Unexpected species diversity of Malagasy primates (Lepilemur spp.) in the same biogeographical zone: a morphological and molecular approach with the description of two new species

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Abstract

**Background:** The lemurs of Madagascar provide an excellent mammalian radiation to explore mechanisms and processes favouring species diversity and evolution. Species diversity, in particular of nocturnal species, increased considerably during the last decade. However, the factors contributing to this high diversity are not well understood. We tested predictions derived from two existing biogeographic models by exploring the genetic and morphological divergence among populations of a widely distributed lemur genus, the sportive lemur (Lepilemur ssp.) along a 560 km long transect from western to northern Madagascar.

**Results:** By using the phylogenetic analyses of mtDNA sequence data, molecular diagnostic sites and phenotypic morphometric traits, we uncovered two previously undetected species whose distributions contradict the two existing biogeographic models. Brief species descriptions are provided and a new biogeographic model is proposed (the "large river model").

**Conclusion:** According to the "large river model", large rivers in north and northwestern Madagascar acted as geographical barriers for gene flow and facilitated speciation events on a much smaller spatial scale than previously thought. Thereby, this study does not only show that species diversity in nocturnal Malagasy primates is continuously underestimated but aims to emphasize the need for conservation actions if those species with small ranges shall not face extinction in the near future.

Background

Malagasy lemurs constitute one of six major radiations of extant primates [1]. Lemurs show a remarkable species diversity, both numerically and in terms of adaptations making them an excellent mammalian radiation to explore mechanisms and processes underlying speciation and evolution. During the last decade, species diversity in lemurs increased from 33 to currently 74 [2,3]. In relation to the small surface area of Madagascar, diversity of species within this primate radiation is quite high. Individual
lemur species tend to have small geographic ranges in comparison to other primates. Because of such limited geographic ranges and the high rate of deforestation, the need for conservation action including genetic monitoring and effective management policies is particularly urgent [2,4]. Two major models have been proposed to explain diversity of Malagasy mammals.

The "Martin model" divided northern and northwestern Madagascar into four biogeographical zones (circles in Fig. 1) [5,6]. The western zone (W1) covers the area between the two major rivers Tsiribihina and Betsiboka. The northwestern zone 1 covers the area between the two major rivers Betsiboka and Maevarano (NW), the northwestern zone 2 the area between the rivers Maevarano and Mahavavy (X). The northern zone (N) covers the area between the rivers Mahavavy and Fanambana. These riverine barriers were hypothesized to form geographical boundaries to gene flow and consequently favour allopatric speciation. This model of speciation within Madagascar was refined [6] and it was shown that it is compatible with a reconstruction of speciation within the families Lemuridae, Cheirogaleidae and Indridae [7].

The recent "Wilmé model" explained the process of explosive speciation on the island using a mechanistic model [8]. Madagascar's rivers and associated watersheds with sources at relatively low elevations suggested to be zones of isolation that led to the evolution of locally endemic taxa, whereas those at higher elevations were proposed to have functioned as zones of retreat and dispersion and contain a lower level of microendemism. Wilmé et al. (2006) divided northern and northwestern Madagascar into six centres of endemism (squares in Fig. 1). The western zone (zone 8) covers the area between the two major rivers Tsiribihina and Betsiboka, corresponding to Martin's W1. One large northwestern zone, zone 9, corresponded to Martin's NW. Two smaller northwestern zones, zone 10, between the two rivers Maevarano and Sambirano, and zone 11 between the two rivers Sambirano and Mahavavy were suggested. In addition, two northern zones, one (zone 12), between the river Mahavavy and the continental divide between eastern and western draining watersheds, and another (zone 1), between the continental divide and the river Bemarivo, divided the N-zone of Martin into two partitions.

The geographical settings in northwestern and northern Madagascar are perfect to test if allopatric speciation of a widely distributed lemur genus follows one of the models. Each model predicts a different minimum number of species in this region and divergent distributions. Whereas the "Martin model" predicts four species, the "Wilmé model" proposes six species.

Sportive lemurs (Lepilemur spp.) are an excellent lemur group to test these two models of mammalian distribution in Madagascar, because they occur in almost all forested regions on the island. They are cat-sized vertical climbers and leapers with powerful hind legs. They are nocturnal and totally arboreal. They live in dispersed pairs and have an elaborated vocal repertoire [9-11]. Because differences in pelage colouration and other external characteristics between species are inconspicuous, their early classification [12,13] based on morphological features was disputed until comprehensive cytogenetic approaches and molecular studies allowed the recognition of twelve species [11,14-18].

The aim of this study is to test the predictions from the models with the largest available genetic and morphological data set of a larger-sized lemur. We sequenced three mitochondrial genes of particular diagnostic importance for phylogeography (D-loop, Cytochrome B and NADH-dehydrogenase subunit 4) of individuals captured in 14 different localities that covered a 560 km transect and the area between eight large rivers (Inter-River-Systems, IRS) from western to northern Madagascar. In addition, morphometric data were analysed in order to explore, to

Figure 1
Zonation of northwestern Madagascar described by Martin (1972), zones marked with letters, and by Wilmé et al. (2006), zones marked with numbers.
which extent genetic differentiation coincides with morphological diversification. As in similar studies [e.g. [19,20]] we favour the phylogenetic species concept [21,22], where fixed molecular differences among parapatric populations indicate the existence of species barriers.

Results

Phylogenetic relationships

The 48 sequences available for the D-loop (43 own sequences + five reference sequences), after having cut out the hypervariable part, varied from 388 to 390 bp in length. 128 characters were constant, 201 variable characters are parsimony-uninformative and 66 were parsimony-informative. There were 17 different haplotypes. The 72 sequences available for the partial cytochrome B (43 own sequences + 29 reference sequences) were 352 bp long, with no indels. 211 characters were constant, 17 variable characters are parsimony-uninformative and 124 were parsimony-informative. There were 32 different haplotypes. The 50 sequences available for the partial ND4 (43 own sequences + seven reference sequences) varied from 630 to 631 bp in length. 408 characters were constant, 72 variable characters were parsimony-uninformative and 153 were parsimony-informative characters. There were 19 different haplotypes. Table 1 shows the best-fit models for the three loci selected by the hierarchical likelihood ratio test (hLRT) implemented in Modeltest 3.5.mac. Based on the single-gene-trees, derived for the new and reference sequences, the samples in this study could be classified as follows: The individuals found in IRS 0 clustered with L. aeeclis, the individuals found in IRS I with L. edwardsi, the individuals found in IRS IV with L. sahamalazensis, the individuals found in IRS V and VI with L. dorsalis, the individuals found in IRS VII with L. ankaranensis, the individuals from Kirindy with L. ruficaudatus and the individuals from Mantadia with L. mustelinus. The individuals from IRS II and III did not cluster with any of the reference sequences. No sampled individual clustered with the reference sequences of L. leucopus, L. microdon, L. randrianasoli or L. septentrionalis.

In order to reconstruct the phylogenetic relationships within the genus Lepilemur, we combined these three loci to one concatenated sequence, 1380 bp in length. 768 characters were constant, 333 variable characters were parsimony-uninformative, and 279 were parsimony-informative. There were 21 different haplotypes. The best-fit model selected by hLRT in Modeltest 3.5.mac was the HKY+I+G model (Table 1). Figure 2 shows the Neighbour-Joining tree based on the concatenated sequence. All populations within each IRS clustered together, so that each IRS (including the populations Kirindy (West) and Mantadia (East)) built separate terminal clades, supported by high bootstrap values (Fig. 2). The phylogram consists of four major clades, a western, a northwestern, a northern clade, and the clade of L. mustelinus. L. mustelinus branched off first, followed by the western clade that consisted of IRS 0 and the individuals found in Kirindy (West) (bootstrap values between 93 and 96). The northern clade consisted of IRS IV, V, VI and VII (bootstrap values of 100), and the northwestern clade of IRS I, II and III (bootstrap values of 100). All so far recognized species formed distinct terminal clades with moderate (L. ankaranensis, L. dorsalis) to large (L. mustelinus, L. ruficaudatus, L. aeeclis) branch lengths. Branch lengths among IRS I, II and III in the northwestern clade were in the same scale as these between L. ankaranensis and L. dorsalis.

The absolute pairwise distances within an IRS ranged from zero to seven characters (Fig. 3). The absolute pairwise distances among IRSs (including Kirindy (West) and Mantadia (East)) varied from 18 to 199 characters. The largest absolute pairwise distance (199 characters) existed between L. aeeclis and L. mustelinus. The smallest absolute pairwise distance (18 to 23 characters) among IRSs existed between IRS V and IRS VI, both of them were previously supposed to give home to L. dorsalis. The relative genetic distance between these two IRSs can be defined as intermediate between the intra-IRS differences (0–7 bp) and the interspecific differences (32–199 bp). This level of differentiation could indicate the presence of two subspecies of L. dorsalis. In accordance with the deep phylogenetic

Table 1: Best-fit mutation model for the three mitochondrial loci and the concatenated sequence selected by the hierarchical likelihood ratio test (hLRT) implemented in Modeltest 3.5.mac.

| Locus   | Method | Model    | Base         | Nst | Alpha | Pinvar | TRatio |
|---------|--------|----------|--------------|-----|-------|--------|--------|
| D-loop  | ML, NJ | HKY+G    | 0.3066 0.2151 0.1828 0.2955 | 2   | 0.1752 | 0      | 3.2075 |
| Cyt B   | ML, NJ | HKY+I+G  | 0.3092 0.3229 0.1219 0.2460 | 2   | 3.5808 | 0.5469 | 12.5738 |
| ND4     | ML, NJ | HKY+G    | 0.3372 0.2697 0.1127 0.2804 | 2   | 0.2736 | 0      | 8.1268 |
| concatenated | ML, NJ | HKY+I+G  | 0.3124 0.2661 0.1377 0.2838 | 2   | 0.8801 | 0.3833 | 6.1953 |

Nst: number of substitution types
Pinvar: assumed proportion of invariable sites
Alpha: shape parameter
TRatio: transition/transversion ratio
splits in the Lepilemur tree among the western, northwestern, and northern clade, absolute pairwise distances were always largest when crossing borders among neighbouring biogeographic zones (L. aeeclis to L. edwardsi and IRS III to L. sahamalazensis, Fig. 3). When examining the absolute pairwise distances among the IRSs within the northwestern clade, they were the same size or even larger than between L. dorsalis and L. ankaranensis, which are accepted species (Fig. 3).

Additional files 1, 2, 3 show the molecular diagnostic sites for each terminal clade in each of the three genes. Recognized species had total number of one (L. dorsalis) to 73 (L. mustelinus) sites that allowed to identify them unmistakably. The two terminal clades containing the individuals of IRS II and III had a total of eleven and seven diagnostic sites, respectively. The absolute pairwise distances as well as the analysis of the diagnostic sites indicate the presence of two new Lepilemur species in northwestern Madagascar, one in IRS II and one in IRS III. Consequently, the geographic range of L. edwardsi is much smaller than previously assumed, and limited exclusively to IRS I. By mapping each of these species with respect to their geographical setting (IRS), it can be concluded that all large rivers act as genetic barriers in this genus (Fig. 4).

Figure 2
Neighbour-Joining tree based on the concatenated sequences of the three loci. The branch lengths indicate the number of substitutions, the numbers at the nodes indicate bootstrap values for internal branches (top: NJ, middle: MP, bottom: ML).

Figure 3
Absolute pairwise distances (minimum-maximum and mean) within IRSs, between neighbouring IRSs/species and among the seven already described species.

Morphometry
The means and standard deviations of all morphometric variables for the nine phylogenetically defined species are provided in Table 2. All variables showed significant differences for species in the ANOVA. Post-hoc tests revealed that ear length and intraorbital distance was significantly different in 15 of the 28 possible pairs of species and thereby the two most distinct variables, followed by snout length (14/28), weight (12/28), head width and lower leg.
Table 2: Descriptive statistics (mean ± SD) for 10 morphometric variables from each species.

| Variable               | L. aeeclis (n = 5) | L. edwardsi (n = 11) | L. sp. nov. in IRS II (n = 6) | L. sp. nov. in IRS III (n = 8) | L. sahamalazensis (n = 7) | L. dorsalis (n = 30) | L. ankaranensis (n = 26) | L. mustelinus (n = 7) | Results of ANOVA |
|------------------------|--------------------|-----------------------|-------------------------------|--------------------------------|---------------------------|----------------------|-------------------------|----------------------|-----------------|
| Ear length [mm]        | 30.72 ± 0.83       | 31.75 ± 1.93          | 33.60 ± 0.99                  | 33.41 ± 2.10 **                | 26.73 ± 1.79               | 25.59 ± 1.52 **       | 28.79 ± 1.71 **       | 31.93 ± 3.11       | F = 37.081, p = 0.000 |
| Head width [mm]        | 36.38 ± 0.83       | 34.93 ± 1.36 *        | 37.88 ± 2.24 **               | 37.61 ± 2.29 **               | 34.00 ± 1.37               | 34.69 ± 1.72          | 34.12 ± 1.97          | 38.36 ± 2.33       | F = 9.105, p = 0.000 |
| Snout length [mm]      | 15.50 ± 1.13       | 17.52 ± 2.25 **       | 21.00 ± 0.83 **               | 19.34 ± 1.45 **               | 15.10 ± 1.05               | 14.74 ± 1.69          | 13.97 ± 1.67          | 18.66 ± 2.14       | F = 23.894, p = 0.000 |
| Intraorbital distance [mm] | 13.60 ± 0.65       | 14.51 ± 0.79          | 15.40 ± 1.04                  | 15.29 ± 1.12 **               | 13.31 ± 0.96 **            | 12.08 ± 0.89          | 12.34 ± 0.76          | 12.79 ± 1.17       | F = 25.088, p = 0.000 |
| Interorbital distance [mm] | 36.98 ± 0.80       | 37.80 ± 1.33          | 36.05 ± 1.92                  | 36.88 ± 0.82 **               | 34.96 ± 1.39 **            | 36.82 ± 1.45 **       | 35.48 ± 1.13 **       | 38.41 ± 1.77       | F = 7.470, p = 0.000 |
| Lower leg length [mm]  | 96.30 ± 1.37       | 96.22 ± 5.37          | 101.37 ± 3.63                 | 105.44 ± 2.91                 | 99.27 ± 1.40               | 98.05 ± 5.88          | 99.74 ± 3.69          | 113.30 ± 3.94      | F = 12.874, p = 0.000 |
| Hind foot length [mm]  | 48.14 ± 2.98       | 51.96 ± 2.68          | 50.23 ± 1.43                  | 51.76 ± 3.45                  | 49.00 ± 2.01               | 49.83 ± 2.31          | 48.90 ± 2.51          | 54.94 ± 1.45       | F = 6.953, p = 0.000 |
| 3rd toe length [mm]    | 21.16 ± 0.90       | 22.24 ± 1.47          | 22.37 ± 1.62                  | 23.75 ± 1.60                  | 21.70 ± 0.82               | 20.61 ± 2.38          | 20.80 ± 1.13          | 25.76 ± 2.91       | F = 9.127, p = 0.000 |
| Tail length [mm]       | 260.00 ± 16.58     | 279.73 ± 14.64        | 253.00 ± 13.58 **             | 280.63 ± 15.24 **             | 257.57 ± 11.87             | 263.40 ± 15.53        | 267.88 ± 17.73        | 252.57 ± 16.16     | F = 4.006, p = 0.001 |
| Tail circumference [mm]| 34.80 ± 0.84       | 35.73 ± 3.26          | 40.33 ± 3.27 *                | 35.25 ± 3.58                  | 33.14 ± 4.38               | 34.40 ± 2.19          | 34.42 ± 1.60          | 42.43 ± 6.83       | F = 9.046, p = 0.000 |
| Body mass [g]          | 795.20 ± 80.79     | 934.73 ± 109.06       | 938.50 ± 116.15               | 939.50 ± 96.97 **             | 673.57 ± 120.13            | 713.07 ± 93.24        | 706.31 ± 61.71 **    | 964.57 ± 96.27     | F = 20.680, p = 0.000 |

Two asterisks indicate significant differences (p ≤ 0.05) and one asterisk indicate a statistical trend (0.05 < p < 0.1) between the neighbouring species/columns.
length (10/28), tail circumference (9/28), 3rd toe length (8/28), interorbital distance and hind foot length (6/28) and tail length (4/28). Six variables showed tendencies (0.05 ≤ p < 0.1) in one to two possible pairs of species. The Post-hoc tests revealed significant differences between the northwestern and northern clade and between the northern clade and L. mustelinus. Moreover, it could distinguish between all neighbouring species (established and proposed), except between L. aeelcis from the western clade and L. edwardsi from the northwestern clade (Table 2).

The discriminant function analysis used five variables for model calculation, ear length, snout length, lower leg length, interorbital distance and intraorbital distance. Two functions were computed explaining a significant part of the morphometric variability between the six established and two proposed species (Wilk’s λ = 0.006; F(35,120) = 8.355; p < 0.000). Table 3 shows the classification matrix, with correct classification in 82.5% of the cross-validated cases. The differences between the classification accuracy of each species ranged from 60% to 100%. The individuals of L. aeelcis, L. sp. nova in IRS III and L. mustelinus were correctly classified in 100% of the cases. The individuals of L. sp. nova in IRS II and L. ankaranesis were correctly classified in 80%, and the individuals of L. edwardsi, L. sahamalazensis and L. dorsalis had the smallest percentage of correct classifications (60%). All misclassifications occurred within each major clade, indicating again cryptic speciation within the genus Lepilemur.

Discussion

Revised phylogeny of the genus Lepilemur

Molecular methods, such as DNA sequencing provide powerful tools to understand diversity and phylogeny [7,17,23-30]. This could be confirmed by our study in sportive lemurs. The phylogenetic trees distinguished all previously described species. Moreover, it provided evidence for two previously unknown species in northwestern Madagascar. The absolute pairwise distances between all species were in the range of those observed in other lemur genera such as Mirza [31], Microcebus [31-33], Hapalemur [34,35] and Propithecus [7,36]. The two new taxa occurred in a single IRS (II and III) each. Their phylogenetic position in the tree, the genetic distances and the number of diagnostic sites, suggest a separation at the species level. Similar conclusions were drawn with comparable approaches in other taxa [17,20,27,31,33].

Besides the molecular evidence, the discriminant function analysis of the morphometric data further supported the species status of the two new Lepilemur taxa (IRS II and III) in northwestern Madagascar. Between 80% (IRS II) and 100% (IRS III) of the animals were correctly classified into their IRS of origin. The ANOVA of the morphometric data detected significant differences between the northwestern and northern clade and between the northern clade and L. mustelinus. Moreover, it could differentiate between the neighbouring species (established and proposed) within the northwestern and northern clade, but it could not distinguish between L. edwardsi and L. aeelcis that are geographically separated by the river Betsiboka. The differentiation between the northwestern and northern clade, and between the northern clade and L. mustelinus is stronger than between the species within these major clades. Although the lack of differentiation between the western and northwestern clade may also partly be due to a sample size effect, it may also suggest cryptic speciation events in the genus Lepilemur not only within major clades as it is known in other taxa [37-39], but also between major clades.

The revised phylogeny of the genus Lepilemur is based on the combination of molecular differences (genetic distances and diagnostic sites) and morphometric traits. Diagnostic sites are also routinely used in DNA barcoding, which is becoming an increasingly important tool in species identification [40]. Although DNA barcoding requires a large and nearly complete database of sequences to which individuals can be compared [41,42], the diagnostic sites we identified can be seen as first step towards such a database in Lepilemur.

Table 3: Classification matrix of the discriminant function analysis.

| % correct | L. aeelcis | L. edwardsi | L. sp. nov. in IRS II | L. sp. nov. in IRS III | L. sahamalazensis | L. dorsalis | L. ankaranesis | L. mustelinus |
|-----------|------------|-------------|----------------------|----------------------|------------------|------------|---------------|--------------|
| L. aeelcis | 100.0      | 5           | 0                    | 0                    | 0                | 0          | 0             | 0            |
| L. edwardsi | 80.0      | 1           | 4                    | 0                    | 0                | 0          | 0             | 0            |
| L. sp. nov. in IRS II | 80.0 | 0          | 0                    | 4                    | 1                | 0          | 0             | 0            |
| L. sp. nov. in IRS III | 100.0 | 0          | 0                    | 0                    | 0                | 0          | 0             | 0            |
| L. sahamalazensis | 60.0      | 0           | 0                    | 0                    | 3                | 1          | 1             | 0            |
| L. dorsalis | 60.0      | 0           | 0                    | 0                    | 0                | 0          | 3             | 1            |
| L. ankaranesis | 80.0      | 0           | 0                    | 0                    | 0                | 0          | 1             | 4            |
| L. mustelinus | 100.0     | 0           | 0                    | 0                    | 0                | 0          | 0             | 5            |
| total      | 82.5       | 6           | 4                    | 4                    | 6                | 3          | 5             | 6            |
Description of two new species

Lepilemur otto sp. nov

Holotype
Individual 02y04bibo, adult male captured in Ambodimahabibo on 1st August 2004 by M. Craul (Fig. 5, 6, 7).

Material
Tissue and hair samples, morphometric measurements as well as photographs of 02y04bibo are stored at the Institute of Zoology of the University of Veterinary Medicine Hannover, Hannover, Germany.

Type locality
Madagascar: Province de Mahajanga, Ambodimahabibo (15°29’54,2”S, 47°28’47,2”E).

Paratype
Individuals 01y04bibo, 03y04bibo and 04y04bibo were captured in Ambodimahabibo by M. Craul in 2004. Tissue and hair samples, morphometric measurements as well as photographs of each paratype are stored at the Institute of Zoology of the University of Veterinary Medicine Hannover, Hannover, Germany.

Description
The dorsal pelage, including shoulders and the upper and lower arms, is predominantly grey-brown. A dark diffuse line runs from the middle of the upper skull down the spine, ending in the middle or at the lower part of the back, but is never present on the tail. The ventral pelage is generally grey to creamy. The coloration of the tail is grey-brown to deep brown, sometimes with a white tail tip. The face and forehead are essentially grey.

Figure 5
Lepilemur otto, portrait of individual 02y04bibo (photograph by M. Craul).

Figure 6
Lepilemur otto, body of individual 02y04bibo (photograph by M. Craul).

Figure 7
Lepilemur otto, back of individual 02y04bibo (photograph by M. Craul).
Diagnosis
The sequenced mtDNA of *Lepilemur otto* has eleven diagnostic sites, eight in the ND4 (positions 42 = G, 57 = T, 123 = G, 255 = A, 306 = C, 630 = A, 631 = T, 632 = :C; see additional file 2: Diagnostic sites of the ND4 region for each terminal clade.), and three in the D-loop (positions 20 = C, 22 = A, 23 = T; see additional file 3: Diagnostic sites of the D-loop region for each terminal clade.). *L. otto* differs from its closest relative, *L. edwardsi*, in 2.92–2.99% and from its sister taxon *L. manasamody* in 3.50–3.57% in the sequenced mtDNA, respectively. The few morphometric data, which are available at the moment indicate that *L. otto* has a significant longer snout than the neighbouring species south of the Mahajamba River, *L. edwardsi*. The tail is significant short compared to the neighbouring species north of the Sofia River, *L. manasamody* and to *L. edwardsi*. *L. otto* shows a tendency to have a wider head than *L. edwardsi* and a bigger tail circumference than *L. manasamody*.

Distribution
The known distribution range of *Lepilemur otto* is so far limited to the sample site of Ambodimahabibo. This site is situated in the IRS II, which is limited by the Mahajamba River in the west and the Sofia River in the north. Intensive surveys are now required in this vastly deforested area to obtain additional information about the location and viability of other remaining populations, so that conservation measures can be proposed.

Etymology
The name *Lepilemur otto* was chosen to acknowledge the donation of Dr. Michael Otto for the purpose of research and conservation of Malagasy lemurs.

Vernacular name
Otto’s sportive lemur or Lépilemur de Otto.

*Lepilemur manasamody* sp. nov

**Holotype**
Individual 16y03amb, adult female captured in Ambongabe on 20th September 2003 by M. Craul (Fig. 8, 9, 10).

**Material**
Tissue and hair samples, morphometric measurements as well as photographs of 16y03amb are stored at the Institute of Zoology of the University of Veterinary Medicine Hannover, Hannover, Germany.

**Type locality**
Madagascar: Province de Mahajanga, Ambongabe (15°19'38.3"S, 46°40'44.4"E) and Anjamangirana I (15°09'24.6"S, 47°44'06.2"E).

*Figure 8*
*Lepilemur manasamody*, portrait of individual 16y03amb (photograph by M. Craul).

*Figure 9*
*Lepilemur manasamody*, body of individual 16y03amb (photograph by M. Craul).
Paratype
Individuals 14y03amb and 15y03amb were captured in Ambongabe and individuals 07y03anji, 08y03anji and 09y03anji in Anjiamangirana I by M. Craul in 2003. Tissue and hair samples, morphometric measurements as well as photographs of all paratypes are stored at the Institute of Zoology of the University of Veterinary Medicine Hannover, Hannover, Germany.

Description
The dorsal pelage is predominantly grey-brown, including shoulders, the upper and lower arms. The ventral pelage is generally grey to creamy. The face and forehead are essentially grey. From the middle of the upper skull, a dark diffuse line runs down the spine, ending in the middle of lower part of the back. This line is never present on the tail. The tail is grey-brown to deep brown, sometimes with a white tail tip.

Diagnosis
The sequenced mtDNA of *Lepilemur manasamody* has seven diagnostic sites, two of them in the Cytochrome B (positions 86 = G, 140 = G; see additional file 1: Diagnostic sites of the Cytochrome B region for each terminal clade.), three in the ND4 (positions 171 = T, 201 = G, 333 = A; see additional file 2: Diagnostic sites of the ND4 region for each terminal clade.), and two in the D-loop (positions 75 = G, 156 = G; see additional file 3: Diagnostic sites of the D-loop region for each terminal clade.). *L. manasamody* differs from its sister taxa *L. otto* in 3.50–3.57% and from *L. edwardsi* in 2.77–2.92% in the sequenced mtDNA, respectively. The few morphometric data, which are available at the moment, indicate that *L. manasamody* has a significantly longer tail than *L. otto*. *L. manasamody* has significantly longer ears and a longer snout, a significantly wider head and bigger intraorbital distance and is heavier than the neighbouring species to the north, *L. sahamalazensis*. It also shows a tendency to have a smaller tail circumference than *L. otto*.

Distribution
The known distribution range of *Lepilemur manasamody* is so far limited to the sample sites of Ambongabe and Anjiamangirana I. Both sites are situated in the IRS III, which is limited by the Sofia River in the south and the Maevvarano River in the north. Intensive surveys are now required to obtain additional information about the location and viability of the remaining populations, so that conservation measures can be proposed.

Etymology
The name *Lepilemur manasamody* was chosen after the forest region Manasamody, west of Anjiamangirana I between the Sofia and Maevvarano River.

Vernacular name
Manasamody sportive lemur or Lépilemur de Mana-samody.

Conclusion
Our results showed that all species, except for *L. mustelinus* from the East, grouped in three major clades (western, northwestern and northern). Taking into account the species diversity within each major clade, however, we can define seven biogeographic zones in northern and northwestern Madagascar. When compared to the predictions derived from the "Martin model" and the "Wilmé model", we find several inconsistencies to our data.

The "Martin model" defined four biogeographic zones from western to northern Madagascar (W1, NW, X and N). They corresponded well to the three deep phylogenetic splits, that gave rise to the western, northwestern and northern clade in our study. However, the species diversity within each major clade could not be explained by this model.

The "Wilmé model" defined six biogeographic zones from western to northern Madagascar (numbers 8, 9, 10, 11, 12 and 1). This model may also explain the deep splits between the three major clades, but it proposed two more splits. One between IRS V and VI, divided by the Sambirano River. This split could be confirmed by our study, although it seems not to be a species barrier for sportive lemurs. The level of absolute pairwise distances is intermediate and may rather suggest a variation on a subspecies level. Very recently however, Rabarivola et al. (2006) proposed species status for the individuals in IRS V based on cytogenetics [18]. They collected samples in IRS V at a...
locality further north of Mahilaka and the number of chromosomes differed between individuals from IRS V (2N = 24) and the neighbouring Lepilemur sahamalazensis (2N = 26) and Lepilemur dorsalis (2N = 26). The second additional split indicated by the "Wilmé model" is that between zone 12 and 1. This split could not be confirmed by our study, since all individuals found in IRS VII (corresponding to Willem's zones 12 and 1) clustered together and belonged to the species L. ankaranensis. One major discrepancy exists between our findings and the "Wilmé model". The "Wilmé model" predicts one centre of endemism in northwestern Madagascar (zone 9), which should correspond to one Lepilemur species in that area. Our study provided evidence, however, for three species of sportive lemurs between the Betsiboka and Maevvarano River, each restricted to one of the three IRSs. Thus, we showed that each IRS is represented as a separate terminal clade in the phylogenetic trees, building distinct phylogenetic units. At least six of the seven large rivers act as species barriers for Lepilemur. Therefore, we propose a new model, the "large river model" to explain the biogeography of this larger-sized nocturnal lemur genus. Large rivers acted as insurmountable barriers for gene flow, leading to cryptic speciation within larger biogeographic units. Except for IRSs V and VI, the genetic distances among all IRSs reach species level.

The deep splits between the major clades may indicate initial colonization events, with the Betsiboka and Maevvarano River playing a major role in long-term and continuous isolation of western, northwestern and northern Madagascar. The splits within each major clade however, indicate younger cryptic speciation events. Populations, initially belonging to one founder species, entered the IRSs I-III and VI-VII respectively, and were subsequently separated from each other by the rivers Mahajamba and Sofia, and the Andranomalaza, respectively. Quaternary paleoclimatic variation may have played another important role in shaping biogeography and speciation events on Madagascar. The climate during periods of glaciation was cooler and drier than today [8,43,44]. Rivers with year-round water course could have acted as retreats/refugia in times of aridification. All seven large rivers in northwestern and northern Madagascar should have belonged to this category, since the genetic isolation of the IRSs would otherwise not have persisted over time and signs of repeated introgression should be detectable. Subsequent recolonization of the IRSs should thereby have originated from small and isolated refugia, which further promoted genetic differentiation between the IRSs.

In conclusion, we presented evidence for an unexpected species diversity of sportive lemurs in northwestern and northern Madagascar. Current biogeographic models were not sufficient to explain the underlying processes of speciation. We therefore suggest a new model of biogeographical zonation, the "large river model". In this model, biogeographic zones are separated and maintained over time by all large rivers with permanent water bodies that may have provided retreat zones during periods of aridification and may have harboured founder populations for subsequent recolonization. The importance of large rivers as biogeographic barriers was previously emphasized for mouse lemurs [3], but also for neotropical primates [45,46]. Further studies are now needed to test the relevance of this model for other terrestrial taxa, such as the insectivores, rodents, or other lemurs.

**Methods**

**Fieldwork**

A total of 157 Lepilemur individuals were captured at 14 different localities along a 560 km transect from western to northern Madagascar (Fig. 4, Table 4). This region is divided by eight large rivers (over 50 km wide 20 km inlands) into eight Inter-River-Systems (IRS 0 to IRS VII, Fig. 4). Six localities were sampled by Mathias Craul (MC) and eight localities were sampled by Solofo Rasohariharina (SR) and Blanchard Randrianambinina (BR). At each site we performed daily and nightly surveys to capture the animals. At daytime we used a net to capture animals out of their sleeping holes and briefly anesthetised them with Ketaset-5 (Selectavet). At night time we anesthetised the animals using a blowpipe (TELINJECT B22T) with Ketaset-5 (Selectavet). Each captured sportive lemur was then characterised with regard to sex, skin colour, reproductive status (testis size or form of vulva), 13 external morphometric measures (ear length, ear width, head length, head width, snout length, interorbital distance, intraorbital distance, lower leg length, hind foot length, 3rd toe length, body length, tail length, tail circumference) and body mass [10,47]). In addition, a small biopsy from one or both pinnae was taken as tissue samples. Tissue samples were stored in Queen's lysis buffer [48] for later DNA extraction and genetic analyses.

**Molecular methods and analyses**

DNA from the tissue of 37 individuals was isolated with the DNeasy Tissue Kit (Qiagen), or extracted using a standard protease K digestion followed by a Phenol/Chloroform protocol [49] and stored at -20°C. In addition, we analysed the DNA of two individuals of Lepilemur ruficaudatus (Kirindy forest, western Madagascar) provided by Yves Rumpler and of four individuals from Mantadia (eastern Madagascar) sampled previously by SR and BR. We sequenced the mitochondrial genes D-loop, cytochrome B and NADH-dehydrogenase subunit 4 (ND4), because reference sequences from all eleven recognized species were available for these particular markers. The complete D-loop was amplified with the oligonucleotide
primers DLp-1.5: 5'-GCC CCC AAA GCT GAR RTT CTA-3' and DLp-5: 5'-CCA TCG WGA TGT CTT ATT TAA GRG GAA-3' [19]. Standard PCRs were carried out in a 25 μl reaction with a final concentration of 1 μM for each primer, 1.5 mM for MgCl₂, 0.2 mM for each dNTP, 1 x NH₄ reaction buffer (50 mM Tris-HCl pH 8.8, 16 mM (NH₄)₂ SO₄, 0.1% Tween® 20), 1.25 units of Taq DNA polymerase, and 1 μl of DNA. Successful amplifications were obtained using the following protocol: 35 cycles of denaturing at 94°C for 60 seconds, primer annealing at 47°C for 60 seconds and extension at 72°C for 90 seconds. The partial Cytochrome B was amplified with the oligonucleotide primers L14841: 5'-AAA AAG CTT CCA TCC AAC ATC TCA GCA TGA TGA AA-3' and H15149: 5'-AAA CTG CAG CCC CTC AGA ATG ATA TTT GTC CTC A-3' [50]. Standard PCRs were carried out in a 25 μl reaction with a final concentration of 1 μM for each primer, 1.5 mM for MgCl₂, 0.2 mM for each dNTP, 1 x NH₄ reaction buffer (50 mM Tris-HCl pH 8.8, 16 mM (NH₄)₂ SO₄, 0.1% Tween® 20), 1.25 units of Taq DNA polymerase, and 1 μl of DNA. Successful amplifications were obtained using the following protocol: 35 cycles of denaturing at 94°C for 60 seconds, primer annealing at 47°C for 60 seconds and extension at 72°C for 90 seconds. The partial NADH-dehydrogenase subunit 4 was amplified with the oligonucleotide primers LepiP1: 5'-TTG ATG TAG TAT GAC TRT TCC-3' and LepiR1: 5'-GCC AAA CCG ATG GCT GCT TCA CAG GCT TCC AG-3' [51]. Standard PCRs were carried out in a 25 μl reaction with a final concentration of 1 μM for each primer, 1.5 mM for MgCl₂, 0.2 mM for each dNTP, 1 x NH₄ reaction buffer (50 mM Tris-HCl pH 8.8, 16 mM (NH₄)₂ SO₄, 1.25 units of Taq DNA polymerase, and 1 μl of DNA. Successful amplifications were obtained using the following protocol: 40 cycles of denaturing 95°C for 30 seconds, primer annealing at 60°C for 60 seconds and extension at 72°C for 60 seconds. The PCR products were cleaned with the Invisorb Spin PCRapid Kit (Invitek) or Quick-Clean (Bioline) and checked for successful amplification by running an aliquot on a 1.5% agarose gel, stained with 1.3 × 10⁻⁴ mg/ml ethidium bromide. After cleaning the PCR products, cycle sequencing reactions were carried out using DYEnamic™ ET dye terminator kit (Amersham Biosciences) and the primers indicated above. After a second cleaning with ammonium acetate, provided with the DYEnamic™ ET dye terminator kit, the PCR products were sequenced on a MegaBACE™ 1000 DNA Sequencing System (Amersham Biosciences). The respective sequences were deposited in GenBank (Table 5).

For a comprehensive phylogenetic analysis of the sequence data, we expanded our data set with reference sequences from all eleven recognized species available from GenBank (Table 5). As outgroup for phylogenetic tree reconstructions, we selected Propithecus diadema. Sequences were aligned using the program CLUSTALX [52] and checked by eye. Tree reconstructions of each single gene were carried out to phylogenetically classify the sampled individuals within the genus Lepilemur. Because of the lack of reference sequences of single individuals for all three genes, further phylogenetic tree reconstructions based on all three genes were performed only with our own data set consisting of 43 sequences. Phylogenetic tree reconstructions were carried out with the maximum-parsimony (MP), neighbour-joining (NJ) and maximum-likelihood (ML) algorithms as implemented in PAUP4.0b10 [53]. Throughout the analyses, all characters were treated as unordered and equally weighted. Gaps were considered as missing data in NJ and ML, but were treated as fifth character in MP analysis. The NJ and ML trees were constructed using the best-fit model selected by the hierarchical likelihood ratio test (hLRT) in Modeltest.

### Table 4: Details of study sites.

| Locality   | Abbreviation | Coordinates          | Origin |
|------------|--------------|----------------------|--------|
| Madirovalo | madi         | 16°22'45.6"S, 46°29'01.9"E | IRS 0  |
| Ampijoroa  | jbb          | 16°17'5, 46°48'E      | IRS I  |
| Mariarano  | mari         | 15°28'50.3"S, 46°41'19.0"E | IRS I  |
| Tananavaoao| tan          | 15°28'15.5"S, 46°39'59.4"E | IRS I  |
| Ambodimahabibo | bibo      | 15°29'54.2"S, 47°28'47.2"E | IRS II |
| Ambongabe  | amb          | 15°19'38.3"S, 47°40'44.4"E | IRS III|
| Anjamangirana I | anji    | 15°09'24.6"S, 47°44'06.2"E | IRS III|
| Ankarafa   | sah          | 14°22'47.8"S, 47°45'26.3"E | IRS IV |
| Mahilaka   | mah          | 14°17'12.0"S, 48°12'12.0"E | IRS V  |
| Lokobe     | lok          | 13°23'23.9"S, 48°20'31.0"E | IRS VI |
| Manehoka   | oka          | 13°25'49.0"S, 48°47'51.0"E | IRS VI |
| Ankavana   | anka         | 12°46'55.7"S, 49°22'27.4"E | IRS VII|
| Ankarana   | kar          | 12°58'05.0"S, 49°08'18.0"E | IRS VII|
| Analabe    | ana          | 12°45'13.8"S, 49°30'03.9"E | IRS VII|
| Kirindy    | kir          | 20°03'5, 44°37'E      | West   |
| Mantadia   | man          | 18°47'5, 48°25'E      | East   |
| Species  | Locality | Abbreviation | Origin | Sample type | D-loop  | Cyt B  | ND4    |
|----------|----------|--------------|--------|-------------|---------|--------|--------|
| L. ruficaudatus | Kirindy  | kir1         | west   | DNA         | EF686766 | EF686723 | EF686680 |
| L. ruficaudatus | Kirindy  | kir2         | west   | DNA         | EF686767 | EF686724 | EF686681 |
| L. ruficaudatus | Kirindy  |             |        | sequence    | DQ109013– | DQ109015, | AF224596 |
| L. randrianasoli | Andramasay |             | west   | sequence    | AY321456, | DQ109019, | DQ234891– |
| L. randrianasoli | Ambalarano |             | west   | sequence    | Q234894,  | DQ234890  |        |
| L. aeeclis | Madirovalo | madi1        | IRS 0  | tissue      | EF686768 | EF686725 | EF686682 |
| L. aeeclis | Madirovalo | madi2        | IRS 0  | tissue      | EF686769 | EF686726 | EF686683 |
| L. aeeclis | Antafiala- | IRS 0        | sequence | EF6866768 |
| L. edwardsi | Ampijoroa | jbb1         | IRS I  | tissue      | EF686756 | EF686713 | EF686670 |
| L. edwardsi | Ampijoroa | jbb2         | IRS I  | tissue      | EF686757 | EF686714 | EF686671 |
| L. edwardsi | Marirano  | mari1        | IRS I  | tissue      | EF686760 | EF686717 | EF686674 |
| L. edwardsi | Marirano  | mari2        | IRS I  | tissue      | EF686761 | EF686718 | EF686675 |
| L. edwardsi | Tananaoaoa | tan1         | IRS I  | tissue      | EF686758 | EF686715 | EF686672 |
| L. edwardsi | Tananaoaoa | tan2         | IRS I  | tissue      | EF686759 | EF686716 | EF686673 |
| L. edwardsi | Andofombomb |             | IRS I  | sequence    | DQ109006, | DQ109004, | AF224595 |
| L. edwardsi | Andofombomb |             | IRS I  | sequence    | DQ234888, |
| L. sp. nov. IRS II | Ambodimahabibo | bibo1    | IRS II | tissue      | EF686762 | EF686719 | EF686676 |
| L. sp. nov. IRS II | Ambodimahabibo | bibo2    | IRS II | tissue      | EF686763 | EF686720 | EF686677 |
| L. sp. nov. IRS II | Ambodimahabibo | bibo3    | IRS II | tissue      | EF686764 | EF686721 | EF686678 |
| L. sp. nov. IRS II | Ambodimahabibo | bibo4    | IRS II | tissue      | EF686765 | EF686722 | EF686679 |
| L. sp. nov. IRS III | Ambongabe | amb1        | IRS III | tissue      | EF686753 | EF686710 | EF686667 |
| L. sp. nov. IRS III | Ambongabe | amb2        | IRS III | tissue      | EF686754 | EF686711 | EF686668 |
| L. sp. nov. IRS III | Anjamanga | anji1       | IRS III | tissue      | EF686755 | EF686712 | EF686669 |
| L. sahamalazensis | Ankarafa | sah1        | IRS IV  | tissue      | EF686750 | EF686707 | EF686664 |
| L. sahamalazensis | Ankarafa | sah2        | IRS IV  | tissue      | EF686751 | EF686708 | EF686665 |
| L. sahamalazensis | Ankarafa | sah3        | IRS IV  | tissue      | EF686752 | EF686709 | EF686666 |
| L. sahamalazensis | Sahamalaza |             | IRS IV  | sequence    | DQ108990– | DQ108992, | DQ234882, |
| L. sahamalazensis | Sahamalaza |             | IRS IV  | sequence    | DQ234883  | DQ234880, |
| L. dorsalis | Mahilaka  | mah1         | IRS V   | tissue      | EF686746 | EF686703 | EF686660 |
| L. dorsalis | Mahilaka  | mah2         | IRS V   | tissue      | EF686747 | EF686704 | EF686661 |
| L. dorsalis | Mahilaka  | mah3         | IRS V   | tissue      | EF686748 | EF686705 | EF686662 |
| L. dorsalis | Ambanja   |             | IRS V   | sequence    | DQ108995– | DQ108997, | DQ234886, |
| L. dorsalis | Ambanja   |             | IRS V   | sequence    | DQ234887  |        |        |
| L. dorsalis | Mahilaka  | mah4         | IRS V   | tissue      | EF686749 | EF686706 | EF686663 |
| L. dorsalis | Lokobe    | lok1         | IRS VI  | tissue      | EF686740 | EF686677 | EF686654 |
| L. dorsalis | Lokobe    | lok3         | IRS VI  | tissue      | EF686741 | EF686698 | EF686655 |
| L. dorsalis | Manohoka  | oka1         | IRS VI  | tissue      | EF686744 | EF686701 | EF686658 |
| L. dorsalis | Manohoka  | oka2         | IRS VI  | tissue      | EF686742 | EF686699 | EF686656 |
| L. dorsalis | Manohoka  | oka3         | IRS VI  | tissue      | EF686743 | EF686700 | EF686657 |
| L. dorsalis | Manohoka  | oka4         | IRS VI  | tissue      | EF686745 | EF686702 | EF686659 |
3.5. mac [54]. Relative support of internal nodes was provided by bootstrap analyses with 1,000 replications for MP and NJ and 100 replications for ML. Absolute pairwise distances were calculated using PAUP4.0b10 [50] and ARLEQUIN 1.1 to describe the variation among taxa. To determine fixed molecular differences among terminal clades (indicating barriers for gene flow), diagnostic sites for each terminal clade to all others were identified using the program MEGA 3.1 [55].

### Statistical analyses of morphometric data
Quantitative analyses of morphometric data were carried out with two different sample sizes. The ANOVA was conducted with 100 individuals. After removing two variables that differed among researchers (1-way ANOVA, STATISTICA 6.0, Statsoft, Inc.), the 11 remaining variables were tested for normality using the Kolmogorov-Smirnov test (Statistica 6.0, Statsoft, Inc.) at a level of $p \leq 0.05$. All were normally distributed. A MANOVA revealed no differences in sex. The variables were then tested for correlation. All variables had an $r < 0.75$ and were therefore defined as sufficiently independent to be used in a discriminant function analysis [56]. This analysis was limited to five adult individuals per species (established and proposed) in order to equilibrate the samples. The discriminant function analysis tested only for species differences and for differences between the IRSs. A stepwise forward method (statistic: Wilk’s $\lambda$) with the criteria $F_{\text{to enter}} = 3.84$ and $F_{\text{to remove}} = 2.71$ and a tolerance level of $p \leq 0.01$ was used to calculate the discriminant function model. The computed discriminant functions were used to classify cases with regard to their group membership. All cases were cross-validated by the “leave-one-out” method, where each case in the analysis is classified by the functions derived from all cases other than that case. The discriminant function analysis was carried out with the program SPSS 13.0 (SPSS, Inc.).

### Footnote
During the review process of this paper, Louis Jr. et al. (2006) described a new sportive lemur species in IRS III [57]. It was named *Lepilemur grewcocki*. This might be a
synonym to *L. manasamody*, as our sampling sites of this species were in the same IRS. However, a joint phylogenetic analysis is still needed to verify the identity of both forms.

**Authors' contributions**

MC participated in the design of the study, conducted part of the field work and all lab work, performed the comparative genetic and morphometric analyses and wrote the MS.

EZ initiated, financed and designed the study. She organized and conceptualized the field work, supervised data analyses and critically revised the MS several times and approved its final version.

SR conducted part of the field work including capturing, measuring and sampling *L. ankaranensis*, *L. dorsalis*, *L. mustelinus* and *L. aeeclis*.

BR conducted part of the field work including capturing, measuring and sampling *L. ankaranensis*, *L. dorsalis*, *L. mustelinus* and *L. aeeclis*.

UR designed the study. She organized and conceptualized the field work, supervised data analyses and critically revised the MS several times and approved its final version.

**Additional material**

Additional file 1

Diagnostic sites of the Cytochrome B region for each terminal clade. Dashes (-) indicate deletions. Points (.) indicate identical bases.

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Additional file 2

Diagnostic sites of the ND4 region for each terminal clade. Dashes (-) indicate deletions. Points (.) indicate identical bases.

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Additional file 3

Diagnostic sites of the D-loop region for each terminal clade. Dashes (-) indicate deletions. Points (.) indicate identical bases.

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