Aipl1 is required for cone photoreceptor function and survival through the stability of Pde6c and Gc3 in zebrafish

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Genetic mutations in aryl hydrocarbon receptor interacting protein-like 1 (AIPL1) cause photoreceptor degeneration associated with Leber congenital amaurosis 4 (LCA4) in human patients. Here we report retinal phenotypes of a zebrafish aipl1 mutant, gold rush (gosh). In zebrafish, there are two aipl1 genes, aipl1a and aipl1b, which are expressed mainly in rods and cones, respectively. The gosh mutant gene encodes cone-specific aipl1, aipl1b. Cone photoreceptors undergo progressive degeneration in the gosh mutant, indicating that aipl1b is required for cone survival. Furthermore, the cone-specific subunit of cGMP phosphodiesterase 6 (Pde6c) is markedly decreased in the gosh mutant, and the gosh mutation genetically interacts with zebrafish pde6c mutation eclipse (els). These data suggest that Aipl1 is required for Pde6c stability and function. In addition to Pde6c, we found that zebrafish cone-specific guanylate cyclase, zGc3, is also decreased in the gosh and els mutants. Furthermore, zGc3 knockdown embryos showed a marked reduction in Pde6c. These observations illustrate the interdependence of cGMP metabolism regulators between Aipl1, Pde6c, and Gc3 in photoreceptors.

In humans, genetic mutations of aryl hydrocarbon receptor interacting protein-like 1 (AIPL1) are linked to Leber congenital amaurosis 4 (LCA4), one of the inherited retinopathies1–4. LCA accounts for nearly 5% of all retinal dystrophies5. In general, patients who suffer from LCA are congenitally blind. Including AIPL1, 24 genes are associated with LCA (see the https://sph.uth.edu/Retnet). Typical symptoms of LCA include a progressive loss of photoreceptors, markedly reduced or no response of photoreceptors to light such as the electroretinogram (ERG) and nystagmus. Genetic mutations of the human AIPL1 gene cause LCA, and in certain mutations result in dominant cone-rod dystrophy and juvenile retinitis pigmentosa12,6. However, mechanisms that underlie such diverse forms of retinal degeneration due to AIPL1 mutations, are not fully understood.

Retinal phenotypes in mice with an Aipl1 mutation were studied as an animal model for human LCA4. As in LCA4 patients, Aipl1 null mutant mice show rapid degeneration of rod and cone photoreceptors in early postnatal stages7–9. Analyses of Aipl1 mutant mice revealed that AIPL1 is required for maintenance of cGMP phosphodiesterase 6 (PDE6) α and β subunits (also referred to as PDE6a and PDE6b), which mediate rod-specific phototransduction. In Aipl1 mutant mice combined with the knockdown of neural retina leucine zipper (NRL), almost all retinal photoreceptors are specified as cones10, which fail to mediate cone phototransduction via reduction of a cone-specific PDE6c′ (also known as PDE6c) and undergo degeneration11. Furthermore, Aipl1 mutant mice expressing human AIPL1 under the control of the rod-specific NRL promoter (Aipl1−/−;Tg[NRL:AIPL1]) showed defects in cone phototransduction and maintenance12. Even though both Aipl1−/−;Nrl−/− and Aipl1−/−;Tg[NRL:AIPL1] retinas provide artificially cone-dominated environments, these observations exclude the possibility that cone loss in Aipl1 mutant mice results entirely from the loss of rods. Rather, they suggest that AIPL1 is cell-autonomously required for cone function and survival.

Further biochemical analyses revealed that, in the absence of Aipl1, rod and cone PDE6 subunits are synthesized normally, but are degraded through the ubiquitin-proteasome system11,13. AIPL1 regulates folding of rod and cone PDE6 subunits as a chaperone, and catalyzes their prenylation for membrane anchoring and assembly13. However, it remains to be determined whether AIPL1 is required only for PDE6, and whether retinopathies in LCA4 are due solely to PDE6 dysfunction.
Photoreceptor degeneration is associated with genetic mutations in components of the phototransduction cascade, including rhodopsin, PDE6b, guanylate cyclase activating proteins (GCAPs), and cyclic nucleotide-gated (CNG) channels. In the photoreceptor, under the dark condition, cGMP concentration is high, and high cGMP levels open CNG channels on plasma membranes to maintain a steady influx of Na⁺ and Ca²⁺ ions. In the light condition, PDE6 is activated to hydrolyze cGMP, resulting in closure of CNG channels. Arrest of cation influx hyperpolarizes photoreceptors. Since GCAPs are Ca²⁺ binding proteins and Ca²⁺ binding to their EF-hand motifs inhibits GCAP-medi­ated activation of guanylate cyclases (GC), decreased intracellular Ca²⁺ concentration caused by closure of CNG channels activates GC. Dysfunctions of PDE6, GCAPs, and CNG channels are expected to increase cGMP levels in photoreceptors, which could subsequently trigger photoreceptor cell death. However, it is unknown whether an elevated cGMP concentration induces photoreceptor cell death, and if so, what signaling pathway underlies this pathology.

Zebrafish are an animal model for research on human diseases. Moreover, zebrafish are ideal for studying cone photoreceptors since they have cone-rich retinas, and cone visual acuity can be evaluated by an optokinetic response (OKR) at 5 day-post-fertilization (dpf). Previously we isolated the zebrafish eclipse (els) mutant, which shows no OKR, and found that the els gene encodes Pde6c. The els mutant shows a progressive degeneration of cones, suggesting that dysfunction of cone Pde6 induces cone degeneration in zebrafish. Another group identified the zebrafish gold rush (gosh) mutant, which shows no OKR, and no optomotor response (ORM) [12]. Here, we investigated retinal phenotypes of the gosh mutant, in which cones undergo progressive degeneration. We found that the gosh gene encodes cone-specific Aipl1, suggesting that gosh mutant zebrafish provide a good model for human LCA4. Further genetic and biochemical analyses revealed interdependence between Aipl1, Pde6c, and Gc3, which supports cone photoreceptor function and survival in zebrafish.

**Results**

**Zebrafish gosh mutant shows defects in cone opsin transport and structural integrity of subcellular photoreceptors organs.** gosh was identified as a mutant that shows no visual behavior due to photoreceptor depletion [13]. However, retinal phenotypes of gosh mutant were not investigated. Here, we examined retinal phenotypes of the gosh mutant histologically. Compared with wild-type retinas, the outer nuclear layer (ONL) was thinner; photoreceptors lost their columnar shape (becoming round), and the outer segment (OS) was abnormally shorter at 7 dpf (Fig. 1a). We examined cell number of the ONL, inner nuclear layer (INL) and retinal ganglion cell layer (GCL) within 75 μm along the line parallel to each layer in wild-type and gosh mutant central retinas (Fig. 1a). Only, ONL cells were significantly less numerous in gosh mutants, suggesting photoreceptor-specific defects.

Next, to examine whether all classes of cones and rods are maintained in the gosh mutant, we examined expression of cone opsins (green-, red-, blue- and UV-sensitive opsins) and rhodopsin. In wild type at 7 dpf, cone opsins and rhodopsin are expressed and localized to the OS (Fig. 1b). In the gosh mutant, opsin and rhodopsin expression was detected in the OS. However, the opsin-positive area was reduced for green, red and UV opsins, while there was no difference in the areas expressing blue opsin and rhodopsin (Fig. 1b). Furthermore, opsin expression was often not restricted to the OS, but abnormally spread to the basal region (arrows in Fig. 1b). Thus, transport of cone opsins to the OS is compromised in the gosh mutant.

Next, we examined retinal phenotypes at 12 weeks-post-fertilization (wpf). Semi-thin sections of wild-type retinas enabled us to distinguish rods and cones according to their nuclear position and shape. Rod nuclei had small, round shapes, and formed a 3–4 cell layer above the outer plexiform layer (OPL) (“rod” in Fig. 2a). The cone layer consists of four types of cones in zebrafish, and is located above the rod nuclear layer (“cone” in Fig. 2a). Rods are elongated through the cone layer and their OS regions are located in the outer-most retinal region that associates with pigmented epithelium (“OS” in Fig. 2a). In the gosh mutant, almost all nuclei of the ONL showed rod-like shapes, and formed a 3–4-cell layer in the central retina. Cone-like photoreceptors were observed as a single row (Fig. 2a). In gosh mutant CMZs, both rods and cones formed, and showed relatively normal columnar shapes (Fig. 2a). However, green opsin expression was mislocalized in gosh mutant peripheral retinas (Fig. 2a), while rhodopsin expression was maintained. Thus, expression of visual pigments is more severely compromised in cones than in rods in the gosh mutant.

To examine how embryonic photoreceptor cell death influences later retinal development, we examined gosh mutant retinas at 4 and 12 weeks-post-fertilization (wpf). At 4 wpf we examined expression of green opsin and rhodopsin, counterstaining with zpr1 antibody, which labels double cones. The ONL was markedly thinner in the central retinas of the gosh mutant than in wild type retinas (Fig. S1a). Zpr1 signals were detected, but green opsin expression was almost absent, while rhodopsin expression was reduced but still observed in the gosh mutant (Fig. S1a). In the ciliary marginal zone (CMZ) of wild-type retinas, retinal stem cells generated all retinal cell-types, including rods and cones (Fig. S1a). In gosh mutant CMZs, both cones and rods formed, and showed relatively normal columnar shapes (Fig. S1a). However, green opsin expression was mislocalized in gosh mutant peripheral retinas (Fig. S1a), while rhodopsin expression was maintained. Thus, expression of visual pigments is more severely compromised in cones than in rods in the gosh mutant.

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Figure 1. Photoreceptor phenotypes of the gosh mutant at 7 dpf. (a) (Upper) Sections of 7 dpf wild-type and gosh mutant retinas. In the gosh mutant, the ONL is thinner with abnormally shaped OS. However, INL and GCL appear to be normal. (Lower) Histogram of cell number of the ONL, INL, and GCL at 7 dpf in wild type (black) and gosh mutant (red). Numbers of nuclei within 75 μm length for each layer were counted (n = 3). Only ONL cells are significantly reduced in the gosh mutant (p = 0.026, students’ t-test). (b) (Upper) Labeling of wild-type and gosh mutant retinas with antibodies against green, red, blue, UV opsins, and rhodopsin (green), and zpr1 antibody (magenta). In the gosh mutant, the opsin-localized OS area is small, and opsins are often mislocalized to plasma membranes of cell bodies or synaptic areas (arrows). In contrast, rhodopsin localization to the OS seems to be normal. (Lower) Histogram of visual pigment-positive area at 7 dpf in wild type (left
were more numerous, while GCL cells were less so (Fig. 2a'). Interestingly, number of rods in gosh mutant central retinas was 74.3 ± 3.21/100 μm (n = 3), similar to wild-type (68.0 ± 2.0/100 μm, n = 3, p = 0.078), suggesting that most of the ONL cells lost in the gosh mutant were cones, and that rod cell number recovers by 12 wpf.

Next, to confirm cone-specific degeneration in the gosh mutant, we examined expression of green opsin and rhodopsin, counterstaining with the zpr1 antibody. In gosh mutant CMZs, both green opsin and rhodopsin expression were observed (Fig. 2b). However, in mutant central retinas, green opsin expression and zpr1-signals were completely absent, but most of the OS area was labeled with rhodopsin antibody (Fig. 2b). Thus, cones progressively degenerate, but rods are retained to occupy the ONL in the gosh mutant.

We further examined ultrastructure of 12 wpf gosh mutant retina using electron microscopy. In wild-type retinas, the ONL consists of rod nuclei, short single cones, long single cones, double cones and rod OS along the inner-outer axis (Fig. 2c). In the gosh mutant, the ONL consisted of rod nuclei and rod OS; cones were mostly absent, except for a discontinuous single row of cone-like cells (Fig. 2d). Thus, most photoreceptors in 12 wpf gosh mutant retinas are rods.

These cone-specific degeneration phenotypes are very similar to those of zebrafish pde6c mutants. In the els mutant, retinal regeneration was observed at 5 wpf14. In the zebrafish pde6c null allele, enhanced proliferation of INL cells occurs from 7 to 15 dpf16. We evaluated if retinal regeneration starts in the gosh mutant at 2 wpf, using a proliferating cell marker, proliferating cell nuclear antigen (PCNA)16. PCNA expression was observed in the CMZ, and also in the ONL and the INL, which correspond to rod progenitors and Müller cell-derived regenerating retinal progenitors in wild type. However, PCNA signals were reduced in the CMZ and in rod progenitor cells in gosh mutants (Fig. 51b). Although apoptosis is increased in the gosh mutant at 7 dpf, retinal regeneration is not active at 2 wpf. It is likely that rod number is recovered by retinal regeneration later than 2 wpf in the gosh mutant.

**gosh mutant gene encodes Aipl1b.** Photoreceptor phenotypes in the gosh mutant resemble those of the zebrafish els mutant. However, gosh mutant retinal phenotypes, including OKR defects, were rescued by a complementation test between gosh and els mutants (all embryos showed normal OKR: n = 30 for progeny of a cross between a gosh+/− male and an els+/− female, n = 20 for progeny of the cross between a gosh+/− male and an els+/− male), suggesting that the gosh mutation gene is different from pde6c. To understand the molecular basis of the gosh mutation, we cloned the gosh mutant gene.

First, we mapped the gosh mutant locus onto chromosomes. Analysis using SSLP markers restricted the gosh mutation between 41.72 and 42.85 Mb on chromosome 14. The zebrafish genomic database indicated that 7 genes were annotated in this genomic region. Among them, we focused on a gene encoding Aryl Hydrocarbon Receptor-interacting Protein-like 1 (Aipl1), namely CT027835.1 (Clone-Based Ensemble, ENSDARG00000014095.4, Ensemble Zv9 release 78), because aipl1 is the causative gene for LCA4 in humans and is required for PDE6 function in photoreceptors. Sequencing of this aipl1 cDNA from gosh mutant embryos revealed that a nonsense mutation occurred in Gln413 (Fig. 3a). Since another zebrafish Aipl1 gene was annotated on chromosome 15 (ENSDARG00000075067.3, Ensemble Zv9 release 78), we named this gene and the gosh mutant gene Aipl1a and Aipl1b, respectively. A phylogenetic tree for AIPL1 protein (Mega 6.06) revealed that zebrafish Aipl1a is more similar to human AIPL1 than to zebrafish Aipl1b (Fig. 3b). Amino acid identity between human AIPL1 and zebrafish Aipl1a was 57%, whereas 45% between human AIPL1 and zebrafish Aipl1b. Amino acid identity between zebrafish Aipl1a and Aipl1b was 39%. These data imply that the gosh mutation gene encodes Aipl1.

To confirm that photoreceptor phenotypes in the gosh mutant are caused only by the aipl1b mutation, we generated another aipl1b null allele, in which an extra 17 bp DNA fragment was inserted into the exon1 of the aipl1b gene (Fig. S2a), using the genome editing CRISPR/Cas9 system17. We confirmed that there were no OKR and photoreceptor degeneration phenotypes in the trans-heterozygotes for aipl1a and this new aipl1b mutant allele goshake (Fig. S2b–c).

**aipl1b mRNA is expressed in cone photoreceptors.** In human LCA4 patients and mouse AIPL1 mutants, both rods and cones degenerate. In humans, AIPL1 is expressed in rods and cones during embryonic development, but is restricted to rods in the adults18,19. To determine why cones exclusively degenerate in gosh mutant retinas, we examined expression of zebrafish aipl1a and aipl1b genes using whole-mount in situ hybridization. In zebrafish, aipl1b mRNA was detected at the 8-cell stage, suggesting maternal expression. At 24 hpf, aipl1b mRNA was detected in the developing brain, but then expression began to be restricted to the retina (Fig. S3a). At 72 hpf, aipl1b mRNA was detected exclusively in both retinal and pineal photoreceptors, whereas aipl1a mRNA was expressed in the brain including the retina (Fig. 3c). Next, we examined mRNA expression of aipl1a and
Figure 2. Cones are progressively eliminated in gosh mutants at 12 wpf. (a) (Upper) Sections of 12 wpf wild-type and gosh mutant retinas. In wild-type central retinas, rods and cones are distinguished. In gosh mutant central retinas, most ONL nuclei have a rod-like shapes, except for a single row of short single cones (red arrowheads), and an enlarged OS area. (a’) In the gosh mutant CMZ, both rods and cones are present. (Lower) Cell number of the ONL, INL, and GCL at 12 wpf in wild type (black) and the gosh mutant (red). Numbers of nuclei within 100 μm length for each layer were counted (n = 3). The ONL, INL, and GCL cells were reduced in the central retina of the gosh mutant. In the peripheral retina, ONL and INL cells was increased, but GCL cells
were decreased in the gosh mutant. Students’ t-test: \(*p < 0.05; **p < 0.01. (b) Labeling of 12 wpf wild-type and gosh mutant retinas with anti-green opsin and rhodopsin antibodies (green), and zpr1 antibody (magenta). In wild-type central retina, green opsin and rhodopsin are located in cone and rod OSs, respectively. However, in the gosh mutant, green opsin is absent, while most of the OS is labeled with rhodopsin. (b) In the gosh mutant CMZ, both green opsin and rhodopsin are detected, although green opsin is mislocalized in some cells (arrows). A dotted line indicates the interface between the ONL and the OPL. (c) Twelve wpf wild-type ONL, which consists of small, compact rod nuclei, short and long single cones, and double cones (left), rod OS (right upper), and synapses (arrows show ribbon, right bottom). An arrowhead indicates the outer limiting membrane. (d) Twelve wpf gosh mutant ONL, which consists of rod nuclei and single row of cone-like cells (arrows, left), rod OS (right upper), and synapses (a white arrow shows the synaptic ribbon, right bottom). The outer limiting membrane is maintained at the outer border of rod nuclei (arrowhead). OS, the outer segment; INL, inner nuclear layer; IPL, inner plexiform layer; OPL, outer plexiform layer; olm, outer limiting membrane; ros, rod outer segment; cos, cone outer segment; e, ellipsoid; cc, connecting cilium.

**aipl1b** in adult retina. Both genes were expressed in the ONL. aipl1b mRNA was expressed in all types of cones, whereas aipl1a mRNA was expressed in rods and probably UV cones (Fig. 3d). Double labeling of wild-type adult retinas with RNA probes for UV opsin and aipl1a or aipl1b confirmed that both aipl1a and aipl1b mRNAs are expressed in UV cones (Fig. 3b). Thus, the two aipl1 genes are differentially expressed in zebrafish, and aipl1b mRNA is expressed exclusively in cones.

Next, we examined expression of aipl1a and aipl1b mRNAs in gosh mutant retinas. At 72 hpf, aipl1b mRNA expression was absent in gosh mutant photoreceptors (Fig. 3e). In contrast, aipl1a mRNA expression was detected in both wild-type and gosh mutant embryos in the ventral ONL, where rod photoreceptors start to differentiate at 72 hpf (Fig. 3e). Thus, only aipl1b mRNA expression is reduced in the gosh mutant, probably due to the nonsense mutation-mediated mRNA decay\(^6,8\). Cone-specific expression of Aipl1b explains cone opsin mislocalization and later cone degeneration in the gosh mutant.

**Pde6c protein is markedly decreased in the gosh mutant.** Aipl1 functions as a chaperone for rod and cone PDE6, and promotes its protein stability, its membrane anchoring via prenylation, and PDE6 subunit assembly\(^7,8,11\). Indeed, retinal phenotypes in the gosh mutants are similar to those of zebrafish pde6c mutants. To clarify whether gosh mutant phenotypes link to PDE6 dysfunction, we examined Pde6c levels. Here we used 7 dpf embryos, because at this stage, gosh mutants still retain photoreceptors. Western blots of wild-type zebrafish retinas using anti-Pde6c antibody detected a band of \(~100\)kDa, corresponding to the molecular weight of Pde6c. However, in the gosh mutant, the band for Pde6c was undetectable (Fig. 4a). Semi-quantitative PCR indicated that the mRNA level of Pde6c did not differ between wild type and the gosh mutant (Fig. 4b), suggesting that protein synthesis or stability of Pde6c is compromised in the absence of Aipl1b.

**The gosh mutation genetically interacts with the els mutation.** To examine whether Aipl1b directly interacts with Pde6c in vivo, we examined genetic interactions between gosh and els mutations. We compared photoreceptor degeneration phenotypes among 7 dpf retinas of the following three genotypes: els\(^+/+\)/gosh\(^+/+\), els\(^+/−\)/gosh\(^+/+\), and els\(^+/−\)/gosh\(^+/−\), by immuno-labeling with zpr-1 antibody (Fig. 4c). The fraction of the zpr1-positive area relative to total retinal area in els\(^+/−\)/gosh\(^+/+\) retinas was significantly smaller than that of els\(^+/−\)/gosh\(^+/−\) retinas, indicating that a half dose of Aipl1b activity enhanced photoreceptor degeneration in the els mutant. We also confirmed that aipl1a mRNA level in the gosh or els mutant was similar to that of wild type, excluding the possibility that aipl1a mRNA expression is modified by some complementary effect or genetic feedback mechanism in both mutants (Fig. 5a). These data suggest that Aipl1b directly interacts with Pde6c in vivo.

**cGMP concentration is not drastically increased at 4 and 7 dpf in retinal photoreceptors of gosh and els mutants.** It is likely that cGMP concentration is elevated in both gosh and els mutant photoreceptors because of severe reduction of Pde6c. It was reported that cGMP levels are elevated in a non-sense mutation allele of zebrafish pde6c, pde6c\(^wel\), at 4 dpf\(^21\) and in Aipl1 knockdown mice\(^7\). However, cGMP concentration was also reportedly decreased in AIPL1 knockdown mice\(^6\) and in AIPL1;NRL double mutant mice\(^2\). We examined cGMP concentration in wild-type, gosh, and els mutant retinas at 7 dpf, using a specific antibody against formaldehyde-fixed cGMP\(^22,23\). In light-adapted conditions, no antibody signal was observed in wild-type photoreceptors (Fig. 5a), suggesting that this antibody cannot detect background levels of cGMP. Surprisingly, we did not detect signals in either gosh or els mutant retinas in light-adapted conditions, although we sometimes observed a very strong signal in only a few photoreceptors near the CMZ (Fig. 5a). Since such positive cells overlapped with a cone marker, zpr1, these high cGMP-leveled cells are cones. In contrast to retina, we observed very strong signals in gosh and els mutant pineal glands, while no cGMP signal was observed in wild-type pineal gland. Thus, cGMP concentration is increased in pineal photoreceptors, but not in retinal photoreceptors, in both gosh and els mutants.

To investigate whether cGMP is elevated transiently during early photoreceptor differentiation but declines to low levels by 7 dpf, we labelled 4 dpf gosh and els mutant retinas with anti-cGMP and zpr1 antibodies. As at 7 dpf, we could not detect antibody signals in photoreceptors, although we occasionally observed strong signals in some photoreceptors (Fig. S5). cGMP level is not increased early in photoreceptor differentiation in either gosh or els mutants.
**Gc3 is markedly reduced in both gosh and els mutant photoreceptors.** cGMP concentration is determined by the balance between PDE6 and GC activities in photoreceptors. In zebrafish, there are three retinal Gcs, zGc1–3. zGc3 is expressed in all cones, whereas zGc1–2 are expressed in rods and UV cones\(^2\). We examined zGc3 levels in gosh and els mutants by western blotting using a specific antibody against zGc3\(^3\). In wild type, the zGc3 antibody cross-reacted with a band of 120 kDa, which corresponds to the molecular weight of zGc3 (Fig. 5b). In contrast, the band was completely absent in gosh and els mutants. Semi-quantitative PCR revealed that zGc3 transcriptional level of gosh and els mutants is similar to that of wild type (Fig. 5c). Thus, Aipl1 and Pde6c are required for protein synthesis or stability of zGc3, suggesting interdependence between these cGMP metabolic enzymes in zebrafish photoreceptors.
Pde6c is markedly reduced in the absence of Gc3. In mice, deletion of GC1 results in no PDE6c, Transducin, and GRK1 in cone photoreceptors, suggesting that GC is required for stability of phototransduction molecules. To examine whether zGc3 is required for Pde6c protein stability, we knocked-down zGc3 by injecting morpholino antisense oligos, and performed western blots of 4 dpf zebrafish heads using the anti-Pde6c antibody. First we confirmed that both ATG- and splicing morpholinos effectively inhibited zGc3 protein production (Fig. 6a). We found that Pde6c level was drastically reduced in both ATG- and splicing-zGc3 morphant embryos, whereas Pde6c was detected at normal levels in control morpholino-injected embryos (Fig. 6b). These data suggest interdependence between the regulators of cGMP metabolism, Pde6c and zGc3, in zebrafish cone photoreceptors.

Discussion
In this study, we investigated a zebrafish gosh mutant that was originally identified as a visual behavior-defective mutant. During embryonic stages, mutant photoreceptors show abnormal organelle shapes, defects in opsin transport, and undergo apoptosis. During adult stages, most cone photoreceptors are eliminated, but rods are retained at normal numbers and with normal structures. We found that the gosh mutant gene encodes cone-specific aipl1b, suggesting that Aipl1b is required for phototransduction and survival of cone photoreceptors.

Genetic mutations of AIPL1 cause congenital blindness, as in LCA4 in humans and rapid degeneration of rods and cones in the early postnatal period in mice. Thus, photoreceptor degeneration phenotypes in the zebrafish gosh mutant seem to be consistent with those caused by human and mouse AIPL1 mutations. In the
zebrafish gosh mutant, photoreceptors show abnormal proportions of subcellular structures and increased cell death at 7 dpf, suggesting a possible role of Aipl1 in photoreceptor differentiation. For the future development of therapy for human LCA4 patients, it is important to address whether photoreceptor differentiation is compromised in the absence of Aipl1 activity. The zebrafish gosh mutant may provide a good model for studying this issue.

Figure 5. zGc3 expression is coupled to Aipl1 and Pde6c in zebrafish photoreceptors. (a) Labeling of wild-type, gosh, and els mutant retinas and pineal eyes at 7 dpf with anti-formaldehyde-fixed cGMP antibody (green) and zpr1 antibody (magenta). Control wild-type retinas show an undetectable level of cGMP. gosh and els mutant retinas show similar results, except that a few photoreceptors near the CMZ show very high accumulation of cGMP (arrow). On the other hand, pineal photoreceptors are positive for cGMP in gosh and els mutants, but not in wild type fish. (b) Western blot of wild-type, gosh and els mutant heads with anti-zGc3 antibody. A band of ~120 kDa is detected in wild-type heads, but disappears in both gosh and els mutant heads. (c) Semi-quantitative PCR of gc3 mRNA expression in wild-type, and gosh or els mutant embryos. mRNA levels are similar between wild-type, and gosh or els mutant embryos.
Photoreceptor phenotypes in the gosh mutant are similar to those of zebrafish pde6c mutants. AIPL1 promotes stability of PDE6c, and membrane anchoring via prenylation. We found that Pde6c is markedly decreased or absent in gosh mutants. However, its mRNA level is not affected. Thus, the absence of AIpl1 compromises Pde6c protein synthesis post-transcriptionally or protein stability. Furthermore, genetic interaction between the gosh and els mutations indicates that Aipl1b directly interacts with Pde6c in vivo. These data support the model that AIPL1 functions as a chaperonin for PDE6. Thus, the requirement of AIPL1 in PDE6 functions is conserved from fish to humans. We have no data indicating that Aipl1 is required for rod Pde6 functions in zebrafish, because aipl1a mutants have not been reported. It will be interesting to examine whether rod Pde6 levels are reduced in zebrafish aipl1a mutants.

It was reported that genetic mutations of phototransduction molecules, including PDE6b, GCAPs, and CNG channels, cause photoreceptor degeneration. In all these cases, cGMP accumulations are expected, and these may trigger photoreceptor degeneration. However, decreased cGMP concentrations were also reported in Aipl1 mutant mice. Thus, it is important to examine cGMP levels in gosh mutant photoreceptors. We found that cGMP concentration is not drastically elevated in retinal photoreceptors in light-adapted gosh and els mutants at 4 and 7 dpf. This is surprising because cGMP level are reportedly elevated in the zebrafish pde6c null mutant, pde6c<sup>−/−</sup>, at 4 dpf. This discrepancy may be due to differences in the anti-cGMP antibodies used, or allelic differences between pde6c<sup>−/−</sup> and our allele, els (pde6c<sup>el69</sup>). In the latter case, cGMP concentration may be increased in gosh and els mutants, but less than the threshold level that the antibody can detect. However, since zGc3 is almost absent in gosh and els mutants at 7 dpf, it is likely that this reduced zGc3 level suppresses chronic elevation of cGMP. In Aipl1<sup>−/−</sup> Nrl double knockout mice, PDE6c was not maintained, but cGMP level was reduced in dark-adapted conditions. In this double mutant, RetGC1 protein level is also reduced. Thus, the absence of PDE6c activity is directly linked to instability of GC proteins throughout vertebrate photoreceptors.

Both rod and cone PDE6 subunits are markedly decreased or absent in Gc1<sup>−/−</sup>Gc2<sup>−/−</sup> double mutant mice. Thus, reduction of GC activity causes PDE6 instability in mice. We found that Pde6c is reduced in zGc3 morphants at 4 dpf, suggesting interdependence between Pde6c and zGc3. At the moment, the molecular mechanism underlying the coupling of Pde6 and Gc3 maintenance in photoreceptors is unknown. One possibility is that intracellular transport of Pde6c and Gc3 is shared in zebrafish photoreceptors. PrBP/δ, also known as PDE6d, which was originally identified as a rod PDE6 subunit, regulates transport of rhodopsin kinase (GRK1) and PDE6c to the OS. Furthermore, both PDE6c and GRK1 were undetectable in the absence of GC1, raising the possibility that these proteins are transported to the OS with GC1-containing transport vesicles. However, RD3, mutations of which cause LCA12 in humans, binds to GC1 and GC2, and regulates their transport to the OS in mice. In RD3 knockdown mice, PDE6c levels are normal, suggesting that transport of PDE6c to the OS is independent of RD3, and probably different from that of GC1. In the latter case, coupling of PDE6c and GC3 maintenance may be mediated by a physiological condition, rather than by a direct interaction between them. It is important to address this topic in the future.

Another important question is how photoreceptor cell death is induced in the gosh mutant. Similar retinal phenotypes between gosh and els mutants suggest a common cell death mechanism, which is presumably linked to the absence of Pde6c. In the mouse Pde6b mutant, rod cell death is induced by apoptosis in a cell-autonomous manner, whereas cone cell death is induced cell-non-autonomously by necroptosis, which depends on receptor-interacting protein kinases 1 and 3 (RIPl and RIP3)<sup>29</sup>. Recently, it was reported that photoreceptor cell death in the zebrafish pde6c null mutant, pde6c<sup>−/−</sup>, depends on Rip3<sup>31</sup>. The authors showed that rip3 knockdown rescues cell death, and visual responses in zebrafish pde6c mutant. They speculated that rod Pde6ub subunits may be artificially expressed in pde6c mutant cones, where they mediate cone phototransduction even in the absence of Pde6c. However, we did not observe OKR in zebrafish gosh or els (pde6c<sup>−/−</sup>) mutant embryos injected with morpholino antisense of rip3 (data not shown). Furthermore, it was reported that cone cell death in zebrafish pde6c<sup>−/−</sup> mutants is induced cell-autonomously<sup>30</sup>, in contrast to the cell-non-autonomous necrototic cone cell death in gosh and els mutants. It is thus important to examine whether rod Pde6b subunits are expressed in cone cells.
death model\textsuperscript{31}. Further investigation is required to understand cell death mechanisms in zebrafish aipl1b and pde6c mutants.

The secondary loss of healthy neurons surrounding dead or dying neurons harboring primary genetic defects is known as bystander effect\textsuperscript{32}. Such a bystander effect was reported in human retinitis pigmentosa, in which rod photoreceptors firstly die, followed by later cone degeneration. However, cone-dystrophy diseases do not induce the secondary death of rods\textsuperscript{33}. In the gosh mutant, cones progressively degenerate. At the embryonic stages, rods are deformed and some of them are likely to degenerate. However, at the adult stages, rod number is recovered to a normal level. In the pde6c null mutant, pde6c\textsuperscript{w59}, rod degeneration is limited to areas with a low rod density\textsuperscript{34}. A cone-specific knockdown of Ran-binding-protein-2 (Ranbp2) in mice induces cone degeneration, but the secondary loss of rods only occurs when degenerating cones are present\textsuperscript{35}. Thus, the early transient rod degeneration in the gosh mutant may associate with the presence of degenerating cones, although its underlying mechanism is unknown.

Unlike mammals, zebrafish can regenerate a damaged retina\textsuperscript{16}. Although photoreceptors undergo apoptosis at 7 dpf in the gosh mutant, retinal regeneration is not initiated at 2 wpf. This is a bit surprising, because INL cell proliferation is elevated in the null allele of zebrafish pde6c mutant at 7 dpf\textsuperscript{16}. Although our western blot did not detect Pde6c protein in the gosh mutant, some residual Pde6c activity may delay retinal regeneration. Consistently, we observed enhanced INL cell proliferation in the gosh mutant retinas at 7 wpf (data not shown). Since such later regeneration can explain the early transient rod degeneration in the gosh mutant, it is important to investigate the interrelationship between cone degeneration and rod regeneration.

Material and Methods

Ethics statement. All zebrafish experiments were performed in accordance with the Animal Care and Use Program of Okinawa Institute of Science and Technology Graduate University (OIST), which is based on the Guide for the Care and Use of Laboratory Animals by the National Research Council of the National Academies and has been accredited by the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC International). All the experimental protocols were approved by the OIST Institutional Animal Care and Use Committee (Approval ID: 2014-83–86).

Fish. Zebrafish (\textit{Danio rerio}) were maintained according to standard procedures\textsuperscript{96}. Okinawa wild type (\textit{oki}) was used as a wild-type strain. WIK was used for mapping the gosh mutation. The gosh mutant was originally isolated in the screening of zebrafish visual mutants using a chemical mutagen, N-ethyl-N-nitrosourea (ENU)\textsuperscript{15}. A \textit{WIK} was used for mapping the \textit{gosh} mutation. The \textit{gosh} gene was identified in our previous screening of zebrafish visual mutants\textsuperscript{14}. \textit{gosh}\textsuperscript{66}, \textit{gosh}\textsuperscript{kn}, and \textit{els}\textsuperscript{w576} were used for our analyses. OKR was performed to identify visual mutants at 5–7 dpf following the published method\textsuperscript{37}. Larvae were partially immobilized in a petri dish containing methylcellulose. A drum with black and white vertical stripes (18°) was placed around the petri dish, and turned around at 10–20 rpm.

Mapping, and cloning of the gosh mutant gene. Mapping of the gosh mutation was conducted as described previously\textsuperscript{96}. Two polymorphic markers, CH174K20 and ZK519, were designed using genomic database information (Ensemble Zv9 release 78) at positions 41.72 and 42.85 Mb on chromosome 14, respectively. A zebrafish \textit{pde6c} mutant, \textit{els}, was identified in our previous screening of zebrafish visual mutants\textsuperscript{14}. \textit{gosh}\textsuperscript{66}, \textit{gosh}\textsuperscript{kn}, and \textit{els}\textsuperscript{w576} were used for our analyses. OKR was performed to identify visual mutants at 5–7 dpf following the published method\textsuperscript{37}. Larvae were partially immobilized in a petri dish containing methylcellulose. A drum with black and white vertical stripes (18°) was placed around the petri dish, and turned around at 10–20 rpm.

Isolation of a new aipl1b mutant allele using the CRISPR/Cas9 genome editing system. Guide RNA (gRNA) was designed using the webpage http://crispr.mit.edu/. A 20 bp sequence targeted to exon 1 of the \textit{aipl1b} gene, ACGTATCTGCTAAACCATCC, was selected as gRNA-1. A chimeric sgRNA, consisting of pT7-gRNA\textsuperscript{17} and the AIPL1b target sequence, was generated by \textit{in vitro} transcription using a MEGAscript T7 kit (Ambion), and purified with an RNeasy kit (Qiagen). For Cas9 preparation, we used pCS-nCas9n, a PCS2 plasmid construct that expresses nls-zCas9-nls\textsuperscript{17}. Capped mRNA encoding nls-zCas9-nls was synthesized using a mMESSAGE mMACHINE SP6 kit (Ambion). Poly(A) tailing was added with \textit{E. coli} Poly (A) Polymerase (NEB# M0276), and then purified by column purification (QIAprep, Qiagen). A mix of 30 ng/μL of sgRNA and 75 ng/μL nls-zCas9-nls RNA was injected into one-cell-stage zebrafish embryos. The F1 generation was produced by a pair-wise cross of the F0-injected adult fish. F1 fish in which deletion or insertion occurred at the target region of exon1 were identified by PCR amplification of the target region. By this means, we established a fish line that has a 17 bp insertion, as a new allele of the \textit{aipl1b} gene.

Morpholino antisense-mediated knockdown of zGc3. We used two morpholino antisense oligonucleotides for gc3 (GeneTools, Inc., Philomath, OR), which targeted the 5′-untranslated region (ATG), or splicing site (splicing), respectively. Standard morpholino was used as a control. Morpholino was dissolved in water at 0.25 mM and injected into wild-type embryos at the one-cell stage. MO sequences are shown below.

MO-ge3-ATG, 5′-TTGGTGAATATCAGAGACGAAGCGA-3′
MO-ge3-splicing, 5′-ATGATGAACACTGACACAACAAAT-3′
Standard MO, 5′-CCTCTTACCTCGTACATTAGAC-3′.
Histology. In situ hybridization, plastic sectioning, immunolabeling of cryosections and paraffin sections were performed as described previously[18]. Paraffin sections were pretreated with heat (120°C, 20 minutes, in 10 mM citrate buffer pH 6.0). zpr1 antibody (ZIRC, Eugene, Oregon; 1:100), anti-zebrafish rhodopsin (1:5000), red opsin, green opsin, blue opsin, and UV opsin (1:200–1000)[26] and PCNA (clone PC10, Sigma P8825; 1:200) were used. Antibody against formaldehyde-fixed cGMP[20] (1:1000) was used for immunolabeling. TUNEL was performed using an In Situ Cell Death Detection Kit (Roche) and carried out as previously described[14]. Nuclear staining was performed using 50 nM SYTOX Green (Molecular Probes) or 1 nM TOPRO3 (Molecular Probes). Images were scanned using a confocal laser scanning microscope (Carl Zeiss, LSM510 and LSM710). EM analyses were performed as previously described[14], and examined with an electron microscope (JEM 1230; Jeol Co., Ltd., Tokyo, Japan).

Western blots. Pools of 5 heads isolated from 4 or 7 dpf embryos were used for western blotting. Protein purification and SDS-PAGE were conducted as previously described[41]. Anti-Pde6c (Abcam; ab5660), zGc3[25], and histone H3 (Abcam; ab1791) antibodies were used at dilutions of 1:2000, 1:1000 and 1:50000, respectively. Signals were developed using horseradish peroxidase (HRP)-conjugated secondary antibodies (GE Healthcare Life Sciences, NA934VS, 1:5000) and ImmunoStar LD (Wako). Luminescence was quantified using a Fuji LAS 4000 image analyzer (Fuji Photo Film).

Semi-quantitative PCR. Total RNA was prepared from 7 dpf gosli and els mutant embryos. Complementary DNA strands were synthesized from total RNA using an AMV reverse transcriptase and used as a template for semi-quantitative PCR. cDNAs of pde6c, zGc3, aipl1a, and β-actin were amplified with the RNA LA PCR™ kit (TaKaRa Biochemicals), either with 20, 25 or 30 cycles; or 20, 30 and 40 cycles. The following primers were used.

Amounts were quantified using ImageJ (NIH).

pde6c forward, 5′-CTCCACCTGATCGAGGAAAAG-3′
pde6c reverse, 5′-GTAATGTCCTGGACACTTAATGG-3′
zGc3 forward, 5′-GGGACATGGTGAGGATGAACCT-3′
zGc3 reverse, 5′-GTTTCTCTGTAAGACCCCCATGC-3′
aipl1a forward, 5′-GGACTGTCACACGTTCACTT-3′
aipl1a reverse, 5′-GCCACGCTATAGAAAGGCTTCTT-3′
β-actin forward, 5′-GGATTTGTCGCTGAACATCAA-3′
β-actin reverse, 5′-GAAGGTGTCTGCTGGATAC-3′.

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gosh

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M.I., Y.N. and I.M. designed the experiments, analyzed and discussed the data. M.I. and I.M. prepared the manuscript. M.I. performed all the experiment. S.N., M.A. and E.O supported mapping of the gosh mutation, in situ hybridization, making expression constructs, respectively. All authors approved the manuscript.

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