White matter changes in Alzheimer’s disease: a focus on myelin and oligodendrocytes

Sara E. Nasrabady 1*, Batool Rizvi 4, James E. Goldman 2,4 and Adam M. Brickman 3,4

Abstract

Alzheimer’s disease (AD) is conceptualized as a progressive consequence of two hallmark pathological changes in grey matter: extracellular amyloid plaques and neurofibrillary tangles. However, over the past several years, neuroimaging studies have implicated micro- and macrostructural abnormalities in white matter in the risk and progression of AD, suggesting that in addition to the neuronal pathology characteristic of the disease, white matter degeneration and demyelination may be also important pathophysiological features. Here we review the evidence for white matter abnormalities in AD with a focus on myelin and oligodendrocytes, the only source of myelination in the central nervous system, and discuss the relationship between white matter changes and the hallmarks of Alzheimer’s disease. We review several mechanisms such as ischemia, oxidative stress, excitotoxicity, iron overload, Aβ toxicity and tauopathy, which could affect oligodendrocytes. We conclude that white matter abnormalities, and in particular myelin and oligodendrocytes, could be mechanistically important in AD pathology and could be potential treatment targets.

Keywords: Alzheimer’s disease, White matter, Myelin, Oligodendrocyte, Neurodegeneration, Oxidative stress

Introduction

Alzheimer’s disease (AD) is conceptualized as a progressive consequence of two hallmark pathological changes: extracellular neuritic plaques, which are composed of amyloid-beta (Aβ) surrounded by dystrophic neuritic processes, and neurofibrillary tangles, which are intraneuronal aggregates of insoluble cytoskeletal elements, composed mainly of phosphorylated tau protein. These pathological changes are believed to result in neurodegeneration, which can be appreciated with structural neuroimaging as regional and global atrophy [73]. Because of the distribution of this pathology and its associated neurodegeneration, AD is typically considered a disease of the brain’s grey matter. However, over the past several years, neuroimaging studies have implicated micro- and macrostructural abnormalities in white matter in the risk and progression of AD, suggesting that in addition to the neuronal loss characteristic of the disease, white matter degeneration and demyelination may be important pathophysiological features. Myelin loss and the inability of the oligodendrocytes, the cells responsible for the production and maintenance of myelin, to repair myelin damage may be additional central features of AD [5, 53, 55, 60]. Because of the essential role of oligodendrocyte cell lineage in myelin production and remyelination processes, changes in the number of oligodendrocytes or their precursor cells and/or their dysfunction can affect myelin integrity and therefore be potentially implicated in AD pathogenesis.

The purpose of this review is to discuss the evidence for white matter abnormalities in AD with a focus on myelin damage and oligodendrocyte lineage cells and to review the relationship between white matter changes and the pathological hallmarks of AD. In addition, we discuss whether white matter changes are a secondary result of cortical AD pathology or whether they contribute directly or indirectly to the pathogenesis and clinical manifestation of AD.

* Correspondence: se2351@columbia.edu
1Department of Psychiatry, Columbia University, New York, NY, USA
2Department of Neurology, Columbia University, New York, NY, USA
3Department of Neurology, Montreal General Hospital, School of Medicine, McGill University, Montreal, QC, Canada
4Department of Neurology, Hospital for Special Surgery, New York, NY, USA
5Department of Psychiatry, Columbia University, New York, NY, USA

Full list of author information is available at the end of the article

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Evidence of white matter abnormalities from imaging studies

The observation that neuroimaging-defined white matter abnormalities are characteristic of AD is relatively new. Work from our laboratory, for example, demonstrated that the burden of white matter hyperintensities (WMH), distributed signal abnormalities visualized on T2-weighted magnetic resonance imaging (MRI), predicts incident AD [18, 19, 21], the rate of cognitive decline among individuals with prevalent AD [78], and is associated with genetic risk factors for late onset AD [20]. We recently showed in the Dominantly Inherited Alzheimer’s Network that WMH volume is elevated among individuals with autosomal dominant, fully penetrant mutations for AD up to 20 years before the expected onset of symptoms, demonstrating that white matter abnormalities are indeed a core feature of AD. Furthermore, the appearance of WMH in these patients emerges contemporaneously with AD-related cerebrospinal fluid (CSF) amyloid and tau abnormalities [49]. WMH severity also correlates with CSF levels of Aβ fluid (CSF) amyloid and tau abnormalities [49]. WMH severity is associated with CSF tau levels in individuals with mild cognitive impairment [79]. White matter hyperintensity severity is associated with cerebrospinal fluid (CSF) amyloid levels independent of vascular factors for late onset AD [20]. We recently showed in the Dominantly Inherited Alzheimer’s Network that WMH volume is elevated among individuals with autosomal dominant, fully penetrant mutations for AD up to 20 years before the expected onset of symptoms, demonstrating that white matter abnormalities are indeed a core feature of AD. Furthermore, the appearance of WMH in these patients emerges contemporaneously with AD-related cerebrospinal fluid (CSF) amyloid and tau abnormalities [49]. WMH severity also correlates with CSF levels of Aβ1–42 in preclinical AD [49] and predicts increasing CSF tau levels in individuals with mild cognitive impairment [79]. White matter hyperintensity severity is associated with cerebrospinal fluid (CSF) amyloid levels independent of vascular factors for late onset AD [20].

The important role of vascular disease in the development of white matter damage should be emphasized. White matter hyperintensities tend to be distributed in brain areas with relatively low perfusion levels, particularly in the deep, periventricular white matter. The density of vessels in these areas decreases both with normal aging and in AD [23], consistent with reports of decreased blood flow to white matter [69], which could cause hypoxic/ischemic damage in these areas. White matter hyperintensities are related to small vessel disease, and inflammation [26, 61, 64], but comprehensive analysis of postmortem tissue in areas known to be affected by WMH, including evaluation of possible hypoxic damage to oligodendrocyte lineage cells, has not been completed. A recent study reported that paretial WMH pathogenesis in AD is related to axonal loss, through Wallerian-like degeneration, which corresponds to cortical phosphorylated tau burden, and demyelination in patients with AD but to vasculopathy and ischemia (by sclerotic index as a marker of small vessel disease and myelin associated glycoprotein to proteolipid protein ratio as a measure of hypoperfusion) in individuals without AD, suggesting that some degree of WMH is secondary to neurodegeneration in the context of AD [57]. However, neuroimaging studies showed that white matter networks are defective in preclinical AD, at a time when neurodegenerative changes, cortical atrophy, or cortical glucose reduction were not apparent [33]. Vascular and blood brain barrier (BBB) impairments, small hemorrhagic lesions and buildup of iron have been reported in the brains of AD patients even in the preclinical stages of the disease [86].

Histopathological evidence of white matter abnormalities

The neuroimaging studies establish that there is some degree of white matter abnormality in the context of AD, which may be radiological manifestations of more widespread white matter pathological abnormalities. White matter hyperintensities have been associated histopathologically with myelin pallor, myelin loss [31, 38] (as shown in Fig. 1) and the loss of myelinated axons, as well as changes in arteriole adventitia in deep white matter [68]. In our aim to review the widespread white matter abnormalities, we focus on the changes in the myelin sheath and oligodendrocyte lineage cells.

Myelin damage

In the normal development of the central nervous system (CNS), different brain regions are myelinated at different times. Myelination begins in the fourth month of human embryonic development and continues until the third or fourth decade of life [22, 47]. In general, the spinal cord and brain stem myelinate earlier, while other areas, such as the telencephalon, the entorhinal cortex, hippocampus and the amygdala myelinate later [17, 30]. Diffusion tensor imaging studies show the development of association tracts in post-adolescent subjects [47]. In addition, there is a particular susceptibility for demyelination in areas that are myelinated at earlier ages, a phenomenon referred to as “neuropathologic retrogenesis” [8, 12, 17, 66, 76]. Myelin loss has been observed consistently in AD and the later-myelinated areas are also most vulnerable [13, 36, 65]. They demonstrate significantly greater myelin loss compared with areas that myelinate earlier [6]. Analysis of postmortem brain tissue of AD patients has revealed that the white matter is altered chemically, compared with that of patients without dementia: the amounts of total protein, myelin basic protein (MBP), myelin proteolipid protein (PLP), Cyclic nucleotide phosphohydrolase (CNPase), and cholesterol is significantly decreased, indicating a loss of myelin. White matter fatty acid ratios are also altered in AD [67].

White matter and myelin changes in AD need to be taken with respect to changes during aging. For example, the overall hemispheric white matter volume decreases with age [50]. How much of this is due to changes in water content or water infusibility is not completely clear [42]. However, a decrease in the total length of myelinated fibers, reaching a 45% decrease from 20 to 80 year-old individuals and the appearance of thinner axons has been reported [50]. Thus, a number of investigations conclude that with age, myelin production by oligodendrocytes continues but leads to thinner myelin sheaths and shorter internodes [50]. Axon caliber decreases in experimental models of demyelination and remyelination [52]. Thinner
myelin sheaths and smaller axons can lead to functional white matter deficits due to conduction failure and by a greater vulnerability to trauma, oxidative stress, or Aβ toxicity [5]. Axonal loss and demyelination are both associated with white matter abnormalities in AD and are predictors of severity of white matter abnormalities [57]. Future studies with the aim of preventing or repairing myelin damage could elucidate the impact of white matter changes as one of the core pathologies of AD.

Abnormalities of oligodendrocyte lineage cells in AD
The primary role of oligodendrocytes is to produce myelin, but they also play a supportive, modulatory and regulatory role for neurons, including the production of neurotrophic factors, inhibition of neurite growth, and stabilization of neuronal connectivity [44, 70]. The adult central nervous system contains both precursor cells for oligodendrocytes in addition to mature, myelinating cells [10]. Oligodendrocyte
precursor cells (OPCs) in the adult central nervous system are capable of proliferating and migrating and effecting new myelination after demyelinating insults. Animal studies have shown that during adulthood, oligodendrocytes are generated from ventricular-subventricular zone and the new oligodendrocytes progenitors migrate to white matter tracks from there [59]. Mature human oligodendrocytes in the condition of being deprived from their myelin sheath may return into their previous subtype capacity and change their phenotype and their myelination program [37]. The different oligodendrocyte lineage cells express changes in morphology during their development and maturation [10]. Figure 2 shows an example of oligodendrocyte distribution throughout white matter areas in an adult postmortem human brain. In mammals, OPCs, by presence of intrinsic hypoxia-inducible factor (HIF) signaling, control white matter angiogenesis, axonal integrity, and the onset of myelination at postnatal stages [85]. Oligodendrocytes are able to modulate ion homeostasis in the axon environment [60]. OPCs are the specific glial cells that directly make synapses with neurons; they build synapses with glutamatergic neurons in the hippocampus (rat) [15], cortex (mouse) [24], white matter tracks (rat) [46] and other areas. However, they only make postsynaptic connection with neurons. The behavior of OPCs can be controlled by neurotransmitters and by surrounding neuronal demands. For example, in the presence of higher neural activity, oligodendrocytes change the amounts of myelin sheaths, which affects the

![Fig. 2](image_url)

**Fig. 2** An example of Olig2+ oligodendrocyte distribution throughout the white matter from a neurologically-healthy adult, postmortem brain. The insets show Olig2+ nuclei at higher magnification. H&E counterstaining. Arrowheads: Olig2+ nuclei (brown). Dashed line: the border of white and grey matter. LV: lateral ventricle; SVWM: subventricular white matter; DWM: deep white matter; SCWM: subcortical white matter; GM: grey matter. The scale bars in a, b and c are 100 μm and the scale bars in the insets are 10 μm.
electrical transmission in the neuronal network [32]. We could speculate that neuronal AD pathology, which is characterized by neuronal and axonal dysfunction, could alter the amounts of myelin produced by oligodendrocytes. This dynamism of myelin and oligodendrocytes thus could ultimately affect behavior [58].

A number of human and animal studies investigated oligodendrocyte changes in AD. In one study, 6–8-month-old APPPS1 mice showed an increase in the numbers of OPCs, while the number of Olig2+ cells in post-mortem tissues of AD patients was decreased [11]. Another study reported a higher number of MAP-2 positive remyelinating oligodendrocytes in the areas adjacent to periventricular white matter lesions and a higher number of PDGFR-α positive OPCs in white matter lesions [74]. In a PS1 knock-in mouse model, increased vulnerability and death of oligodendrocytes due to glutamate and Aβ were demonstrated and these cells showed a deficit in calcium regulation [60]. This study suggests that the abnormalities of oligodendrocytes in the presence of a PS1 mutation could be an early event in the disease course. Another study demonstrated that MBP and the number of myelinating oligodendrocytes were decreased in 6-month-old triple transgenic mice (3xTg-AD), the number of immature oligodendrocytes remained unchanged and mature non-myelinating cells were increased [29]. The authors reported that myelinating oligodendrocytes are highly sensitive to oxidative stress due to their higher metabolic demand, higher iron, and lipid content. Reduction of the diameter of oligodendrocyte nuclei in AD patients in parahippocampal white matter was also reported [35] while the mean nuclear diameter of neurons remained unchanged. Alzheimer’s-related changes in oligodendrocytes for human studies and animal models are summarized in Table 1.

One of the frequently accepted themes related to oligodendrocyte damage in AD is that these cells suffer from oxidative stress, which can be produced by a wide range of factors. The adult CNS contains oligodendrocyte precursor cells, which can be mobilized to differentiate into myelinating oligodendrocytes [10]. Oxidative stress impairs the differentiation of OPCs, in part by decreasing the levels of expression of genes that promote oligodendrocyte differentiation, such as Shh, Sox10 and HDAC3 [34]. In cell culture, pre-oligodendrocytes show a sensitivity to oxidative stress and glutathione depletion [2]. In rat cell cultures, low antioxidant content and high iron capacity, in addition to excitotoxicity through metabotropic glutamate receptors, makes oligodendrocytes more vulnerable to oxidative stress [27, 77]. Other factors affecting oligodendrocytes are listed below.

**Aβ and tau**: Several studies suggest that Aβ is toxic to oligodendrocytes [28, 29, 41, 48, 84]. For example, a cell culture study of rat oligodendrocytes demonstrated that Aβ-induced oxidative stress can drive oligodendrocyte death and dysfunction [84]. This study also showed that mitochondrial DNA damage and the consequent NF-kB and AP-1 activation are other possible mechanisms of Aβ toxicity for oligodendrocytes [84]. Note that although amyloid plaques are exceedingly rare in AD white matter, the levels of soluble Aβ are elevated in the white matter [25]. Thus, a direct exposure of white matter oligodendrocytes to increased amounts of Aβ is likely. Although there are toxic effects of Aβ on oligodendrocytes, clinical trials that have aimed to remove the Aβ plaque in symptomatic AD patients, did not prevent the progressive neurodegeneration and cognitive decline in AD patients [39, 72]. These findings suggest that this toxic effect needs to be targeted earlier or it could not be the only pathology leading to cell death and atrophy in symptomatic patients. In addition to the effects of amyloid pathology, the impact of tau pathology on white matter needs to be considered. Tau can affect the normal function of neurons through a toxic gain of function or a loss of its normal function in stabilizing microtubules. Although severe neocortical tauopathy occurs in later stages of AD and mostly affects grey matter, phosphorylated tau transforms into neurofibrillary tangles in neurons as well as glial tangles in astrocytes.

| The model and specimen | Oligodendrocyte changes | Source |
|------------------------|-------------------------|--------|
| PS1 knock-in mouse     | Vulnerability and death of OLs | Pak et al. 2003 |
| Postmortem AD          | Increased MAP-2 positive remyelinating OLs adjacent to WM lesions | Simpson et al. 2007 |
| 3xTg-AD mouse          | Decreased myelinating OLs | Desai et al. 2010 |
| Postmortem AD          | Reduced OLs nuclear diameter in parahippocampal white matter | Gagyi et al. 2011 |
| APPPS1 mouse           | Increased Olig2+ | Behrendt et al. 2013 |
| Postmortem AD          | Decreased Olig2+ | Behrendt et al. 2013 |

*Table 1*: This table summarizes the studies, the specimen that was used, and oligodendrocyte alterations in AD animal models and human AD patients in parahippocampal white matter was also reported [35] while the mean nuclear diameter of neurons remained unchanged. Alzheimer’s-related changes in oligodendrocytes for human studies and animal models are summarized in Table 1.

It depicts the verity of the results in different animal models and human studies. PS1 Presenelin-1, OL oligodendrocyte, MAP microtubule associated protein, PDGFR platelet-derived growth factor receptor, OPCs oligodendrocyte progenitor cells, WM white matter, 3xTg-AD triple transgenic AD mouse model. APPPS1 mouse mouse with both APP and PS1 transgenes.
or oligodendroglia [4]. Furthermore, phosphorylated tau in grey matter is associated with white matter abnormalities and demyelination in AD patients [56, 57]. The increased levels of calpain2 in the AD white matter, an indicator of axonal loss, was shown to be associated with increased cortical phosphorylated tau and amyloid [57] and the phosphorylated tau showed to be a predictor for white matter hyperintensities [56].

Iron: During myelination, oligodendrocytes require 2–3 fold higher energy levels than other cell types in the CNS to produce such an extensive amount of membrane. Oligodendrocytes synthesize cholesterol, a process that is highly metabolically demanding, making them vulnerable to hypoperfusion, excitotoxicity, heavy metals, and free radicals that induce oxidative stress. Oligodendrocytes have the highest iron content of all cell types, which increases with age and even further in AD [5]. Oligodendrocytes at all stages of their differentiation, compared with other glial cells contain smaller amounts of antioxidant agents (e.g. glutathione peroxidase) and only half of the glutathione reductase activity [43]. Thus, a high iron content and a low antioxidant content make oligodendrocytes one of the most vulnerable cell classes to oxidative stress in the CNS. If oxidative stress is exacerbated by age, it may lead to increased cell damage [81]. Bartokis by comparing a map of cortical myelination with maps of Aβ deposition hypothesized that age-associated myelin breakdown leads to iron release from oligodendrocytes and that this iron release promotes Aβ oligomerization in the parenchyma [7].

Hypoxia/Ischemia: Deep white matter areas lie at the ends of the CNS arterial circulation, making them susceptible to decreases in blood flow and oxygenation. Some anterior and posterior white matter lies in watershed zones between the anterior cerebral and middle cerebral arteries and middle cerebral and posterior cerebral arteries respectively. Vascular pathology in these regions is greater in patients with AD than in individuals without dementia [23, 69]. Late-stage oligodendrocyte progenitors are more sensitive to hypoxic or ischemic damage than early-stage progenitors and more mature oligodendrocytes [3]. A recent rodent study, using single cell RNA sequencing, identified a population of oligodendrocyte precursors as vascular and leptomeningeal cells. These OPCs are located along vessels and they show similarities with pericyte lineage cells [51]. In addition, to emphasize the importance of relationship between vascular system and oligodendrocyte lineage cells Tsai showed that those OPCs require the physical infrastructure provided by the vascular system to facilitate their migration during development [80].

Excitotoxicity: In general, oligodendrocytes show a great vulnerability to excessive ATP and/or activation of glutamate receptors [55]. Oligodendrocytes express a wide variety of receptors and membrane channels (e.g. ionotropic glutamate and ATP receptors, ligand gated Ca2+ channels and P2X7 receptors). Because of the lack of the GluR2 subunit in oligodendrocyte AMPA receptors, there is a higher permeability to Ca2+ ions compared with neurons [54]. Another example is the sustained activation of p2X7 receptors in oligodendrocytes due to excitotoxicity or to high levels of ATP/ADP/AMP, which leads to excessive Ca2+ in the cytosol and the activation of apoptosis through caspase-3 activation. Extensive activation of these receptors can result in oligodendrocyte damage and subsequently myelin destruction. Back and colleagues showed maturation-dependent vulnerability of oligodendrocytes caused by intracellular glutathione depletion [2]. In addition, as we mentioned, oligodendrocytes and myelin damage due to excitotoxicity and calcium dysregulation could be an early pathological feature of AD [45, 60].

DNA damage: Age related DNA damage in myelinating oligodendrocytes may contribute to myelin loss [81, 82]. Postmortem analysis of white matter lesions obtained from aging individuals shows the presence of oxidative damage (8-OHdG immunoreactivity) in oligodendrocyte nuclear DNA. These cells are also positive for senescence markers such as SA-β-gal [1]. In older adults, excessive DNA damage occurs in vulnerable oligodendrocytes and the DNA repair mechanism becomes overwhelmed. Studies of changes in genomic integrity and genomic instability of oligodendrocytes in the white matter of patients and animal models could illuminate the role of oligodendrocyte in white matter damage and pathology of AD [81, 82]. In addition, oligodendrocyte lineage transcription factor 2 (Olig2) is located on chromosome 21 which is 6.8 Mb telomeric of the amyloid precursor protein (APP) gene. The possibility of these two genes interacting in a context of the disease needs to be studied [75].

Discussion
A variety of structural, histopathological and biochemical pathologies take place in the white matter of AD patients (summarized in Fig. 3). In this review, we have tried to answer two questions:

What changes occur in white matter in the course of AD and what is the relationship between these changes and the pathological hallmarks of the disease?

Radiological, pathological, and molecular changes occur in the white matter of AD patients. Radiological markers of white matter damage occur as early as 22 years before the estimated age of symptom onset in humans who carry AD mutations [49]. These white matter changes are believed to reflect demyelination and axon damage [63]. It is possible that the oligodendrocytes or the precursors responsible for remyelination of these areas are altered in number and in DNA stability or are functionally less efficient in the presence of genetic changes, oxidative stress, increased iron.
levels, and vascular pathology [5, 81]. In addition to gross white matter damage in AD, there are chemical alterations marked by loss of proteins and cholesterol. The decreases in the levels of myelin proteins, such as myelin basic protein (MBP), myelin proteolipid protein (PLP) and CNPase, in white matter reflects the changes in oligodendrocytes and myelin sheaths. In animal models of AD, the white matter disruption and changes in myelin marker expression are among the earliest pathological changes [16, 30]. Although white matter changes are believed to be partly related to neuronal degeneration in the cortex [57], there is also evidence that oligodendrocyte and myelin pathology, which are detected in AD mouse models, are affected prior to appearance of amyloid and cortical pathology. It is not clear if these changes are independent of cortical pathology or the cortical structural damages are beyond the detection limits of methods at early stages of the disease [29, 30, 40] and subtle neurodegenerative changes could precede or promote white matter changes even if they are not detectable with conventional methods. In terms of therapeutic implications, contemporary clinical trials have focused on the removal of fibrillar forms of amyloid protein as a primary target, motivated by the “amyloid hypothesis” that has dominated the field’s conceptualization of the disease for many years [73]. Accordingly, interventions that target the removal of amyloid would arrest the progression of disease and improve cognitive outcomes. However, to date, clinical trials aimed to clear Aβ plaques in AD have not resulted in clinical improvement or reduction in the rates of disease progression. New strategies for disease treatment and prevention are therefore necessary. The consistent observations that implicate white matter abnormalities in AD pathogenesis and progression point to opportunities to target potential novel mechanisms implicated in the disease.

Are the clinical changes in AD secondary only to the cortical hallmarks of the disease or do white matter abnormalities contribute directly or indirectly to the disease?

Beside amyloid hypothesis as the cause of AD, it is crucial to note that there is a weak association between amyloid plaques and AD symptoms [5]. Further, senile plaques can be found in about 20–40% of older adults without symptoms of dementia and cognitive impairment [14, 62]. As discussed above, myelin loss contributes to cognitive decline in humans [67, 83] and in AD animal models, early changes in white matter are followed by the first cognitive impairment detected after intracellular accumulation of Aβ but before Aβ plaques

Fig. 3 This figure summarizes the pathological cascades, and their relation with each other, occurring during the development of Alzheimer's disease in white matter and cortex. While ischemia, excitotoxicity, oxidative stress, and iron overload in white matter damage oligodendrocytes, on one hand, and amyloid toxicity affects them, on the other hand, the iron released from damaged oligodendrocytes promotes amyloid polymerization and deposition in grey matter. The consequent demyelination and axonal loss result in further white matter damage and neuronal dysfunction. Neuronal dysfunction is also a result of amyloid deposition in cortex and a proposed cause for white matter abnormalities in AD patients. White matter hyperintensities are labelled with red in the MRI (FLAIR) scan of an AD patient. Blue arrows: direction of the damages originating in grey matter. Maroon arrows: direction of the damages originating in white matter. LV lateral ventricle.
appear [29]. Altering axonal conduction by demyelination or axonal damage could directly and/or indirectly affect cognition. Future studies with the aim of repairing myelin loss could clarify the impact of white matter changes on AD pathogenesis and may have therapeutic benefit. In addition, dysfunctional oligodendrocytes early in the course of the disease may not be able to play a protective role for neurons and their axons. Hence, the process of remyelination and myelin repair will be affected. Signal conductivity, and synchronicity of impulses, which are required for information processing between neurons, depend on the amount of myelin produced by oligodendrocytes and therefore will be affected. Studies with the aim of repairing myelin damage, besides other efforts focused on the hallmark of Alzheimer’s disease, could elucidate the impact of white matter changes as one of the core pathologies of AD.

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Availability of data and materials

The manuscript has no associated data.

Authors’ contributions

SEN, JEG and AMB conceptualized the review, drafted the manuscript, read and approved the final manuscript. BR contributed towards the creation of the figures, read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Al-Mashhadi S, Simpson JE, Heath PR, Dickman M, Forster G, Matthews FE, Brayne C, Ince PG, Wharton SB (2015) Oxidative glial cell damage associated with white matter lesions in the aging human brain. Brain pathology (Zurich, Switzerland) 25:565–574. https://doi.org/10.1111/bpa.12216

2. Back SA, Gan X, Li Y, Rosenberg PA, Volpe JJ (1998) Maturation-dependent vulnerability of oligodendrocytes to oxidative-stress-induced death caused by glutathione depletion. J Neurosci 18:241–6253

3. Back SA, Han BH, Luo NL, Chrichton CA, Xanthoudakis S, Tam J, Arvin KL, Holtzman DM (2002) Selective vulnerability of late oligodendrocyte progenitors to hypoxia-ischemia. J Neurosci 22:465–463

4. Ballatore C, Lee VM, Trojanowski JQ (2007) Tau-mediated neurodegeneration in Alzheimer’s disease and related disorders. Nat Rev Neurosci 8:663–672. https://doi.org/10.1038/nrn2194

5. Bartozkos G (2011) Alzheimer’s disease as homeostatic responses to age-related myelin breakdown. Neurobiol Aging 32: 1341–1371. Doi 0.1016/j.

6. Bartozkos G, Cummings J, Sultzer D, Henderson WW, Nuechterlein KH, Mintz J (2003) White matter structural integrity in healthy aging adults and patients with Alzheimer disease: a magnetic resonance imaging study. Arch Neurol 60:393–398

7. Bartozkos G, Lu PH, Mintz J (2007) Human brain myelination and amyloid beta deposition in Alzheimer’s disease. Alzheimer’s Dement 3:122–125. https://doi.org/10.1016/j.jalz.2007.01.019

8. Bartozkos G, Sultzer D, Lu PH, Nuechterlein KH, Mintz J, Cummings JL (2004) Heterogeneous age-related breakdown of white matter structural integrity: implications for cortical ‘disconnect’ in aging and Alzheimer’s disease. Neurobiol Aging 25:843–851. https://doi.org/10.1016/j.neurobiolaging.2003.09.005

9. Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, Marcus DS, Cairns NJ, Xie X, Blazer TM et al (2012) Clinical and biomarker changes in

Abbreviations

3xTg-AD: triple transgenic AD mice model; AD: Alzheimer’s disease; AP-1: activator protein 1; ATP: adenosine triphosphate; Aβ: amyloid-beta; BBB: blood brain barrier; CNPase: cyclic nucleotide phosphohydrolase; CNS: central nervous system; CSF: cerebrospinal fluid; HDAC3: histone deacetylase 3; HIF: hypoxia-inducible factor; MBP: myelin basic protein; MRI: magnetic resonance imaging; NF-κB: nuclear factor kappa-light-chain-enhancer of activated B cells; OPC: oligodendrocyte precursor cell; P2X7 receptors: P2X purinoceptor 7; PLP: myelin proteolipid protein; Shh: sonic hedgehog; Sox-10: SRY-related HMG-box; WMC: white matter hyperintensity

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dominantly inherited Alzheimer's disease. N Engl J Med 367: 795–804. doi:10.1056/NEJMoa1202753

10. Baumann N, Pham-Dinh D (2001) Biology of oligodendrocyte and myelin in the mammalian central nervous system. Physiol Rev 81:781–927.

11. Behrens T, Baer K, Buffo A, Curtis MA, Faull RL, Rees MI, Gotz M, Dimou L (2013) Dynamic changes in myelin aberrations and oligodendrocyte generation in chronic amyloidosis in mice and men. Glia 61:273–286. doi:10.1002/glia.22432

12. Bender AR, Volkle MC, Naz R (2016) Differential aging of cerebral white matter in middle-aged and older adults: a seven-year follow-up. NeuroImage 125:74–83. https://doi.org/10.1016/j.neuroimage.2015.10.030

13. Benitez A, Feeramam E, Jensen JH, Falangola MF, Tabesh A, Fennis SH, Helppin JA (2014) White matter tract integrity metrics reflect the vulnerability of late-myelinating tracts in Alzheimer's disease. NeuroImage Clin 4:64–71. https://doi.org/10.1016/j.nicl.2013.11.001

14. Bennett DA, Schneider JA, Anvanatikai Z, Kelly JF, Aggarwal NT, Shah RC, Wilson RS (2006) Neuropathology of older persons without cognitive impairment from two community-based studies. Neurology 66:1837–1844. doi:10.1212/01.wnl.0000216668.81166.66

15. Berghes DE, Roberts JD, Somogyi P, Jahr CE (2000) Glutamatergic synapses on oligodendrocyte precursor cells in the hippocampus. Nature 405:187–191. doi:10.1038/35012083

16. Billings LM, Oddo S, Green KN, McLaugh JL, LaFerla FM (2005) Intraneuronal Abeta causes the onset of early Alzheimer's disease-related cognitive deficits in transgenic mice. Neuron 45:675–688. https://doi.org/10.1016/j.neuron.2005.01.040

17. Bloak H, Broak E (1996) Development of Alzheimer-related neurofibrillary changes in the neocortex inversely recapitulates cortical myelogenesis. Acta Neuropathol 92:197–201

18. Brickman AM (2013) Contemplating Alzheimer's disease and the contribution of white matter hyperintensities. Curr Neurol Neurosci Rep 13: 415. https://doi.org/10.1007/s11916-013-0415-7

19. Brickman AM, Provenzano FA, Muraskin J, Manly JJ, Blum S, Apa Z, Stern Y, Brown TR, Luchsinger JA, Mayeux R (2012) Regional white matter hyperintensity volume, not hippocampal atrophy, predicts incident Alzheimer disease in the community. Arch Neurol 69:1621–1627. https://doi.org/10.1001/archneur.2012.1527

20. Brickman AM, Schupf N, Manly JJ, Luchsinger JA, Provenzano FA, Nakheida E, Radlgiz Q, Collins-Praino I, Arteo S et al (2014) APOE epsilon4 contribution of white matter hyperintensities. Curr Neurol Neurosci Rep 13: 301. doi:10.1007/s11916-014-0490-x

21. Brown WR, Thore CR (2011) Review: cerebral microvascular pathology in ageing and neurodegeneration. Neurophroph Athl Neurobiol 37:56–74. doi:10.10111/j.1365-3990.2010.01199.x

22. Chitrakul R, Aguirre A, Gallo V (2004) NG2-positive cells in the mouse white and grey matter display distinct physiological properties. J Physiol 561:109–122. https://doi.org/10.1111/j.physiol.2004.047252

23. Collins-Praino LE, Francis YL, Griffith EF, Wiegman AF, Urbach J, Lawton A, Hanig LS, Cortes E, Vonsattel JP, Canoll PD et al (2014) Soluble amyloid beta immunisation in Alzheimer's disease: post-mortem quantitative MRI and neuropathology. Brain : a journal of neurology 131:3286–3298. https://doi.org/10.1093/brain/awn265

24. Gogiy G, Kornomi B, Castellanos KJ, Valyi-Nagy K, Korneff D, LoPresti P, Woltjer R, Valyi-Nagy T (2012) Decreased oligodendrocyte nuclear diameter in Alzheimer's disease and Lewy body dementia. Brain pathology (Zürich, Switzerland) 22:820–810. https://doi.org/10.1111/j.1750-3639.2012.00595.x

25. Gogiy G, Cheung RT, Lee TM, Chu LW, Chan YS, Mak HK, Zhang JX, Ou D, Fung G, Cheung C (2011) Possible retrogenesis observed with fiber tracking: an anteroposterior pattern of white matter disintegrity in normal aging and Alzheimer's disease. Journal of Alzheimer's disease : JAD 26:47–58. https://doi.org/10.3233/jad-2011-01178

26. Gogate N, Verma L, Zhou JM, Milward E, Rusten R, O'Connor M, Kufa C, Kim J, Hudson L, Dubois-Dalcq M (1994) Plasticity in the adult human oligodendrocyte lineage. J Neurosci : the official journal of the Society for Neuroscience 14:5471–5487

27. Gouw AA, Seewann A, Vrenken H, van der Flier WM, Rozemuller JM, Barkhof F, Scheltens P, Geurts JJ (2008) Heterogeneity of white matter hyperintensities in Alzheimer's disease: post-mortem quantitative MRI and neuropathology. Brain : a journal of neuroscience : the official journal of the Society for Neuroscience 131:3286–3298. https://doi.org/10.1093/brain/awn265

28. Holmes C, Boche D, Wilkinson D, Yadegaarfar G, Hopkins V, Bayer A, Jones RW, Bulrock R, Love S, Neal J (et al) (2008) Long-term effects of Abeta42 Immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. Lancet (London, England) 372: 216–223. doi:10.1016/j.amlan.2007.07.002

29. Horluch M, Maezaa R, Itoh A, Wakiyama K, Jin LW, Itoh T, Decarli C (2012) Amyloid beta1-42 oligomer versus total amyloid beta protein sheet formation in vivo. Neurobiol Aging 33:499–509. https://doi.org/10.1016/j.neurobiolaging.2010.05.007

30. Jantaratnotai N, Ryu JK, Kim SU, Martinson JG (2003) Amyloid beta peptide-induced corruscal cell damage and glial activation in vivo. Neuroreport 14:1429–1433. https://doi.org/10.1097/00001225-200307040-00027

31. Jongerum AM, Mamer L, Pankenber B (2008) No change in total length of white matter fibers in Alzheimer's disease. Neuroscience 157:878–883. https://doi.org/10.1016/j.neuroscience.2008.06.075

32. Juruulk BH, Thorburn SK, Hertz L (1998) Peroxide-scavenging deficit underlies oligodendrocyte susceptibility to oxidative stress. Glia 22:371–378

33. Kaplanzinger J, Schwab ME (1994) Inverse patterns of myelination and GAP-43 expression in the adult CNS: neurite growth inhibitors as regulators of neuronal plasticity? J Comp Neuro 340:194–206. https://doi.org/10.1002/cne.904020206

34. Kelleher RJ 3rd, Shin J (2017) Presenilin-1 mutations and Alzheimer's disease. Proc Natl Acad Sci U S A 114:629–631. https://doi.org/10.1073/pnas.1619574114

35. Kukley M, Capetillo-Zarate E, Dietrich D (2007) Vescial glutamate release from axons in white matter. Nat Neurosci 10:311–320. https://doi.org/10.1038/nb1850

36. Lebelt C, Beaulieu C (2011) Longitudinal development of human brain wiring continues from childhood into adulthood. J Neurosci 31:10937–10947. https://doi.org/10.1523/jneurosci.3503-10.1018

37. Lee JT, Xu J, Lee JM, Ku G, Han X, Yang DI, Chen S, Hsu CY (2004) Amyloid-beta peptide induces oligodendrocyte death by activating the neutral sphingomyelinase-carediase pathway. J Cell Biol 164:123–131. https://doi.org/10.1083/jcb.200307017
