Protective effects of paeonol on subacute/chronic brain injury during cerebral ischemia in rats

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Abstract. Ischemic stroke is a highly complex pathological process that is divided into acute, subacute and chronic phases. Paeonol is a biologically active natural product with a variety of pharmacological effects, including those on neuronal activity. However, the effects of paeonol on subacute/chronic ischemic stroke have remained to be elucidated. The present study was designed to investigate the effects of paeonol against subacute and chronic cerebral ischemic injury and to explore the possible underlying mechanisms. Male adult Sprague Dawley rats were randomly divided into a sham group (treated with saline), a model group [subjected to middle cerebral artery occlusion (MCAO) and treated with saline] and a paeonol-treated group (MCAO + paeonol at 25 mg/kg). Behavioral impairment, infarct volume and ischemic/contralateral hemispheric ratios were assessed at 72 h and at 28 days after MCAO, respectively. Immunofluorescence was employed to determine the neuronal damage and glial responses after MCAO. Compared with the model group, paeonol treatment significantly attenuated behavioral impairment, ischemic infarct volume and moderate cerebral edema in the ischemic brain at 72 h, as well as brain atrophy at 28 days after reperfusion. Furthermore, paeonol treatment alleviated neuronal damage in the ischemic core and boundary zone regions at 72 h after reperfusion and in the boundary zone at 28 days after reperfusion. In addition, paeonol treatment reduced the proliferation of astrocytes in the boundary zone, and inhibited microglial activation in the ischemic core and boundary zone regions at 72 h and 28 days after reperfusion. These results demonstrated the protective effects of paeonol against subacute/chronic cerebral ischemia, and the mechanism of action may include subacute/chronic microglial activation and astrocyte proliferation.

Introduction

Ischemic stroke is the primary cause of cerebrovascular diseases and remains to be one of the leading causes of death and disability in patients worldwide (1,2). Despite tremendous research efforts leading to improvements in ischemic stroke treatment, the available therapeutic strategies are currently limited, and several promising agents evaluated in extensive preclinical trials failed to enter clinical trials (3,4). At present, thrombolytic therapy remains to be the gold standard for ischemic stroke treatment; however, it is only efficacious within a narrow therapeutic window (1,2,4,5). Hence, beyond the therapeutic window, management of stroke mainly depends on supportive therapy, secondary prevention and rehabilitation (1). Thus, an enhanced understanding regarding the pathological processes may provide novel therapeutic options to promote stroke recovery.

Ischemic stroke is a highly complex pathological process, which induces a series of cellular and molecular events and may be divided into three stages: The acute phase (hours), the subacute phase (hours to days) and the chronic phase (days to months) (6). In the acute stage, metabolic disturbances and excitotoxicity are involved in the progression of neuronal damage; in the subacute phase, inflammation and cell death are the dominant events; in the chronic phase, brain repair is the major response and includes microgliosis, glial scar formation, angiogenesis, as well as neuronal regeneration (6-8). In addition, during the pathological process of ischemic stroke, the subacute and chronic phases are generally termed as late (subacute/chronic) phases (6,9). In the late phase, one of the dominant responses is reactive astrogliosis with subsequent glial scar formation, which not only protects the neurons against harmful substances by isolating the injury area but also obstructs neuronal regeneration by suppressing axonal sprouting (10,11). Another important event is the activation

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of microglia, which endangers neuronal survival by releasing numerous proinflammatory and neurotoxic mediators (8,12,13).

Recently, considerable attention has been paid to Traditional Chinese Medicine, as it includes sources of neuroprotective components (14). Natural products, particularly medicinal plants, may provide an ideal choice for the development of safe and effective drugs for stroke treatment (14). Paeonol, an active component isolated from the Chinese herbal medicine Cortex Moutan, which is the root bark of Paeonia suffruticosa Andr., has been demonstrated to possess diverse pharmacological activities, including anti-oxidant (15), anti-atherosclerotic (16), anti-tumor (17), anti-diabetic (18), and anti-inflammatory effects (19). Regarding its applicability for central nervous system diseases, previous studies have demonstrated that paeonol exerts neuroprotective actions against acute ischemic stroke, as well as Parkinson's and Alzheimer's disease in animal models, (19-22). These studies indicate that paeonol may be a promising drug for the treatment of neurological disorders. However, to the best of our knowledge, the potential effects of paeonol on subacute/chronic ischemic stroke have remained to be determined. The present study pursued to investigate the therapeutic potential of paeonol in middle cerebral artery occlusion (MCAO)-induced subacute/chronic cerebral ischemia. Furthermore, the present study focused on the post-ischemic microglial and astrocyte responses in the rat brain in an attempt to elucidate the underlying mechanisms.

Materials and methods

Experimental animals. A total of 125 adult male Sprague Dawley rats weighing 230-280 g (10-12 weeks old) were obtained from the Animal Science Center of Zhejiang Academy of Medical Sciences, [Hangzhou, China; certificate no. SCXK (Zhe) 2014-0001]. Animals were housed in an animal center at a constant temperature of 22±2°C, a relative humidity of 50±10% and a 12-h light/dark cycle. They were allowed free access to food and water. Behavioral experiments were arranged at 10:00 a.m.-05:00 p.m. during the day. All efforts were made to minimize their suffering. All experimental procedures were performed in accordance with the National Institutes of Health Guidelines for the Care and Use of Experimental Animals and were approved by the Animal Care and Use Committee of Zhejiang University (Hangzhou, China).

Induction of transient focal cerebral ischemia. Transient focal cerebral ischemia was induced by MCAO according to previous methods (23) with certain modifications, such as using the nylon suture with a poly-L-lysine coated and without paeonol-treated group (MCAO + Pae at 25 mg/kg, n=21). For drug delivery, the same volume of saline or paeonol solution was injected i.p. at the onset of MCAO (when the nylon suture was inserted and the MCA origin was blocked), and then injected once a day for 3 days. During the experiment, one part of the animals (n=8) were used for determination of infarct volume, and the remaining animals were prepared to cut frozen sections for immunostaining. The 10-µm frozen sections at 2-mm intervals from the frontal to the occipital poles were cut by cryomicrotomy (CM1900, Leica, Wetzlar, Germany).

To evaluate the effects of paeonol on subacute ischemic injury, the animals were randomly divided into a sham group (saline, n=16), a model group (MCAO + saline, n=23) and a paeonol-treated group (MCAO + Pae at 25 mg/kg, n=21). For drug delivery, the same volume of saline or paeonol solution was injected i.p. at the onset of MCAO (when the nylon suture was inserted and the MCA origin was blocked), and then injected once a day for 3 days. During the experiment, one part of the animals (n=8) were used for determination of infarct volume, and the remaining animals were prepared to cut frozen sections for immunostaining. The 10-µm frozen sections at 2-mm intervals from the frontal to the occipital poles were cut by cryomicrotomy (CM1900, Leica, Wetzlar, Germany).

Drug administration and groups. Paeonol was obtained from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany) with a purity of >99% and dissolved in saline at a concentration of 2 mg/ml. Based on a preliminary experiment, a dose of 25 mg/kg paeonol (i.p.) was used in the present study, which exerted the best neuroprotective effect in acute cerebral ischemia [2,3,5-triphenyltetrazolium chloride (TTC) staining and histopathological results; data not shown]. The regimen of drug administration applied in the present study was established in previous studies (9,24).

To evaluate the effects of paeonol on subacute ischemic injury, the animals were randomly divided into a sham group (saline, n=16), a model group (MCAO + saline, n=23) and a paeonol-treated group (MCAO + Pae at 25 mg/kg, n=21). For drug delivery, the same volume of saline or paeonol solution was injected i.p. at the onset of MCAO (when the nylon suture was inserted and the MCA origin was blocked), and then injected once a day for 3 days. During the experiment, one part of the animals (n=8) were used for determination of infarct volume, and the remaining animals were prepared to cut frozen sections for immunostaining. The 10-µm frozen sections at 2-mm intervals from the frontal to the occipital poles were cut by cryomicrotomy (CM1900, Leica, Wetzlar, Germany).

To evaluate the effects of paeonol on chronic ischemic injury, the animals were randomly divided into a sham group (saline, n=16), a model group (MCAO + saline, n=26) and a paeonol-treated group (MCAO + Pae at 25 mg/kg, n=23) in another separate experiment. For drug delivery, the same volume of saline or paeonol was injected i.p. at the onset of MCAO, then injected once a day from days 2-7, and then once every 2 days from days 8-28.

Behavioral assessment. The neurological deficit score was determined at the indicated time-points according to the established scoring system: In the absence of neurological deficits, the score of 0 was given, upon failure to extend right paw fully, the score of 1 was given, animals circling to the right received the score of 2, falling to the right was scored as 3, and no spontaneous walking and depressed levels of consciousness was scored as 4 (23). The test was repeated for three times and the average value was recorded.

The inclined board test was performed to evaluate the balance and coordination of animals according to previous research (28) with certain modifications, including the
material and size of the board, and the rotation angle (24). Animals were placed on the board (50x30 cm), and once they were stable, the board was inclined from horizontal to vertical at a rate of 2°/sec. The holding angle was defined as the angle at which the animal fell off the board. The test was repeated for three times and the average value was calculated.

One observer who was blinded to the experimental groups performed all of the assessments.

Determination of infarct volume. After the neurological assessment, one part of the animals was re-anesthetized, and the brains were quickly removed and coronally sliced into 6 sections of 2 mm in thickness. The slices were then incubated in 0.5% 2,3,5-triphenyltetrazolium chloride (TTC) at 37°C for 20 min, followed by fixation in 4% paraformaldehyde for 2 h. Subsequently, images of the fixed slices were captured. The infarction area was identified as the unstained area in the brain sections. The infarct volume was calculated by summing up the volumes of the 6 slices, and represented as the percentage of infarction in the total brain hemisphere.

Histological analysis. Following the behavioral tests, the remaining animals were re-anesthetized and then transcardially perfused with 4% paraformaldehyde after a pre-flush with saline. The brains were quickly removed, and were immediately immersed in 4% paraformaldehyde for 24 h, followed by dehydration in 30% sucrose for 3 days, then images were captured with a digital camera. Finally, a series of sections (10 µm) from the frontal to the occipital poles were cut by cryomicrotomy (CM1900; Leica Microsystems, Wetzlar, Germany). The sections were prepared for immunostaining.

To determine the changes in the number of different cell types, immunofluorescence staining was performed. The 10-µm slides were rinsed with PBS and then incubated with 10% normal goat serum (Zhongshan Belling Biotechnology Co., Ltd., Beijing, China) for 2 h at room temperature. Subsequently, the sections were incubated at 4°C overnight with the following primary antibodies: Mouse anti-glial fibrillary acidic protein (GFAP; cat. no. MAB3402; 1:800 dilution; EMD Millipore, Billerica, MA, USA), mouse anti-neuronal nuclei (NeuN; cat. no. MAB377; 1:200 dilution; EMD Millipore) and rabbit anti-ionized calcium binding adaptor molecule 1 (Iba1; cat. no. 019-19741; 1:1,000 dilution; Wako Pure Chemical Industries, Ltd., Wako, Japan). Thereafter, sections were washed three times in PBS and incubated with fluorescein isothiocyanate-conjugated secondary antibody (1:200 dilution; cat. nos. AP124F or AP132F; EMD Millipore) for 2 h at room temperature. For the negative control, PBS was applied instead of the primary antibodies. Images of the stained sections were captured using a fluorescence microscope (Olympus BX51; Olympus, Tokyo, Japan). Eight non-overlapping images for each site of the same rats were randomly selected, and the average value was determined. The neurons and the microglia were calculated according to the average number of stained cells and the astrocytes were calculated according to the average fluorescence intensity.

Statistical analysis. Values are expressed as the mean ± standard error of the mean. Significance of differences was assessed by one-way analysis of variance, followed by Dunnett’s post-hoc test (SPSS 10.0 for Windows; SPSS, Inc., Chicago, IL, USA). The results of the behavioral assessments were analyzed using the Kruskal-Wallis test. P<0.05 was considered to indicate a statistically significant difference.

Results

Physiological changes after the operation. No significant differences in the physiological parameters, including mean arterial blood pressure, partial pressure of CO₂ (PaCO₂), PaO₂, blood glucose, pH and weight were identified between 30 min prior to and after the operation. After cerebral ischemia, the rCBF was ~70% decreased at 30 min after occlusion, and then returned to baseline levels after reperfusion (P<0.01; Table I). Paeonol did not affect these physiological variables, including the reduction of rCBF (Table I). During the 28 days, all of the sham animals were survived. However, no significant difference in the survival rate was identified between the model group (61.54%) and the paeonol-treated group (69.57%), and these rates were consistent with those reported by previous studies (29).

Effect of paeonol on behavioral impairment. In the subacute experiment, behavioral impairments were evaluated at 24 (data not shown) and 72 h after reperfusion (Fig. 1). It was demonstrated that the neurological deficit scores and the holding angle were aggravated at 72 h after reperfusion. Compared with those in the sham group, the neurological deficit scores were significantly increased in the MCAO rats, whereas the holding angle in the inclined board test was significantly decreased (P<0.01; Fig. 1A and C). By contrast, a significant alleviation in the behavioral impairment was observed in the paeonol-treated group compared with that in the model group (P<0.05; Fig. 1A and C).

In the chronic experiment, the measures of behavioral impairment, including the neurological deficit score and the holding angle, were aggravated at 72 h after reperfusion, and then gradually recovered at 7, 14 and 28 days after reperfusion (data not shown). Compared with the model group, paeonol administration led to a more efficient improvement in neurological scores and the holding angle in rats at 72 h after reperfusion. However, no significant difference in the behavioral impairments was identified between these groups from days 28 after reperfusion due to the chronic recovery (P>0.05; Fig. 1B and D).

Effect of paeonol on infarct volumes and ischemic/contra-lateral hemispheric ratio. The representative images of total brains and the TTC-stained coronal slices indicated swelling/atrophy on the surface and infarctions in the hemispheres after cerebral ischemia, respectively (Fig. 2A-B).

The effects of paeonol on subacute brain injury at 72 h after reperfusion were assessed. Compared with those in the sham group, the infarct volume and the ischemic/contra-lateral hemispheric ratio, an index of brain edema, were significantly increased at 72 h after reperfusion (P<0.01; Fig. 1E and G, respectively, and Fig. 2A). However, compared with that in the model group, paeonol treatment significantly alleviated the increase of infarct volume and ischemic/contra-lateral hemispheric ratio at 72 h after reperfusion (P<0.05 or P<0.01;
Fig. 1E and G, respectively, and Fig. 2A). Furthermore, the effects of paeonol on chronic brain injury were assessed at 28 days after reperfusion. Compared with that in the sham group, the ischemic/contralateral hemispheric ratio was markedly decreased at 28 days after reperfusion (P<0.01; Figs. 1H and 2B), indicating brain atrophy, which was consistent with the results of previous studies (9,29,30). By contrast, administration of paeonol markedly alleviated chronic brain atrophy in comparison with that in the model group (P<0.05; Figs. 1H and 2B). However, no significant differences were identified in the infarct volumes between the paeonol-treated and model groups (P>0.05; Figs. 1F and 2B). This may have been due to the apparent atrophy, which hindered the observation of further changes, and the chronic functional recovery. Collectively, these results indicated that paeonol exerted neuroprotective effects on subacute and chronic cerebral ischemic injury.

**Effect of paeonol on neuronal damage.** To assess the effect of paeonol on chronic neuronal injury, the changes of neuronal damage in the ischemic brains after paeonol treatment were observed (Fig. 3). Cerebral ischemia induced evident ischemic lesions exhibiting neuronal injury, which included the shrinkage of cell bodies that were deeply stained using Nissl staining, and the disappearance of Nissl bodies (data not shown). In the ischemic core, compared with that in the sham group, the density of neurons identified by immunofluorescent staining for NeuN was markedly decreased at 72 h after reperfusion, and disappeared at 28 days after reperfusion (P<0.01; Fig. 3A, B and D). In the boundary zone, neuronal density was markedly decreased at 72 h and 28 days after reperfusion (P<0.01; Fig. 3A, C and E). Paeonol treatment significantly ameliorated neuronal loss in the ischemic core, as well as in the boundary zone at 72 h after reperfusion in comparison with that in the model group (P<0.01; Fig. 3A-C). In addition, paeonol treatment markedly inhibited neuronal loss in the boundary zone at 28 days after reperfusion (P<0.01; Fig. 3A and E). However, no significant differences in the neuronal density in the ischemic core area were observed between the paeonol-treated and model groups, as neurons disappeared at 28 days after reperfusion in the ischemic core area (Fig. 3A and D). However, a limited number of degenerated neurons or background staining may be observed (Fig. 3A and D).

**Effect of paeonol on astrocyte proliferation.** To determine the underlying mechanisms involved in the neuroprotective effects of paeonol on subacute and chronic ischemic injury, the present study further assessed the astrocyte responses in the ischemic hemisphere after paeonol administration (Fig. 4). The results indicated that the density of GFAP-positive astrocytes was not significantly changed at 24 h after

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**Table I. Physiological variables prior to and after operation.**

| Variable          | Sham          | Model          | Pae  |
|-------------------|---------------|----------------|------|
| Body weight (g)   | 262.3±11.4    | 272.8±14.3     | 264.4±11.6 |
| MABP (mmHg)       |               |                |      |
| Baseline          | 105.2±10.1    | 102.3±8.9      | 110.0±14.7 |
| 30 min after reperfusion | 111.8±11.2   | 110.1±14.3     | 118.5±10.2 |
| PaO2 (mmHg)       |               |                |      |
| Baseline          | 109.2±4.1     | 105.2±4.5      | 103.8±5.3  |
| 30 min after reperfusion | 104.7±3.0    | 99.8±4.1       | 101.2±5.6  |
| PaCO2 (mmHg)      |               |                |      |
| Baseline          | 38.3±5.1      | 40.4±6.2       | 39.4±4.1   |
| 30 min after reperfusion | 41.2±3.9     | 42.8±4.3       | 44.3±4.9   |
| pH                |               |                |      |
| Baseline          | 7.45±0.38     | 7.38±0.41      | 7.39±0.43  |
| 30 min after reperfusion | 7.40±0.42    | 7.48±0.39      | 7.46±0.36  |
| Glucose (g/l)     |               |                |      |
| Baseline          | 6.58±0.81     | 6.85±0.64      | 6.12±0.53  |
| 30 min after reperfusion | 7.10±0.75    | 6.99±0.68      | 7.08±0.65  |
| rCBF (%)          |               |                |      |
| Baseline          | 100           | 100            | 100   |
| 30 min after ischemia | 98.5±6.2     | 35.8±7.5\*     | 34.6±5.7\* |
| 30 min after reperfusion | 98.9±5.8     | 93.6±8.3       | 95.1±7.9   |

*P<0.01 compared with sham group, analyzed by one-way analysis of variance. The variables were measured 30 min prior to operation (baseline) and 30 min after reperfusion. Values are expressed as the mean ± standard error of the mean for six animals per group. MABP, mean arterial blood pressure; Pa, partial pressure; rCBF, regional cerebral blood flow; Pae, paeonol.
reperfusion (data not shown). However, at 72 h after reperfusion, astrocytes in the boundary zone were significantly increased and hypertrophied, and a glial scar surrounding the ischemic core area had formed at 28 days after reperfusion \((P<0.01; \text{Fig. 4A, C and E})\). By contrast, in the ischemic core area, astrocytes were initially decreased at 72 h after reperfusion, and eventually disappeared at 28 days after reperfusion \((\text{Fig. 4A, B and D})\). Paeonol treatment significantly reduced the density of GFAP-positive astrocytes in the boundary zone at 72 h and 28 days after reperfusion compared with that in the model group \((P<0.01; \text{Fig. 4A, C and E})\). No significant differences were observed in the density of GFAP-positive astrocytes in the ischemic core area between the paeonol-treated and model group at 72 h and 28 days after reperfusion, and the astrocytes gradually disappeared from this area \((\text{Fig. 4A, B and D})\). Overall, the results indicated that paeonol exerted its beneficial effects by reducing post-ischemic astrocyte proliferation.

**Effect of paeonol on microglial activation.** To evaluate whether microglial activation was associated with the neuroprotective effects of paeonol on post-ischemic injury, immunostaining analysis for microglia was performed. Microglia are inflammatory cells that are rapidly activated after brain injury. Activation of microglia involves their proliferation, migration into the injured area, upregulation of various immunomodulators and phagocytosis of the damaged cells and debris. In the cerebral cortex of sham rats, ramified Iba-1-positive microglial cells were diffusely distributed \((\text{Fig. 5A})\). At 24 h after reperfusion, the number of Iba-1-positive microglia was not particularly changed, but the of Iba-1-positive microglia morphology became irregular (data not shown). However, the number of Iba-1-positive cells, including that of the ramified microglia and activated (ameboid/round) microglia/macrophages, was gradually increased at 72 h after reperfusion compared with that in the sham group \((P<0.01; \text{Fig. 5A-C})\). In addition, at 28 days after reperfusion (data not shown).
reperfusion, the number of Iba-1-positive cells, particularly that of activated (ameboid/round) microglia/macrophages, was markedly increased in the ischemic core and boundary zone regions compared with that in the sham group (P<0.01; Fig. 5A, D and E). By contrast, compared with that in the model group, paeonol treatment significantly inhibited the increase in Iba-1-positive microglia and ameliorated the morphological changes in the ischemic core, as well as the boundary zone at 72 h and 28 days after reperfusion (P<0.01; Fig. 5A-E). These results indicated the protective effects of paeonol, which were associated with post-ischemic microglial activation.

Discussion

Traditional Chinese Medicine has been increasingly recognized and has demonstrated a therapeutic significance in the treatment of ischemic stroke (14). Paeonol, an active phenolic component of Cortex Moutan, has been demonstrated to possess diverse biological properties (16,17,19-22). Paeonol has been proved to be effective in treating acute experimental ischemic stroke by inhibiting the excitotoxicity, calcium overload and oxidative stress (19,20). However, to date, the effects of paeonol against subacute/chronic cerebral ischemic injury have remained elusive. Therefore, the present study investigated the effects of paeonol on subacute/chronic cerebral ischemic injury and further determined the underlying mechanisms.

The amphiphilic structure and low molecular weight of paeonol facilitate the easy penetration of the blood brain barrier (19). Thus, in the present study, paeonol was administered to rats by intraperitoneal injection, and its effect against subacute/chronic cerebral ischemic injury was explored. Numerous animal studies on ischemia have established a model where the injury at 24 h following ischemia/reperfusion is regarded as acute injury of cerebral ischemia, that at 72 h to 7 days after reperfusion is regarded as subacute injury and that at 14-35 days following ischemia/reperfusion is regarded as chronic injury (6,9,29,31). Thus, in the present study, the ischemic injury in rats was observed at 72 h and 28 days after cerebral ischemia/reperfusion. The behavioral impairments in these rats were also evaluated in 24 h after reperfusion (data not shown). Consistent with the previous studies, the behavioral impairments were aggravated at 24 h after reperfusion, and paeonol treatment improved their neurological deficits. The major results of the present study demonstrated that the increase in the infarct volume and ischemic/contralateral hemispheric ratio (edema) in the ischemic hemisphere at 72 h after reperfusion and the decrease in the ischemic/contralateral hemispheric ratio (atrophy) at 28 days after reperfusion were significantly attenuated by paeonol treatment. Furthermore, paeonol treatment greatly ameliorated the behavioral impairment at 72 h after reperfusion compared with that in the model group. In addition, paeonol treatment significantly ameliorated neuronal damage in the ischemic core and the boundary zone regions at 28 days after reperfusion. Taken together, it was indicated that paeonol had protective effects on subacute and chronic cerebral ischemic injury.

In addition to the above changes, the present study further investigated the potential mechanisms underlying the protective effects of paeonol on subacute and chronic ischemic stroke. As is known, astrocytes and microglia have pivotal roles in the progression of ischemic stroke (10,32). Accumulating evidence suggests that glial responses require hours to days to fully develop, and may in turn provide potential targets for stroke recovery with longer therapeutic windows compared with those of other treatments (3,13). In addition, post-ischemic inflammation is a crucial step during the process of ischemic stroke (13,33). Thus, the present study subsequently focused on the effects of paeonol on inflammation-associated glial cells.

During the post-ischemic phase, one important event in subacute/chronic ischemic brain injury is reactive astrogliosis and glial scar formation (10,11). As a vital part of the neurovascular unit, astrocytes have an important role in the
Figure 3. Effects of paeonol on neuronal injury at 72 h and 28 days after reperfusion in rats. (A) Representative photomicrographs indicated neurons by immunostaining for NeuN at 72 h and at 28 days after reperfusion (scale bar, 100 µm) and (B-E) quantification results. In the ischemic core, the reduction of neuronal density was significantly attenuated by paeonol treatment at 72 h after reperfusion, whereas NeuN-expressing cells completely disappeared at 28 days after reperfusion. In the boundary zone, the decrease of neuronal density was significantly attenuated by paeonol treatment at 72 h and 28 days after reperfusion. Values are expressed as the mean ± standard error of the mean from eight rats per group. **P<0.01 compared with sham group, ##P<0.01 compared with model group. Pae, paeonol; MCAO, middle cerebral artery occlusion; n.d., not detectable; NeuN, neuronal nuclei.
physiology of the normal brain, which includes blood circulation, extracellular ionic homeostasis, release of energy substrates and growth factors, as well as neurotransmission (3,11). Numerous studies have demonstrated that astrocytes protect the neurons from glutamate excitotoxicity during ischemic stroke (10,11). However, increasing evidence has indicated that astrocytes paradoxically exacerbate ischemic injury with morphological and phenotypic changes; this process is termed reactive gliosis or astrogliosis (7,10). Following focal ischemic injury, reactive astrocytes migrate towards the boundary zone...
and then eventually organize into the glial scar that separates the ischemic core region from healthy tissue (3,7,10). The formation of a glial scar is a critical event in the brain repair responses after ischemic injury, which act as a physical and biochemical barrier that separates the viable and dead tissues, but also obstructs neuronal regeneration by suppressing axonal sprouting (3,10,11). Growing evidence suggests that modulation of reactive astrocytes may be a promising strategy for the

Figure 5. Effects of paeonol on microglial activation at 72 h and 28 days after reperfusion in rats. (A) Representative photomicrographs displaying Iba-1-immunopositive microglia in the ischemic core and boundary zone regions at 72 h and 28 days after reperfusion (scale bar, 100 µm) and (B-E) quantification results. At 72 h after reperfusion, Iba-1-positive microglia initially increased in the ischemic core and the boundary zone regions, and these changes were significantly inhibited by paeonol treatment. At 28 days after reperfusion, microglial cells in the ischemic core and boundary zone regions were significantly increased, which was attenuated by paeonol treatment. Values are expressed as the mean ± standard error of the mean from eight rats per group. **P<0.01 compared with sham group, ##P<0.01 compared with model group. Pae, paeonol; MCAO, middle cerebral artery occlusion; Iba-1, ionized calcium binding adaptor molecule 1.
Guidelines for the early management in the process of chronic cerebral ischemia may lead to the understanding the long-term neuroprotective effects of paeonol. Based on the above results obtained in rats, neuronal loss, and inhibited microglial proliferation and astrocytic activation and astrocyte proliferation. During chronic cerebral ischemia, paeonol markedly ameliorated brain atrophy and subacute/chronic cerebral ischemia, particularly in the late phase of microgliosis. The activation of microglia involves their proliferation and migration into the injured area, upregulation of various immunomodulators, phagocytosis of the damaged cell debris and antigenic substances (8,33,34). However, microglial activation has dual effects, and uncontrolled or overactivated microglia are detrimental for the pathological conditions (35). Activated microglia may aggravate neuronal damage through the release of nitric oxide, reactive oxygen species, protease and proinflammatory cytokines such as tumor necrosis factor-α, as well as interleukin-1β and -6 (35,36). Chronic microglial activation has been considered to be the possible underlying mechanism in the neuronal damage and is associated with numerous neurodegenerative diseases, including ischemic stroke (12,13). Therefore, appropriate identification of agents that inhibit microglial activation may act as effective treatment strategies for providing neuroprotection (12,35). In the present study, paeonol treatment significantly inhibited the increase in Iba-1-positive microglia and ameliorated the morphological changes in the ischemic core and boundary zone areas at 72 h and 28 days after reperfusion. Among the various biological properties, the anti-inflammatory activities of paeonol have been demonstrated in macrophages, microglia and in the BV2 cell line in vitro, which was consistent with the present in vivo results (37,38). It was indicated that paeonol treatment may regulate microglial activation, suggesting the potential use of paeonol in the treatment of post-ischemic microgliosis.

In summary, the present study indicated the protective effects of paeonol and its involvement in the treatment of subacute/chronic cerebral ischemia, particularly in the late phase of microglial activation and astrocyte proliferation. During subacute cerebral ischemia, paeonol effectively alleviated neurological impairment, reduced the infarct volume, cerebral edema and neuronal loss, and suppressed microglial activation and astrocyte proliferation. During chronic cerebral ischemia, paeonol markedly ameliorated brain atrophy and neuronal loss, and inhibited microglial proliferation and astrocyte proliferation. Based on the above results obtained in rats, understanding the long-term neuroprotective effects of paeonol in the process of chronic cerebral ischemia may lead to the development of a novel clinical treatment for ischemic stroke. Overall, the present study broadened the current knowledge on the range of pharmacological properties of paeonol for the treatment of ischemic stroke. However, the precise molecular mechanisms of the neuroprotective effects of paeonol require elucidation in future studies.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

BZ, QJS and HW designed the research. BZ, QJS, ZZZ, HW performed the experiments and data collection. BZ, SYW and XW analyzed the data. BZ, QJS and HW wrote the manuscript. All authors have read and approved this version of the article, and ensure the integrity of the work.

Ethics approval and consent to participate

All experimental procedures were performed in accordance with the National Institutes of Health Guidelines for the Care and Use of Experimental Animals and were approved by the Animal Care and Use Committee of Zhejiang University (Hangzhou, China).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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