Research Article

Synthesis, Characterization, and Antifungal Studies of Cr(III) Complex of Norfloxacin and Bipiridyl Ligand

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A novel slightly distorted octahedral complex of Cr(III) of norfloxacin (Nor) with the formula [CrIII(Nor)(Bipy)Cl2]Cl⋅2CH3OH has been synthesized hydrothermally in the presence of a N-containing heterocyclic compound 2,2′-bipyridyl (Bipy). The complex was characterized with FT-IR, elemental analysis, UV-visible spectroscopy, and X-ray crystallography. Spectral studies suggest that the Nor acts as a deprotonated bidentate ligand. Thermal studies were also carried out. The synthesized complex was screened against four fungi Pythium aphanidermatum (PA), Sclerotinia rolfsii (SR), Rhizoctonia solani (RS), and Rhizoctonia bataticola (RB).

1. Introduction

Quinolones are the broad spectrum synthetic antibiotics containing 4-oxo-3-carboxylic-1,4-dihydroquinoline skeleton, with bactericidal effect, good oral absorption, excellent bioavailability, and good penetration into tissues [1, 2]. Nalidixic acid is the first member of quinolones [3]. Due to the narrow spectrum of nalidixic acid several modifications were made on the basis of structure activity relationship (SARS) to enhance the bactericidal spectrum and improve the pharmacokinetics properties. It has been found that the introduction of fluorine atom at position 6 and a piperazine ring at position 7 without the presence of N at position 8 enhances the biological activity spectrum. The quinolones with these modifications are grouped together as fluoroquinolones. Norfloxacin (Nor) (Figure 1) is the first member of the fluoroquinolones. This quinolone usually acts as bidentate ligand due to the presence of pyridone oxygen at position-4 and one of the carboxylate oxygens at position-3 and it exists in a zwitterionic form. Treatment with this drug leads to uncoiling of double stranded DNA and causes immediate microbial cell death [4, 5].

In the last few years, the synergetic effect of transition metal ions with quinolones has been the subject of active research in bioinorganic chemistry. It has been observed that metal complexes with appropriate ligands are biologically more significant and specific than the metal ions and the ligand itself [6–8]. Utilization of metal ions in treatment of various diseases has been thoroughly studied and a large number of quinolone metal complexes with diverse metal ions have been reported [9]. The majority of the metal-quinolone complexes are neutral with metal ions such as magnesium(II) [10], calcium(II) [11], boron(III) [12], vanadium(IV) [13], manganese(II) [14], iron(III) [15], cobalt(II) [16], nickel(II) [17], copper(II) [18], zinc(II) [19], silver(I) [20], cadmium(II) [21], cerium(III) [22], and lead(II) [23], while for magnesium(II) [24], iron(III) [25], copper(II) [26], zinc(II) [26], platinum(II) [27], and bismuth(III) [28], the complexes are either binary metal-quinolone complexes or ternary complexes with one or more O—(e.g., H2O, MeOH, and DMSO) or N-donor ligands (e.g., pyridine, 2,2′-bipyridine) as coligands. There are some Ru(III) complexes of quinolones with potential antitumor activity that have also been reported [29]. Anticancer potency of Ru(III) complexes containing antibacterial quinolones was reported [30].

In order to investigate the coordination behaviour of Nor we have synthesized a novel mixed ligand complex of Cr(III) metal ion in presence of N-containing heterocyclic ligand Bipy hydrothermally by gradual heating and cooling.
Norfloxacin Zwitterionic form of norfloxacin

**Figure 1:** Molecular structure of Nor and its zwitterion.

During literature survey it has been found that all of the reported synthesised metalloquinolones are screened against animal pathogenic microbes (fungus and bacteria). In this present work we have screened our synthesised complex against four phytopathogenic fungi for suppression of various plant diseases to develop the agriculture science by protecting crops and vegetables for the production of a plentiful supply of high-quality and affordable food.

2. **Experimental**

2.1. **Materials.** Nor was purchased from Sigma Aldrich. Bipy and the metal salt CrCl$_3$·6H$_2$O were obtained from Merck. All the chemicals used for this work were of analytical grade.

2.2. **Synthesis of the Complex.** An equimolar mixture of CrCl$_3$·6H$_2$O and Bipy in 15 mL of 1:1 solvent mixture of methanol and acetone was stirred for 10 minutes on a magnetic stirrer at room temperature. Then 5 mL equimolar solution of Nor in the same solvent mixture was added drop by drop under stirring condition. Further, the resulting mixture was heated in a hydrothermal vessel in programmed temperature oven at 100°C for 24 hrs. Then it was gradually cooled to room temperature after 72 hrs, leading to a bright green needle shaped crystal.

Calc. for C$_{28}$H$_{35}$Cl$_3$CrFN$_5$O$_2$: C, 48.11; H, 5.05; N, 10.02%. Found: C, 48.48; H, 4.91; N, 10.61%.

2.3. **Physical Measurements.** Fourier transform infrared (FT-IR) spectra were recorded on a spectrometer Perkin Elmer Spectrum BX II in the range of 400–4000 cm$^{-1}$ by preparing sample pellets with KBr. Electronic spectra were recorded in solid state on an instrument Shimadzu UV-3101PC spectrometer. C, H, and N elemental analysis was performed on an Elmetor vario ELIII instrument. Thermogravimetric analysis (TGA) measurements were carried out in an oxygen atmosphere from ambient temperature to 900°C using Perkin Elmer Diamond. Single-crystal X-ray diffraction (XRD) was collected in a Bruker D8 diffractometer, using Cu Kα radiation.

2.4. **Microbiological Studies.** The *in vitro* antifungal activities of the ligand and the synthesised complex have been evaluated against four pathogenic fungi, *Pythium aphanidermatum* (PA), *Sclerotinia rolfsii* (SR), *Rhizoctonia solani* (RS), and *Rhizoctonia bataticola* (RB), by the agar plate technique. The compounds are directly mixed with the medium in 0, 0.0125, 0.025, 0.05, and 0.1 mg/mL (in 0.1% of Dimethyl sulfoxide) concentrations. Controls were also run and three replicates were used in each case. The antimicrobial activity is estimated on the basis of the size of the inhibition zone around the dishes after four days and the percentage inhibition was calculated by the following equation:

\[
%\text{inhibition} = \left(1 - \frac{T}{C}\right) \times 100,
\]  

where $T$ is the test sample and $C$ is the control.
Figure 3: ORTEP diagram (50% probability factor for thermal ellipsoid) of the molecule [Cr(Nor)(Bipy)Cl₂]Cl·2CH₃OH; color code: carbon: grey; nitrogen: blue; oxygen: red; fluorine: green; chlorine: dark green; chromium: orange (only hydrogen atoms were not shown for clarity).

where \( C \) and \( T \) are the diameters of the fungal colony in the control and the test plates, respectively [31].

3. Results and Discussion

3.1. Infrared Spectroscopy. FT-IR band assignments were done by comparing the spectra of the synthesised complex with those of the free ligand, Nor. FT-IR spectra of free ligand in KBr disk show that the peak at 1617 cm\(^{-1}\), assigned for pyridine stretch \( \nu(C=N) \) of py, \( \nu(C≡N) \) of pyridine, and \( \nu(C≡C) \) of aromatic ring of free Nor, was slightly shifted to 1630 cm\(^{-1}\) in the complex (Supplementary Figure S1 in Supplementary Materials available online at http://dx.doi.org/10.1155/2014/457478). In case of free ligand, a characteristic absorption band at \( \sim1733 \text{ cm}^{-1} \) which is assigned for carboxylic stretch \( \nu(C=O)_{\text{carb}} \), was replaced in the complex by two characteristic bands at 1587 cm\(^{-1}\) and at 1381 cm\(^{-1}\), assigned as asymmetric \( \nu(O-C-O) \) and symmetric \( \nu(O-C-O) \) stretching vibrations, respectively [6]. This indicates the involvement of the pyridone oxygen and carboxylate oxygen in the coordination with Cr(III) ion. The difference \( \Delta \nu = \nu(O-C-O)_{\text{ complexes}} - \nu(O-C-O)_{\text{free}} \) is the important criteria for the determination of coordination mode of the ligand [32]. \( \Delta \nu \) of the complex was found to be 206 cm\(^{-1}\) that indicates the monodentate interaction of the carboxylate group with metal ion. The FT-IR data of the complex shows a very strong and broad band at 3368 cm\(^{-1}\) and medium to weak bands at 2841 and 2487 cm\(^{-1}\). These three bands confirm the vibration of quaternized nitrogen of the piperazinyl group indicating that the zwitterionic form of free Nor involves during complex formation with the Cr(III) ion.

3.2. Electronic Spectra. Electronic spectra of the synthesized Cr(III) metal complex and the free ligand Nor were recorded in the range of 200–900 nm in solid state. Two bands have been found at 285 nm and 335 nm in case of free ligand. These two bands were assigned to \( \pi-\pi^* \) and \( n-\pi^* \) transitions, respectively. These two transitions were observed due to the presence of aromatic ring containing pyridone oxygen and carboxylate oxygen. Pattern of the electronic spectra of Cr(III) complex is similar to that of the free ligand, indicating that the ligand has not changed its structure during complexation; differences due to Bipy are not easily distinguished [33–36]. The band at 285 nm in the spectra of the complex is shifted hypochromically compared to the free ligand and the band at 335 nm slightly shifted to higher wave length region (Supplementary Figure S2), suggesting that both pyridone oxygen and carboxylate oxygen participate in the complex formation. A broad band was observed in the visible region which is centred at 590 nm due to d-d transition.

3.3. Thermal Analysis. Thermal analysis of metal complex and free ligand was also studied starting from ambient temperature to 900 °C with controlled heating rate of 10°C min\(^{-1}\) under oxygen atmosphere. The temperature ranges, percentage weight loss, eliminated moiety of every decomposition, and melting point are listed in Table 1. The free ligand and the metal complex were found to have three and four stages of weight loss, respectively (Figure 2). In case of free ligand 9% of weight loss was observed in the temperature range of 25–273 °C. Second step of weight loss started at 273 °C and ended at 579 °C with 70.63% of weight loss and third decomposition occurs between 579 °C and 727 °C with a weight loss of 20.25%. In case of metal complex first weight loss was observed in the temperature range 31–101 °C with 2.8% of weight loss. After that second step starts at 101 °C and ends at 304 °C with a mass loss of 5%. The third step was observed in the temperature range 304–508 °C with weight loss of about 47.51% and forth step of weight loss occurs in the temperature between 508 °C and 902 °C with 17.47% of weight loss.

It has been also observed that the melting point of the synthesized complex (372 °C) is more than that of parent
Figure 4: Percentage inhibition of (a) Nor, (b) bipy, and (c) [Cr(Nor)(Bipy)Cl₂]Cl·2CH₃OH against PA, SR, RS, and RB.

Table 1: Thermogravimetric data of Nor and [Cr(Nor)(Bipy)Cl₂]Cl·2CH₃OH.

| Entry                  | Step    | Temp. range (°C) | Weight loss (%) | M. P (°C) |
|------------------------|---------|------------------|-----------------|-----------|
| nor                    | First   | 25–273           | 9.0             | 221       |
|                        | Second  | 273–579          | 70.63           |           |
|                        | Third   | 579–727          | 20.25           |           |
| [Cr(Nor)(Bipy)Cl₂]Cl·2CH₃OH | First  | 31–101           | 2.82            | 372       |
|                        | second  | 101–304          | 3.78            |           |
|                        | Third   | 304–508          | 47.31           |           |
|                        | Fourth  | 508–902          | 17.47           |           |
one of the carboxylate oxygen atoms, two ring N atoms of Bipy, and two chlorine atoms giving a distorted octahedral geometry.

Further details on the crystal structure data may be obtained from Cambridge data base, Oxford, on quoting the depository number CCDC-953445. Alerts shown in check cif are due to disorder of solvent molecule.

3.5. Biological Activity. Antifungal activity of the free ligand and the complex against selected four pathogenic fungi Pythium aphanidermatum (PA), Sclerotinia rolfsii (SR), Rhizoctonia solani (RS), and Rhizoctonia bataticola (RB) was carried out. These four are soil born phytopathogenic fungi and are responsible for several plant diseases like damping-off of seedlings, stem canker, crown blight, root, crown, bulb, tuber, and fruit rots. In the future, our compound may become commercially available in order to improve the crop diseases control. The results of the antifungal tests are illustrated graphically in Figure 4. It has been observed that the antifungal activity of the newly synthesized complex increases with an increase in concentration, but remains the same for the parent ligand.

4. Conclusion

The synthesis and the characterization of neutral mononuclear mixed ligand metal complex of Nor and Bipy with Cr(III) metal ion have been realized with physicochemical and spectroscopic method. Nor coordinated with Cr(III) in a monodentate way and possesses distorted octahedral geometry. The complex shows more activity as compared with the standard ligand indicating that metal complexation enhances the activity of the parent ligand; this may be explained by chelation theory according to which chelation reduces the polarity of the ligand and the central metal atom because of the delocalization of $\pi$ electrons over the whole chelate ring increases, which favours permeation of the complexes through the lipid layer of the cell membrane.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

[1] P. C. Appelbaum and P. A. Hunter, “The fluoroquinolone antibacterials: past, present and future perspectives,” International Journal of Antimicrobial Agents, vol. 16, no. 1, pp. 5–15, 2000.

| Table 2: Single crystal X-ray crystallographic data for compound [Cr(Nor)(Bipy)Cl2]Cl2CH2OH. |
|---------------------------------------------------------------|
| **Empirical formula** | $\text{C}_{28}\text{H}_{34}\text{Cl}_{3}\text{CrF}_{5}\text{N}_{5}\text{O}_{5}$ | **Z** | 2 |
| **Formula weight** | 697.95 | **$\rho_{\text{calc}}$ [g cm$^{-3}$]** | 1.532 |
| **Crystal system** | Triclinic | **$\mu$ [mm$^{-1}$]** | 0.696 |
| **Space group** | P-1 | | |
| **Crystal color** | Green | **Reflection collected** | 15448 |
| **Temp.** | 293 (2) | | |
| **a [Å]** | 10.6127 (6) | **Unique (Rint)** | 4711 |
| **b [Å]** | 12.0733 (7) | **Observed [I > 2$\sigma$(I)]** | 400 |
| **c [Å]** | 13.5624 (8) | **Parameters** | 1035 |
| **$\alpha$ [°]** | 68.548 (5) | | |
| **$\beta$ [°]** | 69.524 (5) | **$R_w=\langle\Sigma w(Fo^2-Fc^2)\rangle/\Sigma w(Fo^2)^{1/2}$** | 0.1352 |
| **$\gamma$ [°]** | 85.285 (4) | | |
| **V [Å$^3$]** | 1513.10 (15) | | |

$^a R = \sum||Fo|-|Fc||/\sum|Fo|$. $^b R_w=\langle\Sigma w(Fo^2-Fc^2)/\Sigma w(Fo^2)^{1/2}\rangle$.

| Table 3: Bond lengths (Å) and angles (°) of [Cr(Nor)(Bipy)Cl2]Cl2CH2OH around Cr(III). |
|-----------------------------------------------|
| **Angles** (Å) | Bonds | **(°)** |
| Cl$\text{−Cr}_1$−Cl$2$ | 177.51(4) | Cl$\text{−Cr}_1$−N$1$ | 2.067(3) |
| Cl$1$−Cr$1$−O$1$ | 90.55(8) | Cl$1$−N$2$ | 2.052(3) |
| Cl$1$−Cr$1$−O$2$ | 88.92(8) | Cl$1$−O$1$ | 1.944(2) |
| Cl$1$−Cr$1$−N$1$ | 90.03(8) | Cl$1$−O$2$ | 1.934(2) |
| N$2$−Cr$1$−Cl$2$ | 90.58(8) | Cl$1$−Cl$2$ | 2.3288(10) |
| N$1$−Cr$1$−N$2$ | 78.63(10) | Cl$1$−Cl$2$ | 2.3284(10) |
| O$2$−Cr$1$−N$1$ | 171.10(10) | | |
| O$1$−Cr$1$−N$2$ | 174.03(10) | | |

quinolone (221°C). From the above thermal study, it is obvious that the synthesized complex is more thermally stable than its parent ligand Nor.

3.4. Crystal Structure of the Complex [Cr(Nor)(Bipy)Cl2]Cl2CH2OH. A single crystal suitable for X-ray diffraction for compound [Cr(Nor)(Bipy)Cl2]Cl2CH2OH was mounted on a capillary tube for indexing and intensity data collection at 183(2) K on an Oxford Xcalibur CCDC single-crystal diffractometer (MoKα radiation, $\lambda = 0.71073$ Å); see Table 2. Routine Lorentz and polarization corrections were applied, and an absorption correction was performed using the ABSSCALE 3 program [37]. Direct methods were used to locate the heavy metal atoms (SHELXS-97). The remaining atoms were located from successive Fourier maps (SHELXL-97) [38].

In the synthesised complex [Cr(Nor)(Bipy)Cl2]Cl2CH2OH, Nor behaves as a bidentate ligand and is coordinated to Cr (III) through pyridone oxygen and one of the carboxylate oxygen atoms. The crystal structure is shown in Figure 3 and selected bond distances and bond angles are listed in Table 3. It has been found that in the complex Cr(III) ion is hexacoordinated surrounded by one pyridone oxygen,
[2] D. C. Hooper, “Clinical applications of quinolones,” *Biochimica et Biophysica Acta—Gene Structure and Expression*, vol. 1400, no. 1-3, pp. 45–61, 1998.

[3] R. Singh, A. Debnath, D. T. Masram, and D. Rathore, “Synthesis and biological activities of selected quinolone-metal complexes,” *Research Journal of Chemical Sciences*, vol. 3, no. 6, pp. 83–94, 2013.

[4] V. Uivarosì, “Metal complexes of quinolone antibiotics and their applications: an update,” *Molecules*, vol. 18, no. 9, pp. 11153–11197, 2013.

[5] K. J. Marians and H. Hiasa, “Mechanism of quinolone action. A drug-induced structural perturbation of the DNA precedes strand cleavage by topoisomerase IV,” *The Journal of Biological Chemistry*, vol. 272, no. 14, pp. 9401–9409, 1997.

[6] A. Debnath, F. Hussain, and D. T. Masram, “Synthesis, characterization and antifungal studies of metalloquinolone [Cd(2-nal)2(phen)2(Cl)2],” *Complex Metals*, vol. 1, pp. 96–102, 2014.

[7] K. C. Skyrianou, E. K. Efthimiadou, V. Psycharís, A. Terzis, D. P. Kessissoglou, and G. Psomas, “Nickel-quinolones interaction. Part 1—nickel(II) complexes with the antibacterial drug sparfloxacin: structure and biological properties,” *Journal of Inorganic Biochemistry*, vol. 103, no. 12, pp. 1617–1625, 2009.

[8] K. C. Skyrianou, F. Perdih, I. Turel, D. P. Kessissoglou, and G. Psomas, “Nickel-quinolones interaction. Part 2—interaction of nickel(II) with the antibacterial drug oxolinic acid,” *Journal of Inorganic Biochemistry*, vol. 104, no. 2, pp. 161–170, 2010.

[9] I. Turel, “The interactions of metal ions with quinolone antibacterial agents,” *Coordination Chemistry Reviews*, vol. 232, no. 1-2, pp. 27–47, 2002.

[10] P. Drevenšek, J. Košmrlj, V. Psycharís, A. Terzis, D. P. Kessissoglou, and G. Psomas, “Nickel-quinolones interaction. Part 2—interaction of nickel(II) with the antibacterial drug oxolinic acid,” *Journal of Inorganic Biochemistry*, vol. 104, no. 2, pp. 161–170, 2010.

[11] Z.-F. Chen, R.-G. Xiong, J. Zhang, X.-T. Chen, Z.-L. Xue, and X.-Z. You, “2D molecular square grid with strong blue fluorescent emission: a complex of norfloxacin with zinc(II),” *Inorganic Chemistry*, vol. 40, no. 16, pp. 4075–4077, 2001.

[12] J.-H. He, D.-R. Xiao, H.-Y. Chen et al., “Two novel entangled metal-quinolone complexes with self-threading and polythreaded characters,” *Inorganica Chimica Acta*, vol. 385, pp. 170–177, 2012.

[13] J.-B. Li, P. Yang, F. Gao, G.-Y. Han, and K.-B. Yu, “Novel lanthanide complexes of ciprofloxacin: synthesis, characterization, crystal structure and *in vitro* antibacterial activity studies,” *Chinese Journal of Chemistry*, vol. 19, no. 6, pp. 598–605, 2001.

[14] Z. An, J. Gao, and W. T. A. Harrison, “Two binuclear complexes containing the enrofloxacin anion: Cd2[Si2N2O6F]4(H2O)2·4H2O and Pb2(C6H5N2O7F)4–4H2O,” *Journal of Coordination Chemistry*, vol. 63, no. 22, pp. 3871–3879, 2010.

[15] I. Turel, I. Leban, M. Zupaničić, P. Bukovec, and K. Gruber, “An adduct of magnesium sulfate with a member of the quinolone family (ciprofloxacin),” *Acta Crystallographica Section C*, vol. 52, no. 10, pp. 2443–2445, 1996.

[16] I. Turel, I. Leban, G. Klintschar, N. Bukovec, and S. Zalar, “Synthesis, crystal structure, and characterization of two metal-quinolone compounds,” *Journal of Inorganic Biochemistry*, vol. 66, no. 2, pp. 77–82, 1997.

[17] I. Turel, K. Gruber, I. Leban, and N. Bukovec, “Synthesis, crystal structure, and characterization of three novel compounds of the quinolone family member (norfloxacin),” *Journal of Inorganic Biochemistry*, vol. 61, no. 3, pp. 197–212, 1996.

[18] P. P. Toffoli, P. Khodadad, and N. Rodier, “Bis (tetra-n-butylammonium) bis [(4-cyanophenyl) dithiocarbimato (2–)nickel(II)],” *Acta Crystallographica Section C*, vol. 54, no. 4, pp. 470–473, 1988.

[19] I. Turel, I. Leban, and N. Bukovec, “Crystal structure and characterization of the bismuth(III) compound with quinolone family member (ciprofloxacin). Antibacterial study,” *Journal of Inorganic Biochemistry*, vol. 66, no. 4, pp. 241–245, 1997.

[20] V. Uivarosì, M. Badea, R. Olar, D. Marinescu, T. O. Nicolescu, and G. M. Nitulescu, “Thermal degradation behavior of some ruthenium complexes with fluoroquinolone derivatives as potential antitumor agents,” *Journal of Thermal Analysis and Calorimetry*, vol. 105, no. 2, pp. 645–650, 2011.

[21] J. Kljun, A. K. Bytzeck, W. Kandioller et al., “Physicochemical studies and anticancer potency of ruthenium 66-p-cymene complexes containing antibacterial quinolones,” *Organometalics*, vol. 30, no. 9, pp. 2506–2512, 2011.

[22] R. V. Singh and A. Chaudhary, “Biologically relevant tetraaza-macrocyclic complexes of manganese: synthetic, spectral, structural and biological properties,” *Journal of Inorganic Biochemistry*, vol. 99, no. 3, pp. 677–689, 2005.
antimicrobial, antifertility and anti-inflammatory approach,” *Journal of Inorganic Biochemistry*, vol. 98, no. 11, pp. 1712–1721, 2004.

[32] D. Anamika and D. T. Masram, “Synthesis and characterization of some transition metal complexes of norfloxacin in presence of 1,10-phenenthroline,” *Der Pharma Chemica*, vol. 5, no. 5, pp. 285–290, 2013.

[33] E. K. Efthimiadou, M. Katsarou, Y. Sanakis et al., “Neutral and cationic mononuclear copper(II) complexes with enrofloxacin: structure and biological activity,” *Journal of Inorganic Biochemistry*, vol. 100, no. 8, pp. 1378–1388, 2006.

[34] E. K. Efthimiadou, Y. Sanakis, C. P. Raptopoulou, A. Karaliota, N. Katsaros, and G. Psomas, “Crystal structure, spectroscopic, and biological study of the copper(II) complex with third-generation quinolone antibiotic sparflaxacin,” *Bioorganic and Medicinal Chemistry Letters*, vol. 16, no. 14, pp. 3864–3867, 2006.

[35] G. Psomas, A. Tarushi, E. K. Efthimiadou, Y. Sanakis, C. P. Raptopoulou, and N. Katsaros, “Synthesis, structure and biological activity of copper(II) complexes with oxolinic acid,” *Journal of Inorganic Biochemistry*, vol. 100, no. 11, pp. 1764–1773, 2006.

[36] E. K. Efthimiadou, H. Thomadaki, Y. Sanakis et al., “Synthesis, characterization, antibacterial and anti-inflammatory activities of enoxacin metal complexes,” *Journal of Inorganic Biochemistry*, vol. 100, pp. 64–67, 2006.

[37] *CrysAlis Pro Software System, Version 171.32*, Oxford Diffraction, Oxford, UK, 2007.

[38] G. M. Sheldrick, “Foundations of crystallography,” *Acta Crystallographica A*, vol. 64, pp. 112–122, 2008.