The Prognostic Role of SOCS3 and A20 in Human Cholangiocarcinoma

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Abstract

As an antagonist of the JAK/STAT pathway, suppressor of cytokine signaling 3 (SOCS3) plays an integral role in shaping the inflammatory environment, tumorigenesis and disease progression in cholangiocarcinoma (CCA); however, its prognostic significance remains unclear. Although tumor necrosis factor α-induced protein 3 (TNFAIP3, also known as A20) can decrease SOCS3 expression and is involved in the regulation of tumorigenesis in certain malignancies, its role in CCA remains unknown. In this study, we investigated the expression of SOCS3 and A20 in human CCA tissues to assess the prognostic significance of these proteins. The expression of SOCS3 and A20 was initially detected by western blot in 22 cases of freshly frozen CCA tumors with corresponding peritumoral tissues and 22 control normal bile duct tissues. Then, these proteins were investigated in 86 CCA patients by immunohistochemistry (IHC) and were evaluated for their association with clinicopathological parameters in human CCA. The results indicated that SOCS3 expression was significantly lower in CCA tumor tissues than in corresponding peritumoral biliary tissues and normal bile duct tissues. Conversely, A20 was overexpressed in CCA tissues. Thus, an inverse correlation between the expression of SOCS3 and A20 was discovered. Furthermore, patients with low SOCS3 expression or high A20 expression showed a dramatically lower overall survival rate. These proteins were both associated with CCA lymph node metastasis, postoperative recurrence and overall survival rate. However, only A20 showed a significant association with the tumor node metastasis (TNM) stage, while SOCS3 showed a significant association with tumor differentiation. Multivariate Cox analysis revealed that SOCS3 and A20 were independent prognostic indicators for overall survival in CCA. Thus, our study demonstrated that SOCS3 and A20 represent novel prognostic factors for human CCA.
Introduction

Cholangiocarcinoma (CCA) is the second most common primary hepatobiliary cancer, arising from the biliary tree with characteristic cholangiocyte differentiation, and epidemiological studies have shown that the incidence of CCA is increasing worldwide [1–4]. Complete surgical resection is still the most preferred and only possible curative treatment for this fatal disease [5]. Unfortunately, most patients are diagnosed at an unresectable stage, where the prognosis of CCA is notoriously poor [6]. Thus, the discovery of effective biomarkers for prognosis, with a view to define the molecular mechanisms underlying CCA tumor development and progression, remains an urgent need.

Chronic biliary inflammation is a confirmed risk factor for CCA, which thus represents a classic model disease to study the relationship between chronic inflammation and the initiation and progression of cancers [7, 8]. The JAK/STAT pathway has been shown to play an integral role in shaping the inflammatory environment of CCA and other cancers [9, 10]. The JAK/STAT pathway regulates a variety of vital processes including innate and adaptive immune function and embryonic development, as well as cell proliferation, differentiation and apoptosis [11], and its key role in regulating human biliary epithelial cell migration has been demonstrated in our prior studies [12].

The suppressors of cytokine signaling (SOCS) proteins function as cytokine signaling inhibitors of the JAK/STAT pathway. Thus far, there have been eight SOCS proteins identified, and these family members possess similar structures but differential mechanisms for inhibiting the JAK/STAT pathway. As part of a classical feedback loop, SOCS3 expression competes with STAT activation by inhibiting its phosphorylation, which is mediated by the stimulation of cytokines or growth factors. Moreover, SOCS3 binds to cytokine receptors that contain JAK-proximal sites, leading to JAK inhibition [13, 14]. Additionally, SOCS3 acts as a negative regulator in the activation of STAT3 and chronic inflammatory processes [15]. Loss of SOCS3 expression has been reported in a variety of malignancies due to epigenetic mechanisms, mostly promoter methylation [16–20]. In CCA, this mechanism was confirmed in an earlier study as well [21]. In liver, lung, and squamous head and neck cancer, as well as a number of hematological malignancies, SOCS3 functions as a classical tumor suppressor [21]. Our recent studies suggested that enhanced expression of SOCS3 could reduce tumor metastasis, the expression of epithelial-to-mesenchymal transition (EMT) markers and STAT3 activation in the absence of interleukin-6 (IL-6) stimulation in CCA cell lines [22]. Very little is known about SOCS3 expression in human CCA tissue and whether SOCS3 may serve as a novel prognostic biomarker for CCA patients.

A20, also known as tumor necrosis factor α-induced protein 3 (TNFAIP3), is a zinc-finger protein that plays a pivotal negative role in the regulation of inflammation and immunity [23]. It was recently discovered in liver regeneration and repair that A20 can increase JAK/STAT3 pro-proliferative signals by decreasing SOCS3 expression, most likely in a miR203-dependent manner; furthermore, A20 can reduce the levels of the cell cycle inhibitor p21 [24]. Moreover, A20 plays an oncogenic role, as indicated by A20 overexpression in several malignancies, such as undifferentiated nasopharyngeal carcinoma, poorly differentiated head and neck squamous cell carcinoma [25], glioma [26, 27], glioblastoma [28], inflammatory breast cancer [29], and hepatocellular carcinoma [30]. Overexpression of A20 in breast cancer cells contributes to resistance to TNFa and tamoxifen, indicating that A20 induces chemoresistance and survival [31]. Although the data above suggest a tumorigenic role for A20 overexpression, loss of A20 function is associated with multiple lymphomas, including B-cell lymphoma [32, 33], non-Hodgkin lymphoma [34], and Hodgkin lymphoma [35]. Taken together, these results have led several investigators to posit that A20 plays a contextual role in tumor biology that may be
tissue-type-dependent [27]. However, the role of A20 expression in human CCA remains unclear but pathophysiologically important.

The aim of the present study was to investigate the status of SOCS3 and A20 expression in human CCA tissue by western blot and immunohistochemistry (IHC), as well as their correlation with clinicopathological parameters. Survival analyses were performed to evaluate the prognostic relevance of these two biomarkers. Therefore, this study has momentous implications for further elucidating the molecular mechanisms of human CCA. Our data suggest that SOCS3 and A20 may serve as novel prognostic biomarkers for CCA patients.

Patients and Methods

Ethics statement

This study and all involved protocols were approved by the ethical committee of the Second Affiliated Hospital of Harbin Medical University. Specimens were obtained with written informed consent, which was signed by patients or their next of kin on behalf of the patients involved in this study.

Fresh-frozen tissue samples

Fresh tumor and corresponding peritumoral biliary tissues were collected from 22 CCA patients who underwent curative surgery and whose CCA was pathologically confirmed intraoperatively between November 2013 and October 2014 at the Second Affiliated Hospital of Harbin Medical University, Heilongjiang, China. Pathologists used frozen tissue examination during the operation to confirm CCA according to following criteria: tumor tissues contained at least 70% tumor cells, while the peritumoral tissues contained no tumor cells. In addition to the CCA specimens, we also obtained 22 normal biliary duct specimens as controls, which were collected from patients undergoing hepatectomy or pancreatoduodenectomy for non-tumor-related diseases (giant hepatic hemangioma, traumatic rupture of duodenum and pancreas trauma), where the selected biliary duct tissues were disease-free and no local inflammation was detected under gross and microscopic observation. All selected tissues were preserved in liquid nitrogen.

Western blot analysis

To determine the expression levels of SOCS3 and A20 in CCA, western blot analyses were carried out as previously described [36] for the 22 cases of CCA where freshly frozen tumors and their corresponding peritumoral tissues were available. A 100 mg sample was resected from each pair of tumor and peritumoral tissues and washed 3 times with ice-cold phosphate-buffered saline (PBS). The sample tissues were ruptured with RIPA buffer (50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate, 0.1% SDS) containing 100 mM phenylmethylsulfonyl fluoride (PMSF; catalog number P0013B, Beyotime Institute of Biotechnology, China) on ice for 30 min. The tissue extracts were centrifuged for 30 min at 14,000×g and 4°C, and the supernatants were collected. The tissue sample supernatants containing equal amounts of protein were resolved by 8% SDS-polyacrylamide gel electrophoresis. Then, the proteins were transferred to polyvinylidene difluoride (PVDF) membranes. These membranes were blocked with 5% skim milk in Tris-buffered saline containing 0.1% Tween-20 (TBST) for 1 h at room temperature and probed overnight at 4°C with the appropriate primary antibodies [anti-SOCS3 rabbit polyclonal antibody (catalog number sc-9023, Santa Cruz Biotechnology, Inc.), anti-A20 mouse monoclonal antibody (catalog number sc-166692, Santa Cruz Biotechnology, Inc.) and anti-β-actin mouse monoclonal antibody (Zhongshan Goldenbridge...
Biotechnology Co. Ltd., China), all at a dilution of 1:1,000]. Then, the membranes were rinsed with TBST before a 1-h incubation with horseradish peroxidase (HRP)-conjugated secondary antibodies [goat anti-rabbit IgG (H+L)-labeled secondary antibody (catalog number A0208, Beyotime Institute of Biotechnology, China) and goat anti-mouse IgG (H+L)-labeled secondary antibody (catalog number A0192, Beyotime Institute of Biotechnology, China), both at a dilution of 1:1,500]. Membranes were washed in TBST solution with 0.5% Tween-20 before enhanced chemiluminescence (ECL) detection with BeyoECL Plus (catalog number P0018, Beyotime Institute of Biotechnology, China). After exposure on X-ray film, protein band densitometry was quantified using the Gel Image Analysis System (Tanon, Shanghai, China), and the antibody against β-actin was used for normalization as previously described [37, 38]. Concurrently, the expression levels of SOCS3 and A20 in the 22 cases of normal biliary duct tissues were detected by western blot analysis according to the above method.

Patients and clinical data
A total of 109 patients who underwent curative surgery, whose CCA (adenocarcinoma subtype) was later pathologically confirmed and who did not have any other malignancy between December 2009 and March 2012 at the Second Affiliated Hospital of Harbin Medical University were retrospectively reviewed. No patients received chemotherapy or radiotherapy before or after surgery in this study. Patients who died of unrelated diseases or within one month after surgery were excluded, leaving 86 patients eligible for this study. These patients consisted of 54 males and 32 females with a median age of 61.5 years (range of 42–83 years). The pathological data for CCA were evaluated according to the 7th edition of the American Joint Committee on Cancer (AJCC). Other clinical data from the patients, such as tumor site, differentiation, histological patterns, lymph node metastasis, vascular invasion, tumor node metastasis (TNM) stage, postoperative recurrence and 3-year survival, were obtained from pathology reports and medical records, along with preoperative serum CEA (carcinoembryonic antigen) and CA19-9 (carbohydrate antigen 19–9) levels and HBV (hepatitis B virus) infection status. Overall survival was calculated as the interval from the date of surgery to death or the date of the latest follow-up for the living patients. Each CCA patient was followed until March 2015 or their date of death, and the median follow-up period was 23 months (ranging from 2 to 48 months) (S1 Table).

Curative surgery for CCA was performed as complete resection of the cancer mass (without a positive surgical margin according to pathological examination), dissection of the regional lymphonodi, and removal of the cancer embolus in the regional vessels and biliary ducts. The CCA specimens were obtained from regions close to the cancer margin. All surgical specimens were made into formalin-fixed paraffin-embedded (FFPE) blocks by pathologists and stored in the pathology department.

IHC
IHC was performed as previously described [4]. FFPE blocks were cut at a thickness of 4 μm, placed and fixed on positively charged slides, and heated at 55°C for 30 min. Then, the slides were deparaffinized in xylene and subsequently hydrated in a graded ethanol solution. The deparaffinized slides were stained with hematoxylin-eosin to identify typical FFPE blocks with tumor morphology. Selected FFPE blocks were cut into serial slides at 4-μm intervals for assessment. All slides were deparaffinized and rehydrated as mentioned above. Then, the slides were incubated with 3% hydrogen peroxide in PBS at room temperature for 20 min to quench endogenous peroxidase activity. After rinsing with PBS, antigen retrieval was carried out in citrate buffer (pH 6.0) for 15 min in a microwave oven. The slides were then incubated in normal goat serum for 30 min at 37°C to block nonspecific binding, followed by incubation with anti-
SOCS3 rabbit polyclonal antibody (catalog number sc-9023, Santa Cruz Biotechnology, Inc.) or anti-A20 mouse monoclonal antibody (catalog number sc-166692, Santa Cruz Biotechnology, Inc.) at a dilution of 1:200 for 30 min at room temperature. Concurrently, PBS was applied instead of the primary antibodies for unstained slides as a negative control. Then, the slides were rinsed 3 times with PBS, followed by incubation with secondary antibody (biotin conjugated goat anti-rabbit or goat anti-mouse immunoglobulin, catalog numbers sc-2040 and sc-2039, Santa Cruz Biotechnology, Inc.) for 30 min. After washing with PBS, the slides were incubated with HRP-streptavidin reagent for 45 min. Diaminobenzidine (DAB) and hematoxylin were utilized for chromogenic detection and counterstaining on all slides.

Assessment of IHC variables

The assessment of IHC variables was performed as described previously [39] with slight modifications. Semi-quantitative expression levels were based on the staining intensity of SOCS3/A20 and the distribution of positive tumor cells. The chromogenic reaction of SOCS3/A20 was classified as four grades by staining intensity using a scale of 0–3, where 0 corresponded to negative staining and 3 to strong staining. The percentage of SOCS3/A20 staining in five random non- overlapping fields of each slide at a final magnification of 400× was also classified by grade as follows: 0 (<25%), 1 (5–25%), 2 (26–50%), 3 (51–75%), and 4 (76–100%). The intensity and percentage scores were multiplied to obtain the final staining score [37]. The staining pattern for all slides was defined as follows: a final staining score of 6–12 was defined as high expression, while scores less than 6 were defined as low expression. Furthermore, each slide was blindly evaluated by two experienced independent pathologists unaware of the clinical data and follow-up information. The final staining scores from the two pathologists were compared, and any inconsistent scores were managed by reappraisal of the slide by both pathologists until a consistent score was reached. Images were obtained using an Olympus BX50 light microscope connected to a charge-coupled device (CCD) camera and the Image-Pro Plus 6.0 software (Media Cybernetics, Inc.).

Statistical analysis

The data analyses were performed using the Statistical Package for the Social Sciences software (SPSS, version 19.0, Chicago, IL). The paired-samples t-test was used to compare the protein expression of SOCS3/A20 between CCA tumors and the corresponding peritumoral tissue samples. The relationship between SOCS3/A20 expression and the clinicopathological features at various stages of CCA progression was elucidated using a chi-square test. Kaplan-Meier analysis was applied to calculate survival curves. As mentioned above, the overall survival was calculated as the interval between the time of surgery and the time of death or the end of follow-up. The relationship between SOCS3 and A20 expression was evaluated by linear regression analysis and Spearman rank correlation. A univariate log-rank test was carried out to determine the significance of the clinicopathological parameters, and then the risk factors filtered by the log-rank test were further analyzed with the Cox hazards regression model to determine independent factors for prognosis. A P value less than 0.05 was considered statistically significant, with all P values based on two-sided statistical analysis.

Results

SOCS3 and A20 expression levels in CCA

The protein expression levels of SOCS3 and A20 were first detected by western blot analysis in freshly frozen tumors and the corresponding peritumoral biliary tissues from 22 CCA cases. The SOCS3 signal was positive in only 27.27% (6 cases) of the tumor tissues, while it was
positive in all corresponding peritumoral biliary tissues. Conversely, the A20 signal was positive in all tumor tissues and in 68.75% (15 cases) of the corresponding peritumoral biliary tissues (Fig 1A). The quantitative analysis of each protein band was standardized against β-actin expression in each sample, and the ratio of the protein of interest to β-actin expression was defined as the expression index. The protein expression level of SOCS3 in CCA tumor tissues was conspicuously lower than that in the corresponding peritumoral biliary tissues (7.56-fold on average, \( P < 0.0001 \)), and it was also lower than that in normal biliary duct tissues (Fig 1B). In contrast, the protein expression level of A20 in CCA tumor tissues was conspicuously higher than that in the corresponding peritumoral biliary tissues (2.23-fold on average, \( P < 0.0001 \)), and it was also significantly higher than the protein expression level of A20 in normal biliary duct tissues (Fig 1C). Linear regression analysis further revealed that the A20 level was inversely correlated with SOCS3 expression in CCA tumor tissues and normal biliary duct tissues (\( R^2 = 0.8232, P < 0.0001 \); Fig 1D).

Next, the expression levels of SOCS3 and A20 were detected in tissue slides from 86 CCA cases by IHC. As mentioned above, SOCS3 expression was found predominately in the cytoplasm and cytomembrane, while A20 expression was detected in the nucleus (Fig 2, scale bar = 100 μm). According to the assessment criteria in this study, 62 (72.1%) of the 86 CCA specimens were classified as showing low expression for SOCS3, whereas 24 (27.9%) showed high expression. Conversely, for A20, 27 (31.4%) of the CCA specimens showed low expression, and 59 (69.6%) showed high expression. Consistent with the western blot analysis, the IHC results indicated decreased expression of SOCS3 and increased expression of A20 in CCA.

To further identify the correlation between the expression of SOCS3 and A20, the 86 CCA specimens were assigned to four groups based on SOCS3 and A20 expression levels (Fig 3). In the 86 CCA samples, 6 (7.0%) showed high expression of both SOCS3 and A20, 9 (10.5%) showed low expression of both SOCS3 and A20, 53 (61.6%) showed high A20 and low SOCS3 expression, and 18 (20.9%) showed low A20 and high SOCS3 expression. Utilizing Spearman rank analysis, a significantly negative correlation between SOCS3 and A20 expression was verified (\( r = -0.585, P < 0.0001 \)). Moreover, the data further confirmed the inverse relationship between the expression levels of SOCS3 and A20 in CCA.

Patients’ characteristics

As shown in Table 1, 86 CCA patients were included in this study, and their average age was 61.6 years. The patients consisted of 32 males and 54 females. Among these patients, 18 had intrahepatic CCA, 33 had perihilar CCA, and 35 cases had distal CCA. In accordance with tumor differentiation, 16 cases were classified as Grade 1 (well differentiated), 38 cases as Grade 2 (moderately differentiated) and 32 cases as Grade 3 (poorly differentiated). Based on histological patterns, 76 cases were histologically classified as tubular adenocarcinoma, 6 cases as papillary adenocarcinoma and 4 cases as mucinous adenocarcinoma. A total of 51 patients suffered from lymph node metastasis, while 12 patients had vascular invasion. According to the 7th edition of the AJCC classification, 12 cases were in stage I, 23 cases were in stage II, 32 cases were in stage III, 19 cases were in stage IV A, and stage IV B patients who were not amenable to surgery were excluded. Each CCA patient was followed until March 2015 or their date of death. The survival time in this study ranged from 2 to 48 months, with a median survival time of 23 months. A total of 52 patients died from recurrence during the follow-up period.

SOCS3/A20 expression level and clinical data

To verify the relationship between SOCS3/A20 expression and major clinical data among the 86 CCA cases, chi-square tests were performed. When the SOCS3/A20 expression level was
classified as high or low, no significant correlation was found between SOCS3/A20 expression and certain clinicopathological parameters, including gender, age, tumor site, histological patterns, vascular invasion, serum CEA and CA19-9 levels and HBV infection status (Table 1, all P values > 0.05). However, statistically significant correlations were found with lymph node metastasis, overall survival rate and postoperative recurrence (P < 0.05) for both SOCS3 and A20. Only A20 showed a statistically significant association with the TNM stage, while SOCS3 merely showed a significant association with tumor differentiation. These results revealed that low SOCS3 expression and high A20 expression in CCA correlated with the status of the disease.

To further investigate the correlation between SOCS3/A20 expression and overall survival in CCA, Kaplan-Meier analysis was applied to calculate survival curves. The results suggested that patients with low SOCS3 levels had a dramatically worse overall survival rate (P = 0.008, Fig 4A), while patients with high A20 levels showed a worse overall survival rate (P = 0.007, Fig 4B).

Univariate and multivariate analysis

To evaluate the role of clinicopathological parameters as probable risk factors for prognosis, univariate analysis was performed to analyze 13 clinicopathological variables in the 86 CCA
patients. As shown in Table 2, the results indicated that tumor differentiation ($P = 0.026$), lymph node metastasis ($P = 0.004$), vascular invasion ($P < 0.001$), TNM stage ($P = 0.008$), SOCS3 expression ($P = 0.001$) and A20 expression ($P < 0.001$) were prognostic factors with statistical significance (Table 2). Then, the 6 risk factors filtered by the log-rank test were further analyzed by multivariate analysis to determine independent factors for prognosis. As presented in Table 3, only lymph node metastasis ($P = 0.007$; HR, 1.142), vascular invasion ($P = 0.001$; HR, 1.370), SOCS3 expression ($P = 0.022$; HR, 2.382) and A20 expression ($P = 0.009$; HR, 6.598) served as independent biomarkers for prognosis, indicating that from all 13 biomarkers, a low SOCS3 level and high A20 level may represent the best prognostic indicators of survival in CCA.

**Discussion**

Although the downregulation of SOCS3 has been regarded as crucial in tumor proliferation and migration [40], few studies have assessed the prognostic role of SOCS3 in human CCA. In addition, to the best of our knowledge, little is known about the role of A20 in predicting CCA patients’ prognosis. Herein, our study suggests that SOCS3 and A20 may serve as prognostic biomarkers in CCA, and our results support the hypothesis that A20 enhances JAK/STAT signaling in CCA by down-regulating SOCS3. Initially, we compared SOCS3 and A20 protein expression levels by western blot analysis in 22 cases of freshly frozen CCA tumors and their corresponding peritumoral biliary tissues. Compared with the corresponding peritumoral biliary tissues, the CCA tumor tissues showed remarkably higher A20 protein expression levels. However, lower SOCS3 protein expression was observed in CCA tumor tissues than the corresponding peritumoral biliary tissues. Linear regression analysis revealed that A20 levels were...
inversely correlated with SOCS3 expression ($R^2 = 0.8232$, $P < 0.0001$). Next, we examined the SOCS3 and A20 protein expression levels in 86 cases of surgical specimens using IHC and detected a correlation between SOCS3 and A20 expression. Our data demonstrated that there was an inverse relationship ($r = -0.585$, $P < 0.0001$) between the expression levels of SOCS3 and A20 in CCA. To investigate the role of prognostic biomarkers, we further analyzed the SOCS3 and A20 protein expression levels with corresponding clinicopathological features in 86 CCA cases. These results suggested that patients with low SOCS3 levels had a remarkably worse overall survival rate compared to those with high SOCS3 levels ($P = 0.001$, Fig 4A), whereas patients with high A20 levels had a worse overall survival rate than those with low A20 levels ($P < 0.001$, Fig 4B).

The JAK/STAT pathway is constitutively activated in various human malignancies, particularly in the tumor subtypes associated with chronic inflammation [41, 42], including CCA, as recently reported [43]. Activation of the JAK/STAT pathway results in tyrosine-phosphorylated STAT3, which is involved in many aspects of tumorigenesis, including proliferation, differentiation, apoptosis, modulation of sensitivity to cytotoxic agents, angiogenesis, recruitment of immune cells, and metastasis [9–12]. Decreased expression of SOCS proteins through promoter methylation significantly contributes to the persistent tyrosine phosphorylation of STAT3 in cancers [44], as well as in CCA [21].

Our recent studies found that high expression of SOCS3 could reduce tumor metastasis, EMT markers and STAT3 activation in the absence of IL-6 stimulation in CCA cell lines [22].
Table 1. SOCS3 and A20 expression status in relation to clinicopathologic features.

| Clinicopathologic features | No. patients | SOCS3 expression level | A20 expression level | P |
|----------------------------|--------------|------------------------|----------------------|---|
|                            |              | High | Low | High | Low |     |
| Age (years)                |              |      |     |      |     |     |
| <60                        | 37           | 9    | 28  | 28   | 9   | 0.185|
| ≥60                        | 49           | 15   | 34  | 31   | 18  | 0.220|
| Gender                     |              |      |     |      |     |     |
| Male                       | 54           | 13   | 41  | 40   | 14  | 0.156|
| Female                     | 32           | 11   | 21  | 19   | 13  |     |
| Tumor site                 |              |      |     |      |     |     |
| Intrahepatic               | 18           | 4    | 14  | 13   | 5   | 0.877|
| Perihilar                  | 33           | 7    | 26  | 23   | 10  |     |
| Distal                     | 35           | 13   | 22  | 23   | 12  |     |
| Differentiation            |              |      |     |      |     |     |
| Grade 1                    | 16           | 10   | 6   | 7    | 9   | 0.160|
| Grade 2                    | 38           | 9    | 29  | 27   | 11  |     |
| Grade 3                    | 32           | 5    | 27  | 25   | 12  |     |
| Histological patterns      |              |      |     |      |     |     |
| Tubular adenocarcinoma     | 76           | 23   | 53  | 51   | 25  | 0.177|
| Papillary adenocarcinoma   | 4            | 1    | 3   | 2    | 2   |     |
| Mucinous adenocarcinoma    | 6            | 0    | 6   | 6    | 0   |     |
| Lymph node metastasis      |              |      |     |      |     |     |
| Positive                   | 51           | 10   | 41  | 42   | 9   | 0.001|
| Negative                   | 35           | 14   | 21  | 17   | 18  |     |
| Vascular invasion          |              |      |     |      |     |     |
| Positive                   | 12           | 1    | 11  | 11   | 1   | 0.128|
| Negative                   | 74           | 23   | 51  | 48   | 26  |     |
| TNM stage                  |              |      |     |      |     |     |
| I                          | 12           | 6    | 6   | 7    | 5   | 0.001|
| II                         | 23           | 6    | 17  | 7    | 10  |     |
| III                        | 32           | 9    | 23  | 30   | 2   |     |
| IVA                        | 19           | 3    | 16  | 15   | 4   |     |
| Postoperative recurrence   |              |      |     |      |     |     |
| Present                    | 50           | 9    | 41  | 39   | 11  | 0.026|
| Absent                     | 36           | 15   | 21  | 20   | 16  |     |
| Survival time              |              |      |     |      |     |     |
| >3 years                   | 34           | 14   | 20  | 19   | 15  | 0.040|
| ≤3 years                   | 52           | 10   | 42  | 40   | 12  |     |
| Serum CEA                  |              |      |     |      |     |     |
| >5 ng/ml                   | 51           | 12   | 39  | 36   | 15  | 0.632|
| ≤5 ng/ml                   | 35           | 12   | 23  | 23   | 12  |     |
| Serum CA199                 |              |      |     |      |     |     |
| >37 U/ml                   | 56           | 15   | 41  | 37   | 19  | 0.489|
| ≤37 U/ml                   | 30           | 9    | 21  | 22   | 8   |     |
| HBV infection              |              |      |     |      |     |     |
| Positive                   | 42           | 11   | 31  | 30   | 12  | 0.581|

(Continued)
In this study, we reinforced the putative prognostic role of SOCS3 in CCA by western blot and IHC in human surgical specimens. Using western blot analysis, the SOCS3 signal was positive in only 27.27% (6 cases) of the freshly frozen CCA tumor tissues, while it was positive in all (22 cases) corresponding peritumoral biliary tissues. The loss of SOCS3 expression in malignancies has been extensively described [16–20], suggesting that it may serve as a general mechanism for activating the JAK/STAT pathway in solid tumors. We found that SOCS3 expression was decreased in 72.1% of the CCA specimens (62 cases) by IHC. Furthermore, we found that SOCS3 expression was remarkably correlated with lymph node invasion and histological type in CCA. Moreover, patients with low SOCS3 levels showed a poor overall survival rate (45.9% vs 73.4%), as evaluated by univariate analysis, and multivariate Cox analysis identified SOCS3 as an independent risk factor for prognosis ($P = 0.022$; HR, 2.382). What was most interesting was that SOCS3 repression was associated with A20 overexpression in CCA tissues.

A20 functions as a tumor enhancer due to its overexpression in multiple malignant solid tumors [25–30] and also as a ubiquitin-editing enzyme that regulates inflammatory responses and cell immunoreaction [23]. In lymphoma, however, A20 repression as well as mutation is commonly observed, which implies its role as a suppressor in tumor biology [32–35]. Therefore, whether A20 acts as an enhancer or a suppressor in tumor biology may depend on the tissue type and tumor stage [27]. According to the results of our western blot analysis, the A20 signal was positive in all (22 cases) freshly frozen CCA tumor tissues and in 68.75% (15 cases) of the corresponding peritumoral biliary tissues. Furthermore, the protein expression levels of A20 in CCA tumor tissues were remarkably higher compared to those in the corresponding peritumoral biliary tissues (2.23-fold on average, $P < 0.0001$) (Fig 1C). By IHC, high A20 expression was detected in 68.6% (59 cases) of the CCA specimens. Patients with high A20 expression also showed a poor overall survival rate (71.4% vs 45.6%), as evaluated by univariate analysis, and A20 was identified as an independent biomarker for prognosis by multivariate Cox analysis ($P = 0.009$; HR, 6.598). These results suggest that A20 may function as an enhancer in the tumor biology of CCA, which is consistent with previous studies of other malignant solid tumors, such as undifferentiated nasopharyngeal carcinoma, poorly differentiated head and neck squamous cell carcinoma [25], glioma [26, 27], glioblastoma [28], inflammatory breast cancer [29], and hepatocellular carcinoma [30], where A20 expression was related to an undesirable prognosis.

We further demonstrated that A20 expression in CCA was significantly associated with tumor differentiation, TNM stage and lymph node metastasis, and multivariate analysis confirmed A20 as an independent prognostic factor according to the Cox proportional hazard regression model. Interestingly, the inverse correlation between A20 overexpression and SOCS3 repression discovered in our study suggests that the mechanism of SOCS3 downregulation in CCA may be similar to that reported in a previous study of liver regeneration [24], which showed that A20 enhanced JAK/STAT signals in hepatocytes by downregulating SOCS3.

| Clinicopathologic features | No. patients | SOCS3 expression level | $P$ | A20 expression level | $P$ |
|---------------------------|-------------|------------------------|-----|----------------------|-----|
|                           |             | High | Low |                | High | Low |
| Negative                  | 44          | 13   | 31  |                | 29   | 15  |

TNM stage, Tumor-Node-Metastasis stage; CEA, carcino embryonie antigen; CA199, carbohydrate antigen 19–9; HBV, Hepatitis B virus.

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in the process of liver regeneration. However, we noticed that 17.5% of the patients in this study did not fit into the category where the relationship between A20 and SOCS3 was inverse (Fig 3), and this result indicates that there might be alternative oncogenic pathways in CCA development for these patients. Moreover, little is known about the mutual impact of SOCS3 and A20 in CCA, and intensive investigation will be performed on this topic in our future study.

Most of the risk factors for CCA, including primary sclerosing cholangitis (PSC), chronic hepatolithiasis and choledocholithiasis, choledochal cysts, chronic hepatitis C virus (HCV) and parasitic bile duct infection, are immediate causes of chronic biliary inflammation. Anomalies in growth regulatory genes have been partially illuminated in established malignant tumors in several studies and may serve as the mechanism for the interaction between biliary chronic inflammation and carcinogenesis [45–47]. During chronic inflammation, activation of the JAK/STAT pathway plays a crucial role in tumor progression, not only in tumor cells but also dendritic cells, myeloid cells, and B and T cells, to establish a chronic inflammatory environment but also in epithelial cells to generate cytokines and chemokines [48]. Tumors originating in these environments maintain secretion of these inflammatory factors most likely because they supply tumor growth factors and favorable survival conditions.

Our early research revealed that IL-6, through activation of JAK/STAT signaling, is involved in the promotion of human biliary epithelial cell proliferation and migration [12]. Subsequently, our recent study confirmed SOCS3 as a negative feedback regulator, controlling the JAK/STAT pathway in relation to cell proliferation and migration through a classic feedback loop in human CCA cell lines [22]. Combined with our present study, these results show the significant role that SOCS3 plays in regulating JAK/STAT signaling during the process of inflammation in the development of CCA, thereby predicting the prognosis of CCA patients. Previous research has demonstrated that A20 regulates the JAK/STAT pathway by upregulating miR203 levels, which decreases SOCS3 mRNA expression. This decrease activates JAK/STAT3 signaling and the transcription of STAT3-dependent mitogenetic genes, such as cyclin A and cyclin D1, while blocking NF-κB signaling, thus decreasing transcription of STAT3-dependent proinflammatory genes [24]. Taken together, our data and previous studies [24]
Table 2. Univariate log-rank test of clinicopathological features of 86 patients with cholangiocarcinoma.

| Clinicopathologic features                  | No. patients | Survival rate (%) | P     |
|--------------------------------------------|--------------|-------------------|-------|
| **Age (years)**                            |              |                   |       |
| <60                                        | 37           | 57.1%             | 0.893 |
| ≥60                                        | 49           | 52.4%             |       |
| **Gender**                                 |              |                   |       |
| Male                                       | 54           | 49.2%             | 0.312 |
| Female                                     | 32           | 65.3%             |       |
| **Tumor site**                             |              |                   |       |
| Intrahepatic                               | 18           | 31.1%             | 0.545 |
| Perihilar                                  | 33           | 59.2%             |       |
| Distal                                     | 35           | 58.6%             |       |
| **Differentiation**                        |              |                   |       |
| Grade 1                                    | 16           | 68.8%             | 0.026 |
| Grade 2                                    | 38           | 60.2%             |       |
| Grade 3                                    | 32           | 33.6%             |       |
| **Histological patterns**                  |              |                   |       |
| Tubular adenocarcinoma                     | 76           | 54.2%             | 0.088 |
| Papillary adenocarcinoma                   | 4            | 50.0%             |       |
| Mucinous adenocarcinoma                    | 6            | 25.0%             |       |
| **Lymph node metastasis**                  |              |                   |       |
| Positive                                   | 51           | 37.0%             | 0.004 |
| Negative                                   | 35           | 68.6%             |       |
| **Vascular invasion**                      |              |                   |       |
| Positive                                   | 12           | 28.4%             | < 0.001 |
| Negative                                   | 74           | 57.8%             |       |
| **TNM stage**                              |              |                   |       |
| I                                          | 12           | 75.0%             | 0.008 |
| II                                         | 23           | 63.6%             |       |
| III                                        | 32           | 49.0%             |       |
| IVA                                        | 19           | 30.2%             |       |
| **Serum CEA**                              |              |                   |       |
| >5 ng/ml                                   | 51           | 57.4%             | 0.774 |
| ≤5 ng/ml                                   | 35           | 52.6%             |       |
| **Serum CA199**                            |              |                   |       |
| >37 U/ml                                   | 56           | 53.2%             | 0.933 |
| ≤37 U/ml                                   | 30           | 56.6%             |       |
| **HBV infection**                          |              |                   |       |
| Positive                                   | 42           | 46.4%             | 0.544 |
| Negative                                   | 44           | 62.9%             |       |
| **SOCS3 expression level**                 |              |                   |       |
| High                                       | 24           | 73.4%             | 0.008 |
| Low                                        | 62           | 45.9%             |       |
| **A20 expression level**                   |              |                   |       |
| High                                       | 59           | 45.6%             | 0.007 |
| Low                                        | 27           | 71.4%             |       |

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support the hypothesis that A20 enhances the activity of the JAK/STAT pathway in CCA by decreasing the SOCS3 level.

CCA is notorious for its devastating prognosis in advanced stages and the lack of conspicuous clinical features in early stages [49]; furthermore, the incidence of CCA is increasing worldwide [50]. Since this disease was first systematically described by Klatskin, advances have been made towards the diagnosis and treatment of CCA in recent decades. Nevertheless, the survival rate of CCA remains unsatisfactory, even for patients who undergo curative surgery [5]. Therefore, the identification of new cancerous biomarkers in surgical specimens is needed for promoting both early diagnosis and prognosis of CCA. This study is the first to provide evidence for the unanticipated interaction of the inflammatory regulators A20 and SOCS3 in CCA. Our data offer vital information for the prediction that A20 and SOCS3 could be used as a novel prognostic biomarkers in CCA and may be regarded as new therapeutic targets for future CCA treatment.

Supporting Information
S1 Table. The follow-up data of 86 CCA patients.
(XLS)

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Table 3. Multivariate Cox regression analysis of clinicopathological features of 86 patients with cholangiocarcinoma.

| Factors                     | Category            | P      | HR    | 95% CI  |
|-----------------------------|---------------------|--------|-------|---------|
| Differentiation             | Grade 3             | 0.285  | 0.396 | 0.203–0.718 |
| Grade 1+ Grade 2            |                     |        |       |         |
| TNM stage                   | III+ IVA            | 0.715  | 0.434 | 0.169–0.835 |
| I+ II                       |                     |        |       |         |
| Vascular invasion           | Positive            | 0.001  | 1.370 | 1.251–1.642 |
| Negative                    |                     |        |       |         |
| Lymph node metastasis       | Positive            | 0.007  | 1.142 | 1.036–1.437 |
| Negative                    |                     |        |       |         |
| SOCS3 expression level      | Low                 | 0.022  | 2.382 | 1.253–4.722 |
| High                        |                     |        |       |         |
| A20 expression level        | High                | 0.009  | 6.598 | 2.584–14.261 |
| Low                         |                     |        |       |         |

HR, hazard ratio; CI, confidence interval.

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Author Contributions
Conceived and designed the experiments: YC YW. Performed the experiments: YW MW QZ HW ZW XZ ST LZ. Analyzed the data: YW. Contributed reagents/materials/analysis tools: YW. Wrote the paper: YW.
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