INTRODUCTION

There are several methods to assess stress levels in horses caused by exercise, including heart rate, lactate density, and cortisol (Marc et al., 2000). Among these, cortisol is frequently used to assess stress levels induced by exercise (Wellhoener et al., 2004; Malinowski et al., 2006; Ferlazzo et al., 2009). Cortisol hormone produced by the adrenal cortex is one of the main chemical substances indicating the level of stress (Strzelec et al., 2011). For example, Wellhoener et al. (2004) reported that cortisol concentration levels in the human body doubled during an hour of exercise, while Linden et al. (1991) demonstrated that cortisol concentration levels in horses increased after show jumping, cross country, and trotting race.

The methods to measure the level of cortisol concentration include the measurement of the hormone concentration through blood serum, saliva, or urine and the measurement of the amount of neurotransmitter (Mostl and Palme, 2002). For ethical considerations, many previous studies on stress have been conducted through the measurement of cortisol concentration in blood and saliva (Shanahan, 2003; Harewood and McGowan, 2005; Strzelec et al., 2011; Larsson et al., 2013; Peeters et al., 2013). For animals, any method using blood serum can cause additional stress during blood collection (Strzelec et al., 2011). The highest peak was confirmed in Exercise 2 (approximately 131%) of RR group and the lowest peak appeared in Exercise 3 (approximately 52%) of ER group. Therefore, resting without any particular exercise can also increase the stress level of horses. Thus, it is better to exercise, as exercise can reduce the stress level, even in cases when riders are clumsy or lack appropriate horse-riding experience. The results of the present study are useful to equestrian center owners and educational riding instructors in that they provide a meaningful insight into a better horse management. (Key Words: Animal Welfare, Horse, Riding Experience, Stress, Salivary Cortisol)
techniques in using reins and aids are crucial in terms of communicating with the horse (Kang et al., 2010). Therefore, it is expected that horse riding as performed by a person with a poor technique can cause stress in the horse. Said differently, a horse can be stressed by excessive pulling of reins, improper use of aids, and unstable riding posture.

Many previous studies have been conducted on stress in horses, such as trailer loading stress in horses (Shanahan, 2003), horses participating in a dressage competition (Moons et al., 2002; Harewood and McGowan, 2005), horses for endurance competition (Larsson et al., 2013), and horses for jumping (Linden et al., 1991; Vincze et al., 2010; Peeters et al., 2013). In these and other relevant studies, cortisol concentrations were measured and verified using the Enzyme Immuno-Assay (EIA) Kit (Moons et al., 2002; Shanahan, 2003; Strzelecka et al., 2011; 2013). Therefore, the present study aimed to estimate the change of stress levels in horses during different types of exercise by analyzing the cortisol concentration in saliva using the EIA Kit (Salimetrics, Webster, TX, USA).

MATERIALS AND METHODS

Experimental animals and physical exercises

All procedures in the present study were performed in accordance with the IACUC (Institutional Animal Care and Use Committee) of Jeju National University, South Korea. A total of 61 clinically healthy Jeju crossbred horses (28 gelding and 33 mare horses) were divided into the following three groups: i) ten geldings and thirteen mares aged from 5 to 20 years old for tourist riding (TR group, horse-riding experience for tourists; 12.61±5.55 years; n = 23); ii) five geldings and nine mares aged from 5 to 22 years old for resting (RR group, in the paddock without any particular exercise and were tied by a rope (approximately 1.2 m long). They had to wait for tourists while being tied by a rope at least during their standard working time. Thus, the horses in RR group were fastened by a rope for three hours in total. ER group participated in 3-hour riding lessons in the morning (1 hour, between 10 to 11 am) and in the afternoon (each hour; between 13 to 14 pm and 14 to 15 pm). A total of 14 horses and 4 instructors were simultaneously involved in the course of ER group. The horses of TR and ER groups had a free time at the paddock except for the exercise time.

Sampling and measurements

All saliva samples for basal values were taken four times a day: at 07:00 (basal) in the paddock of 25×35 m, 11:00 (Exercise 1), 14:00 (Exercise 2), and 16:00 (Exercise 3). Saliva samples of TR and ER groups were collected in the paddock and those of RR group were collected while the horses were tied up in a line outdoors (except for the basal values).

Saliva samples were collected using Salivette tubes (Sarstedt) with cotton wool (Sarstedt, Numbrecht, Germany). The cotton was inserted and rubbed into the horse's mouth and then placed onto the tongue of the horse using metal tweezers. All saliva samples were frozen at –20°C for 2 hours immediately after collection.

The kit for measuring the concentration of cortisol in the saliva samples was assayed by Salimetrics Salivary Cortisol Enzyme Immuno-Assay Kit (Diagnostic System Laboratories Inc., Webster, TX, USA). On the analysis day, the saliva samples were completely thawed, vortexed, and centrifuged at 1,500×g (at 3,000 rpm) for 15 minutes according to the manual instructions.

We followed the EIA-Kit (Item No. 1-3002-5) instructions. The minimal detectable concentration was 0.003 μg/dL (0.08 nmol/L). The intra-assay and inter-assay coefficient of variation was 3.35% and 3.75%, respectively. The mean recovery was 100.8%±3.7%. The absorbance was
analyzed using a Multiskan FC microplate photometer (Thermo Fisher Scientific, Vantaa, Finland) at 450 nm. To estimate cortisol concentration from the measured sample absorbances, a standard curve was constructed using the known concentrations of cortisol. Furthermore, we calculated the percent bound (B/Bo) for each standard, control, and unknown by dividing the average optical density (OD) (B) by the average OD for the zero (Bo). If the cortisol value exceeded 3.0 μg/dL (82.77 nmol/L), the sample was diluted with an assay diluent and rerun for accurate results. We assayed all samples in duplicate and used the average of the duplicates in the subsequent data analyses.

**Statistical analyses**

Statistical analyses were carried out with the SAS version 8 (SAS Institute Inc., Cary, NC, USA). The differences between the group averages and the cortisol data within each group were analyzed using an analysis of covariance (ANCOVA) for repeated measures with times, groups, and ages. All values were expressed as means± standard deviation, with a statistical significance level set at p<0.05.

**RESULTS**

**Changes of cortisol concentration between horse groups**

The changes of cortisol concentration in the three groups according to the measured time are summarized in Table 1 and Figure 1. In a percentage relative to the base value, cortisol levels at Exercise 3 were confirmed to decrease in all groups as compared to the basal value percentage in the following sequence: ER>TR>RR. The highest peak was confirmed at Exercise 2 (ca. 131%) of RR group and the lowest peak appeared in Exercise 3 (ca. 52%) in ER group (Figure 1).

In the within-group comparison in the measured time, cortisol concentration levels in ER group significantly decreased by approximately 48% at Exercise 3 (2.76±1.94 nmol/L) as compared with the basal value (5.31±3.82 nmol/L) (p<0.01). However, cortisol concentration in RR and TR groups did not change significantly, ranging from the minimum value of 3.07±1.80 nmol/L to the maximum value of 4.01±2.51 nmol/L and the minimum value of 2.96±1.19 nmol/L to the maximum value of 2.44±1.16 nmol/L, respectively.

In the comparison of the three groups in the measured time, salivary cortisol concentration at the basal values was significantly higher in ER group (5.31±3.82 nmol/L) than in TR group (2.68±0.93 nmol/L) and RR group (3.07±1.80 nmol/L) (p<0.05). Salivary cortisol concentration at Exercise 1 was remarkably higher in ER group (4.25±2.27 nmol/L) than in TR group (2.96±1.19 nmol/L) and RR (3.61±1.42 nmol/L) (p<0.05). Salivary cortisol concentrations at Exercise 2 varied among the groups, with the value of 2.44±1.16 nmol/L in TR group, 4.01±2.51 nmol/L in RR group, and 3.35±2.27 nmol/L in ER group (p<0.05). No intergroup difference in salivary cortisol concentration at Exercise 3 was observed (2.59±1.40 nmol/L in TR group, 3.14±1.78 nmol/L in RR group, and 2.76±1.94 nmol/L in ER group).

**Changes of cortisol concentration among horses from different age groups**

Age-specific changes of cortisol concentration according to the measured time are summarized in Table 2 and Figure 2. In a percentage relative to the basal value,
cortisol levels at Exercise 3 were confirmed to decrease as compared to the basal value percentage in the following sequence: A4 > A3 > A2 > A1 sequence. The highest peak was confirmed at Exercise 1 (ca. 130%) of A1 group and the lowest peak appeared at Exercise 3 (ca. 54%) of A4 group (Figure 1).

In the comparison by age according to the measured time, the average cortisol concentration ranged between 3.24±3.33 nmol/L and 5.41±3.33 nmol/L in A1; 2.96±1.80 nmol/L and 3.55±1.40 nmol/L in A2; 2.46±0.91 nmol/L and 3.51±1.88 nmol/L in A3, with no significant differences. However, cortisol concentration in A4 was 4.64±4.22 nmol/L at the basal value and considerably decreased (by ca. 98%, to 2.49±0.86 nmol/L) at Exercise 3 (p<0.05).

In the comparison by age groups of the measured time, no statistically significant differences were observed in cortisol concentration at basal values, ranging from 3.34±1.88 nmol/L to 4.64±4.22 nmol/L. There was a significant difference in cortisol concentration at Exercise 1, ranging from 3.06±1.51 nmol/L to 5.41±3.33 nmol/L (p<0.05). There was also a remarkable difference in cortisol concentration between horses from different age groups at Exercise 2, ranging from 2.48±1.64 nmol/L to 5.02±3.50 nmol/L (p<0.05).

DISCUSSION

According to the analysis results of the three groups collected at 7:00 am, the average cortisol concentration at basal values was 3.81±2.86 nmol/L. In a previous research on salivary cortisol concentration measured at roughly the same time, Peeters et al. (2011; 2013) confirmed that cortisol concentration was 0.58 to 1.77 nmol/L at n = 5 and n = 22 with an average of 1.01±0.62 nmol/L and 1.19±0.54 nmol/L, respectively. In addition, Moons et al. (2002) reported the average cortisol concentration of 2.77±0.45 nmol/L (Arabian horse; n = 10); this does not correspond with the results of the present study. However, since the type of work, environment, and sampling protocols in the present study differed from those used in Peeters et al. (2013), it is difficult to straightforwardly compare cortisol concentration in the two studies.

The meaningful finding of present study is that cortisol levels at Exercise 3 were confirmed to decrease as compared to the basal value percentage in the following sequence: ER > TR > RR. Also, the highest peak was confirmed at Exercise 2 (ca. 131%) in RR group and the lowest peak appeared at Exercise 3 (ca. 52%) in ER group (Figure 1). Said differently, stress levels increased in the non-exercise group (RR) as compared to the exercise groups.

Cortisol level percentage was always the highest in RR group at Exercise 1 (ca. 118%), Exercise 2 (ca. 131%), and Exercise 3 (ca. 102%) and the lowest in ER group at Exercise 1 (ca. 80%), Exercise 2 (ca. 63%), and Exercise 3

| Treatment | Measurement time (Mean±SD) | Significance |
|-----------|-----------------------------|--------------|
|           | Basal | Exercise 1 | Exercise 2 | Exercise 3 |            |
| A1        | 4.16±2.57 | 5.41±3.33 | 5.02±3.50 | 3.24±3.33 | NS          |
| A2        | 3.34±1.88 | 3.55±1.40 | 2.46±0.91 | 2.96±1.80 | NS          |
| A3        | 3.51±1.88 | 3.50±1.21 | 2.46±0.91 | 2.55±0.94 | NS          |
| A4        | 4.64±4.22 | 3.06±1.51 | 2.48±1.64 | 2.49±0.86 | *           |

Significance NS * * NS

SD, standard deviation.
1 A1, 1 to 5 years old; A2, 6 to 10 years old; A3, 11 to 15 years old; A4, 16 to 20 years old.
Levels of significance: NS, not significant; * p<0.05.
* Means with different superscripts in the same column significantly differ (p<0.05).
** Means with different superscripts in the same row significantly differ (p<0.05).
groups at Exercise 3. This coheres with Strzelec et al. (2011), cortisol levels are affected by intensity and length.

In our study, for TR, i.e. the horse group used for tourist riding experience, there was no significant intergroup difference in the collected cortisol concentration. For RR, the collected cortisol concentration either increased or decreased with the passage of time; however, little intergroup difference was observed. Since TR group was the group used for horse riding experience, the horses in TR group exercised at a normal or fast speed. Furthermore, ER had a higher level of cortisol concentration at the basal period, our results demonstrate that the concentration highly decreased in A4 (p<0.05). According to Nogueira and Barnabe (1997) analysis of cortisol concentration by age of racehorses, the oldest group had the lowest cortisol concentration. This result can be considered to reflect the ability of horses to adapt through experience.

In conclusion, salivary cortisol levels were lower in the exercise groups than in the non-exercise group. Also, the effects of cortisol levels according to horses’ age were confirmed to decrease in the following order: A4, A3, A2, and A1. Also, cortisol levels of all groups at Exercise 3 were lower than before basal. In other words, exercise intensity and length did not affect the actual stress of the horses. Therefore, resting without any particular exercise can also increase the stress level in horses. It is better to exercise, since exercise can reduce stress level as compared with non-exercise, even in cases when riders are clumsy riders or lack relevant horse-riding experience.

This study has several limitations. First, the sample with which we investigated the changes in the cortisol concentration level according to age of horses was limited, which makes it difficult to accurately assess. Second, the resting environment was not fully considered, because RR group was tied with a rope and did not graze in the pasture during rest. A more accurate estimate can be made if these limitations are addressed in further research. Despite the limitations, the results of the present study contribute to obtaining basal values and provide a meaningful insight for equestrian center owners and educational riding instructors into a better horse management.

**IMPLICATIONS**

In the present study, stress levels in horses involved in different types of work were confirmed by measuring salivary cortisol concentration. Cortisol levels showed a higher level in the non-exercise group than the exercise groups. In comparison with age, cortisol levels were the lowest in the oldest group of horses. Therefore, the differences in exercise intensity, such as those between walking, trotting, and light canter, did not affect the actual stress of the horses.

**CONFLICT OF INTEREST**

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

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