Effect of *Ocimum Sanctum* Bio Compounds against csuE Gene Protein of *Acinetobacter baumannii*

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Authors’ contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

**Background:** *Acinetobacter baumannii* is a Gram-negative bacillus that is aerobic, pleomorphic and non-motile. Multi-drug resistance and biofilm formation contributes to the virulence and pathogenicity of the bacterium. Among many virulence factors, csuE is critical for initiation and assembly, showing much homology to type 1 and P pili. With much propensity of drug resistance, in recent years alternative medications have spurred renewed interest in targeting potent pathogens. *Ocimum sanctum*, also known as holy basil or tulsi possess various bio-active properties and can be used as alternative medicine to treat systemic ailments.

**Aim:** This study was aimed to analyze the drug-ligand interactions between csuE protein of *A. baumannii* and the bio-compounds from *O. sanctum* using in-silico docking analysis.

**Material and Methods:** csuE protein was retrieved and optimisation of protein was done. Ligands were selected and were assessed for drug likeness using molinspiration parameters. Further the compounds were subjected for docking analysis and the interacted molecules were visualized for binding energy and hydrogen bonds.

**Results:** Out of the 9 compounds of *Ocimum sanctum*, benzofuran showed good interaction with csuE protein of *Acinetobacter baumannii* with a least docking energy of -5.31Kcal/Mol.

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Conclusion: The present study recommends benzofuran as the potent candidate for novel drug design to treat the infections caused by *A. baumannii* upon further evaluations for its safety and immunological response.

Keywords: *A. baumannii*; *O. sanctum*; novel benzofuran; drug ligand; innovative interactions; environmental strains.

1. INTRODUCTION

*Acinetobacter baumannii* is a Gram-negative bacterium that is normally small, almost oval, and rod-shaped. It may be an opportunistic pathogen in humans, targeting patients with weakened immune systems, and it is becoming more common as a hospital-acquired infection [1]. These are non-motile but possess a typical motile behaviour through type IV pili, which are pole-like structures that can stretch and retract [2]. Organisms of the genus *Acinetobacter* are often thought to be widespread in nature inhabiting soil and water. *A. baumannii* is a member of the ACB complex (*A. baumannii*, *A. calcoaceticus*, and *Acinetobacter* genomic species 13TU). The unique species of ACB complex members are difficult to identify, and they are the most scientifically important members of the genus. *Acinetobacter baumannii* has also been classified as an ESKAPE pathogen (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* species), which is responsible for the majority of nosocomial infections [3]. Because of its apparent abrupt appearance in military treatment facilities during the Iraq War, *A. baumannii* is colloquially known as "Iraqibacter." [4–15] Extensive knowledge and research experience in the present research area by our team has been translated into high quality publications and has been utilized in the present study [4–15].

Multi-drug resistance and biofilm formation contribute to its virulence and pathogenicity of the bacterium. Among many virulence factors, csuE must be critical for initiation of assembly, showing much homology to type 1 and P pili. csuE is completely abrogated in the pilus assembly and folded into beta barrel domains. The angle between the domains gives the molecule an overall C like shape, bending over domains of csuE. csuE pili is elaborated from four protein subunits, csuA/csuB and csuE via archaic chaperone usher pathway. Archaic CU pillar constitute the foremost important family of CU systems and along with the choice of CU family form the ‘non classical’ branch of CU superfamily. csuE associated virulence is documented in many earlier reports portraying its importance in the pathogenesis of *A. baumannii* [16] It is now a challenging task to treat the patients infected with *A. baumannii* due to its severity of multi-drug resistance and biofilm formation.

In this line, alternative medication plays a vital role in the eradication of the pathogen. The aromatic perennial herb *Ocimum sanctum*, also known as holy basil or tulsi, belongs to the Lamiaceae family. It is indigenous to the Indian subcontinent and is widely grown in the Southeast Asian tropics. *Ocimum Sanctum* is responsible for its various medicinal properties and their effects at the molecular level need to be investigated in more detail [17]. Tulsi is cultivated for its natural oil, as well as for religious and herbal medicine purposes. It is widely used as a herbal tea, is commonly used in Ayurveda, and plays a role in Hinduism’s Vaishnava culture, where devotees practise worship using holy basil plants or leaves. Similar in-silico based studies have been referred to design the present study and the expertise as received from previous literatures were also incorporated for the same. With the advent of science and technology, the drug ligand interaction studies are possible with the wide computational platform of bioinformatics databases and tools. Our team has extensive knowledge and research experience that has translate into high quality publications [18–22]. The present investigation was thus aimed to target the csuE protein of *A. baumannii* using the bio-compounds from *O.sanctum*.

2. MATERIALS AND METHODS

2.1 Study Setting

This is an observational in-silico study done in the Department of Microbiology, Saveetha Dental College and Hospital.

2.2 csuE Retrieval and Optimisation of Protein

The crystal structure of csuE was obtained from RCSB protein data bank. The optimisation of
csuE, which is crystal structure, done by addition of hydrogen atoms. Kollman united atoms force field was used to assign electronic charges to the protein atoms which was done in AutoDock tool-1.5.6 and the RASMOL tool was used for the visualisation of three dimensional structure of csgA protein.

2.3 Optimisation and Ligand Preparation

The structures of the bioactive derivatives of A. baumannii were obtained from the chemsketch software. The generated 3d structures were then optimised. The selected ligands were then subjected to subsequent conversions by open label molecular converter to subsequent conversions by open label molecular converter program. They were then saved in PDB format. The selected ligands were further saved in a molecular file.

2.4 Molinspiration Assessment of the Molecular Properties of Selected Compounds

The counts of hydrogen bond acceptors and donors in relation to the membrane permeability and bio availability of the compounds, log P for partition coefficient, molecular weight of compounds of basic molecular descriptions were assessed with the help of molinspiration assessment program. The characteristics of absorption, distribution, metabolism and elimination of the selected bio compounds were further evaluated on the basis of “The Lipinski’s rule of five”.

2.5 Docking Stimulation

AUTODock tool was used for docking analysis to interpret the affinity between the bio compounds of A. baumannii against csuE protein.

2.6 Docking Visualisation

Using the discovery studio visualiser, the hydrogen bond interaction between the bio compounds of A. baumannii against csuE proteins were visualised. With further docking score assessments, binding affinities, molecular dynamics and energy simulations, the relative stabilities were evaluated.

3. RESULTS

The sequence of csuE from A. baumannii was retrieved from the uniport database and its sequence is Q6XBY2. The structure of csue was not available in the PDB database. Hence it was modeled using swiss model server using the template 6FJY-B chain. The modeled structure was found to be highly plausible as it had 100% sequence identity with that of the template. Moreover, the Ramachandran plot also showed 94% of residues in the disallowed region.

3.1 Structural Retrieval of csuE Protein from A. baumannii

FASTA sequence of csuE from A. baumannii was retrieved from UNIPORT database and its sequence ID was Q6XBY2. Using the swiss model server, the homology model was made with 6FJY-B chain as template. The model was highly plausible with 100% sequence identity with the template. Besides, the Ramachandran plot showed 94% of the residues in the disallowed region. The 3d structure of the csuE was visualised using RASMOL with the pink coloured denoting the alpha helix, yellow arrow denoting the beta sheets and white colour denoting the turns.

3.2 Molinspiration Assessment Towards Drug Likeness

Based on the calculation of the ion channel modulation, GPCR ligand, nuclear receptor ligand, kinase inhibitor, enzyme inhibition and protease inhibition, the bioactivity score prediction of essential compounds of A.indica against A.baumannii towards drug likeness was accessed and tabulated.

4. DISCUSSION

The potent virulence factor of A. baumannii is its ability to form biofilms in a major step process viz, attachment of bacteria to the surface, formation of micro colony, maturation of biofilms and compounds for the in vitro analysis. Selection of bioactive compounds from Ocimum sanctum was done based on the available earlier literature. The compounds are vital for transforming into nano-formulations as well due to their hydophilicity. With the aid of computational bioinformatics tools and databases, the drug ligand interactions were analyzed based on the pose and strength [23]. Drug likeness was highly promising from the molinspiration results with no violations for all the selected bio compounds except bis(2-propyl pentyl) phthalate ester.
Table 1. Showing the molinspiration results of the *O.sanctum* selected compounds

| Compounds                                      | M.wt  | Hydrogen Bond Donor | Hydrogen Bond Acceptor | miLogP | Rotatable bonds | nViolations | TPSA (Å) | Volume  |
|------------------------------------------------|-------|---------------------|------------------------|--------|-----------------|-------------|----------|---------|
| Estragole                                      | 148.21| 0                   | 1                      | 2.82   | 3               | 0           | 9.23     | 154.12  |
| Eugenol                                        | 164.20| 1                   | 2                      | 2.10   | 3               | 0           | 29.46    | 162.14  |
| Methyleugenol                                  | 18.47 | 0                   | 0                      | 2.41   | 0               | 0           | 18.47    | 179.67  |
| Benzofuran, 7-(2,4-dinitrophenoxy)-3-ethoxy2,3-dihydro-2,2-dimethyl | 374.35| 0                   | 9                      | 4.49   | 6               | 0           | 119.35   | 318.05  |
| Hexahydro-1,6-dimethyl-4-(1-methylethyl)-      | 220.36| 0                   | 1                      | 4.66   | 1               | 0           | 17.07    | 238.11  |
| Citral                                         | 152.24| 0                   | 1                      | 3.65   | 4               | 0           | 17.07    | 169.74  |
| Ceftazidime                                    | 546.59| 4                   | 13                     | –5.68  | 9               | 2           | 191.23   | 439.78  |

Table 2. Showing overall docking energies of the drug-ligand interactions

| CsU/E docking with compounds                    | Number of hydrogen bonds | Binding energy | Ligand efficiency | Intermolecular energy | vdW + Hbond + desolv Energy | Electrostatic energy | Torsional energy | Total internal Unbound |
|------------------------------------------------|--------------------------|----------------|-------------------|-----------------------|-----------------------------|----------------------|-------------------|---------------------|
| Estragole                                      | 1                        | -4.9           | -0.45             | -5.79                 | -5.74                       | -0.05                | 0.89              | -0.18               |
| Eugenol                                        | 2                        | -4.59          | -0.38             | -5.78                 | -5.65                       | -0.14                | 1.19              | -0.71               |
| Methyleugenol                                  | 2                        | -4.89          | -0.38             | -6.08                 | -6.00                       | -0.09                | 1.19              | -0.49               |
| Benzofuran, 7-(2,4-dinitrophenoxy)-3-ethoxy2,3-dihydro-2,2-dimethyl | 4                     | -5.31          | -0.2              | -7.1                  | -6.38                       | -0.72                | 1.79              | -0.79               |
| Hexahydro-1,6-dimethyl-4-(1-methylethyl)-      | 1                        | -5.73          | -0.36             | -6.03                 | -5.97                       | -0.06                | 0.3               | -0.28               |
| Citral                                         | 1                        | -4.1           | -0.37             | -5.29                 | -5.28                       | -0.01                | 1.19              | -0.37               |
| Ceftazidime                                    | 5                        | -5.35          | -0.14             | -8.63                 | -7.99                       | -0.65                | 3.28              | -2.41               |
Table 3. showing overall binding interactions among the csuE and *O.sanctum*

| KPSM docking with compounds | Hydrogen bonds interactions | van der Waals interactions | π-σ interactions/ π-π T-shaped interactions/ amide-π stacked interactions | alkyl/π-alkyl interactions | Other interactions |
|-----------------------------|-----------------------------|---------------------------|------------------------------------------------------------------------|---------------------------|-------------------|
| Estragole                   | ASN57                       | SER58, GLY198, GLY59, SER32, SER37 | -                                                                      | LEU200, LEU154, PHE56    | THR36 (carbon hydrogen bond) |
| Eugenol                     | ASN57, SER32                | GLY198, SER58, LEU197, GLY35, SER34 | -                                                                      | LEU154                    | THR36 SER37 (carbon hydrogen bond) |
| Methyleugenol               | ASN57, SER32                | SER61, SER32, GLY59, SER196, LEU197, SER58, GLY198, SER37, GLY35, THR36 | -                                                                      | LEU154                    | -                 |
| Benzofuran, 7-(2,4-dinitrophenoxo)-3-ethoxy2,3-dihydro-2,2-dimethyl Hexahydro-1,6-dimethyl-4-(1-methylethyl)- | ALA139 (2) ARG136 SER53 | GLU51, SER53, ASN52, LEU137, GLY50, GLU48 | ASN49 | PRO138 ARG136 ASN49 | - |
| Citral                      | THR36                       | LEU197, GLY59, ASN57, GLY35, SER37, GLY198, SER199, SER32 | -                                                                      | LEU147 (2) VAL333 ILE274 PHE312 VAL335(2) VAL233 LEU200(2) PHE56 LEU154 | - |
| Ceftazidime                 | ARG236 ILE210 (2) ALA212 (2) | GLU48 GLN213 | LEU211 | ALA212 ILE210 | - |
Fig. 1. Structure Prediction of CsuE by Homology modeling using Swissmodel server

Fig. 2. 3D structures of the *O.sanctum* compounds selected for the study
The drug molecules that have low molecular weight (<500) are transported, diffuse band absorbed without difficulty in comparison to large molecules. Molecular weight is one of the critical aspects in corrective drug action; if it seems to increase correspondingly, which affects the efficiency of the drug. Some hydrogen bond donors and a number of hydrogen bond acceptors are natural compounds as established in Lipinski’s limit range 1-16 and 1-11 that come out to be less than 10 and 5 [24–28].

The molecules are more flexible when the number of rotatable bonds increases and more susceptible to proficient interactions with a precise binding pocket. Selected five commonly used aromatic profiles of essential oil of *ocimum sanctum* were estragole, eugenol, methyl eugenol, benzofuran, hexahydro-1,6 -dimethyl 1-4, citral, ceftazidime. For benzofuran, there are 4 compounds and the docking energy (Kcal/Mol) was -5.31. For citral, there is only one compound and the docking energy was -4.9. For eugenol the number of compounds were two and the docking energy was -4.59 Kcal/Mol. For Methyl eugenol, there were two compounds and docking energy was -4.89. For ceftazidime, there are 5 interactions and docking energy was -5.35. Though ceftazidime is an antibiotic and showed least binding energy, it had violated two out of 5 Lipinski’s rules. The findings of the present study suggest benzofuran as the best candidate against *A. baumannii* on csuE gene as it has -5.31 Kcal/Mol docking energy and has 4 compounds. The results also correlate with our earlier studies [7]. The limitation of the study is that it was done as an computational observational study and the future prospects are set to experimentally evaluate the antimicrobial effects using in-vitro and in-vivo studies [29,1,30].

5. CONCLUSION

The present study was undertaken as a computational approach to evaluate the drug-ligand interactions of the bio-active compounds from *O. sanctum* with csuE of *A.baumannii*. Out of the 5 compounds selected for the study, benzofuran is considered as the best compound with a docking energy of -5.31 Kcal/mol. However, further experimental validations have to be made in-vitro to assess the safety and efficacy of the compounds in the development of novel drugs and to curtail *A.baumannii* associated infections.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

CONSENT

It is not applicable.

ETHICAL APPROVAL

Institution approval for research was obtained (IHEC/SDC/UG-1942/21/155)

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Iswarya Jaisankar A, Smiline Girija AS, Gunasekaran S, Vijayashree Priyadharsini J. Molecular characterisation of csgA gene among ESBL strains of *A. baumannii* and targeting with essential oil compounds from *Azadirachta indica*. Journal of King Saud University - Science 2020; 32:3380–7.
2. Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species. Archives of Oral Biology 2018;94:93–8. Available:https://doi.org/10.1016/j.archoralbio.2018.07.001.
3. As SG, J VP, Paramasivam A. Prevalence of Acb and non-Acb complex in elderly population with urinary tract infection (UTI). Acta Clinica Belgica. 2021;76:106–12. Available:https://doi.org/10.1080/17843286.2019.1669274.
4. Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species. Archives of Oral Biology 2018;94:93–8.
5. Vijayashree Priyadharsini J. In silico validation of the non-antibiotic drugs aceterminophen and ibuprofen as antibacterial agents against red complex pathogens. J Periodontol 2019;90:1441–8.
6. Paramasivam A, Vijayashree Priyadharsini J, Raghunandhakumar S. N6-adenosine methyltransferase (m6A): a promising new molecular target in hypertension and cardiovascular diseases. Hypertens Res 2020;43:153–4.
7. Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. An insight into the emergence of Acinetobacter baumannii as an oro-dental pathogen and its drug resistance gene profile – An in silico approach. Heliyon 2018;4:e01051. Available:https://doi.org/10.1016/j.heliyon.2018.e01051.
8. Paramasivam A, Vijayashree Priyadharsini J. Novel insights into m6A modification in circular RNA and implications for immunity. Cell Mol Immunol 2020;17:668–9.
9. Paramasivam A, Priyadharsini JV, Raghunandhakumar S. Implications of m6A modification in autoimmune disorders. Cell Mol Immunol 2020;17:550–1.
10. Girija ASS, Shankar EM, Larsson M. Could SARS-CoV-2-Induced Hyperinflammation Magnify the Severity of Coronavirus Disease (CoVID-19) Leading to Acute Respiratory Distress Syndrome? Front Immunol 2020;11:1206.
11. Jayaseelan VP, Arumugam P. Exosomal microRNAs as a promising theragnostic tool for essential hypertension. Hypertens Res 2020;43:74–5.
12. Ushanthika T, Smiline Girija AS, Paramasivam A, Priyadharsini JV. An in silico approach towards identification of virulence factors in red complex pathogens targeted by reserpine. Nat Prod Res 2021;35:1893–8.
13. Ramalingam AK, Selvi SGA, Jayaseelan VP. Targeting prolyl tripeptidyl peptidase from Porphyromonas gingivalis with the bioactive compounds from Rosmarinus officinalis. Asian Biomed 2019;13:197–203.
14. Kumar SP, Girija ASS, Priyadharsini JV. Targeting NM23-H1-mediated inhibition of tumour metastasis in viral hepatitis with bioactive compounds from Ganoderma lucidum: A computational study. Pharmaceutical-Sciences 2020;82. Available:https://doi.org/10.36468/pharmaceutical-sciences.650.
15. Mathivadani V, Smiline AS, Priyadharsini JV. Targeting Epstein-Barr virus nuclear antigen 1 (EBNA-1) with Murraya koengii bio-compounds: An in-silico approach. Acta Virol 2020;64:93–9.
16. Girija SA, Priyadharsini JV, Paramasivam A. Prevalence of carbapenem-hydrolyzing OXA-type β-lactamases among Acinetobacter baumannii in patients with severe urinary tract infection. Acta Microbiologica et Immunologica Hungarica 2019;1–7. Available:https://doi.org/10.1556/030.66.20 19.030.
17. As SG, S SGA, J VP. CLSI based antibiogram profile and the detection of MDR and XDR strains of Acinetobacter baumannii isolated from urine samples. Medical Journal of The Islamic Republic of Iran 2019. https://doi.org/10.47176/mjiri.33.3.
18. PradeepKumar AR, Shemes H, Jothilatha S, Vijayabharathi R, Jayalakshmi S, Kishen A. Diagnosis of Vertical Root Fractures in Restored Endodontically Treated Teeth: A Time-dependent Retrospective Cohort Study. J Endod 2016;42:1175–80.
19. Dhinesh B, Isaac Joshua Ramesh Lalvani J, Parthasarathy M, Annamalai K. An assessment on performance, emission and combustion characteristics of single cylinder diesel engine powered by Cymbopogon flexuosus biofuel. Energy Convers Manage 2016;117:466–74.
20. Lekha L, Kanmani Raja K, Rajagopal G, Easwaramoorthy D. Schiff base complexes of rare earth metal ions: Synthesis, characterization and catalytic activity for the oxidation of aniline and substituted anilines. J Organomet Chem 2014;753:72–80.
21. Soh CL, Narayanan V. Quality of life assessment in patients with dentofacial deformity undergoing orthognathic surgery—A systematic review. Int J Oral Maxillofac Surg 2013;42:974–80.
22. Krishnan V, Lakshmi T. Bioglass: A novel biocompatible innovation. J Adv Pharm Technol Res 2013;4:78–83.
23. Girija ASS, Smiline Girija AS, Vijayashree Priyadharsini J, Paramasivam A. Plasmid-encoded resistance to
trimethoprim/sulfamethoxazole mediated by dfrA1, dfrA5, sul1 and sul2 among Acinetobacter baumannii isolated from urine samples of patients with severe urinary tract infection. Journal of Global Antimicrobial Resistance 2019;17:145–6. Available:https://doi.org/10.1016/j.jgar.2019.04.001.

24. Samuel SR, Kuduruthullah S, Khair AMB, Shayeb MA, Elkaseh A, Varma SR. Dental pain, parental SARS-CoV-2 fear and distress on quality of life of 2 to 6 year-old children during COVID-19. Int J Paediatr Dent 2021;31:436–41.

25. Samuel SR. Can 5-year-olds sensibly self-report the impact of developmental enamel defects on their quality of life? Int J Paediatr Dent 2021;31:285–6.

26. Barma MD, Muthupandiyam I, Samuel SR, Amaechi BT. Inhibition of Streptococcus mutans, antioxidant property and cytotoxicity of novel nano-zinc oxide varnish. Arch Oral Biol 2021;126:105132.

27. Teja KV, Ramesh S. Is a filled lateral canal - A sign of superiority? J Dent Sci 2020;15:562–3.

28. Reddy P, Krithikadatta J, Srinivasan V, Raghu S, Velumurugan N. Dental Caries Profile and Associated Risk Factors Among Adolescent School Children in an Urban South-Indian City. Oral Health Prev Dent 2020;18:379–86.

29. Jayaseelan VP, Paramasivam A. Emerging role of NET inhibitors in cardiovascular diseases. Hypertens Res 2020;43:1459–61.

30. Girija AS. Fox3 (+) CD25 (+) CD4 (+) T-regulatory cells may transform the nCoVs′ final destiny to CNS! COMMENT 2021.