Diabetes and aortic aneurysm: current state of the art

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Abstract

Aortic aneurysm is a life-threatening disease due to the risk of aortic rupture. The only curative treatment available relies on surgical approaches; drug-based therapies are lacking, highlighting an unmet need for clinical practice. Abdominal aortic aneurysm (AAA) is frequently associated with atherosclerosis and cardiovascular risk factors including male sex, age, smoking, hypertension, and dyslipidaemia. Thoracic aortic aneurysm (TAA) is more often linked to genetic disorders of the extracellular matrix and the contractile apparatus but also share similar cardiovascular risk factors. Intriguingly, a large body of evidence points to an inverse association between diabetes and both AAA and TAA. A better understanding of the mechanisms underlying the negative association between diabetes and aortic aneurysm could help the development of innovative diagnostic and therapeutic approaches to tackle the disease. Here, we summarize current knowledge on the relationship between glycaemic parameters, diabetes, and the development of aortic aneurysm. Cellular and molecular pathways that underlie the protective effect of diabetes itself and its treatment are reviewed and discussed, along with their potential implications for clinical translation.

Keyword

Diabetes • Aneurysm • Pathophysiology

Introduction

Aortic aneurysm corresponds to a localized enlargement of the aorta resulting from a weakening of the aortic wall. The disease is often asymptomatic but is associated with high rates of mortality, especially when complications such as aortic rupture occur.1 Despite significant advances in our understanding of the disease mechanisms, the only curative treatment available today relies on endovascular or open surgical repair; drug-based therapies are still lacking.2 The disease may occur both in the thoracic and abdominal parts of the aorta, the most frequent localization being the abdominal infrarenal aorta. Thoracic and abdominal aortas differ in structure, biochemical composition, smooth muscle cell origin and biology, and even if abdominal and thoracic aortas share common mechanisms, they both have distinct features.2 The pathologic processes that lead to abdominal aortic aneurysm (AAA) formation are complex, and involve alteration and loss of vascular smooth muscle cells (VSMCs), in association with inflammatory cell infiltration, extracellular matrix (ECM) remodelling, and intraluminal thrombus (ILT) formation.3,4

Thoracic aortic aneurysms (TAA) frequently develop at younger ages, are often linked to genetic mutations such as Marfan syndrome or Loeys-Dietz syndrome, and are frequently associated with aortic dissection.5 Alterations of ECM and VSMC tone play important roles in TAA, but the inflammatory component and VSMC apoptosis are less prominent than in AAA.2,5,6 Despite these differences, epidemiological studies have paradoxically highlighted an inverse relationship between diabetes mellitus (DM) and both TAA and AAA.5,7 The aim of our review is to summarize the current knowledge on the pathophysiological mechanisms that underlie the inverse association between diabetes and aortic aneurysm.

Diabetes and the epidemiology of aortic aneurysm

Diabetes mellitus corresponds to a heterogeneous disease characterized by a chronic hyperglycaemia and is generally classified in several categories including mainly type 1 (T1D) and type 2 diabetes (T2D).8 Although
DM represents a major cardiovascular risk factor,^8^9 the vast majority of epidemiological studies have paradoxically identified an inverse association between DM and the prevalence and incidence of AAA. Indeed, several reports have shown that diabetic patients develop smaller AAA, as demonstrated by significantly lower aortic diameters compared to non-diabetic subjects. Besides, several studies highlighted lower growth rates of AAA in diabetic patients compared to controls.

In practice, the decision to treat AAA takes into consideration the risk of rupture. As large aortic diameter and high growth rate represent major risk factors of rupture, it is not surprising that a negative association between DM and aneurysm rupture was identified.

While DM appears as a protective factor of AAA formation and expansion, the prognosis and outcome after AAA treatment also differs between diabetics and non-diabetic patients. Heterogeneous results were found among different studies, some reports revealing increased operative mortality in diabetics,^11^21 others showing no difference,^12^22 and some reporting lower mortality. Besides, morbidity following AAA repair was analysed, and DM was identified either as a negative or protective factor of specific post-operative outcomes. Higher rates of complications such as myocardial infarction, infection, or pancreatitis were observed in diabetic patients after AAA open repair,^22^ and higher incidence of device-related complications following endovascular AAA repair were found. On the opposite, DM had a protective effect on AAA growth and re-interventions after endovascular repair,^23^ but no significant difference in the occurrence of neck dilatation or type 1 endoleaks was identified between diabetics and controls.24

Similar to AAA, a negative association was identified between DM and TAA. A meta-analysis combining four studies which included patients with ascending TAA, thoraco-abdominal aneurysms, and non-syndromic TAA confirmed the negative association between diabetes and TAA. However, the impact of diabetes on TAA growth is still unknown and its effect on post-operative outcome has been poorly investigated. It will be important to better explore the impact of diabetes in the different forms of TAA distinguishing ascending and descending TAA, thoraco-abdominal aneurysms, as well as TAA caused by genetic disorders in order to fully understand the link between these disease states.

**Glycaemic parameters and aortic aneurysm**

Glucose homeostasis results from a complex mechanism involving two main hormones secreted by the pancreas, insulin, and glucagon, which have opposite effects.8 The impairment of glucose regulation leads to chronic hyperglycaemia, which characterizes DM. In practice, diabetes is biologically defined as fasting plasma glucose (FPG) >7.0 mmol/L (>126 mg/dL) or as a 2-h plasma glucose value after a 75 g oral glucose tolerance test (OGTT) >11.1 mmol/L (>200 mg/dL) or as a glycated haemoglobin A1c (HbA1c) >6.5%.8

It is noteworthy that in the wide majority of epidemiological studies on AAA, DM was defined as a known history of the disease according to the patient’s declaration and to medical records. However, Hjeltestad et al.17 found that according to OGTT and HbA1c values, half of the patients with AAA and DM were unaware of their DM diagnosis. Hence, the exclusive reliance on history of disease and disease records could lead to an underestimation of DM in the studied cohorts. Moreover, most of the studies did not distinguish between T1D and T2D diabetic patients. Given the distinct characteristics of pathophysiological patterns between these two forms of DM, it is possible that their effects and relationships with AAA differ. Also, the relationship between AAA development and the pre-diabetic state is still poorly understood, underlining the need for a better investigation of the association between AAA and the different forms and stages of DM.

Nevertheless, several studies investigated the relationship between glycaemic parameters and AAA characteristics. An inverse correlation was found between fasting glucose and aortic diameter, suggesting a relationship between glucose metabolism and AAA formation. However, no association was found between serum levels of glucose and aortic expansion in a cohort of 198 patients followed up to 3 years. While fasting blood glucose reflects short-term glucose regulation, HbA1c is representative of the glycaemic status over the past 3 months. In this regard, an inverse association between the growth rate of AAA and the level of HbA1c was found both in individuals with and without DM, suggesting a mechanistic relationship between long-lasting elevated blood glucose levels and AAA progression.

Another study found no association between fasting blood glucose measured at mean age 33 and aortic diameter measured at age 65 years. Moreover, Taimour et al. revealed that AAA prevalence did not differ between 65-year-old men who were newly diagnosed as T2D patients and non-diabetic men, and HbA1c level did not correlate with AAA diameter in newly diagnosed T2D patients. These results suggest that mechanisms linking altered glucose metabolism and a decreased risk of AAA are not relevant until late in life or until they are sustained long after the diagnosis of DM.

While short- and long-term glucose levels have been investigated in the context of AAA, the relationships between AAA characteristics and parameters of glucose regulation, such as insulin or glucagon, have so far received little attention. One study did not identify any association between fasting insulin and AAA, whereas another reported a positive correlation between C-peptide and AAA diameter in T2D patients. C-peptide is a biomarker used in medical practice to assess endogenous insulin secretion and is also useful to indirectly evaluate insulin resistance. Nevertheless, the significance and direct role of C-peptide in AAA pathogenesis are still poorly understood. Moreover, the link between glycaemic parameters and TAA has not been investigated so far.

**Animal models of diabetes and aortic aneurysm**

Several experimental models have been developed to understand the impact of DM on aneurysm. Chemically induced AAA models are schematically categorized in non-dissecting and dissecting models and reproduce the main human pathophysiological features even if they present some limits. Non-dissecting AAA models can be created through perfusion or application of porcine pancreatic elastase or calcium chloride. These models reproduce some of the main features of human AAA, including VSMC loss, inflammatory cell infiltration, ECM degradation, and aortic dilatation. However, they are self-contained and do not progress to rupture. Dissecting AAA models are characterized by intramural dehiscence leading to haematoma formation and aortic dilatation and are often associated with dilatation and dissection of the thoracic aorta.

They can be induced by infusion of angiotensin II, with or without inhibition of transforming growth factor (TGF)-β or lysyl-oxidase, or through administration of a mineralocorticoid-receptor agonist and high salt. Animal models of genetically triggered TAA (Marfan or Loeys-Dietz syndromes) have also been developed through mutation or deletion of the incriminated gene, such as fibrillin-1 or TGF-β ligands and receptors.
In parallel, several animal models of DM have been developed, allowing to study the impact of the disease on AAA development. T1D can be induced by administration of streptozotocin or alloxan, which affects pancreatic beta cell survival and leads to chronic hyperglycaemia. Streptozotocin-induced T1D had a protective effect on elastase-induced AAA development, as demonstrated by decreased aortic diameter and decreased AAA growth. Similar results were obtained in calcium phosphate-induced aneurysm model and after angiotensin II perfusion in ApoE−/− mice.

Models of T2D tend to reproduce human features of insulin resistance and/or beta cell failure and can be associated with obesity. Obese animal models of T2D include monogenic models such as mice deficient in leptin signalling pathway or polygenic models. Among them, KK-Ay mice develop severe hyperinsulinaemia and insulin resistance associated with impairment of pancreatic islet function. Non-obese models of T2D, such as Goto-Kakizaki (GK) rats, are also available allowing the study of disease mechanisms in a lean colony. Similarly to humans, T2D exerts a protective effect on AAA development, as demonstrated by decreased aortic diameter and decreased AAA growth. Similar results were obtained in calcium phosphate-induced aneurysm model and after angiotensin II perfusion in ApoE−/− mice.

Table 1 Experimental models used to study aneurysm development in diabetic mice

| Diabetes model       | Aneurysm model                      | Main results                                                                 | References |
|----------------------|-------------------------------------|-------------------------------------------------------------------------------|------------|
| T1D: streptozotocin administration | CaPO4-induced aneurysm in the carotid artery of C57BL/6 mice | Decreased AAA formation                                                       | Tanaka et al.36 |
|                      | Elastase infusion in the abdominal aorta of C57BL/6 mice | Decreased AAA formation                                                       | Dua et al.34 |
|                      | Decreased macrophage infiltration  | Decreased AAA formation                                                       | Miyama et al.35 |
|                      | Decreased macrophage infiltration associated with decreased PAI-1 aortic gene expression | Decreased macrophage infiltration associated with a decrease of mural neovasculartiy and MMP9 activity and a better elastin preservation | Miyama et al.35 |
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| T2D: KK-Ay mice      | CaPO4-induced aneurysm in the carotid artery | Decreased aneurysm formation                                                  | Tanaka et al.36 |

Mechanistic effects of diabetes on aortic aneurysm

ECM remodelling

Several studies addressed the impact of DM on vascular wall structure (Figure 1). In the absence of difference in aortic lumen diameter, aortic wall stress of the abdominal aorta was shown to be 20% lower in T1D patients compared to controls, with 22% larger aortic intima-media thickness, which significantly correlated with the duration of DM. A study integrated protein–protein interaction and genetic interaction to examine biological pathways related to DM, aneurysm, and atherosclerosis. The study identified a set of 16 proteins with high brokerage values (i.e. essential for the interconnectedness of the network), and this network was enriched in proteins involved in cell matrix-adhesion, supporting a role for ECM alterations in the link between DM and reduced risk of aneurysm. ECM is a dynamic network composed of basement membrane, collagen, elastin, and proteoglycans that contribute to tissue organization and structure. Interestingly, proteome analysis of non-atherosclerotic arteries (free of adventitia) from T2D patients showed increased amounts of the basement membrane and ECM components including alpha1-type IV and alpha2-type IV collagen, gamma1, and beta2 laminin. These results suggest distinct properties of ECM in diabetic patients and could partly be involved in the protective effect on AAA development. In addition to the fibrous proteins, the background of the ECM is composed of glycosaminoglycans (GAG), which are complex polysaccharides covalently linked to ECM proteins in the form of proteoglycans. GAGs are categorized into four classes according to their disaccharide units, including chondroitin sulfate/dermatan sulfate, heparan sulfate, keratin sulfate, and hyaluronan. Biglycan, a member of chondroitin sulfate/dermatan sulfate, is abundant in normal human aorta and its expression is decreased in abdominal aortic aneurysmal tissue. Moreover, biglycan gene defects cause an X-linked syndromic form of severe TAA and dissection. To better understand the role of biglycan in aneurysmal pathology, experimental models were used. Biglycan deficiency in Ldlr−/− mice aggravated angiotensin II-induced aneurysm, with increased rates of vascular mortality and rupture, and induced the development of descending TAA. This was associated with breaks in medial elastin as well as a decrease in dense collagen fibre network in the aortic wall. In addition, biglycan deficiency in BALB/cA mice induced...
mortality due to spontaneous aortic dissection and rupture. Transmission electron microscopy and biomechanical testing revealed abnormalities of collagen fibrils with marked variations in size and shape associated with reduced tensile strength. Other studies pointed to interactions between biglycan and cytokines, particularly TGF-β, which plays a central role in both TAA and AAA. Biglycan can regulate TGF-β signalling pathway and TGF-β can enhance biglycan expression, suggesting a mutual and positive feedback interaction to preserve the ECM and protect the arterial wall against aneurysm development and dissection. Although the direct impact of diabetes on biglycan expression in the aneurysmal aortic wall is still uncertain, diabetes is associated with an upregulation of Cell Division Autoantigen 1 (CDA1) which enhances TGF-β signalling pathway and contributes to the protective effect of diabetes on AAA formation. DM can also alter the production, degradation and deposition of other GAGs in the aorta, with additional consequences on ECM remodelling as well as the structural and physical properties of the arterial wall. Decorin, a small leucin-rich repeat proteoglycan, is increased in response to high-glucose concentration, interacts with TGF-β to regulate matrix organization and collagen matrices and may reduce aneurysm formation in part through modulation of ECM remodelling. On the other hand, substantial accumulation of (foci of) GAGs, mainly hyaluronan, aggrecan and versican, is frequently associated with the progression and severity of TAA. These GAG foci generate important osmotic pressures with localized increases in intramural stress, and are thought to initiate medial delamination. We speculate that an imbalance between GAGs and the connective tissue in the aorta of diabetic patients in favour of increased collagen deposition, may limit the susceptibility of the aorta to intralamellar Donnan swelling pressures caused by focal accumulation of GAGs, thereby limiting the risk of aneurysmal dilatation and dissection, particularly in the thoracic aorta.

ECM remodelling also results from a balance between the synthesis of ECM components and their degradation by proteases including cathepsins, metalloproteinases (MMPs), and serine proteases. In some studies, the protective effect of DM on experimental AAA was associated with a decrease of MMP9 expression and activity, and a relative elastin preservation compared to controls. This is corroborated by the observation of decreased MMP9 secretion by activated monocytes exposed to aortic media extracted from diabetic patients compared to non-diabetics. To further address the mechanisms involved, in vitro studies were performed using murine macrophage cell line RAW264.7 treated...
with tumour necrosis factor alpha (TNF alpha) and calcium phosphate (CaPO₄) to mimic conditions of CaPO₄-induced AAA formation. Hyperglycaemic conditions (incubation with glucose 15.5 mmol/L) attenuated TNF/CaPO₄-induced elevation of MMP9 at the gene, protein and activity levels, in a Nr1h2-dependent mechanism. However, others reported an induction of MMP expression and activity, including MMP9 in human monocyte-derived macrophages, and MMP1/MMP2 in human endothelial cells, under higher glucose concentrations (25 mmol/L). MMP-2 and MMP-9 serum concentrations were lower in T2D patients compared to non-diabetics and hyperglycaemia during OGTT induced a decrease in MMP9. However, HaB1C paradoxically correlated with MMP9 in T2D patients. Hence, the effect of hyperglycaemia on MMP regulation is complex and would require further studies to fully understand it. It is also important to note the conflicting evidence regarding the role of MMP collagenases in aneurysm formation, and more particularly the increased susceptibility to angiotensin II-induced dissecting AAA in hypercholesteroleamic mice with MMP9 deficiency or genetically engineered resistance to collagenases, some of these effects being related to altered biomechanical properties of the aortic wall due to altered collagen deposition and fibre orientation. In addition to MMPs, serine proteases such as plasmin represent one of the major proteolytic factors related to altered biomechanical properties of the aortic wall due to altered collagen deposition and fibre orientation. In addition to MMPs, serine proteases such as plasmin represent one of the major proteolytic enzymes involved in ECM remodelling. Plasmin activity results from the activation of plasminogen by plasminogen activators (PA), and this pathway is inhibited by plasminogen activator inhibitor-1 (PAI-1). Interestingly, AAA development was decreased in diabetic mice compared to controls, concomitantly with an increase of plasma PAI-1 level and aortic PAI-1 gene expression, and a decrease of plasmin generation. However, the mechanisms behind PAI-1 modulation in diabetic mice, and the direct link with reduced AAA development have not been explored.

**Glycation and advanced glycation end products**

DM and chronic hyperglycaemia lead to an increase of advanced glycation end products (AGEs), which result from non-enzymatic glycoxidation of proteins and peptides after contact with aldose sugar. These products include non-cross-linking AGEs such as carboxymethyllysine (CML) and carboxyethyllysine (CEL), and cross-linking AGEs such as pentosidine. Interestingly, concentrations of pentosidine, CML and CEL were in general lower in AAA biopsies compared to non-aneurysmal aortic tissues. When comparing diabetic patients to non-diabetics, an increase of pentosidine concentration was observed in AAA aortic samples and its concentration showed a tendency toward a negative correlation with AAA diameter, suggesting a protective effect of cross-linking AGEs. Besides, a significant decrease of carboxyterminal telopeptide of collagen type 1, and a tendency toward a decrease of collagen type 1C-telopeptide was observed in glycated AAA tissues compared to non-glycated AAA tissues. As those collagen telopeptides are products from the degradation of mature type 1 collagen by MMPs and cysteine proteases, and no difference was found in protease activity (including MMP2, MMP9, cathepsin A, B, and S) between AAA biopsies of diabetics compared to non-diabetics, the results suggest that glycation of AAA tissue protects the collagen network and increases its resistance to proteolysis. The precise effects of glycation and cross-linking on ECM remodelling were further investigated by in vitro studies on THP1 cell lines and peripheral blood mononuclear cells (PBMCs). Interestingly, glycated bovine serum albumin (BSA) and glycated monomer collagen enhanced the secretion of MMP9 by THP1 cells. In contrast, glycated collagen lattices and cross-linked collagen lattices reduced the secretion of MMP2 and MMP9 by both THP1 cells and PBMCs. These results underline the complexity of the effect of glycation on monocytes and ECM remodelling, which may differ depending on the type of glycated protein.

In addition to direct modification of protein structure, AGEs can lead to the activation of several signalling pathways through various cell surface receptors. The most known of those receptors is the multi-ligand receptor for advanced glycation end products (RAGE), which has been identified in various cells including monocytes/macrophages, endothelial, and VSMCs. Experimental studies unravelled the complex role of AGEs and their receptor RAGE in angiotensin II-induced AAA model using ApoE⁻/⁻/RAGE⁻/⁻/mice. RAGE deficiency decreased the incidence of AAA, and this was associated with a decrease of MMP9 activity and cellularity of the adventitia. Further *in vitro* studies revealed that stimulation with RAGE ligands increased MMP9 gene expression as well as MMP9 activity in murine macrophage cell line RAW 264.7. Another study using transgenic mice overexpressing the human S100A12 (a ligand of RAGE known to be increased in DM) reported progressive dilatation of the aorta with elastin fibre disruption, fibrosis, and loss of VSMCs within the medial layer, along with an increase of MMP2 protein expression. Besides, analysis of human samples revealed that S100A12 protein was expressed in all cases of thoracic aortic type A dissection, in 25% of stable TAA, but was not detected in non-aneurysmal control aortic tissue, consistent with a potential pathogenic role in human aortic disease. Interestingly, investigation of the association between RAGE gene polymorphisms and AAA revealed that the 825 allele of RAGE was a risk factor for AAA. Taken together, these results suggest that the impact of glycation on protein structure could partly account for the protective effect of DM on AAA formation (*Figure 1*), whereas its effect through RAGE signalling pathway may exert a pathogenic effect on the aortic wall and contribute to AAA development.

As glycation affects AAA development, investigators examined if measurement of AGEs could be of interest as a biomarker in AAA. Interestingly, skin accumulation of AGEs measured by auto fluorescence was found to be higher in patients with AAA compared to controls. However, this association was lost when adjusted for cardiovascular risk factors. No association was found between AAA progression and serum CML concentration. Besides, no correlation was found between pentosidine concentration and AAA diameter, and there was no correlation between plasma levels of pentosidine and its levels in AAA biopsies. It is reasonable to hypothesize that AGEs are formed in other tissues, making the systemic measurement less representative of the local effect in the AAA wall. This field of research is still in its infancy, and new clinical studies are currently investigating the types and locations of AGEs in aortic tissues and in several biological fluids of patients with AAA, and will probably provide a better understanding of the complex role of glycation in this disease.

**Inflammation**

Over the past decades, several studies revealed a close link between metabolism and immunity; alterations of the immune system contribute to the pathogenesis of DM and conversely, metabolic alterations leading to DM modulate the inflammatory response. The enhanced state of immune activation in DM, whether chronic autoimmunity in T1D or chronic low-grade inflammation in T2D with enhanced production of inflammatory mediators, is not expected to protect against the development and/or progression of aortic aneurysm. Most of the studies point to a promoting role of DM on vascular inflammation through several mechanisms, including increased oxidative and endoplasmic reticulum stress responses, and impaired activation of protective anti-inflammatory
pathways (reduced endothelial nitric oxide synthase, insulin resistance in vascular cells, etc.).\textsuperscript{71} It should be kept in mind, however, that this is not a black-and-white situation, and strong counter-regulatory mechanisms (e.g., TGF-\(\beta\)) may still operate and may even be promoted, systemically and locally, under diabetic conditions. Monocyte recruitment to the endothelium is finely regulated by glycaemic conditions. A study in Goto-Kakizaki rats (GK-rats), a genetic non-obese model of T2D, revealed that fluctuation of blood glucose enhanced monocyte adhesion to the thoracic aortic endothelium, an effect possibly due to increased gene expression of connecting segment (CS)-1 and very-late acting antigen (VLA)-4.\textsuperscript{72} On the other hand, administration of insulin and nateglinide (known to stimulate insulin secretion) in GK-rats reduced post-prandial hyperglycaemia, thus attenuating blood glucose fluctuation, and induced a decrease of monocyte adhesion to endothelial cells.\textsuperscript{73} The results suggest a promoting effect of DM on vascular inflammation. However, diabetes alters the differentiation of haematopoietic stem cells towards monocytes and macrophages, in part through upregulation of DNM1 and hypermethylation of major transcription regulators PU.1, NOTCH1, and KLF4,\textsuperscript{74} which may limit the inflammatory process. Furthermore, increased monocyte recruitment may not favour AAA development unless monocytes differentiate into pro-inflammatory macrophages. However, DM or hyperglycaemic conditions have been shown to modulate macrophage phenotype towards either a pro-\textsuperscript{74–76} or anti-inflammatory state,\textsuperscript{77} through various mechanisms including upregulation of acyl-CoA synthetase 1\textsuperscript{75} or regulation of AKT/mTOR and ERK pathways.\textsuperscript{27} The pro-inflammatory phenotype appears to be prevalent after short-term diabetes and in association with a hypercholesterolaemic setting,\textsuperscript{75} whereas the anti-inflammatory phenotype was observed after several months of diabetes induction.\textsuperscript{77} Thus, increased monocyte recruitment into the injured artery wall in DM may not lead invariably to increased accumulation of pro-inflammatory macrophages. Furthermore, glycation has been shown to alter monocyte/macrophage function toward an anti-inflammatory phenotype, reducing interleukin (IL)-6 production as shown in lipopolysaccharide-activated THP1 monocytes incubated with glycated collagen lattices and cross-linked collagen lattices.\textsuperscript{77} As IL-6 deficiency limits both TAA and AAA development in mice, and Mendelian randomization approaches supported the involvement of the IL6 receptor pathway in human AAA,\textsuperscript{78} the effect of glycation on IL-6 may contribute to the protective effect of DM in aortic aneurysm.

Studies in animal models of AAA revealed a decrease of macrophage infiltration in diabetic mice compared to controls.\textsuperscript{14–16} In contrast, a higher macrophage infiltration was observed in AAA wall of diabetic compared to non-diabetic patients who underwent AAA surgical repair and was correlated with serum glucose concentration.\textsuperscript{77} AAA diameter was similar between diabetics and non-diabetics, and no significant difference was observed for T-lymphocyte infiltration. The discrepancy between those human and animal studies underlines the complex impact of diabetes on inflammation and suggests that the protective effect of DM on AAA development and progression may not be linked to its impact on inflammatory pathways (Figure 1). Another plausible explanation is that a higher level of inflammation is required to overcome the resistance of the aorta of diabetic patients to aneurysm development.

**Aortic mural neoangiogenesis**

Aortic mural neoangiogenesis is involved in TAA and AAA development, and in the progression to aortic rupture.\textsuperscript{80} Interestingly, a decrease of mural neovascularity was observed in the aneurysmal wall of diabetic mice compared to controls.\textsuperscript{85} Studies in animal models revealed that DM inhibits neoangiogenesis in aortic tissue, impairs ischaemia-induced neovascularization,\textsuperscript{81} and impairs vascular endothelial growth factor pathway induction by hypoxia-inducible factor 1.\textsuperscript{82} Similar mechanisms may be involved during AAA. Angiogenesis inhibitors limit AAA progression in a dissecting aneurysm II model, possibly through reduced immune cell infiltration\textsuperscript{84} and reduced interstitial oedema,\textsuperscript{55} suggesting that the protective effect of DM on AAA and TAA could partly be attributed to a decrease of neovascularization (Figure 1).

**VSMC homeostasis**

VSMCs have the ability to switch from a contractile phenotype to a secretory phenotype and impact on vascular homeostasis in physiological and pathological states.\textsuperscript{84} Several studies addressed the impact of DM on VSMCs.\textsuperscript{84} First, the morphology of saphenous vein VSMCs collected from T2D patients differ from non-diabetics, with a rhomboid morphology suggesting a dedifferentiated phenotype.\textsuperscript{85} Focal adhesions characterized by vinculin immunostaining were increased in VSMCs from diabetic patients, with a disparate organization of the alpha actin network. Histologic analysis of tissues from patients with AAA suggested decreased staining for alpha-smooth muscle actin in diabetic patients compared with non-diabetics.\textsuperscript{86} The significance of this finding is unclear and may reflect either reduced VSMC number or increased VSMC dedifferentiation. Although some studies reported less proliferation in VSMCs from T2D compared to non-diabetics,\textsuperscript{85} most of the studies are in favour of a promoting effect of DM on VSMC proliferation through activation of ERK signalling pathways and the secretion of mitogenic factors.\textsuperscript{86,87} Note that VSMCs were harvested from the saphenous vein and from the internal mammary arteries, respectively, which may explain, at least in part, the discrepancy between the studies. Experimental studies in animal models revealed persistent phenotypic changes and altered inflammatory gene expression in aortic VSMCs cultured from diabetic mice.\textsuperscript{88} High glucose and palmitic acid induce epigenetic changes in aortic VSMCs that promote VSMC ‘dysfunction’. For example, miR-504 is upregulated in VSMC under those conditions, leading to inhibition of contractile gene expression, while increasing migration, proliferation, and inflammatory gene expression, through effects on Grb10 and Egr2.\textsuperscript{88} The study fell short however of establishing a direct link between diabetic conditions, epigenetic alterations, and VSMC dedifferentiation. Moreover, the term VSMC ‘dysfunction’ is too vague and may be misleading. For example, inflammatory changes in VSMCs under diabetic conditions may be expected to be conducive for aortic aneurysm development and progression. In contrast, acquisition of migratory and proliferative properties by VSMCs may have a protective healing and stabilizing role. Phenotypic switching of VSMC is controlled by the transcription factor KLF4\textsuperscript{90} and is an important feature of aortic aneurysms.\textsuperscript{91} Selective deletion of KLF4 in VSMCs reduces the development and severity of aortic aneurysm in both dissecting and non-dissecting mouse models of the disease.\textsuperscript{91} Intriguingly, hyperglycaemia reduces KLF4 expression in VSMCs and arteries of mice and patients with diabetes, in part through Foxoa2-dependent upregulation of miR-29c.\textsuperscript{92} The results are consistent with the upregulation of miR-143/145 (a major regulator of KLF4 expression\textsuperscript{93}) in VSMCs (from saphenous vein) of patients with diabetes, and suggest a potential protective role of diabetes against AAA and TAA through limitation of KLF4 induction. Along this line of thinking, increased activation of the TGF-\(\beta\) signalling pathway under diabetic conditions, both systemically and locally in aortic VSMCs,\textsuperscript{94,95} may also play an important protective role. TGF-\(\beta\) is required for the induction and maintenance of VSMC differentiation and homeostasis. Genetic mutations in the TGF-\(\beta\) signalling pathway predispose to TAA in humans.\textsuperscript{96}
and selective deletion of TGF-β signalling pathway in VSMCs promotes TAA development and dissection in mice.\textsuperscript{97–99} Beyond a direct effect on VSMC differentiation, TGF-β signalling in VSMCs appears to limit their production of IL-1β,\textsuperscript{100} which further contributes to the protection against aneurysm formation. Another pathway involved in the protective effect of TGF-β may be related to the regulation of VSMC contractility.\textsuperscript{101} Genetic mutations in genes affecting VSMC contractility predispose to TAA in humans,\textsuperscript{102} and alteration of VSMCs contractility has been suggested to contribute to aortic dissection in mice with disrupted TGF-β signalling.\textsuperscript{103} In this regard, it is interesting to note that the reduction of KLF4 expression in VSMCs of diabetic mice and diabetic patients was associated with increased expression of contractile proteins.\textsuperscript{92}

Others have also reported increased expression of contractile genes and proteins in VSMCs under elevated glucose levels, related in part to activation of protein kinase C, Rho/Rho-kinase and actin polymerization.\textsuperscript{104} Given the contribution of VSMC contractility to microvascular tone and ECM contraction, and the proposed roles of the latter physiological pathways in aortic wall homeostasis and its resistance to aneurysm formation and dissection,\textsuperscript{54} those changes in VSMCs induced by, or promoted under diabetic conditions may further account for the protection against aortic aneurysm.

Endoplasmic reticulum (ER) stress induction in aortic VSMCs is thought to promote cell death and aneurysm,\textsuperscript{105} a pathway that might be counter-regulated by induction of autophagy.\textsuperscript{106} High-glucose conditions may favour the induction of ER stress in aortic VSMCs.\textsuperscript{107} However, DM also highly promotes AGE-dependent VSMC proliferation through induction of autophagy,\textsuperscript{108} which may overcome its deleterious effect on ER stress.

Endothelial to mesenchymal transition, which corresponds to the process by which endothelial cells are able to switch to a myofibroblastic phenotype in response to injury, is impaired in TAA.\textsuperscript{109} As diabetes is associated with enhanced endothelial to mesenchymal transition in the vascular tissue, this process could potentially be involved in the inverse association between diabetes and aneurysm. Finally, a recent study unravelled a detrimental role of nitric oxide (as a result of NOS2 activation) in promoting medial aortic degeneration and dilatation in a mouse model of Marfan syndrome.\textsuperscript{110} The mechanisms may involve an impact of nitric oxide on VSMC contractility and MMP9-mediated elastin fragmentation. It is therefore tempting to hypothesize that reduced nitric oxide bioavailability in diabetic setting may limit this pathogenic process. In general, the impact of DM on VSMC phenotype during aneurysm has not been thoroughly investigated so far (Figure 1), and the available studies did not specifically distinguish between VSMCs derived from the thoracic and the abdominal aortic region. This is of major importance given the distinct embryonic origins of VSMCs in AAA and TAA, and the distinct pathological features of aortic aneurysm in those different locations.

**Intraluminal thrombus formation**

ILT formation is a factor contributing to AAA development. The wall underlying the thrombus is thinner and exhibits increased signs of degrada-tion. The rate of thrombus growth also correlates with aneurysm growth and rupture.\textsuperscript{111} Interestingly, glucose concentration is associated with altered clot structure, with a higher clot density and decreased porosity reported in T2D patients compared to controls.\textsuperscript{112} This may confer increased clot resistance to fibrinolysis in diabetic patients. As ILT is a source of protease release and activation, it can be hypothesized that a more compact but less active clot could limit AAA expansion in diabetic patients (Figure 1). In TAA, thrombus may develop within the aortic wall after dissection but it is not considered as an important factor in the initiation of dissection.

**Effects of antidiabetic treatments**

Treatment of DM relies on lifestyle measures associated with blood glucose lowering drugs.\textsuperscript{5} Insulin administration is the main therapy for T1D patients. Pharmacological treatments mostly used in T2D include insulin sensitizers such as biguanides (metformin) or thiazolidinediones (rosiglitazone, pioglitazone), drugs stimulating insulin secretion such as sulfonylureas or meglitinides, and drugs with incretin effects such as glucagon-like-peptide-1 (GLP-1) receptor agonists (i.e. liraglutide, exenatide, lixisenatide) or dipeptidyl peptidase 4 (DPP-IV) inhibitors (i.e. alogliptin).

Intriguingly, epidemiological studies have revealed that mechanisms conferring a protective effect of DM on AAA are not only related to the pathophysiology of DM but also to antidiabetic treatments. A study including 1269 patients with AAA showed that the use of antidiabetic drugs was associated with a 56% reduction in AAA growth rate, and this association was independent of confounding factors including other therapeutic agents.\textsuperscript{113} In addition, a nested case–control analysis including 4468 patients with AAA and 4468 matched controls revealed that metformin, sulfonylurea, and thiazolidinedione use was associated with a lower risk of developing aneurysm.\textsuperscript{114} The negative association between metformin use and AAA enlargement and growth was confirmed in other studies.\textsuperscript{115,116} No significant association was found between metformin use and the risk of rupture.\textsuperscript{117}

Experimental studies were performed to better understand the cellular and molecular pathways underlying the negative association between antidiabetic use and AAA development. Studies using the elastase-induced AAA model in normoglycaemic mice suggest a protective role of metformin, with preservation of VSMCs and aortic medial elastin, associated with decreased inflammatory cell infiltration.\textsuperscript{115} (Figure 2).

However, lower levels of ECM components or basement membrane proteins, including alpha-chains of type IV collagen, collagen type XVIII, laminin-gamma-1, laminin-beta-2, and nidogen-1 were observed in arteries from T2D patients treated by metformin compared to non-metformin users.\textsuperscript{39} Taken together, these results point to a complex effect of metformin on the pathophysiological events of AAA development and progression.

Several experimental studies addressed the impact of thiazolidinediones on AAA development (Figure 2). This drug class improves insulin sensitivity through activation of the nuclear transcription factor peroxi-some proliferator-activated receptor (PPAR)-gamma and plays important roles in regulating inflammation by antagonizing several inflammatory transcription factors AP-1, STAT, and NF-kappaB in various immune cells.\textsuperscript{118,119} Although treatment with thiazolidinediones has been associated with adverse hepatic, cardiovascular, osteogenic, and carcinogenic effects, their use in T2D is still debated.\textsuperscript{119} Treatment with pioglitazone as well as treatment with PPAR-alpha agonists (fenofibrate) reduced aortic diameter in the angiotensin II/ApoE\textsuperscript{−/−} model, and this effect was associated with reduced osteopontin (OPN) staining in the aorta and reduced macrophage infiltration.\textsuperscript{120} As OPN is a chemoattractant factor for macrophages and its deficiency improves AAA,\textsuperscript{121} the protective effect of pioglitazone could be partly due to downregulation of OPN. Treatment with rosiglitazone also reduced AAA formation and abolished death caused by rupture in the angiotensin II/ApoE\textsuperscript{−/−} model.\textsuperscript{122,123} This selective agonist of PPAR-gamma decreased angioten-sin II-induced expression of E-selectin, TNF-α and IL-6 in the aorta,
which may in part explain the protective effect given the known roles of those pro-inflammatory mediators in experimental AAA. In addition, rosiglitazone inhibited angiogenesis II-induced JNK activation, modulated the inflammatory process by downregulating TLR4, and suppressed the production of TLR4/JNK dependent chemokines Monocyte Chemoattractant Protein-1 and MIP1 alpha, which may further explain its protective effect on AAA development. At last, as rosiglitazone reduced serum levels of MMP9 in T2D patients, its protective effect on AAA development could potentially be linked with ECM preservation.

The action of drugs with incretin effect including GLP-1 receptor agonists (i.e. lixisenatide, liraglutide) and DPP-IV inhibitors (i.e. alogliptin, sitagliptin, MKEY0620) has also been investigated. Subcutaneous administration of GLP-1 receptor agonist lixisenatide reduced CaCl2-induced AAA development through antioxidant, anti-inflammatory, and ECM preservation effects, as revealed by decreased reactive oxygen species (ROS), reduced macrophage infiltration and inhibition of TNF, MMP2, and MMP9 gene expression in the aortic wall. These effects were associated with a decrease of ERK expression in the AAA wall suggesting the involvement of this signaling pathway in the protective effect of GLP-1 analogues. The protective effect of this drug was confirmed in another study where administration of lixisenatide reduced AAA formation in Apoe<sup>−/−</sup> mice infused with angiogenesis II. The treatment increased circulating active GLP-1 and was associated with a preservation of elastin content, a decrease of macrophage infiltration, and a reduction of MMP2 and MMP9 expression. In vivo studies on monocytic U937 cells further confirmed the effect of the GLP-1 analogue, where pre-treatment with lixisenatide decreased angiogenesis II-induced ROS formation, MMP2 and MMP9 activities, and abolished angiogenesis II-induced monocyte recruitment in a concentration dependent manner.

Other studies investigated the effect of DPP-IV inhibitors (alogliptin, sitagliptin, MKEY0620) and revealed the protective effect of this class of drugs on AAA development. DPP-IV gene expression was increased in the aneurysmal aortic wall, and immunostaining for DPP-IV (also known as CD26) showed it was expressed in macrophages. Besides, plasma DPP-IV activity was increased, and plasma active GLP-1 concentration decreased in Apoe<sup>−/−</sup> mice after angiotensin II infusion compared to controls. As expected, the administration of DPP-IV inhibitors decreased DPP-IV activity and increased plasma active GLP-1 concentration. Intriguingly, DPP-IV inhibitors did not affect blood glucose concentration and plasma insulin levels, suggesting that the protective effect of DPP-IV inhibitors against AAA development may not be related to an incretin effect. DPP-IV inhibitors induced a dose dependent increase of elastin content, associated with a decrease in ROS production, and a reduction of the expression and gelatinolytic activities of MMP2 and MMP9 in the aneurysmal aortic wall. In another study, DPP-IV inhibitors reduced the incidence of AAA and the protective effect was associated with reduced aortic expression of IL1β and increased tissue inhibitor of metalloproteinase 2 (TIMP2). Additional treatment with an incretin receptor antagonist did not reverse the protective effect of DPP-IV inhibitors, confirming that DPP-IV inhibitors limit AAA formation in an incretin independent manner. In addition to GLP-1, DPP-IV has various substrates, at least in vitro, including gastrointestinal hormones, neuropeptides, cytokines (IL-1β and IL-2), and chemokines (CCL5), which could potentially account for the protective effect of DPP-IV inhibitors.

While oral antidiabetic drugs were associated with a protective effect in experimental AAA, insulin therapy had an intriguing opposite effect, negating the protective effect of hyperglycaemia (induced by streptozotocin injection) on elastase-induced AAA formation.

Figure 2 Hypothesized protective effects of anti-diabetic treatments in AAA. Anti-diabetic treatments may contribute to the negative association observed between diabetes and AAA through protective effects on inflammation, neangiogenesis, extracellular matrix remodelling, and oxidative stress.
To conclude, the mechanism involved in the protective effect of anti-diabetic drugs may not be directly related to an effect on blood glucose or insulin secretion. It may rather be due to improvement of insulin resistance, and modulation of other pathways, including inflammation, oxidative stress, and ECM remodelling (Figure 2). The impact of anti-diabetic treatments on TAA development and progression is still unknown.

Conclusion and perspectives

Epidemiologic and experimental studies demonstrated a negative association between DM and aortic aneurysm formation and expansion. DM contributes to vascular tissue remodelling and has profound impact on several pathways relevant to aneurysm development, including ECM remodelling, inflammation, VSMC homeostasis, aortic mural neoangiogenesis, and ILT formation. Despite sharing a few common mechanisms, AAA and TAA display distinct features, and the main mechanisms underlying the protective effect of DM may slightly differ between AAA and TAA (Figure 1). The protective effect of diabetes on AAA may involve the role of glycation and cross-linking on ECM remodelling, the anti-inflammatory properties of TGF-β signalling pathway, and the impact on ILT formation, neoangiogenesis, and VSMC apoptosis. In addition to DM itself, anti-diabetic drugs interfere with the pathophysiological mechanisms of aneurysm and may contribute partly to the protective effect observed in diabetic patients. In TAA, the protective effect of diabetes may involve the balance between collagen and GAG contents in the media and the resulting consequences on intramural stress distribution, the regulation of neoangiogenesis, the prevention of the deleterious effect of nitric oxide on aortic degeneration through reduced nitric oxide availability, and the promotion of protective TGF-β signalling in VSMCs, VSMC proliferation and endothelial to mesenchymal transition. Note, however, that the effect of anti-diabetic drugs on TAA formation is still unknown, and merits appropriate investigation.

In the limelight of current knowledge, future directions can be proposed. First, DM is a complex disease classified in different types whose pathophysiological mechanisms differ. Future clinical studies should distinguish T1D from T2D patients to better understand the link of each type of DM with aortic aneurysm formation and progression. In parallel, the vast majority of experimental studies were performed in animal models mimicking T1D. More studies are needed in T2D animal models, with detailed investigation of the cellular and molecular mechanisms at play. Besides, the impact of intermediate states such as prediabetes is still unknown and remains to be elucidated. While the effect of hyperglycaemia itself has been investigated, the impact of major hormones involved in glucose homeostasis such as insulin and glucagon is poorly understood. Moreover, the molecular and cellular effects of DM in modulating the distinct pathophysiological mechanisms of AAA and TAA require further investigation. Given the availability of experimental models of DM and aortic aneurysm, and given the potential perspective to identify and develop new therapeutic tools based on modulation of glucose metabolism, the challenge is undoubtedly worth it.

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