The role of *Tal2* and *Tal1* in the differentiation of midbrain GABAergic neuron precursors

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Summary
Midbrain- and hindbrain-derived GABAergic interneurons are critical for regulation of sleep, respiratory, sensory-motor and motivational processes, and they are implicated in human neurological disorders. However, the precise mechanisms that underlie generation of GABAergic neuron diversity in the midbrain–hindbrain region are poorly understood. Here, we show unique and overlapping requirements for the related bHLH proteins Tal1 and Tal2 in GABAergic neurogenesis in the midbrain. We show that Tal2 and Tal1 are specifically and sequentially activated during midbrain GABAergic neurogenesis. Similar to Gata2, a post-mitotic selector of the midbrain GABAergic neuron identity, Tal2 expression is activated very early during GABAergic neuron differentiation. Although the expression of Tal2 and Gata2 genes are independent of each other, Tal2 is important for normal midbrain GABAergic neurogenesis, possibly as a partner of Gata2. In the absence of Tal2, the majority of midbrain GABAergic neurons switch to a glutamatergic-like phenotype. In contrast, Tal1 expression is activated in a Gata2 and Tal2 dependent fashion in the more mature midbrain GABAergic neuron precursors, but Tal1 alone is not required for GABAergic neuron differentiation from the midbrain neuroepithelium. However, inactivation of both Tal2 and Tal1 in the developing midbrain suggests that the two factors co-operate to guide GABAergic neuron differentiation in a specific ventro-lateral midbrain domain. The observed similarities and differences between Tal1/Tal2 and Gata2 mutants suggest both co-operative and unique roles for these factors in determination of midbrain GABAergic neuron identities.

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Key words: Neurogenesis, GABAergic neuron, Ventral tegmental area (VTA), Substantia nigra pars reticulata (SNpr), Midbrain, Dopaminergic neuron, Hindbrain, Rhombomere 1, Transcription factor, Gata, Tal, Sc1, Brain development, Mouse

Introduction
The mammalian midbrain and hindbrain contain neuronal populations that control motivation, motor and sensory function as well as vital autonomic activity and sleep.

Gamma-aminobutyric acid (GABA) is the primary inhibitory neurotransmitter in the mature brain and is used by hundreds of different types of neurons throughout the central nervous system (CNS). In the midbrain, distinct GABAergic precursors are generated in ventro-lateral and dorsal progenitor domains m1–m5 (Nakatani et al., 2007; Kala et al., 2009). These precursors are thought to contribute to diversified GABAergic neurons in the midbrain including superior and inferior colliculi, periaqueductal gray area and nuclei in the midbrain reticular formation (mRF), where they are involved in functions such as processing of sensory information (Tsunekawa et al., 2005; Kala et al., 2009; Peltopuro et al., 2010; Lahti et al., 2013).

Other GABAergic neurons associate with dopaminergic (DA) nuclei, ventral tegmental area (VTA) and substantia nigra (SN), in the ventral midbrain. These ventral midbrain GABAergic neurons (vMB GABAn) are critical for the activity of the DA pathways and regulation of voluntary movements. Furthermore, the vMB GABAn embedded in the VTA and rostromedial tegmental nucleus (RmTg; also called the “tail of VTA”) have recently been found to be central for regulation of motivational states and reward (Jhou et al., 2009; Kaufling et al., 2010; Hong et al., 2011; Cohen et al., 2012; Lahti et al., 2013). Thus, the vMB GABAn are critical for understanding of aetiology and potentially also treatment of neurological and psychiatric disease, such as Parkinson’s disease, mood disorders, addiction and schizophrenia. In contrast to other midbrain GABAergic neurons, which are derived from the midbrain neuroepithelium, the vMB GABAn population originates from the rhombomere 1 (r1) neuroepithelium and migrates rostrally to the midbrain as post-mitotic young neuronal precursors (Achim et al., 2012). In addition, the GABAergic neurons in the diencephalic part of SN
have a distinct origin, likely in the more anterior brain regions. Thus, the developmental history of the midbrain GABAergic neurons is complex and involves contributions from adjacent brain regions, especially the r1.

Underlying the segmental organization of diversified GABAergic neuron development are organizational signals in the dorsal–ventral and anterior–posterior axes that instruct expression of transcription factors in distinct and overlapping patterns (Puelles, 2007; Nakamura et al., 2008). In the midbrain and posterior diencephalon, a zinc-finger transcription factor (TF) Gata2 is expressed upon cell cycle exit of all GABAergic precursors and controls their differentiation (Kala et al., 2009; Willett and Greene, 2011; Virolainen et al., 2012). Here Gata2 appears to function as a terminal selector gene of the GABAergic neuron identity. Without Gata2, all the midbrain derived GABAergic precursors switch to a glutamatergic phenotype. In contrast to the midbrain, Gata2 is not expressed in the anterior forebrain GABAergic precursors (Parras et al., 2002; Petryniai et al., 2007), and while it is expressed in the r1 it is functionally dispensable for GABAergic neurogenesis, possibly because of co-expression of Gata3 (Kala et al., 2009; Achim et al., 2012). In addition to GABAergic neurons, Gata2 and Gata3 are also expressed in the r1 serotonergic neurons and Gata2 is required for their development (Craven et al., 2004; Kala et al., 2009).

Gata TFs can exert different and even opposing effects on the same targets and their functions are highly context-dependent (Wozniak et al., 2008). These differences can be explained by participation of other regulatory factors. Indeed, Gata factors typically function as a part of a multi-protein transcription regulatory complex that may include Friend of Gata (Fog) family proteins Fog1 and Fog2, LIM-only TF Lmo2, LIM-family cofactor Nli (Ldb1), BHLH TF Tal1 (Sc1) and others (reviewed by Cantor and Orkin, 2002; Grosveld et al., 2005). In cultured blood cell progenitors, the presence or absence of Tal1 in the Gata2-complex can define the nature of target gene regulation (Tripic et al., 2009). The Tal1–Gata2 interaction has also been shown to regulate neuronal fate selection in the ventral spinal cord (Zhou et al., 2000; Karunaratne et al., 2002; Muroyama et al., 2005). Here, Tal1 appears to specifically instruct the development of V2b GABAergic neurons from an initially bipotent postmitotic precursor pool (Del Barrio et al., 2007; Peng et al., 2007). The precursor fate segregation also involves a cofactor Lmo4, which mediates the formation of Gata2–Tal1 transcriptional complex that turns on Gata2/3 expression and other targets in V2b cells (Joshi et al., 2009).

Although their cell-type specificity has not been demonstrated, expression of Tal1 and a related gene Tal2 have been detected in the mouse midbrain and r1 raising the possibility of unique or stage specific roles have not been described.

Here, we established the functions of Tal1 and Tal2 in GABAergic neurogenesis in the developing midbrain using single and compound mutant mice. We show that Tal1 and Tal2, together with Gata2 and Gata3, are activated in spatially and temporally specific fashion in the differentiating GABAergic precursors, and that unique functions of these factors are necessary components of GABAergic sub-type specification in distinct regions of the developing brain.

Materials and Methods
Mice
The following mouse lines and their genotyping have been described previously: En1Cre (Kimmel et al., 2000) provided by Wolfgang Wurst, Helmholtz Centre Munich, Germany; Wnt1Cre (Danielian et al., 1998) Jackson Laboratory, Bar Harbor, USA; Gata2lox (Haugas et al., 2010), Tal1lox (Bradley et al., 2006) provided by David Curtis Monash University, Melbourne, Australia and Stuart Orkin, Dana-Farber Cancer Institute, Boston, USA; Tal2flox (Bucher et al., 2000) provided by Terry Rabbits, MRC Weatherall Institute of Molecular Medicine, Oxford, UK, and Gad67GFP (Tamamaki et al., 2003) provided by and Yuchoi Yanagawa, Gunma University Graduate School of Medicine, Maebashi, Japan. For staging, the day of vaginal plug was counted as embryonic day 0.5 (E0.5). For in situ mRNA hybridization (ISH) and immunohistochemistry (IHC) embryos and brains were fixed in 4% paraformaldehyde in 1× PBS at RT for 2–7 days. Samples were dehydrated, embedded in paraaffin (Merek), sectioned at 5 μm, and detected on adjacent slides. All analyses were confirmed using 2–5 biological replicates (e.g. mutant embryos from different litters of same stage). All experiments were approved by the Laboratory Animal Center of the University of Helsinki, Finland and the Institutional Animal Care and Use Committee of the University of California, San Francisco.

In situ mRNA hybridization and immunohistochemistry
mRNA ISH analyses on paraffin sections were performed as described previously (Wilkinson and Green, 1990) using 35S- or digoxigenin-labeled cRNA probes. IHC was performed as described previously (Kala et al., 2008). For combined ISH and IHC, TSA Fluorescence Palette System (PerkinElmer) was used to visualize ISH signal. Additional primary antibodies were added after the ISH signal detection. For defining of the domain boundaries in the midbrain (m3–m5), adjacent sections were analysed for Helt, Nkx2.2 and Nkx6.1 expression. Detailed ISH and IHC protocols are available upon request.

Mouse cDNA probes used for ISH analysis were: Gad1 (Gad65), Slc17a6 (Vglut2) (Guimera et al., 2006), Gata2, Gata3 (Lillevåli et al., 2004), Six3 (IMAGE 761326), Tal1 (IMAGE 6826611) Tal1fox (detects specifically the Tal1 transcript from the wild-type or unrecombined locus; contains the sequences between the LoxP sites in the Tal1fox allele (nt 699–1104 of NM_011527)), Tal2 (IMAGE 4005579). Following antibodies were used in IHC stainings: rabbit anti-Gata2 (Santa Cruz sc-9008, 1:250), mouse anti-Gata3 (Santa Cruz sc-268, 1:200), goat anti-GFP (Abcam ab6673, 1:500), rabbit anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heisile (Helt, 1:500), goat anti-GFP (Abcam ab290, 1:500), mouse anti-Nkx6.1 (1:1000, DSHB F55A10), mouse anti-Sox2 (Millipore AB5603, 1:400), mouse anti-TM (Millipore MAB318, 1:800), rabbit anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), mouse anti-Ascl1 (BD Biosciences 556604, 1:200), mouse anti-Nkx2-2 (DHSB 74A5, 1:250), mouse anti-Nkx6.1 (1:1000, DSHB F55A10), mouse anti-Sox2 (Millipore AB5603, 1:400), mouse anti-TM (Millipore MAB318, 1:800), rabbit anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan), mouse anti-Hu/C (Molecular Probes A21271, 1:1000), goat anti-GFP (Abcam ab290, 1:500), guinea pig anti-Heslike (Helt, 1:500), gift from Ryoichiro Kageyama, Institute for Virus Research, Kyoto University, Japan). For defining of the domain boundaries in the midbrain (m3–m5), adjacent sections were analysed for Helt, Nkx2.2 and Nkx6.1 expression. Detailed ISH and IHC protocols are available upon request.

Microscopy
ISH and IHC staining on paraffin sections were visualized with an Olympus AX70 microscope with Olympus DP70 camera or a Zeiss AxioImager M2 microscope with Axioscan HRc camera. Images were processed and assembled with Adobe Photoshop software. Confocal images were taken as snapshots with Leica SP5 confocal microscope.

Results
Tal1 and Tal2 are co-expressed with Gata2 in developing GABAergic neurons of the midbrain and r1
We first analysed the expression of Tal1 genes in the wild-type midbrain and r1 regions by ISH during GABAergic neurogenesis at E10.5–E12.5. In the midbrain, we detected Tal2 expression in the ventricular zone (VZ) and intermediate zone (IZ; Fig. 1C,H, Fig. 2A, Fig. 3A,C,F), while Tal1 was confined to the IZ and...
mantle zone (MZ; Fig. 1B,G, Fig. 2B, Fig. 3B,D). In addition, both Tal1 and Tal2 were expressed in the GFP+ GABAergic neurons in the Gad67GFP/+ mouse embryos (Fig. 2A–D). In contrast to the midbrain, Tal2 expression was very low and restricted to a small IZ area in the anterior r1 (Fig. 2L,M) while Tal1 expression in r1 was robust and extended from the VZ to MZ (Fig. 2J,K, Fig. 3E).

In both midbrain and r1, expression domains of Tal1/2 and Gata2/3 appeared to overlap extensively (Fig. 1, Fig. 2J–S), with the exception of Gata2/3+ serotonergic neuron precursors in the r1 (Fig. 2K,M,O,Q,S,T). To further study the co-expression of Tal1/2 and Gata2/3, we combined ISH for Tal1/2 and IHC for Gata2/3. This showed that in the midbrain, Gata2 is nearly exclusively co-expressed with Tal2 in the VZ and IZ (Fig. 2E).

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**Tal1** is activated later during differentiation, and thus Gata2 and Tal1 are co-expressed in the IZ (Fig. 2F). **Tal1** positive cells in the MZ most abundantly expressed Gata3 (Fig. 2H). In contrast, we detected only little co-expression of Tal2 and Gata3 in the MZ (Fig. 2G). Both **Tal1** and Tal2 were detected in postmitotic (HuC/D+) neurons (Fig. 3C,D). In the r1, the weak expression of Tal2 precluded fluorescent ISH-based co-localization studies. However, radioactive ISH demonstrated that Tal2 expression was mostly confined to the IZ in the r1 (Fig. 2M). In contrast to the midbrain, **Tal1** expression was detected already in the VZ and early during post-mitotic differentiation of the r1 GABAergic neurons (Fig. 2K, Fig. 3E). In summary, **Tal1** and Tal2 specifically mark the midbrain and r1 GABAergic precursors and co-localize with Gata2 and Gata3. However, the dynamics of **Tal1** and Tal2 expression differ between the midbrain and r1. While Tal2 expression precedes **Tal1** in the midbrain, the opposite is observed in the r1 where **Tal1** is expressed at an earlier stage and co-expressed with Gata2 already in the VZ. Gata2 is required for the expression of **Tal1** but not Tal2 in the midbrain

To study if activation of **Tal1** and/or Tal2 requires Gata2 function, we analysed En1Cre; Gata2flox/flox (Gata2cko) mutants in which both midbrain and r1 are devoid of Gata2 expression (Kala et al., 2009). In the Gata2cko midbrain, **Tal1** was downregulated at E12.5 (Fig. 4C,D) together with GABAergic neuron markers Gad1, Gata3 and Six3 (Fig. 4A,B and data not shown). In contrast, we detected persistent Tal2 mRNA expression (Fig. 4E,F, Fig. 3G). In the r1, expression of Gad1 and Tal1/2 was not affected by the loss of Gata2 function (Fig. 4B,D,F and data not shown), indicating that Gata2 is required for **Tal1** expression in the midbrain but not in the r1, while Tal2 expression is Gata2-independent, at least in the midbrain.

We also analysed Gad1, **Tal1** and Tal2 expression in wild-type and Gata2cko midbrains at E15.5 (Fig. 4G–L). Consistent with expression of **Tal1** in the MZ at E12.5, **Tal1** was abundantly expressed in the wild-type midbrain regions containing GABAergic neurons. In contrast, the expression of Tal2 was restricted to the VZ/IZ of the most dorsal midbrain, where GABAergic neurogenesis continues later than in the ventrolateral regions (Achim et al., 2012). In the Gata2cko midbrain, **Tal1** expression was abolished, except for the r1-derived GABAergic neurons in the SNpr (Fig. 4LJ). As in the ventrolateral midbrain at E12.5, Tal2 was still expressed in the dorsal midbrain of Gata2cko mutants at E15.5, although its pattern of expression was more narrow suggesting more rapid downregulation in the IZ (Fig. 4K,K′,L,L′).

**Tal** factors are required to activate genes characteristic for differentiating GABAergic neurons in the midbrain

To elucidate the importance of different Tal factors in regulation of GABAergic neuron differentiation in the midbrain and r1, we studied the expression of several genes characteristic to midbrain and r1 GABAergic neurons (Gata2, Gata3, Tal1, Six3, Gad1) in embryos mutant for Tal2 (Tal2null/null = Tal2cko), **Tal1** (En1cre+; Tal2flox/flox = Tal1cko), or both **Tal1** and Tal2 (Wnt1cre+; Tal2flox/flox; Tal2null/null = Tal2cko) in comparison with the Gata2cko mutants. Wnt1cre was used for inactivation in Tal1/2cko double mutants due to availability of the mouse strains. In the Tal1cko mutants, En1cre-mediated inactivation of Tal1flox allele occurs throughout the midbrain and r1. In the Tal2cko, the Tal2flox allele is inactivated by Wnt1cre, which is active mostly in the midbrain. Both Wnt1cre- and En1cre-mediated recombination is complete by E8.5, well before the onset of neuronal differentiation (Trokovic et al., 2003). We detected no **Tal1** expression in the midbrain and r1 of the Tal1cko or Tal1/2cko embryos at E11.5–E12.5 (Fig. 1L and data not shown). At E11.5, inactivation of **Tal1** alone (Tal1cko) did not affect GABAergic neurogenesis or the expression of Gad2, Gata3 or Tal2 in the midbrain (Fig. 1K–O).

The VZ progenitors in the midbrain GABAergic regions appeared unaffected also in the Tal2cko (supplementary material Fig. S1) and Gata2 expression was maintained in the early postmitotic precursors at E11.5 (Fig. 5H). In contrast, other post-mitotic GABAergic precursor specific genes were largely downregulated, in Tal2cko embryos (Fig. S1–L). However, in contrast to Gata2cko, some Gad1+ and Gata3+ cells were still detected in the Tal2cko embryos, especially in the m5 domain (Fig. S1L, arrowheads).

Although **Tal1** expression was downregulated in the Tal2cko brains at E11.5 (Fig. S1), and **Tal1** inactivation alone did not affect GABAergic neurogenesis in the midbrain (Fig. 1K–O) (Achim et al., 2012), it is possible that its expression at an earlier stage or at an undetectable level accounts for the survival of the few GABAergic cells observed in the Tal2cko mutants. To address this possible redundancy, we analyzed the Tal1/2cko double mutant midbrains. In these mutants, Gad1 expression was lost completely in E11.5 midbrain (Fig. 5B), along with the GABAergic markers Six3 and Sox14 (Fig. 5D and data not shown). However, we still detected Gata3 expressing cells in the...
m5 domain of the Tal1/2dko midbrains (Fig. 5O), suggesting that the loss of GABAergic identity was still incomplete at least in this domain. Importantly, Gata2 expression was unaffected in the Tal1/2dko (Fig. 3N), and the early GABAergic markers Helt (Fig. 5M) and Ascl1 (supplementary material Fig. S1) were detected in the VZ, confirming that the absence of Tal factors results in alterations only at the postmitotic differentiation stage.

In summary, our results suggest that Tal2 and Gata2 do not require each other for their expression but are independently activated in the midbrain and both control genes required for complete GABAergic differentiation and for the acquisition of correct neuronal identity. In addition, whereas Gata2 is required for GABAergic neurogenesis in the entire midbrain, requirement for the Tal factors for GABAergic development varies between the midbrain subdomains.

Ectopic activation of glutamatergic neuron markers in Tal mutant midbrain

Concomitant to loss of GABAergic identity, we observed ectopic upregulation of glutamatergic neuron markers Pax6 and Slc17a6 (Fig. 6E,F). These results indicate a GABAergic-to-glutamatergic fate transformation specifically in the midbrain of Tal2ko mutants, similar to the Gata2cko phenotype in the midbrain (Kala et al., 2009). However, unlike in Gata2cko mutants where only the m4v domain transforms to Pax6 expressing glutamatergic neurons whereas the m3 domain starts expressing m1–m2 glutamatergic marker Pou4f1 (Kala et al., 2009), in Tal2ko mutants ectopic Pax6 positive cells can be detected extensively in both m4d and m3 domains (Fig. 6E, arrowheads). Similar to the Tal2ko mutants, Pax6 and Slc17a6 expression was upregulated in Tal1/2dko midbrain compared to wild type (Fig. 6H,I). In the wild type, Nkx6.1 is expressed in the VZ of domains m6, m5 and m3 and in the glutamatergic postmitotic precursors in m6. Nkx6.1 did not display ectopic expression in postmitotic Pax6 positive precursors derived from m4 or m3 in either Tal2ko or Tal1/2dko midbrains (Fig. 6A,D,G, arrows). However, especially in the Tal1/2dko mutants, Nkx6.1 expression appeared somewhat expanded in the postmitotic precursors in the m5 domain, similar to what has been observed in Gata2cko mutants (Kala et al., 2009).

Thus, the Tal factors, and especially Tal2, appear to operate as post-mitotic selectors between GABAergic and glutamatergic neuronal phenotypes in the midbrain. In this respect, they share the function of their putative cofactor Gata2. However, Tal and Gata factors also differ in the ways they affect development of neuronal precursors derived from distinct midbrain regions.

Loss of Tal2 and Gata2 results in changes in neuronal identities in the nuclei of perinatal midbrain

Next, we compared the expression of GABAergic and glutamatergic neuron markers in the midbrains of Gata2cko and Tal2ko mutants at perinatal stages. Normally, both Gad1+ GABAergic neurons and Slc17a6+ glutamatergic neurons are found in a variety of nuclei across midbrain (Fig. 7A,B,D). At E15.5–P0, Gad1+ neurons co-expressed Gata3 and Tal1 in...
specific nuclei and midbrain regions including the SN (Fig. 4G,I, Fig. 7B,C and data not shown). In both Gata2^cko and Tal2^cko animals, GABAergic markers were completely lost in the dorsal midbrain including the superior colliculi (Fig. 7A,F,K and data not shown). Similarly, we found that Tal2^cko greatly resembles Gata2^cko in that the expression of GABAergic markers Gad1 (Fig. 7B,G,L), Gata3 and Tal1 (Fig. 7C,H,M and data not shown) was lost (Gata2^cko) or reduced (Tal2^cko) in the mRF. Furthermore, in the SN/VTA marked by TH staining (Fig. 7E,J,O), Gad1, Gata3 and Tal1 expressing cells were retained both in the Gata2^cko and Tal2^cko (Fig. 7B,C,G,H,L,M, arrowheads, and data not shown). Consistent with cell fate re-specification, upregulation of Slc17a6 was observed in the mRF region but not in SN/VTA of Gata2^cko and Tal2^cko midbrains (Fig. 7D,I,N). Similar changes were seen in Tal1^2^cko (supplementary material Figs S2, S3; note for example the glutamatergic red nucleus (RN) which is surrounded by mostly GABAergic neurons in the wild type, but glutamatergic neurons in the mutants in supplementary material Fig. S3B,C,E,F). Thus, the perinatal Tal2 and Gata2 mutants display neuronal fate changes, which are consistent with their embryonic phenotypes. Furthermore, in contrast to Tal1 (Achim et al., 2012), Tal2 or Gata2 are not required by the majority of vMB GABAn.

Discussion
Knowledge of developmental regulatory mechanisms can provide key insights into generation of neuronal diversity and functional specialization in the CNS. Such mechanisms might also be involved in developmental anomalies leading to life-threatening human neurological disorders. For example, mutation of transcription factor PTF1A in humans results in abnormal development of GABAergic neurons in the dorsal r1 and cerebellar agenesis (Sellick et al., 2004; Hoshino et al., 2005). Development of GABAergic neurons is regulated by different molecular mechanisms in different parts of the developing CNS. Our previous studies identified Gata2 as a post-mitotic terminal selector gene during GABAergic neurogenesis in the embryonic midbrain and revealed developmental diversity of the distinct GABAergic neuron groups in the midbrain–r1 region (Kala et al., 2009; Achim et al., 2012). Here, we studied how the putative cofactors of Gata2, Tal1 and Tal2, are involved in the GABAergic neuron differentiation in the midbrain. Our results suggest that Tal2 is important for selection of GABAergic over glutamatergic neuron identity in the midbrain. In contrast, Tal1 marks more mature midbrain GABAergic precursors but is not required for their differentiation. Unlike Gata2, the requirement for Tal factors varies between the midbrain subregions. The ventro-lateral m5 domain appears the most tolerant to Tal2 inactivation. Here, Tal1 and Tal2 may function redundantly to regulate GABAergic neurogenesis (Fig. 8).

Similarities and differences of Gata and Tal factors as the terminal selectors of GABAergic identity
Studies on neuronal differentiation in various models, including C. elegans, have led to the concept of terminal selector genes (Hobert, 2011). These genes are thought to be active in post-mitotic neuronal precursors and regulate expression of gene batteries providing a neuron its identity. An important feature of a terminal selector gene is that its inactivation does not lead to the loss of the neuron but rather the loss, or transformation, of its identity. Our previous studies have suggested that Gata2 fulfills most of the criteria of a terminal selector gene both in the developing midbrain and diencephalon (Kala et al., 2009; Virolainen et al., 2012). The results presented here suggest that the selection of the GABAergic identity also requires Tal2 in the developing midbrain. As Gata and Tal factors are involved in the
same transcriptional complexes in other tissues, they may also form an analogous TF complex in the midbrain. Our results suggest that expression of both Gata2 and Tal2 is initiated very early after cell cycle exit and further demonstrate independent initiation of Tal2 and Gata2 expression, likely triggered by region- and cell-cycle stage specific cues. However, neither the endogenous Tal2 nor Gata2 alone are sufficient to induce all GABAergic neuron specific genes in the embryonic midbrain. Instead, we propose that the coincident expression of these two TFs leads to formation of a terminal selector complex of the GABAergic identity (Fig. 8).

Consistent with the hypothesis above, the midbrain phenotypes of Gata2 and Tal2 mutants are similar. In both cases progenitor proliferation, cell cycle exit and production of neuronal precursors appeared undisturbed, but the postmitotic precursors lost their GABAergic identity and acquired glutamatergic characteristics instead. However, there also are some notable differences. In contrast to Gata2 mutants, the GABA marker genes are not completely lost in the Tal2 mutants. In particular, the m5 domain still retains some of its GABAergic characteristics without Tal2. Studies of the Tal1/2dko mutants suggest that this is partly explained by redundancy between Tal2 and Tal1. However, some GABAergic markers, such as Gata3, were detected in m5 even in the absence of both Tal1 and Tal2. In the m3 domain, pronounced GABAergic-to-glutamatergic phenotypic transformation occurs in both Tal2 and Gata2 mutants. However, the ectopic glutamatergic neurons appear to be of different subtype in the two mutants. Without Gata2, the mutants, the GABAergic neuron specific genes in the embryonic midbrain. In particular, the m5 domain still retains some of its GABAergic characteristics without Tal2. Studies of the Tal1/2dko mutants suggest that this is partly explained by redundancy between Tal2 and Tal1. However, some GABAergic markers, such as Gata3, were detected in m5 even in the absence of both Tal1 and Tal2. In the m3 domain, pronounced GABAergic-to-glutamatergic phenotypic transformation occurs in both Tal2 and Gata2 mutants. However, the ectopic glutamatergic neurons appear to be of different subtype in the two mutants. Without Gata2, the precursors produced in m3 upregulate Pou4f1, resembling the glutamatergic neurons born in m1 and m2 (Kala et al., 2009). In contrast, in the Tal2 mutants, the ectopic glutamatergic neurons in m3 are positive for Pax6, similar to the glutamatergic neurons produced in m4v. Thus, in addition to the putative Gata2–Tal2 complex, the two factors may also have functions independent of each other or their loss can be differentially compensated by other associating factors.

Distinct Gata/Tal complexes regulate GABAergic neurons in different spatio-temporal patterns?

Gata2/3 and Tal1/2 also mark a subset of differentiating GABAergic neurons in the r1. Ventrally this GABAergic subdomain is bordered by Nkx2.2 positive serotonergic region and dorsally it extends to the dorsal boundary of Nkx6.1 expression (Achim et al., 2012). However, requirement for the Gata and Tal factors in the r1 is different from the midbrain. In contrast to the midbrain, Tal1 is required for GABAergic differentiation in the rhombomere 1 (Achim et al., 2012). Similar to Tal2 in the midbrain, Tal1 may perform the selector function in a complex together with Gata factors. It is likely that this variation is used for generating GABAergic neuron subpopulations, which differ in their gene expression patterns and cellular phenotypes.

As discussed above, the Gata and Tal TFs may also operate independent of each other in distinct gene regulatory complexes. This is particularly evident in the r1, where Gata2 and Gata3 are strongly expressed in the developing serotonergic neurons without any detectable Tal1 or Tal2. Therefore, the TF complex regulating serotonergic neuron differentiation must differ from the one regulating the GABAergic phenotype. It will be of interest to analyse whether Gata2 and Gata3 also perform a terminal selector gene function in the serotonergic neuron lineage. If this is the case, Tal1, Tal2 and additional Gata cofactors may differentiate between serotonergic and GABAergic lineages.

Conclusions

Here we demonstrate distinct requirements for the putative Gata cofactors Tal2 and Tal1 in differentiating GABAergic neurons in the midbrain. We suggest that variants of Gata–Tal complex function as terminal selectors identifying different types of postmitotic GABAergic precursors. Although Gata2 and Tal2/Tal1 likely co-operate as GABAergic neuron determinants, differences in the midbrain neuronal differentiation in the absence of Gata2 or Tal1/2 function also suggest that these factors have unique targets. Knowledge on developmental regulation of the subgroups of GABAergic neurons in the midbrain-r1 region will allow later studies of their molecular composition, morphology and function.

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Competing Interests

The authors have no competing interests to declare.

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**Fig. S1.** Loss of Tal2 or Tal1/2 does not affect the gene expression in the ventricular zone. Immunostainings on E11.5 WT (A,B), Tal2ko (C,D) and Tal1/2dko (E,F) midbrains with anti-Sox and anti-HuC/D (A,C,E), and anti-Ascl1 (B,D,F). Dashed lines indicate the borders of midbrain m3–m5 domains, defined by the expression of Nkx2.2 in the MZ (ISH on an adjacent section, not shown). Scale bar: 100 μm.

**Fig. S2.** Expression of GABAergic and glutamatergic marker genes in wild-type, Tal2ko and Tal1/2dko mutant midbrain at E14.5. (A–O) Radioactive ISH on coronal parallel frozen sections of the midbrain with the probes indicated. Few residual cells still express Gad1 in mutants, whereas the expression of glutamatergic marker Slc17a6 is strongly upregulated.

**Fig. S3.** Expression of Gad1, Pou4f1, and Slc17a6 in wild-type and Tal1/2dko midbrains at E18.5. (A–F) ISH on parallel coronal frozen sections of the midbrain with the probes indicated. SC, superior colliculi, PAG, periaqueductal grey, mRF, midbrain reticular formation, RN, red nucleus.