Introduction

Disruption to the triceps brachii tendon is a rarely reported tendon injury in companion animals, with most triceps tendon disruptions secondary to avulsion. Treatment outcomes have been limited to only a few descriptive studies in dogs. This report describes successful treatment of a chronic complete laceration of the triceps brachii tendon in a 38 kg entire male German Shepherd. Surgical management included staged debridement, with delayed primary repair supported by an autogenous thoracolumbar fascia (TLF) onlay graft. To the best of the authors’ knowledge, successful surgical repair of tendinous injury incorporating the use of an autogenous thoracolumbar fascia onlay graft has not previously been described.

Case History

A 5-year-old 38 kg entire male German Shepherd dog was referred for persistent non-weight bearing left thoracic limb lameness 5 weeks after failed triceps tendon repair. The inciting trauma to the triceps tendon occurred when the dog was attacked by a member of the public with a machete. Tendon transection was reported to have occurred 1 cm proximal to the olecranon. Primary repair was attempted by the referring veterinarian 48 hours following this injury and included anastomosis of the triceps tendon using 0-PDS in a combination of simple continuous and interrupted cruciate sutures following wound flushing. Postoperative management included a modified Robert-Jones bandage and exercise...
Continued treatment with meloxicam (Boehringer Ingelheim Pty Ltd, North Ryde, Australia) (0.1 mg/kg per os [PO] every 24 hours) and strict confinement at home failed to improve a non-weight bearing left thoracic limb lameness.

The patient was then referred for further treatment 5 weeks after the initial trauma with no weight bearing reported since the inciting trauma. At that time the patient had been treated with 3 weeks of amoxycillin–clavulanic acid (Apex Laboratories Pty Ltd, Somersby, Australia) (13 mg/kg PO every 12 hours) for a persistent draining tract proximal to the olecranon which was not resolving.

Upon presentation to the referral surgeon (R.G.C.), the patient was found to be non-weight bearing lame in the left forelimb with a purulent draining tract located 2 cm proximal to the olecranon on the caudolateral aspect of the distal brachium. Physical exam revealed a large palpable defect proximal to the olecranon. When the shoulder was fixed in extension, the elbow could be fully flexed, confirming complete incompetence of the triceps mechanism. Cutaneous innervation of the left forepaw remained intact circumferentially, indicating an intact nervous supply to the paw.

At this point, treatment was planned and divided into two stages.

1. Surgical exploration of the traumatized area with resection of sinus tract and interposed fibro-granulomatous tissue, lavage and exit culture in preparation for definitive reconstruction of the triceps brachii tendon.
2. Reconstruction and repair of the triceps brachii tendon once a healthy wound environment was achieved.

The previous surgical scar was excised and associated draining tract explored and debrided. The tract extended to the caudal distal humerus, just over the proximal olecranon. A large amount of proliferative granulation tissue and fibrous scar tissue proximal to the olecranon communicated with a draining tract to the skin. Aggressive en bloc resection of this reparative tissue and sinus was performed. Deep to this tissue was healthy fibrotic scar tissue interposed between the triceps brachii long head muscle belly and the olecranon. Following wound debridement and lavage, an exit culture was taken and submitted for routine culture and sensitivity. The subcutaneous layers and skin were closed routinely. The patient was discharged with instructions to administer meloxicam (Boehringer Ingelheim Pty Ltd, North Ryde, Australia) (0.1 mg/kg PO every 24 hours), codeine (Aspen Pharma Pty Ltd, St. Leonards, Australia) (0.8 mg/kg PO every 12 hours) and amoxycillin–clavulanic acid (Apex Laboratories Pty Ltd, Somersby, Australia) (26 mg/kg PO every 12 hours) pending culture and sensitivity results. Strict exercise restriction was also instructed between surgical exploration and definitive repair.

The results of the final microbiology report showed occasional epithelial cells, with no bacteria seen on Gram/ZN stain and no growth obtained by day two of culture.

Definitive triceps brachii tendon repair was performed weeks after initial injury, 1-week post sinus and partial scar excision. A caudolateral approach to the left mid-distal brachium was performed. Fibrous adhesions between the lateral and long heads of the triceps brachii muscle were broken down. The accessory and medial heads were found encased in fibrous scar tissue along with the anconeus muscle. No attempt was made to dissect out these bellies. The radial nerve was identified and protected during dissection. The dysfunctional tendon remnants and the scar tissue (Fig. 1) that was interposed between the muscle bellies of the lateral and long heads of the triceps were resected until healthy muscle was identified (Fig. 2). Fibrous tissue present at the

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**Fig. 1** Intraoperative photo of excessive scar tissue between the triceps brachii muscle and olecranon.

**Fig. 2** Intraoperative photo following removal of all interposed fibro-adipose scar tissue.
tendon insertion on the olecranon was also debrided, leaving ~5 mm of tendon at the olecranon insertion point. With the elbow placed in full extension and the shoulder partly flexed, the long head muscle belly was attached to the stub of tendon on the olecranon with 0-Nylon using several locking loop sutures and three-loop pulley sutures. Due to marked tension of the anastomosis, the risk of postoperative suture pull-out and gap formation was considered very high. For this reason, a TLF graft was then harvested as previously described (Fig. 3) and placed over the repair as an onlay graft to support the anastomosis as a tension relieving graft.11 The TLF graft was oriented such that when harvested, its original cranial border was now positioned proximally over the triceps lateral head and its caudal border was now positioned over the triceps lateral head. The graft was secured with 0 PDS suture in a Krakow suture pattern running the length of the graft with its dorsal and ventral margins, now oriented cranially and caudally on the olecranon respectively (Fig. 4). The lateral head of the triceps was then secured to the craniodistal border of the long head muscle and the onlay graft using 2-Nylon with a combination of locking loop and three loop pulley sutures. The site was lavaged thoroughly with sterile saline. The subcutaneous tissue was closed with 2–0 Glyconate with skin edges apposed with 3–0 Nylon in a Ford interlocking pattern.

The limb was bandaged, and a spica splint was fashioned from Dynacast Prelude Splinting Material; Roll 10 × 4.6m (BSN medical ANZ, Mulgrave, Australia) and applied postoperatively to protect and immobilize the left thoracic limb with the elbow maintained in an extended position for 6 weeks. A constant rate infusion (CRI) of fentanyl (Hameln Pharmaceuticals GmbH, Hameln Germany) (4 µg/kg/hour) was started intraoperatively and maintained at that rate throughout the procedure and overnight. Postoperatively, a CRI of medetomidine (Jurox Pty Ltd, Rutherford, Australia) (2 µg/kg/hour) was started and maintained at this rate until the patient was discharged from hospital 24 hours after surgery. The patient was discharged with instructions to administer meloxicam (Boehringer Ingelheim Pty Ltd, North Ryde, Australia) (0.1 mg/kg PO every 24 hours) and codeine (Aspen Pharma Pty Ltd, St. Leonards, Australia) (0.8 mg/kg PO every 12 hours) for 7 days, amoxicillin–clavulanic acid (Apex Laboratories Pty Ltd, Somersby, Australia) (26 mg/kg PO every 12 hours) for 14 days following surgery, as well as strict confinement to a small area, exercise restriction to controlled leash walks for elimination purposes only, and bandage care. Four weeks’ worth of acepromazine (Ceva Animal Health Pty Ltd, Glenorie, Australia) (0.6 mg/kg PO every 12 hours) was also dispensed.

Follow-up examinations were performed every 2 weeks until spica splint removal at 6 weeks, 1-week post spica splint removal at 7 weeks and then at 7 months after surgery. At 6 weeks following definitive surgical repair, all external coaptation was removed. The reconstructed triceps tendon was palpated as thick and robust with inability to flex the elbow while maintaining shoulder extension. Owners were instructed to maintain the patient on leash walking for a further 3 weeks, increasing frequency and duration of walks prior to resuming unrestricted off leash activity 9 weeks following definitive surgery. Re-evaluation at 4 and 7 months post definitive repair revealed the patient to be free of lameness with normal stance and elbow range of motion (Video 1).

**Video 1**

Owner’s check in video 3 months post definitive repair of chronic left triceps tendon laceration. Online content including video sequences viewable at:https://www.thieme-connect.com/products/ejournals/html/10.1055/s-0041-1725056.

**Discussion**

In order for a myotendinous apparatus to function effectively, achieving and maintaining appropriate tendon length and strength is of critical importance.12–14 In most cases, surgical

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Fig. 3 Intraoperative photo of the harvesting of the thoracolumbar fascia (TLF) graft. The proposed margins were cranially at the level of the paralumbar fossa/caudal aspect of rib 13; caudally, just cranial to the tuber coxa in alignment with the aponeurosis of the sartorius muscle; dorsally, a parasagittal incision adjacent to the dorsal processes of the lumbar vertebrae and ventrally, dorsal to the aponeurosis of the TLF with the external and internal abdominal oblique muscles.

Fig. 4 The thoracolumbar fascia graft was secured with its dorsal and ventral margins, now oriented cranially (left) and caudally (right) on the humerus respectively.
treatment is performed shortly after a tendon injury. Repair is considered delayed when the disrupted tendon has been neglected for at least 4 weeks. Untreated, the tendinous ends may atrophy and retract from their insertion, leaving inadequate tissue for anatomical reconstruction and a wide gap occupied by excessive fibro-adipose scar tissue with poor suture holding strength, which is needed to be excised before repairing the tendon. In cases where a tendon gap persists or for failed tendon repairs, autogenous, xenogenic and synthetic grafts have been utilized to provide additional support.

In the case reported herein, failure of the primary repair at index surgery to resist tension resulted in a large gap formation with exuberant interposed ‘bridging scar’ tissue formation. Debridement of interposed scar tissue resulted in a 10 cm gap between the triceps long and lateral heads and the olecranon. Definitive surgery to re-attach these muscle bellies to the olecranon resulted in marked anatomosis tension which was protected with an autogenous TLF onlay graft to prevent repeat gap formation occurring.

Onlay augmentation (reinforcement) grafts in human rotator cuff repair models have been shown to increase stiffness and, as a result of load sharing, decrease tendon gapping. Studies have reported conflicting evidence regarding the effect on load to failure. Shea and colleagues report that application of an onlay augmentation was estimated to share 34.7% of the global load applied to the tendon repair during cyclic subfailure loading, transferring some of that load directly to the bone and bypassing the repair. Clinically, demonstration of superior stiffness is a more relevant evaluation of the day-to-day response of the repair construct to subdestructive loads that are frequently seen with the rehabilitation process. Strength, in terms of load to failure, represents the ability of the repair to resist a one off destructive force, which is likely less relevant if the repair is further protected by limb immobilization. Rather in humans, repetitive, low-level loading of the rotator cuff repair with routine muscle contraction, even in a postoperative sling, is thought to be the likely mechanism of gap formation. Use of a polyurethane urea onlay mesh graft has successfully been used to augment the surgical repair of a chronic tendon rupture in the canine. Application of the TLF graft in an onlay (reinforcement) fashion in the dog reported herein is thought to have helped prevent gap formation at the repair site and resist displacement at these subdestructive loads.

In this patient, the decision to reinforce the repair with an onlay free graft of autologous TLF graft was elected due to the increased graft size when compared with tensor fascia lata. Furthermore TLF grafts demonstrate similar tensile strength and stiffness in material properties to that of fascia lata. Additionally, the chronic nature of the discharging wound which communicated with the disrupted tendon was concerning for active infection. For this reason, staged treatment with debridement/lavage and exit culture prior to definitive repair was performed. Active bacterial infection is known to be detrimental to tendon healing. Despite the negative exit culture, an autogenous graft was still favoured over a synthetic graft for risk of ongoing infection. Henderson and colleagues demonstrated that TLF, like fascia lata, acts in an anisotropic manner during tensile testing, with greater strength and stiffness in the dorsoventral direction and proximodistal direction respectively. Due to the inherent shape and useable surface area of TLF, the graft was placed over the repair with its original cranial–caudal orientation now oriented proximal to distally bridging the tendon repair. The authors believe that despite the orientation being perpendicular to the graft’s strongest orientation, physiologic loads likely did not exceed its failure point as suggested by the successful outcome. Further biomechanical testing would be required to assess the TLF as an onlay graft in terms of its applied orientation and useable length. Furthermore, the discrepancy in useable length of the graft when oriented in the cranio-caudal versus dorsoventral planes is significant, which influences how much tissue is covered and incorporated in the onlay repair which ultimately may influence construct strength.

In a triceps tenotomy model in the dog, only 56% of original strength was achieved at 6 weeks following traditional suture repair, justifying the need for postoperative immobilization to protect the repair from excessive, destructive, weight-bearing forces. Early and colleagues reported that transarticular external skeletal fixation and spica splints to be the most commonly applied measure of elbow immobilization following triceps tendon repair in dogs and cats respectively. A spica splint was used successfully for support of the primary tendon reconstruction in this patient, immobilizing the limb in an anatomical position to facilitate early weight bearing and protect the repair by avoiding excessive tension at the suture site. Furthermore, the surgeon did not want transarticular external skeletal fixation pin tracts in the region of the non-vascularized graft for potential increased risk of infection. Other options that have been reported in the veterinary literature are custom-made orthotic braces, long lateral splints and Robert-Jones bandages, with the former two achieving good functional outcome.

Previous reports suggest that chronic disruption of triceps tendon would be associated with a guarded prognosis. However, the outcome reported with the technique described herein resulted in return to excellent function. Although definitive conclusions cannot be drawn from a single case report, the authors believe that consideration for tenorrhaphy augmentation with a free TLF onlay graft should be considered in particularly challenging cases.

Authors’ Contributions
R.G.C. performed the surgery and revised the current manuscript. C.J.W. wrote the manuscript and submitted to the journal.

Conflict of Interest
None declared.

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