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Clogging Issues with Aquifer Storage and Recovery of Reclaimed Water in the Brackish Werribee Aquifer, Melbourne, Australia

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Abstract: As part of an integrated water-cycle management strategy, City West Water (CWW) is conducting research to develop an aquifer storage recovery (ASR) scheme utilizing recycled water. In this contribution, we address the risk of well clogging based on two ASR bore pilots, each with intensive monitoring. Well clogging is a critical aspect of the strategy due to a projected high injection rate, a high clogging potential of recycled water, and a small diameter injection borehole. Microscopic and geochemical analysis of suspended solids in the injectant and backflushed water, demonstrate a significant contribution of diatoms, algae and colloidal or precipitating Fe(OH)3, Al(OH)3 and MnO2. CWW is, therefore, testing additional prefiltration that includes a 20 µm spin Klin disc and 1–5 µm bag filter operating in series. In this paper, we present optimized methods to (i) detect the contribution of the injectant and aquifer particles to total suspended solids in backflushed water by hydrogeochemical analysis; and (ii) predict and reduce the risk of physical and biological clogging, by combination of the membrane filter index (MFI) method of Buik and Willemsen, a modification of the total suspended solids method of Bichara and an amendment of the exponential bacterial growth method of Huisman and Olsthoorn.

Keywords: ASR; recycled water; well clogging; geochemical analysis; filtration; biofouling; risk management

1. Introduction

The use of reclaimed or recycled water is on the rise worldwide, mainly because of (i) water scarcity due to exponential population growth and climate change, and (ii) the need to reduce the pollution load from waste water that ends in surface water systems [1,2].

As part of an integrated water-cycle management strategy for the fast growing city of Melbourne, City West Water (CWW) is conducting research to apply aquifer storage recovery (ASR) utilizing recycled water [3]. This non-potable water is to be injected and stored during winter in a brackish, anoxic sand aquifer at 220–250 m below ground level (BGL), and recovered during peak demand periods in summer. The purpose is to reduce drinking water consumption by supplying recycled water via a third pipe system. The infiltration water will be Class A recycled water, which is fit for high exposure uses (not for drinking water consumption), diluted with desalinated water (by reverse osmosis). Class A recycled water comes from Melbourne Water Corporation’s Western Treatment Plant, which produces recycled water from a series of anaerobic and aerobic lagoons followed by UV disinfection and chlorine disinfection.
Several studies have confirmed the viability of ASR at the trial site (West Werribee, west of Melbourne; Figure 1), based on the acceptable overall quality of the infiltration water [4,5], the transmissivity and dispersivity of the aquifer and background flow [4], geochemistry of aquifer [6,7], the ambient water quality in the target aquifer [5], predicted water quality of recovered water [6,8], and economic, societal and water security implications [3].

Figure 1. Site map showing the location of the aquifer storage recovery (ASR) (production) and monitoring wells. ASR well 5 was the production well during the trial in 2012, with its monitoring well 7 at ca. 40 m distance. Production wells 20–24 form together the future West Werribee ASR plant, but for now they are part of the second pilot, with well 20 as ASR well and 21–24 as monitoring wells. AB = geological section shown in Figure 2.

Nevertheless, two recognized main risks still warrant ongoing investigation of respectively the actual well clogging risk [7,9], and risk caused by undesired water quality changes in the target aquifer [5]. Injection well clogging can be extremely cumbersome leading to high costs due to monitoring, maintenance, premature well replacement, intensification of pretreatment or addition of more wells [9–12].

Common water quality issues with ASR application, especially when dealing with recycled water, consist of (i) mobilization of trace metals and arsenic by pyrite oxidation [13]; (ii) lack of full elimination of high concentrations of NO$_3$ and PO$_4$; (iii) the formation of trihalomethanes and haloacetic acids by chlorination [14]; (iv) the persistence of some pathogens and organic micropollutants in the aquifer [2,15]; and (v) the undesired admixing of ambient groundwater (total dissolved solids (TDS), H$_2$S, natural radionuclides) [5]. There are also concerns about potential changes in the microbiota and stygofauna (taxa that spend their whole life cycle in groundwater) by introducing among others oxygen, algae and xenobiotic compounds [16–18].

In this contribution, we evaluate the risks of ASR well clogging based on a short ASR pilot investigation in 2012, and an ongoing ASR pilot test which started in 2017. We present hydraulic, geochemical and microscopic data, diagnose the main clogging causes, and indicate how to mitigate or prevent the problem.
2. Materials and Methods

2.1. Aquifer Storage Recovery (ASR) Pilot Tests

Two pilot tests were conducted in 2012 and 2017–2019, respectively. The first used one injection bore (well 5) and well 7 served as a monitoring bore (Figure 1). The second test (still running) consists of one injection bore (well 20), wells 21–24 as nearby and wells 12–15 as remote monitoring bores (Figure 1).

Details of the ASR cycling scheme are summarized for both pilot tests in Table 1. The 5 clustered wells 20–24 are part of the future ASR system, which is expected to infiltrate 0.5–1.0 Mm$^3$/year during winter time and to recover ~80% of this volume during summer time, depending on demand. The ASR wells include a stainless steel (316) telescopic design, with internal diameter of 350 mm in the upper section, 200 mm in the lower section (incl. well screen), and screen aperture ranging from 0.4 to 0.8 mm. They are not supplied with a gravel pack nor a downhole flow control valve.
Table 1. Details of ASR cycling scheme during the first and second pilot. CSV = cumulative stored volume; NSV = net stored volume (injected minus recovered); ASV = actual remaining stored volume (bubble volume); RE = recovery efficiency. Red numbers = input data in spreadsheet.

| Cycle No | Injection Period | Storage Period 1 | Recovery Period | Storage Period 2 | Total for Cycle | Time since Start | Injection Rate | Pumping Rate | RE per Cycle | RE cumul. | CSV | NSV | ASV |
|----------|------------------|------------------|-----------------|------------------|-----------------|-----------------|---------------|--------------|-------------|-----------|-----|-----|-----|
| 1        | 10 April–14 June 2012 | 0.83            | 1.05            | 0.99            | 2.97            | 5.8             | 5.8            | 36.0         | 108.0       | 357.6     | 358 | 720 | −1855 |
| 2        |                  | 3.00            | 3.02            | 2.10            | 0.07            | 8.2             | 14.0           | 72.0         | 90.0        | 87.7      | 121 | 5904 | −1216 |
| 3        |                  | 14.88           | 20.00           | 16.17           | 0.00            | 51.0            | 65.1           | 72.0         | 108.0       | 163.0     | 155 | 31,608 | −17,416 |
| SUM (day)|                  | 18.71           | 24.07           | 19.26           | 3.04            | SUM (m$^3$)     | 31,608        | 49,024       | SUM (m$^3$) | SUM (m$^3$) | R$_{END}$ (m) = 0.0 |
| 1        | 10 July 2017–likely September 2019 | 1.00            | 4.00            | 1.00            | 5.00            | 11.0            | 11.0           | 54.0         | 104.4       | 193.3     | 193 | 1296 | −1210 |
| 2        |                  | 7.00            | 21.00           | 7.00            | 7.00            | 42.0            | 53.0           | 54.0         | 104.4       | 193.3     | 193 | 10,368 | −9677 |
| 3A       |                  | 13.00           | 386.00          | 0.00            | 0.00            | 399.0           | 452.0          | 54.0         | 0.0         | 0.0       | 74  | 27,216 | 7171  |
| 3B #     |                  | 34.00           | ??              | ??              | ??              | ONGOING         | 54.0           | 0.0          | ??          | ??        | ??  | 71,280 | ??   |
| SUM (day)|                  | 21.00           | 411.00          | 8.00            | 12.00           | SUM (m$^3$)     | 27,216        | 20,045       | SUM (m$^3$) | SUM (m$^3$) | R$_{END}$ (m) = ?? |

*: Including prefiltration over 20 µm and 1-5 µm; ?? = recovery cycle not yet started; R$_{END}$ = radius injected water at end.
2.2. Characterization of Target Aquifer

The ASR target aquifer consists of the coarser grained sequences within the Lower Werribee Formation (LWF) of Early Tertiary age. It is formed by a heterogeneous sequence of sands, silts, clays and lignite, slightly dipping on average in SE direction. The LWF rests on nearly impervious Lower Paleozoic basement rock, and is overlain by 200 m of (un)consolidated sediments (Figure 2).

In the LWF, the sands are typically found between 220 and 240 m below sea level. They vary in grain size (0.1–0.8 mm) and thickness (3–22 m) over short distances. Most wells have two screen sections separated by a blind casing interval of 2–4 m in the middle, where the aquifer is too silty. Average aquifer characteristics are listed in Table 2, based on pumping tests [4], breakthrough curves [5], sedimentological and geochemical analysis of aquifer cores [7,19].

Table 2. Characteristics of principal wells in both ASR pilots, and of local aquifer. \( r_{MW} \) = radial distance to monitoring well; D = thickness, \( T = \) transmissivity, \( S = \) storativity, \( \varepsilon = \) effective porosity, \( t_{50} \) = travel time, \( \alpha_L \) = longitudinal dispersivity.

| Code       | Screen m BGL | \( r_{MW} \) m | D m | \( T \) m²/day | S | \( \varepsilon \) | \( t_{50} \) day | \( \alpha_L \) m | Geochemistry |
|------------|---------------|----------------|-----|----------------|---|----------------|----------------|----------------|--------------|
| ASR well 5 | 220–236       | 50             | 15.5| 71             | 0.00035 | 0.3          | 0.5            | -              | -            |
| MW 7       | 237–246       | 100            | 20  | 70             | 0.0002  | 0.3          | 13             | 0.4            | 22,214 #     |
| ASR well 20| 242–251       | 100            | 11  | 77             | -         | 0.3          | 60             | 1.0            | <0.1 # <0.2 # |
| MW 21      | 223–235       | 100            | 11  | 77             | -         | 0.3          | 60             | 1.0            | -            |

#: in WTP3, ca. 1720 m south of MW 7. Sample probably representative of silty layer, not sand.

Figure 2. Generalised NNW-SSE geological section showing Werribee Formation and ASR pilot sites. (Left): large scale, modified after [20]; (Right): smaller scale, slightly modified after [21].

2.3. Water Quality Analysis

Samples of infiltration water and of water from wells were taken at the well head, with and without prior filtration over 0.45 µm in the field. The standing volume in well screen and riser was always evacuated 3 times prior to sampling. In the field, \( \text{pH} \), electrical conductivity (EC), temperature, oxidation reduction potential (ORP) and \( \text{O}_2 \) were measured at selected intervals. Sensors of EC, temperature and water pressure, installed at screen depth in the wells, have been registered between 3 and 10 min.

Sample container type, preservation method, holding time and analytical method were as recommended by ALS Environmental [22], which performed the analyses on a wide spectrum of parameters: general chemistry (\( \text{pH} \), EC, turbidity, total suspended solids (TSS), algae), major ions, nutrients, total and dissolved organic carbon (TOC and DOC, respectively), (trace) metals,
gases (O\textsubscript{2}, CO\textsubscript{2}, H\textsubscript{2}S, CH\textsubscript{4}), disinfection by-products, industrial organic micropollutants, pesticides, pharmaceuticals, radionuclides, and microbiological parameters. Sampling frequency was on average on a weekly basis but fine-tuned to the duration of ASR cycles, the breakthrough in monitoring wells, and costs of analytical packages [8].

2.4. Analysis of Suspended Matter during Backflushing

Backflushing of the ASR well, with a 2–3 times higher pumping than injection rate, directly or after juttering, is a method to unclog the well screen or bore hole wall. Samples of the turbid backflush water were taken from ASR well 20 on 4 occasions (A–D): during injection cycle 2 (A), two well redevelopments after cycle 3A (B, C) and during injection cycle 3B (D), respectively. Samples of unfiltered water were taken after evacuation of 9–12 m\textsuperscript{3}, corresponding with ~1 standing well volume. The samples (3.6–4.2 L) were transported to the lab in an upright position, in the dark and cooled at 4 °C, without shaking. In the lab, these samples were kept in the dark at 4 °C for 2 days in upright position without shaking/tremors, in order to let the suspended matter settle. Subsequently as much clear water as possible was slowly decanted or sucked out, containing on average 95% of all water with on average 2.7% of all TSS: 1 mg TSS/L in 0.95 × 4 L and 690 mg TSS/L in 0.05 × 4 L yields indeed 100 × (0.95 × 4 × 1)/(0.95 × 4 × 1 + 0.05 × 4 × 690) = 2.7%, which is neglected in Equation (1).

The decanted water was analyzed, after 0.45 µm filtration, for all main constituents (incl. pH and EC), DOC and trace elements. This yields the dissolved fraction. The remaining fluid (0.2–0.5 L) with 97% of all TSS was first analyzed for TSS, and subsequently, after nearly complete destruction with strong suprapure HNO\textsubscript{3} at 95 °C, analyzed for TOC, total concentration of main constituents (excluding Cl, HCO\textsubscript{3}, NO\textsubscript{3}) and trace elements.

The analytical results of the dissolved and suspended fraction in mg/L were used to calculate the composition of the suspended material as follows:

\[ X_{SS} = 10^6 \frac{X_T - X_{H2O}}{TSS}, \]  

where: \( X_{SS} \) = content of component \( X \) in suspended solids (ppm = mg/kg dry weight); \( X_T \) = total \( X \) concentration in water with suspended solids (mg/L); \( X_{H2O} \) = concentration of \( X \) dissolved in water (mg/L); \( TSS = \) total suspended solids in water (mg/L).

2.5. Clogging Predictor Based on Membrane Filter Index (MFI)

Buik and Willemsen [23] used the membrane filter index (MFI; [10,24,25]), well and hydrogeological parameters to predict the clogging rate of recharge wells, based on the following semi-empirical equation (without dimensional homogeneity):

\[ v_{CLOG} = 2 \times 10^{-6} \times MFI \times t_{EQ} \times V_{DIR}^2 / (K_h / 150)^{1.2}, \]  

with:

\[ K_{h} = 150(D_{50}/1000)^{1.65} \text{ or } K_{h} = T/D, \]  

\[ V_{DIR} = Q_{IN} / (2\pi r_{B}L), \]  

where: \( v_{CLOG} \) = clogging rate (m/a); \( MFI = 0.45 \text{ µm membrane filter index (s/L²)}; t_{EQ} = \) total amount of equivalent full load hours per year (h); \( V_{DIR} = \) entrance velocity on borehole wall (m/h); \( D_{50} = \) median grain size diameter of aquifer (µm); \( Q_{IN} = \) mean infiltration rate when well is recharging (m\textsuperscript{3}/h); \( r_{B} = \) radius of borehole (m); \( L = \) length of well screen (m); \( K_{h} = \) horizontal hydraulic conductivity (m/d).

2.6. Clogging Predictor Based on Total Suspended Solids (TSS)

Another way to predict the physical clogging rate, is to relate the decrease in flow rate of an injection well \( (Q_d/Q_0) \) to the ratio of the total input of suspended solids after a given injection time,
to the open area of the external borehole wall \((A_{\text{OPEN}})\). On the basis of many experimental data
Bichara [26] showed an interesting plot of that relation. Notwithstanding the scatter of his many data,
an average linear trend can be used to estimate the decline in flow rate of an injection well by particle
clogging:

\[
\frac{Q_t}{Q_0} = 100 - 17.5 \times \frac{Q_{\text{IN}}}{t \times A_{\text{OPEN}}}, \quad \text{if } t > 0, \text{ else } 0
\]  

\[
t_{10} = \frac{10}{t_0 - \frac{Q_t}{Q_0}}, \quad \text{if } t > 0, \text{ else } 0
\]

with:

\[
A_{\text{OPEN}} = 2\pi r B \varepsilon L, \quad (m^2)
\]

where: \(Q_t, Q_0\) = injection rate at time \(t\) since start and at start \((t = 0)\), respectively \((m^3/h)\); \(\varepsilon\) = aquifer porosity (-); \(t\) = time since injection start (day); \(t_{10}\) = injection period needed to reduce flow by 10% (day).

2.7. Predictor of Bioclogging

Bioclogging can be simulated by the following equation (modified after Huisman and
Olsthoorn [27], by introducing a lag time), which calculates the buildup of bacteria in infiltration wells:

\[
N_t = \frac{V_{\text{DIR}} N_0 T_D \left(e^{0.693 \frac{t}{t_{\text{LAG}}}} - 1 \right)}{0.693}, \quad (m^{-2})
\]

where: \(N_0\) = average number of bacteria in input \((n/m^3)\); \(N_t\) = average number of bacteria at and
behind bore hole wall, at time \(t\) \((n/m^2)\); \(t\) = time since injection start (day); \(t_{\text{LAG}}\) = lag time during
which bacteria do not reproduce (day); \(T_D\) = doubling time for bacterial population (day); \(V_{\text{DIR}}\) = entrance velocity on borehole wall, see Equation (4) \((m/day)\).

Thus, it is assumed that each input bacterium is filtered out at the borehole wall and starts
multiplying after a specific lag time. The thickness of the biofouled layer \(D_{\text{BAC},t}\) which fully occupies
the aquifer pore space, can be calculated using Equation (9) by assuming that (i) the accumulation of
cells with a specific cell volume is taking place on the borehole wall and behind that, both with open
space \(A_{\text{OPEN}}\), and (ii) there is no open space between bacteria:

\[
D_{\text{BAC},t} = \frac{1000 N_t v_{\text{CELL}}}{(1 - \varepsilon)}, \quad (mm)
\]

where: \(v_{\text{CELL}}\) = spherical volume of each cell \((m^3)\); \(\varepsilon\) = aquifer or gravel pack porosity (-).

A more realistic model sets a limit to bacterial growth, based on e.g., the observed change in DOC
or biodegradable DOC (BDOC; [28]). The available data show a very small decline of the injectant
during short storage or recovery cycles \((1-2\text{ days})\) for DOC \((<0.5\text{ mg C/L})\) and BDOC \((-0.2\text{ mg C/L})\).
This observed consumption of BDOC during a 1–2 days lasting storage or recovery cycle \((\Delta_{\text{BDOC}};\text{ mg}
C/L)\), is the sum of bacterial respiration (BaR) and biomass production (BoP; [29]), so that:

\[
\text{BoP} = f_{\text{BoP}}(10^{-3} \Delta_{\text{BDOC}} Q_{\text{IN}} - \text{BaR}) = f_{\text{BoP}} 10^{-3} \Delta_{\text{BDOC}} Q_{\text{IN}}, \quad \text{ (kg C/day)}
\]

where: \(Q_{\text{IN}}\) = injection rate \((m^3/d)\); \(f_{\text{BoP}} = \text{BoP}/(\text{BoP} + \text{BaR}) = \text{fraction of } \Delta_{\text{BDOC}}\text{ consumed by bacterial}
respiration \((-); \Delta_{\text{BDOC}} = \text{consumption of biodegradable DOC} \text{ (mg C/L)}\).

We assume that \(\Delta_{\text{BDOC}}\) represents steady state, and that it approaches consumption by the total
number of active cells at maximum growth, so that:

\[
\Delta_{\text{BDOC}} = f_{\text{BoP}} 10^{-3} \Delta_{\text{BDOC}} Q_{\text{IN}} / V_{\text{CELL}} C_{\text{CELL}}
\]

\[
N_{\text{MAX}} = \frac{10^{-3} f_{\text{BoP}} \Delta_{\text{BDOC}} Q_{\text{IN}}}{V_{\text{CELL}} C_{\text{CELL}} A_{\text{OPEN}}} = \frac{10^{-3} f_{\text{BoP}} \Delta_{\text{BDOC}} V_{\text{DIR}}}{V_{\text{CELL}} C_{\text{CELL}} \varepsilon}, \quad (m^{-2})
\]
where: \( C_{\text{CELL}} = \) Carbon bacterial mass, 220 kg dry weight/m\(^3\) according to [30].

\( N_{\text{MAX}} \) can also be calculated with Equation (8) by replacing \( t \) by \( t_{\text{MAX}} \), which is defined as the time since start of exponential bacterial growth to the maximum level of growth. Combination of this Equation (8) with Equations (11) and (4) yields after rewriting, for both limitation cases:

\[
t_{\text{MAX}} = \frac{T_D}{0.693 \ln \left( \frac{0.693N_{\text{MAX}}}{N_0 T_D V_{\text{DIR}}} + 1 \right) + t_{\text{LAG}}}, \quad \text{(day)}
\]

(12)

3. Results

3.1. Injection Well Clogging

A typical pattern of impressed head and drawdown, TSS (at well head) and turbidity (in central injection manifold) for ASR well 20 during ASR Cycle 3, is shown in Figure 3. The peaks of TSS and turbidity coincide with 9 short flow reversals due to backpumping (108 m\(^3\)/h) and reinjection (54 m\(^3\)/h). This triggered the mobilization of fines from the borehole wall and probably also from the well itself. TSS peaked during both switches (injection to backpumping, and vice versa), whereas turbidity only peaked during the switch from backpumping to injection, due to the position of the turbidity sensor right after the injection pump. The TSS sensor’s output is maximum 20 mg/L in order to focus on lower level variability, but short peaks of up to 43.5 (=870/20) mg/L have been measured (Table 3).

Table 3. Summary of analytical results of suspended matter sampled during backflush events A–D. The column with heading ‘A–C’ contains the average of events A, B and C (without prefiltration of injectant). Also shown: composition of ASR input TSS (average 6 samples) and of an aquifer core (1 sample), and the calculated fraction of the ASR input contributing to the average composition of samples A–C \( (\alpha) \). Numbers in red indicate a significant trend over time (see Figure 4).

| Event # | Vol. out | Unit TSS Input ASR | SUSPENDED MATTER DURING BACKFLUSHING | Aquifer Core | \( \alpha \) A–C |
|---------|----------|-------------------|-------------------------------------|-------------|----------------|
|         | m\(^3\)  | mg/L              | A 12.0 | B 12.0 | C 9.0 | D 11.0 | A–C  | A–C |
| TSS     |          | -                 | -     | -     | -     | -     | -    | -    |
| \( C_{\text{ORG}} \) | ppm      | 0.23              | -     | -     | -     | -     | -    | -    |
| Si      | ppm      |                   | 23,364 | 8433 | 15,091 | 35,500 | 15,629 | -    | -    |
| S       | ppm      | 1                 | 13,811 | 9591 | 7113   | 31,842 | 3488  | 108  | 0.30 |
| P       | ppm      |                   | 900   | 2180  | 2459   | 10,145 | 1846  | 140  | -    |
| Al      | ppm      | 18,400            | 36,198 | 6632  | 8844   | 12,261 | 17,225 | 22,300 | -    |
| Fe      | ppm      | 32,583            | 29,293 | 19,517 | 18,025 | 26,043 | 22,278 | 15,500 | 0.40 |
| Mn      | ppm      | 11,533            | 1462  | 3994  | 5007   | 31,842 | 3488  | 108  | 0.30 |
| Co      | ppm      | 100               | 35    | 29    | 34     | 121    | 32    | 2.2  | 0.31 |
| Cr      | ppm      | 87                | 144   | 42    | 58     | 111    | 81    | 193  | 1.05 |
| Cu      | ppm      | 200               | 55    | 62    | 38     | 157    | 52    | 11.4 | 0.21 |
| Ni      | ppm      |                   | 53    | 54    | 41     | 155    | 49    | 6.4  | -    |
| Sr      | ppm      |                   | 157   | 52    | 56     | 142    | 88    | 25   | -    |
| Ti      | ppm      | 217               | 1895  | 229   | 277    | 332    | 800   | -    | -    |
| V       | ppm      | 22                | 122   | 26    | 42     | 18     | 63    | 85   | 0.34 |
| Sum $ $  | %        | 6.3               | 11.3  | 5.7   | 6.9    | 17.8   | 8.4   | 5.3  | -    |
| Al/Fe   | ppm      | 0.56              | 1.24  | 0.34  | 0.49   | 0.47   | 0.77  | 1.44 | -    |
| Fe/Mn   | ppm      | 2.8               | 20.0  | 4.9   | 3.6    | 0.8    | 6.4   | 143.5| -    |
| Fe\(_{\text{S}}\) | mmol/kg | 0                 | 215   | 150   | 111    | 45     | 159   | 186  | 0.15 |
| FeFe\(_{\text{S}}\) | ppm | 1 | 12,028 | 8353 | 6194 | 2526 | 8858 | 10,363 | 0.15 |
| Fe-rest | ppm      | 32,582            | 17,265 | 11,164 | 11,830 | 23,517 | 13,420 | 5137 | 0.30 |

$ $: A = 24 Aug. 2017; B = 3 Nov. 2017; C = 13 Dec. 2017; D = 19 Dec. 2018; $ $: Sum = (sum all elements)/10\(^4\); $ $: TSS in ca. 20 times concentrated water sample; A–C injected water without prefiltration; D injected water with prefiltration over 20 \( \mu \)m + 5 \( \mu \)m.

The backflushing is successful in removing the bulk of clogging material, but a growing residual (chronic) clogging is remaining, as evidenced by the rising impressed head (Figure 3) during identical successive injection runs such as 3.7, 3.9 and 3.10 (not fully shown). The clogging process was linear.
during the 15 days lasting injection period 3 of the first ASR trial [4], which is typical for clogging mainly caused by particles [10].

Figure 3. Impressed head/drawdown, total suspended solids (TSS, at well head) and turbidity (in central injection manifold) for ASR well 20, during ASR Cycle 3A (with 10 sub-cycles divided by backflush events), with zooming in on 25 October 2017 10–18 h. Note the TSS cut-off at 20 mg/L, and the logarithmic increase of impressed head.

3.2. Results of Hydrogeochemical Analysis of Suspended Material

The composition of the suspended solids was calculated for each sample taken during the 4 backflushing events A–D, by applying Equation (1). Each sample was taken after on average 1.5 days injection of 1950 m$^3$, and after ~5 min backflushing (corresponding with 11 m$^3$ and 1 standing volume in the well). The results are presented in Table 3, together with a summary of the geochemical analyses of aquifer cores and TSS of the injectant. Zero values mean that $X_T$ equals $X_{H2O}$, so that practically all of $X$ is in dissolved form. Empty cells in Table 3 (with ‘-’) mean no data available.

3.2.1. Average Composition

In Table 3, the average composition is listed for samples of events A–C. The sample of event D was excluded because it refers to a different input (see Section 3.2.2). On average ~8.4% of the suspended
material has thus been identified and quantified. This means that 92% is not covered by the analysis, due to (i) lack of dissolution in the acid and oxidizing leach (notably quartz), (ii) lack of the usual conversion of the elements in their oxide form, and (iii) no inclusion of $H_2O$ which is present in the crystal lattice of some minerals such as amorphous $Fe(OH)_3$ and $Al(OH)_3$.

The highest contents are noticed for the following main constituents, in decreasing order: $Si$ (underestimated due to incomplete extraction), $Fe$, $Al$, $S$, $C$, $Mn$ and $P$. The true $Si$ content is estimated at max 37% (not the 1.6% extracted; 37% = (100% – sum $X_nO$-bound $H_2O$)/$MW_{SiO_2}$, where $X_nO$ = all individual elements, $SiO_2$ excluded, each transformed in its oxide form, e.g., $C$ in $CH_2O$ and $Fe$ in $Fe_2O_3$). $Si$ is mainly linked to $SiO_2$ as quartz and diatoms. Concentrations of $Na$, $K$, $Ca$ and $Mg$ were <0.2%.

The high $S$ content very likely indicates the presence of iron sulfide particles, because gypsum can be excluded and the alternative, organic matter, contains on average only 1.8% $S$ (0.018 $×$ $C_{ORG}$ = 127 ppm $S$). Thus assuming the remaining 10,045 ppm (=10,172 – 127) to be $FeS_2$ (pyrite) requires the presence of enough $Fe$, namely 8858 ppm. This would mean that the $TSS$ contains an excess of 13,420 ppm $Fe$, which could be $Fe(OH)_3$ or $Fe_2O_3$ deriving from the source water directly or indirectly (from biofilms on the transmission pipeline, a storage tank or ASR well casing), and from rust particles from corroding stainless steel parts of the transmission pipeline, storage tank or ASR well casing).

3.2.2. Trends

The composition of samples from backflushing events A–D varies over time, in a more or less systematic way for the following elements (red numbers in Table 3; Figure 4): (i) $P$ and $Mn$ steadily increasing, and (ii) $S$, $FeS_2$, $Fe-FeS_2$, $Fe/Mn$ and $V$ steadily decreasing. This indicates that over time, the infiltration water input is contributing more ($P$ and $Mn$) and aquifer material less ($pyrite and $V$) to $TSS$ in the backflushed water. This agrees with expectations.

![Figure 4](image-url). Changes in element content of suspended solids in a sample of backflushed water from ASR well 20 during backflushing events without (A–C) and with prefiltration (D). Based on Table 4. Arrows indicate significant trend over time.
Table 4. Results of microscopic analysis of suspended material in source water (SW) and backflushed water, in 2017. 1–7 = relative abundance low to very high. Samples 1, 2, 3 = after 5, 15 and 30 ‘equivalent’ minutes of pumping. Samples coded 1 (taken after 5 min) correspond with events A, B and C in Table 3. Sample 1 on 3 November shown in Figure 5.

| Parameter                          | SW 13 Sept. | 24 Aug. | Backflush |
|------------------------------------|-------------|---------|-----------|
| Sample #                           |             |         |           |
| Time to filter 100 cc (s)          | 15          | 30      | 60        |
| Fine amorphous debris              | 3           | 7       | 5         |
| Mainly non-filamentous bacteria    | 3           | 5       | 4         |
| Diatom frustules                   | 2           | 2       | 1         |
| Viable diatoms                     |             | 3       |           |
| Blue-green algae                   | 1           | 1       | 1         |
| Other algae                        | 1           | 1       | 1         |
| Zooplankton                        | 1           |         |           |

The sample from backflushing event D showed a remarkable overall concentration increase (S and V excluded). The reason for this increase is not clear, but could be related to the applied prefiltration step, which resulted in the accumulation of finer grained particles on the borehole wall and behind. These finer grained particles could contain less quartz particles and more soluble material.

3.2.3. Comparison with Aquifer Cores and Infiltration Water

In Table 3, the element content of TSS can be compared with the aquifer core data and with the average TSS composition of infiltration water. We selected the mean TSS composition of backflush events A–C, the injectant (without prefiltration) and the core data from monitoring well 3 (~1 km south of well 15, Figure 1). The core had about the same extraction and geochemical analysis as TSS, and showed a similar aquifer sedimentology as well 20. These data have been used in Table 3 to estimate the fraction of suspended particles in the infiltration water ($\alpha$) and of aquifer particles (1 – $\alpha$) contributing to the mean TSS composition of the backflush sample. The following unmixing equation for 2 end-members was used:

$$\alpha = \frac{X_{SS} - X_{AQUIFER}}{X_{INPUT} - X_{AQUIFER}}.$$  \hspace{1cm} (13)

Fraction $\alpha$ appears lowest (0.15) for S (and associated FeS$_2$ and Fe-FeS$_2$), so that practically all S (pyrite bound) in TSS is derived from the aquifer, as deduced earlier. Fraction $\alpha$ increases slightly for Cu, Mn, Co, V and Fe (from 0.21 to 0.40, respectively), which indicates that the aquifer is forming their mean supply during events A–C. Only P, Cr, Ni and Sr seem to be largely delivered by the injectant during events A–C, Cr perhaps even totally. The calculated $\alpha$ numbers are not very accurate due to temporal variations in the injectant and spatial variations in aquifer geochemistry.

An important assumption in this interpretation is, that the mentioned ions did not (co)precipitate in the water sample during transport from well to the lab and during 2 days of air-tight bottle detention prior to the separation of decantable fluid from the remaining high TSS liquid. We checked this by comparing the infiltration water with the decanted fluid, and the decanted fluid of the first with the last sample. Concentrations of dissolved Al, Fe, Mn NO$_3$, NH$_4$ and PO$_4$ did not show significant changes, indicating that the potential contribution of Fe or Mn flocs formed by oxidation of the backflushed water was rightly ignored.

3.3. Results of Microscopic and Particle Size Analysis of Suspended Material

Suspended material in source water and in backflushed water was examined by microscopic analysis. The results are summarized in Table 4, and a characteristic image is shown in Figure 5.
The diatoms and other algae in the backflushed water are clearly derived from the infiltration water input, because they are too undamaged and some are even still viable.

**Figure 5.** Microscopic image (×100 magnification) of backflushed material on 3 November 2017 (event B in Table 3), showing diatoms (incl. fragments), algae, mostly non-filamentous bacteria, and amorphous flocs.

Results of particle size analysis by Light-Scattering using a Mastersizer 3000 instrument, after sonication or addition of a dispersant, are shown in Figure 6. In the backflushed water, 3 peaks can be observed in the particle size distribution: around 0.5, 7 and 200 µm. In the infiltration water, there is a large peak around 13 µm with 2 small, ill-defined peaks around 0.8 and 800 µm. The difference between both particle size distributions could be not representative due to fluctuations in particle load and particle size distribution of either the infiltration water or backflush water. The peak around 200 µm in the backflush water could be due to either a rather high number of elongate diatoms >200 µm long (Figure 5), which the analyser underestimates due to their shape, or sand intake by the well.
Table 4. Results of microscopic analysis of suspended material in source water (SW) and backflushed water, in 2017. 1–7 = relative abundance low to very high. Samples 1, 2, 3 = after 5, 15 and 30 ‘equivalent’ minutes of pumping. Samples coded 1 (taken after 5 min) correspond with events A, B and C in Table 3. Sample 1 on 3 November shown in Figure 5.

| Parameter               | SW 13 Sept. | Backflush 24 Aug. | Backflush 3 Nov. | Backflush 13 Dec. |
|-------------------------|-------------|-------------------|------------------|-------------------|
| Time to filter 100 cc   | 15          | >600              | 98               | 20                |
| Fine amorphous debris   | 3           | 5                 | 5                | 4                 |
| Mainly non-filamentous bacteria | 3 | 5 | 2 | 1 |
| Diatom frustules        | 2           | 3                 | 1                | 1                 |
| Viable diatoms          | 3           | 1                 | 1                | 1                 |
| Blue-green algae        | 1           | 1                 | 1                | 1                 |
| Other algae             | 1           | 2                 | 1                | 2                 |
| Zooplankton             | 1           |                   |                  |                   |

Results of particle size analysis by Light-Scattering using a Mastersizer 3000 instrument, after sonication or addition of a dispersant, are shown in Figure 6. In the backflushed water, 3 peaks can be observed in the particle size distribution: around 0.5, 7 and 200 μm. In the infiltration water, there is a large peak around 13 μm with 2 small, ill-defined peaks around 0.8 and 800 μm. The difference between both particle size distributions could not be representative due to fluctuations in particle load and particle size distribution of either the infiltration water or backflush water. The peak around 200 μm in the backflush water could be due to either a rather high number of elongate diatoms >200 μm long (Figure 5), which the analyser underestimated due to their shape, or sand intake by the well.

Figure 6. Particle size distribution of infiltration water (top) and backflushed water (bottom), both sampled on 13 December 2017 (event C in Table 3).

3.4. Evaluation of Clogging Risks

3.4.1. Global Evaluation

ASR well clogging can be caused by any of the following, potential clogging processes (or their concerted action): (i) suspended solids in the infiltration water such as clay, algae, diatoms and Fe(OH)₃ flocs, (ii) biofouling due to high concentrations of biodegradable organic matter (TOC, assimilable DOC (AOC), BDOC) and nutrients (PO₄, NO₃), (iii) air entrainment (gas bubbles formed during cascading in the well), (iv) chemical clogging by precipitates of e.g., Fe(OH)₃, MnO₂ or CaCO₃, (v) clay swelling and clay mobilization due to replacing brackish ambient groundwater with high SAR (sodium adsorption ratio) by fresher, lower SAR Injectant, and (vi) permeability reduction by aquifer jamming or aquifer corrosion.

The last four processes are considered of minor importance. Well clogging by gas bubbles is unlikely because of a proper well design [4] and a non-corresponding clogging pattern [10]. The precipitation of minerals by mixing with ambient groundwater during injection phase can be ruled out, because the Fe concentrations in the ambient groundwater are low (0.16 mg/L) and the flow velocity near the well is sufficiently high and reactions are so slow, that reaction products such as iron hydroxide flocks will move relatively far away from the well before being deposited, while the mixing of (sub)oxic water with anoxic ambient groundwater will mainly take place far away from the well. Experiences with and calculations on intentional subterranean iron removal (SIR) [31] indicate that the accumulation of reaction products during both injection and recovery can be neglected on the time scale of one century. SIR is also an unintentional process during ASR [32], which prevents Fe(II) during recovery to reach the ASR well and mix there with any O₂ (if this survived the aquifer detention at some depth). The risk of CaCO₃ precipitation is also very low, because the injectant and ambient groundwater are undersaturated with respect to CaCO₃. Another potential cause of mineral precipitation is chemical instability of the injectant, which can lead to retarded, in-well flocculation of Fe(OH)₃ or MnO₂. This process is hard to distinguish from the advective transport of suspended particles, and therefore not considered further.
Barry et al. [7] investigated the clogging risk by dispersion of the clay minerals within columns filled with aquifer material, especially within the context of potential future changes in the mixing ratio of Class A water and RO desalinated water. They used a method based on the principles of the ‘Emerson Crumb test’ [33–35]. That approach characterizes dispersion using combinations of aquifer materials and source waters by measuring the turbidity increase of the supernatant, relative to that of the water alone, after a period of 48 h. Dispersion of clays in the batch tests was so low that no visible cloudiness was observed in any of the waters tested. Although kaolinite, the predominant clay mineral in the target aquifer, is a non-swelling clay and present within the aquifer storage zone at very low concentrations (<1%), its interaction with low salinity water can still contribute to a reduction in aquifer permeability by detachment from solid surfaces and migration through porous media where it can clog pore spaces [36]. In conclusion, the risk of clay mobilization is (very) low but cannot be ruled out completely, especially not if more RO water is contributing to its mixture with Class A recycled water.

The risk of aquifer permeability losses through aquifer jamming by repeatedly shifting from injection to backflushing, which creates shock waves, is considered low [10]. Aquifer dissolution (not only by the injectant but also by acids applied for well redevelopment) is potentially a risk factor in marley limestone or calcite cemented sandstone aquifers, because the carbonate dissolution will be accompanied by the mobilization of fines that may end up in the pore throats [37,38]. The target aquifer at Werribee is, however, not containing carbonates and thus not vulnerable to this dissolution.

In Table 5, an overview is given of potentially relevant clogging parameters and their levels in the infiltrated Class A recycled water with (second ASR pilot) and without RO water (first pilot), and with (first pilot and cycle 3B of second pilot) and without an onsite prefiltration step (cycles 1–3A of second pilot). The following conclusions are drawn, when comparing the injectant with the given guideline values:

TSS is relatively low (on average 1–2 mg/L), but still above the guideline value for a sandy aquifer (0.1 mg/L). This should lead to physical clogging. Particles containing Fe, Mn and Al probably play a significant role, as deduced from analyzing suspended solids during backflushing, even though the difference between total and dissolved concentrations (the particulate fraction), seems small. MFI is too high, suggesting a high risk of physical clogging. Concentrations of DOC, BDOC ([28]) and AOC ([39]) are high, far above their guideline values. This will lead to biofouling (biological clogging) if without regular backflushing, also because nutrients N + P are not limiting.
Table 5. Overview of parameters that have a potential impact on well clogging, and their average concentration or value in the infiltration water during the three cycles of each pilot. For several parameters a guideline value for deep well recharge is indicated.

| ASR Pilot | TSS | Turb. | DOC | BDOC | AOC | pH | Temp | PO₄-T | NO₃ | NH₄ | O₂ | Fe-T | Fe₅⁺ | Mn-T | Mn₅⁺ | Al-T | Al₅⁺ | Cl₂F | SAR | SAR |
|-----------|-----|-------|-----|------|-----|-----|------|-------|-----|-----|----|------|------|------|------|------|------|-----|-----|-----|
|           | mg/L | NTU | µg C/ac/L | s/L² | °C | mg/L |       |       |  |     |     | | | | | | | | meq/L |
| Guideline $ | <0.1 | <1 | <2 | <0.2 | <10 | <3-5 | | | | | | | | | | | | | |
| ASR 5 100% Class A recycled water | | | | | | | | | | | | | | | | | | | | |
| Cycle 1 | <5 | 2.9 | 10 | 10 | 1.5 | 170 | 6.91 | 22.6 | 113.9 | 0.09 | 8.6 | <0.05 | | <0.01 | | 8.1 |
| Cycle 2 | <1 | 0.8 | 11 | 10 | 2.2 | 160 | 6.94 | 30.4 | 116.1 | 0.03 | 9.6 | <0.05 | | <0.01 | | 8.0 |
| Cycle 3 | <0.1 | 10 | 10 | 2.1 | 185 | 6.9 | 29.7 | 135.1 | 0.21 | 9.2 | <0.05 | | 0.020 | | 7.9 |
| ASR 20 33% Class A recycled water + 67% RO treated Class A recycled water | | | | | | | | | | | | | | | | | | | | |
| Cycle 1 | <2 | 0.50 | 3.90 | 3.90 | 0.50 | | 7.39 | 11.8 | 10.1 | 40.8 | <0.13 | 8.0 | 0.027 | 0.012 | 0.015 | 0.012 | 0.071 | 0.002 | 5.1 |
| Cycle 2 | 0.25 | 3.55 | 3.55 | 0.47 | 47 | 7.16 | 13.0 | 9.3 | 37.4 | 8.8 | 0.099 | 0.020 | 0.014 | 0.008 | 0.029 | 0.023 | 5.0 |
| Cycle 3A | 0.63 | 3.87 | 3.63 | 0.30 | 7.60 | 19.8 | 10.8 | 15.9 | 8.8 | 0.018 | 0.011 | 0.022 | 0.003 | 0.022 | 0.012 | 5.2 |
| Cycle 3B | 2.5 # | 0.69 | 4.41 | 4.32 | 1.35 | 7.62 | 20.2 | 9.3 | 19.6 | 1.03 | 7.5 | 0.032 | 0.012 | 0.038 | 0.021 | 0.047 | 0.020 | 4.8 |

$ = values for deep well injection, partly according to Perez-Paricio & Carrera [40] and Barry et al. [10]; # = prior to prefiltration.
3.4.2. Risk of Clogging by Suspended Particles

We quantified this risk by calculating the clogging rate using the modified Bichara method and the Buik and Willemsen method (Section 2.6). The results of calculation are shown for several scenarios in Table 6 (Bichara’s method) and Table 7 (Buik and Willemsen method).

**Table 6.** Predicted flow reduction ($\frac{Q_t}{Q_0}$ and $\frac{Q_{10}}{Q_0}$) as function of injected particle mass ($\Sigma$TSS = TSS $Q_{IN}$ t) and initial open area of external borehole wall ($A_{OPEN}$). Based on average linearized relation between $\Sigma$TSS/AOPEN and $Q_t/Q_0$ trends in Bichara [26], see Section 2.6. $\psi_0$, $\psi_{MEAS}$ = the impressed head in the injection well as predicted and measured, respectively. Red numbers: significant scenario changes.

| Data Input | Model Output | Verif. |
|------------|--------------|--------|
| TSS mg/L   | ΣTSS kg     | Bichara g/cm² | $\psi_t$ | $\psi_{MEAS}$ |
| Q_{IN} m³/h | A_{OPEN} m² | $Q_t/Q_0$ % | $t_{10}$ day | m |
| $t_{INF}$ d |          |           |         |      |
| L m        |           |           |         |      |
| $r_B$ m    |           |           |         |      |
| n          |           |           |         |      |

| Scenario 1 | 0.0 | 2.5 | 0.000 | 100.0 | - | 0.0 | 0 |
|------------|-----|-----|-------|-------|---|-----|---|
| Scenario 2 | 1.3 | 2.5 | 0.053 | 90.7 | 1.1 | 22.0 | 21.3 |
| Scenario 3 | 2.6 | 2.5 | 0.106 | 81.5 | 1.1 | 24.5 | 24.9 |
| Scenario 4 | 3.9 | 2.5 | 0.159 | 72.2 | 1.1 | 27.7 | 27.1 |
| Scenario 5 | 6.5 | 2.5 | 0.264 | 53.7 | 1.1 | 37.2 | - |
| Scenario 6 | 13.0 | 2.5 | 0.529 | 74.1 | 1.1 | 268.6 | - |
| Scenario 7 | 13.0 | 4.9 | 0.264 | 53.7 | 2.2 | 37.2 | - |
| Scenario 8 | 4.8 | 2.5 | 0.196 | 65.7 | 2.9 | 30.4 | - |
| Scenario 9 | 1.3 | 2.5 | 0.053 | 90.7 | 10.8 | 22.0 | - |

With the modified Bichara method we calculate for ASR well 20 during cycle 3A, resetting the clock after each backflushing event, a flow reduction of about 9% for each day of continuous injection (Table 6). Maintaining the injection rate means that more pressure is needed, which leads to an increase of the impressed head in the well. Intuitively, this increase is proportional to the predicted flow reduction while ignoring temperature effects on viscosity, so that:

$$0.01 \frac{Q_t}{Q_0} = f \frac{\psi_0}{\psi_{t}}$$

so that

$$\psi_{t} = 100 f \frac{\psi_0}{(Q_t/Q_0)} \quad (14)$$

where: $\psi_0$, $\psi_{t}$ = the impressed head in the injection well without (significant) clogging, and with clogging at time $t$ since start, respectively (m); $f$ = empirical fit factor, being zero at $t = 0$, and otherwise $>0$.

Application of Equation (16) to the 3-day-long injection period from 27 October up to and including 30 October 2017 (Figure 3), yields an excellent overlap of calculated with measured $\psi_{t}$ (Table 6), with parameter settings of $f = 2$ (if $t > 0$) and $\psi_0 = 10$ m. Table 6 also shows how much effect can be expected from reducing the injection rate, TSS input or augmenting the backflush frequency or borehole radius.
For ASR well 20 during cycle 3A, with an MFI value of 47 s/L², we calculate with the method of Buik and Willemsen [23] a clogging rate of 520 m/a or 1.42 m/day, assuming 100 days of injection a year (Table 7). This high clogging rate looks much smaller than the one obtained with Bichara’s method (4.3 m versus 27 m in 3 days), but \( v_{\text{CLOG}} \) does not include the water level rise due to unsteady flow as does \( \varphi_t \). Table 7 also shows how much effect can be expected from reducing the injection rate, MFI (by enhanced pretreatment) and equivalent full loading hours, or increasing the bore hole radius or implementing all measures to reduce the clogging rate.

### 3.4.3. Risk of Bioclogging

The results of calculating the number of bacteria (N) and the thickness of the biofouled layer (\( \Delta \text{BAC} \)) are presented for several scenarios in Table 8, and the bacterial growth curve is presented for selected cases in Figure 7. An unrealistic high \( \Delta \text{BACT} \) is predicted for scenarios A and E–G of the second Werribee trial, when an unlimited supply of assimilable carbon (and nutrients and \( \text{O}_2 \)) is assumed, during a long uninterrupted injection run. The observed consumption level of BDOC in the aquifer (\( \Delta \text{BDOC} \)) is believed to represent the biodegradable fraction of DOC, which is then used for both respiration and biomass production. The concentration of \( \Delta \text{BDOC} \) (~0.2 mg C/L) is much lower than the \( \text{NO}_3 \) and \( \text{PO}_4 \) concentrations in the injectant (Table 5) and is, therefore, considered the growth-limiting factor.

**Table 8.** Predicted unlimited growth of bacteria (\( N_\text{t} - N_\text{0} \)) according to Equation (8) and carbon limited growth of bacteria (\( N_\text{tMAX} - N_\text{0} \)) according to Equation (11), and their clogging of open pore space on the borehole wall in terms of thickness of biofouled layer (\( \Delta \text{BAC,t} \) and \( \Delta \text{BAC,max} \); Equation (9)). Also shown is \( t_\text{MAX} \) (Equation (14); time since start of exponential bacterial growth to the maximum level of growth) assuming that carbon supply (with \( \Delta \text{BDOC} \) as indicator) for bacterial growth is the limiting factor. Explanations of parameters in Section 2.7. Red numbers = significant scenario changes.

| Scenario                | Data Input | Calculated | No limitation | + BDOC limitation |
|-------------------------|------------|------------|---------------|-------------------|
|                         | \( Q_\text{IN} \) m³/day | \( L \) m | \( r_B \) m | \( \varepsilon \) Porosity | \( N_\text{0} \) n/m³ | \( T_\text{D} \) day | \( t \) day | \( V_{\text{CELL}} \) m³ | \( t_{\text{LAG}} \) day | \( \Delta \text{BDOC} \) mg/L |
| Werribee well 20, A     | 1296       | 15.5      | 0.10        | 0.30               | 1E+08              | 0.3            | 7.5        | 2.0E-18               | 0.5        | 0.2               |
| Werribee well 20, B     | 1296       | 15.5      | 0.10        | 0.30               | 1E+08              | 0.3            | 3.0        | 1.0E-18               | 0.5        | 0.2               |
| Werribee well 20, C     | 1296       | 15.5      | 0.10        | 0.30               | 1E+09              | 0.3            | 3.0        | 1.0E-18               | 0.5        | 0.2               |
| Werribee well 20, D     | 1296       | 15.5      | 0.10        | 0.30               | 1E+07              | 0.3            | 7.5        | 1.0E-18               | 0.5        | 0.2               |
| Werribee well 20, E     | 1296       | 15.5      | 0.10        | 0.30               | 1E+07              | 1.0            | 3.0        | 1.0E-18               | 0.5        | 0.2               |
| Werribee well 20, F     | 2592       | 15.5      | 0.10        | 0.30               | 1E+07              | 1.0            | 3.0        | 1.0E-18               | 0.5        | 0.2               |
| Werribee well 20, G     | 1296       | 15.5      | 0.20        | 0.30               | 1E+07              | 1.0            | 3.0        | 1.0E-18               | 0.5        | 0.2               |

Additional data input for all scenario’s: \( f_{\text{BCOP}} = 0.5; C_{\text{CELL}} \) (kg/m³) = 220

| Scenario                | Calculated | No limitation | + BDOC limitation |
|-------------------------|------------|---------------|-------------------|
|                         | \( A_{\text{OPEN}} \) m² | \( V_{\text{DER}} \) m³/day | \( N_t \) n/m² | \( D_{\text{BAC,t}} \) mm | \( t_{\text{MAX}} \) day | \( N_{\text{MAX}} \) n/m² | \( D_{\text{BAC,max}} \) mm |
| Werribee well 20, A     | 2.9        | 133          | 6E+16       | 173                 | 4.7        | 1E+14               | 0.29               |
| Werribee well 20, B     | 2.9        | 133          | 2E+12       | 0.003               | 5.0        | 2E+14               | 0.29               |
| Werribee well 20, C     | 2.9        | 133          | 2E+13       | 0.03                | 4.0        | 2E+14               | 0.29               |
| Werribee well 20, D     | 2.9        | 133          | 6E+15       | 8.67                | 6.0        | 2E+14               | 0.29               |
| Werribee well 20, E     | 2.9        | 133          | 1E+18       | 2074                | 17.2       | 2E+14               | 0.29               |
| Werribee well 20, F     | 2.9        | 266          | 3E+18       | 4148                | 17.2       | 4E+14               | 0.58               |
| Werribee well 20, G     | 5.8        | 67           | 7E+17       | 1037                | 17.2       | 1E+14               | 0.14               |
What can we do to lower the risk of biological well clogging? The various scenarios in Table 8 reveal

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Figure 7. Predicted unlimited (-lim) and carbon limited (+BDOC-lim) growth of bacteria (plotted as

log (number of cells)), and their clogging of open pore space on the borehole wall and behind, in terms

of thickness of biofouled layer (D_{BAC}), for scenario A defined in Table 8. Note that right-hand scale

for 100 D-bac has to be divided by 100. Carbon limitation calculated with observed biodegradable
dissolved organic carbon (BDOC) decline during 1–2 days detention in aquifer. t_{MAX} = time since start

of exponential bacterial growth to the maximum level of growth due to carbon limitation.

When we include growth limitation, then much less bacteria are accumulating in a much thinner
biofouled layer: e.g., in case of Werribee scenario A, the calculated total number of bacteria decreases
from $6.1 \times 10^{16}$ to $1.0 \times 10^{14}$ m$^{-2}$ and the thickness of the biofouled layer declines from 173 to 0.29 mm,
in case of BDOC limitation (Table 8).

These predictions are hampered by many imperfections due to among others: (i) inaccuracy
of parameters, especially $\Delta_{BDOC}$; (ii) accumulation of dead bacteria by lack of e.g., oxygen, (iii)
accumulation of suspended particles other than bacteria, (iv) incomplete removal of biofouling
and accumulated fines during backflushing, and (v) erosion by flowing water and predation by
higher organisms.

The presented model calculations thus only serve the purpose of doing a sensitivity or risk analysis.
What can we do to lower the risk of biological well clogging? The various scenarios in Table 8 reveal
that this risk can be lowered by (i) further pretreatment to reduce $N_0$ (requires chlorination, advanced
oxidation or reverse osmosis) and BDOC (requires granular activated carbon filtration or slow sand
filtration), (ii) chlorination to reduce $N_0$ and extend $t_{LAG}$ (this may raise BDOC and the capacity to
oxidize aquifer minerals however), (iii) augmenting the frequency of backflushing (which reduces $t$),
(iv) making wells with a larger $A_{OPEN}$, especially by drilling larger diameter holes, and (v) reducing
the injection rate by augmenting the number of wells.

4. Discussion

Clogging is the enemy of managed aquifer recharge (MAR) operations [41,42], and this holds ‘in extremis’ for injection wells, especially in case of siliclastic aquifers. In Werribee, the results obtained
during the first and the second ASR trial (up to and including cycles 1–3A), the first with and the
second without additional filtration, confirm that well clogging is indeed cumbersome, requiring
frequent backflushing (once per 1–3 days), incidental mechanical well regeneration and intensive
monitoring of hydraulic heads. The ongoing research with the additional filtration steps in place
(Figure 8) shows optimistic preliminary results, to be confirmed when injection phase 3B continues
and more data become available.
In this study, we focused on the 2 main causes of well clogging, i.e., physical clogging by suspended solids in the injectant, and biofouling, after evaluating the risks of 4 other causes of well plugging and the results of column studies by [7] with core material from well 20.

Hydraulic head trends indicate a more or less linear increase of clogging, which pleads for physical clogging as the main process [10]. However, injection runs were relatively short, which prevents biofouling to clearly manifest itself with the typical exponential progress and a typical H₂S smell during backflushing after a long storage period during which biomass will putrefy. Backflushing could be optimized by raising the pumping rate not 2 but 3 times the normal recovery pumping rate, by occasional backflushing several times, and by limiting the backflushing time to the period with a significant turbidity rise so as to reduce the waste water volume (Pyne, written communication). Chlorination has been applied during both pilots, but could also be optimized (see [42]) to reduce biofouling while keeping the formation of disinfection byproducts low.

We developed a simple method to deduce the composition and origin of the suspended solids removed during backflushing, based on readily available methods. A more direct method is desirable, however, and could consist of (i) TSS collection in the injectant from both filtration units (right panel in Figure 8), (ii) on-site filtration of a high volume sample during backflushing, and (iii) analysis of the material retained on the filters. For the injectant a lower minimum detection limit of TSS is needed (to be lowered from <1 to <0.1 mg/L), for the backflush water the additional analysis of loss on ignition of TSS is desired to estimate its organic matter content.

Microscopic analyses proved very useful to demonstrate the ubiquitous presence of diatoms, other algae, fine amorphous debris and bacteria in TSS of both the infiltration water and the backflushed water. Seasonal algae blooms should, therefore, be translated into an injection stop, possibly aided by on line turbidity measurements coupled to intelligent decision support and specific action protocols [43]. Another option is to utilize tangential columns [44] as an early warning system to prevent injection of e.g., an incidental TSS peak.

With the focus on well clogging by suspended particles and biofouling, we calculated these risks by three well clogging predictors: (i) the method of Buik and Willemsen [23] based on the MFI, well and hydrogeological parameters, (ii) a modification of Bichara’s method [26] based on the ratio of the TSS input after a given infiltration time, to the open area of the external borehole wall, and (iii) a modification of the method of Huisman and Olsthoorn [27] which relates biofouling to the exponential growth of bacteria. Application of the first and second method is straightforward. The third is, however, handicapped by scarcity of relevant data on changes in biodegradable organic carbon (e.g.,
during aquifer detention, and difficult to assess processes such as the lag time (period for bacteria to acclimatize) and growth limitation by either biodegradable carbon, nutrients or oxygen.

5. Conclusions

From the 6 main causes of injection well clogging, suspended particles and biofouling are the most common of ASR bore clogging [10,42]. This also holds for the ASR system in Werribee, as demonstrated by: (i) hydraulic data, geochemical and microscopic analysis of total suspended solids in the injectant and backflushed water, and (ii) 3 well clogging predictors, 2 of which were optimized for this study and are fit for general application. Clogging predictors yield an order of magnitude estimate of the clogging rate, but above all they serve the purpose of doing a sensitivity or risk analysis.

Based on these predictors the following recommendations can be given to reduce the risk of well clogging by suspended particles and biofouling: (i) reduce the TSS input by prefiltration (preferably by applying rapid followed by slow sand filtration), (ii) reduce the injection rate (may require additional wells), (iii) augment the backflush frequency and its performance, (iv) chlorinate or optimize the chlorination to reduce N0 and extend tLAG (this may raise BDOC, however; CWW already applies chlorination and occasionally a shock chlorination), (v) reduce BDOC (requires granular activated carbon filtration or slow sand filtration), and (vi) drill larger diameter holes. The strength of the predictors is that they yield insight into the relative effect of measures taken.

The order of recommendations presented above (i–vi) reflects the order of decreasing priority for the Werribee pilot. In case of a new pilot anywhere, we recommend the following general order of descending priority, with numbering as above: vi > ii > i > iii > iv > v. This order is subject to modifications depending on among others the outcome of the three presented clogging predictors.

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Abbreviations

The following abbreviations are used in this manuscript:

- AOC: assimilable organic carbon
- ASR: aquifer storage and recovery
- BDOC: biodegradable dissolved organic carbon
- BGL: below ground level
- BOD5: biological oxygen demand in 5 days at 20 °C
- CWW: City West Water
- D: aquifer thickness
- D50: median grain size diameter of aquifer
- DOC: dissolved organic carbon
- EC: electrical conductivity
- L: length of well screen
- LWF: Lower Werribee Formation
- MAR: managed aquifer recharge
- MFI: 0.45 µm Membrane Filter Index
- Mm³: mega cubic meter
\[ N_t, N_0 \] number of bacteria at time \( t = t \) and \( t = 0 \)

\[ \text{ORP} \] oxidation reduction potential

\[ Q_{IN} \] mean infiltration rate when well is recharging

\[ r_B \] radius of borehole

\[ Q_0, Q_t \] injection rate at time \( t \) since start and at start \( (t = 0) \), respectively

\[ \text{RE} \] recovery efficiency

\[ S \] aquifer storativity

\[ \text{SAR} \] sodium adsorption ratio

\[ t \] time since start (day)

\[ t_{10} \] injection period needed to reduce flow with 10% (day)

\[ T \] aquifer transmissivity

\[ T_D \] doubling time for bacterial population (day)

\[ \text{TDS} \] total dissolved solids

\[ T_{EQ} \] total amount of equivalent full load hours per year

\[ t_{LAG} \] lag time during which bacteria do not reproduce, e.g., due to chlorination (day)

\[ t_{MAX} \] time since start of exponential bacterial growth to maximum growth level (day)

\[ \text{TOC} \] total organic carbon

\[ TSS \] total suspended solids

\[ v_{CLOG} \] clogging rate

\[ v_{DIR} \] entrance velocity on borehole wall

\[ \alpha_L \] longitudinal dispersivity

\[ \varepsilon \] porosity

\[ \Delta BDOC \] consumption of biodegradable DOC

\[ \varphi_0, \varphi_t \] impressed head in injection well without clogging, and with clogging at time \( t = t \)

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