Role of the intracellular receptor domain of gp130 (exon 17) in human inflammatory bowel disease

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INTRODUCTION

The gp130 cytokine family includes interleukin (IL)-6, IL-11, leukemia inhibitory factor, oncostatin M, ciliary neurotrophic factor, novel-neurotrophin-1-B-cell stimulating factor-3, neuroepoetin and cardiotrophin (CT)-1. Signal transduction of gp130 cytokines involves activation of the Janus kinases and signal transducer and activator of transcription (Jak-STAT) signaling cascade pathway as well as the src-homology tyrosine phosphatase 2 (SHP2)-Ras-Erk pathway. Briefly, ligand binding to the gp130 receptor complex induces autophosphorylation of Jak, followed by the phosphorylation of specific tyrosine residues in the cytoplasmatic receptor domain and subsequent tyrosine phosphorylation of STAT proteins. Phosphorylated STAT proteins (pSTATs) translocate as homo- or heterodimeric complexes to the nuclei and act as specific transcription factors by binding to regulatory promoter elements of various genes.

The intracellular domain of the common gp130 receptor subunit harbors several essential motifs involved in signal transduction of gp130 cytokines (Figure 1). The membrane proximal homology region of the intracellular gp130 domain containing the box1 and box2 motifs is important for Jak association. The tyrosine residue Y757/759 (murine/human) in the cytoplasmic domain of activated gp130 is essential for the association of gp130 with src-homology 2 domain-bearing protein tyrosine phosphatase (SHP)-2 and suppressor of cytokine signaling (SOCS)-3. SHP-2 down-regulates STAT signaling and links the Jak-STAT cascade to the ras-dependent mitogen-activated protein kinase pathway. SOCS-3 inhibits the Jak-STAT signaling cascade by directly inhibiting Jak kinase activity as well as by binding to activated intracellular cytokine receptor domains and inhibiting further protein association.

Finally, the tyrosines Y765/767 (murine/human), Y812/814 (murine/human), Y904/905 (murine/human), and Y914/915 (murine/human) in the cytoplasmic domain of activated gp130 are required for the association of STAT proteins with gp130. This association is essential for STAT phosphorylation and activation.

Gp130<sup>−/−</sup> mice harboring a mutated (Y765F and Q768A) and truncated (769stop) intracellular gp130 receptor domain, thus lacking the four specific STAT-binding site motifs YxxQ at tyrosines Y765/812/904/914 in murine gp130, display abrogated STAT3 phosphorylation but elevated activation of SHP2 and Erk1/2 following IL-6 stimulation. Gp130<sup>−/−</sup> mice show impaired colonic mucosal wound healing, mucosal ulcers in the gastric pylorus and coecum, and exaggerated sodium dextran sulfate (DSS)-induced colitis.
Crohn's disease and ulcerative colitis are classified as chronic inflammatory bowel diseases (IBDs) characterized by chronic inflammation and the inability to maintain mucosal integrity in the large and/or small intestine. The human gp130 gene is located on chromosome 5q11[4]. Until now, linkage studies have not reported the g p130-corresponding loci on chromosome 5q11 to be susceptible loci for IBD. Nevertheless, data obtained from Gp130 mice strongly suggest that mutations in the intracellular binding sites of gp130 might play a role in the pathophysiology of IBD[5-7]. Therefore, we performed a candidate gene study, searching for mutations in exon 17 of the gp130 gene in 146 patients with IBD.

### MATERIALS AND METHODS

#### Patients

A total of 146 patients with Crohn's disease ($n = 73$), ulcerative colitis ($n = 63$) or indeterminate colitis ($n = 10$) were recruited through the IBD in-patient and out-patient clinics at our university hospital in Munich. Patient data were obtained by chart review and patient questionnaire and recorded by a senior gastroenterologist. Inclusion criteria were based on homogenous diagnostic parameters including clinical data (physical examination, CDAI[9]/Lichtiger score[10]), laboratory findings, endoscopic criteria (location and degree of inflammation, stenosis, fistulas, transmural involvement in colonoscopy) and radiological findings (CT and/or MRT). At inclusion, all patients had duration of IBD of more than one year. An infectious pathogenesis of gastrointestinal symptoms was excluded in all patients by biopsy and stool culture. EDTA blood samples for molecular genetic studies were taken, after written informed consents were obtained from the patients and/or their parents. The study protocol followed ethical guidelines and was approved by the Institutional Ethical Committee.

#### DNA isolation and sequencing

Genomic DNA was isolated from the peripheral blood leukocytes with the DNA blood mini kit (QIAGEN, Hilden, Germany). Exon 17 of the gp130 gene (GeneBank Accession M57230; Ref.[4]) was amplified by PCR using the intron-specific primer pair 5' AGTTTCAGAGATGCATTAGCTCGTG-3' (sense) and 5'-GGCAGATGTCATCCT CAGAGATG-3' (antisense; nt 68781 to nt 69703 in GeneBank Accession AC016596) in order to avoid amplification of an intronless pseudogene of gp130[4]. Each 50-µL reaction contained approximately 300 ng of genomic DNA, 0.4 µmol/L of each primer, 1.5 mmol/L MgCl$_2$, 5 µL Thermo Start reaction buffer (ABgene, Epsom, UK), 125 µmol/L dNTPs, and 0.25 µL of Thermo Start Taq DNA polymerase (ABgene). PCR was performed for 40 amplification cycles (denaturation at 95 °C for 30 s, annealing at 62 °C for 30 s and extension at 72 °C for 30 s). PCR products were purified using the QIAquick PCR purification kit (QIAGEN, Hilden, Germany) and sequenced with the Big Dye terminator v3.1 ready reaction cycle sequencing kit (Applied Biosystems, Foster City, USA). Data analysis was performed on an ABI Prism 377 DNA sequencer (Applied Biosystems).

### RESULTS

#### Patient characteristics

Seventy-three patients with Crohn's disease (39 males, 34 females), 63 patients with ulcerative colitis (24 males, 39 females) and 10 patients with indeterminate IBD (4 males, 6 females) were examined. The median age at inclusion was 39±10 years in the CD group, 41±15 years in the UC group and 47±19 years in the indeterminate group (mean±SD). All patients included in this study had a history of at least one episode with an elevated CDAI[9] of more than 150 points in Crohn's disease or a Lichtiger[10] score of more than 10 points in ulcerative colitis in combination with characteristic endoscopic and radiological findings.

#### gp130 exon 17 sequencing

The genomic DNA of the 146 patients with IBD, who were classified as Crohn's disease ($n = 73$), ulcerative colitis ($n = 63$) and indeterminate IBD ($n = 10$), was sequenced for mutations in exon 17 of the gp130 gene. None of these 146 patients carried a mutation or a polymorphism in this region of the gp130 receptor gene. Therefore, we could exclude mutations in exon 17 of the gp130 receptor gene being involved in the etiology of Crohn's disease in more than 5% of patients ($P<0.05$). Likewise, we did not find any evidence that mutations in exon 17 of the gp130 receptor gene might be involved in the etiology of ulcerative colitis.

### DISCUSSION

The current study demonstrated a lack of mutations in exon 17 of the human gp130 gene in 146 patients with IBD, e.g., Crohn's disease and ulcerative colitis. Exon 17 of the gp130 gene[8] is spanning most of the intracellular region of the gp130 receptor (Figure 1) and harbors several important signal transduction regions such as the membrane proximal box2 motif for Jak association, the tyrosine residue Y$_{757}$/Y$_{759}$ (murine/human), Y$_{765}$/Y$_{767}$ (murine/human), Y$_{812}$/Y$_{814}$ (murine/human), Y$_{771}$ for SHP2 and SOCS-3 association, and the tyrosine residues Y$_{761}$ for STAT activation, SHP2 activation, and JAK activation.

**Figure 1** Schematic diagram of the intracellular receptor domain of human gp130 (adapted from Refs.[8-10]).
Y9/4/95 (murine/human), and Y9/4/915 (murine/human) for STAT association[6,8].

The animal model of gp130STAT mice demonstrated impaired STAT activation due to a truncated intracytoplasmic gp130 receptor domain lacking the tyrosine residues Y757F/V760A and V760A YxxN motif for SHP-2 and SOCS-3 binding displayed enhanced tissue STAT3 phosphorylation, but abrogated SHP2-Ras-ERK signaling following IL-6 stimulation. Gp130 (757F) mice developed gastric adenomas by 3 mo of age[6,9]. These two gastrointestinal phenotypes are highly similar to the phenotypes exhibited by mice deficient in trefoil factor 1 (pS2/TFF1) and in intestinal trefoil factor (ITF)/TFF3, respectively. TFF1 and TFF3 are members of the trefoil factor (TFF) family; each of them bears a characteristic three-looped structure known as the trefoil domain. TFF1 is primarily expressed in gastric-pit mucus cells. TFF3 is expressed at its highest level in the mucus-secreting goblet cells of the small and large intestine. TFFs are supposed to be involved in the process of healing or restitution, since they are upregulated after injury to the gastrointestinal epithelium. TFF1 knock-out mice could develop gastric hyperplasia, whereas deletion of TFF3 might become visible in an exaggerated response to DSS-mediated epithelial damage. Regulation of TFF1 and TFF3 in the gastrointestinal tract seems to be dependent on SHP2/Erk and STAT1/3 signaling, as reduced gastric levels of TFF1 and TFF3 are observed in gp130D−/− mice[5-7]. Taken together, it seems that the integrity of the gastrointestinal mucosa can be maintained by a balanced activation of both pathways (leading to a certain ratio of TFF1 vs TFF3). According to this model, the imbalance in favor of STAT1/3 signaling leads to excessive antral proliferation, presumably caused by unhindered effects of STAT signaling. An exaggerated activation of the SHP2/Erk signaling cascade results in mucosal healing defects due to a lack of phosphorylated STAT3. Either the decreased level of phosphorylated STAT3 itself or down-regulation of TFF3 leads to impaired epithelial cell migration after the injury of the intestinal epithelium. However, our current study has demonstrated that impaired STAT activation due to a mutated or truncated intracellular gp130 receptor domain is not involved in the pathogenesis of IBD in humans. Nevertheless, mutations at this site might be involved in the pathogenesis of other human diseases, e.g., degenerative joint disease or blastocyst implantation failure[6,8,12,14].

In summary, dysbalance of the Jak-STAT pathway seems to be involved in the pathogenesis of colitis[6,8,12,14]. In the current study, however, we found no evidence that mutations in exon 17 of the gp130 gene, encoding all functionally essential intracellular tyrosine residues of the gp130 receptor (Figure 1), are involved in the pathogenesis of human IBD. Our data suggest that other mechanisms are important for the dysbalance of the Jak-STAT pathway in IBD. The role of STAT protein activation and negative regulation of Jak-STAT signaling merits further investigation. Studies on the putative role of SOCS-3 mutations in the pathogenesis of IBD are currently underway.

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