Molecular phylogeography of *Ficus benghalensis* Linnaeus using nrDNA ITS 1, cpDNA trnL and cpDNA rps16 from the Indian subcontinent

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*Ficus benghalensis* Linnaeus (Moraceae) is the national tree of India and is well known for its pharmacological properties. The present study was aimed to determine the genetic diversity of *F. benghalensis* from the Indian subcontinent using sequence-based multi-locus phylogeography. A total of 20 geographical isolates were collected from different regions, covering major parts of its species range within the country. Sequence data from nuclear-encoded internal transcribed spacer region (ITS1), plastid-encoded trnL-F spacer region (trnL), and ribosomal intron region (rps16) were generated. The trnL-based maximum likelihood phylogram revealed the existence of two haplotypes, whereas ITS1 and rps16-based maximum likelihood analysis did not reveal much variation for this species distributed in the Indian subcontinent. These results depict long-distance random gene flow across the subcontinent, and support the post-glacial population contraction events. To validate the impact of palaeo-historic climatic events on current geographic and genetic distribution, species distribution modelling-coupled phylogeography is suggested.

Keywords: Banyan, genetic heterogeneity, haplotypes, maximum likelihood, phylogeography.

**FICUS** Linn. (Moraceae) is one of the largest genera amongst angiosperms with around 1000 tree species native mainly to the world tropics. Fig trees (*Ficus* spp.) are considered to be the foundation species with major influence on community structure and ecosystem function, as they are often abundant and fruit throughout the year in many tropical forests. Figs are exclusively pollinated by female agaonine wasps that lay their eggs in fig flowers, where wasp larvae thrive on developing seeds. This obligate mutualism is considered to be the basis for co-evolution of fig-wasp species over millions of years. The short-lived (1–2 days) fig wasps are vulnerable to climate change and global warming may lead to their extinction in the near future. Fig plants are the centre for complex web animals, including specialists and generalists, but are at major risk owing to the declining population of fig wasps, which will further affect many trophic levels.

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**ACKNOWLEDGEMENTS.** N.B. thanks The World Green Tea Association, Shizuoka, Japan for award of O-Cha (Tea) Pioneer Academic Research Grant Program. We thank Department of Biotechnology, Government of India for financial support.

Received 20 December 2018; accepted 20 February 2019

doi: 10.18520/cs/v117/i2/288-293

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**RESEARCH COMMUNICATIONS**

*Ficus benghalensis* is an important species of the genus and is known for many variations in the species habits. This fabled tree is well-documented in Ayurveda and holds a sacred place in Hinduism, Jainism and Buddhism. The characteristic copious aerial roots of this tree on reaching the ground grow into a new indistinguishable trunk, forming a miniaturized forest. Being the national tree of India, it is widely grown and revered across the subcontinent. Bioactive molecules useful in wound healing, teeth disorders, diarrhoea, allergies, skin disorders, diabetes, immune dysfunctions and cancer have been reported from banyan tree.

Despite its long-known pharmacological worth, the tree has been ignored largely in terms of molecular assessment. *F. benghalensis* is facing problems of genetic erosion and mislabelling. All the previous studies on molecular phylogeny of figs were done to examine the co-evolutionary patterns with fig-wasps, and were conducted elsewhere in the world. While lot of literature is available on the bioactive potentials of *F. benghalensis*, assessment on either its genetic heterogeneity or phylogeny has never been done for Indian isolates. This important tree species has never been subjected to sequence-based DNA barcoding, despite being the national tree of India. The present study evaluates phylogenetic relationships between various geographical isolates of *F. benghalensis* from India using Internal Transcribed Spacer-1 (ITS1), ribosomal protein s16 (rps16) and trnL markers.

Leaf samples of 20 geographical isolates of *F. benghalensis* were collected from different locations in the Indian subcontinent (Figure 1). Samples were transported in zip-lock polythene bags under cold conditions. Then they were cleaned thoroughly with tap water to remove the overlay dust and contaminant particles, and then cleaned with distilled water for further processing. Leaf morphology was studied, and pressed sample vouchers were prepared, and deposited in the Central National Herbarium, Botanical Survey of India, Kolkata (Index Herbarium code: CAL). Samples for molecular analysis were stored at −80°C.

Apical parts of leaf samples were excised and total genomic DNA was extracted using HiPura™ plant genomic extraction kit (HiMedia Laboratories Pvt Ltd, Mumbai) according to the manufacturer’s protocol. The DNA was quantified on a spectrophotometer at 260 nm and by electrophoresis on 0.8% agarose gel.

PCR amplifications were carried out using primers ITS1, rps16 and trnL (Table 1). The 20 µL reaction mix consisted of 10× PCR buffer, 10 µM of each primer, 3 mM MgCl₂, 10 mM dNTP and 1 unit of Taq DNA polymerase. The reactions were carried out in programmable thermal cycler as optimized for each primer set. Amplified products were electrophoresed on 1.5% agarose gel and positive reactions were selected. These products were purified using ExoSAP-IT® PCR clean-up kit according to the manufacturer’s protocol. Sequencing was performed using Applied Biosystems reagents and a DNA analyzer (ABI 3730 xl, Applied Biosystem, USA). The chimeras were removed and contig alignment was done using CodonCodeAligner (CodonCode Corporation, USA). The obtained DNA sequences were annotated with other pertinent information such as collection date and herbarium voucher accession, and submitted to GenBank with accession nos KT924351–KT924368 and KT884619–KT884657 (Table 2). The FASTA format sequences of all accessions generated in this study are provided under Supplementary Table 1.

Sequences were compared with non-redundant database available at the National Centre for Biotechnology Information (NCBI) using BLASTn. Sequences were first aligned by MUSCLE and CLUSTALW, and alignments were edited manually. Gaps were considered as missing data and indels were removed from the alignments. Step-by-step protocol for phylogenetic analysis, including alignment construction, model test, phylogeny reconstruction using maximum likelihood method and distance analysis was done according to Bast. Best-fitting nucleotide substitution models were tested using maximum

![Figure 1](image-url)  
**Figure 1.** Overlay of sampling locations on the map of India (map outline based on the Survey of India).

| Primer       | Sequence (5’–3’)         |
|--------------|--------------------------|
| ITS1 (ref. 13) | TCCGTTAGGTAAACCTGCGG     |
| tmnLF (ref. 12) | CGAAAATCGGTAGCCTACG      |
| tmnLR (ref. 11) | ATTTGAACTGGTGACACGGAG    |
| rps16 intronF (ref. 26) | GTGTTAGAAGACGACGTGGACCTT |
| Rps16 intronR (ref. 14) | TCGGATCGAACATCAATTGCAAC |
Table 2. Details of geographical isolates analysed in this study

| Voucher no.          | Location               | Collected by             | Collection date | Latitude (N) | Longitude (E) | ITS 1 sequence generated (GenBank Accession no.) | trnL sequence generated (GenBank Accession no.) | rps16 sequence generated (GenBank Accession no.) |
|----------------------|------------------------|--------------------------|-----------------|--------------|--------------|------------------------------------------------|------------------------------------------------|-------------------------------------------------|
| CUPVoucher-FB-2014-1 | Madhya Pradesh (Bina)  | Gajendra Singh Vishwakarma | 02-07-2013      | 24°10'03.9000"N | 078°11'00.4000"E | KT884628                                         | KT884654                                           | –                                               |
| CUPVoucher-FB-2014-2 | AP: Andhra Pradesh     | Dr Anil Kumar Manjesh     | 02-12-2013      | 16°27'36.0000"N | 077°33'36.0000"E | KT884621                                         | KT884650                                         | KT924352                                       |
| CUPVoucher-FB-2014-3 | BR: Bihar (Sheikhpora) | Ravi Shankar Dilip Singh  | 20-10-2013      | 25°08'41.7000"N | 085°52'09.8000"E | KT884627                                         | KT884642                                         | KT924354                                       |
| CUPVoucher-FB-2014-4 | KA: Karnataka (Mysore) | Ashish Kumar Pandey       | 13-11-2013      | 12°18'00.0000"N | 076°42'00.0000"E | –                                               | KT884649                                         | KT924361                                       |
| CUPVoucher-FB-2014-5 | JH: Jharkhand (Palamu) | Ashish Kumar Pandey       | 15-11-2013      | 23°21'00.0000"N | 085°19'48.0000"E | KT884625                                         | KT884644                                         | KT924359                                       |
| CUPVoucher-FB-2014-6 | RJ: Rajasthan (Jhunjhunu) | Prateek Sharma           | 09-10-2013      | 26°34'21.7200"N | 073°50'20.4000"E | KT884637                                         | KT884651                                         | KT924365                                       |
| CUPVoucher-FB-2014-7 | RJ (NRC): Rajasthan (NRC) | Satish Bhushan           | 09-10-2013      | 28°09'36.0000"N | 073°13'12.0000"E | KT884623                                         | KT884648                                         | KT924366                                       |
| CUPVoucher-FB-2014-8 | KL: Kerala (Palamkud)  | Dr Felix Bast             | 10-06-2013      | 12°06'00.0000"N | 075°12'00.0000"E | KT884619                                         | KT884652                                         | KT924362                                       |
| CUPVoucher-FB-2014-9 | DL: Delhi              | Dr Felix Bast             | 25-11-2013      | 28°37'58.0000"N | 077°13'10.5600"E | KT884629                                         | KT884638                                         | KT924355                                       |
| CUPVoucher-FB-2014-10| HR: Haryana (Paltri)   | Renu Yadav                | 9-06-2013       | 28°46'48.0000"N | 076°07'48.0000"E | KT884622                                         | KT884645                                         | KT924358                                       |
| CUPVoucher-FB-2014-11| AS: Assam (Bokajan)    | Anamika Das               | 19-09-2013      | 26°01'12.0000"N | 093°46'48.0000"E | KT884632                                         | KT884639                                         | KT924353                                       |
| CUPVoucher-FB-2014-12| PB: Punjab (Abohar)    | Poornima Rani             | 31-05-2013      | 30°08'00.2400"N | 074°12'30.3600"E | KT884633                                         | KT884643                                         | –                                               |
| CUPVoucher-FB-2014-13| PB: Punjab (Bhatinda)  | Sheetal Sharma            | 27-07-2013      | 30°19'12.0000"N | 076°27'00.0000"E | KT884620                                         | KT884641                                         | KT924364                                       |
| CUPVoucher-FB-2014-14| HP: Himachal Pradesh   | Anil Rana                 | 12-08-2013      | 31°40'48.0000"N | 076°31'12.0000"E | KT884634                                         | KT884640                                         | KT924357                                       |
| CUPVoucher-FB-2014-15| MH: Maharashtra        | Digvijay Singh Yadav      | 28-02-2014      | 18°34'04.8000"N | 073°53'52.2000"E | KT884636                                         | KT884657                                         | KT924363                                       |
| CUPVoucher-FB-2014-16| IX: Jammu (Bantalab)   | Shekhar Amin Poswal       | 03-02-2014      | 32°48'51.0000"N | 074°50'01.0000"E | KT884626                                         | KT884656                                         | KT924360                                       |
| CUPVoucher-FB-2014-17| TN: Tamil Nadu (Muruganagar) | Digvijay Singh Yadav | 18-02-2014      | 12°58'18.0000"N | 080°12'51.0000"E | KT884631                                         | KT884655                                         | KT924368                                       |
| CUPVoucher-FB-2014-18| GJ: Gujrat (Chandkheda) | Digvijay Singh Yadav      | 20-03-2014      | 23°25'60.0000"N | 072°40'00.0000"E | KT884635                                         | KT884646                                         | KT924356                                       |
| CUPVoucher-FB-2014-19| AP: Andhra Pradesh     | Dr Felix Bast             | 07-03-2014      | 17°21'57.6000"N | 078°28'33.6000"E | KT884630                                         | KT884653                                         | KT924351                                       |
| CUPVoucher-FB-2014-20| TN: Tamil Nadu (Adyar)  | Dr Felix Bast             | 20-02-2014      | 13°09'22.6800"N | 080°15'26.6400"E | KT884624                                         | KT884647                                         | KT924367                                       |

*– No data available.*
likelihood model test in MEGA v7. Pairwise distances between sequences were calculated using p-distance model in MEGA v7 (http://www.megasoftware.net/). After analysis, a consensus tree was generated using the consensus tree builder with Geneious v6.1.5.

Morphological assessment of leaves and DNA sequencing data were congruent in the identification of the test samples as *F. benghalensis*. There were no significant intraspecific morphological differences observed in the samples, suggesting that the species is phenotypically least influenced by climatic variations.

In total, 58 sequences for three primers sets, i.e. ITS1, rps16 and trnL were generated. The length of these sequences was varied from 700 to 1300 bp. Ambiguous sequences of low quality were excluded. This study reports the barcoding record of *F. benghalensis* with rps16 intron. Phylogenetic reconstruction using Bayesian inference for rps16 dataset, with *F. formosana, F. erecta, F. vaccinoides, F. benguetensis* and *F. variegata* as outgroup, generated a single monophyletic clade within section *Conosycea* consisting of all isolates of *F. benghalensis* (Figure 2). Likewise, the phylogenetic reconstruction for ITS1 loci with *F. crassiramea* and *F. pubilimba* as out group, generated a single monophyletic clade within section *Conosycea* (Figure 3). Due to the fast-evolving nature of ITS1 and rps16, these primers are generally expected to be more variable than the conserved and slow evolving ones like trnL, but nesting of all sequences in a single clade suggests poor resolution of these markers for *Ficus* spp. The lower genetic variation observed in ITS1 and rps16 as well as the presence of a single widespread haplotype in India are suggestive of long-distance nuclear and chloroplast gene flow via pollen.

However, phylogenetic analysis performed for trnL sequences generated two haplotypes having zero within-group mean distance and $2 \times 10^{-3}$ between-group mean distance, calculated using Tamura-3-parameter with 500 bootstrap replications (Figure 4). The population differentiation into two haplotypes depicted by trnL data suggests that gene flow in this case must have been restricted. This intraspecific variation can possibly be attributed to climatic variations. These two haplotypes may also pertain to two different cohesion species arising from the genetic and ecological factors creating different reproductive communities. However, reproductive lineage data would be required to validate them as two cohesion species.

The present study provides evidence of substantial phylogeographic distribution of *F. benghalensis* into two haplotypes. Although posterior probability values are not convincing, they leave us with the curiosity of further studies on the same. Earlier reports have also emphasized the plastid gene flow; however, conflicts arising because of existing understanding of *Ficus* and nuclear phylogeny could not be resolved. Despite issues of ancient hybridization, plastid-based phylogeny could address more conflicts in comparison to nuclear, suggesting plastid-mediated gene flow. Our result is congruent with the

![Figure 2](image-url) **Figure 2.** Phylogenetic position of *Ficus benghalensis* isolates among other related species in rps16 dataset using Bayesian inference phylogenetic reconstruction (LnL0 = −1308.789) with Hasegawa–Kishino–Yano model of molecular evolution with equal distribution. Numbers near nodes represent Bayesian posterior probabilities. Scale bar represents average nucleotide substitutions per site.

![Figure 3](image-url) **Figure 3.** Phylogenetic position of *F. benghalensis* isolates among other related species in ITS1 dataset using Bayesian inference phylogenetic reconstruction (LnL0 = −525.529) with Jukes–Cantor model of molecular evolution with equal distribution. Numbers near nodes represent Bayesian posterior probabilities. Scale bar represents average nucleotide substitutions per site.
and regression models can be used to implement SDM, such that spatio-temporal distribution of the species can be predicted from environmental data provided through GIS. SDM-coupled phylogeography is also expected to forecast the predictive species distributions of the concerned species on future climate models and help in devising better strategies to preserve the genetic diversity.

This study provides insights into genetic heterogeneity of *F. benghalensis* across India, and contributes to the sequence data of the species by barcoding with rps16. The study also validates the efficacy of trnL for molecular phylogeny for *Ficus* species. For a better understanding of the causes of the lower genetic variation observed, SDM-coupled phylogeography is suggested.

**Conflict of interest:** The authors declare that they have no conflict of interest.

1. Chantararusan, B., Berg, C. C., Kjellberg, F., Ronsted, N., Garcia, M., Baider, C. and van Welzen, P. C., A new classification of *Ficus* subsection *Urostigma* (Moraceae) based on four nuclear DNA markers (ITS, ETS, G3pdh, and ncP GS), morphology and leaf anatomy. *PloS One*, 2015, 10(6); https://doi.org/10.1371/journal.pone.0128289.
2. Machado, C. A., Robbins, N., Gilbert, M. T. P. and Herre, E. A., Critical review of host specificity and its coevolutionary implications in the fig/fig–wasp mutualism. *Proc. Natl. Acad. Sci. USA*, 2005, 102, 6558–6565.
3. Ronsted, N., Weiblen, G. D., Cook, J. M., Salamin, N., Machado, C. A. and Savolainen, V., 60 million years of co-divergence in the fig–wasp symbiosis. *Proc. R. Soc. London, Ser. B*, 2005, 272, 2593–2599.
4. Güven, K. C., Percot, A. and Sezik, E., Alkaloids in marine algae. *Mar. Drugs*, 2010, 8(2), 269–284.
5. Xu, L., Harrison, R. D., Yang, P. and Yang, D. R., New insight into the phylogenetic and biogeographic history of genus *Ficus*: vicariance played a relatively minor role compared with ecological opportunity and dispersal. *Int. J. Syst. Evol. Microbiol.*, 2011, 61(6), 546–557.
6. Bhalerao, S. A., Poojari, A. C. and Sharma, A. C., Ethnobotany, phytochemical properties and pharmacological review of *Ficus benghalensis* Linn. *Indian J. Pharm. Sci.*, 2015, 4(8), 372–381.
7. Bhaskara Rao, K. V., Ojha, V., Preeti, Kumar, G. and Karthik, L., Phytochemical composition and antioxidant activity of *Ficus benghalensis* (Moraceae) leaf extract. *J. Biol. Act. Prod. Nat.*, 2014, 4(3), 236–248.
8. Ronsted, N., Weiblen, G. D., Clement, W. L., Zerega, N. J. C. and Savolainen, V., Reconstructing the phylogeny of figs (*Ficus*, Moraceae) based on four nuclear DNA markers (ITS, ETS, G3pdh, and ncP GS). *Proc. Natl. Acad. Sci. USA*, 2015, 112, 7939–7944.
9. Chowdhary, N., Kaur, M., Singh, A. and Kumar, B., Wound healing activity of aqueous extracts of *Ficus religiosa* and *Ficus benghalensis* leaves in rats. *Ind. J. Res. Pharm. Biotechnol.*, 2014, 2(2), 1071.
10. Govindan, V. and Francis, G. S., Qualitative and quantitative determination of secondary metabolites and antioxidant potential of *Ficus benghalensis* Linn. seeds. *Int. J. Pharm. Pharm. Sci.*, 2015, 7, 118–124.
11. Baraket, G., Olfa, S., Khaled, C., Messaoud, M., Mohamed, M., Mokhtar, T. and Amel, S. H., Chloroplast DNA analysis in Tunisian fig cultivars (*Ficus carica L.*): sequence variations of the trnL–tmF intergenic spacer. *Biochem. Syst. Ecol.*, 2008, 36(11), 828–835.

Figure 4. Phylogenetic position of *F. benghalensis* isolates among other related species in trnL dataset using Bayesian inference phylogenetic reconstruction (LnL0 = −1581.828) with Hasegawa–Kishino–Yano model of molecular evolution with equal distribution. Numbers near nodes represent Bayesian Posterior Probabilities. Scale bar represents average nucleotide substitutions per site.

Previously described notions of molecular markers, further categorizing the geographical isolates into two putative haplotypes. Similar edaphic heterogeneity was also observed in *Protrium* spp. (Burseraceae) across Amazon Basin, where varied degree of edaphic specialization and division of the population in eastern and western zones was reported. There are several reports available for the clonal variations and adaptive differentiation of tree species in quantitative traits reflecting local environmental origins. Structured genetic diversity was observed amongst two groups of foundation species in South Western Australia following by climatic gradient and newly formed and expanded central refugia. The succeeding contraction at the Last Glacial Maximum (LGM) was identified in *Eucalyptus wandoo* Blakely.

Pleistocene glacial events generated periodic contraction and expansion of species ranges, leaving genetic fingerprints on existing populations. The low level of genetic diversity observed in *F. benghalensis* can be attributed to genetic bottlenecks during palaeo-historical events after LGM. However, to validate this hypothesis, species distribution modelling (SDM) based on the principle of climate-restricted species distribution within their evolutionary and biogeographic history is suggested, which can serve as an important tool to provide better understanding about the evolutionary patterns of current populations of *F. benghalensis*. Various niche-modelling algorithms such as artificial neural networks and regression models can be used to implement SDM, such that spatio-temporal distribution of the species can be predicted from environmental data provided through GIS. SDM-coupled phylogeography is also expected to forecast the predictive species distributions of the concerned species on future climate models and help in devising better strategies to preserve the genetic diversity.

This study provides insights into genetic heterogeneity of *F. benghalensis* across India, and contributes to the sequence data of the species by barcoding with rps16. The study also validates the efficacy of trnL for molecular phylogeny for *Ficus* species. For a better understanding of the causes of the lower genetic variation observed, SDM-coupled phylogeography is suggested.

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RESEARCH COMMUNICATIONS

12. Poeaim, A., Poeaim, S., Sotyong, K. and Krajangyuthi, T., Genetic diversity of Ficus carica L. based on non-coding regions of chloroplast DNA. In 8th International Symposium on Biocntrol and Biotechnology, 2012.

13. Rakshith, D., Santosh, P. and Satish, S., Isolation and characterisation of antimicrobial metabolite producing endophytic Phomopsis sp. from Ficus pumila Linn. (Moraceae). Int. J. Chem. Anal. Sci., 2013, 4(3), 156–160.

14. Ovelman, B., Liden, M. and Berglund, D., Chloroplast rps16 intron phylogeny of the tribe Sileaneeae (Caryophyllaceae). Plant. Syst. Evol., 1997, 206(1–4), 393–410.

15. Bast, F., Sequence similarity search, multiple sequence alignment, model selection, distance matrix and phylogeny reconstruction. Nature Protoc. Exchange, 2013; doi:10.1038/protex.2013.065.

16. Honorio Coronado, E. N., Dexter, K. G., Poelchau, M. F., Hollingsworth, P. M., Phillips, O. L. and Pennington, R. T., Ficus insipida subsp. insipida (Moraceae) reveals the role of ecology in the phylogeography of widespread Neotropical rain forest tree species. J. Biogeogr., 2014, 41(9), 1697–1709.

17. Templeton, A. R., Using phylogeographic analyses of gene trees to test species status and processes. Mol. Ecol., 2001, 10(3), 779–791.

18. Bruun-Lund, S., Clement, W. L., Kjellberg, F. and Roesten, N., First plastid phylogenomic study reveals potential cyto-nuclear discordance in the evolutionary history of Ficus L. (Moraceae). Mol. Phylogenet. Evol., 2017, 109, 93–104.

19. Fine, P. V., Zapata, F., Daly, D. C., Mesones, I., Misiewicz, T. M., Cooper, H. F. and Barboza, C. E. A., The importance of environmental heterogeneity and spatial distance in generating phylogeographic structure in edaphic specialist and generalist tree species of Protium (Burseraceae) across the Amazon Basin. J. Biogeogr., 2013, 40(4), 646–661.

20. Alberto, F. J. et al., Potential for evolutionary responses to climate change – evidence from tree populations. Global Change Biol., 2013, 19(6), 1645–1661.

21. Dalmaris, E., Ramalho, C. E., Poot, P., Veneklaas, E. J. and Byrne, M., A climate change context for the decline of a foundation tree species in south-western Australia: insights from phylogeography and species distribution modelling. Ann. Bot., 2015, 116(6), 941–952.

22. Mayol, M. et al., Adapting through glacial cycles: insights from a long-lived tree (Taxus baccata). New Phytol., 2015, 206(3), 973–986.

23. Wolfbriigh, S. A., Whitham, T. G., Gehring, C. A., Allan, G. J. and Bailey, J. K., Climate relicts and their associated communities as natural ecology and evolution laboratories. Trends Ecol. Evol., 2014, 29(7), 406–416.

24. Roberts, D. R. and Hamann, A., Glacial refugia and modern genetic diversity of 22 western North American tree species. Proc. R. Soc. London, Ser. B, 2015, 282(1804), 20142903.

25. Wisz, M. S. et al., The role of biotic interactions in shaping distributions and realised assemblages of species: implications for species distribution modelling. Biol. Rev., 2013, 88(1), 15–30.

26. Khan, S., Al-Qurainy, F., Nadeem, M. and Tarroum, M., Development of genetic markers for Ochrudenus arabicus (Resedaceae), an endemic medicinal plant of Saudi Arabia. Genet. Mol. Res., 2012, 11(2), 1300–1308.

ACKNOWLEDGEMENTS. The study was supported by grant-in-aid ‘Research Seed Money’ scheme of the Central University of Punjab, Bathinda. We thank the Vice-Chancellor, Central University of Punjab for support. We also thank Dr Pankaj Bhardwaj for support in experimental works and all those who helped in sample collection. R.M. thanks the Indian Council of Medical Research, New Delhi for financial support (JRF).

Received 31 October 2017; revised accepted 30 March 2019

doi: 10.18520/cs/v117i2/293-298

Magmatic epidote in the Grenvillian granitoids of North Purulia Shear Zone, Chhotanagpur Gneissic Complex, India and its significance

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Magmatic epidote planel from North Purulia Shear Zone, eastern India, are identified by textural and chemical criteria. The accessory sphere, epidote, allanite and magnetite in the granitoid indicate high O2 during crystallization. Hornblende were crystallized between 1.2 and 4.8 kbar, 753°C and 783°C as well as high O2 (>Ni–NiO buffer). Preservation of magmatic epidote in pluton emplaced at low pressure is due to rapid magma ascent (>3.1 km/year). Fast upward transportation of parental magma of the pluton took place through extensional voids along the regional shear zone.

Keywords. Granitoids, magmatic epidote, oxygen fugacity, shear zone emplacement.

The petrological implications of magmatic epidote (mEp) have been petrographically and experimentally demonstrated by several workers1–4. The presence of mEp in tonalites and granodiorites suggests that the plutos crystallized under lithostatic pressure >6 kbar (refs 2, 3). Survival of mEp in calc-alkaline granitoids implies rapid upward movement of the magma4. Consequently, mEp can be used for estimating crystallization pressure, oxygen fugacity and rate of upward movement of melt. Studies on mEp in the Neoarchaean granitoids of Srinivasapura (Eastern Dharwar Craton) and the Paleoproterozoic Malanjkhand Granitoid (Central India) revealed upward magma migration rate of 27.65 and 0.45 km/year respectively5,6. In the present study, mEp is distinguished from secondary epidote based on textural and chemical criteria in the granitoids of Agarpur pluton lying in the North Purulia Shear Zone (NPSZ) of Chhotanagpur Gneissic Complex (CGC) of eastern India. The significance of mEp on emplacement mechanism of granitoids is also discussed.

The ENE–WSW-trending Central Indian Tectonic Zone and CGC mark the Grenvillian collisional zone between the North Indian Block and the South Indian Block7–9 (Figure 1 a). The CGC is mainly composed of granitoid and migmatitic gneisses with older enclaves of para- and ortho-metamorphic rocks10. The structural trend of the rocks is E-W with steep northern dip. Younger

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