In general, it was observed that amid prodrugs of NSAIDs with sulfanilamide showed more analgesic, antiinflammatory activity and less ulcerogenic potential than their corresponding parent NSAIDs. This may be because sulfanilamide is a prototype of sulfonamides and has sulfonamide group which increases COX⁻2 selectivity and thus acting synergistically. Moreover it also possesses carbonic anhydrase inhibitory activity, which may have shown inhibitory effect on gastric carbonic anhydrase enzyme thus, limiting the acid secretion. Hence, as proposed the prodrugs with greater potency and reduced gastric toxicity than the corresponding parent NSAIDs were synthesized. However, the complete elimination of ulcer formation cannot be avoided, since the prodrug conversion avoids the gastric irritation due to direct contact of acidic group with the gastric mucosa, while the ulcer formation due to inhibition of cytoprotective prostaglandin synthesis is inevitable.

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**Chronopharmacokinetics of Puerarin in Diabetic Rats**

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**Zhang, et al.: Chronopharmacokinetics of Puerarin**

Puerarin injection has been widely used for clinic treatment of diabetes recently. To assess the relationship between the administration time of puerarin and the blood concentration of puerarin as well as its pharmacokinetic parameters, the diabetic rat model was used in current study. The rats were randomly divided into morning and evening groups according to the administration time. After the puerarin injection, blood glucose was tested in order to know whether the efficiency of puerarin was influenced by its concentration and pharmacokinetic parameters. Our results show that the average concentration of puerarin in the evening group is significantly higher than that in the morning group. The numbers of $t_{1/2\alpha}$, $t_{1/2\beta}$, CL and AUC$_{(0-\infty)}$ are significantly different between the morning and evening groups. The blood glucose level in the evening group was lower than that in the morning group. The speed of its onset is higher and the blood glucose level declines much more significantly in the evening group. These findings suggest that the concentration and pharmacokinetic parameters of puerarin affect its efficiency in diabetic rats. Therefore, it might be better to give puerarin in evening than in the morning for the mellitus treatment.

Key words: Chronopharmacokinetics, puerarin injection, blood glucose, variations, diabetic rats

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Puerarin (7-hydroxy-3-(4-hydroxyphenyl)-1-benzopyran-4-one 8-(β-D-glucopyranoside)) is an abundant isoflavone glycoside extracted from the traditional Chinese medicine Puerariae Radix (PR, root of the Pueraria plant). It has been proved to effectively treat various diseases including cardiovascular, neurological and hyperglycemic disorders\cite{1-3}. Its effects on diabetes have been supported by numerous medical studies\cite{1,2}. Traditional Chinese medicine injection is an innovative and quick-acting formulation. A number of these injections have been played very important roles in clinical treatment of diabetes. The puerarin injection is a new formulation of puerarin. Recently it has been extensively used for clinic treatment of diabetes and resulting in a beneficial response in the treated patients.

Usually, the symptoms of diabetes occur in the morning and it has an obvious rhythmic variation. As such, the blood glucose level of diabetic patients exhibits a peak value in the morning\cite{4}. The pathphysiology of this kind disease not only shows biological rhythms, but also impacts the pharmacokinetics. The pharmacokinetics may vary from absorption, distribution, metabolism and elimination of medicine applied. It had been reported that the pharmacokinetics of puerarin were influenced by functional dyspepsia of the disease\cite{5}. The pharmacokinetics may influence the pharmacodynamics and vice versa. Both of them might be responsible for the time-dependent variations in the efficiency of puerarin treatment. Therefore, we speculate that the pharmacokinetics and pharmacodynamics of puerarin injection may be influenced by the rhythms of diabetes.

The first aim of our studies is to verify whether the time of puerarin administration according to the biological rhythms affect the pharmacokinetics of puerarin in diabetic rats. The second aim is to determine whether the efficiency of puerarin is influenced by its concentration and pharmacokinetic parameters by testing the blood glucose levels. Therefore, the appropriate administration time may be determined to obtain the optimal pharmacotherapy of puerarin.

The reference standard of puerarin and daidzein (internal standard) were purchased from the National Institute for the Control of Pharmaceutical and Biological Products, Beijing, China. Puerarin injection dosage was purchased from Baoji Jinsen Pharmaceutical Co., Ltd., Baoji, China. The glucose oxidase assay kit was supplied by Changchun Huili Biotech Co., Ltd., Jilin Province, China. Other chemicals were of analytical or HPLC grade and commercially available ultrapure water was used.

Male clean-grade, diabetic Sprague-Dawley rats (300-350 g) were obtained from the Laboratory Animal Institute of Heilongjiang University of Chinese Medicine, Harbin, China. These rats were housed in a room with 12-h light/dark cycles and fasted for 12 h before experiments. All animal experiments were conducted with the approval of the Animal Care and Use committee of Heilongjiang University of Chinese Medicine.

The rats were randomly divided into two groups and they were administrated with puerarin injection via vein. We chose 7:00 a.m. and 7:00 p.m. as the administration time to mimic the biological rhymes. Blood samples (500 μl) were collected into heparinized tubes via the orbital venous plexus before and after administration at 3, 5, 10, 20, 40, 50, 60, 80, and 100 min.

The diabetic rats in experimental groups were administrated with puerarin injection once a day for 5 consecutive days in the morning or evening. The blood samples were collected every day for blood glucose tests. These blood samples were centrifuged at 3000 rpm for 5 min to separate the plasma fraction. After that, the blood glucose levels were tested by automatic chemistry analyzer (Glamour 2000).

To isolate puerarin from plasma, liquid phase extraction was used. In our experiments, the biological agents were extracted via precipitating protein by acetonitrile. Then we removed non-polar interfering impurities. Each blood sample was transferred right away to a heparinized microtube and centrifuged at 3000 rpm for 5 min. one hundred and fifty microliters of the separated plasma was spiked with the internal standard (daidzein) and 150 μl acetonitrile. After that the mixture was vortex-mixed for 1 min and centrifuged at 3000 rpm for 5 min. Two hundred microliters of the supernatant was collected and then centrifuged at 12000 rpm for 5 min. Then 20 μl of the supernatant was injected into HPLC system for analysis.
The concentration of puerarin in biological samples was determined by HPLC (Shimadzu LC-2010A). Chromatographic experiment was performed on a Diamonsil C\textsubscript{18} reversed-phase column (150×4.6 mm, 5 μm). The mobile phase was consisted of water and acetonitrile. They were filtered through a 0.45-μm millipore filter and degassed prior to use. A gradient elution program was used, which is the proportion of acetonitrile increased from 5 to 55%, giving a total run time 22 min. We chose 250 nm as our experimental wavelength and set 0.8 ml/min as the flow rate with the column temperature set at 30°. The dosage volume was 20 μl each time.

Data from these samples were used to make concentration-time profiles. The pharmacokinetic parameters were calculated by the DAS 2.0. Compartment model was discriminated according to the method of Akaike’s information criterion (AIC). All results were expressed as mean±SD. SPSS software was used to assess statistical significance and a $P<0.05$ was considered to be statistically significant.

Fig. 1 shows the curves of the mean value of plasma puerarin after the intravenous administration to the rats in the morning or evening. The plasma concentration of puerarin versus time data in diabetic rat best fits a two-compartment open model for both the morning and the evening group. Although the plasma puerarin in both groups declines in a bi-exponential fashion with time, we do find that the average plasma concentration in evening group is higher than that in morning group, most of them showing significant differences.

Pharmacokinetic parameters of the blood puerarin in the morning or evening group are shown in Table 1 with statistical analysis of the comparisons. After the administration of puerarin in the evening, we find that $V_c$ is larger and CL smaller, accompanied by a lower $t_{1/2\alpha}$ and a higher $t_{1/2\beta}$. It means that puerarin is distributed faster and eliminated slower in diabetic rats of evening group. However, $t_{1/2}$ does not show significant change, and that could be the reason for the higher plasma concentrations in evening group rats. And we also find that the AUC\textsubscript{(0-∞)} is larger than that in the morning group.

The blood glucose levels of the rats in these two groups before and after puerarin administration are shown in Table 2. The results show that the puerarin injection could significantly reduce blood glucose level in diabetic rats. The blood glucose levels in the evening group are all higher than that in the morning group. It has been reported that the blood glucose level for healthy male SD rat was less than 7.0 mmol/l\cite{6}. In our case, the blood glucose level decreased to normal on the third day in the evening after puerarin administration, while the fourth day in the morning, the onset of effect was earlier and the blood glucose levels declined more significantly in the evening group.

The average concentration of plasma puerarin in the evening group is higher than that in the morning group. There is a significant difference of plasma puerarin between these two groups. We get the same observation for some pharmacokinetic parameters, such as $t_{1/2\alpha}$, $t_{1/2\beta}$, AUC\textsubscript{(0-∞)} and CL. From our studies, we found that AUC\textsubscript{(0-∞)} and $t_{1/2\alpha}$ are the main kinetic

| Parameters        | Morning        | Evening        |
|-------------------|----------------|----------------|
| $t_{1/2\alpha}$ (min) | 7.805±0.431    | 4.898±0.905*   |
| $t_{1/2\beta}$ (min) | 26.577±0.899   | 30.126±1.671*  |
| AUC\textsubscript{(0-∞)} (μg·min/ml) | 4981.950±114.012 | 7401.935±214.582* |
| $V_c$ (ml/kg)      | 0.064±0.014    | 0.072±0.008    |
| CL (ml/min)        | 0.005±0.0001   | 0.004±0.0001*  |
| $t_{1/2}$ (min)    | 12.070±2.364   | 10.212±0.723   |

Each value represents the mean±S.D for $n=6$, *Pharmcokinetic parameters were significantly different ($P<0.05$) from morning group.
parameters influenced by the administration time of puerarin injection ($P<0.01$). $t_{1/2a}$ is lower, and $t_{1/2b}$ is higher after injection of puerarin in the evening, which means that puerarin is distributed more rapidly and eliminated more slowly. However, $t_{1/2a}$ is not significantly changed, resulting in higher plasma concentrations in diabetic rats, so the AUC$_{(0-\infty)}$ is higher than the morning group. Our studies show that there are significant differences of the puerarin pharmacokinetics in diabetic rats between the morning and evening groups.

We also found that the concentration of puerarin in the evening group is higher and puerarin is distributed more rapidly and eliminated more slowly. Meanwhile, the pharmacodynamic studies indicate that the speed of onset is more rapid and the blood glucose levels decrease more significantly in the evening group. It suggests that puerarin injection as a treatment of diabetes might play better role in the evening than in the morning. The results suggest that there is a relationship between the rhythms and efficiency of the puerarin.

Circadian changes have already been found in cytochrome P450 level as well as in the activities of some medical metabolism enzymes, such as phase I and II[7-9]. This fact could partially explain our observation of variations in the different groups. On the other hand, dose- and time-dependent variation in medicine distribution and elimination cannot be explained only by circadian changes of enzyme activities, for circadian rhythmicity of blood flow might play a role as well. Some studies on rats showed that blood flow to kidney, liver and intestine gets maximal level during the active phase and goes down to minimal level in the rest phase[10]. We found that the medicine distribution is faster at night during which time the corresponding medical enzymes are at active phase. The factors influencing the distribution could be the rhythms of blood flow to the organs, the binding ability of medicine to plasma proteins and or the membrane permeability to the medicine. Therefore, the circadian variations of plasma protein binding, medicine distribution and elimination, including rhythms of blood flow to the organs and metabolism enzymes (CYP450) might be the mechanisms of the circadian variations in pharmacokinetics of puerarin. However, further studies on animals and human need to be done to determine the mechanisms of puerarin pharmacokinetics.

Our studies show that the average plasma concentration of puerarin in the evening group is higher than that in the morning group. Puerarin is distributed more rapidly and eliminated more slowly in the evening. These indicate that the efficiency of puerarin injection in the evening is better than in the morning. Therefore, puerarin injection may be better to administrate in the evening than in the morning for the treatment of diabetes. However, more works on its efficiency and toxicity should be done so that security treatment of puerarin injection could be offered to the patients in need. Meanwhile, the influence of administration time on pharmacodynamics and clinical applications of puerarin injection remains to be further studied.

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Nonisothermal Kinetics Analysis of the Dehydration of Ziprasidone Hydrochloride Monohydrate by Thermogravimetry

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In the current work the kinetics of dehydration of ziprasidone hydrochloride monohydrate was studied by nonisothermal thermogravimetry. Ziprasidone hydrochloride monohydrate was heated from 30 to 150° with a heating rate of 5° per min under nitrogen gas atmosphere and weight loss data were collected. Powder X-ray difraction was used to characterize the solid before and after dehydration. The well accepted Coats-Redfern model fitting approach was applied to the thermogravimetry data for the kinetic analysis. Thirteen solid state reaction models were studied; among them one-dimensional diffusion model was found to be the best fit model for this reaction with an excellent correlation 0.9994. The Arrhenius parameters, activation energy, and pre-exponential factor were determined, the values were found to be 28 k.cal/mol and $9.53 \times 10^{13}$ sec$^{-1}$, respectively.

Key words: Model fitting, kinetics, dehydration, thermogravimetry, ziprasidone hydrochloride

Ziprasidone hydrochloride (ZH), chemically 5-[2-[4-(1,2-benzisothiazol-3-yl)-1-piperazinyl]ethyl]-6-chloro-1,3-dihydro-2H-indol-2-one, monohydrochloride, represented in fig. 1, is a typical antipsychotic drug used for the treatment of schizophrenia, a mental disorder. It is commercially available as Geodon® capsules [1]. Anhydrous and monohydrate forms of ZH are known from literature [2].

In view of increasing regulatory concern related to solid form of the drug substance in the recent days, it is highly important to understand the physical stability of different solid forms of a drug substance and their inter-convertibility [3,4]. Understanding the kinetics of solid state reactions in relation to phase transformations of drug substance leaves an opportunity to optimize the operational conditions so as to prevent and have control over the changes in solid form, during manufacturing process and shelf life of the drug product. In the present work dehydration of ZH monohydrate to ZH anhydrous form, as represented in Eqn. (1) was studied by thermogravimetry (TG). ZH.H$_2$O→ZH+$\text{H}_2\text{O}$. (1)

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Fig. 1: Molecular structure of ziprasidone hydrochloride monohydrate.