Long Term Cultivation Effect on Soil Health Inside Poly Houses Under Sub-Tropical Condition of North East India

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Protected farming is becoming increasingly popular in different developed and developing countries of the world due to its multifaceted advantages. Study was carried out to evaluate the long term cultivation effect on soil health under poly houses of varying age groups. It was observed that after 3-5 years of cultivation, the soils under the poly houses were deteriorated due to formation of soil acidity, nutrient imbalance and reduction in microbial diversity and enzyme activity. Key soil enzymes, microbial biomass carbon and microbial population were assessed to understand the effect of poly house cultivation on soil health. pH, organic carbon content and bulk density of poly house soil of varying age groups varied significantly with respect to open field soil. Significantly lower cation exchange capacity and total exchangeable bases were recorded in poly house of >10 years age irrespective of soil depth and seasons. The available N, P₂O₅, K₂O in soils showed an increasing trend up to 3-5 years of poly house age, thereafter decreased significantly. Irrespective of seasons and soil depth, the bacterial and fungal population showed a decreasing trend with advancement of poly house age. The microbial biomass carbon, dehydrogenase activity, phosphononoesterase activity and Flourescent di acetate hydrolysis activity in soils showed a declining trend with increasing poly house age. Study indicated that the soils under poly houses required rejuvenation particularly after 6-8 years for improvement of physical, chemical and biological properties so as sustain soil health as well as crop productivity.

Keywords
Poly house, age groups, soil health, enzymatic activity, soil deterioration

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Introduction
Protected cultivation is gaining popularity across the world due to its multifaceted advantages like higher productivity and profitability, wider window of sowing or planting times, climate smart, safe and quality produce and enhanced resource use efficiency. Cultivation of many high value crops including horticultural crops, vegetables etc. in off season are economically profitable for producers. Crops grown under the poly houses are safer because of less damage by chemical residue, mechanical damage and are
of higher qualities (Chang and Liao, 1989). Besides these, cultivation in poly house protects crops from rain, hailstorm, snow and wind, improves seed bed condition, better seed germination, higher yield and quality (Antill, 1988).

Long term cultivation in poly house leads to deterioration of soil physical, chemical and biological properties which leads to the reduction in crop productivity. Micro environment such as temperature, moisture, aeration inside poly house together with tillage and fertilization cause the soil compaction, acidification, nutrient accumulation and imbalance in the soil (Li et al., 1996).

Continuous poly house cultivation and management can cause reduction in the species diversity of the soil biota and may markedly change the microbial biomass, dehydrogenase activity and basal respiration was (Lee et al., 2004).

Further, low light intensity inside the poly house is known to increase various soils borne pathogens. Increased temperature under polyhouse (2-3°C in summer and 4-5°C in winter) leads to substantial reduction of organic matter emphasizing the need for use of organic manures and crop residues for maximizing profitability and maintaining environment quality (Sen et al., 1984).

Further in winter, all sides of poly house are covered to keep the temperature high inside restricting the free flow of air that might reduce the activity of beneficial organisms. All these conditions affect the soil quality directly and indirectly affect the crops grown under the poly house.

In cognizance with the above, the present study was undertaken to evaluate the long term cultivation effect inside poly house of different age group on physico-chemical and biological properties of soil.

**Materials and Methods**

The present study was conducted in the Horticultural Research Farm of Assam Agricultural University, Jorhat, Assam, during 2013. The farm is located between 26°44'N latitude and 94°12'longitude, at an elevation of 91.0 m above mean sea level. The climate of the area is humid subtropical characterized by hot and wet summer and dry and cool winter.

The data on the mean maximum and minimum temperature, relative humidity at morning and evening, total rainfall and sunshine hours inside the poly houses are presented in Fig.1. The experiment was laid out in Completely Randomized Design with 5 treatments and 4 replications. The vegetable crops were cultivated inside the poly houses round the year. Poly houses of 4 age groups (0-2 yrs., 3-5 yrs., 6-8yrs. and >10yrs.) and four numbers of poly houses of each age group were selected as treatments and replications respectively. Non-poly house soils were taken as control from adjacent open areas of poly houses. Soil samples were collected from five different spots from each of the poly houses at two depths (0-15cm and 15-30cm) in winter and summer seasons.

**Soil physico-chemical characteristics**

The physico-chemical properties of soil was determined by following standard protocols i.e. clod method for bulk density (Blake, 1986), international pipette method for soil texture (Piper, 1966), Glass-electrode pH meter method for soil reaction (1: 2.5) (Jackson, 1973), Walkley and Black method for organic carbon (Jackson, 1973), centrifuge method for cation exchange capacity (Jackson, 1973), Alkaline permanganate
method for available N (Subbiah and Asija, 1956), Bray’s method for available P (Jackson, 1973), flame photometric method for available K (Jackson, 1973).

**Soil biological characteristics**

**Soil microbial population**

The classical serial dilution technique was used for enumeration of microbial population (bacterial and fungal) by spread plate technique on nutrient agar media and rose bengal agar media, respectively. Rhizosphere soil sample of 1g was suspended in 9 ml water blank, followed by serially diluted up to \(10^3\), \(10^4\) and \(10^5\) fungi and bacteria respectively. Aliquots of 100μl of \(10^3\), \(10^4\) and \(10^5\) dilutions were spread over the solidified media in triplicates and plates were incubated at 30±2°C for 2-5 days. The microbial numbers were estimated as colony forming unit (CFU) per gram soil on dry weight basis.

**Enzyme activities**

FDA hydrolysis activities were carried out following the method described by Adam and Duncan (2001). Soil was incubated with the substrate, FDA, at 25°C for 1 hour. The amount of fluoresce formed was determined colorimetrically (Nano Drop 1000 spectrophotometer) following extraction with an organic solvent mixture (2:1 chloroform: methanol). Using the calibration curve the mass of fluoresce in produced in each assay from the corresponding optical density (OD at 490nm) value was calculated on dry weight basis. Fluorescent diacetate hydrolysis activity was expressed as μg fluorescent per gram dry soil per hour.

Dehydrogenase activities were determined by the reduction of triphenyl tetrazolium chloride (TTC) to triphenyl formazan (TPF) as described by Casida et al., (1968). Fresh soil (1g) treated with 1 ml of 3% TTC, and then incubated at 28 °C for 24 hours. To account for any abiotic TTC reductions, sterile controls consisted of autoclaved soil (121°C, 20 minutes. on three consecutive days) were used. Spectrophotometer blanks for both autoclaved and non-autoclaved treatments consisted of soil and TTC replaced with millipore water. Controls and blanks treated like samples. The optical density at 485 nm was compared to that of triphenyl formazan standards. Dehydrogenase activity was expressed on dry weight as μg TPF per gram dry soil per 24hours.

Phosphomonoesterase activities involve the use of an artificial substrate, p-nitrophenyl phosphate (p-NPP). The product of Phosphomonoesterase activity, p-nitrophenol, a chromophore under alkaline conditions were detected colorimetrically following the method of Tabatabai and Bremner (1969) and expressed as μg p-nitrophenol per gram dry soil per hour.

**Microbial biomass carbon**

Soil microbial biomass carbon was determined by chloroform fumigation extraction technique following the method of Vance et al., (1987). Fresh soil (25 g) fumigated with ethanol free chloroform at 25 °C for 24hours. After fumigation, chloroform vapours were removed by repeated evacuation. The soil samples were then extracted with 100 ml 0.5M K₂SO₄ (1:4 soil K₂SO₄). Controls were prepared by extracting soils without fumigation. The soil suspension was filtered through Whatman No 42 filter paper. Total organic carbon content in the soil extract was estimated with dichromate (66.7mM) digestion method (Walkley and Black, 1934). Microbial biomass carbon was calculated from the differences in extractable organic carbon between the fumigated and
non-fumigated soil and expressed as μg per gram dry soil.

The analysis of data obtained was carried out following the statistical method of Completely Randomized Block Design (Panse and Sukhatme, 1967) for statistical significance the difference between the treatment means was tested with appropriate critical difference value at 1% and 5% level of probability. Simple correlation coefficient between soil physico-chemical properties and enzymatic activities were analyzed to establish possible statistical relationship (Chandel, 2004).

Results and Discussion

Physico-chemical characteristics

The significant variation in soil bulk density was recorded inside the poly houses compared to open field condition. Among the different age of the poly houses, highest bulk density was observed in > 10 years age group followed by 6-8 years, 3-5 years and 0-2 years in both surface (0-15 cm) and sub-surface soil (15-30 cm) irrespective of seasons (Fig. 2.1 and 2.2). However, soil bulk density was higher in winter compared to summer in both surface and sub-surface soil. The result is in accordance with that of Patil and Bhagat (2014).

Soils pH ranged from 4.30-5.18 inside the poly houses and in open field irrespective of the season. However the pH of the open field was lower than the poly house soil. Among the different age group, highest soil pH was recorded in 3-5 years age group and lowest was recorded in > 10 years age group (Table 1). Contribution of relatively higher soil pH in middle age group poly houses might be due to the application of fertilizer and organic manures resulting in increased the retention of exchangeable bases and the cation exchange capacity of the soil (Yaduvanshi et al., 1985). Decreased pH in soils of higher age poly houses might be due to excessive cultivation and irrigation that reduced the soil pH.

Cation exchange capacity (CEC) of soils under poly houses and open field condition was mainly influenced by organic matter and clay content of the soil (Table 1). Result revealed higher CEC in surface soils as compared to sub surface soils. Similarly, CEC was higher during summer season compared to winter season. A higher CEC in summer season soils over winter season soils might be due to the faster decomposition of organic matter in summer season resulting in more addition of organic sources to the soil (Norman et al., 2000). Further, the result revealed that the >10 years age group poly houses showed lower CEC, due to loss of soil organic carbon by continuous cropping and application of fertilizer. This result is in accordance with the result of Basumatary (1995), who also reported a decline in CEC and base saturation of soil after 7 years of continuous cropping under chemical amendments.

The available N, P, K content in open field soils was low because of the high rainfall as compared to poly house soils (Table 2). The higher available N content in soil under poly houses might be due to increased microbial population resulting faster decomposition of organic matter in summer season over the winter season. Among the different age groups, significantly higher available N was recorded in 3-5 years age group and the lowest was recorded in poly houses >10 years age group. Further, higher available N in soil was recorded in summer season over winter season (Table 2). The lower available N content in older age group of poly houses than younger age groups indicated the deterioration in organic carbon status, soil reaction and CEC of the soil was also
reported by Singh (1991). In case of open field soils, lower available nitrogen content was recorded due to loss of nutrients through leaching in both winter and summer season.

Similarly, higher available P content was observed in summer season over winter season that might be due to release of P from fixed phosphate with increased in temperature (Sharma and Singh, 1999). Among the poly houses, 3-5 years age group poly houses showed higher available P over the other poly houses of other age groups (Table 2). Higher availability of P in surface soils over the sub-surface soils attributed to the higher organic carbon content of the soils that convert the immobile P into labile P (Das and Ram, 2005).

Unlike available nitrogen and phosphorus, available potash in soil was higher during winter compared to summer season (Table 2). A decline trend of available potash content of higher age of poly houses over the open field soils was observed which might be due to consequent higher removal by crops (Bhardwaj and Omanwar, 1994). Further, the data revealed that potash availability in the soil was increased with the increasing depth, the lowest being in the surface layer which might be due to high clay content as reported by Dutta (1991).

Organic carbon content in the soil under field condition was more over the soil inside the poly during both winter and summer season indicating the higher temperature variation within and outside the poly houses (Fig.3.1 and 3.2). The declining trend of organic carbon in poly houses soil with the age of the poly houses was observed which might be due to faster decomposition of organic matter brought about by increased temperature (±3-5°C in summer and ±5-7°C in winter) inside the poly houses leaving very less residual carbon in soils (Sen et al., 1984). Moreover, the less organic carbon content in summer season attributed to higher rate of decomposition because of high temperature, high population and activity of microbes during summer season also leads to faster decomposition of organic matter as compared to winter season. Higher organic carbon content of the surface (0-15 cm) soil over the sub surface (15-30 cm) was recorded which may be due to accumulation of different organic matter and efficient bacterial activity.

**Soil biological properties**

**Soil microbial population**

The congenial environment in open field soils had higher bacterial and fungal population and their activity over the poly house soils (Table 3). The population size of soil microorganism is directly related to soil organic matter content (Alexander, 1961). Further, the toxic substances produced during microbial activity or due to interaction might not be washed away in poly houses leading to lower microbial population compared to open field condition. The surface soils contained higher microbial population as compared to sub-surface soils during both winter and summer season which might be due to the better soil condition prevailed on upper layer in terms of nutrient status and organic matter content (Gupta and Tripathi, 1988). Higher bacterial and fungal population during summer season as compared to winter season was also earlier reported by many workers (Helmeczi et al., 1984; Voinova-Raikova, 1984 and Das et al., 1991) and attributed to the application of fertilizers, decomposition rate of organic residue as well as favorable temperature in the soil.

**Enzyme activities**

The activity of dehydrogenase and phosphomonoesterase enzymes in soil
increased significantly with increasing the age groups. The Dehydrogenase and phosphomonoesterase activity were highest in 0-2 years age group poly houses (Fig. 4.1, 4.2, 5.1 and 5.2). Flourescent di acetate (FDA) hydrolysis activity was recorded highest in open field soils as compared to poly house soils (Fig 6.1 and 6.2), which revealed the congenial natural environment for their activity and availability of sufficient organic matter in natural environment. This may be due to the fact that FDA hydrolysis activity was mainly affected by the stabilization degree of the added organic matter, and it was unaffected by the disturbance caused to soil micro flora despite the disturbance caused by the exogenous organic matter (Mondini et al., 2007).

**Fig.1** Maximum/minimum temperature and relative humidity inside and outside poly house, total rainfall, bright sunshine hour, number of rainy days during the study period

**Fig.2.1** Effect of different age group of poly houses on soil bulk densities at 0-15 cm soil depth
**Fig. 2.2** Effect of different age group of poly houses on soil bulk densities at 15-30 cm soil depth

**Fig. 3.1** Effect of different age group of polyhouses on organic carbon content at 0-15 cm soil depth

**Fig. 3.2** Effect of different age group of polyhouses on organic carbon content at 15-30 cm soil depths
**Fig. 4.1** Effect of different age group of poly houses on dehydrogenase enzyme activity of soils at 0-15 cm depth

**Fig. 4.2** Effect of different age group of poly houses on dehydrogenase enzyme activity of soils at 15-30 cm depth

**Fig. 5.1** Effect of different age group of poly houses on phosphomonoesterase enzyme activity on soil at 0-15 cm depth
Fig. 5.2 Effect of different age group of poly houses on phosphomonoesterase enzyme activity on soil at 15-30 cm depth

Fig. 6.1 Effect of different age group of poly houses on FDA hydrolysis of soil at 0-15 cm depth

Fig. 6.2 Effect of different age group of poly houses on FDA hydrolysis of soil at 15-30 cm depth
Microbial biomass carbon

Higher microbial biomass carbon in open field soils compared to poly house soils might be due to the higher organic carbon content and microbial population in open soils than the poly house soils. The microbial biomass carbon was lowest in poly houses of above 10 years age group (Table 2) and might be due to the unfavourable environment for the microorganisms. The lesser degree of decomposition of organic matter due to low temperature and moisture revealed less microbial biomass carbon in winter season over the summer season.

Soils under poly houses required rejuvenation particularly after 6-8 years for improvement of physical, chemical and biological properties. Generally, poly house soil is subjected to higher temperature and less light intensity than natural soils for which organic matter, a key component of soil fertility, get decomposed fast in poly house and hence there is a reduction in level of this component. Proper maintenance of organic matter in poly house is therefore required so as to improve microbial activity and build up soil fertility. Appropriate management of nutrient, water, tillage as well as plant protection measures may be developed after 6-8 years of poly house cultivation for sustaining soil health and crop productivity.

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