Human menstrual blood-derived stem cells as immunoregulatory therapy in COVID-19: A case report and review of the literature

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Abstract

BACKGROUND
The coronavirus disease 2019 (COVID-19) caused by novel coronavirus 2019 in December 2019 has spread all around the globe and has caused a pandemic. There is still no current effective guidance on the clinical management of COVID-19. Mesenchymal stem cell therapy has been shown to be one of the therapeutic approaches to alleviate pneumonia and symptoms through their immunomodulatory effect in COVID-19 patients.

CASE SUMMARY
We describe the first confirmed case of COVID-19 in Hangzhou to explore the role of human menstrual blood-derived stem cells (MenSCs) in the treatment of COVID-19. Moreover, we review the immunomodulation effect including non-specific and specific immune functions of MenSCs for the therapy of COVID-19.

CONCLUSION
MenSCs can be helpful to find a promising therapeutic approach for COVID-19.

Key Words: COVID-19; Human menstrual blood-derived stem cells; Immunoregulatory therapy; Inflammatory response; Cytokine storm; Case report

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Core Tip: The coronavirus disease 2019 (COVID-19) is the word that certainly will not be forgotten by everybody who lives in the first half of the twenty-first century. It has led many researchers from different biomedical fields to find solutions or treatments to manage the pandemic. However, there is still no current effective guidance on the clinical management of COVID-19. Mesenchymal stem cells are widely used to treat tissue and organ injuries with effective immunomodulatory and repair capacities, which makes them ideal for allogenic adoptive transfer therapy. In this study, we describe the first confirmed case of COVID-19 in Hangzhou, China to explore the role of human menstrual blood-derived stem cells (MenSCs) in the treatment of COVID-19. Moreover, we review the immunomodulation effect including non-specific and specific immune functions of MenSCs for the therapy of COVID-19, which can be helpful to find a promising therapeutic approach for this disease.

INTRODUCTION
The outbreak of coronavirus disease 2019 (COVID-19) caused by novel coronavirus 2019 (2019-nCoV) in late December 2019 is increasing rapidly in an epidemic scale and has spread to over 200 countries[1,2]. It has rapidly transmitted and become a major concern all over the world. The epidemiology, clinical characteristics, and treatment of COVID-19 have been reported in many cases by many institutions[3]. However, there is still no current effective guidance from the World Health Organization on the clinical management of COVID-19[4].

Mesenchymal stem cells (MSCs) are widely used to treat tissue and organ injuries, with effective immunomodulatory and repair capacities and low immunogenicity. This makes them ideal for allogenic adoptive transfer therapy[5,6]. Human menstrual blood-derived stem cells (MenSCs) have become a promising alternative because they are easy to collect and isolate and do not involve ethical considerations[7,8]. Previously, our group revealed that MenSCs were effective for treating liver failure and lung injury[9,10]. Here, we present the first case of COVID-19 identified in Hangzhou, China, on January 19, 2020, and its treatment, including MenSC therapy, and review the immunoregulatory effect of MenSC in treatment of COVID-19.

CASE PRESENTATION

Chief complaints
A low-grade fever and fatigue accompanied by dizziness for 2 d.

History of present illness
On January 19, 2020, a 32-year-old man was referred to our hospital with a low-grade fever and fatigue accompanied by dizziness for 2 d. Given the COVID-19 outbreak in Wuhan and his symptoms, the local hospital immediately admitted him with suspected COVID-19. He was immediately placed in a quarantine ward and underwent examination. Sputum and throat swab specimens were collected at admission and tested by reverse transcription-polymerase chain reaction for SARS-Cov-2 RNA. The infection was confirmed the day of admission, in accordance with China Centers for Disease Control guidance.

History of past illness
He had no other underlying diseases. He worked and lived in Hangzhou, but had travelled to Wuhan for business one week earlier and contacted two colleagues there.
Personal and family history
There was no noteworthy personal or family medical history.

Physical examination
After hospitalization, the patient’s chief symptoms were an occasional cough, shortness of breath, and chest pain. On physical examination, he had short rough breaths. His temperature was elevated to 38.7 °C, with an arterial oxygen tension (PaO$_2$) of 82 mmHg under ambient air.

Laboratory examinations
Laboratory examinations showed normal leukocytes ($9.3 \times 10^9$/L), neutrophils ($6 \times 10^9$/L), and lymphocytes ($2.3 \times 10^9$/L). The patient’s high-sensitivity C-reactive protein (hs-CRP) level was 10.8 mg/L. Laboratory testing showed sharply increased leukocytes ($25.9 \times 10^9$/L) and neutrophils ($23.2 \times 10^9$/L), and an hs-CRP level of 43.5 mg/L. On day 7, his fever reached 39.4 °C and he developed chest tightness with a PaO$_2$ of 68 mmHg, PaCO$_2$ of 34 mmHg, and oxygenation index of 208 mmHg on nasal oxygen at 3 L/min. The levels of inflammatory cytokines were also increased, with interleukin (IL)-6 71.5 pg/mL, IL-10 7.75 pg/mL, tumor necrosis factor (TNF)-α 84.28 pg/mL, and TNF-γ 34.71 pg/mL on day 11. The sputum 2019-nCoV RNA test turned negative for the first time, although a repeat test was positive on day 20. The leukocyte count ($14.6 \times 10^9$/L), neutrophil count ($10.4 \times 10^9$/L), and inflammatory factor levels were all improved. Liver function tests showed elevated alanine aminotransferase (ALT, 190 U/L) and aspartate aminotransferase (AST, 41 U/L) levels on day 23.

Imaging examinations
The initial chest computed tomography (CT) scan showed a few interstitial changes in both lungs and ground-glass opacities (GGOs) in the subpleural area of the right lower lobe (Figure 1A). Subsequent chest CT showed multifocal peripheral patchy areas of nodular consolidation and new GGOs in the left subpleural area on day 7. Progressive resolution of the parenchymal lesions was seen on follow-up CT, which showed basilar streaky opacities and patchy consolidations, but worsened GGOs in both lungs (Figure 1B) on day 11. Chest CT on day 17 revealed improvement of the infiltrates in both lungs (Figure 1C). On day 24, the patient was discharged on glycyrrhizin tablets for liver protection. No abnormalities were observed on chest CT 9 d post-discharge (Figure 1D).

MULTIDISCIPLINARY EXPERT CONSULTATION
Given the severe pulmonary injury caused by the inflammatory response and side effects, the glucocorticoid, antiviral, and antibiotic therapies were withdrawn. Under the guidance of a specialist group, MenSC therapy was proposed. The therapy was discussed and approved by the hospital ethics committee and the patient and family members provided informed consent before the therapy. Intravenous MenSCs were given at 3000, 2000, and 3000 U on hospital days 11, 12, and 14, respectively; no adverse events were observed in association with the infusion. Over the following 3 d, the patient’s breathing improved, with an intermittent dry cough and decreased chest stuffiness. His temperature decreased to 37.4 °C.

FINAL DIAGNOSIS
The final diagnosis of the presented case was COVID-19 caused by 2019-nCoV.

TREATMENT
The patient was given supportive care including immunoglobulin and antiviral treatment at the first stage of hospitalization. Supportive treatment was strengthened, and methylprednisolone and vaccination were given as empirical treatment during the progression of the disease. Finally, MenSC therapy was proposed while the glucocorticoid, antiviral, and antibiotic therapies were withdrawn.
Figure 1 Chest computed tomography images of the patient with coronavirus disease 2019 in different stages of illness. A: Interstitial changes in both lungs and ground-glass opacities (GGOs) in the subpleural area of the right lower lobe on January 19 (Illness day 1); B: Worsening basal streaky opacities, patchy consolidations, and GGOs in both lungs on January 30 (Illness day 12); C: Significant absorption of patches infiltrating in both lungs on February 4 (Illness day 17); D: Almost no abnormality with a few fibrotic changes left in both lungs on February 19.

OUTCOME AND FOLLOW-UP
The condition of the patient was significantly improved. Regular follow-up revealed that no abnormalities or recurrences were observed on the chest CT after the patient’s discharge (Supplementary Figure 1).
DISCUSSION

This was the first confirmed case of COVID-19 in Hangzhou, Zhejiang Province, China. Most of the initial cases had a history of exposure in the epidemic area and were infected with the virus via human-to-human transmission[14,15]. The main clinical manifestations of COVID-19 infection are cough, fever, fatigue, and gradual dyspnea in some cases and acute respiratory distress syndrome in severe cases, with a 1–14-d period from onset to admission[21]. Our patient developed a mild fever and fatigue 4 d after contact in the epidemic area. As his illness worsened, he developed obvious respiratory symptoms, including a cough and chest discomfort. With treatment, his temperature normalized and the other complaints were relieved.

Our patient showed changes in routine laboratory parameters and inflammatory cytokines consistent with other reports[22,23]. The elevated ALT and AST levels observed late in the patient’s illness were thought to be adverse outcomes of systemic inflammation due to the virus[21,24]. Of note, the stool specimen collected on day 23 was negative for 2019-nCoV RNA, consistent with the sputum specimens[25–27].

Chest CT can help diagnose patients suspected of having COVID-19[19]. Similar to viral pneumonia due to other etiologies, GGOs are the main CT findings in COVID-19. The initial CT showed basilar streaky opacities and GGOs in the right lower lobe[28–30]. As the illness progressed, the size and density of these GGOs or paving patchy consolidations increased. Only slight fibrotic changes remained when the patient recovered.

The most effective medical treatment against COVID-19 remains unknown, though several antiviral therapies were confirmed to be effective in published studies[31,32]. Antibiotic was also adopted to prevent secondary bacterial infection in this case. Apart from the large supportive treatment, stem cell therapy was considered with no side effect after three times of intravenous administration. After the third administration, inflammatory cytokines such as IL-6 and IL-10 decreased significantly. Moreover, patchy consolidations were absorbed in both lungs, accompanied with the gradually well-improved clinical conditions.

The superiority of MSC therapy over therapies based on bone marrow, adipose, umbilical cord, or embryonic tissue is due to the easily accessible source of the cells and their high rate of proliferation, the low invasiveness of the procedure, and the absence of ethical issues[23,24]. MenSCs can be isolated from female uterine blood with readily accessible materials, free of trauma or ethical concerns, and are thus of great potential for treating diseases[35,36]. In vitro, MenSCs can be differentiated into ectoderm and mesoderm, specifically, into fat, bone, cartilage, nerve, and endothelioid cells[37,38]. The expression by MenSCs of OCT-4, specific embryonic antigen SSEA-4, C-kit, and other embryonic stem cell markers suggests that MenSCs are more primitive and have a stronger multidirectional differentiation potential than other MSCs[25–27]. Moreover, with their strong paracrine and angiogenic potential, they may be useful in the repair of damaged tissues[39].

MSCs interact with inhibitory T cells, B cells, natural killer (NK) cells, and regulatory T cells to regulate the immune response, which also have direct effects on NK cells[26–30]. Recent studies indicate that acute respiratory distress syndrome (ARDS), characterized by acute inflammation and edema of the alveolar epithelial cells, is accompanied by the massive release of inflammatory cytokines, such as IL-1, IL-8, interferon, and TNF, which activate macrophages or NK cells, which in a “cytokine storm” trigger a strong immune response to damage alveoli[39]. COVID-19 is associated with abundant complications, with ARDS as the main cause of death[33,34]. By inhibiting over-activated NK cells, MenSCs could confer protection against the highly inflammatory environment while maintaining the NK activities that reduce inflammation during healing. Moreover, immunoregulation by MenSCs via IL-6 and IL-10 has been reported as well[25,26,27].

MenSCs are significant inhibitors of the inflammatory response[27,28]. Their ability to inhibit T lymphocyte proliferation results in stronger immunomodulatory effects than those exhibited by human umbilical cord mesenchymal stem cells or bone marrow mesenchymal stem cells (BM-MSCs)[39,40]. The inhibition of T lymphocyte proliferation by MenSCs is mediated by their secretion of prostaglandin (PGE2) and indoleamine2,3-dioxygenase (IDO)[41]. PGE2 contributes to the transformation of classically activated macrophages (M1) to replace activated macrophages (M2) and may therefore be of interest in ameliorating the over-active inflammatory response in COVID-19[41]. It has also been shown that MenSCs promote the proliferation of CD4+ T lymphocytes in a density-dependent manner that is not affected by the concentration of IDO, in contrast to BM-MSCs, which are sensitive to IDO[42].

In addition to interleukins or PGE2, MenSCs release hepatocyte growth factor,
granulocyte-macrophage colony stimulating factor, and keratinocyte growth factor (KGF), which may protect respiratory epithelial cells in lung injury via activities that include the increased clearance of alveolar fluid, the promotion of endothelial repair, and the inhibition of the inflammatory response. MenSCs downregulate caspase-3 and IL-1β expression and upregulate KGF expression to improve microvascular permeability. In turn, KGF activates alveolar type II epithelial cells to stimulate the synthesis of pulmonary surfactant.

These abilities of MenSCs suggest a capacity to regulate the inflammatory response and thereby overcome the severe cytokine storm associated with COVID-19, while restoring the normal function of immune cells and tissues. Based on their homing potential, MenSCs may enhance the repair properties of immune and other cells, thus allowing the reconstruction of damaged tissues by receptor-mediated interactions. By protecting alveolar epithelial cells and improving the pulmonary microenvironment, MSCs can contribute to preventing lung dysfunction and the development of COVID-19 pneumonia. Through mediators such as interleukin receptors, PGE2, and KGF, MSCs can significantly inhibit the inflammatory response and repair the vascular-alveolar epithelial cell barrier to promote the elimination of alveolar fluid and thereby the infection.

CONCLUSION

This was the first COVID-19 case seen in Hangzhou, China. During the patient’s hospitalization, laboratory abnormalities and chest CT findings changed in synchrony with the clinical illness. The immune system of COVID-19 patients produces a drastic inflammatory response, causing a cytokine storm that includes the overproduction of cytokines or immune cells. MSCs have been reported to enhance cellular reparative and immunomodulatory properties and thereby prevent or reduce the cytokine storm that develops in COVID-19 patients. After the intravenous injection of MSCs, a portion of the cells accumulate in the lungs via the circulation, where they may contribute to the recovery of the pulmonary microenvironment by protecting alveolar epithelial cells, reducing the amount of effusion, and preventing pulmonary fibrosis, thus ameliorating tissue dysfunction and COVID-19 pneumonia. Pulmonary fibrosis is a serious complication of COVID-19, with the first step in its pathogenesis consisting of the intra-alveolar infiltration of proliferating fibroblasts. Proliferating fibroblasts infiltrating intra-alveolar zone was considered the first step in pathogenesis of pulmonary fibrosis. Edema, fibroblast invasion of the pulmonary interspaces, and type II pneumocyte hyperplasia with inflammatory cytokine infiltrations have been seen in the lung sections of patients with COVID-19. MenSCs, alone or combined with other therapeutic agents, might be a promising treatment for COVID-19.

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REFERENCES

1. Li J, Gong X, Wang Z, Chen R, Li T, Zeng D, Li M. Clinical features of familial clustering in patients infected with 2019 novel coronavirus in Wuhan, China. Virus Res 2020; 286: 198043 [PMID: 32502551] DOI: 10.1016/j.virusres.2020.198043

2. Chakraborty C, Sharma AR, Bhattacharya M, Sharma G, Lee SS. The 2019 novel coronavirus disease (COVID-19) pandemic: A zoonotic prospective. Asian Pac J Trop Med 2020; 13: 242-246 [DOI: 10.4103/1995-7645.281613]

3. Hadi AG, Kadhom M, Hairunisa N, Younis E, Mohammed SA. A Review on COVID-19: Origin, Spread, Symptoms, Treatment, and Prevention. Biointerface Res App 2020; 10: 7234-7242 [DOI: 10.33263/BRIAC106.72347242]

4. Bhattacharya M, Sharma AR, Patra P, Ghosh P, Sharma G, Patra BC, Lee SS, Chakraborty C. Development of epitope-based peptide vaccine against novel coronavirus 2019 (SARS-COV-2): Immunoinformatics approach. J Med Virol 2020; 92: 618-631 [PMID: 32108359] DOI: 10.1002/jmv.25736

5. Fan XL, Zhang Y, Li X, Fu QL. Mechanisms underlying the protective effects of mesenchymal stem cell-based therapy. Cell Mol Life Sci 2020; 77: 2771-2794 [PMID: 31965214] DOI:
6 Le Blanc K, Rasmussen I, Sundberg B, Götherström C, Hassan M, Uzunel M, Ringdén O. Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *Lancet* 2004; 363: 1439-1441 [PMID: 15121408 DOI: 10.1016/S0140-6736(04)16104-7]

7 Wu X, Lao Y, Chen J, Pan R, Xiang B, Du X, Xiang L, Shao J, Xiang C. Transplantation of human menstrual blood progenitor cells improves hyperglycemia by promoting endogenous progenitor differentiation in type 1 diabetic mice. *Stem Cells Dev* 2014; 23: 1245-1257 [PMID: 24499421 DOI: 10.1089/scd.2013.0390]

8 Rodrigues MC, Voltarelli J, Sanberg PR, Allickson JG, Kazmín-Nichols N, Garabuzova-Davis S, Borlongan CV. Recent progress in cell therapy for basal ganglia disorders with emphasis on menstrual blood transplantation in stroke. *Neurosci Biobehav Rev* 2012; 36: 177-190 [PMID: 21645544 DOI: 10.1016/j.neubiore.2011.05.010]

9 Chen L, Xiang B, Wang X, Xiang C. Exosomes derived from human menstrual blood-derived stem cells alleviate fulminant hepatic failure. *Stem Cell Res Ther* 2017; 8: 9 [PMID: 28115012 DOI: 10.1186/s13287-016-0453-6]

10 Xiang B, Chen L, Wang X, Zhao Y, Wang Y, Xiang C. Transplantation of Menstrual Blood-Derived Mesenchymal Stem Cells Promotes the Repair of LPS-Induced Acute Lung Injury. *Int J Mol Sci* 2017; 18: 689 [PMID: 28346367 DOI: 10.3390/ijms18040869]

11 Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Qin J, Qiu W, Jiang C, Gao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020; 395: 497-506 [PMID: 31986264 DOI: 10.1016/S0140-6736(20)30183-5]

12 Phan LT, Nguyen TV, Luong QC, Nguyen TV, Nguyen HT, Le HQ, Nguyen TT, Cao TM, Pham QD. Importation and Human-to-Human Transmission of a Novel Coronavirus in Vietnam. *N Engl J Med* 2020; 382: 872-874 [PMID: 31991079 DOI: 10.1056/NEJMoa2001272]

13 Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W; China Novel Coronavirus Investigating and Research Team. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med* 2020; 382: 727-733 [PMID: 31979845 DOI: 10.1056/NEJMoa2001017]

14 Chan JF, Yuan S, Kok KH, To KK, Chu H, Yang J, Xing F, Liu J, Yip CC, Poon RW, Tai SN, Yip SK; Washington State 2019-nCoV Case Investigation Team. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med* 2020; 382: 929-936 [PMID: 32004427 DOI: 10.1056/NEJMoa2001191]

15 Corman VM, Landt O, Kaiser M, Molenkamp R, Meijer A, Chu DK, Bleicker T, Brünink S, Kuhn JH, Schmidt TF, Zinselmeyer B, Buchheister K, Christian TF, Linthicum HX, Niedrig M, Schmitz MS, Walker DJ; Germany 2019-nCoV Case Investigation Team. Diagnostic and Therapeutic Potential of Remdesivir for SARS-CoV-2: A Case Report. *Euro Surveill* 2020; 25: 2000045 [PMID: 31992387 DOI: 10.2807/1560-7917.ES.2020.25.3.2000045]

16 Lei J, Li J, Li X, Qi X. CT Imaging of the 2019 Novel Coronavirus (2019-nCoV) Pneumonia. *Radiology* 2020; 295: 18 [PMID: 32036464 DOI: 10.1148/radiol.2020200236]

17 Pan Y, Guan H. Imaging changes in patients with 2019-nCoV. *Eur Radiol* 2020; 30: 3612-3613 [PMID: 32025790 DOI: 10.1007/s00330-020-06713-2]

18 Chakraborty C, Sharma AR, Sharma G, Bhattacharya M, Lee SS. SARS-CoV-2 causing pneumonia-associated respiratory disorder (COVID-19): diagnostic and proposed therapeutic options. *Eur Rev Med Pharmacol Sci* 2020; 24: 4016-4026 [PMID: 32329877 DOI: 10.26355/eurrev_202004_20871]

19 Lu H. Drug treatment options for the 2019-new coronavirus (2019-nCoV). *Biosci Trends* 2020; 14: 69-71 [PMID: 31966494 DOI: 10.5582/bst.2020.01020]

20 Golchin A, Farahany TZ, Khajasteh A, Soleimaniifar F, Ardeshirylajimi A. The Clinical Trials of Mesenchymal Stem Cell Therapy in Skin Diseases: An Update and Concise Review. *Curr Stem Cell Res Ther* 2019; 14: 223-33 [PMID: 32019006 DOI: 10.2174/1574888313666180913123242]

21 Liu S, Peng D, Qiu H, Yang K, Fu Z, Zou L. Mesenchymal stem cells as a potential therapy for COVID-19. *Stem Cell Res Ther* 2020; 11: 169 [PMID: 32366290 DOI: 10.1186/s13287-020-01678-8]

22 Bozorgmehr M, Moazzemi SM, Salehnia M, Sheikian A, Nikoo S, Zarnani AH. Mesenchymal blood-derived stromal stem cells inhibit optimal generation and maturation of human monocytic-derived dendritic cells. *Immunol Lett* 2014; 162: 239-246 [PMID: 25455606 DOI: 10.1016/j.imlet.2014.10.005]

23 Khatami S, Khannamohammad M, Zarnani AH, Talebi S, Edalatkhah H, Eghtesad S, Nikoee K, Kazemnejad S. Efficient generation of functional hepatocyte-like cells from menstrual blood-derived stem cells. *J Tissue Eng Regen Med* 2015; 9: E124-E134 [PMID: 23505217 DOI: 10.1002term.1715]

24 Gang EJ, Bosnakovski D, Figuereido CA, Visser JW, Perlingeiro RC. SSEA-4 identifies mesenchymal stem cells from bone marrow. *Blood* 2007; 109: 1743-1751 [PMID: 17062733 DOI: 10.1182/blood-2005-11-010504]

25 Greco SJ, Liu K, Rameshwar P. Functional similarities among genes regulated by OCT4 in human
mesenchymal and embryonic stem cells. Stem Cells 2007; 25: 3143-3154 [PMID: 17761754 DOI: 10.1634/stemcells.2007-0351]

27. Yao S, Chen S, Clark J, Hao E, Beattie GM, Hayek A, Ding S. Long-term self-renewal and directed differentiation of human embryonic stem cells in chemically defined conditions. Proc Natl Acad Sci USA 2006; 103: 6907-6912 [PMID: 16632596 DOI: 10.1073/pnas.0602280103]

28. Jiang Z, Hu X, Yu H, Xu Y, Wang L, Chen H, Chen H, Wu R, Zhang Z, Xiang C, Webster KA, Wang JA. Human endometrial stem cells confer enhanced myocardial salvage and regeneration by paracrine mechanisms. J Cell Mol Med 2013; 17: 1247-1260 [PMID: 23837996 DOI: 10.1111/jcmm.12100]

29. Fayyad-Kazan H, Faour WH, Badran B, Lagneaux L, Najar M. The immunomodulatory properties of human bone marrow-derived mesenchymal stromal cells are defined according to multiple immunobiological criteria. Inflamm Res 2016; 65: 501-510 [PMID: 26956767 DOI: 10.1007/s00011-016-0933-2]

30. Chatterjee D, Marquardt N, Tufa DM, Hatlapatka T, Hass R, Kasper C, von Kaisenberg C, Schmidt RE, Jacobs R. Human Umbilical Cord-Derived Mesenchymal Stem Cells Utilize Activin-A to Suppress Interferon-γ Production by Natural Killer Cells. Front Immunol 2014; 5: 662 [PMID: 25584044 DOI: 10.3389/fimmu.2014.00662]

31. Uccelli A, de Rosbo NK. The immunomodulatory function of mesenchymal stem cells: mode of action and pathways. Ann N Y Acad Sci 2015; 1351: 114-126 [PMID: 26152292 DOI: 10.1111/nyas.12815]

32. Peteranderl C, Morales-Nebreda L, Selvakumar B, Lecuona E, Vadász I, Morty RE, Schmidt C, Bespalowa J, Wolff T, Pleschka S, Mayer K, Gattenloehner S, Fink L, Lohmeyer J, Seeger W, Sznajder JI, Mutlu GM, Budinger GR, Herold S. Macrophage-epithelial paracrine crosstalk inhibits lung edema clearance during influenza infection. J Clin Invest 2016; 126: 1566-1580 [PMID: 26999591 DOI: 10.1172/JCI89331]

33. Leng Z, Zhu R, Hou W, Feng Y, Yang Y, Han Q, Shan G, Meng F, Du D, Wang S, Fan J, Wang W, Deng L, Shi H, Li H, Hu Z, Zhang F, Gao J, Liu H, Li X, Zhao Y, Yin K, He X, Gao Z, Wang Y, Yang J, Jin R, Stamblter I, Lim LW, Su H, Moskalev A, Cano A, Chakrabarti S, Min KJ, Ellison-Hughes G, Caruso C, Jin, Zhao RC. Transplantation of ACE2 Mesenchymal Stem Cells Improves the Outcome of Patients with COVID-19 Pneumonia. Aging Dis 2020; 11: 216-228 [PMID: 32552737 DOI: 10.104336/AD.2020.0227]

34. Matthy MA, Calfee CS, Zhuo H, Thompson BT, Wilson JG, Levitt JE, Rogers AJ, Gotts JE, Wiener-Kronish JP, Bajwa EK, Donahoe MP, McVerry BJ, Ortiz LA, Exline M, Jiang Z, Liu KD. Treatment with allogeneic mesenchymal stem cells for moderate to severe acute respiratory distress syndrome (START study): a randomised phase 2a safety trial. Lancet Respir Med 2019; 7: 154-162 [PMID: 30459077 DOI: 10.1016/S2213-2600(19)30418-1]

35. Meftahi GH, Jangravi Z, Sahraei H, Bahari Z. The possible pathophysiology mechanism of cytokine storm in elderly adults with COVID-19 infection: the contribution of "inflame-aging". Inflamm Res 2020; 69: 825-839 [PMID: 32529477 DOI: 10.1007/s00011-020-01372-8]

36. Saha RP, Sharma AR, Singh MK, Samanta S, Bhattacharya M, Lee SS, Chakraborty C. Repurposing Drugs, Ongoing Vaccine, and New Therapeutic Development Initiatives Against COVID-19. Front Pharmacol 2020; 11: 1258 [PMID: 32973505 DOI: 10.3389/fphar.2020.01258]

37. Khoury M, Alcayaga-Miranda F, Illanes SE, Figueroa FE. The promising potential of menstrual stem cells for antenatal diagnosis and cell therapy. Front Immunol 2014; 5: 205 [PMID: 24904569 DOI: 10.3389/fimmu.2014.00205]

38. Alcayaga-Miranda F, Cuenca J, Luz-Crawford P, Aguila-Díaz C, Fernandez A, Figueroa FE, Khoury M. Characterization of menstrual stem cell: angiogenic effect, migration and hematopoietic stem cell support in comparison with bone marrow mesenchymal stem cells. Stem Cell Res Ther 2015; 6: 32 [PMID: 25889741 DOI: 10.1186/s13287-015-0013-5]

39. Cuenca J, Le-Gatt A, Castillo V, Belletti J, Díaz M, Korte G M, Gonzalez PL, Alcayaga-Miranda F, Schuh CMAP, Ezquer F, Ezquer M, Khoury M. The Reparative Abilities of Mesenchymal Stem Cells Modulate the Wound Matrix Signals and Improve Cutaneous Regeneration. Front Physiol 2018; 9: 464 [PMID: 29867527 DOI: 10.3389/fphys.2018.00464]

40. Luz-Crawford P, Torres MJ, Noël D, Fernandez A, Toupet K, Alcayaga-Miranda F, Tejedor MJ, Jorgensen C, Illanes SE, Figueroa FE, Djouad F, Khoury M. The immunosuppressive signature of menstrual blood mesenchymal stem cells entails opposite effects on experimental arthritis and graft vs host diseases. Stem Cells 2016; 34: 456-469 [PMID: 26528946 DOI: 10.1002/stem.2244]

41. Liu J, Feng B, Xu Y, Zhu J, Feng X, Chen W, Sheng X, Shi X, Pan Q, Yu J, Zeng X, Cao H, Li L. Immunomodulatory effect of mesenchymal stem cells in chemical-induced liver injury: a high-dimensional analysis. Stem Cell Res Ther 2019; 10: 262 [PMID: 31443686 DOI: 10.1186/s13287-019-1379-6]

42. Aleahmad M, Ghanavatinejad A, Bozorgmehr M, Shokri MR, Nikoo S, Tavakoli M, Kazemnejad S, Shokri F, Zamanli AH. Menstrual Blood-Derived Stromal Stem Cells Augment CD4+ T Cells Proliferation. Avicenna J Med Biotechnol 2018; 10: 183-191 [PMID: 30090214]

43. Chen L, Qu J, Xiang C. The multi-functional roles of menstrual blood-derived stem cells in regenerative medicine. Stem Cell Res Ther 2019; 10: 1 [PMID: 30606242 DOI: 10.1186/s13287-018-1105-9]
Chen L, Qu J, Cheng T, Chen X, Xiang C. Menstrual blood-derived stem cells: toward therapeutic mechanisms, novel strategies, and future perspectives in the treatment of diseases. Stem Cell Res Ther 2019; 10: 406 [PMID: 31864423 DOI: 10.1186/s13287-019-1503-7]

Horie S, Laffey JG. Recent insights: mesenchymal stromal/stem cell therapy for acute respiratory distress syndrome. F1000Res 2016; 5: F1000 Faculty Rev-1532 [PMID: 27408702 DOI: 10.12688/f1000research.8217.1]

Tang L, Jiang Y, Zhu M, Chen L, Zhou X, Zhou C, Ye P, Chen X, Wang B, Xu Z, Zhang Q, Xu X, Gao H, Wu X, Li D, Jiang W, Qu J, Xiang C, Li L. Clinical study using mesenchymal stem cells for the treatment of patients with severe COVID-19. Front Med 2020; 14: 664-673 [PMID: 32761491 DOI: 10.1007/s11684-020-0810-9]

Gupta A, Kashe S, Gupta M, Rodriguez HC, Gautam SS, Kadam S. Mesenchymal stem cells and exosome therapy for COVID-19: current status and future perspective. Hum Cell 2020; 33: 907-918 [PMID: 32780299 DOI: 10.1007/s13577-020-00407-w]

Leng Z, Zhu R, Hou W, Feng Y, Yang Y, Han Q, Shan G, Meng F, Du D, Wang S, Fan J, Wang W, Deng L, Shi H, Li H, Hu Z, Zhang F, Gao J, Liu H, Li X, Zhao Y, Yin K, He X, Gao Z, Wang Y, Yang B, Jin R, Stambler I, Lim LW, Su H, Moskalev A, Cano A, Chakrabarti S, Min KJ, Ellison-Hughes G, Caruso C, Jin K, Zhao RC. Transplantation of ACE2-Mesenchymal Stem Cells Improves the Outcome of Patients with COVID-19 Pneumonia. Aging Dis 2020; 11: 216-228 [PMID: 32257537 DOI: 10.14336/AD.2020.0228]

Antebi B, Mohammadipoor A, Batchinsky AI, Cancio LC. The promise of mesenchymal stem cell therapy for acute respiratory distress syndrome. J Trauma Acute Care Surg 2018; 84: 183-191 [PMID: 29019797 DOI: 10.1097/TA.0000000000001713]

Wilson AG, Liu KD, Zhou H, Caballero L, McMillan M, Fang X, Cosgrove K, Vojnik R, Calfee CS, Lee JW, Rogers AJ, Levitt J, Wiener-Kronish J, Bajwa EK, Leavitt A, McKenna D, Thompson BT, Matthy MA. Mesenchymal stem (stromal) cells for treatment of ARDS: a phase 1 clinical trial. Lancet Respir Med 2015; 3: 24-32 [PMID: 25529339 DOI: 10.1016/S2213-2600(14)70291-7]

Mehta P, McCauley DF, Brown M, Sanchez E, Tattersall RS, Mansor JJ; HLH Across Speciality Collaboration; UK. COVID-19: consider cytokine storm syndromes and immunosuppression. Lancet 2020; 395: 1033-1034 [PMID: 32192578 DOI: 10.1016/S0140-6736(20)30628-0]

Sun X, Wang T, Cai D, Hu Z, Chen J, Liao H, Zhi L, Wei H, Zhang Z, Qiu Y, Wang J, Wang A. Cytokine storm intervention in the early stages of COVID-19 pneumonia. Cytokine Growth Factor Rev 2020; 53: 38-42 [PMID: 32360420 DOI: 10.1016/j.cytogfr.2020.04.002]

Saha A, Sharma AR, Bhattacharya M, Sharma G, Lee SS, Chakraborty C. Probable Molecular Mechanism of Remdesivir for the Treatment of COVID-19: Need to Know More. Arch Med Res 2020; 51: 585-586 [PMID: 32439198 DOI: 10.1016/j.arcmed.2020.05.001]

El Agha E, Kramer R, Schneider RK, Li X, Seeger W, Humphreys BD, Belluscio S. Mesenchymal Stem Cells in Fibrotic Disease. Cell Stem Cell 2017; 21: 166-177 [PMID: 28777943 DOI: 10.1016/j.stem.2017.07.011]

Tian S, Hu W, Niu L, Liu H, Xu H, Xiao SY. Pulmonary Pathology of Early-Phase 2019 Novel Coronavirus (COVID-19) Pneumonia in Two Patients With Lung Cancer. J Thorac Oncol 2020; 15: 700-704 [PMID: 32114094 DOI: 10.1016/j.jtho.2020.02.010]
