Dear Editor,

Interferons (IFNs) are a group of secreted autocrine and paracrine proteins released by host cells that impart resistance against viral infections. IFNs that are produced by virus-infected cells help other cells in the proximity to heighten their anti-viral defences. Based on their affinity to bind with various receptors, IFNs are classified into three classes, namely, Type I, Type II, and Type III IFNs (Borden et al., 2007). These unique signaling proteins facilitate the transcription of more than 300 IFN-inducible genes, which are also referred to as IFN-stimulated genes (ISGs), that actively participate in the process of viral protein synthesis inhibition (Hsu et al., 2015).
Chronic obstructive pulmonary disease (COPD) is the third leading cause of death worldwide. In a given population aged more than 45 years, COPD affects at least 10 % of them, whereas, it affects more than 50 %, among chronic smokers (Chellappan et al., 2020, Mehta et al., 2020). Chronic airway inflammation in COPD is mediated by pro-inflammatory cytokines which include several IFNs (Chin et al., 2020). Furthermore, the expression levels of interferon stimulated genes – myxovirus resistance gene A (ISG MxA), 2'-5'-oligoadenylate synthase 1, and viperin genes are reduced in severe COPD patients.

Influenza A virus (IAV) infected human primary airway epithelial cells (hPCE) isolated from active smokers have shown impaired antiviral responses compared to hPCE isolated from non-smokers. IAV triggers the release of an array of IFNs, primarily, IFN-λ 1, IFN-λ 2/3 and IFN-β, which in turn stimulate a robust antiviral response in hPCE from non-smokers. In contrast, hPCE from smokers showed inhibition of IFN-λ 1, IFN-λ 2/3 and IFN-β mRNA responses by 96 %, 95 % and 85 %, respectively (Wu et al., 2016). Moreover, there is a downregulation of IFN-β signaling in the cell-free sputum supernatant of smokers, when compared to people who have never smoked.

Cigarette smoke extract (CSE) may activate serine phosphorylation driven ubiquitination and degradation of the IFN alpha/beta receptor 1 subunit of the Type I IFN receptor. This may lead to reduced IFN signaling cascade and a corresponding amount of suppression in the resistance against viral infections (HuangFu et al., 2008). Therefore, COPD patients are likely to experience acute infectious exacerbations leading to an increased risk of mortality/morbidity, as well as, reduced quality of life and poor prognosis. Moreover, immunolabeling scores of IFN-β and its main transcription factor (IRF-7) were reduced up to 65 % and 74 % respectively, in epithelial tissues and alveolar macrophages in COPD patients, when compared with non-COPD patients (Garcia-Valero et al., 2019).

The use of experimental approaches involving in vitro cell culture systems or in vivo animal models have revealed key mechanistic insights, which support the fact that, cigarette smoke dampens Type I IFN responses followed by an exaggerated inflammatory response to viral infections. An in vivo investigation that employed the elastase-induced experimental emphysema mouse model with C57BL/6J and BALB/cJ mouse strains has reported that, the BALB/cJ mice had high levels of interleukin (IL)-17A mRNA and a number of classically (M1) and alternatively (M2) activated macrophage genes, whereas, C57BL/6J mice showed high levels of IFN-γ. This study demonstrated that mice with different genetic backgrounds show contrasting susceptibility to the progression of emphysema, which is a common feature in COPD (Limjunyawong et al., 2015).

Increased airway inflammation was observed in mice exposed to cigarette smoke followed by chronic infection with non-typeable Haemophilus influenzae (NTHi) virus compared to air-exposed controls. NTHi-specific T cells from cigarette smoke-exposed mice generated lower levels of IFN-γ and IL-4 (Lugade et al., 2014). IFN-γ may modulate macrophages to increase the response to toll-like receptor (TLR) ligands, such as LPS. IFN-γ increases the expression levels of TLR2 and TLR4. It further activates the Janus Kinase (JAK)/Signal Transducer and Activator of Transcription (STAT) signaling pathway in alveolar macrophage. IFN-γ mediated cytokine-expression and STAT1 activation were resistant to corticosteroids (dexamethasone) while, JAK/STAT1 inhibition repressed the effect of IFN-γ. Therefore, targeting IFN-γ signaling by JAK inhibitors could be a promising novel therapeutic approach against COPD.

In an ex vivo study conducted with bronchoalveolar lavage (BAL) cells (95 % were macrophages) collected from 7 COPD patients and 10 healthy control subjects, the cells infected with live rhinovirus resulted in a 50 % impairment in IFN-α and IFN-λ release from the COPD group compared to control. Similarly, the level of IFN-β stimulated by rhinovirus was remarkably higher in the healthy control subjects compared with the COPD group. The low levels of IFNs
in COPD were correlated with impairment in the induction of the ISG CXCL10 (Mallia et al., 2011).

Another study that employed primary bronchial epithelial cells isolated from COPD and healthy volunteers, who were infected with Rhinovirus-1B, showed that the cells from COPD patients had a higher expression of interleukin (IL)-6, IFNs-β and λ1 when compared to the cells from healthy subjects. The pre-treatment of IFN-β/λ1 to cells from COPD patients did not change IFN-β expression but resulted in increased levels of IFN-λ1 leading to suppression of viral replication. However, this did not raise pro-inflammatory cytokines (Baines et al., 2013). Moreover, in vitro experiments conducted in both human alveolar basal epithelial adenocarcinoma cell line (A549) and primary bronchial epithelial cells showed that, IFN-β exerts not only a robust, but also a prolonged protective effect against Rhinovirus infection. The viral RNA was found to be reduced in both cell lines after 18 hours of exposure with IFN-β followed by infection with rhinovirus for the next 72 hours (Gaajetaan et al., 2013).

Similarly, in a lung organ culture model, CSE exposure led to inhibition of IFN-inducible protein-10, and mRNA expression. Similarly, there was a reduction of IFN-β mRNA by CSE. This suppression was due to oxidative inhibition of viral-mediated induction of the pattern recognition receptor RIG-I (Wu et al., 2011). Stimulation of the RIG-I/mitochondrial antiviral signaling protein pathway by intracellular delivery of polyinosinic: polycytidylic acid (with liposomes or via nucleofection) upregulated IFN-β, mainly through the TLR3/TRIF pathway in primary airway epithelial cells with low co-stimulation of IL-8. This study highlights the feasibility of augmenting the production of IFN-β by airway epithelial cells without excessive co-stimulation of IL-8 (Dauletbaev et al., 2015).

A collective and comprehensive understanding of the role of IFNs in the pathogenesis of COPD may provide us with a promising platform to understand the in-depth disease mechanism and would provide a logical understanding to investigate potential therapeutic alternatives.

**Conflict of interest**

None.

**REFERENCES**

Baines KJ, Hsu AC, Tooze M, Gunawardhana LP, Gibson PG, Wark PA. Novel immune genes associated with excessive inflammatory and antiviral responses to rhinovirus in COPD. Respir Res. 2013;14:15.

Borden EC, Sen GC, Uze G, Silverman RH, Ransohoff RM, Foster GR, et al. Interferons at age 50: past, current and future impact on biomedicine. Nat Rev Drug Discov. 2007;6:975-90.

Chellappan DK, Yee LW, Xuan KY, Kunalan K, Rou LC, Jean LS, et al. Targeting neutrophils using novel drug delivery systems in chronic respiratory diseases. Drug Dev Res. 2020;81:419-36.

Chin LH, Hon CM, Chellappan DK, Chellian J, Madheswaran T, Zeehan F, et al. Molecular mechanisms of action of naringenin in chronic airway diseases. Eur J Pharmacol. 2020;879:173139.

Dauletbaev N, Cammisano M, Herscovitch K, Lands LC. Stimulation of the RIG-I/MAVS Pathway by polyinosinic:polycytidylic acid upregulates IFN-beta in airway epithelial cells with minimal costimulation of IL-8. J Immunol. 2015;195:2829-41.

Gaajetaan GR, Geelen TH, Vernooy JH, Dentener MA, Reynaert NL, Rohde GG, et al. Interferon-beta induces a long-lasting antiviral state in human respiratory epithelial cells. J Infect. 2013;66:163-9.

Garcia-Valero J, Olloquequi J, Montes JF, Rodriguez E, Martin-Satue M, Texido L, et al. Deficient pulmonary IFN-beta expression in COPD patients. PLoS One. 2019;14(6):e0217803.

Hsu AC, Starkey MR, Hanish I, Parsons K, Haw TJ, Howland LJ, et al. Targeting PI3K-p110alpha Suppresses Influenza Virus Infection in Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med. 2015;191:1012-23.

HuangFu WC, Liu J, Harty RN, Fuchs SY. Cigarette smoking products suppress anti-viral effects of Type I interferon via phosphorylation-dependent downregulation of its receptor. FEBS Lett. 2008;582:3206-10.
Limjunyawong N, Craig JM, Lagasse HA, Scott AL, Mitzner W. Experimental progressive emphysema in BALB/cJ mice as a model for chronic alveolar destruction in humans. Am J Physiol Lung Cell Mol Physiol. 2015;309:L662-76.

Lugade AA, Bogner PN, Thatcher TH, Sime PJ, Phipps RP, Thanavala Y. Cigarette smoke exposure exacerbates lung inflammation and compromises immunity to bacterial infection. J Immunol. 2014;192:5226-35.

Mallia P, Message SD, Gielin V, Contoli M, Gray K, Kebadze T, et al. Experimental rhinovirus infection as a human model of chronic obstructive pulmonary disease exacerbation. Am J Respir Crit Care Med. 2011;183:734-42.

Mehta M, Chellappan DK, Wich PR, Hansbro NG, Hansbro PM, Dua K. miRNA nanotherapeutics: Potential and challenges in respiratory disorders. Future Med Chem. 2020;12:987-90.

Wu W, Patel KB, Booth JL, Zhang W, Metcalf JP. Cigarette smoke extract suppresses the RIG-I-initiated innate immune response to influenza virus in the human lung. Am J Physiol Lung Cell Mol Physiol. 2011;300:L821-30.

Wu W, Zhang W, Booth JL, Hutchings DC, Wang X, White VL, et al. Human primary airway epithelial cells isolated from active smokers have epigenetically impaired antiviral responses. Respir Res. 2016;17:111.