Letter to the Editor

Joint infection due to Elizabethkingia miricola

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Sir,

Elizabethkingia miricola is a non-fermenting Gram-negative rod, non-motile and non-spore-forming which was firstly described in 2003 when it was isolated from condensation water on the space station MIR [1]. Initially named Chryseobacterium miricola, it was reclassified along with Chryseobacterium meningosepticum into the new genus Elizabethkingia, this was due to the phylogenetic analysis, based on the sequencing of the 16S rRNA gene [2]. E. miricola has been demonstrated to be pathogenic, with reports of bacteremia, sepsis and pulmonary abscesses [3-5]. To our best knowledge, we report the first case of infection in a native joint due to E. miricola.

A 52-years-old immunocompetent woman came to the Emergency Department of our hospital due to non-favorable evolution of a catastrophic right foot caused by a traffic accident. The diagnosis was made by means of a foot x-ray and it was observed that the affected bones were the calcaneus, talus, scaphoid and cuboid. Patient refers poor evolution of skin injuries during a month with pain that has slightly subsided. The clinical history was unremarkable and no underlying diseases were reported. No antibiotic treatment during that month was administered, and only wound cures every two days were performed. Due to the infectious aspect of the right ankle, a surgical procedure with extensive debridement was then carried out; also, five bone biopsies from different sites were taken and sent to the microbiology laboratory for culture. The patient did not have indwelling devices or invasive catheters before infection. Considering the clinical and pathologic aspects, the patient was definitively discharged.

Direct Gram staining of the samples showed abundant Gram-negative bacilli, and on the first day of incubation growth of grey colonies in pure culture was observed on all plates above mentioned. The oxidase test was positive (Oxidase Reagent Droppers, Becton Dickinson). The microorganism was identified by mass spectrometry (MALDI-TOF MS, Bruker Biotyper, Billerica, MA, USA) as E. miricola (Log score 2.3). The MIC of different antibiotics was carried out by the MicroScan device (Beckman Coulter). According to the breakpoints of CLSI for non-fermenting Gram-negative rods, E. miricola was susceptible to ciprofloxacin (MIC 0.38 mg/L), levofloxacin (MIC 0.25 mg/L), piperacillin-tazobactam (MIC 16 mg/L) and resistant to amikacin (MIC>256 mg/L), cefepime (MIC>256 mg/L), ceftazidime (MIC>256 mg/L), colistin (MIC>256 mg/L), fosfomycin (MIC>256 mg/L), gentamicin (MIC 32 mg/L), imipenem (MIC>32 mg/L), meropenem (MIC>32 mg/L) and tetracycline (MIC >24 mg/L). MIC to tigecycline was 0.5 mg/L. At this stage, antimicrobial treatment was started with piperacillin-tazobactam 4gr/8h i.v, but after 15 days of treatment the patient had poor renal function without a known cause, and it was decided to change the treatment to oral levofloxacin 500 mg/24h for three weeks. The patient presented good evolution and after going to successive consultations, 4 months later she was definitively discharged.

At this time, the most common microorganisms causing bone infection are staphylococci or Gram-negative bacilli, including Pseudomonas aeruginosa. From cultures of intraoperative specimens, Staphylococcus aureus is the main causative agent of chronic bacterial osteomyelitis, accounting for about two thirds of isolates, followed by Pseudomonas and Enterobacterales [6]. However, other rare microorganisms could be implicated in the etiology of these infections, as in our case, so physicians and microbiologist should be aware about this possibility.
The first case of human infection due to *E. miricola* was reported in 2008 in an adult with mantle cell carcinoma who underwent stem cell transplantation [3]. After this, *E. miricola* has demonstrated to cause bacteremia, sepsis, and pulmonary and tract urinary infections [3-5, 7]. From these patients, three of them had underlying comorbidities such as cancer, alcoholic pancreatitis and cystic fibrosis. In all cases, the isolate was identified by MALDI-TOF MS. Thus, the recent introduction of mass spectrometry for routine identification in the clinical laboratories may help to identify some rare pathogens and to know the true incidence of infections due to these microorganisms.

Currently, there are no CLSI/EUCAST guidelines for *E. miricola*. This bacterium has been found to be multidrug resistant, similar to *E. meningoseptica* which is known to harbor β-lactamases showing resistance to β-lactams and carbapenems. *E. miricola* isolates have been found to be resistant to many antibiotics. A study showed resistance to imipenem, ceftazidime, cotrimoxazole and variable susceptibility to quinolones [8]. In a recent study, all isolates of *E. miricola* were susceptible to tetracyclines and piperacillin-tazobactam. However, 50% of the isolates were susceptible to levofloxacin and tigecycline [9]. Another study showed that 91% and 77% of *Elizabethkingia* spp isolates were resistant to ciprofloxacin and levofloxacin, respectively [10]. The most prevalent alterations were two single mutations in GyrA, Ser83Le and Ser83Arg. In our case, however, the isolate was susceptible to both levofloxacin and ciprofloxacin.

In summary, we here presented the first case of bone infection due to *E. miricola*. Until now, the presence of this pathogen is rare as cause of human infections, but the recent introduction of MALDI-TOF MS can help to identify some microorganisms, as in our case, which rarely produces bone infections. On the other hand, susceptibility to these isolates should be performed due to the fact that several species often exhibit extensive antimicrobial resistance.

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**CONFLICTS OF INTEREST**

The authors declare that there are no conflicts of interest.

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