Antirheumatoid Arthritic Effects of Sabia parviflora Wall. Leaf Extracts via the NF-κB Pathway and Transient Receptor Potential Protein Family

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As an important traditional medicine of Buyi and Miao ethnic groups in Guizhou, Sabia parviflora Wall. provides antiviral properties against hepatitis, eliminates wind and dampness, and exhibits anti-inflammatory and pain relief properties. It has also been shown to treat rheumatoid arthritis (RA) and other diseases. However, the pharmacodynamic mechanism of S. parviflora Wall. for RA has not been reported. In this study, we identified the effective compounds of S. parviflora Wall. leaves against RA and discussed the mechanism against complete Freund’s adjuvant-induced arthritis (AIA) based on inflammatory proteins and transient receptor potential (TRP) proteins. S. parviflora Wall. leaf extracts (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg, once daily) were given orally for 21 days. On the 15th day of complete Freund’s adjuvant-induced RA, the effects of this medicine on RA rats were investigated. S. parviflora Wall. extracts increased body weight, decreased foot swelling, and reduced thymus and spleen indices in model rats. Most of pannus in the synovial tissue of RA rats disappeared upon treatment, and the local inflammatory cells were greatly reduced when given the fraction of n-butanol (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg, once daily) of 70% alcohol-soluble fraction of S. parviflora Wall. leaves. In addition, the release of inflammatory factors such as tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), interleukin-6 (IL-6), interleukin-10 (IL-10), interleukin-15 (IL-15), and vascular endothelial growth factor (VEGF) in the RA rat serum was inhibited. The active compounds inhibited the expression of TNF-α, IL-1β, IL-6, IL-10, IL-15 and nuclear factor kappa-Bp65 (NF-κBp65) inflammatory protein and TRP protein transient receptor potential melastatin-5 (TRPM-5) and transient receptor potential channel-6 (TRPC-6), to reduce the expression of VEGF in synovial tissue of RA rats and relieve redness and edema. High-performance liquid chromatography identified six flavonoids and three triterpenoid saponins as active compounds. These findings suggest S. parviflora Wall. leaves may play a role in RA treatment by inhibiting the release of inflammatory factors as well as participating in the inflammatory protein expression in the NF-κB pathway and TRP protein family.

Keywords: Sabia parviflora Wall., TRP family, NF-κB, rheumatoid arthritis, flavonoids
INTRODUCTION

Rheumatoid arthritis (RA) is a chronic, systemic autoimmune disease with the proliferation of synovial tissue, infiltration of inflammatory cells, and pannus formation. It gradually leads to the irreversible damage of cartilage and bone, resulting in different degrees of the loss of joint function in RA patients (Aletaha et al., 2018; Li et al., 2019). Currently, the exact
pathogenesis of RA remains unclear. Therefore, research on the etiology and pathological mechanism of RA and the development of therapeutic drugs are imperative. Numerous studies have shown that synovial cells, monocytes/macrophages, and lymphocytes can produce a large number of inflammatory cytokines when RA occurs, which have important internal relationships with the emergence and development of the disease (Falconer et al., 2018; Pala et al., 2018; Siouti et al., 2019; Saleem et al., 2020). At present, the drugs used to treat RA on the market are mainly non-steroidal anti-inflammatory drugs, adrenal glucocorticoids, and biological agents. Although these drugs can relieve pain and reduce inflammatory response, they are prone to serious side effects, such as teratogenicity, osteoporosis, and osteonecrosis. (Chatzidionysiou et al., 2017; Schultz et al., 2017). In contrast, Chinese herbal medicine has attracted extensive attention because of its unique curative effects, low toxicity, minimal side effects, and great development potential. It plays a very important role in the treatment of RA and has obvious characteristics and advantages.

Sabia parviflora Wall. is a deciduous woody vine belonging to the genus Sabia of Sabiaceae with small yellow flowers and red fruits. It is also known as “xiaohuaqingfengteng” in Chinese. Its leaves are considered a vital Miao medicine, and it has been widely used to treat RA by Buyi and Miao ethnic groups in southwest Guizhou Province with no scientific validation (Sui et al., 2011; Chen et al., 2020). The crude extract of S. parviflora Wall. has pharmacological activities against acute liver injury and influenza virus (Chen et al., 2015; Zhao et al., 2018). At present, the research on the chemical constituents of S. parviflora Wall. mainly focuses on triterpenoids, alkaloids, flavonoids, and other compounds (Fan et al., 2018; Cui et al., 2022; Zhou et al., 2022). However, the pharmacodynamic mechanism of S. parviflora Wall. leaves for the treatment of RA has not been reported.

In this research, we identified the bioactive compounds of S. parviflora Wall. leaf extracts for the treatment of RA in a rat model. We also measured the inflammatory cytokine levels in the serum of RA rats and the pathogenesis-related protein expression in synovial tissue in order to elucidate the possible mechanism.

**MATERIALS AND METHODS**

**Chemicals and Reagents**

Complete Freund's adjuvant (CFA) was obtained from Sigma Chemical Co. (St.Louis, MO, United States), and dexamethasone was purchased from Hubei Tianyao Pharmaceutical Co. (Wuhan, China). Tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), interleukin-6 (IL-6), interleukin-10 (IL-10), interleukin-15
FIGURE 4 | Contents of IL-18, IL-6, IL-10, IL-15, TNF-α, and VEGF in rat serum of different groups. Data are expressed as (mean ± SD), n = 8. **p < 0.01 vs. control group, ^p < 0.05 and ^ ^ p < 0.01 vs. model group, ♦ p < 0.05 and ♦ ♦ p < 0.01 versus dexamethasone group, ▲ p < 0.05 and ▲ ▲ p < 0.01 vs. different groups of petroleum ether, low-dose, and middle-dose of ethyl acetate group, and ◆ p < 0.05 versus high-dose of ethyl acetate group.
(IL-15), and vascular endothelial growth factor (VEGF) enzyme-linked immunosorbent assay (ELISA) kits were purchased from ZhongShan Golden Bridge Biotechnology Co. (Beijing, China). TNF-α (bs-10802R), IL-1β (bs-6319R), IL-6 (bs-6309R), IL-10 (bs-0698R), IL-15 (bs-1829R), VEGF (bs-1313R), nuclear factor kappa-Bp65 (NF-κBp65) (bs-0465R), transient receptor potential channel-6 (TRPC-6) (bs-2393R), transient receptor potential melastatin-5 (TRPM-5) (bs-9047R) of rabbit polyclonal antibody, and goat anti-rabbit IgG H&L/HRP antibody (bs-40295G-HRP) were purchased from Beijing Biosen Biotechnology Co. (Beijing, China). High-performance liquid chromatography (HPLC)-grade methanol was purchased from American BCR company (Columbus, Ohio, United States). Watsons drinking water (Guangzhou Watsons Food & Beverage Co., Guangzhou, China) was used in this experiment. Other reagents were analytically pure (Tianjin Fuyu Fine Chemical Co., Tianjin, China).

**Plant Material and Extraction**

The plant was collected from Ceheng County, Guizhou, China, in August 2018 and was identified as Sabia parvi flora Wall. by Professor Qingwen Sun. A voucher specimen (number 20180820) was deposited at the Traditional Chinese Medicine and Ethnic Medicine Laboratory of Guizhou University of Traditional Chinese Medicine.

The air-dried and crude powdered leaves of S. parvi flora Wall (12.3 kg) were extracted with 70% ethanol (98 L × three times), each time for 2 hours. The combined filtrate was concentrated under reduced pressure to obtain the black extract (2,361.6 g) with a yield of 19.2%. The ethanol extract was extracted five times with petroleum ether, ethyl acetate, and water-saturated n-butanol, respectively. After vacuum drying under reduced pressure, the dried products of the petroleum ether layer (350.5 g), ethyl acetate layer (301.3 g), and water-saturated n-butanol layer (1,295.0 g) were obtained.

**FIGURE 5 | Protein of IL-1 β (A) expression in rat synovial tissue of different groups and its average optical density (AOD) (B).** A, B, and C represent normal group, model group, and positive group (dexamethasone), respectively. D, E, and F represent low-dose, middle-dose, and high-dose of fraction of n-butanol extract of 70% alcohol-soluble fraction of S. parvi flora Wall, leaves, respectively. Data are expressed as (mean ± SD), n = 8. **p < 0.01 vs. control group, *p < 0.05 and ***p < 0.01 vs. model group, ▲ p < 0.05 vs. dexamethasone group, and ▲▲ p < 0.05 vs. low-dose of n-butanol group.
Animals
SPF Sprague-Dawley (SD) rats (180–200 g) were obtained from Changsha Tianqin Biotechnology Co., Ltd (Changsha, China). The standard rat chow was obtained from Shanghai SLAC Laboratory Animal Co., Ltd. (Shanghai, China). The rats were maintained under standard laboratory conditions (temperature 22 ± 1°C, humidity 60 ± 5%, and with 12 h light-dark cycle) and given standard rodent chow and tap water before initiation of the experiment. All animal experiments were carried out in accordance with the Animal Ethics Committee of Guizhou University of Traditional Chinese Medicine on 25 June 2021 with the Approval No. 20210040.

Replication of CFA-Induced RA Rat Model and Drug Administration
The rats were adaptively fed for 1 week and their body weight and foot diameter were measured every week thereafter. A total of 96 SD rats were randomly divided into 12 groups of eight animals: 1) normal group, 2) model group, 3) positive control group (dexamethasone), 4) petroleum ether low-dose group, 5) petroleum ether middle-dose group, 6) petroleum ether high-dose group, 7) ethyl acetate low-dose group, 8) ethyl acetate middle-dose group, 9) ethyl acetate high-dose group 10) n-butanol low-dose group, 11) n-butanol middle-dose group, and 12) n-butanol high-dose group. The model was carried out for 2 weeks for all groups, excluding the normal group (each animal was given an intradermal injection of 0.2 ml CFA into the left hind footpad) according to a previously described method (Pan et al., 2017; Bao et al., 2019). The extracts were administered via gavage to all groups once a day for 3 weeks, with ordinary feeding after 2 weeks. The dosages of petroleum ether, ethyl acetate, n-butanol (high-, middle-, and low-dose groups), and the dexamethasone positive control groups were 0.64 g/kg, 0.32 g/kg, 0.16 g/kg, and 7.5 mg/kg, respectively.

**FIGURE 6** | Protein of IL-6 (A) expression in rat synovial tissue of different groups and its average optical density (AOD) (B). A, B, and C represent normal group, model group, and positive group (dexamethasone), respectively. D, E, and F represent low-dose, middle-dose, and high-dose of fraction of n-butanol extract of 70% alcohol-soluble fraction of *S. parviflora* Wall. leaves, respectively. Data are expressed as (mean ± SD), n = 8. *p < 0.05 vs. control group, **p < 0.01 vs. control group, #p < 0.05 and ##p < 0.01 vs. model group, ▲p < 0.05 and ▲▲p < 0.01 vs. dexamethasone group.
Measurement of Body Weight and Changes in Ankle Circumference
The body weight of rats in each experimental group and the normal group were measured and recorded every 7 days from day zero to day 35. The changes in the ankle circumference were also evaluated using a Vernier caliper every 7 days from day zero to day 35.

Thymus Index and Spleen Index Assay
After collecting blood from the abdominal aorta of rats, the anesthetized rats were killed, and the thymus and spleen were immediately removed and weighed. The index of the thymus and spleen is calculated as follows: thymus index = thymus mass/body mass × 1,000 and spleen index = spleen mass/body mass × 1,000 (Jing et al., 2019).

Hematoxylin-Eosin Staining Analysis of Synovial Tissue
The rats were killed on day 35, and the ankle joints of rats were cut, fixed in 4% (w/v) paraformaldehyde, and decalcified in 10% ethylene-diamine-tetraacetic acid (EDTA) at 4°C for 1 month. The decalcified ankle tissues were dehydrated, embedded in wax, sectioned, and stained with hematoxylin-eosin. The stained tissues were sealed and then observed and photographed under a microscope using a Ba200 digital micro camera system (magnification, ×100, Motic China Group Co., Ltd., Xiamen, China).

Immunohistochemical Staining
The expressions of inflammatory proteins in synovial tissue were measured according to the immunohistochemical staining method (He et al., 2018). The treated synovial tissues were incubated with
primary antibodies of TNF-α (1:200), IL-1β (1:400), IL-6 (1:400), IL-10 (1:400), IL-15 (1:400), NF-κBp65 (1:500), VEGF (1:500), TRPC-6 (1:400), and TRPM-5 (1:400) overnight in a refrigerator at 4°C. The following day, the tissues were washed with phosphate-buffered solution (PBS) three times for 3 min. The tissues were incubated with the anti-goat antibody for 30 min at 37°C and washed with PBS three times for 3 min. The chromogenic agent, 3,3′-diaminobenzidine (DAB), was added for 10 min. The color was observed at room temperature, and the dyeing state was observed under a microscope. Then, the tissues were stained with hematoxylin for 5 min and examined under a microscope using a Ba200 digital micro camera system (magnification, ×100, Motic China Group Co., Ltd., Xiamen, China). The protein expression in synovial tissue was quantitatively analyzed by the average optical density (AOD) of protein according to a previously described method. (Wang, et al., 2021).

ELISAs for Inflammatory Factors in Serum
The rats were anesthetized by the intraperitoneal injection of 20% urethane, and 4 ml of blood was collected from the abdominal aorta in a labeled red glass. The serum was obtained from blood by cryogenic centrifugation (4°C, 3,000 rpm, and 15 min) and the serum levels of TNF-α, IL-1β, IL-6, IL-10, IL-15, and VEGF were quantified by enzyme-linked immunosorbent assay kits (ELISAs) following the manufacturer instructions.

HPLC Analysis of the n-Butanol Active Fraction
Chromatographic analysis was performed on an Agilent-1260 HPLC system (Agilent Technologies Inc, Santa Clara, California, United States) coupled with a rapid separation diode array detector (DAD). Chromatographic separation
was performed on a YMC-Pack ODS-A column (250 mm × 4.6 mm, 5 μm). The mobile phase consisted of methanol (A) and water (B), and the flow rate was 1.0 ml/min. A linear gradient elution was employed for elution from 35% (A) (v/v) to 100% (A) (v/v) (0–50 min). The DAD detection wavelength was 210 nm, and the column temperature was 25 °C. The injection volume was 10 μL.

Statistical Analysis
Statistical analysis of the experimental data was conducted using GraphPad PRISM version 5.0 software and SPSS 17 software. The results are presented as mean ± standard deviation (SD). The differences between groups were analyzed by one-way analysis of variance (ANOVA) with the post hoc test. A p-value less than 0.05 was considered statistically significant, and a p-value less than 0.01 was considered for significant differences.

RESULTS
Effects of S. parviflora Wall. Leaf Extracts on Body Weight and Ankle Circumference in Rats
The body weight of rats in the model group showed a gradual increase, compared to the normal control from day 14 to day 35 (Figure 1A). The administration of S. parviflora Wall. leaf extracts, only a fraction of n-butanol (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg), and fraction of ethyl acetate (0.64 g/kg) increased body weight from day 21 to day 35, compared to that of the model group (see Supplementary Table S1).

The swelling of the ankle joint of rats from day 7 to day 35 is shown in Figure 1B. The right foot of rats in other groups began to swell on day 7 (p < 0.01) compared to the normal rats.
Compared with the RA model group, the swelling of rats’ paw of fraction of n-butanol (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg) and fraction of ethyl acetate (0.64 g/kg) of 70% alcohol-soluble fraction of \textit{S. parvi}flora Wall. leaves-treated group was alleviated from day 21 (Figure 1B, Figure 1C) (\(p < 0.05\) and \(p < 0.01\)) (see Supplementary Table S2).

**Effects of \textit{S. parvi}flora Wall. Leaf Extracts on the Thymus Index and Spleen Index in Rats**

Compared with the normal group, the thymus index (Figure 2B) and spleen index (Figure 2A) of the RA model group were significantly elevated (\(p < 0.01\)). In addition, the thymus index and spleen index of the n-butanol fractions (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg) (\(p < 0.05\) and \(p < 0.01\)) and ethyl acetate fraction (0.64 g/kg) (\(p < 0.05\)) of 70% alcohol-soluble fraction of \textit{S. parvi}flora Wall. leaves were lower than those of the model group. Also, the thymus index and spleen index of other treatment groups were higher than those of dexamethasone (\(p < 0.05\) and \(p < 0.01\)) (see Supplementary Table S3).

**Effects of \textit{S. parvi}flora Wall. Leaf Extracts on Histopathological Changes in Rat Synovial Tissue**

The cells in the synovial tissue of the normal group were arranged orderly, without inflammatory cell infiltration and vascular proliferation, and the articular cartilage was smooth without damage (Figure 3A). The synovial tissue of the RA model group rats showed obvious synovial hyperplasia, disordered arrangement, and local joint fibrosis. The inflammatory cell infiltration appeared in the synovial stroma and small blood vessels increased in the lower synovial layer, resulting in the formation of a large number of pannus (Figure 3B). Compared

FIGURE 10 | Protein of TNF-\(\alpha\) (A) expression in rat synovial tissue of different groups and its average optical density (AOD) (B). A, B, and C represent normal group, model group, and positive group (dexamethasone). D, E, and F represent the low-dose, middle-dose, and high-dose of fraction of n-butanol extract of 70% alcohol-soluble fraction of \textit{S. parvi}flora Wall. leaves. Data are expressed as (mean ± SD), \(n = 8\). **\(p < 0.01\) vs. control group, *\(p < 0.05\) and **\(p < 0.01\) vs. model group, ^\(p < 0.05\) and ^\(^\circ\)\(p < 0.01\) vs. dexamethasone group, and *\(p < 0.05\) vs. low-dose of n-butanol group.
with the model group, the n-butanol fractions (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg) and ethyl acetate fraction (0.64 g/kg) of the 70% alcohol-soluble fraction of *S. parvi*flora* Wall. leaves reduced the pannus in the synovium and local inflammatory cells after treatment for 3 weeks (Figures 3I–L).

**Effects of *S. parvi*flora* Wall. Leaf Extracts on Inflammatory Cytokine Levels in Rat Serum**

As demonstrated in Figure 4, the inflammatory cytokine levels of IL-1β, IL-6, IL-10, IL-15, TNF-α, and VEGF in the serum of the model group rats were significantly higher than those in the normal group rats. The IL-1β, IL-6, IL-10, IL-15, TNF-α, and VEGF levels in RA rats treated with the n-butanol fractions (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg) (p < 0.05, p < 0.01) and ethyl acetate fraction (0.64 g/kg) (p < 0.05) of the 70% alcohol-soluble fraction of *S. parvi*flora* Wall. leaves decreased. In addition, the positive control dexamethasone (7.5 mg/kg/d) also reduced the release of these inflammatory cytokines with significant efficacy (p < 0.01). In addition, the inflammatory cytokine levels of other treatment groups were higher than those of dexamethasone (p < 0.05 and p < 0.01) (see Supplementary Table S4).

**S. parvi*flora* Wall. Leaf Fraction of n-Butanol Extract Suppresses the Expression of Inflammatory Proteins in Rat Synovial Tissue**

A large number of inflammatory proteins were expressed in RA rat synovial tissue as determined by immunohistochemical examination. Compared with the normal group, the expression of proteins IL-1β, IL-6, IL-10, IL-15, NF-κBp65, TNF-α, TRPC-6, TRPM-5, and VEGF (Figures 5–13) in synovial tissue of RA rats increased significantly (p < 0.01). Compared with the RA model
group, the expression of these inflammatory proteins in synovial tissue decreased upon treatment with the n-butanol fractions (0.64 g/kg, 0.32 g/kg, and 0.16 g/kg) (p < 0.05, p < 0.01) of the 70% alcohol-soluble fraction of *S. parviflora* Wall. leaves. Treatment with dexamethasone (7.5 mg/kg/d) also relieved the inflammatory protein expression in synovial tissue of RA rats, with significant efficacy (p < 0.01). In addition, the expressions of proteins IL-1β, IL-6, IL-10, IL-15, and TNF-α of other treatment groups were higher than those of dexamethasone (p < 0.05) (see Supplementary Table S5 and Supplementary Table S6).

**Identification of Active Components in n-Butanol Active Fraction**

Six flavonoids: rutin (1), isorhamnetin-3-O-rutinoside (2), kaempferol-3-O-rutinoside (3), quercetin-3-O-gentiobioside (4), luteolin (5), and kaempferol 6) and three triterpenoid saponins: saikosaponin a (7), saikosaponin b2 (8), and saikosaponin d 9) were identified from the n-butanol fraction of the 70% ethanol extract by HPLC (Figure 14A,B).

**DISCUSSION**

Rheumatoid arthritis belongs to the category of “arthralgia” (“Bi Zheng” in Chinese) in traditional Chinese medicine, and the main clinical manifestations are pain and numbness in muscles, bones, and joints (Li et al., 2009). Modern medicine shows that the core of RA lesion is the synovial joint, and its main histocytological changes include proliferation of synovial intimal cells, infiltration of inflammatory cells in the subintimal layer of synovium, formation of pannus, which erodes the articular surface, activation of osteoclasts, and mediation of the erosion of mineralized cartilage and soft bone (Schett et al., 2012).

**FIGURE 12 |** Protein of TRPM-5 (A) expression in rat synovial tissue of different groups and its average optical density (AOD) (B). A, B, and C represent normal group, model group, and positive group (dexamethasone). D, E, and F represent low-dose, middle-dose, and high-dose of fraction of n-butanol extract of 70% alcohol-soluble fraction of *S. parviflora* Wall. leaves, respectively. Data are expressed as (mean ± SD), n = 8. 

\[\text{AOD of proteins} \]

\[\text{Normal} \quad \text{Model} \quad \text{Dexamethasone (7.5 mg/kg/d)} \quad \text{N-Butanol (L) } \quad \text{N-Butanol (M)} \quad \text{N-Butanol (H)} \]

\[\text{p} < 0.01 \text{ vs. control group}, \quad \text{p} < 0.05 \text{ and } \text{p} < 0.01 \text{ vs. model group}, \quad \text{p} < 0.05 \text{ vs. dexamethasone group, and } \text{p} < 0.05 \text{ vs. low-dose of n-butanol group.}

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Recent studies have validated the crucial roles of osteoprotegerin (OPG)/receptor activator of nuclear factor-κB ligand (RANKL), mitogen-activated protein kinase (MAPK), c-Jun N-terminal kinase (JNK), and Janus kinase (JAK)/signal transducers and activators of transcription (STAT) signal pathways in RA. Among these pathways, many important signal molecules and transcription factors, including NF-κB, c-fos, and c-jun, are involved in the pathogenesis of RA (Gaafar, et al., 2018; Bao, et al., 2019; Kanai, et al., 2020; Zhao et al., 2021; Zhai., et al., 2019). In this study, we have confirmed the antirheumatoid arthritis effects of *S. parviflora* Wall. leaf extracts on RA rats via the NF-κB pathway and TRP protein family.

Studies have shown that immune disorders in RA patients are mainly caused by the activation of specific signaling pathways by IL-6, IL-1, TNF-α, and other cytokines. NF-κB is one of the important signal transduction pathways, and it plays a decisive role in regulating inflammatory response (Xia et al., 2018; Zuo et al., 2018). At present, researchers have found that inflammatory cytokines, such as TNF-α and interleukin, in the synovial fluid of RA patients can activate the NF-κB pathway through different ways to cause the RA onset and joint destruction. Targeted blocking of the NF-κB pathway can regulate some cell activities and improve the pathological process of RA (Huang et al., 2019; Pandolfi, et al., 2020). In the present study, the high-dose ethyl acetate fraction, and the n-butanol fractions of the 70% alcohol-soluble fraction of *S. parviflora* Wall. leaves reduced the serum levels of inflammatory factors, IL-1β, IL-6, IL-10, IL-15, and TNF-α in RA rats. These treatment groups also exhibited inhibition of IL-1β, IL-6, IL-10, IL-15, TNF-α, and NF-κB protein expressions in RA rat synovial tissue.

In addition, researchers have found a superfamily of membrane proteins that are found in a wide variety of organisms and participate in sensing external stimuli,
namely, TRP. Most subtypes are non-selective Ca\(^{2+}\) permeability channels (David, 2003), and most endothelial cell functions depend on intracellular free Ca\(^{2+}\) concentration and Ca\(^{2+}\) signal transduction. The TRP plays an important role in the regulation of vascular endothelial cell function (Wong et al., 2011). Research on the mechanism of the TRP family, in the regulation of vascular endothelial function, has become a hot spot. Most members of the TRP family have calcium permeability function and are extremely sensitive to exogenous stimuli. Therefore, they have been extensively studied with the aim of revealing the molecular mechanisms of vascular function changes caused by different factors and the occurrence and development of vascular diseases. In particular, TRPC-6 and TRPM-5 proteins are the focus of many studies, as they play key roles in the pathogenesis of vascular diseases (Wu et al., 2020; Shekhar, et al., 2021). In the present work, the n-butanol extract of the 70% alcohol-soluble fraction of S. parvi flora Wall. leaves could prevent the calcium influx by inhibiting the expression of TRPC-6 and TRPM-5 proteins in RA rat synovial tissue.

Many studies have shown the TRP subfamily members can participate in angiogenesis by affecting the concentration of calcium in endothelial cells. For example, by inhibiting the TRPC-6 channel in human umbilical vein endothelial cells, the cells can stagnate in the G2/M phase and prevent endothelial cell proliferation and lumen formation induced by VEGF, but it has no significant effects on endothelial growth factor (EGF)-induced angiogenesis, indicating TRPC-6 plays an important role in VEGF-induced angiogenesis (Ge et al., 2009). As an important vascular growth factor, VEGF is continuously and highly expressed in the early stage of RA, combined with its receptor through signal transduction. It promotes angiogenesis and increases vascular permeability, resulting in continuous synovitis and pannus formation of RA (Kim et al., 2015; Yamaguchi et al., 2019). Studies have shown that many inflammatory factors directly or indirectly regulate the expression of VEGF in RA patients, and some cytokines such as IL-1, TNF-\(\alpha\), and IL-6 can upregulate the expression of VEGF (Bi et al., 2019; Mumtaz et al., 2020). Therefore, anti-VEGF treatment of RA can delay the progress of the disease, improve clinical symptoms, and provide a new
strategy for clinical prevention and treatment of RA. In this work, the n-butanol extract of the 70% alcohol-soluble fraction of *S. parvi*flora Wall. leaves inhibited the formation of pannus and reduced the symptoms of synovitis by affecting the protein function of VEGF.

At present, alkaloids, flavonoids, and terpenes have been isolated and identified from *S. parvi*flora Wall. Among them, pentacyclic triterpenoids are the main type of triterpenoids in *S. parvi*flora Wall., and the predominant type of flavonoids is flavonols, which exhibit antiviral and anti-inflammatory activities (Li, et al., 2020; Sun et al., 2021). In this study, we identified five flavonols, one flavonoid, and three triterpenoid saponins from the chromatographic peaks of the n-butanol extract of 70% alcohol-soluble fraction of *S. parvi*flora Wall. leaves. These results suggest that the flavonols and triterpenoid saponins in the leaves of *S. parvi*flora Wall. may be involved in the treatment of RA. According to chromatographic profile analysis of *S. parvi*flora Wall. leaf extracts, we preliminarily summarized and mapped the possible mechanism of *S. parvi*flora Wall. leaves in the treatment of RA (Figure 15). The flavonoids and triterpenoids in the effective parts of the leaves of *S. parvi*flora Wall. may block the calcium influx by reducing the protein expression of VEGF and inhibiting IL-1β, TNF-α, and other inflammatory factors and protein expression. As a result, the protein expression of the NF-κB pathway is inhibited, playing a role in the treatment of RA.

In this study, we confirmed that the leaves of *S. parvi*flora Wall. contain important bioactive compounds that play a role in the treatment of RA by inhibiting the release of inflammatory factors and inhibiting the expression of TRP family protein to reduce the expression of IL-1β, IL-6, TNF-α, and VEGF inflammatory proteins and further inhibit the expression of NF-κB protein in synovial tissue. The combined effects lead to the mitigation of redness and edema in the foot in RA.

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article/Supplementary Material; further inquiries can be directed to the corresponding authors.
ETHICS STATEMENT

The animal study was reviewed and approved by the Animal Ethics Committee of Guizhou University of Traditional Chinese Medicine.

AUTHOR CONTRIBUTIONS

CZ analyzed the fraction of n-butanol extract of 70% alcohol-soluble fraction of *S. parviflora* Wall. leaves. YZ, HX, and XZ contributed to study design and experiments. YZ wrote the manuscript and performed data interpretation. YZ and YZ revised the manuscript. TZ and YZ supervised the work.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2022.880350/full#supplementary-material

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