Research Article

Colorectal cancer molecular classification using BRAF, KRAS, microsatellite instability and CIMP status: Prognostic implications and response to chemotherapy

Oscar Murcia1++, Miriam Juárez2++, María Rodríguez-Soler1, Eva Hernández-Illán2, Mar Giner-Calabuig2, Miren Alustiza2, Cecilia Egoavil3, Adela Castillejo3, Cristina Alenda4, Víctor Barberá3, Carolina Mangas-Sanjuan1, Ana Yuste5, Luis Bujanda6, Joan Clófent7, Montserrat Andreu8, Antoni Castells9, Xavier Llor10, Pedro Zapater11, Rodrigo Jover1*

1 Servicio de Medicina Digestiva, Hospital General Universitario de Alicante, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain, 2 Unidad de Investigación, Hospital General Universitario de Alicante, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain, 3 Molecular Genetics Laboratory, Hospital General Universitario de Elche, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain, 4 Department of Pathology, Hospital General Universitario de Alicante, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain, 5 Oncology Department, Hospital General Universitario de Alicante, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain, 6 Gastroenterology Unity, Hospital Donostia/Instituto Biodonostia, Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (CIBERehd), Universidad del País Vasco (UPV/EHU), San Sebastián, Spain, 7 Gastroenterology Unit, Hospital de Sagunto, Sagunto, Spain, 8 Gastroenterology Unit, IMIM: Institut Hospital del Mar d’Investigacions Mèdiques, Hospital del Mar, Barcelona, Spain, 9 Gastroenterology Unit, Hospital Clinic, IDIBAPS, CIBERehd, University of Barcelona, Barcelona, Spain, 10 Section of Digestive Diseases, Yale University, Yale New Haven Hospital, New Haven, Connecticut, United States of America, 11 Clinical Pharmacology Department, Hospital General Universitario de Alicante, Instituto de Investigación Sanitaria ISABIAL, Alicante, Spain

These authors contributed equally to this work.
‡ These authors are co-first authors on this work.
* rodrigojover@gmail.com

Abstract

Objective

The aim of this study was to validate a molecular classification of colorectal cancer (CRC) based on microsatellite instability (MSI), CpG island methylator phenotype (CIMP) status, BRAF, and KRAS and investigate each subtype’s response to chemotherapy.

Design

This retrospective observational study included a population-based cohort of 878 CRC patients. We classified tumours into five different subtypes based on BRAF and KRAS mutation, CIMP status, and MSI. Patients with advanced stage II (T4N0M0) and stage III tumours received 5-fluoruracil (5-FU)-based chemotherapy or no adjuvant treatment based on clinical criteria. The main outcome was disease-free survival (DFS).

Results

Patients with the combination of microsatellite stable (MSS) tumours, BRAF mutation and CIMP positive exhibited the worst prognosis in univariate (log rank P<0.0001) and multivariate

---

Citation: Murcia O, Juárez M, Rodríguez-Soler M, Hernández-Illán E, Giner-Calabuig M, Alustiza M, et al. (2018) Colorectal cancer molecular classification using BRAF, KRAS, microsatellite instability and CIMP status: Prognostic implications and response to chemotherapy. PLoS ONE 13(9): e0203051. https://doi.org/10.1371/journal.pone.0203051

Editor: Hiromu Suzuki, Sapporo Ika Daigaku, JAPAN

Received: March 22, 2018
Accepted: August 14, 2018
Published: September 6, 2018

Copyright: © 2018 Murcia et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: This work was supported by the Instituto de Salud Carlos III (PI08/0726, INT-09/208, P111/2630, INT-12-078, INT13-196, P114/01386), FISABIO-ISABIAL foundation (UGP-13-221, UGP-14-265), and the Asociación Española contra el Cáncer (Fundación Científica GCB13131592CAST).
analyses (hazard ratio 1.75, 95% CI 1.05–2.93, P = 0.03) after adjusting for age, sex, chemotherapy, and TNM stage. Treatment with 5-FU-based regimens improved prognosis in patients with the combination of MSS tumours, KRAS mutation and CIMP negative (log rank P = 0.003) as well as in patients with MSS tumours plus BRAF and KRAS wild-type and CIMP negative (log-rank P < 0.001). After adjusting for age, sex, and TNM stage in the multivariate analysis, only patients with the latter molecular combination had independently improved prognosis after adjuvant chemotherapy (hazard ratio 2.06, 95% CI 1.24–3.44, P = 0.005).

Conclusion
We confirmed the prognostic value of stratifying CRC according to molecular subtypes using MSI, CIMP status, and somatic KRAS and BRAF mutation. Patients with traditional chromosomally unstable tumours obtained the best benefit from adjuvant 5-FU-based chemotherapy.

Introduction
Colorectal cancer (CRC) is one of the most prevalent neoplasms and an important cause of death in Western countries [1]. In the last few years, several therapeutic options have improved the survival of CRC patients, due in part to better comprehension of its molecular biology. CRC has largely been considered a heterogeneous disease, with an increasing number of pathways involved in different subtypes. Therefore, it is plausible that these different subtypes exhibit different behaviour in terms of not only overall prognosis, but also the response to adjuvant chemotherapy. Recently, studies demonstrating the role of immunotherapy in MSI tumours highlighted the importance of adequately classifying CRC patients in order to effectively provide the best molecular-based treatment [2].

Phipps et al [3] and Sinicrope et al [4] proposed two molecular classifications with prognostic implications based on different combinations of microsatellite instability (MSI), CpG island methylator phenotype (CIMP) status, and BRAF and KRAS somatic mutations in tumours. Thus, we can classify CRC into five subtypes according to the described pathways of colorectal carcinogenesis [5]. Studies have shown clinical and pathological differences between the subtypes, as well as relevant differences in prognosis [3–7]. On the other hand, Guinney and colleagues proposed a new molecular classification based on gene expression profiling [8,9] but some difficulties have arisen in transferring this classification to clinical practice [10–12]. As the molecular classification based on MSI, CIMP, BRAF, and KRAS [3] is simple and easy to apply, the aim of our study was to reproduce and validate this classification and investigate the response of each of subtypes to chemotherapy.

Material and methods

Study population
We enrolled a population-based cohort of 878 patients with CRC, available tumour tissue and complete genotyping for BRAF, KRAS, CIMP and MSI status, from the nationwide and multicentre EPICOLON I and EPICOLON II projects [13,14] in a retrospective observational study (Fig 1). Patients were included between years 2000–2001 in EPICOLON I and 2006–2007 in EPICOLON II. Adjuvant chemotherapy was administered following standard clinical criteria, regardless the MMR status of the tumours. Oncologists that decided
treatment with adjuvant chemotherapy were blinded to the MMR status of the tumours. The institutional review board (IRB) of the Hospital General Universitario de Alicante approved the study as well as IRBs of all the participant hospitals. All patients provided written informed consent.

**CRC samples**

After the surgery of each CRC, tumor samples were obtained and paraffin-embedded at the different participant hospitals. These paraffin-embedded samples were sent to the Hospital General Universitario of Alicante. There, DNA was extracted from paraffin-embedded tissue with the QIAamp DNA Investigator kit (QIAGEN, Hilden, Germany) and with the E.Z.N.A Forensic DNA kit (OMEGA Biotek, Norcross, GA), according to manufacturer’s protocols.

**Variables**

We analysed patient age, sex, tumour location, date and stage at the time of CRC diagnosis, **BRAF** and **KRAS** mutation status, CIMP, and MSI status, treatment (surgical, chemotherapy, radiotherapy, or biologic), and disease-free survival (DFS) time in months. We analysed the response to adjuvant chemotherapy in patients with advanced stage II (TNM stage II and T4) and stage III tumours. The main outcome was DFS. The follow-up of patients after surgery was performed according to clinical practice guidelines with scheduled CEA measurements, computed tomography and colonoscopy procedures [15;16].

**Mutations in the BRAF and KRAS genes**

The V600E **BRAF** mutation was detected by real-time PCR (ABI PRISM 7500, Applied Biosystems, Foster City, CA) using specific TaqMan probes and allelic discrimination software as
described previously [17]. The reporter fluorophore on the probes was 6-carboxyfluorescein (FAM) for the mutant allele and VIC for the wild-type allele. We used the allelic discrimination software on the ABI Prism 7500 instrument to analyse the fluorescence data. KRAS mutation at exon 1, including codons 12 and 13, was identified by DNA direct sequencing. We assessed both mutations by direct amplicon sequencing with BigDye v1.1 terminators and a 3500 Genetic Analyzer (Applied Biosystems) [18].

CIMP analysis
Genomic DNA was modified with sodium-bisulphite using the EZ Methylation Gold Kit (Zymo Research, Orange, CA). The PCR reaction contained bisulphite modified DNA, HotStar Taq polymerase, forward primers, biotinylated reverse primers, and water. We analysed markers by real-time methylight technology, considering a tumour CIMP-H when three of the five analysed markers (CACNAG1, SOCS1, RUNX3, NEUROG1 and MLH1) harboured aberrant hypermethylation. Each marker was classified as methylated when the mean percentage was greater than 5% for CACNAG1, SOCS1, RUNX3, and MLH1, and 10% for NEUROG1.

MSI analysis
We tested for MSI using either the 5-marker panel proposed by the National Cancer Institute or a pentaplex of mononucleotide repeats (BAT25, BAT26, NR21, NR24, and NR27), classifying tumours as microsatellite stable (MSS) or unstable (MSI). The presence of two or more unstable markers defined a tumour as MSI [19;20].

Chemotherapy regimen
The adjuvant chemotherapy regimen followed standard clinical criteria. Patients with advanced stage II and stage III tumours received 5-fluorouracil (5-FU) based chemotherapy (5-FU, capecitabine, 5-FU plus oxaliplatin (FOLFOX), or 5-FU plus irinotecan (FOLFIRI)) as first line treatment following standard schedules and doses according to the local decision of the responsible oncologist. Oncologists were blinded to the BRAF, KRAS, MSI or CIMP tumour status. The decision to give or not to give chemotherapy drugs was based on the oncologist criteria in each of the participating centres.

Classification of CRC
We classified tumours into five subtypes according to the model proposed by Phipps et al [3] and shown in Table 1. Subtype 1, sporadic serrated and unstable tumours; subtype 2, sporadic serrated tumours without MSI; subtype 3, traditional tumours with KRAS mutations or serrated tumours from the alternative serrated pathway; subtype 4, traditional chromosomally unstable (CIN) tumours without KRAS mutations; and subtype 5, familial unstable tumours.

Table 1. Molecular classification of colorectal cancer. MSI, microsatellite instability; MSS, microsatellite stability.

| Subtype  | MMR | CIMP | BRAF | KRAS |
|----------|-----|------|------|------|
| 1        | MSI | +    | +    | -    |
| 2        | MSS | +    | +    | -    |
| 3        | MSS | -    | -    | +    |
| 4        | MSS | -    | -    | -    |
| 5        | MSI | -    | -    | -    |
Statistical analysis

In order to maximize the number of cases available for analysis, we performed a multiple imputation model to complete cases with missing values at one (n = 141) or two molecular markers (n = 16) [3;21]. We carried out 25 rounds of imputation using the automatic model of SPSS software until an appropriate model was obtained [22]. We considered a deviation proportion with respect to original data <0.1% as appropriate; it reliably reproduced the percentage of alterations at the four markers in the complete cases model. The imputation took into account BRAF and KRAS status, presence of CIMP, MMR status, sex, age, TNM stage, tumour location, treatment with chemotherapy, and DFS time. After imputation, we classified cases into subtypes 1 to 5. Cases with no defined molecular combination were defined as the “unclassified” subtype. We performed several simulation analyses before concluding the validation of the imputation model, including relative risks (RRs) and hazard ratios (HRs). Original data, non-including patients with missing values using the imputation model, are available as Supplementary material and referred as complete-cases model (S1–S4 Tables).

Parametric continuous variables are reported as mean ± standard deviation (SD) and non-parametric continuous variables as median (Q2-Q3 interquartile range). We compared categorical variables using chi-squared.

For overall prognosis, we compared differences in DFS time (interval of time between remission of disease and their reappearance) among the five subtypes by log rank test in a univariate analysis, expressing it graphically with Kaplan-Meier survival curves. The multivariate analysis was performed by adjusting for potential confounder and interaction variables (age, sex, TNM stage, and chemotherapy) in a Cox regression model. Subtype 4 was the subtype of reference.

For the adjuvant chemotherapy response, we performed several univariate analyses for each subtype. The multivariate analysis was adjusted for the different subtypes, age, sex and TNM stage. As comparisons were made in each subgroup between patients receiving and not receiving adjuvant chemotherapy, we did not establish a reference group.

All P-values were two-sided, with P<0.05 considered significant. All data were analysed using SPSS 22.0 software.

Results

Among the 878 patients, complete analysis of the four molecular markers was possible in 721 (Fig 1). A total of 141 cases had one missed marker, and 16 cases had two missed markers. These cases were completed using the multiple imputation model. The median follow-up was 52 months (interquartile range 16–64). A total of 241 cases (27%) had a CIMP-high pattern. A total of 324 cases had a somatic KRAS mutation (37%) and 54 had somatic mutations in BRAF (6%). Eighty cases (10%) had MSI. According to the proposed model, we could classify 677 patients (77%) into subtypes 1 to 5: 25 (3%) subtype 1, 22 (3%) subtype 2, 218 (25%) subtype 3, 388 (44%) subtype 4, and 24 (3%) subtype 5. The remaining cases (n = 201, 23%) were tumours of the “unclassified” molecular subtype (Table 2). The median age was lower in cases with subtype 5 tumours compared to the other subtype groups, which had similar median ages. Sex was homogeneously distributed, except in the subtype 5 group, which had a predominance of women. The majority of tumours were TNM stages II or III at diagnosis, with no significant differences among the subtypes. A majority of subtype 1 tumours were right-sided, whereas subtypes 3 and 4 were preferably located in the left colon. The proportion of patients treated with chemotherapy differed slightly among subtypes (Table 2).

Regarding the “unclassified” subtype, there were observed three major genetic combinations among the 201 cases. Of them, 93 (46.3%) had CIMP-H and KRAS-mutation, which were
equally distributed regarding sex, age and tumor location. Other 68 cases (33.8%) had CIMP-H in combination with any other genetic alteration. These were preferentially men (63%) and located in left colon (71%). Finally, 21 showed CIMP-H and MSI combination with BRAF wild-type (10.4%). Characteristics of the “unclassified” subtype are shown in the S1 Table.

Overall prognosis

Fig 2 shows the Kaplan Meier survival curves for the different CRC subtypes. In the univariate analysis, subtype 2 had the lowest DFS, below 30%. In contrast, subtype 5 had a DFS rate at the end of the follow-up period of up to 80% (log rank $P<0.001$, Table 3).

Multivariate analysis showed that age ($P<0.001$), TNM stage ($P<0.001$), and chemotherapy ($P<0.001$) were significantly influential in regards to DFS. Regarding the different subtypes, only patients with subtype 2 (HR = 1.75, $P = 0.03$) had a different prognosis with respect to the subtype of reference after adjusting for age, sex, chemotherapy, and TNM (Table 3). Subtype 3 also revealed a trend for different prognosis (HR = 1.2, $P = 0.109$). Trends were similar in the complete-cases model, although no significantly independent better prognosis was found in subtype 2 (S2 Table).

Prognosis after adjuvant chemotherapy

We analysed the response to chemotherapy in 324 patients with advanced stage II (T4N0M0) or stage III CRC (Table 4).

In this group, a total of 114 patients did not receive adjuvant chemotherapy. Overall, chemotherapy improved DFS time (log rank $P<0.001$) in the univariate analysis. Regarding CRC subtypes, there was a clear benefit from chemotherapy in subtypes 3 and 4. In both subtypes,
the patients had higher DFS rates when they received chemotherapy (log rank $P = 0.003$ for subtype 3 and $P < 0.001$ for subtype 4, Fig 3A and 3B and Table 5).

A multivariate analysis was also performed for the different subtypes (Table 5). After adjusting for age, sex, and TNM stage, only patients with subtype 4 CRC independently exhibited significant benefit with chemotherapy (subtype 3: HR = 1.93, 95% CI 0.86–4.34, $P = 0.1$; subtype 4: HR = 2.06, 95% CI 1.24–3.44, $P = 0.005$). These results were similar in the complete-cases model (S3 and S4 Tables).

Figure 2. Kaplan-Meier survival curves comparing disease-free survival in colorectal cancer patients by tumour subtype.

![Kaplan-Meier survival curves](https://doi.org/10.1371/journal.pone.0203051.g002)

Table 3. Overall prognosis for different subtypes in the multiple imputation model. Subtype 4 serves as a reference. The analysis was adjusted for sex, age, chemotherapy, and TNM as potential confounder factors. CRC, colorectal cancer; HR, hazard ratio; CI, confidence interval.

| Case participants | Relapse or CRC-death |
|-------------------|----------------------|
|                   | Number | % | Number | % | HR  | 95% CI   | P value |
| Subtype 1         | 25     | 2.8 | 8       | 32.0 | 0.59 | 0.29–1.20 | 0.144 |
| Subtype 2         | 22     | 2.5 | 16      | 76.2 | 1.75 | 1.05–2.93 | 0.032 |
| Subtype 3         | 218    | 24.8 | 117     | 54.9 | 1.21 | 0.96–1.52 | 0.109 |
| Subtype 4         | 388    | 44.2 | 191     | 50.3 | 1.0  | 1.0      | —     |
| Subtype 5         | 24     | 2.7 | 7       | 29.2 | 0.56 | 0.26–1.19 | 0.131 |
| Unclassified      | 201    | 22.9 | 85      | 42.9 | 0.84 | 0.65–1.09 | 0.187 |

https://doi.org/10.1371/journal.pone.0203051.t003
Discussion

Our results confirm that, as suggested in previous studies [3;4], classification of the molecular profile of CRC using BRAF, KRAS, MSI, and CIMP status can identify differences in prognosis. Our results confirm the poorest prognosis found for subtype 2 in previous cohorts, unaffected by age or TNM stage. This lower survival rate for subtype 2 tumours suggests that the combination of CIMP and BRAF mutation confers a poor prognosis in stable tumours, whereas the same combination in tumours with MSI (i.e., subtype 1) evolves in a completely different way. Moreover, our results show that the more common subtypes 3 and 4 benefit from adjuvant 5-FU-based chemotherapy, but only in subtype 4 CRC patients is this benefit independent of
sex, age, and TNM stage. The response in other subtypes cannot be evaluated due to their small representation in our cohort.

Subtypes 1 and 2 belong to the serrated pathway of carcinogenesis, with serrated polyps as the precursor lesion, whereas subtypes 4 and 5 follow the traditional adenoma-carcinoma pathway with MSS (subtype 4) tumours or MSI (subtype 5). On the other hand, subtype 3 includes tumours that can originate in serrated or adenomatous polyps, with the KRAS somatic mutation as their molecular hallmark [5]. Different studies have shown discordant results with respect to the prognostic value of some molecular markers [23;24]. BRAF V600E has traditionally been linked to a poor prognosis, with many authors proposing it as an independent predictor of low survival [25]. However, other studies have found differences according to MMR status, showing better survival rates in tumours with MSI than in MSS tumours. [26] Something similar has been seen with CIMP [27]; though many studies have not attributed any prognostic value [23], others proposed it is a predictor of short survival, and even of high survival if MSI appeared concomitantly [28]. Taken together, all these discordances could be due to the consideration of isolated markers instead of combinations of them [10]. Grouping according to the proposed molecular pathways led to different and well defined survival curves. Recently, Guinney et al [8] proposed a new molecular classification of CRC based on gene expression datasets. This classification divides CRC into four subtypes: MSI immune with MSI, BRAF mutated, and CIMP positive tumours; canonical with WNT and MYC activation; metabolic with KRAS mutated tumours; and mesenchymal, characterized by stromal infiltration and TGF-β activation. The majority of the proposed molecular subtypes somehow coincide with those in our study [10]. However, some patients are very difficult to classify, especially those from the mesenchymal subtype, which lacks convenient molecular makers. Although this classification can be considered the most robust classification system currently available for CRC, with a clear biological basis and comprehensive tumour characterization based on advances in genomic technology, the challenges in data interpretation and clinical application have not yet been adequately overcome [10].

In our knowledge, this is the first study that examines the classification of CRC proposed by Phipps and tries to correlate the response of each subtype to chemotherapy and compare among them. Previous studies using these classifications did not analyse the role of chemotherapy in the evolution of CRC patients in regards to molecular subtypes. Unfortunately, our

Table 5. Chemotherapy response of different subtypes in univariate and multivariate analyses using the multiple imputation model. The multivariate analysis was adjusted for sex, age, and TNM stage for each subtype. CT, chemotherapy; DFS, disease-free survival; HR, hazard ratio; CI, confidence interval.

| Subtype  | CT   | Number of patients | Median of DFS time (months) | P value | HR      | 95% CI   | P value |
|----------|------|--------------------|----------------------------|---------|---------|----------|---------|
| Subtype 1| Yes  | 4                  | 39.3                       | 0.620   | 0.53    | 0.19–1.46| 0.220   |
|          | No   | 5                  | 62.2                       |         |         |          |         |
| Subtype 2| Yes  | 6                  | 44.6                       | 0.068   | 0.40    | 0.04–3.92| 0.430   |
|          | No   | 4                  | 9.6                        |         |         |          |         |
| Subtype 3| Yes  | 47                 | 49.5                       | 0.003   | 1.93    | 0.86–4.34| 0.111   |
|          | No   | 30                 | 27.8                       |         |         |          |         |
| Subtype 4| Yes  | 99                 | 54.2                       | 0.000   | 2.06    | 1.24–3.44| 0.005   |
|          | No   | 43                 | 29.0                       |         |         |          |         |
| Subtype 5| Yes  | 3                  | 63.9                       | 0.107   | 1.01    | 0.31–3.28| 0.991   |
|          | No   | 3                  | 16.9                       |         |         |          |         |
| Unclassified| Yes | 51                 | 58.6                       | 0.015   | 1.71    | 0.74–3.96| 0.208   |
|          | No   | 29                 | 39.0                       |         |         |          |         |

https://doi.org/10.1371/journal.pone.0203051.t005
study does not have enough power to confirm this lack of response of MSI subtypes 1 and 5 due to the small sample size. However, this lack of response to 5-FU-based chemotherapy by tumours with MSI was well established in previous studies [29–32]. Our results highlight that the more common CRC subtypes seem to improve with chemotherapy in the univariate analysis (subtypes 3 and 4). These benefits were only maintained in subtype 4 in the multivariate analysis, revealing an advantage of this subtype in 5-FU-based chemotherapy, and a potential limited effect of this treatment in subtype 3 CRC patients. On the other hand, we also observed a trend of obtaining a benefit from chemotherapy for subtype 2. The hypothesis that patients with molecularly different tumours have different responses to chemotherapy is clearly plausible given the biological diversity found between CRC tumours. It is especially interesting to investigate the response of subtype 2 and 3 CRC, as they are biologically different from the classical CIN CRC. Another point that requires specific research is the possible role of new targeted molecules in the different molecular CRC subtypes. Improved survival using immunotherapy in CRC with MSI was demonstrated recently [2], and there is a biological explanation for both: the lack of effect of conventional 5-FU-based chemotherapy and the benefit from immunotherapy in this subtype of CRC. The role of immunotherapy or other types of chemotherapy could be more appropriately investigated if we were able to adequately classify CRC in different subtypes. Larger cohorts are needed for differentially investigating the role of different drugs in the diverse molecular landscape of CRC.

This study has some limitations that should be considered. First, it is important to note that we could appropriately classify only 71% of these cases using the molecular markers. There are at least two reasons for this. First, biology is not perfect and may include aberrant combinations with an unpredictable course. Second, it is possible that there are other molecular pathways of colorectal carcinogenesis not included in this classification and molecular subtypes of CRC with prognostic implications could include more than those proposed. Another limitation of our study is the small number of patients receiving adjuvant chemotherapy in minority subtypes (i.e., 1, 2, and 5), precluding the possibility of appropriately determining the response to 5-FU-based chemotherapy in these subtypes. In addition, our study shows a higher proportion of CIMP positive tumors (27%) than other studies [3]. This could be likely attributable to the characteristics of our sample, such as the mean age of our unselected consecutive population, with more than 60% older than 70 years-old. Finally, the decision regarding giving chemotherapy in stage II and III patients was based on clinical criteria. Previous studies of these CRC subtypes were nested in randomized clinical trials comparing different chemotherapy regimens [3,4], which precludes obtaining information about the predictive value of this subtype classification. We can investigate the role of adjuvant 5-FU-based chemotherapy only if there is a group of patients that do not receive treatment, and that could be considered a strength of our study. However, basing the treatment decision on clinical criteria can provoke some biases, because it is possible that this clinical decision was based on factors such as age and performance status, avoiding chemotherapy in people in the worst health status. Nevertheless, that scenario otherwise allows us to study the effect of adjuvant chemotherapy in the different subtypes considering the difficulty obtaining other data and given that it could be considered unethical to design a randomized clinical trial with an untreated control group in advanced stage II and stage III CRC patients.

In summary, our results confirm the prognostic value of stratifying CRC according to molecular subtypes using MSI and CIMP status and somatic KRAS and BRAF mutations. The use of this classification can be helpful when making clinical decisions in the management of CRC patients. Serrated CRC with CIMP and somatic BRAF mutation has the worst prognosis. 5-FU-based chemotherapy provides a benefit in subtypes 3 and 4 in regards to DFS time, but this benefit was independent after adjusting for sex, age, and TNM stage only in subtype 4.
tumours representing the traditional CIN pathway. A better understanding of the biology of CRC, description of new pathways of colorectal carcinogenesis, and determination of driver mutations will improve and complete this classification in the near future. Research should focus on molecularly targeted therapy for the different subtypes, especially for those with the worst prognosis.

**Supporting information**

S1 Table. Clinical characteristics of patients according to subtype in complete-cases model. St, subtype. DFS, disease-free survival. (DOCX)

S2 Table. Overall prognosis for different subtypes in the complete cases model. Subtype 4 serves as a reference. The analysis was adjusted for sex, age, chemotherapy, and TNM as potential confounder factors. CRC, colorectal cancer; HR, hazard ratio; CI, confidence interval. (DOCX)

S3 Table. Clinical characteristics of patients with advanced stage II and stage III tumours in complete-cases model. DFST, disease-free survival time. St, Subtype. (DOCX)

S4 Table. Chemotherapy response of different subtypes in univariate and multivariate analyses using the complete-cases model. The multivariate analysis was adjusted for sex, age, and TNM stage for each subtype. CT, chemotherapy; DFS, disease-free survival; HR, hazard ratio; CI, confidence interval. (DOCX)

**Author Contributions**

**Conceptualization:** Eva Hernández-Illán, Montserrat Andreu.

**Data curation:** Adela Castillejo, Xavier Llor.

**Formal analysis:** Adela Castillejo, Cristina Alenda.

**Investigation:** Oscar Murcia, Miriam Juárez, Mar Giner-Calabuig, Víctor Barberá, Carolina Mangas-Sanjuan, Antoni Castells, Rodrigo Jover.

**Methodology:** María Rodríguez-Soler, Mar Giner-Calabuig, Miren Alustiza, Cecilia Egoavil, Ana Yuste, Luis Bujanda, Joan Clófent, Pedro Zapater, Rodrigo Jover.

**Project administration:** Cristina Alenda.

**Writing – review & editing:** Rodrigo Jover.

**References**

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebele M et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015; 136(5): E359–E386. https://doi.org/10.1002/ijc.29210 PMID: 25220842

2. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N Engl J Med* 2015; 372(26):2509–20. https://doi.org/10.1056/NEJMoa1500596 PMID: 26028255

3. Phipps AI, Limburg PJ, Baron JA, Burnett-Hartman AN, Weisenberger DJ, Laird PW et al. Association between molecular subtypes of colorectal cancer and patient survival. *Gastroenterology* 2015; 148 (1):77–87. https://doi.org/10.1053/j.gastro.2014.09.036 PMID: 25280443
4. Sinicrope FA, Shi Q, Smyrk TC, Thibodeau SN, Dienstmann R, Guinney J et al. Molecular markers identify subtypes of stage III colon cancer associated with patient outcomes. *Gastroenterology* 2015; 148(1):88–99. https://doi.org/10.1053/j.gastro.2014.09.041 PMID: 25305506

5. Jass JR. Classification of colorectal cancer based on correlation of clinical, morphological and molecular features. Histopathology 2007; 50(1):113–30. https://doi.org/10.1111/j.1365-2559.2006.02549.x PMID: 17204026

6. Bae JM, Kim JH, Kang GH. Molecular Subtypes of Colorectal Cancer and Their Clinicopathologic Features, With an Emphasis on the Serrated Neoplasia Pathway. *Arch Pathol Lab Med* 2016; 140(5):406–12. https://doi.org/10.5858/arpa.2015-0310-RA PMID: 27128298

7. Samadder NJ, Vierkant RA, Tillmans LS, Wang AH, Weisenberger DJ, Laird PW et al. Associations between colorectal cancer molecular markers and pathways with clinicopathologic features in older women. *Gastroenterology* 2013; 145(2):348–56. https://doi.org/10.1053/j.gastro.2013.05.001 PMID: 23665275

8. Guinney J, Dienstmann R, Wang X, de Reyniès A, Schlicker A, Soneson C et al. The consensus molecular subtypes of colorectal cancer. *Nat Med* 2015; 21(11):1350–6. https://doi.org/10.1038/nm.3967 PMID: 26457759

9. Dienstmann R, Vermeulen L, Guinney J, Kopetz S, Tejpar S, Tabernero J et al. Consensus molecular subtypes and the evolution of precision medicine in colorectal cancer. *Nat Rev Cancer* 2017; 17(4):268.

10. Sinicrope FA, Okamoto K, Kasi PM et al. Molecular Biomarkers in the Personalized Treatment of Colorectal Cancer. *Clin Gastroenterol Hepatol* 2016; 14(5):651–8. https://doi.org/10.1016/j.cgh.2016.02.008 PMID: 26872400

11. Trinh A, Trumpi K, De Sousa E Melo, Wang X, de Jong JH, Fessler E et al. Practical and Robust Identification of Molecular Subtypes in Colorectal Cancer by Immunohistochemistry. *Clin Cancer Res* 2017; 23(2):387–98. https://doi.org/10.1158/1078-0432.CCR-16-0680 PMID: 27459899

12. Dunne PD, McArt DG, Bradley CA, O’Reilly PG, Barrett HL, Cummins R et al. Challenging the Cancer Molecular Stratification Dogma: Infratumoral Heterogeneity Undermines Consensus Molecular Subtypes and Potential Diagnostic Value in Colorectal Cancer. *Clin Cancer Res* 2016; 22(16):4095–104. https://doi.org/10.1158/1078-0432.CCR-16-0032 PMID: 27151745

13. Pinol V, Castells A, Andreu M, Castellví-Bel S, Alenda C, Llor X, et al. Accuracy of revised Bethesda guidelines, microsatellite instability, and immunohistochemistry for the identification of patients with hereditary nonpolyposis colorectal cancer. *JAMA* 2005; 293(16):1986–94. https://doi.org/10.1001/jama.293.16.1986 PMID: 15953432

14. Abuli A, Bessa X, Gonzalez JR, Ruiz-Ponte C, Cáceres A, Muñoz J et al. Susceptibility genetic variants associated with colorectal cancer risk correlate with cancer phenotype. *Gastroenterology* 2010; 139(3):788–96, 796. https://doi.org/10.1053/j.gastro.2010.05.072 PMID: 20638935

15. Van Cutsem EJ. Advanced colorectal cancer: ESMO clinical recommendations for diagnosis, treatment and follow-up. *Ann Oncol* 2007; 18 Suppl 2:ii25–ii26.

16. Van Cutsem EJ, Oliveira J. Colon cancer: ESMO clinical recommendations for diagnosis, adjuvant treatment and follow-up. *Ann Oncol* 2008; 19 Suppl 2:ii25–ii26.

17. Benlloch S, Paya A, Alenda C, Bessa X, Andreu M, Jover R et al. Detection of BRAF V600E mutation in colorectal cancer: comparison of automatic sequencing and real-time chemistry methodology. *J Mol Diagn* 2006; 8(5):540–3. https://doi.org/10.2353/jmoldx.2006.060070 PMID: 17065421

18. Guarinos C, Juarez M, Egoavil C, Rodríguez-Soler M, Pérez-Carbonell L, Salas R et al. Prevalence and characteristics of MUTYH-associated polyposis in patients with multiple adenomatous and serrated polyps. *Clin Cancer Res* 2014; 20(15):5158–68. https://doi.org/10.1158/1078-0432.CCR-13-1490 PMID: 24470512

19. Boland CR, Thibodeau SN, Hamilton SR, Sidransky D, Eshleman JR, Burt RW et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res* 1998; 58(22):5248–57. PMID: 9823339

20. Xiocia RM, Llor X, Pons E, Castells A, Alenda C, Piñol V et al. Performance of different microsatellite marker panels for detection of mismatch repair-deficient colorectal tumors. *J Natl Cancer Inst* 2007; 99(3):244–52. https://doi.org/10.1093/jnci/djk033 PMID: 17284719

21. Gottfredson NC, Sterba SK, Jackson KM. Explicating the Conditions Under Which Multilevel Multiple Imputation Mitigates Bias Resulting from Random Coefficient-Dependent Missing Longitudinal Data. *Prev Sci* 2017; 18(1):12–9. https://doi.org/10.1007/s11121-016-0735-3 PMID: 27866307

22. Pedersen AB, Mikkelsen EM, Cronin-Fenton D, Kristensen NR, Pham TM, Pedersen L et al. Missing data and multiple imputation in clinical epidemiological research. *Clin Epidemiol* 2017; 9:157–66. https://doi.org/10.2147/CLEP.S129785 PMID: 28352203
23. Juo YY, Johnston FM, Zhang DY, Juo HH, Wang H, Pappou EP et al. Prognostic value of CpG island methylator phenotype among colorectal cancer patients: a systematic review and meta-analysis. *Ann Oncol* 2014; 25(12):2314–27. https://doi.org/10.1093/annonc/mdu149 PMID: 24718889

24. Thiel A, Ristimaki A. Toward a Molecular Classification of Colorectal Cancer: The Role of BRAF. *Front Oncol* 2013; 3:281. https://doi.org/10.3389/fonc.2013.00281 PMID: 24298448

25. Farina-Sarasqueta A, van LG, Moerland E, Creemers GJ, Lemmens VE, Rutten HJ et al. The BRAF V600E mutation is an independent prognostic factor for survival in stage II and stage III colon cancer patients. *Ann Oncol* 2010; 21(12):2396–402. https://doi.org/10.1093/annonc/mdq258 PMID: 20501503

26. Samowitz WS, Sweeney C, Herrick J, Albertsen H, Levin TR, Murtaugh MA et al. Poor survival associated with the BRAF V600E mutation in microsatellite-stable colon cancers. *Cancer Res* 2005; 65(14):6063–9. https://doi.org/10.1158/0008-5472.CAN-05-0404 PMID: 16024606

27. Ahn JB, Chung WB, Maeda O, Shin SJ, Kim HS, Chung HC et al. DNA methylation predicts recurrence from resected stage III proximal colon cancer. *Cancer* 2011; 117(9):1847–54. https://doi.org/10.1002/cncr.25737 PMID: 21509761

28. Ogino S, Nosho K, Kirkner GJ, Kawasaki T, Meyerhardt JA, Loda M et al. CpG island methylator phenotype, microsatellite instability, BRAF mutation and clinical outcome in colon cancer. *Gut* 2009; 58(1):90–6. https://doi.org/10.1136/gut.2008.155473 PMID: 18832519

29. Jover R, Zapater P, Castells A, Llor X, Andreu M, Cubiella J et al. The efficacy of adjuvant chemotherapy with 5-fluorouracil in colorectal cancer depends on the mismatch repair status. *Eur J Cancer* 2009; 45(3):365–73 https://doi.org/10.1016/j.ejca.2008.07.016 PMID: 18722765

30. Sargent DJ, Marsoni S, Monges G, Thibodeau SN, Labianca R, Hamilton SR et al. Defective mismatch repair as a predictive marker for lack of efficacy of fluorouracil-based adjuvant therapy in colon cancer. *J Clin Oncol* 2010; 28(20):3219–26. https://doi.org/10.1200/JCO.2009.27.1825 PMID: 20498393

31. Ribic CM, Sargent DJ, Moore MJ, Thibodeau SN, French AJ, Goldberg RM et al. Tumor microsatellite instability as a predictor of benefit from fluorouracil-based adjuvant chemotherapy for colon cancer. *N Engl J Med* 2003; 349:247–57 https://doi.org/10.1056/NEJMoa022289 PMID: 12867608

32. Jover R, Zapater P, Castells A, Llor X, Andreu M, Cubiella J et al. Mismatch repair status in the prediction of benefit from adjuvant fluorouracil chemotherapy in colorectal cancer. *Gut* 2006; 55:848–55. https://doi.org/10.1136/gut.2005.073015 PMID: 16299036