Removal of the 2-Mercaptobenzothiazole from Model Wastewater by Ozonation

Jan Derco, Angelika Kassai, Michal Melicher, and Jozef Dudas

Faculty of Chemical and Food Technology, Slovak University of Technology, Radlinského 9, 812 37 Bratislava, Slovakia
Water Research Institute, Nábr. Arm. Gen. L. Svobodu 5, 812 49 Bratislava, Slovakia

Correspondence should be addressed to Jan Derco; jan.derco@stuba.sk

Received 31 August 2013; Accepted 20 October 2013; Published 23 January 2014

Academic Editors: J. J. Brandner and M. A. Centeno

Copyright © 2014 Jan Derco et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The feasibility of ozonation process for 2-mercaptobenzothiazole (2-MBT) removal follows from results of ozonation of the model wastewater. Total removal of 2-MBT was observed after 20 minutes of ozonation. Very good reproducibility of repeated ozonation trials including sampling and analysis was observed. However, the majority of dissolved organic carbon (DOC) and chemical oxygen demand (COD) remained in the reaction mixture. Benzothiazole (BT) and 2-hydroxybenzothiazole (OBT) intermediates were identified during degradation of 2-MBT with ozone. In addition to the above benzothiazole derivatives, the creation of some other organic compounds follows from results of mass balance. The best fits of experimental data were obtained using the first kinetic model for 2-MBT and zero-order kinetic model for COD and DOC. The reaction time of 60 minutes can be considered as effective with regard to controlled oxidation in order to increase a portion of partially oxidized substances. Higher biodegradability and lower toxicity of ozonation products on respiration activity of activated sludge microorganisms was observed at higher ozonation time.

1. Introduction

Ozone is a very strong oxidant and reacts with most of inorganic and organic pollutants. It is used in water and wastewater treatment. Ozone can react directly with a compound or through reactions of hydroxyl radicals generated in the ozone decomposition that then react with a compound. It appears as an attractive treatment option due to its well-known capacity to oxidise aromatic compounds [1].

Benzothiazoles are toxic and poorly biodegradable pollutants. Benzothiazole and its derivatives are widely used as industrial chemicals in the leather and wood industries, as biocorrosion inhibitors in cooling systems, ingredients in antifreezing agents for automobiles, and mainly as vulcanization accelerators in rubber production. They are often used, as herbicides and fungicides, as anti-fungal drug, as corrosion inhibitors in cooling water, as slimicides in the paper and pulp industry, and mainly as vulcanization accelerators in rubber production [2]. Correspondingly, these xenobiotic compounds are widely distributed in the environment and have been detected in industrial wastewater, as well as in soils, estuarine sediments, and superficial water [3]. Benzothiazole compounds have been detected in various environmental compartments: in wastewaters, soils, estuarine sediments, and superficial waters [2].

Benzothiazoles pose an environmental concern when released into watercourses [4]. These compounds inhibit microorganisms activity in conventional biological wastewater treatment systems and most of them are not readily biodegradable [2, 5]. Moreover, these compounds can be absorbed into cell membranes, leading to bioaccumulation [2]. Unfortunately, conventional biological wastewater treatment cannot effectively remove such contaminants, since they are resistant to biodegradation [2].

2-Mercaptobenzothiazole (2-MBT) is widespread toxic and poorly biodegradable substance. Moreover, 2-MBT inhibits degradation of easily degradable organics and wastewater nitrification is inhibited at far lower concentrations [6]. This compound is known to be toxic to aquatic organisms [7],...
allergenic [8], and inducing tumours [9]. Published data related to biodegradation of MBT are ambiguous. Main prize et al. [10] and Gaja and Knapp [11] reported that it may be metabolised (although incompletely) by microorganisms which have been cultivated on other benzothiazoles. Chudoba et al. [12] suggested MBT as a recalcitrant to biodegradation. The removal of MBT by activated sludge is specified as a nonenzymatic process [11]. It may also hamper wastewater treatment processes at concentrations higher than 600 µmol·L⁻¹ [13]. Fiehn et al. [6] have found MBT to inhibit degradation of easily degradable organics and the process of nitrification was inhibited at a far lower concentration. MBT is readily transformed to oxidation products by ozonation and wastewaters containing MBT should easily be detoxified using ozone [6]. Thus, the development of efficient treatment/pretreatment processes is required in order to eliminate their discharge into aquatic ecosystems.

Puig et al. [14] studied the ozonation of complex wastewater containing eight benzothiazole derivatives. They achieved complete removal of 2-MBT but did not detect any benzothiazole derivative although there was no decrease in dissolved organic carbon (DOC). Fiehn et al. [6] investigated the ozonation of model wastewater containing 600 µmol·L⁻¹ of 2-MBT at pH 3, 7, and 9. All tests were carried out at a constant pH. An aerobically treated tannery effluent was taken for a comparative ozonation of 2-MBT at pH 7. This wastewater sample was also spiked with 600 µmol·L⁻¹ of 2-MBT. During ozonation maximum value of BT, OBT, and BTSO₂ was measured after 6, 12.5, and 15 minutes and the process took over 55 minutes to remove all identified benzothiazole derivatives, including 2-MBT.

No significant differences in the results of ozonation carried out at the above pH values were observed. During the comparative ozonation with aerobically treated tannery effluent spiked with the same amount of 2-MBT, the same intermediates were observed at the same time periods. For BT the same concentration was measured as during ozonation of the model wastewater. OBT concentration reached 1.5 times of the maximum concentration and BTSO₂ was about half compared with the measured values in the ozonation of the model wastewater. Based on the total balance of dissolved organic carbon (DOC), however, the authors stated that at the end of the process about 75% of the residual DOC remained unidentified. Therefore, benzothiazole derivatives may still enter the aquimincetatic environment, if the 2-MBT-containing effluents are not biologically degraded with suitable bacterial sludge after the ozonation [6].

This work aims to study the kinetics of degradation and transformation of 2-MBT with ozone and monitor the impact of ozonation products on activated sludge microorganisms. The aim is to find conditions for controlled ozonation 2-MBT in terms of its partial transformation, complete mineralization, and reduce toxicity to activated sludge microorganisms. Another objective is to determine the specific parameter values for 2-MBT and verify the reproducibility of experimental measurements including analytical determination of indicators 2-MBT, COD, and DOC.

2. Materials and Methods

2.1. Characterisation of Wastewater. The wastewater from rubber industry contains more than 200 mg·L⁻¹ of 2-MBT [1, 15]. Similar content of 2-MBT was measured in the wastewater produced from N-cyclohexyl-2-benzothiazol-sulfenamide [16, 17]. The concentration of MBT in model wastewater varied from 661 to 1055 mg·L⁻¹ for this set of experiments. The chemical was provided by Merck (≥95.5% purity).

2.2. Experimental Equipment and Procedures. Ozonation experiments were carried out in two glass columns, 0.04 m diameter and 1.70 m height. The first column was filled with activated sludge sample, and the other one with solution of potassium iodide. The reason was to specify the amount of residual ozone as well as to destroy residual ozone in the outlet of ozonation column. The effective volume of both columns was 1.0 litre. Schematic diagram of experimental ozonation apparatus is shown in Figure 1. The Lifetech ozone generator with the maximum ozone production 5.0 g·h⁻¹ was used. Mixture of ozone and oxygen was introduced into ozonation column at the bottom, and it was mixed with model wastewater sample through fine-bubble porous aeration element.

The system was operated in batch mode with regard to wastewater samples. Wastewater samples were added to ozonation column at the beginning of trials. Continuous flows of oxygen at 20 L·h⁻¹ (T = 22.0°C, p = 99 800 Pa) were applied for generation of ozone. Ozonation trials were carried out at performance of ozone generator 70% of the power maximum. Ozonation times were from 20 to 60 minutes. At selected time intervals, samples were collected and analyzed for indicators COD and identified benzothiazoles (BT, 2-MBT, OBT).

The outlet gas mixture was conducted into second bubble column through a fine-bubble porous distribution element. The dimensions of the column were identical to ozonation column. The column was filled with a solution of potassium iodide. The excess ozone destruction was carried out in this column. Likewise as the ozonation column, the effective volume of the bubble column was 1.0 L. The concentration of ozone in gas phase was measured using Life ODU 200 Analyzer.

The influence of raw and ozonated wastewater samples on activated sludge microorganisms activity was evaluated based on results obtained by respirometric measurements. Oxygen uptake rates by microorganisms of activated sludge, which was cultivated in semicontinuous laboratory scale activated sludge process were measured [18].

2.3. Analytical Procedures. 2-MBT and other BT derivatives were analysed using Hewlett Packard Liquid Chromatography series II 1090 with DAD detector. Direct injection method was applied using linear gradient of RP-HPLC with UV-DAD detector on the column C18 (Merck). TOC was measured with analyzer Shimadzu TOC-VCPH/CPN (Japan). Analytical control of model wastewater during the treatment
procedures included also COD (chemical oxygen demand) determination [19, 20].

2.4. Mathematical Treatment of Experimental Data. Experimental data were fitted by zero (see (1), first see (2) and then see (3)) order reaction kinetic models. For a batch reaction system, under the assumption of a constant reaction volume, the following relationships are obtained

$$\text{COD}_t = \text{COD}_0 - k_0 t,$$

(1)

$$\text{COD}_t = \text{COD}_0 \exp(-k_1 t),$$

(2)

$$\text{COD}_t = \frac{\text{COD}_0}{1 + \text{COD}_0 k_2 t},$$

(3)

where $\text{COD}_0/(\text{g} \cdot \text{m}^{-3})$ denotes the value of COD in wastewater in time $t$, $\text{COD}_0/(\text{g} \cdot \text{m}^{-3})$ the initial value of COD in wastewater, and $k_0/(\text{g} \cdot \text{m}^{-3} \cdot \text{h}^{-1})$, $k_1/\text{h}^{-1}$, $k_2/(\text{g}^{-1} \cdot \text{m}^3 \cdot \text{h}^{-1})$ the rate constants for the kinetics of zero, the first, and the second order, respectively.

The parameters of the applied kinetic models were calculated by the grid search optimization procedure. The residual sum of squares ($S^2$) between the observed values and the values given by the model divided by its number of degrees of freedom $\nu$ (the number of observations less the number of parameters estimated) was used as the objective function.

2.5. Partial Oxidation and Mineralization. To evaluate the oxidized and mineralized portions of synthetic sewage pollution during ozonation and identification of the highest efficiency of partial oxidation, we used following equation [21, 22]:

$$\alpha_{\text{COD}_{\text{oxi}}} = 1 - \frac{\text{COD}_t}{\text{COD}_0},$$

(4)

$$\alpha_{\text{COD}_{\text{miner}}} = 1 - \frac{\text{DOC}_t}{\text{DOC}_0},$$

(5)

$$\alpha_{\text{COD}_{\text{partoxi}}} = \alpha_{\text{COD}_{\text{oxi}}} - \alpha_{\text{COD}_{\text{miner}}},$$

(6)

$$\mu_{\text{COD}_{\text{partoxi}}} = \frac{\alpha_{\text{COD}_{\text{partoxi}}}}{\alpha_{\text{COD}_{\text{oxi}}}},$$

(7)

where $\alpha_{\text{COD}_{\text{oxi}}}$ is the proportion of oxidized organic pollutants expressed as COD, $\alpha_{\text{COD}_{\text{miner}}}$ is the proportion of oxidized COD, which was completely mineralized, and $\alpha_{\text{COD}_{\text{partoxi}}}$ is only partially oxidized COD of the synthetic wastewater during the process. The effectiveness of partial oxidation $\mu_{\text{COD}_{\text{partoxi}}}$ is proportional to the ratio of decrease in COD due to the transformation of substances intermediates ($\alpha_{\text{COD}_{\text{partoxi}}}$) and the decrease of oxidized COD of the synthetic wastewater ($\alpha_{\text{COD}_{\text{oxi}}}$) in the process.

2.6. Impact on Activated Sludge Microorganisms. Respirometric measurements [22] were conducted to assess the impact of ozonation products on activated sludge microorganisms. The measurement results have been processed using the following Monod equation (see (8), [22], and for Haldan equation see (9)) [23, 24]:

$$r_X = r_{X,\text{max}} \frac{S}{K_S + S},$$

(8)

$$r_X = r_{X,\text{max}} \frac{S}{K_S + S + (S^2/K_I)},$$

(9)

where $r_X$ and $r_{X,\text{max}}$ are, respectively, the specific respiration rate and maximum specific respiration rate [mg·g$^{-1}$·h$^{-1}$], and $S$, $K_S$, and $K_I$ are the substrate concentration, half-saturation constant, and inhibition constant [mg·L$^{-1}$]. The values of kinetic parameters in (8), and (9) were determined using the grid search method.

2.7. Mean Oxidation State of Organic Carbon. Wastewater treatment processes try to convert the environmentally harmful carbon species into a less problematic one, often CO$_2$. A suitable method to monitor the fate and oxidation state of the total organically bound carbon is the mean oxidation number of organic carbon (MOC) as proposed by Stum and Morgan [25] and extended by Vogel et al. [26]. For wastewater containing more pollutants, COD and TOC values can be combined to give a MOC estimate according to (10):

$$\text{MOC} = 4 - 1.5 \frac{\text{COD}_o}{\text{TOC}},$$

(10)

where \text{COD}_o is the organic COD and \text{TOC} is total organic carbon.
3. Results and Discussion

Three independent series of measurements, with reaction times of 20–60 minutes, were performed. Thus, overall 23 experimental values were used for each monitored variable to evaluate the kinetics. The pH values varied in the range from 12.3 to 7.9 during ozonation experiments.

The time dependencies of measured and calculated values of 2-MBT, COD, and DOC are presented in Figures 2 to 4. The boundaries of the 95% confidence intervals, which were calculated on the basis of statistical analysis of repeated measurements, are also marked in these figures.

There can be seen two regions in Figure 2. During the first region with duration of approximately 15 minutes, the conversion of 2-MBT to BT and other BT-derivatives occurs according to mechanisms proposed by Fiehn et al. [6] and results published by Derco et al. [17, 27]. Kinetic data measured for this region were fitted very well by zero-order kinetics. The trend of 2-MBT transformation is very similar with the trends of DOC and COD degradation kinetics (Figures 3 and 4). However, from the values of zero-order kinetic constants (Table 1) it follows that transformation and degradation processes occur simultaneously with obviously higher transformation rate of 2-MBT in comparison to degradation rates of DOC and COD. Relatively very low 2-MBT concentration values and consequently also 2-MBT decline rates are obvious for the second stage of 2-MBT dependence on ozonation time.

From Figure 2 it is evident that during the first 20 minutes of the process performance almost complete elimination of 2-MBT in the model wastewater occurred. However, after this reaction time, 93.8% of the original value of DOC (Figure 3), and 75.5% of the original value of COD (Figure 4) remained in the model wastewater.

It is obvious that 2-MBT was only partially oxidized and majority was transformed to intermediates and products, respectively simple benzothiazole derivatives, which remained in the treated model wastewater. During the next 40 minutes of ozonation performance DOC value declined to 68.5% and COD decreased to 37.6% of the initial values.

Calculated values in Figures 2 to 4 correspond to the best descriptions of the experimental values using conventional kinetic models. The values of kinetic parameters and statistical characteristics (correlation coefficient and residual dispersion) are for individual indicators of pollution and kinetic models with the best description of the experimental values given in Table 1. The best description of the experimental values of DOC and COD was achieved using zero kinetics (Table 1). Removal of 2-MBT corresponds to the first kinetic model (Table 1).

From dependencies pictured in Figures 2 to 4 from the values of the statistical characteristics given in Table 1, and it can be seen that the largest residual dispersion between measured and calculated values shows time dependence of 2-MBT values. In Table 2 there are given the results of statistical treatment of repeated experiments which were carried out to verify the reproducibility of the results achieved. The removals of studied pollution indicators depending on reaction time are graphically illustrated in Figures 2 to 4.

The results in Table 2 show the largest variability of COD values measured after 15 minutes of ozonation. In the case of 2-MBT, the value of 95% confidence interval did not exceed 3% of the average 2-MBT value after 15 minutes of ozonation. The accuracy of repeated measurements for DOC

| Variable | $N$ | $k_n$ | Dimension of $k_n$ | $r^2_{XY}$ | $S^2_Y$ |
|----------|-----|-------|-------------------|-----------|--------|
| MBT      | 0   | 55.59 | mg L$^{-1}$ min$^{-1}$ | 0.9888    | 968    |
| DOC      | 0   | 1.77  | mg L$^{-1}$ min$^{-1}$ | 0.7590    | 248    |
| COD      | 0   | 19.21 | mg L$^{-1}$ min$^{-1}$ | 0.9175    | 8143   |
Table 2: Results of statistical processing achieved removal efficiencies % in repeated experiments aimed to verify the reproducibility of results.

| Reaction time | Variable | Pollutant average efficiency | 95% confidence interval | Pollutant average efficiency | 95% confidence interval | Pollutant average efficiency | 95% confidence interval |
|---------------|----------|------------------------------|-------------------------|------------------------------|-------------------------|------------------------------|-------------------------|
| 15 min        | 2-MBT    | 97.2                         | 2.9                     | 30 min                       | —                       | 95% confidence interval     | —                       |
|               | DOC      | 4.6                          | 1.0                     | 60 min                       | 15.5                    | 1.3                         | 36.5                    |
|               | COD      | 27.1                         | 9.6                     |                              | 41.3                    | 2.3                         | 65.8                    |

Figure 5: Dependence of concentrations (●) 2-MBT, (⧫) BT, and (◼) OBT on ozonation time.

and COD pollution indicators increases at longer reaction times. The limit values for 95% confidence intervals for these indicators of pollution did not exceed 3% of the average values after 60 minutes of ozonation. It can be concluded that the reproducibility of the results of repeated ozonation assays, including sampling and analysis, was very good.

From the statistical analysis of repeated measurements of initial values, the following specific values for 2-MBT have been obtained: COD/2-MBT = 2.008 g·g⁻¹ (limit value for 95% confidence interval is ±0.145 g·g⁻¹) and DOC/2-MBT = 0.463 g·g⁻¹ (limit value for 95% confidence interval is ±0.032 g·g⁻¹).

In Figure 5 are in addition to 2-MBT shown also time dependencies of identified intermediates (BT and OBT) of 2-MBT transformation with ozone. It can be seen from these dependencies that to a maximum decrease of 2-MBT concentration correspond maximum values of the degradation products. After removal/transformation of 2-MBT the content of these derivatives gradually decreases to zero concentration.

Taking into account the above values of DOC and COD, it is obvious that in the sample of ozonated model wastewater there are accumulated so far unidentified reaction products.

Figure 6 shows the dependencies of oxidation, mineralization, partial oxidation portions, and the effectiveness of partial oxidation (see (4)–(7)) on ozonation time during 60 minutes ozonation assay with the model sample of 2-MBT. Of these dependencies it follows that time courses of the portions of COD oxidation (see (4), αCODoxi), mineralization (see (5), αCODmin), and partial oxidation (see (6), αCODpartoxi) exhibit an approximately linear dependence on the reaction time. High partial oxidation efficiency (see (7), μCODpartoxi about 73%) corresponds to the reaction time of 20 minutes. Due to the fact that this response time corresponds to a relatively low proportion of αCODoxi (19.3%), of which the portion of αCODpartoxi is 14.1%, these parameters cannot be considered to be effective in terms of controlled oxidation in order to increase the portion of partially oxidized substances with higher oxygen content prior to completing mineralization processes by economically the most convenient biological wastewater treatment processes. From this aspect an effective reaction time is 60 minutes (αCODoxi = 62.3%), while the proportion of partially mineralized αCODpartoxi is 51.0%.

This is confirmed by the time dependencies of specific consumption of ozone related to unit mass of COD reduction (ΔO₃/ΔCOD) and the mean oxidation number of organic carbon MOC [22, 26], which are shown in Figure 7.

In Figure 8 the time courses of ozone concentrations are shown measured under the same conditions at the inlet and outlet of the ozonation column filled both, with demineralized water and the model wastewater. It can be seen that, during approximately 40 minutes of ozonation of the model wastewater, nearly all supplied ozone was transferred to the liquid phase. After this time, ozone concentration in the gas phase gradually increased, indicating a less consumption of ozone for oxidation of created intermediates and products. This indicates also the reaction conditions, at which the ozonation process is not limited by the amount of ozone supplied. Based on ozone balance can be stated that 98.6% of O₃ was transferred to liquid phase.

The results of respirometric measurements to assess the impact of products generated during the ozonation of the model wastewater containing 2-MBT on the activity of
the activated sludge microorganisms are shown in Figure 9. Respirometric measurements were performed with activated sludge, which was cultivated in semicontinuous lab-scale model of activated sludge process (glucose, peptone, sludge age 10 days). The first ozonation of the model wastewater lasted 15 minutes. From the dependence of specific oxygen uptake rate on COD value it is clear that at higher COD values there exists inhibition of respiratory activity of activated sludge microorganisms. For a description of this dependence Haldane kinetic model was used (see (9)). The second curve in Figure 9 corresponds to the measurement of respiratory activity after the addition of ozonized model wastewater samples containing 2-MBT after 40 minutes ozonation. For a description of this dependence Monod kinetic model was used (see (8)). Parameter values of Monod and Haldane equations and correlation coefficient values are given in Table 3. The maximum values of specific respiration rate obtained for ozonized samples of synthetic sewage of 2-MBT are significantly smaller than for reference sample glucose ($r_{x,m} = 99.5 \text{ mg} \cdot \text{g}^{-1} \cdot \text{h}^{-1}$). On the other hand, respirometric measurements indicate that the inhibitory effect of a model substance and its degradation products to ozone can be reduced by a longer duration of the ozonation.

### Table 3: Kinetic parameters and correlation coefficients.

| Kinetic equation | $r_{x,m}$ (mg g$^{-1}$ h$^{-1}$) | $K_S$ (mg dm$^{-3}$) | $K_I$ (mg dm$^{-3}$) | $r_{xy}^2$ (-) |
|------------------|---------------------------------|---------------------|---------------------|----------------|
| Monod            | 11.6                            | 70.2                | —                   | 0.9767         |
| Haldane          | 18.1                            | 334.4               | 44.1                | 0.9275         |

4. Conclusions

From the evaluation of the results of repeated measurements it can be concluded that the reproducibility of treatment assays including sampling and analysis was very good. The best description of the experimental 2-MBT, COD, and DOC data was obtained by using zero kinetic model. Almost complete removal of 2-MBT was observed after 20 minutes of ozonation. However, in the reaction mixture there were 93.8% of DOC and 75.5% of COD of the original values at that time. These results confirm that at the first stage of 2-MBT oxidation with ozone prevails its transformation into other organic materials. Benzothiazole derivatives OBT and BT were identified as emerging intermediates during decomposition of 2-MBT with ozone. Results of mass balance of carbon indicate that also other organic compounds are created. The reaction time of 60 minutes with 51.0% proportion of partially oxidized COD was found to be effective in terms of controlled oxidation in order to increase the amount of partially oxidized compounds. From the balance of ozone amounts at the inlet and outlet of the reactor it follows that 98.6% of supplied ozone during the ozonation was transferred from gas into liquid phase. The results of respirometric measurements show that the specific respiration rate of the model wastewater sample ozonated for 40 minutes was significantly higher compared to the sample after 15 minutes of ozonation. This can be explained by the fact that the sample ozonated for 15 minutes contained more less ozonated substances, which are toxic on microorganisms of activated sludge at higher concentrations, reducing their respiration activity and having an inhibitory effect.
Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported by the Slovak Research and Development Agency under contract no. APVV-0656-12. The authors wish to thank also the support of the VEGA Grant 1/0734/12.

References

[1] H. Valdés, C. A. Zaror, and M. Jekel, “Kinetic study of reactions between ozone and benzothiazole in water,” Water Science and Technology, vol. 48, no. 11-12, pp. 505–510, 2003.
[2] H. de Wever and H. Verachtert, “Biodegradation and toxicity of benzothiazoles,” Water Research, vol. 31, no. 11, pp. 2673–2684, 1997.
[3] H. Valdés and C. A. Zaror, “Ozonation of benzothiazole saturated-activated carbons: influence of carbon chemical surface properties,” Journal of Hazardous Materials, vol. 137, no. 2, pp. 1042–1048, 2006.
[4] H. Valdés, F. A. Murillo, J. A. Manoli, and C. A. Zaror, “Heterogeneous catalytic ozonation of benzothiazole aqueous solution promoted by volcanic sand,” Journal of Hazardous Materials, vol. 153, no. 3, pp. 1036–1042, 2008.
[5] H. De Wever, S. Weiss, T. Reemtsma et al., “Comparison of sulfonated and other micropollutants removal in membrane bioreactor and conventional wastewater treatment,” Water Research, vol. 41, no. 4, pp. 935–945, 2007.
[6] O. Fiehn, G. Wegener, J. Jochimsen, and M. Jekel, “Analysis of the ozonation of 2-mercaptobenzothiazole in water and tannery wastewater using sum parameters, liquid- and gas chromatography and capillary electrophoresis,” Water Research, vol. 32, no. 4, pp. 1075–1084, 1998.
[7] Y. Yoshioka and Y. Ose, “A quantitative structure-activity relationship study and ecotoxicological risk quotient for the protection from chemical pollution,” Environmental Toxicology and Water Quality, vol. 8, no. 1, pp. 87–101, 1993.
[8] J. H. Jung, J. L. McLaughlin, and J. D. Guin, “Isolation, via activity-directed fractionation, of mercaptobenzothiazole and dibenzothiazyl disulfide as 2 allergens responsible for tennis shoe dermatitis,” Contact Dermatitis, vol. 19, no. 4, pp. 254–259, 1988.
[9] L. Swirsky Gold, T. H. Slone, B. R. Stern, and L. Bernstein, “Comparison of target organs of carcinogenicity for mutagenic and non-mutagenic chemicals,” Mutation Research, vol. 286, no. 1, pp. 75–100, 1993.
[10] J. Mainprize, J. S. Knapp, and A. G. Calley, “The fate of benzothiazole 2 sulphonylic acid in biologically treated industrial effluents,” Journal of Applied Bacteriology, vol. 40, no. 3, pp. 285–291, 1976.
[11] M. A. Gaja and J. S. Knapp, “Removal of 2-mercaptobenzothiazole by activated sludge: a cautionary note,” Water Research, vol. 32, no. 12, pp. 3786–3789, 1998.
[12] J. Chudoba, F. Tucek, and K. Zies, “Biochemischer Abbau von Benzothiazoldervaten,” Acta Hydrochim, vol. 5, pp. 499–501, 1977.
[13] H. De Wever, K. De Moor, and H. Verachtert, “Toxicity of 2-mercaptobenzothiazole towards bacterial growth and respiration,” Applied Microbiology and Biotechnology, vol. 42, no. 4, pp. 631–635, 1994.
[14] A. Puig, P. Ormad, P. Roche, J. Sarasa, E. Gimeno, and J. L. Ovelleiro, “Wastewater from the manufacture of rubber vulcanization accelerators: characterization, downstream monitoring and chemical treatment,” Journal of Chromatography A, vol. 733, no. 1-2, pp. 511–522, 1996.
[15] M. H. Habibi, S. Tangestaninejad, and B. Yadollahi, “Photocatalytic mineralisation of mercaptans as environmental pollutants in aquatic system using TiO2 suspension,” Applied Catalysis B, vol. 33, no. 1, pp. 57–63, 2001.
[16] J. Derco, A. Gulyásová, M. Králik, and L. Mraňková, “Treatment of an industrial wastewater by ozonation,” Petroleum and Coal, vol. 42, no. 2, pp. 92–97, 2001.
[17] J. Derco, M. Melicher, A. Kassai, J. Dudas, and M. Valicková, “Removal of Benzothiazoles by ozone pretreatment,” Environmental Engineering Science, vol. 28, no. 11, pp. 781–785, 2011.
[18] H. Spanjer, P. A. Vanrolleghem, G. Olson, and P. L. Dold, “Respirometry in control of the activated sludge process: principles,” IAWQ Scientific and Technical Report 7, J. W. Arrowsmith, Bristol, UK, 2000.
[19] ISO, 6060, Water Quality—Determination of Chemical Oxygen Demand, International Organisation for Standardisation, Geneva, Switzerland, 1989.
[20] A. E. Greenberg, L. S. Clesceri, and A. Eaton, Eds., Standard Methods for the Examination of Water and Wastewater, American Public Health Association, Washington, DC, USA, 22nd edition, 2005.
[21] J. C. Jochimsen and M. R. Jekel, “Partial oxidation effects during the combined oxidative and biological treatment of separated streams of tannery wastewater,” Water Science and Technology, vol. 35, no. 4, pp. 337–345, 1997.
[22] D. Mantzavinos, E. Lauer, M. Sahibzada, A. G. Livingston, and I. S. Metcalfe, “Assessment of partial treatment of polyethylene glycol wastewaters by wet air oxidation,” Water Research, vol. 34, no. 5, pp. 1620–1628, 2000.
[23] J. R. Sonnad and C. T. Goudar, “Solution of the Haldane equation for substrate inhibition enzyme kinetics using the decomposition method,” Mathematical and Computer Modelling, vol. 40, no. 5-6, pp. 573–582, 2004.
[24] G. Tziotzios, G. Lyberatos, S. Pavlou, and D. V. Vayenas, “Modelling of biological phenol removal in draw-fill reactors using suspended and attached growth olive pulp bacteria,” International Biodeterioration and Biodegradation, vol. 61, no. 2, pp. 142–150, 2008.
[25] W. Stum and J. J. Morgan, Aquatic Chemistry, John Wiley & Sons, New York, USA, 2nd edition, 1981.
[26] F. Vogel, J. Harf, A. Hug, and P. R. Von Rohr, “The mean oxidation number of carbon (MOC)—a useful concept for describing oxidation processes,” Water Research, vol. 34, no. 10, pp. 2689–2702, 2000.
[27] J. Derco, M. Melicher, and A. Kassai, “Removal of selected benzothiazols with ozone,” in Municipal and Industrial Waste Disposal, X.-Y. Yu, Ed., vol. 26, p. 239, INTECH, Rijeka, Croatia, 2012.