Membrane transporter proteins: a challenge for CNS drug development
François Girardin, MD

The consequences of transporters on central nervous system (CNS) drug development are becoming increasingly important, due to their influence on clinical outcome. Membrane transporters provide insight into the mechanisms of treatment failure, adverse drug reactions, and individual differences in the management of neurological and psychiatric disorders.

The presence of uptake and efflux transporters in capillary endothelial cells mediates drug transport from the circulatory system to the brain. These transporters play a significant role in drug absorption and distribution to organic systems, particularly if the organs are protected by blood-organ barriers, such as the blood-brain barrier or the maternal-fetal barrier. In contrast to neurotransmitters and receptor-coupled transporters or other modes of interneuronal transmission, drug transporters are not directly involved in specific neuronal functions, but provide global protection to the central nervous system. The lack of capillary fenestration, the low pinocytic activity, and the tight junctions between brain capillary and choroid plexus endothelial cells represent further gatekeepers limiting the entrance of endogenous and exogenous compounds into the central nervous system. Drug transport is a result of the concerted action of efflux and influx pumps (transporters) located both in the basolateral and apical membranes of brain capillary and choroid plexus endothelial cells. By regulating efflux and influx of endogenous or exogenous substances, the blood-brain barrier and, to a lesser extent, the blood-cerebrospinal barrier in the ventricles, represent the main interface between the central nervous system and the blood, ie, the rest of the body. As drug distribution to organs is dependent on the affinity of a substrate for a specific transport system, membrane transporter proteins are increasingly recognized as a key determinant of drug disposition. Many drug transporters are members of the adenosine triphosphate (ATP)-binding cassette (ABC) transporter superfamily or the solute-linked carrier (SLC) class. The multidrug resistance protein MDR1 (ABCB1), also called P-glycoprotein, the multidrug resistance-associated proteins MRP1 (ABCC1) and MRP2 (ABCC2), and the breast cancer-resistance protein BCRP (ABCG2) are ATP-dependent efflux transporters expressed in the blood-brain barrier. They belong to the superfamily of ABC transporters, which export drugs from the intracellular to the extracellular milieu. Members of the SLC class of solute carriers include, for example, organic ion transporting peptides, organic cation transporters, and organic ion transporters. They are ATP-independent polypeptides principally expressed at the basolateral membrane of brain capillary and choroid plexus endothelial cells that also mediate drug transport through central nervous system barriers.

Keywords: drug transporter; blood-brain barrier; drug distribution; ATP-binding cassette transporter; solute-linked carrier

Author affiliations: Unit of Clinical Psychopharmacology, Geneva University Hospitals, Chênes-Bourg, Geneva, Switzerland

Address for correspondence: Dr François Girardin, Unit of Clinical Psychopharmacology and Department of Community Medicine, Geneva University Hospitals, CH-1225 Chênes-Bourg, Geneva, Switzerland (e-mail: françois.girardin@hcuge.ch)
bloodstream to the organs. For example, these membrane transporters play an essential role in the digestive tract, the kidney, and the liver, as well as other organic systems such as the blood cells, the placenta, and the central nervous system.1

All cells selectively transport endogenous and exogenous compounds across their membrane to maintain an intracellular milieu distinct from the outer one; this is achieved in part by the membrane transporters, which are multispecific transport proteins. These transporters display physiologic functions in terms of transporting endogenous compounds, eg, hormones, amino acids, bile acids, and lipids.2-4 Moreover, membrane transporter mutations may lead to severe genetic disorders such as cystic fibrosis (ABCC7),5 immune deficiency (ABCB2 and ABCB3),6 intrahepatic cholestasis of pregnancy (ABCB11),7 persistent hypoglycemia of infancy (ABCC8),8 X-linked adrenoleukodystrophy (ABCD1),9 X-linked ataxia with sideroblastic anemia (ABCB7),10 and retinal degeneration (ABCA4).11,12

This review focuses on the functional significance of membrane transporters as drug carriers: their role is constantly increasing in current medical practice, as they represent a key factor in clinical outcome.7,13,14

Physiological aspects

The blood-brain barrier (BBB) is a physical barrier, and maintains a given extracellular environment for neurons and glial cells in the CNS. The BBB is formed by the connection of closely sealed tight junctions between the capillary endothelial cells, which are not fenestrated and which display minimal pinocytosis (Figure 1). The capillary endothelial cells form a polarized barrier similar to that located in the retina or in the renal proximal tubule, which regulates diffusion of molecules across the BBB, and limits the entry of xenobiotics via paracellular pathways by intercellular tight junctions.15-17 Not all cerebral blood vessels are closely sealed with tight junctions: fenestrated capillaries are found in the pituitary gland and

Selected abbreviations and acronyms

ABC adenosine triphosphate-binding cassette
ATP adenosine triphosphate
BBB blood-brain barrier
BCRP breast cancer resistance protein
CNS central nervous system
CSB (blood) cerebrospinal barrier
MDR multidrug resistance
MRP multidrug resistance-associated protein
OAT organic ion transporter
OATP organic anion transporting peptide
OCT organic cation transporter
SLC solute-linked carriers

Figure 1. The blood-brain barrier and drug transporters in the capillary endothelial cells.

Figure 2. The cerebrospinal barrier and drug transporters in the choroid plexus cells.
in the circumventricular organs such as the area postrema, the lamina terminalis, and the median eminence.\(^{18}\) The choroid plexus is a highly vascularized epithelial organ which secretes the cerebrospinal fluid and regulates its composition through active and selective transport processes; it has an active role in the cleansing of the cerebrospinal fluid of endogenous and exogenous compounds.\(^{19}\) The blood cerebrospinal barrier (CSB) is considered as the second fluid barrier protecting the central nervous system: it is principally formed by epithelial cells of the choroid plexus in the ventricles and the arachnoid membrane. Like the brain capillary endothelial cells, the choroid plexus epithelial cells are connected by high-resistance tight junctions, which closely separate the blood from the cerebrospinal fluid compartment (Figure 2).\(^{20}\)

Together with the BBB and the CSB, the membrane transporter systems represent further gatekeepers to the CNS; these play a critical role in drug disposition.\(^{21}\) Membrane transporters either enhance or restrict drug distribution to the target organs. Depending on their main function, these membrane transporters are divided into two categories: the efflux (export) and the influx (uptake) transporters. Influx transport proteins facilitate and efflux transporters limit drug passage through membrane barriers such as the BBB or the CSB.\(^{22}\) Several membrane transporters are found at the apical and basolateral epithelial cell membrane of the brain capillary and of the choroid plexus endothelial cells (Figure 3).\(^{23}\)

**Pharmacological aspects**

Drug absorption from the systemic circulation into the CNS was previously considered a passive process that depended on drug physicochemical properties such as molecular size, lipophilicity, and the pK\(_a\) of a drug. Although the physicochemical properties of a medication do affect its absorption and access to target organs, transporter proteins have a major role in the overall drug distribution process through their targeted expression in tissue such as the brain. Until recently, most pharmacokinetic studies focused only on the role of drug-metabolizing enzymes as a key determinants of drug disposition.\(^{24}\)

Phase I enzymatic reactions are mainly represented by the cytochrome P450 mono-oxygenase system, as well as by other enzymes such as pseudocholinesterase or alcohol dehydrogenase. Phase I enzymatic reactions modify the chemical structure of the compound itself with either loss of pharmacological activity or, on the contrary, when prodrugs are administered, enhanced biological activity through biotransformation into an active metabolite.

In contrast, Phase II enzymatic reactions are conjugation reactions that lead to the formation of a covalent linkage with an endogenously derived compound such as glucuronic acid, sulfate, glutathione, or amino acids into highly polar conjugates excreted into the bile or urine: N-acetyltransferase 2 or catechol-O-methyltransferase are examples of enzymes that catalyze phase II reactions.

Drug membrane transporters enable access of compounds to phase I reactions and further elimination after phase II reactions. Thus, drug uptake transporters deliver the drug to an intracellular enzymatic detoxification system, whereas drug efflux transporters decrease the intracellular load from the detoxification system. Inhibition of transmembrane transporters may lead to a lower substrate uptake with a poorer intracellular access of the drug to the enzymatic systems.\(^{25}\)

This synergy between metabolizing enzymes and drug transporters accounts for the distribution to brain tissues, and determines the pharmacokinetic profile of CNS drugs.\(^{26}\) Furthermore, substrate similarity is well documented between membrane transporters and cytochrome P450 enzymes. For instance, substrates of the PGP multidrug resistance protein (MDR1) are usually substrates of the CYP3A4, digoxin and fexofenadine excepted.\(^{27}\)
Characterization of drug transporters

Many drug transporters are members of the adenosine triphosphate (ATP)-binding cassette (ABC) transporter superfamily or the solute-linked carrier (SLC) class. The ABC superfamily (ABC transporters) comprises the main efflux membrane transporters for drug elimination, and the solute-linked carriers (SLC transporters) are involved with influx and efflux transport function for drug uptake and export.28

ABC transporters

The ABC transporters are transmembranous proteins found in all animal species; they require energy from ATP hydrolysis to actively remove compounds from the cell, often against a high concentration gradient. They are composed of two transmembrane domains that form a pathway through which the substrates move, and by two nucleotide-binding domains located at the cytoplasmatic face of the membrane, providing ATP hydrolysis to allow substrate translocation across cellular membranes.29 Members of the ABC superfamily are classified as such according to consensus sequences including both domains (Walkers A and Walkers B ATP-binding motifs), as well as the ABC signature (C motif).30 To date, 49 human ABC family members have been identified, divided into seven different subfamilies. The ABC transporter superfamily includes medically important members such as the multidrug resistance-associated protein 1 (MRP1) located at the basolateral plasma membrane domain (abluminal), the multidrug resistance-associated protein 2 (MRP2), the breast cancer resistance protein (BCRP), and MDR1 (also known as PGP) localized at the apical membrane (luminal). MDR1 (encoded ABCB1), BCRP (encoded ABCG2), MRP1 (encoded ABCC1), and MRP2 (encoded ABCC2) were identified in the BBB at the luminal side of the capillary endothelial cells, MRP1 excepted (Figure 3).

The substrates handled by the ABC transporters include a wide range of endogenous and exogenous compounds and diverse type of molecules, from organic cations and anions to larger molecules such as large polypeptides or therapeutic agents. For instance, MRP1 has preferential transport of anionic compounds such as sulfate conjugates or glutathione, whereas MDR1 shows broader substrate specificity.31,32 Variation of ABC transporter activity by drug-drug interactions, genetic polymorphisms, and overexpression is considered as a major cause of treatment failure, interindividual variability, and adverse drug reactions.23,33 However, randomized controlled studies on these issues with antidepressants, antipsychotics, or mood stabilizers in humans are still lacking.

ABC transporters are considered as the most relevant determinants of efflux transport and provide multiple barriers in the brain capillary and choroid plexus endothelial cells.24 A multidrug resistance feature is associated with a poorer clinical outcome in several CNS disorders.24 Furthermore, several ABC transporters were directly implicated in drug delivery to brain neoplasms and in the response to therapeutic agents.38 For instance, the expression of the ABC transporters ABCC4 and ABCC5 was associated with an astrocytic phenotype with higher chemoresistance of astrocytic tumors compared with oligodendrogliomas.39 Recently, the ABC transporters were also found to be associated with pharmacoresistance to anticonvulsant drugs in patients with intractable mesial temporal lobe epilepsy.37 MDR1 activity significantly decreases during aging, with, consecutively, an increased brain exposure to drugs and toxins in elderly subjects.38 Furthermore, impaired MDR1 function is reported as a predisposing factor in the development of neurodegenerative diseases such as Parkinson’s disease or sporadic Alzheimer’s dementia.39 Pathologic accumulation of amyloid β in Alzheimer’s disease may result from an impaired MDR1 activity, as amyloid β is considered as a substrate for MDR1.13

SLC transporters

The SLC class of solute carriers consists of specific membrane transporters that mediate sodium-independent transmembrane solute transport: it is divided into 43 human families based upon amino acid homology of at least 25% between family members. To date, nearly 300 genes have been identified.40 The Human Genome Organization (HUGO) Nomenclature Committee Database provides information about new transporter families of the SLC gene series (SLC transporters-gene nomenclature, SLCO).41 Members of the SCLO superfamily are not only expressed in the BBB and in the choroid plexus, but also in the small intestine, the liver, the kidney, the blood-testis barrier, and the placenta.42,43 The SLCO superfamily of solute carriers includes carriers,
such as the organic anion transporting protein family (OATPs), the organic anion transporter family (OATs, \( SLCO22A \)), and the organic cation transporter family (OCTs, \( SLCO15A \)). The organic cation/carnitine transporter (OCTN, \( SLCO22A \)), the monocarboxylate transporter family (MCTs, \( SLCO16 \)), and the peptide transporter (PEPT, \( SLCO15A \)) may represent further important SLC families, and their function as CNS barriers is currently under investigation.44,45 For example, \( \alpha \)-hydroxybutyrate (GHB), a therapeutic agent for catalepsy with narcolepsy, undergoes passive diffusion through the BBB but also the MCT1 carrier-mediated process, that is saturable and can be inhibited.46 Proof-of-concept studies are being conducted to provide better insights into GHB therapy and GHB toxicity by means of transport inhibitors.47 Several in vitro and in vivo data indicate that OATP1A2, OATP1C1, and OATP2B1 (members of the \( SLCO21A \) family), and OAT1, OAT2, OCT1, OCT2, and OCT3 (members of the \( SLCO22A \) family) are expressed in the murine and human brain, and mediate drug transport through the CNS barriers.2-4,36,41,48  

The \( SLCO21A \) family is referred to as the OATP family: these transporters consist of 12 transmembrane domain proteins, whose substrates are anionic amphipathic high-molecular-weight molecules that bind to albumin.40 The transport mechanism is based upon anion exchange coupled to cellular uptake of organic compounds with the efflux of bicarbonate, glutathione, and conjugates. The \( SLCO22A \) transporters include OCTs including OCTN, and OATs that also consist of 12 transmembrane domain proteins, but with different substrate specificity. Indeed, OATPs not only mediate uptake of anionic, but also neutral and cationic, compounds. OCT members are mainly unidirectional porters, whereas OAT members act as anion exchangers. The organic anion transporting proteins (OATPs), OATs, and the OCTs represent the major uptake transport systems that mediate organic compound transport activities at the apical and the basolateral plasma membrane domains.

**Drug transporter polymorphisms**

The expression of transport proteins localized in the membranes of various organs are significant determinants of the pharmacokinetics of therapeutic agent including at the level of the CSB and the BBB.33,50-53 There is genetic polymorphism of drug transporters in the structure of genes and in the number of alleles. The \( MDRI \) gene has been particularly well investigated and several \( MDRI \) polymorphisms have been found: many of them determine membrane transporters expression in the BBB and in the CNS with variable drug transport activity.26,54,55 Such medically relevant polymorphisms are called nonsynonymous polymorphisms, as they directly condition the drug transporter function with potentially variable clinical outcomes. The functional significance of different \( MDRI \) expression for drug disposition was mainly studied with \( MDRI \) knockout mice. For instance, \( MDRI \) knockout mice are 50- to 100-fold more sensitive to the neurotoxic pesticide ivermectine, and the accumulation of this drug in the CNS was 80- to 100-fold greater when compared with control mice.56 In humans, several mutations resulting in several \( MDRI \) single nucleotide polymorphisms (SNPs) have been identified, but only those at positions 2677 and 3435 seem to be associated with changes in PGP expression and function.25,57,58 In contrast to the \( P450 \) drug-metabolizing enzymes such as CYP2C9, CYP2C19, and CYP2D6, for which loss of function mutations or gene amplification manifests as distinct phenotypes in the population (eg, poor, intermediate, extensive, or ultrarapid metabolizers), the impact of \( MDRI \) polymorphisms on pharmacokinetics is moderate: no definite \( MDRI \) phenotype is recognized in humans.59 There is no complete loss of transport function when polymorphisms are present: the genotype-related differences in the \( MDRI \) expression between, eg, the 3435 genotype, remains moderate with substantial overlap.59 However, the difference between clinical outcomes may be in some conditions very impressive: patients with drug-resistant epilepsy were much more likely to have the CC genotype at ABCB1 3435 than the TT genotype (odds ratio: 2.66).60 Furthermore, ABC transporter polymorphisms are not only associated with resistance to treatment or failure, for example, for anticonvulsants, cytostatics, or antibiotics, but they also determine the incidence of adverse drug events.50,53,57,60-63 Some examples of clinical effects and potential implications associated with human drug transporter polymorphisms are listed in Table I. Interestingly, the clinical impact of single nucleotide polymorphisms on genetic variability of expression and function of the multidrug resistance-associated proteins (MRPs, \( ABCC \) transporter) is to date rather limited as compared with eg, \( MDRI \).73 Outside the CNS, multiple but rare familial mutations in, eg, the \( ABCC2 \) gene (\( MRP2 \)) are responsible for the recessive inherited Dubin-Johnson syndrome: although hepatic function is
normal, patients with this syndrome have an increased risk of drug-induced liver toxicity.74 Although SLCO transporters are genetically extensively characterized, relevant clinical data about the impact of polymorphisms are still limited. Genetic variants of uptake transporters have predominantly been investigated for OATPs, but a large number of single nucleotide polymorphisms in the OCT1 (SLCO22A1) and OCT2 (SLCO22A2) gene were also found, altering the transport function in vitro.25,75 As OATP1A2 is predominantly localized in the capillary endothelial cells of the brain, genetic variability and polymorphisms of this drug uptake carrier may represent a future pathway for CNS drugs as it is a determinant of brain toxicity.23

**Drug transporter interactions**

Membrane transporters can undergo inhibition or induction, which respectively slow down or accelerate their transporter activity.76-78 In 1951, indirect clinical evidence already suggested the role of specific transport systems at the level of renal cell membranes79: coadministration of probenecid with penicillin resulted in decreased renal clearance, prolonged half-life, and elevated plasma level of penicillin, enabling a substantial reduction in antibiotic dose. The mechanism of this interaction was found several years later: the active penicillin secretion was reduced by OAT inhibition in the basolateral membrane of renal proximal tubule.80 Similarly, coadministration of probenecid with HIV antiviral drugs or with antihypertensive drugs such as the angiotensin-converting enzyme inhibitors also causes a reduction in renal clearance, a prolonged half-life, and elevated plasma levels.81 In humans, digoxin is a high-affinity substrate for MDR1, and most interacting drugs are either inductors, or, more frequently, inhibitors, of MDR1.82 Significant MDR1 inhibition, by administering atorvastatin, clarithromycin, or verapamil as MDR1 inhibitors, was associated with a significant increase in the serum digoxin concentration, ie, more

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### Table I. Examples of genetic polymorphisms in human drug transporters. ABC, adenosine triphosphate-binding cassette; MDR, multi-drug resistance; BCRP, breast cancer resistance protein; SLC, solute-linked carriers; OATP, organic anion transporting peptide; OAT, organic ion transporter; OCT, organic cation transporter; CNS, central nervous system.

| Symbol | Transporter | Polymorphism | Associated clinical effect |
|--------|-------------|--------------|---------------------------|
| ABCB1  | MDR1        | 3435 C>T     | 3435 C epilepsy drug-resistance67  
3435 TT fewer viral resistance in efavirenz patients,64 higher digoxin level,65 and nortriptyline-induced postural hypotension66 |
| ABCB1  | MDR1        | 1236 C>T     | 1236 decreased survival in acute myeloid leukaemia patients67 |
| ABCB1  | MDR1        | 2677 G>T/A   | 2677 T tacrolimus-induced neurotoxicity68 |
| ABCG2  | BCRP        | C421A        | Increased plasma concentration of diflomotecan69 |
| SCL21A3| OATP1A2     | 38T>C, 382A>T| CNS penetration and toxicity of various drugs and substrates52 |
| SCL21A3| OATP1A2     | 404A>T, 516A>C| |
| SCL21A3| OATP1A2     | 559 G>A, 2003 C>G| |
| SLC22A6| OAT1        | 1361G>A, 877C>T| Possible altered renal uptake of cidofovir and adefovir and drug-induced nephrotoxicity66 |
| SLC22A1| OCT1        | 181C>T, 1201G>A| Decreased substrate uptake in vitro71 |
| SLC22A2| OCT2        | 1198C>T, 495G>A| Decreased substrate uptake in vitro72 |

### Table II. Examples of drug-drug interactions involving drug transporters. MDR, multidrug resistance; CNS, central nervous system; OCT, organic cation transporter; OATP, organic anion transporting peptide; OAT, organic ion transporter.

| Drug - substrate | Inhibitor / inducer | Interaction | Effect |
|------------------|---------------------|-------------|--------|
| Loperamide       | Quinidine           | MDR1 inhibition | Increased CNS penetration of loperamide68 |
| Digoxin          | Paroxetine          | MDR1 inhibition | Increased CNS penetration of digoxin75 |
| Cyclosporine     | St John’s wort      | MDR1 induction | Decreased plasma level of cyclosporine8 |
| Desipramine      | Ritonavir           | OCT1 inhibition | Decreased hepatic uptake with poorer access to CYP3A443 |
| Fexofenadine     | Fruit juices        | OATP inhibition | Decreased oral bioavailability44 |
| Penicillin       | Probenecid          | OAT inhibition | Prolonged penicillin half-life by reduced renal clearance41 |
than twice the upper therapeutic limit. Another striking and clinically relevant effect of the PGP-associated interactions was demonstrated by giving healthy volunteers loperamide, an opiate that is not absorbed from the gut, simultaneously with quinidine, a potent MDR1 inhibitor: coadministration of this antidiarrheal agent with quinidine resulted in central opioid effect such as respiratory depression and euphoria, confirming in vivo a major MDR1 inhibition in the intestinal and in the BBB gatekeeper function.

Recently, a population pharmacokinetic analysis of drug-drug interactions between risperidone, bupropion, and sertraline in rodents suggested that sertraline produces significant inhibitory effects on MDR1 transport at the BBB, increasing the brain entry of risperidone and its metabolite 9-OH-risperidone. The order of magnitude was high, and could be clinically significant for humans: sertraline did not change the plasma concentration of risperidone and of its metabolite, but increased the brain area under the plasma concentration curve of risperidone and 9-hydroxy-risperidone 1.5-fold ($P<0.05$) and 5-fold ($P<0.01$), respectively.

Interestingly, another study with rodents showed that the MDR1 localized in the BBB is more resistant to inhibition than in other tissues. In vivo studies in humans are needed to assess the clinical relevance of such differential sensitivity to inhibition.

In vitro techniques for the assessment of drug-drug interactions involving membrane transporters are currently under development. Inhibition on the MDR1 was investigated in vitro with recent antidepressants, displaying different interaction potentials: whereas paroxetine is regarded as a strong PGP inhibitor, citalopram and venlafaxine are considered as antidepressants with low interaction potential with membrane transporters. A case report confirmed the ability of paroxetine to inhibit the PGP in the BBB and in the kidney, causing digitalis intoxication with delirium, visual hallucinations, and disorientation. However, specific data from in vivo studies, substrate specificity, and inhibition or induction potential of psychiatric and neurological medication are still lacking. Examples related to drug-drug interaction at the membrane transporter level are illustrated in Table II.

### Discussion

In vitro and in vivo studies show that drug carriers are expressed in the BBB and in the CSB. They represent major determinants of toxicity and clinical outcome related to drug response. Understanding the functional significance of membrane transporters in the BBB and in the CSB provides further opportunities to improve drug delivery to the CNS.

We propose that the role of transporter proteins should be studied at an early stage of CNS drug development, as there are in vitro methods such as cell cultures to achieve this purpose. Knockout animals are valuable in vivo models, but in vivo methods in humans are few. Direct in vivo determinations of drug concentration and effective transporter function into the brain remain particularly challenging, as invasive techniques are necessary. Neuroimaging techniques should be helpful, since molecules can be measured by positron emission tomography (PET) or by magnetic resonance imaging spectrometry. For example, the latter can be used to assess the pharmacokinetics of some fluoride-containing molecules in the brain.

Although several members of the membrane transporters present in the BBB have been characterized in detail, numerous questions remain open. Firstly, the determination of detailed tissue expressions and in vivo studies of carriers with better specificity are required to target more efficiently therapeutic agents into the CNS and into other organs. Secondly, in order to enhance the potential clinical implications of drug transporter polymorphisms and interactions, the development of specific inducers and inhibitors may represent promising strategies.

Thirdly, future delivery procedures include the use of prodrugs, drug-targeting vector conjugates, or liposomes tagged with targeting vectors to elude physiological barriers. Drug transporter protein studies provide insight into the mechanisms of resistance, treatment failure, and interindividual response to neurological and psychiatric medication. Membrane transporter proteins are not only CNS gatekeepers, but represent determinant partners in CNS drug development strategies. Exploring the functional significance of membrane transporters in drug delivery to the CNS is essential for the treatment of neurological and psychiatric disorders.
Proteínas transportadoras de membrana: un desafío para el desarrollo de fármacos para el sistema nervioso central

Los transportadores de fármacos son proteínas de membrana presentes en varios tejidos como linfocitos, intestino, hígado, riñón, testículos, placenta y el sistema nervioso central. Estos transportadores juegan un papel significativo en la absorción de fármacos y en la distribución a los sistemas del organismo, especialmente si los órganos están protegidos por barreras sangre-órgano, como la barrera hémato-encefálica o la barrera materno-fetal. En contraste con los neurotransmisores y los transportadores acoplados a receptores u otros modos de transmisión interneuronal, los transportadores de fármacos no están directamente involucrados en funciones neuronales específicas, pero proveen una protección global al sistema nervioso central. La falta de capilarización, la baja actividad de los pinocitos, y las uniones estrechas entre los capilares cerebrales y las células endoteliales de los plexos coroideos representan más barreras que limitan la entrada de compuestos endógenos y exógenos al sistema nervioso central. El transporte de fármacos es el resultado de una acción concertada de bombas de entrada y salida (transportadores) ubicadas en las membranas basolaterales y apicales de los capilares cerebrales y de las células endoteliales de los plexos coroideos. En la regulación de la entrada y salida de las sustancias endógenas o exógenas, la barrera hémato-encefálica, y en menor medida la barrera hémato-cerebroespinal en los ventrículos, representan el principal punto de contacto entre el sistema nervioso central y la sangre, es decir, el resto del organismo. Como la distribución de los fármacos a los órganos depende de la afinidad de un sustrato por un sistema de transporte específico, las proteínas transportadoras de membrana están siendo cada vez más reconocidas como un factor determinante en la disponibilidad de los fármacos. Muchos transportadores de fármacos son miembros de la superfamilia del transportador (ABC) unido a adenosín trifosfato (ATP) o de la clase de transportadores unidos a soluto (SLC). La proteína de resistencia a multifármacos MDR1 (ABCB1), también llamada P-glicoproteína, las proteínas asociadas a resistencia a multifármacos MRP1 (ABCC1) y MRP2 (ABCC2), y la proteína de resistencia al cáncer de mama BCRP (ABCG2) son transportadores de salida dependientes de ATP que expresan en la barrera hémato-encefálica. Ellos pertenecen a la superfamilia de transportadores ABC, los cuales llevan fármacos desde el medio intracelular al extracelular. Los miembros de la clase SLC de los transportadores de soluto incluyen, por ejemplo, péptidos transportadores de iones orgánicos, transportadores de cationes orgánicos y transportadores de iones orgánicos. Ellos son polipéptidos independientes de ATP que se expresan principalmente en la membrana basolateral de los capilares cerebrales y las células endoteliales de los plexos coroideos que también median el transporte de fármacos a través de las barreras del sistema nervioso central.

REFERENCES

1. Tamai I, Tsuji A. Transporter-mediated permeation of drugs across the blood-brain barrier. J Pharm Sci. 2000;89:1371-1388.
2. Kullak-Ublick GA, Fisch T, Oswald M, et al. Dehydroepiandrosterone sulfate (DHEAS): identification of a carrier protein in human liver and brain. FEBS Lett. 1998;424:173-176.
3. Kusuhara H, Sugiyama Y. Efflux transport systems for drugs at the blood-brain barrier and blood-cerebrospinal fluid barrier (Part 1). Drug Discov Today. 2001;6:150-156.
4. Kusuhara H, Sugiyama Y. Efflux transport systems for drugs at the blood-brain barrier and blood-cerebrospinal fluid barrier (Part 2). Drug Discov Today. 2001;6:206-212.
5. Vergani P, Basso C, Mense M, Nairn AC, Gadsby DC. Control of the CFTR channel’s gates. Biochem Soc Trans. 2005;33(Pt 5):1003-1007.
6. Herget M, Tampe R. Intracellular peptide transporters in human - compartmentalization of the “peptidome.” Pflugers Arch. 2006;May 18 [Epub ahead of print].
7. Pauli-Magnus C, Lang T, Meier Y, et al. Sequence analysis of bile salt export pump (ABCB11) and multidrug resistance p-glycoprotein 3 (ABCB4, MDR3) in patients with intrahepatic cholestasis of pregnancy. Pharmacogenetics. 2004;14:91-102.
8. Fernandez-Marmiesse A, Salas A, Vega A, Fernandez-Lorenzo JR, Barreiro J, Carracedo A. Mutation spectra of ABCC8 gene in Spanish patients with hyperinsulinism of infancy (HI). Hum Mutat. 2006;27:214.
9. Gueugnon F, Volodina N, Taouil JE, et al. A novel cell model to study the function of the adrenoleukodystrophy-related protein. Biochim Biophys Res Commun. 2006;341:150-157.
10. Bekri S, Kispal G, Lange H, et al. Human ABC7 transporter: gene structure and mutation causing X-linked sideroblastic anemia with ataxia with disruption of cytosolic iron-sulfur protein maturation. Blood. 2000;96:3256-3264.
11. Stenrii S, Battistella S, Fermo I, et al. De novo deletion removes a conserved motif in the C-terminus of ABCA4 and results in cone-rod dystrophy. Clin Chem Lab Med. 2006;44:533-537.
12. Michaelides M, Hardcastle AJ, Hunt DM, Moore AT. Progressive cone and cone-rod dystrophies: phenotypes and underlying molecular genetic basis. Surv Ophthalomol. 2006;51:232-258.
13. Cirrito JR, Deane R, Fagan AM, et al. P-glycoprotein deficiency at the blood-brain barrier increases amyloid-beta deposition in an Alzheimer disease mouse model. J Clin Invest. 2005;115:3285-3290.
14. Stefkova J, Poledne R, Hubacek JA. ATP-binding cassette (ABC) transporters in human metabolism and diseases. Physiol Res. 2004;53:235-243.
15. Redzic ZB, Biringer J, Barnes K, et al. Polarized distribution of nucleoside transporters in rat brain endothelial and choroidplexus epithelial cells. J Neurochem. 2005;94:1420-1426.
Protéines de transport membranaires : un défi pour le développement des médicaments du SNC

Les transporteurs de médicaments (drug transporters) sont des protéines situées dans les membranes cellulaires de différents tissus comme les lymphocytes, l’intestin, le foie, les reins, les testicules, le placenta ou le SNC (système nerveux central). Ces transporteurs jouent un rôle primordial dans l’absorption et la distribution des médicaments dans les organes cibles, surtout s’ils sont protégés par des structures comme la barrière hématoencéphalique (BHE) ou la barrière maternofoetale. Contrairement aux transporteurs liés à des récepteurs ou à la transmission interneuronale, ces transporteurs membranaires ne sont pas directement impliqués dans une fonction neuronale spécifique mais assurent une protection globale du système nerveux central. Le transport de médicaments est la résultante des mécanismes d’efflux (sortie) et d’ influx (entrée) situés au niveau de la membrane apicale et basolatérale des cellules endothéliales des cellules cérébrales et du plexus choroidien. En régulant l’efflux et l’influx des substances endogènes ou exogènes, la BHE et la barrière hémato-cérébro-spinale représentent les principales interfaces entre le système nerveux central et le sang. La distribution d’un médicament vers les organes cibles est dépendante de son affinité pour un système de transport spécifique : les transporteurs membranaires sont ainsi reconnus comme un élément clé dans la disposition des médicaments au niveau du système nerveux central. Plusieurs transporteurs membranaires font partie de la superfamille ABC (ATP-Binding Cassette) ou SLC (Solute Carriers). La Multi-Drug Resistance protein MDR1 (ABCB1) nommée aussi P-glycoprotéine, les Multidrug Resistance-associated Proteins MRPI (ABCC1) et MRP2 (ABCC2), ainsi que la Breast Cancer Resistance Protein BCRP (ABCG2) sont des transporteurs d’efflux ATP-dépendants présents au niveau de la BHE et qui appartiennent à la superfamille des transporteurs ABC. Les OATPs (Organic Anion Transporter Proteins), OCTs (Organic Cation Transporters) et OATs (Organic Anion Transporters) font partie des transporteurs ATP-indépendants de type SLC constitués de polypeptides (solute carriers). Les conséquences cliniques des transporteurs membranaires semblent jouer un rôle de plus en plus important dans le développement de nouveaux médicaments, particulièrement en neurologie et en psychiatrie. Les différents mécanismes qui régulent le transfert de molécules à travers les membranes cellulaires et les barrières organiques sont des éléments clés pour expliquer les mécanismes d’échec ou de résistance au traitement, ainsi que certains effets indésirables et variations individuelles à la réponse thérapeutique attendue.
34. Soontornmalai A, Vlaming ML, Frischy JM. Differential, strain-specific cellular and subcellular distribution of multidrug transporters in murine choroid plexus and blood-brain barrier. *Neuroscience*. 2006;138:159-169.

35. Calatazzo C, Gelati M, Ciucani E, et al. Expression of drug resistance proteins Pgp, MRP1, MRP3, MRPS and GSTP1 in human glioma. *J Neurooncol.* 2005;74:113-121.

36. Bronger H, Konig J, Kopplow K, et al. ABCB drug efflux pumps and organic anion uptake transporters in human glioma and the blood-tumor barrier. *Cancer Res.* 2005;65:11419-11428.

37. Kubota H, Ishihara H, Langmann T, et al. Distribution and functional activity of P-glycoprotein and multidrug resistance-associated proteins in human brain microvascular endothelial cells in hippocampal sclerosis. *Epilepsy Res.* 2006;68:213-228.

38. Toornvliet R, van Berckel BN, Luurtsema G, et al. Effect of age on functional P-glycoprotein in the blood-brain barrier measured by use of (R)–(C)-verapamil and positron emission tomography. *Clin Pharmacol Ther.* 2006;79:540-548.

39. Sasonango L, Link JM, Muzi M, et al. Imaging P-glycoprotein transport activity in the blood-brain barrier with positron emission tomography. *Clin Pharmacol Ther.* 2005;77:503-514.

40. Hagenbuch B, Meier PJ. Organic anion transporting polypeptides of the OATP/SLC21 family: phylogenetic classification as OATP/SLCO superfamily, new nomenclature and molecular functional properties. *Pflugers Arch.* 2004;447:653-665.

41. Hediger MA, Romero MF, Peng J, Rolfs A, Takanaga H, Bruford EA. The ABCG family of solute carriers: physiological, pathological and therapeutic implications of human membrane transport proteins. Introduction. *Pflugers Arch.* 2004;447:465-468.

42. Hagenbuch B, Meier PJ. The superfamily of organic anion transporting polypeptides. *Biochem Biophys Res Acta.* 2003;1609:1-18.

43. Pizzagalli F, Hagenbuch B, Stelger B, Klenk U, Folkers G, Meier PJ. Identification of a novel human organic anion transporting polypeptide as a high affinity thyroxine transporter. *Mol Endocrinol.* 2002;16:2238-2246.

44. Galic S, Schneider HP, Broer A, Deitmer JW, Broer S. The loop between helix 4 and helix 5 in the monocarboxylate transporter MCT1 is important for activity of P-glycoprotein and multidrug resistance-associated proteins in the drug transporter gene ABCB1. *N Engl J Med.* 2003;348:1442-1448.

45. Verstuyft C, Schwab M, Schaeffeler E, et al. Digoxin pharmacokinetics and MDR1 genetic polymorphisms. *Eur J Clin Pharmacol.* 2003;58:809-812.

46. Roberts RL, Joyce PR, Mulder RT, Begg EJ, Kennedy MA. A common P-glycoprotein polymorphism is associated with nortriptyline-induced postural hypotension in patients treated for major depression. *Pharmacogenomics.* 2002;2:191-196.

47. Illmer T, Schuler US, Thiede C, et al. MDR1 gene polymorphisms affect therapy outcome in acute myeloid leukemia patients. *Cancer Res.* 2002;62:4955-4962.

48. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

49. Marzolini C, Tirona RG, Kim RB. Pharmacogenomics of the OATP and OAT family transporters. *Clin Pharmacol Ther.* 2002;71:18-24.

50. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

51. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

52. Verstuyft C, Schwab M, Schaeffeler E, et al. Digoxin pharmacokinetics and MDR1 genetic polymorphisms. *Eur J Clin Pharmacol.* 2003;58:809-812.

53. Roberts RL, Joyce PR, Mulder RT, Begg EJ, Kennedy MA. A common P-glycoprotein polymorphism is associated with nortriptyline-induced postural hypotension in patients treated for major depression. *Pharmacogenomics.* 2002;2:191-196.

54. Illmer T, Schuler US, Thiede C, et al. MDR1 gene polymorphisms affect therapy outcome in acute myeloid leukemia patients. *Cancer Res.* 2002;62:4955-4962.

55. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

56. Marzolini C, Tirona RG, Kim RB. Pharmacogenomics of the OATP and OAT families. *Pharmacogenetics.* 2004;5:273-282.

57. Wakasugi H, Yano I, Marsh S, et al. Diffomotocan pharmacokinetics in relation to ABCG2 421C>A genotype. *Clin Pharmacol Ther.* 2004;76:38-44.

58. Fujita T, Brown C, Carlson EJ, et al. Functional analysis of polymorphisms in the organic anion transporter, SLC22A6 (OAT1). *Pharmacogenet Genomics.* 2005;15:201-209.

59. Kerb R, Brinkmann U, Chatskaia N, et al. Identification of genetic variations of the human organic cation transporter OCT1 and their functional consequences. *Pharmacogenetics.* 2002;12:591-595.

60. Leabman MK, Huang CC, Kawamoto M, et al. Polymorphisms in a human kidney xenobiotic transporter, OCT2, exhibit altered function. *Pharmacogenetics.* 2002;12:495-496.

61. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

62. Marzolini C, Tirona RG, Kim RB. Pharmacogenomics of the OATP and OAT families. *Pharmacogenetics.* 2004;5:273-282.

63. Wakasugi H, Yano I, Marsh S, et al. Diffomotocan pharmacokinetics in relation to ABCG2 421C>A genotype. *Clin Pharmacol Ther.* 2004;76:38-44.

64. Fujita T, Brown C, Carlson EJ, et al. Functional analysis of polymorphisms in the organic anion transporter, SLC22A6 (OAT1). *Pharmacogenet Genomics.* 2005;15:201-209.

65. Kerb R, Brinkmann U, Chatskaia N, et al. Identification of genetic variations of the human organic cation transporter OCT1 and their functional consequences. *Pharmacogenetics.* 2002;12:591-595.

66. Leabman MK, Huang CC, Kawamoto M, et al. Polymorphisms in a human kidney xenobiotic transporter, OCT2, exhibit altered function. *Pharmacogenetics.* 2002;12:495-496.

67. Yamauchi A, Ieiri I, Kataoka Y, et al. Neurotoxicity induced by tacrolimus after liver transplantation: relation to genetic polymorphisms of the ABCB1 (MDR1) gene. *Transplantation.* 2002;74:571-572.

68. Marzolini C, Tirona RG, Kim RB. Pharmacogenomics of the OATP and OAT families. *Pharmacogenetics.* 2004;5:273-282.

69. Wakasugi H, Yano I, Marsh S, et al. Diffomotocan pharmacokinetics in relation to ABCG2 421C>A genotype. *Clin Pharmacol Ther.* 2004;76:38-44.

70. Fujita T, Brown C, Carlson EJ, et al. Functional analysis of polymorphisms in the organic anion transporter, SLC22A6 (OAT1). *Pharmacogenet Genomics.* 2005;15:201-209.

71. Kerb R, Brinkmann U, Chatskaia N, et al. Identification of genetic variations of the human organic cation transporter OCT1 and their functional consequences. *Pharmacogenetics.* 2002;12:591-595.
84. Boyd RA, Stern RH, Stewart BH, et al. Atorvastatin coadministration may increase digoxin concentrations by inhibition of intestinal P-glycoprotein-mediated secretion. J Clin Pharmacol. 2000;40:91-98.

85. Eneroth A, Astrom E, Hoogstraate J, et al. Evaluation of a vincristine resistant Caco-2 cell line for use in a calcein AM extrusion screening assay for P-glycoprotein interaction. Eur J Pharm Sci. 2001;12:205-214.

86. Frohlich M, Albermann N, Sauer A, Walter-Sack I, Haefeli WE, Weiss J. In vitro and ex vivo evidence for modulation of P-glycoprotein activity by progestins. Biochem Pharmacol. 2004;68:2409-2416.

87. Sadeque AJ, Wandel C, He H, Shah S, Wood AJ. Increased drug delivery to the brain by P-glycoprotein inhibition. Clin Pharmacol Ther. 2000;68:231-237.

88. Wang JS, DeVane CL, Gibson BB, Donovan JL, Markowitz JS, Zhu HJ. Population pharmacokinetic analysis of drug-drug interactions among risperidone, bupropion, and sertraline in CF1 mice. Psychopharmacology (Berl). 2006;183:490-499.

89. Keogh JP, Kunta JR. Development, validation and utility of an in vitro technique for assessment of potential clinical drug-drug interactions involving P-glycoprotein. Eur J Pharm Sci. 2006;27:543-554.

90. Koren G, Woodland C, Ito S. Toxic digoxin-drug interactions: the major role of renal P-glycoprotein. Vet Hum Toxicol. 1998;40:45-46.

91. Leabman MK, Huang CC, Kawamoto M, et al. Polymorphisms in a human kidney xenobiotic transporter, OCT2, exhibit altered function. Pharmacogenetics. 2002;12:395-405.

92. Dresser GK, Schwarz UI, Wilkinson GR, Kim RB. Coordinate induction of both cytochrome P4503A and MDR1 by St John’s wort in healthy subjects. Clin Pharmacol Ther. 2003;73:41-50.

93. Dresser GK, Spence JD, Bailey DG. Pharmacokinetic-pharmacodynamic consequences and clinical relevance of cytochrome P450 3A4 inhibition. Clin Pharmacokinet. 2000;38:41-57.