Mast cells are important modifiers of autoimmune disease: with so much evidence, why is there still controversy?

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INTRODUCTION
Autoimmune and allergic diseases share fundamentally important features in that both are the result of “hypersensitive” immune responses directed toward inherently harmless antigens. It is this overzealous response that causes the pathology. In fact, as late as the 1960s many disease models that we now know are autoimmune were labeled as “experimental allergic” neuritis, encephalomyelitis, and orchitis, uveitis, and glomerulonephritis, reflecting this concept were labeled as “experimental allergic” neuritis, encephalomyelitis, and orchitis, uveitis, and glomerulonephritis, reflecting this concept. Similar to their role as first responders in infection settings, mast cells likely orchestrate early recruitment of immune cells, including neutrophils, to the sites of autoimmune destruction. This co-localization promotes cellular crosstalk and activation and results in the amplification of the local inflammatory response thereby promoting and sustaining tissue damage. Despite the evidence, there is still a debate regarding the relative role of mast cells in these processes. However, by definition, mast cells can only act as accessory cells to the self-reactive T and/or antibody driven autoimmune responses. Thus, when evaluating mast cell involvement using existing and somewhat imperfect animal models of disease, their importance is sometimes obscured. However, these potent immune cells are undoubtedly major contributors to autoimmunity and should be considered as important targets for therapeutic disease intervention.

Keywords: mast cells, autoimmunity, neutrophils, MC-T cell crosstalk, mast cell deficient mice

There is abundant evidence that mast cells are active participants in events that mediate tissue damage in autoimmune disease. Disease-associated increases in mast cell numbers accompanied by mast cell degranulation and elaboration of numerous mast cell mediators at sites of inflammation are commonly observed in many human autoimmune diseases including multiple sclerosis, rheumatoid arthritis, and bullous pemphigoid. In animal models, treatment with mast cell stabilizing drugs or mast cell ablation can result in diminished disease. A variety of receptors including those engaged by antibody, complement, pathogens, and intrinsic danger signals are implicated in mast cell activation in disease. Similar to their role as first responders in infection settings, mast cells likely orchestrate early recruitment of immune cells, including neutrophils, to the sites of autoimmune destruction. This co-localization promotes cellular crosstalk and activation and results in the amplification of the local inflammatory response thereby promoting and sustaining tissue damage. Despite the evidence, there is still a debate regarding the relative role of mast cells in these processes. However, by definition, mast cells can only act as accessory cells to the self-reactive T and/or antibody driven autoimmune responses. Thus, when evaluating mast cell involvement using existing and somewhat imperfect animal models of disease, their importance is sometimes obscured. However, these potent immune cells are undoubtedly major contributors to autoimmunity and should be considered as important targets for therapeutic disease intervention.

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FIGURE 1 | Mast cells amplify autoimmune responses through multiple mechanisms. (1) Naïve autoreactive B and T cells are activated in the secondary lymphoid organs via antigen presentation by APCs. (2) B and T cells undergo differentiation and proliferation leading to a large pool of autoreactive effector cells. Plasma cells secrete self-reactive IgG and/or IgE antibodies. (3) T cells and antibody direct the immune response to the target tissue by entering the bloodstream and trafficking to sites of relevant autoantigen expression. (4) Mast cells are activated early in disease either through TLR/NLR engagement or T cell-mast cell crosstalk via cell–cell contact or soluble mediators. FcR and C3aR/C5aR engagement on mast cells may also play a role. The possibility of activation through cross-talk with other cell types has not been verified. (5) T cells are reactivated through antigen encounter within the target tissue or through cross-talk with mast cells and other resident immune cells. (6) The subsequent release of mediators by mast cells and T cells results in increased vascular permeability and in the recruitment of other inflammatory cells, including neutrophils. (7) The resulting increasing cascade of mediators released by the cells that have co-localized at target tissue sites contribute to amplified inflammation that can directly or indirectly sustain tissue damage.

It is this secondary, albeit critical, role of innate cells that often makes it difficult to definitively determine their relative contribution because of the overlaps in immune cell function. The reliance on inherently imperfect animal models of human disease to define the interplay between adaptive and innate immune cells exacerbates this problem. The spontaneous or actively induced development of disease in animals is rarely physiologic and is often associated with an overly robust autoreactive B or T cell response, one that does not recapitulate the gradual inflammatory processes that characterize the development of most chronic human autoimmune illnesses. Thus, the contribution of modulatory cells can be masked, especially if they exert their critical effects early in disease. Such limitations have sometimes led to inconsistent results and controversy in the field about which cells make substantial contributions to disease pathogenesis. The challenge in convincingly implicating such accessory cells is perhaps best illustrated by the studies of mast cells in autoimmunity. Despite a wealth of supportive evidence, the contribution of mast cells to autoimmunity, particularly to rheumatoid arthritis (RA) and multiple sclerosis (MS), is still debated.

MAST CELLS: POTENT IMMUNOMODULATORY CELLS

Mast cells are c-kit+ FcεRI+ granular cells of the innate immune system that reside in most tissues, but do not circulate in a mature form in the blood. They are most prevalent at sites that interface with the external environment and are often found in close association with blood vessels and nerves (reviewed in Rao and Brown, 2008). Depending on the mode of activation, mast cells can release unique arrays of immune modulating molecules that exist preformed in granules within minutes. Activation also induces the new synthesis of cell surface associated and soluble immune mediators that can exert prolonged effects on a response (Galli et al., 2005; Metz et al., 2007). Although cross-linking of the high affinity IgE receptor (FcεRI) is the best studied mode of mast cell stimulation, the last few years have been transformative in terms of identifying a range of other mast cell activating receptors that act in this capacity leading to either protective or pathologic outcomes. Indeed, there are few infection or disease states in which these potent immunomodulatory cells have not been directly or indirectly implicated (Rao and Brown, 2008) and thus it follows that mast cells are most certainly modifiers of autoimmune disease as well.

MAST CELLS AS MODIFIERS OF AUTOIMMUNITY I: CORRELATIVE EVIDENCE

The idea that mast cells are involved in the initiation and sustaining events of autoimmunity is not based on conventional wisdom, but rather on substantial data from studies of both human disease and animal models (Benoist and Mathis, 2002; Lee et al., 2002; Gregory and Brown, 2006; Rao and Brown, 2008; Sayed et al., 2008; Walker et al., 2011). Here we focus primarily on studies of RA, bullous pemphigoid (BP), type I diabetes and multiple...
sclerosis (MS), where the evidence for a mast cell contribution is particularly convincing.

In RA the primary targets of the autoantibody-directed immune response are the synovial joints (Lee and Weinblatt, 2001; Lee et al., 2002). Although mast cells are normal residents within the synovium, their numbers are increased in human disease and this is associated with the local production of several mast cell-derived mediators (reviewed in Nigrovic and Lee, 2007). Tryptase, a preformed mast cell-specific protease, is thought to initiate the inflammatory response by complexing with heparin to induce the release of neutrophil chemotactic factors such as TNF, IL-1β, and IL-17, as well as IL-33 by synovial fibroblasts (Shin et al., 2009; Hueber et al., 2010). IL-33, in turn, acts directly on mast cells to initiate the expression of additional inflammatory cytokines and chemokines (Xu et al., 2008, 2010; Verri et al., 2010). Tryptase can also directly activate synovial fibroblasts through its interaction with the protease-activated receptor 2 (PAR2) enhancing their ability to express proteases that degrade cartilage and bone (Palmer et al., 2007; Sawamukai et al., 2010). In human RA, a 6- to 25-fold expansion of mast cells has been observed in affected joints when compared to numbers in normal joints (Nigrovic and Lee, 2007). Mast cell numbers also increase more than threefold in multiple animal models of RA (Aloe et al., 1993; Kakizoe et al., 1999; Shin et al., 2006) and it has been postulated that they are essential for the joint-specific vascular leak phenomenon involving both histamine and serotonin (Binstadt et al., 2006). In collagen-induced arthritis, increased mast cell numbers and the associated neutrophil influx to the joints as well as exacerbated clinical severity are reduced by treatment with the mast cell stabilizer, sodium nedocromil (Pimentel et al., 2011).

Bullous pemphigoid is a skin blistering disease that can be induced in mice by injection of an antibody (anti-BP180) directed to hemidesmosomes, structures involved in cell–cell adherence (Kasperkiewicz and Zillikens, 2007). Early events in this model of disease include massive degranulation of skin mast cells that precede the characteristic neutrophil influx and blister formation. Cromolyne sodium, another mast cell stabilizing drug, blocks these events (Chen et al., 2001; Navi et al., 2007). In human disease, degranulated mast cells are prominent in lesions where histamine, tryptase, and several mast cell-associated chemokines are detected in blister fluid (Baba et al., 1976; Dvorak et al., 1982; Katayama et al., 1984; D’Auria et al., 2000).

In Type 1 diabetes, CD8+ T cells are thought to play a predominant role in the destruction of the insulin-producing islet cells of the pancreas although CD4+ T cells and antibodies are also implicated. The resulting loss of blood glucose regulation can lead to retinopathy, nephropathy, cardiovascular disease, and neuropathy (Bluestone et al., 2010). Gene expression profiling of the pancreatic lymph nodes in the spontaneous BB rat model revealed activated mast cell transcripts that correlate with disease and treatment with cromolyn sodium, results in a significant delay in disease onset (Geoffrey et al., 2006). Inhibitors of tyrosine kinase can also inhibit and reverse spontaneous disease in the NOD mouse model of Type 1 diabetes, however, specific inhibitors of c-kit, a tyrosine kinase-associated receptor required for mast cell development, have marginal effects on limiting disease progression (Louve et al., 2008). Paradoxically, a protective role for mast cells in this disease has been proposed based on results of studies in a model of spontaneous diabetes in NOD mice. Treatment of NOD mice with anti-FceRI antibodies, which activate mast cells and basophils to produce IL-4, delays diabetes (Hubner et al., 2011). It is postulated that in this model system IL-4 is exerting anti-inflammatory effects by skewing CD4+ T helper cells to a Th2 dominated response.

Multiple sclerosis is a CD4+ T cell-dependent disease in which myelinated nerves in the relatively immunologically sequestered CNS are targeted for destruction leading to variable neurologic dysfunction (Sospedra and Martin, 2005; Hauser and Oksenberg, 2006). Mast cells are observed in plaques of patients assessed post-mortem as are transcripts encoding tryptase, histamine, and FceRI, identified by microarray analyses (Lock et al., 2002). Tryptase and histamine are also detected in the cerebral spinal fluid of some patients (Tuomisto et al., 1983; Rozniecki et al., 1995). In experimental autoimmune encephalomyelitis (EAE), a mouse model of MS induced by immunization with myelin peptides in CFA, treatment of mice with proxicromil an inhibitor of mast cell degranulation, ciproheptadine, a serotonin receptor antagonist or reserpine, which depletes mast cell granules, inhibits disease (Dietzsch and Hinrichs, 1989; Dimitriadou et al., 2000).

MAST CELLS AS MODIFIERS OF AUTOIMMUNITY II: CONFOUNDING EVIDENCE FROM IN VIVO STUDIES OF MAST CELL DEFICIENT MICE

Although compelling, measures of increased mast cell numbers, mast cell activation profiles, or efficacy of mast cell-inhibitory drugs are not proof of mast cell involvement. The most definitive studies implicating mast cells have relied on the use of mast cell deficient mice, most commonly those with mutations in Kit, the gene encoding c-kit, the stem cell factor (SCF) receptor (W/W\(^\text{sh}\), W\(^{-\text{sh}}\) mice) or in Kit\(\varepsilon\), the gene encoding the c-kit ligand, SCF (Sl/Sl mice). The differentiation and long term survival of mast cells is dependent on sustained high level SCF/c-kit signaling and mice with W/W\(^\text{W}\), W\(^{-\text{sh}}\)/W\(^{-\text{sh}}\), or Sl/Sl mutations that compromise this signaling exhibit defects in mast cell development (Galli and Kitamura, 1987; Grimbaldeston et al., 2005). These mice are imperfect models because they also bear other hematologic and non-hematologic defects including neutropenia, anemia, pigment production defects, reduced intestinal mobility and fertility (W/W\(^\text{W}\)), or time-dependent loss of mast cells, cardiac abnormalities, and neutropenia (W\(^{-\text{sh}}\)/W\(^{-\text{sh}}\); Galli and Kitamura, 1987; Grimbaldeston et al., 2005; Nigrovic et al., 2008). Sl/Sl mice also develop lymphocytic leukemia at a high rate and have shortened life spans (Galli and Kitamura, 1987). Systemic or local reconstitution of Kit mutant mice with bone marrow derived mast cells (BMMC) selectively restores most mast cell populations, although not always to wild type levels, but does not correct other mutant Kit related abnormalities. If a wild type phenotype is re-established in Kit mutant mice by selective repopulation of tissue mast cells, it is generally accepted that mast cells are implicated.

Several recent reports have described the generation of unique strains of mast cell deficient mice that do not appear to have the accompanying profound hematologic defects of Kit mutant mice, although their characterization is still incomplete (Dudeck et al., 2011; Feyerabend et al., 2011; Lilia et al., 2011; Otsuka et al., 2011). These mice provide the promise that a more unequivocal assessment of mast cell contributions in vivo can be achieved.
Of particular interest to this discussion of autoimmune disease are the Cpa3<sup>Cre<sup>+</sup></sup> mice. Carboxypeptidase 3 (Cpa3) is a mast cell specific protease expressed early in development and Cpa3<sup>Cre<sup>+</sup></sup> mice contain a transgene encoding cre-recombine under the control of the carboxypeptidase 3 (Cpa3) promoter (Feyerabend et al., 2011). It is presumed that the high level expression of Cre-recombine in mast cell precursors that express Cpa3 is toxic, causing genomic instability and the ultimate demise of this cell lineage at an early stage in development. With the exception of a small diminution in basophil numbers, no other cell lineages appear to be affected. Below, we review the controversial evidence that supports and refutes a role for mast cells in autoimmunity using mast cell deficient mice.

### EVIDENCE FOR MAST CELL INFLUENCE IN AUTOIMMUNITY USING Kit<sup>W/Wv</sup> MICE

Results in Kit<sup>W/Wv</sup> mice have provided the most consistent supportive evidence for mast cell involvement in autoimmunity. Of note, in most autoimmune disease models examined using these mice, including RA, BP, and EAE, the mast cell-dependent neutrophil recruitment discussed above is an important pathogenic event, one that is absent in Kit<sup>W/Wv</sup> mice (D’Auria et al., 2000; Chen et al., 2006; Kasperkiewicz and Zillikens, 2007; Monach et al., 2010; Sayed et al., 2010; Kaplan, 2011; Walker et al., 2011; Warde, 2011). Despite the inherent neutropenia of Kit<sup>W/Wv</sup> mice, restoration of mast cells results in normal disease-related neutrophil influx to inflamed tissues indicating that the few neutrophils present in these mice are mobilized in a mast cell-dependent manner. The relationship of mast cells and neutrophils was first defined in infection settings where mast cells, acting as first responders, release multiple mediators including histamine, PAF, TNF, CXCL2, and IL-8 within minutes of activation and initiate the influx of neutrophils (Zhang et al., 1992; Malaviya et al., 1996a; Biedermann et al., 2000). As in infection, in autoimmune disease, neutrophils likely promote disease-associated changes in vascular permeability, but also contribute directly to self-tissue damage through the release of proteases, reactive oxygen intermediates, and chemotactants that regulate the influx of other inflammatory cells, contributing to a cascade of damaging events (reviewed in Kumar and Sharma, 2010; Mantovani et al., 2011).

Mast cell dependence has been demonstrated in models of RA (K/BxN serum transfer-induced and antigen-induced RA) using (WB xC57BL/6)F1 (WBB6) Kit<sup>W/Wv</sup> mice where, as in human RA, neutrophils are the dominant lineage in synovial effusions (Negovic and Lee, 2007). K/BxN mice exhibit an early and spontaneous onset of RA-like symptoms due to the presence of glucose-6-phosphate reactive autoantibodies and transfer of K/BxN serum into wild type but not WBB6 Kit<sup>W/Wv</sup> mice, induces disease (Corr and Crain, 2002; Lee et al., 2002). Disease susceptibility is restored to Kit<sup>−/−</sup> mutant mice by BMMC reconstitution. Mast cell reconstitution with IL-1β<sup>−/−</sup> BMMCs fails to restore disease susceptibility in the K/BxN serum transfer model, indicating that disease pathogenesis is mediated by mast cell-derived IL-1β (Negovic et al., 2007). Mast cells also directly modulate access of arthritogenic antibodies to the joint in the hyperacute phase of K/BxN serum transfer arthritis, a feature likely directly or indirectly related to the effects of mast cells on vascular permeability. Labeled antibodies were specifically deposited in the joint of wild type mice within minutes after transfer, while deposition was delayed and reduced in W/W<sup>+</sup> animals (Wipke et al., 2002, 2004). In antigen-induced arthritis, a chronic arthritis model induced by the direct injection of methylated bovine serum albumen into the joint, there is little difference in early disease severity in wild type and Kit<sup>W/Wv</sup> mice, but mast cell deficient mice develop significantly reduced cartilage damage over time, a phenotype reversed by BMMC reconstitution (van den Broek et al., 1988). However, in the collagen-induced arthritis model, there are no clinical disease differences leading the authors to suggest that either this model is mast cell-independent or that mast cells have a minor role (Pitman et al., 2011).

In the BP model, WBB6 Kit<sup>W/Wv</sup> mice exhibit reduced neutrophil accumulation in the skin and diminished skin blistering upon transfer of serum autoantibodies compared to wild type mice, whereas a wild type-like disease with associated neutrophil influx is restored upon mast cell reconstitution (Chen et al., 2002). Notably, administration of IL-8, a neutrophil chemoattractant, can overcome the mast cell-deficiency (Chen et al., 2001). Evidence of mast cell involvement in EAE pathogenesis using Kit<sup>W/Wv</sup> mice has been less consistent. EAE is induced by immunization of mice with myelin peptide in CFA (Mix et al., 2010). Original studies of WBB6 Kit<sup>W/Wv</sup> mice and their wild type littermates in a chronic model of MS induced by myelin oligodendrocyte glycoprotein35–55 peptide (MOG)35–55 immunization revealed that the absence of mast cells does not abolish disease, but there is significantly reduced severity (Secor et al., 2000). This susceptibility is restored to wild type levels by BMMC reconstitution, a procedure that does not correct the c-kit-related anemia in these mice. Mast cells that normally reside in the meninges are essential for this fulminant disease and act to regulate BBB integrity and inflammatory cell entry into the CNS (Sayed et al., 2010). Neutrophils are among the first cells to enter the CNS in EAE and their requirement for disease pathogenesis was confirmed by studies of mice lacking the chemokine receptor, CXCR2, expressed on neutrophils. CXCR2<sup>−/−</sup> mice cannot mobilize neutrophils from the bone marrow in response to inflammatory signals and fail to develop EAE (Carlson et al., 2008). Mast cells and mast cell-derived TNF are required for neutrophil influx into the CNS (Sayed et al., 2010). Despite the convincing data in mouse models, neutrophil involvement in MS is still controversial. Neutrophils are short-lived cells and likely exert effects during the initiation of disease or in flares associated with relapsing-remitting disease. These cells are not detected in post-mortem assessment of the meninges or CNS as expected, leading many to dismiss them as participants in MS pathogenesis.

Severe PLP<sub>131–159</sub>-induced disease in SJL mice is also mast cell-dependent. This model is considered by some to be more relevant because it mimics the most commonly presenting relapsing-remitting form of human disease, one observed in at least 85% of patients. SJL Kit<sup>W/Wv</sup> mice exhibit a significantly reduced disease course that corresponds to decreased infiltration of inflammatory cells to the CNS and mast cell reconstitution restores wild type disease susceptibility (Sayed et al., 2011). While there has been independent corroboration that mast cells are important in disease by other groups using Kit<sup>W/Wv</sup> mice (Stelekati et al., 2009; Piconese et al., 2011), other studies report no disease differences
The basis of this disparity is still not clear, but there are several potential explanations including age-related differences in susceptibility, the use of distinct disease induction protocols that result in overwhelming T cell responses and unique housing- or diet-related gut microbiota that alter the general immune responsiveness of the mice.

STUDIES USING KitW−/−sh/W−/− MICE FAIL TO REVEAL A MAST CELL INFLUENCE IN RA AND EAE

C57BL/6 KitW−/−sh/W−/− mice were originally touted as better subjects for in vivo studies of mast cell-dependent events because they are on a pure C57BL/6 background and are fertile (Grimbaldeston et al., 2005). However, contradicting the results of studies in KitW+/W+ mice, there is no decrease in EAE or RA disease severity in KitW−/−sh/W−/− suggesting mast cells are not important (Zhou et al., 2007; Piconese et al., 2011). These results may be explained by the fact that unlike neutropenic KitW+/W+ mice, KitW−/−sh/W−/− mice have a baseline pro-inflammatory phenotype that includes neutrophilia, splenomegaly, and increased mast cell precursors in the spleen. Thus, their pre-existing inflammatory state, particularly the neutrophilia, likely obviates the need for the mast cell-dependent neutrophil recruitment that affects early disease-promoting events.

STUDIES USING Cpa3cre/+ MICE FAIL TO IMPLICATE MAST CELLS IN RA AND EAE

A recent study by Feyerabend et al. (2011) using the newly described Cpa3cre/+ mice also fails to corroborate a role for mast cells in EAE and RA. Cpa3cre/+ mice are refractory to induction of anaphylaxis and fail to exhibit the normal protective expansion of intestinal mast cells associated with N. Brasiliensis infection supporting the validity of using these mice in models in which IgE-activated mast cells are the major effector cells. Yet Cpa3cre/+ mice are susceptible to RA induction by transfer of K/BxN serum, in contrast to KitW+/W+ mice. In addition, no diminution of EAE disease severity was observed in either Cpa3cre/+ or KitW+/W+ mice when compared to their wild type littermate controls. Two conclusions were made based on these results: (1) mast cells are dispensable in both models of RA and MS (EAE) and, (2) other c-kit-regulated events influence the susceptibility of mice to RA. A corollary to these conclusions is that mast cells are only relevant in IgE-dependent allergic disease (Katz and Austen, 2011).

Yet these conclusions may be premature for a number of reasons. For example, mast cells express a myriad of other activating receptors that have been documented to play important physiologic roles in many inflammatory responses outside allergy (Rao and Brown, 2008; Sayed et al., 2008). In addition, as discussed earlier mast cells are not the major effector cells in T and/or B cell-dependent autoimmune responses as they are in IgE-dependent responses, thus they are relegated to accessory roles. The strong disease induction conditions used in the EAE experiments in this study (corroborated by high morbidity in all groups Feyerabend et al., 2011) make it likely that a vigorous T cell response masks the mast cell contribution. There is abundant evidence supporting mast cell contributions to autoimmune inflammation and it is important to delineate the underlying reasons for this conflicting data.

 POTENTIAL MECHANISMS OF MAST CELL ACTIVATION IN AUTOIMMUNE DISEASE

There are a multitude of receptors expressed on mast cells that are candidates for involvement in autoimmune modulation (Figure 2). As discussed below, many of these receptors and their ligands have been implicated in autoimmune disease, but whether they are acting to activate mast cells in the context of disease is still in question.

PATHOGEN- AND DANGER-ASSOCIATED MOLECULAR PATTERN MOLECULES IN INFECTION-ASSOCIATED AUTOIMMUNITY: OVERCOMING TOLERANCE

It is well established that the activation and expansion of self-reactive T and B cells can occur only under defined conditions: either the normal mechanisms of negative selection fail to delete particular self-reactive clones during development, there is a failure of normal peripheral mechanisms that maintain tolerance to self, the normal sequestration of antigens that prevent lymphocyte access to self antigens is breached or infection with certain pathogens elicits an adaptive immune response that cross-reacts with self antigens, a phenomenon termed molecular mimicry (Nurieva et al., 2011; Pillai et al., 2011).

Although not definitively established in humans, the development of several autoimmune diseases has been linked to exposure to infectious agents (Kivity et al., 2009). For example, prior EBV infections are postulated to increase the predisposition to Systemic lupus erythematosus (SLE), RA, MS, Sjogren’s syndrome, and polyarteritis nodosa (Barzilai et al., 2007), while autoimmune diabetes susceptibility is associated with enterovirus infections (Richardson et al., 2009). Bacterial infections including H. pylori are implicated in susceptibility to autoimmune gastritis and Sjogrens syndrome (Annibale et al., 2001; D’Elios et al., 2004). In addition to eliciting a cross-reactive pathogen-specific adaptive immune response, infectious agents can engage a class of pathogen- and danger-associated receptors on both adaptive and innate immune cells, including mast cells. These receptors, including Toll-like receptors (TLRs) and Nod-like receptors (NLRs), are expressed either within the plasma membrane or in endosomal compartments and recognize conserved components of bacteria, viruses, and protozoa (such as lipopolysaccharide, LPS), double stranded RNA, flagellin, and non-methylated CpG DNA, collectively termed pathogen-associated molecular patterns (PAMPs; reviewed in Kawai and Akira, 2011). Activation of immune cells through TLRs and NLRs induces the expression of multiple inflammatory mediators that can ultimately result in local tissue damage causing the release of normally sequestered tissue antigen. Subsequent recognition of these antigens by T or B cells will induce activation and initiate autoimmunity. Alternatively, infection-induced activation of accessory immune cells, including mast cells, macrophages, and neutrophils, can boost inflammation and transform a relatively modest autoreactive response into one that causes detectable pathology.

Toll-like receptors/NLRs are also highly effective at sensing and responding to tissue injury, such as that induced in autoimmunity (Zhang et al., 2010; Kufer and Sansonetti, 2011). Just as pathogens are detected by virtue of “Stranger” signals, damaged cells release “Danger” signals, designated as alarmins or Damage Associated Molecular Patterns (DAMPs) that are also sensed by
TLRs and NLRs (reviewed in Bianchi, 2007; Piccinini and Midwood, 2010; Marek and Kagan, 2012). In mammals, TLR3, TLR7, and TLR9 detect microbial nucleic acids, as well as those released from necrotic cells. For example, damaged cells release heat shock proteins such as Hsp60, Hsp70, and alpha-β crystalline that are recognized by TLR2 and TLR4 as well as endogenous cellular products such as uric acid crystals and aggregated peptides that activate NLRs. ATP release from damaged cells acts through purinergic receptor-regulated channels to cause cytosolic ion fluxes that are detected by NLRs. The resulting signals generated through such cell injury can activate immune cells, contributing to the persistence of the inflammatory response and promoting more tissue destruction.

There is emerging evidence that TLRs/NLRs are important receptors in autoimmune diseases (Fischer and Ehlers, 2008; Mills, 2011). Activation through TLR2, 3, 4, and 9 is associated with disease in animal models of RA (Ronaghy et al., 2002; Abdollahi-Roodsaz et al., 2008; Hoffmann et al., 2011; Goh and Midwood, 2012) and TLR4 engagement can overcome the requirement for IL-1β in the serum transfer model of this disease (Choe et al., 2003). Similarly, TLR1, 2, 3, and 7 signals mediate islet inflammation in rat and mouse models of type 1 diabetes (Lien and Zipris, 2009). TLR1, 2, 3, and 4 are expressed in the salivary glands of patients with Sjögren’s syndrome and when engaged can trigger the expression of a number of activation molecules, including CD40 and ICAM-1, on salivary gland epithelial cells (Spachidou et al., 2007). TLR2 ligation specifically activates Th17 cytokine production in the peripheral blood mononuclear cells of Sjögren’s syndrome patients (Kwok et al., 2012). Based on studies in MS models where TLR2, 4, 7, and 9 have been implicated in exacerbated disease (Marta et al., 2009) and TLR3 is protective (Touil et al., 2006), the targeting of these molecules for therapy has been proposed (Gambuzza et al., 2011).

With the exception of TLR5, there is evidence that most other TLRs are expressed on mast cells and are functional (Marshall et al., 2003a, b; Marshall, 2004). For example, activation of mature mast cells through TLR2 results in their production of several pro-inflammatory cytokines critical in autoimmune including IL-17, IFNγ, TNF, and IL-1β (Mrabet-Dahbi et al., 2009). Mast cells also express the NLR, NLRP3, and activation of this intracellular receptor activates the inflammasome, a multiprotein complex that regulates the processing and secretion of the IL-1 family of cytokines including IL-1β and IL-18 through the activation of caspase 1 (Nakamura et al., 2009). Mast cells clearly are capable of responding to PAMP and DAMP signals and it will be critical to determine whether this is an important mode of activation in autoimmune disease.
**FcεRI AND FcγR ACTIVATION OF MAST CELLS: COMMON MECHANISMS LINKING AUTOIMMUNITY AND ALLERGY**

The connection between IgE- and IgG-mediated mast cell responses and autoimmunity is still a tenuous one and is just beginning to gain some traction (Valenta et al., 2009). As discussed earlier, allergy and autoimmunity are both considered hypersensitive diseases that result from inappropriate responses to innocuous antigens. However, it is notable that self-reactive IgG and IgE antibodies are commonly detected in both allergic and autoimmune diseases including atopic dermatitis, RA, systemic lupus erythematosus, Hashimoto’s thyroiditis, Graves’ disease, BP and MS, and their animal models (Permin and Wiik, 1978; Bartholomew et al., 1991; Valenta et al., 1991; Cooper et al., 1993; Ghohestani et al., 1998; Pedotti et al., 2001; Metcalfe et al., 2002; Concha et al., 2004; Smith et al., 2005; Fairley et al., 2007). In atopy, these autoreactive antibodies are primarily detected in those individuals with severe and chronic diseases such as atopic dermatitis, but not in patients with mild respiratory allergies (Valenta et al., 1991). Many of the self antigens that react with IgE are homologs of environmental antigens, including the actin binding protein, profilin, serum albumin, collagen and desmoplakin, and responses to these are thought to result from cross-reactivity (Patterson et al., 1986; Bartholomew et al., 1991; Valenta et al., 1991; Cooper et al., 1993; Spitzauer et al., 1994; Ghohestani et al., 1998). Alternatively there is speculation that inflammation associated with IgE-dependent mast cell responses to environmental allergens induces tissue damage and in a Th2 dominated immune environment these self antigens, no longer sequestered from the immune system, elicit an IgE response (Valenta et al., 2009).

Mast cells also express multiple IgG Fc receptors. Important participants in protective immune responses to pathogens, IgG receptor cross-linking elicits mast cell activation in allergy and also plays critical roles in autoimmunity. In fact, IgG autoantibodies are hallmarks of many autoimmune diseases and have been detected in BP, RA, SLE, and MS (reviewed in Sayed et al., 2008). IgG-specific Fc receptors include both the activating forms, FcγRI and FcγRIII, as well as the inhibitory FcγRIIB. FcγRIIB engagement down regulates both FcγRII- and FcγRII-induced mast cell mediator release (Miyajima et al., 1997) and the loss of this inhibitory receptor renders mice susceptible to induced autoimmune arthritis and spontaneous SLE and leads to exacerbated EAE (Abdul-Majid et al., 2002; Lock et al., 2002; Pedotti et al., 2003; Robbie-Ryan et al., 2003). Kit^W/W^ mice reconstituted with either FcγRIIa^−/−^ mast cells (which lack functional FcγRII, FcεRI, and FcγRI) or with mast cells lacking only FcγRII exhibit a reduced and delayed EAE disease course, suggesting that activation of mast cells through these receptors modulates disease (Robbie-Ryan et al., 2003). Mast cell-derived IL-1β, which is necessary for disease in the K/BxN model, is produced after stimulation through autoantibody engagement of FcγRII (Nigrovic et al., 2007).

Clearly such findings, particularly the presence of self-reactive IgE antibodies in both allergy and autoimmunity, blur the distinction between these classes of disease and suggest that allergy and autoimmunity share similar antibody-dependent mechanisms to activate mast cells and elicit tissue damage in disease.

**Complement**

The complement system, named for its ability to complement the action of antibodies and phagocytes in an immune response, consists of several proteins synthesized by the liver that exist as inactive pro-proteins. Under inflammatory conditions, a proteolytic cascade is initiated that results in the cleavage and generation of active components, some of which act as direct chemoattractants or bind to specific G protein-coupled receptors on cells and stimulate their activation (reviewed in Ricklin et al., 2010). Complement activation is triggered by the direct binding of complement proteins to microbial surfaces (alternative pathway) or via complement binding to complement-fixing antibody isotypes that have been cross-linked by antigen (classical pathway). A mannose binding lectin-induced pathway also exists (lectin pathway) and these processes of complement fixation, chemoattraction, and cell activation are essential for protection from infection.

Mast cells have an intimate relationship with complement in that they can activate complement components, but they also express surface receptors that directly bind the breakdown products of this proteolytic cascade. It has been speculated that mast cell degranulation induces release of chemoattractants (e.g., LTβ4), vasoactive amines (e.g., histamine), and proteases (e.g., tryptase; (Dawicki and Marshall, 2007). C3 and C5 complement proteins are recruited through the permeable local vasculature to sites of mast cell residence and are cleaved through the action of mast cell proteases to C3a and C5a (Schwartz et al., 1983; Fukuoka et al., 2008). These byproducts can activate mast cells through the engagement of complement receptors expressed on mast cells, C3aR and C5aR, leading to further chemokine/cytokine production and release (Wyatt et al., 1979; Biesecker et al., 1981; Falk et al., 1983).

Not unexpectedly, the best evidence of complement activation in autoimmunity comes from studies of antibody-mediated diseases such as RA, BP, and SLE. Mice depleted of complement are refractory to collagen-induced arthritis (Backlund et al., 2002). Antibodies specific for double stranded DNA form immune complexes and their deposition in the glomeruli of patients with SLE activates complement, generating C3a and C5a (Wyatt et al., 1979; Biesecker et al., 1981; Falk et al., 1983). Although complement activation normally promotes immune complex clearance, the deposition of complement activation products also contributes to inflammation and local tissue injury (Rothfield and Stoller, 1967; Couser, 1985; Couser et al., 1985; Perkinsson et al., 1985; Mackworth-Young et al., 1986). In BP, COL17-specific autoantibodies target human type XVII collagen (Robson et al., 2003). Access of these antibodies to the basement membrane elicits an inflammatory cascade that includes complement activation, infiltration of neutrophils and eosinophils, and degranulation of dermal mast cells (Oikarinen et al., 1983; Stahle-Backdahl et al., 1994; Chen et al., 2001; Robson et al., 2003; Liu et al., 2008). In a mouse model of BP induced by administration of anti-hCOL17 antibody, altering the capacity of the antibody to fix complement abrogates its disease-causing ability (Li et al., 2010). In SLE, complement activation deposition of complement at sites of tissue damage is often observed, however, deficiencies in the early components of the complement cascade are strongly associated with disease. In animal models of disease, the lack of C1q or C4 results in loss of apoptotic cell clearance, increasing the supply of...
autoantigens that are targets of the immune response and promoting a predisposition to SLE-like disease (Cook and Botto, 2006). In EAE, complement acts in concert with autoantibody to promote demyelination (Uriah et al., 2006). However, while C3, but not C5, is required for disease, the deletion of both C3aR and C5aR has no effect on disease severity, perhaps illustrating another instance where immune redundancy or strong induction of disease in this experimental setting obscures the role for these molecules (Reiman et al., 2002; Boos et al., 2004; Morgan et al., 2004; Ramos et al., 2009).

There is only limited data demonstrating a direct link between complement-activated mast cells and autoimmune disease. For example, in the K/BxN model, C5aR expression on mast cells is necessary for neutrophil recruitment and subsequent disease development (Nigrovic et al., 2010). The development of BP in mice requires the expression of C5aR on mast cells, which is required for their degranulation (Heimbach et al., 2011). In EAE, the data are equivocal. Targeted deletion of receptors for C3a and C5a does not protect mice from disease, yet deletion of C3 convertase regulator complement receptor 1-related protein y (Cryy) exacerbates disease (Reiman et al., 2002; Ramos et al., 2009; Ramaglia et al., 2012). There is also evidence that C5 may actually promote remyelination and prevent gliosis in the CNS (Weerth et al., 2003).

**Immune cell crosstalk**

The concept that cell–cell interactions result in cross-activation has now become a prevailing paradigm in immunology. There are many examples of mast cell interactions with T cells, DCs, and neutrophils, although the consequences of most of these interactions have not yet been ascribed to autoimmune disease pathogenesis. Mast cell activation is likely to be an early and initiating event that triggers cell co-localization and facilitates such interactions, given that many mast cell mediators released early in a response act to recruit cells to sites of inflammation. As alluded to above, mast cell derived TNF, CXCL2, and LTβ4, for example, recruit neutrophils to inflamed tissues (Ramos et al., 1991, 1994; Zhang et al., 1992, 1995; Mercer-Jones et al., 1999; Biedermann et al., 2000; Wang and Thorlacius, 2005; Nakamura et al., 2009). Similarly, mast dependent neutrophil recruitment to affected tissues has been demonstrated in animal models of autoimmunity including the synovium in RA, the skin in BP, and the meninges in EAE (Chen et al., 2001, 2002; Sayed et al., 2010; Pimentel et al., 2011). A subset of these same mediators, particularly TNF and LTβ4 also recruit DCs and T cells while IL-16, lymphotactin (MIP-1α), and MIP-1β are chemotactic for CD4+ and CD8+ T cells (reviewed in Mekori and Metcalfe, 1999). In a model of cutaneous contact hypersensitivity, mast cell activation is essential for DC migration to the skin and histamine and PGE2, both produced by mast cells, induce the expression of Th2-recruiting cytokines by DCs (Mcllroy et al., 2006). Mast cells also affect DC function in a skin allograft model where mast cell derived GM-CSF sustains the DC population that is required for tolerance at the graft site (de Vries et al., 2011). Here we will focus on mast cell interactions with effector T cells and regulatory T cells, as these are arguably the best-described interactions and also the mostly likely to play a significant role in autoimmunity.

There is an abundance of evidence that mast cells affect the activation state and functionality of T cells. Included among the many tissue sites where mast cells reside are the secondary lymphoid organs and a number of studies have demonstrated that their numbers increase in the lymph nodes and spleen in response to some inflammatory signals (Tanzola et al., 2003; Kashiwakura et al., 2004; Sayed et al., 2010). At these sites of T cell priming as well as other tissue sites where T cells are reactivated, bystander mast cells can potentially influence the differentiation phenotype of T cells through their ability to express a myriad of T cell polarizing cytokines including IL-4 and histamine (Mekori and Metcalfe, 1999). Mast cell-derived TNF, in concert with OX40L, enhances T cell activation and proliferation and also contributes to the polarization of Th2 cells in a model of airway hyperreactivity (Nakae et al., 2006, 2007a,b). Mast cells express MHC Class I and inducible MHC Class II molecules and thus have the potential to act as antigen presenting cells (APCs) and there are several reports that substantiate mast cell APC function both in vitro and in vivo (Frandji et al., 1993, 1995; Fox et al., 1994; Malaviya et al., 1996b; Kambayashi et al., 2008, 2009). However, if mast cells serve in this capacity, it is likely that they are more relevant in recall responses and not in the priming of naïve T cells. Given the emerging appreciation of superantigens in allergic pathogenesis, particularly those derived from Staphylococcus (Barnes, 2011), it is tempting to speculate that mast cells “present” superantigens to T cells and initiate their activation. The ability of mast cells to present staphylococcal enterotoxin B to CD8+ T cells supports this idea (Frandji et al., 1996). Through the expression of co-stimulatory molecules such as CD80, CD86, CD40, CD40L, GITR, and 4-1BB as well as the inhibitory molecules PD-L1, PD-L2, and Fas, mast cells can alter T cell effector function (Nakae et al., 2005, 2007; Lu et al., 2006; Stelekati et al., 2007, 2009; Picone et al., 2009; Hershko and Rivera, 2010).

A newer concept is that T cells can activate and alter mast cell function. Many of the studies to support this idea have been performed in vitro using membranes isolated from activated T cells and mechanisms that underlie T cell mediated mast cell activation have not been extensively explored. However, activated T cells induce human mast cells to release MMP-9, an event that is blocked by inhibiting TNF (Baram et al., 2001). Human mast cells also produce the neutrophil chemoattract IL-8 upon interaction with activated, but not resting, T cells (Salamon et al., 2005). Oncostatin M, a multifunctional cytokine of the IL-6 family, is released by human mast cells after interaction with activated T cells, but not after cross-linking through the IgE receptor (Salamon et al., 2008). Two receptor-ligand interactions have been implicated in T cell-driven mast cell activation. Upon engagement of the lymphotixin-β receptor (LTβR), expressed on mast cells, with its ligand, expressed on activated T cells, mast cells release pro-inflammatory cytokines including IL-4, IL-6, TNF, MIP-2, and RANTES (Stopfer et al., 2004). Interaction of LFA-1, expressed on activated T cells, and ICAM-1, expressed on mast cells, induces MC degranulation (Imamura et al., 1998).

Mast cells have a paradoxical relationship with T regulatory (Treg) cells. Most studies that have investigated mast cell-T regulatory cell crosstalk were done using a skin allograft model. In this model Treg cell-derived IL-9 recruits and activates mast cells to the
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