Association of trauma exposure with proinflammatory activity: a transdiagnostic meta-analysis

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Exposure to psychological trauma (for example, childhood/early life adversity, exposure to violence or assault, combat exposure, accidents or natural disasters) is known to increase one’s risk of developing certain chronic medical conditions. Clinical and population studies provide evidence of systemic inflammatory activity in trauma survivors with various psychiatric and nonpsychiatric conditions. This transdiagnostic meta-analysis quantitatively integrates the literature on the relationship of inflammatory biomarkers to trauma exposure and related symptomatology. We conducted random effects meta-analyses relating trauma exposure to log-transformed inflammatory biomarker concentrations, using meta-regression models to test the effects of study quality and psychiatric symptomatology on the inflammatory outcomes. Across k = 36 independent samples and n = 14,991 participants, trauma exposure was positively associated with C-reactive protein (CRP), interleukin (IL)-1β, IL-6, and tumor necrosis factor (TNF)-α (mean r = 0.2455, 0.3067, 0.2890, and 0.2998, respectively). No significant relationships were noted with fibrinogen, IL-2, IL-4, IL-8, or IL-10. In meta-regression models, the presence of psychiatric symptoms was a significant predictor of increased effect sizes for IL-1β and IL-6 (β = 1.0175 and 0.3568, respectively), whereas study quality assessment scores were associated with increased effect sizes for IL-6 (β = 0.3812). Positive correlations between inflammation and trauma exposure across a range of sample types and diagnoses were found. Although reviewed studies spanned an array of populations, research on any one specific psychiatric diagnosis was generally limited to one or two studies. The results suggest that chronic inflammation likely represents one potential mechanism underlying risk of health problems in trauma survivors.

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INTRODUCTION

Chronic inflammation may be a hallmark of many chronic diseases, including cardiovascular disease (CVD), diabetes, and chronic pain disorders, among others. A history of exposure to traumatic events (for example, early life adversity, exposure to violence or assault, combat exposure, accidents, or natural disasters) is known to increase one’s risk of developing chronic medical problems,1,2,4 and research on inflammatory biomarkers has begun to elucidate some of the potential mechanisms underlying this increased risk.

Physiological mechanisms linking the experience of psychological stressors to immune functioning are complex, with influences exerted through various pathways. Briefly, in response to a traumatic stressor, the biological stress systems (including sympathetic/parasympathetic, catecholamine, hypothalamic-pituitary-adrenal axis, and immune system components) assist in promoting adaptive behavioral and physiological responses to the stressor.2,3 Severe, repeated and prolonged exposure to traumatic stressors, however, can lead to chronic dysregulation of these basic biological systems. Chronic, systemic inflammation has been posited as one mechanism underlying psychiatric symptomatology, across a range of disorders,6-8 as well as with increased risk of many physical health problems.2,9,10

Historically, research into the physiological mechanisms occurring within the context of psychopathology has been segregated according to diagnostic classification. However, despite differing symptom presentations, mounting evidence of neurobiological, genetic, and physiological mechanisms underlying a range of physical and psychological disorders has led to increasing recognition that current diagnostic classifications may no longer provide an adequate framework for psychobiological research or for translating this research into clinical practice.11,12 This awareness has led to transdiagnostic initiatives such as the US National Institutes of Mental Health (NIMH) Research Domain Criteria project.13 Indeed, studies of immune activity across multiple psychiatric disorders, including posttraumatic stress disorder (PTSD),5 major depression,14 and bipolar disorder15,16 have all identified disruptions in proinflammatory cytokines (such as interleukin (IL)-6, tumor necrosis factor (TNF)-α, and IL-1β), among symptomatic individuals, as compared with healthy control participants. In addition, a convergence of evidence has identified lifetime trauma exposure, particularly early life adversity, as a major risk factor for a range of chronic physical and psychiatric conditions,1,2 and systemic inflammation has been suggested as one potential mechanism mediating this association.2,4,5,17

Despite a rapidly growing body of literature on the relationships between trauma exposure and inflammatory biomarkers (including cytokines and acute-phase proteins, such as C-reactive protein (CRP) and fibrinogen) in both clinical and nonclinical samples, few

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attempts have been made to quantitatively integrate this research. To our knowledge, all existing meta-analyses have been limited to disorder-specific comparisons of symptomatic vs asymptomatic individuals, such as in depressive disorders,14,18–21 or bipolar disorder.15,16 In addition to the narrow focus on specific psychiatric diagnoses, prior meta-analyses have faced methodological challenges, including skewed biomarker concentrations in primary studies, disparate statistical techniques used to evaluate data, and systematic exclusion of large, population-based regression studies, which are often better equipped to statistically control for covariates, such as body mass index (BMI), age, sex, and the use of medications or other substances, all of which have previously been identified as important in biobehavioral research.22 Using meta-analysis and meta-regression models, the present study therefore describes the relationships between trauma history and in vivo inflammatory biomarkers (that is, cytokines and acute-phase proteins) from a transdiagnostic perspective.

**MATERIALS AND METHODS**

**Protocol**

This study adhered to PRISMA guidelines for meta-analysis.23 Search strategy and data extraction were informed by a preliminary review of the literature and further specified on the basis of data availability and methodological variation, including limiting the present analysis to unstimulated in vivo inflammatory biomarkers (that is, cytokines and acute-phase proteins). Inclusion/exclusion criteria To address the relevant transdiagnostic theoretical questions, we included all studies of adult participants that analyzed unstimulated, in vivo (circulating) inflammatory biomarkers in blood samples (that is, cytokines, CRP, or fibrinogen) in relation to measures of trauma exposure. We excluded child and adolescent samples due to evidence that cytokine production in children differs substantially from that of adults, even among healthy populations.24–26 Trauma exposure was defined either through self-report measures of trauma or abuse (for example, Adverse Childhood Experiences questionnaire26) or by exposure to events meeting criterion A for PTSD.27 Studies that only included pre-trauma cytokine measurements were excluded. The statistical analyses assessed the impact of trauma exposure, and meta-regression models assessed the impact of relevant covariates, including study quality and the presence or absence of psychiatric symptomatology within the samples. As trauma exposure can be assessed either as a continuous or dichotomous variable, our inclusion criteria were intentionally inclusive of either design. Because only a small number of studies had adequate data to test the relative contribution of posttraumatic symptomatology within trauma-exposed populations (that is, comparing trauma-exposed symptomatic individuals to trauma-exposed controls), such analyses were underpowered and the therefore reported solely as supplemental analyses (see Supplementary Material). Thus, we chose to exclude studies that did not include either a continuous measure of trauma exposure or a non-trauma-exposed comparison group.

**Study selection**

Studies were identified through systematic searches of PubMed, PsycINFO, PILOTS (Published International Literature on Traumatic Stress), and Ovid MEDLINE databases. Search criteria included peer-reviewed articles published in English, using terms related to inflammation (for example, interleukin, cytokine) and either traumatic event exposure (for example, trauma* stress, child* maltreatment) or psychiatric conditions commonly linked to trauma exposure (for example, PTSD, depression), limiting the results to human samples. Additional selected references were identified through limited update searches, citations in other papers, and personal communications with authors (see Supplementary Material for full search strategy).

**Study characteristics and data extraction**

**Data extraction.** A single effect size estimate was calculated from each fully independent sample of participants for each biomarker. Potential duplicate publications were identified (within each biomarker outcome) through cross-checking authors’ names and sample locations. Evaluation of inclusion/exclusion criteria and extraction of relevant data were conducted systematically by one of two coders, with a selection evaluated by both coders. Any disagreements (less than 5%) regarding eligibility or extracted data were settled by consensus.

Effect size and other study data were extracted from each of the published reports. Forms were piloted and revised as needed for extraction of relevant information. All assessments were made at the outcome level, with separate biomarkers analyzed independently. In the cases for which published data were insufficient, at least three attempts were made to contact authors. In total, the authors of 38 studies were contacted requesting information about sample independence, study eligibility or effect size calculation. Response rate was 76%, although some of these studies were not included in the present analyses (due to inclusion/exclusion criteria).

**Data preparation and statistical analysis**

**Effect size preparation.** A single effect size was calculated for each biomarker measured in an independent sample. Seven articles were identified by study authors as one or more to be duplicate samples, and a single report contained two independent subsamples (that is, African American and Caucasian participants), which were treated individually. Due to inclusion of both continuous and dichotomous predictors, we used correlation coefficients to synthesize the literature.29

As inflammatory biomarker distributions tend to be skewed, authors generally take one of three approaches to statistical testing and reporting of data. Use of nonparametric statistical methods, dichotomization according to clinically-relevant cut points (most frequently CRP = 3 mg/L30), and logarithmic transformations to normalize the distributions before using parametric statistics. Because data are log-transformed before statistical aggregation (individual data are converted into an exponent-scale before taking an arithmetic mean), effects based on log-transformed data cannot be mixed in the same analyses as raw (non-transformed) data. Log-transformed biomarker data are more likely to meet assumptions of normality than raw data; therefore, we chose to convert all raw effect sizes to estimated log2-transformed effect sizes,31 which were then converted to a log10 scale. When log-transformations were applied, any zero values were set to the smaller of the assay detection limit or a raw value of 1.0 (corresponding to a log-transformed value of 0), to allow for transformation and avoid artificially inflating the effect size estimate. Effect sizes for one study,32 for which we were unable to obtain effect size estimates by usual means, were extrapolated from a published scatterplot of individual participant data using WebPlotDigitizer, version 2.6 (http://arohatgi.info/WebPlotDigitizer/). Other effect size conversions were performed according to standard methods.29,33,34

**Synthesis of results.** We produced meta-analytic models for all analyses including at least three independent effect size estimates. Correlation coefficients were converted to Fisher's Z-values for analyses and back-converted to correlations for interpretation. All meta-analysis and meta-regression models were fitted using Wilson's meta-analysis macros for SPSS.15,35,36 Because we expected at least a moderate level of heterogeneity across studies, we used non-iterative method of moments random-effects models to integrate study findings. This approach produces wider confidence intervals and, thus, a more conservative estimate than either fixed- or other random-effects models.37 Forest plots were created,37 and heterogeneity analyses were conducted using the Q-test and I2 statistic.38

**Publication bias and sensitivity analyses.** Publication bias was assessed using funnel plots relating effect size to precision (inverse of standard error), two-tailed rank correlation tests,39 and trim and fill techniques.40 using the Comprehensive Meta-Analysis software, version 2.0. To provide protection against Type I error, publication bias was assessed only in those analyses which consisted of at least five studies. To evaluate the stability of results, we conducted sensitivity analyses by excluding samples consisting primarily of patients with known psychiatric conditions (for example, CVD, migraine, pregnant samples), as the inflammatory processes could differ from those in the general population, despite study-level
control. These post hoc models were fitted if there remained at least two studies for the analysis.

**Study-level risk of bias.** Risk of study-level bias was assessed with a checklist modeled after the Quality Assessment Tool for Quantitative Studies (QAT), modified slightly to fit our observational research question (see Supplementary Materials for details). We calculated both a categorical global rating (that is, high, moderate or low risk of bias), and an average QAT score (average of the ordinal domain ratings). For each study, the percentage of recommended covariates (for example, age, sex, BMI, and the use of medications or other substances, among others) controlled either methodologically (that is, through exclusion or matching of subjects) or statistically was documented and entered as part of the QAT score (see Supplementary Material). Each domain was scored on a three-point likert scale (strong/moderate/weak); thus, both categorical and average QAT scores ranged between 1.0 and 3.0, with higher scores indicating greater potential for bias.

**Meta-regression models.** For each analysis, we tested the effect of average QAT scores using univariate models. We then conducted multivariate meta-regression models assessing the impact of a study’s inclusion of symptomatic individuals, while statistically controlling for study-level bias. Finally, we ran a series of dummy-coded meta-regression models to assess the effect of psychiatric diagnosis or symptom type. Due to the small number of studies of non-PTSD psychiatric populations, we used models comparing samples of non-comorbid PTSD participants to all other samples (including those using other psychiatric diagnoses and non-psychiatric samples). Any models that displayed significant differences between the two groups (PTSD vs other samples) were then subjected to post hoc analyses to determine which symptom subgroups differed.

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**Figure 1.** Study screening and eligibility.

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One study contained two independent samples, so 36 samples from 35 studies were included.
Table 1. Studies included in the meta-analysis

| Study                  | Total N | Group n (T/NT) | Country     | Type of study (Sample name) | Risk of bias | %F Age (years) | Sample type | Biomarkers included | Biomarkers excluded |
|------------------------|---------|----------------|-------------|-----------------------------|--------------|----------------|--------------|---------------------|---------------------|
| Spitzer, et al.⁴²       | 3047    | 1653/1394      | Germany     | Population study (SHIP): T/NT | Low          | 52             | Serum CRP   | CRP                 | IL-18               |
| Baumert, et al.⁴³       | 3012    | 1224/1788      | Germany     | Population study (KORA): T/NT | Low          | 52             | Blood, N/S CRP | CRP                 | IL-18               |
| Cho, et al.⁴⁴           | 2716    | (Regress.)     | USA         | Population study (CARDIA): T/NT | Low          | 55             | Range Plasma IL-6, CRP | IL-18               |
| O'Donovan, et al.⁴⁵     | 979     | (Regress.)     | USA         | Stable CVD: trauma           | Low          | 19             | 67 (11) Serum IL-6, CRP | IL-6, TNF-α, CRP | Resistin            |
| Danese, et al.⁴⁶        | 886     | 315/551        | New Zealand | Prospective birth cohort (Dunedin): T/NT | Low          | 47             | All 32 years Serum Blood, N/S CRP | IL-6, CRP fibrogen | IL-18               |
| Slopen, et al. (C)⁴⁸    | 822     | (Regress.)     | USA         | Population study, subsample (MIDUS): ELA/no ELA | Low          | 53             | 59 (12) Serum CRP, IL-6, CRP | IL-6, CRP fibrogen | IL-18               |
| Bertone-Johnson, et al.⁴⁷ | 700    | 452/248        | USA         | Population study (Nurses' health study): T/NT | Low          | 100            | 44 (5) Plasma IL-6, CRP | IL-6, CRP TNFR2     | IL-18               |
| Rooks, et al.⁴⁸         | 482     | 245/237        | USA         | Population (VET twin registry): T/NT | Low          | 0              | 55 (3) Plasma IL-6, CRP | IL-6, CRP          | IL-18               |
| Plantinga, et al.⁴⁹     | 476     | 108/368        | USA         | Population (VET twin registry): T/NT | Low          | 0              | 56 (3) Plasma Fibrinogen IL-6, CRP | IL-6, CRP fibrogen | IL-18               |
| Slopen, et al. (AA)⁵⁰  | 177     | (Regress.)     | USA         | Population study (AA) (MIDUS + Mike): T/NT | Low          | 67             | 54 (11) Serum CRP, IL-6, fibrogen | IL-6, CRP fibrogen | IL-18               |
| Tietjen, et al.⁵¹       | 141     | 90/51          | USA         | Migraine patients (+HC): T/NT | Moderate      | 100            | 33 (7) Blood, N/S CRP | IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α, IFN-α | IL-18               |
| Smith, et al.⁵²         | 138     | 95/43          | USA         | Primary care/GYN: ELA/no ELA | Moderate      | 50             | 46 (11) Plasma IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α | IL-18               |
| Blackmore, et al.⁵³     | 137     | 50/80          | USA         | Pregnant women: T/NT         | Low          | 100            | 25 (4) Serum IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α | IL-18               |
| Gouin, et al.³⁶         | 130     | 57/73          | USA         | Dementia caregivers: ELA/no ELA | High         | 82             | 65 (13) Serum IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α | IL-18               |
| Guo, et al.⁵⁴           | 100     | 50/50          | China       | PTSD/NTC                     | High         | 53             | 42 (13) Serum IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α | IL-18               |
| Carpenter, et al.⁵⁵     | 92      | 21/71          | USA         | ELA/no ELA                   | Moderate      | 51             | 31 (9) Plasma CRP | IL-6, CRP          | IL-18               |
| Gill, et al.⁵⁶          | 77      | 53/24          | USA         | 26 PTSD, current; 27 PTSD, recovery: T/NT | Low          | 100            | 34 (8) Plasma IL-6, CRP | IL-6, CRP          | IL-18               |
| Danielson, et al.⁵⁷     | 75      | 40/35          | Canada      | Undergraduate students: current IPV/ no current IPV | Moderate      | 100            | 20 (2) Plasma IL-6, IL-10 | IL-6, CRP          | IL-18               |
| Cankaya, et al.⁵⁸       | 75      | 64/11          | USA         | Primary care: traumatic loss/ no loss | Low          | 100            | > 40 Serum IL-6, CRP | IL-6, CRP TNF-α | IL-18               |
| Dennison, et al.⁵⁹      | 72      | 24/48          | Ireland     | Schizophrenia: 24 PSY+ELA; 16 PSY – ELA: 32 NTC | High         | 54             | 37 (10) Plasma IL-1, IL-6, IL-8, TNF-α | IL-18               |
| Tucker, et al.⁶⁰        | 68      | 54/14          | USA         | Hurricane or mixed trauma: T/NT | High         | 35             | 36 (12) Serum IL-2, IL-6 | IL-6, CRP          | IL-18               |
| Heppul, et al.⁶¹        | 63      | 25/38          | UK          | First episode psychosis: 10 PSY+ELA; 15 HC+ELA; 18 PSY – ELA; 20 HC – ELA | High         | 29             | 27 (2) Plasma CRP | IL-6, CRP          | IL-18               |
| Cohen, et al.⁶²         | 61      | 48/13          | Israel      | Prospective musculoskeletal injury: injured/NTC | Moderate      | 28             | 35 (13) Serum IL-4, IL-6, IL-8, IL-10 | TGF-β               | IL-18               |
| Gola, et al.⁶³          | 60      | 22/25          | Germany     | PTSD± MDD/HC                  | Low          | 55             | 31 (10) Plasma IL-6, IL-8, IL-10, TNF-α | MCP-1, IL-1, IL-2, IL-4, IL-6, IL-10, TNF-α, IL-13, IL-15, IFN-γ, IP-10-others | IL-18               |
| Hoge, et al.⁶⁴          | 56      | 28/28          | USA         | PTSD/HC                      | Moderate      | 50             | 41 (11) Plasma IL-6, CRP, TNF-α | IL-18               |
| Grassi-Oliveira, et al.⁶² | 49     | 30/19          | Brazil      | MDD outpatients/HC: ELA regress. | Moderate      | 100            | 38 (8) Plasma TNF-α | TNFR1, TNFR2 | IL-18               |
| Maes, et al.⁶⁵          | 45      | 13/32          | Belgium     | PTSD/HC                      | Moderate      | 58             | 38 (8) Plasma TNF-α | TNFR1, TNFR2 | IL-18               |
| Janusek, et al.⁶⁶       | 40      | (EUA regres.) | USA         | Breast cancer                | High          | 80             | 46 (8) Serum IL-6 | IL-6, CRP, TNF-α | IL-1, IL-2, IL-4, IL-6, IL-8, IL-10, TNF-α, IL-15, IFN-γ, IP-10-others | IL-18               |
| Symes, et al.⁶⁷         | 39      | 25/14          | USA         | CVD: IPV/no IPV              | High          | 100            | 57 (NR) Serum IL-6, CRP, TNF-α | IL-18               |
| Spivak, et al.⁶⁸        | 38      | 19/19          | Israel      | PTSD± MDD/NTC                 | Moderate      | 0              | 29 (11) Serum IL-1β | sIgG2 | IL-6, CRP, TNF-α | IL-18               |
| Gill, et al.⁶⁹          | 36      | 22/14          | USA         | PTSD± MDD/NTC                 | Moderate      | 42             | 36 (12) Plasma IL-6, CRP | IL-6, CRP | IL-18               |
| Groer, et al.⁷⁰         | 31      | 15/16          | USA         | Sexual assault crisis center: raped/HC | Moderate      | 100            | Range Serum IL-6, IL-10, CRP, IFN-γ | TNFR1, TNFR2 | IL-18               |
| Pace, et al.⁷¹          | 28      | 14/14          | USA         | MDD+ELA/NTC                  | Moderate      | 0              | 30 (9) Plasma IL-6 | IL-6, CRP | IL-18               |
significantly from the PTSD samples. To increase power and interpretability, all meta-regression models were fitted only if at least four samples were included in the analysis.

RESULTS

Study selection

A total of 3647 unique articles were identified through our searches, the majority of which were excluded through title or abstract screening (n = 2919). Reasons for exclusion of full-text articles are shown in Figure 1. We identified 40 independent samples measuring 29 different cytokines, receptors or acute-phase proteins that met initial inclusion criteria. Of these, only biomarkers measured by at least three samples were included in our analyses. Three studies that failed to provide sufficient information to calculate log-transformed effect size estimates were not included. Thus, nine biomarkers from 36 independent samples were included in the meta-analysis (see Table 1).

None of the meta-analysis models for the anti-inflammatory cytokines (IL-4 and IL-10) was statistically significant (see Supplementary Figure 1), so we focus on presenting analyses of the acute-phase proteins (CRP and fibrinogen) and the proinflammatory cytokines (IL-1β, IL-2, IL-6, IL-8, and TNF-α).

The results of the baseline models, based on 36 independent samples, are presented in Figure 2. As expected, there was evidence of significant heterogeneity across studies for all biomarkers. Although rank correlation tests revealed a significant correlation between the standardized effect size and the standard error for IL-6 (r = 0.2963, z = 2.1889, P = 0.0302), no imputed ‘missing studies’ were identified using the trim and fill method.46

Acute phase proteins

CRP. Analyses for CRP included 16 studies and a total of 13 374 participants. As shown in Figure 2, a significant association between CRP concentration and trauma exposure was detected (mean r = 0.2507, P = 0.0030).

Fibrinogen. Analysis of four studies involving 1890 participants showed no significant correlation between trauma exposure and fibrinogen (mean r = 0.0675, P = 0.1860; see Figure 2).

Proinflammatory cytokines

IL-1β. Meta-analysis of four studies (304 participants) showed a significant relationship between IL-1β and trauma exposure (mean r = 0.3169, P = 0.0322), displayed in Figure 2.

IL-2. Overall analyses of four studies (362 participants) revealed no statistically significant association between IL-2 and trauma exposure (mean r = 0.3627, P = 0.1256; see Figure 2).

IL-6. As shown in Figure 2, analysis of 26 studies with 7295 participants showed a significant relationship between trauma exposure and IL-6 (mean r = 0.3029, P < 0.0001).

IL-8. No significant overall correlation relating trauma exposure to IL-8 was detected, based on five studies with 349 participants (mean r = 0.4649, P = 0.1609; see Figure 2).

TNF-α. As presented in Figure 2, TNF-α was significantly associated with trauma exposure in analysis of 11 studies with 1899 participants (mean r = 0.2998, P = 0.0288).

Meta-regression models and sensitivity analyses

As shown in Table 2, risk of study-level bias (defined as the average of the applicable QAT domains: selection bias, study
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**Acute Phase Proteins**

|        | Total N | $Q$  | $I^2$ | Mean $r$ | Lower 95% | Upper 95% | $p$   |
|--------|---------|------|-------|----------|-----------|-----------|-------|
| CRP    | 16      | 13,374 | 1200.8312 | 99.98%  | .2507     | .0854     | .4159 | .0030 |
| Fibrinogen | 4 | 1890 | 9.6126 | 90.26%  | .0675      | -.0325    | .1659 | .1860 |

**Proinflammatory Cytokines**

|         | Total N | $Q$  | $I^2$ | Mean $r$ | Lower 95% | Upper 95% | $p$   |
|---------|---------|------|-------|----------|-----------|-----------|-------|
| IL-1β   | 4       | 304  | 17.1445 | 96.94%  | .3169     | .0269     | .6070 | .0322 |
| IL-2    | 26      | 362  | 56.0579 | 99.71%  | .3627     | -.1015    | .8269 | .1256 |
| IL-6    | 7       | 295  | 382.5809 | 93.47%  | .3094     | .1974     | .4084 | <.0001 |
| IL-8    | 5       | 349  | 144.6798 | 99.92%  | .4649     | -.1851    | 1.1148 | .1609 |
| TNF-α   | 11      | 1,899 | 228.8252 | 99.85%  | .2998     | .0310     | .5687 | .0288 |

* $p$ ≤ .05, † $p$ ≤ .001, ‡ $p$ ≤ .0001
$k$ = Number of studies included in analysis; Total $N$ = total number of participants

Figure 2. Meta-analysis summary statistics.

Design, covariate control, assessment reliability/validity, and statistical analysis) was a significant predictor of heterogeneity in meta-regression models for the proinflammatory cytokines IL-6 and IL-8, such that higher risk of bias was associated with larger effect size estimates. In models controlling for risk of bias, studies that included participants with psychiatric disorders yielded larger effects for IL-1β and IL-6.

None of the meta-regression models comparing PTSD samples with all other samples was statistically significant, with the exception of fibrinogen (which did not include any psychiatric samples) and IL-6 (see Supplementary Table 2). Post hoc comparisons coding non-PTSD psychiatric samples separate from asymptomatic samples revealed no differences in effect sizes for IL-6 between non-comorbid PTSD vs other types of psychiatric disturbance.

Sensitivity analyses showed that the findings were robust to exclusion of medical samples, which did not significantly change the pattern or significance of findings for any of the biomarkers except TNF-α (see Supplementary Figure 1). Although of increased magnitude, the TNF-α mean effect size was no longer statistically significant, once medical samples were excluded ($k = 6$, mean $r = 0.4579$, $P = 0.0857$).

**DISCUSSION**

To our knowledge, this is the first meta-analysis of trauma exposure as a risk factor for inflammation to utilize a transdiagnostic perspective. We integrated all available studies measuring trauma exposure, rather than limiting the analysis to a single diagnostic category. Further, to reflect the true status of the current literature, we incorporated both continuous and categorical measures. Although this procedure is likely to increase the observed heterogeneity in effect sizes across studies, it is essential to represent the diversity of study designs to obtain an accurate and comprehensive review of inflammation in trauma survivors. A growing body of evidence suggests that there may be a dose-response relationship between physical health problems and increased levels of exposure to traumatic events. Within the field of trauma research, however, it has been difficult to arrive at a single adequate method of quantifying cumulative trauma exposure, taking into account differences in number, frequency and severity of potentially traumatic experiences. As a result, there was insufficient literature to assess the potential impact of continuous vs categorical measures or to assess variability across different continuous measures of trauma exposure.

As expected, increased inflammation in trauma-exposed samples was found across a range of biomarkers. In particular, we noted moderate-to-large correlations relating trauma exposure to circulating concentrations of the proinflammatory cytokines IL-1β, IL-6, and TNF-α and of the acute-phase protein CRP. On the basis of our meta-regression models, the relationship between trauma exposure and the proinflammatory cytokines IL-1β and IL-6 was especially pronounced in samples that included, at least in part, clinical populations, as compared with samples which did not specifically recruit symptomatic individuals. No significant differences were observed as a function of the specific psychiatric diagnosis, although the statistical power was likely insufficient, at least in most analyses, to detect such relationships if they exist.

Because it was not possible to obtain log-transformed effect sizes for all studies, we produced log-transformed effect size estimates following accepted techniques. When we were unable to confirm with authors the base of the logarithm applied, we assumed the use of log-10 transformations, in accordance with similar studies. In keeping with prior recommendations for biomarker research, we advocate for greater consistency in reporting of methods, including transformations applied, sample demographics and characteristics, and covariates controlled in statistical analyses. Although it is beyond the scope of this study to recommend specific statistical techniques, other authors have endorsed the use of log-transformation when data are log-normally distributed, due to its flexibility in allowing for the use of traditional (parametric) statistical techniques and due to the ability to back-transform (exponentiate) geometric means to facilitate interpretation and comparison.
Despite recent interest in these questions, research on the relationships among trauma exposure, psychiatric symptomatology, and inflammatory biomarkers has historically been limited to the PTSD literature. Thus, our power to detect differences between diagnostic groups was somewhat limited by a relative scarcity of reporting or measurement of trauma histories in non-PTSD psychiatric samples. Given the evidence that trauma exposure, particularly when prolonged and/or occurring early in life, is associated with a wide range of chronic medical and psychological health problems, even after accounting for the effects of psychiatric disorders, it would be useful for future research to routinely include measures of trauma exposure in both medical and psychiatric populations.

Our analyses were also limited by the number of published biomarker studies providing information on participants’ trauma history. Although the numbers of studies included in our meta-analyses are comparable with other early meta-analyses on inflammation and psychiatric disorders, further research is necessary to determine whether replicable relationships exist between trauma exposure and those biomarkers with relatively few published studies (for example, fibrinogen, IL-1β, IL-2, and IL-8).

Our findings suggest that the risk of study-level bias, especially related to the control of relevant covariates, such as medication use, BMI, and comorbid medical conditions, was significantly related to heterogeneity of effect sizes observed across studies. That is, higher QAT scores (indicating greater risk of bias) were associated with larger correlations between trauma exposure and those biomarkers with relatively few published studies (for example, fibrinogen, IL-1β, IL-2, and IL-8).

Another important question concerns the longitudinal course of inflammation in trauma-exposed individuals. We were unable to evaluate the effects of treatment or to examine the longitudinal course of inflammation following traumatic events in our analyses. However, there is preliminary evidence to suggest that successful treatment of trauma-related psychopathology may lead to improvements in the chronic immune dysregulation observed in our analyses, although results to date have been somewhat contradictory and unclear. This lack of consistency may be due, at least in part, to methodological differences among studies. Further research is needed to better understand the time course of inflammatory biomarkers following successful psychological or pharmacological treatment.

### CONCLUSIONS

The relevance of inflammation in the pathophysiology and consequences of psychiatric disorders and general medical conditions has been increasingly recognized within research, clinical and public health arenas. The results of this meta-analysis are in keeping with a growing body of cross-disciplinary evidence which provides a framework for examination of transdiagnostic relationships among psychiatric risk factors (such as trauma exposure) and both psychological and physiological dysfunction. In our review of 36 samples with 14 991 participants, we found moderate correlations between inflammatory biomarker concentrations (IL-1β, IL-6, TNF-α, and CRP) and trauma exposure (mean \( r = 0.2455, 0.3067, 0.2890, \) and 0.2998, respectively) across 36 independent samples with a total of 14 991 participants. Further research is needed to confirm this association in a broader range of psychiatric and general medical populations, and to determine whether these findings extend to other inflammation-related biomarkers.

Although prior systematic reviews on inflammatory biomarkers in PTSD have provided a qualitative synthesis of the literature, to our knowledge, this is the first meta-analysis to examine the relationship of trauma to proinflammatory cytokines...
and acute-phase proteins. Meta-analyses of inflammatory activity observed within other psychiatric disorders (for example, depression\textsuperscript{1,4,18–21} and bipolar disorder\textsuperscript{15,16}) have found evidence of systemic inflammation, but none have yet examined the impact of trauma on these relationships. Our findings are also consistent with the results of two recent studies\textsuperscript{9,61} (included within our analysis), which found significantly higher inflammatory biomarkers only in those psychotic patients with a trauma history, as compared with healthy control participants. In both studies, patients without a trauma history did not significantly differ from controls. However, the majority of studies in psychiatric populations that we screened did not assess participants’ histories of trauma exposure, so further research is needed to confirm and clarify these findings. We therefore endorse routine assessment and reporting of trauma exposure within immunological research studies.

**CONFLICT OF INTEREST**

The authors declare no conflict of interest.

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