Structural Diversity and Role of Phytochemicals against P38-α Mitogen-activated Protein Kinase and Epidermal Growth Factor Receptor Kinase Domain: A Privileged Computational Approach

Amar Shankar1, S.M. Gopinath2, Shiva Prasad Kollur3, P. Sushma4, Anisha S. Jain5, Sharanagouda S. Patil6, Chandrashekar Srinivasa2, Shivalingaiah7 and Chandan Shivamallu4*∗

1Department of Food Technology, School of Engineering and Technology, JAIN (Deemed-to-be University), Bangalore - 562 112, Karnataka, India.
2Department of Studies in Biotechnology, Davangere University, Shivangotri, Davangere – 577002, Karnataka, India.
3Department of Sciences, Amrita School of Arts and Sciences, Mysuru, Amrita Vishwa Vidyapeetham – 570 026, Karnataka, India.
4Department of Biotechnology and Bioinformatics, School of Life Sciences, JSS Academy of Higher Education and Research, Mysuru - 570015, Karnataka, India.
5Department of Microbiology, School of Life Sciences, JSS Academy of Higher Education and Research, Mysuru - 570015, Karnataka, India.
6ICAR-National Institute of Veterinary Epidemiology and Disease Informatics (NIVEDI), Yelahanka, Bengaluru -560064, Karnataka, India.
7Department of Botany, Maharani's Science College for Women, Mysuru- 570005, India.

Abstract

Computational databases and tools in recent times have been proved to provide an essential aid for anticancer studies in the field of oncology. Molecular docking studies facilitate the structural diversity of plant-derived phytomolecules having anticancer properties against receptor proteins involved in cancer signaling pathways. The current study involves the investigation of phytocompounds-agasthisflavone, anacardic acid, zaopatanolide A, a purified product of the plant extract Amarogopinois546 were subjected to docking studies on p38-α MAPK and EGFR Kinase domain. The effectiveness of this study was evaluated by comparing the docking interactions of a standard drug, doxorubicin against the receptor molecules. The docking study is analyzed by compound estimated with lowest binding energy is considered to have the highest affinity towards the active site of the receptor proteins. The isolated plant compound Amarogopinois546 displayed the least binding score with a large number of hydrogen bonds and hydrophobic interactions towards the P38α MAP kinase receptor in comparison with the EGFR kinase domain. This preliminary result can strongly be supported for carrying out experimental evaluation in near future.

Keywords: Phytomolecules, Docking, Binding Energy, Hydrophobic, MAPK, EGFR

*Correspondence: chandans@jssuni.edu.in; +91 9538500423 (Received: May 28, 2021; accepted: August 20, 2021)
INTRODUCTION

Ayurveda is a sacred precept of rule or commandments which guides us about all aspects of life and it is considered to be the oldest healing science in India. Anacardium occidentale commonly known as Cashew belongs to the family Anacardiaceae considered as a multi-purpose tree since the leaves and the bark of the tree are of therapeutic importance and the fruit is an edible resource of high value. Anacardium occidentale native of Brazil is an all-purpose tree that is widely distributed in India, Brazil, Africa, Mexico, Malaysia, and Turkey. Vitamin B2 and B3, vitamin E, vitamin C, pantothenic acid and magnesium are all abundant in cashews. Zoapatanolide A, Agasthisflavone, and anacardic acid are the biological components identified in substantial quantities in the powdered leaf extract and Cashew nutshell liquid. These phytochemicals were checked for their in-silico anticancer activity. Molecular Docking is a computational in-silico approach that helps in predicting the preferred orientation of a molecule (ligand) when it is made to interact at the binding pocket of a macromolecule (enzyme/protein). This knowledge of orientation prediction may in turn help in predicting the binding affinity or the strength of association between two molecules. The in-silico molecular docking has become an efficient technique in the field of drug discovery for structure-based medication disclosure. The most popular and commonly used software for molecular docking purposes is AutoDock (v.4.0). It is freely available open-source software for virtual screening of small molecules to receptors and computational docking. To obtain preliminary data, docking studies were conducted using the screened phytocompounds as ligands and the target molecules p38-MAPK (p38-mitogen-activated protein kinase) and EGFR (Epidermal growth factor receptor) Kinase domain (3W2S). The target protein, p38 mitogen-activated protein kinase belongs to serine/threonine protein kinases. The human p38 MAP kinase is classified into four isoforms: p38α, p38β, p38γ and p38δ, with p38α being the most well-studied of the four. Phosphorylation of nuclear and cytoplasmic targets is carried out by the p38 MAP kinase. p38 MAP kinase is a signaling pathway that regulates a complex network of proteins involved in cellular activity, cell senescence, cell cycle arrest, cell differentiation, tumor suppression, apoptosis and cytokine synthesis.

MATERIALS and METHODS

Protein preparation

The structure of proteins, 4FA2 (mitogen-activated protein kinase) and 3W2S (epidermal growth factor receptor) were obtained from Protein Data Bank (PDB) and proteins devoid of the ligand is represented in Fig. 1. The protein structures were validated through the Ramachandran plot depicted in Fig. 2. Only structures which had at least ~96% of residues in favored regions and ~2% of residues in allowed regions were fit for molecular interaction studies. Ramachandran plot prediction for the structure of

Fig. 1. 3D structure of P38α MAP kinase (4FA2) and EGFR Kinase domain (3W2S).
p38α MAP kinase protein showed 97.9% residues in favored regions and for EGFR Kinase domain protein 97.7% residues in favored regions.

**Ligand Preparation**

The structure of agasthisflavone, anacardic acid, zoapatanolide A and standard drug doxorubicin were obtained from Pubchem. The obtained SDF format of the above-mentioned ligands was converted to .pdb format using Open Babel (2.4.0) by adding hydrogens and generating 3D coordinates. The purified product of the plant extract Amarogopinois was drawn using Chemsketch and saved in .mdl format which was further converted into .pdb format. The geometry of all these PDB structures was cleaned and refined using Argus Lab (4.0.1). The validated structure which obeyed structure-activity relationships (SAR) was selected and the structures of agasthisflavone, anacardic acid, zoapatanolide A, doxorubicin and the Amarogopinois are represented in Fig. 3.

The binding residues for the proteins P38α MAP kinase (4FA2) and EGFR Kinase domain (3W2S) were predicted using the GalaxySite tool of GalaxyWEB server. Each protein showed the involvement of more than 6 residues in the formation of the active site (Table 1).

**Molecular Docking**

The predicted protein structures docked using Autodock suite (v.4.0). Autodock is a set of automated docking tools, which predicts the interactions between a small molecule and its receptor proteins, which proposes several binding modes of a docked complex. It employs the Lamarckian genetic algorithm, with Gasteiger performing the charge computations. All of the chosen ligands were docked using a Genetic algorithm, in which the ligands were subjected to conformational modifications to find the lowest energy conformation, considered as the most stable structure. The binding modes with the lowest energies were screened and grouped. The chosen compounds were docked against the target proteins to identify and evaluate the binding affinity of the docked complexes. The molecular docking complexes were further visualized using the UCSF Chimera visualization tool to manually pick the best feasible docking pose.

**Table 1.** The details of amino acid residues of both 4FA2 and 3W2S

| No. | Protein PDB ID | Binding site residues |
|-----|----------------|-----------------------|
| 1   | 4FA2           | 30V, 38V, 53K, 71E, 75L, 84I, 104L, 106T, 109M, 110G, 111A, 167L, 168D, 169F |
| 2   | 3W2S           | 718L, 726V, 743A, 745K, 795F, 796G, 797C, 800D, 801Y, 841R, 842N, 844L, 854T, 997F |

**Fig. 2.** Ramachandran plot prediction for structure 4FA2 and 3W2S.
RESULTS AND DISCUSSION

The molecular interaction studies performed through molecular docking using Autodock (4.2.6) of agathisflavone, anacardic acid, zoapatanolide A, doxorubicin and the standard drug Amarogopinois546 against the P38α MAP kinase (4FA2) and EGFR Kinase domain (3W2S) exhibited binding affinity and molecular interactions with proteins (Table 3 and 4). Whereas, binding affinity towards P38α MAP kinase (4FA2) showed

![Image of molecules A to E](image-url)

**Fig. 3.** 3D structure of A: Agathisflavone, B: Anacardic acid, C: Zoapatanolide A, D: Doxorubicin and E: Amarogopinois546.

**Table 2.** Residues of the target molecule p38α MAP kinase interacting with the three ligands: Zoapatanolide A, Agathisflavone and Anacardic acid, a standard drug Doxorubicin and Amarogopinois546.

| Name of the Biomarker | Ligands | H-bond interaction No. of bonds | Amino acid Residues (Hydrophobic interactions) |
|-----------------------|---------|---------------------------------|-----------------------------------------------|
| Agathisflavone        | 4       | Ala 157, Leu 156, Asn 155, Pro 153, Asp 112, Ser 154, Gly 170, Leu 171, Leu 104, Lys 53, Val 52, Val 38, Gly 31, Val 30, Ser 32 |
| Anacardic acid        | 1       | UNL                             | Leu 74, Leu 75, Ile 84, Ile 166, Leu 167, Asp 168, Phe 169, Glu 71, Thr 106, Ala 157 Ser 154, Lys 152, Asn 155, Leu 171 |
| Zoapatanolide A       | 2       | UNL                             | Phe 169, Gly 170, Lys 53, Leu 171, Gly 33, Ser 32, Ser 154, Gly 31, Val 163, Lys 162 |
| Doxorubicin           | 4       | UNL                             | Thr 106, His 107, Leu 108, Ile 84, Lys 165, Gly 110, Leu 167, Val 158, Ala 157, Leu 156, Ala 111, Asn 155, Ser 154, Pro 153, Asp 112, Leu 113, Lys 156, Leu 171, Gly 33, Thr 106, His 107, Leu 108, Ile 84, Lys 165, Gly 110, Leu 167, Val 158, Ala 157, Leu 156, Ala 111, Asn 155, Ser 154, Pro 153, Asp 112, Leu 113, Lys 156, Leu 171, Gly 33, Ser 32 |
| Amarogopinois546      | 6       | UNL                             | Thr 106, His 107, Leu 108, Ile 84, Lys 165, Gly 110, Leu 167, Val 158, Ala 157, Leu 156, Ala 111, Asn 155, Ser 154, Pro 153, Asp 112, Leu 113, Lys 156, Leu 171, Gly 33, Thr 106, His 107, Leu 108, Ile 84, Lys 165, Gly 110, Leu 167, Val 158, Ala 157, Leu 156, Ala 111, Asn 155, Ser 154, Pro 153, Asp 112, Leu 113, Lys 156, Leu 171, Gly 33, Ser 32 |

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Table 3. Residues of the target molecule EGFR Kinase domain interacting with Zoapatanolide A, Agasthisflavone, Anacardic acid, a standard drug Doxorubicin and Amarogopinois546

| Name of the Biomarker | Ligands         | H-bond interaction | Amino acid residues of target proteins (Hydrophobic interactions) |
|-----------------------|-----------------|--------------------|----------------------------------------------------------------|
|                       |                 | No. of bonds       | Residues                                                       |
| Agasthisflavone       | 4               | UNL                | Ala 157, Leu 156, Asn 155, Pro 153, Asp 112, Ser 154, Gly 170, Leu 171, Leu 104, Lys 53, Val 52, Val 38, Gly 31, Val 30, Ser 32 |
|                       |                 | UNL                | Leu 74, Leu 75, Ile 84, Ile 166, Leu 167, Asp 168, Phe 169, Glu 71, Thr 106, Ala 157 |
| Anacardic acid        | 3               | Ala 722            | Ser 154, Lys 152, Asn 155, Leu 171 |
| EGFR Kinase domain    | 2               | Gly 724            | Ser 154, Lys 152, Asn 155, Leu 171 |
| (3w2s)                |                 | Gly 724            | Ser 154, Lys 152, Asn 155, Leu 171 |
| Doxorubicin (standard | 4               | UNL                | Phe 169, Gly 170, Lys 53, Leu 171, Gly 33, Ser 32, Ser 154, Gly 31, Val 163, Lys 162 |
| drug)                 |                 | UNL                | Leu 718, Gly 719, Ser 720, Val 726, Phe 997, Met 793, Gly 796, Asp 800, Arg 803, Leu 1001, Leu 844, Lys 745, Asp 855 |
| Amarogopinois546      | 5               | LYS 745            | Leu 718, Gly 719, Ser 720, Val 726, Phe 997, Met 793, Gly 796, Asp 800, Arg 803, Leu 1001, Leu 844, Lys 745, Asp 855 |

Fig. 4. Docked pose of the ligand shown with p38α MAP kinase protein and Hydrophobic interactions displayed within the solid surface of the docked complex between the binding residues of the p38α MAP kinase receptor and the small molecule A: Agasthisflavone, B: Anacardic acid, C: Zoapatanolide A, D: Doxorubicin and E: Amarogopinois546.
Ligands docked with EW2S

Fig. 5. Docked pose of the ligand shown with EGFR Kinase protein and the binding pocket of the EGFR protein fits the ligand by interactions and bonds A: Agasthisflavone, B: Anacardic acid, C: Zoapatanolide A, D: Doxorubicin and E: Amarogopinois546

CONCLUSION

The in silico studies were designed by selecting screened phytocompounds as ligands and target proteins as p38-α MAPK (p38-α mitogen-activated protein kinase) and EGFR Kinase domain (3W2S). The selected ligands were compared with the isolated compound Amarogopinois546 and standard drug doxorubicin. The objective concluded that Amarogopinois546 showed a high binding affinity towards the P38α MAP kinase receptor (4FA2) when compared to the EGFR Kinase domain (3W2S). Thereby concluding all the predefined objectives of this investigation with the following future aspects.

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CONFLICT OF INTEREST
The authors declare that there is no conflict of interest.

AUTHORS’ CONTRIBUTION
AS, GSM, SP, ASJ, SSP, CS, and CS conceptualized and gathered the data with regard to this work. AS, CS and SP gave the necessary inputs towards the designing of the manuscript.

FUNDING
None.

DATA AVAILABILITY
All datasets generated or analyzed during this study are included in the manuscript.

ETHICS STATEMENT
Not applicable.

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