Comparative analysis of karyotypes of *Chironomus solitus* Linevich & Erbaeva, 1971 and *Chironomus anthracinus* Zetterstedt, 1860 (Diptera, Chironomidae) from East Siberia

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**Academic editor:** V. Golgina | Received 9 February 2015 | Accepted 17 April 2015 | Published 3 June 2015

**Citation:** Proviz V (2015) Comparative analysis of karyotypes of *Chironomus solitus* Linevich & Erbaeva, 1971 and *Chironomus anthracinus* Zetterstedt, 1860 (Diptera, Chironomidae) from East Siberia. Comparative Cytogenetics 9(2): 237–248. doi: 10.3897/CompCytogen.v9i2.4702

**Abstract**

A comparative chromosome banding analysis of *Chironomus solitus* Linevich & Erbaeva, 1971 and *Chironomus anthracinus* Zetterstedt, 1860 from East Siberia (Lakes Baikal, Gusinoe, Arakhley and Irkutsk Reservoir) showed close similarity of banding sequences. *Ch. solitus* differs from *Ch. anthracinus* in one species-specific sequence of arm B. Arms C (43%) and D (30%) had inversion banding sequences previously reported in *Ch. anthracinus*. The similarity of karyotypic features of *Ch. solitus* and *Ch. anthracinus* in combination with morphological features of larvae provide evidence in favour of including *Ch. solitus* in the *C. anthracinus* group of sibling species long with *Ch. reservatus* Shobanov, 1997.

**Keywords**

Karyotype, banding sequences, inversion, *Chironomus solitus*, *Chironomus anthracinus*

**Introduction**

*Chironomus solitus* Linevich & Erbaeva, 1971 and *Chironomus anthracinus* Zetterstedt, 1860 are abundant chironomid species (Diptera: Chironomidae), inhabiting the silty bottoms of various water bodies in Pribaikalye and Zabaikalye. *Ch. solitus* was first
registered in the Irkutsk Reservoir as well as in the Angara River, Bratsk, Ust-Ilimsk water reservoirs (Linevich and Erbaeva 1971; Linevich 1981; Proviz et al. 1991; Erbaeva and Safronov 2009), in lakes and rivers of the Barguzin River basin (Buyantuev 1999) and recently encountered in the near-shore zone of Lake Baikal. *Ch. anthracinus* is a widespread Holarctic species known from the Angara River and its tributaries, Irkutsk Reservoir, lakes of Western Zabaikal (Linevich and Erbaeva 1971; Linevich 1981; Kiknadze et al. 2005) and the basin of the Barguzin River (Buyantuev 1999).

*Ch. solitus* and *Ch. anthracinus* live in the single type water environments (lakes, water reservoirs), and are characterized by similar larval morphology in the features used in the distinction of *Chironomus* species, which makes their differentiation complicated. Thus, accurate identification of these species requires analysis of their karyotypes, rather than only external larval morphology. Until recently, the *Ch. solitus* karyotype had only been examined in one population from the Irkutsk Reservoir. The first data were reported by Bukhteeva (1979); later, banding chromosome patterns and polymorphisms were described (Proviz 2009). Karyological analysis was made of *Ch. anthracinus* from many Palearctic and Nearctic regions (Belyanina 1983; Kiknadze et al. 1991, 1996; Shobanov 1996; Petrova and Rakisheva 2003; Kiknadze et al. 2005). In East Siberia, karyotypes of larvae from Lake Shchuchie (Buryatia) were briefly reported by Bukhteeva (1979, 1980). Later, Kiknadze and co-authors (Kiknadze et al. 1991, 1996) described the karyotypic of *Ch. anthracinus* from the Vilyuy Reservoir (Yakutia).

The present work is aimed at comparative analysis of *Ch. solitus* and *Ch. anthracinus* karyotypes from the largest lakes of East Siberia, Baikal, Gusinoe, Arakhley and Irkutsk Reservoir, and determination of cytogenetic features for their identification.

**Material and methods**

Fourth instar larvae of *Ch. solitus* were collected in January 1992 in the Irkutsk Reservoir (depth 3 m, 52 larvae), and in June 2008 in Lake Baikal opposite the Bolshye Koty Settlement (depth 6 m, 12 larvae). *Ch. anthracinus* were collected in May 2013 in Lake Gusinoe (10–22 m, 65 larvae), and in March 2014 in Lake Arakhley (10–17 m, 78 larvae). Larvae were fixed in a 3:1 mixture of 96% ethanol and glacial acetic acid. Karyological preparations were made using the ethyl-orcein method (Demin and Shobanov 1990). In 1992 and 2008, chromosomes were photographed by a micro-camera unit MCU-1 with 90x zoom magnification; in 2013–2014, this was performed using an Axiostar plus (Zeiss) microscope (Centre for Microscopic Analysis LIN SB RAS) with AxioVision Rel. 4.7.1 software. Mapping of arms A, C, D, E, and F of *Ch. anthracinus* chromosomes was performed according to Kiknadze et al. (2005) based on piger-standard (Keyl 1962, Devai et al. 1989), while standard map of *Ch. plumosus* suggested by Shobanov (1994, 1996) was used for mapping of arm B. Symbols designating banding sequences are as follow: distribution areas marked by p’ for Palearctic,
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n’ for Nearctic, and h’ for Holarctic zoogeographical regions (Kiknadze et al. 2005) and followed by abbreviated species name (sol), arm designation (A) and banding sequence number–p’solx1 (in homozygote–p’solA1.1).

**Results**

**Larval morphology**

Both species have a light yellow (from the dorsal part) cephalic capsule, including the frontal sclerite. Abdominal segment VIII bears two pairs of long ventral appendages; lateral appendages on segment VII are absent (bathophilus type after: Lenz 1926). Premandible with two uneven teeth. Fourth lateral cusp of mentum is smaller than fifth cusp. Third antennal segment is shorter than the fourth. The colour of the fourth lower mandibular tooth varies; that of *Ch. solitus* is dark yellow, while the remaining teeth are dark brown. The results of our examination of the population from Lake Arakhley showed that *Ch. anthracinus* tooth was either dark yellow or of the same colour as the rest of the teeth.

**Karyotype characteristics**

Karyotypes of *Ch. solitus* (Fig. 1) and *Ch. anthracinus* (Fig. 2) have common morphological features: 2n=8. A combination of chromosome arms is typical for species from “thummi” cytocomplex. Chromosomes AB and CD are metacentric, EF, submetacentric, and G, telocentric. The species differ in the size of cenromeric heterochromatin. The centromeric areas of *Ch. solitus* are well defined, and the centromeres of *Ch. anthracinus* look like thin disks. Arm G homologues are unconjugated and carry a Balbiani Ring (BR) and a nucleolus (N). In *Ch. anthracinus* there is a second nucleolus in the arm F.

**Banding sequences**

**Arms A** of *Ch. anthracinus* and *Ch. solitus* are monomorphic with a single identical banding sequence h’antA1=h’solA1 (Fig. 3, a, b): h’antA1= h’solA1 1a-2c 10a-12a 13ba 4a-c 2g-d 9e-4d 2h-3i 12cb 13c-19f C

**Arms B** of *Ch. anthracinus* and *Ch. solitus* are monomorphic with banding sequences h’antB1 (Fig. 3, c) and p’solB1 (Fig. 3, d) differ by a simple inversion:

h’antB1 25s-24i 18c-16b 22b-21a 23l-24h 18d-20n 23k-d 15m-16a 22c-23c 15l-12v C

p’solB1 25s-24i 18c-16b 18d 24h-23l 21a-22b 19a-20n 23k-d 15m-16a 22c-23c 15l-12v C
Figure 1. Karyotype of *Chironomus solitus*. h’solA1.1, p’solB1.1 etc.—genotypic combinations of banding sequences in chromosomal arms; N – nucleolus; BR – Balbiani Ring; p – puff, arrows show centromeric bands.

Figure 2. Karyotype of *Chironomus anthracinus*. The designations are the same as in Fig. 1.
Figure 3. Homozygous banding sequences in the arms A and B of Chironomus anthracinus and Chironomus solitus. a h’antA1.1 b h’solA1.1 c h’antB1.1 d p’solB1.1. Numbers and small letters under chromosome arm correspond to banding sequences, brackets near chromosome arms show inversions.
In addition to the inversion, *Ch. solitus* differs from *Ch. anthracinus* by the presence of a puff in the region 17. The banding sequence h’antB2 (Kiknadze et al. 2005) were found in Palearctic and Nearctic *Ch. anthracinus* populations. The borders of this inversion located close to the borders of inversion that differ banding sequence p’solB1 from h’antB1. Standard mapping of *Ch. plumosus* (Shobanov, 1994) allows it to be represented as follows:

h’antB2 25s-24i 18c-17a 23l 21a-22b 16b 24h 18d 19a-20n 23k-d 15m-16a 22c-23c 15l-12v C

**Arm C** of *Ch. anthracinus* is monomorphic, with a single banding sequence h’antC1 (Fig. 4, a). Arm C of *Ch. solitus* is polymorphic and has two banding sequences—p’solC1 (Fig. 4, b) and h’solC2—differing by one simple inversion (Fig. 4, c). Inversion heterozygotes p’solC1.h’solC2 made up 25% and 43% of Baikal and Irkutsk Reservoir populations, respectively. The same banding sequences, h’antC1 (=h’solC2) and p’antC2 (=p’solC1), were also registered in *Ch. anthracinus* populations from other localities within this area, although in somewhat different proportions:

h’antC1= h’solC2 1a-2c 2d-6b 11c-8a 15ed 15c-11d 6gh 17a-16a 7d-a 6f-c 17b-22g C  
p’solC1=p’antC2 1a-2C 15de 8a-11c 6b-2d 15c-11d 6gh 17a-16a 7d-a 6f-c 17b-22g C

h’antC1 sequence dominated in all of the populations studied, while p’antC2 was less common and occurred in both homo- and heterozygous states (Kiknadze et al. 2005).

**Arm D** of *Ch. anthracinus* is monomorphic, with one h’antD1 banding sequence (Fig. 5, a). Arm D of *Ch. solitus* is polymorphic and has two banding sequences—h’solD1 (Fig. 5, b, c), identical to h’antD1, and p’solD2 (Fig. 6), which differs by a simple inversion. Inversion heterozygotes h’solD1.p’solD2 were found in 17% of specimens from the Baikal population, and in 30% from the Irkutsk Reservoir. *Ch. anthracinus* from western parts of Palearctic also had a p’antD2 banding sequence identical to that of p’solD2 and was found in homo- and heterozygous states (Kiknadze et al. 2005):

h’antD1=h’solD1 1a-3g 14g-16e 8c-7g 5d-7f 18d-17a 8d-10a 13a-11a 14f-13b 10b-e 4a-5c 18e-24g C  
p’antD2=p’solD2 1a-3g 14g-16e 5c-4a 10e-b 13b-14f 11a-13a 10a-8d 17a-18d 7f-5d 7g-8c 18e-24g C

**Arms E** of *Ch. anthracinus* (Fig. 7, a) and *Ch. solitus* (Fig. 7, b) are monomorphic and have an identical banding sequence:

h’antE1= h’solE1 1a-3e 5a-10b 4h-3f 10c-13g C

**Arms F** of *Ch. anthracinus* (Fig. 7, c) and *Ch. solitus* (Fig. 7, d) also have an identical banding sequence that is only found in a homozygous state in East Siberia:
Figure 4. Banding sequences in the arm C of *Chironomus anthracinus* and *Chironomus solitus*. a homozygotes h’antC1.1. b homozygotes p’solC1.1 c heterozygous inversions p’solC1.h’solC2. Designations as in Fig. 3.
Figure 5. Homozygous banding sequences in the arm D of *Chironomus anthracinus* and *Chironomus solitus*. **a** h’antD1.1 **b** and **c** h’solD1.1. Designations as in Fig. 3.
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**Figure 6.** Inversion heterozygote h’solD1.p’solD2 in the arm D *Chironomus solitus*. Designations as in Fig. 3.

h’antF1= h’solF1 1a-8e 9c-17d 18a-23f C

A second nucleolus in Arm F of *Ch. anthracinus* is a species-specific feature of *Ch. anthracinus* that makes it different from *Ch. solitus* with a single nucleolus in arm G.

**Arms G** of *Ch. anthracinus* and *Ch. solitus* (Figs 1, 2) have similar morphology: unconjugated homologues with a constriction, unclear banding pattern, similar location of Balbiani Ring and nucleolus. In general, homologues have ectopic contacts in active loci.

**Discussion**

As a result of comparative analysis of banding patterns of *Ch. solitus* and *Ch. anthracinus* from East Siberia, the similarity of these species in morphological features of larvae as well as karyotypes was revealed. Most of the chromosomal arms, A, D, E and F, have identical banding sequences, and a similar structure of arm G. The principal distinctive features of *Ch. solitus* karyotype are the species-specific p’solB1 sequence and the absence of a nucleolus in arm F. Previous investigators (Belyanina 1979, Kiknadze et al. 1991, 1996, Shobanov 1996, Petrova and Rakisheva 2003, Kiknadze et al. 2005) reported a low level of chromosome polymorphisms in *Ch. anthracinus*. Analysis of the populations with standard banding sequences from Lakes Gusinoe and Arakhley also confirmed these observations. The overall banding sequence pool of *Ch. anthracinus* from other regions includes h’antC2, h’antC1 and p’antD2 sequences, which are identical to p’solC1, h’solC2 and p’solD2 from East Siberia; this is suggestive of karyological similarity of *Ch. solitus* and *Ch. anthracinus*.

There is one more species of the genus Chironomus – *Ch. reservatus* Shobanov, 1997, which has close similarity of karyotypic and morphological features at all developmental instars of *Ch. anthracinus* (Shobanov, 1997). Based on these results, the author included the two species in the *C. anthracinus* group. Banding sequence p’reS1,
Figure 7. Homozygous banding sequences in the arms E and F of *Chironomus anthracinus* and *Chironomus solitus*. a h’ant E1.1 b h’solE1.1 c h’antF1.1 d h’solF1.1.
alongside h’antB2, localised close to p’solB1, and is regarded one of the species-specific markers:

p’reB1 25s-24i 18c-16b 22b-18d 24h-23d 15m-16a 22c-23c 15l-12v C

The morphology of Ch. anthracinus and Ch. solitus imagines from East Siberia is insufficiently studied (Linevich and Erbaeva 1971), therefore, it is possible to compare only several characteristics of these species. For instance, AR of Ch. solitus (3.8) is most closely related to Ch. anthracinus (4.14–4.43) from the European part (Shobanov 1996), and Ch. anthracinus from East Siberia (5.0) – to Ch. reservatus (4.8–5.6) (Shobanov 1997). Further research into metamorphosis of these species should be conducted to make reliable conclusions.

The results of our investigation, similarity of karyotypic features of Ch. solitus and Ch. anthracinus in combination with morphological features of larvae provide evidence in favour of their close similarity and enable us to include Ch. solitus as well as Ch. reservatus in the C. anthracinus group.

Acknowledgement

This study was supported by the State Project VI.51.1.10 “Current state, biodiversity and ecology of the shore zone of Lake Baikal” (headed by Prof O.A. Timoshkin).

The author offers sincere gratitude to N.V. Bazova, P.V. Matafonov and D.V. Matafonov who kindly provided larvae from Lakes Gusinoe and Arakhley.

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