Characterization of beta-thalassemia mutations in patients from the state of Rio Grande do Norte, Brazil

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Abstract

35 unrelated individuals were studied for characterization as either heterozygous or homozygous for beta-thalassemia. Molecular analysis was done by PCR/RFLP to detect the mutations most commonly associated with beta-thalassemia (β^0 IVS-I-1, β^+ IVS-I-6, and β^39). In the patients who showed none of these mutations, the beta-globin genes were sequenced. Of the 31 heterozygous patients, 13 (41.9%) had the β^+ IVS-I-6 mutation, 15 (48.4%) the β^0 IVS-I-1 mutation, 2 (6.5%) the β^+ IVS-I-110 mutation and 1 (3.2%) the β^+ IVS-I-5 mutation. IVS-I-6 was detected in the four homozygotes. The mutation in codon 39, often found in previous studies in Brazil, was not detected in the present case. This is the first study aiming at identifying mutations that determine beta-thalassemia in the state of Rio Grande do Norte.

Key words: hereditary hemoglobinopathies, beta-thalassemia, mutations, PCR-RFLP, Brazilian population.

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Beta-thalassemia is a group of hemoglobin diseases caused by a reduction (β^− thalassemia) or absence (β^0 thalassemia) in the synthesis of beta-globin chains. More than 200 different types of mutations have been described as being responsible for this disease. Affected individuals can be heterozygous, compound heterozygous, or homozygous for beta-thalassemia, or even have interactions with other hemoglobinopathies. Their phenotypes include microcytic and hypochromic anemia, raised HbA2 levels, and various syndromes caused by the combination of β^0 and β^+ alleles (Thein, 1998; Weatherall, 2001).

The frequency of heterozygotes in Brazil is around 1% (Freitas and Rocha, 1983; Ramalho et al., 1999). Studies in the South and Southeast of Brazil have shown that the most frequent mutations are β^0 39 (C→T) and β^+ IVS-I-110 (G→A) (Martins et al., 1993; Reichert et al., 2008), whereas in the Northeast, an entirely different pattern was observed, the most frequently encountered allele being β^+ IVS-I-6 (T→C), followed by β^0 IVS-I-1 (G→A) (Araújo et al., 2003).

Due to the lack of information regarding the types of beta-thalassemia mutations encountered in the state of Rio Grande do Norte, an effort was made to characterize the disease in patients who were homozygous and heterozygous for beta-thalassemia, by way of hematological and molecular tests.

The sample consisted of 35 unrelated individuals (13 males and 22 females) with HbA2 levels above 3.5%. Age was between 1 and 70 years and all were born in the state of Rio Grande do Norte. The subjects were recruited from individuals referred to the Integrated Laboratory Clinical Analysis, at Rio Grande do Norte Federal University, between April, 2008 and October, 2009, by hematologists from the public and private sectors for the diagnosis of possible anemia. The study was approved by the Research Ethics Committee of Rio Grande do Norte Federal University (CEP-UFRN, protocol number 015/08) in accordance with...
the standards laid down in the National Health Council res-
olution 196/96. All the participants or their legal guardians
were informed of the aim of the study and signed a volun-
tary informed-consent form.

Samples of venous blood from each patient were col-
clected in two sterile tubes, one with EDTA and the other
without an anticoagulant. The aliquot of blood containing
EDTA was used for measurement of red blood cell indices
(ABX Diagnostics, Montpellier, France), alkaline hemo-
globin electrophoresis (Dacie and Lewis, 1995), measure-
ment of HbA2 by elution (Bezerra, 1984), quantification of
HbF by alkaline denaturation (Betke et al., 1959), and DNA
extraction using the illustra blood genomicPrep Mini Spin
commercial kit (GE Healthcare, Little Chalfont, Buckin-
ghamshire, UK). The aliquot without anticoagulant was used
to measure serum ferritin by chemiluminescence (Im-
mulite, Diagnostics Products Co., Los Angeles, CA, USA).

The mutations most commonly associated with the
disease, viz., β^0IVS-I-1 (G→A), β^+IVS-I-6 (T→C) and
β^β39 (C→T), were investigated by PCR-RFLP. PCR was
carried out in a GeneAmp 9700 thermocycler (Applied
Biosystems, Foster City, CA, USA), using the primers
EA74 (5’-GTTTGAAGTCCAUCTCT-3’) and EA72
(5’-CCTCAGTGTGGCAAAGGTG-3’) as described by
Sirvent et al. (1998). The amplified products and digested
samples were visualized by electrophoresis in 2% agarose
gels followed by staining with ethidium bromide, the re-
results then being recorded in a photodocumentation system
(Sambrook et al., 1989). A previously sequenced standard
sample for homozygosis and heterozygosis was used for
each mutation investigated.

Complete beta-gene sequencing, from the 5’ pro-
moter region to the 3’ untranslated region (3’ UTR), was
undertaken when no mutation was otherwise identified by
PCR-RFLP. Sequencing was with primers described by
Miranda et al. (1997), as well as the DYEnamic ET Dye
Terminator Kit (GE Healthcare, Little Chalfont, Buckin-
ghamshire, UK), and a MegaBace 1000 DNA Analysis
System automatic sequencer (Molecular Dynamics,
Amersham Pharmacia Biotech, Sunnyvale, CA, USA).

Descriptive analysis (mean and standard deviation),
as well as the Student’s t-test, Kolmogorov-Smirnov test,
and Levene test were used for statistical analysis with SPSS
for Windows version 10.0 and Microsoft Office Excel
2007. A significance level of (p < 0.05 was used in all the
tests.

Of the 35 patients included in the study, 31 were het-
erozygous and 4 homozygous for beta-thalassemia. Of the
31 heterozygotes, 13 (41.9%) bore the β^0IVS-I-6 (T→C)
mutation and 15 (48.4%) the β^+IVS-I-1 (G→A) mutation,
whereas in 3 (9.7%) no mutation could be identified by
PCR-RFLP. DNA from the latter three patients was sub-
mitted to beta-globin gene sequencing, thereby revealing
two to be heterozygous for the β^+IVS-I-110 (G→A) muta-
tion and one for the β^+IVS-I-5 (G→C) mutation. All the
four homozygous patients bore the β^+IVS-I-6 (T→C) muta-
tion.

Of the 39 thalassemia alleles investigated, 53.8%
bore the IVS-I-6 mutation, 38.5% the IVS-I-1, 5.1% the
IVS-I-110, and 2.6% the IVS-I-5 mutation.

Comparison of hematological analysis between tha-
lassemia patients without iron deficiency and heterozygous
for β^+ IVS-I-6 (T→C) and β^0 IVS-I-1 (G→A) mutations,
revealed a significant difference as regards MCV
(p = 0.023), MCH (p = 0.007), and Hb A2 (p = 0.001) quanti-
fication. Nevertheless, the comparison of laboratory anal-
yses between patients heterozygous and homozygous for
the IVS-I-6 mutation, revealed a statistically significant
difference (p < 0.05) for all the parameters analyzed (Ta-
ble 1).

### Table 1 - Comparison of β^0IVS-I-1 (G→A) and β^+IVS-I-6 (T→C) mutations found in this study with blood indices

| Blood indices                  | Type of mutation | β^0IVS-I-1 (N = 12)* | β^+IVS-I-6 (N = 4) | p value(1,a) | p value(1,b) |
|-------------------------------|------------------|----------------------|-------------------|--------------|--------------|
| Hemoglobin (g/dL)             |                  | 10.0 ± 1.0           | 7.8 ± 0.8         | < 0.001      | < 0.001      |
| Red blood cells (x10³/L)      |                  | 5.21 ± 0.71          | 4.39 ± 0.51       | 0.320        | 0.001        |
| MCV (fL)                      |                  | 63.3 ± 3.8           | 60.0 ± 5.8        | 0.023        | 0.027        |
| MCH (pg)                      |                  | 19.3 ± 1.7           | 17.9 ± 1.5        | 0.007        | 0.006        |
| Hemoglobin A (%)              |                  | 93.3 ± 1.8           | 83.4 ± 5.0        | 0.009        | 0.018        |
| Hemoglobin A₂ (%)             |                  | 5.3 ± 0.6            | 5.7 ± 0.6         | < 0.001      | 0.010        |
| Hemoglobin F (%)              |                  | 1.4 ± 1.8            | 10.9 ± 5.2        | 0.333        | 0.029        |

1(a) p value in the Student t-test for independent samples.
1(b) heterozygous for β^0IVS-I-1 and β^+IVS-I-6 mutations.
2(heterozygous and homozygous for the β^+IVS-I-6 mutation.
3Seven patients (3 heterozygotes with the IVS-I-1 mutation and 4 with the IVS-I-6 mutation) with iron deficiency or whose serum ferritin levels had not been measured, were excluded.
Orkin et al. (1982) identified the pattern of beta-thalassemia mutations in individuals of Mediterranean origin, wherein βIVS-I-110 (G→A), β39 (C→T), βIVS-I-1 (G→A), and βIVS-I-6 (T→C) mutations were the most common. In Portugal, studies have shown that the most frequently found mutations are β39 (C→T), IVS-I-1 (G→A) and IVS-I-6 (T→C), with frequency varying in accordance with the region (Tamagnini et al., 1983; Cabeda et al., 1999; Faustino et al., 1999).

In Brazil, the pattern of beta-globin mutations varies according to the region. In the South and Southeast, the β39 (C→T) and βIVS-I-110 (G→A) mutations are very frequent (Martins et al., 1993; Bertuzzo et al., 1997; Fonseca et al., 1998; Reichert et al., 2003), whereas in the Northeast, the most frequent mutations are βIVS-I-6 (T→C) and βIVS-I-1 (G→A) (Araújo et al., 2003).

This is the first study to determine the profile of beta-thalassemia mutations in the population of Rio Grande do Norte State. Through PCR/RFLP analysis, IVS-I-6 and IVS-I-1 mutations were detected in 36 (92.3%) of the 39 thalassemia alleles analyzed. The remaining three (7.7%) were characterized by beta-globin gene sequencing.

The Brazilian population is one of the most heterogeneous in the world, due to five centuries of interethnic crossing of peoples from three continents, namely, European colonizers, mainly represented by the Portuguese, African slaves and autochthonous Amerindians (Reichert et al., 2008). As regards European immigration, it is estimated that about 500,000 Portuguese arrived in the country between 1500 and 1808. Significantly, in the approximately 100-year-period from 1872 to 1975, Brazil received ever increasing numbers of immigrants from various parts of the world, viz., Italians (34%), Portuguese (29%), Spanish (14%), Japanese (5%), Germans (4%), Lebanese and Syrians (2%), and others (12%) (Pena et al., 2009). The population of Rio Grande do Norte is the result of miscegenation between Amerindians, sub-Saharan Africans, and European colonizers, the African influence having been insignificant. Of the Europeans, the Portuguese exerted the greatest influence, followed by the French. Although the Dutch were also present in the state, their contribution to the genetic makeup of the population was not significant (Cascudo, 1980).

The high frequency of IVS-I-6 (53.8%) and IVS-I-1 (38.5%) mutations appears to have resulted from the Portuguese contribution to the genetic makeup of the population of Rio Grande do Norte. Both these mutations were also frequent in Pernambuco, thereby demonstrating the thus far heterogeneity of beta-thalassemia in Brazil (Araújo et al., 2003).

In the present study, the IVS-I-110 (G→A) mutation was identified in two out of the three patients whose β genes were sequenced. In Brazil, this mutation was the most commonly found in the southern and southeastern region (Martins et al., 1993; Bertuzzo et al., 1997; Reichert et al., 2008). The βIVS-I-5 (G→C) mutation, found in the third patient, and which could not be characterized by PCR-RFLP, was also encountered, with a frequency of 9.3%, in a population studied by Araújo et al. (2003). This mutation is very common in Southeast Asia, especially Malaysia and Indonesia, as well as in various regions of India (Thein, 1998).

PC-RFLP, through its usefulness in identifying the most common beta-thalassemic alleles in the population studied, represents a practical alternative in situations where sequencing is unavailable. As there are no records to date of the types of beta-thalassemia mutations found in Rio Grande do Norte, and as studies have shown that the pattern of such mutations varies according to ethnic influence in the different regions of Brazil, the authors believe this study will play an important role in acquiring a greater understanding of the molecular profile of beta-thalassemia in Brazil.

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