Early-light embryonic stimulation suggests a second route, via gene activation, to cerebral lateralization in vertebrates

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Genetic factors determine the asymmetrical position of vertebrate embryos allowing asymmetric environmental stimulation to shape cerebral lateralization. In birds, late-light stimulation, just before hatching, on the right optic nerve triggers anatomical and functional cerebral asymmetries. However, some brain asymmetries develop in absence of embryonic light stimulation. Furthermore, early-light action affects lateralization in the transparent zebrafish embryos before their visual system is functional. Here we investigated whether another pathway intervenes in establishing brain specialization. We exposed chicks' embryos to light before their visual system was formed. We observed that such early stimulation modulates cerebral lateralization in a comparable vein of late-light stimulation on active retinal cells. Our results show that, in a higher vertebrate brain, a second route, likely affecting the genetic expression of photosensitive regions, acts before the development of a functional visual system. More than one sensitive period seems thus available to light stimulation to trigger brain lateralization.

A symmetry along the left-right axis is a feature common to all vertebrates. Heart and liver are placed to the left and right side respectively1–4 and even paired-symmetric organs display some degree of asymmetry (e.g. lungs differ in the number of lobes). The brain exhibits profound anatomical and functional asymmetries (review7). How anatomical asymmetry is imposed on a seemingly bilaterally symmetric structure, the vertebrate neural tube, is however still largely obscure. Selective expression of the transforming growth factor (TGF) family member Nodal, a signal transduction pathway, on the left side of the early embryo seems to mediate the asymmetrical morphogenesis and placement of the internal organs through activation of a signaling cascade6,7. Whilst such a Nodal cascade controls ventral forebrain development, its effect on lateralization is on epiphyseal gene expression in the dorsal forebrain (e.g. in zebrafish, the asymmetry of the diencephalic habenular nuclei and the photoreceptive pineal complex8,9).

Lateralization mediated by Nodal cascade seems to operate in the brain, triggered by asymmetric sensory stimulation in embryo. The processes underlying the asymmetric morphology and positioning of the viscera are accompanied by a slight torsion of the embryo whose forehead points to the right10. Such a rightward spinal torsion seems to occur in all amniotes11, including human embryos, which also display a right-turn of their head12. Asymmetric turning associated with Nodal signals may set the stage for either direct asymmetrical sensory stimulation of the embryo (because of its placement in utero or in ovo13,14) or by constrained motor patterns that in turn may promote asymmetrical stimulation (for instance, a slight preference to move the right arm because of turning of the embryo can then be enhanced by an increased eye-hand contact on the right side15).

This has been investigated in detail in the avian brain. During incubation, the birds’ embryos bend so that the head is asymmetrically tilted with the right eye placed below the egg surface and the left eye leant below the wing16. In this position, environmental light penetrating the eggshell acts on the retina of the right eye only, producing structural asymmetries on the ascending visual projections and thus modulating functional cerebral specialization13,17–19. Environmental illumination during latest stages of embryonic development is crucial in the determination of brain asymmetries through the selective action on the fully-formed visual receptors20: domestic chicks hatched from dark incubated eggs lack any asymmetry in the visual pathways21,22. Furthermore, swapping the exposed eye by withdrawing the embryo’s head from the egg (i.e., making the left rather than the right eye to...
receive illumination) reverses the pattern of asymmetry in both chicks\textsuperscript{22,23} and pigeons\textsuperscript{24,25}. Environmental illumination affects also functions of the left (unstimulated) eye and associated contralateral brain structures (as shown in attack, copulation and detection of predator\textsuperscript{26} and visuo-spatial abilities\textsuperscript{27}) by modifying inter-hemispheric cross-talk\textsuperscript{28,29}.

Despite light being such a strong environmental triggering factor, some asymmetries in birds are unaffected by embryonic light exposure. For instance, uni-hemispheric sleep patterns\textsuperscript{30}, lateralized mechanisms of social recognition\textsuperscript{31}, and the neural mechanisms underlying imprinting\textsuperscript{32,33} are apparent also in dark-incubated birds. Besides, work on zebrafish has shown only partial correspondence between the reversal of the visceral situs and diencephalic asymmetries and the reversal of lateralized behaviours\textsuperscript{34}. Similarly, in rare cases of spontaneous situs inversion in humans only some of the brain and behavioural lateralities change the direction of their asymmetry (e.g. language dominance continued to be a feature of the left hemisphere, i.e., not reversed\textsuperscript{35,36}). All this is suggestive of multiple genetic/environmental routes to brain lateralization.

In zebrafish environmental illumination applied early in development (at day one after fertilization) is needed to generate left/right cerebral asymmetries. After light stimulation, the left eye shows more interest in motivating stimuli\textsuperscript{37–38}. At this early stage of development, a motor (at day one after fertilization) is needed to generate left/right cerebral asymmetries and the reversal of lateralized behaviours\textsuperscript{39}. Similarly, in rare cases of spontaneous situs inversion in humans only some of the brain and behavioural lateralities change the direction of their asymmetry (e.g. language dominance continued to be a feature of the left hemisphere, i.e., not reversed\textsuperscript{35,36}). All this is suggestive of multiple genetic/environmental routes to brain lateralization.

Here we show for the first time that early environmental illumination provided to chicks’ embryo at an age in which light cannot exert any effect on fully-formed retinal receptors which seem not to be in place\textsuperscript{40} may nonetheless cause cerebral lateralization, likely involving the second developmental pathway, i.e., by affecting the genetic expression of undifferentiated cells of photosensitive regions.

Results

In Experiment 1, chicks incubated in darkness (Di-chicks), exposed to light during the first 3 days after fertilization (EarlyLi-chicks) and the last 3 days before hatching (LateLi-chicks) were free to peck at food grains, scattered in an array of identical vertical sectors: a central one, 8 left and 8 right sectors (Figure 1). The total amount of pecks in each sector over a 3 minutes period was scored for each chick. In a repeated measures ANOVA, Hatch (Di-, EarlyLi- and LateLi-chicks) Side (Left and Right) and Distance (1 to 8 sectors) were analyzed as factors. The ANOVA showed a significant effect of Hatch (F\textsubscript{(2,85)} = 4.046, P = 0.021) and a significant effect of Distance (F\textsubscript{(7,595)} = 1093.597, P < 0.001) with decreased pecking with increasing distance from the center; there was also a significant main effect of Side (F\textsubscript{(1,85)} = 10.657, P = 0.002). The interaction between Side and Hatch was significant (F\textsubscript{(2,85)} = 4.841, P = 0.010) with Di-chicks choosing equally for the left and the right side of the grid (Di- toward left: 2.353 ± 0.128; toward right: 2.427 ± 0.156, t\textsubscript{(28)} = 0.533, P = 0.598, Paired Samples t-Test) and both EarlyLi- and LateLi-chicks preferring significantly the left side (respectively, EarlyLi-: toward left: 2.531 ± 0.133; toward right: 2.165 ± 0.133, t\textsubscript{(27)} = −3.731, P = 0.001; LateLi-: toward left: 2.907 ± 0.067; toward right: 2.573 ± 0.128; t\textsuperscript{(30)} = −3.599, P = 0.001) as shown in Figure 2. The interaction between Distance and Hatch was significant (F\textsubscript{(14,595)} = 3.322, P < 0.001), as well as the interaction between Side and Distance (F\textsubscript{(7,595)} = 4.173, P < 0.001).

In Experiment 2, Di-, EarlyLi- and LateLi-chicks were left free to run from a starting box to a feeder located on the opposite ends of a long runway (Figure 3(a)). The routes covered by the chicks were scored with a video analysis software (Videopoint\textsuperscript{®}) and then analyzed with Matlab\textsuperscript{®} to determine the direction (left vs. right) of each route. We estimated the area (expressed in m\textsuperscript{2}) between the real route covered by the chick and the optimal route both for leftward and rightward trajectories as visible in Figure 3(b). A repeated measures ANOVA with Hatch (Di-, EarlyLi- and LateLi-chicks) and Route Direction (Left vs. Right) as factors showed no difference across hatching conditions (F\textsubscript{(2,58)} = 1.115, P = 0.335). By contrast, the main effect Route Direction was significant (F\textsubscript{(1,58)} = 153.913, P < 0.001) with all animals running more to the left than to the right (respectively, 0.049 m\textsuperscript{2} ± 0.006 vs. 0.016 m\textsuperscript{2} ± 0.004; t\textsubscript{(60)} = 12.514, P < 0.001, Paired Samples t-Test). When an obstacle was inserted in the center of the runway (along the midline of the optimal straight route as shown in Figure 4(a)) and the chick was allowed to reach the target 8 consecutive times, an ANOVA with Hatch as between-subject factor and Detour Direction as dependent variable revealed a significant heterogeneity between groups (F\textsubscript{(2,54)} = 3.626, P = 0.033). Di-chicks chose to detour the obstacle leftward (t\textsubscript{(19)} = 3.644, P = 0.002, One-Sample t-Test), whereas EarlyLi- and LateLi-chicks showed no systematic preference for a direction (respectively, t\textsubscript{(17)} = −0.676, P = 0.508; t\textsubscript{(18)} = 0.468, P = 0.645) as shown in Figure 4(b).

Discussion

Chicks hatched from eggs exposed to ambient illumination for three days either at an early or a late stage of embryonic development performed in a comparable way in two different tasks. In the first experiment, both EarlyLi- and LateLi-chicks showed a leftward bias when allowed to peck at crumbs scattered on a grid in front of them. Chicks maintained in dark, in contrast, directed their pecks uniformly toward the left and the right hemi-spaces. The result that LateLi-chicks have a left bias in attending the target position whereas Di-chicks show no systematic asymmetrical preference confirms
Figure 2 | Means (+ s.e.m.) pecks at grains made by Di-chicks (top), EarlyLi-chicks (middle) and LateLi-chicks (bottom) at the cancellation grid toward left (L) and right (R) from the central midline (C) in 3 minutes of activity. Lighter parts indicate the greater amount of pecks toward left. Single grains of food and the chick’s head viewed from above are superimposed in post-editing to the graph for representational purposes only.
Such a late light action is well-known and it is thought to affect brain structures by a retinal route, stimulating photoreceptors of the right eye during a critical period (last three days before hatching) in which retinal ganglion cells start to be active.

The result that an early light stimulation may induce a comparable behavioural lateralization provides the first evidence that illumination also play a role on structures of the nervous system that are not the fully-formed eye.

In the second experiment, we found again a striking similarity of the effects of the early and the late exposure to light. Both EarlyLi- and LateLi-chicks showed no bias when they had to detour an obstacle in order to reach a target, moving around either to the left or the right equally. On the contrary, Di-chicks showed a bias, choosing to detour the obstacle significantly more often to the left. Note that both chicks stimulated by light and chicks incubated in the dark run in a comparable way when they had only to reach for a target, meaning that there were no differences in motor behaviour depending on the different incubation conditions. Rather, a difference in control of attention may explain the results. Both EarlyLi- and LateLi-chicks, having decided on approach to the target, are able to ignore the obstacle altogether, whereas Di-chicks have to actively sustain approach to the target by using the right eye to view the obstacle. Di-chicks seem less able in sustaining attention than the groups of chicks exposed to light much as light chicks use the right eye in initial selection of target. There is evidence that many vertebrates view potential danger with the right eye in order to sustain examination and assessment (reviews).

Here we showed that two different lateralized behaviours are affected by environmental illumination: both the early and the late stimulation seem to affect the lateralized behaviours in the same direction. Despite the observed effects involve visuo-spatial behaviours mediated primarily by visual processing, functions like sustained attention or inhibition of responses are also crucial in performing these tasks. These functions may be differently modulated by asymmetries of other brain regions than the visual pathways on which the late light stimulation operates. The responsible mechanism of the early stimulation needs to be investigated, but we hypothesize that one route to the development of cerebral asymmetries may involve the genetic expression of photosensitive regions.

The early photosensitivity in the epiphyseal area of zebrafish has been proposed as responsible for this early action of light, via gene activation, in cells which are not specialized retinal photoreceptors. It has been demonstrated that these photosensitive cells respond to light very early in fish embryonic development, well before retinal photoreceptors. To our knowledge it is not known whether the epiphyseal photoreceptors are active during the first 3 days of embryonic development in the chick, though it can be reasonably assumed that a similar developmental pattern may be shared across vertebrates. However, it is not possible to exclude that other photosensitive molecules in the developing telencephalon and diencephalon in early stages of development may be directly involved.

We know that during the time-window in which we applied the early-light, the chick embryo still lies in a symmetric position within the egg but asymmetric processes are likely to be already at play.
Kuan and collaborators\textsuperscript{3} report that lateralized Nodal signaling influences the directional asymmetry of the parapineal which in turn mediates habenular asymmetry in fish. The asymmetry of early action of light may be due to asymmetry of gene expression, presumably in the area where the parapineal develops and where photoreceptors start to be active early. A comparable cascade may be at work in the chick brain, possibly involving different areas outside the habenula. Indeed, in this species the epiphysis is forming as a separate knob at 60 hours after fertilization\textsuperscript{4}. The epiphysial area could be the target substrate for the early illumination to trigger visuospatial asymmetry comparable to the late activity of light on the optic nerve.

Although this explanation has to be taken cautiously, it is clear that structures other than the eye may be modulated by light exposure. Evolution may have provided organisms with more than one sensitive temporal window as a chance for the light to exert its important ecological role in triggering brain asymmetry.

\begin{itemize}
  \item \textbf{Methods.} The study was carried out in compliance with the European Community and the Italian national animal experiments by the Ministry of Health, under the authorization of the Ethical Committee of the University of Trieste (protocol number 385 pos II/9 dd 16.03.2012).
  \item \textbf{Subjects.} Two hundred and six Hybro (White Leghorn) chicks (\textit{Gallus gallus}) hatched in our laboratory under controlled conditions were used. The eggs were obtained from a local commercial hatchery immediately after fertilization; thereafter, some eggs (n = 70) were kept in complete darkness until the hatching day (Di-chicks) in an incubator FIEM snc, MG 100 H (45 cm wide \times 58 cm high \times 43 cm deep), under controlled temperature (36.7°C) and humidity (about 50–60%) conditions; some eggs (n = 66) were exposed to light from day 1 to day 3 (nearly 70 hours) after fertilization (EarlyLi-chicks) and thereafter remained in the dark; some others (n = 70) were maintained in darkness and exposed to light from day 18 of incubation (LateLi-chicks) to day 21 of hatching. A 60 W incandescent light bulb provided light within of the incubator. Immediately after hatching, chicks were reared singly in metal cages (22 cm wide \times 30 cm high \times 40 cm deep) illuminated by 30 W fluorescent lights (12 L: 12 D cycle) and located in a separate room at 28–30°C.
  \item \textbf{Apparatus.} The apparatus was the same used in previous experiments\textsuperscript{5,6} and consisted of a white uniform wooden enclosure (50 cm wide \times 58 cm high \times 43 cm deep) with a brown ground. A white cardboard box (14 cm wide \times 14 cm high \times 14 cm deep) served as confining box; it was fixed at the middle of the rear wall and presented on its frontal wall a circular window measuring 2 cm in diameter. A Poliplack\textsuperscript{7} rear wall and presented on its frontal wall a circular window measuring 2 cm in diameter. A Poliplack\textsuperscript{7} (17 columns of 10 compartments each) containing a single grain of chicks (1 cm (17 columns of 10 compartments each) containing a single grain of chicks) 1 cm (17 columns of 10 compartments each) containing a single grain of chicks. A Poliplack\textsuperscript{7} indicated the presence of a small yellow and green rectangular plastic A Poliplack\textsuperscript{7} (12 L: 12 D cycle) and located in a separate room at 28–30°C. A Poliplack\textsuperscript{7} and the spatial substrates provided by two lamps of 25 W placed exactly on the top of the starting box and of the landmark. The behavior was videorecorded by a Panasonic\textsuperscript{8} NV-GS27 camera and scored offline. In order to keep track of the chick’s movements within the apparatus, a black removable piece of paper was temporarily attached on the chick’s back.
  \item \textbf{Procedure.} On day 3 of age, after 3 hours of food deprivation, all the chicks were first positioned inside the starting box and left free to move around in order to get acquainted with the novel environment by reaching the target and finding mealworm larvae (\textit{Tenebrio molitor}). Chicks did this quite spontaneously since the landmark was conspicuous and caught their interest. The habituation phase lasted in average 30 minutes: each chick was placed in the starting box and left free to walk toward the landmark to find the reward. The next day, after overnight of food deprivation, each chick was in turn placed within the apparatus in the starting box and left free to run toward the landmark once. No food was available in the feeder. The trial started as soon as the chick came out of the starting box and ended when the chick rested at the feeder. The positions of the starting box and the target were counterbalanced between subjects.
  \item \textbf{Procedure.} The same procedure was used for the animals that underwent the running task in previous experiments\textsuperscript{9} placed in the obstacle placed in the centre of the apparatus (this time 80 cm wide). The obstacle consisted of 2 identical black plastic cylinders (2 cm in diameter and 30 cm high), spaced 2 cm from one another and blocked by a plastic bar positioned just underneath the sawdust. The proportion of the left routes over the total 8 routes was calculated to determine detour direction.
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Author contributions

C.C. and G.V. wrote the manuscript text. C.C., G.V. and R.J.A. conceived the experiments. C.C. and J.G. performed the experiments, analyzed the data and prepared the figures. C.C., R.J.A. and G.V. reviewed the manuscript.

Additional information

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