Oxygen kinetics and debt during recovery from expiratory flow-limited exercise in healthy humans

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Accepted: 3 November 2006 / Published online: 6 December 2006 © Springer-Verlag 2006

Abstract In healthy subjects expiratory flow limitation (EFL) during exercise can lower O2 delivery to the working muscles. We hypothesized that if this affects exercise performance it should influence O2 kinetics at the end of exercise when the O2 debt is repaid. We performed an incremental exercise test on six healthy males with a Starling resistor in the expiratory line limiting expiratory flow to ~1 l s–1 to determine maximal EFL exercise workload (Wmax). In two more square-wave exercise runs subjects exercised with and without EFL at Wmax for 6 min, while measuring arterial O2 saturation (% SaO2), end-tidal pressure of CO2 (PETCO2) and breath-by-breath O2 consumption (V̇O2) taking into account changes in O2 stored in the lungs. Over the last minute of EFL exercise, mean PETCO2 (54.7 ± 9.9 mmHg) was significantly higher (P < 0.05) compared to control (41.4 ± 3.9 mmHg). At the end of EFL exercise %SaO2 fell significantly by 4 ± 3%. When exercise stopped, EFL was removed, and we continued to measure V̇O2. Recovery V̇O2 kinetics after EFL exercise was best characterized by a double-exponential function with fundamental and slow time constants of 27 ± 11 and 1,020 ± 305 s, compared to control values of 41 ± 10 and 1,358 ± 320 s, respectively. EFL O2 debt was 52 ± 22% greater than control (2.19 ± 0.58 vs. 1.49 ± 0.38 l). We conclude that EFL exercise increases the O2 debt and leads to hypoxemia in part due to hypercapnia.

Keywords Exercise tolerance · Oxygen debt · COPD models

Introduction

In spite of the fact that dynamic hyperinflation (Calverley and Koulouris 2004; O’Donnell et al. 2001) and peripheral muscle deconditioning (Maltais et al. 1998; Richardson et al. 1999) are important factors limiting exercise in COPD, there is evidence that inadequate oxygen delivery to working locomotor and respiratory muscles may also play an important role (Aliverti and Macklem 2001; Aliverti et al. 2005a; Iandelli et al. 2002).
This situation has been modeled in normal humans by imposing expiratory flow-limitation (EFL) during exercise which limits exercise performance by intense dyspnea (Aliverti et al. 2002; Iandelli et al. 2002), while decreasing arterial oxygen saturation and cardiac output (Aliverti et al. 2005a). This in turn leads to premature competition between respiratory and locomotor muscles for available energy supplies and the early onset of mixed respiratory and metabolic acidosis (Aliverti and Macklem 2001; Kayser et al. 1997).

As a result of these previous experiments (Aliverti and Macklem 2001; Aliverti et al. 2005a; Kayser et al. 1997) we predicted that EFL exercise in normal subjects should increase the oxygen debt by decreasing systemic oxygen delivery to exercising muscles. Accordingly, to test this hypothesis, we measured O$_2$ kinetics during repayment of the O$_2$ debt after control and EFL exercise in young, normal, male subjects. Our results revealed that EFL increased O$_2$ net debt by an average 52% and led to hypoxemia in part due to hypercapnia.

### Material and methods

#### Subjects

We studied six healthy male individuals whose baseline characteristics are shown in Table 1. All subjects gave signed–informed consent and the protocol was approved by the University of Athens Ethics Committee, where the experiments were conducted.

#### Pulmonary function tests

Spirometry for the determination of forced expiratory volume in 1 s (FEV$_1$) was performed by a $V_{\text{max}229}$ system (Sensor Medics, Anaheim, CA, USA) throughout both control and EFL exercise and recovery runs. Flow displaced at the mouth was also measured by a screen type pneumotachograph (3813 Hans Rudolph, Kansas City, MO, USA) and from these measurements the following parameters were obtained: minute ventilation, ($\dot{V}_E$); tidal volume, ($V_T$); breathing frequency, ($f_b$); inspiratory ($T_i$), expiratory ($T_e$), total respiratory

### Exercise protocols

Three exercise tests were performed for each subject on an electromagnetically braked cycle ergometer (Ergoline 800, Sensor Medics) with the subjects maintaining a pedaling frequency of 60 rpm. At the beginning an incremental test was performed to establish the individual subject’s maximal workload when EFL was imposed ($W_{\text{maxEFL}}$) by the application of a Starling resistor in the expiratory line limiting flow to approximately 1 l s$^{-1}$. After 3 min of baseline measurements, followed by 3 min of unloaded pedaling, the work rate was increased every min by 20 W to the limit of tolerance (i.e. the point where the work rate could not be tolerated due to severe sensation of dyspnea, see Table 1). On a separate day, two square-wave protocols were performed: the first without EFL (control) and the second with EFL. After 3 min of baseline measurements, followed by 3 min of unloaded pedaling, the work rate was set and maintained at ($W_{\text{maxEFL}}$) for 6 min in both protocols. At the end of exercise, EFL was removed and recovery was monitored for an additional 6 min (Fig. 1).

### Gas exchange and flow measurements

Instantaneous inspired and expired O$_2$ and CO$_2$ concentrations, end-tidal values for O$_2$ and CO$_2$ ($P_{\text{ETO}_2}$, $P_{\text{ETCO}_2}$, respectively) and flow at the mouth were recorded breath-by-breath ($V_{\text{max}229}$, Sensor Medics) throughout both control and EFL exercise and recovery runs. Flow displaced at the mouth was also measured by a screen type pneumotachograph (3813 Hans Rudolph, Kansas City, MO, USA) and from these measurements the following parameters were obtained: minute ventilation, ($\dot{V}_E$); tidal volume, ($V_T$); breathing frequency, ($f_b$); inspiratory ($T_i$), expiratory ($T_e$), total respiratory

### Table 1 Subjects’ characteristics

| Subject | Age (years) | Height (m) | Weight (kg) | FEV$_1$ (l) | FEV$_1$ (%predicted) | ($W_{\text{maxEFL}}$) | $\dot{V}_{O2EFL}$ (l min$^{-1}$) | Dyspnea (Borg scale) |
|---------|-------------|------------|-------------|-------------|---------------------|----------------------|-------------------------|---------------------|
| 1       | 38          | 1.76       | 78          | 3.90        | 97                  | 155                  | 1.7                     | 5                   |
| 2       | 22          | 1.73       | 75          | 4.23        | 100                 | 170                  | 2.5                     | 8                   |
| 3       | 19          | 1.69       | 59          | 3.93        | 97                  | 100                  | 1.6                     | 7                   |
| 4       | 22          | 1.94       | 86          | 5.47        | 112                 | 125                  | 2.5                     | 5                   |
| 5       | 22          | 1.88       | 80          | 5.73        | 109                 | 100                  | 2.0                     | 5                   |
| 6       | 34          | 1.66       | 69          | 3.52        | 96                  | 90                   | 1.8                     | 7                   |
| Mean    | 26          | 1.77       | 73          | 4.46        | 102                 | 123                  | 2.1                     | 6                   |
| SD      | 8           | 0.11       | 11          | 0.91        | 7                   | 33                   | 0.4                     | 1                   |

FEV$_1$, forced expiratory volume in 1 s in absolute and as percentage predicted; ($W_{\text{maxEFL}}$), maximal workload when EFL was imposed during the incremental cycling test; $\dot{V}_{O2EFL}$, maximal oxygen uptake when EFL was imposed during the incremental test.
(T_{\text{vol}}) cycle time; and duty cycle (T_i/T_{\text{Tot}}). Cardiac frequency (f_c) and percentage arterial oxygen saturation (% SaO_2) were determined using the R–R interval from a 12-lead on-line electrocardiogram (Marquette Max, Marquette Hellige GmbH, Germany) and a pulse oximeter (Nonin 8600, Nonin Medical, USA), respectively. The modified Borg Scale (Borg 1982) was used to rate the magnitude of dyspnea and leg discomfort every 2 min throughout exercise.

Operational chest-wall volume measurements

Simultaneously, throughout the square wave exercise and recovery protocols the volume of the chest wall was measured by optoelectronic plethysmography (OEP system, BTS, Milan, Italy) as previously described (Aliverti et al. 1997; Cala et al. 1996; Vogiatzis et al. 2005). In brief, the motion of 89 retro-reflective markers placed front and back over the chest wall from clavicles to pubis was recorded. Each marker was tracked by six video cameras, three in front of the subject and three behind. Subjects grasped handles positioned at mid-sternum level, which lifted the arms away from the rib cage, so that lateral markers could be visualized. Dedicated software, which recognizes the markers on each camera in real time, reconstructed their 3D co-ordinates by stereophotogrammetry and using Gauss’s theorem, calculated inspiratory and expiratory tidal volume variations. Thus, breathing pattern was measured both plethysmographically and by integration of flow. Data acquisition from the OEP system was interrupted for 1 min every 2 min (Fig. 1) in order to allow data storage.

Calculation of breath-by-breath oxygen consumption

The volume of O_2 exchanged at the mouth differs from the volume of O_2 taken up by pulmonary capillaries if the amount of O_2 stored within the lung changes. This occurs if the volume inspired is different from the volume expired in a given respiratory cycle, and/or if alveolar concentrations of O_2 change (Capelli et al. 2001). This is particularly important during on- and off-exercise transients (Cautero et al. 2002). The changes in O_2 stored within the lung must be subtracted from the volume of O_2 exchanged at the mouth in order to obtain the amount of O_2 exchanged at the alveolar level. This can be done by combining OEP with independent measures of the subdivisions of lung volume and continuously measuring changes in the absolute gas volume by OEP throughout all exercise tests. This method was recently described in detail (Aliverti et al. 2004a, 2005a). Accordingly, in the present study, breath-by-breath oxygen consumption (\dot{V}O_2) was computed throughout using the data acquired by OEP, flow at the mouth and the continuous inspired and expired O_2 and CO_2 concentration recordings. Using this method corrections were made for (1) pneumotachograph integrator drift, (2) the sampling rates of the O_2 and CO_2 meters, (3) the time delay of the O_2 analyzer and the flow through the O_2 and CO_2 sampling lines, (4) precise synchronization of O_2 concentration and volume signals and (5) any changes in O_2 stored within the lung. Thus, in contrast to commercial devices, we measured \dot{V}O_2 at the level of the pulmonary capillaries, not at the mouth.

Curve fitting

To characterize the kinetics of the \dot{V}O_2 response during square-wave exercise and recovery single and double-exponential functions were applied, respectively, to the data using a non-linear least-squares fitting procedure (Figs. 2, 3) without omitting the first 20 s of the response for either the on- or off-transient. The equations applied (Linnarsson 1974; Ozyener et al. 2001) were

For the on-transient: \dot{V}O_2(t) = a \cdot (1 - \exp^{-b \cdot t}), \quad (1)

For the off-transient: \dot{V}O_2(t) = a \cdot \exp^{-b \cdot t} + c \cdot \exp^{-d \cdot t}, \quad (2)

where in Eq. 2, the response is the sum of two components, each described by an exponential decay with two time constants: \tau_1 = 1/b (the fundamental \dot{V}O_2
component) and \( \tau_2 = 1/d \) (the slow \( VO_2 \) component). In the same equation, \( a \) and \( c \) are the initial values of these two components and their sum \((a + c)\) represents the initial value of the total response, i.e. the value of \( VO_2 \) at time zero. The adequacy of the fitting model was tested by calculating \( R^2 \) values and residuals for single and double exponentials.

Measurement of oxygen debt

Oxygen debt was calculated by integrating, for each subject, the fitting curves between time zero (start of recovery) and an arbitrary time of 400 s (Fig. 3). Then, the value of mean \( VO_2 \) at baseline, multiplied by the same period of time, was subtracted (Aliverti et al. 2005a).

Estimation of cardiac output and arterio-venous oxygen difference

To estimate the cardiac output (CO) and the arterio-venous oxygen (\( \text{a-v} \) \( \text{O}_2 \)) difference during EFL and control exercise the following equations (Stringer et al. 1997) were used:

Estimated \( \text{CO} = VO_2/[5.72 + (0.105 \times \% \text{VO}_2\text{max})] \),

\( (3) \)

Estimated \( \text{a-v} \) \( \text{O}_2 = 5.72 + (0.105 \times \% \text{VO}_2\text{max}) \).

\( (4) \)

Statistical analysis

Data are presented as mean ± SD. Significant differences in recorded variables over the last minute of the square-wave exercise protocols between control and EFL tests were assessed by the Wilcoxon signed-rank test as were recovery data from control and EFL exercise tests. Linear regression analysis was performed using the least-squares method. A statistical significance of 0.05 was used for all analyses.

Results

Square-wave protocols

Figure 2 displays a typical example of breath-by-breath \( VO_2 \) during control (closed circles) and EFL exercise (open circles) and recovery. Upon completion of exercise the Starling resistor was removed. Note that time is aligned on the precise instant in which the recovery started (time zero).

![Fig. 2](image-url)

Fig. 2 Typical example of breath-by-breath \( VO_2 \) during control (closed circles) and EFL exercise (open circles) and recovery. Data are fitted with double-exponential curves

Figure 3 displays a typical example of breath-by-breath \( VO_2 \) during recovery from control (closed circles) and EFL exercise (open circles). Data are fitted with double-exponential curves.

![Fig. 3](image-url)

Fig. 3 Typical example of breath-by-breath \( VO_2 \) during recovery from control (closed circles) and EFL exercise (open circles). Data are fitted with double-exponential curves

Over the last minute of EFL exercise, mean values for \( f_c \), \( P_{ETCO_2} \) (54.7 ± 9.9 mmHg) and dyspnea scores were significantly larger compared to control exercise (Fig. 4), whereas mean \( P_{ETO_2} \) (87.0 ± 11.3 mmHg) was significantly lower (Fig. 4). The mean fall (\( P = 0.044 \)) in %\( \text{SaO}_2 \) at the end of EFL exercise was 4 ± 3%. (Fig. 4). Leg discomfort at the end of EFL exercise was not different to that at the end of control exercise (Fig. 4). The on-transient data were best fitted with a mono-exponential function where the time constant of the fundamental \( VO_2 \) component was not significantly
Table 2  Exercise data at the end of the 6 min protocols with and without EFL.

|                           | Control exercise | EFL exercise |
|---------------------------|------------------|--------------|
| Work rate (W)             | 111 ± 36         | 109 ± 30     |
| VO₂ (l min⁻¹)             | 2.08 ± 0.31      | 1.93 ± 0.42  |
| VCO₂ (l min⁻¹)            | 2.14 ± 0.38      | 1.88 ± 0.49* |
| fᵢ (beats min⁻¹)         | 152 ± 19         | 163 ± 19*    |
| Vₑ (l min⁻¹)             | 59.8 ± 10.2      | 38.8 ± 15.1* |
| Vᵣ (l)                   | 2.08 ± 0.48      | 2.05 ± 0.45  |
| i/Tot (%)                | 1.00 ± 0.10      | 0.85 ± 0.15  |
| Tᵣ (s)                   | 1.15 ± 0.23      | 2.02 ± 0.43* |
| Tᵣ/Tot (%)              | 46 ± 5           | 31 ± 4*      |

Values are means ± SD
VO₂: oxygen uptake; VCO₂: carbon dioxide output; fᵢ, cardiac frequency; Vₑ, minute ventilation; Vᵣ, tidal volume; fᵢ, breathing frequency; Tᵣ, inspiratory time; Tₑ, expiratory time; Tᵣ/Tot, duty cycle
*Significant differences (P < 0.05) between EFL and control exercise

...different during EFL exercise as compared to control exercise (Table 3).

Recovery oxygen kinetics and oxygen debt

Figure 3 displays a typical example of breath-by-breath VO₂ data fitted with a double-exponential function during recovery from control and EFL exercise square-wave tests. The R² values of the fitted curves applied to the recovery breath-by-breath VO₂ data were not significantly different between EFL (0.89 ± 0.07) and control exercise (0.86 ± 0.09). Immediately after removing the Starling resistor at the end of EFL exercise there was a step increase in VO₂, so that repayment of O₂ debt started at a higher VO₂ after EFL than control. Figure 5 demonstrates individual values of the a, c, and a + c parameters obtained from Eq. 2. All three parameters were significantly higher after EFL compared to control exercise (a: P = 0.04; c: P = 0.0002; a + c: P = 0.02). Also shown in Fig. 5 is the total O₂ debt which was 2.19 ± 0.58 l after EFL exercise compared to the control value of 1.49 ± 0.38 l, an increase of 52 ± 22% (P = 0.01) caused by EFL. In order to verify possible problems originated by the curve fitting, oxygen debt and deficit were also calculated directly from the original breath-by-breath data and did not differ significantly compared to the data obtained by the fitted curves (average difference 4.3 ± 3.1%).

Individual and mean values for the off-transient time constants of the fundamental and slow VO₂ components are shown in Table 3. The fundamental time constant was significantly (P = 0.028) shorter during recovery from EFL compared to control exercise (Table 3). The off-transient slow component during recovery from EFL tended (P = 0.068) to be shorter compared to that after control exercise; however, in recovery from control exercise the slow component was not evident in subjects 3 and 4 (Table 3). Table 4 reports the individual and mean data of the on and off VO₂ transients for EFL and control exercise.

Estimation of cardiac output and arterio-venous oxygen difference

From Eq. 3, \( CO_{EFL} = [1.9 \text{ l min}^{-1}/5.72 + (0.105 \times 95^-1)] = 12.5 \text{ l min}^{-1} \).

From Eq. 4, \(a-v O₂_{EFL} = 5.72 + (0.105 \times 95) = 15.2 \text{ ml O₂ per 100 ml of blood}. In addition, \(a-v O₂_{control} = 5.72 + (0.10 \times 60) = 5.72 + 5.9 = 11.7 \text{ ml O₂ per 100 ml of blood}.

Based on the above calculations, the mixed venous blood O₂ content (CvO₂) for EFL would be equivalent to 4.8 ml per 100 ml blood or 48 ml O₂ l⁻¹. For control, CvO₂ would be equivalent to 8.3 ml O₂ per 100 ml blood or 83 ml O₂ l⁻¹.

Taking into account that 70% of blood volume is located in the venous compartment at rest and during moderate exercise that figure falls to approximately 50% (Astrand and Rodahl 1986), then during EFL exercise with a cardiac output of 12.5 l min⁻¹ the CvO₂ will be \([(12.5 \text{ l min}^{-1} \times 0.5) \times 0.048 \text{ l O₂} \times 0.3 = 0.1 \text{ l}.\]

For control exercise, the CvO₂ will be \([(13.75 \text{ l min}^{-1} \times 0.5) \times 0.083 \text{ lO₂} = 0.571 \text{ l} \times 570 \text{ ml O₂}. Thus, the difference in CvO₂ between EFL and control will be 570 – 300 = 270 ml. Hence from the 0.70 l difference in oxygen debt between EFL and control calculated in Table 5 (2.19 – 1.49 = 0.70 l), 0.27 l (i.e. approximately one-third) would account for the replenishment of the blood oxygen stores.

Discussion

There is growing evidence that an inadequate oxygen supply to meet demand may play a more dominant role (Astrand and Rodahl 1986).
role in limiting exercise capacity in some patients with advanced COPD than impaired lung function (Aliverti et al. 2004b, 2005b; Maltais et al. 1998; Oelberg et al. 1998; Potter et al. 1971; Richardson et al. 1999; Stark-Leyva et al. 2004). In healthy subjects, limiting expiratory flow by a Starling resistor or increasing expiratory load by imposing a constant pressure at the mouth during expiration, have been shown to significantly reduce cardiac output during exercise (Aliverti et al. 2005a; Stark-Leyva et al. 2004). This results from the enforced decrease in the velocity of shortening of expiratory muscles and hypercapnia, both of which contribute to increased expiratory muscle force and decreased duty cycle so that expiration acts like a Valsalva maneuver with inadequate time to recover during inspiration (Aliverti et al. 2002, 2005a; Iandelli et al. 2002). Furthermore, the decreased duty cycle magnifies the effect of high expiratory pressures when averaged over the whole respiratory cycle. Our present results show that EFL increased O₂ debt by an average 52% and led to hypoxemia in part due to hypercapnia. This supports our hypothesis that the reduction in cardiac output during EFL exercise decreases O₂ supply to working locomotor and respiratory muscles resulting in an increased O₂ debt. We conclude that the decreased cardiac output is important in limiting EFL exercise performance in healthy subjects.

Fig. 4 Individual values for heart rate (f, a), %SaO₂ (b), P_{ET}O₂ (c), P_{ET}CO₂ (d), dyspnea (e) and leg discomfort (f) at the end of control and EFL exercise protocols.
EFL as a model for COPD

The experimental model used in this study was designed to simulate, in part, the flow-limitation commonly experienced by COPD patients during exercise. The pros and cons of this model have been previously extensively discussed (Aliverti et al. 2002; Iandelli et al. 2002). The model has demonstrated that exercise with EFL induces intolerable dyspnea, CO2 retention, impaired exercise performance, expiratory muscle recruitment (Kayser et al. 1997), blood shifts from trunk extremities, a reduced duty cycle, arterial desaturation, and a decrease in cardiac output (Aliverti et al. 2005a).

In the present study, we confirmed many of these results and showed that the increased O2 debt was accompanied by a reduction in VO2 by 8% and a fall in SaO2 by 4% at the end of EFL exercise, thus leading to a reduction in systemic O2 delivery of ~12%, in close agreement with the 15% reduction reported by Ali-

Table 3 Individual and mean data on the time constant of the fundamental and slow component of the VO2 response for on- and off-transients

| Subject | On-transient τ1 (s) | Off-transient τ1 (s) | Off-transient τ2 (s) |
|---------|---------------------|----------------------|----------------------|
|         | Control | EFL     | Control | EFL     | Control | EFL     |
| 1       | 75.8    | 66.2    | 34.7    | 29.2    | 1,429   | 1027.5  |
| 2       | 50.5    | 62.9    | 32.1    | 23.1    | 1,428   | 930.7   |
| 3       | 49.0    | 54.6    | 33.9    | 32.8    | *       | 976.1   |
| 4       | 89.3    | 87.0    | 53.4    | 30.9    | *       | 1605.8  |
| 5       | 32.5    | 41.2    | 36.1    | 6.9     | 909     | 855.0   |
| 6m      | 61.7    | 72.5    | 53.1    | 36.6    | 1,666   | 727.7   |
| Mean    | 59.8    | 64.1    | 40.6    | 26.6    | 1358.0  | 1020.5  |
| SD      | 20.4    | 15.6    | 9.9     | 10.6    | 319.5   | 305.2   |
| P value | 0.276   | 0.028   | 0.068   | 0.068   |

Fundamental (τ1) time constant of the on-transient VO2 response during control and EFL exercise (first and second columns). Fundamental (τ1) and slow (τ2) time constants of the off-transient VO2 response after control and EFL exercise (third to sixth column).

*Values > 10⁶

Fig. 5 Individual values of the parameters given in Eq. 1: a (a), c (b), a + c (c) and calculated gross values for the oxygen debt (d) after the EFL and control exercise protocols.
It is reasonable to assume that the reduction in systemic O₂ delivery during EFL exercise would be associated with the progressive recruitment of fast-twitch fibers and hence the premature onset of the lactate threshold. The latter has been shown to be the case during exercise with EFL (Aliverti et al. 2005a). When exercise workloads exceed the lactate threshold, energy supplies are inadequate to meet demands. The resulting competition between working locomotor and respiratory muscles for the available energy supplies, regulated by autonomic reflex mechanisms (Harms et al. 1998), would be substantially worsened by the decrease in the available O₂ and should further increase the lactate production. It is therefore likely that the muscle and blood lactate levels (not measured in the present study) would be appreciably higher during EFL exercise and consequently, the lactate related metabolic cost could significantly contribute to the repayment of the O₂ debt in recovery from EFL exercise.

In addition to the reduction in systemic O₂ delivery, the present study shows that application of EFL during exercise significantly decreased minute ventilation compared to control exercise, causing hypercapnia. This confirms earlier results that show that a vicious circle is induced whereby increasing central ventilatory drive increases expiratory pressure which further reduces alveolar ventilation and cardiac output (Aliverti et al. 2002, 2005a; Iandelli et al. 2002). Furthermore, the observed elevated heart rate during EFL exercise possibly reflects a reduction in stroke volume secondary to the decrease in venous return. In the study by Stark-Leyva et al. (2004), expiratory loading during exercise increased heart rate in an attempt to minimize the effects of the reduced stroke volume on cardiac output. Thus, the reduced cardiac output secondary to the high expiratory pressures would also be expected to contribute importantly to the greater oxygen debt that was measured after EFL exercise.

In summary, EFL exercise in healthy subjects reproduces many features of COPD including acute respiratory failure, a condition resembling cor pulmonale, dyspnea, and impairment of exercise performance. Furthermore, it has led to testable predictions, one of which is the rationale for this study and which to date have been proven to be accurate (Aliverti et al. 2004b, 2005a, b).

### Off-transient O₂ kinetics

The increase in O₂ debt by 52% that we found during EFL exercise impacted, as predicted, on O₂ kinetics during recovery. Accordingly, the repayment of O₂ debt after EFL exercise started at a higher VO₂ compared to control and the off-transient time constant of the fundamental and slow VO₂ components were shorter after EFL exercise. Collectively, the findings describing the EFL off-transient O₂ kinetics reflect a more rapid replenishment of blood O₂ stores (fundamental component) and a faster repayment of O₂ tissue debt (slow component), both of which could result from the greater O₂ deficit during EFL exercise as opposed to control exercise. Indeed, if we considered...
that cardiac output during EFL exercise was lower than control and \( \dot{V}O_2 \) was not significantly different between EFL and control, then it would be reasonable to expect that during exercise with EFL, that is known to decrease pulmonary blood flow (Aliverti et al. 2005a), the arterial to mixed venous blood \( O_2 \) difference would be larger and the mixed venous blood \( O_2 \) content would be lower. This notion was confirmed by our calculations of arterial to mixed venous blood \( O_2 \). When at the end of exercise EFL was removed, the sudden increase in cardiac output and in blood perfusing the lungs brought about a refilling of the \( O_2 \) stores of mixed venous blood; hence the sudden increase of \( \dot{V}O_2 \) (marking the replenishment of the body’s deprived oxygen stores), a faster \( \dot{V}O_2 \) kinetics and a larger \( O_2 \) debt in EFL (Table 5). The larger depletion of the inner oxygen stores is also suggested by the longer time constants of the on- and off-transient \( \dot{V}O_2 \) response during EFL exercise (Table 3). Our calculations showed that approximately 30% of the \( O_2 \) debt difference between EFL and control exercise was due to the blood \( O_2 \) replenishment (Table 5) caused by the reduced cardiac output during EFL, whereas the rest was due to other possible mechanisms described below.

In line with the results of Cunningham et al. (2000) and Ozyener et al. (2001) describing \( O_2 \) kinetics after heavy exercise, the off-transient \( \dot{V}O_2 \) data after EFL exercise were best fitted by a two component exponential function. The fundamental off-transient component after control exercise (40.6 s) was similar to that (~33 s) described by Cunningham et al. (2000) and Ozyener et al. (2001) following very heavy exercise, lasting as in the present study for 6 min. On the other hand, the time constant of the fundamental component after EFL exercise (26.6 s) was significantly shorter than that of control exercise, thus confirming replenishment of the body’s deprived oxygen stores upon removal of EFL. In addition, the shorter time constant calculated after EFL is the result of fitting the data from higher starting points as evidenced by the significantly higher \( a + c \) values shown in Table 4 and Fig. 5c.

Furthermore, the time constant of the slow component after EFL exercise (1,020 s) was twofold longer than the one (460 s) described previously for very heavy exercise (Ozyener et al. 2001) possibly reflecting the additive effects of the EFL-induced reduction in systemic \( O_2 \) delivery on the repayment of tissue \( O_2 \) debt. Although at present the mechanism(s) of the \( \dot{V}O_2 \) slow component is not fully understood, there are important factors that could influence the slow component after EFL exercise. These include the decline lactate concentration (Poole et al. 1994), the influence of the metabolic acidosis on the \( HbO_2 \) dissociation curve (Wasserman et al. 1991), the increased respiratory and cardiac muscle energy requirement associated with EFL exercise (Aaron et al. 1992; Harms et al. 1998), the progressive recruitment of type-II fibers (Coyle et al. 1992) and to a lesser extent the increased levels of circulating catecholamines (Gaesser et al. 1994) associated with the greater cardiovascular response during EFL exercise. Importantly, excessive expiratory muscle recruitment has been shown in patients with airflow limitation to increase the oxygen cost of breathing threefold (Aliverti et al. 2004b). Accordingly, it is reasonable to assume that the higher \( O_2 \) cost of breathing during EFL exercise would significantly enhance the \( O_2 \) debt as compared to exercise without EFL.

In the present study, we utilized a double-exponential function to describe the off-transient \( \dot{V}O_2 \) kinetics not only for EFL exercise but also for control exercise [typically fitted by a mono-exponential function (Cunningham et al. 2000; Ozyener et al. 2001)], in order to allow adequate comparisons of relevant parameters of recovery for both exercise tests. Accordingly, the slow component observed after control exercise was not discernible in two subjects using the double-exponential function. This is in accordance with previously reported data by Cunningham et al. (2000) who exercised healthy subjects at a similar work rate (100 W), yielding similar exercise \( \dot{V}O_2 \) (1.7 l min\(^{-1}\)) as the one reached in the present study during control exercise. In conditions where exercise is sustained at a moderate intensity, as in the control test, the off-transient slow component is often not discernible such that \( \dot{V}O_2 \) kinetics can retain first-order characteristics (Cunningham et al. 2000; Gerbino et al. 1996; Ozyener et al. 2001).

In conclusion, the results of the present study provide further evidence that expiratory flow limitation during exercise reduces systemic \( O_2 \) delivery, enhances the \( O_2 \) debt and leads to hypoxemia in part due to hypercapnia.

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