A novel method for mapping village-scale outdoor resting microhabitats of the primary African malaria vector, *Anopheles gambiae*

Julius R. Dewald1*, Douglas O. Fuller1, Günter C. Müller2 and John C. Beier3

**Abstract**

**Background:** Knowledge of *Anopheles* resting habitats is needed to advance outdoor malaria vector control. This study presents a technique to map locations of resting habitats using high-resolution satellite imagery (world view 2) and probabilistic Dempster-Shafer (D-S) modelling, focused on a rural village in southern Mali, West Africa where field sampling was conducted to determine outdoor habitat preferences of *Anopheles gambiae*, the main vector in the study area.

**Methods:** A combination of supervised and manual image classification was used to derive an accurate land-cover map from the satellite image that provided classes (i.e., photosynthetically active vegetation, water bodies, wetlands, and buildings) suitable for habitat assessment. Linear fuzzy functions were applied to the different image classes to scale resting habitat covariates into a common data range (0–1) with fuzzy breakpoints parameterized experimentally through comparison with mosquito outdoor resting data. Fuzzy layers were entered into a Dempster-Shafer (D-S) weight-of-evidence model that produced pixel-based probability of resting habitat locations.

**Results:** The D-S model provided a highly detailed suitability map of resting locations. The results indicated a significant difference ($p < 0.001$) between D-S values at locations positive for *An. gambiae* and a set of randomly sampled points. Further, a negative binomial regression indicated that although the D-S estimates did not predict abundance ($p > 0.05$) subsequent analysis suggested that the D-S modelling approach may provide a reasonable estimate locations of low-to-medium *An. gambiae* density. These results suggest that that D-S modelling performed well in identifying presence points and specifically resting habitats.

**Conclusion:** The use of a D-S modelling framework for predicting the outdoor resting habitat locations provided novel information on this little-known aspect of anopheline ecology. The technique used here may be applied more broadly at different geographic scales using Google Earth, Landsat or other remotely-sensed imagery to assess the malaria vector resting habitats where outdoor control measures can reduce the burden of the disease in Africa and elsewhere.

**Keywords:** Species distribution modeling, *Anopheles*, Dempster-Shafer modeling, Mali, Resting habitats

*Correspondence: j.dewald@umiami.edu
1 Department of Geography and Regional Studies, University of Miami, Coral Gables, FL, USA
Full list of author information is available at the end of the article

© 2016 The Author(s). This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.
Background

Malaria remains one of the most serious public health problems in the developing world, and is considered a high priority for control and elimination within endemic regions [1–4]. Programmes such as the Roll Back Malaria (RBM) have helped reduce malaria by 47 % between 2000 and 2013 globally and by 54 % in the World Health Organization (WHO) African Region [5]. Further, the WHO has set new goals for global malaria reduction by 2030, which include the reduction of global malaria incidence and mortality rates by at least 90 %, as well as elimination of the disease in at least 35 endemic countries [6]. In West Africa, the disease remains particularly problematic, with 15 out of 18 West African countries having the highest malaria transmission rates on the continent.

The spatially extensive nature of malaria in Africa can be explained by the ecology and behaviour of a highly competent vectorial system of Anopheles mosquitoes, primarily Anopheles gambiae, Anopheles arabiensis of the An. gambiae complex, and Anopheles funestus [7]. Complicating control strategies, these vector species often display different habitat preferences and life histories. For example, adult An. gambiae and An. funestus feed frequently and predominantly on humans, rest mainly inside houses (i.e., endophyly), and can survive for relatively longer periods (39–44 days) relative to adult An. arabiensis, which typically survive between 11 and 17 days [8, 9]. Peak densities of An. gambiae and An. arabiensis follow seasonal patterns of rainfall and both use a range of freshwater larval habitats to breed [10]. An. funestus typically proliferates in permanent swamps and reaches peak densities after seasonal rains into the dry season [10]. An. arabiensis, which has the most extensive geographic range in all of dominant vectors in sub-Saharan Africa, typically favours more arid habitats relative to the other dominant African vectors [11]. This species has proven difficult to control because of its outdoor (exophilic) resting and combined anthropophilic and zoophilic feeding behaviours [12, 13]. This study focused only on An. gambiae, which was the most common species in the study area. Although studies have shown that An. gambiae is primarily endophilic [14–17], other studies have also shown that adult An. gambiae females also prefer outdoor environments [18–21]. The ability of An. gambiae to survive indoors as well as outdoor environments reveals a gap in the understanding of this species’ ecology.

In recent years, much of the effort to control anopheles has focused on development of outdoor strategies that target larvae [22–24]. Relatively novel methods such as attractive toxic sugar baits (ATSB) provide effective means to control adult mosquito populations within and around settlements where mosquitoes typically rest. The use of ATSB, for example, has been shown to decrease male and female An. gambiae populations by 90 % and eliminate most older females [25, 26].

Efficient use of these adult targeted outdoor vector control strategies largely depends on knowledge of where the adult mosquitoes are within the environment surrounding communities at risk. Resting sites of adult mosquitoes in general are poorly documented in the literature, despite their importance in the life cycles of these organisms [27–29]. Resting habitats for mosquitoes are defined as areas where mosquitoes remain after emergence, after taking blood meals and/or before oviposition or during periods of inactivity during the daylight hours. Identification of high-probability resting habitats can advance vector control in rural areas by leading to more efficient targeting of vector control interventions around rural villages and large settlements [30]. Thus, highly detailed maps that display the location of resting habitats can provide a key tool to guide outdoor control efforts, particularly placement of ATSB bait stations, insecticide spraying, and larvicides that reduce mosquito populations.

As small poikilothermic organisms, mosquitoes [31] seek shaded areas for the majority of the day to avoid severe desiccation and heat stress that may result in direct sun in tropical locations [32]. This suggests that the daytime resting habitats for these organisms are usually associated with dense vegetation, which provides shade and thus a cooler microclimate relative to open areas such as bare soil, agricultural fields, and built-up spaces. Furthermore, proximity to water bodies and to blood meals provides important environmental resources for Anophelines, as they require shallow, temporary bodies of fresh water that may contain floating and submerged algae, emergent grass, or rice to deposit their eggs [11, 33–39]. The proximity to water is thus considered an important factor when identifying resting habitats [40–43].

Various methods exist to map habitat suitability and presence of mosquitoes from point observations, including Maxent [44] and genetic algorithms [45]. However, these methods are often employed in conjunction with multiple bioclimatic and topographic covariates to map probable presence over large regions or at continental scales. Moreover, to map habitat preferences at local (i.e., village) scales at high resolution (<10 m), different methods may be required in developing countries where high-resolution climate and topographic data are often unavailable at the local scales and spatial resolutions needed to accurately guide vector control activities. However, land cover maps derived from multispectral satellite imagery can provide a range of potential covariates that are specific to mosquito habitat preferences.
This study employs Dempster-Shafer (D-S) weight-of-evidence modelling, a variant of Bayesian Theory to generate probability surfaces of habitat suitability. Unlike the Bayesian Theory, the D-S approach does not assume that one has full information and thus D-S models work well when one has incomplete knowledge that changes over time [46, 47] as validation data and other information become available. As the results are based on a small set of land cover classes derived from multispectral imagery, this study demonstrates the feasibility of modelling probability of vector presence and potentially abundance using a more limited set of covariates than are typically employed in other species distribution models.

Methods

Study area

The study area was centered around the village of Kenieroba (see Fig. 1), which is located 71 km southwest of Bamako, Mali. The study area covers approximately a 25 square km surrounding the village, which contained approximately 1000 houses as visually estimated from a high resolution WorldView 2 image of the area. A floodplain of about 2 km wide separates Keineroba from the Niger River. During the rainy season (June–September) villagers use parts of the floodplain for rice agriculture, which may be used by vectors as larval habitat. This study site was selected due to its mixture of various land covers, which include a close proximity to the Niger River, dense agricultural plots and sparse natural vegetation, along with areas of human occupation. Focus was placed on the prediction of resting habitats during the early dry season when adult vector densities are generally low relative to the wet season. During the dry season larval habitats become limited as the surface area of water bodies decreases owing to evaporation, whereas these habitats increase sharply at the onset of the rainy season and become abundant and extensive where micro-depressions are found in the landscape [48]. Focusing on mosquito control efforts during the dry season, when vector populations are typically limited by low water availability, may provide a more effective way to control local mosquito populations relative to control efforts during the wet season [49].

Field data

Field teams collected adult mosquitoes using drop-net collection methods during November and December 2013. The drop-net captured adult mosquitoes resting in grasses and other herbaceous vegetation near woody canopies as well as in open areas that were not underneath any trees or shrubs. Each drop-net enclosed a 2 × 2 m area and was deployed at predetermined microhabitat locations representing a variety of outdoor habitats. Each drop-net contained a suspended Center for Disease Control light trap (CDC-UV light trap) to capture the resting mosquitoes in the enclosed space within the net. The locations of each sampled location can be seen in Fig. 1, and close up views can be seen in Fig. 2.

Study area A was situated next to an existing wetland area by the River Niger, and specifically the sampling locations in this area radiated out from a temporary lagoon. Sampling areas B and C were located in and around riverine forests at opposite banks of the Niger River. The remaining sampling locations were placed at points of interest across the study area and have been grouped together as Area D. These points of interest include mango plantations, open agricultural fields, sandy islands, and various savanna habitats. Sampling in each microhabitat type was done during a 2-to-3 day period and a total of 36 drop-nets were employed in each sampling location with a total of 18 placed in mornings and 18 in the afternoons. After 12 h, the traps were removed and the Anophelines were counted for each site to provide an estimate of abundance at each microhabitat location (see Table 1). Furthermore, the mosquito species collected were identified using PCR (polymerase chain reaction) [50]. Only An. gambiae were included in this study. The locations for these microhabitat sampling locations are shown in Fig. 2. These sampling locations
were chosen using a non-random sampling design that aimed to assess different habitats where anopheline mosquitoes were likely to be found. To assess spatial autocorrelation in the field observations, a Moran’s I statistic was applied to the abundance data.

**Remotely-sensed imagery**

A cloud-free, high-resolution WorldView 2 satellite image from March 5, 2013 (2.0 m spatial resolution) was used to map land cover around the study site. The imagery contained four multispectral bands covering the

---

**Fig. 2** Detailed images showing sampling areas and exact locations where drop nets were placed. The numbers refer to the ID given to each sampling location and correspond with the ID values found in Table 1.
red (0.63–0.69 μm), green (0.51–0.58 μm), blue (0.45–0.51 μm), and near infrared (0.705–0.745 μm) wavelengths. Although image archives did not yield cloud-free WorldView 2 images coincident with the field sampling, the imagery covered the same dry season during which field collections occurred. A classified map was generated from the WorldView 2 imagery using a combination of supervised classification and an image segmentation algorithm to identify discrete objects such as clumps of dense woody vegetation, metal-roofed buildings, water bodies, open fields, and bare earth. Owing to spectral similarity, it was not possible to accurately separate woody vegetation and photosynthetically active grass canopies. However, most areas classified as dense vegetation were associated with tree crowns as the majority of grasses had senesced during this time of the year. A segmentation algorithm was applied to improve classification results as this approach often outperforms per-pixel classification approaches [51, 52]. The segmentation merged pixels into non-overlapping homogeneous objects, which was then used to assign land cover groups for a classification method [53]. An additional 'wetlands' class was also included in the final classified image by identifying low-lying areas of areas of alluvial soils that were inundated during field sampling.

### Classification accuracy

Accuracy assessment of the classified image was done using a colour composite image and 280 random points created in ArcMap. The points were randomly stratified across the different classes and care was taken to ensure that the points used for accuracy assessment and training were mutually exclusive. A nearest neighbour analysis was employed to confirm that the points were randomly distributed. The wetlands class was not included in the classification accuracy assessment as it was created independent of the classification algorithms. To further increase class accuracy, a modal filter with a 3 × 3 window was used to eliminate isolated, misclassified pixels. Obviously misclassified features were corrected manually using Google Earth imagery as a cross-reference. Specifically, certain shallow water bodies were classified as metal-roofed buildings by the segmentation algorithm due to the spectral similarity across the four bands. It is also important to note that only buildings with metal roofs were identified in the classified map. These structures comprised approximately 45 % of the total buildings, with the remainder having roofs constructed from thatch materials that could not be distinguished spectrally from senesced vegetation. Furthermore, the size of

| ID | Area | Mosquito sum | Sex: F/M | ID | Area | Mosquito sum | Sex: F/M | ID | Area | Mosquito sum | Sex: F/M |
|----|------|--------------|----------|----|------|--------------|----------|----|------|--------------|----------|
| 1  | A    | 0            | 0/0      | 23 | C    | 1            | 1/0      | 45 | D    | 0            | 0/0      |
| 2* | A    | 3            | 2/2      | 24 | C    | 2            | 2/0      | 46 | D    | 0            | 0/0      |
| 3  | A    | 115          | 55/60    | 25* | C    | 323          | 241/82   | 47* | D    | 0            | 0/0      |
| 4* | A    | 58           | 38/20    | 26 | C    | 6            | 5/1      | 48 | D    | 0            | 0/0      |
| 5  | A    | 188          | 113/75   | 27 | C    | 128          | 104/24   | 49 | D    | 0            | 0/0      |
| 6* | A    | 38           | 23/15    | 28 | C    | 201          | 143/58   | 50 | D    | 0            | 0/0      |
| 7  | A    | 1            | 1/0      | 29 | C    | 182          | 143/39   | 51 | D    | 0            | 0/0      |
| 8  | A    | 0            | 0/0      | 30 | C    | 21           | 15/6     | 52 | D    | 0            | 0/0      |
| 9  | B    | 4            | 3/1      | 31 | C    | 2            | 2/0      | 53 | D    | 0            | 0/0      |
| 10 | B    | 33           | 23/10    | 32* | C    | 0            | 0/0      | 54 | D    | 0            | 0/0      |
| 11 | B    | 215          | 164/51   | 33 | C    | 7            | 6/1      | 55* | D    | 0            | 0/0      |
| 12 | B    | 113          | 89/24    | 34 | D    | 0            | 0/0      | 56* | D    | 0            | 0/0      |
| 13*| B    | 19           | 16/3     | 35 | D    | 0            | 0/0      | 57 | D    | 0            | 0/0      |
| 14 | B    | 44           | 36/8     | 36* | D    | 0            | 0/0      | 58 | D    | 0            | 0/0      |
| 15 | B    | 1            | 1/0      | 37* | D    | 0            | 0/0      | 59* | D    | 0            | 0/0      |
| 16 | B    | 4            | 4/0      | 38 | D    | 0            | 0/0      | 60 | D    | 0            | 0/0      |
| 17 | B    | 209          | 143/66   | 39 | D    | 0            | 0/0      | 61* | D    | 0            | 0/0      |
| 18 | C    | 1            | 1/0      | 40 | D    | 0            | 0/0      | 62 | D    | 0            | 0/0      |
| 19*| C    | 3            | 2/1      | 41* | D    | 0            | 0/0      | 63* | D    | 0            | 0/0      |
| 20 | C    | 14           | 11/3     | 42 | D    | 0            | 0/0      | 64 | D    | 0            | 0/0      |
| 21 | C    | 116          | 96/20    | 43 | D    | 0            | 0/0      | 65 | D    | 0            | 0/0      |
| 22 | C    | 138          | 112/26   | 44 | D    | 0            | 0/0      | 66 | D    | 0            | 0/0      |

Pearson Correlation Coefficient of male to female resting site preference: 0.916

* Points that were excluded from the D-S model training and reserved for testing
these buildings was very small, the majority of the buildings being only 3–5 m in diameter.

**D-S modelling**

Two hypotheses were used to construct a framework for D-S modelling of resting habitat probability. Firstly, as vegetation can provide substantial amounts of shade, there was an assumption that resting habitats are associated with areas consisting of savanna trees, shrubs, and grasses. Further, the hypothesis was formulated based on the assumption that resting areas are found close to water and that such locations will likely contain higher densities of resting mosquitoes than areas distant (e.g., >1 km) from water bodies. While proximity to the potential blood meals should also influence resting site selection, this hypothesis was excluded from the D-S modelling framework as no outdoor-resting anopheles mosquitoes were found in any of the sampling locations near human habitations.

The classes of dense vegetation, open water, and wetlands were extracted from the classified image and transformed into distance layers using the distance module in IDRISI GIS software [46]. The distance module calculates the distance of each pixel in the study area to the nearest set of target pixels (e.g., vegetation, open water, or wetlands). These distance maps were then converted into fuzzy classes that were scaled from 0 to 1 representing the degree of class membership [54]. A key consideration was the establishment of breakpoints and function selection for fuzzy set membership with breakpoints established where membership began to fall below 1 (breakpoint C) to where membership becomes 0 (breakpoint D). The different fuzzy functions (sigmoidal, J-shaped curve, and user-defined linear) determined the shape of the transition from full to zero class membership (i.e., 1–0).

48 field sample sites (75 %) were randomly selected for in the creation of the D-S model (see Table 1). To create the D-S model, each parameter was entered into the Belief module in the IDRISI GIS software with an indication of their supported hypothesis (resting habitat or not). The module then accumulated evidence in support of each hypothesis by calculating a state of knowledge using the Dempster-Shafer rule of combination, from which a belief image representing the total support for the hypothesis was produced. Multiple iterations of D-S models were created to adjust the fuzzy breakpoint values. The D-S probability values of multiple D-S model iterations were plotted against distance for sample sites that had zero mosquito counts. Each plot revealed how changes in the breakpoint values affect the D-S probability values for each land cover while the other variables are kept constant. The distance at which the probability value began to rise above zero (p > 0) was plotted and as a way to determine an appropriate breakpoint for each fuzzy function included in the D-S model and the mean distance where the condition p > 0 was true for each set of locations with zero mosquito counts was used to establish the final breakpoint value. In this way, commission errors were limited to zero. In the case of curves representing distances from vegetation (n = 6), water (n = 8), and wetland (n = 4), the mean p > 0 values were 17.5 (sd = 8.22), 568.75 (sd = 402.61), and 77.5 (sd = 20.21) respectively. These distances were then used as the final D-S break points values for the fuzzy layers. Figure 3 provides a graphical example of how breakpoint determination was done using this approach.

As a way to validate the results, a t test was performed for independent samples (assuming unequal variances) to compare D-S values for the 30 locations that were positive for *An. gambiae* with a sample of 50 random points. Further, since the field data provided mosquito counts in different locations, a negative binomial model (SPSS Version 22, IBM Corp. 2013. Armonk, NY: IBM Corp) was used to assess whether the D-S model may be related to abundance of *An. gambiae* in the landscape. In so doing, the assumption was made that D-S probability should be positively related to mosquito abundance. The negative binomial regression used the remaining 25 % (16 locations) of sampling locations that were omitted from the D-S model.

**Results**

Only *An. gambiae* mosquitoes were found in field sampling. 34 of the sampled points produced no resting *An. gambiae* mosquitoes during any point of the sampling period and at sampling locations where *An. gambiae* mosquitoes were found the number of mosquitoes captured range from a single mosquito up to 323 mosquitoes. In total, 2190 *An. gambiae* mosquitoes were collected, with 1595 females and 595 males. During morning sampling, a total of 1219 mosquitoes were collected, and 971 were obtained during the afternoons. PCR analysis of 50 adults indicated that all collected specimens belonged to *An. gambiae s.s*. No attempt was made to distinguish between *An. gambiae* and *Anopheles coluzzii*, [55]. Figure 4 provides a depiction of mosquito catches per sampling location. Further, Moran’s I produced values of 0.091, a variance of 0.008, a z-score of 1.17, and a p value of 0.24, which indicates that the spatial structure of the field data did not appear to be significantly different from random. Further, a Pearson correlation was performed between the average amount of female and male mosquitoes captured per site. The results of this analysis are shown in Table 1. A high Pearson correlation coefficient of 0.916 suggests that there was no significant difference in resting site preference between males and females.
The nearest neighbour analysis of points used to validate the imagery indicated a spatially random distribution ($z$-score $= -1.60$ and a p value of 0.109) (Fig. 5). The error matrix revealed that the overall accuracy of the classified map was 82%. Individual class accuracies related to anopheline habitats were generally high (Table 2). For example, the accuracies for dense vegetation, water, and metal-roofed buildings were of 96, 97.7, and 90.9 % respectively. In addition, the open field class had an accuracy of 95.6 %, while the bare earth class had an accuracy of 60.7 %.

**D-S model analysis**

Figure 6 shows the D-S model belief output, which represents expected outdoor resting habitats. Meanwhile, Table 3 provides the parameters pertaining to the break.
points and the functions used for the individual land covers used in the D-S model creation. The model surface in Fig. 6 indicates that resting habitats were generally restricted to specific locations near the Niger River and other water bodies (wetlands and open water), particularly densely vegetated areas. The findings from the belief output are supported by previous studies indicating that shade and moisture availability are primary factors for exophilic resting habitats [22, 23, 30–32, 35]. The majority of the study site was found to be unsuitable for resting, despite possible proximity to human settlements (i.e., a proxy for blood meal availability). Direct comparisons between the D-S model output and the mosquito data are shown in Figs. 7 and 8.

Consistent with the qualitative analysis of the model surface (Fig. 6), the t test comparing mean values for positive locations (n = 30) with 50 randomly sampled points indicated a significant difference (p < 0.001) between these two samples (Fig. 9). Further, the negative binomial regression failed to indicate a significant relationship (p > 0.05) between mosquito counts and the D-S surface values. However, when the same analysis was performed after removal of an outlier where a high mosquito count was obtained (n = 323 catches), the relationship between abundance and D-S model values was significant (Wald Chi Square = 18.86, p = 0.00, df = 14). This suggests that the D-S model results produced a relatively realistic depiction of both presence and abundance of An. gambiae at low to moderate density, but appeared insufficient to estimate areas of very high density populations. This assumption is consistent with the work of other researchers [56, 57] who have found positive relationships between predicted probability of presence and abundance within a range of taxa when using species distribution models such as, Maxent.

**Discussion**

This study is the first to employ D-S modelling to assess probability of outdoor resting habitats of An. gambiae at a scale that is relevant for guiding vector management. Both the t test and a negative binomial model suggest that the D-S model performed well in distinguishing between suitable and unsuitable outdoor resting microhabitats as well as predicting the abundance of resting anopheline

### Table 2 Classification accuracy

| Class                   | Accuracy (%) |
|-------------------------|--------------|
| Dense vegetation        | 96           |
| Water                   | 97.7         |
| Metal roofed buildings  | 90.9         |
| Open field              | 95.6         |
| Bare earth              | 60.7         |
| Overall accuracy        | 82           |

### Table 3 D-S model parameters

| Parameter                  | Land cover feature          | Variant        | Function       | C-breaking point (m) | D-breaking point (m) |
|----------------------------|-----------------------------|----------------|----------------|----------------------|----------------------|
| Wetlands                   | Wetlands                    | Decreasing     | Linear         | 0                    | 77.5                 |
| Dense vegetation near open water* | Open water          | Decreasing     | Sigmoidal      | 0                    | 569.75               |
| Dense vegetation**         |                            | Decreasing     | Linear         | 0                    | 17.5                 |

Land cover feature describes the land cover that has been extracted from the classified image for the creation of the parameter. C-breaking point refers to the distance from the land cover at which the fuzzy value begins to drop from 1 down to a value of 0 with that distance indicated by breaking point D. The decreasing variant described that the fuzzy value approached 0 rather than 1. The function refers to what method of slope was used to create the fuzzy layer.

* Parameter relied on two separate fuzzy layers overlaid through multiplication

** User-defined linear relationship, where the maximum predictive value was set at 0.50
Fig. 7  Comparison of mosquito presence/absence with the final D-S model

Fig. 8  Mosquito abundances at the field sites compared to final D-S model
mosquitoes at these sites at low to moderate densities ranging from about 0–60 specimens per site. This study’s D-S model surface thus helps to advance understanding of exophilic resting ecology of Anopheles mosquitoes. Specifically the model indicates that shaded areas nearby water sources are important to exophilic resting habitat selection. This can be seen in the breakpoints used for the fuzzy layers of dense vegetation and open water shaded areas in Table 3. Dense vegetation surrounding open water are likely to have higher amounts of soil moisture. The optimal resting sites were found to be shaded areas close to shallow water bodies such as those represented by the wetlands class of the classified image.

While the D-S model used distances from major land cover type as a way of predicting location of resting habitats, land cover per se is largely a proxy for microclimatic conditions that favour mosquito thermoregulation. These conditions include areas of moist soil that are shaded by a dense over-story of trees and shrubs and are proximate to water sources or influenced by run-on from sheet flow. Further analysis of plant species diversity within the dense vegetation class may yield further insights on resting habitat suitability especially flowering plants that provide sugar-feeding opportunities. Additional investigation may also include assessment of leaf-area index as way to understand how variable sub-canopy light environments provided by trees and shrubs may provide suitable resting sites within clumps of woody vegetation. This study assumes that all areas covered by the dense vegetation class are spatially homogeneous in their provision of shade and sugars. However, it is likely that some tree species may provide more shade and feeding opportunities than others due to different leaf areas, and this may affect the quality of resting habitats. Furthermore, flowering phenology may control the distribution of sugar resources available at different points in the seasonal cycle and therefore the distribution and density of anophelines may shift in response to the availability of plant nectars. Thus, flowering patterns would likely impact mosquito concentrations and would likely lead to the development of more robust spatial predictions of habitat suitability during the seasonal cycle. The inability for the current D-S model to accurately take these conditions into consideration may account for the model’s limited ability to predict areas of high mosquito density. Finally, this study sampled mosquitoes in the herbaceous layer only and further sampling of resting habitats in the tree and shrub canopies may advance understanding of how resting anophelines are distributed among different canopy layers.

Despite several limitations, the D-S model driven by a small set of satellite-derived land-cover classes provided a relatively simple way to map probability of presence at high spatial resolution. Previous research for mapping mosquito presence at the village scale in Africa, [57] evaluated similar spatial patterns. A number of other studies have employed 30-m Landsat imagery [58, 59] to derive robust surfaces that indicate where transmission and breeding is likely. In contrast, this study utilized very high-resolution satellite imagery to pinpoint localized environmental factors that are likely to affect vector presence in the environment, which were transformed through the D-S modelling process to a high-resolution probability surface. Overall, however, this approach was consistent with Landsat-based approaches [57, 58] that have demonstrated a clear association between moist, alluvial locations near villages and occurrence of infective anophelines.

**Conclusion**

The insights into resting habitat selection, as revealed by the D-S model provided valuable information for guiding control of anophelines at local scales, which often target specific villages and peri-urban environments where transmission remains problematic. This study also suggests that D-S modelling of resting habitats may be applicable using coarser resolution imagery such as Landsat (i.e., 30 m), which covers much larger areas relative to the WorldView 2 imagery used in our study. Landsat imagery may be used to produce classified maps that delineate land cover features, such as small waterbodies, and associated wetlands, as well as patches of dense woody
vegetation, with high degrees of accuracy [60]. In this way, more spatially extensive maps may be generated to extend mapping of vector resting and sugar-feeding habitats in sub-Saharan Africa.

Abbreviations
ATSB: attractive toxic sugar baits; D-S: Dempster Shafer; IRS: indoor residual spraying; ITNs: insecticide-treated nets.

Authors’ contributions
JRD drafted manuscript, created models, performed statistical analysis. DOF conceived the study along with JCB, and participated in its design and coordination and helped to draft the manuscript and perform statistical analysis. GCM reviewed manuscript and led field data collection. JCB conceived of study, reviewed manuscript, and led field data collection. All authors read and approved the final manuscript.

Author details
1 Department of Geography and Regional Studies, University of Miami, Coral Gables, FL, USA. 2 Kuvin Center for the Study of Tropical and Infectious Diseases, Hadassah Medical School, Hebrew University, Jerusalem, Israel. 3 Department of Public Health Sciences, Miller School of Medicine, University of Miami, Miami, FL, USA.

Acknowledgements
Research reported in this publication was supported by the National Institute of Allergy And Infectious Diseases of the National Institutes of Health under Award Number R01AI109068. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Competing interests
The authors declare that they have no competing interests.

Availability of data and materials
All field data used for this study will be submitted to the Malaria Atlas Project and the imagery produced in this study (land cover and D-S predictive surface) will be submitted to DRYAD upon acceptance of this manuscript.

Consent for publication
All authors have read and consented to the submission of this manuscript for publication.

Received: 14 June 2016   Accepted: 13 September 2016
Published online: 22 September 2016

References
1. Tanner M, Savigny D. Malaria eradication back on the table. Bull World Health Organ. 2008;86:82.
2. Mendis K, Rietveld A, Warsame M, Bosman A, Greenwood B, Weistroffer WH. From malaria control to eradication: the WHO perspective. Trop Med Int Health. 2009;14:802–9.
3. Bousema T, Drakeley C. Epidemiology and infectivity of Plasmodium falciparum and Plasmodium vivax gametocytes in relation to malaria control and elimination. Clin Microbiol Rev. 2011;24:377–410.
4. Cotter C, Sturrock HJW, Hsiang MS, Liu J, Phillips AA, Hwang J, et al. The changing epidemiology of malaria elimination: new strategies for new challenges. Lancet. 2013;382:900–11.
5. WHO. World Malaria report 2014. Geneva: World Health Organization; 2014. http://www.who.int/malaria/publications/world_malaria_report_2014/en/. Accessed 7 Jan 2016.
6. WHO. Global technical strategy for malaria 2016–2030. Geneva: World Health Organization; 2015. http://www.who.int/malaria/areas/global_technical_strategy/en/. Accessed 7 Jan 2016.
7. Coetzee M, Craig M, Le Sueur D. Distribution of African malaria mosquitoes belonging to the Anopheles gambiae complex. Parasitol Today. 2000;16:74–7.
8. Afrane YA, Zhou G, Lawson BW, Githeko AK, Yan G. Effects of microclimatic changes due to deforestation on the survivorship and reproductive fitness of Anopheles gambiae in western Kenya highlands. Am J Trop Med Hyg. 2006;74:772–8.
9. Afrane YA, Zhou G, Lawson BW, Githeko AK, Yan G. Life-table analysis of Anopheles arabiensis in western Kenya highlands: effects of land covers on larval and adult survivorship. Am J Trop Med Hyg. 2007;77:660–6.
10. White GB. Anopheles-gambiae complex and disease transmission in Africa. Trans R Soc Trop Med Hyg. 1974;68:278–302.
11. Sinka ME, Rubio-Palis Y, Manguin S, Patil AP, Temperley WH, Gething PW, et al. The dominant anopheline mosquitoes of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis. Parasit Vectors. 2010;3:72.
12. Coluzzi M. Heterogeneities of the malaria vectorial system in tropical Africa and their significance in malaria epidemiology and control. Bull World Health Organ. 1984;62:107–13.
13. WHO Study Group. Malaria vector control and personal protection. World Health Organ Tech Rep Se. 2006;936:1–62.
14. Githeko AK, Service MW, Mbogo CM, Atelli FK, Juma FO., Origin of blood meals in indoor and outdoor resting malaria vectors in western Kenya. Acta Trop. 1994;58:307–16.
15. Diatta M, Spiegel A, Lochouarn L, Fontenelle D. Similar feeding preferences of Anopheles gambiae and A. arabiensis in Senegal. Trans R Soc Trop Med Hyg. 1998;92:270–2.
16. Antonio-Nkondjio C, Awoono-Ambene P, Toto JC, Meunier JY, Zebaze-Kemieu S, Nyambam R, et al. High malaria transmission intensity in a village close to Yaounde, the capital city of Cameroon. J Med Entomol. 2002;39:350–5.
17. Dabire KR, Diabate A, Pare-Toe L, Rouamba J, Duan A, Fontenille D, et al. Year to year and seasonal variations in vector bionomics and malaria transmission in a humid savannah village in west Burkina Faso. J Vector Ecol. 2008;33:70–5.
18. Bockarie MJ, Service MW, Toure YT, Traore S, Bamish G, Greenwood BM. The ecology and behaviour of the forest form of Anopheles gambiae s.s. Parasitology. 1993;5:5–8.
19. Githeko AK, Adunjo NI, Kanjam DA, Hawley WA, Vulliez MJ, Seroney IK, et al. Some observations on the biting behavior of Anopheles gambiae s.s., Anopheles arabiensis, and Anopheles funestus and their implications for malaria control. Exp Parasitol. 1996;82:106–15.
20. Duchemin JB, Tsy JM, Rabarison P, Roux J, Coluzzi M, Costantini C. Zoophagy of Anopheles gambiae s.l. in coastal regions of the Ivory Coast. Acta Trop. 1994;58:307–16.
21. Cano J, Berzosa PJ, Roche J, Rubio JM, Moyano E, Guerra-Neira A, et al. Malaria vectors in the Bioko Island (Equatorial Guinea): estimation of vector dynamics and transmission intensities. J Med Entomol. 2004;41:158–61.
22. Kilian GF, Fillinger U, Knols BGJ. Advantages of larval control for African malaria vectors: low mobility and behavioral responsiveness of immature mosquito stages allow high effective coverage. Malar J. 2002;1:8.
23. Gu WD, Novak RJ. Habitat-based modeling of impacts of mosquito larval interventions on entomological inoculation rates, incidence, and prevalence of malaria. Am J Trop Med Hyg. 2005;73:546–52.
24. Fillinger U, Ndenga B, Githeko A, Lindsay SW. Integrated malaria vector control with microbial larvicides and insecticide-treated nets in Western Kenya: a controlled trial. Bull World Health Organ. 2009;87:655–65.
25. Muller GC, Jumilla A, Schlein Y. Effective control of adult Culex pipiens by spraying an attractive toxic sugar bait solution in the vegetation near larval habitats. J Med Entomol. 2010;47:63–6.
26. Schlein Y, Mueller GC. An Approach to mosquito control: using the dominant attraction of flowering Tamarix jordanis trees against Culex pipiens. J Med Entomol. 2008;45:384–90.
27. Muller GC, Beier JC, Traore SF, Toure MB, Traore NM, Bah S, et al. Successful field trial of attractive toxic sugar bait (ATSB) plant-spraying methods against malaria vectors in the Anopheles gambiae complex in Mali West Africa. Malar J. 2010;9:210.
28. Burkett-Cadena N, Graham SP, Giovanetto LA. Resting environments of some Costa Rican mosquitoes. J Vector Ecol. 2013;38:12–9.
29. Gillies MT. Studies in house leaving and outside resting of *Anopheles gambiae* Giles and *Anopheles funestus* Giles in East Africa. The outside resting population. Bull Entomol Res. 1954;45:361–74.

30. Gillies MT. The density of adult *Anopheles* in the neighbourhood of an East African village. Am J Trop Med Hyg. 1955;4:1103–13.

31. Afane VA, Klinkenberg E, Drechsel P, Owusu-Daaku K, Garsa R, Kruppa T. Does irrigated urban agriculture influence the transmission of malaria in the city of Kumasi, Ghana? Acta Trop. 2004;89:125–34.

32. Pajamans KP, Thomas MB. The influence of mosquito resting behaviour and microclimate for malaria risk. Malar J. 2011;10:183.

33. May M. Insect thermoregulation. Annu Rev Entomol. 1979;24:313–49.

34. Sumba LA, Okoth K, Deng AL, Githure J, Knols BG, Beier JC, et al. Daily oviposition patterns of the African malaria mosquito *Anopheles gambiae* Giles (Diptera: Culicidae) on different types of aqueous substrates. J Circadian Rhythms. 2004;2:1–7.

35. Edillo FE, Toure YT, Lanzaro GC, Dolo G, Taylor CE. Spatial and habitat distribution of *Anopheles gambiae* and *Anopheles arabiensis* (Diptera: Culicidae) in Banambani village, Mali. J Med Entomol. 2002;39:70–7.

36. Bogh C, Clarke SE, Jawara M, Thomas CJ, Lindsay SW. Localized breeding of the *Anopheles gambiae* complex (Diptera: Culicidae) along the river Gambia West Africa. Bull Entomol Res. 2003;93:279–87.

37. Koenraadt CJM, Githeko AK, Takken W. The effects of rainfall and evapotranspiration on the temporal dynamics of *Anopheles gambiae* s.s. and *Anopheles arabiensis* in a Kenyan village. Acta Trop. 2004;90:141–53.

38. Minakawa N, Sonye G, Mogi M, Yan G. Habitat characteristics of *Anopheles gambiae* s.s. larvae in a Kenyan highland. Med Vet Entomol. 2004;18:301–5.

39. Awolola TS, Oduola AO, Obansa JB, Chukwurar NJ, Unyimadu JP, Gillies MT. Studies in house leaving and outside resting of *Anopheles gambiae* Giles and *Anopheles funestus* Giles in East Africa. The outside resting population. Bull Entomol Res. 1954;45:361–74.

40. Trape JF, Lefebvre-Zante E, Legros F, Ndiaye G, Bouganali H, Druilhe P, et al. Daily oviposition patterns of the African malaria mosquito *Anopheles gambiae* Giles (Diptera: Culicidae) on different types of aqueous substrates. J Circadian Rhythms. 2004;2:1–7.

41. Edillo FE, Toure YT, Lanzaro GC, Dolo G, Taylor CE. Spatial and habitat distribution of *Anopheles gambiae* and *Anopheles arabiensis* (Diptera: Culicidae) in Banambani village, Mali. J Med Entomol. 2002;39:70–7.

42. Bogh C, Clarke SE, Jawara M, Thomas CJ, Lindsay SW. Localized breeding of the *Anopheles gambiae* complex (Diptera: Culicidae) along the river Gambia West Africa. Bull Entomol Res. 2003;93:279–87.

43. Koenraadt CJM, Githeko AK, Takken W. The effects of rainfall and evapotranspiration on the temporal dynamics of *Anopheles gambiae* s.s. and *Anopheles arabiensis* in a Kenyan village. Acta Trop. 2004;90:141–53.

44. Minakawa N, Sonye G, Mogi M, Yan G. Habitat characteristics of *Anopheles gambiae* s.s. larvae in a Kenyan highland. Med Vet Entomol. 2004;18:301–5.

45. Foley DH, Klein TA, Kim HC, Brown T, Wilkerson RC, Rueda LM. Validation of ecological niche models for potential malaria vectors in the Republic of Korea. J Am Mosq Control Assoc. 2010;26:210–3.

46. Malpica JA, Alonso MC, Sanz MA. Dempster-Shafer theory in geographic information systems: a survey. Expert Syst Appl. 2007;32:67–55.

47. Gordon J, Shortliffe EH. A method for managing evidential reasoning in a hierarchical hypothesis space. Artif Intell. 1985;26:323–57.

48. Charlwood JD, Vij R, Billingsley PF. Dry season refugia of malaria-transmitting mosquitoes in a dry savannah zone of East Africa. Am J Trop Med Hyg. 2000;62:726–32.

49. Baber I, Keita M, Sogoba N, Konate M, Diallo M, Doumbia S, et al. Population size and migration of *Anopheles gambiae* in the Bancoumana region of Mali and their significance for efficient vector control. PLoS ONE. 2010;5:e10270.

50. Scott JA, Brogdon WG, Collins FH. Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction. Am J Trop Med Hyg. 1993;49:320–9.

51. Lu D, Weng Q. A survey of image classification methods and techniques for improving classification performance. Int J Remote Sens. 2007;28:823–70.

52. Benz UC, Hofmann P, Willhauck G, Lingenfelder I, Heynen M. Multi-resolution, object-oriented fuzzy analysis of remote sensing data for GIS-ready information. ISPRS J Photogramm Remote Sens. 2004;58:239–58.

53. Thomas N, Hendrix C, Congalton RG. A comparison of urban mapping methods using high-resolution digital imagery. Photogramm Eng Remote Sens. 2003;69:963–72.

54. Jiang H, Eastman JR. Application of fuzzy measures in multi-criteria evaluation in GIS. Int J Geogr Inf Sci. 2000;14:173–84.

55. Coetzee M, Hunt R, Wilkerson R, Torre AD, Coulibaly M, Besansky N. *Anopheles coluzzii* and *Anopheles arabiensis*, new members of the *Anopheles gambiae* complex. Zootaxa. 2013:3619:246–74.

56. VanDerWal J, Shoo LP, Johnson CN, Williams SE. Abundance and the environmental niche: environmental suitability estimated from niche models predicts the upper limit of local abundance. Am Nat. 2009;42:229–40.

57. Ricotta EE, Frese SA, Choobwe C, Louis TA, Shiff CJ. Evaluating local vegetation cover as a risk factor for malaria transmission: a new analytical approach using ImageJ. Mol J. 2014:13:94.

58. Bogh C, Lindsay SW, Clarke SE, Jawara M, Dean A, Thomas CJ. Monitoring mosquitoes in a dry savannah zone of East Africa. Am J Trop Med Hyg. 2003;69:244–6.

59. Clennon JA, Kamanga A, Musapa M, Shiff C, Glass GE. Identifying malaria vector breeding habitats with remote sensing data and terrain-based landscape indices in Zambia. Int J Health Geogr. 2010;5:58.

60. Frazier PS, Page KJ. Water body detection and delineation with Landsat TM data. Photogramm Eng Remote Sens. 2000:66:1461–7.