TECHNOLOGY REPORT

First siRNA library screening in hard-to-transfect HUVEC cells

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ABSTRACT

Meaningful RNAi-based data for target gene identification are strongly dependent on the use of a biologically relevant cell type and efficient delivery of highly functional siRNA reagents into the selected cell type. Here we report the use of the Amaxa® Nucleofector® 96-well Shuttle® System for siRNA screening in primary cells. Lonza’s Clonetics® HUVEC-Human Umbilical Vein Endothelial Cells were transfected with Thermo Scientific Dharmacon siGENOME® siRNA Libraries targeting protein kinases and cell cycle related genes and screened for genes important for cell viability. Of the 37 primary hits, down-regulation of 33 led to reduced proliferation or increased cell death, while down-regulation of two allowed for better cell viability. The validated four genes out of the 16 strongest primary hits (COPB2, PYCS, CDK4 and MYC) influenced cell proliferation to varying degrees, reflecting differing importance for survival of HUVEC cells. Our results demonstrate that the Nucleofector® 96-well Shuttle® System allows the delivery of siRNA libraries in cell types previously considered to be difficult to transfect. Thus, identification and validation of gene targets can now be conducted in primary cells, as the selection of cell types is not limited to those accessible by lipid-mediated transfection.

KEYWORDS: Nucleofection, RNAi, siRNA, primary cell, screening, transfection, HUVEC

INTRODUCTION

RNAi-based library screening has become a powerful in vitro tool to identify drug targets that play a role in disease development and progression (Martin and Caplen, 2007). Successful screening experiments using siRNA require efficient delivery of highly functional and specific siRNA molecules into appropriate cells. While lipid-mediated transfection is a common approach for siRNA delivery, many cell types, including suspension cell lines and primary cells, are not compatible with this technology (Merkerova et al, 2007). This limitation prevents analysis of many biologically relevant cell types and restricts siRNA library screenings mainly to transformed, adherent cells that often exhibit phenotypic and genetic anomalies after extended periods of culturing lines (MacKeigan et al, 2005; Bartz et al, 2006; Whitehurst et al, 2007). Ideally, the diversity of biological questions requires the use of appropriate cell types, typically primary cells. In addition to this issue, several of the lipid delivery reagents can cause cytotoxicity and are capable of inducing a potent interferon response and/or altering gene expression profiles (Marques and Williams, 2005; Fedorov et al, 2005; Wang, 2006). These unintended phenotypes can significantly affect experimental outcomes and drastically interfere with identifying relevant genes and understanding a gene’s function. Human Umbilical Vein
Endothelial Cells (HUVEC), a difficult-to-transfect cell type, were screened with an siRNA library delivered using the Amaxa® Nucleofector® 96-well Shuttle® System. The screen targeted protein kinases and genes associated with the cell cycle to identify target genes important for cell viability.

**MATERIALS AND METHODS**

The siRNA reagents used were Dharmacon Human siGENOME® SMARTpool® siRNA Libraries for Protein Kinases (targeting 779 genes) and Cell Cycle Regulation (targeting 111 genes) (Thermo Fisher Scientific). Clontech® HUVEC Cells (Lonza) were cultured in Clontech® EGM® Endothelial Growth Medium (Lonza) at 37°C, 5% (v/v) CO₂ and transfected according to the recommendations in the respective Optimized Protocol for 96-well Nucleofection® (Amaxa). Briefly, 2 x 10⁴ HUVEC cells were transfected with 20 pmol siRNA (if not noted differently). For optimal assay conditions, post-transfection HUVEC cells were plated in 96-well culture plates at a density of 2 x 10⁴ cells per well (100 μl). Outer wells of culture plates were filled with media only in order to avoid edge effects in the phenotypic assays. HUVEC cells were analyzed 72 hrs post-transfection for cell viability.

For the primary screen (n=3 independent experiments), Clontech® HUVEC cells were transfected with the respective libraries or control siRNAs and analyzed for phenotypic effects (cell viability). Data from each screen were analyzed by statistical means: the Z’ factors (Zhang et al, 2002; Reagan-Shaw and Ahmad, 2005). CHEK-1 is expected to decrease cell viability. Using PLK-1, post-transfection plating densities were adjusted to allow for significant discrimination of positive and negative control samples on the phenotypic level. This was achieved by plating HUVEC cells at a low cell density of 2 x 10⁴ per well for 3-4 days after transfection (Figure 1A). As shown in Figure 1B, the phenotypic effect for PLK-1 silencing was weaker and built up slower than silencing of CHEK-1, thus representing potential differences expected for “strong” and “weak” library targets. It has been reported earlier that PLK-1 depletion by siRNA transfection exerts a strong effect on cancer cell lines, but not primary cells (Spänkuch-Schmitt et al, 2002; Reagan-Shaw and Ahmad, 2005). However, as the down-regulation of PLK-1 mRNA was not demonstrated in the previous reports, it could not be excluded that the underlying cause was more of an issue of inefficient transfection of the siRNA rather than the significance of PLK-1 roles in cell survival in primary cells. Nevertheless, despite a PLK-1 mRNA knockdown of more than 90% (data not shown), cell survival of HUVEC cells was diminished only to approximately 50% after 96 hrs, suggesting that persistent PLK-1 may indeed be a factor in survivability of cancer cells. An analysis time point of 72 hrs suited well for both targets. In pilot screens for further determination of assay robustness, controls were plated into the central 60 wells of a 96-well culture plate and analyzed for cell viability. Wells in the outer rows were filled with medium to avoid potential edge effects. Z’ factors of both positive controls (CHEK-1: 0.55; PLK-1: 0.22; Figure 1C; data for PLK-1 not shown) reflected a suitable window for discrimination of potential hits with different phenotypic strength in the subsequent screen from background.

**RESULTS AND DISCUSSION**

**Viability assay optimization**

For the kinase and cell cycle screen in HUVEC Cells, siRNA reagents targeting polo-like kinase 1 (PLK-1) and Cell Cycle Check-point Kinase 1 (CHK-1 or CHEK-1) were selected as positive controls to set up the viability assay. PLK-1 is a key regulator of mitotic progression in mammalian cells and the knock-down of PLK-1 is known to induce apoptosis in cancer cells (Spänkuch-Schmitt et al, 2002; Reagan-Shaw and Ahmad, 2005). CHEK-1 is involved in the DNA damage response and is also required for cell proliferation and survival. CHEK-1 knockdown by siRNA has been reported to induce mitotic arrest (Tang et al, 2006). As such, down-regulation of PLK-1 and/or CHEK-1 is expected to decrease cell viability. Using PLK-1, post-transfection plating densities were adjusted to allow for significant discrimination of positive and negative control samples on the phenotypic level. This was achieved by plating HUVEC cells at a low cell density of 2 x 10⁴ per well for 3-4 days after transfection (Figure 1A). As shown in Figure 1B, the phenotypic effect for PLK-1 silencing was weaker and built up slower than silencing of CHEK-1, thus representing potential differences expected for “strong” and “weak” library targets. It has been reported earlier that PLK-1 depletion by siRNA transfection exerts a strong effect on cancer cell lines, but not primary cells (Spänkuch-Schmitt et al, 2002; Reagan-Shaw and Ahmad, 2005). However, as the down-regulation of PLK-1 mRNA was not demonstrated in the previous reports, it could not be excluded that the underlying cause was more of an issue of inefficient transfection of the siRNA rather than the significance of PLK-1 roles in cell survival in primary cells. Nevertheless, despite a PLK-1 mRNA knockdown of more than 90% (data not shown), cell survival of HUVEC cells was diminished only to approximately 50% after 96 hrs, suggesting that persistent PLK-1 may indeed be a factor in survivability of cancer cells. An analysis time point of 72 hrs suited well for both targets. In pilot screens for further determination of assay robustness, controls were plated into the central 60 wells of a 96-well culture plate and analyzed for cell viability. Wells in the outer rows were filled with medium to avoid potential edge effects. Z’ factors of both positive controls (CHEK-1: 0.55; PLK-1: 0.22; Figure 1C; data for PLK-1 not shown) reflected a suitable window for discrimination of potential hits with different phenotypic strength in the subsequent screen from background.

**Primary screen and hit validation**

HUVEC Cells were transfected with siRNA pools targeting individual genes in the Human siGENOME® siRNA Libraries for protein kinases and cell cycle regulation. Multiple independent screening experiments (n=3) were performed to confirm the reproducibility of individual primary hits. Robust Z-score for cell viability was calculated for each of the 890 targets in the three independent experiments. As an example, the robust Z-scores of one screening experiment are shown in Figure 2A. A substantial proportion of targets displayed a median absolute deviation (MAD) below -3 or above 3 (MAD >3) including our positive controls PLK-1 and CHEK-1, which are members of both libraries. Thirty-five targets plus CHEK-1 and PLK-1 had a mean MAD greater than 3 in the three screening experiments and thus were considered as potential hits (Figure 2F). Eighteen of these targets showed a robust phenotype with MAD of greater than 3 in all three screening experiments (Figure 2B), while 24 targets were significant in two of the three experiments (Figure 2C) and 123 targets showed MAD greater than 3 in just one experiment (Figure 2D). The remaining 725 targets showed no response beyond the threshold in any experiment. The categories of targets with one or two experiments with MAD greater than 3 showed the highest standard deviations due to the occurrence of outliers. These categories also contained targets with a small standard deviation, suggesting that their importance for survival of HUVEC was not sufficiently strong for the
Figure 1. Determination of optimal assay conditions. In three independent experiments, HUVEC cells were transfected with 20 pmol SMARTpool® siRNA targeting PLK-1 (A, B) or CHEK-1 (B, C) and siGENOME® non-targeting control. Cell viability was analyzed at different time points post Nucleofection® (A/B: 24, 48, 72 and 96 hrs; C: 72 hrs). Values were normalized to the negative control samples (A, B) or to untreated cells (C). The rightmost dots in C represents the mean and SD of the 60 individual values.

chosen assay conditions (Figure 2B and C). Every sixth target fell into these categories of unclear importance, thus five of six targets were reproducibly classified as either important or not relevant for survival of HUVEC (Figure 2C and D). Generally, a considerable degree of variation was seen in the data, including the most significant hit category (Figure 2B). The most striking examples were MYC, which nevertheless could be validated, and CHEK1, which was a member of the library and also served as a positive control (Figure 2B and F). The degree of data variation might be attributed to not fully standardized cell culture conditions during the preparation of the experiments and thus argues for our strategy to select the primary hits from repeated screening experiments. While a higher number of primary hits for validation lowered the probability for erroneously classified false negatives, it also lowered the validation rate. Of the 35 identified primary hits, 33 had a pro-proliferative/anti-apoptotic function, as their down-regulation led to increased cell death, while 2 had an anti-proliferative effect as their knockdown allowed for better cell viability. The 16 strongest of the 33 pro-proliferative hits plus the two positive controls were selected for further evaluation (Table 1).

Five of the 18 selected targets (COPB2, CDK4, PYCS, MYC and PLK-1) were validated by demonstrating that the phenotype could be reproduced with multiple individual siGENOME® siRNAs from the original SMARTpool® and further with different siRNAs, i.e., an ON-TARGETplus® SMARTpoolV (Figure 3A, Table 1). The phenotypes could be correlated to the knockdown on mRNA level (Figure 3B) and to the amount of transfected siRNA (Figure 3C; only COPB2 shown). Both results, redundant phenotypic effects with an alternative siRNA pool and linking these phenotypes to proven specific mRNA knockdown, suggest that these were not off-target effects.

The reduction in cell survival with the ON-TARGETplus® SMARTpool® was mostly comparable to the corresponding siGENOME® SMARTpool®, but weaker than the strongest single siGENOME® siRNAs (Figure 3A and B). In the case of CDK4, the phenotypic effect of the ON-TARGETplus® SMARTpool® was comparable to siGENOME® siRNA duplex 4 despite a lower knock-down of the target mRNA (Figure 3A and B). For siRNAs of the same type, e.g., siGENOME® siRNAs, the phenotypic effects correlated well with the observed knock-down efficiencies (Figure 3A and B). Six of the 18 selected targets were confirmed with two of four single siGENOME® siRNAs, three of which could not be confirmed with the ON-TARGETplus® SMARTpool®, still suggesting them as potential hits that require further efforts for validation, such as expanding the set of tested single siRNAs. As discussed above for ON-TARGETplus® siRNAs, the insufficient phenotypic effect of the siRNAs, which could not be validated, may be explained by lower knock-down efficiencies or the reduction of off-target effects that contributed to the overall phenotype.
Figure 2. Primary Screen. HUVEC cells were transfected with 20 pmol of the combined Human siARRAY® SMARTpool® siRNA Libraries for Kinases (targeting 779 genes) and Cell Cycle Regulators (targeting 111 genes). Cell viability was analyzed 72 hrs post-Nucleofection®. (A) Representation of robust Z-scores of cell viability measures from 1 screening experiment. (B-E) Robust Z-scores of all primary hits with MAD of >3 in all three (B), two of three (C), or one of three (D) independent experiments and with MAD of <3 in all three experiments (E). (F) Robust Z-scores of the top 37 primary hits (with MAD of >3) from three independent experiments.
Figure 3. Hit validation. HUVEC cells were transfected with 20 pmol (if not indicated differently) siGENOME® (siG) SMARTpool® or single siRNA #1 - 4 (from the de-convoluted pool) targeting CDK4, COPB2, MYC or PYCS. CHEK-1 and siGENOME® Non-Targeting siRNA #1 (control siRNA) and siRNA targeting CHEK-1 (CHEK-1) served as controls. 72 hrs post-Nucleofection® cell viability was analyzed and normalized to control siRNA (A, C) and mRNA levels were determined for CDK4 (B) and COPB2 (B, C) and normalized to cyclophilin B mRNA and the control siRNA.

Five of the eighteen selected targets were considered “false positives”, because neither the ON-TARGETplus® SMARTpool®, nor more than one of four single siRNAs reproduced the phenotype seen with the original siGENOME® SMARTpool®. Thus, it is likely that these were the result of off-target effects of individual siRNA sequences. As for the above mentioned primary hits that showed a higher rate of validated single siRNAs, testing further single siRNAs with proven mRNA knockdown may allow a more definitive hit stratification.

Previous reports demonstrate the involvement of most of the identified primary hits in cell cycle regulation, cell survival or apoptosis (Table 1). No such evidence could be found for the kinases IRAK3, considered as false positive, and RPS6K1, which has been validated with two of four single siGENOME® siRNAs. Hence, five of the classified “false positives” have been reported in earlier literature, but their importance for HUVEC cell survival and proliferation remain unclear. For a majority of the identified primary hits, earlier reports describe their importance for cancer cells, while little information is available with respect to primary cells.

Cyclins, cyclin-dependent kinases and their substrates play pivotal roles during cell cycle progression and control proliferation of normal cells. Cyclin-dependent kinase 4 (CDK4) phosphorylates retinoblastoma protein (Rb) and other Rb-related proteins (Ewen et al, 1993; Kato et al, 1993; Leng et al, 2002) ultimately promoting the expression of various genes essential for cell cycle phase G1-S transition (Nevins, 2001). Low molecular-weight inhibitors of CDK4 lead to a delayed G2/M progression with reduced cell growth and mitosis rates in a number of cell lines (Burgess et al, 2006). However, CDK4, as well as the functionally redundant CDK6 and the associated D-type cyclins (D1, D2 and D3) are not essential for cell proliferation in mammalian cell types (Kozar and Sicinski, 2005; Malumbres, 2004). This explains the moderate, albeit consistent, reduction in cell viability after CDK4 knockdown in our experiments. Nevertheless, inhibitors specific for CDK4 and CDK6 can show significant antiproliferative activity against Rb-positive tumor cells (Fry, 2004).

The proto-oncogene c-MYC has been intensively studied and its deregulation by various mitogens leads to the genesis of diverse human cancers (Oster, 2002). The gene product of c-MYC is a transcription factor, involved in the regulation of cell cycle related genes, such as CDK4 or Cyclin B1 (Menssen, 2002). While deregulation of c-MYC results in hyperproliferation, antisense oligo-mediated...
Table 1. Protein kinases and cell cycle regulators in HUVEC cells. Top hits were selected from the primary screen. Hits are shown in descending order of MADs. “k” indicates members of the siRNA library against kinases, “cc” indicates members of the siRNA library against cell cycle related genes.

| gene | library | number of validated single siGENOME® siRNAs [of 4] | with GTP pool siRNAs | hit category | Previous evidence for involvement in cell cycle, cell survival or apoptosis | evidence | reported cell type / tissue | author |
|------|---------|-------------------------------------------------|----------------------|--------------|-------------------------------------------------|----------|-----------------------------|--------|
| COPB2 | k       | 3                                               | X                    | validated hit | indirect | CLL-B-cells | see discussion |
| CMKSR1 | k       | 1                                               | false positive       | indirect | HeLa, Cos cells | Jaffe et al., 2004 |
| RAPGFE3 | k       | 3                                               | candidate            | direct | HUVEC | Namkoong et al., 2009 |
| CDK4 | k,cc | 2                                               | X                    | validated hit | direct | various | see discussion |
| CHEK1 | k,cc | 2                                               | X                    | validated positive control | direct | various cancer cells | see discussion |
| PYCS | k       | 3                                               | X                    | validated hit | indirect | cancer cell lines | see discussion |
| IRAK3 | k       | 1                                               | false positive       | none | cancer cells | Yalcin et al., 2009 |
| PFKFB3 | k       | 1                                               | false positive       | indirect | cancer cells | Emami et al., 2009 |
| NLK | k       | 2                                               | candidate            | direct | prostate | Emami et al., 2009 |
| MYC | cc | 3                                               | validated hit       | direct | cancer cell lines | see discussion |
| RPS6KL1 | k       | 2                                               | candidate            | none | cancer cell lines | see discussion |
| EDN2 | k       | 2                                               | X                    | candidate | direct | melanocytes | Yada et al., 1991 |
| PKM2 | k       | 3                                               | candidate            | indirect | Cos-7 cells | Steták et al., 2007 |
| MS1R | k       | 1                                               | false positive       | indirect | cancer cell lines | Follenzi et al., 2000 |
| RIOK1 | k       | 1                                               | false positive       | indirect | colon cancer | Line et al., 2002 |
| MARK3 | k       | 2                                               | candidate            | indirect | Cos cells | Müller et al., 2001 |
| CENPE | cc | 1                                               | false positive       | direct | HeLa cells | Kittler et al., 2004 |
| PLK1 | k,cc | 3                                               | X                    | validated positive control | direct | cancer cell lines | see discussion |

CONCLUSIONS

The presented data demonstrate that the Nucleofector® 96-well Shuttle® System allows the delivery of siRNA libraries, e.g., Dharmacon siGENOME® Libraries, in cell types previously considered difficult to transfect. Preserved cell functionality and efficient mRNA knockdown allow the identification and validation of gene targets in primary cells, which reflect a higher biological relevance for certain pathways.

Most of the identified and validated targets have been reported earlier to be implicated in the cell cycle, cell survival or apoptosis in cancer cells, while little has been known about their importance in normal primary cells.

Our study suggests that overcoming the limitation of using transformed cell lines for functional screens and rather studying the relevant primary cells pledge the potential to identify novel drug target candidates.

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COMPETING INTERESTS

MZ, LMA NUES, SS, SMO, SBSD, AT and HAMH work for Lonza Cologne AG R&D. ASA and DL work for and/or have financial interests in Thermo Fisher Scientific.
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