A rare case of recurrent ovarian cancer with NTRK1-TPM3 gene rearrangement

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Abstract

Background:

*NTRK* gene fusion is rare in gynecologic cancer. Entrectinib is a novel targeted drug which is a potent inhibitor of TRK A, B and C. Here, we present a case of recurrent ovarian cancer with *NTRK1-TPM3* rearrangement, which was detected by next-generation sequencing (NGS) and treated with entrectinib.

Case Presentation:

A 56-year-old woman was diagnosed as having stage IV ovarian cancer due to pleural effusion. Neoadjuvant chemotherapy and interval debulking surgery followed by chemotherapy were performed. Ten months after completion of chemotherapy, the patient’s disease recurred. She was treated with multimodal therapy for recurrence. DNA-based NGS detected *NTRK1-TPM3* rearrangement and entrectinib was started. However, the patient’s disease progressed despite six weeks’ administration of entrectinib, and one month after discontinuation of entrectinib, she died. After her death immunohistochemistry with a pan-Trk monoclonal antibody was performed to determine the expression of *NTRK*. However, immunohistochemistry was negative for *NTRK*.

Conclusion:

We presented a rare case of recurrent ovarian cancer with *NTRK1-TPM3* gene fusion, in which entrectinib was not effective. While *NTRK* gene fusion was detected by DNA-based NGS, immunohistochemistry was negative for *NTRK*. Immunohistochemistry should be performed for confirmation of NTRK protein expression before entrectinib administration.

Background

*NTRK* gene fusions are consistently detected in rare types of cancers (secretory breast carcinoma, mammary analogue secretary carcinoma, congenital infantile fibrosarcoma, and congenital mesoblastic nephroma) and they are novel therapeutic targets across multiple tumor types. [1–3] On the other hand, these gene fusions are rare in common adult cancers. [1, 2] In gynecologic oncology, *NTRK* gene fusion is also rare, although there are several reports of uterine sarcoma with this fusion gene. [4–7] A previous cohort study showed that *NTRK1-TPM3* is most frequent in *NTRK1* fusions across multiple histologies. [2] Immunohistochemistry (IHC) staining, fluorescence in situ hybridization, reverse transcriptase polymerase chain reaction, DNA-based next-generation sequencing (NGS) and RNA-based NGS are used to identify patients with *NTRK* gene fusion cancer. Each method to detect *NTRK* gene fusion has its own characteristics. [3]

Entrectinib is a potent inhibitor of *TRKA, TRKB, TRKC, ROS1, and ALK*, and is specifically designed to have systemic activity. In gynecologic oncology, treatment using entrectinib is rare because of the low
frequency of \textit{NTRK} fusions. [1] Here, we report a case of recurrent ovarian cancer (OC) with \textit{NTRK1-TPM3} gene fusion and that was treated with entrectinib.

\section*{Case Presentation}

In September 2013, a 56-year-old woman was referred to our hospital with bilateral ovarian tumor, multiple disseminations in the peritoneum, bilateral pleural effusion, and multiple swellings of the pelvic, and paraaortic lymph nodes. Her serum level of cancer antigen 125 (CA125) was elevated to 1740 U/ml. She was diagnosed as having stage IIIC OC according to the International Federation of Gynecology and Obstetrics (FIGO) 1988 because pleural effusion cytology was positive. Paclitaxel (175 mg/m\(^2\)) and carboplatin (area under the curve 6) were started as neoadjuvant chemotherapy. After four courses of chemotherapy, computed tomography (CT) revealed a reduction in tumor size. Interval debulking surgery including abdominal hysterectomy, bilateral salpingo-oophorectomy, omentectomy, and pelvic and paraaortic lymphadenectomy, was performed. Histopathological diagnosis was high-grade serous carcinoma. Following this surgery, another three courses of the same regimen were administered, and the patient achieved clinical complete response.

Ten months after the last therapy, OC recurred. Because of the disease persistence, she was treated with nine regimens of chemotherapy combined with two surgeries for recurrent tumors; the first was partial hepatectomy due to dissemination to the liver, and the second was enterectomy due to recurrence in the mesentery. Microsatellite stability was detected in specimens from the second recurrence resection.

Six years after initiation of therapy, FoundationOne® CDx (Foundation Medicine, Cambridge, MA), which is DNA based NGS and covers 324 genes, was performed based on the patient's archival tumor tissue from the second recurrence resection. This revealed a missense variant of \textit{TP53} (c.731G > A) and a rearrangement between exon 11 of \textit{NTRK1} (NM_002529) and somewhere around exon 2–3 of \textit{TPM3} (pos1="chr1:156844554–156844771", pos2="chr1:154155588–154155822"). Oral entrectinib (600mg/day) was started after discussing with experts. Six weeks after initiation of entrectinib, the patient's serum CA125 level elevated to 4360 U/ml, although it was 1712 U/ml before initiation of entrectinib and CT revealed progression of liver metastasis. (Fig. 1) Adverse events during entrectinib administration comprised grade 2 dysgeusia. One month after discontinuation of entrectinib, the patient died. (Fig. 2)

After the patient's death, IHC staining with a pan-Trk monoclonal antibody (mAB) clone EPR17341 (Abcam, Cambridge, MA) was performed to assess for \textit{TRKA}, \textit{TRKB}, and \textit{TRKC} expression as previously described. [8] This mAB clone is most commonly used and has been investigated thoroughly. In addition, this mAB clone reacts with a conserved proprietary peptide from the C-terminus of \textit{TRKA}, \textit{TRKB} and \textit{TRKC}, and is therefore reactive to any oncogenic \textit{NTRK} fusion. [3] IHC was negative for all specimens from the primary site resection, as well as the first and second recurrent site resections. (Fig. 3)

\section*{Discussion}
Here we presented a case of recurrent OC with \textit{NTRK1-TPM3} rearrangement. Additionally, the present case demonstrates the discrepancy between gene rearrangement detected by NGS and protein expression. This discrepancy may be a biomarker for predicting the ineffectiveness of entrectinib for cancers with \textit{NTRK} rearrangement detected by NGS.

In the current case, NGS revealed \textit{NTRK1-TPM3} rearrangement and a missense variant of \textit{TP53}. There are few approved therapies for \textit{TP53}, although almost all cases of ovarian high-grade serous carcinoma (95%) have somatic \textit{TP53} variants. [9] On the other hand, \textit{NTRK} fusions are oncogenic drivers and novel targets. Doebele et al. reported the safety and activity of entrectinib in adult patients with advanced or metastatic \textit{NTRK} fusion-positive cancer across three clinical trials (ALKA-372-001, STARTRK-1 and STARTRK-2). In these trials, only one ovarian cancer patient was included. They showed that the objective response rate, which included complete response and partial response, was 57% (95% CI 43.2–70.8). The median duration of response was 10 months (95% CI 7.1 to not estimable) and the percentage of progressive disease (PD) was only 7%. However, the characteristics of cases with PD remained unclear in their report. [1]

In the present case, entrectinib was administered because NGS revealed \textit{NTRK1-TPM3} rearrangement and entrectinib was recommended after a discussion among experts. However, this novel target drug was ineffective. \textit{NTRK} protein was not expressed, although IHC testing with a pan-Trk mAB clone [EPR17341] was performed. A previous study reported that gene fusions involving \textit{NTRK1, 2, and 3} and their partner genes result in a constitutive activation or overexpression of TRK receptors, potentially leading to oncogenesis. [10] Additionally, other reports have shown that pan-Trk IHC yielded a sensitivity of 75–95.2%, and a specificity of 92–100% and that the sensitivity of pan-Trk IHC for \textit{NTRK1} was 96.2%. [3, 8, 11, 12] Pan-Trk IHC is a reliable screening method for the detection of \textit{NTRK} gene fusions based on this date. Moreover, pan-Trk IHC can rapidly assess malignancies which may harbor possible \textit{NTRK} fusions in order to determine eligibility of patients for targeted therapy with TRK inhibitors. [8] However, it should be considered that there are \textit{NTRK} rearrangements which are found to be negative by IHC, and can only be detected by NGS, such as in the present case.

Drilon et al. reported the efficacy of larotrectinib, which is a selective inhibitor of TRKA, TRKB and TRKC. In their study, six of an initial 55 patients showed primary resistance to larotrectinib. Three patients had tumor material available for central analysis, and in all three cases, pan-Trk IHC did not reveal the presence of TRK protein expression. This indicated that the rearrangements detected by NGS were false positives or that the identified fusion genes were not expressed at the protein level. [13] It is considered that entrectinib has the same characteristics as larotrectinib with regard to discrepancy between gene fusion and protein expression, as observed in the current case, and that this finding may be a key to predict the ineffectiveness of entrectinib for cancers with \textit{NTRK} rearrangement detected by NGS.

To the best our knowledge, this is the first case report of OC with \textit{NTRK} rearrangement. It is known that a small percentage of common adult cancers carry fusions of \textit{NTRK} genes. [2] A large cohort study revealed that the frequency of \textit{NTRK} gene fusions was 0.25% of general cancers. [2, 12] Therefore,
physicians have few chances to experience this molecular characteristic. However, physicians should be aware of the pitfall that NTRK protein may not expression even if NGS reveals \textit{NTRK} rearrangement.

\textbf{Conclusion}

In conclusion, we here presented a rare case of recurrent OC with \textit{NTRK1-TPM3} fusion. Physicians should be aware of the discrepancy of DNA rearrangement and protein expression, and IHC should be performed for confirmation of NTRK protein expression before entrectinib administration.

\textbf{Abbreviations}

\begin{itemize}
  \item IHC \quad \text{immunohistochemistry}
  \item NGS \quad \text{next-generation sequencing}
  \item OC \quad \text{ovarian cancer}
  \item CA-125 \quad \text{cancer antigen 125}
  \item FIGO \quad \text{International Federation of Gynecology and Obstetrics}
  \item CT \quad \text{computed tomography}
  \item mAB \quad \text{monoclonal antibody}
  \item PD \quad \text{progressive disease}
\end{itemize}

\textbf{Declarations}

\textbf{Ethics approval and consent to participate.}

This study was approved by the Ethical Committee of Fukushima Medical University.

\textbf{Consent for publication}

Written informed consent was obtained from the patient for publication of this case report and the accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

\textbf{Availability of data and materials}
The data used or analyzed are all included in this published article

**Competing interests**

The authors declare that they have no competing interests.

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**Author's contribution**

YE, TW and MS drafted the manuscript. YE, TW, RS, HS, YN, ES, MU, NK, SF, SS, SS and KF managed this patient. MS, KS and KK performed immunohistochemical staining. All authors have read and approved the final manuscript.

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Figures

Figure 1

Computed tomography of liver metastasis a. Before administration of entrectinib b. Six weeks after beginning administration of entrectinib.
Figure 2

Clinical flowchart of this patient CA125: Cancer antigen-125; MSI: microsatellite instability test; NGS: next generation sequencing; DOD: dead of disease; S, surgery; TC, paclitaxel and carboplatin; TP, paclitaxel and cisplatin; BV, bevacizumab; PLD, pegylated liposomal doxorubicin; GEM, gemcitabine; NGT, nogitecan; wPTX, weekly paclitaxel; VP-16, etopocide; CDGP, nedaplatin; Entr, entrectinib.
Figure 3

Histopathological and immunohistochemical features (× 100) a, b. Interval debulking surgery specimen (tumor resection following neoadjuvant chemotherapy) c, d. Liver dissemination specimen (first resection for recurrence) e, f. Mesentery dissemination specimen (second resection for recurrence) a, c, e. Hematoxilin and eosin staining b, d, f. Immunohistochemical stain for pan-Trk. Immunohistochemistry was negative for pan-TRK among all specimens.