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Gut Microbiota Features Associated With *Campylobacter* Burden and Postnatal Linear Growth Deficits in a Peruvian Birth Cohort

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(See the Major Article by Rouhani et al on pages 989–99 and the Editorial Commentary by Colin Stine on pages 1008–9.)

**Background.** *Campylobacter* infection is associated with impaired growth of children, even in the absence of symptoms. To examine the underlying mechanisms, we evaluated associations between *Campylobacter* infection, linear growth, and fecal microbial community features in a prospective birth cohort of 271 children with a high burden of diarrhea and stunting in the Amazonian lowlands of Peru.

**Methods.** *Campylobacter* was identified using a broadly reactive, genus-specific enzyme-linked immunosorbent assay. 16S rRNA-based analyses were used to identify bacterial taxa in fecal samples at ages 6, 12, 18, and 24 months (N = 928). Associations between infection, growth, and gut microbial community composition were investigated using multiple linear regression adjusting for within-child correlations, age, and breastfeeding. Indicator species analyses identified taxa specifically associated with *Campylobacter* burden.

**Results.** Ninety-three percent (251) of children had *Campylobacter* present in asymptomatic fecal samples during the follow-up period. A 10% increase in the proportion of stools infected was associated with mean reductions of 0.02 length-for-age z scores (LAZ) at 3, 6, and 9 months thereafter (P < .01). We identified 13 bacterial taxa indicative of cumulative *Campylobacter* burden and 14 taxa significantly associated with high or low burden of enteric aggregative *Escherichia coli*, norovirus, or *Giardia*.

**Conclusions.** *Campylobacter* infection is common in this cohort and associated with changes in microbial community composition. These results support the notion that disruptions to the fecal microbiota may help explain the observed effects of asymptomatic infections on growth in early life.

**Keywords.** *Campylobacter*; enteropathy; microbiota; child growth.

Linear growth faltering, or stunting, affects more than 160 million children annually and underlies a large proportion of childhood mortality and disability worldwide [1]. In low- and middle-income countries (LMICs), interactions between enteric infections, undernutrition, and diarrhea contribute to stunting [2, 3]. Evidence suggests that chronic or repeated enteropathogen infections without overt symptoms lead to intestinal inflammation, impaired gut barrier function, and blunted immune responses that, in turn, drive linear growth failure in the absence of diarrhea [4–8]. This may partly account for the persistence of stunting in areas where programs to prevent and treat clinical diarrhea and provide nutritional supplementation are well implemented [9–11].

*Campylobacter* may be an example of how enteropathogens can impair growth and immunity independently of clinical diarrhea [12]. Extensive literature links *Campylobacter* to Guillain-Barré syndrome, reactive arthritis, and irritable bowel diseases, demonstrating that infections can impact immunological tolerance and gut function beyond cessation of acute symptoms [13–18]. Data from large, multisite cohorts in LMICs reveal associations between *Campylobacter*, inflammation, gut permeability, and impaired linear growth [12, 19, 20], and studies have shown increased presence of *Campylobacteraceae* in the gut among undernourished children [21]. These observations are especially relevant given recent estimates of higher asymptomatic carriage than previously appreciated [22, 23].

In the present study, we test the hypothesis that *Campylobacter* is associated with specific changes in the developing gut microbiota. The rationale for exploring this issue comes from the increasing appreciation of the impact of *Campylobacter* on the intestinal environment and recent observations linking the gut microbiota to child growth. Birth cohorts have shown that assembly of the gut microbial community during early postnatal life exhibits shared features across individuals and geographic...
regions [24] and that this assemblage is critical for maturation of the gut mucosal immune system, barrier integrity, and other features of adaptive and innate immunity [25–27]. Moreover, studies have demonstrated impaired development of the gut microbiota in undernourished children in LMICs [28] and have used mouse models to provide evidence for a causal relationship between microbial composition and growth faltering [29].

In the current study, we use data from a birth cohort of 271 children to describe temporally structured associations between Campylobacter infection, fecal microbial diversity and composition, and linear growth. In our companion article presented in this issue of Clinical Infectious Diseases, we report that all-cause diarrhea provokes enduring changes to the gut microbial environment that, in turn, may impact subsequent growth acquisition and risk of illness. Here, we complement this analysis by determining whether asymptomatic Campylobacter infections incur independent effects on the gut microbiota in a manner that impacts intestinal integrity and child growth.

**METHODS**

Study Design

This study was conducted as part of the Etiology, Risk Factors and Interactions of Enteric Infections and Malnutrition and the Consequences for Child Health and Development Study, a birth cohort exploring enteropathy, nutrition, and child growth in 8 countries with high burdens of diarrhea and undernutrition. This study was conducted in the Amazonian lowlands near Iquitos, Peru. Details of demographic and biomedical data collection methods have been published previously [30–32]. Children were enrolled within 17 days of birth and followed for 24 months (n = 271). For surveillance of asymptomatic enteric infections, children contributed fecal samples and weight and length were measured monthly. Fieldworkers visited homes twice-weekly to record histories of illness, antibiotic use, breastfeeding, and dietary intake; additional specimens were collected during diarrheal episodes. Routine fecal samples obtained at postnatal months 6, 12, 18, and 24 were retrospectively selected for profiling of gut bacterial community composition. In total, 146 boys and 125 girls contributed 6096 months of observations, 6011 surveillance fecal samples, and 2440 diarrheal samples to the study.

Diagnostics and Microbial Analyses

Swabs from fecal samples were placed into Cary-Blair transport medium and processed for culture on Campylobacter Agar base with Blaser’s supplement (Beckton Dickinson, Sparks, MD). Cultures were incubated at 42°C for 48 hours under microaerophilic conditions (5% oxygen, 10% carbon dioxide, and 85% nitrogen). Gram-negative colonies were tested for oxidase, catalase, and hippurate hydrolysis. Hippurate-positive Campylobacter strains were characterized as Campylobacter jejuni and hippurate-negative isolates as other Campylobacter species. Campylobacter testing directly on fecal samples stored at −70°C was done using a broadly reactive genus-specific ProSpecT enzyme-linked immunosorbent assay (ELISA) [33]. Details of diagnostic assays for >40 other enteropathogens are described elsewhere [34].

Procedures for isolating DNA frozen fecal samples, polymerase chain reaction–based amplification, and sequencing of the V4 region of bacterial 16S rDNA genes are published elsewhere [28, 29]. DNA sequences were oriented, trimmed to remove primer sequences, and paired using bbtools (37.02; https://sourceforge.net/projects/bbmap/). DADA2 (1.8.0) was used to remove chimeric sequences and identify and quantify amplicon sequence variants (ASVs) [35]. Taxonomic assignments were made using the RDP Naive Bayesian Classifier algorithm and the GreenGenes (13.8) training set [36]. Identified ASVs with taxonomic assignments are listed in Supplementary Table 1. ASVs were used to construct a neighbor-joining phylogenetic tree in phangorn (2.4.0) [37]. Metrics of community diversity and richness were estimated using the phyloseq [38] and picante (1.7) [39] packages. Three extreme outliers (>4 standard deviations for richness) and 2 samples taken during “exclusive breastfeeding” were removed before analysis.

**Definition of Covariates**

Diarrhea was defined as ≥3 loose stools in a 24-hour period, with distinct episodes separated by 2 diarrhea-free days. Severity was measured using a community diarrheal assessment tool [40]. Asymptomatic infection was defined as detection of pathogens in surveillance stools in the absence of diarrhea or fever, and “pathogen pressure” was calculated as the mean number of enteropathogens per stool. Standard anthropometric and breastfeeding categories were based on World Health Organization (WHO) definitions [41, 42] and frequency of exposure to breastmilk was calculated as the cumulative mean number of feeds in the previous 24-hour period. Dietary diversity was measured as the number of WHO food groups (0–7) to which children were exposed [41].

We considered the number of distinct Campylobacter-positive diarrheal episodes and the cumulative proportion of surveillance stools with Campylobacter detected per child as exposure variables denoting symptomatic and asymptomatic infection, respectively. Diarrheal duration was calculated as the cumulative mean number of days per episode. The cumulative proportion of positive surveillance stools was divided into quartiles to compare the microbiota of children with the highest and lowest burdens. Persistent carriage was defined as 3 consecutive asymptomatic infections from monthly stools. We assessed gut bacterial diversity using Shannon’s and Simpson’s diversity indices; bacterial richness was assessed using the Chao1 index and phylogenetic diversity (see companion article for more details). We identified ASVs associated with high and low infection burden for Campylobacter and other enteropathogens with
high prevalence in this population: *Giardia*, entero-aggregative *Escherichia coli* (EAEC), and norovirus.

**Statistical Analyses**

To assess relationships between *Campylobacter* burden and specific ASVs, we conducted indicator species analysis (ISA). ISA identifies taxa indicative of particular habitats by calculating indicator species values for each ASV in each habitat [43], defined here as categorical groups of children with high or low enteropathogen burden. Each value is the product of the proportion of samples in 1 “habitat” (high or low burden) in which the ASV is detected and its mean relative abundance in that habitat, normalized by the sum of mean relative abundances across all habitat types. Indicator species values are bounded by 0 and 1, with 0 indicating the absence of an ASV from 1 group, and 1 indicating that an ASV occurs in every member of a group and only that group. Hypothesis tests were performed by permutation tests [43].

ISA was performed separately for each age category (6, 12, 18, and 24 months), comparing samples from children in low- and high-burden groups for *Campylobacter*, norovirus, EAEC, and *Giardia*. Within each ISA, P values were adjusted using the Benjamini-Hochberg method to control false discovery rates. Only ASVs found in at least 20% of the children in at least 1 enteropathogen burden group and having a mean percent abundance of 0.1% when present were included.

Associations between *Campylobacter*, length-for-age Z-scores (LAZ), bacterial diversity and richness, and indicator ASVs were investigated using linear regression with generalized estimating equations and linear mixed-effects models to account for longitudinal sampling. We modeled asymptomatic and symptomatic *Campylobacter* infections as predictors of LAZ at the current time point and 3, 6, and 9 months thereafter; *Campylobacter* infections and diarrheal frequency, severity, and duration as predictors of intestinal diversity and richness; and the presence and abundance of indicator ASVs on LAZ at the time of sampling and 1 and 3 months thereafter. All models were adjusted for age, breastfeeding frequency and category, pathogen pressure, dietary diversity, days using antibiotics, and diarrheal frequency. Models of diversity metrics were also adjusted for the number of sequencing reads assigned to ASVs in order to control for differences in depth of sequencing. Models with “current” or “future” LAZ outcomes were additionally adjusted for LAZ at birth or current time point, respectively.

**RESULTS**

*Campylobacter* and Child Growth

*Campylobacter* was detected by ELISA in 912 (22%) of surveillance stools and 758 (31%) of diarrheal specimens (Figure 1, Table 1). Only 22% (n = 364) of ELISA-positive samples were positive by culture-based methods, and this was consistent among surveillance (n = 197; 22%) and diarrheal samples (n = 167; 22%). *Campylobacter jejuni* accounted for nearly half of culture-positive diarrheal (45%) and asymptomatic fecal specimens (48%). By age 2 years, *Campylobacter* had been detected in ≥1 sample from each of 251 (93%) asymptomatic children. Mean time to first infection was 7.8 months, and detection increased with each month of age in the first year (odds ratio [OR], 1.21; P < .001). Fifty-five children (20.3%) experienced persistent infections. Of 260 children experiencing diarrhea, 221 (85%) had >1 *Campylobacter*-positive episode. The majority (n = 169; 64%) had *Campylobacter*-positive surveillance stools prior to clinical symptoms. Mean age of first *Campylobacter*-diarrhea was 9.3 months (Figure 1), with a mean of 2.6 episodes during the study. Twenty-seven percent of episodes were treated with antibiotics. Children who were exclusively or predominantly breastfed had significantly reduced odds of *Campylobacter*-diarrhea (OR, 0.39; P < .001) and asymptomatic detections (OR, 0.48; P < .001) after accounting for age.

By 24 months, 181 (66.8%) of children were stunted (LAZ < −2), and 22.1% were severely stunted (LAZ < −3) on ≥1 visit. The point prevalence of stunting at 24 months was 40%. *Campylobacter* infections were negatively associated with linear growth. An increase of 10% in the proportion of surveillance stools with *Campylobacter* was associated with a 0.02 reduction in LAZ at 3, 6, and 9 months thereafter (β = −0.02; P < .01 across all time points). Controlling for LAZ at birth, each episode of *Campylobacter*-diarrhea was associated with a reduction of 0.03 in current LAZ (95% confidence interval [CI], −0.04 to −0.01; P = .002). *Campylobacter*-positive diarrheal episodes were not predictive of subsequent LAZ scores.

*Campylobacter* and the Gut Microbiota

There were no significant associations between fecal bacterial diversity and richness and frequency, duration, or severity of *Campylobacter*-positive diarrheal separately from all-cause diarrheal. In contrast, asymptomatic *Campylobacter* detections were
significantly positively associated with all bacterial diversity and richness measures (Table 2).

Thirteen ASVs were indicators of either high or low Campylobacter burden at 6, 12, or 24 months (Figure 2, Table 3, Supplementary Table 2). Nine of these ASVs were indicative of high Campylobacter burden, including Ruminococcus toraus (ASV23) at month 6, a member of the genus Dialister (ASV26) at month 12, and ASVs assigned to Prevotella (ASV204 and ASV275), Succinivibrio (ASV52), Catenibacterium (ASV57), Phascolarctobacterium (ASV254), Coriobacteriaceae (ASV304), and Eubacterium biforme (ASV171) at 24 months. Four ASVs were indicative of low Campylobacter burden at month 24: Bacteroides ovatus (ASV40), Ruminococcus toraus (ASV242), members of Bacteroides (ASV27), and Lachnospiraceae (ASV39).

Several additional ASVs had indicator species values with adjusted \( P < .1 \), including 1 assigned to Streptococcus (ASV69; Supplementary Table 2), which was marginally indicative of low burden of Campylobacter at 6 months. A query of the GenBank 16S rDNA databank revealed that this ASV shares 100% sequence identity with Streptococcus lactarius, which has been isolated from the breast milk of healthy mothers [44].

The most abundant ASV assigned to the genus Bifidobacterium (ASV1) had a marginally significant association with low Campylobacter burden at 6 months (Supplementary Table 1). This ASV was detected in 97%–98% of the children in both high and low Campylobacter burden groups; its moderate association with low Campylobacter burden reflects higher relative abundance in that group. This ASV shares perfect sequence identity with published 16S rDNA reference sequences of constituents of the early developing gut microbiota during breastfeeding (Bifidobacterium longum subsp. infantis, B. longum, Bifidobacterium breve [24]).

Comparing the indicator species of high and low burden for Campylobacter with those for other enteropathogens revealed remarkably little overlap. Indicator species were not shared across high- and low-burden groups of the 4 enteropathogens assessed, with few exceptions. At age 6 months, R. toraus (ASV23) was a high-burden indicator for Campylobacter, EAEC, and norovirus; Morganella (ASV141) was an indicator for high burden of EAEC and norovirus (but not Campylobacter; Figure 2, Table 3); and the

**Table 1. Campylobacter Infections Among 271 Children Aged 0–24 Months in the Peruvian Birth Cohort**

| Campylobacter in the Population | Age, months |
|--------------------------------|-------------|
| % of children (n) ever infected | 6  | 12  | 18  | 24  |
| % of children (n) experiencing Campylobacter-positive diarrhea | 46.2 (115) | 82.3 (181) | 91.6 (185) | 94.7 (178) |
| Mean (range) of Campylobacter-positive diarrheal episodes by enzyme-linked immunosorbent assay | 0.26 (0–4) | 0.97 (0–4) | 1.9 (0–11) | 2.6 (0–15) |
| Mean (SD) proportion of surveillance stools per child with Campylobacter detected | 12.6 (17.4) | 19.2 (16.9) | 21.3 (15.7) | 22.5 (15.6) |
| Mean (SD) proportion of diarrheal samples collected from each child with Campylobacter detected | 20.5 (31.4) | 30.3 (21.9) | 32.6 (25.5) | 33.2 (24.4) |

Abbreviation: SD, standard deviation.

**Table 2. Evidence of Independent Associations Between Campylobacter Infections and Community Bacterial Diversity and Richness**

| Predictor Variable | Mean Change in Gut Microbial Diversity—β (95% Confidence Interval; Backtransformed β) |
|--------------------|----------------------------------------------------------------------------------|
| Asymptomatic infection (per 10% increase in proportion of Campylobacter-positive fecal samples) | 0.048 (.023 to .072); 0.050*** 0.069 (.031 to .107); 0.035 (.016 to .053); 3.511*** 0.041 (.016 to .066); 0.323** |
| No. of episodes of Campylobacter-positive diarrhea | −0.012 (.042 to .017); −0.013 0.027 (.07 to .15); −0.001 (.022 to .02); −0.07 .011 (.024 to .048); 0.088 |
| Mean severity of Campylobacter-positive diarrhea | −0.016 (.054 to .022); −0.015 0.017 (.062 to .029); −0.003 (.027 to .022); −0.278 .009 (.051 to .033); −0.088 |
| Mean duration of Campylobacter-positive diarrhea | −0.025 (.056 to .006); −0.023 0.022 (.06 to .17); −0.008 (.029 to .012); −0.828 .025 (.058 to .008); −0.189 |

Models were adjusted for age, breastfeeding frequency and category, pathogen pressure excluding Campylobacter, dietary diversity, antibiotic exposure, and the number of amplicon sequence variant-assignable DNA reads per sample. Coefficients for Shannon’s diversity and phylogenetic diversity are presented as the change in standard deviations and then backtransformed. For the Chao1 index, coefficients are in log-transformed units, and the backtransformed units represent percent change.

*Model adjusted for the proportion of surveillance stools infected with other enteropathogens.

*Model adjusted for the number of Campylobacter-negative diarrheal episodes experienced.

*Model adjusted for the mean severity.

*Model adjusted for the duration of Campylobacter-negative diarrheal episodes to assess whether Campylobacter infection and diarrhea were independently associated with microbial diversity metrics.

**, \( P < .01 \); ***, \( P < .001 \).
Streptococcus ASV69 marginally associated with Campylobacter was also marginally associated with norovirus and EAEC.

Overall, 26 ASVs had significant indicator species values for low or high burden of at least 1 enteropathogen. As shown in Figure 2, despite the low degree of sharing of particular indicator ASVs, groups of ASVs behave similarly across enteropathogen burden and age groups. Hierarchical clustering divides these organisms into 2 major clusters, 1 predominated by ASVs indicative of high enteropathogen burden at 18 and 24 months and a second composed of ASVs indicative of high enteropathogen burden at 6 months and low burden thereafter. Moreover, differences between indicator species values reveal correlations within each age bin, suggesting that several ASVs correlated with high or low burden for multiple enteropathogens, even if their indicator species values were only statistically significant for 1. For example, at 6 months, the differences between indicator species values for high and low Campylobacter burden are correlated with those for norovirus ($r = 0.67$, $P = .001$), Giardia ($r = 0.60$, $P = .005$), and EAEC ($r = 0.84$, $P < .001$). At 24 months, the differences in indicator species values for high and low Campylobacter burden groups were correlated with those for norovirus ($r = 0.55$, $P = .004$) and Giardia ($r = 0.80$, $P < .001$) but not for EAEC ($r = 0.06$, $P = .759$).

The presence and abundance of each ASV listed in Table 3 were modeled as predictors of LAZ at the time of sampling and 1 month and 3 months thereafter across the entire 2-year sampling period, as well as separately at 6, 12, 18, and 24 months. After correction for false discovery rates, no single indicator ASV exhibited a statistically significant association with LAZ. However, in a linear model adjusting for LAZ at birth and sequencing depth, the aggregate number of ASVs indicative of high Campylobacter burden (Figure 2) was significantly associated with LAZ; specifically, each additional indicator of high Campylobacter burden detected in a sample at 24 months was associated with a reduction of 0.08 in LAZ score ($P = .021$).

**DISCUSSION**

This study provides evidence for the hypothesis that asymptomatic Campylobacter burden is associated with changes to the gut microbial community that, in turn, may impact child growth in a setting of high enteric disease burden and stunting. Our concurrent work in the same Peruvian cohort demonstrates that all-cause diarrhea was associated with reductions in bacterial diversity and richness, that children with the most severe stunting profiles experience the greatest perturbations and longest recovery times following an episode of diarrhea, and...
In the present study, we provide evidence that Campylobacter burden is associated with linear growth faltering and changes to the gut microbial community in childhood in the absence of diarrhea. These observations suggest that alterations to gut microbiota associated with Campylobacter are independent from diarrhea-related purging and accelerated transit of upper gut flora. These results lend further evidence to the emerging hypothesis that asymptomatic Campylobacter infections contribute to childhood growth faltering in LMICs, possibly through alterations to the developing gut microbiota.

The development of the human gut microbiota in the first 2 years of life follows a path defined by several shared features. Increasingly, deviations from this normal developmental program have been associated with childhood growth deficits. Our observation that asymptomatic Campylobacter infections are associated with increased diversity among children aged 0–2 years, independent of other enteropathogens, may reflect a disruption of that developmental process. Indeed, breast milk exerts a suppressive effect on diversity in the infant gut, and limited diversity in early life may denote a healthy predominance of bacteria that metabolize breast milk and confer resilience to external insult [45–48]. This is consistent with the protective effects of exclusive or predominant breastfeeding against Campylobacter reported in this cohort [12]. Here, we also observed associations between low Campylobacter burden and a Streptococcus ASV that is found in breast milk, together with an ASV comprising several Bifidobacteria, including B. infantis. Bifidobacterium infantis contains

### Table 3. Bacterial Amplicon Sequence Variants Significantly Associated With Low and High Asymptomatic Enteropathogen Burden at Aged 6, 12, 18, and 24 Months

| Age (months) | Amplicon Sequence Variant Number | Taxonomy           | Enteropathogen | High | Low | Frequency of Detection | Adjusted P Value |
|--------------|---------------------------------|--------------------|----------------|------|-----|------------------------|------------------|
| 6            | 23                              | Ruminococcus gnavus| Campylobacter   | 0.333| 0.032| 0.415                  | 0.802            |
| 12           | 26                              | Dialister          | Campylobacter   | 0.288| 0.004| 0.305                  | 0.943            |
| 24           | 304                             | Coriobacteriaceae  | Campylobacter   | 0.222| 0.003| 0.240                  | 0.926            |
| 24           | 204                             | Prevotella         | Campylobacter   | 0.270| 0.004| 0.300                  | 0.900            |
| 24           | 254                             | Phasolorcotobacterium| Campylobacter | 0.286| 0.016| 0.340                  | 0.840            |
| 24           | 171                             | Eubacterium biforme| Campylobacter   | 0.289| 0.004| 0.320                  | 0.902            |
| 24           | 275                             | Prevotella         | Campylobacter   | 0.320| 0.000| 0.320                  | 1.000            |
| 24           | 52                              | Succinivibrio      | Campylobacter   | 0.353| 0.002| 0.360                  | 0.980            |
| 24           | 57                              | Catenibacterium    | Campylobacter   | 0.535| 0.066| 0.720                  | 0.743            |
| 24           | 242                             | Ruminococcus torques| Campylobacter | 0.002| 0.337| 0.040                  | 0.044            |
| 24           | 27                              | Bacteroides        | Campylobacter   | 0.032| 0.430| 0.260                  | 0.123            |
| 24           | 40                              | Bacteroides ovatus | Campylobacter   | 0.057| 0.494| 0.360                  | 0.159            |
| 24           | 39                              | Lachnospiraceae    | Campylobacter   | 0.251| 0.646| 0.800                  | 0.313            |
| 6            | 141                             | Morganella         | EAEC           | 0.190| 0.018| 0.240                  | 0.791            |
| 6            | 38                              | Collinsella aerofaciens | EAEC      | 0.234| 0.006| 0.260                  | 0.900            |
| 6            | 87                              | Clostridium difficile | EAEC       | 0.251| 0.032| 0.320                  | 0.784            |
| 6            | 23                              | Ruminococcus gnauus| EAEC           | 0.326| 0.042| 0.420                  | 0.776            |
| 12           | 31                              | Bacteroides        | EAEC           | 0.330| 0.018| 0.404                  | 0.817            |
| 18           | 299                             | Clostridium colinum| Giardia        | 0.278| 0.000| 0.278                  | 1.000            |
| 18           | 130                             | Clostridaceae      | Giardia        | 0.000| 0.242| 0.019                  | 0.005            |
| 24           | 438                             | Prevotella         | Giardia        | 0.217| 0.000| 0.217                  | 1.000            |
| 24           | 120                             | Clostridium        | Giardia        | 0.042| 0.518| 0.239                  | 0.177            |
| 6            | 141                             | Morganella         | Norovirus      | 0.185| 0.030| 0.273                  | 0.679            |
| 6            | 111                             | Pseudoramibacter/Eubacterium | Norovirus | 0.208| 0.025| 0.258                  | 0.809            |
| 6            | 112                             | Clostridium saccharogumia | Norovirus | 0.210| 0.033| 0.273                  | 0.771            |
| 6            | 23                              | Ruminococcus gnauus| Norovirus      | 0.261| 0.049| 0.348                  | 0.748            |
| 6            | 18                              | Clostridium        | Norovirus      | 0.336| 0.101| 0.485                  | 0.694            |
| 6            | 12                              | Streptococcus      | Norovirus      | 0.334| 0.645| 0.970                  | 0.344            |
| 24           | 108                             | Lachnospiraceae    | Norovirus      | 0.057| 0.482| 0.292                  | 0.197            |

Species indicative of high and low burden are shown in red and green, respectively. Expanded results with associations with adjusted P values < .1 are shown in Supplementary Table 1. Abbreviation: EAEC, entero-aggregative Escherichia coli.
genes involved in the uptake and metabolism of human milk oligosaccharides and has an important role in healthy gut community assembly in early childhood [24, 49]. There were additional strong correlations between indicator ASVs and high burden of enteropathogens at 6 months. A number of these ASVs are typically not observed in the developing gut microbiota of healthy children until the second postnatal year [28]. This observation lends further support to the hypothesis that early deviations in microbiota assembly away from the lower diversity, predominantly breast milk–associated community may be associated with adverse health outcomes, including infection with enteropathogens.

In summary, we report changes to gut microbial populations associated with asymptomatic Campylobacter infections in a longitudinal study with a large sample size, generating testable hypotheses to improve control of a highly prevalent enteropathogen with adverse effects on growth in infancy and early childhood. A large proportion of infants in this setting are already exposed to Campylobacter by age 6 months. Our study is limited by the fact that we did not analyze samples prior to 6 months, precluding our ability to characterize the microbiota before infection. Results emphasize the need to address the question of cause and effect, that is, is Campylobacter uniquely equipped to disrupt community assembly or do specific community features predispose to Campylobacter invasion and perturbations in the normal functional maturation of the microbiota? Future studies should be extended to determine the generalizability of the results to populations in geographical settings with different cultural and dietary practices. Advancing these research objectives will help assess whether interventions designed to deliberately target the microbiota may be useful in mitigating the deleterious effects of Campylobacter on growth.

Supplementary Data
Supplementary materials are available at Clinical Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes
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Potential conflicts of interest. J. G. is a co-founder of Matatu, Inc, a company that characterizes the role of diet-by-microbiota interactions in animal health. All other authors report no potential conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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