BPC 157 as a Therapy for Retinal Ischemia Induced by Retrobulbar Application of L-NAME in Rats

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Providing NO-system importance, we suggest that one single application of the NOS-blocker L-NAME may induce retinal ischemia in rats, and that the stable pentadecapeptide BPC 157 may be the therapy, since it may interact with the NO-system and may counteract various adverse effects of L-NAME application. A rat retinal ischemia study was conducted throughout 4 weeks, including fundoscopy, behavior presentation, tonometry, and histology assessment. Retrobulbar L-NAME application (5 mg/kg; 0.5 mg/0.1 ml saline/each eye) in rats immediately produced moderate generalized irregularity in the diameter of blood vessels with moderate atrophy of the optic disc and faint presentation of the choroidal blood vessels, and these lesions rapidly progressed to the severe stage. The specific L-NAME–induced vascular failure points to normal intraocular pressure (except to very transitory increase upon drug retrobulbar administration). When BPC 157 (10 μg; 10 ng/kg, as retrobulbar application, 1 μg; 1 ng/0.1 ml saline/each eye) is given at either 20 min after L-NAME or lately, at 48 h after L-NAME, the regular retrobulbar L-NAME injection findings disappear. Instead, fundoscopy demonstrated only discrete generalized vessel caliber irregularity with mild atrophy of the optic disc, and then, quite rapidly, normal eye background and choroidal blood vessels, which remain in all of the subsequent periods. Also, histology assessment at 1, 2, and 4 weeks shows that BPC 157 counteracted the damaged inner plexiform layer and inner nuclear layer, and revealed normal retinal thickness. The poor behavioral presentation was also rescued. Thus, while further studies will be done, BPC 157 counteracted L-NAME–induced rat retinal ischemia.

Keywords: retina, ischemia, BPC 157, L-NAME, rats

INTRODUCTION

We focused on the stable gastric pentadecapeptide BPC 157 (Sikiric et al., 2014; Sikiric et al., 2018; Sikiric et al., 2020) and retinal ischemia in rats. Reasonably, providing an essential physiological role of the nitric oxide (NO) system (Cudeiro and Rivadulla, 1999; Toda and Nakanishitoda, 2007; Guthrie and Kang-Mieler, 2014), its inhibiting may directly lead to the retinal ischemia and definitively debilitated function.
Thus, we suggest that one single application of the NO-synthase (NOS) blocker N(G)-nitro-L-arginine methyl ester (L-NAME) may induce retinal ischemia in rats, and that the stable pentadecapeptide BPC 157 may be the therapy, since it may interact with the NO-system and may counteract various adverse effects of L-NAME application (see Sikiric et al., 2014). This particular point supports the recent evidence that vasomotor tone is carried out through BPC 157-specific activation of the Src-Caveolin-1-endothelial NOS (eNOS) pathway (Hsieh et al., 1997). Likewise, BPC 157 has a modulatory role (Sikiric et al., 2014) (in particular, to the blood pressure (Sikiric et al., 1997) and thrombocytes function maintenance (Stupnisek et al., 2012,2015; Konosic et al., 2019)) and additional interaction with several molecular pathways (Tkalcic et al., 2007; Chang et al., 2011, 2014; Hsieh et al., 2017; 2020; Kang et al., 2018; Vukojevic et al., 2018; Vukojevic et al., 2020; Park et al., 2020; Wu et al., 2020) [i.e., activation of the VEGFR2-Akt-eNOS signaling pathway without the need of other known ligands or shear stress (Hsieh et al., 2017)]. To illustrate its modulatory role on NO-system functions (Sikiric et al., 2014), BPC 157 may counteract L-NAME hypertension and prothrombotic effect as well as L-arginine hypotension and antiangiotic effect (Sikiric et al., 1997; Stupnisek et al., 2015), and induce its own NO-release (Sikiric et al., 1997; Turkovic et al., 2004). Also, BPC 157 may be the free radical scavenger to counteract reperfusion-induced free radical injury (Belusic Halle et al., 2017; Duzel et al., 2017; Luetic et al., 2017; Amic et al., 2018; Drmic et al., 2018; Vukojevic et al., 2018; Sever et al., 2019; Sucic et al., 2019; Kolovrat et al., 2020; Park et al., 2020). In addition, BPC 157, as an agent with cytoprotective effect in the entire gastrointestinal tract (and thereby, innate endothelium protection) (Sikiric et al., 2018; Sikiric et al., 2020), prevents and reverses thrombosis formation (Hrelec et al., 2009; Vukojevic et al., 2018; Gojkovic et al., 2020; Kolovrat et al., 2020). The evidence is provided after abdominal aorta anastomosis (Hrelec et al., 2009) or major vein occlusion (Vukojevic et al., 2018; Gojkovic et al., 2020; Kolovrat et al., 2020). Also, BPC 157 maintains thrombocytes function (Konosic et al., 2019) without interfering with coagulation pathways (Konosic et al., 2019). Finally, when confronted with major vessel occlusion, BPC 157 administration recruits vessels to rapidly activate the collateral pathway (Duzel et al., 2017; Amic et al., 2018; Drmic et al., 2018; Vukojevic et al., 2018; Cesar et al., 2020; Gojkovic et al., 2020; Kolovrat et al., 2020). These responses effectively negate the harmful effect of L-NAME–NOS blockade (Duzel et al., 2017; Amic et al., 2018; Drmic et al., 2018; Cesar et al., 2020). Furthermore, there is evidence that BPC 157 counteracted stroke, given in reperfusion, after clamping of the common carotid arteries (i.e., both early and delayed neural hippocampal damage, and achieving full functional recovery (the Morris water maze test, the inclined beam-walking test, and the lateral push test)) (Vukojevic et al., 2020). Likewise, BPC 157 may ameliorate other peripheral and central neuronal damage (Ilic et al., 2009; Ilic et al., 2010; Ilic et al., 2011a; Ilic et al., 2011b; Gjurasin et al., 2010; Tudor et al., 2010; Kliceck et al., 2013; Sikiric et al., 2013; Sikiric et al., 2016; Lojo et al., 2016; Drmic et al., 2017; Medvidovic-Grubisic et al., 2017; Perovic et al., 2019; Wang et al., 2019). This may suggest that BPC 157 administration would counteract retinal disturbances also.

In the previous eye research studies, BPC 157 counteracts atropine-mydriasis, but it also opposes an immediate and hour-lasting miotic effect of L-NAME in rats and guinea pigs, and participates in pupil control potentially via NO-mediated and cholinergic mechanisms (Kokot et al., 2016). Likewise, there are the adverse effects of NO-substrate and L-arginine–induced NO overstimulation, which may be counteracted by the application of BPC 157 as well (Kokot et al., 2016). BPC 157 provided a particular healing and vascular effect since it maintained corneal transparency (Lazic et al., 2005; Masnec et al., 2015). Illustratively, total debridement of corneal epithelium as well as perforating corneal incisions (Masnec et al., 2015) was cured with no corneal neovascularization and no new vessels (Lazic et al., 2005; Masnec et al., 2015).

This may be essential for the effective counteraction of the damaging effect of the retrobulbar L-NAME application. In support, intravenous or intravitreous L-NAME application, given before, would aggravate the harmful effect of the major noxious procedure (i.e., ischemia by increasing intraocular pressure above systolic pressure by the infusion into the anterior chamber; ligating optic nerve) (Imai et al., 1997; Ostwald et al., 1997; Hangai et al., 1999; Sakamoto et al., 2006).

Therefore, by retrobulbar L-NAME (and/or BPC 157 application as therapy), we investigated the degree of ischemia of the retina with fundus images of both eyes, from normal presentation until the strong generalized irregular diameter of blood vessels with severe atrophy of the optic disc, and extremely barely visible (extremely faint presentation) choroidal blood vessels (bright fundus background color). Upon sacrifice, we assessed retinal disturbances, particularly those in the inner nuclear layer and the inner plexiform layer. Likewise, after retrobulbar L-NAME application, we focused on the corresponding animal behavior. The contribution of intraocular pressure elevation in the procedure was also evaluated. BPC 157 was given in the early post-L-NAME time, or later, in a more advanced injury course, at 48 h after L-NAME.

**MATERIALS AND METHODS**

**Animals**

Study protocols were conducted in male albino Wistar rats, body weight 200 g, 13 weeks old, in-house bred—animal facility, Department of Pharmacology, School of Medicine, Zagreb, Croatia. Animal facility is registered by the directorate of veterinary, Reg. No: HR-POK-007. Laboratory rats were acclimated for 5 days and randomly assigned to their respective treatment group. Laboratory animals were housed in PC cages in conventional laboratory conditions at the temperature of 20–24°C, relative humidity of 40–70%, and noise level of 60 dB. Each cage was identified using the following data: number of study, group, dose, number, and sex.
of each animal. Fluorescent lighting provided illumination 12 h per day. Standard good laboratory practice diet and fresh water were provided ad libitum. Animal care was in compliance with standard operative procedures of Pharmacology Animal facility, the European convention for the protection of vertebrate animals used for experimental and other scientific purposes (ETS 123). Ethical principles of the study ensured compliance with European Directive 2010/63/E, the Law on Amendments to the Animal Protection Act (Official Gazette 37/13, the Animal Protection Act (official Gazette 135/06), Ordinance on the protection of animals used for scientific purposes (Official Gazette 55/13), FELASA recommendations, and recommendations of the Ethics Committee, School of Medicine, University of Zagreb. All experiments received specific approval from the local Ethics Committee at the School of Medicine (the University of Zagreb, Zagreb, Croatia). We randomly assigned 10 rats per experimental group and period for all experiments.

Drugs
Pentadecapeptide BPC 157 (manufactured by Diagen, Ljubljana, Slovenia) (Ilic et al., 2009; Ilic et al., 2010; Ilic et al., 2011a; Ilic et al., 2011b; Gjurasin et al., 2010; Tudor et al., 2010; Klicek et al., 2013; Sikiric et al., 2013; Sikiric et al., 2016; Lojo et al., 2016; Drmic et al., 2017; Medvidovic-Grubisic et al., 2017), GEPPPGKPADDAGLV, M.W. 1,419, partial sequence of human gastric juice protein BPC, peptide with 99% high-pressure liquid chromatography (HPLC) purity, expressing 1-des-Gly peptide as an impurity, freely soluble in water at pH 7.0 and in saline) was dissolved in distilled water. L-NAME (Sigma United States) was commercially purchased.

Procedure
Before retrobulbar application and/or fundus imaging (filmed with USB microscope camera “Velho Discovery VMS-004 Deluxe” and “VOLK” Digital Wide Field Lens for indirect ophthalmoscopy), the animals were deeply anesthetized with diazepam (5 mg/kg b.w. i.p.) and sodium thiopental (5 mg/kg b.w. i.p.) were used for anesthesia.

Alternatively, to perceive the effect of the given medication itself (without L-NAME), rats received only BPC 157 (total dose 10 μg/kg or 10 ng/kg as retrobulbar application in each eye, 1 μg/0.1 ml saline/each eye, or 1 ng/0.1 ml saline/each eye) or only saline via retrobulbar application in each eye (0.1 ml/each eye) (as an additional control sham group) at the corresponding time point.

Assessment procedure (the fundus images were filmed for 1 min) included several specific time points as follows: i) normal rats before application; ii) before L-NAME retrobulbar administration and immediately thereafter; iii) when the rats underwent early treatment at 20 min after L-NAME administration: before medication (a) and after medication (b, c, d, e, f, and g), immediately after (b), at the days 1 (c), 2 (d), 7 (e), 14 (f), and 28 (g) L-NAME time; iv) these mentioned points were also included in the assessment of the rats which only underwent BPC 157 or saline retrobulbar administration; and v) in the rats, which underwent delayed therapy, at 48 h post-L-NAME administration: immediately before medication (A) and after application (B, C, D, E, F, and G), immediately after (B), and after 1 and 2 days, corresponding to day 3 (C) and day 4 (D) L-NAME time, and at the days 7 (E), 14 (F), and 28 (G) L-NAME time.

During recording, the animals received artificial tears (Isopto Tears, Alcon Pharmaceuticals, United Kingdom) in the eyes. Moistening of the cornea allows better visualization of the fundus and clearer images of the retina.

The degree of ischemia of the retina was scored 1–4 by taking images of the animals fundus as follows: 1: orderly eye background, normal presentation of the choroidal blood vessels (normal reddish fundus background color); 2: discrete generalized irregularity in the diameter of the blood vessels with mild atrophy of the optic disc, normal presentation of the choroidal blood vessels (normal reddish fundus background color); 3: moderate generalized irregularity of the diameter of blood vessels with moderate atrophy of the optic disc, barely visible (faint presentation) choroidal blood vessels (brighter fundus background color); and 4: strong generalized irregularity of the diameter of the blood vessels with severe atrophy of the optic disc, barely visible (extremely faint presentation) choroidal blood vessels (bright fundus background color). The images were processed with software purchased with the USB microscope camera “Velho Discovery VMS-004 Deluxe”.

Behavior
The behavior of the animals was recorded from the second day, after the last retrobulbar application. To illustrate animal behavior after retrobulbar application of the agents, each of the rats was mounted on an elevated surface (50 cm long, 20 cm width, and 30 cm high) and filmed for 1 min. We recorded the number of rats that maintained the initial position of firmly standing on the surface with posterior legs, with only limited movements around, as well as the number of those demonstrating no freezing behavior, quick exploration, and escaping from the elevated surface.
Intraocular Pressure Measuring

Intraocular pressure was measured using a calibrated applanation tonometer Tonopen XL by Reichert Technologies (the transducer was placed on the same spot of the cornea for each measurement). The rats were deeply anesthetized with diazepam (5 mg/kg b.w. i.p.) and sodium thiopental (5 mg/kg b.w. i.p.), and furthermore, tetracaine drops were administered in both eyes. Intraocular pressure was assessed in normal rats, and values between 13 and 19 mmHg were considered normal, in accordance with previous rat data (Mermoud et al., 1994).

To assess the effect of retrobulbar administration itself, the values were assessed before retrobulbar application, after retrobulbar application of saline (0.1 ml/each eye), BPC 157 1 μg/0.1 ml saline/each eye, or L-NAME 0.5 mg/0.1 ml saline/each eye, at 1, 2, 4, 6, 8, and 10 min thereafter.

To evaluate the effect of the L-NAME–induced retinal injury protocol and the medication given after L-NAME administration, intraocular pressure was assessed before L-NAME retrobulbar administration as well as thereafter at described intervals.

For the early treatment at 20 min, we assessed intraocular pressure before medication as well as thereafter at described intervals. Further intraocular pressure assessment was performed at days 1 and 2, and after 1, 2, and 4 weeks.

For the delayed therapy, intraocular pressure was assessed as described above, at 48 h post–L-NAME administration, before medication application, and thereafter at described intervals. We performed further intraocular pressure assessment at 20 min after medication, after days 1 and 2, and after 1, 2, and 4 weeks.

Besides the initial short-lasting increase of the intraocular pressure upon retrobulbar application, intraocular pressure was continuously normal, and the data were not specifically shown.

Microscopy

In the histopathological evaluation at the end of 1, 2, 3, and 4 weeks, the enucleated eyes were fixed in 4% phosphate-buffered formaline, and transverse sections passing through the optic nerve were obtained. The specimens were processed on paraffin wax, and 5-μm-thick paraffin sections were obtained and stained with hematoxylin and eosin or immunohistochemically for FVIII (Dako, Glostrup, Denmark) using Dako Autostainer, according to the manufacturer’s protocols. For the thickness of the retinal tissue, internal plexiform layer, and internal nuclear layer, the measurements (ISSA program (VAMSTEC, Zagreb, Croatia) from the area of maximal tissue damage detected using semi-serial sections, five high-power fields were randomly selected for analysis, were made with an ocular micrometer at 20 magnification within 0.5 mm from the optic disc. Signs of histopathologic changes such as edema, vacuolar degeneration and pyknosis, and polymorphonucleated leucocytes (PMNL) infiltration were noted if present. The histological examination was performed by the pathologist, who was unaware of the treatment.

Statistical Analysis

We used Statistica 12.1. for Windows to perform statistical analysis. Distribution of data normality was tested by the Kolmogorov–Smirnov test. The data were expressed as arithmetic mean ± standard deviation (SD) and minimum/medium/maximum. The statistical difference among groups was compared using one-way ANOVA followed by the post hoc Student–Newman–Keuls’ test or the Kruskal–Wallis test followed by the post hoc Mann–Whitney U test (where appropriate). Qualitative data between the control and treatment groups were analyzed by the Fisher test. The differences between the groups were considered statistically significant if p < 0.05.

RESULTS

In general, the retrobulbar L-NAME application procedure induces only short-lasting mild increase of intraocular pressure. Of note, with normal intraocular pressure, the severe眼 damage rapidly appears and persists as an inescapable downhill course. If rats receive BPC 157 therapy, a counteraction occurs. We demonstrated BPC 157 therapy effect by fundoscopy, behavior presentation, and histology.

Intraocular Pressure

As increased intraocular pressure is an integral feature of ischemic retinal damage in rats, we measured intraocular pressure using a calibrated applanation tonometer. Besides the initial short-lasting increase of intraocular pressure upon retrobulbar application, which appeared equally upon retrobulbar application of saline, L-NAME, and BPC 157 (Figure 1), intraocular pressure was continuously normal, and it was not otherwise changed upon application of BPC 157 or saline (the data are not specifically shown). Thus, all the damages after retrobulbar application of L-NAME appear within the normal range of intraocular pressure in rats.

Fundoscopy

We demonstrated the specific damaging effect of L-NAME application. On the other hand, much like in the healthy rats (Figure 2A), we demonstrated that immediately after saline or BPC 157 administration, there was consistent normal eye background and normal presentation of the retinal and choroidal blood vessels (IS and IB). These remained the same at 20 min (Figure 1) and remained undisturbed until the end of the experiment.

In contrast, with the administration of L-NAME, there was a moderate generalized irregularity in the diameter of blood vessels with moderate atrophy of the optic disc, faint presentation of the choroidal blood vessels appears immediately, and these lesions (Figure 2, IN, IIN) rapidly progress (Figure 3, A→C; Figure 4; D→G; Figure 5). Subsequently, strong generalized irregularity diameter blood vessels with severe atrophy of the optic disc and extremely poor presentation of the choroidal blood vessels appeared.

BPC 157 retrobulbar application was given as therapy after L-NAME at 20 min or at 48 h, as early therapy (i.e., after L-NAME at 20 min), or as delayed therapy (after L-NAME at 48 h); BPC 157 retrobulbar application stopped this downhill course and instantly reversed back to the normal presentation.
FIGURE 1 | Intraocular pressure was measured using a calibrated applanation tonometer Tonopen XL by Reichert Technologies (the transducer was placed on the same spot of the cornea for each measurement). Intraocular pressure was assessed in normal rats, and values between 13–19 mmHg were considered as normal. Then, the values were assessed before retrobulbar application, and after retrobulbar application of saline (0.1 ml/each eye) (control), BPC 157 1 μg/0.1 ml saline/each eye, or L-NAME 0.5 mg/0.1 ml saline/each eye, at 1, 2, 4, 6, 8, and 10 minutes thereafter. No difference vs. control was noted. 10 rats per experimental group and period for all experiments.

FIGURE 2 | Presentation of the rat retina before (H) and after retrobulbar administered agents (saline (S), BPC 157 (B), and L-NAME (N)), immediately (I), and at 20 min (II) (IS, IB, IN; IIS, IIB, and IIN). H, before agent’s application (H—healthy). Normal eye background, normal presentation of the retinal and choroidal blood vessels (A). I, immediately after agent’s application (IS, IB, and IN) (B). Normal eye background, normal presentation of the retinal and choroidal blood vessels (saline (IS), BPC 157 (IB); moderate generalized irregularity in diameter of blood vessels with moderate atrophy of the optic disk, faint presentation of the choroidal blood vessels (IN). II, 20 min after agent’s application (IIS, IIB, and IIN) (C). Normal eye background, normal presentation of the retinal and choroidal blood vessels (saline (IIS), BPC 157 (IIB) (same presentation maintained till the end of the experiment, data not specifically shown); moderate generalized irregularity diameter blood vessels with moderate atrophy of the optic disk, faint presentation of the choroidal blood vessels (IIN). The images are processed with software purchased with a USB microscope camera “Veho Discovery VMS-004 Deluxe.”
(Figure 3, A→C; Figure 4 D→G; a→g, Figure 5). The beneficial effect of BPC 157 retrobulbar application seems to be permanent (Figure 3 A→C; Figure 4 D→G; a→g, Figure 5).

**Microscopy**

Retrobulbar application of L-NAME resulted in degeneration of ganglion cells and in nerve cell layer narrowing of the blood vessel lumen (using immunohistochemistry for Factor VIII). There was particularly damaged inner plexiform and inner nuclear layer, less thickness, along with complete retinal damage, less thickness, noted 1 week after L-NAME retrobulbar administration. These layers progressively became more damaged toward the end of the 2 and 4 weeks (Figure 6, Figure 7).

BPC 157 retrobulbar application was given as therapy after L-NAME at 20 min or at 48 h. Along with the beneficial effect noted with fundoscopy assessment, all BPC 157 rats (both μg- and ng-regimen, those that received medication at 20 min after L-NAME, and those that received medication at 48 h after L-NAME) commonly presented preserved plexiform and inner nuclear layer and retinal thickness (Figure 6, Figure 7). They did not show narrowing of the blood vessel lumen.

**Behavior**

Illustrative animal behavior on days and weeks after retrobulbar application of L-NAME when mounted on an elevated surface was that rats maintained the initial position of firmly standing on to the surface with posterior legs, almost “frozen,” with limited movements only (Figure 8, Figure 9).

BPC 157 retrobulbar application was given as therapy after L-NAME at 20 min or at 48 h. By contrast, BPC 157–treated rats demonstrated no freezing behavior, but quick exploration and escaping from the elevated surface (Figure 8, Figure 9).

**DISCUSSION**

With the role of the NO-system (Cudeiro and Rivadulla, 1999; Toda and Nakanishitoda, 2007; Guthrie and Kang-Mieler, 2014) and consequent role of the retrobulbar application of the NOS-blocker L-NAME in rats, there is a particular noxious chain of events invariably leading to rapid severe retinal damage and function failure. Likewise, providing the high range of the stable gastric pentadecapeptide BPC 157 regimens (for review, see Sikiric et al., 2014; Sikiric et al., 2020), there is a consistent evidence that BPC 157 application, given after L-NAME challenge, may instantly break this chain of events, both at its early stage and an already advanced stage. Thus, in addition to the general BPC 157 interaction with the NO-system and counteraction of the adverse effects of NO-agents [including prolonged miosis in rats and guinea pigs (for review, see Sikiric et al., 2014; Kokot et al., 2016)], these particular beneficial findings may also indicate a specific counteraction of the L-NAME–induced noxious events, leading to rat retinal ischemia. Applied after L-NAME, BPC 157 administration would equally oppose the L-NAME vascular effect, and consequences thereof, at the early stage, that is, 20 min after L-NAME application (and also, still the presence of L-NAME at that time (Avontuur et al., 1998)), as well as at 48 h, at the already advanced course, with already more established lesions. It means that BPC 157, in accordance with its particular effect on the NO-
leads to the death of retinal neurons, and thereby, beneficial effects of NOS blockers (Adachi et al., 1998; Ju et al., 2000)). This may override the NO-negative significance (overproduction of NO interacting with oxygen radicals leads to the death of retinal neurons, and thereby, beneficial effect of NOS blockers (Adachi et al., 1998; Ju et al., 2000)).

system (Hsieh et al., 1997; Sükiric et al., 2014), may reestablish NO-system functioning in the retinal injury, which may be an early injury or lately, a protracted injury, both counteracted.

Unless therapy was given, we demonstrated the characteristic chain of noxious events after L-NAME in all rats. L-NAME rats exhibited the moderate generalized irregularity in the diameter of blood vessels with moderate atrophy of the optic disc, faint presentation of the choroidal blood vessels appearing immediately, and these lesions (Figure 2, IN, IIIN), and a rapid progress to the severe stage (Figure 3, A→C; Figure 4 D→G; Figure 5). This appeared as a direct consequence of the retrobulbar L-NAME application. Thereby, particularly damaged inner plexiform and inner nuclear layer, noted at week 1, and thereby thinned retina, appear as a result of the L-NAME course. Together, providing the innate harmful effect of NOS blocker L-NAME itself, these findings may emphasize the positive significance of NO in protecting retinal neurons (and thereby, harmful effect of NOS-blockers (Ostwald et al., 1995, 1997; Imay et al., 1997; Hangai et al., 1999; Sakamoto et al., 2006)). This may override the NO-negative significance (overproduction of NO interacting with oxygen radicals leads to the death of retinal neurons, and thereby, beneficial effect of NOS blocker (Adachi et al., 1998; Ju et al., 2000)).

Furthermore, it was demonstrated that NO is continuously produced in the retina and essential for maintenance of retinal blood flow (Nagaoka et al., 2002). Likewise, it was shown that without affecting main blood pressure, intravitreous L-NAME application in cats had a detrimental retinal effect that started at 40 min (i.e., decreased retinal blood flow and decrease of vessel diameters at 100 min), lasting for 180 min (Nagaoka et al., 2002). Thus, it seems that with retrobulbar application of L-NAME in rats and mentioned disturbances, this harmful effect is even stronger and appears quite immediately. L-NAME retrobulbar application produced a definitive particular vascular defect in the retina responsible for the further damaging course. This course goes, as ophthalmoscopy showed, with quickly disabled arteries, with the central retinal artery affected first, and then the central retinal vein along with the central retinal artery (Hayreh et al., 1978; Osborne et al., 2004). The subsequent poor presentation of the choroidal blood vessels (and thereby ciliary arteries affected as well) would appear as a further progress of damaging course of events. Of note, the evidence that the damaged inner plexiform layer and inner nuclear layer are supplied through the central retinal artery verifies that L-NAME (i.e., vasoconstriction) directly affected the central retinal artery for a firm time to
produce/initiate the chain of damaging events. Further progress (i.e., rat retinal thickness, inner plexiform, and inner nuclear layer thickness from week 1 to week 2 to week 4) may indicate the progressive NO-system failure. Evidently, this initial L-NAME–induced defect and subsequent course prompted a particular chain of events. As mentioned above, this may be comparable to the complete occlusion (ligation)-induced defects produced in the other vessels and consequent syndromes (Vukojevic et al., 2018; Gojkovic et al., 2020; Kolovrat et al., 2020). In particular experiments, the infrarenal inferior caval vein (Vukojevic et al., 2018), suprahepatic inferior caval vein (Gojkovic et al., 2020), superior anterior pancreaticoduodenal vein (Amic et al.,...
2018), portal vein and hepatic artery (Kolovrat et al., 2020), and left colic artery and vein (Duzel et al., 2017) were occluded. Consequently, we noted portal and caval hypertension, ECG disturbances, arterial and venous thrombosis, and organ lesions (Vukojevic et al., 2018; Gojkovic et al., 2020; Kolovrat et al., 2020). Therefore, confronted with vessel occlusion (Duzel et al., 2017; Amic et al., 2018; Drmic et al., 2018; Vukojevic et al., 2018; Cesar et al., 2020; Gojkovic et al., 2020; Kolovrat et al., 2020), BPC 157 therapy effect was ascribed to the rapid opening of the collateral pathways to reestablish blood flow and compensate permanent occlusion of the vessel. Accordingly, the concomitant syndromes were fully counteracted (Vukojevic et al., 2018; Gojkovic et al., 2020; Kolovrat et al., 2020). The BPC 157 therapy effect may be analogous in the L-NAME-rats, confronted with the particular damaging retinal effect of L-NAME, and thereby with the particular local effect.

Thereby, while the specific mechanism remains to be further elucidated, a rapid recovering effect on the events going on in the central retinal artery in L-NAME retrobulbar rats may also be possible. Further argument that BPC 157 had conducted a prompt counteraction in the face of L-NAME retinal ischemia may be the mentioned, quite general, counteracting potential of BPC 157 on the L-NAME–induced damaging effects in different models and species (Sikiric et al., 2014). Evidently, BPC 157 counteracted those associated with the arrhythmias and hypertension (Sikiric et al., 1997; Balenovic et al., 2009; Barisic et al., 2013), vascular injuries (Drmic et al., 2017, 2018; Duzel et al., 2017; Amic et al., 2018), pulmonary hypertension (Grabarevic et al., 1997), and pro-thrombotic effect (Stupnisek et al., 2018).
et al., 2015). Also, BPC 157 counteracted L-NAME-induced prolonged miosis (Kokot et al., 2016) and sphincter dysfunction (Djakovic et al., 2016; Belosic Halle et al., 2017; Becejac et al., 2018), muscle weakness (Medvidovic-Grubisic et al., 2017), and catalepsy (Zemba Cilic et al., 2021). Finally, BPC 157 counteracted L-NAME induced many organ lesion aggravation, mostly associated with free radical formation (Lojo et al., 2016; Luetic et al., 2017; Sucic et al., 2019). Furthermore, from the BPC 157-L-NAME retinal ischemia evidence’s point of view, the ideal solution to the combination of these effects was precisely the chosen L-NAME and BPC 157 protocol in agreement with previous studies in all respects similar to the noted outcome. This common protocol point (and thereby, regular counteraction of the L-NAME effects by BPC 157 administration) may effectively integrate the BPC 157-L-NAME-NO-system relationship in the present retinal ischemia study (Sikiric et al., 1997; Balenovic et al., 2009; Barisic et al., 2013; Stupnisek et al., 2015; Djakovic et al., 2016; Kokot et al., 2016; Lojo et al., 2016; Belosic Halle et al., 2017; Drmic et al., 2017, 2018; Duzel et al., 2017; Luetic et al., 2017; Medvidovic-Grubisic et al., 2017; Amic et al., 2018; Becejac et al., 2018; Sucic et al., 2019; Zemba Cilic et al., 2021). Likewise, BPC 157 induced the NO-release of its own, which is resistant to L-NAME (Sikiric et al., 1997; Turkovic et al., 2004). Also, BPC 157 (in aortal ring) induced vasodilatation (even when endothelium is removed) and regulated vasomotor tone through BPC 157–specific activation of the Src–Caveolin-1–endothelial NOS (eNOS) pathway (Hsieh et al., 1997). Activation of the VEGFR2-Akt-eNOS signaling pathway goes without the need of other known ligands or shear stress (Hsieh et al., 2017). In addition, BPC 157 may act as a free radical scavenger (Belosic Halle et al., 2017; Duzel et al., 2017; Luetic et al., 2017; Amic et al., 2018; Drmic et al., 2018; Vukojevic et al., 2018; Sever et al., 2019; Sucic et al., 2019; Kolovrat et al., 2020; Park et al., 2020). In particular, BPC 157 reestablished NO-values in the occluded inferior caval vein and ischemic colon tissue, as well as counteracted increased MDA values (Duzel et al., 2017; Vukojevic et al., 2018). Accordingly, there is an important common understanding that any form of treatment that can enhance the natural cellular antioxidant defense system should have a neuroprotective action in retinal ischemia (Osborne et al., 2004). Finally, the role of BPC 157 as the stabilizer of cellular junction is well founded (Park et al., 2020). It acts via increasing tight junction protein ZO-1 expression and transepithelial resistance. Specifically, BPC 157 inhibited the mRNA of inflammatory mediators (iNOS, IL-6, IFNγ, and TNF-α),
increased the expression of HSP 70 and 90, and antioxidant proteins (such as HO-1, NQO-1, glutathione reductase, glutathione peroxidase 2, and GST-pi) (Park et al., 2020). Consequently, it may effectively counteract ischemia-induced increased capillary permeability also in the rats damaged by retrobulbar L-NAME administration.

Further, we should appreciate the noted methodology point (i.e., the vascular failure obtained by retrobulbar L-NAME injection and by therapy counteracted, as specifically findings, and the normal intraocular pressure—the severe eye damage). This would exclude the possible nonspecific contribution of the additional volume (as it may be the infusion into the anterior chamber to increase intraocular pressure above systolic pressure to induce ischemia (Ostwald et al., 1995, 1997; Imai et al., 1997; Sakamoto et al., 2006)). Upon drug retrobulbar application, intraocular pressure increase was very transitory in all rat groups. The changes occurred only in L-NAME rats, while normal eye background and no change in the normal presentation of the retinal and choroidal blood vessels were observed in the rats that received saline (IS) or BPC 157 (IB) as sham treatment. With the advanced L-NAME time, that is, 20 min or 48 h, the specific vascular failure points the prompt aggravation immediately after retrobulbar saline application and prompt recovering effect after retrobulbar BPC 157 application. This would also exclude the possible contribution of the applied concomitant procedure and given agents on the intraocular pressure. Otherwise, different effects were reported for the agents used in anesthesia {diazepam [increase (Kovalcuka et al., 2013)] and sodium thiopental [decrease (Alipour et al., 2014)]}, for mydriasis (tropicamide, a short-acting anticholinergic agent [increase (Portney and Purcell, 1975)], and local anesthesia {tetracaine drops [increase (Faghihi et al., 2020) or decrease (Sarchahi and Bozorgi, 2012)]}.

Finally, the obtained findings should be summarized with the previous eye evidence. It may be found that BPC 157 participates in ocular control (i.e., BPC 157 counteracted L-NAME-miosis and atropine-mydriasis) (Kokot et al., 2016). This effect goes via NO-mediated and cholinergic mechanisms (Kokot et al., 2016), which were accepted as key in ophthalmic arterial dilatation (Osborne et al., 2004). This may be the common point for the transparency maintained, fully rescued total debridement of corneal epithelium (Lazic et al., 2005), and perforating corneal incisions (Masnec et al., 2015), and thereby antagonization of L-NAME–induced retinal ischemia. Thereby, as L-NAME uniformly affected all control rats, along with the fundoscopic and microscopic findings, the final clue may be the highly expressed freezing behavior in all control rats, which was completely recovered after BPC 157 therapy was given. In support, we evidenced the pentadecapeptide BPC 157 (Sikiric et al., 2013; Sikiric et al., 2016) centrally mediated counteraction...
of the L-NAME–induced catalepsy as well as BPC 157, L-NAME, L-arginine, and NO-relation, in the suited acute and chronic rat behavioral models resembling “positive-like” symptoms of schizophrenia (Zemba Cilic et al., 2021). In terms of the suited rat behavioral models, we mentioned the counteracting potential of BPC 157 on both dopamine- (Zemba Cilic et al., 2021) and NO-system–induced central disturbances (Zemba Cilic et al., 2021). This can explain the reversed freezing rat behavior as consequent to the L-NAME–induced retinal ischemia, which was recovered by BPC 157 administration. In principle, this may be analogous to the demonstrated BPC 157–NO-system relations in many models and species (see Balenovic et al., 2009; Barisic et al., 2013; Sikiric et al., 2014; Kokot et al., 2016; Lojo et al., 2016; Belosic Halle et al., 2017; Drmic et al., 2017, 2018; Duzel et al., 2017; Luetic et al., 2017; Medvidovic-Grubisic et al., 2017; Amic et al., 2018; Sucic et al., 2019; Zemba Cilic et al., 2021). Besides, BPC 157 therapy effect always exhibited the functional recovery as well. This was noted with peripheral nerve injury (Gjurasin et al., 2010) and central nervous disturbances (i.e., traumatic brain injury (Tudor et al., 2010), various encephalopathies (Ilic et al., 2009; Ilic et al., 2010; Ilic et al., 2011a; Ilic et al., 2011b; Kliceck et al., 2013; Lojo et al., 2016; Drmic et al., 2017; Medvidovic-Grubisic et al., 2017), stroke (Vukojevic et al., 2020), and spinal cord injury (Perovic et al., 2019). Likewise, there are many functional and structural similarities between the retina and brain (Osborne et al., 2004), and BPC 157 can likely cross the blood–brain barrier to induce neurotransmitter release (Tohyama et al., 2004; Sikiric et al., 2016).

On the other hand, for the L-NAME retrobulbar application–induced retinal ischemia model and BPC 157 therapy in rats, we should emphasize the point that animal studies per se may be cautious regarding their results, although the role of an animal model is indispensable (Minhas et al., 2015; Vestergaard et al., 2019). Also, we should emphasize the relative paucity of the BPC 157 clinical data (Seiwerth et al., 2018; Sikiric et al., 2018; Sikiric et al., 2020; Gwyer et al., 2019). However, it should be noted that BPC 157 was proven to be efficacious in the ulcerative colitis, both in clinical settings (Veljaca et al., 2003; Ruenzi et al., 2005) and in experimental rat studies (see Duzel et al., 2017), and has very safe profile (LD1 could be not achieved) (Seiwerth et al., 2018), a point recently confirmed (Xu et al., 2020). Thus, while the additional studies will be done, we could suggest, based on the role of NO-system (Cudeiro and Rivadulla, 1999; Toda and Nakashihotoda, 2007; Guthrie and Kang-Mieler, 2014), known BPC 157–NO-system close relations (Sikiric et al., 2014; Hsieh et al., 2017), and consequent role of the retrobulbar application of the NOS-blocker L-NAME and BPC 157 therapy in rats, that BPC 157 may consistently counteract retinal ischemia in rats. Hopefully, providing that it may mimic the occlusion of the central retinal artery in patients (Minhas et al., 2015; Chronopoulos and Schutz, 2019), all together, we can envisage the further clinically effective treatments of the retinal ischemia using the stable gastric pentadecapeptide BPC 157.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusion of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The animal study was reviewed and approved by the Ethics Committee School of Medicine, the University of Zagreb.

AUTHOR CONTRIBUTIONS

MZ and TK: conceptualization and methodology. AK and IB: validation. MP and LV: formal analysis. SM and KM: investigation. DD and BR: resources. SS and FS: writing—original draft, review, and editing.

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