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Lipopolysaccharide inhibits hepatic gluconeogenesis in rats: The role of immune cells

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ABSTRACT

Aims/Introduction: Bacterial septicemia has diverse clinical symptoms including severe hypoglycemia. However, sepsis-induced hypoglycemia has not yet been examined in detail. The aim of the present study was to investigate the mechanisms underlying hypoglycemia in sepsis.

Materials and Methods: We induced endotoxin shock in rats using lipopolysaccharide (LPS). After an intraperitoneal injection of LPS, we measured gluconeogenesis using the pyruvate tolerance test. The effects of LPS on glucose metabolism were investigated in perfused livers and isolated hepatocytes. Furthermore, its effects on the production of inflammatory cytokines were examined in isolated splenocytes. The interaction between splenocytes and hepatocytes in response to LPS was investigated in vitro using a co-culture of splenocytes and hepatocytes.

Results: In the pyruvate tolerance test, the pretreatment with LPS decreased gluconeogenesis. The in vivo pretreatment of rats with LPS did not inhibit glucose production in perfused livers. The in vitro treatment of isolated hepatocytes with LPS did not decrease hepatic gluconeogenesis. Although LPS increased the production of inflammatory cytokines (tumor necrosis factor-α, interferon-γ, interleukin-1β, interleukin-6 and interleukin-10) and nitric oxide in isolated splenocytes, only nitric oxide significantly inhibited gluconeogenesis in isolated hepatocytes. When splenocytes and hepatocytes were co-cultured in medium containing LPS, the messenger ribonucleic acid expression of glucose-6-phosphatase in hepatocytes was suppressed.

Conclusions: LPS reduced hepatic gluconeogenesis, at least in part, by stimulating the production of nitric oxide in splenocytes. This effect could contribute to the mechanisms responsible for septicemia-induced hypoglycemia.

INTRODUCTION

Sepsis is a systemic response caused by pathogenic microorganisms, and is accompanied by a number of clinical symptoms¹. These symptoms include endotoxin shock caused by lipopolysaccharide (LPS), a component of the outer membrane of Gram-negative bacteria. Septicemia causes not only hyperglycemia, but also hypoglycemia. Although the incidence of hypoglycemia is markedly lower than that of hyperglycemia, the mechanisms underlying hypoglycemia in sepsis must be considered. Endotoxic hypoglycemia plays an important role in predicting the severity and prognosis of sepsis².

Several studies examined the mechanisms responsible for the development of hypoglycemia in animals or humans with sepsis. LPS was shown to increase insulin sensitivity³, and inhibit hepatic gluconeogenesis⁴. To the best of our knowledge, the mechanism underlying hypoglycemia in sepsis has not yet been elucidated. LPS stimulates immune cells and increases the secretion of inflammatory cytokines, such as tumor necrosis factor-α.
Phosphoenolpyruvate carboxykinase (PEPCK) is an enzyme that catalyzes the rate-limiting step of gluconeogenesis, and glucagon increases its activity. TNF-α, a protein induced by LPS, inhibits glucagon-stimulated PEPCK gene expression and gluconeogenesis. LPS also increases hepatic insulin sensitivity, and accordingly, might exert hypoglycemic effects. In contrast, Sugita et al. previously reported that hyperglycemia and insulin resistance were induced by LPS. Thus, glucose metabolism during septic shock has not yet been examined in detail.

Macrophages activated by LPS generate nitric oxide (NO), a highly reactive molecule with a number of important biological actions. NO is synthesized by three different NO synthase (NOS) isoforms: inducible NOS (iNOS), endothelial NOS and neuronal NOS. iNOS is primarily expressed in immunocytes, and produces large quantities of NO when stimulated by LPS, leading to marked increases in plasma NO concentrations. Horton et al. showed that the addition of NO donors to incubation medium inhibited gluconeogenesis in isolated hepatocytes, which supports increases in plasma NO levels in vivo also inhibiting hepatic gluconeogenesis. We herein investigated the mechanisms underlying glucose dyshomeostasis in sepsis, and clarified whether LPS inhibits hepatic gluconeogenesis. Our results showed that LPS failed to exert any direct effects on gluconeogenesis in isolated hepatocytes or perfused livers. Furthermore, using isolated splenocytes, we measured the production of representative inflammatory cytokines (TNF-α, interferon [IFN]-γ, interleukin [IL]-1β, IL-6 and IL-10) and NO after stimulation with LPS. Our results showed that LPS increased the secretion of TNF-α, IFN-γ, IL-1β, IL-10 and NO from splenocytes. We measured glucose production in isolated hepatocytes after stimulations with TNF-α, IFN-γ, IL-1β, IL-6 and NO, and found that gluconeogenesis in primary hepatocytes was suppressed by the addition of sodium nitroprusside (SNP), a NO donor, but not by TNF-α, IFN-γ, IL-1β, IL-6 or IL-10. Our results showed that LPS suppressed the messenger ribonucleic acid (mRNA) expression levels of G6Pase, a master regulator of glucose production, in hepatocytes co-cultured with splenocytes. Therefore, LPS might inhibit hepatic gluconeogenesis, at least partly, by increasing the release of NO from the spleen, and does not appear to act directly on the liver.

METHODS

Animals

Male Wistar rats, weighing approximately 180 g (8 weeks-of-age), were purchased from Japan SLC, Inc. (Hamamatsu, Japan) and maintained in a temperature-controlled (25 ± 1°C) environment with a 12-h light/dark cycle. Rats had free access to standard laboratory chow and water, and were maintained and used in accordance with the Guidelines for Animal Experiments of Tezukayama Gakuin University.

Effects of LPS on blood glucose

Blood glucose levels were measured, followed by an intraperitoneal injection of saline (control) or LPS from Escherichia coli serotype O111:B4 (0.1–0.5 mg/kg; Sigma Aldrich, St. Louis, MO, USA). Blood glucose levels were measured 2, 4, 6 and 8 h after the LPS injection using the glucose oxidation method (glucose CII test; Wako, Osaka, Japan).

Pyrurate tolerance test

LPS from 0.1–0.5 mg/kg or saline (control) was given in a single intraperitoneal injection to 16-h fasted rats weighing approximately 250 g. After 4 h, glucose levels were measured in blood collected from the tail vein, and pyruvate (1 g/kg) was then injected intraperitoneally into rats. Blood was serially collected at 30-min intervals after the pyruvate injection. In NOS inhibitor pretreatment experiments, rats were injected intravenously with N(G)-monomethyl-L-arginine (L-NMMA; 10 mg/kg) 20 min before the intraperitoneal injection of LPS (0.5 mg/kg).

Preparation and incubation of hepatocytes and splenocytes

Hepatocytes were isolated from male Wistar rats fasted for 16 h by collagenase digestion as described previously. Isolated hepatocytes (>90% viable by the trypan blue exclusion test) were seeded on 24-well plastic plates (Corning, New York, NY, USA) at a density of 5.0 × 10⁵ cells per well. Cells were cultured in Krebs–Ringer buffer and incubated in 95% O₂/5% CO₂ at 37°C for 2 h in the presence of 0.24 mmol/L 3-isobutyl-1-methylxanthine and gluconeogenic precursors (10 mmol/L lactate and 1 mmol/L pyruvate) with/without LPS (0.02–20.0 μg/mL). After the incubation, medium was centrifuged at 50 g for 3 min to separate supernatants and pellets.

Splenocytes were harvested from the spleens of rats by homogenization. Crude cells were left to sink to the bottom of tubes for 1 min, and the supernatant was then centrifuged at 500 g for 3 min. Ammonium chloride was added to the precipitate in order to lyse red blood cells. Then, 30 s later, cells were washed twice with Dulbecco’s Modified Eagle’s Medium (DMEM). Isolated splenocytes were seeded on 10-cm plastic dishes at a density of 1.0 × 10⁶ cells/mL, and cultured in DMEM supplemented with 100 μmol/L 2-mercaptoethanol with/without LPS (0.01–1.0 μg/mL).

Gluconeogenesis in perfused rat livers

Rat liver perfusion was carried out with the flow-through method, as described previously. Details are shown in Appendix S1.

Effects of LPS on nitrate + nitrite, TNF-α, IFN-γ, IL-1β, IL-6 and IL-10 production in splenocytes

The concentrations of the LPS-induced cytokines, NO, IFN-γ, IL-1β, IL-6 and IL-10 in the culture medium of splenocytes

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Lipopolysaccharide and hypoglycemia

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were measured after 4, 8, 16, and 24 h. NO is hydrolyzed in medium, and this is followed by the formation of NO stable metabolites (nitrate + nitrite [NOx]). NOx concentrations were measured using the Griess method (NO2/NO3 Assay Kit-CII; Dojindo, Kumamoto, Japan). Enzyme-linked immunosorbent assay was carried out to measure the levels of TNF-α (IBL, Gunma, Japan), IFN-γ (R&D systems, Minneapolis, MN, USA), IL-1β (R&D systems), IL-6 (Arigobio, Hsinchu, Taiwan) and IL-10 (R&D systems).

**Effects of inflammatory cytokines on hepatic gluconeogenesis in vitro**

Primary hepatocytes were prepared as described above, and cultured with SNP, TNF-α, IFN-γ, IL-1β, IL-6 and IL-10. SNP was used as a NO donor. After a 2-h incubation, the concentration of glucose in the supernatant was measured as described above.

**Immunoblotting analysis**

Western blotting was carried out as described previously with slight modifications. Details are shown in Appendix S2.

**Co-culture of splenocytes and hepatocytes**

Isolated splenocytes were seeded on a polycarbonate membrane on the bottom of insert cups for 24-well plastic plates (Thermo Fisher Scientific, Roskilde, Denmark) at a density of 3 x 10⁶ cells/cup, and were cultured for 24 h in DMEM supplemented with 100 μmol/L 2-mercaptoethanol with/without LPS (1.0 μg/mL). Freshly isolated hepatocytes were then co-cultured in 24-well plates with the above-described splenocytes for an additional 2 h in DMEM supplemented with/without LPS (1.0 μg/mL), without direct contact between splenocytes and hepatocytes.

**Total RNA extraction, complementary deoxyribonucleic acid synthesis and real-time reverse transcription polymerase chain reaction**

Total RNA was extracted from hepatocytes using Sepazol (Nacalai Tesque, Kyoto, Japan), and complementary deoxyribonucleic acid synthesis was synthesized using the PrimeScript RT reagent Kit (Takara Bio) was used for real-time polymerase chain reaction. The rat sequences for forward and reverse primers to detect PEPCK, G6Pase and β-actin as an inner control are shown in Table S1. Polymerase chain reaction was carried out with the Step One Plus Real-Time PCR System (Applied Biosystems, Foster City, CA, USA) under the following conditions: initial denaturation at 95°C for 10 s, followed by 40 cycles at 95°C for 5 s and 60°C for 34 s.

**Effects of LPS on blood NOx concentrations**

Blood NOx concentrations were measured, followed by an intraperitoneal injection of saline (control) or LPS (0.1–0.5 mg/kg). Blood was collected after 2, 4, 8 and 24 h, and the levels of NOx produced were measured as described above. In NOS inhibitor pretreatment experiments, rats were injected intravenously with L-NMMA (10 mg/kg) 220 min after the injection of LPS (0.5 mg/kg).

**Statistical analysis**

Comparisons between two groups were carried out using an unpaired Student’s t-test. A one-way ANOVA followed by Dunnett’s post-hoc test was carried out for more than two groups. A value of P < 0.05 was considered significant.

**RESULTS**

**Time-course of effects of LPS on blood glucose levels**

As the first step to elucidate the effects of LPS on glucose homeostasis, we examined the time-course of changes in blood glucose levels using rats. The intraperitoneal injection of LPS (0.1–0.5 mg/kg) or saline (control) did not significantly alter blood glucose levels (Figure 1).

**Effects of LPS on the pyruvate tolerance test**

We next evaluated the effects of LPS on hepatic gluconeogenesis using the pyruvate tolerance test. Four hours after the injection of LPS or saline intraperitoneally, elevations in blood glucose levels after the intraperitoneal administration of pyruvate were significantly suppressed in the LPS groups (Figure 2). This suppressive effect of LPS (0.5 mg/kg) on gluconeogenesis was significant 30 min (control, 6.3 ± 0.49 mmol/L; LPS, 4.6 ± 0.51 mmol/L, P < 0.05 vs control), 60 min (control, 6.1 ± 0.49; LPS, 3.6 ± 0.41 mmol/L, P < 0.01 vs control) and 120 min (control, 5.5 ± 0.15 mmol/L; LPS, 3.7 ± 0.42 mmol/L, P < 0.01 vs control).

**Figure 1**

No significant differences in blood glucose levels after an injection of lipopolysaccharide (LPS) or saline. Blood glucose levels of randomly fed Wistar rats were measured, followed by an injection of saline (control) or LPS (0.1–0.5 mg/kg bodyweight) intraperitoneally. Blood glucose levels were measured 2, 4, 6 and 8 h after the injection. Rats were provided with food ad libitum, even after the LPS injection. White circles, control; white triangles, LPS (0.1 mg/kg bodyweight); black circles, LPS (0.5 mg/kg bodyweight). Values are means ± standard error (n = 5).
$P < 0.01$ vs control) after the pyruvate injection. This suppressive effect of LPS (0.1 mg/kg) was significant 30 min (LPS, 4.2 ± 0.48 mmol/L, $P < 0.01$ vs control) and 120 min (LPS, 4.3 ± 0.47 mmol/L, $P < 0.05$ vs control) after the pyruvate injection. Blood glucose levels after the intraperitoneal injection of vehicle (saline), but without pyruvate injection were not significantly affected. These results show that LPS decreases gluconeogenesis from pyruvate.

Effects of LPS on gluconeogenesis in isolated hepatocytes
In order to investigate the direct effects of LPS on hepatocytes, we examined gluconeogenesis using isolated hepatocytes. The effects of LPS on glucose synthesis from 10 mmol/L lactate plus 1 mmol/L pyruvate in a 2-h incubation of hepatocytes with LPS are shown in Figure 3. LPS did not exert suppressive effects on hepatic gluconeogenesis in freshly isolated hepatocytes at concentrations ranging between 0.02 and 20.0 µg/mL. Glucose production by hepatocytes was significantly reduced by approximately 85% and 40% in the absence of gluconeogenesis precursors in incubation medium and in the presence of 5 mmol/L metformin, a representative inhibitor of gluconeogenesis, respectively.

Effects of LPS on gluconeogenesis in perfused livers
The effects of LPS on gluconeogenesis from lactate were investigated in perfused rat livers. Glucose output into the perfusate was stabilized under 2 µmol/g/h. After the perfusate had been changed from basic buffer to Krebs–Ringer buffer containing 2 mmol/L lactate from 15 to 105 min, glucose output rapidly increased and reached a steady state at approximately 10 µmol/g/h. After the washout of lactate, glucose output decreased to 2 µmol/g/h. An intraperitoneal injection of LPS (0.5 mg/kg) 4 h before liver perfusion did not significantly change the glucose output from that in the control (Figure 4a). In Figure 4b, LPS (0.1–0.5 mg/kg/30 min) was infused continuously from 55 to 85 min with perfusion buffer, and had no significant effect on glucose output. Metformin (5 mmol/L) was infused continuously from 55 to 85 min with perfusion buffer, and significantly suppressed glucose output. These results suggest that the effects of LPS on glucose output in perfused rat livers were not significantly different from those in the controls.

Time-course of the production of cytokines induced by LPS in splenocytes
As the suppressive effects of LPS on hepatic gluconeogenesis might be due to its indirect effects on the liver, we examined the production of cytokines in immunocytes with the addition of LPS. The main cytokines, NO, TNF-α, IFN-γ, IL-1β, IL-6 and IL-10, induced by LPS in splenocytes were measured. The exposure of splenocytes to LPS significantly increased the production of NO, TNF-α, IFN-γ, IL-1β and IL-10 in a concentration-dependent manner (Figure 5).

Figure 5 shows the time-course of increases in NOx, TNF-α, IFN-γ, IL-1β and IL-10 production after the application of LPS at concentrations of 0.01–1.0 µg/mL. IL-6 was undetectable in the absence and presence of LPS (0.01–1.0 µg/mL) at all sampling times (data not shown).
Effects of cytokines on gluconeogenesis in hepatocytes

The effects of cytokines (NO, TNF-α, IFN-γ, IL-1β, IL-6 and IL-10), which are known to be induced by LPS, on gluconeogenesis in isolated hepatocytes were examined. After a 120-min incubation with cytokines, glucose concentrations were measured in order to investigate the effects of cytokines on gluconeogenesis in hepatocytes (Figure 6). The inclusion of SNP in incubation medium resulted in the inhibition of glucose synthesis without insulin. This inhibition was significant at concentrations >50 μmol/L SNP (Figure 6a). TNF-α, IFN-γ, IL-1β, IL-6 and IL-10 had no significant effects on gluconeogenesis in hepatocytes (Figure 6b–f). Glucose production was significantly reduced in the absence
of gluconeogenesis precursors in incubation medium or in the presence of 5 mmol/L metformin. These results show that LPS-induced NO is a key regulator of hepatic gluconeogenesis.

Effects of NO on adenosine monophosphate-activated protein kinase-α phosphorylation in hepatocytes

In order to investigate the underlying mechanisms by which NO suppresses gluconeogenesis, we examined the effects of NO and metformin on adenosine monophosphate-activated protein kinase-α (AMPKα) using isolated hepatocytes. The therapeutic effects of metformin are mediated by the activation of AMPK, leading to a decrease in hepatic gluconeogenesis. After a 2-h exposure, 50 μmol/L SNP and 5 mmol/L metformin stimulated the phosphorylation of Thr172 of AMPKα, suggesting the activation of AMPK (Figure 7).

Effects of LPS on co-cultured splenocytes and hepatocytes

The interaction between splenocytes and hepatocytes in response to LPS was investigated in vitro. After splenocytes and hepatocytes were co-cultured for 2 h in culture medium with/without LPS (1.0 μg/mL), the mRNA expression levels of PEPCK and G6Pase, master regulators of glucose production, were examined in hepatocytes. G6Pase mRNA expression levels were significantly lower in LPS-treated hepatocytes than in LPS-untreated hepatocytes, whereas those of PEPCK were lower in LPS-treated hepatocytes than in LPS-untreated hepatocytes (Figure 8).
Time-course of blood NOx concentrations after the LPS injection

In order to investigate the in vivo effects of LPS on NO production, we examined the time-course of blood NOx concentrations after the LPS injection using rats. Blood NOx concentrations were significantly higher in the LPS group (0.5 mg/kg) than in the control group from 4 h to 24 h (Figure 9). NOx concentrations were slightly higher in the LPS group (0.1 mg/kg) than in the control group. NOx concentrations at 4 h were significantly lower in the presence of L-NMMA (10 mg/kg), which was intravenously injected 220 min after LPS (0.5 mg/kg i.p.), than in its absence. These results show that LPS significantly increased the in vivo production of NO, which could suppress hepatic gluconeogenesis.

Inhibitory effects of L-NMMA on LPS action in the pyruvate tolerance test

In the pyruvate tolerance test, the pretreatment of L-NMMA (10 mg/kg i.v.) significantly reduced the inhibitory effects of LPS (0.5 mg/kg i.p.) on gluconeogenesis 60 min after the pyruvate injection (pretreatment, 5.5 ± 0.23 mmol/L; no pretreatment, 3.6 ± 0.41 mmol/L, P < 0.05 vs pretreatment).

DISCUSSION

The results of the present study showed that LPS, a substance that produces endotoxic shock, decreased gluconeogenesis from pyruvate in vivo. We also showed that the LPS injection increased the plasma levels of TNF-α and IFN-γ, which are inflammatory cytokines. A previous study reported that TNF-α prevented enhancements in glucagon-stimulated gluconeogenesis from lactate in isolated hepatocytes. In that study, hepatocytes were pre-incubated for 4 h in culture medium containing fetal calf serum and insulin. Hepatocytes were then cultured for 44 h in the absence of fetal calf serum, and stimulated with glucagon for an additional 4 h in medium M199 supplemented with TNF-α and [14C]lactate. The rate of gluconeogenesis was then assessed radiochemically. However, another study reported that TNF-α stimulated gluconeogenesis by increasing plasma...
glucagon levels in vivo. In that study, rats were injected intraperitoneally with TNF-α, followed by collagenase digestion of the liver to isolate hepatocytes. Isolated hepatocytes were incubated for 2 h in culture medium containing alanine as a glucogenic amino acid, and the rate of hepatic gluconeogenesis was measured. The present results showed that TNF-α failed to affect gluconeogenesis in freshly isolated hepatocytes. In the present study, isolated hepatocytes were incubated for 2 h in Kreb–Ringer buffer with gluconeogenic precursors (lactate and pyruvate) in the presence of TNF-α. The reasons for the discrepancy between the present results and the findings of Christ et al. and Blumberg et al. remain unknown, but might be attributed to differences in experimental protocols and conditions.

Regarding IFN-γ, Wang et al. reported that a deficiency in IFN-γ reduced hepatic G6Pase activity and enhanced glucose tolerance in mice. In that study, the bodyweight of and food intake by IFN-γ knockout mice were lower, whereas their physical activity was greater than the control. Nevertheless, the present results showed that the addition of IFN-γ to culture medium did not affect gluconeogenesis in isolated rat hepatocytes. The reason for the discrepancy in our results and the findings of Wang et al. remains unclear, but again might be ascribed to differences in the methods and species used.

Chronic inflammatory signaling is generally regarded as a key cause of insulin resistance. We herein showed that the acute exposure of hepatocytes to TNF-α and IFN-γ did not have any effects on gluconeogenesis. Therefore, the acute and chronic effects of inflammatory cytokines appear to differ.

iNOS, known as an LPS-stimulated protein, has beneficial roles in host immunity. iNOS mutant mice have an increased susceptibility to infection, showing the importance of iNOS in host defenses against infectious disease. Excessive amounts of NO produced by iNOS might react with the superoxide anion to generate the peroxynitrite radical, which has detrimental effects on cells. Regarding glucose metabolism, iNOS was shown to reduce glucose tolerance and increase insulin resistance. Conversely, in septic shock, the genetic deletion of iNOS in mice maintained hepatic gluconeogenesis, leading to the prevention of hypoglycemia. Therefore, the impact of iNOS on glucose metabolism remains controversial.

The disruption of endothelial NOS causes insulin resistance, whereas its activation ameliorates insulin resistance in...
diabetic mice. Therefore, endothelial NOS is currently attracting attention for its central role in the regulation of glucose metabolism. The production of an appropriate amount of NO might be important in the regulation of glucose metabolism and prevention of infection in patients with diabetes.

Endotoxic hypoglycemia has been reported in patients with liver cirrhosis and kidney failure, implying that a dysfunction in gluconeogenic organs is associated with septic hypoglycemia.

In the present study, after the LPS injection, gluconeogenesis in vivo decreased in the pyruvate tolerance test. Blood NOx concentrations also significantly increased 4 h after the LPS injection. Regarding our study in vitro, SNP suppressed gluconeogenesis in isolated hepatocytes, whereas LPS stimulated the production of NOx in isolated splenocytes. In co-cultures of splenocytes and hepatocytes, LPS suppressed G6Pase mRNA expression in hepatocytes. Collectively, these results suggest that NO produced by splenocytes is involved in hypoglycemia with endotoxic shock. LaNoue et al. reported that the intraperitoneal administration of LPS to rats reduced G6Pase activity, which is consistent with the present study.

Regarding the LPS dosage applied in vivo experiments, Raetzsch et al. reported that the intraperitoneal application of LPS at dosages of 0.05–0.5 mg/kg reduced blood glucose levels in a dose-dependent manner, with the maximal response being observed at 0.5 mg/kg. In that study, the intraperitoneal application of LPS (5 mg/kg) was lethal. Consistent with their study, we used LPS dosages of 0.1–0.5 mg/kg to examine the effects of LPS on glycemia and the pyruvate tolerance test.

The spleen is a lymphatic organ that is located in the left hypochondriac region. Blood in this organ is rich in macrophages. The splenic vein directly connects to the portal vein. LPS is known to induce splenomegaly, which is associated with a larger number of immune cells than that in a healthy spleen. Thus, in endotoxin shock, NO produced by splenocytes is more likely to play an important role in the inhibition of hepatic gluconeogenesis.

The limitations of the present study need to be considered. In this study, we did not carry out experiments using a co-culture of hepatocytes and Kupffer cells, which have been shown to release NO by various stimuli including LPS. NO produced by Kupffer cells in response to LPS might inhibit hepatic gluconeogenesis. Further research is warranted. Another limitation is that we carried out liver perfusion using the flow-through method, not the re-circulation method. In our perfusion method, some of the NO produced by Kupffer cells in response to LPS might have been washed out by fresh perfusion solution, thereby decreasing its effects on hepatocytes. Goto et al. carried out liver perfusion using the re-circulation method in 10-day-old rats, and reported that the intraperitoneal pretreatment of LPS reduced hepatic gluconeogenesis. The reasons for the discrepancy between the present results and the findings of Goto et al. remain unknown, but might be attributed to differences in the experimental protocols used. In the present study, the suppression of glucose output was not observed in the liver perfusion experiment with LPS. Therefore, further studies are required.

Decreased NO levels in blood have been reported with diabetes in humans and rats. The regulation of NOS-mediated NO signaling might open a new avenue for the treatments for diabetes.

In conclusion, high blood concentrations of NO in endotoxemia might cause hypoglycemia through the inhibition of hepatic gluconeogenesis in an insulin-independent pathway. The regulation of iNOS-mediated NO signaling could lead to new treatments for hypoglycemia in endotoxemia.

**DISCLOSURE**

The authors declare no conflict of interest.

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SUPPORTING INFORMATION
Additional Supporting Information may be found in the online version of this article:

Appendix S1 | Gluconeogenesis in perfused rat livers.
Appendix S2 | Immunoblotting analysis.
Table S1 | Primer sequences used in quantitative reverse transcription polymerase chain reaction