Monoamine Oxidase Inhibition by Plant-Derived β-Carbolines; Implications for the Psychopharmacology of Tobacco and Ayahuasca

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The monoamine oxidases (MAOs) are flavin-containing amine oxidoreductases responsible for metabolism of many biogenic amine molecules in the brain and peripheral tissues. Whereas serotonin is the preferred substrate of MAO-A, phenylethylamine is metabolized by MAO-B, and dopamine and tyramine are nearly ambivalent with respect to the two isozymes. β-Carboline alkaloids such as harmine, harmine, and norharmane are MAO inhibitors present in many plant materials, including foodstuffs, medicinal plants, and intoxicants, notably in tobacco (Nicotiana spp.) and in Banisteriopsis caapi, a vine used in the Amazonian ayahuasca brew. The β-carbolines present in B. caapi may have effects on neurogenesis and intrinsic antidepressant properties, in addition to potentiating the bioavailability of the hallucinogen N,N-dimethyltryptamine (DMT), which is often present in admixture plants of ayahuasca such as Psychotria viridis. Tobacco also contains physiologically relevant concentrations of β-carbolines, which potentially contribute to its psychopharmacology. However, in both cases, the threshold of MAO inhibition sufficient to interact with biogenic amine neurotransmission remains to be established. An important class of antidepressant medications provoke a complete and irreversible inhibition of MAO-A/B, and such complete inhibition is almost unattainable with reversible and competitive inhibitors such as β-carbolines. However, the preclinical and clinical observations with synthetic MAO inhibitors present a background for obtaining a better understanding of the polypharmacologies of tobacco and ayahuasca. Furthermore, MAO inhibitors of diverse structures are present in a wide variety of medicinal plants, but their pharmacological relevance in many instances remains to be established.

Keywords: tobacco, Nicotiana, ayahuasca, Banisteriopsis caapi, monoamine oxidase (MAO), β-carbolines, harmine, dimethyltryptamine (DMT)
INTRODUCTION

The monoamine oxidases (MAOs EC 1.4.3.4) are flavin-containing amine oxidoreductases that occur in the outer mitochondrial membrane of most mammalian cells. MAOs deaminate their endogenous substrates in the presence of molecular oxygen; this generates the corresponding aldehyde-intermediates, which further metabolize to carboxylic acids or alcohol derivatives. The deamination step releases the amine nitrogen as ammonia and with the production of hydrogen peroxide. As such, MAO is a double-edged sword, ridding the organism of excess biogenic monoamine compounds, while also producing potentially toxic metabolites. MAO occurs in MAO-A and MAO-B isozymes with slightly divergent amino acid sequences, and having similar genetic organization with 15 exons on the X-chromosome (Shih and Chen, 2004). Apparently having arisen by a gene duplication event early in the mammalian lineage, the two forms have differing substrate specificities. Whereas serotonin is the preferred substrate of MAO-A, phenylethylamine is the preferred substrate for MAO-B; dopamine and tyramine are nearly ambivalent with respect to the two forms of MAO (Schoepf and Azzaro, 1981; Yang and Neff, 1973). These metabolic activities point to a key role of MAO in maintaining homeostasis of biogenic monoamine neurotransmitters, but by no means does MAO expression occur only in the central nervous system (CNS); MAO-A is present in the liver, pulmonary vasculature, the gastrointestinal tract (more about this later) and the placenta, whereas MAO-B is present in blood platelets. As shall be seen below, heterocyclic β-carbolines (9H-pyrido[3,4-b]indole) alkaloids such as harmine, harmame, and norharmane1 are MAO inhibitors present in many plant materials, including tobacco and its smoke, and in the decoctions known as ayahuasca or yagé used in Indigenous Amazonian medical systems and practices. In the context of the psychopharmacology of ayahuasca, it is widely believed that β-carbolines may potentiate the psychoactive potency of N,N-dimethyltryptamine (DMT); although from a traditional Amazonian perspective this may not be the only essential mechanism of action, and ayahuasca brew samples need not invariably contain DMT (McKenna 1984a; Callaway et al., 2005; Ona et al., 2020; Politi et al., 2021), we present for reference purposes the chemical structures of DMT and key naturally occurring β-carbolines (Figure 1).

THE CLASSES OF MAO INHIBITORS

MAO inhibitors fall into various categories with respect to the nature of the binding to the enzyme. Certain molecules are MAO substrates that can exert competitive inhibition of the enzyme by virtue of their occupation of the binding pocket, and other compounds, like the β-carbolines, are reversibly binding competitive inhibitors that are not substrates for MAO. Still other compounds, like pargyline, clorgyline, and rasagline, are irreversible MAO inhibitors, also known as suicide substrates. This class of MAO inhibitor is substrates that mechanistically form a covalent bond with the enzyme via the propargyl moiety; this reaction permanently kills the enzyme. To this day, there is a prevailing notion that pharmacologically blocking the breakdown of dopamine, serotonin, and other MAO substrates should underlie the antidepressant property of MAO inhibitors by rectifying some neurochemical deficiency, despite the paucity of evidence for any such deficiency syndrome in patients with depression (Gryglewski, 2014). Be that as it may, irreversible MAO inhibitors were among the first effective antidepressant medications (Quitkin et al., 1979), including the non-selective (MAO-A/B) inhibitors phenelzine (Nardil), isocarboxazid (Thase et al., 1995), and tranylcypromine (Ulrich et al., 2017). While effective against depression and anxiety symptoms, the irreversible MAO inhibitors have fallen out of favor due to the risk of toxic interactions arising from complete blockade of MAO. Certain foods and beverages contain high levels of pharmacologically active substances such as tyramine that normally undergoes inactivation by MAO-A in the gut. Consuming certain cheeses rich in tyramine can thus provoke a dangerous hypertensive crisis in people treated with irreversible MAO inhibitors, known as the “cheese effect” (Youdim and Weinstock, 2004). While individuals free of MAO inhibition can consume tyramine by the spoonful without ill effect, patients treated with tranylcypromine are at risk for a hypertensive crisis after consuming as little as 10 mg of tyramine (Gillman, 2001; Ulrich et al., 2017). This is unlikely to occur with use of irreversible MAO-B inhibitors such as rasagline (an antiparkinsonian compound), or with reversible MAO-A inhibitors such as moclobemide, an antidepressant that has largely supplanted irreversible inhibitors in clinical practice, despite arguments for their greater antidepressant efficacy (Shulman et al., 2013). In addition to the perhaps overstated cardiovascular risks, MAO inhibition has effects on the chemistry of the CNS. For example, treatment with pargyline increased the vesicular concentrations of dopamine and noradrenaline in rodent brain (Buu and Lussier, 1989). Acute treatment with pargyline enormously increases the brain concentrations of the trace amines phenylethylamine and tryptamine (Durden and Philips, 1980) as well as tyramine (Suzuki et al., 1979) and other MAO substrates that are normally present at low concentrations in brain. There is a diverse family of trace amine-associated receptors (TAAR), which can exert a variety of actions in the brain and peripheral tissues (Gainetdinov et al., 2018). It follows that treatment of patients with MAO inhibitors should exert actions extending beyond effects on the far more abundant neurotransmitters such as dopamine, noradrenaline, and serotonin.

THE BEHAVIORAL EFFECTS AND NEUROCHEMISTRY OF MAO INHIBITION

While MAO inhibition should increase the levels of a wide variety of endogenous and xenobiotic substances, the threshold for

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1Harmame/norharmane are often also written as harman/norharman; we follow the former convention in this manuscript.
physiologically relevant MAO inhibition is not well established. In particular, is MAO inhibition from tobacco smoke merely an epiphenomenon, or does it contribute to the psychopharmacology of tobacco use, its experienced effects, and/or dependence? Some work indicates that the norharmane level in plasma of habitual smokers was inversely related to tobacco craving in smokers with a relatively low dependence on tobacco (Van Den Eijnden et al., 2003). This suggests that certain β-carbolines may have intrinsic effects on drug-seeking behavior, unrelated to any direct interaction with (for example) nicotine. Mice treated with the irreversible MAO inhibitor tranylcypromine showed an increased liking for nicotine, based on their self-administration behavior (Villégier, 2007). Similar studies in rats showed that clorgyline or norharmane (5 mg/kg/day) potentiated the reinforcing effects of nicotine, whereas the MAO-B selective inhibitor L-deprenyl had no such effect (Guillem, 2006). Yet another study confirmed the potentiation of nicotine self-administration in rats pretreated with tranylcypromine, but failed to show a comparable effect from a cocktail of tobacco alkaloids including harmine and norharmane, perhaps due to insufficient dose (Smith, 2015). On the other hand, treatment of rats with three β-carbolines (harmine, norharmane, and harmine) evoked dose-dependent increases in the threshold for intracranial self-stimulation (ICSS) (Harris et al., 2020). In this experimental paradigm, trained rats press a lever in order to get a (rewarding) electrical stimulation of mesolimbic dopamine release. The increasing ICSS thresholds suggest that β-carbolines in isolation exerted aversive/anhedonic effects. Conditioned place preference (CPP) is another paradigm to measure “drug-liking”; if an animal finds a drug treatment rewarding, they will tend to return to the very spot where the drug was first administered, which might be described as magical thinking, or simple pragmatism. Pretreatment of rats with clorgyline at a low dose (1 mg/kg) did have some interaction with the CPP induced by the powerful psychostimulant methamphetamine (Kitanaka, 2006), and a still lower clorgyline dose (0.1 mg/kg) more clearly interfered with the methamphetamine CPP, while selectively decreasing the concentration in brain of the noradrenaline metabolite 3-methoxy-4-hydroxyphenylglycol (MHPG) (Kitanaka, 2010). Thus, partial MAO inhibition in brain may be relevant to the reinforcing and/or aversive properties of drugs such as nicotine or methamphetamine.

Genetic ablation of MAO gives insight into the perturbations produced by pharmacological MAO inhibition. Thus, knockout of MAO-A in mice results in elevated brain levels of dopamine, serotonin, and noradrenaline (Shih and Chen, 1999), along with a behavioral phenotype including aggression, perseveration, and social behavior deficits, which have been likened to autism spectrum disorder (Bortolato, 2013), and which were partially rescued by treatment with serotonin reuptake inhibitors (Godar, 2014). Perhaps similarly, males in a Dutch kindred with a loss of function mutation in the MAO-A gene showed a pattern of intellectual disability as well as abnormal behavior including aggression, arson, and various other forms of criminality (Brunner, 1993). However, mice with ablation of the MAO-B gene do not show the aggressive behavior typical of MAO-A knockouts, but have the expected increases in tissue levels of phenylethylamine (Shih and Chen, 1999). Two individuals with Norrie disease involving deletion of the MAO sequences on the X-chromosome showed markedly reduced formation of the noradrenaline metabolite MHPG and enormously elevated urine levels of the MAO-B substrates phenylethylamine and tyramine (Murphy, 1990). In some sense, these developmental abnormalities may mirror the consequences of pharmacological MAO inhibition, with the caveat that genetic ablation must result in a divergent pathway of brain development.

How might MAO inhibition alter cerebral function by increasing levels of neurotransmitters? The cerebral microdialysis technique enables the detection of dynamic changes in interstitial neurotransmitter levels after drug treatments and across behavioral states. In one such study, treatment with a high dose
of pargyline non-significantly increased the interstitial level of dopamine in rat striatum by about 50% (Cumming, 1992), while enormously increasing the levels of the default metabolite 3-methoxytyramine (Brown, 1991). A subsequent study seemed to show that pargyline-evoked increases in dialysates from rat striatum were likely an artefact of the procedure for implanting the dialysis probes (Blaha et al., 1996). We may suppose that dopamine fibers autoregulate their rate of dopamine synthesis and release following MAO inhibition, despite the net increased tissue concentration of dopamine. While dopamine is ambivalent with respect to the form of MAO in vitro, other microdialysis studies involving clorgyline or L-deprenyl treatment indicate that MAO-A normally metabolizes dopamine in mouse striatum, but that MAO-B also contributes under conditions of elevated dopamine levels (Fornai, 1999). This may not hold for the rat, in which brain dopamine metabolism is apparently exclusively mediated by MAO-A; the case is less clear for humans, in whom the expression of MAO-B increases with age (Shih et al., 1999). Various microdialysis studies showed that interstitial serotonin levels were two-fold higher in MAO-A knockout mice compared to wild-type mice, and that other perturbations included down-regulation of serotonin transporters and loss of autoregulation of serotonin release. These adaptive phenomena might underlie the antidepressant effects of irreversible MAO blockers, but the incomplete inhibition obtained with plant-derived MAO inhibitors may not suffice to produce chronic adaptive changes in dopamine and serotonin pathways. On the other hand, the presence of 2,3,6-trimethyl-1,4-naphthoquinone and β-carbolines in tobacco smoke may be responsible for the protection against Parkinson’s disease afforded by smoking. A follow-up study of 30,000 male British doctors lasting 65 years indicated a 30–50% lower incidence of Parkinson’s disease among the smokers (Mappin-Kasirer, 2020). As in the case of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced neurotoxic parkinsonism, uninhibited MAO may generate a toxic metabolite or excess amounts of hydrogen peroxide, thus promoting the senescence of vulnerable dopamine neurons (Castagnoli and Murugesan, 2004).

NATURAL SOURCES OF MAO INHIBITORS; THE CASE OF TOBACCO

Substances acting as MAO inhibitors occur in a wide variety of lichen, fungi, and plants. For example, 5-hydroxy-2-methylchroman-4-one from a lichen fungus (Daldinia sissa) inhibits MAO-A/B at μM concentrations (Jeong, 2021), as do a variety of plant-derived and synthetic coumarin derivatives (Koyiparambath, 2021). The pigment derived from madder root (purpurin; 1,2,4-trihydroxyanthraquinone) is another MAO inhibitor with reputed antidepressant effects in a rodent model (Ma, 2020). Even licorice (Glycyrrhiza glabra) extracts are significantly potent inhibitors of MAO-B in vitro (Ramadan, 2021), perhaps due to the sweet tasting saponin compound glycyrrhizin. St. John’s Wort (Hypericum perforatum) has long served as an antidepressant in traditional medicine, an effect often attributed to MAO-inhibition by one or more of its constituents. Floral extracts indeed inhibited MAO-A in vitro (IC₅₀ 65 μg/ml), which proved to be due to quercetin and flavonoids rather than hypericin (Herraz and Guillén, 2018), much as shown in an earlier study (Bladt and Wagner, 1994); the more recent study indicated 1000-fold higher inhibition of MAO-A by Peganum harmala extracts, which contain high levels of harmaline and harmine. MAO-inhibitory activity in a plant extract need not suffice to evoke significant MAO inhibition upon consumption. However, concentrations of the β-carbolines harmaline and norharmane in sesame seeds or their oil may partially inhibit MAO after consumption (Liu, 2021).

Recent years has seen burgeoning interest in the pharmacological and therapeutic effects of β-carbolines, which are present at significant concentrations in extracts of P. harmala, a rue native to the Mediterranean region (Moloudizargari, 2013), and in the South American vine Banisteriopsis caapi. The latter plant is the key constituent of the ayahuasca decoction widely used in Indigenous Amazonian social and medical practice. In that context, MAO-inhibition among other things serves to potentiate central action of the hallucinogenic component NN-dimethyltryptamine (DMT), which is a hallucinogenic serotonin 5-HT₂A agonist otherwise subject to rapid deamination in the gut following oral administration (Cumming, 2021), as attested by a series of self-experiments (Ott, 1999). DMT also forms endogenously in the living brain and might thus properly be termed a trace amine neurotransmitter comparable to tyramine, phenylethylamine, and tryptamine. The latter trace amines normally occur at very low concentrations in brain, being vulnerable to rapid metabolism by MAO; it is currently unknown if MAO inhibition increases the brain concentration of endogenous DMT to a relevant extent. Certainly, irreversible MAO inhibitors are not hallucinogenic, but their administration might still increase brain levels of DMT to a degree evoking some physiological effects or interactions. However, DMT is naturally present in a wide variety of plant sources; notably the south American plant Psychotria viridis (Carbonaro and Gatch, 2016), and likewise reportedly in Diplopterys cabrerana (McKenna 1984) and Mimosa tenuiflora (Simão, 2019). While orally administered DMT is essentially inactive, conjoint consumption of the ayahuasca decoction’s constituents (e.g., B. caapi and P. viridis) (Simão, 2019), evokes an intense psychedelic and visionary experience. However, these two plants have highly variable alkaloid contents; 32 B. caapi samples collected in Brazil had β-carboline concentrations ranging from 0.3 to 8.4 mg/g dry weight for harmine and 0.03–8.3 mg/g dry weight for harmaline (Callaway et al., 2005). In the same study 36 samples of P. viridis had DMT contents ranging from nil to 18 mg/g dry weight. Other constituents of ayahuasca rituals may sometimes include the potent tobacco species Nicotiana rustica, which contains unknown levels of β-carbolines and other alkaloids. Hallucinogenic preparations of myristaceous bark and leaf samples contained DMT and its congener 5-methoxy,N,N-dimethyltryptamine (5-MeO-DMT), but only traces of β-carbolines (McKenna, 1984b), an observation which may call for qualifying the necessity of conjoint administration of DMT along with an MAO inhibitor.

According to Paracelsus, dosis sola facit venenum (the dose makes the poison). In our considerations of MAO interactions, we must be mindful that MAO inhibitors, despite their ubiquity in plants, may not be sufficiently active to have pronounced effects on biogenic amine metabolism in the living organism. For example, sensitive
analyses reveal the presence of harmame and norharmame at low concentrations (<50 ng/g) in a wide variety of foodstuffs, and moderately high levels in coffee beverages (200 ng/ml). The first attestation of the presence of harmame and norharmame in tobacco, and its apparent pyrolytic formation from tryptophan in the tobacco, dates to 1962 (Poindexter and Carpenter, 1962). Janiger and de Rios likewise noted 40–100-fold higher harmame and norharmame levels in tobacco smoke than in the uncombusted tobacco and raised the possibility of inherent psychoactive effects of these compounds (Janiger and de Rios, 1973). More recent reports indicate impressively high concentrations of these alkaloids in cigarette smoke (up to 3,000 ng/cigarette) (Herraiz, 2004). In keeping with the notion of pyrogenic formation of β-carbolines, others reported concentrations of around 0.5 ng/g harmane and harmine in the leaves and flowers of Nicotiana tabacum (Tarkowská, 2020). Interestingly, the authors of that study also reported relatively high concentrations (10 ng/g) of the alpha2 antagonist yohimbine, which has a pressor and anxiogenic action; administration of yohimbine increased the reinforcing efficiency of nicotine in an intravenous self-administration paradigm, but only in female rats (Li, 2014).

As noted above, mainstream smoke from commercial tobacco contains roughly 2 µg harmame and 4 µg norharmame, but smoke from barley tobacco can have several-fold higher concentrations (Zhang et al., 2011). The authors of that study also reported on the presence in smoke of “exotic” heterocyclic compounds derived from tryptophan, namely 3-amino-1,4-dimethyl-5H-pyrido[4,3-b] indole and 3-amino-1-methyl-5H-pyrido[4,3-b]indole. Tobacco smoke particulate has very high nicotine concentrations, greatly exceeding the amounts of harmame and norharmame, which seemed insufficient to inhibit completely MAO-A/B in smokers (Truman et al., 2017); the authors postulated that unidentified substances might account better for the MAO-inhibitory effect of smoking. Nonetheless, β-carbolines such as norharmame are clearly present in blood of tobacco smokers, attaining a concentration of 200 pg/ml after smoking a single cigarette (Breyer-Pfaff, 1996). Further work demonstrated an association with smoking, but no such effect of alcohol consumption by non-smokers (Spijkerman, 2002). Norharmame and harmame are present in tobacco smoke; whereas norharmame inhibits MAO-A and MAO-B with low µM Ki, harmame inhibits MAO-B with much greater potency, having a Ki of 55 nM (Herraiz and Chaparro, 2005). Surprisingly, flavored e-liquids favored by some tobacco smokers as a less harmful nicotine delivery system proved to have significant MAO-A/B inhibitory action, which was attributed to the presence of vanillin (4-hydroxy-3-methoxybenzaldehyde) (Truman, 2019). Indeed, vanillin inhibits MAO-A (IC50 20 µM) and MAO-B (IC50 45 µM), as does eugenol (2-methoxy-4-(prop-2-en-1-yl)phenol), the main odorant of clove oil (Dhimam et al., 2019).

As suggested above, components of tobacco smoke other than β-carbolines may contribute to the net inhibition of MAO in brain of smokers. For example, analysis of cured tobacco leaf revealed the presence of the non-selective MAO inhibitor 2,3,6-trimethyl-1,4-naphthoquinone and the selective MAO-B inhibitor farnesylacetone (Khall et al., 2006); in vitro enzyme competition assays indicated Ki values close to 1 µM, which may or may not be physiologically relevant. A Philip Morris study confirmed the presence of 2,3,6-trimethyl-1,4-naphthoquinone in tobacco or smoke, along with other potential MAO inhibitors such as anabasine, 2-naphthylamine, and farnesylacetone (van der Toorn, 2019).

**PET IMAGING OF MAO IN VIVO**

Molecular imaging by positron emission tomography (PET) affords a window on the living brain, whereby the uptake and binding of selective ligands reveals the molecular targets. We can properly understand PET as a binding assay in vivo, where the requirement for external detection by the tomograph calls for integration of certain positron-emitting radionuclides into the tracer molecule of interest. In practice, this is most commonly accomplished using cyclotron-generated fluorine-18 (physical half-life 109 min) or carbon-11 (physical half-life 20 min). The radioactive decay with emission of a positron in the medium of the brain is followed swiftly by annihilation of the positron upon meeting its counterpart electron; the entire energy bound up in the mass of the electron-positron pair (E = mc2) then converts into two photons of enormously high energy, approximately 512 keV. These photons, commonly known as gamma rays, fly apart at an angle of just about 180°. If such an emission occurs when an individual’s head is within the aperture of the PET instrument, this counts as “an event”. The accumulating recording of millions of such events over time gives rise to a dynamic PET image, analysis of which maps out the uptake and distribution of binding sites for the tracer molecule. Arguably, MAO-PET was one of the earliest successful uses of this molecular imaging technology to yield new information about human physiology (Fowler, 1987; Fowler, 1988; Kumlien, 1995; Bergström et al., 1997).

In fact, the very first PET study with an MAO ligand employed the neurotoxin MPTP labelled with carbon-11 in the 1-position, i.e., [11C]MPTP (Moerlein, 1986). Like MPTP, this radiotracer undergoes oxidative deamination by MAO-B present in astrocytes and some specific neuronal populations, yielding the product [11C]-MPP+. That charged metabolite remains inside living cells, thus imparting an increasing PET signal proportional to the local MAO-B activity, as evidenced by the effect of pretreatment with the irreversible MAO inhibitor tranylcypromine, which blocked the specific PET signal in brain of non-human primates. Others confirmed the MAO-B specificity of [11C]MPTP in non-human primate brain by blocking its retention through pretreatment with the MAOA/B suicide substrate pargylne, whereas brain trapping was unaffected by pretreatment with the MAO-A-prefering substrate storglypine (Hartvig, 1986). Other studies in non-human primates with MPTP-induced parkinsonism showed no change in the intensity of [11C]-deprenyl uptake in the dopamine depleted striatum, thus indicating that the enzymatic conversion does not take place in the dopamine fibers per se (Leenders, 1988). Indeed, it emerged that the toxic MPTP metabolite MPP+ is formed elsewhere (perhaps in astrocytes), whereas it gains access to the vulnerable dopamine neurons via the dopamine uptake site and accumulates to toxic levels via the ionic gradient of living neurons; blockers of dopamine uptake protect against MPTP-induced parkinsonism. Unfortunately, doses of MPTP as low as 1 mg/kg cause an irreversible syndrome of parkinsonism, due to the toxic degeneration of brain dopamine...
neurons. While the microgram doses of $[^{11}C] \text{MPTP}$ used in PET studies might be safe, there was a general feeling that better tracers should be developed for human use. Indeed, there soon followed a report on the uptake in mouse and human brain of the MAO-B ligand $[^{11}C] \text{N,N-dimethylphenylethylamine}$ (Shinotoh, 1987). Its uptake in mouse brain was blocked by L-deprenyl but not clorgyline pretreatment, and the PET tracer showed abundant accumulation in human brain, especially in the thalamus; the mechanism underlying its trapping in living brain seems to entail enzymatic conversion, as was the case with $[^{11}C] \text{MPTP}$.

Initial PET studies in human volunteers showed abundant cerebral uptake of $L-[^{11}C] \text{deprenyl}$, an MAO-B-prefering suicide substrate, which was blocked by pretreatment with non-radioactive L-deprenyl (Fowler, 1987). In serial pig PET studies, MAO-B was blocked by treatment with L-deprenyl and the time course of the subsequent return of $L-[^{11}C] \text{deprenyl}$ binding in brain was monitored (Oredland, 1990). This approach indicated a half-life for the turnover of MAO-B in living brain to be about 1 week, which was one of the first such estimates for a specific brain protein. Other pharmacological studies measured the uptake of $L-[^{11}C] \text{deprenyl}$ in brain of healthy volunteers, in conjunction with various doses of the then novel reversible MAO-B inhibitor, Ro 19–6,327 (Bench, 1991; Lammertsma, 1991). Kinetic analyses of the scans with various doses indicated an ID$_{50}$ of 0.3 mg/kg, i.e. the dose of Ro 19–6,327 sufficient to block MAO-B activity in brain by 50%. This competitive inhibition application has since emerged as an important tool in medicinal chemistry for establishing appropriate dose regimens, for example with respect to the necessary frequency of dosing with Ro 19–6,327 required to maintain a constant blockade of MAO-B (Fowler, 1993). The quantitation of $L-[^{11}C] \text{deprenyl}$ uptake in brain is a bit problematic, as it binds so rapidly and irreversibly to brain MAO-B that its uptake is partially limited by cerebral perfusion rate; the greater the blood flow, the greater the PET signal. Aiming to resolve this, the pioneering researchers at Brookhaven National Laboratory and the Karolinska Institute modified $L-[^{11}C] \text{deprenyl}$ by the addition of a deuteron (heavy hydrogen) in the molecule (Fowler, 1988). Since breaking the carbon-deuteron bond by MAO-B is disfavored thermodynamically, the rate of irreversible trapping in brain was slowed substantially compared to ordinary $L-[^{11}C] \text{deprenyl}$, which allowed the physiological separation of MAO-B binding from cerebral blood flow effects (Fowler, 1995).

Alternatives exist, for example the highly specific reversible MAO-B inhibitor 5-[(4-[(benzylxoy)phenyl]-3-(2-cyanoethyl)-1,3,4-oxadiazol-5-[13]C]-2 (3 H)-one, which has nM affinity in vitro (Bernard, 1996). PET studies in nonhuman primate indicated a good signal for the detection of MAO-B in brain.

Preliminary work showed the promising reversible MAO-A ligand $[^{11}C] \text{brofaromine}$ to be largely unsuitable for PET studies due to insufficient specific binding (Ametamey, 1996). The irreversible ligand $[^{11}C] \text{clorgyline}$ binds to MAO-A in living brain, albeit with a non-MAO-A signal present in the cerebral white matter that may impede quantitation (Fowler, 2001). Nonetheless, the PET study with the suicide-substrate $[^{11}C] \text{clorgyline}$ PET revealed a substantial inhibition of MAO-A in brain and peripheral organs of habitual smokers (Fowler, 1996). That remarkable observation was the first evidence that MAO-inhibition might be an aspect of the psychopharmacology of tobacco addiction. The finding was confirmed in a completely independent PET study with the alternate MAO-A ligand $[^{11}C] \text{bexofaxetine}$ (Leroy, 2009). That study used the arguably superior endpoint of binding potential (BP$_{PD}$), showing a mean 60% reduction of MAO-A activity in cerebral cortex of tobacco smokers, versus a 40% reduction in subcortical regions. Deuterium-substituted $[^{11}C] \text{deprenyl}$ PET revealed a similar reduction of MAO-B availability in brain of smokers (Fowler, 1998). Further to the question of $H. \text{perforatum}$, a [11C]harmine PET study did not indicate any MAO-A inhibition in brain of volunteers consuming the product at the dose recommended for depression (Sacher, 2011). This may present an example of the Paracelsus dictum; the presence of MAO-inhibitory substances in extracts of $H. \text{perforatum}$ may not suffice to inhibit the enzyme in vivo, unless consumed in enormous quantities.

Smoking one or two cigarettes increased plasma levels not just of the MAO-B inhibitor norharmarone, but also increased levels of the MAO-A inhibitor harmarnone to 20–30 pg/ml (0.1 nM), then declining with a half-life of about 1 h (Romnellsapcher, 2002). This low concentration may explain why smoking a single cigarette was without discernible effect on the cerebral binding of deuterated $L-[^{11}C] \text{deprenyl}$ (Fowler, 1999); the harmarnone dose is simply too low to evoke much in the way of MAO-B inhibition in brain. Furthermore, there is a pool of endogenous norharmarone in human blood, which may be derived from metabolism of plasma tryptophan (Fekkes, 2001); it remains to be established if this endogenous pool has any physiological relevance. The broader action of MAO is indicated by its ubiquity in various organs other than the brain; the aforementioned deuterium isotope effect can be used to confirm specificity of $[^{11}C] \text{deprenyl}$ binding for MAO-B in various organs of the human body, with a rank order of abundance kidneys $\geq$ heart $>$ lungs = spleen (Fowler, 2002). This ubiquity makes it difficult to assign a singular role of MAO in physiology.

Early work in vitro indicated the $\beta$-caroline $[^{11}C] \text{harmine}$ to have good properties for detection of MAO-A sites in tissue specimens (Goller, 1995). PET studies in pigs showed a very abundant distribution of $[^{11}C] \text{harmine}$ binding throughout brain, which was entirely blocked by pretreatment with pargylne (Jensen, 2006). That experiment aimed to test a specific hypothesis that blockade of MAO-A would potentiate the effect of amphetamine on dopamine release in brain. Insofar as MAO-A activity contributes to the control of brain dopamine concentrations, and given that amphetamine acts by releasing the intracellular dopamine pool and flushing the synapse, we expected that pargylne pretreatment would potentiate the amphetamine-induced displacement of the dopamine D2 receptor ligand $[^{11}C] \text{raclopride}$ by dopamine. There was no evidence for any such potentiation according to the PET competition paradigm in anesthetized pigs. Very similarly, pretreatment of rats with pargylne did nothing to enhance the displacement of striatal $[^{11}C] \text{raclopride}$ binding evoked by amphetamine (Pedersen, 2007). This may stand in contrast to clinical investigations as cited above that MAO inhibition is a salient aspect of tobacco addiction (van Amsterdam, 2006). Similarly, a $[^{11}C] \text{raclopride}$ PET study in depressed smokers versus healthy controls did not indicate any main effect of depression,
although tobacco addiction in depressed individuals tended to decrease the amphetamine-evoked dopamine release (Busto, 2009). That result, in accord with the pig and rat PET studies described above, does not support a strong role for the modulation of amphetamine-induced dopamine release by MAO inhibition from tobacco smoke. Others used \(^{18}\text{F}\)altanserin PET to measure availability of serotonin 5-HT\(_{2A}\) receptors in brain of depressed individuals; treatment with clomipramine, a re-uptake inhibitor likely to increase interstitial serotonin levels, did reduce \(^{18}\text{F}\)altanserin binding, but there was no comparison made with healthy controls. Therefore, that study cannot support any claims about altered serotonin release as a biomarker of depression, or indeed any interaction with MAO-inhibition due to smoking (Larisch, 2003).

Further \(^{11}\text{C}\)harmine PET studies in humans indicated that acute serotonin depletion decreased MAO-A binding, whereas pharmacological activation of dopamine synthesis had the opposite effect of increasing MAO-A availability (Sacher, 2012). The authors interpreted these results to indicate a dynamic regulation of MAO-A activity serving to accommodate homeostasis, but did not provide a compelling mechanism to account for that phenomenon. Electroconvulsive therapy for depression, which causes massive serotonin and dopamine release in brain, had only a slight effect in reducing \(^{11}\text{C}\)harmine binding (Baldinger-Melich, 2019), which might argue against homeostatic control of MAO-A activity in brain. On the other hand, remission from major depression in patients treated with SSRI medications was not associated with any decline in the elevated \(^{11}\text{C}\)harmine binding, suggesting persistence of a trait that might tend to deplete cerebral levels of serotonin and other substances.

\(^{18}\text{F}\)Fluoroethylharmol was developed as an alternative to \(^{11}\text{C}\)harmine, which presents logistic difficulties due to its very brief physical half-life (Maschauer, 2015). Using a synthesis of findings in vivo and in vitro, \(^{18}\text{F}\)fluoroethylharmol binding results in rat brain suggested a dopamine concentration of 0.4 \(\mu\text{M}\) in the striatal compartment containing MAO-A, which seems insufficient to reduce the ligand binding by competition. This might support the implication of the previously described finding (Sacher, 2012) that endogenous MAO substrates do not have directly competitive effects on the activity of MAO in living brain. On the other hand, sevoflurane anesthesia, which is not a direct inhibitor of MAO, decreased deuterated \(L-\text{[}^{1}\text{C}\]deprenyl uptake in non-human primate brain by 80%, which is certainly consistent with some novel regulation of MAO-B availability (Varnäs, 2021). Hormones may also play a role here; testosterone treatment for gender dysphoria slightly decreased (~10%) the \(^{11}\text{C}\)harmine binding in human brain (Kranz, 2021).

Returning to the theme of \(^{11}\text{C}\)harmine PET, studies in nonsmoking patients with major depression indicated a mean 34% greater distribution volume in brain compared to non-depressed controls; this increase was present throughout cerebral cortex, hippocampus, and in subcortical structures (Meyer, 2006). Examination of the scatter plots in Figure 2 of that paper indicated a large effect size (Cohen’s d > 1). While an impressive finding for biological psychiatry, the increased MAO-A availability was not pathognomonic of depression, as there was still considerable overlap in \(^{11}\text{C}\)harmine uptake between the two groups, even in the regions of highest binding, i.e., thalamus and cingulate cortex. One might presume that increased MAO-A levels in depressed individuals would predict decreased brain levels of its preferred substrate, i.e., serotonin. This scenario might match with the widely held consideration that major depression is in some sense a serotonin deficiency syndrome, as suggested by the supposed antidepressant efficacy of SSRIs. Furthermore, decreased tonic serotonin levels might alter phasic serotonergic signaling, and could be a factor the meta-analytic finding of slightly (10%) decreased levels of serotonin transporters in brain of depressed individuals compared to age-matched controls (Gryglewski, 2014).

THE PHYSIOLOGICAL RELEVANCE OF MAO INHIBITION; THE CASE OF AYAHUASCA

We have focused in the preceding section on the molecular imaging of MAO, and its possible relation to neurotransmission and behavior. The physiological relevance of partial MAO-inhibition is a key consideration. Therapy with MAO inhibitors usually aims to obtain nearly complete inhibition of MAO in brain (and consequently in peripheral organs). How important, then, is the transient and partial MAO inhibition seen in the context of tobacco smoking? More dramatically, harmine or other \(\beta\)-carboline alkaloids potentiate the hallucinogenic action of DMT in the context of ayahuasca, presumably by a peripheral action of blocking DMT metabolism in the gut. However, central MAO inhibition might be an independent aspect of the psychopharmacology of harmine in ayahuasca mixtures. Unfortunately, there is scant information about the pharmacokinetics of harmine. An early study followed the disposition of harmine and its metabolites following intravenous injection in humans (0.5 mg/kg) and rats (5 mg/kg) (Slotkin et al., 1970). Much as seen in PET studies with \(^{11}\text{C}\)harmine at tracer doses (Jensen, 2006), there was a rapid decline in harmine concentration in human blood (Slotkin et al., 1970). The harmine level in human plasma declined to about 4 \(\mu\text{g}/\text{ml}\) (10 \(\mu\text{M}\)) within minutes after injection, remaining around this level for about 1 h, but falling to about 0.8 \(\mu\text{g}/\text{ml}\) (2 \(\mu\text{M}\)) at 4 h after intravenous injection. Given the affinity of harmine for MAO-A in vitro (circa 1 nM), we can predict substantial MAO-A inhibition throughout the body and lasting several hours after a dose of 0.5 mg/kg harmine.

While \(B.\ caapi\) is an essential component of ayahuasca, the importance of DMT and other alkaloids in \(P.\ viridis\) is less clear. In an ayahuasca study, a group of 15 male volunteers (mean body weight 75 kg) consumed 150 ml of a decoction containing 30 mg harmaline, 250 mg harmine and 159 mg tetrahydroharmine, as well as 35 mg DMT (Callaway, 1999). Pharmacokinetic analysis indicated peak plasma levels of 90 ng/ml (500 nM) for harmine at 30 min post-ingestion, which declined with a half-life of 116 min. Tetrahydroharmine attained a peak plasma level of 80 ng/ml (400 nM) at 180 min post ingestion and declining with a half-life of 530 min. The peak plasma level of DMT was 12 ng/ml (60 nM) at 120 min post ingestion, and then declining with a half-life of 260 min. A similar study involving oral administration of ayahuasca to healthy volunteers similarly indicated peak plasma levels of DMT (10 ng/ml) harmaline (3 ng/ml) and harmol (12 ng/...
suggests a non-linear dose-response, making it difficult to equate harmaline and harmine being barely detectable in plasma. This relationship is supported by cardiovascular signs (Achour, 2012). We cannot attribute these findings of DMT (20 µg/L; 100 nM) the toxic effects of 5-hydroxyindoleacetic acid (5-HIAA), nor was there any decrease in the plasma levels of some of the β-carbolines. More importantly perhaps, the subjective effects of the ayahuasca peak at about 2 hrs after ingestion, and had declined almost to baseline 4 hrs later. Thus, there is a close alignment of the plasma concentrations of DMT and some β-carbolines with the intensity of the psychotropic effects. Brito-da-Costa et al. (2020) have recently presented a detailed review of the literature on DMT and β-carbolines kinetics presenting a wealth of details. Wang (2019) produced a similarly detailed investigation of the pharmacokinetics of various alkaloids in rats fed with seeds of P. harmala, the rue plant mentioned above. A single rather high dose of the seeds (150 mg/kg) resulted in plasma harmaline levels above 10 ng/ml persisting for at least 12 hrs, whereas harmine levels were around 100 ng/ml for a similar period, but harmol levels were generally below 10 ng/ml. The corresponding concentrations were much lower in rats treated with a low P. harmala dose (15 mg/kg), with harmaline and harmine being barely detectable in plasma. This suggests a non-linear dose-response, making it difficult to extrapolate these results to human ingestion of P. harmala; there is a case report of fatal toxicity due to P. harmala ingestion (Ghizlane, 2021), whereas a study from Morocco indicated a 6% fatality rate in a series of 200 cases of poisoning marked by neurological, gastrointestinal and cardiovascular signs (Achour, 2012). We cannot attribute these effects to β-carbolines per se. However, the causal relationship may become clearer in a fatality following consumption of unspecified extracts, with post mortem findings of DMT (20 µg/L; 100 nM) the toxic effects of 5-methoxy-N,N-dimethyltryptamine (2 mg/L; 10 µM) and harmine (0.2 mg/L; 1 µM) (Sklarov, 2005). However, that case report was critiqued in the contemporary literature (Callaway, 2006). Severe poisoning by β-carbolines is certainly possible, but seems to be a rare occurrence, and might properly be attributable to interactions with other pharmacologically active substances, as in the case of the “cheese effect” discussed above.

CONCLUSION

In summary, β-carbolines and other classes of MAO inhibitors are present in tobacco and many other plants, and likely contribute to the psychopharmacology of tobacco usage and perhaps dependence. However, there is incomplete documentation of the dose-response relationships; a low degree of MAO inhibition may potentiate the reinforcing properties of nicotine or amphetamine, but complete MAO blockade may be aversive. There is a particular involvement of MAO-A inhibition in the pharmacology of ayahuasca, which is generally attributed to its potentiation of the bioavailability of the actively hallucinogenic ingredient DMT. However, not all ayahuasca preparations contain significant amounts of DMT (McKenna 1984a; Callaway et al., 2005), and some Indigenous groups of the Amazon employ B. caapi without admixture plants, in the context of healing and initiatory rites (Rodd, 2008; Politi et al., 2021); we need to consider better the polypharmacology of ayahuasca and the possible independent effects of β-carbolines on brain function. The β-carbolines present in B. caapi were found for instance to stimulate neurogenesis in adult mice (Moraes-Garcia, 2017) and harmane and norharmane (i.e., the alkaloids present in tobacco) were associated with antidepressant-like effects in the mouse forced swim test (Farzin and Mansouri, 2006). Furthermore, it remains unknown if MAO inhibition increases the brain concentration of endogenous DMT, as is well established for the classical trace amines. In the context of tobacco use, what accounts for the composite effect on MAO-activity? Self-rolled cigarettes produce particulate with higher MAO-inhibitory activity than is the case in tailor-made cigarettes (Lewis, 2012). Is this due to the variety of tobacco species used, or its curing process? What is the alkaloid content of Indian beedis and do the wrapping leaves (Diospyros melanoxylon) contain pharmacologically active compounds? Clove cigarettes (kretes), which are popular in Indonesia, contain half as much nicotine as typical cigarettes, but their aromatic quality may encourage deeper inhalation and thus greater absorption of alkaloids (Malson, 2003). Where there is smoke, there is fire; this aphorism does not apply to snuff and chewing tobacco, which have not been addressed in this brief contribution and are scantily investigated with respect to their psychopharmacology. The composition of MAO-inhibiting alkaloids in different tobacco species also remains unknown. Nicotiana rustica, for instance (“Aztec tobacco,” in Peru known as “Mapacho”) in the Amazon region is considered an important medicinal plant and is a frequent component of Amerindian ethnobotanical mixtures, also elsewhere. In traditional Amazonian medicine, this tobacco species is prepared as a liquid remedy and is ingested orally to treat mental health and other ailments (Berlowitz, 2020); the rapid first pass clearance of oral nicotine could mean that unidentified alkaloids other than nicotine play a role in the psychopharmacology of this orally taken ethnomedical application of N. rustica.

In this account, we have emphasized the family resemblance of tobacco and ayahuasca; both substances contain a mixture of alkaloids, with considerable potential for interaction, especially through the inhibition of MAO by β-carbolines and other competitors. We may inadvertently have raised more questions than we have answered, but we see a clear need for more systematic examination of the separate and combined contributions of polypharmacy in the contexts of tobacco or ayahuasca consumption.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

FUNDING

This research was funded by the Swiss National Science Foundation grant number 320030_204978.
REFERENCES

Achour, S. (2012). Peganum harmala L. Poisoning in Morocco: about 200 Cases. *Therapie* 67 (1), 53–58. doi:10.2521/therapie/2012003

Ametamey, S. M. (1996). Radiosynthesis of [11C]brofaromine, a Potential Tracer for Imaging Monoamine Oxidase A. *Nucl. Med. Biol.* 23 (3), 229–234. doi:10.1016/1096-8051(95)02051-9

Balinger-Melich, P. (2019). The Effect of Electroconvulsive Therapy on Cerebral Monoamine Oxidase A Expression in Treatment-Resistant Depression Investigated Using Positron Emission Tomography. *Brain Stimul* 12 (3), 714–723. doi:10.1016/j.brs.2018.12.976

Bench, C. J. (1991). Measurement of Human Cerebral Monoamine Oxidase Type B (MAO-B) Activity with Positron Emission Tomography (PET): A Dose Ranging Study with the Reversible Inhibitor Ro 19-6327. *Eur. J. Clin. Pharmacol.* 40 (2), 169–173. doi:10.1007/BF00280072

Bergström, M., Westerberg, G., and Långström, B. (1997). 11C-harmine as a Tracer for Monoamine Oxidase A (MAO-A): In Vitro and In Vivo Studies. *Nucl. Med. Biol.* 24 (4), 287

Berlowitz, I. (2020). Tobacco Is The Chief Medicinal Plant In My Work*: Therapeutic Uses Of Tobacco In Peruvian Amazonian Medicine Exemplified By the Work Of a Maestro Tabaquero. *Frontal. Pharmacol.* 11. doi:10.3389/fphar.2020.594591

Bernard, S. (1996). Synthesis and Bladt, S., and Wagner, H. (1994). Inhibition of MAO by Fractions and Constituents *Neurochem. Res.* 19, 303 (3), 145–150. doi:10.1016/0306-4522(96)00329-9

Berlowitz et al. 2021. (2021). Molecular and Functional Imaging Studies of Psychodelic Drug Action in Animals and Humans. *Molecules* 26 (9). doi:10.3390/molecules26092451

Bhargava, I., Malik, N., and Khatkar, A. (2019). Lead Optimization for Promising Monoamine Oxidase Inhibitor from Eugenia for the Treatment of Neurological Disorder: Synthesis and In Silico Based Study. *BMC Chem.* 13 (1), 38. doi:10.1186/s13305-019-0552-4

Bodur, D. A., and Phillips, S. R. (1980). Kinetic Measurements of the Turnover Rates of Phenylylamine and Tryptamine In Vivo in the Rat Brain. *J. Neurochem.* 34 (6), 1725–1732. doi:10.1111/j.1471-4159.1980.732434x

Farzin, D., and Mansouri, N. (2006). Antidepressant-like Effect of Harmame and Other β-carbolines in the Mouse Forced Swim Test. *Eur. Neuropsychopharmacol.* 16 (5), 324–328. doi:10.1016/eunapeuro.2005.08.005

Fekkes, D., (2001). Tryptophan: a Precursor for the Endogenous Synthesis of Norharman in Man. *Neurosci. Lett.* 303 (3), 145–148. doi:10.1016/s0304-9801(01)01750-5

Fornai, F. (1999). Striatal Dopamine Metabolism in Monoamine Oxidase B-Deficient Mice: a Brain Dialysis Study. *J. Neurochem.* 73 (6), 2344–2440. doi:10.1111/j.1471-4159.1999.732434x

Fowler, J. S. (1996). Brain Monoamine Oxidase A Inhibition in Cigarette Smokers. *Proc. Natl. Acad. Sci. U S A.* 93 (24), 14065–14069. doi:10.1073/pnas.93.24.14065

Fowler, J. S., (1987). Mapping Human Brain Monoamine Oxidase A and B with 11C-Labeled Suicide Inactivators and PET. *Science* 235 (4876), 481–485. doi:10.1126/science.3099392

Fowler, J. S., (1988). Mechanistic Positron Emission Tomography Studies: Demonstration of a Deuterium Isotope Effect in the Monoamine Oxidase-Catalyzed Binding of [11C]-Deprenyl in Living Baboon Brain. *J. Neurochem.* 51 (5), 1524–1534. doi:10.1111/j.1471-4159.1988.801121.x

Fowler, J. S. (1993). Monoamine Oxidase B (MAO B) Inhibitor Therapy in Parkinson’s Disease: the Degree and Reversibility of Human Brain MAO B Inhibition by Ro 19 6527. *Neurology* 43 (10), 1984–1992. doi:10.1212/wnl.43.10.1984

Fowler, J. S., (1991). Neuropharmacological Actions of Cigarette Smoke: Brain Monoamine Oxidase B (MAO B) Inhibition. *J. Addict. Dis.* 17 (1), 23–34. doi:10.1002/jad.2500110013

Godar, S. C. (2014). The Aggression and Behavioral Abnormalities Associated with Monoamine Oxidase A Deficiency Are Rescued by Acute inhibition of Serotonin Reuptake. *J. Psychiatr. Res.* 56, 1–9. doi:10.1016/j.jpsychires.2014.04.014

Castagnoli, K., and Murugesan, T. (2004). Tobacco Leaf, Smoke and Smoking, MAO Inhibitors, Parkinson’s Disease and Neuroprotection; Are There Links? *Neurotoxicology* 25 (1-2), 279–291. doi:10.1016/j.neuro.2003.01017-4

Cumming, P., (1992). Formation and Clearance of Interstitial Metabolites of Dopamine and Serotonin in the Rat Striatum: an In Vivo Microdialysis Study. *J. Neurochem.* 59 (5), 1905–1914. doi:10.1111/j.1471-4159.1992.711026x

Durden, D. A., and Philips, S. R. (1980). Kinetic Measurements of the Turnover Rates of Phenylylamine and Tryptamine In Vivo in the Rat Brain. *J. Neurochem.* 34 (6), 1725–1732. doi:10.1111/j.1471-4159.1980.732434x

Fowler, J. S., (1996). Brain Monoamine Oxidase A Inhibition in Cigarette Smokers. *Proc. Natl. Acad. Sci. U S A.* 93 (24), 14065–14069. doi:10.1073/pnas.93.24.14065

Fowler, J. S., (1988). Mapping Human Brain Monoamine Oxidase A and B with 11C-Labeled Suicide Inactivators and PET. *Science* 235 (4876), 481–485. doi:10.1126/science.3099392

Fowler, J. S., (1988). Mechanistic Positron Emission Tomography Studies: Demonstration of a Deuterium Isotope Effect in the Monoamine Oxidase-Catalyzed Binding of [11C]-Deprenyl in Living Baboon Brain. *J. Neurochem.* 51 (5), 1524–1534. doi:10.1111/j.1471-4159.1988.801121.x

Fowler, J. S. (1993). Monoamine Oxidase B (MAO B) Inhibitor Therapy in Parkinson’s Disease: the Degree and Reversibility of Human Brain MAO B Inhibition by Ro 19 6527. *Neurology* 43 (10), 1984–1992. doi:10.1212/wnl.43.10.1984

Fowler, J. S., (1991). Neuropharmacological Actions of Cigarette Smoke: Brain Monoamine Oxidase B (MAO B) Inhibition. *J. Addict. Dis.* 17 (1), 23–34. doi:10.1002/jad.2500110013

Godar, S. C. (2014). The Aggression and Behavioral Abnormalities Associated with Monoamine Oxidase A Deficiency Are Rescued by Acute inhibition of Serotonin Reuptake. *J. Psychiatr. Res.* 56, 1–9. doi:10.1016/j.jpsychires.2014.04.014
Riba, J. (2003). Human Pharmacology of Ayahuasca: Subjective and Cardiovascular Effects. *Pharmacol. Exp. Ther.* 306 (1), 73–83. doi:10.1124/pet.103.098882

Rodd, R. (2008). Reassessing the Cultural and Psychopharmacological Significance of Banisteriopsis Caapi: Preparation, Classification and Use Among the Piaroa of Southern Venezuela. *J. Psychoactive Drugs* 40 (3), 301–307. doi:10.1080/002236408020760645

Rommelspacher, H., (2002). The Levels of Norharman Are High Enough after Smoking to Affect Monoamineoxidase B in Platelets. *Eur. J. Pharmacol.* 441 (1-2), 115–125. doi:10.1016/s0014-2999(02)01452-8

Sacher, J. (2012). Dynamic, Adaptive Changes in MAO-A Binding after Alterations in Substrate Availability: an *In Vivo* [(11)C]-harmine Positron Emission Tomography Study. *J. Cereb. Blood Flow Metab.* 32 (3), 443–446. doi:10.1038/jcbfm.2011.184

Sacher, J. (2011). Monoamine Oxidase A Inhibitor Occupancy during Treatment of Major Depressive Episodes with Moclobemide or St. John’s Wort: an [(11)C]-Harmine PET Study. *J. Psychiatry Neurosci.* 36 (6), 375–382. doi:10.1503/jpn.100117

Schoepp, D. D., and Azzaro, A. J. (1987). Kinetics of [11C]N,N-dimethylphenylethylamine in Mice and Humans: Potential for Measurement of Brain MAO-B Activity. *J. Pharmacol. Exp. Ther.* 281 (6), 1006. doi:10.1124/jpet.103.049882

Shih, J. C., and Chen, K. (2004). Regulation of MAO-A and MAO-B Gene Expression. *Carr. Med. Chem.* 11 (15), 1995–2005. doi:10.2174/0929670633647575

Shih, J. C., and Chen, K. (1999). MAO-A and -B Gene Knock-Out Mice Exhibit Distinctly Different Behavior. *Neurobiology (Bp)* 7 (2), 235

Shih, J. C., Chen, K., and Ridd, M. J. (1999). Monoamine Oxidase: from Genes to Behavior. *Annu. Rev. Neurosci.* 22, 197–217. doi:10.1146/annurev.neuro.22.1.1197

Shinotoh, H. (1987). Kinetics of [(11)C]N,N-dimethylphenylethylamine in Mice and Humans: Potential for Measurement of Brain MAO-B Activity. *J. Nucl. Med.* 28 (6), 1006.

Shulman, K. L., Herrmann, N., and Walker, S. E. (2013). Current Place of Monoamine Oxidase Inhibitors in the Treatment of Depression. *CNS Drugs* 27 (10), 789–797. doi:10.1007/s10026-013-0997-3

Simón, A. Y. (2019). Toxicological Aspects and Determination of the Main Components of Ayahuasca: A Critical Review. *Medicines (Basel)* 6 (4).

Sklarow, I. (2005). A Fatal Intoxication Following the Ingestion of 5-Methoxy-N,N-Dimethylytryptamine in an Ayahuasca Preparation. *J. Anal. Toxicol.* 29 (8), 838–841. doi:10.1093/jat/29.8.838

Slovak, T. A., DiSestefano, V., and Au, W. Y. (1970). Blood Levels and Urinary Excretion of Harman and its Metabolites in Man and Rats. *J. Pharmacol. Exp. Ther.* 173 (1), 26–30.

Smith, T. T. (2015). Effects of MAO Inhibition and a Combination of Minor Alkaloids, β-carbolines, and Acetaldehyde on Nicotine Self-Administration in Adult Male Rats. *Drug Alcohol Depend* 155, 243–252. doi:10.1016/j.drugalcdep.2015.07.002

Spijker, R. (2002). The Impact of Smoking and Drinking on Plasma Levels of Norharman. *Eur. Neuropsychopharmacol.* 12 (1), 61–71. doi:10.1016/s0924-977x(01)00143-2

Suzuki, O., Oya, M., and Katsumata, Y. (1979). Oxidation of P-, M- and O-Tyramine by Type A and Type B Monoamine Oxidase. *Biochem. Pharmacol.* 28 (17), 2682–2684. doi:10.1016/0006-2952(79)90050-9

Tarkowska, D. (2020). A Fast and Reliable UHPLC-MS/MS-Based Method for Screening Selected Pharmacologically Significant Natural Plant Indole Alkaloids. *Molecules* 25 (14), 3274. doi:10.3390/molecules25143274

Thase, M. E., Trivedi, M. H., and Rush, A. J. (1995). MAOs in the Contemporary Treatment of Depression. *Neuropsychopharmacology* 12 (3), 185–219. doi:10.1016/s0893-133x(94)00058-8

Truman, P., Grounds, P., and Brennan, K. A. (2017). Monoamine Oxidase Inhibitory Activity in Tobacco Particulate Matter: Are Harman and Norharman the Only Physiologically Relevant Inhibitors? *Neurotoxicology* 59, 22–26. doi:10.1016/j.neuro.2016.12.010

Truman, P. (2019). Monoamine Oxidase Inhibitory Activity of Flavoured E-Cigarette Liquids. *Neurotoxicology* 75, 123–128. doi:10.1016/j.neuro.2019.09.010

Ulrich, S., Rücken, R., and Adli, M. (2017). Tranylcypromine in Mind (Part I): Review of Pharmacology. *Eur. Neuropsychopharmacol.* 27 (8), 697–713. doi:10.1016/j.euroneuro.2017.04.003

van Amsterdam, J., (2006). Contribution of Monoamine Oxidase (MAO) Inhibition to Tobacco and Alcohol Addiction. *Life Sci.* 79 (21), 1969–1973. doi:10.1016/j.lfs.2006.06.010

Van Den Eijnden, R., Spijkerman, R., and Fekkes, D. (2003). Craving for Cigarettes Among Low and High Dependent Smokers: Impact of Norharman. *Addict. Biol.* 8 (4), 463–472. doi:10.1080/1355621031000164457

van der Toorn, M., (2019). Comparison of Monoamine Oxidase Inhibition by Cigarettes and Modified Risk Tobacco Products. *Toxicol. Rep.* 6, 1206–1215. doi:10.1016/j.toxrep.2019.11.008

Varnás, K. (2021). Effects of Sevoflurane Anaesthesia on Radioligand Binding to Monoamine Oxidase-B *In Vivo*. *Br. J. Anaesth.* 126 (1), 238–244. doi:10.1016/j.bja.2020.08.052

Villégier, A. S. (2007). Involvement of Alpha1-Adrenergic Receptors in Tranylcypromine Enhancement of Nicotine Self-Administration in Rat. *Psychopharmacology (Berl)* 193 (4), 457–465. doi:10.1007/s00213-007-0799

Wang, Y. (2019). Subchronic Toxicity and Concomitant Toxicokinetics of Long-Term Oral Administration of Total Alkaloid Extracts from Seeds of Peganum Harmala Linna: A 28-day Study in Rats. *J. Ethnopharmacol.* 238, 111866. doi:10.1016/j.jep.2019.111866

Yang, H. Y., and Neff, N. H. (1973). Beta-phenethylamine: a Specific Substrate for Type B Monoamine Oxidase of Brain. *J. Pharmacol. Exp. Ther.* 187 (2), 365–371.

Youdim, M. B., and Weinstock, M. (2004). Therapeutic Applications of Selective and Non-selective Inhibitors of Monoamine Oxidase A and B that Do Not Cause Significant Tyramine Potentiation. *Neurotoxicology* 25 (1-2), 243–250. doi:10.1016/s0196-325x(03)00103-7

Zhang, L., Ashley, D. L., and Watson, C. H. (2011). Quantitative Analysis of Six Heterocyclic Aromatic Amines in Mainstream Cigarette Smoke Condensate Using Isotope Dilution Liquid Chromatography-Electrospray Ionization Tandem Mass Spectrometry. *Nicotine Tob. Res.* 13 (2), 120–126. doi:10.1093/ntr/ntq219

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