Abstract

The identification, purification and characterization of cancer stem cells (CSCs) holds tremendous promise for improving the treatment of cancer. Mounting evidence is demonstrating that only certain tumour cells (i.e. the CSCs) can give rise to tumours when injected and that these purified cell populations generate heterogeneous tumours. While the cell of origin is still not determined definitively, specific molecular markers for populations containing these CSCs have been found for leukaemia, brain cancer and breast cancer, among others. Systems approaches, particularly molecular profiling, have proven to be of great utility for cancer diagnosis and characterization. These approaches also hold significant promise for identifying distinctive properties of the CSCs, and progress is already being made.

Keywords: transcriptomics • proteomics • molecular signature • networks

Introduction

Cancer is the second deadliest disease in the United States, accounting for one out of every four deaths. Despite tremendous efforts dedicated to conquering this disease, the age-adjusted mortality rate has remained almost unchanged for most cancers for the past five decades, with one compelling instance being a devastating brain tumour – glioblastoma – which carries a median survival rate of merely 9–12 months regardless of treatment. Among the conglomeration of cellular, molecular and genetic complications contributing to the persistence of cancer, the cellular origin provides a promising avenue that could
lead to targeted elimination of cancer. Cancers often occur in tissues with constant proliferation. A paradigm learned from developmental biology suggests that only the clonogenic stem cells in the hierarchical tissue developmental processes are long-lived, whereas their differentiated progenies lose proliferating potential. Emerging evidence suggests that cancers may actually be a malignance derived from a small cell population dubbed cancer stem cells (CSCs), among the heterogeneous tumour cells, that demonstrate self-renewal and multiple lineage differentiation properties [1]. Discovered initially in leukaemia [2], cancer stem cell populations have been isolated from several tissues, including breast, brain and colorectal cancers [3–5]. The identification of these cancer stem cell populations sets the stage for systems level analysis, and holds potentially tremendous ramifications in terms of our understanding of the molecular pathogenesis of cancer, patient stratification, and therapeutic intervention.

As with any complex biological system, cancer (including CSCs) can be viewed and interrogated at the genome-scale using systems biology approaches. Systems approaches stress three concepts regarding biological information [6]. First, there are two fundamental types of biological information – the digital information of the genome and the environmental information that is outside our DNA. Second, the digital genome information encodes two types of biological networks – protein interactions and gene regulatory networks. Protein networks transmit and use biological information for development, physiology and metabolism. Gene regulatory networks – transcription factors and RNAs that regulate networks of other transcription factors and other RNAs – receive information from, for example, signal-transduction networks, integrate and modulate it, and convey the processed information to networks of genes and proteins that execute developmental and physiological functions. In biological systems, these two types of networks are closely integrated. The organization of these networks can be inferred from various different types of measurements including, for example, global measurements of dynamically changing levels of mRNAs and proteins during developmental and physiological responses, as well as large-scale measurements of protein-protein and protein-DNA interactions. Third, there are many hierarchical levels of organization and information (for example, DNA, RNA and protein networks, cell signalling and metabolic networks and organization and responses of organ systems). To understand biological systems, information must be gathered from as many information levels as possible and integrated into models that generate testable hypotheses about how biological systems function.

In this review article we delineate our view for investigating CSCs using powerful integrated transcriptomic, proteomic and computational approaches. We will focus on gene expression profile signatures which will provide fundamental insights into the networks in the application of cancer diagnosis, patient stratification, and treatment management. We will also discuss emerging new experimental technologies and computational modelling approaches that empower systems strategies for tackling cancer stem cell challenges.

Molecular profiling for cancer classification

Systems approaches have proven of great utility in the study of cancer, with increasing power expected to continue to emerge in the future. Despite notable and significant challenges that remain [7, 8], one area that has shown significant promise is in the mining of global gene expression data sets to identify molecular signatures that can be used for diagnosis and treatment selection [9].

These studies typically involve the collection of samples from two or more classes (e.g. cancer versus normal, or responsive versus non-responsive to treatment) and the use of a set of data on which to train the classifier and another set on which to test. In the absence of a true test set, re-sampling methods such as cross-validation are generally used to estimate likely performance of the classifier on future data. Challenges often arise in these studies when different measurement platforms are used in training and test sets. The ability to generate an accurate classifier is a function of factors such as (1) the size of the training set relative to the number of features, (2) the computational method used and (3) the inherent distinctness of the selected phenotypes. Typically, the number of samples is far less than the number of transcripts, leading to overfitting being a significant problem. This leads to the need for computational methods that aid in avoiding overfitting when selecting a
classifier. A variety of methods have been applied to cancer diagnoses including approaches based on support vector machines [10, 11] and relative expression reversals [12–15], among many others.

Application of these methods has led to the discovery of molecular classifiers of varying degrees of accuracy to identify prognostic signatures for breast cancer [16–32], ovarian cancer [33–35], colon cancer [36, 37], prostate cancer [14, 38–43] and brain cancer [44, 45], among others. Given the success of such global approaches to identify signatures for cancer tumours, there is reason to suppose that such approaches will prove very useful in uncovering new biology, markers, and treatments in the emerging area of CSCs. Indeed, progress in this direction is already being made.

Molecular profiling of cancer stem cells

Following on the successes of molecular profiling in identifying prognostic signatures for many cancers, researchers have begun to perform profiling of CSCs as well. We will discuss such efforts in the context of three cancers: leukaemia, brain and breast. In addition to profiling for signatures of specific CSCs, interesting work has also been done to find general signatures for ‘stem-ness’ in tumours. For example, an 11-gene signature for ‘stem-ness’ in multiple cancer types has been identified [46] that predicts short interval to disease recurrence, distant metastasis and death from cancer. This signature reflects a BMI-1 oncogene-driven gene expression pathway, where the BMI-1 gene is essential for the self-renewal of haematopoietic and neural stem cells. Using Kaplan–Meier analysis, this signature for ‘stem-ness’ was found to show predictive ability in 11 different cancers, including five epithelial cancers (prostate, breast, lung, ovariina and bladder) and six non-epithelial (lymphoma, mesothelioma, medulloblastoma, glioma and acute myeloid leukaemia). Thus, there is evidence that the property of ‘stemness’ (defined with this signature) is predictive of outcome in a wide variety of tumours. If validated, this observation could have a major impact on patient care [47]. Recent studies have also shown that cancer and normal stem cells share the same self-renewal mechanisms, such as the Bmi1 and Wnt canonical pathways [48, 49], further strengthening the link between stem cells and CSCs.

Historical perspective: cancer stem cells in leukaemia

The fundamental concept of CSCs came from early studies of blood cancer (leukaemia) and the blood forming haematopoietic stem cells (HSC). Elegant work by Till and McCulloch in the early 1960s established the existence of bone marrow HSC capable of forming myelo-erythroid colonies in the spleen of lethally irradiated hosts [50]. These cells were later isolated by Weissman and colleagues and shown to be able to self-renewal and exhibit multi-potent differentiation giving rise to all the blood lineages [51, 52]. Studies of human leukaemia using in vitro and in vivo colony-formation assays demonstrated that only a small subset of leukaemia cells possess extensive proliferative capability [53, 54] suggesting that leukaemia may actually be derived from a small leukaemic stem cell (LSC) population. This concept was further proved by the successful isolation of myeloid leukaemia-initiating cells using cell surface phenotype CD34+CD38− and subsequent in vivo transplantation into severe combined immune-deficient (SCID) mice [2]. One intriguing question that remained unanswered until very recently is whether the LSCs are derived from normal HSCs or from their downstream committed progenitor cells that regain the self-renewal property. Using mouse genetics and clinical studies, Weissman and colleagues demonstrated that both mechanisms exist in leukaemia: while chronic myeloid leukaemia can be derived from LSC residing at the HSC stage [55], the more differentiated progenitor – granulocyte-macrophage progenitor (GMP) – can also gain LSC property during the blast-crisis of myeloid leukaemia [56]. One common characteristic between stem cells and cancer is the unlimited self-renewal capability. The hierarchical organization and the cellular heterogeneity in the haematopoietic systems, and lessons learned from LSC have inspired later isolation of CSCs from solid tumours [3], and will continue to provide theoretical framework for investigating tumouregenesis.

Molecular profiling of brain cancer stem cells

Brain cancer is a highly heterogeneous disease consisting of multiple tumour types affecting both children and adults [57]. The most malignant form of
brain cancer, glioblastoma multiforme (GBM), is characterized by a diverse cellular phenotypic and genetic heterogeneity which has been a significant impediment to the development of effective targeted medical therapies. Traditional treatment for GBM consists of surgical resection to reduce the tumour mass followed by radiation and chemotherapy selective for highly-proliferating tumour cells. Despite continuous refinement over the past three decades, these treatments have not significantly impacted time to tumour recurrence or long-term survival in GBM patients. Given the low-proliferate rate of normal brain tissue, it has long been hypothesized that brain cancer may arise from neural stem or progenitor cells which were thought to be more sensitive to oncogenic transformation [58]. Using neural stem cell culture techniques, two different types of malignant brain tumours were shown to possess sub-populations of tumour cells with characteristic stem cell-like properties, such as self-renewal, potential for differentiation and capacity to form neurospheres [59]. The cell (or cells) of origin for these putative brain CSCs has not been determined definitively, but evidence on this subject is starting to accumulate. For example, a number of groups have confirmed that brain tumours contain a small fraction of cells that can be separated on the basis of the cell-surface marker CD133 and that have clonogenic potential as measured by the neurosphere assay. In a key in vivo experiment, a minority population of cells from brain tumours was shown to be tumourigenic by implantation into immuno-compromised mouse brain. In this model, injection of as few as 100 CD133+ cells could generate tumours in vivo, whereas injecting of as many as $10^5$ CD133+ cells, which comprise the majority of the GBM tumour cells, did not lead to any viable tumour growth. Interestingly, CD133 had previously been found to be a marker of HSC [60–62] and has now been implicated in the identification of a number of putative tumour stem cells from a variety of human cancers, including leukaemia [63], prostate cancer [64] and colon cancer [4]. At this point, evidence supports that the sub-population of tumourigenic brain cancer cells is a subset of CD133+ cells as not all CD133+ cells are tumourigenic [65]. Several labs are currently refining molecular markers to refine tumour stem cell identification and purification.

The paradigm that brain tumours possess a small fraction of cancer-initiating tumour stem cells which can reconstitute the tumour’s cellular and functional hierarchy has important implications for treatment. To further explore the potential differences in sub-populations of GBM cells, Lee et al. used high-throughput genome-wide molecular profiling to characterize GBM-derived tumour stem cells cultured in serum-free conditions optimized to sustain neural stem cells [66]. Genome-wide expression and genotyping studies revealed that these GBM-derived tumour stem cells had extensive similarities to normal neural stem cells and possessed the capacity to initiate brain tumours in a xenograft mouse model. Significantly, the genotype and phenotype of the mouse xenograft brain tumours resulting from implantation of tumour stem cells cultured in serum-free conditions matched that of the parent human GBM tumours from which they were derived. When GBM-derived tumour cell lines were generated using standard serum-based culture conditions not conducive to stem cell growth, the resulting cells lacked the ability to initiate or propagate tumours in the mouse xenograft model. Genome-wide transcription analysis and genotyping studies revealed significant differences between these serum-cultured cell lines and their corresponding parent GBM tumours and matched serum-free cultured tumour stem cell lines. Of note, the serum-cultured cell lines did manifest extensive similarities to the most commonly utilized immortalized cancer cell lines. These studies provided further evidence that GBM has a distinct sub-population of tumour cells with stem cell-like properties, and that it is these cells that are biologically relevant in initiating and sustaining tumour growth. Unfortunately, most preclinical studies have historically been based on targeting therapies to features identified in serum-cultured immortalized cancer cell lines. This landmark large-scale molecular profiling study raises significant concerns about this approach and suggests the use of serum-free cultured tumour-stem cells as a more appropriate pre-clinical model [66].

There is great interest in identifying effective therapies that can target the tumour-initiating and tumour-maintaining sub-population of tumour stem cells identified as CD133+. Two recent studies used molecular profiling to characterize the differential response of CD133+ cells to therapeutic intervention compared to CD 133− cells. After radiation treatment, GBM tumours exhibited an increase in the fraction of CD133+ cells, more robust activation of DNA repair pathways in CD 133+ than CD 133− cells, and more aggressive growth characteristics on serial
transplantation [67]. Pre-treatment with an inhibitor of cell-cycle checkpoints CHK1/CHK2 sensitized CD133+ cells to radiation, suggesting a role for heightened DNA repair in the relative resistance of CD133+ tumour stem cells to radiation-induced DNA damage. CD133+ cells also retain the ability to differentiate into more terminally-differentiated and less aggressive cells, a feature lacking in CD133- cells. Bone morphogenic protein (BMP), a potent differentiating molecule, led to significant differentiation of CD133+ cells and decreased tumour growth in an in vivo mouse model [68]. This suggests that the use of differentiating agents may play an important role against tumour stem cells which, which unlike CD 133- cells, retain important differentiation capacity – presumably because different triggering leads to the loss of neoplastic potential. Another area of fertile discovery is the role of epigenetic regulation in the control of stem cell maintenance, self-renewal and differentiation. Transcriptional changes that result from the pharmacological reversal of epigenetic gene silencing, typically with either a histone deacetylase inhibitor or a demethylating agent, allows for the identification of epigenetically-regulated genes on a genome-wide scale using molecular profiling techniques [69, 70]. Recent large-scale genomic studies have confirmed the importance of epigenetic gene silencing in the biological behaviour of GBM, including response to therapy [69, 71, 72]. Using a similar approach, several investigators have identified key genes which are epigenetically regulated during differentiation in neural stem cells and have been implicated in neoplasia [73–75]. Epigenetic modifications make attractive therapeutic targets as they are readily reversed and the underlying DNA sequence is not altered. Taken together, these studies highlight the importance of targeting therapies to the CD 133+ sub-population of cells which can exploit the tumour stem cell properties and vulnerabilities important in the effective therapeutic treatment of GBM.

The fraction of cells in a GBM tumour that express CD133 has been observed to be as little as 1% in low-grade tumours and as high as 30% for high-grade glioblastoma. However, in any particular grade, such as glioblastoma, there is high variability in this fraction [65]. Interestingly, time to progression and overall survival are not greatly affected by extent of resection in GBM patients. Despite the removal of the bulk tumour, consisting mostly of CD133- cells, the tumour recurs within 1 year on average. Thus, it is not yet clear whether the fraction of CD133+ cells in the bulk tumour will prove to be useful as a prognostic marker. It could be that the important neoplastic-associated stem cells constitute only a small fraction of the total CD133+ cells. Assuming that CD133+ cells (or a sub-fraction) are the biologically active tumour stem cells responsible for tumour recurrence, identifying and targeting the location of these cells in vivo, within the tumour or in the surrounding brain after tumour resection, will be of great importance. A recent study indicates that tumour stem cells exist in the 'perivascular niche', and that endothelial cells potentiate the tumour stem cells capacity to initiate and maintain tumour growth [76]. Application of inhibitors of endothelial cells significantly depleted tumour blood vessels resulting in a dramatic reduction of tumour stem cells but not CD 133- cells in the bulk tumour. This suggests that targeting the unique vulnerabilities of tumour stem cell populations should include further characterization of the micro-environment that support and sustains tumour stem cell self-renewal and differentiation. Large-scale molecular profiling of purified sub-populations of tumour cells from GBM will play a critical role in further defining these molecular targets [77].

**Molecular profiling of breast cancer stem cells**

CSCs have also been identified and characterized for breast tumours, a much more common malignancy than brain tumours. Breast cancer tumours contain a sub-population of highly tumourigenic cells characterized by CD44 expression and no or low CD24 expression (CD44+CD24-low), as demonstrated by their distinguishing capability to generate tumours in immunodeficient mice [3]. These CD44+CD24-low cells have also been demonstrated to have the capacity to give rise to the various cell types characteristic of the bulk tumour. As with CD133+ cells in brain tumours, the CD44+CD24-low cells in breast tumours contain the population of CSCs, but may not uniquely identify them. Indeed, additional cell-surface markers may further distinguish the tumourigenic breast cell population. For example, it has been shown that among CD44+CD24-low cells, those that were also ESA had enriched tumour generating potential compared with those that were ESA+. Thus, one hypothesis would be that ESA+CD44+CD24-low cells may represent a purer
population of a subset of the CSCs [3]. This interesting question for all different tumour stem cells is the extent to which neoplastic stem cell population may be purified by additional cell-surface markers. Also of interest is that CD44⁺CD24⁻/low cells generate additional CD44⁺CD24⁻/low cells as well as phenotypically distinct cells, providing evidence that breast CSCs have the hallmark stem cell characteristics of self-renewal and the capacity to generate heterogeneous populations.

It has been shown that these tumourigenic breast cancer cells can propagate in culture and maintain properties of normal human mammary gland stem/progenitor cells that have the distinct ability to grow in selective culture conditions as non-adherent spherical clusters of cells [78, 79]. Most importantly, after in vitro culturing these cells were found to remain CD44⁺CD24⁻/low and maintain their ability to generate tumours in immunodeficient mice with as few as 1000 injected cells [78, 79]. Thus, there is strong evidence that breast CSCs have indeed been isolated and can even be propagated in culture while maintaining their cancer stem cell characteristics.

Machine-learning from global gene expression data has been used to identify a gene signature from tumourigenic breast-cancer cells that is associated with both overall and metastasis-free survival in patients with breast cancer [24]. This signature was found through comparing the gene-expression profiles of the CD44⁺CD24⁻/low breast CSCs, which demonstrate enriched invasiveness [80], with normal breast epithelium. The differentially expressed genes between these two groups were then used to generate a 186-gene signature for ‘invasiveness’ that showed a significant association with overall and metastasis-free survival of patients with breast cancer [24]. This invasiveness signature was then also shown to be associated with prognosis in other cancers, including medulloblastoma, lung cancer and prostate cancer, demonstrating the general features at the gene expression level of invasiveness. It will be interesting to convert these tumourigenic gene signatures into their corresponding biological networks so as to begin to understand the unique and shared features of CSCs.

Emerging technologies for proteome characterization

While transcriptomic approaches have proven useful for identifying informative molecular signatures for cancers and CSCs, proteomic characterizations have lagged behind. The reason for this is clearly the more difficult challenge of measuring proteins compared to transcripts. Emerging proteomics technologies hold the promise of greatly improving our ability to make detailed assessments of protein-based molecular signatures, similar to those that have had success thus far for gene expression. One key advantage of protein signatures relative to gene expression is that proteins can be found in the blood and other accessible bodily fluids more readily due to slower degradation rates than their mRNA counterparts. Thus, protein signatures represent an important class of molecular signature for disease diagnosis. For CSCs, these approaches will help to elucidate the differences between the proteome of CSCs relative to their non-tumourigenic counterparts. Ultimately, the reconstruction of protein and gene regulatory networks of CSCs holds the promise of rationally-designed therapeutics for personalized medicine by allowing the therapeutic chosen to be governed by informative molecular signatures measured from each patient at the time of diagnosis. A few promising proteomics technologies are described below (Fig. 1).

Isobaric tagging for relative and absolute quantitation (iTRAQ)

Stable isotope labelling enables the quantitative analysis of protein concentrations through mass spectrometry (MS). One state-of-the-art technique for quantitative MS is iTRAQ [81], which uses stable isotope labelling of proteolytic peptides. This technique modifies primary amino acid groups of peptides by linking a mass balance group (carbonyl group), and a reporter group (based on N-methylpiperazine) by forming an amide bond. The iTRAQ reagents are designed to be isobaric, which enables differentially labelled peptides to appear as single peaks in MS, which is important for reducing peak overlapping in the MS scans. When MS/MS is used for analysis with iTRAQ-tagged peptides, the mass balancing carbonyl moiety is released as a neutral fragment, thereby liberating isotope-encoded reporter ions that provide relative quantitative information on protein abundance. Because four different iTRAQ reagents are currently available, comparative analysis of a set of two to four samples is feasible.
within a single MS run [82]. The, iTRAQ technology represents the state-of-the-art in quantitative proteomics and represents a promising technology for using proteomics to differentiate key differences in protein networks of CSCs from normal stem cells or other cells in the tumour.

**Glyco-peptide capture**

MS-based methods will allow for the identification of proteins spanning approximately three orders of magnitude in concentration from a given sample. Therefore, methods that can select specified fractions of the proteome are important for simplifying the sample sufficiently to identify the proteins of interest. One recently developed approach is the shotgun glycopeptides capture approach [83]. This approach selects for N-linked or O-linked glycosylated peptides, which are enriched for secreted proteins and cell-surface markers. Thus, this approach could be used to identify candidates for unique cell-surface markers for CSCs that differ from the bulk tumour by comparing the glyco-captured proteomes from both sample sets.

**Antibody microarrays using surface plasmon resonance imagining (SPRI)**

Protein chip methods hold potential for broad quantitative screens of proteins, and a variety of techniques have been developed based on antibody binding [84–86]. Antibody microarrays have been used for biomarker discovery and protein profiling of serum
from patients with prostate, lung, pancreas and bladder cancer [87–90]. One emerging approach with tremendous promise is SPRI [82, 91, 92], which enables real-time, label-free measurement of protein expression. SPR-based chips have a detection sensitivity of 10–100 times less than ELISA [82], but have a spatial resolution down to approximately 4 µm [93]. It is thus possible to print up to 800 unique antibodies on Lumera Nanocapture Gold™ microarray slides and monitor the abundance of the target proteins in real time [82]. Because the same slide can be regenerated for reuse many times (Z. Hu, C. Lausted, unpublished observations), this means that the approach has the capacity necessary to screen through hundreds of patient samples. Thus, this approach holds tremendous promise in the case of CSCs to be able to screen through large numbers of proteins, including secreted proteins and cell-surface markers and not only measure their presence, but also abundance and the dynamics of their binding. The limitation of this technique is its dependence on the affinity and specificity of the antibodies it employs for detection – cross re-activities in complex protein mixtures (like blood) can pose significant problems.

**DNA-encoded antibody libraries (DEAL)**

One recently developed technique that offers great potential for analyses of CSCs is DEAL, which enables cell localization and single-cell measurements of protein, RNA, and single-stranded DNA simultaneously on a single chip [94]. DEAL is a highly sensitive measurement technique, with a reported detection limit of 10 fM for the protein IL-2-150 times more sensitive than ELISA. This sensitivity can be applied to the isolation of rare cells based on combinations of cell-surface markers, enabling the isolation and addressing of individual CSCs. DEAL can also be used to make single cell measurements of secreted proteins from each of these isolated single cells. Thus, DEAL offers superb sensitivity and the ability to perform spatially multiplexed detection for characterization of CSCs.

**Biomolecular networks in cancer**

Systems approaches to cancer [95–99] require not only the identification of the key components of a system through global analyses, but also information about how these components interact in biological networks. Network models of multiple types have been applied to cancer systems. The most commonly applied to cancer are interaction networks, including protein–protein interaction networks, protein-DNA interaction networks and so forth. Gene expression data can be used to identify differentially expressed genes in which can then be visualized on interaction networks, as has been done for lung cancer [100]. Various properties of these networks have been studied [99], with reported findings including, for example, the enrichment of cancer-related genes among the ‘hubs’ of the networks. While these interaction networks are very useful tools for visualizing large data sets, they are not computable, predictive network models, which are those that hold the most promise for predictive medicine and drug development. Predictive models stemming from mathematical descriptions of biochemical reaction networks and statistical influence models should prove highly useful [101].

Another area of network modelling that should prove very beneficial in research of cancer and CSCs is that of metabolic networks. Key metabolic differences have been shown to exist in cancers which can be exploited using Positron Emission Tomography (PET) to do in vivo imaging of tumours [102] and even to predict treatment response [103, 104]. If key metabolic differences can be found between CSCs and the rest of the tumour, such approaches could potentially even be used to identify the location of cancer stem cell populations in vivo. One enabling resource for large-scale quantitative modelling of metabolic networks in cancer is the recent stoichiometric reconstruction of known human metabolism at the genome-scale [105]. With this global reconstruction, gene expression and other data can be used to create initial models of the genome-scale metabolic networks of a variety of human cell types, including for CSCs. These biochemical reaction networks can be used to make numerous quantitative simulations that have been shown previously to match well with experimental data in model organisms [106]. These successes with model organisms have also been extended to models of simple human systems such as the erythrocyte [107, 108] and mitochondria [109], with the global metabolic reconstruction poised to allow for larger human metabolic networks to now be modelled.
These studies may well provide insights into the unique metabolic features of cancer cells – allowing one to identify both metabolic features that are shared among cancer cells and features that are unique to individual types of cancer.

More detailed dynamic models of specific biochemical networks in cancer have been made for important signalling networks in cancer, leading to insightful biological observations for, among many others, the NF-κB signalling network [110–112]. As isolated cancer stem cell populations become better characterized, it will be possible to model these systems to identify differences in their regulation in CSCs and further identify possible therapeutic targets. Dynamic simulations of large-scale signalling networks in cancer cells has also been performed [113].

Large amounts of high-throughput data (i.e. transcriptomes) can also be used to infer networks that can explain statistical dependencies seen in the data, indicate candidate novel interacting partners, and quantitatively predict the gene expression resulting from knockouts or environmental perturbation. For model systems, such approaches are now being successfully applied at the genome-scale for gene-regulatory networks [114, 115]. Such approaches are now also being applied to mammalian systems as was done for normal and cancerous B cells with the development of an algorithm called Reconstruction of Accurate Cellular Networks (ARACNe) [116]. As cancer stem cell populations are profiled extensively, these same approaches will be useful to identify predictive networks for CSCs. Comparing these networks to those in normal stem cells and other tumour cells should prove highly informative for identifying drug targets unique to the cancer stem cell population of interest. By generating networks of CSCs in particular and comparing them with networks of normal stem/progenitor cells we should be able to greatly enhance our understanding of what could lead to these cells becoming cancerous.

Computational modelling and systems approaches will be key to catalyzing the future of drug discovery [117, 118], and drug discovery focused specifically on CSCs offers tremendous promise for advancing cancer therapies. Thus, computational modelling of cancer stem cell networks to identify potential therapeutic targets and to predict the effect of drug-induced perturbations is critical to this field moving forward.

**Perspective and concluding remarks**

The identification and prospective isolation of CSCs from leukaemia and a number of solid tumours has spawned a new paradigm in cancer research. From the perspective of systems biology – with the goal of predictive, preventive, personalized, and participatory (P4) medicine – we envision increasingly global assessment of CSCs and their microenvironments (niche) at the level of complete transcriptome, proteome and epigenome, using empowering new high-throughput technologies. The resulting gene expression profile signatures of cancer stem cell would serve as more accurate indicative for cancer diagnosis and prognosis. Emerging proteomic technologies employing MS and protein chip platforms would allow for identification of better cell-surface markers and their interaction with the resident stem cell niche and potential diagnostic markers from both body fluids and tumour tissues. Incorporating these data into biological networks will provide fundamet insights into the biology of CSCs and their abilities for renewal and differentiation. These combined efforts will ultimately lead to new therapeutic strategy specifically targeting CSCs for unprecedented personalized cancer therapy.

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