Long-term immunogenicity and safety of a non-typeable *Haemophilus influenzae*-Moraxella catarrhalis vaccine: 4-year follow-up of a phase 1 multicentre trial

Philippe De Smedt, Geert Leroux-Roels, Corinne Vandermeulen, Annaelisa Tasciotti, Gennaro Di Maro, Marie Dozot, Daniela Casula, Margherita Annaratone, Daniele Riccucci, Ashwani Kumar Arora

*Centre for the Evaluation of Vaccination, Vaccine and Infectious Disease Institute, University of Antwerp, Antwerp, Belgium*

*Centre for Vaccinology, Gent University and Gent University Hospital, Gent, Belgium*

*Leuven University Vaccinology Centre, Department of Public Health and Primary Care, KU Leuven, Leuven, Belgium*

*GSK, Siena, Italy*

*GSK, Rixensart, Belgium*

**A R T I C L E  I N F O**

**Article history:**
Received 31 May 2021
Received in revised form 9 September 2021
Accepted 29 October 2021
Available online 3 November 2021

**Keywords:**
Acute exacerbation
Antibody persistence
Clinical trial
COPD
*Haemophilus influenzae*
*Moraxella catarrhalis*

**A B S T R A C T**

A multicomponent vaccine has been developed to reduce the frequency of acute exacerbations of COPD associated with non-typeable *Haemophilus influenzae* (NTHi) and *Moraxella catarrhalis* (Mcat) infections, containing NTHi (PD and PE-PilA) and Mcat (UspA2) surface proteins. In a randomised, observer-blind, placebo-controlled study with two steps (NCT02547974), the investigational vaccine had good immunogenicity and no safety concerns were identified. In step 2, 90 adults aged 50–71 years with smoking history received two doses 60 days apart of one of two AS01E-adjuvanted formulations containing 10 μg of each antigen (10–10-AS01) or 10 μg NTHi antigens and 3.3 μg UspA2 (10–3-AS01), or placebo. Long-term persistence of antigen-specific humoral antibodies was assessed in 81 participants during 3 years of follow-up after the initial 14-month study (NCT03201211).

Antigen-specific antibody concentrations were measured in blood samples taken every 6 months. Safety monitoring evaluated serious adverse events (SAEs) and potential immune-mediated disease (pIMD).

Immune responses against NTHi antigens persisted up to 4 years post-vaccination. For PD, PE and PilA, at each follow-up time point, adjusted antibody geometric mean concentrations (GMCS) were higher (non-overlapping 95% confidence intervals [CIs]) in the vaccine groups versus placebo and versus pre-vaccination. Antibody GMC point estimates were higher with 10–3–AS01 than with 10–10–AS01. For UspA2, 95% CIs included 1 for GMC ratios of 10–10–AS01 or 10–3–AS01 to placebo at each time point. During follow-up, SAEs were reported in nine (11.1%) participants, one of which was fatal (lung cancer, 607 days after second 10–10–AS01 dose). One non-serious pIMD, trigeminal neuralgia, was reported 771 days after second 10–3–AS01 dose. The SAEs and pIMD were considered not related to vaccination.

Immune responses against NTHi antigens persisted for 4 years after two-dose vaccination with the investigational NTHi-Mcat vaccine. There was no persistent response against the Mcat antigen. No safety concerns were identified during the long-term follow-up.

© 2021 GlaxoSmithKline Biologicals S.A. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

**Abbreviations:** AECOPD, acute exacerbations of chronic obstructive pulmonary disease; ANCOVA, analysis of covariance; AS01E, Adjuvant System AS01E, containing 3-O-desacyl-4′-monophosphoryl lipid A, QS-21 (*Quillaja saponaria* Molina, fraction 21) and liposome; CI, confidence interval; COPD, chronic obstructive pulmonary disease; ELISA, enzyme-linked immunosorbent assay; EU, enzyme-linked immunosorbent assay units; GMC, geometric mean concentration; GMR, geometric mean ratio; LLOQ, lower limit of quantification; Mcat, *Moraxella catarrhalis*; MPL, 3-O-desacyl-4′-monophosphoryl lipid A; NTHi, non-typeable *Haemophilus influenzae*; PD, protein D; PE, protein E; PilA, Pilin A; pIMD, potential immune-mediated disease; QS-21, *Quillaja saponaria* Molina, fraction 21; SAE, serious adverse event; UspA2, ubiquitous surface protein A2.

*Corresponding author at: Clinical Research & Development, GSK, via Fiorentina 1, 53100 Siena, Italy.

E-mail address: ashwani.k.arora@gsk.com (A.K. Arora).
Introduction

Chronic obstructive pulmonary disease (COPD) is the third leading cause of death globally [1], with an estimated prevalence of 12% in people aged 30 years or more [2]. Acute exacerbations of COPD (AECOPD) are periods of worsened respiratory symptoms, beyond that seen with day-to-day variation, that increase the risk of myocardial infarction, stroke, pulmonary embolism and death [3]. No vaccine is approved for the prevention of AECOPD, although influenza and pneumococcal vaccines, which are routinely recommended to COPD patients [4], may have some effect on the frequency of exacerbations [5].

Bacterial infection is frequently associated with AECOPD, most commonly non-typeable Haemophilus influenzae (NTHi), Moraxella catarrhalis (Mcat) and Streptococcus pneumoniae infections [6–9]. Targeting the major bacterial species associated with AECOPD may be a viable strategy for vaccine development. There is evidence that NTHi and Mcat can act as co-pathogens in respiratory tract infections and COPD, as indicated by protection of NTHi from complement-mediated killing via complement resistance factors on outer membrane vesicles produced by Mcat [10]. Increased resistance to antibiotics and host clearance also appears to be promoted by NTHi and Mcat co-infection [11,12].

An adjuvanted multicomponent vaccine has been developed to reduce the frequency of moderate and severe AECOPD associated with NTHi and Mcat. The investigational NTHi-Mcat vaccine contains four surface proteins involved in the virulence mechanisms of both bacterial pathogens [13]. Three are from NTHi, a free recombinant protein D (PD) and a recombinant fusion protein combining protein E and Pilin A (PE-PilA), and the fourth from Mcat, ubiquitous surface protein A2 (UspA2). Evidence from animal studies suggest anti-PD antibodies have opsonic activity [14] and protect against H. influenzae infection [15], while antibodies generated by the PE-PilA fusion protein inhibit the binding of PE to vitronectin (which may protect the bacterium from complement attack [16,17]) and inhibit the formation of NTHi biofilms [18], as previously described for anti-PilA antibodies [19]. Anti-UspA2 antibodies significantly reduced the lung bacterial load in mice challenged with homologous or heterologous Mcat strains [20] and have been shown to be bactericidal and cross-reactive [20,21]. A vaccine formulation containing the NTHi proteins had an acceptable safety and reactogenicity profile and induced antigen-specific immune responses in phase 1 studies of healthy adults 18–40 years of age. The population group of adults with a smoking history was chosen to immunologically match the COPD population, with evidence suggesting that alterations in the immune system start early in smokers, before COPD is diagnosed [22–25]. NTHi vaccine formulations that included the Adjuvant System AS01e [26] produced the highest humoral and cellular immune responses in older adults [22]. A phase 2 study of the adjuvanted NTHi vaccine in adults with COPD showed no safety concerns and good immunogenicity [27].

In the first clinical assessment of the safety, reactogenicity and immunogenicity of the investigational NTHi-Mcat vaccine, two doses were given 60 days apart and the study was conducted in two steps [13]. In step 1, healthy adults aged 18–40 years received a non-adjuvanted vaccine formulation or placebo. In step 2, older adults with a smoking history received one of two AS01e-adjuvanted formulations (one containing 10 μg PD, 10 μg PE-PilA and 10 μg UspA2 and the other 10 μg PD, 10 μg PE-PilA and 3.3 μg UspA2) or placebo. No safety concerns were identified with the NTHi-Mcat vaccine and the formulation containing 3.3 μg UspA2 induced the best humoral response against NTHi antigens [13]; this response was consistent with that induced by the adjuvanted NTHi vaccine in the phase 2 study of patients with COPD [27]. The anti-UspA2 immune response was moderate and transient with both formulations. In the present study, we assessed the long-term persistence of antigen-specific humoral antibodies in the groups included in step 2 of the initial study. In a follow-up period of 3 years (i.e. up to 4 years after the second vaccine dose), safety monitoring also continued, specifically evaluation of serious adverse events (SAEs) and potential immune-mediated disease (pIMD).

Methods

Study design and participants

This was a 3-year open-label follow-up study (ClinicalTrials.gov identifier: NCT03201211) of a 14-month phase 1, randomised, observer-blind, placebo-controlled study conducted in Belgium between August 2015 and March 2017 (NCT02547974). The methods of the phase 1 study were described previously [13] and a study summary is available at www.gsk-studysregister.com (study identifier, 204913). Briefly, participants were randomised to receive two vaccine formulation doses 60 days apart. There were two steps: in step 1, 30 healthy adults aged 18–40 years received a non-adjuvanted vaccine formulation containing 10 μg of each NTHi antigen and 10 μg of UspA2 antigen per dose or placebo. In step 2, 90 healthy adults aged 50–71 years with a smoking history of at least 10 pack-years received an AS01e-adjuvanted formulation containing either 10 μg of each antigen (10–10–AS01) or 10 μg of each NTHi antigen and 3.3 μg of Mcat UspA2 (10–3–AS01), or placebo. The NTHi antigens were described previously [22]. AS01e is an Adjuvant System containing 3-O-desacyl-4-monophosphoryl lipid A (MPL), QS-21 (Quillaja saponaria Molina, fraction 21; licensed by GSK from Antigenics LLC, a wholly owned subsidiary of Agenus Inc., a Delaware, USA corporation) and liposome (25 μg MPL and 25 μg QS-21) [26].

In this follow-up study, the primary objective was to evaluate the persistence of humoral antibodies in the groups of participants included in step 2 of the initial study, up to 3 years after the last study visit. The last study visit of step 2 occurred 14 months after the study start and 12 months after the second vaccine dose. The secondary objective was to assess the long-term safety of the investigational NTHi-Mcat vaccine by monitoring the occurrence of SAEs and pIMD over the 3-year follow-up period. Eligible participants had participated in step 2 of the initial study, had received two doses, and were able to return for follow-up visits. Participants were excluded if they received or were scheduled to receive any investigational or non-registered vaccine or drug during the study. Other exclusion criteria included receipt of an immune-modifying drug, immunoglobulins or blood products during the study period, and any acute or chronic condition that could interfere with the results of the study.

The study was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice. The protocols and associated documents were reviewed and approved by an independent ethics committee. All participants provided written informed consent before study entry.

Antibody measurement

Immunoglobulin G antibody concentrations to each vaccine antigen were measured by enzyme-linked immunosorbent assay (ELISA), developed by GSK Biologicals and validated (for PD) or qualified (qualification in parallel with study conduct for PE, PilA and UspA2) on blood samples collected every 6 months during the 3-year follow-up, which started 14 months after the start of the study (12 months since the second vaccine dose), i.e. at 20,
26, 32, 38, 44 and 50 months from the start of the study. Sera were stored at −20 °C until assayed. Standardised procedures and in-house-made reference serum were used for each assay. The cut-off of the assays (i.e. lower limit of quantification, LLOQ) was 153 ELISA units (EU)/mL, 25 EU/mL, 16 EU/mL and 38 EU/mL for anti-PD, anti-PE, anti-PilA and anti-UspA2, respectively, for the first two time points, and 153 EU/mL, 16 EU/mL, 8 EU/mL and 28 EU/mL, respectively, for the remaining time points (post-qualification).

Safety analyses

During the entire follow-up study, the occurrence ofSAEs was recorded, defined as any untoward medical occurrence that resulted in death, was life-threatening, required hospitalisation or prolongation of hospitalisation, resulted in disability or incapacity, or was a congenital anomaly or birth defect in the child of a subject. Data on pIMD, which included autoimmune and other inflammatory or neurological disorders of interest [28], were also recorded regardless of seriousness. Investigators or site staff were responsible for detecting, documenting and reporting events meeting the criteria of a SAE or pIMD. Safety oversight was provided by a safety review team through ongoing routine safety monitoring.

Statistical analysis

It was planned to enrol all eligible participants from step 2 of the initial study [13]. The persistence of humoral antibodies was analysed in the per-protocol cohort for immunogenicity, which consisted of all eligible participants who were compliant with study procedures and had at least one assay result for at least one of the follow-up time points.

Descriptive exploratory analyses characterised the differences between groups in humoral immune response, as measured by ELISA geometric mean concentration (GMC). For analysis purposes, samples with concentrations that fall below the LLOQ were assigned a value of half the LLOQ to calculate the GMC. The difference between groups in antibody GMC was evaluated by calculating the 95% confidence intervals (CIs) of the ratio of antibody GMCs between groups, using a one-way analysis of covariance (ANCOVA) model on the log_{10} transformation of antibody concentrations. The ANCOVA model included group category as fixed effect and the baseline antibody log_{10} concentration (before the first vaccine dose in the initial study) as covariate. Geometric mean ratios (GMRs) with 95% CIs were calculated to describe the mean change in antibody concentration at a specific time point with respect to the pre-vaccination antibody concentration for each group. GMRs were calculated using a one-way analysis of variance model on the ratio between the log_{10} antibody concentration at a specific time point and the baseline log_{10} antibody concentration, with group category as a fixed effect. Any differences between groups or time points should be interpreted with caution, as no adjustment for multiplicity was performed when computing the CIs.

The safety analysis was performed on all enrolled participants. Statistical analyses were performed using Statistical Analysis System Version 9.4 on Life Science Analytics Framework 4.3 (SAS Institute Inc., Cary, NC, USA).

Results

Study population

Eighty-one healthy adults with a smoking history were enrolled in this follow-up study (27 in the 10–10-AS01 group, 26 in the 10–3-AS01 group and 28 in the placebo group). Immunogenicity results were available for at least one time point for all enrolled participants. Demographic characteristics were similar between groups (Table 1). All participants were white (European heritage).

Immunogenicity

Immune responses against theNTHi antigens (PD, PE and PilA) persisted during the 3-year follow-up period after initial study completion (Fig. 1; supplementary Fig. S1). At each follow-up time point, adjusted antibody GMCs against each NTHi antigen were higher in the vaccine groups versus placebo, as indicated by non-overlapping 95% CIs (Fig. 1). GMC ratios for vaccine group versus placebo were higher than or equal to 3.1 (95% CI: 2.0–4.9) for PD, 22.3 (13.5–36.8) for PE and 5.5 (3.5–8.7) for PilA (Fig. 2). In the vaccinated groups, GMRs at each time point were higher than or equal to 3.4 (95% CI: 2.5–4.8) for PD, 23.2 (15.7–34.3) for PE and 4.4 (3.1–6.3) for PilA (Fig. 1). Antibody GMC point estimates for the NTHi antigens were higher in the 10–3-AS01 group than in the 10–10-AS01 group at each time point (Fig. 1).

For the Mcat antigen, UspA2, GMC point estimates were higher in the vaccine groups at each follow-up time point compared to placebo (Fig. 1, supplementary Fig. S1). GMRs in the vaccinated groups were between 1.2 and 1.9 at each time point, apart from in the 10–3-AS01 group at months 44 and 50 (GMR 0.9); all GMC 95% CIs included 1 (Fig. 1). Antibody GMC ratios for 10–10-AS01 versus placebo and 10–3-AS01 versus placebo were 1.7 (95% CI: 1.2–2.2) or lower (Fig. 2).

Safety

At least one SAE was reported in nine (11.1%) participants: five reported nine SAEs in the 10–10-AS01 group, two reported two SAEs in 10–3-AS01 group, and two reported two SAEs in the placebo group (Table 2). No SAE category (preferred term) was reported more than once (Table 2) and none of the SAEs were assessed as related to vaccination. Eight participants experienced unsolicited adverse events leading to hospitalisation: four in the 10–10-AS01 group, two in the 10–3-AS01 group and two in the placebo group.

There was one death, which was due to lung cancer and occurred in the 10–10-AS01 group 607 days (approximately 20 months) after receiving the second vaccine dose. This was considered not related to study vaccination. One non-serious pIMD, trigeminal neuralgia, was reported in the 10–3-AS01 group in an individual 771 days (over 2 years) after the second vaccination. The pIMD was of moderate intensity, lasted 37 days, and the participant recovered fully. This event was not considered to be causally related to vaccination.

| Characteristic | 10–10-AS01 (N = 27) | 10–3-AS01 (N = 26) | Placebo (N = 28) |
|----------------|---------------------|---------------------|------------------|
| Age (years) at dose 1, mean (SD) | 59.7 (6.3) | 59.0 (5.9) | 58.2 (6.5) |
| Age group (years), n (%) | 50–59 | 14 (51.9) | 15 (57.7) | 18 (64.3) |
| 60–70 | 13 (48.1) | 11 (42.3) | 10 (35.7) |
| Male sex, n (%) | 15 (55.6) | 14 (53.8) | 19 (67.9) |
| Smoking status, n (%) | Current smoker | 10 (37.0) | 8 (30.8) | 10 (35.7) |
| Former smoker | 17 (63.0) | 18 (69.2) | 18 (64.3) |

10–10-AS01, group that received vaccine containing 10 µg of each non-typeable *Haemophilus influenzae* (NTHi) antigen and 10 µg of *Moraxella catarrhalis* (Mcat) antigen with AS01E; 10–3-AS01, group that received vaccine containing 10 µg of each NTHi antigen and 3.3 µg of Mcat antigen with AS01E; N, number of participants; n, number of participants in a specific category; SD, standard deviation.
Discussion

This is the first report of the long-term persistence of antibodies induced by vaccination against NTHi antigens, PD and PE-PilA, and Mcat antigen, UspA2. The results show the adjuvanted NTHi-Mcat vaccine formulations induced persistent immune responses against the NTHi antigens up to 4 years after the second vaccine dose, with the 10–3-AS01 formulation inducing the best humoral responses. There was no persistent response against UspA2. These findings are consistent with those from the initial 12-month follow-up period [13].

The good long-term persistence of antibody against NTHi antigens is possibly due to the adjuvant used in the vaccine, AS01E. The liposome-based AS01 Adjuvant System has been shown to enhance adaptive immune responses against antigens from pathogens causing complex diseases including malaria, hepatitis and shingles [26,29]. There is also evidence that inclusion of an Adjuvant System in the vaccine can produce persistent immune responses against H. influenzae. A study of an investigational vaccine containing three protein antigens, including H. influenzae PD, reported persistent anti-PD antibodies in healthy adults 1 year after two vaccine doses and higher responses when the vaccine was administered with an Adjuvant System containing α-tocopherol and squalene in an oil-in-water emulsion, AS03 [30].

In the initial study, there was a moderate but transient specific response against the Mcat antigen [13]. In the subsequent 3-year follow-up, all GMC ratio and GMR 95% CIs included 1 for each UspA2 assessment, indicating no differences between groups or differences from baseline. This lack of persistent immune response could be caused by several factors. It may have been due to natural boosting by repetitive exposure or colonisation by Mcat, and the conserved nature of UspA2, allowing successful detection before

Fig. 1. Geometric mean concentrations (GMCs with 95% CIs, adjusted for baseline antibody log_{10} concentrations) and geometric mean ratios (GMRs with 95% CIs) of log_{10} antibody concentrations at each time point versus pre-vaccination during follow-up (per-protocol immunogenicity cohort). Months 20, 26, 32, 38, 44 and 50 equate to 18, 24, 30, 36, 42 and 48 months after the second vaccine dose. Number of participants with available results at each time point: between 24 and 27 in 10–10-AS01 group, 23 and 26 in 10–3-AS01 group, 26 and 28 in placebo group. EU, enzyme-linked immunosorbent assay units; PD, protein D; PE, protein E; PilA, Pilin A; UspA2, ubiquitous surface protein A2; 95% CI, 95% confidence interval; 10–10-AS01, group that received vaccine containing 10 μg of each non-typeable Haemophilus influenzae (NTHi) antigen and 10 μg of Moraxella catarrhalis (Mcat) antigen with AS01E; 10–3-AS01, group that received vaccine containing 10 μg of each NTHi antigen and 3.3 μg of Mcat antigen with AS01E.
vaccination in the ELISA. The concentration of anti-UspA2 antibodies before vaccination was relatively high in all groups (384.1–572.5 EU/mL in the initial study [13]; see also Fig. S1), suggesting strong natural exposure. An inverse association between pre-vaccination titre and vaccine response has been reported in studies of other vaccines, such as influenza and respiratory syncytial virus vaccines, and oral cholera vaccines [31–33]. Also, it is clear from individual subject listings that the immune response against UspA2 was highly variable (data not shown). While the UspA2 protein induces a bactericidal and cross-reactive immune response [21], recent evidence indicates that it has a variable sequence and structure [34,35], which may have an impact on antibody recognition and binding.

As reported in the initial 14-month study [13], in the 4-year follow-up, the vaccine formulation containing 3.3 µg UspA2 induced higher specific responses (higher antibody GMC point estimates) against the NTHi antigens than the formulation containing 10 µg UspA2. Conversely, anti-UspA2 point estimates were lower with the 3.3 µg UspA2 formulation than with the 10 µg formulation. This is suggestive of immunological interference, with a higher quantity of UspA2 leading to a higher response against the Mcat antigen but lower response against NTHi antigens, and the formulation with lower UspA2 quantity inducing a lower UspA2 response but better responses against NTHi antigens.

No safety concerns were identified during the long-term follow-up period. Thirteen SAEs were reported over 3 years, with no SAE (preferred term) reported more than once, and one non-serious pIMD was reported, occurring in the 10–3-AS01 group. One death occurred in the 10–10-AS01 group. All were assessed as not causally related to vaccination.

This study is limited by its open-label design and small sample size. Also, the long-term persistence of antibodies may differ in the target population for the vaccine (COPD patients). A strength of the study was the enrolment of most participants (81) from the cohort of 90 included in step 2 of the initial study.
### Table 2
Participants with at least one serious adverse event (SAE), by Medical Dictionary for Regulatory Activities (MedDRA) preferred term.

| MedDRA preferred term                  | Number of participants (percentage; 95% CI) |
|----------------------------------------|---------------------------------------------|
| **At least one SAE**                   |                                             |
| 10–10-AS01 (N = 27)                    |                                             |
| 10–3-AS01 (N = 26)                     |                                             |
| Placebo (N = 28)                       |                                             |
| At least one SAE                       | 5 (18.5; 6.3–38.1)                         |
| Humerus fracture                       | 1 (3.7; 0.1–19.0)                          |
| Tibia fracture                         | 0                                           |
| Tendon rupture                         | 1 (3.7; 0.1–19.0)                          |
| Post-procedural fever                  | 1 (3.7; 0.1–19.0)                          |
| Lung neoplasm malignant                | 1 (3.7; 0.1–19.0)                          |
| Schwannoma                             | 1 (3.7; 0.1–19.0)                          |
| Ileus                                  | 1 (3.7; 0.1–19.0)                          |
| Ileus paralytic                        | 1 (3.7; 0.1–19.0)                          |
| Intestinal obstruction                 | 1 (3.7; 0.1–19.0)                          |
| Post-operative wound infection         | 1 (3.7; 0.1–19.0)                          |
| Spinal stenosis                        | 1 (3.8; 0.1–19.0)                          |
| Ischaemic stroke                       | 0                                           |
| Nephrolithiasis                        | 0                                           |

10–10-AS01, group that received vaccine containing 10 μg of each non-typeable *Haemophilus influenzae* (NTHi) antigen and 10 μg of *Moraxella catarrhalis* (Mcat) antigen with AS01E; 10–3–AS01, group that received vaccine containing 10 μg of each NTHi antigen and 3.3 μg of Mcat antigen with AS01E. N, number of participants for each study group; 95% CI, exact 95% confidence interval.

Post-procedural fever, Schwannoma and post-operative wound infection reported in a single subject; ileus, ileus paralytic and intestinal obstruction reported in a single subject.

In conclusion, in the 4-year period after two-dose vaccination with the investigational NTHi–Mcat vaccine, there was long-term persistence of immune responses against the NTHi protein antigens. This was not observed for the Mcat antigen, UspA2, although a decline in anti-UspA2 immune response was already present at the end of the initial 14-month study. No safety concerns were identified.

### Declaration of Competing Interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: PDS declares no financial or non-financial relationships and activities and no conflicts of interest. GL-R, CV, AT, GDM, MD, DC, MA, DR and AKA declare no other financial or non-financial relationships which may be considered as potential competing interests. PDS declares no financial or non-financial relationships associated with the development and publication of this manuscript.

### Author contributions

DC and AKA were involved in the study conception and design. PDS, GL-R, CV and MD were involved in acquisition and generation of data and/or performed the study. AT, GDM, MD, DC, MA, DR and AKA were involved in data analysis and data interpretation. All authors had full access to the data and contributed substantially to the development of the manuscript and gave final approval before submission. All authors attest they meet the International Committee of Medical Journal Editors criteria for authorship.

### Funding

GlaxoSmithKline Biologicals SA funded this study and was involved in all stages of study conduct, including analysis of the data. GlaxoSmithKline Biologicals SA also took charge of all costs associated with the development and publication of this manuscript.

### Data sharing statement

Anonymised individual participant data and study documents can be requested for further research from www.clinicalstudydatarequest.com.

### Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.jvacx.2021.100124.

### References

[1] GBD 2017 Causes of Death Collaborators. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980–2017: a systematic analysis for the Global Burden of Disease Study 2017. Lancet 2018;392:1736–88. doi: 10.1016/s0140-6736(18)32203-7.

[2] Adeloye D, Chua S, Lee C, Basquill C, Papana A, Theodoratou E, et al. Global and regional estimates of COPD prevalence: systematic review and meta-analysis. J Glob Health 2015;5:https://doi.org/10.7189/jogh.05-020415.

[3] Celli BR, Wedzicha JA. Update on clinical aspects of chronic obstructive pulmonary disease. N Engl J Med 2019;381(13):1257–66. https://doi.org/10.1056/NEJMoa1900550.
[4] Global Initiative for Chronic Obstructive Lung Disease (GOLD), Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease, 2021 report, 2020. https://goldcopd.org/wp-content/uploads/2020/11/GOLD-REPORT-2021-v1.0-16Nov20_WMV.pdf. (Accessed 20 Apr 2021).

[5] Fros E, Roche N, Blasi F. Pneumococcal vaccination and chronic respiratory diseases. Int J Chron Obstruct Pulmon Dis 2017;12:3457–68. https://doi.org/10.2147/copd.s140378.

[6] Huang YJ, Erb-Downward JR, Curtis JL, Huffnagle GB, Han MK. Pulmonary diseases. Int J Chron Obstruct Pulmon Dis 2017;12:3457–68. https://doi.org/10.2147/copd.s140378.

[7] Mayhew D, Devos N, Lambert C, Brown JR, Clarke SC, Kim VL, et al. Longitudinal profiling of the lung microbiome in the AERS study demonstrates repeatability of bacterial and eosi

[8] Wilkinson TMA, Aris E, Bourne S, Clarke SC, Peeters M, Pascal TG, et al. A prospective, observational cohort study of the seasonal dynamics of airway pathogens in the aetiology of exacerbations in COPD. Thorax 2017;72(10):919–27. https://doi.org/10.1136/thoraxjnl-2016-209073.

[9] Sethi S, Evans N, Grant BBJ, Murphy TF. New strains of bacteria and exacerbations of chronic obstructive pulmonary disease. N Engl J Med 2002;347(7):645–751. https://doi.org/10.1056/NEJMoa020561.

[10] Tan TT, Morgen G, Forsgren A, Riesbeck K. Haemophilus influenzae survival during complement-mediated attacks is promoted by Moraxella catarrhalis outer membrane vesicles. J Infect Dis 2007;195(11):1661–70. https://doi.org/10.1086/524701108661176111.

[11] Armstrong CE, Hong W, Pang B, Wiener KED, Juneau RA, Turner J, et al. Indirect pathogenicity of Haemophilus influenzae and Moraxella catarrhalis in polymicrobial otitis media occurs via interspecies quorum signaling. mBio 2016;7(1). https://doi.org/10.1128/mBio.00102-16.

[12] Schaar V, Nordström T, Mørgen G, Riesbeck K. Moraxella catarrhalis outer membrane vesicles carry beta-lactamase and promote survival of Streptococcus pneumoniae and Haemophilus influenzae by inactivating amoxicillin. Antimicrob Agents Chemother 2011;55:3845–53. https://doi.org/10.1128/aac.01772-10.

[13] Van Damme P, Leroux-Roels C, Vandermeulen C, De Ryck I, Tasciotti A, Dozot J, et al. Safety and immunogenicity of non-typeable Haemophilus influenzae-Moraxella catarrhalis vaccine. Vaccine 2019;37(23):3113–22. https://doi.org/10.1016/j.vaccine.2019.04.041.

[14] Davoudi Vijeh Motlagh A, Siadat SD, Abedian Kenari S, Mahdavi M, Behrouzi A, et al. Safety and immunogenicity of a respiratory syncytial virus fusion glycoprotein and hepatitis B surface antigen in healthy adult volunteers. Vaccine 2008;26(10):1375–86. https://doi.org/10.1016/j.vaccine.2007.12.038.

[15] Berglund J, Vink P, Tavares Da Silva F, Koster E, Veitch T, Pasquet M, et al. Safety, reactogenicity and immunogenicity of a investigational non-typeable Haemophilus influenzae and Moraxella catarrhalis cross-protective vaccine antigen. Vaccine 2021.https://doi.org/10.1016/j.vaccine.2021.08.002.

[16] Desai SN, Cravioto A, Sur D, Kanungo S. Maximizing protection from use of oral cholera vaccines in developing country settings: an immunological review of potential immune mediated disorders in clinical trials of new vaccines. Vaccine 2013;31(14):1870–6. https://doi.org/10.1016/j.vaccine.2013.01.042.

[17] Didierlaurent AM, Laupèze B, Di Pasquale A, Herliq N, Cillignon G, Caron N. Adjuvant system AS01: helping to overcome the challenges of modern vaccines. Expert Rev Vaccines 2017;16(1):55–63. https://doi.org/10.1080/14760588.2016.1213632.

[18] Didierlaurent AM, Schenardi S, Brightling C, Bakerly ND, Lewis K, MacNee W, et al. Non-typeable Haemophilus influenzae protein E recognizes the C-terminal domain of vitronectin and protein D of Haemophilus influenzae in mice and chinchillas. Infect Immun 2019;87(8). https://doi.org/10.1128/IAI.00345-19.

[19] Forsgren A, Riesbeck K, Janson H. Protein D of Haemophilus influenzae and Moraxella catarrhalis outer molecules mediates the interaction with pneumococcal polysaccharide vaccines in developing country settings: an immunological review of potential immune mediated disorders in clinical trials of new vaccines. Vaccine 2013;31(14):1870–6. https://doi.org/10.1016/j.vaccine.2013.01.042.