New isoquinoline alkaloid from *Carduus crispus* L.

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**Abstract:** A new isoquinoline alkaloid, along with other known 4 bioactive compounds (1 isoquinoline alkaloid, 2 flavonoids, 1 coumarin), were isolated from the ethanol extract of aerial parts of *Carduus crispus* L. Crispine A N-oxide was newly isolated compound, two of them (quercetin and rutin) were compounds isolated for the first time from indicated plant. The structures of these compounds were elucidated and confirmed with spectroscopic methods e.g. ¹H-NMR, ¹³C-NMR, EI-MS, COSY, HSQC, HMBC, and by comparison with literature data.

**Keywords:** ethanol extract, chromatography, chloroform, crispine A N-oxide

**Introduction**

The genus *Carduus* (*Compositae*) comprises 95 species all over the world [1]. Two species (*C.crispus* L. & *C.nutans* L.) are widely distributed in Mongolia [2]. Investigations of chemical constitutes showed that flavonoids, alkaloids and coumarins are the main components of this genus [1,3,4,5]. *C.crispus* is used in Mongolian traditional medicine for treatment of gland’s cancer and as analgestics [6]. It has been shown that this plant has antioxidant and cell wall strengthening activity. It has been established the cytotoxic activity of the alcoholic extract of *C.crispus*. [3].

Isoquinoline alkaloids (crispine A-E, carcrisine A, B) and flavone glycoside were isolated from this plant collected in Inner Mongolia have been reported [1,3,5]. Herewith, we report about the isolation and structural elucidation of a new isoquinoline alkaloid; crispine A N-oxide. The ¹H and ¹³C NMR spectral data of this compound are given for the first time.

**Experimental**

**Apparatus**

Melting point was determined on a Stuart SMP 10 apparatus. FABMS were measured on Bruker APEX II spectrometer. ¹H NMR and ¹³C NMR (400 MHz) spectra (all in CDCl₃) were recorded with a Bruker AM 400, using TMS as internal standard. Silica gel 60 (Merck 0.063-0.200 mesh) was used for column chromatography. Al₂O₃ plates were used for TLC. Plates were visualized by spraying with Dragendorff’s reagent.

**Plant material**

Aerial parts of *C. crispus* of its full flowering stage were collected in July 2003-2004, from Handgait place, near Ulaanbaatar city, Mongolia. A species was identified by Prof. Ch.Sanchir, Institute of Botany, Mongolian Academy of Sciences. Aerial parts of the
samples were dried at room temperature and grinded.

**Extraction and isolation**

The plant material (4.5 kg) was weighed and extracted with 95% ethanol three times. The ethanol extract was concentrated by reducing pressure to dryness. After was added 5% sulfuric acid to adjust pH 1 and thoroughly shaked. The yielding alkaloidal acidic solution was filtered and treated with NaHCO₃ to adjust pH 5. Basic solution was extracted with chloroform. Distillation of chloroform solution gave extract-A 25 g. Extracted with chloroform. Distillation of NaHCO₃ solution was filtered and treated with sulfuric acid to adjust pH 1 and thoroughly shaked. The yielding alkaloidal acidic solution was filtered and treated with NaHCO₃ to adjust pH 5. Basic solution was extracted with chloroform. Distillation of chloroform solution gave extract-A 25 g. Adding to acidic solution Na₂CO₃ to adjust pH 8 and following extraction with chloroform after distillation yield extract-B, 23 g. The extract-B was chromatographed over a silica gel with gradient CHCl₃–MeOH (100→99:1→70:30) to give 85 fractions. Fraction 35-39 were chromatographed on Al₂O₃ column eluted with CHCl₃–MeOH (98:2) to yield compound CC-1 (64 mg).

**Results and Discussion**

**The structural identification**

Isoquinoline alkaloids were isolated from the ethanol extract of the aerial parts of *C. crispus*. Compound CC-1, white needles, gave positive result in the Dragendorff test, mp. 218–220°C; molecular ion peak at m/z 248,2645 [M]+ (calcd for C₁₄H₁₉NO₃: 249.3081) was observed in the EI-MS spectrum. EIMS: m/z = 232, 231, 218, 202, 137, 113 and 81.

The ¹H NMR spectrum demonstrated signals for 19 protons in total. Two singlet at δH 6.46 (1H, s) and 6.58 (1H, s), indicating of the presence of 8, 9, 6a, 10a tetra substituted phenyl ring, might be attributed to aromatic protons (H-7 and H-10). The signals at δH 3.77 (3H, s) and 3.79 (3H, s) were assigned to two methoxy groups and the other 11 signals at δH 1.96 – 4.72 were due to aliphatic protons.

The ¹³C NMR indicated 14 signals, and DEPT experiment showed signals for two methoxy groups (δC 56.18, 56.02), five methylenes (δC 20.16, 25.07, 31.44, 58.39, 67.90), three methines (δC 76.40, 109.32, 110.97) and four quaternary carbons (δC 121.62, 124.31, 148.52, 148.72). The chemical structure of compound CC-1 was identified by comparison of this ¹H and ¹³C NMR spectral data with those of crispine A [3]. As shown in Table 1 carbon atoms C-3, C-5, C-10b linked to nitrogen atom of compound CC-1 have 10.19 -14.84 ppm. downfield chemical shifts in comparison with molecule of crispine A (Tabl.1).

**Table 1.** ¹H and ¹³C NMR (400MHz) and DEPT spectral data of Crispine A N-oxide and Crispine A (δ ppm)*

| Carbon atom | ¹³C     | DEPT   |
|-------------|---------|--------|
|             | Crispine A N-oxide | Crispine A |
| C-1         | 51.44   | 50.49  | CH₂   |
| C-2         | 20.16   | 22.17  | CH₂   |
| C-3         | 67.90 (+14,84) | 53.06  | CH₂   |
| C-5         | 58.39 (+10,19) | 48.20  | CH₂   |
| C-6         | 25.07   | 27.84  | CH₂   |
| C-7         | 110.97  | 111.22 | CH    |
| C-8         | 148.72  | 147.27 | C     |
| C-9         | 148.52  | 147.17 | C     |
| C-10        | 109.32  | 108.76 | C     |
| C-6a        | 121.62  | 126.03 | CH    |
| C-10a       | 124.31  | 130.63 | C     |
| C-10b       | 76.40 (+13,51) | 62.89  | CH    |
| C-OCH₃      | 56.18   | 55.81  | CH₃   |
| C-OCH₃      | 56.02   | 55.92  | CH₃   |

* CDCl₃ as solvent for CC-1.

Researchers have established that, chemical shifts of carbon atoms adjacent to N atom undergo a downfield shift due to oxidation of nitrogen compounds to N-oxides [7]. This proves conversion of nitrogen atom in crispine A into N-oxide form. Because of absence of any information in literature about crispine A N-oxide we assign that crispine A N-oxide is new naturally occurring compound (Fig.1).
Chemical method have used for provement of chemical structure of crispine A N-oxide. We have carried out reduction (deoxygenation) of crispine A N-oxide and convert it to crispine A. The spectral data of newly synthesized product was similar to spectral data of crispine A and proves that crispine A is a precursor of crispine A N-oxide.

**Method of deoxygenation of crispine A**

Crispine A N-oxide was dissolved in 5ml 5% sulfuric acid, added Zn powder, shaked permanently during 24 hours for providing chemical reaction. After end of reaction, the solution was filtered. Filtrate was alkalified by 5% sodium hydroxide to adjust pH 8-9 and was extracted by chloroform. Chloroform extract was concentrated in vacuo until dryness. Residue was chromatographed by using TLC with pure crispine A. Only one spot of crispine A have been detected in chromatogram, that prove that crispine A N-oxide have been completely converted to crispine A by deoxygenation reaction (Fig.2). It is known to be that this method widely used for deoxygenation of N-oxides [7].

**Conclusion**

New isoquinoline alkaloid has been isolated from the aerial parts of *C.crispus* L. It's structure was confirmed by chemical and spectral analysis and named as crispine A N-oxide.

**References**

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