Evaluation of antioxidant potential and total phenolic content of exotic fruits grown in Colombia

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INTRODUCTION
The generation of reactive oxygen species (ROS) by the body’s metabolism has a beneficial role in killing infectious agents and regulating cell signaling pathways, as well as genes related to the inflammatory and immune response (Khurana et al., 2013). Nevertheless, the imbalance produced by this intracellular accumulation of ROS induces oxidative stress, which is a cellular state in which DNA, carbohydrates, proteins, and lipids are oxidized, resulting in impaired cellular function, mutations, and cell death. These cellular alterations have been related to the initiation and progression of diseases such as diabetes, cancer, cardiovascular disease, and Alzheimer’s disease, among others that are classified as noncommunicable diseases (NCDs) (Lee et al., 2015; Liguori et al., 2018). In this sense, substances capable of inhibiting the excessive production of ROS could become an alternative to reduce the incidence of NCDs.

Results obtained in epidemiological studies showed a correlation between the high consumption of fruits and the prevention of NCDs, especially cardiovascular diseases and cancer (Aune et al., 2017; Boeing et al., 2012; Wang et al., 2014). Based on these studies, the World Health Organization has recommended the consumption of five servings of fruits and vegetables daily as per the Strategy for the Prevention and Control of NCDs (Aune et al., 2017). Consistent with this recommendation, the relationship between fruit consumption and health promotion has been evidenced by the increase in the number of bibliographic references. These studies concluded that the positive effect of fruits on health is associated with their content of phytochemicals, mainly phenolic compounds (Côté et al., 2010; Hidalgo and Almajano, 2017; Kalaycıoğlu and Erim, 2017; Skrovankova et al., 2015). These compounds have a recognized antioxidant effect...
and, therefore, have been associated with the prevention of NCDs (Y Aboul-Enein et al., 2013; Zhang et al., 2015).

The demand for fruits in national and international markets has continuously grown, perhaps by the publications that show that low consumption of fruits is among the main factors associated with worldwide mortality. On the contrary, fruit intake is related to a decreased risk of NCDs (Kho et al., 2011; Slavin and Lloyd, 2012). Colombia is one of the countries that is considered as megadiverse, since it has a great variety of exotic fruits (Contreras-Calderón et al., 2011). Fruits such as yellow pitaya (Hylocereus megalanthus), banana passion (Passiflora cumbalensis), purple passion fruit (Passiflora edulis), golden berry (Physalis peruviana), tamarillo (Solanum betaceum), lulo (Solanum quitoense), soursop (Annona muricata), kalipatti sapota (Manilkara zapota), sapote (Pouteria sapota), and sour guava (Psidium friedrichsthalianum) are widely employed in folk medicine as a poultice and tisane. Colombia is one of the countries that is considered as megadiverse, since it has a great variety of exotic fruits (Contreras-Calderón et al., 2011). Fruits such as yellow pitaya (Hylocereus megalanthus), banana passion (Passiflora cumbalensis), purple passion fruit (Passiflora edulis), golden berry (Physalis peruviana), tamarillo (Solanum betaceum), lulo (Solanum quitoense), soursop (Annona muricata), kalipatti sapota (Manilkara zapota), sapote (Pouteria sapota), and sour guava (Psidium friedrichsthalianum) are commonly consumed in Colombia and are used extensively to treat various diseases in different regions of the world. For example, soursop and purple passion fruit are used to treat cancer, hypertension, inflammation, among other diseases (Daddiouaissa and Amid, 2018; Nanda et al., 2013; Taïwe and Kuete, 2017). Sapote and tamarillo are useful remedies for patients’ gastrointestinal problems (Angulo et al., 2012; Ngomle et al., 2020; Stanley et al., 2009). Golden berry is used to treat asthma, rheumatism, cancer, and leukemia (Hassanien, 2011; Wu et al., 2004a). Lulo is used to treat nerve problems and skin diseases (Bussmann et al., 2018). Kalipatti sapota is used in the treatment of pulmonary diseases and also inhibits the growth of breast and colon cancers (Bano and Ahmed, 2017). Yellow pitaya has a mild laxative effect (Fratoni et al., 2019; Sudha et al., 2017). Leaves and flowers of P. cumbalensis are widely employed in folk medicine as a poultice for gastrointestinal problems (Jerves-Andrade et al., 2014). However, no reports of the species of P. friedrichsthalianum were found in the literature.

Regarding studies of antioxidant activity, in the literature, there are reports of the antioxidant potential of some fruits included in this study that are evaluated individually. For example, the content of total phenolic compounds and the uptake of radicals 2,2'-diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonic acid) diaminonitrosourea (ABTS) have already been reported for all fruits included in this work (Bravo et al., 2016; dos Reis et al., 2018; Flores et al., 2013; Gordillo et al., 2012; Prabhu et al., 2018; Torres Grisales et al., 2017; Vasco et al., 2008; Woo et al., 2013), while for yellow pitaya and sour guava, there are no activity reports for the ferric-reducing antioxidant power (FRAP) or the oxygen radical absorbance capacity-fluorescein (ORAC-FL) (Bravo et al., 2015, 2016; Espin et al., 2016; González et al., 2012; Gordillo et al., 2012; Penaloza et al., 2017; Shafi et al., 2017; Vasco et al., 2008; Woo et al., 2013; Yahia and Barrera, 2009; Yahia et al., 2011; Zapata et al., 2014). Moreover, the scavenging of nitric oxide radical and the inhibition of the generation of intracellular ROS have been reported only for golden berry, kalipatti sapota, soursop, tamarillo, and purple passion fruit (Gomathy et al., 2013; González et al., 2016; Kou et al., 2009; Maruki-Uchida et al., 2013; Orqueda et al., 2020; Rop et al., 2012; Tan et al., 2018; Wu et al., 2004b; Zamudio-Cuevas et al., 2014). Despite this background, it is complicated to determine which would be the most promising fruits as antioxidants, since the fundamental differences are observed in the extraction methods regarding the used methodology and in the expression of the results. Few reports have studied the antioxidant properties of these fruits under the same experimental conditions (Bravo et al., 2016; Contreras-Calderón et al., 2011). Taking into account the high demand for these fruits, their ethnomedicinal usage, and antioxidant history, in this work, we evaluated the antioxidant potential of ten exotic fruits grown in Colombia to identify the promising fruits for the treatment or prevention of NCDs by quantifying the content of total phenolic compounds, as well as their free radical scavenging capacity (DPPH, ABTS, ROO, NO). Additionally, the reduction capacity of the extracts was measured with the FRAP assay and the inhibition of the generation of ROS in lipopolysaccharide (LPS)-stimulated RAW 264.7 mouse macrophages was measured using the DCFH-DA method.

**MATERIALS AND METHODS**

**Reagents**

Gallic acid, Folin–Ciocalteu, sodium nitroprusside dihydrate, sulfanilamide, horseradish peroxidase, guaiacol, catechin, fluorescein, quercetin, 6-hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid (Trolox), ABTS, DPPH, 2,2'-azobis(2-methylpropionamide) dihydrochloride (AAPH), 2,4,6-Tris(2-pyridyl)-s-triazine (TPTZ), 5,5-dimethyl-1-pyrrole N-oxide, N-(1-naphthyl)ethylenediamine dihydrochloride, 2,7'-dichlorofluorescein diacetate (DCFH-DA), pyrogallol red, ferric chloride, ferrous sulfate heptahydrate, magnesium sulfate heptahydrate, potassium persulfate, and lipopolysaccharide (LPS) were obtained from Sigma-Aldrich (St. Louis, MO). Phosphoric acid, potassium chloride, and sodium chloride were obtained from JT Baker (Phillipsburg, NJ). Methanol, ethanol, dimethyl sulfoxide (DMSO), Dulbecco’s modified eagle medium (DMEM), and fetal bovine serum (FBS) were obtained from Thermo Fisher Scientific (Pittsburgh, PA). Caffeic acid, aluminum chloride, ascorbic acid, sodium hydroxide, sodium nitrite, D-glucose, calcium chloride, potassium phosphate monobasic, potassium phosphate dibasic, sodium phosphate dibasic, sodium phosphatemonobasic, hydrochloric acid, acetic acid, hydrogen peroxide, and 3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) were obtained from Merck Millipore (Burlington, MA).

**Fruit samples and determination of physicochemical parameters**

Fruits included in this study were obtained from Fusagasugá (Cundinamarca) and Turbana (Bolivar), which involved yellow pitaya (H. megalanthus), banana passion (P. cumbalensis), purple passion fruit (P. edulis), golden berry (P. peruviana), tamarillo (S. betaceum), lulo (S. quitoense), soursop (A. muricata), kalipatti sapota (M. zapota), sapote (P. sapota), and sour guava (P. friedrichsthalianum). Representative samples of each fruit were used to determine the soluble solids (°Brix), titratable acidity, and pH values, which were conducted following the guidelines established by the Association of Official Analytical Chemists (Helrich, 1990). The concentration of the solid soluble content (°Brix) was measured using a digital refractometer (Sper Scientific 300034) at 25°C and the pH values were determined by a pH meter (Starter 3100, OHAUS) at 25°C.
Preparation of extracts and determination of total phenolics content

Fruits were washed under tap water, weighed, homogenized, frozen at −80°C, and lyophilized. The dried material was extracted with ethanol (96%) at room temperature in stoppered containers for a defined period of time with frequent agitation until exhaustion and the obtained extract was concentrated in a rotary evaporator at controlled temperatures (38°C – 40°C) and reduced pressure. The Folin–Ciocalteu method was used to quantify the total phenolic compounds (Rivera et al., 2018). Phenolic compounds were calculated as mg gallic acid equivalents (GAE)/100 g of fresh weight.

Antioxidant activity assays

DPPH and ABTS free radical assays

Free radical DPPH and ABTS scavenging was determined using the standard methods with some modifications (Castro et al., 2018). For the uptake of the DPPH radical, 75 µl of different concentrations of the fruit extracts were mixed with 150 µl of a methanolic solution of DPPH (70 µg/ml). The mixture was allowed to incubate for 30 minutes and the absorbance at 517 nm was measured. While for the ABTS radical, 20 µl of the different concentrations of fruit extracts were mixed with 180 µl of the ABTS solution. After 30 minutes of incubation, the absorbance was measured at 734 nm in a Multiskan Go microplate reader (Thermo Scientific, Waltham, MA). A Trolox standard curve was used to determine the equivalent antioxidant capacity and the results were expressed as µmol Trolox equivalents/g of fresh weight.

Ferric-reducing antioxidant power assay

In this work, we used the standard method with some modifications (Castro et al., 2014). The FRAP reagent was prepared by mixing 10 volumes of 300 mM acetate buffer (pH 3.6) with 1 volume of 10 mM TPTZ in HCl (40 mM) and with 1 volume of 20 mM FeCl3·6H2O. Different concentrations of fruit extracts (10 µl) were mixed with the FRAP reagent (290 µl), incubated independently at 37°C, and the absorbance was determined at 593 nm after 60 minutes of the reaction using an EnSpire Multimode microplate reader (PerkinElmer, Waltham, MA). The FRAP value was determined by plotting a standard curve made by the addition of ferrous sulfate to the FRAP reagent. Results were expressed as µmol of Fe²⁺/100 g of fresh weight.

Oxygen radical absorbance capacity-fluorescein

The ability of fruit extracts to trap the peroxyl radical was determined according to the method proposed by Ou et al. (2001) and validated by Bisby et al. (2008). The peroxyl radicals generated by the thermal decomposition of AAPH reacted with fluorescein to produce a nonfluorescent product over time. Briefly, 20 µl of different concentrations of fruit extracts were preincubated for 5 minutes with 120 µl of fluorescein (100 nM) in a 96-well flat-bottom black plate. After which, 60 µl of AAPH (600 mM) was added to each well and the fluorescence was recorded using a Fluoroskan Ascent equipment (Thermo Scientific, Waltham, MA) every 56 seconds for 150 minutes at excitation and emission wavelengths of 485 and 538 nm, respectively. The results were expressed as µmol Trolox equivalents/g of fresh weight.

Hydrogen peroxide scavenging activity

The method described by Doerge et al. (1997), with some modifications, was used to determine the hydrogen peroxide scavenging activity of fruit extracts. Briefly, 900 µl of HBSS buffer, 100 µl of H2O2 (10 mM), and 10 µl of fruit extracts at different concentrations were incubated for 30 minutes. After incubation, 50 µl of guaiacol and 10 µl of horseradish peroxidase (0.6 µg/ml) were added and incubated again for 20 minutes. The scavenging activity was determined in a Multiskan EX microplate reader (Thermo Scientific, Waltham, MA) at 450 nm. Results were expressed as µmol Trolox equivalents/g of fresh weight.

Scavenging of the nitric oxide radical

Nitric oxide is generated by solubilizing sodium nitroprusside in an aqueous solution, where it is transformed into nitrite by the oxygen present in the medium. The produced nitrates are easily quantified by the spectrophotometric assay based on the Griess reaction (Rao, 1997). We incubated 990 µl of sodium nitroprusside solution (10 mM) in PBS with 10 µl of different concentrations of fruit extracts for 4 hours at room temperature. After incubation, equal volumes of Griess reagent and the samples were mixed. The nitric oxide concentration was determined by spectrophotometry at 550 nm in a Multiskan Go microplate reader (Thermo Scientific, Waltham, MA) by comparing to a standard curve of NaNO2 (1–200 µM). Results were expressed as µmol of results were expressed as µmol of caffeic acid equivalents/g of fresh weight.

Cellular ROS determination by DCFH-DA

Cell culture

Murine macrophage RAW cell lines (ATCC® TIB-71T™, Rockville, MD) were maintained in the DMEM supplemented with 10% heat-inactivated FBS in a humidified 5% CO2 atmosphere at 37°C.

Assessment of cell viability

The toxicity of fruit extracts on RAW 264.7 macrophages was evaluated using the 3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) colorimetric method (Scudiero et al., 1988). RAW 264.7 macrophages were seeded in sterile 96-well plates (2×10⁵ cells/well) and incubated at 37°C for 48 hours, then the medium was removed, and cells were washed with phosphate buffer saline (PBS) and treated for 30 minutes with different concentrations of fruit extracts, after which, they were activated with LPS (1 µg/ml) and incubated again for 20 minutes. The toxicity was determined in a Multiskan EX microplate reader (Thermo Scientific, Waltham, MA). In each trial, a group of cells not exposed to the extracts was included as the negative control group as 100% viability. The viability percentages were calculated, considering the negative control group as 100% viability.

DCFH assay

The formation of intracellular ROS was determined using the oxidation-sensitive dye DCFH-DA according to the protocol
described by Park et al. (2017), with some modifications. For that, macrophages RAW 264.7 (10×10⁴ cells/well) were incubated in 96-well sterile black plates. After 24 hours of incubation, the cells were washed twice with sterile PBS to remove nonadherent cells. The remaining cells were incubated for 30 minutes with different concentrations of fruit extracts and activated with LPS (1 μg/ml). After 24 hours of incubation, the cells were washed twice with sterile PBS to remove nonadherent cells. The remaining cells were incubated for 30 minutes with different concentrations of fruit extracts and activated with LPS (1 μg/ml). After 24 hours of incubation, the cells were washed three times with sterile PBS, stained with DCFH-DA (20 μM) in PBS, and incubated for 30 minutes at 37°C in the dark. Fluorescence intensity was measured in a 96-well Fluoroskan Ascent plate reader (Thermo Scientific, Waltham, MA) at an excitation wavelength of 485 nm and an emission wavelength of 538 nm. Quercetin was used as the positive control.

Statistical analysis

Antioxidant activity results of three independent assays were reported as mean ± SEM. A one-way analysis of variance was used to analyze the data, followed by Tukey’s and Dunnett’s comparison tests. P-values less than 0.05 were considered significant.

RESULTS AND DISCUSSION

Fruits, like vegetables, provide nutrients, such as vitamins and minerals, as well as nonnutritive secondary metabolites with biological activity that can contribute to improving people’s health and reducing the risk of chronic NCD. Among these metabolites, phenolic compounds stand out, which have several pharmacological properties and are recognized for their potent antioxidant activity (Liu, 2013). In this work, we evaluated the antioxidant potential as well as the content of total phenolic compounds of ten extracts of fruits grown in Colombia using seven different methods. The information of fruits selected for this study and their state of maturation is presented in Tables 1 and 2.

Quantification of phenolic compounds

Results of the quantification of phenolic compounds of fruits under study are presented in Table 3. Considering the broad range of variations in the total phenolic compounds, the fruits were classified into three groups: with a high content (>80 mg GAE/100 g FW), a moderate content (80> mg GAE/100 g FW >40), and a low content (<40 mg GAE/100 g FW). The results show that only banana passion and sour guava presented high contents of phenolic compounds. On the contrary, yellow pitaya, golden berry, purple passion fruit, sapote, kalipatti sapota, tamarillo, and soursop showed moderate content. Lulo had the lowest content of these compounds.

DPPH, ABTS, and FRAP assay

Table 3 presents the results of the scavenger capacity of DPPH and ABTS free radicals as well as the FRAP of the ten fruit extracts under study. It can be observed that the extracts of banana passion and sour guava presented the best scavenging activity of radicals DPPH and ABTS and the best activity of ferric ion reduction. On the contrary, the extracts of yellow pitaya, golden berry, purple passion fruit, sapote, kalipatti Sapota, tamarillo, soursop, and lulo had a moderate activity. These results are related to the content of phenolic compounds, showing high correlation coefficients in all cases (DPPH: \( r = 0.978 \); ABTS: \( r = 0.979 \); and FRAP: \( r = 0.981 \)). These results are similar to those reported by other studies, where

| Fruits              | Part used       | Extract yields (%) | Voucher number | Collection site                  |
|---------------------|-----------------|--------------------|----------------|----------------------------------|
| Yellow pitaya       | Pulp and seed   | 24.54              | 199083         | Fusagasugá (Cundinamarca)        |
| Hylocereus megalanthus |                |                    |                | (4°20′14″ N, 74°21′52″ O)        |
| Banana passion      | Pulp            | 9.78               | 199085         | Altitude: 1,800 masl             |
| P. cumbalensis      | Pulp            | 13.04              | 199081         | Precipitation: 950 mm of rain per year |
| Purple passion fruit| Pulp            | 18.08              | 199086         | Climate: 8°C–28°C                |
| P. edulis           | Whole fruit      | 10.10              | 199080         |                                  |
| Golden berry        | Pulp            | 32.27              | 201955         | Turbana (Bolivar)                |
| P. peruviana        | Pulp            | 26.26              | 201955         | (10°16′35″ N, 75°26′19″ O)        |
| Tamarillo           | Pulp            | 12.01              | 201952         | Altitude: 138 masl               |
| S. quitoense        | Pulp and seed   | 5.46               | 199084         | Precipitation: 778 mm of rain per year |
| Soursop             | Pulp            | 13.12              | 201951         | Climate: 23°C–32°C               |
| A. muricata         | Pulp            | 32.27              | 201953         |                                  |
| Kalipatti sapota    | Pulp            | 26.26              | 201952         |                                  |
| M. zapota           | Pulp            | 12.01              |                |                                  |
| Sapote              | Pulp            | 13.12              |                |                                  |
| P. sapota           | Pulp            | 32.27              |                |                                  |
| Sour guava          | Pulp            | 26.26              |                |                                  |
| P. friedrichstallianum | Pulp          | 12.01              |                |                                  |

The part of the fruit used to carry out the study and prepare the extracts was selected based on the way people eat the fruit. Yields were calculated as: [(grams of lyophilized pulp/grams of fresh weight) × 100].
the free radical scavenging capacity of phenolic compounds is evident; being able to react with highly ROS, donating a hydrogen atom (HA), or transferring a single electron (SE) is all due to the low redox potential that it possesses (Kurek-Górecka et al., 2014). In fact, according to reaction mechanisms, antioxidant activity tests can be classified as a transference assay of an HA or an SE. FRAP and ABTS have been included in the category as the HA assays, while the DPPH assay can be classified in both groups since both reactions can occur, and the antioxidant agent determines the final mechanism (Hidalgo and Almajano, 2017). It should be noted that although the extracts of golden berry, purple passion fruit, and sapote showed a similar content of phenolic compounds, sapote extract presented better free radical scavenging capacity. This may happen because the activity of phenolic compounds also depends on the concentration on the number of hydroxyl groups and the position of the aromatic ring (Kurek-Górecka et al., 2014).

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) scavenging assays

The excessive production of ROS causes cell toxicity and target organ damage, causing the initiation of NCDs. The results of ROS scavenging activity by those fruit extracts showed that the extracts of yellow pitaya, sour guava, and banana passion exhibited potent peroxyl radical scavenging activity (Table 4). Extracts of sour guava and banana passion significantly decrease the amount of $H_2O_2$, which is an important precursor of hydroxyl radicals among other free radicals.

The NO radical can act directly as an effector molecule or as a second messenger, intervening in numerous signaling pathways (Zhang et al., 2017). When it is produced in excess, it can cause noxious effects, which are related to the capacity of this mediator to react with oxygen, superoxide, and transition metals, generating NOx, peroxyxinitrite, and heme-complexes, respectively. Peroxyxinitrite is a powerful, extremely cytotoxic oxidizing agent used by cells of the immune system within its defense mechanisms. However, when produced in excess, it can damage a wide range of molecules in cells, including proteins, DNA, and lipids, which leads to the initiation of a recognized pathophysiological mechanism causing the appearance of chronic NCDs (Pacher et al., 2007). Extracts of sour guava and banana passion presented an important nitric oxide radical scavenging effect (Table 4). It is remarkable that, once again, the fruit extracts with the major activity were those with the highest content of phenolic compounds.

### Table 2. Physicochemical parameters of the fruits.

| Fruits         | Brix   | pH     | Total acidity (g/100 ml) |
|----------------|--------|--------|--------------------------|
| Yellow pitaya  | 17.68 ± 0.27 | 4.58 ± 0.01 | 0.11 ± 0.01               |
| Banana passion | 10.62 ± 0.11  | 3.41 ± 0.03 | 1.23 ± 0.04               |
| Purple passion | 15.04 ± 0.46  | 3.13 ± 0.04 | 2.97 ± 0.25               |
| Golden berry   | 14.20 ± 0.06  | 3.52 ± 0.006 | 2.15 ± 0.01               |
| Tamarillo      | 9.61 ± 0.14   | 3.61 ± 0.004 | 2.18 ± 0.03               |
| Lulo           | 9.08 ± 0.16   | 3.13 ± 0.007 | 3.33 ± 0.09               |
| Sour Guava     | 12.57 ± 0.38  | 4.06 ± 0.04 | 0.62 ± 0.04               |
| Kalipatti sapota | 25.37 ± 0.30 | 5.25 ± 0.08 | 1.30 ± 0.18               |
| Sapote         | 22.33 ± 0.37  | 6.75 ± 0.05 | 1.07 ± 0.02               |
| Sour Guava     | 8.43 ± 0.20   | 2.86 ± 0.02 | 3.80 ± 0.09               |

The results represent mean ± SEM (n = 9) from three independent experiments.

### Table 3. Quantification of phenolic compounds and antioxidant potential of the extracts by the DPPH, ABTS, and FRAP methods.

| Fruits         | Phenolic compounds | DPPH          | ABTS          | FRAP          |
|----------------|--------------------|---------------|---------------|---------------|
| Banana passion | 249.5 ± 0.064      | 1959.7 ± 1.288| 4257.3 ± 0.622| 5962.6 ± 3.241|
| Sour guava     | 80.9 ± 0.010       | 374.5 ± 0.083 | 822.3 ± 0.110 | 1607.4 ± 0.220|
| Yellow pitaya  | 59.1 ± 0.013       | 177.1 ± 0.073 | 323.8 ± 0.026 | 811.2 ± 0.201 |
| Golden berry   | 53.3 ± 0.011       | 78.9 ± 0.018  | 228.6 ± 0.034 | 419.7 ± 0.125 |
| Purple passion | 50.3 ± 0.006       | 93.3 ± 0.030  | 209.8 ± 0.027 | 312.4 ± 0.149 |
| Sapote         | 49.0 ± 0.004       | 136.5 ± 0.030 | 310.7 ± 0.015 | 549.2 ± 0.094 |
| Kalipatti sapota | 44.7 ± 0.008     | 73.2 ± 0.034  | 193.9 ± 0.044 | 412.2 ± 0.156 |
| Tamarillo      | 41.9 ± 0.007       | 72.0 ± 0.018  | 197.6 ± 0.023 | 427.2 ± 0.070 |
| Soursop        | 40.4 ± 0.008       | 64.3 ± 0.018  | 183.8 ± 0.023 | 204.1 ± 0.091 |
| Lulo           | 31.8 ± 0.006       | 84.5 ± 0.036  | 144.2 ± 0.008 | 341.2 ± 0.147 |

Phenolic compounds were calculated as mg gallic acid/100 g of fresh weight. Results of DPPH and ABTS were expressed as micromoles of Trolox equivalents (TEs) per 100 g of fresh weight (μmol of TEs/100 g of FW) and FRAP results as micromoles of Fe$^{2+}$/100 g of fresh weight (μmol of Fe$^{2+}$/100 g of FW). Results represent mean ± SEM (n = 12) from three independent experiments.
Production of intracellular ROS

The DPPH, ABTS, FRAP, and ORAC-FL assays are widely used to evaluate the antioxidant capacity of fruits (Hidalgo and Almajano, 2017). However, in recent years, antioxidant activity assays that include cellular models have been widely accepted since it is considered that the results obtained with them could better predict the antioxidant power of fruits.

In order to deepen the understanding of the antioxidant potential of the ten fruit extracts, their capacity to inhibit the generation of intracellular ROS was measured using DCFH-DA. The extracts of golden berry, banana passion, and yellow pitaya were the most active (Fig. 1). The extracts of sour guava, purple passion fruit, sapote, kalipatti sapota, tamarillo, and lulo showed a low inhibition on the formation of intracellular ROS (data not shown). The extract of soursop fruit was not evaluated because it had a toxic effect on the cells. Note that the extract of golden berry was more active than expected, as it was not the extract with the highest content of phenolic and it did not have the best free radical scavenging effect. The opposite happened with the extract of sour guava, which showed the best free radical scavenging activity and the highest content of phenolic compounds, but it did not exhibit a high capacity to inhibit the production of ROS in the cell assay. In such a way that there is no evidence of a correlation between antioxidant activity at the cellular level and the content of phenolic compounds, contrary to what happens with DPPH, ABTS, and FRAP free radical scavenging assays. It could be because the antioxidant properties of polyphenols depend on the physicochemical environment, which is very complex within a biological matrix, while it is more stable in

Table 4. Scavenging effect on reactive oxygen and nitrogen species.

| Fruits                  | ORAC         | H₂O₂         | NO            |
|-------------------------|--------------|--------------|---------------|
| Banana passion          | 2409.82 ± 120.74 | 38.04 ± 1.71 | 70.81 ± 8.58  |
| Sour guava              | 2602.08 ± 51.94 | 36.51 ± 1.90 | 168.81 ± 3.60 |
| Yellow pitaya           | 2999.77 ± 80.56 | 2.38 ± 0.01  | ulq           |
| Golden berry            | 350.13 ± 5.54  | 4.78 ± 0.44  | ulq           |
| Purple passion fruit    | 471.69 ± 4.91  | 13.79 ± 3.80 | ulq           |
| Sapote                  | 143.87 ± 2.65  | 2.63 ± 0.15  | 31.50 ± 4.22  |
| Kalipatti sapota        | 697.69 ± 7.82  | 0.66 ± 0.14  | ulq           |
| Tamarillo               | 169.71 ± 3.15  | 4.94 ± 0.21  | 18.41 ± 3.75  |
| Soursop                 | 311.24 ± 3.69  | 3.84 ± 0.01  | ulq           |
| Lulo                    | 133.19 ± 1.84  | 1.99 ± 0.06  | 37.81 ± 1.44  |

ORAC and H₂O₂ scavenging capacities were expressed as micromoles of Trolox equivalents per gram of fresh weight (μmol of TEs/g of FW). NO scavenging capacity was expressed as micromoles of caffeic acid equivalents per gram of fresh weight (μmol of caffeic acid/g of FW). Results represent mean ± SEM (n = 12) from three independent experiments. Ulq = Under limit of quantification.

Figure 1. Effect of fruit extracts on the production of intracellular ROS in LPS-stimulated RAW 264.7 macrophages (1 μg/ml) for 24 hours. ROS are expressed as fold increase of 2′,7′-dichlorofluorescein fluorescence over LPS-untreated controls. Fruit extracts were evaluated to 100, 1,000, and 2,000 μg/ml. Quercetin (Q) (100 μM) was employed as the positive control. The results represent mean ± SEM. (*p < 0.05; **p < 0.01; ****p < 0.0001 are statistically significant compared to the LPS-treated group).
ABTS, DPPH, and FRAP free radical scavenging assays (Scalbert et al., 2005). Furthermore, it is also possible that the antioxidant activity shown by these fruit extracts is due to the presence of other secondary metabolites.

This study provides new information on the antioxidant properties of yellow pitaya, banana passion, lulo, sapote, and sour guava. The results of the scavenging effect on DPPH, ABTS, NO, and peroxyl radicals, as well as the inhibition of the generation of intracellular ROS in macrophages, showed that the total ethanolic extracts obtained from the edible parts of banana passion (P. cumbalensis), yellow pitaya (H. megalanths), and golden berry (P. peruviana) make them the most promising fruits as antioxidants. These results support the ethnopharmacological use of these three fruits, concerning the treatment of various chronic NCDs, in addition to being an essential foundation to continue promoting their consumption.

CONCLUSION

The fruits like banana passion (P. cumbalensis), yellow pitaya (H. megalanths), and golden berry (P. peruviana) are a significant source of antioxidant compounds. With the results obtained in this work, we can propose these fruits as functional foods that can be used to enhance health. Nevertheless, these fruit extracts should be evaluated in in vivo models to ensure the bioavailability of the active compounds and should be studied more in-depth regarding their phytochemical characterization to identify the compounds responsible for this activity.

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AUTHORS’ CONTRIBUTIONS

RS and LF conceived the study; RS, YO, CD, JC, and LF supervised the study and designed the experiments; NM and JC carried out the experiments; JC and LF wrote the manuscript. All authors read and approved the final manuscript.

CONFLICT OF INTEREST

The authors state that there is no conflict of interest.

REFERENCES

Angulo A, Rosero R, Gonzales M. Estudio etnobotánico de las plantas medicinales utilizadas por los habitantes del corregimiento de Genoy, Municipio de Pasto, Colombia. Rev Univ Salud, 2012; 14(2): 168–85.
Aune D, Giovannucci E, Boffetta P, Fadnes LT, Keum N, Norat T, Greenwood DC, Riboli E, Vatten LJ, Tonstad S. Fruit and vegetable intake and the risk of cardiovascular disease, total cancer and all-cause mortality—a systematic review and dose-response meta-analysis of prospective studies. Int J Epidemiol, 2017; 46(3):1029–56.
Bano M, Ahmed B. Manilkara zapota (L.) P. Royen (Sapodilla): a review. Int J Adv Res Ideas Innov Technol, 2017; 3:1364–71.
Bisby RH, Brooke R, Navaratnam S. Effect of antioxidant oxidation potential in the oxygen radical absorption capacity (ORAC) assay. Food Chem, 2008; 108(3):1002–7.
Boeing H, Bechthold A, Bub A, Ellinger S, Haller D, Kroke A, Leschik-Bonnet E, Müller MJ, Oberritter H, Schulze M. Critical review: vegetables and fruit in the prevention of chronic diseases. Eur J Nutr, 2012; 51(6):637–63.
Bravo K, Alzate F, Osorio E. Fruits of selected wild and cultivated Andean plants as sources of potential compounds with antioxidant and anti-aging activity. Ind Crops Prod, 2016; 85:341–52.
Bravo K, Sepulveda-Ortega S, Lara-Guzman O, Navas-Arboleda AA, and Osorio E. Influence of cultivar and ripening time on bioactive compounds and antioxidant properties in Cape gooseberry (Physalis peruviana L.). J Sci Food Agric, 2015; 95(7):1562–9.
Bussmann RW, Zambrana NYP, Romero C, Hart RE. Astonishing diversity—the medicinal plant markets of Bogotá, Colombia. J Ethnobiol Ethnomed, 2018; 14(1):43.
Castro C, Mura F, Valenzuela G, Figueuera C, Salinas R, Zuñiga MC, Torres JL, Fuguet E, Delciorte P. Identification of phenolic compounds by HPLC-ESI-MS/MS and antioxidant activity from Chilean propolis. Food Res Int, 2014; 64:873–9.
Castro J, Rivera D, Franco LA. Topical anti-inflammatory activity in TPA-induced mouse ear edema model and in vitro antibacterial of Cordia alba flowers. J Pharm Invest, 2019; 49(3): 331–336.
Contreras-Calderón J, Calderón-Jaimes L, Guerra-Hernández E, García-Villanovo B. Antioxidant capacity, phenolic content and vitamin C in pulp, peel and seed from 24 exotic fruits from Colombia. Food Res Int, 2011; 44(7):2047–53.
Côté J, Callet S, Doyon G, Sylvain J-F, Lacroix M. Bioactive compounds in cranberries and their biological properties. Crit Rev Food Sci Nutr, 2010; 50(7):666–79.
Daddiouaissa D, Amid A. Anticancer activity of acetogenins from Annona muricata fruit. JIMM Med J Malays, 2018; 17(3):103–112.
Doerge DR, Divi RL, Churchwell MI. Identification of the colored guaiacol oxidation product produced by peroxidases. Anal Biochem, 1997; 250(1):10–7.
Fos Reis LCR, Faccio EMP, Salvador M, Flóres SH, de Oliveira Rios A. Antioxidant potential and physicochemical characterization of yellow, purple and orange passion fruit. J Food Sci Technol, 2018; 55(7):2679–91.
Espín S, Gonzalez-Manzano S, Tao V, Poveda C, Ayuda-Durán B, Gonzalez-Paramas AM, Santos-Buelga C. Phenolic composition and antioxidant capacity of yellow and purple-red Ecuadorian cultivars of tree tomato (Solanum betaceum Cav.). Food Chem, 2016; 194:1073–80.
Freses G, Dastmalchi K, Wu S-B, Whalen K, Dabo AJ, Reyneertson KA, Forony RF, Jeanine M, Kennelly EJ. Phenolic-rich extract from the Costa Rican guava (Psidium krietshchthalanum) pulp with antioxidant and anti-inflammatory activity. Potential for COPD therapy. Food Chem, 2013; 141(2):889–95.
Fratiotoni MJ, da Silva KRR, Moreira A. NPK application in yellow pitaya seedlings grown on sand and organic compost. Semina, 2019; 40(5 Supl 1):2179–88.
Gomathy K, Baskar R, Kumaresan K. Comparison of antioxidant potential in pulp and peel extracts of Manilkara zapota (L.) P. Royen. Afr J Biotechnol, 2013; 12(31):4936–43.
González E, Fernández A, Sáyago-Ayerdi S, Estrada R, Vallejo L, Manikara zapota. Antioxidant capacity of crude extracts and acetogenin fraction of soursp fruit pulp. Pharm Anal Acta, 2016; 7(484):2.
González E, Vaillant F, Pérez AM, Rojas G. In vitro cell-mediated antioxidant protection of human erythrocytes by some common tropical fruits. J Nutr Food Sci, 2012; 2:3.
Gordillo JC, Ortiz D, Larrahollo JE, Mejia MS, Pachon H. Actividad antioxidante en guanábana (Annona muricata L.): una revisión bibliográfica. Bol Latinoam Caribe Plant Med Aromát, 2012; 11(2):111–26.

Hassannien MFR. Physalis peruviana: a rich source of bioactive phytochemicals for functional foods and pharmaceuticals. Food Rev Int, 2011; 27(3):259–73.

Helrich K. 1990. Official methods of analysis of the AOAC. 15th edition, Methods 932.06, 925.09, 985.29, 923.03. Association of Official Analytical Chemists, Arlington, VA, 1990.

Hidalgo G-I, Almajano M. Red fruits: extraction of antioxidants, phenolic content, and radical scavenging determination: a review. Antioxidants, 2017; 6(1):7.

Jerves-Andrade L, Cuzco N, Tobar V, Ansaloni R, Maes L. Wilches. Medicinal plants used in south Ecuador for gastrointestinal problems: an evaluation of their antibacterial potential. J Med Plants Res, 2014; 8(45):1310–20.

Kalaycıoğlu Z, Erım FB. Total phenolic contents, antioxidant activities, and bioactive ingredients of juices from pomegranate cultivars worldwide. Food Chem, 2017; 221:496–507.

Khoo H-E, Prasad KN, Kong K-W, Jiang Y, Ismail A. Carotenoids and their isomers: color pigments in fruits and vegetables. Molecules, 2011; 16(2):1710–38.

Khurana S, Piche M, Hollingsworth A, Venkataraman K, Tai T. Oxidative stress and cardiovascular health: therapeutic potential of polyphenols. Can J PharmSci, 2013; 91(3):198–212.

Kou M-C, Yen J-H, Hong J-T, Wang C-L, Lin C-W, Wu M-J. Cyphomandra betacea Sendt. phenolics protect LDL from oxidation and PC12 cells from oxidative stress. Food Sci Technol, 2009; 42(2):458–63.

Kurek-Görecka A, Rzepecka-Stojko A, Görecki M, Stojko J, Sosada M, Świerczek-Zięba G. Structure and antioxidant activity of polyphenols derived from propolis. Molecules, 2014; 19(1):78–101.

Lee Y-M, Han S-I, Song BC, Yeum K-J. Bioactives in commonly consumed cereal grains: implications for oxidative stress and inflammation. J Med Food, 2015; 18(11):1179–86.

Ngomle S, Eko R, Kanwat M, Kalita H, Moyon N. Eating from the wild: an insight into the indigenous wild edible plants consumed by the Digaru Mishmi tribe of Arunachal Pradesh. Indian J Tradit Knowl, 2020; 19(2):360–9.

Orqueda ME, Torres S, Zampini IC, Cattaneo F, Díez de C, Sánchez J, Muñoz S, Ochoa C, Fuentes J, Leiva D. Activity of piceatannol from passion fruit (Passiflora edulis) seeds in UVB-irradiated keratinocytes. Biol Pharm Bull, 2013; 36(5):845–9.

Pachor P, Beckman JS, Liaudet L. Nitric oxide scavenging through activation of Nrhl2/ho-1 signaling pathway in RAW 264.7 macrophages. BMC Complement Altern Med, 2017; 17(1):374.

Penaloza J, Canas J, Rojano B, Maldonado M. Antioxidant capacity, polyphenol content and antiproliferative activity of yellow, tropical and andean berries extracts obtained from mixtures of blackberry (Rubus glaucus), passion fruit (Passiflora edulis and Passiflora ligularis), pineapple (Ananas comosus), guava (Psidium guajava L. Radd) and pineapple guava (Acca sellowiana). Curr Top Nutraceutical Res, 2017; 15(3–4):163.

Prabhj JS, Selvam AP, Rajeswari VD. Effective anti-cancer property of Pouteria sapota leaf on breast cancer cell lines. Biochem Biophys Rep, 2018; 15:39–44.

Rao M. Nitric oxide scavenging by curcuminoids. J Pharm Pharmacol, 1997; 49(1):105–7.

Rivera DE, Ocampo YC, Castro JP, Barrios L, Díaz F, Franco LA. A screening of plants used in Colombian traditional medicine revealed the anti-inflammatory potential of physalis angulata calyces. Saudi J Biol Sci, 2019; 26(7):1758–1766.

Rop O, Mlekč J, Juríkova T, Valšíkova M. Bioactive content and antioxidant capacity of cape gooseberry fruit. Open Life Sci, 2012; 7(4):672–9.

Salcibert A, Manach C, Morand C, Rémésy C, Jiménez L. Dietary polyphenols and the prevention of diseases. Crit Rev Food Sci Nutr, 2005; 45(4):287–306.

Sudha K, Basham С, Malek EA, Ismail M. Phytochemical and antioxidant properties of Manilkara zapota (L.) Proen fruit extracts and its formulations for cosmeceutical application. Asian J Plant Sci Res, 2017; 7(3):29–41.

Skrovinkova S, Sumczynski D, Mlekč J, Juríkova T, Sochor. Bioactive compounds and antioxidant activity in different types of berries. Int J Mol Sci, 2015; 16(10):24673–706.

Slavin JL, Lloyd-B. Health benefits of fruits and vegetables. Adv Nutr, 2012; 3(4):506–16.

Stanley R, Dietzen R, Gidley, M. Proceedings of the tropical fruits in human nutrition and health conference 2008. Held at Couran Cove Island Resort, Gold Coast, Australia.

Sudha K, Baskaran D, Ramasamy D, Siddarth M. Evaluation of functional properties of hydrolyzed undatus (White dragon fruit). Int J Agric Sci Res, 2017; 7(5):451–6.

Tawe G, Kutea V. Physalis edulis. Medicinal spices and vegetables from Africa. Elsevier, Amsterdam, Netherlands, pp 513–26.

Tan BL, Norhaizan ME, Chan LC. Manilkara zapota (L.) R. Royen leaf water extract induces apoptosis in human hepatocellular carcinoma (HePG2) cells via ERK1/2/Akt1/NFκB signaling pathways. Evid Based Complement Alternat Med, 2018; 2018:17.

Torres Grisales Y, Melo Sabogal DV, Sofia Torres-Valenzuela L, Andrea Serna-Jiménez J, Sanín Villarreal A. Evaluation of bioactive compounds with functional interest from yellow pitahaya (Selenicereus megalanthus Haw.). Rev Fac Nac Agronomía Medellín, 2017; 70(3):8311–8.

Vasco C, Ruales J, Kamal-Eldin A. Total phenolic compounds and antioxidant capacities of major fruits from Ecuador. Food Chem, 2008; 111(4):816–23.

Wang X, Ouyang Y, Liu J, Zhu M, Zhao G, Bao W, Hu FB. Fruit and vegetable consumption and mortality from all causes, cardiovascular disease, and cancer: systematic review and dose-response meta-analysis of prospective cohort studies. BMJ, 2014; 349:g4490.

Woo P, Yim H, Khooh H, Sia C, Ang Y. Effects of extraction conditions on antioxidant properties of sapodilla fruit (Manilkara zapota). J Food Res Int, 2013; 20(5):2065.
Y Aboul-Enein H, Berczynski P, Kruk I. Phenolic compounds: the role of redox regulation in neurodegenerative disease and cancer. Mini Rev Med Chem, 2013; 13(3):385–98.

Yahia E, Barrera A. Antioxidant capacity and correlation with phenolic compounds and carotenoids in 40 horticultural commodities. VI Int Postharvest Symp, 2011; 877.

Yahia EM, Gutiérrez-Orozco F, Arvizu-de Leon, C. Phytochemical and antioxidant characterization of maneye (Pouteria sapota Jacq. HE Moore & Stearn) fruit. Food Res Int, 2011; 44(7):2175–81.

Zamudio-Cuevas Y, Diaz-Sobac R, Vázquez-Luna A, Landa-Solis C, Cruz-Ramos M, Santamaría-Olmedo M, Martínez-Flores K, Fuentes-Gómez A, López-Reyes A. The antioxidant activity of soursop decreases the expression of a member of the NADPH oxidase family. Food Funct, 2014; 5(2):303–9.

Zapata S, Piedrahita AM, Rojano B. Capacidad atrapadora de radicales oxígeno (ORAC) y fenoles totales de frutas y hortalizas de Colombia. Perspectivas en Nutrición Hum, 2014; 16(1):25–36.

Zhang N, Diao Y, Hua R, Wang J, Han S, Li J, Yin Y. Nitric oxide-mediated pathways and its role in the degenerative diseases. Front Biosci, 2017; 22:824–34.

Zhang Y-J, Gan R-Y, Li S, Zhou Y, Li A-N, Xu D-P, Li H-B. Antioxidant phytochemicals for the prevention and treatment of chronic diseases. Molecules, 2015; 20(12):21138–56.

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