Bone morphogenetic protein-4 and bone morphogenetic protein receptors expressions in the adult rat eye

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Abstract

We investigated the expressions of bone morphogenetic protein-4 (BMP4) and its receptors, bone morphogenetic protein receptor IA (BMPRIA), bone morphogenetic protein receptor IB (BMPRIIB) and bone morphogenetic protein receptor II (BMPRII) in the adult rat eye. Interestingly differences in expression profile were observed between BMPRIA and BMPRIIB in the retina. BMPRIA-like immunoreactivity (IR) was very intensely seen in the photoreceptor layer, while BMPRIIB-IR was mainly observed in the other layers. In the cornea, BMP4, BMPRIA, BMPRIIB and BMPRII-IRs were abundantly seen in the cell body of basal cells in the corneal epithelium, and endothelium. In the lens, BMP4, BMPRIA, BMPRIIB and BMPRII-IRs were observed in epithelial cells, lens cortical fiber cells, however they were not seen in the capsule and the central region of the lens. In the iris and ciliary body, strong BMP4 and BMPRIIB-IRs were observed in nonpigmented epithelium. These results suggest that different kinds of BMP signaling should be needed in different areas in the adult eye to keep the shapes, differentiation levels, and functions of various cells.

Introduction

Bone morphogenetic protein-4 (BMP4) is a member of the transforming growth factor β (TGF-β) superfamily. BMP4 signaling has been shown to play an important role in multiple biological events, including neural induction, tissue patterning, epithelial-mesenchymal interactions underlying organogenesis, lineage selection, and in the creation of stem cell ‘niches’ in developing and adult organs. BMP4 is synthesized as a large precursor, and subsequently cleaved to yield a carboxy-terminal mature protein. BMP4 exerts its biological functions by interacting with membrane-bound receptors belonging to the serine/threonine kinase family including bone morphogenetic protein receptor type I (BMPRIA, BMPRIIB) and type II (BMPRII). These receptors form heteromeric complexes of type I and type II receptors in which type II is the ligand binding subunit which phosphorylates type I resulting in intracellular cascade events.

Also in the early stages of the developing eye, BMP4 has been reported to play pivotal roles, including the lens formation, topographic retinotectal projection, and apoptotic cell death. Although BMP4 and its receptors expressions have been well described in the early stages of the developing eye, little information is available for BMP4 and its receptors expressions in the adult eye. In the present study, we thus investigated the expression of BMP4 and its receptors, BMPRIA, BMPRIIB and BMPRII in the adult eye. In the present study, we show that they are abundantly and differentially expressed in the adult eye.

Materials and Methods

Animals

Male Wistar rats at 7 weeks (n=5) were used. For immunohistochemistry, five rats were perfused transcardially under deep anesthesia with saline followed by 0.1 M phosphate buffer (PB) containing 4% paraformaldehyde. Ten eyes (n=5) were removed rapidly, postfixed in the same fixative for 2 h at 4°C and dehydrated in 100% ethanol. They were embedded in routinely paraffin wax, and were sectioned at 4 μm and mounted on glass slides, and these sections were preserved at -30°C until use to prevent the attenuation of immunoreactivity. All experiments conformed to the Guidelines for Animal Experimentation at Hamamatsu University School of Medicine on the ethical use of animals.

Immunohistochemistry

Paraffin sections were deparaffinized in xylene, dehydrated in decreasing alcohol solutions, and endogenous peroxidase was blocked with 0.3% H2O2 in methanol for 20 min. For BMP4 staining, the sections were treated with 1% normal goat serum, 2% bovine serum albumin (BSA) and 0.02% Triton X-100 in 0.1 M PB for 2 h at room temperature, and incubated further in monoclonal mouse anti-BMP4 (1:100 dilution; NCL-BMP-4, Novocastra, Newcastle, UK) overnight at 4°C. After washing with 0.1 M PB, sections were incubated in goat anti-mouse IgG with peroxidase complex (Histofine Simple Stain Rat MAX-PO(M); Nichirei, Tokyo, Japan) for 2 h at room temperature. For BMPRIA, BMPRIIB and BMPRII staining, the sections were treated with 10% normal rabbit serum, 2% BSA and 0.2% Triton X-100 in 0.1 M PB for 2 h at room temperature, and incubated further in polyclonal goat anti-BMPRIA (1:50 dilution; sc-5676, Santa Cruz Biotechnology, Santa Cruz, CA, USA), anti-BMPRIIB (1:100 dilution; sc-5679, Santa Cruz Biotechnology) and goat anti-BMPRII (1:50 dilution; sc-5683, Santa Cruz Biotechnology) overnight at 4°C, respectively. After washing with 0.1 M PB, sections were incubated in rabbit anti-goat IgG with peroxidase complex (Histofine Simple Stain Rat MAX-PO(G); Nichirei) for 2 h at room temperature. After the final wash with 0.1 M PB, immunoreaction was visualized with 3,3'-diaminobenzidine (DAB) (Wako, Osaka, Japan). The anti-
BMP4 antibody was a mouse monoclonal antibody raised against the recombinant mouse bone morphogenetic protein-4. The anti-BMPRIA antibody and the anti-BMPRIB antibody were affinity purified goat polyclonal antibodies raised against the N-terminal domains of each BMPR protein of human origin, respectively. The anti-BMPRII antibody was an affinity purified goat polyclonal antibody raised against the cytoplasmic domain of BMPRII protein of human origin. For BMP4 pre-absorption controls, 1 nmol BMP4 recombinant protein was added to the diluted primary antibody (100 μL), and incubated overnight at 4°C. For BMPRIA, BMPRIB and BMPRII pre-absorption controls, 40 μg of each blocking peptide (sc-5676 P, sc-5679 P and sc-5683 P; Santa Cruz Biotechnology, respectively) which were linked with 20 μL of NHS-activated Sepharose 4 Fast Flow (GE Healthcare Bio-Science KK, Tokyo, Japan), were added to the diluted primary antibodies (100 μL), and incubated overnight at 4°C. And then, each supernate was used for immunostaining. Three cross-sections of eyes at 80-μm intervals were used for each immunostaining and observation per rat. Brightfield images were obtained with a microscope (Vanox-AHBS3; Olympus, Tokyo, Japan). They were further processed in a graphic editing program (Photoshop; Adobe, Tokyo, Japan) to obtain figures.

cDNA fragments encoding the C-terminal domain of rat BMP4 (aa 293-408, AY184241) were generated by RT-PCR using rat brain total RNA and subcloned into the pGEX-5X-1 vector. After transfection into BL21 E. coli, GST-BMP4 protein was expressed and purified with glutathione-sepharose 4B (Amersham Biosciences, Uppsala, Sweden) according to the manufacturer’s instructions.

**Results**

### Specificity of anti-BMP4, BMPRIA, BMPRIB and BMPRII antibodies

The specificity of antibodies used to detect BMP4, BMPRIA, BMPRIB and BMPRII proteins has been validated in our previous reports. Furthermore, to further confirm the specificity of these antibodies, we performed pre-absorption test in the retina and lens. In both regions, pre-absorption of the antisera with the corresponding antigens completely abolished the immunostaining. These data indicate that the antibodies specifically recognize BMP4 and BMP receptor proteins in the eye.

![Expressions of BMP4, BMPRIA, BMPRIB and BMPRII in the retina](image)

**Figure 1.** Pre-absorption tests for anti-BMP4 and BMP receptors antibody. BMP4 (a), BMPRIA (b), BMPRIB (c) and BMPRII (d) were immunostained with the corresponding antibodies in the adult rat retina. (e–h) Pre-absorption of the antibodies for BMP4, BMPRIA, BMPRIB and BMPRII using the corresponding antigens completely abolished the immunostainings, respectively. Note that intense BMP4-IR and BMPRIB-IR are observed in the nerve fiber layer and ganglion cell layer (arrowheads in a and c). In addition, note that, in the photoreceptor cell layer, intense BMPRIA-IR and BMPRII-IR are detected in the photoreceptor cell layer (arrowheads in b and d). Ch, choroidea; GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; NFL, nerve fiber layer; ONL, outer nuclear layer; PE, pigmented epithelium; PR, Photoreceptor cell layer; Sc, sclera. Scale bars, 50 μm (a–h).
Moderate BMP4-IR was seen in the inner plexiform layer, outer nuclear layer and pigmented epithelium (Figure 1a). BMP4 expression levels were weak in the inner nuclear layer and the outer segment of the photoreceptor cell layer (Figure 1a). Interestingly differences in expression profile were observed between BMPRIA and BMPRIB in the retina. BMPRIA was very intensely expressed in the outer segment of the photoreceptor cell layer (arrowheads in Figure 1b), while BMPRIB expression was weak in this layer (Figure 1c). In contrast, intense BMPRIB expression was observed in the nerve fiber layer, ganglion cell layer, inner plexiform layer (arrowheads in Figure 1c), and pigmented epithelium, while BMPRIA expression in these areas were weak. BMPRII were intensely expressed in the outer segment of the photoreceptor cell layer (arrowheads in Figure 1d), and weakly expressed in the other layers (Figure 1d). In the choroid, BMP4, BMPRIA, BMPRIB and BMPRII were weakly expressed (Figure 1 a-d). Closer observation further revealed their detailed expression profiles.

In the nerve fiber layer, where axons of ganglion cells and astrocytes exist, strong BMP4 and BMPRIB-IRs were observed (arrowheads in Figure 2 a,c), while BMPRIA and BMPRII-IRs were weak (arrowheads in Figure 2 b,d). In the ganglion cell layer, ganglion cell bodies and their neurites were strongly stained with anti-BMP4 and anti-BMPRIB antibodies (arrowheads in Figure 2 a,c), while BMPRIA and BMPRII-IRs were weak (arrowheads in Figure 2 b,d). In the inner plexiform layer, where neurites from bipolar cells, amacrine cells and ganglion cells make synapses, many line-like structures were stained intensely with anti-BMPRII antibody and moderately with anti-BMP4 antibody (Figure 2 a,c). In contrast, BMPRIA-IR and BMPRII-IRs were moderately observed in epithelial cells (arrowheads in Figure 4 a-d).

Expressions of BMP4, BMPRIA, BMPRIB and BMPRII in the lens

We observed intense BMP4, BMPRIA, BMPRIB and BMPRII-IRs in the lens (Figure 4 a-d). Pre-absorption of the antisera with the antigens for BMP4, BMPRIA, BMPRIB and BMPRII completely abolished these immunoreactivities (Figure 4 e-h). The mature lens is a polarized structure consisting of a mitotic epithelial layer that covers the anterior surface and terminally differentiated lens fibers that occupy the interior volume and the posterior surface. BMP4, BMPRIA, BMPRIB and BMPRII-IRs were moderately observed in epithelial cells (arrowheads in Figure 4 a-d).

Interestingly strong BMP4, BMPRIA, BMPRIB and BMPRII-IRs were observed in the lateral edge of the equatorial region, and gradually

Figure 2. BMP4 and BMP receptors expressions in the adult rat retina. BMP4, BMPRIA, BMPRIB and BMPRII expressions in the upper layers (a-d, respectively) and lower layers (e-h, respectively) of the adult rat retina. Note that strong BMP4 and BMPRIB-IRs are observed in the nerve fiber layer and ganglion cell layer (arrowheads in a and c), while BMPRIA and BMPRII-IRs are weak there (arrowheads in b and d). In addition, note that, in the photoreceptor cell layer, the inner segment shows strong BMPRIA-IR, moderate BMP4 and BMPRIB-IRs and weak BMPRII-IR (arrowheads e-h), and the outer segment exhibits very strong BMPRIA-IR, strong BMPRII-IR, and weak BMP4 and BMPRIB-IR (arrowheads in e-h). Ch, choroid; GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; IS, inner segment of photoreceptor cell layer;NFL, nerve fiber layer; ONL, outer nuclear layer; OPL, outer plexiform layer; OS, outer segment of photoreceptor cell layer; PE, pigmented epithelium. Scale bar, 20 µm (a-h).
decreased from the lateral edge to the center of the lens. Nuclei of lens cortical fiber cells were devoid of BMP4, BMPRIA, BMPRIB and BMPRII-IRs (arrowheads in Figure 4 i-l). They were not expressed in the capsul and the central region of the lens (Figure 4 i-l).

Expressions of BMP4, BMPRIA, BMPRIB and BMPRII in the iris and ciliary body

In the iris, strong BMP4 and BMPRIB-IRs were observed in nonpigmented epithelium, while BMPRIA and BMPRII-IRs were weakly seen there (arrowheads in Figure 5 a-d). In the stroma, smooth muscles and pigmented epithelium in the iris, BMP4-IR was observed at moderate level, while BMPRIA, BMPRIB and BMPRII-IRs were observed at weak level (Figure 5 a-d). In the ciliary body, strong BMP4 and BMPRIB-IRs were also observed in nonpigmented epithelium, while BMPRIA and BMPRII-IRs were weakly seen there (arrowheads in Figure 5 e-h). In the other areas in the ciliary body, BMP4, BMPRIA, BMPRIB and BMPRII-IRs were weak (Figure 5 e-h) (Table 1).

Discussion

It is noteworthy that BMPRIA and BMPRIB were complementary expressed in the retina. BMPRIA was very intensely expressed in the photoreceptor cell layer, while BMPRIB was abundantly expressed in the other layers. Every photoreceptor cell has four regions: an outer segment, an inner segment, a cell body, and a synaptic terminal. The outer segment contains a stack of membranous disks, where light signals are converted into electronic signals. Our results indicate that, in a photoreceptor cell, BMPRIA expression was dominant in the outer segment and inner segment, while BMPRIB expression was dominant in the cell body and synaptic terminal, showing that BMPRIA and BMPRIB are differentially expressed in a single photoreceptor cell. Although the functional differences between BMPRIA and BMPRIB have not been clearly elucidated so far, we speculate that to keep the complicated structure of photoreceptor cells, different kinds of BMP signaling should be needed in different areas in photoreceptor cells.

Fischer et al. have reported that in the adult chick intraocular injections of BMP4 on the days immediately before a neurotoxic insult greatly reduced the amount of cell death induced by NMDA.11 Furthermore, in the rodent retina, the absence of BMPRIB results in elevated apoptotic cells in the inner nuclear layer, suggesting that BMP4-mediated signaling is normally required to promote the survival of inner retinal neurons.14 Interestingly, in the present study, we observed BMP4, BMPRIB and BMPRII expressions in the inner nuclear layer, further supporting the possibility that BMP4 may act as a survival factor via BMPRIB and BMPRII in this region.

We found that BMP4 was very intense-
Figure 4. BMP4, BMPRIA, BMPRIB and BMPRII expressions in the adult rat lens. Intense BMP4, BMPRIA, BMPRIB and BMPRII-IRs are observed in the lens (a-d). Note that BMP4, BMPRIA, BMPRIB and BMPRII-IRs are moderately seen in epithelial cells (arrowheads in a-d). In addition, note that abundant BMP4, BMPRIA, BMPRIB and BMPRII expressions are also observed in lens fiber cells, while the nuclei of lens fiber cells are devoid of stainings (arrowheads in e-h). FC, fiber cells; Ir, iris; L, lens; N, nuclei of lens fiber cells. Scale bars, 0.5 mm (a-d) and 20 µm (e-h).
Figure 5. BMP4, BMPRIA, BMPRIB and BMPRII expressions in the adult rat iris and ciliary body. Note that strong BMP4 and BMPRIB-IRs are observed in the nonpigmented epithelium of the iris and ciliary body (arrowheads in a, c, e and g), while BMPRIA and BMPRII-IRs were weakly seen there (arrowheads in b, d, f, and h). CT: connective tissue; L, lens; NE, nonpigmented epithelium; PE, pigmented epithelium; SE, subcapsular epithelium; SM, smooth muscle, St, stroma; Scale bars 20 µm.
BMP ligands and BMP receptors play critical roles in lens development. Faber et al. demonstrated that BMP signaling is required for development of primary lens fiber cells in the mouse. Noggin could suppress primary fiber cell elongation and mouse lens size in explant culture. When dominant negative BMPRIB was expressed in transgenic mice, the mice showed defects in the differentiation of primary lens fiber cells. In the present study, we observed abundant BMP4, BMPRIA, BMPRIIB and BMPRII-IRs in epithelial cells and lens cortical fiber cells, indicating that BMP4 signaling is also important to regulate the differentiation processes of primary lens fiber cells even in the adult rat lens.

The aqueous humor is formed by the bilateral ciliary epithelium. The pigmented epithelium (PE) rests on the connective tissue stroma and the non-pigmented epithelium (NPE) is polarized with its basal lamina facing the posterior chamber of the eye. Interestingly, we found that strong BMP4 and BMPRIB expressions, and weak BMPRIA and BMPRII expressions in non-pigmented epithelium. Chang and colleagues showed that a heterozygous deficiency of BMP4 resulted in anterior segment dysgenesis, elevated intraocular pressure, and optic nerve abnormalities. Although the detailed mechanisms of BMP4 signaling in the adult ciliary body has been not elucidated, BMP4 signaling may be involved in regulating intraocular pressure conditions and be associated with the pathogenesis of glaucoma. In this research, no quantitative analysis has been carried out, focusing on only qualitative analysis. Therefore, no conclusion about quantitative differences of BMPs and BMPRs was obtained from the results of this study, which is limitation of this research. In the future, quantitative investigation of BMPs and BMPRs in eyes will be needed.

Table 1. Distribution and intensity of BMP4, BMPRIA, BMPRIIB and BMPRII-IRs in the adult rat eye.

| Area                  | BMP4 | BMPRIA | BMPRIIB | BMPRII |
|-----------------------|------|--------|---------|--------|
| Retina                |      |        |         |        |
| Nerve fiber layer     | +++  | +      | +++     | +      |
| Ganglion cell layer   | +++  | +      | +++     | +      |
| Inner plexiform layer | ++   | +      | +++     | +      |
| Inner nuclear layer   | +    | +      | +       | +      |
| Outer plexiform layer | +    | +      | +       | +      |
| Outer nuclear layer   | ++   | +      | +       | +      |
| Photoreceptor cell layer | ++  | ++++   | +       | +      |
| inner segment         | +    | ++++   | +       | +      |
| outer segment         | +    | ++++   | +       | +      |
| Pigmented epithelium  | ++   | +      | +++     | +      |
| Choroidesa            | +    | +      | +       | +      |
| Sclera                | +    | +      | +       | +      |
| Cornea                |      |        |         |        |
| Epithelium upper      | +    | +      | +       | +      |
| base                  | +++  | ++     | +++     | +      |
| Bowman's membrane     | +    | +      | +       | +      |
| Stroma                | +    | +      | +       | +      |
| Descemet's membrane   | +    | +      | +       | +      |
| Endothelium           | +    | +      | +       | +      |
| Lens                  |      |        |         |        |
| Subcapsular epithelium| +++  | ++++   | +++     | +++    |
| Lens fibers           | +++  | ++++   | +++     | +++    |
| Lens capsule          | n.d. | n.d.   | n.d.    | n.d.   |
| Iris                  |      |        |         |        |
| Stroma                | +    | +      | +       | +      |
| Pigmented epithelium  | +    | +      | +       | +      |
| Nonpigmented epithelium| +++ | +      | +++     | +      |
| Ciliary body          |      |        |         |        |
| Connective tissue     | +    | +      | +       | +      |
| Pigmented epithelium  | +    | +      | +       | +      |
| Nonpigmented epithelium| +++ | +      | +++     | +      |

Relative intensities were estimated by visual comparison of immunostained slides: n.d., not detected; +, low; ++, moderate; ++++, strong; ++++, very strong.

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