Review Article

Postmyocardial Infarct Remodeling and Heart Failure: Potential Contributions from Pro- and Antiaging Factors

Halliday A. Idikio

Department of Laboratory Medicine and Pathology, University of Alberta, Room 5B4.11 WCM-HSC, 8440-112th Street, Edmonton, AB, Canada T6R 2B7

Correspondence should be addressed to Halliday A. Idikio, hidikio@ualberta.ca

Received 20 November 2010; Revised 14 February 2011; Accepted 14 February 2011

Academic Editor: Gerasimos Filippatos

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Myocardial infarction and adverse postinfarct remodeling in older persons lead to poor outcome and need greater understanding of the contributions of age-related factors on abnormal cardiac function and management. In this perspective, how normal aging processes could contribute to the events of post-myocardial infarction and remodeling is reviewed. Post-myocardial infarction and remodeling involve cardiomechanical factors and neurohormonal response. Many factors prevent or accelerate aging including immunosenescence, recruitment and regeneration of stem cells, telomere shortening, oxidative damage, antiaging hormones klotho and melatonin, nutrition, and Sirtuin protein family, and these factors could affect post-MI remodeling and heart failure. Interest in stem cell repair of myocardial infarcts to mitigate post-MI remodeling needs more information on aging of stem cells, and potential effects on stem cell use in infarct repair. Integrating genomics and proteomics methods may help find clinically novel therapy in the management of post-MI remodeling and heart failure in aged individuals.

1. Introduction

Aging is universal and is governed by many factors, the basis of which many be theoretical. The fundamental argument is whether aging is programmed or simply determined by interactions between environmental and genetic factors [1] (Table 1). Many are searching for human longevity genes but the efforts are complicated or compounded by the role of interdependent environmental factors. The factors suggested include insulin/insulin-like growth factor-1 (IGF-1), and Fox-head transcription factors (FOXOs) [2]. Klotho gene is a recent description of antiaging gene [3, 4]. On the other side the Wnt canonical and alternate signaling pathways are participants in aging mainly in their regulation of stem cell renewal [5]. Moreover, the elderly experience worse heart failure postMI and are major demographic contributors to increase in heart failure burden [6]. The offsprings of centenarians have an advantage as they fare better with age, have less risk of cardiovascular disease and stroke [7]. Centenarian offsprings have longer telomeres and their experience suggests close interaction of environment and cellular repair systems[8]. Model systems have provided a wealth of information on the roles of altered genetic factors on aging [9]. Gene expression changes with aging in mice is available [10] and changes in gene expression in all organs with age are noted including 23 genes and 10 gene sets changed with aging mouse heart.

The present paper aims to summarize relevant ideas of normal aging process and factors involved in promoting or protecting tissue and organ aging. The changes in cardiac function and morphology with aging before onset of myocardial infarction (MI), myocardial infarction and remodeling and effects of aging factors and the search for ways to ameliorate the adverse outcome of postMI remodeling in older individuals including use of “stem cells” for infarct repair.

2. Pro- and Antiaging Contributors

Multiple factors, genetic and environmental, affect the aging/longevity process in cells, tissues and organ systems of humans. The theoretical factors of aging include somatic mutation, telomere loss, mitochondrial oxidative damage, and altered proteins, network theories; these factors do not
adequately explain differences in longevity in model, wild animals and humans and as some organisms show no aging and as such studies in model organisms may not reproduce human aging adequately [1].

2.1. ROS-Induced Mitochondrial and DNA Damage. Reactive oxygen species (ROS) are generated from the mitochondria under normal conditions and increases with cellular stress. NADPH oxidase family of proteins (Nox1, Nox2, Nox3, Nox4, Nox5, Duox) are the most important for generating ROS in the heart and other tissues and cells [11, 12]. Nox family oxidases contribute to cardiac hypertrophy and fibrosis [12]. ROS can damage DNA, proteins and can lead to cell death (apoptosis). ROS-induced DNA damage generate mutations as seen in mice and humans. There are intrinsic antioxidants such as the superoxide dismutases (copper/zinc and manganese). In animal models, the roles of molecules/genes in ageing are defined by using knockouts and mutations of individual molecules and the models generally lead to premature aging. Inadequate or poor DNA-damage and repair response leads to tissue stem cell depletion, senescence and apoptosis and ultimately aging [13]. Antioxidative enzymes such as superoxide dismutase (SOD) and catalase are also antiaging in model systems, though their contribution longevity in humans is debated [2].

2.2. Protein Misfolding and Aging. Heat shock proteins and their signaling pathways involved in cellular stress responses, responding protein misfolding, also promote aging when their functions decline [14].

2.3. Antiaging Hormone. A recently described antiaging gene in mouse knockout model, Klotho has many functions that include inhibition of insulin/insulin-like growth factor (IGF) signaling and oxidative stress [4, 15]. Klotho interacts with and dampens Wnt signaling and loss of klotho l leads to more Wnt signaling [16]. Klotho can be measured in serum and cerebrospinal fluid. Klotho expression variations exist within populations with cardiac implications [17].

2.4. Diabetes and Aging. Mouse models of diabetes leads to aging of “cardiac stem cells” due to generation of ROS, DNA damage and cell death [18].

2.5. Micronutrient Inadequacy. The Sirtuin family of proteins [19] (nuclear Sirt 1, 6, and 7, cytoplasmic Sirt 2, and mitochondrial Sirt 3–5) are involved in the response of cells to environmental and dietary stress. Sirtuins are NAD-dependent deacetylases and monoadenosine diphosphate-ribosyl transferases that regulate lipid and glucose metabolism, DNA repair, control cell survival, thermogenesis, insulin secretion, endothelial nitric oxide synthetase (eNOS), p53, and possibly mitochondrial flux [19]. Model organisms have suggested a role of Sirtuins in aging [19]. Sirt 1 regulates autophagy under starvation conditions. In aging model animals, Sirt 1 is increased with stress in myocytes and may serve as antiaging agent [20]. Sirt 3 in excess prevents cardiac hypertrophy by reducing ROS-dependent signaling [21]. Many metabolic changes, such as accumulation of altered proteins, advanced glycation end-products (AGE), toxic methylglyoxal, occur with aging [22] and these changes contribute to abnormal mitochondrial function.

2.6. Immunologic Aging. The participating cells of the innate and adaptive immune responses are affected by aging (immunosencence) due to reduction of thymic cell population, affinity of B-cell-derived immunoglobulins, and impaired function of CD3 + T-cells [23]. Other participants in innate immune response such as natural killer cells (NK cells) show reduced numbers and cytotoxic activity, dendritic cells and monocytes show reduced phagocytic functions, FasL, IL-12, and tumor necrosis factor (TNF-)alpha, and increased IL-10 with age [24]. The serum levels of the inflammatory mediator and chemokine, interleukin-6 (IL-6) increases with age [25] while its receptor declines though not directly responsible for rising IL-6 and may be influenced by diet and exercise.

2.7. p53 and Aging. The genome protector and complex transcriptional factor p53 has a role in aging. p53 is involved in many signaling pathways (DNA damage and repair signaling) and functions in cell death pathways(apoptosis and autophagy) and determines cell fate [26, 27]. p53 promotes aging via its induction of cell senescence that could compromise tissue renewal and pro-oxidant functions; it is antiageing by its stabilizing genome integrity and restraining glycosylation and many mouse models confirm the contribution of p53 in protecting genome integrity. p53 is a participant in stem cell fate and potential differentiation.

2.8. Sestrins and Aging. Sestrin family of proteins (sestrin 1, sestrin 2, and sestrin 3) were described in the past decade and are induced by oxidative stress, and hypoxia. Sestrins are present in all adult tissues. Sestrins interact with and inhibit mTOR, interact and regulated by FoxO and p53, and reduce effects of oxidative stress by generating peroxiredoxins; these sestrins increase autophagy, reduce ROS and cell growth [28, 29].
3. Telomeres, Telomerase and Aging

Telomeric DNA length is dependent on genetic inheritance, oxidative stress and telomerase activity [30]. The terminal (TTAGGG)n repeats vary from 5–15 kb in length and losses occur with each cell division. Telomere length is maintained by telomerase activity. Telomerase consists of telomerase reverse transcriptase (hTERT), dyskerin and telomerase RNA(hTR) and telomerase adds repeats of (TTAGGG)n to ends of telomeres. Epigenetic changes in telomeres due to methylation and retinoblastoma protein affect telomere length and recombination [31]. It is known that telomeres are transcribed by RNA polymeraseII to create noncoding RNAs (TERRA or TelRNA) and accumulation of these noncoding telRNAs occurs with aging [31]. Telomerase activity is high in embryonic cells, multipotent germ cells and cancer cells. The continuing loss of telomeres over time reduces replicative capacity of dividing cells. Telomere length varies among individuals, within cells and this variation depends on subtelomeric sequences. The minimum length is 2 kb but can be as low as 300 bp. Model organisms such as mice have longer telomeres. Telomere length, measured by confocal telomere quantitative fluorescence (Q-FISH) is longest in stem cell niches [32]. Gender- and organ-related telomere length is described in model organisms [33]. Telomere length in young persons with paternal history of early myocardial infarction is generally shorter than controls [34].

Telomere loss can subject cells to replicative senescence, apoptosis by activating DNA damage signaling, p53-p21-retinoblastoma axis signaling pathways and hence genome instability [35]. Telomere attrition as a trigger for cellular senescence may subsequently affect the heart by (a) in the wound healing response by reduced inflammatory response and (b) limiting adult stem cell life span [36]. Telomere induced senescence leads to the so-called “senescence secretome” pathway that is characterized by (i) reduced IGF1 and Wnt2, (ii) increased IL-6 and its receptor, (iii) increased interferon gamma and cell death and (iv) increased matrix metalloproteinases 1 and 3, reduced fibronectin and collagen.

In both aging humans with telomere attrition and model mouse strains with TERC+/−, atherosclerosis and blood pressure increase with age, left ventricular function and wound healing response are impaired [37]. In population based study of telomere length and longevity of elderly persons, telomere length was associated with years of healthy life (YHL) but nor cardiovascular disease [38]. In human studies, telomere reduction may contribute to premature myocardial infarction [39].

The factors that are involved in human longevity have been sought through twin studies and multiple epidemiologic analysis. These studies, though imperfect and mainly done in a single country or a defined population groups, have found contribution of many of the factors to human longevity as summarized above for model organisms including heat shock proteins, IL-6, Sirtuins, Klotho [40].

4. Aging Heart before Myocardial Infarction (Part of the Aging Cardiovascular System)

Normal aging affects many components of the cardiovascular system. The heart undergoes increased left ventricular thickness, increased atrial size, reduced reserve, increased vascular diameter and wall stiffness and endothelial dysfunction [41, 42]. In addition, standing heart rate decreases with age [43]. Precursor/stem cells (c-kit and sca-1 positive) in human myocardium age and promote heart failure [44]. Age-related gene expression changes in human are described but the findings are limited by sample numbers [45]. In mouse models of wild-type and IGF-1 transgenic mice, aging is accompanied by myocyte senescence, increased apoptosis, necrosis, myocyte hypertrophy, loss of telomere-related protein TRF2 but not TRF1 and reducing telomere length [46]. In rat hearts, aging leads to increase in cytosolic cytochrome C, drop in apoptosis markers Bcl-2 and Bax, and increased lipid peroxidation [47]. Furthermore, aging hearts show defective autophagy in clearing accumulating damaged and oxidized lipids, damage and “large mitochondria” [48]. The aged heart, therefore is disadvantaged before any onset of myocardial ischemia and the extra burden of reperfusion.

5. Effects of Aging on Myocardial Infarction and Remodeling

Myocardial infarction involves several phases (i) myocyte damage and cell death (apoptosis, autophagy and necrosis), (ii) activation of and recruiting of inflammatory cells, (iii) mesenchymal/stem/progenitor cell response, recruitment and migration to infarct site and (iv) wound healing response and collagen deposition [49]. For the preconditioned and aging heart, the events of myocardial infarction are clearly additional burden and heavy stress. In addition ROS induces mitochondrial deficits and ATP depletion after myocardial infarction and reperfusion [50].

5.1. Effects on Cell Death. Myocardial infarction is associated with all cell death types-necrosis is best recognized but apoptosis and autophagy contribute to border zone and remote zone cell death [51]. In the aging heart, cell death is already elevated and renewal of lost cardiomyocytes is limited [44].

5.2. Effects on Inflammation. The inflammatory activation after MI involves both the innate and adaptive immune responses to damaged cells, DNA and membrane lipids. Recent studies in aging mouse models of MI demonstrated reduced migration of inflammatory cells and hence reduced cytokine expression and their downstream effects on wound repair [52].

5.3. Effects on Wound Repair. Mesenchymal stem cells are recruited to the infarct zone (IZ) as part of the wound repair response [53]. Although cardiac stem cells are recognized, they are few and have long turn-over interval [54]. The studies on diseased and aging human hearts showed attrition of c-kit+ve cells compared to nondiseased hearts and contributed to heart failure [44]. This finding and the general
knowledge of aging hematopoietic and mesenchymal stem cells may add to the impairment of recruiting stem cells to infarct zone. Wnt signaling also participates in wound healing [5].

5.4. Effects on Cardiac Hypertrophy. Post-MI remodeling is defined by (a) Infarct expansion and acute dilatation of infarct zone; (b) dilatation of noninfarct zone and increased sphericity, leading to poor function; (c) ventricular (myocyte) hypertrophy, myocytolysis, myocyte loss, myocyte lengthening, altered extracellular matrix [55]. The model for postMI remodeling in humans has been characterized as the “Bio-mechanical model” [56] which involves structural, functional changes in the heart and the activation of neuro-hormonal response [57, 58] and are the cornerstone for targeted therapeutics and clinical management. Myocardial hypertrophy is complex and promoted by stress signaling, calcium levels, growth factors, phosphoinositide-3-kinase (PI3K) and mitogen-activated protein kinase (MAPK) pathways, and fibroblasts. There are intrinsic negative regulators of cardiac hypertrophy which include Glucose synthase kinase (GSK), caveolin-3 and calcineurin inhibitors [59]. Aging mice after myocardial infarction show reduced collagen deposition, and myofibroblast induction with progression of remodeling and limited response to transforming growth factor (TGF-beta) [52]. In aged rats, postmyocardial infarction mortality and all left ventricular function parameters (reduced cardiac index, peak pressure and end-diastolic pressure and increased end-diastolic volume index) were impaired [60]. The emerging contributions of microRNAs (cardiac-specific) to angiogenesis, myocyte proliferation, myocyte apoptosis and fibrosis are not fully explored in relation to the aging heart and remodeling [61]. It is logical therefore that all components of the initiating MI and postMI remodeling are subject to aging effect and need full attention (Table 2).

6. Future and Emerging Approaches to Post-MI Remodeling and Heart Failure Management

In the search for “aging” factors, Human Aging Genomic Resources is now available and expanding [62]. The information generated should help promote more appropriate approaches to the so-called “aging therapies” [63]. The complex interactions of many signaling pathways in heart failure clearly calls for integrated approach to preclinical studies of aging effects on postinfarct remodeling. Although the conceptual framework for postinfarct remodeling is still strong and relevant for clinical management, newer models that incorporate all available information may be needed [64]. A systems based approach using all “omics” tools may be the way for better understanding of aging in general and specifically for cardiovascular discovery (Figure 1) [65, 66]. The suggestion that signaling pathways modulate with age [67] also means that system wide assessment of the effects of pro- and antiaging factors on cardiomyocyte function will be the optimal approach to the discovery of age-modulating pathways.

6.1. Do We Target Telomerase Function and Telomeres?

Telomerase inhibition (using nucleoside analogs or Glucose synthase kinase 3 inhibitors) are proposed for other conditions but obviously not needed to modify aging and stem cells [68].

6.2. Do We Promote Wound Healing and Reduce Cardiac Hypertrophy?

6.2.1. Emerging Use of Stem Cells for Infarct Repair. There are many attempts to use hematopoetic, peripheral blood or mesenchymal stem cells to mitigate the wound repair in
Infarct hearts [69]. The studies and clinical trials have used chemokines (Granulocyte-colony stimulating factor-GCSF) to promote engraftment combined with direct implantation into infarct zone via the coronary artery and have produced results in model animals and humans ([70, 71] but need careful assessments of cell types used and long-term regenerative potential [72]. In addition, the aged cardiac environment may influence the capacity of mobilized, transplanted, or implanted stem/progenitor cells to differentiate and function [73].

Induced pluripotent cells (iPSCs) generated by retroviral vectors carrying Oct4, Sox2, and Klf4, are now a potential source cells for cardiac repair [74, 75]. Biological and molecular profiles of iPSCs and potential drawbacks on their use have been reviewed recently [76]. Alternate method for generating same cells, RNA-induced pluripotent cells (RiPSCs) is described [77]. iPSCs have been used in mouse models of myocardial infarction and repair and the cells can repair infarcts by producing functional cardiomyocytes, smooth muscle cells and vessels [78]; a side effect is promoting teratomas.

A recent study in mice illustrates the need to verify the stem cell capacity to generate myocytes and repair infaracts and reestablish function [79]. The progenitor and stem cells used in studies and clinical trials probably have varying potentials and have differing stem cell biomarkers such as isl1, sca-1 like, Nksx2.5, GATA4, Lin, c-kit and others [80]. The cells with regenerative capacity used in infarct repair include (a) mesenchymal (b) hematopoietic (circulating), embryonic stem cells and (d) residual cardiac stem cells in the myocardium. Many mouse models have reported functional improvement after infusion or activation of stem cells of varying potential or type (cardiac progenitors, mesenchymal stem cells, human hematopoietic stem cells, or embryonic stem cells) [81–84]. Studies in humans including clinical trials are providing support for the mouse models [85]. Some studies show no improvement, as in the swine model of large infarcts using mononuclear cells [86] and older mouse models that had poorer engraftment of stem cells [87]. In reviewing five clinical trials [88], it was found that gains in ejection fraction were not universal and depended on length of study and time of assessment. Additional meta-analyses have highlighted the varied methods of delivery including intracoronary and endocardial, use of different stem cells [89] embryonic, adult) and the varied observation periods; the average increase left ventricular ejection fraction (LVEF) was 3% in ten randomized trials. The human studies have not separated young and older infarct patients to determine differences in engraftment and functional recovery. In order to properly compare results in model animals and in humans, complete or comparable data sets on the use of stem cells should be available.

6.2.2. Other Ways to Reverse Cardiac Hypertrophy. Melatonin, the pineal hormone and antioxidant [90], is a protector of cardiac reperfusion injury [91]. Melatonin, potentially are antiproliferative and antiapoptotic [92]. Ischemia and reperfusion related ROS-induced myocyte damage is reduced by melatonin in mouse models [93]. Sirt3 overexpression in mouse models can inhibit cardiac hypertrophy [21] by activation of Foxo3a-dependent antioxidants and inhibition of reactive oxygen species (ROS) dependent signaling. Melatonin also reduced reperfusion induced fibrosis and arrhythmias [94]. Some investigators suggest targeting the ROS-induced damages after reperfusion [50] and the use of statins, selective xanthine oxidase inhibitor oxipurinol [95].

6.2.3. Age Well to Improve the Heart. This is the simple part as long as ageing is not reformulated as an illness or disease. As aging is partly dependent on the environment, dietary approaches are simple. Dietary/calorie restriction is a well-defined way to promote longevity [96] and leads to activation of nutrient sensors (TOR-target of rapamycin) and Sirtuins which promote cell cycle progression, improve detoxification and mitochondrial biogenesis. Zinc in diet can limit aging in Klotho knockout mice [97]. Lipoic acid in model mice reduced oxidative stress [98]. Melatonin is also projected to improve aging-related immune senescence and change both the T-cell and cytokine profile [99]. An activator of Sirtuin and AMPK pathways, Resveratrol [100] has been promoted as antiaging supplement. Healthy habits such as diet including vitamins [101] and antioxidants such as blue-berries [102] are proposed to promote healthy aging. Tea-derived flavonoids (catechins and theaflavins) via their proposed antioxidant, antiinflammatory could protect from age-related effects [103]. Exercise [104] and other prevention could assist in healthy aging.

7. Summary

The contributions of the multiple pro- and antiaging factors to human aging are of great interest and are now better understood. The contributions of environmental, genetic and biologic factors (telomere attrition, sestrins, Sirt 1, klotho, diabetes, p53) to the function of the aging heart and their possible effects on the outcome of myocardial infarction, postMI remodeling and heart failure are important areas of research. An integrated approach to future studies in model animals and humans with myocardial infarction may uncover new therapeutic targets. There are emerging therapeutic options such as “stem cell therapy” for myocardial infarction. All of these advances will certainly benefit all aging individuals in future.

Acknowledgments

The author appreciates and acknowledges the support and encouragement of his wife, Mrs. Augusta. E Idikio, RN, BN. Many thanks are due to Professor Y. Coe (Cardiology) for reading the paper. He is also grateful to Prof. B. I. Jugdutt for the opportunity to participate in this very exciting area of postMI remodeling.
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