Associations of Mitochondrial DNA 3777-4679 region mutations with maternally inherited essential hypertensive subjects in China.

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Abstract

Background: Nuclear genome or family mitochondrial screening system has become the hot focus of studies into essential hypertension. The role of mitochondrial DNA (mtDNA) in sporadic Chinese patients with hypertension has not been fully understood. The study was to evaluate the associations of mtDNA mutations with maternally inherited essential hypertensive subjects in China. Methods: From June 2009 to June 2016, a total of 800 gender-matched Chinese patients with maternally inherited essential hypertension (MIEH) and control group were 1:1 enrolled in this case-control study. Genomic DNA was extracted from each person’s peripheral blood cells. The main mtDNA locations for MIEH were screened with oligodeoxynucleotides 3777-4679bp, analyzed and compared with the updated consensus Cambridge Sequence. Pathogenic mtDNA mutations were identified from the mitochondrial map. Results: MIEH subjects presented significantly higher values than those of control group in abdominal circumference (AC), waist circumference (WC), body mass index (BMI), fasting blood glucose (FBG), triglyceride (TG), low-density lipoprotein cholesterol (LDL) and renal function (P < 0.05). MIEH subjects carried more amino acid changes and coding sequence variants (P < 0.01) than control group. The allele frequencies of the eight single nucleotide polymorphisms (SNPs) were significantly different between the two groups, including m.3970 C>T, m.4048G>A, m.4071C>T, m.4086C>T, m.4164A>G and m.4248T>C in ND1 gene, and m.4386T>C and m.4394C>T in tRNA Gln gene (P < 0.001). Fifty-five homoplasmic or heteroplasmic mutations were detected in 5 genes: ND1, tRNA Ile, tRNA Met, tRNA Gln and ND2 gene. The ND1 gene was the main mutation site, where the most mtDNA mutation was m.3970 C>T. Conclusions: The mtDNA mutations were involved in the process of MIEH. We identified mitochondrial genetic characteristics in MIEH patients in China. The present research serves as a solid foundation for further detailed research on the association between MIEH and mitochondrial dysfunction, and their causal relationship in Chinese and other populations with a similar lifestyle.

Background

Essential hypertension (EH) is a common cardiovascular disorder, influencing about 1 billion people around the world [1]. It is generally believed that the interaction between genic and environmental
factors affected EH, which may be caused by single-gene defects or multifactorial conditions[2]. Maternally inherited essential hypertension (MIEH) is EH that is consistent with the pattern of maternal inheritance [3]. Mitochondrial DNA (mtDNA) can cause mitochondrial diseases, which are transmitted from the mother exclusively. MtDNA mutations were marked in several pathogenic disorders including mitochondrial myopathy, stroke-like attacks, encephalopathy and maternally hereditary diabetes[4]. In addition, mutations in mtDNA have also been observed to play a role in the pathogenesis of MIEH [5]. The repair and protection systems of mtDNA are less efficient compared to that of nuclear DNA [6]. MtDNA mutations have been linked to MIEH through modifying several functional tRNAs [7]. In particular, a previous study identified that the mutation may reduce the steady-state level of mt-tRNA^{Gln} m.4375C>T and subsequently cause the mitochondrial dysfunction that is responsible for hypertension[8]. So far, the mechanism of mtDNA mutations in MIEH has not been completely elucidated, especially on the interplay between mtDNA mutations and other risk factors, such as the development of blood pressure, nuclear genes, and environmental conditions[9]. While people are more concerned about the role of the nuclear genome[10], investigating the role of mtDNA sequence alteration may help to understand the genetic pathogenesis of MIEH. Mitochondrial genes in 3777-4679 region were proposed to be hot spots for mutations associated with hypertension as described previously[11]. In order to better understand the pathogenic mechanisms underlying MIEH, we studied clinical and genetic evidence to investigate the association between the mtDNA mutations in 3777-4679 region and MIEH. In this study, we focused on the Han Chinese population, as there is a limited amount of study on this racial group and they might be overlooked for lacking medical knowledge and regular examinations[12].

Methods

Subjects

The current case-control study was based on 400 unrelated patients with MIEH and 400 healthy control individuals in the Jiangsu Province of China. The MIEH patients were recruited according to the following inclusion criteria:
(1) in-patients or outpatients who have undergone regular medical check-up at the Department of Cardiology in Northern Jiangsu People’s Hospital from June 2009 to June 2016;
(2) more than 18 years old;
(3) with a diagnosis of primary hypertension;
(4) diagnosed with MIEH on the basis of the maternal transmission of EH within generations, which was transmitted by the mother or her relatives, rather than by the father.

Participants were excluded if they were diagnosed as follows:
(1) secondary hypertension (e.g. aortic coarctation, renal arterial stenosis, hyperaldosteronism, and pheochromocytoma);
(2) congenital cardiovascular disease;
(3) organic valve diseases.

Another 400 gender-matched healthy individuals were recruited to the current study as controls. Furthermore, the controls were unrelated healthy subjects from the same area who received annual examination in physical examination center of Northern Jiangsu People’s Hospital. They were collected randomly from the physical examination list. The control group included the following criteria:
(1) systolic blood pressure (SBP) of <130mmHg and diastolic blood pressure (DBP) of <85mmHg.
(2) no personal or family history of hypertension.

Hypertension in one or both biologic parents was considered to be a positive family history of EH. All subjects in the study were interviewed to identify both personal and family medical histories of clinical abnormalities. Verbal Informed Consent, medical history, clinical assessment and genetic analysis were obtained from each individual under protocols involved in the study. Verbal consent is that the mtDNA analysis is used only for diagnosis, not for treatment. It was of no harm to anyone. The protocol was implemented in accordance with the Declaration of Helsinki and approved by the ethics committee of the Northern Jiangsu People’s Hospital.

**Data collection**

Body mass index (BMI) refers to a person’s body mass in kilograms divided by height in square
meters (kg/m²). Patients reporting cigarette use within one year prior to examination were considered as smokers. Blood pressure was measured by an experienced physician who was blinded to the study according to the criteria of the World Health Organization (WHO) [13]. Three measurements of systolic and diastolic blood pressure were taken and the mean value was used as the measurement. According to the 2010 Chinese Hypertension Management, hypertension was diagnosed as follows [14]: the SBP > 140 mm Hg and/or DBP > 90 mm Hg measured three times on different days or a history of hypertension with current antihypertensive medications. All participants also underwent laboratory tests on hypertension risk factors. 12 hours after fasting, lipid profile, fasting blood glucose (FBG), and kidney function test were performed by an automatic biochemistry analyzer (Hitach 7600DDP, Japan).

**Mitochondrial DNA analysis**

Genomic DNA was extracted from each person’s peripheral blood using standard protocols [15]. MtDNA was isolated by Promega Wizard Genomic DNA Purification Kit (Madison, WI, USA). The main chromosome locations for hypertension as described previously [16] were screened using oligodeoxynucleotides 3777-4679bp. The mitochondrial tRNA^{ille} gene was amplified by Polymerase chain reaction (PCR) using the primer sequences as follows: forward: 5′- TGGCTCCTTTAACCTCTCCA-3′ and reverse: 5′- AAGGATTATGGATGCGGTTG -3′. PCR cycle program was carried out in a 9700 Thermocycler (Perkin-Elmer Applied Biosystems, Norwalk, USA). Each fragment had been purified and sequenced by ABI 3730 Sequence Analysis software (Applied Biosystems, Inc., Foster City, CA, USA) using the BigDye Terminator v1.1 kit (ABI Company, Carlsbad, CA, USA), and subsequently SeqWeb program GAP(GCG) was analyzed and compared with the updated consensus Cambridge Sequence [17, 18]. Pathogenic variants were identified from the mitochondrial map (http://www.mitomap.org/) [19].

**Statistical analysis**

Statistical analysis was performed using R and SPSS software (version 22.0; SPSS Inc., Chicago, IL, USA). Continuous variables were tested for normal distribution by Kolmogorov-Smirnov test and then
expressed by mean ± standard deviation (SD). The relationship between potential continuous and discrete factors and MIEH were analyzed with Student’s t-test and Fisher’s exact t-test. A P-value ≤0.05 was considered to be statistically significant.

Results

Clinical evaluation of baseline characteristics

Table 1 shows the main characteristics of all participants. In the present study, age and SBP did not differ between the two groups. There were significant differences in BMI, waist circumference(WC), abdominal circumference (AC), total cholesterol(TC), triglycerides(TG), low-density lipoprotein cholesterol (LDL), FBG, uric acid(UA), creatinine(CR) as well as blood urea nitrogen(BUN) (P<0.05) between the two groups.

mtDNA analysis

The distribution of mutations in the mtDNA 3777-4679 bp of all participants is shown in Figure 1. Table 2 shows a comparison of the frequency of mtDNA sequence analyses in the 400 balanced cases and controls. MtDNA analysis revealed 55 mutation sites in the 400 MIEH subjects (Table 3). The ND1 gene was the main mutation site, where the highest mutation frequency was m.3970 C>T Figure 2a. These results showed that the mtDNA in the MIEH group had more variations than the control group. The allele frequencies of eight single nucleotide polymorphisms (SNPs) were significantly (P<0.001) different between the two groups, including m.3970 C>T, m.4048G>A, m.4071C>T, m.4086C>T, m.4164A>G and m.4248T>C (Figure 2b) in ND1 gene, and m.4386T>C and m.4394C>T in tRNA^Gln gene. 55 heteroplasmic or homoplasmic mutations were detected in 5 genes: ND1, tRNA^Ile, tRNA^Met, tRNA^Gln and ND2 gene. We found 45 homoplasmic mutations in 267 subjects of MIEH. Ten heteroplasmic mutations were found in 11 MIEH subjects. MIEH subjects carried more amino acid changes and coding sequence variants (P<0.01) compared with normotensive(NT) individuals. An interesting observation in MIEH patients was that we found 4 deletions: m.4563 delG (Figure 2c), m.4576 delA, m.4611 delA (Figure 2d) and m.4612 delT mutations in MIEH group patients and the site of highest deletion frequency (10/400=0.025) was m.4611 delA mutation in 10 MIEH subjects. The results suggested that mtDNA mutations were positively correlated with MIEH.
Discussion

The purpose of this study was to investigate mutations in mtDNA 3777-4679 region of the Chinese MIEH population. The results showed that the group of MIEH had more mtDNA variations, which mainly located at ND1 site and the highest mutation site was m.3970C>T. Mitochondrial dysfunction is an important factor in cardiovascular disorders [20]. As far as we know, this is one of the first large-scale Han Chinese population-based studies about and the potential role of mtDNA in MIEH. The systematic screen of the association between MIEH and mtDNA mutation is not only essential to further our understanding of the specific mechanism of the mutation in disease etiology, but can also improve the diagnosis and treatment of hypertension.

In an attempt to determine whether mtDNA was involved in the biochemical indicators of the MIEH individuals, we compared and analyzed the biochemical abnormalities of all the participants. Clinical evaluation of all the participants suggested that MIEH subjects presented significantly higher values than those of control group in BMI, WC, AC, LDL, TG, FBG and renal function. Notably, MIEH individuals were obese or overweight in comparison to NT group. High BMI for EH was reported as a strong predictor of hypertension in a multivariate analysis [21]. Therefore, a normal body weight (BMI 18.5-24.9 kg/m²) should be recommended for the prevention and management of MIEH [22]. These factors might contribute to the occurrence and development of MIEH, or they might occur as a result of the progress of MIEH, which results in the damage of the target organ.

Many studies have depicted the function of inherited mtDNA mutations in MIEH family, such as the analysis of maternally transmitted hypertension in a large Han Chinese cohort [23]. Here, we investigated the mtDNA 3777-4679 region using PCR amplification and the sequence analysis. Current results showed that eight SNPs were significantly different between the MIEH and the control groups: m.3970C>T, m.4048G>A, m.4071C>T, m.4086C>T, m.4164A>G and m.4248T>C in ND1 gene, m.4386T>C and m.4394C>T in tRNA^Gln^ gene. ND1 gene is a hotspot for mutations linked to MIEH. Notably, the impaired synthesis of ND1 (subunits of OXPHOS complex I) may be specifically responsible for the decreased activities of complex I [24]. Mitochondrial encoded complex I is a crucial component in the respiratory chain. The ND1 variants in complex I early in the OXPHOS process,
which may contribute to the production of reactive oxygen species (ROS) and may participate in key functional development processes of EH[25].

Mutations of mtDNA may lead to disease, and the significant determinant of their clinical manifestation are susceptible to associated with mutations arising in mtDNA [26]. In this study, unlike mild elevation of total cellular ROS production, an increase in the cells carrying the m.4248T>C and m.3970 C>T mutations suggested that the defective mitochondria are the major producers of ROS. In turn, the increased levels of cytosolic ROS may produce damage to mitochondrial proteins, nucleic acids, stimulating a forward feeding loop of mitochondrial ROS generation and aggravated cell damage [27]. The reduced levels of mitochondrial proteins were observed in cell lines carrying hypertension associated homoplasmic tRNA^Gln m.4386T>C and m. 4394 C>T mutations belonging to mitochondrial haplogroup M. Ancestral variations in human mtDNA define population-specific mtDNA lineages or haplogroups. Importantly, these were used to trace the origins of different races and provide a foundation for mitochondria based evolutionary medicine [28]. The association between mtDNA haplogroup/SNPs and hypertension has been attributed to a significant alteration in the structure of tRNA and may decrease the steady-state levels of tRNA and oxygen consumption rates that may affect protein levels [29]. Failing to keep the balance of oxygen consumption and production induces mitochondrial dysfunction which was implicated in hypertension pathophysiology, as mentioned earlier [30]. Therefore, mtDNA mutations are promising novel biomarkers for the early detection, prevention, and management of MIEH.

MtDNA mutations, incorporating point mutations, deletions that affect transcription and translation of mtDNA are implicated in various mitochondrial disorders. Rocha et al. have previously revealed a strong correlation between mitochondrial genetics and respiratory chain deficiency in patients with single, large-scale mtDNA deletion [31]. In line with the previous report, our intriguing observation is that there were also a lot of sporadic single mtDNA deletions in MIEH patients [32]. The mean onset time of hypertension for patients with mtDNA point mutations and deletions was gradually ahead of schedule. Thus, it suggests that mtDNA is probably the molecular cause of this disorder. Deletions arise in the mtDNA of post-mitotic cells in patients with mtDNA maintenance disorders, especially
during aging. The impaired mitochondrial function may contribute to increased blood pressure of aging with the accumulated multiple mtDNA deletions later in life. The later onset of clinical symptoms due to mtDNA deletions is likely due to the multiple copies of mtDNA within cells. Therefore, the time for mtDNA deletions exceeded an acritical biochemical threshold in proportion, which is necessary to cause a bioenergetic deficit. The mtDNA deletions are thought to be caused by copying errors or repair, or a combination of the both, with the rate of mtDNA deletion formation likely depends on the disorder, the tissue and mechanism of mtDNA replication [32]. We found that the mtDNA deletions disrupted most of the MT-ND2 genes. This suggests there may be duplications with breakpoints that affect complex I, explaining their isolated complex I defect, or there are other deletions we have not found yet. A possible explanation of mtDNA defects in hypertension is that clonally expanded mtDNA deletions are known to be an underlying cause of mitochondrial OXPHOS deficiency in post-mitotic cells. There are studies demonstrating higher levels of mtDNA deletion heteroplasmy in cytochrome c oxidase deficient cells, ultimately initiating hypertension [33]. Some specific deletions in mitochondria DNA may have an underlying role in Chinese hypertensives due to their dysfunction. How mitochondrial mutations and/or deletions interact synergistically in the phenotypic manifestation of hypertension is the next question, which needs further investigation. However, there is still uncertainty regarding the exact mechanisms by which mtDNA deletions clonally expand in patients with mtDNA maintenance disorders. The present study was the primitive step in evaluating the role of mitochondria in unrelated Chinese hypertension patients.

We observed some limitations in this study. Our result from the single-center with relatively small sample size needs further validation. We studied the mtDNA from peripheral blood, which may not represent the mitochondrial activity in other tissues [34]. Another limitation was the lack of functional experiments. Further investigations should be performed to determine the mitochondrial dysfunctions caused by mtDNA mutations, including assessment of ROS production, ATP production and so on.

Conclusions
The mtDNA mutations were involved in the pathological process of MIEH. Additionally, we identified the mitochondrial genetic characteristics of MIEH patients in Chinese Han population. The
investigation of the role of mitochondrial dysfunction in MIEH provides new insights into the understanding and treatment of the disorder. The present research serves as a solid foundation for further detailed research on the association between MIEH and mitochondrial dysfunction, and their causal relationship in Chinese and other populations with a similar lifestyle.

**Abbreviations**

MtDNA: Mitochondrial DNA; MIEH: Maternally inherited essential hypertension; EH: Essential hypertension; SBP: Systolic blood pressure; DBP: Diastolic blood pressure; BMI: Body mass index; WC: Waist circumference; AC: Abdominal circumference.

**Declarations**

**Ethics approval and consent to participate**

Verbal informed consent was obtained from the subjects involved in the study. The protocol was conducted in accordance with the Declaration of Helsinki and approved by the institutional ethical board of Northern Jiangsu People’s Hospital, Jiangsu Province, China.

**Consent to publish**

Not applicable.

**Availability of data and materials**

The datasets analysed during the current study are available at [http://www.mitomap.org](http://www.mitomap.org) and [http://www.mtdb.igp.uu.se/](http://www.mtdb.igp.uu.se/). The datasets analysed during the current study available from the corresponding author on reasonable request.

**Competing interests**

The authors declare that they have no competing interests.

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**Authors' contributions**

YZ, JY and XG carried out the studies, participated in collecting data, and drafted the manuscript. YZ
and CX performed the statistical analysis and participated in its design. CX helped to draft the manuscript. All authors read and approved the final manuscript.

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References

1. Wang L, Dong Z, Lin W, Gao R, Chen C, Xu J: Molecular characterization of a pedigree carrying the hypertension associated mitochondrial tRNAGln T4363C mutation. Molecular medicine reports 2017, 16(5):6029-6033.

2. Shen Y, Peng X, Wang M, Zheng X, Xu G, Lu L, Xu K, Burstrom B, Burstrom K, Wang J: Family member-based supervision of patients with hypertension: a cluster randomized trial in rural China. J Hum Hypertens 2017, 31(1):29-36.

3. Qiu Q, Li R, Jiang P, Xue L, Lu Y, Song Y, Han J, Lu Z, Zhi S, Mo JQ et al: Mitochondrial tRNA mutations are associated with maternally inherited hypertension in two Han Chinese pedigrees. Human mutation 2012, 33(8):1285-1293.

4. Schapira AH: Mitochondrial disease. Lancet (London, England) 2006, 368(9529):70-82.

5. Inaba Y, Chen JA, Bergmann SR: Carotid plaque, compared with carotid intima-media thickness, more accurately predicts coronary artery disease events: a meta-analysis. Atherosclerosis 2012, 220(1):128-133.

6. Liu P, Demple B: DNA repair in mammalian mitochondria: Much more than we thought? Environmental and molecular mutagenesis 2010, 51(5):417-426.

7. Lu Z, Chen H, Meng Y, Wang Y, Xue L, Zhi S, Qiu Q, Yang L, Mo JQ, Guan MX: The tRNAMet 4435A>G mutation in the mitochondrial haplogroup G2a1 is
responsible for maternally inherited hypertension in a Chinese pedigree.

*European journal of human genetics : EJHG* 2011, **19**(11):1181-1186.

8. Chen H, Sun M, Fan Z, Tong M, Chen G, Li D, Ye J, Yang Y, Zhu Y, Zhu J:

*Mitochondrial C4375T mutation might be a molecular risk factor in a maternal Chinese hypertensive family under haplotype C*. *Clin Exp Hypertens* 2018, **40**(6):518-523.

9. Kirichenko TV, Sobenin IA, Khasanova ZB, Orekhova VA, Melnichenko AA, Demakova NA, Grechko AV, Orekhov AN, Ble Castillo JL, Shkurat TP: *Data on association of mitochondrial heteroplasmy and cardiovascular risk factors: Comparison of samples from Russian and Mexican populations*. *Data Brief* 2018, **18**:16-21.

10. Deng H, Yuan L: *Molecular genetics of congenital nuclear cataract*. *Eur J Med Genet* 2014, **57**(2-3):113-122.

11. Zhu HY, Wang SW, Martin LJ, Liu L, Li YH, Chen R, Wang L, Zhang ML, Benson DW:

*The role of mitochondrial genome in essential hypertension in a Chinese Han population*. *European journal of human genetics : EJHG* 2009, **17**(11):1501-1506.

12. Han GX, Song Y, Chen L, Zhai YZ, Dong J, Li TS, Zhu HY: *The role of mitochondrial DNA mutations in a Han Chinese population on sepsis pathogenesis*. *Eur Rev Med Pharmacol Sci* 2017, **21**(14):3247-3252.

13. Chalmers J, MacMahon S, Mancia G, Whitworth J, Beilin L, Hansson L, Neal B, Rodgers A, Ni Mhurchu C, Clark T: *1999 World Health Organization-International Society of Hypertension Guidelines for the management of hypertension*. Guidelines sub-committee of the World Health Organization. *Clinical and experimental hypertension (New York, NY : 1993)* 1999, **21**(5-6):1009-1060.

14. Liu LS: *2010 Chinese guidelines for the management of hypertension*.

*Zhonghua xin xue guan bing za zhi* 2011, **39**(7):579-615.
15. Zhu HY, Wang SW, Liu L, Li YH, Chen R, Wang L, Holliman CJ: **A mitochondrial mutation A4401G is involved in the pathogenesis of left ventricular hypertrophy in Chinese hypertensives.** *European journal of human genetics: EJHG* 2009, **17**(2):172-178.

16. Liu Y, Li Y, Gao J, Zhu C, Lan Y, Yang J, Li Z, Guan M, Chen Y: **Molecular characterization of a Chinese family carrying a novel C4329A mutation in mitochondrial tRNAlle and tRNAGln genes.** *BMC medical genetics* 2014, **15**:84.

17. Echem C, Costa TJD, Oliveira V, Giglio Colli L, Landgraf MA, Rodrigues SF, Franco M, Landgraf RG, Santos-Eichler RA, Bomfim GF et al: **Mitochondrial DNA: A new driver for sex differences in spontaneous hypertension.** *Pharmacol Res* 2019, **144**:142-150.

18. Zhu Y, Gu X, Xu C: **Mitochondrial DNA 7908-8816 region mutations in maternally inherited essential hypertensive subjects in China.** *BMC Med Genomics* 2018, **11**(1):89.

19. Ingman M, Gyllensten U: **mtDB: Human Mitochondrial Genome Database, a resource for population genetics and medical sciences.** *Nucleic acids research* 2006, **34**(Database issue):D749-751.

20. Samuels DC, Li C, Li B, Song Z, Torstenson E, Boyd Clay H, Rokas A, Thornton-Wells TA, Moore JH, Hughes TM et al: **Recurrent tissue-specific mtDNA mutations are common in humans.** *PLoS genetics* 2013, **9**(11):e1003929.

21. Bruno RM, Pucci G, Rosticci M, Guarino L, Guglielmo C, Agabiti Rosei C, Monticone S, Giavarini A, Lonati C, Torlasco C et al: **Association Between Lifestyle and Systemic Arterial Hypertension in Young Adults: A National, Survey-Based, Cross-Sectional Study.** *High Blood Press Cardiovasc Prev* 2016, **23**(1):31-40.

22. Chobanian AV, Bakris GL, Black HR, Cushman WC, Green LA, Izzo JL, Jr., Jones DW,
Materson BJ, Oparil S, Wright JT, Jr. et al: **Seventh report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure.** *Hypertension* (Dallas, Tex : 1979) 2003, 42(6):1206-1252.

23. Chen X, Zhang Y, Xu B, Cai Z, Wang L, Tian J, Liu Y, Li Y: **The mitochondrial calcium uniporter is involved in mitochondrial calcium cycle dysfunction:** Underlying mechanism of hypertension associated with mitochondrial tRNA(Ile) A4263G mutation. *The international journal of biochemistry & cell biology* 2016, 78:307-314.

24. Zhou M, Wang M, Xue L, Lin Z, He Q, Shi W, Chen Y, Jin X, Li H, Jiang P et al: **A hypertension-associated mitochondrial DNA mutation alters the tertiary interaction and function of tRNA(Leu(UUR)).** *The Journal of biological chemistry* 2017, 292(34):13934-13946.

25. Zhao Y, Chen X, Li H, Zhu C, Li Y, Liu Y: **Mitochondrial genome mutations in 13 subunits of respiratory chain complexes in Chinese Han and Mongolian hypertensive individuals.** *Mitochondrial DNA Part A, DNA mapping, sequencing, and analysis* 2018, 29(7):1090-1099.

26. Shimizu A, Mito T, Hayashi C, Ogasawara E, Koba R, Negishi I, Takenaga K, Nakada K, Hayashi J: **Transmitochondrial mice as models for primary prevention of diseases caused by mutation in the tRNA(Lys) gene.** *Proceedings of the National Academy of Sciences of the United States of America* 2014, 111(8):3104-3109.

27. Mohanty K, Dada R, Dada T: **Neurodegenerative Eye Disorders: Role of Mitochondrial Dynamics and Genomics.** *Asia-Pacific journal of ophthalmology (Philadelphia, Pa)* 2016, 5(4):293-299.

28. Zhou H, Nie K, Qiu R, Xiong J, Shao X, Wang B, Shen L, Lyu J, Fang H: **Generation and Bioenergetic Profiles of Cybrids with East Asian mtDNA Haplogroups.**
Oxid Med Cell Longev 2017, 2017:1062314.

29. Crescenzo R, Bianco F, Falcone I, Coppola P, Liverini G, Iossa S: Increased hepatic de novo lipogenesis and mitochondrial efficiency in a model of obesity induced by diets rich in fructose. European journal of nutrition 2013, 52(2):537-545.

30. Singh A, Pezeshki A, Zapata RC, Yee NJ, Knight CG, Tuor UI, Chelikani PK: Diets enriched in whey or casein improve energy balance and prevent morbidity and renal damage in salt-loaded and high-fat-fed spontaneously hypertensive stroke-prone rats. The Journal of nutritional biochemistry 2016, 37:47-59.

31. Rocha MC, Rosa HS, Grady JP, Blakely EL, He L, Romain N, Haller RG, Newman J, McFarland R, Ng YS et al: Pathological mechanisms underlying single large-scale mitochondrial DNA deletions. Annals of neurology 2018, 83(1):115-130.

32. Lehmann D, Tuppen HAL, Campbell GE, Alston CL, Lawless C, Rosa HS, Rocha MC, Reeve AK, Nicholls TJ, Deschauer M et al: Understanding mitochondrial DNA maintenance disorders at the single muscle fibre level. Nucleic acids research 2019.

33. Moskalev AA, Aliper AM, Smit-McBride Z, Buzdin A, Zhavoronkov A: Genetics and epigenetics of aging and longevity. Cell cycle (Georgetown, Tex) 2014, 13(7):1063-1077.

34. Herbers E, Kekalainen NJ, Hangas A, Pohjoismaki JL, Goffart S: Tissue specific differences in mitochondrial DNA maintenance and expression. Mitochondrion 2019, 44:85-92.

Tables
Table 1 Comparison of baseline clinical data between the MIEH and control groups
| Subjects               | MIEH group   | Control group | P-value |
|-----------------------|--------------|---------------|---------|
| gender M/F            | 400202/198   | 400197/203    | 1       |
| age at test (years)   | 68.65±8.34   | 65.36±6.75    | 1.38e-9 |
| age at onset (years)  | 48.56±6.7    | NA            |         |
| SBP (mmHg)            | 148.5±19.8   | 145.6±18.6    | 0.033   |
| DBP (mmHg)            | 94.8±8.9     | 88.4±12.5     | 3.72e-16|
| BMI (kg/m²)           | 25.90±3.60   | 23.52±3.03    | 1.11e-22|
| WC (cm)               | 86.90±10.78  | 78.08±8.71    | 8.52e-34|
| AC (cm)               | 89.50±11.15  | 80.28±7.79    | 2.02e-37|
| Alcohol, n (%)        | 100(25)      | 41(10)        | 5.13e-8 |
| Smoking, n(%)         | 90(23)       | 52(13)        | 5.84e-4 |
| TG (mmol/L)           | 1.86±1.20    | 1.37±0.88     | 8.65e-11|
| TC (mmol/L)           | 4.58±1.96    | 4.25±1.78     | 0.0129  |
| LDL (mmol/L)          | 2.63±1.24    | 2.02±1.39     | 1.04e-10|
| FBG (mmol/L)          | 5.19±2.18    | 4.33±1.84     | 2.54e-09|
| UA (umol/L)           | 368.04±127.28| 323.39±78.92  | 4.02e-09|
| Cr (ummol/L)          | 105.39±33.71 | 87.38±30.71   | 9.40e-15|
| BUN (mmol/L)          | 5.76±2.08    | 4.84±1.79     | 3.86e-11|

Abbreviations: F: female; M: male. SBP: Systolic blood pressure; DBP: Diastolic blood pressure; BMI:
Body mass index; WC: waist circumference; AC: abdomen circumference; TG, triglyceride; TC, total cholesterol; LDL, low-density lipoprotein cholesterol; FBG, fasting blood glucose; UA, uric acid; Cr, creatinine; BUN, blood urea nitrogen:*: A P value <0.05 was marked by a star.

Table 2 Distribution of mtDNA sequence analyses at positions 3777-4679.

| Gene | Position | Length | Control group (n(%)) | MIEH group (n(%)) | Fisher's exact P value |
|------|----------|--------|----------------------|-------------------|-----------------------|
| ND1  | 3777-4262 | 486    | 16                   | 237               | <2.2e-16              |
| tRNA^{Ile} | 4263-4331 | 69     | 0                    | 6                 | 0.03                  |
| tRNA^{Gln} | 4329-4400 | 72     | 0                    | 19                | 3.065e-1               |
| tRNA^{Met} | 4402-4469 | 68     | 0                    | 4                 | 0.12                  |
| ND2  | 4470-4679 | 210    | 1                    | 12                | 0.003                 |

Table 3 Mutation sites of mtDNA in MIEH individuals and controls

| Site of mutation | Gene | Replacement | Homoplasmic/heteroplasmic variants | Number of mutations | MIEH | (Controls) | P value |
|------------------|------|-------------|-----------------------------------|--------------------|------|------------|---------|
| 3902             | ND1  | A to T      | heteroplasmic                     | 1                  | 1    |            | 1       |
| 3905             | ND1  | T to A      | heteroplasmic                     | 1                  | 0    | 0          | 1       |
| 3915             | ND1  | G to A      | homoplasmic                       | 6                  | 1    | 0.1234     |
| 3918             | ND1  | A to G      | homoplasmic                       | 3                  | 1    | 0.6241     |
| 3941             | ND1  | A to G      | homoplasmic                       | 1                  | 0    | 1          |
| 3948             | ND1  | A to G      | homoplasmic                       | 1                  | 0    | 1          |
| 3970             | ND1  | C to T      | homoplasmic                       | 67                 | 5    | 4.72e-16   |
| 4017             | ND1  | C to T      | homoplasmic                       | 1                  | 0    | 1          |
| 4025             | ND1  | C to T      | homoplasmic                       | 3                  | 1    | 0.6241     |
| 4038             | ND1  | A to G      | homoplasmic                       | 2                  | 0    | 0.4994     |
| 4047             | ND1  | T to C      | homoplasmic                       | 1                  | 0    | 1          |
| 4048             | ND1  | G to A      | homoplasmic                       | 20                 | 2    | 9.93e-05   |
| 4071             | ND1  | C to T      | homoplasmic                       | 24                 | 2    | 7.77e-06   |
| 4083             | ND1  | T to C      | homoplasmic                       | 1                  | 0    | 1          |
| 4086             | ND1  | C to T      | homoplasmic                       | 19                 | 1    | 3.29e-05   |
| 4092             | ND1  | G to A      | homoplasmic                       | 1                  | 0    | 1          |
| 4093             | ND1  | A to G      | homoplasmic                       | 1                  | 0    | 1          |
| 4104             | ND1  | A to G      | homoplasmic                       | 1                  | 0    | 1          |
| 4113             | ND1  | G to A      | homoplasmic                       | 1                  | 0    | 1          |
|    |   |    |                  |     |   |     |
|----|---|---|-----------------|-----|---|-----|
| 4116 | ND1 | C to T | homoplasmic | 1 | 0 | 1   |
| 4117 | ND1 | T to C | homoplasmic | 1 | 0 | 1   |
| 4129 | ND1 | A to G | homoplasmic | 3 | 0 | 0.2491 |
| 4131 | ND1 | A to T | heteroplasmic | 1 | 0 | 1   |
| 4134 | ND1 | A to T | heteroplasmic | 1 | 0 | 1   |
| 4135 | ND1 | T to C | homoplasmic | 1 | 0 | 1   |
| 4136 | ND1 | A to G | homoplasmic | 1 | 0 | 1   |
| 4140 | ND1 | C to T | homoplasmic | 1 | 0 | 1   |
| 4161 | ND1 | C to T | homoplasmic | 1 | 0 | 1   |
| 4164 | ND1 | A to G | homoplasmic | 14 | 1 | 8.83e-4 |
| 4170 | ND1 | C to T | homoplasmic | 1 | 0 | 1   |
| 4176 | ND1 | A to G | homoplasmic | 2 | 0 | 0.4994 |
| 4200 | ND1 | A to T | heteroplasmic | 2 | 0 | 0.4994 |
| 4203 | ND1 | A to G | homoplasmic | 4 | 0 | 0.1241 |
| 4216 | ND1 | T to C | homoplasmic | 4 | 0 | 0.1241 |
| 4227 | ND1 | A to G | homoplasmic | 1 | 0 | 1   |
| 4233 | ND1 | T to C | homoplasmic | 1 | 0 | 1   |
| 4245 | ND1 | C to T | homoplasmic | 2 | 0 | 0.4994 |
| 4248 | ND1 | T to C | homoplasmic | 39 | 2 | 3.25e-10 |
| 4254 | ND1 | T to C | homoplasmic | 1 | 0 | 1   |
| 4314 | tRNA^{Ile} | T to C | homoplasmic | 4 | 0 | 0.1241 |
| 4317 | tRNA^{Ile} | A to G | homoplasmic | 2 | 0 | 0.4994 |
| 4336 | tRNA^{Gln} | T to C | homoplasmic | 8 | 0 | 0.0075 |
| 4386 | tRNA^{Gln} | T to C | homoplasmic | 10 | 0 | 0.0018 |
| 4401 | tRNA^{Met} | A to G | homoplasmic | 1 | 0 | 1   |
| 4435 | tRNA^{Met} | A to G | homoplasmic | 1 | 0 | 1   |
| 4452 | tRNA^{Met} | T to C | homoplasmic | 1 | 0 | 1   |
| 4457 | tRNA^{Met} | C to A | heteroplasmic | 1 | 0 | 1   |
| 4491 | ND2 | G to A | homoplasmic | 4 | 1 | 0.3734 |
| 4515 | ND2 | G to C | heteroplasmic | 1 | 0 | 1   |
| 4535 | ND2 | A to C | heteroplasmic | 2 | 0 | 0.4994 |
| 4562 | ND2 | A to G | homoplasmic | 2 | 0 | 0.4994 |
| 4563 | ND2 | G to T | heteroplasmic | 1 | 0 | 1   |
| 4563 | ND2 | delG | heteroplasmic | 1 | 0 | 1   |
| 4574 | ND2 | A to T | heteroplasmic | 1 | 0 | 1   |
| 4576 | ND2 | delA | heteroplasmic | 1 | 0 | 1   |
| 4580 | ND2 | G to A | homoplasmic | 1 | 0 | 1   |
| 4611 | ND2 | delA | heteroplasmic | 10 | 0 | 0.0018 |
| 4612 | ND2 | delT | heteroplasmic | 2 | 0 | 0.4994 |

*aH/B/M/X means human/bovine/mouse/xenopus.*

bSee [http://www.mitomap.org](http://www.mitomap.org) and [http://www.genpat.uu.se/mtDB/](http://www.genpat.uu.se/mtDB/).

**Figures**

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Figure 1

Distribution histogram of the number of variants in mtDNA 3777-4679 bp among 5 genes.

mutant

Wild type

mutant

m.3970C>T

m.4248T>C
Figure 2

a. Identification of the m.3970C>T mutation in the mitochondrial ND1 gene. Arrow indicates
the position of the gene mutation. b. Identification of the m.4248T>C mutation in the
mitochondrial ND1 gene. Arrow indicates the position of the gene mutation. c. Identification
of the m.4563 delG mutation. Arrow indicates the position of the deletion mutation. d.
Identification of the m.4611delA mutations. Arrow indicates the position of the deletion
mutation.