Clinical equivalence with G-CSF biosimilars: methodologic approach in a (neo)adjuvant setting in non-metastatic breast cancer

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Received: 26 June 2017 / Accepted: 16 August 2017 / Published online: 20 September 2017
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Abstract Biosimilars are biological medicines that have been shown to be similar to a reference biological medicine that has already been approved for use. Development of biosimilars is based on a “totality of evidence” approach that involves a series of steps by which biosimilars must demonstrate similarity to a reference product in all aspects of the drug and eliminate any remaining uncertainties. Clinical studies are then considered confirmatory and are performed to show that there are no clinically meaningful differences compared with the reference product in a sensitive patient population. The recombinant human granulocyte colony-stimulating factor (G-CSF) biosimilar EP2006/Zarxio® (filgrastim-sdnz) became the first FDA-approved biosimilar in 2015. This review evaluates how clinical equivalence can be demonstrated with G-CSF biosimilars through the identification of “sensitive” study populations and endpoints. Patients with non-metastatic breast cancer treated in the (neo)adjuvant setting represent a potentially homogenous population, making this a suitable sensitive indication for assessing filgrastim and pegfilgrastim biosimilars compared with reference products. This review includes clinical trials of G-CSF biosimilars in breast cancer, focusing on key aspects of the trials that were necessary to accurately demonstrate clinical equivalence and enable extrapolation to relevant indications, based on guidelines and biostatistical principles.

Keywords Biosimilars · Filgrastim · Granulocyte colony-stimulating factors · Breast cancer · Extrapolation

Introduction

Biosimilars are biological medicines that have been shown to be similar to a reference biological medicine that has already been approved for use [1]. As patents for biological drugs expire, there are increased opportunities for the development of biosimilars and, as such, biosimilars are becoming increasingly available, particularly in oncology [2]. The development of biosimilars is significantly more complex than the development of small molecule generic drugs [2], but the principles for their development and approval, based on a “totality of evidence” approach, are now well established. Totality of evidence involves a series of steps by which biosimilars must demonstrate similarity to a reference product in all aspects of the drug and eliminate any remaining uncertainties [3]. This sequential process must include comparative structural and functional characterization, nonclinical evaluation, and clinical studies to compare human pharmacokinetic (PK) and pharmacodynamic (PD) data, clinical safety and immunogenicity data, and typically comparative clinical efficacy studies [1, 4]. This stepwise approach is essential because clinical studies are generally the least sensitive means to detect differences between two biological products. Clinical studies are nonetheless vital to confirm that there are no clinically meaningful differences in terms of biological activity, safety, and immunogenicity compared with the reference product [5].

In Europe, 12 biosimilars have been approved in oncology indications since the approval of the first biosimilar in this field, Binocrit® (epoetin alfa), in 2007 [6]. Although biosimilars have been slower to enter the US market, the recombinant human granulocyte colony-stimulating factor (G-CSF) biosimilar EP2006/Zarxio® (filgrastim-sdnz) became the first FDA-approved biosimilar in 2015 [7]. This review
Extrapolation

The European Medicines Agency (EMA) defines extrapolation as “extending information and conclusions available from studies in one or more subgroups of the patient population (source population) … to make inferences for another subgroup of the population (target population), or condition or product, thus reducing the need to generate additional information… to reach conclusions for the target population” [8]. Extrapolation must be scientifically justified in order to support a determination of biosimilarity for each additional indication; this is dependent on multiple factors that must be consistent across each indication, including similarity in structural and functional properties, the knowledge of the mechanism of action, similar PK and bio-distribution, immunogenicity, and expected toxicities [4].

An important consideration regarding extrapolation is that it is already an established scientific principle in drug regulation, for example, in a case of a major change of a manufacturing process, or when data from intravenous formulations are extrapolated to a new subcutaneous formulation [3]. Furthermore, extrapolation is also well accepted when a drug has been evaluated in a randomized clinical trial setting with strict inclusion and exclusion criteria, and then is extrapolated to real-world patients outside the restrictions of a clinical trial.

Guidelines for biosimilar development stipulate that in order to permit extrapolation, the clinical data collected using the totality of evidence approach must be in a “sensitive indication” [1, 2, 4] (Table 1). A sensitive indication is a patient population in which the treatment being assessed has a large effect on the relevant endpoint so that a difference between biosimilar and reference product will most easily be detected. Likewise, an immunocompetent study population is required to allow the meaningful evaluation of immunogenicity [1, 4].

Epoetin is a good example of selecting a sensitive indication in biosimilar development, which enables extrapolation to other indications. Binocrit® (epoetin alfa biosimilar) has been approved in Europe since 2007 [6] for the treatment of chemotherapy-induced anemia and renal anemia [9]. Patients with renal anemia without any major complications or comorbidities that may alter the response to epoetin offer a sensitive population to assess biosimilarity since potential differences in efficacy between the reference and biosimilar may be more easily shown in this population rather than in cancer patients undergoing chemotherapy who may be immunosuppressed and have variable responses to epoetin [10]. Furthermore, this indication was particularly relevant since renal anemia patients are the population at risk of developing pure red cell aplasia (PRCA) with epoetin treatment and no cases of PRCA have been reported in oncology patients receiving epoetin [3]. Extrapolation of the use of epoetin from renal anemia to cancer patients was scientifically justified since the biological effect is mediated by the same mechanism of action. The totality of evidence approach established that biosimilar epoetin alfa is similar to the reference medicine and, as such, extrapolation of renal anemia to chemotherapy-induced anemia was permitted [9].

Rationale for (neo)adjuvant breast cancer as a sensitive indication for assessment of G-CSF biosimilarity

G-CSF has numerous indications, including reduction in neutropenia/febrile neutropenia in patients receiving cytotoxic chemotherapy; mobilization of peripheral blood progenitor cells; treatment of severe congenital, cyclic, or idiopathic neutropenia; and treatment of persistent...
neutropenia in HIV patients [11–16]. However, G-CSF acts via the same mechanism of action across all associated patient populations, through selective binding of the G-CSF receptor [5, 17]. Therefore, if a study directly compares reference and biosimilar G-CSF in a sensitive population and demonstrates similarity, this supports extrapolation across all indications as part of the totality of evidence concept [3, 18].

Selection of a sensitive population in which to investigate potential differences between a reference medicine and a proposed biosimilar includes identification of a homogeneous population. Homogeneous populations allow any difference in response between reference and biosimilar to be attributed to product characteristics and reduce the likelihood that it is due to individual variation [19]. Increased homogeneity within a population contributes to increased sensitivity, allowing more accurate assessment of similarity compared with heterogeneous populations.

One of the indications for G-CSF is to decrease the risk of febrile neutropenia in patients with non-myeloid malignancies undergoing chemotherapy. Within this indication, patients receiving (neo)adjuvant treatment for breast cancer can be considered a sensitive cohort in which to assess biosimilar compared with reference G-CSF since it provides a homogenous patient population [18–20]. A key feature of this homogeneity is that unlike patients with metastatic breast cancer, patients with (neo)adjuvant disease have not received prior chemotherapy. This means that they exhibit less inter-patient variation in terms of potential for treatment-related toxicity and other confounding factors such as disease burden, location of metastases, and phenotype of metastatic cells [21]. This also means that patients with (neo)adjuvant breast cancer are, in general, representative of breast cancer patients worldwide, provided disease and treatment characteristics are similar [18–21]. Furthermore, these patients have not yet received treatment that likely differs from region to region. In addition, unlike previously treated patients, (neo)adjuvant patients have not experienced previous chemotherapy-induced immunosuppression and, as such, are a more sensitive population in which to assess risk of immunogenicity.

TAC (docetaxel, doxorubicin, and cyclophosphamide) chemotherapy is recommended in international treatment guidelines as one of the standard (neo)adjuvant chemotherapy regimens for patients with breast cancer due to its documented efficacy [22]. TAC has a proven dose-limiting hematological toxicity with grade 3–4 neutropenia in 65.5% patients [23], a median duration of severe (grade 4) neutropenia (DSN) of 7 days [24], and febrile neutropenia reported in 24–34% of patients [23–26] without G-CSF support. Treatment guidelines require primary prophylaxis with G-CSF as supportive care for TAC chemotherapy [27–29] with a proven substantial effect in this setting, reducing mean DSN to 1.4 days (95% confidence interval [CI] 1.1, 1.7) [30].

### Demonstrating clinical equivalence of G-CSF during randomized controlled trials in (neo)adjuvant breast cancer

Endpoints measured are a key consideration when planning confirmatory clinical studies comparing biosimilar and reference medicines. Sensitive endpoints should assess biological activity of the proposed biosimilar, as opposed to treatment outcomes, to allow similarity to be assessed more accurately [1]. DSN can be considered a sensitive endpoint in assessing biosimilarity of G-CSF in (neo)adjuvant breast cancer. Due to its dependence on G-CSF efficacy, any variation in DSN between homogeneous treatment groups can be considered to be a direct consequence of differences between activity of reference and biosimilar rhG-CSF. This sensitivity compared with other endpoints (e.g., infections, febrile neutropenia) can also be attributed to its continuous nature and frequent repeat sampling. Furthermore, risk of infection is directly proportional to severity and duration of neutropenia [31], making DSN a clinically relevant endpoint.

Clinical studies designed to assess potential differences between a reference medicine and a proposed biosimilar are typically designed to show equivalence of the two treatments. Equivalence in this sense means that the efficacies of the two products under assessment are similar to the extent that neither could be considered superior or inferior to the other [32]. In equivalence trials, the objective is to demonstrate that the

| Hypothesis to be evaluated: \( \mu_t - \mu_b \leq d \) days vs H1: \( \mu_t - \mu_b > d \) days |
|---------------------------------------------|------------------|------------------|------------------|------------------|
| Assumptions: CI, 97.5%; expected difference in means 0.25 days; randomization 1:1 |
| Power | 80% | 90% |
| Non-inferiority limit | 1 day | 0.5 days | 1 day | 0.5 days |
| Standard deviation | 1.5 | 2 | 1.5 | 2 | 1.5 | 2 |
| N per group | 24 | 42 | 64 | 113 | 32 | 55 | 86 | 151 |
| Paper | Reference G-CSF | Proposed biosimilar G-CSF | Patients randomized to treatment | Primary endpoint | Study power/sample size assumptions |
|-------|----------------|--------------------------|---------------------------------|-----------------|-----------------------------------|
| Holmes FA, et al. J Clin Oncol. 2002;20(3):727–31 [35] | Filgrastim (Neupogen®, Amgen) | Pegfilgrastim (Neulasta®, Amgen) | N/A | N = 310 (filgrastim, N = 156; pegfilgrastim, N = 154) | Mean DSN in cycle 1, (number of days with grade 4 neutropenia with an ANC < 0.5 x 10^9/L) | Equivalence was assessed using an upper 97.5% one-sided confidence interval limit (95% two-sided) with an equivalence limit of < 1 day for the difference in mean DSN. Based on similar assumptions for the test, this would mean 90% power. |
| Holmes FA, et al. Ann Oncol. 2002;13:903–9 [36] | Filgrastim (Neupogen®, Amgen) | Pegfilgrastim (Neulasta®, Amgen) | N/A | N = 154 (filgrastim, N = 25; pegfilgrastim, N = 129) | Mean DSN in cycle 1, (number of days with grade 4 neutropenia with an ANC < 0.5 x 10^9/L) | Equivalence was assessed using a two-sided 95% confidence interval with an equivalence limit of ± 1 day for the difference in mean DSN. For the dose groups comparison, assumptions presented gave a power of about 80% per group comparison with no adjustment of alpha. Sample size was based on a non-inferiority design, and the mean duration and standard deviation (SD) of severe neutropenia observed in the phase II study. Equivalence was assessed using an upper 97.5% one-sided confidence interval limit (95% two-sided) with an equivalence limit of < 1 day for the difference in mean DSN. The phase II publication only presented confidence intervals (not mean and SD) but it seems likely that 90% power was used. |
| Green MD, et al. Ann Oncol. 2003;14:29–35 [34] | Filgrastim (Neupogen®, Amgen) | Pegfilgrastim (Neulasta®, Amgen) | N/A | N = 157 (filgrastim, N = 77; pegfilgrastim, N = 80) | Mean DSN in cycle 1, (number of days with grade 4 neutropenia with an ANC < 0.5 x 10^9/L) | Equivalence was assessed using a non-inferiority design, and the mean duration and standard deviation (SD) of severe neutropenia observed in the phase II study. Equivalence was assessed using an upper 97.5% one-sided confidence interval limit (95% two-sided) with an equivalence limit of < 1 day for the difference in mean DSN. The number of patients per treatment group was not based on formal statistical sample size calculations but was considered adequate to allow for determination of an optimal pegfilgrastim dose for phase III clinical studies. Comparison of DSN to show non-inferiority was performed post hoc and the study was not powered for this analysis but 95% confidence limits were used for the difference in the adjusted mean DSN. Based on similar assumptions for the test, this would mean 90% power. |
| Del Giglio A, et al. BMC Cancer 2008;8:332 [37] | Filgrastim (Neupogen®, Amgen) | Filgrastim (XM02, Teva) | N = 348 (reference filgrastim, N = 140; biosimilar filgrastim, N = 136; placebo N = 72) | Mean DSN in cycle 1, (number of days with grade 4 neutropenia with an ANC < 0.5 x 10^9/L) | Equivalence was assessed using a two-sided 95% confidence interval with an equivalence limit of ± 1 day for the difference in mean DSN. Based on similar assumptions for the test, this would mean 90% power. |
| Waller CF, et al. Onkologie. 2010;33(10):504–11 [38] | Filgrastim (Neupogen®, Amgen) | Filgrastim (Nivestim®, Hospira) | N = 279 (reference filgrastim, N = 95; biosimilar filgrastim, N = 184) | Mean DSN (ANC < 0.5 x 10^9/L) in treatment cycle 1 | Equivalence was assessed using a two-sided 95% confidence interval with an equivalence limit of ± 1 day for the difference in the adjusted mean DSN. Based on similar assumptions for the test, this would mean 90% power. |
| Buchner A, et al. Breast Cancer Res Treat. 2014;148:107–16 [39] | Pegfilgrastim (Neulasta®, Amgen) | Lipegfilgrastim (XM22, Teva) | N = 208 (pegfilgrastim, N = 54; lipegfilgrastim 3 mg, N = 53; lipegfilgrastim 4.5 mg, N = 51; lipegfilgrastim 6 mg, N = 50) | Mean DSN (number of consecutive days from ANC < 0.5 x 10^9/L to ≥ 0.5 x 10^9/L) during cycle 1 | The number of patients per treatment group was not based on formal statistical sample size calculations but was considered adequate to allow for determination of an optimal lipegfilgrastim dose for phase III clinical studies. Comparison of DSN to show non-inferiority was performed post hoc and the study was not powered for this analysis but 95% confidence limits were used for the difference in mean duration with an equivalence margin of ± 0.5 day. For the dose groups comparisons, assumptions presented led to a power of about 80% per group comparison with no adjustment of alpha. Assuming an expected mean difference in DSN of 0.25 days with a common standard deviation of 1.5 days, a 10% dropout rate and a | |
| Blackwell K, et al. Ann Oncol. 2015;26:33–40 [40] | Filgrastim (EP2006, Sandoz) | | N = 218 (reference filgrastim, N = 54; biosimilar filgrastim, | Mean DSN (number of consecutive days from ANC < 0.5 x 10^9/L to ≥ 0.5 x 10^9/L) during cycle 1 | |
Table 3 (continued)

| Paper | Reference G-CSF | Proposed biosimilar G-CSF | Patients randomized to treatment | Primary endpoint | Study power/sample size assumptions |
|-------|----------------|---------------------------|---------------------------------|-----------------|------------------------------------|
| 1948–53 [18] | Filgrastim (Neupogen®, Amgen) | Filgrastim (Neupogen®, Amgen) | N = 54; reference then biosimilar, N = 55; biosimilar then reference, N = 55 | days from ANC < 0.5 × 10^9/L to ≥ 0.5 × 10^9/L during cycle 1 | non-inferiority margin of −1 day; 96 patients per group (biosimilar or reference) were required to have 90% power for showing non-inferiority based on a one-sided 97.5% CI. |
| Blackwell K, et al. Oncologist 2016; 21:789–94 [40] | Pegfilgrastim (Neulasta®, Amgen) | Pegfilgrastim (LA-EP2006, Sandoz) | N = 308 (reference pegfilgrastim, N = 153; biosimilar pegfilgrastim, N = 155) | Mean DSN (number of consecutive days from ANC < 0.5 × 10^9/L to ≥ 0.5 × 10^9/L) during cycle 1 | A sample size of 302 patients was considered sufficient to achieve 90% power for testing of equivalence (two one-sided tests) at the 2.5% significance level, assuming no difference in mean DSN between treatments with a common SD of 1.6 days. |
| Harbeck N, et al. Future Oncol. 2016;12(11): 1359–67 [41] | Pegfilgrastim (Neulasta®, Amgen) | Pegfilgrastim (LA-EP2006, Sandoz) | N = 316 (reference pegfilgrastim, N = 149; biosimilar pegfilgrastim, N = 146) | Mean DSN (number of consecutive days from ANC < 0.5 × 10^9/L to ≥ 0.5 × 10^9/L) during cycle 1 | A sample size of 302 patients was considered sufficient to achieve 90% power for testing of equivalence (two one-sided tests) at the 2.5% significance level, assuming no difference in mean DSN between treatments with a common standard deviation (SD) of 1.6 days. |
| Hegg R, et al. Clinics (Sao Paulo). 2016;71(10):586–92 [42] | Filgrastim (Granulokine®, Roche) | Filgrastim (Fiprimas®, Eurofarma) | N = 217 (reference filgrastim, N = 108; biosimilar filgrastim, N = 109) | Rate of grade 4 neutropenia during cycle 1 | Sample size was calculated using the historical incidence of grade 4 neutropenia after the first chemotherapy cycle. This was 73–83% in studies involving the two eligible chemotherapy regimens. Considering a one-tailed alpha of 5% and a statistical power of 80% for the study to obtain a maximum absolute difference of 15% in the rate of grade 4 neutropenia between the two groups, and assuming that this rate would be 80% in the control group, 88 patients should be included in each study group. Assuming a dropout rate of approximately 20%, 110 patients were anticipated in each arm, for a total of 220 patients. |
| Waller CF et al. 2017. European CanCer Organization (ECCO) 2017 European Cancer Congress. January 27–30, 2017, Amsterdam, Netherlands [43] | Pegfilgrastim (Neulasta®, Amgen) | Pegfilgrastim (MYL-1401H, Mylan) | N = 164 (reference pegfilgrastim, N = 67; biosimilar pegfilgrastim, N = 127) | DSN in cycle 1 (days with absolute neutrophil count [ANC] < 0.5 × 10^9/L) | Equivalence was assessed using a two-sided 95% confidence interval with an equivalence limit of ±1 day for the difference in the adjusted mean DSN. Based on similar assumptions for a test, this would mean 90% power. |

The table also includes RCTs comparing reference pegfilgrastim with reference filgrastim in patients with breast cancer. Neupogen® and Neulasta® are registered trademarks of Amgen; Nivestim® is a registered trademark of Hospira; Granulokine® is a registered trademark of Amgen/Roche; Fiprimas® is a registered trademark of Eurofarma. N/A not applicable.
biosimilar (b) is not meaningfully different to the reference (r), in terms of an endpoint (μ):

**Null hypothesis (the therapies are not equivalent)**

\[ |μ_r - μ_b| ≥ Δ \]

**Alternative hypothesis (the therapies are equivalent)**

\[ |μ_r - μ_b| < Δ \]

where Δ represents the equivalence margin, defined as “the maximum tolerable difference considered to be clinically acceptable” [32].

When assessing biosimilarity, it is essential that a clinically relevant and meaningful equivalence margin is established, i.e., the range over which the efficacies can be considered equivalent [32]. Identification of an appropriate equivalence margin is dependent on the specific characteristics of the reference product and its therapeutic class [19]. Therapeutic equivalence is concluded if the 95% confidence interval is completely contained within the equivalence margin. This is statistically equivalent to calculating two independent one-sided tests at a 2.5% alpha level (one in each direction), of which both have to be successful [33].

A second key consideration when assessing biosimilarity is ensuring that the trial is sufficiently powered to avoid making a type II error, i.e., incorrectly claiming that there is no difference between two treatment groups. This is dependent on factors including level of type I error (typically \( p = 0.05 \)), level of type II error (\( p = 0.10 \) or 0.20), standard deviation (estimated from published or preliminary data), an estimation of the true value of \( μ_r - μ_b \), and the equivalence margin [33]. Based on these biostatistical considerations, calculations were performed to identify an appropriate equivalence margin and sample size necessary to assess clinical equivalence in DSN between reference and biosimilar G-CSF in patients with (neo)adjuvant breast cancer (Table 2). Using these calculations, it can be established that at a significance level of 0.05%, a power of 90%, an equivalence limit of 0.5 days difference in DSN, and a standard deviation of 1.5, 86 patients are required per treatment group in order to robustly assess equivalence.

Patients with breast cancer represent a sensitive population for clinically evaluating all G-CSF medicines, including biosimilars. In accordance with these considerations, multiple randomized controlled trials (RCTs) have been conducted to demonstrate equivalence between biosimilar and reference G-CSF in breast cancer (Table 3). Clinical studies were also performed to compare reference pegfilgrastim with reference filgrastim in patients with breast cancer [34–36]. Sensitive endpoints include DSN, incidence of hospitalization due to febrile neutropenia; incidence of infections; depth and time of absolute neutrophil count (ANC) nadir; and time to ANC recovery [5, 18].

Based on the totality of evidence provided, and the use of sensitive patient populations, studies demonstrating clinical equivalence of biosimilar G-CSF in breast cancer can be extrapolated to support clinical equivalence with reference filgrastim in other tumor types and indications. Given the availability for clinically relevant PD parameters for G-CSF treatment (ANC, CD34+ cell count), highly sensitive PK/PD studies can waive the need for a comparative phase III trial for regulatory approval including full extrapolation in Europe under certain circumstances. For example, following demonstration of comparability in structural and functional attributes and in PK/PD characteristics compared with reference filgrastim in healthy volunteers, with a confirmatory safety single-arm phase III trial in patients with breast cancer, the biosimilar filgrastim Zarzio® was approved by the EMA for the same indications as reference biosimilar [13, 16]. In the USA, the FDA requested an additional randomized controlled clinical trial. Therefore, and following a further head-to-head comparator study in patients with breast cancer vs reference filgrastim [18], Zarzio® (marketed as Zarzio® in the USA) was subsequently approved for the same indications by the FDA [5].

This approach, taken to confirm the equivalence of reference and biosimilar G-CSF in a sensitive population, is now being used to show equivalence between biosimilar and reference pegfilgrastim. To date, two confirmatory phase III trials, PROTECT-1 and PROTECT-2, have provided evidence of therapeutic equivalence according to the abovementioned sensitive endpoints in a total of 622 patients with (neo)adjuvant breast cancer [40, 41]. However, regulatory authorities have determined that further trials are necessary to address unanswered questions, such as a potential lack of equivalence in the concentrations of pegfilgrastim compared with biosimilar pegfilgrastim in blood [44].

### Conclusions

Using the rigorous “totality of evidence” approach, clinical equivalence between reference and biosimilar products can be established in a single sensitive population and reliably extrapolated to further indications. (Neo)adjuvant, non-metastatic breast cancer is a suitable sensitive patient population for assessing filgrastim and pegfilgrastim biosimilars compared with reference products.

### Acknowledgements

Editorial support was provided by Caroline McGown of the Spirit Medical Communications Ltd., supported by the Sandoz GmbH, Kundl, Austria. Final approval of the manuscript rested solely with the scientific authors.
Funding  This work was supported by the Sandoz GmbH, Kundl, Austria. Authors received no monetary compensation in return for their work on this manuscript.

Compliance with ethical standards

Conflict of interest  Matti Aapro: Amgen (honoraria, expert testimony), Hexal AG (employment), Sandoz (consultancy, research funding), Hospira (consultancy, research funding), Teva (consultancy, research funding), Merck (KGAa, advisory role), Merck (advisory role), Sandoz (advisory role, research funding), Pierre Fabre Medicament (advisory role, research funding), Vifor Pharma (advisory role, speakers bureau). Nadia Hoebel: Hexal AG (employment). Andriy Krendyukov: Hexal AG (employment).

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