Antioxidant Potential of Pyrimidine Derivatives against Oxidative Stress

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Nair et al.: Antioxidant Potential of Pyrimidine Derivatives

The human body is under constant attack from oxidative stress which in turn induces the excessive production of free radicals, particularly the reactive oxygen species in the body which causes damage to cells, proteins and deoxyribonucleic acid. Free radicals are associated with progression of numerous pathological disruptions in humans including cancer, inflammation, atherosclerosis, Alzheimer’s disease, Parkinson’s disease and aging as well. Antioxidants are the so called “free radical scavengers” of the body, with ability to reduce the risk of chronic diseases by diminishing oxidative damage. Pyrimidines are heterocyclic molecule with ability to be utilized in therapeutics as antioxidant. Recently, novel approaches have been made and significant progress has occurred in last few years to overcome damage caused by free radical generation using pyrimidine derivatives. The current review focuses on synthesizable pyrimidine derivatives that show promise towards future development as antioxidants with therapeutic purpose.

Key words: Antioxidant, pyrimidines, free radicals, oxygen scavenger, superoxides

Heterocyclic compounds have contributed to development of large number of novel drugs till now[1]. Many heterocyclic compounds exist in nature as part of structural unit of bioactive molecules like alkaloids, vitamins, hormones, haemoglobin and amino acids whereas synthetic heterocyclic compounds like pyridine[2], piperidines[3], pyrimidines[4] have shown to have significant biological activities. Pyrimidine and N-base analogues of uracil, thymine and cytosine have a significant role in constitutional part of Deoxyribonucleic Acid (DNA) and Ribonucleic Acid (RNA) in the form of nitrogenous bases as well as several coenzymes in the body[5]. The importance of the pyrimidine and its derivative compounds may be established by the fact that many drugs contain pyrimidine nucleus[6]. As a result, several pyrimidine derivatives have gained attention due to interesting pharmacological activities such as antitumor[7], antiviral[8], antioxidant[9], anti-malarial[10,11], antibacterial[12-14], anti-fungal[15,16], anti-inflammatory[17] and anti-hypertensive activity[18].

Reactive Oxygen Species (ROS) like peroxides, superoxide and hydroxy peroxides are produced either as the metabolic by-products of oxidative metabolism as a consequence of both enzymatic and nonenzymatic reactions of different substrates like fatty acids, glucose and amino acids to produce energy in the body or from external sources such as exposure to X-rays, ozone, cigarette smoking, air pollutants and industrial chemicals[19,20]. Free radicals produced by oxidative metabolism may undergo a chain reaction in presence of water, which may lead to oxidative damage to the cell structures, vital biomolecules-lipids, nucleic acids and proteins and thus have deleterious effect on the body by bringing about cell damage and homeostatic disruption. ROS such as superoxide, hydrogen peroxide, hydroxyl radical and their metabolites play an important role in carcinogenesis. ROS causes crosslinking and induce DNA damage. Free radicals and other reactive species are also produced by radiolysis or by direct action of radiation on DNA; these effects cause cell mutation and cancer. Lipid peroxides are also considered responsible for carcinogenesis[19,20].

Antioxidants are free radical scavengers which play a crucial role in prophylaxis and prevent propagation...
of diseases due to excessive oxidative stress in the body\cite{21}. However, to prevent those reactions there are many protective compounds present or produced in the cell such as superoxide dismutase, catalase, glutathione peroxidase, tocopherol, ascorbic acid, glutathione and uric acid\cite{22}. These antioxidants trap the free radicals by donating an electron to the rampant free radical and neutralize it, thus reducing its capacity to damage and prevent the irreversible adverse effects on the cells, while they may become new free radical which is less active and less dangerous\cite{23}. They also inhibit the reaction involved in lipid peroxidation and due to this property, they have found an important place in therapy and control of several diseases like atherosclerosis, DNA damage leading to mutation and cancer, neurodegenerative disease like Alzheimer’s disease, Parkinsonism and ageing process\cite{24}. Small molecules such as vitamin C, vitamin E, uric acid and glutathione play an important role as intracellular antioxidants whereas synthetic antioxidants such as tert-butyl hydroxyl-toluene, Tert-Butylhydroxyanisole (TBHA) and Tert-Butylhydroquinone (TBHQ) have been widely used in the food industry to retard lipid peroxidation. However, these synthetic antioxidants are not preferred for pharmacological use due to toxicological concerns\cite{25}. Thus, focus is shifted more on identification, synthesis and production of safe and non-toxic antioxidant for medicinal use. This review focuses on current trend in the search of non-toxic molecules, synthesisable pyrimidine derivatives that show promising potency and can serve as potential antioxidants.

**CLINICALLY USED ANTIOXIDANTS HAVING PYRIMIDINE RING**

As observed in studies, excess production of free radicals leads to many deleterious effects on the body and in order to overcome this oxidative stress the body synthesizes many free radical scavengers\cite{26}. With progression of age, the defensive mechanism of body is unable to do so and the antioxidants have to be supplemented from outside the body. To serve this purpose, many antioxidants with pyrimidine nucleus such as riboflavin, thiamine, pyrazolopyrimidine, triazolopyrimidines have been synthesized in past years (fig. 1). In spite of the availability of many antioxidants with pyrimidine nucleus only few of them are used clinically due to their toxic effects\cite{27}.

Riboflavin is commonly known as “B group water soluble vitamin” or specifically as B2 vitamin. Glutathione is a crucial antioxidant produced by human body but it is active only in reduced form. Riboflavin and the glutathione reductase enzyme are both used to convert oxidized glutathione to the reduced form thereby depicting antioxidant activity. Riboflavin plays an important role in maintaining the levels of other antioxidant enzymes, such as superoxide dismutase, catalase and glutathione peroxidase and also helps in the prevention and treatment of many eye disorders\cite{28}. The clinically used riboflavin formulations available in the market are riboflavin injection, Photrexa® (eyedrop), capsules (Bacmin, Biovita), Alertonic, Active FE, B-combo, B-Plex, C-Nate DHA and Centratex.

Thiamine another B family vitamin commonly known as B1 vitamin, possess remarkable antioxidant activity. It has been shown to inhibit lipid peroxidation in rat liver microsomes in vivo and free radical oxidation of oleic acid in vitro. Thiamine act by transfer of 2H+ from NH2 group of pyrimidine ring and H+ from thiazole ring to reactive free radical species. Thiamine in turn reacts with ROS and itself gets oxidized to thiochrome and thiamine disulphide. Formulations are available which can be used in deficiency of thiamine or therapeutically
for the treatment of polyneuropathies and cardiovascular disease that are exaggerated by oxidative stress[29]. Some clinical formulations of thiamine used recently include benfotiamine, which is an S-acyl derivative, allithiamine, synthetic sulbutiamine, betaxin and fursultiamine (thiamintetrahydrofurfuryl disulphide). Benfotiamine being lipid insoluble formulations shows maximum bioavailability than other water soluble ones.

**EVALUATION OF ANTIOXIDANT ACTIVITY**

Evaluation of antioxidant activity is necessary for determining the potency of existing and newly synthesized compounds for comparing their relative activities. Numerous methods have been identified for evaluation of antioxidants based on various principles.

**In vitro methods:**

A single antioxidant test method is inefficient in determining the absolute activity of the compound therefore several methods are used for evaluation of antioxidant activities. 2,2-diphenyl-1-picrylhydrazyl (DPPH) method is the most commonly used in vitro method, however different methods have their own advantages and disadvantages.

**DPPH method:** DPPH is the simplest and widely reported method. DPPH radical scavenging test is a standard and rapid technique for screening of Radical Scavenging Activity (RSA)[17]. It measures the ability to donate hydrogen or electron to RSA for evaluation of antioxidant activity. In this method, the extent of bleaching of the purple colour of methanolic solution of DPPH is used to indicate the antioxidant activity of the test compound. The absorbance of observed colour is established under wavelength 517 nm using spectrophotometric method. DPPH is a stable free radical due to delocalisation of spare electron on it and do not allow its dimerization like other free radicals.

\[ \text{Percentage scavenged H}_2\text{O}_2 = \left( \frac{A_i - A_t}{A_i} \right) \times 100 \]

Where, \(A_i\) is absorbance of control and \(A_t\) is absorbance of test compound[30].

**Nitric Oxide (NO) scavenging test:** In this method NO free radicals were generated from sodium nitroprusside at physiological pH which is then combined with oxygen to form stable compound. This NO free radical is determined quantitatively using Griess reagent[31]. The absorbance is measured by colorimetric method at 546 nm. The percentage inhibition of NO radicals is given by the equation:

\[ \text{Percentage inhibition (NO radical)} = \left( \frac{A_0 - A_i}{A_0} \right) \times 100 \]

Where \(A_0\) is absorbance before reaction and \(A_i\) is the absorbance measured with Griess reagent[32].

**Ferric reducing antioxidant power assay:** In this method the ability of a compound to reduce ferric ion to ferrous ion at low pH, is measured to evaluate antioxidant activity. A diode-array spectrophotometer is used to measure the absorbance at 593 nm. During the course of reaction, colour changes from various shades of green and blue due to reduction which depends upon the reducing ability expressed as mm of Fe²⁺ equivalents per kg of the material. This method was developed by Benzie et al.[33].

**Hydrogen peroxide (H₂O₂) scavenging activity:** H₂O₂ is generated in the body during various metabolic processes and is rarely reactive unless it is converted to reactive hydroxyl radical (OH⁻) on exposure to Ultraviolet (UV) which may lead to various deleterious effects[34]. H₂O₂ assay is the measure of ability of the sample to scavenge H₂O₂. The hydroxyl radicals are reacted with dimethyl sulfoxide to yield formaldehyde. The released formaldehyde produces intense yellow colour with Nash reagent (2 M ammonium acetate with 0.05 M acetic acid and 0.02 M acetyl acetone in distilled water). The intensity of the colour is measured at 412 nm spectrophotometrically against reagent blank. Formaldehyde produces strong yellow blank reagent[35].

\[ \text{Percentage scavenged H}_2\text{O}_2 = \left( \frac{A_0 - A_i}{A_0} \right) \times 100 \]

Where, \(A_i\) is absorbance of control and \(A_t\) is absorbance of test.

**Total radical-trapping antioxidant parameter method:** This method is based on the fact that antioxidants are involved in protection of fluorescence decay of R-phycocerythrin (R-PE) during peroxidation reaction and decrease in fluorescence of R-PE by a radical generator 2,2'-azobis(2-amidinopropane) (ABAP) is measured in the presence of antioxidant. The antioxidative activity is evaluated by measuring the decay in decolouration. According to Ghiselli et al.[36] a diluted sample of 120 µl is added to phosphate buffer (pH 7.4, 2.4 ml), diluted R-PE (30 µl), bi-distilled water (375 µl) and 75 µl of ABAP; the reaction kinetics method the ability of a compound to reduce ferric ion to ferrous ion at low pH, is measured to evaluate antioxidant activity. A diode-array spectrophotometer is used to measure the absorbance at 593 nm. During the course of reaction, colour changes from various shades of green and blue due to reduction which depends upon the reducing ability expressed as mm of Fe²⁺ equivalents per kg of the material. This method was developed by Benzie et al.[33].

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Oxygen Radical Absorbance Capacity (ORAC) method: ORAC is a revolutionary new test tube analysis for estimating antioxidant power of foods and chemical substances. It is performed using b-Phycoerythrin (b-PE) or fluorescein as target molecule. Trolox is employed as a standard in this test to determine the Trolox Equivalent (TE). The ORAC value is then calculated using TE and expressed as ORAC units or value and higher the ORAC value, greater the scavenging capability against free radicals. Based on generation of free radical using 2,2-azobis(2-amidinopropane) dihydrochloride (AAPH), this assay measures the decrease in fluorescence in the presence of free radical scavengers. Litescu et al.\cite{38} described an automated ORAC assay. In this assay b-PE was used as a target, Trolox as standard control and AAPH as a peroxy free radical generator where on addition of AAPH to the test solution, the fluorescence is recorded and the antioxidant activity is expressed as TE.

In vivo methods:

These methods consist of administration of the test compound into the testing animals in a definite dose. After a specified time, the tissues and cells from the animals are subjected to evaluation by sacrificing the animals.

Reduced glutathione method: Glutathione-SH (GSH) is a reductase enzyme present inside the cells of the body. It protects the cell structure from ROS, peroxides and other toxic compounds. Tissue homogenate is added to Ellman’s reagent and the absorbance is measured at 412 nm against blank. The absorbance of test compounds is compared with standard curve of GSH. The method was developed by Owens et al.\cite{39}.

Ferric reducing ability of plasma: This method is based on the same principle as in vitro ferric reducing power assay described in the in vitro method. It includes measurement of increase in the absorbance due to formation of ferrous ions from ferric reducing ability of plasma reagent containing 2,4,6-Tri(2-pyridyl)-s-triazine (TPTZ) and Ferric Chloride Hexahydrate (FeCl₃.6H₂O). The absorbance is measured at 593 nm.\cite{40}

Catalase assay: Aebi method can be used to determine catalase activity in erythrocyte lysate by using a spectrophotometer. At 240 nm, the catalase activity is measured for 1 min and the molar extinction coefficient of H₂O₂, which is 43.6 M per cm, is used to determine the activity, where 1 unit of activity is equal to 1 mmol of H₂O₂ degraded per minute and is expressed as units per milligram of protein.

Superoxide Dismutase (SOD) method: This method is given by McCord and Fridovich. The method can be used for estimating antioxidant activity in the erythrocyte lysate which is prepared from the 5 % Red Blood Corpuscles (RBC) suspension by adding 50 μl of the erythrocyte lysate to 75 mM of Tris-HCl buffer (pH 8.2), followed by 30 mM Ethylenediaminetetraacetic Acid (EDTA) and 2 mM of pyrogallol and the absorbance is measured in spectrometer for 3 min which shows an increase in absorbance at 420 nm. One enzyme activity exhibited 50 % inhibition of the rate of autooxidation of pyrogallol as determined by change in absorbance/min at 420 nm. The activity of SOD is expressed as units per mg of protein.\cite{41}

Lipid Peroxidation (LPO) assay: LPO assay is an autacatalytic process generally followed by cell death. This process may lead to peroxidative tissue damage and cause inflammation, cancer and toxicity of xenobiotics and aging. Malondialdehyde (MDA) is one of the end products in the lipid peroxidation process, which is accepted as an indicator of lipid peroxidation. In this method described by Okhawa, the tissues are homogenized in 0.1 M buffer of pH 7.4 with a Teflon glass homogenizer. Amounts of MDA produced primarily are used to determine the lipid peroxidation in the homogenate. Tissue homogenate of 8.1 % sodium dodecyl sulphate, acetic acid and 2-thiobarbituric acid are added then heated at 95°. Then tubes are cooled down to room temperature and final volume made to 5 ml in each tube. 5 ml of butanol:pyridine (15:1). The upper organic layer is taken after centrifugation and its optical density is measured at 532 nm against an appropriate blank. The lipid peroxide level can be expressed in n moles of thiobarbituric acid reactive substances/mg protein using an extinction coefficient of 1.56×10⁵ MLcm⁻¹.\cite{19}

CURRENT TRENDS IN EVALUATION OF ANTIOXIDANT ACTIVITY

DPPH method for assay gives more accurate results but there are few methods available which have other advantages and are used very often for carrying out the assay procedure of antioxidant compounds.

Trolox Equivalent Antioxidant Capacity (TEAC) assay:

TEAC method is a widely accepted method for this purpose. TEAC method provides the measure of
relative antioxidant activity of a given substance with reference to a standard substance, Trolox\[^{[41]}\]. Three methods for TEAC have been developed over the time. These methods may be used interchangeably which may create significant variability in the measurements of the antioxidants. The ‘pre-addition technique’ (employed by adding antioxidants before radical generation) for TEAC I measured the delaying ability of radical formation and scavenging property of the compound. 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid (ABTS)/TEAC method is used widely today because of its operational simplicity, reproducibility, diversity and most importantly it provides flexible use in hydrophilic and lipophilic antioxidant capacity\[^{[42]}\].

**LEAD COMPOUND AS ANTIOXIDANT TEMPLATES**

Chandrashekaraiah *et al.*\[^{[43]}\] synthesized a series of pyrimidine-azitidinone analogues. All compounds are tested for antioxidant property by DPPH, NO and \(H_2O_2\) methods. DPPH method for assay of antioxidants measures the hydrogen donating capacity of the molecules in the sample. Compounds 1a1, 1a2 and 1a3 (fig. 2a), due to the presence of mild electron donating groups such as chloro groups attached to the rings, showed good radical scavenging activity in all three methods when compared with the standard drug ascorbic acid. The Half-maximal inhibitory concentration (IC\(_{50}\)) value of the standard ascorbic acid in DPPH method was found to be 15.15 at 25 \(\mu g/ml\) whereas the IC\(_{50}\) values of the compounds 1a1, 1a2 and 1a3 were found to be 17.72, 17.21 and 16.92 \(\mu g/ml\) respectively\[^{[43]}\].

Rangaswamy *et al.*\[^{[44]}\] synthesized series of novel 2,4,6-substituted pyrimidine derivatives conjugated by sulfonyl chloride attached with benzoferan at C-6. All the newly synthesized compounds were evaluated for their antioxidant activity by using three *in vitro* assays DPPH radical scavenging activity\[^{[17]}\], N,N-dimethyl-p-phenylenediamine (DMPD) and ferric ion reducing power assay. Among the synthesized compounds, 1b and 1b1 (fig. 2b) possessed maximum radical activity as well as were more potent than the standard when 4-hydroxyphenyl substitution at S-linked C-2 and C-6 positions of pyrimidine was present as well as with 4-hydroxy-3-methoxyphenyl substitution at C-6 position of pyrimidine. IC\(_{50}\) (DPPH) values of these compounds were 08 and 10 micro molar respectively\[^{[44]}\].

Khoobi *et al.*\[^{[45]}\] synthesized novel series of 4H-chromenes containing a pyrimidine-2-thione. The antioxidant activities of the synthesized chromenopyrimidinethiones were evaluated by two *in vitro* methods (DPPH and ABTS) in order to compare the results. Two controls, ascorbic acid and Trolox are included. All pyrimidinethione derivatives

![Fig. 2: (a) Compounds 1a1, 1a2 and 1a3 and (b) Compounds 1b1 and 1b2](image-url)
exhibited significant DPPH radical scavenging activity against their precursors with IC\textsubscript{50} values less than 42 \(\mu\)g/ml. The ABTS radical cation scavenging capacity of chromenopyrimidinethiones demonstrated that all compounds have good ability to trap free radicals. The IC\textsubscript{50} values of synthesized compounds were in the range of 21.28-72.98 \(\mu\)g/ml. Among the chromenopyrimidinethiones, compound 2a (fig. 3a) exhibited the most potent antioxidant activity in both DPPH and ABTS methods. Although compounds 2a was not as potent as reference drug Trolox, but its antioxidant activity could be considered as associated property of the title compound\cite{45}.

Kotaiah \textit{et al.}\cite{21} synthesized a series of 1,3,4-oxadiazole tagged thieno[2,3-d]pyrimidine derivatives. All the compounds are tested for \textit{in vitro} antioxidant property by DPPH, NO and \(\text{H}_2\text{O}_2\) methods respectively. Compounds 2b, 2c, 2d and 2e (fig. 3b) showed good radical scavenging activity in all three methods due to the presence of mild electron donating groups such as difluoro, fluoro and chloro-fluoro groups attached to the benzene rings when compared with the standard drug, ascorbic acid. The IC\textsubscript{50} value of the standard ascorbic acid in DPPH method was found to be 15.11 mg/ml at 25 mg/ml whereas the IC\textsubscript{50} values of the compounds 2b, 2c, 2d and 2e were found to be 16.35, 16.91, 17.25 and 17.70 mg/ml respectively\cite{21}.

Rani \textit{et al.}\cite{46} synthesized new series of tetrahydroimidazo[1,2-\(\alpha\)]pyrimidine-6-carboxamide analogues. The antioxidant activity of the synthesized compounds was evaluated using free radical scavenging DPPH assay \textit{via} spectrophotometer and it was demonstrated that compounds 3a and 3b (fig. 4a) exhibited excellent activity due to the presence of electron releasing groups on benzylidene moiety. At absorbance of 517 nm the compounds show IC\textsubscript{50} values of 46.31 and 48.81 respectively which compares with standard drug ascorbic acid. These compounds may be used as a lead for development of new age antioxidant\cite{46}.

Mansouri \textit{et al.}\cite{47} synthesized some novel Biginelli-type pyrimidines which are 4-(furan-2-yl)-6-methyl-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carboxylate esters. Their antioxidant potential was assessed using three different methodologies, namely DPPH free radical scavenging, reducing power and hydrogen peroxide scavenging assays and two standards, ascorbic acid and gallic acid were included. Compound 3c (fig. 4b) was the most potent antioxidant with the IC\textsubscript{50} of 0.6 mg/ml. The results of reducing power assays proved 3d and 3e as the moderate reducing agents. All of the studied compounds were very weak compared with gallic acid in scavenging hydrogen peroxide\cite{47}.

Bhalgat \textit{et al.}\cite{48} synthesized a series of novel pyrimidines and its triazole fused derivatives. All of these synthesized compounds when screened for \textit{in vitro} antioxidant activity by various methods such as scavenging of hydrogen peroxide, scavenging of nitric oxide.

\begin{table}[h]
\centering
\begin{tabular}{|c|c|c|c|c|}
\hline
\(\text{R}\) & 2b & 2c & 2d & 2e \\
\hline
R & 4-F & 4-F & 2,4-dif & 2,4-dif \\
R\(_1\) & 4-H & 4-Cl & 4-H & 4-Cl \\
\hline
\end{tabular}
\caption{Chemical structures of compounds 2a and 2b, 2c, 2d and 2e}
\end{table}
oxide radical, lipid peroxidation inhibitory activity revealed that some of the tested compounds showed moderate to good antioxidant activity. 4a has shown good antioxidant activity as compared to standard by scavenging of nitric oxide radical and scavenging of hydrogen peroxide method, while 4b showed most potent antioxidant activity by scavenging of nitric oxide radical method (fig. 5)[48].

Abu-Hashem et al.[49] synthesized a series of thiazolopyrimidines, pyrrolothiazolopyrimidines and triazolopyrrolo-thiazolopyrimidines. The newly developed compounds were subjected for their antioxidant activity; out of the synthesized compounds, 5a and 5b (fig. 6) manifested potent antioxidative activity in the lipid peroxidation assay. Novel thiazolopyrimidine derivatives incorporated with carbohydrazide, amino, oxadiazol and other moieties also possess potential antioxidant activities[49].

Quiroga et al.[50] synthesized a new compound of 5-aryl-4-oxo-3,4,5,8-tetrahydropyrido[2,3-d] pyrimidine-7-carboxylic acids and conduct their antioxidant activity by DPPH radical scavenging method. Compounds 6a and 6b (fig. 7) showed good antioxidant activity compared to standard drugs[50].

Mondal et al.[51] synthesized a series of 1-(2-mercapto-6-(substituted phenyl)pyrimidine-4-yl)-3-(2-substituted phenylimino)indoline-2-one and 1-(2-amino-6-(substituted phenyl)pyrimidine-4-yl)-3-(2-substituted phenylimino)indolin-2-ones from different substituted chalconised indole-2,3-diones. Compounds 7a2, 7a3, 7b2, 7b3, 7a6, and 7b6 (fig. 8) show more promising antioxidant activity due to the substitution of the Sulphhydryl (SH) and NH$_2$ groups at the second position of the pyrimidine ring[51].

Gressler et al.[52] synthesized 4-trifluoromethyl-2-(5-aryl-3-styryl-1H-pyrazol-1yl)-pyrimidine derivatives. These molecules were screened for their in vitro antioxidant activity using the DPPH- which measures the hydrogen-donating capacity of the molecules and Horseradish Peroxidase (HRP)/luminol/ H$_2$O$_2$ chemiluminescence assay method based on the light emission produced by a chemical reaction. When SH group is present at R$_3$ position, compound 8a shows potent antioxidant (fig. 9a)[52].

Kumar et al.[53] synthesized a novel series of 4,6-bisaryl-pyrimidin-2-amine derivative. The synthesized compounds, when evaluated for their antioxidant activity by nitric oxide and hydrogen peroxide free radical scavenging method using ascorbic acid as a standard drug, compound and compound 8b4 (fig. 9b) show potent antioxidant activity as compare with the standard drug due to the presence of -Cl and -Br as the electron withdrawing group at positions R$_1$ and R$_2$. Compound 8b4 was evaluated using nitric oxide free radical scavenging method and by hydrogen peroxide free radical scavenging method. IC$_{50}$ values of the compound were are 0.019 and 0.020 mol/l respectively[53].

Mohana et al.[54] reported novel series of pyrimidine derivatives which were evaluated for antioxidant activity by DPPH method. The structural analysis of all these molecules was done on the basis of Fourier transform infrared spectroscopy, Proton Nuclear Magnetic Resonance (°H-NMR) and mass spectral
data. All the compounds showed DPPH radical scavenging activity, whereas, compounds 9a, 9b and 9c (fig. 10) exhibited best radical scavengers due to presence of electron donating methoxy group at different position (ortho, meta and para)[54].

Panda et al.[55] synthesized a novel series of indolyl-pyrimidine derivatives and all the synthesized molecules were screened for antioxidant activity using DPPH radical scavenging method. Compounds 10a, 10b, 10c, 10d, 10e, 10f and 10g (fig. 11) have exhibited good antioxidant activity which is comparable with that of standard drug, ascorbic acid[55]. Table 1 compares these pyrimidine derivatives based on their antioxidant activity.

**DRUG CANDIDATES UNDER CLINICAL TRIAL**

The reactive oxygen project team of Pharmacia and Upjohn has identified series of novel pyrrolo[2,3-d] pyrimidine derivatives to be orally active antioxidants and inhibitor of lipid peroxidation which may be used in myocardial infarction, Parkinson's disease, Alzheimer's
Disease, multiple sclerosis, amyotrophic lateral sclerosis and asthma. The latest in this series 1-[(2,4-dipyrrolidinyl-9H-pyrimido[4,5-b]indol-9-yl)-acetyl] pyrrolidine monohydrochloride (PNU-142731A), which has shown promising antioxidant activity orally and is currently undergoing phase I clinical trials.\textsuperscript{[36]}

Fig. 8: Mercapto-pyrimidine and amino-pyrimidine derivatives of indoline-2-one

\[
\begin{array}{|c|c|c|}
\hline
R & Cl & NO_2 \\
\hline
 & NO_2 & OH & OCH_3 \\
\hline
\end{array}
\]

Fig. 9: (a) Compound 8a: 2-(4,5-Dihydro-1-H-pyrazol-1-yl)-pyrimidine and (b) Compounds 8b1-8b4
In recent studies it has been found that excess of tyrosine kinase Mitogen-Activated Protein Kinase (MEK) 1 and MEK2 leads to tumour. On this basis orally active kinase inhibitors like trametinib and ibrutinib (fig. 12) has been approved by Food and Drug Administration (FDA) to be used as antitumor
in patients with inoperable or metastatic melanoma in early 2012. Trametinib is now used in combination with dabrafenib. Ibrutinib has undergone extensive set of clinical trials in 2013 and FDA granted approval for use of this drug in specific lymphoma. Trametinib and ibrutinib are available in the market by trade name MEKINIST® and IMBRUVICA®.[57]

**CONCLUSION**

The antioxidants have the ability to trap as well as scavenge ROS and may be of great value in preventing the onset and propagation of different diseases. Pyrimidine is a unique molecule and has a long and distinguished history extending from

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TABLE 1: COMPARATIVE ANTIOXIDANT ACTIVITY OF SYNTHETIC PYRIMIDINE DERIVATIVES

| S. No | Lead compounds as good antioxidants | Evaluation methods | Antioxidant activity |
|-------|-------------------------------------|--------------------|---------------------|
| 1     | Pyrimidine-azetidinone analogues (1a1,1a2,1a3) | DPPH, NO and H₂O₂ | Good               |
| 2     | Benzofuran-Gathered C-2,4,6-substituted pyrimidine derivatives conjugated by sulfonyl chlorides (1b1,1b2) | DPPH, DMPD and ferric ion reducing power assay | Maximum            |
| 3     | Chromenopyrimidinethiones (2a) | DPPH and ABTS assay | Most potent         |
| 4     | 1,3,4-oxadiazole tagged thieno[2,3-d]pyrimidine derivatives (2b,2c,2d,2e) | DPPH, NO and H₂O₂ | Good               |
| 5     | Tetrahydroimidazo[1,2-a] pyrimidine derivatives (3a,3b) | DPPH | Excellent          |
| 6     | 4-(furan-2-yl)-6-methyl-2-thioxo-1,2,3,4-tetrahydropyrimidine 5-carboxylate ester (3c) | DPPH, H₂O₂ and reducing power assay | Most potent         |
| 7     | Triazole fused pyrimidine derivatives (4a,4b) | H₂O₂, Lipid peroxidation inhibitory activity | Most potent         |
| 8     | Thiazolopyrimidines, Pyrrolothiazolopyrimidines and Triazolopyrrolothiazolopyrimidines derivatives (5a,5b) | ABTS, erythrocyte hemolysis and bleomycin dependent DNA damage | Potent              |
| 9     | 5-aryl-4-oxo-3,4,5,8-tetrahydropyrido[2,3-d]pyrimidine-7-carboxylic acids (6a,6b) | DPPH, ORAC | Good               |
| 10    | Mercapto-pyrimidine and amino-pyrimidine derivatives of indoline-2-one (7a2, 7a3, 7b2, 7b3, 7a4 and 7b4) | DPPH | Promising          |
| 11    | 4-trifluoromethyl-2-(5-aryl-3-styryl-1H-pyrazol-1-yl)-pyrimidine derivatives (8a) | DPPH and HRP/luminol/H₂O₂/chemiluminescence assay | Potent              |
| 12    | 4,6-bisaryl-pyrimidin-2-amine derivative (8b1-8b4) | NO and H₂O₂ method | Potent              |
| 13    | New series of pyrimidine derivatives (9a,9b, 9c) | DPPH | Best               |
| 14    | Indolyl-pyrimidine derivatives (10a-10g) | DPPH | Good               |

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**Fig. 12: Drug candidates under clinical trials**
the days of discovery as important constituents of nucleic acid to their wide clinical applications including in cancer. Various pyrimidine derivatives have shown promising antioxidant activity. Majority of compounds shown significant radical scavenging properties due to the presence of electron donating substituent on the pyrimidine nucleus which enhances the radical scavenging activity and serves to improve the potency. The antioxidants bearing pyrimidine nucleus are not able to enter in the lipid moiety of low-density lipoprotein, hence these types of compounds portray very low penetration power, in turn reducing effectiveness. Substitution with electron donating groups like Br, NO₂, CF₃ and Cl on the pyrimidine nucleus may increase the penetration of molecules into the lipid membrane which increases the antioxidant activity by combining with the ROS, which is generated in different disease conditions. However, the electron withdrawing substituent may decrease the antioxidant activity. In short, high antioxidant activity can be correlated with low electron density of ring system owing to good antioxidant activity of substituted pyrimidine.

Conflict of interests:

The authors declared no conflicts of interest.

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