REVIEW ARTICLE

Selecting postoperative adjuvant systemic therapy for early stage breast cancer: A critical assessment of commercially available gene expression assays

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Risk stratification of patients with early stage breast cancer may support adjuvant chemotherapy decision-making. This review details the development and validation of six multi-gene classifiers, each of which claims to provide useful prognostic and possibly predictive information for early stage breast cancer patients. A careful assessment is presented of each test's analytical validity, clinical validity, and clinical utility, as well as the quality of evidence supporting its use.

KEYWORDS
adjuvant, biomarkers, breast neoplasms, chemotherapy, clinical decision-making, gene expression, genomics risk, prognosis, tumor

1 INTRODUCTION

For 20 years, breast cancer mortality rates have steadily fallen.1 Although adjuvant chemotherapy (CTx) has been a strong contributor to the overall effect, only a small proportion of patients individually benefit from the treatments they receive.2 Commercially available gene expression assays are promoted as a window into intrinsic tumor biology allowing a more accurate determination of prognosis and treatment benefit than traditional clinicopathologic classifiers.

Haddow and Polamaki’s framework for the evidence-based evaluation of genomic assays identifies components essential for any new test.3,4 Analytic validity represents an assay’s ability to accurately and reproducibly measure biomarkers of interest. Clinical validity requires the assay to provide accurate and reliable information regarding outcomes of relevance.5 Clinical utility reflects an assay’s ability to favorably alter patient outcome.
The Tumor Marker Utility Grading System (TMUGS) grades evidence on a progressive scale. The best evidence comes from completely prospective, controlled clinical trials (Level A). A lower level of evidence (LOE) are prospective-retrospective studies (Level B) with a prospective study design of archived tissue from a previous prospective clinical trial. Two concordant Level B trials can achieve Level I evidence. Prospective evaluation of previously collected patient registry data (level C), represents a lower LOE, but provides opportunity to confirm findings from higher LOE studies. Post hoc investigations, using convenience samples, in the absence of defined treatment, data acquisition, or follow-up, are prone to bias and represent the lowest LOE (level D). These should only be used for general confirmation of higher-level data or for hypothesis generation. TMUGS is summarized in Table 1.

Currently there are six commercial gene expression assays available worldwide for patients with early stage breast cancer (ESBC). This review identifies the relevant supporting data for these assays graded by TMUGS LOE.

2 | MATERIALS AND METHODS

2.1 | Article sources

A systematic search of the scientific literature was conducted in September 2016 that identified primary published studies describing the clinical validation of one of the six commercially available genomic classifiers for breast cancer (Table 2.). Databases searched included Medline and the Cochrane Library. In addition, relevant previously published review articles were consulted to be certain that appropriate reports were not overlooked.

2.2 | Search strategy

A keyword search strategy was implemented for the initial search of databases that included “BCI,” “Breast Cancer Index,” “HOXB13/IL17BR,” “MGI,” “Molecular Grade Index,” “IHC4,” “Oncotype,” “21 gene,” “Recurrence Score,” “MammaPrint,” “70 gene,” “BluePrint,” “TargetPrint,” “PAM 50,” “intrinsic subtype,” “Endopredict,” and “Prosigna.”

2.3 | Study selection and data extraction

Only English-language publications were selected. Studies analyzed original data relevant to clinical validity or clinical utility and included a formal statistical test of pre-specified outcome endpoints. Epidemiological, exploratory, and purely descriptive studies were excluded. As this review focuses on decision-making in the immediate postoperative adjuvant setting, we have not included an evaluation of gene expression assays in the setting of neoadjuvant or extended adjuvant therapy, even when the LOE was high (e.g., Sgroi et al.7). Excluded also were publications not relating to female invasive breast cancer and studies without peer-reviewed publication. Final reports published in E-format were eligible. All eligible articles were fully reviewed by the investigators to determine that the study met the prespecified requirements. The PRISMA flow diagram for this review is presented in Fig. S1.

2.4 | Study analysis

A total of 34 articles were identified for inclusion in this analysis (Table S1). Data from each article selected was captured in a structured abstraction form. The framework from Simon and colleagues was used to evaluate the LOE of the identified studies. Each element of the
| Characteristic | Breast cancer index | Endopredict | IHC4 | MammaPrint | OncotypeDX breast cancer assay | Prosigna |
|---------------|---------------------|-------------|------|------------|-------------------------------|----------|
| Vendor        | bioTheranostic      | Myriad Genetics | Genoptix | Agendia BV | Genomic Health, Inc. | Nanostring, Inc. |
| Validated in  | ER+, LN−, ESBC treated with TAM | ER+ HER2− ESBC treated with TAM from ABCSG-6 (tam arm only) and ABCSG-8 | ER+ ESBC treated with TAM or anastrozole | Stage I, II; ≤5.0 cm LN−; ER+ or ER−; HER2−, negative or positive; < 53 years of age | ER+ Node + or −, ESBC treated with TAM | ER+, node negative or positive, ESBC in post-menopausal women |
| Genes included | HOX B13/IL17BR ratio, Molecular Grade Index (5 genes) (Jenkowitz BCR 2011) | 9 risk-related plus 3 normalization; Combined with clinical indicators for EPClin score (Filipits Clin Cancer Res 2011) | 4 breast cancer related (Cuzick J Clin Oncol 2011) | 70 (van de Vijver N Engl J Med 2002) | 5 proliferation-related, 2 invasion-related, 4 ER-related, 5 Other, 5 reference (Paik N Engl J Med 2004) | 50 PAM50-related, 8 normalization (Nielsen BMC Cancer 2014) |
| Analytical platform | Central by vendor | Central by vendor | Immunohistochemistry | Microarray, central lab | RT-PCR | nCounter system from Nanostring Technologies, Inc. |
| Sample requirements | FFPE block or slides | FFPE block or slides | FFPE slides or tissue microarrays | FFPE block or slides | FFPE block, slides, cores | FFPE blocks or slides |
| Assay readout | Two risk categories | Continuous, two risk categories | Continuous risk score | Two risk categories | continuous risk score, three risk categories | Three risk categories plus continuous risk score |
| Referencing guidelines | ASCO (ER+ LN− ESBC, for prognosis); St. Gallen (ESBC for prognosis), ESMO (ER+ primary breast cancer for prognosis and prediction), AGO (ER+ LN− ESBC for prediction) | ASCO (ER+ LN− ESBC, for prognosis); ESMO (ER+ primary breast cancer for prognosis and prediction), St. Gallen (ESBC for prognosis), NCCN (ESBC for prognosis) | None | ESMO (primary breast cancer for prognosis and prediction), St. Gallen (ER+ ESBC for prognosis and prediction), NCCN (ER+ ESBC for prognosis and prediction), ASCO (ER+ LN− ESBC, for prognosis and prediction), NICE (ER+ LN− ESBC for prognosis and prediction), AGO (ER+ LN− ESBC for prognosis) | ESMO (ER+ primary breast cancer for prognosis and prediction), St. Gallen (ER+ ESBC for prognosis and prediction), NCCN (ER+ ESBC for prognosis and prediction), ASCO (ER+ LN− ESBC, for prognosis and prediction), NICE (ER+ LN− ESBC for prognosis and prediction), AGO (ER+ LN− ESBC for prognosis) | St. Gallen (ESBC for prognosis), NCCN (ER+ ESBC for prognosis), AGO (ER+ LN− ESBC for prognosis) |
| Commercial availability | Yes | Yes | Yes | Yes | Yes | Yes |
reviewed articles was given a rating according to the best fit against the four study types of the Simon model (Table 1). If individual elements of a single study were not all rated as the same category, the lowest category of an individual element was used as the study's overall rating.

3 | RESULTS

Key elements of each study and the assigned Simon LOE are summarized in Table 3.

3.1 Breast cancer index

The Breast Cancer Index (BCI) is a multi-gene quantitative reverse transcriptase-polymerase chain reaction (RT-PCR) assay with two components, the HOXB13:IL17BR ratio (H/I) and the Molecular Grade Index (MGI). The H/I ratio was derived by evaluating differential gene expression in 60 tamoxifen-treated (TAM+) estrogen receptor-positive (ER+) postmenopausal ESBC patients. The MGI component of BCI was developed by identifying genes differentially expressed between tumors of low and high grade, focusing on those involved in invasive growth. The H/I ratio and the MGI have been the subject of independent prognostic validation studies. As the individual components are not used separately in commercial assays, their predictive value is not reviewed here.

The BCI assay was trained in 314 samples of postmenopausal TAM+ ER+ lymph node-negative (LN−) ESBC patients from the Stockholm trial of adjuvant tamoxifen (TAM). This resulted in a continuous risk score from 0 to 10. The Stockholm trial, conducted between 1976 and 1990, randomized postmenopausal LN− patients to TAM for 2-5 years, versus no treatment. BCI was validated in 274 patients from the tamoxifen-untreated (TAM−) arm of the trial. BCI was significantly prognostic (P < 0.001) for distant recurrence (DR), independent of tumor size, grade, progesterone receptor (PR) status, and HER2/neu status.

A new BCI algorithm was subsequently developed from 283 postmenopausal subjects using the TAM− arm of the same Stockholm trial, and then validated in 317 TAM+ trial patients. Use of the same patient cohorts in these two separate studies, with different algorithms, diminishes the independence of the two studies and lessens their value for clinical validation. Patients were classified into low (64%), intermediate (20%), and high-risk (16%) groups using the new algorithm with 5-year distant recurrence free survival (DRFS) of 98%, 95.2%, and 87.8%. There was a significant (P = 0.0063) association of risk group with DRFS. External validation was performed in a multi-institutional convenience sample of 358 pre- (30%) and post-menopausal patients with ER+ LN− tumors. Treatment was heterogeneous in this group, with 32% having received CTx on an ad hoc basis. Univariate, and Cox multivariable analysis, demonstrated BCI as the only significant prognostic indicator among age, tumor size, tumor grade, PR-status, or CTx treatment in this sample.

Sgroi et al evaluated 665 primary tumor samples from postmenopausal ER+ LN− patients entered into the randomized ATAC trial of TAM and anastrozole. The RNA analyzed had been prepared by Genomic Health for another study with the ATAC group using its proprietary technology. In the primary analysis, use of the cubic model of the BCI as a continuous variable was not associated with DR (interquartile HR 1.39 [95%CI 0.990-3.70]; LR-Δγ² = 3.70; P = 0.054), even though the pre-specified categorical risk groups were associated with DR (P < 0.0001). In a secondary analysis, a revised linear model of BCI was utilized, demonstrating significant differences in absolute DR rate among the BCI risk groups (P < 0.0001). DR at 10 years for the low, intermediate, and high-risk groups was 4.8% (95%CI, 3.0-7.6), 18.3% (95%CI, 12.7-25.8), and 29% (95%CI, 21.1-39.1), respectively.

The cubic model of BCI was again tested in 292 patient samples from both arms of the NCIC MA14 clinical trial of TAM and octreotide, versus TAM alone. Octreotide proved to be an inactive agent in the parent study. All patients were postmenopausal and 92% had ER+ disease. A total of 51% were LN− and 35% received adjuvant CTx. BCI risk groups had a significant univariate association with relapse free survival (RFS) (stratified log-rank P = 0.005). A stratified Cox stepwise multivariate model only demonstrated a significant hazard ratio for T-stage ≥ T2 (HR, 2.22 95%CI 1.22-4.07; P = 0.01) and for higher continuous BCI (P = 0.004). The BCI low-risk (49.7%), intermediate-risk (23.6%), and high-risk (26.7%) groups had 10-year RFS rates of 87.5%, 83.9%, and 74.7%, respectively. BCI was prognostic for both LN− and lymph node-positive (LN+) patients.

There are no high LOE studies of BCI that predict the value of initial postoperative adjuvant ETx or CTx in ESBC patients.

3.2 Endopredict

The Endopredict assay was developed using microarray gene expression data from 964 ER+, HER2/neu-negative (HER2-), LN− tumors from TAM+ patients. RT-PCR expression of eight cancer-related genes and three standardization genes were combined algorithmically to produce a continuous risk measurement. A cutpoint of independent prognostic validation studies. As the individual components are not used separately in commercial assays, their predictive value is not reviewed here.

Clinical validation using a prospective-retrospective design evaluated 1702 postmenopausal ER+, LN− or LN+ samples from ETx-only individuals in the ABCSG-6 (378 patients) or ABCSG-8 (1324 patients) trials. ABCSG-8 included subjects with low to moderate risk features, excluding grade three tumors. The study suggested that EP and EPclin scores were significant predictors of DR. The 10 year DR rates for patients with EP-low and EP-high were 8% and 22% in ABCSG-6 (P < 0.001) and 6% and 15% in ABCSG-8 (P < 0.001). More than 51% of the combined study population were EP-high despite the low to moderate clinical risk features of the ABCSG-8 patients. Nonetheless, multivariable Cox models demonstrated continuous EP score and nodal status to be independent predictors of DR.

When the same postmenopausal ABCSG-6 and ABCSG-8 populations were assessed in a study by Dubsky, EPclin, produced a greater difference between risk groups (HR 5.11, 95%CI 3.48-7.51; log rank P < 0.001) than National Comprehensive Cancer Network (NCCN) 2007 guidelines (HR 2.16, 95%CI 0.80-5.85; P = 0.119), German S3 2008 guidelines (HR 2.20, 95%
| Assay          | Focus                        | Study                                      | Setting | Pre-menopausal | Post-menopausal | ER+ | ER- | LN+ | LN- | Simon category |
|---------------|------------------------------|--------------------------------------------|---------|----------------|-----------------|-----|-----|-----|-----|----------------|
| Blue print    | Prognostic                   |                                            |         |                |                 |     |     |     |     | D              |
|               |                              | Krijgsman Breast Cancer Res Treat 2011     | Various | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
| Breast Cancer Index | Prognostic                   |                                            |         |                |                 |     |     |     |     | B              |
|               |                              | Goetz Clin Cancer Res 2006                 | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Jerevall Br J Cancer 2011                  | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Sgroi Breast Cancer Res 2016               | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Prognostic and predictive    |                                            |         |                |                 |     |     |     |     | B              |
|               |                              | Sgroi Lancet Oncol 2013                    | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Zhang Clin Cancer Res 2013                 | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
| Endopredict   | Prognostic                   |                                            |         |                |                 |     |     |     |     | B              |
|               |                              | Filipits Clin Cancer Res 2011              | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Dubsky Ann Oncol 2013                      | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Martin Breast Cancer Res 2014              | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Buus J Natl Cancer Inst 2016               | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
| IHC4          | Prognostic                   |                                            |         |                |                 |     |     |     |     | B              |
|               |                              | Cuzick J Clin Oncol 2011                   | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Park Oncology 2014                         | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | C              |
|               |                              | Sgroi Lancet Oncol 2013                    | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               |                              | Stephen Br J Cancer 2014                   | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
| MammaPrint    | Prognostic                   |                                            |         |                |                 |     |     |     |     | D              |
|               |                              | Bueno-de-Mesquita Lancet Oncol 2007        | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
|               |                              | Buyse J Natl Cancer Inst 2006               | No Rx   | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | C              |
|               |                              | Cardoso N Engl J Med 2016                  | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | A              |
|               |                              | Mook Ann Oncol 2010                        | AET     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
|               |                              | Mook Breast Cancer Res Treat 2009          | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
|               |                              | van de Vijver N Engl J Med 2002            | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
|               |                              | Wittner Clin Cancer Res 2008               | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |

(Continues)
| Assay          | Focus                              | Study                                | Setting | Pre-menopausal | Post-menopausal | ER+ | ER− | LN+ | LN− | Simon category |
|---------------|------------------------------------|--------------------------------------|---------|----------------|-----------------|-----|-----|-----|-----|----------------|
| Predictive    | Simon category                     |                                      |         |                |                 |     |     |     |     | D              |
|               | Knauer Breast Cancer Res Treat 2010 | ACT                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | D              |
| Oncotype      | D                                   |                                                     |         |                |                 |     |     |     |     | B              |
| Prognostic    | D                                   | Oncotype                               | ACT     | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Dowsett J Clin Oncol 2010           | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Palk N Engl J Med 2004              | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Sparano N Engl J Med 2015           | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Gluz J Clin Oncol 2016              | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | A              |
| Prognostic and predictive |                                  |                                      |         |                |                 |     |     |     |     | A              |
| Predictive    | D                                   | Albain Lancet Oncol 2010             | AET/ACT | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
| PAM50         | D                                   |                                      |         |                |                 |     |     |     |     | B              |
| Prognostic    | Chia Clin Cancer Res 2012           | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Dowsett J Clin Oncol 2013           | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Gnant Ann Oncol 2014                | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Gnant Ann Oncol 2015                | AET                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
| Predictive    | Cheang Clin Cancer Res 2012         | ACT                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |
|               | Liu Breast Cancer Res Treat 2015    | ACT                                  | ✓       | ✓              | ✓               | ✓   | ✓   | ✓   | ✓   | B              |

AET, adjuvant ETx; ACT, adjuvant CTx; ER+, estrogen receptor positive; ER−, estrogen receptor negative; LN+, lymph node positive; LN−, lymph node negative; LOE, level of evidence (Simon J Natl Cancer Inst 2009).
CI 1.16–4.19; P = 0.014), or St. Gallen 2011 guidelines (HR 2.78 95% CI 1.50–5.14; P < 0.001).\(^{20}\) EPClin reassigned 58 (61%) of patients guideline-classified as high-risk to EPClin low-risk. However, the clinical guidelines of the era classified over 80% of the patients as high-risk to begin with. Ultimately, the authors suggested that the preponderance of truly low-risk patients in this cohort limited the statistical power of the study, proposing that the analyses for the remaining high-clinical-risk subgroup be regarded as exploratory. The authors suggested CTx for the high-risk group, but there was no evidence presented to support EPClin as a predictor of CTx benefit.

Martin studied the prognostic properties of EP and EPClin in an independent cohort of 555 patients with ER+ LN+ disease from the 1246 patient prospective GEICAM 9906 study treated with CTx and ETx.\(^{21}\) Both EP and EPClin-defined risk groups were significantly associated with distant metastasis free survival (DMFS) (log rank P < 0.0001). For EP, 25% (141) of evaluable patients were low-risk with estimated DMFS at 10 years of 93%. Among the 75% (414) of high-risk patients, 10-year DMFS was 70%. The absolute difference in risk between low-risk and high-risk EP groups was 23% (HR, 4.8 (95%CI 2.45–9.55), P < 0.0001). Using EPClin, only 13% were low-risk with 100% DMFS at 10 years. The remaining high-risk patients had a 10-year DMFS of 72%.

GEICAM 9906 included 300 (54%) premenopausal and 255 (46%) postmenopausal patients, with prognostic value evident in both groups (premenopausal, HR, 6.7; P < 0.0001; postmenopausal, HR, 3.3; P = 0.0069). EPClin was similarly prognostic for both premenopausal (P = 0.0006) and postmenopausal subsets as well. (P = 0.0023). As all patients in GEICAM 9906 received CTx, EP as a predictor of CTx benefit could not be separately assessed. To date, there are no LOE studies of EP or EPClin that identify a role for the assays in predicting CTx or ETx benefit.

The prognostic capability of EP and EPClin were compared to Oncotype DX using 928 RNA samples from the ATAC study, previously prepared by Genomic Health.\(^{22}\) In a multivariate analysis, the EP, EPClin, and the Oncotype DX assay were all prognostic for 10-year DR. EP provided more prognostic information than the Oncotype DX Recurrence Score (RS) in the 5-10 year interval, but the assays were equally prognostic from 0 to 5 years. Clinical factors in the EPClin score resulted in prognostic information that exceeded both the RS and EP. However, clinical factors were not generated by the assay and were not similarly incorporated into the RS data, making any direct comparison of EPClin and Oncotype DX problematic.

The genomic component of these assays sorts patients into low and high-risk groups of different sizes: EP classified more patients as high-risk (58.4%) compared to the Oncotype DX assay, which classified 38.3% with a “non-low” RS (RS ≥ 18). The hazard ratio for 10-year DRS between the low-risk and non-low risk patients was 2.98 (95%CI 1.94-4.58, P < 0.001) for EP compared to HR = 2.73 (95%CI 1.91-3.89, P < 0.001) for Oncotype DX). Although the HRS are similar and the confidence intervals overlap, a larger proportion of EP patients would be considered high-risk and might receive recommendation for CTx.

Martin, et al, more recently evaluated patients from the same GEICAM 9906 study and compared prognostic features of EP and EPClin with a non-commercial research version of PAM50.\(^{23}\) However, the comparison does not represent a real-world evaluation of the Prosigna assay, providing little value for practical decision-making.

### 3.3 IHC4

IHC4 is an integrated immunohistochemistry (IHC) assay that utilizes four breast cancer protein biomarkers (ER, PR, HER2, and Ki-67) to assess risk in patients with ER+ ESBC.\(^{24}\) Initial analysis was performed in a single laboratory using unique antibodies for IHC, careful H-scoring for ER, and specialized image analysis. Expression levels of these proteins are incorporated into the IHC4 algorithm. A clinical treatment score (CTS), derived from clinicopathologic features, may be added to obtain an overall risk score. The assay is performed with formalin-fixed paraffin-embedded (FFPE) tissue.

Development and validation was based on FFPE tissue samples from the ATAC trial.\(^{24}\) The biomarkers were evaluated by likelihood ratio testing and shown to contribute prognostic information to time to DR (TTDR). In the ER+, LN− discovery cohort of ATAC, IHC4 coupled with CTS had greater prognostic value than CTS alone. (IHC4 LR=3.4 95%CI 2.93-3.37).\(^{25}\)

IHC4 was also evaluated in a convenience sample of 786 ER+, post-menopausal, and pre-menopausal ESBC patients from Nottingham who were either untreated or treated with ETx alone.\(^{24}\) Due to manual reading and the use of a different antibody, Ki-67 levels were nearly 2.5 times higher than in the ATAC cohort, requiring rescaling. After adjustment, the modified IHC4 score was prognostic for outcome when added to the clinical score (HR, 3.9; P < 0.0001).

Sgroi et al evaluated IHC4 in Cuzick’s LN− ATAC population, the same population in which IHC4 was developed, to confirm the prognostic value of the immunohistochemical assay.\(^{24}\) In the 915 patients, the authors examined the likelihood ratio for both IHC4 and BCI; IHC4 appeared prognostic for recurrence to 5 years in multivariable analysis. Stephen and colleagues also showed that IHC4 was prognostic to 5 years using independent Cox analysis in samples from the Edinburgh BCI series and TEAM trial.\(^{25}\) However, others have shown that a nomogram constructed from St. Gallen guidelines and Adjuvant Online (AOL), combined with IHC4 + CTS, provides prognostic information beyond that from IHC4 + CTS alone.\(^{26-28}\)

IHC4 has not been validated to predict therapeutic benefit for ETx or CTx adjuvant regimens. Furthermore, Cuzick and colleagues have acknowledged that variations in consistency and qualitative output across distributed laboratories have limited the value of the assay.\(^{24}\) Commercialization of the assay by Genoptix, may address some of these issues, but validation data for the commercial assay is currently unavailable.

### 3.4 MammaPrint

MammaPrint is a microarray-based assay that uses the expression levels of 70 selected genes to classify tumors as “poor signature” or “good signature.” Investigators used frozen tumor samples from 78 women with ESBC from archives at the Netherlands Cancer Institute
Clinical validation was performed in 295 consecutive NKI patients, younger than 53 years of age with ER+ (76.6%) and ER− (23.4%) early stage breast cancer. There was significant heterogeneity among this population, with 49% LN+, 44% having received some form of adjuvant ETx and/or CTx. Nearly 21% (61/295) were in the original developmental group of patients, introducing potential bias. Overall, poor-signature patients had a higher rate of distant metastasis at 10 years (HR, 5.1, 95%CI (2.9-9.0); \( P < 0.001 \)) than good-signature patients with DMFS of 50.6 ± 4.5%, and 85.2 ± 4.3%, respectively. A subsequent validation study used 307 samples from ER+ (71%) and ER− (29%) subjects ≤ 60 years of age enrolled in the TRANSBIG registry who had not received adjuvant therapy. In this study, 85 of the 90 patients with ER− disease had a high-risk gene signature. The HR for overall survival (OS) by risk group was 2.79 (95%CI 1.60-4.87, \( P < 0.001 \)). Also significant was TTDR (HR, 2.32, 95%CI 1.35-4.00, \( P = 0.002 \)) and disease free survival (DFS) (HR, 1.50, 1.04-2.16, \( P = 0.032 \)). In each case the 70-gene assay provided more robust prognostic information than clinicopathologic assessments, including AOL.

MammaPrint was also assessed in LN+ subjects from a multi-institution convenience sample that identified significant association between the gene signature and BCSS (log rank \( P < 0.001 \)) or 10-year DMFS (log rank \( P = 0.001 \)). In another convenience sample, Wittner demonstrated a negative predictive value of 100% in 100 LN− postmenopausal patients from the Massachusetts General Hospital. However, the corresponding positive predictive value was only 12%. Furthermore, there was no significant difference in TTDR between the gene signature groups. Mook, used a sample of postmenopausal LN− patients from the NKI to show significant association between the gene signature and BCSS (log rank \( P = 0.036 \)), but not DMFS (log rank \( P = 0.07 \)).

MammaPrint is one of only two assays for which there is published Level I-A evidence supporting prognosis. The MINDACT trial was a multicenter prospective randomized study that enrolled 6693 women with invasive ESBC, after screening 11 288. Of the 4595 screening failures, 26% were reported due to technical failure of the assay. Of enrolled patients, all underwent clinical risk assessment with a modified version of AOL, and had genomic risk assessment performed with MammaPrint. Low-clinical risk was defined as a 10-year BCSS in excess of 88% for ER+ patients, and of 92% for ER− patients. The difference between the ER+ and ER− thresholds was to account for a presumed absolute 4% 5-year benefit of adjuvant ETx.

The MINDACT study utilized a complex design with multiple arms, randomizations, and study subpopulations. Analysis was made more challenging by risk assessment errors in 275 patients (4% of the total population). The clinical risk assessment was faulty in 103 cases, largely due to changes in local site assessment of tumor size, grade, or nodal status. However, 177 patients had a change in genomic assessment due to an assay quality control error. Additional complexity was introduced by 15% patient noncompliance with treatment assignment.

The primary endpoint of MINDACT was survival without distant metastasis at 5 years. This was assessed in a pre-defined “primary test population” derived from clinically high-risk genomic low-risk (cHgL) patients. The 644 patients in the primary test population, representing 9.6% of MINDACT, were cHgL and did not receive CTx. Excluded were 21 patients who had changes in risk assessment and 85 patients who did not comply with treatment recommendations. The rate of survival free of distant metastasis was 94.7% (95%CI, 92.5-96.2), exceeding the prospectively set non-inferiority criterion of 92% and suggesting prognostic value for the MammaPrint assay.

Several studies have suggested an association between MammaPrint risk signature and CTx benefit. In a retrospective meta-analysis study by Knauer using pooled data from previous non-trial clinical reports of adjuvant therapy, DDFS was 93% without CTx and 99% with CTx in good-signature patients (HR, 0.26, 95%CI 0.03-2.02; \( P = 0.20 \)), and 76% versus 88%, in poor signature patients (HR, 0.35, 95%CI 0.17-0.71; \( P < 0.01 \)). The HR reduction was similar for the two groups, despite differences in significance, in this very underpowered study. The small sample sizes and the absence of randomized controlled CTx administration in any of the component studies limit any conclusion. Straver analyzed stage II and III patients with response to neoadjuvant chemotherapy assessed as a function of MammaPrint risk grouping. However, neoadjuvant studies cannot be directly extrapolated to predict degree of long-term benefit in the absence of late outcome information.

The prospective observational RASTER study, conducted in 16 community hospitals in the Netherlands, has been proposed to support MammaPrint use for therapy selection. However, treatment decisions were made by the patient and treating physician, in a discretionary fashion, based on a combination of MammaPrint, prognostic indicators, treatment guidelines, and personal preferences. The heterogeneity of treatment and the absence of MammaPrint as the treatment discriminator make interpretation of the outcome data difficult.

MINDACT has provided the only high LOE data available to assess MammaPrint’s role in therapeutic treatment benefit. CTx or no-CTx, was determined by randomization of 2745 patients in each of the discordant clinical and genomic risk assessment arms, on an “intent to treat” basis. The 5-year DMFS rate among the cHgL patients receiving CTx was 95.9% (95%CI, 94.0-97.2) and 94.4% (95%CI, 92.3-95.9) for those not receiving CTx. A 1.5% advantage in 5-year DMFS (with overlapping confidence intervals) was noted for those receiving CTx. In the per protocol population, the hazard rate reduction for CTx in the cHgL group ranged from 34 to 37% for DMFS, DFS, and OS. Notable was the statistically significant impact on DFS in this underpowered analysis (HR 0.64; 95%CI, 0.43-0.95, \( P = 0.03 \)). For the clinical low-risk genomic high-risk (cLgH) discordant group, there was no apparent advantage to genomic assay-directed CTx, with 5-year survival free of distant metastasis 95% for both. In the per protocol population, there was no statistical advantage in DMFS, DFS, or OS for CTx administration. When reviewing both discordant groups, these data cannot exclude a small advantage for CTx in the cHgL group, but suggest no benefit for the use of chemotherapy in the cLgH group. As a result, MINDACT data provides no evidence of MammaPrint predictive CTx value in poor prognosis patients.
Two companion microarray-based assays are marketed with MammaPrint. TargetPrint offers quantitative assessment of ER, PR, and HER2 expression; Blueprint, when used with MammaPrint, classifies tumors into molecular subsets. Concordance between Blueprint/MammaPrint and the original intrinsic gene set from Perou et al is 92%. However, analysis of the data suggests that these approaches characterize overlapping but distinct populations of luminal patients.

There are no studies with long-term outcome endpoints to assess the value of the MammaPrint/Blueprint combination. Whitworth has suggested a value in intrinsic subset reassignment with a resultant improvement in neoadjuvant pCR rates, evaluated in 403 patients from the NBRST prospective registry. However, the study had no long-term outcome endpoints, relying only on the surrogate endpoint of pCR. Variations in the degree of pCR cannot be correlated to quantitative outcome differences across varied populations, particularly in hormone receptor positive (HR+) HER2− breast cancer.

### 3.5 Oncotype DX

Oncotype DX is an RT-PCR-based assay that measures expression levels of 21 genes, in RNA from FFPE tissue, to assess the risk of DR in ER+ ESBC. The assay was developed from 250 prognostic breast cancer genes, using a total of 447 patients from three independent studies. The 21 genes included 16 cancer-related genes, and five reference genes. A weighted algorithm produced a continuous RS from 0 to 100 which can be used to assign patients to low (0-17), intermediate (18-30), or high-risk (31) categories. Alternate risk cutpoints have been proposed.

Validation of Oncotype DX was performed in a prospective-retrospective study using 668 archived tissue samples from the TAM+ arm of the NSABP B-14 trial. The 10-year DR free rate (DRFR) was retrospective study using 668 archived tissue samples from the TAM+. The 10-year DR free rate (DRFR) was evaluated in 403 patients from the NBRST prospective registry. However, the study had no long-term outcome endpoints, relying only on the surrogate endpoint of pCR. Variations in the degree of pCR cannot be correlated to quantitative outcome differences across varied populations, particularly in hormone receptor positive (HR+) HER2− breast cancer.

The value of the Oncotype DX assay as a predictor of benefit from systemic chemotherapy was addressed in two separate prospective-retrospective studies, yielding Level 1-B validation evidence from both studies. Paik analyzed samples from ER+ LN− subjects in the NSABP B-20 trial that had randomized ER+ LN− patients to TAM, versus TAM. The continuous RS was a significant predictor of DFS (HR per 50 units, 2.64, 95%CI 1.33-5.27; P = 0.006). The continuous RS was also prognostic for OS over a 10-year period, with HR 4.42 (95%CI 1.96-9.97; P < 0.001) adjusting for nodes.

The highest level of evidence for prognosis currently available comes from the prospective Cooperative Group trial, TAILORx, which was initiated in 2006 and which was specifically designed to evaluate Oncotype DX. The study population of 10 253 eligible ER+, HER2−, LN− Oncotype DX tested women was divided into three predetermined risk groups with RS cutpoints at 11 and 25. Cutpoints lower than the prospectively defined cutpoints for the initial clinical validation studies were utilized to avoid patient undertreatment by accounting for the ends of the 95% confidence intervals for 10-year risk of recurrence rather than just the means. Patients in the RS < 11 group were assigned ETx alone, those in the mid-range group with RS 11-25 were randomized to CTx followed by ETx versus ETx alone, and those in the RS > 25 group were assigned CTx followed by ETx. Results from the mid-range group randomization are expected in 2017. However, data from the low-risk group of patients with RS < 11 have been published, providing TMUGS Level I-A clinical evidence for prognosis in this population. Of the 1626 low-risk patients who were assigned ETx alone, the 5-year invasive DFS was 93.8% (95%CI, 92.4-94.9), the proportion free of DR was 99.3% (95%CI, 98.7-99.6), and the OS was 98.0% (95%CI, 97.1-98.6). For DFS, second primary breast cancers and deaths from other causes exceeded the 5-year risk of recurrence. Multivariate analysis that included age, tumor size, histologic grade, and surgery type showed no significant association with the rate of invasive DFS or freedom from DR. Only histologic grade showed any statistically significant association (P = 0.02) with freedom from any recurrence between intermediate and low grade (HR 8.07; 95%CI 1.06-61.45), and between high and low grade (HR 4.73; 95%CI 0.29-76.42).

Prospective data are also available from the West German Study Group Plan B trial. This study, initiated in 2009, recruited 3198 patients. It was designed to assess alternative CTx regimens in patients with clinically high-risk LN− and LN+ ESBC. Oncotype DX testing was successfully performed in 2577 patients, representing 98% of the total HR+ population with tumor tissue samples. After an early protocol amendment, ETx treatment alone was recommended and accepted by 348 HR+, HER2− patients with pN0-1 disease and RS ≤ 11. Although 31.4% were LN+ and 20% had grade three disease, the 3-year DFS rate was 98.4% (95%CI 97.0-99.8%). Among the intermediate-risk group (RS12-25), the 3-year DFS was 97.5% (95%CI, 95.9-99.0), and among the high-risk group (RS > 25) the 3-year DFS was 94.9% (95%CI, 91.4-98.4), with all patients in both groups assigned adjuvant CTx. Although only a 3-year DFS endpoint has been published, the authors suggested that the favorable outcome with ETx alone in the low-risk group precluded any potential benefit of CTx.

The value of the Oncotype DX assay as a predictor of benefit from systemic chemotherapy was addressed in two separate prospective-retrospective studies, yielding Level 1-B validation evidence from both studies. Paik analyzed samples from ER+ LN− subjects in the NSABP B-20 trial that had randomized ER+ LN− patients to TAM, versus TAM.
plus CTx. Subjects in the high-risk group (RS ≥ 31) had a significantly reduced DR when CTx was utilized (RR, 0.26; 95%CI, 0.13-0.53). In contrast, there was no significant benefit of CTx for the low (RS < 18) (RR, 1.31; 95%CI, 0.46-3.78) or intermediate-risk (RS18-30) (RR, 0.61; 95%CI, 0.24-1.59) groups. Cox models demonstrated significant interaction between the continuous RS and CTx (P = 0.038).

Albain analyzed the Oncotype DX assay in 367 (40%) available samples from the SWOG-8814 study randomized to sequential CAF and TAM+, versus TAM+ alone. Log rank test of CTx benefit, stratified by number of positive nodes, revealed a trend toward a DFS benefit for the entire population of analyzed patients (stratified log rank P = 0.054) consistent with the parent trial. However, when analyzed by the Oncotype DX test, DFS improvement for patients receiving CAF was confined to the high-risk group with RS ≥ 31 (log rank P = 0.033; HR = 0.59, 95%CI 0.35-1.01). There was no apparent DFS benefit for the addition of CTx to patients with RS < 18 (log rank P = 0.97; HR = 1.02, 95%CI 0.54-1.93) or RS18-30 (log rank P = 0.48; HR = 0.72, 95%CI 0.39-1.31). OS benefit was similarly restricted to the high-risk group (stratified log rank P = 0.027), without improvement for the low (stratified log rank P = 0.63) or intermediate-risk (stratified log rank P = 0.85) groups.

### 3.6 | Prosigna

The Prosigna breast cancer test is a multiplex assay that measures the expression of 50 classification genes and eight housekeeping genes in order to assign a “risk of recurrence” (ROR) score to patients with ESBC. The assay, initially termed PAM50, was adapted and commercialized by Nanostring, Inc. for use in their nCounter analytic system, consisting of a single hybridization reaction with specific labeled probes. Gene expression of 46 of the genes is used to compare a specimen to prototypical intrinsic biologic subsets originally described by Perou. A proprietary algorithm combines the gene expression data, a proliferation score, and tumor size to calculate an ROR score that corresponds to the 10-year risk of DR. Patients are assigned to low, intermediate, or high-risk groups, based on pre-determined cut-points. Unlike other tests, the cutpoints vary by clinicopathologic feature. All patients with ≥4 LN+ are considered high-risk. Patients with 1-3 LN+ and ROR ≤ 15, or LN− patients with ROR ≤ 40, are classified as low-risk. Satisfactory analytic validity has been reported when the test is conducted using standardized procedures in local molecular biology laboratories.

Dowsett evaluated Prosigna in 940 RNA samples previously extracted by Genomic Health from the ATAC trial. The prospective-retrospective study demonstrated that a continuous ROR score added prognostic value for DR beyond CTS for all patients in the LN− and LN+, ER+, postmenopausal population treated with ETx (LR-Δχ² = 33.9, P < 0.001). In addition, Kaplan-Meyer analysis showed ROR risk groups were prognostic for 10-year DR, varying by LN status.

Two prospective-retrospective reports of Prosigna utilized RNA and data from the previously described ABCSG-8 study. Each study analyzed the ABCSG-8 population of postmenopausal patients with grade 1-2 tumors either alone or with additional patients from ATAC. In their primary validation study, Ganet and colleagues analyzed 1478 patients from ABCSG-8 and developed a prognostic clinical linear predictor (CLP) based on tumor grade, tumor size, and nodal status. The ROR score provided a significant additional increase in prognostic information over the CLP (log −likelihood test: ΔLRy2 = 53.49; P = 0.0001). Subgroup analysis suggested that additional information was provided for all LN− and LN+ patients except those with HER2-positive disease. Pre-specified ROR risk groups were also prognostic for DFS with a 10-year probability of 96.7% (95%CI 94.6-98.0) for the low-risk group, 91.3% (95%CI 88.1-93.8) for the intermediate-risk group, and 79.9% (95%CI 75.7-83.4) for the high-risk group. Classification to luminal A versus luminal B subtype was prognostic for 10-year DMFS (HR, 2.85; 95%CI 2.04-4.00; P < 0.0001).

Evaluation of ROR in LN+ ER+ post-menopausal women was conducted in relevant patients from ABCSG-8 and ATAC. Analysis revealed prognostic value for DMFS in both the single LN+ subgroup (high- versus low-risk: HR, 3.56, 95%CI 1.62-7.80, P = 0.0016) and the 2-3 LN+ subgroup (high- versus low/intermediate-risk: HR, 3.023, 95%CI 1.462-6.249, P = 0.0028) using pre-defined cutoffs. However, the LN+ number itself had influence on subgroup assignment beyond the actual gene expression though its impact on the ROR subgroup cutoff. Intrinsinc subtype analysis revealed improved DMFS for patients with Luminal A versus Luminal B tumors, in both N1 and N2-3 subgroups.

Chia further evaluated PAM50 in 398 premenopausal Stage I-III HR+ and hormone receptor-negative (HR−) patients who received adjuvant CTx as part of NCIC MA. Unlike the commercial assay, RT-PCR rather than NanoString n-Counter technology was used. HR+ individuals represented 73% of patients, while 94% were either Stage I or II. Nearly 75% had positive nodes. In adjusted multivariate analysis PAM50 intrinsic subtyping was prognostic beyond clinicopathologic variables for both DFS (P = 0.02) and OS (P = 0.02). Continuous ROR, combined with a proliferation score and tumor size correlated with 10-year relapse in both LN− and LN+ patients using ROC analysis (DFS C-index = 57.6%, 95%CI 52.5-62.1%, and for OS C-index = 61.1%, 95%CI 55.8-66.3%). However, the study’s primary focus was on intrinsic subtype classification and did not adequately pre-specify the specific ROR model that would be utilized. The study did suggest prognostic value for PAM50 in CTx treated premenopausal populations. However, as CTx was mandated in all patients, impact of CTx on outcome cannot be fully appreciated.

Cheang evaluated the prognostic properties of the quantitative RT-PCR version of PAM50 in 476 LN+ premenopausal patients from the NCIC MA.5 randomized controlled trial who received one of two CTx regimens. Correlation between the assay, expressed as ROR-subtype, and the clinical outcome showed the ROR risk classifier was associated with a distinct 5-year RFS and OS prognostic advantage (P < 0.0001; log rank test). However, as all of the randomized arms of the original study received some form of CTx, any predictive value of ROR on selecting adjuvant PCTx, could not be assessed.

The only large prospective-retrospective study to evaluate PAM50 for adjuvant taxane therapy benefit prediction was published by Liu and associates, based on the NCIC CTG MA.21 clinical trial. The original study enrolled 2104 patients, with 1105 available for...
PAM50 analysis. Pre-menopausal and post-menopausal women up to
60 years of age with LN+, or high-risk LN− breast cancer were enrolled
between 2000 and 2005. Patients were randomized to doxorubicin,
cyclophosphamide, and paclitaxel (AC/T); dose-intense cyclophos-
phamide, epirubicin, and fluorouracil (CEF); or dose-dense, dose-
intense epirubicin, cyclophosphamide, and paclitaxel (EC/T). None of
the patients were randomized to ETx without CTx. CEF and EC/T
proved superior to AC/T (P = 0.01). Although continuous ROR was
associated with poorer RFS (P = 0.03), categorical ROR comparing the
high ROR with the combined low and intermediate ROR risk groups
was neither prognostic nor predictive. The lack of prognostic
performance of the categorical ROR may have been related to the
finding that 78.7% of patients were classified into the ROR high-risk
group and only 3.4% were identified as ROR low-risk. The
PAM50-based intrinsic subtypes also provided no predictive value
for treatment selection or benefit. Only in the evaluation of RFS
through multivariate analysis did intrinsic subtype and continuous
ROR demonstrate prognostic, but not predictive, associations
(P = 0.03 and P = 0.002, respectively).

4 | DISCUSSION

Molecular assays provide an opportunity for more accurate assessment
of tumor biology than traditional clinicopathologic classifiers. The six
commercially available gene expression assays reviewed in this report all
provide prognostic information designed to estimate an individual's risk
of tumor recurrence and/or mortality. Some manufacturers also suggest
a predictive role for their tests in selecting among different systemic
therapy options. However, each assay is based on different gene sets,
has been developed in a unique population of varying heterogeneity,
and was tested in studies of varying quality.

When assessing a genomic test, an appreciation of the analytic
validity, clinical validity, and clinical utility are paramount. Peer-reviewed
published evidence, rather than podium presentations or a marketing
clearance, should be the prime consideration. Studies should be logical
and easily interpreted, with unequivocal primary and secondary
endpoints. Any assessment of chemotherapy benefit prediction should
be conducted in a randomized clinical trial that compared patients treated
with chemotherapy to patients who did not receive chemotherapy.
Although differences in neoadjuvant pCR may suggest assay value,
differences in the rate of pCR do not directly correlate with long-term
outcome, particularly when assessing heterogeneous populations.59,60
Clinical endpoint data should capture short-term and relevant long-term
cancer events.

Analytic validation reflects the standardized performance and
reproducibility of a specific assay technique. Ideally, an assay's
methodology should be fixed prior to conducting definitive clinical
validation studies so that the validation evidence adequately reflects
performance of the commercial test. Dissonance between a validation
test and the commercially marketed assay is illustrated by the BCI,
which has undergone several iterations. The first two key BCI studies
used opposite arms of the same Stockholm trial for development and
validation.14,16 This type of crosstalk between the studies weakens
their independence and, therefore, their validation value. Further-
more, the assays developed from each strategy were, in effect,
different tests. The third validation study in ATAC patients utilized two
different approaches to scoring (BCI-cubic and BCI-linear), with the
BCI-cubic (primary analysis) a non-significant prognostic test, and the
BCI-linear (secondary analysis) highly prognostic. Although the ATAC
BCI evaluation by Biotheranostics used commercial assay protocols,
Genomic Health had performed the original ATAC RNA isolation with
its own proprietary technology. As a result, BCI, as well as subsequent
Prosinga and EP evaluations of patient cohorts from ATAC,
incorporated a technique not reproduced in their ultimate commercial
test.8,22,53 Similar variations between validation study procedure and
the commercial assay are noted in the Prosinga test. The final Prosinga
 assay gene panel differed from the initial PAM50 assay by several gene
 panel deletions and additions, and the assay was performed using RT-PCR in some validation studies and using NanoString technology in
 others.56,61

IHC4 analytic validity has also come under scrutiny. Because of
the variability in IHC assays, particularly Ki-67, performed locally with
varying reagents, and cutpoints, a fixed commercial version has been
developed, marked by Genoptix as NexCourse Breast. However, there
is no published validation to link the Genoptix NexCourse Breast
commercial results with the previously published IHC4 validation data.

Although the genes selected and the algorithm utilized for the
MammaPrint assay have remained unchanged throughout its
development, there have been significant procedural changes,
including a shift from fresh frozen/RNA-preserved tissue to
formalin-fixed/paraffin-embedded tissue, and variations in the
microarray plates utilized between studies and the commercial
test. A study performed by Sapino and colleagues evaluated
MammaPrint-FFPE to MammaPrint-fresh sample pairs.63 The
results for the two types of tissue preservation had a Pearson
 correlation coefficient of 0.93 (95%CI 0.92-0.94) with the majority
of the discordant samples in both studies within a 10% range of the
cutpoints. This discordance, typically centering about the cutpoints,
calls into question the strategy of a strict high-risk/low-risk
reporting model promoted for this assay.

Of all the commercially available assays, only the Oncotype DX
test made use of the commercial assay procedure for its entire suite of
validation testing. The Oncotype DX assay was developed in FFPE
tissue and all validation studies have used the same algorithm.
Although clinical parameters may be used with the assay for clinical
treatment decision-making, these parameters were not required for
the assay's validation and are not incorporated into the RS calculation.
Analytic validity should reflect both the quality controls related to
pathology, histology, RNA extraction, and other pre-assay procedures
and the precision capabilities of the test itself.
EPclin and Prosigna ROR Score add clinical parameters to the core genomic assay in order to produce a prognostic test with added precision. However, the primary purpose of genomic assays is to provide new information not otherwise available by an assessment of common clinical parameters. Clinical features should always be considered for ultimate real-world decision-making, but should be added to, rather than intrinsic to, the genomic assay. Combined scores may be misleading as noted in comparisons of EP and EPclin with the Oncotype DX test result. The Kaplan-Meier analysis showed a similar HR separation between low and non-low-risk groups for both the EP and Oncotype DX tests for 10 years. The Oncotype DX assay characterized 48% more low-risk patients than EP alone, but only 5% more than EPclin. However, the one-sided use of recognized clinical prognostic features limits the value of such evaluations and provides little information about the quality of the core genomic component.

An important component of analytic validity is the ability of the assay to produce a reliable result. In the MINDACT trial, 26% of ineligible patients, or 10% of the total patients screened, suffered an analytic failure of the assay. Few of the remaining assays have published failure rates. However, the final analytic failure rate of the Oncotype DX test has been reported at less than 1% of 103 863 sequential specimens.6 The MammaPrint assay has also suffered an important quality control (QC) lapse that was unrecognized for several months. This led to misreporting of results at both its European and American laboratory sites. There was a resulting over-reporting of risk and inappropriate chemotherapy recommendation for hundreds of patients. This quality control issue led to an FDA recall for the California laboratory, and complicated the conduct of the MINDACT assay in Europe.35,65 Although the error was reportedly due to a supplier change in a reagent used for RNA extraction, the episodes demonstrate the importance of internal ongoing QC checks for single tests that may have a dramatic impact on patient care.

Clinical validation requires clinical trials that are consistent with an assay’s target population and that address relevant clinical questions. The study population must be sufficiently well defined to allow reasonable translation to common disease settings. Outcome statistics should not be confounded by widely varying treatments and group heterogeneity. Given the value of ETx in HR+ ESBC, appropriate prognostic validation studies in this population should include ETx as a base upon which chemotherapy decisions may be evaluated.66,67

An example of confounding population heterogeneity is evident in the original MammaPrint validation study that included an assortment of different patient populations receiving an assortment of varying treatments.30 Of the 295 patients in the study, 144 were node positive and 151 were node negative. ER+ was a characteristic in 69, with 226 ER−. Nearly 37% happened to receive adjuvant CTx, while 56% had been untreated. No patient treatments were randomized or protocol-directed. In the subsequent study of Buyse, none of the patients received any systemic adjuvant treatment.31 In the BCI validation study, using the MA.14 clinical trial, 35% of patients received adjuvant CTx, confounding the prognostic value of the assay. As a result, the impact of CTx on the prognostic value of BCI remains uncertain.18

The TMUGS process for assessing genomic assays, is designed to standardize validation by assigning levels of evidence to all supporting published studies.5,7 The best support for any gene expression assay comes from a prospectively designed and prospectively conducted study. A single well-designed prospective study, conducted in a relevant population, with sufficient follow-up, can serve as the highest level of evidence. However, the cost of such studies may be prohibitive, and they may not always be feasible. The prospective-retrospective study provides an alternative for developing high level of evidence, when confirmed by two or more such studies with consistent results.

To date, only three completely prospective studies have been reported (TAILORx, PlanB and MINDACT). Peer-reviewed publications are available for the low-risk population of TAILORx (RS < 11), the German Plan B (RS ≤ 11) trial, and for several subsets of MINDACT.35,44,47 TAILORx and the Plan B studies are based on the Oncotype DX assay, whereas MINDACT is based on the 70-gene MammaPrint assay. All of the studies demonstrate excellent outcomes for CTx untreated patients in their pre-determined genomic low-risk populations. Prior prospective-retrospective studies have suggested that there is no benefit to CTx in the low-risk Oncotype DX group. However, MINDACT suggests that there may be a small CTx benefit (approximately 1.5% absolute) in the MammaPrint-determined genomic low-risk discordant group, with an associated improvement in DFS. In an editorial accompanying the MINDACT publication, Hudis suggested that this small benefit may be meaningful for some patients.68 Beyond MINDACT, there are no TMUGS-defined high-LOE studies for MammaPrint. Although suggestive of prognostic value, the remaining studies are flawed by suboptimal design, population heterogeneity, as well as significant potential for treatment and detection bias. The remaining four genomic assays do have high-level evidence from at least one or more of prospective-retrospective studies suggesting their prognostic value. However, validation has been in varied patient populations without purely prospective trial data.

In the past year, a published prospective registry study of more than 38 000 Oncotype DX-evaluated ER+, LN−, HER2− patients in the United States produced results that paralleled those of prior prospective-retrospective studies.69 In the SEER database, 21 023 low-risk LN− Oncotype DX patients (RS < 18) had a 5-year breast cancer specific mortality (BCSM) of 0.4% (95% CI 0.3%–0.6%) with only 7% known to have received CTx.69 Even among SEER patients with positive nodes (microscopic to three nodes), 5-year BCSM was only 1.0% (95% CI 0.5–2.0) with 23% known to have received CTx. This registration study provides "real world" confirmation of the clinical utility of the Oncotype DX assay, and provides outcomes evidence that is remarkably consistent with results from earlier validation studies.

But, is the identification of very low-risk group by a well-validated genomic assay sufficient? Should all remaining patients above an assay’s low-risk cutoff be expected to benefit from CTx? Prospective-retrospective data used to evaluate the Oncotype DX assay in the NSABP B-20 study showed no benefit for CTx in patients with low or intermediate RS results. However, there was a disproportionate benefit of CTx in patients with high-risk RS results (relative risk 0.26; 95%CI 0.13–0.53), and a statistically significant test for interaction
between chemotherapy treatment and the RS result. This relationship between the RS and CTx benefit was also found in the prospective- retrospective SWOG 8814 LN+ study with CTx/ETx compared to ETx alone.\textsuperscript{46} Together these studies suggest a patient population (RS ≥ 31) for whom chemotherapy may be remarkably active, providing recurrence risk reductions of up to 75%. As a quantum transition of CTx benefit at RS ≥ 31 is unlikely, this data suggest that some small portion of the intermediate-risk population might be CTx responsive, perhaps at the highest RS levels within this group. However, the majority of intermediate group patients (RS18-30) do not appear to benefit from adjuvant CTx.

None of the remaining assays has TMUGS high-level evidence to convincingly predict CTx benefit in subgroups of tested subsets. MINDACT provided an opportunity to prospectively demonstrate the predictive value of MammaPrint. However, an analysis of the cHLG group showed a consistent risk reduction of 35% when chemotherapy was used in this group. This benefit was statistically significant for DFS, even in this underpowered subgroup. Conversely, the cLGH group provided no evidence of a benefit to chemotherapy, even when driven by the high-genomic risk. Thus, although 48% of the cHLG patients may potentially be spared chemotherapy, 47% of the cLGH patients might be recommended to receive unnecessary chemotherapy, without change in outcome measure. Although a very low-risk group for any genomic assay might not have sufficient risk or benefit to justify chemotherapy, predictive value does matter with increased risk, providing the Oncotype DX assay with the strongest clinical utility argument for deciding on CTx use in patients with ER+ and HER2– ESBC.

Some have suggested that individual prognosis, and the correlation between recurrence risk and CTx responsiveness, are common across all the assays. However, the assays do not classify patients similarly, and outcomes cannot be extrapolated from one test to the next. Poulet and colleagues performed a prospective comparison between the commercial versions of the Oncotype DX and MammaPrint tests in 57 ER+ LN− patients with ESBC.\textsuperscript{70} The disagreement was significant. Many of the MammaPrint high-risk patients were Oncotype DX low-risk, and had high ER expression. In this case, because ER expression correlates with ETx benefit, a poor-prognosis MammaPrint patient might do very well on ETx alone. Poor concordance was also evident (r = 0.08, 95%CI –0.2-0.35; Spearman correlation) between the Oncotype DX and Prosigna commercial assays in 52 patients studied by Alvarado and colleagues. They found a risk-classification concordance of only 54%.\textsuperscript{71}

More recently, the OPTIMA Prelim Trial evaluated five assays, including four of the tests discussed in this review (Oncotype DX, MammaPrint, Prosigna, and NexCourse Breast-IHC4). Each assay was performed in its manufacturer’s own commercial laboratory.\textsuperscript{72} Remarkably, there was significant disagreement between assays, with the overall level of agreement only “moderate” (κ = 0.40–0.59). The authors were perplexed that multiple assays, each with independent validation for prognosis, could have such disagreement. They suggested that assay concordance was no better than classic pathologic grading. However, OPTIMA Prelim did not measure long-term patient outcomes and could not provide independent validation of each of the competing assays, nor could it identify the most accurate of the disparate tests. The poor concordance between all tests suggests that one well validated assay cannot reliably be replaced with others with less robust validation.

The challenge of medical decision-making is to provide the best care for today’s patients while continually assessing new diagnostic and treatment paradigms for those in the future. Adjuvant therapy provides an opportunity to reduce the chance of DR and death, but carries its own risk of short and long-term toxicity. Modern therapies also generate significant costs to individuals and healthcare systems. Genomic classifiers that provide accurate prognostic information and reliable benefit prediction personalize the selection of treatment. They improve the likelihood of favorable long-term outcome while conserving healthcare resources. However, the choice should be made on the basis of a rigorous analysis of the ever-changing supporting data.

5 CONCLUSIONS

Multiple marketed gene expression assays are now available to provide prognostic and predictive information in ESBC. A review of the available studies suggests that each of the six assays has some evidence supporting their use as prognostic tools in varying sub-populations. However only two assays, Oncotype DX and MammaPrint, have published evidence from large prospective clinical trials for prognosis relevant to many ESBC patients. Both of these assays identify a unique subpopulation with low-risk for which chemotherapy would add little benefit. To date, however, only Oncotype DX predicts the benefit of CTx in a broad population of HR+, pre-and postmenopausal, LN− and LN+ patients validated through adjuvant CTx trials that compared randomized CTx-treated with CTx-untreated populations in long-term follow-up. It is this combination of strong prognostic and reliable predictive information that provides an assay with maximum clinical utility.

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CONFLICTS OF INTEREST

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