INTRODUCTION

Antimicrobial resistance (AMR) is a growing social problem for public, animal and environmental health worldwide (World Health Organization, 2014). The use of antimicrobial drugs in livestock farms and in the rearing of domestic animals is suspected to play an important role in spreading AMR (Nhung et al., 2015). Previous studies have shown that AMR in wildlife tends to be more prevalent closer to human settlements where wild animals could have contact with livestock or companion animals in which antibacterial drugs are used as clinical therapies or growth stimulants (Kozak, Boerlin, Janecko, Reid-Smith, & Jardine, 2009; Yagasaki, 1986). Direct and/or indirect contact among farm or pet animals and wildlife could be involved in the transmission of AMR, facilitating the dissemination of AMR into natural environments. Many attempts are being made to understand how AMR is spread. Antimicrobial-resistant strains of Escherichia coli have been used as an indicator of the routes of AMR spread (Luo et al., 2011; Van den Bogaard & Stobberingh, 2000).
The Okinawa rail (Gallirallus okinawae) is a flightless bird in the family Rallidae and the species inhabit the evergreen laurel forest zone, called “Yambaru,” located in the northern part of Okinawa Main Island, Japan (Figure 1). Wild populations of the Okinawa rail are declining because of habitat loss and predation by invasive alien species such as the small Indian mongoose (Herpestes auropunctatus), feral cats (Felis catus) and feral dogs (Canis lupus) (Arcilla, Choi, Ozaki, & Lepczyk, 2015; Harato & Ozaki, 1993; Yamada & Sugimura, 2004). The Okinawa rail is currently listed as an endangered species in The International Union for Conservation of Nature Red List (IUCN, 2017) and highly protected under Act on Conservation of Endangered Species of Wild Fauna and Flora and Programs for the Rehabilitation of Natural Habitats and Maintenance of Viable Populations of Japan. Recently, it has been observed that the Okinawa rail population is slightly increasing around some livestock farms, where they possibly have easy access to food sources, including earthworms and insects (Ogura, Iijima, Ozaki, Nagamine, & Kuwana, 2009). Therefore, it is assumed that the Okinawa rail is exposed to AMR through food and through water contaminated with the faeces of livestock animals.

The aim of this study was to understand whether the Okinawa rail, an endangered species, plays an important role in carrying and spreading AMR, acting as a vehicle between human habitation and the natural environment in Okinawa Main Island.

2 | MATERIALS AND METHODS

Fifty-nine avian faecal samples which were suspected of wild Okinawa rails were collected between August 2012 and May 2014. The survey was carried out in Kunigami village, Okinawa Main Island, Japan located at 26°45′N and 128°17′E (Figure 1). Freshly excreted faeces on the asphalt road were collected using a sterilized swab or spoon in the early morning. After sampling, the surface of the road at the collection site was washed for the next sampling. Thirty faecal samples (18 samples collected in 2012, 10 samples collected in 2013 and two samples collected in 2014) were collected from around livestock farm areas (abbreviated LA), including human settlements, where Okinawa rails were frequently observed and 29 faecal samples were collected from a forest area (abbreviated FA) in 2014, located about 3–7 km from the LA sites (Figure 1). It is said that the territory size of Okinawa rails is about 1 km, but it is unpublished (private message from observer). Samples were stored at 4°C until laboratory examination.

To identify whether faecal samples were derived from Okinawa rails, we first screened for its characteristic odour in the sampling area. Secondly, molecular methods were employed to confirm that the faecal samples were from the desired species. The white-breasted waterhen (Amaurornis phoenicurus), in the family Rallidae, inhabits the same area as the Okinawa rail and faeces from this species were used for discrimination. White-breasted waterhen faecal samples were collected from captive individuals at Neo Park Okinawa, Okinawa, Japan, on November 4, 2013. For polymerase chain reaction (PCR) amplification, a primer set targeting a conserved region of the Okinawa rail mitochondrial ATP6 gene was designed using available sequence data (GenBank accession no. AP010821). The primers used were as follows: yanbaru kuina F (5′-ATGGGCCCTAACACTCTCCT-3′; nucleotides 12,945 to 12,964) and yanbaru kuina R (5′-GGAGACTGCGGGTATGATGG-3′; nucleotides 13,311 to 13,292). PCR products were purified using the QIAquick PCR Purification Kit® (QIAGEN, USA) and direct-sequenced using an ABI 3130xl sequencing system (Applied Biosystems, USA). The described primer set was also used for sequencing.

Okinawa rail faecal samples, identified using PCR, were cultured on MacConkey agar (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan) and screened for E. coli using Triple Sugar Iron and Motility-Indole-Lysine media (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan), followed by identification using the ID-Test/EB20 (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan). The drug susceptibility of the isolates was determined according to the Clinical and Laboratory Standards Institute methods (Clinical and Laboratory Standards Institute, ). The Kirby–Bauer disk diffusion method was used to determine the antimicrobial agent sensitivity profiles of E. coli isolates for the following 17 antimicrobial agents: Ampicillin (ABPC),...
Piperacillin (PIPC), Cefozopran (CZOP), Kanamycin (KM), Gentamicin (GM), Oxytetracycline (OTC), Ofloxacin (OFLX), Chloramphenicol (CP), Nalidixic acid (NA), Fosfomycin (FOM), Sulfamethoxazole-trimethoprim (ST), Streptomycin (SM), Ceftazidime (CAZ), Ceftriaxone (CTX), Cefotaxime (CTX), Cephalothin ( CET) and Cefuroxime (CXM) (Clinical and Laboratory Standards Institute, ). These antimicrobial drugs were chosen based on their clinical and agricultural uses for domestic animals and as feed additives. The diameters (in millimetres) of the clear zones of growth inhibition around the antimicrobial agent disks were measured using precision calipers. To categorize isolates as resistant or non-resistant to each antimicrobial agent, we used the standard method (Clinical and Laboratory Standards Institute, ). The E. coli strain ATCC 25,922 (American Type Culture Collection) was used for quality control.

The minimum inhibitory concentration (MIC) was determined using the ETEST® (SYSMEX bioMérieux Co., Ltd.), according to the manufacturer’s recommendations. The inoculum was matched to a McFarland standard of 0.5, and, using a sterilization swab, a lawn of bacteria was created on Mueller-Hinton agar plates. Two ETEST® strips were placed onto each Mueller-Hinton agar plate. The plates were incubated for 18 hr at 35°C. The MIC was interpreted as the value at which the inhibition zone intersected the scale on the ETEST® strip.

Chi-squared testing was used to compare the prevalence of antimicrobial-resistant bacteria from faecal samples obtained from the LA and FA samples. Antimicrobial resistance was expressed as odds ratios with 95% confidence intervals.

### 3 RESULTS AND DISCUSSION

For species identification, the Okinawa rail mitochondrial ATP6 gene was successfully amplified using PCR in 81% (48/59) of faecal samples. Of these 48 samples, 22 samples were collected from LA and 26 samples collected from FA.

*E. coli* were isolated from 31 of 48 (65%) samples, and 14 of 31 (45%) samples were antimicrobial resistant. Among the 22 faecal samples collected around LA, *E. coli* was isolated from 16 (73%) and 11 (69%) of 16 samples showed antimicrobial resistance. *E. coli* was isolated from 15 (58%) of the 26 faecal samples collected around FA and three (20%) of the 15 *E. coli*-positive samples showed antimicrobial resistance (Table 1). The prevalence of AMR *E. coli* in faecal samples from wild Okinawa rails from LA (69%) was significantly higher than those from FA (20%) (p = .006). The MICs of antimicrobials (in μg/ml) are shown in Table 2. Furthermore, five different multiple drug resistance (MDR) patterns were found in LA samples and three different MDR patterns were found in FA samples (Table 3).

Previous research investigating AMR in Japanese wild birds revealed that antimicrobial-resistant strains of *E. coli* were found in 12.5% of wild green pheasants (*Phasianus versicolor*) and in 15.8% of wild bamboo partridges (*Bambuscosa thoracicus*) (Kanai, Hashimoto, & Mitsushashi, 1981; Nakamura, Yoshimura, & Koeda, 1982). By comparison, the prevalence of AMR in Okinawa rails from LA was much higher, and it is likely that livestock farms could be a source of AMR exposure. Indeed, it is indicated that resistant strains of bacteria which was produced in the livestock body by inadequate use of antibotics for the treatment of clinical disease, as feed additives for the prevention of disease, or for nutritional purposes (Blanco, Lemus, & Grande, 2009) and/or which was produced by remained antibiotics in their excrements and feed have mainly emerged from livestock farming to the environment. Our results suggest that Okinawa rails are exposed to AMR *E. coli* at livestock farms. Therefore, it is suspected that the movement of Okinawa rails between LA and FA transmitted AMR.

The most frequently detected AMR strain of *E. coli* from wild Okinawa rails was OTC-resistant. OTC is widely used in the rearing of domestic animals to prevent and treat infectious diseases and to promote their growth (Asai, 2010; Horie & Takegami, 2006). OTC-resistant strains have been detected in hooded cranes (*Grus monacha*) that inhabit cattle farms in the Kagoshima Prefecture, Japan. The greatest OTC MIC value in hooded cranes was 128 μg/ml (Kitadai, Obi, Yamashita, Murase, & Takase, 2012); in Okinawa rails, we detected a higher resistance to OTC (MIC ≥ 256 μg/ml). Oonaka, Furuhata, Kiuchi, Hara, and Fukuyama (2004) compared AMR bacterium MIC values in samples from the environment and of human clinical origin, and found higher MIC values in human clinical isolates. Although the use of ABPC in human therapy is quite rare (Terada, Miyake, & Urase, 2012), 16 t/year of bulk powder has been sold in Japan for use with domestic and pet animals (National Veterinary Assay Laboratory Ministry of Agriculture, Forestry, & Fisheries of Japan, 2012). OTC and ABPC resistance in *E. coli* from wild Okinawa rails inhabiting the area around LA indicates that the livestock industry is likely the source origin of AMR for these antimicrobials. Furthermore, CP resistance was detected in rails collected in FA at high rates (67%). CP has been frequently used as a therapeutic

| Table 1 | Proportion of Okinawa Rail fecal samples demonstrating resistance to each antimicrobial. No resistance to CZOP, GM, FOM, SM, CAZ, CTRX, CTX and CXM was detected in Escherichia coli isolates from Okinawa rail faecal samples in both areas |
|-------------------|-------------------------------|-------------------------------|
| Antimicrobial     | LA No. of detected/total sample (%) | FA No. of detected/total sample (%) |
| OTC              | 9/11 (81.8)                   | 3/3 (100)                     |
| CET              | 5/11 (45.5)                   | 1/3 (33.3)                    |
| ABPC             | 4/11 (36.4)                   | 0/3 (0)                       |
| ST               | 4/11 (36.4)                   | 1/3 (33.3)                    |
| PIPC             | 3/11 (27.3)                   | 0/3 (0)                       |
| NA               | 2/11 (18.2)                   | 1/3 (33.3)                    |
| OFLX             | 2/11 (18.2)                   | 1/3 (33.3)                    |
| CP               | 2/11 (18.2)                   | 2/3 (66.7)                    |
| KM               | 1/11 (9.1)                    | 0/3 (0)                       |
agent for domestic animals in livestock farms in Japan because of its broad-spectrum activity against pathogenic bacteria (Harada, 2009). However, this drug is toxic to humans, causing aplastic anaemia, and the Japanese government has permanently banned the use of CP for the treatment of disease in food-producing animals (Gilmore, 1986; Harada, 2009). The presence of CP-resistant E. coli isolated from the Okinawa rail suggests that CP-resistant bacteria may have remained at the sampling areas more than 10 years after the use of the drug had ceased, or that CP may still be used for livestock animals at this location. Moreover, synthetic antimicrobial agents, including NA, OFLX and ST are substances that are created chemically, and are not naturally occurring antibiotics (Ministry of agriculture, forestry, & fisheries of Japan, 2014), and they have remained relatively stable in the environment (Sarmah, Meyer, & Boxall, 2006). There were 2–8 multiple drug resistance patterns, including synthetic antimicrobial agents, in resistant E. coli isolated from FA and LA.

It is suggested that AMR occurs as a result of interactions between microbial agents, host organisms and the environment (Delport, Harcourt, Beaumont, Webster, & Power, 2015). The indiscriminate use of certain antimicrobials in human and veterinary medicine has become a significant public health concern because it may select for resistant bacterial strains (Hiltunen, Virta, & Laine, 2017). This study indicates that wildlife, even critically endangered species such as the Okinawa rail, may play an important role as host reservoirs and potential vectors for the spread of AMR into the natural environment. However, we assembled samples from different areas

| Antimicrobial | Sampling area | MIC (µg/ml) | 1 | 8 | 16 | 32 | 64 | 96 | 128 | ≧256 |
|---------------|--------------|-------------|---|---|----|----|----|----|----|------|
| OTC           | FA           | 1           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 2    |
|               | LA           | 2           | 7 | 1 | 1  | 1  | 1  | 1  | 1   | 2    |
| CET           | FA           | 1           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 2           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| ABPC          | FA           |             |   |   |    |    |    |    |     |      |
|               | LA           | 4           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| ST            | FA           | 1           | 4 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 1           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| PIPC          | FA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 2           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| NA            | FA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| OFLX          | FA           | 1           | 1 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| CP            | FA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 2           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
| KM            | FA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |
|               | LA           | 1           | 2 | 1 | 1  | 1  | 1  | 1  | 1   | 1    |

**TABLE 2** MICs of AMR E. coli from Okinawa Rail faecal samples in LA and FA

| Sampling area | Resistance pattern | Resistance antimicrobial | No. of detected samples |
|---------------|--------------------|--------------------------|-------------------------|
| LA            | Single-drug resistance | OTC                      | 4                       |
|               |                     | CET                      | 2                       |
|               | Multi-drug resistance | OTC-ST                   | 1                       |
|               |                     | ABPC-PIPC-KM-OTC-CET     | 1                       |
|               |                     | ABPC-PIPC-OTC-ST-CET     | 1                       |
|               |                     | ABPC-OTC-NA-OFLX-CP-ST   | 1                       |
|               |                     | ABPC-PIPC-OTC-NA-OFLX-CP-ST-CET | 1 |
| FA            | Multi-drug resistance | OTC-CP                   | 1                       |
|               |                     | OTC-CET                  | 1                       |
|               |                     | OTC-NA-OFLX-CP-ST        | 1                       |

**TABLE 3** MDR patterns of AMR E. coli isolated from faecal samples of Okinawa rail collected in LA and FA
in different year and our sample size was small. Therefore, more samples need to be collected and genetic analysis of AMR E. coli using the PFGE (Pulsed-Field Gel Electrophoresis) method needs to be performed. Conclusively, it is imperative that attention has to be given to the prevalence of AMR among endangered species in programmes of in situ/ex situ conservation that include reintroduction, in order to maintain public and ecological health.

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CONFLICT OF INTEREST

The authors have no conflict of interests to declare.

ORCID

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