**RESEARCH ARTICLE**

**Meis1: effects on motor phenotypes and the sensorimotor system in mice**

Aaro V. Salminen1,*, Lillian Garrett2,3,*, Barbara Schormair1, Jan Rozman3,4, Florian Giesert2, Kristina M. Niedermeier2, Lore Becker3, Birgit Rathkolb3,4,5, Ilidikó Rácz3,6, German Mouse Clinic Consortium7, Martin Klingenspor7, Thomas Klopstock8,9,10, Eckhard Wolf8, Andreas Zimmer8, Valérie Gailus-Durner3, Miguel Torres11, Helmut Fuchs3, Martin Hrabé de Angelis3,4,12, Wolfgang Wurst2,9,10,13, Sabine M. Höltér2,3 and Juliane Winkelmann1,10,14,15,‡

**ABSTRACT**

**MEIS1** encodes a developmental transcription factor and has been linked to restless legs syndrome (RLS) in genome-wide association studies. RLS is a movement disorder leading to severe sleep reduction and has a substantial impact on the quality of life of patients. In genome-wide association studies, **MEIS1** has consistently been the gene with the highest effect size and functional studies suggest a disease-relevant downregulation. Therefore, haploinsufficiency of **Meis1** could be the system with the most potential for modeling RLS in animals. We used heterozygous **Meis1**-knockout mice to study the effects of **Meis1** haploinsufficiency on mouse behavioral and neurological phenotypes, and to relate the findings to human RLS. We exposed the **Meis1**-deficient mice to assays of motor, sensorimotor and cognitive ability, and assessed the effect of a dopaminergic receptor 2/3 agonist commonly used in the treatment of RLS. The mutant mice showed a pattern of circadian hyperactivity, which is compatible with human RLS. Moreover, we discovered a replicable prepulse inhibition (PPI) deficit in the **Meis1**-deficient animals. In addition, these mice were hyposensitive to the PPI-reducing effect of the dopaminergic receptor agonist, highlighting a role of Meis1 in the dopaminergic system. Other reported phenotypes include enhanced social recognition at an older age that was not related to alterations in adult olfactory bulb neurogenesis previously shown to be implicated in this behavior. In conclusion, the **Meis1**-deficient mice fulfill some of the hallmarks of an RLS animal model, and revealed the role of Meis1 in sensorimotor gating and in the dopaminergic systems modulating it.

**KEY WORDS:** Meis1, Prepulse inhibition, Restless legs syndrome, Sensorimotor system, Mouse model, Pramipexole

**INTRODUCTION**

**MEIS1** encodes a TALE homeobox transcription factor known to play a role in hematopoiesis and vascular patterning (Azcoitia et al., 2005; Carramolino et al., 2010; Hisa et al., 2004), as well as in the development of the proximodistal limb axes (Mecardier et al., 1999), the nervous system (Barber et al., 2013; Spieler et al., 2014) and the development of various organs, such as the heart (Mahmoud et al., 2013), eyes (Marcos et al., 2015) and pancreas (Zhang et al., 2006). **MEIS1** cooperates with other transcription factors to perturb myeloid differentiation in leukemogenesis (Nakamura et al., 1996) and, in genetic association studies, it has been linked with restless legs syndrome (RLS) (Winkelmann et al., 2007, 2011), periodic leg movements during sleep (Moore et al., 2014; Stefansson et al., 2007), symptoms of insomnia (Lane et al., 2016), PR interval (Butler et al., 2012) and waist-to-hip ratio (Shungin et al., 2015). Structurally, **MEIS1** consists of a Pbx interaction motif and a TALE homeodomain near the C-terminus, responsible for DNA interaction. It has two major functional splice variants, **MEIS1A** and **MEIS1B**, which diverge in the C-terminus of the protein (Huang et al., 2005). These isoforms are differentially expressed during development and are likely to differ in their genomic targets and effects on neural differentiation (Maeda et al., 2001). **MEIS1** binds transcription start sites and genomic enhancers alone and in interaction with **Pbx** and **HOX** proteins, regulating the expression of genes involved in the processes of axon guidance, protein phosphorylation and cell differentiation, among others (Marcos et al., 2015; Penkov et al., 2013).

**RLS** is a sensorimotor movement disorder causing severe sleep loss and impaired quality of life (Trenklewalder and Paulus, 2010). The hallmark symptoms include an urge to move the legs, relieved by voluntarily moving the legs. The symptoms are predominantly

---

1Institute of Neurogenomics, Helmholtz Zentrum München, 85764 Neuherberg, Germany. 2Institute of Developmental Genetics, Helmholtz Zentrum München, 85764 Neuherberg, Germany. 3German Mouse Clinic, Institute of Experimental Genetics, Helmholtz Zentrum München, 85764 Neuherberg, Germany. 4German Center for Diabetes Research (DZD), 85764 Neuherberg, Germany. 5Institute of Molecular Animal Breeding and Biotechnology, Gene Center, Ludwig-Maximilians-University Munich, 81377 Munich, Germany. 6Institute of Molecular Psychiatry, Medical Faculty, University of Bonn, 53127 Bonn, Germany. 7Chair of Molecular Nutritional Medicine, Technical University Munich, EKFZ – Else Kröner Fresenius Center for Nutritional Medicine, Gregor-Mendel-Str. 2, 85350 Freising, Germany. 8Department of Neurology, Friedrich-Baur-Institute, Klinikum rechts der Isar, Technische Universität München, Ziemssenstr. 1a, 80336 Munich, Germany. 9Deutsches Zentrum für Neurodegenerative Erkrankungen e. V. (DZNE), 81377 Munich, Germany. 10Munich Cluster for Systems Neurology (SyNergy), Adolf-Butenandt-Institut, Ludwig-Maximilians-Universität München, 81377 Munich, Germany. 11Centro Nacional de Investigaciones Cardiovasculares (CNIC), 28029 Madrid, Spain. 12Chair of Experimental Genetics, School of Life Science Weihenstephan, Technische Universität 85354 Freising, Germany. 13Chair of Developmental Genetics, Faculty of Life and Food Sciences, Ludwig-Maximilians-Universität München, 81377 Munich, Germany. 14German Center for Diabetes Research (DZD), 85764 Neuherberg, Germany. 15Neurologic Clinic, Klinikum rechts der Isar, Technische Universität München, 81675 Munich, Germany.

*These authors contributed equally to this work.

†The members of the German Mouse Clinic Consortium are listed in the Acknowledgements.

‡Author for correspondence (juliane.winkelmann@tum.de)

© 2017. Published by The Company of Biologists Ltd | Disease Models & Mechanisms (2017) 10, 981-991 doi:10.1242/dmm.030080

**This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.**

**Received 28 March 2017; Accepted 19 June 2017**
present in the evening and night time, and at rest. Epidemiological evidence shows that RLS is more common in females compared to males (Berger and Kurth, 2007), but the sex difference could be explained by the number of previous pregnancies (Pantaleo et al., 2010). In the context of RLS, MEIS1 contributes to the heritability of the disease through both common and rare genetic variation. In genome-wide association studies (GWAS), MEIS1 has consistently been the gene with the largest effect size, with an odds ratio up to 1.74 (Winkelmann et al., 2007, 2011). In large-scale sequencing and family studies, putative causal loss-of-function rare variants have been identified (Schulte et al., 2014). A MEIS1 intronic haplotype linked to RLS risk is associated with decreased expression of MEIS1 (Xiong et al., 2009). Therefore, Meis1 haploinsufficiency could be considered the most promising animal model system for human RLS. Meis1 knockout mice, in which Meis1 has been inactivated by knocking in a modified ERT2 domain (Meis1tm1Mtor), die during development due to a fetal hematopoietic failure (Azcoitia et al., 2005). Heterozygous animals, however, show a hyperactive phenotype potentially compatible with components of human RLS (Spieler et al., 2014). In addition, the mutant animals show low fertility, low body mass at birth and various eye-related phenotypes (Azcoitia et al., 2005; Marcos et al., 2015).

In a bid to provide a more comprehensive understanding of the effect of Meis1 haploinsufficiency on the central nervous system, we exposed these heterozygous Meis1-deficient mice to a series of motor, sensorimotor and cognitive behavioral and neurological tests. To further disentangle what role this gene plays in the brain and how it could be involved in RLS, we also determined the effect of a commonly used RLS therapeutic dopaminergic receptor 2/3 (D2/3R) agonist, pramipexole, as well as performing concomitant semi-quantitative measures of dopaminergic neuron number, iron metabolism, believed to play an important role in the pathogenesis of RLS, and adult neurogenesis, the birth and assimilation of new neurons.

RESULTS
Meis1 haploinsufficiency leads to sex-dependent motor alterations at middle age

We have shown previously (Spieler et al., 2014) that Meis1 haploinsufficiency leads to increased locomotor activity in the open field in both male and female young adult mutant mice at the age of 9 weeks. Furthermore, these animals were hyperactive as indexed by a tendency towards increased locomotor activity over a 21-h period in the home-cage environment of PhenoMaster cages (Spieler et al., 2014). In the current study, we determined whether the severities of these effects are maintained with age. Thus, we assessed the same locomotor activity parameters in middle-aged mice (9-11 animals per genotype and sex) from the age of 38-40 weeks. At this older age, Meis1 haploinsufficiency leads to locomotor alterations that dichotomize based on sex. This was manifest as a significant locomotor increase in the male mutant mice in the open field, which was not evident in the females (2-way ANOVA, sex×genotype interaction effect: $F_{1,43}=5.64$, $P=0.022$, Fig. 1C).

Locomotor activity alterations in response to the open field could reflect an anxiety reaction to the novelty of the arena even though this was not evident in the anxiety-related index of center time in the open field (2-way ANOVA, sex×genotype interaction effect: $F_{1,43}=1.70$, $P=0.20$, Fig. 1C).

Fig. 1. Measures of locomotor activity, motor coordination and balance. Measures of locomotor activity in the PhenoMaster cages (females: A, males: B), open field (C) and voluntary running wheels (females D, males E), and motor coordination and balance in the rotarod (F; seconds). *$P<0.05$ vs +/- controls. (B,D) Boxes indicate an increase in activity just after lights on in the rest phase seen in female +/- mice in voluntary running wheels (D) and in male +/- mice in home-cage activity (B). The cohort included 21 female (9 +/- and 12 +/-) and 23 male (10 +/- and 13 +/-) mice. Results are plotted as means±s.e.m. rev, revolutions.
would be expected from an RLS animal model. With this analysis, see whether the locomotor differences are also present when are permitted to habituate to the surroundings. It is then possible to longer-term observation in the PhenoMaster cages where the mice this hyperactivity in the male mutant mice could be confirmed with

Table 1. Results of the nociceptive phenotype screening (hotplate test) in our first mouse cohort

|                | Meis1+/+ | Meis1−/− | Meis1+/+ | Meis1−/− | P-value |
|----------------|----------|----------|----------|----------|---------|
|                | Female   | Male     | Female   | Male     |         |
| First response time (s) | 10.53±3.11 | 10.84±3.29 | 29.41±3.05 | 27.42±2.5 | 0.402   |
| First response type: licking (no.) | 8        | 10       | 4        | 11       |         |
| First response type: shaking (no.) | 7        | 5        | 11       | 4        |         |
| Second response time (s) | 14.25±3.72 | 14.33±3.71 | 13.37±2.47 | 13.72±1.85 | 0.787   |
| Second response type: licking (no.) | 7        | 5        | 11       | 4        |         |
| Second response type: shaking (no.) | 8        | 10       | 4        | 11       |         |
|                | Click ABR | ABR      | Click ABR | ABR      |         |
| Threshold at 6 kHz (dB) | 40 [40, 44] | 48 [45, 50] | 40 [35, 40] | 35 [35, 40] | 0.104   |
| Threshold at 12 kHz (dB) | 28 [25, 34] | 30 [26, 34] | 20 [20, 25] | 15 [15, 20] | 0.612   |
| Threshold at 18 kHz (dB) | 35 [30, 40] | 45 [35, 49] | 30 [25, 35] | 25 [20, 30] | 0.908   |
| Threshold at 24 kHz (dB) | 55 [46, 59] | 52 [46, 55] | 40 [32, 40] | 35 [30, 45] | 0.876   |
| Threshold at 30 kHz (dB) | 62 [48, 70] | 65 [56, 75] | 50 [45, 55] | 55 [45, 60] | 0.115   |

The nociception results are presented as means±s.d. and P-values are calculated using 2-way ANOVA with sex and genotype as factors with interaction term. The auditory brainstem response (ABR) results are presented as median [25%, 75%] and the P-values calculated with Wilcoxon rank sum test. All P-values represent the genotype effect. n=15 per sex per genotype.
Meis1 deficiency confers insensitivity to D2/3R-agonist effects on sensorimotor gating

Given this robust and replicable effect of Meis1 haploinsufficiency on sensorimotor gating ability, we wanted to determine the effect of a dopaminergic receptor agonist used in the treatment of RLS on this phenotype. Thus, the effect of pramipexole, a preferential D2/3R agonist, at two doses [1 and 3 mg/kg bodyweight intraperitoneal (i.p.)] on PPI was assessed with saline vehicle control. In the vehicle-treated mice, the mutant animals showed a higher reactivity to acoustic startle than the wild types at higher sound intensities. PPI was decreased in the mutant mice after vehicle injection, replicating the findings of the first two cohorts. The 1 mg/kg dose of pramipexole had an effect at high sound intensities in the mutant group, making their startle response even stronger. In the control group, the startle response was only altered at lower sound intensities. The 3 mg/kg dose of pramipexole, on the other hand, increased the startle response of the wild-type mice at higher stimulus intensities but had no effect on the mutant group. The results are shown in Fig. S1.

PPI was affected by pramipexole in a more consistent manner. In the wild-type group, PPI was significantly reduced by both high and low doses of pramipexole. The PPI in these animals was reduced to the level of the vehicle-treated mutant mice. In contrast, in the heterozygous mutants, the PPI was not significantly altered by pramipexole (Fig. 2C).

To investigate whether this insensitivity to the D2/3R agonist pramipexole was influenced by the number of dopaminergic neurons, which could lead to altered D2/3R signaling, using optical fractionator estimates we quantified the number of tyrosine hydroxylase (TH)-positive cells in the substantia nigra pars compacta (SNpc) and the ventral tegmental area (VTA). We did not observe clear differences in the total number of TH+ cells in either the SNpc (2-way ANOVA, genotype effect: F_{1,12}=0.401, P=0.538, Fig. 3C) or the VTA (2-way ANOVA, genotype effect: F_{1,12}=0.291, P=0.599, Fig. 3F) between genotypes to indicate that neuron number differences lead to the altered signaling. Furthermore, we assessed iron homeostasis by measuring the plasma levels of iron, ferritin (iron storage) and transferrin (iron binding) (Table 2) because iron has been suggested to play a major role in RLS pathogenesis in humans (Earley et al., 2014). There were no clear differences between the groups in any of the three measures except for a tendency towards a decrease in the iron-storage protein ferritin in the female Meis1-deficient mice (2-way ANOVA, sex×genotype interaction effect: F_{1,56}=3.32, P=0.074).

Fig. 2. Prepulse inhibition (PPI). The PPI results from the first screening cohort (A, n=15 per sex per genotype) and the replication cohort (B, n=15 per sex per genotype). The results show that the Meis1 knockout genotype effect on PPI was replicable when compared to wild-type littermates. Results are plotted as means±s.e.m. Genotype effects: *P<0.05, **P<0.005, ***P<0.001 calculated by two-way ANOVA with sex as a factor, including interaction term. (C) The global effect of pramipexole 1.0 mg/kg body weight (left) and 3.0 mg/kg body weight (right) on the PPI of wild-type and heterozygous animals is shown. Results are plotted as means±s.e.m. Genotype effects: **P<0.005 calculated by two-way ANOVA with sex as a factor, including interaction term.
**Meis1 haploinsufficiency alters social memory**

The olfactory social recognition index was increased in both female and male mutant mice compared to wild types (Fig. 4A; 2-way ANOVA, genotype effect: $F_{1,41}=9.48$, $P=0.004$, sex effect: $F_{1,41}=0.58$, $P=0.45$, sex×genotype interaction effect: $F_{1,41}=1.45$, $P=0.24$), suggesting better social discrimination memory. There were no genotype-related differences in social investigation time during the sample phase of the test (data not shown). The social recognition index showed no correlation with distance traveled in the open field ($r=−0.050$, $P=0.749$), with time spent in the middle of the arena ($r=−0.166$, $P=0.282$) or with total activity in the running wheel ($r=−0.073$, $P=0.679$). None of the parameters showed significant correlation when the sexes were tested separately (data not shown). We have shown previously that decreased social memory can be due to decreased adult neurogenesis in mice (Garrett et al., 2015). Thus, to determine whether the behavioral alterations were associated with modifications of the adult neurogenesis in Meis1-deficient mice, a stereological approach was used to quantify the number of doublecortin (DCX)-positive cells in the granular cell layer of the olfactory bulb (Fig. 4B). DCX expression is an immature neuron marker in the adult brain that has also been shown to reflect overall levels of neurogenesis, and DCX+ cell quantification can be used as an alternative to BrdU pulse/chase analysis (Couillard-Despres et al., 2005). Optical fractionator estimates showed that there was no significant effect of genotype on the number of DCX+ cells in the granular cell layer of the olfactory bulbs in these mice (unpaired $t$-test: $t_{10}=0.401$, $P=0.697$).

**DISCUSSION**

Haploinsufficiency of Meis1 is a potential model system for simulating RLS in mice. Here, we show that this Meis1 deficit causes a sex- and context-dependent RLS-related phenotype at middle age, impaired sensorimotor gating ability that was refractory to the D2/3R agonist pramipexole as well as increased social memory. These behavioral abnormalities occurred without differences in thermal sensitivity, plasma iron concentrations or dopaminergic neuron number in the SNpc/VTA. Thus, although Meis1 haploinsufficiency causes the RLS-related phenotype of motor restlessness in mice, the detailed nature of these effects is complex. Furthermore, it is characteristically distinct from other genetic RLS models (e.g. Btbd9 knock-out), highlighting the heterogeneous nature of the effects of these different risk genotypes in this disease.

Motor restlessness in these Meis1-haploinsufficient mice was detected previously in young adult male and female mice (Spieler et al., 2014). RLS is, however, more prevalent at an older age in humans (Berger et al., 2004) and therefore we also tested middle-aged mice. Human RLS occurs in a circadian fashion where the patients are typically affected in the evening hours, and therefore a circadian hyperactivity pattern specific to the beginning of the inactive period should be expected from an RLS animal model. So, although we have shown here that the overall locomotor effects in the open field and PhenoMaster cages deviate per the sex of the animal in middle age, both the male and female Meis1-haploinsufficient mice still show such an RLS-like phenotype: specific increases in activity...
during the initial stage of the rest phase. In the males, this occurs in the PhenoMaster cages and, in the females, in their voluntary wheel-running activity pattern. In males, however, hyperactivity is also observed during the active period, making the effect less specific to the RLS-relevant time bins.

Meis1-haploinsufficiency thus leads to RLS-like circadian motor restlessness in middle age that, in the case of the females, requires amplification with an additional environmental stimulus. The latter could represent the urge to move, a key feature of human RLS. This outcome further reveals that RLS associated with Meis1 risk haplotypes is likely to produce sexually dimorphic motor effects in middle age.

In terms of sensory function, the Meis1 deficiency did not alter thermal sensitivity in response to the hotplate, which is incongruous with the effects in the Btbd9 mouse RLS model (DeAndrade et al., 2012). Nevertheless, augmented thermal sensitivity is a feature of patients with secondary RLS, whereas patients with idiopathic RLS experience different types of sensitivity (Bachmann et al., 2010). We also report here the first evidence to indicate that a Meis1 deficit leads to a reproducibly decreased PPI response. This deficit occurs in the absence of ABR alterations, suggesting that the PPI phenotype is not secondary to impaired hearing in the mice. PPI of the acoustic startle reflex is a commonly used rodent readout for sensorimotor gating and integration. Impairment of PPI has been observed in neurological disorders (Kohl et al., 2013) and it is most frequently used as a readout in schizophrenia research (Swerdlow et al., 2001). To test whether it may be a causative factor in the insensitivity to pramipexole in these mice we quantified the number of TH+ neurons in the SNpc and VTA. Nevertheless, our results did not indicate significant differences in the numbers of dopaminergic neurons in these areas. It remains possible that the expression of dopamine receptors is reduced with Meis1 risk haplotypes. This prospect is also supported by the lack of PPI differences in homozygous Btbd9-knockout mice (Btbd9tm1b(EUCOMM)Wtsi) (Meehal et al., 2017; see also: www.mousephenotype.org/data/experiments?geneAccession=MGI:1916625). Thus, PPI should be studied in RLS patients who also are carriers of the MEIS1 risk allele.

In our treatment experiment, pramipexole, a D2/3R agonist, did not rescue the PPI phenotype in Meis1-deficient mice. On the contrary, it reduced the PPI in wild-type mice to the level of the mutants. The effect of pramipexole on wild-type mice contradicts an earlier study reporting no pramipexole effect on PPI in C57BL/6 mice (Chang et al., 2010). However, the Meis1-deficient mice were bred on a C57BL/6JOlaHsd substrain background and this different outcome could be explained by the high strain sensitivity of the dopaminergic effect on PPI (Swerdlow et al., 2004). The lack of effect on the mutant animals might indicate that the Meis1-deficient mice are hyposensitive to pramipexole, which may signify the involvement of Meis1 in the dopaminergic system. It has been demonstrated in neuropathological studies that there is a higher level of TH, the dopaminergic synthetic enzyme, in the substantia nigra of RLS sufferers and fewer D2 receptors in the putamen correlated with RLS severity (Connor et al., 2009). Thus, given that PPI is regulated by dopaminergic systems of the nucleus accumbens and striatum (Swerdlow et al., 2004), to test whether it may be a causative factor in the insensitivity to pramipexole in these mice we quantified the number of TH+ neurons in the SNpc and VTA. Nevertheless, our results did not indicate significant differences in the numbers of dopaminergic neurons in these areas. It remains possible that the expression of dopamine receptors is reduced.

Fig. 4. Behavioral alterations are not associated with modifications of adult neurogenesis in Meis1-deficient mice. (A) Social recognition test revealed an increased social recognition index in the mutant animals compared to wild types. The cohort included 21 female (9 heterozygous and 12 wild-type) and 23 male (10 heterozygous and 13 wild-type) mice. **P<0.01, 2-way ANOVA genotype effect: F1,41=9.48, P=0.004, Meis1+/− vs Meis1+/+.

(B) No clear differences were detected in the level of adult neurogenesis in the olfactory bulb granule cell layer (OB) of the mutant mice as indexed by the number of doublecortin (DCX)+ cells in this region. Data from males and females were pooled for this analysis (n=6+/+, n=6 +/−). Results are plotted as means±s.e.m. Representative photomicrographs are also depicted showing DCX+ cells (black arrowheads) in the OB of wild type (+/+) and heterozygous mutant (+/−) mice. Scale bars: 100 µm.
in Meis1-deficient mice and this will require further study. There were also no differences in plasma iron homeostasis. This again distinguishes the Meis1-haploinsufficient mice from the Btbd9 mice, where the latter also shows decreased serum iron levels (DeAndrade et al., 2012). Nevertheless, the Btbd9 mice did not show differences in brain-tissue iron levels. Thus, it would be propitious to quantify striatal iron levels in these Meis1-deficient mice.

We have shown here, for the first time, a non-sensorimotor cognitive phenotype associated with Meis1 deficiency. The Meis1-deficient mice had improved social memory as indexed by enhanced social recognition ability. There is limited conflicting evidence available concerning the cognitive ability of RLS patients with reports of deficits (Kim et al., 2014), enhanced ability (Moon et al., 2014) and no alterations in different types of memory (Driver-Dunckley et al., 2009; Fulda et al., 2010). These inconsistencies could relate to the type of memory assessed, the age and sex of the patient, the type of RLS (idiopathic versus secondary), medication status and, as previously alluded, patients in these studies have not been stratified according to risk haplotypes. Moon and colleagues have shown that drug-naive idiopathic RLS patients, predominantly at middle age, exhibit enhanced verbal fluency and word memory (Moon et al., 2014). They conjectured that this increased ability may be an adaptation to sleep disturbance manifesting as heightened alertness with an unknown underlying mechanism. We have demonstrated previously involvement of adult neurogenesis in social discrimination memory in the DCXCreER2; DTA model where loss of DCX+ cells and hence decreases in adult neurogenesis reversibly impaired social recognition memory (Garrett et al., 2015). Therefore, given the impaired social recognition memory, we hypothesized that abnormal neurogenesis in the olfactory bulbs of Meis1-deficient mice may underlie this deficit. Our quantification of DCX+ cells in the olfactory bulb did not, however, support this hypothesis and no differences were observed. This indicates that alternative processes likely mediate the enhanced social memory consequent to Meis1 deficiency. Recent evidence showing zonal Meis1 expression in the main olfactory epithelium could signify one potential avenue for further investigation in this regard (Parrilla et al., 2016). Furthermore, in light of this new evidence, a more detailed analysis of cognitive ability in RLS patients with the MEIS1 risk haplotype is warranted.

In summary, Meis1-deficient mice show an RLS-like motor-restlessness phenotype, a sensorimotor gating deficiency refractory to a D2/3R agonist and some heretofore unknown Meis1-deficiency-related effects. Other proposed RLS-related genetic mouse models such as Btbd9 (DeAndrade et al., 2012) and Ptprd (Drgonova et al., 2015) exhibit degrees of motor restlessness. There are features of Meis1-deficient mice, however, that are distinct and could be explained by the genetic architecture of the disease. For example, when a motor phenotype (periodic leg movements during sleep) is used to assess RLS in human genetic studies, the BTBD9 locus has the highest effect size (Moore et al., 2014; Stefansson et al., 2007). When the sensory RLS symptoms are assessed, the MEIS1 locus is the top hit (Winkelmann et al., 2007, 2011). This indicates that the specific disease characteristics in patients will depend on the combination of risk alleles at different genetic risk loci and this is supported by the current data. RLS is a genetically heterogeneous complex trait with high prevalence but large phenotypic variability, and both idiopathic and secondary forms contribute to prevalence statistics. Thus, fine-mapping and further genetic studies in RLS patients may reveal differential effects of the various risk loci as observed in these mice and in other studies. In future investigations, an RLS-related mutation within Meis1 may provide a more disease-relevant disruption of Meis1 function in mice and therefore a better RLS phenotype (Allen et al., 2017).

Materials and Methods

Animals and breeding

The Meis1flm1Mtor mice used in the experiments were created in Madrid, Spain (Azcoitia et al., 2005) and have been bred on the C57BL/6JoAldh (Envigo, Horst, The Netherlands) background. In this strain, Meis1 has been inactivated by knocking in a modified ERT2 domain. The result is predicted to encode an inactive protein product. The mouse line has been bred in Munich, Germany, backcrossing to wild-type C57BL/6JoAldh every generation. Mouse husbandry was performed according to the FELASA recommendations (Nicklas et al., 2002) (http://www.felasa.eu) in a controlled specific pathogen free (SPF) hygiene standard environment. The mice had access to ad libitum standard feed and water always. All tests were carried out with approval for the ethical treatment of animals by the responsible authority of the Regierung von Oberbayern (Government of Upper Bavaria).

Analysis of Meis1flm1Mtoror mice

Four different mouse cohorts were bred for the experiments described here and the corresponding tests and order performed are shown in Fig. 5. In all experiments, heterozygous animals (Meis1+/−) were used, homoygotes were embryonic lethal (Azcoitia et al., 2005). The mice from the different cohorts were tested together with their wild-type (Meis1+/+) littermates as controls. Animals were tested with paraformaldehyde (PFA) and used for brain-tissue stereological analysis of dopaminergic neuron number (TH+ cells) and adult neurogenesis (DCX+ cells). The mice (30 animals per sex per genotype) from cohort 4 were tested for sensitivity to pramipexole in PPI (at 10 weeks) for 1 mg/kg body weight and at 11 weeks for 3 mg/kg body weight. Mice from the first cohort also underwent the standardized phenotype screening at the German Mouse Clinic (GMC; www.mouseclinic.de). The test battery included additional screening of the mice in the areas of dysmorphology, clinical chemistry, energy metabolism, cardiovascular studies, eye function, immunology, allergy, steroid metabolism, lung function and pathology. The screens were performed as described in previous publications (Fuchs et al., 2009; Gailus-Durner et al., 2005, 2009) and the results of this analysis can be found on the GMC phenomap (tools.mouseclinic.de/phenomap/jsp/ annotation/public/phenomap.jsp).

Open field

The open-field analysis was carried out as described previously (Garrett et al., 2012; Hölter et al., 2013; Zimprich et al., 2014). It consisted of a transparent and infrared-light-permeable acrylic test arena with a smooth floor (internal measurements: 45.5×45.5×39.5 cm). Illumination levels were set at approx. 150 lux in the corners and 200 lux in the middle of the test arena. Data were recorded and analyzed using the ActiMot system (TSE, Bad Homburg, Germany).

PPI of the ASR

Mice were housed with food and water available ad libitum under standard laboratory conditions. Animals were separated based on sex, but not genotype. PPI was assessed using a startle apparatus setup (Med Associates Inc., VT, USA) including four identical sound-attenuating cubicles. The
protocols were written using the Med Associates ‘Advanced Startle’ software. Experiments were carried out between 08:30 h and 17:00 h. Background noise was 65 dB, and startle pulses were bursts of white noise (40 ms). A session was initiated with a 5-min acclimation period followed by five presentations of leader startle pulses (110 dB) that were excluded from statistical analysis. Trial types for the PPI included four different prepulse intensities (67, 69, 73, 81 dB); each prepulse preceded the startle pulse (110 dB) by a 50-ms inter-stimulus interval. Each trial type was presented ten times in random order, organized in ten blocks, each trial type occurring once per block. Inter-trial intervals varied from 20 to 30 s. This protocol is based on the protocol used in IMPRESS from the IMPC (see www.mousephenotype.org/impress), adapted to the specifications of our startle equipment.

Drug preparation
Pramipexole (pramipexole dihydrochloride, Sigma Aldrich, St Louis, MO, USA) was prepared in saline and injected at two different doses (1 and 3 mg/kg body weight). For the PPI test with drug treatment, Meis1 and control mice were assigned to receive either pramipexole (1 and 3 mg/kg body weight) or saline vehicle and this was balanced for genotype, startle chamber and treatment. The mice were tested for PPI 1 h after administration of the treatment and the i.p. injection volume was 10 ml/kg body weight. The experimenter was not blinded during the PPI testing as the test is automated and there was no possibility for observer bias.

ABR
ABR measurements were performed using an RP2.1 workstation (Tucker-Davis Technologies, FL, USA) as previously described (Lee et al., 2015). In short, animals were anesthetized and electrodes were placed on the vertex (reference) and ventrolateral to the ears (active and ground) of the animals. ABR potentials, as responses to broadband clicks or pure-tone frequencies, were recorded. Frequencies ranged from 6 to 30 kHz with a 6 kHz stepping, and sound pressure levels ranged from 0 to 85 dB with 5 dB steps. Hearing thresholds were determined for each frequency as the lowest sound pressure level (SPL) producing a measurable ABR pattern response.

Hotplate
In nociception, mice were screened using a hotplate assay. A mouse was placed on a 28-cm-diameter metal surface maintained at 52+0.2°C surrounded by a 20-cm-high Plexiglass wall (TSE, Bad Homburg, Germany). Mice remained for 30 s on the plate or until they performed one of three behaviors regarded as indicative of nociception: hind-paw licking, hind-paw shake/flutter or jumping. The latency of the first sign of pain was compared.

Social discrimination
The social discrimination procedure consisted of two 4-min exposures of stimulus animals (ovariectomized 129Sv females) to the test animal in a fresh cage to which the test animal had been moved 2 h prior to testing. During the first exposure, one stimulus animal was exposed to the test animal. After a retention interval of 2 h, this stimulus animal was re-exposed to the test animal together with an additional, not previously presented stimulus animal. The duration of investigatory behavior of the test animal towards the stimulus animals was recorded by a trained observer with a hand-held computer. A social recognition index was calculated as time spent investigating the unfamiliar stimulus mouse/time spent investigating both the familiar and unfamiliar stimulus mouse.
Rotarod
The rotarod (Bioseb, Chaville, France) was used to measure forelimb and hindlimb motor coordination, balance and motor learning ability. The machine was set up in an environment with minimal stimuli such as noise and movement. The rotarod device is equipped with a computer-controlled motor-driven rotating rod. The unit consists of a rotating spindle and five individual lanes, one for each mouse. In general, the mouse is placed perpendicular to the axis of rotation, with head facing the direction of the rotation. All mice were placed on the rotarod at an accelerating speed from 4 to 40 rpm for 300 s with 15 min between each trial. In motor-coordination testing, mice were given four trials at the accelerating speed in one day. The mean latency to fall off the rotarod during the trials was recorded and used in subsequent analysis. Before the start of the first trial, mice were weighed.

Locomotor activity in PhenoMaster cages
For the evaluation of home-cage locomotor activity, single mice were kept in respirometry cages (PhenoMaster System, TSE Systems, Germany) (Gaulius-Durner et al., 2009). The measurement started at 13:00 h (CET) after a 2-h adaptation to the cages and continued until 10:00 h the next morning. The setup allowed the analysis of locomotor activity of individual mice every 20 min, resulting in 63 readings per individual and trial. Two infrared-light-beam frames allowed the monitoring of physical activity (lower frame: distance traveled per 20 min, upper frame: number of rearings per 20 min).

Voluntary wheel running
All mice were singly housed and provided with an angled running wheel (diameter 15.50 cm) complete with a wireless-controlled activity counter (Wheel Manager software, Med Associates Inc., VT, USA) for 21 days.

Plasma iron level analysis
Blood samples were collected in the morning at 14 or 15 weeks of age by retrobulbar puncture under isoflurane anesthesia. Determination of clinical chemistry parameters in heparinized plasma samples was done using an AU480 (Beckman-Coulter, Krefeld, Germany) and reagent kits provided by Beckman-Coulter, as described previously (Rathkolb et al., 2013). Hematomalyses analyses were performed with an ABC animal blood counter (Scil animal care company, Viernheim, Germany) on whole-blood samples collected in EDTA-coated tubes.

Tissue preparation
Animals were deeply anesthetized using CO₂ and perfused transcardially with 4% PFA in 0.1 M phosphate buffer. Brains were dissected from the skulls, post-fixed overnight in 4% PFA at 4°C and then transferred to a 30% sucrose solution at ice-cooled block with a sliding microtome (Leica, Bensheim) into 40-µm-(w/v) sucrose solution until saturated. Brains were then sectioned on a dry-skulls, post-fixed overnight in 4% PFA at 4°C and then transferred to a 30% sucrose solution at ice-cooled block with a sliding microtome (Leica, Bensheim) into 40-µm-(w/v) sucrose solution until saturated. Brains were then sectioned on a dry-skulls, post-fixed overnight in 4% PFA at 4°C and then transferred to a 30% sucrose solution at ice-cooled block with a sliding microtome (Leica, Bensheim) into 40-µm-(w/v) sucrose solution until saturated. Brains were then sectioned on a dry-skulls, post-fixed overnight in 4% PFA at 4°C and then transferred to a 30% sucrose solution at ice-cooled block with a sliding microtome (Leica, Bensheim) into 40-µm-(w/v) sucrose solution until saturated. Brains were then sectioned on a dry-skulls, post-fixed overnight in 4% PFA at 4°C and then transferred to a 30% sucrose solution at ice-cooled block with a sliding microtome (Leica, Bensheim) into 40-µm-(w/v) sucrose solution until saturated.

Immunostaining
For immunostaining of DCX and TH, an Avidin-Biotin Complex (ABC) method like that employed previously (Garrett et al., 2012) was used. For the DCX staining, a primary rabbit polyclonal anti-DCX antibody (1:500, Abcam, cat. no: ab18723) was used on this occasion with a biotinylated goat anti-rabbit IgG (1:300; Jackson ImmunoResearch Laboratories Inc.) and 3,3’-diaminobenzidine (DAB) as the chromogen. For the TH staining, a primary rabbit anti-TH antibody (1:1000, Pel-Freeze, cat. no: P40101) was used with a biotinylated goat anti-rabbit IgG (1:300; Jackson ImmunoResearch Laboratories Inc.) and DAB as the chromogen.

Quantification of TH+ and DCX+ neurons
Estimation of the total number of TH+ and DCX+ cells was determined using a method based on the stereological optical fractionator principle and the semiautomatic StereoInvestigator system (MicroBrightField Inc., Williston, VT, USA). For TH+-cell estimation, the SNpc and VTA were traced in every 6th section. For DCX+-cell estimation, the olfactory bulb granular cell layer (OB) was traced in every 6th section. Immunopositive cells were quantified by systematic random sampling using the following settings: TH+ cells in the SNpc and VTA: a scan grid size of 200×200 µm and a counting frame size of 70×70 µm; DCX+ cells in the OB: a scan grid size of 200×200 µm and a counting frame of 100×100 µm. Cells that intersected the uppermost focal plane or the lateral exclusion borders of the counting frame were not quantified. These settings were sufficient to keep the Gundersen coefficient of error (m=1) less than 0.1 for each animal.

Statistics
Tests for genotype effects were done either by t-test, Wilcoxon rank sum test, linear models or ANOVA. The choice of the method depended on the assumed distribution of the parameter and the nature of the question posed to the data. Correlations in the aged cohort were evaluated using Pearson correlation coefficient. A P-value of 0.05 was used as level of significance without correction for multiple testing. The analyses were performed using the programming language R (version 3.0.2).

Competing interests
The authors declare no competing or financial interests.

Author contributions
Conceptualization: A.V.S., L.G., B.S., J.R., F.G., K.M.N., L.B., B.R., I.R., GMCC, M.K., T.K., E.W., A.Z., V.G.-D., H.F., M.H.d.A., W.W., S.M.H., J.W.; Methodology: A.V.S., L.G., B.S., J.R., F.G., K.M.N., L.B., B.R., I.R., GMCC, M.K., T.K., E.W., A.Z., V.G.-D., H.F., M.H.d.A.; Formal analysis: A.V.S., L.G., B.S., W.W.; Resources: B.S., M.T., W.W., S.M.H., J.W.; Writing - original draft: A.V.S., L.G., B.S., W.W., S.M.H., J.W.; Writing - review & editing: F.G., K.M.N., M.T.; Visualization: A.V.S., L.G.; Supervision: L.G., J.R., L.B., B.R., I.R., GMCC, M.K., T.K., E.W., A.Z., V.G.-D., H.F., M.H.d.A., W.W., S.M.H., J.W.; Project administration: B.S., J.W.; Funding acquisition: W.W., S.M.H., J.W.

Funding
A.V.S. was supported by the Emil Aaltonen Foundation, Tampere, Finland (Emil Aaltonen Säätiö). J.W. and M.T. were partly funded by the European Commission through an ERA-NET NEURON grant. This work has been funded by the German Federal Ministry of Education and Research (Bundesministerium für Bildung und Forschung) to the GMC (Infrafrontier grant 01KX1012), to the German Center for Diabetes Research (DZD e.V.) and through the Integrated Network MitoPD (Mitochondrial endophenotypes of Morbus Parkinson), under the auspices of the eMed Programme (grant 031A430E), as well as by the Heilmholz Portfolio Theme ‘Supercomputing and Modelling for the Human Brain’ (STMBH) to W.W. This work was also supported by the German Science Foundation Collaborative Research Centre (CRC) (Deutsche Forschungsgemeinschaft) 870.

Supplementary information
Supplementary information available online at http://dmm.biologists.org/lookup/doi/10.1242/dmm.030080-supplemental

This article has an associated First person interview with the first author(s) of the paper available online at http://dmm.biologists.org/lookup/doi/10.1242/dmm.030080.supplemental

References
Allen, R. P., Donelson, N. C., Jones, B. C., Li, Y., Manconi, M., Rye, D. B., Sanyal, S. and Winkelmann, J. (2017). Animal models of RLS phenotypes. Sleep Med. 31, 23-28.
Huang, H., Rastegar, M., Bodner, C., Goh, S.-L., Rambaldi, I. and Featherstone, M. (2005). MEIS C termini harbor transcriptional activation domains that respond to cell signaling. J. Biol. Chem. 280, 10119-10127.

Kim, S. M., Choi, J. W., Lee, C., Lee, B. U., Koo, Y. S., Kim, K. H. and Jung, K.-Y. (2014). Meis1 regulating memory deficits in patients with restless legs syndrome: an event-related potential study. Sleep Med. 15, 808-815.

Kohl, S., Heekeren, K., Klosterkötter, J. and Kuhn, J. (2013). Prefrontal inhibition in psychiatric disorders – Apart from schizophrenia. J. Psychiatr. Res. 47, 445-452.

Lane, J. M., Liang, J., Vlasac, I., Anderson, S. G., Bechtold, D. A., Bowden, J., Emsley, R., Gill, S., Little, M. A., Luik, A. I. et al. (2016). Genome-wide association analyses of sleep disturbance traits identify new loci and highlight shared genetics with neuropsychiatric and metabolic traits. Nat. Genet. 49, 274-281.

Lee, T., Rose, S., Metodiev, M. D., Becker, L., Vernoaken, A., Klopstok, T., Gailus-Durner, V., Fuchs, H., HablìÄŒ, M., Douthwaite, S. and Larsson, N. G. (2015). Overexpression of the mitochondrial methyltransferase TFB1M in the mouse does not impact mitobonsomal methyltransferation status or hearing. Hum. Mol. Genet. 20, 7286-7294.

Leoni, S., Winkel, E. F., Peckham, E., León-Ariza, D. S., Bara-Janemita, J. and Hallett, M. (2015). Auditory and lower limb tactile prepulse inhibition in primary restless legs syndrome: clues to its pathophysiology. J. Clin. Neurophysiol. 32, 369-374.

Maeda, R., Mood, K., Jones, T. L., Aruga, J., Buchberg, A. M. and Daar, I. O. (2015). Xmeis 1, a protooncogene involved in specifying neural crest cell fate in Xenopus embryos. Oncogene 20, 1329-1342.

Mahmod, A. I., Kocabas, F., Muralidhar, S. A., Kimura, W., Koura, A. S., Thet, S., Porrello, E. R. and Sadek, H. A. (2013). Meis1 regulates postnatal cardiac progenitor cell fate and ensures normal heart function. Nature 497, 245-251.

Marcos, S., Gonzalez-Lazaro, M., Beccari, L., Carramolino, L., Martin-Bermejo, M. J., Amarante, O., Mateos-San Martin, D., Torroja, C., Bogdanovic, O., Doohan, R. et al. (2015). Meis 1 coordinates a network of genes implicated in eye development and microphthalmia. Development 142, 3009-30020.

Meehan, T. F., Conte, N., West, D. B., Jacobsen, J. O., Mason, J., Warren, J., Chen, C. K., Tudose, I., Relac, M. et al. (2017). Disease model discovery from 3,328 gene knockouts by The International Mouse Phenotyping Consortium. Nat. Genet. doi:10.1038/ng.3901 [Epub ahead of print].

Mecardner, N., Leonardo, E., Azpilzueta, S., Serrano, A., Morata, G., Martinez-A. C. and Torres, M. (1999). Conserved regulation of proximodistal limb axis development by Meis1/th4. Nature 402, 425-429.

Moon, Y. J., Song, J.-Y., Lee, B. U., Koo, Y. S., Lee, S. K. and Jung, K.-Y. (2014). Comparison of cognitive function between patients with restless legs syndrome and healthy controls. Sleep Med. Res. 5, 20-24.

Moon, S.-I., Winkel, E. F., Lin, L., Fine, L., Peppard, P. and Mignot, E. (2014). Periodic leg movements during sleep are associated with polymorphisms in BTBD9, TOX3/BC034767, MEIS1, MAP2K5/SKOR1, and PTPRD. Hum. Mol. Genet. 23, 2713-2739.

Nakamura, T., Largaespada, D. A., Shaughnessy, J. D., Jenkins, N. A. and Copeland, N. G. (1996). Cooperation of Hoxa and Pbx1-related genes in murine myeloid leukemias. Nat. Genet. 12, 149-153.

Nicklas, W., Baneux, P., Boot, R., Decelle, T., Deeny, A. A., Fumanelli, M., Ilgen-Wilcke, B. and FELASA (Federation of European Laboratory Animal Science Associations Working Group on Health Monitoring of Rodent and Rabbit Colonies) (2002). Recommendations for the health monitoring of rodent and rabbit colonies in breeding and experimental units. Lab. Anim. 36, 20-42.

Pantaleo, N. P., Hening, W. A., Allen, R. P. and Earley, C. J. (2010). Pregnancy accounts for most of the gender difference in prevalence of familial RLS. Sleep Med. 11, 310-315.

Parrilla, M., Chang, I., Dell’ignocenti, A. and Omura, M. (2016). Expression of homeobox genes in the mouse olfactory epithelium. J. Comp. Neurol. 524, 2713-2739.

Penkov, D., Sanmartin, D. M., Fernandez-Diaz, L. C., Rosselli, C. A., Torroja, C., Salcedo-Cabo, F., Warnatz, H. J., Sultan, M., Yaso, M. L., Gabrieli, A. et al. (2013). Analysis of the DNA-binding profile and function of TALE homeoproteins reveals their specialization and specific interactions with hox genes/proteins. Cell Rep. 3, 1321-1333.

Richard, B., Heins, W., Prehn, C., Fuchs, H., Galius-Durner, V., Aigner, B., Adamski, J., Wolf, E. and HablìÄŒ, M. (2013). Clinical chemistry and other laboratory tests on mouse plasma or serum. In Current Protocols in Mouse Genetics, pp. 69-100. Hoboken, NJ, USA: John Wiley & Sons, Inc.

Schulte, E. C., Kousi, M., Tan, P. L., Tilch, E., Knauf, F., Lichtner, P., Trenkwalder, C., Bang, F., Frauscher, B., Berger, K. et al. (2014). Targeted resequencing and systematic in vivo functional testing identifies rare variants in MEIS1 as significant contributors to restless legs syndrome. Am. J. Hum. Genet. 95, 85-95.

Shungin, D., Winkler, T. W., Croteau-Chonka, D. C., Ferreira, T., Locke, A. E., Magi, R., Strawbridge, R. J., Pers, H. F., Fischer, K., Justice, A. E. et al. (2015). New genetic loci link adipose and insulin biology to body fat distribution. Nature 518, 187-196.
Spieler, D., Kaffe, M., Knauf, F., Bessa, J., Tena, J. J., Giesert, F., Schormair, B., Tilch, E., Lee, H., Horsch, M. et al. (2014). Restless Legs Syndrome-Associated intronic common variant in Meis1 alters enhancer function in the developing telencephalon. Genome Res. 24, 592-603.

Stefansson, H., Rye, D. B., Hicks, A., Petursson, H., Ingason, A., Thorgersson, T. E., Palsson, S., Sigmundsson, T., Sigurdsson, A. P., Eiriksdottir, I. et al. (2007). A genetic risk factor for periodic limb movements in sleep. N. Engl. J. Med. 357, 639-647.

Swerdlow, N. R., and Light, G. A. (2015). Animal models of deficient sensorimotor gating in schizophrenia: are they still relevant? In Translational Neuropsychopharmacology. Current topics in Behavioral Neurosciences (Eds, T. W. Robbins and B. J. Sahakian), pp. 305-325. Springer.

Swerdlow, N. R., Geyer, M. A. and Braff, D. L. (2001). Neural circuit regulation of prepulse inhibition of startle in the rat: current knowledge and future challenges. Psychopharmacology (Berl.) 156, 194-215.

Swerdlow, N. R., Shoemaker, J. M., Crain, S., Goins, J., Onozuka, K. and Auerbach, P. P. (2004). Sensitivity to drug effects on prepulse inhibition in inbred and outbred rat strains. Pharmacol. Biochem. Behav. 77, 291-302.

Trenkwalder, C. and Paulus, W. (2010). Restless legs syndrome: pathophysiology, clinical presentation and management. Nat. Rev. Neurol. 6, 337-346.

Winkelmann, J., Schormair, B., Lichtner, P., Ripke, S., Xiong, L., Jailizadeh, S., Fulda, S., Pütz, B., Eckstein, G., Hauk, S. et al. (2007). Genome-wide association study of restless legs syndrome identifies common variants in three genomic regions. Nat. Genet. 39, 1000-1006.

Winkelmann, J., Czamara, D., Schormair, B., Knauf, F., Schulte, E. C., Trenkwalder, C., Dauvilliers, Y., Polo, O., Högl, B., Berger, K. et al. (2011). Genome-Wide association study identifies novel restless legs syndrome susceptibility loci on 2p14 and 16q12.1. PLoS Genet. 7, 1-10.

Xiong, L., Catoire, H., Dion, P., Gaspar, C., Lafreniere, R. G., Girard, S. L., Levchenko, A., Riviere, J. B., Fiori, L., St-Onge, J. et al. (2009). MEIS1 intronic risk haplotype associated with restless legs syndrome affects its mRNA and protein expression levels. Hum. Mol. Genet. 18, 1065-1074.

Zhang, X., Rowan, S., Yue, Y., Heaney, S., Pan, Y., Brendolan, A., Selleri, L. and Maas, R. L. (2006). Pax6 is regulated by Meis and Pbx homeoproteins during pancreatic development. Dev. Biol. 300, 748-757.

Zimprich, A., Garrett, L., Deussing, J. M., Wotjak, C. T., Fuchs, H., Gailus-Durner, V., de Angelis, M. H., Wurst, W., Höfle, S. M. (2014). A robust and reliable non-invasive test for stress responsivity in mice. Front. Behav. Neurosci. 8, 125.