Evaluating Complementary Therapies for Canine Osteoarthritis
Part I: Green-lipped Mussel (Perna canaliculus)

Anna Hielm-Björkman¹, Riitta-Mari Tulamo¹, Hanna Salonen² and Marja Raekallio¹

¹Faculty of Veterinary Medicine, Department of Equine and Small Animal Medicine, University of Helsinki, PO Box 57, Fi-00014, Finland and ²Huhtakoukku 16, 02340, Espoo, Finland

A green-lipped mussel (GLM) preparation was evaluated in a randomized, double-controlled and double-blinded clinical trial. It was hypothesized that the treatment effect would be less than that of the positive control (carprofen) but more than that of the negative control (placebo). Forty-five dogs with chronic pain and a radiographic diagnosis of osteoarthritis that were randomly allocated into one of three groups completed the study. All dogs were fed the test products or placebo for 8 weeks. The dogs were evaluated four times, at 4-week intervals. Six different variables were assessed: veterinary-assessed mobility index, two force plate variables, owner-evaluated chronic pain index and pain as well as locomotion visual analogue scales (VASs). Intake of extra carprofen was also evaluated. A chi-squared and a Mann–Whitney test were used to determine significance between groups. When changed to dichotomous variables, there were more dogs in the GLM than in the placebo group that improved, according to veterinary-assessed mobility, owner-evaluated chronic pain index and pain VAS (P = 0.031, P = 0.025, P = 0.011, respectively). For the same three, the odds ratio and their confidence interval were over one. The extent of improvement was significantly different between the GLM and the control in veterinary-assessed mobility (P = 0.012) and pain VAS (P = 0.004). In conclusion, GLM alleviated chronic orthopedic pain in dogs although it was not as effective as carprofen. As no side-effects were detected, GLM may be beneficial in dogs e.g. when non-steroidal anti-inflammatory drugs cannot be used.

Keywords: Controlled – dog – Lyproflex® – nutraceutical – OA – placebo

Introduction

Over the last few years, there has been a growing interest in new treatment options for osteoarthritis (OA), both for humans and pets, especially dogs. The so-called nutraceuticals have become available, since some patients cannot tolerate or do not want to take the risk of non-steroidal anti-inflammatory drugs (NSAIDs) because of their side-effects (1,2). Nutraceuticals have been described as naturally occurring, biologically effective nutritional supplements that can confer some degree of health benefit and there is a whole new science referred to as ‘bioprospecting’ (3) that explores and introduces these new herbs—or animal molecules or products. Currently, several nutraceuticals on the market are claiming to relieve arthritic symptoms. These products generally fall mainly into two distinct product groups, including glucosamine and chondroitin sulfate combinations or polyunsaturated fatty acids (PUFAs), particularly Omega-3 series PUFAs, such as those derived from marine sources. One of the nutraceuticals that may benefit OA is a product based on Perna canaliculus, or the green-lipped mussel (GLM) that...
Green-lipped mussel for canine osteoarthritis

has been thoroughly examined by Halpern (4) [see also (5)]. The early GLM-based products were often produced from rejected mussel meat from human food processing, which typically included steam processing as part of the manufacturing process and the early trials using these unstable GLMs, showed poor results. In the 1980s, a method of temperature-controlled cold processing and stabilization of GLM by adding organic acids to prevent oxidation and freeze-drying was patented (6). After 1986, this new stabilized GLM has been in use and is documented to be efficacious in treating experimental arthritis in rats (7,8), clinical arthritis in humans (9,10) and more recently also in dogs (11,12).

The GLM product used in this study, which originates from mussel farms in the Pacific Ocean, has been harvested when the mussels are 12 to 18-months old, and is stabilized, freeze-dried at −40°C and packed immediately thereafter. The GLM product is a rich source of nutrients, including glycosaminoglycans (GAGs), such as chondroitin sulfates, vitamins, minerals and Omega-3 series PUFAs. It is not totally clear how the products function (13) [this is not totally clear for NSAIDs either (14,15)], despite substantial research. Although there has been only a few studies published on the use of stabilized GLM on dogs, it is already available for dogs with OA, as a powder, as capsules or incorporated into pebbles.

The aim of this study was to evaluate a GLM product as an OA nutraceutical for dogs in a randomized, double-controlled and double-blinded trial. We expected the positive control carprofen to significantly reduce pain and locomotion difficulties, and the negative control (placebo) to have no such effect. The effect of the nutraceutical was hypothesized to lie somewhere between those of the positive and negative. Objective, semi-objective and subjective variables were used for assessment. Since the GLM is also used in humans, the treatment outcome is of interest for many people as OA is becoming one of the most prevalent and costly diseases in our aging society. Due to the side-effects associated with NSAIDs, 60–90% of dissatisfied human arthritis patients are reported to seek complementary therapies for their disease (16).

Methods

Dogs

Inclusion criteria were that dogs had clinical signs and a radiographic diagnosis of OA, in either a hip joint or an elbow joint. The owner had to have described at least two of the following signs as being frequent: difficulty in lying down and/or in getting up from a lying position, difficulty in jumping or refusing to jump, difficulty in walking up or down stairs, or definite lameness. Dogs were excluded from the study if they had had prior surgery of the evaluated joint, inadequate clinical symptoms, systemic or infectious disease, neurological deficits, lameness from articular infection, or recent trauma.

Sixty-eight dogs were chosen based on 124 telephone interviews with owners. Of these, 51 dogs will be presented here and the remaining 17 constituted a treatment group for another study (17). Six dogs (two in each group) were excluded from the study at some point for the following reasons: having had a previous operation of the affected hip joint, having a transverse vertebra (n = 2), sustaining a cruciate ligament injury (n = 2) and diagnosed with degenerative myelopathy. There were 15 dogs in each group that finished the study: 12 dogs with canine hip dysplasia (CHD) and three dogs with elbow OA in each group, all confirmed by radiographs. Twenty-five dogs were male and 20 were female. The median age was 6 years (range 1–11) and the median body weight (BW) was 34 kg (range 18–56). There were both uni- and bi-laterally affected dogs. All dogs had either moderate or severe radiological changes in the worst-affected hip or elbow joint (18).

Owners were asked not to give the dogs NSAIDs or corticosteroids for at least 30 days and no Na-pentosan polysulfate (Carthrophen®, Biopharm Pty. Ltd., Australia) for at least 90 days prior to the study. However, this proved not to be always the case, as some owners felt that their dogs were in such pain that they therefore gave them NSAIDs. The use of the additional analgesic pre-trial was, however, recorded in the questionnaire.

Test Products

The product tested in this study was the GLM (Lyproflex® 500 mg; ICENI, OMNI nutraceuticals, Cambridgeshire/Biofarm Oy, Finland). The capsules contain 46–52% protein, 5.9–9.3% fat, 8.0–21.5% carbohydrates, 3.9–5.8% moisture, 14–25% ash, >12 mg eicosatetraenoic acid (ETA)/100 g, >500 mg eicosapentaenoic acid (EPA)/100 g and >400 mg docosahexaenoic acid (DHA)/100 g. The initial dose was 4 (for dogs ≤40 kg BW) or 6 (for dogs >40 kg BW) capsules/day for 10 days, then continuing with half of the loading dose (i.e. two or three capsules) for the rest of the study. This meant an initial dose of 20–49 mg/kg/day depending on the BW of the dog.

Two control groups were included: the established positive control carprofen (Rimadyl® 50 mg; Pfizer, Helsinki, Finland) at a dose of 2 mg/kg twice a day and a negative control that received all products as placebos. Carprofen was a white pill without the usual stamp and its placebo an identical lactose tablet, GLM was...
appointments, and at this first visit (W0), the dogs were
CONSORT guidelines (19). A secretary made the first
controlled, double-blinded clinical trial using the
The study was designed as a randomized double-
questionnaire.
of additional carprofen doses used was recorded in the
dog of 31–40 kg and three tablets for a dog of 41–60 kg)
if they felt the dog was in considerable pain. The number
additional carprofen doses used was recorded in the
questionnaire.

Study Protocol
The study was designed as a randomized double-
controlled, double-blinded clinical trial using the
CONSORT guidelines (19). A secretary made the first
appointments, and at this first visit (W0), the dogs were
assigned into groups, in order of arrival using a
computer-generated random list. Only the location of
the diseased (hip or elbow OA) was stratified for in the
randomization. Initial clinical, orthopedic and neurologi-
cal examinations were performed and diagnostic criteria
included decreased range of motion and pain on
stretching the hip or flexing the elbow. Radiographs
were taken of the dogs’ hips and/or elbows and other
joints if needed. The W0 evaluation and W0 questionnaire
was set as baseline, except for pre-trial analgesic
medication, where the assessment was made at W−4, as
the owners were told not to use them anymore between
W−4 and W0. Follow-up visits with questionnaires for
reassessment were at 4, 8 and 12 weeks (W4, W8 and
W12). The dogs were given the products orally for
8 weeks, from W0 to W8. At W12, the dogs had been
off all medication for 4 weeks and were evaluated to
determine long-term effects of the different treatments
as follow up. All evaluators (veterinarians and owners)
and all technical assistants were blinded. Owners of
the dogs were required to sign informed consent forms.
The study protocol was approved by the Ethics
Committee of the University of Helsinki.

Veterinary Evaluation
Two veterinarians subjectively assessed three parameters
at W0, W4, W8 and W12: locomotion, jumping and
walking stairs using 0–4 descriptive scales. The three
scores assigned by the two veterinarians were summed to
form a veterinary-assessed mobility index, with a possible
minimum score of 0 and a maximum of 24 (2 × 3 × 0–4).

Owner Assessment
Four weeks before the first visit (W−4) and during each
following evaluation, owners answered a three-part
questionnaire. The first part used a descriptive scale of
0–4 and contained questions about attitude, behavior and
locomotion. Of these, 11 questions were combined to
form a combined owner-assessed chronic pain index, as
described previously (20). The second part contained two
10 cm visual analogue scales (VASs): one for pain and
the other for locomotion. The end of the lines to the
left represented no pain or no difficulties in locomotion,
and to the right, the worst possible pain or the most
severe difficulties in locomotion. The third part consisted
of questions about possible adverse reactions to treat-
ment, including change in appetite, vomiting, diarrhea
and atopic skin reactions. The question about additional
analgesics was not a continuous variable but used
the following scale: ‘during the last four weeks additional
carprofen was given 1 = not at all, 2 = 1–2 times,
3 = about once a week, 4 = about 3–5 times a week,
5 = daily/almost daily’.

Objective Evaluation of Gait
Gait was analyzed by force plate gait analysis (Kistler
forceplate, Type 9286, Kistler Instrumente AG
Winterthur, CH-8408, Switzerland), which assesses weight
bearing of limbs. The force plate was submerged into the
concrete floor so that the plate and floor surfaces were on
the same level. The floor was then covered with a 2 mm
thick rubber mat that extended from 7 m before to 7 m
after the plate, forming a 14 m walkway. A hole was cut
in the mat over the force plate and a 3–4 mm gap was
left between the force plate mat and the rest of the mat.
The signal from the plate was processed and stored using
a computer-based software program, and velocities
and acceleration were determined by three photoelectric
cells placed exactly 1 m apart and a start-interrupt
timer system (Aquire 6.0, Sharon Software Inc.,
DeWitt, MI, USA).

Dogs guided by their owners trotted over the walkway
from left to right. The speed was one comfortable
for each dog in trot and had to be in the same range
(± 0.5 m/s) for the dog each time the test was
performed (at W0, W4, W8 and W12). The acceleration
was < 0.5 m/s/s and contact had to be made with the
plate first by the forelimb and shortly after with the hind
limb of the same side for the evaluation to be valid. The
test was repeated until sufficient valid results were
obtained for both left and right limbs.

Three valid measurements for each side and for each
visit were then chosen by a blinded assistant (one
not otherwise participating in the study) according to
speed, acceleration and with no interferences, such as
gait abnormalities or extra body movements. The mean
of these three measurements was used for analysis. The ground reaction forces were normalized for each dog’s BW and mean peak vertical force (PVF) and mean vertical impulse were used as variables. Only measurements from the most severely affected leg at time W₀ were used in the analysis.

**Blood Samples**

Blood samples were collected from the dogs at each visit. Blood urea nitrogen (BUN), creatinine, serum alanine aminotransferase (ALAT), alkaline phosphatase (AFOS), total protein and albumin were analyzed.

**Statistical Analysis**

The number of dogs needed in each group was calculated for a two-tailed test (Fisher). The sample size \( n = 15/\text{group} \) was sufficiently large to detect a 47% difference (11) in treatment outcome (effective versus not effective) with a statistical power of 0.8 and allowing for a 5% \( \alpha \)-error.

To counteract the effect of the extra NSAID on dogs that at W₈ had used extra carprofen more than three times per week, their W₈/C₀ values were changed into the most negative value measured for any dog at that time. This enabled us to use the whole data in the statistical analyses.

For calculating the percentage of dogs/group that improved between baseline and W₈ and the odds ratio, the results of each variable were converted into dichotomous responses of ‘improved’ and ‘not improved’. Dogs that deteriorated and dogs with no change in the evaluated variable were considered ‘not improved’. The difference between the treatment groups and the placebo was calculated using a chi-squared test. The odds ratio was calculated using the common Mantel–Haenszel odds ratio estimate and the confidence interval (CI) was set to 95%. An odds ratio more than 1.0 indicated a beneficial effect of the test treatments.

The changes from baseline to W₈ were also calculated for each variable. The difference between the GLM and placebo group was analyzed using the Mann–Whitney test. The changes from W₀ to W₈ in the force plate variables were proportional in the front and hind legs, although the values were different. Therefore, force plate data of all four legs were analyzed together. The dogs, for which no force plate results could be registered, were considered ‘not improved’ in the dichotomous evaluations and excluded in the median analyses. A correlation test was used to evaluate the association between the assessments of the two veterinarians. Statistical significance was set at \( P < 0.05 \). Statistical tests were preformed using SPSS 12.0 for Windows (SPSS Inc., Chicago, IL, USA).

**Results**

**Baseline Values**

Baseline variable median (range) values were: for the veterinary-assessed mobility index: 6 (0–18), PVF: 71.21 (54.7–135.25), vertical impulse: 9.11 (6.02–19.9), owner-evaluated chronic pain index: 16 (4–25), pain VAS: 3.55 (0–8.4) and locomotion VAS: 4.8 (0–8.3). There was no statistical bias between the groups at baseline. The evaluations of the two veterinarians correlated well \( (R = 0.853, P < 0.01) \).

**Dichotomous Responses**

There were four dogs (all from the placebo group) that had used extra carprofen more than three times per week at W₈. For three of the variables [veterinary-assessed mobility index \( (P = 0.031) \), chronic pain index \( (P = 0.028) \) and pain VAS \( (P = 0.011) \)] there were significantly more improved dogs in the GLM group compared to the placebo group (Table 1). The odds ratio for the veterinary-assessed mobility index was 5.5 (95% CI 1.14–26.41) indicating that a dog that had received the GLM product was 5.5 times more likely to have a positive response compared to a dog that had received the placebo. The odds ratio for the force plate PVF was 2.50 (95% CI 0.52–11.93), for the force plate impulse 2.40 (95% CI 0.52–10.99), for the owner-assessed chronic pain index was 6.0 (95% CI 1.17–30.72), for the pain VAS 8.0 (95% CI 1.52–42.04) and for the locomotion VAS 4.12 (95% CI 0.88–19.27).

**Medians of the Change from W₀ to W₈**

All variables showed a similar trend of improvement, with carprofen being the most efficient, placebo the least and GLM being between these two (Table 1). There was a significant difference between the GLM and the placebo in two variables [veterinary-assessed mobility index \( (P = 0.012) \) and pain VAS \( (P = 0.004) \)] and a third variable was close to significant [locomotion VAS \( (P = 0.057) \)].

**Extra Carprofen**

At W₄, before the owners were requested to stop all medication, 14% of the carprofen group, 13% of the GLM group and 8% of the placebo group were given NSAIDs once a week or more. At W₈, (Fig. 1) 0, 7 and 27% of the respective groups were given additional carprofen once a week or more. At follow-up (W₁₂), the respective numbers were 33, 14 and 29%. The differences between both GLM and carprofen compared to the placebo group at time W₈ were significant \( (P = 0.021 \text{ and } P = 0.008, \text{ respectively}) \).
Complications and Side-effects

Three dogs (one in the GLM group, two in the placebo group) were so lame during the visits that no usable data were obtained from the force plate. Two of these dogs (one in the GLM group, one in the placebo group) were euthanized between W8 and W12 due to severe pain.

In populations, neither our findings of clinical side-effects nor clinical chemistry in any of the blood parameters were severe or related to any particular group. Palatability was never a concern.

Discussion

In our study, dogs showed a beneficial clinical response to treating OA-induced pain and locomotion difficulties with GLM. More dogs improved in the GLM group compared to the placebo group and the extent of treatment effects was between that of our two control groups, as can be seen from the median values in Table 1. The carprofen had in previous studies shown 56–80% of improvement in dogs with OA (graded by veterinarians and owners) whereas the placebo in the same studies showed improvement in only 23–38% of the cases (21, 22). As these numbers are close to the results we obtained in our study for the two control groups (Table 1), they indicate that our cohort reflects reality well and that we can trust the results of our treatment group. The fact that extra carprofen was used significantly more often in the placebo group at W8 is also a positive result for the tested product.

This positive outcome opens a discussion about possible working mechanisms of the GLM (Fig. 2). In the early GLM clinical trials on human patients, the outcomes were not good and often contradictory (23,24). Twenty years later, possibly after having stabilized the product by freeze-drying and lyophilizing, the results of clinical trials for GLM have been significantly promising.

Table 1. Percentage of improved dogs and median (range) of improvement for evaluated variables, per group from W0 to W8

| Variable                  | Carprofen (n = 15) | GLM (n = 15) | Placebo (n = 15) |
|---------------------------|-------------------|--------------|-----------------|
| Improved                 | Improvement       | Improved     | Improved        |
| Veterinary mobility index | 66.7% (P = 0.031) | 66.7% (P = 0.031) | 26.7% (P = 0.031) |
| Force plate PVF           | 66.7% (P = 0.031) | 46.7% (P = 0.031) | 26.7% (P = 0.031) |
| Force plate Impulse       | 80.0% (P = 0.011) | 53.3% (P = 0.011) | 33.3% (P = 0.011) |
| Chronic pain index        | 80.0% (P = 0.028) | 80.0% (P = 0.028) | 40.0% (P = 0.028) |
| Pain VAS                  | 85.7% (P = 0.001) | 66.7% (P = 0.001) | 20.0% (P = 0.001) |
| Locomotion VAS            | 85.7% (P = 0.002) | 60.0% (P = 0.002) | 26.7% (P = 0.002) |

For each treatment group: First column: Percentage of dogs in the group that improved. Below: P = Difference in percentage of improved between treatment groups and placebo. Second column: Median (with range) of change from W0 to W8 [(+, improvement; (-), deterioration] in evaluated variables for the carprofen-, GLM- and placebo-groups. P = Difference in improvement between treatment groups and placebo (the force plate values do not include three dogs for whom no results were obtained). n, number of patients per group; GLM, green-lipped mussel; PVF, peak vertical force; VAS, visual analogue scale.

Figure 1. At the end of the treatment period (W8), there was 4/15 dogs in the placebo group given extra carprofen 3–7 days/week (n = number of dogs).
The lyophilizing process might have been more important as in fact, the difference in lipid, sterol or fatty acid composition of frozen and freeze-dried GLM has been shown to be non-existent; the only major difference was between total lipid composition on a dry weight basis because of the removal of water in the deep-frozen product (25). The potent anti-inflammatory activity of GLM powder was confirmed in vivo using the established rat paw oedema model; rats fed mussel lipids perorally developed neither adjuvant-induced polyarthritis nor collagen-induced auto-allergic arthritis (8). However, these lipids showed only marginal inhibition of carrageenan-induced paw edema in rats (acute irritation assay, which is the standard test for NSAIDs), indicating that they do not mimic rapid-acting NSAIDs (8,13). Macrides and others (7) found that the ETAs of GLM had considerable anti-inflammatory activity. In vitro, the extracted lipids have been shown to possess significant cyclo-oxygenase (COX) and lipoxygenase (LOX) inhibitory activity; hence, the GLM seems to be working on the same mechanisms as newer NSAIDs (8).

This dual inhibition of both LOX- and COX metabolic pathways may offer an explanation for the reported clinical efficacy and favorable gastrointestinal tolerability of GLM. Platelet aggregation remains unaltered and the lipid fraction be non-gastrotoxic in fasted disease-stressed arthritic rats at a dose of 300 mg/kg (treatment dose 20 mg/kg) (8,13). This shows that the GLM does not have the negative side-effects of the NSAIDs. Recently, new GLM extracts were tested (26) and a Tween-20 extract (that draws out membrane-bound proteins by a cationic detergent) effectively inhibits both COX-1 and COX-2 activity. It also induced a significant reduction in TNF-α, IL-1, IL-2 and IL-6 and decreased IgG levels, indicating that GLM may regulate the immune system and promote humoral and cellular activity (26).

The active components possess a molecular weight above 100 kDa and when a proteolytic enzyme was added to the extract, it eliminated the component effective against inflammatory cytokines, suggesting that at least part of the active substance resides in the protein moiety associated with the glycogen, probably as a glucose polymer. GLM, green lipped mussel; Th, T helper cell; IL, interleukin; TNF, tumor necrosis factor.

Figure 2. Main active constituents of the green lipped mussel and their effect on the inflammation pathways of osteoarthritis. The main active constituents, according to how we understand their working mechanisms now: The Omega-3 PUFAs (especially the ETAs) have anti-inflammatory activity; they possess significant cyclo-oxygenase (COX 1 and 2) and lipoxygenase (LOX-5) inhibitory activity. Due to their glycosaminoglycan content (especially chondroitin sulphate with its high glucosamine content), the GLM may have chondroprotective properties. The vitamins and minerals are needed in cartilage anabolism. GLM, green lipped mussel; Th, T helper cell; IL, interleukin; TNF, tumor necrosis factor.
glycosaminoglycans (GAGs) of the GLM, further work as building blocks in cartilage anabolism; glucosamine is one of their main constituents. They help the joint capsule to hold water and to adapt to changes in pressure, thereby absorbing shock induced by abnormal joint stress (28). The role of minerals and vitamins has not been studied, but it is possible that they also contribute to the positive effects of the GLM. As seen earlier, the GLM probably acts through several different working mechanisms.

Three studies exist on stabilized GLM as a treatment for canine OA and our findings are consistent with two of them. Bierer and Bui (11) conducted three 6-week, randomized, double-blinded trials, in which they compared three different GLM dog feeds with control feeds. They used a total arthritic score by summing eight variables. As in our study, all individual variables showed no significant improvement, although a significant change was observed in the total arthritis score in favor of all three GLM test groups. In our study, a different set of variables was used. The veterinarians evaluated only mobility and not range of motion, crepitus, etc., as we had noted that owner compliance was much higher if provocations that hurt the dogs were not used.

Two force plate measurements were chosen as objective variables. Force plate has been used in similar studies to evaluate treatments of hip (22,29–31) and elbow (22,31,32) joints. The best variables for these conditions were considered to be PVF and vertical impulse, both of which were included here. The change from baseline to end of treatment in vertical impulse and PVF in our study was in the same range as in previous studies (30) but the range was larger. Furthermore, we used three owner-assessed variables: two VAS scores, one of them slightly and the CI were large, making an exact interpretation difficult: they started showing more signs of pain when the weather changed (42,43) to a humid, raw cold (our placebo group became worse between W0 to W8) and later in the spring when the weather turned warm and dry (W8 to W12) (Fig. 1) and later in the spring when the weather turned warm and dry (W8 to W12) the dogs were better.

In future studies, to obtain an optimal effect from the GLM product, should reconsider some aspects of the treatment regime. As OA often is a clinically variable disease, not having homogeneous groups is a major drawback, and this likely influenced the results. However, although the cohort of dogs was non-homogeneous, observed as a wide range even at the start of the trial, the documented trend of improvement was clear and similar for all variables and may even have been more evident, if we had more dogs. The positive effects of the GLM could eventually have been underestimated, rather than overestimated: if a non-articular concurrent pain such as spondylosis or secondary muscle pain, etc. was present, there would have been a positive analgesic effect for these also in the NSAID carprofen group, while the GLM might have helped primarily in arthritic disorders (9–12).
The choice of patients and the treatment time might also have an impact on the results. Radiographically, all dogs in our study had moderate or severe OA. Although the radiological data does not correlate well with the true clinical picture, at least not in dog hip joint (20), these dogs might have been too seriously affected to benefit optimally from the product. In one of the older human studies, severity of the disease was shown to have an impact on treatment outcome; mild and moderate knee OA responded very well to GLM treatment, whereas patients with severe knee OA did not benefit from treatment (34). Also, our 2-month study period might have been too short, as some earlier human studies have been unable to show a significant improvement compared to controls until patients had ingested fatty acid products for 3–6 months (34,38–41).

In conclusion, our results suggest that the modern stabilized and freeze-dried GLM is more effective than the placebo in treating chronic pain due to moderate to severe OA and that it has no side-effects. For dogs that can not use NSAIDs or corticosteroids and for patients who need analgesic support over extended periods of time, oral GLM may be an acceptable alternative for treating chronic arthritis pain, although it does not alleviate pain as well as carprofen. As dogs are used as models for human OA, we hope these promising results will stimulate new human research in this area.

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References

1. FDA. US ADE (Adverse Drug Experience) reports summary 1998. Animal Pharm 1999;435-9.
2. MacPhail CM, Lappin MR, Meyer DJ, Smith SG, Webster CRL, Armstrong PJ. Hepatocellular toxicosis associated with administration of carprofen in 21 dogs. J Am Vet Med Assoc 1998;212:1895-901.
3. Cooper EL. Bioprospecting: a CAM frontier. Evid Based Complement Alternat Med 2005;2:1-3.
4. Halpern GM. The Inflammation Revolution: A Natural Solution for Arthritis, Asthma & Other Inflammatory Disorders. NY: SquareOne Publishers, 2005.
5. Cooper EL. The amazing science behind nature’s ‘miracle from the sea’. Evid Based Complement Alternat Med 2005;2:569-70.
6. Broadbent JM, Kosuge Y. Stabilized mussel extract. New Zealand patent 211928 (April 29, 1985); Australian patent PG 4775/84 (May 1, 1984), 1985.
7. Macrides TA, Treschow AP, Kalafatis N, Wright PFA. The anti-inflammatory effects of Omega-3 triaenolic fatty acids isolated from a lipid extract from the New Zealand green-lipped mussel. In: Proceedings of the 88th American Oil Chemists Society Annual Meeting; May 1997, Seattle; 1997.
8. Whitehouse MW, Macrides TA, Kalafatis N, Betts WH, Haynes DR, Broadbent J. The anti-inflammatory activity of a lipd fraction from the New Zealand green lipped mussel. Inflammopharmacology 1997;5:237-46.
9. Gibson SLM, Gibson RG. The treatment of arthritis with a lipid extract of Perna canaliculus: a randomized trial. Complement Ther Med 1998;6:122-6.
10. Cho SH, Jung YB, Seong SC, Park HB, Byun KY, Lee DC, et al. Clinical efficacy and safety of Lyprinol, a patented extract from New Zealand green-lipped mussel (Perna canaliculus) in patients with osteoarthritis of the hip and knee: a multicenter 2-month clinical trial. Allergy Immunol 2003;6:212.
11. Bierer TL, Bui LM. Improvement of arthritic signs in dogs fed green-lipped mussel (Perna canaliculus). J Nutr 2002;132 (Suppl): 1634-6.
12. Pollard B, Guilford WG, Ankenbauer-Perkins KL, Hedderley D. Clinical efficacy and tolerance of an extract of green-lipped mussel (Perna canaliculus) in dogs presumptively diagnosed with degenerative joint disease. New Zel Vet J 2006;54:114-8.
13. Rainsford KD, Whitehouse MW. Gastroprotective and anti-inflammatory properties of green lipped mussel (Perna canaliculus) preparation. Arznei Forsch 1980;30:2128-32.
14. Lees P, May SA, White D. Pharmacokinetics and dosage regimens of anti-inflammatory drugs. Ann Rech Vet 1990;21 (Suppl): 73-8.
15. Fox SM, Johnston SA. Use of carprofen for the treatment of pain and inflammation in dogs. J Am Vet Med Assoc 1997;210:1493-8.
16. Ahmed S, Anuntyjo J, Malemud CJ, Haqqi TM. Biological basis for the use of botanicals in osteoarthritis and rheumatoid arthritis: a review. Evid Based Complement Alternat Med 2005;2:301-8.
17. Helm-Björkman A, Tulamo R-M, Salonen H, Raekallio M. Evaluating a complementary therapy for moderate to severe canine osteoarthritis. Part II: a homeopathic combination preparation (Zeel®). eCAM, doi:10.1093/ecom/nem143.
18. Federation Canine International. FCI Hip Dysplasia and Elbow classification, workshop in Dortmund, 14.6 1991 (brochure) 1991.
19. Moher D, Schulz KF, Altman DG. The CONSORT statement: revised recommendations for improving the quality of reports of parallel-group randomized trials. Lancet 2001;357:1191-4.
20. Helm-Björkman AK, Kuusela E, Liman A, Markkola A, Saarto E, Huitunen P, et al. Evaluation of methods for assessment of pain associated with chronic osteoarthritis in dogs. J Am Vet Med Assoc 2003;222:1552-8.
21. Holsinger RH, Parker RB, Beale BS, Friedman RL. The therapeutic efficacy of carprofen (Rimadyl®-V) in 209 clinical cases of canine degenerative joint disease. Vet Comp Orthop Traumatol 1992;5:140–4.
22. Vasseur PB, Johnson AL, Budsberg SC, Lincoln JD, Toombs JP, Whitehaur JG, et al. Randomized, controlled trial of the efficacy of carprofen, a nonsteroidal anti-inflammatory drug, in the treatment of osteoarthritis in dogs. J Am Vet Med Assoc 1995;206:807–11.
23. Gibson RG, Gibson SLM, Conway V, Chappell D. Perna canaliculus in the treatment of arthritis. Practice 1980;224:955-60.
24. Huskisson EC, Scott J, Bryans R. Seatone is ineffective in the treatment of osteoarthritis. J Am Vet Med Assoc 1993;202:288–302.
26. Mani S, Lawson JW. In vitro modulation of inflammatory cytokine and IgG levels by extracts of *Perna canaliculus*. *BMC Complement Alternat Med* 2006;6:3.
27. Miller TE, Dodd J, Ormrod DJ, Geddes R. Anti-inflammatory activity of the glycogen extracted from *Perna canaliculus* (NZ green-lipped mussel). *Agents Actions* 1993;38:C139–42.
28. Bauer JE. Evaluation of nutraceuticals, dietary supplements, and functional food ingredients for companion animals. *J Am Vet Med Assoc* 1999;218:1755–60.
29. Budsberg SC, Chambers JN, Van Lue SL, Foutz TL, Reece L. Prospective evaluation of ground reaction forces in dogs undergoing unilateral total hip replacement. *Am J Vet Res* 1996;57:1781–5.
30. Budsberg SC, Johnston SA, Schwarz PD, De Camp CE, Claxton R. Efficacy of etodolac for the treatment of osteoarthritis of the hip joints in dogs. *J Am Vet Med Assoc* 1999;214:206–10.
31. Moreau M, Dupuis J, Bonneau M, Desnoyers M. Clinical evaluation of a nutraceutical, carprofen and meloxicam for the treatment of dogs with osteoarthritis. *Vet Rec* 2003;152:323–9.
32. Theyse LFH, Hazewinkel HA, Van Den Brom WE. Force plate analyses before and after surgical treatment of unilateral fragmented coronoid process. *Vet Comp Orthop Traumatol* 2000;13:335–40.
33. Dobenecker B, Beitz, Kienzle E. A placebo-controlled double-blind study on the effect of nutraceuticals (chondroitin sulfate and mussel extract) in dogs with joint diseases as perceived by their owners. *J Nutr* 2002;132 (Suppl): 1690–1.
34. Audeval B, Bouchacourt P. Etude contrôlée en double aveugle contre placebo, de l'extrait de moule *Perna canaliculus* (moule aux orles vertes) dans la gonarthrose. *Gaz Medicale* [in French] 1986;93:111–6.
35. Bassleer C, Henrotin Y, Franchimont P. In vitro evaluation of drugs proposed as chondroprotective agents. *Int J Tissue React* 1992;14:231–41.
36. Korthauer W, Torre J. Treatment of deforming arthropathy in working dogs with ‘canosan’, a new glycosaminoglycan preparation. *Kleintierprax* 1992;37:467–78.
37. Bucci LR. Chondroprotective agents: glucosamine salts and chondroitin sulfates. *Townsend Lett Dr* 1994;1:52–4.
38. Cleland LG, French J, Betts HW, Murphy G, Elliot M. Clinical and biochemical effects of dietary fish oil supplements in rheumatoid arthritis. *J Rheumatol* 1988;15:1471–5.
39. Kremer JM. Clinical studies on Omega-3 fatty acid supplementation in patients who have rheumatoid arthritis. *Rheum Dis Clin North Am* 1991;17:391–402.
40. Volker D, Garg M. Dietary N-3 fatty acid supplementation in rheumatoid arthritis- mechanisms, clinical outcomes, controversies, and future directions. *J Clin Biochem Nutr* 1996;20:83–97.
41. Cobb CS, Ernst E. Systematic review of a marine nutriceutical supplement in clinical trials for arthritis: the effectiveness of the New Zealand green-lipped mussel *Perna canaliculus*. *Clin Rheumatol* 2005;25:275–84.
42. Aikman H. The association between arthritis and the weather. *Int J Biometeorol.* 1997;40:192–9.
43. Strusberg I, Mendelberg RC, Serra HA, Strusberg AM. Influence of weather conditions on rheumatic pain. *J Rheumatol* 2002;29:335–8.

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