An Atlas to Support the Progressive Control of Tsetse-transmitted Animal Trypanosomosis in Burkina Faso

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Abstract

Background

African animal trypanosomosis (AAT), transmitted by tsetse flies, is arguably the main disease constraint to integrated crop-livestock agriculture in sub-Saharan Africa, and African Heads of State and Government have adopted a resolution to rid the continent of this scourge. In order to sustainably reduce or eliminate the burden of AAT, a progressive and evidence-based approach is need, which must hinge on harmonized, spatially-explicit information on the occurrence of AAT and its vectors.

Methods

A digital repository containing tsetse and AAT data collected in Burkina Faso between 1990 and 2019 was assembled. Data were collected either in the framework of control activities or for research purposes. Data were systematically verified, harmonized, georeferenced and integrated into a database (PostgreSQL). Entomological data on tsetse were mapped at the level of individual monitoring trap. When this was not possible, mapping done was at the level of site or location. Epidemiological data on AAT were mapped at the level of location/village.

Results

Entomological data showed that presence of four tsetse species in Burkina Faso. *Glossina tachinoides* was the most widespread and abundant species (56.35% of the catches), present from the eastern to the western part of the country. *Glossina palpalis gambiensis* was the second most abundant species (35.56%), and it was mainly found in the West. *Glossina morsitans submorsitans* was found at lower densities (6.11%), with a patchy distribution in the southern parts of the country. One only cluster of *G. medico* was detected (less than 0.1%), located in the Southwest. For the AAT component, data for 54,948 animal blood samples were assembled from 218 geographic locations. The samples were tested with a variety of diagnostic methods. AAT was found in all surveyed departments, including the tsetse-free areas in the North. *Trypanosoma vivax* and *T. congolense* infections were the dominant species 6.11±21.56% and 5.19±18.97% respectively, and to a lesser extend *T. brucei* infections (0.00±0.10%).

Conclusions

The atlas provides a synoptic view of the available information on tsetse and AAT distribution in Burkina Faso. Data are very scanty for most of the tsetse-free areas in the northern part of the country. Despite this limitation, this study generated a robust tool to aid the targeting of future surveillance and control activities. The development of the atlas also strengthened the collaboration between the different institutions involved in tsetse and AAT research and control in Burkina Faso, which will be crucial for future updates and the sustainability of the initiative.

Background

Tsetse flies are the cyclical vectors of African trypanosomosis, a parasitic disease that affects both humans (sleeping sickness) [1] and animals (nagana) [2]. These diseases have historically hindered the development of the African continent [3], and they persist today as a major constraint to mixed crop-livestock agriculture, food security and human health [4]. The annual economic losses attributed to African animal trypanosomosis (AAT) are measured in billions of dollars [5, 6], while more than 50 million people are still considered at risk of contracting human African trypanosomosis (HAT) [7, 8].

In Burkina Faso, the expenditure on the trypanocides used for the preventive and curative treatment of livestock, mainly cattle, have been estimated to be 3.9 million US$ per annum [9]. Two million and six hundred thousand doses of trypanocidal drugs are used every year to control the disease [9, 10], and 9 out of the 13 Regions of the country are affected by tsetse flies. More than 60% of the country's cattle population is raised in zones with high trypanosomosis risk [11, 12]. In particular, many zones where grazing capacities for the cattle are high and human activities are limited, also provide suitable habitats for tsetse flies [12].

In 2000, the Government of Burkina Faso embarked with the other African countries in the Pan African Tsetse and Trypanosomosis Eradication Campaign (PATTEC), and initiative led by the African Union [13]. At the country level, this first phase of the campaign was implemented between 2006 and 2013, and used an integrated approach combining insecticide treated targets (ITT), insecticide-treated cattle (ITC), the sequential aerosol technique (SAT) and mass livestock treatment with trypanocides [14]. The project generated a vast amount of data, while other activities and projects implemented by other stakeholders also collected field data on the occurrence of AAT and its vectors in Burkina Faso. Unfortunately, the data generated by these activities and institutions have not been harmonized and centralized.

National atlases of tsetse and AAT are considered indispensable tools for efficient disease management in the context of the progressive control pathway for AAT [15]. In 2013, the Food and Agriculture Organization of the United Nations (FAO) launched a multi-national project to support six AAT-affected countries (i.e. Burkina Faso, Mali, Ghana, Kenya, Ethiopia and Uganda), and one of its goals was to promote the development of the national atlases. The Insectary of Bobo Dioulasso – Tsetse and Trypanosomosis Eradication Campaign (Insectarium de Bobo Dioulasso-
Methods

The methodology to develop the atlas of tsetse and AAT in Burkina Faso is broadly based on the FAO continental atlas [16–18], and a similar approach has already been used at the national level in Sudan [19], Mali [20], Kenya [21] and Zimbabwe [22]. The main difference between the continental and the national atlases is that the former is solely based on publicly available data from peer-reviewed journals, while the latter include all published or unpublished data collected in a given country.

The atlas in Burkina Faso covers a period of 30 years (1990–2019). Data were provided by all the institutions involved in tsetse and trypanosomosis research and control at the national level. In addition to IDB-CETT, these are the School of tsetse control (Ecole de lutte anti-tsé-tsé – ELAT), the International Research and Development Centre on Livestock farming in the Subhumid Zone (Centre international de recherche pour le développement sur l’Élevage en zone Subhumide – CIRDES), the French Agricultural Research Centre for International Development (Centre de coopération internationale en recherche agronomique pour le développement – CIRAD), the French Research Institute for Development (Institut de recherche pour le développement – IRD), the International Atomic Energy Agency (IAEA) and FAO.

Input data

Input data were collected either in the framework of tsetse and trypanosomosis control operations or in the context of research activities. Control operations, mostly carried out, provided a large amount of unpublished data, including both baseline cross-sectional surveys (i.e. pre-intervention) and longitudinal surveys (i.e. during or post-intervention). The raw data from research activities were obtained from the authors of published articles, using as a basis the systematic review of the literature carried out by FAO for the continental atlas. The full list of publications that contributed to the national atlas in Burkina Faso are provided in Additional file 1: Text_S1. When the raw data underpinning publications could not be retrieved, information on AAT and tsetse occurrence was extracted directly from the papers, as done for the continental atlas.

Tsetse data

In Burkina Faso, entomological surveys are mainly carried out with biconical traps [23]; boosted or not with odour attractants [24]. However, other traps, such as monoconical Vavoua trap [25] [26, 27], N’Zi traps, adhesive targets and E-targets were also occasionally used in experimental settings [28]. The trapping period is usually 3 days for the surveys carried out in the framework of control activities, and 1 to 3 days for research activities. All traps were geo-referenced with GPS.

Entomological data are recorded at the level of individual trap. Data are initially captured in hard-copy recording sheets, which are subsequently converted into digital format (e.g. Microsoft Excel or Access). Field recording sheets include information such as the name of the surveyed location, its coordinates and administrative units (which in Burkina Faso are called Regions, Provinces and Departments), the date of the survey, the time of trap deployment and removal (and the corresponding duration of trapping), and the number, species and sex of trapped tsetse flies. Complementary information (e.g. type of trap, use of odour attractants, etc.) is normally available in mission or project reports. For research activities linked to scientific publications, raw datasets at the trap level were normally obtained. When these were not available, information was mapped at the site level.

African animal trypanosomosis data

As for tsetse, AAT data are mainly collected in the framework of control activities, thus including both baseline and monitoring/longitudinal investigations. Supplementary data are generated in the context of research.

AAT surveys in Burkina Faso target the main susceptible animal species such as cattle, sheep, goats and donkeys, but data on horses and camels are also occasionally collected. In most surveys, the sampling locations and the animals are selected randomly. However, a non-random sampling is sometimes performed, for example in studies of trypanocidal drug resistance [29, 30], or for the isolation of trypanosomes. Cross-sectional surveys provided the majority of datasets included in the atlas, but longitudinal investigations are also included [14].

Field data recording sheets for AAT include the data source and the institution in charge, the name, geographical coordinates and administrative units pertaining to the survey village or site, the date of the survey and the sample size (i.e. the number of animals tested). The sheets also include parasitological results such as the number of AAT-positive animals by trypanosome species, the haematocrit (packed cell volume - PCV) and the possible use of trypanocidal drugs (including the type of drug).

AAT data are recorded at the level of individual animal in the parasitological recording sheets, and information at the animal level was retained in the atlas. When the animal level data were not available, aggregated information at the herd/site level was recorded.
Structure of the atlas

The atlas is composed of a structured data repository and two databases (tsetse distribution and AAT distribution respectively). Data on tsetse infections are not yet included. Open-source PostgreSQL is used as the main database management system, together with its spatial extension PostGIS. Microsoft Access © is used to facilitate a broader data utilization.

Data repository

The data repository includes digital copies of all input files used to build the atlas, i.e., spreadsheets, databases, reports, thesis and scientific articles. At a first level, the repository is organized by type of data (i.e. tsetse or AAT), at a second level by geographical area (i.e. province), and last by individual data source or publication. File names are standardized as author name and year of publication for scientific publications (i.e. Surname_YEAR.pdf) and as title of activity and year for unpublished works.

Tsetse database

Entomological data recorded in the tsetse database are organized in six tables: 1) data sources; 2) geographical data; 3) entomological surveys (trap level); 4) tsetse catches (trap level); 5) entomological surveys (site level); and 6) tsetse catches (site level). As in all relational databases, records in each table are given a unique identifier, the ‘primary key’, which allows them to be linked to records in other tables through ‘foreign keys’.

The table on ‘data sources’ summarizes information on the input files stored in the repository. In particular, it records the following information: institution, author’s name, title, year of production/publication, presence of information on tsetse or AAT, availability of raw data and whether the source is published or not. The ‘geographical data’ table includes the location or village name, its geographical coordinates, and the corresponding administrative units. The table on ‘entomological surveys’ (trap level) includes the code of the trap (as per survey recording sheet), the type of trap, the attractant used (if any), the geographical coordinates of the trapping site (latitude and longitude in decimal degrees on WGS84 datum), the trapping period and its duration. Information on the possible presence of tsetse control activities in the surveyed area is also recorded. The table on ‘tsetse catches’ (trap level) summarizes the results of the trapping and it includes the tsetse species, sex and the number of flies caught.

The tables on ‘entomological surveys’ and ‘tsetse catches’ (site level) record data that were directly extracted from scientific publications when raw data at the trap level could not be obtained. These site-level tables of the national atlas for Burkina Faso have a similar structure to those of the continental atlas [17].

The detailed structure of the tsetse database, including the relationships between the tables, is provided in Additional file 2: Text_S2.

African animal trypanosomosis database

The database for AAT contains seven tables: 1) data sources; 2) geographical data; 3) animal species; 4) diagnostic method; 5) AAT data (animal level); 6) AAT data (herd level); and 7) chemotherapy.

The tables ‘data sources’ and ‘geographical data’ are similar to those of the tsetse database. The table ‘animal species’ records the species and breed of tested animals, and the table ‘diagnostic method’ provides details on the diagnostic test used to detect AAT. The table ‘AAT data’ (animal level) includes the code of the animal (as per survey recording sheet), its age, sex, haematocrit/PCV, health status, treatment with trypanocidal drugs, date of last treatment, husbandry system, date of survey, presence or absence of trypanosomes, species of trypanosomes (including subspecies/groups), type of animal sampling (random or purposeful), type of survey (longitudinal or cross-sectional) and possible presence of vector control operations during the survey. The table ‘AAT data’ (herd level) is used to capture data extracted from scientific publications when the raw data at the animal level could not be obtained; as such, it follows the structure of the continental atlas [16]. The last table (chemotherapy) provides details on the possible treatment of animals with anti-trypanosomal drugs before the survey. It includes the date and cause of treatment, the drug used, the doses and the person who administered the treatment.

The detailed structure of the AAT database, including the relationships between the tables, is provided in Additional file 3: Text_S3.

Development of the atlas

The first step in the development of the atlas was to collect input data from all partner institutions. Workshops and meetings were organized to facilitate the process. When input data could only be retrieved as hard copy, Microsoft Excel © was used to enter data in digital format.

Merging data from a variety of sources into the atlas databases required a systematic process of verification and harmonization. Geographical coordinates were standardized as latitude and longitude (decimal degrees on WGS84 datum). When Universal Transverse Mercator (UTM) coordinates were reported in the input files, coordinate transformation was performed. Extensive harmonization was also needed for data on
animal breeds, husbandry systems, names of geographical locations and the related administrative units. For tsetse data, the names of locations were often missing. For AAT data, a high proportion of epidemiological records lacked geographic coordinates; in these cases, the names of the survey sites were used to extract geographic coordinates from other sources (i.e. Geographical Institute of Burkina, Google Earth and Google Maps). Also, input data sheets often lacked information on whether interventions against tsetse were taking place in the study area at the time of the survey, or whether trypanocidal drugs were being used. In these cases, reports and publications were often useful to fill the gaps. When raw data could not be retrieved, information was extracted directly from the papers; for example, the maps included in the publication were imported in the GIS and georeferenced to estimate the geographical coordinates of the study sites.

Results

A total of 2261 data recording sheets were included in the atlas repository (i.e. 723 for the tsetse component and 1538 for the AAT component). The review of the published and grey literature also enabled to identify 105 eligible documents, including scientific papers, PhD and MSc thesis and activity reports.

The results of the atlas are summarized in Fig. 1, which shows the locations where tsetse flies and AAT were detected in Burkina Faso in the period 1990–2019. Locations where surveys were conducted but tsetse and AAT were not detected are not shown in Fig. 1, although they are included in the database of the atlas.

Tsetse fly distribution

Entomological data on tsetse fly distribution were assembled for a total of 19,960 tsetse trapping events in 10,022 distinct trapping locations, including 9768 locations at the trap level and 254 locations at the site level. ‘Site’ locations normally refer to groups of traps, and they were used to map data from publications when the underlying trap-level data could not be obtained. The overall trapping intensity was 48,345 trap days.

The bulk of the entomological data included in the atlas are from the western and south-western part of the country. In particular, intensive baseline and monitoring surveys were conducted between 2007 and 2013 in the region at the Loop of the Mouhoun River (‘Boucle du Mouhoun’), in the framework of an attempted large-scale elimination campaign [24, 14]. Entomological surveys are limited in most of the other regions, and especially in the north-eastern parts of the country that are believed to be free of tsetse.

The device most commonly used to trap flies was the biconical trap [23], which was used in more than 95% of the surveys and often boosted with an odour attractant (77.31% of traps). Other trapping devices used were sticky targets [27], E-targets [28] and Nzi traps [31].

A total of 126,849 tsetse flies were caught in all the surveys included in the atlas, representing all three taxonomical groups (subgenera) historically present in Burkina Faso. The vast majority of catches belong to the palpalis group (91.91%), in particular *G. palpalis gambiensis* (35.56%) and *G. tachinoides* (56.35%). Flies of the morsitans group were represented by *G. morsitans submorsitans*, which accounted for 6.11% of the overall catches. *G. medicorum* of the fusca group accounted for less than 0.1% of the catches. Approximately 1.34% of flies collected were not identified to the species level.

In terms of geographic distribution, *G. palpalis gambiensis* and *G. tachinoides* were found to be widespread in the western part of the country (Fig. 2), in particular along the major river systems (Mouhoun, Bourgouriba and Comoe) and their main tributaries. In the eastern part of the country *G. palpalis gambiensis* was detected in only one location of Arly National Park, while *G. tachinoides* was found in several locations, and especially in protected areas (e.g. Nazinga game ranch and Arly National Park) [32]. The distribution of flies of the morsitans and fusca groups was more circumscribed. *Glossina morsitans submorsitans* was present in few locations in the southern parts of the country, and in particular in protected areas, whereas *G. medicorum* was only found in one protected area in the South-East (Diefoula Forest and Logonieugue Forest).

African animal trypanosomosis distribution

In the AAT component of the atlas, results of AAT diagnosis for 54,948 animal’s blood sample were included, originating from 218 distinct geographic locations. Cattle account for 90.37% (n = 49,654) of animals tested, followed by sheep (5.20 %; n = 2859), goats (2.29 %; n = 1258) and donkeys (2.10 %; n = 1154). Very limited data was found on horses (n = 21) and camels (n = 2).

AAT was confirmed to be present in all the tsetse-infested areas, in particular in the southern and south-western parts of the country. Hotspots of transmission appear to be located in the areas bordering Côte d’Ivoire and Mali, and in the Mouhoun River basin. To a lesser extent, transmission occurs in the eastern part of the country, especially in the areas bordering Togo and Benin near the Arli National Park (Fig. 3). Serological prevalence of the disease was also found in some of the tsetse-free areas in the northern part of the country [33], at a distance of over 200 km from the tsetse-infested belt [32]. However, very limited surveillance is carried out in these tsetse-free areas, and the true extent of the AAT problem in these regions cannot be ascertained with the available data.
Buffy Coat Technique (BCT) [34] was the mostly diagnostic method used for Trypanosomal infections identification (62.59 % of all samples, n = 34,390), followed by molecular technique, in particular PCR (8.78%, n = 4825) [35]. Serological techniques, more specifically ELISA was most often used on BCT positive sample (28.63% of all samples, n = 15,733) [12, 36] (Fig. 4). Trypanosomosis prevalence was variable among hosts and diagnostic techniques. For cattle, an overall prevalence of 6.06% for BCT (1881/31,048), 45.83% for ELISA (6316/13,781) and 30.86% for PCR (1489/4825) was found (Table 1).

Three species of trypanosomes pathogens for livestock, Trypanosoma vivax, T. congolense and T. brucei were found. Parasitological techniques showed an overall prevalence of 5.58% (1921 infections/34390), 30.68% (1489 infection/4825) for molecular techniques while serological technique showed an overall prevalence of 41.90% (6592 infection/15,733). T. vivax and T. congolense were the most common species responsible for AAT. The parasitological prevalence by BCT for T. vivax was 6.11 ± 21.56% and 5.19 ± 18.97% (673 infection) for T. congolense. Trypanosoma brucei was detected at smaller rates (0.00 ± 0.10; 7 infections). With PCR, the prevalence were 13.98 ± 22.71% and 9.16 ± 19.29% for T. vivax and T. congolense. The prevalence of T. brucei was 0.35 ± 2.24%.

### Table 1

| Diagnostic method | Animal Species | Number of infection [n] | Mean of prevalence [%] | Standard Deviation [%] |
|-------------------|----------------|-------------------------|------------------------|------------------------|
|                   |                | Tc  | Tv  | Tb  | T | Tc  | Tv  | Tb  | T | Tc  | Tv  | Tb  | T |
| BCT               | Donkey        | 673 | 5   | 7   | 0 | 12 | 0.74 | 1.84 | 0.00 | 2.58 | 4.82 | 10.70 | 0.00 | 11.79 |
|                   | Cattle        | 31,048 | 712 | 642 | 7 | 1881 | 7.86 | 6.43 | 0.00 | 13.69 | 24.26 | 21.15 | 0.11 | 30.54 |
|                   | Camels        | 1   | 0   | 0   | 0 | 0   | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
|                   | Goats         | 801 | 2   | 6   | 0 | 8   | 0.10 | 0.40 | 0.00 | 0.50 | 0.78 | 2.82 | 0.00 | 3.28 |
|                   | Horses        | 17  | 0   | 0   | 0 | 0   | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
|                   | Sheep         | 1850 | 2   | 18  | 0 | 20  | 0.05 | 0.97 | 0.00 | 1.03 | 0.84 | 4.92 | 0.00 | 5.05 |
|                   | **Subtotal**  | 34,390 | 721 | 673 | 7 | 1921 | 6.11 | 5.19 | 0.00 | 10.94 | 21.56 | 18.97 | 0.10 | 27.64 |
| PCR               | Cattle        | 4825 | 591 | 605 | 29 | 1489 | 9.16 | 13.98 | 0.35 | 22.16 | 19.29 | 22.71 | 2.24 | 28.87 |
| Elisa             | Donkey        | 481 | 7   | 26  | 6 | 29  | 1.28 | 5.51 | 1.46 | 5.99 | 5.16 | 15.20 | 6.75 | 15.29 |
|                   | Cattle        | 13,781 | 2444 | 4258 | 1069 | 6316 | 24.39 | 40.00 | 11.85 | 47.21 | 32.93 | 35.20 | 24.68 | 35.70 |
|                   | Camels        | 1   | 1   | 1   | 1 | 1   | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
|                   | Goats         | 457 | 5   | 25  | 16 | 43  | 0.82 | 4.84 | 2.13 | 7.48 | 3.51 | 13.95 | 6.09 | 15.76 |
|                   | Horses        | 4   | 0   | 0   | 0 | 0   | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
|                   | Sheep         | 1009 | 98  | 162 | 48 | 203 | 6.01 | 9.48 | 2.27 | 12.40 | 15.22 | 23.68 | 7.56 | 25.81 |
|                   | **Subtotal**  | 15,733 | 2555 | 4472 | 1140 | 6592 | 18.78 | 31.17 | 9.26 | 37.03 | 30.28 | 34.94 | 21.97 | 36.55 |
| Total             | Donkey        | 1154 | 12  | 33  | 6 | 41  | 0.96 | 3.32 | 0.59 | 3.95 | 4.96 | 12.80 | 4.33 | 13.39 |
|                   | Cattle        | 49,654 | 3747 | 5505 | 1105 | 9686 | 13.04 | 17.44 | 3.64 | 24.51 | 27.80 | 30.37 | 14.67 | 35.31 |
|                   | Camels        | 2   | 1   | 1   | 1 | 1   | 50.00 | 50.00 | 50.00 | 50.00 | 70.71 | 70.71 | 70.71 | 70.71 |
|                   | Goats         | 1258 | 7   | 31  | 16 | 51  | 0.39 | 2.18 | 0.85 | 3.29 | 2.32 | 9.33 | 3.98 | 10.82 |
|                   | Horses        | 21  | 0   | 0   | 0 | 0   | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
|                   | Sheep         | 2859 | 100 | 180 | 48 | 223 | 2.32 | 4.21 | 0.86 | 5.36 | 9.83 | 15.65 | 4.79 | 17.29 |
| **General Total** | 54,948 | 3867 | 5750 | 1176 | 10,002 | 10.47 | 14.32 | 3.02 | 20.12 | 25.24 | 28.14 | 13.23 | 33.05 |

Tc: T. congolense, Tv: T. vivax, Tb: T. brucei, T: Number of animals infected; [n]: Number; [%]: Prevalence

### Database completeness

A high level of completeness was achieved in the atlas database. Regarding the tsetse component, the required information items could be found for the vast majority of the entomological records, in particular trap coordinates (83.57%), administrative units (100%) [i.e. region, province, department], location names (89.69%), date of trapping (99.52%), duration of trapping (97.25%), type of trap (99.25%), tsetse species (98.65%),...
tsetse apparent densities (97.25%), and control interventions against tsetse (76.34%). The majority of trap coordinates were in UTM (75.72%), so they had to be converted into geographical coordinates (Decimal Degrees). Regarding the AAT component, a similarly high level of completeness was achieved for geographical coordinates (87.12%), administrative units (90.72), location names (90.49%), survey period (97.40%), sample size (100%), number and species of infected animals (84.89%), sampling strategy (i.e. random or purposeful) (95.93%), chemotherapy (67.77%) and possible interventions against tsetse (84.37).

Discussion

Our systematic review of the available published and unpublished data provided a synoptic picture of the distribution of tsetse fly and AAT in Burkina Faso for the period 1990–2019. The atlas updates the latest national tsetse distribution map produced in 1977 [37], whose northern limit had been updated in 2010 [32]. Detailed distribution maps of the four tsetse species present in Burkina Faso are also presented. As far as AAT is concerned, the distribution maps presented in this paper, based on the testing of over 50,000 animals, are the first of their kind in Burkina Faso.

However, as observed in similar national mapping exercises [19–21, 18, 22], a number of gaps not attributable to the data compilation and processing still affect our knowledge of the distribution of tsetse fly and AAT in Burkina Faso. The gaps are related to the inherent limitations of the existing datasets, and in particular to the geographical coverage, range of animal species tested and diagnostic methods.

With regard to the geographical coverage, the main gap in the atlas is the severe lack of information from the north, north-east, north-west, centre and extreme east of the country. This situation is explained by the fact that institutions involved in tsetse and AAT research and control in Burkina Faso focus their investigations and efforts on the regions of highest tsetse infestation [14, 38]. As already observed in Mali [20], this lack of information on a large part of the national territory hides the true extent of the AAT problem in the country.

As regards the animal species tested for AAT, more than 90% of atlas data are on cattle. However, in Burkina Faso small ruminants (i.e. sheep and goats) far exceed cattle in terms of population numbers. Furthermore, the diagnostic technique used to test blood samples was the BCT technique for more than 60% of the samples. While parasitological techniques based on blood concentration may be appropriate for treatment decisions [39], they are known to have low sensitivity and to underestimate the true prevalence of the disease [36].

Despite the limitations, the atlas provides a fairly accurate assessment of the geographic distribution of three tsetse species of veterinary importance in Burkina Faso: Glossina palpalis gambiensis, Glossina tachinoides, and Glossina morsitans submorsitans. Glossina medicorum, because of its limited distribution, has a very low impact on AAT transmission in Burkina. A fifth species, G. longipalpis, had been reported in Burkina Faso, in particular in the South West Region in the zone of Batié [37]. However, the last observation was in 1961 [37], while recent data from this region are insufficient to rule out its possible persistence.

Thanks to their adaptation capacity to land cover changes and vegetation degradation, tsetse species of the riverine group (i.e. palpalis) are confirmed to be the most widespread in Burkina Faso [37]. In particular, the geographical range of G. tachinoides extends from the East to the West of the country, and the species is now found also in some previously free areas (e.g. western part of the Orodara Department) [37].

As regards savannah species, their distribution is fragmented and confined to a few protected areas. This patterns has been observed in other countries in western Africa [18], as a result of habitat degradation, land cover changes and the depletion of the wild fauna, their preferred hosts [40]. Glossina morsitans submorsitans is found along the southern border between Burkina Faso and Ghana and Benin, as well as in the pastoral zones in the western part of the country (Samorogouan and Sideradougou). Glossina medicorum was reported from one province in the south-western part of the country (Comoé), while it had been previously reported also along the borders with Ghana, Togo and Benin [37].

Overall, our study seems to corroborate the notion that in recent years the distribution of tsetse species in Burkina Faso has been affected by droughts and demographic pressure [41].

Regarding animal trypanosomosis, the three major trypanosome species infective for livestock (T. vivax, T. congolense and T. brucei) are widespread in the country. Serologically positive animals, not confirmed parasitologically, have been reported also from the north of the country (Djibo Department) [33]. The absence of parasitological confirmations in these areas is ascribed to the combined effect of the absence of cyclical vectors and the trypanocidal drug treatments administered by livestock keepers before, during and after transhumance in the AAT enzootic area [33]. In this context, it is unclear whether any of the serological positive animals identified in the North area have been infected locally.

In Burkina Faso, T. vivax is the most widespread trypanosome species follow by T. congolense. This could be related to the ability of T. vivax to be mechanically transmitted by Stomoxys and Tabanids [42]. Mechanically transmitted of T. congolense is also well-know [42]. These parasites are also transmitted mechanically by tsetse flies [43, 42] and especially by many other haematophagous insects [44, 42]. In some contexts, these mechanical transmissions act as an amplifying factor for the disease [45–47].
*T. brucei* has been found confined in protected areas of the country, in the west, south-west and Centre-east. These infested areas are economic interest to the country, including pastoral areas, a park and a reserve. These areas would be inhabited by wild or domestic animals that are reservoirs of the disease.

The highest serological prevalence show in this study indicates that many animals have been in contact with the trypanosomes even from animals of tsetse free area.

The AAT prevalence obtained in this study may be underestimated because of that most of the diagnosis methods used was by BCT.

**Conclusion**

Our study showed that animal trypanosomosis is widespread in Burkina Faso, even though tsetse flies are mainly confined to the West, South-West and East of the country. The disease therefore poses constraints to the development of the livestock and agricultural sectors at the national level. The prevalence of AAT is highest in the west, which is considered the breadbasket of the country. The newly developed national atlas represents a valuable tool to support the progressive control of AAT at the national level, in particular to target and monitor surveillance and control activities. The development of the atlas also helped to develop capacities for data management of all the involved partners. It has promoted synergies among national institutions for data collection and sharing, and the establishment of mechanisms for the regular updating and upgrading of the atlas. With a view to filling the present geographical gaps, AAT data collection could include the East, North-east and the Centre where data are not available. Entomological surveys could also include the Batié area update the status of *G. longipalpis*. Additional work may be carried out to include tsetse fly infections into the atlas.

**Abbreviations**

AAT: African animal trypanosomosis; AfDB: African Development Bank; BCT: buffy coat technique; I PCR: polymerase chain reaction ; IDB-CETT: Insectarium de Bobo-Dioulassi – Campagne d’Eradication de la Lutte contre les Mouches tsé-tsé et de la Trypanosomose (Coordination Unit for Tsetse and Animal Trypanosomosis Control); INERA: Institut National de l’Environnement et de la Recherche Agronomique; FAO: Food and Agriculture Organization of the United Nations; IRD: Institut de recherche pour le développement; CIRAD: Centre de coopération internationale en recherche agronomique pour le développement; CIRDES: Centre International de Recherche-Développement sur l’Elevage en zone Subhumide; ELAT: Ecole de Lutte anti-tsetse; GPS: Global Positioning System; HAT: human African trypanosomosis; IAEA: International Atomic Energy Agency; LNE: Laboratoire National d’Elevage; PATTEC: Pan-African Tsetse and Trypanosomosis Eradication Campaign; PCP: progressive control pathway; PCV: packed cell volume; UTM: Universal Transverse Mercator; WGS84: World Geodetic System 1984.

**Declarations**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Availability of data and materials**

Relevant data are within the paper and its Additional files. The bulk of the data on the tsetse and AAT occurrence in Burkina Faso is the property of the Government of Burkina Faso, Ministère des Ressources Animales et Halieutiques/Insectarium de Bobo-Dioulasso – Campagne d’Eradication de la Lutte contre les Mouches tsé-tsé et de la Trypanosomose, (IBD-CETT), and data can be requested to: Direction Generale, PO Box: 1087, Bobo-Dioulasso 01, Burkina Faso, Phone: +226 20 970955/971521, Website: [www.pattec.bf](http://www.pattec.bf)

**Competing interests**

The authors declare that they have no competing interests.

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Authors’ contributions

LP, JBR, MP, IS, MO and GC made substantial contributions to conception and design the study; LP, MP, SPH, MO, JBR, GG, SP, AS and GC participated to the data collection; LP, JBR, MP, and GC analysed the data; and LP, MP, JBR, GG, SP, SPH, IS and GC participated in the writing the manuscript. All authors read and approved the final manuscript.

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Figures

Figure 1

The occurrence of African animal trypanosomosis and tsetse flies (genus: Glossina) in Burkina Faso. Data collection period: 1990–2019
Figure 2

Presence and absence of capture of G. palpalis gambiensis, G. tachinoides and G. morsitans submorsitans and G. medicorum in Burkina Faso. Data collection period: 1990–2019.
Figure 3

Presence and absence of African Animal Trypanosomosis in Burkina Faso. Data collection period: 1990 –2019.
Figure 4

Presence and absence and prevalence of African Animal Trypanosomosis in Burkina Faso with different diagnostic methods. Data collection period: 1990 – 2019.

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