SCREENING OF ASPARAGUS OFFICINALIS L. SEEDS FOR OCCURRENCE AND PLOIDY OF TWIN EMBRYOS

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We used germination tests to assess the frequency of polyembryony in 9 asparagus cultivars with a high propensity to produce double embryos with different ploidy levels: Alpha, Andreas, Boonlim, Cipres, Eposs, Helios, Limbras, Ravel and Sartaguda. Twin embryos inside a single seed were found in 3 cultivars: Eposs 2n, Ravel 2n and Sartaguda 2n, at 0.60% frequency (15 seeds with twin embryos out of 2500 seeds). Of 30 obtained seedlings, 14 were separated diploid-diploid twins, 6 were conjoined diploid pairs, 8 were separated diploid-haploid and 2 were diploid-haploid pairs conjoined in the hypocotyl region. Some embryos showed unilateral dominance of one embryo (size and shape). The haploid status of the smallest embryo was confirmed by chromosome number (n=x=10) and flow cytometry (nuclear C DNA amount 1.95 pg). The haploid obtained in this manner possessed enough vegetative vigor to undergo chromosome doubling.

**Key word:** Polyembryonic seeds, haploid seedlings, Asparagus officinalis, flow cytometry, chromosome number, morphological analysis.

INTRODUCTION

Asparagus officinalis L. (Liliaceae) belongs to a large genus containing 200 perennial species. Numerous species are used for ornamental purposes [A. densiflorus (Kunth) Jessop cv. Plumosus; A. densiflorus cv. Sprengeri and A. falcatus L.] but the most important agricultural crop is A. officinalis L. (Valdes, 1964). About 100 different edible cultivars are grown in Europe and many genotypes are held in local collections (Knaflowski, 1996) and in a large collection of doubled haploids (Ricardi et al., 2011). A. officinalis is a dioecious species with homomorphic sex chromosomes. Female plants are homogametic (XX), whereas males are heterogametic (XY). The high yield and homogeneity of male cultivars (with XY karyotype) makes them particularly desirable; molecular markers have been used to identify them prior to flowering (Gebler et al., 2007). Some morpho-agronomic traits of asparagus as a dioecious and perennial plant have been described, such as the structure of shoots, inflorescences, fruits and seeds (Ellison, 1986; Stajner et al., 2002; Ito et al., 2007). The diploid chromosome number of A. officinalis is 2n = 2x = 20 but some cultivars are also tetraploid 2n = 4x = 40 (Randall and Rick, 1945; Kunitake et al., 1998). Breeding programs have created the triploid cv. Hiroshima Green (2n = 3x = 30), which has larger shoots and a longer harvest period than diploid cultivars (Ozaki et al., 2004; Moreno et al., 2006).

Asparagus officinalis is becoming increasingly important in agriculture. Breeding efforts to introduce disease resistance from wild relatives of cultivars are therefore needed (Marcellan and Camadro, 1999). In crop breeding programs the role of haploids in creating homozygotic dihaploids is particularly important (Malepszy, 2009; Riccardi et al., 2011). Many positive features can be captured and valuable traits can be obtained through double haploidization and multiple cross combinations with male asparagus plants (Thevenin, 1968; Ellison, 1986; Kunitake et al., 1998) but such processes are labor-intensive, time-consuming and often inefficient. Haploids created by androgenesis often display greater variability, lower fitness and heterozy-
Haploids have been obtained in situ parthenogenetically from unfertilized egg cells or after apogamic processes of embryo development from antipodal and synergid cells. The subsequent development of haploid embryos is highly dependent on the endosperm (Zenkteler and Nitzsche, 1984). Monoploids in *Zea* and polyhaploids in *Triticum*, *Solanum* and *Gossypium* were obtained by apogamy, the suitability of which for breeding was evaluated after double haploidization or stimulation by pollination of pollen inactivated after radiation with UV or γ rays (Malepszy et al., 1989). Gynogenic haploids of *Beta*, *Helianthus*, *Oryza* and *Allium* have been successfully produced in breeding programs (Forster et al., 2007). Haploidization also takes place by somatic reduction of chromosomes during mitosis of hybrid cells a few days after double fertilization, which subsequently eliminates male chromosomes from the hybrid embryo and from the endosperm. The main cause of this is loss of balance between the genomes of both partners of distant crosses, as has been well documented in crosses of *Hordeum vulgare × Hordeum bulbosum* (Zenkteler and Straub, 1979).

In this study we assessed the frequency of natural polyembryony and the possibility of obtaining haploid plants from screened germination of seeds of asparagus cultivars growing in Polish plantations.

### MATERIALS AND METHODS

#### PLANT MATERIAL

Seeds from 9 randomly selected cultivars of asparagus obtained from the collection of the Department of Vegetable Crops of Poznań University of Life Sciences were analyzed after germination tests for the prevalence of twin embryos (Tab. 1), between 2006 and 2008. The germination data are means ± SD of three replicates.

We made ovule squash trials to find the best screening method and establish the origin of twin embryos. Immature fruits were harvested and three developmental stages of cv. Eposs fruits (equatorial diameter of 0.3, 0.4 and 0.6 cm) compared in order to select the optimal stage for ovule isolation (Fig. 1a–c). Stage 1 (Fig. 1a) proved to be the best size for squashing ovules in 1% acetocarmine. The ovules usually contained a visible single embryo; very rarely we found twin embryos in the same ovule (Fig. 1d).

The germination tests were performed with seeds sown in semi-sterile conditions at 23±2°C, RH 60%, with 1 week of dark incubation. The seeds were surface-disinfected in 70% (v/v) ethanol for 1 min, rinsed five times with sterile distilled water and sown on wet filter paper in 20 cm Ø Petri dishes. The seeds were morphologically screened 14 and 20 days after sowing. The frequency per cultivar and morphology of twin seedlings in one seed were recorded.

#### ROOTING AND ACCLIMATIZATION

After germination the twin seedlings were separated from seeds, transplanted to plastic pots with a mixture of autoclaved soil with sand and perlite (1:1:0.3/l), then watered and fertilized once a week with 1/2 MS macrosalt solution (Murashige and Skoog, 1962). The potted plantlets were covered with polyethylene bags to maintain high humidity. After 1 week the covers were removed progressively. About 95% of diploid and all haploid plantlets survived and showed normal growth inside the culture room (23°C, 60% RH, 16 h photoperiod). Two

### TABLE 1. Frequency of polyembryonic seeds in asparagus cultivars screened in germination test

| Cultivars | Ploidy | Sex* | Number of seeds | Monoearyony % | Number of seeds with twin embryos |
|-----------|--------|------|-----------------|---------------|----------------------------------|
| Alpha     | 2n     | ♂/♀ | 425             | 83.7 ± 9.1b   | 0                                |
| Andreas   | 2n     | ♂    | 640             | 90.0 ± 6.5    | 0                                |
| Boonim    | 2n     | ♂    | 510             | 91.8 ± 6.1a   | 0                                |
| Cipres    | 2n     | ♂    | 435             | 91.1 ± 6.9    | 0                                |
| Eposs     | 2n     | ♂/♀ | 1500            | 84.6 ± 7.3    | 8                                |
| Helios    | 2n     | ♂/♀ | 400             | 88.0 ± 7.6    | 0                                |
| Limbras   | 2n     | ♂    | 800             | 89.1 ± 8.1    | 0                                |
| Ravel     | 2n     | ♂    | 450             | 92.4 ± 6.2a   | 5                                |
| Sartaguda | 2n     | ♂/♀ | 550             | 90.8 ± 6.6    | 2                                |
| Total     |        |      | 5710            |               | 15                               |

Data are means ± SD. Different letters indicate significant differences (P< 0.05); *Dioecious or male.
months after planting in garden conditions, one spear was removed from each acclimatized plant for flow cytometry (FCM) analysis.

NUCLEAR DNA CONTENT

Juvenile phylloclades collected from twin seedlings were assessed by flow cytometry for identify the haploids and their nuclear DNA amount. *Asparagus* samples together with an internal standard of *Pisum sativum* (2C value 9.11 pg) were chopped with a razor blade in an isolation buffer according to Doležel et al. (1989). After filtering the suspension through 30 μm nylon mesh, measurements were made with a Partec PAS flow cytometer (Partec GmbH, Germany). The UV spectrum excited with an HBO 100W/2 lamp was measured with a GG 435 long-pass filter. Partec software (DPAC V2.1.) was used to determine the G1/G0 peaks of the samples.

CHROMOSOME COUNTING

Root tips of asparagus seedlings 0.5 cm in length were fixed in cool (+4°C) AA solution (96% ethanol and glacial acetic acid, 3:1) for 24 h, then hydrolysed for 15 min in 1N HCl at 60°C and stained with 1% acetocarmine (w/v) for 25 min. Meristematic tissues were squashed and the chromosomes in the metaphase plate was counted and photographed under a Carl Zeiss Axiostar microscope with a digital camera.

RESULTS

ANALYSIS OF SQUASHED OVULES

Only 3 of the 500 immature ovules of cv. Eposs (stage 1, Fig. 1a) that we squashed, acetocarmine-stained and checked by LM contained twin pairs of embryos. In the majority of ovules the embryo sacs contained a single embryo; only 0.6% of the ovules contained double embryos in the same embryo sac.
The frequency of ovules containing no embryo at all (although their endosperm developed normally) was 19%. From the globular twin embryos (three pairs) obtained from immature fruits, two pairs were of equal size and one pair had an embryo that was larger than the other (Fig. 1d). All of the twin embryos were smaller than the normal single ones that developed at the same time.

**MORPHOLOGICAL ANALYSIS**

The seedlings obtained from germination tests were screened in Petri dishes. The seeds of only three cultivars (Eposs, Ravel, Sartaguda) had some double seedlings (15 twins from 2,500 seeds, 0.6%). Diploids constituted 83.3% of all the plants derived from twin seedlings, and the remainder (16.7%) were haploids. Twin seedlings are shown in Figure 2a–d. In the 450 screened seeds of cv. Ravel (Fig. 2a,b) only 5 pairs of double seedlings were found. The number of double seedlings was highest in cv. Eposs; many fewer were found in Ravel and Sartaguda, and none in the remaining cultivars (Tab. 1). Eleven of the pairs were separated (each plantlet had its own cotyledon and root), 3 pairs were conjoined in the hypocotyl region, and 1 pair had a common hypocotyl and shared one primary root (Fig. 2c). Two equal-sized pairs were fasciated.
at 3–5 cm along the length of their young phylloclades (Tab. 2). After 6 weeks the conjoined diploid twins separated from each other.

The growth and survival of diploid plantlets derived from twins were high, yielding 20 strong seedlings. The haploid seedlings grew less vigorously and represented their characteristic root and shoot phenotype. Two haploid and some diploid seedlings were lost during acclimatization from pots to the garden.

NUCLEAR DNA CONTENT

Relative nuclear DNA content as revealed by the position of the DNA distribution peaks indicated nuclei in the G1 phase of the cell cycle. The peaks depended on the cultivar analyzed. FCM revealed that different-sized pairs had different ploidy levels, reflected in plantlet morphology. The DNA content of 10 haploid/diploid seedlings ranged from 3.71 to 3.75 pg in the largest seedlings and from 1.89 to 1.95 pg in the smallest. The majority of seedlings derived from equal-sized twin embryos had nuclear DNA content (2C) ranging from 3.68 to 3.77 pg. The two diploid twins did not differ in ploidy level (Tab. 2).

SEEDLING CHROMOSOME NUMBER

Fifteen polyembryonic seeds of asparagus gave rise to thirty twin seedlings (Tab. 2). Only five of the pairs had mixed ploidy level (diploid and haploid), and the rest were diploid only. Metaphase plate analysis of root meristem cells revealed that both (diploid and haploid) chromosome sets were medium-sized, uniform, without morphological differences between them. The majority of double seedlings were dipo- loids with somatic chromosome number 2n = 2x = 20 (Fig. 2d,a; Tab. 2). Only shorter and thinner roots from small seedlings were found upon cytological examination to be essentially haploids containing n = x = 10 chromosomes in their somatic cells (Fig. 2d,b; Tab. 2).

DISCUSSION

In this evaluation of the possibility of obtaining haploid Asparagus plants by separating them from natural polyembryonic seeds we found the frequency of twins to be cultivar-dependent. Our results are much lower than those obtained in lemon by Perez-Tornerod and Porras (2008) (1–3 embryos per seed), tangerine (6–10 embryos) and orange (10–15 embryos), whose seeds are up to 43% polyembryonic (Moreira et al., 1947). Broad germination screening of seeds has also been performed in the Allium genebank for 92 species; polyembryony was detected in 26 species and tended to be species-specific. High rates of twin seedlings, up to 32%, have been found in Allium splendens 2n = 4x = 48. In the genus Allium the tendency for twin embryo formation may be higher because of the high ploidy level of the species (Specht et al., 2001).

The number of twin embryos per seed was highest in cv. Eposs (8/1500, 0.50%). Germination tests of cv. Mary Washington 500W (Uno et al., 2002) revealed 0.34% double embryos (34/9925). Other authors have reported frequencies of twins obtained from various asparagus genotypes of 0.13–3.54% (Randall and Rick, 1945) 0.22%, and 1.79% in a haploid (Thevenin, 1968). Those authors concluded that Asparagus officinalis has only a moderate tendency to produce polyembryonic seeds (Webber, 1940; Randall and Rick, 1945). Very few haploid plants have been obtained by this method. Their poor viability and high mortality present a major obstacle in breeding efforts.

In a comprehensive review of polyembryony in Dactylis, Gossypium, Nicotiana, Phleum, Poa, Solanum and Triticum, Webber (1940) stated that there are three origins of polyembryony: nucellar, monozygotic and dizygotic. Nucellar polyembryony is a well known phenomenon in Citrus (Frost, 1926; Moreira et al., 1947). The process starts outside the embryo sac: a few nucellar cells next to the micropyle undergo mitotic divisions and give rise to adventitious embryos, and the embryos develop into plants genetically identical to the mother plant. Such a reproductive strategy assures the preservation and wide distribution of the maternal plant genotype (Zenktele and Guzowska, 1967). This phenomenon would have important evolutionary significance as a mechanism of adaptation and speciation (Ito et al., 2007).

Asparagus frequently has been the subject of research on the nature and origin of twin haploid-diploid embryos (Ellison, 1986; Kunitake et al., 1998; Randall and Rick, 1945; Uno et al., 2002). We can only speculate about the origin of additional embryos in polyembryonic seeds of Asparagus. Because polyembryony occurs at low frequency it has been extremely difficult to target the earliest stage of twin embryo development by microtome ovule sectioning. In our attempt to locate embryos at an immature stage of ovule development by the squash method we aimed to avoid the risk of early abortion or starvation of the haploid during competition with the second, larger embryo. We do not know whether these were mono- or dizygotic twins. It is not easy to distinguish morphologically nucellar from zygotic seedlings.

In asparagus, sex is determined by heterozygous Mm (male) or homozygous recessive mm (female) genes. The progeny of selfed andromonoecious plants segregate 1:2:1: mm (female), Mm
(male) and MM (supermale of great importance for its yield, longevity and resistance to diseases) (Riccardi et al., 2011). When haploids are obtained from microspores, mm and MM plants can be formed and the hybrids between them are also male (Doré 1990). When haploids are extracted from seeds with twin embryos only female lines can be created.

Following the results of others (Thevenin, 1968), we suggest that twins may originate (i) from a fertilized egg cell that split (2n/2n), (ii) one from the egg cell and the second as an adventitious embryo from the nucellus or integument (2n/2n), or (iii) from the fertilized reduced egg cell and simultaneously from an apogamously developing element of the embryo sac (2n/n). A special version of twin embryo formation results from meiotic defects which often give rise to unreduced gametes which may be involved in polyembryony. When chromosome pairing and recombination during meiotic prophase are disrupted, some meiotic mutants with unreduced gametes are produced (Brownfield and Köhler, 2011). Such gametes may participate in fertilization and lead to an attempt at embryo multiplication.

Another possibility for twin formation is early cleavage at the proembryonic stage, resulting in identical monozygotic plantlets, as in Theobroma cacao (Martinson, 1972). An extremely rare case is the formation of two gametophytes within single ovules, leading to dizygotic development of twins differing in genotype when the twin embryos arise by additional fertilization of synergids or antipodal cells (in Allium: Specht et al., 2001). Two-embryonic seeds were induced chemically by treatment of pepper flowers with 2,4-D or IBA (0.001% water solutions) separately or combined with BAP; both growth regulators had a marked effect (1.41% twins) on the increase in frequency of additional embryos (Nowaczyk and Nowaczyk, 1996; Jędrzejczyk and Nowaczyk, 2009). Ascorbic acid (Asc) injected into ovaries (50 μl on the first two days after pollination) promotes proembryo cell division and regulates cell polarity, giving rise to polyembryony and polycotyly in Nicotiana tabacum cv. Xanthi; the high level of dehydroascorbate reductase (DHAR), which recycles Asc, induces monozygotic twinning (Chen and Gallie, 2012).

The female gametophyte of asparagus conforms to the eight-nucleate Polygonum type. The mature embryo sac is asymmetrical. The micropylar end, which contains a three-celled egg apparatus, is much larger than the chalazal end, which is almost filled by the three antipodal cells (Webber, 1940). It is the antipodal cells which often give rise to the additional embryos in the early stage of polyembryonic development in the isolated embryo sac of Allium tuberosum (Specht et al., 2001). Twin progeny, one haploid embryo and the other diploid, developed simultaneously in the ovules of lily and tobacco hybrids; twins were found arising from a zygote and from a synergid stimulated to parthenogenetic divisions and to haploid plantlet development (Cooper, 1943).

Nowaczyk and Nowaczyk (2006) made an original attempt to explain the origin of twin embryos in Lycopersicum. Their experiments were based on segregation of marker characters in the F₁ and F₂ generations of hybrids. The presence of different recessive characters in each of the twins revealed adventitious embryo formation and splitting in tomato polyembryony. Most of the gross structural alterations involved in cleavage probably occurred during seed germination, although the process may also start in earlier stages of embryo development.

Interspecific hybrids of Carthamus palaestinus and C. tinctorius showed genetic linkage between stem fasciation and twin embryo seed development in F₁ (Singh et al., 2010). Mutation of the TWN1 gene disrupts suspensor differentiation and development in early embryogenesis in Arabidopsis. Developmental suspensor mutants give rise to a high percentage of seeds containing twin embryos (Vernon et al, 2001).

Ovules with an unequal pair of embryos contain two ontogenetically different types of embryo, one more developed and the other smaller (Fig. 1d). In early development one of the twins competes less well for space and nutrients, and as a result becomes smaller and less vigorous. Some evidence from the squashed ovules indicates that twin embryo sacs may produce dizygotic twin seedlings. This, along with an analysis of the combinations of chromosome numbers and the relative lengths of twin seedlings, indicates that the remaining ones probably originate from cleavage of a single initial embryo.

As determined by FCM, the DNA content of haploids was half that of somatic cells in diploids. Analysis of the nuclear DNA content of plantlets derived from twin embryos revealed differences in content between diploid and haploid embryos. The values found for haploids (1.95 pg) in this study suggest that low DNA content is a property of the genus Asparagus.

In this study we obtained haploid plantlets by germinating twin embryo seeds of asparagus, and confirmed their haploid state by morphological comparison, chromosome counts and genome size analysis. Some of the haploids grew well after acclimatization but did not develop flowers, so we could not confirm their sex. Haploid progeny of cultivars such as Ravel and Sartaguda can provide an important source of breeding material.
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REFERENCES

CHEN Z, and GALLIE DR. 2012. Induction of monozygotic
twinning by ascorbic acid in tobacco. PLoS ONE 7/6 : 1–12.

BROWNFIELD L, and KOHLER C. 2011. Unreduced gamete for-
mation in plants: mechanisms and prospects. Journal of
Experimental Botany 65(5): 1659–1668.

COOPER DC. 1943. Haploid-diploid twin embryos in Lillium
and Nicotiana. American Journal of Botany 30: 408–413.

Doležel J, Binarova P, and Lucretti S. 1989. Analysis of
dual nature of plant cells by flow cytometry. F1 Hybrids. Biologia Plantarum 31: 113–120.

DÖRÉ C. 1990. Asparagus Anther Culture and Field Trial of
Dihaploids and Biotechnology in Agriculture and Forestry. 323–333, vol. 12. Springer Verlag, Berlin Heidelberg.

ELLISON JH. 1986. Asparagus breeding. In: Basset MJ [ed.],
Breeding Vegetable Crops. 521–569. AVI Publishing
Company Inc., Westport.

Frost HB. 1926. Polylembryony, heterozygocity and chimeras
in citrus. Hilgardia 1 : 365–402.

FORSTER BP, HEBERLE-BARS E, KASHA KJ, and TURAEV A. 2007.
The resurgence of haploids in higher plants. Trends in
Plant Science 12: 368–375.

GEBLER P, WOLKO L, and KNAFLEWSKI M. 2007. Identification of
molecular markers for selection of supermale (YY)
aspargus plants. Journal of Applied Genetic 48: 129–131.

ITO T, ASHIZAWA T, SHIMODATE T, FUKUDA T, KAMEYA T, and
KANNO A. 2007. Production and analysis of reciprocal
hybrids between Asparagus officinalis L. and A. schobere-
rioides Kunth. Genetic Resources of Crop Evolution 54
(5): 1063–1071.

JĘDRZEJCZYK I, and NOWACZYK P. 2009. In vivo polylembryony
induction in species of Capsicum. Acta Biologica
Cracoviensia Series Botanica 51/1: 55–60.

KNAFLEWSKI M. 1996. Genealogy of asparagus cultivars. In: Nichols M and Swain D [eds.], Proceedings VIII Int.
Asparagus Symp.: 87–91.

KUNITAKE H, NAKASHIMA T, MORI K, and TANAKA M. 1998.
Somaclonal and chromosomal effects of genotype, ploidy
and culture duration in Asparagus officinalis L. Euphytica 102: 309–316.

MALEPSZY S, NIEMIROWICZ -SZCZYT K, and PRZYBECKI Z. 1989.
Biotechnologia w Genetyce i Hodowli Roślín. 151–196. PWN, Warszawa.

MALEPSZY S. 2009. Rośliny w zaspokojaniu potrzeb człowieka.
In: Malepszy [ed.], Biotechnologia Rośl. 3–11. PWN, Warszawa.

MARCELLAN ON, and CAMADRO EL. 1999. Formation and develop-
ment of embryo and endosperm in intra- and inter-
specific cross of Asparagus officinalis and A. densiflorus
(cv. ’Sprengeri’. Scientia Horticulturae 81: 1–11.

MARTINSON V. 1972. Polylembryony in Theobroma cacao L.
Annals of Botany 36(5): 947–951.

MOREIRA S, GURGEL JTA, and ARRUDA LF. 1947. Polylembrionia
em citrus. Bragantia 10: 69–106.

MORENO R, ESPEJO JA, CABRERA A, MILLAN T, and GIL J. 2006.
Ploidy and molecular analysis of ‘Morado de Huelva’
aspargus (Asparagus officinalis L.) population; a spanish
tetraploid landrace. Genetic Resources of Crop
Evolution 53: 729–736.

MURASHIGE T, and SKOOG F. 1962. A revised medium for rapid
growth and bioassays with tobacco tissue cultures. Physiologia Piantarum 15: 473–497.

NOWACZYK P, and NOWACZYK L. 1996. The influence of growth
regulators on the frequency of polylembryony in pepper
(Capsicum annuum L.). Genetica Polonica 37A: 204–207.

NOWACZYK P, and NOWACZYK L. 2006. Genetic analysis of tomato
(Lycopersicon esculentum Mill.) twin forms. Acta
Biologica Cracoviensia Series Botanica 48/1: 53–58.

OZAKI Y, NARIBKO Y, FLUITA C, and OKUBO H. 2004. Ploidy
variation of progenies from intra- and inter-ploidy crosses
with regard to trisomic production in Asparagus offici-
inalis L. Sexual Plant Reproduction 17: 157–164.

PÉREZ-TORNERO O, and PORRAS I. 2008. Assessment of polylembryony
in lemon rescue and in vitro culture of immature embryos. Plant Cell Tissue and Organ Culture
93(2): 173–181.

RANDALL TE, and RICK CM. 1945. A cyto genetic study of polylembryony in Asparagus officinalis L. American Journal of Botany 32: 560–569.

RICCARDI P, CASALI PE, MERCATI F, FALAVIGNA A, and SUNSERI F.
2011. Genetic characterization of asparagus doubled
haploids collection and wild relatives. Scientia
Horticulturae 130: 691–700.

SHIGA I, UNO Y, KANECHI M, and INAGAKI N. 2009.
Identification of polyploidy of in vitro anther-derived
shoots of Asparagus officinalis L. by flow cytometric
analysis and measurement of stomatal length. Journal of
Japan Society for Horticultural Science 78/1: 103–108.

SINGH V, AKADE JH, and NIMBARKAR N. 2010. Inheritance of stem
fasciation and twin/multi-embryonic seeds and genetic
linkage between them in safflower. Indian Journal of
Genetics and Plant Breeding 70: 281–287.

SPECHT CE, MEISTER A, KELLER ERJ, KORZUN L, and BORNER A.
2002. Genetic variability of economically important Asparagus species as revealed by
gene size analysis and rDNA ITS polymorphisms. Plant Science 162: 931–937.

THEVENIN L. 1968. Les problèmes d’amélioration de l’asperge
(Asparagus officinalis L.). Annales de l’ Amélioration des
Plantes 18: 327–365.

UNO Y, LI Y, KANECHI M, and INAGAKI N. 2002. Haploid produc-
tion from polylembryonic seeds of Asparagus. Proc. X
Int. ISHS Symposium on Asparagus. Acta Horticulturae
589: 212–224.

VALDES B. 1964. Asparagus L. In: Tutin TG, Heywood VH, Burges NA, Moore DM. Valentine DH, Walters SM and
WEBB DA [eds.], *Flora Europaea*, 71–73. Cambridge University Press.

VERNON DM, HANSON MJ, LE M, and FORSTHOEFL NR. 2001. An expanded role for the TWN1 gene in embryogenesis: defects in cotyledon pattern and morphology in the *twn1* mutant of *Arabidopsis* (Brassicaceae). *American Journal of Botany* 88: 570–582.

WEBBER JM. 1940. Polyembryony. *Botanical Review* 6: 575–598.

ZENKTELER M, and GUZOWSKA I. 1967. O niektórych zagadnieniach eksperymentalnej embriologii roślin. I. Poliembrionia. *Wiadomości Botaniczne* 11: 181–190.

ZENKTELER M, and NITZSCHE W. 1984. Wide hybridization experiments in cereals. *Theoretical Applied of Genetics* 68: 311–315.

ZENKTELER M, and STRAUB J. 1979. Cytoembryological studies on the process of fertilization and the development of haploid embryos of *Triticum aestivum* L. (2n=47) after crossing with *Hordeum bulbosum* (2n=14). *Zeitschrift für Pflanzenzüchtung* 82: 32–44.

ZHANG C-J, WANG H-L, MA Y, and KANG Y-Q. 1994. Regeneration of haploid plants from isolated microspores of asparagus (*Asparagus officinalis* L.). *Plant Cell Reports* 13: 637–640.