In 2011, vaccinia virus caused an outbreak of bovine vaccinia, affecting dairy cattle and dairy workers in Brazil. Genetic and phenotypic analyses identified this isolate as distinct from others recently identified, thereby reinforcing the hypothesis that different vaccinia virus strains co-circulate in Brazil.

Throughout most of Brazil, vaccinia virus (VACV), family Poxviridae, is the etiologic agent of bovine vaccinia (1). Outbreaks often occur on unhygienic rural properties and cause mild to severe rashes on teats and udders of dairy cows and various locations on humans (1,2). Dairy workers usually seek medical care for the painful lesions, but rarely are they hospitalized. Some studies suggest an association between these outbreaks of bovine vaccinia and the VACV strains used during the World Health Organization smallpox eradication campaign (3). Since 1999, VACV strains in Brazil have been investigated (2–8); biological and molecular approaches indicated 2 distinct groups of these viruses (9,10). In 2011, a bovine vaccinia outbreak occurred in Serro County, Minas Gerais state, in southeastern Brazil, one of the largest milk-producing regions in Brazil. The outbreak affected 91 dairy cows and 3 dairy workers, 1 of whom was hospitalized (online Technical Appendix Figure 1, panel A, wwwnc.cdc.gov/EID/pdfs/12-0145.pdf). Our aim was to elucidate the genetic and phenotypic characteristics of this VACV isolate.

The Study

The outbreak affected 2 farms, 91 cows, and 3 humans (Figure 1). On day 1, on farm 1, the index case was a sick cow with ulcerative lesions on the teats and udder. On day 3, the owner of farm 1 (patient A), who had had direct contact with the sick cow, noticed lesions on his hand. On day 5, another 6 cows on farm 1 became sick, and patient A went to farm 2 and handled healthy cattle. On day 7, all 14 cows on farm 1 were sick. On day 9, some cows on farm 2 became sick. On day 12, the owner of farm 2 (patient B) and his employee (patient C) became sick. On day 15, patient C was hospitalized with high fever, lymphadenopathy, prostration, and painful vesicular–pustular lesions on his hands and arms. He received clinical support and remained hospitalized for 10 days. He had no immunologic disorders and took no medications that could be associated with his severe clinical condition. According our investigation, only those 3 patients had direct contact with the infected cattle during the outbreak.

By day 18, all 77 cows on farm 2 were sick. On day 24, veterinary surveillance teams isolated these farms for 8 days. On day 26, patient C returned to work although lesions remained on his hands and arms (online Technical Appendix Figure 1, panel B). By day 28, all cattle were recovering.

To identify the etiologic agent responsible for the outbreak, on day 27 we collected swab samples from lesions of patients B and C (not from patient A, whose lesions were healing) and from 1 infected cow (from farm 2). Samples were placed in Vero cells for virus isolation as described (2) and then purified in a sucrose gradient (11). The isolates from the patients B and C and the cow were named VACV Serro human 1/2011 (SH1V/2011), VACV Serro human 2/2011 (SH2V/2011), and VACV Serro bovine 1/2011 (SB1V/2011), respectively.

The isolates were examined by PCR for the A56R gene (hemagglutinin [HA]), and the fragments obtained (950 bp) were sequenced and analyzed as described (2,12,13). The nucleotide sequences showed 100% identity among...
all VACV Serro 2011 isolates and exhibited high identity with VACV strains from Brazil, particularly SPAn232 (99.8% identity) and GuaraniP1 (99.5% identity) viruses. The HA phylogenetic tree clustered most VACV isolates from Brazil together, mainly because of the presence of the deletion signature (group 1). However, VACV Serro 2011 isolates did not exhibit this signature and instead clustered with VACV strains that are less frequently isolated during outbreaks in Brazil (group 2) (Figure 2, panel A).

To further characterize the virus, we also sequenced the A26L gene (ati) (14). Because HA sequences were identical for SH1V/2011, SH2V/2011, and SB1V/2011 (and hypothetically represent the same isolate), we selected SH2V/2011 to analyze for ati and virulence in BALB/c mice. The ati gene is highly polymorphic in VACV strains from Brazil, and some strains exhibit a large deletion in the ati gene (15). Therefore, we used ati to characterize the VACV strains from Brazil. A 1,600-bp fragment that also did not contain the deletion was amplified from SH2V/2011. This virus exhibits no deletion in either the HA or ati genes in the analyzed regions of these genes. The ati phylogenetic tree (Figure 2, panel B) showed that SH2V/2011 also clustered with SPAn232 virus (group 2) and was segregated from the other group 1 VACV strains.

Figure 2. Phylogenetic analysis of vaccinia virus (VACV) isolates. A) Phylogenetic tree based on the nucleotide sequence of the orthopoxvirus hemagglutinin gene. VACV Serro bovine (SB1V), VACV Serro human 1 and 2 (SH1V and SH2V) grouped with VACV group 2 isolates, far from other VACV group 1 members. These isolates grouped far from (outliers) Serro-2 virus, a VACV isolated in the same geographic region. B) Phylogenetic tree based on the nucleotide sequence of the orthopoxvirus ati gene. The hemagglutinin tree shows SH2V grouping with VACV Western Reserve (WR) and SPAn232 virus, members of VACV group 2, not close to other isolates from Brazil. The neighbor-joining method with the Tamura-Nei model of nucleotide substitutions in the MEGA4 software program (www.megasoftware.net/) was used. Bootstrap confidence intervals are shown on branches (1,000 replicates) with GenBank accession numbers. MPXV, monkeypox virus; CPXV, cowpox virus; HSPV, horsepox virus; VARV, variola virus. Black dots indicate samples isolated during the 2011 outbreak of bovine vaccinia in Brazil. Scale bars indicate nucleotide substitutions per site.
nonvirulent controls, respectively. Another group was inoculated with phosphate-buffered saline. The SH2V/2011 sample was highly virulent in BALB/c mice; morbidity rate was high (online Technical Appendix Figure 2), thereby supporting the grouping of this sample in the virulent cluster. The animals that were infected with SH2V/2011 exhibited ruffled fur, arched backs, and weight loss, much like those infected with VACV-GuaraniP1. No clinical signs were observed in mice that had been inoculated with either VACV-GuaraniP2 or phosphate-buffered saline.

To further characterize the virus, we also performed a plaque phenotype assay in BSC-40 cell cultures. The VACV strains from Brazil that exhibited virulence in a BALB/c model usually formed large plaques in BSC-40 cell cultures. This assay showed that, in contrast to VACV-GuaraniP1, in BSC-40 cell cultures, SH2V/2011 induces the formation of large plaques that are similar to those induced by VACV-Western Reserve and VACV-GuaraniP1 (online Technical Appendix Figure 3).

Conclusions
Our results indicate that VACV Serro-2011 is a new mouse-virulent VACV strain associated with an outbreak that affected cows and humans. Recently, several VACV strains have been isolated in Brazil, most exhibiting a signature deletion in A56R, few or no deletions in the ati gene, and low virulence in mouse models. In contrast, the VACV isolated from the outbreak reported here, affecting cattle and humans, exhibited virulence in mice but no deletions in either the A56R or ati genes. This strain is genetically and phenotypically distinct from the Serro-2 strain that was isolated from the same region in 2005 (7); thus, >1 VACV might be circulating in Serro and possibly other regions of Brazil (7,8).

During outbreaks of bovine vaccinia, hospitalization of humans, especially for several days, is unusual. Unfortunately, despite the development of in vivo models for study and differentiation of VACV from Brazil, no clear association between viral genetics and disease severity in humans and cattle has been shown. Although preliminary, the data presented here indicate a possible association between these 2 factors, considering the hospitalization of patient C to be long term. Although the surveillance and characterization of VACV have advanced in recent years, observation and description of any clinical characteristics of infected humans and cattle are still helpful. Such observations might be associated with genotypic and phenotypic features of VACV, which could influence surveillance and control strategies for the management of VACV outbreaks.

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Group 2 Vaccinia Virus, Brazil

Technical Appendix

Technical Appendix Figure 1. A) Location of 2011 vaccinia virus outbreak in Serro County, a dairy production region of Minas Gerais state, southeastern Brazil. Square in inset, Minas Gerais state; circle on main map, Serro region. B) Ulcerative lesion on dairy worker’s hand (patient C).

Technical Appendix Figure 2. Clinical signs of BALB/c mice infected by intranasal inoculation with $10^6$ PFU/10 μL vaccinia virus (VACV). A) Median weight loss relative to initial weight. Inoculation with VACV
GuaraniP1 virus (GP1V) and VACV Serro human 2/2011 (SH2V) showed similar weight loss (67.5% and 66.3% respectively). B) Control mouse inoculated with VACV GuaraniP2 (GP2V) virus. No clinical signals were observed in control mice inoculated with phosphate-buffered saline (NC-PBS) and GP2V. C) Mouse inoculated with SH2V, showing clinical signs. Severe clinical signs, such as weight loss, ruffled fur, arched back, closed eyes with inflammation, and facial edema were observed in mice inoculated with VACV GuaraniP1 and SH2V.

Technical Appendix Figure 3. Results of phenotypic plaque-forming testing of vaccinia viruses (VACVs) on epithelial kidney cells BSC-40. A) VACV GuaraniP1 virus; B) VACV GuaraniP2 virus; C) VACV Serro human; D) VACV Serro bovine; E) VACV Western Reserve. Viruses in panels C and D show similar large cytopathic effects, like VACV GuaraniP1 virus, members of VACV group 2, and VACV Western Reserve. VACV virus shows smaller plaques on cell culture, like other VACV group 1 viruses.