ABSTRACT

Dry rot of yam (Dioscorea spp.) caused by Scutellonema bradys and Pratylenchus spp. has restricted yam production in Brazil. Plant-based products can be useful reducing nematode damages. The objectives of this study were to evaluate the nematostatic and nematicidal in vitro activities of pyroligneous extracts from Cocos nucifera, Syagrus cearensis, S. coronata, and Wodetia bifurcata palms against S. bradys and to evaluate the effect of C. nucifera pyroligneous extract in the treatment of yam tubers infected by S. bradys and Pratylenchus sp. under greenhouse conditions. The pyroligneous extracts obtained at pyrolysis temperatures of 400, 500, 600, and 700 °C, and at different concentrations, were tested for nematode immobility and mortality in Kline slides. Cocos nucifera pyroligneous extract obtained at 400 °C and 1% concentration was tested on yam tubers, under different immersion periods. Six months after yam planting the nematode populations were evaluated. All pyroligneous extracts inhibited the mobility and caused mortality to S. bradys, however 100% inhibition of both variables was achieved at concentrations varying from 0.75 to 2%, depending on the pyrolysis temperatures. The reproduction factor was reduced in 43% by treating infected yam tubers with pyroligneous extract from C. nucifera.

Keywords: alternative control; Arecaceae; Dioscorea spp.; Scutellonema bradys; Pratylenchus sp.
plant pests and diseases (Azevedo et al., 2013; Trindade et al., 2014; Pieta, 2017), including plant-parasitic nematodes (Corbani, 2008; Santos et al., 2017). The antimicrobial activity of pyroligneous extracts is attributed to the presence of phenolic compounds, carbonyls and organic acids (Campos, 2018). Some species of Arecaceae commonly found in Brazil and not yet known in relation to their biological activity on plant pathogens have showed antimicrobial effects on human pathogens, for instance, ouricuri [(Syagrus coronata (Mart.) Becc)] (Hughes et al., 2013; Bessa et al., 2016), and coconut (Cocos nucifera L.) (Igwe & Ugwunnaji, 2016). Studies related to the biological potential of pyroligneous extracts obtained from palm trees regarding plant pathogens, should be performed.

The objectives of this study were: i) to test the nematostatic and nematicidal effects of pyroligneous extracts from coconut, catolé (Syagrus cearensis Noblick), ouricuri and foxtail palms (Wodetia bifurcata Irvine) against the Yam nematode, and ii) to evaluate the effect of coconut pyroligneous extract in the treatment of Yam tubers naturally infected by S. bradys and Pratylenchus sp. under greenhouse conditions.

MATERIAL AND METHODS

Experimental site

The experiments were performed at the Laboratory of Plant Pathology and greenhouse of the Research Center for Agricultural Sciences, Federal University of Alagoas (UFAL), Rio Largo, AL, Brazil.

Pyroligneous extracts origin

The pyroligneous extracts were obtained at the Chemical Department from the Federal University of Alagoas, Brazil. All extracts were obtained from the fruit endocarp (agro-industrial waste) of the palm species and under different pyrolysis temperatures from 400°C to 700°C, heating rate of 10°C min⁻¹ and residence time of 2 hours (Veira, 2019).

In vitro assays of the pyroligneous extracts on Scutellonema bradys

Yam tubers (Dioscorea cayenensis Lam,) exhibiting typical dry rot symptoms were collected in the field, in the state of Alagoas, Brazil, and then the nematodes were extracted according to the method of Coolen & D’Herde (1972). Nematode identification was based on their morphological characteristics according to Mai & Mullin (1996).

The pyroligneous extracts from C. nucifera, S. cearensis, S. coronata, and W. bifurcata, obtained at four different pyrolysis temperatures (400, 500, 600, and 700 °C), were diluted in water at concentrations of 0, 0.25, 0.50, 0.75, 1.0, and 2.0%. In Kline plates 200 µL of each extract and for each concentration were added before individually transferring 20 nematodes (juveniles and adults) from the extraction suspension to each cavity with the aid of a fine insect pin. Subsequently, plates were placed in plastic boxes containing filter paper soaked in distilled water, in order to maintain the humidity, and kept at room temperature. After 24 h of incubation the nematode specimens remaining motionless were counted under an inverted light microscope and transferred to distilled water; those which did not recover motility after 24 hours were considered as dead.

Four assays were performed to test each extract in a completely randomized design in factorial scheme (4 pyrolysis temperatures x 6 extract concentrations) with four replicates. The data were analyzed by analysis of variance using the statistical software SAEG 9.1, and response surface graphics were generated using the software Statistics 12.0.

In vivo assay of the pyroligneous extracts of coconut on Scutellonema bradys and Pratylenchus sp.

Under greenhouse conditions, Yam tuber seeds (D. cayenensis) with symptoms of dry rot disease were collected from Yam production areas in the state of Alagoas. Only the pyroligneous extract obtained at a pyrolysis temperature of 400°C from the fruit endocarp of C. nucifera was tested, due to the better nematostatic and/or nematicidal effect demonstrated in vitro, and due to the availability of the material. Samples of 1g of each tuber peel were processed according to Coolen & D’Herde (1972) method, to determine the initial population (Pi). Nematode quantification was performed in Peters slides, under an inverted light microscope.

Sprouting Yam seed tubers were immersed for 2, 3, 4, and 5 hours, based on a preliminary test. Subsequently, the tubers were planted in plastic pots, with 8 kg capacity, containing sterilized soil. Six months after planting, roots and tubers were collected to determine the root and tuber peel fresh weight. After weighing, the nematodes were extracted according to Coolen & D’Herde (1972) method, while 100 cm³ of soil were processed according to Jenkins (1964). The total number of nematodes was divided by the root or tuber peel weight, to obtain the number of nematodes g⁻¹ tissue. The reproduction factor (RF) of the nematodes [RF = Final population (roots + tubers + soil) / Initial population from the tubers] was also calculated for each replicate. The experiment was performed under completely randomized design with four treatments plus the control, and eight replicates. Data were submitted to analyses of variance and, when statistically significant (P ≤ 0.05), to regression analysis using the statistical software SAEG 9.1.
RESULTS

In vitro assays of the pyroligneous extracts on Scutellonema bradys

Significance was observed for all the products tested concerning the percentage of S. bradys immobility (PIM) and mortality (PMORT), depending on the concentrations and the pyrolysis temperatures (Table 1). The exposition of the nematodes to pyroligneous extracts resulted on morphological alterations similar to vacuoles, which were observed in high numbers in all nematodes. No apparent variations were observed on the nematode body from control treatment (Figure 1).

There was a higher effect of PIM and PMORT when the concentrations increased for all pyroligneous extracts tested. Moreover, increasing concentrations resulted in a tendency to stabilize the nematostatic and nematicidal effects, which reached the maximum point (100%) at a concentration between 0.75 and 2%, depending on the pyrolysis temperatures (Figures 2 and 3).

Concerning the pyroligneous coconut extract, increasing concentration resulted in higher PIM, regardless the pyrolysis temperature (Figure 2A). As the regression effect was quadratic for concentration, the maximum value for nematode immobility was reached between 0.75 and 1.0%, depending of the pyrolysis temperature. However, for temperatures between 400 and 600 °C, the maximum was observed at a concentration of 0.75%, and at 700 °C the maximum immobility was observed near the concentration of 1.0%. Therefore, the pyrolysis temperature showed a negative linear effect on PIM, according to the adjusted model. Thus, higher temperatures caused a slight reduction in PIM, allowing to achieve a maximum value at higher concentrations. Furthermore, increasing the concentration of the pyroligneous extract from coconut correspondingly increased PIM, independent of the pyrolysis temperature (Figure 2B). As the effect was quadratic for concentration, the maximum nematode mortality was reached between 0.75 and 1.0%, depending on the pyrolysis temperature. For lowest temperatures (400 and 500 °C) the maximum PMORT occurred at 0.75%, and at higher temperatures (600 and 700 °C), the maximum value was close to 1.0%. In addition, the pyrolysis temperature showed a negative linear effect on PMORT, according to the adjusted model, showing the increase in temperature caused a slight reduction on PMORT, allowing to achieve a maximum value at higher concentrations.

In the case of foxtail palm, the PIM was also concentration-dependent, as PIM increased with an increase in the concentration of the pyroligneous extract, independent of the pyrolysis temperature (Figure 2C). As the effect was quadratic for concentration, the maximum...
nematode immobility was reached at 1%, for pyrolysis temperature between 400 and 600 ºC. At 700 ºC, the maximum PIM occurred at 0.75%, due to the quadratic effect of the temperature, as a decrease of PIM occurred with the increase of temperature from 400 to 500 ºC, and an increase in PIM occurred between 600 and 700 ºC. Similar results were observed for PMORT (Figure 2D).

In catolé, the PIM was influenced by the extract concentration, and the maximum value was reached at 1.0%, independent of the pyrolysis temperature (Figure 3A). The temperature showed a negative linear effect on PIM, according to the adjusted model, that is, the increase on the temperature triggered a slight decrease in PIM, however, did not interfere in the maximum concentration determined (1%). Similar results were observed for PMORT (Figure 3B).

Concerning the pyroligneous extract from ouricuri, the increase of PIM was concentration-dependent, and independent of the pyrolysis temperatures (Figure 3C). As the effect was square root model for concentration, the maximum nematode immobility was reached near 1%, for temperatures between 400 and 600 ºC. At 700 ºC, the concentration resulting in a maximum PIM fell to 0.75%. This behavior was due to the effect of the model, that is to say, there was a decrease in PIM with the increase of the temperature between 400 and 500 ºC, and an increase in PIM between 600 and 700 ºC. Similar behavior was also observed for PMORT (Figure 3D).

**In vivo assay of the pyroligneous extracts of coconut on Scutellonema bradys and Pratylenchus sp.**

Only *S. bradys* was detected in the initial population, however, mixed population formed by *S. bradys* (92.86%) and *Pratylenchus* sp. (7.13%) was observed in the final population. This is likely due to the low population density of *Pratylenchus* sp. in the seed tubers, associated to the low weight (1 g) of the tuber skin used for extraction in the determination of the initial population of the nematodes. Among the analyzed variables, significant difference was

![Figure 1: Characteristics observed within Scutellonema bradys bodies: A - after 24 hours of incubation in pyroligneous extracts, showing morphological changes similar to vacuolation (arrow) (200X magnification), B - control (100X magnification).](image-url)
observed only for the reproduction factor (RF), which was reduced by 43% from the immersion period of 5 hours when compared to the control. This variation can be represented by the linear model with the coefficient of determination ($R^2$) of 0.985 (Figure 4).

**DISCUSSION**

Results from the present study demonstrated that all pyroligneous extracts caused immobility and mortality to *S. bradys*, mostly independent of the pyrolysis temperatures. These results did not corroborate those published by Booker et al. (2010) who observed the activity of bio-oils from tobacco leaves (*Nicotiana tabacum* L.) ranged according to the pyrolysis temperatures, having a decreasing activity with the increase of temperature up to 550 °C; and at 450 °C, the greatest growth inhibition was observed for *Streptomyces scabies* (Thaxter) Waksman & Henrieci, *Clavibacter michiganensis* (Smith) Davis and *Pythium ultimum* Trow. According to these authors, this could be attributed to the active components being cracked into smaller, inactive components due to the increase of the temperature.

It is important to emphasize that the chemical nature of the liquid fraction, as well as the other components obtained from the pyrolysis process (solids and gases), depends on the feedstock and other variables as the temperature, residence time, and heating rates (Czernik & Bridgwater, 2004; Mohan et al., 2006). For instance, cellulose degradation occurs at 240-350 °C to produce anhydrocellulose and levoglucosan; hemicellulose decomposes at temperatures of 200-260 °C and its degradation produces carboxylic acids; and lignin decomposes when heated at 280 to 500 °C, and consists mainly of phenolic compounds (Mohan et al., 2006; Guedes et al., 2010). According to Campos (2018) the antimicrobial activity of pyroligneous extracts is due to the presence of phenolic compounds, carboxyls, and organic acids.

Pyroligneous extracts of coconut and ouricuri obtained at pyrolysis temperature of 400 °C and analyzed by gas

---

**Figure 2:** Response surface graphics for pyrolysis temperatures (400, 500, 600, and 700 °C), and pyroligneous extracts concentrations (0, 0.25, 0.50, 0.75, 1.0, and 2.0%) on percentage of immobility (PIM) and mortality (PMORT) in *Scutellonema bradys*. Coconut (a, b) and foxtail palm (c, d). Regression equations - PIM: Coconut = 25.87 + 155.1650*C – 57.7616*C*C – 0.0148*T ($R^2$ = 0.849, P < 0.05); foxtail palm = 49.83 + 154.0530*C – 53.2227*C^2 – 0.1834*T + 0.0001924*T^2 ($R^2$ = 0.950, P < 0.05. PMORT: Coconut = 24.02 + 157.374*C – 57.9692*C*C – 0.0173*X ($R^2$=0.864, P < 0.05); foxtail palm = 61.20 + 155.3660*C – 53.1017*C^2 – 0.2364*T + 0.000219*T^2 ($R^2$=0.950, P < 0.05); T= temperature; C= concentration.
chromatography coupled to mass spectrometry showed the following chemical groups: aldehyde, phenolic aldehyde, ketone, ether, phenol, hydrocarbons, and terpenes (coconut), and carboxylic acid, organic acid, aldehyde, phenolic aldehyde, ketone, ether, and phenol (ouricuri) (Vieira, 2019). Among these compounds the chemical groups of aldehydes, phenols, terpenes, or organic acids, are known as nematicidal substances against *Bursaphelenchus xylophilus* (Steiner et Buhrer) Nickle, *Panagrellus redivivus* (L.) Goodey, *Caenorhabditis elegans* (Maupas) Dougherty, *Heterodera zeae* Koshy, Swarup & Sethi (Faizi et al., 2011); *C. elegans* (Abdel-Rahman et al., 2013) and *Meloidogyne incognita* Kofoid & White (Aoudia et al., 2012; Caboni et al., 2013; Seo & Kim, 2014).

The organic acids, acetic acid and lactic acid were tested alone and in combination, promoting 100% mortality of *M. incognita* second-stage juveniles at a concentration of 0.1% (acetic acid and mixtures) and 0.5% (lactic acid). Furthermore, the nematode bodies were disrupted severely and moderately by vacuolation at 0.5% mixed and single organic acids, respectively (Seo & Kim, 2014). This suggests that probably, the nematostatic/nematicidal effect observed in the present study, was due to the occurrence of some organic acids present in the pyroligneous extracts, considering that vacuolation observed was similar to those previously described.

High predominance of phenolic compounds was also observed in bio-oils from coconut fibers (Almeida et al., 2013) and coconut shell (Gao et al., 2016; Hadanu & Apituley, 2016) at pyrolysis temperatures varying from 350 -700 ºC. Probably, the presence of these compounds in the pyroligneous extracts of coconut used in the present study contributed for their effect on *S. bradys*. However, the bio-activity of *C. nucifera* on nematodes is poorly documented. As example, the studies of coconut extracts against intestinal nematodes (Oliveira et al., 2009; Costa et al., 2010).

The RF decrease observed in the present study (3.6 to 2.04) is a valuable information for the management of these

---

**Figure 3:** Response surface graphics for pyrolysis temperatures (400, 500, 600, and 700 ºC), and pyroligneous extracts concentrations (0, 0.25, 0.50, 0.75, 1.0, and 2.0%) on percentage of immobility (PIM) and mortality (PMORT) in *Scutellonema bradys*. Catolé (a, b) and ouricuri (c, d). Regression equations - PIM: catolé = 23.50 + 155.1790*C – 53.443*C^2 – 0.0323*T (R^2 = 0.937; P < 0.05); ouricuri = 346.33 + (176.2720*Sqrt(C)) – 77.2415*C – 29.9975* Sqrt(T) + 0.6491*T (R^2=0.974; P < 0.05). PMORT: catolé = 21.84 + 155.2850*C – 53.4715*C^2 – 0.0321*T (R^2 = 0.949; P < 0.05); ouricuri = 460.1630 + 171.5900*Sqrt(C) – (73.2614*C) – 39.8209* Sqrt(T) + 0.8575*T (R^2=.962; P < 0.05). T= temperature; C= concentration.
plant-parasitic nematodes on yam. However, a RF below 1.0 is desirable to reduce nematode populations in the area (Alves et al., 2011). Then, the pyroligneous extracts of coconut should be investigated in complementary studies, in order to achieve robust results regarding its nematicidal potential.

Studies focused on pyroligneous extracts have shown important bio-activity for other plant-parasitic nematodes, and the efficacy of these products can also occur through indirect action by the plant-induced resistance. The pyroligneous extract Biopirol® (aqueous phase obtained in the distillation of eucaliptus tar), for example, was used as resistance inducer and showed efficacy to reduce the number of eggs and the reproduction factor of *M. incognita* in tomato (*Solanum lycopersicum* L.), at the lowest concentration tested (0.5%) in all application periods (Melo et al., 2012). In another study, Biopirol® reduced the hatching of second stage juveniles of *M. incognita, M. javanica* (Treub) Chitwood, and *Tylenchulus semipenetrans* Cobb, and also the formation of galls by *M. incognita* and *M. javanica* in tomato roots (Corbani, 2008). The use of Biopirol at 5% resulted in high suppressive effect on population densities of *Pratylenchus* and *Meloidogyne* in sugarcane fields (Rossi & Lima, 2007). On lettuce (*Lactuca sativa* L.), efficient nematostatic and nematicidal action of the product against *M. incognita* was observed for all tested concentrations, however, a phytotoxic effect was verified (Santos et al., 2017). Probably, this phytotoxic effect is due to the chemical characteristics of the pyroligneous extract, such as the low pH, that may negatively affect the availability of soil nutrients to the plants. Researchers have pointed out that the application of the pyroligneous extract PiroQualis® (produced from eucalyptus wood) at 4 % (v/v) and 8 % (v/v) induced an increase in the potential acidity, as well as a decrease in pH, base saturation, total cation exchange capacity and calcium concentration in the 0-20 cm soil layer (Togoro et al., 2014). In the present study, in spite of the acidity (pH value of 3.07) of the coconut pyroligneous extract, yam plants did not show symptoms of phytotoxicity.

Despite some information documented in the literature regarding the use of pyroligneous extract against plant-parasitic nematodes, positive findings observed for pyroligneous extract obtained from palm trees is now documented for the first time. However, further studies are compulsory in order to evaluate different extract concentrations and immersion periods in the treatment of the yam propagative material infected by the dry rot disease nematodes.

**CONCLUSIONS**

Pyroligneous extracts from the palm trees *Cocos nucifera, Syagrus cearensis, S. coronata*, and *Wodetia bifurcata* showed nematostatic and nematicidal effect for *S. bradys*.

Pyroligneous extract from *C. nucifera* reduced the reproduction factor of a mixed population formed by *S. bradys* and *Pratylenchus sp.* on yam plants.

**ACKNOWLEDGEMENTS AND FULL DISCLOSURE**

This study was financed in part by the Coordination for the Improvement of Higher Education Personnel - Brazil (CAPES) - Finance Code 001. The authors inform that there is no conflict of interest in carrying the research and publishing this manuscript.
REFERENCES

Abdel-Rahman FH, Alaniz N & Saleh MA (2013) Nematicidal activity of terpenoids. Journal of Environmental Science and Health, 48:16-22.

AGROFITT (2020) Sistemas de Agrotóxicos Fitosanitários. Produtos formulados. Available at: http://agrofit.agricultura.gov.br/agrofit_cons/principal_agrofit_cons. Accessed on: February 20th, 2020.

Almeida TM, Bispo MD, Cardoso AR T, Migliorini MV, Schena T, Campos MCV, Machado ME, López Já, Krause LC & Caramão EB (2013) Preliminary studies of bio-oil from fast pyrolysis of coconut fibers. Journal of Agricultural and Food Chemistry, 61:6812-6821.

Alves TCU, Silva RA, Borges DC, Motta LCC & Kobayast L (2010) Biomass gasification and pyrolysis: practical design and theory. Burlington, Academic Press. 376p.

Basu P (2010) Biomass gasification and pyrolysis: practical design and theory. Burlington, Academic Press. 376p.

Bessa CMAS, Nascimento RS, Alves RCC, Anselmo JM, Silva APSA, Silva AG, Lima VLM, Tavares JF, Silva LCN, Silva MV & Correa MTS (2016) Syngenta corn seed oils have antimicrobial action against multidrug-resistant Staphylococcus aureus. Journal of Medicinal Plants Research, 10:310-317.

Campos AD (2007) Técnicas para produção de extrato pirolênho. Available at: http://agrofit.agricultura.gov.br/agrofit_cons/principal_agrofit_cons. Accessed on: February 20th, 2020.

Campos AD (2018) Informação técnica sobre extrato pirolênho. Pelotas, Embrapa Clima Temperado. 59p. (Circular Técnica, 65).

Corbani RZ (2008) Estudo do extrato pirolênho Biopiro® no manejo de nematodes in cana-de-açúcar, olérficas e citros, em diferentes ambientes. Tese de Doutorado. Universidade Estadual Paulista "Júlio de Mesquita Filho", Jaboticabal. 55p.

Costa CTC, Bevilacqua CML, Morais SM, Camarúca-Vasconcelos ALF, Maciel MV, Braga RR & Oliveira LMB (2010) Anthelmintic activity of Cocos nucifera L. on intestinal nematodes of mice. Research in Veterinary Science, 88:101-103

Czernik S & Bridgewater AV (2004) Overview of applications of biomass fast pyrolysis oil. Energy & Fuels, 18:590-598.

Faizi S, Fayyaz S, Bano S, Iqbal EY, Ahmed L, Siddiqui H & Naz A (2011) Isolation of nematicidal compounds from Tagetes patula L. yellow flowers: structure-activity relationship studies against cyst nematode Heterodera zeae infective stage larvae. Journal of Agricultural and Food Chemistry, 59:9080-9093.

Gao Y, Yang Y, Qin Z & Sun Y (2016) Factors affecting the yield of bio oil from the pyrolysis of coconut shell. SpringerPlus, 5:333.

Guedes CLB, Adão DC, Quesada TP, Borsato D, Galão OF, Mauro ED, Pérez JMM & Rocha JD (2010) Avaliação de biocombustível derivado do bio-óleo obtido por pirólise rápida de biomassa lignocelulósica como aditivo para gasolina. Química Nova, 33:781-786.

Hadaru R & Apirutey DAN (2016) Volatile compounds detected in coconut shell liquid smoke through pyrolysis at a fractioning temperature of 350-420 °C. Makara Journal of Science, 20:95-100.

Hughes AF, Lima FG, Lucchese AM, Neto AG & Uetanabaho APT (2013) Antimicrobial activity of Syagrus coronata (Martius) Beccari. Brazilian Archives of Biology Technology, 56:269-274.

Igwe OU & Ugwunnaji IP (2016) Phytochemistry, antioxidant and antimicrobial studies of endosperm tissues of Cocos nucifera L. International Journal of Chemical, Material and Environmental Research, 3:78-83.

Jenkins WRA (1964) A rapid centrifugal-floatation technique for separating nematodes from soil. Plant Disease Reporter, 48:692.

Li G-H, Dang L-Z, Liu F-F, Li L, Liu Y-J & Zhang K-Q (2009) Nematicidal activity of honokiol and magnolol isolated from Magnolia tripetula. Journal of Phytopathology, 157:390-392.

Lima RS, Muniz MFS, Costa JG, Silva KB, Beheading A (2019) Extratos aquosos de Annona spp. e Croton heliotropifolius sobre Scutellonema bradyi e prospecção química dos compostos. Summa Phytopathologica, 45:223-224.

Mai WF & Mullin PG (1996) Plant-parasitic nematodes: a pictorial key to genera. 5th edition. New York, Cornell University. 288p.

Melo TA, Serra IMRS, Silva GS & Sousa RMS (2012) Produtos naturais aplicados para manejo de Meloidogyne incognita em tomateiros. Summa Phytopathologica, 38:223-227.

Mohan D, Pittman CU & Steele PH (2006) Pyrolysis of wood/ biomass for bio-oil: a critical review. Energy & Fuels, 20:848-889.

Moura RM (2016) Doenças do inãhme–da–coasta (Dioscorea cayennensis). In: Amorim L, Rezende JAM, Bergamin Filho A & Camargo LEA (Eds). Manual de fitopatologia - doenças das plantas cultivadas. 5th edition. Ouro Fino, Agronômica Ceres. p. 477-483.

Oliveira LMB, Bevilacqua CML, Costa CTC, Macedo ITF, Barros RS, Rodrigues ACM, Camarúca-Vasconcelos ALF, Morais SM, Lima YC, Vieira LS & Navarro AMC (2009) Anthelmintic activity of Cocos nucifera L. against sheep gastrointestinal nematodes. Veterinary Parasitology, 159:55-59.

Pieta S (2017) Eficácia de extratos pirolênhos de cana-de-açúcar (Saccharum officinarum L.) e eucalipto (Eucalyptus spp.) no controle in vitro de patógenos da soja. Dissertação de Mestrado. Universidade Federal da Grande Dourados, Dourados. 59p.

Rev. Ceres, Viçosa, v. 67, n.6, p. 482-490, nov/dec, 2020
Rossi CE & Lima CB (2007). Controle alternativo de nematóides em cultura orgânica de cana-de-açúcar. Revista Brasileira de Agroecologia, 2:1545-1548.

Santos AM, Almeida FA, Fonseca WL, Leite MLT, Pereira FF & Carvalho RM (2017) Ácido pirolenholso no manejo de nematóides das galhas na cultura da alface. Espacios, 38:01-09.

Seo Y & Kim YH (2014) Control of Meloidogyne incognita using mixtures of organic acids. Plant Pathology Journal, 30:450-455.

Shaw M (2006) Pyrolysis of lignocellulosic biomass to maximize bio-oil yield: an overview. In: 2006 ASABE Annual Meeting. Edmonton, ASAB. p. 1-14.

Togoro AH, Silva JAS & Cazetta JO (2014) Chemical changes in an oxisol treated with pyroligneous acid. Ciência e Agrotecnologia, 38:113-121.

Trindade RCP, Palmeira LH, Sant’ana AEG, Sousa RS, Costa APA & Amorim EPR (2014) Atividade do extrato pirolenholso sobre lagartas de Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae). Revista Brasileira de Agroecologia, 9:84-89.

Vieira WT (2019) Caracterização cromatográfica e avaliação da atividade antimicrobiana do extrato pirolenholso obtido a partir de biomassas residuais. Dissertação de Mestrado. Universidade Federal de Alagoas, Maceió. 138p.