Research of Biological Dose Conversion Platform Based on a Modified Linear Quadratic Model

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Abstract
The study aimed to develop a novel dose conversion platform by improving linear-quadratic (LQ) model to more accurately describe radiation response for high fraction/acute doses. This article modified the LQ model via piecewise fitting the biological dose curve using different fractionated dose and optimizing the consistency between mathematical model and experimental data to gain a more reasonable transform. That mathematical development of the LQ model further amended certain deviations of various cell curves with high doses and implied the rationality of the present model at low dose range. The modified biologically effective dose model that solved the dilemma of inaccurate LQ model had been used in comparing between hypofractionated and conventional fractioned dose. It has been verified that the calculated values are similar in the treatment of same efficacy, no matter what $a/b$ is, and provided a more rational explanation for significant differences among various hypofractionations. The equivalent uniform dose based on the subsection function could represent arbitrary inhomogeneous dose distributions including high-dose fractions, providing a foundation for the implementation of detailed evaluation of different cell dose effects.

Keywords
dose conversion, linear-quadratic (LQ) model, biologically effective dose (BED), equivalent uniform dose (EUD), stereotactic body radiation therapy (SBRT)

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Introduction

The exploration of ideal dose delivery pattern has greatly evolved due to the improvement of stereotactic conformal radiotherapy technique. Stereotactic body radiation therapy (SBRT) or stereotactic radiosurgery (SRS) irradiates both small and/or large complex-shaped lesions while minimizing the dose to adjacent radiosensitive tissues. Large doses per fraction has been used in ongoing clinical trials designed to explore the use of that pattern for different tumor sites; however, the radiosensitivity of different time-dose fraction schemes should be estimated accurately due to the deviation of radiobiological assessment.

The biologically effective dose (BED) based on linear-quadratic (LQ) model has been widely used to calculate tissue effective of various dose fractions since it was formulated by Douglas and Fowler. The fractioned dose and the gradient of biological effective determined the relative effectiveness in the formula, which shows the ratio of the initial slope and the slope at high dose range. The assumption has been proved biologically by Barendsen GW.

Some studies indicated that caution should be exercised in applying and interpreting results when using the LQ model with high doses per fraction. Alternating mechanisms in addition to DNA strand breaks and/or chromosome aberrations may involve in response of tumors to SBRT or SRS, Autophagy, characterized with the prominent formation of autophagic vacuoles in the cytoplasm, is a novel response of cancer cells, remaining largely elusive. Vascular damage may play an important role in the response of human tumors to high-dose hypofractionated irradiation, damaging the intratumor micro-environment and leading to indirect tumor cell death. The biological radiation effect cannot be theorized precisely in total dose range, as little effect of quantization is known about such mechanisms as radiation clonogenic capacity in the target.

The LQ model has been widely used in modeling the effect of total dose and dose per fraction in conventionally fractionated radiotherapy for decades. The LQ model solely depends on the expected incidence of direct interactions of radiation with specific cellular targets. The model generated by much of the data obtained in vitro has well-documented predictive properties for fractionation/dose-rate effects in the laboratory. Recent experimental studies have verified the inadequacy of the LQ model in converting hypofractionated doses into single doses and as a result may overestimate the effect of high fractional doses of radiation. No biological interpretation of the LQ parameters was proposed to explain together the radiation response in a wide dose range.

The validity of the LQ model for calculating iso-effect doses in radiation therapy has been intensively debated. Because sublethal damage repair takes place, the LQ survival curve continuously bends downward with increasing radiation dose. Some study improved the goodness of fit by removing the low dose data and high dose points. Another showed that higher order terms may be present to respond the heterogeneity of irradiated cell population while there is a bias in estimating values of α and β.

Material and Methods

The Extension of the Linear–quadratic Model at High-Dose Range

The equation about cell biological radiation effects \( E \) and single irradiation dose \( d \) can be derived using Taylor’s expansion.

\[
E(d) = E(d_0) + E(d_0)(d - d_0) + \frac{E(d_0)}{2!}(d - d_0)^2 + \cdots + R_n(d).
\]

A quadratic function got by developing the Taylor equation, using a logarithmic of survival rates for \( E \).

\[
\ln(SF) = f(d_0) + \alpha_1(d - d_0) + \beta_1(d - d_0)^2.
\]

The traditional LQ function is obtained when \( d_0 \) is zero.

\[
\ln(SF) = \alpha(d - 0) + \beta(d - 0)^2.
\]

If \( d = d_0 \), the derivations of both formula are equal, then

\[
\alpha_1 = \alpha + 2\beta d_0.
\]

\[
f(d_0) = \alpha d_0 + \beta d_0^2.
\]

The formula of cell curve can be written as a piecewise function

\[
\ln(SF)_m = \begin{cases} 
\alpha d + \beta d^2 & (d \leq d_0) \\
\alpha d_0 + \beta d_0^2 + (\alpha + 2\beta d_0)(d - d_0) + \beta_1(d - d_0)^2 & (d > d_0)
\end{cases}
\]
Grouping constants and terms linear and quadratic in the variable, one gets:

\[-\ln(SF)_m = \begin{cases} 
\alpha d + \beta d^2 & (d \leq d_0) \\
(\beta_1 - \beta)d_0^2 + [\alpha + 2(\beta - \beta_1)d_0]d + \beta_1 d^2 & (d > d_0)
\end{cases} \]  

(7)

**Compare the Goodness of Fit for Cell Survival Curve With Different Models**

\[-\ln(SF) = \alpha d + \beta d^2 + \gamma d^3.\]  

(8)

Methods in vitro cultured 800 human fibroblasts, A375, A549, were inoculated in 10-cm dishes, respectively. The clonogenic assay was carried out by using A375 melanoma cells, A549 human non-small cell lung cancer (NSCLC) cells, and hFB human skin fibroblasts, respectively. Briefly, cells were harvested and 800 cells per dish were reseeded in a 100-mm dish. The cells were treated with radiation at different doses from 0 Gy to 11 Gy at 12 hours after reseeding. Following 10 to 14 days of incubation, cells were fixed and stained with crystal violet, and colonies containing at least 50 cells were scored. The linear accelerator irradiation parameters were 6 MV X ray, and 180 degree irradiation. The 2 cm equivalent solid water module was placed below.

The survival curves of the 3 cells were fitted by the Equations 3, 6, 8, respectively.

Coefficient of determination, R-square, was used for the measures of goodness of fit.

\[R - \text{square} = \frac{\sum_{i=1}^{n}(\hat{y}_i - \bar{Y}_i)^2}{\sum_{i=1}^{n}(y_i - \bar{Y}_i)^2}.\]  

(9)

Where \(n\) is the number of samples, \(y_i\) is the estimated value, \(\bar{Y}_i\) is observed data, and \(\bar{Y}_i\) is sample mean.

Root mean squared error (RMSE) was employed to evaluate the precision and robust of different model systems.

\[\text{RMSE} = \sqrt{MSE} = \sqrt{\frac{\text{SSE}}{n}} = \sqrt{\frac{1}{n} \sum_{i=1}^{n} w_i(y_i - \bar{y}_i)^2}.\]  

(10)

Where \(n\) is the number of samples, \(w_i\) is data weight, \(\bar{y}_i\) is the estimated value, and \(\bar{Y}_i\) is observed data.

We compared the imitative effect of models, including another LQ model, whose \(\alpha\) and \(\beta\) were confirmed experimentally at a low dose range.

**Chi-Square Test of Mathematical Models**

The deviations of survival curves from the optimal models’ predictions and the stability of models were determined by using \(\chi^2\) test. 

\(\chi^2/df\) performed a \(\chi^2\) goodness-of-fit test that the data were a random sample from a normal distribution with mean and variance estimated with per degree of freedom. The \(P\) value was the probability of observing the given result, or one more extreme, by chance if the null hypothesis was true.

**Biologically Effective Dose Based on the Modified LQ Model**

The BED formula introduced by Fowler

\[\text{BED} = \frac{-n\ln(SF)}{\alpha} = nd\left(1 + \frac{d}{\alpha/\beta}\right) = D_T \times RE.\]  

(11)

The improved BED formula at high dose range based on the modified LQ model (Equation 7)

\[\text{BED}_{m} = \frac{-n\ln(SF)_m}{\alpha} = nd\left(1 + \frac{d}{\alpha/\beta_1}\right) + nd_0(2d - d_0)\left(1 + \frac{1}{\alpha/\beta} - 1\right).\]  

(12)

**Biologically Effective Dose Comparison of Various Dose-Fractionations**

Chang et al compared the curative effect between hypofraction and conventional fractionation for the treatment of melanoma. The study showed the hypofractionation (6 Gy×5 Fx) and the conventional fractionation (2 Gy×30 Fx) are equally efficacious in 5-year in-field local regional control, 5-year freedom from distant metastases, 5-year cause-specific, and overall survival (OS).

The randomized clinical trial of SPACE showed there were no statistically significant differences between SBRT (22 Gy×3 Fx) and conventional fractioned radiotherapy (CFRT) (2 Gy×35 Fx). The BED values of 4 dose fractionations above were calculated by Equation 11 with the \(\alpha/\beta\) fitted in Equation 3 at low dose range, and Equation 12 with the \(\alpha/\beta\) and \(\alpha/\beta_1\) best fitted in Equation 6, respectively.

Haque et al investigated more dose fractionations of SBRT and CFRT, which demonstrated the survival benefit to hypofraction. The BED computed by Equation 12 with the \(\alpha/\beta\) and \(\alpha/\beta_1\) best fitted in Equation 6, showed the differences of various treatments.

Stephans et al detailed and analyzed tumor control for common SBRT dose fractionation regimens in stage I NSCLC, of which BED would be recalculated by Equation 12 to explore the relationship between dose fractionation and local control (LC).

**Equivalent Uniform Dose Based on the Modified LQ Model**

The OS fraction is the weighted average of the survival fractions taken over all near homogeneously irradiated subvolumes
of the target, where colognes are uniformly distributed across the volumes.

\[
SF_{(D)} = \frac{\sum_{j=1}^{n} v_i \cdot \rho_i \cdot SF_{(D_j)}}{\sum_{j=1}^{n} v_i \cdot \rho_i},
\]

(13)

Assuming a constant rate of proliferation, it can be shown that the overall surviving fraction for per fraction dose \(d\) given in \(n\) fractions is

\[
\ln(SF) = \begin{cases} 
-n(\alpha d + \beta d^2) + \frac{\ln2 \cdot t}{T_{pot}} & (d \leq d_0) \\
-n[\alpha d_0 + \beta d_0^2 + (\alpha + 2\beta d_0)(d - d_0) + \beta_1(d - d_0)^2] + \frac{\ln2 \cdot t}{T_{pot}} & (d > d_0),
\end{cases}
\]

(14)

Here \(t\) is the overall treatment time after the start of proliferation, and \(T_{pot}\) is the potential doubling time of colognes.

The average of the survival fractions can be rewritten as a complex form.

\[
\overline{SF} = \frac{\sum_{j=1}^{P} v_j \rho_j e^{-n(\alpha d_j + \beta d_j^2) + \frac{\ln2 \cdot t}{T_{pot}} + \sum_{k=1}^{Q} v_k \rho_k e^{-n[\alpha d_0 + \beta d_0^2 + (\alpha + 2\beta d_0)(d_k - d_0) + \beta_1(d_k - d_0)^2] + \frac{\ln2 \cdot t}{T_{pot}}}}}{\sum_{i=1}^{P+Q} v_i \rho_i},
\]

where \(d_j < d_0; d_k > d_0;\)

\[
\overline{SF}_{ref} = \frac{\sum_{i=1}^{J} v_i \rho_i e^{-n(\alpha d_0 + \beta d_0^2) + \ln2_{ref}}}{\sum_{i=1}^{J} v_i \rho_i},
\]

(15)

where \(n_{ref}d_{ref} = D_{ref};\)

**Results**

Table 1 showed the colony formation efficiency of hFB, A375, A549 at different dose. respectively. Tables 2, 3, and 4, showed the fitting results of above 3 cells survival curves with different functions.

| D (Gy) | 0   | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  |
|--------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| SF     |     |     |     |     |     |     |     |     |     |     |     |     |
| hFB    | 1   | 0.781 | 0.595 | 0.447 | 0.143 | 0.072 | 0.031 | 0.015 | 0.004 |     |     |     |
| A375   | 1   | 0.857 | 0.784 | 0.522 | 0.325 | 0.200 | 0.071 | 0.027 | 0.004 |     |     |     |
| A549   | 1   | 0.670 | 0.424 | 0.277 | 0.154 | 0.072 | 0.031 | 0.015 | 0.004 |     |     |     |

Abbreviation: SF, survival functions.

Figure 1 showed the clonogenic survival curve of A375 within a range of 0 to 8 Gy and the comparison of predictions of Equation 3 (LQ), Equation 3-L (LQ model fitted at low-dose range(0-4 Gy)), Equation 6 (\(d_0 = 5\) Gy), and Equation 8. Equation 6 fit well in low dose range similar to Equation 3-L, and as
good as Equation 8 in high dose range. There was a perceived overprediction of cell killing by the LQ model.

The optimal parameters of LQ model were chosen at low dose range and those of Equation 6 had been obtained when the values of $a$ and $b$ were positive and RMSE was the lowest. It was possible to test the hypothesis that the 2 functions described the data ($H_0$: not rejected) by assuming that the measured values were independent and normally distributed, and the errors were known as well as standard normal distributed. For that, the results for total dose range are shown in Table 5. The table showed the both functions with optimal parameters could describe the measured survival data of all cells. However, an improvement of the Equation 6 was achieved with considerable difference of $\chi^2/df$ in magnitude with LQ model and an increment of the $P$ values which were greater than 95\% in 3 cells, far higher than those of LQ.

Table 6 showed the BEDs of 6 Gy/C2 for the treatments of melanoma. The disparities of calculated results were lower with the conventional formula and increased slightly with the modified one.

Table 7 shows the BEDs of 22 Gy/C2 for the treatments of NSCLC. The figures of SBRT were well above CFRT by taking the conventional equation and became nearly identical by the new one.

Table 8 shows the modified BED values of various hypofractioned NSCLC treatments. The biological effects were vastly different among various hypofractionations corrected the deviation of LQ model.

Table 9 presents the new BED values of the hypofractionations applied in Stephans et al’s study and the reported LC ratio. There was a distinctive linear relationship between them.

**Discussion**

Given closer analysis of the survival behavior of CHOA8, U373MG, DU145, and CP3 cells in an extensive dose range for X-rays irradiation, Garcia et al found that the fit quality of LQ model was related to the selected dose region. The outcome was adapted to a small scale, which was different in various cell types.\(^1\)

$\chi^2$-statistics test in this study also presented a deterioration of goodness of LQ model fit at high-dose ranges, especially in the curves of tumor cells. The survival model showed more suitable for the normal tissue cells, which reflects a special in vitro proliferation existing in the tumor cells.

The parameters $a$ and $\beta$ from LQ model have been widely used in different ways of fraction dose radiotherapy, while the BED with $a$ and $\beta$ was applied as a comparison parameter among various kinds of treatment plans despite its limitations and imperfections.

A curve function describing the survival of cells well in the full fractionation range without changing LQ model at all is a clinical imperative, which can absorb practical experience in conventionally fractionated treatment and be fitted to hypofractionated radiotherapy outcomes.
The curve fitting result of Equation 8 indicated it was not sufficient to solely depend on increasing orders. The RMSE in cell A549 curve fitting of Equation 8 was higher than Equation 3 (0.1269 vs 0.1216), meanwhile, \( \alpha/\beta \) value was 84.4, far higher than the maximum, 10, what is normally considered. The \( \beta \) value in cell A375 curve fitting became negative, which could not be explained as the mean probability per unit square of the dose in linear quadratic therapy.

This study added second-order coefficient \( \beta_1 \) in the improved curve function, retaining \( \alpha \) and \( \beta \) parameters, without increasing the order, thus better results had been gained.

The added parameter could be explained in terms of radiobiology. The average probability of both particles interaction should be modified due to the increasing of fraction dose and electron scattering.

More biological explanation of the parameters in the modified LQ function can be further explored, the constant, \( \alpha d_0 + \beta d_0^2 \), means the total biological radiation effect when the fraction dose is \( d_0 \). The variable, \( (\alpha + 2\beta d_0)(d - d_0) \), means the effect of one particle interaction caused by the dose exceed \( d_0 \), the parameter, \( \alpha + 2\beta d_0 \), is the average probability per unit one particle interaction, which should increase with electron yield and absorbed dose. \( \beta_1 \) is the correction probability, \( \beta_1(d - d_0)^2 \) means the correction effect of both particles interaction to modified the deviation caused by the first 3 variables and can be either positive or negative.

In the fitting of A549 survival curve, if \( d_0 = 2 \), \( \beta_1 \) was close to zero (\( \beta_1 = -0.00275 \)). The Equation 6 could be rewritten as a simple style, \( \alpha d + 2\beta d_0 d - \beta d_0^2 \), if a fraction dose was greater than \( d_0 \), \( \chi^2 \) of independence analyses showed that there was no statistical difference between fitted and actual data (\( P = 0.7449 \)).

The difference of BED values between SBRT and CFRT increased slightly in the modified model due to the small deviation of LQ model with lower \( \alpha/\beta \) values.

The conventional BED value of SBRT showed doubles that of CFRT for similar NSCLC treatments, which seemed hard to explain.\(^{16}\) Some researchers proposed it as “overkill” of SBRT,\(^{20}\) that would mean no needing to increase the dosage to get more curative effect. There is a huge problem in comparing differentia at various dose fractionations, resulting from the theoretical defect of LQ at high dose range. Biologically effective dose model, correcting the deviation, will obtain the further expansion in clinical practice’s value. It is reliable as Table 7 shows, in similar treatments of tumors with high \( \alpha/\beta \) values, BEDs are close and the differences can be seen as the benefits of short protocols overcoming cells’ accelerate repopulation.

Haque et al concluded SBRT had more benefit in OS\(^{18}\) which seemed to contradict with Chang et al’s study. Actually, not all of what Haque investigated could be considered as an ablative radiotherapy, and when BEDs had been recalculated by the modified model, and huge disparities were found among them. The conclusion would be supported when there were a large sample of hypofractionations with BEDs, that is, more than 71 (20 Gy × 3 Fx, \( d_0 = 2 \)), otherwise, the results would be similar to the SPACE study.
Another retrospective analysis on 508 cases with SBRT revealed gross tumor volume BED was associated with in-field failure and the cutoff value should be more than 110 ($\alpha/\beta = 10$). The lower prescription doses (ie, 12 Gy $\times$ 4 or 10 G $\times$ 5) compared with 18 Gy or 20 Gy $\times$ 3 should be avoided for squamous cell carcinomas. The validated results of modified model as Table 8 indicated that the BEDs of lower ones were

**Figure 1.** The clonogenic survival curve of A375 within a range of 0 to 8 Gy. Comparison of predictions of Equation 3 (LQ), Equation 3-L (LQ model fitted at low-dose range [0-4 Gy]), Equation 6 ($d_0 = 5$ Gy), and Equation 8. Equation 6 fit well in low-dose range similar with Equation 3-L, and as good as Equation 8 in high-dose range.

**Table 5.** $\chi^2$ test for Actual and Fitted Data of Equation 3 With Parameters got at Low-Dose Range and Equation 6 With Optimal Parameters in Several Cell Lines.

| Static Value | Equation | hFB ($d_0 = 3$ Gy) | A375 ($d_0 = 5$ Gy) | A549 ($d_0 = 2$ Gy) | A549 ($d_0 = 2$ Gy, $b_1 = 0$) |
|--------------|----------|---------------------|---------------------|---------------------|-------------------------------|
| Hypothesis test | Equation 3 (with parameters got at low-dose range) | $0.1763$ | $0.3015$ | $1.1349$ |
| ($95\%$ confidence) | Equation 6 (with optimal parameters) | $0.0034$ | $0.0033$ | $0.0015$ | $0.1059$ |
| $\chi^2/df$ | Equation 3 (with parameters got at low-dose range) | $0.8384$ | $0.5829$ | $0.2867$ |
| | Equation 6 (with optimal parameters) | $0.9536$ | $0.9541$ | $0.9692$ | $0.7449$ |
| $P$ | Equation 3 (with parameters got at low-dose range) | $0.8384$ | $0.5829$ | $0.2867$ |
| | Equation 6 (with optimal parameters) | $0.9536$ | $0.9541$ | $0.9692$ | $0.7449$ |

**Table 6.** The BED Values of 6 Gy $\times$ 5 Fx and 2 Gy $\times$ 30 Fx for the Treatments of Melanoma.

| No. | Dose Fraction | BED | $d_0 = 4$, $(\alpha/\beta = 2.05, \alpha/\beta_1 = 0.52)$ | $d_0 = 5$, $(\alpha/\beta = 0.67, \alpha/\beta_1 = 0.18)$ |
|-----|---------------|-----|---------------------------------|---------------------------------|
| CFRT | 2 Gy $\times$ 30 Fx | 152 | 119 | 239 |
| SBRT | 6 Gy $\times$ 5 Fx | 167 | 147 | 319 |
| BED Ratio (SBRT/CFRT) | 1.1 | 1.2 | 1.3 |

**Table 7.** The BED Values of 22 Gy $\times$ 3 Fx and 2 Gy $\times$ 35 Fx for the Treatments of NSCLC.

| No. | Dose Fraction | BED | $d_0 = 2$, $(\alpha/\beta = 8.5, \alpha/\beta_1 = 0.67, \alpha/\beta_1 = 0.18)$ | $d_0 = 3$, $(\alpha/\beta = 19.8, \alpha/\beta_1 = 0.67, \alpha/\beta_1 = 0.18)$ |
|-----|---------------|-----|---------------------------------|---------------------------------|
| CFRT | 2 Gy $\times$ 35 Fx | 79 | 86 | 77 |
| SBRT | 22 Gy $\times$ 3 Fx | 157 | 76 | 67 |
| BED Ratio (SBRT/CFRT) | 2.0 | 0.9 | 0.9 |

Abbreviations: BED, biologically effective dose; CFRT, conventional fractioned radiotherapy; SBRT, stereotactic body radiation therapy.

Another retrospective analysis on 508 cases with SBRT revealed gross tumor volume BED was associated with in-field failure and the cutoff value should be more than 110 ($\alpha/\beta = 10$). The lower prescription doses (ie, 12 Gy $\times$ 4 or 10 G $\times$ 5) compared with 18 Gy or 20 Gy $\times$ 3 should be avoided for squamous cell carcinomas. The validated results of modified model as Table 8 indicated that the BEDs of lower ones were
only 62 and 66 far below the CFRT (2 Gy × 30), which was close to 20 Gy × 3. The threshold of 110 would not be used prevalently as SBRT lacked explicit definition, while the modified model was suitable for various dose fractionations.

The reliability of modified model could be validated via the linear relationship between new BEDs and LC, which was congenial with reason and common sense and should be more obviously when the LCs of 7.5 Gy × 8 Fx and 5 Gy × 10 Fx in central type were analyzed respectively.

A new EUD model was derived to compare treatment plans with different time-dose fraction schemes and dose distributions. The sectional function solves the problem that the LQ model is not accurate and applicable to high doses per fraction. Based on this mathematical model, a new EUD model derived is instructive to compare the radiation effects of volumes with various dose distributions.

As the clinic application of hypofraction and SBRT gains popularity rapidly, correcting the deviation of LQ and BED models in high dose area grows more important. This research improved cell survival curve fitting method, whose merit lay in that it broke up the dose range of α, β parameters limit on the previous method. Such equation promotes hands-on clinic experience retaining α, β arguments, expands judicious use of BED attaching a correction parameter of hypofraction dose. We established the formula of modified BED and EUD model, based on the piecewise curve functions, which could serve for the transformation of different dose fraction equivalents and the standardization of heterogeneity distribution comparison. Quite close analogous calculation results are obtained by applying modified BED model to the different dose fractions, whose efficacy showing similar and BED-based LQ model showing big variations in the cancers with high α/β values. The modified models are worthy to be testified and brought into wide use.

### Table 8. The Modified BED Values of Various Hypofractioned NSCLC Treatments.

| Dose Fraction | \( \text{BED}_{m} (d_0 = 2) \) | \( \text{BED}_{m} (d_0 = 3) \) |
|---------------|-----------------|-----------------|
| 2 Gy × 30 Fx  | 74              | 66              |
| 2 Gy × 35 Fx  | 86              | 77              |
| 15 Gy × 3 Fx  | 57              | 45              |
| 12 Gy × 4 Fx  | 62              | 50              |
| 10 Gy × 5 Fx  | 66              | 54              |
| 5 Gy × 10 Fx  | 67              | 60              |
| 20 Gy × 3 Fx  | 71              | 55              |
| 11 Gy × 5 Fx  | 72              | 58              |
| 6 Gy × 10 Fx  | 81              | 72              |
| 7.5 Gy × 8 Fx | 81              | 72              |
| 7 Gy × 10 Fx  | 94              | 78              |

Abbreviations: BED, biologically effective dose; NSCLC, non-small cell lung cancer.

### Table 9. The Modified BED Values of Various Hypofractionations and Local Control in Stage I NSCLC Treatments.

| Type of NSCLC | Peripheral | Central | Dose Fraction | \( \text{BED}_{m} (d_0 = 2) \) | \( \text{BED}_{m} (d_0 = 3) \) | Local Control (95% CI) |
|---------------|------------|---------|---------------|-----------------|-----------------|----------------------|
| 20 Gy × 3 Fx  | 71         | 66      | 55            | 81              | 72              | 97%                  |
| 10 Gy × 5 Fx  | 66         | 54      | 54            | 67              | 60              | 85%                  |
| 12 Gy × 4 Fx  | 62         | 50      | 50            | 66              | 54              | 85%                  |

Abbreviations: BED, biologically effective dose; CI, confidence interval; NSCLC, non-small cell lung cancer.
Authors’ Note
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