Drug–Drug Interactions with Antiretroviral Drugs in Pregnant Women Living with HIV: Are They Different from Non-Pregnant Individuals?

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Published online: 22 July 2020
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Abstract
Background and Objective Although the separate effects of drug–drug interactions and pregnancy on antiretroviral drug pharmacokinetics have been widely studied and described, their combined effect is largely unknown. Physiological changes during pregnancy may change the extent or clinical relevance of a drug–drug interaction in a pregnant woman. This review aims to provide a detailed overview of the mechanisms, magnitude, and clinical significance of antiretroviral drug–drug interactions in pregnant women.

Methods We performed a literature search and selected studies that compared the magnitude of drug–drug interactions with antiretroviral drugs in pregnant vs non-pregnant women.

Results Forty-eight papers examining drug–drug interactions during pregnancy were selected, of which the majority focused on pharmacokinetic boosting. Other selected studies examined the drug–drug interactions between efavirenz and lumefantrine, efavirenz and tuberculosis drugs, etravirine and tenofovir disoproxil fumarate, atazanavir and tenofovir disoproxil, and mefloquine and nevirapine in pregnant compared to non-pregnant women. The clinical significance of antiretroviral drug–drug interactions changed during pregnancy from a minimal effect to a contra-indication. In almost all cases, the clinical significance of a drug–drug interaction was more relevant in pregnant women, owing to the combined effects of pregnancy-induced physiological changes and drug–drug interactions leading to a lower absolute drug exposure.

Conclusions Multiple studies show that the clinical relevance of a drug–drug interaction can change during pregnancy. Unfortunately, many potential interactions have not been studied in pregnancy, which may place pregnant women living with human immunodeficiency virus and their newborns at risk.

Key Points

More attention is needed from both researchers and clinicians to demonstrate which drug–drug interactions (DDIs) are clinically relevant in special populations.

The extent of a DDI may differ if the exposure of the perpetrator changes during pregnancy. DDIs at the level of intestinal transporters do not seem to be different in pregnant women.

The clinical significance of most DDIs in this review changed during pregnancy when looking at the absolute drug exposure in pregnant women. The combined effects of the DDI and other pregnancy-related physiological changes may lead to a more pronounced reduction in drug exposure, thereby exposing to the risk of therapeutic failure.
1 Introduction

All patients living with human immunodeficiency virus (HIV) must be exposed to an adequate concentration of antiretroviral drugs (ARVs) to achieve and maintain virologic suppression. In pregnancy, the exposure, and thus the efficacy of ARVs, can be modified by several factors such as drug–drug interactions (DDIs) and pregnancy-induced physiological changes. These factors have to be taken into consideration when selecting an individual ARV regimen in addition to side effects, resistance testing, previous antiviral regimens, and practical aspects [1].

Several studies have demonstrated the clinically relevant effect of DDIs on the pharmacokinetics of ARVs. Protease inhibitors (PIs), non-nucleoside reverse transcriptase inhibitors, and the integrase strand inhibitor (INSTI) elvitegravir (EVG) are extensively metabolized by the cytochrome P450 enzyme (CYP450) system leading to a variety of relevant DDIs [2, 3]. For example, the exposure of multiple ARVs is decreased as a result of CYP induction by rifampicin [4]. However, ARVs can also influence other co-medications: efavirenz (EFV) induces CYP3A4 activity and can, thereby, decrease the efficacy of several co-medications [5]. We also make use of interactive effects, for example CYP3A4 inhibition by ritonavir or cobicistat to increase the exposure to PIs and EVG (pharmacokinetic boosting) [6]. Drug–drug interactions are not limited to CYP interactions, but extend to non-CYP metabolism, drug transporters, and the absorption, distribution, and elimination of drugs. Inhibition of efflux transporter proteins in the gastrointestinal tract by cobicistat can increase the absorption of substrates of this transporter [7]. Drug absorption can also be affected by chelation with another drug and by pH-influencing drugs [8, 9]. Plasma protein displacement can affect the total plasma concentration and distribution of some highly protein-bound drugs [10].

It is also clearly demonstrated that physiological changes in pregnancy influence the pharmacokinetics of ARVs and co-medications [11]. Drug absorption may be altered because of increased gastric pH and slower intestinal motility [12, 13]. Physiological processes affect the ARV distribution and changes in plasma protein concentrations can affect the unbound ARV fraction of highly protein-bound drugs [14]. Finally, changes in hepatic blood flow, metabolic enzyme activity, glomerular filtration rate, and renal blood flow may alter the metabolism and elimination of ARVs and co-medication [14, 15].

In general, dosing recommendations and contra-indications of ARVs are based on DDI studies in healthy volunteers or individuals living with HIV, while pregnant women are excluded. However, pregnancy-related physiological changes may influence the magnitude or clinical relevance of a DDI. A non-clinically relevant DDI leading to a marginally lower exposure of the ARV in a non-pregnant woman may become relevant when adding the effect of pregnancy causing a potential risk for virologic failure and mother-to-child transmission. As an example, labels were recently updated to discourage the use of cobicistat-boosted regimens during pregnancy owing to a reduced exposure of cobicistat leading to insufficient concentrations of the boosted ARV during pregnancy, whereas this effect is not observed in non-pregnant women [16].

Antiretroviral drug exposure is difficult to predict in pregnant women, especially when the women are receiving interacting co-medications. The ARV exposure is determined by the different pharmacokinetic processes in the body. A pharmacokinetic process, for example hepatic metabolism by CYP3A4 enzymes, can be influenced by a DDI and physiological pregnancy changes concurrently. Pregnancy hormones induce CYP3A4 activity, while an interacting drug can inhibit CYP3A4 activity [14]. This is further complicated if the DDI degree is concentration dependent because the concentration of the interacting drug can be impacted by the DDI itself and by the physiological changes related to pregnancy. These interrelated effects make it difficult to predict the magnitude of DDIs in pregnant women, and studies examining DDIs in pregnant women are, therefore, necessary to ensure adequate ARV exposure and efficacy. This article provides an overview of the current knowledge on DDIs with ARVs in pregnant women living with HIV. The inter-related effects of DDIs and physiological changes in pregnancy are further elucidated in the results section with the examples from the literature. We discuss the mechanisms leading to altered ARV exposure and the effect in terms of clinical relevance.

2 Search Methodology

PubMed and Embase searches were performed to find the relevant articles about DDIs in pregnant women living with HIV. The following terms were used: (Pregnancy OR pregnant) AND drug interaction* AND anti human immunodeficiency virus agents. As boosting is not always referred to as being an interaction, a second search term was used: ‘Pharmacokinetic*’ AND ‘HIV’ AND ‘pregnant OR pregnancy’ AND ‘ritonavir OR cobicistat’. In addition, registration information from the European Medicines Agency and the US Food and Drug Administration was used and abstracts of relevant international conferences (Conference on Retroviruses and Opportunistic Infections, International Workshop on Clinical Pharmacology of Antiviral Therapy, European AIDS Conference and International AIDS Conference) were screened.

We selected studies comparing drug exposure in pregnant women with exposure in women postpartum or non-pregnant women. Drug–drug interactions in pregnant women...
are discussed for enzyme inhibition, enzyme induction, transporter inhibition, and for other unknown mechanisms. For every interaction mechanism, three scenarios are discussed: (1) the effect of DDIs in non-pregnant individuals (mostly studied in men); (2) the effect of pregnancy on drug exposure in the absence of an interacting drug; and (3) the combined effect of DDIs and pregnancy on drug exposure in pregnant women. In addition, changes in the DDI magnitude and the related clinical significance in pregnant women are discussed.

### 3 Studied Interactions in Pregnant Women Living with HIV

In total, 48 relevant papers and abstracts were selected. The majority of these studies examined the DDI between a booster and PIs or EVG in pregnant compared to postpartum women. Other selected studies examined the DDI between EFV and lumezantrine (LF) \([n=1]\), EFV and tuberculosis drugs \((n=1)\), etravirine (ETR) and tenofovir disoproxil fumarate (TDF) \([n=1]\), tenofovir alafenamide (TAF) and cobicistat \((n=2)\), atazanavir (ATV) and TDF \((n=3)\), and mefloquine and nevirapine \([n=1]\) in pregnant compared to non-pregnant women.

#### 3.1 Enzyme Inhibition

##### 3.1.1 Boosters and Protease Inhibitors/Elvitegravir

The DDI between boosters (i.e., cobicistat and ritonavir) and PIs/EVG has been extensively studied in pregnant women living with HIV. The effect of pregnancy-related physiological changes on the DDI was evaluated by comparing the PI/EVG exposure during pregnancy and postpartum in the same women. A decreased PI/EVG exposure was repeatedly confirmed in pregnant women during the first, second, and third trimester compared to postpartum. The average area under the curve (AUC) decrease had a range of 5–56% for PIs and 7–44% for EVG (Table 1) [17–53].

Figure 1 discusses the mechanism of this DDI in pregnant women considering three scenarios, which all compare to non-pregnant women using PIs or EVG without boosters. Figure 1a depicts the DDI in the non-pregnant situation where the PI/EVG exposure is greatly increased owing to CYP3A4 inhibition by the booster [54–56]. Figure 1b shows the probable effect of pregnancy on PI/EVG and booster exposures in the absence of the DDI. Hormonal changes in pregnancy induce CYP3A4 activity, which results, likely in combination with other physiological pregnancy changes, in decreased PI/EVG and booster exposures [14]. Pharmacokinetic assessments of unboosted indinavir, a PI primarily metabolized by CYP3A4, have been performed in pregnant women living with HIV, mimicking the situation in Fig. 1b. These studies showed a 62–86% decrease in indinavir AUC \(0–8\) during pregnancy compared with postpartum [57–59]. In one study, the decreased indinavir exposure corresponded to an increase in the urinary metabolic ratio of cortisol, a reliable marker for CYP3A4 activity, supporting that induced CYP3A4 activity contributed to decreased PI/EVG exposure during pregnancy [59]. Another study reported the metabolic ratio of cortisol to be 35% (1–81%) [median (95% confidence interval [CI])] higher during the third trimester of pregnancy compared to postpartum [60].

As described in Fig. 1c and Table 1, the combined effects (pregnant women taking a boosted PI/EVG regimen) lead to lower PI/EVG exposures than non-pregnant women taking a boosted PI/EVG regimen. The median (range) boosted indinavir AUC\(_{0–12\text{ h}}\) was 27.1 (18.6–44.7) μg*h/mL during postpartum compared with 16.1 (7.5–39.9) μg*h/mL in the third trimester, showing a 40% decrease for ritonavir-boosted indinavir during pregnancy (Table 1) [41]. The ritonavir AUC\(_{0–12\text{ h}}\) was also decreased by 60% during pregnancy, as a result of both pregnancy-related CYP induction and other physiological changes (i.e., decreased plasma albumin) [Fig. 1c, Table 1] [41, 46]. The comparison of unboosted and boosted indinavir pharmacokinetics shows that pregnancy has a smaller impact on ritonavir-boosted indinavir (~40%) than unboosted indinavir (62–86%) and that the absolute indinavir exposure is higher for ritonavir-boosted indinavir than for unboosted indinavir in pregnant women [41, 57–59]. This suggests that CYP3A4 inhibition by ritonavir, despite a reduction in ritonavir exposure during pregnancy, compensates, to some extent, the CYP3A4 induction in pregnancy (Fig. 1c).

Physiological changes in pregnancy seem to have a different effect on ritonavir- and cobicistat-boosted regimens, as shown in Table 1. The darunavir (DRV) trough concentration (\(C_{\text{trough}}\)) seems to be particularly lower during pregnancy in cobicistat- compared to ritonavir-boosted DRV 800 mg once-daily regimens; boosting with cobicistat resulted in a 71–92% decrease in \(C_{\text{trough}}\) vs 24–64% with ritonavir boosting [25, 26, 28–31, 33, 34]. Similarly, the ATV \(C_{\text{trough}}\) is lower in cobicistat- compared to ritonavir-boosted regimens, and EVG \(C_{\text{trough}}\) is also excessively decreased during pregnancy on average by 81–89% [17–24, 35–38]. Because of the pronounced reduction in \(C_{\text{trough}}\), which is considered to be mostly related to efficacy, cobicistat-boosted regimens are no longer recommended during pregnancy [16]. The difference in booster efficacy in pregnant women can be caused by multiple processes.

First, the decrease in cobicistat AUC and especially the \(C_{\text{trough}}\) seems larger compared to the decrease in ritonavir AUC and \(C_{\text{trough}}\) (Table 1). Cobicistat concentrations in pregnant women largely drop below the half maximal inhibitory concentration for CYP3A4 inhibition (i.e., 0.11 mg/L) to
Table 1 Overview of studies that reported antiretroviral drugs (ARVs) and boosters pharmacokinetic parameters in pregnant women compared to non-pregnant women. Postpartum women were assumed to be physiologically similar to non-pregnant women

| Reference | Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|-----------|------------|-----------------|------------|---------------------------------------------------------------|---------|
|           |            |                 |            | AUC | C\_trough | C\_max | AUC | C\_trough | C\_max |
| Atazanavir 300 mg QD+ritonavir 100 mg QD | | | | | | |
| [17] | n = 103, 88% African, sparse sampling, fasted/ fed unknown | Self-calculated ratio | 1 T vs PP (intra) | – | 0.65* | – | – | 1.38 | – |
| | | | 2 T vs PP (intra) | – | 0.69* | – | – | 0.86 | – |
| | | | 3 T vs PP (intra) | – | 0.72* | – | – | 1 | – |
| [18] | n = 10, 70% white, intensive sampling, fed without concomitant TDF, 1 patient with 400/100 mg | LSM (90%CI) | 3 T vs PP (intra) | 0.66 (0.53–0.83) | 0.64 (0.47–0.87) | 0.65 (0.52–0.82) | – | – | – |
| [19] | n = 37, mostly Hispanic, intensive sampling, fed without concomitant TDF | Self-calculated ratio | 2 T vs PP (intra) | 0.63* | 0.54* | 0.69* | 0.34* | 0.71 | 0.38* |
| [20] | n = 20, 85% black, intensive sampling, fasted/fed unknown | GMR (90% CI) | 2 T vs PP (intra) | 0.79 (0.61–1.03) | 1.00 (0.65–1.54) | 0.83 (0.65–1.06) | – | – | – |
| | | | 3 T vs PP (intra) | 0.79 (0.63–0.99) | 1.01 (0.73–1.39) | 0.73 (0.56–0.96) | – | – | – |
| [21] | n = 18, 67% black, intensive sampling, fed without concomitant TDF | Self-calculated ratio | 3 T vs PP (intra) | 0.72* | 0.58* | 0.88* | 0.44* | BLOQ | 0.44* |
| [22] | n = 25, 52% Caucasian, sparse sampling, fasted/ fed unknown | Self-calculated ratio | 1 T vs PP (intra) | – | 0.53 | – | – | 0.43 | – |
| | | | 2 T vs PP (intra) | – | 0.85* | – | – | 0.35* | – |
| | | | 3 T vs PP (intra) | – | 0.66* | – | – | 0.25* | – |
| [23] | n = 17, 53% African, intensive sampling, fasted/fed unknown | GMR (90% CI) | 3 T vs PP (intra) | 0.94 (0.80–1.11) | 0.96 (0.82–1.13) | 0.94 (0.74–1.11) | – | – | – |
| Atazanavir 300 mg QD+cobicistat 150 mg QD | | | | | | |
| [24] | n = 6, 67% black, intensive sampling, fasted/fed unknown | Self-calculated ratio | 2 T vs PP (intra) | 0.77 | 0.36 | – | – | – | – |
| | | | 3 T vs PP (intra) | 0.62 | 0.27 | – | – | – | – |
| Reference | Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|-----------|------------|-----------------|------------|------------------------------------------------------------------|---------|
|           |            |                 |            | AUC                  | $C_{\text{trough}}$ | $C_{\text{max}}$ | AUC                  | $C_{\text{trough}}$ | $C_{\text{max}}$ |
| 1.  GMR (90% CI) 3 T vs PP (intra) | 25 | n = 6, Europe (66% black), intensive sampling, fed | 0.78 (0.60–1.00) | 0.89 (0.58–1.36) | 0.76 (0.53–1.11) | 0.74 (0.65–0.84) | 0.72 (0.56–0.92) | 0.63 (0.53–0.76) |
| 2.  Self-calculated ratio 3 T vs PP (intra) | 26 | n = 34, USA (41% Caucasian), intensive sampling, fasted/fed unknown | 0.74* | 0.84# | 0.72* | – | – | – |
| 3.  LSM ratio (90% CI) or self-calculated ratio 3 T vs PP (intra) | 27 | n = 16, USA and Puerto Rico (63% black), intensive sampling, fasted/fed unknown | 0.76 (0.63–0.90) | 0.92 (0.66–1.30) | 0.72 (0.61–0.86) | 0.72 (0.44–1.17) | 0.57 | 0.66 (0.41–1.08) |
| 4.  Self-calculated ratio 2 T vs PP (intra) | 28 | n = 16, USA and Puerto Rico (41% white), intensive sampling, fasted/fed unknown | 0.66 (0.60–0.74) | 0.87 (0.67–0.85) | 0.70 (0.61–0.86) | 0.67 (0.43–1.04) | 0.58 | 0.63 (0.40–0.98) |
| 5.  GMR (90% CI) 3 T vs PP (intra) | 29 | n = 18, European (50% Caucasian), intensive sampling, fed | 0.67 (0.56–0.82) | 0.58 (0.44–0.78) | 0.78 (0.65–0.95) | 0.59 (0.42–0.83) | 0.72 (0.47–1.10) | 0.59 (0.42–0.82) |

**Table 1 (continued)**

| Reference | Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|-----------|------------|-----------------|------------|------------------------------------------------------------------|---------|
| 1.  GMR (90% CI) 3 T vs PP (intra) | 25 | n = 6, Europe (66% black), intensive sampling, fed | 0.78 (0.60–1.00) | 0.89 (0.58–1.36) | 0.76 (0.53–1.11) | 0.74 (0.65–0.84) | 0.72 (0.56–0.92) | 0.63 (0.53–0.76) |
| 2.  Self-calculated ratio 3 T vs PP (intra) | 26 | n = 34, USA (41% Caucasian), intensive sampling, fasted/fed unknown | 0.74* | 0.84# | 0.72* | – | – | – |
| 3.  LSM ratio (90% CI) or self-calculated ratio 3 T vs PP (intra) | 27 | n = 16, USA and Puerto Rico (63% black), intensive sampling, fasted/fed unknown | 0.76 (0.63–0.90) | 0.92 (0.66–1.30) | 0.72 (0.61–0.86) | 0.72 (0.44–1.17) | 0.57 | 0.66 (0.41–1.08) |
| 4.  Self-calculated ratio 2 T vs PP (intra) | 28 | n = 16, USA and Puerto Rico (41% white), intensive sampling, fasted/fed unknown | 0.66 (0.60–0.74) | 0.87 (0.67–0.85) | 0.70 (0.61–0.86) | 0.67 (0.43–1.04) | 0.58 | 0.63 (0.40–0.98) |
| 5.  GMR (90% CI) 3 T vs PP (intra) | 29 | n = 18, European (50% Caucasian), intensive sampling, fed | 0.67 (0.56–0.82) | 0.58 (0.44–0.78) | 0.78 (0.65–0.95) | 0.59 (0.42–0.83) | 0.72 (0.47–1.10) | 0.59 (0.42–0.82) |
Table 1 (continued)

| Reference | Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|-----------|------------|-----------------|------------|---------------------------------------------------------------|---------|
|           |            |                 |            | AUC (90% CI) | C_{trough} (90% CI) | C_{max} (90% CI) | AUC | C_{trough} | C_{max} |
| [30] n = 5, Spain (50% Caucasian), intensive sampling, fed | GMR (90% CI) | 3 T vs PP (intra) | Total: 0.69 (0.35–1.35) | Total: 0.53 (0.19–1.52) | Total: 0.73 (0.41–1.32) | – | – | – |
|           |            |                 |            | Unbound: 0.76 (0.44–1.32) | Unbound: 0.76 (0.26–2.25) | Unbound: 0.71 (0.44–1.16) | – | – | – |
| [31] n = 33, 67% black, sparse sampling, fed | Self-calculated ratio | 1 T vs PP (intra) | 1.08 | 0.51* | 0.46* | – | – | – |
|           |            |                 | 2 T vs PP (intra) | 0.62 (0.44–0.88)* | 0.66 (0.39–1.12)* | 0.81 (0.64–1.01)* | 0.88 (0.61–1.27)* | 0.85 (0.67–1.08)* | 0.92 (0.68–1.23)* |
|           |            |                 | 3 T vs PP (intra) | 0.64 (0.55–0.73)* | 0.58 (0.49–0.69)* | 0.71 (0.62–0.81)* | 0.65 (0.52–0.82)* | 0.67 (0.41–1.11)* | 0.67 (0.54–0.83)* |
| Darunavir 800 mg BID + ritonavir 100 mg BID | GMR (90% CI) | 2 T vs PP (intra) | 0.73 (0.37–0.79) | 0.29 (0.10–0.81) | 0.41 (0.30–0.57) | 0.62 (0.46–0.84) | 0.76 (0.37–0.85) | 0.39 (0.21–0.7) | 0.72 (0.49–1.1) |
|           |            |                 | 3 T vs PP (intra) | 0.56 (0.42–0.73) | 0.11 (0.081–0.15) | 0.72 (0.55–0.93) | 0.41 (0.30–0.57) | 0.35 (0.23–0.47) | 0.62 (0.46–0.84) |
| Darunavir 800 mg QD + cobicistat 150 mg QD | LSM (90% CI) | 2 T vs PP (intra) | 0.67 (0.34–1.33) | 0.29 (0.10–0.81) | 0.74 (0.44–1.26) | – | – | – |
|           |            |                 | 3 T vs PP (intra) | 0.52 (0.37–0.74) | 0.25 (0.13–0.49) | 0.64 (0.50–0.82) | – | – | – |
| Elvitegravir 150 mg QD + cobicistat 150 mg QD | GMR (90% CI) | 2 T vs PP (intra) | 0.76 (0.57–1.0) | 0.19 (0.10–0.35) | 0.92 (0.71–1.2) | 0.56 (0.37–0.85) | 0.39 (0.21–0.7) | 0.72 (0.49–1.1) |
|           |            |                 | 3 T vs PP (intra) | 0.56 (0.42–0.73) | 0.11 (0.081–0.15) | 0.72 (0.55–0.93) | 0.41 (0.30–0.57) | 0.35 (0.23–0.47) | 0.62 (0.46–0.84) |
| [32] n = 24, 50% black, intensive sampling, fasted/fed unknown | GMR (90% CI) | 2 T vs PP (intra) | 0.64 (0.55–0.73)* | 0.58 (0.49–0.69)* | 0.71 (0.62–0.81)* | 0.65 (0.52–0.82)* | 0.67 (0.41–1.11)* | 0.67 (0.54–0.83)* |
|           |            |                 | 3 T vs PP (intra) | 0.67 (0.55–0.73)* | 0.58 (0.49–0.69)* | 0.71 (0.62–0.81)* | 0.65 (0.52–0.82)* | 0.67 (0.41–1.11)* | 0.67 (0.54–0.83)* |
| [33] n = 7, 71% black, intensive sampling, fed | LSM (90% CI) | 2 T vs PP (intra) | 0.77 (0.57–1.0) | 0.19 (0.10–0.35) | 0.92 (0.71–1.2) | 0.56 (0.37–0.85) | 0.39 (0.21–0.7) | 0.72 (0.49–1.1) |
|           |            |                 | 3 T vs PP (intra) | 0.58 (0.42–0.73) | 0.11 (0.081–0.15) | 0.72 (0.55–0.93) | 0.41 (0.30–0.57) | 0.35 (0.23–0.47) | 0.62 (0.46–0.84) |
| [34] n = 29, USA (62% black), fasted/fed unknown | GMR (90% CI) | 2 T vs PP (intra) | 0.67 (0.34–1.33) | 0.29 (0.10–0.81) | 0.41 (0.30–0.57) | 0.62 (0.46–0.84) | 0.76 (0.37–0.85) | 0.39 (0.21–0.7) | 0.72 (0.49–1.1) |
|           |            |                 | 3 T vs PP (intra) | 0.52 (0.37–0.74) | 0.25 (0.13–0.49) | 0.64 (0.50–0.82) | – | – | – |
| Elvitegravir 150 mg QD + cobicistat 150 mg QD | GMR (90% CI) | 2 T vs PP (intra) | 0.76 (0.57–1.0) | 0.19 (0.10–0.35) | 0.92 (0.71–1.2) | 0.56 (0.37–0.85) | 0.39 (0.21–0.7) | 0.72 (0.49–1.1) |
|           |            |                 | 3 T vs PP (intra) | 0.56 (0.42–0.73) | 0.11 (0.081–0.15) | 0.72 (0.55–0.93) | 0.41 (0.30–0.57) | 0.35 (0.23–0.47) | 0.62 (0.46–0.84) |
| [35] n = 30, USA (67% black), intensive sampling, fed | GMR (90% CI) | 3 T vs PP (intra) | 0.93 | 0.30 | 1.28 | 0.94 | 1.45 | 0.76 | 0.67 |
|           |            |                 | Unbound: 1.28 | Unbound: BLOQ | Unbound: 1.45 | – | – | – |
| [36] n = 1, intensive sampling, fed | Self-calculated ratio | 3 T vs PP (intra) | 1.07 | 0.4 | 1.09 | 0.57 | BLOQ | – | – |
|           |            |                 | Unbound: BLOQ | Unbound: BLOQ | Unbound: BLOQ | – | – | – |
| [37] n = 1, black, intensive sampling, fasted/fed unknown | Self-calculated ratio | 3 T vs PP (intra) | 0.66 (0.53–0.83) | 0.23 (0.14–0.39) | 0.79 (0.65–0.97) | – | – | – |
| [38] n = 14, European (71% black), intensive sampling, fed | GMR (90% CI) | 3 T vs PP (intra) | 0.66 (0.53–0.83) | 0.23 (0.14–0.39) | 0.79 (0.65–0.97) | – | – | – |
Table 1 (continued)

| Reference Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|----------------------|----------------|------------|------------------------------------------------|----------|
|                      |                |            | AUC | $C_{\text{trough}}$ | $C_{\text{max}}$ | AUC | $C_{\text{trough}}$ | $C_{\text{max}}$ |
| Fosamprenavir 700 mg BID + ritonavir 100 mg BID |                   |            |     |               |               |     |               |               |
| [39] $n=10$, 50% black, 50% Hispanic, intensive sampling, fasted/fed unknown | GMR (90% CI) | 2 T vs PP (intra) | 0.65 (0.47–0.89) | 0.64 (0.45–0.93) | 0.63 (0.47–0.85) | 0.38 (0.23–0.64) | 0.40 (0.23–0.71) | 0.32 (0.17–0.60) |
| [39] $n=10$, 50% black, 50% Hispanic, intensive sampling, fasted/fed unknown |                       | 3 T vs PP (intra) | 0.75 (0.57–1.00) | 0.62 (0.48–0.92) | 0.81 (0.62–1.06) | 0.91 (0.58–1.71) | 0.86 (0.52–1.42) | 1.00 (0.58–1.71) |
| [40] $n=29$, 52% Hispanic, intensive sampling, fasted/fed unknown | GMR (90% CI) | 2 T vs PP (intra) | 0.68 (0.44–1.04) | 0.48 (0.14–1.65) | 0.83 (0.56–1.22) | 0.51 (0.28–0.91) | 0.71 (0.24–2.14) | 0.65 (0.37–1.12) |
| [40] $n=29$, 52% Hispanic, intensive sampling, fasted/fed unknown |                       | 3 T vs PP (intra) | 0.60 (0.49–0.72) | 0.56 (0.43–0.72) | 0.74 (0.58–0.93) | 0.72 (0.55–0.95) | 0.70 (0.53–0.91) | 0.83 (0.61–1.11) |
| Indinavir 400 mg BID + ritonavir 100 mg BID |                   |            |     |               |               |     |               |               |
| [41] $n=26$, Thailand, intensive sampling, fasted and fed | Self-calculated ratio | 2 T vs PP (intra) | 0.55 | 0.46 | 0.72 | 0.40 | 0.44 | 0.37 |
| [41] $n=26$, Thailand, intensive sampling, fasted and fed |                       | 3 T vs PP (intra) | 0.59 | 0.46 | 0.67 | 0.41 | 0.50 | 0.32 |
| Lopinavir 400 mg BID + ritonavir 100 mg BID |                   |            |     |               |               |     |               |               |
| [42] $n=27$, Brazil tablet, intensive sampling, fasted/fed unknown | Self-calculated ratio | 2 T vs PP (intra) | 0.72 | 0.74 | 0.75 | 0.57 | 0.47 | 0.62 |
| [42] $n=27$, Brazil tablet, intensive sampling, fasted/fed unknown |                       | 3 T vs PP (intra) | 0.71 | 0.71 | 0.76 | 0.60 | 0.56 | 0.59 |
| [43] $n=33$, 61% Hispanic, tablet, intensive sampling, fed | Self-calculated ratio | 2 T vs PP (intra) | 0.54* | 0.51* | 0.58* | 0.61 | 0.74 | 0.53 |
| [44] $n=42$, 70% black, tablet or soft-gel, sparse sampling, fasted/fed unknown | Self-calculated ratio | 1 T vs PP (intra) | Total: 0.88 | Unbound: 1.09 | – | – | – | – |
| [44] $n=42$, 70% black, tablet or soft-gel, sparse sampling, fasted/fed unknown |                       | 2 T vs PP (intra) | Total: 0.73 | Unbound: 0.85 | – | – | – | – |
| [44] $n=42$, 70% black, tablet or soft-gel, sparse sampling, fasted/fed unknown |                       | 3 T vs PP (intra) | Total: 0.74 | Unbound: 0.74 | – | – | – | – |
| [45] $n=41$, 83% Caucasian tablet, sparse sampling, fed | GMR (95% CI) | 3 T vs non-pregnant (inter) | - | 0.87 (0.62–1.32) | – | – | – | – |
| [46] $n=12$, 75% black, tablet, intensive sampling, fed | Self-calculated ratio | 2 T vs PP (intra) | Total: 0.63 | Unbound: 0.62 | – | – | – | – |
| [46] $n=12$, 75% black, tablet, intensive sampling, fed |                       | 3 T vs PP (intra) | Total: 0.71 | Unbound: 0.69 | – | – | – | – |
| Reference | Population | Outcome measure | Comparison | Ratio pregnant/non-pregnant women of ARV pharmacokinetic parameter | Booster |
|-----------|------------|----------------|------------|---------------------------------------------------------------|---------|
|           |            |                |            | AUC | C_{trough} | C_{max} | AUC | C_{trough} | C_{max} |
| [47]      | n = 10, 50% black, soft-gel, 4 samples, fed | GMR (95% CI) 3 T vs PP (intra) | 0.60 (0.25–1.43) | 0.36 (0.14–0.92) |
| [48]      | n = 17, 41% black, 41% Hispanic, soft-gel, intensive sampling, fed | GMR (90% CI) 3 T vs PP (intra) | 0.72 (0.54–0.96) | 0.44 (0.32–0.60) | 0.77 (0.61–0.97) | 0.84 (0.65–1.08) | 0.82 (0.56–1.18) | 0.86 (0.64–1.16) |
| Lopinavir soft-gel 533 mg BID + ritonavir 133 mg BID |            | Self-calculated ratio 3 T vs PP (intra) | 0.58* | 0.53* | 0.58* |
| [49]      | n = 26, 42% black, intensive sampling, fed | Lopinavir tablet 600 mg BID + ritonavir 150 mg BID |            | | 0.91 | 0.87 | 0.95 | 0.90 | 0.85 | 0.98 |
| [50]      | n = 14, 71% white, intensive sampling, fed | Saquinavir 1000 mg BID + ritonavir 100 mg BID | Self-calculated ratio 2 T vs PP (intra) | 0.86 | 1.08 | 1.19 | 0.47 | 0.34 | 0.63 |
| [51]      | n = 28, Germany, intensive sampling, fed | | GMR (90% CI) 3 T vs non-pregnant (inter) | 0.65 (0.37–1.14) | 0.75 (0.68–0.90) | 0.65 (0.37–1.16) | 0.53 (0.33–0.86) | 0.61 (0.39–0.97) | 0.50 (0.29–0.89) |
| [52]      | n = 37, 38% black, intensive sampling, fed | Tenofovir alafenamide 10 mg QD + cobicistat 150 mg QD | Self-calculated ratio 2 T vs PP (intra) | 0.94 | 1.05 | 0.92 | 0.74 | 0.90 | 0.79 |
| [53]      | n = 27, USA (52% Hispanic), intensive sampling, fasted/fed unknown | Tenofovir alafenamide 25 mg QD + cobicistat 150 mg QD QD/ritonavir 100 mg QD | GMR (90% CI) 2 T vs PP (intra) | 0.79 (0.50–1.27) | – | – | – | – | – |
|           |            | | 3 T vs PP (intra) | 0.86 (0.66–1.12) | – | – | – | – | – |
|           |            | Tenofovir alafenamide 25 mg QD + cobicistat 150 mg QD | 3 T vs PP (intra) | 0.94 (0.38–1.33) | – | – | – | – | – |

1 T first trimester, 2 T second trimester, 3 T third trimester, AUC area under the curve, BID twice-daily dosing, BLOQ below level of quantification, CI confidence interval, C_{max} maximum concentration, C_{trough} trough concentration, GMR geometric mean ratio, inter inter-individual, intra intra-individual, LSM least-squares mean, PP postpartum, QD once-daily dosing, self-calculated ratio ratio of published mean values calculated by the authors

*Reported significant difference
#Reported non-significant difference
A. Only interaction

B. Only pregnancy

C. Pregnancy + interaction

Fig. 1  Hypothesis of the drug–drug interaction of protease inhibitors (PIs)/elvitegravir (EVG) and boosters in pregnant women explained using three situations. The depicted situations are in comparison to PI/EVG exposure in non-pregnant women without booster co-treatment. a Co-treatment with boosters increases PI/EVG exposure in non-pregnant individuals. b Physiological changes by pregnancy are expected to decrease PI/EVG exposure, as well as booster exposure. c Co-treatment with a booster in pregnant women increases the PI/EVG exposure, although less than in non-pregnant women because of a decreased booster exposure resulting in less cytochrome P450 (CYP) 3A4 inhibition. indicates a positive effect, ↓ indicates a negative effect, — indicates causes, --→ indicates a decreased positive effect, −→indicates a decreased negative effect, *, ** and *** show inter-related effects

Although there is a clinically different interaction in pregnant women taking a cobicistat-boosted regimen, some experts also believe that the result of pregnancy-related physiological changes are clinically relevant for some ritonavir-boosted regimens [70, 71]. Alternative dosing regimens are recommended for ritonavir-boosted ATV, DRV, and lopinavir to ensure an adequate C\textsubscript{trough} during pregnancy [70, 71]. Because of the abundance of alternative regimens, no increased dose of EVG or additional cobicistat dosing has been evaluated in pregnant women treated with a cobicistat-boosted EVG-based regimen. However, it would be interesting to evaluate whether dosing modification of EVG or cobicistat leads to adequate EVG exposure during pregnancy.

3.2 Enzyme Induction

3.2.1 Efavirenz and Lumefantrine

Adegbola et al. examined the DDI between the antiretroviral EFV and the antimalarial LF in pregnant women compared to women postpartum [72]. Lumefantrine is primarily metabolized by CYP3A4 and EFV induces CYP3A4 activity, resulting in a decreased LF exposure in non-pregnant patients as showed in Fig. 2a [72, 73]. Pregnant women treated with LF have ~30% lower LF concentrations at day 7, a marker for therapeutic efficacy, compared with non-pregnant women [74–77]. A higher metabolic or elimination rate was observed in some of the pharmacokinetic studies, indicating induced CYP3A4 activity during pregnancy [14, 75–77]. Altered distribution of the lipophilic LF is also hypothesized to decrease LF exposure during pregnancy and is represented by other physiological changes in Fig. 2b [74]. The study of Adegbola et al. also showed that the EFV mid-dose concentration (C12h) significantly decreased in pregnant women treated with EFV compared with non-pregnant women (median 1820 ng/mL vs 2760 ng/mL, respectively).
The largest study of EFV pharmacokinetics during pregnancy showed a more modest effect. The 24-h concentration was significantly and marginally lowered during pregnancy (geometric mean ratio [90% CI] 0.92 [0.77–1.09], p = 0.01), while no-effect on the AUC$_{0-24h}$ was seen (geometric mean ratio [90% CI] 1.09 [0.90–1.32]) [78]. This modest reduction in EFV pharmacokinetics is supported by other intensive pharmacokinetic studies and occurs possibly because of decreased plasma proteins or CYP2B6 induction as depicted in Fig. 2b [79, 80].

A smaller reduction in LF exposure is surprisingly observed in pregnant women compared with postpartum co-treated with EFV in the study of Adegbola et al. This suggests that the magnitude of the DDI is mitigated during pregnancy. The authors hypothesized that the reduced EFV exposure during pregnancy may lead to less CYP3A4 induction, as shown in Fig. 2c. This suggests that the decreased CYP3A4 induction by EFV is not compensated by the hormone-related CYP3A4 induction in pregnancy (Fig. 2c). Although CYP3A4 induction by EFV has been showed to be concentration dependent, it is debated if the modest EFV during pregnancy leads to less induction of CYP3A4 [81]. The lack of an appropriate control group in the study by Adegbola et al. may have led to an inadequate conclusion. Efavirenz is primarily metabolized by CYP2B6 and the CYP2B6 metabolizer status of the pregnant and non-pregnant control group was not determined. Possibly, the non-pregnant control group consisted of more CYP2B6 slow metabolizers leading to higher EFV concentrations, which is correlated with lower LF concentrations [82].

Importantly, a considerable proportion of pregnant women were shown to have a LF concentration 7 days post-drug intake below the 280-ng/mL cut-off previously considered for LF therapeutic efficacy [82]. Thus, the DDI between EFV and LF remains clinically relevant in pregnant women.

### 3.2.2 Efavirenz plus an Anti-Tuberculosis Drug: Additional Influence of Genotype

The CYP-metabolizing genotype may also influence the clinical significance of a DDI in pregnant women. This is illustrated by the example of EFV, isoniazid, and rifampicin. Efavirenz is metabolized predominantly by CYP2B6 and to a lesser extent by CYP2A6 and CYP3A4. Efavirenz plasma concentrations are highly variable, and most variability can be explained by the CYP2B6 genotype [83].

Co-treatment with rifampicin, a CYP inducer, was shown to decrease EFV concentrations modestly and to increase inter-patient variability as shown in Fig. 3a [83, 84]. The effect of rifampicin was shown to vary depending on the CYP2B6 genotype; in patients with an extensive CYP2B6 genotype, rifampicin caused a larger DDI magnitude [85]. Current treatment guidelines suggest that a dose adjustment of EFV is not necessary in the general population [83]. In pregnant women, EFV exposure is modestly decreased likely as a result of CYP2B6 induction and decreased plasma proteins and does not warrant a dose adjustment (Fig. 3b) [79, 80]. However, Dooley et al. demonstrated that pregnant women with an extensive CYP2B6 genotype are at a particular high risk for low EFV concentrations [83] and, therefore, this special patient population might require an increase in EFV dose. The cumulative effects of rifampicin interaction, physiological pregnancy alterations, and an extensive CYP2B6 genotype may result in a clinically relevant interaction (Fig. 3d). It has been reported that 36% of the pregnant women treated with rifampicin, isoniazid, and EFV had a minimum concentration < 1 μg/mL, compared with 23% of the pregnant women not treated with tuberculosis drugs [83]. Thus, the modest DDI between EFV and rifampicin may become relevant in pregnant women with an extensive CYP2B6 genotype, justifying an EFV dose increase. Additionally, the magnitude of the EFV and rifampicin DDI may be increased as rifampicin exposure is modestly increased.

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Fig. 2 Hypothesis of the drug–drug interaction of efavirenz (EFV) and lumefantrine (LF) in pregnant women explained using three situations. The depicted situations are in comparison to EFV exposure in non-pregnant women without the drug–drug interaction. a Co-treatment with EFV decreases LF exposure. b Physiological changes by pregnancy decrease LF exposure, as well as EFV exposure slightly. c Co-treatment with EFV in pregnant women decreases the LF exposure less than in non-pregnant women because of a decreased EFV exposure.
during pregnancy, possibly as a result of decreased clearance due to cholestatic changes [86].

Isoniazid, a CYP2A6 inhibitor, may further increase EFV concentrations in pregnant women with a CYP2B6 slow-metabolizer genotype, as in this situation CYP2A6 metabolism becomes more relevant [83]. A median minimum concentration of 6.54 μg/mL has indeed been reported in co-infected pregnant women with a slow-metabolizer genotype receiving tuberculosis treatment, making EFV-related side effects of concern [83].

### 3.3 Transporter Inhibition

#### 3.3.1 Etravirine and Tenofovir Disoproxil Fumarate: An Additional Compartment

Another pitfall is that additional interactions may exist during pregnancy because an additional compartment is present in pregnant women: the fetus. Transplacental transport to the fetus is susceptible to DDIs. Research showed for example that ETR inhibits placental ABC transporter BCRP (ABCG2) in vitro, and ritonavir inhibits several placental ABC transporters at physiological concentrations [87, 88]. Etravirine and ritonavir can, therefore, increase the fetal exposure of co-administered PIs or lamivudine, which are known substrates of the placental ABC transporters [88].

#### 3.3.2 Cobicistat and Tenofovir Alafenamide

Although cobicistat is contraindicated in pregnancy since 2018, we discuss this interaction as this provides insight into transporter interactions during pregnancy. Interaction data in pregnancy are available from prior to 2018. When combined with cobicistat, TAF exposure is strongly increased by the inhibition of efflux transporter proteins in the gastrointestinal tract (i.e., P-glycoprotein and breast cancer resistance protein) as shown in Fig. 4a [7]. Therefore, a dose reduction from 25 to 10 mg of TAF is recommended in patients co-treated with cobicistat in fixed-dose combinations [89]. In contrast to this interaction, pregnancy-related physiological changes decrease TAF exposure (Fig. 4b); TAF exposure decreased by approximately 35% in the third trimester in women treated with 25 mg of unboosted TAF [53, 90]. This decrease probably does not result from an alteration in transporter activity as indicated by an animal study showing similar P-glycoprotein activity in pregnant and non-pregnant mice [14].

In the combined situation of pregnant women co-treated with cobicistat, there is a marginal and insignificant decrease in TAF exposure compared with postpartum, see Table 1 and Fig. 4c [53, 90, 91]. This suggests that the DDI compensates for the reduced exposure as a result of the physiological pregnancy changes (Fig. 4b). Theoretically, this is inconsequent because cobicistat exposure is greatly reduced during pregnancy, as mentioned earlier. Possibly the magnitude of the DDI is not influenced by physiological pregnancy changes and the decreased cobicistat exposure. The intestinal transporter inhibition by cobicistat might be caused by local (intestinal) cobicistat concentrations, and to a lesser extent by (decreased) systemic cobicistat concentrations [92]. This inconsistency can also be explained by the large variability and uncertainty in these data because TAF plasma concentrations are difficult to measure and often below the limit of quantification. It is unclear yet whether the intracellular concentration of the active tenofovir anabolite is sustained during pregnancy.

The clinical significance of the interaction between TAF and cobicistat is lower in pregnant compared with non-pregnant women. The exposure of TAF is similar in pregnant women co-treated with TAF 25 mg and cobicistat (median AUC 0–24 h 335 ng*h/mL [91]) and non-pregnant patients treated with TAF 25 mg (median AUC 0–24 h 308 ng*h/mL [93]), which is considered to be a safe and regular treatment option. Based on these limited data, TAF exposure is not clinically significantly altered in pregnant women co-treated with cobicistat.

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**Fig. 3** Hypothesis of the drug–drug interaction of efavirenz (EFV) and rifampicin (RIF) in pregnant women with an extensive cytochrome P450 (CYP) 2B6 genotype explained using four situations. The depicted situations are in comparison to EFV exposure in non-pregnant women without the drug–drug interaction. a Co-treatment with RIF decreases EFV exposure. b Physiological changes by pregnancy decrease EFV exposure slightly, and increase RIF exposure slightly [86]. c Co-treatment with RIF in pregnant women decreases the EFV exposure more than in non-pregnant women. d Co-treatment with RIF in pregnant women and an extensive CYP2B6 genotype decreases the EFV exposure even more, owing to a higher CYP metabolism activity, indicates a positive effect. Δ indicates a negative effect, *, ** and *** show inter-related effects.
Fig. 4 Hypothesis of the drug–drug interaction of cobicistat and tenofovir alafenamide (TAF) in pregnant women explained using three situations. The depicted situations are in comparison to TAF exposure in non-pregnant women without co-treatment with boosters. a Co-treatment with cobicistat increases TAF exposure. b Physiological changes by pregnancy decrease TAF exposure, as well as cobicistat exposure. c Co-treatment with cobicistat in pregnant women increases the TAF exposure, but this increase is less than in non-pregnant women with a cobicistat-boosted regimen (situation 4A). indicates a positive effect, ↓↓ indicates a negative effect. *, ** and *** show inter-related effects.

### 3.4 Unknown Mechanism

#### 3.4.1 Tenofovir Disoproxil Fumarate and Atazanavir

Several studies examined the DDI between TDF and ATV in pregnant women. Most studies observed a ~25% decrease in ATV exposure in non-pregnant patients co-treated with TDF [94–96], but a non-altered ATV exposure was also observed in one study [97]. The mechanisms of this DDI is unknown until today. As tenofovir (TFV) is renally cleared and ATV undergoes CYP-mediated metabolism, an interaction at the biotransformation level is unlikely. The DDI could possibly occur at the absorption level as a result of induction of intestinal P-glycoprotein by TFV or owing to a physicochemical interaction in the gut [94].

A decrease in ATV exposure has also been confirmed in pregnant women living with HIV, as shown in Table 2 and depicted in Fig. 5b [18–21]. The ATV exposure is likely decreased in pregnant women owing to a combination of decreased absorption, increased clearance, and reduced booster exposure [19]. Tenofovir exposure is also decreased during pregnancy because of an increased volume of distribution and increased renal excretion [98, 99]. Three studies confirmed that the ATV exposure is lower in pregnant women treated with a TDF-based regimen compared with both non-pregnant women co-treated with a TDF-based regimen and pregnant women co-treated with a non TDF-based regimen, see Table 2 and Fig. 5c [18, 19, 21].

The magnitude of the DDI does not seem to change during pregnancy based on our recent data and third-trimester data from Mirochnick et al. (Table 2) [18, 21]. Interestingly, the study of Kreitchmann et al. reports a change in the magnitude of DDIs during pregnancy, which seems to relate to non-adherence in the non-TDF arm in this study [19]. Thus, the similar DDI magnitude in pregnant women, despite the reduced TFV exposure, suggests that this DDI is not concentration dependent. This supports the hypothesis of a local interacting effect of TDF on the absorption of ATV. Furthermore, if the DDI is not influenced by physiological changes in pregnancy, there is probably no competing effect of pregnancy and TFV on ATV absorption (Fig. 5c).

Although the extent of the interaction is similar in non-pregnant and pregnant women, the cumulative effect of physiological pregnancy changes (Fig. 5c) and the DDI lead to a more clinically significant DDI during pregnancy. Lower ATV C_{trough} is observed in pregnant women with the DDI; a geometric mean (95% CI) of 0.44 (0.31–0.62) mg/mL is observed in pregnant women co-treated with ATV and TDF vs a geometric mean (95% CI) of 0.89 (0.59–1.32) mg/mL in non-pregnant women co-treated with ATV and TDF. Nevertheless, co-treatment with ATV and TDF is not contraindicated during pregnancy because the observed C_{trough} is not lower than the 0.15-mg/L cut-off value considered for therapeutic efficacy [100, 101].

#### 3.4.2 Nevirapine and Mefloquine

Haaland et al. examined ARV plasma concentrations in a group of pregnant women living with HIV co-treated with mefloquine, which showed a high rate of mother-to-child transmission [102]. A significant reduction in median nevirapine concentrations (approximately 25%) was observed in pregnant women receiving co-treatment with mefloquine, although the nevirapine concentrations remained above the previously reported target values to prevent HIV replication [102]. This reduction was not observed in pregnant women receiving nevirapine with placebo. The mechanism of this DDI remains unclear, and the DDI may be observed in pregnant women owing to pregnancy-related physiological changes that could amplify the magnitude of the DDI. This interaction has never been examined in non-pregnant women, and, therefore, the magnitude of the DDI in the non-pregnant situation is unknown. Despite adequate nevirapine trough concentrations, more mother-to-child transmissions.
Table 2  Overview of reported pharmacokinetic parameters of atazanavir (ATV) in pregnant women compared to non-pregnant women with and without tenofovir disoproxil fumarate (TDF) co-treatment

| Reference           | Population                                                                 | Parameter                  | Non-pregnant individuals/postpartum women | Pregnant women |
|---------------------|-----------------------------------------------------------------------------|----------------------------|------------------------------------------|----------------|
|                     |                                                                             | AUC (h*ng/L)               | Without TDF | With TDF | Self-calculated ratio | Without TDF | With TDF | Self-calculated ratio |
| Taburet et al. [94] | HIV-infected non-pregnant patients, *n* = 11, France, intensive sampling after 4 weeks with ATV and 2 weeks co-treatment with TDF, Mean (SD) | AUC (h*ng/L)               | 53.8 (35.3) | 39.3 (23.0) | 0.73 | 32.08 (21.1–48.7) | 28.8 (22.2–37.4) | 0.90 |
|                     |                                                                             | C<sub>max</sub> (µg/mL)    | 5.2 (3.0)  | 3.4 (1.4)  | 0.66 | 2.93 (1.95–4.40) | 2.92 (2.21–3.84) | 1.00 |
|                     |                                                                             | C<sub>min</sub> (µg/mL)    | 0.86 (0.8) | 0.58 (0.4) | 0.67 | 0.58 (0.32–1.05) | 0.44 (0.31–0.62) | 0.76 |
| Colbers et al. [18] | HIV-infected pregnant patients, *n* = 31 (third trimester), *n* = 25 (postpartum), 52% black, intensive sampling, fed, GM (95% CI) | AUC (h*ng/L)               | 49.2 (34.7–69.8) | 46.1 (36.2–58.6) | 0.94 | 30.6 (19.9–39.0) | 26.2 (19.9–38.4) | 0.86 |
|                     |                                                                             | C<sub>max</sub> (µg/mL)    | 4.58 (3.31–6.34) | 4.17 (3.42–5.08) | 0.91 | 3.11 (1.95–4.06) | 2.73 (1.65–4.02) | 0.88 |
|                     |                                                                             | C<sub>trough</sub> (µg/mL) | 0.90 (0.47–1.71) | 0.89 (0.59–1.32) | 0.99 | 0.71 (0.47–1.00) | 0.44 (0.28–0.59) | 0.62 |
| Kreitchmann et al. [19] | HIV-infected pregnant patients, *n* = 31 (second trimester), *n* = 68 (postpartum), 44% Hispanic, intensive sampling, fed, Median (IQ) | AUC (h*ng/L)               | 48.8 (34.9–67.5) | 58.7 (41.4–80.5) | 1.20 | 30.6 (19.9–39.0) | 26.2 (19.9–38.4) | 0.86 |
|                     |                                                                             | C<sub>max</sub> (µg/mL)    | 4.52 (2.31–5.61) | 5.43 (3.81–6.80) | 1.20 | 3.11 (1.95–4.06) | 2.73 (1.65–4.02) | 0.88 |
|                     |                                                                             | C<sub>trough</sub> (µg/mL) | 0.90 (0.39–1.33) | 1.26 (0.69–1.95) | 1.40 | 0.71 (0.47–1.00) | 0.44 (0.28–0.59) | 0.62 |
| Mirochnick et al. [21] | HIV-infected pregnant patients, *n* = 38 (third trimester), *n* = 32 (postpartum), fed, Median (IQ) | AUC (h*ng/L)               | 57.9 (47.1–64.8) | 39.6 (21–54.9) | 0.68 | 41.9 (27.4–60.8) | 28.8 (15.4–34.5) | 0.69 |
|                     |                                                                             | C<sub>max</sub> (µg/mL)    | 4.1 (3–5.8)  | 4.1 (1.7–4.5) | 1.00 | 3.6 (2.8–5.1)  | 2.5 (1.6–3)  | 0.69 |
|                     |                                                                             | C<sub>trough</sub> (µg/mL) | 1.2 (1.1–2)  | 0.8 (0.6–1.2)  | 0.67 | 0.7 (0.5–1.1)  | 0.5 (0.4–0.7) | 0.71 |

* AUC area under the curve, CI confidence interval, C<sub>max</sub> maximum concentration, C<sub>min</sub> minimum concentration, C<sub>trough</sub> trough concentration, GM geometric mean, HIV human immunodeficiency virus, IA interaction, IQ interquartile range, SD standard deviation, self-calculated ratio ratio of published mean values calculated by the authors.
were observed, making this DDI likely of clinical relevance for HIV and malaria co-infected pregnant women.

### 4 Common Unstudied Interactions in Pregnant Women Living with HIV

Other commonly used drugs in pregnancy that can interact with ARV include: proton pump inhibitors, iron and mineral supplements, antacids, domperidone, and macrolide antibiotics [103, 104]. Drug–drug interactions with these co-medications have not been studied in pregnant women, while having the potential for clinically relevant interactions. They also illustrate possibly altered interaction mechanisms during pregnancy at the level of distribution, absorption, and non-CYP metabolism.

For instance, caution is needed when interpreting serum concentrations of highly protein-bound ARVs (such as PIs and INSTIs) in pregnancy, especially when co-administered with other highly protein-bound drugs. A DDI at the level of distribution can theoretically change in pregnant women because the amount of plasma proteins decreases during pregnancy [14]. Competition for plasma proteins with a highly protein-bound concomitant drug and a decrease in plasma proteins can co-exist, resulting in even lower total ARV concentrations.

Another common DDI mechanism involves the chelation of INSTIs with divalent cations such as iron, requiring staggered administration [8, 105–110]. The magnitude of the physiochemical interaction in the gut is not affected by physiological changes in pregnancy. However, this DDI is likely to be more clinically relevant in pregnant women because of the additive effects of chelation and pregnancy-related physiological changes potentially leading to a lower absolute INSTI exposure. Additionally, the time between INSTI and iron administration may need to be increased during pregnancy because gastrointestinal motility is decreased in pregnant women [14]. The limited decrease of raltegravir and dolutegravir (DTG) exposure in pregnant women is not considered to be of clinical significance [111–113], but may become of clinical significance in pregnant women co-treated with iron.

Furthermore, the common DDI between gastric acid-neutralizing drugs and ATV or rilpivirine (RPV) is yet unstudied in pregnant women living with HIV. The absorption of RPV/ATV decreases with increasing gastric pH, thus co-administration with proton pump inhibitors is contraindicated in non-pregnant women and it is recommended to stagger administration with histamine H2 antagonists (cimetidine, famotidine, ranitidine) [96, 114]. However, a different recommendation might be needed in pregnant women as pregnancy-related physiological changes include a possible increase in gastric pH that will also result in a decrease in RPV/ATV exposure [13, 115–117]. Thus, it is possible that the absorption of ATV/RPV is worse in pregnant women treated with an H2 antagonist because of the additive effect of the drug- and pregnancy-related physiological changes on the gastric pH. Research is needed to show if a contra-indication for ATV/RPV and H2 antagonists is required in pregnant women.

Hepatic DDIs concerning non-CYP enzymes can be potentially altered during pregnancy as well. A common example is the DDI between rifampicin and DTG, as the prevalence of HIV-tuberculosis co-infection is high in third-world countries and DTG-containing regimens are preferred according to the World Health Organization guidelines [118]. Glucuronidation to the main metabolite of DTG is catalyzed by UGT1A1, which can be induced by rifampicin. A dose increase to 50 mg of DTG twice daily is needed in non-pregnant individuals when co-administered with rifampicin [118, 119]. In pregnant women, DTG exposure is modestly decreased and UGT1A1 induction and
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5 Future Perspectives

More research on DDIs in pregnant women is needed, but the study design remains challenging. Pregnancy pharmacology studies lend themselves to investigate DDIs, as well as the systematic collection of therapeutic drug monitoring samples. In both cases, accurate registration of co-medication, dose, and time of intake make it possible to determine the absolute exposure of ARVs in pregnant women with DDIs, but a sufficient sample size is crucial for the interpretation. Observational pharmacology studies in pregnancy generally focus on the exposure to the ARV; however, in the presence of potential DDIs with non-ARVs, the plasma concentration of these agents should be systematically determined. Physiologically based pharmacokinetic (PBPK) models have been applied to predict drug exposure in special populations and to simulate DDIs. The validity of pregnancy PBPK models has improved tremendously during the recent years owing to the increased interest in these models [124]. Although more in vitro and clinical studies are needed to fill knowledge gaps, for example on UGT and drug transporter activity, the models are already widely used to understand and predict pharmacokinetic changes during pregnancy [124]. The knowledge from DDI and pregnancy PBPK models can be combined to improve our understanding of this complex situation. In particular, DDIs on the level of placental drug transport are difficult to investigate in clinical studies as only sampling from the umbilical cord during delivery is considered to be ethical. Knowledge on pregnancy PBPK models including the placenta and the fetus is gained rapidly and a combination of these models can offer a solution for predicting placental drug transporter DDIs [125].

Clinically relevant DDIs apply to pregnant women, but are also a factor of consideration for other special patient population such as the elderly, children, and patients with hepatic impairment. The management of DDIs has to be conducted on an individual basis. In the era of precision medicine, DDIs cannot be ignored and are an opportunity for a more effective and individualized drug treatment. More attention is needed from both researchers and clinicians to demonstrate which DDIs are clinically relevant in special populations.

6 Conclusions

Drug–drug interactions in pregnant women remain a complex situation because the extent and clinical significance of a DDI can change during pregnancy. Only a few studies examined this complex situation; therefore, the management of DDIs in pregnant women is mostly based on theoretical assumptions. This makes decisions regarding the optimal dose and ARV regimen very difficult, and places pregnant women living with HIV and their newborns at risk.

The extent of a DDI can change during pregnancy, although it is very challenging, if not impossible, to distinguish the influence of individual processes. The extent of a DDI may differ if the exposure of the perpetrator changes during pregnancy as suggested for concentration-dependent DDIs such as the DDI between cobicistat and PIs/EVG (inhibition of metabolism). Drug–drug interactions at the level of intestinal transporters do not seem to be different in pregnant women (i.e., TAF co-administered with cobicistat).

More importantly, the clinical significance of most DDIs in this review changed during pregnancy when looking at the absolute drug exposure in pregnant women. The combined effects of the DDI and other pregnancy-related physiological changes may lead to a more pronounced reduction in drug exposure, thereby exposing to the risk of therapeutic failure. Only the DDI between cobicistat and PIs/EVG in pregnancy has led to a change in drug labels; currently, the use of cobicistat-boosted regimens in pregnancy is no longer recommended [16]. There are not enough data for the other reviewed DDIs to change the official recommendations, although some DDIs resulted in drug concentrations below the target values in a large proportion of the pregnant women.

Compliance with Ethical Standards

Funding No specific funding was received for the preparation of this article.

Conflict of interest DMB has received research grants to his institution from Merck, BMS, Janssen/Tibotec, ViiV Healthcare, and Gilead, has received educational grants from Merck, was a speaker for a symposia for Merck, and is a member of the advisory boards of ViiV Healthcare and Merck. VEB, AC, CM, and JM have no conflicts of interest that are directly relevant to the content of this article.

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