Potential roles of mediator Complex Subunit 13 in Cardiac Diseases

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Abstract

Mediator complex subunit 13 (MED13, previously known as THRAP1 and TRAP240) is a subunit of the cyclin-dependent kinase 8 (CDK8) kinase module in the eukaryotic mediator complex. MED13 has been known to play critical roles in cell cycle, development, and growth. The purpose of this review is to comprehensively discuss its newly identified potential roles in myocardial energy metabolism and non-metabolic cardiovascular diseases. Evidence indicates that cardiac MED13 mainly participates in the regulation of nuclear receptor signaling, which drives the transcription of genes involved in modulating cardiac and systemic energy homeostasis. MED13 is also associated with several pathological conditions, such as metabolic syndrome and thyroid disease-associated heart failure. Therefore, MED13 constitutes a potential therapeutic target for the regulation of metabolic disorders and other cardiovascular diseases.

Key words: MED13; cardiovascular diseases; energy metabolism

Introduction

Aberrant gene transcription causes serious human diseases [1]. The mediator complex (MED) acts as a structural and functional bridge between DNA-binding transcription factors (TFs) and the basal transcriptional machinery [2]. It helps regulate the expression of RNA polymerase II-transcribed genes, as well as transcription initiation and elongation [3]. Mediator subunit 13 (MED13, known as THRAP1 and TRAP240) is part of the MED [4]. MED13 maintains the transcription of some essential proteins involved in growth and development, and MED13 gene mutations are associated with various diseases, including cardiac diseases. MED13 plays critical roles in diseases like metabolic syndrome and cardiovascular disease. This review will cover the basic structure and functions of MED13 and its potential roles in metabolic and non-metabolic cardiac diseases. Understanding MED13 biology is critical for its optimal manipulation as a potential therapeutic target.

MED and MED13 structure and function

Eukaryotic transcription requires RNA polymerase II and various transcriptional repressors and activators, including TFs, that bind to specific DNA sequences in gene enhancer regions. TFs function is regulated by transcriptional co-regulators [5]. The MED acts as a co-regulator and contains multiple subunits (approximately 25 and 30 subunits in yeast and humans, respectively). It functions as an adaptor between TFs bound to upstream regulatory elements and the transcription machinery, which includes RNA polymerase II and general TFs (GTFs), such as transcription initiation factor II (TFII) A, TFIIIB, TFIID, TFIIe, TFIIF, and TFIIF [5-7]. The MED is recruited by TFs to regulatory regions and binds to RNA polymerase II and GTFs that have been recruited to the core promoter to drive formation of the
preinitiation complex. The MED mainly transmits signals from TFs to RNA polymerase II (Figure 1) and regulates the transcription of specific genes, such as those for nuclear receptors (NRs) [8, 9].

Electron microscopy data indicate that the MED is composed of four modules: the head, middle, tail, and cyclin-dependent kinase 8 (CDK8) kinase module (CKM) [5, 10]. The CKM comprises CDK8, cyclin C (Cyc C/CCNC), MED13, and MED12 [11] (Figure 1). Three CKM proteins, CDK8, MED13, and MED12, have paralogs that can replace them in the kinase domain, although they do not perform exactly the same functions [12]: cyclin-dependent kinase 19, mediator subunit 13 like protein (MED13L, originally called PROSIT240 or THRAP2), and mediator subunit 12 like protein (MED12L). MED13L is similar in size to MED13, with which it shares 50% identity at the protein level [13]. CKM can regulate transcription by blocking the MED–RNA polymerase II association, thereby preventing transcription initiation or reinitiation [14]. The MED undergoes a conformational change and forms a dynamic bridge between enhancers and promoters via reversible dissociation of CKM from the MED [11, 15] (Figure 1).

MED13 plays a connective role between the CKM and the MED core [16]; therefore, targeting MED13 may affect MED transcriptional co-regulatory function. In yeast, Caenorhabditis elegans, zebrafish, Drosophila, and mouse, the MED13 sequence is conserved, and the protein regulates gene expression in numerous physiological growth and developmental processes [12, 17-21] (Table 1). Moreover, MED13 is critical in myogenesis, zygotic genome activation, prevention of mitochondrial fission, and programmed cell death [12, 21, 22]. In mice, MED13/MED13L has an essential function in embryonic development. Snijders Blok et al. [23] reported three missense mutations (p.Pro327Ser, p.Thr326Ile, and p.Pro327Gln) and one in-frame deletion (p.Thr326del) in a conserved phosphodegron of human MED13. Normally, this phosphodegron is recognized by the S-phase kinase-associated protein (Skp)-Cullin-F-box (SCF) F-box and WD repeat domain-containing 7 (Fbw7) ubiquitin ligase complex (SCF/Fbw7), resulting in MED13 protein ubiquitination and degradation [24], suggesting that these mutations may affect MED13 protein turnover. Some MED13 gene mutations are reportedly associated with abnormal heart development [25, 26], neurodevelopmental disorders, nervous system disorders, craniofacial dysmorphisms, and muscle diseases [23, 27, 28] (Table 1). MED13L gene mutations are also associated with these phenotypes, but with more variable features [13, 26, 29-36]. MED13 not only plays a critical role in necessary physiological processes, including organismal growth and development, but is also involved in pathophysiological processes and diseases.

**Figure 1. The main structure and function of the mediator complex (MED).** The MED is composed of four parts: the head, middle, tail, and CKM. The CKM includes CDK8/19, Cyc C/CCNC, MED12/12L, and MED13/13L. (A) The CKM associates with the MED via interaction between MED13 and a “hook” at the end of the MED (middle module). Targeted degradation of MED13 can affect the association between the CKM and the MED. (B) An activator (Act) recruits the MED to an enhancer and then recruits GTFs. (C) The MED helps to recruit RNA polymerase (Pol) II to the promoter via interaction with the non-phosphorylated carboxy-terminal domain (CTD) of RNA polymerase II. MED also helps recruit other factors to promote the formation of the preinitiation complex. (D) This process includes CKM dissociation from the MED. (E) Then, the CTD of RNA polymerase II is phosphorylated (P-CTD) by the GTF TFIIH, which is accompanied by MED dissociation, thereby allowing RNA polymerase II to escape from the promoter and initiate transcription. The MED also regulates RNA polymerase II pausing and elongation. (CKM, cyclin-dependent kinase 8 (CDK8) kinase module; GTF, general transcription factor.)
MED13 degradation under oxidative stress

Elevated levels of reactive oxygen species can be observed during aging and after exposure to environmental stress [22]. In yeast, in response to oxidative stress, Cyc C is released from the nucleus into the cytoplasm, where it interacts with mitochondria to regulate cell death [44, 45]. This process is dependent on MED13 degradation. Under normal conditions, MED13 is required for Cyc C retention in the nucleus, binding Cyc C via intrinsic disordered regions (IDRs) [22, 46]. SCF protein GRR1 ubiquitin ligase complex (SCF<sub>Grr1</sub>)-mediated degradation of MED13 targets its IDRs. CDK8 phosphorylates MED13 in the absence of stress. However, in response to oxidative stress, the cell wall integrity/MAP kinase/Slt2 pathway is activated and Slt2 phosphorylates Cyc C and MED13. The phosphorylated sites are within the same MED13 IDRs that are associated with Cyc C and SCF<sub>Grr1</sub> activity. CDK8- and Slt2-mediated phosphorylation of IDRs in MED13 release Cyc C from MED13, leading to its recognition by SCF<sub>Grr1</sub> and subsequent degradation [47]. In addition, the AMP kinase Snf1 activates another SCF<sub>Grr1</sub>-responsive degron of MED13 [48]. Thus, MED13 degradation plays a critical role in mediating Cyc C involvement in mitochondrial fragmentation and cell death following oxidative stress. However, in mammalian cells, only 10% of total Cyc C is released, and the overall level of Cyc C protein expression does not change under oxidative stress [46]. In human cells, SCF<sub>FBw7</sub> regulates MED13 degradation and ubiquitylation, which may use a similar two-kinase mechanism of MED13 degradation [24, 47]. Additional research investigating changes in MED13 during oxidative stress in higher eukaryotes is warranted.

Pathophysiological role of MED13 in the heart

MED13 is ubiquitous in human tissues, with highest MED13 expression found in the placenta, skeletal muscle, heart, pancreas, and brain [49]. MED13/MED13L plays an important role in physiological heart development. Pediatric patients with cyanotic congenital heart disease exhibit higher rates of adenosine-to-inosine (A-to-I) RNA editing of MED13 intronic segments than healthy individuals, which likely affects gene expression by altering RNA splicing and mRNA nuclear retention, among other mechanisms [25, 50]. Moreover, defects in MED13L are related to transposition of the great arteries and coarctation of the aorta [13, 26, 33]. Mutations of the human MED13L gene lead to MED13L haploinsufficiency syndrome, which includes congenital heart defects [40, 51]. Cardiac MED13 is involved in systemic energy metabolism, and is associated with obesity, diabetes, and other diseases linked to energy metabolism. In addition, cardiac MED13 participates in regulation of cardiac function with thyroid hormone (TH).

Cardiac MED13 modulates energy metabolism through NR signaling

Cardiac overexpression of MED13 in transgenic (α-myosin heavy chain [αMHC]-MED13-TG or MED13-cTG) mice promotes sensitivity to insulin and increases energy expenditure, thereby preventing high-fat diet (HFD)-induced obesity. Conversely, cardiac-specific deletion of MED13 promotes obesity in response to HFD and leads to metabolic syndrome, characterized by glucose intolerance and fatty liver [42]. Moreover, the expression of cardiac MED13 mRNA is lower in Zucker diabetic fatty rats than in their lean counterparts [52]. This finding is consistent with the function of cardiac MED13 in metabolic regulation. In addition, the expression of cardiac MED13 mRNA in female Zucker lean rats is higher than that in males, which may explain why female rats are able to maintain insulin sensitivity and glucose tolerance while male rats are not. This sex-specific difference in MED13 expression indicates that MED13 is a protective factor in the hearts of healthy female rats [52].
Figure 2. Role of cardiac-specific MED13 in systemic energy metabolism. Nebivolol (Neb) and rapamycin (Rap) can inhibit expression of miR-208a by suppressing activation of mTORC1. The expression of MED13, a downstream target gene of miR-208a, is downregulated by miR-208a. Overexpression of cardiac MED13 inhibits the association of RNA polymerase II (Pol II) with the MED, thereby repressing transcription and expression of NR target genes involved in cardiac energy metabolism. Changes in cardiac energy metabolism alter systemic energy metabolism. Another regulator of cardiac MED13 is KLF5, which can promote MED13 expression in the heart. (MED, mediator complex; NRs, nuclear receptors; NRE, NR response element; T3, triiodothyronine).

The heart requires a constant supply of energy, which is maintained by communication between the heart and energy storage depots in the body. Thus, heart energy metabolism affects whole-body energy metabolism and homeostasis (Figure 2). Expression of cardiac energy metabolism genes can be regulated by NRs [53]. The MED reportedly interacts with NRs and NR response element (NRE) as a transcriptional co-activator when triiodothyronine (T3, a thyroid hormone) is present [54]. MED13 in the heart inhibits MED-dependent NR signaling [42] to suppress the transcription of some NR-responsive metabolic genes (Figure 2). Deletion of these downregulated NR-responsive genes can result in resistance to HFD-induced obesity and protection against insulin resistance [42]. In addition, other TFs involved in regulation of lipid biosynthesis and metabolism, such as NCoR, SREBP, and PPARγ [53, 55, 56], can be modulated by cardiac MED13 as responsive genes [42]. Therefore, cardiac overexpression of MED13 represses metabolic genes in the heart to drive whole-body energy metabolism.

Signaling pathways that potentially regulate cardiac MED13 expression

There are two potential pathways that regulate cardiac MED13 involvement in energy metabolism: the mammalian target of rapamycin complex 1 (mTORC1)/miR-208a/MED13 pathway and the Krüppel-like transcription factor 5 (KLF5)/MED13 pathway. Regarding the first potential pathway, chronic over-nutrition can lead to excessive activation of mTORC1 and downstream signaling, thereby inducing metabolic disorder [57]. Therefore, increased mTORC1 activation is a key initiator of obesity [57, 58]. Rapamycin is a canonical inhibitor of mTORC1. Nebivolol can also suppress mTORC1 activation in the hearts of Zucker obese rats [59]. Gul et al. [60] demonstrated that rapamycin and nebivolol, which confer resistance to obesity in animal models, suppress mTORC1 activation and miR-208a expression in HL-1 cardiomyocytes. These results suggest that mTORC1 promotes the expression of miR-208a (Figure 2), a cardiac-specific small noncoding RNA encoded by an intron in the α-MHC gene. Deletion of miR-208a gene can increase MED13 expression, suggesting that miR-208a is negatively regulating MED13 expression [61] (Figure 2). Conversely, upregulating MED13 expression confers resistance to obesity in animal models upon pharmacologic miR-208a inhibition or rapamycin treatment. These results suggest that miR-208a is an upstream target of MED13, and reports have confirmed a miR-208a binding target site in the 3′ untranslated region (UTR) of the MED13 gene [61-63]. Taken together, these findings suggest that the mTORC1/miR-208a/MED13 axis plays a role in regulating cardiac and systemic energy metabolism. In agreement with these results, exercise training can prevent obesity and cardiac pathological hypertrophy.

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by increasing MED13 expression via regulation of miR-208a [64].

The second potential pathway is the KLF5/MED13 signaling pathway. KLF5 as one of the KLF family is a TF to involve in regulating lipid metabolism [65]. KLF5 in the heart reportedly modulates the expression of cardiac fatty acid metabolism-related genes [66, 67]. Moreover, deletion of cardiac KLF5 results in susceptibility to diet-induced obesity in mice, which can be suppressed by expression of cardiac MED13. Cardiac MED13 expression decreases in αMHC-KLF5-/- mice. Furthermore, KLF5 overexpression increases MED13 expression in cardiomyocytes [68] (Figure 2). Thus, MED13 may be the key downstream target of KLF5 for metabolic regulation. Using promoter analysis, two KLF5-binding sites were identified in mouse and human MED13 promoters. Furthermore, chromatin immunoprecipitation experiments with HL-1 cells suggest the presence of a KLF5-binding site in the mouse MED13 promoter [68]. These results indicate that KLF5 is a positive regulator of cardiac MED13 mediated metabolic regulation.

**Coordination of cardiac MED13 and thyroid hormone to regulate cardiac transcription**

There is a close relationship between thyroid hormone [TH, including T3 and thyroxine (T4)] and cardiovascular system [69-71]. High level of TH increases heart rate, cardiac output, and myocardial contractility. Decreased TH causes opposite functions [71]. Clinical and animals evidence indicate that hypothyroidism can induce heart failure and is associated with poor prognosis [72-74]. The main role of TH in the heart is to regulate cardiac-specific gene transcription by interacting with TH receptors (TRs), and TRs bind to thyroid hormone response elements (TREs) in gene promoter regions [71]. Cardiac contractility function can be affected by expressions of the α-MHC (myh6) and β-MHC (myh7) genes. TH stimulates expression of the α-MHC gene and inhibits expression of the β-MHC gene in the heart after birth [75] (Figure 3). Conversely, hypothyroidism suppress α-MHC and induce β-MHC expression, which is associated with cardiac dysfunction [63, 71]. Although β-MHC is the main isofrom in the ventricles of healthy adult humans, even small changes in expression of the two isoforms can meaningfully alter cardiac function. TH also positively regulates sarcoplasmic reticulum Ca2+ ATPase (Serca2a) gene expression [69, 76, 77] (Figure 3). Serca2a is an important Ca2+ pump for the maintenance of Ca2+ homeostasis, which is associated with myocardial calcium handling and plays a critical role in maintaining cardiac function [78, 79].

To investigate whether miR-208a is required for hypothyroidism-up-regulating β-MHC, van Rooij et al. treat miR-208a-/- mice with propylthiouracil (PTU, an inhibitor of T3) to make a hypothyroidism model, and found that β-MHC is not upregulated in response to hypothyroidism [63], suggesting that miR-208a is required for increased expression of β-MHC by hypothyroidism. MED13 (known as thyroid hormone receptor associated protein 1), a downstream target of miR-208a [63], can regulate transcription and the activity of TRs via RNA polymerase II and GTF recruitment [42, 63, 80, 81]. Therefore, in response to hypothyroidism, there would be prediction that overexpression of miR-208a inhibits MED13 and additional targets expression, thereby enhancing β-MHC expression via reducing the repressive activity of TRs towards β-MHC and altering cardiac function (Figure 3).

To further test the impact of cardiac MED13 on the response of the heart to low level of TH (hypothyroidism), Olson et al. treated MED13-cTG mice with PTU, which displayed increased cardiac systolic function compared with wild-type mice [42] (Figure 3). Moreover, in the context of hypothyroidism, deletion of cardiac MED13 exacerbates cardiac dysfunction. The expressions of myh7 and Serca2a genes are lower in cardiac MED13 knockout (MED13-cKO) mice than wild-type mice [82], suggesting that cardiac MED13 plays a role in cardiac function conservation in hypothyroidism, as myh7 and Serca2a expressions are necessary to maintain normal cardiac function.

The expression cardiac MED13 is increased in response to hypothyroidism and the increased MED13 expression might repress cardiac pro-inflammatory and pro-fibrotic gene transcriptions [82], which may be a potential mechanism for MED13 modulating cardiac transcription in the context of hypothyroidism (Figure 3).

**Involvement of MED13 in non-metabolic cardiovascular diseases**

Cardiac MED13 expression is mainly involved in energy homeostasis. However, whether the effects of MED13 on cardiac energy metabolism influence cardiac function remains unclear. In addition, miR-208b-3p/MED13/Wnt signaling plays a role in cardiomyocyte apoptosis in the hypoxic/reoxygenated state [83]. miR-208 is downregulated during right ventricular failure, which activates the MED13/NCoR1 pathway and inhibits myocyte enhancer factor 2 (MEF2) [84]. Additional research is required to determine whether cardiac MED13 is involved in other cardiovascular diseases and if it regulates cardiac function.
Potential MED13 crosstalk among organs

The interactions between metabolic organs and the central nervous system are crucial for maintaining whole-body energy homeostasis and may be altered in metabolic diseases such as obesity and diabetes. The central nervous system and peripheral organs produce bioactive molecules to communicate with other tissues, including cytokines and hormones [85]. Cardiac and skeletal muscles are the key organs involved in regulating energy metabolism. MED13 is expressed in multiple organs associated with energy metabolism that participate in crosstalk.

Heart-adipose tissue/liver crosstalk

The heart requires a considerable energy supply, and may signal peripheral organs to convey its needs. Therefore, cardiac MED13, which affects cardiac energy metabolism, may control whole-body energy homeostasis. Olson et al. [42] reported that overexpression of MED13 in the mouse heart reduces fat accumulation in peripheral tissues. Deletion of cardiac MED13 in mice or Drosophila increases their susceptibility to HFD-induced obesity [42, 86]. In addition, MED13-cTG mice exhibit 60% greater lipid clearance rate in the blood than that in wild-type mice.

Figure 3. Possible mechanism of MED13-mediated regulation of cardiac function in response to hypothyroidism. Top panel: The T3 induces α-MHC gene transcription through TR binding a positive TRE and inhibits β-MHC transcription via TR binding a negative TRE. The intron in the α-MHC gene encodes miR-208a. In the context of hypothyroidism (PTU inhibits T4 synthesis and suppresses conversion from T4 to T3), miR-208a may inhibit cardiac MED13 and additional targets expression, which regulates TH receptor activity and β-MHC gene expression. T3 also enhances expression of Serca2a via TR binding TREs in its regulatory region. Changes in the level of α/β-MHC and Serca2a expression could affect cardiac function in the context of hypothyroidism. Lower panel: Cardiac contractility function is suppressed by PTU. Overexpression of cardiac MED13 ameliorates cardiac dysfunction induced by hypothyroidism, whereas deletion of cardiac MED13 aggravates hypothyroidism-related cardiac dysfunction. The expression of serca2a and myh7 are lower in the hearts of MED13-cKO mice than wild-type mice treated with PTU and pro-inflammatory/pro-fibrotic genes expression are higher in MED13-cKO mice. These changes are associated with worse cardiac dysfunction in MED13-cKO mice than in wild-type mice. (PTU, propylthiouracil; T4, thyroxine; T3, triiodothyronine; TR, thyroid hormone receptor; TRE, thyroid hormone response element).
Lipid oxidation in the white adipose tissue (WAT) and liver is also higher in MED13-cTG mice than in wild-type mice. Furthermore, lipid uptake, fatty acid \( \beta \)-oxidation, mitochondrial content, energy consumption, and many genes involved in fatty acid metabolism and the Krebs cycle are upregulated in the WAT and liver of MED13-cTG mice, leading to a lean phenotype [88]. The heart appears to send metabolic signals to peripheral tissues via circulating factors (Figure 4) because heterotypic wild-type parabiosis have a lean phenotype in heterotypic parabiosis experiments [88]. Several cardiokines, including atrial natriuretic factor and B-type natriuretic peptides, are involved in cardiac physiological and pathological processes [87]. However, the serum levels of metabolic hormones and cardiokines are not elevated in MED13-cTG mice, and other circulating factors levels need to be confirmed [88]. Moreover, in \( \alpha \)MHC-KLF5-/- mice, cardiac MED13 expression is lower than in wild-type mice, whereas the levels of FGF21 in the plasma and heart are higher in HFD-fed \( \alpha \)MHC-KLF5-/- mice.

Muscle-adipose tissue/liver crosstalk

Muscle plays a central role in systemic energy metabolism, and functions as a secretory organ. MED13 mediates crosstalk between skeletal muscle and adipose tissue of Drosophila, and between muscle and adipose tissue/liver of mice. Olson et al. [86] reported that expression of MED13 in muscle in Drosophila controls obesity. In addition, muscle-specific knockdown of MED13 in mice increases their susceptibility to obesity. Therefore, the function of MED13 in the muscle of Drosophila is the same as its role in the heart, with regard to adipose tissue crosstalk. The muscle-secreted obesity-associated factor Wingless (Wg) functions as a downstream target of MED13 to mediate muscle-adipose crosstalk and repress obesity [86] (Figure 4).

Figure 4. Crosstalk among energy metabolism organs mediated by MED13. Overexpression of cardiac MED13 in mice increases liver and WAT fatty acid metabolism and reduces lipid accumulation in these tissues via regulation of circulating factors. Genetic deletion of KLF5 in the mouse heart induces lower MED13 expression and higher FGF21 expression in the plasma and heart, which activates the PPAR\( \gamma \) pathway in WAT and induces obesity. Expression of hepatic MED13 and mitochondrial fatty acid metabolism are higher in miR-378/378* knockout mice than in wild-type controls. However, opposite effects are observed upon overexpression of muscle-specific MED13 in mice and Drosophila. Upregulation of muscle MED13 in Drosophila activates the Wnt/\( \beta \)-catenin pathway in the muscle and adipose tissues. Wingless (Wg) acts downstream of muscle MED13 in Drosophila to enhance lipid metabolism and suppress obesity. In mice, genetic deletion of muscle MED13 improves glucose tolerance and metabolism, and confers resistance to hepatic steatosis. Moreover, skeletal muscle MED13 represses the NURR1/MEF2 cooperative pathway, thereby reducing the expression of the glucose-handling gene GLUT4 (WAT, white adipose tissue).
The effect of murine muscle MED13 on metabolic homeostasis is opposite to that of cardiac MED13. Deletion of muscle MED13 in mice (MED13-mKO) fed HFD improves their glucose tolerance, enhances muscle glucose metabolism, and prevents hepatic steatosis [89] (Figure 4). In addition, RNA sequencing data reveals that gene expressions of nuclear receptor NURR1, salt-inducible kinase 1 (which activates the TF MEF2), and glucose transporter type 4 (GLUT4) are upregulated in MED13-mKO muscle. Muscle-specific MED13 primarily represses the MEF2/NURR1 cooperative pathway, thereby suppressing expression of glucose handling genes, such as GLUT4 [89-91]. The opposite functions of MED13 in the heart and muscle in systemic energy metabolism suggest tissue-specific functions of MED13 in the heart and muscle in handling genes, such as GLUT4 [89-91]. The opposite functions of MED13 in the heart and muscle in systemic energy metabolism suggest tissue-specific functions for MED13 via various signaling pathways.

Liver-adipose tissue crosstalk

The liver is also an organ associated with energy metabolism, and MED13 mediates crosstalk between the liver and adipose tissue. The 3' UTR of the MED13 gene includes three sites that can be recognized by sequences of miR-378*, and a luciferase reporter assay demonstrates repression of the MED13 3' UTR in response to increasing expression of miR-378/ miR-378* [43]. In addition, mice with genetic deletion of miR-378/miR-378* exhibit higher hepatic MED13 expression than wild-type mice, and hepatic MED13 regulates energy homeostasis via the transcriptional network regulated by the transcriptional co-regulator PGC-1β [43, 50]. The miR-378/miR-378* knockout mice display resistance to HFD-induced obesity, with less fat accumulation in the WAT than their wild-type counterparts [43] (Figure 4). These findings suggest that hepatic MED13 is a target of miR-378*, and that hepatic MED13 could influence adipose tissue and obesity.

Furthermore, WAT and brown adipose tissue can be interconverted through a process known as adipose organ remodeling. MED13 is expressed in various energy metabolism organs and mediates crosstalk via circulating factors or other signaling pathways. Therefore, MED13-mediated crosstalk may participate in adipose organ remodeling through endocrine factors [92].

The role of MED12 in cardiac diseases and its link with MED13

MED12, as a part of CKM, is closely related to MED13. MED12 also plays a critical role in the development and growth of organisms just like MED13 [19, 93-97]. Reports have showed that mutation of the MED12 gene in zebrafish and mouse affected heart development [98, 99]. In the MED12 gene mutant mouse embryonic stem (ES) cell model, embryos live no more than 10.5 days and exhibit heart malformations, including enlarged hearts and cardiac dysfunction. Furthermore, the canonical Wnt and Wnt/β-catenin signaling is impaired in this model [99]. In addition, MED12 was reported to interact with Nanog to regulate Nanog target genes in mouse ES cells, but these results cannot be reproduced in other models [99, 100]. An alternative transcript of the MED12 gene may be involved in endothelial differentiation [101]. Reportedly mutations of the MED12 gene in humans cause similar phenotypes as mutations in the MED13 and MED13L genes. MED12-associated disorders include Opitz-Kaveggia syndrome (FG syndrome) type 1, Lujan syndrome, and X-linked Ohdo syndrome, and all that share cardiac malformations [102-106].

MED12 and MED13 have similar functions in controlling obesity in Drosophila, especially heart and muscle MED12 and MED13. Knockdown of cardiac MED12 and MED13 in Drosophila increases fat accumulation and induces obesity, which suggests that like MED13, cardiac MED12 also can control metabolism homeostasis. In addition, deletion of muscle MED12 and MED13 in Drosophila also results in similar obesity phenotypes [86].

Conclusions and Perspective

MED13 regulates transcription and influences organismal growth and development [107]. In humans, MED13 gene mutations are associated with abnormal organ development, leading to congenital heart defects and neurological disorders. To date, the functions of MED13 have been mainly characterized in cardiac and systemic energy metabolism, through which it is associated with metabolic diseases such as obesity and diabetes. Understanding the molecular mechanisms underlying MED13-associated disorders and pathological conditions will facilitate the identification of potential therapeutic targets for clinical application. For example, miR-208a and miR-378/378* reportedly downregulate MED13 expression. Therefore, pharmacological agents targeting these miRNAs, such as nebivolol and rapamycin, may prove therapeutic by modulating expression of cardiac MED13 or MED13-regulated cardiokines in metabolic disorders [60, 87].

Nevertheless, we must consider that these therapeutic interventions may cause unintended effects due to the diverse functions of MED13 in different tissues. miRNA antagonists or agonists will affect many targets and thus require thorough investigation for adverse effects. Dexmedetomidine has been shown to protect cardiomyocytes from apoptosis through the miR-208b-3p/MED13/Wnt
signaling pathway under hypoxic conditions. Furthermore, MED13 regulates cardiac transcription with TH under hypothyroid conditions. In conclusion, MED13 provides a possible therapeutic target in metabolic and non-metabolic cardiac diseases. However, many challenges to their clinical application remains, in particular, the accompanying adverse effects need to be elucidated.

Abbreviations

MED13: Mediator complex subunit 13; MED13L: Mediator subunit 13 like protein; CDK8: Cyclin-dependent kinase 8; MED: Mediator complex; TFs: Transcription factors; GTFs: General transcription factors; TFII: Transcription initiation factor II; NRs: Nuclear receptors; CKM: Cyclin-dependent kinase 8 kinase module; Cyc C/CCNC: Cyclin C; MED12: Mediator complex subunit 12; MED12L: Mediator subunit 12 like protein; CDK19: Cyclin-dependent kinase 19; RNA Pol II: RNA polymerase II; SCF: S-phase kinase-associated protein-Cullin-F-box; Fbw7: F-box and WD repeat domain-containing 7; IDRs: Intrinsic disordered regions; A-to-I: Adenosine-to-inosine; TH: Thyroid hormone; HFD: High fat diet; KLF5: Krüppel-like transcription factor 5; UTR: Untranslated region; TRs: Thyroid hormone receptors; TREs: Thyroid hormone-response elements; MHC: Myosin heavy chain; Serca2a: Sarcoplasmic reticulum Ca²⁺ ATPase; WAT: White adipose tissue; MEF2: Myocyte enhancer factor 2; ES cells: Embryonic stem cells.

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Authors’ contributions

WZ and HC conceptualized, drafted and reviewed the manuscript. JL,HX,XW, and HM searched and summarized the literature and reviewed the manuscript. YZ conceptualized, edited, and reviewed the manuscript, and acquired funding. LC edited and reviewed the manuscript.

Competing Interests

The authors have declared that no competing interest exists.

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