Thyrotropin and Obesity: Increased Adipose Triglyceride Content Through Glycerol-3-Phosphate Acyltransferase 3

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Epidemiological evidence indicates that thyrotropin (TSH) is positively correlated with the severity of obesity. However, the mechanism remains unclear. Here, we show that TSH promoted triglyceride (TG) synthesis in differentiated adipocytes in a thyroid hormone-independent manner. Mice with subclinical hypothyroidism, which is characterized by elevated serum TSH but not thyroid hormone levels, demonstrated a 35% increase in the total white adipose mass compared with their wild-type littermates. Interestingly, Tshr KO mice, which had normal thyroid hormone levels after thyroid hormone supplementation, resisted high-fat diet-induced obesity. TSH could directly induce the activity of glycerol-3-phosphate-acyltransferase 3 (GPAT3), the rate-limiting enzyme in TG synthesis, in differentiated 3T3-L1 adipocytes. However, following either the knockdown of Tshr and PPARγ or the constitutive activation of AMPK, the changes to TSH-triggered GPAT3 activity and adipogenesis disappeared. The over-expression of PPARγ or the expression of an AMPK dominant negative mutant reversed the TSH-induced changes. Thus, TSH acted as a previously unrecognized master regulator of adipogenesis, indicating that modification of the AMPK/PPARγ/GPAT3 axis via the TSH receptor might serve as a potential therapeutic target for obesity.

Obesity, the excess accumulation of white adipose tissue (WAT), is a major health hazard worldwide, and the epidemic incidence of obesity is increasing. Obesity is an established risk factor for metabolic diseases, including type 2 diabetes, hypertension, and nonalcoholic fatty liver disease1,2. Obesity-related metabolic diseases are accompanied by abnormal serum lipid parameters3, which are usually caused by higher TG levels and ectopic TG accumulation. WAT contains white adipocytes, which specialize in the storage of energy as triglycerides (TGs) and in the secretion of numerous adipokines that affect numerous aspects of metabolism4. The etiology of obesity is complicated, with both genetic and environmental factors influencing its development and susceptibility5. In this study, we identified a hormone involved in obesity.

Thyrotropin (thyroid stimulating hormone, TSH) is a hypophysial hormone, the major role of which is to stimulate thyrocyte proliferation, iodide uptake, hormonogenesis, and the release of thyroid hormones (T4 and T3). Numerous studies have confirmed the association between serum TSH levels and obesity. Epidemiological evidence has indicated a positive correlation between elevated serum TSH concentrations and body mass index (BMI) in euthyroid subjects6–11. In patients with metabolic syndrome and/or subclinical hypothyroidism (SCH, a type of thyroid disease accompanied only by elevated serum TSH levels), serum TSH levels are correlated with the severity of obesity12,13. Furthermore, obese people exhibit higher serum TSH levels than non-obese people14–16. However, the molecular mechanism by which TSH affects obesity has not been fully elucidated.

Expression of the TSH receptor (TSHR), once thought to be limited to thyrocytes, has been detected in numerous extrathyroidal tissues, including liver17 and adipose tissues18,19. Our previous study indicated that TSH promotes 3T3-L1 preadipocyte differentiation. Additionally, knocking down Tshr blocked the effects of TSH on preadipocyte differentiation20. Similar results were observed in rat preadipocytes21, human orbital preadipocyte fibroblasts22,23, and mouse embryonic stem cells23.

Obesity is a hypertrophic disease resulting from an increase in the number or size of individual adipocytes. We previously demonstrated that TSH could increase the number of adipocytes by promoting preadipocytes to differentiate into mature adipocytes20. The number of adipocytes is set during childhood and adolescence24.

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Adipocyte hypertrophy due to increased TG synthesis was recently shown to be the determinant of the development of adult obesity4,5. Brook showed an increase in adipose cell size among all obese subjects, but the total number of adipose cells was only increased in obese children and in adults who developed obesity during childhood6. Thus, adipocyte size is a major determinant of obesity in adults. Still, the role of TSH in regulating TG synthesis among differentiated adipocytes has not been completely established.

Glycerol-3-phosphate acyltransferase (GPAT) is the rate-limiting enzyme involved in TG synthesis27. GPAT3 is the major GPAT isoform expressed in adipocytes and plays a crucial role in adipogenesis28. The over-expression of GPAT3 in mammalian cells resulted in increased TG formation29, whereas the targeted knockdown of GPAT3 in 3T3-L1 cells significantly impaired adipogenesis30. Recent studies have indicated that the expression and activity of GPAT3 are regulated by insulin29, alcohol30, the glucocorticoid receptor31, and thiazolidinediones (peroxisome proliferator-activated receptor γ agonists)29. However, whether TSH affects the adipogenesis of differentiated adipocytes through GPAT3 has not been previously investigated. In the present study, we tested the hypothesis that TSH might upregulate GPAT3 expression, resulting in adipogenesis and obesity. This study provides a more comprehensive understanding of the pathophysiological effects of TSH on adipogenesis and suggests a novel interpretation of the mechanism that causes metabolic syndrome (MS) or SCH patients to develop obesity and obesity-related metabolic diseases.

Results

TSH regulates triglyceride content in adipose tissue. Based on the characteristic elevated serum TSH levels only in SCH, we first established the SCH mouse model. The SCH mice exhibited equal serum fT4 levels and higher TSH levels compared with their littermate controls. In addition, both the Epi-WAT TG content and serum TG levels were increased in SCH compared with those in control mice (Fig. 1C). Epididymal weight ratio revealed a 25% relative increase in SCH mice compared with their littermate controls. In both the Epi-WAT TG content and serum TG levels were increased in SCH mice. The serum concentration of adiponectin, an adipokine specifically secreted by adipocytes that is known to regulate glucomlipid metabolism, was also reduced.

To confirm the direct effect of TSH on adipocytes, we established the differentiated 3T3-L1 adipocyte model. TSH stimulation significantly increased the number of lipid droplets (Fig. 1E), and the differentiated 3T3-L1 adipocytes has not been completely established.

Through molecular screening, we noted that the level of GPAT3, the rate-limiting enzyme of TG synthesis in adipocytes34, was significantly decreased in Tshr−/− mice. Consistently, the GPAT3 protein levels were significantly decreased in the Epi-WAT of Tshr−/− mice (Fig. 2B).

However, after treating differentiated 3T3-L1 adipocytes with TSH, the GPAT3 mRNA levels were enhanced (Fig. 2C), and the protein levels increased in a dose- and time-dependent manner (Fig. 2D). Similar changes were observed in the primary adipocytes of mice and humans (Figs. S1A and B). Immunofluorescence analysis indicated that after TSH treatment, the increased GPAT3 protein was mainly located in the cytoplasm in a dose-dependent manner (Fig. 2E).

We next determined the response of the promoter region of GPAT3 to TSH. When GPAT3 promoter constructs linked to a luciferase reporter gene were transfected into HEK293 cells, the luciferase activity of the GPAT3 promoter sharply increased following TSH challenge in a dose-dependent manner (Fig. 2F).

To assess whether TSH induces adipogenesis through GPAT3, we transfected GPAT3 siRNA into differentiated 3T3-L1 adipocytes for 48 hours, followed by an additional incubation with TSH for 24 hours. GPAT3 silencing markedly attenuated TSH-induced adipogenesis. The Oil Red O staining was weak (Fig. 2G) and the TG content significantly dropped (Fig. 2H) relative to the group treated with only TSH.

PPARγ is indispensable in TSH-induced adipogenesis. We observed that the PPARγ mRNA and protein levels were decreased in the Tshr−/− mice in vivo (Fig. 2A and Fig. 3A). Consistently, in differentiated adipocytes in vitro, TSH promoted the expression and nuclear accumulation of PPARγ protein (Fig. 3B) in a dose- and time-dependent manner (Fig. 3C, Figs. S1A and B). Additionally, TSH promoted PPARγ mRNA expression (Fig. 3D).

To confirm the effect of TSH on PPARγ activity, HEK293 cells were cotransfected with a luciferase reporter gene construct (PPRE3-tk-luc; PPRE) and a PPARγ over-expression plasmid. As expected, the PPARγ inhibitor (GW9662) reduced the luciferase reporter activity to a greater extent than that observed in the control. In contrast, TSH alone elevated the reporter activity in a dose-dependent manner. However, preincubating the cells with GW9662 prevented this increase in activity (Fig. 3E).

To reconfirm that TSH-induced adipogenesis is mediated by PPARγ, we used RNA interference to block PPARγ expression in differentiated 3T3-L1 adipocytes. PPARγ siRNA significantly knocked down PPARγ mRNA expression (p < 0.05) (data not shown). TSH-stimulated adipogenesis and GPAT3 upregulation were significantly attenuated by PPARγ siRNA (Figs. 3F and G). In addition, van Beekum reported that the phosphorylation of PPARγ at Ser112 resulted in decreased transcriptional activity in the reporter assays35. After the cells were transfected with the PPARγ siRNA plasmid (Ser112 mutated to alanine), the regulation of GPAT3 by TSH was enhanced (Fig. 3H).
PPARγ inhibitor ameliorates high-fat diet (HFD)-induced obesity in vivo. To determine whether PPARγ is the upstream factor involved in the effects of GPAT3 on HFD-induced obesity in vivo, GW9662-administered mice were used. HFD-fed C57/BL6 mice (13 weeks old) were supplemented with or without GW9662 for 5 weeks. Compared with vehicle, GW9662 ameliorated the HFD-induced increase in weight, Epi-WAT index and adipocyte size (Figs. 4A and B). Furthermore, GW9662 inhibited GPAT3 expression (Fig. 4C), decreased intracellular and plasma TG levels (Fig. 4D and Fig. S2A) and suppressed adipose TG synthesis in vivo. In addition, the function of the adipose tissue was changed due to the suppression of adiponectin release by GW9662 (Fig. 4D). These results indicated that PPARγ is essential for the regulation of TG synthesis via TSH.

TSH-stimulated adipogenesis in differentiated adipocytes is dependent on AMPK. As an energy sensor in metabolic tissues, AMP-activated protein kinase (AMPK) plays an important role in the regulation of PPARγ activity in adipocytes. In Tshr^−/−^ mice, AMPK Thr172 phosphorylation was increased compared with that in Tshr^+/+^ mice (Fig. 5A); these effects have been reported to correlate with kinase activity. Meanwhile, the key downstream targets of AMPK (i.e., SREBP-1c, FAS, SCD, and ACC) were reduced in Tshr^−/−^ mice (Fig. 2A). Based on these changes, we
hypothesized that the functions of TSH are activated in association with AMPK.

As shown in Fig. 5B (Figs. S1A and B), the expression of phosphorylated AMPK decreased in mature adipocytes in vitro in a dose- and time-dependent manner following TSH treatment. Similarly, AMPK activity was inhibited by TSH according to a SAMS peptide phosphorylation assay (Fig. 5C).

To determine whether TSH functions by modulating AMPK activity, we transfected differentiated 3T3-L1 adipocytes with either a constitutively active form of AMPK (CA-AMPK) to increase their AMPK activity or a dominant negative mutant of AMPK (DN-AMPK) to reduce their AMPK activity. Transfection with CA-AMPK significantly ameliorated TSH-stimulated TG and lipid droplet production, and transfection with DN-AMPK enhanced the effects of TSH (Fig. 5D and Fig. S3).

In addition, TSH-stimulated PPARγ and GPAT3 upregulation were significantly inhibited by transfection with CA-AMPK but were enhanced by transfection with DN-AMPK (Figs. 5E and F). This finding suggests that TSH stimulates PPARγ and GPAT3 expression via AMPK in vitro.

Thus, we analyzed whether TSH induces adipogenesis through AMPK in vivo by chronically treating mice with AICAR. As shown

Figure 2 | TSH promotes adipocyte GPAT3 activity, leading to elevated TG synthesis. Homozygous Tshr-knockout mice (Tshr−/−) were supplemented with T4 (serum T4 levels were similar to that in the wild type mice, \( p > 0.05 \)) to avoid the effects of the thyroid hormones. Tshr−/− mice were used as controls. (A) RT-PCR analysis of adipogenesis-related gene expression in the Epi-WAT. * \( p < 0.05 \) compared with Tshr−/− mice. (B) The expression of GPAT3 protein was detected by western blot. (C) RT-PCR analysis of GPAT3 mRNA in TSH (2 \( \mu \)M, for 48 h)-treated differentiated 3T3-L1 cells. * \( p < 0.05 \) compared with control. (D) The time-dependent and dose-dependent effects of TSH on GPAT3 expression in differentiated 3T3-L1 cells. (E) Immunofluorescence staining of GPAT3 in differentiated 3T3-L1 cells treated with the indicated concentrations of TSH for 24 h. The blue color depicts the DAPI-stained nuclei, and the red shows the GPAT3 protein (TRITC-conjugated). (F) HEK293 cells were transfected with GPAT3 luciferase reporter construct vectors for 48 h, then treated with vehicle or TSH (1 or 2 \( \mu \)M) for an additional 24 h. GPAT3 activity was detected by dual luciferase reporters. * \( p < 0.05 \) compared with control; * \( p < 0.05 \) compared with 1 \( \mu \)M TSH at 24 h. (G–H) RNAi of GPAT3 inhibited TSH-induced TG synthesis. After transfection with non-targeting or GPAT3 siRNA for 48 h, the differentiated 3T3-L1 cells were incubated with TSH (2 \( \mu \)M) or vehicle for another 24 h. (G) Lipid droplet contents and (H) intracellular TG contents were assayed. The results are expressed as the mean ± SD. # \( p < 0.05 \) compared with the non-targeting siRNA group; * \( p < 0.05 \) compared with TSH group. All panels above are representative of 3 independent experiments. All the gels were run under the same experimental conditions, and key data cropped blots are used here. The full-length gel images are available in Supplementary Fig. 6.
in Fig. 5G, chronic exposure to AICAR increased AMPK phosphorylation. Moreover, a significant decrease in PPARγ, GPAT3 mRNA and protein expression occurred in the Epi-WAT of AICAR-treated Tshr1/1 and Tshr2/2 mice (Figs. 5G and H).

**Tshr ablation ameliorates HFD-induced obesity in vivo.** To address whether TSH-stimulated adipogenesis depended on TSHR, we transfected 3T3-L1 adipocytes with Tshr siRNA and found that Tshr mRNA expression was significantly reduced. Compared with the non-targeting siRNA, Tshr siRNA inhibited the effects of TSH on the regulation of TG levels and lipid droplet content (Figs. 6A and B). Furthermore, Tshr siRNA inhibited the effects of TSH on the upregulation of PPARγ and GPAT3 expression, although AMPK phosphorylation increased (Fig. 6C).

For the *in vivo* study, we fed Tshr−/− and Tshr+/+ mice (8 weeks old) a HFD for 10 weeks, and measured relevant obesity-related parameters. Although the body lengths and food intake of Tshr−/− mice were similar to those of the Tshr+/+ mice (data not shown), the body weights of mice on the HFD (30.86 ± 1.03 g) were 23% less than those of the Tshr+/+ mice (39.41 ± 1.99 g) at 18 weeks of age (Fig. 6D). In Tshr−/− mice, due to the absence of TSHR expression, the Epi-, retroperitoneal-adipose mass and adipocyte size were sig-
GW9662 at a dose of 0.35 mg/kg per day in their drinking water for 5 weeks (GW9662 group, n = 8) or to a corresponding volume of vehicle (control group, n = 8). (A) Increases in body fat were suppressed by GW9662 treatment, and the adipose mass of Epi fat, retroperitoneal fat (RP fat) and adipocyte size were diminished in the GW9662 group. (B) Epi-WAT weight, WAT index, weight at 18 weeks and weight changes were measured. *p < 0.05 compared with control group. (C) The protein expression of GPAT3 was measured using western blots. The gels were run under the same experimental conditions, and cropped blots are used here. The full-length gel images are available in Supplementary Fig. 6. (D) Intracellular TG levels and plasma adiponectin were detected, respectively. The results are expressed as the mean ± SD. *p < 0.05 compared with control group. All of the above panels are representative of 3 independent experiments.

Figure 4 | Antagonism of PPARγ prevents high-fat-diet-induced obesity in vivo. High-fat-diet (HFD) chowed C57/BL6 mice were subjected to GW9662 at a dose of 0.35 mg/kg per day in their drinking water for 5 weeks (GW9662 group, n = 8) or to a corresponding volume of vehicle (control group, n = 8). (A) Increases in body fat were suppressed by GW9662 treatment, and the adipose mass of Epi fat, retroperitoneal fat (RP fat) and adipocyte size were diminished in the GW9662 group. (B) Epi-WAT weight, WAT index, weight at 18 weeks and weight changes were measured. *p < 0.05 compared with control group. (C) The protein expression of GPAT3 was measured using western blots. The gels were run under the same experimental conditions, and cropped blots are used here. The full-length gel images are available in Supplementary Fig. 6. (D) Intracellular TG levels and plasma adiponectin were detected, respectively. The results are expressed as the mean ± SD. *p < 0.05 compared with control group. All of the above panels are representative of 3 independent experiments.

Discussion

Here, we discovered a novel role of TSH in regulating adipogenesis and obesity. TSH increased TG accumulation via TSHR, in which GPAT3 activation is induced by the AMPK/PPARγ signaling pathway. SCH mice manifested obesity, whereas the ablation of Tshr in mice caused resistance to high fat diet-induced obesity. Our findings reveal a novel effect of TSH on TG metabolism in adipose tissue and might partially explain the pathogenesis of obesity in SCH patients.

Adipose tissue serves as a regulator of various physiological pathways and is the key tissue involved in the regulation of energy balance and lipid homeostasis. Previous studies have reported that human, mouse, and rat preadipocytes differentiation is accompanied by TSHR expression. Toyoshi cloned Tshr cDNA from the white adipose tissue of rats and determined that the Tshr cDNA from adipose tissue was almost identical to that from the thyroid glands. The expression levels of TSHR in the epididymal adipose tissue and the thyroid were similar. We previously demonstrated that TSHR expression increases as 3T3-L1 preadipocytes differentiate and that TSHR knockdown delays adipocyte differentiation. All of these studies have focused on the effects of TSH on the adipocyte differentiation. However, whether TSH regulates TG synthesis in differentiated adipocyte had never been explored.

The functions of differentiated adipocytes include lipid storage, energy flow, and the production of adipokines. The hypertrophy of differentiated adipocytes results in dysfunction and induces an inflammatory state and abnormal glucolipid metabolism, in which altered adipokine secretions occur. Moreover, mature adipocytes are the most important form in human adipose tissues. In the present study, we observed that the expression of a panel of genes known to play a critical role in adipogenesis was decreased in Tshr−/− mice. Therefore, we believe that deciphering the regulation of this process could provide new insight into the regulation of conventional adipogenesis.

The expression of TSHR in adipocytes is not constant. Thus, it is inappropriate to use the differentiation process within the 3T3-L1 preadipocytes model to study the effect of TSH on adipogenesis. To determine the functional role of TSH in adipocytes, we studied 3T3-L1 adipocytes that were cultured for 16 days following the induction of differentiation. 3T3-L1 cells were the first non-thyroidal cell line reported to express TSHR, and TSHR expression can also be induced in these cells. 3T3-L1 cells subjected to long-term culture are considered to be representative of mature adipocytes, in which the expression of proteins linked to adipocyte differentiation, including TSHR, does not significantly change. Furthermore, TSHR expression remains unchanged in TSH-stimulated differentiated adipocytes. In addition, using primary fat cells provides a suitable system for exploring fat cell biology and numerous regulatory mechanisms at the cellular level.

GPAT has been suggested to be the rate-limiting enzyme in the TG biosynthesis pathway because of its low specific activity and its position as the initiating enzyme. Although GPAT catalyzes the con-
Figure 5 | TSH-stimulated adipogenesis was dependent on AMPK. (A) Phosphorylated protein at Thr172 of AMPKα subtype (p-AMPK) was detected by western blot in the Epi-WAT of Tshr−/− and Tshr+/+ mice. (B) The time-dependent and dose-dependent effects of TSH on p-AMPK protein expression in differentiated 3T3-L1 cells. (C) After cells were incubated with TSH (1 μM), AICAR (1 mM) or vehicle for 24 h, the differentiated 3T3-L1 cells underwent AMPK activity assays with SAMS peptides. *p < 0.05 compared with control; **p < 0.05 compared with TSH group. (D–F) Differentiated 3T3-L1 cells were transfected with constitutively activated AMPK (CA-AMPK) or dominant negative mutant of AMPK (DN-AMPK) plasmids for 48 h and then stimulated with or without 1 μM TSH for another 24 h. (D) Intracellular TG contents were detected. *p < 0.05 compared with control; **p < 0.05 compared with TSH group; ***p < 0.05 compared with CA-AMPK group; ****p < 0.05 compared with CA-AMPK + TSH group. GPAT3 and PPARγ expression and location were detected by (E) western blotting or (F) immunofluorescent staining, respectively. (G–H) Tshr−/− or Tshr+/+ mice were injected intraperitoneally with AICAR (n = 5, respectively) or PBS (n = 5, respectively) for 3 weeks. (G) The p-AMPK, total AMPK, GPAT3 and PPARγ protein were detected by western blot. (H) GPAT3 and PPARγ mRNA levels were detected by RT-PCR. *p < 0.05 compared with Tshr−/− + AICAR group; **p < 0.05 compared with Tshr−/− + Vehicle group. All of the panels above are representative of 3 independent experiments. All the gels were run under the same experimental conditions, and key data cropped blots are used here. The full-length gel images are available in Supplementary Fig. 6.
version of glycerol-3-phosphate to lysophosphatidic acid, Igal reported that GPAT directed the incorporation of fatty acids into TGs. GPAT3 is the major GPAT isoform in adipocytes and plays a crucial role in adipogenesis and TG storage. Because TGs are primarily stored in adipose tissue, we evaluated the TG contents and adipocyte sizes in vivo, as well as intracellular TG and lipid droplets contents in vitro, to estimate TG synthesis. These results illustrated that the changes in TG content were consistent with the shifts in GPAT3 expression and activity. Furthermore, we used TSH stimulation, siRNA-mediated depletion of TSHR, SCH and Tshr2/2 mice models to show that TSH specifically promoted GPAT3 expression and TG synthesis. In the present study, we demonstrated...
for the first time that TSH regulates GPAT3 expression and activity. Based on our previous study and this current work, our results strongly suggest that TSH promotes TG accumulation, both by increasing adipocyte differentiation and by augmenting TG synthesis in differentiated adipocytes, resulting in the development of obesity and metabolic-related disorders.

In this study, we found that TSH promotes adipocyte hypertrophy by countering the inhibitory effect of AMPK on PPARγ/GPAT3 signaling. AMPK is directly regulated by cAMP/PKA and functions by inhibiting fatty acid synthesis, adipogenesis and activating fatty acid oxidation. Numerous studies have established that TSH is involved in lipid metabolism through the cAMP/ PKA or PI3K/Akt pathway, but the role of TSH in the AMPK pathway was not fully elucidated. A better understanding of the molecular mechanisms by which TSH regulates AMPK activity in adipose tissues may provide new insights into the global modulation of adipogenesis. Our present data indicate that AMPK acts as the pivot in TSH-induced adipogenesis, as shown by the following results: 1) TSH functions as a potent suppressor of AMPK activity according to the SAMs peptide phosphorylation assay; 2) the selective PKA inhibitor, H89, activated AMPK in a dose- and time-dependent manner in C57/BL6 mice (Fig. S5); 3) CA-AMPK inhibited the nuclear accumulation of PPARγ, suppressed GPAT3 expression and resulted in reduced intracellular TG contents; 4) transfection with DN-AMPK resulted in diametrically opposed effects were observed; and 5) the potency of the proadipogenic effect of TSH was shown to be highly dependent on AMPK.

The PPARs are transcription factors that regulate diverse physiological and pathological processes, especially glucolipid metabolism. PPARγ is the most important PPAR species in adipose tissue; it is essential for the differentiation and function of white adipocytes and promotes lipid accumulation. Previous studies consider PPARγ to be involved in lipid metabolism and adipogenesis. However, recent studies have shown that PPARγ is essential for the survival of mature adipocytes. In addition, adipose-specific PPARγ-knockout mice displayed diminished TG stores in WAT. Our results showed that PPARγ could increase GPAT3 expression through its involvement in TSH-stimulated TG synthesis. Meanwhile, the inhibition of PPARγ promotes the expression of the adipokine adiponectin. Therefore, we believe that PPARγ plays an important role in regulating the function and adipogenesis of mature adipocytes, not only in adipocyte differentiation. The characterization of PPARγ, a potential regulator in mature adipocytes, could open the door for novel approaches for the treatment of obesity and other disorders of mature adipocytes.

Thyroid hormones should be considered as important confounding factors when discussing the relationship between TSH and lipid metabolism. In our study, we used Tshr−/+ mice to evaluate the effects of TSH on lipid profiles, independent of the thyroid hormone levels. Because the deletion of TSHR can cause TSH malfunction, Tshr−/− mice were compared with Tshr+/− mice to investigate the function of TSH in vivo. Meanwhile, to study the function of TSH in vitro, we stimulated differentiated 3T3-L1 adipocytes with TSH or the vehicle, further used RNAi Tshr. Through the above-mentioned approaches, we excluded the role of thyroid hormones both in vivo and in vitro, confirming the relationship between TSH and TG synthesis in adipocytes. Although strong clinical and experimental evidence supports the pivotal role of TSH in the regulation of metabolism, the molecular mechanisms involved in these actions remain largely unknown. In this study, we demonstrated that TSH, by binding to TSHR, inhibits AMPK Thr172 phosphorylation, activates the PPARγ transcriptional program, and subsequently increases GPAT3 expression in mature adipocytes. In summary, our data indicate that TSH is linked with the promotion of adipocyte hypertrophy, thus contributing to the pathogenesis of obesity. The modification of the AMPK/PPARγ/GPAT3 axis via TSH might therefore serve as a potential therapeutic strategy for obesity-associated metabolic syndrome and SCH.

Methods
Primary adipocytes culture. The procedure to obtain primary normal human and mice adipocytes has been carefully standardized. Human primary adipocytes were isolated from subcutaneous adipose tissue of male patients from Shandong Provincial Hospital Affiliated to Shandong University. Mice primary adipocytes were isolated from the epididymal fat pads of normal male C57/BL6 mice. In brief, excised adipose tissues were digested in D-hanks' buffer with 1 mg/ml collagenase type I, 100 mg/ml penicillin and 100 mg/ml streptomycin for 30 min at 37.8°C. Subsequently, the cell suspension was filtered sequentially through 100 mm nylon mesh and centrifuged at 200 g for 10 min. In the laminar flow hood, remove and discard the underlying pellet (containing preadipocytes, fibroblasts, and erythrocytes) and media. Resuspend the remaining fat layer in 20 ml D-hanks' and centrifuge for 10 min at 200 g. Repeat this step two additional times. After the last centrifugation step, transfer the fatty layer (containing the mature adipocytes) to 35 mm cell culture dish. Dishes were washed with DMEM/F12 + 10% FBS. The culture medium was replaced with fresh medium every 48 h. All the patients were properly informed, and written informed consent was obtained from each individual. Our research was approved by the Ethics Committee of Shandong Provincial Hospital (Jinan, China). All animal experiments were performed according to the Shandong Provincial Hospital Animal Care and Use Committee (Jinan, China).

Animals and treatment. All the mice were kept in a SPF room in which lighting was controlled (12 hours on, 12 hours off), and the temperature was maintained at 23°C. SCH mice. Male C57/BL6 mice (8 weeks old) were obtained from Vital River Corporation (Beijing, China). Mice were divided into two groups: one group were given methimazole, a drug that inhibits thyroid hormone synthesis (MMI, 0.04 mg/kg BW) in their drinking water for 5 weeks (GW9662 group, n = 8); and the second group received solvent only (control group, n = 8). MMI was administered for 12 weeks, the mice were fasted for 6 h and then euthanized using pentobarbital sodium. Serum samples were collected immediately prior to sacrificing the mice and were tested for T4 and TSH levels.

GW9662 treated mice. High-fat-diet (HFD) chowed male C57/BL6 mice (13 weeks old) were subjected to GW9662 at a dose of 0.35 mg/kg BW in their drinking water for 5 weeks (GW9662 group, n = 8) or to a corresponding volume of vehicle (control group, n = 8).

HFD chowed Tshr−/− and Tshr+/− mice. Male Tshr−/− and Tshr+/− mice were obtained from Jackson Laboratory (USA). All the information of these mice could get from our previous research. Tshr−/− (n = 8) and Tshr+/− (n = 8) mice (8 weeks old) fed HFD for 10 weeks.

H89 treated mice. Male C57/BL6 mice (8 weeks old) were gavaged with H89 at a dose of 20 nmol for 3 or 5 days (n = 3, respectively), or 40 nmol for 3 days (n = 3), or with a corresponding volume of vehicle (n = 3).

AICAR treated Tshr−/− and Tshr+/− mice. Male Tshr−/− and Tshr+/− (8 weeks old) mice were injected intraperitoneally with AICAR at a dose of 0.5 μg/kg BW·d for 3 weeks (n = 5, respectively), or with a corresponding volume of vehicle (n = 5, respectively).

For each animal experiment, Epi-WAT weight, Epi-WAT weight/body weight, Epi-WAT triglyceride contents were measured. Blood from animals was then obtained for analyses of serum TG and adiponectin (SCH mice also detected BMI, f4 and TSH levels). In addition, adipose tissue from all animals were collected and immediately frozen for assay.

AMPK activity assay. AMPK activity was measured after immunoprecipitation with polyclonal rabbit AMPK α1/2 antibodies raised against synthetic peptides of AMPK. AMPK activity was measured using the SAMS peptide phosphorylation assay. Briefly, the kinase assay was performed in 30 μl kinase buffer containing 200 μM SAMS peptide (HRMRSMSGLHVKKR, Upstate Technology), 200 μM AMP, 5 mM MgCl2, 200 μM ATP, and 10 μCi [γ-32P]ATP (specific activity 3,000 Ci/mmol) at 30°C for 15 min with continuous shaking. 20 μl aliquots of the reaction mixture were spotted on half-cut P81 Whatman phosphocellulose paper. The P81 paper was washed three times for 15 min in 1% phosphoric acid with gentle stirring to remove free ATP and washed once with acetone for 5 min. The papers air-dried and radioactivity was counted. 32P-labeled incorporation into SAMS peptide in 4 ml of scintillation liquid by scintillation counting. Control immunoprecipitation that contained no AMPK antibody was performed to ensure that SAMS peptide phosphorylation was dependent on specific immunoprecipitation of AMPK. AMPK activity was calculated as picomoles per minute per milligram of protein.

Materials. 3T3-L1 cells culture, Oil Red O staining, Triglyceride assay, H&E staining, Adiponectin assay, Cell transfection, Knockdown of Tshr, PPARγ or
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**Author contributions**
The work presented here was carried out in collaboration between all authors. S.M. performed the experiments, S.M. and F.J. wrote the main manuscript text, J.Z., Q.G. and L.G. designed the experiments, L.Z. prepared all the animals, C.X. and Y.L. prepared figures 1–3, Y.S. and C.Y. prepared figures 4–6. D.J. prepared supplemental figures. All authors reviewed the manuscript.

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