The Potential Role of Genetics, Environmental Factors, and Gut Dysbiosis in the Aberrant Non-Coding RNA Expression to Mediate Inflammation and Osteoclastogenic/Osteogenic Differentiation in Ankylosing Spondylitis

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Ankylosing spondylitis (AS) or radiographic axial spondyloarthritis is a chronic immune-mediated rheumatic disorder characterized by the inflammation in the axial skeleton, peripheral joints, and soft tissues (enthesis, fascia, and ligament). In addition, the extra-skeletal complications including anterior uveitis, interstitial lung diseases and aortitis are found. The pathogenesis of AS implicates an intricate interaction among HLA (HLA-B27) and non-HLA loci [endoplasmic reticulum aminopeptidase 1 (ERAP1), and interleukin-23 receptor (IL23R), gut dysbiosis, immune plasticity, and numerous environmental factors (infections, heavy metals, stress, cigarette smoking, etc.) The latter multiple non-genetic factors may exert a powerful stress on epigenetic regulations. These epigenetic regulations of gene expression contain DNA methylation/demethylation, histone modifications and aberrant non-coding RNAs (ncRNAs) expression, leading to inflammation and immune dysfunctions. In the present review, we shall discuss these contributory factors that are involved in AS pathogenesis, especially the aberrant ncRNA expression and its effects on the proinflammatory cytokine productions (TNF-α, IL-17 and IL-23), T cell skewing to Th1/Th17, and osteoclastogenic/osteogenic differentiation. Finally, some potential investigatory approaches are raised for solving the puzzles in AS pathogenesis.

Keywords: ankylosing spondylitis, HLA-B27, endoplasmic reticulum aminopeptidase, gut dysbiosis, IL-23/IL-17 axis, enthesitis, circular RNA, osteogenesis/osteoclastogenesis
**INTRODUCTION**

Ankylosing spondylitis (AS) and allied diseases, also known as radiographic axial spondyloarthritis (r-AxSpA), are a set of common immune-mediated inflammatory arthritides mainly affecting axial skeleton, particularly the sacroiliac joints, non-synovial spinal joints and enthesis (the connection between tendon and bone). Chronic pain with ankylosis of the spine and disability are the characteristics of AS/r-AxSpA. It is estimated that 0.5% of the world population are affected by AS, rendering it an important health-care and socioeconomic issue. Although the exact etiology and pathogenesis of AS/r-AxSpA remain obscure, genetic predisposition by human leukocyte antigen (HLA)-B27 subtypes is known to have a strong association with the disease. However, other genetic loci of non-major histocompatibility complex (MHC) including endoplasmic reticulum aminopeptidase (ERAPI) and interleukin 23 receptor (IL-23R), gut microbiome, local immunometabolomics in gastrointestinal (GI) tract and joints, as well as T cell plasticity may also be implicated in its pathogenesis (Voruganti and Bowness, 2020; Hwang et al., 2021). In addition to musculoskeletal manifestations, the extra-articular complications such as anterior uveitis (AU), interstitial pulmonary fibrosis, osteoporosis, syndesmophyte formation, and cardiovascular diseases (e.g., aortitis) may also occur (El Maghraoui, 2011; Stolwijk et al., 2015; Redeker et al., 2020). On the other hand, the effects of some environmental factors have been recognized through studying the relationships of different urinary tract infections and the contaminating inorganic compounds in the urine of AS patients. Shiue (2015) have reported significantly higher urine concentrations of cadmium, antimony, tungsten, uranium, and trimethylarsine in patients with AS. In a prospective cohort study, Zeboulou-Ktorza et al. (2013) have demonstrated a moderate statistical significance

![A model depicting the possible pathogenetic elements in patients with ankylosing spondylitis](image)

A model depicting the possible pathogenetic elements in patients with ankylosing spondylitis: Many genetic, environmental and personal factors are involved in the induction of gut dysbiosis and aberrant epigenetic regulation. Subsequently, these induced pathological changes may lead to intestinal inflammation, immunometabolic alterations and immune dysfunction. Finally, the musculo-skeletal and extra-articular manifestations ensue.
between vaccination and an elevation of Bath ankylosing spondylitis disease activity index (BASDAI) in AS. In spite of these factors, the investigations on the prevalence and influence of multi-morbidities in the disease activity of patients with AS revealed no direct cause-effect relationship between environmental factors and aberrant epigenetic regulation. Fitzgerald et al. (2020) have found that only the individual factors of metabolic syndrome are associated with more severe disease. Based on these backgrounds, we’ll discuss consecutively the possible molecular bases underlying the development of AS from viewpoints of genetics, gut dysbiosis, aberrant epigenetic regulation, and immune dysregulation to further understand the immunopathogenesis of AS. A proposed multifactorial co-morbid model in causing AS is depicted in Figure 1.

PROPOSED MOLECULAR PATHOGENETIC MECHANISMS FOR AS

At least five hypotheses to account for the molecular pathogenesis in AS patients have been proposed: 1) arthritogenic peptide stimulation, 2) unfolding protein response, 3) HLA-B*27 homodimer formation, 4) dysfunction of endoplasmic reticulum aminopeptidase (ERAP), and 5) gut inflammation caused by microbiota dysbiosis (Sharip and Kunz, 2020). We’ll first discuss in detail the link of genetics (HLA subtypes and non-HLA loci) with these hypotheses in the next section.

IMPLICATION OF GENETICS IN THE AS PATHOGENESIS

HLA-B27 positivity is present in 85–95% of patients with AS/r-AxSpA in different ethnicities in the world (Jamalyaria et al., 2017; Reveille, et al., 2019). However, only 5% of HLA-B27 (+) individuals in the general population have AS/r-AxSpA or undifferentiated spondyloarthropathy (USpA) (Akkoc and Khan, 2005). Nevertheless, HLA-B27 is still considered to be an important genetic factor highly associated with the development of AS. Six mechanisms have been suggested for the disease association; (A) The presentation of an arthritogenic peptide (Faham et al., 2017) to CD8+ T lymphocyte enriched in the inflamed joint (Gracey et al., 2020); (B) The presence of subtype HLA-B27 heavy chain, B*27:02, with a greater tendency to fold erroneously and accumulate in endoplasmic reticulum (ER)-derived vesicles. This may lead to a response to an unfolded peptide that can activate intracellular biochemical events and upregulate proinflammatory cytokines such as interferon-γ (IFN-γ), IL-17 and IL-23 (Jeanty et al., 2014; Jah et al., 2020; Navid et al., 2021); (C) A striking tendency of HLA-B27 heavy chains to adhere with each other, forming homodimers. These homodimers on the cell surface can be recognized by killer cell immunoglobulin-like receptor (KIR) and leukocyte immunoglobulin-like receptor (LILR) on natural killer (NK) cells (Lim Kam Sian et al., 2019); (D) HLA-B27 bearing individuals show impaired intracellular killing of pathogenic microorganisms that can lead to the persistence of intracellular bacterial pathogens and consequently stimulating proinflammatory cytokine production (Sahlberg et al., 2012); (E) Trimolecular complex (B27 heavy chain, α2 microglobulin and peptide) of HLA-B27 itself, free heavy chain, or homodimers of HLA-B27 may be recognized as neoantigens by the T cell receptor on CD8+ T lymphocytes in activating autoimmune responses (Boyle et al., 2001), (F) HLA-B27 bearing individuals generate an altered intestinal microbiome to increase HLA-B27 subtype expression involved in the immunopathogenesis of AS. These six potential effects conferred by HLA-27 subtypes are illustrated in Figure 2.

In addition to the HLA-B27 subtypes, genome-wide association studies (GWAS) have also identified more than 100 non-HLA-B27 loci associated with AS. These loci include machinery for antigen presentation (ERAP1 and ERAP2), some loci in Th17 cells (encoding IL-6R, IL-23R, TYK1 and STAT3) and others in macrophages and T cells (encoding IL-7R, CSF2, RUNX3 and GPR65) (Hanson and Brown, 2017; Kavadichanda et al., 2021; Wordsworth et al., 2021).

While over 90% of AS patients have an HLA-B*27 haplotype, only around 5% of individuals bearing HLA-B*27 develop AS. This implies a presence of additional risk factors to facilitate the disease development. It has been reported that strong epistatic gene-gene interactions between HLA-B27 and specific ERAP1 variants may work (Reveille, 2012). In addition, one of the major functions of ERAP1 is to trim endogenous peptides before their binding onto MHC-class I molecules. The epistatic interactions between ERAP1 and HLA allele may be involved in AS pathogenesis (Evans et al., 2011; Cortes et al., 2015). Furthermore, some reports have unveiled that ERAP1 can suppress both innate and adaptive immune responses (Goto et al., 2011; Aldhaman et al., 2013; Aldhamen et al., 2015). In experiments to evaluate bone morphogenesis of the axial skeletons in ERAP1−/− mice, the authors have found that ERAP1−/− mice can serve as a useful model for AS to observe spinal ankyloses, osteoporosis and inflammation. In addition, it can reduce both T helper 1 and T helper 2 differentiation and function (Wakkach et al., 2003). Finally, inflammation in the spine happens. The two ERAP genes, ERAP1 and ERAP2, are ubiquitously, zinc-dependent, and multifunctional, playing a role in several HLA-class I-mediated diseases in addition to AS (McGonagle et al., 2015; de Castro et al., 2016; Vitulano et al., 2017; Hansen et al., 2018). However, it is quite interesting that only functional polymorphisms of ERAP1 affect AS risk in HLA-B27 bearing individuals (Evans et al., 2011). ERAP2 appears independent from HLA-B27 (Robinson et al., 2015). Recently, authors discovered that a single nucleotide polymorphism (SNP), rs75862629, in the ERAP2 promoter region can influence the ERAP2 expression that can be counteracted by a higher expression of ERAP1 (Paladini et al., 2018). Furthermore, this SNP was found capable of modulating simultaneously the expression of both ERAP1 and ERAP2 and protecting hosts from AS in HLA-B27-positive individuals in Sardinia populations (Paladini et al., 2019). In contrast, a significant association between ERAP1 polymorphisms, rs30187 and rs27037, conferred an increased risk for AS in
A meta-analysis of ERAP1 gene polymorphism unveiled that SNPs rs27044 and rs30187 are significantly associated with AS susceptibility in Caucasians rather than Asians (Gao et al., 2020). Silencing of ERAP1 suppresses HLA-B27-free heavy chain expression and Th17 responses in AS (Chen L. et al., 2016; Nakamura et al., 2021). Moreover, the effects of ERAP1 polymorphisms on the proinflammatory and anti-inflammatory cytokine expressions in AS patients have also been investigated. It was found that T allele of rs30187 and C allele of rs2287987 were associated with risk of HLA-B27-positive AS development by significant overexpression of proinflammatory cytokines (IL-17A, IL-17F, IL-23, TNF-α and IFN-γ) and under-expression of anti-inflammatory cytokines (IL-10 and TGF-β) in PBMC of HLA-B27 (+) AS patients (Babaie et al., 2020).

On the other hand, the studies exploring the association between TNF polymorphisms and AS remain inconclusive. A meta-analysis has revealed that the A allele in TNF-238 and TNF-308, the C allele in TNF-1031, the T-allele in TNF-850 and rs769138 are significantly associated with AS susceptibility in the total population (Hu et al., 2021). In addition, IL-12B gene polymorphism (Ivanova et al., 2019), IL-23R SNPs and IL-10-819 polymorphism (Xia et al., 2018) are associated with AS pathogenesis.

In short summary, the aminopeptidases, ERAP1 and ERAP2, trim the peptides to a length suitable for fitting into the groove of MHC class I molecules for protection from viral infection. The epistatic interactions between HLA-B27 peptide repertoires can determine the innate immunological function of HLA-B27 such as antigen presentation to T cells. However, the process also produces a by-product, an intracellular misfolded HLA-B27, or an HLA-B27 homodimer on the cell surface, which can elicit ER stress responses, autophagic engulfment, or innate immune responses. These aberrant interplays between HLA-B27 and...
ERAP1/ERAP2 result in a deviation of the physiological function from defending against infections to pathological induction of spondyloarthritis (Vitulano et al., 2017). Moreover, the accumulation of misfolded HLA-B27 heavy chain along with β2-microglobulin and ER chaperones (calnexin, calreticulin, BiP, 94kD glucose-regulated protein) into ER-derived vesicles is different from the peptide-loading complex. These abnormal behaviors may become the unique features of HLA-B27 subtypes predisposing to AS (Jah et al., 2020).

**ROLES OF HAZARD HLA ALLELE-ASSOCIATED GUT DYSBIOSIS AND ITS METABOLITES IN THE PATHOGENESIS OF AS**

Sufficient evidence has demonstrated the pathogenic roles of gut microbiome in inflammatory arthritides including SpA. Human gut is colonized with bacteria, viruses, and fungi, which actively interact with each others (Li M. et al., 2019). More than 100 trillion bacteria reside in the mammalian gut to establish a symbiotic relationship. This intimate association can influence many aspects of the host’s metabolism, physiology and immunity. Accordingly, intestinal dysbiosis may play an important role in the development of AS by altering intestinal permeability, stimulating immune responses, and exerting molecular mimicry (Yang et al., 2016).

To determine whether AS patients’ guts carry a distinct microbial signature from that in the healthy individuals, 16S ribosomal RNA sequences of microbiome in the terminal ileum were analyzed. A higher abundance of five families of bacteria (Lachnospiraceae, Ruminococcaceae, Rikenellaceae, Porphyromonadaceae, and Bacteroidaceae) were reported (Costello et al., 2015). In addition, a quantitative metagenomic study based on the shotgun sequencing of the gut microbial DNA in Chinese AS patients has revealed the increases in Prevotella melaninogenica, Prevotella coppei, and Prevotella spp. C561, but a decrease in Bacteroides spp. In addition, Bifidobacterium genus, which is commonly used in probiotics, was found accumulated in AS patients (Wen et al., 2017). An amplicon gene sequencing study on 16S ribosomal RNA disclosed the genus Dializer as a microbial marker for disease activity in SpA (Tito et al., 2017). Recently, by using real-time polymerase chain reaction (PCR) to analyze intestinal microbiota in stool samples, investigators have found that the total quantity of bacteria is decreased in patients with AS. The intestinal dysbiosis is associated with a more severe articular disease as evidenced by the findings that Bifidobacterium and Lactobacillus were increased in active AS patients (Cardoneanu et al., 2021). Besides, gut microbiota and mycobiota in AS patients were detected by 16S rRNA gene and ITS2-based DNA sequencing. It revealed that Proteobacteria was increased and Bacteroides was decreased. This abnormality was resulted from enrichment of Escherichia-Shigella, Veillonella, Lachnospiraceae NK4A136 group, and reduction of Prevotella strain 9, Megamonas, and Fusobacterium. On the other hand, the mycobiota in alimentary tract of AS patients, as shown by 16S rRNA gene analyses, exhibited higher expression of Ascomycota (especially the class of Dothideomycetes) and lower expression of Basidiomycota, mainly because of a decrease in Agaricales (Li X. et al., 2019). These results may imply that decreased ITS2/16s biodiversity ratios and altered bacterial-fungal inter-kingdom networks somewhat contribute to the pathogenesis of AS. The gut dysbiosis in AS may cause bowel inflammation as reflected by increased fecal calprotectin levels (Klinberg et al., 2019).

Since HLA-B27-associated SpA relevant to an altered gut microbiota and bowel inflammation, it is interesting to search for the cause-effect relationships among HLA-B27, gut dysbiosis and host/bacterial metabolites. Sufficient evidence indicates HLA alleles (HLA-B27 and HLA-DRB1) can affect gut microbiota as reported in rat models of SpA and RA (Asquith et al., 2017; Gill et al., 2018) as well as human AS and RA (Asquith et al., 2019; Xu and Yin, 2019). The HLA-B27-induced gut dysbiosis and zonulin upregulation may change microbiota-derived metabolites and antigens. These changes can further alter vascular barrier of gut epithelium (Ciccia et al., 2017) and deregulate intestinal immune system to activate IL-23/IL-17, IFN, TNF-α and IL-1 expression (Gill et al., 2018) in AS patients. In rat model of SpA, both microbial and host metabolites are altered. These metabolites include amino acid, carbohydrate, xenobiotics and medium-chain fatty acid. Thus, upregulation of histidine, tyrosine, supermidine, N-acetylmuramate and glycercate can be found in HLA-B27/β2m rats. The HLA-B27 presentation is also associated with altered host expression of microbial metabolite receptor genes such as FFAR2, FFAR3 and NACR1 (Asquith et al., 2017). The studies on fecal signatures of AS patients with either gender have further revealed differences in steroid metabolites (He et al., 2019). These experiments have shown that male-specific fecal signatures include cholestan-3-ol, tocopherol, stigmastan-3,5-diene, cholest-3-ene, cholest-4-en-6-one and 1-heptatriacotanol. In contrast, the female-specific fecal signatures are ergost-5-en-3-ol acetate and D-myo-inositol. These results may indicate gender-attributed fecal signature differences between males and females, reflecting AS gender features. Metagenome-wide association study on the alterations in the gut composition has disclosed that AS subjects harbor more bacterial species associated with carbohydrate metabolism and glycan biosynthesis in their feces than normal individuals. They also express bacterial profiles with less liability to degrade xenobiotics biologically or to synthesize and transport vitamins (Huang et al., 2020). To understand thoroughly the molecular mechanisms linking intestinal microbial dysbiosis, intestinal inflammation and Th17 immunity in patients with AS, Berlingberg et al. (2021) conducted LC-MS-based metabolomic screening and shotgun metagenomic measurements in paired colon biopsies and fecal specimens. The authors also showed significant alterations in metabolites in tryptophan pathway that can increase indole-3-acetate (IAA) and indole-3-acetaldehyde (I3Ald) in AxSpA. The shotgun metagenomics confirmed abundance of numerous enzymes involved in tryptophan metabolism such as indole pyruvate decarboxylase. These enzymes can enhance significantly the generation of IAA and I3Ald to facilitate tryptophan synthesis. Tryptophan and its metabolites in this particular gut microbiome may disturb immune functions and further expedite the development of...
AxSpA. Moreover, gut microbiome may evolve together with the human hosts and provide them with a myriad of molecules such as microbe-associated molecular patterns (MAMPs) to augment intestinal inflammatory processes. In association with damage-associated molecular patterns (DAMPs), the inflammatory processes may be triggered further in the already ongoing pathological status such as HLA-B27-associated acute anterior uveitis (Rosenbaum and Asquith, 2018; Lu Yang et al., 2021). These data support that the invisible “essential organs” can communicate with each other and “talk” across with host immune cells in healthy individuals as well as in AS patients. The effects of HLA-B27 risk alleles on gut microbiota and metabolomic changes in patients with AS are depicted in Figure 3.

**IMMUNE DYSFUNCTIONS IN PATIENTS WITH AS**

Many studies have shown that various immune-related cells, via their secreting cytokines and molecules, may play crucial roles in
AS pathogenesis (Mei et al., 2011; Liu et al., 2015; Madej et al., 2015; Sveas et al., 2015; Wang et al., 2016; Yang et al., 2016; Wang et al., 2018). These data indicate that both innate and adaptive immune cells are involved in AS pathogenesis. The innate immune cells include dendritic cells, macrophages and natural killer cells. The adaptive immune cells include helper T, Treg, CD8\(^+\)T, and B cells (Rezaianemesh, et al., 2018). Much evidence has demonstrated that certain phenotypes of resting and activated macrophages expressing scavenger receptor, CD163, can link between immune alterations of the gut and synovial inflammation in AS (Baeten et al., 2002; Slobodin et al., 2012; Talpin et al., 2014; Wright et al., 2016; Rezaianemesh et al., 2017). The abnormal polarization of macrophages induced by IL-4 was found in AS patients (Lin et al., 2015). Besides, the studies on immune dysfunctions of T cell subpopulations are prosperous. These results revealed increased frequency of Th2 (Yang et al., 2004) and Th17 (Jandus et al., 2008; Shen et al., 2009; Xueyi et al., 2013), abnormal form of HLA-B27 expression on CD4\(^+\) T cells (Boyle et al., 2004), defective function of CD24\(^+\)CD38\(^+\) regulatory B cells (Chen M. et al., 2016), and expansion of CD4\(^+\)CD28\(^{high}\) Treg cells (Ciccia et al., 2010). Recently, two molecules on T lymphocytes, T cell immunoglobulin and mucin-domain-containing molecule 3 (Tim-3) and programmed death-1 (PD-1) for negative regulation of immune responses, attracted investigators to focus on AS pathogenesis (Zhou et al., 2015). PD-1 expression in T cells was reported to inversely relate to the spinal radiologic changes in Taiwanese patients with AS (Chen et al., 2011). Tim-3 polymorphism could result in down-regulation of the expression of itself and be involved in AS susceptibility (Wang et al., 2014). Furthermore, other investigations revealed a decreased expression of PD-1 on CD8\(^{+}\) T cells in AS patients. The deficiency may activate immune responses in AS patients (Duan et al., 2017). More recent investigations have disclosed that Tim-3\(^{+}\)CD8\(^{+}\) and PD-1\(^{+}\)CD8\(^{+}\) T cells can produce more IL-10 than other subsets.

Another studies have revealed that either low percentage (Wu et al., 2011; Zhao et al., 2011; Xueyi et al., 2013) or functional impairment (Guo et al., 2016; Wang et al., 2018) in CD4\(^+\) Treg may be present in AS patients. On the contrary, meta-analyses have unveiled that only the proportions of CD4\(^+\)CD25\(^{high}\)FOXP3\(^{+}\) and PD-1\(^{+}\)CD8\(^{+}\) T cells can produce more IL-10 than other subsets. These IELs can secrete increased amount of IL-1\(\beta\) in addition, the intra-epithelial lymphocytes (IELs) are the T cells within intestinal epithelium which can also secrete IL-17A and other proinflammatory cytokines (IL-1\(\beta\), TNF-\(\alpha\), and IFN-\(\gamma\) to mediate tissue inflammation. Furthermore, both regulatory T cell (Treg) and regulatory B cell (Breg) hypofunctions may also participate in the immune dysregulation in AS patients.

Several studies have been published in recent years demonstrating the pivotal role of gut-microbiota and IL-23/IL-17 axis in the AS pathogenesis (Jandus et al., 2008; Shen et al., 2014; Babiae et al., 2018). Enthesis inflammation (enthesitis) was demonstrated to be IL-23-dependent (Sherlock et al., 2012; Chen et al., 2014) and likewise IL-17-dependent (Shabgah et al., 2017). Moreover, investigations from the blood samples of AS patients revealed the abundance of Th17 (Shen et al., 2009; Zhang et al., 2012), Th22 (Zheng et al., 2012) and γ/δ T cells (Kenna et al., 2012) with high levels of IL-17 in the circulation (Wendling et al., 2007; Mei et al., 2011; Lin et al., 2015). The innate lymphoid cells (ILCs) can stimulate inflammation in the gut with respect to AS. NKp44\(^{+}\) ILC3 cells were found expanded in the intestine, synovial fluid, bone marrow and peripheral blood of patients with AS to produce IL-17, IL-22 (Ciccia et al., 2015), and granulocyte-macrophage colony-stimulating factor (GM-CSF) but not IL-17A in the inflamed joints (Blijdorf et al., 2019). The intraepithelial lymphocytes (IELs) are another T cell subpopulation within the intestinal epithelium in close contact with bacteria that can be affected by intestinal microbiota. The total number of IELs is significantly decreased in AS due to a decrease in TCR \(\beta\) IELs. These IELs can secrete increased amount of IL-1\(\beta\), IL-17A and IFN-\(\gamma\) in Crohn’s disease and significantly enhance the amount of TNF-\(\alpha\) in AS (Regner et al., 2018). All of these results may suggest a correlation between altered microbiota and IEL function in AS.

The pathogenesis of AS is characterized by a predilection of adaptive immunity toward IL-23/IL-17 axis with the presence of a polarization stimulator for Th17 response. As a result, the IL-17 and TNF-\(\alpha\) production are enhanced. However, failure of IL-23 blockade in the treatment of spinal polyarthritis but not...
peripheral enthesitis has ever been encountered (McGonagle et al., 2021). Thus, the importance of IL-23 pathway in AS pathogenesis awaits further evaluations. A recent study has even demonstrated high levels of IL-7 mRNA and peptide in the peripheral type SpA (Rihl et al., 2008). IL-7 belongs to a hematopoietin cytokine family with a molecular weight of 17.4 kDa (Sutherland et al., 1989). It can stimulate Th17 cells, innate immune cells like γ/δ T cells (Michel et al., 2012) and mucosa-associated invariant T (MAIT) cells (Tang et al., 2013) to produce proinflammatory cytokines including IL-17. In addition, these innate-like T cells rather than Th17 cells have been proved to be the main source of IL-17A (Venken and Elewaut, 2015; Debusschere et al., 2016). It appears that IL-7 is more important than IL-23 in the polarization of type 17 (IL-17) signature because IL-7 receptor is present in the key cells of innate immunity that are essential for the polarization of type 3 (IL-3) response and SpA (Goncalves and Duarte, 2019). The immune dysfunctions in patients with AS are illustrated in Figure 4.

PATHOGENESIS OF ENTHESISITIS IN AS PATIENTS

Enthesis is regarded as the region where tendon attaches to bone. In broad sense, the enthesal tissues include fibrocartilage, bursa, fat pad, deeper fascia, adjacent trabecular bone networks and enthesis. These tissues play an anchorage between mobile organs and stress resistance (Benjamin and McGonagle, 2009). It has been recognized that enthesitis become the primary pathological process underlying SpA-associated skeletal inflammation (Watad et al., 2018). Normal enthesal tissues contain group 3 NKp44+ ILCs, γ/δ T cells, conventional CD4+ and CD8+ T cells, and cells of myeloid lineage. These cells are capable of producing prostaglandins, different growth factors and proinflammatory cytokines (TNF-α and IL-17) for physiological tissue repair and homeostasis (Reinhardt et al., 2016; Cuthbert et al., 2017; Cuthbert et al., 2019; Watad et al., 2021). It appears that IL-7 is more important than IL-23 in the polarization of type 17 (IL-17) signature because IL-7 receptor is present in the key cells of innate immunity that are essential for the polarization of type 3 (IL-3) response and SpA (Goncalves and Duarte, 2019). The immune dysfunctions in patients with AS are illustrated in Figure 4.

PATHOGENESIS OF OSTEOPOROSIS AND OSTEOGENESIS IN AS PATIENTS

A mystery of skeletal damage in patients with AS is the consequence of bone destruction followed by the new bone formation. The inflammation-induced osteoporosis in the spine and peripheral bones is quite common in the early stage of AS. It can cause trabecular bone weakness and lead to an increase in spinal fracture rate (Davey-Ranasinghe and Deodhar, 2013). Enhanced expression of IL-17 in the serum and synovial fluid has been reported implicating in the bone loss of AS patients (Akgöl et al., 2014). However, another puzzle of proinflammatory cytokine, IL-17 family, effecting on the bone metabolism, arises since IL-17A can enhance both bone loss and osteogenesis.

Investigations have unraveled that IL-17 can activate osteoclasts (OCs) to express RANK ligand (RANKL), which then reciprocally stimulates OCs themselves by RANK-RANKL interaction and induces bone absorption (Page and Miossec, 2005; Miossec, 2009). IL-17A-stimulated miR214 expression in OCs is an important inhibitor for bone formation in AS patients (Wang et al., 2013; Zhao et al., 2015; Liu et al., 2020). Nevertheless, adding of exogenous IL-17A into cultured normal primary bone-derived cells (BdCs) promotes OC activity and differentiation as evidenced by increased alkaline phosphatase (ALP) activity through JAK2/STAT3 pathway (Jo et al., 2018a; Wang et al., 2018).

On the other hand, abnormal bone remodeling with excessive new bone formation can cause syndesmophytes or even “bamboo spine” to limit the spinal motility in AS patients. Many studies have revealed that higher bone morphogenetic proteins (BMP), lower Dickkopf-1 (DKK-1) levels (Liao et al., 2018), and increased ALP activity (Jo et al., 2019) in serum are parallel to the accelerated osteogenesis in the BdCs derived from AS patients (AS-BdCs) (Jo et al., 2018b; Kang et al., 2018). For investigating the relevance of ALP to the regulation of osteoblast (OB) differentiation in AS patients, ALP was inhibited in AS-BdC culture. A remarkable suppression of the master transcriptional factor in OB, RUNX2, was observed. This implies that RUNX2 can regulate promoter activity of ALP through a positive ALP-RUNX2 feedback mechanism (Jo et al., 2019). To further identify the role of HLA-B27 in syndesmophyte formation in AS, the mesenchymal stem cells (MSCs) obtained from enthesis of AS patients were studied. The results demonstrated that HLA-B27-mediated activation of the SXBP1/RARB/TNAP (tissue nonspecific alkaline phosphatase) axis is essential in the development of syndesmophyte in AS patients (Liu C.-H. et al., 2019). Furthermore, the expression of miR-146a is upregulated and DKK1 is down-regulated respectively in capsular tissue of the hip in AS patients. Therefore, a negative correlation was displayed between the expressions of miR-146a and DKK1. Further investigations disclosed that miR-146a could inhibit DKK1 expression by directly targeting 3′-UTR region of DKK1 (Di et al., 2018). In addition, the level of miR-17-5p is significantly elevated in fibroblasts and ligament tissues, which is assumed to be targeting the 3′-UTR of ankylosis protein homolog (ANKH). This may subsequently increase osteogenesis in AS patients. Down-regulation of miR-17-5p slowed AS progression via...
regulation of DKK1 and VEGF. These findings have verified the role of miR-17-5p-ANKH axis in regulating heterotopic ossification in AS patients (Qin et al., 2019).

The transgenic SpA-associated and non-SpA-associated HLA-B27 subtypes in Drosophila revealed an antagonistic interaction of HLA-B27 against activin receptor-like kinase-2 (ALK2). This antagonistic interaction may exert inhibitory effects on TGF-β1/BMP signaling pathway at the cross-road between inflammation and ossification and become a putative mechanism for HLA-B27-mediated SpA development (Grandon et al., 2019). The molecular basis underlying the dilemma in inflammation-mediated bone metabolism in AS patients is shown in Figure 6.

In addition to the above mentioned factors contributing to AS pathogenesis, it is believed that the environmental factors might affect the hereditable epigenetic regulation of the down-stream gene expression in developing human diseases. These may include infectious, autoimmune/inflammatory, or neoplastic diseases. Recently, (Ghafouri-Fard et al., 2021a), have unveiled the interaction between ncRNAs and Toll-like receptors (TLRs) in transducing both MyD88-dependent and TRIF-dependent signaling cascades to induce human inflammatory and autoimmune disorders. Furthermore, the same group have found in the literature that both miRs and lncRNAs can regulate bone development processes including osteogenesis. Both intramembranous and endochondrial ossification of osteogenesis were observed (Ghafouri-Fard et al., 2021b). miRs were found to exert their actions through both Wnt/β-catenin and TGF-β/BMP pathways whereas lncRNAs worked as molecular sponge for binding miRs to directly affect these pathways and osteogenic transcription factors. The examples include MALAT1/miR-30, MALAT1/miR-214, LEF1-AS1/miR-24-3P, MCF2L-AS1/miR-33a, MSC-AS1/miR-140-5P, and KCNQ1OT1/miR-214. It is quite interesting that nuclear factor-kappa B (NF-kB) represents a group of inducible transcription factors (TFs) to regulate gene expression implicated in the immune responses. NF-kB can functionally interact with ncRNAs to construct an intricate NF-kB-miRs-lncRNAs network in regulating down-stream gene expression in different aspects. This type of network interactions among miR-146a/b, MALAT1, NKILA and NF-kB have been reported in the pathogenesis of some inflammatory conditions (Ghafouri-Fard et al., 2021c). Collectively, these data can provide some clues to support the interactions among environmental factors, gut

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**FIGURE 5** Dissection of immune-related cells in normal enthesis and AS enthesitis. In normal enthesis, many different cell populations may release physiological amount of prostaglandins (PGs), TNF-α, IL-17 and various growth factor (GFs) to maintain tissue repair and homeostasis in these biomedical stressed-entheses. However, in HLA-B27 risk allele (+) AS patients, the aberrant IL-17 production can cause inflammation in enthesis (enthesitis) as well as extra-articular manifestations such as aortitis, acute anterior uveitis, and interstitial lung fibrosis. PAMP, pathogen-associated molecular pattern; DAMP, damage-associated molecular pattern; NKP44, Natural cytotoxicity triggering receptor 2 or CD336.
dysbiosis, aberrant ncRNA expression and inflammation/autoimmunity in the development of AS. We are going to discuss in detail the aberrant epigenetic regulation in AS pathogenesis in the next section.

**ABERRANT EPIGENETIC REGULATION IN AS**

Epigenetics is a study on the heritable changes in gene expression which takes place without an alteration in DNA sequence but with modulations of chromatin-associated molecules caused by environmental factors. These environmental factors include dietary nutrition, lifestyle, exercise/physical activity, drugs/toxins and other miscellaneous contributory factors (Abdul et al., 2017; Heinbockel and Csoka, 2018; Martin and Fry, 2018). In general, epigenetic study may include DNA methylation, histone acetylation/deacetylation and circular RNA (cRNA) regulation (Berlingberg and Kuhn, 2020).

DNA methylation is an epigenetic modification with addition of methyl groups to cysteine or adenine residue to control gene transcription (Dor and Cedar, 2018). Many factors can affect DNA methylation such as age, sex, smoking, medications, alcohol and nutrition-diet (Whyte et al., 2019). DNA methyltransferase 1 (DNMT1) is an enzyme that regulates methylation of cytosine residues. Decreased DNMT1 expression can increase gene expression. Previous studies have shown that the expression level of DNMT1 in AS patients is significantly down-regulated which is associated with hypermethylation of the promoter region of DNMT1 (Aslani et al., 2016). These results suggest that the dysregulation of DNMT1 expression via altered methylation level of the other target genes may contribute to AS pathogenesis. Recently, the genome-wide DNA methylation profile analysis identified many altered DNA methylation sites in the peripheral

![Diagram of epigenetic regulation in AS](https://example.com/diagram.png)
blood mononuclear cells (PBMCs) of AS patients. These sites include hypermethylation of HLA-DQB1 (Hao et al., 2017), hypermethylation of CpG3 and CpG5 in B-cell chronic lymphocytic leukemia/lymphoma 11B (BCL11B) (Karami et al., 2017), and hypermethylation of GTPase-related genes (Coit et al., 2019). However, HLA-B27 bearing patients with AS were found to have some hypomethylated DNA promoters in HCP5 tubulin folding cofactor A (TBCA) and phospholipase D family member 6 (PLD6) (Coit et al., 2019).

To elucidate the epigenetic regulation of immune dysfunctions in AS, the DNA methylation profile of blood cells was analyzed. Hypermethylation of the promoter in interferon regulatory factor 8 (IRF8) (Chen et al., 2019) and 2CpG islands of IL-12B (IL12B-1 and IL12B-2) (Zhang et al., 2019) were found. Both IFN-γ and IL-12 are crucial cytokines in suppressing Th17 cell proliferation and differentiation, which contribute in consequence to reduce severity of AS. Further investigations disclosed that hypermethylated miR-34b promoter leads to upregulation of miR-34b, thus inhibiting the IL-12B gene expression and alleviating disease activity of AS (Meng et al., 2021).

On the other hand, histone modification allows activation (euchromatin) and deactivation (heterochromatin) of chromatin by two enzymes, histone acetyltransferase (HATs) and histone deacetylase (HDACs) (Allis and Jenuwein, 2016). In PBMC study, decrease in and imbalance between HAT and HDAC activities were present in AS patients, compared to the healthy controls (Toussirot et al., 2013). The aberrant DNA methylation and histone modifications in PBMC are depicted in Figure 7.

FIGURE 7 | The contribution of aberrant epigenetic regulations in AS pathogenesis including histone modifications, DNA methylation and ncRNA expression. Decreased both HATs and HDACs expression, and therefore imbalanced HATs/HDACs ratio are found in AS. Moreover, hypermethylation of HLA-DQB1, CpG3 and CpG5 isolates of BCL11B (B cell chronic lymphocytic leukemia IIIB) and GTPase-related genes are found. In contrast, DNA methyltransferase (DNMT1) promoter region hypermethylation was found in AS patients. This can cause hypermethylation in the promoter region of IFN-γ and IL-12 and conversely enhance IL-17 production. On the contrary, hypermethylation of miR-34b promoter increase IL-12 production to suppress IL-17 expression. The abnormal ncRNA expressions in the intracellular and extracellular parts with their modes of action are presented in Tables 1–3.

The Characteristics of Non-Coding RNAs and Their Roles in the Pathogenesis of and Clinical Applications in AS Patients
ncRNAs are single-stranded RNAs composed of microRNAs (miRs, with 20-24 nucleotides) and long non-coding RNAs (lncRNAs, with more than 24 nucleotides and less than 300 nucleotides). They regulate gene expression and therefore are involved in physiological and pathophysiological processes. They exist in the cells, extracellular fluid and cell-derived exosomes in a stable form. In addition, lncRNAs can act as sponge for modulating miR functions. Accordingly, ncRNA expression profiles can serve as biomarkers for disease activity, pathogenesis, prognosis and therapeutic monitoring of the diseases (Mohammadi et al., 2018; Berlingberg and Kuhn, 2020; Motta et al., 2020). Besides their molecular stability, the characteristic tissue specificity, easy obtainability from different biological fluids (plasma, saliva, urine, synovial fluid and other tissue fluid), and powerful discrimination render ncRNA profiling a useful tool in studying autoimmune, inflammatory and neoplastic diseases (Park et al., 2009; Pauley et al., 2009). Therefore, we will discuss in detail the contribution of ncRNAs to the deranged T cell responses, inflammation, altered bone homeostasis and monitoring of disease activity in AS.
Lai et al. (2013) discovered that three ncRNAs, miR-16, miR-221 and let-7i, were over-expressed in T cells from AS patients (AS-T). TLR-4 has been confirmed to be the target molecule of let-7i in AS-T cells. In addition, increased expression of let-7i enhanced IFN-γ production in AS patients. miR-221 and let-7i were also associated with disease activity of lumbar spine (as calculated by BASRI) in AS. Huang et al. (2014) found that significant higher expression of miR-29 in PBMCs of AS patients, although not correlated to disease activity, could be used as a useful diagnostic biomarker in new bone formation. These data imply that miR-29 might become a useful marker for new bone formation in AS patients as evidenced by its ability to regulate DKK-1 in Wnt signaling pathway.

Fogel et al. (2019) investigated the miR expression in both CD14+ monocytes and CD4+ T lymphocytes from AS patients. The group found downregulated miR-361-3p, miR-223-3p, miR-384, and miR-16-5p in monocytes and upregulated miR-16-1-3p, miR-28-5p, miR-199a-5p, miR-128-3p in T lymphocytes that might contribute to AS pathophysiology. Li X et al. (2019) reported elevated expression of miR-17-5p, miR-27a, miR-29a and miR-126-3p in PBMCs of axial SpA, which might be regarded as useful diagnostic markers in AS. Furthermore, Yang et al. (2019) provided evidence that miR-335-5p, miR-27a and miR-218 would predispose syndesmophyte formation in AS patients. Recently, Ni and Leng, 2020 have found that miR-495 in PBMC, whole blood, and serum is downregulated because its promoter region is highly methylated in AS patients. Besides, the miR-495 expression is negatively associated with programmed cell death protein 10, PDCD10. This may indicate PDCD10 expression can be targeted by miR-495 in AS. Bioinformatic analyses and signaling pathway studies have revealed that miR-495 can down-regulate β-catenin and TGF-β1. The intracellular ncRNA expressions in immune cells (PBMCs, T cells or monocytes), their modes of action and clinical applications in AS are summarized in Table 1.

The extracellular ncRNAs may include those in circulatory tissue fluid (serum, plasma, etc.) and tissue-derived (exosomes) ones. Qian et al. (2016) unraveled that serum miR-146a and miR-155 were significantly up regulated in AS. Moreover, the serum level of miR-155 is associated with disease activity and the severity of thoracolumbar kyphosis secondary to AS. Prajzlerová et al. (2017) found that miR-625-3p can reflect disease activity in AS with spinal involvement. Moreover, miR-29a-3p, miR-146a-5p and miR-222-3p are involved in extracellular matrix formation and inflammation, and are associated with spinal changes and disease activity (BASDAI) in AS patients. These dysregulated miRs are also suggestive of their potential as biomarkers for disease progression. Perez-Sanchez et al. (2018) unclosed that higher expression levels of miR-146a-5p, miR-125a-5p, miR-151a-3p and miR-22-3p and lower expression levels of miR-150-5p and miR-451a were present in the AS plasma. Bioinformatic analysis has revealed that these six miRs target proinflammatory and bone remodeling genes. Besides, miR-125a-5p, miR-151a-3p, miR-150-5p and

| Cell/Body fluid | ncRNA | Mode of action | Biomarker | References |
|-----------------|-------|----------------|-----------|------------|
| T cell          | miR-16↑ | IFN-γ↑ | Disease activity in lumbar spine | Lai et al. (2013) |
|                 | miR-221↑ |          |           |            |
|                 | IncRNA let-7i↑ |        |           |            |
| PBMC            | miR-29↑ | DKK-1↑ | Disease activity | Huang et al. (2014) |
|                 |         | β-catenin↑ |          |            |
|                 |         | RUNX2↑ |          |            |
|                 |         | GSK-3β↑ |          |            |
| CD4+ Mφ         | miR-361-3p↓ |        | AS pathogenesis | Fogel et al. (2019) |
|                 | miR-223-3p↓ |        |           |            |
|                 | miR-484↓ |          |           |            |
|                 | miR-16-5p↓ |          |           |            |
|                 | miR-16-1-3p↓ |        |           |            |
| CD4+ T          | miR-28-5p↑ |          |           |            |
|                 | miR-199a-5p↑ |        |           |            |
|                 | miR-128-3p↑ |        |           |            |
| PBMC            | miR-17-5p↑ |          | Diagnostic biomarker | Li et al. (2019a) |
|                 | miR-27a↑ |          |           |            |
|                 | miR-29a↑ |          |           |            |
|                 | miR-128-3p↑ |        |           |            |
| PBMC            | miR-335-5p↑ |          | New bone formation | Yang et al. (2019) |
|                 | miR-27a↑ |          |           |            |
|                 | miR-218↑ |          |           |            |
| PBMC/ Serum     | miR-495↑ | PDCD10↑ | New bone formation | Ni and Leng, (2020) |
|                 |         | β-catenin↑ |           |            |
|                 |         | TGF-β1↑ |           |            |

Mφ, macrophage; PBMC, peripheral blood mononuclear cell; CD, cluster of differentiation; IFN, interferon; RUNX, Runt related transcriptional factor; PDCD, programmed cell death protein; GSK, glycogen synthase kinase; TGF, transforming growth factor; DKK, Dickkopf related protein; ↑ also appearing in serum.
miR-451a expression are related to the presence of syndesmophytes in AS. Accordingly, these six plasma miR signature can become gorgeous non-invasive biomarkers for AS diagnosis. Recently, Reyes-Loyola et al. (2019) assessed plasma levels of ncRNAs in Mexican AS patients. They found plasma Inc let-7 was higher in patients and might serve as a diagnostic biomarker in Mexican AS patients. On the other hand, plasma miR-16 level is inversely correlated to ASDAS-CRP score and MMP-1 level, thus, serving as disease activity marker. Li Y et al. (2020) have further found that plasma IncRNA MEG3 is downregulated and negatively correlated to the levels of IL-1β, IL-6 and TNF-α in AS patients, and can block the inflammatory response of the immune cells in AS patients. Conversely, plasma miR-146a was upregulated and positively correlated to the proinflammatory IL-1β, IL-6 and TNF-α. The authors also clarified that over expression of miR-146a could revert the inhibitory effect of abnormal MEG3 expression on inflammatory cytokines. These data imply that IncRNA MEG3 plays an anti-inflammatory role via targeting miR-146a and thus can provide a new potential therapeutic role for AS treatment. In further elucidation of the molecular mechanism for the therapeutic potential of IncRNA MEG3, Ma et al. (2020) found that the expression levels of MEG3 and sclerostin (SOST) are decreased but IncRNA let-7 is increased in AS patients. Their results confirmed that MEG3 can interact with (sponge) let-7i in AS fibroblast and promotes SOST expression to restrain the progression of AS. This would provide a new treatment modality in AS. Fotoh et al. (2020) discovered a higher expression of miR-125a and a lower expression of miR-451a in the plasma from active Egyptian AS patients. Interestingly, both miRs were able to distinguish AS patients with a structural damage and could be used as sensitive diagnostic, prognostic and disease burden biomarkers for AS patients. The extracellular (serum or plasma) ncRNA expression, their modes of action and clinical application in AS patients are listed in Table 2.

In summary, these aberrant soluble extracellular ncRNAs expression in AS patients can be divided into three categories in clinical practice as shown below;

1) Biomarkers for disease diagnosis: IncRNA let-7, miR-146a-3P, miR-125a-5P, miR-151a-3P, miR-22-3P, miR-150-5P, and miR-451a.
2) Biomarkers for disease activity: miR-146a, miR-155, miR-625-3P, miR-29a-3P, miR-146a-5P, miR-22-3P, and IncRNA-MEG.
3) Biomarkers for both diagnosis and disease activity: miR-16a, miR-146a, miR-125a, and miR-451a

### The Role of ncRNAs in Enthesitis and Ligament Inflammation

Enthesis is the characteristic sites where pathological processes occur in AS patients, causing enthesitis. To reflect ossifications more realistically in the ligaments of AS patients, the epigenetic regulation of the cultured ligament-derived fibroblasts were analyzed for osteogenic differentiation. Zhang et al. (2017) compared miR, IncRNA and mRNA profiles in hip joint ligament tissues from AS patients. The authors identified that miR-17-5p and miR-27b-3p could increase the potentials of osteogenic differentiation in ligament fibroblasts of the hip joint. Tang et al. (2018) isolated ligament fibroblasts from AS patients and induced them to differentiate into osteoblast (OB). During osteogenic differentiation, miR-124, β-catenin, osteorix and RUNX2 expression gradually increased, while that of GSK-3β gradually declined. Zhao et al. (2020) have demonstrated that miR-204-5p can negatively regulate NOTCH 2 expression in osteogenic differentiation in the ligament fibroblasts derived from AS patients in vitro. These results may provide a

| Source | ncRNA | Mode of action | Biomarker | References |
|--------|-------|----------------|-----------|------------|
| Serum  | miR-146a | Extracellular matrix formation and inflammation | Disease activity and kyphosis | Qin et al. (2016) |
| Serum  | miR-155 | Disease activity | Prajzlerová et al. (2017) |
| Plasma | miR-146a-3p | Target inflammatory and bone remodeling | Diagnostic and new bone formation | Perez-Sanchez et al. (2018) |
| Plasma (Mexico) | IncRNA let-7 | MMP-1, CRP | Diagnostic | Reyes-Loyola et al. (2019) |
| Plasma | IncRNA MEG3 | IL-1β, IL-6, TNF-α | Disease activity | Li et al. (2020b) |
| Plasma (Egypt) | miR-125a | Structural damage | Diagnostic, prognostic and disease burden | Fotoh et al. (2020) |

IL, interleukin; MMP, matrix metalloprotein; CRP, C-reactive protein; TNF, tumor necrosis factor.

**Table 2**: Extracellular (serum, plasma) ncRNA expressions, their modes of action and clinical applications in patients with AS.
therapeutic basis for the effective treatment for patients with AS. In the OBs isolated from murine model of AS, Ma et al. (2019) found that miR-96 expressed at a high level while sclerostin (SOST) expressed at a low level. Actually, miR-96 was observed to target and negatively regulate SOST. Furthermore, the over-expressed miR-96 activated the Wnt signaling pathway and increased proinflammatory cytokines (IL-6, TNF-α, and IL-10), ALP activity, calcium nodule formation and OB viability. These results indicated that the overexpression of miR-96 can enhance OB differentiation and subsequent bone formation in AS mice via Wnt signaling pathway. In contrast (Liu W et al., 2019), showed that mesenchymal stem cells (MSCs) derived from AS patients exhibited a strong capacity to inhibit osteoclastogenesis and secreted more CXCL5. Further studies showed that down-regulation of miR-4284 in AS-MSCs resulted in increased CXCL5, indicating that osteoclastogenesis may be markedly suppressed via miR-4284/CXCL5 axis. By experiments with human fibroblast-like synovial cells (HFLSs) isolated from AS tissues, Du et al. (2019) showed that miR-495 and dishevelled 2 (DVL-2) molecule were negatively correlated with each other in AS. Both molecules can inhibit inflammation by down-regulating proinflammatory cytokines, IL-1, IL-6 and TNF-α and facilitate bone differentiation by up-regulating osteoprotegerin (OPG) and RANKL levels in HFLS. Besides, miR-495 and siRNA, si-DVL-2, enhanced expression of wnt3α, RUNX2 and β-catenin and reduced β-catenin phosphorylation. Collectively, miR-495 depresses inflammatory responses and promotes bone differentiation of HFLSs via Wnt/β-catenin/RUNX-2 pathway by targeting DVL-2.

In addition to miRs and lncRNAs, circular RNAs (circRNAs) are a particular class of endogenous ncRNAs with a covalently closed circular structure (Hsu and Coca-Prados, 1979; Hansen et al., 2013; Memczak et al., 2013). Different from the linear RNAs, circRNAs lack free 3′-end poly A tail and 5′-end cap which prevent them from being degraded by nucleic acid endonuclease (Qu et al., 2015). Accordingly, the closed circular structure of circRNAs makes them extremely stable in the extracellular milieu and able to regulate the expression of target miRs by their sponge effects in human diseases (SanterBar and Thum, 2019; Wang et al., 2019). Kou et al. (2020) analyzed the circRNA expression profile of the spinal ligament tissue in AS patients. The authors found 57 circRNAs were up-regulated and 66 were down-regulated which were mainly involved in the regulation of biological processes of peptidyl-serine phosphorylation and immune system relevant to AS pathogenesis. In addition, the circRNA-miR interactions may provide new clues for understanding the mechanisms, diagnosis and potential molecular targets for the treatment in AS patients.

Another interesting findings by Tabrizi et al. (2017) were from the investigations on expression levels of the maturing microprocessor complex of miR in PBMCs from AS patients. It is believed that major enzymes responsible for miR maturation are Dicer, Drosha, and Drosha assistant DGCR8. Their results revealed that both Dicer and DGCR8 mRNA expression were

### Table 3: Intracellular expressions of ncRNAs in ligament-derived fibroblast, osteoblasts, bone marrow-derived mesenchymal stem cells (MSCs) and human fibroblast-like synovial cells (HFLSs) from AS patients.

| Source               | ncRNA     | Mode of action                                      | References                        |
|----------------------|-----------|-----------------------------------------------------|-----------------------------------|
| Fibroblast           | miR-17-3p | Osteogenic differentiation†                         | Zhang et al. (2017)               |
| Fibroblast           | miR-27b-3p| β-catenin†                                          | Tang et al. (2018)                |
| Fibroblast           | miR-124   | Wnt signaling†                                      |                                   |
| Fibroblast           | miR-204-5p| IL-6†                                               |                                   |
| Fibroblast           | miR-96    | TNF-α†                                              | Ma et al. (2019)                  |
| MSC                  | miR-4284  | Osteoclastogenesis†                                 | Liu W et al. (2019)               |
| HFLS                 | miR-495   | Inflammation†                                       | Du et al. (2019)                  |
|                     | DVL-2     |                                                     |                                   |

RUNX, Runt related transcriptional factor; TNF, tumor necrosis factor; IL, interleukin; ALP, alkaline phosphotase; Wnt, wingless and Int-1; SOST, sclerostin gene; CXCL, ligand for cysteine-X-cysteine chemokine; RANKL, Receptor activator of nuclear factor kappa-B ligand; GSK, glycogen synthase kinase. DVL, segment polarity protein dishevelled homolog.
down-regulated whereas Drosha mRNA expression was not influenced in AS. These data suggest that the down-regulated miR maturation components may probably contribute to the pathogenesis of AS. The intracellular expression of ncRNAs in ligament-derived fibroblasts of AS patients and their modes of action are summarized in Table 3.

CONCLUSION AND PERSPECTIVES

Genetic and environmental factors intriguingly interact with each other in affecting epigenetic modifications in patients with AS. More than 100 genes have been identified to contribute to AS susceptibility. Among them, HLA-B27 subtypes, polymorphic ERAP, and IL-23R mutation seem to be significantly associated. It is also conceivable that the microtrauma in the entheses may trigger the onset of AS. Besides, the arthritogenic peptides (misfolded HLA-B27 antigen and HLA-B27 homodimer), gut dysbiosis, abnormal intestinal metabolomic products, and immune plasticity can also induce soft tissue, articular and extra-articular inflammation. The inflammation-induced osteoporosis, the subsequent osteogenesis, and finally the new bone formation may cause skeletal disability. Extra-musculoskeletal manifestations of AS include mainly anterior uveitis, aortitis and interstitial fibrosis of the upper lungs. The florid immune dysfunctions of innate and adaptive immune responses resulted from the abnormal IL-23/IL-17 axis is paramount crucial. Nevertheless, the aberrant presentations of upstream epigenetic regulatory mechanisms are the culprits of similar importance for the abnormal immune regulation in AS. That is to say, these abnormalities of ncRNAs (i.e., miRs, lncRNAs and circRNAs) and microRNA maturing microprocessor complex (Dicer and Drosha) in PBMCs, serum/plasma and tissues play a crucial role in disrupting innate and adaptive immune responses implicated in multiple pathological processes in patients with AS. Although, the molecular mechanisms for these characteristic pathogenic events including enthesisitis, osteoporosis, osteogenesis, anterior uveitis, aortitis and interstitial lung fibrosis have been discussed in detail above, the real causes of AS pathogenesis remain elusive. Some perspective investigations helpful for the understanding of this complicated epigenetic regulation are suggested:

1) A molecular basis for the induction of gut dysbiosis by HLA-B27 subtypes with its subsequent abnormal epigenetic regulation and immune dysregulation should be clarified.

2) Clinically applicable serum ncRNAs as biomarkers for measuring disease activity and assessing therapeutic response in AS patients should be identified on the level of high sensitivity and specificity, compared to the nonspecific serum CRP and stool calprotectin currently available.

3) The role of aberrant ncRNA expression with subsequent abnormal immune responses in stressful enthesis induced by denatured hyaluronan-I needs to be explored.

4) Molecular and cellular bases for the absence of rheumatoid factors in AS patients that may be relevant to deranged ncRNA expression need to be unveiled.

5) The production of anti-CD74 autoantibody with its immunopathological roles in AS patients that may be relevant to aberrant ncRNA expression should also be delineated.

AUTHOR CONTRIBUTIONS

C-LY and C-YT supervised the writing of the manuscript. C-YT and H-TL prepared the manuscript and wrote the draft together. T-HL and S-CH prepared the figure drafts. C-YS, C-HW, H-TL, C-HL, C-CL, Y-MK, Y-SS, K-JL, and C-TC actively participated in the discussions and suggestions for the manuscript. All authors have read and agreed the final version of the manuscript.

FUNDING

This work is supported partially by grants from Taipei Veterans General Hospital (V108-C-203) and the Ministry of Science and Technology, Executive Yuan (MOST-107-2314-B-075-051-MY3), Taiwan.

ACKNOWLEDGMENTS

The authors thank all of the individuals who participated in this investigation.

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