Acquired Factor V Inhibitor After Antibiotic Therapy: A Clinical Case Report and Review of the Literature

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Abstract

Acquired factor V inhibitor (aFVi) is an exceptionally rare hematologic condition that can range from incidental laboratory abnormalities to life-threatening hemorrhage. Bovine thrombin was formerly the most common cause of this condition; however, the decreased use of bovine thrombin in surgical procedures has led to a shift in the cause of aFVi toward antibiotic use and malignancies. Here we present a case of an 80-year-old Caucasian female on long-term warfarin therapy who presented with epistaxis and an elevated international normalized ratio, and a history of cephalosporin antibiotic use. We review the published literature beginning in 2016 to identify the evolving causes of aFVi. Additionally, we propose that stress-mediated immune regulation may contribute to antibody formation, preventing the interaction between factor V and the damaged phospholipid membranes. This case highlights the evolving causes of aFVi and should prompt physicians to consider this diagnosis in coagulopathies that do not correct with traditional therapies.

Introduction

Acquired hemophilia is a rare and clinically challenging scenario as patient presentation can range from clinically asymptomatic laboratory abnormalities to life-threatening hemorrhage. Acquired factor V inhibitors (aFVis) were first identified by Hördor in 1955 and twice more by Ferguson et al. in 1957 and 1958 [1]. Since the initial reports, physicians have become aware of the associations between surgical procedures, antibiotics, immunosuppressants, and bovine serum proteins, and this rare condition. In their review of 105 previous cases in 1998, Knobl and Lechner found that from the identified factors, the majority of cases were secondary to the use of bovine thrombin as a hemostatic agent [1]. Bovine thrombin has largely been replaced since the FDA approval of recombinant human thrombins in both 2007 and 2008 [2]. The use of a less immunogenic thrombin has removed the largest cause of aFVi, further decreasing the incidence of this already exceedingly rare disease. Recently, there has been a shift in the associated causes of aFVi toward antibiotics, malignancies, and autoimmune disorders [3]. There is not an accepted mechanism as to how antibodies are formed against factor V (FV) in these situations. Two retrospective studies found that aFVi had an incidence of between 0.06 and 0.09 cases per one million people per year [4]. The true incidence of aFVi is likely higher due to the presence of asymptomatic carriers, which were found to represent between 19% and 31% of cases in literature reviews [3-5]. Here we present a case of a patient who was prescribed cephalosporin antibiotics after a podiatric procedure and subsequently presented with increased international normalized ratio (INR) and epistaxis and was found to have developed
Case Presentation

An 80-year-old obese Caucasian female with a relevant past medical history significant for deep vein thromboses in the 1970s being managed with warfarin presented to the emergency department with epistaxis for 24 hours and INR > 10 on her home monitor. Prior to this, the patient denied any recent dose change to her warfarin or taking any additional doses. The patient denied any changes to diet or activity. The patient was seen by a podiatrist three weeks prior for a left hallux total nail removal and was prescribed a course of cephalexin 500 mg two times daily for seven days, which she completed.

The patient was well appearing on presentation in the emergency department (ED); she denied hematemesis, melena, or hematuria. The patient denied any changes to medications except for the recent cephalexin use. The patient denied any known family bleeding disorders. Physical exam was significant for dried blood in the nares, and blood was seen in the posterior oropharynx. Additionally, it was noted she had a large ecchymosis on her right breast. Vital signs were unremarkable in the ED. Lab results were significant for hemoglobin level of 14.1 g/dL (reference range: 11.3-15.1 g/dL), red blood cell count of 5.0 x 10^6/μL (reference range: 3.7-5.2 x 10^6/μL), white blood cell count of 8.2 x 10^3/μL (reference range: 3.9-11.2 x 10^3/μL), and platelet count 218 x 10^3/μL (reference range: 165-366 x 10^3/μL). A coagulation profile revealed INR, prothrombin time (PT), and activated partial thromboplastin time (APTT) above detectable limits (Table 1). In the ED, she was given 2.5 mg oral vitamin K. The patient was then admitted to the general medicine service for further evaluation and management.
| Day of admission | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 |
|------------------|---|---|---|---|---|---|---|---|
| Treatment        | 2.5 mg PO vitamin K, 10 mg PO vitamin K, 10 mg SQ vitamin K, 10 mg IV vitamin K | PCC | 100 mg PO prednisone | 100 mg PO prednisone | Rituximab 375 mg/m² x 2.26 m² (did not finish), 100 mg PO prednisone | Rituximab (finished), 100 mg PO prednisone | 100 mg PO prednisone |
| PT (9.5-11.2 seconds) | >90 | >90 | >90 | >90 | >90 | >90 | >90 | >90 |
| INR              | >10 | >10 | >10 | >10 | >10 | >10 | >10 | >10 |
| APTT (22-29 seconds) | >139 | >139 | >139 | >139 | >139 | >139 | >139 | >139 |
| PT mix (9.5-11.2 seconds) | 44.7 | 44.7 | 44.7 | 44.7 | 44.7 | 44.7 | 44.7 | 44.7 |
| APTT mix (22.0-29.0 seconds) | 84 | 84 | 84 | 84 | 84 | 84 | 84 | 84 |
| Thrombin time (0.0-23.9 seconds) | 19.1 | 19.1 | 19.1 | 19.1 | 19.1 | 19.1 | 19.1 | 19.1 |
| Factor V inhibitor | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units | 30 inhibitor units |

**TABLE 1: Selected Labs and Treatment of First Admission**

PO, oral administration; SQ, subcutaneous; PCC, prothrombin complex concentrate; PT, prothrombin time; INR, international normalized ratio; APTT, activated partial thromboplastin time; PT mix, prothrombin time mixing study; APTT mix, activated partial thromboplastin mixing time

On day 1 of hospital admission, the ecchymosis on the patient’s right breast was noted to be increasing in size from hospital day 0. On days 1 and 2 of admission, the patient was treated with multiple doses of oral and intravenous Vitamin K with no significant change in laboratory studies (Table 1). The patient continued to be hemodynamically stable. During this time, multiple ecchymoses on her right breast appeared, and the initial ecchymosis continued to increase in size. The hematology service was then consulted for further management.

On day 3 of hospitalization, mixing studies were ordered, which showed partial correction for both PT (corrected to 44.7 seconds [9.5-11.2 seconds]) and APTT (corrected to 84 seconds [22.0-29.0 seconds]). Factor VIII was 270 (64-189 %ACT). Factors II, VII, IX, and X were within normal limits. Disseminated intravascular coagulation (DIC) panel was negative, and thrombin time was within normal limits. FV activity was <5% (62-140%), and a standard Bethesda assay confirmed aFVı by the presence of an FV inhibitor with 30 inhibitor units (Table 1). The patient
continued to be stable hemodynamically and had unremarkable vital signs. There were increased ecchymoses bilaterally on her upper and lower extremities and at the sites of IV access. Treatment was begun with prednisone 100 mg PO (oral) daily on day 4 of hospitalization and rituximab 375 mg infusion was begun on day 5, with the plan for weekly infusions for a total of four weeks. The first infusion was begun inpatient, and the patient began to show symptoms of an infusion reaction; therefore, the infusion was stopped and finished the next day. The patient was discharged from the hospital hemodynamically stable with INR > 10, PT > 90 seconds (9.5-11.2 seconds), and APTT > 139 seconds (22-20 seconds) with plans to see hematology outpatient and continue with rituximab infusions weekly on an outpatient basis.

Since the first admission, the patient has been re-admitted twice, once for melenic stools and a second time for pulmonary embolism. The first readmission was four days following the patient’s initial discharge. Her labs on admission were significant for a hemoglobin level of 6.3 g/dL, red blood cell count of 2.2 x 10^6/μL, white blood cell count of 18.4 x 10^3/μL, and platelet count of 378 x 10^3/μL, and coagulation labs also were collected (Table 2). On day 1 of admission, the patient underwent an upper endoscopy, which showed no active source of bleeding. On day 4, FV activity was measured as 13% (62-140%). Labs at the time of discharge were obtained (Table 2). The patient was readmitted a second time for shortness of breath, which upon investigation was found to be a pulmonary embolism. On day 3, FV activity was at 106% (62-140%). She was discharged in stable condition on day 12 of admission after being treated symptomatically. The patient’s coagulation labs on admission and discharge were drawn (Table 3). The patient continues to see the hematology service in the outpatient clinic.

![Table 2: Selected Coagulopathy Labs of Second Admission](image)

| Day of admission | 0 | 7 |
|------------------|---|---|
| PT (9.5-11.2 seconds) | 62 | 12.7 |
| INR | 6.8 | 1.2 |
| APTT (22-29 seconds) | 77.7 | - |

**TABLE 2: Selected Coagulopathy Labs of Second Admission**

PT, prothrombin time; INR, international normalized ratio; APTT, activated partial thromboplastin time

![Table 3: Selected Coagulopathy Labs of Third Admission](image)

| Day of admission | 0 | 12 |
|------------------|---|---|
| PT (9.5-11.2 seconds) | 11.3 | 14.7 |
| INR | 1.1 | 1.4 |
| APTT (22-29 seconds) | 25.4 | - |

**TABLE 3: Selected Coagulopathy Labs of Third Admission**

PT, prothrombin time; INR, international normalized ratio; APTT, activated partial thromboplastin time
Discussion

aFVi is a rare hematologic disorder with widely variable presenting symptoms. Commonly associated conditions are autoimmune disorders (13%), cancer (22%), and the use of antibiotics (42%) including beta-lactams, aminoglycosides (especially streptomycin), tetracyclines, and fluoroquinolones (especially ciprofloxacin) [3]. As patients can range from clinically asymptomatic to experiencing serious life-threatening bleeding diathesis, laboratory tests are critical to making the diagnosis of aFVi, and the interpretation of the tests can be difficult. The most common lab identification of aFVi is with grossly prolonged PT and APTT, which are only partially corrected with mixing studies. The inhibitor can then be confirmed through further laboratory testing. A number of treatments have been reported to reverse coagulopathy in case reports. Corticosteroids are typically used with success, and, in recent years, immunomodulating medications such as rituximab have also been utilized [3].

A literature search of previous reviews was conducted. In their 2010 systematic review, Franchini and Lippi reviewed cases of aFVi not associated with bovine protein. Of the 154 total cases identified beginning from 1955, the authors found 78 cases not associated with bovine thrombin [3]. Of these cases 42% (n = 33) were associated with antibiotics. Among antibiotics, the most represented were beta-lactams and aminoglycosides [3].

In a subsequent review, Boland and Shreenivas used a similar approach to examine cases from 2010 to 2016. Again, cases involving bovine thrombin were excluded to reflect the switch to human recombinant thrombin. Boland and Shreenivas found an additional 47 cases [5]. Infections were the most commonly implicated cause (n = 16), whereas antibiotics were the second most associated cause of aFVi (n = 15). The most common antibiotic implicated were beta-lactams. Combined with the previous study by Franchini and Lippi, the total number of cases up to 2016 not involving bovine thrombin is 125; and the most commonly reported association is antibiotic use, which is present in 58% of all cases [5].

A review of case reports in the previous five years was conducted. To provide continuity with both of the previous systematic reviews, cases involving bovine serum were excluded. Additionally, any cases included in the previous 125 reports were excluded. Cases were found by using the search terms “acquired factor v inhibitors” and “acquired coagulation inhibitors” on PubMed. Initially, 28 new cases were found from 26 published case reports. Of these, nine were included in the Boland and Shreenivas’s review and excluded from the table. Overall, 12 new cases were found on PubMed from 2016 to 2020 after removing congenital causes of FV deficiency and articles without full text availability (Table 4). The most common association was with antibiotics (5 out of 12), with beta-lactams being implicated in all antibiotic-related cases (Table 4).

| Study          | Age (years)/gender | Hemorrhagic symptoms | Reported association            | Treatment                                                                 | Outcome                        |
|----------------|--------------------|----------------------|--------------------------------|---------------------------------------------------------------------------|--------------------------------|
| Andreadis et al. [6] | 78/M               | Ecchymosis of the left hemithorax | β-lactam, surgery             | Steroids, rituximab, cyclophosphamide, rFVIIa                             | Remission (did not recheck inhibitor levels) |
| Nakata et al. [7]    | 72/F               | None                 | unknown                        | Steroids, cyclophosphamide, plasma exchange                               | Remission                      |
| Quek et al. [8]        | 57/F               | Knee hemarthrosis    | Light chain myeloma            | Bortezomib, cyclophosphamide, Remission                                    |                                |
| Authors          | Sex | Age  | Bleeding Site / Cause                                      | Treatment               | Outcomes                                      |
|------------------|-----|------|------------------------------------------------------------|-------------------------|-----------------------------------------------|
| Fujita et al. [9] | M   | 82   | Gum bleeding                                               | Myelodysplastic syndrome, Steroids, platelet transfusions | Remission                                    |
| Ogawa et al. [10] | M   | 87   | Recurrent bleeding from hemodialysis sites                | β-lactam, fluoroquinolone, Steroids, plasma exchange | Death due to aspiration pneumonia and general debility four months after second hospitalization |
| Taniwaki et al. [11]| M   | 78   | None                                                      | β-lactam, fluoroquinolone, meropenem, micafungin, cefcapene pivoxil | Discontinued antibiotics                      |
| Li et al. [12]   | M   | 51   | 1) Oral mucosal bleeding 2) Intracranial hemorrhage       | 1) β-lactam 2) Idiopathic, 1) Steroids 2) FPP | 1) Remission 2) Remission                     |
| Li et al. [12]   | F   | 82   | None                                                      | Idiopathic              | FFP                                           |
| Sakatoku et al. [14] | M   | 80   | Nasal hemorrhage, hematuria                                | Prasugrel hydrochloride, Discontinued prasugrel, rFVIIa, steroids | Remission                                    |
| Mihara et al. [15] | M   | 74   | Subcutaneous hemorrhage on the left upper chest           | Idiopathic              | Steroids, cyclophosphamide                    |
| Yamanishi et al. [16] | M   | 84   | Subcutaneous hemorrhage                                   | β-lactam                | Steroids                                      |

**TABLE 4: Compiled Literature of Acquired Factor V Inhibitors from 2016 to 2020**

rFVIIa, recombinant factor VIIa; FPP, fresh frozen plasma

aFVi treatment involves two steps: first is to control symptomatic bleeding and second is to remove the inhibitor. Fresh frozen plasma (FFP), prothrombin complex concentrates (PCC), and platelet transfusions have been used to varying degrees of success [4]. In our case, we used vitamin K and PCC to control bleeding, which did not improve the patient’s worsening ecchymoses. The use of PCC and FFP generally are thought to have low utility in treatment because of their low levels of FV. A trend in recent literature is the use of platelet transfusions as first-line treatment to control acute bleeding. The rationale behind this is that roughly 20% of a human’s FV is stored in the platelets and platelets have shown to be relatively unaffected by FV inhibitors, possibly by the distinct features of plasma and platelet FV [17]. Ang et al. showed the ability of platelet concentrates to normalize FV in 11 out of 16 cases, whereas Boland and Shreenivas showed 7 out of 12 had normalization of FV with platelet...
concentrates [4]. We found in our literature review that 1 out of 12 cases utilized platelet transfusions in their treatment plan, which resulted in remission in the single case (Table 4).

The second step in treatment is to remove the inhibitor, which can be done by a variety of methods. High-dose intravenous immunoglobulin (IVIG) has been shown to decrease anti-FV inhibitor titers [18]. Boland and Shreenivas showed that 88% of patients who received immunosuppression with corticosteroids alone achieved remission [5]. Rituximab has also been utilized in addition to corticosteroids, and there have been positive results, including our presented case. However, it is hard to ascertain if these results would have been equally obtained with steroids alone [5]. We found in our literature review that steroids were utilized in 9 out of 11 cases in which medication was given as part of treatment, whereas only one of the cases utilized rituximab (Table 4).

Theorized pathophysiolog
A number of theories have been suggested for antibiotic-related aFVi. These include stress-based dysregulation of the immune system prior to antibiotic administration and simple homology between antibiotics and FV.

Prior research has indicated that the characteristics of the FVi (antibody) have been shown to best correlate with clinical bleeding risk rather than titers of the inhibitors. These characteristics include the inhibitor’s capacity to access FV stored in the alpha granules of platelets as well as to recognize the epitope in the C2 domain of the FV light chain [3]. Normally, FV acts by binding exposed procoagulant phospholipid-phosphatidylserine on activated platelets or endothelial cells through this C2 domain of the light chain [3]. This complex then binds to von Willebrand factor and enhances the coagulation process by activating factor X, which cleaves prothrombin to thrombin. Similarly, there is evidence of some acquired factor VIII inhibitors having inhibitors against the C2 domain of the light chain (as well as the A2 and A3 domains of the heavy chain), which suggests that there may be a similar mechanism causing both FVIII and FV inhibitor formation [19]. Interestingly, there is a strong body of evidence linking antibiotic classes (beta-lactams and aminoglycosides) strongly associated with acquired FV inhibitors to interactions with similar phospholipid residues [20]. In particular, many of the associated antibiotics are known to interact with phospholipids in the membranes of the bacterial cells they target. We suggest that in a heightened immunologic state of cancer, autoimmunity, or infection, the immune system may aberrantly produce antibodies against these antibiotics, which may themselves have some homology to the C2 region of the clotting factors’ light chains. However, further research into the inciting molecular and cyto-biologic mechanisms of aFVi formation is needed in light of many complex clinical presentations, a multitude of different associations, and a lack of in-depth antibody sequence analyses.

Conclusions
aFVi is a rare and potentially fatal coagulopathy that is thought to be underreported in the literature. There are a limited number of cases reported on this condition, with variable causes. We report this case to increase awareness among practicing clinicians of this condition. Furthermore, we compiled a summary of aFVi cases published in the past five years to better characterize the common causes of this coagulopathy. Finally, we hypothesize that stress-mediated immune dysregulation may contribute to antibody formation blocking interactions between the C2 epitope of FV and the damaged phospholipid membrane residues. Further sequencing analysis of the antibodies and homology analyses must be performed to confirm or refute our hypothesized mechanism.

Additional Information
Disclosures

Human subjects: Consent was obtained by all participants in this study. Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following:
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References

1. Knobl P, Lechner K: Acquired factor V inhibitors. Baillieres Clin Haematol. 1998, 11:305-318. 10.1016/s0950-3536(98)80050-4
2. Cheng CM, Meyer-Massetti C, Kayser SR: A review of three stand-alone topical thrombins for surgical hemostasis. Clin Ther. 2009, 31:32-41. 10.1016/j.clinthera.2009.01.005
3. Franchini M, Lippi G: Acquired factor V inhibitors: a systematic review. J Thromb Thrombolysis. 2011, 31:449-457. 10.1007/s11239-010-0529-6
4. Ang A, Kuperan P, Ng C, Ng H: Acquired factor V inhibitor: a problem-based systematic review. Thromb Haemost. 2009, 101:852-859. 10.1160/TH08-11-0767
5. Boland F, Shreenivas A: Acquired factor V inhibitors: a review of literature. Ann Hematol Oncol. 2017, 4:1168.
6. Andreadis P, Kafantari K, Agapidou A, Vakalopoulou S, Vlachaki E: Successful outcome of severe intra-cerebral bleeding associated with acquired factor V inhibition: utilization of multiple therapeutic agents. Balk Med J. 2018, 35:112-115. 10.4274/balkanmedj.2017.0158
7. Nakata K, Ueda S, Matsunaga H, et al.: High titer of acquired factor V Inhibitor presenting with a pseudo-deficiency of multiple coagulation factors. Intern Med J. 2018, 57:595-597. 10.2169/internalmedicine.9150-17
8. Quek J, Wong W, Tan C, Tha MH, Nagarajan C, Lee LH, Ng HJ: Acquired factor V deficiency in a patient with myeloma and amyloidosis. Thrombosis Research. Thromb Res. 2018, 164:1-3. 10.1016/j.thromres.2018.01.045
9. Fujita Y, Nakazato T, Ito C, Masuda K, Osada Y, Aisa Y, Mori T: A rare case of an acquired factor V inhibitor in a patient with myelodysplastic syndrome during azacitidine treatment. Ann Hematol. 2018, 97:2009-2010. 10.1007/s00277-018-3548-z
10. Ogawa H, Souri M, Kanouchi K, et al.: A high titer of acquired factor V inhibitor in a hemodialysis patient who developed arterial thrombosis. International J Hematol. 2019, 109:214-220. 10.1007/s12185-018-2561-9
11. Taniwaki M, Katsutani S, Yamasaki M, et al.: Acquired factor V inhibitor after antibiotic treatment in a patient with pneumonia: a case report. Ann Hematol. 2019, 98:1989-1990. 10.1007/s00277-019-05638-6
12. Li F, Shi K, Liu J, et al.: Acquired factor V inhibitor with symptoms and titer of inhibitor differences: report of two cases. Acta Haematol. 2019, 141:148-150. 10.1159/000496080
13. Meidert AS, Kinzinger J, Mohnle P, et al.: Case report perioperative management of a patient with severe factor V deficiency presenting with chronic subdural hematoma: a clinical report. World Neurosurg. 2019, 127:409-413. 10.1016/j.wneu.2019.04.080
14. Sakatoku K, Takakuwa T, Miura A, Araki T, Fujitani Y, Yamamura R: Acquired factor V inhibitor with hemorrhagic symptoms after prasugrel hydrochloride treatment [Online ahead of print]. Acta Haematol. 2019, 10.1159/000502406
15. Mihara M, Ogawa Y, Nagasaka M, et al.: Successful management of acquired factor v inhibitor by monitoring factor V activity, antigen, and inhibitor values during immunosuppressive therapy [Online ahead of print]. Acta Haematol. 2019, 10.1159/000502730
16. Yamanishi M, Nishimi Y, Usui M, Miki K, Imashuku S: Acquired factor V deficiency associated with CFPM administration. Clin Lab. 2019, 65:190240. 10.7754/clin.lab.2019.190240
17. Gould WR, Silveira JR, Tracy PB: unique in vivo modifications of coagulation factor V produce a physically and functionally distinct platelet-derived cofactor. J Biol Chem. 2004, 279:2383-2393. 10.1074/jbc.M308600200
18. de Raucourt E, Barbier C, Sinda P, Dib M, Peltier J, Ternisien C: High-dose intravenous immunoglobulin treatment in two patients with acquired factor V inhibitors. Am J Hematol. 2003, 190:187-190. 10.1002/ajh.10420

19. Barrow RT, Healey JF, Jacquemin MG, Lollar P: Antigenicity of putative phospholipid membrane-binding residues in factor VIII. Blood. 2001, 97:169-174. 10.1182/blood.v97.1.169

20. Epand RM, Walker C, Epand RF, Magarvey NA: Molecular mechanisms of membrane targeting antibiotics. Biochim Biophys Acta. 2016, 1858:980-987. 10.1016/j.bbamem.2015.10.018