Value of anti-p53 antibody as a biomarker for hepatocellular carcinoma
Evidence from a meta-analysis

Yue Chang, MS \(^{a,b,}\), Baiqing Liu, MS \(^{a,b,}\), Haiyan Niu, MS \(^{a,b,}\), Zhenguow Wang, MS \(^{b,}\), Shihai Xia, MD \(^{a,b,}\), Hai Li, MD \(^{a,b,}^{*}\)

Abstract

Introduction: The incidence of hepatocellular carcinoma (HCC) ranks sixth in the world, but its mortality is the third highest due to the lack of early diagnostic markers. Nowadays, the increase of autoantibody levels has been found in many cancers, and many studies have begun to pay attention to the detection of anti-p53 antibodies in HCC. The purpose of this study is to quantitatively and comprehensively analyze the potential diagnostic value of anti-p53 autoantibodies in HCC.

Methods: English articles up to November 2019 were collected. The overall sensitivity and specificity were calculated. Besides, the positive likelihood ratio, negative likelihood ratio (NLR), diagnostic odds ratio (DOR), and summary receiver operating characteristic curves of the overall diagnostic accuracy of anti-p53 antibody were calculated by STATA software. Finally, according to the heterogeneity of the results, the subgroup analysis, and the publication bias were performed.

Results: A total of 16 eligible studies were incorporated into this meta-analysis, including 1323 patients with HCC and 1896 control. The pooled sensitivity was 0.28 (0.17–0.41) and specificity was 0.98 (0.95–0.99). The pooled DOR was 10.44 (6.31–17.29) and the pooled NLR was 0.74 (0.63–0.86). The area under ROC curve of symmetrical ROC was 0.840.

Conclusions: The anti-p53 antibody has a high specificity for HCC, but the low sensitivity is not perfect and would limit the clinical application. The anti-p53 antibody would help rule out HCC but not help rule in HCC for early diagnosis. Whether combined as a diagnostic panel with other biomarkers or laboratory tests may prove useful requires further study.

Abbreviations: AFP = alpha-fetoprotein, AUC = area under ROC curve, CH = chronic hepatitis, DOR = diagnostic odds ratio, ELISA = enzyme-linked immunosorbent assay, FN = false negative, FP = false positive, HCC = hepatocellular carcinoma, LC = liver cirrhosis, LR = likelihood ratio, NHS = normal healthy serum, NLR = negative likelihood ratio, PLR = positive likelihood ratio, Sp = specificity, SROC = summary receiver operating characteristic curves, TN = true negative, TP = true positive.

Keywords: biomarker, diagnosis, hepatocellular carcinoma, meta-analysis, p53 antibody

1. Introduction

Hepatocellular carcinoma (HCC) is the sixth most common malignant tumor in the world, and most patients will die in 1 year after the clear diagnosis of HCC.\(^1\) The third high mortality rate of HCC is partly due to the lack of effective early diagnosis technology for HCC around the world. There is a high incidence of HCC, about 50.5% of new patients and 51.4% HCC-related deaths globally occur in China every year.\(^2\) In current clinical practice, alpha-fetoprotein (AFP) and imaging characteristics are the most commonly used modality for diagnosis and monitoring of HCC.\(^3,4\) However, AFP cannot be effective in differentiating HCC and other liver diseases, suggesting that the effect of screening is generally in the early diagnosis of HCC.\(^5\) As many as 40% of patients with normal AFP levels cannot be accurately detected in early stage and depends on definitive imaging results. This often leads to misdiagnosis or a delay in the correct diagnosis and management.\(^6\) Therefore, it is urgent to find a novel noninvasive or less invasive biomarker, known as “liquid biopsy,” to detect HCC at an early stage.\(^7\)

An increase in serum autoantibody levels has been shown to precede the occurrence and development of cancers. Immuno-logic processes causing autoantibody production are believed to be generated by the immune system in response to mutations, overexpression of proteins, degradation, or others.\(^8,9\) Therefore, there is a growing interest in autoantibody levels in patient blood...
2. Materials and methods

2.1. Literature search strategy

This meta-analysis was registered on PROSPERO (www.crd.york.ac.uk, ID: CRD42020171966). Two collectors (YC and BL) searched the studies on the serological detection of p53 antibody in human hepatocellular carcinoma in both PubMed, Cochrane library, and Embase database. To ensure the integrity of searches, we mainly used 3 keywords, “hepatocellular carcinoma,” “liver cell cancer,” “anti-p53 antibody,” “blood or serum,” which limits were the time of January 1990 to December 2019 and non-English articles in both databases. Also, the relevant references in the articles were put into our study to increase the integrity of the included literatures. The online Supplemental Digital Content presents our search strategies (see Table S1, Supplemental Content, which illustrates the search strategies used, http://links.lww.com/MD/E732).

2.2. Eligibility criteria

Figure 1 showed the search flow chart. All the records retrieved were completed independently by 2 different filters and disagreements were resolved by consensus. First of all, no important records, reviews, case studies, systematic reviews, studies of animal or cell lines, and other cancer literature studies are excluded. Then the inclusion criteria are as follows: The experimental group was diagnosed with HCC, and the clinical symptoms of the control group were different from that in the experimental group; anti-p53 antibody in serum or plasma of patients with liver cancer was detected using uncertain methods; the gold standard can be distinguished from the experimental group; methodology evaluation of anti-p53: the test method of anti-p53 antibody, sample source, the value of cutoff, TP, FP, FN, TN. Not available was used to represent the contents if they were not mentioned in the original articles, and it should be noted that we have not contacted the author for more detailed information.

2.3. Evaluation of quality of literature

QUADAS (quality assessment for studies of diagnostic accuracy), recommended by Cochrane, which can be used in systematic reviews of diagnostic accuracy studies, was used to evaluate the methodological quality of the included studies by 2 independent reviewers. If there were differences in the processing, we would track it through the third as well. The QUADAS tool is based on a consensus reached by the 9 experts in the field of diagnosis including 14 evaluation items, respectively, the disease spectrum, gold standard, disease progression bias, confirmation bias, clinical evaluation bias, combined bias, case withdrawal, uncertain results, and detailed guidelines for scoring each of the items included in the tool. The evaluation of the tool has 3 results: “yes,” “no,” “not clear.” If the result of the evaluation is “yes,” the score of 1 article will be increased by 1 point, otherwise, it would get none.

2.4. Information extraction

Table 1 is completed independently by 2 data filters. YC and BL were respectively extracted data from the including 14 literatures with the standard form. Some of the differences were discussed with both 2. In the present study, the following features mainly include. Basic information: the first author, published years, country, region the collection time of the sample; the characteristics of the experimental group, the gold standard, sample group sex ratio, age, tumor classification, etiology; the disease spectrum in the control group; methodology evaluation of anti-p53: the test method of anti-p53 antibody, sample source, the value of cutoff, TP, FP, FN, TN. Not available was used to represent the contents if they were not mentioned in the original articles, and it should be noted that we have not contacted the author for more detailed information.

2.5. Statistical analyses

Through the 2 meta-data analysis methods in the diagnostic tests, Se and Sp are calculated and the forest map and the ROC curve of the subjects for each data including articles were drawn. The potential problem associated with Se and Sp was handled by adding 0.5 values to all cells of the diagnostic 2 × 2 table. Also, we are very interested in calculating the positive likelihood ratio (PLR), negative likelihood ratio (NLR), diagnostic odds ratio (DOR), and the 95% confidence interval (95% CI) of them. Q test can be used statistically whether there is the presence of heterogeneity between different results of these studies, and I² can be used to reflect the effect size of the heterogeneity. I² < 25%, showed no heterogeneity; I² = 25% to 50%, indicating a moderate heterogeneity; I² > 75%, indicating a high degree of heterogeneity. To investigate the source of heterogeneity among different experiments, meta-regression and subgroup meta-analysis were done by our group. All statistical procedures were performed in Stata 15.1 (Corp. STATA, Station college, TX). The P values of the <.05 values of all the sets are statistically significant.

3. Results

3.1. Study characteristics and quality assessment

A total of 16 articles were included in this meta-analysis which was based on inclusion and exclusion criteria after completely searching. Table 1 shows the main information of the incoming
articles, there are 1323 patients with HCC and the control group of 1896 patients with non-HCC, including liver cirrhosis, hepatitis, or normal healthy people in the meta-analysis. The gold standard of 12 studies was identified by histopathology as HCC and the other 4 studies were determined by clinical information, imaging, and AFP. Six were from Asia, 6 studies were from Europe, 3 studies were from Africa, and only 1 from the United States in all the 16 studies. In the aspect of etiology, the main causes of HCC were mainly found by virus infection (hepatitis B virus [HBV], hepatitis C virus [HCV]), alcoholic liver disease, and some other unknown factors. In particular, the HCV infection rate of the 2 studies in Africa that reached 50% was much higher than in other regions. Tumor grade here does not represent the same classification, which uses Barcelona Clinic Liver Cancer (6 articles were used Child-Pugh graded, and the other 3 articles were graded by other classifications). The control group was mainly composed of liver cirrhosis, hepatitis, and healthy people, but each of the personnel structures is not consistent, 10 contains easily confused liver disease and 3 had normal healthy people in the control group. The average score of the studies was 9 points, 4 articles were more than 9 points, and the other 2 were less than 9 points. The remaining articles were all of 9 points in the full 14 points score of the QUADAS tool. None of studies mentioned the statement of potential conflict of interests.
### Table 1
Characteristics of all included studies.

| No. | Author (year)          | Country | Time                  | Gender (M/F) of HCC | Mean age  | Tumor stage | HBV/HCV/both/none/alcohol | Characteristics of control group | Characteristics of HCC group | Gold standard | Assay method | TP | FP | FN | TN | QUADAS |
|-----|------------------------|---------|-----------------------|---------------------|------------|-------------|---------------------------|---------------------------------|-----------------------------|---------------|--------------|----|----|----|----|--------|
| 1   | Volkmann et al (1993)  | EU      | 1983–1991             | 64/16               | 58.5       | NA          | 10/5/**/23/other           | Histology                       | 80                          | 29            | 35            | 3  |    |    |    | 20     |
| 2   | Ryder and Rizzi (1996) | EU      | 1985–1993             | NA                  | NA         | 23/14/24/8 | 19/10/5/**/4              | Image and AFP                   | 38                          | 34            | NA            | NA |    |    |    | 14     |
| 3   | Shiota et al (1997)    | Japan   | 1988–1995             | 69/17               | NA         | 104/57/120 | 28/46/11/1/42             | Histology                       | 86                          | 20            | 20            | NA |    |    |    | 28     |
| 4   | Raedle et al (1998)    | Germany | 1994–1996, 1994–1996  | NA                  | NA         | 19/26/22/13| NA                         | Histology                       | 75                          | 259           | 377           | NA |    |    |    | 17     |
| 5   | Saffroy et al (1999)   | EU      | 1994–1995             | 100/30              | 55         | 25/16/22/8 | 4/*/4/*/*                   | Histology                       | 78                          | 36            | NA            | 40 |    |    |    | 8      |
| 6   | Muller et al (2000)    | EU      | 1993–1991             | 64/16               | 59         | NA          | NA                        | History                         | 80                          | NA            | NA            | 80 |    |    |    | 19     |
| 7   | Loi et al (2006)       | China   | NA                    | NA                  | NA         | 51/32/22/14| NA                        | Other                           | 77                          | NA            | NA            | 82 |    |    |    | 7      |
| 8   | Zhang et al (2007)     | China   | NA                    | 116/26              | 57         | NA          | 108/16/*/*/*              | History                         | 142                         | 30            | 30            | 82 |    |    |    | 20     |
| 9   | Gadelhak et al (2009)  | Egypt   | 2001–2003             | NA                  | 54         | NA          | 2/*/*/*/*                  | Histology                       | 52                          | 27            | NA            | 10 |    |    |    | 38     |
| 10  | Chen et al (2010)      | China   | NA                    | 57/20               | 58         | 32/13/8    | 58/8/4/7/8              | History                         | 77                          | 30            | 30            | 82 |    |    |    | 14     |
| 11  | Liu et al (2012)       | China   | NA                    | 58/22               | 63.7       | NA          | 48/17/5/10/*              | Histology                       | 70                          | NA            | 50            | 70 |    |    |    | 9      |
| 12  | El Amr et al (2013)    | Egypt   | 2009–2012             | 22/29               | 52 ±11.7   | 57/18/7    | 26/16/8/*/*/*              | Image and AFP                   | 281                         | 20            | NA            | NA |    |    |    | 194    |
| 13  | Dai et al (2014)       | USA     | NA                    | 50/26               | 57 ±11.2   | 52/16/8/*/*/*             | Histology                       | 76                          | 30            | 30            | 89 |    |    |    | 10     |
| 14  | Middleton et al (2014) | EU      | NA                    | 78/18              | 65         | NA          | 4/24/*/16/23             | Other                           | 96                          | 73            | 97            | 96 |    |    |    | 7      |
| 15  | Abdel-Aziz et al (2016)| Saudi Arabia | NA                | 73/13              | 47         | 60/0/0     | 274/*/*/*/*              | History                         | 68                          | 18            | NA            | 20 |    |    |    | 45     |
| 16  | Attallah et al (2017)  | Egypt   | 2014–2015             | 87/33              | 56(50–102) | NA          | */120/*/*/*            | History                         | 60                          | 0             | 59            | NA |    |    |    | 47     |

CH = chronic hepatitis, ELISA = enzyme-linked immunosorbent assay, EU = Europe, F = female, FP = false positive, FN = false negative, HBV = hepatitis B virus, HCC = hepatocellular carcinoma, HCV = hepatitis C virus, LC = liver cirrhosis, M = male, NA = not available, No = number, QUADAS = quality assessment of diagnostic accuracy studies, TP = true positive, TN = true negative, WB = Western blot.
3.2. Diagnostic accuracy

There was found the sensitivity of range 6% to 88% with a large difference and the pooled specificity was almost 95% after the summary of the 16 articles. The pooled sensitivity was 0.28(95% CI: 0.17–0.41, \(P < .001, I^2 = 96.85\%\)), and the pooled specificity was 0.98 (95% CI: 0.95–0.99, \(P < .001, I^2 = 93.45\%\)) (Fig. 2). Generally, it is considered that the sensitivity and specificity have a great relationship with the value of cutoff and may be unfavorable for evaluating the accuracy of the diagnosis experiment as the main index in the current study. Because the heterogeneity of the \(I^2\) and \(Q\) tests showed a little large heterogeneity, the random effect model was adopted in these articles. From the results of Figure 3, we can find that the pooled DOR was 10.44, 95% CI (6.31–17.29), \(I^2 = 45.6\%\) \((P = .024)\), suggesting that including studies contained heterogeneity. Similarly, the pooled PLR was 11.53, 95% CI (5.86–22.69), \(I^2 = 79.4\%\), \(P < .001\), also showed a heterogeneity (see Fig. 4). The pooled NLR was 0.74 > 0.1, 95% CI (0.63–0.86), \(I^2 = 90.3\%\), and also showed a large heterogeneity. In Figure 5, the AUC of symmetrical ROC was 0.840 > 0.7, indicating the medium value of the anti-p53 antibody in the diagnostic for HCC.

3.3. Possible sources of heterogeneity

Meta-regression and the subgroup analyses were used to explore the possible sources of heterogeneity, which include the region, proportion of HBV patients, proportion of HCV patients, number of HCC patients, the stage I% of the participants, and the style of negative control group composition. Meta-regression indicated that the above variables region, the proportion of HBV patients, and tumor stage I% may be the sources of heterogeneity for anti-p53 antibody in HCC serum \((P < .001)\). The results of the subgroup analysis were shown in Table 2, which contained region, HCV, HBV, stage I%, number of HCC patients, and control style. It should be noted that the subgroup of African studies, tumor stage I<50%, HBV patients \(\geq 50\%\) had a low degree of heterogeneity. Besides this, subgroup performed in HCV patients proportion \(\geq 50\%\) has a higher sensitivity (0.58, 95% CI: 0.32–0.81), despite the heterogeneity.

3.4. Publication bias and sensitivity analysis

Overall, it may appear to be a certain degree of publication bias in different aspects of these studies. The most obvious is that the sensitivity of Asian and European studies showed lower than that in the African studies. However, Deeks’ funnel plots for publication bias also showed symmetry in Figure 6, indicating that these studies had no publication bias \((P = .270)\). Sensitivity analysis was conducted in terms of statistical analysis methods, sample size, QUADAS, and so on. However, the results produced no obvious changes by using a random effect model to replace the fix effect model.

4. Discussion

In the present study, 16 studies on the detection of serum anti-p53 antibody were collected for HCC by our group. According to the...
fndings, we could regard it as the first meta-analysis of anti-p53 for HCC. The detection of anti-p53 antibody in the diagnosis has a moderate value for hepatocellular carcinoma (AUC = 0.840) in the summary receiver operating characteristic curves (sROC), suggests that the anti-p53 antibody may become an effective marker for the diagnosis of HCC. However, the low sensitivity is not perfect and would limit the clinical application.

Usually, the pathological examination is the gold standard for HCC. However, liver cancer patients with the same clinic pathologic features often display different results, which suggest that there are several complex molecules and cells involved in the development of hepatocellular carcinoma. It’s still lack of effective diagnostic markers for the early diagnosis. Although the current tumor markers, such as AFP, AFP-L3, and DCP, play an important role in the diagnosis of HCC, they are mostly used in conjunction due to lack of specificity. Along with the antigens of tumor cells were easily changing, anti-tumor-associated antigens (TAA) may be detected before symptoms appear in 5 years, demonstrating that the body could be an immune reaction to that abnormal auto-antigen early. In the past few years, an increasing number of studies have focused on the immune response of the body to the tumor, and the detection of autoantibodies to tumor-associated antigens has been carried out in which the highest focused point is the anti-p53 antibody in various cancers.[15] As a tumor suppressor gene, the mutation of the p53 gene is found not only in HCC but also in many other cancers. The anti-p53 antibody is not specific for HCC, the other as esophageal carcinoma,[39] gastric carcinoma,[40] colorectal carcinoma,[41] and other cancer patients can be detected as well. P53 gene mutation and antibody can be coexisting, but not completely consistent in different cancers, suggesting that the anti-p53 antibody may be one of the diagnostic indicators of some cancers.

As a whole, the Sp of anti-p53 antibody for HCC is generally coming to 98% (P < .001), but Se had a wide range of 6% to 88% in each study (P < .001). The problem of heterogeneity between NLRs is a key point to our Meta-Analysis. A gold standard is one of the important steps of the experimental diagnosis. The 4 included articles of them did not fully use pathological to diagnosis which would inevitably affect the results of the experiment. Enzyme-linked immunosorbent assay (ELISA) is one of the most convenient and simple tests which is the only way to detect serum anti-p53 antibodies in these included studies excluding the western bolt. Another reason for heterogeneity is the difference of population selection in the control group. There were 2 studies only using the normal healthy samples as the control group significantly increased the ability of anti-p53 antibody in the diagnosis of HCC in different liver diseases.[26,28] In addition, it can be seen in the study of tumor grade (only 7 studies) that the diagnostic efficacy of anti-p53 in patients with primary hepatocellular carcinoma was slightly higher than that in late stage, but the difference between them was not obvious.
Other factors such as age, the sex ratio in the analysis of our meta-regression also express a certain significance. In our opinion, these factors mentioned above are the causes of heterogeneity. Although we try our best to find the heterogeneity, it cannot come to the completely reliable conclusion because of the lack of information. In fact, HCC is extremely heterogeneous cancer, so it cannot thoroughly eliminate the heterogeneity of Se, Sp, and NLRs in all studies.

In particular, the Se of anti-p53 antibodies in several African studies are much higher than the Asian studies, but their specificity was significantly lower than in other regions. Some studies show that the HCV core protein gene can induce the mutation of the p53 gene and HCV Core protein can be involved in the regulation of p53 gene expression, which leads to the generation of abnormal p53 protein in human body.\footnote{42}

We found that the HCV infection rate in Egyptian patients with liver disease leads to a relatively high positive rate, which is an important reason for the impact of heterogeneity.\footnote{21,34,43,44}

Simultaneously, HBV was one of the most high-risk factors of HCC which infected about 90% HCC patients in Asia especially China. Some researchers suggested that HBV X protein through inhibiting p53 gene expression\footnote{45} may be caused by the lower rate detection of anti-p53 antibody in the serum of HCC.

After analyzing the heterogeneity of the study, it cannot ignore the value of PLR, DOR (Fig. 4), and sROC curve (Fig. 5) of the

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**Figure 4.** Forest plot of estimates of the positive likelihood ratio (PLR) and negative likelihood ratio (NLR) for anti-p53-antibody in the diagnosis of HCC. The point estimates of the positive likelihood ratio from each study are shown as solid circles. Error bars are 95% confidence intervals. HCC = hepatocellular carcinoma.

**Figure 5.** The sROC for anti-p53 antibody in the serum in the diagnosis of HCC. Each solid circle represents each study in the meta-analysis. HCC = hepatocellular carcinoma, SROC = summary receiver operating characteristic curves.
Subgroup (n) Se (95% CI) Sp (95% CI) PLR (95% CI) NLR (95% CI) DOR (95% CI) I²

Region
Asian (6) 0.19 (0.10, 0.34) 0.98 (0.91, 1.00) 9.16 (3.02, 27.40) 0.83 (0.73, 0.94) 11.97 (4.33, 32.68) 97.95
EU and USA (7) 0.16 (0.12, 0.20) 0.99 (0.95, 1.00) 20.23 (5.51, 72.2) 0.83 (0.76, 0.90) 24.49 (475, 15332) 80.95
African (3) 0.71 (0.66, 0.75) 0.91 (0.84, 0.95) 7.05 (5.12, 15.69) 0.31 (0.21, 0.44) 24.48 (586, 71.06) 12.50

Gold standard
Histology (12) 0.23 (0.14, 0.35) 0.98 (0.95, 1.00) 12.06 (4.83, 29.61) 0.76 (0.60, 0.90) 15.94 (6.95, 38.24) 97.95
Image and AFP (4) 0.45 (0.14, 0.76) 0.96 (0.86, 0.99) 11.16 (4.58, 25.65) 0.57 (0.33, 1.01) 19.77 (7.01, 59.31) 95.95

Stage
≥50% (5) 0.37 (0.15, 0.66) 0.95 (0.90, 0.98) 7.35 (4.24, 12.96) 0.66 (0.44, 1.01) 11.04 (5.20, 24.64) 95.95
<50% (3) 0.14 (0.10, 0.20) 0.98 (0.96, 1.00) 5.71 (2.77, 11.64) 0.87 (0.82, 0.92) 7.91 (3.89, 14.86) 0.00

HBV
≥50% (6) 0.38 (0.18, 0.63) 0.98 (0.88, 1.00) 15.80 (4.83, 28.52) 0.64 (0.44, 0.91) 25.74 (6.63, 91.31) 97.95
<50% (5) 0.18 (0.13, 0.26) 0.99 (0.95, 1.00) 20.23 (5.51, 72.2) 0.83 (0.76, 0.90) 24.49 (475, 15332) 80.95

HCV
≥50% (5) 0.58 (0.32, 0.81) 0.90 (0.76, 0.96) 5.74 (2.63, 12.68) 0.47 (0.26, 0.64) 12.63 (4.58, 35.55) 96.95
<50% (6) 0.16 (0.13, 0.25) 0.99 (0.95, 1.00) 28.22 (5.44, 22.47) 0.82 (0.76, 0.89) 34.14 (4.69, 29.14) 55.95

No. of HCC
≥80 (6) 0.25 (0.13, 0.44) 0.99 (0.89, 1.00) 47.43 (2.21, 1043.59) 0.53 (0.35, 0.81) 70.55 (5.19, 1094.47) 98.95
<80 (10) 0.29 (0.15, 0.49) 0.96 (0.92, 0.98) 8.37 (4.63, 15.04) 0.73 (0.58, 0.92) 11.20 (6.06, 22.16) 97.95

Control style
Health + Disease (10) 0.48 (0.36, 0.70) 0.99 (0.74, 1.00) 37.41 (2.02, 709.84) 0.53 (0.35, 0.81) 70.55 (5.19, 1094.47) 98.95
80 (6) 0.29 (0.15, 0.49) 0.96 (0.92, 0.98) 8.37 (4.63, 15.04) 0.73 (0.58, 0.92) 11.20 (6.06, 22.16) 97.95
50% (8) 0.23 (0.13, 0.37) 0.98 (0.94, 0.99) 9.68 (4.32, 21.88) 0.79 (0.69, 0.91) 12.61 (5.47, 28.31) 97.95
80 (10) 0.29 (0.15, 0.49) 0.96 (0.92, 0.98) 8.37 (4.63, 15.04) 0.73 (0.58, 0.92) 11.20 (6.06, 22.16) 97.95
50% (8) 0.18 (0.13, 0.25) 0.99 (0.95, 1.00) 28.22 (5.44, 22.47) 0.82 (0.76, 0.89) 34.14 (4.69, 29.14) 55.95
50% (3) 0.14 (0.10, 0.20) 0.98 (0.96, 1.00) 5.71 (2.77, 11.64) 0.87 (0.82, 0.92) 7.91 (3.89, 14.86) 0.00

5. Conclusion
Current evidence suggests that the anti-p53 antibody may be a potential indicator of potential value in the diagnosis of hepatocellular carcinoma, and has a certain clinical value. Despite the high specificity of the anti-p53 antibody for HCC, the low sensitivity is not perfect and would limit the clinical application. The anti-p53 antibody would help rule out HCC but not help rule in HCC for early diagnosis. Thus, a combination of panels of TAAs to detect multiple different and specific antibodies is the current goal to increase their diagnostic potential for HCC.

Author contributions
YC and HL contributed to the study design. YC and BQL generated the literature strategy and filtered through the identified studies. HYN and SHX evaluated study quality. Study results were entered in duplicate by HL, SHX, and YC. All authors read and approved the final manuscript.

References
[1] Villanueva A. Hepatocellular carcinoma. N Engl J Med 2019;380:1450–62.
[2] Hong Y, Huang J. Autoantibodies against tumor-associated antigens for detection of hepatocellular carcinoma. World J Hepatol 2015;7:1581–5.
[3] Sauzay C, Petit A, Bourgeois AM, et al. Alpha-fetoprotein (AFP): a multi-purpose marker in hepatocellular carcinoma. Clin Chim Acta 2016;463:39–44.
[4] De Stefano F, Chacon E, Turcios L, et al. Novel biomarkers in hepatocellular carcinoma. Dig Liver Dis 2018;50:1115–23.
[5] Ertle JM, Heider D, Wichert M, et al. A combination of alpha-feto protein and des-gamma-carboxy prothrombin is superior in detection of hepatocellular carcinoma. Digestion 2013;87:121–31.
[6] Sutherland T, Watts J, Ryan M, et al. Diffusion-weighted MRI for hepatocellular carcinoma screening in chronic liver disease: direct comparison with ultrasound screening. J Med Imaging Radiat Oncol 2017;61:34–9.
Attallah AM, Shiha GE, Ismail H, et al. Expression of p53 protein in liver carcinoma. J Transl Med 2014;12:121.

Wang T, Liu M, Zheng SJ, et al. Tumor-associated autoantibodies are useful biomarkers in immunodiagnosis of alpha-fetoprotein-negative hepatocellular carcinoma. World J Gastroenterol 2017;23:3496–504.

Atallah AM, Shiu GE, Ismail H, et al. Expression of p53 protein in liver and sera of patients with liver fibrosis, liver cirrhosis or hepatocellular carcinoma associated with chronic HCV infection. Clin Biochem 2009;42:455–61.

Lei QQ, Liu JW, Zheng H. Potential role of anti-p53 antibody in diagnosis of lung cancer: evidence from a bivariate meta-analysis. Eur Rev Med Pharmacol Sci 2013;17:3012–8.

Qi LN, Bai T, Chen ZS, et al. The p53 mutation spectrum in hepatocellular carcinoma from Guangxi, China: role of chronic hepatitis B virus infection and aflatoxin B1 exposure. Liver Int 2015;35:999–1009.

Soussi T. The humoral response to the tumor-suppressor gene product p53 in human cancer: implications for diagnosis and therapy. Immunol Today 1996;17:354–6.

Soussi T. P53 antibodies in the sera of patients with various types of cancer: a review. Cancer Res 2000;60:1777–88.

Whiting P, Rutjes AW, Reitsma JB, et al. The development of QUADAS: a tool for the quality assessment of studies of diagnostic accuracy included in systematic reviews. BMC Med Res Methodol 2003;3:25.

Chu H, Guo H. A unification of models for meta-analysis of diagnostic accuracy studies. Biostatistics 2009;10:201–3.

Higgins JP, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. BMJ 2003;327:557–60.

Middleton CH, Irving W, Robertson JF, et al. Serum autoantibody measurement for the detection of hepatocellular carcinoma. PLoS One 2014;9:e103867.

Dai L, Ren P, Liu M, et al. Using immunoinformatic approach to enhance tumor-associated autoantibody detection in diagnosis of hepatocellular carcinoma. Clin Immunol 2014;152:127–39.

El Azm AR, Yousel M, Salah R, et al. Serum anti-P53 antibodies and alpha-fetoprotein in patients with non-B non-C hepatocellular carcinoma. Springerplus 2013;2:69.

Liu H, Zhang J, Wang S, et al. Screening of autoantibodies as potential biomarkers for hepatocellular carcinoma by using T7 phase display system. Cancer Epidemiol 2012;36:82–8.

Chen Y, Zhou Y, Qiu S, et al. Autoantibodies to tumor-associated antigens combined with abnormal alpha-fetoprotein enhance immunodiagnosis of hepatocellular carcinoma. Cancer Lett 2010;289:32–9.

Gadeltakh NA, Gadelkh SA, El-Morsi DA, et al. Prognostic significance of three hepatitis markers (p53 antibodies, vascular endothelial growth factors and alpha fetoprotein) in patients with hepatocellular carcinoma. Hepatogastroenterology 2009;56:1417–24.

Zhang JY, Meglinoirro R, Peng XX, et al. Antibody detection using tumor-associated antigen mini-array in immunodiagnosing human hepatocellular carcinoma. J Hepatol 2007;46:107–14.

Looi K, Meglinoirro R, Shi FD, et al. Humoral immune response to p16, a cyclin-dependent kinase inhibitor in human malignancies. Oncol Rep 2006;16:1105–10.

Saffroy R, Leong JC, Azoulay D, et al. Clinical significance of circulating anti-p53 antibodies in European patients with hepatocellular carcinoma. Br J Cancer 1999;79:604–10.

Muller M, Meyer M, Schilling T, et al. Testing for anti-p53 antibodies increases the diagnostic sensitivity of conventional tumor markers. Int J Oncol 2006;29:973–80.

Radele J, Oremek G, Truschwotnch M, et al. Clinical evaluation of autoantibodies to p53 protein in patients with chronic liver disease and hepatocellular carcinoma. Eur J Cancer 1998;34:1198–203.

Shiota G, Kishimoto Y, Suyama A, et al. Prognostic significance of serum anti-p53 antibody in patients with hepatocellular carcinoma. J Hepatol 1997;27:661–8.

Ryder SD, Rizzi PM, Volkmann M, et al. Use of specific ELISA for the detection of antibodies directed against p53 protein in patients with hepatocellular carcinoma. J Clin Pathol 1996;49:295–9.

Volkmann M, Muller M, Hofmann WJ, et al. The humoral immune response to p53 in patients with hepatocellular carcinoma is specific for malignancy and independent of the alpha-fetoprotein status. Hepatology 1993;18:559–65.

Atallah AM, El-Far M, Abdelrezak MA, et al. Combined use of nuclear phosphoprotein c-Myc and cellular phosphoprotein p53 for hepatocellular carcinoma detection in high-risk chronic hepatitis C patients. Br J Biomed Sci 2017;74:170–5.

Abdel-Aziz MM, Elshal MF, Abas AT, et al. Comparison of AFP-L3 and p53 antigen concentration with alpha-fetoprotein as serum markers for hepatocellular carcinoma. Clin Lab 2016;62:1121–9.

Mann J, Reeves HL, Feldstein AE. Liquid biopsy for liver diseases. Gut 2018;67:2204–12.

Shakado S, Sakisaka S, Chayama K, et al. Alpha-fetoprotein and des-gamma-carboxy-prothrombin at twenty-four weeks after interferon-based therapy predict hepatocellular carcinoma development. World J Hepatol 2015;7:2757–64.

Sakamoto N. NX-PVKA assay, a conventional but refined prognostic biomarker for hepatocellular carcinoma. J Gastroenterol Hepatol 2013;28:755–6.

Dohi C, Nousu K, Miyahara K, et al. Potential of alpha-fetoprotein as a prognostic marker after curative radiofrequency ablation of hepatocellular carcinoma. Hepatol Res 2015;46:916–23.

Blanchard P, Quero L, Piacuti V, et al. Prognostic significance of anti-p53 and anti-KRas circulating antibodies in esophageal cancer patients treated with chemoradiotherapy. BMC Cancer 2012;12:119.

Kunizaki M, Fukuda A, Wakata K, et al. Clinical significance of serum p53 antibody in the early detection and poor prognosis of gastric cancer. Anticancer Res 2017;37:1979–84.

Liu S, Tan Q, Song Y, et al. Anti-p53 autoantibody in blood as a diagnostic biomarker for colorectal cancer: a meta-analysis. Scand J Immunol 2019;91:e12829.

Swiatek-Koscielna B, Kaluzna EM, Januszkiewicz-Lewandowska D, et al. HCV infection and interferon-based treatment induce p53 Gene transcription in chronic hepatitis C patients. Viral Immunol 2015;28:434–41.

Atta MM, El-Masry SA, Abdel-Hameed M, et al. Value of serum anti-p53 antibodies as a prognostic factor in Egyptian patients with hepatocellular carcinoma. Clin Biochem 2008;41:1131–9.

El-Emshaty HM, Gadelkh SA, Abdelaziz MM, et al. Serum P53 Abs in HCC patients with viral hepatitis—type C. Hepatogastroenterology 2014;61:1688–95.

Yeom S, Jeong H, Kim SS, et al. Hepatitis B virus X protein activates proapoptotic activator 28 gamma expression via upregulation of p53 levels to stimulate virus replication. J Gen Virol 2018;99:655–66.

Parasole R, Izzo F, Perrone F, et al. Prognostic value of serum biological markers in patients with hepatocellular carcinoma. Clin Cancer Res 2001;7:3504–9.

Siriuk V, Vayss J, Chevre S, et al. Prevalence and prognostic value of serum anti-p53 antibody-ies in hepatocellular carcinoma. A study of 159 patients. Gastroenterol Clin Biol 2000;24:1159–63.

Muller M, Meyer M, Schilling T, et al. Testing for anti-p53 antibodies increases the diagnostic sensitivity of conventional tumor markers. Int J Oncol 2006;29:973–80.

Chaturkun N, Tangkijvanich P, Voravud N, et al. Clinical significance of p53 antigen and anti-p53 antibodies in the sera of hepatocellular carcinoma patients. J Gastroenterol 2001;36:830–6.