Fine-tuning the electronic structure of heavy-atom-free BODIPY photosensitizers for fluorescence imaging and mitochondria-targeted photodynamic therapy†

Suje Qi, a Nahyun Kwon, a Yubin Yim, a Van-Nghia Nguyen b * † a and Juyoung Yoon b * a

Theranostics that combines both diagnosis and therapy into a single platform has recently emerged as a promising biomedical approach for cancer treatment; however, the development of efficient theranostic agents with excellent optical properties remains a challenge. Here, we report novel mitochondria-targeting BODIPY photosensitizers (R-BODs) that possess considerable singlet oxygen generation capabilities and good fluorescence properties for imaging-guided photodynamic therapy (PDT). The incorporation of sulfur atoms into the π-conjugated skeleton of BODIPY along with the introduction of different functional groups at the meso-position of the BODIPY core is essential for tuning the photophysical and photosensitizing properties. Notably, the MeOPh-substituted thiophene-fused BODIPY (MeO-BOD, \( R = p \)-methoxyphenyl) displayed the highest singlet oxygen generation capability (\( \Phi_S \approx 0.85 \) in air-saturated acetonitrile) and a moderate fluorescence quantum yield (\( \Phi_F = 17.11 \)). Furthermore, MeO-BOD showed good biocompatibility, low dark toxicity and superior fluorescence imaging properties in living cells. More importantly, the PDT efficacy of mitochondria-specific anchoring of MeO-BOD was remarkably amplified with an extremely low half-maximal inhibitory concentration (IC_{50}) value of 95 nM. We believe that the incorporation of an electron-donating group at the meso-position of the thiophene-fused BODIPY platform may be an effective approach for developing theranostic agents for precision cancer therapy.

Introduction

Photodynamic therapy (PDT) is an effective clinical treatment strategy for malignant tumors. In the PDT process, a photosensitizer (PS) is activated under light irradiation, and the excited PS subsequently interacts with molecular oxygen to generate cytotoxic reactive oxygen species (ROS), which can oxidize biomolecules resulting in cancer cell death. Theranostics, which combines diagnostic and therapeutic capabilities in one dose to achieve the real-time and precise monitoring of the therapeutic effect of the drug. Moreover, PDT combined with image-guided diagnosis has attracted considerable attention owing to its prominent advantages such as high spatiotemporal selectivity, non-invasiveness, and fewer side effects. Therefore, the design of novel PSs that can effectively produce both fluorescence and ROS is in high demand.

To date, numerous organic dyes, such as porphyrins, phthalocyanines, cyanine, squaraine, diketopyrrolopyrrole (DPP), and boron dipyrromethane (BODIPY) derivatives, have been developed as theranostic agents. Among these, BODIPY dyes have attracted great interest as theranostic agents in photodynamic cancer therapy due to their excellent photophysical stability, good biocompatibility, high molar extinction coefficients, high quantum efficiencies of fluorescence and facile modification. Photodynamic therapy utilizing BODIPY dyes, the most popular approach is the introduction of heavy halogen atoms (Br and I) to promote spin-orbit coupling (SOC), which enhances intersystem crossing (ISC) and improves the singlet oxygen \( ^1\text{O}_2 \) generation capability. However, the incorporation of heavy atoms increases the dark-toxicity and quench fluorescence. Thus, BODIPY PSs without heavy halogen atoms are preferred as theranostic agents. Recently, several approaches to enhance the ISC, such as the use of double excited states, spin converters, and photoinduced electron transfer (PET), have been implemented in the development of PSs without...
heavy atoms; however, the search for novel PSs is still required for PDT to reach its full potential.

Intriguingly, one alternative strategy to strengthen the ISC with an efficient triplet population is to increase the SOC while decreasing the singlet–triplet energy gaps; to do this, a thiophene moiety was introduced into the 7- coupled system of BODIPY,

but the oncological applications of this approach have not been fully studied.

Furthermore, the introduction of electron-donating groups at the meso-site of the BODIPY platform is also an emerging strategy for achieving efficient PSs.

The abovementioned factors inspired us to consider designing PSs with different functional substituents incorporated at the meso-position of thiophene-fused BODIPY to achieve compounds suitable for clinical use. To the best of our knowledge, there have been no attempts to design efficient theranostic agents by taking advantage of the above two strategies.

Herein, we designed and synthesized a series of heavy-atom-free thiophene-BODIPY derivatives by incorporating different functional groups (pyridinyl (PY), phenyl (PH), p-methoxyphenyl (MeO) and N,N-dimethylaminophenyl (DMA)) at the meso-position of the thiophene-BODIPY platform (Scheme 1).

The experimental results revealed that the \(^{1}O_2\) generation ability of these thiophene-BODIPY derivatives through a type II process gradually increased with increasing electron-donating ability of the substituent (\(\Phi_{\Delta 5}^{*}\): PY-BOD < PH-BOD < MeO-BOD). Fortunately, MeO-BOD had prominent dual functions, showing both a high \(^{1}O_2\) quantum yield and moderate fluorescence intensity.

Furthermore, the cellular experimental results showed that MeO-BOD could be utilized as a mitochondria-specific diagnostic agent to reinforce the PDT effect.

**Results and discussion**

**Synthesis and photophysical properties**

The different thiophene-fused BODIPY derivatives were synthesized through the trifluoroacetic acid-catalyzed condensation of thiophene-fused pyrroles with different benzaldehydes, followed by oxidation and complexation (Scheme 1).

Detailed experimental procedures are provided in the ESI.† The structures of all these thiophene-fused BODIPY derivatives were fully confirmed by \(^{1}H\) and \(^{13}C\) NMR spectroscopy and high-resolution mass spectrometry (Fig. S1–S14†). UV-vis absorption and fluorescence spectra of the new thiophene-BODIPY derivatives were acquired (Fig. 1, S15 and S16†). These key photophysical property data are summarized in Table 1.

The absorption and fluorescence spectra of all the thiophene-fused BODIPY derivatives were similar in shape to those of H-BOD, indicating that the R substituents had little effect on the HOMO–LUMO energy gap of the thiophene-fused BODIPY chromophore because they are nearly orthogonal to the thiophene-fused BODIPY core (Fig. 1a). All the studied compounds retained high absorption molar extinction coefficients \((\varepsilon \sim 100\,000\,\text{M}^{-1}\,\text{cm}^{-1})\), and the maximum absorptions were shifted to longer wavelengths \((\lambda_{\text{abs}} \sim 565\,\text{nm})\) compared with that of unmodified BODIPY. The absorption spectra of all the thiophene-BODIPY derivatives showed a slight blueshift with increasing solvent polarity. This phenomenon indicates that the dipole moments of the ground state of these thiophene-BODIPY derivatives might be larger than those of the first singlet excited states. On the other hand, PY-BOD, PH-BOD and MeO-BOD exhibited moderate fluorescence intensities, while DMA-BOD was poorly emissive due to a PET mechanism.

The fluorescence quantum yields of all the thiophene-BODIPY derivatives were evaluated in toluene (Table 1), and PY-BOD, PH-BOD and MeO-BOD were shown to be excellent candidates for fluorescence imaging.

**\(^{1}O_2\) generation ability**

Subsequently, the singlet oxygen generation capability of PY-BOD, PH-BOD, MeO-BOD and DMA-BOD was assessed in air-saturated acetonitrile (ACN) under 560 nm irradiation. A commercial \(^{1}O_2\) probe, 1,3-diphenylisobenzofuran (DPBF), was used as an indicator, and rose bengal (RB, \(\Phi_{\Delta 5}^{*} = 0.54\) in ACN) was used as the reference. As shown in Fig. 2a and S17,† the absorbance at 410 nm of DPBF decreased gradually in the presence of the thiophene-BODIPY derivatives under continuous light irradiation. According to the linear relationship of the decay curves (Fig. 2b), the \(^{1}O_2\) quantum yields of PY-BOD, PH-BOD, MeO-BOD and DMA-BOD were calculated to be 0.52, 0.69, 0.85 and 0.04, respectively (Table 1). The strongest \(^{1}O_2\) generation ability of MeO-BOD among all the R-BODs suggested

![Scheme 1: Synthetic route to the thiophene-fused BODIPY derivatives.](image)

![Fig. 1: (a) Design of thiophene-fused BODIPY derivatives, (b) UV-Vis absorption and (c) emission spectra of PY-BOD, PH-BOD, MeO-BOD and DMA-BOD in toluene at c = 5.0 \(\mu\text{M}\) and \(\lambda_{\text{ex}} = 540\,\text{nm}\).](image)
the electron donating group (MeOPh−) in the thiophene-fused BODIPY derivatives played an important role in enhancing the \( ^1\text{O}_2 \) generation. Based on these results, we propose a plausible mechanism for MeO-BOD. The introduction of the donating group probably favors \( ^1\text{O}_2 \) generation first by increasing the formation of the charge transfer (CT) state via photoinduced charge transfer (PCT) (\( \text{BOD}^\text{Donor} \rightarrow \text{BO}\text{D}^\text{Donor} \)). The charge recombination of the CT state further triggers the production of \( \text{T}_1 \) of the thiophene-fused BODIPY derivatives (\( \text{BOD}^{\text{Donor}} \rightarrow \text{BO}\text{D}^\text{Donor} \)). Finally, \( ^1\text{O}_2 \) is generated by energy transfer from \( \text{T}_1 \) to molecular oxygen (\( \text{BO}\text{D}^\text{Donor} + \text{O}_2 \rightarrow \text{BO}\text{D}^\text{Donor} + ^1\text{O}_2 \)) (Scheme S11). Unfortunately, DMA-BOD showed almost no \( ^1\text{O}_2 \) generating ability in ACN, which implied that the electron-donating group could also lead to a PET process, which triggered the quenching of fluorescence and forbidding of non-radiative transitions such as ISC, thereby prohibiting the \( ^1\text{O}_2 \) generation. Thus, the \( ^1\text{O}_2 \) formation efficiency could be controlled by finely tuning the electronic properties of the meso-substituent on the thiophene-BODIPY platform, especially by the introduction of a suitable electron-donating group. Taken together, these results indicate that MeO-BOD has potential as a theranostic agent for cancer treatment.

**Cellular fluorescence imaging and subcellular colocalization**

Given the above inspiring results, we attempted to investigate the fluorescence (FL) imaging and PDT efficiency of MeO-BOD in living cells. To this end, MeO-BOD nanoparticles (NPs) were prepared by adding the DMSO stock solution of MeO-BOD into water. The transmission electron microscopy (TEM) image in Fig. 2c indicated that the MeO-BOD NPs had a regular spherical morphology with a diameter of approximately 72 nm. The size distribution of the nanoparticles was determined using dynamic light scattering (DLS), which showed that the average size was \( 68 \pm 7 \) nm (Fig. S18). These particles are suitably sized for passive targeting through the enhanced permeability and retention (EPR) effect. In addition, the evident decrease in absorbance and the broad absorption spectrum and the redshift of the fluorescence spectrum of MeO-BOD in deionized water (DW) suggested that the formation of MeO-BOD NPs might be due to J-aggregation (Fig. 2d and S19b†). The MeO-BOD NPs could be disassembled and the instinctive fluorescence peak of MeO-BOD could be restored in the presence of FBS (10%) in DW (Fig. S19†).

Then, we further explored the cellular uptake of MeO-BOD in HeLa cells by using confocal laser scanning microscopy. As illustrated in Fig. 3a, MeO-BOD could be rapidly internalized by living cells, and the FL images showed strong emission in the cell cytoplasm. Therefore, MeO-BOD could be employed as an imaging-guided PDT agent. Furthermore, to test the main organelle locations of MeO-BOD, we co-stained HeLa cells with MeO-BOD and commercial MitoTracker Green (MTG) or LysoTracker Green (LTG). The colocalization experiments indicated that MeO-BOD were mainly localized in mitochondria, as indicated by the high Pearson’s coefficient (0.98), instead of the lysosomes (Pearson’s coefficient 0.62) (Fig. 3b–e). In addition, the time-dependent colocalization fluorescence imaging of MeO-BOD with MTG in HeLa cells was performed. As shown in Fig. S20† MeO-BOD almost internalize into the mitochondria of HeLa cells after 1 h. Recent reports have demonstrated that the BODIPY dye itself could localize into the mitochondria of cells due to its low electron density character. The unmodified BODIPY platform has a low electron density (HOMO) character. Also, the incorporation of sulfur atoms into the π-conjugated skeleton of BODIPY perhaps led to more reduction of electron
density of the BODIPY core,\textsuperscript{46} which might facilitate the mitochondrial accumulation of MeO-BOD. Subcellular organelles are indispensable in maintaining cellular biological function.\textsuperscript{19} In particular, the generation of $^{1}\text{O}_2$ in mitochondria can induce direct dysfunction and trigger cell apoptosis. Thus, mitochondria-targeted theranostic agents could maximize cancer treatment efficiency.

**In vitro PDT efficacy evaluation**

To demonstrate the PDT efficacy of MeO-BOD in living cells, we first tested its cytotoxicity by the methyl thiazolyltetrazolium (MTT) assay in HeLa cells. As indicated in Fig. 4a, MeO-BOD had negligible cytotoxicity in the dark, revealing its excellent biocompatibility in vitro. Under 560 nm light irradiation (0.1 W cm\(^{-2}\), 5 and 10 min), the viability of HeLa cells gradually decreased with the increasing concentration of MeO-BOD, and the growth inhibition ratio reached ~88% even at a very low concentration of 0.15 µM (Fig. 4b). The half-maximal inhibitory concentration (IC\(_{50}\)) of MeO-BOD for HeLa cells was as low as 95 nM under 560 nm light irradiation (0.1 W cm\(^{-2}\), 10 min). The extremely low IC\(_{50}\) of MeO-BOD could be attributed to the high $^{1}\text{O}_2$ quantum yield and efficient mitochondria-specific ROS generation upon light irradiation. To clarify the cytotoxicity in the PDT process, we tracked the morphological variations of HeLa cells in the presence of MeO-BOD under 559 nm laser irradiation using confocal laser scanning microscopy. As described in Fig. S21,\textsuperscript{†} with increasing irradiation time (0–10 min), the morphology of the HeLa cells preincubated with MeO-BOD obviously changed; gradual thinning of the cell membrane and the formation of numerous blebs (red line) were observed.

In contrast, the cells not exposed to MeO-BOD exhibited no appreciable morphological changes under the same laser irradiation.

Additionally, to intuitively establish the PDT efficacy of MeO-BOD (0.5 and 1.0 µM), live/dead cell co-staining of HeLa cells with calcein AM (green, live cells) and PI (red, dead cells) was performed to visualize the cell viability (Fig. 4c). The control group showed strong green fluorescence for live cells, which also verified the low dark toxicity and good biocompatibility of MeO-BOD. In comparison, almost all the HeLa cells treated with MeO-BOD were killed, and intense red fluorescence was observed. All these results confirmed that MeO-BOD could be employed as a theranostic agent for cancer treatment in vitro.

**Apoptosis mechanism of MeO-BOD-mediated PDT**

Finally, to elucidate the potential therapeutic mechanism at the cellular level, we further evaluated the cellular $^{1}\text{O}_2$ generation capability of MeO-BOD by using 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA) as the $^{1}\text{O}_2$ indicator in HeLa cells (Fig. 5a). The HeLa cells treated with only DCFH-DA and only MTG showed almost no fluorescence, whereas the group pretreated with MeO-BOD before light irradiation and then incubated with DCFH-DA showed apparent green fluorescence from oxidized 2,7-dichlorofluorescein (DCF). These experimental results validated the $^{1}\text{O}_2$ generation ability of MeO-BOD in living cells. As mentioned above, the production of $^{1}\text{O}_2$ in mitochondria causes mitochondrial destruction, resulting in cell apoptosis. The reduction of mitochondrial membrane potential (MMP) is
Conclusions

We synthesized thiophene-fused BODIPY analogues with different functional groups at the meso-position of the BODIPY core as mitochondria-targeted theranostic agents for imaging and PDT. The varying R-substituents had little effect on the absorption and emission maxima. PY-BOD, PH-BOD and MeO-BOD exhibited moderate fluorescence quantum yields, while that of DMA-BOD was much lower due to a PET process. Among the analogues, MeO-BOD exhibited excellent dual functionality, both considerable $^{1}\text{O}_2$ generation ability and high brightness. The cell experiments manifested that MeO-BOD offered many advantages: good biocompatibility, mitochondria-specific fluorescence imaging, and a very low IC$_{50}$ value ($\approx 95$ nM). Therefore, MeO-BOD could be employed as an imaging-guided PDT agent for cancer treatment. In addition, the apoptosis mechanism of light-induced PDT might be a result of ROS-induced damage to mitochondria, which was demonstrated by detecting the changes of the mitochondrial membrane potential. Briefly, finely tuning the electronic structure of the substituent at the meso-site of a heavy-atom-free thiophene-fused BODIPY core is a promising strategy for developing highly efficient theranostic agents with minimal side effects to accomplish the integration of diagnosis and therapy.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was supported by grants from the National Research Foundation of Korea (NRF) funded by the Korean government (MSIP) (No. 2012R1A3A2048814 for J. Y.). Mass spectral data were obtained from the Korea Basic Science Institute (Daegu) on a Jeol JMS 700 high resolution mass spectrometer.

Notes and references

1. X. Li, S. Lee and J. Yoon, Chem. Soc. Rev., 2018, 47, 1174–1188.
2. P. Agostinis, K. Berg, K. A. Cengel, T. H. Foster, A. W. Girotti, S. O. Gollnick, S. M. Hahn, M. R. Hamblin, A. Juzeniene, D. Kessel, M. Korbelik, J. Moan, P. Mroz, D. Nowis, J. Piette, B. C. Wilson and J. Golab, Ca-Cancer J. Clin., 2011, 61, 250–281.
3. D. E. J. G. J. Dolmans, D. Fukumura and R. K. Jain, Nat. Rev. Cancer, 2003, 3, 380–387.
4. J. Zhang, L. Ning, J. Huang, C. Zhang and K. Pu, Chem. Sci., 2020, 11, 618–630.
5. J. Dai, Y. Li, Z. Long, R. Jiang, Z. Zhuang, Z. Wang, Z. Zhao, X. Lou, F. Xia and B. Z. Tang, ACS Nano, 2020, 14, 854–866.
6. Y. Yuan, C.-J. Zhang, M. Gao, R. Zhang, B. Z. Tang and B. Liu, Angew. Chem., Int. Ed., 2015, 54, 1780–1786.
7. Y. Cai, W. Si, W. Huang, P. Chen, J. Shao and X. Dong, Small, 2018, 14, 1704247.
8. R. Kumar, W. S. Shin, K. Sunwoo, W. Y. Kim, S. Koo, S. Bhuniya and J. S. Kim, Chem. Soc. Rev., 2015, 44, 6670–6683.
9. X. Yi, F. Wang, W. Qin, X. Yang and J. Yuan, Int. J. Nanomed., 2014, 9, 1347–1365.
10. X. Li, C. H. Kim, S. Lee, D. Lee, H.-M. Chung, G. Kim, S.-H. Heo, C. Kim, K.-S. Hong and J. Yoon, J. Am. Chem. Soc., 2017, 139, 10880–10886.
