Transgenic mouse model harboring the transcriptional fusion ccl20-luciferase as a novel reporter of pro-inflammatory response.

Martina Crispo, Laurye van Maele, Julien Tabareau, Delphine Cayet, Agustina Errea, Ana María Ferreira, Martin Rumbo, Jean Claude Sirard

To cite this version:

Martina Crispo, Laurye van Maele, Julien Tabareau, Delphine Cayet, Agustina Errea, et al.. Transgenic mouse model harboring the transcriptional fusion ccl20-luciferase as a novel reporter of pro-inflammatory response.. PLoS ONE, 2013, 8 (11), pp.e78447. 10.1371/journal.pone.0078447 . inserm-00925644

HAL Id: inserm-00925644
https://inserm.hal.science/inserm-00925644

Submitted on 9 Jan 2014

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Transgenic Mouse Model Harboring the Transcriptional Fusion Ccl20-Luciferase as a Novel Reporter of Pro-Inflammatory Response

Martina Crispo1, Laury Van Maele2,3,4,5, Julien Tabareau2,3,4,5, Delphine Cayet2,3,4,5, Agustina Errea6, Ana María Ferreira7, Martin Rumbo6, Jean Claude Sirard2,3,4,5*

1 Unidad de Animales Transgénicos y de Experimentación – Instituto Pasteur de Montevideo, Montevideo, Uruguay, 2 Institut Pasteur de Lille, Centre d’Infecti and d’Immunité de Lille, F-59000 Lille, Region Nord-Pas de Calais, France, 3 Institut National de la Santé et de la Recherche Médicale, U1019, F-59000 Lille, Region Nord-Pas de Calais France, 4 Centre National de la Recherche Scientifique, UMR 8204, F-59000 Lille, Region Nord-Pas de Calais France, 5 Univ Lille Nord de France, F-59000 Lille, Region Nord-Pas de Calais, France, 6 Laboratorio de Investigaciones del Sistema Inmune (LISIN) – National University of La Plata, Provincia de Buenos Aires, Argentina, 7 Catedra de Inmunología, Facultad de Ciencias/Facultad de Química, Universidad de la República, Montevideo, Uruguay

Abstract

The chemokine CCL20, the unique ligand of CCR6 functions as an attractant of immune cells. Expression of CCL20 is induced by Toll-like Receptor (TLR) signaling or proinflammatory cytokine stimulation. However CCL20 is also constitutively produced at specific epithelial sites of mucosa. This expression profile is achieved by transcriptional regulation. In the present work we characterized regulatory features of mouse Ccl20 gene. Transcriptional fusions between the mouse Ccl20 promoter and the firefly luciferase (luc) encoding gene were constructed and assessed in in vitro and in vivo assays. We found that liver CCL20 expression and luciferase activity were upregulated by systemic administration of the TLR5 agonist flagellin. Using shRNA and dominant negative form specific for mouse TLR5, we showed that this expression was controlled by TLR5. To address in situ the regulation of gene activity, a transgenic mouse line harboring a functional Ccl20-luc fusion was generated. The luciferase expression was highly concordant with Ccl20 expression in different tissues. Our data indicate that the transgenic mouse model can be used to monitor activation of innate response in vivo.

Introduction

Chemokines are small proteins that play a major role in the chemotactic movement of cells along the body. They are critical for the proper positioning of immune cells and the development of immune responses. Thus, the expression of chemokines is highly regulated [1]. Chemokines can be divided into constitutive and inducible chemokines with regards to their function. Constitutively expressed chemokines contribute at steady state to positioning of immune cells in lymphoid and non lymphoid organs [2]. Inducible chemokines are swiftly and transiently expressed upon stimulation by external factors and promotes the migration of immune cells to stimulation sites [3]. In pro-inflammatory immune responses triggered during infection, inducible chemokines orchestrate the entry of leukocytes such as neutrophils or inflammatory monocytes into the site of injury.

Some chemokines may participate in both homeostatic and alarm-driven functions. The chemokine CCL20 (or LARC for liver- and activation-regulated chemokine) is constitutively expressed in specific sites such as follicle-associated epithelium of Peyer’s patches, isolated lymphoid follicles and tonsils. In addition, the production of CCL20 is strongly upregulated by various danger stimuli in different organs such as liver, intestine or lung and also mucosal sites [4–7]. The gene coding for CCL20 represents a stereotypical signature of innate response to infection as determined by meta-analysis of microarray data [8,9]. CCR6, the unique receptor of CCL20, is expressed by B cells, activated T cells (particularly Th17), regulatory T cells, immature DC or innate lymphoid cells [10–14]. How CCL20 participates in constitutive and inducible cells of immunity is supported by the analysis of the promoters from human (CCL20) and mouse (Ccl20) genes [15–18]. Conserved motifs specific for binding of transcription regulators were identified, including the canonical NF-xB and the Ets (E-twenty six) family binding sites [15,17,19]. The promoters of CCL20 encoding genes are interesting candidates for the establishment of reporter systems. We have previously characterized the CCL20 human promoter gene and generated stable reporter human intestinal cell lines [19,20] to investigate how microbial products regulate the transcriptional activity of the gene [21,22].

The flagellin family, the major constituent of bacteria flagella, forms the unique agonist of Toll-like receptor 5 (TLR5) signaling [23]. Whereas there are differences in human and mouse TLR5 sequences, the specific recognition of flagellins seems to require similar TLR5 determinants [24]. Remarkably, a stop mutation of
Figure 1. Functional characterization of Ccl20-luc transcriptional fusions. A. Alignment of human and mouse CCL20 promoters. Position +1 indicates initiation site for transcription and first codon is highlighted in grey. * indicates homology in the sequence alignment. Critical features and
transcription factor binding sites are underlined. B. Schematic representation of the different reporter constructs. (luc: firefly luciferase gene, pA: polyadenylation signal). "IRES" stands for internal ribosome entry site. The cDNA of luc encodes the enhanced yellow fluorescent protein. Positions related to promoter are coincident with panel A. C. Functional analysis of Ccl20-luc in vivo. BALB/c mice were injected for 48 h by hydrodynamic shock with Ccl20-luc and TK-renilla plasmids, treated next i.v. for 8 h with 15 μg flagellin or PBS, and liver luciferase activity was determined. Results are expressed as relative luciferase activity using PBS-treated mice as reference and renilla activity as normalizer for transfection efficiency. Data are given as mean ± SD (n = 6–8; * p < 0.05 Mann-Whitney test) and are representative of three experiments. D. NF-κB site but not Ets1 site is required for the activation of transcriptional fusion by flagellin. Experiments were conducted as described in C using different luciferase reporter Ccl20-luc (−299/+6) plasmids harboring deletions or mutations as shown in B. E. Liver response induced by flagellin depends on TLR5. Experiments were conducted as described in C using Ccl20-luc, TK-renilla and dominant negative TLR5Δ393apo or control pcDNA3 plasmids. Data are given as mean ± SD (n = 6–8; * p < 0.05 n.s.: non significant differences, Mann-Whitney test) and are representative of three experiments. F. Liver response induced by flagellin is TLR5 dependent. Experiments were conducted as described in C using Ccl20-luc-IRES-eyfp. Data are given as mean ± SD (n = 8; * p < 0.05 Mann-Whitney test) and are representative of two experiments.

doi:10.1371/journal.pone.0078447.g001

human TLR5 (1–392) acts as a dominant negative mutant on TLR5 signaling [25]. MyD88 is an essential adaptor molecule for TLR5 signaling since responses are totally abolished in MyD88−/− mice [26,27]. TLR5 stimulates transcription of pro-inflammatory genes dependent on NF-κB and MAPKs. Interestingly, flagellins signal very efficiently in both hematopoietic cells including dendritic cells, and stromal cells, especially intestinal epithelial cells or liver cells [6,28,29]. Thus, flagellins are a potent inducer of CCL20 in human and mouse systems [30], thereby constituting an appropriate way to address regulation of gene coding CCL20.

In this work, we characterized regulatory features of the mouse Ccl20 gene and generated a mouse reporter system. To this aim, transcriptional fusions between the Ccl20 promoter and the firefly luciferase (luc) encoding gene were constructed and assessed in vitro and in vivo assays. Flagellin was used to evaluate the function of promoter and the specificity of regulation with shRNA against Th5 and a dominant form of mouse TLR5 (TLR5Δ393apo). A functional Ccl20-luc fusion was finally integrated in a mouse line to address in vivo the regulation of gene activity. We showed that this model can be used to monitor activation of innate response in vivo, with maximal expression in the liver.

Materials and Methods

Animals

BALB/cj, C57BL/6j and the transgenic line U68 harboring the Ccl20-luc-IRES-EYFP construct were bred at the SPF animal facility of the Transgenic and Experimental Animal Unit of Institut Pasteur de Montevideo or Institut Pasteur de Lille (agreement #A59-35009). NFκB-RE-luc mice were purchased from Taconic (Germantown, NY, USA). Experimental procedures complied with international FELASA guidelines, national regulations and ethical guidelines (French protocol CEEA 052011 and Uruguayan Animal Care Committee) and all experimental procedures involving animals were approved by the corresponding Institutional Animal Care Committees of either Institut Pasteur de Lille or Institut Pasteur de Montevideo.

Plasmid Constructs

The Ccl20 gene (about 1.4 kb of promoter and 1.0 Kb of transcribed gene) was originally amplified using the following primers: CTCGAGCAGAATGGAGAGTGGCA and AAGCCTTATGGTACCGAGGCGACATCG (pHindIII). The promoter was then cloned using XhoI and the endogenous KpnI site close to the start of transcription (Figure 1A) into the firefly luciferase reporter pGL-3 basic vector (Promega, USA). The resulting construct pCcl20-luc (−1377/+11) harbors 1388 bp (1377 bp upstream transcription start site and 11 bp of transcribed region) of the Ccl20 promoter region. To generate the transgenic animals, an IRES-EYFP cassette obtained from Dr. Richard M. Locksley [31] was cloned downstream the luc cDNA of pCcl20/luc to give rise to the plasmid pCcl20-luc-IRES-eYFP. We also used the reporter plasmids harboring Ccl20 promoters −299/+6, −85/+6, −299/+4 (NF-κB mutant) and luc fusions previously described by Fujiie et al. [15]. An Ets-1 mutant fusion was constructed using the −299/+6 reporter fusion as template to change the (−156) GAGCAGAAGT (−147) Ets-1 site in GCATTGACT. To define the TLR5 specificity of flagellin response, two sets of constructs were used. First, plasmids expressing functional shRNA under the control of 78K promoter targeting Th5 or coding for a scrambled shRNA sequence (Invivogen, France). Second, the cDNA coding for the recombinant mouse TLR5 (residues 1 to 392 with codon 393 changed in a stop codon), a sequence equivalent to the dominant negative human TLR5 [25] was cloned into the HindIII and NotI sites of the vector pcDNA3 using the PCR fragment obtained with primers: 4AGCCTTGGATCATGACATGTCACCATGGACTTGC and GCAGGGGCTCAGAGTCTGAAGGTGTGTAAGC. All plasmids were verified by sequencing.

Hydrodynamic Shock Injections

BALB/c mice were injected with combination of plasmids diluted in 1.4 mL Ringer solution (147 mM NaCl, 4 mM KCl, 1.13 mM CaCl2, pH 7.5) via tail vein as described previously [32]. Typically, 25 μg Firefly luciferase reporter plasmid (Ccl20 promoter) and 2.5 μg Renilla luciferase plasmid pRL-TK (Promega) for the normalization of transfection efficiency ±25 μg plasmid for interference or dominant negative or control plasmids, were used for each animal. Plasmid injection was completed in less than 5 s and the level of activation of reporter plasmids was assessed 40 h later. For this purpose, animals were injected i.v. with 200 μL PBS ±5 μg flagellin as described previously [19,21]. Different organs including the liver, spleen, or lung were sampled 6 h later to measure luciferase activity (Firefly and Renilla), using Dual-Luciferase Assay (Promega), following manufacturer instructions.

Generation and Analysis of Transgenic Animals

Transgenic animals were generated on a C57BL/6j genetic background at the Institut Clinique de la Souris in Strasbourg, France. Briefly, the plasmid pCcl20-luc-IRES-eYFP was purified and injected in fertilized eggs from C57BL/6j. Several offspring founders were generated and identified by PCR genotyping and bred with C57BL/6j to produce F1. The first screening to identify interesting reporter animals was performed on the F2 generation by administration of flagellin and measurement of luciferase activity.
activity in different organs (see below). Classical genotyping of mice was performed on tail genomic DNA using the following set of primers for control (Cpxm1-F: ACTGGGATCTTCTGAACTCTTTGGAC and Cpxm1-R: GATGTTGGGG-CACTGCTCATTCACC), 5' amplification of construct (D-F: GTGACAGGATGAGGCAGATTAATCAATGGG and D-R: CTTTACCAACAGTACCGGAATGCCAAG), and 3' amplification of construct (F-F: GCCCAGAAGGTACCTTGTATG and F-R: GTTTACGTCGCCGTCCGCTC). In some assays, genomic DNA was assessed using real-time quantitative PCR with the following primers specific for luc (LUC-F: TGAGTACTTCGAAATGTCCGTTC and LUC-R: GTATTCCAGCCCATATCGTTTCAT) and Actb (Actb-F: CGTCATCCATGGCGAACTG and Actb-R: GCTTCTTTGCAGCTCCTTCGT). Since Ccl20-luc-IRES-eyfp expression was stable over F1 to F8 (not shown), one can consider that the germline transmission of transgene is effective and not subjected to attenuation of expression.

**Figure 2. Characterization of Ccl20-luc-IRES-eyfp transgenic line.** Transgenic mice were treated with 5 μg flagellin or PBS i.v. and were sacrificed at indicated times. Organs were recovered for determination of luciferase activity or Ccl20 expression. A. Analysis of fusion expression in several tissues after flagellin stimulation. Luciferase activity was measured 4 h post-treatment. Results are expressed as relative luciferase activity using PBS-treated mice as reference. Data (n = 8) are given as mean ± SD and are representative of two experiments. B. Flagellin-specific upregulation of Ccl20 transcription in tissues. Ccl20 mRNA levels were measured in different organs by RT-qPCR 4 h post-treatment. Results are expressed as relative mRNA levels using to PBS treated mice as reference. Data (n = 5) are given as mean ± SD and are representative of two experiments. C. Kinetics of liver activation by flagellin as measured by fusion expression. Luciferase activity was measured in liver at different times after treatment. Results (n = 7) are expressed as in A. D. Kinetics of liver activation by flagellin as measured by mRNA levels. Ccl20 mRNA level measured by RT-qPCR at 4 h and 24 h. Results (n = 5) are expressed as in C.

doi:10.1371/journal.pone.0078447.g002
Transgenic animals were stimulated with heat-inactivated Salmonella enterica (10⁶ CFU) or flagellin from Salmonella typhimurium (5 µg/animal) (FLA-ST, InvivoGen, San Diego, CA, USA) via i.p. or i.n. routes for 4 h or as indicated.

Luciferase Assays

At indicated times, mice were sacrificed by cervical dislocation or pentobarbital injection and samples of liver, spleen, lung and small intestine were homogenized in lysis buffer (Luciferase Assay System, Promega, MA, USA) via i.p. or i.v routes for 4 h or as indicated.

Gene Expression Analysis

After stimulation, animals were sacrificed, organs recovered and homogenized in RA1 lysis buffer (GE Healthcare, Pittsburgh, USA) and total RNA was extracted with Nucleo Spin RNA II kit (Macherey-Nagel, Duren, Germany). Reverse transcription was performed using 200 U M-MLV reverse transcriptase (Invitrogen, USA), 0.5 mM dNTPs, 1 µM random primers, 20 U RNAse-OFF, 7.5 mM DTT, 1X reaction buffer and 400 ng of RNA in a final volume of 20 µl. The RT reaction was performed at 25°C for 10 minutes followed by 37°C for 60 minutes. For real-time PCR, SYBR® Green Master Mix (Invitrogen) was used. A standard protocol of 40 cycles of denaturation 15 sec at 90°C and annealing/extension 1 min 60°C amplification was used. Primers for pro-inflammatory CCL20 and mouse NF-κB were previously designed and described [20,33]. Relative mRNA expression of each chemokine was evaluated through the 2-ΔΔCt method and the β-actin gene was used as normalization gene [20].

In vivo Imaging Experiments

A comparative analysis of the effect of the two agonists of the innate immunity described before was done when administered via i.p. and i.v. in CCL20-luc-IRES-eyfp and NFκB-luc control mice of 8 to 12 weeks of age. Activity of the reporter gene was studied in different organs by in vivo imaging at 4, 8, 12 and 24 h post-stimulation, through injection of 150 mg/kg of luminogenic substrate for luciferase (D-Luciferin Firefly, Caliper Life Sciences, MA, USA), keeping the animal anesthetized with isoflurane (Isoforine, Cristalia, SP, Brazil) for 5 min in in vivo imaging analysis system (MS FX PRO, Carestream Health Inc, USA). Intensity of innate response activation in different organs was evaluated using the software Carestream molecular imaging v 5.3.3.17476.

For Ccl20-luc reporter mice, an imaging capture protocol of 5 min exposure was established, in comparison of a shorter protocol used for the NF-κB model. In both cases, a previous X-ray exposure followed by the bioluminescence filter was performed. Images obtained by each animal were regionalized (ROI) in three for Ccl20-luc-IRES-eyfp mice: thorax, upper abdomen and middle abdomen, and in five for NF-κB mice: head and neck, thorax, upper abdomen, middle abdomen and lower abdomen. For each ROI the software calculated net and mean intensity of photon emission per mm², plotting average ± SE for each treatment and acquisition time.

Results

Construction and Characterization of Mouse Ccl20 Reporter Fusions

With the aim to generate a reporter mouse strain, the Ccl20 gene promoter was linked to the Firefly luciferase gene luc, giving rise to a transcriptional fusion (Figure 1). The 1388 bp fragment that was used as the Ccl20 promoter contains 1377 bp upstream the initiation of transcription site of murine Ccl20 gene as defined previously by Fujie et al. [15] (Figure 1A-B). By transient transfection on human epithelial cell lines Caco-2 and HEK293, functional activity of reporter upon pro-inflammatory stimulation by the TLR5 agonist flagellin was confirmed (data not shown). The activity of the reporter fusion was next assessed in vivo using hydrodynamic shock injection followed by stimulation with flagellin (Figure 1C). This approach allowed a transient transfection of mouse liver cells with the plasmid(s) injected by the i.v. route. The flagellin stimulation was able to induce >100-fold the luciferase activity in liver in contrast to PBS-stimulation of transfected animals. These results are in accordance with previous reports indicating that flagellin increases liver pro-inflammatory transcription [29].

Additional experiments were performed to further characterize the activity of the reporter in response to flagellin stimulation. First, we defined that NF-κB site is essential to the activity of the Ccl20 promoter using the constructs of Fujie et al. [15]. Indeed, reporter fusions deleted (–85/+6) or mutated within the NF-κB site were totally abrogated in the capacity to respond to flagellin (Figure 1D). We also generated a fusion harboring a mutation in the Ets-1 site (–147). This mutation did not affect the response to flagellin, thereby indicating that the liver response is dominated by pro-inflammatory NF-κB signaling (Figure 1D). Next, we assessed the role of direct TLR5 stimulation on liver luciferase activity (Figure 1E-F). To this aim, Th5-specific shRNA encoding plasmids were co-administered with the Ccl20-luc reporter construction by hydrodynamic shock and mice were stimulated with flagellin. Under these conditions, impairment of liver TLR5 expression abrogated completely the reporter expression in response to flagellin (p<0.001) but not LPS (Figure 1E).

In contrast, the control (scramble) shRNA plasmid still displayed inducible luciferase activity independently of stimulation. Finally, the requirement of TLR5 signaling was studied using the mouse counterpart of human dominant negative TLR5, e.g. mTLR5D33stop. Injection of the plasmid encoding mTLR5D33stop was associated to >90% decrease response to flagellin (Figure 1F). In conclusion, the direct flagellin-TLR5 interaction on liver cells
**CCL20-Luciferase Reporter Mouse Strain**

**Figure A**

- NFκB-luc
- Ccl20-luc

**Figure B**

- Head
- Torax
- Upper abdomen
- Middle abdomen
- Lower abdomen

**Figure C**

- CCL20-luc (flagellin i.v. treatment)
- NFκB-luc (flagellin i.v. treatment)
- CCL20-luc (Salmonella i.v. treatment)
seems to be the major pathway that control Ccl20 early expression upon systemic flagellin delivery.

Generation and Functional Analysis of Ccl20 Reporter Mice

The pCcl20-luc plasmid was further engineered to incorporate the IRES-eyfp cassette downstream of luc cDNA. The resulting plasmid pCcl20-luc-IRES-eyfp was found to respond as effectively as pCd20-luc in the hydrodynamic shock model (Figure 1G). However, we were not able to detect increase of EYFP fluorescence in liver of flagellin-stimulated mice (data not shown). The generation of a stable reporter mouse strain by transgenesis was then started using the pCcl20-luc-IRES-eyfp DNA at the Institut Clinique de la Souris. Among several PCR positive founders, 5 clones were selected for expansion as F1 generation and characterization. The F2 mice derived from three selected F1 founders (clones 61, 68 and 72) were analyzed for the relative levels of luciferase activity in various organs upon stimulation with flagellin (Figure S1). The animals derived from clone 61 were selected for further expansion and analysis. In several tissues of C20-luc mice derived from clone 61, the luciferase activity was low or close to background signals at the steady state or after mock treatment (data not shown). In contrast, the flagellin treatment was consistently able to induce the expression of reporter fusion as detected by the increase of the luciferase activity (Figure 2). The maximal activation was associated with the liver although significant luciferase activity was observed in lung, spleen, ileum and colon (Figure 2A). However, C20 reporter fusion expression was almost undetectable in blood, duodenum and jejunum. These results highly correlated with the levels of C20 transcripts (Figure 2B). Indeed, liver C20 mRNA levels were increased 140±30 fold upon flagellin stimulation, whereas the increase was 30±8 and 8±3 fold in lung and ileum, respectively. The fusion expression in liver was also assessed at different times after flagellin stimulation. We observed a fast induction of luciferase activity (200±50 fold) 4 h post-treatment associated to a gradual decrease at later time points (Figure 2C). This rapid activation correlates with a rise in C20 mRNA levels of around 150 fold at 4 h post-treatment that return to basal levels at 24 h (Figure 2D). Interestingly, qPCR analysis of genomic DNA derived from F1 to F7 generations revealed that the transgene copy number is stable as defined by relative levels of genomic DNA specific for luciferase and beta actin genes (data not shown). Altogether, our data show that the C20-luc reporter mice represent an appropriate model to monitor in real time the activity of C20 promoter.

As CCL20 can be induced at various mucosal sites in response to pro-inflammatory stimuli [14], the activation of the reporter in lung mucosa after intranasal and systemic flagellin administration was compared. A significant induction of the luciferase activity was observed in both cases (Figure 3A).

Real Time Bio-imaging of Ccl20 Promoter Activity

A major use of luciferase reporter fusion is the real time monitoring of gene expression by bio-imaging [34]. The C20-luc reporter mouse was used for in vivo imaging studies at different times after systemic stimulation, i.e. i.v./i.p. administration of flagellin or heat-inactivated Salmonella (Figure 4). The NF-kB-luc reporter mice strain that harbors artificial promoter with tandem NF-kB motifs was used as control to evaluate the intensity of the pro-inflammatory response [35]. In 4 h-treated NF-kB-luc mice, flagellin induced a strong luciferase activity in the abdominal region (liver and gut) and the respiratory tract compared to untreated mice (Figure 4A). In the same conditions, luciferase activity was detected in C20-luc-IRES-eyfp mice but was restricted to the abdomen. The system used for in vivo imaging allows the quantification of the luminescence emission in defined areas. Different anatomic regions were defined to get a comparative analysis of regionalized activation (Figure 4B). The kinetic analysis performed in C20-luc-IRES-eyfp mice showed that upon flagellin stimulation, the maximal activity was mainly limited to upper abdomen, with a peak at 4 h and a progressive decrease to basal levels at 24 h. In the middle abdomen, a lower luminescence and a different kinetics (maximum at 8 h) was observed (Figure 4C). The magnitude of luciferase activity was more important in NF-kB-luc mice than in C20-luc-IRES-eyfp mice, with significant differences in kinetics and anatomical expression. When C20-luc-IRES-eyfp mice were stimulated with heat-inactivated Salmonella, luciferase expression patterns were found to be similar to flagellin-induced treatment although the global magnitude was higher (Figure 4C and data not shown). Altogether these results showed that the C20-luc-IRES-eyfp mice could be used for in vivo real time imaging. Furthermore, the C20-luc-IRES-eyfp mice showed a specific pattern of expression to systemic challenge characterized by specific activation in the liver.

Discussion

In the present work we have characterized some features of the C20 promoter and generated the transgenic reporter mouse line C20-luc-IRES-eyfp that recapitulates the transcriptional activity of C20 in various organs. We have shown that the transgenic mouse line can be used for in vivo imaging of C20 response, representing a novel tool for the examination of physiopathological situations that may include the C20-CCR6 axis as a major component.

From the initial descriptions of mouse C20 and human CCL20 promoters, the conservation of several features was appreciated. First, the proximal NF-kB binding site was found to be critical to the promoter activity in steady state (constitutive) and pathological (inducible) conditions [15,16,36,37]. We have previously shown that the NF-kB binding site can bind different NF-kB combinations [19]. When the canonical p65/p50 pro-inflammatory NF-kB dimer binds the site, a highly inducible C20 response is triggered as a feature of pro-inflammatory stimuli. Besides, the same site can bind RelB/p52 NF-kB that is involved in the NF-kB alternative pathway function, thereby promoting a sustained
transcription of CCL20 encoding gene after engagement of several TNFR superfamily members such as LTβR or TWEAKR [20,30]. This pathway has been involved in the constitutive expression of CCL20, particularly in sites such as the follicle associated epithelium in mucosal sites. By targeted mutations, Kwou et al. showed that ETS binding sites in the proximal promoter region were critical for CCL20 expression in intestinal epithelium [39]. In this work, we have shown that in the mouse promoter the NF-κB binding site is critical for in vivo liver inducible activation; however the ETS site positioned in −156/−147 is dispensable to this activity. Among the various ETS transcription factors, the intestinal epithelium specific ESE-1 transcription factor was shown to bind the CCL20 promoter [39] and contribute to the inducible expression in intestinal epithelium [20]. The high inducible activity observed in liver may be contributed by other specific tissue transcription factors that are not acting on ESE-1 binding site and have not yet been identified.

In concordance with previous reports [37,40], we have observed that CCL20 expression upon different pro-inflammatory stimuli is maximal in liver. LPS, as well as proinflammatory cytokines such as TNFα, are potent inducers of liver CCL20 expression [37,41]. Since flagellin stimulation may trigger the production of TNFα by myeloid cells [42], it may be expected that by systemic administration of flagellin, CCL20 expression can be contributed initially by direct PRR-PAMP interaction and secondary by indirect activation by cytokines such as TNF that are secreted upon PAMP stimulation. However, we have shown that impairing TLR5 activity in liver in vivo abrogates completely the CCL20 liver response, indicating that direct PRR-PAMP interaction is the main trigger of CCL20 liver expression in this case (Figure 1). The kinetics of CCL20 expression observed in vivo upon flagellin or heat-killed salmonella stimulation supports this hypothesis (Figures 2 and 4).

We have shown that the transgenic reporter mice are suitable for in vivo imaging experiments. The capacity of doing a whole body scanning for luciferase activity allows determining that upon proinflammatory stimulation, CCL20 production is maximal in the liver, in coincidence to what was shown by determination of enzymatic activity and mRNA in a case by case approach (Figure 2). Other organs that were not included in the screening such as skin or thymus were not detected as major sites of CCL20 enzymatic activity and mRNA in a case by case approach.

We have shown that ETS binding sites in the proximal promoter region are critical for CCL20 expression in intestinal epithelium by genome profiling and microdissection. Gastroenterology 129: 321–327. 5. Tanaka Y, Imai T, Baba M, Ishikawa I, Uehira M, et al. (1999) Selective expression of liver and activation-regulated chemokine (LARC) in intestinal epithelium in mice and humans. European Journal of Immunology 29: 635–642.

References

1. Le Y, Zhou Y, Iribarren P, Wang J (2004) Chemokines and chemokine receptors: their manifold roles in homeostasis and disease. Cell Mol Immunol 1: 95–104.
2. Ansel KM, Cytryn JG (2001) Chemokines in lymphopoiesis and lymphoid organ development. Current Opinion in Immunology 13: 172–179.
3. Mootz A, Van Donme J, Proost P (2012) Overview of the mechanisms regulating chemokine activity and availability. Immuno Lett 145: 2–9.
4. Andreu P, Rubio M, Serró F, Mansourian R, Michetti P, et al. (2005) Novel markers of the human follicle-associated epithelium identified by genomic profiling and microdissection. Gastroenterology 129: 321–327.
5. Tanaka Y, Imai T, Baba M, Ishikawa I, Uehira M, et al. (1999) Selective expression of liver and activation-regulated chemokine (LARC) in intestinal epithelium in mice and humans. European Journal of Immunology 29: 635–642.

CCL20-Luciferase Reporter Mouse Strain

The in vivo imaging approach allowed us to perform a simple comparison of activity against the NF-κB-luciferase reporter mouse strain that harbors an artificial promoter containing tandem NF-κB binding sites [35] and was widely used to monitor in vivo NF-κB activation [44]. Although CCL20 expression is highly dependent on NF-κB activation, the luciferase activity observed was not equivalent in C20-luc and NFκB-luc mice. For instance, NFκB-luc mice showed high luciferase activity at steady state and widespread tissue expression upon pro-inflammatory stimulation compared to C20-luc-IREs:LUC mice. The differences observed between the two reporter mice strains reflect the activity of their respective promoters. The native mouse C20 promoter contains several regulatory regions that respond to several transcriptional modulators including NF-κB. As described previously, specific combinations of transcription factors are required to activate C20 expression with respects to nature of tissue/cell and environmental signals.

We have shown that the C20-luc-IREs:LUC reporter mice recapitulate CCL20 expression in tissue- and stimuli-specific manner. This mouse model may constitute an important tool for research on liver inflammation and on any diseases where CCL20 may constitute an important element of the pathophysiology, such as NASH [45] autoimmune hepatitis [46] sterile inflammation and atherosclerosis [47] or ischemia reperfusion injury [48].

Supporting Information

Figure S1 Functional screening on several C20-luc-IREs:LUC mice lines originated from different founders. C20-luc-IREs:LUC mice from founders #61, #68 and #72 were treated i.p. with 5 µg flagellin or PBS. Tissues were sampled 4 h post-treatment and luciferase activity was determined on tissue lysates. Results are expressed as mean luminescence (n = 5–7).

Acknowledgments

We thank Dr. R. Locksley for IRES-LUC plasmid.

Author Contributions

Conceived and designed the experiments: MR, JCS. Analyzed the data: MC LVM MR JCS AMF. Contributed reagents/materials/analysis tools: MC MR JCS AMF. Wrote the paper: MC JCS MR. Conducted most of the in vivo experiments with the reporter mouse strain: MC. Conducted most of the characterization of the modulation of CCL20 promoter response in vitro and in vivo: LVM. Conducted the cloning of CCL20 promoter: JT. Conducted the characterization of the CCL20 promoter in vitro: DC. Conducted the characterization of the in vivo response of the reporter mouse strain: AE. Conducted the in vivo experiments with the reporter mouse strain: MR AMF. Conducted the characterization of the in vivo response of the reporter mouse strain: JCS.

References

1. Le Y, Zhou Y, Iribarren P, Wang J (2004) Chemokines and chemokine receptors: their manifold roles in homeostasis and disease. Cell Mol Immunol 1: 95–104.
2. Ansel KM, Cytryn JG (2001) Chemokines in lymphopoiesis and lymphoid organ development. Current Opinion in Immunology 13: 172–179.
3. Mootz A, Van Donme J, Proost P (2012) Overview of the mechanisms regulating chemokine activity and availability. Immuno Lett 145: 2–9.
4. Andreu P, Rubio M, Serró F, Mansourian R, Michetti P, et al. (2005) Novel markers of the human follicle-associated epithelium identified by genomic profiling and microdissection. Gastroenterology 129: 321–327.
5. Tanaka Y, Imai T, Baba M, Ishikawa I, Uehira M, et al. (1999) Selective expression of liver and activation-regulated chemokine (LARC) in intestinal epithelium in mice and humans. European Journal of Immunology 29: 635–642.
6. Sierro F, Dubois B, Coste A, Kaisermani D, Kraehenbuhl JP, et al. (2001) Flagellin stimulation of intestinal epithelial cells triggers CCL20-mediated migration of dendritic cells. Proc Natl Acad Sci U A 98: 13722–13727.
7. Chiappa M, Rusciano M, Huang AY, Germain RN (2006) Dynamic imaging of dendritic cell extension into the small bowel lumen in response to epithelial cell TLR engagement. J Exp Med 203: 2841–2852.
8. Jenner RG, Young RA (2005) Insights into host responses against pathogens from transcriptional profiling. Nat Rev Microbiol 3: 281–294.
9. Boldrick JC, Alizadeh AA, Diefen M, Dudoit S, Liu CL, et al. (2002) Stereotyped and specific gene expression programs in human innate immune responses to bacteria. Proc Natl Acad Sci U A 99: 972–977.
27. Gewirtz AT, Navas TA, Lyons L, Godowski PJ, Madara JL (2001) Bacterial protein-3alpha/CCL20 gene transcription via the NF-kappa B receptor 5- and lymphotoxin beta receptor-dependent epithelial Ccl2 0 kDa luciferase reporter gene expression involves the same NF-kappaB binding site but distinct NF-kappaB pathways and dynamics. Biochem Biophys Acta 1789: 386–394. Epub 2009 Mar 12.

28. Didierlaurent A, Ferrero I, Otten LA, Dubois B, Reinhardt M, et al. (2004) Flagellin promotes myeloid differentiation factor 8-dependent development of Th2-type response. J Immunol 172: 6922–6930.

29. Burdelya LG, Brackertz CM, Kojouharov B, Glinin, H, Leonova KI, et al. (2013) Central role of liver in anticancer and radioprotective activities of Toll-like receptor 5 agonist. Proc Natl Acad Sci U A S A 110: E1857–1866.

30. Anderle P, Rumbio M, Sierro F, Mansourian R, Michetti P, et al. (2005) Novel markers of the human follicle-associated epithelium identified by genomic profiling and microdissection. Gastroenterology 129: 321–327.

31. Reinhardt RL, Hong S, Kang SJ, Wang ZE, Lockley RM (2006) Visualization of IL-12/23p40 in vivo reveals immunostimulatory dendritic cell migrants that promote Th1 differentiation. J Immunol 177: 1618–1627.

32. Lewis DL, Hagstrom JR, Loomis AG, Wolff JA, Herweijer H (2002) Efficient delivery of siRNA for inhibition of gene expression in postnatal mice. Nat Genet 32: 107–108.

33. Roberts R, Moreno G, Bottero D, Gaillard ME, Fingermann M, et al. (2008) Outer membrane vesicles as archetypal vaccine against pertussis. Vaccine 26: 4639–4646. Epub 2008 Jul 4618.

34. Polk M, De A (2012) Bioinformatics based in vivo screening technologies. Curr Opin Pharmacol 12: 592–600.

35. Carlsen H, Moskaug JO, Fromm SH, Blomhoff R (2002) In vivo imaging of NF-kappaB activity. Journal of Immunology 168: 1441–1446.

36. Battaglia F, Delfino S, Merello E, Puppo M, Piva R, et al. (2008) Hypoxia transcriptionally induces macrophage-inflammatory protein-salpha/CCL-20 in primary human mononuclear phagocytes through nuclear factor (NF-kappaB). J Leukoc Biol 83: 648–662.

37. Sugita S, Kohno T, Yamamoto K, Imaizumi Y, Nakajima H, et al. (2002) Induction of macrophage-inflammatory protein-salpha gene expression by TNF-dependent NF-kappaB activation. Journal of Immunology 168: 5621–5628.

38. Saitoh T, Nakayama M, Nakano H, Yagita H, Yamamoto N, et al. (2003) TWEAK induces NF-kB2 p100 processing and long-lasting NF-kB activation. J Biol Chem 278: 873–874. Epub 2002 Oct 3031.

39. Hieshima K, Imai T, Odenakker G, Van Damme J, Kusuda J, et al. (1997) Molecular cloning of a novel human CC chemokine liver and activation-regulated chemokine (LARC) expressed in liver. Chemotactic activity for lymphocytes and gene localization on chromosome 2. J Biol Chem 272: 5846–5853.

40. Iwamoto S, Kidoo M, Aoki N, Nishiura H, Marnoka R, et al. (2013) TNF-alpha is essential in the induction of fatal autoimmune hepatitis in mice through upregulation of hepatic CCL20 expression. Clin Immunol 146: 15–25.

41. Sanderson CJ, Moore DA, 3rd, Williams IR, Gewirtz AT (2008) Both radioresistant and hemopoietic cells promote innate and adaptive immune responses to flagellin. J Immunol 180: 7184–7192.

42. Doyle TC, Burns SM, Contag CH (2004) In vivo bioluminescence imaging for integrated studies of infection. Cell Microbiol 6: 303–317.

43. Kelkar M, De A (2012) Bioinformatics based in vivo screening technologies. Curr Opin Pharmacol 12: 592–600.

44. Anderle P, Rumbio M, Sierro F, Mansourian R, Michetti P, et al. (2005) Novel markers of the human follicle-associated epithelium identified by genomic profiling and microdissection. Gastroenterology 129: 321–327.

45. Larrain S, Rinella ME (2012) A myriad of pathways to NASH. Clin Liver Dis 16: 525–548.

46. Della Corte C, Sartorelli MR, Comparcola D, Alterio A, Giorgio V, et al. (2012) Liver injury is induced by oncofetal antigen TWEAK. J Biol Chem 1: 1.

47. Kubes P, Mehal WZ (2012) Sterile inflammation in the liver. Gastroenterology 143: 1158–1172.

48. van Gelen RF, van Gulik TM, Heger M (2012) The sterile immune response during hepatic ischemia/reperfusion. Cytokine Growth Factor Rev 23: 69–84.