Review Article

Targeting Molecular Mediators of Ferroptosis and Oxidative Stress for Neurological Disorders

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With the acceleration of population aging, nervous system diseases including Alzheimer’s disease (AD), Parkinson’s disease (PD), Huntington’s disease (HD), anxiety, depression, stroke, and traumatic brain injury (TBI) have become a huge burden on families and society. The mechanism of neurological disorders is complex, which also lacks effective treatment, so relevant research is required to solve these problems urgently. Given that oxidative stress-induced lipid peroxidation eventually leads to ferroptosis, both oxidative stress and ferroptosis are important mechanisms causing neurological disorders, targeting mediators of oxidative stress and ferroptosis have become a hot research direction at present. Our review provides a current view of the mechanisms underlying ferroptosis and oxidative stress participate in neurological disorders, the potential application of molecular mediators targeting ferroptosis and oxidative stress in neurological disorders. The target of molecular mediators or agents of oxidative stress and ferroptosis associated with neurological disorders, such as reactive oxygen species (ROS), nuclear factor erythroid 2-related factor-antioxidant response element (Nrf2-ARE), n-acetylcysteine (NAC), Fe2+, NADPH, and its oxidases NOX, has been described in this article. Given that oxidative stress-induced ferroptosis plays a pivotal role in neurological disorders, further research on the mechanisms of ferroptosis caused by oxidative stress will help provide new targets for the treatment of neurological disorders.

1. Introduction

As a novel mode of regulated cell death (RCD), ferroptosis was firstly proposed by Dixon in 2012 [1]. Ferroptosis is an iron-dependent and nonapoptotic cell death mode, which is caused by excessive accumulation of ROS and imbalance of cell lipid oxide metabolism. In 2018, the Nomenclature Committee on Cell Death (NCCD) advised defining ferroptosis as RCD caused by oxidative stress in the cell microenvironment which can be regulated by glutathione peroxidase 4 (GPX4). Moreover, ferroptosis can be inhibited by iron chelators, iron intake inhibitors, and lipophilic antioxidants [2, 3]. Recent studies have shown that ferroptosis is obviously different from traditional programmed cell death. It has distinctive morphological features and biochemical features [4], such as morphological changes of mitochondrial, accumulation of iron, and lipid reactive oxygen species (ROS). At present, it is believed that the main cause of ferroptosis to cell death is the inactivation of the cellular antioxidant system, which includes the inhibition of cystine/glutamate antiporter system (system Xc−) and GPX4, disrupted iron homeostasis, and lipid peroxidation. Ultimately, the decrease of antioxidant capacity and the accumulation of intracellular lipid ROS lead to oxidative cell death [5].

Although there are many studies about ferroptosis, the mechanism of ferroptosis is not clear yet. Ferroptosis was identified to involve in oxidative stress-induced cell death [6]. During the process of ferroptosis, cellular over-accumulation of ROS eventually generates oxidative stress and cell death. According to current research, the possible mechanisms of ferroptosis include oxidative stress, lipid peroxidation, iron metabolism, and other mechanisms to be explored. The process of oxidative stress in ferroptosis is complex that is regulated by a variety of regulatory factors.

Neurological disorders have become the major leading cause of disability and the second leading cause of death...
worldwide [7]. Over the past few decades, due to the lack of effective precaution and treatment, the mortality and the disabled owing to neurological diseases have increased, particularly in low-income and middle-income countries [8]. The death and disability caused by neurological disorders have caused a huge burden on families and countries. Meanwhile, it has become a serious social problem. In order to alleviate the burden, there are increasing studies on how to treat or prevent the process of neurological disorders.

Mounting evidence has shown that oxidative stress-induced ferroptosis plays a significant role in neurological disorders [3]. Oxidative stress is both an important process of ferroptosis and the pathogenesis of neurological disorders. Hence, oxidative stress has become a critical linking between ferroptosis and neurological disorders. Up to now, the inhibitors and activators of ferroptosis [9–14], as well as other involved signaling pathways [15–17], have been widely studied. Furthermore, the mediators of oxidative stress might become a potential therapeutic target for the treatment of neurological disorders by regulating the process of ferroptosis. In this review, we will briefly summarize the mechanisms of ferroptosis, and highlight the role of targeting molecular mediators of oxidative stress in neurological disorders, so as to provide an option for the therapeutic application of ferroptosis in neurological disorders.

2. The Mechanism of Ferroptosis

Ferroptosis is a unique kind of nonapoptotic-regulated cell death, which is distinct from apoptosis, autophagy, and necrosis in morphology, biochemistry, and genetics. Iron metabolism and lipid peroxidation are two essential events in ferroptosis [2]. Additionally, system Xc⁻ and GPX4 are considered to be the primary signaling pathways [18]. Here, we summarize the main mechanisms involved in the process of ferroptosis, such as oxidative stress, lipid peroxidation, iron metabolism, and the signaling pathways (Figure 1).

2.1. Oxidative Stress in Ferroptosis. Oxidative stress was formulated in 1985 [19], which is an important cause leading to neurological disorders that mainly arise from the imbalance between depletion of antioxidants and production of peroxides [20]. Ferroptosis is a unique, oxidative stress-induced cell death pathway characterized by glutathione depletion and lipid peroxidation. Based on current research, ferroptosis can be regulated by system Xc⁻ [21]. System Xc⁻ can exchange glutamate and cystine inside and outside of the cell. Glutathione is an important free radical scavenger and antioxidant in vivo, which can be categorized as either reduced (GSH) or oxidized (GSSG). Intracellular cystine can be converted into reduced GSH by a series of biochemical reactions. GPX4 is a member of the glutathione peroxidase family which can convert GSH to GSSG, and GSH/GSSG constitutes an antioxidant system and provides reducing equivalents to eliminate oxidative species [22]. GPX4 is a redox enzyme and can also inhibit ferroptosis by decreasing the level of lipid peroxides. GPX4 plays a crucial role in reducing reactive aldehydes (PUFAs-OH) to their alcohol form (PUFAs-OH) which can reduce the content of ROS [18].

ROS are some of the most common oxidants in cells. ROS includes superoxide (O₂⁻), hydrogen peroxide (H₂O₂), lipid peroxides (ROOH), or the corresponding hydroxyl (HO•) and peroxyl radicals (ROO•) [23]. Accumulation of ROS can lead to oxidative stress, causing oxidative stress-induced lipid peroxidation. What is worse, lipid peroxidation may further generate ROS or degrade into reactive compounds capable of crosslinking DNA and proteins [24]. Oxidative stress-induced lipid peroxidation eventually leads to ferroptosis. Meanwhile, ferroptosis can be induced by oxidative stress directly (Figure 1).

2.2. Iron Metabolism in Ferroptosis. Metabolism is essential for the biochemical process in cells and goes awry in many diseases. Iron metabolism is an important mechanism of ferroptosis [2]. Iron is a significant trace element in the body [25]. Abnormal distribution and content of iron in the body can affect the normal physiological processes. Fe²⁺ is absorbed by the intestine or degraded by erythrocyte and then can be oxidized to Fe³⁺ by ceruloplasmin [26]. Fe³⁺ can combine with transferrin (Tf) on cytomembrane to form Fe³⁺-Tf, which can be endocytosed into cells to form a complex with membrane protein TF receptor (Tfr) [26]. The complex enters the endosome and is divided into TF-TFR and Fe³⁺ [26]. Fe³⁺ is then reduced to Fe²⁺ by the six-transmembrane epithelial antigen of the prostate 3 (STEAP3), and Fe²⁺ is stored in the unstable iron pool (LIP) and ferritin [26]. Nevertheless, under pathological conditions, iron homeostasis is destroyed, and the overwhelming production of Fe²⁺ exceeds the limit, breaking cell homeostasis [27]. Excess Fe²⁺ can combine with H₂O₂, reducing H₂O₂ to hydroxyl radical (OH⁻) by Fenton reaction [28, 29], which was first described by Fenton [30], who in 1894 reported on the oxidation of malic acid by hydrogen peroxide in the presence of ferrous ions.

2.3. Role of Mitochondria in Ferroptosis. Mitochondria, through the tricarboxylic acid cycle (TCA) and electron transport chain (ETC) activity, are the principal sources of cellular energy production [31]. Meanwhile, ROS can be produced during the process of TCA which is an important cause of ferroptosis-induced lipid peroxides [32]. In addition, mitochondrial morphological changes are considered to be an important basis for the diagnosis of ferroptosis [1]. The change of mitochondrial membrane potential is also an important discovery in the study of ferroptosis [33, 34]. To date, abnormal mitochondrial architecture including mitochondrial fragmentation, rupture of mitochondrial outer membrane, and mitochondrial shrinkage, as well as vanished mitochondrial cristae, is regarded as the typical morphological characteristic of ferroptosis [1]. In summary, mitochondria play a significant role in ferroptosis [35], and molecular research targeting mitochondria is of great significance to intervene in the development of ferroptosis (Figure 1).

2.4. Other Mechanisms. As a novel form of cell death, the mechanism of ferroptosis is complex. Ferroptosis is regulated by numerous pathways and implicated in many
diseases. In addition to the mechanisms mentioned above, there are other possible mechanisms which have been proposed, such as the antioxidant mechanisms which include GPX4 pathway [36], FSP1 pathway [37], and other pathways. FSP1-NADPH-CoQ pathway [38] and nuclear factor erythroid 2-related factor 2 (Nrf2) pathway are potential regulatory pathways of ferroptosis [39]. Hence, the mechanism of ferroptosis remains to be explored.

## 3. Ferroptosis in Neurological Disorders

In recent years, the incidence of neurological disorders increases substantially. Because of the lack of effective treatment, sequelae caused by neurological disorders have brought an enormous burden on society. Ferroptosis has been proved to be closely related to the occurrence and development of various neurological disorders [40–43]. Therefore, figuring out the role of ferroptosis in neurological disorders will provide an important basis for the treatment of diseases (Figure 1, Table 1).

### 3.1. Ferroptosis in Parkinson’s Disease

Parkinson’s disease (PD), also called paralysis agitans, is the second most common neurodegenerative disorder worldwide. It is typical of neuronal death in the substantia nigra pars compacta (SNpc), which adjusts motor function. PD causes clinical symptoms such as resting tremor, rigidity, bradykinesia, postural instability, and other motor symptoms [44–46].
Iron is considered to be an important target of neurodegenerative diseases. Besides, iron metabolism is closely related to the pathogenesis of neurodegenerative diseases [34, 47]. Researchers found that patients with PD suffered from GSH depletion, ROS elevation, and lipid peroxidation [48–50]. Ferritin heavy chain 1 (FTH1), a main iron storage protein, can affect intracellular iron metabolism and then trigger ferroptosis. As an important ferroptosis-related protein, FTH1 is differentially expressed in rats with PD compared with normal rats. Overexpression of FTH1 can reduce the effect of ferroptosis in PD-related cells [51]. These findings indicate that ferroptosis may be used as a therapeutic target for PD [52, 53]. Neuroimaging and post-mortem examination report that much iron accumulation in substantia nigra (SN) results in an increase in iron content in the residual dopaminergic neurons [54]. α-Synuclein causes iron cytotoxicity by acting on mitochondria in the neurodegeneration of PD. A study found that cyclosporine A, a blocker of mitochondrial permeability transition pore (mPTP), could prevent iron-induced mitochondrial damage and cell death. Furthermore, knocking down α-synuclein expression by siRNA can play a similar role [55]. α-Synuclein is the main component of Lewy bodies and is abundantly expressed in the nervous system, which is strongly related to PD’s pathophysiology [56]. Besides, it has been recently shown that α-synuclein causes lipid peroxidation by producing ROS, leading to increased calcium influx and consequent cell death [57]. Ferroptosis inhibitors like ferrostatin or iron chelators [58] can inhibit cell death caused by the above method, supporting the hypothesis that ferroptosis participates in this process and may harbor therapeutic potential. Studies have also shown that ferroptosis occurs earlier than apoptosis in the development of PD and ferric ammonium citrate (FAC)-induced ferroptosis was dependent on p53, not MAPK signaling pathway [54].

### 3.2. Ferroptosis in Alzheimer’s Disease

Alzheimer’s disease (AD) is one of the most common neurodegenerative diseases worldwide. Pathological features of AD include cerebral atrophy, intraneuronal accumulation of hyperphosphorylated tau in neurofibrillary tangles, extracellular deposition of amyloid-β peptide in senile plaques, oxidative stress, chronic inflammation, and loss of neurons and synapses [59]. There is evidence that iron excess and homeostasis disorder can lead to neurodegeneration of AD [60, 61]. The increase of brain iron content in patients with AD has been confirmed in many studies [62–67]. When brain iron regulation in AD patients is disturbed, excess Fe$^{2+}$ can not only produce hydroxyl radicals in Fenton reaction but also induce neuroinflammation, resulting in oxidative stress and neurodegeneration mediated by ferroptosis [68–72]. The research found that the increase of ferritin light chain (FTL) is related to the decrease of GPX4 level in AD which indicates that dysfunctional ferritin will reduce the antioxidant capacity of brain and the level of glutathione will decrease in AD patients [73]. Increased light subunit (xCT) expression in cells of patients with AD [74] and xCT can be upregulated by Nrfa [75, 76]. Furthermore, mice with a conditional deletion of GPX4 show cognitive impairment and hippocampal degeneration similar to patients with AD and have the neurodegenerative characteristics of ferroptosis. Simultaneously, these symptoms can be improved by ferroptosis inhibitors [77], which provide a basis for the association between AD and ferroptosis. Recently, researchers observed that tau can stabilize ferroportin (FPN1) which is the only iron export channel found in mammalian cells [78, 79]. FPN decreased significantly in the brain tissue of AD patients. According to MMSE (mini-mental state examination) scores, FPN is involved in the cognitive impairment and brain atrophy of AD [80]. In patients with AD and a mouse model of AD, NADPH oxidases 4 (NOX4) causes lipid peroxidation and promotes ferroptosis of astrocytes via the damage of mitochondrial metabolism in AD [81]. Bhatia et al. [82] summarized a variety of multilayer targeted ligands with iron chelation properties, which is potentially useful in AD. Ashraf et al. [83] summarized the clinical cases involving iron chelators, antioxidants, NAC, and selenium in the treatment of AD and provided a feasible basis for the treatment of AD.
These studies have shown that ferroptosis plays an important role in AD, in which a variety of regulatory factors are involved, and provide many potential therapeutic targets for the treatment of AD.

3.3. Ferroptosis in Huntington’s Disease. Huntington’s disease (HD) is an autosomal dominant, late-onset, and fatal neurodegenerative disorder [84], which is caused by an abnormal CAG repeat in the huntingtin gene [85]. It is characterized by highly selective striatal injury, leading to dance-like movement, progressive dementia, and dystonia. In 2001, Pigeon et al. [86] first identified the association between hepcidin and iron homeostasis. Studies found that hepcidin could promote ferroptosis through iron metabolism [87] and could prevent erastin-induced ferroptosis by degrading Fpn [88]. Qian et al. [89] summarized the therapeutic potential of hepcidin in neurodegenerative diseases, including HD, indicating that there is a link between ferroptosis and HD. A study found that there is an accumulation of toxic iron in neurons of mouse model with HD and the symptoms of HD can be alleviated by deferoxamine, indicating that the accumulation of iron may contribute to the neurodegenerative process [90]. Studies have shown that HD patients will cause oxidative stress and neurotoxicity to neurons in the striatum, resulting in neuronal death, and eventually leading to motor and cognitive impairment [91]. HD patients have shown higher levels of plasma lipid peroxidation and lower GSH levels [92]. Oxidative stress and lipid peroxidation are important mechanisms leading to ferroptosis. In 2017, Cardoso et al. [93] proposed that as a key factor in regulating ferroptosis, GPX4 could provide protective mechanisms against neurodegeneration. Skouta et al. [94] found that ferroptosis inhibitor Fer-1 and its derivatives can prevent cell death in HD brain slice model. Mi et al. [85] summarized the evidence of ferroptosis involved in HD in different animal models or human patients. It revealed the close connection between ferroptosis and HD which would provide an important potential target for the treatment of HD.

3.4. Ferroptosis in Anxiety and Depression. With the development of society, the incidence rate of anxiety and depression is increasing year by year. The research on the mechanism and the treatment of anxiety and depression have also attracted extensive attention. Jiao et al. [95] found that the contents of total iron and ferrous ion in the hippocampus of chronic unpredictable mild stress (CUMS) model mice increased, and GPX4, FTH1, ACSL4, and COX-2 also changed significantly, which proved the evidence of ferroptosis in the hippocampus of the depression mouse model. Previous research reported that sodium hydrosulphide (NaHS) can alleviate the depressive and anxiety-like behaviors induced by type 1 diabetes mellitus (T1DM) [96]. Wang et al. [97] revealed that NaHS reduced ferroptosis in the prefrontal cortex (PFC) of the T1DM mouse model by reducing iron deposition and oxidative stress, increasing the expression of GPX4 and SLC7A11, thus alleviating the anxiety-like and depression-like behavior of the mouse model with T1DM. Moreover, there is increasing evidence that oxidative stress has close contact with anxiety and depression [98, 99]. As a free radical scavenger [100], Edaravone (3-methyl-1-phenyl-2-pyrazolin-5-one, EDA) can improve depression and anxiety-like behavior, as well as inhibit oxidative stress and neuroinflammation in a mouse model of depression, and knocking down of GPX4 expression in CSDs mouse model can inhibit the therapeutic effect of EDA on depression and anxiety [101]. The study has shown that GPX4 can affect the role of EDA in mouse models of anxiety and depression, which suggests that GPX4-mediated ferroptosis may be a potential mechanism affecting anxiety and depression.

3.5. Ferroptosis in Stroke. Stroke is mainly divided into ischemic stroke and intracerebral hemorrhage (ICH). Zhang et al. [102] summarized the multiple roles of ferroptosis in stroke. Jin et al. [104] reviewed some neuroprotectants that show protective effects in stroke models. These substances have been recently validated as ferroptosis inhibitors. These researches indicate that ferroptosis plays a key role in the progression and toxicity of stroke. Chen et al. [105] analyzed and identified ferroptosis-related differentially expressed genes (DEGs) in ischemic stroke by bioinformatics, which provided more evidence for the important role of ferroptosis in ischemic stroke. NAC is considered as a potential therapeutic agent of ferroptosis. Recently, studies have found that NAC can reduce neuronal death and improve functional recovery in mouse models with ICH by inhibiting ferroptosis [106]. Clinical trials and experimental studies have reported that treatment with natural compounds could reduce oxidative stress in ischemic stroke [107]. Therefore, reducing the production of ROS during reperfusion is the key to the treatment of ischemic stroke. Guan et al. found that natural product carvacrol can increase the expression of GPX4 to inhibit ferroptosis, protecting the cognitive function of mice with ischemia-reperfusion injury [108]. In addition, Cui et al. revealed that ACSL4 can exacerbate ischemic stroke by promoting ferroptosis-induced brain injury and neuroinflammation [109]. The incidence rate of ICH is low, but the deformity and mortality rates are significantly higher than ischemic stroke [110]. ICH can lead to the rupture of blood vessels in the brain and release red blood cells to produce a large amount of free iron and ROS, which result in oxidative stress and ultimately neuronal damage [111]. Iron chelator deferoxamine (DFO) can effectively alleviate ICH-induced neuronal injury in mice [112], which also provides evidence for the above process. The same characteristics of ferroptosis were observed in the animal model of ICH [113]. Furthermore, selenium can drive adaptive transcription against ferroptosis to protect neurons and provide direction for the treatment of stroke [114].

3.6. Ferroptosis in TBI. As one of the acute central nervous system (CNS) trauma [115], traumatic brain injury (TBI) is a major cause of death and disability worldwide [116]. Owing to the difficulty of treatment and serious sequelae, TBI has always been a huge problem in the medical field and caused a huge burden on families and society [41]. Analyzing the mouse TBI model [117] and clinical cases with TBI [118], abnormal regulation of ferroptosis after TBI was found both in experimental model and clinical cases.
Interestingly, Xie et al. found that the content of iron in damaged cortical cells increased, the function of iron metabolism was impaired, lipid ROS accumulation was increased, and mitochondrial atrophy was enhanced. The above symptoms can be alleviated by intraventricular administration of ferroptosis inhibitor Fer-1 in a mouse model with TBI [119], which proves that ferroptosis is one of the causes of TBI. Previous studies have shown that mir-212-5p is highly expressed in the brain [120] and is associated with a variety of neurological diseases [77, 119, 121, 122]. The use of mir-212-5p in the CCI mouse model can target PTGS2 to reduce ferroptosis and protect the brain [123]. Moreover, Cheng et al. [124] found that the use of iron uptake inhibitor ferritin II in mouse model with TBI can inhibit the formation of neurological diseases [77, 119, 121, 122]. The use of miR-212-5p in the CCI mouse model can target PTGS2 to reduce ferroptosis and protect the brain [123]. Moreover, Cheng et al. [124] found that the use of iron uptake inhibitor ferritin II in mouse model with TBI can inhibit the formation of iron proteasomes and alleviate TBI injury in brain. For a long time, due to the lack of clear understanding, the treatment of TBI is very difficult. These studies provide a potential theoretical basis for the treatment of TBI.

4. Mediators of Oxidative Stress in Neurological Disorders

Oxidative stress can spread, which aggravates the production of ROS. Due to the high oxygen demand and high energy demand of the brain, but its antioxidant capacity is weak, it is more vulnerable to oxidative stress [125], which will lead to various neurological diseases. Therefore, inhibiting or weakening oxidative stress might effectively reduce the damage to the nervous system. Studies for targeting molecular mediators of oxidative stress are considerable and will provide effective treatment for neurological disorders (Figure 2).

4.1. Nrf2-ARE-Mediated Oxidative Stress. Nuclear factor erythroid 2-related factor (Nrf2) is a transcription factor belonging to the basic leucine zipper (bZIP) transcription factor family that can participate in the regulation of oxidative stress. The antioxidant capacity of Nrf2 works by inducing glutathione biosynthesis [126]. Studies found that Nrf2 can play a neuroprotective role in central nervous system (CNS) diseases [127–129]. In addition to the transcriptional regulation of Nrf2, current hot research directions for the activity of Nrf2 are Kelch-like ECH-associated protein 1 (Keap1)/Nrf2/antioxidant response element (ARE) pathway. Keap1/Nrf2/ARE can be divided into two parts, one in the cytoplasm and the other in the nucleus. Normally, Keap1 and Nrf2 bind in the cytoplasm and are in an inactive state. If they are not activated all the time, Nrf2 will be ubiquitinated and then degraded. Under certain stimulation, the binding of Keap1-nrf2 will be unstable. Nrf2 will be released, transferred to the nucleus, combined with ARE, activate the transcription of downstream genes, and then translate a series of related proteins, which can induce various antioxidant enzymes, including heme oxygenase 1 (HO-1), NAD (P) H: quinone oxidoreductase 1 (NQO1), superoxide dismutase (SOD), glutathione peroxidase (GPX), and catalase (CAT). These enzymes can play physiological functions [130]. Targeted regulators of Nrf2 are of great significance in the treatment of oxidative stress-induced injury in neurological disorders. Nrf2 plays a neuroprotective role by regulating the antioxidant system of cells. The regulation will have an impact on glutathione (GSH), thioredoxin (TXN), NADPH, and various other ROS-related enzymes [131, 132].

Keap1/Nrf2/ARE pathway has been proved to be an important antioxidant target. Keap1 can perceive ROS levels through cysteine-rich regions [133–135]. In the steady state, Nrf2 in the cytoplasm can be degraded by Keap1, and Nrf2 increases rapidly under stress and transfers to the nucleus for transcriptional activity [136]. Mitoquinone (MitoQ) is a mitochondrial-targeted antioxidant and can cause a sharp decline in Keap1 in cells, prompting more Nrf2 to be transmitted to the nucleus to produce an antioxidant response. The effect was confirmed in subarachnoid hemorrhage (SAH) animal model experiments [137]. Nrf2 can also play a protective role as a therapeutic target in astrocytes. There are seven functional domains of Nrf2 (neh1-neh7) that regulate its transcriptional activity and stability [138]. Kelch-like ECH-associated protein 1 (Keap1) is an inhibitor of Nrf2 [139, 140] and interacts with Nrf2 through neh2 domain [141]. Early studies believed that Keap1 inhibition was the main mechanism to activate astrocytes. Recent studies have found that mild oxidative stress (nonlethal oxidative stress) can also activate Nrf2 independently through neh5 transactivation domain [142]. In addition to the various regulatory factors mentioned above, there are other Keap1/Nrf2 pathway regulators such as microRNA-592 (miR-592) [143], ericidocytol [144], and tripartite motif 16 (TRIM16) [145]. These regulatory factors can act on Keap1/Nrf2/ARE pathway to play the role of antioxidant stress, so as to treat or alleviate neurological disorders (Figure 2).

4.2. NADPH- and NOX-Mediated Oxidative Stress. Nicotinamide adenine dinucleotide-phosphate (NADPH) is a coenzyme, which is the product of the final electron acceptor NADP + after receiving electrons. It participates in a variety of biochemical reactions as a hydrogen transmitter in vivo. Studies have confirmed that NADPH can resist oxidative stress and improve energy metabolism, so as to play a neuroprotective role in stroke [146]. NADPH oxidases (NOXs) have 7 kinds of subtypes.

H₂O₂ can interact with mutant huntingtin in the brain of HD patients [147]. The activity of NOX can be measured by lucigenin-enhanced luminescence method [148]. The heterozygous huntingtin (HTT) and Cd can induce cytotoxicity which will cause NOXs-mediated oxidative stress. As an exogenous antioxidant and NOX inhibitor, apocynin can block the process of oxidative stress in the process of HD [149]. So far, there are still few studies on the role of NOX in the process of HD. As a potential therapeutic target of HD, the mechanism of NOX and its way of alleviating HD patients deserve further exploration.

Studies found that the cerebellum is an important participant in mental illness and is affected by oxidative stress. Schiavone et al. found that inhibition of NOX during brain maturation can prevent the development of psychotic behavioral dysfunction. Administration of ketamine to the postpartum mouse model can enhance oxidative stress and reduce IL-10 in this region [150]. In recent years, many compounds that inhibit NOX have been studied for the
treatment of neurological disorders, and Ganguly et al. [191] reviewed the relationship between NOX and AD, and summarized drug trials of NOX inhibitors in Alzheimer’s disease and its precursor conditions. The results displayed that the activity of NOX increased in the brain of mouse models with neurodegenerative diseases and AD patients [192-194]. However, the effects of NOX inhibitors and their precursors are not satisfactory, and the therapeutic results of these naturally obtained compounds for AD are quite variable, including berberine and blueberry-derived polyphenols. Because of the complexity of neurological disorders, the study of NOX inhibitors can provide a new direction for the treatment of neurological disorders (Figure 2).

4.3. Other Mediators and Potential Therapeutic Agents. In this part, we mainly summarize three common regulators of targeted oxidative stress in neurological disorders. In addition to the above three common regulators, other regulators can be used to treat or relieve the symptoms of neurological disorders, such as PINK1, Ca²⁺, Sigma-1 receptor (sig-1R), peroxidase (prx), toll-like receptors (TLRs), and alarmins/c-jun N-terminal kinase (JNK) [151]. PINK1 is a mitochondrial-targeted E3 ubiquitin ligase, which deficiency can lead to mitochondrial damage, oxidative stress, and exert neuroprotective effects by activating Parkin. Ca²⁺ can consume GSH to regulate cell death [152]. Sigma-1 receptor (sig-1R) [153, 154] is a mitochondrial endoplasmic

![Figure 2: The top schematic is a summary of oxidative stress mechanisms in neurological disorders. Under homeostatic conditions, Nrf2 binds with Keap1 and remains inactive. Nrf2-keap1 is also regulated by PI3K-Akt pathway. Under pathological conditions (neurological disorders) cause oxidative stress, Nrf2 and Keap1 will be separated. Nrf2 enters the nucleus and combines with MAF and ARE to activate antioxidant metabolic genes and play an antioxidant role. In addition, Nrf2 can activate GPX4 and be inhibited by Keap1. Both TCA cycle in mitochondria and Fenton reaction involving Fe²⁺ can cause oxidative stress by producing ROS. NAC, as a precursor of GPX4 and L-cysteine, affects oxidative stress through systemXc⁻ and GPX4. In addition, NAC can interfere with glutamate homeostasis and participate in the occurrence of a variety of nervous system diseases. NOX can produce ROS and cause oxidative stress under the action of a variety of regulatory factors. The bottom table is a summary of several molecules that affect oxidative stress in neurological disorders, including their action targets, action modes, and action results. Abbreviations: Akt: protein kinase B; ARE: antioxidant response element; ETC: electron transport chain; GPX4: glutathione peroxidase 4; Keap1: Kelch-like ECH-associated protein 1; Maf: musculoaponeurotic fibrosarcoma oncogene homolog; NAC: N-acetylcysteine; NOX: nicotinamide adenine dinucleotide phosphate oxidases; NADP: nicotinamide adenine dinucleotide phosphate; Nrf2: NF-E2-related factor 2; PI3K: phosphatidylinositol-3-kinase; ROS: reactive oxygen species; TCA: tricarboxylic acid.](image-url)
As an important regulator of oxidative stress, N-acetylcysteine (NAC) is a synthetic derivative of the endogenous amino acid L-cysteine and a precursor of GSH. GSH is a key factor in the clearance of ROS. Studies have shown that NAC mainly relies on GSH to exert its indirect antioxidant capacity, but this capacity is weak [158–160]. The direct antioxidant capacity of NAC depends on its nucleophilic carriers in multiple brain injury models [175].

In addition, Toll-like receptors (TLRs) are also closely related to the inflammatory response [166, 167], and NAC administration can reduce the level of ROS by increasing GSH [168], and oral NAC treatment within 24 hours after injury can reduce the level of ROS by increasing GSH [168], and oral NAC treatment within 24 hours after injury can reduce the level of ROS by increasing GSH [168], and oral NAC treatment within 24 hours after injury can reduce the level of ROS by increasing GSH [168].

As a precursor of GSH, NAC plays a beneficial role in the pathology and sequelae of TBI. Previous studies have shown that the administration time of NAC affects the therapeutic effect of NAC on TBI. Previous studies have shown that the administration time of NAC affects the therapeutic effect of NAC on TBI. In the 3-day TBI model, NAC administration 15–60 minutes after the injury can reduce the inflammatory response [166, 167], and NAC administration 12 hours after controlled cortical impact (CCI)-induced injury can reduce the level of ROS by increasing GSH [168], and oral NAC treatment within 24 hours after injury can alleviate the sequelae of TBI, such as confusion, headache, and abnormal sleep [169]. Recent studies have also found that the use of antioxidant dual therapy targeting antioxidant stress has a significant effect on the symptoms of TBI and improves the functional outcome [170]. For example, the use of NAC and sulforaphane (SFN) dual therapy can reduce TNF, IL, and other neuroinflammatory markers. In addition, the dual therapy of NAC + SPF has also been proved to be effective in alleviating epilepsy after SE [171]. In recent studies, c57bl/6 (wild type, WT) mice and CCL5 knockout (CCL5-KO) mice were used to induce mild brain injury and establish a weight drop model [172], which is a novel TBI model that can simulate the full spectrum of human TBI, mainly focusing on simulating diffuse brain injury. In the weight drop model, NAC can significantly alleviate the symptoms of decreased memory and learning ability after trauma and increase the level of Gpx1 in the hippocampus of mice [173]. With the application of nanomaterials in biomedicine, it is found that embedding NAC eluted poly (d,l-lactide-co-glycolide) (PLGA) nanofibers into scaffolds can effectively improve the shelf life of drugs and reduce systemic side effects. At the same time, it can give full play to the neuroprotective effect of NAC and the cell proliferation of nanosystems [174], which provides a broad prospect for the application of nanosystems in nerve repair. For example, neutral hydroxyl-terminated polyamide (PAMAM) dendrimers have shown great potential as nanocarriers in multiple brain injury models [175].

5. Conclusion

In this review, we summarize the relevant mechanisms of ferroptosis and some regulators of targeted oxidative stress in the treatment of neurological disorders. In the process of searching the literature, we find that oxidative stress and neurological disorders are closely related to the role of mitochondria (Figure 2), but the connection is very complex and lacks a systematic and clear mechanism. Ferroptosis may be a key mechanism to study the connection (Figure 1). For future research on neurological disorders, the role of mitochondria and the mechanism of ferroptosis will be the focus and hotspot of research. In addition, in view of the harmfulness of neurological disorders and the complexity of oxidative stress and ferroptosis, future research on the molecular mediators targeting oxidative stress and ferroptosis should pay more attention to the underlying mechanisms, so as to provide a theoretical basis for the treatment of neurological disorders.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Authors’ Contributions

Jing Li, Bowen Jia, and Ying Cheng contributed equally to this work.

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References

[1] S. J. S. J. Dixon, K. M. Lemberg, M. R. Lamprecht et al., “Ferroptosis: An Iron-Dependent Form of Nonapoptotic Cell Death,” Cell, vol. 149, no. 5, pp. 1060–1072, 2012.
[2] X. Chen, J. Li, R. Kang, D. J. Klionsky, and D. Tang, “Ferrop-tosis: machinery and regulation,” Autophagy, vol. 17, no. 9, pp. 2054–2081, 2021.

[3] Y. Yu, Y. Yan, F. Niu et al., “Ferrop-tosis: a cell death connecting oxidative stress, inflammation and cardiovascular diseases,” Cell Death Discovery, vol. 7, no. 1, p. 193, 2021.

[4] S. Xu, Y. He, L. Lin, P. Chen, M. Chen, and S. Zhang, “The emerging role of ferrop-tosis in intestinal disease,” Cell Death & Disease, vol. 12, no. 4, 2021.

[5] J. Li, F. Cao, H. L. Yin et al., “Ferrop-tosis: past, present and future,” Cell Death & Disease, vol. 11, no. 2, 2020.

[6] J. Ren, C. Li, X. L. Yan, Y. Qu, Y. Yang, and Z. N. Guo, “Cross-talk between oxidative stress and ferrop-tosis/oxytosis in ischemic stroke: possible targets and molecular mechanisms,” Oxidative Medicine and Cellular Longevity, vol. 2021, Article ID 6643382, 13 pages, 2021.

[7] V. L. Feigin, T. Vos, E. Nichols et al., “The global burden of neurological disorders: translating evidence into policy,” The Lancet Neurology, vol. 19, no. 3, pp. 255–265, 2020.

[8] A. S. Winkler, “The growing burden of neurological disorders in low-income and middle-income countries: priorities for policy making,” Lancet Neurology, vol. 19, no. 3, pp. 200–202, 2020.

[9] A. Schmitt, W. Xu, P. Bucher et al., “Dimethyl fumarate induces ferrop-tosis and impairs NF-κB/STAT3 signaling in DLBCL,” Blood, vol. 138, no. 10, pp. 871–884, 2021.

[10] Y. Li, X. Wang, Z. Huang et al., “CISD3 inhibition drives cystine-deprivation induced ferrop-tosis,” Cell Death & Disease, vol. 12, no. 9, p. 839, 2021.

[11] G. Lei, L. Zhuang, and B. Gan, “mTORC1 and ferrop-tosis: regulatory mechanisms and therapeutic potential,” BioEssays, vol. 43, no. 8, p. 2100093, 2021.

[12] P. Koppula, L. Zhuang, and B. Gan, “Cystine transporter SLC7A11/xCT in cancer: ferrop-tosis, nutrient dependency, and cancer therapy,” Protein & Cell, vol. 12, no. 8, pp. 599–620, 2021.

[13] W. Qu, Y. Cheng, W. Peng et al., “Targeting iNOS alleviates early brain injury after experimental subarachnoid hemor-rage via promoting ferrop-tosis of M1 microglia and reducing neuroinflammation,” Molecular Neurobiology, vol. 59, no. 5, pp. 3124–3139, 2022.

[14] Y. Zhao, Y. Li, R. Zhang, F. Wang, T. Wang, and Y. Jiao, “The role of erastin in ferrop-tosis and its prospects in cancer therapy,” OncoTargets and therapy, vol. Volume 13, pp. 5429–5441, 2020.

[15] H. Zhang, W. Jiao, H. Cui, Q. Sun, and H. Fan, “Combined exposure of alumina nanoparticles and chronic stress exacerbates hippocampal neuronal ferrop-tosis via activating IFN-γ/ ASK1/JNK signaling pathway in rats,” Journal of Hazardous Materials, vol. 411, p. 125179, 2021.

[16] Y. Wang, Y. Zhao, T. Ye, L. Yang, Y. Shen, and H. Li, “Ferrop-tosis signaling and regulators in atherosclerosis,” Frontiers in cell and developmental biology, vol. 9, pp. 809457–809457, 2021.

[17] J. Liu, R. Kang, and D. Tang, “Signaling pathways and defense mechanisms of ferrop-tosis,” The FEBS Journal, 2021.

[18] S. Xu, B. Wu, B. Zhong et al., “Naringenin alleviates myocardial ischemia/reperfusion injury by regulating the nuclear factor-erythroid factor 2-related factor 2 (Nrf2) /system xc-/ glutathione peroxidase 4 (GPX4) axis to inhibit ferrop-tosis,” Bioengineered, vol. 12, no. 2, pp. 10924–10934, 2021.

[19] H. J. Forman and H. Zhang, “Targeting oxidative stress in disease: promise and limitations of antioxidant therapy,” Nature Reviews. Drug Discovery, vol. 20, no. 9, pp. 689–709, 2021.

[20] A. Singh, R. Kukreti, L. Saso, and S. Kukreti, “Oxidative stress: a key modulator in neurodegenerative diseases,” Molecules, vol. 24, no. 8, p. 1583, 2019.

[21] L. Wang, Y. Liu, T. du et al., “ATF3 promotes erastin-induced ferrop-tosis by suppressing system Xc−,” Cell Death and Differentiation, vol. 27, no. 2, pp. 662–675, 2020.

[22] M. Maiorino, M. Conrad, and F. Ursini, “GPx4, lipid peroxi-dation, and cell death: discoveries,rediscoveries, and open issues,” Antioxidants & Redox Signaling, vol. 29, no. 1, pp. 61–74, 2018.

[23] S. Yang and G. Lian, “ROS and diseases: role in metabolism and energy supply,” Molecular and Cellular Biochemistry, vol. 467, no. 1-2, pp. 1–12, 2020.

[24] R. Tripathi, R. Gupta, M. Sahu et al., “Free radical biology in neurological manifestations: mechanisms to therapeutics interventions,” Environmental Science and Pollution Research International, 2021.

[25] S. Ni, Y. Yuan, Y. Kuang, and X. Li, “Iron metabolism and immune regulation,” Frontiers in Immunology, vol. 13, 2022.

[26] S. Wang, J. Luo, Z. Zhang et al., “Iron and magnetic: new research direction of the ferrop-tosis-based cancer therapy,” American Journal of Cancer Research, vol. 8, no. 10, pp. 1933–1946, 2018.

[27] Z. Geng, Z. Guo, R. Guo, R. Ye, W. Zhu, and B. Yan, “Ferrop-tosis and traumatic brain injury,” Brain Research Bulletin, vol. 172, pp. 212–219, 2021.

[28] D. Haschka, A. Hoffmann, and G. Weiss, “Iron in immune cell function and host defense,” Seminars in Cell & Developmental Biology, vol. 115, pp. 27–36, 2021.

[29] T. Tsuneda, “Fenton reaction mechanism generating no OH radicals in Nafion membrane decomposition,” Scientific Reports, vol. 10, no. 1, p. 18144, 2020.

[30] H. J. H. Fenton, “LXXIII.—Oxidation of tartaric acid in presence of iron,” Journal of the Chemical Society, vol. 65, pp. 899–910, 1894.

[31] H. Wang, C. Liu, Y. Zhao, and G. Gao, “Mitochondria regu-lation in ferrop-tosis,” European Journal of Cell Biology, vol. 99, no. 1, p. 151058, 2020.

[32] J. R. Friedman and J. Nunnari, “Mitochondrial form and function,” Nature, vol. 505, no. 7483, pp. 335–343, 2014.

[33] B. S. Connolly and A. E. Lang, “Pharmacological treatment of Parkinson disease,” JAMA, vol. 311, no. 16, pp. 1670–1683, 2014.

[34] C. Moreau, J. A. Duce, O. Rascol et al., “Iron as a therapeutic target for Parkinson’s disease,” Movement Disorders, vol. 33, no. 4, pp. 568–574, 2018.

[35] J. E. Chipuk, J. N. Mohammed, J. D. Gelles, and Y. Chen, “Mechanistic connections between mitochondrial biology and regulated cell death,” Developmental Cell, vol. 56, no. 9, pp. 1221–1233, 2021.

[36] C. Xu, S. Sun, T. Johnson et al., “The glutathione peroxidase Gpx4 prevents lipid peroxidation and ferrop-tosis to sustain Treg cell activation and suppression of antitumor immunity,” Cell Reports, vol. 35, no. 11, p. 109235, 2021.

[37] M. Yang, M. G. Tsui, J. K. W. Tsang et al., “Involvement of FSP1-CoQ10-NADH and GSH-GPx-4 pathways in retinal pigment epithelium ferrop-tosis,” Cell Death & Disease, vol. 13, no. 5, p. 468, 2022.
[38] S. Doll, F. P. Freitas, R. Shah et al., “FSPI1 is a glutathione-independent ferroptosis suppressor,” Nature (London), vol. 575, no. 7874, pp. 693–698, 2019.

[39] X. Sun, Z. Ou, R. Chen et al., “Activation of the p62-Keap1-NRF2 pathway protects against ferroptosis in hepatocellular carcinoma cells,” Hepatology, vol. 63, no. 1, pp. 173–184, 2016.

[40] Q. Tan, Y. Fang, and Q. Gu, “Mechanisms of modulation of ferroptosis and its role in central nervous system diseases,” Frontiers in Pharmacology, vol. 12, 2021.

[41] T. Rui, H. Wang, Q. Li et al., “Deletion of ferritin H in neurons counteracts the protective effect of melatonin against traumatic brain injury-induced ferroptosis,” Journal of Pineal Research, vol. 70, no. 2, article e12704, pp. 212–219, 2021.

[42] Y. Tian, J. Lu, X. Hao et al., “Ferroptosis is a mechanism in Parkinson’s disease,” Cell, vol. 142, no. 6, pp. 857–867, 2010.

[43] K. Jellinger, W. Paulus, I. Grundke-Iqbal, P. Riederer, and M. B. H. Youdim, “Brain iron and ferritin in Parkinson’s and Alzheimer’s diseases,” Journal of Neurotransmission. Parkinson’s Disease and Dementia Section, vol. 2, no. 4, pp. 327–340, 1990.

[44] M. A. Smith, P. L. R. Harris, L. M. Sayre, and G. Perry, “Ferroptosis and its role in cell death caused by iron overload and its underlying mechanism in Parkinson’s disease,” Free Radical Biology & Medicine, vol. 152, pp. 227–234, 2020.

[52] S. J. Guiney, P. A. Adlard, A. I. Bush, D. I. Finkelstein, and S. Ayton, “Ferroptosis and cell death mechanisms in Parkinson’s disease,” Neurochemistry International, vol. 104, pp. 34–48, 2017.

[53] A. Weiland, Y. Wang, W. Wu et al., “Ferroptosis and its role in diverse brain diseases,” Molecular Neurobiology, vol. 56, no. 7, pp. 4880–4893, 2019.

[54] P. Zhang, L. Chen, Q. Zhao et al., “Ferroptosis was more initial in cell death caused by iron overload and its underlying mechanism in Parkinson’s disease,” Free Radical Biology & Medicine, vol. 152, pp. 227–234, 2020.
[70] T. I. Williams, B. C. Lynn, W. R. Markesbery, and M. A. Lovell, “Increased levels of 4-hydroxynonenal and acrolein, neurotoxic markers of lipid peroxidation, in the brain in Mild Cognitive Impairment and early Alzheimer’s disease,” Neurobiology of Aging, vol. 27, no. 8, pp. 1094–1099, 2006.

[71] D. A. Butterfield, H. F. Poon, D. St. Clair et al., “Redox proteomics identification of oxidatively modified hippocampal proteins in mild cognitive impairment: insights into the development of Alzheimer’s disease,” Neurobiology of Disease, vol. 22, no. 2, pp. 223–232, 2006.

[72] P. J. Derry, M. L. Hegde, G. R. Jackson et al., “Revisiting the intersection of amyloid, pathologically modified tau and iron in Alzheimer’s disease from a ferroptosis perspective,” Progress in Neurobiology, vol. 184, p. 101716, 2020.

[73] P. K. Mandal, S. Saharan, M. Tripathi, and G. Murari, “Brain Glutathione Levels - A Novel Biomarker for Mild Cognitive Impairment and Alzheimer’s Disease,” Biological Psychiatry, vol. 78, no. 10, pp. 702–710, 2015.

[74] A. Ashraf, J. Jeandriens, H. G. Parkes, and P. W. So, “Iron dyshomeostasis, lipid peroxidation and perturbed expression of cysteine/glutamate antiporter in Alzheimer’s disease: Evidence of ferroptosis,” Redox Biology, vol. 32, p. 101494, 2020.

[75] E. Habib, K. Linher-Melville, H. X. Lin, and G. Singh, “Expression of xCT and activity of system x₅ are regulated by Nrf2 in human breast cancer cells in response to oxidative stress,” Redox Biology, vol. 5, pp. 33–42, 2015.

[76] F. Correa, E. Ljunggren, C. Mallard, M. Nilsson, S. G. Weber, and M. Sandberg, “The Nrf2-inducible antioxidant defense in astrocytes can be both up- and down-regulated by activated microglia: involvement of p38 MAPK,” Glia, vol. 59, no. 5, pp. 785–799, 2011.

[77] W. S. Hambright, R. S. Fonseca, L. Chen, R. Na, and Q. Ran, “Ablation of ferroptosis regulator glutathione peroxidase 4 in forebrain neurons promotes cognitive impairment and neurodegeneration,” Redox Biology, vol. 12, pp. 8–17, 2017.

[78] J. N. Stankowski, V. L. Dawson, and T. M. Dawson, “Ironing out tau’s role in parkinsonism,” Nature Medicine, vol. 18, no. 2, pp. 197–198, 2012.

[79] P. Lei, S. Ayton, D. I. Finkelstein et al., “Tau deficiency induces parkinsonism with dementia by impairing APP-mediated iron export,” Nature Medicine, vol. 18, no. 2, pp. 291–295, 2012.

[80] W. D. Bao, P. Pang, X. T. Zhou et al., “Loss of ferroportin induces memory impairment by promoting ferroptosis in Alzheimer’s disease,” Cell Death and Differentiation, vol. 28, no. 5, pp. 1548–1562, 2021.

[81] M. W. Park, H. W. Cha, J. Kim et al., “NOX4 promotes ferroptosis of astrocytes by oxidative stress-induced lipid peroxidation via the impairment of mitochondrial metabolism in Alzheimer’s disease,” Redox Biology, vol. 41, p. 101947, 2021.

[82] R. Bhatia, S. S. Chakrabarti, U. Kaur, G. Parashar, A. Banerjee, and R. K. Rawal, “Multi-target directed ligands (MTDLs): promising coumarin hybrids for Alzheimer’s disease,” Current Alzheimer Research, vol. 18, no. 10, pp. 802–830, 2021.

[83] A. Ashraf and P. W. So, “Spotlight on ferroptosis: iron-dependent cell death in Alzheimer’s disease,” Frontiers in Aging Neurosciences, vol. 12, p. 196, 2020.

[84] C. A. Ross and S. J. Tabrizi, “Huntington’s disease: from molecular pathogenesis to clinical treatment,” Lancet Neurology, vol. 10, no. 1, pp. 83–98, 2011.

[85] Y. Mi, X. Gao, H. Xu, Y. Cui, Y. Zhang, and X. Gou, “The emerging roles of ferroptosis in Huntington’s disease,” Neuromolecular Medicine, vol. 21, no. 2, pp. 110–119, 2019.

[86] C. Pigeon, G. Ilyin, B. Courselaud et al., “A New Mouse Liver-specific Gene, Encoding a Protein Homologous to Human Antimicrobial Peptide Hepcidin, Is Overexpressed during Iron Overload,” The Journal of Biological Chemistry, vol. 276, no. 11, pp. 7811–7819, 2001.

[87] H. Zhang, R. Ostrowski, D. Jiang et al., “Hepcidin promoted ferroptosis through iron metabolism which is associated with DMT1 signaling activation in early brain injury following subarachnoid hemorrhage,” Oxidative Medicine and Cellular Longevity, vol. 2021, Article ID 9800794, 19 pages, 2021.

[88] N. Geng, B. J. Shi, S. L. Li et al., “Knockdown of ferroportin accelerates erastin-induced ferroptosis in neuroblastoma cells,” European Review for Medical and Pharmacological Sciences, vol. 22, no. 12, pp. 3826–3836, 2018.

[89] Z. M. Qian and Y. Ke, “Hepcidin and its therapeutic potential in neurodegenerative disorders,” Medicinal Research Reviews, vol. 40, no. 2, pp. 633–653, 2020.

[90] J. Chen, E. Marks, B. Lai et al., “Iron accumulates in Huntington’s disease neurons: protection by deferoxamine,” PLoS One, vol. 8, no. 10, article e77023, 2013.

[91] B. D. Paul, J. I. Sbodio, R. Xu et al., “Cystathionine γ-lyase deficiency mediates neurodegeneration in Huntington’s disease,” Nature, vol. 509, no. 7498, pp. 96–100, 2014.

[92] N. Klepac, M. Relja, R. Klepac, S. Hećimović, T. Babić, and V. T. Kukulja, “Oxidative stress parameters in plasma of Huntington’s disease patients, asymptomatic Huntington’s disease gene carriers and healthy subjects: a cross-sectional study,” Journal of Neurology, vol. 254, no. 12, pp. 1676–1683, 2007.

[93] B. R. Cardoso, D. J. Hare, A. I. Bush, and B. R. Roberts, “Glutathione peroxidase 4: a new player in neurodegeneration?,” Molecular Psychiatry, vol. 22, no. 3, pp. 328–335, 2017.

[94] R. Skouta, S. J. Dixon, J. Wang et al., “Ferrostatins inhibit oxidative lipid damage and cell death in diverse disease models,” Journal of the American Chemical Society, vol. 136, no. 12, pp. 4551–4556, 2014.

[95] H. Jiao, H. Yang, Z. Yan et al., “Traditional Chinese formula Xiaoyaosan alleviates depressive-like behavior in CUMS mice by regulating PEBP1-GPX4-mediated ferroptosis in the hippocampus,” Neuropsychiatric Disease and Treatment, vol. -Volume 17, pp. 1001–1019, 2021.

[96] W. L. Chen, B. Xie, C. Zhang et al., “Antidepressant-like and anxiolytic-like effects of hydrogen sulfide in behavioral models of depression and anxiety,” Behavioural Pharmacology, vol. 24, no. 7, pp. 590–597, 2013.

[97] Y. Wang, S. Wang, Y. Xin et al., “Hydrogen sulfide alleviates the anxiety-like and depressive-like behaviors of type 1 diabetic mice via inhibiting inflammation and ferroptosis,” Life Sciences, vol. 278, p. 119551, 2021.

[98] C. N. Black, M. Bot, P. G. Scheffer, P. Cuijpers, and B. W. J. H. Penninx, “Is depression associated with increased oxidative stress? A systematic review and meta-analysis,” Psychoneuroendocrinology, vol. 51, pp. 164–175, 2015.

[99] S. Bhatt, A. N. Nagappa, and C. R. Patil, “Role of oxidative stress in depression,” Drug Discovery Today, vol. 25, no. 7, pp. 1270–1276, 2020.
Oxidative Medicine and Cellular Longevity

[100] C. S. Sriram, A. Jangra, S. S. Gurjar, P. Mohan, and B. K. Bezbarauh, "Edaravone abrogates LPS-induced behavioral anomalies, neuroinflammation and PARP-1," *Physiology & Behavior*, vol. 154, pp. 135–144, 2016.

[101] R. Dang, M. Wang, X. Li et al., "Edaravone ameliorates depressive and anxiety-like behaviors via Sirt1/Nrf2/HO-1/Gpx4 pathway," *Journal of Neuroinflammation*, vol. 19, no. 1, p. 41, 2022.

[102] E. J. Benjamini, P. Muntnner, A. Alonso et al., "Heart disease and stroke statistics-2019 update: a report from the American Heart Association," *Circulation*, vol. 139, no. 10, pp. e56–e528, 2019.

[103] Y. Zhang, X. Lu, B. Tai, W. Li, and T. Li, "Ferroptosis and its multifaceted roles in cerebral stroke," *Frontiers in Cellular Neuroscience*, vol. 15, p. 615372, 2021.

[104] Y. Jin, Y. Zhuang, M. Liu, J. Che, and X. Dong, "Inhibiting ferroptosis: a novel approach for stroke therapeutics," *Drug Discovery Today*, vol. 26, no. 4, pp. 916–930, 2021.

[105] G. Chen, L. Li, and H. Tao, "Bioinformatics identification of ferroptosis-related biomarkers and therapeutic compounds in ischemic stroke," *Frontiers in Neurology*, vol. 12, p. 745240, 2021.

[106] S. S. Karuppagounder, L. Alin, Y. Chen et al., "N-acetylcysteine targets 5 lipopxygenase-derived, toxic lipids and can synergize with prostaglandin E2 to inhibit ferroptosis and improve outcomes following hemorrhagic stroke in mice," *Annals of Neurology*, vol. 84, no. 6, pp. 854–872, 2018.

[107] I. Olmez and H. Ozyurt, "Reactive oxygen species and ischemic cerebrovascular disease," *Neurochemistry International*, vol. 60, no. 2, pp. 208–212, 2012.

[108] X. Guan, X. Li, K. Yang, et al., "The neuroprotective effects of carvacrol on ischemia/reperfusion-induced hippocampal neuronal impairment by ferroptosis mitigation," *Life Sciences*, vol. 235, p. 116759, 2019.

[109] Y. Cui, Y. Zhang, X. Zhao et al., "ACSL4 exacerbates ischemic stroke by promoting ferroptosis-induced brain injury and neuroinflammation," *Brain, Behavior, and Immunity*, vol. 93, pp. 312–321, 2021.

[110] S. J. An, T. J. Kim, and B. W. Yoon, "Epidemiology, risk factors, and clinical features of intracerebral hemorrhage: an update," *Journal of Stroke*, vol. 19, no. 1, pp. 3–10, 2017.

[111] X. Y. Xiong, J. Wang, Z. M. Qian, and Q. W. Yang, "Iron and intracerebral hemorrhage: from mechanism to translation," *Translational Stroke Research*, vol. 5, no. 4, pp. 429–441, 2014.

[112] M. Okauchi, Y. Hua, R. F. Keep, L. B. Morgenstern, T. Schallert, and G. Xi, "Deferoxamine treatment for intracerebral hemorrhage in aged rats: therapeutic time window and optimal duration," *Stroke*, vol. 41, no. 2, pp. 375–382, 2010.

[113] M. Zille, S. S. Karuppagounder, Y. Chen et al., "Neuronal death after hemorrhagic stroke in vitro and in vivo shows features of ferroptosis and necroptosis," *Stroke*, vol. 48, no. 4, pp. 1033–1043, 2017.

[114] I. Alim, J. T. Caulfield, Y. Chen et al., "Selenium drives a transcriptional adaptive program to block ferroptosis and treat stroke," *Cell*, vol. 177, no. 5, pp. 1262–1279.e25, 2019.

[115] X. Hu, Y. Xu, H. Xu et al., "Progress in understanding ferroptosis and its targeting for therapeutic benefits in traumatic brain and spinal cord injuries," *Frontiers in Cell and Developmental Biology*, vol. 9, 2021.

[116] P. E. Vos and R. Diaz-Arrastia, *Traumatic brain injury*, John Wiley & Sons Inc., Chichester, West Sussex, UK; Hoboken, NJ, 2015.

[117] V. A. Tyurin, Y. Y. Tyurina, G. G. Borisenko et al., "Oxidative stress following traumatic brain injury in rats: quantitation of biomarkers and detection of free radical intermediates," *Journal of Neurochemistry*, vol. 75, no. 5, pp. 2178–2189, 2000.

[118] H. Bayir, V. E. Kagan, Y. Y. Tyurina et al., "Assessment of antioxidant reserves and oxidative stress in cerebrospinal fluid after severe traumatic brain injury in infants and children," *Pediatric Research*, vol. 51, no. 5, pp. 571–578, 2002.

[119] B. S. Xie, Y. Q. Wang, Y. Lin et al., "Inhibition of ferroptosis attenuates tissue damage and improves long-term outcomes after traumatic brain injury in mice," *CNS Neuroscience & Therapeutics*, vol. 25, no. 4, pp. 465–475, 2019.

[120] A. Wanet, A. Tacheny, T. Arnould, and P. Renard, "miR-212/132 expression and functions: within and beyond the neuronal compartment," *Nucleic Acids Research*, vol. 40, no. 11, pp. 4742–4753, 2012.

[121] M. Quiles Del Rey and J. D. Mancias, "NCOA4-mediated ferritonophagy: a potential link to neurodegeneration," *Frontiers in Neuroscience*, vol. 13, 2019.

[122] S. Sun, X. Han, X. Li et al., "MicroRNA-212-5p prevents dopaminergic neuron death by inhibiting SIRT2 in MPTP-induced mouse model of Parkinson’s disease," *Frontiers in Molecular Neuroscience*, vol. 11, 2018.

[123] X. Xiao, Y. Jiang, W. Liang et al., "miR-212-5p attenuates ferroptotic neuronal death after traumatic brain injury by targeting Ptg2s," *Molecular Brain*, vol. 12, no. 1, p. 78, 2019.

[124] Y. Cheng, W. Qu, J. Li et al., "Ferristatin II, an iron uptake inhibitor, exerts neuroprotection against traumatic brain injury via suppressing ferroptosis," *ACS Chemical Neuroscience*, vol. 13, no. 5, pp. 664–675, 2022.

[125] A. J. Hulbert, R. Pamplona, R. Buffenstein, and W. A. Butte, "Life and death: metabolic rate, membrane composition, and life span of animals," *Physiological Reviews*, vol. 87, no. 4, pp. 1175–1213, 2007.

[126] K. Jung and M. Kwak, "The Nrf2 system as a potential target for the development of indirect antioxidants," *Molecules*, vol. 15, no. 10, pp. 7266–7291, 2010.

[127] L. Liu, L. M. Locascio, and S. Doré, "Critical role of Nrf2 in experimental ischemic stroke," *Frontiers in Pharmacology*, vol. 10, 2019.

[128] L. Zhang and H. Wang, "Targeting the NF-E2-related factor 2 pathway: a novel strategy for traumatic brain injury," *Molecular Neurobiology*, vol. 55, no. 2, pp. 1773–1785, 2017.

[129] M. J. Calkins, D. A. Johnson, J. A. Townsend et al., "The Nrf2/ARE pathway as a potential therapeutic target in neurodegenerative disease," *Antioxidants & Redox Signaling*, vol. 11, no. 3, pp. 497–508, 2009.

[130] F. Y. Yang, Q. K. Guan, Y. H. Cui, Z. Q. Zhao, W. Rao, and Z. Xi, "NAD(P)H quinone oxidoreductase 1 (NQO1) genetic C609T polymorphism is associated with the risk of digestive tract cancer: a meta-analysis based on 21 case-control studies," *European Journal of Cancer Prevention*, vol. 21, no. 5, pp. 432–441, 2012.

[131] A. C. Wild, H. R. Moimina, and R. T. Mulcahy, "Regulation of γ-gluamylycysteine synthetase subunit gene expression by the transcription factor Nrf2," *Journal of Biological Chemistry*, vol. 274, no. 47, pp. 33627–33636, 1999.
Oxidative Medicine and Cellular Longevity

[132] H. R. Moinova and R. T. Mulcahy, “Up-regulation of the human γ-glutamylcysteine synthetase regulatory subunit gene involves binding of Nrf-2 to an electrophile responsive element,” Biochemical and Biophysical Research Communications, vol. 261, no. 3, pp. 661–668, 1999.

[133] D. D. Zhang and M. Hannink, “Distinct cysteine residues in Keap1 are required for Keap1-dependent ubiquitination of Nrf2 and for stabilization of Nrf2 by chemopreventive agents and oxidative stress,” Molecular and Cellular Biology, vol. 23, no. 22, pp. 8137–8151, 2003.

[134] M. McMahon, N. Thomas, K. Itoh, M. Yamamoto, and J. D. Hayes, “Redox-regulated Turnover of Nrf2 Is Determined by at Least Two Separate Protein Domains, the Redox-sensitive Neh2 Degron and the Redox-insensitive Neh6 Degron,” Journal of Biological Chemistry, vol. 279, no. 30, pp. 31556–31567, 2004.

[135] M. Kobayashi, L. Li, N. Iwamoto et al., “The antioxidant defense system Keap1-Nrf2 comprises a multiple sensing mechanism for responding to a wide range of chemical compounds,” Molecular and Cellular Biology, vol. 29, no. 2, pp. 493–502, 2009.

[136] M. Dodson, M. Redmann, N. S. Rajasekaran, V. Darley-Usmar, and J. Zhang. “KEAP1–NRF2 signalling and autophagy in protection against oxidative and reductive proteotoxicity,” Biochemical Journal, vol. 469, no. 3, pp. 347–355, 2015.

[137] T. Zhang, P. Wu, E. Budbazar et al., “Mitophagy reduces oxidative stress via Keap1 (Kelch-like epithorohydrin-associated protein 1)/Nrf2 (nuclear factor-E2-related factor 2)/PHB2 (prohibitin 2) pathway after subarachnoid hemorrhage in rats,” Stroke, vol. 50, no. 4, pp. 978–988, 2019.

[138] L. F. M. F. Cardozo, L. M. Pedruzzi, P. Stenvinkel et al., “Nutritional strategies to modulate inflammation and oxidative stress pathways via activation of the master antioxidant switch Nrf2,” Biochimie, vol. 95, no. 8, pp. 1525–1533, 2013.

[139] K. Itoh, N. Wakabayashi, Y. Katoh et al., “Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain,” Genes & Development, vol. 13, no. 1, pp. 76–86, 1999.

[140] N. Wakabayashi, K. Itoh, J. Wakabayashi et al., “Keap1 -null mutation leads to postnatal lethality due to constitutive Nrf2 activation,” Nature Genetics, vol. 35, no. 3, pp. 238–245, 2003.

[141] M. Yamamoto, T. W. Kensler, and H. Motohashi, “The KEAP1–NRF2 system: a thiol-based sensor–effecter apparatus for maintaining redox homeostasis,” Physiological Reviews, vol. 98, no. 3, pp. 1169–1203, 2018.

[142] B. R. Al-Mubarak, K. F. Bell, S. Chowdhry et al., “Non-canonical Keap1-independent activation of Nrf2 in astrocytes by mild oxidative stress,” Redox Biology, vol. 47, p. 102158, 2021.

[143] G. D. Wu, Z. H. Li, X. Li, T. Zheng, and D. K. Zhang, “microRNA-592 blockade inhibits oxidative stress induced Alzheimer’s disease astrocytes via the KIAA0319-mediated Keap1/Nrf2/ARE signaling pathway,” Experimental Neurology, vol. 324, p. 113128, 2020.

[144] P. He, S. Yan, X. Wen et al., “Eriodictyol alleviates lipopolysaccharide-triggered oxidative stress and synaptic dysfunctions in BV-2 microglial cells and mouse brain,” Journal of Cellular Biochemistry, vol. 120, no. 9, pp. 14756–14770, 2019.

[145] X. Ren, J. Yu, L. Guo, and H. Ma, “TRIM16 protects from OGD/R-induced oxidative stress in cultured hippocampal neurons by enhancing Nrf2/ARE antioxidant signaling via downregulation of Keap1,” Experimental Cell Research, vol. 391, no. 1, p. 111988, 2020.

[146] X. Wang, F. Wang, G. H. Mao et al., “NADPH is superior to NADH or edaravone in ameliorating metabolic disturbance and brain injury in ischemic stroke,” Acta Pharmacologica Sinica, vol. 43, no. 3, pp. 529–540, 2022.

[147] A. Bertoni, P. Giuliano, M. Galgani et al., “Early and Late Events Induced by PolyQ-expanded Proteins;,” The Journal of Biological Chemistry, vol. 286, no. 6, pp. 4727–4741, 2011.

[148] G. A. Dominah, R. McMinimy, S. Kallon, and G. F. Kwakye, “Acute exposure to chlorpyrifos caused NADPH oxidase mediated oxidative stress and neurotoxicity in a striatal cell model of Huntington’s disease,” Neuropathology (Park Forest South), vol. 60, pp. 54–69, 2017.

[149] G. F. Kwakye, J. A. Jiminez, M. G. Thomas et al., “Heterozygous huntingtin promotes cadmium neurotoxicity and neurodegeneration in striatal cells via altered metal transport and protein kinase C delta dependent oxidative stress and apoptosis signal- ing mechanisms,” Neurotoxicology, vol. 70, pp. 48–61, 2019.

[150] S. Schiavone, P. Tucci, L. Trabace, and M. G. Morgese, “Early castration administration prevents ketamine-induced psychotic-like behavioral dysfunctions, oxidative stress and IL-10 reduction in the cerebellum of adult mice,” Molecules, vol. 24, no. 21, p. 3993, 2019.

[151] S. K. Barodia, R. B. Creed, and M. S. Goldberg, “Parkin and PINK1 functions in oxidative stress and neurodegeneration,” Brain Research Bulletin, vol. 133, pp. 51–59, 2017.

[152] P. Maher, K. van Leyen, P. N. Dey, B. Honrath, A. Dolga, and A. Mether, “The role of Ca2+ in cell death caused by oxidative glutamate toxicity and ferroptosis,” Cell Calcium, vol. 70, pp. 47–55, 2018.

[153] D. Sun, X. Chen, G. Gu, J. Wang, and J. Zhang, “Potential roles of mitochondria-associated ER membranes (MAMs) in traumatic brain injury,” Cellular and Molecular Neurobiology, vol. 37, no. 8, pp. 1349–1357, 2017.

[154] T. Hayashi and T. P. Su, “Sigma-1 receptor chaperones at the ER-mitochondrion interface regulate Ca(2+) signaling and cell survival,” Cell, vol. 131, no. 3, pp. 596–610, 2007.

[155] Y. Kim and H. H. Jang, “Role of cytosolic 2-Cys Prx1 and Prx2 in redox signaling,” Antioxidants, vol. 8, no. 6, p. 169, 2019.

[156] L. Li, C. Acigolu, R. F. Heary, and S. Elkabes, “Role of astroglial toll-like receptors (TLRs) in central nervous system infections, injury and neurodegenerative diseases,” Brain, Behavior, and Immunity, vol. 91, pp. 740–755, 2021.

[157] N. D. Anfinojenova, M. T. Quinn, J. A. Schepetkin, and D. N. Atochin, “Alarmins and c-Jun N-terminal kinase (JNK) signaling in Neuroinflammation,” Cell, vol. 9, no. 11, p. 2350, 2020.

[158] Y. Samuni, S. Goldstein, O. M. Dean, and M. Berk, “The chemistry and biological activities of N-acetylcysteine,” Biochimica et Biophysica Acta (BBA) – General Subjects, vol. 1830, no. 8, pp. 4117–4129, 2013.

[159] G. F. Rushworth and I. L. Megson, “Existing and potential therapeutic uses for N-acetylcysteine: the need for conversion to intracellular glutathione for antioxidant benefits,” Pharmacology & Therapeutics, vol. 141, no. 2, pp. 150–159, 2014.
[160] G. Aldini, A. Altomare, G. Baron et al., “N-Acetylcysteine as an antioxidant and disulphide breaking agent: the reasons why,” Free Radical Research, vol. 52, no. 7, pp. 751–762, 2018.

[161] G. Tardiolo, P. Bramanti, and E. Mazzon, “Overview on the effects of N-acetylcysteine in neurodegenerative diseases,” Molecules, vol. 23, no. 12, p. 3305, 2018.

[162] G. J. Zipfel, D. J. Babcock, J. M. Lee, and D. W. Choi, “Neuronal apoptosis after CNS injury: the roles of glutamate and calcium,” Journal of Neurotrauma, vol. 17, no. 10, pp. 857–869, 2000.

[163] D. A. Baker, K. McFarland, R. W. Lake et al., “Neuroadaptations in cystine-glutamate exchange underlie cocaine relapse,” Nature Neuroscience, vol. 6, no. 7, pp. 743–749, 2003.

[164] R. J. Bridges, N. R. Natale, and S. A. Patel, “System xc-cystine/glutamate antiporter: an update on molecular pharmacology and roles within the CNS,” British Journal of Pharmacology, vol. 165, no. 1, pp. 20–34, 2012.

[165] S. Spencer and P. W. Kalivas, “Glutamate transport: a new bench to bedside mechanism for treating drug abuse,” The International Journal of Neuropsychopharmacology, vol. 20, no. 10, pp. 797–812, 2017.

[166] G. Chen, J. Shi, Z. Hu, and C. Hang, “Inhibitory effect on cerebral inflammatory response following traumatic brain injury in rats: a potential neuroprotective mechanism of N-acetylcysteine,” Mediators of Inflammation, vol. 2008, Article ID 716458, 8 pages, 2008.

[167] N. Senol, M. Naziroglu, and V. Yuruker, “N-acetylcysteine and selenium modulate oxidative stress, antioxidant vitamin and cytokine values in traumatic brain injury-induced rats,” Neurochemical Research, vol. 39, no. 4, pp. 685–692, 2014.

[168] Y. Xiong, P. L. Peterson, and C. P. Lee, “Effect of N-acetylcysteine on mitochondrial function following traumatic brain injury in rats,” Journal of Neurotrauma, vol. 16, no. 11, pp. 1067–1082, 1999.

[169] M. E. Hoffer, C. Balaban, M. D. Slade, J. W. Tsao, and B. Hoffer, “Amelioration of acute sequelae of blast induced mild traumatic brain injury by N-acetyl cysteine: a double-blind, placebo controlled study,” PLoS One, vol. 8, no. 1, article e54163, 2013.

[170] J. Kyyriäinen, N. Kajevu, I. Bañuelos et al., “Targeting oxidative stress with antioxidant duotherapy after experimental traumatic brain injury,” International Journal of Molecular Sciences, vol. 22, no. 19, p. 10555, 2021.

[171] A. Pauli, G. Terrone, T. Shekh-Ahmad et al., “Targeting oxidative stress improves disease outcomes in a rat model of acquired epilepsy,” Brain, vol. 142, no. 7, pp. e39–e39, 2019.

[172] B. T. Kalish and M. J. Whalen, Weight drop models in traumatic brain injury, Springer New York, New York, NY, 2016.

[173] M. Ho, C. H. Yen, T. H. Hsieh et al., “CCL5 via GPX1 activation protects hippocampal memory function after mild traumatic brain injury,” Redox Biology, vol. 46, p. 102067, 2021.

[174] G. D. Mahumane, P. Kumar, V. Pillay, and Y. E. Choonara, “Repositioning N-acetylcysteine (NAC): NAC-loaded electrospun drug delivery scaffolding for potential neural tissue engineering application,” Pharmaceutics, vol. 12, no. 10, p. 934, 2020.

[175] A. Sharma, K. Liaw, R. Sharma, Z. Zhang, S. Kannan, and R. M. Kannan, “Targeting mitochondrial dysfunction and oxidative stress in activated microglia using dendrimer-based therapeutics,” Theranostics, vol. 8, no. 20, pp. 5529–5547, 2018.