Mitochondrial DNA copy number, metabolic syndrome, and insulin sensitivity: Insights from the Sugar, Hypertension, and Physical Exercise Studies

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Abstract:

Mitochondrial DNA copy number (mtDNA-CN) measured in blood has been associated with many aging-related diseases, with higher mtDNA-CN typically associated with lower disease risk. Exercise training is an excellent preventative tool against aging-related disorders and has been shown to increase mitochondrial function in muscle. Using the Sugar, Hypertension, and Physical Exercise cohorts (N = 105), we evaluated the effect of 6-months of exercise intervention on mtDNA-CN measured in blood. Although there was no significant relationship between exercise intervention and mtDNA-CN change (P = 0.29), there was a nominally significant association between mtDNA-CN and metabolic syndrome (P = 0.04), which has been seen in previous literature. We also identified a nominally significant association between higher mtDNA-CN and higher insulin sensitivity (P = 0.02).

Introduction:

Mitochondria are well-known for their essential roles in ATP production, though they perform additional functions such as calcium homeostasis, apoptosis signaling, and lipid metabolism[1–3]. ATP synthesis and supply is crucial for skeletal muscle contraction during exercise, and thus, functional mitochondria are necessary for aerobic exercise. Mitochondria contain their own genomes (mtDNA), which can range from tens to thousands of copies per cell. This variation in quantity is referred to as mitochondrial DNA copy-number (mtDNA-CN), and widely differs across cell types and individuals. mtDNA-CN levels are associated with mitochondrial membrane potential, respiratory enzyme function, and energy reserves [4,5], suggesting that mtDNA-CN may be a marker of mitochondrial health. Lower mtDNA-CN in buffy coat has been
associated with frailty, often characterized by decreased muscle tone [6]. Exercise has also been shown to be an excellent preventive tool for many of the aging-related disorders associated with lower mtDNA-CN [7,8]. These findings suggest a relationship between exercise training, which can increase muscle density [9] and mtDNA-CN measured in blood. Indeed, persistent exercise training has been shown to increase mitochondrial function and mitochondrial volume in skeletal muscle [10,11]. Additionally, Lanza et al has shown that mtDNA-CN in muscle for endurance-trained individuals is higher than that of sedentary subjects [8]. However, mtDNA-CN in skeletal muscle is difficult to obtain, as muscle biopsy is required. We hypothesize that exercise intervention can increase mtDNA-CN in blood. To answer this question, we use two randomized controlled exercise intervention cohorts within studies of exercise training.

Results

Exercise increases VO$_2$max and decreases BMI in the SHAPE cohorts

The Sugar, Hypertension, and Physical Exercise (SHAPE) cohorts are a set of randomized controlled studies that aimed to evaluate the effects of exercise and diet interventions on blood pressure and other secondary outcomes [12]. Briefly, participants were randomized into two intervention groups, which varied based on the specific SHAPE study (Table 1). Interventions were for 6 months, and blood samples were drawn at both baseline (pre-intervention) and final (post-intervention) visits.
| Study  | N   | Comorbidities                      | Group1            | Group2            | Completed Protocol |
|--------|-----|------------------------------------|-------------------|-------------------|--------------------|
| SHAPE3 | 77  | Overweight/obese + prediabetes/diabetes | Diet              | Diet + Exercise   | 55                 |
| SHAPE5 | 77  | Obese, otherwise healthy           | Exercise + Low CHO| Exercise + Low Fat| 60                 |

Table 1. Cohort data for the SHAPE3 and SHAPE5 cohorts. Low CHO = low carbohydrate weight loss diet, Low Fat = low fat weight loss diet.

Maximal oxygen uptake (VO\textsubscript{2}\text{max}), is a measurement of an individual’s aerobic capacity and increases after exercise training [13,14]. To confirm the efficacy of the exercise intervention, we examined associations between the 6-month change in VO\textsubscript{2}\text{max} and the number of exercise sessions that an individual attended. There was a positive association (R = 0.38, P = 7.44 x 10\textsuperscript{-5}) between more exercise sessions and a 6-month increase VO\textsubscript{2}\text{max} (Supplemental Figure 1). Additionally, the number of exercise sessions was significantly associated with a 6-month decrease in BMI (R = -0.29 , P = 0.002, Supplemental Figure 2). Taken together, these correlations indicate that exercise intervention was effective.

Supplemental Figure 1. The number of exercise sessions attended is significantly associated with a 6-month increase in VO\textsubscript{2}\text{max}.
Supplemental Figure 2. There was a significant association between a greater number of exercise sessions attended and decreased BMI over the 6-month intervention period.

Measurement and validation of mtDNA-CN in the SHAPE cohorts

mtDNA-CN was measured from whole blood samples obtained at baseline and final visits. Briefly, a monochrome qPCR assay with a nuclear target (albumin) and a mitochondrial target (D-loop) was used to measure the proportion of mitochondrial DNA relative to nuclear DNA [15]. To avoid batch effects, samples derived from the same individual were run on the same plate, and the final mtDNA-CN metric was adjusted for plate as a random effect. The final data was then centered and scaled.

mtDNA-CN is typically higher in females and decreases with age [16,17]. To validate our mtDNA-CN metric, we evaluated associations with these two known covariates, using only baseline (pre-intervention) samples (N = 145). Despite the small sample size, age and sex were both significantly associated in the expected directions [18] (Figure 1). As it has been shown that the relationship between mtDNA-CN and age is nonlinear [19], We modeled the effect of age on mtDNA-CN using a natural spline, yielding a knot at 52.6 years. However, using a log likelihood test, the spline age term did not perform significantly better than a linear term (P =
0.42), potentially due to the small sample size. Adding sex as a covariate significantly improved the model (P = 0.008).

Figure 1. Baseline mtDNA-CN is associated with age and sex in the expected directions.

These effect estimates are consistent across the two different SHAPE studies, confirming that the associations are not driven by any one study (Supplemental Figure 3).
Supplemental Figure 3. Associations between mtDNA-CN and age and sex are in the expected directions when stratifying by SHAPE study. SHAPE3 on the left, SHAPE5 on the right.
During quality control checks, we discovered that higher baseline mtDNA-CN was significantly associated with study dropout ($R = 0.21$, $P = 0.01$, Supplemental Figure 4). This association persisted even when stratifying the analysis by SHAPE study (Supplemental Figure 5).

Examination of other covariates was unable to account for the variation in dropout explained by baseline mtDNA-CN, and we currently do not have a biological explanation for this finding.

Supplemental Figure 4. Individuals who dropped out of the study had significantly higher mtDNA-CN.
Supplemental Figure 5. Individuals who dropped out of the study have significantly higher mtDNA-CN than those who were retained (On left, SHAPE3, on right, SHAPE5).

mtDNA-CN is correlated between visits

Because baseline and final measurements are only separated by six months, we expected baseline and final mtDNA-CN to be correlated. After correcting for plate effects, the Pearson correlation was 0.578 (N = 105) and is consistent between the two studies (Figure 2).

Figure 2. Baseline (pre-intervention) and final (post-intervention) mtDNA-CN measurements taken six months apart are well-correlated.

No significant change in mtDNA-CN after 6 months of study intervention

To calculate the change in mtDNA-CN, we subtracted baseline mtDNA-CN from final mtDNA-CN. As such, positive values indicate an increase in mtDNA-CN over the 6-month period. We found that more extreme baseline mtDNA-CN measurements were likely to have larger 6-month changes, suggesting a reversion to the mean (Supplemental Figure 6). To account for this, all
analyses evaluating associations with 6-month change in mtDNA-CN are adjusted for baseline mtDNA-CN as a covariate.

Supplemental Figure 6. Baseline mtDNA-CN is associated with a change in mtDNA-CN in the direction of the mean. Purple points denote individuals with final mtDNA-CN measurements closer to the mean than their baseline measurements, while yellow is vice versa. Significantly more individuals move towards the mean (chi-squared p = 0.004). For the purple samples, absolute magnitude of baseline mtDNA-CN is positively correlated with the absolute value of 6-month change, however, this association is not significant (P = 0.51).

When comparing the change in mtDNA-CN between exercisers and non-exercisers, there was no significant difference in the mtDNA-CN change (P = 0.29, Figure 3). We also analyzed associations between the number of exercise sessions attended and change in mtDNA-CN in the group of individuals who exercised and found no significant relationship (P = 0.45).

However, both analyses were consistent with a positive correlation between mtDNA-CN change
and exercise. With our current sample size (N = 105), we had 80% power to detect a 0.234
difference in means between exercisers and non-exercisers.

Figure 3. There was no significant association between exercise intervention and 6-month
change in mtDNA-CN.

We also examined associations between mtDNA-CN and VO_2max, a measure of
cardiorespiratory fitness. A linear mixed model, adjusting for individual as a random effect and
age, sex, visit, and study as fixed effects, found no significant associations between mtDNA-CN
and VO_2max (P = 0.44).

There was also no association between 6-month change in mtDNA-CN and 6-month change in
VO_2max (Supplemental Figure 7).
Supplemental Figure 7. There was no significant association between 6-month change in VO$_2$ max and 6-month change in mtDNA-CN.

Evaluating associations between secondary outcomes

In addition to exercise and diet, we were interested in associations between mtDNA-CN and secondary outcomes such as muscle mass, insulin sensitivity, and resting metabolic rate.

| Secondary outcome         | Effect size estimate | Standard error | P-value | FDR-adjusted P-value |
|---------------------------|----------------------|----------------|---------|----------------------|
| Muscle mass               | -0.25                | 0.38           | 0.51    | 0.63                 |
| **Insulin sensitivity**   | **0.005**            | **0.002**      | **0.02**| **0.10**             |
| Resting metabolic rate    | 0.33                 | 7.95           | 0.97    | 0.97                 |
| Baseline glycemia         | -1.14                | 1.80           | 0.53    | 0.63                 |
| **Metabolic syndrome**    | **-0.81**            | **0.38**       | **0.04**| **0.11**             |
| HbA1c                     | 0.09                 | 0.06           | 0.13    | 0.26                 |

Table 2). To leverage data from both baseline and final visits, we utilized a linear mixed model, adjusting for age, sex, visit, and individual.
Table 2. Effect size estimates, standard errors, and p-values for secondary outcomes of interest.

| Secondary outcome       | Effect size estimate | Standard error | P-value | FDR-adjusted P-value |
|-------------------------|----------------------|----------------|---------|-----------------------|
| Muscle mass             | -0.25                | 0.38           | 0.51    | 0.63                  |
| Insulin sensitivity     | 0.005                | 0.002          | 0.02    | 0.10                  |
| Resting metabolic rate  | 0.33                 | 7.95           | 0.97    | 0.97                  |
| Baseline glycemia       | -1.14                | 1.80           | 0.53    | 0.63                  |
| Metabolic syndrome      | -0.81                | 0.38           | 0.04    | 0.11                  |
| HbA1c                   | 0.09                 | 0.06           | 0.13    | 0.26                  |

Of these secondary outcomes, insulin sensitivity and metabolic syndrome were both associated with mtDNA-CN prior to multiple-testing correction. Associations between mtDNA-CN and metabolic syndrome have been previously reported, supporting this finding [19]. As individuals with prevalent diabetes are known to have lower mtDNA-CN [20] and type 2 diabetes is a disease primarily characterized by decreased insulin sensitivity [21], we re-examined this association after adjusting insulin sensitivity for diabetes status. The association between mtDNA-CN and insulin sensitivity remained, even after accounting for diabetes status (P = 0.007).

As many of these secondary outcomes are known to be linked with metabolic syndrome, we repeated the analysis, adjusting for metabolic syndrome as a covariate. Results did not significantly change, suggesting that metabolic syndrome does not mediate relationships between mtDNA-CN and these secondary outcomes (Supplemental Table 1).
Supplemental Table 1. Effect size estimates for secondary outcomes after including metabolic syndrome status as a covariate. Results generally stay the same, indicating that metabolic syndrome is not mediating the effects of mtDNA-CN on these outcomes.

| Secondary outcome | Effect size estimate | Standard error | P-value | FDR-adjusted P-value |
|-------------------|----------------------|----------------|---------|----------------------|
| HbA1c             | 0.09                 | 0.06           | 0.09    | 0.23                 |

Discussion

In the SHAPE cohorts, metabolic syndrome and insulin sensitivity were nominally significantly associated with mtDNA-CN, with lower mtDNA-CN associated with metabolic syndrome and lower insulin sensitivity. mtDNA-CN from baseline and final visits were well-correlated, indicating that while mtDNA-CN may change over time, measures taken six months apart are relatively consistent.

We were powered to detect a difference of 0.234 standard deviations for 6-month change in mtDNA-CN between exercising and non-exercising groups and did not observe a significant difference in our dataset. Previous literature has described a significant increase in blood mtDNA-CN after exercise, as well as a significant association between mtDNA-CN and VO₂max [22]. However, these methods do not normalize mtDNA content to a nuclear DNA target, normalizing instead to a spike-in standard DNA target. As such, the metrics used in the aforementioned study do not adjust for the number of cells and cell type differentials present in each sample.

After evaluating associations between mtDNA-CN and several secondary outcomes, insulin sensitivity as estimated from the QUICKI score was significantly associated with mtDNA-CN, with higher mtDNA-CN associated with increased insulin sensitivity. Loss of mitochondrial
function in elderly subjects has been shown to lead to lipid accumulation and ultimate insulin resistance, corroborating our finding [23].

Since we do not have cell-type composition data, it is difficult to determine whether observed changes in mtDNA-CN are due to changes in mitochondrial content or changes in cell type composition, as aerobic exercise is known to cause decreases in neutrophils [24] and monocyte-platelet aggregates [21], but also causes increases in overall platelet count[25]. mtDNA-CN is known to be confounded by cell-type composition, with increased platelet count leading to higher mtDNA-CN and increased neutrophil count leading to lower mtDNA-CN [26].

As such, these findings must be interpreted with this limitation in mind.

An additional constraint to this study is the varying comorbidities between SHAPE3 and SHAPE5. Although both SHAPE cohorts are comprised of obese and overweight individuals, SHAPE3 recruited subjects with type 2 diabetes or having prediabetes, while SHAPE5 individuals were otherwise healthy except for having abdominal obesity. As diabetes is known to cause abnormalities in mitochondrial function [20,27,28], this may affect the relationship between exercise and mtDNA-CN in individuals with diabetes. However, with the limited sample size in this study, there did not appear to be an association between exercise and mtDNA-CN, and addition of diabetes status as a covariate did not change results.

In summary, we do not detect a significant change in mtDNA-CN after exercise intervention in these study cohorts, despite marked improvements in fitness and substantial weight loss. After examining secondary outcomes, we uncovered a significant association between mtDNA-CN and insulin sensitivity, likely driven by biological pathways that connect mitochondria, lipid accumulation, and insulin resistance.
Methods

Participant Recruitment

This study was approved by the Johns Hopkins Medicine IRB under retrospective application IRB0007178. Written informed consent was obtained for all participants, and all DNA samples and associated phenotype data were de-identified prior to analysis. All studies are listed under ClinicalTrials.gov (SHAPE3: NCT00928005, SHAPE5: NCT00990457).

SHAPE3

Subjects were overweight or obese (BMI between 26 and 42 kg/m²), sedentary men and women (n=77), 30-65 years, with prediabetes or diabetes, according to American Diabetes Association criteria (fasting glucose > 126 mg/dl, casual plasma glucose > 200 mg/dl, or 2-hour plasma glucose > 200 mg/dl after a 75-gram oral glucose load). Individuals with uncontrolled diabetes, defined as fasting blood glucose over 300 mg/dl or A1C > 11% were excluded.

SHAPE5

Subjects were overweight or obese, sedentary men and women (n=77), BMI 25-42 kg/m², 30-65 years, who were otherwise healthy.

Exercise and Diet Intervention

SHAPE3

Diet intervention for SHAPE3 was a nutritionally balanced, moderately hypocaloric diet with reduced saturated fat consistent with American Diabetes Association guidelines. The diet was adjusted to produce a 600 kcal deficit/day for each individual, using resting metabolic rate calculated from the Mifflin-St Jeor equation [29].
Exercise intervention for SHAPE3 was designed based on guidelines from the American College of Sports Medicine and the American Diabetes Association, consisting of warm-up, 45 minutes of aerobic exercise, several resistance training exercises, and cool-down. Exercise sessions were supervised by exercise physiologists to ensure safety and that the exercises were carried out properly. Individuals assigned to exercise intervention were asked to exercise 3 times a week over a 26-week period.

**SHAPE5**

The low-carbohydrate (CHO) group adhered to the New Atkins for Life diet, consisting initially of 15% CHO, 30% protein, and 55% fat, followed by a gradual shift to 40% CHO, 20% protein, and 40% fat.

The low-fat group followed American Heart Association (AHA) and National Cholesterol Education Program (NCEP) guidelines, following a diet of 30% fat, 50-55% CHO, and 15-20% protein.

All subjects in SHAPE5 participated in 3 times per week supervised exercise training following ACSM guidelines for moderate intensity aerobic and resistance training, consisting of 45 minutes of aerobic exercise and 2 sets of 7 resistance exercises.

**Measurement of study variables**

Maximal oxygen uptake (VO2 max ml/kg/min). A Cardinal Health Metabolic/EKG system was used to measure VO2 max. The exercise began at 3 mph, 0% grade, and increased 2.5% grade every 3 minutes. There was continuous EKG and cardiorespiratory monitoring. The 12 lead ECG was recorded at every stage. BP was measured during the last 30 seconds and the Rating of perceived exertion (RPE), using the Borg 6 to 20 scale, was obtained during each stage. An RPE
of 18-20 and a respiratory exchange ratio > 1.1 were considered as indicators of maximal effort.

The highest observed value of VO\textsubscript{2} was recorded as VO\textsubscript{2} max.

Muscle mass was measured using Dual Energy X-Ray Absorptiometry (DEXA) with a GE Lunar Prodigy. DEXA lean mass measurements were utilized as a representation for muscle mass.

Insulin sensitivity was calculated from fasting glucose and fasting insulin measurements using the quantitative insulin sensitivity check index (QUICKI) formula [30]. Insulin, glucose, and Hb1ac levels were measured from a fasting blood draw. Anthropometry was performed to obtain height and weight measurements with a balance scale and stadiometer. BMI was then calculated using these measurements. Waist and hip measurements were taken using a tape measure. Resting metabolic rate was estimated using the Mifflin-St Jeor equation [29].

Subjects were categorized as having metabolic syndrome if they had ≥ 3 of the 5 factors: central obesity with a waist circumference of > 40 inches (M) or > 35 inches (F); hyperglycemia, fast glucose ≥ 100 mg/dl or taking medications; dyslipidemia, triglycerides ≥ 150 mg/dl or taking medications; dyslipidemia (2\textsuperscript{nd}, separate criteria, HDL cholesterol ≤ 40 mg/dl (M) or ≤ 50 mg/dl (F) or taking medication for both sexes; hypertension, ≥130 mm Hg systolic or ≥85 mm Hg diastolic, or taking medications.

**mtDNA-CN measurement**

mtDNA-CN was measured using a monochrome qPCR method [15]. Previous work comparing this assay with mtDNA-CN derived from whole genome sequencing has shown that individuals with polymorphisms in the D-loop primer region have unreliable mtDNA-CN monochrome assay measurements [31]. As such, samples that had deltaCT (difference between nuclear and mitochondrial probe CTs) less than 7 were filtered out due to assumed polymorphisms in the D-
loop primer (7 total samples). One outlier individual was removed due to a baseline and a final mtDNA-CN value that was greater than 3 SD from the mean.

**Genetic fingerprinting**

Genetic fingerprinting using the Agena iPLEX Pro SampleID Panel was used to identify sample swaps and confirm that baseline and final samples originated from the same individual. Seven samples (two total individuals) were removed due to duplicated sample IDs with non-matching genetic information, as there was no way to match which sample corresponded to the correct individual. Four samples were removed due to poor sample quality, with greater than 50% missingness on the array. Finally, two samples (one individual) were removed due to fewer than 90% matching calls between baseline and final samples.

**Code and Data Availability**

De-identified data is available on the Arking lab website: https://www.arkinglab.org/. Code and scripts are available on the ArkingLab Github: https://github.com/ArkingLab/SHAPE_scripts.

**Statistical Analyses**

All statistical analyses were performed with R version 4.1.1 [32]. Linear mixed models were performed using the lme4 package and plots were created with ggplot2 [33,34].

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