Rice (Oryza) hemoglobins [v2; ref status: indexed, http://f1000r.es/4vp]

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Abstract

Hemoglobins (Hbs) corresponding to non-symbiotic (nsHb) and truncated (tHb) Hbs have been identified in rice (Oryza). This review discusses the major findings from the current studies on rice Hbs. At the molecular level, a family of the nshb genes, consisting of hb1, hb2, hb3, hb4 and hb5, and a single copy of the thb gene exist in Oryza sativa var. indica and O. sativa var. japonica. Hb transcripts coexist in rice organs and Hb polypeptides exist in rice embryonic and vegetative organs and in the cytoplasm of differentiating cells. At the structural level, the crystal structure of rice Hb1 has been elucidated, and the structures of the other rice Hbs have been modeled. Kinetic analysis indicated that rice Hb1 and 2, and possibly rice Hb3 and 4, exhibit a very high affinity for O₂, whereas rice Hb5 and tHb possibly exhibit a low to moderate affinity for O₂. Based on the accumulated information on the properties of rice Hbs and data from the analysis of other plant and non-plant Hbs, it is likely that Hbs play a variety of roles in rice organs, including O₂-transport, O₂-sensing, NO-scavenging and redox-signaling. From an evolutionary perspective, an outline for the evolution of rice Hbs is available. Rice nshb and thb genes vertically evolved through different lineages, rice nsHbs evolved into clade I and clade II lineages and rice nshbs and thbs evolved under the effect of neutral selection. This review also reveals lacunae in our ability to completely understand rice Hbs. Primary lacunae are the absence of experimental information about the precise functions of rice Hbs, the properties of modeled rice Hbs and the cis-elements and trans-acting factors that regulate the expression of rice hb genes, and the partial understanding of the evolution of rice Hbs.
Amendments from Version 1

Major differences between the revised and the published version of this review are: (1) a figure (Figure 3) for rice non-symbiotic Hb1-5 and rice and Arabidopsis tHbs sequence alignments was included, (2) a table (Table 2) for the rate and equilibrium constants for the reaction of oxygen from rice Hb1 and 2 and selected plant and non-plant Hbs was included, (3) structural and biophysical properties for rice tHb are discussed in a separate subsection within section “Structure and biophysical properties of rice hemoglobins”, (4) in all cases we indicated that NO binds to oxyHb, (5) care was taken when properties for predicted structures of rice Hb2-5 and tHb were postulated/hypothesized by preceding postulates/hypothesis with “possible”, “probable”, “suggest”, etc., (6) a statement on the biological relevance of the folding pathways for rice Hb1-5 was included, (7) similarity values for rice Hb1-5 and tHb between O. sativa var. japonica and O. sativa var. indica were included, and (8) the number of references was reduced.

See referee reports

Abbreviations
2,4-D, 2,4-dichlorophenoxyacetic acid; ARR1, Arabidopsis response regulator 1; BCIP, 5-bromo-4-chloro-3-indolyphosphate; Hb, hemoglobin; Lb, leghemoglobin; MIP1, macrophage inflammatory protein 1; mya, million of years ago; NBT, nitro-blue tetrazolium; nsHb, non-symbiotic hemoglobin; nsHb-1, non-symbiotic hemoglobin type 1; nsHb-2, non-symbiotic hemoglobin type 2; nsHb-I, clade I non-symbiotic hemoglobin; nsHb-II, clade II non-symbiotic hemoglobin; RT-PCR, reverse transcriptase-polymerase chain reaction; SNP, sodium nitroprusside; tHb, truncated (2/2) hemoglobin.

Introduction
Two decades ago Taylor and co-workers reported the cloning and sequencing of a hemoglobin (Hb) cDNA from barley. This was the first report about the existence of Hbs in monocotyledonous plants. Since then Hbs have been identified in a number of monocots, including rice, maize and wheat. Rice Hbs and genes coding for these proteins are rather well characterized, thus in some aspects rice Hbs are a model to understand monocot and other land plant Hbs. However, the accumulated information on rice Hbs over the last seventeen years is scattered. This review discusses major findings from the study of rice Hbs including a historical perspective, and proposes biochemical and physiological mechanisms for rice Hbs based on information available about rice Hbs and other monocot and land plant Hbs. For general aspects and the biochemistry, physiology and evolution of plant Hbs, we recommend to the reader reviews published elsewhere.

Generalities on hemoglobins
Hb is known to the reader because this protein is responsible for the red color of vertebrates’ blood. However, Hbs are widely distributed in living organisms, ranging from bacteria to mammals. The tertiary structure of Hbs consists of a specific arrangement of 6 to 8 α-helices (designated with letters A to H) known as the globin-fold. This protein folding forms a hydrophobic pocket where a heme prosthetic group is located. Two structural types of the globin-fold have been identified in Hbs: the 2/2- and 3/3-folding. In the 2/2-Hbs, helices B and E overlap to helices G and H and in the 3/3-Hbs helices A, E and F overlap to helices B, G and H. Likewise, three evolutionary families have been identified in Hbs: the M, S and T Hb families. The M Hbs, which exist in bacteria and eukaryotes, include flavoHbs and single domain globins, the S Hbs, which exist in bacteria and some fungi, include globin-coupled sensors, protoglobins and single domain globin sensors, and the T Hbs, which exist in bacteria, unicellular eukaryotes and plants, include truncated Hbs (tHbs). Canonical T Hbs from bacteria and unicellular eukaryotes are ~100 to 120 amino acids in length, however plant T Hbs are longer than canonical T Hbs because of the existence of extra amino acids at the N- and C-terminal. The M and S Hbs fold into the 3/3-folding whereas the T Hbs fold into the 2/2-folding.

A variety of ligands bind to the heme iron of Hbs, including O2 and NO. Reversible binding of O2 is closely associated to the major function of Hbs in organisms, which is the transport of O2. Binding of NO by oxygenated Hbs is essential to NO-detoxification via a NO-dioxygenase activity. Several additional functions have been reported for Hbs, including dehaloperoxidase activity and reaction with free radicals, binding and transport of sulfide and lipids, and O2-sensing. This indicates that in vivo Hbs might be multifunctional proteins.

Land plant hemoglobins
Land plant Hbs were first identified by Kubo in soybean root nodules. Few years after Kubo’s discovery these proteins were named as leghemoglobins (Lbs) by Virtanen and Lane because they were only found in the symbiotic (N2-fixing-) nodules of the leguminous plants. Lbs are the most abundant soluble proteins in nodules (e.g. in soybean nodules their concentration is as high as 3 mM). The x-ray analysis of lupin Lb revealed that the tertiary structure of Lbs was remarkably similar to that of the sperm whale myoglobin. This evidence demonstrated that Lbs are plant Hbs and indicated that plant and animal Hbs evolved from a common ancestor more than 600 mya. Subsequent work led to the identification of Lb-like (or symbiotic) Hbs in nodules of actinorhizal plants, purification of an Hb from the root nodules of the dicotyledonous non-legume Parasponia andersonii, cloning and sequencing of an Hb gene from the non-nodulating dicot Trema tomentosa and detection of Hbs in non-symbiotic organs from several land plants, including primary bryophytes and evolved angiosperms. Until now three types of Hbs have been identified in land plants: the symbiotic Hbs, which include Lbs, that are specifically located within nodules of the N2-fixing land plants, and the non-symbiotic (nsHbs) and truncated (tHbs) Hbs, that are located within non-symbiotic and symbiotic organs of primitive and evolved land plants. Based on sequence similarity the nsHbs are further classified into type 1 and type 2 nsHbs (nsHb-1 and nsHb-2, respectively). Distribution of hemoglobins in monocotyledonous plants
Monocots are a large family of flowering plants that includes cereals, such as rice, maize and wheat, and are the main source of food for humans. Because of this, during that last decade the genomes of a number of cereals have been sequenced. This allowed the identification of novel cereal Hbs. The search of Hb genes in databases by G. Rodriguez-Alonso and R. Arredondo-Peter.
revealed that nsHb and tHb sequences exist in the Brachypodium distachyon, Hordeum vulgare (barley), Oryza glaberrima (rice), O. rufipogon (rice), O. sativa (rice) var. indica, O. sativa (rice) var. japonica, Panicum virgatum (switchgrass), Setaria italica (foxtail millet), Sorghum bicolor (sorghum), Triticum aestivum (wheat) and Zea mays ssp. mays (maize) genomes. The highest number of nsHbs (5) exists in O. sativa var. indica and O. sativa var. japonica, whereas one to three nsHbs exist in barley, Brachypodium, foxtail millet, maize, O. glaberrima, O. rufipogon, sorghum, switchgrass and wheat. Also, with the exception of wheat, which contains two copies of the tthb gene, a single copy of tthb was identified in the genome of Brachypodium, barley, O. sativa var. indica, O. sativa var. japonica, switchgrass, foxtail millet, sorghum and maize. Little is known about Hbs from non-cultivated monocots. The only Hb reported from a non-cultivated monocot is that of teosinte (Z. mays ssp. parviglumis)\(^1\), which is postulated as the ancestor of maize.\(^2\) Analysis by Southern blot using the teosinte hb gene as probe showed that apparently a single copy of hb exists in teosinte (J. Sáenz-Rivera and R. Arredondo-Peter, unpublished results). Sequence comparison revealed that maize and teosinte Hb polypeptides are identical\(^3\).

Early search and identification of rice hemoglobins

Monocots were a target for searching Hbs after these proteins were detected in non-symbiotic organs of dicotyledonous plants (see subsection above). At that time, monocot genomes had not been sequenced. Searching approaches consisted in detecting Hb polypeptides and hb genes by spectroscopy and molecular biology methods, respectively. Attempts to detect absorption maxima in the Soret (~410 nm) and Q (~500 to 550 nm) regions, which are characteristic of ferric (Fe\(^{3+}\)) and ferrous (Fe\(^{2+}\)) and liganded Hbs\(^5\)\(^6\), were unsuccessful (R. V. Klucas and C. A. Appleby, unpublished results) mostly due to the very low Hb concentration (~50 to 100 nM) in plant non-symbiotic organs\(^7\). At the molecular level a consensus probe designed from legume and non-legume (T. tomentosa, P. andersonii and Casuarina glauca) Hb sequences\(^8\) hybridized with hb-like sequences from rice and other monocot total DNAs (Figure 1). This observation suggested that hb sequences exist in monocots, however hybridizing fragments were not subsequently cloned and sequenced in order to verify if they actually corresponded to hb genes.

Rice Expressed Sequence Tags (ESTs) were first deposited in databases early in the 1990’s. The first rice Hb (Hb1 and Hb2) sequences were detected from ESTs deposited in the DNA Data Bank of Japan (DDBJ) database\(^9\). Rice Hb1 and Hb2 corresponded to clones C741 and C2576 with DDBJ accession number D15507 and D38931, respectively. Rice hb1 and hb2 genes were subsequently amplified by PCR, cloned and sequenced. Sequence analysis revealed that rice hb1 codes for non-symbiotic Hb1 and that rice hb2 codes for non-symbiotic Hb2\(^2\). Afterwards, sequencing of the rice (O. sativa L. ssp. indica) genome more than a decade ago\(^10\) allowed the identification of a family of rice nshb genes and a single copy of the rice tthb gene (see subsection below).

Molecular biology of rice hemoglobins

Rice hemoglobin genes

The O. sativa var. indica and O. sativa var. japonica genomes are fully sequenced, and the O. glaberrima and O. rufipogon genomes are partially sequenced. Rice genome sequences are mainly available from the GenBank (www.ncbi.nlm.nih.gov) and Phytome (http://www.phytozome.org/) databases. Search of Hb sequences in the

![Figure 1. Early (1991) detection of rice, maize, sorghum and wheat hb-like sequences by dot-blot hybridization (R. Arredondo-Peter, unpublished results).](image-url) Approximately 20 μg of undigested total DNA was used as template and a consensus oligonucleotide for legume and non-legume plant Hbs\(^10\) was used as probe. Sequence of the consensus probe was 5'-GTA GCC TAT GAT GAA TTG GCA GCT GCA ATT AAG-3'. The probe was labeled by nick translation with Biotin-dATP using a Bionick labeling system (Gibco BRL). The membrane was prehybridized with SSC 2x for 4h at 42°C, hybridized overnight at the same temperature, washed at high stringency (SSC 2x/SDS 0.1% for 3 min at room temperature, SSC 0.2x/SDS 0.1% for 15 min at room temperature and SSC 0.16x/SDS 0.1% for 15 min at 65°C) and incubated with the streptavidin-alkaline phosphatase conjugate and the BCIP/NBT mix to develop color. Animal (salmon sperm and calf thymus) and legume DNAs were included as negative and positive controls, respectively.
above databases showed that a family of the nshb genes, consisting of hb1, hb2, hb3, hb4 and hb5, and a single copy of the thb gene exist in the O. sativa var. indica and O. sativa var. japonica genomes. A single copy of the nshb gene was detected in the O. glaberrima and O. rufipogon genomes, however thb genes have not yet been detected in these plants\(^\text{[2]}\). Given that the sequencing of the O. glaberrima and O. rufipogon genomes is in progress the identification of hb genes in these genomes is incomplete. Thus, the following discussion will focus on the O. sativa var. indica and O. sativa var. japonica hbs. However, we must clarify to the reader that the sequence of Hb1, Hb2, Hb3, Hb4 and thb and Hb5 polypeptides are 100% and 97% identical between O. sativa var. indica and O. sativa var. japonica, respectively. Therefore, the subsequent discussion on the O. sativa hbs will indistinctively correspond to either O. sativa var. indica or O. sativa var. japonica.

The structure of known rice hb genes corresponds to four exons and three introns, with introns located at similar position as all of the known plant hb genes\(^\text{[3]}\). Canonical TATA boxes and a variety of potential promoters exist upstream of the rice hb genes which suggests that rice hbs are functional and that the regulation of the hb genes in this plant is complex\(^\text{[4,5,6]}\). Figure 2 shows the localization of hbs in the O. sativa chromosomes and mapping of hbs in the O. sativa genome. Rice hb1, hb3 and hb4 cluster forming the hb1-hb4 cluster\(^\text{[7]}\) which is localized in chromosome 3. Rice hb2 is also localized in chromosome 3 but 467 kb upstream of the hb1-hb4 cluster. In contrast, rice hb5 and thb genes are localized in chromosomes 5 and 6, respectively (Figure 2A). Rice hbs are flanked by a variety of genes with known and unidentified functions (Figure 2B). However, with the exception of genes coding for a ternary complex factor macrophage inflammatory protein MIP1 and an ubiquitin fusion protein which are located 239 and 411 nucleotides up- and downstream of the hb1-hb4 cluster, respectively, distance of flanking genes to hbs is >1 kb. This suggests that co-expression of hb and flanking genes is unlikely.

Gene expression and localization of hemoglobins in rice organs

The expression of hb genes and localization of Hb polypeptides have been analyzed in rice growing under normal and stressed conditions. Under normal conditions the expression level of rice nshbs was low\(^\text{[8,9]}\). However, analysis by RT-PCR revealed that hb1, hb2 and hb5 genes were expressed in embryonic and vegetative organs obtained from rice plants grown under a normal environment\(^\text{[10,11]}\). Specifically, transcripts for rice Hb1 were detected in embryos, seminal roots, leaves and roots, transcripts for rice Hb2 were detected in embryos, coleoptiles, seminal roots and leaves, and transcripts for rice Hb5 were detected in embryos, coleoptiles, seminal roots, leaves and roots. Likewise, evaluation of the β-glucuronidase (GUS) activity from a construct containing the rice nshb2 gene promoter that is responsive to the cytokinin-regulated ARR1 trans-acting factor showed that this promoter is activated in roots, the vasculature of young leaves, flowers and the pedicel/stem junction of transgenic Arabidopsis\(^\text{[12]}\). In addition, a variety of potential promoters was identified upstream of the rice nshb genes, such as those involved in the ethylene synthesis, photoregulation, heat shock response and plant defense signaling\(^\text{[13,14]}\). However the activities of these promoters have not been determined.

Transcriptomic analyses revealed that nsHb and tHb transcripts coexist in rice embryonic and vegetative organs (Table 1). This evidence suggests that nsHb (i.e. Hb1, Hb2, Hb3, Hb4 and Hb5) and tHb polypeptides coexist and probably function in rice organs. Immunoanalysis by Western blot and confocal microscopy using a polyclonal anti-rice Hb1 antibody revealed that Hb polypeptides exist in rice seeds and in rice leaves and roots from 2 to 14 weeks after seed germination. These analyses also revealed that Hb polypeptides exist in the cytoplasm of differentiating cells of the root cap, schlerenchyma, aleurone, and in the vasculature, principally in the differentiating xylem\(^\text{[15,16,17]}\). However, the anti-Rice Hb1 antibodies cross-react with different rice Hbs (G. Sarath and E. J. H. Ross, unpublished results) and thus it is not known which Hb polypeptides were detected in the above analyses by the anti-rice Hb1 antibodies.

It is well documented that land plant hb genes are either up- or down-regulated by stress conditions\(^\text{[18,19,20]}\). Table 1 shows that Hb transcripts coexist in rice growing under cold, drought and salt stress conditions. Also, Ohwaki and co-workers\(^\text{[21]}\) reported that

| Rice organs     | Hb1 transcripts | Hb2 | Hb3 | Hb4 | Hb5 | tHb |
|-----------------|-----------------|-----|-----|-----|-----|-----|
| Normal conditions |                 |     |     |     |     |     |
| Seed            |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Vegetative rice |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Leaves          |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Stems           |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Roots           |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Reproductive rice |                | ✓   | ✓   | ✓   | ✓   | ✓   |
| Inflorescence   |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Leaves          |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Stems           |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Roots           |                 | ✓   | ✓   | ✓   | ✓   | ✓   |
| Reproductive organs |         | ✓   | ✓   | ✓   | ✓   | ✓   |
| Lemma           |                 | ✓   | ✓   | ✓   | ✓   |
| Anther          |                 | ✓   | ✓   | ✓   | ✓   |
| Palea           |                 | ✓   | ✓   | ✓   | ✓   |
| Ovary           |                 | ✓   | ✓   | ✓   | ✓   |
| Pistil          |                 | ✓   | ✓   | ✓   | ✓   |
| Stress conditions |                | ✓   | ✓   | ✓   | ✓   | ✓   |
| Reported as part of the plant response to stress |     | ✓   | ✓   | ✓   | ✓   | ✓   |

Table 1. Detection of Hb transcripts in organs from rice growing under normal and (cold, drought and salt) stressed conditions.
Figure 2. Localization of hbs in the O. sativa chromosomes (A) and mapping of hb genes into the O. sativa genome (B). The hb genes were localized in the rice chromosomes by BlastN analysis using the rice (O. sativa) genome resource from the GenBank database as template and the sequence for the rice hb1, hb3 and hb4 (GenBank accession number AF335504), hb2 and hb5 (GenBank accession numbers AF335503 and EF061459, respectively) and thb (GenBank accession number NM_001064507) genes as probes. The hb (black boxes) and flanking (gray boxes) genes were mapped into 50 kb fragments of the O. sativa genome by BlastN2.2.26+ analysis using the Phytozome V9.1 server (www.phytozome.org) and the above hb sequences as probes. Arrows indicate the transcription orientation. Information for each gene corresponds to predicted protein (following the Phytozome nomenclature), locus name in the O. sativa genome and position at the O. sativa chromosome. Gene sizes and distance between genes are not shown at scale. Pltd, chloroplast chromosome; MT, mitochondrial chromosome.
nsbh1 and nshb2 are induced by nitrate, nitrite and NO in cultured rice cells. These observations indicate that rice hb genes response to a variety of stress conditions. However, the detection of Hb polypeptides by Western blot using the anti-rice Hb1 antibodies showed that level of Hbs increased in rice etiolated leaves and flooded roots, but not in rice plants subjected to oxidative (H2O2), nitrosative (SNP) and hormonal (2,4-D) stresses. These observations suggest that rice Hbs do not appear to be a part of a generalized stress response, but may be functional in plant organs subjected to specific stress conditions.

Structure and biophysical properties of rice hemoglobins

Rice hb genes are functional and code for Hb polypeptides with a predicted molecular mass of ~16 to 19 kDa. Also, sequences among rice nsHb polypeptides are highly similar: Hb1 and Hb2 are 93% similar to each other, Hb3 and Hb4 are 87.1% similar to each other, and 85.5% and 84.7%, and 79.2% and 82.2% similar to Hb1 and Hb2, respectively, and rice Hb1 and Hb5 are 67% similar to each other (Figure 3A).

**Figure 3.** Sequence alignment of rice Hbs. (A) Sequence alignment of rice nsHbs. Note the 11 amino acids deletion in rice Hb5 at position 75–85. Modified from Garrocho-Villegas et al. (B) Sequence alignment of rice and Arabidopsis (GenBank accession number AAK55409) tHbs. Distal and proximal His in rice nsHbs (H79 and H114, respectively) and proximal His (H100) and proposed distal Tyr/Tre (Y46/W113) in Arabidopsis and rice tHbs and conserved amino acids are shown with black and gray background, respectively. Helices are indicated with letters A to H based on the crystal structure of rice Hb1 in rice nsHbs and on the crystal structure of Arabidopsis tHb in rice tHbs. Left- and right-oriented arrows within a black circle in the rice and Arabidopsis tHbs sequence alignment delimit the globin domain.
Rice Hb1 was the first monocot nsHb whose crystal structure was elucidated. This protein crystallizes as a dimer when its concentration is ≥1 mM. After the elucidation of the rice Hb1 structure the tertiary structure of rice Hb2, Hb3, Hb4 and Hb5 (CASPUR PMDB ID PM0075009, PM0075873, PM0076005 and PM0075011, respectively) was predicted using computational methods and rice Hb1 (PDB ID 1D8U) as the structural homolog. The crystal structure of rice Hb1 and that of predicted rice Hb2, Hb3 and Hb4 is highly similar. The tertiary structure of these proteins consists of six helices that fold into the 3/3-folding (see subsection on Generalities on hemoglobins). However, the structure of rice Hb1 to 4 is characterized by the existence of a short pre-helix A located at the N-terminal and an extended and poorly ordered CD-loop. The heme pocket in these proteins differs from that in "traditional" Hbs because the proximal and distal His side chains coordinate the heme iron forming a hemichrome (Figure 4), resulting in that heme iron from rice Hb1 to 4 is hexacoordinate. Also, the amino acid residues (V50, S53, E125, V126, F129 and A130 from Figure 3A) located at the monomer-monomer interface of dimeric rice Hb1 are highly conserved in rice Hb2 to 4. This suggests that rice Hb1 to 4 can potentially form homo- or hetero-dimers if the hbl1 to 4 genes coexpress in rice organs. The tertiary structure of rice Hb5 also consists of six helices that fold into the 3/3-folding. However, rice Hb5 differs from rice Hb1 to 4 in missing 11 amino acids in helix E (Figure 3A) which results in that the length of the CD-loop and helix E in the predicted Hb5 structure are unusually long and short, respectively. An apparent consequence from this characteristic is that distal His is located far away (13.92 Å, compared to 2.11 Å in rice Hb1) from the heme iron within the predicted Hb5 structure, resulting in that heme iron from rice Hb5 could be pentacoordinate. The amino acid residues located at the monomer-monomer interface of dimeric rice Hb1 are poorly conserved in rice Hb5 (Figure 3A) which suggests that rice Hb5 exists in vivo as a monomer.

The folding pathway and kinetics of rice nsHbs were predicted using the Average Distance Map (ADM) method. This analysis indicated that rice Hb1 and Hb2 could fold in the C → N direction at a moderate rate, that rice Hb3 could fold in the N → C direction at a fast rate, and that rice Hb4 and Hb5 could fold in the N → C direction at a moderate rate. Thus, it appears that the predicted folding pathway and kinetics among rice nsHbs are diverse. Also, the ADM analysis showed that pre-helix A and CD-loop apparently do not play a role during the folding of rice nsHbs. The physiological relevance of the folding pathways for rice nsHbs, including the polypeptide association with the heme, is still not known.

**Spectroscopic characteristics of rice non-symbiotic hemoglobins**

Visible spectroscopy (see subsection Early search and identification of rice hemoglobins) is a tool to analyze the redox state of and ligand-binding to the heme iron of Hbs. Rice Hb1 is the only rice nsHb that has been spectroscopically characterized. This protein exhibits spectral characteristics that are similar to other Hbs. However, rice Hb1 exhibits distinctive absorption maxima in the deoxyferrous form: the unligated ferrous state exhibits maxima at 526 and 556 nm which are characteristic of hexacoordinate heme iron... This is in contrast to pentacoordinate Hbs which display a broad peak centered at 556 nm in their deoxyferrous form. The distal ligand that coordinates the heme iron in rice Hb1 was identified as His74 by site directed mutagenesis. Absorbance spectra of the ferric and deoxyferrous forms of an H74L mutant of rice Hb1 showed no evidence of His coordination. Also, the addition of exogenous imidazole to ferric and deoxyferrous forms of an H74L mutant of rice Hb1 showed no evidence of His coordination. In contrast to pentacoordinate Hbs which display a broad peak centered at 556 nm in their deoxyferrous form, the distal ligand that coordinates the heme iron in rice Hb1 was identified as His74 by site directed mutagenesis. Absorbance spectra of the ferric and deoxyferrous forms of an H74L mutant of rice Hb1 showed no evidence of His coordination. Also, the addition of exogenous imidazole to ferric and deoxyferrous forms of an H74L mutant of rice Hb1 showed no evidence of His coordination.

![Rice Hb1](image1.png)

**Rice Hb1**

![Rice tHb](image2.png)

**Rice tHb**

**Figure 4.** Crystal structure of rice Hb1 (PDB ID 1D8U) and predicted structure of rice tHb. The tertiary structure of rice tHb was modeled using the automated mode of the I-Tasser server (http://zhanglab.ccmb.med.umich.edu/I-TASSER) and the crystal structure of the Thermobifida fusca tHb (PDB ID 2BMM) as the structural homologue. Model for the rice tHb is deposited in the Caspur Protein Model Database (http://bioinformatics.cineca.it/PMDB/main.php) under the ID number PM0079484. Helices are indicated with letters A to H. Note the overlapping of helices A, E and F to helices B, G and H in (3/3-folding) rice Hb1, and overlapping of helices B and E to helices G and H in (2/2-folding) rice tHb. Heme prosthetic group is shown in dark green color and proximal and distal His are shown in light brown color.
deoxyferrous rice Hb5, resulting in that heme iron in rice Hb5 could be pentacoordinate.

Rate and equilibrium constants for the reaction of oxygen from rice non-symbiotic hemoglobins

Analysis of ligand-association and -dissociation rate constants of penta- and hexacoordinate Hbs using stopped-flow methods indicated that these proteins exhibit low to moderate and high affinity for O₂, respectively. Rice Hb1 is hexacoordinate and apparently rice Hb2 to 4 are hexacoordinate and rice Hb5 is pentacoordinate. The O₂-association rate constants for rice Hb1 and 2, and possibly for rice Hb3 and 4, are rather similar to those of other O₂-transport and-storage proteins, such as the sperm whale myoglobin and soybean Lba (Table 2). However, in rice Hb1² and 2¹⁴, and possibly in rice Hb3 and 4, the bound O₂ is stabilized by distal His after binding to the heme iron, which results in very low O₂-dissociation rate constants. The O₂-association and -dissociation rate constants of hexacoordinate rice Hb1 and 2, and possibly of rice Hb3 and 4, result in that the affinity of these proteins for O₂ is very high (Table 2). In the absence of biochemical data it becomes difficult to evaluate the O₂-binding characteristics of rice Hb5, however, its predicted pentacoordinate structure would suggest a low to moderate affinity for O₂.

Table 2. Rate and equilibrium constants for the reaction of O₂ from rice Hb1 and 2. Constants for arbitrarily selected plant and non-plant Hbs are included for comparison.

| Protein     | k\(^{-}O₂\) (μM\(^{-1}\)s\(^{-1}\)) | kO₂ (s\(^{-1}\)) | kO₂ (μM\(^{-1}\)) | Reference |
|-------------|----------------------------------|-----------------|-----------------|-----------|
| Plant Hbs   |                                  |                 |                 |           |
| nsHbs       |                                  |                 |                 |           |
| Rice Hb1    | 68                               | 0.038           | 1800            | 2         |
| Rice Hb2    | 50                               | 0.038           | 1316            | 14        |
| Barley Hb   | 2.4                              | 0.028           | 86              | 134       |
| Arabidopsis AtGLB1 | 74                         | 0.12            | 617             | 49        |
| Arabidopsis AtGLB2 | 1                           | 0.17            | 7               | 49        |
| Lotus Glb1-1 | 187                           | 0.004           | 20,250          | 111       |
| Lotus Glb1-2 | 300                           | 0.27            | 1111            | 111       |
| tHbs        |                                  |                 |                 |           |
| Arabidopsis AtGLB3 | 0.2                         | 0.3             | 0.66            | 69        |
| Symbiotic Hbs |                                |                 |                 |           |
| Soybean Lba | 130                              | 5.6             | 23              | 135       |
| Non-plant Hbs |                              |                 |                 |           |
| Sperm whale myoglobin | 14                       | 11              | 1.3             | 136       |
| Ascaris Hb   | 1.5                              | 0.004           | 375             | 137       |
| Paramphistomum Hb | 108                    | 0.033           | 3270            | 137       |
| Synecocystis tHb | 240                        | 0.011           | 21,818          | 138       |

k\(^{-}O₂\) is the O₂-association rate constant; kO₂ is the O₂-dissociation rate constant; kO₂ (k\(^{-}O₂\)/kO₂) is the O₂-affinity constant.

Postulated migration routes for gaseous ligands to the heme iron in rice Hb1

The bis-histidyl hexacoordinated form of rice Hb1 displays a hydrophobic distal cavity which appears to be connected with the external solvent through the position of Phe44 (also known as FB10 because it occupies the tenth position in helix B). It was suggested that this amino acid regulates the migration of small ligands in rice Hb1, for example in ligand binding to the heme iron, ligand migration through internal docking sites and ligand release into the external solvent. Kinetic analysis after laser flash photolysis of rice Hb1 encapsulated in silica gel combined with computational analysis revealed the existence of two channels in the rice Hb1 CO-bound species. The first channel is located in the distal region of the heme pocket and is connected with a secondary channel that is directly connected with the external solvent. Apparently, the position of FB10 in hexacoordinated rice Hb1 leaves the distal heme pocket accessible to the external solvent, however after the ligand entrance the phenyl ring rotates closing the cavity and thus hindering the exit of the bound ligand. Thus, together with distal His (see subsection Spectroscopic characteristics of rice non-symbiotic hemoglobins) and aromatic amino acids that are located in the distal region of the heme pocket, FB10 appears to regulate hexacoordination and functioning of rice Hb1.

Rice truncated hemoglobin, predicted structure and properties

Rice (O. sativa) tHb (GenBank accession number NP_001057972) is 172 amino acids in length, which corresponds to a globin domain (position 26 to 147) flanked by N- and C-terminal extensions (Figure 3B). No monocot tHb has been analyzed by x-ray crystallography, however the tertiary structure of a rice tHb was predicted using computational methods (Figure 4). The predicted structure of rice tHb is highly similar to the crystal structure of an Arabidopsis thaliana tHb. The globin domain from rice and A. thaliana tHbs folds into the 2/2-folding (see subsection on Generalities on hemoglobins). Similarly to the A. thaliana tHb structure, flanking regions to the globin domain of predicted rice tHb correspond to an N-terminal helical extension and a C-terminal unfolded extension (Figure 4). The high similarity between the crystal structure of A. thaliana tHb and the predicted structure of rice tHb suggests that the biochemical properties and function of dicot and monocot tHbs are similar.

Rice tHb has not been subjected to spectral analysis, however the predicted structure of this protein (Figure 4) is highly similar to the crystal structure of an A. thaliana tHb (see above). The absorption spectra of an A. thaliana tHb showed that heme iron from this protein is pentacoordinate. Thus, it is likely that heme iron in rice tHb is pentacoordinate and that the rate and equilibrium constants for the reaction of O₂ of rice tHb are similar to those of the Arabidopsis tHb (Table 2), i.e. the O₂-association and -dissociation rate constants are low to moderate.

Postulated functions for rice hemoglobins

While data on the localization, kinetics, regulation and structure of rice Hbs have accumulated, little work has been performed to fully understand the function of these proteins in rice organs. However,
previous work from other plant and non-plant Hbs provides data that enable us to propose potential functions for rice Hbs. Rice Hbs could potentially function within cells through O$_2$-transport and -signaling, binding to small molecules (most notably NO) and other as yet undetermined mechanisms. Here we evaluate the evidence for and against these modes of action.

Oxygen transport is a major function of many Hbs. This process requires that the kinetics of O$_2$-binding do not limit the O$_2$-diffusion process$^{47-50}$. Based on the concentration of Hb polypeptides in rice organs ($\sim$50 to 100 nM)$^{57}$, the O$_2$-association rate constant of rice Hb1 and 2 (Table 2) and possibly that of rice Hb3 to 5 and tHb (see subsections Rate and equilibrium constants for the reaction of oxygen from rice non-symbiotic hemoglobins and Rice truncated hemoglobin, predicted structure and properties), and the free O$_2$ concentration in aerated rice roots (<1.4 $\mu$M)$^{51}$, it is likely that Hbs would be substantially oxygenated in rice organs. However, the O$_2$-dissociation rate constants of rice Hb1 and 2 (Table 2), and possibly that of rice Hb3 and 4, are extremely low. These data do not support the O$_2$-transport function for rice Hb1 to 4 because these proteins would not release O$_2$ after oxygenation.

It was reported that hexacoordinate Hbs interact with either organic molecules or protein partners$^{52-54}$ and thus a possibility is that such interactions could impact the kinetic constants, particularly the O$_2$-dissociation rate constants, of hexacoordinate nsHbs$^{55}$. There have been no direct biochemical evaluations of this hypothesis in rice or in other plants, precluding definitive answers. However, their unique structural features could result in as yet undiscovered interactions.

Rice Hbs may function in O$_2$-signaling if they easily bind and release O$_2$. Appleby and co-workers$^3$ proposed that under normal conditions Hbs would be oxygenated and under O$_2$-limiting conditions the concentration of deoxyHb would increase triggering an anaerobic response. It was reported that levels of Hbs increase in rice roots from flooded plants indicating that the synthesis of rice Hbs increases under O$_2$-limiting conditions$^{56}$. Rice is a flooding resistant crop, thus under flooding (i.e. hypoxia) conditions rice Hbs could sense low O$_2$-concentrations and trigger an anaerobic metabolism for rice growth. To act as a signaling molecule, rice Hbs will need to bind directly to the DNA, to additional proteins, such as transcription factors, or catalyze some unique reactions that could potentially function within cells through O$_2$-transport and -signaling. There are reports of nuclear-localized Hbs$^{34}$, but no direct evidence for a function arising from translocation of Hbs from the cytoplasm to the nucleus currently exist.

The NO dioxygenase activity exhibited by oxygenated Hbs is well documented$^{55-57}$. NO is a hormone-like radical that modulates several aspects of the plant physiology, including plant immunity, seed germination, de-etiolation, apoptosis, stomata guard cells opening/closure and the rhizobia-legume symbiosis$^{96-109}$. Scavenging of NO is considered a function of plant Hbs$^{100,101-104}$. During this process, oxygenated plant Hbs react with NO producing nitrate and ferric Hb. Ferric plant Hbs are subsequently reduced to ferrous Hb by enzymatic$^{103,106}$ and non-enzymatic$^{107-111}$ mechanisms. This process regenerates (oxy) ferrous Hb which is able to bind NO in a cyclic pathway referred to as the Hb/NO cycle$^{104,112}$. The operation of this cycle appears to be involved in maintaining an active metabolism in the plant cells$^{105}$. Rice Hb1 exhibits NO dioxygenase activity (k$_{obs}$ NO = 90 s$^{-1}$)$^{112}$, thus a possible function of Hbs into the rice physiology is modulating levels of NO by scavenging NO. However, the inability of rice Hb1 to substitute the NO scavenger activity in a flavoHb knockout Escherichia coli$^{113}$ and the observation that levels of Hbs did not change in rice seeds germinated under nitrosative stress$^{66}$ suggest that the NO dioxygenase activity of rice Hbs is limited in vivo.

A consequence of the operation of the Hb/NO cycle could be the maintenance of cell respiration and energy status. Based on the studies on over- and under-expressing barley nsHb in maize cells, it was proposed that under hypoxic conditions barley nsHb is involved in the ATP metabolism, particularly in maintaining the energy status under O$_2$-limiting conditions$^{113}$. Immunolocalization data showed that rice Hbs are localized in differentiating cells (see subsection on Gene expression and localization of hemoglobins in rice organs)$^{47}$. The metabolism of these cells is redirected in response to differentiation signals, such as a change in the cell redox state. Rice Hbs could be involved in redox signaling if the redox state of the heme is functional$^{34}$. Thus, under these conditions rice Hbs may function by sensing or maintaining redox environments that promote specific cell metabolisms$^{54}$.

It was proposed that one of the functions of plant Hbs could be related to the peroxidase activity$^{49,51}$. This is of interest because peroxidase activity modulates the levels of reactive oxygen species and a variety of cellular processes$^{114-121}$. In plants, evaluation of the peroxidase activities of Arabidopsis Hbs (AtGLB1, AtGLB2 and AtGLB3) revealed that these proteins oxidize Amplex Red, DHR123 and guaiacol substrates$^{122}$ and overexpression of AtGLB1 increased tolerance of Arabidopsis to H$_2$O$_2$ stress$^{123}$. These observations suggested that Arabidopsis Hbs function as antioxidants. However, levels of Hb polypeptides did not change in rice seeds germinated under H$_2$O$_2$ stress$^{56}$. Also, the analysis of the peroxidase activity of rice Hb1 compared to that from horseradish peroxidase (HRP) showed that the catalytic efficiency of rice Hb1 for the oxidation of guaiacol using H$_2$O$_2$ as electron donor is several orders of magnitude lower than that of HRP (k$_c$/K$_m$ = 15.8 and 44,833 mM$^{-1}$ min$^{-1}$, respectively). Additionally, it was observed that recombinant rice Hb1 poorly protects E. coli from H$_2$O$_2$ stress$^{24}$. This evidence indicates that it is unlikely that rice Hbs function in vivo as peroxidases.

Based on gene expression (Table 1), protein localization and structural and kinetic properties of rice Hbs and data from the analysis of other plant and non-plant Hbs it is likely that Hbs play a variety of roles in rice plants growing under normal and stressed conditions. These functions may include O$_2$-transport, O$_2$-sensing, NO-scavenging and redox-signaling. Future work on rice Hbs should focus on testing the above potential functions as well as newly proposed functions that emerge from novel observations.
Evolution of rice hemoglobins

Hbs are widely distributed in land plants, ranging from primitive bryophytes to evolved angiosperms. The outline of plant Hb evolution subsequent to land colonization was clarified. Briefly, a phylogenetic analysis showed that plant and animal *hb* genes diverged 900–1400 mya, that land plant *nshb* and *thb* genes vertically evolved through different lineages from algal ancestors, that *nsHbs-1* and *nsHbs-2* are monophyletic and evolved via a gene duplication event prior to the divergence of monocots and dicots at ca. 140 mya, and that symbiotic *hbs* originated from *nshb* genes at ca. 94 mya. Likewise, the structural analysis of primitive *nsHbs* and *Lbs* revealed that changes during the evolution of *nsHbs* to *Lbs* were a hexacoordinate to pentacoordinate transition at the heme prosthetic group, a length decrease at the CD-loop and N- and C-terminal regions, and a compaction of the protein into a globular structure.

In contrast, the evolution of rice Hbs is partially understood owing to the limited availability of Hb sequences from a wide variety of wild and cultivated rice. However, the outline of monocot Hb evolution is rather well understood. Thus, in this section we will discuss the evolution of rice Hbs within the context of major events that occurred during the evolution of monocot Hbs. A major event during the evolution of land plant *nsHbs* was the duplication of an ancestral *nsHb* into *nsHb-1* and *nsHb-2* prior to the monocot-dicot divergence. Sequence analysis revealed that *nsHb-1* and *nsHb-2* genes exist in dicots and that apparently only *nsHb-1* genes exist in monocots. Subsequent phylogenetic analysis of monocot *nsHb* sequences revealed that apparently only *nsHb-1* evolved within monocots, that *nsHb-1* duplicated early in the evolution of monocots originating clade I and clade II *nsHbs* (*nsHbs-I* and *nsHbs-II*, respectively), that *nsHbs-I* correspond to dicot *nsHbs-1*, and that *nsHbs-II* diversified into regular *nsHbs-II*, post-helix H-containing *nsHbs-II* and 11 amino acids deletion-containing *nsHbs-II*.

This analysis also showed that *O. sativa* var. indica and *O. sativa* var. japonica *Hb1 to 4* and *Hb5* cluster within clade I and clade II, respectively, and that *O. glaberrima* and *O. rufipogon* (whose all *nshb* copies remain unidentified because their genome sequencing is in progress) *nsHbs* cluster within clade I. Thus, apparently clade I and clade II lineages remain conserved during the evolution of rice *nsHbs*.

Evaluation of the rate of divergence of selected land plant Hbs revealed that evolutionary rates slowed down previous to the origin of magnoliophyta and that the rate of divergence was slower in rice *Hb1* than in rice *tHb*. This observation suggested that rice *Hb1* (and conceivably other rice *nsHbs*) evolved under the effect of the stabilizing selection. However, the estimation of the variability of the *O. sativa* var. indica, *O. sativa* var. japonica, *O. glaberrima* and *O. rufipogon* *nshb* and *thb* genes revealed that in these plants variability is higher in *nshb* than in *thbs* and that these genes evolved under the effect of neutral selection. Currently the effect of rates of divergence and gene variability on the Hbs function during the rice evolution is not known.

Concluding remarks and future directions

In the preceding sections of this review we summarized major findings from the study of rice Hbs. This review also reveals some major lacunae in our ability to completely understand rice Hbs, more specifically the lack of information about the precise functions of Hbs in rice organs. The proposed functions for rice Hbs are mostly based on the analysis of other plant and non-plant Hbs. Thus, future work should evaluate the Hb activities (e.g. the NO-binding and -detoxifying activities) in either rice organs or rice cell cultures under a variety of growing conditions. Elucidating the functions of rice Hbs also requires the identification of organic molecules and protein partners that interact with rice Hbs. Other lacunae are the absence of biochemical, biophysical and cellular data on the properties of rice *Hb2* to 5 and *tHb*. Generating recombinant rice *Hb2* to 5 and *tHb* should provide Hb polypeptides for a variety of analyses that reveal the biochemical and biophysical properties of these proteins.

With the exception of rice *hb2*, a lacuna is the absence of experimental information about the cis-elements and trans-acting factors that regulate the expression of rice *hb*. This information may help to integrate the *hb* gene expression into the rice metabolisms, including those that are modulated by plant hormones.

A final lacuna is the incomplete understanding of the evolution of rice Hbs. Sequencing of the *O. glaberrima* and *O. rufipogon* genomes will be completed soon and most likely a number of rice genomes (including that of *O. barthii*, which is postulated as the ancestor of *O. glaberrima*) will be sequenced within the near future. This will provide new Hb sequences for phylogenetic analysis and the understanding of the evolution of rice Hbs, including the identification of ancestral rice Hbs and the evaluation of the effect of rice domestication and breeding during the evolution of rice Hbs.

Author contributions

RAP conceived this review and prepared the first draft of the manuscript. RAP, JFM and GS were involved in the revision of the draft manuscript and prepared the revised version from the Reviewers’ evaluation.

Competing interests

No competing interests were disclosed.

Grant information

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into the early evolution of plant non-symbiotic hemoglobins. Mol Biol Evol. 2008; 25(7): 1482–1487.

Published Abstract | Publisher Full Text

48. Smagghie BJ, Hoy JA, Percifield R, et al.: Review: correlations between oxygen affinity and sequence classifications of plant hemoglobin. Biopolymers. 2009; 91: 1083–1096.

Published Abstract | Publisher Full Text

49. Trevaskis B, Watts RA, Andersson SR, et al.: Two hemoglobin genes in Arabidopsis thaliana: the evolutionary origins of leghemoglobins. Proc Natl Acad Sci U S A. 1997; 94(22): 12230–12234.

Published Abstract | Publisher Full Text | Free Full Text

50. Radford AE: Fundamentals of plant systematics. New York, NY: Harper & Row, Inc. 1986; 498.

Reference Source

51. Rodríguez-Alonso G, Arredondo-Peter R: Expression and evolution of functionally distinct haemoglobin genes in plants. (PubMed Abstract)

52. Rodríguez-Alonso G, Arredondo-Peter R: Phylogenetic analysis reveals an apparent duplication of the non-symbiotic hemoglobin 1 gene early in the evolution of monocotyledonous plants. ScienceJ. 2012; 1: 27.

Reference Source

53. Antonini E, Rossi-Bernardi L, Chiancone E: editors. Hemoglobins and myoglobin in their reactions with ligands. Amsterdam, London: North-Holland Pub. Co. 1971; 21: 457.

Reference Source

54. Yu J, Hu S, Wang J, et al: A draft sequence of the rice genome (Oryza sativa L. ssp. indica). Science. 2002; 296(5565): 79–92.

Published Abstract | Publisher Full Text

55. Vinogradov SN, Fernandez I, Hoogwijk D, et al.: Phylogenetic relationships of 3/3 and 2/2 hemoglobins in Archaeplastida genomes to bacterial and other eukaryote hemoglobins. Mol Plant. 2011; 4(1): 42–58.

Published Abstract | Publisher Full Text

56. Larrañaga V, Rosa E, Sarah G, et al: Mapping and analysis of a hemoglobin gene family from rice (Oryza sativa). Plant Physiol Biochem. 2002; 40: 199–202.

Reference Source

57. Ross EJ, Stone JM, Elowsky CG, et al.: Activation of the Oryza sativa non-symbiotic haemoglobin-2 promoter by the cytokinin-regulated transcription factor, ARR1. J Exp Bot. 2004; 55(403): 1721–1731.

Published Abstract | Publisher Full Text

58. Larrañaga V, Ruiz-Kubili M, Arredondo-Peter R: Expression of non-symbiotic hemoglobin 1 and 2 genes in rice (Oryza sativa) embryonic organs. Comm Integr Biol. 2011; 4(4): 457–458.

Published Abstract | Publisher Full Text

59. Larrañaga V, Sarah G, Kuciaus RV, et al.: Synthesis of hemoglobins in rice (Oryza sativa var. Jackson) plants growing in normal and stress conditions. Plant Sci. 2001; 161(2): 279–287.

Published Abstract | Publisher Full Text

60. Ioantescu AI, DeWilde S, Kiger L, et al.: Characterization of non-symbiotic tomato hemoglobin. Bioch J. 2005; 384(3): 2628–2639.

Published Abstract | Publisher Full Text | Free Full Text

61. Watts RA, Hunt PW, Hvitved AN, et al.: A hemoglobin from plants homologous to truncated hemoglobins of microorganisms. Proc Natl Acad Sci U S A. 2001; 98(18): 10119–10124.

Published Abstract | Publisher Full Text | Free Full Text

62. Ohashi K, Kawai-Chibayashi M, Wakisaka K, et al.: Induction of class-1 non-symbiotic hemoglobin genes by nitrate, nitrite and nitric oxide in cultured rice cells. Plant Cell Physiol. 2005; 46(3): 324–331.

Published Abstract | Publisher Full Text

63. Hargrove M, Brucker EA, Stec B, et al.: Crystal structure of a nonsymbiotic plant hemoglobin. Structure. 2000; 8(9): 1005–1014.

Published Abstract | Publisher Full Text

64. Goodman MD, Hargrove MS: Quaternary structure of rice nonsymbiotic hemoglobin. J Biol Chem. 2001; 276(9): 6834–6839.

Published Abstract | Publisher Full Text

65. Gopalasubramaniam SK, Ganoch-Villegas V, Gustos BG, et al.: Use of in silico (computer) methods to predict and analyze the tertiary structure of plant hemoglobins. Meth Enzymol. 2008; 438: 393–410.

Published Abstract | Publisher Full Text

66. Ishimura T, Kukuchi T: Analysis of the differences in the folding kinetics of structurally homologous proteins based on predictions of the gross features of residue contacts. Proteins: Struct Funct Biol. 2003; 51(4): 515–530.

Published Abstract | Publisher Full Text

67. Kukuchi T, Némethy G, Scheraga HA: Prediction of the location of structural domains in globular proteins. J Prot Chem. 1988; 7(4): 427–471.

Published Abstract

68. Navajna S, Alzayed-Saleh E, Kukuchi T, et al.: Prediction of folding pathway and kinetics among plant hemoglobins using an average distance map method. Proteins: Struct Funct Biol. 2005; 61(3): 500–506.

Published Abstract | Publisher Full Text

69. Appleby CA, Bergersen FJ: Preparation and experimental use of leghemoglobin. In: Bergersen FJ, editor. Methods for Evaluating Biological Nitrogen Fixation. Chichester: Wiley; 1980: 315–335.

Published Abstract

70. Olson JS: Stopped-flow, rapid mixing measurements of ligand binding to hemoglobin and red cells. Meth Enzymol. 1981; 7B: 631–652.

Published Abstract

71. Appleby CA: Leghemoglobin. In: Quispel A, editor. The Biology of Nitrogen Fixation. New York: American Elsevier Publishing Co. 1974: 521–554.

Published Abstract | Publisher Full Text

72. Arredondo-Peter R, Moran JF, Sarah G, et al.: Molecular cloning of the cowpea leghemoglobin II gene and expression of its cDNA in Escherichia coli. Purification and characterization of the recombinant protein. Plant Physiol. 1997; 114(2): 493–500.

Published Abstract | Publisher Full Text | Free Full Text

73. Sarmah BJ, Kundu S, Hoy JA, et al.: Role of phenylalanine B10 in plant nonsymbiotic hemoglobins. Biochemistry. 2006; 45(32): 9735–9745.

Published Abstract | Publisher Full Text

74. Spyrakis F, Lusque FJ, Vippiani C: Structural analysis in nonsymbiotic hemoglobins: what can we learn from inner cavities? Plant Sci. 2011; 181(1): 8–13.

Published Abstract

75. Bacht KN, Abruzzetti S, Uppal S, et al.: Ligand migration and hexacoordination in type 1 non-symbiotic rice hemoglobin. Biochim Biophys Acta. 2011; 1814(8): 1042–1053.

Published Abstract | Publisher Full Text

76. Reeder BJ, Hough MA: The structure of a class 3 nonsymbiotic plant haemoglobin from Arabidopsis thaliana reveals a novel N-terminal helical extension. Acta Crystallogr D Biol Crystallogr. 2014; 70(Pt 5): e141–e148.

Published Abstract | Publisher Full Text | Free Full Text

77. Wittenberg JB: Myoglobin-facilitated oxygen diffusion: role of myoglobin in oxygen entry into muscle. Physiol Rev. 1970; 50(4): 559–636.

Published Abstract

78. Wittenberg JB, Wittenberg BA: Mechanism of cytoplasmic hemoglobin and myoglobin function. Annu Rev Biophys Chem. 1990; 19: 217–241.

Published Abstract | Publisher Full Text

79. Wittenberg JB: Facilitated oxygen diffusion. The role of leghemoglobin in nitrogen fixation by bacteroids isolated from soybean root nodules. J Biol Chem. 1974; 249(13): 4057–4066.

Published Abstract

80. Wittenberg JB: On optima: the case of myoglobin-facilitated oxygen diffusion. Gene. 2007; 385(1–2): 156–161.

Published Abstract | Publisher Full Text

81. Armstrong W, Gaynard T: The critical oxygen pressure for respiration in intact plants. Plant Physiol. 1976; 37(3): 200–206.

Published Full Text

82. Wakisaka K, Nakano T, Kitatsuji C, et al.: Human neuroglobin interacts with flavilin-1, a lipid raft microdomain-associated protein. Biochem Biophys Res Comm. 2004; 318(2): 450–460.

Published Abstract | Publisher Full Text

83. Sainz-Rivera J, Sarah G, Arredondo-Peter R: Modeling the tertiary structure of a maize (Zea mays ssp. mays) nonsymbiotic hemoglobin. Plant Physiol Biochem. 2004; 42(11): 891–897.

Published Abstract

84. Sengoku C, Mustardy L, Ayydin F, et al.: Nuclear localization of a hypoxia-inducible novel non-symbiotic hemoglobin in cultured alfalfa cells. FEBS Lett.
Role of active oxygen species and NO in plant defence responses.
Open Peer Review

Current Referee Status: ✔️ ✔️

Version 2

Referee Report 19 December 2014

doi:10.5256/f1000research.6325.r7011

Juliette T. J. Lecomte, Eric Johnson
T. C. Jenkins Department of Biophysics, Johns Hopkins University, Baltimore, MD, USA

We appreciate the authors consideration of our comments and we agree with the revisions to this paper. This review effectively highlights the current state of knowledge for this field and points out additional research that would make valuable contributions.

We have read this submission. We believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

Version 1

Referee Report 20 November 2014

doi:10.5256/f1000research.5905.r6522

Juliette T. J. Lecomte, Eric Johnson
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The manuscript "Rice (Oryza) hemoglobins" by Arredondo-Peter, Moran and Sarath reviews the current status of research on the group of proteins found within rice plants belonging to the hemoglobin (Hb) superfamily. There is growing interest in Hbs, particularly non-mammalian Hbs, owing to their potential role as mediators of reactive nitrogen molecules, such as nitric oxide. Such a review is welcome, as rice is an important cultivar and adaptation to stress during growth plays an important role in the survival of the plant. The review covers a wide range of relevant topics, such as phylogenetic aspects, cellular localization, and chemical properties.

Within this review, the literature on rice Hbs is covered thoroughly, as witnessed by the extensive bibliography that accompanies the manuscript. However, closer inspection of that bibliography reveals an underlying issue with this manuscript. Although over 150 citations are given, a minority of those papers reference research performed specifically on rice plant or rice Hbs. This is not meant as a slight against
the authors, rather as an observation that current research on rice Hbs is still in its infancy and extensive literature on the topic simply does not exist. With that in mind we would suggest altering this manuscript to acknowledge this shortage of experimental evidence in the following ways.

- By far the majority of experimental evidence exists of Hb1, yet throughout the manuscript Hb1 is treated more-or-less equally to Hb2-Hb5. The authors should focus on Hb1 in presenting the available evidence, then in a separate section (perhaps “correlations to other Hb genes”) the authors could examine homology models and hypothetical behavior without the danger of having conjecture misinterpreted by the reader as published fact.

- tHb belongs to a separate class of Hbs than the other Hbs in rice (Hb1 through Hb5), and throughout the manuscript it is often treated as an afterthought. As an addendum to the above comment we would also suggest a separate section be reserved for tHb. Though little experiment information may be available for this protein, a short stand-alone section would be more informative than a series of trailing paragraphs.

- Great care must be taken by the authors when postulating biophysical characteristics of rice Hbs using only homology models based upon a single crystal structure. Simple variations in structure can carry significant changes in heme iron coordination, reactivity and binding affinity. If such conjecture is used in this manuscript it must be plainly stated with the caveat that this is not published fact, or experimental evidence.

Additional specific comments

1. To orient the reader it would be helpful to include a two-panel figure containing (a) the amino acid sequence alignment of Hbs 1 through 5 using Hb1 as the anchor, and (b) the alignment of tHb using Arabidopsis thaliana GLB3 as the anchor. In both panels, the differences between each Hb and Hb1 (a) or tHb and GLB3 (b) could be emphasized (e.g., with color). The secondary structure of Hb1 (a) and GLB3 (b) could be indicated as well. In the text the % sequence identity for the relevant pairs should be mentioned. Whether one or the other cultivar is used should be clearly indicated throughout the review (for example, in Figure 2).

2. Apoprotein folding pathways are discussed on page 7. An additional statement as to the biological relevance of the folding pathway and information about association with the heme (when, where) would be interesting.

3. In general, it is not possible to predict the thermodynamic or chemical properties of a heme protein based on its primary structure. The case of Parasponia andersonii and Trema tomentosa hemoglobins provides one illustration of the difficulty within the plant world. Predictions can be inaccurate even when a three-dimensional structure is available. This shortcoming of sequence analyses and modeling could be emphasized with a discussion of specific examples and used to advocate the need for additional hemoglobin research.

4. Reaction with nitric oxide is a likely function of many hemoglobins. According to the work of Gardner and colleagues, the NO dioxygenase reaction begins with the binding of dioxygen followed by combination with NO to produce nitrate. In this mechanism, binding of NO to the iron is not necessary. In this regard, the statement on page 3 (bottom left) should be clarified. Likewise,
the Hb/NO cycle as proposed by Igamberdiev and Hill involves oxyHb. In contrast, the description on page 9 suggests that one turnover occurs with oxyHb, followed by formation of Hb-NO.

5. Speculations regarding the formation of homodimeric or heterodimeric structures should also be qualified, since the concentration of the hemoglobins (sub micromolar) appears to be much lower than the projected Kd (mM).

6. In the "Postulated functions" section, a proposal is made that Hb5 and tHb are O₂ transporters. Could the authors elaborate on this function? (Transport to what and for what purpose, and is it consistent with the cellular concentrations?)

7. Ligand binding is central to the function of hemoglobins. The "kinetic" section provides little such information while the second paragraph of the “Postulated functions” section offers numbers. It would be useful to consolidate the kinetic data with a table containing the measured equilibrium and rate constants for Hb1 and Hb2 (CO, NO, O₂), as published in various primary references (for example, reference 97), and for Hb relatives mentioned in the text.

We have read this submission. We believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however we have significant reservations, as outlined above.

Competing Interests: No competing interests were disclosed.

Author Response 29 Nov 2014

Raul Arredondo-Peter, Universidad Autonoma del Estado de Morelos, Mexico

We thank Drs. Lecomte and Johnson for evaluating the first version of this review and for providing useful comments and suggestions. We agree with them and incorporated the suggested changes into the revised version of the review.

Competing Interests: No competing interests were disclosed.

Referee Report 13 November 2014

doi:10.5256/f1000research.5905.r6520

Martino Bolognesi
Department of Biosciences, University of Milan, Milan, Italy

The Review by Arredondo-Peter et al. presents our current knowledge on the fascinating field of plant non-symbiotic (nsHb) and truncated hemoglobins (tHb), for which substantial, but scattered, information has accumulated over the past twenty years. The Review deals specifically with rice Hbs. Work from several distinct worldwide groups has so far provided information on the gene families for five nsHbs, and for a single tHb in rice; moreover, expressed proteins have been located to various plant organs and developmental stages. Crystal structures and kinetic analyses have helped delineating the potential roles of rice Hbs in plant physiology, highlighting different O₂ binding affinities that differentiate the various Hbs. A main question that remains unanswered concerns the in vivo functions carried over by the five distinct
rice nsHbs and tHb; hints reviewed from the literature include O₂ sensing, signaling, O₂ transport, NO scavenging, and NO dioxygenase pseudo-enzymatic activities (others may also be plausible). The Review includes evolutionary considerations (and the effects of rice selection through domestication) that will be reinforced by the forthcoming completion of rice genomes.

Overall, the Review provides a useful compendium over a subject whose reunification into a coherent presentation will indeed support deeper exploration of the functional aspects, whose scientific and practical relevance cannot be underestimated.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

**Competing Interests:** No competing interests were disclosed.

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Author Response 16 Nov 2014

**Raul Arredondo-Peter,** Universidad Autonoma del Estado de Morelos, Mexico

We thank Dr. Bolognesi for evaluating this review and his comments.

**Competing Interests:** No competing interests were disclosed.