**ITPKC** Single Nucleotide Polymorphism Associated with the Kawasaki Disease in a Taiwanese Population

Ho-Chang Kuo1,3, Kuender D. Yang1,3, Suh-Hang Hank Juo4,6, Chi-Di Liang3, Wei-Chiao Chen4, Yu-Shiuan Wang4, Chih-Hung Lee11, Edward Hsi8, Hong-Ren Yu1,3, Peng-Yeong Woon9, I-Chun Lin2, Chien-Fu Huang5, Daw-Yang Hwang10, Chiu-Ping Lee1, Li-Yan Lin1, Wei-Pin Chang7, Wei-Chiao Chang4,5,6*

1 Division of Allergy, Immunology and Rheumatology, Chang Gung Memorial Hospital-Kaohsiung Medical Center, Kaohsiung, Taiwan; 2 Division of Cardiology, Department of Pediatrics, Chang Gung Memorial Hospital-Kaohsiung Medical Center, Kaohsiung, Taiwan; 3 Graduate Institute of Clinical Medical Science, Chang Gung University College of Medicine, Kaohsiung, Taiwan; 4 Department of Medical Genetics, College of Medicine, Kaohsiung Medical University, Kaohsiung, Taiwan; 5 Center of Excellence for Environmental Medicine, Kaohsiung Medical University, Kaohsiung, Taiwan; 6 Cancer Center, Kaohsiung Medical University Hospital, Kaohsiung, Taiwan; 7 Department of Healthcare Management, Yuanpei University, HsinChu, Taiwan; 8 Graduate Institute of Medicine, College of Medicine, Kaohsiung Medical University, Kaohsiung, Taiwan; 9 Department of Molecular Biology and Human Genetics, Tzu Chi University, Hualien, Taiwan; 10 Division of Nephrology, Department of Medicine, Kaohsiung Medical University Hospital, Kaohsiung, Taiwan; 11 Department of Dermatology, Graduate Institute of Medicine, Kaohsiung Medical University, Kaohsiung, Taiwan

**Abstract**

Kawasaki disease (KD) is characterized by systemic vasculitis with unknown etiology. Previous studies from Japan indicated that a gene polymorphism of **ITPKC** (rs28493229) is responsible for susceptibility to KD. We collected DNA samples from 1,531 Taiwanese subjects (341 KD patients and 1,190 controls) for genotyping **ITPKC**. In this study, no significant association was noted for the **ITPKC** polymorphism (rs28493229) between the controls and KD patients, although the CC genotype was overrepresented. We further combined our data with previously published case/control KD studies in the Taiwanese population and performed a meta-analysis. A significant association between rs28493229 and KD was found (Odds Ratio: 1.36, 95% Confidence Interval 1.12–1.66). Importantly, a significant association was obtained between rs28493229 and KD patients with aneurysm formation (P = 0.001, under the recessive model). Taken together, our results indicated that C-allele of **ITPKC SNP** rs28493229 is associated with the susceptibility and aneurysm formation in KD patients in a Taiwanese population.

**Introduction**

Kawasaki disease (KD) is a multi-systemic vasculitis with unknown etiology which was first reported by Kawasaki et al. [1] from Japan in 1974 in the English language literature. It occurs worldwide and mainly affects children less than 5 years old especially in Asian countries, Japan, Korea and Taiwan with the incidence from 69 to 213 cases per 100,000 for children <5 years of age [2,3]. The clinical characteristics of KD include prolonged fever, bilateral non-purulent conjunctivitis, diffuse mucosal inflammation, polymorphous skin rashes, indurative angioedema of the hands and feet, and non-suppurative central lymphadenopathy [3,4]. The most serious complication of KD is coronary artery lesions (CAL) including myocardial infarction, coronary artery dilatation, coronary fistula or coronary artery aneurysm [5,6]. In developed countries, KD is the leading cause of acquired heart diseases in children.

While these clinical features of KD are recognized, their underlying immunopathogenic mechanisms are still unclear; in particular, the culprit for developing CAL in patients with KD remains as yet to be uncovered. KD is regarded as an immune disorder of host rather than an infectious disease. From our previous reports, monocytosis and T helper 2 (Th2) immune response were associated with CAL formation [6,7], and intravenous immunoglobulin (IVIG) treatment response in KD [8]. These results highlight the involvement of dysregulated immune response in the pathogenesis of KD.

Genetic association of KD with inositol 1,4,5-trisphosphate 3-kinase C (ITPKC) in a genome wide scan has been recently reported by Onouchi et al. They first indicated a functional SNP in the **ITPKC** gene associated with KD susceptibility and the development of coronary artery lesions [9]. The polymorphism of **ITPKC** (rs28493229) located in intron 1 that resulted in the different transcriptional level of mature mRNA by interfering the RNA splicing efficiency. **ITPKC** is a kinase that phosphorlates inositol 1,4,5-trisphosphate (IP$_3$). IP$_3$ is an important factor in the initiating of calcium release from endoplasmic reticulum [10].

The empty of endoplasmic further evokes store-operated calcium influx. Store-operated calcium influx has been shown to involve in the activation of T cells. For example, IP$_3$-mediated
High level expression of ITPKC can attenuate the Ca2+ pathways via over phosphorylation of IP3 that resulted in reduction of T cell activation and IL-2 production. ITPKC is controversial [11,12].

Genetic polymorphisms of ITPKC were associated with the susceptibility of Kawasaki disease and CAL formation. However, the replication (rs28493229) was confirmed to associate with the susceptibility of Kawasaki disease and CAL formation. Patients studied

Materials and Methods

Patients studied

All subjects studied were children with the diagnosis of KD and were admitted at Chang Gung Memorial Hospital at Kaohsiung between 2001 and 2009. All patients were treated with a single infusion of IVIG (2 g/kg) administered over a 12-hour period. Aspirin (3–5 mg/kg/day) was administered until all signs of inflammation were resolved or regression of CAL was detected under two-dimensional (2D) echocardiography; these conditions were the same as that in our previous studies [6,7,8,13,14]. We excluded patients who did not meet the diagnostic criteria for KD. CAL was defined by the internal diameter of the coronary artery being at least 3 mm (4 mm, if the subject was over the age of 5 years) or the internal diameter of a segment being at least 1.5 times that of an adjacent segment, as observed in the echocardiogram [15,16,17]. KD patients with coronary artery ectasia or dilatation which was disappearing within the initial 2 months after the onset of illness were defined as transient ectasia (or transient CAL) and not judged as CAL. IVIG responsiveness was defined as defervescence 48 h after the completion of IVIG treatment and no fever (temperature, >30°C) recurrence for at least 7 days after IVIG with marked improvement or normalization of inflammatory signs. A total of 1190 control subjects were recruited from the general population who volunteered to participate in our study while receiving a health screening examination at the Kaohsiung Medical University Hospital. All the subjects gave the consent form. The study protocol conformed to the Declaration of Helsinki and study was approved by the Institute Review Board of Hospital.

DNA extraction

Blood cells were subjected to DNA extraction by treating them first with 0.5% SDS lysis buffer and then protease K (1 mg/ml) for digestion of nuclear protein for 4 h at 60°C. Total DNA was harvested by using the Gentra extraction kit followed by 70% alcohol precipitation.

Genotyping

Genomic DNA was extracted from whole blood samples by using the standard method. Genotyping was carried out using the TaqMan Allelic Discrimination Assay (Applied Biosystems, Foster city, CA). Briefly, the polymerase chain reaction (PCR) was performed by using a 96-well microplate with the ABI 9700 Thermal Cycler. The thermal cycle conditions were as follows: denaturing at 95°C for 10 min, followed by 40 cycles of denaturing at 92°C for 15 s and annealing and extension at 60°C for 1 min. After PCR, fluorescence was measured and analyzed using the System SDS software version 1.2.3. Average genotyping successful rate in our lab is around 95.7%, so some individuals are without genotype data.

Statistical analysis

SAS 9.1 for Windows was used for analysis. The statistical differences between cases and controls in genotype and allele frequency were assessed by the χ2-test or the Fisher exact test. The statistical differences in the genotype and allele frequency of KD patients with and those without CAL formation and patients responding to IVIG and those showing resistance were assessed using the χ2-test. For the meta-analysis, sufficient data were available from other two KD association studies, and the odds ratios with 95% confidence intervals were calculated for the risk factors investigated. Combined odds ratios were calculated using Mantel–Haenszel method with a fixed effect model for each outcome measure. Statistical analysis was completed using the rmeta package of R software.

Table 1. Basal characteristics of patients with Kawasaki disease (KD) and of normal controls.

| Characteristics                  | Patients with KD | Normal Control |
|----------------------------------|------------------|----------------|
| Number of subjects               | 341              | 1190           |
| Gender: male, No (%)             | 226 (66.3%)      | 681 (57.2%)    |
| Age (years)                      | 1.6 ± 1.4        | 26.0 ± 22.6    |
| Range                            | 3.5–144 (months) | 1–85 (years)   |
| Coronary artery lesions (CAL)    | 35 (10.3%)       |                |
| Aneurysm formation               | 14 (4.1%)        |                |
| Intravenous immunoglobulin (IVIG) resistant | 43 (12.6%) |                |

CAL: coronary artery lesions; IVIG: intravenous immunoglobulin; SD: standard deviation.

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Table 2. Genotype frequencies for ITPKC SNP and Kawasaki disease susceptibility.

| Genotype | Case (%) (n = 334) | Control Subjects (%) (n = 1131) | Allele | Case (%) (n = 334) | Control Subjects (%) (n = 1131) | Genotype P Value | Dominant P Value | Recessive P Value | Allelic P Value |
|----------|--------------------|-------------------------------|--------|-------------------|-------------------------------|-----------------|-----------------|-----------------|-----------------|
| rs28493229 |                    |                               |        |                   |                               |                 |                 |                 |                 |
| CC       | 2 (0.6)            | 8 (0.7)                       | C      | 54 (8.1)          | 158 (7.0)                    | 0.509           | 0.283           | 0.832           | 0.335           |
| CG       | 50 (15.0)          | 142 (12.6)                    | G      | 614 (91.9)        | 2104 (93.0)                  | 0.062*          | 0.891*          | 0.065*          |                 |
| GG       | 282 (84.4)         | 981 (86.7)                    |        |                   |                               |                 |                 |                 |                 |

*Adjusted the effects of age and gender.

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Results

Meta-analysis shows that *ITPKC* SNP rs28493229 is associated with the susceptibility of KD

Table 1 shows the characteristics of the subjects. A total of 341 KD patients and 1190 controls were recruited in this study. 66.3% of cases and 57.2% of controls were male. The mean age of patients and controls was 1.6±1.4 months (standard deviation [SD]) and 26.0±22.6 years, respectively. Children were predominant in the study population, an observation that is in agreement with the reported incidence of KD. The prevalence of KD is less than 1/1,000 children in Taiwan. Therefore, we assumed that there was no KD case in the control group. There were 10.3% of KD with CAL formation, and 4.1% with aneurysm formation. Among these KD patients 12.6% were resistant to initial IVIG treatment. The distribution of *ITPKC* SNP genotypes was in accordance with the Hardy-Weinberg equilibrium for both cases and controls (Table 2). However, the SNP tested in this study doesn’t show a significant association with KD under three genetic models (Dominant model; Recessive model and Allelic model). To assess the role of variation in *ITPKC* SNP rs28493229 more extensively, we therefore performed a meta-analysis on all published KD association studies in the Taiwanese population including the present data. The results showed a significant association between *ITPKC* SNP rs28493229 and the susceptibility of KD (OR = 1.36, 95% CI 1.12–1.66) in the Taiwanese population (Figure 1).

No association between *ITPKC* SNP rs28493229 and CAL formation or IVIG treatment response in KD patients

A total of 341 KD patients were included in this study, of which 35 patients (10.3%) had CAL formation and 43 patients (12.6%) were resistant to initial IVIG treatment (Table 1). We further evaluate the relationship between rs28493229 and the risk of CAL formation or IVIG resistance. The data is shown in Table 3 and Table 4, respectively. The frequency of C allele was higher in the patients with CAL formation (10.3% vs. 7.6%) or IVIG resistance (11.6% vs. 7.6). However, the genotype or allele frequency of rs28493229 still failed to reach any significant association with CAL formation or IVIG resistance.

SNP rs28493229 of *ITPKC* significant associated with aneurysm formation in KD

We further identify the role of SNP rs28493229 of *ITPKC* in the pathogenesis of CAL in KD patients. A subset analysis where cases reported having aneurysm formation was performed. In this study, 14 patients (4.1%) had aneurysm formation (aneurysm was defined

| Table 3. Genotyping and allele frequency of *ITPKC* SNP in patients with Coronary artery lesion (CAL) and without CAL. |
|---|
| Genotype | CAL (%) (n = 34) | Without (%) (n = 295) | Allele | CAL (%) (n = 34) | Without (%) (n = 295) | Genotype P Value | Dominant P Value | Recessive P Value | Allelic P Value |
| Rs28493229 |  |  |  |  |  |  |  |  |  |
| CC | 1 (2.9) | 1 (0.3) | C | 7 (10.3) | 45 (7.6) | 0.181 | 0.674 | 0.065 | 0.440 |
| CG | 5 (14.7) | 43 (14.6) | G | 61 (89.7) | 545 (92.4) |  |  |  |  |
| GG | 28 (82.4) | 251 (85.1) |  |  |  |  |  |  |  |
as internal diameter >4 mm or in children ≥5 years of age, the internal diameter of a segment being at least 1.5 times that of an adjacent segment and that was persistent after at least one year followed up) (Table 1). As shown in the Table 5, SNP rs28493229 of ITPKC was associated with the KD patients with aneurysm formation. KD patients carry rs28493229 homozygous C/C genotype had a 24.5-fold ($P=0.001$) increased risk of aneurysm formation when compared with those with C/G and G/G genotypes.

**Table 4. Genotyping and allele frequency of ITPKC SNP in patients with resistant and responsive to Intravenous immunoglobulin (IVIG) treatment.**

| Genotype   | Resistant (%) (n = 43) | Responsive (%) (n = 291) | Allele | Genotype $P$ Value | Dominant $P$ Value | Recessive $P$ Value | Allelic $P$ Value |
|------------|------------------------|--------------------------|--------|---------------------|-------------------|---------------------|------------------|
| Rs28493229 | CC                     | 0 (0.0)                  | 2 (0.7) | C                   | 10 (11.6)          | 44 (7.6)            | 0.266            |
|            | CG                     | 10 (23.3)                | 40 (13.7)| G                   | 76 (88.4)          | 538 (92.4)          | 0.159            |
|            | GG                     | 33 (76.7)                | 249 (85.6)|                    |                   |                    | 0.581            |

As shown in Table 2, although rs28493229 didn’t have significant association with KD susceptibility and BCG scar reactivation during the acute phase but not associated with the develop or severity of CAL formation. As shown in Table 2, although rs28493229 didn’t have significant association with the KD, higher C allele frequency in KD cases (8.1% vs 7.0%) was seen in this study. To increase the analysis power, we therefore performed a meta-analysis on all published KD association studies in the Taiwanese population. Consistent with the first ITPKC studies by Onouchi [11,12], results from meta-analysis strongly indicated that C allele of ITPKC rs28493229 is a risk allele in the KD susceptibility in the Taiwanese population. In this study, we also first analysis the association between rs28493229 with IVIG treatment response and aneurysm formation. Our results indicated that C allele of ITPKC rs28493229 had significant higher risk to have aneurysm formation. Although the exact role of ITPKC in the development of KD is unclear, it may function as the negative regulator in the regulation of immune genes or to be involved in the T cell activation mechanisms. The results also, at least, reflect the facts that complex genetic, ethnic, and environmental factors are involved in the association with KD [11,12]. Regarding to the different results obtained for ITPKC genetic association studies in the Taiwanese population, we attribute this to the population migration between Taiwan and China, Thailand, Indonesia, Malaysia, Vienna and other Asia areas, due to the increase of genetic diversity between cities in the south or north of Taiwan.

ITPKC is able to effect cellular expression level of IP$_3$ via phosphorylation steps. Two possible signaling pathways that may involve in the increase of intracellular calcium concentration can be triggered by IP$_3$. First, upon binding to the IP$_3$ receptor (IP$_3$R) on the endoplasmid reticulum, IP$_3$ triggers the release of intracellular calcium from stores. Second, store empty resulted in the activation of STIM1-mediated store-operated calcium channels. Both calcium releases from stores or calcium influx through store-operated calcium channels have been confirmed to initiate signaling pathways to regulate inflammatory cytokines and immune diseases. In T-cells, agents block calcium influx through store-operated calcium channel attenuated the translocation of NFAT or NFkB and the expression of immune related genes [18]. The polymorphism of ITPKC (C allele) exhibited lower gene expression that may result in the increase of global intracellular calcium concentration via IP$_3$-mediated pathways. Over-activation of calcium-dependent inflammatory cytokines may damage coronary vessel wall and cause to the aneurysm formation. Our results implied that C-allele of ITPKC SNP rs28493229 may passively increase the activation of calcium-dependent signaling pathways that lead to aneurysm formation in KD patients.

KD is considered as one of the immune-mediated diseases. Several genes have been reported to be associated with

**Table 5. Genotyping and allele frequency of ITPKC SNP in patients with aneurysm or without aneurysm.**

| Genotype   | Aneurysm (%) (n = 14) | Without (%) (n = 320) | Allele | Aneurysm (%) (n = 14) | Without (%) (n = 320) | Genotype $P$ Value | Dominant $P$ Value | Recessive $P$ Value | Allelic $P$ Value |
|------------|------------------------|------------------------|--------|------------------------|------------------------|---------------------|-------------------|---------------------|------------------|
| Rs28493229 | CC                     | 1 (7.1)                | 1 (0.3) | C                     | 3 (10.7)               | 51 (8.0)           | 0.004*            | 0.892               | 0.001*           |
|            | CG                     | 1 (7.1)                | 49 (15.3)| G                     | 25 (89.3)              | 589 (92.0)         |                   | 0.602               |                   |
|            | GG                     | 12 (85.8)              | 270 (84.4)|                      |                        |                    |                   |                     |                   |

*Significant ($P<0.05$) values are in bold.

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susceptibility to KD and/or CAL formation including ITPKC, CASP3, CTLA4, TGF-beta pathways and MMP family... etc [9,16,19,20,21,22,23]. Genes of ITPKC and CASP3 are involved in the Ca^{2+}/NFAT pathways. These two genes have been shown to associate with susceptibility to KD in the Japanese and European Americans. Consistent with this, we still observe a significant association in the KD patients with aneurysm formation even though the allele frequency is low. What might be the relevance of our findings to the KD? KD patients who are resistant with initial IVIG therapy have higher risk of developing CAL or aneurysm formation. Many inhibitors for Ca^{2+}/NFAT pathways are already in the clinical use. From pharmacology prospective, to identify novel inhibitors against Ca^{2+}/NFAT pathways seems to be promising in the treatment of KD. Further pharmacogenomic studies to confirm to the effects of inhibitors for Ca^{2+}/NFAT pathways such as Cyclosporine A in the IVIG resistant patients may should be considered.

In conclusion, we reported that ITPKC (rs28493229) had significant association with the susceptibility of KD in the Taiwanese population. Importantly, KD patients carry ITPKC rs28493229 C allele had significant higher risk to have aneurysm formation.

Author Contributions
Conceived and designed the experiments: H-CK K-DY S-HHJ W-CC. Performed the experiments: H-CK W-CC Y-CW D-YH. Analyzed the data: E-H, HR-Y P-YW I-CL C-FH C-PL W-PC W-CC. Wrote the paper: H-CK W-PC W-CC.

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