Asymmetric dimethylarginine: A novel biomarker of gastric mucosal injury?

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Abstract

Nitric oxide (NO), a multifunctional endogenous gas molecule, is metabolized from L-arginine by enzymatic reaction in the presence of nitric oxide synthase. NO, an important gas signaling molecule, is a gastric mucosa protective factor that contributes significantly to maintain normal gastric mucosa integrity. NO increases gastric mucosa blood flow, regulates the secretion of mucus and bicarbonate, and inhibits the secretion of gastric juice. Asymmetric dimethylarginine (ADMA) is derived from the catabolism of L-arginine and is identified as the major endogenous inhibitor of nitric oxide synthase. The function of ADMA is to decrease NO production via inhibiting nitric oxide synthase activity. Besides inhibiting NO synthesis, ADMA also directly induces oxidative stress and cell apoptosis, and participates in inflammation reaction. Its systemic accumulation was observed in conjunction with several cardiovascular and metabolic diseases. ADMA also mediates gastric ulcer injury induced by ethanol, stress, helicobacter pylori and indomethacin. The mechanism of ADMA directly producing adverse effect in gastric mucosa is incompletely understood. It is widely accepted that NO bioavailability decrease is the majority reason. Promotion of apoptosis and aggravation of inflammation may be other important mechanisms of ADMA-induced gastric injury. ADMA might be a novel clinical and experimental biomarker related to gastric mucosa disorder. Although therapeutic tool targeting to ADMA is available in multiple cardiovascular diseases, it is unknown in gastrointestinal disease. The strategy to inhibit ADMA is beneficial to gastric ulcer induced by ethanol in rats. Thus, ADMA might be a candidate of therapeutic target in gastric mucosa damage.

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Key words: Asymmetric dimethylarginine; Mucosal injury; Nitric oxide

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INTRODUCTION

Nitric oxide (NO), a multifunctional endogenous gas molecule, is metabolized from L-arginine by enzymatic reaction in the presence of nitric oxide synthase (NOS). It is accepted as the most critical messenger and functional executant in a variety of biologic activities. Numerous researches have reported the beneficial effects of NO, including endothelium protection, inhibition of local inflammation and cell pathoproliferation. NO bioavailability is most directly regulated by the abundance and/or activity of NOS. NO can be competed by substrates, endogenous L-arginine and asymmetric dimethylarginine (ADMA). The function of ADMA is to decrease NO production via inhibiting the NOS activity. ADMA is derived from the catabolism of
proteins containing methylated arginine residues\textsuperscript{[3]}. Arginine methylation of cellular proteins is catalyzed by protein arginine methyltransferases\textsuperscript{[5]}. ADMA is hydrolyzed by dimethylarginine dimethylaminohydrolase (DDAH)\textsuperscript{[3]}.

Growing evidence has shown that excessive accumulation of ADMA can decrease NO bioavailability in cells/tissues, induce domain dysfunctions or produce detrimental effects in multiple systems\textsuperscript{[8]}. Besides inhibiting NO synthesis, ADMA can directly induce oxidative stress and cell apoptosis, and participate in the inflammation reactions\textsuperscript{[14,15]}. It is well documented that ADMA is involved in a wide range of cardiovascular disorders. ADMA impairs endothelial functions, thus leading to hypertension, atherosclerosis, coronary heart disease, diabetes mellitus, pulmonary hypertension and renal failure\textsuperscript{[3,16]}. Although it is well established that ADMA is an independent risk factor of cardiovascular diseases, the pathological role of ADMA in other diseases is still unknown, including gastric diseases.

**NO AND GASTRIC MUCOSAL PROTECTION**

Gastric mucosal diseases, including gastric ulcers, affect 25%-30% of the world’s population. The mechanism of gastric mucosal damage and protection is yet unclear. NO, an important gas signaling molecule, is a gastric mucosal protective factor that contributes significantly to maintaining normal gastric mucosal integrity\textsuperscript{[7,8]}. Gastric mucosa is composed of gastric epithelial cells and glands. Gastric epithelial cells can secrete multiple active substances, including NO. There is substantial evidence that NO may be an important defensive factor of gastric mucosa\textsuperscript{[108]}. In the animal models of gastric mucosal injury induced by ethanol, endothe-lin, ischemia/reperfusion, cold-stress, water immersion restraint, indomethacin and aspirin, NO exerts protective effects on gastric mucosal integrity\textsuperscript{[111-113]}. The mechanisms underlying the protective effects of NO could be explained as follows: (1) NO increases gastric mucosal blood flow; (2) NO regulates the secretion of mucus and bicarbonate, which compose the first level of gastric tissue defense; and (3) NO inhibits the secretion of gastric juice\textsuperscript{[14,13]}. 

**ADMA AND GASTRIC MUCOSAL DAMAGE**

*Helicobacter pylori* (H. pylori) infection is a definitive cause of gastroduodenal ulcer. Epidemiological analysis has shown that 70%-80% gastric ulcers and 95%-100% duodenal ulcers are attributed to *H. pylori* infection. In a clinical investigation of *H. pylori*-positive and -negative volunteers, *H. pylori* infection was found to increase the tissue contents of ADMA\textsuperscript{[16,46]}. More importantly, a laboratory research has shown that proteolysis of *H. pylori* extract also results in a substantial accumulation of ADMA, indicating that *H. pylori* infection must be taken into account as a cause of increased ADMA levels\textsuperscript{[17]}. Strong associations have also been found between ADMA and gastric mucosal dysfunction. Exogenous administration with ADMA inhibits the mucosal alkaline response to acid exposure, which is regarded as one of the important beneficial factors of gastric mucosa complaint. And an increase in ADMA mediates the water extract of *H. pylori*-induced acid-base imbalance in duodenal tissues\textsuperscript{[171]}. In our *in vitro* experiment, we found that intervention of *H. pylori* stimulated ADMA release from cultured gastric epithelial cells\textsuperscript{[46]}. We also observed that *H. pylori* infection markedly exacerbated ADMA-produced gastric tissue injury in rats\textsuperscript{[19]}.

Besides *H. pylori* invasion, gastric mucosal function is also disturbed by other factors, such as ethanol, indomethacin and cold-stress. Our recent studies have explored the role of ADMA in gastric mucosal injury induced by other causes in addition to *H. pylori* infection. The results have shown that ADMA levels are increased in gastric juice in three separate gastric mucosal models, including ethanol, indomethacin and cold-stress initiated mucosal injury, accompanied with a decrease in NO level and DDAH activity\textsuperscript{[19]}. Pretreatment with L-arginine, an antagonist of ADMA, markedly reduces the degree of gastric injury and elevates NO bioavailability. Administration of exogenous ADMA significantly exacerbates gastric injury, concomitantly with a decrease in plasma level of NO. These findings indicate that ADMA may participate in the gastric mucosal injury process induced by various deleterious factors. Multiple exogenous NOS inhibitors, such as L-NMA, L-NAME and L-NMMA intensify gastric mucosal injury by decreasing gastric mucosal blood flow, while NO donor (nitroprusside sodium) or substrate (L-arginine) reverses such effects\textsuperscript{[118,28]}. 

Cigarette smoking is one of the risk factors provoking gastroduodenal ulceration\textsuperscript{[21]}. It increases both the incidence and relapse rate of peptic ulcer diseases and delays ulcer healing. Besides reducing bioavailability of NO, nicotine, a major component of tobacco, aggravates gastric mucosal injury due to some factors such as ethanol or stress\textsuperscript{[22]}. Recently, we have found that in the rats treated with ethanol, the ulcer index and ADMA level were increased and the NO level was decreased, and these effects of ethanol were augmented by pre-treatment with nicotine\textsuperscript{[23]}. Nicotine alone did not show significant impact on ulcer index, ADMA level and NO level. In *in vitro* incubation of epithelial cells with ethanol induced cell injury and increased ADMA level in the cultural medium, an effect that was amplified in the presence of nicotine. Therefore, the

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**Figure 1** The hypothesized role of asymmetric dimethylarginine in gastric mucosal injury and the underlying mechanisms. ADMA: Asymmetric dimethylarginine; NO: Nitric oxide.
accelerated effect of nicotine on gastric mucosal dysfunction is associated with endogenous ADMA.

The mechanism of ADMA directly producing adverse effect in gastric mucosa is incompletely understood. It is widely accepted that NO bioavailability decrease is the major reason. Apoptosis reaction produced by ADMA has been observed in different types of cells, such as endothelial cell and smooth muscle cell. Cell apoptosis has also been observed in cultured gastric epithelial cells when exposed to an exorbitant concentration of ADMA. ADMA could directly induce the production of inflammatory cytokines tumor necrosis factor (TNF-α) and soluble intercellular adhesion molecule-1 via activation of p38 MAPK and ERK1/2 pathways in cultured endothelial cells. We deduce that inflammatory reaction may be another important factor for ADMA-induced gastric injury because TNF-α level, the marker of inflammation, is significantly elevated when cells are damaged by ADMA in gastric mucosal epithelial cells. The hypothesized role of ADMA in gastric mucosal injury and the underlying mechanisms are summarized in Figure 1.

PERSPECTIVES AND CLINICAL IMPLICATIONS

ADMA might be a novel clinical and experimental biomarker related to gastric mucosal disorder. The mechanisms of impairment involve NO generation inhibition, inflammatory reaction and apoptosis promotion. Although the therapeutic tool targeting ADMA is available in multiple cardiovascular diseases, it is unknown in gastrointestinal diseases. The strategy to inhibit ADMA is beneficial to the treatment of gastric ulcer induced by ethanol in rats. Thus, ADMA might be a candidate of therapeutic target in gastric mucosal damage.

REFERENCES

1. Beltowski J, Kedra A. Asymmetric dimethylarginine (ADMA) as a target for pharmacotherapy. Pharmacol Rep 2006; 58: 159-178
2. Rizzo NO, Maloney E, Pham M, Luttrell I, Wessells H, Tateya S, Daum G, Handa P, Schwartz MW, Kim F. Reduced NO-cGMP signaling contributes to vascular inflammation and insulin resistance induced by high-fat feeding. Arterioscler Thromb Vasc Biol 2010; 30: 758–765
3. Fiedler L. The DDAH/ADMA pathway is a critical regulator of NO signalling in vascular homeostasis. Cell Adh Migr 2008; 2: 149–150
4. Greenlof M, Zheng C, Werle E, Fiehn W, Ulmer HE. Plasma levels of asymmetric dimethyl-L-arginine in patients with congenital heart disease and pulmonary hypertension. J Cardiovasc Pharmacol 2001; 37: 489–492
5. Wells SM, Holian A. Asymmetric dimethylarginine induces oxidative and nitrosative stress in murine lung epithelial cells. Am J Respir Cell Mol Biol 2007; 36: 520–528
6. Yuan Q, Jiang DJ, Chen QQ, Wang S, Xin HY, Deng HW, Li YJ. Role of asymmetric dimethylarginine in homocysteine-induced apoptosis of vascular smooth muscle cells. Biochem Biophys Res Commun 2007; 356: 880–885
7. Böger RH. Asymmetric dimethylarginine (ADMA) and cardiovascular disease: insights from prospective clinical trials.
8. Visc Med 2005; 10 Suppl 1: S19–S25
9. Vallance P, Leone A, Calver A, Collier J, Moncada S. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. Lancet 1992; 339: 572–575
10. Brown JF, Hanson PJ, Whittle BJ. Nitric oxide donors increase mucus gel thickness in rat stomach. Eur J Pharmacol 1992; 223: 103–104
11. Lopez-Belmonte J, Whittle BJ, Moncada S. The actions of nitric oxide donors in the prevention or induction of injury to the rat gastric mucosa. Br J Pharmacol 1993; 108: 73–78
12. Masuda E, Kawano S, Nagano K, Tsuji S, Takei Y, Tsuji M, Oshita M, Michida T, Kobayashi I, Nakama A. Endogenous nitric oxide modulates ethanol-induced gastric mucosal injury in rats. Gastroenterology 1995; 108: 58–64
13. Franco L, Doria D. Nitric oxide enhances prostaglandin production in alcohol-induced gastric mucosal injury in rats. Eur J Pharmacol 1998; 348: 247–256
14. Ohta Y, Nishida K. L-arginine protects against stress-induced gastric mucosal lesions by preserving gastric mucus. Clin Exp Pharmacol Physiol 2002; 29: 32–38
15. Petersson J, Phillipson M, Jansson EA, Fatzak A, Lundberg JO, Holm L. Dietary nitrate increases gastric mucosal blood flow and mucosal defense. Am J Physiol Gastrointest Liver Physiol 2007; 292: G718–G724
16. Kato S, Kitamura M, Korolikiewicz RP, Takeuchi K. Role of nitric oxide in regulation of gastric acid secretion in rats: effects of NO donors and NO synthase inhibitor. Br J Pharmacol 1998; 123: 839–846
17. von Bohmer C, Edebo A, Lönnroth H, Olle L, Pettersson A, Fändriks L. Helicobacter pylori infection inhibits antral mucosal nitric oxide production in humans. Scand J Gastroenterol 2002; 37: 404–408
18. Fändriks L, von Bohmer C, Johansson B, Holm M, Bölin I, Pettersson A. Water extract of Helicobacter pylori inhibits duodenal mucosal alkaline secretion in anesthetized rats. Gastroenterology 1997; 113: 1570–1575
19. Zhang Z, Zou YY, Zhou Y, Zhou H, Li YJ. The aggravatory effect of nicotine on Helicobacter pylori-induced gastric mucosal injury: role of asymmetric dimethylarginine. J Clin Gastroenterol 2009; 43: 261–266
20. Wang L, Zhou Y, Peng J, Zhang Z, Jiang DJ, Li YJ. Role of endogenous nitric oxide synthase inhibitor in gastric mucosal injury. Can J Physiol Pharmacol 2008; 86: 97–104
21. Lamarque D, Dutreuil C, Dhumeaux D, Delchier JC. Increased gastric bicarbonate secretion in portal hypertensive anesthetized rats: role of prostaglandins and nitric oxide. Dig Dis Sci 1997; 42: 743–750
22. Maity P, Biswas K, Roy S, Banerjee RK, Bandyopadhyay U. Smoking and the pathogenesis of gastroduodenal ulcer—recent mechanistic update. Mol Cell Biochem 2003; 253: 329–338
23. Qui BS, Mei QB, Liu L, Tchou-Wong KM. Effects of nitric oxide on gastric ulceration induced by nicotine and cold-restraint stress. World J Gastroenterol 2004; 10: 594–597
24. Zhang Z, Zhou Y, Zou YY, Wang L, Yang ZC, Guo R, Li D, Peng J, Li YJ. Detrimental effects of nicotine on the acute gastric mucosal injury induced by ethanol: role of asymmetric dimethylarginine. Can J Physiol Pharmacol 2008; 86: 835–840
25. Jiang DJ, Dai S, Li YJ, Asymmetric dimethylarginine induces apoptosis via p38 MAPK/caspase-3-dependent signaling pathway in endothelial cells. J Mol Cell Cardiol 2006; 40: 529–539
26. Jiang JL, Wang S, Li NS, Zhang XH, Deng HW, Li YJ. The inhibitory effect of simvastatin on the ADMA-induced inflammatory reaction is mediated by MAPK pathways in endothelial cells. Biochem Cell Biol 2007; 85: 66–77
27. Liu YZ, Zhou Y, Li D, Wang L, Hu GY, Peng J, Li YJ. Reduction of asymmetric dimethylarginine in the protective effects of ruteacarpine on gastric mucosal injury. Can J Physiol Pharmacol 2008; 86: 675–681

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