Review

Harnessing Synthetic Lethal Interactions for Personalized Medicine

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Abstract: Two genes are said to have synthetic lethal (SL) interactions if the simultaneous mutations in a cell lead to lethality, but each individual mutation does not. Targeting SL partners of mutated cancer genes can kill cancer cells but leave normal cells intact. The applicability of translating this concept into clinics has been demonstrated by three drugs that have been approved by the FDA to target PARP for tumors bearing mutations in BRCA1/2. This article reviews applications of the SL concept to translational cancer medicine over the past five years. Topics are (1) exploiting the SL concept for drug combinations to circumvent tumor resistance, (2) using synthetic lethality to identify prognostic and predictive biomarkers, (3) applying SL interactions to stratify patients for targeted and immunotherapy, and (4) discussions on challenges and future directions.

Keywords: biomarker; cancer; genetic interaction; precision medicine; synthetic lethal

1. Introduction

Two genes are called synthetic lethal (SL, a type of genetic interaction) when a simultaneous mutation of both genes leads to cell death, but a single mutation of either does not; see Figure 1 for an illustration. Although synthetic lethality was first observed in fruit flies by Calvin Bridges, the concept of synthetic lethality can be applied to exploit cancer-cell specific mutations for therapeutics as indicated in seminal papers [2,3]. Targeting SL partners of mutated cancer genes will selectively kill cancer cells but spare normal cells. Therefore, the synthetic lethality strategy offers a way to treat cancer cells with non-druggable mutant tumor suppressor genes (TSGs) and stability genes, e.g., TP53 and BRCA1, by targeting their SL partners. The clinical relevance of synthetic lethality has been rapidly recognized. For example, pioneering studies of SL partners in BRCA1 and BRCA2-deficient cancer cells identified PARP1. PARP inhibitors (PARPi) have become the first clinically approved drugs exploiting the synthetic lethality concept. The US FDA approved PARPi for ovarian cancer in 2014, breast cancer in 2018 and prostate cancer in 2020. Notwithstanding, it has taken more than 15 years since the concept of synthetic lethality was first indicated for cancer therapies [2] to develop these PARP inhibitors, which are used to treat breast cancer and high-grade serous ovarian cancer patients with homologous recombination deficiency (HRD), which includes mutation in BRCA1/2, RAD51C/D, or PALB2, hyper-methylation of the BRCA1 promoter, or a series of yet to be defined causes [4].

It has been reported that PARP inhibitors are more effective for patients with BRCA1- and BRCA2-mutant ovarian cancer than for breast cancers [5], which shows that genetic context is crucial for functional genomic target screening. Advances in biotechnology such as CRISPR [6,7] have been expedited, which shall support the discovery of genetic contexts under which SL-based inhibitors work. Furthermore, there are many resources to mine novel SL interactions [8], e.g., Project DRIVE [9] and Project Achilles [10], which used a large set of human cell lines to uncover SL interactions, and multi-omics and clinical data of 33 cancer types at The Cancer Genome Atlas (TCGA). Additionally, a database named...
The Network Data Exchange (NDEx) [11] was built to organize the published SL interactions. As there are so many drug combinations, there are simply not sufficient patients to recruit for clinical trials, in addition to a huge amount of costs and time-consuming. Thus, adapting computational approaches, in particular machine learning-based approaches [11], will greatly benefit research on translating SL interactions to personalized medicine, as discussed later.

Here, we review applications of the SL concept to translational medicine in cancer over the past five years. In particular, we focus on the applications of synthetic lethality to personalized medicine. The scope of the article includes (1) exploiting the SL concept for drug combinations to circumvent tumor resistance, (2) using synthetic lethality to identify prognostic and predictive biomarkers, and (3) applying SL interactions to stratify patients for targeted and immunotherapy. We close with discussions on challenges and future directions.

Figure 1. A graphic illustration of synthetic lethality. (a,c) Gene A and gene B are synthetic lethal when simultaneous mutation of gene A and B leads to cell death, but a single mutation of either does not. (b) The SL concept can be exploited to inhibit the SL partner (Gene A) of a mutant Gene B in a tumor cell.

2. Exploiting SL Interactions for Drug Combinations

Taking tumor heterogeneity into account, combinations of drugs will be more effective than single agent approaches [12]. Further, resistance may also be developed by a single targeted agent in a tumor, thus identification of effective drug combinations which target resistance pathways will also be crucial [13]. Experimentally validated SL pairs could lead to clinically relevant drug combinations provided that the genetic context of a tumor is well understood. This has been demonstrated by the success of PARP inhibitors and other SL combined drugs currently undergoing phase III clinical trials, e.g., olaparib combined with AR-pathway targeting in patients with metastatic castration-resistant prostate cancer (CRPCa) [14].

Jariyal and colleagues [15] reviewed several SL interactions which have been under clinical trial studies, in addition to olaparib and other PARP inhibitors. For example, Wee1 and TP53 is a known SL pair [16]. Hirai and colleagues showed that Wee1 inhibition (MK-1775) combined with DNA damage agents, such as gemcitabine, carboplatin, and cisplatin, led to apoptosis in p53-deficient cells [17]. This result provided a scientific basis for the phase II trial on MK-1775 in combination with carboplatin for patients with p53-mutant ovarian cancer [NCT01164995]. Moreover, ATM and TP53 are also known SL. Durant and colleagues showed...
that ATM inhibitor (ATMi, AZD1390) combined with radiation therapy improved survival of preclinical brain tumor models [18], which led to a phase-1 clinical trial (NCT03423628). Thus, exploiting verified SL interactions to select patients with the associated mutations will optimize the patient outcomes and make clinical trials more efficient.

Currently, there are 342 clinical trial studies, registered at the US clinicaltrials.gov (accessed on 10 December 2021), for olaparib or olaparib-based drug combinations. In addition to breast cancer and ovarian cancer, there are also several clinical trials on PARP inhibitors for BRCA1/2- or ATM- mutant patients with prostate cancer [19] or cervical cancer [NCT04641728]. Recent studies of gene signatures for response to the PD-L1 inhibitor therapy (atezolizumab) in urothelial cancer (UC) [20–22], non-small cell lung cancer (NSCLC), and renal cell carcinoma (RCC) [23] revealed that the pathways most significantly associated with tumor mutation burden (TMB, a known factor correlated with response to immunotherapy checkpoint inhibitors), were cell cycle, DNA replication, and DNA damage response (DDR). DDR genes include BRCA1/2, ATM, RAD51, and others, and the former four genes are SL partners of PARP1. This provided a foundation for clinical trials on checkpoint inhibitors (CPIs) combined with PARPi in UC, NSCLC and RCC. Indeed, olaparib combined with pembrolizumab has been used for recurrent or metastatic cervical cancer patients in a phase II trial (NCT04641728). Several on-going clinical trials on combinations of olaparib and immunotherapies are listed in Table 1 (collected till 10 December 2021, from clinicaltrials.gov). Some studies include additional targeted therapies. We note that there is no phase III trial on combinations of olaparib and immunotherapy. However, there are phase III clinical trials on other PARP inhibitors, e.g., the JAVELIN Ovarian PARP 100 trial (NCT03642132), which failed due to no improvement of progression-free survival in patients. Additionally, another phase III clinical trial was withdrawn (NCT03806049).

| No. | Immunotherapy (and Targeted Therapies) | Investigated Cancer Types | Clinical Phase | Refs/Clinical Trial No. |
|-----|--------------------------------------|---------------------------|---------------|-------------------------|
| 1   | Olaparib, AZD6738 or Durvalumab       | TNBC \(^1\)               | Phase II      | NCT03740893             |
| 2   | Olaparib and Pembrolizumab           | Pancreatic Cancer         | Phase II      | NCT04548752             |
| 3   | Olaparib, Durvalumab and Tremelimumab| Solid Cancers             | Phase II      | NCT04169841             |
| 4   | Olaparib and Pembrolizumab           | Cervical Cancer           | Phase II      | NCT04483544             |
| 5   | Olaparib and Pembrolizumab           | TNBC                      | Phase II      | NCT04683679             |
| 6   | Olaparib and Pembrolizumab           | Breast Cancer             | Phase II      | NCT03025035             |
| 7   | Olaparib and Tremelimumab            | Peritoneal Cancer, Fallopian Cancer and Ovarian Cancer | Phase II | NCT04034927 |
| 8   | Olaparib, Nilotinib, Everolimus, Sorafenib, Lapatinib, Pazopanib, Durvalumab and Tremelimumab | Solid Neoplasms | Phase II | NCT02029001 |
| 9   | Olaparib and Pembrolizumab           | Pancreatic Cancer         | Phase II      | NCT05093231             |
| 10  | Olaparib and Atezolizumab            | Breast Cancer             | Phase II      | NCT02849496             |
| 11  | Olaparib and Durvalumab              | Prostate Cancer           | Phase II      | NCT04336943             |
| 12  | Olaparib and Durvalumab              | Bladder Cancer            | Phase II      | NCT04579133             |
| 13  | Olaparib and Ramucirumab             | Gastric Cancer Esophageal Cancer | Phase I/II | NCT03008278 |
| 14  | Olaparib and Tremelimumab            | Ovarian Cancer            | Phase I/II    | NCT02571725             |
| 15  | Olaparib and Pembrolizumab           | Melanoma                  | Phase II      | NCT04633902             |
| 16  | Olaparib and Bevacizumab, Cediranib and Cediranib Maleate | Glioblastoma             | Phase II      | NCT02974621             |

\(^1\) Note that TNBC denotes triple-negative breast cancer.
Resistance also develops after patients are treated with PARP inhibitors, and resistance is important to the initiation of therapy. For example, olaparib prevents DNA repair. Thus, these tumor cells gain a growth advantage due to accumulated mutations, leading to clinical resistance to PARP inhibitors eventually [25]. Furthermore, most patients with advanced cancer eventually develop resistance to targeted therapies [26–28]. This acquired resistance may be treated by a secondary drug uncovered by synthetic rescue (SR) interactions [29]. Note that both primary and secondary resistance can be mediated by SR mechanisms. SR interaction refers to a functional interaction where a fitness reduction of cancer cells due to the inactivation of one gene, called a vulnerable gene, is compensated for by the altered expression of another, called a rescuer gene. Sahu and colleagues developed a computational approach (INCISOR), which successfully associated gene pairs with SR interactions. The inhibition of predicted rescuer genes sensitized resistant tumor cells to therapies, which was validated in vitro. Thus far, there is no clinical validation on SR interactions. The concept of SR-interactions has the potential for future basic research.

After effective drug combinations are discovered, these drug combinations could be used to identify patient populations that would respond. For example, prostate cancer patients with BRCA1- mutations could be selected for clinical trials and for treatments with olaparib and ATM inhibitor, provided that the drug combination is approved by the FDA. Note that the combination of olaparib and AZD0156 (an ATM inhibitor) is currently undergoing a phase-I clinical trial (CT02588105) for patients with advanced cancer.

3. SL Interactions to Uncover Prognostic and Predictive Biomarkers

In general, two types of biomarkers are investigated: prognostic and predictive [30]. A panel of prognostic biomarkers identified for cancers could enable the selection of patients best suited for intensive adjuvant therapies in clinics. Thus, prognostic biomarkers could help advance personalized medicine.

Using published SL gene pairs, Shieh and colleagues developed a systematic approach to uncover IHC prognostic markers in colorectal cancer, lung adenocarcinoma and oral squamous cell carcinoma [31–33]. Specifically, they utilized a list of 643–742 SL pairs collected from the literature, gene expression data of the corresponding cancer under study, IHC expression, and clinical data on local cancer patients, to develop a computational approach to identify IHC prognostic biomarkers for the aforementioned three cancers. The authors first screened the collected SL pairs, most of which were validated by genome-wide RNAi screenings in various cancers [34,35], using microarray gene expression data of cancerous and non-cancerous tissues. They sorted the SL pairs by the fractions of the (up, up), (up, down), (down, up), and then (down, down) patterns, which were computed using patients’ gene expression of the associated cancers. About 20 genes with high fractions in the (up, up) pattern were selected for IHC staining. Next, they successfully identified single and combined IHC prognostic markers by correlating the single/paired IHC with overall survival of cancer patients via univariate Cox regression analysis [36]. The predicted prognostic markers were further verified by at least one external data set, e.g., TCGA lung adenocarcinoma for [32]. The approach revealed IHC marker pairs when neither single IHC was a marker. Furthermore, several of the identified prognostic markers with components involved in different pathways, e.g., the pair CK1e(C)-Rb1(N) was revealed to be a prognostic biomarker [33], but phosphorylation of CK1e is involved in the p53 pathway [37], which is different from the Rb pathway. As most of methods to uncover IHC markers to date have been mainly based on one or two proteins [38] or one pathway [39], their approach improved the current state-of-the-art for IHC markers. The flowchart of their approach is presented in Figure 2.

Srivas and colleagues [40] applied the conserved tumor suppressor genes (TSGs) from yeast to humans to identify TSGs interacting with the target of an FDA-approved drug. In particular, they identified ATM-irinotecan (inhibitor of TOP1 [41]). To date, both FOLFIRI (5-flourouracil plus irinotecan) and FOLFOX, which is a chemotherapy regimen made up of folinic acid, fluorouracil and oxaliplatin, have been indicated to treat metastatic colorectal
cancer (mCRC) patients with an approximately 40% response rate. Nevertheless, there is no diagnostic test to guide selections from the aforementioned regimen for better response. Applying the SL combination \(\text{ATM}\) mutation and irinotecan, they found six out of 16 mCRC patients with \(\text{ATM}\) mutations, who were treated with irinotecan, had improved survival (44 months versus 29 months). Therefore, \(\text{ATM}\) could be a predictive biomarker to stratify mCRC patients for FOLFIRI [15].

Figure 2. A graphical display of the approach in [31–33] to discover prognostic biomarkers. Gene expression of cancerous versus non-cancerous tissues was used to select SL gene pairs relevant to a cancer under study, from the collected SL pairs. This procedure resulted in ~20 genes for immunohistochemistry (IHC). Then combinations of IHC and overall survival of patients were analyzed by Cox regression to yield prognostic markers, which were further validated by at least one external data set such as TCGA.

4. Synthetic Lethality Applied to Stratify Patients for Targeted and Immunotherapy

4.1. Selection of Patients for Clinical Trials

Medical doctors determine whether new treatments are safe and more effective than current treatments through clinical trials. SL interactions can be applied to stratify patients for clinical trials of targets therapies as follows. In clinical trials of targeted drugs, if only a small proportion of patients have the required genetic context for response in a trial. Strong signals in individual patients will be diluted by patients without the necessary genetic context, and the trial may fail. In order to conduct clinical trials effectively, patients with proper genetic contexts should be selected. For example, de Bono and colleagues reported
that only ~10% of patients enrolled showed mutations in the homologous recombination repair (HRR) biomarkers [42]. This lack of specificity poses a significant problem in clinical trials [19]. On the other hand, the first major biomarker study in prostate cancer (PCA) (the PROfound study) reported that 17.6% of the 4425 patients had mutations in at least one of the predefined 15 HRR genes, which included BRCA1, BRCA2, and ATM [42]. De Bono and colleagues revealed that PCA patients with BRCA1, BRCA2, or ATM mutations responded better to therapy and had increased progression-free survival and overall survival, whereas patients with long-tail HRR alterations such as FANCL or RAD51C did not have significant clinical benefits [43]. Analysis of DNA or RNA profiling of cancer patients and exploiting verified SL interactions will help prioritize candidates for clinical trials on SL-based drugs.

One example of how to select patients for a clinical trial using known SL pairs, e.g., TP53-WEE1, is the case of small cell lung cancer. As 100% of small cell lung cancer has the TP53 mutation, it is expected that most small cell lung cancers have lost the G1 checkpoint and have a high probability of depending on Wee1 for proper DNA repair and cell cycle progression. Thus, we could select patients with p53-mutant small cell lung cancer for a clinical trial of Wee1 inhibitor, which is also an ongoing phase II clinical trial (NCT026688907).

4.2. Synthetic Lethality Applied to Stratify Patients for Immunotherapy

Almost 90% of human cancer deaths are due to metastases. Immunotherapy is one of the most effective treatments for patients with certain metastatic cancer types to date. For instance, the PD-L1 inhibitor atezolizumab can treat certain patients with metastatic urothelial tumors [20,22]. Nevertheless, for some cancer types, e.g., HGSOC, CPIs did not work well [44]. Note that the objective response rate (ORR) for several cancers remain very low, for example ORR for urothelial cancer is only about 10% and less than 30% for non-small cell lung cancer. Thus, the identification of biomarkers to select patients for immunotherapy is important and very useful in clinical practice.

In addition to uncover biomarkers for checkpoint inhibitor immunotherapy, synthetic lethality can be exploited to direct immune cells specifically to tumor cells and destroy them as follows. The accumulation of genetic alterations in cancer cells, results in neoantigens. In theory, the immune system should generate T cell responses to recognize and kill nascent cancer cells. Nevertheless, tumor cells can escape immune pressure by evolving intrinsic genetic changes, for which [45] provided evidence. Zaretsk and colleagues found that loss of function mutations in JAK1 could enhance immune evasion and confer anti-PD1 resistance in patients treated with a checkpoint inhibitor [45].

Recent studies also suggest that many oncogenes and TSGs may be involved in immune evasion. For example, LKB1 was reported to be a putative tumor-intrinsic immune evasion gene [46]. Skoulidis and colleagues showed that Lkb1/Stk11 loss promoted PD-1/PD-L1 inhibitor resistance, using Kras- mutant murine lung adenocarcinoma models. Furthermore, they found that patients with LKB1 loss, correlating with reduced PD-L1 expression, did not respond to treatment with PD-1 inhibitors. This indicated that LKB1 was a genuine suppressor of immune evasion. Other genetic alterations that correlated with immune evasion include MYCN amplification [47], CASP8 loss of function [48], and PTEN loss of function [49]. CASP8 loss of function was found to rescue cancer cells from T cell-killing by blocking the TNF pathway, while PTEN loss of function promoted resistance to T cell-mediated immunotherapy. Therefore, a functional evaluation of all known cancer genes may lead to the identification of drug targets to reverse immune evasion phenotype, and these targets can also serve as biomarkers to select patients for clinical trial of immunotherapy. For instance, the inhibition of over-expressed SL partner(s) of LKB1, MYCN, CASP8, and PTEN may reverse the immune evasion. Note that the identification of biomarkers through immune evasion targets has a great advantage, as biomarkers for immunotherapy have been difficult to find.

Discovery of immune evasion targets requires two procedures: (1) identification of a genetic context that gives rise to immune evasion, and (2) identification of drug targets that
can reverse such immune evasion. As previously reviewed [5], SL-based CRISPR screening can be applied to tackle the first step. After an immune evasion genetic context has been uncovered, target screening can be performed in vitro with PD-L1 expression or other relevant immune readout, which may be the most efficient way to identify drug targets, as reported [5]. For example, identification of the immune evasion context for JAK1 loss of function can be performed using cell lines harboring a JAK1 loss of function mutation to measure PD-L1 expression after interferon stimulation. After the genetic context of immune evasion is revealed, SL-based target discovery approaches can be applied to discover the targets, which can reverse the immune evasion phenotype when knocked out. Advances in CRISPR technology [5] will enable the integration of cancer genetics (including the concept of synthetic lethality) and immune-oncology to elucidate the mechanism of immune evasion and make immunotherapy more clinically effective in the future.

5. Discussion and Future Directions

As mentioned in Section 1, there are too many drug combinations to recruit patients for clinical trials. Therefore, adapting computational approaches, followed by functional genomics screening to validate and confirm the SL interactions will be efficient. To achieve this aim, some useful computational algorithms and databases are available. Mining Synthetic Lethals (MiSL) [50] was developed to predict SL partners using multi-omics data, such as DNA mutation, copy number alteration, and gene expression, from 12 TCGA cancers. Sinha and colleagues exploited conserved TSGs from yeast to humans and used pancancer data to identify SL combinations [40]. The Network Data Exchange platform (NDEx) organizes published SL interactions [11] and is machine readable and searchable, so machine learning and database algorithms can be applied. As it encompasses a large volume of data, namely SL interactions in humans and other species and multi-omics data of ~33 cancer types, it will be efficient for predicting new SL interactions and the genetic context in which an SL-based drug will affect a particular cancer, through a machine learning approach. In addition to the big volume of omics data of various types of cancer, many validated genetic (including SL) interactions, protein interactions and prior knowledge are available, all of which can be used to train a machine learning algorithm. For instance, a deep learning algorithm was shown to predict drug combination effectively [51]. One can foresee that, once DNA sequencing and/or gene expression data of a patient’s tumor is profiled, and the data are fitted into a pre-trained machine learning algorithm, it will be possible to output the genetic context of the tumor and suggest an SL-based drug or a drug combination, in the near future. Then, a medical doctor can prescribe the suggested treatment to the patient accordingly. For example, olaparib combined with an immune CPI therapy can be prescribed to patients with BRCA2-mutant breast cancer.

Although Project DRIVE [9] and Project Achilles [10] have discovered SL interactions using a large set of human cell lines, the uncovered and fully verified drug targets remain limited. Project Score [52] has provided compelling evidence that there are still many drug targets for discovery, and they can be uncovered using a functional genomics approach. Synthetic lethality, ML-based computational algorithms, and the recent advances in biological science/technology, e.g., the powerful CRISPR-based functional screening, will enable the discovery of new SL interactions and new drug targets in cancer.

As SL-interactions work in a context-dependent fashion, the genetic context under which a targeted therapy or immunotherapy is added to the primary treatment (chemotherapy or targeted therapy) is critical to develop combined therapy. A future direction is to incorporate a computational approach to mine SL-based drug combinations, followed by validation using CRISPR, single cell techniques, and patient-derived organoids, which will enable the discovery of tumor heterogeneity underlying the primary drug resistance and secondary resistance driven by tumor cell evolution. This will eventually lead to combined drugs.
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**Data Availability Statement:** Table 1 was queried from clinicaltrials.gov (accessed on 10 December 2021).

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**References**

1. Mani, R.; St Onge, R.P.; Hartman, J.L., IV; Giaever, G.; Roth, F.P. Defining genetic interaction. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 3461–3466. [CrossRef] [PubMed]
2. Hartwell, L.H.; Szankasi, P.; Roberts, C.J.; Murray, A.W.; Friend, S.H. Integrating genetic approaches into the discovery of anticancer drugs. *Science* **1997**, *278*, 1064–1068. [CrossRef]
3. Kaelin, W.G., Jr. The concept of synthetic lethality in the context of anticancer therapy. *Nat. Rev. Cancer* **2005**, *5*, 689–698. [CrossRef] [PubMed]
4. Miller, R.E.; Leary, A.; Scott, C.L.; Serra, V.; Lord, C.J.; Bowtell, D.; Chang, D.K.; Garsed, D.W.; Jonkers, J.; Ledermann, J.A.; et al. ESMO recommendations on predictive biomarker testing for homologous recombination deficiency and PARP inhibitor benefit in ovarian cancer. *Ann. Oncol.* **2020**, *31*, 1606–1622. [CrossRef] [PubMed]
5. Huang, A.; Garraway, L.A.; Ashworth, A.; Weber, B. Synthetic lethality as an engine for cancer drug target discovery. *Nat. Rev. Drug Discov.* **2020**, *19*, 23–38. [CrossRef] [PubMed]
6. Morgens, D.W.; Deans, R.M.; Li, A.; Bassik, M.C. Systematic comparison of CRISPR/Cas9 and RNAi screens for essential genes. *Nat. Biotechnol.* **2016**, *34*, 634–636. [CrossRef]
7. Evers, B.; Jastrzebski, K.; Heijmans, J.P.; Gernrum, W.; Beijersbergen, R.L.; Bernards, R. CRISPR knockout screening outperforms shRNA and CRISPRi in identifying essential genes. *Nat. Biotechnol.* **2016**, *34*, 631–633. [CrossRef]
8. O’Neil, N.J.; Bailey, M.L.; Hieter, P. Synthetic lethality and cancer. *Nat. Rev. Genet.* **2017**, *18*, 613–623. [CrossRef]
9. McDonald, E.R., 3rd; de Weck, A.; Schlabach, M.R.; Billy, E.; Mavrikis, K.J.; Hoffman, G.R.; Belur, D.; Castelletti, D.; Frias, E.; Gampa, K.; et al. Project DRIVE: A Compendium of Cancer Dependencies and Synthetic Lethal Relationships Uncovered by Large-Scale, Deep RNAi Screening. *Cell* **2017**, *170*, 577–592.e510. [CrossRef]
10. Barretina, J.; Caponigro, G.; Stransky, N.; Venkatesan, K.; Margolin, A.A.; Kim, S.; Wilson, C.J.; Lehár, J.; Kryukov, G.V.; Sonkin, D.; et al. The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. *Nature* **2012**, *483*, 603–607. [CrossRef]
11. Pratt, D.; Chen, J.; Welker, D.; Rivas, R.; Pilkis, R.; Rynkov, V.; Ono, K.; Miello, C.; Hicks, L.; Szalma, S.; et al. NDEX, the Network Data Exchange. *Cell Syst.* **2015**, *1*, 302–305. [CrossRef] [PubMed]
12. Williams, S.P.; McDermott, U. The Pursuit of Therapeutic Biomarkers with High-Throughput Cancer Cell Drug Screens. *Cell Chem. Biol.* **2017**, *24*, 1066–1074. [CrossRef]
13. Sun, C.; Fang, Y.; Yin, J.; Chen, J.; Ju, Z.; Zhang, D.; Chen, X.; Vellano, C.P.; Jeong, K.J.; Ng, P.K.; et al. Rational combination therapy with PARP and MEK inhibitors capitalizes on therapeutic liabilities in RAS mutant cancers. *Sci. Transl. Med.* **2017**, *9*.[CrossRef] [PubMed]
14. de Bono, J.S.; Mehra, N.; Scaglotti, G.V.; Castro, E.; Dorff, T.; Stirling, A.; Stenzl, A.; Fleming, M.T.; Higano, C.S.; Saad, F.; et al. Talazoparib monotherapy in metastatic castration-resistant prostate cancer with DNA repair alterations (TALAPRO-1): An open-label, phase 2 trial. *Lancet Oncol.* **2022**, *23*, 1250–1264. [CrossRef]
15. Jariyal, H.; Weinberg, F.; Acheva, J.; Nagarath, D.; Srivastava, A. Synthetic lethality: A step forward for personalized medicine in cancer. *Drug Discov. Today* **2020**, *25*, 305–320. [CrossRef] [PubMed]
16. Wang, Q.; Fan, S.; Eastman, A.; Worland, P.F.; Sausville, E.A.; O’Connor, P.M. UCN-01: A potent abrogator of G2 checkpoint function in cancer cells with disrupted p53. *J. Natl. Cancer Inst.* **1996**, *88*, 956–965. [CrossRef]
17. Hirai, H.; Iwasawa, Y.; Okada, M.; Arai, T.; Nishihata, T.; Kobayashi, M.; Kimura, T.; Kaneko, N.; Ohtani, J.; Yamanaka, K.; et al. Small-molecule inhibition of Wee1 kinase by MK-1775 selectively sensitizes p53-deficient tumor cells to DNA-damaging agents. *Mol. Cancer Ther.* **2009**, *8*, 2992–3000. [CrossRef]
18. Durant, S.T.; Zheng, L.; Wang, Y.; Chen, K.; Zhang, L.; Zhang, T.; Yang, Z.; Riches, L.; Trinidad, A.G.; Fok, J.H.L.; et al. The brain-penetrant clinical ATM inhibitor AZD1390 radiosensitizes and improves survival of preclinical brain tumor models. *Sci. Adv.* **2018**, *4*, eaat1719. [CrossRef]
19. Werdt, A.v.; Brandt, L.; Schäfer, O.D.; Rubin, M.A. PARP Inhibition in Prostate Cancer With Homologous Recombination Repair Alterations. *JCO Precis. Oncol.* **2021**, *5*, 1639–1649. [CrossRef]
20. Mariathasan, S.; Turley, S.J.; Nickles, D.; Castiglioni, A.; Yuen, K.; Wang, Y.; Kadel, E.E.; III; Koeppen, H.; Astarita, J.L.; Cubas, R.; et al. TGFβ3 attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. Nature 2018, 554, 544–548. [CrossRef]

21. Teo, M.Y.; Seier, K.; Ostrovnaya, I.; Regazzi, A.M.; Kania, B.E.; Moran, M.M.; Cipolla, C.K.; Bluth, M.J.; Chaim, J.; Al-Ahmadie, H.; et al. Alterations in DNA Damage Response and Repair Genes as Potential Marker of Clinical Benefit From PD-1/PD-L1 Blockade in Advanced Urothelial Cancers. J. Clin. Oncol. 2018, 36, 1685–1694. [CrossRef] [PubMed]

22. Nassar, A.H.; Moun, K.W.; Jegede, O.; Shinagare, A.B.; Kim, J.; Liu, C.J.; Pomertz, M.; Harshman, L.C.; Van Allen, E.M.; Wei, X.K.; et al. A model combining clinical and genomic factors to predict response to PD-1/PD-L1 blockade in advanced urothelial carcinoma. Br. J. Cancer 2020, 122, 555–563. [CrossRef]

23. Banchereau, R.; Leng, N.; Zill, O.; Sokol, E.; Liu, G.; Pavlick, D.; Maund, S.; Liu, L.-F.; Kadel, E.; Baldwin, N.; et al. Molecular determinants of response to PD-L1 blockade across tumor types. Nat. Commun. 2021, 12, 3969. [CrossRef]

24. Fong, C.Y.; Gilan, O.; Lam, E.Y.; Rubin, A.F.; Ftouni, S.; Tyler, D.; Stanley, K.; Sinha, D.; Yeh, P.; Morison, J.; et al. BET inhibitor resistance emerges from leukaemia stem cells. Nature 2015, 525, 538–542. [CrossRef] [PubMed]

25. Miyamoto, D.T.; Zheng, Y.; Wittner, B.S.; Lee, R.J.; Zhu, H.; Brannigan, B.W.; Trautwein, J.; et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. Science 2015, 349, 1351–1356. [CrossRef]

26. Sahu, A.D.; Lee, J.S.; Wang, Z.; Zhang, G.; Iglesias-Bartolome, R.; Tian, T.; Wei, Z.; Miao, B.; Nair, N.U.; Ponomarova, O.; et al. Genome-wide prediction of synthetic rescue mediators of resistance to targeted and immunotherapy. Mol. Syst. Biol. 2019, 15, e8323. [CrossRef]

27. Motter, A.E.; Gulbahce, N.; Almaas, E.; Barabási, A.L. Predicting synthetic rescues in metabolic networks. Mol. Syst. Biol. 2008, 4, 168. [CrossRef]

28. Cho, S.H.; Jeon, J.; Kim, S.I. Personalized medicine in breast cancer: A systematic review. J. Breast Cancer 2012, 15, 265–272. [CrossRef]

29. Barbie, D.A.; Tamayo, P.; Boehm, J.S.; Dunn, I.F.; Schinzel, A.C.; Sandy, P.; Meylan, E.; Scholl, C.; et al. Genome-wide prediction of synthetic rescue mediators of resistance to targeted and immunotherapy. Mol. Syst. Biol. 2019, 15, 441–450. [CrossRef] [PubMed]

30. Fumet, J.D.; Limagne, E.; Thibaudin, M.; Truntzer, C.; Bertaut, A.; Rederstorff, E.; Ghiringhelli, F. Precision medicine phase II study evaluating the efficacy of a double immunotherapy by durvalumab and tremelimumab combined with olaparib in patients with solid cancers and carriers of homologous recombination repair genes mutation in response or stable after olaparib treatment. BMC Cancer 2020, 20, 748. [CrossRef]

31. Teo, M.Y.; Seier, K.; Ostrovnaya, I.; Regazzi, A.M.; Kania, B.E.; Moran, M.M.; Cipolla, C.K.; Bluth, M.J.; Chaim, J.; Al-Ahmadie, H.; et al. Alterations in DNA Damage Response and Repair Genes as Potential Marker of Clinical Benefit From PD-1/PD-L1 Blockade in Advanced Urothelial Cancers. J. Clin. Oncol. 2018, 36, 1685–1694. [CrossRef] [PubMed]

32. Banchereau, R.; Leng, N.; Zill, O.; Sokol, E.; Liu, G.; Pavlick, D.; Maund, S.; Liu, L.-F.; Kadel, E.; Baldwin, N.; et al. Molecular determinants of response to PD-L1 blockade across tumor types. Nat. Commun. 2021, 12, 3969. [CrossRef]

33. Fong, C.Y.; Gilan, O.; Lam, E.Y.; Rubin, A.F.; Ftouni, S.; Tyler, D.; Stanley, K.; Sinha, D.; Yeh, P.; Morison, J.; et al. BET inhibitor resistance emerges from leukaemia stem cells. Nature 2015, 525, 538–542. [CrossRef] [PubMed]

34. Miyamoto, D.T.; Zheng, Y.; Wittner, B.S.; Lee, R.J.; Zhu, H.; Brannigan, B.W.; Trautwein, J.; et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. Science 2015, 349, 1351–1356. [CrossRef]

35. Sahu, A.D.; Lee, J.S.; Wang, Z.; Zhang, G.; Iglesias-Bartolome, R.; Tian, T.; Wei, Z.; Miao, B.; Nair, N.U.; Ponomarova, O.; et al. Genome-wide prediction of synthetic rescue mediators of resistance to targeted and immunotherapy. Mol. Syst. Biol. 2019, 15, e8323. [CrossRef]

36. Motter, A.E.; Gulbahce, N.; Almaas, E.; Barabási, A.L. Predicting synthetic rescues in metabolic networks. Mol. Syst. Biol. 2008, 4, 168. [CrossRef]

37. Cho, S.H.; Jeon, J.; Kim, S.I. Personalized medicine in breast cancer: A systematic review. J. Breast Cancer 2012, 15, 265–272. [CrossRef]

38. Fumet, J.D.; Limagne, E.; Thibaudin, M.; Truntzer, C.; Bertaut, A.; Rederstorff, E.; Ghiringhelli, F. Precision medicine phase II study evaluating the efficacy of a double immunotherapy by durvalumab and tremelimumab combined with olaparib in patients with solid cancers and carriers of homologous recombination repair genes mutation in response or stable after olaparib treatment. BMC Cancer 2020, 20, 748. [CrossRef]

39. Barbie, D.A.; Tamayo, P.; Boehm, J.S.; Dunn, I.F.; Schinzel, A.C.; Sandy, P.; Meylan, E.; Scholl, C.; et al. Genome-wide prediction of synthetic rescue mediators of resistance to targeted and immunotherapy. Mol. Syst. Biol. 2019, 15, e8323. [CrossRef] [PubMed]

40. Fong, C.Y.; Gilan, O.; Lam, E.Y.; Rubin, A.F.; Ftouni, S.; Tyler, D.; Stanley, K.; Sinha, D.; Yeh, P.; Morison, J.; et al. BET inhibitor resistance emerges from leukaemia stem cells. Nature 2015, 525, 538–542. [CrossRef] [PubMed]

41. Miyamoto, D.T.; Zheng, Y.; Wittner, B.S.; Lee, R.J.; Zhu, H.; Brannigan, B.W.; Trautwein, J.; et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. Science 2015, 349, 1351–1356. [CrossRef] [PubMed]

42. Sahu, A.D.; Lee, J.S.; Wang, Z.; Zhang, G.; Iglesias-Bartolome, R.; Tian, T.; Wei, Z.; Miao, B.; Nair, N.U.; Ponomarova, O.; et al. Genome-wide prediction of synthetic rescue mediators of resistance to targeted and immunotherapy. Mol. Syst. Biol. 2019, 15, e8323. [CrossRef]

43. Motter, A.E.; Gulbahce, N.; Almaas, E.; Barabási, A.L. Predicting synthetic rescues in metabolic networks. Mol. Syst. Biol. 2008, 4, 168. [CrossRef] [PubMed]
44. Borella, F.; Ghisoni, E.; Giannone, G.; Cosma, S.; Benedetto, C.; Valabrega, G.; Katsaros, D. Immune Checkpoint Inhibitors in Epithelial Ovarian Cancer: An Overview on Efficacy and Future Perspectives. *Diagnostics* 2020, 10, 146. [CrossRef] [PubMed]
45. Zaretsky, J.M.; Garcia-Diaz, A.; Shin, D.S.; Escuin-Ordinas, H.; Hugo, W.; Hu-Lieskovan, S.; Torrejon, D.Y.; Abril-Rodriguez, G.; Sandoval, S.; Barthly, L.; et al. Mutations Associated with Acquired Resistance to PD-1 Blockade in Melanoma. *N. Engl. J. Med.* 2016, 375, 819–829. [CrossRef]
46. Skoulidis, F.; Goldberg, M.E.; Greenawalt, D.M.; Hellmann, M.D.; Awad, M.M.; Gainor, J.F.; Schrock, A.B.; Hartmaier, R.J.; Trabucco, S.E.; Gay, L.; et al. STK11/LKB1 Mutations and PD-1 Inhibitor Resistance in KRAS-Mutant Lung Adenocarcinoma. *Cancer Discov.* 2018, 8, 822–835. [CrossRef]
47. Layer, J.P.; Kronmüller, M.T.; Quast, T.; van den Boorn-Konijnenberg, D.; Effern, M.; Hinze, D.; Althoff, K.; Schramm, A.; Westermann, F.; Peifer, M.; et al. Amplification of N-Myc is associated with a T-cell-poor microenvironment in metastatic neuroblastoma restraining interferon pathway activity and chemokine expression. *Oncoimmunology* 2017, 6, e1320626. [CrossRef]
48. Kearney, C.J.; Vervoort, S.J.; Hogg, S.J.; Ramsbottom, K.M.; Freeman, A.J.; Lalaoui, N.; Pijpers, L.; Michie, J.; Brown, K.K.; Knight, D.A.; et al. Tumor immune evasion arises through loss of TNF sensitivity. *Sci. Immunol.* 2018, 3, eaar3451. [CrossRef]
49. Peng, W.; Chen, J.Q.; Liu, C.; Malu, S.; Creasy, C.; Tetzlaff, M.T.; Xu, C.; McKenzie, J.A.; Zhang, C.; Liang, X.; et al. Loss of PTEN Promotes Resistance to T Cell-Mediated Immunotherapy. *Cancer Discov.* 2016, 6, 202–216. [CrossRef]
50. Sinha, S.; Thomas, D.; Chan, S.; Gao, Y.; Brunen, D.; Torabi, D.; Reinisch, A.; Hernandez, D.; Chan, A.; Rankin, E.B.; et al. Systematic discovery of mutation-specific synthetic lethals by mining pan-cancer human primary tumor data. *Nat. Commun.* 2017, 8, 15580. [CrossRef]
51. Flobak, Å.; Baudot, A.; Remy, E.; Thommesen, L.; Thieffry, D.; Kuiper, M.; Lægreid, A. Discovery of Drug Synergies in Gastric Cancer Cells Predicted by Logical Modeling. *PLoS Comput. Biol.* 2015, 11, e1004426. [CrossRef] [PubMed]
52. Behan, F.M.; Iorio, F.; Picco, G.; Gonçalves, E.; Beaver, C.M.; Migliardi, G.; Santos, R.; Rao, Y.; Sassi, F.; Pinnelli, M.; et al. Prioritization of cancer therapeutic targets using CRISPR-Cas9 screens. *Nature* 2019, 568, 511–516. [CrossRef] [PubMed]