Abstract: Meat and meat products have been described as a very good source of angiotensin I converting enzyme (ACEI)-inhibitory peptides. The generation of bioactive peptides can occur through the action of endogenous muscular enzymes during processing, gastrointestinal digestion, or by using commercial enzymes in laboratory or industry under controlled conditions. Studies of bioavailability are necessary in order to prove the positive health effect of bioactive peptides in the body as they should resist gastrointestinal digestion, cross the intestinal barrier, and reach bloodstream and target organs. However, in order to better understand their effect, interactions, and bioavailability, it is necessary to consider food matrix interactions and continue the development of quantitative methodologies in order to obtain more data that will enable advances in the field of bioactive peptides and the determination of their influence on health.

Keywords: meat; peptides; antihypertensive; peptidomics

1. Introduction

Bioactive peptides derived from food proteins can exert different effects after their absorption in the human body, such as prevention of diseases or physiological modulation. Physiological properties such as antihypertensive, antioxidant, antithrombotic, or hypocholesterolemic activity in the cardiovascular system [1,2]; mineral binding, antidiabetic, antimicrobial, or anti-inflammatory effects in the gastrointestinal system [3,4]; cytomodulatory or immunomodulatory actions in the immune system [5]; and opioid agonist or antagonist activity in the nervous system [6] have been recently described to be exerted by different food-derived peptides [7].

The activity of angiotensin I-converting enzyme (ACEI) inhibitors has been extensively studied over the last decade. The main reason for this interest is the relevance of hypertension in the development of cardiovascular diseases, which is the most important public health problem of this century. In this respect, different synthetic drugs are available on the market for the treatment of hypertension but their numerous side effects have focused researchers’ interest on the search of alternative non-toxic and naturally generated peptides for controlling blood pressure [8].

ACEI is a dipeptidyl carboxypeptidase enzyme that participates in the renin–angiotensin system (RAS) and converts angiotensin-I into the vasoconstrictor angiotensin-II by cleaving two amino acids at the same time, thus inactivating the vasodilator bradykinin. The role of ACEI inhibitors is to maintain the balance between the vasoconstrictive and salt-retentive effects of angiotensin-II and vasodilator effects of bradykinin (Figure 1). Thus, the main interest for studying ACEI-inhibitory natural peptides is due to their capacity to inhibit ACEI, which lead to a decrease in blood pressure by inactivating the formation of angiotensin-II [9].
Figure 1. The renin-angiotensin system (RAS). ACEI: angiotensin I-converting enzyme; ACEII: angiotensin II-converting enzyme.

In this article, the generation of ACEI-inhibitory peptides from meat and meat products and their identification by empirical and in silico approaches have been reviewed, as well as the latest studies on bioavailability from the point of view of their health relevance. A discussion about current limitations and challenges to be overcome in order to advance in the state-of-the-art of this field is also included.

2. Generation of Meat-Derived ACEI-Inhibitory Peptides

ACEI-inhibitory peptides are usually small peptides with sizes comprising between 2 and 20 amino acids. Their function depends on the protein source, hydrolysis conditions, degree of hydrolysis, molecular mass, and amino acid composition as well as the position of amino acids in the peptide sequences. In this respect, ACEI-inhibitory peptides have been described that have hydrophobic and branched-chain amino acids in their structure. According to the literature, the type of amino acids located in the three positions close to the C-terminal end of the ACEI-inhibitory peptide is important for activity. The presence of aromatic, positively-charged, and basic amino acids in these positions is important for competitive binding to the ACEI active site. In fact, milk-derived tripeptides containing prolines might have different cis/trans configurations of bonds which could influence their access/binding to the ACEI complex [10–14]. Milk proteins have been described as a very good source of antihypertensive peptides, released during gastrointestinal digestion or food processing. The tripeptides Ile-Pro-Pro and Val-Pro-Pro, that are released from casein during the fermentation of milk, have been described as antihypertensive in several animal models as well as in clinical studies [15].

ACEI-inhibitory peptides obtained from food sources are inactive within the intact parent protein but can exert their activity once they are released by hydrolysis. Different ways of generating ACEI-inhibitory peptides have been utilised, as shown in Figure 2.
2.1. Bioactive Peptides Generated during Gastrointestinal Digestion

Gastrointestinal digestion (GI) is the last step for the generation of bioactive peptides from foods. After food ingestion, gastrointestinal peptidases such as pepsin, trypsin, or chymotrypsin are the main proteases responsible for the generation of multiple peptides, including bioactive sequences. In the laboratory, gastrointestinal digestion can be simulated using specific commercial enzymes and controlled conditions of pH and temperature. Thus, a simulated gastrointestinal digestion of raw pork meat using pepsin and pancreatin indicated that the physiological digestion of pork proteins could generate peptides with biological activity [16]. These results were later confirmed in vitro with the ACEI-inhibitory peptides KAPVA (Lys-Ala-Pro-Val-Ala) and PTPVP (Pro-Thr-Pro-Val-Pro), showing half maximal inhibitory concentration (IC$_{50}$) values of 46.56 and 256.41 µM, respectively [17], using the ACEI-inhibitory method described by Sentandreu and Toldrá (2006) [18]. Later, it was confirmed that these peptides also produced in vivo a decrease in the systolic blood pressure (SBP) of spontaneously hypertensive rats (SHRs) of 33.72 ± 8.01 mmHg and 25.66 ± 6.84 mmHg, respectively, after single oral administration [19]. A recent study evaluated the digestion of beef proteins by studying the kinetics of peptide release in vivo by regularly sampling the gastric contents using a cannula. The obtained results were evaluated with bioinformatics tools in order to identify potentially bioactive peptides [20].

On the other hand, GI simulation has also been used in studies of bioavailability of certain peptide sequences in order to demonstrate whether they could exert a positive health effect in the body, as they should resist further GI digestion, cross the intestinal barrier, and reach the bloodstream and target organs.

Finally, necessary treatments of meat before consumption such as cooking could facilitate the later generation of bioactive peptides due to denatured proteins being more susceptible to be hydrolysed by the enzymes of the intestinal tract.

2.2. Hydrolysis Treatments with Commercial Enzymes

The most used methodology for the generation of bioactive peptides is the hydrolysis of proteins with commercial enzymes. Proteases from different sources such as of microbial, plant, or animal origin, have been used for the hydrolysis of food proteins. In meat and meat products, Flavourzyme
from Aspergillus oryzae, and Neutrase and Alcalase from Bacillus subtilis and Bacillus lincheniformis, respectively, have been the most used in the generation of bioactive peptides. In addition, proteases from plant origin such as bromelain and papain have been described as interesting enzymes for the hydrolysis of meat proteins by their contribution to meat tenderisation. These enzymes show a wider specificity in comparison with other enzymes such as trypsin or pepsin, cleaving peptide bonds from a wide variety of regions and frequently acting as either endopeptidases, or as exopeptidases hydrolysing amino acids from N- and C-terminal sites. In fact, the activity and hydrolytic specificity of many commercial peptidases is not clearly defined by manufacturers and thus, the degree of hydrolysis and final content of peptides is difficult to predict [8].

Several studies have reported the generation and identification of ACEI-inhibitory peptides resulting from hydrolysates of pork [17,21,22], chicken [23,24], and beef [25]. However, proteins obtained from by-products constitute good substrates that can be used to obtain bioactive peptides through this methodology [26,27], giving an extra added value to these products as well as reducing their environmental impact. In fact, this is the most commonly used procedure when the objective is to obtain high amounts of bioactive peptides for commercialisation, because its efficiency is optimised in a laboratory and later scaled up for pilot plant and industrial production.

2.3. Bioactive Peptide Generation during Ageing and the Processing of Meat

Bioactive peptides can be generated through the action of endogenous enzymes in ageing and curing processes as well as in combination with microbial peptidases such as in fermentation processes. Proteolysis by endogenous proteases is the most important phenomena occurring in the ageing of meat that influences its final characteristics with endogenous peptidases as main figures. Broadly speaking, endopeptidases such as calpains and cathepsins are first responsible for the hydrolysis of proteins into large fragments and oligopeptides, which affect the texture of meat during ageing and the initial steps of curing processes. Later, the activity of exopeptidases such as aminopeptidases and carboxypeptidases will generate small peptides and free amino acids, responsible for the characteristic flavour of dry-cured products. Some of the generated small peptides have also been described as bioactive peptides, exerting activities such as ACEI-inhibitory activity and antihypertensive, antioxidant, antilisteria, dipeptidyl peptidase IV (DPP-IV) inhibitory, and anti-inflammatory activity.

Dry-fermented sausages are elaborated using shorter processes with microorganisms such as lactic acid bacteria (LAB) (as a starter), yeasts, or moulds that are responsible for fermentation followed by ripening/drying. Lactic acid bacteria such as Lactobacillus sakei, Lactobacillus curvatus, Lactobacillus plantarum and Lactobacillus casei alone or in combination with staphylococci, Kocuria, yeast, or moulds, exert proteolysis through the action of endo- and exopeptidases. In general, these fermentation processes are involved in the liberation of small peptides and free amino acids that not only affect flavour development but also contribute to the generation of bioactive peptides [28].

The presence of ACEI-inhibitory peptides naturally generated during the processing of meat products such as dry-cured hams or dry-fermented sausages has also been described [29–33].

3. Identification of ACEI-Inhibitory Peptides

Traditionally, empirical approaches have been the method of choice for the identification of bioactive peptides from food matrices. However, it is very challenging when the objective is to generate specific peptide sequences that are able to exert certain activity. Then, the experimental design can be simplified by using bioinformatics for computer simulation in silico.

Empirical approaches used for the identification of bioactive peptides including ACEI-inhibitory peptides in complex sample matrices such as meat and meat products involve: (1) the release of the bioactive sequences from the parent protein; (2) preliminary in vitro assays to screen for bioactivity; (3) purification and separation through the use of high-resolution techniques, such as chromatography; (4) additional in vitro assays to determine the most active fractions; (5) identification of peptides by mass spectrometry (MS) techniques; (6) selection and synthesis of potential bioactive peptides; and (7) in vitro
and in vivo confirmation of the bioactivity [34]. A scheme of the traditional empirical procedure for the identification and confirmation of bioactive peptides from food matrices is shown in Figure 3.

In vitro ACEI-inhibitory activity is typically measured by monitoring the conversion of a specific substrate by ACEI in the presence and absence of inhibitors. Spectrophotometric and chromatographic methods have been commonly used to measure the hydrolysis of substrates such as Hippuryl-His-Leu (HHL) or the fluorogenic o-aminobenzoylglycyl-p-nitrophenylalanylproline. However, the inhibitory activities of these peptides on ACEI activity do not always correlate with antihypertensive effects. In this regard, SHRs are the animal model most frequently used to verify the in vivo efficacy of ACEI-inhibitory peptides. Some studies have evaluated the effects on SBP of SHRs after oral administration of meat hydrolysates or peptide extracts showing ACEI-inhibitory activity [19,35–37]. Table 1 shows the antihypertensive effects of meat-derived peptides after single oral administration to SHR [38–44]. As a last step, human clinical trials are the most accurate method to assess the efficacy and physiological functions of meat-derived antihypertensive peptides, although few studies have been done in this respect due to the complexity and expensive costs. Hodgson et al. (2006) suggested that a partial substitution of carbohydrate intake with protein-rich foods such as lean red meat may lower SBP in hypertensive persons [45], whereas a clinical study done by Saiga-Egusa et al. (2009) using chicken collagen hydrolysate observed a SBP reduction in mildly hypertensive subjects by inhibiting ACEI and plasma renin activity [46]. Additionally, it has been reported that the regular consumption of dry-cured ham would not increase blood pressure despite its high salt content, and even could exert other beneficial effects on cardiovascular health related to glucose and lipid metabolism, and inflammatory processes [47–49].

**Figure 3.** Scheme of the traditional empirical procedure for the identification and confirmation of bioactive peptides from food matrices. SEC: size-exclusion chromatography; CE: capillary electrophoresis; LC: liquid chromatography; IEF: isoelectric focusing; HPLC: high performance liquid chromatography; MS/MS: mass spectrometry in tandem.
Table 1. Angiotensin I-converting enzyme (ACEI)-inhibitory peptides identified in meat and meat products with antihypertensive effects in spontaneously hypertensive rats.

| Source            | Peptide Sequence | Parent Protein         | Hydrolysis Treatment | IC<sub>50</sub> (µM)  | Dose (mg/kg BW) | SBP (mmHg)  | Time (h) | Reference |
|-------------------|------------------|------------------------|----------------------|------------------------|----------------|------------|----------|-----------|
| Chicken muscle    | IKW              | —                      | Thermolysin          | 0.21                   | 60             | −0.17      | 4        | [23]      |
| Chicken muscle    | LKP              | Aldolase               | Thermolysin          | 0.32                   | 60             | −0.18      | 4        | [23]      |
| Chicken muscle    | FKGRYP           | Creatine kinase        | Thermolysin          | 0.55                   | 60             | 0          | —        | [23]      |
| Chicken muscle    | GA(Hyp)GL(Hyp)GP | Collagen               | Proteases            | 29.4                   | 4.5            | −0.18      | 6        | [39]      |
| Chicken bone      | YYRA             | Immunoglobulin heavy chain | Pepsin | 57.2                   | 10             | −0.20      | 6        | [40]      |
| Porcine muscle    | MNPPK            | Myosin                 | Thermolysin          | 945.5                  | 1              | −0.23      | 6        | [35]      |
| Porcine muscle    | ITTNP            | Myosin                 | Thermolysin          | 549                    | 1              | −0.21      | 6        | [35]      |
| Porcine muscle    | VKKVLGNP         | Myosin light chain     | Pepsin               | 29                     | 10             | −0.24      | 3        | [41]      |
| Porcine muscle    | KROKYDI          | Troponin               | Pepsin               | 26.2                   | 10             | −0.9       | 6        | [42]      |
| Porcine muscle    | KRVTGY           | Myosin heavy chain     | Pepsin               | 6.1                    | 10             | −0.23      | 6        | [43]      |
| Porcine muscle    | VKAGF            | Actin                  | Pepsin               | 20.3                   | 10             | −0.17      | 6        | [43]      |
| Porcine muscle    | RPR              | Nebulin                | Pepsin + pancreatin  | 382                    | 1              | −0.33      | 6        | [19]      |
| Porcine muscle    | KAPVA            | Titin                  | Pepsin + pancreatin  | 46.56                  | 1              | −0.33      | 6        | [19]      |
| Porcine muscle    | PTPVP            | Titin                  | Pepsin + pancreatin  | 256.41                 | 1              | −0.25      | 6        | [19]      |
| Porcine skin      | GF(Hyp)GP        | Collagen               | Aspergillus protease | 91                     | 10             | −0.20      | 8        | [44]      |
| Goat muscle       | FQPS             | —                      | Protamex® + Flavourzyme® | 27.0              | 2.39           | −0.10      | 8        | [45]      |
| Spanish dry-cured ham | AAATP           | Allantocase            | No treatment         | 100                    | 1              | −0.26      | 8        | [19]      |

<sup>a</sup> IC<sub>50</sub> value is the peptide concentration that inhibits 50% of ACE activity; <sup>b</sup> Oral administration of the peptide expressed as mg/kg body weight of rat; <sup>c</sup> Maximum decrease in systolic blood pressure (SBP) after administration of the peptide to spontaneously hypertensive rats; <sup>d</sup> Time after peptide administration to exert the maximum decrease in systolic blood pressure.
In addition to empirical approaches, the use of in silico analyses that combine bioinformatics tools and peptide databases has been increasingly used as a cost- and time-effective alternative. This predictive strategy enables to obtain biological and chemometric information on peptide sequences to be obtained through a series of steps: (1) selection of proteins of interest with known amino acid sequences by predicting their potential as precursors of novel bioactive peptides; (2) in silico protein digestion by selected proteolytic enzymes; (3) in silico identification and characterisation of peptides; (4) bioactivity prediction using a combination of sequence biochemical properties and databases of known bioactive peptides; (5) peptide synthesis; and (6) in vitro or in vivo confirmation of the bioactivity [50]. Figure 4 shows the main steps of in silico approaches and suggests open access databases and bioinformatics tools for the selection of the protein, hydrolysis simulation and bioactivity prediction. In this regard, BIOPEP is a widely used database for the study and identification of food-derived bioactive peptides as well as for in silico digestion and prediction of their bioactivities. In addition, computational models such as quantitative structure-activity relationships (QSAR), quantitative structure-property relationships (QSPR), and molecular docking simulations allow the discovery and characterisation of structural and physical-chemical properties such as hydrophilicity-hydrophobicity, molecular size, and electronic and steric characteristics, and results in very useful information to evaluate the potential affinity between the biopeptide sequence of interest and the target [50,51].

**Figure 4.** Main steps of in silico approaches and open access databases for the selection of the protein, hydrolysis simulation and bioactivity prediction.

In silico approaches have shown that some bovine, porcine and chicken proteins such as collagen, connectin, and myosin are good sources of ACEI-inhibitory peptides, which could be released from the parent protein through the action of determined enzymes [52–54]. Moreover, computer simulations are fundamental for understanding molecular mechanisms and ACEI-peptide interactions such as the fact that the C-terminal tripeptide sequence, hydrophobicity, and positive charge of the amino acid residues in this region of the peptide have a major influence on ACEI inhibition [55].

At the end, empirical and in silico approaches converge in the need for the confirmation of both the identity of the generated/predicted peptide sequences and their activity in the complexity of the matrix (see Figure 4).
4. Bioavailability of ACEI-Inhibitory Peptides

Bioavailability studies are necessary to assess whether the bioactive peptide can reach its target site in active form and sufficient quantity to exert health effects in the organism. The action of gastrointestinal enzymes, intestinal absorption, cellular uptake, and action of blood plasma peptidases can modify the structure of ACEI-inhibitory peptides or hydrolyse them leading to a loss, maintenance, or gain of bioactivity [11,56].

The use of digestion models and LC-MS techniques combined with in silico and in vitro/in vivo approaches have enabled evaluation of the stability of ACEI-inhibitory peptides from beef, chicken and pork meat in gastrointestinal digestion as well as the identification and quantification of the resulting products [17,20,30,57–59]. On the other hand, cell models such as heterogeneous human epithelial colorectal adenocarcinoma cells (Caco-2 cell) monolayers have been useful to study the transepithelial transport of ACEI-inhibitory peptides derived from meat proteins, being able to determine structural changes and amount of peptides transported or the involved transport pathway [60–62]. The ability of peptides to resist enzymatic degradation and be transported across intestinal membranes into blood circulation depends on their characteristics, length, and amino acid composition. In this regard, proline-rich peptides are more resistant to be attacked by gastrointestinal enzymes, and di- and tri-peptides could be absorbed intact by peptide transporter systems and hydrolysed later [63]. The low transport ability of oligopeptides compared to di- and tri-peptides is probably due to their length and involve paracellular route, while the hydrophobicity of peptides does not seem to influence absorption [64]. Additionally, the absorption of peptides could be affected by co-existing peptides and food components, which can share the transport pathway or participate in its regulation [64].

The bioavailability and bioaccessibility of bioactive peptides can also be affected by processing/storage conditions and food matrix-peptide interactions that can lead to peptide modifications with changes in its native structure and activity [65]. Several studies have evaluated the stability of ACEI-inhibitory peptides after household cooking preparations of pork and beef meat [66], different temperatures and pH used when processing meat products [30,67], and the effect of ageing under industrial conditions (vacuum-packed and chilled-storage) and cooking of beef meat [68].

5. Challenges and Limitations

Currently, with the basis of knowledge for the identification of bioactive peptides already clear, it is necessary to continue the research in bioactive peptides to achieve a better understanding of their effect, interactions, and bioavailability. In this sense, several authors have established the need for serious consideration of food matrix interactions, especially when the objective is to use the bioactive peptides as a functional ingredient [65]. Increasingly, once the peptide has been identified in a food matrix, it is synthesised and characterised as an individual molecule. However, the expected in vitro and/or in vivo activity may differ when the peptide interacts with the complex mixture of compounds that are taking part of any food.

On the other hand, increased effort on the development of quantitative methodologies for a better understanding of hydrolysis, bioactivity, and/or bioavailability is necessary. Data such as the quantity of specific naturally generated peptides in the original food and the dose of a bioactive peptide needed to exert an effect in vivo, as well as the final sequences and amount present in bloodstream and target organ after GI digestion are key data for advancing in the field of bioactive peptides and their health influence. In fact, determining the quantity of ACEI-inhibitory peptides in the meat sample that are able to reach the target site in the human system is of fundamental importance in bioavailability studies to better understand the effects and mechanisms of action of these peptides. The main limitation for quantitation is the nature of sample: small peptides often comprise fewer than four amino acids at low abundance, and there is high complexity of the matrix [69]. Current advances in mass spectrometry instrumentation, bioinformatics tools, and updated protein databases are contributing to progress in quantitative peptidomics [68].
6. Conclusions

Meat and meat products have been described as a very good source of ACEI-inhibitory peptides. With proteins being a major constituent of meats, the generation of bioactive peptides from meat proteins has been described as occurring either through the action of endogenous muscular enzymes during processing, during GI digestion, or by using commercial enzymes in the laboratory or in industrial processes under controlled conditions. The identification of ACEI-inhibitory peptides has been traditionally done using empirical approaches, although currently there is an increasing interest in in silico approaches based on bioinformatics as they are less time-consuming and cheaper methodologies. However, despite the identification of bioactive peptides being clear, there is an increasing need to study food matrix interactions, especially when the objective is to use the bioactive peptides as a functional ingredient. The quantitation of these peptides for a better understanding of their health influence and bioavailability is necessary to advance in this field.

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