ABSTRACT

Background Both pathogenic bacteria and viruses are frequently detected in the nasopharynx (NP) of children in the absence of acute respiratory infection (ARI) symptoms. The aim of this study was to estimate the aetiological fractions for ARI hospitalisation in children for respiratory syncytial virus (RSV) and influenza virus and to determine whether detection of specific respiratory pathogens on NP samples was associated with ARI hospitalisation.

Methods 249 children up to 5 years of age hospitalised for ARI (following a symptom-based case definition) and 306 hospital controls were prospectively enrolled in 16 centres across seven European Union countries between 2016 and 2019. Admission day NP swabs were analysed by multiplex PCR for 25 targets.

Results RSV was the leading single cause of ARI hospitalisations, with an overall population attributable fraction (PAF) of 33.4% and high seasonality as well as preponderance in younger children. Detection of RSV on NP swabs was strongly associated with ARI hospitalisation (OR adjusted for age and season: 20.6, 95% CI: 9.4 to 45.3). Detection of three other viral pathogens showed strong associations with ARI hospitalisation: influenza viruses had an adjusted OR of 6.1 (95% CI: 2.5 to 14.9), parainfluenza viruses (PIVs) an adjusted OR of 4.6 (95% CI: 2.5 to 8.7), and metapneumovirus, on NP swabs can establish aetiology with high probability. PAFs for RSV and influenza virus are highly seasonal and age dependent.

Conclusion RSV is the predominant cause of ARI hospitalisations in young children in Europe and its detection, as well as detection of influenza virus, PIV or metapneumovirus, on NP swabs can establish aetiology with high probability. PAFs for RSV and influenza virus are highly seasonal and age dependent.

INTRODUCTION

The Global Burden of Disease Study 2015 estimates that acute respiratory infections (ARI) cause more than 15% of under-five mortality. Approximately 102 million cases of...
pneumonia resulting in 0.7 million deaths occurred each year in children under 5 years of age.4 While compared with low-resource settings case fatality in paediatric ARI in Europe is much lower,3 ARI remains the most frequent reason for hospitalisation in children.4 In children under the age of 5, respiratory viruses are detected in up to 80% of ARI cases.5 6 While Streptococcus pneumoniae remains the most prevalent bacterial pathogen,7 8 the proportion of bacterial ARI and prevalence of bacterial colonisation have declined with widespread use of conjugate vaccines.4 9–11 However, the frequency of treatment of ARI with antibiotics has not declined accordingly.12

Both in paediatric clinical care and research, nasopharyngeal (NP) samples are the only microbiological specimen that can routinely be obtained.13 Children’s upper airways are regularly colonised by potentially pathogenic bacteria or they are asymptomatic carriers of respiratory viruses.14 In a study on community-acquired pneumonia (CAP) among children in the USA, bacterial pathogens were identified in a majority of cases, with viruses, particularly respiratory syncytial virus (RSV) in younger and human rhinovirus (HRV) in older children, being identified in the majority.15 Codetection of viruses and bacteria is common.16–18 Codetection has been associated with more severe disease in some,17 18 yet not in other studies.19 20 Viral–bacterial codetection has mostly been shown to be associated with more severe disease, while codetection of several viruses was more often unrelated to severity.21 For S. pneumoniae, prevalence of upper airway colonisation peaks at around 3 years of age.22

Recent studies in Africa, Asia and North America, conducted in an era of widespread routine vaccination against encapsulated bacteria, show that the bulk of disease is caused by viral infections (predominantly RSV and HRV).15 23 The Pneumonia Etiology Research for Child Health study compared more than 4000 children admitted for CAP with more than 5000 community controls in seven countries in sub-Saharan Africa and South-East Asia and in contrast to earlier findings, showed that RSV had by far the highest aetiological fraction (31%) of all studied pathogens.23 Surprisingly, comparable data are not available for Europe. Both RSV and influenza viruses show distinct seasonal patterns in Europe.24 According to the European Centre for Disease Prevention and Control, pneumococcal conjugate vaccines (PCVs) have been introduced into routine infant vaccination schedules in the majority of European countries. However, influenza vaccines are only part of vaccination plans for children with increased risk, for example, due to comorbidities. Pronounced seasonality of ARI in Europe and potentially differing and changing epidemiology warranted a multicentre European study including cases hospitalised with ARI and healthy controls.

PREPARE (Platform for European Preparedness Against (Re-)emerging Epidemics) is a European Commission-funded network for harmonised large-scale clinical research studies on infectious diseases, prepared to rapidly respond to any severe infectious disease outbreak. The Multi-centre EuRopean study of MAjor Infectious Disease Syndromes (MERMAIDS) is the part of the PREPARE platform including case-control and cohort studies on aetiology and management of ARI.

The aim of this study was to estimate the aetiological fractions for ARI hospitalisation in children for RSV and influenza virus, both pathogens with a strongly seasonal occurrence in Europe, and to determine whether detection of specific respiratory pathogens in upper airway samples was associated with ARI hospitalisation. In line with the general mission of PREPARE, a secondary study aim was to build a European network of paediatric research sites trained and equipped for studies on ARIs in emergency care.

METHODS

Study design

The study was a case–control study with collection of clinical data and samples at baseline and follow-up of cases until discharge. Primary objective was to estimate the proportion of cases attributable to specific respiratory pathogens. Cases and controls were enrolled continuously throughout the year between September 2016 and March 2019 at 16 secondary or tertiary hospitals in seven European countries (Belgium, Germany, Greece, Italy, Lithuania, Spain and the UK). PCV was part of routine vaccination schedules within the first 6 months of life in all of these countries and influenza vaccines were not recommended as routine vaccines in any. Participants were recruited as they presented to paediatric departments, where they were screened for eligibility, mainly during daytime hours with high level of staffing. Total numbers of presenting eligible patients and number of screened patients were not systematically collected. Parents of all participants gave written informed consent and the study was approved by the responsible ethics committees at all sites. The study was designed to be as minimally invasive as possible and only pathogen detection samples were taken from all children. This protocol was written without patient involvement. Patients or guardians were not invited to comment on the study design or to contribute to the writing or editing of this document for readability or accuracy.

The protocol is available on the PREPARE website: https://prepare.ersnet.org/trials-protocols.aspx.

Case and control groups

Participants were otherwise healthy children under 6 years of age. Patients with temperature ≥38°C hospitalised due to a new episode of ARI according to a clinical definition were included in the study as cases. Controls were afebrile children attending the same hospitals for scheduled procedures or visits not related to infections and who did not fulfil clinical criteria for the case group but may have had mild symptoms of ARI. Hospital controls rather than community controls were chosen.
for feasibility. To be included for the case–control study, an NP sample (as specified in the section on pathogen detection) had to be obtained. Eligibility criteria are listed in Table 1. Matching of controls to cases by age, season and site of recruitment was attempted but not consistently successful.

Pathogen detection
NP swabs for pathogen detection were collected for cases and controls at admission (within 24 hours), or for some controls during an outpatient clinic visit. Samples were frozen at −80°C until shipment to the central study laboratory at the University of Antwerp, where specimens were stored at −80°C until analysis. NP samples were extracted with the NucliSens EasyMag (bioMérieux, France) by using the specific A protocol. The FTD Respiratory Pathogens 21 plus (Fast Track Diagnostics, Ltd, Luxembourg) was applied according to the instructions of the manufacturer for the qualitative detection of influenza A, influenza B, influenza A-H1N1, human coronaviruses NL63, 229E, OC43 and HKU1, parainfluenza viruses (PIVs) 1, 2, 3 and 4, human metapneumovirus A and B, HRV, RSV A and B, adenovirus, enterovirus, parechovirus, bocavirus, Mycoplasma pneumoniae, Chlamydia pneumoniae, S. pneumoniae, Haemophilus influenzae b and Staphylococcus aureus.

Table 1  Complete inclusion and exclusion criteria

| Case group                              | Control group                               |
|-----------------------------------------|---------------------------------------------|
| **Inclusion criteria**                  |                                             |
| Clinical suspicion of a new episode of ARI within the last 7 days | One of the following 1. Attending for an elective or semielective procedure requiring general anaesthesia or moderate-deep sedation (including, eg, surgery, radiological examinations etc). 2. Well and otherwise healthy children attending an outpatient clinic for a non-emergency clinical assessment. |
| The attending physician has decided that the child requires hospitalisation |                                             |
| Primary reason for hospital admission was clinical suspicion of a new episode of ARI |                                             |
| Temperature ≥38°C measured by any method (reported within 24 hours or at presentation) | Afebrile on the day of enrolment |
| And at least two of the below (with at least one of 1 or 2): | No evidence of severe infection as judged by attending physician |
| 1. Signs of lower respiratory tract infection: cough, abnormal sounds on chest auscultation (crackles, reduced breath sounds, bronchial breathing, wheezing), dyspnoea (chest indrawing, nasal flaring, grunting). |  |
| 2. Signs of upper respiratory tract infection: coryza, nasal congestion, sore throat, pharyngitis, myringitis, acute otitis media. |  |
| 3. Signs of respiratory dysfunction: tachypnoea for age or brady/apnoea or decreased oxygen saturation (<92% in room air). |  |
| 4. Signs of reduced general state: poor feeding, vomiting, lethargy/drowsiness. |  |

**Exclusion criteria**

Inpatient care for 24 hours or more for any condition within the previous 30 days, except for routine postnatal care

Immunocompromised infant (stem cell transplant, solid organ transplant, HIV, AIDS, immunosuppressive therapy, primary immunodeficiency, haemodialysis)

Presence of complex chronic comorbidities

Body weight <3 kg on day of assessment and/or corrected gestational age <37 weeks

Aetiology other than infection (such as trauma, autoimmune disorder, malignancy) is suspected to be the primary cause of the current illness episode

Any signs and symptoms suggesting a clear primary focus of infection, such as urinary tract infection, open wounds, indwelling catheters, reactivation of previously diagnosed infectious or inflammatory condition

Dehydration due to previous illness episode such as diarrhoea and vomiting

ARI, acute respiratory infection.
Data management and statistics

Demographic data, focused medical history, clinical characteristics on admission, focused aspects of patient management and final outcome were entered into electronic case report forms hosted by the Julius Center at the University of Utrecht. Data were checked for completeness at the end of the study and data queries completed within 6 months of inclusion of the last participant.

The sample size was determined before the study started based on estimates for the proportions of children who were positive for RSV and influenza virus. With an expected positive proportion for influenza of 10% in cases and 3% in controls and aiming for 90% power and alpha=5% for an OR larger than 1.0, the required sample size for influenza was 320 per group. The required sample size for RSV was 40 per group (expected 25% positive in cases and 1% in controls). As these sample sizes were not additional, the total required sample size followed the calculation for influenza and was therefore 640 (320 cases and 320 controls).

Data handling and statistical operations were performed in Stata V.14. Complete records analysis was done throughout, as missing data were rare (below 5% on individual variables, none on pathogen detection and no dropped observations on covariate analyses). For categorical variables, p values were obtained by χ² test and for continuous variables by Wilcoxon rank-sum test due to the non-normal distribution of their values. Necessitated by incomplete matching, adjusted ORs were calculated by logistic regression with age and season of recruitment as covariates, p values were obtained by Wald test. For adjusted ORs, sensitivity analyses were performed to explore if these differed based on (1) coadjusting for country of inclusion, because proportions of cases and controls among the participants differed by country and (2) using a control group restricted to completely asymptomatic children. The population attributable fraction (PAF) was calculated by substituting the OR for the risk ratio where detection of the pathogen in the control group was sufficiently rare to allow for this substitution.

RESULTS

A total of 349 cases and 306 controls were enrolled from 16 sites in seven countries during the study period, with approximately three-quarters of cases and two-thirds of controls enrolled in the main European ARI season between October and March. Sixty-one per cent of participants were male, with cases slightly younger than controls. The proportions of cases and controls included in the participating countries differed (table 2).

Clinical course

In cases, the median time to presentation in the emergency department, from first day of onset of symptoms was 2 days (IQR: 1–4). The most common symptom on presentation was poor feeding, seen in 241 (69.3%), 190 (54.4%) had chest recessions, 137 (39.4%) wheezing and 109 (31.2%) had a fever of >39°C. A relevant proportion of cases had signs of more severe disease: 26 (7.5%) required supplemental oxygen on admission and 12 (3.5%) had a central capillary refill time of >2 s. Of the controls, 33 (10.8%) had mild illness as reported by parents during the previous 7 days but did not fulfil inclusion criteria as a case.

The majority of cases had disease clinically classified as lower respiratory tract infection (LRTI). About 39.3% of cases were treated with antibiotics during their admission. Among cases diagnosed as LRTI, 68.6% received antibiotics, compared with 34.8% among children with upper respiratory tract infection (URTI) and 35.7% with unspecified infection.

Detected potential pathogens in cases and controls

Figure 1A shows prevalence of detection of respiratory pathogens in study samples. S. pneumoniae was the most prevalent target detected in cases and controls. The second most frequent potentially pathogenic bacteria found was S. aureus, which was the only one weakly associated with being a control rather than a case (OR for case status: 0.7, 95% CI: 0.5 to 1).

Respiratory viruses were detected in >80% of cases and >40% of controls. The respiratory virus most frequently detected was HRV, which was found in >20% of both cases and controls. Eighty per cent (32 of 40) of the detected influenza viruses were influenza A, among these 22 (68.8%) were influenza A-H1N1. All influenza B were isolated from children in the case group, but due to the small number of isolates this may likely be random.

Human coronaviruses were detected at similar frequencies, between 5% and 10%, in both cases and controls and showed no predominance for any age (median age in years among coronavirus negative: 1.39, IQR: 0.49 to 3.20; among coronavirus positive: 1.34, IQR: 0.59 to 2.40; p=0.642). All four endemic coronaviruses were evenly distributed between cases and controls (229E in 7 cases and 5 controls, NL63 in 3 cases and 5 controls, HKU1 in 4 cases and 7 controls and OC43 in 8 cases and 10 controls).

Codetection of potentially pathogenic bacteria and viruses was common in both groups (51.9% in cases and 28.8% in controls). Among the 187 cases in which both bacteria and viruses were found, RSV was found in 74 and influenza virus in 16. S. pneumoniae was the bacterial potential pathogen found in most codetection cases (83.4%) and S. aureus was detected in 32.1%. In contrast, among controls with detection of both viruses and bacteria simultaneously, only very few carried RSV or influenza virus (7 and 3, respectively) but 43 (48.9%) carried rhinovirus and proportions positive for S. pneumoniae (87.5%) and S. aureus (38.6%) were similar as in cases.

Other respiratory pathogens were detected in only very low proportions of participants: M. pneumoniae in three
cases (0.9%) and no controls, *C. pneumoniae* was not detected in any cases but in three controls (1.0%) and *H. influenzae b* was not detected in either group.

No potential respiratory pathogen was detected in only 27.1% of controls and 4.3% of cases. Figure 1B illustrates clear evidence of a higher frequency of detection of RSV (OR: 23.0, 95% CI: 10.5 to 59.2), influenza virus (OR: 5.4, 95% CI: 2.2 to 15.9), PIV (OR: 4.5, 95% CI: 1.8 to 13.5) and metapneumovirus (OR: 4.2, 95% CI: 1.2 to 23.1) in cases compared with controls.

Proportions of patients with detection of no pathogen, only bacteria, only viruses or both viruses and bacteria did not change depending on prior duration of symptoms in days (p=0.502).

There was no difference in the frequency of antibiotic prescriptions when comparing different groups of detected pathogens. Antibiotics were given in 62.5% of children with no pathogen detected, 65.7% with only bacteria detected, 63.6% with only viruses detected and 52.9% with viruses and bacteria detected (p=0.158).

There was no difference in prevalence of antibiotics in children who tested positive for RSV (61.3% received antibiotics, compared with 56.5% in RSV negative, p=0.637) or positive for influenza virus (56.9% received antibiotics, compared with 55.8% in influenza negative, p=0.479).

**Age and season dependency of pathogen detection**

Table 3 presents the breakdown of detection of RSV, influenza, parainfluenza, metapneumovirus and *S. pneumoniae*. As expected, RSV was more commonly found in younger children and almost 90% of detection of influenza virus occurred between January and March. PIV was more often detected in children of the two younger age groups and in autumn or summer as compared with
Detection of metapneumovirus was evenly distributed among age groups and seasons. Additionally, online supplemental table 1 provides numbers and proportions of participants with detection of all pathogens presented in figure 1 by case or control status. Parechovirus was more often detected in cases in autumn (6.6%, compared with 1.1% in winter and 0% in summer, p=0.011) and enterovirus more often in summer (8.8%, compared with 1.3% in autumn and 1.7% in winter, p=0.011). Other pathogens showed no distinct seasonality. Because both age and season of recruitment were distributed differently between cases and controls, we calculated OR estimates adjusted for these covariates. As shown in table 4, the adjusted ORs did not differ markedly from the crude ORs presented in figure 1B.

Detection of RSV was very strongly associated with being a case (OR: 20.6). To a lesser degree, this is also true for influenza virus (OR: 6.1), PIV (OR: 4.6) and metapneumovirus (OR: 4.5). For S. pneumoniae, the association of detection and case status was weaker (OR: 1.7). The sensitivity analysis including country of inclusion...
as a covariate indicated that, after multivariate adjustment, country of inclusion had no effect on the association between pathogens and case status (table 5). When restricting the analysis to controls without any symptoms of respiratory infection, ORs for *S. pneumoniae* and influenza virus remained similar, but the association between detection of RSV or metapneumovirus and case status became even more pronounced (RSV—OR: 42.6, metapneumovirus—OR: 6.2).

### Age-specific and season-specific PAF estimates

As *S. pneumoniae* (and some respiratory viruses including HRV) were commonly detected in controls, PAF was not estimated. For RSV, the overall PAF was 33.4% and for influenza virus 7.9%. Due to the specific distribution characteristics of these viruses there were, however, marked differences between the PAF estimates for different age groups and at different times of the year (table 6). For PIV, the overall PAF was 6.5% and for metapneumovirus 3.0%. Estimation of age-group and season-specific PAF was unfeasible due to low overall detection numbers.

Between the months of October and March, RSV caused close to 50% of ARI hospitalisation in children under the age of 1 and a third of ARI hospitalisation in children aged 2–3 years in this study. These fractions did not differ between the autumn months (October–December) and winter months (January–March). In contrast, the PAF for influenza virus was highest in winter and in children over the age of 3 years, where influenza virus caused a third of ARI hospitalisations.

### DISCUSSION

The aetiology of severe ARI requiring hospitalisation in children from this European case-control study is consistent with previously reported results from other settings.15 23 These findings show that RSV was the dominant respiratory pathogen in preschool children during the study period, causing a third of ARI

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**Table 4** OR of detection in cases compared with controls by multiplex PCR in nasopharyngeal swabs, by pathogen, adjusted for age and season (logistic regression)

| Pathogen                        | Adjusted OR | 95% CI         | P value |
|---------------------------------|-------------|----------------|---------|
| *Streptococcus pneumoniae*      | 1.7         | 1.2 to 2.3     | 0.002   |
| RSV                             | 20.6        | 9.4 to 45.3    | <0.001  |
| Influenza virus                 | 6.1         | 2.5 to 14.9    | <0.001  |
| Parainfluenza virus             | 4.6         | 1.8 to 11.3    | 0.001   |
| Metapneumovirus                 | 4.5         | 1.3 to 16.1    | 0.021   |
| No pathogen detected            | 0.1         | <0.1 to 0.2    | <0.001  |

P values were obtained by Wald test. RSV, respiratory syncytial virus.

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**Table 5** Sensitivity analyses for OR of detection in cases compared with controls, by pathogen, adjusted for age and season (logistic regression)

| Pathogen                        | Additionally adjusted for country of inclusion | Excluding mildly symptomatic controls |
|---------------------------------|-----------------------------------------------|-------------------------------------|
|                                 | Adjusted OR | 95% CI | P value | Adjusted OR | 95% CI | P value |
| *Streptococcus pneumoniae*      | 1.63         | 1.17 to 1.27 | 0.003 | 1.95         | 1.39 to 2.73 | <0.001 |
| RSV                             | 20.7         | 9.38 to 45.82 | <0.001 | 42.6         | 13.31 to 136.29 | <0.001 |
| Influenza virus                 | 6.38         | 2.56 to 15.88 | <0.001 | 6.38         | 2.42 to 16.81 | <0.001 |
| Parainfluenza virus             | 4.87         | 1.93 to 12.28 | 0.001 | 4.02         | 1.63 to 9.96  | 0.003  |
| Metapneumovirus                 | 4.04         | 1.10 to 14.79 | 0.035 | 6.20         | 1.37 to 28.02 | 0.018  |
| No pathogen detected            | 0.12         | 0.07 to 0.22  | <0.001 | 0.10         | 0.05 to 0.18  | <0.001 |

P values were obtained by Wald test. RSV, respiratory syncytial virus.

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**Table 6** PAF matrix for RSV and influenza

| Age group (PAF%) | RSV              | Influenza virus |
|------------------|------------------|-----------------|
| <12 Months       | 48.0             | *               |
| 12–<36 Months    | 38.1             | 8.0             |
| ≥36 Months       | 20.0             | 30.3            |

RSV, respiratory syncytial virus.

No estimates for *S. pneumoniae* are presented due to its high prevalence in the control group (see Methods for details) and for PIV and metapneumovirus due to low overall detection numbers.

*None detected in cases or controls.*

PAF, population attributable fraction; RSV, respiratory syncytial virus.
hospitalisations. Detection of RSV in healthy children was rare. In our study, half of ARI hospitalisations could be attributed to the four pathogens with the strongest association with ARI hospitalisation, that is, RSV, influenza virus, PIV and metapneumovirus.

Both RSV and influenza virus are highly seasonal, and their prevalence was associated with patient age. However, the strength of their association with being hospitalised for ARI did not differ between age groups and seasons. This suggests that their detection is highly predictive for aetiology of an ARI episode even in patients who are not part of the typical risk group (ie, infants for RSV) or who present outside the pathogen’s main season. Compared with data from studies set in sub-Saharan Africa and South-East Asia, influenza viruses were more commonly detected in children hospitalised for ARI in this study. While the PAF is therefore higher in Europe, the association of detected virus with clinical disease was similar across settings. PIV was more often detected in autumn and summer compared with the winter months when most patients with ARI are hospitalised. This suggests that it may have a high PAF in months with a lower frequency of cases. However, numbers of PIV detection were too low in this study to demonstrate this.

* S. pneumoniae was the most frequently detected respiratory pathogen in NP swabs from both children hospitalised for ARI and healthy controls but was not a strong predictor of hospitalisation for ARI. This is consistent with previous findings of an age-related dynamic with high nasopharyngeal colonisation rate by *S. pneumoniae* in infancy and early childhood. Therefore, detection of *S. pneumoniae* on upper airway samples is unable to reliably establish the aetiology of an ARI episode. *S. aureus* was more frequently found in control children. *S. aureus* may cause LRTIs and is especially common as a cause of superinfections following influenza. However, its detection in NP specimens was not indicative of ARI. An inverse relationship between carriage of *S. pneumoniae* and *S. aureus* may explain this finding due to *S. pneumoniae* being a more common cause of ARI. This has been suggested, but study results are contradictory.

The higher proportion of codetection of viruses and bacteria in cases as compared with controls was almost exclusively accounted for by higher proportions of children infected with RSV or influenza virus. Thus, the association between codetection of viruses and bacteria with being hospitalised for ARI in our study likely reflects the importance of viruses in causing severe respiratory symptoms in young children rather than suggesting true co-infections.

The low rate of detection of less-common causes of pneumonia, such as *M. pneumoniae* and *C. pneumoniae*, is likely due to the generally low prevalence of these pathogens in the studied age group. The absence of *H. influenzae* b is not surprising, as carriage and infections have virtually disappeared since the introduction of the vaccine. This study ended before the beginning of the current COVID-19 pandemic. The four previously endemic coronaviruses were found in similar and low proportions in both cases and controls and were not associated with case status. Therefore, although they may have caused ARI hospitalisation in some children, their detection on NP swabs was not sufficient to establish them as causative pathogens of ARI.

The study’s aim of capacity building proved to be crucial regarding the current pandemic. With the onset of the pandemic, the study network was instantly able to provide information on patient management strategies and to facilitate site participation in the WHO-initiated ISARIC Clinical Characterisation Protocol. Further, several study sites are participating in subsequently launched consortia funded by the European Commission.

A limitation of this study is that we only analysed upper airway samples. Previous multiplex PCR-based studies confirmed a high concordance between respiratory pathogens found in upper and lower airways, but especially with regard to specific pathogens discordance has been reported. By restricting the samples to NP samples, the study may still have missed a stronger association of lower airway carriage of bacteria with ARI hospitalisation. Nevertheless, restricting samples to NP swabs best reflects the situation in paediatric clinical care.

Another important limitation is that we did not systematically record numbers of screened patients and that both cases and controls were convenience samples. Because of differing thresholds for patient admission it is difficult to compare severity of disease in hospitalised children with ARI between high-resource and low-resource settings, but on average hospitalised children in high-resource settings have milder disease. Very few children in our sample required intensive care unit treatment on admission, and this is consistent with the low case fatality of childhood ARI in high-resource settings. Poor feeding was the most frequently observed clinical ARI sign. Poor feeding can be seen in the majority of children with RSV bronchiolitis and is often the cause for hospitalisation.

In terms of disease severity, the group of cases in our study does not seem to differ from children included in other studies on hospitalisations for ARI (including, but not exclusively, CAP) in comparable settings. Although overall case severity showed a good representation of the intended group, it is nonetheless possible that convenience sampling biased associations between detected pathogens and hospitalisation in either direction.

The study was conducted in a setting where in all places PCV was part of routine infant vaccine schedules and influenza vaccine was not. Therefore, the study cannot provide evidence on the effect of these vaccines on aetiology of childhood ARI hospitalisation and may not be generalisable to settings with differing vaccination schedules.
Hospital controls are less likely to have symptoms than children in the community, as those with symptoms may not attend hospital visits or may postpone elective procedures. Children were still included in the control group if they showed symptoms of mild disease as long as they did not meet case criteria as this has been shown to result in a control group most representative of the general population. Exclusion of controls with mild respiratory symptoms would lead to an overestimation of the association between pathogens and hospitalisation, especially for pathogens rarely found in asymptomatic children. Some overestimation of these associations may still have occurred due to the choice of hospital controls rather than community controls.

This study is the first to present findings on the aetiology of hospitalisations for ARI applying the same protocol across sites in seven European countries. The results illustrate that the causes of severe ARI requiring hospital admission do not differ profoundly between these European and other global settings. However, the pronounced seasonality of circulation of RSV and influenza virus as major causal agents leads to differing probabilities of infection with these pathogens in an individual patient, strongly depending on age and time of year.

Previous studies have shown that reduction of inappropriate antibiotic prescriptions in emergency departments can be achieved with antibiotic stewardship programmes. Our study provides evidence that detection of RSV and influenza virus on NP samples can strongly support a suspected viral aetiology of ARI and may therefore help to reduce antibiotic prescriptions.

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Authors 1Department of Paediatric Cardiology, Pulmonology and Intensive Care Medicine, University Children’s Hospital Tübingen, Tübingen, Germany 2Clinical Pathways and Epidemiology Unit, IRCCS Bambino Gesù Children’s Hospital, Rome, Italy 3Translational Paediatrics and Infectious Diseases, Hospital Clínico Universitario de Santiago, Servizo Galego de Saúde, Santiago de Compostela, Spain 4Genetics, Vaccines and Infectious Diseases Research Group, Instituto de Investigación Sanitaria de Santiago, Universidade de Santiago de Compostela, Santiago de Compostela, Spain 5Clinic of Children’s Diseases, Institute of Clinical Medicine, Vilnius University, Vilnius, Lithuania 6Department of Paediatrics, St-Pierre Hospital Brussels, Brussels, Belgium 71st Department of Paediatrics, National and Kapodistrian University of Athens (NKUA) School of Medicine, Agia Sophia Children’s Hospital of Athens, Athens, Greece 8Department of Paediatric Infectious Diseases, Alder Hey Children’s Hospital, Liverpool, UK 9Department of Paediatric Infectious Diseases Department, La Paz University Hospital, Madrid, Spain 10Department of Paediatrics, University General Hospital of Patras, Patras Medical School, Patras, Greece 11Division of Paediatric Infectious Diseases and Rheumatology, Department of Paediatrics and Adolescent Medicine, University Medical Centre, Medical Faculty, University of Freiburg, Freiburg, Germany 12National Heart and Lung Division, Faculty of Medicine, Imperial College London, London, UK 13Department of Medical Microbiology, Amsterdam UMC, Amsterdam, The Netherlands 14Department of Viroscience, ErasmusMC, Rotterdam, The Netherlands 15Sophia Children’s Hospital, ErasmusMC, Rotterdam, The Netherlands 16Department of Infectious Diseases and Vaccinology, University of Basel Children’s Hospital (UKBB), Basel, Switzerland
REFERENCES

1. GB-D 2015 Child Mortality Collaborators. Global, regional, and national, and selected subnational levels of stillbirths, neonatal, infant, and under-5 mortality, 1980-2015: a systematic analysis for the global burden of disease study 2015. *Lancet* 2016;388:1725-74.

2. Beletew B, Bimer DL, Mengesha A, et al. Prevalence of pneumonia and its associated factors among under-five children in East Africa: a systematic review and meta-analysis. *BMJ Pediatr* 2020;20:254.

3. Nair H, Simões EA, Rudan I, et al. Global and regional burden of hospital admissions for severe acute lower respiratory infections in young children in 2010: a systematic analysis. *Lancet* 2013;381:1380-90.

4. McAllister DA, Liu L, Shi T, et al. Global, regional, and national estimates of pneumonia morbidity and mortality in children younger than 5 years between 2000 and 2015: a systematic analysis. *Lancet Glob Health* 2019;7:e47-57.

5. Jiang W, Wu M, Zhou J, et al. Etiologic spectrum and occurrence of coinfections in children hospitalized with community-acquired pneumonia. *BMJ Infect Dis* 2017;17:787.

6. Wong-Chew RM, García-León ML, Noyola DE, et al. Respiratory viruses detected in Mexican children younger than 5 years old with community-acquired pneumonia: a national multicenter study. *Int J Infect Dis* 2017;62:32-8.

7. Bénét T, Sánchez Picot V, Messaadou M, et al. Microorganisms Associated With Pneumonia in Children <5 Years of Age in Developing and Emerging Countries: The SABGIEL Pneumonia Multicenter, Prospective, Case-Control Study. *Clin Infect Dis* 2017;65:604-12.

8. Lavi E, Breuer O. The impact of prior antibiotic therapy on outcomes in children hospitalized for community-acquired pneumonia. *Curr Infect Dis Rep* 2016;18:3.

9. Greigbrech D, Givon-Lavi N, Ben-Shimol S, et al. Impact of PCV7/ PCV13 introduction on community-acquired alveolar pneumonia in children <5 years. *Vaccine* 2015;33:4623-9.

10. Izurieta P, Bahety P, Adegbola R, et al. Public health impact of pneumococcal conjugate vaccine introduction: assessment of invasive pneumococcal disease burden and serotype distribution. *Expert Rev Vaccines* 2018;17:478-93.

11. Kleyhans J, Cohen C, McMorrow M, et al. Can pneumococcal meningitis surveillance be used to assess the impact of pneumococcal conjugate vaccine on total invasive pneumococcal disease? A case-study from South Africa, 2005-2016. *Vaccine* 2019;37:5724-30.

12. Jackson C, Haia Y, BieickJI, et al. Estimating global trends in total and childhood antibiotic consumption, 2011-2015. *BMJ Glob Health* 2019;4:e001241.

13. Levine OS, O’Brien KL, Deloria-Knoll M, et al. The pneumonia etiology research for child health project: a 21st century childhood pneumonia etiology study. *Clin Infect Dis* 2012;54 Suppl 2:S93-101.

14. Higdon MM, Hammitt LL, Deloria Knoll M, et al. Should controls with respiratory symptoms be excluded from case-control studies of pneumonia etiology? Reflections from the PERCH study. *Clin Infect Dis* 2017;64:S205-12.

15. Jain S, Williams DJ, Arnold SR, et al. Community-Acquired pneumonia requiring hospitalization among U.S. children. *N Engl J Med* 2015;372:835-45.

16. Cebe-López M, Herberg J, Pardo-Seco J, et al. Viral co-infections in pediatric patients hospitalized with lower tract acute respiratory infections. *PLoS One* 2015;10:e0136526.

17. Zhang X, Chen Z, Gu W, et al. Viral and bacterial co-infection in hospitalised children with refractory Mycoplasma pneumoniae pneumonia. *Epidemiol Infect* 2018;146:1384-8.

18. Hoffmann J, Machado D, Terrier O, et al. Viral and bacterial coinfection in severe pneumonia triggers innate immune responses and specifically enhances IP-10: a translational study. *Sci Rep* 2016;6:38532.

19. Brand KH, de Groot R, Galama JMD, et al. Infection with multiple viruses is not associated with increased disease severity in children with bronchiolitis. *Pediatr Pulmonol* 2012;47:393-400.

20. Asner SA, Rose W, Petroch A, et al. Is virus coinfection a predictor of severity in children with viral respiratory infections? *Clin Microbiol Infect* 2015;21:264.e1–264.e6.

21. Cebe-López M, Herberg J, Pardo-Seco J, et al. Does viral co-infection influence the severity of acute respiratory infection in children? *PLoS One* 2016;11:e0152481.

22. Bogaert D, De Groot R, Hermans PWI. Streptococcus pneumoniae colonisation: the key to pneumococcal disease. *Lancet Infect Dis* 2004;4:144-54.

23. Pneumonia Etiology Research for Child Health (PERCH) Study Group. Causes of severe pneumonia requiring hospital admission in children without HIV infection from Africa and Asia: the PERCH multicountry case-control study. *Lancet* 2018;394:757-79.

24. Li Y, Reeves RM, Wang X, et al. Global patterns in monthly activity of influenza virus, respiratory syncytial virus, parainfluenza virus, and metapneumovirus: a systematic analysis. *Lancet Glob Health* 2019;7:e1031-45.

25. Hammitt LL, Feikin DR, Scott JAG, et al. Addressing the analytic challenges of cross-sectional pediatric pneumonia etiology data. *Clin Infect Dis* 2017;64:S197-204.

26. Dunne EM, Smith-Vaughan HC, Robins-Browne RM, et al. Nasopharyngeal microbial interactions in the era of pneumococcal conjugate vaccination. *Vaccine* 2013;31:2333-42.

27. Skevaki CL, Tsialta P, Trochosou Al, et al. Associations between viral and bacterial potential pathogens in the nasopharynx of children with and without respiratory symptoms. *Pediatr Infect Dis J* 2015;34:1296-301.

28. Dietrich A, Hirsch HH, Decker M-L, et al. Mycoplasma pneumoniae detection in children with respiratory tract infections and influence on management - a retrospective cohort study in Switzerland. *Acta Paediatr* 2020;109:375-80.

29. Jacups SP. The continuing role of Haemophilus influenzae type b carriage surveillance as a mechanism for early detection of invasive disease activity. *Hum Vaccin* 2011;7:1254-60.

30. Kohns Vasconcelos M, Renk H, Pielapska J, et al. SARS-CoV-2 testing and infection control strategies in European paediatric emergency departments during the first wave of the pandemic. *Eur J Pediatr* 2021;180:1-7.
31 Vasconcelos MK, Epalza C, Renk H, et al. Harmonisation preserves research resources. *Lancet Infect Dis* 2021;21:e71.

32 Azadeh N, Sakata KK, Saeed A, et al. Comparison of respiratory pathogen detection in upper versus lower respiratory tract samples using the BioFire FilmArray respiratory panel in the immunocompromised host. *Can Respir J* 2018;2018:2685723:1–6.

33 Boonyaratanakornkit J, Vivek M, Xie H, et al. Predictive value of respiratory viral detection in the upper respiratory tract for infection of the lower respiratory tract with hematopoietic stem cell transplantation. *J Infect Dis* 2020;221:379–88.

34 Loens K, Van Heirstraeten L, Malhotra-Kumar S, et al. Optimal sampling sites and methods for detection of pathogens possibly causing community-acquired lower respiratory tract infections. *J Clin Microbiol* 2009;47:21–31.

35 Nascimento-Carvalho CM. Community-Acquired pneumonia among children: the latest evidence for an updated management. *J Pediatr* 2020;96 Suppl 1:29–38.

36 Schuh S, Babi FE, Dalziel SR, et al. Practice variation in acute bronchiolitis: a pediatric emergency research networks study. *Pediatrics* 2017;140:e20170842.

37 Tannous R, Haddad RN, Torbey P-H. Management of community-acquired pneumonia in pediatrics: adherence to clinical guidelines. *Front Pediatr* 2020;8:302.

38 Donà D, Barbieri E, Daverio M, et al. Implementation and impact of pediatric antimicrobial stewardship programs: a systematic scoping review. *Antimicrob Resist Infect Control* 2020;9:3.

39 Feudtner C, Feinstein JA, Zhong W, et al. Pediatric complex chronic conditions classification system version 2: updated for ICD-10 and complex medical technology dependence and transplantation. *BMC Pediatr* 2014;14:199.