differed between sham and IOFC rats at any point. Only differences to safety and danger were observed, and these only occurred during probability shifts. LOFC lesions could have made the cues more difficult to separate, but this should have resulted in uniform changes across all cues during upshift and downshift, instead of the asymmetrical generalization pattern we observed. The most parsimonious account of the overall pattern of fear is that IOFC lesions did not impair encoding or
separation of cues during initial discrimination, but altered fear generalization during probability shifts.

The role of the IOFC in fear regulation may be conceptualized by the reward construct of ‘credit assignment’ [39, 40]. Credit assignment reflects the need to attribute discrepancies between expected and received rewards (prediction errors) to specific causes (e.g. cues or choices) [41]. If one makes a series of choices (A, B, and C), and only choice C produces more reward than expected, credit needs to be selectively assigned to C. Rhesus macaques with OFC intact correctly assign credit (to choice C), whereas macaques with OFC damage randomly assign credit (to choices A, B and C) [39]. Viewed through the lens of credit assignment, increasing shock probability induces a positive error, and in normal rats, positive errors are selectively assigned to uncertainty. Decreasing shock probability induces a negative error that is globally assigned. LOFC damage inverts this assignment; positive errors are now globally assigned, while negative errors are more selectively assigned. The net result of the IOFC lesion-induced inversion is to more globally enhance or maintain fear when a variable predictor of threat worsens or improves.

A role for the IOFC to regulate prediction error assignment is accordant with previous findings. Rats receiving OFC inactivation with muscimol following discriminative cue conditioning show excessive fear to safety in a recall test [42]. Rats with OFC lesions fail to acquire discriminative fear to a context paired with shock versus a shock-free context, showing excessive fear to shock-free context [13, 20]. In terms of prediction error assignment, normal rats selectively assign positive prediction error (surprising shock receipt) to the shock context, while OFC-lesioned rats inappropriately assign positive error to the shock-free context, as well as the shock context. Further, it has been shown that OFC lesions [20] or OFC DREADD inactivation [15] result in sustained cued fear in the face of extinction. This is consistent with inappropriate assignment of negative prediction error when shock is surprisingly omitted in extinction. Use of inhibitory DREADDs [15] specifically during probability shift sessions would have permitted direct demonstration of the IOFC contribution to prediction error assignment. However, this temporally-focused approach would not have permitted disruption of IOFC function spanning fear regulation and alcohol consumption, a requirement for testing the partitioning hypothesis. Our hypothesized role for the OFC in prediction error assignment unites function in fear and reward settings and is consistent with findings from the fear conditioning literature.

LOFC contribution to alcohol consumption

LOFC lesions increased alcohol consumption over that of an isocaloric control solution. This finding is in general accordance with a recent study showing that IOFC DREADD inhibition or lesions increased alcohol consumption in alcohol-dependent mice [17]. Our finding that the IOFC regulates alcohol consumption is line with work demonstrating that alcohol drinking induces OFC opioid release in people, and alterations in OFC binding correlate with excessive alcohol consumption [19]. One caveat is that we did not measure drinking from the water bottle. The water bottle was designed to be leaky to ensure any and all alcohol consumption was entirely voluntary. Thus, the present data cannot speak to a preference for alcohol over water, nor can we entirely rule out IOFC lesions affecting overall fluid consumption. The pattern of consumption in the current study, initially high levels of consumption followed by a decrease across sessions, differs from other reported patterns of consumption where intake escalates over time [21, 43]. However, differences in observed drinking patterns are not uncommon and can be due to a variety of methodological differences such as age, bottle type, housing conditions, and the nature of previous alcohol experience [25, 44]. The pattern we observed is highly consistent with previous reports from our laboratory irrespective of [27, 45, 46] and following
fear discrimination [47]. Of course, we cannot rule out that prior fear conditioning influenced alcohol consumption. The current results support a role for the IOFC in moderating and reducing voluntary alcohol consumption. Behavioral procedures assessing alcohol learning [48], as well as motivational properties of alcohol [46] and alcohol-associates cues [49], are likely to uncover an even greater role for the IOFC.

**LOFC partitions mechanisms for fear regulation and alcohol consumption**

Fear regulation and alcohol consumption became inextricably linked when the IOFC was damaged. Extreme increases in fear to uncertainty following upshift, and extreme decreases in fear to uncertainty following downshift, predicted excessive alcohol consumption. This is in contrast to the clear partitioning exhibited by IOFC-intact shams. While admittedly idiosyncratic, the IOFC as partitioner is broadly consistent with a proposal for the OFC as a 'cognitive map' [50, 51]. The cognitive map view stems from findings that, within a structured reward task, the OFC organizes and separates task-relevant components. This includes observable events [52] and hidden states, which must be inferred from the task structure [50, 53]. This map is sufficiently detailed such that the OFC informs the current 'location' within the structure [50]. Partitioning is inherent to distinguishing task-relevant from task-irrelevant information and discerning location within a larger cognitive map.

Our proposal can be seen as the application of the cognitive map hypothesis to a more macro level. The partitioning hypothesis, and our data, suggest that a primary result of IOFC damage is to alter information processing pertaining to fear and alcohol in a larger neural network. LOFC projects to a variety of brain regions critical to processes of fear and alcohol, including prelimbic cortex [54–56], basolateral amygdala [45, 57, 58], central amygdala [58–60], and nucleus accumbens [61–63]. Just as the IOFC organizes and separates information within a single reward task, the IOFC may organize and separate information across distinct motivational mechanisms (fear versus alcohol) in single neurons, or neural ensembles, in a wider neural network. A primary result of reduced OFC function would be common processing of fear and alcohol information in a wider neural network.

**LOFC and comorbid anxiety disorders/alcohol use disorder**

Our results provide mechanisms by which OFC dysfunction (hypoactivity) could individually contribute to each disorder. For anxiety disorders and stress disorders [64], OFC hypoactivity would encourage generalization of aversive prediction errors in a way that increases fear, or maintains fear, when a variable threat predictor became more or less predictive. For alcohol use disorder, OFC hypoactivity would promote alcohol consumption and/or alcohol seeking. Most novel, OFC hypoactivity may be a key mechanism by which neurobehavioral circuits for fear and alcohol, which are normally partitioned, become linked. OFC dysfunction initially triggered by disordered anxiety could drive excessive alcohol consumption, and/or OFC dysfunction initially triggered by disordered drinking could drive extreme fear regulation. A testable prediction of this hypothesis is that greater OFC hypoactivity (during a fear task, alcohol task, or at baseline) would be found in individuals with comorbid anxiety disorder/alcohol use disorder, compared to those with only one disorder or the other. Even more, fear-related and alcohol-related cues/processes should produce more similar patterns of network activity in comorbid individuals than health controls or individuals with only one disorder.

**Conclusion**

Here we have revealed the IOFC is essential to partitioning of fear regulation and alcohol consumption. These results provide a plausible source of neural dysfunction that directly...
contributes to anxiety disorder and alcohol use disorder independently and most notably, their comorbid relationship. Future studies examining fear and alcohol information processing in amygdalar, striatal, and prefrontal neural ensembles [65] with IOFC intact or impaired, will provide valuable insight into normal IOFC partitioning. Such studies are likely to accelerate understanding of the neural dysfunction driving comorbidity of anxiety and alcohol use disorder, hastening the development of therapies to restore function.

Acknowledgments
We would like to thank Daniel Pimpinelli and Alyssa Russ for technical assistance and Rachel Zacharias for comments on the manuscript.

Author Contributions
Conceptualization: Madelyn H. Ray, Michael A. McDannald.
Data curation: Madelyn H. Ray, Emma Hanlon.
Formal analysis: Madelyn H. Ray, Michael A. McDannald.
Funding acquisition: Michael A. McDannald.
Investigation: Madelyn H. Ray.
Methodology: Madelyn H. Ray.
Resources: Michael A. McDannald.
Supervision: Michael A. McDannald.
Writing – original draft: Madelyn H. Ray, Michael A. McDannald.
Writing – review & editing: Madelyn H. Ray, Michael A. McDannald.

References
1. Kushner MG, Abrams K, Borchardt C. The relationship between anxiety disorders and alcohol use disorders: a review of major perspectives and findings. Clin Psychol Rev. 2000; 20(2):149–71. PMID: 10721495.
2. Smith JP, Book SW. Comorbidity of generalized anxiety disorder and alcohol use disorders among individuals seeking outpatient substance abuse treatment. Addictive Behaviors. 2010; 35(1):42–5. https://doi.org/10.1016/j.addbeh.2009.07.002 PubMed PMID: WOS:000271788900008. PMID: 19733441
3. Kessler RC, Crum RM, Warner LA, Nelson CB, Schulenberg J, Anthony JC. Lifetime co-occurrence of DSM-III-R alcohol abuse and dependence with other psychiatric disorders in the national comorbidity survey. Archives of General Psychiatry. 1997; 54(4):313–21. PubMed PMID: WOS:A1997WT58100003. PMID: 9107147
4. Petrakis IL, Gonzalez G, Rosenheck R, Krystal JH. Comorbidity of alcoholism and psychiatric disorders—An overview. Alcohol Research & Health. 2002; 26(2):81–9. PubMed PMID: WOS:000223806600001.
5. Milad MR, Rauch SL. The role of the orbitofrontal cortex in anxiety disorders. Linking Affect to Action: Critical Contributions of the Orbitofrontal Cortex. 2007; 1121:546–61. https://doi.org/10.1196/annals.1401.006 PubMed PMID: WOS:000252265300032. PMID: 17698998
6. Etkin A, Wager TD. Functional neuroimaging of anxiety: a meta-analysis of emotional processing in PTSD, social anxiety disorder, and specific phobia. Am J Psychiatry. 2007; 164(10):1476–88. https://doi.org/10.1176/appi.ajp.200707030504 PubMed PMID: 17698396; PubMed Central PMCID: PMC3318959.
7. Volkow ND, Fowler JS. Addiction, a disease of compulsion and drive: Involvement of the orbitofrontal cortex. Cerebral Cortex. 2000; 10(3):318–25. https://doi.org/10.1093/cercor/10.3.318 PubMed PMID: WOS:000083865600011. PMID: 10731226
8. Dom G, Sabbe B, Hulstijn W, Van Den Brink W. Substance use disorders and the orbitofrontal cortex—Systematic review of behavioural decision-making and neuroimaging studies. British Journal of
9. Claus ED, Ewing SWF, Filbey FM, Sabbineni A, Hutchinson KE. Identifying Neurobiological Phenotypes Associated with Alcohol Use Disorder Severity. Neuropsychopharmacology. 2011; 36(10):2086–96. https://doi.org/10.1038/npp.2011.99 PubMed PMID: WOS:000293893100013. PMID: 21677649

10. Koob GF, Volkow ND. Neurocircuitry of addiction. Neuropsychopharmacology. 2010; 35(1):217–38. https://doi.org/10.1038/npp.2009.110 PMID: 19710631; PubMed Central PMCID: PMC2805560.

11. Spoormaker VI, Andrade KC, Schroter MS, Sturm A, Goya-Maldonado R, Samann PG, et al. The neural correlates of negative prediction error signaling in human fear conditioning. Neuroimage. 2011; 54(3):2250–6. https://doi.org/10.1016/j.neuroimage.2010.09.042 PubMed PMID: WOS:000286302000047. PMID: 20869454

12. Tabbert K, Stark R, Kirsch P, Vaitl D. Hemodynamic responses of the amygdala, the orbitofrontal cortex and the visual cortex during a fear conditioning paradigm. International Journal of Psychophysiology. 2005; 57(1):15–23. https://doi.org/10.1016/j.ijpsycho.2005.01.007 PubMed PMID: WOS:000229862300003. PMID: 15935259

13. Trow JE, Hong NS, Jones AM, Lapointe J, Macphail JK, McDonald RJ. Evidence of a Role for Orbital Prefrontal Cortex in Preventing over-Generalization to Moderate Predictors of Biologically Significant Events. Neuroscience. 2017; 345:49–63. https://doi.org/10.1016/j.neuroscience.2016.10.017 PubMed PMID: WOS:000394726700006. PMID: 27743344

14. Mitchell JM, O’Neil JP, Janabi M, Marks SM, Jagust WJ, Fields HL. Alcohol Consumption Induces Endogenous Opioid Release in the Human Orbitofrontal Cortex and Nucleus Accumbens. Science Translational Medicine. 2012; 4(116). doi: ARTN 116ra6 10.1126/scitranslmed.3002902. PubMed PMID: WOS:000299010000110. PMID: 23314382

15. Simms JA, Steensland P, Medina B, Abernathy KE, Chandler LJ, Wise R, et al. Intermittent access to 20% ethanol induces high ethanol consumption in Long-Evans and Wistar rats. Alcohol Clin Exp Res. 2008; 32(10):1816–23. https://doi.org/10.1111/j.1530-0277.2008.00753.x PubMed PMID: WOS:0002138769000118. PMID: 18671810; PubMed Central PMCID: PMC3151464.

16. Gallagher M, McManan RW, Schoenbaum G. Orbitofrontal cortex and representation of incentive value in associative learning. J Neurosci. 1999; 19(15):6610–4. PubMed PMID: WOS:000081648400038. PMID: 10414988

17. Pickens CL, Saddoris MP, Setlow B, Gallagher M, Holland PC, Schoenbaum G. Different roles for orbitofrontal cortex and basolateral amygdala in a reiner devaluation task. Journal of Neuroscience. 2003; 23(35):11078–84. Epub 2003/12/06. doi: 10.1523/JNEUROSCI.5499-10.2011 PubMed PMID: WOS:000273929000039. PMID: 21325538

25. Doremus TL, Brunell SC, Rajendran P, Spear LP. Factors influencing elevated ethanol consumption in adolescent relative to adult rats. Alcohol Clin Exp Res. 2005; 29(10):1796–808. PMID: 16269909.
26. Berg BA, Schoenbaum G, McDannald MA. The dorsal raphe nucleus is integral to negative prediction errors in Pavlovian fear. European Journal of Neuroscience. 2014; 40(7):3096–101. https://doi.org/10.1111/ejn.12676 PMID: 25041165

27. DiLeo A, Wright KM, McDannald MA. Sub-second fear discrimination in rats: Adult impairment in adolescent heavy alcohol drinkers. Learning & Memory. 2016; 23:618–22.

28. Wright KM, DiLeo A, McDannald MA. Early adversity disrupts the adult use of aversive prediction errors to reduce fear in uncertainty. Front Behav Neurosci. 2015; 9:227. https://doi.org/10.3389/fnbeh.2015.00227 PMID: 26379520; PubMed Central PMCID: PMCPMC4550796.

29. Pickens CL, Golden SA, Adams-Deutsch T, Nair SG, Shaham Y. Long-lasting incubation of conditioned fear in rats. Biological Psychiatry. 2009; 65(10):881–6. Epub 2009/01/27. doi: S0006-3223(08)01592-8 [pii] https://doi.org/10.1016/j.biopsych.2008.12.010 PMID: 19167702.

30. Arico C, McNally GP. Opioid receptors regulate blocking and overexpectation of fear learning in conditioned suppression. Behav Neurosci. 2014; 128(2):199–206. https://doi.org/10.1037/a0036133 PMID: 24773439.

31. Anglada-Figueroa D, Quirk GJ. Lesions of the basal amygdala block expression of conditioned fear but not extinction. Journal of Neuroscience. 2005; 25(42):9680–5. Epub 2005/10/21. doi: 25/42/9680 [pii] https://doi.org/10.1523/JNEUROSCI.2600-05.2005 PMID: 16237172.

32. Schoenbaum G, Nugent SL, Saddoris MP, Setlow B. Orbitofrontal lesions in rats impair reversal but not acquisition of go, no-go odor discriminations. Neureport. 2002; 13(6):885–90. Epub 2002/05/09. PMID: 11997707.

33. Izquierdo A, Suda RK, Murray EA. Bilateral orbital prefrontal cortex lesions in rhesus monkeys disrupt choices guided by both reward value and reward contingency. Journal of Neuroscience. 2004; 24 (34):7540–8. https://doi.org/10.1523/JNEUROSCI.1921-04.2004 PubMed PMID: WOS:000223521200014. PMID: 15329401

34. Teitelbaum H. A Comparison of Effects of Orbitofrontal and Hippocampal Learning Upon Discrimination Learning and Reversal in the Cat. Exp Neurol. 1964; 9:452–62. PMID: 14188532.

35. Jones B, Mishkin M. Limbic lesions and the problem of stimulus—reinforcement associations. Exp Neurol. 1972; 36(2):362–77. PMID: 4626489.

36. Izquierdo A. Functional Heterogeneity within Rat Orbitofrontal Cortex in Reward Learning and Decision Making. Journal of Neuroscience. 2017; 37(44):10529–40. https://doi.org/10.1523/JNEUROSCI.1678-17.2017 PubMed PMID: WOS:000414190300001. PMID: 29093055

37. Fellows LK, Farah MJ. Ventromedial frontal cortex mediates affective shifting in humans: evidence from a reversal learning paradigm. Brain. 2003; 126(Pt 8):1830–7. https://doi.org/10.1093/brain/awg180 PMID: 12821528.

38. Clarke HF, Robbins TW, Roberts AC. Lesions of the medial striatum in monkeys produce perseverative impairments during reversal learning similar to those produced by lesions of the orbitofrontal cortex. J Neurosci. 2008; 28(43):10972–82. https://doi.org/10.1523/JNEUROSCI.1521-08.2008 PMID: 18945905; PubMed Central PMCID: PMC3391893.

39. Walton ME, Behrens TEJ, Buckley MJ, Rudebeck PH, Rushworth MFS. Separable Learning Systems in the Macaque Brain and the Role of Orbitofrontal Cortex in Contingent Learning. Neuron. 2010; 65 (6):927–39. https://doi.org/10.1016/j.neuron.2010.02.027 PubMed PMID: IS000276148900019. PMID: 20346766

40. Walton ME, Behrens TEJ, Noonan MP, Rushworth MFS. Giving credit where credit is due: orbitofrontal cortex and valuation in an uncertain world. Critical Contributions of the Orbitofrontal Cortex to Behavior. 2011; 1239:14–24. https://doi.org/10.1111/j.1749-6632.2011.06257.x PubMed PMID: WOS:000299228800002. PMID: 22145871

41. Stalnaker TA, Cooch NK, Schoenbaum G. What the orbitofrontal cortex does not do. Nature Neuroscience. 2015; 18(5):620–7. https://doi.org/10.1038/nn.3982 PubMed PMID: WOS:000353636900007. PMID: 25919962

42. Sarlito MC, Foiib AR, Christianson JP. Inactivation of the Ventrolateral Orbitofrontal Cortex Impairs Flexible Use of Safety Signals. Neuroscience. 2018; 379:350–8. https://doi.org/10.1016/j.neuroscience.2018.03.037 PMID: 29604383.

43. Mill DJ, Bito-Onon JJ, Simms JA, Li R, Bartlett SE. Fischer rats consume 20% ethanol in a long-term intermittent-access two-bottle-choice paradigm. PLoS One. 2013; 8(11):e79824. https://doi.org/10.1371/journal.pone.0079824 PMID: 24244567; PubMed Central PMCID: PMCPMC3826209.

44. Gilpin NW, Karanikas CA, Richardson HN. Adolescent binge drinking leads to changes in alcohol drinking, anxiety, and amygdalar corticotropin releasing factor cells in adulthood in male rats. PLoS One. 2012; 7(2):e31466. https://doi.org/10.1371/journal.pone.0031466 PMID: 22347484; PubMed Central PMCID: PMCPMC3275622.
45. Moaddab M, Mangone E, Ray MH, McDannald MA. Adolescent Alcohol Drinking Renders Adult Drinking BLA-Dependent: BLA Hyper-Activity as Contributor to Comorbid Alcohol Use Disorder and Anxiety Disorders. Brain Sci. 2017; 7(11). https://doi.org/10.3390/brainsci7110151 PMID: 29135933; PubMed Central PMCID: PMC55704158.

46. DiLeo A, Wright KM, Mangone E, McDannald MA. Alcohol gains access to appetitive learning through adolescent heavy drinking. Behav Neurosci. 2015; 129(4):371–9. https://doi.org/10.1037/bne0000075 PMID: 26052793.

47. Zacharias RA, Andreansky C, Ray MH, McDannald MA. Early adolescent adversity inflates threat estimation in females and promotes alcohol use initiation in both sexes Behavioral Neuroscience. In Press.

48. Remedios J, Woods C, Tardif C, Janak PH, Chaudhri N. Pavlovian-conditioned alcohol-seeking behavior in rats is invigorated by the interaction between discrete and contextual alcohol cues: implications for relapse. Brain and Behavior. 2014; 4(2):278–89. https://doi.org/10.1002/brb3.216 PubMed PMID: WOS:000346973200015. PMID: 24683519

49. Corbit LH, Janak PH. Ethanol-associated cues produce general pavlovian-instrumental transfer. Alcohol Clin Exp Res. 2007; 31(5):766–74. https://doi.org/10.1111/j.1530-0277.2007.00359.x PMID: 17378919.

50. Schuck NW, Cai MB, Wilson RC, Niv Y. Human Orbitofrontal Cortex Represents a Cognitive Map of State Space. Neuron. 2016; 91(6):1402–12. https://doi.org/10.1016/j.neuron.2016.08.019 PubMed PMID: WOS:000386760000022. PMID: 27657452.

51. Wikenheiser AM, Schoenbaum G. Over the river, through the woods: cognitive maps in the hippocampus and orbitofrontal cortex. Nature Reviews Neuroscience. 2016; 17(8):513–23. https://doi.org/10.1038/nrn.2016.56 PubMed PMID: WOS:000380656900009. PMID: 27256552.

52. Lopatina N, Sadacca BF, McDannald MA, Styer CV, Peterson JF, Cheer JF, et al. Ensembles in Medial and Lateral Orbitofrontal Cortex Construct Cognitive Maps Emphasizing Different Features of the Behavioral Landscape. Behavioral Neuroscience. 2017; 131(3):201–12. https://doi.org/10.1037/brn0000195 PubMed PMID: WOS:000402048900001. PMID: 28541078.

53. Corbit LH, Janak PH. Ethanol-associated cues produce general pavlovian-instrumental transfer. Alcohol Clin Exp Res. 2007; 31(5):766–74. https://doi.org/10.1111/j.1530-0277.2007.00359.x PMID: 17378919.

54. Schuck NW, Cai MB, Wilson RC, Niv Y. Human Orbitofrontal Cortex Represents a Cognitive Map of State Space. Neuron. 2016; 91(6):1402–12. https://doi.org/10.1016/j.neuron.2016.08.019 PubMed PMID: WOS:000386760000022. PMID: 27657452.

55. Wikenheiser AM, Schoenbaum G. Suppression of Ventral Hippocampal Output Impairs Integrated Orbitofrontal Encodinng of Task Structure. Neuron. 2017; 95(5):1197-+. https://doi.org/10.1016/j.neuron.2017.08.003 PubMed PMID: WOS:000408687900023. PMID: 28823726.

56. Willcocks AL, McNally GP. The role of medial prefrontal cortex in extinction and reinstatement of alcohol seeking in rats. European Journal of Neuroscience. 2013; 37(2):259–68. https://doi.org/10.1111/jen.12031 PubMed PMID: WOS:000313857000009. PMID: 23106416.

57. Fanselow MS, LeDoux JE. Why we think plasticity underlying Pavlovian fear conditioning occurs in the basolateral amygdala. Neuron. 1999; 23(2):229–32. Epub 1999/07/10. doi: S0896-6273(00)80775-8 [pii]. PMID: 10399930.

58. Shi CJ, Cassell MD. Cortical, thalamic, and amygdaloid connections of the anterior and posterior insular cortices. Journal of Comparative Neurology. 1998; 399(4):440–68. https://doi.org/10.1002/(Sici)1096-9861(19981006)399:4<440::Aid-Cne2>3.0.Co;2–1 PubMed PMID: WOS:000075648000002. PMID: 9741477.

59. Gilpin NW, Herman MA, Roberto M. The Central Amygdala as an Integrative Hub for Anxiety and Alcohol Use Disorders. Biological Psychiatry. 2015; 77(10):859–69. https://doi.org/10.1016/j.biopsych.2014.09.008 PubMed PMID: WOS:000353559600005. PMID: 25433901.

60. Kapp BS, Frysinger RC, Gallagher M, Haselton JR. Amygdala central nucleus lesions: effect on heart rate conditioning in the rabbit. Physiology & Behavior. 1979; 23(6):1109–17. Epub 1979/12/01. PMID: 542252.

61. Wright CI, Groenewegen HJ. Patterns of overlap and segregation between insular cortical, intermedio-dorsal thalamic and basal amygdaloid afferents in the nucleus accumbens of the rat. Neuroscience. 1996; 73(2):359–73. https://doi.org/10.1016/0306-4522(95)00592-7 PubMed PMID: WOS: A1996UQ40300004. PMID: 8783254.

62. Chaudhri N, Sahuque LL, Schairer WW, Janak PH. Separable Roles of the Nucleus Accumbens Core and Shell in Context- and Cue-Induced Alcohol-Seeking. Neuropsychopharmacology. 2010; 35(3):783–91. https://doi.org/10.1038/npp.2009.187 PubMed PMID: WOS:000273635100019. PMID: 19724113
63. Muschamp JW, Van’t Veer A, Parsegian A, Gallo MS, Chen M, Neve RL, et al. Activation of CREB in the Nucleus Accumbens Shell Produces Anhedonia and Resistance to Extinction of Fear in Rats. Journal of Neuroscience. 2011; 31(8):3095–103. https://doi.org/10.1523/JNEUROSCI.5973-10.2011 PubMed PMID: WOS:000287670100037. PMID: 21414930

64. American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorders. Washington, DC 2013.

65. Liberti WA, Markowitz JE, Perkins LN, Liberti DC, Leman DP, Guitchounts G, et al. Unstable neurons underlie a stable learned behavior. Nature Neuroscience. 2016; 19(12):1665–71. https://doi.org/10.1038/nn.4405 PubMed PMID: WOS:000389011900026. PMID: 27723744