Abstract: L-Arginine (Arg) is oxidized to l-citrulline and nitric oxide (NO) by the action of endothelial nitric oxide synthase (NOS). In contrast, protein-incorporated Arg residues can be methylated with subsequent proteolysis giving rise to methylarginine compounds, such as asymmetric dimethylarginine (ADMA) that competes with Arg for binding to NOS. Most ADMA is degraded by dimethylarginine dimethylaminohydrolase (DDAH), distributed widely throughout the body and regulates ADMA levels and, therefore, NO synthesis. In recent years, several studies have suggested that increased ADMA levels are a marker of atherosclerotic change, and can be used to assess cardiovascular risk, consistent with ADMA being predominantly absorbed by endothelial cells. NO is an important messenger molecule involved in numerous biological processes, and its activity is essential to understand both pathogenic and therapeutic mechanisms in kidney disease and renal transplantation. NO production is reduced in renal patients because of their elevated ADMA levels with associated reduced DDAH activity. These factors contribute to endothelial dysfunction, oxidative stress and the progression of renal damage, but there are treatments that may effectively reduce ADMA levels in patients with kidney disease. Available data on ADMA levels in controls and renal patients, both in adults and children, also are summarized in this review.

Keywords: asymmetric dimethylarginine (ADMA); arginine (Arg); children; dimethylarginine dimethylaminohydrolase (DDAH); endothelial dysfunction; kidney; methylarginines; nitric oxide; oxidative stress; renal failure
**Abbreviations:** ADMA, asymmetric dimethylarginine; Arg, L-arginine; cGMP, cyclic guanosine monophosphate; Citr, citrulline; cNOS, constitutive nitric oxide synthase; CRP, C reactive protein; DDAH, dimethylarginine dimethylaminohydrolase; eNOS, endothelial nitric oxide synthase; GFR, glomerular filtration rate; Hcys, homocysteine; iNOS, inducible oxide nitric synthase; LDL, low density lipoproteins; NMMA, N-monomethyl-L-arginine; nNOS, neuronal oxide nitric synthase; NO, nitric oxide; NOS, nitric oxide synthase; PRMT, protein arginine methyltransferase; RTx, renal transplantation; SDMA, symmetric dimethylarginine.

1. Introduction

In addition to the traditional cardiovascular risk factors, other factors associated with renal failure involved in common metabolic pathways have recently been studied in detail. The metabolism of amino acids tends to be impaired in patients with chronic renal failure due to abnormalities in their synthesis or excretion. For example, one of the alterations of amino acid metabolism in patients with chronic renal failure is hyperhomocysteinaemia; this is considered to be an independent risk factor for cardiovascular disease [1] and occurs in more than 85% of patients after kidney transplant (RTx) and in those with end-stage renal failure [2].

The new metabolic pathways currently being described may help, to some extent, to group these conditions by pathophysiological characteristics and to identify possible alternative therapeutic approaches. In particular, it seems interesting to study the metabolism of arginine and associated metabolic pathways. For this, it is useful to determine the levels of arginine and its methylated derivatives as part of the analysis of the cardiovascular risk and endothelial dysfunction of renal disease.

2. Nitric Oxide and the Kidney

The vascular endothelium has many functions and, accordingly, endothelial dysfunction is responsible for numerous health problems including atherosclerosis, high blood pressure, sepsis, thrombosis, vasculitis, and bleeding, among others. One of the most important functions of the endothelium is to secrete nitric oxide (NO), a relatively unstable diatomic free radical, which can be synthesized by a broad range of organisms. This molecule has a role as a messenger in many biological processes in humans including involvement in the regulation of neurone communication, antimicrobial activity, ventilation, hormone secretion, inflammation and immune responses as well as vascular tone [3]. Indeed, it is a potent vasodilator and levels are often reduced when endothelial function is impaired, making it a vascular risk factor and, in particular, together with dyslipidaemia, a risk factor for coronary disease. Nitric oxide was first described as an endothelium-derived vascular relaxant factor, but its role in vasoilation depends on an increase in the levels of cyclic guanosine monophosphate (cGMP) in smooth muscle cells (Figure 1). In this case, NO is synthesized from arginine by the enzyme nitric oxide synthase (NOS), which has been identified in neurons, endothelial cells, macrophages and hepatocytes in various different isoforms: inducible (iNOS) and constitutive (cNOS), which contains neuronal (nNOS), and endothelial (eNOS).
Figure 1. Nitric oxide (NO) synthesis in vascular endothelium and its diffusion to smooth-muscle cells where soluble guanylyl cyclase (sGC) is stimulated resulting in enhanced synthesis of cyclic guanosine monophosphate (GMP).

In the kidney, where the synthesis of arginine occurs mainly in the proximal tubules, cNOS has been found in the glomerules, vessels and tubular segments including the macula densa and inner medullary segments of the collecting duct system [4,5]. The iNOS isoform has been found in the smooth muscle cells of blood vessels, the distal end of the efferent arteriole and the medullary area of the ascending limb of the loop of Henle [6]. Cytokines that stimulate this inducible form (iNOS) have been found in cultures from proximal tubules, inner medullary segments of the collecting duct system and the mesangium [7].

Renal medullary blood flow varies in response to endothelium-dependent vasodilation and endothelial cells of the vasa recta renis are capable of producing NO, which can have an influence on transport within the collecting duct system. The inhibition of NO synthesis in the kidney may have numerous consequences for patients:

- Reduced glomerular blood flow, together with an increase in the vascular resistance of the afferent and efferent arterioles;
- Reduced ultrafiltration, renal blood flow and glomerular filtration rate (GFR);
- Decreased secretion of renin, a hormone involved in the sodium and water balance in the body;
- Reduced ability to excrete sodium under normal conditions;
- Increased blood pressure and deterioration in renal function;
- Lack of stimulation for Na⁺ and HCO₃⁻ transport in the nephron proximal tubules mediated by cGMP;
- Production of oxygen reactive species;
- Production of nitric peroxide when exposed to superoxide anions.

In brief, it has relatively recently been discovered that NO is an important messenger molecule involved in numerous biological processes, and knowledge about its activity is essential to help us understand both pathogenic and therapeutic mechanisms in kidney disease.
2.1. Endothelial Dysfunction and Oxidative Stress in Kidney Disease

Several theories indicate that endothelial dysfunction is what predisposes individuals to rapid atherosclerosis that, in turn, is involved in the pathogenesis of high blood pressure and associated with renal function decline [8]. On the other hand, it has been suggested that a reduction in NOS activity or an increase in its degradation rate, due to oxidative stress, may cause endothelial damage [9].

Oxidative stress in patients who have undergone RTx is mainly due to endothelial dysfunction caused by the inhibition of NO synthesis. It should, however, be taken into account that the use of immunosuppressants also influences oxidative stress in patients after RTx, since cyclosporine and tacrolimus cause post-transplant hypertension, which weakens defenses against oxidative stress [10,11]. In the case of tracrolimus, it has been described how the immunosuppressant can cause inhibition of the activity and transcription of eNOS [12], leading to a reduction in NO levels. Reduced production of endothelial NO could also be a direct consequence of therapy with cyclosporine since it induces calciuria, and this would increase the risk of early atherothrombosis in patients after RTx [13].

It is very important to study the role of arginine in this type of patient, since NOS activity in kidney failure is determined by arginine concentration [14], the key role of the kidney in the production of the NO precursor making renal patients prone to NO deficiency.

2.2. Arginine-Nitric Oxide Metabolism

L-arginine (Arg) can be broken down by several metabolic pathways: apart from its transformation into guanidinoacetate and creatine, it can be oxidized to L-citrulline (Citr) and NO by endothelial NOS (its main substrate) (Figure 2a). In line with this, acute or chronic administration of Arg increases the production of endothelial nitric oxide and improves endothelial function, decreasing the risk of atherothrombosis [15].

**Figure 2. (a)** Arginine-nitric oxide metabolic pathway. The abbreviations stand for the following compounds: nitric oxide synthase (NOS), asymmetric dimethylarginine (ADMA), nitric oxide (NO), protein arginine methyltransferase (PRMT) and dimethylarginine dimethylaminohydrolase (DDAH); (b) Molecular diagram illustrating the inhibitory role of asymmetric dimethylarginine (ADMA) on nitric oxide (NO) production.
On the other hand, protein Arg residues can be methylated in various ways such that their subsequent proteolysis gives rise to three types of compounds: asymmetric dimethylarginine (ADMA), symmetric dimethylarginine (SDMA) and N-monomethyl-L-arginine (NMMA) (Figure 2b). Arg methylation is catalysed by type I and II protein arginine methyltransferase (PRMT), whose activity also depends on S-adenosylmethionine as a donor of methyl groups. Type I PRMT leads to ADMA, which is produced in all body cells and competes with Arg for the NOS isoforms, while type II PRMT gives raise to NMMA and SDMA, which does not directly affect NOS activity, but may compete with Arg in the transport system $y^+$. Type I PRMT is expressed predominately in the endothelium and smooth muscle cells of the cardiovascular system [16]. Under oxidative stress, low-density lipoproteins (LDLs) and oxidized LDLs may increase the activity of the enzyme in endothelial cells, leading to an increase in the production of ADMA [17]. Unmetabolized ADMA is released to the blood stream by the membrane transport system $y^+$. In addition, a small percentage (10%) of the
clearance of ADMA occurs by urinary excretion. Further, although SDMA does not inhibit NO synthesis directly, it may compete with the cationic amino acid transporter of the membrane of the endothelial cells, accentuating the Arg deficiency and indirectly altering the level of NO production, it having been reported that SDMA itself significantly reduces Arg uptake in the loop of Henle of the rat nephrons [18]. For these reasons and the fact that SDMA is mainly eliminated through urine, many research groups are studying its correlation with renal function [19–21], with creatinine excretion [22], with inflammatory parameters [23], and with mechanisms that regulate blood pressure [24], and it is considered to be a marker for cardiovascular disease as others guanidinio compounds [25,26].

Most ADMA, but not SDMA, is degraded to Citr and dimethylamine (Figure 1a) by dimethylarginine dimethylaminohydrolase (DDAH), which is widely distributed throughout the body, particularly in the liver and kidney [27]. DDAH has two isoforms (I and II), DDAH I predominantly in tissue expressing nNOS and DDAH II in tissues containing eNOS [28]. The two isoforms are expressed differently depending on the tissue, and in this way regulate ADMA levels and hence NO synthesis [29]. For these reasons, reduced activity DDAH may be the main mechanism underlying an increase in ADMA levels.

3. ADMA as a Cardiovascular Risk Factor

Several studies have found that the plasma levels of ADMA in healthy subjects with no apparent cardiovascular disease are related to age, blood pressure, insulin resistance, and carotid intima-media thickness [30,31]. These findings suggest that an increase in ADMA levels is a marker for arteriosclerotic changes. For this reason, ADMA levels have been used for assessing cardiovascular risk [32,33]. Further, it is known that ADMA is mainly absorbed by endothelial cells, extracellular levels being 5 to 10 times lower. This means that slight changes in plasma levels of ADMA are sufficient to significantly change intracellular ADMA levels and, thereby, modify NO production and contribute to the development of cardiovascular disease. Under pathological conditions, there may be a 3- to 9-fold increase in plasma levels of ADMA, and this would inhibit NO production by 30% to 70% [34]. This explains why this metabolite can be used as a cardiovascular marker, as a decrease in NO production by the some of the endothelium leads to adverse vascular effects (Figure 3) such as dysregulation of blood pressure, prevention of vasodilation, loss of antithrombotic activity, problems in homeostasis and fibrinolysis, and inhibition of platelet aggregation, among others [35,36]. Given this, the effect of ADMA on the Arg-NO pathway is of particular importance in conditions associated with vascular wall damage, such as hypercholesterolaemia, high blood pressure, smoking, diabetes and obesity.
Cardiovascular disease is responsible for 70% of deaths in patients with type 2 diabetes [37], but the levels of the conventional markers for cardiovascular disease are not sufficiently elevated to explain the excess cardiovascular risk in these patients. Given this, the plasma levels of ADMA have been analyzed in individuals with glucose intolerance [38], insulin resistance [39] and diabetes [40], elevated levels of the metabolite being associated with reduced DDAH activity, throughout a polymorphism of the $DDAH2$ gene in case of type 2 diabetes [41]. Several authors have reported a direct relationship between high blood pressure and ADMA levels [42,43]. Further, the relationship between the two factors has already been demonstrated in healthy subjects, who were administered methylarginine in order to increase their ADMA concentration to higher than normal levels, and in patients with end-stage renal disease in whom elevated ADMA levels were directly associated with carotid atherosclerosis and cardiovascular mortality [44,45].

Elevated ADMA levels have also been linked to oxidative stress [46] and hypercholesterolaemia [47,48], through a reduction in DDAH activity. It has recently been confirmed that smoking increases the levels of ADMA and decreases those of Arg, and this may be explained by impairment of the y+ transport system by tobacco smoke [49].

ADMA levels are elevated in humans and other animals with heart failure, specifically in patients with dysfunction and left ventricular hypertrophy [50], and also in patients with chronic kidney failure [51,52]. More recently, reduced DDAH activity has also been observed in the coronary endothelium, smooth muscle and cardiac myocytes in an animal model of kidney failure [53]. Therefore, this metabolite could be used as a marker for kidney failure as well as for cardiovascular risk.

**Relationship between ADMA and Homocysteine (Role of the DDAH Enzyme)**

Various studies have failed to find a relationship between the levels of cardiovascular marker homocysteine (Hcys) and ADMA [54], both of which are elevated in paediatric patients after RTx. It is well known that Hcys causes endothelial dysfunction, and this could be due to the formation of disulphide bonds and generation of hydrogen peroxide and superoxide anions, which would increase...
the oxidative degradation of NO [55]. Hcy may also decrease NO synthesis through the degradation of NOS. A recent study demonstrated that an increase in Hcy causes a decrease in the activity of DDAH, which is able to degrade ADMA [56]. The resulting accumulation of ADMA impairs NOS function leading to a reduction in NO synthesis and accelerates the progression of atherosclerosis [46].

DDAH has four cysteine residues, whose sulfhydryl groups are of great importance for its activity, agents such as p-chloromercuribenzene acid and mercury chloride that block these sulfhydryl groups being potent inhibitors of the enzyme. Similarly, Hcy also has very reactive sulfhydryl groups that are responsible for the production of Hcy dimers and the formation of covalent bonds between Hcy and proteins. Therefore, it is quite possible that Hcy forms disulphide bonds to the DDAH enzyme, blocking the binding of ADMA to the active site of the enzyme and, thereby, slowing the degradation of the dimethylarginine molecule. Stühlinger et al. [46] indicated that the bonds between the Hcy and DDAH modify the spatial configuration of the enzyme, inhibiting its activity.

Further, the high reactivity of Hcy may cause the degradation of proteins, destabilizing their structure and increasing the level of oxidative stress. In this way, methylated arginine residues arising from the proteins are released giving rise to ADMA molecules. The increase in the plasma level of this methylated arginine is a better cardiovascular marker than Hcy: some authors have not found Hcy to be associated with cardiovascular events or carotid intima-media thickness in renal patients [57]; while, in contrast, analyzing the relationship between levels of ADMA and the survival of patients with chronic kidney disease using multivariate Cox models, elevated levels were found to be significantly associated with cardiovascular-related mortality [58].

Other studies have suggested that elevated levels of oxidized LDLs and lysophosphatidylcholine, their major component, lead to a reduction in DDAH activity thus increasing ADMA levels [59,60]. On the other hand, if LDL cholesterol levels are normal, this inhibitory mechanism can be ruled out as an explanation of the elevated ADMA levels in patients with renal dysfunction.

The fact that ADMA is correlated with few other variables, and in particular that it is not correlated with Hcy, reinforces the theory that ADMA is an independent cardiovascular biomarker [61]. In fact, it has been confirmed that the predictive cardiovascular value of ADMA is similar to that of B-type natriuretic peptide (a biomarker of heart failure) and C reactive protein (CRP, an inflammatory mediator). There are even predictive models based on these three markers, which provide greater predictive value for potential adverse cardiovascular events [62]. Higher concentrations of ADMA have also been found to be associated with a reduction in the urinary excretion of creatinine, and this illustrates the sense in which ADMA is a biomarker for kidney disease, levels being directly related to renal function [30,63].

Arginine acts as a natural precursor for NO production in the endothelium and other types of tissue. Accordingly, it would be interesting to compare the ratio between Arg and ADMA in patients with renal disease and a control group, as this value may indicate a potential problem with NO production. Some authors have already observed a reduction in the Arg/ADMA ratio in patients with renal disease, and this was associated with clinical manifestations of atherosclerosis and, accordingly, also with the regulation of processes such as blood pressure control, vasodilation, antithrombotic activity, homeostasis and fibrinolysis [64,65]. This ratio is of great importance for the absorption of Arg by cells and subsequent NO production, not only in renal tubular epithelial cells but also in endothelial cells.
4. Asymmetric Dimethylarginine in Kidney Disease

It has been observed that endothelial NO production is reduced in renal patients, in part due to limited synthesis of the substrate Arg but also because the levels of ADMA, a competitive inhibitor of NOS, are elevated [15]. These factors contribute to endothelial dysfunction, cardiovascular risk and the progression of renal damage.

ADMA accumulates in patients with chronic renal failure [64,66], inhibiting the activity of NOS, which may explain, at least to some extent, the high blood pressure and endothelial dysfunction seen in these patients. There have also been other observations that suggest the great biological importance of the activity of ADMA as a cause of endothelial dysfunction, reducing NO production via decreased eNOS phosphorylation [67,68]. In a recent review of the role of ADMA in chronic kidney disease [65], four different main mechanisms were identified to explain the accumulation of ADMA in this type of patient, and this analysis may be extrapolated to RTx [69]:

- Higher levels of protein methylation;
- Increased rate of protein turnover;
- Impaired activity of DDAH, which degrades ADMA;
- Impaired renal excretion.

It is worth noting that ADMA levels are elevated in patients with early phases of renal disease, even before glomerular filtration is significantly reduced [63]. This suggests that only a very small fraction of circulating ADMA is excreted through urine, although it is negatively correlated with GFR [32]. The importance of high levels of methylated arginine in renal disease is evidenced by the fact that patients have been diagnosed with chronic kidney disease despite having normal blood creatinine levels, and therefore GFR was not measured [70]. Some studies found lower DDAH activity in an animal model with kidney failure, showing increased levels of ADMA [53,71], these depending more on the activity of the enzyme that degrades it than on the GFR.

In some studies, elevated ADMA levels have not been found to be directly related to cardiovascular events either in the general population or in renal patients. This can be attributed to a state of malnutrition or an increase in inflammation together with oxidative stress [72]. In general, however, renal patients do have higher ADMA levels than control groups; accordingly, NO production is inhibited, contributing to endothelial dysfunction and cardiovascular risk in renal patients and limiting survival, given the association with heart disease [73,74]. Table 1 summarizes the values of Arg, ADMA and SDMA measured in control subjects and/or renal patients from various studies in recent years.
### Table 1. Concentrations of Arg and its methylated metabolites in plasma of healthy human subjects and patients with renal failure.

| Subjects                        | Age (Years) | ADMA (µM) | SDMA (µM) | Arg (µM) | Arg/ADMA | Method          |
|---------------------------------|-------------|-----------|-----------|----------|----------|----------------|
| Marescau et al. 1997 [75]       | Controls    | 23–86     | 0.41 ± 0.09 | 0.38 ± 0.10 | 110 ± 24 | LC-Fluorescence |
|                                 | CKD: CC > 40| 23–86     | 0.60 ± 0.10 | 0.83 ± 0.22 | 111 ± 24 | LC-Fluorescence |
|                                 | CKD: CC = 40–20 | 23–86 | 0.72 ± 0.22 | 1.41 ± 0.63 | 119 ± 33 | LC-Fluorescence |
|                                 | CKD: CC = 20–10 | 23–86 | 0.84 ± 0.15 | 2.24 ± 0.74 | 122 ± 36 | LC-Fluorescence |
|                                 | CKD: CC < 10 | 23–86     | 0.80 ± 0.14 | 3.17 ± 1.05 | 129 ± 37 | LC-Fluorescence |
| Pi et al. 2000 [76]             | Controls    | 23–35     | 0.30 ± 0.05 | 0.34 ± 0.06 | 60.7 ± 19.0 | LC-Fluorescence |
| Tsikas et al. 2003 [77]         | Controls    | 35.6 ± 11.4 | 0.39 ± 0.06 |          |          | GC-MS/MS       |
| Martens-Lobenhoffer et al. 2004 [78] | Controls | 20–56     | 0.36 ± 0.07 | 0.46 ± 0.09 | 63.9 ± 23.9 | LC-MS/MS       |
|                                 | CKD         | 36–78     | 0.67 ± 0.13 | 3.16 ± 0.91 | 48.1 ± 18.5 | LC-MS/MS       |
| Schwedhelm et al. 2005 [79]     | Controls    | >18       | 0.55 ± 0.14 | 0.69 ± 0.23 | 65.6 ± 23.4 | 132 ± 55       | LC-MS/MS       |
| Martens-Lobenhoffer et al. 2006 [80] | Controls | 22–32     | 0.37 ± 0.06 | 0.45 ± 0.06 | 60.6 ± 18.3 | LC-MS/MS       |
| Wilcken et al. 2006 [81]        | Controls    | 34.6 ± 11.7 | 0.49 ± 0.07 | 0.40 ± 0.07 | 87.9 ± 19.5 | 181.9 ± 56.1  | LC-Fluorescence |
|                                 | CBS         | 34.2 ± 12.6 | 0.55 ± 0.08 | 0.39 ± 0.09 | 73.5 ± 18.8 | 132.9 ± 24.7  | LC-Fluorescence |
| Bishop et al. 2007 [82]         | Controls    | >18       | 0.66 ± 0.12 |          | 87 ± 35 | 142 ± 81       | LC-MS/MS       |
| Schwedhelm et al. 2007 [83]     | Controls    | >18       | 0.46 ± 0.09 | 0.37 ± 0.07 | 74 ± 19 | 166 ± 50       | LC-MS/MS       |
| Weaving et al. 2008 [84]        | Controls    | 20.9 ± 2.5 | 0.40 ± 0.14 | 0.47 ± 0.06 | 162 ± 76 | SPE-MS/MS      |
| Zhang et al. 2009 [85]          | Controls    | 46.1 ± 13.2 | 0.49 ± 0.12 | 0.24 ± 0.08 |          | LC-Fluorescence |
|                                 | CKD         | 45.7 ± 14.2 | 2.36 ± 0.89 | 0.48 ± 0.11 |          | LC-Fluorescence |
|                                 | RTx         | 45.7 ± 14.2 | 0.70 ± 0.24 | 0.26 ± 0.07 |          | LC-Fluorescence |
| El-Khoury et al. 2012 [86]      | Controls    | 19–64     | 0.36–0.67 | 0.32–0.65 | 53.1–129.7 | LC-MS/MS       |

CBS: Cystathionine β-synthase deficiency; CKD: Chronic kidney disease; RTx: Renal transplant; CC: Creatinine clearance (mL/min).
In young people, the metabolic pathway involving ADMA has been little studied despite evidence that levels of this molecule are clinically relevant [87]. Among the few studies reported to date, elevated levels of ADMA have been detected in children and adolescents with high blood pressure [42], and its levels have started to be investigated in children with chronic renal failure [24,88], and in children and adolescents with hypercholesterolaemia [89], diabetes [90] and phenylketonuria [91]. Table 2 summarizes reported measurements of ADMA in the paediatric population: levels are higher than in adults due to the immaturity of the enzyme system, affecting DDAH activity in children and adolescents.

Table 2. Concentrations of Arg and its methylated metabolites in plasma of children and adolescents.

| Subjects          | Age (Years) | ADMA (µM) | SDMA (µM) | Arg (µM) | Arg/ADMA | Method |
|-------------------|-------------|-----------|-----------|----------|----------|--------|
| Wang et al. 2007  | Controls    | 12.6 ± 1.0| 0.78 ± 0.16| 0.71 ± 0.23| 65.3 ± 21.3| LC-MS/MS |
| Brooks et al. 2009| Controls    | 11.3 ± 4.7| 1.10 ± 0.35| 2.06 ± 1.11| 57.9 ± 22.1| LC-MS/MS |
| Andrade et al. 2011| Controls  | 7–18      | 0.41–0.96  | 2.1 ± 1.1 | 57.9 ± 22.1| 62.4 ± 27.7| LC-MS/MS |
| Huemer et al. 2012 | Controls   | 11.6 ± 3.7| 0.64 ± 0.66| 59 ± 59  | 59 ± 91  | ELISA  |

CKD: Chronic kidney disease; RTx: Renal transplant.

4.1. Asymmetric Dimethylarginine and Proteinuria

Proteinuria and also microalbuminaemia are potent and independent markers of cardiovascular risk, and there is evidence of their close relationship with endothelial dysfunction and inhibition of NO production [93]. Given that ADMA has also been associated with endothelial dysfunction in animal models and patients with chronic kidney disease [94], it is conceivable that the development of proteinuria may have a role in the accumulation of ADMA via the overexpression of PRMT that facilitates the synthesis of ADMA [95]. In line with this, positive correlations have been observed between ADMA and proteinuria [96] and, in an animal model, some researchers have observed a decrease in the activity of the DDAH enzyme [97], which is abundantly expressed in the renal tubular cells.

4.2. Asymmetric Dimethylarginine after Renal Transplantation

So far, there have been very few studies of ADMA levels in patients undergoing RTx. This is despite the fact that, given the association with cardiovascular risk, it is a metabolite of great interest in this group of patients. Yilmaz et al. [98] reported that, during their study follow-up period of four weeks, RTx greatly decreased ADMA levels, compared to before transplantation. Other studies have also highlighted that ADMA levels decrease during the first month after transplantation, but they remain elevated compared to a control population [85]. This fall results in an increase in NO synthesis and associated improvement in endothelial function, NO synthesis being closely associated with atherosclerosis in chronic kidney disease [99]. The fact that patients start immunosuppressive therapy does not seem to affect the improvement in endothelial function. This study also indicated that the levels of adiponectin, inflammatory and anti-atherogenic hormone are not correlated with the levels of
ADMA or CRP before and after RTx, despite all the values corresponding to these variables having decreased after transplantation. Thus, as well as poor renal function, other factors may contribute to increasing levels of adiponectin in patients with chronic kidney disease. Further, the decrease in the inflammatory state after RTx may mask a correlation between GFR and adiponectin values.

The levels of the metabolites considered in this review must be analyzed taking into account that if the conversion of Arg into guanidinoacetate is impaired, the accumulation of Arg and its asymmetric derivatives may compromise NOS function and, in turn, have a negative effect on endothelial function; this would enhance the cardiovascular risk inherent to the use of immunosuppressive agents, increasing mortality and morbidity in these patients.

The mechanisms by which plasma ADMA levels may increase include an increase in the methylation of proteins and/or in proteolysis, and impairment of renal excretion of ADMA and/or of DDAH activity. In a patient population, it is relatively important to assess whether methylation activity is significantly enhanced (S-adenosylmethionine/S-adenosylhomocysteine ratio) in order to relate this to an increased methylation of proteins. In a recent study, we observed a significant increase in ADMA levels and a decrease in methylation activity in transplant recipients compared to a control population [92]. Other authors have suggested that obesity after RTx may be directly associated with elevated ADMA levels and a decrease in adiponectin, increasing cardiovascular risk [100,101]. Moreover, ADMA was found to be a significant risk factor for graft failure, so these plasma levels could predict morbidity, mortality and the deterioration of graft function RTx [102].

5. Treatments to Reduce ADMA Levels

Oxidative stress may increase the synthesis of ADMA, by stimulating the expression of methyltransferases in the body or reducing DDAH activity. Specific ADMA-lowering therapies are not yet available, but different dialysis models have different clearance capability on plasma ADMA [103], even angiotensin converting enzyme inhibitors and angiotensin receptor blockers may help to improve endothelial function and to prevent ADMA-induced pathology [104]. On the other hand, treatment with antioxidants of cells with very elevated intracellular ADMA levels restores the activity of DDAH. Given this, several studies have been initiated focusing on the use of antioxidants, such as tea and vegetable consumption [105], to re-establish endothelial production of NO.

One of these treatments includes the use of vitamin E that reduces lipid peroxidation and improves NO production, inhibiting the production of free radicals. Saran et al. [106] observed a 23% reduction in ADMA levels in patients with non-diabetic chronic kidney disease, after a period of eight weeks on supplementation with vitamin E. This confirms the success of this strategy of using antioxidants to reduce ADMA levels, increasing DDAH activity and reducing the levels of free radicals. The antioxidant power of vitamin E seems not to be strong enough to change the levels of Hcys and CRP, at least not with the doses and duration of treatment in the aforementioned study. Further, vitamin E hardly reduces ADMA levels at all in patients with high cardiovascular risk but, given its antioxidant nature, does normalize the levels in patients with chronic kidney disease and hyperhomocysteinaemia, in which there is elevated oxidative stress [107].

Impaired renal function and the presence of proteinuria or microalbuminuric patients are commonly associated with endothelial dysfunction in patients with chronic kidney disease, in whom levels of
ADMA are elevated and positively correlated with the reduction in renal function. For this reason, Teplan et al. [108] explored treating obese patients with chronic kidney disease with a low protein diet supplemented with keto-amino acids to maintain the nitrogen balance and prevent malnutrition. This type of treatment achieved a reduction in proteinuria, stabilization of renal function and a decrease in ADMA levels in these patients, methylarginine mainly coming from the proteolysis of proteins. Other studies suggest that a soy protein-rich diet can contribute to improving endothelial function by normalizing the Arg/ADMA ratio in kidney transplant recipients [109] using isoflavones, this treatment being easy to implement given that it just involves consumption soy protein in place of some animal protein.

New therapeutic approaches are currently being developed, in particular, based on the administration of intravenous $N$-acetylcysteine. This treatment not only lowers Hcys levels, but also reduces blood pressure and improves endothelial function [110], as well as decreasing ADMA levels in patients on haemodialysis [111]. $N$-acetylcysteine breaks disulphide bonds; the reason why this thiol has a direct antioxidant effect, decreasing the levels of oxygen reactive species and increasing the bioavailability of DDAH, is that it releases the enzyme from any molecule binding to it, such as Hcys, thus re-establishing normal enzymatic activity. In this way, it is possible to decrease levels of ADMA, as this molecule will be degraded to amine groups and Citr.

5.1. Statins

Given the high cardiovascular risk associated with elevated levels of ADMA and Hcys, it is logical to consider possibility of treating such patients with statins: in the long term, these agents having been found to improve the oxidative status of adult renal transplant patients and even increase their levels of HDL cholesterol, which has great antioxidant properties [112].

It is known that statins significantly reduce the number of episodes of acute transplant rejection, while improving graft survival and renal function. These effects are correlated with a decrease in the levels of triglycerides and a blocking of the synthesis of cholesterol and non-steroidal compounds derived from mevalonate, an early precursor of cholesterol. Further, statins may have an additional immunosuppressive effect through the inhibition of interleukin-6, in this way altering the initial stages of T-cell activation and, thereby, interrupting cellular rejection. This effect is also associated with a decrease in the levels of triglycerides mediated by interleukin-6 [113].

Initial studies in patients with chronic renal insufficiency have indicated that treatment with statins does not reduce ADMA levels to the normal range [107,114], although they do have a cardiovascular protective effect. While some studies have associated ADMA with LDL cholesterol [59,60], it seems that only certain types of statins reduce plasma levels of ADMA, since the vascular effects of these agents depend on the expression of the regulatory gene NOS [35]. Specifically, it has been reported that rosuvastatin contributes to reducing ADMA levels in children [47].

On the other hand, there are various concerns with respect to the use of statins given their most common adverse effects including fatigue, gastrointestinal problems including constipation and cramps. Accordingly, alternatives have started to be tested such as the effect on endothelial function of kidney transplanted patients of the consumption of fish rich in long-chain omega-3 fatty acids, which may also have a protective effect against the nephrotoxicity of some immunosuppressive agents [115].
While the dose-response relationship for omega-3 fatty acid supplementation remains to be clarified, it has already been demonstrated that this treatment is not associated with Hcys levels in patients with chronic kidney disease [116]. Similarly, the administration of statins to kidney transplant patients did not have an impact on plasma levels of Hcys, although it does manage to reduce cardiovascular risk by decreasing the levels of total and LDL cholesterol and triglycerides, without affecting the levels of HDL cholesterol [117]. For this reason, in the case of children and adolescents with renal insufficiency or those subject to transplantation, there are more doubts over the usefulness of statin treatment if the levels of triglycerides, total cholesterol and its subfractions are within normal ranges. In particular, if these patients have hyperhomocysteinaemia, the treatment would not reduce Hcys levels given that there is no association between the levels of Hcys and statins.

5.2. Arginine

In recent years, it has been demonstrated that Arg supplementation restores NO production, improves renal function and reduces inflammation in kidney transplant patients [118]. A key objective in renal patients is to decrease ADMA levels [119]. For this reason, arginine supplementation has been used, reversing ADMA-induced inhibition of NO synthesis. According to several studies, arginine also improves endothelial dysfunction in patients on haemodialysis by regulating ADMA levels [120]; that is, the inhibitory action of ADMA on NOS is reverted by arginine loading.

Arginine supplementation may increase the production of guanidinoacetate and creatine, through the conversion of S-adenosylmethionine to S-adenosylhomocysteine and Hcys, which may themselves have adverse effects on endothelial function [121]. In kidney transplant recipients, however, in whom there is already endothelial dysfunction for various reasons, supplementation with arginine contributes to improving renal function, normalizing the Arg/ADMA ratio, increasing NO production, and eliminating nitrites and nitrates, which improves endothelial function [122]. It has even been demonstrated that the intake of Citr may have similar effects since it is converted into Arg, with no adverse effects and with a good tolerance [123].

On the other hand, in patients with high plasma Arg and Citr levels, the efficacy of supplementation of this amino acid has been questioned due to its potential adverse effects. It should also be taken into account that Arg is an acidifying agent that may alter the acid-base equilibrium in patients with reduced GFRs; accordingly, in such cases, it should be administered in the form of arginine salts other than chlorates. This may explain why data on the effects of the arginine supplementation are inconsistent, some studies having found no significant beneficial effects on vascular function and others finding no differences in endothelial NO synthesis with respect to placebo [32].

6. Conclusions

Kidney disease is directly associated with elevated levels of the cardiovascular marker ADMA and a significant reduction in the Arg/ADMA ratio, such that NO synthesis is inhibited, worsening renal dysfunction and increasing cardiovascular risk.

The lack of correlation between ADMA and other variables strengthens the theory that it is an independent marker of cardiovascular risk. It has also been observed that ADMA levels tend to be higher in children, due to immaturity of the enzyme system, affecting DDAH activity.
Treatments to reduce ADMA levels may be useful in patients with kidney disease, but decisions should be taken on a case-by-case basis, including a complete analysis of the corresponding metabolic pathways and lipid profiles.

Acknowledgements

This work was financially supported by Grants from the Department of Health of Basque Government and Carlos III Spanish Health Institute. We are also grateful to the Basque Foundation for Health Innovation and Research for its support.

References

1. Clarke, R.; Daly, L.; Robinson, K.; Naughten, E.; Cahalane, S.; Fowler, B.; Graham, I. Hyperhomocysteinemia: An independent risk factor for vascular disease. *N. Engl. J. Med.* 1991, 324, 1149–1155.

2. Aldámiz-Echevarría, L.; Sanjurjo, P.; Vallo, A.; Aquino, L.; Perez-Nanclares, G.; Gimeno, P.; Rueda, M.; Ruiz, I.; Urreizti, R.; Rodriguez-Soriano, J. Hyperhomocysteinemia in children with renal transplants. *Pediatr. Nephrol.* 2002, 17, 718–723.

3. Nitsch, D.D.; Ghilardi, N.; Muhl, H.; Nitsch, C.; Brune, B.; Pfeilschifter, J. Apoptosis and expression of inducible nitric oxide synthase are mutually exclusive in renal mesangial cell. *Am. J. Pathol.* 1997, 150, 889–900.

4. Tojo, A.; Welch, W.J.; Bremen, V.; Kimoto, M.; Kimura, K.; Omata, M.; Ogawa, T.; Vallance, P.; Wilcox, C.S. Colocalization of demethylating enzymes and NOS and functional effects of methylarginines in rat kidney. *Kidney Int.* 1997, 52, 1593–1601.

5. Holmqvist, B.; Olsson, C.F.; Svensson, M.L.; Svanborg, C.; Forsell, J.; Alm, P. Expression of nitric oxide synthase isoforms in the mouse kidney: Cellular localization and influence by lipopolysaccharide and Toll-like receptor 4. *J. Mol. Histol.* 2005, 36, 499–516.

6. Morrissey, J.J.; McCracken, R.; Kaneto, H.; Vehaskari, M.; Montani, D.; Klahr, S. Location of an inducible nitric oxide synthase mRNA in the normal kidney. *Kidney Int.* 1994, 45, 998–1005.

7. Raij, P.; Shultz, P.J. Endothelium-Derived relaxing factor, nitric oxide: Effects on and production by mesangial cells and glomerulus. *J. Am. Soc. Nephrol.* 1993, 3, 1435–1441.

8. Perticone, F.; Maio, R.; Perticone, M.; Sciacqua, A.; Shehaj, E.; Naccarato, P.; Sesti, G. Endothelial dysfunction and subsequent decline in glomerular filtration rate in hypertensive patients. *Circulation* 2010, 122, 379–384.

9. Crespo, M.; Campistol, J.M. Disfunción endotelial en transplante renal. *Nefrología* 2003, 23, 52–58.

10. Caló, L.A.; Dall’Amico, R.; Pagnin, E.; Bertipaglia, L.; Zacchello, G.; Davis, P.A. Oxidative stress and post-transplant hypertension in pediatric kidney-transplanted patients. *J. Pediatr.* 2006, 149, 53–57.

11. Sahin, G.; Akay, O.M.; Bal, C.; Yalcin, A.U.; Gulbas, Z. The effect of calcineurin inhibitors on endothelial and platelet function in renal transplant patients. *Clin. Nephrol.* 2011, 76, 218–225.

12. Trapp, A.; Weis, M. The impact of immunosuppression on endothelial function. *J. Cardiovasc. Pharmacol.* 2005, 45, 81–87.
13. Morris, S.T.; McMurray, J.J.V.; Rodger, R.S.; Farmer, R.; Jardine, A.G. Endothelial dysfunction in renal transplant recipients maintained on cyclosporine. *Kidney Int.* 2000, 57, 1100–1106.

14. Schmidt, R.J.; Yokota, S.; Tracy, T.S.; Sorkin, M.I.; Baylis, C. Nitric oxide production is low in end-stage renal disease patients on peritoneal dialysis. *Am. J. Physiol.* 1999, 276, 794–797.

15. Loscalzo, J. L-Arginine and atherothrombosis. *J. Nutr.* 2004, 134, 2798S–2800S.

16. Vallance, P.; Leiper, J. Cardiovascular biology of the asymmetric dimethylarginine: Dimethylarginine dimethylaminohydrolase pathway. *Arterioscler. Thromb. Vasc. Biol.* 2004, 24, 1023–1030.

17. Böger, R.H.; Sydow, K.; Borlak, J.; Thum, T.; Lenzen, H.; Schubert, B.; Tsikas, D.; Bode-Böger, S.M. LDL cholesterol upregulates synthesis of asymmetrical dimethylarginine in human endothelial cells: Involvement of S-adenosylmethionine-dependent methyltransferases. *Circ. Res.* 2000, 87, 99–105.

18. Tojo, A.; Welch, W.J.; Bremer, V.; Kimoto, M.; Kimura, K.; Omata, M.; Ogawa, T.; Vallance, P.; Wilcox, C.S. Colocalization of demethylating enzymes and NOS and functional effects of methylarginines in rat kidney. *Kidney Int.* 1997, 52, 1593–1601.

19. Kielstein, J.T.; Salpeter, S.R.; Bode-Boeger, S.M.; Cooke, J.P.; Fliser, D. Symmetric dimethylarginine (SDMA) as endogenous marker of renal function—A meta-analysis. *Nephrol. Dial. Transpl.* 2006, 21, 2446–2451.

20. Kielstein, J.T.; Zoccali, C. Asymmetric dimethylarginine: A novel marker of risk and a potential target for therapy in chronic kidney disease. *Curr. Opin. Nephrol. Hypertens.* 2008, 17, 609–615.

21. Kiechl, S.; Lee, T.; Santer, P.; Thompson, G.; Tsimikas, S.; Egger, G.; Holt, D.W.; Willeit, J.; Xu, Q.; Mayr, M. Asymmetric and symmetric dimethylarginines are of similar predictive value for cardiovascular risk in the general population. *Atherosclerosis* 2009, 205, 261–265.

22. Oner-Iyidogan, Y.; Oner, P.; Kocak, H.; Gurdol, F.; Bekpinar, S.; Unlucerci, Y.; Caliskan, Y.; Cetinalp-Demircan, P.; Kocak, T.; Turkmen, A. Dimethylarginines and inflammation markers in patients with chronic kidney disease undergoing dialysis. *Clin. Exp. Med.* 2009, 9, 235–241.

23. Schepers, E.; Barreto, D.V.; Liaubeuf, S.; Glorieux, G.; Eloit, S.; Barreto, F.C.; Massy, Z.; Vanholder, R. European Uremic Toxin Work Group (EUTox). Symmetric dimethylarginine as a proinflammatory agent in chronic kidney disease. *Clin. J. Am. Soc. Nephrol.* 2011, 6, 2374–2383.

24. Brooks, E.R.; Langman, C.B.; Wang, S.; Price, H.E.; Hodges, A.L.; Darling, L.; Yang, A.Z.; Smith, F.A. Methylated arginine derivatives in children and adolescents with chronic kidney disease. *Pediatr. Nephrol.* 2009, 24, 129–134.

25. Bode-Böger, S.M.; Scalera, F.; Kielstein, J.T.; Martens-Lobenhoffer, J.; Breithardt, G.; Fobker, M.; Reinecke, H. Symmetrical dimethylarginine: A new combined parameter for renal function and extent of coronary artery disease. *J. Am. Soc. Nephrol.* 2006, 17, 1128–1134.

26. Schepers, E.; Glorieux, G.; Dou, L.; Cerini, C.; Gaynard, N.; Louvet, L.; Maugard, C.; Preus, P.; Rodriguez-Ortiz, M.; Argiles, A.; et al. European Uremic Toxin Work Group (EUTox). Guanidino compounds as cause of cardiovascular damage in chronic kidney disease: An in vitro evaluation. *Blood Purif.* 2010, 30, 277–287.
27. Nijveldt, R.J.; Teerlink, T.; van Guldener, C.; Prins, H.A.; van Lambalgen, A.A.; Stehouwer, C.D.; Rauwerda, J.A.; van Leeuwen, P.A. Handling of asymmetrical dimethylarginine and symmetrical dimethylarginine by the rat kidney under basal conditions and during endotoxaemia. *Nephrol. Dial. Transpl.* **2003**, *18*, 2542–2550.

28. Leiper, J.M.; Santa Maria, J.; Chubb, A.; MacAllister, R.J.; Charles, I.G.; Whitley, G.S.; Vallance, P. Identification of two human dimethylarginine dimethylaminohydrolases with distinct tissue distributions and homology with microbial arginine deiminases. *Biochem. J.* **1999**, *343*, 209–214.

29. Torondel, B.; Nandi, M.; Kelly, P.; Wojciak-Stothard, B.; Fleming, I.; Leiper, J. Adenoviral-Mediated overexpression of DDAH improves vascular tone regulation. *Vasc. Med.* **2010**, *15*, 205–213.

30. Sydow, K.; Fortmann, S.P.; Fair, J.M.; Varady, A.; Hlatky, M.A.; Go, A.S.; Iribarren, C.; Tsao, P.S. ADVANCE Investigators. Distribution of asymmetric dimethylarginine among 980 healthy, older adults of different ethnicities. *Clin. Chem.* **2010**, *56*, 111–120.

31. Perticone, F.; Sciacqua, A.; Maio, R.; Perticone, M.; Galiano Leone, G.; Bruni, R.; Di Cello, S.; Pascale, A.; Talarico, G.; Greco, L.; *et al.* Endothelial dysfunction, ADMA and insulin resistance in essential hypertension. *Int. J. Cardiol.* **2010**, *142*, 236–241.

32. Wilcken, D.E.; Sim, A.S.; Wang, J.; Wang, X.L. Asymmetric dimethylarginine (ADMA) in vascular, renal and hepatic disease and the regulatory role of L-arginine on its metabolism. *Mol. Genet. Metab.* **2007**, *91*, 309–317.

33. Sibal, L.; Agarwal, S.C.; Home, P.D.; Boger, R.H. The role of asymmetric dimethylarginine (adma) in endothelial dysfunction and cardiovascular disease. *Curr. Cardiol. Rev.* **2010**, *6*, 82–90.

34. Cardounel, A.J.; Cui, H.; Samouilov, A.; Johnson, W.; Kearns, P.; Tsai, A.L.; Berka, V.; Zweier, J.L. Evidence for the pathophysiological role of endogenous methylarginines in regulation of endothelial NO production and vascular function. *J. Biol. Chem.* **2007**, *282*, 879–887.

35. Böger, R.H. Asymmetric dimethylarginine (ADMA): A novel risk marker in cardiovascular medicine and beyond. *Ann. Med.* **2006**, *38*, 126–136.

36. Bermúdez, V.; Bermúdez, F.; Acosta, G.; Acosta, A.; Añez, J.; Andara, C.; Leal, E.; Cano, C.; Manuel, V.; Hernández, R.; *et al.* Molecular mechanisms of endothelial dysfunction: From nitric oxide synthesis to ADMA inhibition. *Am. J. Ther.* **2008**, *15*, 326–333.

37. Laakso, M. Hyperglycemia and cardiovascular disease in type 2 diabetes. *Diabetes* **1999**, *48*, 937–942.

38. Miyazaki, H.; Matsuoka, H.; Cooke, J.P.; Usui, M.; Ueda, S.; Okuda, S.; Imaizumi, T. Endogenous nitric oxide synthase inhibitor: A novel marker of atherosclerosis. *J. Cardiol.* **1999**, *33*, 105–106.

39. Stühlinger, M.C.; Abbasi, F.; Chu, J.W.; Lamendola, C.; McLaughlin, T.L.; Cooke, J.P.; Reaven, G.M.; Tsao, P.S. Relationship between insulin resistance and an endogenous nitric oxide synthase inhibitor. *JAMA J. Am. Med. Assoc.* **2002**, *287*, 1420–1426.
40. Tarnow, L.; Hovind, P.; Teerlink, T.; Stehouwer, C.D.; Parving, H.H. Elevated plasma asymmetric dimethylarginine as a marker of cardiovascular morbidity in early diabetic nephropathy in type 1 diabetes. *Diabetes Care* **2004**, *27*, 765–769.

41. Andreozzi, F.; Presta, I.; Mannino, G.C.; Scarpelli, D.; Di Silvestre, S.; Di Pietro, N.; Succurro, E.; Sciacquar, A.; Pandolfi, A.; Consoli, A.; *et al*. A functional variant of the dimethylarginine dimethylaminohydrase-2 gene is associated with insulin sensitivity. *PLoS One* **2012**, *7*, e36224.

42. Goonasekera, C.D.; Rees, D.D.; Woolard, P.; Frend, A.; Shah, V.; Dillon, M.J. Nitric oxide synthase inhibitors and hypertension in children and adolescents. *J. Hypertens.* **1997**, *15*, 901–909.

43. Ueda, S.; Yamagishi, S.; Kaida, Y.; Okuda, S. Asymmetric dimethylarginine may be a missing link between cardiovascular disease and chronic kidney disease. *Nephrology* **2007**, *12*, 582–590.

44. Mallamaci, F.; Zoccali, C. Clinical implications of elevated asymmetric dimethylarginine in chronic kidney disease and end-stage renal disease. *J. Ren. Nutr.* **2009**, *19*, 25–28.

45. Alsagaff, M.Y.; Thaha, M.; Aminuddin, M.; Yogiarto, R.M.; Yogiantoro, M.; Tomino, Y. Asymmetric dimethylarginine: A novel cardiovascular risk factor in end-stage renal disease. *J. Int. Med. Res.* **2012**, *40*, 340–349.

46. Stühlinger, M.C.; Tsao, P.S.; Her, J.H.; Kimoto, M.; Balint, R.F.; Cooke, J.P. Homocysteine impairs the nitric oxide synthase pathway: Role of asymmetric dimethylarginine. *Circulation* **2001**, *104*, 2569–2575.

47. Lu, T.M.; Ding, Y.A.; Leu, H.B.; Yin, W.H.; Sheu, W.H.; Chu, K.M. Effect of rosuvastatin on plasma levels of asymmetric dimethylarginine in patients with hypercholesterolemia. *Am. J. Cardiol.* **2004**, *94*, 157–161.

48. Hasanoğlu, A.; Okur, I.; Oren, A.C.; Bıberoğlu, G.; Oktar, S.; Eminoğlu, F.T.; Tümer, L. The levels of asymmetric dimethylarginine, homocysteine and carotid intima-media thickness in hypercholesterolemic children. *Turk. J. Pediatr.* **2011**, *53*, 522–527.

49. Zhang, W.Z.; Venardos, K.; Chin-Dusting, J.; Kaye, D.M. Adverse effects of cigarette smoke on NO bioavailability: Role of arginine metabolism and oxidative stress. *Hypertension* **2006**, *48*, 278–485.

50. Napora, M.; Graczykowska, A.; Próchniewska, K.; Zdrojewski, Z.; Całka, A.; Górny, J.; Stompór, T. Relationship between serum asymmetric dimethylarginine and left ventricular structure and function in patients with end-stage renal disease treated with hemodialysis. *Pol. Arch. Med. Wewn.* **2012**, *122*, 226–234.

51. Zoccali, C.; Mallamaci, F.; Maas, R.; Benedetto, F.A.; Tripepi, G.; Malatino, L.S.; Cataliotti, A.; Bellanuova, I.; Böger, R. Left ventricular hypertrophy, cardiac remodeling and asymmetric dimethylarginine (ADMA) in hemodialysis patients. *Kidney Int.* **2002**, *62*, 339–345.

52. Shi, B.; Ni, Z.; Zhou, W.; Yu, Z.; Gu, L.; Mou, S.; Fang, W.; Wang, Q.; Cao, L.; Yan, Y.; *et al*. Circulating levels of asymmetric dimethylarginine are an independent risk factor for left ventricular hypertrophy and predict cardiovascular events in pre-dialysis patients with chronic kidney disease. *Eur. J. Intern. Med.* **2010**, *21*, 444–448.
53. Chen, Y.; Li, Y.; Zhang, P.; Traverse, J.H.; Hou, M.; Xu, X.; Kimoto, M.; Bache, R.J. Dimethylarginine dimethylaminohydrolase and endothelial dysfunction in failing hearts. *Am. J. Physiol. Heart. Circ. Physiol.* 2005, 289, 2212–2219.

54. Notsu, Y.; Nabika, T.; Bokura, H.; Suyama, Y.; Kobayashi, S.; Yamaguchi, S.; Masuda, J. Evaluation of asymmetric dimethylarginine and homocysteine in microangiopathy-related cerebral damage. *Am. J. Hypertens.* 2009, 22, 257–262.

55. Lang, D.; Kredan, M.B.; Moat, S.J.; Hussain, S.A.; Powell, C.A.; Bellamy, M.F.; Powers, H.J.; Lewis, M.J. Homocysteine-induced inhibition of endothelium-dependent relaxation in rabbit aorta: Role for superoxide anions. *Arterioscler. Thromb. Vasc. Biol.* 2000, 20, 422–427.

56. Dayal, S.; Rodionov, R.N.; Arning, E.; Bottiglieri, T.; Kimoto, M.; Murry, D.J.; Cooke, J.P.; Faraci, F.M.; Lentz, S.R. Tissue-Specific downregulation of dimethylarginine dimethylaminohydrolase in hyperhomocysteinemia. *Am. J. Physiol. Heart Circ. Physiol.* 2008, 295, 816–825.

57. Van Guldener, C.; Nanayakkara, P.W.; Stehouwer, C.D.A. Homocysteine and asymmetric dimethylarginine (ADMA): Biochemically linked but differently related to vascular disease in chronic kidney disease. *Clin. Chem. Lab. Med.* 2007, 45, 1683–1687.

58. Young, J.M.; Terrin, N.; Wang, X.; Greene, T.; Beck, G.J.; Kusek, J.W.; Collins, A.J.; Sarnak, M.J.; Menon, V. Asymmetric dimethylarginine and mortality in stages 3 to 4 chronic kidney disease. *Clin. J. Am. Soc. Nephrol.* 2009, 4, 1115–1120.

59. Ito, A.; Tsao, P.S.; Adimoolam, S.; Kimoto, M.; Ogawa, T.; Cooke, J.P. Novel mechanism for endothelial dysfunction: Dysregulation of dimethylarginine dimethylaminohydrolase. *Circulation* 1999, 99, 3092–3095.

60. Feng, M.; Liu, L.; Guo, Z.; Xiong, Y. Gene transfer of dimethylarginine dimethylaminohydrolase-2 improves the impairments of DDAH/ADMA/NOS/NO pathway in endothelial cells induced by lysophosphatidylcholine. *Eur. J. Pharmacol.* 2008, 584, 49–56.

61. Rocha, M.S.; Teerlink, T.; Janssen, M.C.; Kluijtmans, L.A.; Smulders, Y.; Jakobs, C.; Tavares de Almeida, I.; Rivera, I.; Castro, R.; Blom, H.J. Asymmetric dimethylarginine in adults with cystathionine β-synthase deficiency. *Atherosclerosis* 2012, 222, 509–511.

62. Schnabel, R.; Blankenberg, S.; Lubos, E.; Lackner, K.J.; Rupprecht, H.J.; Espinola-Klein, C.; Jachmann, N.; Post, F.; Peetz, D.; Bickel, C.; et al. Asymmetric dimethylarginine and the risk of cardiovascular events and death in patients with coronary artery disease: Results from the AtheroGene Study. *Circ. Res.* 2005, 97, 53–59.

63. Kielstein, J.T.; Böger, R.H.; Bode-Böger, S.M.; Frölich, J.C.; Haller, H.; Ritz, E.; Fliser, D. Marked increase of asymmetric dimethylarginine in patients with incipient primary chronic renal disease. *J. Am. Soc. Nephrol.* 2002, 13, 170–176.

64. Kielstein, J.T.; Böger, R.H.; Bode-Böger, S.M.; Schäffer, J.; Barbey, M.; Koch, K.M.; Frölich, J.C. Asymmetric dimethylarginine plasma concentrations differ in patients with end-stage renal disease: Relationship to treatment method and atherosclerotic disease. *J. Am. Soc. Nephrol.* 1999, 10, 594–600.

65. Jacobi, J.; Tsao, P.S. Asymmetrical dimethylarginine in renal disease: Limits of variation or variation limits? A systematic review. *Am. J. Nephrol.* 2008, 28, 224–237.
66. Tripepi, G.; Mattace Raso, F.; Sijbrands, E.; Seck, M.S.; Maas, R.; Boger, R.; Witteman, J.; Rapisarda, F.; Malatino, L.; Mallamaci, F.; et al. Inflammation and asymmetric dimethylarginine for predicting death and cardiovascular events in ESRD patients. *Clin. J. Am. Soc. Nephrol.* **2011**, *6*, 1714–1721.

67. Dobrian, A.D. ADMA and NOS regulation in chronic renal disease: Beyond the old rivalry for L-arginine. *Kidney Int.* **2012**, *81*, 722–724.

68. Kajimoto, H.; Kai, H.; Aoki, H.; Yasuoka, S.; Anegawa, T.; Aoki, Y.; Ueda, S.; Okuda, S.; Imaizumi, T. Inhibition of eNOS phosphorylation mediates endothelial dysfunction in renal failure: New effect of asymmetric dimethylarginine. *Kidney Int.* **2012**, *81*, 762–768.

69. Zoccali, C.; Kielstein, J.T. Asymmetric dimethylarginine: A new player in the pathogenesis of renal disease? *Curr. Opin. Nephrol. Hypertens.* **2006**, *15*, 314–320.

70. Kielstein, J.T.; Martens-Lobenhoffer, J.; Vollmer, S.; Bode-Böger, S.M. L-Arginine, ADMA, SDMA, creatinine, MDRD formula: De tour to renal function testing. *J. Nephrol.* **2008**, *21*, 959–961.

71. Chen, G.F.; Moningka, N.C.; Sasser, J.M.; Zharikov, S.; Cunningham, M., Jr.; Tain, Y.L.; Schwartz, I.F.; Baylis, C. Arginine and asymmetric dimethylarginine in puromycin aminonucleoside-induced chronic kidney disease in the rat. *Am. J. Nephrol.* **2012**, *35*, 40–48.

72. Busch, M.; Fleck, C.; Wolf, G.; Stein, G. Asymmetrical (ADMA) and symmetrical dimethylarginine (SDMA) as potential risk factors for cardiovascular and renal outcome in chronic kidney disease—Possible candidates for paradoxical epidemiology? *Amino Acids* **2006**, *30*, 225–232.

73. Muscheites, J.; Meyer, A.A.; Drueckler, E.; Wigger, M.; Fischer, D.C.; Kundt, G.; Kienast, W.; Haffner, D. Assessment of the cardiovascular system in pediatric chronic kidney disease: A pilot study. *Pediatr. Nephrol.* **2008**, *23*, 2233–2239.

74. Lu, T.M.; Chung, M.Y.; Lin, C.C.; Hsu, C.P.; Lin, S.J. Asymmetric dimethylarginine and clinical outcomes in chronic kidney disease. *Clin. J. Am. Soc. Nephrol.* **2011**, *6*, 1566–1572.

75. Marescau, B.; Nagels, G.; Possemiers, I.; de Broe, M.E.; Because, I.; Billiouw, J.M.; Lornoy, W.; de Deyn, P.P. Guanidino compounds in serum and urine of nondialyzed patients with chronic renal insufficiency. *Metabolism* **1997**, *46*, 1024–1031.

76. Pi, J.; Kumagai, Y.; Sun, G.; Shimojo, N. Improved method for simultaneous determination of L-arginine and its mono- and dimethylated metabolites in biological samples by high-performance liquid chromatography. *J. Chromatogr. B* **2000**, *742*, 199–203.

77. Tsikas, D.; Schubert, B.; Gutzki, F.M.; Sandmann, J.; Frölich, J.C. Quantitative determination of circulating and urinary asymmetric dimethylarginine (ADMA) in humans by gas chromatography-tandem mass spectrometry as methyl ester tri(N-pentafluoropropionyl) derivative. *J. Chromatogr. B* **2003**, *798*, 87–99.

78. Martens-Lobenhoffer, J.; Krug, O.; Bode-Böger, S.M. Determination of arginine and asymmetric dimethylarginine (ADMA) in human plasma by liquid chromatography/mass spectrometry with the isotope dilution technique. *J. Mass. Spectrom.* **2004**, *39*, 1287–1294.

79. Schwedhelm, E.; Tan-Andresen, J.; Maas, R.; Riederer, U.; Schulze, F.; Böger, R.H. Liquid chromatography-tandem mass spectrometry method for the analysis of asymmetric dimethylarginine in human plasma. *Clin. Chem.* **2005**, *51*, 1268–1271.
80. Martens-Lobenhoffer, J.; Krug, O.; Bode-Böger, S.M. Fast and efficient determination of arginine, symmetric dimethylarginine, and asymmetric dimethylarginine in biological fluids by hydrophilic-Interaction liquid chromatography–electrospray tandem mass spectrometry. *Clin. Chem.* **2006**, *52*, 488–493.

81. Wilcken, D.E.; Wang, J.; Sim, A.S.; Green, K.; Wilcken, B. Asymmetric dimethylarginine in homocystinuria due to cystathionine beta-synthase deficiency: Relevance of renal function. *J. Inherit. Metab. Dis.* **2006**, *29*, 30–37.

82. Bishop, M.J.; Crow, B.; Norton, D.; Paliakov, E.; George, J.; Bralley, J.A. Direct analysis of un-derivatized asymmetric dimethylarginine (ADMA) and L-arginine from plasma using mixed-mode ion-exchange liquid chromatography-tandem mass spectrometry. *J. Chromatogr. B* **2007**, *859*, 164–169.

83. Schwedhelm, E.; Maas, R.; Tan-Andresen, J.; Schulze, F.; Riederer, U.; Böger, R.H. High-Throughput liquid chromatographic-tandem mass spectrometric determination of arginine and dimethylated arginine derivatives in human and mouse plasma. *J. Chromatogr. B* **2007**, *851*, 211–219.

84. Weaving, G.; Rocks, B.F.; Bailey, M.P.; Titheradge, M.A. Arginine and methylated arginines in human plasma and urine measured by tandem mass spectrometry without the need for chromatography or sample derivatisation. *J. Chromatogr. B* **2008**, *874*, 27–32.

85. Zhang, W.; Zhou, C.; Xie, J.; Chen, B.; Chang, L. Serum asymmetric dimethylarginine and endothelial function after renal transplantation. *J. Cent. South Univ. (Med. Sci.)* **2009**, *34*, 289–294.

86. El-Khoury, J.M.; Bunch, D.R.; Reineks, E.; Jackson, R.; Steinle, R.; Wang, S. A simple and fast liquid chromatography-tandem mass spectrometry method for measurement of underivatized L-arginine, symmetric dimethylarginine, and asymmetric dimethylarginine and establishment of the reference ranges. *Anal. Bioanal. Chem.* **2012**, *402*, 771–779.

87. Tain, Y.L.; Huang, L.T. Asymmetric dimethylarginine: Clinical applications in pediatric medicine. *J. Formos. Med. Assoc.* **2011**, *110*, 70–77.

88. Wang, S.; Vicente, F.B.; Miller, A.; Brooks, E.R.; Price, H.E.; Smith, F.A. Measurement of arginine derivatives in pediatric patients with chronic kidney disease using high-performance liquid chromatography-tandem mass spectrometry. *Clin. Chem. Lab. Med.* **2007**, *45*, 1305–1312.

89. Jehlička, P.; Stožický, F.; Mayer, O., Jr.; Varvařovská, J.; Racek, J.; Trefil, L.; Siala, K. Asymmetric dimethylarginine and the effect of folate substitution in children with familial hypercholesterolemia and diabetes mellitus type 1. *Physiol. Res.* **2009**, *58*, 179–184.

90. Heilman, K.; Zilmer, M.; Zilmer, K.; Kool, P.; Tillmann, V. Elevated plasma adiponectin and decreased plasma homocysteine and asymmetric dimethylarginine in children with type 1 diabetes. *Scand. J. Clin. Lab. Invest.* **2009**, *69*, 85–91.

91. Huemer, M.; Simma, B.; Mayr, D.; Möslinger, D.; Mühl, A.; Schmid, I.; Ulmer, H.; Bodamer, O.A. Free asymmetric dimethylarginine (ADMA) is low in children and adolescents with classical phenylketonuria (PKU). *J. Inherit. Metab. Dis.* **2012**, in press.
92. Andrade, F.; Rodríguez-Soriano, J.; Prieto, J.A.; Aguirre, M.; Ariceta, G.; Lage, S.; Azcona, I.; Prado, C.; Sanjurjo, P.; Aldamíz-Echevarría, L. Methylation cycle, arginine-creatine pathway and asymmetric dimethylarginine in paediatric renal transplant. *Nephrol. Dial. Transpl.* 2011, 26, 328–336.

93. Arcos, M.I.; Fujihara, C.K.; Sesso, A.; de Almeida Prado, E.B.; de Almeida Prado, M.J.; de Nucci, G.; Zatz, R. Mechanisms of albuminuria in the chronic nitric oxide inhibition model. *Am. J. Physiol. Renal. Physiol.* 2000, 279, 1060–1066.

94. Okubo, K.; Hayashi, K.; Wakino, S.; Matsuda, H.; Kubota, E.; Honda, M.; Tokuyama, H.; Yamamoto, T.; Kajiyama, F.; Saruta, T. Role of asymmetrical dimethylarginine in renal microvascular endothelial dysfunction in chronic renal failure with hypertension. *Hypertens. Res.* 2005, 28, 181–189.

95. Kaida, Y.; Ueda, S.; Yamagishi, S.I.; Nakayama, Y.; Ando, R.; Iwatani, R.; Fukami, K.; Okuda, S. Proteinuria elevates asymmetric dimethylarginine levels via protein arginine methyltransferase-1 overexpression in a rat model of nephrotic syndrome. *Life Sci.* 2012, in press.

96. Caglar, K.; Yilmaz, M.I.; Sonmez, A.; Cakir, E.; Kaya, A.; Acikel, C.; Eyileten, T.; Yenicesu, M.; Oguz, Y.; Bilgi, C.; et al. ADMA, proteinuria, and insulin resistance in non-diabetic stage I chronic kidney disease. *Kidney Int.* 2006, 70, 781–787.

97. Matsumoto, Y.; Ueda, S.; Yamagishi, S.; Matsuguma, K.; Shibata, R.; Fukami, K.; Matsuoka, H.; Imaizumi, T.; Okuda, S. Dimethylarginine dimethylaminohydrolase prevents progression of renal dysfunction by inhibiting loss of peritubular capillaries and tubulointerstitial fibrosis in a rat model of chronic kidney disease. *J. Am. Soc. Nephrol.* 2007, 18, 1525–1533.

98. Yilmaz, M.I.; Saglam, M.; Caglar, K.; Cakir, E.; Ozgurtas, T.; Sonmez, A.; Eyileten, T.; Yenicesu, M.; Acikel, C.; Oguz, Y.; et al. Endothelial functions improve with decrease in asymmetric dimethylarginine (ADMA) levels after renal transplantation. *Transplantation* 2005, 80, 1660–1666.

99. Zoccali, C. Endothelial damage, asymmetric dimethylarginine and cardiovascular risk in end-stage renal disease. *Blood Purif.* 2002, 20, 469.

100. Teplan, V.; Schück, O.; Racek, J.; Siroka, R.; Haluzik, M.; Kudla, M.; Vitko, S. Asymmetric dimethylarginine and adiponectin after renal transplantation: Role of obesity. *J. Ren. Nutr.* 2008, 18, 154–157.

101. Teplan, V.; Malý, J.; Gürlich, R.; Teplan, V., Jr.; Kudla, M.; Pit’ha, J.; Racek, J.; Haluzik, M.; Senolt, L.; Stollová, M. Muscle and fat metabolism in obesity after kidney transplantation: No effect of peritoneal dialysis or hemodialysis. *J. Ren. Nutr.* 2012, 22, 166–170.

102. Abedini, S.; Meinitzer, A.; Holme, I.; März, W.; Weihrauch, G.; Fellstrom, B.; Jardine, A.; Holdaas, H. Asymmetrical dimethylarginine is associated with renal and cardiovascular outcomes and all-cause mortality in renal transplant recipients. *Kidney Int.* 2010, 77, 44–50.

103. Zhang, D.L.; Liu, J.; Liu, S.; Zhang, Y.; Liu, W.H. The differences of asymmetric dimethylarginine removal by different dialysis treatments. *Ren. Fail.* 2010, 32, 935–940.

104. Maas, R. Pharmacotherapies and their influence on asymmetric dimethylarginine (ADMA). *Vasc. Med.* 2005, 10, S49–S57.
105. Goralczyk, T.; Tisonczyk, J.; Fijorek, K.; Undas, A. High tea and vegetable consumption is associated with low ADMA generation in older healthy subjects. *Metabolism* **2012**, *61*, 1171–1176.

106. Saran, R.; Novak, J.E.; Desai, A.; Abdulhayoglu, E.; Warren, J.S.; Bustami, R.; Handelman, G.J.; Barbato, D.; Weitzel, W.; D’Alessy, L.G.; *et al.* Impact of vitamin E on plasma asymmetric dimethylarginine (ADMA) in chronic kidney disease (CKD): A pilot study. *Nephrol. Dial. Transpl.* **2003**, *18*, 2415–2420.

107. Nanayakkara, P.W.; Kieft-de Jong, J.C.; ter Wee, P.M.; Stehouwer, C.D.; van Ittersum, F.J.; Olthof, M.R.; Teerlink, T.; Twisk, J.W.; van Guldener, C.; Smulders, Y.M. Randomized placebo-controlled trial assessing a treatment strategy consisting of pravastatin, vitamin E, and homocysteine lowering on plasma asymmetric dimethylarginine concentration in mild to moderate CKD. *Am. J. Kidney Dis.* **2009**, *53*, 41–50.

108. Teplan, V.; Schück, O.; Racek, J.; Mareckova, O.; Stollova, M.; Hanzal, V.; Malý, J. Reduction of plasma asymmetric dimethylarginine in obese patients with chronic kidney disease after three years of a low-protein diet supplemented with keto-amino acids: A randomized controlled trial. *Wien. Klin. Wochenschr.* **2008**, *120*, 478–485.

109. Cupiste, A.; Ghiadoni, L.; D’Alessandro, C.; Cardas, I.; Morelli, E.; Panichi, V.; Locati, D.; Morandi, S.; Saba, A.; Barsotti, G.; *et al.* Soy protein diet improves endothelial dysfunction in renal transplant patients. *Nephrol. Dial. Transpl.* **2007**, *22*, 229–234.

110. Urquhart, B.L.; House, A.A. Assessing plasma total homocysteine in patients with end-stage renal disease. *Perit. Dial. Int.* **2007**, *27*, 476–488.

111. Thaha, M.; Widodo, P.W.; Yogiantoro, M.; Tomino, Y. Intravenous N-acetylcysteine during hemodialysis reduces asymmetric dimethylarginine level in end-stage renal disease patients. *Clin. Nephrol.* **2008**, *69*, 24–32.

112. Ruiz, M.C.; Moreno, J.M.; Ruiz, N.; Vargas, F.; Asensio, C.; Osuna, A. Effect of statin treatment on oxidative stress and renal function in renal transplantation. *Transpl. Proc.* **2006**, *38*, 2431–2433.

113. Trimarchi, H.M.; Brennan, S.; González, J.M.; Suki, W.N. Effects of the statins in kidney transplantation. *Medicina (B. Aires)* **2000**, *60*, 457–465.

114. Panichi, V.; Mantuano, E.; Paololetti, S.; Santi, S.; Manca Rizza, G.; Cutrupi, S.; Pizzini, P.; Spoto, B.; Tripepi, G.; Zoccali, C. Effect of simvastatin on plasma asymmetric dimethylarginine concentration in patients with chronic kidney disease. *J. Nephrol.* **2008**, *21*, 38–44.

115. Lim, A.; Manley, K.J.; Roberts, M.A.; Fraenkel, M.B. Fish oil treatment for kidney transplant recipients: A meta-analysis of randomized controlled trials. *Transplantation* **2007**, *83*, 831–838.

116. Beavers, K.M.; Beavers, D.P.; Bowden, R.G.; Wilson, R.L.; Gentile, M. Omega-3 fatty acid supplementation and total homocysteine levels in end-stage renal disease patients. *Nephrology (Carlton)* **2008**, *13*, 284–288.

117. Małyszko, J.; Małyszko, J.S.; Brzóska, S.; Pawlak, K.; Myśliwiec, M. Effects of fluvastatin on homocysteine and serum lipids in kidney allograft recipients. *Ann. Transpl.* **2002**, *7*, 52–54.

118. Vos, I.H.; Rabelink, T.J.; Dorland, B.; Loos, R.; van Middelaar, B.; Grone, H.J.; Joles, J.A. L-Arginine supplementation improves function and reduces inflammation in renal allografts. *J. Am. Soc. Nephrol.* **2001**, *12*, 361–367.
119. Baylis, C. Arginine, arginine analogs and nitric oxide production in chronic kidney disease. *Nat. Clin. Pract. Nephrol.* **2006**, *2*, 209–220.

120. El-Mesallamy, H.O.; Abdel Hamid, S.G.; Gad, M.Z. Oxidative stress and asymmetric dimethylarginine are associated with cardiovascular complications in hemodialysis patients: Improvements by l-arginine intake. *Kidney Blood Press. Res.* **2008**, *31*, 189–195.

121. Bodamer, O.A.; Sahoo, T.; Beaudet, A.L.; O’Brien, W.E.; Bottiglieri, T.; Stockler-Ipsiroglu, S.; Wagner, C.; Scaglia, F. Creatine metabolism in combined methylmalonic aciduria and homocystinuria. *Ann. Neurol.* **2005**, *57*, 557–560.

122. Schramm, L.; La, M.; Heidbreder, E.; Hecker, M.; Beckman, J.S.; Lopau, K.; Zimmermann, J.; Rendl, J.; Reiners, C.; Winderl, S.; *et al.* L-Arginine deficiency and supplementation in experimental acute renal failure and in human kidney transplantation. *Kidney Int.* **2002**, *61*, 1423–1432.

123. Schwedhelm, E.; Maas, R.; Freese, R.; Jung, D.; Lukacs, Z.; Jambrecina, A.; Spickler, W.; Schulze, F.; Böger, R.H. Pharmacokinetic and pharmacodynamic properties of oral L-citrulline and L-arginine: Impact on nitric oxide metabolism. *Br. J. Clin. Pharmacol.* **2008**, *65*, 51–59.

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