Review

FOURIER TRANSFORM INFRARED SPECTROSCOPY IN MONITORING THE WINE PRODUCTION

ESPECTROSCOPÍA DE INFRAVERMELHO COM TRANSFORMADA DE FOURIER NA MONITORIZAÇÃO DA PRODUÇÃO DE VINHO

Vasiliki Thanasi1*, Sofia Catario1,2, Jorge Ricardo-da-Silva1

1LEAF (Linking Landscape Environment Agriculture and Food) Research Center, Instituto Superior de Agronomia, Universidade de Lisboa, Tapada da Ajuda, 1349-017 Lisboa, Portugal.
2CeFEMA (Centre of Physics and Engineering of Advanced Materials) Research Center, Instituto Superior Técnico, Universidade de Lisboa, Av. Rovisco Pais, 1, 1049-001 Lisboa, Portugal.

* Corresponding author: Tel.: + 306972097472; e-mail: vasilikithanasi@isa.ulisboa.pt

(Received date. 20.12.2021 Accepted date. 13.05.2022)

SUMMARY

The complexity of the wine matrix makes monitoring of the winemaking process from the grapes to the final product crucial for the wine industry. In this context, analytical methodologies that can combine good accuracy, robustness, high sample throughput, “green character”, and by preference real-time analysis, are on-demand to create high-quality vitivinicultural products. In the last years, Fourier-transform Infrared Spectroscopy (FTIR) combined with chemometric analysis has been evaluated in several studies as an effective analytical tool for the wine sector. Some applications of FTIR spectroscopy have been already accepted by the wine industry, mainly for the prediction of basic oenological parameters, using portable and non-portable instruments, but still many others are waiting to be thoroughly developed. This literature review aims to provide a critical synopsis of the most important studies assessing grape and wine quality and authenticity, and to identify possible gaps for further research, meeting the needs of the modern wine industry and the expectations of most demanding consumers. The FTIR studies were grouped according to the main sampling material used - 1) leaves, stems, and berries; 2) grape must and wine applications - along with a summary of the basic limitations and future perspectives of this analytical technique.

RESUMO

A complexidade da matriz do vinho torna a monitorização da sua produção, desde a maturação da uva até o produto final, fundamental para a indústria do vinho. Neste contexto, metodologias analíticas que possam combinar boa acurácia, robustez, elevado rendimento de amostras, menos penalizadoras para o meio ambiente, e possam ser capazes de fornecer resultados em tempo real, são muito importantes para a obtenção de produtos vitivinícolas de alta qualidade. Nos últimos anos, a Espectroscopia de Infravermelho com Transformada de Fourier (FTIR) combinada com a análise quimiométrica têm sido avaliada em diversos estudos por ser uma ferramenta analítica apropriada para o setor vitivinícola. Algumas aplicações de FTIR já foram adoptadas pela indústria do vinho, principalmente para a previsão de parâmetros enológicos básicos, através de instrumentos portátiles e não portátiles, mas há ainda um enorme potencial de desenvolvimento a explorar.

A presente revisão da literatura tem como objetivo fornecer uma sinopse crítica dos estudos mais importantes realizados para avaliação da qualidade e autenticidade do vinho, identificando possíveis lacunas para investigação futura, indo ao encontro das necessidades da indústria vitivinícola moderna e das expectativas dos consumidores mais exigentes. Os estudos sobre FTIR foram agrupados de acordo com o principal material de amostragem - 1) folhas, engaços e bagos; 2) mostos e vinhos - juntamente com informação sobre as limitações básicas e perspectivas futuras desta técnica analítica.

Keywords: FTIR spectroscopy, wine, winemaking monitoring, quality control, authenticity assessment.

Palavras-chave: Espectroscopia FTIR, vinho, monitorização da produção de vinho, controlo da qualidade, avaliação da autenticidade.

INTRODUCTION

Wine is considered the result of multiple biochemical and physicochemical reactions, which are responsible for its complex chemical composition. First of all, the grape composition at the harvest plays an important role in the future quality of the wine. Therefore, careful monitoring of specific grape quality parameters during ripening is essential for the wine industry. Monitoring of grape ripening by winemakers is mainly based on the measurement of total soluble solids (TSS), reducing sugars, pH, and titratable acidity. Generally, wine grapes with a high accumulation of sugars, phenolics (red grapes) and aromatic compounds,
intense colour (red grapes) and suitable acid content are the main goals. However, in some cases, these attributes are not sufficient to assess the wine quality. More assessments considering the maturation stage of grapes, the identification of the vineyard plots from which the grapes come, and the quality assessment of the grapes when they are delivered to the winery are needed (Dambers et al., 2015). Furthermore, considering that winemaking is based on several techniques and technologies to transform grapes into wine, there is a need for continuous quality control during these oenological procedures. The first important step of the quality control begins with the monitoring of the vinification (including alcoholic fermentation when glucose and fructose are converted mainly to ethanol and carbonic gas). It also includes other important transformations and chemical reactions due to the extraction and microbial metabolism of grape components. Moreover, malolactic fermentation (MLF), which can occur simultaneously with primary fermentation or sequentially, is another important step in the winemaking process for some wine styles. Furthermore, production techniques can be different according to the kind of grapes (white or red grapes) and the desired style of wine produced. For example, some fortified wines can be produced with the addition of wine spirit to stop alcoholic fermentation and enhance the final alcohol content. Finally, after the fermentation a series of important steps are also carried out, which include operations of clarification, stabilisation, maturation/ageing and packaging of the wine (Waterhouse et al., 2016).

Nowadays, several analytical techniques and methods used in the wine industry are considered to be non-environmentally friendly as they are using hazardous chemical solvents, or time-consuming, consisting of many analytical steps. In this context, FTIR technique is applied in the wine industry due to various advantages that allow to overcome previous analytical limitations, namely practicability, of the traditional techniques. Each spectroscopic technique has different and limited frequency ranges. The Infrared (IR) region is between 50-14000/cm and is divided into three areas: near-infrared (NIR) 4000-14000/cm; 2500-715 nm), mid-infrared (MIR) 400-4000/cm; 2500-2500 nm), and far-infrared (FIR) (50-400/cm; 200000-25000 nm) (Ferreira, 2015). FTIR is a non-destructive, time- and cost-effective technique that does not use hazardous chemical reagents, is characterised by high degree of automation, and short preparation time. Most applications in the wine industry are based on MIR and NIR spectroscopy. More specifically, FTIR instrumentation in combination with state-of-the-art software, designed specifically for grape and wine analysis, has recently received much attention. For most FTIR measurements, the MIR region (400-4000/cm) is investigated. FTIR can be used for qualitative and quantitative measurements, analysing at the same time several oenological parameters with a precision equal to the traditional techniques. Moreover, it is possible to combine portable and non-portable devices for monitoring the winemaking process in different locations (Ferreira, 2015).

Many applications based on IR spectroscopy of portable and non-portable devices are used to assess the maturation stage and to make decisions for the appropriate harvest time (Power et al., 2019, Felix instruments, 2021). Furthermore, IR spectroscopy, and specifically FTIR, is already applied in routine analysis for quality control of wines by a high number of laboratories both from wine companies and control entities, such as a Portuguese one, IVDP (Instituto dos Vinhos do Douro e do Porto) under the accreditation of NP EN ISO/IEC 17025. Furthermore, this technique has been considered in an inter-laboratory essay (physicochemical and instrumental analysis) for laboratory performance assessment, involving a significant number of participants (Alabe-Associação dos Laboratórios de Enologia, 2021). However, to improve the technique, several validations and optimisation procedures took place along with the development of new databases for analytical calibration aims (Moreira et al., 2002; Ferreira et al., 2009; Brandão, 2019). Moreover, OIV has established specific guidelines on infrared analysers in the oenology sector, concerning the exploitation of the characteristic absorptions of the organic compounds in the IR region of wines and musts for quantification purposes (OIV, 2010).

In this context, this review aims to give a brief introduction to the basic concepts of IR spectroscopy chemometric tools, and recent applications of FTIR spectroscopy to monitor each step of wine production. For the literature search, some of the most recent references describing applications and developments were used. Moreover, some oldest scientific sources of great interest were analysed. Published articles on the web of science, science direct, and other credible sources were used, along with some books.

**FTIR: BASIC CONCEPTS**

**Instrumentation**

The principle of the IR technique relies on recording absorption changes of IR radiation by molecules after vibrational and rotational modes due to absorption of energy. Each molecule contains a quantity of different functional groups (for example a carbonyl group or an amide group). Each functional group imparts a characteristic IR absorption at a specific frequency range. The
functional groups will vibrate when they will be exposed to IR radiation (Stuart, 2004).

Specifically, FTIR spectroscopy is based on the interference of radiation between two beams that results in the creation of an interferogram. The interferogram is a signal produced according to the change of path length between the two beams. This generated signal from the interferometer can then be reconverted into the two domains of distance and frequency that form a signal, by the mathematical method of Fourier transformation. A typical FTIR spectrometer includes a source, sample cell, detector, amplifier, analog-to-digital (A/D) converter, and computer. Radiation derived from the sources passes through the interferometer before reaching the detector. An A/D converter and amplifier convert the signal into a digital form before being transferred to the computer where the Fourier transform is taking place (Titus et al., 2019). The basic components of an FTIR spectrometer are shown in Figure 1.

![Figure 1. Schematic representation of FTIR equipment (adapted from Griffiths and De Haseth, 2007).](image)

**FTIR sampling methods**

The most commonly used sampling techniques for Vibrational Spectroscopy are as follows: 1) Transmission (TR) or Reflectance; 2) Attenuated Total Reflection (ATR); 3) Diffuse Reflectance (DRIFT).

In TR spectroscopy, the sample is placed directly into the (IR) beam. As the IR beam passes through the sample, the transmitted energy is measured and a spectrum is generated. The transmission technique can be used alone or in conjunction with accessories such as microscopes, and liquid or gas cells to analyse. Typically, in the case of liquid samples, fixed-length transmission cells are used. In the case of solids, the most common preparation is either the nujol mull (paraffin oil) or alkali halide (KBr) pellets (Mendes and Duarte, 2021).

In ATR spectroscopy, measurement is based on the changes that occur in an internally reflected IR beam when it contacts with a sample. An IR beam is directed onto a high refractive index (high optical density) crystal at a certain angle. Most often this material is either diamond, Zinc selenide (ZnSe), or Germanium (Ge). This internal reflectance creates an evanescent wave that extends beyond the crystal’s surface and penetrates the sample. In regions of the IR spectrum where the sample absorbs energy, the evanescent wave will be attenuated. The detector records the attenuated IR beam as an interferogram signal, which can then be used to generate an IR spectrum (Blum and John, 2012).

In DRIFT, the IR energy is directed into a sample cup filled with a mixture of the sample and an IR transparent matrix (such as KBr). The IR radiation interacts with the particles and then reflects off their surfaces, causing the light to diffuse, or scatter, as it moves throughout the sample. The output mirror then directs this scattered energy to the detector in the spectrometer. The detector records the altered IR beam as an interferogram signal, which can then be used to generate a spectrum (Olale et al., 2017).
Multivariate statistical techniques

The FTIR measurements produce multivariate responses and, as a result, generate a large amount of data. Chemometrics combined with FTIR is used for calibration, validation, and comparison in order to facilitate the analysis of such complex matrix interactions. Chemometrics is a discipline that uses mathematical and statistical methods to extract, represent, and display maximum chemical information from a sample. The chemometric analysis involves three main methods: Mathematical pre-treatments; Classification methods; Regression methods (Jamwal et al., 2021).

The Mathematical spectral pre-processing treatments are taking place in order to improve the acquired spectra, enhance the signal-to-noise ratio and reduce irrelevant spectra information. The most applied methods for this purpose can be divided into scatter correction methods and derivation methods. Scatter correction methods include Multiplicative Signal or Scatter Correction (MSC) (extended MSC, inverse MSC, inverse extended MSC and detrending), Standard Normal Variate Scaling (SNV), normalisation, and baseline correction. The derivation methods include finite difference, Savitzky–Golay, and Norris–Williams (Rinnan et al., 2009).

The multivariate classification methods can be unsupervised (exploratory data analysis) and supervised; the former is carried out with unknown data to identify the similarities and differences between samples by reducing the data dimensionality. The most widely used unsupervised methods are Principal Component Analysis (PCA) and Hierarchical Cluster Analysis (HCA). The supervised methods are mainly used for classification studies and the most widely known methods are Linear Discriminant Analysis (LDA), Soft Independent Modeling by Class Analogy (SIMCA), and Partial Least Squares Discriminant Analysis (PLS-DA) (Zhao and Liu, 2007; Jamwal et al., 2021).

Finally, for quantification purposes, Regression methods are used. The most widely used regression models are Multiple Linear Regression (MLR), Principal Component Regression (PCR), Partial Least Squares Regression (PLS-R), and Orthogonal Partial Least Squares Regression (OPLS). All above are linear methods applicable to first-order data and suitable for Parallel Factor Analysis (PARAFAC). For second-order data N-way, Partial Least Squares Regression (N-PLS) is considered more effective (Guillen-Casla et al., 2011; Mendes and Duarte, 2021). Non-linear methods can also be used, in which the variables show a non-linear relationship to the target characteristics (output variables). Artificial Neural Networks (ANN), Wavelet Neural Networks (WNN), and Support Vector Machines (SVM) are the most commonly used non-linear regression methods (Alexandidis and Zapranis, 2013; Moldes et al., 2017; Costa et al., 2019).

FTIR analytical calibration

The complexity and amount of the data obtained by FTIR make the development of analytical calibration procedures very important. The analytical methodologies based on FTIR measurements are not considered absolute methods of analysis, and therefore the obtained calibration models need to be validated in comparison with reference methods. The analytical calibration procedure consists of fitting the model to the data in order to obtain reliable estimations. Wine samples can differ extremely in the origin, vintage, cultivar, viticulture, winemaking procedures, and wine style, making important the selection of the calibration data to be considered representative (in terms of concentration ranges and sample matrix). Usually, to determine different chemical parameters, a selection process of specific spectral wavelengths is performed. Furthermore, the validation consists of examining the accuracy, precision, and prediction ability of the model for future samples deriving from the same population as the calibration samples. To estimate realistically the performance of the calibration model, an independent set of samples is required (Bauer et al., 2008; Ferreira, 2015).

FTIR APPLICATIONS IN WINE PRODUCTION

Analysis of grapes and leaves

Determination of basic oenological parameters

The first important step in the winemaking procedure, which is illustrated in Figures 2 and 3, is the collection of the grapes at the appropriate time, when they have reached the desired maturation stage according to the winemaking target. At this stage, the most usual evaluations, strongly correlated with the grape quality, are the determination of sugar content, acidity-related parameters, and phenolic content (red grapes). These parameters, which showed variations across the vineyard, making an easy, quick, and precise determination very important for the wine industry (Watson, 2003). In this context, some of the most interesting studies are summarised in Table I and analysed below.
In a chemometric exploratory analysis, Fourier Transformed Mid-infrared spectroscopy (FT-MIR) was applied to a grape must sample from South Africa (Swanepoel et al., 2007). The purpose of the study was the simultaneous quantification of total soluble solids (TSS, measured as °Brix), pH, and titratable acidity (TA, expressed as g tartaric acid/L) using PCA and PLS. The spectra region used for the calibrations was 1474-2685/cm. The results obtained were satisfactory for qualification analysis, but the analytical accuracies were not good enough to quantify these oenological parameters. This fact was ascribed to an insufficient fit of the South African grape samples to the global FT-MIR WineScan calibrations (Swanepoel et al., 2007). In a similar study, FT-MIR in combination with PCA analysis was used to monitor the evolution of red berries from véraison to harvest. MIR spectra provided more reliable discrimination between the berry samples from the different developmental stages than NIR spectra. ATR FT-MIR spectra from fresh homogenised berry samples were more discriminatory than with the frozen homogenised berry samples. The key to discrimination in between-stage variation was the sugar-organic acid absorption band, underlining the role of organic acid and sugars concentration as berry growth biomarkers. However, the discrimination of the last two stages of berry maturity faced more difficulties indicating the need for additional information deriving from the identification of other important compounds such as terpenoids (e.g., monoterpenes, sesquiterpenes, and carotenoids) or phenolics (e.g., flavonols, anthocyanins, and tannins) (Musingarabwi et al., 2016).
A classification study took place using ATR FT-IR and ANN. Specifically, spectra derived from grape skins and leaves were used for the classification of the samples according to their variety and ripeness degree. Pectin (associated with a peak at 825/cm) present in grapes influenced that classification for both variety and ripeness identification, while fructose content was associated with the ripeness degree. Polyphenols did not contribute to a great extent to the samples’ classification (Murru et al., 2019).

**Determination of phenolic compounds**

In order to enhance the spectra response, grape extracts were enriched with distinct phenolic compounds of different chemical structures (Fragoso et al., 2011). In this way, FT-MIR spectroscopy combined with PLS regression was able to quantify phenolic compounds for monitoring the ripening process in red grapes during the harvest period (Fragoso et al., 2011). Indeed, this method allowed evaluating total phenolic compounds, total anthocyanins, and condensed tannins simultaneously. Moreover, in this study, an attempt of developing models for individual varieties was quite promising. These results suggest the possible application of FT-MIR by each winery to design specific models for the varieties they are using. In each case, more samples are needed to cover a wider range of values (Fragoso et al., 2011).

The phenolic content of the wines depends on the extraction of phenolic compounds from the grapes. It is known that when the sugar content (degree of ripening) increases, the cell wall material decreases, affecting the phenolic compounds’ extractability (Ortega-Regules et al., 2008). For this reason, ATR-FTIR and Raman spectra of grape skin have been recorded and linked to the extractability of phenolic compounds (anthocyanins, flavanols, and total phenolics) of the inner grape skin surface. Concerning the ATR-FTIR data, the spectra peaks showed a significant influence of the number of polysaccharides and the degree of methyl esterification of pectins on the phenolic extractability levels of grape skin tissue, which were found mainly at 3324, 1732, 1018/cm. However, these correlations were only possible using the spectra of the inner skin surface of the grapes. The result was assigned to possible low amounts of phenolic compounds on the external skin surface (Nogales-Bueno et al., 2017).
In addition, due to an increasing interest in the full exploitation of the oenological by-products, determination of the quality parameters of grape seeds is important. Phenolic compounds are located in all the grape cluster: stems and berries (skins, seeds, pulp) (Sun et al., 1999). The extraction of phenolic compounds from the skins is faster for phenolic acids and anthocyanins, but for flavanols it requires more time. Moreover, when more extended maceration (red winemaking) is taking place, the process is more complex (Sun et al., 1999). In this case, the phenolic composition of the grape seeds is of great interest for the winemakers. The quantification of total phenolic and condensed tannins contents of grape seeds was possible with the application of a chemoometric analysis of FTIR spectra of grape seeds samples. The samples were lyophilised before analysis and as for the quantification of the phenolic content, the second derivative of the spectral region 1170-1560/cm was examined (Kyraleou et al., 2015). In the same context, seeds of different grapevine cultivars were analysed by FTIR and chemometrics to be discriminated according to their fatty acid and phenolic composition. For this purpose, lyophilised seed samples of three different Italian cultivars grown in an experimental vineyard (Lazio) were used. The reference values for the method.

### Table I

Representative examples of FTIR analysis of grapes, and leaves

| Aim of the analysis | Type of sample/Preparation mode | Wavenumber range (/cm) | Spectral pre-treatment | Statistical method | Reference |
|---------------------|--------------------------------|------------------------|------------------------|--------------------|-----------|
| Optimization of the quantification of total soluble solids, pH and titratable acidity | Fresh homogenates of berries | 964-1542, 1717-2969 | NM | PCA, PLS | Swaneepol et al. (2007) |
| Assessment of grape maturity | Fresh homogenates of berries | 926-5012 | 1st derivative (Savitzky-Golay) | PCA, PLS, PLS-DA | Piguë et al. (2010) |
| Estimation of pH of grape seeds procyanidins | Grape seeds powder | 700-1800 | SNV | PCT-PLS, O-PLS | Passos et al. (2010) |
| Quantification of phenolic acids | Fresh homogenates of berries | 1168-1457 | NM | PCA, PLS | Fragoso et al. (2011) |
| Determination of nitrogen | Centrifuged berries | 600-1800 | NM | PCA, PLS | Piguë et al. (2010) |
| Monitoring nitrogen and carbon content in grape vine | Wood and root grape tissue | 374-7496 | SNV, PL, Der, MC | PLS, SVR | Schmidike et al. (2012) |
| Quantification of Botrytis bunch rot | Fresh homogenates of berries | 3717-12500, 388-3984 | (Savitzky-Golay) | PLS, PCA | Hill et al. (2013) |
| Measurement of the concentration of nutrients in grapevine petioles | Dried grape leaves petioles | 375-7500 | SNV | PCA, PLS | Smith et al. (2014) |
| Analysis and discrimination of grape spoilage via volatiles | Whole berries | 600-4000 | Baseline correction, low pass filtering and smoothing | PCA, SIMCA | Dong et al. (2014) |
| Quantification of Tannin and G+G in Grape Seeds | Freeze dried and ground grape seeds | 400-1200 | 2nd derivative | PLS | Pappas et al. (2015) |
| Quantification of total phenolics and condensed tannins in grape seeds | Freeze dried and ground grape seeds | 400-1200 | 2nd derivative | PLS | Kyraleou et al. (2015) |
| Quantitative and qualitative evaluation of grape berries at various stages of maturity | Fresh and frozen homogenates of berries | 900-3800, 5000-12000 | NM | MVDA, PCA, OPLS-DA, PLS | Musingarabwi et al. (2016) |
| Determination of phenolic extractability | Whole berries, frozen berry skin layers | 600-4000 | Mean centre derivation | PCA | Nogales-Bueno et al. (2017) |
| Differentiation according to ripeness degree and grape variety | Berry skin layer, leaves | 600-4000 | NM | ANNs | Murr et al. (2019) |
| Discrimination of Aspergillus spp., Botrytis cinerea, and Penicillium expansum | Frozen homogenates of berries | 375-1850 | SNV, Mean centered data | PCA, SIMCA | Schmitz et al. (2019) |
| Discrimination of geographical origin and year of harvest, prediction of oenological parameters | Grape homogenates, grape skin | 400-4000 | Normalization, 1st derivative | PLS-DA, PLS | Lemos et al. (2020) |
| Determination of yeast assimilable nitrogen content | Grape juice | 4000-12500, 600-4000, 929-4000, 4000-5011 | 1st derivative, constant offset elimination | PLS | Petrovic et al. (2020) |
| Qualitative analysis of grape seeds | Freeze dried grape seeds | 500-4000 | NM | PCA | Lucarinia et al. (2020) |
| Characterisation of red grapes skin extracts | Hydrolysates of skin extracts | 4000-4000 | NM | PCA, LDA, PC, LDA | Alesca et al. (2020) |

**Notes:**
- NM=Not mentioned; PCA=Principal Component Analysis; PLS=Partial Least Squares Regression; PLS-DA=Partial Least Squares-Discriminant Analysis; PCT-PLS=Principal Components Transform-Partial Least Squares Regression; O-PLS=Orthogonal Partial Least Squares; SNV=Standard Normal Variate; SVR=Support Vector Regression; MSC=Multiple Scatter Correction; MC=Mean Centering; SIMCA=Soft Independent Modeling by Class Analog; MVDA=Multivariate Data Analysis; ANNs=Artificial Neural Networks; OPLS-DA=Orthogonal Projections to Latent Structures Discriminant Analysis; KNN=K-Nearest Neighbors; RFM=Recency, Frequency, and Monetary value; PC-LDA=Principal Components-Linear Discriminant Analysis; LDA=Linear Discriminant Analysis.
validation were obtained by conventional analysis HPLC/DAD/MS and GC/MS. The method allowed differentiating successfully the grape seeds from the different grapevine cultivars (Lucarini et al., 2020).

Another specific factor correlated in the long term with wine quality is the type of proanthocyanidins’ structures. Proanthocyanidins with a different mean degree of polymerisation (mDP; the average number of units in the polymer) and different degree of galloylation (% G; percentage of subunits containing gallic acid esters) correlate to different organoleptic properties (Vidal et al., 2003). The average degree of polymerisation of procyanidins (DPn) derived from red and white grape seeds was examined by FTIR and PLS regression. Before analysis, the samples underwent an extraction pre-treatment process and then a fractionation step was carried out using methanol/chloroform. Thiolysis was used as pre-treatment followed by HPLC-UV and MS detection to obtain reference values. The method allowed to correlate the increase of the absorbance peaks at 1203-1099/cm with an increase of DPn, and this was ascribed to the greater substitution in the aromatic ring of the polymerised procyanidin molecules (Passos et al., 2010). DRIFT spectra were combined with the PLS regression in a quantification study of tannin mean degree of polymerisation and percentage of galloylation in grape seeds. For this purpose, seed samples from two different vintages, two different Greek grapevine varieties (‘Maurotragano’ and ‘Xinomavro’) were used after freeze-drying. The developed models were compared with reference values from samples treated by phloroglucinolysis followed by HPLC-UV and LC-MS analyses. The second derivative of 1832-1416/cm and 918-739/cm spectral regions were used (values varied from 1.57 to 11.77) for quantification of mDP, while the second derivative of the spectral area 1813-607/cm (values ranges of %G from 2.98 to 15.85) was examined for %G quantification (Pappas et al., 2015).

Determination of other quality parameters

Usually at the reception of the grapes in the wineries, the basic grapes’ quality indicators (TSS, acidity-related parameters) are insufficient for a comprehensive quality control. In addition, phytosanitary status of the grapes influences their quality. Some types of fungi developing in grapes can lead to the formation of undesirable metabolites (e.g., 1-octen-3-ol and 1-octen-3-one), mycotoxins dangerous for human health, and wine oxidation through the production of laccase (Dewey et al., 2008). As a result, it is of great importance to identify infected grapes in the winery’s receiving line.

In this context, a differentiation study of four types of fungal mycelia in samples of ‘Chardonnay’ grapes using FTIR spectroscopy and a series of modelling approaches was performed. Support vector modelling was chosen as the best chemometric prediction modelling technique for pathogen and control samples (Schmidtke et al., 2019). However, the developed method needs to be applied in samples where the pathogens have occurred naturally and consider a larger sample set (Schmidtke et al., 2019). A study of Botrytis bunch rot in white grapes was focused on the suitability of FT-NIR and MIR spectroscopy combined with PLS regression for quantification purposes. Interesting results were attained, highlighting the importance of 7299-7937/cm region for the identification of important compounds produced by Botrytis cinerea. However, the method did not give sufficiently accurate results concerning the ranges that are relevant for quality control in the wineries. More wide sampling for validating similar methods deriving from different regions and varieties is required (Hill et al., 2013). Furthermore, it is known that specific volatile compounds, such as carbon dioxide, ethanol, and esters, are released from grapes during spoilage and their concentration can gradually change over storage time. A long optical path FTIR and sensor arrays (carbon dioxide and ethanol sensors) along with chemometric tools were investigated as possible ways of determining grape spoilage. The method was evaluated positively as a tool for the identification of different grape spoilage stages based on their volatile status. In addition, sensor arrays allowed to discriminate between healthy and decayed grapes as well (Dong et al., 2014).

Yeast assimilable nitrogen (YAN) content, including the nitrogen sources [free amino nitrogen (FAN) and ammonia] available in the grape juice matrix for the yeasts, is another crucial parameter strongly correlated with the healthy evolution of the fermentation process. Some amino acids (branched-chain and aromatic) are considered precursors of specific volatile compounds, influencing the quality of the final product Bell and Henschke, 2005). FT-MIR combined with PLS regression was applied to determine assimilable nitrogen in grape juice derived from red and white grapevine varieties. The method showed good accuracy and precision, and the results were confirmed using the formol titration as reference method, highlighting its feasibility for quantification purposes in the wineries (Skoutelas et al., 2011). Contextual information derived from the quantification of YAN content in grape juice was investigated using FT-IR, FT-NIR, and ATR-MIR and chemometrics. The best results were attained through FT-IR. The study considered a large and variable data set, and the developed models were evaluated by an independent data set. The developed method allowed predicting the YAN status of samples from a new vintage, highlighting the feasibility of this technology for industrial scale usage, providing quick and useful information for
nutrient supplementation decisions by the winemakers (Petrovic et al., 2020).

The nutritional status of the grapevine, correlated with mineral nutrient deficiency or toxicity, is of relevance for winemaking (Robinson, 1992). ATR-FTIR spectra with chemometric tools were applied in order to determine the nutrient status of petioles. The method was successful in determining the macronutrients in grapevine petiole tissue with good accuracy, but not adequate for certain micronutrients (Smith et al., 2014).

The nitrogen and starch contents in the perennial parts of the grapevine are correlated with the yield, the sugar accumulation, and the secondary metabolite production at the stage of berry ripening (Holzapfel et al., 2010). ATR-FTIR, and PLS or SV regression were applied to grapevine tissues to assess starch and nitrogen reserves. The results showed sufficient precision and accuracy, with support vector SV regression giving the best prediction values. The method could give useful information about the vine growth cycle and development (Schmidtke et al., 2012).

**Discrimination studies**

In addition, bearing in mind the influence of different spectral pre-treatments and sample preparation in many studies, ‘Tempranillo’ grape clones were analysed by ATR FT-MIR combined with multivariate analysis, resorting to different spectral pre-treatments (first derivative; normalised; normalised plus first derivative) to discriminate them in terms of geographical origin and year of harvest. At the same time, it was possible the prediction of the basic oenological parameters of interest, TSS, pH, and titratable acidity (Lemos et al., 2020). According to the authors, ATR FT-MIR combined with PLS-DA gave the most satisfactory results for the aforementioned discrimination. The better classification was obtained using the normalised spectra for the grape homogenates, and the normalised plus 1st derivative spectra for the skins (Lemos et al., 2020). Moreover, specific intervals were chosen to eliminate spectral interferences. Regarding the spectra regions, 750-1900/cm and 2650-3850/cm were selected for the homogenates, while those of 450-1900/cm and 2600-3700/cm were selected for the skins, corresponding to the fundamental vibrational modes of interest of the compounds under study (Lemos et al., 2020). In another discrimination study, based on a chemometric analysis (PC-LDA) of FTIR and Raman spectra, hydroalcoholic skin extracts from four different red grapevine varieties and two different types of vineyards (conventional and organic) were used. According to the chemometric analysis of the spectra, the method was able to discriminate between vineyards, but not the antioxidant activity levels and total phenolics content (Alecu et al., 2020).

**Analysis of grape must under alcoholic fermentation**

**Determination of basic oenological parameters**

Wine production is based on the biochemical process called alcoholic fermentation. This process includes the transformation of sugars (glucose and fructose) into ethanol and carbon dioxide, and usually is carried out by yeasts of the Saccharomyces genus. During winemaking, the parameters that are routinely measured daily to monitor the healthy evolution of the fermentation are sugars, total acidity, assimilable N, temperature, density, and pH (Ribereau-Gayon et al., 2021). Many studies have been focused on the determination of these parameters simultaneously using FTIR spectroscopy in combination with chemometrics, as shown in Table II.

FT-NIR coupled with chemometrics was applied for the determination of sugars, (FAN), malic acid, lactic acid, and ethyl carbamate content. The prediction values were very good for the sugars’ concentration but were not satisfactory for the remaining parameters. However, must samples with different FAN levels and wine samples with different organic acid and ethyl carbamate content could be classified when the discriminatory SIMCA method was applied to the spectra (Manley et al., 2001). FTIR with chemometric tools was used to quantify the volatile acidity, glycerol, ethanol, reducing sugars, and glucose content in fermented natural and model musts deriving from small-scale fermentations. The obtained results showed that the method can be considered effective in the simultaneous determination of these parameters. The prediction values for the volatile acidity showed the highest accuracy (Nieuwoudt et al., 2006). FTIR modelled with two-dimensional correlation techniques was proven to be an efficient tool for monitoring physicochemical changes occurring during red wine fermentation (chemical reactions and their dynamics). The method allowed to show that the conversion reaction rate of glucose to alcohol was more rapid than the one of fructose (Wynne et al., 2007). FT-NIR and FTIR spectroscopy was applied for more detailed monitoring of a maceration/fermentation classical red winemaking process. The results after different chemometric treatments were quite promising. The method made it possible to simultaneously determine sugars, ethanol, glycerol, and phenolic content, and the samples could be classified according to their stage of fermentation (Di Egidio et al., 2010). These results were confirmed by a similar study in which FTIR and FT-NIR spectroscopy were applied again to identify molecular changes involved in wine fermentation. In addition, to combine more information for taste and aroma profiles of the samples, electronic tongue and electronic nose were used. Furthermore, the
chemometric analysis of the spectral, electronic nose and electronic tongue data was able to underline specific critical points during the fermentation process, which is useful for quality control of the final product (Buratti et al., 2011).

| Aim of the analysis | Type of sample/Preparation mode | Wavenumber range (μm) | Spectral pre-treatment | Statistical method | Reference |
|---------------------|---------------------------------|-----------------------|------------------------|-------------------|-----------|
| Determination of analytical parameters | Muts | 4000-10000 | MSC | PLS, SIMCA | Manley et al. (2001) |
| Authentication of white grape musts | Muts | 4000-10000 | MSC | PLS-DA, FDA | Roussel et al. (2003) |
| Monitoring of wine fermentation | Frozen fermentation musts | 4552-5800 | 1st and 2nd derivative | MLR, PLS | Urtubia et al. (2004) |
| Determination of compositional dynamics of wine fermentation | Frozen musts | 900-4000 | NM | dimensional correlation | Wynne et al. (2007) |
| Monitoring of red fermentation | Centrifugated musts | 3600-12500 | SNV, MSC, 1st and 2nd derivative | PCA, LDA | Di Edigio et al., (2010) |
| Monitoring of alcoholic fermentation | Centrifugated musts | 3600-12500 | SNV, Golay derivative | PCA | Buratti et al. (2011) |
| Surface measurement of mannoproteins and β-glucans of yeast cell walls during wine fermentation | Powdered natural and synthetic musts | 650-4000 | baseline correction | PCA, OPLS-DA, ANOVA | Moore et al. (2015) |
| Monitoring Saccharomyces cerevisiae grape must fermentation | Centrifugated musts | 800-4000 | Mean centered, 2nd derivative | PCA, SIMCA | Puzea et al. (2015) |
| Determination of anthocyanins | Digested, blended, squeezed mus | 926-5012 | NM | PCA, PLS-R | Rasines-Perea et al. (2015) |
| Strain typing of yeasts | Muts | 400-4000 | NM | HCA | Gerhardt et al. (2015) |
| Study of inter-and intraspecific biodiversity of cultivable non-Saccharomyces yeasts | Muts | 400-4000 | NM | HCA | Grangenteau et al. (2016) |
| Monitoring of wine process and prediction of its parameters | Frozen musts and wines | 650-4000 | WOS, OSCW, 1st and 2nd derivative | PCA, PLS | Canal and Ozen. (2017) |
| Determination of calcium | Filtered musts | 4348-9091 | SNV, MSC, 1st derivative | PLSR | Véstia et al. (2019) |
| Early detection of undesirable deviations in must fermentation | Diluted musts | 649-3999 | Smoothing and normalization | PCA, PLSR, PLS-DA | Cavagli et al. (2019) |
| Monitoring wine fermentation deviations | Homogenized, centrifugated musts | 650-4000 | 1st and 2nd derivative | PCA, PLSR, PLS-DA, SPV | Cavagli et al. (2020a) |

Table II
Representative examples of FTIR analysis in musts

NM=Not mentioned; MSC=Multiple Scatter Correction; PLS=Partial Least Squares Regression; SIMCA=Soft Independent Modeling by Class Alogory; GA=Genetic Algorithms; PLS-DA=Partial Least Squares-Discriminant Analysis; FDA=Functional Data Analysis; MLR=Multiple Linear Regression; SNV=Standard Normal Variate; MSC=Multiple Scatter Correction; PCA=Principal Component Analysis; LDA=Linear Discriminant Analysis; ANOVA=Analysis of Variance; OPLS-DA=Orthogonal Projections to Latent Structures Discriminant Analysis; PLS-RL=Partial-Least-Squares Regression; HCA=Hierarchical Cluster Analysis; WCS=Wavelet Compression of Spectra; WOSC=Orthogonal Signal Correction; OSCW=Orthogonal Signal Correction in Combination with Wavelet; PLSR=Partial-Least-Squares Regression; MSPC=Multivariate Statistical Process Control.

However, when deviations occur implying sluggish or stuck fermentations or other unwanted contaminations by microorganisms, distinct time-consuming chemical analyses are required. As a result, there is a growing need for real-time information delivered by methods, such as vibrational spectroscopy, for the appropriate readjustments to take place in a timely manner before the process ends. IR spectroscopy was used as an analytical tool to distinguish between normal and problematic fermentation. The developed calibration models showed sufficient accuracy for the determination of glucose, fructose, glycerol, ethanol, and most of the organic acids (tartaric, succinic, lactic, acetic, and citric) during fermentation of ‘Cabernet Sauvignon’ musts. The only exception was malic acid, for which the developed calibration was not good enough to distinguish between normal and problematic behaviour. When the calibration models were applied to the other varieties under study, the results were not so adequate for any of the parameters (Urtubia et al., 2004). ATR-FTIR spectra derived from a portable device were used to discriminate between normal fermentation and problematic in terms of accumulative yeast nitrogen deficiencies. Several models have been developed to monitor the evolution of fermentation and, at the same time, discriminate between normal and problematic fermentations. The outcomes showed that the
method could be applied in the wine industry for monitoring the sugar content and identifying nutrient deficiencies early (Cavaglia et al., 2019). Spectra coming from a portable ATR-MIR device and multivariate analysis were used to control the alcoholic fermentation process and detect wine fermentation problems. Generally, the method succeeded to predict density and pH in fermenting must samples. The spectra recorded during fermentations with LAB (Lactobacillus)-inoculations (ICF) were performed showed only minor changes than in normal fermentation conditions (NFC). This fact was assigned to the small concentration changes involved in the malolactic fermentation process. In most samples, the deviations in L-malic acid were between 0.7 to 0.8 g/L concentrations resulting in a small pH increase. The novelty of this ATR FT-MIR method was the detection of no NFC before the end of malolactic fermentation (Cavaglia et al., 2020b). In a different chemometric approach, detection of lactic bacteria spoilage during fermentation took place with the use of a portable ATR FT-MIR instrument and Multivariate Statistical Process Control charts (MSPC). For this purpose, samples from alcoholic fermentation in normal conditions (NOC) and alcoholic fermentation with the addition of lactic bacteria (MLF). MSPC charts based on Q residuals and Hotelling’s T2 statistics proved to be efficient in detecting lactic bacteria spoilage before the end of the alcoholic fermentation (Cavaglia et al., 2020a). The analysis of the cell wall composition of yeasts is an indirect approach for monitoring the fermentation. ATR FT-MIR and chemometrics was used to evaluate the cell wall composition during fermentation. Different yeast strains were investigated, including Saccharomyces (laboratory and industrial) and non-Saccharomyces strains. PCA analysis was able to differentiate Saccharomyces strains from the non-Saccharomyces ones, and industrial wine yeast strains from laboratory ones. The fingerprint region (1768-770/cm) was associated with lipid, protein (including CHO and amide groups), mannans, nucleotide, phospholipid, and glucan functional groups. Mannoproteins were more abundant in industrial strains, whereas mannan and glucan polymers were more plentiful in laboratory strains (Moore et al., 2015). In the same context, ATR-FTIR coupled with SIMCA was applied in fermented ‘Grenache Blanc’ and ‘Chardonnay’ musts to determine the physiological state (exponential and stationary) of three commercial strains of Saccharomyces cerevisiae (ES454, E491, and ES181). The different physiological states were possible to be identified based on the different cell wall components of the strains. At the exponential phase mainly glucans, mannoproteins, and lipids were identified, while at the stationary phase glucans and mannans were observed (Puxeu et al., 2015).

**Determination of other quality parameters**

Moreover, during the fermentation process, other important quality parameters need to be determined. The prediction of calcium is of great importance for the winemakers, and especially for the sparkling wine industry, due to the risk of aggregation with alginate capsules when this technique is used. Usually, the calcium content is determined by atomic absorption spectrometry (AAS). In a different approach, FT-NIR was applied in combination with chemometrics for the prediction of calcium content in base wine samples and musts. AAS was used as a reference method, after pretreatment of the samples by dry ashing. High concentrations of calcium were observed.. FT-NIR–PLSR models could be used in real-time industrial monitoring for quantitative analysis of calcium in wines (Véstia et al., 2019). Another pivotal quality parameter is the anthocyanins content. FT-MIR in combination with PLS-R evaluated the prediction of 12 anthocyanins (five non-acetylated, three acetylated, three p-coumaroylated and one caffeoylated 3-O-glucosides) in red grapes musts. Internal and external validation sample sets were used to confirm the obtained results. Unfortunately, when must samples from a new harvest were used the results were not satisfactory. However, when the model was built from samples derived from all the different considered harvests, the prediction values were improved. This fact demonstrates the potential of the method for a quick semi-quantitative determination of anthocyanin content in the wineries but more studies are needed to improve the prediction values (Rasines-Perea et al., 2015).

The final quality of the product is known to be influenced by the ‘terroir’, including soil, climate, grapevine plant, and viticultural and oenological practices (OIV, 2010). However, there is a need for information about site-specific differences of vineyard yeasts and how they can influence fermentation. This topic was investigated by analysing spontaneous fermentations from six different vineyards. FTIR spectra were acquired from randomly isolated yeasts to discriminate at the strain level. The yeasts coming up from the vineyard were discriminated from those winery related. The latter showed the biggest influence on the spontaneous fermentation, without following a constant pattern (Gerhards et al., 2015). Similar results were attained by FT-IR spectra in an inter- and intra-biodiversity study of non-Saccharomyces yeasts. The pyrosequencing method was used as a reference to compare the obtained results. In terms of global diversity, the method identified efficiently seven different genera (Aureobasidium, Candida, Cryptococcus, Hanseniaspora, Issatchenkia, Metschnikowia and Pichia). Concerning the intraspecific analysis, 19
different strains from 58 isolates were identified. The FT-IR spectroscopy proved to be an effective tool for yeast strain characterisation and monitoring of yeast starter strains (Saccharomyces and/or non-Saccharomyces) during fermentation (Grangeteau et al., 2016).

**Authentication studies**

Bearing in mind a continuous need for authenticity confirmation of the products of the wine industry, an authentication study in terms of varietal classification of musts was made using FTIR, ultraviolet spectrophotometry, and analysis by electronic nose. Genetic Algorithms were used as a pre-processing technique to improve the interpretation of the data. FTIR spectroscopy matched with multivariate chemometric techniques proved to be superior in comparison to ultraviolet spectrophotometry, and analysis by electronic nose (Roussel et al., 2003).

FT-MIR spectra combined with multivariate analysis were used for the determination of chemical (ethanol, glycerol, organic acids, titratable acidity, °Brix, sugars, total phenolics, and anthocyanins content) and microbiological parameters of red, rose and white wines during the whole winemaking process. Regarding the pretreatment of the spectra, the second derivative seemed to be the most appropriate option for the determination of most of the chemical parameters, except for ethanol. The results were satisfactory for the prediction of almost all the chemical parameters (except pH and organic acids). On the other hand, the prediction of microbiological parameters was not accurate enough. In addition, the method was able to differentiate the beginning of the fermentation process from the remaining winemaking steps (Canal and Ozen, 2017).

**Analysis of wines and wine spirits**

**Determination of basic oenological parameters**

Monitoring of winemaking process is a dynamic task that examines continuously the wine quality after the fermentation, from the ageing stage to the bottling. The basic routine analysis includes determination of alcoholic strength, sulphur dioxide (SO$_2$), volatile acidity, and pH. Some representative examples of the literature are presented in Table III.

Optimisation of a FTIR method applied for general analysis of wine samples from red, white, rose, and sparkling Portuguese wines was performed (Moreira et al., 2002). Global calibrations were built for the simultaneous determination of various oenological parameters (density, dry extract, total acidity, volatile acidity, total SO$_2$); total sugars were determined through an independent calibration. The outcomes revealed good precision for most of the parameters in comparison with the reference methods, except for total SO$_2$. Comparing the accuracy of the global calibration and the specific calibration for the total sugars, in the second case the results were more satisfactory (Moreira et al., 2002). A series of changes in the oenological parameters of white wines packed in bag-in-box (BIB) were monitored by chemometric-based FTIR analysis. The developed calibration models, based on a PLS algorithm, successfully predicted the colour, free and total SO$_2$ contents, total phenolics and aldehydes contents, and the storage time in bag in box. However, a potential optimisation of the method’s accuracy requires the use of a larger range of samples (Fu et al., 2009). FT-MIR spectroscopy in combination with chemometrics was used for determination of several quality parameters (pH, total phenolic content, anthocyanin content, titratable acidity, sugar content, electrical conductivity, and some colour parameters) of alcoholic beverages made of twice-distilled grapes and anise (Raki) and wine. The developed PLS models for wines showed good prediction values for total phenolics, anthocyanin contents, pH, °Brix, and colour intensity. Concerning the Raki samples, total phenolics, sugar content, and pH, were predicted successfully (Ozturk et al., 2012). The ATR-FTIR wine spectra provided contextual information. Some common parameters of wine were predicted (alcoholic content, sugars, total acids), while the determination of others did not provide satisfactory results (SO$_2$ and volatile acidity) (He et al., 2013). In a similar study in grape-derived spirits, FTIR–ATR combined with chemometrics was applied for the determination of their alcoholic strength, methanol, acetaldehyde, and fusel alcohols content. Most of the parameters were predicted with good accuracy, except for 2-butanol (Anjos et al., 2016). In this context, one of the most recent studies evaluated the feasibility of FTIR combined with Network analysis in an Eigenspace layout for wine analysis. A network can be defined as a collection of nodes connected through links. Each sample is represented by each node in the network. Each edge (formed by the links) represents the difference between the two connected nodes (dissimilarity weight). The FTIR data sets derived from different wine samples, lead to the creation of a networking in the Eigenspace layout, giving specific chemical significance to the positioning nodes. The method allowed to identify compositional differences among the samples, classifying them into two groups, and assessing their inter- and intra-group homogeneity. The wines were separated into two groups based on the differences found in the spectra region 970-1100/cm, suggesting different levels of ethanol, phenolics, phosphates, phenyl derivatives, unsaturated lipids and saccharides (Kumar et al., 2021).
Table III

| Aim of the analysis | Type of sample/Preparation mode | Wavenumber range / (cm⁻¹) | Spectral pre-treatment | Statistical method | Reference |
|---------------------|---------------------------------|---------------------------|------------------------|-------------------|-----------|
| Optimisation of wine analysis | Degassed wines | 926-5012 | NM | NM | Moreira et al. (2002) |
| Analysis of wine poly saccharides | White wine poly saccharide extracts | 800-1200 | Autocorrelation (Mean centred, standardized) | PCA, CCA, PLS | Coimbra et al. (2002) |
| Quantitative analysis of red wine tannins | Wines purified by SPE | 650-4000 | Mean-centring, 1st and 2nd derivatives | PLS | Fernández and Agossin (2007) |
| Analysis of wine poly saccharides | Red wine poly saccharide extracts | 950-1850 | Linear standardization | PLS | Boulet et al. (2007) |
| Identification of Geographical regions for the quantification of red wine tannins | Wines | 926-5012 | NM | PLS | Jensen et al. (2008) |
| Discrimination of Greek red wines based on the grapevine variety | Phenolic extracts of red wines | 900-1800 | 2nd derivative (Savitsky-Golay) | Match value | Tarantilis et al. (2008) |
| Determination of oenological parameters | Wines | 600-4000 | Mean centre and baseline corrections | PLS | Fu et al. (2009) |
| Prediction of total antioxidant capacity of red wine | Wines | 600-4400 | Mean-centring | PLS | Versari et al. (2010) |
| Discrimination of oenological tannins | Solution of oenological tannin| 950-1500 | NM | NM | Laghi et al. (2010) |
| Authentication of Cypriot sweet wine | Freeze-dried and concentrated wines under nitrogen flow | 400-4000 | NM | PCA, CA, LDA, CART, MLA | Ioannou-Papayianni et al. (2011) |
| Determination of anthocyanin content | Degassed wines | 4000-12500 | 1st and 2nd derivatives, Detrend, smoothing, MSC, SNV, mean centring | PCA, PLS, DA, WILMA-D | Ferrari et al. (2011) |
| Direct determination of oenological parameters | Filtered wines and wine derived products | 1000-7895 | NM | PLS | Regmi et al. (2012) |
| Measuring of wine routine parameters | Filtered and thermostated wines | 926-5012 | NM | PCA, PLS | Romero-Fernández et al. (2012) |
| Discrimination between subzones inside a controlled designation of origin | Heated Wines | 4000-52632 | SNV, 2nd derivative (Savitsky-Golay) | PCA, LDA, SIMCA, SVM | Martelo-Vidal et al. (2013) |
| Prediction of total phenolics and flavonoid contents and antioxidant capacity | Wines | 650-4000 | NM | PLS | Silva et al. (2014) |
| Quality control of grape derived spirits | Grape derived spirits | 400-4000 | MSC, VecNor, MinMax, SLS, 1st and 2nd derivatives | PCA, PLS | Anjos et al. (2016) |
| Determination of chloride and sulphate | Filtered and degassed wines | 1000-3050 | NM | PCA, PLS | Teixeira dos Santos et al. (2016) |
| Analysis of ash in wines | NM | 3rd derivative analysis | | | |
| Characterisation of Marsala wines and ageing monitoring | Incubated wines | 350-4000 | Baseline correction | ANOVA, PCA, LDA | Conduru et al. (2018) |
| Romanian wines | Centrifugated wines | 400-4000 | 2nd derivative (Savitsky-Golay) | PCA | Topala and Tataru (2018) |
| Discrimination of sweet wines | NM | 400-4000 | 2nd derivative (Savitsky-Golay) | PCA | Topala and Tataru (2019) |
| Analysis of ellagitannins | Wines | 950-1820 | Baseline correction (Savitsky–Golay polynomial filter) | PLS | Basakelou et al. (2019) |

NM=Not mentioned; PCA=Principal Component Analysis; CCA=Canonical-Correlation Analysis; PLS=Partial Least Squares Regression; CA=Correspondence Analysis; LDA=Linear Discriminant Analysis; CART=Classification and Regression Tree Analysis; RDA=Redundancy Analysis; PLS-DA=Partial Least Squares-Discriminant Analysis; WILMA-D=A Novel Algorithm based on the Fast Wavelet Transform (FWT); MSC=Multiple Scatter Correction; SNV=Standard Normal Variate; SIMCA=Soft Independent Modeling by Class Analogy; SVM=Support Vector Machine; ANOVA=Analysis of Variance.
As shown above, most of the studies were focused on the simultaneous assessment of several wine parameters, but often the results are not enough satisfactory for the prediction of all of them. Due to spectral interferences, the analytical calibration of compounds that are found in low concentrations in the wine matrix, in comparison with other abundant components like ethanol and organic acids, is facing difficulties. Independent studies have been performed on the determination of organic acids to achieve better performances. For this reason, FTIR measurements took place in samples spiked with known organic acid concentrations (tartaric, malic, lactic, acetic, and citric acids). The obtained recoveries for the total acidity and the individual organic acids (for the same spiked additions) were compared. Nevertheless, validation procedures should not be forgotten and should be fully performed. The estimations for total acidity recovery were good, but for the individual organic acid concentrations the recovery results revealed a lack of accuracy. This fact was assigned to spectral similarities between the organic acids (Moreira and Santos, 2005). In another chemometric approach, FTIR combined with PLS regression was used to build calibration models for tartaric acid, malic acid, lactic acid, succinic acid, citric acid, and acetic acid in wines, vinegars, and spirits. HPLC was used as a reference method for the validation of the obtained results. Excellent performance was observed for the determination of tartaric acid, malic acid, succinic acid, and lactic acid at high concentrations, except for tartaric acid in vinegar samples. The method was also efficient for the assessment of malic and citric acid at low concentrations and acetic acid in red and white wine and brandy samples. This fact was considered to be correlated with the predicting performance of the reference method and the composition of the data set. Moreover, it was not possible to obtain a global calibration due to the different nature of the samples under study (Regmi et al., 2012). White wines obtained from different maceration and pectic enzyme clarification procedures were used to obtain polysaccharide extracts in order to develop a method for monitoring polysaccharide composition. The samples resulting from wine concentration, dialysis and lyophilisation were fractionated by graded ethanol precipitation. Through chemometric analysis of the FTIR spectra, the wine extracts were differentiated according to their polysaccharide content and correlated with the wine extracts were differentiated according to the chemical nature of the samples under study (Regmi et al., 2012). Contextual information was obtained in chemometric investigation of red wine polysaccharides with FTIR application. The samples were obtained from the purification of red wine. The information extracted from the spectral region 950-1850/cm, allowed to identify the different polysaccharide families (mannoproteins, arabinogalactan-proteins, RG-I, and RG-II) (Boulet et al., 2007).
**Determination of phenolic compounds**

In addition to the basic monitoring analysis, other relevant chemical markers are needed to ensure the quality of the final product, such as the phenolic content, chloride and sulphate content, antioxidant capacity, and chemical age of the wine. However, the quantification of red wine tannins faces specific difficulties due to spectral interferences from other components of the wine matrix. To overcome these difficulties, some wines were purified by a Solid-Phase extraction column before analysis. Furthermore, six different predictive models were evaluated in combination with spectral preprocessing procedures to develop a quantification method for tannins with the use of FT-MIR. Besides, two different reference methods, protein precipitation, and phloroglucinolysis were examined as well. The best combination for tannin quantification and prediction of the mean degree of polymerisation was the one using PLS regression of the spectrum full range (650-4000/cm), the second derivative of the spectra, and phloroglucinolysis as the reference method. The samples used were from ‘Carménére’ grapevine cultivar and resulting from two different vintages and different Chilian wineries (Fernández and Agosín, 2007). In another similar study, four different variable selection tools were evaluated to identify the specific FT-MIR spectra area for tannin quantification in order to avoid the time-consuming pre-treatment extraction. The two regions selected and identified by all the tools under examination were 1485-1425/cm and 1060-995/cm. Based on these regions, a successful calibration model was developed using a large data set (128 commercial red wines from different vintages, grapevine varieties, and production countries) for the quantification of red wine tannins (Jensen et al., 2008).

ATR-FTIR was compared with electronic tongue in terms of their discrimination capability of phenolic content. PLS-1 was applied to the data set derived from both techniques and led to good estimations for simultaneous determination of Total Phenolics Index (TPI), CIELab, and Glories parameters. Both techniques were considered robust enough to be applied for a quick and efficient assessment of phenolic composition but electronic tongue showed better prediction values (higher coefficients of correlations and lower residual errors) (Garcia-Hernandez et al., 2020). Determinations of twelve anthocyanins and three different groups of anthocyanins were examined with the application of FT-IR combined with PLS-R, and HPLC-DAD was used as a reference method. One hundred fifty-three Rioja wines (young wines of 2004 and 2005, “Crianza”, and “Reserva” wines) were analysed for the calibration model, which was evaluated by internal and external data sets. Unfortunately, only the anthocyanin content of young wines was able to be predicted with a low error. These results were assigned to the higher concentration level of anthocyanins in young wines than in the aged ones (Romera-Fernández et al., 2012). In another work, ATR-FTIR was used for the prediction of total phenolic and flavonoid contents and antioxidant activities (DPPH and FRAP assays) in ‘Moscatel’ dessert wines. Through chemometric analysis of the spectral region 900-1800/cm, a good determination capability for total phenolic and flavonoid composition was observed, but the results were not enough satisfactory regarding the determination of antioxidant activities. This fact was attributed to the poor specificity of the DPPH and FRAP methods and the possible influence of interferent molecules, not included in the study (for example polysaccharides) (Silva et al., 2014). Another research was focused more specifically on the determination of the total antioxidant capacity of red wines. FTIR matched with PLS regression was applied in the fingerprint region 965-1543/cm and the obtained results showed promising estimation values. However, more studies in extended data sets are needed for the validation of the method and its application in the industry (Versari et al., 2010). The use of FTIR spectra from samples of ‘Cabernet Sauvignon’ Mexican wines in combination with multivariable statistical analysis led to the creation of an optimised chemometric model for the determination of total bioactive phenolic compounds and antioxidant capacities (ABTS and DPPH assays). The spectral area used was 824-1550/cm (Grijalva-Verdugo et al., 2018). In a comparison study between FTIR and UV-vis spectroscopy, FTIR showed more accurate results for the prediction of tannin concentration, while UV-vis stood out for the prediction of anthocyanins. However, the analysis of some wavelengths in the visible region in combination with FTIR spectrum contributed to a better determination of anthocyanin content (Miramont et al., 2020). The quantification of hydrolysable tannins is of great importance for the wine industry because they are responsible for very important modifications in the wine’s organoleptic characteristics (Michel et al., 2011). The determination capability of FTIR spectroscopy in combination with chemometrics was investigated by analysing the spectral region 950-1821/cm deriving for wine samples aged in different wooden barrels. The method was an effective tool for the prediction of total ellagitannins concentration in samples aged in different French and American oak barrels or being in contact with oak sticks of different types of oak. However, the estimations for samples aged in chestnut barrels were not good enough. This observation could be related to the existence of ellagic acid derivatives (Basalekou et al., 2019). In a more detailed study considering different samples derived from all the winemaking stages, ATR-FTIR and UV–Vis were combined to investigate the fingerprint region of polyphenols (900-1800/cm). For this purpose, the wine samples of
red and white varieties underwent a solid-phase extraction. Both spectra derived from ATR-FTIR and UV-Vis were analysed by a Heat Map to establish correlations between the spectra. The method was able to identify peaks correlated with anthocyanins and flavanols for the red wines, non-flavanoids and flavonoids for the white wines, and glycosylated phenolics for both wines (Scano, 2021).

**Determination of other quality parameters**

Another important analysis for certified wines to enter the market is the assessment of the wine ash. FTIR matched with multivariate analysis with different spectral pre-treatments was evaluated for its applicability in the prediction of ash content in wines. The use of four wavelengths and a centred average proved to be the most efficient solution. The validation of the method provided good estimations values and, according to the authors, after enlargement of the data set the method could be applied on an industrial scale (Jug et al., 2017). In addition, several other chemical elements, like chloride and sulphate anions, are considered important quality markers of the wine. FTIR spectroscopy combined with chemometrics was used successfully for the determination of chloride and sulphate in a wide range of Portuguese wine types. The calibration models were able to achieve quantification of the sulphate and semi-quantification of the chloride (Teixeira-dos Santos et al., 2016).

**Authentication studies**

Moreover, due to the globalisation of the wine market and due to many cases of mislabelling and adulterations, a series of authentication studies have been carried out to guarantee the quality and authenticity of the final product. The related studies have resorted to chemical compounds that are considered as authentication “markers” and are characteristic and unique for each parameter or comparisons with known and authentic samples (Basalekou et al., 2020).

A chemometric approach took place comparing the performance of FT-NIR and $^1$H NMR in differentiating adulterated wine samples. The adulterated samples derived from a blend with wine very rich in anthocyanins (called “Rossissimo”) or from the addition of anthocyanins extracted from black rice. The results achieved by NIR spectroscopy gave less satisfactory results than NMR, but still, a relationship with the anthocyanins content and the NIR spectra was noticeable. These results were ascribed to the low sensitivity of the method in determining low concentrations levels and to the possible matrix effect (Ferrari et al., 2011). FTIR-ATR combined with the multivariate analysis was used for the discrimination of “Marsala” wine from Sicily (Italy) based on the grapevine cultivar, production technology, and ageing. The samples were differentiated according to their sugar concentration, and a clear discrimination between the high-quality samples (Virgin) and the remaining ones was found. Moreover, “Marsala” wine samples with different ageing times were discriminated successfully using the spectral area 1058-1076/cm (Condurso et al., 2018). Successful differentiation of Greek red wines was performed using FT-MIR spectroscopy and specific software. For this purpose, extracts of wine phenolic components were obtained by solid-phase extraction with C-18 columns and elution by methanol. The study allowed to discriminate different grapevine cultivars based on the absorption characteristics in the fingerprint spectral region of 900-1800/cm, leading to the creation of libraries for each cultivar. The characterisation of a new unknown sample took place using the spectra bank data set (Tarantilis et al., 2008). FTIR Spectroscopy and chemometrics were used in order to distinguish between sweet wines in terms of sugar content, acidity, and total polyphenol content. Through PCA analysis of MIR spectra derived from some Romania wine samples, it was possible to discriminate the red from the white samples and the dry and the half-dry ones (Topala and Tataru, 2018). In a similar study, Romanian sweet wines and Canadian ice wines were differentiated according to their MIR spectra correlated with their different chemical composition. The chemometric approach underlined the biggest differences in terms of sugar content due to the very high sugar concentration of the ice wines, while they did not express significant differences in terms of antiradical activity (Topala and Tataru, 2019). An identification study of oenological tannins took place using different analytical techniques. Among them, FTIR was applied and was able to identify five of the six samples of oenological tannins (three of them condensed tannins and two mixtures of hydrolysable tannins). The difficulties of identifying and quantifying the tannins under analysis were assigned to the different botanical origin and to the extraction/purification processes, and could be overcome with the enlargement of the database (Lagh et al., 2010). In the same context, two different analytical approaches were examined for red wine authentication in terms of varietal and vintage year identification. Through chemometric analysis of the data, UV-Vis spectroscopy was considered more appropriate for varietal discrimination and FTIR spectroscopy for vintage year discrimination. However, both multiparametric techniques allowed to identify red wine samples aged for more than six years, correlating spectral differences with the formation of new compounds during ageing (Geană et al., 2019). Moreover, successful differentiation of wine spirit samples aged through different technologies was possible with the use of FTIR (Anjos et al., 2020).
functional data analysis (FDA) was applied to ATR-FTIR spectra, and was proven efficient in discriminating the studied wine spirits. The observed results were confirmed with the parallel determination of various analytical parameters, including chromatic characteristics (CIExLab method), total phenolic index, concentrations of furfural, ellagic acid, vanillin, and coniferaldehyde, and total content of low molecular weight phenolic compounds (HPLC). In this context, another discrimination study was performed using NIR spectra combined with UV-Vis spectra, and chemometrics. Samples from different subzones of the same Protected Designation of Origin (PDO) were differentiated and also samples that did not belong to this PDO were identified (Martelo-Vidal et al., 2013). The discrimination capacity of FTIR was evaluated in terms of authentication of sweet wines from Cyprus, (“Commandaria”) through chemometric analysis of FTIR spectra of 65 sweet wines. Differentiation of several types of Commandaria wine (non-fortified, fortified and homemade) from Cyprus and other countries was also made (Ioannou-Papayianni et al., 2011). A series of different analytical approaches were used in combination with chemometrics for the characterisation of sparkling wines (cava wines) from different grapevine cultivars, wine blends, and wine-making stages. FTIR spectroscopy was applied for the analysis of total reducing sugars, pH, acetic acid, total acidity, malic acid, lactic acid, and alcoholic strength. The models were established mainly depending on the polyphenol and organic acid content. Rosé wine samples and ‘Chardonnay’ wines samples showed the highest levels of phenolic acids. Concerning the samples underwent malolactic fermentation, the discrimination was mainly based on malic acid and lactic acid contents (Izquierdo-Llopard and Saurina, 2021). Tri-step IR spectroscopy in combination with electronic nose was applied for an authentication study of ‘Cabernet Sauvignon’ wines. Through chemometric analysis of the fingerprint region of ethanol and sugar dry wines (DW), semi-dry wines (SD), semi-sweet wines (SS), and sweet wines (SW), were discriminated Furthermore, it was possible a quantitative determination of ethanol and total sugar levels based on PLS regression (Wang et al., 2022).

CONCLUDING REMARKS
Several studies found in the literature have been focused on the applicability of FTIR in monitoring the wine production from the grapes to the final product. The wine industry demands constant product monitoring and requires process control from the grape ripening until bottling. IR spectroscopy is already being applied in the wine industry from the vineyard, assessing the grape maturation, during the winemaking process, in real-time analysis of the general chemical parameters in musts and wines, until bottling. The measurements have been performed with portable and non-portable instrumentation coupled with calibration models already built by the manufacturer. However, other IR applications combined with chemometric approaches could also be considered a promising tool for the determination of more specific parameters and applied widely in all the winemaking stages. Methodologies that depend on IR analysis demand minimal or zero sample preparation, are environmentally friendly, easy to use, fast, and more economical than the traditional techniques. As a result of all these advantages, FTIR measurements are already applied in the wine sector, but many challenges still remain. The most important limitations and critical aspects of the studies that were mentioned in the present work are summarised below.

First of all, it was noticeable that the different preparation modes can influence the spectra and the determination of the desired oenological parameters. Therefore, careful consideration of the appropriate mode for each factor should be taken. Moreover, most of the developed methodologies found in the literature depend on feasibility studies using a limited number of samples (less than 100 in most cases) and the validation of the methods took place with the cross-validation tests. External validation using independent sets of samples is needed. Besides, developed models were not applied to different grapevine cultivars, harvests, and types of wines, and led to a poor representativity of the samples. Furthermore, many developed methods were focused on a specific oenological parameter or chemical compound or they have been applied to a specific stage of the winemaking process. In fact, the simultaneous determination of a large number of parameters was more challenging in most cases. Moreover, compounds with a concentration higher than 1 g/L are easier to be determined by FTIR due to their ability of engaging absorption phenomena.

Finally, the complexity of the wine matrix and the chemical similarity of the compounds under study makes the interpretation of the spectra very difficult. In addition, the presence of water and ethanol leads to interference in the IR spectra due to their large absorption band.

In the context of all these critical points, it is of great importance that the initial commercial analytical calibration be updated by each winery, using samples representative of those under study. Most applications in the wine industry consider the use of techniques based on FTIR for the evaluation of the basic chemical composition. In the future, more applications evaluating more specific factors (for example aroma compounds or tannins) or for
authenticity purposes would be of great interest to the contemporary wine industry.

CONFLICTS OF INTEREST: The authors declare no conflict of interest.

REFERENCES
Alabe, 2021. Available at: https://www.alabe.pt (accessed on 17.12.2021).
Alecu G.C., Olteanu R.L., Radulescu C., Stirbescu R. M., Necula I., Boboaca-Mihaescu D.N., 2020. Characterization of red grapes skin extracts using vibrational spectroscopy and chemometrics. J. Sci. Arts, 51, 475-490.
Alexandridis A.K., Zapranis, A.D., 2013. Wavelet neural networks: A practical guide. Neural Netw., 42, 1-27.
Anjos O., Santos A.J.A., Estevinho L.M., Caldeira I., 2016. FTIR–ATR spectroscopy applied to quality control of grape-derived spirits. Food Chem., 205, 28-35.
Anjos O., Comesaña M.M., Caldeira I., Pedro S.L., Oller P E., Canas S., 2020. Application of functional data analysis and FTIR-ATR spectroscopy to discriminate wine spirits aging technologies. Mathematix, 8, 1-21.
Basalekou M., Kallithraka S., Tarantilis P.A., Kotsiridis Y., 2019. Ellagitannins in wines: future prospects in methods of analysis using FT-IR spectroscopy. LWT, 101, 48-53.
Basalekou M., Pappas C., Tarantilis P.A., Kallithraka S., 2020. Wine authenticity and traceability with the use of FT-IR. Beverages, 6, 1-13.
Bauer R., Nieuwoudt H., Bauer F.F., Kossmann J., Koch K. R., Esbenshade K.H., 2008. FTIR spectroscopy for grape and wine analysis. Anal. Chem., 80, 1371-1379.
Bell S.J., Henschke P.A., 2005. Implications of nitrogen nutrition for grapes, fermentation, and wine. Aust. J. Grape Wine Res., 11, 242-295.
Blum M.M., John H., 2012. Historical perspective and modern applications of attenuated total reflectance- Fourier transform infrared spectroscopy (ATR-FTIR). Drug Test. Anal., 4, 298-302.
Boulet J.C., Williams P., Doco T., 2007. A Fourier transform infrared spectroscopy study of wine polysaccharides. Carbohydr. Polym., 69, 79-85.
Brandão A.F.V.A., 2019. Aplicação de Metodologia FTIR para controlo de Qualidade em Vinhos e Aguardentes. Master thesis, Universidade do Porto.
Buratti S., Ballabio D., Giovanelli G., Zuluanga C.M., Dominguez A.M., Moles A., Benedetti S., Sinelli N., 2011. Monitoring of alcoholic fermentation using near-infrared and mid-infrared spectroscopy combined with electronic nose and electronic tongue. Anal. Chim. Acta, 697, 67-74.
Canal C., Ozén B., 2017. Monitoring of wine process and prediction of its parameters with mid-infrared spectroscopy. J. Food Process Eng., 40, 1-10.
Cavaglia J., Giussani B., Mestres M., Puxeu M., Busto O., Ferre J., Boqué R., 2019. Early detection of undesirable deviations in must fermentation using a portable FTIR-ATR instrument and multivariate analysis. J. Chemom., 33, 1-11.
Cavaglia J., Schorn-García D., Giussani B., Ferre J., Busto O., Acena L., Mestres M., Boque R., 2020a. Monitoring wine fermentation deviations using an ATR-MIR spectrometer and MSPC charts. Chemom. Intell. Lab. Syst., 201, 1-7.
Cavaglia J., Schorn-García D., Giussani B., Ferre J., Busto O., Acena L., Mestres M., Boqué R., 2020b. ATR-MIR spectroscopy and multivariate analysis in alcoholic fermentation monitoring and lactic acid bacteria spoilage detection. Food Control, 109, 1-7.
Coimbra M.A., Gonçalves F., Barros A.S., Delgadillo L., 2002. Fourier transform infrared spectroscopy and chemometric analysis of white wine polysaccharide extracts. J. Agric. Food Chem., 50, 3405-3411.
Condurso C., Cincotta F., Tripodi G., Verzera A., 2018. Characterization and aging monitoring of marsala dessert wines by a rapid FTIR-ATR method coupled with multivariate analysis. Eur. Food Res. Technol., 244, 1073-1081.
Costa N.L., García Llobodanin L.A., Castro I.A., Barbosa R., 2019. Using Support Vector Machines and neural networks to classify Merlot wines from South America. Inf. Process. Agric. 6, 265-278.
Dambers R., Gisht M., Cozolino D., 2015. A review of the state of the art, limitations, and perspectives of infrared spectroscopy for the analysis of wine grapes, must, and grapevine tissue. Appl. Spectrosc. Rev., 50, 261-278.
Dewey F. M., Hill M., DeSchenzo R., 2008. Quantification of Botrytis and laccase in wine grapes. Am. J. Enol. Vitic., 59, 47-54.
Di Egidio V., Sinelli N., Giovaneli G., Molea A., Casiraghi E., 2010. NIR and MIR spectroscopy as rapid methods to monitor red wine fermentation. Eur. Food Res. Technol., 230, 947-955.
Dong D., Zheng W., Wang W., Zhao X., Jiao L., Zhao C., 2014. Analysis and discrimination of grape spoilage via volatiles: a comparison between long optical path Fourier-transform-infrared spectroscopy and sensor arrays. Analyst, 139, 5028-5034.
Felix instruments, 2021. Available at https://felixinstruments.com/ (accessed on 17.12.2021)
Fernández K., Agosin E., 2007. Quantitative analysis of red wine tannins using Fourier-transform mid-infrared spectrometry. J. Agric. Food Chem., 55, 7294-7300.
Ferreira M.L., Costa A.M., Ribeiro N., Simões T., 2009. Quality control in FTIR wine analysis, acceptance of analytical Results. Ciência Téc. Vitiviníc., 24, 47-53.
Ferreira M.L., 2015. Automatização da análise de vinhos-FTIR. In: Química enológica-métodos analíticos. Curvelo-García A. S., Barros P., (ed.). Publindustria, Edições Técnicas, LDA., Porto.
Ferrari E., Foca G., Vignali M., Tassi L., Ulrici A., 2011. Adulteration of the anthocyanin content of red wines: perspectives for authentication by Fourier transform-infrared and 1H NMR spectroscopies. Anal. Chim. Acta, 701, 139-151.
Fragoso S., Acena L., Guasch J., Mestres M., Busto O., 2011. Quantification of phenolic compounds during red winemaking using FT-MIR spectroscopy and PLS-regression. J. Agric. Food Chem., 59, 10795-10802.
Fu Y., Lim L.T., Mcnicholas P.D., 2009. Changes on enological parameters of white wine packaged in bag-in-box during secondary shelf life. J. Food Sci., 74, 608-618.
Garcia-Hernandez C., Salvo-Comino C., Martin-Pedrosa F., Garcia-Cabezón C., Rodriguez-Mendez M.L., 2020. Analysis of red wines using an electronic tongue and infrared spectroscopy. Correlations with phenolic content and color parameters. LWT, 118, 1-8.
Geană E.L., Ciucure C.T., Apetrei C., Artem V., 2019. Application of spectroscopic UV-Vis and FT-IR screening techniques coupled with multivariate statistical analysis for red wine authentication: varietal and vintage year discrimination. *Molecules*, 24.

Gerhards D., Buchl N., Wenning M., Scherer S., Von Wallbrunn C., 2015. Terroir of yeasts? application of FTIR spectroscopy and molecular methods for strain typing of yeasts. *BIO Web Conf.*, 5, 02001.

Grangeteau C., Gerhards D., Terrat S., Deguited S., Alexandre H., Guilloux-Benatter M., Von Wallbrunn C., Rousseaux S., 2016. FT-IR spectroscopy: a powerful tool for studying the inter-and intraspecific biodiversity of cultivable non-Saccharomyces yeasts isolated from grape must. *J. Microbiol. Methods*, 121, 50-58.

Griffiths P.R., de Haseth J.A., 2007. *Fourier Transform Infrared Spectrometry*, John Wiley & Sons, Inc.

Grijalva-Verdugo C., Hernández-Martínez M., Meza-Márquez O.G., Gallardo-Velázquez T., Osorio-Revilla G., 2018. FT-MIR spectroscopy and multivariate analysis for determination of bioactive compounds and antioxidant capacity in Cabernet Sauvignon wines. *CYTA - J. Food*, 16, 561-569.

Guillen-Casla V., Rosales-Conrado N., Perez-Arribas L.V., Polo-Diez L.M., 2011. Principal component analysis (PCA) and multiple linear regression (MLR) statistical tools to evaluate the effect of E-beam irradiation on ready-to-eat food. *J. Food Comp. Anal.*, 24, 456-464.

He Z., Duan X., Ma Z., 2013. Measuring routine parameters of wine by ATR-MIR spectroscopy. *Appl. Mech. Mater.*, 397, 1749-1752.

Hill G. N., Evans K. J., Beresford R. M., Dambergs R. G., 2013. Near and mid-infrared spectroscopy for the quantification of botrytis bunch rot in white wine grapes. *J. Near Infrared Spectrosc.*, 21, 467-475.

Holzapfel B. P., Smith J. P., Field S. K., Hardie W. J., 2010. Dynamics of carbohydrate reserves in cultivated grapevines. *Hort. Rev.*, 36, 143-211.

Ioanou-Papayianni E., Kokkinotra R. I., Theocharis C. R., 2011. Authenticity of Cypriot sweet wine commandaria using FT-IR and chemometrics. *J. Food Sci.*, 76, 420-427.

Isquierdo-Llopart A., Saurina J., 2021. Multi-sensor characterization of sparkling wines based on data fusion. *Chemosensors*, 9, 1-18.

Jamwal R., Amit, Kumari S., Sharma S., Kelly S., Cannavan A., Singh D. K., 2021. Recent trends in the use of FTIR spectroscopy integrated with chemometrics for the detection of edible oil adulteration. *Vib. Spectrosc.*, 113, 103222.

Jensen J. S., Egeo B., Meyr A.S., 2008. Identification of spectral regions for the quantification of red wine tannins with Fourier transform mid-infrared spectroscopy. *J. Agric. Food Chem.*, 56, 3493-3499.

Jug T., Boni S., Kosmerl T., 2017. FTIR analysis of ash in wine. *BIO Web Conf.*, 9, 02023.

Kumar K., Giehl A., Schweiggert R., Patz C.D., 2021. Network analysis on Fourier-transform infrared (FTIR) spectroscopic data sets in an Eigenspace layout: Introducing a novel approach for analyzing wine samples. *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 251, 1-9.

Kyraréou M., Pappas C., Voskidi E., Kotseridis Y., Basalekou M., Tarantilis P.A., Kalithraka S., 2015. Diffuse reflectance Fourier transform infrared spectroscopy for simultaneous quantification of total phenolics and condensed tannins contained in grape seeds. *Ind. Crops Prod.*, 74, 784-791.

Laghi L., Parpinello G.P., Del Rio D., Calani L., Mattioli A. U., Versari A., 2010. Fingerprint of enological tannins by multiple techniques approach. *Food Chem.*, 121, 783-788.

Lemos A.M., Machado N., Egea-Cortines M., Barros A.I., 2020. ATR-MIR spectroscopy as a tool to assist 'Tempranillo' clonal selection process: geographical origin and year of harvest discrimination and oenological parameters prediction. *Food Chem.*, 325, 1-8.

Lucarini M., Durazzo A., Kiefer J., Santini A., Lombardi-Boccia G., Stoot E.B., Romani A., Lampe A., Ferrari Nicoli S., Gabrielli P., Bevilacqua N., Campo M., Morassut M., Cecchini F., 2020. Grape Seeds: chromaticographic profile of fatty acids and phenolic compounds and qualitative analysis by FTIR-ATR spectroscopy. *Foods*, 9, 1-14, 10.

Manley M., Van Zyl A., Wolf E.H., 2001. The evaluation of the applicability of Fourier transforms near-infrared (FT-NIR) spectroscopy in the measurement of analytical parameters in must and wine. *S. Afr. J. Enol. Vitic.*, 22, 93-100.

Martelo-Vidal M.J., Domínguez-Agís F., Vázquez M., 2013. Ultraviolet/visible/near-infrared spectral analysis and chemometric tools for the discrimination of wines between subzones inside a controlled designation of origin: a case study of Rías Baixas. *Aust. J. Grape Wine Res.*, 19, 62-67.

Mendes E., Duarte N., 2021. Mid-infrared spectroscopy as a valuable tool to tackle food analysis: a literature review on coffee, dairies, honey, olive oil, and wine. *Foods*, 10, 1-32, 477.

Michel J., Jourdes M., Silva M.A., Giordanengo T., Mourey, N., Teissedre P.L., 2011. Impact of concentration of ellagitannins in oak wood on their levels and organoleptic influence in red wine. *J. Agric. Food Chem.*, 59, 5677-5683.

Miramont C., Jourdes M., Teissedre P.L., 2020. Development of UV-vis and FTIR partial least squares models: comparison and combination of two spectroscopy techniques with chemometrics for polyphenols quantification in red wine. *OENO Oné*, 4, 779-792.

Moore J.P., Zhang S.L., Nieuwoudt H., Divol B., Trygg J., Bauer F.F., 2015. *J. Agric. Food Chem.*, 63, 10054-10063.

Moldes O.A., Mejuto J.C., Rial-Otero R., Simal-Gándara J. 2017. A critical review on the applications of artificial neural networks in winemaking technology. *Crit. Rev. Food Sci Nutr.*, 57, 2896–2908.

Moreira J.L., Marcos A.M., Barros P., 2002. Analysis of Portuguese wine by Fourier transform infrared spectrometry. *Ciência. Tec. Vitiv.*, 17, 27-33.

Moreira J.L., Santos L., 2005. Analysis of organic acids in wines by Fourier-transform infrared spectroscopy. *Anal. Bioanal. Chem.*, 382, 421–425.

Murr C., Chimeno-Trinchet C., Díaz-García M.E., Badía-Lañoñ R., Fernández-González A., 2019. Artificial neural network and attenuated total reflectance-fourier transform infrared spectroscopy to identify the chemical variables related to ripeness and variety classification of grapes for protected designation of origin wine production. *Comput. Electron. Agric.*, 164, 1-6.

Musingarabwi D.M., Nieuwoudt H.H., Young P.R., Eyéghé-Bickong H. A., Vivier M. A. D. M., 2016. A rapid qualitative and quantitative evaluation of grape berries at various stages of development using Fourier-transform infrared spectroscopy and multivariate data analysis. *Food Chem.*, 190, 253-262.

Nieuwoudt H.H., Pretorius I.S., Bauer F.F., Nel D.G., Prior B.A.H., 2006. Rapid screening of the fermentation profiles of wine yeasts by Fourier transforms infrared spectroscopy. *J. Microbiol. Methods*, 67, 248-256.
Nogales-Bueno J., Baca-Bocanegra B., Rooney A., Hernández-Hierro J.M., José Heredia F., Byrne H.J., 2017. Linking ATR-FTIR and Raman features to phenolic extractability and other attributes in grape skin. *Talanta*, **167**, 44-50.

OIV, resolution OIV/VITI 333/2010. Definition of vitivinicultural ‘terroir’. International organization of vine and wine, Paris, France.

OIV, resolution OIV/OENO 390/2010. Guidelines on infrared analyzers in oenology. International organization of vine and wine, Paris, France.

Olale K., Walyambahilah W., Mohammed S.A., Sila A., Shepherd K., 2017. Application of DRIFT-FTIR spectroscopy for quantitative prediction of simple sugars in two local and two 34 Floridian mango (*Mangifera indica L.*) cultivars in Kenya. *J. Anal. Sci. Technol.*, **8**, 1-13.

Ortega-Regules A., Ros-García J.M., Bautista-Ortín A.B., López-Roca J.M., Gómez-Plaza E., 2008. Changes in skin cell wall composition during the maturation of four premium wine grape varieties, *J. Sci. Food Agric.*, **88**, 420–428.

Ozturk B., Yucesoý D., Ozen B., 2012. Application of mid-infrared spectroscopy for the measurement of several key parameters of alcoholic beverages, wine, and raki. *Food Anal. Methods*, **5**, 1435-1442.

Pappas C., Kyraleou M., Voskidi E., Kotseridis Y., Taranilis P.A., Kalithraka S., 2015. Direct and simultaneous quantification of tannin mean degree of polymerization and percentage of galloylation in grape seeds using diffuse reflectance Fourier transform-infrared spectroscopy. *J. Food Sci.*, **80**, 298-306.

Passos C., Cardoso S.M., Barros A.S., Silva C.M., Cомнbra M.A., 2010. Application of Fourier transform infrared spectroscopy and orthogonal projections to latent structures/parametric least squares regression for estimation of procyanidins average degree of polymerization. *Anal. Chem. Acta*, **661**, 143-149.

Petrovic G., Aleixandre-Tudo J.L., Buica A., 2020. Viability of IR spectroscopy for the accurate measurement of yeast assimilable nitrogen content of grape juice. *Talanta*, **206**, 1-7.

Picque D., Lieben P., Chretien P., Beguin J., Guerin L., 2010. Assessment of maturity of Loire valley wine grapes by mid-infrared spectroscopy. *J. Int. Sci. Vigne Vin.*, **44**, 219-229.

Power A., Truong V. K., Chapman J., Cozzolino D., 2019. From the laboratory to the vineyard-evolution of the measurement of grape composition using NIR spectroscopy towards high-throughput analysis. *High-Throughput*, **8**, 1-9.

Preserova J., Ranc V., Milde D., Kubistov V., Stavek J., 2015. Study of phenolic profile and antioxidant activity in selected Moravian wines during winemaking process by FT-IR spectroscopy. *J. Food Sci. Tech.*, **52**, 6405-6414.

Puz exu M., Andorra L., De Lamo-Castellví S., 2015. Monitoring Saccharomyces cerevisiae grape must fermentation process by attenuated total reflectance spectroscopy. *Food Bioproc. Tech.*, **8**, 637-646.

Rasines-Perea Z., Prieto-Perea N., Romera-Fernández M., Beruet L.A., Gallo B., 2015. Fast determination of anthocyanins in red grape musts by Fourier transform mid-infrared spectroscopy and partial least squares regression. *Eur. Food Res. Technol.*, **240**, 897-908.

Regmi U., Palma M., Barroso C.G., 2012. Direct determination of organic acids in wine and wine-derived products by Fourier transform infrared (FT-IR) spectroscopy and chemometric techniques. *Anal. Chem. Acta*, **732**, 137-144.

Ribereau-Gayon P., Dubourdieu D., Donèche B., Lonvaud A., 2021. Handbook of Enology: Volume 1: The Microbiology of Wine and Vinifications (3rd edition). John Wiley & Sons, Ltd.,Chichester.

Rinnan A., Nargaard L., Van den Berg, F., Thygesen J., Bro R., Engelsen S.B., 2009. Data Pre-processing. In: Infrared Spectroscopy for Food Quality Analysis and Control, Sun D.W. (ed.) Elsevier Inc.

Robinson J.B., 1992. Grapevine nutrition. In: Viticulture, Vol. 2. Practices, 178–208. Coombe, B.G. and Dyr., P.R., (eds.), Winetitles: Adelaide, SA, Adelaide.

Romera-Fernández M., Beruetta L. A., Garmón-Lobato S., Gallo B., Vicente F., Moreda J.M., 2012. Feasibility study of FT-MIR spectroscopy and PLS-R for the fast determination of anthocyanins in wine. *Talanta*, **88**, 303–310.

Rousset S., Bellon-Maurel V., Roger J.M., Grenier P., 2003. Authenticating white grape must variety with classification models based on aroma sensors, FT-IR, and UV spectrometry. *J. Food Eng.*, **60**, 407-419.

Scano P., 2021. Characterization of the medium infrared spectra of polyphenols of red and white wines by integrating FT IR and UV–Vis spectral data. *LWT*, **147**, 1-6,111604.

Schmidtke L.M., Smith J.P., Müller M.C., Holzapfel B.P., 2012. Rapid monitoring of grapevine reserves using ATR–FT-IR and chemometrics. *Anal. Chim. Acta*, **732**, 16-25.

Schmidtke L.M., Schwarz L.J., Scheuermann C., Steel C.C., 2019. Discrimination of Aspergillus spp., Botryts cinerea, and Penicillium expansum in grape berries by ATR-FTIR spectroscopy. *Am. J. Enol. Vitic.*, **70**, 68-76.

Silva S. D., Feliciano R. P., Boas L. V., Bronze M. R., 2014. Application of FTIR–ATR to muscatel dessert wines for prediction of total phenolic and flavonoid contents and antioxidant capacity. *Food Chem.*, **150**, 489-493.

Skoutelas D., Ricardo-Da-Silva M., Laureano O., 2011. Validation and comparison of formol and FT-IR methods for assimilable nitrogen in vine grapes. *S Afr. J. Enol. Vitic.*, **32**, 262-266.

Smith J. P., Schmidtke L.M., Muller M.C., Holzapfel B.P., 2014. Measurement of the concentration of nutrients in grapevine petioles by attenuated total reflectance Fourier transform infrared spectroscopy and chemometrics. *Aust. J. Grape Wine Res.*, **20**, 99-309.

Stuart B., 2004. Experimental Methods. In: Infrared Spectroscopy. Fundamentals and Applications B., 15-44. John Wiley & Sons.

Sun B.S., Pinto T., Leandro M.C., Ricardo-da-Silva J.M. and Spranger M.I., 1999. Transfer of catechins and procyanidins from solid parts of the grape cluster into wine. American Journal of Enology and Viticulture. *Am. J. Enol. Vitic.*, **50**, 179-184.

Swanepoel M., du Toit M., Nieuwoudt H.H., 2007. Optimization of the quantification of total soluble solids, pH, and titratable acidity in south African grape must using Fourier transform mid-infrared Spectroscopy. *S Afr. J. Enol. Vitic.*, **28**, 140-149.

Tarantilis P.A., Troianou V.E., Pappas C.S., Kotseridis Y. S., Polissiou M.G., 2008. Differentiation of Greek red wines on the basis of grape variety using attenuated total reflectance Fourier transform infrared spectroscopy. *Food Chem.*, **111**, 192-196.

Teixeira-dos Santos C.A., Páscoa R.M.N.J., Porto P.A.L.S., Cerdeira A.L., Lopes J.A., 2016. Application of Fourier-transform infrared spectroscopy for the determination of chloride and sulfate in wines. *LWT*, **67**, 181-186.
Titus D., Samuel J.J., Roopan S.M., 2019. Nanoparticle characterization techniques. In: Green synthesis, characterization, and applications of nanoparticles. 313-314. Elsevier Inc.

Topala C.M., Tataru L.D., 2018. Rapid method for the discrimination of Romanian wines based on mid-infrared spectroscopy and chemometrics. Rev. Chim., 69, 469-473.

Topala C.M., Tataru L.D., 2019. ATR-FTIR spectroscopy coupled with chemical and chemometric analysis to distinguish between some sweet wines. Rev. Chim., 70, 2355-2361.

Urtubia A., Pérez-Correa J.R., Meurens, M., Agosin E., 2004. Monitoring large scale wine fermentations with infrared spectroscopy. Talanta, 64, 778-784.

Versari A., Parpinello G.P., Scazza F., Del Rio D., 2010. Prediction of total antioxidant capacity of red wine by Fourier transform infrared spectroscopy. Food Control, 21, 786-789.

Véstia J., Barroso J.M., Ferreira H., Gaspar L., Rato A.E., 2019. Predicting calcium in grape must and base wine by FT-NIR spectroscopy. Food Chem., 276, 71-76.

Vidal S., Francis L., Guyot S., Marnet N., Kwiatkowski M., Gawel R., 2003. The mouth-feel properties of grape and apple proanthocyanidins in a wine-like medium. J. Sci. Food Agric., 83, 564-573.

Wang S., Hu X.Z., Liu Y.Y., Tao N.P., Lu Y., Wang X. C., Lam W., Lin L., Xu C.H., 2022. Direct authentication and composition quantitation of red wines based on Tri-step infrared spectroscopy and multivariate data fusion. Food Chem., 372, 1-8.

Waterhouse A.L., Sacks G.L., Jeffery D.W., 2016. Understanding Wine Chemistry, John Willey and Sons, Inc.

Watson B., 2003. Evaluation of wine grape maturity. In: Oregon Viticulture. 235-245. Hellman, E., (ed.), 1233 Oregon State University Press, Corvallis.

Wynne L., Clark S., Adams M.J., Barnett N.W., 2007. Compositional dynamics of a commercial wine fermentation using two-dimensional FTIR correlation analysis. Vib. Spectrosc., 44, 394-400.

Zhao Z., Liu H., 2007. Spectral feature selection for supervised and unsupervised. ICML ’07, 1151-1157.