The Methanol Dehydrogenase Gene, *mxaF*, as a Functional and Phylogenetic Marker for Proteobacterial Methanotrophs in Natural Environments

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Abstract

The *mxaF* gene, coding for the large (α) subunit of methanol dehydrogenase, is highly conserved among distantly related methylotrophic species in the Alpha-, Beta- and Gammaproteobacteria. It is ubiquitous in methanotrophs, in contrast to other methanotroph-specific genes such as the *pmoA* and *mmoX* genes, which are absent in some methanotrophic proteobacterial genera. This study examined the potential for using the *mxaF* gene as a functional and phylogenetic marker for methanotrophs. *mxaF* and 16S rRNA gene phylogenies were constructed based on over 100 database sequences of known proteobacterial methanotrophs and other methylotrophs to assess their evolutionary histories. Topology tests revealed that *mxaF* and 16S rDNA genes of methanotrophs do not show congruent evolutionary histories, with incongruencies in methanotrophic taxa in the Methylococcaceae, Methylocystaceae, and Beijerinckiaceae. However, known methanotrophs generally formed coherent clades based on *mxaF* gene sequences, allowing for phylogenetic discrimination of major taxa. This feature highlights the *mxaF* gene’s usefulness as a biomarker in studying the molecular diversity of proteobacterial methanotrophs in nature. To verify this, PCR-directed assays targeting this gene were used to detect novel methanotrophs from diverse environments including soil, peatland, hydrothermal vent mussel tissues, and methanotroph isolates. The placement of the majority of environmental *mxaF* gene sequences in distinct methanotroph-specific clades (Methylocystaceae and Methylococcaceae) detected in this study supports the use of *mxaF* as a biomarker for methanotrophic proteobacteria.

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Introduction

Atmospheric methane contributes to ~20% of the total radiative forcing by long-lived greenhouse gases globally. Due to the relatively short lifetime of atmospheric methane (~9 yrs) compared to CO2, reductions in atmospheric methane flux would have an immediate impact on global warming [1]. Microbial methane oxidation is the only major biological sink of methane [2]. With the exception of members of the phylum Verrucomicrobia [3], and the yet to be cultured anaerobic methanotrophs of the ‘NC10’ phylum, aerobic proteobacterial methanotrophs are the only known proteobacterial methanotrophs and other methylotrophs to assess their evolutionary histories. The similarities in physiological characteristics, and the generally highly conserved nature of the 16S rRNA and functional gene sequences of proteobacterial methanotrophs, have enabled the use of targeted PCR primers for gene amplification to describe methanotroph diversity [15,16]. However, the 16S rRNA gene is non-protein coding (i.e., not linked directly to methanotroph physiology), and therefore does not directly determine function. For example, it cannot be determined whether 16S rDNA environmental sequences placed close to, but outside of, known other C1 compounds (e.g., methanol, methylated amines, formate, or formamide) [9,10]. The vast majority of known aerobic methanotrophs belong to the Proteobacteria. Known exceptions include the phylum Verrucomicrobia, whose pathways for methanotrophy are still poorly understood [3,11,12], and enrichments of the uncultured methanotrophs such as *Candidatus Methylobacterium oxyfera* affiliated with the ‘NC10’ phylum, which appear to be incapable of oxidizing methane at low (~3%) oxygen levels in the laboratory [7,8,13]. Proteobacterial methanotrophs are placed in the families Methyloccaceae and Beijerinckiaceae in the Alphaproteobacteria [14,15].

The similarities in physiological characteristics, and the generally highly conserved nature of the 16S rRNA and functional gene sequences of proteobacterial methanotrophs, have enabled the use of targeted PCR primers for gene amplification to describe methanotroph diversity [15,16]. However, the 16S rRNA gene is non-protein coding (i.e., not linked directly to methanotroph physiology), and therefore does not directly determine function. For example, it cannot be determined whether 16S rDNA environmental sequences placed close to, but outside of, known

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monophyletic methanotroph clades are indeed methanotrophic [17,19]. In comparison, functional genes for enzymes found in methanotroph metabolic pathways are directly related to physiology [19]. The initial step of methane oxidation to methanol by proteobacterial methanotrophs is mediated by particulate and/or soluble methane monooxygenases (MMOs) [15,20]. Genes in this step, such as pmoA [encoding a subunit of the particulate methane monooxygenase (pMMO)] and mmox [encoding a subunit of the soluble methane monooxygenase (sMMO)] have been used for describing methanotroph diversity [17]. However, the pmoA gene is not present in the genera Methylocella and Methyloferax [21,22,23], and the mmox is present only in a few strains of methanotrophs [15,24]. Hence, the use of both pmoA and mmox genes in PCR-based studies results in the underestimation of methanotroph diversity and suggests that an alternative is needed.

The mxaF gene was first proposed as a functional gene probe for methyloths by McDonald and Marrelli [19]. Methane oxidation gene cluster Δ(mxaF) encodes the α-subunit of the enzyme methanol dehydrogenase (MDH), an enzyme containing a pyrroloquinoline quinone (PQQ) cofactor that oxidizes methanol to formaldehyde in the second step of the methane oxidation pathway, following oxidation of methane to methanol [20,25,26,27,28]. Methanol dehydrogenase is common to all methanotrophs in the Proteobacteria [15,19], but is absent in members of the phylum Verrucomicrobia, which possess a homolog of the mxaF gene, called xoxF gene, a gene of unknown function [29,30,31,32]. Sequences of the mxaF gene are highly conserved in methylotrophic species in the Alpha-, Beta- and Gamma-proteobacteria, reflecting 16S rRNA phylogeny [19,32,33]. In studies thus far based on the mxaF gene, a limited number of representative methanotrophs (<15 taxa) have been placed in generally distinct and coherent clades separate from the other methylotrophs [15,19,24,32]. These studies indicate the mxaF gene can be a useful phylogenetic marker for the classification of methanotrophs.

However, the extent to which mxaF gene sequences of all known proteobacterial methanotrophs can be placed in separate distinct phylogenetic clades has yet to be determined. This may affect its accuracy as a reliable functional biomarker and potentially lead to incorrect inferences when determining phylogenetic relationships, notably if horizontal gene transfer has occurred across these taxa. To date, no extensive phylogenetic comparisons of proteobacterial methylotroph (including methanotroph) mxaF gene sequences have been conducted. In the only study of mxaF phylogeny of the family Methylocystaceae (genera Methylosinus and Methylocystis), representative mxaF genes sequences from this family clustered within a distinct clade, but did not separate according to genus-specific subclades, suggesting that horizontal transfer of this gene may have occurred across this family. However the study did not indicate the extent of horizontal gene transfer in other proteobacterial methanotrophs, and which taxa were involved [17]. Previous studies of mxaF revealed that it is related to quinone alcohol dehydrogenases (ADHs), which utilize a variety of primary and secondary alcohols, but not methanol, as substrates [34], and that mxaF is related to xoxF [32].

Here, we examined whether the mxaF and 16S rRNA gene phylogenies of methanotrophs from the GenBank [35] database reflect congruent evolutionary histories. Though Candidatus M. oxyfera (and members of NC10 phylum) possesses an mxaF-like gene, it is placed outside of known proteobacterial mxaF gene clusters (data not shown). Candidatus M. oxyfera’s genome lacks any known PQQ-biosynthesis pathways and thus may not be able to oxidize methanol on its own [8]. Hence it was not included in our analyses.

This study addresses the following questions, (a) Does the extensive mxaF gene phylogeny discriminate between methanotrophic and other methylotrophic sequences available in genetic databases? (b) Are the mxaF and 16S rRNA gene phylogenies congruent for methanotrophs in the families Methylococaceae, Methylococaceae, and Beijerinckiaceae? (c) Can we tell whether mxaF gene retrieved from the environment belong to methanotrophs or methylotrophs? These questions are addressed using phylogenetic analyses involving sequences from published databases, as well as mxaF sequences obtained in this study from surveys of diverse environments (forest soils, peat and Sphagnum moss, and symbiont-hosting mussel gills) and from methanotroph cultures. Overall, these analyses advance the study of methanotroph diversity by showing that the mxaF gene consistently places methanotroph sequences in resolved phylogenetic clusters for all known members of the families Methylococaceae and Methylococaceae, and has the potential to elucidate the roles methanotrophs play in natural environments.

**Materials and Methods**

**Cultures**

Two mxaF gene sequences were determined in this study for cultured species of the Methylococaceae, *Methylophilus rubra* and *Methylococcus palustre*, provided by J. Semrau and R. Knowles, respectively. Positive control cultures used in this study included two members of the Methylococaceae and a member of the Methylococcaceae for which mxaF gene sequences are already available: *Methylosinus trichosporium* OB3b, *Methylcystis parvis* OBPP, *Methylococcus album* BG8, respectively.

**Collections and Site Descriptions**

Samples for DNA extraction or bacterial isolation were collected from four distinct habitats for DNA extraction or bacterial isolation: (a) Four soil samples, weighing ~15 g, from the organic horizon of long-term nitrogen-amended and control pine and hardwood forest soils of the Harvard Forest Long-Term Ecological Research (LTER) site, previously shown to oxidize methane at atmospheric levels, in Petersham, Massachusetts (42°30′N, 72°10′W) in 2004 [18,36], (b) *Sphagnum recurvum* peat and peat from Crystal Bog (Vilas county, Wisconsin), a 7 ha poor fen enclosing a 2.5 m deep, 0.54 ha, dystrophic lake on the North Temperate Lakes LTER site in 2005 (46°00′30″N 89°36′30″W) [37], (c) two species of methanotroph-hosting mussels (n = 3 per site) from the Mid-Atlantic Ridge (MAR) deep-sea vent sites sampled in 2003 using DSV Alvin: Bathymodiolus azoricus from Lucky Strike (LS; 37°17′N, 32°16′W; 1693 m deep) and Rainbow (RB; 36°13′N, 33°54′W, 2300 m deep), and Bathymodiolus puteoserpentis from Logatchev (LO; 14°45′N, 44°58′W; 3027 m deep) [38,39], and (d) the Halls Brook Holding Area (HBHA), an artificial lake in the Aberjona Watershed, sampled in 2004, near Boston, MA, which becomes stratified during the summer months, whereby the bottom depths become anoxyic [40]. No specific permissions were required for collecting samples in these locations because samples did not involve endangered or protected species, Harvard Forest is owned by Harvard University and permission is granted to research employees, and Crystal Bog and Halls Brook Holding Area are located on public land.

**Methanotroph Isolation, DNA Extraction, and Purification**

Methanotrophic HBHA isolate 1 and HBHA isolate 2 were isolated from water collected at the oxic-anoxic interface at HBHA (~1 m depth) using sterile Tygon® tubing connected to a peristaltic pump, and injected into sterile flasks containing Nitrato

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Mineral Salts (NMS) minimal medium [41], under 90:10 air: methanedeheadspace, and incubated with shaking at room temperature. Both isolates failed to grow in the absence of methane and oxygen. Purified DNA from methanotrophic cultures was obtained using the Wizard® Genomic DNA Purification Kit (Promega Inc.). DNA from endosymbiont-containing mussel gill tissue and forest soils was obtained as previously described [18,30]. DNA was extracted from HBHA isolates, peat and Sphagnum moss following the method as previously described [10].

**Primer Design, PCR Amplification, Cloning, and Sequencing**

Methylothroph mmxF-specific PCR primers F1003 and R1561 ([33]; Table 1) sequences were verified through BLAST [42] to determine their specificity to proteobacterial methanotrophs in the GenBank [35] database, and used to amplify partial mmxF gene sequences from soils, peat, Sphagnum moss, methanotrophic HBHA isolates 1 and 2, and control methanotroph cultures. The amplified region encompasses three amino acid residues (out of eight) in the MDH active sites – Asn-261, Asp-303 and Arg-331 – based on the amino acid sequence of Methylobacterium extorquens MDH [26]. BLAST searches of the primer pair F1003 and R1561 retrieved only proteobacterial mmxF genes, but not that of *Candidatus ‘Methylomirabilis oxyfera.’*

Additionally, degenerate primers F1003degen and R1561degen (Table 1) were created after comparing amino acid sequences from MDH retrieved from the GenBank database and identifying degenerate base positions where more than one nucleotide codon specified the same amino acid residue. The degenerate primers were used to PCR amplify mmxF genes from DNA extracts of mussel tissues, as primers F1003 and R1561 failed to PCR amplify mmxF genes from the same DNA extracts. The same primers were also used to PCR amplify mmxF genes from Sphagnum moss in order to verify these primers also PCR amplify mmxF sequences outside of the Methyloococaceae. All PCR reaction mixtures contained 1 × PCR buffer (Qiagen Inc.), 2.5 mM MgCl₂, 300 mM final concentration of each dNTP, a 1.0 µM final concentration of each primer, 1.0 U of Taq polymerase, and approximately 300–800 ng of template DNA in a final volume of 25 µl. PCR conditions were: denaturation at 94°C for 45 sec, primer annealing at 60°C for 1 min, and extension at 72°C for 1.5 min for 30 cycles, and a final 10 min extension at 72°C. PCR products of the expected size (~550 bp for mmxF gene) were purified, cloned, and sequenced (~3 clones per sample).

**Cloning and Sequencing**

PCR products of the expected size (~550 bp for mmxF) were purified (QIAquick PCR Purification Kit, Qiagen Inc.), cloned (pCR® 2.1-TOPO vector from the TOPO TA Cloning Kit, Invitrogen Corp.), using chemical method on TOP 10 competent cells, and plated on LB agar plates containing 50 µg ml⁻¹ kanamycin, and 40 mg ml⁻¹ X-Gal (5-bromo-4-chloro-3-indolyl-β-D-Galactopyranoside). Colonies were screened for inserts using the respective primers via PCR. Plasmid DNA was isolated from positive clones using the QIAprep® Spin MiniPrep Kit (Qiagen Inc.). Sequencing reactions were performed using the ABI PRISM® Big Dye Terminator Cycle Sequencing kit (version 3.1, Applied Biosystems®) and an ABI model 3100 automated sequencer (Applied Biosystems®) according to the manufacturer’s instructions. Recombinants were sequenced in both directions using M13 forward and reverse primers. For environmental DNA extracts, at least five clones were sequenced from each PCR reaction. For *Bathymodiolus* mussels, three clones each were sequenced from frozen gill tissue of three B. azoricus mussels from LS, three B. azoricus mussels from RB, and three B. putosoperetis mussels from LO, for a total of 27 sequences.

**Phylogenetic Analyses of Methylothrophic Bacterial Sequences from GenBank Database**

The ADH gene and/or 16S rRNA gene sequences of *Solobacter umidilis* Ellin 6076 (GenBank Accession no. NC_008536/ CP000473, Phylum Acidobacteria) were used as outgroup in all phylogenetic analyses. *mxaF* gene sequences were aligned using ClustalW2 ([http://www.ebi.ac.uk/Tools/msa/clustalw2/](http://www.ebi.ac.uk/Tools/msa/clustalw2/)) and manually inspected in MacClade 4.08 (Sinauer Associates Inc., Sunderland, MA). Phylogenetic reconstruction was implemented using PAUP 4.0B10 ([43]) and Geneious 4.8.5 ([http://www.geneious.com/](http://www.geneious.com/)) with PAUP plug-in. Unless stated otherwise, statistical support for all trees were obtained from 1,000 bootstrap replicates under the same initial settings (only bootstrap values >50% are reported). Pairwise base comparisons of *mxaF* nucleotide sequences within and between phylogenetic groups were determined using ClustalW2 and reported as % identity values.

(a) **Congruency tests between mxaF and 16S rRNA gene tree topologies.** The program Modeltest [44] was used to determine the best fit substitution model under Maximum Likelihood (ML) analysis (from PAUP software) for each dataset, and searched heuristically for the best model of nucleic acid sequence evolution that best fits our data. The topology of the best tree from ML analyses of each dataset was saved and then enforced as a topological constraint during subsequent paired ML phylogenetic analyses for each dataset. One-tailed Templeton and Shimodaira-Hasegawa (SH) tests were used to compare the constrained and unconstrained topologies using reestimated log likelihoods (RELL) simulation [45,46] and full optimization distributions using 10,000 bootstrap replicates under the likelihood tree scores menu. Uncorrected and Bonferroni-corrected P-values were reported for one-tailed Templeton and SH tests [45]. Confidence intervals were determined for the null hypotheses: the unconstrained and constrained ML tree of each dataset tested have significantly different likelihood scores. The data consisted of nineteen methanotroph taxa spanning three families (Methylocystaceae, Beijerinckiaceae, and Methylococcaceae) and one outgroup for which both genes were available: (a) seven members of the Methylocystaceae – *Methylosinus trichosporium* BF1, *M. trichosporium* O19/1, *M. trichosporium* K821, *Methylosinus sporium* F10/1b, *Methylycitis echinoides* IMET10491, *Methylcystis* sp. IMET10490, and *Methylcystis* sp. IMET10494, (b) four members of the methanotrophic Beijerinckiaceae – *Methylactella silvestris*, *M. tundra*, *M. palaeis*, and *Methylcopra acidiphila*, and (c) eight members of the Methylococcaceae – *Methylomacoccus capsulatus* Bath, *Methylolocaldum* sp. E10a, *Methylomonas methanica*, *M. albus*, *M. rubra*, *Methylolobacter luvis*, *Methylobacter* sp. 5FB, and *Methylomicrobium* sp. For the Methylococcaceae, there are insufficient *mxaF* sequences in GenBank.

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**Table 1.** PCR Primers sets used in this study.

| Name   | Sequence (5' → 3')                      | Reference |
|--------|----------------------------------------|-----------|
| F1003  | GCGGCACCACTTGGGCGCTGCT                | 33        |
| R1561  | GACGGAGCCTCAGTGGTCC                    | 33        |
| F1003degen | GGGCANACTCTGGGGGNTGTT                  | This study |
| R1561degen | GGGGACGCTTTGAGTTGCTGCC                 | This study |

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hence the mxaF of Methylomicrobium album str. BG8 (GenBank Accession no. L33682) was matched to the 16S rDNA gene sequence of Methylomicrobium sp. (GenBank Accession no. D89279). Separate ML phylogenetic analyses were conducted on three datasets: (a) mxaF gene sequences (~513 bp) of these methanotrophs, (b) the corresponding 16S rRNA gene sequences (~1471 bp) of the same taxa and (c) the concatenated mxaF and 16S rRNA gene sequences (~1984 bp).

(b) Phylogenetic analyses of cultured methanotrophs and other related methylotrophs. Maximum parsimony (MP) phylogenies of known methylotrophs (including methanotrophs), and related xoxF PQP-linked dehydrogenase genes, which have recently been described [32] and were included to verify their phylogenetic placement relative to mxaF genes, were constructed based on nucleotide and on inferred amino acid sequences (n = 145) available in GenBank [35]. Alignments for mxaF nucleotide sequences included sequences with and without third nucleotide positions. Phylogenetic analyses (MP) implemented inferred mxaF without third nucleotides to assess third codon position bias) and most parsimonious trees, obtained from a heuristic search.

Separate ML phylogenetic analyses were conducted on three sequence of Methylomicrobium album (GenBank Accession no. L33682) was matched to the 16S rRNA gene sequences (~1984 bp) available in Genbank [35]. Alignments for mxaF and xoxF gene sequences (n = 145) are available in Genbank [35].

Results

Nucleotide Sequence Accession Numbers

The mxaF gene nucleotide and inferred amino acid sequences of Methylomonas rubra and Methylobacter luteus, HBHA isolates 1 and 2, and uncultured clones have been deposited in GenBank, under accession numbers JX312966-JX313018.

Phylogenetic Analyses of Environmental mxaF Gene Sequences

All environmental mxaF nucleotide sequences detected in this study were verified through BLAST searches, and mxaF nucleotide sequences of their closest relatives (identified through database searches) were included in the assembly. The final alignments for mxaF nucleotide sequences, mxaF sequences without third nucleotides and inferred mxaF amino acid sequences comprised 513, 342, and 171 characters, respectively for 145 taxa. The resulting tree was obtained via random stepwise addition of sequences, as consensus of 4 saved, most parsimonious trees, obtained from a heuristic search.

Phylogenetic Analyses of PCR-amplified Environmental mxaF Gene Sequences

All inferred amino acid sequences translated from mxaF genes amplified in this study possessed the three amino acid residues involved in the interactions of the active site. MP phylogenetic trees based on all nucleotides, 1st and 2nd codon positions, and inferred mxaF amino acid sequences were identical in topology; the tree based on all three nucleotides is shown in Fig. 3. Of the mxaF clones amplified from Harvard Forest soils (labeled P_C, P_F, and H_F), one was placed in the Methylomonadaceae and five were placed in the Methylloccaceae, with the remaining three (P_F#1;#2;#3;#6;#10, P_F#4, and P_F#5) closely related to environmental clones detected in other soils. Three sequences detected in Sphagnum moss (labeled Sphag, Fig. 3) clustered with the methylophoroth Methylobacterium multivorans, although this clade lacked bootstrap support. Of the four sequences detected in peat (labeled Peat) in the northern WI bog, one (Peat#4) clustered with mxaF sequences from clones detected in acidic forest and acidic
Figure 1. Congruency tests between \textit{mxaF} and 16S rRNA gene nucleotide sequences of methanotrophs from GenBank database. Phylogenetic trees for congruency tests based on maximum likelihood (ML) analysis of \textit{mxaF} (~513 bp) and 16S rRNA gene nucleotide sequences (~1471 bp) from methanotrophs in GenBank, including the \textit{mxaF} nucleotide sequences of \textit{Methylomonas rubra} and \textit{Methylobacter luteus} sequenced in this study. The ADH gene of \textit{Solibacter usitatus Ellin 6076} was used as outgroup. Methanotrophs (in the Methylococcaceae, Methylocystaceae and Beijerinckiaceae) are indicated by shaded clusters. Accession numbers of \textit{mxaF} and 16S rRNA gene sequences downloaded from GenBank are indicated in parentheses. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if $>50$). The scale bar represents the number of nucleotide changes.

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Figure 2. Simplified phylogenetic tree of methanotrophs and their close relatives based on mxaF nucleotide sequences from GenBank database. Unrooted phylogenetic tree based on maximum parsimony (MP) analysis of known proteobacterial partial mxaF and xoxF/xoxF-like nucleotide sequences (~513 bp) from GenBank and the mxaF nucleotide sequences (in bold) of *Methylomonas rubra* and *Methylobacter luteus* sequenced in this study. The ADH gene of *Solibacter usitatus* Ellin 6076 was used as outgroup. Accession numbers of sequences downloaded...
from GenBank are indicated in parentheses. Bootstrap values from 1,000 replicates are indicated at the nodes of branches (if >50). The three bacterial families containing methanotrophs (Methylococcaceae, Methylocystaceae and methanotrophic members of the Beijerinckiaceae) are indicated by shaded clusters and the other alphaproteobacterial and betaproteobacterial methyloprokaryotes are delineated by lines. The identity of mxaF and mxaF-like sequences from the “Methylobacterium cluster (within cluster 2)”, “Mainly Hyphomicrobiurn (Cluster 3)”, “β proteobacterial methyloprokaryotes (Cluster 5)”, and “mxaF/mxaF-like genes” is shown in Table 2. The scale bar represents the number of nucleotide changes. The complete phylogenetic tree of methanotrophs and their close relatives based on mxaF nucleotide sequences is shown in Supplement S1.

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Discussion

Establishing the link between microbial phylogeny and physiology is complicated by the high level of physiological diversity in most microbial taxa (e.g., many microbes utilizing several carbon sources), and the potential for horizontal gene transfer – the movement of microbial genes between divergent genomes. Consequently, it is necessary to evaluate the accuracy of candidate functional genes as a diagnostic of key metabolic processes, as well as being accurate markers of evolutionary history. Doing so requires evaluating the result of phylogenetic reconstruction of gene sequences from all cultured organisms possessing the gene, before analyses of sequences from uncultured environmental clones. Methanotrophs are unique due to their preference for methane as a metabolic substrate and their possession of functional genes involved in this process. Here, we investigated the phylogeny, based on the mxaF gene, of all known proteobacterial methanotrophs from GenBank database and highlighted the mxaF gene’s ability to detect most methanotrophic bacteria and describe their molecular diversity in natural environments.

mxaF and 16S rRNA Gene Phyllogenies

In this study, the topological differences between 16S rRNA and mxaF gene trees, as seen in incongruencies between both trees (Fig. 1), suggest multiple occurrences of horizontal gene transfer in the mxaF genes of many methanotrophic taxa in the Methylococcaceae, Methylocystaceae, and Beijerinckiaace. Phylogenetic analyses based on mxaF genes placed the vast majority of cultivated proteobacterial methanotrophs, excluding members of the genera Methyloptopium, Methylocella, and Methyloferula, in distinct and coherent clades representing the Methylococcaceae and Methylocystaceae, with higher nucleotide identity between taxa within these clades than between members of different clades. These family-level clade distinctions are consistent with patterns evident in the 16S rDNA phylogeny. However, below the family level, mxaF-based analyses failed to differentiate between distinct subclades based on genera.

The mxaF gene has poor resolving power for methanotrophs within the Beijerinckiaace. Our mxaF gene tree (Fig. 2) indicates that these methanotrophs, composed of the genera Methyloptopium, Methylocella, and Methyloferula, are polyphyletic, sharing common ancestry with other alphaproteobacterial methyloprokaryotes. This pattern corroborates previous studies, which assessed fewer methyloprokaryote taxa [23,49,53], and showed different topologies between mxaF and 16S rRNA genes, for example, in the genus Methyloptopium [22,50,53]. The more extensive mxaF gene phylogeny inferred here (compared to previous studies) suggests that
methanotrophy (a) arose once in the Beijerinckiaceae and was lost by some methylotrophic taxa, (b) arose separately in more than one taxon in the Beijerinckiaceae, and/or (c) multiple occurrences of horizontal gene transfers have occurred in the common ancestor.

Table 2. Taxa included (but not shown) in the phylogenetic analyses for Fig. 2.

| Cluster name                  | Taxa and GenBank Accession number (in parentheses)                                                                 |
|-------------------------------|---------------------------------------------------------------------------------------------------------------|
| Methylobacterium cluster      | Methylobacterium nodulans strain ORS2060 (AF220764)                                                         |
| (within cluster 2)            | Methylobacterium aquatilis DSM1371 (EF562464)                                                              |
|                               | Methylobacterium sp. MP3 (EF030549)                                                                          |
|                               | Methylobacterium kusitamawanum strain MP2 (EF030548)                                                         |
|                               | Methylobacterium rhodesianum strain DSM5687 (EF562473)                                                       |
|                               | Methylobacterium hispanicum strain DSM16372 (EF562468)                                                       |
|                               | Methylobacterium jeotgalii strain 52R03-9 (EF031552)                                                         |
|                               | Methylobacterium sp. MPS (EU047511)                                                                          |
|                               | Methylobacterium mesophilicum strain DSM1708 (EF562470)                                                       |
|                               | Methylobacterium organophilum strain DSM5687 (EF562471)                                                       |
|                               | Methylobacterium radiotolerans JCM2831 (EF562472)                                                            |
|                               | Methylobacterium fujisawaense strain KACC10744 (EF562467)                                                    |
|                               | Methylobacterium oryzae strain CBMB20 (EF562478)                                                             |
|                               | Methylobacterium fujisawaense strain MP1 (EF030547)                                                           |
|                               | Methylobacterium oryzae strain CBMB110 (EF562476)                                                            |
| Mainly Hyphomicrobium          | Hyphomicrobium sp. strain B 314 (HSMXAF314)                                                                 |
| (Cluster 3)                   | Hyphomicrobium sp. strain B 327 (HSMXAF327)                                                                 |
|                               | Hyphomicrobium sp. strain DPB 2c (HSMXAF2C)                                                                  |
|                               | Hyphomicrobium aestuarii (HMXAF)                                                                            |
|                               | Hyphomicrobium sp. strain B 583 (HSMXAF583)                                                                  |
|                               | Hyphomicrobium sp. strain B 520 (HSMXAF520)                                                                  |
|                               | Hyphomicrobium sp. strain B 294 (HSMXAF294)                                                                  |
|                               | Hyphomicrobium sp. strain P 324 (HSMXAF324)                                                                  |
|                               | Hyphomicrobium zavarzinii (HZMXAF)                                                                           |
|                               | Hyphomicrobium sp. strain B 69 (HSMXAF69)                                                                    |
|                               | Hyphomicrobium vulgare (HVMXAF)                                                                             |
|                               | Hyphomicrobium sp. strain P 768 (HSMXAF768)                                                                   |
|                               | Hyphomicrobium sp. strain P 495 (HSMXAF495)                                                                   |
|                               | Hyphomicrobium sp. strain B 495 (HSMXAF495)                                                                   |
|                               | Hyphomicrobium sp. P 495 (HSMXAF495)                                                                          |
|                               | Hyphomicrobium sp. P 768 (HSMXAF768)                                                                          |
|                               | Hansschlegelia plantiphila strain S1 (DQ652143)                                                               |
|                               | Hansschlegelia plantiphila strain S2 (DQ652144)                                                               |
|                               | Hansschlegelia plantiphila strain S4 (DQ652145)                                                               |
|                               | Paracoccus kondratievae NCMIB 13773 (AJ878072)                                                                |
|                               | Methylosulfonomonas methylovora (MMU70525)                                                                    |
|                               | Ancylobacter aquaticus strain IAM 12364 (AB455976)                                                            |
|                               | Allobacter methyloparas DSM 13819 (AJ878096)                                                                  |
|                               | Methylophilaceae ATCC 700716 (AJ878071)                                                                       |
| β-proteobacterial Methylotrophs| Flavobacterium sp. Vu-144 (EU912489)                                                                          |
| (Cluster 5)                   | Duganella sp. B41 (EU439303)                                                                                 |
|                               | Methylobacterium methylovorum (AM004097)                                                                      |
|                               | Hyphomicrobium sp. P 495 (HSMXAF495)                                                                          |
|                               | Hyphomicrobium sp. P 768 (HSMXAF768)                                                                          |
|                               | Hansschlegelia plantiphila strain S1 (DQ652143)                                                               |
|                               | Hansschlegelia plantiphila strain S2 (DQ652144)                                                               |
|                               | Hansschlegelia plantiphila strain S4 (DQ652145)                                                               |
|                               | Paracoccus kondratievae NCMIB 13773 (AJ878072)                                                                |
|                               | Methylosulfonomonas methylovora (MMU70525)                                                                    |
|                               | Ancylobacter aquaticus strain IAM 12364 (AB455976)                                                            |
|                               | Allobacter methyloparas DSM 13819 (AJ878096)                                                                  |
|                               | Methylophilaceae ATCC 700716 (AJ878071)                                                                       |

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of methanotrophic Beijerinckiaceae. It is therefore difficult to ascertain whether microorganisms with \( \text{mxaF} \) gene sequences placed near the polyphyletic Beijerinckiaceae genera \( \text{Methylocapsa} \), \( \text{Methylocella} \), and \( \text{Methylotrenena} \) are indeed methanotrophic.

Overall, our data suggest that the partial \( \text{mxaF} \) gene (~550 bp) amplified by the primer set mentioned is a useful phylogenetic marker and provides sufficient resolution to broadly discriminate between known proteobacterial methanotroph families via the Methylococccaceae and Methylcystaceae clusters, which together encompass the vast majority of known methanotrophs. However, it has poor resolution at the sub-family level across the Methylococccaceae (not shown in previous studies) and Methylcystaceae, and ambiguities exist between the methanotrophic Beijerinckiaceae and other methyloths in the Alphaproteobacteria, due possibly to horizontal transfers of the \( \text{mxaF} \) gene.

**\( \text{mxaF} \) Sequences from Diverse Environments, Isolates and Endosymbiotic Methanotrophs**

The \( \text{mxaF} \) gene was used here to examine the phylogenetic placement and diversity of over fifty novel sequences of putative methanotrophs from a range of natural environments and from cultured isolates in light of evolutionary information from the \( \text{mxaF} \) gene phylogeny determined above. The \( \text{mxaF} \) datasets from Harvard Forest soil, peat and \( \text{Sphagnum} \) moss from northern Wisconsin, and the HBHA water column contained diverse sequences that clustered primarily in the Methylococccaceae and Methylcystaceae, as well as sequences related to methanotrophic Beijerinckiaceae and other alphaproteobacterial methyloths. Notably, the sample of \( \text{Sphagnum} \) moss contained three sequences (\( \text{Sphag} \#1 \), \( \text{Sphag} \#2 \), and \( \text{Sphag} \#3 \)), suggestive of three strains, most closely related to the methyloph \( \text{Methylyrhabdus multivorans} \). However, these three sequences are also closely related to the methanotrophic Beijerinckiaceae genus \( \text{Methylceila} \) (Fig. 3). Given the uncertainty with which \( \text{mxaF} \) discriminates between methanotrophic Beijerinckiaceae and certain alphaproteobacterial methyoths discussed above, the sequences detected in peat, forest soils and \( \text{Sphagnum} \) moss could represent methanotrophic bacteria.

Our analyses also provide the first insights into the phylogenetic placement and biogeography of the previously unknown \( \text{mxaF} \) genes from putative methanotrophic endosymbions of deep-sea hydrothermal vent Bathymodiod mussels. \( \text{Bathymodiolus azoricus} \) and \( \text{B. puteoserpentis} \) are the dominant species of mussels within the two spatially separate mussel hybrid zones of Lucky Strike and Rainbow, and Logatchev on the Mid-Atlantic Ridge [34]. Here, dual bacterial endosymbions provide nutrition to the mussel hosts through thiotrophy and methanotrophy, but the mode of methanotrophic symbiont transmission, whether vertical (symbionts are passed from parent to offspring) or horizontal (symbionts are taken up from the environment, or from co-occurring hosts) is not known [38,39,53,56]. Here, we show that these novel \( \text{mxaF} \) gene sequences of putative endosymbiotic methanoths belong to the Methylcocccaceae, in agreement with their placement in the Gammaproteobacteria with other methanotrophs based on 16S rDNA phylogeny [6]. \( \text{B. azoricus} \) mussels at the adjacent Lucky Strike and Rainbow vent sites harbored a closely related putative methanotrophic symbionts, with most mussels possessing a heterogeneous population of putative endosymbions, based on the closely-related but unique \( \text{mxaF} \) gene sequences we detected. This result is consistent with the hypothesis of environmental acquisition of mussel symbionts [38,55,57,58], where each mussel may be expected to contain multiple genetic variants.

In contrast, \( \text{B. puteoserpentis} \) mussels at Logatchev, a vent site over 2600 km away, harbored a separate monophyletic group of closely related putative methanotrophic symbionts. The phylogenetic clustering of \( \text{B. azoricus} \) and \( \text{B. puteoserpentis} \) symbions in two distinct clades suggests either taxon-specific differences in the specificity of the symbiont-host relationship (i.e., each host species associates with a unique symbiont strain) or that \( \text{B. azoricus} \) mussels at Lucky Strike and Rainbow acquire putative symbionts from an environmental pool that is genetically distinct from that available to \( \text{B. puteoserpentis} \) mussels at Logatchev vent site. Our phylogenetic analyses of \( \text{mxaF} \) gene sequences indicate that the putative endosymbiotic methanoths have diverged from an ancestral sequence into two monophyletic groups, either in response to host-symbiont co-speciation or to geographic separation. Indeed, the fragmented distribution of deep-sea hydrothermal vents may promote spatial isolation that acts as a barrier to symbiont dispersal [59]. With more information, the extent to which symbiont diversification is driven by either geographic isolation and symbiont-host specificity and co-evolution, can be assessed.

Proteobacterial methanotrophs currently constitute the vast majority of known aerobic methane oxidizing bacteria. In this study, we demonstrate the usefulness of the \( \text{mxaF} \) gene in studying proteobacterial methanotroph diversity in non-anoxic environments. Studying the evolutionary history of this gene from known methanotrophs extensively may provide insights into the placement of novel taxa detected in different environments and avoid incorrect inferences from their phylogenetic placement. Our results indicate the \( \text{mxaF} \) gene can be a functional and phylogenetic marker for proteobacterial methanotrophs, providing more information about an important group of microorganisms involved in the global biogeochemical methane cycle.

**Supporting Information**

Supplement S1 (DOCX)

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Author Contributions

Conceived and designed the experiments: EL MCF. Performed the experiments: EL MCF. Analyzed the data: EL MCF PAS CMC. Contributed reagents/materials/analysis tools: EL MCF PAS CMC. Wrote the paper: EL MCF PAS CMC.
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