Semi-refined carrageenan induces eryptosis in a Ca²⁺-dependent manner

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Abstract

Background: Semi-refined carrageenan (food additive E407a) is a widely used thickener, which has been reported to exert toxic and pro-inflammatory effects. In particular, there is accumulating evidence that it induces eryptosis, i.e. a programmed cell death of erythrocytes, via ROS-mediated pathways. However, the role of Ca²⁺-dependent mechanisms in E407a-induced eryptosis is not elucidated.

Material and methods: Semi-refined carrageenan at concentrations of 0 mg/ml, 1 mg/ml, 5 mg/ml and 10 mg/ml was incubated with blood of intact female WAG rats (n=9) for 24 h in RPMI and fetal bovine serum. After 24 h, the samples were used to obtain erythrocyte suspensions. The obtained suspensions were stained with a Ca²⁺-sensitive FLUO4 AM probe (30 min, 2.5 µM). The fluorescence of FLUO4 in erythrocytes was detected by a BD FACSCanto II flow cytometer.

Results: The intracellular Ca²⁺ levels are proportional to the fluorescence of FLUO4. The mean fluorescence intensities (MFI) were compared. Low levels (1 mg/ml) of E407a had no impact on Ca²⁺ concentrations in erythrocytes (p>0.05). On the contrary, high concentrations (5 mg/ml and 10 mg/ml) of this food additive promoted an increase in the intracellular Ca²⁺ levels. The MFI values were 2.3- and 2.5-fold higher, respectively (p<0.0001). In addition, the exposure to E407a at concentrations of 5 mg/ml and 10 mg/ml (p<0.0001) increased the percentage of cells with high FLUO4 fluorescence.

Conclusion: Food additive E407a induces eryptosis in a Ca²⁺-dependent manner.

Key words: calcium ion, erythrocytes, food additives, FLUO4

Introduction

The lifespan of erythrocytes is approximately 120 days. After that period, they are eliminated from the bloodstream by the fixed macrophages. However, in response to unfavorable factors, including lack of energy, hyperosmolality, accumulation of reactive oxygen species (ROS), they undergo cell death referred to as eryptosis [1, 2]. In some ways, it resembles apoptosis of nucleus-containing cells and is characterized by blebbing, shrinkage and plasma phospholipid membrane scrambling [3]. The activation of eryptosis is mediated by oxidative stress (ROS overproduction), elevation of intracellular calcium ions, ceramide formation, etc. [4]. The physiological aim of eryptosis is to prevent destruction of damaged cells by hemolysis, which is associated with membrane rupture and release of alarms or damage-associated molecular patterns (DAMPs). Erythrocyte-derived DAMPs such as hemoglobin, ATP, interleukin-33 and heat shock protein 70 are strongly pro-inflammatory [5]. In addition to endogenous factors, eryptosis is induced by a plethora of xenobiotics, including drugs [6, 7].

Eryptosis contributes significantly to the pathophysiology of many diseases, since its activation is associated with the reduced lifespan of red blood cells and impaired blood clotting, which alters microcirculation [8]. Accelerated erythrocyte removal from the bloodstream via eryptosis favors the development of anemia in thalassemia [4], end stage renal disease [9], malaria [10], sickle cell anemia [2], etc. This suggests that the approaches used to manipulate eryptosis can be applied as therapeutic strategies in diseases associated with anemia [11].

It is important to note that nowadays eryptosis parameters are considered to be markers of biocompatibility and cytotoxicity [12-14]. In particular, eryptosis indices can be used to assess the cytotoxicity of food additives [15]. The safety of one of the numerous food additives available in the market called carrageenan, which is registered in EU countries as either E407 (refined or native carrageenan) or E407a (semi-refined carrageenan), is debating. Carragenans are thickeners and gelling agents whose share in the global food market has been increasing for decades. Their content in food...
and daily intake may vary significantly. In some dairy products, the content of carrageenan can reach up to 0.5% of weight [16]. According to some estimates, the amount of daily ingested carrageenan can reach up to 0.5% of weight [16]. According to some estimates, the amount of daily ingested carrageenan can be up to 7.2 g and this amount has been growing for years due to the prevalence of carrageenan-containing processed foods in Western diet [17]. However, carrageenans have been suggested to induce inflammation [16, 18-24]. The proinflammatory effects attributed to them contributed to setting up a programme for re-evaluating carrageenan safety for consumers (EFSA-Q-2018-00771). Controversial data on the findings of toxicity studies indicate that novel models for assessing the carrageenan safety are required. Semi-refined carrageenan has been shown to induce eryptosis in a dose-dependent way with the involvement of ROS-mediated pathways in activation of this cell death mod [15]. However, it is not clear whether this food additive can promote eryptosis via calcium-mediated pathways.

The aim of this study was to analyze the role of Ca\(^{2+}\)-dependent mechanisms in eryptosis induced by the common food additive E407a.

**Materials and methods**

**Incubation of E407a with blood**

Blood specimens were taken from nine intact adult female WAG rats weighing up to 200 g. To ensure prevention of blood clotting, K2EDTA Vacutainers (IMPROVACUTER Evacuated EDTA K2 Spray Dried PET Tubes, Guangzhou, China) were used. Then blood samples (50 µl) were incubated horizontally with 5 ml RPMI-1640 medium with stable glutamine (Biowest, France) and 5% fetal bovine serum (BioWhittaker®, Lonza, Belgium) during 24h in sterile SPL 15 ml conical tubes. Four tubes were used per rat (E407a concentrations: 0 mg/ml, 1 mg/ml, 5 mg/ml and 10 mg/ml). After incubation, blood samples were centrifuged. Supernatants were discarded. Cell pellets were used to obtain erythrocyte suspensions, which was performed by double washing using phosphate buffer saline (PBS). Thereafter, 2 µl of red blood cell mass was used for FLUO4 AM loading. The study design is demonstrated in Figure 1.

The study was carried out in conformity with the Directive 2010/63/EU for the Protection of Animals Used for Scientific Purposes and the European Convention for the Protection of Vertebrate Animals used for Experimental and other Scientific Purposes (EST 123). The Commission on Ethics and Bioethics (Kharkiv National Medical University, Kharkiv, Ukraine; minutes #5 dated September 17, 2019) approved of the study.

**FLUO4 AM staining**

FLUO4 AM staining was used to assess intraerythrocytic Ca\(^{2+}\) levels by flow cytometry. FLUO4 AM dye powder purchased from Becton Dickinson (USA) was stored at - 20 °C. Prior to the experiment, it was warmed up to room temperature and dissolved in anhydrous dimethyl sulfoxide (Sigma Aldrich, USA) to obtain 5 mM stock solution. The stock solution was used to stain erythrocyte suspensions containing 2 µl of erythrocytes incubated with various concentrations of E407a and 98 µl PBS. The final concentration of Ca\(^{2+}\)-sensitive probe in working solutions was 2.5 µM. The FLUO4-loaded erythrocyte suspensions were incubated in the dark for 30 min. Then 400 µl PBS was added to each tube. Fluorescence was acquired by BD FACScanto™ II system. In each sample, 300 000 events were collected. The fluorescence of FLUO4 was analyzed (FL1 = 530/30 BP). Erythrocyte suspensions with no added FLUO4 were used as negative controls. Hydrogen peroxide-treated (0.1 mM) erythrocytes were used as positive controls.

**Statistical analysis**

The Shapiro-Wilk test allowed assessing the distribution normality. Four independent invariables were compared using the Kruskal-Wallis and post-hoc Dunn’s tests. The mean fluorescence intensities (MFI) of FLUO4 were represented as the median (Me) and interquartile range (IQR; 25%–75%) with p values below 0.05 considered statistically significant. All statistical calculations were performed with Graph Pad Prism 5.0 (USA).

**Results**

Since calcium release into the cytosol of erythrocytes plays a pivotal role in induction of eryptosis, its concentration in red blood cells exposed to E407a was determined by FLUO4 staining. Quantitatively, intracellular Ca\(^{2+}\) levels were estimated by comparing MFI values of FLUO4, which depend on Ca\(^{2+}\) concentrations, and the percentage of erythrocytes with the increased FLUO4 fluorescence.

Representative histograms and MFI values of FLUO4 in all the studies groups of samples are shown in Figures 2 and 3.

![Figure 1 - Diagram of the study design](image)

**Figure 1 - Diagram of the study design**

Blood samples were collected from 9 intact adult female WAG rats in K\(_2\)EDTA vacutainers

Blood aliquots (50 µl) were incubated with various concentrations of E407a (0 mg/ml, 1 mg/ml, 5 mg/ml and 10 mg/ml) in 5 ml RPMI and 5% FBS for 24 h (n=9)

Erythrocyte suspensions obtained from the incubated blood were stained with FLUO4 AM to detect intracellular calcium ion levels

Fluorescence of FLUO4 was acquired by flow cytometry

![Figure 2 - Representative side scatter (SSC) & green fluorescence (FL1) histograms demonstrate a more intense fluorescence of FLUO4 in erythrocytes exposed to the highest concentrations of semi-refined carrageenan (E407a)](image)
The lowest concentration of semi-refined carrageenan, i.e. 1 mg/ml did not promote the elevated FLUO4 fluorescence, evidenced by no statistically significant changes in MFI values of FLUO4 (p=0.25). At the same time, a 2.3-fold elevation of this index was observed in response to 5 mg/ml (p=0.004). The highest amount of semi-refined carrageenan used in this study (10 mg/ml) increased FLUO4 fluorescence over 2.5 times, p<0.0001 (Figure 3).

In addition to the MFI values of FLUO4, we analyzed the amount of erythrocytes with high FLUO4 fluorescence. Low levels of E407a (1 mg/ml) were revealed to have no effects on this parameter (p>0.05), whereas an increase in the concentration of this food additive promoted the increase in the percentage of such cells either 14.6-fold (5 mg/ml) or 20.2-fold (10 mg/ml). These data are represented in Figure 4. It should be noted that the difference is statistically significant (p<0.0001). Thus, changes in both parameters of FLUO4 fluorescence indicate an increase in the intracellular Ca^2+ levels in erythrocytes exposed to over 5 mg/ml E407a.

Discussion
Reports on the mechanism of carrageenan toxicity are multiple, but controversial. There is accumulating evidence that pro-inflammatory effects of E407a are mediated by TLR4 (toll-like receptor 4) signaling, NF-kB (nuclear factor kappa light chain enhancer of activated B cells) transcription factor, ROS-activated pathways and NLRP3 (NOD-, LRR- and pyrin domain-containing protein 3) inflammasome activation [25-27]. All these pathways eventually result in the upregulation of pro-inflammatory cytokines. However, more research efforts should be made to close the knowledge gap on the molecular targets and pro-inflammatory pathways activated by carrageenans.

The study has several strengths. Firstly, FLUO4 AM staining with data acquisition via flow cytometry is a widely recognized and modern technique to assess intracellular Ca^2+ levels. Secondly, two parameters for FLUO4 fluorescence were used. Thirdly, simple experimental design can be easily replicated. The study limitations include: application of only one method to assess intracellular Ca^2+ levels; human erythrocytes were not used.

Conclusion
Semi-refined carrageenan (E407a) promotes eryptosis in a Ca^2+-dependent manner.

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