Introduction

Fish culture has become an important industry and the world’s fastest growing sector of agricultural business Villa-Cruz et al. [1]. FAO [2] reported that the total world fishery production decreased slightly and the human consumption for aquatic product increased over the years. The need to enhance growth performance, feed efficiency and disease resistance of cultured organisms is substantial for various sectors of this industry EL-Haroun et al. [3]. Diet supplementation is an important aspect in aquaculture management especially in intensive or in semi-intensive fish culture and is promising for increasing fish production. EL-Haroun, [4] pointed out that diet is often the single largest operating cost item that represent over 50% of the operating costs in intensive aquaculture. Traditional use of antibiotics and other chemotherapeutics in fish culture has been criticized because of the potential development of multiple antibiotic resistant bacteria, environmental pollution and the accumulation of residues in fish Ringo et al. [5]. Scientists have intensified efforts to identify and develop safe dietary supplements and additives that enhance the life activity, health and immune system of farmed fish Ji et al. [6]; Shim et al. [7].

World Health Organization encourages using of medicinal herbs and plants to substitute or minimize the use of chemicals through the global trend to go back to nature. With the shifting of attention from synthetic drugs to natural plant products, the use of plant extracts for enhancing growth performance in animals is now on the increase. Plants that were once considered of no value are now being investigated, evaluated and developed into drugs with little or no side effects. One of such plant is Talinum triangulare (Water leaf). T. triangulare is a cosmopolitan weed belonging to the Talinum genus, family Portulacaceae that grows best under humid conditions Burkill [8]. It is one of the vegetables widely cultivated and consumed in Africa especially Southern Nigeria Imoh and Julia [9]. Ezekwe et al. [10]...
reported that T. triangulare is a rich source of vitamins, β-carotene, minerals (such as calcium, potassium and magnesium), pectin, proteins and vitamins. It has also been found to possess useful medical potentials such as laxative, purgative, treatment of diarrhea, gastrointestinal illnesses Oguntona [11]; Mensor et al [12] as well as in the management of cardiovascular diseases such as: stroke and obesity Aja et al. [13].

Previous studies on rats Ezekwe et al. [14] showed that inclusion of T. triangulare methanolic extract suggest a lot of health benefits in human and animal. However, the potential of health benefits of T. triangulare in fish has not been investigated. Clarias gariepinus (Burchell, 1822) is the most important cultivated fish in Nigeria. This species has shown considerable potential as a fish suitable for use in intensive aquaculture. In spite of the remarkable aquaculture potential of C. gariepinus, the size obtained under culture is still poor compared to its hybrid and Heterobranchus spp Dada & Ikuerowo [15]. There is need to enhance the culture of this valuable species for sustainable aquaculture production and also to meet the demand of the ever increasing population in the country. The objective of this study was to investigate the effects of varying dietary supplementation of T. triangulare additive on growth performance of African catfish, C. gariepinus.

Materials and Methods

Experimental Fish

One hundred and forty four fingerlings of Clarias gariepinus were obtained from private fish farm in Oloko, Oyo State of Nigeria. The fish were acclimatized in 50 litres plastic bowls for two weeks. During the period of acclimatization the fish were fed ad libitum Anibeze and Eze, [16] at 5% body weight twice daily Okoye et al. [17] with a formulated diet of 35% crude protein without herbal extract and no history of herbal feeding from the farm where fish were sourced. At the end of the acclimatization period, the fish were randomly selected and assigned to four experimental groups of 0% (controls), 0.5%, 1.0% and 3% concentrations of T. triangulare powder in diet at 36 fish per treatment. Twelve plastic aquariums each measuring 40 cm x 27 cm x 27 cm with each aquarium holding 12 fish were set up to maintain three replicates per treatment. Feeding was suspended 24 hours before the feeding trial to increase appetite and reception for new diet Madu & Akiolo [18].

Experimental Diet

Fishmeal, maize, wheat bran, soya bean meal, calcium carbonate and limestone sourced from local markets were used to formulate basal diets (35% CP) without herbal plants. All ingredients were ground into powdery form using a mechanical grinder, and then mixed with mineral mixture and vegetable oil. The ingredients and proximate chemical composition of basal feed was estimated by the methods described by the AOAC [19]. Four different diets with or without additives, representing four dietary variants (Table 1) were then prepared by incorporated T. triangulare powder into basal diet at levels of 0.0%, 5.0%. 1.0% and 3.0%. The ingredients were then thoroughly mixed together by hand. Warm water was added to the premixed ingredients and homogenized to a dough-like paste. The diets were then pelletised using 0.5 mm pellet press. The diets were sun-dried for 4 days and labeled appropriately and stored in airtight containers throughout the experimental period.

Table 1: Composition of experimental diet.

| Ingredients | Concentration of Talinum triangulare in diet |
|-------------|---------------------------------------------|
|             | 0%  | 0.5% | 1.0% | 3.0% |
| Yellow corn (g) | 35.0 | 35.0 | 35.0 | 35.0 |
| Soybean meal (44%)(g) | 28.5 | 28.5 | 28.5 | 28.5 |
| Fish meal (65%)(g) | 17.0 | 17.0 | 17.0 | 17.0 |
| Wheat bran(g) | 9.5 | 9.5 | 9.5 | 9.5 |
| Tt Powder(g) | 0.0 | 0.5 | 1.0 | 3.0 |
| Calcium Carbonate(g) | 0.3 | 0.3 | 0.3 | 0.3 |
| Ground lime stone(g) | 0.7 | 0.7 | 0.7 | 0.7 |
| Vegetable Oil(ml) | 6.5 | 6.5 | 6.5 | 6.5 |
| Mineral mixture (g) | 1.7 | 1.7 | 1.7 | 1.7 |
| Vitamin mixture(g) | 1.0 | 1.0 | 1.0 | 1.0 |

Experimental Procedure

Feeding of Fish: Fish in all the treatments were fed two times a day (morning 9 am and evening 4 pm) at the rate of 5% body weight per day. Feeding allowance was adjusted in accordance with increase in body weight (Hogendoorn & koops, 1983) and diet allotments were increased weekly after the length-weight determination. The feeding last for a period of 70 days. Feed not consumed and faecal matter were siphoned out every day. Dead fish were picked daily and recorded.

Monitoring of Water Quality: Water quality parameters were managed and maintained as recommended by Boyd and Lickotcooper [20]. Water quality parameters were taken weekly throughout period of the experiment. Water temperature (°C), Dissolved Oxygen (DO), pH, Total Dissolved solids (TDS), Ammonia (NH3) and Nitrite (NO2) were measured using a portable HannaR HI98186 meter and Aqua chek® (USA) water quality test stripes. The water in the tanks was drained and replaced every day.

Data Collection: Fish in each tank were batch weighed at the commencement of the study and weekly thereafter with digital electronic weighing balance to the nearest gram. The ration was adjusted every week when new mean weights of fish for the various experimental units were determined. The standard length was determined with a graduated tape. Data on performance such as body weight changes, relative weight gain, specific growth rate, feed conversion ratio, condition factors Bagenal, [21] and survival percentage Fasakin et al., [22] were determined using the following formulae:

\[ \text{Weight gain (g)} = \text{Final weight of fish} - \text{Initial weight of fish} \]  
\[ \text{Relative weight gain (RWG, %)} = \frac{\text{Weight gain}}{\text{Initial weight}} \times 100 \]  
\[ \text{Specific growth rate (SGR)} = \text{calculated as:} \]
a. SGR (% per day) = \frac{(\text{netLogW}_2 - \text{netLogW}_1)}{t_2 - t_1} \times 100 \quad (3)

Where:
\[
\begin{align*}
\text{W}_2 &= \text{Weight of fish at time T2 (final)} \\
\text{W}_1 &= \text{Weight of fish at time T1 (initial)}
\end{align*}
\]

ii. Mortality (M) was calculated as:
\[
M (\%) = \frac{(N_o - N_t)}{N_o} \times 100
\]

Table 2: Water quality fluctuations in experimental tanks containing the African catfish, Clarias gariepinus.

| Treatment | Parameters | 0.0% | 0.5% | 1.0% | 3.0% |
|-----------|------------|------|------|------|------|
|           | Temp(0C)   | 28.0±0.28 a | 28.0±0.30 a | 27.97±0.33 a | 28.00±0.30 a |
|           | DO (mg/l)  | 6.4±0.06 a | 6.5±0.04 a | 6.45±0.02 a | 6.45±0.02 a |
|           | pH         | 7.1±0.0 a | 7.1±0.0 a | 7.10±0.0 a | 7.07±0.03 a |
|           | NH3(mg/l)  | 0.0±0.0 a | 0.0±0.0 a | 0.0±0.0 a | 0.0±0.0 a |
|           | NO2(mg/l)  | 0.01±0.01 a | 0.01±0.01 a | 0.01±0.01 a | 0.02±0.01 a |
|           | TDS (ppm)  | 7.06±5.46 a | 7.56±1.20 a | 7.26±0.67 a | 7.63±1.45 a |

Note: Average of 10 weeks readings. Mean values with the same superscript letter in the same row are not significantly different (p>0.05). Control (0.0%).

Growth Performance

Growth performance of the Clarias gariepinus fingerlings fed locally made feed containing varying quantities of T. triangulare powder as feed additives over a 70 days period is presented in Table 3. Average of initial body weight of C. gariepinus fed the experimental diets at the start did not differ, indicating that groups were homogenous. Fish were able to utilize the test diets at varying degrees but average weight gain (AWG), Feed Conversion Ratio (FCR), Specific Growth Rate (SGR), Mortality (M), Condition Factor (K) of fish were only marginally different (P>0.05). However, the mean final body weight (MFBW), was significantly (P<0.05) higher than the initial fish weight in all diet treatment groups (Table 3). The best growth performance, Feed Conversion Ratio and Condition Factor were observed in the group of fish fed 1.0% T. triangulare. However, highest survival rate (SR) of 83.34±8.33% was observed in the group of fish fed 0.5% T. triangulare while lowest SR of 75.00±4.81% were observed in both control group and in the group of fish fed 3.0% T. triangulare.

Table 3: Growth performance of Clarias gariepinus fingerlings fed different concentration of Talinum triangulare as feed additives.

| Parameters | 0.0% | 0.5% | 1.0% | 3.0% |
|------------|------|------|------|------|
| MBW (g)    | 1.07±0.24 a | 1.05±0.029 a | 1.10±0.015 a | 1.09±0.049 a |
| MFBW (g)   | 1.91±0.127 a | 2.14±0.084 a | 2.38±0.096 a | 1.86±0.439 a |
| AWG (%)    | 0.84±0.107 a | 1.09±0.113 a | 1.28±0.085 a | 0.87±0.110 a |
| RWG4 (%)   | 78.9±8.87 a | 104.9±13.63 a | 116.3±6.82 a | 97.9±5.69 a |
Discussion

The need to understand the roles of phyto-additives in aquaculture has led to various investigation of different herbal plants in aquaculture feed. To date a variety of herbs and spices have been successfully used in fish culture as growth promoters and immune stimulants Irkin et al. [24]. When medicinal plants are used in fish diets, one of the common problems encountered is the acceptability of the feed by fish, and this frequently relates to the palatability of the diet Rodriguez et al. [25]. In this present investigation, the experimental diet variants were accepted by Clarias gariepinus fingerlings, indicating that the levels of incorporation of herbal plant used did not affect the palatability of the diets. This might be attributed to the processing technique which involved air drying techniques that might have reduced the anti-nutrient factors that may be present in this plant thereby not affecting the palatability of the diets. This observation corroborates the works of Fagbenro [26], Francis et al. [27] and Siddharaju & Becker [28]. These workers reported that reduction in anti-nutrient by different processing like soaking, and drying techniques resulted in better palatability and growth in fish. Medicinal plants have received increasing attention as spices for human and additives in diets for animals. However, only few studies have been done on the use of feed additives in fish nutrition Lawhavinit et al. [29] and more so medicinal plants as feed additives in rearing of African catfish in this part of the world. The results in this experiment suggest that dietary feed additive of Talinum triangulare had no effect on growth of Clarias gariepinus fingerlings as evidenced by similar body weight gain among treatment groups fed supplemented diet. However, all treated groups showed improved weight gain, Feed Conversion Ratio and Condition Factor and higher survival rate (SR) compared to group of fish fed diet that did not supplemented (control diet) and best results were observed in the group of fish fed 10g/kg of Talinum triangulare. More so, final fish weights were higher than the initial weights in all the groups, this indicates no negative growth as a result of diet supplementation. Ndong and Fall [30] reported that garlic supplemented diet resulted in decreased body weight gain in juvenile hybrid tilapia (Oreochromis niloticus x Oreochromis aureus) fed diets supplemented with 0.5g/kg garlic over 4 weeks. The slow weight gains recorded in this experiment might be attributed also to the fact that fish were fed exclusively on the formulated feeds without having access to natural feed as may be found in pond or riverine conditions. The African catfish is omnivorous and feeds from a wide array of organisms under natural conditions.

Conclusion and Recommendation

This study has shown that 0.5 - 3.0% waterleaf supplement in fish feeds enhance growth of C. gariepinus, however the inclusion rate at 1.0% proved to be more beneficial than other inclusion rates used in the experiment. Further research is recommended to investigate toxicity of this plant at varying inclusion levels of 0.5% to 3.0% in the Clarias gariepinus culturing to prove the safety of this plant [31]. It is also recommended that good aquaculture practice which include good quality water management among others be ensured in aquaculture practice.

Acknowledgment

The authors wish to acknowledge various sources of references for this work. More so, the management and staff of F. M Sanusi Aqualife Farms, Oloodo Ibadan, Oyo State, Nigeria were appreciated for supplying fish used in this study and for their technical advice and encouragement.

References

1. Villa Cruz V, Davilla J, Viana MT Vazquez Duhalt R (2009) Effect of broccoli (Brassica oleracea) and its phytochemical sulforaphane in balanced diets on the detoxification enzymes levels of tilapia (Oreochromis niloticus) exposed to a carcinogenic and mutagenic pollutant. Chemosphere 74: 1145-1151.
2. FAO (2010) The State of World Fisheries and Aquaculture. p. 197.
3. EL Haroun ER, Ma A, Geda S, Kabir Chowdhury MA (2006) Effect of dietary probiotic Biogens supplementation as growth promoter on growth performance and feed utilization of Nile tilapia, Oreochromis niloticus (L.). Aquac Res 37: 1473-1483.
4. EL Haroun ER (2007) Improved growth rate and feed utilization in farmed African catfish Clarias gariepinus (Burchell 1822) thought a growth promoter Biogen® supplementation. Jour Fish Aquit Sci 2: 319-327.
5. Ringo E, Oken RE, Gifstad T0, Dalmo RA, Amkund H, et al. (2010) Prebiotics in aquaculture: a review. Aquaculture Nutrition 16: 117-136.
6. Ji SC, Takasoka O, Jeong GS, Lee SW, Ishimaru K, et al. (2007) Dietary medicinal herbs improve growth and some non-specific immunity of red sea bream Pagrus major. Fisheries Science 73: 63-69.
7. Shim SM, Ferruzzi MG, Kim YC, Janie EM, Santerre CR (2009) Impact of Phytochemical-rich foods on bioaccessibility of mercury from fish. Food Chemistry 112: 46-50.
8. Burkhill HM (1994) The Usehl Plants of West Tropical Africa. Volume 2: Families EL Kew. Royal Botanic Gardens, Kew.
9. Imoh EU, Julia OM (2000) Nutrient requirement for the growth of water leaf (Talinum triangulare) in Uyo Metropolis, Nigeria. The Environ 21: 153-159.
10. Ezekwe MO, Besong SA, Igbokeke PE (2001) Beneficial influence of purslane and waterleaf supplement to Humans. FASEB J 16(4): A639.
11. Oguntola T (1998) Green Leafy Vegetables. In: Nutritional Quality of Families EI. Kew. Royal Botanic Gardens, Kew.
12. Mensor LL, Fabio SM, Gildor GL, Alexander SR, Tereza CD, et al. (2001) Screening of Brazilian plant extracts for antioxidant activity by the use of DPPH free radical methods. Phytother Res 15(2): 127-130.
Cite this article: Tiamiyu Adebisi Musefiu. Growth Performance of the African Catfish, Clarias gariepinus, Fed Varying Inclusion Levels of Talinum triangulare as Feed Additives. Biomed J Sci & Tech Res 9(3)-2018. BJSTR. MS.ID.001812. DOI: 10.26717/BJSTR.2018.09.001812.