Serum Resistin Levels in Adult Patients with Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-analysis

Dongli Han1#, Jie Chen2#, Shousheng Liu3, Zengzhi Zhang2, Zhenzhen Zhao3, Wenwen Jin1* and Yongning Xin1,4*

1Department of Infectious Disease, Qingdao Municipal Hospital, Dalian Medical University, Dalian, Liaoning, China; 2Qingdao City Shinan District Centers for Disease Control and Prevention, Qingdao, Shandong, China; 3Clinical Research Center, Qingdao Municipal Hospital, Qingdao, Shandong, China; 4Digestive Disease Key Laboratory of Qingdao, Qingdao, Shandong, China

Abstract

Background and Aims: Previous studies reported that serum resistin levels were remarkably changed in patients with nonalcoholic fatty liver disease (NAFLD) but the conclusions were inconsistent. The aim of this study was to investigate accurate serum resistin levels in adult patients with NAFLD. Methods: A complete literature research was conducted in the PubMed, Embase, and Cochrane Library databases, and all the available studies up to 7 May 2020 were reviewed. The pooled standardized mean difference (SMD) values were calculated to investigate the serum resistin levels in patients with NAFLD and healthy controls. Results: A total of 28 studies were included to investigate the serum resistin levels in patients with NAFLD. Patients with NAFLD had higher serum resistin levels than controls (SMD = 0.522, 95% confidence interval [CI]: 0.004–1.040, I² = 95.9%). Patients with nonalcoholic steatohepatitis (NASH) had lower serum resistin levels than the healthy controls (SMD = −0.44, 95% CI: −0.91–0.23, I² = 74.5%). In addition, no significant difference of serum resistin levels was observed between patients with NAFLD and healthy controls (SMD = −0.34, 95% CI: −0.91–0.23, I² = 79.6%) and between patients with NAFL and NASH (SMD = 0.15, 95% CI: −0.06–0.36, I² = 0.0%). Furthermore, subgroup and sensitivity analyses suggested that heterogeneity did not affect the results of meta-analysis. Conclusions: This meta-analysis investigated the serum resistin levels in adult patients with NAFLD comprehensively. Patients with NAFLD had higher serum resistin levels and patients with NASH had lower serum resistin levels than healthy controls. Serum resistin could serve as a potential biomarker to predict the development risk of NAFLD.

Citation of this article: Han D, Chen J, Liu S, Zhang Z, Zhao Z, Jin W, et al. Serum resistin levels in adult patients with nonalcoholic fatty liver disease: A systematic review and meta-analysis. J Clin Transl Hepatol 2021;9(4):484–493. doi: 10.14218/JCTH.2021.00018.

Introduction

Nonalcoholic fatty liver disease (NAFLD) is defined as hepatic steatosis by imaging or histology without secondary factors of hepatic fat aggregation, such as significant alcohol consumption and long-term use of a steatogenic medication.1 NAFLD ranges from nonalcoholic fatty liver (NAFL), which is characterized as simple benign hepatic steatosis, to nonalcoholic steatohepatitis (NASH), the histologic features of which are macrovesicular steatosis, hepatocellular ballooning, lobular inflammation, and pericellular fibrosis. NASH can progress to the more severe fibrosis, that is defined as the accumulation of extracellular matrix proteins in the liver interstitial space, cirrhosis, and even the hepatocellular carcinoma.2 Nowadays, NASH-associated cirrhosis has become the second leading cause for liver transplantation in the USA. Meanwhile, NAFLD increases the risk of developing type 2 diabetes, cardiovascular disease, and chronic kidney disease.3

NAFLD has been certainly become the most predominant chronic liver disease in the world, with the highly shocking prevalence of 25.24% among the global population. In fact, the prevalence is predicted to become even higher in the next decade.4 Up to now, the diagnostic golden standard for NAFLD is liver biopsy. As an invasive technology, some adverse events can occur during liver biopsy diagnosis of patients, such as pain, infection, bleeding and even death.5 Therefore, there is an urgent need to develop a novel biomarker to predict and diagnose NAFLD conveniently and accurately.

Resistin belongs to the family of resistin-like molecules, also known as “found in inflammatory zone” (FIIZ), and functions as a pro-inflammatory adipokine.6 Resistin is mainly produced by adipose tissue, inflammatory cells, such as macrophages and monocytes, and hepatic stellate cells.7 Previous reports have suggested that resistin could be up-regulated by proinflammatory cytokines, including TNF-α, IL-6, IL-1β. In turn, resistin can activate the nuclear factor-kappa B (NF-κB) signaling pathway and promote the synthesis of TNF-α, IL-6 and other pro-inflammatory agents.8 Regarding the association of serum resistin levels in pa-
tients with NAFLD, the studies showed conflicting results so far. Some researchers have reported that serum resistin levels are high in patients with NAFLD, NAFL, and NASH compared to healthy subjects. However, other researchers have suggested that no significant difference exists for serum resistin levels in patients with NAFLD, NAFL, NASH, and healthy controls. In the comparison between patients with NASH and NAFL, some studies have found higher serum resistin levels in patients with NASH, whereas others found similar levels of serum resistin in patients with NASH and NAFL. Meanwhile, some researchers have reported lower serum resistin levels in patients with NASH compared to patients with NAFL or healthy controls.

In consideration of the inconsistence of serum resistin levels in patients with NAFLD, it is worthwhile to investigate the exact performance of serum resistin levels in patients with NAFLD according to the available studies. The aim of this study was to conduct a systematic review of the available studies and comprehensively analyze the relationship between serum resistin levels and the degree of NAFLD.

Methods

Search strategy

To obtain the relevant studies for this meta-analysis, a complete literature search was conducted in the databases of PubMed, Embase, and Cochrane Library by the following strategy: ((((((((Nonalcoholic Fatty Liver Disease) OR Non alcoholic Fatty Liver Disease) OR NAFLD) OR Nonalcoholic Fatty Liver Disease) OR Fatty Liver, Nonalcoholic) OR Fatty Livers, Nonalcoholic) OR Liver, Nonalcoholic Fatty) OR Livers, Nonalcoholic Fatty) OR Nonalcoholic Fatty Liver Disease) OR Nonalcoholic Steatohepatitis) OR Nonalcoholic Steatohepatitides) OR Steatohepatitides, Nonalcoholic) OR Steatohepatitis, Nonalcoholic) AND (Resistin) OR Adipocyte Cysteine-Rich Secreted Protein FIIZ3) OR Adipocyte Cysteine Rich Secreted Protein FIIZ3. All the potentially relevant studies in English language and published before 7 May 2020 were reviewed. In case of data missed, we tried to contact the corresponding authors to obtain the original data.

Inclusion and exclusion criteria

Clinical studies which performed comparison of serum resistin levels between NAFLD (NAFL or NASH) patients and healthy controls were suitable for this meta-analysis. Studies were included if they conformed to the following criteria: (1) original full-text publications; (2) NAFLD diagnosed with biopsy, ultrasound, liver enzymes or computerized tomography; and (3) serum resistin levels compared. Studies were excluded according to the following principles: (1) patients with other causes of chronic liver disease (alcoholic fatty liver disease, viral or autoimmune hepatitis); (2) subjects included in more than one study; (3) some necessary data missing and not obtainable from the authors; (4) quality of publication too low; (5) reviews, editorials, case reports, letters, hypotheses, book chapters, studies on animals or cell lines, and unpublished data or abstracts; or (6) participants with other medical conditions, such as diabetes and coronary heart disease.

Data extraction and quality assessment

Two authors (HDL and CJ) evaluated each article and extracted the data independently. The controversy was solved by discussion with a third author (LSS). The study quality was evaluated using the Newcastle-Ottawa scale (NOS), as approved by the Cochrane Collaboration. The NOS uses a star system to decide the quality of a study in three realms: collection, comparability, and outcome/exposure. The NOS assigns four stars for selection, two stars for comparability, and three stars for outcome/exposure. Any study that received a score of 6 or more stars was regarded as being at low risk of bias (the highest quality), and lesser stars indicated a risk of bias.

Statistical analysis

The meta-analysis was conducted using Stata/SE 15.0. Serum resistin levels in the NAFLD group and controls were extracted as mean difference-standard deviation (SD) and the pooled values were expressed as standardized mean difference (SMD) with 95% confidence interval (CI). Forest plots were constructed to evaluate the heterogeneity of included studies by I² statistic. According to Higgins and Thompson, I² values of approximately 25% represented low heterogeneity, approximately 50% represented medium heterogeneity, and approximately 75% represented high heterogeneity. In this meta-analysis, continuous-weighted fixed-effects model analysis was used when the I²≤50%. Otherwise, the random-effects model was used. The possibility of publication bias was evaluated using funnel plot and the Egger’s regression asymmetry test. The sensitivity analysis, subgroup analysis, and meta-regression analysis were conducted to explore the possible sources of (expected) heterogeneity among the eligible studies. The GRADE approach was used to evaluate the quality of the pooled results of serum resistin levels in the NAFLD group vs. controls, NASH group vs. controls, NAFL group vs. controls, and NAFL group vs. NASH group.

Results

Characteristics of the included studies

According to the search strategy, a total of 448 studies were obtained ((PubMed (n=103), Cochrane (n=328), and Embase (n=13)). After removing 109 duplicates, 339 articles were retrieved. After removing reviews, conference abstracts, letters, editorials, conference papers, notes and short surveys, 159 potential studies were retrieved. After full-text evaluation, 28 studies were included eventually for this meta-analysis (Fig. 1). The main characteristics of the included studies are summarized in Table 1. All the included studies were cross-sectional or case-control studies. Patients with NAFLD in 22 studies9,10,12,15–33 were assessed by liver histology, and 5 studies34–38 evaluated NAFLD by ultrasonography. One study did not specifically describe the diagnosis of NAFLD. Among these studies, 10 were carried out in Asia, 6 in North America, and 10 in Europe. Two studies were carried out in South America. Among the 28 included studies, 25 had no the risk of bias and 3 had risk of bias.

Comparison of the serum resistin levels in NAFLD patients and controls

A total of 1,934 patients with NAFLD and 1,240 controls were included in this study. Only 18 of the included 28 studies investigated the serum resistin levels in NAFLD patients...
Han D. et al: Serum resistin in adult patients with NAFLD

(NAFLD patients were not divided into the NAFL or NASH) and healthy controls. Random-effects model was used to conduct the meta-analysis and the results showed that patients with NAFLD had higher serum resistin levels than controls (SMD=0.522, 95% CI: 0.004–1.040, $I^2=95.9\%$) (Fig. 2A). Ten studies investigated the serum resistin levels in patients with NASH and healthy controls. Random-effects model was used to conduct the meta-analysis and the results showed that patients with NASH had lower serum resistin levels than the healthy controls (SMD=−0.44, 95% CI: −0.83–0.55, $I^2=74.5\%$) (Fig. 2B). Seven studies investigated the serum resistin levels in patients with NAFL and healthy controls. Random-effects model was used to conduct the meta-analysis and no significant difference of serum resistin levels was observed between patients with NAFL and healthy controls (SMD=−0.34, 95% CI: −0.91–0.23, $I^2=79.6\%$) (Fig. 2C). Nine studies investigated the serum resistin levels in patients with NAFL and NASH. Fixed-effects model was used to conduct the meta-analysis and the results showed that there was no significant difference of serum resistin levels between patients with NAFL and NASH (SMD=0.15, 95% CI: −0.06–0.36, $I^2=0.00\%$) (Fig. 2D).

**Sensitivity and subgroup analyses**

In consideration of significant heterogeneity existing between the NASH group vs. controls, NAFL group vs. controls, and NAFL group vs. NASH group, sensitivity analysis was carried out to explore the possible sources of heterogeneity in the included studies. Each study was evaluated by exclusion in turn, and then the summarized SMD of the remaining studies were calculated. Only when the

---

**Fig. 1. Flow chart of the literature search process.**

Records identified through database searching = 448

292 articles excluded:
Duplicates = 109
Reviews = 120
Conference abstracts = 35
Letters = 11
Editorials = 9
Conference papers = 4
Notes = 3
Short survey = 1

Potentially relevant studies identified for evaluation = 156

110 articles excluded:
Without necessary data = 66
Gene polymorphisms = 21
Other liver diseases = 15
Cells or animals studies = 8

Eligible studies identified for evaluation = 46

18 articles excluded:
With other type of date = 10
Subjects were children = 6
Patient with DM = 2

Studies included in this meta-analysis = 28
| First author, Year | Group | n (M/F) | Age in years | BMI in kg/m² | Country | Study design | Diagnose of NAFLD | Biopsy on controls | Measurement method of resistin | NOS |
|--------------------|-------|---------|--------------|--------------|---------|--------------|-------------------|-------------------|-------------------------------|-----|
| Argentou et al. 2009<sup>10</sup> | Control | 9 (2/7) | 37.11±9.78 | 55.22±8.6 | Greece | Cross sectional | Liver biopsy | Yes | ELISA | 8 |
| | NAFLD | 41 (15/26) | 38.88±9.19 | 56.70±8.06 | | | | | | |
| | SS | 31 (9/22) | 38.06±9.23 | 56.27±8.45 | | | | | | |
| | NASH | 10 (6/4) | 41.04±9.07 | 58.02±6.99 | | | | | | |
| Auguet et al. 2013<sup>30</sup> | Control | 19 | 44.1±10.7 | 49.5±7.0 | Spain | Case-control | Liver biopsy | Yes | ELISA | 6 |
| | NAFLD | 69 | 46.79±10.3 | 48.2±6.6 | | | | | | |
| Auguet et al. 2014<sup>26</sup> | Control | 16 | 44±3.2 | 48.6±2.6 | Spain | Cross sectional | Liver biopsy | Yes | ELISA | 7 |
| | SS | 28 | 47.4±3.5 | 48.1±7.8 | | | | | | |
| | NASH | 28 | 45.9±1.4 | 47.5±5.4 | | | | | | |
| Bostrom et al. 2011<sup>19</sup> | Control | 40 (10/30) | 44 (24–67) | NA | Sweden | Case-control | Liver biopsy | No | ELISA | 4 |
| | NAFLD | 48 (24–65) | | | | | | | | |
| Pagano et al. 2006<sup>15</sup> | Control | 33 (30/3) | 42±3 | 26.9±1.0 | Italy | Case-control | Liver biopsy | No | ELISA | 8 |
| | NAFLD | 28 (26/2) | 45±2 | 27.3±0.6 | | | | | | |
| Cengiz et al. 2010<sup>18</sup> | Control | 24 | 38±10 | 25.6±1.1 | Turkey | Case-control | Liver biopsy | No | ELISA | 7 |
| | NAFLD | 39±9 | 30.1±4.5 | | | | | | | |
| Eminler et al. 2014<sup>21</sup> | Control | 40 (18/22) | NA | NA | Turkey | Case-control | Liver biopsy | No | ELISA | 6 |
| | NAFLD | 40 (21/19) | NA | | | | | | | |
| Floreani et al. 2008<sup>32</sup> | Control | 137 (12/125) | 60.2±10.4 | USA | Case-control | Liver biopsy | No | ELISA | 4 |
| | NASH | 30 (0/30) | 49.9±3.7 | | | | | | | |
| Musso et al. 2005<sup>28</sup> | Control | 25 (23/2) | 38±2 | 25.2±0.6 | Italy | Case-control | Liver biopsy | No | ELISA | 7 |
| | NAFLD | 25 (23/2) | 37±2 | 25.3±0.2 | | | | | | |
| Jarrar et al. 2008<sup>12</sup> | Control | 38 (5/33) | 40±9.5 | USA | Case-control | Liver biopsy | Yes | ELISA | 6 |
| | NAFLD | 45 (13/32) | NA | | | | | | | |
| | SS | 19 (2/17) | 37±9.2 | 47.2±7.5 | | | | | | |
| | NASH | 26 (11/15) | 43.9±11.4 | 47.5±8.3 | | | | | | |
| Jiang et al. 2009<sup>34</sup> | Control | 43 | 51.1±12.5 | 24.81±1.91 | China | Case-control | Ultrasound | No | ELISA | 7 |
| | NAFLD | 43 | 52.6±10.8 | 25.75±1.91 | | | | | | |
| Jamali et al. 2016<sup>31</sup> | Control | 18 (13/5) | 30.44±10.11 | 29.28±3.89 | Iran | Case-control | Liver biopsy | No | ELISA | 7 |
| | NAFLD | 18 (13/5) | 34.5±8.85 | 31.58±3.94 | | | | | | |
| Krawczyk et al. 2009<sup>33</sup> | Control | 16 | NA | 22.6±2.5 | Poland | Case-control | Liver biopsy | No | ELISA | 6 |
| | NAFLD | 18 (16/2) | 42.55±21 | 26.6±4 | | | | | | |
| Musso et al. 2013<sup>35</sup> | Control | 51 (33/18) | 56±1 | Italy | Cross sectional | Ultrasound | No | ELISA | 7 |
| | NAFLD | 161 (101/60) | 56±1 | 27.3±0.5 | | | | | | |

(continued)
| First author, Year | Group | n (M/F) | Age in years | BMI in kg/m² | Country | Study design | Diagnose of NAFLD | Biopsy on controls | Measurement method of resistin | NOS |
|-------------------|-------|---------|--------------|--------------|---------|-------------|-----------------|-------------------|------------------------|-----|
| Magalhaes et al. 2014<sup>26</sup> | Control | 36 | 37.9±1.3 | 36.7 (30.3–55.4) | Brazil | Cross sectional | Ultrasound | No | ELISA | 5 |
| Musso et al. 2017<sup>30</sup> | Control | 75 (61/14) | 50±1 | 25.9±0.2 | Italy | Cross sectional | Ultrasound | No | ELISA | 7 |
| Musso et al. 2012<sup>25</sup> | Control | 40 | 50±3 | 25.1±1.6 | USA | Case-control | Liver biopsy | No | ELISA | 7 |
| Perseghin et al. 2006<sup>39</sup> | Control | 47 (38/9) | 36±8 | 26.8±3 | USA | Case-control | NA | No | ELISA | 6 |
| Polyzos et al. 2016<sup>27</sup> | Control | 25 (5/20) | 53.6±1.8 | 30.5±0.8 | Greece | Cross sectional | Liver biopsy | No | ELISA | 7 |
| D’Incao et al. 2017<sup>29</sup> | Control | 4 | 38.5±10.85 | 49±6.73 | Brazil | Cross sectional | Liver biopsy | Yes | ELISA | 7 |
| Jamali et al. 2016<sup>23</sup> | SS | 2 (2/0) | 27±2.82 | 28.09±7.77 | Iran | Cross sectional | Liver biopsy | Yes | ELISA | 6 |
| Sanal et al. 2009<sup>17</sup> | Control | 18 | 44±8 | 43±14 | India | Case-control | Liver biopsy | No | ELISA | 6 |
| Senators et al. 2012<sup>9</sup> | Control | 66 (33/33) | 39±9 | 23±4 | Turkey | Case-control | Liver biopsy | No | ELISA | 7 |
| Shen et al. 2014<sup>22</sup> | Control | 43 (29/14) | 45±14 | 22±1.8 | China | Cross sectional | Liver biopsy | No | ELISA | 7 |
| Wong et al. 2006<sup>16</sup> | Control | 41 (17/24) | 42±10 | 24.1±6.8 | China | Case-control | Liver biopsy | No | ELISA | 7 |
| Younossi et al. 2011<sup>10</sup> | SS | 39 (3/36) | 40.51±10.28 | NA | USA | Cross sectional | Liver biopsy | Yes | ELISA | 7 |
| Zhu et al. 2016<sup>27</sup> | Control | 86 (57/29) | 52.9±13.07 | 22.86±2.94 | China | Cross-sectional | Ultrasound | No | ELISA | 7 |
| Younossi et al. 2008<sup>24</sup> | Control | 32 (13/19) | 39.3±9.8 | 47±9.1 | USA | Cross-sectional | Liver biopsy | Yes | ELISA | 7 |

Data are presented in numbers or mean±SD or medians and interquartile ranges. BMI, body mass index; ELISA, enzyme-linked immunosorbent assay; NA, not available.
study conducted by Polyzos et al. was removed, the heterogeneity was significantly reduced, which indicated that this study was the main source of heterogeneity. In order to investigate whether this study affected the results of the meta-analysis, the meta-analysis were reperformed after removal of the study (Polyzos et al., 2016) with the fixed-effects model. The results showed that patients with NASH had lower serum resistin levels than controls (SMD = -0.23, 95% CI: -0.43–0.04) (Fig. 3A); there was no significant difference of serum resistin levels between patients with NAFL vs. controls (SMD = 0.03, 95% CI: -0.24–0.29) (Fig. 3B), and between patients with NAFL vs. NASH patients (SMD = 0.14, 95% CI: -0.09–0.36) (Fig. 3C). These results indicated that the heterogeneity did not affect the results of the meta-analysis.

The same method was used to explore the source of heterogeneity in the meta-analysis of studies for NAFLD patients vs. controls, but no study was found to contribute to the heterogeneity. In addition, the subgroup analysis was conducted according to the diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores was entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Meta-regression and quality evaluation**

To further explore the source of heterogeneity between NAFLD and control groups, the effect of potential confounders were evaluated by meta-regression analysis (based upon random-effects) when ≥10 comparisons were available. Diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores were entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Fig. 2. Forest plots of serum resistin levels between (A) NAFLD patients vs. controls, (B) NASH patients vs. controls, (C) NAFL patient vs. controls, (D) NAFL patients vs. NASH patients.**

patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Meta-regression and quality evaluation**

To further explore the source of heterogeneity between NAFLD and control groups, the effect of potential confounders were evaluated by meta-regression analysis (based upon random-effects) when ≥10 comparisons were available. Diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores were entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Meta-regression and quality evaluation**

To further explore the source of heterogeneity between NAFLD and control groups, the effect of potential confounders were evaluated by meta-regression analysis (based upon random-effects) when ≥10 comparisons were available. Diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores were entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Meta-regression and quality evaluation**

To further explore the source of heterogeneity between NAFLD and control groups, the effect of potential confounders were evaluated by meta-regression analysis (based upon random-effects) when ≥10 comparisons were available. Diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores were entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.

**Meta-regression and quality evaluation**

To further explore the source of heterogeneity between NAFLD and control groups, the effect of potential confounders were evaluated by meta-regression analysis (based upon random-effects) when ≥10 comparisons were available. Diagnosis methods, ethnicity, mean age, types of study design, mean body mass index, biopsy on controls, and NOS scores were entered separately as covariates. As Table 2 shows, all of these factors failed to account for the heterogeneity between NAFLD and controls (Table 2). The GRADE approach was used to evaluate the quality of the evidence, and the results showed that the quality of results of serum resistin levels in NAFLD patients vs. controls was low, and moderate in NASH patients vs. controls, NAFL patient vs. controls, and NAFL patients vs. NASH patients. As Figure 4 shows, no obvious publication bias was observed.
Discussion

Resistin is a significant pro-inflammation adipokine and the role of its serum levels in patients with NAFLD remain controversial. This study systematically analyzed the serum levels of resistin in patients with NAFLD, especially in those with NAFL and NASH. The results suggested that patients with NAFLD had higher serum resistin levels than healthy controls, but low serum resistin levels were observed in patients with NASH when compared to healthy controls. In addition, no significant difference of serum resistin levels was observed between patients with NAFL and healthy controls, and between patients with NAFL and NASH. A reasonable explanation may be that all the patients with NASH and NAFL were diagnosed by liver biopsy, and patients with NAFLD were diagnosed by liver biopsy or ultrasound. The difference of diagnostic methods may contribute to these outcomes.

Some previous studies reported that serum levels of resistin in patients with NAFLD were higher,\textsuperscript{15} lower,\textsuperscript{39} or of no significant difference\textsuperscript{40} compared to healthy controls, accompanied by the different diagnosis methods used for NAFLD. Zhu et al.\textsuperscript{37} investigated the levels of serum protein as the diagnostic biomarkers for NAFLD, and they found that serum resistin was significantly higher in patients with NAFLD than in healthy controls. However, Magalhaes et al.\textsuperscript{36} investigated the serum levels of resistin in obese NAFLD patients and controls, but they found that the serum levels of resistin were negatively associated with the risk of NAFLD; that is, the serum resistin levels were low in NAFLD patients compared to controls. Except for the above reports, other research investigations also provided findings that precluded making a definitive conclusion. In this meta-analysis, we analyzed all the available studies which investigated the serum resistin levels in patients with NAFLD and controls, and we found that serum resistin levels were significant higher than in the healthy controls. Notably, all the patients with NAFLD were diagnosed by liver biopsy or ultrasound, and the NAFLD patients were not divided by NAFL and NASH stage. In consideration of the high heterogeneity in the meta-analysis, sensitivity analysis was conducted. Interestingly, when the study by Polyzos et al.\textsuperscript{27} (2016) was removed, the heterogeneity was markedly decreased, but the results of meta-analysis were unchanged. These results indicated that an individual study may contribute to the heterogeneity, but whether the results of meta-analysis were affected should be further investigated.

Resistin up-regulates the expression of proinflammatory cytokines such as TNF-\textalpha, IL-6, IL-12, and monocyte chemotactant protein-1 in monocytes, macrophages, and hepatic stellate cells via the NF-\textkappaB pathway.\textsuperscript{41} Serum resistin levels in patients with NASH and the association of serum resistin levels with the risk fibrosis remains inconsistent. Argentou et al.\textsuperscript{10} investigated the relationship of serum resistin...
in levels with some individual histopathological parameters, global activity grade, and fibrosis stage in NASH patients, but no significant association was observed. However, Tsochatzis et al.\(^4\) reported the serum levels in chronic hepatitis B and chronic hepatitis C patients, which suggested that serum resistin levels were negatively related to the degree of fibrosis. In this meta-analysis, the serum resistin levels in patients with NASH were significantly lower than in healthy controls, which was consistent with the previous study by Tsochatzis et al.\(^4\) to some degree. The probable reason may be that patients with NASH possess different degrees of fibrosis, usually, and the serum resistin levels could be negatively associated with the fibrosis. In this study, however, all the patients with NASH had NAFLD-related NASH, and the cause of fibrosis in NASH patients was different from that of the chronic hepatitis B/C patients. Whether the relationship of serum resistin levels with fibrosis was affected by the cause of fibrosis remains unknown and further studies are needed to clarify it.

Our results suggested that patients with NAFLD had higher serum resistin levels than healthy controls, but low serum resistin levels were observed in the patients with NASH compared to healthy controls. This is an interesting finding because resistin levels seem to rise with the progression of NAFLD, from healthy to NAFL, but decline when NAFL progresses to NASH. The same phenomenon occurred in patients with type 2 diabetes. In 2020, Galla et al.\(^4\) reported that patients with prediabetes had higher levels of resistin than patients with type 2 diabetes and healthy controls, as found in their 20-year follow-up study. In addition, a large number of cohort studies and meta-analysis suggested that resistin is a risk factor for cardiovascular disease.\(^4\) Acute coronary syndromes often occur in patients with high resistin levels, while chronic stable angina pectoris is more common in patients with low resistin levels.\(^4\) Given that pre-diabetes and coronary heart disease are a large part of the hidden population,\(^4\) patients with NAFLD are more likely to suffer from the type 2 diabetes and coronary heart disease, which may have affected the results of this study. In addition, whether reduced resistin levels will reduce the risk of NAFL, type 2 diabetes and coronary heart disease is unknown, and more research is needed in the future.

This meta-analysis has strengths and limitations that may have affected its conclusions. This is the first meta-analysis to systematically investigate the serum resistin levels in patients with NAFLD. The serum resistin levels were evaluated in patients with NAFLD, including patients with NAFL and NASH. In addition, this work is based on 28 high-quality studies. The limitations, however, include that some NAFLD patients were diagnosed by ultrasound other than liver biopsy in the included studies. Second, higher heterogeneity may disturb the accuracy of the results. Third, the association of serum resistin levels with fibrosis was not investigated in detail in this study. Fourth, although every
**Table 2. Meta regression analysis of possible sources of heterogeneity in NAFLD vs. control group (18 studies)**

| Effect size                  | Coefficient | Standard error | t     | p > | 95% CI         |
|------------------------------|-------------|----------------|-------|-----|----------------|
| Diagnosis methods            | 0.403       | 0.325          | −1.13 | 0.276 | 0.073–2.225    |
| Ethnicity                    | 0.175       | 0.223          | −1.58 | 0.133 | 0.175–1.287    |
| Types of study design        | 1.527       | 1.318          | 0.49  | 0.631 | 0.245–9.513    |
| Mean age (30–40, 40–50, ≥50) | 0.814       | 0.346          | −0.48 | 0.635 | 0.331–2.003    |
| Mean BMI (>30)               | 0.467       | 0.246          | −1.44 | 0.168 | 0.153–1.428    |
| Biopsy on controls           | 0.367       | 0.393          | −0.94 | 0.363 | 0.038–3.552    |
| NOS score                    | 1.996       | 0.783          | 1.76  | 0.097 | 0.869–4.586    |

step of this meta-analysis was carried out in strict accordance with the requirements, this meta-analysis was not registered on relevant websites in advance.

**Conclusions**

In summary, this study systematically investigated the serum resistin levels in adult patients with NAFLD for the first time. The results suggest that patients with NAFLD have higher serum resistin levels than healthy controls, but patients with NASH have lower serum resistin levels than healthy controls. In addition, no significant differences of serum resistin levels were observed between the patients with NAFL and controls, nor the patients with NAFL and NASH. Although a little incoherence between the results of this study and several previous studies existed, it remains reasonable to illustrate the variation of serum resistin levels in patients with NAFLD. In consideration of the present results, serum resistin possesses the potential to serve as a biomarker to predict the development risk of NAFLD, and the diagnostic sensitivity and specificity should be improved by excluding the interference of other factors. Further studies should be conducted to clarify the serum resistin levels in healthy controls and patients with NAFLD that is diagnosed by liver biopsy.

**Funding**

This study was supported by a grant from the National Natural Science Foundation of China (No. 31770837), which plays a significant role in the design of the study and data collection, analysis or interpretation, nor in the writing of the manuscript.

**Conflict of interest**

The authors have no conflict of interests related to this publication.

**Author contributions**

Study concept and design (YX, WJ), acquisition and analysis of data (DH, JC, SL, ZeZ, ZhZ), drafting and writing of the manuscript (DH, JC), and revision of the manuscript (YX, WJ). All authors approved the final manuscript.

**Data sharing statement**

All data are available upon request.

**References**

[1] Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, et al. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. Hepatology 2018;67(1):328–357. doi:10.1002/hep.29367.

[2] Sheka AC, Adely O, Thompson J, Harneed B, Crawford PA, Ikramuddin S. Nonalcoholic steatohepatitis: A review. JAMA 2020;323(12):1175–1183. doi:10.1001/jama.2020.2298.

[3] Wong VW, Chan WK, Chaitu-S, Chanl Y, Dan YY, Duseja A, et al. Asia-Pacific Working Party on Non-alcoholic Fatty Liver Disease guidelines 2017- Part 1: Definition, risk factors and assessment. J Gastroenterol Hepatol 2018;33(1):70–85. doi:10.1111/jgh.13857.

[4] Lonardo A, Byrne CD, Caldwell SH, Cortez-Pinto H, Targher G. Global epidemiology of nonalcoholic fatty liver disease: Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology 2016;64(4):1388–1389. doi:10.1002/hep.28584.

[5] Boyd A, Cain G, Chauhan A, Webb GJ. Medical liver biopsy: background, indications, procedure and histopathology. Frontline Gastroenterol 2020;11(1):40–47. doi:10.1136/fgastro-2018-101139.

[6] Polyszos SA, Kountouras J, Mantzoros CS. Adipokines and cytokines in non-alcoholic fatty liver disease. Aliment Pharmacol Ther 2008;27(5):412–421. doi:10.1111/j.1365-2036.2007.03582.x.

[7] Colica C, Atenovoli L. Resistin Levels in Non-alcoholic Fatty Liver Disease Pathogenesis. J Transl Med 2018;16(1):52–53. doi:10.2478/jtm-2018-0011.

[8] Shoelson SE, Herrero L, Naa A. Obesity, inflammation, and insulin resistance. Gastroenterology 2007;132(6):2169–2180. doi:10.1053/j.gastro.2007.03.059.

[9] Senates E, Colak Y, Yesil A, Coskunpinar E, Sahin O, Kahraman OT, et al. Circulating resistin is elevated in patients with non-alcoholic fatty liver disease and is associated with steatosis, portal inflammation, insulin resistance and nonalcoholic steatohepatitis scores. Minerva Med 2012;103(5):369–376.

[10] Argentino M, Tiriakos DG, Karanikolos M, Melachrinou M, Makri MG, Kittas C, et al. Adipokine serum levels are related to liver histology in severely obese patients undergoing bariatric surgery. Obes Surg 2009;19(9):1311–1323. doi:10.1007/s11695-009-9912-9.

[11] Westier E, Swan J, Sanderson S, Krishnan A, Watt K, Charlton M. Growth hormone, dehydroepiandrosterone and adipokine levels in non-alcoholic steatohepatitis: an endocrine signature for advanced fibrosis in obese patients. Liver Int 2012;32(2):272–286. doi:10.1111/j.1478-3231.2011.02637.x.

[12] Jarrar MH, Baranova A, Collantes R, Ranard B, Stepanova M, Bennett C, et al. Adipokines and cytokines in non-alcoholic fatty liver disease. Aliment Pharmacol Ther 2008;27(5):412–421. doi:10.1111/j.1365-2036.2007.03586.x.

[13] Fitzpatrick E, Dew TK, Quaglia A, Sherwood RA, Mityr RR, Dhawan A. Analysis of adipokine concentrations in paediatric non-alcoholic fatty liver disease. Pediatr Obes 2012;7(6):471–479. doi:10.1111/j.2047-6316.2012.00082.x.

[14] Weils G, Shea B, O’Connell D, Robertson J, Peterson J, Welch V, et al. The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta analyses. Available from: http://www3.med.unipmn.it/dispensa_etsm/2009-2010/Corso%20Perfezionamento%20ENBM_Faggiano/NOS_oxford.pdf.

[15] Pagano C, Soardo G, Hidon C, Mincio M, Basan L, Milan G, et al. The Newcaste-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta analyses. Available from: http://www3.med.unipmn.it/dispensa_etsm/2009-2010/Corso%20Perfezionamento%20ENBM_Faggiano/NOS_oxford.pdf.

[16] Pagano C, Soardo G, Pinol C, Milocco C, Basan L, Milan G, et al. Increased serum resistin in nonalcoholic fatty liver disease is related to liver disease severity and not to insulin resistance. J Clin Endocrinol Metab 2006;91(3):1081–1086. doi:10.1210/jc.2005-1056.

[17] Wong VW, Hui AY, Tsang SW, Chan JH, Tse AM, Chan KF, et al. Metabolic and adipokine profile of Chinese patients with nonalcoholic fatty liver disease. Clin Gastroenterol Hepatol 2006;4(9):1154–1161. doi:10.1016/j.cgh.2006.06.011.

[18] Sanal MG, Sarin SK. Serum adipokine profile in Indian men with nonalcoholic steatohepatitis: Serum adipokine is paradoxically decreased in lean versus obese patients. Diabetes Metab Syndr 2009;3(4):198–203. doi:10.1016/j.dsx.2009.07.012.

[19] Cengiz C, Ardiciglu B, Bulut S, Boyacioglu S. Serum retinol-binding protein 4 in patients with nonalcoholic fatty liver disease: does it have a significant
Han D. et al: Serum resistin in adult patients with NALFD

impact on pathogenesis? Eur J Gastroenterol Hepatol 2010;22(7):813-819. doi:10.1097/MEG.0b013e32833823bc.

[19] Boström ÉE, Ekstedt M, Kezagiti Y, Sjöwall C, Bokarewa MI, Almer S. Resistin is associated with breach of tolerance and anti-nuclear antibodies in patients with hepatobiliary inflammation. Scand J Immunol 2011;74(5):463–470. doi:10.1111/j.1365-3083.2011.02592.x.

[20] Augest T,terra X, Porras JA, Orellana-Gavalda JM, Martinez S, Aguilar C, et al. Plasma visfatin levels and gene expression in morbidly obese women with associated fatty liver disease. Clin Biochem 2013;46(3):202–208. doi:10.1016/j.clinbiochem.2012.11.006.

[21] Emirin AT, Aygün C, Konduk T, Kocaman O, Senturk O, Randhawa M, Afendy M, Stepanova M, Jamali R, Hatami N, Kosari F. The correlation between serum adipokines and liver damage in non-alcoholic fatty liver disease. J Res Med Sci 2014;19(11):1058–1061.

[22] Shen C, Zhao CY, Wang W, Wang YD, Sun H, Cao W, et al. Serum adipokine levels in patients with nonalcoholic fatty liver disease. BMC Gastroenterol 2014;14:39. doi:10.1186/1471-230X-2014-39.

[23] Jamali R, Hatami N, Kosari F. The correlation between serum adipokines and liver damage in non-alcoholic fatty liver disease. Hepat Mon 2016;16(5):e37412. doi:10.5812/hepatmon.37412.

[24] Younossi ZM, Jarrar M, Nugent C, Randhawa M, Afendy M, Stepanova M, et al. The relationship between resistin and ghrelin levels with fibrosis in nonalcoholic fatty liver disease. J Clin Biochem 2013;46(3):202–208. doi:10.1016/j.clinbiochem.2012.11.006.

[25] Younossi ZM, Page S, Rafiq N, Birerdinc A, Stepanova M, Hossain N, et al. Plasma visfatin levels and gene expression in morbidly obese women with associated fatty liver disease. Obes Surg 2017;27(8):2151–2158. doi:10.1007/s11695-017-2627-4.

[26] Magalhães GC, Feitoza FM, Moreira SB, Carmo AV, Souto FJ, Reis SR, et al. Hypoadiponectinemia in nonalcoholic fatty liver disease obese women is associated with infrequent intake of dietary sucrose and fatty foods. J Hum Nutr Diet 2014;27(Suppl 2):301–312. doi:10.1111/jhn.12110.

[27] Zhu JZ, Zhu HT, Dai YN, Li CX, Fang ZY, Zhao DJ, et al. Serum resistin is a potential biomarker for non-alcoholic fatty liver disease: a case-control study. Endocrine 2016;51(1):91–100. doi:10.1007/s12020-015-0735-2.

[28] Musso G, Cassader M, De Michieli F, Paschetta E, Pinach S, Saba F, et al. MERTK rs4374383 variant predicts incident nonalcoholic fatty liver disease and diabetes: role of mononuclear cell activation and adipokine response to dietary fat. Hum Mol Genet 2017;26(9):1747–1758. doi:10.1093/hmg/ddw400.

[29] Persseghin L, Lattuada G, De Cobelli F, Nati G, Esposito A, Bursa K, et al. Serum resistin and hepatic fat content in nondiabetic individuals. J Clin Endocrinol Metab 2006;91(12):5122–5125. doi:10.1210/jc.2006-1368.

[30] Cho YK, Lee WY, Oh SY, Park JH, Kim JH, Park DI, et al. Factors affecting the serum levels of adipokines in Korean male patients with nonalcoholic fatty liver disease. Hepatogastroenterology 2007;54(77):1512–1516.

[31] Andjelkovic K, Jelic-Ivanovic Z. Circulating resistin protein and mRNA concentrations in patients with NASH: postprandial lipid metabolism as a link between adiponectin and liver histology findings in non-alcoholic fatty liver disease. World J Gastroenterol 2015;21(2):242–251. doi:10.3748/wjg.v21.i2.242.

[32] Dragoa AJ, Litiulo A, Kiviniemi H, Kesäniemi YA, Ukkola O. Peptide hormones and risk for future cardiovascular events among prediabetics: a 20-year follow-up in the OPERA study. Ann Med 2020;52(3-4):85–93. doi:10.1080/00365520.2020.1741673.