Intraspecific variation in response to magnitude and frequency of freeze-thaw cycles in a temperate grass

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Abstract. Winter warming and its accompanying predicted decrease in snow pack for northern temperate regions may increase frost damage to plants induced by an increase in freeze-thaw cycles (FTCs) due to reduced insulation. FTC frequency, minimum temperature during freezing and pre-existing local adaptations potentially all influence site-specific plant responses to future climatic changes. Within a chamber experiment, frost sensitivity towards recurrent FTCs was determined in 12 Dactylis glomerata populations from various European sampling sites differing in temperature and precipitation. After winter hardening, plants were frozen at −4 and −8 °C at frequencies of one, three and seven FTCs within a 1-week treatment phase. The control was kept at 4.5 °C. Plant survival, leaf elongation, chlorophyll content and above-ground net primary productivity (ANPP) decreased with lower minimum temperatures and higher FTC frequencies, while lower freezing temperatures generally proved more influential than increased freezing frequencies. Plant survival rates correlated with the amount of annual precipitation at seed origin, as individuals from comparably drier sites exhibited higher survival rates. This response, however, was limited in its effect to low freezing temperatures (−8 °C) and low and medium freezing frequencies (1 and 3 FTCs). In the set of surviving plants, water availability at seed origin best explained the plants’ growth responses to FTC treatment. The observed intraspecific variation emphasizes the ecological importance of potential local adaptations within a more variable future winter climate.

Keywords: Above-ground net primary productivity (ANPP); central Europe; climate change; cocksfoot; Dactylis glomerata; freeze-thaw cycles; freezing-thawing; winter ecology.

Introduction

Climate change scenarios suggest an increase in the frequency of soil freeze-thaw cycles (FTCs) in many northern temperate regions, currently insulated and thermally stabilized by snow cover (Henry 2008; Pauli et al. 2013).

As strong changes in seasonality and precipitation patterns accompany global warming, air frost (~30 to ~45 %) and snow cover days (~30 to ~40 %) will be strongly reduced across northern Europe (Jylhä et al. 2008). With ~50.5 % of the northern hemisphere’s total land area
seasonally frozen (averaged for 1950–96; Zhang et al. 2003) a rapidly changing winter climate leaves low-stat-ure ecosystems exposed to frost events, as the typically occurring and insulating snow cover decreases (Jylhä et al. 2008). Therefore, ecological processes related to snow cover and soil frost regimes are becoming increasingly important drivers of ecosystem functioning and composition within a warming and increasingly variable climate (Kreyling 2010; Pauli et al. 2013).

Freeze-thaw cycles can disrupt soil microorgan-isn and soil aggregate dynamics, affecting below-and above-ground ecological processes (Oztas and Fayetorbay 2003; Schadt et al. 2003; Six et al. 2004; Sjursen et al. 2005; Vankoughnett and Henry 2013). Their effect on local plant populations however differs significantly depending on their timing, duration, severity and frequency. Short-term warming during winter may deharden acclimatized plant tissue within several days (Strimbeck et al. 1995; Kalberer et al. 2006) or even hours (Rapacz et al. 2000) increasing soil frost-related root injury upon (re-)freezing and in turn negatively affecting fine-root dynamics, primary productivity and nutrient cycling on-site (Fitzhugh et al. 2001; Tierney et al. 2001, 2003; Kreyling et al. 2008, 2010).

Within Europe’s temperate zone, perennial grass species are geographically widely distributed and grow under diverse climatic conditions (Beierkuhnlein et al. 2011). For several grass species, local adaptations to, e.g., precipitation patterns, climatic extremes and winter air temperatures have already been reported (Jentsch and Beierkuhnlein 2010; Kreyling et al. 2010, 2011; Malyshhev et al. 2016), whereas local adaptations to FTCs have not yet been extensively studied. While winter seasonality typically evokes long-term physiological adaptations (Kreyling et al. 2010, 2011), short-term climatic events yield the potential to decimate entire populations (e.g. Jentsch et al. 2007). Thus, inherent local adaptation becomes a valuable strategy against increasing climatic variability (Kreyling et al. 2011).

Here, we tested for local preadaptation of 12 ecotypes to FTCs in cold-acclimated individuals of the temperate grass species Dactylis glomerata, spanning a pan-European climatic gradient within a climate chamber experiment. We hypothesized that (i) plant survival, chlorophyll content, leaf elongation and above-ground net primary productivity (ANPP) decrease with increasing magnitude and frequency of FTC manipulations, while plant injury increases. Furthermore, we also expected (ii) frost adaptation to vary among ecotypes with inherited local adaptation in frost tolerance being related to climatic conditions at seed origin, such as temperature, altitude and/or precipitation.

Methods

Plant material
In June 2013 (July for Swedish seeds), D. glomerata seeds were collected from European grasslands along a broad climatic gradient, where, to expert knowledge, the target species was autochthonous (Fig. 1). At least 20 individuals were sampled per site, with minimum sampling distances between sampled plants being >1 m. Corresponding climate data for each seed source were extracted from the WorldClim data sets (Hijmans et al. 2005; Table 1).

Spanish and Ukrainian seeds originated from two locations, Swedish seeds from four different locations within their respective country of origin. The maximal distance separating these locations averaged 18 km for the Swedish, 19.5 km for the Spanish and 200 km for the Ukrainian seeds. The remaining nine ecotypes were represented by one location exclusively. Swedish, Spanish and Ukrainian climate parameters were averaged across their respective locations due to their close geographical proximity and/or comparative climate on site. The climatic data sets varied by a maximum of 0.4 °C mean annual temperature and 4 mm in annual precipitation for both the Swedish and Spanish locations. Ukrainian sampling sites exhibited a difference of 0.3 °C and 43 mm, respectively.

Plants growth and cold acclimation
Individuals from all populations were grown under standardized conditions at the University of Bayreuth, Germany. Following germination in February 2014, plants were grown in seedling trays outside (protected from frost by glass frames) before being transplanted into pots measuring 5 cm in diameter and 7 cm in depth, filled with homogenized ED 73 substrate (Einheitserde Werkverband e.V.). Plants were watered at a rate that kept the soil constantly moist.

Six weeks after transplantation plants were cut to a height of 2 cm and placed into seven identical plant seed trays, later randomly partitioned to represent the control and the six treatment groups with five individuals per each of 60 plants per tray. Trays positions were rearranged every other day. Plant arrangement within each tray was systematic, in order to control for edge and neighbouring effects between ecotypes. Plant cold acclimation commenced with a 10-h photoperiod, initiating cold acclimation in separate climate chambers (Adaptis by Conviron, model number: A1000) for all trays. The temperature was set to 6 °C during the night and 12 °C during the day. Plants were watered weekly and light intensity ranged between 70 and
Figure 1. Map displaying the 12 seed origins of Dactylis glomerata used in the experiment. Colouring indicates the lowest temperature (°C) in January above European landmasses, as extracted from the WorldClim data set (Hijmans et al. 2005). Multiple sites of seed origin for single countries are displayed (see UA) but overlay for SE and ES.

Table 1. Geographic and climatic characteristics at the 12 seed sampling sites, as extracted from WorldClim data sets (Hijmans et al. 2005). Shown are latitude (°, LAT, Northern direction), longitude (°, LON, Eastern direction), precipitation variability (PV), mean annual precipitation (mm, MAP), temperature of the warmest month (°C, TWM), temperature of the coldest month (°C, TCM), temperature of the warmest quarter (°C, TWQ), temperature of the coldest quarter (°C, TCQ), mean annual temperature (°C, MAT), altitude (m, ALT), temperature range (TR) and mean precipitation during the warmest quarter (mm, PWQ) and coldest quarter (mm, PCQ) for each site. Ukrainian and Spanish seeds originated from two, Swedish seeds from four locations with maximal distances between sampling sites of 200, 19.5 and 18 km, respectively.

| Country | LAT  | LON  | PV  | MAP  | TWM  | TCM  | TWQ  | TCQ  | MAT  | ALT  | TR  | PWQ | PCQ |
|---------|------|------|-----|------|------|------|------|------|------|------|-----|-----|-----|
| Austria | 47.07| 11.18| 28  | 1142 | 13.3 | −10  | 8.3  | −6.2 | 1.1  | 2041 | 7   | 406 | 146 |
| Belgium | 51.15| 4.40 | 12  | 778  | 22.4 | −0.2 | 17.1 | 3    | 10.2 | 9    | 7.7 | 198 | 75  |
| Switzerland | 46.44| 8.94 | 25  | 1417 | 18.6 | −5.3 | 13.3 | −1.8 | 5.7  | 1290 | 6.9 | 468 | 163 |
| Germany | 49.92| 11.58| 20  | 674  | 22.5 | −3.7 | 16.3 | −0.3 | 8    | 426  | 8.4 | 221 | 76  |
| Spain   | 42.77| −3.29| 20  | 798  | 23.9 | 1    | 17.5 | 4.6  | 10.8 | 753  | 8.8 | 151 | 89  |
| France  | 45.38| 2.44 | 18  | 806  | 24.1 | −1.4 | 16.9 | 2.8  | 9.9  | 596  | 10.3| 218 | 94  |
| Hungary | 46.46| 19.58| 25  | 552  | 27.1 | −8.3 | 20.2 | 0.5  | 10.8 | 123  | 9.6 | 176 | 72  |
| Italy   | 43.10| 13.05| 18  | 881  | 26.6 | 0.5  | 20.5 | 4    | 12   | 649  | 7.5 | 194 | 101 |
| Ukraine | 48.44| 30.13| 31  | 590  | 24.7 | −8   | 18.7 | −3.8 | 7.7  | 195  | 8.2 | 214 | 83  |
| Sweden  | 50.28| 30.27| 29  | 633  | 24.5 | −8.4 | 18.8 | −4.1 | 7.7  | 175  | 8.1 | 228 | 87  |
| Turkey  | 62.94| 17.79| 24  | 656  | 20   | −13  | 14.1 | −7.9 | 3    | 105  | 8.5 | 191 | 74  |
| UK      | 62.88| 18.11| 24  | 659  | 20.3 | −12.8| 14.4 | −7.8 | 3.2  | 81   | 8.3 | 188 | 75  |
|         | 62.87| 18.11| 24  | 659  | 20.3 | −12.8| 14.4 | −7.8 | 3.2  | 81   | 8.3 | 188 | 75  |
|         | 62.85| 18.09| 24  | 659  | 20.3 | −12.8| 14.4 | −7.8 | 3.2  | 81   | 8.3 | 188 | 75  |
|         | 38.56| 27.39| 73  | 769  | 31.6 | 1.1  | 26.3 | 5.8  | 14.4 | 503  | 11.4| 35  | 157 |
|         | 51.07| −2.16| 19  | 804  | 20.8 | 0.3  | 15.3 | 3.5  | 9.1  | 145  | 7.5 | 169 | 90  |
After 2 weeks, to enhance cold acclimation, the light duration was reduced to 8 h and the temperature to 4 °C during both day and night. Cold acclimation continued for 1 month, with 10 plants dying during this period.

**FTC manipulation**

The FTC manipulations were administered in a three-factorial design, manipulating: (i) FTC frequency (zero for control plants and one, three and seven cycles each for the respective treatment trays), (ii) different minimum temperatures (4.5 °C for the control and −4 and −8 °C for each freezing treatment) and (iii) plant ecotypes (12 ecotypes). Each factorial combination was replicated five times, adding up to 420 plants in total.

Freeze-thaw cycle manipulations lasted a total of 7 days (Fig. 2). In order to simulate natural conditions, plants experienced thawing at 4.5 °C during the day and night-time freezing down to −4 or −8 °C in separate climate chambers (Fig. 2). For treatment, plants were moved from the control chamber (4.5 °C) to one of two treatment chambers set at either −4 or −8 °C. The temperature inside the chambers was measured using temperature loggers (HOBO Pro v2, Onset Computer Corporation, MA, USA). The last freezing cycle overlapped with the last day of treatment to affect all plants equally. Following the freezing treatments, plants were transferred into the greenhouse for a growth period of 2 weeks at an average mean air temperature of 20 °C and 43 % mean relative humidity during the day in order to simulate natural spring conditions.

**Response parameters**

Initial plant survival was determined at the end of the 2-week post-frost growing phase in the greenhouse and transformed into relative survival rates for each factorial combination. Among the surviving plants, ANPP was quantified as the complete, dried harvest of above-ground biomass. In order to accurately estimate plant injury as a response to FTC manipulations, plant tissue was separated into categories of ‘living’ (green tissue) and ‘dead’ (brown tissue) based on visual inspection. It was subsequently dried for 48 h at 75 °C and weighed separately. Plant injury itself was later determined as percent dry mass of ‘dead’ tissue relative to overall ANPP.

Leaf lengths of predominantly green leaves were measured at the end of the treatment phase and at the end of the growth phase in the greenhouse. Both measurements were averaged for three different leaves of each plant. Leaf elongation was calculated by subtracting the mean leaf length at the end of the treatment phase from the average leaf length at the end of the greenhouse growing period to clearly distinguish treatment effects from pre-existing morphologies.

Leaf chlorophyll content was estimated using a SPAD 502 Plus Chlorophyll Meter (Konica Minolta). Measurements were repeated once at three different leaves at 1 (t = 1) and 2 weeks (t = 2) upon treatment.
completion and averaged, after we found no significant differences between the two sampling dates.

**Data analysis**

In order to test for the effects of FTC frequency (freq), magnitude (magn) and seed origin (orig) on plant survival rates and within the set of surviving plants on ANPP, plant injury, leaf elongation and chlorophyll content three-factorial analyses of variance (ANOVAs) relating survival to freq, magn and orig were applied, with the latter being treated as a categorical factor. These analyses also tested for statistical interactions between all three factors and estimated their relative effect sizes ($\eta^2$, Cohen 1988) within the three-factorial ANOVA. Separate analyses to subsets of plants frozen at either −4 or −8 °C were applied in order to check for frequency or origin effects potentially masked in the three-factorial analyses.

To further identify the impact of inherited local adaptation on plant survival and plant parameter variation, the categorical factor of seed origin was transformed into numeric parameters of ecological importance, as fixed effects in interaction with freq and magn. Readily available factors used herein were: altitude, temperature of the coldest and warmest months, temperature of the coldest and warmest quarters, annual precipitation and its variability, mean annual temperature, annual temperature range (max. temperature of the warmest month − min. temperature of the coldest month) and the average precipitation values of the coldest and warmest quarters as extracted from the WorldClim data sets (Hijmans et al. 2005). The colinearity of these factors prohibited a direct comparison among them, but allowed for an independent identification of the variable(s) best explaining plant treatment responses.

In order to control for a possibly inflated type I error as a result of multiple comparisons, we applied false discovery rate testing (Storey et al. 2004). Subsequently, results with a $P$-value of $\alpha \leq 0.05$ and a corresponding $q$-value of $\alpha > 0.05$ were disregarded as non-significant, as were results with a $P$-value of $\alpha > 0.05$.

All response parameters were checked for their normal distribution using the Shapiro–Wilk test, visual inspection of frequency histograms and normal q-q plots. Both ANPP and leaf elongation values were consequently square-root transformed for the ANOVA analyses (non-transformed values are shown in the figures). All analyses were performed using R 3.4.3 (R Core Team 2017), installing additional ‘raster’ (Hijmans 2017), ‘heplots’ (Fox et al. 2017), ‘qvalue’ (Dabney and Storey 2017) and ‘rgdal’ packages (Bivand et al. 2017).

**Table 2.** Absolute number of plant deaths following the treatment phase in response to both treatment type (FTC frequency and magnitude) and country of seed origin ($n = 5$ per factorial combination). Treatment combinations are sorted by temperature (4.5, −4 and −8 °C) and freezing frequency: low (one FTC), medium (med, three FTCs) and high (seven FTCs). The respective countries are ranked by the amount of annual precipitation received, ranging from lowest (Hungary) to highest (Switzerland), as extracted from the WorldClim data sets (Hijmans et al. 2005). The amount of annual precipitation on-site proved to be most influential on survival patterns and mortality following freezing treatment. In total, 72 out of 410 individuals died during treatment.

| Country | 4.5 °C | −4 °C | −8 °C | Σ = 72 |
|---------|--------|-------|-------|--------|
|         | Control | Med   | High  | Low    | Med   | High  | Σ = 72 |
| Hungary | 1       | 1     |       | 2      | 1     | 5     | 72     |
| Ukraine |        |       |       | 2      | 1     | 3     |        |
| Sweden  |        |       |       | 3      | 4     | 7     |        |
| Germany |        | 1     | 2     | 2      | 1     | 3     | 8      |
| Turkey  | 1       | 1     | 3     | 3      |      | 8     |        |
| Belgium | 1       | 1     | 3     | 1      | 5     |      |        |
| Spain   | 2       | 2     | 4     |        |       |       |        |
| UK      | 2       | 1     | 2     | 5      |       |       |        |
| France  | 2       | 1     | 4     | 7      |       |       |        |
| Italy   | 1       | 1     | 1     | 2      | 2     | 7     |        |
| Austria | 1       | 3     | 2     | 6      |       |       |        |
| Switzerland | 1 | 3 | 3 | 3 | 10 | | |
| Σ = 72 | 2 | 1 | 2 | 4 | 9 | 27 | 27 | 72 |
Results

Post-frost plant survival—FTC magnitude vs. frequency

The control plants exhibited the highest survival rate (96.7 %), followed by plants undergoing freezing at −4 °C (96.1 %) and −8 °C (62.5 %; Table 2). Survival rates among plants experiencing freezing at −4 °C decreased with increasing freezing frequency from 98.3 to 96.7 and 93.2 %. This effect was only partly observed in the −8 °C category: following the addition of two additional freezing cycles (one FTC vs. three FTCs) survival rates decreased by two-thirds, but did not further decrease upon the addition of four additional freezing cycles (three FTCs vs. seven FTCs).

Post-frost plant survival—climatic influence at seed origin

Mean annual precipitation at seed origin was the most influential climatic parameter explaining survival patterns across ecotypes (Fig. 3). Annual precipitation at the site of seed origin (P ≤ 0.01, $\eta^2_p = 0.08$) and freezing magnitude (P ≤ 0.001, $\eta^2_p = 0.27$) proved to have the strongest effect on survival rates. Freezing frequency × severity interactions (P ≤ 0.001, $\eta^2_p = 0.36$) were significant. Typically, higher precipitation values at seed origin coincided with a decreased survival rate, as seen, for example, in the comparison of Hungary, exhibiting fairly low annual precipitation values with Switzerland representing the site with the highest amount of annual precipitation and the lowest survival rate overall (Table 2; Fig. 3). The effect of precipitation at seed origin on plant survival was most pronounced at low freezing temperatures (−8 °C) and low and medium freezing frequencies (three FTCs). Hence, ecotypes with a lower amount of annual precipitation at seed origin tended to be more resilient to freezing manipulations.

Post-frost plant responses—FTC magnitude vs. frequency

Within the set of surviving plants (n = 338) all measured response parameters (ANPP, plant injury, leaf elongation and chlorophyll content) were affected by the factors seed origin (orig), freezing magnitude (magn) and frequency (freq) and any factorial combination thereof (orig × magn, orig × freq, magn × freq, orig × freq × magn). Bold face characters connote statistically significant responses. The significance level is set to $P = 0.05$.

Table 3. Statistical effects ($P$, $F$, $\eta^2_p$) of the three-factorial ANOVA for ANPP, plant injury, leaf elongation and chlorophyll content in dependence of the categorical factors seed origin (orig), freezing magnitude (magn) and frequency (freq) and any factorial combination thereof (orig × magn, orig × freq, magn × freq, orig × freq × magn). Bold face characters connote statistically significant responses. The significance level is set to $P = 0.05$.

|                      | ANPP (g) | Plant injury (%) | Leaf elongation (cm) | Chlorophyll content |
|----------------------|----------|------------------|----------------------|---------------------|
|                      | $P$ | $F$ | $\eta^2_p$ | $P$ | $F$ | $\eta^2_p$ | $P$ | $F$ | $\eta^2_p$ |
| orig                 |    |     |            |    |     |            |    |     |            |
| magn                 |< 0.001 | 73.8 | 0.15       |< 0.001 | 97.2 | 0.19       |< 0.001 | 57.7 | 0.16       |
| freq                 | 0.090  | 2.9  | 0.01       |< 0.05 | 5.6  | 0.02       | 0.126  | 2.4  | 0.01       |
| orig × magn          | 0.921  | 0.5  | 0.01       | 0.982 | 0.3  | 0.01       | 0.388  | 1.1  | 0.04       |
| orig × freq          | 0.622  | 0.8  | 0.03       | 0.541 | 0.9  | 0.03       | 0.324  | 1.1  | 0.05       |
| magn × freq          |< 0.001 | 31.2 | 0.10       |< 0.001 | 49.6 | 0.14       |< 0.001 | 38.7 | 0.13       |
| orig × freq × magn   | 0.424  | 1.0  | 0.04       | 0.778 | 0.7  | 0.02       | 0.459  | 1.0  | 0.04       |

Figure 3. Survival rates for individuals frozen at −8 °C in dependence of mean annual precipitation values at the site of seed origin. Overall, plant survival decreases with an increase in mean annual precipitation values at the site of seed origin ($P < 0.01$, $r^2 = 0.17$, for individuals frozen at −8 °C independent of freezing frequency). Individuals adapted to a lower amount of annual precipitation at seed origin tended to be more resilient at lower freezing temperatures (see Table 2: Hungary, Ukraine, Sweden, Germany).
elongation, chlorophyll content) were significantly influenced by freezing magnitude and magnitude × frequency interactions (Table 3). In addition, chlorophyll content and plant injury were significantly affected by freezing frequency as a stand-alone factor.

Above-ground net primary productivity responded negatively to an increase in freezing magnitude, as it exhibited its lowest values at −8 °C. The effect of freezing frequency on ANPP resulted in a significantly increasing loss of primary productivity of 29.9, 41.2 and 50.0% with increasing freezing frequency levels at low freezing temperatures (−8 °C; Fig. 4). Plant injury increased with increasing FTC magnitude and frequency. Compared to freezing frequency, the effect sizes of freezing magnitude proved to be higher for both ANPP (η^2_p = 0.15 vs. η^2_p = 0.01) and plant injury (η^2_p = 0.19 vs. η^2_p = 0.02). Plants frozen at −8 °C exhibited significant leaf elongation losses of 50.5, 95.2 and 92.9%, after one, three and seven FTCs, respectively. Increasing the FTC frequency from one to three cycles proved to have a more pronounced effect on leaf elongation variation than the further increase to seven cycles. The effect of freezing temperature (η^2_p = 0.16) outweighed that of freezing frequency (η^2_p = 0.01).

Mean chlorophyll content decreased with increasing freezing frequencies for both plants frozen at −4° and −8 °C in comparison to untreated individuals, with the latter exhibiting a lower chlorophyll content (Fig. 4). Chlorophyll content positively correlated with ANPP under freezing conditions. Here, freezing magnitude exhibited similar effect sizes (η^2_p = 0.03) as freezing frequency (η^2_p = 0.03).

Post-frost plant responses—climatic influence at seed origin
Following the FTC manipulations, ecotypes from comparatively colder climates grew faster than their counterparts (Fig. 5). Leaf elongation correlated negatively with the temperature of the warmest month, the warmest quarter and the annual mean temperature, as well as the temperature and precipitation of the coldest quarter at seed origin. Significant interactions between freezing treatment and climate conditions at seed origin were found for mean annual precipitation and precipitation of the warmest quarter with freezing frequency (Table 4).

Plant injury positively correlated with the amount of mean annual precipitation at seed origin (P ≤ 0.05, η^2_p = 0.02). Ecotypes experiencing a comparatively low amount of annual precipitation tended to be less sensitive to injury at −4 °C.

**Discussion**
Survival of D. glomerata individuals across populations of different geographic origin was negatively affected by lower freezing temperatures and increasing freezing
frequencies, most likely due to frost-induced tissue damage and dieback (Bokhorst et al. 2009; Wipf et al. 2009). Notably, increasing freezing frequencies from three to seven FTCs did not further decrease survival chances for plants frozen at −8 °C, which suggests that freezing frequency is limited in its effect, with additional FTCs not being more detrimental following three FTCs. This supposed limitation of freezing frequency was mirrored in subsequent response analyses of chlorophyll content, leaf elongation, plant injury and ANPP among the surviving individuals, as freezing magnitude outweighed freezing frequency in its respective effect size.

Overall, survival rates, primary production, leaf elongation and chlorophyll content decreased with freezing magnitude, while plant injury increased. This increase of frost damage at temperatures likely to occur within a snow-free environment stresses the importance of diminishing soil insulation. Without snow, the lack of insulation leaves above- and below-ground biota exposed to freezing temperatures (Fitzhugh et al. 2001; Hardy et al. 2001; Maljanen et al. 2007). This may result in an increase of fine-root necromass and an overall decrease of root biomass within the organic layer (Groffman et al. 2001; Gaul et al. 2008). A plant’s ability to take up nutrients is in turn reduced (Campbell et al. 2014). In agreement with our findings, recent field experiments in temperate grasslands indicate that this loss of insulation and an exposure to cyclic freezing result in a decrease in ANPP for at least one growing season (Schuerings et al. 2014).

Out of the 11 climatic parameters used to characterize the locations of seed origin, the amount of annual precipitation correlated best with a population’s survival chances. Plants from comparably drier sites exhibited higher survival rates. While this effect was prominent at −8 °C and low and medium freezing frequencies, independent treatment effects increased at high freezing frequencies. This indicates that, while certain plants are better equipped to survive a short amount of time (here 12 h) under frost stress, conditions of recurrent freezing at −8 °C exceed ecotype-specific tolerances. Previous experiments with shrubs (Boorse et al. 1998) and woody plants (Nardini et al. 2000; Nilsson 2001; Rehfeldt et al. 2002) have revealed patterns of intraspecific variation in the cold sensitivity of plants corresponding to the environmental conditions of their origin. In addition, Poirier et al. (2012) showed that following a growing season under conditions of restricted water supply, temperate D. glomerata populations exhibited lower amounts of induced frost damage the following
Table 4. Overview over plant responses to treatment, including the control. The three-factorial ANOVA considers climatic conditions at the sites of seed origin, as well as freezing frequency and magnitude co-variates. Shown here are singular ecotype effects, as well as interactions between ecotype and either freezing magnitude (magn), freezing frequency (freq) or both (magn × freq). Effect directionality is denoted by +/−, with ‘+’ indicating a positive correlation between the environmental and plant parameters and ‘−’ a negative one. n.a. stands for ‘not available’ and denotes non-significant results. Environmental factors used herein are extracted from the WorldClim data sets (Hijmans et al. 2005) and include precipitation variability (PV), mean annual precipitation (mm, MAP), temperature of warmest month (°C, TWM), temperature of coldest month (°C, TCM), temperature of warmest quarter (°C, TWQ), temperature of coldest quarter (°C, TCQ), mean annual temperature (°C, MAT), altitude (m, ALT), temperature range (°C), and mean precipitation during the warmest quarter (mm, PWQ) and coldest quarter (mm, PCQ). Bold face characters indicate a statistically significant relationship. The significance level is set to $P = 0.05$ and $q = 0.05$. 

|              | ANPP (g) | Plant injury (%) | Leaf elongation (cm) | Chlorophyll content |
|--------------|----------|-----------------|----------------------|---------------------|
|              | $P$      | $F$             | $\eta^2$             | +/-                 | $P$      | $F$             | $\eta^2$             | +/-                 | $P$      | $F$             | $\eta^2$             | +/-                 |
| PV           | 0.549    | 0.4             | 0.00 n.a.            | 0.934 0.0 0.00 n.a. | 0.250 1.3 0.01 n.a. | 0.279 1.2 0.00 n.a. |
| PV × magn    | 0.519    | 0.4             | 0.00 n.a.            | 0.448 0.6 0.00 n.a. | 0.443 0.6 0.00 n.a. | 0.771 0.1 0.00 n.a. |
| PV × freq    | 0.773    | 0.1             | 0.00 n.a.            | 0.863 0.0 0.00 n.a. | 0.883 0.0 0.00 n.a. | 0.108 2.6 0.01 n.a. |
| PV × magn × freq | 0.870    | 0.0             | 0.00 n.a.            | 0.756 0.1 0.00 n.a. | 0.134 2.3 0.01 n.a. | 0.325 1.0 0.00 n.a. |
| MAP          | 0.320    | 0.9             | 0.00 n.a.            | <0.05 5.8 0.02 +   | 0.056 3.7 0.01 n.a. | 0.846 0.0 0.00 n.a. |
| MAP × magn   | 0.993    | 0.0             | 0.00 n.a.            | 0.603 0.3 0.00 n.a. | 0.169 1.9 0.02 n.a. | 0.134 2.3 0.01 n.a. |
| MAP × freq   | 0.781    | 0.1             | 0.00 n.a.            | 0.222 1.5 0.00 n.a. | <0.01 6.2 0.02 n.a. | 0.819 0.1 0.00 n.a. |
| MAP × magn × freq | 0.493    | 0.5             | 0.00 n.a.            | 0.227 1.5 0.00 n.a. | 0.092 2.9 0.01 n.a. | 0.455 0.6 0.00 n.a. |
| TWM          | 0.736    | 0.1             | 0.00 n.a.            | 0.236 1.4 0.01 n.a. | <0.01 10.3 0.04 n.a. | 0.312 1.0 0.00 n.a. |
| TWM × magn   | 0.433    | 0.6             | 0.00 n.a.            | 0.555 0.4 0.00 n.a. | 0.334 0.9 0.00 n.a. | 0.293 1.1 0.00 n.a. |
| TWM × freq   | 0.397    | 0.7             | 0.00 n.a.            | 0.786 0.1 0.00 n.a. | 0.064 3.5 0.01 n.a. | 0.565 0.3 0.00 n.a. |
| TWM × magn × freq | 0.969    | 0.0             | 0.00 n.a.            | 0.264 1.3 0.00 n.a. | 0.671 0.2 0.00 n.a. | 0.929 0.0 0.00 n.a. |
| TCM          | 0.479    | 0.5             | 0.00 n.a.            | 0.215 1.5 0.01 n.a. | 0.341 0.9 0.00 n.a. | 0.853 0.0 0.00 n.a. |
| TCM × magn   | 0.449    | 0.6             | 0.00 n.a.            | 0.833 0.0 0.00 n.a. | 0.062 3.5 0.01 n.a. | 0.413 0.7 0.00 n.a. |
| TCM × freq   | 0.957    | 0.0             | 0.00 n.a.            | 0.263 1.3 0.00 n.a. | 0.838 0.0 0.00 n.a. | 0.514 0.4 0.00 n.a. |
| TCM × magn × freq | 0.877    | 0.0             | 0.00 n.a.            | 0.917 0.0 0.00 n.a. | 0.966 0.0 0.00 n.a. | 0.703 0.1 0.00 n.a. |
| TWQ          | 0.779    | 0.1             | 0.00 n.a.            | 0.295 1.1 0.00 n.a. | <0.001 12.8 0.05 n.a. | 0.252 1.3 0.00 n.a. |
| TWQ × magn   | 0.461    | 0.5             | 0.00 n.a.            | 0.469 0.5 0.00 n.a. | 0.293 1.1 0.00 n.a. | 0.331 0.9 0.00 n.a. |
| TWQ × freq   | 0.341    | 0.9             | 0.00 n.a.            | 0.811 0.1 0.00 n.a. | 0.072 3.3 0.01 n.a. | 0.391 0.7 0.00 n.a. |
| TWQ × magn × freq | 0.960    | 0.0             | 0.00 n.a.            | 0.194 1.7 0.01 n.a. | 0.662 0.2 0.00 n.a. | 0.896 0.0 0.00 n.a. |
| TCQ          | 0.352    | 0.9             | 0.00 n.a.            | 0.407 0.7 0.00 n.a. | <0.01 10.4 0.04 n.a. | 0.380 0.8 0.00 n.a. |
Table 4. Continued

|                | ANPP (g) | Plant injury (%) | Leaf elongation (cm) | Chlorophyll content |
|----------------|----------|------------------|----------------------|---------------------|
|                | p        | F                | $\eta^2$             | +/-                 | p        | F    | $\eta^2$ | +/-     | p        | F    | $\eta^2$ | +/-     |
| TCQ × magn     | 0.511    | 0.4              | 0.00                 |                      | 0.747    | 0.1  | 0.00     |          | 0.123    | 2.4  | 0.00     |          | 0.752    | 0.1  | 0.00     |
| TCQ × freq     | 0.286    | 1.2              | 0.00                 |                      | 0.261    | 1.3  | 0.00     |          | 0.119    | 2.4  | 0.01     |          | 0.252    | 1.3  | 0.00     |
| TCQ × magn × freq | 0.387 | 0.8              | 0.00                 |                      | 0.415    | 0.7  | 0.00     |          | 0.486    | 0.5  | 0.00     |          | 0.928    | 0.0  | 0.00     |
| MAT × freq     | 0.485    | 0.5              | 0.00                 | n.a.                | <0.001   | 13.9 | 0.05     | –        | 0.925    | 0.0  | 0.00     | n.a.     | 0.0      | 0.0  | 0.00     |
| MAT × magn     | 0.401    | 0.7              | 0.00                 |                      | 0.567    | 0.3  | 0.00     |          | 0.136    | 2.2  | 0.00     |          | 0.465    | 0.5  | 0.00     |
| MAT × freq × magn | 0.625 | 0.2              | 0.00                 |                      | 0.226    | 1.5  | 0.00     |          | 0.563    | 0.3  | 0.00     |          | 0.985    | 0.0  | 0.00     |
| ALT × freq     | 0.080    | 3.1              | 0.01                 | n.a.                | 0.244    | 1.4  | 0.01     | n.a.     | 0.932    | 0.0  | 0.00     | n.a.     | 0.707    | 0.1  | 0.00     |
| ALT × magn     | 0.770    | 0.1              | 0.00                 |                      | 0.395    | 0.7  | 0.00     |          | 0.240    | 1.4  | 0.01     |          | 0.147    | 2.1  | 0.01     |
| ALT × freq × magn | 0.855 | 0.0              | 0.00                 |                      | 0.470    | 0.5  | 0.00     |          | 0.143    | 2.2  | 0.01     |          | 0.729    | 0.1  | 0.00     |
| ALT × magn × freq | 0.858 | 0.0              | 0.00                 |                      | 0.150    | 2.1  | 0.01     |          | 0.253    | 1.3  | 0.00     |          | 0.515    | 0.4  | 0.00     |
| TR × freq      | 0.371    | 0.8              | 0.00                 | n.a.                | 0.052    | 3.8  | 0.01     | n.a.     | 0.965    | 0.0  | 0.00     | n.a.     | 0.968    | 0.0  | 0.00     |
| TR × magn      | 0.325    | 1.0              | 0.00                 |                      | 0.657    | 0.2  | 0.00     |          | 0.769    | 0.1  | 0.00     |          | 0.152    | 2.1  | 0.00     |
| TR × freq × magn | 0.947 | 0.0              | 0.00                 |                      | 0.711    | 0.1  | 0.00     |          | 0.074    | 3.2  | 0.01     |          | 0.178    | 1.8  | 0.01     |
| PWQ × freq     | 0.275    | 1.2              | 0.00                 | n.a.                | 0.107    | 2.6  | 0.01     | n.a.     | 0.787    | 0.1  | 0.00     | n.a.     | 0.846    | 0.0  | 0.00     |
| PWQ × magn     | 0.784    | 0.1              | 0.00                 |                      | 0.781    | 0.1  | 0.00     |          | 0.555    | 0.4  | 0.01     |          | 0.169    | 1.9  | 0.01     |
| PWQ × freq × magn | 0.745 | 0.1              | 0.00                 |                      | 0.533    | 0.4  | 0.00     | <0.05    | 6.4      | 0.02  |          |          | 0.895    | 0.0  | 0.00     |
| PCQ × freq     | 0.960    | 0.0              | 0.00                 | n.a.                | 0.377    | 0.8  | 0.00     | n.a.     | <0.05    | 5.4  | 0.02     | –        | 0.956    | 0.0  | 0.00     |
| PCQ × magn     | 0.638    | 0.2              | 0.00                 |                      | 0.416    | 0.7  | 0.00     |          | 0.228    | 1.5  | 0.00     |          | 0.837    | 0.0  | 0.00     |
| PCQ × freq × magn | 0.858 | 0.0              | 0.00                 |                      | 0.598    | 0.3  | 0.00     |          | 0.963    | 0.0  | 0.00     |          | 0.343    | 0.9  | 0.00     |
| PCQ × magn × freq | 0.436 | 0.6              | 0.00                 |                      | 0.721    | 0.1  | 0.00     | <0.05    | 4.1      | 0.01  |          |          | 0.226    | 1.5  | 0.01     |
winter than Mediterranean populations. Physiological plant responses to drought and frost stress are similar at cellular levels (Beck et al. 2007), implying cross-stress tolerance (Walter et al. 2013), i.e. drought-adapted plants being more frost-tolerant. Detecting species and ecotypes which show superior cross-stress tolerance and stress memory might therefore be a potential adaptation strategy in agriculture against the projected increase in climate-driven extreme weather events (IPCC 2012).

Furthermore, our results imply that while temperature might be an influential factor in determining overall plant growth patterns, a populations’ reaction to freezing is much more likely to be dependent on precipitation patterns at seed origin, as interactions between freezing manipulations and all precipitation-related parameters (MAP, PWQ, PCQ) proved significant. Northern ecotypes grew faster following freezing, likely due to a higher frost tolerance accumulated prior to treatment. Recently, Malyshiev et al. (2014) have shown that following exposure to colder temperatures southern grass ecotypes cold acclimate at a slower rate, maintaining growth longer than their northern counterparts. This potentially beneficial growth advantage however increases the plants’ vulnerability, as subsequent frost events threaten to injure lesser cold-acclimated tissue. This is of special importance, as FTCs are likely to increase especially in regions where the current mean winter temperature is close to the freezing point (Henry 2008).

Conclusions

Herein we show that FTCs, which will become more common in vast regions where winter snow cover is decreasing, negatively affect plant performance. Freezing magnitude proved more detrimental than freezing frequency. Our results also suggest that a populations’ survival chances are likely to be dependent on precipitation patterns at seed origin, as individuals from drier origins exhibited higher survival rates. Among surviving plants, leaf elongation patterns were linked to water availability year-round, as well as during the warmest and coldest quarters. For plant injury, ANPP and chlorophyll content, treatment responses were similar across ecotypes. Local adaptations and within-species variation therefore might only provide a certain degree of buffering against adverse effects of (winter) climate change.

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Contributions by the Authors

A.J. and J.K. conceived the project; A.V.M., J.K. and C.C.D. designed and set up the manipulation experiment; C.C.D. sampled and analysed the data; C.C.D. wrote the paper with substantial input from A.V.M. and contributions from all authors.

Conflict of Interest

None declared.

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Literature Cited

Beck EH, Fettig S, Knake C, Hartig K, Bhattarai T. 2007. Specific and unspecific responses of plants to cold and drought stress. Journal of Biosciences 32:501–510.

Beierkuhnlein C, Thiel D, Jentsch A, Willner E, Kreyling J. 2011. Ecotypes of European grass species respond differently to warming and extreme drought. Journal of Ecology 99:703–713.

Bivand R, Keitt T, Rowlingson B. 2017. rgdal: Bindings for the ‘Geospatial’ Data Abstraction Library. R package version 1.2-16. https://CRAN.R-project.org/package=rgdal (6 December 2017).

Bokhorst SF, Bjerke JW, Tømmervik H, Callaghan TV, Phoenix GK. 2009. Winter warming events damage sub-Arctic vegetation: consistent evidence from an experimental manipulation and a natural event. Journal of Ecology 97:1408–1415.

Boose GC, Ewers FW, Davis SD. 1998. Response of chaparral shrubs to below-freezing temperatures: acclimation, ecotypes, seedlings vs. adults. American Journal of Botany 85:1224–1230.

Campbell JL, Socci AM, Templer PH. 2014. Increased nitrogen leaching following soil freezing is due to decreased root uptake in a northern hardwood forest. Global Change Biology 20:2663–2673.

Cohen J. 1988. Statistical power analysis for the behavioral sciences, 2nd edn. Hillsdale, NJ: Lawrence Earlbaum Associates.

Dabney A, Storey JD. 2017. qvalue: q-value estimation for false discovery rate control. R package version 1.43.0. https://bioconductor.org/packages/release/bioc/html/qvalue.html (6 December 2017).

Fitzhugh RD, Driscoll CT, Groffman PM, Tierney GL, Fahey TJ, Hardy JP. 2001. Effects of soil freezing disturbance on soil solution nitrogen, phosphorous, and carbon chemistry in a northern hardwood ecosystem. Biogeochemistry 56:215–238.

Fox J, Friendly M, Monette G. 2017. heplots: Visualizing Tests in Multivariate Linear Models. R package version 1.3-4. https://CRAN.R-project.org/package=heplots (6 December 2017).
Dietrich et al. – Responses to freeze-thaw cycles in Dactylis glomerata

Gaul D, Hertel D, Leuschner C. 2008. Effects of experimental soil frost on the fine-root system of mature Norway spruce. Journal of Plant Nutrition and Soil Science 171:690–698.

Groffman PM, Driscoll CT, Fahey TJ, Hardy JP, Fitzhugh RD, Tierney GL. 2001. Colder soils in a warmer world: a snow manipulation study in a northern hardwood forest ecosystem. Biogeochemistry 56:135–150.

Hardy JP, Groffman PM, Fitzhugh RD, Henry KS, Weilman AT, Demers JD, Fahey TJ, Driscoll CT, Tierney GL, Nolan S. 2001. Snow depth manipulation and its influence on soil frost and water dynamics in a northern hardwood forest. Biogeochemistry 56:151–174.

Henry HAL. 2008. Climate change and soil freezing dynamics: historical trends and projected changes. Climatic Change 87:421–434.

Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005. Very high resolution interpolated climatological surfaces for global land areas. International Journal of Climatology 25:1965–1978.

Hijmans RJ. 2017. raster: Geographic Data Analysis and Modeling. R package version 2.6-7. https://CRAN.R-project.org/package=raster (6 December 2017).

IPCC. 2012. Summary for policymakers. In: Field CB, Barros V, Stocker TF, Qin D, Dokken DJ, Ebi KL, Estrada YO, Estrada YO, Kirtman B, Kantar T, Mastrandrea MD, Miles L, Murola K, et al. Climate Change 2012: Impacts, Adaptation, and Vulnerability. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge, UK: Cambridge University Press, 1–19.

Jentsch A, Beierkuhnlein C. 2010. Simulating the future – responses of ecosystems, key species, and expected European provenances to expected climatic trends and events. Nova Acta Leopoldina 112:89–98.

Jentsch A, Kreyling J, Beierkuhnlein C. 2007. A new generation of climate change experiments: events, not trends. Frontiers in Ecology and the Environment 5:365–374.

Jylhä K, Fronzek S, Tuomenvirta H, Carter TR, Ruosteenoja K. 2008. Changes in frost, snow and Baltic sea ice by the end of the twenty-first century based on climate model projections for Europe. Climatic Change 86:441–462.

Kalberer SR, Wiesniewski M, Arora R. 2005. Deacceleration and reacceleration of cold-hardy plants: current understanding and emerging concepts. Plant Science 171:3–16.

Kreyling J. 2010. Winter climate change: a critical factor for temperate vegetation performance. Ecology 91:1939–1948.

Kreyling J, Beierkuhnlein C, Jentsch A. 2010. Effects of soil freeze-thaw cycles differ between experimental plant communities. Basic and Applied Ecology 11:65–75.

Kreyling J, Beierkuhnlein C, Pritska K, Schiote M, Jentsch A. 2008. Recurrent soil freeze-thaw cycles enhance grassland productivity. New Phytologist 177:938–945.

Kreyling J, Thiél D, Simmacher C, Willner E, Jentsch A, Beierkuhnlein C. 2011. Geographic origin and past climatic experience influence the response to late spring frost in four common grass species of Central Europe. Ecography 34:1–8.

Maljanen M, Kohonen AR, Virkajärvi P, Martikainen PJ. 2007. Fluxes and production of N2O, CO2 and CH4 in boreal agricultural soil during winter as affected by snow cover. Tellus, Series B: Chemical and Physical Meteorology 59:853–859.

Malyshev AV, Arfin Khan MAS, Beierkuhnlein C, Steinbauer MJ, Henry HA, Jentsch A, Dengler J, Willner E, Kreyling J. 2016. Plant responses to climatic extremes: within-species variation equals among-species variation. Global Change Biology 22:449–464.

Malyshev AV, Henry HAL, Kreyling J. 2014. Relative effects of temperature vs. photoperiod on growth and cold acclimation of northern and southern ecotypes of the grass Arrhenatherum elatius. Environmental and Experimental Botany 106:189–196.

Nardini A, Salleso L, Lo Gullo MA, Pitt F. 2000. Different responses to drought and freeze stress of Quercus ilex L. growing along a latitudinal gradient. Plant Ecology 148:139–147.

Nilsson JE. 2001. Seasonal changes in phonological traits and cold hardiness of F1-populations from plus-trees of Pinus sylvestris and Pinus contorta of various geographical origins. Scandinavian Journal for Forest Research 16:7–20.

Oztas T, Fayetorbay F. 2003. Effect of freezing and thawing processes on soil aggregate stability. Catena 52:1–8.

Pauli JN, Zuckerberg B, Whitman JP, Porter W. 2013. The sub-nivium: a deteriorating seasonal refugium. Frontiers in Ecology and the Environment 11:260–267.

Poirier M, Durand J-L, Volaire F. 2012. Persistence and production of perennial grasses under water deficits and extreme temperatures: importance of intraspecific vs. interspecific variability. Global Change Biology 18:3632–3646.

R Core Team. 2017. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. https://www.R-project.org/ (6 December 2017).

Rapacz M, Plazek A, Niemczyk E. 2000. Frost de-acclimation of barley (Hordeum vulgare L.) and meadow fescue (Festuca pratensis Huds.). Relationship between soluble carbohydrate content and resistance to frost and the fungal pathogen Bipolaris sorokiniana (Sacc.) Shoem. Annals of Botany 86:539–545.

Rehfelt GE, Tchebakova NM, Parfenova VL, Wykoff WR, Kuzmina NA, Milyutin LI. 2002. Intraspecific responses to climate in Pinus sylvestris. Global Change Biology 8:912–929.

Schadt CW, Martin AP, Lipson DA, Schmidt SK. 2003. Seasonal dynamics of previously unknown fungal lineages in tundra soils. Science 301:1359–1361.

Schuering S, Jentsch A, Walter J, Kreyling J. 2014. Winter warming pulses differently affect plant performance in temperate heathland and grassland communities. Ecological Research 29:561–570.

Six J, Bossuyt H, Degryze S, Denef K. 2004. A history of research on the link between (micro)aggregates, soil biota, and soil organic matter dynamics. Soil and Tillage Research 79:7–31.

Sjursen H, Michelsen A, Holmstrump M. 2005. Effects of freeze-thaw cycles on microarthropods and nutrient availability in a sub-Arctic soil. Applied Soil Ecology 28:79–93.

Storey JD, Taylor JE, Siegmond D. 2004. Strong control, conservative point estimation, and simultaneous conservative consistency of false discovery rates: a unified approach. Journal of the Royal Statistical Society, Series B 66:187–205.

Strimbeck GR, Schaberg PG, DeHayes DH, Shane JB, Hawley GJ. 1995. Midwinter dehardening of montane red spruce during a natural thaw. Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere 25:2040–2044.

Tierney GL, Fahey TJ, Groffman PM, Hardy JP, Fitzhugh RD, Driscoll CT. 2001. Soil freezing alters fine root dynamics in a northern hardwood forest. Biogeochemistry 56:175–190.
Tierney GL, Fahey TJ, Groffman PM, Hardy JP, Fitzhugh RD, Driscoll CT, Yavitt JB. 2003. Environmental control of fine root dynamics in a northern hardwood forest. Global Change Biology 9:670–679.

Vankoughnett MR, Henry HAL. 2013. Combined effects of soil freezing and N addition on losses and interception of N over winter and summer. Ecosystem 16:694–703.

Walter J, Jentsch A, Beierkuhnlein C, Kreyling J. 2013. Ecological stress memory and cross stress tolerance in plants in the face of climate extremes. Environmental and Experimental Botany 94:3–8.

Wipf S, Stoeckli V, Bebi P. 2009. Winter climate change in alpine tundra: plant responses to changes in snow depth and snowmelt timing. Climatic Change 94:105–121.

Zhang T, Barry RG, Knowles K, Ling F, Armstrong RL. 2003. Distribution of seasonally and perenially frozen ground in the Northern Hemisphere. In: Phillips M, Springman SM, Arenson LU, eds. Permafrost. Lisse: Swets & Zeitlinger, 1289–1294.