Correlation of $^{18}$F-fluoroethyl tyrosine positron-emission tomography uptake values and histomorphological findings by stereotactic serial biopsy in newly diagnosed brain tumors using a refined software tool

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Abstract

Objectives: Magnetic resonance imaging (MRI) is the standard neuroimaging method to diagnose neoplastic brain lesions, as well as to perform stereotactic biopsy surgical planning. MRI has the advantage of providing structural anatomical details with high sensitivity, though histological specificity is limited. Although combining MRI with other imaging modalities, such as positron-emission tomography (PET), has proven to increment specificity, exact correlation between PET threshold uptake ratios (URs) and histological diagnosis and grading has not yet been described.

Background: Magnetic resonance imaging (MRI) is the standard neuroimaging method to diagnose neoplastic brain lesions, as well as to perform stereotactic biopsy surgical planning. MRI has the advantage of providing structural anatomical details with high sensitivity, though histological specificity is limited. Although combining MRI with other imaging modalities, such as positron-emission tomography (PET), has proven to increment specificity, exact correlation between PET threshold uptake ratios (URs) and histological diagnosis and grading has not yet been described.

Objectives: The aim of this study was to correlate exactly the histopathological criteria of the biopsy site to its PET uptake value with high spatial resolution (mm³), and to analyze the diagnostic value of PET using the amino acid O-(2-[18F]fluoroethyl)-L-tyrosine ($^{18}$F-FET) PET in patients with newly diagnosed brain lesions in comparison to histological findings obtained from stereotactic serial biopsy.

Patients and methods: A total of 23 adult patients with newly diagnosed brain tumors on MRI were enrolled in this study. Subsequently to diagnoses, all patients underwent a $^{18}$F-FET PET-guided stereotactic biopsy, using an original newly developed software module, which is presented here. Conventional MRI, stereotactic computed tomography series, and $^{18}$F-FET PET images were semiautomatically fused, and hot-spot detection was performed for target planning. UR was determined using the uptake value from the biopsy sites in relation to the contralateral frontal white matter. UR values $\geq 1.6$ were considered positive for glioma. High-grade glioma (HGG) was suspected with URs $\geq 3.0$, while low-grade glioma (LGG) was suspected with URs between 1.6 and 3.0. Stereotactic serial biopsies along the trajectory at multiple sites were performed in millimeter steps, and the FET URs for each site were correlated exactly with a panel of 27 different histopathological markers. Comparisons between FET URs along the biopsy trajectories and the histological diagnoses were made with Pearson product-moment correlation coefficients. Analysis of variance was performed to test for significant differences in maximum UR between different tumor grades.

Results: A total of 363 biopsy specimens were taken from 23 patients by stereotactic serial biopsies. Histological examination revealed eight patients (35%) with an LGG: one with a World Health Organization (WHO)-I lesion and seven with a WHO-II lesion. Thirteen (57%) patients revealed an HGG (two with a WHO-III and three with a WHO-IV tumor), and two patients (9%) showed a process that was neither HGG nor LGG (group X or no-grade group).

The correlation matrix between histological findings and the UR revealed five strong correlation coefficients. Analysis of variance was performed to test for significant differences in maximum UR between different tumor grades.

Running head recto: New hot spot-detection software for stereotactic biopsy

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Introduction

Intracerebral tumors represent a significant health problem and major imaging challenge, comprising a heterogeneous group of tumors, mainly represented by gliomas (50%), other neuroepithelial tumors, lymphomas, and tumors of the myelin sheath. Additional important differential diagnoses include brain metastases, infectious diseases, and demyelinating disorders. The overall annual incidence rate of primary malignant and benign brain tumors in developed countries is approximately 15 per 100,000 individuals, and for primary malignant brain tumors it is seven per 100,000. Brain-tumor incidence and mortality have increased by up to 300% over the past 3 decades, primarily in people aged over 75 years.

Differentiating between a glioma and nonglioma disease, as well as being aware of the exact histopathological grading, is still not possible, even with sophisticated neuroimaging techniques. Given its wide distribution and prevalence, the study of gliomas is one of the pillars of neuro-oncology. It is known that the histological differentiation of gliomas and grading are predictors for both aggressiveness and outcome. High-grade gliomas (HGGs) represent the increased complexity in neuroimaging diagnostics, by evaluating aberrant microvascular proliferations, large tissue invasion, necrosis, and perilesional edema, among others. Such heterogeneity represents a diagnosis and therapeutic challenge when facing a stereotactic biopsy or surgical resection decision. In the former case, it is a priority to sample areas of high malignancy, in order to avoid tumor subgrading. In the latter, it is the definition of tumor borders for surgical resection. Another challenge, of equal magnitude, is postoperative imaging control, since discernment between tumor remnants, treatment-induced tissue changes (ie, necrosis, pseudoprogression and pseudoregression), and tumor proliferation is difficult.

Stereotactic serial biopsy (SSB) is a minimally invasive procedure to obtain tissue from an intracerebral lesion, with low morbidity and mortality. In neuro-oncological patients, SSB has been used to establish histopathological diagnosis prior to therapeutic decisions. At present, magnetic resonance imaging (MRI) is the standard imaging modality which is employed to guide for surgical planning of a brain tumor biopsy or resection in neurosurgery. This is so in part due to the large anatomical detail that the method offers. Advantages of MRI include its accurate morphological and structural neural tissue evaluation, however this method does not provide information about biological tumor activity.

Law et al evaluated the value of conventional and sophisticated MRI techniques like perfusion MRI and proton MR spectroscopy imaging for noninvasive grading of gliomas. Sensitivity, specificity, positive predictive value (PPV), and negative predictive values (NPV) for determining an HGG with conventional MRI were 72.5%, 65.0%, 86.1%, and 44.1%, respectively. Moreover, the integration of cerebral blood-volume measurements, metabolic ratios (choline:creatine, choline:N-acetylaspartate, and N-acetylaspartate:creatine) resulted in increased sensitivity, specificity, PPV, and NPV of 93.3%, 60.0%, 87.5%, and 75.0%, respectively. Differentiating grade III from grade IV tumors in the high-grade group, as well as differentiating grade II from grade I tumors in the low-grade group, was not performed, nor was the cellular origin of the glioma defined (astrocytes, oligodendrocytes, etc).

All these findings may have a significant impact on therapeutic decisions. Therefore molecular imaging using positron-emission tomography (PET) is an interesting alternative that might overcome these limitations of MRI. Based on the tracer used to perform PET, this image modality can provide information about glucose, amino acid, thymidine, and choline metabolism, thereby allowing identification of hypometabolic zones and determining the extent of a proliferation activity.

This method has been used to evaluate patients with gliomas since 1970; however, it is not present in every center around the world, when compared to MRI which is generally cheaper and more available worldwide. PET can characterize the biological process from the metabolic to molecular level, and may define the limits of the tumor.
There are many radiotracers used, depending on the information to be obtained. The use of radiolabeled amino acids in PET, such as O-2-[\(18^F\)]fluoroethyl]-L-tyrosine (\(18^F\)-FET) allows a better delineation of tumoral margins, improving target biopsy and radiotherapy sites, as well as the surgical plan. Other imaging agents used in PET for evaluation of gliomas are: 3,4-dihydroxy-6-[\(18^F\)]fluoro-L-phenylalanine (\(18^F\)-DOPA) and \(^{11}C\)-methionine.

The objective of this original work was to explore the diagnostic value of PET using amino acid \(18^F\)-FET in patients with newly diagnosed brain lesions by an exact comparison of FET uptake ratios (URs) to histological findings obtained from SSB with high spatial resolution. The literature dealing with the diagnostic value of metabolic imaging does not consider this aspect, as the exact definition of URs to the biopsy site has not been possible with conventional stereotactic software tools.

**Patients and methods**

A total of 23 adult patients with newly diagnosed brain lesions on MRI were enrolled in this study, performed at the Neurosurgery Department of Freiburg University Medical Center, Germany. The study was presented to and approved by the Freiburg University ethics committee. An informed consent form was signed by all patients. All patients underwent \(18^F\)-FET PET examination and subsequently PET-guided stereotactic biopsy, using an originally developed software package. Conventional MRI, stereotactic frame computed tomography (CT) images and \(18^F\)-FET PET images were semiautomatically fused, and automatic hot-spot detection within the tumor was performed. The URs along the trajectory and biopsy were automatically calculated in 1 mm steps and analyzed to correlate URs exactly with 27 different histopathological criteria: coagulated blood, diapedesis bleeding, acute hemorrhage, eduate, edema, gliosis, tissue-infiltration zone, low cell density, moderate cell density, strong cell density, pathological pattern of proliferation, prominent blood vessels, mitosis, necrosis, GFAP (GFAP immunoreactivity highlights the glioblastomatous component of the tumor), NogoA (oligodendrocytic marker, strongly expressed in 71% of oligodendrogliaomas), MAP2 (marker for mature neurons), Olig2 (oligodendroglial lineage marker, universally expressed in diffused gliomas, higher in anaplastic oligodendrogliomas versus glioblastomas), p53 (tumor-suppressor gene), tibor peroxidase–antiperoxidase silver impregnation for reticulin (increased reticulin staining highlights the spindle-cell sarcomatous component tumor), vimentin (positive in glial processes of cerebellar pilocytic astrocytomas, in Schwann cell neurinomas, and in endothelial cells of all oncotypes), strong gliosis, central nervous system with cell proliferation, macrophages, lateral fluid percussion (tumor-necrosis factor), periodic acid–Schiff stain (used to detect fungal organisms and cytoplasmic accumulation of glycogen) positive myelin, reactive changes, and lymphocyte infiltration.

**Definition of uptake ratio**

\(18^F\)-FET UR is determined by calculating the relationship between the mean maximum standardized uptake value from the biopsy area and the mean standardized uptake value from an arbitrary reference area. For this work, we employed the contralateral frontal lobe white matter (Figures 1–7). A UR value \(\geq 1.6\) was considered positive for glioma. HGG was suspected with UR \(\geq 3.0\), while low-grade glioma (LGG) was suspected with UR between 1.6 and 3.0.

**Preoperative evaluation**

SSB was indicated as the final diagnostic step to obtain a histopathological diagnosis for brain lesions when clinical
and neuroradiological findings were inconclusive, for lesions not amenable for resection, diffusely infiltrating lesions, and small deep-seated or multiple lesions. In patients with tumors nearby functionally relevant areas and an unclear radiological diagnosis, SSB was first indicated for establishing a histopathological diagnosis and to plan further therapeutic steps, and afterward to offer the patient alternatives to a sometimes risky resection.

Figure 2 Automated hot-spot detection on surgical planning of 18F PET-guided stereotactic serial biopsy.
Notes: (A) 18-F FET-PET guided stereotactic serial biopsy of an insular lesion. The red line indicates the biopsy trajectory until the target point inside the contrast enhancing area. (B) The software allows image fusion between CT-MRI and FET PET. Biopsy target was achieved by means of an automated hot-spot detection system.
Abbreviations: FET, fluoroethyl tyrosine; PET, positron-emission tomography; CT, computed tomography; MRI, magnetic resonance imaging.

Figure 3 Box-and-whisker plots displaying distribution and quartiles.
Notes: Post hoc comparisons using Tukey's honest significant-difference test for the first (A) and second (B) analysis of variance test results, and their 95% family-wise confidence intervals (C and D, respectively). (A) Mean URs of group L (LGG; mean 1.4, SD 0.56) was significantly different from group H (HGG; mean 2.28, SD 0.44), but not from group X (no-grade lesion; mean 1.65, SD 0.64). (B) There were no significant differences between mean URs of group I (WHO tumor grade I; mean 1), group 2 (WHO tumor grade II; mean 1.46, SD 0.57), group 3 (WHO tumor grade III; mean 1.95, SD 0.35), group 4 (WHO tumor grade IV; mean 2.35, SD 0.44) and group X, represented as “0” (no tumor grade; mean 1.65, SD 0.64). The mean score of group 4 was significantly different from that of group 2.
Abbreviations: UR, uptake ratio; LGG, low-grade glioma; HGG, high-grade glioma; SD, standard deviation; WHO, World Health Organization.
For SSB, a modified Riechert–Mundinger stereotactic system was used, and serial biopsies were taken with microforceps with a diameter of 1.4 mm as previously described. Preoperative workup included 1.5 or 3.0 T MRI of 1 mm slices, sagittal T1-weighted MRI after gadolinium enhancement, and additionally a T2-weighted sequence of 1 mm sagittal slices in the case of non-contrast-enhancing lesions.

MRI and 18F-FET PET were taken the day before surgery. Prior to PET, patients fasted for at least 6 hours. Euro PET GmbH (Freiburg, Germany) provided the FET PET medium. Scanning was performed with an ECAT Exact 922 scanner (CTI/Siemens, Knoxville, TN, USA). Per case, a total of 250 MBq of FET was intravenously infused. Emission scans were performed in three-dimensional mode from 10 to 25 minutes after FET injection, with measured attenuation correction. PET data were iteratively reconstructed using the ordered subset-expectation maximization-reconstruction algorithm.

On the day of operation, cranial CT after contrast enhancement with the stereotactic frame mounted to the head of the patient was performed and semi-automatically fused with the MRI and 18-FET-PET images.

**Trajectory planning**

Stereotactic planning was performed using the Inomed planning system (TatraMed SRO, Bratislava, Slovak Republic; Inomed, Emmendingen, Germany). Stereotactic CT was transformed, coregistered, and fused with preoperative MRI and FET PET through the software package.

The biopsy target point (highest FET uptake inside the hot spot) was defined automatically by the software hot spot-detection tool, selecting the area that presented the
Figure 5 FET PET-guided stereotactic biopsy.

Notes: FET UR was retrospectively determined using the uptake value from the biopsy sites in relation to the contralateral frontal white matter. UR ≥1.6 was considered positive for glioma. High-grade glioma was suspected with UR ≥3.0, and low-grade glioma suspected with UR between 1.6 and 3.0. FET PET findings were compared to histological examinations.

Abbreviations: FET, fluoroethyl tyrosine; PET, positron-emission tomography; UR, uptake ratio; R, right; L, left.

Figure 6 18F-FET PET-guided stereotactic serial biopsy of an insular lesion.

Notes: The red line indicates the biopsy trajectory until the target point inside the contrast-enhancing area (A), which coincided with the area of highest metabolic activity in FET PET (B). Due to increased UR and contrast enhancement a high-grade glioma was suspected. Histological analysis, however, revealed the presence of a nonspecific inflammatory process.

Abbreviations: FET, fluoroethyl tyrosine; PET, positron-emission tomography; UR, uptake ratio.
strongest metabolic activity; the trajectory was manually adopted to avoid prominent blood vessels or eloquent brain areas (Figures 4–7).

Statistical analysis

$^{18}$F-FET PET URs were compared with histological examinations. Descriptive statistics were calculated first for the following histopathological characteristics: coagulated blood, diapedesis bleeding, acute hemorrhage, exudate, edema, gliosis, tissue-infiltration zone, low cell density, moderate cell density, strong cell density, pathological pattern of proliferation, prominent blood vessels, mitosis, necrosis, GFAP, NogoA, GFAP, MAP2, Olig2, p53, tibor peroxidase–antiperoxidase silver impregnation for reticulin, vimentin, strong gliosis, central nervous system with cell proliferation, macrophages, lateral fluid percussion- or periodic acid–Schiff-positive myelin, reactive changes, and lymphocyte infiltrates.

The $^{18}$F-FET PET uptake values of all biopsy sites taken along the stereotactic trajectory were calculated exactly with the aid of the newly developed software tool. In order to assess linear dependencies between histological findings and the calculated cortical URs, the Pearson product-moment correlation coefficient was calculated for each finding. We considered $P$-values $<0.05$ as indicators for a significant relationship between UR and histopathological characteristics.

In a second step, significant differences in URs between the groups of HGG, LGG, and other pathological entities were assessed using analysis of variance (ANOVA). Here also, $P<0.05$ was considered to indicate statistical significance.

Results

Patient data

A total of 23 adult patients with image-based newly diagnosed brain tumors were enrolled in this study at the Department...
of Neurosurgery, University of Freiburg, Germany. The population consisted of five females (22%) and 18 males (78%) with ages between 26 and 84 years (average 56 years, median 60 years).

**Histopathology**
A total of 363 tissue samples were taken from these 23 patients by frame-based SSB, with a range of seven to 50 samples per trajectory and an average of 16 samples taken per patient (median 15). Eight patients (35%) were diagnosed with an LGG: one with a World Health Organization (WHO) grade I lesion and seven with a WHO grade II lesion. Another 13 (57%) patients revealed an HGG: two a WHO grade III and eleven a WHO grade IV tumor. Two patients (9%) showed a process that was neither graded HGG nor LGG, called group X or the no-grade group.

**Linear dependencies**
The correlation matrix among histological findings and cortical UR revealed five strong correlations (Table 1). Low cell density in tissue samples was found to have a significant negative correlation with measured cortical UR \( r = -0.43, P = 0.02 \), as well as moderate cell density \( r = -0.48, P = 0.02 \). Additionally to those two negative correlations, we found three positive and significant associations. Pathological pattern of proliferation showed a coefficient of 0.37 \( P = 0.04 \), GFAP also correlated at 0.37 \( P = 0.04 \), and Olig2 showed a strong positive correlation with cortical UR \( r = 0.36, P = 0.05 \).

**Differences between means**
A major objective in the assessment of the diagnostic value of PET is to look for differences between the cortical uptake rates per tumor grading. Consequently, two ANOVAs\(^2\) were calculated, the first to see if there were differences between three grading groups on cortical uptake rate: LGG, HGG, and the no-grade group. The second ANOVA was calculated over all four WHO tumor grades plus the no-grade group.

**First ANOVA (three groups: H, L, and X)**
A one-way ANOVA was calculated to test for differences in cortical UR among three different groups of brain lesions (X, LGG, HGG). Average cortical UR differed significantly across the three types \( F = 8.27, P = 0.002 \) (Table 2).

Post hoc comparisons using Tukey’s honest significant difference test indicated that the mean score of the LGG group (1.4, standard deviation [SD] 0.56) differed significantly \( P = 0.002 \) from the HGG group (2.28, SD 0.44). The mean score of group X (no-grade lesion; 1.65, SD 0.64) was neither significantly different from that of the HGG group \( P = 0.23 \) nor of the LGG group \( P = 0.8 \) (Table 3 and Figure 3).

**Second ANOVA (five groups: 1, 2, 3, 4, and X)**
Another one-way between-group ANOVA was run to find differences in cortical UR among five different grade groups (WHO grades I–IV and group X). Again, a significant

### Table 1 Correlation matrix between histopathological findings and FET uptake values

| Sample | Findings                  | Correlation \( r \) | \( P \)-value (one-sided) |
|--------|---------------------------|---------------------|--------------------------|
| 1      | Coagulated blood          | -0.08               | 0.37                     |
| 2      | Diapedesis/bleeding       | 0.23                | 0.30                     |
| 3      | Acute hemorrhage          | -0.13               | 0.28                     |
| 4      | Exudate                   | 0.09                | 0.35                     |
| 5      | Only edema                | 0.13                | 0.28                     |
| 6      | Gliosis                   | -0.23               | 0.15                     |
| 7      | Tissue-infiltration zone  | 0.17                | 0.22                     |
| 8      | Low cell density          | -0.43               | 0.02*                    |
| 9      | Moderate cell density     | -0.48               | 0.02*                    |
| 10     | Strong cell density       | 0.10                | 0.33                     |
| 11     | Pathological pattern of proliferation | 0.37     | 0.04*                    |
| 12     | Prominent blood vessels   | 0.17                | 0.22                     |
| 13     | Mitosis                   | 0.22                | 0.16                     |
| 14     | Necrosis                  | 0.30                | 0.08                     |
| 15     | GFAP                      | 0.37                | 0.04*                    |
| 16     | NogoA                     | -0.02               | 0.47                     |
| 17     | MAP2                      | -0.23               | 0.15                     |
| 18     | Olig2                     | 0.36                | 0.05*                    |
| 19     | p53                       | -0.33               | 0.06                     |
| 20     | Tibor PAP silver impregnation for reticulin | -0.20  | 0.17                     |
| 21     | Vimentin                  | 0.01                | 0.48                     |
| 22     | Strong gliosis            | 0.13                | 0.28                     |
| 23     | CNS with cell proliferation | 0.04             | 0.43                     |
| 24     | Macrophages               | 0.04                | 0.42                     |
| 25     | LFP- or PAS-positive myelin | 0.04             | 0.43                     |
| 26     | Reactive changes          | -0.02               | 0.46                     |
| 27     | Lymphocyte infiltrates (TILs) | 0.01          | 0.48                     |

**Note:** \( ^*P < 0.05 \).

**Abbreviations:** PAP, peroxidase-antiperoxidase; CNS, central nervous system; LFP, lateral fluid percussion; PAS, periodic acid–Schiff; TILs, tumor-infiltrating lymphocytes; FET, fluoroethyl tyrosine; GFAP, glial fibrillary acidic protein; MAP2, microtubule-associated protein 2; p53, Proteina tumorale 53.
difference between groups was found: \( F=4.5, P=0.01 \) (Table 4). However, Tukey’s honest significance test (Table 5) indicated that the group 1 mean (WHO grade I; 1) was neither different from group 2 (WHO grade II; 1.46, SD 0.57), group 3 (WHO grade III; 1.95, SD 0.35), group 4 (WHO grade IV; 2.35, SD 0.44), nor group X (1.65, SD 0.64). The only significant difference found here was between tumor grades II and IV \( (P=0.012) \) (Figure 3, B and D and Tables S1 and S2).

### Discussion

The aim of this study was to correlate exactly URs of \(^{18}\)F-FET PET examinations to detailed histomorphological criteria in patients with newly diagnosed brain lesions. Amino acid radiotracers used in PET are highly taken up by glial tumors compared to normal brain tissue, resulting in a high tumor-to-brain contrast. Several radiotracers have been suggested to evaluate gliomas for different purposes, such as diagnostic, grading, recurrence, planning, and monitoring therapy. \(^{18}\)F-FET is an artificial amino acid that is increasingly taken up by unregulated tumor cells due to increased metabolism, though not incorporated into proteins.\(^{24,25}\) However, inflammatory and ischemic lesions can also have enhanced amino acid uptake in humans, which could compromise the specificity of the technique. Other imaging agents used in evaluations of gliomas are \(^{18}\)F-DOPA\(^{19}\) and \(^{11}\)C-methionine.\(^{20}\)

In a systematic review and meta-analysis\(^{25}\) on the performance of \(^{18}\)F-FET PET for the differential diagnosis of primary brain tumors, PET showed great performance for newly diagnosed brain lesions, with sensitivity of 82% and 84% and average specificity of 76% and 75% for the diagnosis of brain tumor and glioma, respectively. Etiologic diagnosis based on MRI has high sensitivity, but limited specificity of approximately 68%. The ability of PET to assess tumor nature is superior to that provided by single MRI, and combined PET–MRI-guided diagnostic biopsy increases specificity of 53% for MRI alone and to 94% when combined.\(^{26}\) When both were combined with MR spectroscopy, a specificity of even 97% was achieved.\(^{27}\)

Several authors have evaluated the usefulness of multimodal imaging approaches in the diagnosis of gliomas. Gempt et al.\(^{10}\) evaluated the performance of such an approach for newly diagnosed brain lesions to distinguish the different areas of malignant gliomas and to investigate the predictive power of such an approach toward the histology of the lesions. They performed conventional MRI (MR spectroscopy, perfusion-weighted imaging) and PET of brain lesions prior to surgical resection, established eight points of biopsy sites, and evaluated the performance of each modality both individually and jointly, while biopsy samples were histopathologically assessed. MIB-1 and CD31 markers correlated with findings defined as “tumor” (T2 hyperintensity, contrast enhancement, increased choline:creatine ratios, relative cerebral blood volume, and pathological T:B-cell ratios). Significances in correlation between N-acetylaspartate:creatine ratio and PET tracer uptake were found, and also between choline:creatine, cell density and N-acetylaspartate:creatine ratio. The presence of MIB-1 was also predominant in biopsies marked as “tumor” when obtained by MRI + PET, but not when MRI or PET were applied individually. The authors concluded that lesions targeted with a multimodal imaging approach

### Table 4 The second one-way analysis of variance test\(^{23}\) was used to evaluate differences in cortical uptake rate among five different grades of brain lesions (WHO-I, II, III, IV, and no-grade group)

| Treatment | df | Sum of squares | Mean square | P-value |
|-----------|----|----------------|-------------|---------|
| Treatment | 4  | 4.495         | 1.121       | 4.551   | 0.01   |
| Residuals | 18 | 4.434         | 0.246       |         |        |

Note: "Snedecor distribution.

**Abbreviation:** WHO, World Health Organization.
increased positive correlations with histological indices of malignancy (eg, MIB-1) compared to those targets only biopsied based on amino acid uptake or MRI contrast enhancement alone.

In this study, a significant negative relationship between low-to-moderate cell density and $^{18}$F-FET UR was observed. On the other hand, a significant and positive relationship between $^{18}$F-FET UR and patterns of pathological proliferation, as well as expression of GFAP and Olig-2, was observed. Interestingly, no other histopathological criteria showed a strong correlation with $^{18}$F-FET URs. This is a further indicator that $^{18}$F-FET is selectively incorporated by proliferating astrocytes (GFAP expression) or oligodendrocytes (Olig-2). Based on the results after the implementation of the first ANOVA test, we found that the diagnostic value of $^{18}$F-FET PET is to allow for differentiation between HHG and LGG, but not between the X group and HGG or LGG. The latter probably was due to the fact that inflammation might also accumulate $^{18}$FET. While Tukey’s honest significance test for ANOVA 2 did not show significance between the WHO-I group and group X ($P<0.82$), this can reasonably be explained by the insufficient sample size ($n=1$). According to the observations reported in this paper and those already supported by the literature, the use of PET is not limited to preoperative evaluation, but refines the determination of biopsy targets, improves tumor delineation, and facilitates radiation targets. So far, static $^{18}$FET PET still cannot provide a gradation with acceptable accuracy. Therefore, histological evaluation remains essential in making treatment decisions. However, dynamic FET-PET techniques may overcome these limitations in the future.

**Conclusion**

Although MRI is the standard imaging modality for tumor biopsy or resection in neurosurgery, stereotactic biopsy guided only by this modality may lead to inadequate tumor grading, resulting in undertreatment. $^{18}$F-FET PET provides identification of pathological areas within a newly diagnosed brain lesion, and thus defines the most reliable biopsy target. It also achieves differentiation of LGGs from HGGs with high statistical significance. However, $^{18}$F-FET PET was not able to offer acceptable grading accuracy or cell-type differentiation, which makes histological analysis still indispensable for therapeutic decisions.

**Disclosure**

Inoned partly sponsored this work. The authors report no other conflicts of interest in this work.

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Supplementary materials

Table S1 Histopathological findings, classified and clustered

| No | Findings                      | Samples | Total samples | Sample/total (%) | UR average | UR SD | WHO-I | WHO-II | WHO-III | WHO-IV | Other | LGG | HGG | Patients | All patients | Patients all (%) |
|----|-------------------------------|---------|---------------|------------------|------------|-------|-------|-------|--------|--------|-------|-----|-----|---------|---------------|-----------------|
| 1  | Coagulated blood             | 12      | 363           | 3                | 1.4        | 0.5   | 0     | Yes   | Yes    | Yes    | Yes   | 0    | Yes | 4    | 23            | 17              |
| 2  | Diapedesis bleeding          | 49      | 363           | 13               | 1.8        | 0.6   | 0     | Yes   | Yes    | Yes    | Yes   | 12   | Yes | 52              |                |
| 3  | Acute hemorrhage             | 27      | 363           | 7                | 1.4        | 0.6   | 0     | Yes   | Yes    | Yes    | Yes   | 8    | Yes | 35              |                |
| 4  | Exudate                      | 3       | 363           | 1                | 1.4        | 0.4   | 0     | Yes   | 0      | 0      | Yes   | 1    | Yes | 24              |                |
| 5  | Only edema                   | 2       | 363           | 1                | 1.9        | 0.1   | 0     | Yes   | 0      | Yes    | 0     | Yes | 1    | 23            | 4              |
| 6  | Gliosis                      | 3       | 363           | 1                | 0.8        | 0     | 0     | Yes   | 0      | Yes    | 0     | Yes | 1    | 23            | 4              |
| 7  | Tissue-infiltration zone     | 25      | 363           | 7                | 1.6        | 0.6   | Yes   | Yes   | Yes    | Yes    | Yes   | 10   | Yes | 43              |                |
| 8  | Small cell presence          | 69      | 363           | 19               | 1.1        | 0.6   | Yes   | Yes   | Yes    | Yes    | 0     | Yes | 10   | 23            | 43             |
| 9  | Moderate cell presence       | 77      | 363           | 21               | 1.2        | 0.6   | Yes   | Yes   | Yes    | Yes    | 0     | Yes | 14   | 23            | 61             |
| 10 | Strong cell presence         | 74      | 363           | 20               | 1.7        | 0.6   | Yes   | Yes   | Yes    | Yes    | 0     | Yes | 12   | 23            | 52             |
| 11 | Pathologic pattern of proliferation | 45  | 363           | 12               | 1.9        | 0.6   | 0     | Yes   | Yes    | Yes    | Yes   | 0    | Yes | 9    | 23            | 39             |
| 12 | Prominent blood vessels      | 3       | 363           | 1                | 2.3        | 0.5   | 0     | Yes   | 0      | Yes    | 0     | Yes | 2    | 23            | 9              |
| 13 | Mitosis                      | 24      | 363           | 7                | 1.9        | 0.5   | 0     | Yes   | Yes    | Yes    | 0     | Yes | 7    | 23            | 30             |
| 14 | Necrosis                     | 34      | 363           | 9                | 1.9        | 0.6   | 0     | Yes   | Yes    | Yes    | 0     | Yes | 10   | 23            | 43             |
| 15 | GFAP                         | 23      | 363           | 6                | 1.5        | 0.6   | Yes   | Yes   | Yes    | Yes    | Yes   | Yes | 21   | 23            | 91             |
| 16 | NogoA                        | 2       | 363           | 1                | 1.5        | 0.6   | Yes   | 0     | Yes    | 0      | Yes | 0    | 2    | 23            | 9              |
| 17 | MAP2                         | 3       | 363           | 1                | 1.2        | 0.6   | 0     | Yes   | 0      | Yes    | 0     | Yes | 3    | 23            | 13             |
| 18 | Olig2                        | 17      | 363           | 5                | 2.2        | 0.6   | 0     | Yes   | 0      | Yes    | 0     | Yes | 2    | 23            | 9              |
| 19 | p33                          | 15      | 363           | 4                | 1.3        | 0.6   | Yes   | Yes   | Yes    | Yes    | Yes   | Yes | 11   | 23            | 48             |
| 20 | Tibor PAP silver impregnation for reticulin | 13  | 363           | 4                | 1.3        | 0.6   | 0     | Yes   | 0      | Yes    | 0     | Yes | 5    | 23            | 22             |
| 21 | Vimentin                     | 2       | 363           | 1                | 1.5        | 0.2   | 0     | Yes   | 0      | Yes    | 0     | Yes | 2    | 23            | 9              |
| 22 | Strong gliosis               | 3       | 363           | 1                | 2          | 0.1   | 0     | Yes   | 0      | Yes    | 0     | Yes | 1    | 23            | 4              |
| 23 | CNS with cell proliferation  | 9       | 363           | 2                | 1.6        | 0.6   | 0     | Yes   | Yes    | 0      | Yes | 4    | 23            | 17             |
| 24 | Macrophages                  | 7       | 363           | 2                | 1.6        | 0.6   | 0     | Yes   | Yes    | 0      | Yes | 4    | 23            | 17             |
| 25 | LFP- or PAS positive myelin  | 5       | 363           | 1                | 1.8        | 0.6   | 0     | Yes   | 0      | Yes    | 0     | Yes | 2    | 23            | 9              |
| 26 | Reactive changes             | 7       | 363           | 2                | 1.3        | 0.6   | 0     | Yes   | Yes    | 0      | Yes | 2    | 23            | 9              |
| 27 | Lymphocyte infiltrates (TILs) | 6       | 363           | 2                | 1.3        | 0.6   | 0     | Yes   | 0      | Yes    | 0     | Yes | 2    | 23            | 9              |

Abbreviations: UR, uptake ratio; SD, standard deviation; LGG, low-grade glioma; HGG, high-grade glioma; WHO, World Health Organization; PAP, peroxidase–antiperoxidase; CNS, central nervous system; LFP, lateral fluid percussion; PAS, periodic acid–Schiff; TILs, tumor-infiltrating lymphocytes; No, number.

Analysis of variance 1
Groups: three (other, LGG, HGG)
F=36.28
P<0.001

Post hoc comparison
There were significant differences between groups with regard to UR value:

a) other versus LGG
   ➢ P=0.15
   ➢ no significant difference in UR between other and LGG

b) other versus HGG
   ➢ P=0.0032
   ➢ significant difference in UR between other and HGG

c) LGG versus HGG
   ➢ P<0.01
   ➢ significant difference in UR between LGG and HGG

Analysis of variance 2
Groups: five (other, WHO-I, WHO-II, WHO-III, WHO-IV)
F=40.37
P<0.01

Post hoc comparison
There were significant differences between groups with regard to UR value:

a) other versus WHO-I
   ➢ P<0.01
   ➢ significant difference in UR between other and WHO-I

b) WHO-I versus WHO-II
   ➢ P<0.01
   ➢ significant difference in UR between WHO-I and WHO-II

c) WHO-II versus WHO-III
   ➢ P=0.36
   ➢ no significant difference in UR between WHO-II and WHO-III
Table S2 Analysis of variance (descriptive per group)

| Tumor grade | UR average | UR median | UR SD  |
|-------------|------------|-----------|--------|
| WHO-I       | 0.68       | 0.6       | 0.18   |
| WHO-II      | 1.34       | 1.18      | 0.53   |
| WHO-III     | 1.26       | 1.22      | 0.46   |
| WHO-IV      | 1.87       | 1.89      | 0.52   |
| Other       | 1.34       | 1.21      | 0.52   |
| LGG         | 1.18       | 1.06      | 0.55   |
| HGG         | 1.7        | 1.68      | 0.57   |
| All         | 1.5        | 1.52      | 0.61   |

Abbreviations: UR, uptake ratio; SD, standard deviation; WHO, World Health Organization; LGG, low-grade glioma; HGG, high-grade glioma.

d) WHO-III versus WHO-IV

- $P<0.01$
- significant difference in UR between WHO-III and WHO-IV