Pan-Cancer Analysis Reveals that the SARS-CoV-2 Receptor ACE2 is a Protective Factor for Cancer Progression

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Research

**Keywords:** ACE2 expression, pan-cancer, tumor immunity and immunotherapy, tumor progression, survival prognosis

**DOI:** https://doi.org/10.21203/rs.3.rs-42534/v1

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Abstract

**Background:** The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has infected more than 13 million people and has caused more than 570,000 deaths worldwide as of July 13, 2020. The SARS-CoV-2 human cell receptor ACE2 has recently received extensive attention for its role in SARS-CoV-2 infection. Many studies have also explored the association between ACE2 and cancer. However, a systemic investigation into associations between ACE2 and oncogenic pathways, tumor progression, and clinical outcomes in pan-cancer remains lacking.

**Methods:** Using cancer genomics datasets from the Cancer Genome Atlas (TCGA) program, we performed computational analyses of associations between ACE2 expression and antitumor immunity, immunotherapy response, oncogenic pathways, tumor progression phenotypes, and clinical outcomes in 12 cancer cohorts. We also identified co-expression networks of ACE2 in cancer.

**Results:** ACE2 upregulation was associated with increased antitumor immune signatures and PD-L1 expression, and favorable anti-PD-1/PD-L1/CTLA-4 immunotherapy response. ACE2 expression levels inversely correlated with the activity of cell cycle, mismatch repair, TGF-β, Wnt, VEGF, and Notch signaling pathways. Moreover, ACE2 expression levels had significant inverse correlations with tumor proliferation, stemness, and epithelial-mesenchymal transition (EMT). ACE2 upregulation was associated with favorable survival in pan-cancer and in multiple individual cancer types.

**Conclusions:** ACE2 upregulation was associated with increased antitumor immunity and immunotherapy response, reduced tumor malignancy, and favorable survival in cancer, suggesting that ACE2 is a protective factor for cancer progression. Our data may provide potential clinical implications for treating cancer patients infected with SARS-CoV-2.

Background

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has infected more than 13 million people and has caused more than 570,000 deaths worldwide as of July 13, 2020 (https://coronavirus.jhu.edu/map.html). SARS-CoV-2 uses the angiotensin-converting enzyme 2 (ACE2) as a host cell receptor to infect humans [1, 2, 3, 4]. ACE2 plays an important role in regulating cardiovascular and renal function [5]. This protein has recently received extensive attention for its role in SARS-CoV-2 infection [1, 2, 4]. Our recent study revealed that ACE2 is expressed in various human tissues [6], suggesting that SARS-CoV-2 may invade various human organs besides the lungs. Also, many studies have investigated the association between ACE2 and cancer [7–13]. For example, Yu-Jun et al. analyzed ACE2 expression in various cancers and revealed a positive association between ACE2 expression and survival prognosis in liver cancer [7]. Cai et al. described the genetic alteration, mRNA expression, and DNA methylation of ACE2 in over 30 cancer types and revealed genetic and epigenetic variations of ACE2 in various cancers [8]. Several studies demonstrated that ACE2 had antitumor effects by inhibiting tumor angiogenesis [10, 11, 13]. A recent study [14] showed that ACE2 expression was associated with
increased tumor immune infiltration and was a positive prognostic factor in uterine corpus endometrial and renal papillary cell cancers. Nevertheless, a systemic investigation into the association between ACE2 expression and antitumor immunity, oncogenic pathways, tumor progression phenotypes, and clinical outcomes in pan-cancer remains lacking.

In this study, we investigated associations between ACE2 expression and antitumor immune signatures in 12 human cancer cohorts from the Cancer Genome Atlas (TCGA) program (https://cancergenome.nih.gov/). We also explored associations between ACE2 expression and multiple tumor phenotypes, including cell proliferation, stemness, epithelial-mesenchymal transition (EMT), oncogenic signaling, and clinical outcomes in these cancer cohorts. We also investigated the association between ACE2 expression and immunotherapy response in four cancer cohorts receiving the immune checkpoint blockade therapy. This study aimed to provide new insights into the association between ACE2 and cancer and the potential association between cancer and SARS-CoV-2 infection.

**Methods**

**Datasets**

From the genomic data commons data portal (https://portal.gdc.cancer.gov/), we obtained RNA-Seq gene expression profiling datasets (level 3 and RSEM normalized) for 12 TCGA cancer cohorts. The 12 cancer cohorts included cervical squamous-cell carcinoma (CESC), colon adenocarcinoma (COAD), esophageal carcinoma (ESCA), head and neck squamous cell carcinoma (HNSC), kidney renal clear cell carcinoma (KIRC), kidney renal clear cell carcinoma (KIRP), lung adenocarcinoma (LUAD), lung squamous cell carcinoma (LUSC), skin cutaneous melanoma (SKCM), thymoma (THYM), uterine corpus endometrial carcinoma (UCEC), and ovarian carcinoma (OV). We log2-transformed all RSEM-normalized gene expression values before further analyses. Besides, we obtained gene expression profiling and clinical data in four cancer cohorts receiving anti-PD-1/PD-L1/CTLA-4 immunotherapy from their related publications, including Nathanson (melanoma) [15], Topalian (melanoma) [16], Ascierto (renal cell carcinoma) [17], and Snyder (bladder cancer) cohorts [18]. A summary of these datasets is presented in Supplementary Table S1 (Additional file 1).

**Evaluating the enrichment levels of immune signatures, pathways, and tumor phenotypes**

We evaluated the enrichment level of a pathway or tumor phenotype in a tumor sample by the single-sample gene-set enrichment analysis (ssGSEA) score [19]. The gene set included all marker genes of a pathway or tumor phenotype. A total of six cancer-associated pathways (cell cycle, mismatch repair, TGF-β, Wnt, VEGF, and Notch signaling) and three tumor phenotypes (cell proliferation, stemness, and EMT) were analyzed. We presented the marker genes of these pathways and tumor phenotypes in Supplementary Table S2 (Additional file 2).

**Gene-set enrichment analysis**
We defined high-ACE2-expression-level (upper third) and low-ACE2-expression-level (bottom third) tumors in each cancer type based on ACE2 expression profiles. We identified the KEGG [20] pathways highly enriched in both groups of tumors using GSEA [21] with a threshold of adjusted \( p \)-value < 0.05. Moreover, we used WGCNA [22] to detect the gene modules (gene ontology) differentially enriched between the high- and low-ACE2-expression-level tumors in pan-cancer. We identified the hub genes as the genes connected to at least 5 other genes with a connectedness weight greater than 0.25 in a gene module and built their co-expression network.

**Statistical analysis**

We used Spearman’s correlation test to evaluate the correlation (\( \rho \)) of ACE2 expression levels with the enrichment levels of pathways or tumor phenotypes, which were not normally distributed. We used Pearson’s correlation test to evaluate the correlation (\( r \)) of ACE2 expression levels with the ratios of immune signatures, which was the log2-transformed values of the ratios between the mean expression levels of all marker genes in immune signatures and was normally distributed. We used the Benjamini and Hochberg method [23] to calculate the FDR for adjusting for multiple tests. We compared overall survival (OS), disease-specific survival (DSS), progression-free interval (PFI), and disease-free interval (DFI) between the high- and low-ACE2-expression-level tumors. We utilized Kaplan-Meier curves to display survival time differences and the log-rank test to evaluate the significance of survival time differences. The R package "survival" was used to perform the survival analyses.

**Results**

**Association of ACE2 expression with immune signatures and immunotherapy response in cancer**

GSEA [21] identified many immune-related pathways highly enriched in the high-ACE2-expression-level tumors at least 5 cancer types. These pathways included cytokine-cytokine receptor interaction, hematopoietic cell lineage, viral myocarditis, natural killer cell-mediated cytotoxicity, chemokine signaling, Jak-STAT signaling, primary immunodeficiency, antigen processing and presentation, autoimmune thyroid disease, T cell receptor signaling, intestinal immune network for IgA production, B cell receptor signaling, systemic lupus erythematosus, Leishmania infection, NOD-like receptor signaling, and epithelial cell signaling in Helicobacter pylori infection (Fig. 1A). Moreover, we found that ACE2 expression levels positively correlated with the pro-/anti-inflammatory ratios in pan-cancer (Pearson’s correlation test, \( r = 0.26, p = 3.31 \times 10^{-74} \)) and in 11 individual cancer types (adjusted \( p \)-value (FDR) < 0.05) (Fig. 1B). This suggests that ACE2 expression has a stronger positive association with the pro-inflammatory signature than the anti-inflammatory signature in these cancer types. Altogether, these results suggest a prominent positive association between ACE2 expression and antitumor immune signatures in cancer. We found that ACE2 had a positive expression correlation with PD-L1 in pan-cancer and in 6 individual cancer types (FDR < 0.05) (Fig. 1C). We expected that the ACE2 expression would have a positive association with the response to anti-PD-1/PD-L1/CTLA-4 immunotherapy. We confirmed the anticipation in four cancer cohorts receiving immune checkpoint blockade therapy. In these cohorts, the
high-ACE2-expression-level (> median) tumors displayed a higher rate of immunotherapy response than the low-ACE2-expression-level (< median) tumors (67% versus 17%, 80% versus 40%, 40% versus 20%, and 46% versus 25% for Nathanson (melanoma), Topalian (melanoma), Ascierto (renal cell carcinoma), and Snyder (bladder cancer) cohorts, respectively) (Fig. 1D). As a result, the former had better overall survival (OS) than the latter in the Nathanson cohort, which had related data available (log-rank test, \( p = 0.036 \)) (Fig. 1E). These results suggest that the ACE2 expression is likely to be a positive predictor for anti-PD-1/PD-L1 immunotherapy.

**Association of ACE2 expression with oncogenic pathways and tumor phenotypes in cancer**

We quantified the activity of a pathway using the single-sample gene-set enrichment analysis (ssGSEA) [19] score of the set of genes included in the pathway. We found that ACE2 expression levels inversely correlated with the activity of cell cycle, mismatch repair, TGF-\( \beta \), Wnt, VEGF, and Notch signaling pathways in 9, 6, 9, 7, 6, and 7 individual cancer types, respectively (Spearman's correlation test, FDR < 0.05) (Fig. 2A). Moreover, we found that ACE2 expression levels had a significant inverse correlation with the expression levels of MKI67, which is a tumor proliferation index marker, in pan-cancer and 8 individual cancer types (Pearson's correlation test, FDR < 0.05) (Fig. 2B). Tumor stemness represents a stem cell-like tumor phenotype associated with tumor progression, metastasis, immune evasion, and drug resistance. We found that ACE2 expression levels showed a marked negative correlation with tumor stemness scores (ssGSEA scores) in pan-cancer and in 9 individual cancer types (FDR < 0.05) (Fig. 2C). EMT plays an outstanding role in facilitating malignant transformation, tumor progression, and metastasis. We observed a marked negative correlation between ACE2 expression levels and EMT signature scores (ssGSEA scores) in 11 individual cancer types (FDR < 0.05) (Fig. 2D). Overall, these data indicate that ACE2 is a protective factor for cancer progression. Indeed, survival analyses showed that ACE2 upregulation was associated with favorable survival in pan-cancer (log-rank test, \( p < 0.001 \) for OS, DSS, PFI, and DFI) and in KIRC, KIRP, LUSC, and OV (log-rank test, \( p < 0.05 \) for OS, DSS, PFI, and/or DFI) (Fig. 2E). Furthermore, we found that ACE2 expression levels significantly increased with the tumor advancement in KIRC (two-sided Student's \( t \) test, \( p < 0.01 \), fold change > 1.5 for high-grade (G3-4) versus low-grade (G1-2), late-stage (stage III-IV) versus early-stage (stage I-II), large tumor size (T3-4) versus small tumor size (T1-2), without regional lymph nodes (N0) versus with lymph nodes (N1-3), and no metastasis (M0) versus metastasis (M1)) (Fig. 2F).

**Identifying interaction networks of ACE2 in cancer**

We identified 200 and 24 genes having marked positive and negative expression correlations with ACE2 in pan-cancer, respectively (\(| r | > 0.5 \)) (Fig. 3A). WGCNA [22] identified four gene modules (indicated in yellow, red, pink, and turquoise color, respectively) highly enriched in the high-ACE2-expression-level tumors and three gene modules (indicated in black, blue, and green color, respectively) highly enriched in the low-ACE2-expression-level tumors (Fig. 3B). The GO terms highly enriched in the high-ACE2-expression-level tumors mainly included immune response, induction of bacterial agglutination, regulation of microvillus length, and epidermal cell differentiation. In contrast, the GO terms highly
enriched in the low-ACE2-expression-level tumors mainly included cell cycle, nervous system process, and microtubule-based process (Fig. 3B). Again, these results indicate that ACE2 expression has a significant positive association with antitumor immune response and a significant negative association with the cell cycle in cancer, suggesting the protective role of ACE2 from cancer progression.

From the yellow gene module, we identified 103 hub genes mainly associated with immune-related pathways. Among the 103 hub genes, three transcription factor (TF) genes, including EOMES, IRF4, and TBX21, were co-expressed with many other immune-related genes, such as PDCD1, TIGIT, GZMK, IL21R, and IL2RG (Fig. 3C). The association between these TFs and immune regulation has been well recognized, such as EOMES (Eomesodermin) mediating the CD8+ T cell differentiation [24], IRF4 (interferon regulatory factor 4) regulating immune cell development [25], and TBX21 (T-bet) playing a pivotal role in regulating Th1 cell development [26].

**Discussion**

We investigated the association of ACE2 expression with immune signatures, oncogenic pathways, and tumor phenotypes in diverse cancer cohorts. Our results indicate that ACE2 is a protective factor for cancer progression. In particular, the ACE2 downregulation correlates with worse survival and tumor advancement in KIRC, also known as clear cell renal cell carcinoma (ccRCC). Previous studies demonstrated that ACE2 exerts antitumor effects by inhibiting tumor angiogenesis [10] and promoting tumor immune infiltration [14]. Our results are consistent with these previous findings. Besides, we found that ACE2 upregulation was associated with reduced cell proliferation, stemness, and EMT, as well as the downregulation of oncogenic pathways, such as cell cycle, mismatch repair, TGF-β, Wnt, and Notch signaling. Moreover, we found that ACE2 had a negative expression correlation with PD-L1, an immunosuppressive molecule, and a predictive marker for an active response to immune checkpoint inhibitors. As a result, ACE2 upregulation correlates with a favorable response to anti-PD-1/PD-L1/CTLA-4 immunotherapy.

ACE2 also plays a protective role in hypertension and heart disease [27]. Moreover, ACE2 deficiency may exacerbate outcomes in patients with SARS-CoV-2 infection [27]. Indeed, a recent study showed that ACE2 was downregulated in virus-infected lung tissue [14], indicating a potential protective role of ACE2 in patients with SARS-CoV-2 infection. Thus, using ACE2 inhibitors for preventing and treating SARS-CoV-2 infections may not be an advisable strategy for individuals with hypertension, heart disease, or cancers.

**Conclusions**

ACE2 upregulation was associated with increased antitumor immunity and immunotherapy response, reduced tumor malignancy, and favorable survival in cancer, suggesting that ACE2 is a protective factor for cancer progression. Our data may provide potential clinical implications for treating cancer patients infected with SARS-CoV-2.
Abbreviations

ACE2: angiotensin-converting enzyme 2; DFI: disease-free interval; DSS: disease-specific survival; EMT: epithelial-mesenchymal transition; FDR: false discovery rate; GO: gene ontology; GSEA: gene set enrichment analysis; OS: overall survival; PFI: progression-free interval; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2; TCGA: The Cancer Genome Atlas; TF: transcription factor; WGCNA: weighted gene co-expression network analysis; CESC: cervical squamous-cell carcinoma; COAD: colon adenocarcinoma; ESCA: esophageal carcinoma; HNSC: head and neck squamous cell carcinoma; KIRC: kidney renal clear cell carcinoma; KIRP: kidney renal papillary cell carcinoma; LUAD: lung adenocarcinoma; LUSC: lung squamous cell carcinoma; OV: ovarian carcinoma; SKCM: skin cutaneous melanoma; THYM: thymoma; UCEC: uterine corpus endometrial carcinoma

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data and material

From the genomic data commons data portal (https://portal.gdc.cancer.gov/), we obtained RNA-Seq gene expression profiling datasets (level 3 and RSEM normalized) and clinical data for 12 TCGA cancer cohorts. We performed all the computational and statistical analyses using R programming (https://www.r-project.org/).

Competing interests

The authors declare that they have no competing interests.

Funding

This work was supported by the China Pharmaceutical University (grant numbers 3150120001 to XW).

Authors' contributions
ZZ performed data analyses and helped in manuscript preparation. LL performed data analyses and helped in manuscript preparation. ML performed data analyses and helped in manuscript preparation. XW conceived the study, designed analysis strategies, and wrote the manuscript. All the authors read and approved the final version of the manuscript.

Acknowledgments

Not applicable.

References

1. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu NH, Nitsche A et al: SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. Cell 2020, 181(2):271-280 e278.

2. Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, Lu G, Qiao C, Hu Y, Yuen KY et al: Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2. Cell 2020, 181(4):894-904 e899.

3. Yan R, Zhang Y: Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2. 2020, 367(6485):1444-1448.

4. Lan J, Ge J, Yu J: Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. 2020, 581(7807):215-220.

5. Danilczyk U, Penninger JM: Angiotensin-converting enzyme II in the heart and the kidney. Circ Res 2006, 98(4):463-471.

6. Li MY, Li L, Zhang Y, Wang XS: Expression of the SARS-CoV-2 cell receptor gene ACE2 in a wide variety of human tissues. Infect Dis Poverty 2020, 9(1):45.

7. Dai YJ, Hu F, Li H, Huang HY, Wang DW, Liang Y: A profiling analysis on the receptor ACE2 expression reveals the potential risk of different type of cancers vulnerable to SARS-CoV-2 infection. Ann Transl Med 2020, 8(7):481.

8. Chai P, Yu J, Ge S, Jia R, Fan X: Genetic alteration, RNA expression, and DNA methylation profiling of coronavirus disease 2019 (COVID-19) receptor ACE2 in malignancies: a pan-cancer analysis. J Hematol Oncol 2020, 13(1):43.

9. Winkler T, Ben-David U: Elevated expression of ACE2 in tumor-adjacent normal tissues of cancer patients. Int J Cancer 2020.

10. Zhang Q, Lu S, Li T, Yu L, Zhang Y, Zeng H, Qian X, Bi J, Lin Y: ACE2 inhibits breast cancer angiogenesis via suppressing the VEGFa/VEGFR2/ERK pathway. J Exp Clin Cancer Res 2019, 38(1):173.

11. Feng Y, Wan H, Liu J, Zhang R, Ma Q, Han B, Xiang Y, Che J, Cao H, Fei X et al: The angiotensin-converting enzyme 2 in tumor growth and tumor-associated angiogenesis in non-small cell lung cancer. Oncol Rep 2010, 23(4):941-948.
12. Xu J, Fan J, Wu F, Huang Q, Guo M, Lv Z, Han J, Duan L, Hu G, Chen L et al: The ACE2/Angiotensin-(1-7)/Mas Receptor Axis: Pleiotropic Roles in Cancer. *Front Physiol* 2017, 8:276.

13. Feng Y, Ni L, Wan H, Fan L, Fei X, Ma Q, Gao B, Xiang Y, Che J, Li Q: Overexpression of ACE2 produces antitumor effects via inhibition of angiogenesis and tumor cell invasion in vivo and in vitro. *Oncol Rep* 2011, 26(5):1157-1164.

14. Yang J, Li H, Hu S, Zhou Y: ACE2 correlated with immune infiltration serves as a prognostic biomarker in endometrial carcinoma and renal papillary cell carcinoma: implication for COVID-19. *Aging (Albany NY)* 2020, 12(8):6518-6535.

15. Nathanson T, Ahuja A, Rubinstein A, Aksoy BA, Hellmann MD, Miao D, Van Allen E, Merghoub T, Wolchok JD, Snyder A et al: Somatic Mutations and Neoepitope Homology in Melanomas Treated with CTLA-4 Blockade. *Cancer Immunol Res* 2017, 5(1):84-91.

16. Ascierto ML, Makohon-Moore A, Lipson EJ, Taube JM, McMiller TL, Berger AE, Fan J, Kaunitz GJ, Cottrell TR, Kohutek ZA et al: Transcriptional Mechanisms of Resistance to Anti-PD-1 Therapy. *Clinical cancer research: an official journal of the American Association for Cancer Research* 2017, 23(12):3168-3180.

17. Ascierto ML, McMiller TL, Berger AE, Danilova L, Anders RA, Netto GJ, Xu H, Pritchard TS, Fan J, Cheadle C et al: The Intratumoral Balance between Metabolic and Immunologic Gene Expression Is Associated with Anti-PD-1 Response in Patients with Renal Cell Carcinoma. *Cancer immunology research* 2016, 4(9):726-733.

18. Snyder A, Nathanson T, Funt SA, Ahuja A, Buros Novik J, Hellmann MD, Chang E, Aksoy BA, Al-Ahmadi H, Yusko E et al: Contribution of systemic and somatic factors to clinical response and resistance to PD-L1 blockade in urothelial cancer: An exploratory multi-omic analysis. *PLoS Med* 2017, 14(5):e1002309.

19. Hanzelmann S, Castelo R, Guinney J: GSVA: gene set variation analysis for microarray and RNA-seq data. *BMC Bioinformatics* 2013, 14:7.

20. Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K: KEGG: new perspectives on genomes, pathways, diseases and drugs. *Nucleic acids research* 2017, 45(D1):D353-D361.

21. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A, Pomeroy SL, Golub TR, Lander ES et al: Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 2005, 102(43):15545-15550.

22. Langfelder P, Horvath S: WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics* 2008, 9:559.

23. Benjamini Y, Hochberg Y: Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society B* 1995, 57:289-300.

24. Knox JJ, Cosma GL, Betts MR, McLane LM: Characterization of T-bet and eomes in peripheral human immune cells. *Front Immunol* 2014, 5:217.

25. Nam S, Lim JS: Essential role of interferon regulatory factor 4 (IRF4) in immune cell development. *Arch Pharm Res* 2016, 39(11):1548-1555.
26. Szabo SJ, Kim ST, Costa GL, Zhang X, Fathman CG, Glimcher LH: A novel transcription factor, T-bet, directs Th1 lineage commitment. *Cell* 2000, 100(6):655-669.

27. Verdecchia P, Cavallini C, Spanevello A, Angeli F: The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med* 2020, 76:14-20.

**Figures**
Figure 1
Association of ACE2 expression with immune signatures and immunotherapy response in cancer. A. Immune-related pathways upregulated in high- (upper third) versus low-ACE2-expression-level (bottom third) tumors in at least 5 cancer types identified by GSEA [21] (adjusted p-value (FDR) < 0.05). B. Significant positive correlations of ACE2 expression levels with the ratios of pro-/anti-inflammatory cytokines in pan-cancer and in 11 individual cancer types. The Pearson correlation coefficient (r) and p- or FDR-value are shown. C. The positive expression correlation between ACE2 and PD-L1 in pan-cancer and in 6 individual cancer types. D. Higher rate of immunotherapy response in the high-ACE2-expression-level (> median) than in the low-ACE2-expression-level (< median) tumors in four cancer cohorts receiving immune checkpoint blockade therapy. E. Kaplan-Meier survival curves showing better survival in high-ACE2-expression-level (> median) than in low-ACE2-expression-level (< median) cancer patients with immune checkpoint blockade therapy. The log-rank test p-value is shown. FDR: false discovery rate. * FDR < 0.05; ** FDR < 0.01; *** FDR < 0.001. They also apply to the following figures.
Association of ACE2 expression with oncogenic pathways and tumor phenotypes in cancer. ACE2 expression levels are likely to inversely correlate with the activity of oncogenic pathways (A), MKI67 expression levels (B), stemness scores (C), and EMT signature scores (D) in cancer. Kaplan-Meier survival curves showing that ACE2 upregulation is associated with favorable survival in pan-cancer and multiple individual cancer types. Log-rank test p-values are shown. EMT: epithelial-mesenchymal transition; OS: overall survival; DSS: disease-specific survival; PFI: progression-free interval; DFI: disease-free interval. E. ACE2 expression levels significantly increase with tumor advancement in KIRC. KIRC: kidney renal clear cell carcinoma.
Figure 3

Interaction networks of ACE2 in cancer. A. 200 and 24 genes having marked positive and negative expression correlations with ACE2 in pan-cancer, respectively (|r| > 0.5). B. Gene modules (gene ontology) enriched in high-ACE2-expression-level and low-ACE2-expression-level pan-cancer. C. Co-expression
subnetwork of the immune response module enriched in high-ACE2-expression-level pan-cancer centered on three transcription factor genes (in yellow).

**Supplementary Files**

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- Supplementarytables.xlsx