ABSTRACT

The cranial arterial pattern of artiodactyls deviates significantly from the typical mammalian pattern. One of the most striking atypical features is the rete mirabile epidurale: a subdural arterial meshwork that functionally and anatomically replaces the arteria carotis interna. This meshwork facilitates an exceptional ability to cool the brain, and was thought to be present in all artiodactyls. Recent research, however, has found that species of mouse deer (Artiodactyla: Tragulidae) endemic to the Malay Archipelago possess a complete a. carotis interna instead of a rete mirabile epidurale. As tragulids are the sister group to pecoran ruminants, the lack of a rete mirabile epidurale in these species raises intriguing evolutionary questions about the origin and nature of artiodactyl thermoregulatory cranial vasculature. In this study, cranial arterial patterns are documented for the remaining species within the Tragulidae. Radiopaque latex vascular injection, computed tomography (CT-scanning), and digital 3-dimensional anatomical reconstruction are used to image the cranial arteries of a Sri Lankan spotted chevrotain, Moschiola meminna. Sites of hard and soft tissue interaction were identified, and these osteological correlates were then sought in nine skulls representative of the remaining tragulid species. Both hard and soft tissue surveys confirm that the presence of an a. carotis interna is the common condition for tragulids. Moreover, the use of a 3-D, radiographic anatomical imaging technique enabled identification of a carotico-maxillary anastomosis that may have implications for the evolution of the artiodactyl rete mirabile epidurale.

INTRODUCTION

Among living ruminant artiodactyls, perhaps the most enigmatic clade is the family Tragulidae. Commonly known as mouse deer or chevrotains for their exceptionally small size, Tragulidae is one of the least diverse groups within the artiodactyl suborder Ruminantia (sensu Spaulding, O’Leary & Gatesy, 2009), containing only 3 genera: Tragulus and Moschiola native to South East Asia, and the single African genus Hyemoschus.
(sensu Rössner, 2007). Ranging in length from 45 to 80 cm and 1.5 to 15 kgs in mass, tragulids are the smallest hoofed mammals alive today. When compared to the majority of ruminants, tragulids can be characterized by a number of primitive behaviors and traits. They are, for example, largely nocturnal, lack cranial appendages, and possess a 4-chambered stomach with rudimentary subdivisions and a greatly reduced ommatum (Milne-Edwards, 1864; Carlsson, 1926; Langer, 1974; Dubost, 1975). Biogeographically, they can be found in relict, disjunct populations in the tropical forests of Southeast Asia and West Africa (Grubb, 1993). This suite of primitive traits (detailed in Rössner, 2007) and relict distribution have led to the designation of the group as “living fossils” (Janis, 1984). Because of this, tragulids are commonly used as a model for early ruminant evolution (Métais & Vislobokova, 2007).

Corroborating the notion that tragulids are living fossils is the apparent lack of a rete mirabile epidurale in the lesser and greater mouse deer, Tragulus javanicus and T. napu, respectively (Fig. 1A; Fukuta et al., 2007). The rete mirabile epidurale is a thermoregulatory cranial arterial meshwork that enables artiodactyls to have one of the most advanced capacities for selective brain cooling (reviewed by Caputa, 2004). This structure provides the predominant supply of oxygenated blood to the brain by anatomically and functionally replacing the a. carotis interna, such that in mature individuals, only the pars intracranialis of the a. carotis interna remains (Fig. 1B; Daniel, Dawes & Prichard, 1953). Due to the combined effects of the rete’s high surface area and the reservoir of maxilloturbinate-cooled blood in the rete’s cavernous venous sinus housing, heat is rapidly transferred from arterial blood destined for the brain into to the venous blood that returns to the trunk (Jessen, 1998). As cooled blood flows directly to the hypothalamus, this dissipation of heat is an effective mechanism for delaying heat stress and evaporative water loss, thereby conserving water (Jessen & Pongratz, 1979; Kühnen, 1997; Jessen, 1998; Mitchell et al., 2002; Ostrowski, Williams & Ismael, 2003). Prior to the study conducted by Fukuta and colleagues (2007), all artiodactyls were thought to possess this advantageous structure. The absence of a rete in the genus Tragulus may be the result of (1) their exceptionally small body size, (2) the basal status of the Tragulidae among ruminants, or (3) an adaptation to their unique ecology. Small body size is an unlikely explanation for the tragulid cranial arterial pattern, as other small-bodied ruminants possess a rete (Fukuta et al., 2007). Moreover, scaling functions for continuous endothermy (McNab, 1983) and evaporative water loss (Altman & Dittmer, 1968) indicate that smaller-bodied mammals have a relatively higher thermoregulation-specific metabolic rate and require more water on a mass-specific basis (McNab, 1990). With body size an unlikely explanation, investigations into the evolutionary hypotheses of plesiomorphy or apomorphy may help shed light on why tragulids have an a. carotis interna in lieu of a rete mirabile epidurale. These studies are, however, hampered by a lack of comparative anatomical data.

In this study, the cranial arterial pattern of a second genus of mouse deer, Moschiola, is described in order to confirm the aberrant tragulid cranial arterial pattern outside of Tragulus. Next, this data was used to identify osteological correlates for tragulid cranial arteries that could be sought in a phylogenetically comprehensive sample of much more
Figure 1 Comparison of *Tragulus* and *Capra* arteries of the cranial base, internal surface. (A) The cranial arterial supply of the lesser mouse deer, *Tragulus javanicus*, derives from a large, tubular a. carotis interna which enters the braincase through the anterior foramen lacerum and courses rostrally along the internal surface of the os basisphenoidale. (B) The cranial arterial supply of the domestic goat, *Capra hircus*, derives from a rete mirabile epidurale. The rete is supplied by the (continued on next page...)
readily available skulls. Through this survey of hard and soft tissue cranial morphology, it can be concluded that the absence of a rete mirabile epidurale is common to extant tragulid species. Furthermore, the use of modern anatomical data collection methods enabled the identification of a possible evolutionary mechanism underlying the presence of the rete mirabile epidurale in the derived ruminant condition.

**MATERIALS AND METHODS**

**Soft tissue data collection**

A single individual of *Moschiola meminna* was obtained on loan from the American Museum of Natural History Department of Mammalogy (AM-201747). Tragulids are rare in the wild (Rössner, 2007), and utilization of an alcohol-preserved specimen was the most feasible option for obtaining a specimen. Due to the terms of the specimen loan, physical dissection was not performed in order to maintain the integrity of the specimen for future study. Soft tissue data collection therefore followed non-destructive digital dissection methods.

The left and right arteria carotis communis were dissected, low in the neck, from the whole-body specimen of *Moschiola*, and the left arteria carotis communis was cannulated with an 18-gauge angiographic cannula (Beckton-Dickinson, Worldwide). The cannula was fixed in place with surgical ligature and adhesive. The right a. carotis communis was ligated as a pressure-outlet and means of monitoring injection progress (O’Brien & Williams, 2014). To clear coagulated blood from the vascular tree, the arterial system was manually flushed with warm water for 10 min, followed by perfusion with 90 mL of 40% One-Point anticoagulant solution. A 10% concentration of anticoagulant solution is recommended for most injections, however, due to the long-term storage of the specimen (over a century), a higher concentration of 40% was used in effort to break up large blood clots. Following initial specimen preparation, radiopaque latex vascular injection was conducted, following the protocol of Holliday et al. (2006) and the perfusion criterion of O’Brien & Williams (2014). The specimen was injected with 5 mL of 40% Liquid Polibar Plus barium sulfate suspension (BaSO₄, E-Z-Em, Westbury, NY, USA) in red liquid latex injection medium (Ward’s, Rochester, NY, USA). Perfusion continued until latex emerged from the contralateral a. carotis communis. Acetic acid (10% glacial acetic acid solution) was used to set any extravasated latex.

Following radiopaque latex injection, the specimen was CT scanned at the Holzer Clinic in Athens, Ohio, on a Philips Brilliance 64-slice CT scanner. By scanning at 0.67 mm slice thickness, 150 kV, and 80 mA, a voxel size of 0.693359 × 0.693359 × 0.5 mm
was achieved. The resultant data were up-sampled to a size of 0.1 × 0.1 × 0.1 mm during post-processing in Avizo (version 7.0; VSG). Up-sampling does not affect the inherent quality of the data, and is a reliable technique for generating a visually smoother surface upon reconstruction. Because a 40% barium concentration yields stark contrast between hard tissues, the skull and arteries were segmented based on distinctive gray-scale values. Manual segmentation was then employed to verify and edit the accuracy of the model. Segmented morphology was then rendered in three dimensions (Figs. 2 and 3; http://figshare.com/s/e22fde06688711e5babe06ec4bbcf141; http://figshare.com/s/f1f3cb9a688711e5ab6506ec4bbcf141).

**Hard tissue data collection**

Skulls of mature individuals representing the four conventional species of tragulids were examined for osteological correlates (Table 1; for taxonomy see discussion in Rössner, 2007). Presence or absence of each correlate was entered into a character matrix using Mesquite (v. 2.75, Maddison & Maddison, 2011; Table 1). The cranial osteology of tragulids was compared with artiodactyl species known to possess a rete mirabile epidurale, including the domestic goat (Capra hircus) and the white-tailed deer (Odocoileus virginianus), as well as the domestic horse, Equus ferus caballus, a perissodactyl with an a. carotis interna (Table 1).

Osteological correlates are defined as a “causal relationship” between soft tissues and their surrounding osteology (Witmer, 1995), so the ontogeny and homology of the a. carotis interna were taken into account. The embryonic development of cranial vasculature in Tragulus javanicus has been detailed by Wible (1984). Because skull development continues post-partum, skulls of Tragulus javanicus (3) and Hyemoschus aquaticus cottoni (1) neonates and juveniles were obtained on loan from the American Museum of Natural History Department of Mammalogy. These specimens ranged in relative age from neonate (AM-107113) and early juvenile (AM-188310) to sub-adult (AM-106302 and AM-53634), as approximated by extent of cartilage, degree of suture closure, and skull length (as in Powers, 1962; Morris, 1972; Mitchell, 1973; Meindl & Lovejoy, 1985; Brunner, 1998; Sanchez-Villagra, 2010).

**RESULTS**

**Soft tissue results: cranial arterial patterns of Moschiola meminna**

**Major distribution of the carotid arteries**

In Moschiola, the arteria carotis communis (CCA) ascends through the neck and bifurcates into substantial arteria carotis externa and interna (ECA and ICA, respectively) approximately 1 cm caudal to the apparatus hyoideus (Fig. 2). In other ruminants, the a. carotis communis continues as the ECA and the pars extracraniialis of the ICA obliterates throughout early ontogeny. As expected for non-artiodactyl mammals, the ICA ascends without branching to the foramen lacerum, through which the vessel enters the braincase (Fig. 2). Immediately after passing through the foramen lacerum, the ICA courses rostrally over the internal surface of the os basisphenoidale (Fig. 3), leaving deep arterial grooves.
Figure 2 Cranial arteries of *Moschiola memmina*. (A) Left lateral view; (B) Sagittal section, left medial view. Abbreviations—ALI, arteria labialis inferior; ALS, arteria labialis superior; APL, arteria profunda linguae; ASL, arteria sublingualis; BA, arteria basilaris; BUC, arteria buccalis; CAC, circulus arteriosus cerebri; cAUR, arteria auricularis caudalis; CCA, arteria carotis communis; CRR, arteria cerebri rostralis; ECA, arteria carotis externa; eICA, arteria carotis interna pars extracranialis; ETI, arteria ethmoidalis interna; IO, arteria infraorbitalis; FA, arteria facialis; II CA, arteria carotis interna pars intracranialis; LA, arteria lingualis; MA, arteria maxillaris; OC, arteria occipitalis; OE, arteria ophthalmica externa; ST, arteria temporalis superficialis; TFL, truncus linguofacialis.

O’Brien (2015), PeerJ, DOI 10.7717/peerj.1451
The pars intracranialis of the ICA then distributes branches to the circulus arteriosus cerebri.

The a. carotis externa gives rise to a number of distributing vessels. At the level of its bifurcation with the ICA, the ECA gives off the rostrally-coursing arteria lingualis, the laterally-coursing arteria facialis (these vessels arise together from a short truncus linguofacialis), and two large rostral and caudal terminations—the arteria maxillaris (MA) and the arteria temporalis superficialis, respectively (Fig. 2). Near its departure from the ECA, the a. temporalis superficialis gives rise to the arteria auricularis caudalis, such that these two vessels share a very short common trunk (Fig. 2). The a. auricularis caudalis then ascends close to the caudal border of the meatus acusticus externus, superficial to the processus jugularis (Fig. 2). Smaller branches arise from the a. auricularis caudalis.
Table 1 Specimens examined and osteological correlates. In addition to mature, adult specimens, specimens of sub-adult, early juvenile, and neonate tragulids were examined to account for ontogenetic changes in osteology. The following character states were used: Carotid groove, os basisphenoidale—Character state [1]: sulcus caroticus present on the internal surface of the os basisphenoidale. Correlates to presence of the a. carotis interna. Character state [0]: internal surface of the os basisphenoidale is smooth medial to the lateral sellar compartment, and lacks a sulcus caroticus. Correlates to presence of the rete mirabile epidurale. Medial bullar groove—Character state [1] sulcus caroticus present on the medial surface of the bulla tympanica. Correlates with presence of the a. carotis interna. Character state [0]: sulcus caroticus absent on the medial surface of the bulla tympanica. Correlates with presence of the rete mirabile epidurale. Foramen orbitotorundum—Character state [1]: ventromedial notch in the wall of the presphenoid/foramen orbitotorundum is present. Correlates with a tubular a. maxillaris and indicates the presence of an a. carotis interna. Character state [0]: foramen orbitotorundum un-notched. Correlates with the diffuse ramus that supply the rete mirabile epidurale from the a. maxillaris and indicates the presence of the rete mirabile epidurale. Foramen lacerum—Character state [1]: the foramen lacerum is notched due to the presence of an a. carotis interna. Character state [0]: the foramen lacerum is absent or un-notched, indicating the absence of an a. carotis interna. Foramen ovale—Character state [1]: foramen ovale is notched and/or trapezoidal in contour. Correlates with rami supplying the rete mirabile epidurale from the a. maxillaris. Character state [0]: foramen ovale is un-notched and oval in contour. Correlates with the absence of rami supplying the rete mirabile epidurale.

| Specimen information | Osteological correlates |
|----------------------|-------------------------|
| **Taxon** | **Specimen no.** | **Approximate age** | **Carotid groove:** BSPH | **Medial bullar groove** | **Foramen orbitotorundum** | **F. orbitotorundum:** ventromedial notch | **For. lacerum:** notched | **For. ovale:** notched |
| Hyemoschus aquaticus | AM-89414 | Adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Hyemoschus aquaticus | AM-146844 | Adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Hyemoschus aquaticus | CM-2692 | Adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Hyemoschus aquaticus | AM-53634 | Sub-adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Tragulus javanicus | AM-106302 | Sub-adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Tragulus javanicus | AM-107113 | Neonate | 0 | 0 | 1 | 0 | 0 | 0 |
| Tragulus javanicus | AM-188310 | Early Juvenile | 1 | 0 | 1 | 0 | 0 | 0 |
| Tragulus napu | CM-88224 | Adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Moschiola memminna | AM-201747 | Adult | 1 | 1 | 1 | 1 | 1 | 0 |
| Odocoileus virginianus | OUVC | Adult | 0 | 0 | 1 | 0 | 0 | 1 |
| Capra hircus | OUVC | Adult | 0 | 0 | 1 | 0 | 0 | 1 |
| Equus ferus caballus | OUVC | Adult | 1 | – | 0 | – | 1 | 0 |

that distribute to the mastoid and occipital regions until the artery terminates on the dorsal surface of the os temporale pars squamosa. The pinna of the tragulid ear is small in relative and absolute terms, and the latex failed to perfuse the cartilaginous and soft tissue portions of the ear, perhaps due to the reduced caliber of arteries that supply this region. The a. stylomastoideus is a small lateral branch of the a. auricularis caudalis that enters the foramen stylomastoideus, courses through the canalis facialis, and ramifies the middle ear.

The a. temporalsuperficialis is the dorsal termination of the ECA. This artery follows the caudal border of the ramus mandibulae toward the processus temporalis of the os zygomaticum (Fig. 2). The largest branch of the a. temporalsuperficialis is the arteria transversa faciei, which courses rostrally over the m. masseter (Fig. 2). Smaller muscular branches perforate the caudal portion of the m. masseter and m. temporalis. A minute a. auricularis rostralis splits from the a. temporalsuperficialis before the parent artery terminates within the m. temporalis. The a. transversa faciei crosses the caudal border of the ramus mandibulae ventral to the processus condylaris (Fig. 2). The a. transversa faciei
does not supply the articulatio temporomandibularis as is common among non-tragulid ruminants.

**Superficial arteries of the face and scalp**

The a. facialis branches from the truncus linguofacialis somewhat higher than the arteria lingualis (Fig. 2). Near its origin, the a. facialis distributes several branches to the glandulae parotis and deeper parenchyma caudal and lateral to the border of the ramus mandibulae. The artery then crosses over the margo ventralis of the corpus mandibulae to course dorsally, superficial to the parotid region. The a. facialis terminates as the a. labialis inferior and the a. labialis superior. The a. labialis inferior, in turn, has multiple branches, both superficial and deep, as it courses superficial to the mandibular dentition to the inferior labium. The a. labialis superior parallels the maxillary dentition, ultimately dispersing within the muscle, skin, and mucosa of the superior labium.

The arteria occipitalis departs directly from the medial surface of the ECA, etching a groove along the posteromedial surface of the bulla tympanica as it ascends. Unlike the a. auricularis caudalis, the a. occipitalis passes medial to the processus jugularis (Fig. 2). The a. occipitalis gives off arteria condylaris and meningea caudalis branches before distributing over the occipital region. The a. meningea caudalis passes through the mastoid foramen to supply oxygenated blood to the caudal meninges.

**Branches and distribution of the maxillary artery**

The a. maxillaris is the largest rostral termination of the ECA, and as is common to other ruminants, supplies the brain, orbit, palate, nasal vestibule, and the superficial and deep structures of the face dorsal to the maxillary dentition and labium. In this specimen of Moschiola, which is both over a century old and preserved in alcohol, several branches of the a. maxillaris common to other ruminants did not perfuse with latex. These include the a. auricularis profunda, the a. tympanica rostralis, the a. masseterica, and the a. palatina descendens and a. sphenopalatina. As with the a. auricularis caudalis, the minute pinna of the tragulid auricle renders these arteries less likely to perfuse due to their small internal diameter. The a. tympanica is likewise expected to be small.

The first main branch of the a. maxillaris is the arteria alveolaris inferior. From the ventral surface of the a. maxillaris, the a. alveolaris inferior courses toward and enters the foramen mandibularis. The a. alveolaris inferior did not perfuse past the foramen mandibularis and into the canalis mandibulae.

Following the a. alveolaris inferior and the small arteries that supply the musculi pterygoidei, m. constrictors pharynges rostrales, and mucosae pharyngeus, the a. maxillaris gives off a large branch from its dorsal surface directly into the foramen orbitotorundum. Upon entering the braincase, this artery immediately anastomoses with the a. carotis interna (Figs. 2 and 4). Proximal to the orbital portion of the a. maxillaris arise the arteriae temporalis profunda caudalis and rostralis. The a. temporalis profunda caudalis courses superiorly, deep to the rostral edge of the processus coronoides. This artery is the primary contributor of blood to the m. temporalis. The a. temporalis profunda rostralis arises shortly after the division of the a. temporalis profunda caudalis, and courses caudal to the processus zygomaticus of the os frontale.
Figure 4  Simplified arterial tree of *Moschiola memmina*, illustrating carotid-maxillary anastomosis. (A) Right lateral view, illustrating the a. carotis interna ascending to the basicranium without branching, and a large ramus from the a. maxillaris entering the foramen orbitorotundum. (B) Right medial sagittal section. The a. carotis interna enters the braincase through the anterior foramen lacerum, and courses rostrally to anastomose with the a. maxillaris. Abbreviations—CCA, arteria carotis commumis; eICA, arteria carotis interna pars extracranialis; for, foramen orbitorotundum; iICA, arteria carotis interna pars intracranialis; MA, arteria maxillaris; OE, arteria ophthalmica externa.

In this specimen, radiopaque latex perfused only three major vessels extending from the rostral extent of the a. maxillaris: the a. ophthalmica externa, the a. infraorbitalis, and the a. buccalis. The a. ophthalmica externa is the dorsal continuation of the a. maxillaris in the orbit (Fig. 2). This artery traverses the periorbita, supplying the m. levator anguli oculi medialis as it travels toward the os ethmoidale. The injection medium did not perfuse the rostral termination of the artery. The a. buccalis is a major lateral branch of the a. maxillaris. The a. buccalis supplies the superficial face by
crossing the tuber maxillae. Once it becomes superficial, the a. buccalis courses rostrally, paralleling the crista facialis (Fig. 2). Throughout its course, the a. buccalis supplies the ventral extraorbital region, the musculi pterygoidei, the m. buccinator, and the deep surface of the m. masseter. It does not, however, give rise to the a. temporalis profunda rostralis, as in many ruminants. The a. infraorbitalis is the rostral continuation of the a. maxillaris (Fig. 2). After coursing underneath the eye and periortibita, the a. infraorbitalis enters the canalis infraorbitalis. The maxillary dentition is supplied by the rami dentales before the artery exits the canal through the foramen infraorbitalis.

**Blood supply to the eye and orbit**
The a. ophthalmica interna contributes blood supply to the globe of the eye. Originating on the rostral face of the circulus arteriosus cerebri, the a. ophthalmica interna exits the braincase through the midline canalis opticus and is transmitted to the eye via the nervus opticus. The a. ophthalmica externa, as described above, perfuses the medial and inferior periortibita, inclusive of the extraocular muscles (Fig. 2). Smaller branches supplying the eye and orbit, such as the a. centralis retinae, the a. lacrimalis, and the ae. ciliares, were not perfused.

**Arterial blood supply to the brain**
The a. carotis communis bifurcates into the ECA and ICA ~1 cm ventral to the hyoid. The ICA ascends toward the basicranium without branching (Figs. 2 and 4). Before entering the braincase through the foramen lacerum, this vessel scours the medial surface of the bulla tympanica, leaving a noticeable groove. This groove is not present in extant artiodactyls with a rete mirabile epidurale. Once inside the braincase, the ICA courses rostrally along the internal surface of the corpus of the os basisphenoidale, again leaving deep grooves (Fig. 3). The circulus arteriosus cerebri (CAC) forms from the anastomosis of the ICA and the a. maxillaris (Figs. 3 and 4). The arteria cerebri rostralis forms the rostrolateral quadrant of the CAC, and continues rostrally in the median plane. Coursing along the nervus olfactorius, the a. cerebri rostralis terminates as rami corticales to the fissure longitudinalis of the cortex cerebri and bulbus olfactorius. The pars rostralis of the cortex cerebri, the nervus olfactorius, and the bulbus olfactorius are supplied by the a. cerebri rostralis. The arteria cerebri media is the lateral continuation of the ICA via the CAC. This artery courses dorsally through the sulcus rhinalis lateralis, between the lobus piriformis and gyri insulae and sylvii. From the caudolateral surface of the CAC (arteria communicans caudalis), three arteries emerge: the a. cerebri caudalis, the rami choroidei caudalis, and the a. cerebrelli rostralis (in order from rostral to caudal). The arteria cerebri caudalis courses deep to the lobus piriformis to supply caudal portions of the cerebrum. The a. cerebri caudalis also courses deep to the lobus piriformis, closer to the cerebellum. Ultimately, this vessel supplies the tectum mesencephali. The arteria cerebrelli rostralis branches from the CAC near the pars petrosal of the os temporale, and courses dorsolaterally along the pars rostralis of the cerebellum. The caudal-most branch of the circulus arteriosus cerebri is the arteria basilaris, which forms at the confluence of the arteriae communicans caudalis. The a. basilaris is situated on the midline, along the ventral surface of the metencephalon and myelencephalon, tapering into the a. spinalis ventralis.
after giving off the a. cerebelli caudalis. The former arteries are the main sources of blood to the cerebellum.

**Hard tissue results: osteological correlates for ruminant cranial arteries**

Five hard tissue features were identified that correlate the presence or absence of an a. carotis interna or rete mirabile epidurale. These features correlate directly with the presence of an arterial feature. In most cases, the presence of a correlate for one arterial state is indicative of the absence of that arterial state in the other. This is because artiodactyls effectively replace a patent a. carotis interna with the rete mirabile epidurale. Osteological correlates were sought both internally and externally. The presence and absence of these correlates is summarized in Table 1.

**Carotid groove, os basisphenoidale**

Inside the braincase, the presence of an a. carotis interna leaves a pronounced sulcus caroticus on the internal surface of the os basisphenoidale in *Tragulus, Hyemoschus, Moschiola,* and *Equus.* This groove deepens throughout development, as the cranial base develops and ossifies in tandem with the development of the cranial arteries (*Padget, 1948; O’Brien, 2012*). In artiodactyls with a rete mirabile epidurale, such as the domestic goat (*Capra hircus*), this groove is absent.

**Medial bullar groove**

Note: This correlate is only applicable to ruminants. In *Moschiola,* the a. carotis interna leaves a groove on the medial wall of the bulla tympanica as it ascends toward the basicranium. This groove was not identified in the neonate or early juvenile specimens, and can be slight in sub-adults. The groove is most clearly identifiable where it contacts the distal-most extent of the extracranial a. carotis interna. This groove is not identified in *Equus,* as the a. carotis interna and foramen lacerum are positioned well rostral of the bulla.

**Foramen orbitoretundum**

Typically, euungulates that do not possess a rete mirabile epidurale do not join the foramen rotundum with the fissura orbitalis into a “foramen orbitoretundum,” and thus have one more alisphenoid foramen (*Sisson & Grossman, 1967*). This is exemplified in *Equus.* In ruminants with a rete mirabile epidurale, the rami connecting the a. maxillaris to the rete mirabile epidurale enter the skull through the large, confluent foramen orbitoretundum. Although all ruminants examined possess a foramen orbitoretundum, the morphology of this foramen differed based on presence or absence of an a. carotis interna in tragulids. In *Moschiola,* the foramen orbitoretundum has a pronounced notch on the ventromedial border, due to contact with the a. carotis interna. This notch was found on skulls of all contemporary tragulids. In *Odocoileus* and *Capra,* the foramen orbitoretundum is smooth and accommodates the network of arteries that supply the rete mirabile epidurale.

**Foramen lacerum**

In *Equus* and *Moschiola,* the a. carotis interna enters the braincase through the anterior foramen lacerum. In both cases, the foramen lacerum possess notches that correspond with the a. carotis interna.
**Foramen ovale**

In ruminants with a rete mirabile epidurale, the foramen ovale frequently has a subtle, oblique notch that corresponds to a ramus or rami that supply the rete mirabile epidurale. This notch yields a foramen ovale that is trapezoidal in shape. In the tragulids examined, the foramen ovale lacked such a notch.

**DISCUSSION**

Among artiodactyls with described cranial arterial patterns, only the tragulids *T. javanicus* and *T. napu* were previously found to possess a substantial arteria carotis interna. This study documents another example of an artiodactyl without a rete mirabile epidurale, with osteological data suggesting that this condition is common to all tragulids. The use of 3-D anatomical data collection methods, combined with digital dissection, have revealed an extension of previous findings (*Fukuta et al., 2007*) that may have developmental implications for artiodactyls with and without a rete mirabile epidurale. Developmental studies of prenatal specimens indicate that the a. carotis interna of rete-bearing artiodactyls obliterates during development, such that the brain is not supplied by a major branch of the 3rd aortic arch (*Tandler, 1899; Tandler, 1901; Tandler, 1906; Daniel, Dawes & Prichard, 1953; Baldwin, 1964; Gillilan, 1974; Bamel, Dhingra & Sharma, 1975; Wible, 1984*). In lieu of a patent a. carotis interna, the brain is largely supplied through the rete mirabile epidurale by the a. maxillaris rostrally (through the foramen orbitosphenoidale) and the a. pharyngea ascendens caudally (through the foramen ovale, foramen lacerum, or canalis caroticus; see e.g., *Daniel, Dawes & Prichard, 1953; Sisson & Grossman, 1967; Smuts & Bezuidenhout, 1987*). In several artiodactyls, particularly suids, the a. pharyngea ascendens (or, more appropriately the pharyngocarotid anastomosis *sensu Wible, 1984*) is frequently identified as the a. carotis interna, however embryological studies indicate that the adult vessel is not homologous (*Daniel, Dawes & Prichard, 1953; Wible, 1984*). In tragulids, the persistent a. carotis interna is likely homologous to that of other vertebrates (*Fukuta et al., 2003*), and the a. pharyngea ascendens does not hypertrophy.

Overall, tragulids exhibit a cranial arterial pattern that is similar to other mammals, with key exceptions that are relevant to their status as the sister group of ruminants. First, the a. basilaris of non-ruminant mammals is formed at its caudal extent by the merger of contralateral branches of the occipital artery (see e.g., *Hofmann, 1900; Gillilan, 1974; Sisson & Grossman, 1967*). This results in a caudal to rostral direction of blood flow through the a. basilaris, such that its parent artery, the a. occipitalis, provides collateral circulation to the CAC (*Daniel, Dawes & Prichard, 1953; Gillilan, 1974*). In ruminants, now inclusive of tragulids, the a. basilaris does not significantly anastomose with the a. occipitalis, instead diminishing in caliber as it passes caudally, away from the CAC (*Daniel, Dawes & Prichard, 1953; Gillilan, 1974; *Fukuta et al., 2007*; this study). Consequently, blood flows from rostral to caudal through the a. basilaris of ruminants. This morphology has important functional implications in ruminants with a rete mirabile epidurale: the vast majority of blood distributed through the CAC must first pass through
the retial heat exchanger, ensuring that nearly all blood bound for the cerebrum and cerebellum is cooled when selective brain cooling is active (Gillilan, 1974).

The second important distinction that can be identified in the arterial pattern of Moschiola meminna is the substantial intracranial connection between the a. maxillaris and a. carotis interna. In the typical mammalian condition, the a. maxillaris does not enter the cranial cavity. In Moschiola, however, the a. maxillaris gives rise to a ramus that enters the braincase via the foramen orbitorotundum, where it directly anastomoses with the a. carotis interna as it scours the internal surface of the sphenoid (Fig. 4). This anastomosis is typical of non-suiform artiodactyls, which supply the carotid rete through rami from the a. maxillaris (see e.g., Daniel, Dawes & Prichard, 1953; Gillilan, 1974; Sisson & Grossman, 1967). In tragulids, this connection persists throughout the animal’s life; however, in other artiodactyls for which cranial arterial patterns are documented, a substantial a. carotis interna obliterates early in development (Wible, 1984). This intracranial carotico-maxillary anastomosis may be a vital prerequisite for the development and evolution of the rete mirabile epidurale and the efficient selective brain cooling that is facilitated by this vascular feature. Without another substantial influx of oxygenated blood to the brain, the developmental loss of a functional pars extracranialis of the ICA could be deleterious. Thus, it is this aberrant anastomosis that may have enabled the evolution of the artiodactyl rete mirabile epidurale.

In evolutionary terms, the complete, patent a. carotis interna of tragulids has been hypothesized as either a plesiomorphy for ruminants, or an apomorphic feature of tragulids that has arisen due to adaptation (Fukuta et al., 2007). If the tragulid condition is plesiomorphic for ruminants, it suggests that the rete mirabile epidurale may have evolved several times among artiodactyls. If the lack of a rete is an apomorphy for the Tragulidae, then the driving selective or developmental pressures should be investigated further. By surveying the cranial arterial pattern of all extant tragulids, either through soft tissue dissection or identification of hard tissue correlates, the presence of an a. carotis interna can be confirmed throughout the family. Although this finding does not provide direct evidence for either evolutionary hypothesis, it is important to note that extant mouse deer have experienced a diverse suite of selective pressures across their extensive evolutionary history. Not the least of these is habitat, with respect to temperature and hydrological budget. For example, H. aquaticus, the water chevrotain, is semi-aquatic and inhabits the dense, equatorial forests of Africa (Dubost, 1978). The Asian taxa, Tragulus and Moschiola, are found in more hydrologically variable environments, with some species preferring more open, drier habitats and others preferring dense vegetation and more moist habitats (Pickford, Senut & Mourer-Chauviré, 2004; Groves & Meijaard, 2005). From a temporal perspective, divergence between the genera Tragulus and Hyemoschus likely occurred in the early Miocene (Hassanin et al., 2013), as the ancestor of Hyemoschus entered the African landmass pursuant to the formation of connections between Europe and Africa (Pickford, 2001; Sanchez et al., 2010). The geographic, ecological, and temporal segregation between living tragulids should result in different suites of selective pressures. If the lack of a rete mirabile epidurale is an apomorphy among tragulids, this condition was almost certainly
established prior to the separation of modern groups. Overall, both this study and that of Fukuta and colleagues (2007) emphasize the need for additional developmental and evolutionary studies into the origins of ruminant cranial vascular patterns.

ACKNOWLEDGEMENTS
RDE MacPhee, E Westwig, N Duncan, and E Hoeger at the American Museum of Natural History for collection and specimen access; K Wheeler skillfully assisted in specimen preparation and injection; C Dennis, B Keener, and J Sands at the Holzer Clinic in Athens, Ohio, for CT scanning.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding
This work was funded by the American Society of Mammalogists Grants-in-Aid, the American Museum of Natural History Collection Study Grant, and the Ohio University Student Enhancement Award. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures
The following grant information was disclosed by the author:
American Society of Mammalogists Grants-in-Aid.
American Museum of Natural History Collection Study Grant.
Ohio University Student Enhancement Award.

Competing Interests
The author declares there are no competing interests.

Author Contributions
• Haley D. O’Brien conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.

Animal Ethics
The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):
This work was conducted using a cadaveric museum specimen, and required no institutional review.

Data Availability
The following information was supplied regarding data availability:
FigShare:
http://figshare.com/s/e22fde06688711e5babe06ec4bcbf141
http://figshare.com/s/f1f3cb9a688711e5ab6506ec4bcbf141.
REFERENCES

Altman PL, Dittmer DS. 1968. Biological handbooks: metabolism. Bethesda: Federation of American Societies for Experimental Biology.

Baldwin BA. 1964. The anatomy of arterial supply to the cranial regions of the sheep and ox. American Journal of Anatomy 115:108–118 DOI 10.1002/aja.1001150107.

Bamel SS, Dhingra LD, Sharma DN. 1975. Anatomical studies on the arteries of the brain of the buffalo (Bubalus bubalis). Anatomischer Anzeiger 137:440–446.

Brunner S. 1998. Skull development and growth in the southern fur seals Arctocephalus forsteri and A. pusillus doriferus (Carnivora: Otariidae). Australian Journal of Zoology 46:43–66 DOI 10.1071/ZO97019.

Caputa M. 2004. Selective brain cooling: a multiple regulatory mechanism. Journal of Thermal Biology 29:691–702 DOI 10.1016/j.jtherbio.2004.08.079.

Carlsson A. 1926. Ueber die Tragulidae und ihre Beziehungen zu ubringen Artiodactyla. Acta Zoologica Stockholm 7:69–100 DOI 10.1111/j.1463-6395.1926.tb00924.x.

Daniel PM, Dawes JDK, Prichard MML. 1953. Studies of the carotid rete and its associated arteries. Philosophical Transactions of the Royal Society of London B 237:173–204 DOI 10.1098/rstb.1953.0003.

Dubost G. 1975. Behavior of the African chevrotain (Hyemoschus aquaticus). Its ecological and phylogenetic significance. Zeitschrift für Tierpsychologie 37:449–501 DOI 10.1111/j.1439-0310.1975.tb00890.x.

Dubost G. 1978. Un aperçu sur l’écologie du Chevrotain africain Hyemoschus aquaticus Olgiby, Artiodactyle Tragulidé. Mammalia 42:1–61 DOI 10.1515/mamm.1978.42.1.1.

Fukuta K, Kudo H, Sasaki M, Kimura J, Bin Ismail D, Endo H. 2007. Absence of carotid rete mirabile in small tropical ruminants: implications for the evolution of the arterial system in artiodactyls. Journal of Anatomy 210:112–116 DOI 10.1111/j.1469-7580.2006.00667.x.

Fukuta K, Orui T, Sasaki M, Kimura J, Endo H, Bin ID, Kudo H. 2003. Arterial branches from aortic arch and blood supply to the brain in the lesser and greater mouse deer. Exp Herbivora 27:35–44.

Gillilan LA. 1974. Blood supply to the brains of ungulates with and without a rete mirabile caroticum. Journal of Comparative Neurology 153:275–290 DOI 10.1002/cne.901530305.

Groves C, Meijaard E. 2005. Intraspecific variation in Mosciola, the Indian Chevrotain. The Raffles Bulletin of Zoology 12:413–421.

Grubb P. 1993. Order Artiodactyla. In: Wilson DE, Reeder DAM, eds. Mammal species of the world: a taxonomic and geographic reference. 2nd edition. Washington, D.C.: Smithsonian Institution Press, 377–414.

Hassanin A, Delsuc F, Ropiquet A, Hammer C, Van Vuuren BJ, Matthee C, Ruiz-Garcia M, Catzeflis F, Areskoug V, Nguyen TT, Couloux A. 2013. Pattern and timing of diversification of Cetartiodactyla (Mammalia, Laurasiatheria), as revealed by a comprehensive analysis of mitochondrial genomes. Comptes Rendus Biologies 335:32–50 DOI 10.1016/j.crvi.2011.11.002.

Holliday CM, Ridgely RC, Balanoff AM, Wittmer LM. 2006. Cephalic vascular anatomy in flamingos (Phoenicopterus ruber) based on novel vascular injection and computed tomographic imaging analysis. The Anatomical Record Part A 288A:1031–1041 DOI 10.1002/ar.a.20374.

Hofmann M. 1900. Zur vergleichenden Anatomie der Gehirn- und Rückenmarksarterien der Vertebraten. Zeitschrift für Morphologie und Anthropologie 2:246–322.
Janis C. 1984. Tragulids as living fossils. In: Eldredge N, Stanley SM, eds. *Living fossils*. New York: Springer-Verlag, 87–94.

Jessen C. 1998. Brain cooling: an economy mode of temperature regulation in artiodactyls. *News in Physiological Sciences* 13:281–286.

Jessen C, Pongratz H. 1979. Air humidity and carotid rete function in thermoregulation of the goat. *Journal of Physiology* 292:469–479 DOI 10.1113/jphysiol.1979.sp012865.

Kuhnen G. 1997. Selective brain cooling reduces respiratory water loss during heat stress. *Comparative Biochemistry and Physiology Part A* 118:891–895 DOI 10.1016/S0300-9629(97)00235-1.

Langer P. 1974. Stomach evolution in the Artiodactyla. *Mammalia* 38:295–314 DOI 10.1515/mamm.1974.38.2.295.

Maddison WP, Maddison DR. 2011. *Mesquite: a modular system for evolutionary analysis*. Version 2.75. Available at http://mesquiteproject.org.

McNab BK. 1983. Food habits, energetics, and the population biology of mammals. *Journal of Zoology, London* 199:1–29 DOI 10.1111/j.1469-7998.1983.tb06114.x.

McNab BK. 1990. The physiological significance of body size. In: Damuth J, MacFadden BJ, eds. *Body size in mammalian paleobiology: estimation and biological implications*. New York: Cambridge University Press, 11–23.

Meindl RS, Lovejoy CO. 1985. Ectocranial suture closure: a revised method for the determination of skeletal age at death based on lateral-anterior sutures. *American Journal of Physical Anthropology* 68:57–66 DOI 10.1002/ajpa.1330680106.

Métails G, Vislobokova I. 2007. Basal ruminants. In: Prothero DR, Foss SE, eds. *The evolution of artiodactyla*. Baltimore: Johns Hopkins University Press, 189–212.

Milne-Edwards A. 1864. *Recherches anatomiques zoologiques, et paleontologiques, sur la famille des Chevrotains, Annales des Sciences Naturelles 5me Serie*. Paris: Tome II.

Mitchell J. 1973. Determination of relative age in the dugong *Dugong dugon* (Müller) from a study of skulls and teeth. *Zoological Journal of the Linnean Society* 53:1–23 DOI 10.1111/j.1096-3642.1973.tb01409.x.

Mitchell D, Maloney SK, Jessen C, Laburn HP, Kamerman PR, Mitchell G, Fuller A. 2002. Adaptive heterothermy and selective brain cooling in arid-zone mammals. *Comparative Biochemistry and Physiology Part B* 131:571–585 DOI 10.1016/S1096-4959(02)00012-X.

Morris P. 1972. A review of mammalian age determination methods. *Mammal Review* 2:69–104 DOI 10.1111/j.1365-2907.1972.tb00160.x.

O’Brien HD. 2012. Ontogeny and phylogeny of cranial vascular patterns in the Tragulidae (Artiodactyla: Ruminantia). *Integrative and Comparative Biology* 52:e304 DOI 10.1093/icb/icb079.

O’Brien HD, Williams SH. 2014. Using biplanar fluoroscopy to guide radiopaque vascular injections: a new method for vascular imaging. *PLoS ONE* 9:e97940 DOI 10.1371/journal.pone.0097940.

Ostrowski S, Williams JB, Ismael K. 2003. Heterothermy and the water economy of free-living Arabian oryx (*Oryx leucoryx*). *The Journal of Experimental Biology* 206:1471–1478 DOI 10.1242/jeb.00275.

Padget DH. 1948. The development of the cranial arteries in the human embryo. *Contributions to Embryology* 32:205–262.
Pickford M. 2001. Africa's smallest ruminant: a new tragulid from the Miocene of Kenya and the biostratigraphy of East African Tragulidae. *Geobios* 34:437–447 DOI 10.1016/S0016-6995(01)80007-3.

Pickford M, Senut B, Mourer-Chauviré C. 2004. Early Pliocene Tragulidae and peafowls in the Rift Valley, Kenya: evidence for rainforest in East Africa. *Comptes Rendus Palevol* 3:179–189 DOI 10.1016/j.crpv.2004.01.004.

Powers R. 1962. The disparity between known age and age as estimated by cranial suture closure. *Man* 62:52–54 DOI 10.2307/2796987.

Rössner GE. 2007. Family Tragulidae. In: Prothero DR, Foss SE, eds. *The evolution of artiodactyls*. Baltimore: Johns Hopkins University Press, 213–220.

Sanchez IM, Quiralte V, Morales J, Pickford M. 2010. A new genus of tragulid ruminant from the Early Miocene of Kenya. *Acta Palaeontologica Polonica* 55:177–187 DOI 10.4202/app.2009.0087.

Sanchez-Villagra MR. 2010. Suture closure as a paradigm to study late growth in recent and fossil mammals: a case study with giant deer and dwarf deer skulls. *Journal of Vertebrate Paleontology* 30:1895–1898 DOI 10.1080/02724634.2010.521218.

Sisson S, Grossman JD. 1967. *The anatomy of the domestic animals*. 4th edition. Philadelphia, London: WB Saunders.

Smuts MMS, Bezuidenhout AJ. 1987. *Anatomy of the dromedary*. Oxford: Clarendon Press.

Spaulding M, O'Leary MA, Gatesy J. 2009. Relationships of Cetacea (Artiodactyla) among mammals: increased taxon sampling alters interpretations of key fossils and character evolution. *PLoS ONE* 9:e7062 DOI 10.1371/journal.pone.0007062.

Tandler J. 1899. Zur verleichenden Anatomie der Kopfarterien bei den Mammalia. *Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Klasse* 67:677–784.

Tandler J. 1901. Zur vergleichenden Anatomie der Kopfarterien bei den Mammalia. *Anatomische Hefte* 18:327–373 DOI 10.1007/BF02246378.

Tandler J. 1906. Zur Entwicklungsgeschichte der arterilien Wundernetze. *Anatomische Hefte I Abteilung* 31:236–267.

Wible JR. 1984. The ontogeny and phylogeny of the mammalian cranial arterial pattern. PhD Dissertation, Duke University.

Witmer LM. 1995. The extant phylogenetic bracket and the importance of reconstructing soft tissues in fossils. In: Thomason JJ, ed. *Functional morphology in vertebrate paleontology*. New York: Cambridge University Press, 19–33.