Changes in Expression of the CLOCK Gene in Obstructive Sleep Apnea Syndrome Patients Are Not Reverted by Continuous Positive Airway Pressure Treatment

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Purpose: Metabolic syndrome and cardiovascular disease are strongly associated with obstructive sleep apnea syndrome (OSAS), which causes substantial changes to normal circadian physiological functions, including metabolic pathways. Because core clock genes are known to be modulated by sleep/vigilance cycles, we asked whether the expression level of mRNA coding for clock genes is altered in non-treated OSAS patients and if it can be corrected by standard continuous positive airway pressure (CPAP) treatment.

Methods: Peripheral blood was collected from male patients diagnosed with severe OSAS (apnea-hypopnea index ≥ 30/h) before and after treatment initiation. qPCR was used to measure mRNA levels of genes associated with the central circadian pacemaker including CLOCK, BMAL1, Cry1, Cry2, and three Period genes (Per 1, 2, 3) in peripheral blood mononuclear cells (PBMCs).

Results: We found statistically significant differences for CLOCK (p-value = 0.022) expression in PBMCs of OSAS patients which were not reverted by treatment with CPAP. We have also found a substantial decrease in the slow wave sleep (SWS) content in OSAS patients (p-value < 0.001) that, contrary to REM sleep, was not corrected by CPAP (p-value = 0.875).

Conclusion: CPAP treatment does not correct substantial changes in expression of core clock genes in OSAS patients. Because CPAP treatment is also unable to normalize the SWS in these patients, it is likely that additional therapeutic interventions that increase SWS content and complement the benefits of CPAP are required to more effectively reduce the known increased cardiovascular risk associated with OSAS patients.

Keywords: obstructive sleep apnea syndrome, clock genes, continuous positive airway pressure, slow wave sleep, Metabolic Syndrome, Circadian Rhythms

INTRODUCTION

The obstructive sleep apnea syndrome (OSAS) is a frequent sleep disorder that constitutes an independent risk factor for the development of metabolic syndrome and cardiovascular disease (1). Most of the gene components of critical metabolic pathways and their regulators exhibit circadian variation at the mRNA and protein levels (2). They are regulated by core clock genes, which in turn are modulated...
by sleep/vigilance cycles (3). Importantly, single genetic disruption of
the core clock genes CLOCK (4) and BMAL1 (5, 6) cause full-
blown metabolic syndrome phenotypes in mice. Milder metabolic
syndrome phenotypes have also been observed in double or triple,
but not single, inactivation of additional core clock genes, includ-
ing Per1, 2, 3, and Cry1 and 2 (7). In this work, we investigated
whether the expression level of mRNA coding for clock genes is
altered in non-treated OSAS patients and if it can be corrected by
continuous positive airway pressure (CPAP) treatment.

**MATERIALS AND METHODS**

**Study Population**

Using polysomnography (Alice 5, Respironics, Inc., Murrysville,
PA, USA) scored according to the 2007 AASM manual for the
scoring of sleep and associated events and stringent clinical
criteria (patients with addiction habits, cancer, hematological
disorders, active infection, or shift work were excluded), we have
identified a group of 13 male patients with severe OSAS [aver-
age apnea-hypopnea index (AHI) of 66.7/h, all ≥ 30/h] and 7
matched controls for sex (average AHI of 2, all < 5/h). Patients
and controls, without the exclusion criteria, were recruited at the
sleep consultation of Santa Maria Hospital (Lisbon, Portugal),
from those with a high degree of suspicion to have Sleep Apnea,
based on their clinical history. Patients and controls were distin-
guished on the basis of the PSG results. Sleepiness was evaluated
using the Stanford Sleepiness Scale and the Epworth Sleepiness
Scale (ESS). See **Table 1** for sleep study results in the healthy
and OSAS group. Cortisol levels were within normal limits for
all subjects (data not shown). For a subset of patients, PSG was
repeated after treatment with CPAP to assess its impact on REM
and slow wave sleep (SWS) content. This study was approved by
the Ethics Committee Review Board of Centro Hospitalar Lisboa
Norte, EPE (Lisboa, Portugal), and written consent obtained for
every patient and control sample included in this study.

**Measurement of Gene Expression**

We have used quantitative RT-PCR to systematically quantify
the expression of the core clock genes in peripheral blood cells of
severe OSAS patients in comparison with that of matched non-
affected control volunteers, before and after 1 month of treatment
with CPAP. Several studies have demonstrated that clock gene
expression profiles in human peripheral blood mononuclear cells
(PBMCs) can be used as surrogate measures of central circadian
rhythm expression changes in patients and healthy subjects (8–11).
Therefore, peripheral blood was collected from the study subjects
before and after 1 month of treatment initiation. Collections were
always performed between 8 and 10 a.m. Blood was then used to
perform routine biochemical analyses (including cortisol levels)
and to isolate PBMCs. The following hematologic and biochemi-
cal parameters were investigated in all participants: full peripheral
blood cell count; hepatic enzymes (aspartate transaminase, alanine
transaminase, and gamma-glutamyltransferase); renal parameters
(creatinine, blood urea nitrogen); standard metabolic parameters
(uric acid, glucose, total cholesterol, low density lipoprotein, high
density lipoprotein, triglycerides, and C-reactive protein). RNA
was isolated and quantitative RT-PCR used to measure mRNA
levels of genes associated with the central circadian pacemaker
including CLOCK, BMAL1, two Cry (Cry 1, 2), and three Period
genes (Per 1, 2, 3). The selected patients were then evaluated
1 month after therapy initiation with CPAP and the mRNA level of
the same genes tested. The sequence of the oligos used for RT-PCR
can be found Table S2 in the Supplementary Material.

**Statistical Analysis**

A non-parametric approach was chosen for the data analysis. We
used median and quartiles for the descriptive statistical analysis.
To test hypotheses related to differences between the control
group and the patient group, we used the Wilcoxon signed-rank
test. To investigate relationships between gene expression and
other parameters, we implemented a correlation analysis based
on Spearman’s rank correlation coefficient.

**RESULTS**

Using Wilcoxon signed-rank test, we found statistically significant
differences between patients and control volunteers in the ESS
($W = 51.5, p$-value $= 0.007$) and in polysomnography parameters

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**TABLE 1** | Polysomnography characterization of healthy controls and obstructive sleep apnea syndrome patients, before continuous positive airway pressure treatment.

|                   | Control group | Patient group |
|-------------------|---------------|---------------|
|                   | 7             | 13            |
|                   | 1st Quartile  | Median        | Mean          | 3rd Quartile  |
| Age (years)       | 35.50         | 40.00         | 44.57         | 54.50         |
| Oxygen desaturation index | 0.10         | 0.30         | 1.16         | 1.60         |
| Time spent with SpO2 < 90%, (min) | 0.00         | 0.00         | 0.21         | 0.00         |
| Lowest SpO2 (%)   | 91.25         | 92.50         | 91.50         | 93.00         |
| Mean SpO2 (%)     | 96.00         | 96.00         | 96.00         | 96.00         |
| ESS               | 0.00          | 2.00          | 2.60          | 3.00          |
| Sleep efficiency (%) | 76.70       | 90.00         | 84.69         | 94.30         |
| Total sleep time (min) | 366.40      | 377.20        | 368.70        | 389.20        |
| REM (min)         | 40.12         | 60.25         | 58.48         | 76.62         |
| Slow wave sleep (min) | 53.50      | 58.25         | 58.75         | 75.38         |
| Arousal index (events/h) | 2.70         | 7.20          | 5.32          | 7.20          |
| Apnea-hypopnea index (events/h) | 0.85        | 1.50          | 2.00          | 3.05          |
(Table 1) including in AHI (W = 91, p-value < 0.0001), oxygen desaturation index (W = 91, p-value < 0.001), SpO2 time < 90% in minutes (W = 90, p-value < 0.001), minimal SpO2 (W = 1, p-value < 0.0001), and average SpO2 (W = 1.5, p-value < 0.001). Statistically significant differences were also found for SWS in minutes (W = 0, p-value < 0.001), REM sleep in minutes (W = 0, p-value = 0.0001), and arousal index (W = 60, p-value < 0.002), but not for sleep efficiency (W = 29.5, p-value = 0.310) or total sleep time (W = 34, p-value = 0.892).

On the comparison between control volunteers and patients before treatment initiation, we found statistically significant differences for CLOCK (W = 15, p-value = 0.022) expression in PBMCs of patients, but not for Cry 1 and 2 or the three Period genes tested (Table 2). The differences of BMAL1 expression between control volunteers and patients nearly reached statistical significance (W = 20, p-value = 0.068), an outcome that is likely, should the population size be larger.

Patient treatment with CPAP did not change abnormal expression of clock genes (Table 3). It also did not improve the reduced levels of SWS observed in non-treated OSAS patients (p-value = 0.875, Table 4), contrary to REM sleep. The persistence of the abnormal expression pattern of the CLOCK gene cannot be attributed to the lack or sub-optimal adherence levels to CPAP treatment as demonstrated by the metrics of CPAP adherence of our patients (Table 5; Table S1 in Supplementary Material). Our data show that adherence to CPAP treatment is well above the minimum required level recommended by the most recent proposal of the American Thoracic Society (12) that points to the need of CPAP use in more than 70% of nights and for more than 4 h per night.

**DISCUSSION**

In our study, as expected, control volunteers and OSAS patients could be easily distinguished by the statistical analysis of the ESS results and polysomnography parameters. They could also be distinguished on the basis of expression levels of CLOCK, but not any of the other core clock genes, with the possible exception of

### Table 2: Characterization of the gene expression between the control group (C) and the patient group at t1 (P1), as well as the results of hypothesis tests for comparison of gene expression between these groups.

|          | Min  | 1st Quartile | Median | 3rd Quartile | Max  | Mean | SD | Hypothesis test |
|----------|------|--------------|--------|--------------|------|------|----|-----------------|
| BMAL1    | P1   | 0.54         | 0.72   | 0.81         | 1.27 | 5.01 | 1.26| W = 20, p-value = 0.068 |
|          | C    | 0.91         | 1.09   | 1.38         | 1.95 | 2.27 | 1.52| 0.04 |
| CLOCK    | P1   | 0.16         | 0.42   | 0.99         | 1.32 | 2.53 | 1.03| W = 15, p-value = 0.022 |
|          | C    | 1.18         | 1.29   | 2.20         | 2.31 | 3.72 | 2.04| 0.90 |
| CRY1     | P1   | 0.96         | 1.18   | 1.52         | 2.12 | 2.85 | 1.68| W = 22, p-value = 0.230 |
|          | C    | 1.26         | 1.58   | 2.13         | 2.57 | 3.31 | 2.14| 0.73 |
| CRY2     | P1   | 0.19         | 1.30   | 1.53         | 1.96 | 3.08 | 1.58| W = 41, p-value = 0.601 |
|          | C    | 0.82         | 1.01   | 1.33         | 1.81 | 1.97 | 1.39| 0.47 |
| PER1     | P1   | 0.05         | 0.32   | 0.40         | 0.59 | 0.98 | 0.48| W = 27, p-value = 0.591 |
|          | C    | 0.26         | 0.37   | 0.61         | 0.80 | 0.85 | 0.58| 0.25 |
| PER2     | P1   | 0.83         | 1.58   | 2.61         | 2.88 | 3.51 | 2.35| W = 35, p-value = 0.438 |
|          | C    | 2.06         | 2.26   | 2.52         | 3.98 | 6.20 | 3.32| 1.63 |
| PER3     | P1   | 1.20         | 2.45   | 2.83         | 4.42 | 11.46| 4.52| W = 53, p-value = 0.088 |
|          | C    | 1.77         | 2.22   | 2.27         | 2.59 | 3.74 | 2.48| 0.62 |

Only the CLOCK gene showed statistically significant changes.

### Table 3: Characterization of the gene expression within the patient group between the values recorded at t1 (P1) and the ones recorded at t2 (P2), as well as the results of hypothesis tests for comparison of gene expression between these time-points.

|          | Min  | 1st Quartile | Median | 3rd Quartile | Max  | Mean | SD | Hypothesis test |
|----------|------|--------------|--------|--------------|------|------|----|-----------------|
| BMAL1    | P1   | 0.54         | 0.72   | 0.81         | 1.27 | 5.01 | 1.26| W = 57, p-value = 0.872 |
|          | P2   | 0.21         | 0.87   | 1.13         | 1.28 | 1.65 | 1.03| 0.45 |
| CLOCK    | P1   | 0.16         | 0.42   | 0.99         | 1.32 | 2.35 | 1.03| W = 58, p-value = 0.923 |
|          | P2   | 0.14         | 0.87   | 0.98         | 1.20 | 1.95 | 1.07| 0.54 |
| CRY1     | P1   | 0.96         | 1.18   | 1.52         | 2.12 | 2.85 | 1.68| W = 32, p-value = 0.813 |
|          | P2   | 0.85         | 1.20   | 1.55         | 2.38 | 3.86 | 1.92| 1.06 |
| CRY2     | P1   | 0.19         | 1.30   | 1.53         | 1.95 | 3.08 | 1.58| W = 36, p-value = 0.756 |
|          | P2   | 0.87         | 1.23   | 1.46         | 2.65 | 4.03 | 2.04| 1.29 |
| PER1     | P1   | 0.05         | 0.32   | 0.40         | 0.59 | 0.98 | 0.48| W = 50, p-value = 1.000 |
|          | P2   | 0.21         | 0.37   | 0.39         | 0.50 | 0.74 | 0.43| 0.17 |
| PER2     | P1   | 0.83         | 1.58   | 2.61         | 2.88 | 3.51 | 2.35| W = 65, p-value = 0.733 |
|          | P2   | 1.07         | 2.24   | 2.74         | 2.79 | 5.42 | 2.70| 1.11 |
| PER3     | P1   | 1.20         | 2.45   | 2.83         | 4.42 | 11.46| 4.52| W = 27, p-value = 0.475 |
|          | P2   | 2.42         | 2.94   | 4.36         | 4.96 | 8.44 | 4.43| 2.07 |

These comparisons did not reveal any significant differences.
BMAL1. Interestingly, these results are strikingly reminiscent of metabolic syndrome phenotypes found in the knockout of the corresponding genes in mice as described earlier. The levels of expression of the CLOCK gene were altered in OSAS patients before and after CPAP treatment, suggesting that either the standard therapeutic management of these patients is insufficient to correct the disruption of the circadian patterns of genes with expression conditioned by cycles of sleep/vigilance or changes in severe sleep apnea patients are structural, for example, as a consequence of irreversible cerebral vascular lesions.

Before patients were treated, we observed consistently and substantially reduced levels of REM and SWS content as compared to healthy controls. Perhaps significantly, while all the patients who initiated CPAP recovered to normal levels of REM sleep, SWS content remained invariably abnormal, at much lower levels than controls (Table 4). It was previously shown that even in young healthy adults, selective suppression of SWS causes pronounced decrease of insulin sensitivity, reduced glucose tolerance, and higher diabetes risk (13).

While it is clear that CPAP substantially improves cognitive and behavioral aspects of the disease (14), metabolic parameters and other OSAS-related risk factors are only partially modified (15), which might explain the incomplete reduction of cardiovascular risk in OSAS. These facts point to the need to complement CPAP therapy with other therapy interventions that are aimed at the sleep structure and circadian rhythm normalization and are likely to have a substantial impact on the natural history of the disease. Drugs that effectively improve the SWS content of patients might constitute prime candidates for this goal.

Study Limitations
While our major conclusions are supported by robust statistically significant results, our study would benefit from increasing the size of the population of both patients and controls. It is likely that with more samples, statistically significant associations will be found for additional clock genes, especially in the case of BMAL1, where a trend is already clear with the current analysis.

INFORMED CONSENT
Informed consent was obtained from all individual participants included in the study.

ETHICS STATEMENT
All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional (Centro Hospitalar Lisboa Norte, EPE) and national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

AUTHOR CONTRIBUTIONS
SM and LM conceived and designed the study; SM, RR, NP, ANC, DP, CP, DF, JR, and CB collected the data. RR, AB, and LM analyzed the data. LM wrote the manuscript.

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SUPPLEMENTARY MATERIAL
The Supplementary Material for this article can be found online at http://www.frontiersin.org/article/10.3389/fmed.2017.00187/full#supplementary-material.

REFERENCES
1. Yaggi HK, Concato J, Kernan WN, Lichtman JH, Brass LM, Mohsenin V. Obstructive sleep apnea as a risk factor for stroke and death. N Engl J Med (2005) 353:2034–41. doi:10.1056/NEJMoa043104
2. Yang X, Downes M, Yu RT, Bookout AL, He W, Straume M, et al. Nuclear receptor expression links the circadian clock to metabolism. Cell (2006) 126:801–10. doi:10.1016/j.cell.2006.06.050
3. Eckel-Mahan K, Sassone-Corsi P. Metabolism and the circadian clock converge. Physiol Rev (2013) 93:107–35. doi:10.1152/physrev.00016.2012
4. Turek FW, Joshu C, Kohsaka A, Lin E, Ivanova G, McDearmon E, et al. Obesity and metabolic syndrome in circadian clock mutant mice. Science (2005) 308:1043–5. doi:10.1126/science.1108750
5. Bunger MK, Wilsbacher LD, Moran SM, Clendenin C, Radcliffe LA, Hogenesch JB, et al. Mop3 is an essential component of the master circadian pacemaker in mammals. Cell (2000) 103:1009–17. doi:10.1016/S0092-8674(00)00205-1
6. Rudic RD, McNamara P, Curtis AM, Boston RC, Panda S, Hogesnesh JB, et al. BMAL1 and CLOCK, two essential components of the circadian clock, are involved in glucose homeostasis. PLoS Biol (2004) 2:e377. doi:10.1371/journal.pbio.0020377

7. Maury E, Ramsey KM, Bass J. Circadian rhythms and metabolic syndrome: from experimental genetics to human disease. Circ Res (2010) 106:447–62. doi:10.1161/CIRCRESAHA.109.208355

8. Takata M, Burioka N, Ohdo S, Takane H, Terazono H, Miyata M, et al. Daily expression of mRNAs for the mammalian clock genes Per2 and clock in mouse suprachiasmatic nuclei and liver and human peripheral blood mononuclear cells. Jpn J Pharmacol (2002) 90:263–9. doi:10.1254/jjp.90.263

9. Takimoto M, Hamada A, Tomoda A, Ohdo S, Ohmura T, Sakato H, et al. Daily expression of clock genes in whole blood cells in healthy subjects and a patient with circadian rhythm sleep disorder. Am J Physiol Regul Integr Comp Physiol (2005) 289:R1273–9. doi:10.1152/ajpregu.00126.2005

10. Fukuya H, Emoto N, Nonaka H, Yagita K, Okamura H, Yokoyama M. Circadian expression of clock genes in human peripheral leukocytes. Biochem Biophys Res Commun (2007) 354:924–8. doi:10.1016/j.bbrc.2007.01.063

11. Burioka N, Takata M, Endo M, Miyata M, Takeda K, Chikumi H, et al. Treatment with beta2-adrenoceptor agonist in vivo induces human clock gene, Per1, mRNA expression in peripheral blood. Chronobiol Int (2007) 24:183–9. doi:10.1080/07420520601140043

12. Schwab RJ, Bade SM, Epstein LJ, Gay PC, Gozal D, Kohler M, et al. An official American Thoracic Society statement: continuous positive airway pressure adherence tracking systems. The optimal monitoring strategies and outcome measures in adults. Am J Respir Crit Care Med (2013) 188:613–20. doi:10.1164/rccm.201307-1282ST

13. Tasali E, Leprout R, Ehrmann DA, Van Cauter E. Slow-wave sleep and the risk of type 2 diabetes in humans. Proc Natl Acad Sci U S A (2008) 105:1044–9. doi:10.1073/pnas.0706446105

14. Kushida CA, Nichols DA, Holmes TH, Quan SF, Walsh JK, Gottlieb DJ, et al. Effects of continuous positive airway pressure on neurocognitive function in obstructive sleep apnea patients: the Apnea Positive Pressure Long-term Efficacy Study (APPLES). Sleep (2012) 35:1593–602. doi:10.5665/sleep.2226

15. Tasali E, Ip MS. Obstructive sleep apnea and metabolic syndrome: alterations in glucose metabolism and inflammation. Proc Am Thorac Soc (2008) 5:207–17. doi:10.1513/pats.200708-139MG

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