Insights to SARS-CoV-2 life cycle, pathophysiology, and rationalized treatments that target COVID-19 clinical complications

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Abstract

Background: Gaining further insights into SARS-CoV-2 routes of infection and the underlying pathobiology of COVID-19 will support the design of rational treatments targeting the life cycle of the virus and/or the adverse effects (e.g., multi-organ collapse) that are triggered by COVID-19-mediated adult respiratory distress syndrome (ARDS) and/or other pathologies.

Main body: COVID-19 is a two-phase disease being marked by (phase 1) increased virus transmission and infection rates due to the wide expression of the main infection-related ACE2, TMPRSS2 and CTSB/L human genes in tissues of the respiratory and gastrointestinal tract, as well as by (phase 2) host- and probably sex- and/or age-specific uncontrolled inflammatory immune responses which drive hyper-cytokinemia, aggressive inflammation and (due to broad organotropism of SARS-CoV-2) collateral tissue damage and systemic failure likely because of imbalanced ACE/ANGII/AT1R and ACE2/ANG(1–7)/MASR axes signaling.

Conclusion: Here we discuss SARS-CoV-2 life cycle and a number of approaches aiming to suppress viral infection rates or propagation; increase virus antigen presentation in order to activate a robust and durable adaptive immune response from the host, and/or mitigate the ARDS-related "cytokine storm" and collateral tissue damage that triggers the severe life-threatening complications of COVID-19.

Keywords: ACE2, ARDS, COVID-19, SARS-CoV-2, TMPRSS2

Introduction

Coronavirus disease 2019 (COVID-19) is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and has resulted in more than 1.45 million of deaths worldwide as of November 30, 2020. While the majority of SARS-CoV-2 infected patients will not require hospitalization, a minority will present with more severe symptoms requiring hospitalization and may experience severe life-threatening complications, including acute respiratory distress syndrome (ARDS), which may trigger a systemic multi-organ collapse [1]. Since SARS-CoV-2 is a new virus and there are few (e.g., Remdesivir; an antiviral drug initially used against hepatitis C virus) [2] anti-viral drugs that have been re-purposed for COVID-19 treatment [3], a better understanding of the underlying COVID-19 pathobiology is required in order to design prophylactic and/or therapeutic strategies. SARS-CoV-2 infects human cells by binding to the cell surface protein angiotensin-converting enzyme 2 (ACE2) through the Receptor Binding Domain (RBD) of its spike (S) protein (Fig. 1) [4]. In addition, the cellular transmembrane serine protease...
2 (TMPRSS2) is required for the priming of the virus S protein [4, 5], while virus entry in the cell may also depend on the endosomal/lysosomal cysteine proteases cysteine B and L (CTSB, CTSL) although their activity is likely not essential [4]. More recently, it was found that furin protease is also involved in the infection process since SARS-CoV-2 contains an unusual for coronaviruses furin cleavage site in the S protein [6], and that the cellular receptor neuropilin-1 (NRP1, binds furin-cleaved substrates) potentiates SARS-CoV-2 infectivity providing also a pathway into the central nervous system [7]; SARS-CoV-2 may also utilize the putative alternative receptor CD147 (expressed in high levels in the brain) to infect cerebral nervous system [8, 9].
The critically balanced ACE/ANGII/AT1R and ACE2/ANG(1–7)/MASR axes

ACE2 is a main component of the renin-angiotensin system (RAS) which maintains fluid and salt balance, as well as blood pressure homeostasis [10]. Renin, angiotensinogen (AGT), angiotensin-converting enzyme (ACE), angiotensin II (ANGII) and the ANGII type 1 and type 2 receptors (AT2R1 and ATR2) (Fig. 1; AT2R is not shown) are major components of RAS. ACE generates ANGII which is a key effector peptide causing vasoconstriction. Overactivation of RAS has been implicated in the pathophysiology of atherosclerosis, heart failure, hypertension, diabetes, renovascular disorders, pulmonary hypertension, pneumonia, fibrosis, and sepsis [11, 12]. On the other hand, ACE2 which has considerable homology (40% identity and 61% similarity) to ACE metalloprotease [13, 14], functions as a negative regulator of the RAS system. Specifically, ACE2 reduces ANGII levels by cleaving it to the sorter ANG(1–7) peptide, which can then activate the vasodilation-promoting and anti-inflammatory MAS receptor (MASR) (Fig. 1) [15]. Furthermore, it has been reported that ACE2 links amino acid malnutrition to intestinal inflammation, as it is a key regulator of innate immunity, dietary amino acid homeostasis and gut microbial ecology [16]. Overall, the balance between the ACE/ANGII/AT1R and the opposing ACE2/ANG(1–7)/MASR axes is central in (among others) the physiological regulation of cardiovascular, blood pressure, neural and kidney functions [10, 12, 15].

It can be assumed that increased ACE2 expression or the co-expression at high levels of the ACE2, TMPRSS2 and CTSB/L proteins in SARS-CoV-2 targeted cells/tissues will correlate with higher risk of viral infection. Reportedly, the ACE2, TMPRSS2 and CTSB/L genes/proteins are widely expressed in human tissues; being particularly enriched in kidney, heart, as well as in tissues of the respiratory and gastrointestinal tract [17]. The ACE2 and TMPRSS2 genes are minimally expressed in blood cells and tend to be co-regulated [17]; it was also found that the SARS-CoV-2 entry factors are expressed at high levels in nasal epithelial cells [18]. These observations suggest that even in the absence of underlying co-morbidities most vital human organs are potentially vulnerable to SARS-CoV-2 infection. It was also found that the ACE2/TMPRSS2 genes are (among others) downregulated by tumor necrosis factor (TNF) and are induced by several pro-inflammatory conditions including Barrett’s esophagus, gastric infection by Helicobacter pylori, obesity, diabetes, autoimmune diseases, as well as by viral infections, cigarette smoking, growth factors, interferons (IFNs) and androgens [17]. In support, ACE2 expression was stimulated by a type I Interferon (IFN-a) gene in human airway epithelial cells [19] and thus, SARS-CoV-2 could (indirectly) exploit IFN-driven upregulation of ACE2 to enhance infection rate in target tissues.

In pathologies like diabetes, obesity, hypertension, respiratory or cardiovascular disease which have all been found to associate with high-risk severe COVID-19 [20], the pro-inflammatory ACE/ANGII/AT1R axis is overactivated triggering the overexpression of the counteracting ACE2 pathway increasing thus SARS-CoV-2 available binding sites. In most cases these patients are treated with anti-hypertensive drugs including ACE inhibitors or ANGII receptor blockers (Fig. 1) [21]; consistently, prescription of anti-hypertensives was more frequent among patients with COVID-19 [22]. Interestingly, it has been shown in mice that SARS-CoV-1 (the coronavirus that caused the SARS epidemic in 2003) infection downregulates ACE2 protein (but not ACE) contributing to severe lung injury [23]. The ACE2-dependent pathogenicity of SARS-CoV-2 has been also confirmed in mice expressing human ACE2 [24]. Suppressed ACE2 expression and locally increased ANGII production can induce leakage of pulmonary blood vessels (a hallmark in ARDS pathogenesis) via AT1R stimulation [25]. Notably, in a model of lung injury being mediated by direct binding of nanoparticles to ACE2, which led to suppression of ACE2 expression levels and activity, administration of losartan (an AT1R antagonist) ameliorated nanoparticle-induced lung injury [26]. Likewise, extensive lung infection by SARS-CoV-2 in COVID-19, triggers capillary leakage which if sustained may lead to viremia (i.e., the presence of infectious virus in the circulation), local over-activation of the ACE/ANGII/AT1R signaling due to ACE2 diminishment, extensive inflammation and the so-called “cytokine storm” (Fig. 2a). Worth mentioning is however, that the etiology of “cytokine storm” remains largely elusive and may be well triggered by mechanisms not directly related to ACE2 through modulation of pulmonary macrophages, dendritic cells and/or neutrophils [27–30]. The alarming “cytokine storm”-related pro-inflammatory signals spread throughout the body most likely triggering ACE2 overexpression and thus increased ACE2/ANGII/AT1R signaling as a counterbalancing effect. Given the extensive expression of ACE2 in most human organs, which is now exaggerated because of the pro-inflammatory alarming cytokines, the potentially (in cases of viremia) circulating virus can attack most vital organs (e.g., kidneys and heart). This vicious cycle may then accelerate due to infection-related locally increased ANGII production, which exaggerates ACE/ANGII/AT1R signaling causing systemic failure. Consistently to this hypothesis, postmortem examination of patients with COVID-19 revealed the existence of SARS-CoV-2 in multiple (apart from the lung) organs including pharynx,
heart, liver, brain, and kidneys further supporting the broad organotropism of the virus [31, 32]. Worth noting is, that although there is seemingly no detectable viremia during asymptomatic infection or in the absence of clinical disease [33], SARS-CoV-2 RNA has been detected in blood samples from patients with mild symptoms [34] and detectable viral RNA in blood is a strong prognostic factor for clinical deterioration [35].

In line with these notions, the enrichment of all SARS-CoV-2 infection-related cellular modules (i.e., ACE2, TMRSS2 and CTSB, CTSL) in the gastrointestinal tract [17, 36] explain diarrhea as a major symptom of COVID-19 and SARS-CoV-2 RNA isolation from stool [33, 37, 38] (Fig. 2b). Given that SARS-CoV-2 productively infects human gut enterocytes [36] or human intestinal organoids [39] it is plausible that human intestinal tract represents a major entry and replication site for SARS-CoV-2 due to consumption of contaminated food. In support, intra-gastric inoculation of SARS-CoV-2 in a mouse model expressing human ACE2 caused productive infection and most interestingly led to pulmonary pathological changes [40]. A significant association between liver dysfunction and mortality of COVID-19 patients has been also reported [41, 42], which may
relate to direct viral infection (still questionable due to relatively low ACE2 expression levels in the liver [17]); to indirect damage because of drug-induced liver injury or because of COVID-19-triggered systemic inflammation [43]. Analyses of severe COVID-19-induced biochemical alterations in the liver have shown the elevation of liver enzymes, such as alanine aminotransferases and aspartate aminotransferases, and significantly lower albumin levels [43, 44] and thus, liver markers should be monitored continuously during COVID-19 evolution. ACE2 and TMPRSS2 are highly expressed in gallbladder [17], whereas regarding pancreas ACE2 is expressed in exocrine tissue microvasculature and in a subset of pancreatic ducts with TMPRSS2 expression being restricted to ductal cells [45, 46]. Notably, both ACE2 and TMPRSS2 are rarely expressed in single pancreatic β cells from donors with or without diabetes [45, 46] suggesting that SARS-CoV-2 cannot directly infect β cells.

Similarly, COVID-19 impact to the nervous system may relate to SARS-CoV-2 infection-mediated systemic imbalance of the neuroprotective ACE2/ANG(1–7)/MASR axis signaling [47, 48] or to direct effects mediated by SARS-CoV-2 neurotropism. SARS-CoV-2 canonical cell entry factors i.e., ACE2 and TMPRSS2 are expressed in glial cells, neurons, endothelial and arterial smooth muscle cells in the brain [49], while neuropilin-11 or CD147 may also facilitate SARS-CoV-2 entry into the central nervous system [7, 9] (Fig. 2c). In support SARS-CoV-2 can directly target neurons of 3D human brain organoids [50, 51] and the virus has been found in cerebrospinal fluid and neuronal cells [49, 52] indicating that SARS-CoV-2 is neuroinvasive, neurotropic and neurovivulent. The two main infection pathways are likely the hematogenous and the neuronal, with the olfactory route (where nasal cell express high levels of ACE2 [18]), along with the lymphatic tissue and the cerebrospinal fluid likely playing a significant role in SARS-CoV-2 neuroinvasion [52]. Most common COVID-19 neurological symptoms are headache, dizziness, hypogeusia and hyposmia, with rarer being severe symptoms like acute cerebrovascular disease, meningitis/encephalitis, acute necrotizing hemorrhagic encephalopathy, or even acute Guillain–Barré syndrome [52]. SARS-CoV-2 can also affect neuronal function indirectly by extensive inflammation-mediated increase of circulating cytokines which can penetrate the damaged blood brain barrier [52].

Conclusively, regarding primary sites of SARS-CoV-2 infection although lungs (Fig. 2a) and likely the gastrointestinal tract (Fig. 2b) are grounds zero during the infection process, SARS-CoV-2 and/or COVID-19 also tear multiple organ systems, with major targets (because of high ACE2, TMPRSS2 expression) being the heart and kidneys.

We propose that (a) the imbalance in the action of ACE- and ACE2-derived peptides, i.e., the increased ANGII versus ANG(1–7) ratio, which occurs due to SARS-CoV-2 binding-mediated diminishment of ACE2 expression, along with, (b) the high ACE-2 expression levels-related increased tropism of the virus to vital human organs (e.g., kidneys and heart) are major drivers of COVID-19 pathobiology. Thus, those at high risk for severe COVID-19 (e.g., the elderly or those with underlying morbidities) should probably be (among others; see below) on prophylactic treatment with RAS inhibitors (e.g., AT1R antagonists or ACE inhibitors) to decrease systemic damage risk and thus blunt COVID-19-associated morbidity and mortality.

Given the aforementioned sequence of events, tissues affected and downstream pathologies, the design of COVID-19 therapeutics (until the discovery of an effective highly specific anti-viral drug and/or a vaccine) may be complex, but it also presents with several potentially druggable opportunities. Overall, it is now understood that acute COVID-19 is a two-phase disease, including (a) infection and spreading of the virus mainly in the respiratory and gastrointestinal tracts, and, (b) ARDS (which can occur after a temporal improvement) and the uncontrolled immune response of the host [53] which can then lead to worsening of ARDS, development of multi-organ pathologies and systemic failure (Fig. 2) [54]. Effective therapeutic treatments should thus probe both SARS-CoV-2 inhibition through better understanding of its life cycle and also the side-effects induced by COVID-19 due to immune system overactivation and organ dysfunction caused by the broad organotropism of SARS-CoV-2.

Targeting the life cycle of the virus

For prophylactically targeting the virus life cycle (phase 1 of the disease), the magic bullet will be the development of an effective vaccine which can induce SARS-CoV-2-specific neutralizing antibodies. Indeed, more than 90 vaccines are being developed against SARS-CoV-2 by researchers in companies and universities worldwide where research teams are trialing different technologies, some of which have not been used in a licensed vaccine before [55]. Most of these vaccines were found to induce protective neutralizing antibodies and CD8+ T cell responses to wild-type (D614) and D614G mutant SARS-CoV-2 in mice, rats, guinea pigs, rabbits, and non-human primates [56–62], while some of them are already being tested in advanced clinical trials with encouraging results [63–65] indicating a potential to provide protection against COVID-19 [66, 67]. Alternatively, the isolation of virus-specific human monoclonal antibodies exerting SARS-CoV-2 neutralization activity from memory B cells (e.g., by high-throughput single-cell sequencing) of
SARS-CoV-2 infected and recovered individuals [68–74] or from genetically-humanized mice [73] can be potentially applied as prophylactic treatment in individuals at high risk for infection or as a post-exposure therapy to limit or treat severe disease. To this end noncompeting antibody cocktails (e.g., REGN10987/REGN10933 or S2E12/S2M11) that target nonoverlapping epitopes on the SARS-CoV-2 S protein can prevent the generation of escape S mutants [75] and are highly effective in both hamsters and rhesus macaques COVID-19 models [76, 77]; additional cocktail (or not) neutralizing antibody preparations were found to be protective in both mice and non-human primates COVID-19 models [78–80] as well as in an interim analysis of a phase 2 trial [81]. Worth mentioning is that beyond the potential prophylactic and/or therapeutic usage of SARS-CoV-2 neutralizing antibodies it would be essential to monitor SARS-CoV-2 seroprevalence and neutralizing activity in donors’ and patients’ blood during the on-going second wave of the COVID-19 pandemic, as well following the initiation of community vaccination; pseudovirus [82] or virus-, cell-free ([83], recently received FDA authorization for use) neutralization assays can be used in this screening.

In another approach, soluble ACE2 (e.g., rhACE2; APN01, GSK2586881), although with less affinity to the virus and half-life as compared to well-selected specific antibodies, can be used as a decoy to neutralize the virus due to competitive binding with cellular ACE2. Supportively, SARS-CoV-2 infections were suppressed in engineered human tissues by clinical-grade soluble human ACE2 [84]. SARS-CoV-2 direct binding to ACE2 can be also suppressed by antibodies or small molecules that target ACE2, such as SSAA09E2, which inhibits SARS-CoV-1 interaction with ACE2 [85]. To reduce the available ACE2 viral binding sites, approaches that suppress ACE2 and/or TMPRSS2 gene expression (e.g., TNF or androgen inhibitors) [17] may be employed. To test this hypothesis, it would be interesting to investigate e.g., whether prostate cancer patients on anti-androgens have less severe COVID-19 due to TMPRSS2 gene suppression. Regarding ACE2 and given the anticipated toxic effects of systemic loss-of ACE2/ANG(1–7)/MASR signaling, any relevant intervention should be transient and with close monitoring of the patients’ clinical condition. An alternative and probably safer, versus ACE2 inhibition, approach would be the use of TMPRSS2 inhibitors. TMPRSS2 is druggable and camostat mesylate partially blocked SARS-CoV-2 cell infection [4] while preliminary reports showed that it reduced the severity of COVID-19 sepsis [86]; thus, several clinical trials are currently ongoing to test whether camostat mesylate could be repurposed and utilized to combat the current pandemic [87]. TMPRSS2 inhibition could also reduce viral tropism at the initial site of SARS-CoV-2 infection and enhance anti-viral humoral immune responses of the host [88, 89]. Additional steps to be targeted at SARS-CoV-2 life cycle include its fusion with cell membrane as well as clathrin-mediated endocytosis. In these pathways, iakuragumycin (clathrin-mediated endocytosis inhibitor; [90]), dynasore or its analogs (dynam inhibitor; [91]), as well as latrunculin B (actin depolymerizing drug, [92]) can be tested in cell-based or preclinical models. Additionally, the small molecule SSAA09E3 was found to suppress the fusion of the host cellular membrane with the virus membrane in a SARS-CoV-1 infection model [85]. Other fusion inhibitors that can be considered include 25-hydrocholesterol which showed broad anti-coronavirus activity by blocking membrane fusion and inhibiting SARS-CoV-2 infection in lung epithelial cells and viral entry in human lung organoids [93].

In the cell, novel SARS-CoV-2-specific antiviral drugs can target the virus’ main protease (Mpro) due to its critical role in processing the polyproteins that are translated from the viral RNA [94, 95] or the SARS-CoV-2 RNA-dependent RNA polymerase (RdRp) that is used for viral genome replication and transcription of viral genes [96]. As a temporary alternative, existing antiviral drugs can be tested (repurposing) since according to molecular docking studies they bind tightly to SARS-CoV-2 RdRp [97]. Here, various anti-polymerase drugs that have been approved for use against other viruses are currently tested and Remdesivir (an RdRp inhibitor) [2] has shown promising clinical effects in patients with severe COVID-19 [98, 99] and in October 22 it became the first COVID-19 drug approved by FDA; worth mentioning, however, is that some well-designed studies, including the WHO’s giant Solidarity trial, challenge its therapeutic value [100]. In general, as was found with other viral pathogens, targeting either Mpro or RdRp are currently the most promising anti-SARS-CoV-2 approaches. Additional intracellular modules of the virus life cycle that can be targeted include tubulin and/or CTSB/L (Fig. 1). Colchicine is an efficient inhibitor of tubulin polymerization [101] and thus of virus loaded endosomes and could be a promising treatment in inhibiting the early phases of virus infection. Similarly, the lysosomotropic compounds chloroquine or hydroxychloroquine inhibit (non-specifically) the activity of endosomal/lysosomal compartments likely reducing the initial phases of viral infection; also, by suppressing lysosomal activity these compounds may also suppress renin production [17] further relaxing the potentially harmful ACE/ANGII/AT1R overactive signaling. Both chloroquine and hydroxychloroquine have been already used as therapeutics against COVID-19 [102]. Yet, given their cardiotoxic effects [103] they should probably be used only in the context of
clinical trials or monitored conditions, and likely in early phases of the infection. Indeed, the RECOVERY trial announced the ending of its hydroxychloroquine arm, as it was concluded that the drug had no clinical benefit for hospitalized patients with COVID-19 [104], while in an open label, randomized controlled trial, adverse events were higher in hydroxychloroquine recipients than in non-recipients [105]. Several existing CTSB/L specific inhibitors, including intracellular regulators (e.g., cathepsins) of cathepsin B activity [106], can be also considered. Notably, cathepsins B/L activity is upregulated during aging [107] and has been correlated with atherosclerotic vascular disease and arterial stiffening [108]; these findings further support the notion that age and cardiovascular morbidities are main risk factors for COVID-19. At the dark side of inhibiting the acidic cellular endosomal/lysosomal compartments or CTSB/L is their functional involvement in MHC class II antigen presentation (Fig. 1) [109]. It can be assumed that the usage of chloroquine/hydroxychloroquine; of CTSB/L specific inhibitors or even the excessive loading of the endosomal compartments by the virus could reduce MHC class II antigen presentation. Suppressed antigen presentation along with the presence of O-linked glycans at the surface of the virus [110] could result in evasion of the immune system. Thus, virus clearance may proceed via proteasome-mediated MHC class I-related antigen cross presentation [111] and the activity of MHC-I dependent cytotoxic T cells. Proteasome activity is downregulated during aging [112] and also proteasome is less responsive to IFN-γ in senescent cells [113], which would then result in reduced antigen presentation by MHC class I molecules and therefore reduced immune responses in aged patients. In this context, small molecules and/or drugs that activate proteasome and in parallel suppress cathepsins B/L activity could provide useful therapeutics against COVID-19. Finally, the finding that SARS-CoV-2 infection reshapes essential cellular pathways, such as nucleic acid metabolism, splicing, translation and carbon metabolism in human cells [114] indicates that a number of small molecule inhibitors that target these pathways could contribute to preventing viral replication in human cells.

**Targeting the adverse effects of COVID-19**

In relation to phase 2 of the disease, i.e., ARDS induction and the alarming “cytokine storm” (Fig. 2a) which can then lead to systemic failure (see above), the ANG(1–7) peptide (or non-peptide analogs) can likely rescue local SARS-CoV-2 infection-related ACE2 loss and the subsequent ANGII accumulation, leading to reactivation of the anti-inflammatory MASR signaling pathway (Fig. 1). Consistently, ANG(1–7) protects endothelial cells from inflammation and high glucose-mediated injury [115], enhances insulin action [116] and was protective in heart failure [117] and stroke [118]. Because of SARS-CoV-2 infection-induced overactivation of the pro-oxidative ACE/ANGII/AT1R axis, which can trigger endothelial dysfunction due to unbalanced reactive oxygen species and nitric oxide ratios in the vessel wall [119], the use of drugs (or small molecules) that activate anti-oxidant cellular defenses [e.g., the Nuclear factor erythroid 2-related factor 2 (Nrf2), pathway] or act as radical scavengers could be an additional prophylactic intervention. Reportedly, COVID-19 may also cause endotheliitis in several organs as a direct consequence of both viral involvement and of the host inflammatory response [120]. Also, COVID-19 correlates with venous thromboembolism and disseminated intravascular coagulation [121, 122]; in this context anticoagulant medications can be used as either prophylactic and/or therapeutic treatment.

Finally, regarding COVID-19-induced “cytokine storm”, i.e., the uncontrolled systemic inflammatory response that relates to the release of high amounts of pro-inflammatory cytokines along with complement components, coagulation dysfunction and immunological “misfiring” [68, 123–126]; the idea of adjunct immunotherapies which inhibit key pro-inflammatory pathways such as IL-6 signaling [127, 128] is a reasonable approach. More specifically, studies in animal models and cell-based assays following SARS-CoV-2 infection, as well as serum and transcriptional profiling of COVID-19 patients, revealed an exaggerated abnormal inflammatory response being marked by reduced levels of type I and III IFNs, along with increased chemokines and IL-6 expression [129]. Also, a single-cell atlas of immune responses in patients with severe COVID-19 revealed a reconfiguration of peripheral immune cells phenotype during life-threatening COVID-19, including HLA class II downregulation, a heterogeneous IFN-stimulated gene signature and a developing neutrophil population that relates to plasmablasts which appear in patients developing ARDS and requiring mechanical ventilation [130]. Interestingly, patients with life-threatening COVID-19 pneumonia (but not with asymptomatic or mild SARS-CoV-2 infection) had neutralizing IgG auto-Abs against IFNs [131] or errors of TLR3- and IRF7-dependent type I IFN immunity [132] suggesting that inborn errors of type I IFN immunity underlies severe COVID-19. Across these lines of research it was found that coordinated CD4+, CD8+ T cells and antibody responses are protective, whereas uncoordinated responses frequently fail to control disease, with a connection between aging and impaired adaptive immune responses to SARS-CoV-2.
Furthermore, lymphocytopenia (especially reduced CD8\(^+\) and CD4\(^+\) T cell counts upon admission), was predictive of disease progression and correlated with high levels of IL-8 and IL-6 also in patients with severe or critical disease [134]. Interestingly, disease severity seems to mostly stem from host factors such as lymphocytopenia, the associated "cytokine storm" and age, whereas genetic variation of the virus was not shown to associate with patients’ clinical outcome [134]. Nonetheless, an excessive inhibition of the immune system by corticosteroids should be avoided since dampening of a cytokine response could allow excessive viral replication. In support, the UK RECOVERY trial found that dexamethasone (a common steroid) could reduce COVID-19 fatalities by as much as one-third when administered to patients who require supplemental oxygen or are on ventilators (RECOVERY); however, dexamethasone treatment has not been shown to offer a benefit for people with mild COVID-19 who do not need oxygen support, possibly because it weakens defenses against the virus itself [135]. Moreover, the observations that IFNs induce ACE2 gene expression [17, 19] prompts for an urgent detailed analysis of how key effectors of the immune system regulate the ACE2, TMPRSS2 and CTSB/L genes and hence, tropism and infection rates of the virus in targeted human tissues.

Interestingly, it has been recently reported the existence of SARS-CoV-2-specific T cells in individuals with

| Table 1 Possible targets to alleviate the life-threatening complications of COVID-19 |
|-------------------------------------------------------|
| **Pre-Phase 1** – Vaccine (e.g., against the SARS-CoV-2 S protein) 1  |
  (for the various technologies employed see also text and [149–152])  |
| **Phase 1 of the disease: Life cycle of the virus (extracellular – early steps of infection)**  |
  - SARS-CoV-2 neutralizing monoclonal antibodies 1  |
    (see text and [153–155]; because antibody-dependent enhancement of disease [156] cannot be reliably predicted after either vaccination or treatment with antibodies, the on-going clinical trials for COVID-19 immune interventions should depend on careful analyses for safety in humans; also, preferentially the development of neutralizing antibodies after vaccination should be monitored) – (neutralizing antibodies from Eli Lilly and Regeneron Pharmaceuticals Inc. have received FDA emergency use authorization and GlaxoSmithKline/Vir Biotechnology has moved an anti-SARS-CoV-2 mAb into Phase 3 clinical trials)  |
  - Soluble ACE2 (decoy for virus) 2  |
    (a recent development in this field is the production of engineered human ACE2 with optimum binding to the S protein of SARS-CoV-2 [157])  |
  - Antibodies or small molecules that target ACE2 2  |
  - Treatments that suppress ACE2 and/or TMPRSS2 genes expression 2  |
  - TMRPSS2 protease inhibitors 3  |
  - Inhibitors of membrane fusion and/or clathrin-mediated endocytosis 4  |
| **Phase 1 of the disease: Life cycle of the virus (intracellular)**  |
  - Tubulin polymerization inhibitors 4 4* 7  |
  - Inhibitors of the endosomal/lysosomal compartments 4,4*  |
  - CTSB/L specific inhibitors 4  |
| **Phase 2 of the disease: adverse effects of COVID-19**  |
  - ACE inhibitors, AT1R blockers 10–12  |
  - The ANG(1–7) peptide (or non-peptide analogs) 10–12  |
  - Antioxidants or radical scavengers 10–12  |
  - Adjunct immunotherapies (or corticosteroids) to mitigate "cytokine storm" (e.g., inhibition of IL-6 signaling) 10–12  |
    (notably, the use of hydrocortisone [160] or dexamethasone [161] showed some beneficial effects on mortality, organ support, days alive and free of mechanical ventilation in patients with severe COVID-19)  |
  - Anticoagulant medications to alleviate intravascular coagulation  |
    (not shown)  |
  - Additional life-supporting measures (e.g., ventilation or intubation)  |
    (not shown)  |

Numbers in bold italics (see, respective red color numbers in Figs. 1, 2) indicate major components in SARS-CoV-2 life cycle and in COVID-19 progression and pathology.
| Intervention | Study type | Biologic efficacy | References |
|-------------|------------|-------------------|------------|
| *Induction of SARS-CoV-2-specific neutralizing antibodies* | | | |
| Recombinant Novel Coronavirus Vaccine | Phase 3 clinical trial (viral two-vector vaccine based on the human adenovirus fused with the S protein of SARS-CoV-2) | Unknown | [162, 163] |
| Adsorbed COVID-19 (inactivated) Vaccine | Phase 3 clinical trial (absorbed inactivated SARS-CoV-2) | Unknown | [164, 165] |
| mRNA-1273 vaccine | Phase 3 clinical trial (mRNA-based vaccine that encodes for a full-length, prefusion stabilized S protein of SARS-CoV-2) | Unknown | [166] |
| **SARS-CoV-2 neutralizing monoclonal antibodies** | | | |
| COV2-2196, COV2-2130 | In vitro and in vivo study (mouse) | Reduce viral burden and level of inflammation in mouse's lungs | see text |
| P2C-1F11, P2B-2F6, P2C-1A3 | In vitro (antibodies derived from B individuals infected with SARS-CoV-2) | Substantial neutralizing activities against SARS-CoV-2 infection | see text |
| CB6 | In vivo (specific human antibodies administered in rhesus macaques) | Prophylactic group: prevention of SARS-CoV-2 infection. Treatment group: reduced SARS-CoV-2 titre | see text |
| **Soluble angiotensin converting enzyme 2 (ACE2) (decoy for virus)** | | | |
| GSK2586881 | Phase 2 clinical trial (recombinant human ACE2 in ventilated patients with ARDS) | Unknown | [167] |
| RhACE2 APN01 | Ongoing phase 2 clinical trial (recombinant human ACE2) | Unknown | [168] |
| **Antibodies or small molecules that target ACE2** | | | |
| SSA09E2 | In vitro (small molecule added to 293 T and Vero cells) | Inhibits fusion of the SARS-S envelope with the host cellular membrane | see text |
| COV2-2196 COV2-2381 | In vivo (monoclonal antibodies administered in rhesus macaques) | Prophylactic group: prevention of SARS-CoV-2 infection | see text |
| **TMRPSS2 protease inhibitors** | | | |
| Camostat mesylate | In vitro (lung cell line) | Blocks SARS-CoV-2 infection of lung cells | see text |
| **Inhibitors of membrane fusion and/or clathrin-mediated endocytosis** | | | |
| Ikarugamycin | In vitro (H1299 cells) | Acutely inhibits clathrin-mediated endocytosis (CME) | see text |
| Dynasore, Dyngo 4a, Dyngo 6a | In vitro | Inhibit specifically dynamin and clathrin-mediated endocytosis | see text |
| Latrunculin b | In vitro | Inhibits Australian bat lyssavirus G-mediated entry into HEK293T cells through actin depolymerization | see text |
| SSA09E3 | In vitro (small molecule added to 293 T and Vero cells) | Prevents fusion of the SARS-CoV-2 membrane with the host cellular membrane | see text |
| **Virus’ RNA-dependent RNA polymerase (RdRp) inhibitors** | | | |
| Setrobuvir, IDX-184, YAK | In vitro | Bind to RdRp tightly and hence may contradict the polymerase function | see text |
| **Cathepsin L inhibitors** | | | |
| SSA09E1, Oxocarbazate, MDL-28170, K11777, EST | In vitro (293 T cells) | Blocks SARS-CoV-2 entry | see text, [169] |
| **Inhibitors of cellular pathways reshaped by SARS-CoV-2 infection** | | | |
| Cycloheximide | In vitro (human Caco2 cells) | Inhibits translation elongation and SARS-CoV-2 replication | see text |
| Emetine | In vitro (human Caco2 cells) | Inhibits the 40S ribosomal protein S14 and SARS-CoV-2 replication | see text |
| Pladienolide B | In vitro (human Caco2 cells) | Inhibits splicing factor SF3B117 and SARS-CoV-2 replication | see text |
| 2-Deoxy-D-glucose | In vitro (human Caco2 cells) | Blocks glycolysis and inhibits SARS-CoV-2 replication | see text |
no history of SARS, COVID-19 or contact with individuals who had SARS or COVID-19; these T cells target (among others) SARS-CoV-2 N protein [136]. Consistently, it was found that S-reactive CD4+ T cells in SARS-CoV-2-unexposed healthy donors reacted primarily to C-terminal S protein epitopes, which show a higher homology to spike glycoproteins of human endemic coronaviruses versus N-terminal epitopes. Moreover, S-reactive T cell lines that were generated from SARS-CoV-2-naive SARS-CoV-2-unexposed healthy donors were found to respond similarly to the S protein (C-terminus) of the human endemic coronaviruses OC43 and 229E and, interestingly enough, of SARS-CoV-2, demonstrating the likely presence of S-cross-reactive T cells, probably generated during past infections with endemic coronaviruses [137]. The presence of S protein cross-reactive T cells in a significant portion of the general population is of critical importance as apart from affecting the dynamics of the current pandemic it may also have important implications for the design and analysis of upcoming COVID-19 vaccine trials.

Towards the development of an effective vaccine, it is encouraging that SARS-CoV-2 is likely mutating slowly with most variants with amino acid changes at RBD being less infectious [138]. Nonetheless, variants which impact on virus infectivity such as the D614G mutation which alters SARS-CoV-2 fitness by increasing its replication ex vivo and transmission in vivo [139–141] or others (e.g., variants A475V, L452R, V483A, F490L) that affect reactivity to neutralizing antibodies and sera from convalescent patients [141, 142] have emerged. Furthermore, it was found that the majority of glycosylation deletions were less infectious, whereas deletion of both N331 and N343 glycosylation drastically reduced infectivity, further supporting the importance of glycosylation for viral infectivity; notably, N234Q was markedly resistant to neutralizing antibodies, whereas N165Q became more sensitive [142]. Thus, a close monitoring of novel SARS-CoV-2 variants with a possible fitness advantage is needed.

### Table 2 (continued)

| Intervention     | Study type                      | Biologic efficacy                                      | References |
|------------------|---------------------------------|--------------------------------------------------------|------------|
| Ribavirin        | In vitro (human Caco2 cells)    | Inhibits inosine monophosphate dehydrogenase and SARS-CoV-2 replication | see text  |
| NMS-873          | In vitro (human Caco2 cells)    | Inhibits the AAA ATPase p97 and SARS-CoV-2 replication | see text  |
| ANG(1–7) peptide | Ongoing Phase 3 clinical trial  | Unknown                                                | [170, 171]|

### Conclusions

COVID-19 is a two-phase disease being marked by (phase 1) rapid virus propagation due to the wide expression of ACE2, TMPRSS2 and CTSB/L genes (along with the other putative alternative receptors and/or attachment factors) in tissues of the respiratory and gastrointestinal tract, as well as by (phase 2) host- [143], and probably sex- [144] and/or age-specific [145, 146] uncontrolled inflammatory immune responses which drive aggressive inflammation, hyper-cytokinemia, and (due to the broad organotropism of SARS-CoV-2) collateral tissue damage and systemic failure because of imbalanced ACE/ANGII/AT1R and ACE2/ANG(1–7)/MASR axes signaling. Discussed notions provide a basis of resources for (a) future investigations of COVID-19 pathogenesis and (b) possible combinatorial therapeutic approaches (Table 1); lists of non-FDA and FDA approved potential treatments targeting specific mechanisms that likely mediate COVID-19 complications are reported in Tables 2, 3 respectively.

It is suggested that the evidence-based (i.e., from both pre-clinical models and clinical trials) use of specific therapeutic approaches/drugs that target modules in pathways 1–12 (Table 1) shown in Figs. 1 and 2, can provide possible means to alleviate the life-threatening complications of COVID-19. In addition, the emerged issues of convalescent plasma treatment effectiveness in severe COVID-19 patients [147]; the duration of anti-SARS-CoV-2 antibodies persistence in mild or severe COVID-19 recovered patients [148] and the possibility of auto-antibodies development against type I IFNs [131] or inborn errors of type I IFN immunity [132] in patients with life-threatening COVID-19 should be further investigated and addressed in adequately powered, randomized controlled trials. Finally, given the fact that COVID-19 has certainly an age-related component [145, 146], as clinical complications mostly develop in the elderly and in patients with non-communicable (age-related) diseases, additional efforts should also focus in those pathways that reportedly become dysfunctional.
Table 3  FDA approved drugs/therapies which can likely target SARS-CoV-2 life cycle and/or COVID-19 complications (see also, Table 1)

| Intervention                                      | Drug category           | Approved indication                                                                 | Biologic and/or clinical efficacy                               | Serious adverse events                                                                 | References |
|---------------------------------------------------|-------------------------|-------------------------------------------------------------------------------------|------------------------------------------------------------------|----------------------------------------------------------------------------------------|------------|
| Suppression of ACE2 gene expression                | Dutasteride 5-a reductase inhibitors | Benign prostate hyperplasia                                                          | Reduces ACE2 and inhibits internalization of SARS-CoV-2 S protein in vitro | None reported                                                                         | [172, 173] |
| Anti-TNF agents                                    | Anti-TNF agents Monoclonal antibodies | Autoimmune diseases                                                                  | Lower incidence of COVID-19 disease in patients with Inflammatory Bowel Disease (Retrospective cohort study and SECURE-IBD database) | Serious infections, demyelinating disorders, drug-induced lupus, may increase risk of malignancies | [174–176] |
| Suppression of TMPRSS2 gene expression             | Homoharringtonine (Omacetaxine) | Protein translation inhibitor Chronic myeloid leukemia                                 | Reduces SARS-CoV-2 pseudoviral entry, in vitro                  | Myelosuppression                                                                       | [177–180] |
|                                                     | Verteporfin Photosensitizer for photodynamic therapy | Subfoveal choroidal neovascularisation                                               | Reduces SARS-CoV-2 pseudoviral entry, in vitro                  | Visual disturbances                                                                    | [177–180] |
|                                                     | Cilnidipine Calcium channel blocker | Hypertension                                                                         | Reduces SARS-CoV-2 pseudoviral entry, in vitro                  | None reported                                                                          | [177–180] |
|                                                     | Dasatinib Tyrosine kinase inhibitor | Chronic myeloid leukemia, acute lymphoblastic leukemia                                | Reduces SARS-CoV-2 pseudoviral entry, in vitro                  | Cytopenia, pleural effusion                                                            | [177–180] |
|                                                     | Venetoclax Selective BCL-2 inhibitor | Chronic lymphocytic leukemia, small lymphocytic lymphoma                              | Reduces SARS-CoV-2 pseudoviral entry, in vitro                  | Neutropenia, lymphopenia, reactivation of hepatitis B, interaction with CYP3A inhibitors and azithromycin | [177–180] |
| Inhibition of TMPRSS2 protease                     | Nafamostat mesylate Serine protease inhibitor | Pancreatitis, anticoagulant during extracorporeal circulation supportive treatment | Inhibits SARS-CoV-2 S-mediated entry into host cells, in vitro  | Bleeding                                                                               | [181, 182] |
| Inhibition of clathrin-mediated endocytosis        | Umifenovir Antiviral | Influenza A and B                                                                     | Its combination with lopinavir/ritonavir ends to better outcome in COVID-19 patients versus only lopinavir/ritonavir | Hepatotoxicity in combination with lopinavir/ritonavir                                 | [183, 184] |
|                                                     | Chlorpromazine Antipsychotic | Schizophrenia, bipolar disorders                                                      | In vitro inhibition of viral replication of coronaviruses      | Parkinsonism, hypotension                                                              | [185, 186] |
| Inhibition of virus' main protease                 | Glecaprevir Antiviral | Hepatitis C                                                                           | Binds with high affinity to SARS-CoV-2 main protease and inhibits it, in vitro | None reported                                                                          | [187]      |
**Table 3 (continued)**

| Intervention | Drug category | Approved indication | Biologic and/or clinical efficacy | Serious adverse events | References |
|--------------|---------------|----------------------|-----------------------------------|------------------------|------------|
| Maraviroc    | Antiviral     | Human Immunodeficiency Virus | Binds with high affinity to SARS-CoV-2 main protease and inhibits it, in vitro | None reported | [187] |

Inhibition of virus’ RNA-dependent RNA polymerase (RdRp)

| Ribavirin    | Antiviral     | Respiratory Syncytial Virus infection, Hepatitis C, some viral hemorrhagic fevers | Bind to the SARS-CoV-2 RdRp tightly in vitro, suppressing its function | Neutropenia, thrombocytopenia, neuropsychiatric toxicity | see text, [188] |
| Remdesivir   | Antiviral     | Broad-spectrum antiviral medication | Bind to the SARS-CoV-2 RdRp tightly in vitro, contradicting its function | Hepatotoxicity, nephrotoxicity | see text, [184] |
| Sofosbuvir   | Antiviral     | Hepatitis C | Bind to the SARS-CoV-2 RdRp tightly in vitro, contradicting its function | None reported | see text |
| Galidesivir  | Antiviral     | Hepatitis C | Bind to the SARS-CoV-2 RdRp tightly in vitro, contradicting its function | None reported | see text, [184] |
| Tenofovir    | Antiviral     | Hepatitis B, Human Immunodeficiency Virus | Bind to the SARS-CoV-2 RdRp tightly in vitro, contradicting its function | Renal and bone toxicity | see text, [189] |
| Favipiravir  | Antiviral     | Influenza | Increases clinical recovery over 7 days and reduces fever, cough, and respiratory problems in COVID-19 patients | Teratogenicity, embryotoxicity | [190] |

Inhibition of tubulin polymerization

| Colchicine   | Anti-inflammatory | Gout, rheumatic diseases, pericarditis | Improves time to clinical deterioration in COVID-19 patients | Diarrhea | see text, [191] |

Inhibition of the endosomal/lysosomal compartments

| Chloquine    | Anti-malarial   | Malaria, lupus erythematosus, rheumatoid arthritis | COVID-19 load reduction/disappearance with hydroxychloroquine | Cardiac arrest, retinotoxicity | see text, [184, 192] |

Cathepsin L inhibitor

| Teicoplanin  | Antibacterial   | Gram positive bacteria (methicillin-resistant Staphylococcus aureus, Enterococcus faecalis) | Inhibits SARS-CoV-2 replication cycle in vitro | None reported | [190, 193] |

Antioxidants

| Thymoquinone | Nutritional supplements | Oxidative stress, vitamin deficiency | Combination of the 3 compounds may prevent and/or decrease SARS-CoV-2 infection severity through activation of Nrf2 transcription factor | None reported | [194] |
| Intervention | Drug category | Approved indication | Biologic and/or clinical efficacy | Serious adverse events | References |
|--------------|---------------|---------------------|-----------------------------------|------------------------|------------|
| Zinc         | Nutritional supplement | Oxidative stress, zinc deficiency | Clinical improvement in COVID-19 patients | None reported | [195, 196] |
| Free radical scavenger | Ergothioneine | Nutritional supplement | Oxidative stress | Potential reduction of severity and mortality of COVID-19 disease | None reported | [197] |
| Immunotherapies to mitigate “cytokine storm” | Tocilizumab | IL-6 receptor blocking monoclonal antibody | Connective tissue diseases | Rapid improvement in clinical and laboratory measures of hyper-inflammation in hospitalized patients with COVID-19 | Neutropenia, thrombocytopenia | [198] |
| Sarilumab | IL-6 receptor blocking monoclonal antibody | Rheumatoid arthritis | Rapid improvement in respiratory function and normalization of inflammatory markers | Neutropenia, thrombocytopenia, upper respiratory and urinary tract infection | [199] |
| Dexamethasone | Corticosteroid | Variety of medical uses | Increase in the number of ventilator-free days in COVID-19 patients with ARDS | Acne, insomnia, vertigo, increased appetite, weight gain, depression | [161] |
| Angiotensin converting enzyme (ACE) inhibitors and angiotensin II type 1 receptor (AT2R1) blockers | ACE inhibitors | Anti-hypertensives | Hypertension | Reduce risk of 28-day death among hospitalized COVID-19 patients with coexisting hypertension and coronary artery disease Decrease the mortality of COVID-19 | Angioedema, anemia | [200–203] |
| | AT1 R blockers | Anti-hypertensives | Hypertension | Decrease mortality in COVID-19 patients with hypertension | Angioneurotic edema, anemia, liver damage, renal failure, aggravation of angina and migraine | [200–203] |
during aging (e.g., reduced proteasome functionality that causes minimized viral MHC antigen presentation, immune senescence, age-related increased inflammation, unbalanced ACE/ANGII/AT1R and ACE2/ANG(1–7)/MASR regulatory axes, etc.) and likely exaggerate the clinical complications of severe COVID-19.

Abbreviations

ACE: Angiotensin converting enzyme; ACE2: Angiotensin converting enzyme 2; AGT: Angiotensinogen; ANGI: Angiotensin II; ARDS: Acute respiratory distress syndrome; AT2R1: ANGII type 1 receptor; AT2R: ANGII type 2 receptor; COVID-19: Coronavirus disease 2019; CTSB: Cathepsin B; CTSL: Cathepsin L; HIV: Human immunodeficiency virus; IFN: Interferon; MASR: Vasodilation-promoting receptor MAS; Mpro: Virus’ main protease; NRP1: Neuropilin-1; RAS: Renin-angiotensin system; RdRp: RNA-dependent RNA polymerase; RBD: Receptor binding domain; S: Spike protein; SARS-CoV-1: Severe acute respiratory syndrome coronavirus 1; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; TMPRSS2: Transmembrane serine protease 2; TNF: Tumor necrosis factor.

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