Bulk Modification of Poly(lactide) (PLA) via Copolymerization with Poly(propylene glycol) Diglycidylether (PPGDGE)

Sandra Castillejos 1, Jorge Cerna 1,*†, Francisco Meléndez 1, María Eugenia Castro 2, Rocío Aguilar 1, César Márquez-Beltrán 3 and Maykel González 4

1 Facultad de Ciencias Químicas, Benemérita Universidad Autónoma de Puebla, Ciudad Universitaria, Puebla 72570, Mexico; sandra.castillejos@alumno.buap.mx (S.C.); francisco.melendez@correo.buap.mx (F.M.); raguilar@ifuap.buap.mx (R.A.)
2 Instituto de Ciencias de la Universidad Autónoma de Puebla, Benemérita Universidad Autónoma de Puebla, Ciudad Universitaria, Puebla 72570, Mexico; mareug.castro@correo.buap.mx
3 Instituto de Física, Benemérita Universidad Autónoma de Puebla, Ciudad Universitaria, Puebla 72570, Mexico; cmarquez@ifuap.buap.mx
4 Tecnológico de Monterrey, Escuela de Ingeniería y Ciencias, Campus Puebla, Puebla 72453, Mexico; mkgc@gmail.com
* Correspondence: jorge.cerna@correo.buap.mx; Tel.: +52-1-(222)-229-5500 (ext. 7390)

Received: 2 October 2018; Accepted: 18 October 2018; Published: 24 October 2018

Abstract: Copolymers of L-lactide and poly(propylene glycol) diglycidyl ether (PPGDGE380) were synthesized by ring opening polymerization (ROP). Stannous octoate was used as the catalyst and 1-dodecanol as the initiator. The effect of the variables on the thermal properties of the copolymers was investigated by differential scanning calorimetry (DSC). Contact angle measurements were made in order to study the wettability of the synthesized copolymers. The copolymers differed widely in their physical characteristics, ranging from weak elastomers to tougher thermoplastics, according to the ratio of L-lactide and PPGDGE380. The results showed that the copolymers were more hydrophilic than neat Poly(lactide) (PLA) and the monomer ratio had a strong influence on the hydrophilic properties.

Keywords: Poly(lactide); copolymerization; PPGDGE; contact angle; hydrophilic

1. Introduction

Recently, the increasing emphasis on sustainability in the polymer industry has become a necessity and the reason why great efforts have been made to replace existing polymers with biodegradable and biorenewable alternatives [1–5]. The most representative thermoplastic in this area is poly(lactic acid) (PLA), a bio-based polyester derived from lactic acid or lactide that is biodegradable, recyclable and compostable [6–8].

The PLA homopolymer has physical properties that make it suitable for a wide range of applications. It has good mechanical properties, thermal stability, transparency, gas impermeability and is easy to process. Its range extends from commodity [9–13] to biomedical [14–18] and pharmaceutical ones [19–21].

However, it has drawbacks that limit its use, such as brittleness, poor melt strength, low heat deflection temperature (HDT), narrow processing and low thermal stability. In some areas, it is limited by its hydrophobicity and degradation rates. The most common strategies to enhance the versatility of PLA and overcome these limitations is through bulk or surface modifications. By following these strategies, the resulting copolymer has shown improved strength and toughness, in addition...
to controlled degradable and hydrophilic properties. It is also possible to obtain numerous new
copolymers with different macromolecular architectures (linear, branch, star and dendron) [22–25].
In recent years many studies have been dedicated to the copolymerization of PLA with more
hydrophilic monomers or polymers due to its high demand in the biomedicine industry. In this field,
one important technology is 3D printing. PLA and its copolymers have proven to be easy to process
and create complex 3D structures [26–28]. Copolymers such as poly(lactic acid-co-caprolactone)
(PLCL) [29–33], poly(lactic acid-glycolic acid) (PLGA) [34–36] and poly(lactic acid-glutamic acid)
(PLGM) [37–39] have been developed. The purpose of the present work is to synthesize a new
copolymer of lactide and poly(propylene glycol) diglycidyl ether (PPGDE) in order to improve the
hydrophilic properties of PLA. We explore these properties through measuring the contact angle.
Further, we aim to develop a biomaterial with an important building block that could help to tailor
different structures and properties.

2. Materials and Methods

2.1. Materials

The L-lactide (98%, L-LA) (Scheme 1a) was purchased from Corbion (Amsterdam, The Netherlands),
and was purified by recrystallization from toluene. Poly(propylene glycol) diglycidyl ether (PPGDE380)
(Scheme 1b), tin(II) 2-ethylhexanoate (Sn(Oct)2) and 1-dodecanol were purchased from Aldrich (Saint Louis,
MS, USA); and were used as-received. Toluene, acetone and methanol were purchased from J.T. Baker
(Mexico City, Mexico); reactive grade.

Scheme 1. Structures of (a) L-Lactide and (b) Poly(propylene glycol)diglycidyl ether.

2.2. Copolymerization

The general process for the bulk copolymerization was as follows: We add the corresponding
concentration of monomer ratio (L-Lactide/PPGDE380) into a glass ampule, then, the catalyst
(Sn(Oct)2) and the initiator (1-dodecanol) were added to the glass ampule and it was sealed. The vial
was placed in a benchtop muffle furnace for a period of two hours. Once the reaction period was
over, the resulting copolymers were kept in a vacuum chamber at 50 ˚C for four days to evaporate
monomer residues. In the first set of experiments, different concentrations of monomer ratio were
used (90/10, 80/20 and 70/30 wt % in feed). In the second set of experiments the monomer ratio
concentration was kept constant and the catalyst/initiator ratio was varied. The cat/ini ratio was
varied as follows: 0.01/0.1, 0.01/0.15, 0.01/0.2, 0.03/0.15, 0.05/0.15 wt %. The temperature used
was 190 ˚C and the reaction period was set in two hours. For convenience and accuracy in weighing
Sn(Oct)2 and 1-dodecanol were prepared as a 1 M and 3 M solution in toluene, respectively.

2.3. Characterization

The chemical structures of the copolymers were analyzed by Fourier transform infrared (FTIR)
spectra and by 1H NMR spectrometry. To obtain the FTIR spectra to the investigated copolymers in
the mid IR region (wavelength range: 400–4000 cm⁻¹) an Excalibur FTS 3000 spectrometer provided
by Mettler Toledo (Columbus, OH, USA); was used. The attenuated total reflectance (ATR) was used,
utilizing a ZnSe crystal. The NMR spectroscopy was obtained from a Bruker of 500 MHz spectrometer.
Deuterated chloroform (CDCl$_3$) was used as the solvent at room temperature. Thermal properties were obtained using a TA Q100 DSC calorimeter (TA Instruments-Waters LLC, Colonia Acacias, Mexico). The device for the contact angle measurements was designed at the university, and consisted of a digital camera (Nikon, Melville, NY, USA), holder for the sample, a Gastight Glass Syringe to deposit a 12 µL drop of water, a UV light source that emits at 365 nm and a visible radiation lamp emitting at 580 nm. The angle was measured through a Java-based image processing program ImageJ, developed at the National Institutes of Health and the Laboratory for Optical and Computational Instrumentation, Wisconsin, United States. The calibration process was made using a glass slide and a water drop deposited in the surface, with any film. Copolymer films were prepared, to change the contact angle of the glass slide, by dissolving the samples into toluene and allowing it to evaporate in a desiccator to obtain a homogeneous film. The contact angle measurements were made using the static sessile drop method.

This method is a static contact angle measurement method which consists of putting down a liquid drop on the solid plate surface we want to characterize, then measuring the contact angle made by the drop on this surface. Indeed, when a drop of liquid is deposited down on a solid surface, three phases occur: solid, liquid and gas (Figure 1).

![Figure 1. Static contact angle measurement with the sessile drop method.](image)

The drop’s profile is being changed depending on the physicochemical characters of the solid surface, on the adhesion forces newly created at the interface solid/liquid and on the cohesion forces of the liquid. This change will affect the contact angle value revealing the surface state (hydrophobic or hydrophilic, rough or smooth, homogeneous or heterogeneous) and the different forces occurred are linked together according to Young’s Equation (1):

$$\gamma_{sv} = \gamma_{sl} + \gamma_{lv} \cos \theta$$

where $\gamma_{sv}$, $\gamma_{sl}$ and $\gamma_{lv}$ represent the “surface tensions” of the interface solid/gas, solid/liquid and liquid/gas, respectively, and $\theta$ represents the contact angle.

### 3. Results

The PLA-PPGDGE copolymers were synthesized by the ring opening polymerization of successively added PPGDGE$_{Mn=380}$ and L-lactide in the presence of 1-dodecanol, using tin(II) 2-ethylhexanoate as a catalyst. These copolymers differ greatly in appearance, from plastic like to gummy according to the monomer composition. The conversion of the polymerization was about 88–92%, with the residual monomer mainly L-lactide. These conversion measurements were based on monomer evaporation from the polymerized samples in a vacuum chamber.

To examine the existence of the copolymer, FTIR experiments were performed and compared with those of the neat PLA and PPGDGE. Figure 2 shows the FTIR spectra of PLA, PPGDGE and the PLA-co-PPGDGE copolymer. For neat PLA, the peaks of interest are the C=O stretching at 1745 cm$^{-1}$ and the O–C=O stretching at 1185 cm$^{-1}$ which correspond to the characteristic ester bonds. It is possible to observe the C–O–C symmetric and symmetric band in the range of 1079–1181 cm$^{-1}$, the symmetric bending of the C–H group at 1360 cm$^{-1}$, the asymmetric bending of –CH$_3$ at 1451 cm$^{-1}$ and the symmetric and asymmetric bands to the –C–H stretching vibrations of CH$_3$ groups in the side chain at 2992 and 2948 cm$^{-1}$. For the plain PPGDGE there are three peaks observed at 907, 839 and 756 cm$^{-1}$.
cm$^{-1}$ corresponding to symmetric and asymmetric stretches of the epoxide ring [40] and the O-H stretching band at 3500 cm$^{-1}$. The synthesis of the copolymer was confirmed by the disappearance of the epoxide ring bands in the copolymer spectra. It is possible for the formation of cross-linking and chain branching formation between diepoxy groups with both the end carboxyl and hydroxyl groups of the PLLA copolymer to produce primary ester and ether bonds, however this band was overlapped by the C–O–C asymmetric and symmetric band corresponding of PLA (1079–1181 cm$^{-1}$). The spectra of the PLA base copolymer correspond to a copolymer with a monomer ratio of PLA/PPGDGE 80/20.

Figure 2. FTIR spectra of: PLA, PPGDGE, PLA and PLLA-co-PPGDGE. Copolymer ratio PLA/PPGDGE 80/20.

For the second set of experiments, it was necessary to keep the monomer ratio constant. The FTIR spectrums of the copolymers with different monomer ratios were analyzed (Figure 3). The spectra of the 90/10 (l-lactide/PPGDGE wt %) copolymer presents some unreacted PPGDGE because the symmetric and asymmetric stretches of the epoxide ring are present (907, 839, 756 cm$^{-1}$) and the broad band of the O–H stretching of the hydroxyl groups at 3500 cm$^{-1}$ can be seen. With 80/20 and 70/30 these bands completely disappear due to a complete reaction of the epoxy groups present in the PPGDGE. In appearance, the 70/30 (l-lactide/PPGDGE) copolymer was rubberier than the 80/20, because of the inclusion of more PPDGE chains into the copolymer. Based on this evidence, the monomer ratio chosen to carry on the second set of experiments was the 80/20 (l-lactide/PPGDGE).

The $^1$H NMR spectrum was taken in deuterated chloroform. It shows the typical signals of PLA and furthermore the PPGDGE signals (Figure 4). Signals at 5.1 ppm were assigned to methane protons (–CH) from the PLA block, the signals at 4.3 ppm belongs to methane protons of the carbon that is attached to the OH group that is formed when the epoxide ring is opened, the signals at 3.5 belonged to methylene protons (–CH$_2$) from PPGDGE and, finally, the methyl protons (–CH$_3$) from PPGDGE and PLA block were at 1.6–1.4 ppm.

Once the monomer ratio was set, the cat/ini ratio influence on the thermal behavior of the PLA based copolymers was studied. First the wt % of the catalyst was modified (0.01, 0.03, 0.05) and the wt % of the initiator was set in 0.15 wt %, after the wt % of the initiator was modified (0.1, 0.15, 0.2) and the catalyst wt % was set in 0.01%. All these modifications were made based on previous literature reports [41].
To evaluate the influence of the cat/ini ratio on the thermal behavior of copolymers differential scanning calorimetry (DSC) was used. All of the thermograms were obtained at a heating rate of 10 °C/min. The samples were initially heated to 115 °C, quenched to −60 °C and then heated from −60 °C to 115 °C at a heating rate of 10 °C/min. The first heating scan data were discarded because it included the previous thermal history. The results are listed in Table 1. The glass transition temperature ($T_g$) for all of the materials was set as the temperature corresponding to the midpoint of the heat capacity increment.

### Table 1. Glass transition temperatures of PLA copolymers with different cat/ini ratio.

| Sample | Cat/Ini (wt %) | $T_g$ (°C) |
|--------|---------------|------------|
| 1      | 0.01/0.1      | 3.59       |
| 2      | 0.01/0.15     | −39.19     |
| 3      | 0.01/0.2      | −1.35      |
| 4      | 0.03/0.15     | ---        |
| 5      | 0.05/0.15     | −3.61      |

As observed, in Figure 5, at the thermogram (a) the amount of initiator (1-dodecanol) used to carry out the reaction was modified, it is observed that with concentrations around 0.15 wt % the formation of more chains of PPGDGE than PLA is being favored since the $T_g$ is around −39.2 °C.
In the thermogram (b) the amount of Sn(Oct)$_2$ used was varied and it is found that when a greater amount of catalyst is added, the $T_g$ can reach $-3.61^\circ$C, indicating in this case that raising the amount of Sn(Oct)$_2$ would favor the formation of more PLA chains within the copolymer.

The data obtained indicates the effect of both the catalyst and initiator. For these copolymers, it was observed that there were ratios where the copolymer was formed mainly by chains of PPGDGE, and ratios where the copolymer chains were formed mostly by PLA. This could be related directly to the formation of the tin-alkoxide complex formed prior to initiation. According to different studies about the reaction mechanism [42–45] it is a consensus that before polymerization begins, a complex between the Sn(Oct)$_2$ and the alcohol initiator will be formed. This complex is formed as shown in Figure 6, where the (a) Stannous octoate first reacts with two alcohol molecules to (b) form a tin alkoxide bond by ligand exchange.

Although the formation of this complex is very important for the final results, it must be kept in mind that this reaction is more complex because a copolymer was formed, and the presence of the other monomer played an important role. In addition, the stannous octoate hydrolyzes in the presence of impurities, like water, and is difficult to remove from the monomers. The 2-ethylhexanoic acid from the chemically polluted stannous octoate was also an issue.

Finally, the wettability properties of the synthesized copolymers were evaluated using measures of contact angle. As seen in Figure 7, a decrease in contact angle measurements were exhibited in all the copolymers compared with neat PLA. An obvious hydrophilic improvement with the increasing constitution of the PPGDE in the copolymer is observed. It is known that the PPGDGE is a polymer with alternate hydrophobic and hydrophilic groups in molecular chains, but in this case the hydrophilic parts of the PPGDGE had a greater influence within the copolymer. The hydrophilicity modifications are confirmed by the rapid contact angle decreasing in the first 10 s, and it serves as an evidence for the copolymer hydrophilic improvement when PPGDGE is introduced. When 30% of this monomer was added, the contact angle decreased almost 38$^\circ$ in comparison with pristine PLA. As can be seen,
the increases of PPGDGE in the monomer ratio decreases the contact angle, this is in accordance with what was previously described in the literature. Alli et.al., found a similar behavior [46], as they included more PPG inside the study copolymer, the contact angle was decreasing.

![Figure 7. Contact angle of PLA-co-PPGDGE by varying monomer ratio. Cat/ini ratio 0.01/0.15 wt %.
](image)

The change in the cat/ini ratio does not have a strong influence in the contact angle (Figure 8), however as the cat/ini rate increases, the contact angle decreases. But this decrement is very slight; the change is about 1°. As analyzed previously by Xiao et al. [47] for the polymerization of 2-ethoxy-2-oxo-1,3,2-dioxaphospholane initiated by Sn(Oct)$_2$, further increase of Sn(Oct)$_2$ or dodecanol does not provide any more active species. That is why we cannot see a drastic change in the contact angle when we use those cat/ini rates.

![Figure 8. Contact angle of PLA-co-PPGDGE by varying cat/ini ratio. Copolymer 80/20 (L-lactide/PPGDGE).
](image)

All the results of the contact angle measurements agreed with the glass transition data obtained through DSC. For example, the copolymer with monomer ratio 80/20 and cat/ini ratio of 0.01/0.15 wt % was the one with the lowest glass transition temperature ($T_g$). The $T_g$ was observed at $-39.15$ °C and the contact angle was $24.8°$, confirming that this copolymer had more PPGDGE chains than PLA chains, which made the material more hydrophilic.

4. Conclusions

It was concluded that it is possible to tailor the hydrophilic properties of the copolymers of PLA and PPGDGE through the variation of monomer ratios, depending of the properties of interest. The inclusion of the PPGDGE into the PLA also modified the mechanical properties, as it was observed through the $T_g$, less rigid copolymers were obtained in comparison with pure PLA. The medical
importance of these kind of materials make them very important, so additional knowledge about their behavior may enable increased usage.

**Author Contributions:** Formal analysis, S.C.; Funding acquisition, J.C. and F.M.; Investigation, S.C.; Resources, J.C., F.M. and C.M.; Writing—original draft, S.C.; Writing—review & editing, J.C., F.M., M.E.C., R.A., C.M. and M.G.

**Funding:** This research was funded by CONACYT for the scholarship 40010, VIEP and DITCo for its financial support.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

**References**

1. Koronis, G.; Silva, A.; Fontul, M. Green composites: A review of adequate materials for automotive applications. *Compos. Part B Eng.* 2013, **44**, 120–127. [CrossRef]
2. Hillmyer, M.; Tolman, W. Aliphatic Polyester Block Polymers: Renewable, Degradable, and Sustainable. *Acc. Chem. Res.* 2014, **47**, 2390–2396. [CrossRef] [PubMed]
3. Malinconico, M.; Cerruti, P.; Santagata, G.; Immirzo, B. Natural Polymers and additives in Commodity and Specialty Applications: A Challenge for the Chemistry of Future. *Macromol. Symp.* 2014, **337**, 124–133. [CrossRef]
4. Senthil, K.P.; Gunasundari, E. Green Chemistry in Textiles. In *Sustainable Innovations in Textile Chemistry and Dyes. Textile Science and Clothing Technology*; Muthu, S., Ed.; Springer Nature Singapore: Singapore, 2018.
5. Yadav, A.; Mangaraj, S.; Singh, R.; Kumar, N.; Simran, A. Biopolymers as packaging material in food and allied industry. *Int. J. Chem. Stud.* 2018, 6, 2411–2418.
6. Kale, G.; Auras, R.; Singh, S.P.; Narayan, R. Biodegradability of polylactide bottles in real and simulated composting conditions. *Polym. Test.* 2007, **26**, 1049–1061. [CrossRef]
7. Maharana, T.; Mohanty, B.; Negi, Y.S. Melt-solid polycondensation of lactic acid and its biodegradability. *Prog. Polym. Sci.* 2009, **34**, 99–124. [CrossRef]
8. Duigou, A.; Pillin, I.; Bourmaud, A.; Davies, P.; Baley, C. Effect of recycling on mechanical behavior of biocompostable flax/poly(L-lactide) composites. *Compos. Part A Appl. Sci. Manuf.* 2008, **39**, 1471–1478. [CrossRef]
9. Bhatia, A.; Gupta, R.; Bhattacharya, S.; Choi, H. Compatibility of biodegradable poly (lactic acid) (PLA) and poly (butylene succinate) (PBS) blends for packaging applications. *Korea Aust. Rheol. J.* 2007, **19**, 125–131.
10. Jandas, P.J.; Mohanty, S.; Nayak, S.K. Surface treated banana fiber reinforced poly (lactic acid) nanocomposites for disposables applications. *J. Clean. Prod.* 2013, **52**, 392–401. [CrossRef]
11. Nofar, M.; Park, C.B. Poly (lactic acid) foaming. *Prog. Polym. Sci.* 2014, **39**, 1721–1741. [CrossRef]
12. Gu, J.; Xiao, P.; Zhang, L.; Wang, H.; Dai, L.; Song, L.; Huang, Y.; Zhang, J.; Chen, T. Functionalization of biodegradable PLA non-woven fabric as superoleophilic and superhydrophobic material for efficient oil absorption and oil/water separation. *Appl. Mater. Interfaces* 2017, 9, 5968–5973. [CrossRef] [PubMed]
13. Yang, J.; Shi, G.; Bai, J.; Wang, S.; Cao, Y.; Shang, O.; Yang, G.; Wan, W. Fabrication and surface modification of macroporous poly(lactic acid) and poly(l-lactic-co-glycolic acid) (70/30) scaffolds for human skin fibroblast cell culture. *J. Biomed. Mater. Res.* 2002, **62**, 438–446. [CrossRef] [PubMed]
14. Singha, L.; Kumara, V.; Ratnher, B.D. Generation of porous microcellular 85/15 poly(l-lactide-co-glycolide) foams for biomedical applications. *Biomaterials* 2004, **25**, 2611–2617. [CrossRef]
15. Jeong, S.J.; Ko, E.K.; Yum, J.; Jung, C.H.; Lee, Y.M.; Shin, H. Nanofibrous poly(lactic acid)/hydroxipapatite composite scaffolds for guided tissue regeneration. *Macromol. Biosci.* 2008, **8**, 328–338. [CrossRef] [PubMed]
16. Pinto, A.M.; Moreira, S.; Goncalves, I.C.; Gama, F.M.; Mendes, A.M.; Magalhaes, F.D. Biocompatibility of poly(lactic acid) with incorporated graphene-based materials. *Colloids Surf. B Biointerfaces* 2013, **104**, 229–238. [CrossRef]
17. Ramot, Y.; Haim-Zada, M.; Domb, A.J.; Nyska, A. Biocompatibility and safety of PLA and its copolymers. *Adv. Drug Deliv. Rev.* 2016, **107**, 153–162. [CrossRef] [PubMed]
18. Fishbein, I.; Chorny, M.; Rabinovich, L.; Banai, S.; Gati, I.; Golomb, G. Nanoparticulate delivery system of a tyrosostin for the treatment of restenosis. *J. Control. Release* 2000, **65**, 221–229. [CrossRef]
19. Matsusue, Y.; Yamamuro, T.; Yoshih, S.; Oka, M.; Ikada, Y.; Hyon, S.; Shikinami, Y. Biodegradable screw fixation of rabbit tibia proximal osteotomies. *J. Appl. Biomater.* 1991, **2**, 1–12. [CrossRef] [PubMed]
20. Dev, A.; Binulal, N.S.; Anitha, A.; Nair, S.V.; Furuike, T.; Tamura, H.; Jayakumar, R. Preparation of poly(lactic acid)/chitosan nanoparticles for anti-HIV drug delivery applications. Carbohydr. Polym. 2010, 80, 833–838. [CrossRef]

21. Jain, A.; Kundurur, K.R.; Basu, A.; Mizrahi, B.; Domb, A.J.; Khan, W. Injectable formulations of poly(lactic acid) and its copolymers in clinical use. Adv. Drug Deliv. Rev. 2016, 107, 213–227. [CrossRef] [PubMed]

22. Cicero, J.A.; Dorgan, J.R.; Garrett, J.; Runt, J.; Lin, J.S. Effects of Molecular Architecture on Two-Step, Melt-Spun Poly(Lactic Acid) Fibers. J. Appl. Polym. Sci. 2002, 86, 2839–2846. [CrossRef]

23. Shaver, M.P.; Cameron, D.J.A. Tacticity Control in the Synthesis of Poly(lactic acid) Polymer Stars with Dipentaerythritol Cores. Biomacromolecules 2010, 11, 3673–3679. [CrossRef] [PubMed]

24. Qian, X.; Long, L.; Shi, Z.; Liu, C.; Qiu, M.; Sheng, J.; Pu, P.; Yuan, X.; Ren, Y.; Kang, C. Star-branched amphiphilic PLA-b-PDMAEMA copolymers for co-delivery of miR-21 inhibitor and doxorubicin to treat glioma. Biomaterials 2014, 35, 2322–2335. [CrossRef] [PubMed]

25. Ya, L.; Weijun, Z. Preparation and Performance of Poly(Lactic Acid)-γ-Cyclodextrin Inclusion Complex-Poly(Lactic Acid) Multibranched Polymers by the Reactive Extrusion Process. Polym. Plast. Technol. Eng. 2018, 57, 836–849.

26. Murphy, C.; Collins, M. Microcrystalline Cellulose Reinforced Poly(lactic acid) Biocomposite Filaments for 3D Printing. Polym. Compos. 2018, 39, 1311–1320. [CrossRef]

27. Kao, C.; Lin, C.; Chen, Y.; Yeh, C.; Fang, H.; Shie, M. Poly(ethylene glycol) coating of 3D printed poly(lactic acid) scaffolds for bone tissue engineering. Mater. Sci. Eng. C. Mater. Biol. Appl. 2015, 56, 165–173. [CrossRef] [PubMed]

28. Liu, X.; Ma, P. Polymeric Scaffolds for Bone Tissue Engineering. Ann. Biomed. Eng. 2004, 32, 477–486. [CrossRef] [PubMed]

29. Plackett, D.V.; Holm, V.K.; Johansen, P.; Ndomi, S.; Nielsen, P.V.; Sipilainen-Malm, T.; Södergard, A.; Verstichel, S. Characterization of L-Polylactide and L-Polylactide-Poly lactide-CoPolymer Films for use in Cheese-Packaging Applications. Packag. Technol. Sci. 2006, 19, 1–24. [CrossRef]

30. Wang, Y.F.; Guo, H.F.; Ying, D.J. Multilayer scaffold of electrospun PLA-PCL-collagen nanofibers as a dural substitute. J. Biomed. Mater. Res. B. Appl. Biomater. 2013, 101, 1359–1366. [CrossRef] [PubMed]

31. Hu, X.; Liu, S.; Zhou, G.; Huang, Y.; Xie, Z.; Jing, X. Electrospinning of polymeric nanofibers for drug delivery applications. J. Control. Release 2014, 185, 12–21. [CrossRef] [PubMed]

32. Rosen, T.; Goldeberg, I.; Navarra, G.; Venditto, V.; Kol, M. Block-Steeoblock Copolymers of Poly(ε-Caprolactone) and Poly(Lactic Acid). Angew. Chem. Int. Engl. 2018, 57, 7191–7195. [CrossRef] [PubMed]

33. Pisani, S.; Dorati, B.; Modena, T.; Bruni, G.; Genta, I. Design of copolymer PLA-PCL electrospun matrix for biomedical applications. React. Funct. Polym. 2018, 124, 77–89. [CrossRef]

34. Grayson, A.; Voskerician, G.; Lynn, A.; Anderson, J.M.; Cima, M.J.; Langer, R. Differential degradation rates in vivo and in vitro of biocompatible poly(lactic acid) and poly(glycolic acid) homo- and co-polymers for a polymeric drug-delivery microchip. J. Biomater. Sci. Polym. Ed. 2004, 15, 1281–1304. [CrossRef] [PubMed]

35. Cantón, I.; McLean, R.; Charnley, M.; Blackwood, A.; Fiorica, C.; Ryan, A.J.; MacNeil, S. Development of an Ibuprofen-Releasing Biodegradable PLA/PGA Electrospun Scaffold for Tissue Regeneration. Biotechnol. Bioeng. 2010, 105, 396–408. [CrossRef] [PubMed]

36. Mannaa, S.; Donnellib, A.M.; Kavalb, N.; Al-Rjoubac, M.F.; Augsburgerc, J.; Banerjee, R.K. Improved design and characterization of PLGA/PLA-coated Chitosan based micro-implants for controlled release of hydrophilic drugs. Int. J. Pharm. 2018, 547, 122–132. [CrossRef] [PubMed]

37. Deng, C.; Tian, H.; Zhang, P.; Sun, J.; Chen, X.; Jing, X. Synthesis and Characterization of RGD Peptide Grafted Poly(ethylene glycol)-b-Poly(L-lactide)-b-Poly(L-glutamic acid) Triblock Copolymer. Biomacromolecules 2006, 7, 590–596. [CrossRef] [PubMed]

38. Pandita, D.; Kumar, S.; Lathe, V. Hibrid poly(lactic-co-glycolic acid) nanoparticles: Design and delivery perspectives. Drug Discov. Today 2015, 20, 95–104. [CrossRef] [PubMed]

39. Chien-Hsin, C.; Ming-Chien, Y.; Da-Guang, Y.; Chi-Hsiung, Y. Effect of immobilization of poly(γ-glutamic acid) on the biocompatibility of electrospun poly (l-lactide) mats. J. Polym. Res. 2018, 25, 92. [CrossRef] [PubMed]

40. Gonzáles, M.; Cabanelas, J.C.; Baselga, J. Applications of FTIR on Epoxy Resins—Identification, Monitoring the Curing Process, Phase Separation and Water Uptake. In Infrared Spectroscopy-Materials Science, Engineering and Technology; Theophile, P.T., Ed.; InTech: London, UK, 2012.
41. De Vos, S.; Jansen, P.; Biochem, P. Industrial-Scale PLA Production from PURAC Lactides. Available online: https://www.researchgate.net/profile/Sicco_De_Vos/publication/228836284_IndustrialScale_PLA_Production_from_PURAC_Lactides/links/563efbb508ae8d65c01460a0.pdf (accessed on 22 September 2018).

42. Kowalski, A.; Duda, A.; Penczec, S. Mechanism of Cyclyc Ester Polymerization Initiated with Tin (II) Octoate. 2. Macromolecules Fitted with Tin(II) Alkoxide Species Observed Directly in MALDI-TOF Spectra. Macromolecules 2000, 33, 689–695. [CrossRef]

43. Kricheldorf, H.R. Tin-initiated Polymerizations of Lactones: Mechanistic and Preparative Aspects. Macromol. Symp. 2000, 153, 55–65. [CrossRef]

44. Ryner, M.; Stridberg, K.; Albertsson, A. Mechanism of Ring-Opening Polymerization of 1,5-Dioxepan-2-one and L-Lactide with Stannous 2-Ethylhexanoate. A Theoretical Study. Macromolecules 2001, 34, 3877–3881. [CrossRef]

45. Pack, J.W.; Kim, S.H.; Park, S.Y.; Lee, Y.W.; Kim, Y.H. Kinetic and mechanistic studies of L-Lactide Polymerization in Supercritical Chlorodifluoromethane. Macromolecules 2003, 36, 8923–8930. [CrossRef]

46. Alli, A.; Hazer, B.; Menceloflu, Y.; Süzer, Ş. Synthesis, characterization and surface properties of amphiphilic polystyrene-b-polypropylene glycol block copolymers. Eur. Polym. J. 2006, 42, 740–750. [CrossRef]

47. Xiao, C.S.; Wang, Y.C.; Du, J.Z.; Chen, X.S.; Wang, J. Kinetics and Mechanism of 2-Ethoxy-2-oxo 1,3,2-dioxaphospholane Polymerization Initiated by Stannous Octoate. Macromolecules 2006, 39, 6825–6831. [CrossRef]

© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).