ERSEM 15.06: a generic model for marine biogeochemistry and the ecosystem dynamics of the lower trophic levels

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Abstract. The ERSEM model is one of the most established ecosystem models for the lower trophic levels of the marine food web in the scientific literature. Since its original development in the early nineties it has evolved significantly from a coastal ecosystem model for the North Sea to a generic tool for ecosystem simulations from shelf seas to the global ocean. The current model release contains all essential elements for the pelagic and benthic part of the marine ecosystem, including the microbial food web, the carbonate system and calcification. Its distribution is accompanied by a testing framework enabling the analysis of individual parts of the model. Here we provide a detailed mathematical description of all ERSEM components along with case studies of mesocosm type simulations, water column implementations and a brief example of a full-scale application for the North-West European shelf. Validation against in situ data demonstrates the capability of the model to represent the marine ecosystem in contrasting environments.

1 Introduction

Over the last two decades a number of marine ecosystem models describing ocean biogeochemistry and the lower trophic levels of the food web have emerged in a variety of contexts ranging from simulations of batch cultures or mesocosms over estuarine and coastal systems to the global ocean (e.g. Fasham et al., 1990; Flynn, 2010; Geider et al., 1997; Wild-Allen et al., 2010; Zavatarelli and Pinardi, 2003; Aumont et al., 2003; Follows et al., 2007; Yool et al., 2013; Stock et al., 2014). Some of them have matured with the years into sound scientific tools in operational forecasting systems and are used to inform policy and management decisions regarding essential issues of modern human society, such as climate change, ecosystem health, food provision and other ecosystem goods and services (e.g. Lenhart et al., 2010; Glibert et al., 2014; van der Molen et al., 2014; Doney et al., 2012; Bopp et al., 2013; Chust et al., 2014; Barange et al., 2014). Given the importance of these applications, transparent descriptions of the scientific contents of these models are necessary in order to allow full knowledge and assessment of their strength and weaknesses, as well as maintenance and updating according to scientific insight and progress.

Here we provide a full description of one of these models, ERSEM (European Regional Seas Ecosystem Model), developed in the early nineties (Baretta et al., 1995; Baretta, 1997) out of a European collaborative effort, building on previous developments (Radford and Joint, 1980; Baretta et al., 1988). Subsequent development of the model has occurred in separate streams leading to individual versions of the model, the main ones being the ERSEM version described in Allen et al. (2001); Blackford and Burkill (2002); Blackford et al. (2004) and the version of Vichi et al. (2004, 2007); Leeuwen et al. (2012); van der Molen et al. (2014); http://www.nioz.nl/northsea_model, also referred to as the Biogeochemical Flux Model. The present release is based on the former development stream (Blackford et al., 2004). It has since the beginnings of ERSEM gradually evolved into what is now the principal model for shelf-seas applications.

1 The two given references are the introductions to two special issues published on the original model versions ERSEM I and II, representing the entire volumes. More specific reference to single papers within these volumes are given in the relevant process descriptions.
within the UK and beyond. It is part of the operational suite of the UK Met Office, and the biogeochemical component for the North-West European shelf seas within the European Copernicus Marine Service.

While it was originally created as a scientific tool for the North Sea ecosystem (hence the name), it has since evolved considerably in its scientific content, broadening the scope of the model to coastal systems across the globe as well as the open ocean. Allen et al., 2001 adopted the model for simulations across the entire North-West European shelf sea, further extended in Holt et al., 2012 and Artioli et al., 2012 to include the North East Atlantic. Blackford et al., 2004 applied the model across six different ecosystem types across the globe, Barange et al., 2014 used applications of the model in the major coastal upwelling zones of the planet, and Kwiatkowski et al., 2014 assessed the skill of the model, demonstrating its competitiveness with respect to other established global ocean models. The model has been subject to validation on various levels ranging from basic statistical metrics of point-to-point matches to observational data (Shutler et al., 2011; de Mora et al., 2013) to multivariate analysis (Allen et al., 2001; Allen and Somerfield, 2009) and pattern recognition (Saux Picart et al., 2012).

The model has been applied in a wide number of contexts that include short-term forecasting (Edwards et al., 2012), ocean acidification (Blackford and Gilbert, 2007), climate change (Holt et al., 2012), coupled climate-acidification projections (Artioli et al., 2013), process studies (Polimene et al., 2012), biogeochemical cycling (Wakelin et al., 2012), habitat (Villarino et al., 2015) and end-to-end modelling (Barange et al., 2014). The wide range of applications and uses of the model coupled with developments since earlier manuscripts documenting the model (Baretta-Bekker, 1995; Blackford et al., 2004) make a thorough and integral publication of its scientific ingredients overdue.

Being an evolution of former models within the ERSEM family that emerged in parallel to other, separate development streams of the original model, the core elements of the current model version closely resemble earlier versions even if presented in much more detail compared to previous works. We present a model for ocean biogeochemistry, the planktonic and benthic parts of the marine ecosystem that includes explicitly the cycles of the major chemical elements of the ocean (carbon, nitrogen, phosphorus, silicate and iron); it includes the microbial food web, a sub-module for the carbonate system, calcification and a full benthic model.

Our main objective with this paper is to provide a full description of all model components, accompanied by simple case studies with low resource requirements that illustrate the model capabilities and enable the interested reader to implement our model and reproduce the test cases shown. To this purpose we present the examples of a mesocosm type framework and three vertical water-column implementations of opposing character complemented with basic validation metrics against in-situ observations. All material required to replicate these test cases, such as parameterisation and input files, are provided in the Supplement. In addition, a brief illustration of a full scale three dimensional implementation is given to show the model in a large scale application.

The next section gives an overview of the model and its philosophy while the two following sections contain the descriptions of the pelagic and benthic components, describe the air–sea and seabed interfaces and detail some generic terms that are used throughout the model. The model description is complemented by two sections that present different implementations of the model and illustrate the testing framework. We complete the work with a section on optional choices of model configuration and a section on the technical specifications of the software package, licence and instructions of where and how to access the model code.

2 The ERSEM model

ERSEM is, since its origins, an ecosystem model for marine biogeochemistry, pelagic plankton and benthic fauna. Its functional types (Baretta et al., 1995; Vichi et al., 2007) are based on their macroscopic role in the ecosystem rather than species or taxa and its state variables are the major chemical components of each type (carbon, chlorophyll a, nitrogen, phosphate, silicate and optionally iron). It is composed of a set of modules that compute the rates of change of its state variables given the environmental conditions of the surrounding water body, physiological processes and preditor–prey interactions. In the simplest case the environmental drivers can be provided offline, or through a simple 0-dimensional box model. However, for more realistic representations, including the important processes of horizontal and vertical mixing (or advection) and biogeochemical feedback, a direct (or online) coupling to a physical driver, such as a 3-D hydrodynamic model, is required.

The organisms in the model are categorised along with the main classes of ecosystem function into primary producers, consumers and bacterial decomposers, particulate and dissolved organic matter (POM, DOM) in the pelagic and consumers, bacterial decomposers, particulate and dissolved organic matter in the benthos. Most of these classes are further subdivided into sub-types to allow for an enhanced plasticity of the system in adapting the ecosystem response to the environmental conditions in comparison to the classical NPZD type models. Importantly, ERSEM uses a fully dynamic stoichiometry in essentially all its types (with the exception of mesozooplankton, benthic bacteria and zoobenthos which use fixed stoichiometric ratios). The model dynamics of a living functional type are generally based on a standard organism that is affected by the assimilation of carbon and nutrients into organic compounds by uptake, and the generic loss processes of respiration, excretion, release, predation and non predatory mortality (Fig. 1; see also Vichi et al., 2007) – “2. Towards a generic formalism for pelagic
Mathematically, the set of prognostic equations describing the dynamics of marine biogeochemical states is generally given by:

\[
\frac{\partial c_p}{\partial t} + \mathbf{u} \cdot \nabla c_p + w_{sp} \frac{\partial c_p}{\partial z} = \nu \frac{\partial^2 c_p}{\partial z^2} + \frac{\partial c_p}{\partial t} \bigg|_{\text{bgc}} \quad (1)
\]

\[
\frac{\partial c_b}{\partial t} = \frac{\partial c_{bgc}}{\partial t} \bigg|_{\text{bgc}},
\]

where \(c_p\) are the pelagic concentrations (per volume) and \(c_b\) the benthic contents (per sediment surface area) of each chemical component of the organic model types or the inorganic model components. \(w_{sp}\) is the velocity of gravitational sinking of particles in the water column. \(\mathbf{x}\) represents the vector of spatial coordinates of which \(z\) is the vertical coordinate being 0 at sea surface and increasing downwards.

The set of equations is closed by the horizontal boundary conditions of the system generally given by the air–sea fluxes \(F|_{\text{air}}\) and the fluxes across the seafloor \(F|_{\text{ben}}\) and lateral boundary conditions if present in the given configuration.

ERSEM computes the biogeochemical rates of change in pelagic (\(\frac{\partial c_p}{\partial t} |_{\text{bgc}}\)) and benthic (\(\frac{\partial c_{bgc}}{\partial t} |_{\text{bgc}}\)) systems, the gas transfer across sea-surface (\(F|_{\text{air}}\) for oxygen and carbon) and the fluxes across the seabed (\(F|_{\text{ben}}\)). The actual numerical integration of these rates along with the advection-diffusion processes that solves Eqs. (1) and (2) needs to be addressed appropriately through an external driver as e.g. discussed in (Butenschön et al., 2012).

2.1 Nomenclature and units

Pelagic state variables in ERSEM are concentrations and are referred to as \(c_p\). When indicating a specific class or type, they are denoted by upper case letters (\(P\): phytoplankton, \(Z\): zooplankton, \(B\): bacteria, \(R\): organic matter, \(O\): gases, \(N\): nutrients), with the chemical component in the subscript in blackboard style (\(C\): carbon, \(N\): nitrogen, \(P\): phosphorus, \(S\): silicon, \(F\): iron with the exception of the chlorophyll \(a\) components which are distinguished by using \(C\), as chlorophyll \(a\) is not a chemical element but a compound), and the specific type in the super-script, e.g. \(P_{\text{C}}\) for diatom carbon. Correspondingly, benthic states use \(c_b\), for generic contents and the specific states (\(H\): bacteria \(Y\); zoobenthos, \(Q\): organic matter, \(G\): gases, \(K\): nutrients, \(D\): states of vertical distribution). Primed (‘) mark available concentrations or contents to loss processes (see Sect. 2.2). Where equations are valid for more than one specific functional type \(\chi, \psi, \Psi\) are used as place holders for functional types and the chemical components may be given as a comma separated list, implying that an equation is valid for all these components, e.g. \(P_{\text{C,N,P}}\) rep-
Figure 2. ERSEM schematic showing how model components interact with or influence each other. Blue connectors represent inorganic carbon fluxes, red represents nutrient fluxes, yellow represents oxygen, black represents predator-prey interactions and green represents fluxes of non-living organics. Dashed arrows indicate the influence of carbonate system variables.

represents the carbon, phosphorus and nitrogen content of each phytoplankton type. The physical environment is given in roman letters, e.g. $T$ for temperature.

Parameters are represented by lower case letters with $r$ for specific rates, $q$ for quotas or fractions, $l$ for limitation or regulating factors, $h$ for half-saturation constants and $p$ for most others. Food preferences of predators on their prey are given as $f_{ZP}$, $Z$ being the preference of predator $Z$ on food $P$.

Fluxes between state variables are given as $F_{A,B}$ for the flux from $A$ to $B$. Specific rates are notated using $S$. Dynamic internal quotas of two components $A$ and $B$ are given by the notation $q_{A,B}$, e.g. $q_{\text{N:C}}$ being the internal nitrogen to carbon quota of diatoms $P_{\text{dia}}$. Derived quotas or fractions are given by a caligraphic $Q$.

The coordinate system used describes the horizontal coordinates in $x$ and $y$, while the vertical coordinate is given by $z$, 0 at the sea surface increasing downwards. The corresponding velocity fields are given by $u$, $v$ and $w$. We are referring to Cartesian coordinates in this publication for simplicity.

The sediment depth coordinate is given by $\zeta$, which is 0 at the sediment surface increasing downwards.

All equations are given as scalar equations for a single pixel of the model domain.

Rates of change of the biogeochemical state variables due to individual subprocesses or groupings of these are given as $\frac{\partial \phi}{\partial t}_{\text{subprocess}}$, where the following abbreviations are used for the subprocesses: bgc = biogeochemical fluxes, bur = burying, calc = calcification, decom = decomposition, denit = denitrification, diss = dissolution, exer = excretion, mort = mortality, net = comprehensive net fluxes, nitr = nitrification, pred = predation, rel = release, remin = remineralisation, resp = respiration, scav = scavenging, sed = sedimentation, upt = uptake.

In equations that hold for multiple functional groups or components squared brackets are used for terms that are only valid for a single functional group or component.

Units in the model for all organic and inorganic nutrient concentrations are in $\text{mmol m}^{-3}$ with the exception of iron being in $\mu\text{mol m}^{-3}$. All forms of organic carbon are in $\text{mg m}^{-3}$ while all species of inorganic carbon are in $\text{mmol m}^{-3}$ with the exception of the internal calculations of the carbonate system where they are converted to $\mu\text{mol kg}^{-1}$. Corresponding benthic contents are two-dimensional and consequently given in $\text{mmol m}^{-2}$, $\text{mg m}^{-2}$ and $\mu\text{mol m}^{-2}$. The penetration depth and depth horizons in the sediments are given in $m$. Temperatures are generally considered in $^\circ\text{C}$, salinity in psu, sea-water density in $\text{kg m}^{-3}$ and pressure is given in $\text{Pa}$, with the exception of the internal calculations of the carbonate system where temperature is converted to absolute temperature in $K$ and pressure to bar. Partial pressure of carbon dioxide is used in ppm.
2.2 Dependencies on the physical environment

Several processes in the model depend directly on the physical environment that the model states are exposed to:

– Metabolic processes depend on the sea-water temperature.

– Primary production relies additionally on the photosynthetically active radiation (PAR) as energy input which should be computed from shortwave radiation at the sea surface $I_{surf}$, taking into account the attenuation coefficients given in Sect. 3.9. Note, that the model requires the average light in each discrete model cell, which is not given by the light at the cell centre, but by the vertical integral of the light curve divided by the cell depth.

– Empirical regressions for alkalinity, saturation states and chemical equilibrium coefficients of the carbonate system reactions require temperature $T$, salinity $S$, pressure $p$ and density $\rho$ of the sea-water.

– Air–sea fluxes of carbon dioxide and oxygen depend on temperature $T$ and the absolute wind speed $u_{wind}$ near the sea-surface.

– Deposition of organic matter on the sea floor and resuspension depend on the shear stress at the sea floor $\tau_{bed}$.

– The optional light attenuation model based on inherent optical properties requires the geographical coordinates of each model pixel and the current simulation date and time in order to compute the zenith angle.

2.3 States and negativity control

In order to avoid the occurrence of negative concentrations or contents in the integration process and reduce the vulnerability to numerical noise all state variables include a lower buffer $\epsilon_{p,b}$, based on a carbon concentration of 0.01 mg m$^{-3}$ modified adequately for the various state variables using reference stoichiometric quotas and unit conversions. This buffer is not accessible to the loss processes of the biogeochemical dynamics. Consequently all processes that diminish the biomass of each state are based on the available concentrations or contents given by $\epsilon_{p,b} = \epsilon_{p,b} - \epsilon_{p,b}$. These small resilient buffers additionally support the spawning of new biomass as soon as favourable conditions occur, similar to the low overwintering biomass limits in Fennel [1995].

Note that when calculating the overall budget of a domain, these background concentrations should be subtracted in order to give adequate results.

3 The pelagic system

In its current form the pelagic part of ERSEM comprises 4 functional types for primary producers, originally defined as diatoms, nanoflagellates, picophytoplankton and dinoflagellates. This classification was historically coined for the North Sea but has since been widened to a broader interpretation almost exclusively based on the single trait size (with the exception of the requirement of silicate by diatoms and an implicit calcification potential of nanoflagellates) leading to the classes of picophytoplankton, nanophytoplankon, microphytoplankon and diatoms. Similarly the zooplankton pool is divided into heterotrophic nanoflagellates, microzooplankton and mesozooplankton. Particular organic matter is treated in three size classes (small, medium and large) in relation to its origin. Dissolved organic matter is treated according to its decomposition time scales into a labile dissolved inorganic state, semi-labile and semi-refractory carbon (see Sect. 3.3).

The inorganic state variables of the pelagic model are dissolved oxidised nitrogen, ammonium, phosphate, silicic acids, dissolved inorganic iron, dissolved inorganic carbon, dissolved oxygen and calcite. In addition the model holds a state variable for alkalinity subject to fluctuations generated from the modelled biogeochemical processes (see Sect. 3.8 and Artioli et al. [2012]. The complete list of pelagic state variables is given in Table 1.

The recently implemented iron cycle (following largely the implementation of Vichi et al. [2007] and the silicate cycle are abbreviated for simplicity; their pathways by-pass the predators and decomposers by turning grazing of phytoplankton iron or silicate directly into detritus and remineralising iron implicitly from detritus into the dissolved inorganic form, while silicate is not remineralized in the water column. Chlorophyll $a$ takes a special role in between the chemical components of the model: being a compound of other elements it is not strictly conserved by the model equations but rather derived from assimilation of carbon and subsequent decomposition of organic compounds. The addition of chlorophyll $a$ states to the model allows for dynamic chlorophyll $a$ to carbon relationships in the photosynthesis description and a more accurate comparison to observations of biomass or chlorophyll $a$.

The growth dynamics in the model are generally based on mass-specific production and loss equations that are expressed in the currency of each chemical component, regulated and limited by the availability of the respective resources.

3.1 Primary producers

The phytoplankton dynamics are modelled for each phytoplankton type as a net result of source and loss processes (Varela et al. [1995]). The carbon and chlorophyll $a$ component is given by uptake in the form of gross primary production and the losses through excretion, respiration, predation by zooplankton and mortality in the form of lysis, while the nutrient content is balanced by uptake, release, predation and
mortality in the form of lysis:

\[
\frac{\partial \chi_{c,c}}{\partial t} = \frac{\partial \chi_{c,c}}{\partial t} _{bgc} - \frac{\partial \chi_{c,c}}{\partial t} _{gpp} - \frac{\partial \chi_{c,c}}{\partial t} _{excr} - \frac{\partial \chi_{c,c}}{\partial t} _{resp} - \frac{\partial \chi_{c,c}}{\partial t} _{pred} - \frac{\partial \chi_{c,c}}{\partial t} _{mort},
\]

(3)

\[
\frac{\partial \chi_{n,p,f,[s]}}{\partial t} = \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{bgc} - \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{upt} - \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{rel} - \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{pred} - \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{mort},
\]

(4)

with \( \chi \) in (pico, nano, micro, dia) and where the silicate component (S) is only active for diatoms.

The formulation of photosynthesis combines the form originally presented in Baretta-Bekker et al. (1997) for the balance of carbon assimilation, excretion and respiration with the negative exponential light harvesting model based on Jassby and Platt (1976), Platt et al. (1982) and Geider et al. (1997) in order to describe the total specific carbon fixation. In this formulation the gross carbon assimilation is assumed to be not depending on nitrogen and phosphorus. Total gross primary production (GPP) is assumed to be composed of a fraction which is assimilated (cellular GPP) through photosynthesis and a fraction which is not utilisable, e.g. due to nutrient limitation, and excreted. A similar approach can be found in Falkowski and Raven (2007). The idea behind this assumption is that nutrient (or specifically nitrogen and phosphorus) limitation affects more the assimilation of newly fixed carbon into cellular biomass (assimilation) than the photosynthesis itself.

Table 1. Pelagic functional types and their components (squared brackets indicate optional states) – chemical components: C carbon, N nitrogen, P phosphorus, F iron, S silicate, C chlorophyll a.

| Symbol | Code | Description |
|--------|------|-------------|
| pico   | P3c,n,p[f],Chl3 | Picophytoplankton (<2µm) |
| nano   | P2c,n,p[f],Chl2 | Nanophytoplankton (2-20µm) |
| micro  | P4c,n,p[f],Chl4 | Microphytoplankton (>20µm) |
| dia    | P1c,n,p[f],Chl1 | Diatoms |
| HET    | Z6c,n,p | Heterotrophic Flagellates |
| ZC     | Z5c,n,p | Microzooplankton |
| BC,N,P | Z4c  | Mesozooplankton |
| lab    | B1c,n,p | Heterotrophic Bacteria |
| R(slab)| R1c,n,p | Labile dissolved organic matter |
| R_C    | R2c  | Semi-labile organic matter |
| R_C    | R3c  | Semi-refractory organic matter |
| R_C    | R4c  | Small particulate organic matter |
| med    | R6c  | Medium size particulate organic matter |
| large  | R8c  | Large particulate organic matter |
| [calc  | [L2c] | Calcite |
| O_D    | O2o  | Dissolved oxygen |
| OC     | O3c  | Dissolved inorganic carbon (DIC) |
| N_P    | N1p  | Phosphate |
| ex     | N3n  | Oxidised nitrogen |
| N_am   | N4n  | Ammonium |
| N_S    | N5s  | Silicate |
| [N_F]  | [N7f] | Dissolved iron |
| [bioAlk]| [bioAlk] | Bioalkalinity |

\( \chi \) is the nitrogen mass concentration in the water column, \( \partial t \) is the time derivative, and \( \partial \chi_{c,c} \) is the change in nitrogen mass concentration in the carbon component. The decomposition of the nitrogen mass concentration into assimilation, excretion and respiration is described by\( \frac{\partial \chi_{c,c}}{\partial t} _{bgc} \),\( \frac{\partial \chi_{c,c}}{\partial t} _{gpp} \),\( \frac{\partial \chi_{c,c}}{\partial t} _{excr} \),\( \frac{\partial \chi_{c,c}}{\partial t} _{resp} \),\( \frac{\partial \chi_{c,c}}{\partial t} _{pred} \) and \( \frac{\partial \chi_{c,c}}{\partial t} _{mort} \) respectively.

The decomposition of the nitrogen mass concentration into uptake, limitation and respiration is described by\( \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{bgc} \),\( \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{upt} \),\( \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{rel} \),\( \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{pred} \) and \( \frac{\partial \chi_{n,p,f,[s]}}{\partial t} _{mort} \) respectively.
Phytoplankton mass-specific gross primary production is then computed as
\[
\chi_{gpp} = \chi_{\max} \frac{\chi}{T} \chi_{\phi} \left( 1 - e^{-\frac{\chi_{\max} \chi_{\phi}}{\chi_{\max} \chi_{\phi} + \chi_{\phi}}} \right) e^{-\frac{\chi_{\max} \chi_{\phi}}{\chi_{\max} \chi_{\phi} + \chi_{\phi}}},
\]
(5)

based on the formulation by Geider et al. (1997) modified for photoinhibition according to Blackford et al. (2004). The symbols in this equation represent the chlorophyll \(a\) to carbon quota of each functional type \(\chi_{c,c}\) and the metabolic response to temperature \(\chi_{T}\) (see Eq. 233) and the silicate and iron limitation factors \(\chi_{S,F} \in [0,1]\) (see Eqs. 233 and 234). The \(\chi_{\max}\) are the maximum potential photosynthetic rate parameters in unlimited conditions at reference temperature. Note, that these are different to the maximum potential growth rates usually retrieved in physiological experiments (e.g. in the work of Geider et al. 1997) or measured at sea, in that they are exclusive upper bounds of the specific growth rate function. In fact, the products of the exponential terms in Eq. (5) have a maximum of
\[
1.0 - \left( 1 - \frac{\chi_{\max} \chi_{\phi}}{\chi_{\max} \chi_{\phi} + \chi_{\phi}} \right) - \frac{\chi_{\max} \chi_{\phi}}{\chi_{\max} \chi_{\phi} + \chi_{\phi}} < 1.
\]
In addition, we refer to gross primary production here as total carbon fixation, a fraction of which is directly excreted. Other parameters are the initial slope \(\chi_{\phi}\) and the photoinhibition parameter \(\chi_{\phi}\) of the light saturation curve (Platt et al. 1982).

A fraction of the specific gross production is directly excreted to the dissolved organic carbon (DOC) pool as a fixed fraction \(\chi_{excr}\) augmented according to the combined nitrogen and phosphorus limitation up to the total gross production:
\[
\chi_{excr} = \frac{\chi_{excr}}{\chi_{\max}} (1 - \frac{\chi_{excr}}{\chi_{\max}}),
\]
(6)

where \(\chi_{excr}\) is the combined nitrogen-phosphorus limitation factor defined in Eq. 232, based on the internal nutrient to carbon quotas according to Droop (1974).

The second generic sink term is given by lysis which occurs proportional to the current biomass by the constant specific rate \(\chi_{mort}\) augmented by nutrient stress according to:
\[
\chi_{mort} = \frac{1}{\min(\chi_{excr},\chi_{S}) + 0.1} \chi_{excr}.
\]
(7)

The carbon and chlorophyll \(a\) dynamics of each phytoplankton type in Eq. 3 are then specified by the following terms:

Carbon is assimilated according to
\[
\frac{d\chi_{C}}{dt} = \chi_{gpp} \chi_{C},
\]
(8)

The synthesis rate of chlorophyll \(a\) is given by:
\[
\frac{d\chi_{a}}{dt} = \chi_{(NP)} \chi_{gpp} \chi_{C},
\]
(9)

where \(\chi_{a}\) is the ratio of chlorophyll \(a\) synthesis to carbon fixation under nutrient replete conditions. It is given by:
\[
\chi_{a} = \left( \frac{\chi_{c,max} - q_{min,c}}{\alpha_{NP} E_{PAR}} \right) \chi_{gpp} + q_{min,c},
\]
(10)

where \(\chi_{c,max}\) are the maximum achievable chlorophyll \(a\) to carbon quota for each type, \(q_{min,c}\) is the minimum chlorophyll \(a\) to carbon ratio.

This formulation differs from the original formulation of Geider et al. (1997) in its asymptotic limit of the carbon to chlorophyll \(a\) synthesis at high PAR. In the original formulation the ratio is unbound, while in this formulation it is bound by the inverse minimum chlorophyll \(a\) to carbon ratio \(q_{min,c}\) in order to avoid excessive quotas not observed in nature.

As opposed to the previous formulation of Blackford et al. (2004), the relative synthesis of chlorophyll \(a\) is directly limited by the internal nutrient quota in order to compensate for the enhanced demand required to maintain the cell structure leading to a reduced investment into the light harvesting capacity.

The excretion of phytoplankton in terms of carbon and chlorophyll \(a\) is given by:
\[
\frac{d\chi_{C}}{dt} = \chi_{excr} \frac{d\chi_{C}}{dt} \chi_{C},
\]
(11)

Respiration of phytoplankton is split into respiration at rest, that is proportional to the current biomass by the constant specific rate \(\chi_{resp}\) complemented with an activity related term that is a fraction \(\chi_{aresp}\) of the assimilated amount.
of biomass per time unit after excretion:

\[
\frac{\partial Q_{C,C}}{\partial t}_{\text{resp}} = \chi_{\text{resp}} P'_{C,C} + \frac{\partial Q_{C,C}}{\partial t}_{\text{excr}}
\]

The losses of phytoplankton by lysis are given by

\[
\frac{\partial Q_{C,C}}{\partial t}_{\text{mort}} = S_{\text{mort}} P'_{C,C}
\]

while the individual terms of loss through predation of predator \( \Psi \) in

\[
\frac{\partial Q_{C,C}}{\partial t}_{\text{pred}} = \sum_{\Psi} F_{\Psi} P'_{C,C}
\]

are specified in the sections of the respective predators in Eqs. (11) and (16). \(^{32}\)

Nutrient uptake of nitrogen, phosphorus and iron is regulated by the nutrient demand of the phytoplankton group, limited by the external availability. Excretion is modelled as the disposal of non-utilisable carbon in photosynthesis while the release of nutrients is limited to the regulation of the internal stoichiometric ratio. This approach is consistent with observations that nutrient excretion plays a minor role in the phytoplankton fluxes (Puyo-Pay et al., 1997). Consequently, demand of nutrients may be positive or negative in sign in relation to the levels of the internal nutrient storages and the balance between photosynthesis and carbon losses, so that:

\[
\frac{\partial Q_{N,P,F}}{\partial t}_{\text{upt}} = \begin{cases} 
\text{min} & \left( F_{\text{demand}} Q_{N,P,F}, F_{\text{avail}} Q_{N,P,F} \right) \\
0 & \text{if } F_{\text{demand}} Q_{N,P,F} > 0 \\
0 & \text{if } F_{\text{demand}} Q_{N,P,F} < 0 
\end{cases}
\]

\[
\frac{\partial Q_{N,P,F}}{\partial t}_{\text{ref}} = \begin{cases} 
0 & \text{if } F_{\text{demand}} Q_{N,P,F} > 0 \\
0 & \text{if } F_{\text{demand}} Q_{N,P,F} < 0 
\end{cases}
\]

Nutrient demand (with the exception of silicate) is computed from assimilation demand at maximum quota \( \chi_{\text{max}}(N,P,F) \) complemented by a regulation term relaxing the internal quota towards the maximum quota and compensating for rest respiration:

\[
F_{\text{demand}} \left| Q_{N,P,F} \right| = S_{\text{gpp}} \left( 1 - Q_{\text{excr}} \right) \left( 1 - Q_{\text{arex}} \right) \chi_{\text{max}}(N,P,F) P'_{C,C} + r_{\text{lux}} \chi_{\text{max}}(N,P,F) P'_{C,C}
\]

where \( r_{\text{lux}} \) is the rate of nutrient luxury uptake towards the maximum quota.

Note, that these terms may turn negative when rest respiration exceeds the effective assimilation rate \( S_{\text{gpp}} \left( 1 - Q_{\text{excr}} \right) \left( 1 - Q_{\text{arex}} \right) \chi_{\text{max}}(N,P,F) P'_{C,C} \) or the internal nutrient content exceeds the maximum quota resulting in nutrient excretion in dissolved inorganic form. The maximum quota for nitrogen and phosphorus may exceed the optimal quota allowing for luxury storage while it is identical to the optimum quota for iron and silicate.

The uptake is capped at the maximum achievable uptake depending on the nutrient affinities \( N_{\text{aff}}(P,F,N) \) and the external dissolved nutrient concentrations:

\[
F_{\text{avail}} Q_{N,P,F} = \chi_{\text{aff}} N_{P,F} \chi_{\text{aff}} N_{P,F}
\]

where the nitrogen need is satisfied by uptake in oxidised and reduced form in relation to the respective affinities and external availability.

This purely linear formulation of maximum uptake proportional to the affinity is in contrast to the more widely used saturation assumption of Michaelis-Menten type (Aksnes and Egge, 1991). It is justified here as ERSEM treats phytoplankton in pools of functional groups, rather than individual species with defined saturation characteristics (Franks, 2009).

Lysis and predation losses are computed analogous to the carbon component:

\[
\frac{\partial Q_{N,P,F}}{\partial t}_{\text{mort}} = S_{\text{mort}} P'_{N,P,F},
\]

\[
\frac{\partial Q_{N,P,F}}{\partial t}_{\text{pred}} = \sum_{\Psi} F_{\Psi} P'_{N,P,F}.
\]

The variability of the internal silicate quota of diatoms reported in literature is small and there’s little evidence of luxury uptake capacity for this element (Brzezinski, 1985 [33], Moore et al., 2013). The silicate dynamics of diatoms are

\(^{32}\) Note that the dimensions of these are \( \text{volume}^1 \ast \text{mass}^{-1} \ast \text{time}^{-1} \) as opposed to \( \text{time}^{-1} \) as for most other rates.
therefore modelled by a simple relaxation towards the optimal quota given by the equations:

\[
\frac{\partial P_S}{\partial t} \bigg|_{\text{dia}} = \max \left( \frac{\partial}{\partial t} q_{\text{ref}, C} S_{\text{growth}}, 0 \right), \tag{22}
\]

\[
\frac{\partial P_S}{\partial t} \bigg|_{\text{rel}} = \max \left( \frac{\partial}{\partial t} \left( P'_S - \frac{\partial}{\partial t} q_{\text{ref}, C} P'_C, 0 \right) \right), \tag{23}
\]

\[
\frac{\partial P_S}{\partial t} \bigg|_{\text{mort}} = \frac{\partial}{\partial t} S_{\text{mort}} \frac{\partial}{\partial t} P'_S, \tag{24}
\]

\[
\frac{\partial P_S}{\partial t} \bigg|_{\text{pred}} = \sum_{\psi} \mathcal{F}_{\psi} \psi \frac{\partial}{\partial t} P'_S, \tag{25}
\]

where \( q_{\text{ref}, C} \) is the reference silicate to carbon quota of diatoms.

A formulation to model the impact of an increased atmospheric \( p_{\text{CO}_2} \) on phytoplankton carbon uptake that was introduced in [Artioli et al., 2014] is available via the CENH preprocessing option. In this case gross carbon uptake (Eq. 8) and activity respiration (the second term in Eq. 12) are enhanced by the factor \( \gamma_{\text{enhC}} \) defined as:

\[
\gamma_{\text{enhC}} = 1.0 + (p_{\text{CO}_2} - 379.48) \times 0.0005, \tag{26}
\]

where \( p_{\text{CO}_2} \) has the units ppm.

### 3.2 Predators

Predator dynamics are largely based on the descriptions of [Baretta-Bekker et al., 1995; Broekhuizen et al., 1995] and [Heath et al., 1997] described by the equations:

\[
\frac{\partial Z_C}{\partial t} \bigg|_{\text{bgc}} = \frac{\partial Z_C}{\partial t} \bigg|_{\text{upt}} - \frac{\partial Z_C}{\partial t} \bigg|_{\text{excr}} - \frac{\partial Z_C}{\partial t} \bigg|_{\text{resp}} - \frac{\partial Z_C}{\partial t} \bigg|_{\text{pred}} - \frac{\partial Z_C}{\partial t} \bigg|_{\text{mort}}, \tag{27}
\]

\[
\frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{bgc}} = \frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{upt}} - \frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{excr}} - \frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{rel}} - \frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{pred}} - \frac{\partial Z_{N,P}}{\partial t} \bigg|_{\text{mort}}. \tag{28}
\]

Note, that the iron and silicate cycles are simplified in a way that the iron/silicate content of phytoplankton subject to predation is directly turned into particulate organic matter (see Eqs. 69 and 70).

The pelagic predators considered in ERSEM are composed of three size classes of zooplankton categorised as heterotrophic flagellates, microzooplankton and mesozooplankton. According to size, these are capable of predating on different prey types including cannibalism as illustrated in Fig. 3.

The total prey available to each zooplankton type \( \chi \) is composed of the individual prey types \( \psi \) using type II Michaelis–Menten type uptake capacities ([Chesson, 1983; Gentleman et al., 2003]) as

\[
\mathcal{P}_{\chi,N,P} = \sum_{\psi} f_{\psi} \frac{\psi_{\chi}'}{\psi_{\chi} + h_{\min}}, \tag{29}
\]

where \( f_{\psi} \) are the food preferences and \( h_{\min} \) is a food half-saturation constant reflecting the detection capacity of predator \( \chi \) of individual prey types.

The prey mass specific uptake capacity for each zooplankton type \( \chi \) is then given by:

\[
S_{\chi,\text{growth}} = g_{\max} \frac{\chi}{T_{\chi}} \frac{2_{\chi}}{\mathcal{P}_{\chi}} \frac{1}{\chi + h_{\min}}, \tag{30}
\]

where \( g_{\max} \) is the maximum uptake capacity of each type at the reference temperature, \( t_{\chi} \) is the metabolic temperature response (Eq. 229), \( h_{\min} \) is a predation efficiency constant limiting the chances of encountering prey. Introducing the prey mass specific fluxes from prey \( \psi \) to predator \( \chi \)

\[
\mathcal{F}_{\psi} = S_{\chi,\text{growth}} f_{\psi} \frac{\psi_{\chi}'}{\psi_{\chi} + h_{\min}}, \tag{31}
\]

the zooplankton uptake can then be written as:

\[
\frac{\partial Z_{C,N,P}}{\partial t} \bigg|_{\text{upt}} = \sum_{\psi} \mathcal{F}_{\psi} \frac{\psi_{\chi}'}{\psi_{\chi} + h_{\min}}. \tag{32}
\]

This formulation is similar to the approach used in [Fasham et al., 1990], but introduces additional Michaelis–Menten
terms for individual prey types. The purpose here is to include sub-scale effects of pooling as prey of different types can be assumed to be distributed in separate patches in the comparatively large cell volume. Consequently, individual prey patches below a certain size are less likely to be grazed upon compared to the larger patches, which is expressed by the $h_{\text{min}}$ parameter.

Note, that in contrast to previous parametrisations, we now normalise the sum of the food preferences for each predator $Z_{\text{C,N,P}}$ to

$$\sum_{\psi} f_{\psi} \tilde{\psi} = 1,$$  \hspace{1cm} (33)

as non-normalised preferences lead to a hidden manipulation of the predation efficiency and at low prey concentrations of the maximum uptake capacity $\tilde{g}_{\text{max}}$.

The ingestion and assimilation of food by the predators is subject to inefficiencies that, given the wide diversity of uptake mechanisms within the zooplankton pools, is for simplicity taken as a fixed proportion of the gross uptake $1 - \tilde{\eta}_{\text{eff}}$. These losses are attributed to the excretion of faeces as a constant fraction $\chi_{\text{excr}}$ and activity costs in form of enhanced respiration $(1 - \tilde{\eta}_{\text{excr}})$.

The excretion term in Eq. 27 is then given by:

$$\frac{\partial \chi_{\text{C,N,P}}}{\partial t} \bigg|_{\text{excr}} = \frac{1}{\tilde{g}_{\text{eff}}} \frac{\partial \chi_{\text{C,N,P}}}{\partial t} \bigg|_{\text{upt}}.$$  \hspace{1cm} (34)

Respiration losses are composed of the activity costs and a basal respiration term required for maintenance and hence proportional to the current biomass by the constant factor $\chi_{\text{resp}}$ multiplied with the metabolic temperature response (Eq. 229):

$$\frac{\partial \chi_{\text{C}}}{\partial t} \bigg|_{\text{resp}} = (1 - \tilde{\eta}_{\text{eff}}) (1 - \tilde{\eta}_{\text{excr}}) \frac{\partial \chi_{\text{C}}}{\partial t} \bigg|_{\text{upt}} + \chi_{\text{resp}} \chi_{\text{T}} Z_{\text{C}}.$$  \hspace{1cm} (35)

This simple formulation of assimilation losses is closely related to the phytoplankton losses described in the previous section following the concept of the standard organism (Baretta et al. 1995) pending a better understanding of the underlying physiological mechanisms (Anderson et al. 2013).

Nitrogen and phosphorus are released regulating the internal stoichiometric quota:

$$\frac{\partial \chi_{\text{N,P}}}{\partial t} \bigg|_{\text{rel}} = \min \left( 0, \chi_{\text{N,P}} - \chi_{\text{N,P,C}} \chi_{\text{C}} \right) \chi_{\text{relNP}},$$  \hspace{1cm} (36)

where $\chi_{\text{relNP}}$ are the relaxation rates of release into dissolved inorganic form (see Eqs. 105 and 108).

Mortality is proportional to biomass based on a basal rate $\chi_{\text{p}}$ enhanced up to $\chi_{\text{p}} + \chi_{\text{mort}}$ under oxygen limitation $\chi_{\text{O}}$ (Eq. 239) as:

$$\frac{\partial \chi_{\text{N,P}}}{\partial t} \bigg|_{\text{mort}} = \left( 1 - \chi_{\text{O}} \right) \chi_{\text{p}} + \chi_{\text{mort}}) \chi_{\text{C,N,P}}.$$  \hspace{1cm} (37)

Biomass lost to other predators $\Psi$ is computed as:

$$\frac{\partial \chi_{\text{C,N,P}}}{\partial t} \bigg|_{\text{pred}} = \sum_{\Psi} \chi_{\Psi} \chi_{\text{C,N,P}}.$$  \hspace{1cm} (38)

**Mesozooplankton**

The top-level predator mesozooplankton takes a special role in the predator group in three respects:

- Its internal nutrient to carbon quota is assumed fixed (Gismervik 1997; Walve and Larsson 1999).
- it is capable of scavenging on particulate organic matter,
- at low prey it can enter a hibernation state (optional) at which its maintenance metabolism is reduced (Blackford et al. 2004).

The resulting overall balance of the mesozooplankton dynamics is in principle identical to the other zooplankton types (Eqs. 27, 28) with the exception of an additional release term for carbon in order to maintain the fixed internal stoichiometric quota:

$$\frac{\partial \chi_{\text{C}}}{\partial t} \bigg|_{\text{bgc}} = \frac{\partial \chi_{\text{C}}}{\partial t} \bigg|_{\text{upt}} - \frac{\partial \chi_{\text{C}}}{\partial t} \bigg|_{\text{excr}} + \chi_{\text{resp}} \chi_{\text{T}} Z_{\text{C}}.$$  \hspace{1cm} (39)

The differences to the heterotrophic flagellates and microzooplankton are given by the release terms for stoichiometric adjustments for carbon, nitrogen and phosphate (Eqs. 258 and 259) that replace nutrient release terms of the other two types (Eq. 36) and enhanced excretion for the scavenging on
particulate matter $\eta_{\text{res}}$ with respect to the uptake of living prey:

\[
\frac{\partial}{\partial t} Z^\text{MESO}_{C,N,P} = \left( 1 - \eta_{\text{eff}} \right) \frac{\partial Z^\text{MESO}_{C,N,P}}{\partial t} \psi_{\text{upt}}^\text{med} R^\text{MESO} \int_{\text{seafloor}}^{0} \frac{\partial}{\partial z} Z^\text{MESO}_{C,N,P} \, \text{d}z + \eta_{\text{res}} \frac{\partial}{\partial t} Z^\text{MESO}_{C,N,P} \psi_{\text{upt}}^\text{med} R^\text{MESO} \int_{\text{seafloor}}^{0} \frac{\partial}{\partial z} Z^\text{MESO}_{C,N,P} \, \text{d}z.
\] (40)

The hibernation formulation (optionally activated by the switch $Z_4_{\text{OW\_SW}}$) for over-wintering is triggered when the vertically integrated prey availability to mesozooplankton computed according to

\[
Pr_{\text{sw}} = \int_0^{\text{seafloor}} Pr_C \, \text{d}z
\] (41)

falls below the threshold $Pr_{\text{sw}}$.

In hibernation (overwintering) state the only active processes for mesozooplankton are respiration and mortality and using reduced the basal rates ($r_{\text{owresp}}$ and $r_{\text{owmort}}$) with respect to the active state:

\[
\frac{\partial}{\partial t} Z_C^\text{MESO} \bigg|_{\text{resp}} = r_{\text{owresp}} Z_C \left( Z_C \right) \quad \text{(42)}
\]

\[
\frac{\partial}{\partial t} Z_C^\text{MESO} \bigg|_{\text{mort}} = r_{\text{owmort}} Z_C \left( Z_C \right) \quad \text{(43)}
\]

### 3.3 Heterotrophic bacteria

Two alternative sub-modules for decomposition of organic material by bacteria are available in the ERSEM model involving different levels of decomposition of organic matter in the microbial food-web:

#### 3.3.1 Original version

In this version (Allen et al. [2002], Blackford et al. [2004], Baretta-Bekker et al. [1997]) bacteria feed explicitly only on labile dissolved organic matter $\dot{R}$. This is sufficient to create microbial loop dynamics in the model opening the pathway from dissolved organic matter (DOM) over bacteria to zooplankton, while the other forms of substrate are recycled implicitly (see Eq. [67]).

The biogeochemical dynamics of heterotrophic bacteria are here given by the equations:

\[
\frac{\partial B_C}{\partial t} \bigg|_{\text{hgc}} = \frac{\partial B_C}{\partial t} \bigg|_{\text{upt}} - \frac{\partial B_C}{\partial t} \bigg|_{\text{resp}} - \frac{\partial B_C}{\partial t} \bigg|_{\text{pred}} - \frac{\partial B_C}{\partial t} \bigg|_{\text{mort}},
\] (44)

\[
\frac{\partial B_{N,P}}{\partial t} \bigg|_{\text{hgc}} = \frac{\partial B_{N,P}}{\partial t} \bigg|_{\text{upt}} - \frac{\partial B_{N,P}}{\partial t} \bigg|_{\text{rel}} - \frac{\partial B_{N,P}}{\partial t} \bigg|_{\text{pred}} - \frac{\partial B_{N,P}}{\partial t} \bigg|_{\text{mort}}.
\] (45)

Bacterial uptake of DOM is given by a substrate mass specific turn-over rate $B_{\text{lab}}$ for labile dissolved organic matter when substrate is scarce and by a maximum bacteria mass specific potential uptake regulated by temperature and limited by nutrient and oxygen conditions when substrate is abundant and the uptake per bacteria is saturated, regulated by the ratio of bacteria over substrate biomass:

\[
B_{\text{upt}} = \min\left( B_{\text{lab}}, B_{\text{max}} T_T B_{\text{lab}} \min\left( l_{\text{Q}}, l_{\text{Q}, N} \right) B_C R_C \right),
\] (46)

\[
\frac{\partial B_{C,N,P}}{\partial t} \bigg|_{\text{upt}} = B_{\text{upt}} \frac{B_C}{R_C} \bigg|_{\text{upt}},
\] (47)

where $B_{\text{max}}$ is the maximum bacteria mass specific uptake of bacteria.

Mortality is given as a constant fraction of bacteria biomass:

\[
\frac{\partial B_{C,N,P}}{\partial t} \bigg|_{\text{mort}} = r_{\text{mort}} B_{C,N,P},
\] (48)

where $r_{\text{mort}}$ is a constant mass specific mortality rate for bacteria.

Bacterial respiration is computed according to activity respiration as an investment of activity in growth dependent on the oxygen state and a basal part:

\[
\frac{\partial B_{C}}{\partial t} \bigg|_{\text{resp}} = \left( 1 - \frac{B_{\text{resp}}}{\dot{R}} \right) l_0 - \frac{B_{\text{resp}}}{\dot{R}} \left( 1 - l_0 \right) \frac{\partial B_C}{\partial t} \bigg|_{\text{upt}} + \frac{B_{\text{resp}}}{\dot{R}} l_T B_C,
\] (49)

where $B_{\text{resp}}$ is the mass specific basal respiration rate at rest (representing the maintenance cost of the metabolism in absence of uptake activity) and $B_{\text{resp}, \text{low}}$ are the bacterial efficiencies at high and low oxygen levels.

Poor nutritional quality of the substrate may result in deprivation of nitrogen or phosphorus resulting in nutrient uptake in competition with phytoplankton for external dissolved nutrient sources, otherwise bacteria releases superfluous nutrients to the environment. The internal stoichiometric
quota of phosphorus is consequently balanced according to:

\[
\frac{\partial B_{\text{upt}}}{\partial t} = \begin{cases} 
B_{\text{rel}} \left( B_{\text{upt}} - B_{\text{max},\text{upt}} \right) B_{\text{C}} \frac{N_{\text{upt}}}{N_{\text{upt}} + h_{\text{N}}} & \text{if } B_{\text{upt}} < B_{\text{max},\text{upt}} \\
0 & \text{if } B_{\text{upt}} = B_{\text{max},\text{upt}} \\
0 & \text{if } B_{\text{upt}} > B_{\text{max},\text{upt}} \end{cases}
\]

with \( q_{\text{max},\text{upt}} \) being the optimal phosphorus to carbon quota of bacteria and \( r_{\text{B}} \) being the mass specific release rate.

For nitrogen the internal stoichiometric quota is balanced using ammonium:

\[
\frac{\partial B_{\text{N,upt}}}{\partial t} = \begin{cases} 
B_{\text{rel}} \left( B_{\text{upt}} - B_{\text{max},\text{upt}} \right) B_{\text{C}} \frac{N_{\text{upt}}}{N_{\text{upt}} + h_{\text{N}}} & \text{if } B_{\text{upt}} < B_{\text{max},\text{upt}} \\
0 & \text{if } B_{\text{upt}} = B_{\text{max},\text{upt}} \\
0 & \text{if } B_{\text{upt}} > B_{\text{max},\text{upt}} \end{cases}
\]

\[
\frac{\partial B_{\text{N,rel}}}{\partial t} = \begin{cases} 
B_{\text{rel}} \left( B_{\text{rel}} - B_{\text{max},\text{rel}} \right) B_{\text{C}} & \text{if } B_{\text{rel}} < B_{\text{max},\text{rel}} \\
0 & \text{if } B_{\text{rel}} = B_{\text{max},\text{rel}} \end{cases}
\]

Predation on bacteria occurs only by heterotrophic flagellates and is given by:

\[
\frac{\partial B_{\text{C,P,F,N}}}{\partial t} = \begin{cases} 
B_{\text{F}} \left( B_{\text{C,P,F,N}} - B_{\text{C,P,F,N}} \right) B_{\text{C}} & \text{if } B_{\text{C,P,F,N}} < B_{\text{C,P,F,N}} \\
0 & \text{if } B_{\text{C,P,F,N}} = B_{\text{C,P,F,N}} \end{cases}
\]

The balance equations for bacteria here are mostly identical to the previous formulation (Eqs. 43 and 44) with the addition of the release of recalcitrant carbon:

\[
\frac{\partial B_{\text{C}}}{\partial t} = \frac{\partial B_{\text{C}}}{\partial t} - \frac{\partial B_{\text{C}}}{\partial t} - \frac{\partial B_{\text{C}}}{\partial t} - \frac{\partial B_{\text{C}}}{\partial t}
\]

\[
\frac{\partial B_{\text{N,P}}}{\partial t} = \frac{\partial B_{\text{N,P}}}{\partial t} - \frac{\partial B_{\text{N,P}}}{\partial t} - \frac{\partial B_{\text{N,P}}}{\partial t} - \frac{\partial B_{\text{N,P}}}{\partial t}
\]

and an alternative formulation of uptake as in this formulation bacteria feed on all forms of particulate and dissolved organic matter:

\[
R_{\text{C,P,N}} = R_{\text{C,P,N}} + q_{\text{lab}} R_{\text{C,P,N}} + q_{\text{lab}} R_{\text{C,P,N}} + q_{\text{lab}} R_{\text{C,P,N}} + q_{\text{lab}} R_{\text{C,P,N}} + q_{\text{lab}} R_{\text{C,P,N}}
\]

\[
+ q_{\text{med}} R_{\text{C,P,N}} + q_{\text{med}} R_{\text{C,P,N}} + q_{\text{med}} R_{\text{C,P,N}} + q_{\text{med}} R_{\text{C,P,N}} + q_{\text{med}} R_{\text{C,P,N}}.
\]

The parameters \( q_{\text{M}} \) are non-dimensional turn-over rates relative to \( R_{\text{C,P,N}} \) turn-over, leading to the following equations for substrate specific and absolute uptake:

\[
S_{\text{upt}} = \min \left( B_{\text{upt}} \frac{B_{\text{C}}}{B_{\text{upt}}} \right)
\]

\[
\frac{\partial B_{\text{C,N,P}}}{\partial t} = \frac{\partial B_{\text{C,N,P}}}{\partial t} - \frac{\partial B_{\text{C,N,P}}}{\partial t} - \frac{\partial B_{\text{C,N,P}}}{\partial t} - \frac{\partial B_{\text{C,N,P}}}{\partial t}
\]

In this case carbon uptake is not nutrient limited as the internal stoichiometric quota of bacteria is balanced directly through the regulating fluxes releasing carbon into semi-labile organic matter.

The release of recalcitrant carbon in the form of capsular semi-refractory material is assumed proportional by a factor of \( q_{\text{refr}} \) to the activity respiration representing the metabolic cost of the uptake activity:

\[
\frac{\partial B_{\text{C}}}{\partial t} = \frac{\partial B_{\text{C}}}{\partial t} \left( 0, \max \left( 1 - q_{\text{C,F}}, 0 \right) \right) B_{\text{C}}
\]

\[
+ q_{\text{refr}} \left( 1 - \frac{B_{\text{C}}}{B_{\text{C}}} \right) \frac{1 - \frac{B_{\text{C}}}{B_{\text{C}}}}{1 - \frac{B_{\text{C}}}{B_{\text{C}}}} \frac{\partial B_{\text{C}}}{\partial t}.
\]

3.3.2 Dynamic decomposition version

In this version, activated with the DOCDYN preprocessing definition, the decomposition of particulate organic matter is directly mediated by bacteria and the partition between labile dissolved organic matter and dissolver matter with longer degradation time scales (including the additional state of semi-refractory carbon) occurs in relation to the nutritional status of bacteria as opposed to the fixed, parametric decomposition and partition of particles in the standard model. (see also the following sections on the fluxes of particulate and dissolved organic matter (Sects. 3.5.3.4). The formulation includes the bacteria mediated production of recalcitrant DOC (Hansell et al., 2003) and therefore provides the conceptual framework for an implementation of the microbial carbon pump (Jiao et al., 2014; 2010). However, the fractions of recalcitrant DOC with long turnover time (\( \gg 1 \text{ year} \)) are not considered in the current formulation. The sub-model is an extended version of the formulation in Polimene et al. (2006; 2007).

3.4 Particulate organic matter

The particulate matter \( \chi \) (\( \chi = \text{small, medium or large} \)) fluxes resulting from the above processes are composed of excretion and mortality inputs and decomposition and scavenging losses (for medium size particulate matter only) complemented by inputs resulting from mesozooplankton regulation of the internal stoichiometric ratio for large particulate
Figure 4. The microbial cycling of organic material for the standard bacteria model (left) and the dynamic decomposition model (right).

matter. As the consumer types for simplicity do not include an internal component for iron or silicate, the corresponding component fluxes resulting from predation are directed to particulate matter as indirect excretion.

\[
\frac{\partial \chi_{RC}}{\partial t} = \frac{\partial \chi_{RC}}{\partial t}_{bgc} + \frac{\partial \chi_{RC}}{\partial t}_{excr} - \frac{\partial \chi_{RC}}{\partial t}_{decomp}
\]

Only the excretion by zooplankton (Eq. 34) results in particulate matter by a fraction of \(1 - q_{dloss}\), while mortality of phytoplankton (Eqs. 12 and 20) and zooplankton (Eqs. 37 and 43) both have a particulate component (\(Q_{pmort}\) or \(1 - q_{dloss}\) respectively):

\[
\frac{\partial \chi_{RC,N,P}}{\partial t} = \frac{\partial \chi_{RC,N,P}}{\partial t}_{bgc} + \frac{\partial \chi_{RC,N,P}}{\partial t}_{excr} - \frac{\partial \chi_{RC,N,P}}{\partial t}_{decomp}
\]

where \( \text{lab}_{pcyt_{N,P}} \) reflects the relative nitrogen or phosphorus content of cytoplasm with respect to the structural components assuming that the dissolved losses of zooplankton through excretion are largely of cytoplasm origin and \(q_{dloss}\) is the dissolved fraction of zooplankton losses. The partition of phytoplankton lysis for each functional type is given as

\[
Q_{pmort} = \min \left( \frac{\chi_{q_{min_{N}}}}{\chi}, \frac{\chi_{q_{min_{P}}}}{\chi} \right)
\]

The size classes of particulate organic matter \( \chi \) in these equations originate from the phytoplankton types \( P \) and zooplankton types \( Z \) as given in Table 2.

Scavenging of mesozooplankton on medium size particulate organic matter results from Eq. (31):

\[
\frac{\partial R_{C,N,P,F,S}}{\partial t}_{\text{scav}} = \frac{F}{MESO} \frac{Z}{Z_{med}} R_{med}
\]

Additional large particulate organic matter may result from the mesozooplankton release flux

\[
\frac{\partial R_{C,N,P,F,S}}{\partial t}_{\text{rel}} = \frac{MESO}{MESO} R_{C,N,P,F,S}
\]
Table 2. Particulate organic matter and its origin.

| POM type                        | Originating from                                      |
|---------------------------------|------------------------------------------------------|
| Small particulate organic matter (R) | Nano- and picophytoplankton (P),\(\text{HET}\) \(\text{nano}\) micro\(\text{dia}\) microzooplankton (Z) |
| Medium size particulate organic matter (R) | Microphytoplankton and diatoms (P),\(\text{MICRO}\) \(\text{med}\) \(\text{mesozooplankton}\) (Z) |
| Large particulate organic matter (R) | Mesozooplankton (Z) \(\text{large}\) |

The decomposition of particulate matter is dependent on the bacteria sub-model applied. In case of the standard bacteria model (Sect. 3.3.1) it is converted to dissolved organic matter proportionally to the amount of substrate available by the rate \(\lambda_{\text{decomp}}\) and modified by the nutritional status of the substrate in relation to the Redfield Ratio \(q_{\text{ref:C:N}}\):

\[
\frac{\partial \chi_{\text{C},N,P,F}}{\partial t}_{\text{decomp}} = q_{\text{ref:C:N}} \frac{\chi_{\text{R}}}{\lambda_{\text{decomp}}} R_{\text{C},P,N,F}.
\]  

For the model with dynamic decomposition (Sect. 3.3.2) directly mediated by bacteria, the decomposition fluxes are given by the bacterial uptake resulting from Eqs. (55), (56), (57) as:

\[
\frac{\partial \chi_{\text{C},N,P,F}}{\partial t}_{\text{decomp}} = -S_{\text{growth}} \frac{\chi_{\text{R}}}{\lambda_{\text{M}}} R_{\text{C},P,N,F}.
\]  

The iron and silicate component of phytoplankton taken up by zooplankton in Eqs. (21) and (25) are for simplicity directly converted to particulate matter:

\[
\frac{\partial \chi_{\text{E}}}{\partial t}_{\text{excr}} = \sum_{\psi,\Psi} F_{\text{Z}}^{\chi_{\text{E}}}_{P_{\Psi}} P_{\Psi}.
\]  

\[
\frac{\partial \chi_{\text{S}}}{\partial t}_{\text{excr}} = \sum_{\psi,\Psi} F_{\text{Z}}^{\chi_{\text{S}}}_{P_{\Psi}} P_{\Psi}.
\]  

In the case of silicate the particulate organic matter types are determined by the predator that ingested the prey and directly releases the silicate contained in the frustule. They are consequently distributed analogous to the zooplankton excretion:

- Small particulate organic matter \(\text{small}\) \(\text{HET}\) \(\text{HET}\) (Z): heterotrophic flagellates (Z),
- Medium size particulate organic matter \(\text{med}\) \(\text{MICRO}\) \(\text{MICRO}\) (Z): microzooplankton (Z),
- Large particulate organic matter \(\text{large}\) \(\text{MESO}\) \(\text{MESO}\) (Z): mesozooplankton (Z).

For iron, on the contrary, the size of particulate iron is given by the prey size class and taken analogous to phytoplankton lysis reflecting the assimilation of iron into the cytoplasm:

- Small particulate organic matter \(\text{small}\) \(\text{nano}\) \(\text{nano}\) (P): nano- and pico-phytoplankton (P),
- Medium size particulate organic matter \(\text{med}\) \(\text{micro}\) \(\text{micro}\) (P): microphytoplankton and diatoms (P),
- Large particulate organic matter \(\text{large}\) \(\text{large}\) (Z): none.

3.5 Dissolved organic matter

The partition of labile dissolved, semi-labile and semi-refractory carbon originating from bacteria substantially differs in between the standard bacteria model (Sect. 3.3.1) and the bacteria model with dynamic decomposition (Sect. 3.3.2).

For the standard bacteria model the fluxes of dissolved organic matter are affected by uptake, excretion, mortality, decomposition and remineralisation:

\[
\frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{lab}} = \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{bgc}} + \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{excr}} + \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{mort}} + \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{decomp}} - \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{upt}} - \frac{\partial \chi_{\text{C},N,P}}{\partial t}_{\text{rem}}.
\]  

\[
\frac{\partial \chi_{\text{C}}}{\partial t}_{\text{lab}} = \frac{\partial \chi_{\text{C}}}{\partial t}_{\text{bgc}} + \frac{\partial \chi_{\text{C}}}{\partial t}_{\text{excr}} + \frac{\partial \chi_{\text{C}}}{\partial t}_{\text{mort}} - \frac{\partial \chi_{\text{C}}}{\partial t}_{\text{decomp}}.
\]  

The losses of bacteria, phytoplankton and zooplankton in dissolved carbon are fractionated at a constant quota \(q_{\text{dis}}\) in between labile and semi-labile DOC. Excretion towards the
dissolved forms of organic matter may originate from phytoplankton (Eq. 11), or zooplankton (Eq. 34):

\[
\frac{\partial R_C}{\partial t}_{\text{lab}} = q_{\text{dis}} \left( \sum \psi \frac{\partial P_C}{\partial t} \right)_{\text{excr}} + \sum \frac{\partial Z_C}{\partial t} \psi_{dloss} \right)_{\text{excr}} \\
\frac{\partial R_C}{\partial t}_{\text{lab}} = (1 - q_{\text{dis}}) \left( \sum \psi \frac{\partial P_C}{\partial t} \right)_{\text{excr}} + \sum \frac{\partial Z_C}{\partial t} \psi_{dloss} \right)_{\text{excr}} \right)_{\text{lab}} + \frac{\partial R_C}{\partial t}_{\text{lab}}_{\text{N.P}} = \frac{\partial R_C}{\partial t}_{\text{lab}}_{\text{N.P}} - \frac{\partial R_C}{\partial t}_{\text{lab}}_{\text{N.P}} = \frac{\partial R_C}{\partial t}_{\text{lab}} \left|_{\text{upt}} \right| - \frac{\partial R_C}{\partial t}_{\text{lab}} \left|_{\text{rem}} \right|
\]

where \( \psi_{dloss} \) is the dissolved fraction of the zooplankton losses.

Mortality input may originate from all three trophic levels (Eqs. [48], [13], [57], [42]):

\[
\frac{\partial R_C}{\partial t}_{\text{lab}} = q_{\text{lab}} \left( \frac{\partial B_C}{\partial t} \right)_{\text{lab}} + \sum \psi \left( 1 - Q_{\text{pmort}} \right) \frac{\partial P_C}{\partial t} \left|_{\text{mort}} \right) + \sum \psi \left( Q_{\text{pmort}} \right) \frac{\partial Z_C}{\partial t} \left|_{\text{mort}} \right)
\]

In addition, the decomposition of the particulate matter types (\( R : \Psi \) = small, medium or large, Eq. [67]) and of semi-labile dissolved organic carbon \( R_C \) is directly converted to labile dissolved organic matter \( R_C \) according to

\[
\frac{\partial R_C}{\partial t}_{\text{slab}} = (1 - q_{\text{lab}}) \left( \frac{\partial B_C}{\partial t} \right)_{\text{lab}} + \sum \psi \left( 1 - Q_{\text{pmort}} \right) \frac{\partial P_C}{\partial t} \left|_{\text{mort}} \right) + \sum \psi \left( Q_{\text{pmort}} \right) \frac{\partial Z_C}{\partial t} \left|_{\text{mort}} \right)
\]

Here, the fractionation of dissolved organic matter originating from bacteria and phytoplankton is based on the originating process. This reflects the capacity of bacteria to utilise different forms of substrate with lysis/mortality contributing to the labile DOM pool, while excretion of carbon occurs in semi-labile form, and discarding the less digestible forms adding semi-refractory organic matter to the set of state variables. Zooplankton losses are treated identically with respect to the standard bacteria model.

Excretion of DOC may originate from the phyto- and zooplankton excretion (Eqs. [11] and [34], the regulation of the bacterial stoichiometric quota (Eq. [58]) and excess bacterial
growth:

\[
\frac{\partial R_C}{\partial t} = \sum \psi q_{diss} \frac{\partial C}{\partial t} \bigg|_{excr} + \sum \psi \frac{\partial P_C}{\partial t} \bigg|_{excr} + \sum \psi (1 - q_{diss}) q_{diss} \frac{\partial Z_C}{\partial t} \bigg|_{excr} \]

(81)

while the non-particulate part of mortality/lysis is split according to:

\[
\frac{\partial R_C}{\partial t} \bigg|_{growth} = \frac{\partial B_C}{\partial t} \bigg|_{rel} + \sum \psi \frac{\partial P_C}{\partial t} \bigg|_{excr} + \sum \psi (1 - q_{diss}) q_{diss} \frac{\partial Z_C}{\partial t} \bigg|_{excr} \]

(82)

\[
\frac{\partial R_C}{\partial t} \bigg|_{calc} = \frac{\partial B_C}{\partial t} \bigg|_{rel} + \sum \psi \frac{\partial P_C}{\partial t} \bigg|_{excr} + \sum \psi (1 - q_{diss}) q_{diss} \frac{\partial Z_C}{\partial t} \bigg|_{excr} \]

(85)

Uptake of labile dissolved matter by bacteria is given by

\[
\frac{\partial R_C}{\partial t} \bigg|_{upt} = S_{growth} R'_{C,N,P} \]

(86)

where the substrate mass specific uptake of bacteria \( S_{growth} \) is given in Eq. (46) for the standard decomposition model and in Eq. (56) for the dynamic decomposition model.

The remaining terms are identical for both decomposition sub-models. Excretion and mortality of nitrogen and phosphorus result in the dissolved fluxes

\[
\frac{\partial R_{N,P}}{\partial t} \bigg|_{excr} = \sum \psi q_{diss} P_{cyt} \Psi \frac{\partial N}{\partial t} \bigg|_{excr} + \sum \psi q_{diss} q_{diss} \frac{\partial P_{N,P}}{\partial t} \bigg|_{excr} \]

(87)

\[
\frac{\partial R_{N,P}}{\partial t} \bigg|_{mort} = \frac{\partial B_{N,P}}{\partial t} \bigg|_{mort} + \sum \psi (1 - q_{p,mort}) q_{p,mort} \frac{\partial P_{N,P}}{\partial t} \bigg|_{mort} + \sum \psi q_{diss} q_{diss} \frac{\partial Z_{N,P}}{\partial t} \bigg|_{mort} \]

(88)

Remineralisation of dissolved organic nutrients into inorganic form is given by fixed mass specific remineralisation rates \( r_{rem,N,P} \):

\[
\frac{\partial R_{N,P}}{\partial t} \bigg|_{rem} = r_{rem,N,P} \frac{\partial N}{\partial t} \bigg|_{rem} \]

(89)

### 3.6 Calcification

The model in its current form does not include calcifiers as a dedicated functional group given the limited knowledge of the physiological constraint of calcification. Therefore, the process of calcification is not directly modelled, but is treated implicitly by considering part of the nanophytoplankton to act as calcifiers. Calcification processes are inferred from the system dynamics based on the assumption of a given ratio between particulate inorganic carbon over particulate organic carbon in sedimenting material, usually referred to as rain-ratio. Here this ratio is used as a proxy for the calcite production matching the local increase of POC originating from nanophytoplankton. Since the rain ratio has been defined for the sinking fluxes and calcite is the more resistant mineral, we limit the description to calcite in this part of the model, neglecting aragonite. This approach is similar to the implementations in other biogeochemical models, e.g. PISCES (Gehlen et al., 2007) or MEDUSA (Yool et al., 2013).

In this context the local rain-ratio is based on a reference ratio \( q_{\text{rain0}} \) that varies according to the regulating factors \( l_C \), \( l_T \) and \( l_{NP} \) given in Eqs. (250) or (252), (254) and (255):

\[
q_{\text{rain}} = \max \left( \frac{1}{200} q_{\text{rain}0} \frac{\text{calc}}{\text{calc}} \frac{\text{calc}}{\text{calc}} \frac{\text{calc}}{\text{calc}} \right) \]

(90)

The calcite dynamics are then described by the equation:

\[
\frac{\partial L_C}{\partial t} \bigg|_{hgc} = \frac{\partial L_C}{\partial t} \bigg|_{mort} + \frac{\partial L_C}{\partial t} \bigg|_{pred} + \frac{\partial L_C}{\partial t} \bigg|_{sed} - \frac{\partial L_C}{\partial t} \bigg|_{dis} \]

(91)

The contribution of nanophytoplankton lysis to calcite production is proportional to the particulate fraction of lysis.
Ingestion of nanophytoplankton and subsequent dissolution in zooplankton guts contributes with a fraction \( q_{\text{gut diss}} \) of the excreted part of nanophytoplankton uptake by the various zooplankton groups (compare Eqs. 14 and 34):

\[
\frac{\partial L_{\text{calc}}}{\partial t} = q_{\text{rain}} q_{\text{gut diss}} (1 - q_{\text{eff}}) \hat{q}_{\text{excr}} \sum \psi P_{\text{calc}} \frac{\partial P_{\text{calc}}}{\partial t} \tag{93}
\]

As sedimentation of nanophytoplankton contributes to the organic carbon considered in the rain-ratio the matching contribution to calcite production is computed as

\[
\frac{\partial L_{\text{calc}}}{\partial t} = q_{\text{rain}} q_{\text{gut diss}} P_{\text{calc}} \frac{\partial P_{\text{calc}}}{\partial t} \tag{94}
\]

with the sinking rate \( \frac{\partial P_{\text{calc}}}{\partial t} \) given in Eq. 138.

Dissolution of calcite is proportional to the current concentration of calcite with a maximum rate of \( q_{\text{dis}} \), regulated by \( l_{\text{calc}} \) (Eqs. 251 or 253):

\[
\frac{\partial L_{\text{calc}}}{\partial t} = q_{\text{dis}} \frac{\partial L_{\text{calc}}}{\partial t} \tag{95}
\]

Note, that while the calcification rates are implicitly derived from the rain-ratio and not directly modelled processes, this formulation is still conservative as all sources and sinks of calcite are balanced by dissolved inorganic carbon (DIC).

\[ \text{DIC} \]

\[ \text{calc} \]

The solution of the calcite dynamics is optional and activated by the preprocessing switch \( \text{CALC} \).

### 3.7 Inorganic components

The dynamics of dissolved inorganic nutrients in the model are given by uptake of phytoplankton and bacteria and are re-supplied locally by remineralisation and excretion. Dissolved inorganic iron is additionally subject to scavenging.

\[
\frac{\partial N_{\text{ox}}}{\partial t} = \psi \left( \frac{\partial N_{\text{ox}}}{\partial t} \right)_{\text{upt}} - \psi \left( \frac{\partial N_{\text{ox}}}{\partial t} \right)_{\text{scav}} \tag{100}
\]

Oxidised nitrogen in the water-column is taken up only by the four phytoplankton types \( P \) following Eq. (15) according to external availability:

\[
\frac{\partial N_{\text{ox}}}{\partial t} = \sum \psi \left( \frac{\partial N_{\text{ox}}}{\partial t} \right)_{\text{upt}} \tag{101}
\]

and regenerated exclusively by nitrification:

\[
\frac{\partial N_{\text{ox}}}{\partial t} = B_{\text{upt}} l_{\text{calc}} l_{\text{N}} l_{\text{PH}} N_{\text{ox}} \tag{102}
\]

where \( B_{\text{nitr}} \) is the maximum ammonium mass specific nitrification rate at reference temperature. In the absence of explicit nitrifiers, nitrification is modelled as an implicit process depending on multiple environmental factors, based on temperature, oxygen and availability ammonium taking into account the poor competitiveness of nitrifying microbes with respect to other pelagic consumers of ammonium (\text{Ward} 2008). The various regulation and limitation factors \( l_{T}, l_{O}, l_{N} \) and \( l_{\text{PH}} \) are given in Sect. 6.1.

Ammonium is taken up by phytoplankton as the reduced part of total nitrogen uptake (Eq. 15) and bacteria when ni-
Ammonium is released by the phytoplankton types $\psi$ (Eq. 15) when respiration exceeds photosynthesis or when above their optimal quota

$$\frac{\partial N_{N_{\psi}}}{\partial t}_{\text{upt}} = \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{N'_{N_{\psi}}}{N'_{N_{\psi}} + r_{\text{aff}_{\psi}}} \left( \frac{\partial P_{N}}{\partial t} \right)_{\text{upt}} + \frac{\partial B_{N}}{\partial t} \bigg|_{\text{upt}}$$

(103)

and remineralised according to Eq. 89

$$\frac{\partial N_{N_{\psi}}}{\partial t}_{\text{rem}} = r_{\text{rem}_{\psi}} \frac{R_{N}}{R'_{N}}.$$  

(104)

Ammonium concentrations may be further reduced by nitrification:

$$\frac{\partial N_{N_{\psi}}}{\partial t}_{\text{nitr}} = B_{\text{nitr}} \frac{B}{l_{\text{am}} l_{\text{ntr}} l_{\text{m}} l_{\text{ph}} N'_{N_{\psi}}}.$$  

(106)

Phosphorus dynamics are analogous to nitrogen dynamics but simplified with only one dissolved inorganic pool being considered in the model. It is taken up according to Eqs. 15 and 50

$$\frac{\partial N_{P}}{\partial t}_{\text{uptake}} = \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial P_{N}}{\partial t} \bigg|_{\text{upt}} + \frac{\partial B_{P}}{\partial t} \bigg|_{\text{upt}},$$

(107)

released following Eqs. 15, 50, 36 and 37

$$\frac{\partial N_{P}}{\partial t}_{\text{rel}} = \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial Z_{P}}{\partial t} \bigg|_{\text{rel}} - \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial P_{N}}{\partial t} \bigg|_{\text{rel}} + \frac{\partial B_{P}}{\partial t} \bigg|_{\text{rel}},$$

(108)

and remineralised as given in Eq. 51

$$\frac{\partial N_{P}}{\partial t}_{\text{rem}} = r_{\text{rem}_{\psi}} \frac{R'_{P}}{R_{P}}.$$  

(109)

Iron is taken up only by phytoplankton (Eq. 15)

$$\frac{\partial N_{F}}{\partial t} \bigg|_{\text{upt}} = \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial P_{F}}{\partial t} \bigg|_{\text{upt}},$$

(110)

and subject to scavenging due to hydroxide, treated similarly as in Aumont et al. 2003 and Vichi et al. 2007:

$$\frac{\partial N_{F}}{\partial t}_{\text{scav}} = r_{\text{Fscav}} \max \left( 0, N'_{F} \right),$$

(111)

where $r_{\text{Fscav}}$ is a threshold concentration over which scavenging occurs, here fixed at 0.6 $\mu$mol m$^{-2}$.

Iron is released by phytoplankton (Eq. 15)

$$\frac{\partial N_{F}}{\partial t} \bigg|_{\text{rel}} = \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial P_{F}}{\partial t} \bigg|_{\text{rel}},$$

(112)

and implicitly remineralised by mesozooplankton scavenging of particulate organic matter (Eq. 66) and bacterial consumption of particulate matter (Eqs. 67 and 68)

$$\frac{\partial N_{F}}{\partial t}_{\text{rem}} = \frac{F_{\text{ZMESO}}}{R} \frac{\text{med}}{R'_{F}} + \frac{\partial \overline{R_{F}}}{\partial t}_{\text{decomp}},$$

(113)

It is assumed here that the feeding activity of scavenging zooplankton increases the bio-availability and accelerates the decomposition of particulate iron.

Silicate is taken up

$$\frac{\partial N_{S}}{\partial t}_{\text{uptake}} = \text{dia} \frac{q_{\text{ref}_{\text{S}}}}{\text{dia}} S_{\text{growth}},$$

(114)

and released

$$\frac{\partial N_{S}}{\partial t}_{\text{rel}} = \text{dia} \frac{P'_{S}}{\text{dia}} - \text{dia} \frac{P_{S}}{\text{ref}_{\text{S}}}$$

(115)

exclusively by diatoms (Eq. 22). It is not remineralised in the pelagic part of the system.

This neglection of silicate conversion into inorganic form in the water column is based on observations that the recycling of this element in particulate form while sinking down the water column is much lower than for the other nutrients, such that most of its remineralisation is confined to the seafloor (Broecker and Peng 1982; Dugdale et al. 1995).

The dynamics of DIC are given by photosynthesis and respiration of the organisms considered and calcification and dissolution of calcite:

$$\frac{\partial C_{\text{DIC}}}{\partial t}_{\text{bgc}} = \frac{\partial B_{C}}{\partial t}_{\text{resp}} + \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial P_{C}}{\partial t} + \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial Z_{C}}{\partial t} - \sum_{\psi} \frac{\psi}{r_{\text{aff}_{\psi}}} \frac{\partial L_{C}}{\partial t} - \frac{\partial L_{C}}{\partial t}_{\text{calc}} + \frac{\partial L_{C}}{\partial t}_{\text{calc}},$$

(116)

where the respiration terms $\frac{\partial B_{C}}{\partial t}_{\text{resp}}$, $\frac{\partial P_{C}}{\partial t}_{\text{resp}}$, $\frac{\partial Z_{C}}{\partial t}_{\text{resp}}$ and $\frac{\partial L_{C}}{\partial t}_{\text{calc}}$ are given in Eqs. 49, 12, 55 and 42. Synthesis of carbon is given in Eq. 8, the dissolution of calcite is given in Eq. 95 and precipitation of DIC into calcite is given by the sum of the calcification terms

$$\frac{\partial L_{C}}{\partial t}_{\text{calc}} = \frac{\partial L_{C}}{\partial t}_{\text{calc}} + \frac{\partial L_{C}}{\partial t}_{\text{calc}} + \frac{\partial L_{C}}{\partial t}_{\text{calc}},$$

(117)

given in Eqs. 92, 93 and 94.
Rates of change of oxygen are implied from the corresponding carbon fluxes converted by stoichiometric factors taking into account different efficiencies for respiration $O_{\text{resp}}$ and photosynthesis $O_{\text{gpp}}$.

The pelagic oxygen cycle is reduced to the consumption of dissolved oxygen in respiration (Eqs. 49, 12, 35 and 42) and the production of dissolved oxygen in photosynthesis (Eq. 8):

$$
\frac{\partial O_{\text{gpp}}}{\partial t}_{\text{bge}} = -p_O \frac{\partial B_C}{\partial t}_{\text{resp}} - p_O \sum_{\psi} \frac{\partial P_C}{\partial t}_{\text{resp}} - p_O \sum_{\psi} \frac{\partial P_C}{\partial t}_{\text{gpp}}.
$$

3.8 The carbonate system

The model for the carbonate system incorporated in ERSEM was introduced in Blackford and Burkall (2002) and further developed in Blackford and Gilbert (2007); Artioli et al. (2012). In this model, the speciation of carbon is calculated from dissolved inorganic carbon $O_C$, total alkalinity $A_{\text{tot}}$ (which can be computed diagnostically, semi-diagnostically or prognostically, see below) and total boron $B_{\text{tot}}$ (which is calculated from a linear regression of salinity). It assumes chemical equilibrium between the inorganic carbon species justified by the fast reaction time scales of the underlying chemical reaction compared to the biological and physical rates on the spatial scales the model operates on. The comprehensive set of equations to describe the carbonate system and ways to solve it have been given in various references. The model requires the complete set of known quantities to be used in the model: $O_C$, $A_{\text{tot}}$, $B_{\text{tot}}$, partial pressure of carbon dioxide $p_{CO_2}$, carbonic acid, carbonate and bicarbonate concentrations ($c[H_2CO_3]$, $c[CO_2]$ and $c[HCO_3^-]$) and pH (using the seawater scale) at chemical equilibrium. The model utilises the four equilibrium constants for saturation of carbon dioxide and for the dissociation of carbonic acid, bicarbonate and boric acid derived from empirical environmental relationships (Millero, 1995; Mehrbach et al., 1973; Weiss, 1974; Dickson, 1990) that are detailed in the Supplement for reference. The resulting set of equations to solve is then given by:

$$
O_C = c[CO_2^-] + c[HCO_3^-] + c[CO_3^{2-}] \quad (118)
$$

$$
A_{\text{tot}} = c[HCO_3^-] + 2c[CO_3^{2-}] + c[B(OH)_3^-] \quad (119)
$$

$$
B_{\text{tot}} = c[B(OH)_3] + c[B(OH)_4^-] \quad (120)
$$

$$
c[B(OH)_3] = \frac{c[H^+]c[B(OH)_3^*]}{k_b} \quad (121)
$$

$$
c[CO_2^-] = \frac{c[H^+]c[HCO_3^-]}{k_1} \quad (122)
$$

$$
c[HCO_3^-] = \frac{c[H^+]c[CO_3^{2-}]}{k_2} \quad (123)
$$

$$
pH = -\log_{10}(c[H^+]) \quad (124)
$$

$$
p_{CO_2} = k_o c[CO_2] \quad (125)
$$

The system is solved using the HALTAFALL algorithm (Ingri et al., 1967) by using the equilibrium relations 121 to 123 to eliminate the unknowns $c[B(OH)_3^*]$, $c[CO_2^-]$ and $c[HCO_3^-]$. The balance equations for DIC and total boron are then used to express $c[CO_2^-]$ and $c[B(OH)_3^*]$ in the balance equation for alkalinity (Eq. 119) as functions of the only remaining unknown $c[H^+]$. This equation is solved for the logarithm of the unknown variable (allowing only positive real numbers as solutions) applying a combination of the bisection method to narrow down the solution to a sufficiently small interval in $c[H^+]$ to permit linear approximation followed by the bisection method reducing the solution residual to the desired tolerance.

Calcite saturation is computed from the product of calcium and carbonate concentrations ($c[Ca^{2+}]$ and $c[CO_3^{2-}]$) divided by their product in chemical equilibrium $k_{calc}$

$$
\Omega_{calc} = \frac{c[Ca^{2+}]c[CO_3^{2-}]}{k_{calc}} \quad (126)
$$

The variability of this ratio is dominated by $c[CO_3^{2-}]$ as $c[Ca^{2+}]$ is nearly constant in sea water (Kleypas et al., 1999) and therefore fixed in the model at the oceanic mean value of 0.01028 mol kg$^{-1}$.

Similarly, the aragonite saturation state is determined by the equation

$$
\Omega_{calc} = \frac{c[Ca^{2+}]c[CO_3^{2-}]}{k_{arag}} \quad (127)
$$

Two different modes to compute total alkalinity are provided with the model:

- A diagnostic mode, that computes alkalinity from salinity or salinity and temperature. This mode is non conservative and the field of alkalinity is recomputed at each time step without physical transport. It does not include changes to alkalinity by the biogeochemical processes of the model.
A prognostic model, that includes biogeochemical changes to alkalinity. It is fully conservative and adds a state variable for alkalinity that is subject to physical transport.

As a third semi-diagnostic option, these two modes can be combined as a sum by setting the prognostic alkalinity state to 0, so that the diagnostic mode provides the background field and the prognostic mode gives a trace of the contribution of biogeochemical processes to the total alkalinity.

The recommended option is the semi-diagnostic option for coastal applications and shelf seas, where reliable and robust regressions exist or the fully prognostic mode, where no single reliable regression is available, e.g. in global simulations. (For further detail the reader is referred to Artioli et al., 2012)

The changes of alkalinity due to biological processes are accompanied by the effect of riverine inputs (see Artioli et al., 2012)

In three dimensional simulations, these changes are accompanied by the effect of riverine inputs (see Artioli et al., 2012)

The different variants of alkalinity regressions available from the scientific literature (Borges and Frankignoulle, 1999; Bellerby et al., 2005; Millero et al., 1998; Lee et al., 2006), the total boron regression and the empirical equilibrium constants k are given in the Supplement.

3.9 Light extinction

Light in the water column is attenuated according to the Beer–Lambert formulation computing PAR as:

$$E_{\text{PAR}} = q_{\text{PAR}} I_{\text{surf}} e^{\int K_d(\xi) d\xi},$$

(130)

where $I_{\text{surf}}$ is the short-wave radiation at sea-surface level, $q_{\text{PAR}}$ is a parameter for the photosynthetically active fraction and $K_d$ is the spatially varying attenuation coefficient. The latter incorporates light attenuation by the modelled living and non-living optically active components as well as background extinction due to clear sea-water and other components not explicitly modelled. Two alternative models are available for the computation of $K_d$:

1. a model based on mass specific attenuation coefficients for the relevant functional types, non-modelled forms of inorganic matter and the background attenuation of clear sea-water; this model is used in previous ERSEM versions (Blackford et al., 2004) and is the default choice,

2. a model based on broadband inherent optical properties (absorption and backscatter), activated by the preprocessing definition IOPMODEL.

For the default model based on specific attenuation coefficients $K_d$ is computed according to:

$$K_d = \sum_\chi \lambda_{\chi, p} \tilde{P}_C + \sum_\Psi \lambda_\Psi \tilde{R}_C + \lambda_{\text{susp}} R_{\text{susp}} + \Lambda_{\text{sea}},$$

(131)

where the $\lambda$s are the specific attenuation coefficients of the optically active components, i.e. the phytoplankton types $\chi$ and the particulate organic matter types $\Psi$. $\Lambda_{\text{sea}}$ is the background attenuation of sea water and $R_{\text{susp}}$ is the concentration of non-modelled optically active substances, mostly suspended matter.

The model based on inherent optical properties (activated by the preprocessing switch IOPMODEL) uses the light attenuation model proposed in Lee et al. (2005):

$$K_d = (1 + 0.005\theta_{\text{zen}})a + 4.18 \left(1.0 - 0.52e^{-10.8a}\right) b_b,$$

(132)

where $\theta_{\text{zen}}$ is the zenith angle at the given time and location. Absorption $a$ and backscatter $b_b$ are composed as:

$$a = \sum_\chi a_{\chi, p} \tilde{P}_C + \sum_\Psi a_{\Psi, R} \tilde{R}_C + a_{\text{M susp}} + a_{\text{sea}},$$

(133)

$$b_b = \sum_\chi b_{\chi, p} \tilde{P}_C + \sum_\Psi b_{\Psi, R} \tilde{R}_C + b_b + b_{\text{sea}}$$

(134)

with $a^*$ and $b^*$ being the mass specific absorption and backscatter coefficients of the respective components, $a_{\text{sea}}$ and $b_{\text{sea}}$ being the broadband absorption and backscatter of clear sea-water, $a_{\text{M susp}}$ the constant absorption of non-modelled suspended matter and $b_b$ a constant amount of background backscatter in the water column.

In both optical models the attenuation of optically active matter that is not modelled by ERSEM ($R_{\text{susp}}$, mostly inorganic suspended particulate matter) can be provided homogeneously through a namelist parameter or spatially variable through the physical driver by filling and updating the ESS variable.

The combination of the attenuation of particulate organic matter and the non-modelled particles may be provided externally through the physical driver using the preprocessing definition ADYTRACER. This option introduces the state
variable \( a_{\text{ady}} \) and Eq. (131) reduces to

\[
K_d = \sum_{\chi} \rho_p \chi P_C + a_{\text{ady}} + a_{\text{sea}}, 
\tag{135}
\]

or in case of the model based on inherent optical properties

\[
a = \sum_{\chi} \rho_p \chi P_C + a_{\text{ady}} + a_{\text{sea}}, 
\tag{136}
\]

\[
b_b = \sum_{\chi} b_{b, k} P_C + b_{b, \text{sea}}, 
\tag{137}
\]

neglecting the backscatter component of particulate and non-modelled matter (see Eqs. 133 and 134).

The two models can be calibrated to give comparable results, but the latter formulation based on inherent properties has the advantage to be based on quantities that are frequently measured, which helps in constraining the parameterisation, validation and enables the direct assimilation of optical data.

3.10 Gravitational sinking

The sinking of model states is incorporated using a simple upwind scheme for the equation

\[
\frac{\partial c_p}{\partial t}_{\text{sed}} = \frac{c_p}{w_{\text{sed}}} \frac{\partial c_p}{\partial z} 
\tag{138}
\]

and adding the resulting rate to the biogeochemical rates that are passed to the physical driver for integration.

The sedimenting states in the model are given by the particulate organic types \( \chi \), phytoplankton types \( \chi P_C, P_N, P_F, P_S \) and calcite \( L_C \). Sinking velocities are constant velocities \( w_0 \) for each particulate matter type \( \psi \), while for the phytoplankton states \( \chi P_C \) they are composed of a constant velocity complemented by a variable component subject to nutrient limitation beyond the threshold \( \tilde{p}_{\text{sink}} \):

\[
\chi w_{\text{sed}} = w_0 + \chi \left( w_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{139}
\]

4 The benthic system

The benthic model in ERSEM is predicated on muddy sediments of the continental shelf, including zoobenthos, bacteria, different forms of organic matter and implicit vertical distribution of material within the sea-bed. It explicitly describes the main functions of the sediment such as benthic predation, decomposition and recycling of organic matter, bioirrigation and bioturbation. As an alternative to using a full benthic model, the benthic-pelagic interface can be described by a simple benthic closure given in Sect. 5.1.5

This scheme adsorbs depositing particulate matter and phytoplankton and returns dissolved inorganic nutrients and carbon to the water column at a given time scale reducing the sediments to a simple buffer layer of organic matter recycling, that however does not involve any explicit benthic processes. It is computationally considerably lighter compared to the full model, but the computational effort in both cases is negligible compared to the pelagic component. While the full benthic model is more adequate for shelf seas application that are dominated by the sediment type it represents with a close connection to the productive upper ocean, the simplified closure scheme is more suitable in deep domains under oligotrophic conditions, where the sediment processes are of lesser importance.

4.1 Benthic model structure

The full benthic model is a simplified version (Blackford, 1997; Kohlmeier, 2004) of the more complex original model introduced in the original version of ERSEM (Ruardj and Van Raaphorst, 1995; Ebenhöh et al., 1995) assuming near-equilibrium conditions for the inorganic components. Organisms are distinguished in classes on a more functional and less size oriented base than in the pelagic part.

The model includes the functional types of aerobic and anaerobic bacteria as decomposers of organic material, three types of benthic predators (suspension feeders, deposit feeders and meiobenthos), dissolved organic matter and three forms of particulate detritus classified according to their size, while for the phytoplankton states \( \chi P_C \) they are composed of a constant velocity complemented by a variable component subject to nutrient limitation beyond the threshold \( \tilde{p}_{\text{sink}} \):

\[
\chi w_{\text{sed}} = w_0 + \chi \left( w_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{139}
\]

\[
\chi w_0 = \left( \chi_{\text{ed}} + \chi_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{140}
\]

\[
\chi w_{\text{lim}} = \left( \chi_{\text{ed}} + \chi_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{141}
\]

4.1 Benthic model structure

The full benthic model is a simplified version (Blackford, 1997; Kohlmeier, 2004) of the more complex original model introduced in the original version of ERSEM (Ruardj and Van Raaphorst, 1995; Ebenhöh et al., 1995) assuming near-equilibrium conditions for the inorganic components. Organisms are distinguished in classes on a more functional and less size oriented base than in the pelagic part.

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\[
\chi w_{\text{sed}} = w_0 + \chi \left( w_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{139}
\]

\[
\chi w_0 = \left( \chi_{\text{ed}} + \chi_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{140}
\]

\[
\chi w_{\text{lim}} = \left( \chi_{\text{ed}} + \chi_{\text{lim}} \max \left( 0, \frac{\tilde{p}_{\text{sink}}}{\chi} \right) \right) 
\tag{141}
\]
Table 3. Benthic functional types and their components (squared brackets indicate option states) – chemical components: C carbon, N nitrogen, P phosphorus, F iron, S silicate.

| Symbol | Code | Description |
|--------|------|-------------|
| DEPO | Y2c | Deposit feeders |
| SUSP | Y3c | Suspension feeders |
| MEIO | Y4c | Meiobenthos |
| for | H1c | Aerobic bacteria |
| anaer | H2c | Anaerobic bacteria |
| diss | Q1c | Dissolved organic matter |
| degr | Q6c,n,p{f},s | Degradable organic matter |
| ref | Q7c,n,p,s | Refractory organic matter |
| burQ | Q17c,n,p | Buried organic matter |
| [bcalc | [bL2c | Calcite |
| GO | G2o | Dissolved oxygen |
| GC | G3c | Dissolved inorganic carbon (DIC) |
| GN | G4n | Dinitrogen |
| KP | K1p | Phosphate |
| KO | K2m | Oxidised nitrogen |
| KN | K3n | Ammonium |
| KS | K5s | Silicate |
| oxy | D1m | Depth of oxygen horizon |
| denit | D2m | Depth of oxidised nitrogen horizon |
| refCO | D3m | Average penetration depth of refractory carbon |
| refCN | D4m | Average penetration depth of refractory nitrogen |
| refCP | D5m | Average penetration depth of refractory phosphorus |
| degrC | D6m | Average penetration depth of degradable carbon |
| degrN | D7m | Average penetration depth of degradable nitrogen |
| degrP | D8m | Average penetration depth of degradable phosphorus |
| degrS | D9m | Average penetration depth of degradable silicate |

A complete list of benthic state variables is given in Tables.

4.2 Implicit vertical distribution of inorganic states in the benthos

In order to determine the dynamics of the oxygen and oxidised nitrogen horizons as well as the inorganic fluxes across the seabed (Sect. 5.1.3), the inorganic components of the benthos are assumed to be close to their equilibrium distributions, in which all source and sink terms of the pore water concentrations of the inorganic components $c_{pw}$ inside the
sediments are perfectly balanced by diffusion:

\[
\nu_{\text{diff}} \frac{\partial^2 c_{\text{pw}}}{\partial \zeta^2} = \frac{1}{\Delta d} \frac{\partial c_b}{\partial t} \bigg|_{\text{bgc}}
\]  

(140)

where \(c_b\) is the layer content. This partial differential equation has a general parabolic solution in \(\zeta\) taking the source-sink term \(\frac{\partial c_b}{\partial \zeta} \bigg|_{\text{bgc}}\) as a fixed equilibrium rate independent of time. This is a reasonable assumption when the diffusive rates are significantly faster than the biogeochemical processes (\(\nu_{\text{diff}}\) is the diffusivity of dissolved inorganic components in the benthos depending on bioirrigation, see Eq. 205). The equations apply to each of the three sediment layers and the resulting system of piece-wise parabolic continuous profiles can be solved using two boundary conditions per layer: the surface concentration at the upper boundary starting with the sediment surface concentration and the flux across the bottom boundary which is equal to the sum of all source and sink processes below the layer under consideration (by definition, no fluxes of dissolved matter can occur across the bottom of the sediments so that all sources and sinks have to be compensated from above).

The sediment surface concentration \(c_{\text{bed}}\) required as a boundary condition to the production-diffusion balance above is generally not equal to the concentration at the centre of the lowest pelagic discretisation cell \(c_p\), as diffusion across the sediment surface will be attenuated by the bottom boundary layer. In the simplest case the difference between cell centre and sediment surface concentrations can be estimated assuming a linear diffusive flux as positively proportional to the biogeochemical net change in the sediments. However, a problem arises for this formulation when the sediments act as net sink, as the calculated differences may exceed the cell centre concentration suggesting negative concentrations at the sediment interface. Therefore, for negative net sinks in the sediments the formulation suggested by Patankar (1980), Burchard et al. (2003) is applied, leading to the equation:

\[
c_{\text{bed}} = \begin{cases} 
    c_p + P_{\text{mix}} \frac{\partial c_b}{\partial t} \bigg|_{\text{bgc}} & \text{if } \frac{\partial c_b}{\partial t} \bigg|_{\text{bgc}} > 0 \\
    c_p - P_{\text{mix}} \frac{\partial c_b}{\partial t} \bigg|_{\text{bgc}} & \text{if } \frac{\partial c_b}{\partial t} \bigg|_{\text{bgc}} < 0
\end{cases}
\]  

(141)

where \(P_{\text{mix}}\) is an inverse mixing velocity constant.

The resulting equilibrium pore water concentrations \(c_{\text{pw}}\) in each layer are converted into the full equilibrium layer contents using the layer thickness and the conversion factor

\[
\nu_{\text{N,\%}} = P_{\text{poro}} P_{\text{ads}}
\]  

(142)

where \(P_{\text{poro}}\) and \(P_{\text{ads}}\) are porosity and adsorption factors that may vary spatially in case of porosity and adsorption of phosphorus while they are constants for all other adsorptions.

The dynamics of the oxygen and oxidised nitrogen horizons are determined by a relaxation towards their equilibrium values \(d_{\text{eq}}\) and \(d_{\text{eq}}\), which are the depths where the pore water equilibrium concentrations are 0. Their time evolution is then described by

\[
\frac{\partial D}{\partial t} = \frac{1}{\tau_{\text{oxy}}} \left( d_{\text{eq}} - D \right)
\]  

(143)

\[
\frac{\partial D}{\partial t} = \frac{1}{\tau_{\text{denit}}} \left( d_{\text{eq}} - D \right)
\]  

(144)

where \(\tau_{\text{oxy}}\) and \(\tau_{\text{denit}}\) are the respective relaxation time scales.

### 4.3 Implicit vertical distribution of organic matter in the benthos

The penetration of organic matter type \(\psi\) into the sediments is assumed as exponential decay of a concentration \(\tilde{c} (\zeta)\) from a sediment surface value \(c_0\) as a function of the e-folding depth \(\lambda\):

\[
\tilde{c} (\zeta) = c_0 e^{-\frac{\zeta}{\lambda}}.
\]  

(145)

Total content \(c_b\) is then given by the integral

\[
c_b = c_0 \int_0^{d_{\text{tot}}} e^{-\frac{\zeta}{\lambda}} d\zeta
\]  

(146)

and the penetration depth \(D\) of matter \(\psi\) is defined accordingly as

\[
D = \frac{\psi}{c_b} \int_0^{d_{\text{tot}}} \zeta e^{-\frac{\zeta}{\lambda}} d\zeta.
\]  

(147)

For \(d_{\text{tot}} \rightarrow \infty\) the two integrals of Eqs 146 and 147 yield

\[
\lambda = D = \frac{c_b}{c_0},
\]  

(148)

i.e. the mean penetration depth is given by the e-folding depth of the distribution function:

\[
\tilde{c} (\zeta) = c_0 e^{-\frac{\zeta}{D}} = \frac{c_b}{\tilde{c}_b} e^{-\frac{\zeta}{\tilde{c}_b}}.
\]  

(149)

The change of penetration depth due to vertically distributed sources and sinks \(f(\zeta)\) can then be calculated by the formula:

\[
\frac{dD}{dt} = \int_0^\infty (\zeta - D) f(\zeta) d\zeta.
\]  

(150)

As the model is not vertically explicit, but, based on the model assumptions, processes can be attributed to layers (e.g.
activity of aerobic bacteria to the aerobic layer), the changes $F_i$ caused in a given layer can be attributed to discrete depth levels being the centre of the layer $\zeta_i$.

The changes of penetration depth due to source and sink terms are complemented by the physical displacement of organic matter by the process of bioturbation, so that the total change is given by the equation:

$$\frac{\partial \psi D}{\partial t} = \sum_i \left( \frac{D'}{\psi D} f_i \frac{D'}{\psi D} + \frac{\partial \psi D}{\partial t} \right)_{\text{burb}}.$$  \hfill (151)

Bioturbation smoothes the concentration gradient and is therefore implemented as diffusive flux proportional to the difference in concentrations between 0 and a bioturbation length scale $\delta_{\text{burb}}$

$$\left. \frac{\partial \psi D}{\partial t} \right|_{\text{burb}} = \nu_{\text{burb}} \left( \frac{\psi D_0 - \psi D}{\psi D} \right),$$  \hfill (152)

where $\nu_{\text{burb}}$ is the bioturbation diffusivity of particulate matter (Eq. 207). Still assuming that $\psi D \ll d_{\text{tot}}$, this takes the form

$$\left. \frac{\partial \psi D}{\partial t} \right|_{\text{burb}} = \frac{\nu_{\text{burb}}}{\psi D} \left( 1 - e^{-\frac{\delta_{\text{burb}}}{D}} \right).$$  \hfill (153)

The fraction of organic matter contained between two given depth levels can then be computed as

$$\frac{\psi D}{\psi D_0} \frac{d_{\text{tot}}}{d_{\text{up}}} = \frac{1}{\psi D_0} \int_{d_{\text{up}}}^{d_{\text{tot}}} \gamma(\zeta) d\zeta = \frac{e^{-\frac{d_{\text{up}}}{D}} - e^{-\frac{d_{\text{tot}}}{D}}}{1 - e^{-\frac{d_{\text{tot}}}{D}}},$$  \hfill (154)

where the total content was approximated as

$$\frac{\psi D}{\psi D_0} = \int_{d_{\text{up}}}^{d_{\text{tot}}} \gamma(\zeta) d\zeta = \psi D_0 \left( 1 - e^{-\frac{d_{\text{tot}}}{D}} \right).$$  \hfill (155)

For consistency with the model assumptions and to avoid numerical issues the penetrations depths are constrained to values between $D_0$ and $d_{\text{tot}}$.

Dissolved organic matter is assumed to reside entirely in the oxygenated layer.

### 4.4 Heterotrophic bacteria

Benthic decomposers consist of aerobic bacteria living in the upper sediment layer down to the oxygen horizon and anaerobic bacteria living in the denitrification layer and anoxic layer. Their dynamics are summarised by the equations

$$\frac{\partial \psi C}{\partial t} = \frac{\partial \psi C_\text{upt}}{\partial t} - \frac{\partial \psi C_\text{excr}}{\partial t} - \frac{\partial \psi C_\text{pred}}{\partial t} + \frac{\partial \psi C_\text{rel}}{\partial t},$$  \hfill (156)

$$\frac{\partial \psi H_C}{\partial t} = \frac{\partial \psi H_C_\text{upt}}{\partial t} - \frac{\partial \psi H_C_\text{excr}}{\partial t} - \frac{\partial \psi H_C_\text{pred}}{\partial t} + \frac{\partial \psi H_C_\text{rel}}{\partial t},$$  \hfill (157)

where $r_{\text{upt}}$ and $r_{\text{excr}}$ are the bacteria and substrate mass specific reference uptake rates. These are generally high for the dissolved form and low for refractory matter. Decomposition of degradable matter has a slow basal component complemented by a fast component $r_{\text{fast}}$ subject to nutrient regulation.

To obtain the uptake rates these substrate mass specific rates are multiplied by the substrate concentrations available in the respective layer (given by Eq. 154):

$$\left. \frac{\partial \psi C}{\partial t} \right|_{\text{upt}} = \sum_i \mathcal{F}_{Q_i} \int_{d_{\text{up}}}^{d_{\text{tot}}} \gamma(\zeta) d\zeta,$$  \hfill (159)

where the layer limits $d_{\text{low}}, d_{\text{up}}$ are 0, $D$ for aerobic bacteria and $D$, $d_{\text{tot}}$ for anaerobic bacteria. Aerobic bacteria feed on dissolved and particulate substrate, while anaerobic bacteria feed exclusively on the particulate form.

The uptake of organic nitrogen and phosphorus is enhanced by a nutrient preference factor $p_{\text{pre}}$ supported by observations that the relative nutrient content of benthic DOM decreases under bacteria production. [van Duyl et al., 1993]
It is complemented by the uptake of inorganic forms when organic matter is nutrient-poor with respect to the fixed bacterial stoichiometric ratio. Inorganic uptake of nutrients by each bacteria type is regulated by Michaelis–Menten terms of the pore water inorganic nutrient content within the oxygenated or oxidised layer with the Redfield equivalent of carbon uptake as the half-saturation term:

\[
\frac{\partial N_{\text{upt}}}{\partial t} = \sum_{\psi} \frac{\chi}{P_{\text{upt}}} F_{\text{upt}} \left| Q_{\text{upt}} \right| \left| d_{\text{upt}} \right| + \chi_{\text{refl,upt}} \frac{\partial N_{\text{upt}}}{\partial t} \left| d_{\text{upt}} \right| + \chi_{\text{refl,upt}} \frac{\partial N_{\text{upt}}}{\partial t} \left| d_{\text{upt}} \right|, \quad (160)
\]

where \( K_{\text{up}} \left| d_{\text{up}} \right| \) and \( K_{\text{low}} \left| d_{\text{low}} \right| \) are the respective layer contents of ammonium or phosphate between the depth \( d_{\text{up}} \) and \( d_{\text{low}} \) and \( \nu_{\text{upt}} \) is a volume correction factor (Eq. [142]) reducing the total layer content to the pore water content.

Anaerobic bacteria is feeding on and excreting only in particulate form, so that the above rates are for gross uptake in the case of aerobic bacteria followed by excretion in dissolved form, while for anaerobic bacteria they are net rates with no subsequent excretion. Excretion occurs at fixed fractions \( q_{\text{dext}} \) and \( q_{\text{dext}} \) of the aerobic bacteria uptake according to:

\[
\frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{excr} \right| = q_{\text{dext}} F_{\text{upt}} \left| Q_{\text{upt}} \right| \left| d_{\text{upt}} \right| + q_{\text{dext}} F_{\text{upt}} \left| Q_{\text{upt}} \right| \left| d_{\text{upt}} \right| + q_{\text{dext}} F_{\text{upt}} \left| Q_{\text{upt}} \right| \left| d_{\text{upt}} \right|. \quad (161)
\]

Respiration of bacteria is given by activity respiration as a fraction of gross uptake \( q_{\text{resp}} \) and temperature regulated basal respiration at rest proportional to the bacteria biomass by the factor \( \chi_{\text{resp}} \):

\[
\frac{\partial H_{\text{upt}}}{\partial t} \left| \text{resp} \right| = \chi_{\text{resp}} \frac{\partial H_{\text{upt}}}{\partial t} \left| \text{upt} \right| + \chi_{\text{resp}} \frac{\partial H_{\text{upt}}}{\partial t} \left| \text{upt} \right|, \quad (163)
\]

Bacterial mortality is fully regulated by oxygen (see Eq. [242] and proportional to the bacteria biomass by factor \( \chi_{\text{mort}} \):

\[
\frac{\partial H_{\text{upt}}}{\partial t} \left| \text{mort} \right| = \chi_{\text{mort}} \left( 1 - \chi_{\text{O}} \right) H_{\text{upt}} \left| \text{mort} \right|, \quad (164)
\]

where aerobic bacteria use oxygen in dissolved form while anaerobic bacteria satisfy their oxygen requirements from oxidised nitrogen.

Benthic bacteria are held at a fixed stoichiometric quota \( Y_{\text{refl,upt}} \), so that any chemical component flux in excess of the reference quota is released according to Eqs. [258] and [259], in dissolved form for the nutrients and in the form of organic matter for carbon.

### 4.5 Predators

The general biogeochemical dynamics of the zoobenthos types \( \chi \) are given by the equations

\[
\frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{hgc} \right| = \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{upt} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{excr} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{resp} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{pred} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{mort} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{rel} \right|, \quad (165)
\]

\[
\frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{bgh} \right| = \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{upt} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{excr} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{resp} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{opt} \right| - \frac{\partial Y_{\text{upt}}}{\partial t} \left| \text{rel} \right|. \quad (166)
\]

The benthic predators considered in ERSEM are deposit feeders, suspension feeders and meiothons, distinguished by their prey fields and preferences, the depth section they live in and their respective metabolic rates. The prey fields available to each type are given in Fig. 5 where organic matter is scavenged only in the depth sections accessible to each predators given by three parameters as follows:

- suspension feeders: \( 0 \leq \zeta \leq d_{\text{SUSP}} \),
- deposit feeders: \( d_{\text{SUSP}} \leq \zeta \leq d_{\text{DEPO}} \),
- meiothons: \( 0 \leq \zeta \leq d_{\text{MEIO}} \).
An additional parameter $d_{\text{SUSP}}$ indicates the range of suspension feeders into the water column assuming homogenous prey distribution over this scale.

The total prey available to each zoobenthos type $\chi$ is composed of the individual prey types $\psi$ as

$$p_{\text{C},\psi} = \sum_\psi f_{\text{pr}} |\hat{Y}_\psi| \cdot \frac{f_{\text{pr}} |\hat{Y}_\psi| \psi_C}{f_{\text{pr}} |\hat{Y}_\psi| \psi_C + h_{\text{min}}} \cdot \hat{Y}_{\text{C},\psi},$$  \hspace{1cm} (165)

where $f_{\text{pr}} |\hat{Y}_\psi|$ are the food preferences and $h_{\text{min}}$ is a food half-saturation constant limiting the detection capacity of predator $\chi$ of individual prey types similar to the zooplankton predation (Eq. 29). In contrast to the pelagic form the detection capability for the benthic fauna is assumed to vary by food-source assuming that benthic predators search their food more actively. The prey contents in the half-saturation term are consequently multiplied by the food-preferences.

The prey mass specific uptake capacity for each zooplankton type $\chi$ is then given by:

$$\hat{Y}_{\chi,\psi} = \hat{Y}_{\text{max}} \cdot I_T \cdot I_O \cdot I_{\text{crowd}} \cdot Y_C \cdot C_{\text{upt}} + h_{\text{upt}},$$  \hspace{1cm} (166)

where $\hat{Y}_{\text{max}}$ is the maximum uptake capacity of each type at reference temperature, $I_T$ is the metabolic temperature response (Eq. 229), $I_O$ is the limitation of oxygen (Eq. 240), $I_{\text{crowd}}$ is a growth limiting penalty function accounting for overcrodding effects (Eq. 257) absent for meiobenhos as this type is capable of feeding on itself), $h_{\text{upt}}$ is a predation efficiency limiting the chances of encountering the prey available ($p_{\text{C}}$).

Introducing the prey mass specific fluxes from prey $\psi$ to predator $\chi$:

$$F_{\psi,\chi} = S_{\text{upt}} \cdot f_{\text{pr}} |\hat{Y}_\psi| \frac{f_{\text{pr}} |\hat{Y}_\psi| \psi_C}{f_{\text{pr}} |\hat{Y}_\psi| \psi_C + h_{\text{min}}} \cdot Y_C$$  \hspace{1cm} (167)

with $f_{\text{pr}} |\hat{Y}_\psi|$ being the food preference of predator $\chi$ for prey $\psi$, $h_{\text{min}}$ being a half-saturation constant reflecting the detection capacity of predator $\chi$, the zooplankton uptake can then be written as:

$$\frac{\partial Y_{\chi,\psi,\psi}}{\partial t} = \sum_\psi F_{\psi,\chi} \psi_C \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi},$$  \hspace{1cm} (168)

Zoobenthos excretion is given by:

$$\frac{\partial Y_{\chi,\psi,\psi}}{\partial t} \psi_C \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi}$$  \hspace{1cm} (169)

$$\frac{\partial Y_{\chi,\psi,\psi}}{\partial t} \psi_C \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi}$$  \hspace{1cm} (170)

where $\psi_{\text{excr}}$ is a fixed proportion of gross uptake excreted and $\psi_{\text{excr}}$ an additional dilution coefficient taking into account a reduced amount of nutrients in the fecal pellets with respect to the uptake quota.

Respiration of zoobenthos is given by activity respiration as a fraction of net uptake $\chi_{\text{aresp}}$ and temperature regulated respiration at rest proportional to the zoobenthos biomass by the factor $\psi_{\text{aresp}}$:

$$\frac{\partial Y_{\chi,\psi,\psi}}{\partial t} \psi_C \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi}$$  \hspace{1cm} (171)

Zoobenthos mortality is regulated by temperature and oxygen and composed of a basal part enhanced under oxygen deficiency and cold temperatures by the factors $\psi_{\text{mortO}}$, $\psi_{\text{mortT}}$:

$$\frac{\partial Y_{\chi,\psi,\psi}}{\partial t} \psi_C \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi} \cdot \psi_{\chi,\psi}$$  \hspace{1cm} (172)

Also, zoobenthos types are kept at a fixed stoichiometric quota $\psi_{\text{refO,P,C}}$ according to Eqs. 258 and 259 resulting in...
the release of nutrients in inorganic form and carbon in the form of degradable organic matter.

4.6 Organic matter

The cycling of carbon, nitrogen and phosphorus through the benthic food web by the processes of uptake, scavenging, excretion, mortality, release and burial results in the following organic matter fluxes:

The dissolved organic matter is produced by excretion and mortality and reduced by bacterial uptake

\[
\frac{\partial Q_{C,N,P}}{\partial t} = \frac{\partial Q_{C,N,P}}{\partial t}^{\text{dis}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{excr}} - \frac{\partial Q_{C,N,P}}{\partial t}^{\text{mort}} - \frac{\partial Q_{C,N,P}}{\partial t}^{\text{upt}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{refr}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{bur}}
\]

Degradable matter is generated by excretion and mortality and release fluxes, taken up by bacteria, and scavenged by zoobenthos

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{degr}} = \frac{\partial Q_{C,N,P}}{\partial t}^{\text{degr}}_{\text{bgc}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{degr}}_{\text{excr}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{degr}}_{\text{mort}} - \frac{\partial Q_{C,N,P}}{\partial t}^{\text{upt}} + \frac{\partial Q_{C,N,P}}{\partial t}^{\text{scav}}
\]

Refractory matter is taken up by bacteria and modified by burying across the total depth horizon

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{refr}} = \frac{\partial Q_{C,N,P}}{\partial t}^{\text{refr}}_{\text{bgc}} - \frac{\partial Q_{C,N,P}}{\partial t}^{\text{refr}}_{\text{upt}} - \frac{\partial Q_{C,N,P}}{\partial t}^{\text{refr}}_{\text{bur}}
\]

The abbreviated cycles for iron and silicate condensate all biogeochemical processes in the benthos into a simple remineralisation of degradable organic matter into dissolved inorganic iron or silicate at a fixed rate \(r_{\text{Fremin}}\) or \(r_{\text{Silremin}}\):

\[
\frac{\partial Q_f}{\partial t} = -r_{\text{Fremin}} \frac{\partial Q_f}{\partial t}^{\text{degr}}_{\text{bgc}}
\]

\[
\frac{\partial Q_S}{\partial t} = -r_{\text{Silremin}} \frac{\partial Q_S}{\partial t}^{\text{degr}}_{\text{bgc}}
\]

In these equations the partitioning in between the different forms of organic matter occurs in the following manner:

Uptake of all forms of organic matter by bacteria is given by Eqs. (156–158) as

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{upt}} = F_{\text{upt}}^{\text{C,N,P}} Q_{C,N,P}^{\text{upt}} + F_{\text{upt}}^{\text{H}} Q_{C,N,P}^{\text{upt}}. \tag{178}
\]

The excretion of aerobic bacteria is directed to dissolved organic matter, while for the zoobenthos types \(Y\) it is directed to degradable matter:

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{excr}} = \frac{\partial Q_{C,N,P}}{\partial t}^{\text{excr}}_{\text{aer}} + \sum_{\psi} \frac{\partial Q_{C,N,P}}{\partial t}^{\text{excr}}_{\text{anaer}} \tag{180}
\]

using Eqs. (161), (162), (169), (170).

The mortality of aerobic bacteria is partitioned between a particulate part directed to degradable matter and a dissolved part \(q_{dmort}\), while for the zoobenthos types \(Y\) and anaerobic bacteria it is entirely directed to degradable matter:

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{mort}} = q_{dmort} \frac{\partial Q_{C,N,P}}{\partial t}^{\text{mort}}_{\text{aer}} + \sum_{\psi} \frac{\partial Q_{C,N,P}}{\partial t}^{\text{mort}}_{\text{anaer}} \tag{182}
\]

using Eqs. (164) and (172).

Degradable matter is scavenged by zoobenthos according to Eq. (167)

\[
\frac{\partial Q_{C,N,P}}{\partial t}^{\text{scav}} = \sum_{\psi} F_{\text{scav}}^{\psi} \frac{\partial Q_{C,N,P}}{\partial t}^{\text{degr}}_{\text{bgc}}. \tag{183}
\]

In addition, degradable carbon may be produced by the stoichiometric adjustment (Eq. 258) of bacteria or zoobenthos.
the total depth horizon, we can use the displacement time
of living organisms in the model $d_{\text{tot}}$ may be interpreted as
burial flux (activated by the ISWbur switch), as material is
removed from the biogeochemical active part of the model.

To derive this flux we use a simple geometric argument
here: it is assumed that the diffusive process will preserve the
local concentration $C_{c,n,p}$, and the displacement rate of the
exponential profile at the given level. Specifically, we know that the local displacement rate at the level of the
penetration depth is precisely the change of penetration depth due to
bioturbation $\frac{\partial H_{\text{tot}}}{\partial t}_{\text{bur}}$.

To derive the local displacement rate of the exponential
profile at the total depth we can use the displacement time scale at $d_{\text{tot}}$, that is independent of the local concentration:

$$\frac{1}{\tau_{\text{bur}}(\zeta)} = \frac{1}{\tau_{\text{bur}}^{\text{refr}}(\zeta)} \frac{\partial \zeta_{\text{refr}}}{\partial t} = \frac{\zeta - \zeta_{\text{refr}}}{\zeta_{\text{refr}} D} \tau_{\text{bur}}(d_{\text{tot}} D) \frac{\partial D}{\partial t}_{\text{bur}}$$

(185)

Scaling the displacement rate using this scale the flux of
matter at $d_{\text{tot}}$, and hence the burial flux, can be computed as:

$$\frac{\partial Q_{c,n,p}}{\partial t}^{\text{refr}} = c_{c,n,p}(d_{\text{tot}}) \tau_{\text{bur}}(d_{\text{tot}} D) \frac{\partial D}{\partial t} \tau_{\text{bur}}(\zeta_{\text{refr}}) \frac{\partial D}{\partial t}_{\text{bur}}$$

(186)

This result can be formally confirmed by a straightforward, but fairly lengthy derivation of the time derivative of the integrated content of refractory matter between the sediment surface and $d_{\text{tot}}$ using Eqs. [149] and Eq. [155].

Note that this process removes biomass from the biogeo-
chemically active part of the model, as there are no processes
connected to buried organic matter and the model currently
does not consider remobilisation. This means that during
long term simulations the loss of nutrients needs to be com-
pensated, e.g. by riverine inputs or atmospheric deposition
(carbon is restored by air–sea exchange).

### 4.7 Inorganic components

The dynamics of benthic nutrients are given by the following
equations (see Eq. [177] for the remineralisation of silicate):

$$\frac{\partial K_{\text{N}}^{\text{ox}}}{\partial t}_{\text{bgc}} = \frac{\partial K_{\text{N}}^{\text{ox}}}{\partial t}_{\text{nitr}} - \frac{\partial K_{\text{N}}^{\text{ox}}}{\partial t}_{\text{denit}}$$

(187)

$$\frac{\partial K_{\text{N}}^{\text{amm}}}{\partial t}_{\text{bgc}} = - \frac{\partial K_{\text{N}}^{\text{amm}}}{\partial t}_{\text{nitr}} + \frac{\partial K_{\text{N}}^{\text{amm}}}{\partial t}_{\text{upt}} + \frac{\partial K_{\text{N}}^{\text{amm}}}{\partial t}_{\text{rel}}$$

(188)

$$\frac{\partial K_{P}}{\partial t}_{\text{bgc}} = - \frac{\partial K_{P}}{\partial t}_{\text{nitr}} + \frac{\partial K_{P}}{\partial t}_{\text{upt}} - \frac{\partial K_{P}}{\partial t}_{\text{rel}}$$

(189)

$$\frac{\partial K_{S}}{\partial t}_{\text{bgc}} = \tau_{\text{remin}} \frac{\partial C_{S}}{\partial t}$$

(190)

while the biogeochemistry of dissolved carbon, oxygen
dinitrogen are given by

$$\frac{\partial G_{C}}{\partial t} = \frac{\partial G_{C}}{\partial t}_{\text{resp}} \frac{\partial G_{O}}{\partial t} - \frac{\partial G_{O}}{\partial t}_{\text{resp}} \frac{\partial G_{\text{nitr}}}{\partial t}$$

(191)

$$\frac{\partial G_{N}}{\partial t}_{\text{bgc}} = \frac{\partial G_{N}}{\partial t}_{\text{resp}} + \frac{\partial G_{N}}{\partial t}_{\text{denit}}$$

(192)

(193)

The respiration terms of dissolved inorganic carbon and
dissolved oxygen are given by Eqs. (163) and (171) as

$$\frac{\partial G_{C}}{\partial t} = \sum_{\Psi} \frac{\partial Y_{\Psi}}{\partial t}_{\text{resp}} + \sum_{\Psi} \frac{\partial Y_{\Psi}}{\partial t}_{\text{resp}}$$

(194)

$$\frac{\partial G_{O}}{\partial t} = - q_{O:C} \left( \frac{\partial H_{C}}{\partial t}_{\text{resp}} + \sum_{\Psi} \frac{\partial Y_{\Psi}}{\partial t}_{\text{resp}} \right)$$

(195)

where $q_{O:C}$ is the oxygen to carbon conversion coefficient.

Nitrification in the benthos is computed similar to the
pelagic nitrification from a maximum ammonium mass
specific nitrification rate $r_{\text{nitr}}$ at reference temperature depending
on the ammonium available in the oxygenated layer, approximated as $\frac{m_{\text{amm}}}{d_{\text{tot}} K_{N}^{H}}$. 

\[ \frac{\partial K_{N}^{\text{ox}}}{\partial t} = \tau_{\text{remin}} \frac{d_{\text{tot}}}{D} \]
\[
\frac{\partial K_{\text{ox}}}{\partial t} \bigg|_{\text{nitr}} = \frac{\partial K_{\text{an}}}{\partial t} = \frac{H}{l_{\text{an}}^{\text{nitr}}} K'_{\text{an}}, \quad (196)
\]

\[
\frac{\partial G_{\text{an}}}{\partial t} = 2 \frac{\partial K_{\text{an}}}{\partial t} \bigg|_{\text{nitr}} \quad (197)
\]

where \( l_{\text{nitr}} \) and \( l_{\text{T}} \) are the nitrification limitation factors due to the presence of high concentrations of oxidised nitrogen and the temperature regulation factor (Eqs. 247 and 229).

Denitrification is calculated from the oxidised nitrogen reduction equivalent required for anaerobic bacteria respiration:

\[
\frac{\partial K_{\text{ox}}}{\partial t} \bigg|_{\text{nitr}} = \frac{\partial K_{\text{an}}}{\partial t} = \frac{H}{q_{\text{red}} q_{\text{ox}} C_{\text{denit}}} \frac{\partial H_{\text{C}}}{\partial t}, \quad (198)
\]

where \( q_{\text{red}} \) is the maximum fraction of anaerobic bacteria respiration resulting in oxidised nitrogen reduction, \( q_{\text{denit}} \) is the fraction of reduction subject to denitrification as opposed to ammonification and 2, \( \frac{5}{2} \) are the stoichiometric coefficients of oxygen demand per reduction equivalent for the ammonification and denitrification reactions respectively.

The actual reduction of oxidised nitrogen by denitrification is then further limited by availability of oxidised nitrogen \( (l_{\text{nitr}}, \text{Eq. 248}) \) resulting in the following denitrification fluxes:

\[
\frac{\partial K_{\text{ox}}}{\partial t} = l_{\text{nitr}} \frac{\partial K_{\text{an}}}{\partial t} \quad (199)
\]

\[
\frac{\partial K_{\text{an}}}{\partial t} = (1 - q_{\text{red}} N_{2}) \frac{\partial K_{\text{an}}}{\partial t} \quad (200)
\]

\[
\frac{\partial G_{\text{nitr}}}{\partial t} = q_{\text{red}} G_{\text{nitr}} \quad (201)
\]

where \( q_{\text{red}} G_{\text{nitr}} \) is the fraction of reduction directed to dinitrogen. As nitrogen fixation is currently not considered in the model, losses of oxidised nitrogen by denitrification are removed from the active cycle and need to be compensated in long term runs by riverine or atmospheric inputs, otherwise denitrification needs to be switched off.

Release of nutrients caused by stoichiometric adjustment (Eq. 258) of bacteria or zoobenthos are given by:

\[
\frac{\partial K_{\text{an}}}{\partial t} = \sum_{\chi} \frac{\partial H_{\text{C}}}{\partial t} + \sum_{\Psi} \frac{\partial Y_{\text{C}}}{\partial t} \quad (202)
\]

\[
\frac{\partial K_{\text{an}}}{\partial t} = \sum_{\chi} \frac{\partial H_{\text{P}}}{\partial t} + \sum_{\Psi} \frac{\partial Y_{\text{P}}}{\partial t} \quad (203)
\]

### 4.8 Bioirrigation

The diffusivity of dissolved inorganic states is given by a basal diffusivity \( \nu_{\chi} \) for each layer \( \chi \): aer, den, anox that is increased for bioirrigation by the factor \( p_{\text{bmino}} \). The activity of deposit feeders and meiofauna cause further enhancement to yield the total bioirrigation diffusivity \( \nu_{\text{diff}} \) (used in Eq. 140):

\[
\nu_{\text{diff}} = \nu_{\chi} \left( p_{\text{bmino}} + p_{\text{bienh}} \frac{S_{\text{burr}}}{S_{\text{burr}} + h_{\text{burr}}} \right), \quad (205)
\]

where \( q_{\text{burr}} \) and \( q_{\text{burr}} \) are the fractions of deposit feeder and meio-benthos uptake contributing to bioirrigation, \( h_{\text{burr}} \) is a half-saturation rate for bioirrigation enhancement and \( p_{\text{bienh}} \) is the maximum bioirrigation enhancement factor of dissolved inorganic diffusion in the benthos.

### 4.9 Bioturbation

For particulate matter in the benthos sediment diffusivity \( \nu_{\text{burb}} \) in Eq. 153 is based on a background diffusivity \( \nu_{\text{part}} \) and an enhancement factor of Michaelis–Menten type depending on the bioturbation caused by deposit feeder activity (see Eq. 168):

\[
\nu_{\text{burb}} = \nu_{\text{part}} \left( 1 + p_{\text{bienh}} \frac{S_{\text{burb}}}{S_{\text{burb}} + h_{\text{burb}}} \right), \quad (207)
\]

where \( q_{\text{burb}} \) is the fraction of deposit feeder uptake contributing to bioturbation, \( h_{\text{burb}} \) is a half-saturation rate for bioturbation enhancement and \( p_{\text{bienh}} \) is the maximum bioturbation enhancement factor of particulate matter diffusion in the benthos.

### 5 Horizontal interfaces

#### 5.1 The benthic-pelagic interface

The boundary condition at the seabed is given by the deposition of sinking particulate organic material, phytoplankton...
and calcite on the seafloor, the diffusion of inorganic chemical components between the pore water and the pelagic water column and resuspension of organic matter. All other state variables generally have no flux conditions at the pelagic-benthic interface.

5.1.1 Deposition of organic matter and phytoplankton

Deposition fluxes are taken analogous to the gravitational sinking rates in Eq. (138) where the sinking velocity is replaced by the deposition velocity $w_{dep}$, according to the seabed shear stress $\tau_{bed}$:

$$c_{p} w_{dep} = \max \left(1 - \frac{\tau_{bed}}{\tau_{crit}}, 0\right) c_{p} w_{sed}, \quad (208)$$

leading to the deposition fluxes

$$F^\text{ben}\left|_{dep}\right. = c_{p} w_{dep} c'_{p}. \quad (209)$$

As for gravitational sinking the only state variables sedimenting onto the seafloor are particulate organic matter, the phytoplankton components and calcite ($M_{C,N,P,F,S}$, $\chi_{C,N,P,F,S,C}$, $L_{C}$). The absorption of deposited carbon, nitrogen and phosphorus components into the sediments then results in separation of the organic material into dissolved, degradable and refractory matter according to

$$F^\text{lab}\left|_{dep}\right. = \sum_{\chi} \left(1 - q_{\text{depo}} \chi_{\text{depo}} - \chi_{\text{depo}}\right) F^\text{ben}\left|_{p}\right. + q_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{R}\right. + \tilde{F}_{\text{lab}}\left|_{P}\right. \quad (210)$$

$$F^\text{part}\left|_{dep}\right. = \chi_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{P}\right. + q_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{R}\right. + \tilde{F}_{\text{part}}\left|_{P}\right. \quad (211)$$

$$F^\text{refr}\left|_{dep}\right. = \chi_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{P}\right. \quad (212)$$

where $q_{\text{depo}}$ and $\chi_{\text{depo}}$ are the dissolved and refractory fractions of deposing material. For nitrogen and phosphorus the portioning is modified according to the relative cytoplasm nutrient contents $\chi_{\text{lab}}$, $\chi_{\text{part}}$, $\chi_{\text{degr}}$, $\chi_{\text{refr}}$:

$$F^\text{lab}\left|_{dep}\right. = \sum_{\chi} \left(1 - q_{\text{depo}} \chi_{\text{depo}} - \chi_{\text{depo}}\right) F^\text{ben}\left|_{p}\right. + q_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{R}\right. \quad (213)$$

$$F^\text{part}\left|_{dep}\right. = \sum_{\chi} \chi_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{P}\right. + q_{\text{depo}} \sum_{\chi} q_{\text{depo}} F^\text{ben}\left|_{R}\right. \quad (214)$$

$$F^\text{refr}\left|_{dep}\right. = \sum_{\chi} \chi_{\text{depo}} \chi_{\text{depo}} \sum_{\chi} F^\text{ben}\left|_{P}\right. \quad (215)$$

The iron and silicate components and phosphorus are entirely directed to degradable matter, the only state considered for these components in the benthic model:

$$F^\text{lab}\left|_{dep}\right. = \sum_{\chi} F^\text{ben}\left|_{P}\right. + F^\text{ben}\left|_{R}\right. \quad (216)$$

$$F^\text{part}\left|_{dep}\right. = F^\text{ben}\left|_{P}\right. + F^\text{ben}\left|_{R}\right. \quad (217)$$

Calcite deposition is given by

$$F^\text{calc}\left|_{dep}\right. = w_{calc} L_{C} \quad (218)$$

5.1.2 Resuspension

In the case of strong shear stress $\tau_{bed}$ at the seafloor part of the sediments may get resuspended into the water column. The erosion flux is calculated proportional to the excess shear stress over a critical threshold $\tau_{crit}$ by a reference erosion flux $r_{er}$ Erosion in terms of particulate organic matter is then approximated as a fraction of the total sediment matter $\rho_{Q}$:

$$S = r_{er} \max \left(\frac{\rho_{Q}}{\tau_{Q}}, 1\right) \quad (219)$$

$$\rho_{Q} = \frac{\rho_{Q}}{\tau_{Q}} + Q_{C} \quad (220)$$

The values and approximations used for the three parameters $\tau_{crit}$, $r_{er}$ and $\rho_{Q}$ are given in the Supplement.

5.1.3 Inorganic fluxes across the seabed

The diffusion of dissolved inorganic states across the benthos is derived from the equilibrium conditions described in Sect. 4.2. Based on the tendency of the system towards equilibrium the total flux across the sea-bed is then given by the
sum of all sources and sinks and a relaxation towards equilibrium.

\[ - \frac{\chi}{F} = \frac{\partial \chi}{\partial t} + \frac{1}{\tau_{eq}} \left( \chi - \chi_{pore,pCads} \right), \quad (221) \]

where \( \chi \) represents the inorganic states of oxygen, DIC, oxidised nitrogen, ammonium, phosphate and silicate.

For phosphorus, ammonium, silicate and DIC the relaxation fluxes towards equilibrium are computed by assuming a parabolic vertical distribution of excess biomass with 0 surface concentration and 0 bottom flux and assuming contributions to the generation of the excess proportional to the layer depth. The compensation flux across the seabed is then again computed from the production-diffusion balance in Eq. (144). For oxidised nitrogen and oxygen the procedure requires modification for two reasons: the separation depths of the oxygenated layer and denitrification layer given by the dissolved oxygen horizon and the horizon of oxidised nitrogen may be considered as fixed parameters for the diffusion-production balance of the other state variables, but not so for dissolved oxygen and oxidised nitrogen whose biogeochemical changes affect the dynamics of these horizons directly. In addition, the system imposes a third boundary condition on the balance equation, i.e. that the concentration at the respective horizon has to be zero by definition (and no sources and sinks exist below these limits), which renders the system overdetermined. For these two variables the relaxation time scale is therefore approximated by the fixed parameters \( \tau_{ox} \) and \( \tau_{denit} \) also used to determine the dynamical evolution of oxygen and oxidised nitrogen horizon in Eqs. (143) and (144).

The recycling of iron in the benthos is abbreviated, as there is very little information on the iron cycle in the sea-bed. The only form of iron considered in the benthos is the degradable matter, which is implicitly remineralised and returned to the water column in dissolved form at a fixed remineralisation rate \( \chi_{\text{remin}} \):

\[ \text{remin}_{\text{pel}} \left|_{\bar{Q}_{\chi}} \right. = r_{\text{remin}} \chi, \quad (222) \]

**5.1.4 Remineralisation of calcite**

No processes related to the formation or dissolution of calcite in the benthos are currently included in the model, the benthic cycle of calcite is resolved purely implicitly similar to iron as simple linear release to the water column of the calcite deposited onto the sediments:

\[ \text{remin}_{\text{calc}} \left|_{\bar{Q}_{\chi}} \right. = r_{\text{remin}} \chi, \quad (223) \]

**5.1.5 Benthic remineralisation sub-model**

As an alternative to the full benthic model described in the Sect. 4, a simple benthic closure is available, that implicitly remineralises benthic substrate into dissolved inorganic states, analogous to the treatment of iron and calcite above. The treatment of deposition and resuspension of organic matter on the sea floor in this case is identical to the full benthic model, while the recycling of organic matter occurs as a linear function of the benthic content at a given remineralisation rate \( \chi_{\text{remin}} \):

\[ \text{remin}_{\text{pel}} \left|_{\bar{Q}_{\chi}} \right. = r_{\text{remin}} \chi, \quad (224) \]

For nitrogen the remineralisation flux is split regenerating oxidised nitrogen and ammonium using the fixed fraction \( \chi_{\text{remin}} \):

\[ \text{remin}_{\text{nit}} \left|_{\bar{Q}_{\chi}} \right. = \left( 1 - \chi_{\text{remin}} \right) \chi_{\text{remin}} Q_{\text{N}} \]

With this option no other biogeochemical processes are considered in the benthos. The treatment of iron and calcite is identical between the full benthic model and this simplified benthic closure.

**5.2 Sea surface fluxes**

The only two boundary fluxes computed in the standard setup at the air–sea interface are the exchange of oxygen and carbon dioxide. Other processes like atmospheric deposition of nutrients and riverine inputs require spatially varying surface fields and are best provided through the physical driver. (Implementations of this type have been used in Artioli et al. 2012; Edwards et al. 2012; Holt et al. 2012; Wakelin et al. 2012)

Oxygen is exchanged based on the difference from the saturation state, which is estimated according to Weiss (1970):

\[ F_{O,\text{sea}}^\text{air} = k_{\text{air}} \left( T, S, u_{\text{wind}} \right) \left( O_{\text{sat}} - s_{\text{O}} \right). \quad (227) \]

The regression formula for \( s_{\text{O}} \) is given in the Supplement.

The exchange of carbon dioxide is based on the difference in partial pressures

\[ F_{C,\text{sea}}^\text{air} = \rho_{\text{sea}} k_{\text{air}} \left( T, u_{\text{wind}} \right) \left( p_{\text{CO}_2} - p_{\text{CO}_2}^\text{air} \right), \quad (228) \]

where \( p_{\text{CO}_2}^\text{air} \) maybe be provided by the physical driver or a constant parameter \( p_{\text{CO}_2}^\text{air} \).

The empirical gas transfer coefficients \( k_{\text{air}} \) and \( k_{\text{air}} \) are taken from Weiss (1970), Nightingale et al. (2000) and given in the Supplement.
6 Generic terms

6.1 Regulation and limitation factors

The regulation of metabolic processes by temperature is modelled using the $Q_{10}$ function introduced in Blackford et al. [2004] that strongly increases at low temperatures and decreases slower at high temperatures representing enzyme degradation:

$$
\chi_{\text{f}} = \frac{N_{\text{f}}}{N_{\text{f}} + h_{\text{f}}},
$$

where $T$ [$^\circ$C] is the water temperature in degrees Celsius and $\chi$ represents the respective process or state.

Nitrogen and phosphorus limitation factors for each of the four phytoplankton types are based on Droop-kinetics (Droop 1974) and computed as:

$$
\chi_{I_{\text{p}}} = \min \left( 1, \max \left( 0, \frac{\chi_{I_{\text{p}}} - \chi_{\text{min},\text{p},\text{c}}}{\chi_{\text{ref},\text{p},\text{c}} - \chi_{\text{min},\text{p},\text{c}}} \right) \right),
$$

$$
\chi_{I_{\text{N}}} = \min \left( 1, \max \left( 0, \frac{\chi_{I_{\text{N}}} - \chi_{\text{min},\text{N},\text{c}}}{\chi_{\text{ref},\text{N},\text{c}} - \chi_{\text{min},\text{N},\text{c}}} \right) \right),
$$

where $\chi$ represents any phytoplankton type (dia, micro, nano, pico), $\chi_{\text{ref},\text{p},\text{c}}$ is its reference internal quota and $\chi_{\text{min},\text{p},\text{c}}$ is its minimal internal quota. These two factors are combined to three alternative forms of co-limitation $\chi_{I_{\text{NP}}}$:

$$
\chi_{I_{\text{NP}}} = f \left( \chi_{I_{\text{N}}}, \chi_{I_{\text{p}}} \right),
$$

switchable through the namelist switch LimnutX:

- **LimnutX = 0**: $\chi_{I_{\text{NP}}}$ is the geometric mean of $\chi_{I_{\text{N}}}$ and $\chi_{I_{\text{p}}}$,
- **LimnutX = 2**: $\chi_{I_{\text{NP}}}$ is the harmonic mean of $\chi_{I_{\text{N}}}$ and $\chi_{I_{\text{p}}}$,
- **LimnutX = 1**: $\chi_{I_{\text{NP}}}$ is the minimum of $\chi_{I_{\text{N}}}$ and $\chi_{I_{\text{p}}}$.

The silicate limitation factor for diatoms is computed from the external availability of dissolved silicate $N_{\text{d}}$, based on a Michaelis–Menten term with half-saturation $h_{\text{d}}$:

$$
\chi_{\text{d}} = \frac{N_{\text{d}}}{N_{\text{d}} + h_{\text{d}}},
$$

The iron limitation factor is computed in the same way as the factors for nitrogen and phosphorus:

$$
\chi_{\text{f}} = \min \left( 1, \max \left( 0, \frac{\chi_{I_{\text{f}}} - \chi_{\text{min},\text{f},\text{c}}}{\chi_{\text{ref},\text{f},\text{c}} - \chi_{\text{min},\text{f},\text{c}}} \right) \right),
$$

with $\chi_{\text{ref},\text{f},\text{c}}$ as its reference internal quota and $\chi_{\text{min},\text{f},\text{c}}$ as its minimal internal quota.

Phosphorus and nitrogen limitation $l_{\text{f}}, l_{\text{N}}$ for the standard model of bacteria mediated decomposition can be based on the availability of the resource in dissolved inorganic form ($\text{ISWBLimX} = 1$) and substrate or only in inorganic form ($\text{ISWBLimX} = 2$):

$$
B
l_{\text{f}} = \begin{cases} 
\min \left( \frac{N_{\text{f}}}{N_{\text{f}} + h_{\text{f}}}, \frac{\chi_{\text{ref},\text{f},\text{c}} - \chi_{\text{min},\text{f},\text{c}}}{\chi_{\text{ref},\text{f},\text{c}} - \chi_{\text{min},\text{f},\text{c}}} \right) & \text{if } \text{ISWBLimX} = 1 \\
\frac{N_{\text{f}} + h_{\text{f}}}{N_{\text{f}} + h_{\text{f}} + h_{\text{p}}} & \text{if } \text{ISWBLimX} = 2
\end{cases}
$$

and analogous:

$$
B
l_{\text{N}} = \begin{cases} 
\min \left( \frac{N_{\text{N}}}{N_{\text{N}} + h_{\text{N}}}, \frac{\chi_{\text{ref},\text{N},\text{c}} - \chi_{\text{min},\text{N},\text{c}}}{\chi_{\text{ref},\text{N},\text{c}} - \chi_{\text{min},\text{N},\text{c}}} \right) & \text{if } \text{ISWBLimX} = 1 \\
\frac{N_{\text{N}} + h_{\text{N}}}{N_{\text{N}} + h_{\text{N}} + h_{\text{p}}} & \text{if } \text{ISWBLimX} = 2
\end{cases}
$$

where $h_{\text{P},\text{N}}$ are the half–saturation constants for phosphorus and nitrogen limitation.

Nutrient regulation of benthic bacteria occurs based on the nutritional state of the substrate

$$
l_{\text{H}} = \min \left( 1, \frac{\chi_{\text{degr},\text{f},\text{p},\text{c}} - \chi_{\text{ref},\text{f},\text{p},\text{c}}}{\chi_{\text{ref},\text{f},\text{p},\text{c}} - \chi_{\text{min},\text{f},\text{c}}} \right),
$$

where $\chi$ are aerobic and anaerobic bacteria within the layers described in Sect. 4.4.

Oxygen limitation of zooplankton ($\chi$: HET, MICRO, MESO) is computed as function of the relative oxygen saturation state

$$
s_{\text{relO}} = \min \left( 1, \frac{G_{\text{O}}}{s_{\text{O}}} \right)
$$

$$
l_{\text{O}} = \frac{s_{\text{relO}} + s_{\text{relO}} \chi}{s_{\text{relO}} + \chi},
$$

where the oxygen saturation concentration $s_{\text{O}}$ is estimated according to Weiss [1970]. (The regression formula used is given in the Supplement.)

For zoobenthos ($\chi$: DEPO, SUSP, MEIO) it is given by a cubic Michaelis–Menten response to the oxygen concentration in the overlying water body in relation to a minimum...
oxygen threshold \( \tilde{y}_{O_{\text{min}}} \) for each species:

\[
\chi_l = \frac{\max \left( G_\Omega - \tilde{y}_{O_{\text{min}}, \Omega} \right)^3}{\max \left( G_\Omega - \tilde{y}_{O_{\text{min}}, \Omega} \right)^3 + \chi_l^3}.
\]  

(240)

For pelagic bacteria it is given by a simple Michaelis–Menten term of the relative oxygen saturation state (Eq. 238)

\[
B_l = \frac{s_{\text{rel}O}}{s_{\text{rel}O} + h_l}.
\]  

(241)

For benthic bacteria oxygen regulation occurs through the oxygen and oxidised nitrogen horizons

\[
l_l = \frac{O_3}{O_3 + h_l} \quad \text{for aerobic nitrification} \quad a_{\text{aer}} = \frac{\text{oxy}}{D + d_{\text{ref}}} \quad \text{and aerobic denitrification} \quad a_{\text{anaer}} = \frac{\text{denit}}{D - D + d_{\text{ref}}},
\]  

(242)

where \( d_{\text{ref}} \) is the aerobic half saturation depth and \( d_{\text{ref}} \) the oxidised half saturation depth for oxygen regulation.

\( l_l \) is the oxygen limitation factor for nitrification:

\[
l_l = O_3 \quad \text{with } h_l \text{ being the cubic saturation constant for oxygen limitation of nitrification}.
\]  

(243)

\( l_l \) is the substrate limitation factor for nitrification:

\[
l_l = \frac{\text{amm}^3}{N_N + \text{nitr}} \quad \text{with } h_{\text{nitr}} \text{ being the cubic saturation constant for substrate limitation of nitrification and } l_{\text{pH}} \text{ is the pH-limitation factor for nitrification:}
\]  

\[
l_{\text{pH}} = \min (2, \max (0, 0.6111 \text{pH} - 3.8889)).
\]  

(245)

Benthic nitrification is inhibited at high benthic content of oxidised nitrogen according to

\[
K_N^{\text{nit}} = \frac{\text{ox}}{D + \frac{\text{denit} \cdot \text{oxy}}{D - D}}.
\]  

(246)

\[b_{\text{nitr}} = \frac{h_{\text{nitr}}}{h_N + K_N^{\text{nit}}},\]

(247)

where \( h_N \) is the oxygenated layer concentration of oxidised nitrogen at which nitrification is inhibited by 50%.

Here, it is assumed that some oxidised nitrogen penetrates into the denitrification layer, so that the oxygenated layer concentration is on average three times higher compared to the denitrification layer.

Based on the same assumption, denitrification in the oxidised layer uses a Michaelis–Menten response to the assumed layer content of oxidised nitrogen

\[
K_N^{\text{denit}} = \frac{1}{3} \frac{\text{oxy}}{\text{oxy} + \frac{\text{ox}}{D + \frac{\text{denit} \cdot \text{oxy}}{D - D}}}.
\]  

(248)

\[l_N^{\text{denit}} = \frac{h_{\text{denit}}}{h_N + h_N^{\text{denit}}},\]

(249)

where \( h_N^{\text{denit}} \) is a denitrification half saturation constant.

Calcification and dissolution of calcite occur in relation to the calcite saturation state of the water \( \Omega_{\text{calc}} \geq 1 \) (Eq. 126). The regulating factor of the rain ratio for calcification and the regulation factor for dissolution of calcite can be calculated in two alternative ways chosen by the ISWCAL = 1 namelist switch. The first option \( \text{(ISWCAL = 1)} \) is based on an exponential term:

\[
l_C^{\text{calc}} = \max \left(0, (\Omega_{\text{calc}} - 1)^{n_{\text{calc}}} \right),
\]  

(250)

\[
l_C^{\text{dis}} = \max \left(0, (1 - \Omega_{\text{calc}})^{n_{\text{dis}}} \right),
\]  

(251)

where \( n_{\text{calc}}, n_{\text{dis}} \) are calcification/dissolution exponents (Ridgwell et al. 2007; Keir 1980).

The second option \( \text{(ISWCAL = 2)} \) uses a Michaelis–Menten term:

\[
l_C^{\text{calc}} = \max \left(0, \frac{\Omega_{\text{calc}} - 1}{\Omega_{\text{calc}} - 1 + h_{\text{calc}}} \right),
\]  

(252)

\[
l_C^{\text{dis}} = \max \left(0, \frac{1 - \Omega_{\text{calc}}}{1 - \Omega_{\text{calc}} + h_{\text{calc}}} \right),
\]  

(253)

where \( h_{\text{calc}} \) is the half-saturation constant for calcification and dissolution of calcite (Blackford et al. 2010 Gehlen et al. 2007).

The rain ratio (Eq. 90) is regulated by nutrient limitation and temperature to reflect the dependency of the calcifying fraction of nanophytoplankton on the environmental conditions. Temperature regulation is given by

\[
l_T^{\text{calc}} = \max \left(0, \frac{T [^\circ C]}{T [^\circ C] + h_{\text{calc}}} \right),
\]  

(254)

where the half-saturation constant is set to \( h_{T} = 2 {^\circ C} \). As coccolithophores are reported to have generally higher phosphorus affinity but lower nitrogen acquisition capacity with respect to other phytoplankton (Riegman et al. 2000; Pausche 1998), limitation of these nutrients has an opposed
impact on the rain ratio. This is reflected in our combined nutrient limitation factor for calcification which is obtained from the phosphorus and nitrogen limitation of nanophytoplankton (Eqs. 231 and 230) as

$$l_{(P,N)} = \min \left(1 - \frac{l_{P}^{\text{nano}}}{l_{P}^{\text{nano}}}, 1 - \frac{l_{N}^{\text{nano}}}{l_{N}^{\text{nano}}} \right).$$  (255)

Uptake limitation of suspension and deposit feeders by overcrowding is given by a nested Michaelis–Menten response to the respective biomass:

$$\dot{\chi}_{\text{crowd}} = \max \left(0, \frac{\chi^{\text{C}} - \dot{\chi}_{\text{C}}}{\chi^{\text{C}} - \dot{\chi}_{\text{C}} + \dot{\chi}_{\text{sat}}} \right).$$  (256)

$$\chi_{\text{crowd}} = 1 - \frac{\chi_{\text{crowd}}}{\dot{\chi}_{\text{crowd}} + \dot{\chi}_{\text{C}}},$$  (257)

### 6.2 Stoichiometric adjustments

For states $\chi$ with fixed stoichiometric quota $q_{\text{N}, P; \text{C}}$ (mesozooplankton, benthic bacteria and predators) the process rates are complemented by release fluxes that regulate imbalances in order to preserve the fixed reference quotas as follows:

$$\frac{\partial \chi_{\text{C}}}{\partial t}_{\text{rel}} = \max \left( \frac{\partial \chi_{\text{C}}}{\partial t}_{\text{bgc}} - \frac{1}{\chi} \frac{\partial \chi_{\text{P}}}{\partial t} \right),$$

$$\frac{\partial \chi_{\text{P}}}{\partial t}_{\text{bgc}} = - \frac{1}{\chi} \frac{\partial q_{\text{N}, P; \text{C}}}{\partial t}_{\text{bgc}},$$

$$\frac{\partial \chi_{\text{P}}}{\partial t}_{\text{net}} = \max \left( \frac{\partial \chi_{\text{P}}}{\partial t}_{\text{net}} - q_{\text{N}, P; \text{C}} \frac{\partial \chi_{\text{C}}}{\partial t} \right).$$

(258)

(259)

where $\frac{\partial \chi_{\text{P}}}{\partial t} _{\text{net}}$ are the comprehensive biogeochemical process rates prior to adjustments

$$\frac{\partial \chi_{\text{P}}}{\partial t} _{\text{net}} = \frac{\partial \chi_{\text{P}}}{\partial t} _{\text{rel}} - \frac{\partial \chi_{\text{P}}}{\partial t} _{\text{net}}.$$  (260)

### 7 Implementations

Most ecosystem models are tightly bound to a specific physical, hydrodynamic driver that is usually three-dimensional and consequently computationally heavy and cumbersome to test and implement. The ERSEM model comes as an independent library and can in principle be coupled to any physical driver with comparatively little effort. In fact, coupled configurations exist for a variety of drivers in one or three-dimensional settings amongst which are the NEMO ocean engine (Madec 2008), the POLCOMS model for shelf seas (Holt and James 2001), and the GOTM/GETM model (Burchard et al. 2006). While for realistic implementations a full-scale three-dimensional configuration is required, for the stages of process development and qualitative analysis of the functioning of the modelled ecosystem, zero- or one-dimensional frameworks are often beneficial as they provide a light-weight implementation that is easier to grasp, much faster to run, amenable to sensitivity analysis and quicker to analyse.

The model distribution itself includes drivers for two idealised systems: the first is a simple zero dimensional implementation of mesocosm type called the ERSEM-Aquarium with a pelagic box overlying a benthic box, each of them with internally homogeneous conditions. This is essentially a test environment for new users and fast process assessment requiring no external software for the ocean physics. The second is a driver for the vertical one-dimensional GOTM model (http://www.gotm.net – Burchard et al. 2006). It is a more realistic system allowing for full vertical structures in a comparatively lightweight software environment that is capable of running in serial mode on any standard desktop or laptop. It requires a copy of the GOTM code with minor modifications to accommodate ERSEM, which can be obtained for the stable release or the development release of GOTM (see Sect. 10). Here, we use the 0-D framework to illustrate the carbon fluxes through the model food web under contrasting environmental conditions (Sect. 7.1) and the 1-D implementation to demonstrate the model capacity to reflect the lower trophic level of the marine ecosystem under varying conditions at three different sites, underpinned by a brief validation against in situ time-series data (Sect. 7.2).

Beyond these simpler test cases, the ERSEM model has been implemented in various full-scale three dimensional applications from coastal to global scales, cited above. The descriptions of these configurations would exceed the scope and volume of this paper and are given in the respective publications, but for completeness we give a short example of a simulation based on a previously published configuration in order to illustrate the full potential of the model (Sect. 7.3).

All simulations presented in this section were performed using the same parametrisation, which is given in the Supplement. This parametrisation was developed using size as the main trait to scale the metabolic rates of the pelagic functional groups more widely than in previous parametrisations (Baretta-Bekker et al. 1997) and extends the conventional restriction of the food matrix suggested in Eq. (13). A table with all parameter values, their mathematical symbols as used in Sect. 2 to 6 and the corresponding name in the model code and namelists is given in the Supplement.
7.1 ERSEM-Aquarium

The simulation of mesocosm type environments is supported through the ERSEM-Aquarium model. The model simulates two 0-D boxes, a pelagic box, which is characterised by its mid-depth below the surface and by the geographical location, and a benthic box beneath it. Seasonal variations in temperature and salinity can be imposed as cosine functions between an extreme value at the first of January in the beginning of the simulation and a second extreme after half a year. The light field can be imposed in the same way as cosine oscillation between two prescribed extreme values, or extracted from the prescribed geographical position using standard astronomical formula ignoring cloud cover. Additionally diurnal oscillations of temperature and light can be superimposed in cosine form by prescribing a daily excursion between midday and midnight. It should be noted that this framework is not designed to deliver realistic simulations of the marine environment in a particular location, but rather to aid the development and quick evaluation of process studies, or to study the model system behaviour in a simplified context without additional complicating factors.

Figure 6 illustrates the carbon fluxes between model compartments for two different simulations using the ERSEM-Aquarium. The first is configured as a representation of tropical oligotrophic conditions characterised by deep and warm waters with high irradiance and low nutrients, while the second roughly corresponds to the shallow coastal eutrophic waters of the Southern North Sea with strong nutrient supply and comparatively low light. Both configurations are run for a thousand years in order to achieve full equilibrium between the benthic and pelagic environments. The former uses the simple benthic closure scheme for remineralisation (Sect. 5.1.5), which is more appropriate for deep water configurations where the impact of the benthos is of lesser importance, while the latter uses the full benthic model (Sect. 4). All configuration files necessary to replicate these runs are given in the Supplement. Figure 6 gives flux magnitudes in the modelled food web directly scaled from the annual average of the last year of each simulation. The experiment highlights the substantial quantitative production differences in between the two systems. In addition, it clearly shows the qualitative shift in the model food web under the contrasting conditions. In the oligotrophic case most of the gross production is excreted to dissolved matter due to strong nutrient limitation. This leads to a microbe dominated scenario with bacteria as the main food-source for the predators and only small amounts of carbon entering the second trophic level leading to negative community production and low deposition of biomass to the sediments. In the eutrophic case production levels are increased by an order of magnitude. The assimilated carbon is used more efficiently by phytoplankton fueling substantial secondary production with autotrophs as the main food source of zooplankton and significantly more biomass exported to the sediments resulting in positive community production.

7.2 GotmErsem – a model Framework for the water column

The GotmErsem framework provides the possibility to include a more realistic physical environment into the simulations with opposing gradients of nutrient supply from depth and short-wave radiation attenuated as it penetrates through the water column. The GOTM model is a one-dimensional water column model including a variety of turbulence closure schemes for vertical mixing (Burchard et al., 2006). Here, we show three implementations using this framework in contrasting environments to demonstrate the portability of the ERSEM model, one for the Oyster Grounds in the Southern North Sea, a typical shelf sea site; one at the L4 site in the Western English Channel representative of a mid-latitude site with mixed waters of both oceanic and coastal origin; and one in the oligotrophic sub-tropics at the Bermuda Atlantic Time-series Study site. Each of these sites is supported by extensive in situ data sets for model evaluation. Full configuration files to run these simulations are provided in the Supplement. The validation against in situ data was performed by sub-sampling the daily averaged model output for each in situ data sample. It is presented in target diagrams (Jolliff et al., 2009) for each site showing statistically robust metrics (e.g. Daszykowski et al., 2007) to account for the underlying non-Gaussian asymmetric data distributions and in order to avoid spurious over-weighting of outliers. The metrics provided are the median bias (median \((M_i - D_i)\); \(M_i\): model sample, \(D_i\): data sample) on the ordinate and the unbiased median absolute error (MAE′, median \(|\text{abs}(M_i - D_i - \text{median}(M_i - D_i))|\)) on the abscissa. Both are normalised with the inter-quartile range (IQR) for the scale of the in situ data and the Spearman or rank correlation is represented by the colour code for each data set. The sign on the abscissa is given by the relation of IQRs (sign \((\text{IQR}(M_i) - \text{IQR}(D_i))\)).

All three sites are forced with data from the ERA-interim reanalysis (Dee et al., 2011) at the atmospheric boundary condition. The L4 and Oyster Ground configurations use surface pressure data to introduce tidal mixing into the idealised one-dimensional set-ups. The BATS and L4 site were additionally relaxed towards temperature and salinity profiles from CTD measurements (BATS – Steinberg et al., 2001; L4 – Harris, 2010) in order to compensate for the missing hydrodynamic impacts of lateral advection and diffusion. Initial conditions for the sites were derived from the concurrent in situ data where available. As for the ERSEM-Aquarium simulations the benthic remineralisation closure was used for the deep, oligotrophic BATS site, while for the shallow eutrophic sites L4 and Oyster Grounds the full benthic model was used.
7.2.1 Oyster Grounds – (54°24'36'' N, 4°1'12'' E)

This site is located in the Southern North Sea and is influenced by the English Channel and surrounding coastal waters, with seasonal stratification in most summers and an accentuated spring bloom at the onset of stratification that depletes the nutrients from the comparatively stable and isolated water surface layer (Baretta-Bekker et al., 2008).

A comparison with smart buoy data for the years 2000–2009 (Greenwood et al., 2010) reveals a good representation of the local seasonal cycle (Fig. 7). Simulations do not show significant bias in any of the variables, while the MAE’ is significantly lower than the in situ data variability (≈ 0.75 of the IQR of the in situ data for chlorophyll a, ≈ 0.25 silicate and phosphate and virtually 0 for oxidised nitrogen). Correlations are high for the nutrients (> 0.6), but comparatively low for chlorophyll a (> 0.2). The lower skill for the latter is partly caused by a weaker secondary bloom in summer in the simulations compared to the observations and comparatively low observational coverage over the first years of the simulation leading to potential overestimation of singular events in the data sampling and giving a spurious picture of the seasonal cycle when compared to the more consistently covered last three years of the period shown. In addition, some deficiencies, in the model simulations are to be expected as the Oyster Ground site is characterised by strong lateral influences including estuarine, coastal and channel waters that include strong direct impacts on the nutrient concentrations in the area that can not be captured in this idealised setting. Particularly in the stratified season in summer these lateral effects are dominating the surface water signal while the deeper part of the depression is essentially isolated from the surface layer (Weston et al., 2008).

7.2.2 L4 – Western English Channel (50°15’ N, 4°13’ W)

The L4 site is a long-term monitoring station near the Northern coast of the Western English Channel. Similar to the Oyster Grounds site, it is seasonally stratified and generally nutrient depleted in summer, but highly affected by episodic events of freshwater inputs of riverine origin (Smyth et al., 2010).

Figure 8 shows the seasonal cycles of oxidised nitrogen, phosphate and chlorophyll a at the sea surface for the model simulations and for the in situ data (Smyth et al., 2010 – http://www.westernchannelobservatory.org.uk/) for the years 2007–2011. The model follows the seasonal cycle of nutrient depletion in summer and nutrient resupply in winter revealed by the data in all three nutrients shown. Also the results for chlorophyll a follow the bulk seasonality represented by the in situ data, but show deficiencies in capturing the episodic peaks, which appear misplaced with respect to the measurements. Possible reasons for these short-comings include the absence of physical and biogeochemical impacts of lateral processes in such an idealised 1-dimensional setting as well as a sub-optimal representation of the local plankton community by the parametrisation adopted consistently across the contrasting environments. Nevertheless, the model skill expressed in the overall statistics is considerable. The bias and MAE’ for all 4 variables fall well below the variability of the in situ data. Chlorophyll a shows a relative bias of about 0.25 and a relative unbiased error of little less than 0.5, while the three nutrients show an error and bias very close to 0.

7.2.3 BATS – Bermuda, Sargasso Sea (31°40’ N, 64°10’ W)

This site in the Sargasso Sea is characterised by a weak geostrophic flow with net down-welling. Strong stratification separates the nutrient-poor surface waters from the nutrient-
rich deep water, with the exception of the passing of cold fronts in winter which cause substantial convective mixing with accompanying nutrient entrainment (Steinberg et al., 2001). This is illustrated in the left panel of Fig. 9 which shows the seasonal cycle of chlorophyll $a$ from model simulations (on top) and in situ data. The mixing events triggering autotrophic growth initially spread over the upper part of the water column, but they are limited to a rather marked deep-chlorophyll $a$ maximum at around 100 m depth when stratification sets in. Interannual variability at the site is dominated by the varying strength of the sub-tropical storm events in spring that cause strong vertical mixing which can reach up to 200 m depth resulting in variable levels of nutrient entrainment, largely captured by the model. A summary of the validation against the extensive in situ data available at BATS (Bermuda Time Series Study—Steinberg et al., 2001) for the years 1990–2008 is given in the target diagram on the right of Fig. 9. In contrast to the two shallow sites, in situ data in this case is vertically resolved, which was respected in the matching procedure.

Bias and MAE’ for all variables do not exceed the variability of the in situ data. Both metrics are very close to zero for the nitrate, phosphate and chlorophyll $a$ and in general most metrics stay below 50% of the in situ variability with the exception of the bias for oxygen and the MAE’ for phosphate. The latter are caused by an underestimated aeriation of the water column and a weaker vertical gradient in phosphate for the model (not shown). However, some weaknesses in the simulation of the vertical distributions are to be expected given the absence of explicit lateral dynamics and the resulting vertical flows. Correlations lie between 0.4 and 0.6 reflecting the overall satisfactory model performance.

Figure 7. Simulation results vs. in situ data at the Oyster Grounds – left: model time series (red lines) vs. in situ measurements (black dots) for oxidised nitrogen, phosphate, silicate and chlorophyll $a$ (top to bottom); right: target diagram with bias (abscissa), MAE’ (ordinate) and Spearman correlation (colour code) for oxidised nitrogen (NO3), phosphate (PO4), silicate (Sil) and chlorophyll $a$ (Chl). The observations consist of ship-based data collected by Rijkswaterstaat as part of the Dutch national monitoring programme MWTL (see [publicwiki.deltares.nl/display/OET/Dataset+documentation+MWTL]) and SmartBuoy data collected by Cefas in collaboration with Rijkswaterstaat (Greenwood et al., 2010; http://www.cefas.co.uk/publications-data/smartbuoys).

Figure 8. Simulation results vs. in situ data at the L4 site – left: model time series (red lines) vs. in situ measurements (black dots) for oxidised nitrogen, phosphate, silicate and chlorophyll $a$ (top to bottom); right: target diagram with bias (abscissa), MAE’ (ordinate) and Spearman correlation (colour code) for oxidised nitrogen (NO3), phosphate (PO4), silicate (Sil) and chlorophyll $a$ (Chl).
7.2.4 Properties emerging from simulations at all three sites

In order to give an impression of the functioning of the ecosystem dynamics across the three sites, Fig. 10 shows a comparison between some ecosystem properties emerging from data meta-analysis and model simulations, namely the internal stoichiometric quotas of nutrients with respect to carbon in phytoplankton and the phytoplankton community structure. On the left of Fig. 10 we show the range of the internal stoichiometric quotas of nitrogen, phosphorus, silicate and iron with respect to carbon on the abscissa plotted against the average quotas for phytoplankton on the ordinate as an indicator of the modelled phytoplankton plasticity in response to nutrient limitation. Quotas from the simulations (circles) are compared to the results of a meta-analysis (diamonds) provided by Moore et al. (2013) based on observed internal stoichiometric phytoplankton quotas from scientific literature. Results for the three macronutrients are consistent in that the average quotas are well matched while the stoichiometric range is underestimated by approximately half an order of magnitude. This is to be expected given that the case studies included in the model simulations don’t cover the full range of natural variability of marine environments. Results for iron show substantial differences in range and average state. The mismatch in average state can be attributed to the fact that the present parametrisation of the iron cycle took into consideration the works of Timmermans et al. (2005) and Veldhuis et al. (2005), which reported comparatively low iron to carbon quotas, but weren’t considered in the above meta-analysis, while the huge discrepancy in range is caused by the absence of substantial iron limitation in the sites of the case studies.

The right hand side panel of Fig. 10 shows the size fractioned contribution of each phytoplankton group to total chlorophyll a across the three sites as a running average over the ordered model samples from all three sites collectively. The procedure is analogous to the meta-analysis provided by Hirata et al. (2011). The results show a domination of the phytoplankton community by picophytoplankton at low chlorophyll a and by large phytoplankton at high chlorophyll a. Nanophytoplankton is present throughout the chlorophyll a range, reaching a maximum at intermediate values. The emerging modelled community structure compares well to the meta-analysis (compare Fig. 2a–c therein) particularly considering the limited range of marine environments considered in this exercise.

7.3 A full scale implementation for the North-West European Shelf

The previous case studies demonstrate the capability of the model to represent the marine ecosystem with a focus on small scale ecosystem processes. Nevertheless, the full potential of the model unfolds in full-scale applications of coupled dynamical systems linked to hydrodynamic models capturing the full advection and diffusion of the biogeochemical states and thus providing a complete synoptic picture of the large scale biogeochemical cycles and the marine environment. A full description of these systems would exceed the scope of this particular paper. Nevertheless, we give here a brief overview of the model performance on a simulation of the North-West European Shelf Seas using the POLCOMS model for shelf sea circulation (Holt and James, 2001), based on a hindcast configuration identical to the one used and described in Holt et al. (2012) and Artioli et al. (2012), but using the most recent model version presented in this work and the same parametrisation as in the above examples.

The left hand side panel of Fig. 11 shows the mean optical-depth-averaged chlorophyll a field of the area to illustrate the model domain as used in the validation exercise, and also to give an idea of the ecosystem characteristics of the area. Model simulations were validated against in situ data for oxidised nitrogen, phosphorus, chlorophyll a, oxygen and salinity retrieved from the ICES data base (ICES, 2009) for the period of 1970–2004 using the same metrics as above, sum-
Figure 10. Emergent properties of the simulations across the three 1-D sites. Left: range (ordinate) and mean (abscissa) of internal stoichiometric ratios of phytoplankton – nitrogen (yellow), silicate (blue), phosphorus (green) and iron (red). Data (diamonds, Moore et al., 2013), assembled 1-D model simulations (circles); right: community fraction of total chlorophyll $a$ from assembled 1-D model simulations. Picophytoplankton (red), nanophytoplankton (green) and microphytoplankton and diatoms (cyan).

Figure 11. The ERSEM model in a simulation for the North West European Shelf Seas – left: optical-depth-averaged chlorophyll $a$; right: hindcast simulation vs. in situ data.

Results are consistent with the validation results of the 1-D sites with both bias and MAE $'$ generally less than 50% of the in situ variability, and correlations $> 0.4$ for all variables confirming the good performance of the model dynamics in a realistic large-scale simulation.

8 Development and testing framework

In addition to the 0- and 1-D ERSEM implementations a framework is provided with the model that allows developers and users of the code to analyse and plot the result of calls to individual ERSEM procedures from Python. This facility is supported through Fortran–C interoperability, that arrived with the Fortran 2003 standard (ISO/IEC 1539-1:2004(E)), and the Python Ctypes package. ERSEM test harnesses consist of the ERSEM library and a set of C wrappers, which are jointly compiled as a shared library. A Python interface to the shared library permits access to Fortran data structures and procedures from Python. This allows developers and users of the code to quickly interrogate the validity and behaviour of individual procedures, without first reimplementing them in a second language, and without running the full model. Here we illustrate this feature by examining the photosynthesis model implemented in ERSEM.

The photosynthesis model used in ERSEM is based on Geider et al. (1997), and is described in Sect. 3.1. In the model, photosynthetic cells are able to regulate their chlorophyll $a$ to carbon ratio in response to changes in irradiance, temperature and silicate (in the case of diatoms) by modifying the proportion of photosynthate that is directed towards chlorophyll biosynthesis ($\chi \rho$; see Eq. [10]). Balanced growth is achieved when cells are fully acclimated, in which case:

$$\frac{d}{dt} \left( \frac{\chi}{\rho} \right) = 0$$

(261)

Chlorophyll $a$ biosynthesis is assumed to be up-regulated in response to a reduction in irradiance and down regulated in response to an increase in irradiance. Through this process, cells are able to balance the rate of energy supply through light absorption, and energy demands for growth.
The maximum, light saturated photosynthesis rate $\chi_g(T)$ is assumed to be independent of changes in irradiance, which is consistent with observations which indicate Rubisco content is relatively invariant with respect to changes in irradiance (Sukenik et al., 1987), and the hypothesis that these cells are adapted to survive and reproduce in dynamic light environments (Talmy et al., 2014).

Using the ERSEM testing framework, it is possible to investigate this process in isolation. Model cells can be artificially acclimated to a given set of environmental conditions by finding a value for $q_{C:C}$ which satisfies Eq. (261). Figure 12 shows a plot of $q_{C:C}$ vs. $I_{PAR}$ for fully photo-acclimated diatoms in ERSEM. Cells were acclimated to a given irradiance by holding cellular carbon fixed and varying the cellular chlorophyll $a$ content within the range $\chi_{\text{min},C} \leq \chi \leq \chi_{\text{max}}$ in order to achieve balanced growth. Using the testing framework, the model can be compared with observations in order to sanity check the validity of the implementation, or parameterised against observations using curve fitting procedures. In Fig. 12 observations for the diatom T. Pseudonana have been overlaid. No attempt was made to fit the curve to this particular set of observations, although the fit appears reasonable. The parameter set is the same as used in the simulations of Sect. 7 and is given in the Supplement.

Diatoms are a physiologically and morphologically diverse group, which are characterised by their requirement for silicate, which they use to construct their cell wall. It is perhaps unsurprising that model fits to photosynthesis-irradiance curves for different diatom species result in a range of parameter values, including differences in the maximum light saturated carbon specific photosynthesis rate as a function of temperature, and the initial slope of the photosynthesis-irradiance curve (e.g. Geider et al., 1997). Ultimately, many of these differences arise due to differences in organism morphology and physiology, with, for example, different pigment complements or levels of investment in biosynthesis, being reflected in derived parameter values. These within group variations pose a perennial problem to the development of marine ecosystem and biogeochemical models. The diatom group in ERSEM is designed to be representative of diatoms as a whole, and to reflect the important biogeochemical role these organisms perform in nature.

ERSEM includes four phytoplankton functional groups: diatoms, which are characterised by their requirement for silicate, and three further groups which are characterised according to their size. These are the pico-, nano-, and microphytoplankton. The choice to characterise groups according to their size reflects the importance of size as a physiological trait (Litchman et al., 2007, 2010), which influences an organism’s competitive ability through its effect on nutrient acquisition, carbon and nutrient storage, the intracellular transport of solutes, photosynthesis rates through pigment packaging effects, and susceptibility to predation (e.g. Chisholm, 1992; Finkel et al., 2010).

Using ERSEM’s testing framework it is possible to demonstrate how this classification impacts the competitive ability of the four photosynthetic groups represented in the model. Figure 13 shows photosynthesis-irradiance curves for ERSEM’s four phytoplankton groups under the condition of balanced growth. As with the diatoms, the use of a single parameter set for each size-based group ignores within group variations that are observed in nature. It is important to take such abstractions into consideration when interpreting model outputs.
This example illustrates how ERSEM’s testing framework can be used to study and check the implementation of different processes within the code. Importantly, this is achieved without having to rewrite sections of the code in a second language with visualisation capabilities, which is an inherently error prone procedure. This capability is designed to complement the 0-D and 1-D drivers that simulate more complex time-varying environments, in which it is often difficult to study processes in isolation.

9 Optional choices

In the following section we provide an overview of the main optional choices in the model configuration. Options that involve major structural changes which alter the number of state variables or add substantial functionality are activated by preprocessor definitions, that need to be included at compile time. These include:

- The model of bacterial decomposition.
- The inclusion of the iron cycle.
- The light attenuation model.
- The calcification model.

Other options can be triggered at runtime via namelist parameters in the files include/ersem_pelagic_switches.nml and include/ersem_benthic_switches.nml without the need for a recompilation of the model code. These include the choice for the alkalinity description of the model and the choice of the benthic model.

9.1 The iron cycle

The use of the iron cycle in the model including growth limitation of phytoplankton by iron is activated by the preprocessor key IRON. It involves additional state variables for dissolved inorganic iron and iron components of the four phytoplankton types, two particulate matter types in the pelagic and one particulate matter type in the benthos.

9.2 Calcification

The use of the calcification sub-module (Sect. 3.6) is activated by the preprocessor key CALC. It’s computational impact is limited adding a single pelagic and a single benthic state to the list of state variables.

9.3 The model of bacterial decomposition

Two options are included for the modelling of the decomposition of organic matter (see Sect. 3.3). By default, the bacteria sub-model presented in Allen et al. (2002) with a basic microbial food-web and implicit decomposition is used. Enabling the preprocessing key DOCDYN the model for dynamic decomposition of organic matter is activated which uses fully explicit recycling of organic matter and includes the recalcitrant fraction of the DOC pool at the cost of an additional state variable.

9.4 The light attenuation model

Two options for light attenuation are available. The default choice is the legacy model based on apparent optical properties in the form of specific attenuation coefficients, while the recently developed model using inherent optical properties in the form of specific adsorption and backscatter coefficients and zenith angle needs to be activated by the IOPMODEL preprocessor key (see Sect. 3.9). The computational effort of the two models is comparable, but the latter involves the computation of the zenith angle and therefore requires the geographical coordinates and the current simulation date and time from the physical driver.

9.5 Alkalinity

The description of alkalinity in the model is given by the combination of two switches. The prognostic mode using an ocean tracer modified by biogeochemical processes affecting alkalinity is activated by setting ISWbioalk in include/ersem_pelagic_switches.nml to 1. The diagnostic mode deriving alkalinity from salinity (and optionally temperature) is enabled by activating an adequate alkalinity regression by setting ISWtalk to a value between 1 and 3. (The different regression options are specified in the Supplement.) The recommended use for these modes is a combination of both modes or the purely prognostic option with ISWbioalk = 1 and ISWtalk=0 (see Sect. 3.8).

9.6 The benthic model

The full benthic model (Sect. 4) is activated by setting the ibenXin parameter in include/ersem_benthic_switches.nml to 2, while for ibenXin = 1 (see Sect. 5.1.5) the benthic closure scheme is used. While the latter involves considerably less state variables and computations, the computational impact of this choice is largely negligible in 1D and 3D simulations as the computational cost is dominated by the advection and diffusion of the pelagic states.

10 Technical Specifications and Code Availability

The ERSEM 15.06 model is written in FORTRAN using the 2008 standard. Output is entirely based on netCDF and the output parsing scripts generating I/O FORTRAN code from plain text lists of variables are written in python.
The model is distributed under the open-source GNU Lesser General Public License through a gitlab server and freely available upon registration through the web-portal www.shelfseasmodelling.org. There are no restrictions or conditions for the registration of individual users, the registration is merely implemented in order to keep track of the user base. The code repository is fully version controlled (using git) and features a bug tracking system open to users. The release code of this publication is available in the master branch of the repository as tag ERSEM-15.06. The GOTM version used in the simulations of this work is also tagged as “ERSEM-15.06” on the ERSEM enabled fork of the development version of GOTM which can be downloaded from the same repository server. A quick start guide and user’s reference manual are also provided along with the code.

The versioning convention used with this software refers to the year and month of the release.

11 Conclusions

In this paper we have provided a full mathematical description of an updated version of ERSEM, one of the most established marine ecosystem models currently in use in the scientific community and in operational systems. Case studies ranging from a mesocosm type zero-dimensional experiment through three one-dimensional water column implementations to a brief three-dimensional full-scale example have illustrated the model dynamics in varying environments.

Quantitative and qualitative validation with in situ data for the basic ecosystem state variables chlorophyll a and the macronutrients has demonstrated the capability of the model to represent ecosystems ranging from oligotrophic open oceans to eutrophic coastal conditions. An integral validation of each single component would exceed the scope of this paper, the main purpose of which is the detailed description of the model ingredients as a reference for scientists, developers and users. Nevertheless, examples of component validations have been published previously and are available in literature (Artioli et al. 2012, Allen and Somerfield 2009, Allen et al. 2007, de Mora et al. 2013). In addition the testing framework supplied within the model distribution allows for targeted analysis and validation of individual parts of the model down to the level of single equations directly without rewriting or extracting the model code. We have demonstrated this capability here on the example of the PI-curve for phytoplankton growth.

The ERSEM 15.06 model is to our knowledge the only model currently available that provides the structure for simulating in one coherent system the biogeochemical cycles of carbon, the major macronutrients and iron (using variable stochiometric relationships), the carbonate system and calcification, the microbial food web and the benthic biogeochemistry.

While the range of processes included in the model brings the advantage of suitability for a whole range of applications as different as process studies, regional or global budgets of different chemical elements, habitat maps or risk assessment of environmental hazard, it also points to one of the major drawbacks of the model, i.e. a comparatively heavy structure and high number of parameters, that render it difficult to access for new users and hard to calibrate and parametrise. These problems are being addressed in a fully modular version of the model with streamlined process descriptions that is currently under development. It will allow for an arbitrary number of functional groups and easy replacement of individual sub-models, which can be tuned to the specific application at run-time. These developments will be made available with the next release of the model.

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