Targeting the adaptive molecular landscape of castration-resistant prostate cancer

Alexander W Wyatt* & Martin E Gleave**

Abstract

Castration and androgen receptor (AR) pathway inhibitors induce profound and sustained responses in advanced prostate cancer. However, the inevitable recurrence is associated with reactivation of the AR and progression to a more aggressive phenotype termed castration-resistant prostate cancer (CRPC). AR reactivation can occur directly through genomic modification of the AR gene, or indirectly via co-factor and co-chaperone deregulation. This mechanistic heterogeneity is further complicated by the stress-driven induction of a myriad of overlapping cellular survival pathways. In this review, we describe the heterogeneous and evolvable molecular landscape of CRPC and explore recent successes and failures of therapeutic strategies designed to target AR reactivation and adaptive survival pathways. We also discuss exciting areas of burgeoning anti-tumour research, and their potential to improve the survival and management of patients with CRPC.

Keywords androgen receptor; castration-resistant prostate cancer; stress response; survival pathways; tumour heterogeneity

DOI 10.15252/emmm.201303701 | Received 17 January 2015 | Revised 12 March 2015 | Accepted 26 March 2015 | Published online 20 April 2015

EMBO Mol Med (2015) 7: 878–894

See the Glossary for abbreviations used in this article.

Introduction

Normal prostate epithelial cells become malignant through deregulation to context-specific tumour suppressors and oncogenes (Taylor et al., 2010; Barbieri et al., 2012; Baca et al., 2013). Although the precise combination of genomic aberrations is notoriously heterogeneous between patients (Wyatt et al., 2014), at diagnosis the most common events include rearrangements affecting ETS gene family members, mutations in the ubiquitin ligase SPOP, disruption to the PI3K antagonist PTEN and copy number gain of oncogenic transcription factor MYC (Taylor et al., 2010; Barbieri et al., 2012). However, true to its hormone-regulated non-malignant ancestor, prostate cancer cells remain dependent on a ligand-activated androgen receptor (AR) to facilitate mitogenic responses enabled by genomic aberration. Despite enormous genomic heterogeneity, this biological homogeneity means that almost all tumours will initially respond to ligand depletion of the AR. Consequently, surgical or chemical castration, and subsequent elimination of most circulating testosterone, has been a mainstay of prostate cancer treatment for over 70 years (Huggins et al., 1941).

Unfortunately, due again to an innate ancestral ability, castration induces adaptive stress responses in prostate cancer cells that insulates against apoptosis. This, together with the steady accrual of further genomic and epigenomic aberration (Grasso et al., 2012), facilitates inevitable progression to a more aggressive tumour capable of growing in castrate levels of testosterone and thus termed castration-resistant prostate cancer (CRPC). The majority of CRPC re-initiate mitogenesis by reactivation of the AR: meaning that analogous targeting of the AR signalling axis is efficacious, at least for a short duration, in a large proportion of CRPC patients (Fizazi et al., 2012; Scher et al., 2012; Ferraldeschi et al., 2015). The precise nature and speed of AR reactivation is governed by existing and accruing genomic aberration, and with increasing cycles of AR axis targeting, this aberration can become more critical in enabling growth and converting innate adaptive survival responses into hard-wired assets. In fact, in the modern era of effective AR axis inhibition at different stages of progression, it is now relatively common to find late-stage CRPC where the AR has become incidental to a tumour’s successful growth and evolution (Beltran et al., 2012; Aparicio et al., 2013; Pezaro et al., 2014a).

Until 2010, only the cytotoxic docetaxel had demonstrated a clear survival benefit in patients progressing on first-line androgen deprivation therapy and first-generation AR antagonists (compounds that compete with endogenous ligand binding the AR) (Tannock et al., 2004). However, the revelation of AR reactivation in CRPC led to development of a wave of agents designed to better inhibit the AR signalling axis, with abiraterone acetate and enzalutamide the first to be approved (Fizazi et al., 2012; Scher et al., 2012). This review explores adaptive survival responses and genomic aberrations that facilitate AR reactivation in CRPC, and the recent successes and failures of strategies designed to exploit such changes. We describe the evolving landscape of treatment resistance that is developing in response to new agents, and discuss novel targets of therapeutic potential. Finally, we highlight the need for predictive biomarkers and evaluate the promise of liquid biopsies to help guide development and implementation of emerging therapeutics.
Glossary

Adaptive stress response
A mechanism by which cells can induce the expression of protective proteins to prevent cell damage

Androgen receptor (AR) antagonists
Compounds that compete with endogenous steroid hormones to bind the AR and thereby inhibit protein function

AR variants
Shortened forms of the AR protein that are missing the ligand-binding domain and therefore do not require activation by steroid molecules

Castration-resistant prostate cancer (CRPC)
Progression of prostate cancer to a form that can grow in very low levels of circulating testosterone

Cross-resistance
Tolerance of cancer cells to a normally effective drug due to prior resistance to a drug with a similar mechanism of action

Immunotherapy
Drugs that induce or enhance the body’s anti-tumour immune response

Liquid biopsy
Non-invasive blood tests that detect tumour cells or pieces of tumour DNA that are circulating in the blood

Stereoidogenesis
Enzymatic process by which cholesterol is converted to steroid hormones

Tumour heterogeneity
Different tumour cells within a single patient can show different genotypic and phenotypic profiles

Tumour microenvironment
The normal cells that surround tumour cells and provide them with nutrients and support

Direct reactivation of the androgen receptor

Renewed androgen synthesis in the castrate setting
Despite castrate levels of circulating testosterone, CRPC tissue often has higher levels of intra-tumoural androgens than non-CRPC counterparts, implying restoration of ligand for the AR (Mostaghel et al, 2007; Montgomery et al, 2008). This is due in part to an innate feedback mechanism inherited from non-malignant prostate epithelial cells, enabling them to adapt to varying levels of steroid. Indeed, a suite of steroidalogenic enzymes is epigenetically up-regulated in CRPC (Stanbrough et al, 2006; Mitsiades et al, 2012), particularly those resulting in accumulation of dihydrotestosterone (DHT, the testosterone metabolite preferred by the AR). This expression program can leverage adrenal androgens (Montgomery et al, 2008), or even initiate de novo synthesis of testosterone (Locke et al, 2008). At the extreme end of the spectrum are tumours with mutations in HSD3B1 (an enzyme governing a rate-limiting step in DHT synthesis) that facilitate accumulation of protein and hence increased DHT synthesis (Chang et al, 2013).

Ultimately, testosterone and DHT synthesis are dependent on catalytic conversion of cholesterol by members of cytochrome P450 (CYP) family of enzymes. CYP17A1 is a pivotal enzyme in this process, required for both canonical and alternative androgen synthesis, and has consequently been the focus of concerted drug development over the past decade (Fig 1). This strategy was vindicated when phase III trials of the CYP17A1 inhibitor abiraterone acetate in metastatic CRPC (mCRPC) patients post- and pre-chemotherapy demonstrated an improved median overall survival (Fizazi et al, 2012; Ryan et al, 2013). The success of abiraterone has accelerated development of other CYP17A1 inhibitors, particularly compounds that specifically inhibit the 17,20-lyase activity of CYP17A1 and thereby render glucocorticoid co-admission unnecessary (Ferraldeschi et al, 2013). One such agent, orteronel (TAK-700; Kaku et al, 2011), was recently evaluated in two large phase III trials in mCRPC patients pre- and post-chemotherapy (NCT01193244; NCT01193257), but failed to demonstrate an overall survival benefit (Dreicer et al, 2014; Saad et al, 2015). These trials were likely confounded by post-study availability of abiraterone, and moreover, since both trials co-administered orteronel with prednisone, its vaunted 17,20-lyase specificity was not exploited. Two further next-generation CYP17A1 inhibitors, VT-464 (Toren et al, 2015) and galeterone (TOK-001) (Handratta et al, 2005) are currently undergoing phase I and II development, respectively (NCT02012920; NCT01709734) (Table 1). VT-464 demonstrated anti-cancer activity in preclinical models of advanced CRPC, significantly lowering tumoural androgen levels in castrate mice, and enforcing greater suppression of the AR signalling axis compared to abiraterone (Toren et al, 2015). Galeterone showed promising phase I activity in chemotherapy-naïve CRPC patients (Taplin et al, 2012) and has the convenient side-effect of AR cross-inhibition.

Although CYP17A1 is a critical hub in stereoidogenesis, there is a clear rationale to co-target other members in the pathway. Indeed, dutasteride, which inhibits 5-alpha-reductase (SRD5A1) catalysis of testosterone to DHT, is currently being tested in combination with abiraterone (NCT01393730). Similarly, AKR1C3 is a promising target given its significant up-regulation in CRPC and key role reducing androstenedione to testosterone (Adeniji et al, 2013). Although the selective oral AKR1C3 inhibitor ASP9521 (Kikuchi et al, 2014) showed no response in phase I/II trials in mCRPC (Loriot et al, 2014), it likely requires use in combination given the alternative pathways to generate DHT.

Genomic modification of the androgen receptor
Even in a castrate-sensitive cell, ligand depletion triggers an innate feedback response leading to increased transcription of the AR gene (Wolf et al, 1993; Cai et al, 2011; Knuutila et al, 2014; Wyatt et al, 2014). The consequent overexpression of AR in CRPC tissue confers hypersensitivity to low levels of androgen as well as facilitating antagonism to agonist conversion for some first-generation AR antagonists (Chen et al, 2004). In over 60% of initial CRPC, AR overexpression is driven by X chromosome rearrangement and subsequent focal copy number gain (Grasso et al, 2012). The persistent transcriptional pressure on the AR gene caused by ligand depletion probably confers susceptibility to DNA breakage (Mathas & Misteli, 2009) and is likely to be partly responsible for AR amplification.

Therefore, in parallel to the clinical development of CYP17A1 inhibitors, awareness of AR up-regulation in CRPC led to wave of second-generation AR antagonists that compete more effectively with androgen for the ligand-binding domain (LBD) of the AR (Fig 1). The first to enter the clinic was enzalutamide, conferring an overall survival improvement for both post- and pre-chemotherapy...
CRPC patients in phase III trials (Scher et al., 2012; Beer et al., 2014). The analogous compound ARN-509 has showed encouraging activity in phase I trials in chemotherapy-naïve CRPC patients before and after abiraterone treatment (Rathkopf et al., 2013b) and is currently under phase III evaluation in non-metastatic CRPC (NCT01946204), with a primary endpoint of delay in progression to M1 disease. A third potent AR antagonist, ODM-201, has a distinctly different structure than enzalutamide or ARN-509 and having shown promise in recent phase I/II trials (Fizazi et al., 2014), will now be evaluated in a phase III trial in men with high-risk non-metastatic CRPC (NCT02200614). The success of these new AR-LBD antagonists reflects more potent inhibition of DHT binding to the AR
and prevention of AR nuclear translocation and binding to target promoters (Tran et al., 2009; Clegg et al., 2012).

This increasing arsenal of more potent anti-AR drugs will provide more tools to combat the non-synonymous AR mutations that are detectable in 10–20% of initial CRPC patients (Grasso et al., 2012; Beltran et al., 2013). The majority of documented mutations fall within the LBD or cofactor binding regions (Gottlieb et al., 2012), reducing binding specificity and permitting activation of the AR by adrenal androgens or other steroid metabolites. Certain LBD mutations are also sufficient to convert AR antagonists to AR agonists and are likely responsible for the 15–30% of patients that exhibit a withdrawal syndrome after cessation of first-generation therapies (e.g. bicalutamide, flutamide) (Small et al., 2004). However, since individual mutations do not tend to confer pan-antagonist resistance, the development of real-time strategies to monitor for AR mutation emergence could guide rational sequencing of AR pathway inhibitors. Interestingly, although a recently reported AR mutation (F876L) can drive resistance to both enzalutamide and ARN-509 in vitro (Balbas et al., 2013; Joseph et al., 2013; Korpal et al., 2013), few cases of enzalutamide withdrawal syndrome have been reported to date (Rodriguez-Vida et al., 2015). Optimistically, this may suggest conversion of enzalutamide from antagonist to agonist is rare, but it may simply reflect that the contemporary metastatic landscape can have multiple independent tumour clones, each responding differently to therapy (Carreira et al., 2014). AR mutations will also be a mechanism of resistance to CYP17A1 inhibitors, especially agents requiring prednisone co-admission, since certain mutations (e.g. L702H) can repurpose glucocorticoids as AR ligand (Carreira et al., 2014). Furthermore, abiraterone increases progesterone levels and appears to select for the progesterone-activating mutation T877A (Chen et al., 2013), promoting AR nuclear translocation and binding to target promoters (Tran et al., 2009; Clegg et al., 2012).

### Table 1. Selected ongoing clinical trials for novel treatments of patients with CRPC.

| Agent(s) | Activity | Phase | Trial ID |
|----------|----------|-------|---------|
| VT-464   | Lyase-selective inhibitor of CYP17 | I/II   | NCT02012920 |
| Galetone (TOK-001) | Dual CYP17 inhibitor and AR antagonist | II | NCT01709734 |
| ARN-509  | Second-generation AR antagonist | III (SPARTAN) | NCT01946204 |
| ODQ-201  | Second-generation AR antagonist | III (ARMS) | NCT02200614 |
| Enzalutamide + abiraterone | Second-generation AR antagonist; CYP17 inhibitor | II | NCT01650194 |
| ARN-509 + abiraterone | Second-generation AR antagonist; CYP17 inhibitor | Ib | NCT01792687 |
| Enzalutamide ± abiraterone | Second-generation AR antagonist; CYP17 inhibitor | III | NCT01949337 |

### Targeting adaptive survival pathways

| Agent(s) | Activity | Phase | Trial ID |
|----------|----------|-------|---------|
| OGX-427 + abiraterone | HSP27 inhibitor; CYP17 inhibitor | II | NCT01681433 |
| AT13387 + abiraterone | HSP90 inhibitor; CYP17 inhibitor | I/II | NCT01685268 |
| CDC-0068 + abiraterone | Pan AKT inhibitor; CYP17 inhibitor | Ib/II | NCT01485861 |
| BEZ235 + abiraterone | Dual PI3K and mTOR inhibitor; CYP17 inhibitor | Ib | NCT01634061 |
| BKM120 + abiraterone | Pan PI3K inhibitor; CYP17 inhibitor | Ib | NCT01634061 |
| AZD8186 | PI3K beta and delta inhibitor | I | NCT01848285 |
| GSK2636771 | PI3K beta inhibitor | I/IIa | NCT01458067 |
| Cabozenab + abiraterone | Tyrosine kinase inhibitor; CYP17 inhibitor | I | NCT01574937 |
| Dasatinib ± abiraterone | Tyrosine kinase inhibitor; CYP17 inhibitor | II (randomized) | NCT01685125 |
| Sunitinib or dasatinib ± abiraterone | Tyrosine kinase inhibitors; CYP17 inhibitor | II (randomized) | NCT01254864 |
| Tivozanib + enzalutamide | VEGF inhibitor; Second-generation AR antagonist | II | NCT01885949 |
| Dovitinib + abiraterone | Tyrosine kinase inhibitor; CYP17 inhibitor | II | NCT01994590 |
| OGX-011 ± cabazitaxel | CLU inhibitor; microtubule inhibitor | III (AFFINITY) | NCT01578655 |
| Alisertib + abiraterone | AURKA inhibitor; CYP17 inhibitor | I/II (randomized) | NCT01848067 |

### Inhibiting DNA repair

| Agent(s) | Activity | Phase | Trial ID |
|----------|----------|-------|---------|
| Olaparib | Selective PARP1 inhibitor | II | NCT01682772 |
| Veliparib ± abiraterone | PARP inhibitor | I/II (randomized) | NCT01576172 |

### Immunotherapy

| Agent(s) | Activity | Phase | Trial ID |
|----------|----------|-------|---------|
| PROSTVAC | | III (Prospect) | NCT01322490 |

### Targeting neuroendocrine prostate cancer

| Agent(s) | Activity | Phase | Trial ID |
|----------|----------|-------|---------|
| Alisertib (MLN8237) | AURKA inhibitor | II | NCT01799278 |
increasingly potent AR therapeutics augment the selective pressure for AR aberration, which can drive primary resistance to the next agent in line. Drug sequence strategies have been further compounded by suggestion that part of the activity of docetaxel in CRPC patients can be attributed to a microtubule-dependent effect on AR activity (Gan et al., 2009; Zhu et al., 2010; Darshan et al., 2011). Indeed, the activity of docetaxel is significantly reduced after abiraterone treatment (Mezynski et al., 2012), although this effect may not be due to cross-resistance per se (Azad et al., 2014; de Leeuw et al., 2015). Early evidence also suggests that enzalutamide elicits only a modest response rate in the post-docetaxel and post-abiraterone population (Bianchini et al., 2014; S Chand et al., 2015) and equally and that abiraterone treatment in the post-enzalutamide population has limited activity (Noonan et al., 2013). Similarly ARN-509, which appeared to have greater in vivo activity than enzalutamide (Clegg et al., 2012), showed indications of diminished activity post-abiraterone compared to the abiraterone-naïve population (Rathkopf et al., 2013a). Ultimately, oncologists are faced with the sobering reality of primary resistance to first- and second-line abiraterone or enzalutamide in 20–40% and 60–80% of CRPC patients, respectively. Even those patients that enjoy initial responses will eventually acquire resistance (Scher et al., 2010, 2012; de Bono et al., 2011).

AR copy number gain and mutation undoubtedly contribute to AR pathway inhibitor resistance [for example, AR gain has recently been linked to lack of response to abiraterone (Carreira et al., 2014)], but they cannot explain the entire landscape of progressing disease. It is now widely recognized that the increased transcriptional pressure on the AR gene during CRPC progression can lead to the generation of truncated isoforms of the AR: coding only for the DNA binding and transactivation domains. Since these AR variants are missing the LBD, they are constitutively active and DNA binding and transactivation potential to specifically inhibit the intrinsically disordered N-terminal domain. In theory, inhibition of AR mRNA via antisense oligonucleotide (ASO) interference is a simple strategy to reduce expression of all AR species. Although a phase I trial of ASO EZN-4167 in advanced CRPC showed minimal activity and concerning toxicity (Bianchini et al., 2013), the drug binds to exon 4, which is missing from the most common AR-Vs. More recently, Yamamoto and colleagues demonstrated that an ASO targeting AR exon 1 (ISIS-ARRx) is sufficient to suppress both ARFL and AR-Vs in enzalutamide-resistant preclinical models (Yamamoto et al., 2015). Furthermore, clear anti-tumour activity of ISIS-ARRx in these models provides preclinical support for AR-ASO strategies as a rational third-line approach for AR pathway inhibitor-resistant CRPC.

With abiraterone and enzalutamide now standard of care in first-line CRPC, novel AR-targeted agents, regardless of mechanism of action, will need to show activity in patients progressing on AR pathway inhibitors who will have higher levels of AR expression and aberration.

**Indirect reactivation of the androgen receptor**

**Deregulation of androgen receptor co-chaperones**

In the absence of ligand in a non-malignant prostate cell, the AR protein forms a complex in the cytoplasm with heat-shock proteins (e.g. HSP70, HSP90) acting as molecular chaperones to maintain the AR in a stable conformation for ligand binding and protect it from proteolysis (Chmelar et al., 2007) (Fig 1). Ligand binding elicits an AR conformation change, and the receptor is then trafficked to the nucleus (Cano et al., 2013). However, the autonomous stress response induced by castration induces activity of heat-shock proteins and helps insulate the AR axis from degradation (Azad et al., 2015c). For example, the stress-induced chaperone, HSP27, is induced by castration and recruited to promote the nuclear transport of the AR (Zoubedi et al., 2007). Overexpression of HSP27 in vivo suppresses apoptosis and is sufficient to confer castrate resistance (Rocchi et al., 2005; Zoubedi et al., 2010b). Thus, inhibiting HSP90 and/or HSP27 may disrupt the AR foldingome and could sensitize AR-targeted agents.

Indeed, a recent phase II trial of OGX-427 (a second-generation ASO against HSP27) in mCRPC patients demonstrated promising results, doubling the PSA (prostate-specific antigen) response (>50% decline) rate compared to prednisone alone (Chi et al., 2012). Conversely however, phase I/II trials of HSP90 inhibitors in CRPC patients have been disappointing, despite preclinical activity in CRPC models (Heath et al., 2008; 2013; Lamoureux et al., 2011b; Pacey et al., 2011). Alternative targeting strategies are in development, including the use of sulfuraphane to inhibit HDAC6 and thereby prevent HSP90 acetylation (Gibbs et al., 2009). Overall, the efficacy of HSP90 inhibition is limited by the functional redundancy of molecular chaperones and adaptive feedback mechanisms including the activation of HSF1 [a heat-shock transcription factor that induces, amongst others, HSP70, HSP27 and CLU (clusterin)] (Azad et al., 2015c).

Since molecular chaperones play key roles in endoplasmic reticulum stress responses and protein homeostasis, co-targeting two or more may overwhelm the ability of cancer cells to regulate their misfolded protein burden. Accordingly, the preclinical activity of HSP90 inhibitors is enhanced via simultaneous
targeting of HSF1, HSP27, or CLU (Lamoureux et al., 2011a, 2014; Chen et al., 2013). Co-targeting the AR simultaneously with HSP90 or HSP27 is also a rational combination strategy: results from phase II trials combining the HSP90 inhibitor AT13387 (NCT01685268) or the HSP27 inhibitor OGX-427 (NCT01681433) with abiraterone will be enlightening (Table 1).

Genomic modification of androgen receptor co-activators

Once in the nucleus, the functions of the AR are mediated by a milieu of co-factors and co-activators capable of modulating the selection and expression of downstream targets (Chmellar et al., 2007; Heemers et al., 2009; Xu et al., 2009). In the context of CRPC, the P160 SRC (steroid receptor co-activator) family genes, NCOA1, NCOA2 and NCOA3 (also known as SRC1-3), have received considerable attention. Overexpression of NCOA1 or NCOA2 can drive increased AR transactivation in castrate conditions (Gregory et al., 2001), and depletion of NCOA2 prevents CRPC development in PTEN-deficient mice (Qin et al., 2014). Interestingly, NCOA2 up-regulation in CRPC combines genomic and adaptive mechanisms, since it is de-repressed by androgen depletion but also frequently amplified in advanced prostate cancer (Agoulnik et al., 2006; Taylor et al., 2010). NCOA3 is also linked to prostate cancer cell proliferation and survival (Zhou et al., 2005; Yan et al., 2008) and is a key target of the ubiquitin ligase SPOP [mutated in 6–15% of prostate cancer (Barbieri et al., 2012)]. Wild-type SPOP, but not mutant, promotes ubiquitination (and subsequent degradation) of NCOA3 (Geng et al., 2013), and interestingly the AR as well (An et al., 2014; Geng et al., 2014) (Fig 1). Although targeting SPOP in prostate cancer constitutes a considerable challenge, inhibiting downstream proteins escaping ubiquitination in SPOP mutant tumours may be more feasible, lending further credence to strategies targeting the P160 SRC family.

The forkhead protein FOXA1 is a critical interacting partner of the AR, functioning as a pioneer factor to modulate chromatin accessibility and facilitate transcription (Jozwik & Carroll, 2012). In prostate cancer, FOXA1 is capable of specifying unique AR binding sites and has an AR-independent function as a metastasis regulator (Jin et al., 2013; Sahu et al., 2013). Although it can be genomically amplified, deleted, or mutated in CRPC patients, suggesting complex context-dependent activity (Taylor et al., 2010; Barbieri et al., 2012; Grasso et al., 2012), the precedent set by the development of a FOXM1 inhibitor suggests that forkhead protein modulation in prostate cancer might hold promise (Gormally et al., 2014). Interestingly, FOXA1 and AR co-localize on chromatin with GATA2, a transcription factor that enhances recruitment of NCOAs to the AR complex (He et al., 2014). Additionally, at the transcriptional level, there appears to be a complex feedback balance between GATA2 and the AR itself, since GATA2 is repressed by the AR and androgen, but is necessary for optimal expression of the AR (He et al., 2014). High GATA2 expression predicts poor outcome in prostate cancer patients and further promotes the concept of therapeutically targeting the AR transcriptional complex in CRPC patients. A promising contemporary strategy to disrupt AR in this manner is to use bromodomain inhibitors (e.g. JQ1) to inhibit the chromatin reader BRD4 that interacts with the N-terminal domain of the AR (Asangani et al., 2014). Preclinical studies have shown that JQ1 disrupts AR-mediated gene transcription in CRPC models, significantly reducing tumour volume relative to controls (Asangani et al., 2014).

An interesting alternative approach to inhibit AR co-activators is to specifically target their interaction with the AR. Recently, potent inhibitors of the AR Binding Factor 3 (BF3) pocket have been developed that demonstrate activity in enzalutamide-resistant preclinical models (Minuganti et al., 2014). A novel class of small organic molecules without a peptide backbone (peptidomimetics) have also been recently shown to disrupt AR co-activator interactions and are candidates for clinical development (Ravindranathan et al., 2013).

Adaptive induction of compensatory pathways

The therapeutic targeting of driver aberration (e.g. an overactive AR) can activate adaptive survival pathways, leading to apoptosis inhibition, tumour cell plasticity, and the emergence of treatment resistance (Zoubeidi et al., 2010a). Unfortunately, the functional redundancy and heterogeneity of CRPC mean that no single pathway is relevant to all tumours and therefore that therapeutic targeting of a specific pathway is likely of limited benefit. However, combination strategies inhibiting molecules involved in crosstalk between multiple pathways have potential to induce conditional lethality (Carver et al., 2011; Azad et al., 2015c). Consequently, in concert with AR inhibition, complimentary strategies co-targeting signalling pathways that cooperatively activate the AR, or stress response pathways that maintain homeostasis, represent exciting opportunities to stimulate a high therapeutic index. Only through precise characterization of the stress-induced adaptive response, and the rational development of combinatorial co-targeting strategies, will the full potential of AR pathway inhibition be achieved.

Activation of kinase-dependent signalling pathways

The profound effect of AR axis inhibition on prostate cancer cells has ramifications for many kinase signalling pathways, particularly those that have accumulated genomic aberration during disease progression. The PI3K/AKT/MTOR pathway is frequently altered in advanced prostate cancer, particularly through deletion of PTEN (> 50% CRPC), but also through mutation (e.g. of PI3KCA) or overexpression of upstream tyrosine kinases (Taylor et al., 2010; Grasso et al., 2012). PTEN loss leads to greater PI3K activity in castrate conditions and cell proliferation, thereby providing a potential escape route from AR inhibition (Wang et al., 2003). In theory, the PI3K/AKT/MTOR pathway marks a putative Achilles heel to target with rational drug design, but it is riddled with functional redundancy and complex compensatory mechanisms. For example, in the absence of PTEN, the AR and PI3K pathways cross-regulate each other via reciprocal feedback, at least in model systems (Carver et al., 2011; Mulholland et al., 2011). A recent study in breast cancer demonstrated that PTEN loss on the background of PI3KCA mutation actually conferred resistance to a PI3K inhibitor (Juric et al., 2015). Perhaps unsurprisingly, single agent targeting of the PI3K pathway in CRPC has been singularly underwhelming, with minimal activity in phase II trials of mTOR inhibitors ridaforolimus, temsirolimus and everolimus (Amato et al., 2012; Krucezk et al., 2013; Templeton et al., 2013). Trials combining PI3K and AR inhibition are more promising in terms of preventing compensatory feedback, but may encounter toxicity issues (Thomas et al., 2013) (NCT01485861;NCT01634061).
Preclinical data suggest the involvement of other signalling molecules including IGF1, HER2, MET and SRC kinases in the progression of segmented populations after second-line AR pathway inhibition (reviewed in Lorente & De Bono, 2014). However, while kinase-targeted strategies are a successful example of precision therapy for several cancers, the majority fail to produce long-term durable response or complete remission (Zhang et al, 2009). Furthermore, tyrosine kinases are rarely altered at the genomic level in CRPC (Grasso et al, 2012). As such, it is unlikely that prostate cancer cells are inherently addicted to specific kinase activation (c.f. EGFR mutations in lung cancer) and can instead adapt accordingly to monotherapy inhibition. For example, although up-regulation of HER2 in CRPC results in increased AR transcriptional activity (Mellinghoff et al, 2004), lapatinib (a non-selective HER2 inhibitor) showed very limited activity in CRPC patients (Whang et al, 2013). Other tyrosine kinase inhibitors targeting MET (cabozantinib), VEGF (sunitinib), endothelin (atasentan, zibotentan) and SRC (dasatinib) failed in phase III trials to improve overall survival of post-docetaxel CRPC patients (Lorente & De Bono, 2014; Michaelsen et al, 2014; Sridhar et al, 2014). These failures are likely driven by the aforementioned lack of absolute kinase dependency in CRPC, and subsequent bypass via compensatory pathways. A non-ideal choice of co-targeting drug (e.g. docetaxel) may also have played a role. More promisingly, there are several ongoing phase I/II trials evaluating tyrosine kinase inhibitors in combination with abiraterone or enzalutamide, and it is conceivable that eliminating context-dependent tyrosine kinase activation as a compensatory mechanism for AR inactivation will enhance the efficacy of AR-targeted agents (Table 1).

Activation of stress response pathways
Cellular stress can drive the evolution and adaptation of cancer cells. The stress response that is activated by castration in AR-driven prostate cancers includes up-regulation of molecular chaperones that regulate protein homeostasis and diverse survival signalling and transcriptional survival networks (Garrido et al, 2006; Dai et al, 2007; Zoubeidi & Gleave, 2012; Matsumoto et al, 2013). It is unsurprising therefore that certain chaperones are frequently up-regulated in prostate and other cancers and their expression correlates with metastases, treatment resistance, and poor survival (Azad et al, 2015c). Outside of the AR signalling axis (that is insulated by the heat-shock proteins discussed above), CLU is the most credentialed molecular chaperone capable of driving treatment resistance in CRPC cells. CLU is induced by stress-activated transcription factors, including EGR1, HSF1 and YBX1, to constrain apoptosis through inhibition of activated Bax and suppression of protein aggregation via autophagy activation (Zoubeidi & Gleave, 2012; Zhang et al, 2014). Accordingly, CLU inhibition potentiates activity of anti-cancer therapeutics in preclinical models (Sowery et al, 2008; Zoubeidi et al, 2010a).

Since CLU is challenging to target with traditional small-molecule inhibitors, the ASO drug OGX-011 (custirsen) was developed to instead inhibit mRNA translation. A phase II trial reported a 7-month overall survival benefit of OGX-011 in combination with docetaxel, compared to docetaxel alone, in chemotherapy-naïve mCRPC (Chi et al, 2010), leading to the initiation of randomized phase III studies: SYNERGY (NCT01188187) and AFFINITY (NCT01578655). SYNERGY randomized 1,022 men with mCRPC to OGX-011 in combination with docetaxel and prednisone to docetaxel and prednisone alone. Survival results were first presented at ESMO in 2014 (Chi et al, 2014) and indicated that addition of OGX-011 did not meet the primary endpoint of a statistically significant improvement in overall survival in men with CRPC compared to docetaxel/prednisone alone (median survival 23.4 versus 22.2 months, respectively; hazard ratio 0.93 and P-value 0.207). AFFINITY, which assesses the second-line indication comparing cabazitaxel with or without OGX-011 in post-docetaxel-treated CRPC, has completed enrolment and should read out by end of 2015 (Table 1). The failure of OGX-011 in SYNERGY, despite robust preclinical proof-of-principle, phase I on-target suppression data (Chi et al, 2005), and phase II survival signals (Chi et al, 2010), illustrates the challenge of selecting appropriate combinations based on phase II signals, and how changes in treatment landscape (in this case the approval of abiraterone and enzalutamide) mid-development may alter outcomes. Given its location on chromosome 8p proximal to the prostate cancer tumour suppressor gene NKX3-1, the CLU gene is homozygously deleted in ~20% of CRPC patients (Grasso et al, 2012), a population that likely confounded OGX-011 evaluation.

Somatic deregulation to DNA repair and cell cycle machinery
At diagnosis, the genomic landscape of prostate cancer is very heterogeneous, bearing a heavy burden of genomic rearrangement replete with rare combinations of tumour suppressor inactivation and oncogene activation (Taylor et al, 2010; Barbieri et al, 2012; Grasso et al, 2012; Baca et al, 2013; Wyatt et al, 2014). Furthermore, since the AR program regulates the transcriptional programs of DNA repair genes in prostate cancer cells, AR axis inhibition has the potentially undesirable side-effect of promoting genomic instability (Polkinghorn et al, 2013). In theory however, as the life-expectancy of contemporary CRPC patients rises, there is increasing opportunity for accumulating genomic aberration to render certain cellular functions useless. For example, it is now recognized that 12% of advanced prostate cancers have accrued defects to DNA mismatch repair machinery and have consequently developed a “hypermutated” genotype similar to that observed in microsatellite instability in colon cancer (Pritchard et al, 2014).

The shedding of specific cellular machinery can remove aspects of functional redundancy that shields cancer cells from monotherapy. Homozygous somatic aberration to key mediators of homologous recombination in DNA repair, including BRCA2 and ATM, now appears common in advanced CRPC (Grasso et al, 2012). This discovery proposed the intriguing hypothesis that targeting DNA repair machinery via poly (ADP-ribose) polymerase (PARP) inhibition will induce synthetic lethality exclusive to tumour cells. PARP1 is a particularly attractive target in CRPC since it also plays a key role supporting both AR function (Schiewer et al, 2012b) and ETS transcription factor activity (Brenner et al, 2011). Remarkably, durable responses have been reported for CRPC patients treated with niraparib, a PARP1 and PARP2 inhibitor (Sandhu et al, 2013). Considerable optimism surrounds the current phase II evaluation of olaparib (a selective PARP1 targeted agent) as a monotherapy in CRPC (NCT01682772), and a phase II trial of veliparib (a PARP1 and PARP2 inhibitor) in combination with abiraterone and prednisone (NCT01576172).

© 2015 The Authors
In prostate cancer, the AR facilitates cell proliferation via effects on the cyclin/cyclin-dependent kinase (CDK)/retinoblastoma (RB) pathway (Schiewer et al, 2012a). This critical cell cycle machinery is frequently deregulated in CRPC, with CCND1 amplification and/or RB1 loss the most recognized genomic contribution, detectable in 5–8 and 30% of CRPC, respectively (Bubendorf et al, 1999; Grasso et al, 2012). In tumours with functional RB1, inhibition of CDK4 and CDK6 activity has a significant suppressive effect on cell proliferation (Comstock et al, 2013). The future success of trials evaluating CDK4/6 inhibitors in CRPC is likely to hinge on enrichment with patients whose tumours exhibit cyclin/CDK activation in the background of intact RB1. Conversely, tumours with complete RB1 loss have hard-wired activation of cyclin/CDK-mediated cell proliferation. Although this renders CDK4/6 inhibition redundant, cells are presumably less able to modulate proliferation. In apparent support of this concept, recent evidence suggests that RB1-depleted tumours are more sensitive to the novel taxane chemotherapy cabazitaxel (de Leeuw et al, 2014, 2015), which was recently approved for patients progressing on docetaxel after demonstrating a survival benefit over mitoxantrone and prednisone (de Bono et al, 2010). Importantly, cabazitaxel does not appear to exhibit cross-resistance with AR-targeted agents, suggesting effects are independent of the AR pathway (Al Nakouzi et al, 2014; Pezaro et al, 2014b; van Soest et al, 2014; de Leeuw et al, 2015). Genomic mechanisms of resistance to taxane-based chemotherapy are unclear, although recent data suggest that ERG affects microtubule dynamics and that ERG overexpressing tumours are consequently more resistant to docetaxel (Galletti et al, 2014). Interestingly, ERG itself represents a potential therapeutic target in tumours with ERG rearrangements (Wang et al, 2014). Since the majority of rearrangements place ERG under direct control of an AR-regulated promoter (e.g. from the TMPRSS2 gene), ERG is overexpressed as a direct consequence of AR reactivation in CRPC (Cai et al, 2009). Although the clinical development of specific ERG inhibitors has proved elusive to date, a recent study showed that inhibition of the ERG-stabilizing deubiquitinase USP9X results in ERG depletion and may represent a novel therapeutic strategy (Wang et al, 2014).

The protective role of the microenvironment

Cancer cells reside within a complex microenvironment that can either compromise or augment survival and growth (Sun & Nelson, 2012). Furthermore, as prostate cancer switches from an endocrine-driven disease to a paracrine- or autocrine-driven disease after CRPC development, tumour cells become increasingly reliant on the microenvironment for survival. For example, prostate-cancer-associated stromal cells can facilitate androgen biosynthesis in tumour cells under castrate conditions (Arnold et al, 2008; Mizokami et al, 2009; Sillat et al, 2009). More recently, an elegant study demonstrated that in the aftermath of genotoxic therapy, the innate DNA damage response in benign stromal cells stimulates secretion of cytokines, growth factors and proteases that ultimately promote therapy resistance in tumour cells (Sun et al, 2012).

Strategies designed to target the tumour microenvironment are attractive, not least since normal cells cannot easily evolve to a resistant state. The most effective approach in many solid malignancies has been to interfere with VEGF-mediated blood vessel recruitment to tumour tissue. Unfortunately, attempts to repurpose anti-angiogenic drugs for CRPC have failed. For example, the VEGF-targeted agents bevacizumab, albintercept and lenalidomide all failed to improve the overall survival conferred by docetaxel in large phase III trials (Kelly et al, 2012; Petrylak et al, 2012; Tannock et al, 2013), collectively implying that VEGF-mediated angiogenesis is not the sole driver of progression in bone-predominant mCRPC. Similarly, endothelin receptor (END1) targeting agents, atrasentan and zibotentan, also failed in phase III studies (Carducci et al, 2007; Nelson et al, 2012), despite biologic and preclinical proof-of-principle as well as signals of activity in phase II studies. Overall, the development of these angiogenesis inhibitors was challenged by lack of single agent activity that compromised detection and/or interpretation of robust activity signals. In the case of zibotentan, a randomized phase II versus placebo in men with M1 CRPC demonstrated improved markers of bone turnover and initially signalled significantly improved survival but with maturation this benefit disappeared (James et al, 2009, 2010). Based on the initial survival benefit, a phase III trial enrolled 594 patients, but survival was not significantly prolonged, in part due to insufficient sample size.

A more successful strategy has been to exploit the remarkable propensity of tumour cells to form metastatic deposits in the bone. Despite decades of availability, crude radiopharmaceuticals have demonstrated only limited uptake due to incidental bone marrow toxicity from errant beta-particles. However, in 2013, the calcium mimetic radium-223 dichloride (Xofigo) was approved for the treatment of bone metastatic CRPC (Parker et al, 2013). Activity is reliant on the potent effect, but short range of alpha radiation emitted from radium-223 decay: reducing peripheral damage to healthy tissue while maintaining powerful anti-tumour efficacy. Overall, although radiopharmaceuticals do not strictly target the microenvironment, their rational use has demonstrated that it is possible to elicit overall survival gains by selectively targeting the bone niche.

Transient cell populations that migrate in and out of the ecosystem can also influence tumour dynamics. Tumours arise in an immunocompetent environment, interacting with innate and adaptive branches of the host immune system (May et al, 2011). Although the host immune system is capable of mounting an anti-tumour response, tumour cells frequently enjoy an excess of regulatory and suppressor T cells, blunting the effector response. In the apoptotic aftermath of initial androgen deprivation therapy, leukocytes are further recruited to tumour tissue, but rather than reacting to the cancer, they may promote progression to CRPC (Luo et al, 2007; Ammirante et al, 2010). The field of immunotherapeutics seeks to exploit the potent and intact anti-tumour response and is reviewed in-depth elsewhere (Madan et al, 2013; Makkouk & Weiner, 2015; May et al, 2011). The most advanced clinical strategies for CRPC are therapeutic vaccines that induce a novel anti-tumour response, and immune checkpoint modulators that prevent suppression of the existing response. Sipuleucel-T (provenge) is a therapeutic vaccine generated by ex vivo stimulation of antigen presenting cells. It became the first immunotherapy approved for use in prostate cancer after demonstrating a significant overall survival benefit in asymptomatic or minimally symptomatic mCRPC (Kantoff et al, 2010a). Similarly, the vector-based vaccine PSA-TRICOM (PROSTVAC), which generates an in vivo response against cells expressing PSA, showed an overall survival benefit for
mCRPC patients in phase II trials (Gulley et al., 2010; Kantoff et al., 2010b) and is currently under phase III evaluation in men with asymptomatic or minimally symptomatic mCRPC (NCT01322490). Ipilimumab (yervoy) is an antibody that binds to CTLA-4 and prevents the suppression of cytotoxic T cells, resulting in a more aggressive anti-tumour immune response (Slovik et al., 2013). Although a phase III trial of ipilimumab in mCRPC did not show a significant gain in overall survival (Kwon et al., 2014), there was evidence of activity in patients with favourable prognoses (Drake et al., 2014). This is particularly relevant given that patients with the least aggressive disease appear to receive the most benefit from sipuleucel-T and PSA-TRICOM.

Despite overall survival gains, immunotherapeutics have not demonstrated impact on short-term progression. Rather, preliminary analyses suggest their efficacy in prostate cancer is via long-term alterations in tumour growth kinetics (Beer et al., 2011; Gulley et al., 2013), potentially explaining the observations of increased activity in less advanced disease. Furthermore, since an anti-tumour response tends to be sustained and can even evolve over time to target more antigens [known as antigen cascade (Disis et al., 2004)], there is a strong rationale to evaluate immunotherapies earlier in disease progression. Combining vaccines with checkpoint inhibition may also enhance the anti-tumour response and is currently being evaluated (Jochems et al., 2014) (NCT01832870).

Inactivation of the androgen receptor

The continual accrual of genomic aberration together with the de-differentiating force of sustained AR inhibition provides opportunities for tumour cells to escape dependence on AR signalling. One potential escape route is via up-regulation of compensatory steroid receptors that show high homology to the AR, suggesting a degree of functional redundancy. Indeed, oestrogen receptor (ER) alpha and beta are frequently upregulated in advanced prostate cancer, but whereas preclinical data support the use of ER targeted agents, there is no evidence of clinical response to ER modulation in CRPC patients (Nelson et al., 2014). A recent study demonstrated that the glucocorticoid receptor (GR) can regulate a proportion of the AR cistrome and its up-regulation in AR-repressed conditions may represent a mechanism to re-initiate mitogenesis in CRPC (Arora et al., 2013; Sahu et al., 2013).

Tumours can also evolve or adapt to become a completely AR-independent disease. In the contemporary disease setting of potent AR targeting, it has become common to observe progression of advanced CRPC in the absence of high serum PSA (a marker of AR activity) and with atypical visceral metastases (Beltran et al., 2012; Aparicio et al., 2013; Pezaro et al., 2014a). Predictably, AR-independent prostate cancer is highly heterogeneous, but a major established subtype is neuroendocrine prostate cancer (NEPC) (Beltran et al., 2011; Epstein et al., 2014). Typical NEPC expresses a dominant and irreversible neuronal phenotype (Lin et al., 2014), which complicates attempts to delineate malignant (and therefore targetable) aspects of the disease. Importantly however, since the AR is either incidental to growth or no longer expressed at all, conventional CRPC therapies are redundant and platinum-based chemotherapy are effective, although relapse is rapid and overall survival remains poor (Aparicio et al., 2013).

A large body of evidence suggests that NEPC arises from prostatic adenocarcinoma cells via an adaptive process termed “neuroendocrine transdifferentiation” (Guo et al., 2011; Lotan et al., 2011; Williamson et al., 2011; Lin et al., 2014). However, even in the contemporary setting, less than a quarter of patients harbour NEPC foi at death, implying that only certain tumours are capable of attaining a proliferative NEPC state. Although prostatic adenocarcinoma cells are adapted to their primary niche, they are presumably poorly adapted to a neuronal niche that utilizes distinct mechanisms of innate tumour suppression. Therefore, to attain a proliferative NEPC tumour, additional and specific genomic perturbation to neuronal tumour suppressors and/or oncogenes is likely required (Fig 2). Emerging data suggest these predisposing aberrations include loss of RB1 and TP53, and gain of MYCN and AURKA (Beltran et al., 2011; Chen et al., 2012; Tan et al., 2014). Although identification of the latter led to initiation of a phase II trial of the AURKA inhibitor MLN8237 in NEPC (NCT01799278), there have been few novel leads for targeting this lethal disease variant, partly due to a paucity in preclinical models. Recently, a first-in-field patient-derived xenograft model of neuroendocrine transdifferentiation has been described (Lin et al., 2014). Molecular characterization of this model led to the discovery that PEG10, a placental gene, is de-repressed during the adaptive response to AR inhibition and highly up-regulated in clinical NEPC (Akamatsu et al., 2014). PEG10 is regulated by the AR and promotes growth and invasion of cancer cells in the context of RB1 and TP53 loss. Furthermore, since expression of PEG10 in adult tissue is extremely limited, it represents a strong candidate for therapeutic targeting. Discoveries such as AURKA, MYCN and PEG10 are likely to be the first of in a wave of mechanistic insights into the development of NEPC, and their critical role during progression suggests that the optimal strategy for select patients may rely on intervening prior to transformation to NEPC.

Future directions

It is plausible that the existing armamentarium of novel agents can elicit greater anti-tumour responses when used at earlier stages in disease, or in rational combination with each other. Indeed, strong responses have been observed after neo-adjuvant or front-line enzalutamide or abiraterone administration. Trials combining these two agents will also prove informative (NCT01650194, NCT00268476). However, genomic biomarkers of response and resistance emerging from prospective studies must be integrated into clinical trial design in order to improve chances of success. As described in this review, in the aftermath of cytotoxic and potent second-line AR-targeted agents, the CRPC landscape is riddled with aggressive, heterogeneous and adaptive clones. Therefore, to facilitate trial enrichment or optimal drug sequencing, patient tumours must be monitored for novel molecular changes. Unfortunately, logistical difficulties and significant morbidity have long precluded routine collection of mCRPC tissue biopsies. Even where possible, one is faced with tumour cellularity issues (particularly in bone metastases), and under-sampling of a single site amongst a highly heterogeneous ecosystem of metastases. Although remarkable efforts from Prostate Cancer Foundation/Stand Up To Cancer Dream Teams have established robust protocols for mCRPC tissue collection, the likelihood
for widespread adoption is low, and a more minimally invasive strategy is urgently required.

Since tumour material is continually shed into the bloodstream, liquid biopsy (extensively reviewed in Crowley et al., 2013; Diaz & Bardelli, 2014; Heitzer et al., 2013; Joosse et al., 2014) holds great promise for improving CRPC patient management. From a trial enrichment/biomarker perspective perhaps the most exciting developments have emerged from studies of tumour-derived cell-free DNA (cfDNA) extracted from patient plasma. Compared to circulating tumour cells (CTCs), cfDNA analyses provide a global survey of tumour status, and the recent application of targeted sequencing to cfDNA extracted temporally from 16 advanced prostate cancer patients was able to accurately monitor the dynamics of lethal tumour clones (Carreira et al., 2014). Allowing for the compromising dilution effect of “normal” cfDNA, relatively simple next-generation sequencing approaches are sufficient to robustly detect amplifications and mutations, making cfDNA analyses ideal for monitoring AR status longitudinally in CRPC patients. Indeed, a recent study used combination of copy number profiling and next-generation sequencing to identify AR amplifications and mutations in the cfDNA of mCRPC patients progressing on novel systemic agents (Azad et al., 2015b). Interestingly, AR alterations accompanied enzalutamide resistance, and the presence of pre-treatment AR amplifications or mutations were predictive for adverse outcomes on enzalutamide (Azad et al., 2015b; Gleave & Chi, 2015). CTCs will remain critical for basic and clinical research alike, due to the ability to profile the transcriptome (e.g. for truncated AR variants) and the potential of establishing cultures. However, the broad utility of
PENDING ISSUES

(i) Can we develop treatment predictive biomarkers that will allow optimal drug sequencing strategies tailored to individual patients?

(ii) How can we develop therapies for patients with AR-negative CRPC?

(iii) Will the new wave of novel AR-targeted therapies for CRPC prove even more effective when moved earlier in disease progression?

References

Adeniji AO, Chen M, Penning TM (2013) AKR1C3 as a target in castrate resistant prostate cancer. J Steroid Biochem Mol Biol 137: 136–149

Agoulnik IU, Vaid A, Nakka M, Alvarado M, Bingman WE III, Erdem H, Frolov A, Smith CL, Ayala GE, Ittmann MM (et al) (2006) Androgens modulate expression of transcription intermediary factor 2, an androgen receptor coactivator whose expression level correlates with early biochemical recurrence in prostate cancer. Cancer Res 66: 10594–10602

Akamatsu S, Wyatt A, Lin D, Lysakowski S, Zhang F, Kim S, Fazli L, Beltran H, Rubin M, Zoubedi A et al (2014) MP31-09 identification of a retinoid/paraoxonase derived gene associated with progression to neuroendocrine prostate cancer. J Urol 191: e325

Al Nakouzi N, Le Moulec S, Albiges L, Wang C, Beuzeboc P, Gross-Goupil M, De La Motte Rouge T, Guillot A, Gajda D, Massard C et al (2014) Cabazitaxel remains active in patients progressing after docetaxel followed by novel androgen receptor pathway targeted therapies. Eur Urol doi: 10.1016/euro.2014.04.015

Amato RJ, Wilding G, Buley G, Loeyv J, Haluska F, Gross ME (2012) Safety and preliminary efficacy analysis of the mTOR inhibitor ridaforolimus in patients with taxane-treated, castration-resistant prostate cancer. Clin Genitourin Cancer 10: 232–238

Amirante M, Luo JI, Grivennikov S, Nedospasov S, Karin M (2010) B-cell-derived lymphotixin promotes castration-resistant prostate cancer. Nature 464: 302–305

An J, Wang C, Deng Y, Yu L, Huang H (2014) Destruction of full-length androgen receptor by wild-type SPOP, but not prostate-cancer-associated mutants. Cell Rep 6: 657–669

Andersen RJ, Mawji NR, Wang J, Wang G, Haile S, Myung JK, Watt K, Tam T, Yang YC, Banuelos CA et al (2010) Regression of castrate-recurrent prostate cancer by a small-molecule inhibitor of the amino-terminal domain of the androgen receptor. Cancer Cell 17: 535–546

Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roesser JC, Chen Y, Mohammad TA, Chen Y, Fedor HL et al (2014) AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. N Engl J Med 371: 1028–1038

Aparicio AM, Harzstark AL, Corn PG, Wen S, Araujo JC, Tu SM, Pagliaro LC, Kim J, Millickan RE, Ryan C et al (2013) Platinum-based chemotherapy for variant castrate-resistant prostate cancer. Clin Cancer Res 19: 3621–3630

Arnold JT, Gray NE, Jacobowitz K, Viswanathan L, Cheung PW, McFann KK, Le H, Blackman MR (2008) Human prostate stromal cells stimulate increased PSA production in DHEA-treated prostate cancer epithelial cells. J Steroid Biochem Mol Biol 111: 240–246

Arora VK, Schenkein E, Murali R, Subudhi SK, Wongvipat J, Balbas MD, Shah N, Cai L, Efstathiou E, Logothetis C et al (2013) Glucocorticoid receptor confers resistance to androgeners by bypassing androgen receptor blockade. Cell 155: 1309–1322

Asangani IA, Dommetti VL, Wang X, Malik R, Cieslik M, Yang R, Escara-Wilke J, Wilder-Romans K, Dhanireddy S, Engelke C et al (2014) Therapeutic targeting of BET bromodomain proteins in castration-resistant prostate cancer. Nature 510: 278–282

Azad AA, Leibowitz-Amit R, Ejgl BJ, Lester R, Wells JC, Murray RN, Kollmannsberger C, Heng DV, Joshua AM, Chi KN (2014) A retrospective, Canadian multi-center study examining the impact of prior response to abiraterone acetate on efficacy of docetaxel in metastatic castration-resistant prostate cancer. Prostate 74: 1544–1550

Azad AA, Ejgl BJ, Murray RN, Kollmannsberger C, Chi KN (2015a) Efficacy of enzalutamide following abiraterone acetate in chemotherapy-naive metastatic castration-resistant prostate cancer patients. Eur Urol 67: 23–29

Azad AA, Volik SV, Wyatt AW, Haegert A, Le Bihan S, Bell RH, Anderson S, McConeghy B, Shukin R, Bazov J et al (2015b) Androgen receptor gene mutations in circulating cell-free DNA: biomarkers of therapeutic resistance in castration-resistant prostate cancer. Clin Cancer Res doi: 10.1158/1078-0432.CCR-14-2666

Azad AA, Zoubedi A, Gleave ME, Chi KN (2015c) Targeting heat shock proteins in metastatic castration-resistant prostate cancer. Nat Rev Urol 12: 26–36

Baca SC, Prandi D, Lawrence MS, Mosquera JM, Romanel A, Dier Y, Park K, Kitabayashi N, MacDonald TY, Chandl M et al (2013) Punctuated evolution of prostate cancer genomes. Cell 153: 666–677

Balbas MD, Evans MJ, Hosfield DJ, Wongvipat J, Arora VK, Watson PA, Chen Y, Greene GL, Shen Y, Sawyers CL (2013) Overcoming mutation-based resistance to androgeners with rational drug design. Elife 2: e00499

Barbier CE, Baca SC, Lawrence MS, Demichelles F, Blattner M, Theurillat JP, White SA, Stojanov P, Van Allen E, Stranksy N et al (2012) Exome sequencing identifies recurrent SPOP, FOXA1 and MED12 mutations in prostate cancer. Nat Genet 44: 685–689

Beer TM, Bernstein GT, Corman JM, Glode LM, Hall SJ, Poll WL, Schellhammer PF, Jones LA, Xu Y, Kylstra JW et al (2011) Randomized trial of autologous cellular immunotherapy with sipuleucel-T in androgen-dependent prostate cancer. Clin Cancer Res 17: 4558–4567

Beer TM, Armstrong Aj, Sternberg CN, Higano CS, Iversen P, Lorig Y, Rathkopf DE, Bhattacharya S, Carles J, De Bono JS et al (2014) Enzalutamide in men with chemotherapy-naive metastatic prostate cancer (mCRPC): results of phase III PREVAIL study. J Clin Oncol 32(Suppl. 4): abstr LBA1

Conflict of interest

The authors declare that they have no conflict of interest.
Beltran H, Rickman DS, Park K, Chae SS, Sboner A, MacDonald TY, Wang Y, Sheikh KL, Terry S, Tagawa ST et al. (2011) Molecular characterization of neuroendocrine prostate cancer and identification of new drug targets. Cancer Discov 1: 487 – 495

Beltran H, Tagawa ST, Park K, MacDonald T, Milowsky MI, Mosquera JM, Rubin MA, Nanus DM (2012) Challenges in recognizing treatment-related neuroendocrine prostate cancer. J Clin Oncol 30: e386 – e389

Beltran H, Yelensky R, Frampton GM, Park K, Downing SR, MacDonald TY, Jarosz M, Lipson D, Tagawa ST, Nanus DM et al. (2013) Targeted next-generation sequencing of advanced prostate cancer identifies potential therapeutic targets and disease heterogeneity. Eur Urol 63: 920 – 926

Bianchini D, Omlin A, Pezaro C, Lorente D, Ferraldeschi R, Mukherji D, Crespo M, Figueiredo I, Miranda S, Riisnaes R et al. (2013) First-in-human Phase I study of EZN-4176, a locked nucleic acid antisense oligonucleotide to exon 4 of the androgen receptor mRNA in patients with castration-resistant prostate cancer. Br J Cancer 109: 2579 – 2586

Bianchini D, Lorente D, Rodriguez-Vida A, Omlin A, Pezaro C, Ferraldeschi R, Zivi A, Attard G, Chowdhury S, de Bono JS (2014) Antitumour activity of enalutamide (MDV3100) in patients with metastatic castration-resistant prostate cancer (CRPC) pre-treated with docetaxel and abiraterone. Eur J Cancer 50: 78 – 84

de Bono JS, Oudard S, Ozuguroglu M, Hansen S, Machiels JP, Kokac I, Gravis G, Bodrogi I, Mackenzie MJ, Shen L et al. (2010) Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. Lancet 376: 1147 – 1154

de Bono JS, Logothetis CJ, Molina A, Fizazi K, North S, Chu L, Chi KN, Jones RJ, Goodman OB Jr, Saad F et al. (2011) Abiraterone and increased survival in metastatic prostate cancer. N Engl J Med 364: 1995 – 2005

Brand Lj, Olson ME, Ravindranathan P, Guo H, Kempema AM, Andrews TE, Chen X, Raj GV, Hariki DA, Dehm SM (2013) EPI-001 is a selective peroxisome proliferator-activated receptor-gamma modulator with inhibitory effects on androgen receptor expression and activity in prostate cancer. Oncotarget 6: 3811 – 3824

Brenner JC, Ateeq B, Li Y, Yocum AK, Cao Q, Asangani IA, Patel S, Wang X, Liang H, Yu J et al. (2013) Mechanistic rationale for inhibition of poly(ADP-ribose) polymerase in ETS gene fusion-positive prostate cancer. Cancer Cell 19: 664 – 678

Bubendorf L, Kononen J, Kivisto P, Schraml P, Moch H, Gasser TC, Willi N, Mihatsch MJ, Sauter G, Kallioniemi OP (1999) Survey of gene amplifications during prostate cancer progression by high-throughput fluorescence in situ hybridization on tissue microarrays. Cancer Res 59: 803 – 806

Cai C, Wang H, Xu Y, Chen S, Balk SP (2009) Reactivation of androgen receptor-regulated TMPRSS2:ERG gene expression in castration-resistant prostate cancer. Cancer Res 69: 6027 – 6032

Cai C, He HH, Chen S, Coleman I, Wang H, Fang Z, Chen S, Nelson PS, Liu XS, Brown M et al. (2011) Androgen receptor gene expression in prostate cancer is directly suppressed by the androgen receptor through recruitment of lysine-specific demethylase 1. Cancer Cell 20: 457 – 471

Cano LQ, Lavery DN, Bevan CL (2013) Mini-review: f Bodesumol: a breakdown of androgen receptor action in prostate cancer. Mol Cell Endocrinol 369: 52 – 62

Carducci MA, Saad F, Abrahamsson PA, Deanaley DP, Schulman CC, North SA, Sleep DJ, Isaacson JD, Nelson JB, Atrasentan Phase IIIISGI (2007) A phase 3 randomized controlled trial of the efficacy and safety of atrasentan in men with metastatic hormone-refractory prostate cancer. Cancer 110: 1959 – 1966

Carreira S, Romanel A, Goodall J, Grist E, Ferraldeschi R, Miranda S, Prandi D, Lorente D, Frenel JS, Pezaro C et al. (2014) Tumor clone dynamics in lethal prostate cancer. Sci Transl Med 6: 254ra125

Carver BS, Chapinski C, Wongvipat J, Hieronymous H, Chen Y, Chandarlapaty S, Arora VK, Ke C, Kouletcher J, Scher H et al. (2011) Reciprocal feedback regulation of PI3K and androgen receptor signaling in PTEN-deficient prostate cancer. Cancer Cell 19: 575 – 586

Chang KH, Li R, Kuri B, Lotan Y, Roehrborn CG, Liu J, Vessella R, Nelson PS, Kapur P, Guo X et al. (2013) A gain-of-function mutation in DHT synthesis in castration-resistant prostate cancer. Cell 154: 1074 – 1084

Chen CD, Welsbie DS, Tran C, Baek SH, Chen R, Vessella R, Rosenfeld MG, Sawyers CL (2004) Molecular determinants of resistance to antiandrogen therapy. Nat Med 10: 33 – 39

Chen H, Sun Y, Wu C, Magyar CE, Li X, Cheng L, Yao JL, Shen S, Osunkoya AO, Liang C et al. (2012) Pathogenesis of prostatic small cell carcinoma involves the inactivation of the PS5 pathway. Endocr Relat Cancer 19: 321 – 331

Chen Y, Chen J, Loo A, Jaeger S, Bagdasarian L, Yu J, Chung F, Korn J, Ruddy D, Guo R et al. (2013) Targeting HSFL1 sensitizes cancer cells to HSP90 inhibition. Oncotarget 4: 816 – 829

Chen E, Sowalsky AG, Gao S, Cai C, Voznesenskyy O, Schaefer R, Loda M, True LD, Ye H, Troncoso P et al. (2013) Abiraterone treatment in castration-resistant prostate cancer selects for progesterone responsive mutant androgen receptors. Clin Cancer Res 21: 1273 – 1280

Chi KN, Eisenhauer E, Fazioli L, Jones EC, Goldstein SL, Powers J, Tu D, Gleave ME (2005) A phase I pharmacokinetic and pharmacodynamic study of OGX-011, a 2′-methoxethyl antisense oligonucleotide to clusterin, in patients with localized prostate cancer. J Natl Cancer Inst 97: 1287 – 1296

Chi KN, Hotte SJ,Yu EY, Tu D, Eigi BJ, Tannock I, Saad F, North S, Powers J, Gleave ME et al. (2010) Randomized phase II study of docetaxel and prednisone with or without OGX-011 in patients with metastatic castration-resistant prostate cancer. J Clin Oncol 28: 4247 – 4254

Chi KN, Hotte SJ, Ellard S, Gingerich JR, Joshua AM, Yu EY, Gleave ME (2012) A randomized phase II study of OGX-427 plus prednisone (P) versus P alone in patients (pts) with metastatic castration-resistant prostate cancer (CRPC). J Clin Oncol 30(Suppl): abstract 4514

Chi K, Higano C, Reeves J, Feyerabend S, Gravis G, Ferrero J, Jacobs C, Barnett-Griness O, Pande A, de Bono J (2014) 7550A randomized phase 3 study comparing first-line docetaxel/prednisone (DP) to DP plus custirsen in men with metastatic castration-resistant prostate cancer (mCRPC). Ann Oncol 25: iv256 – iv256

Chmela R, Buchanan G, Need EF, Tilley W, Greenberg NM (2007) Androgen receptor coregulators and their involvement in the development and progression of prostate cancer. Int J Cancer 120: 719 – 733

Clegg NJ, Wongvipat J, Joseph JD, Tran C, Ouk S, Dilhas A, Chen Y, Grillot K, Bischoff ED, Cai I et al. (2012) ARN-509: a novel antiandrogen for prostate cancer treatment. Cancer Res 72: 1494 – 1503

Comstock CE, Augello MA, Goodwin JF, de Leeuw R, Schiewer MJ, Ostrander WF Jr, Burkhardt RA, McClendon AK, McCue PA, Trabulsi EJ et al. (2013) Targeting cell cycle and hormone receptor pathways in cancer. Oncogene 32: 5481 – 5491

Crowley E, Di Nicolantonio F, Lopukis F, Bardelli A (2013) Liquid biopsy: monitoring cancer-genetics in the blood. Nat Rev Clin Oncol 10: 472 – 484

Dai C, Whitesell L, Rogers AB, Lindquist S (1999) Heat shock factor 1 is a powerful multifaceted modifier of carcinogenesis. Cell 130: 1005 – 1018

Dalal K, Roshan-Moniri M, Sharma A, Li H, Ban F, Hessein M, Hsing M, Singh K, LeBlanc E, Dehm S et al. (2014) Selectively targeting the DNA-binding
domain of the androgen receptor as a prospective therapy for prostate cancer. J Biol Chem 289: 33877
Darshan MS, Loftus MS, Thadani-Mulero M, Levy BP, Escuin D, Zhou XK, Gyerzi A, Chanel-Vos C, Shen R, Tagawa ST et al (2011) Taxane-induced blockade to nuclear accumulation of the androgen receptor predicts clinical responses in metastatic prostate cancer. Cancer Res 71: 6019–6029
Dehm SM, Tindall DJ (2011) Alternatively spliced androgen receptor variants. Endocr Relat Cancer 18: R183–R196
Diaz LA Jr, Bardelli A (2014) Liquid biopsies: genotyping circulating tumor DNA. J Clin Oncol 32: 579–586
Disis ML, Goodell V, Schiffman K, Knutson KL (2014) Immoral epoetin-spreading following immunization with a HER-2/neu peptide based vaccine in cancer patients. J Clin Immunol 24: 571–578
Drake CG, Kwon ED, Fizazi K, Bossi A, van den Eertwegh AJ, Logothetis C, Scher HI, Beer TM, McHenry B, Liu D (2014) Results of subset analyses on overall survival (OS) from study CA184-043: ipilimumab (ipi) versus placebo (Pbo) in post-docetaxel metastatic castration-resistant prostate cancer (mCRPC). J Clin Oncol 32(Suppl. 4); abstr 2b
Dreicer R, Jones R, Oudard S, Efstathiou E, Saad F, De Wit R, De Bono J, Shi Y, Tejura B, Agus DB et al (2014) Results from a phase 3, randomized, double-blind, multicenter, placebo-controlled trial of orteronel (TAK-700) plus prednisone in patients with metastatic castration-resistant prostate cancer (mCRPC) that has progressed during or following docetaxel-based therapy (FLM-PC 5 trial). J Clin Oncol 32(Suppl. 4); abstr 7b
Epstein JI, Amin MB, Beltran H, Lotan TL, Mosquera JM, Reuter VE, Robinson BD, Troncoso P, Rubin MA (2014) Proposed morphologic classification of prostate cancer with neuroendocrine differentiation. Am J Surg Pathol 38: 756–767
Ferraldeschi R, Sharifi N, Auchus RJ, Attard G (2013) Molecular pathways: inhibiting steroid biosynthesis in prostate cancer. Clin Cancer Res 19: 3535–3559
Ferraldeschi R, Weito J, Luo J, Attard G, de Bono JS (2015) Targeting the androgen receptor pathway in castration-resistant prostate cancer: progresses and prospects. Oncogene 34: 1745–1757
Fizazi K, Scher HI, Molina A, Logothetis CJ, Chi KN, Jones RJ, Staffurth JN, North S, Vogelzang NJ, Saad F et al (2012) Abiraterone acetate for treatment of metastatic castration-resistant prostate cancer: final overall survival of the COU-AA-301 randomised, double-blind, placebo-controlled phase 3 study. Lancet Oncol 13: 983–992
Fizazi K, Massard C, Bono P, Jones R, Kataja V, James N, Garcia JA, Protheroe A, Tammela TL, Elliott T et al (2014) Activity and safety of ODM-201 in patients with progressive metastatic castration-resistant prostate cancer (ARADES): an open-label phase 1 dose-escalation and randomised phase 2 dose expansion trial. Lancet Oncol 15: 975–985
Galletti G, Matov A, Beltran H, Fontugne J, Miguel Mosquera J, Cheung C, MacDonald TY, Sury M, O’Toole S, Kench JG et al (2014) ERG induces taxane resistance in castration-resistant prostate cancer. Nat Commun 5: 5548
Gan L, Chen S, Wang Y, Watahiki A, Bohrer L, Sun Z, Wang Y, Huang H (2009) Inhibition of the androgen receptor as a novel mechanism of taxol chemotherapy in prostate cancer. Cancer Res 69: 8386–8394
Garrido C, Brunet M, Didelet C, Zermati Y, Schmitt E, Kroemer G (2006) Heat shock proteins 27 and 70: anti-apoptotic proteins with tumorigenic properties. Cell Cycle 5: 2592–3601
Geng C, He B, Xu L, Barbieri CE, Eedunuri VK, Chew SA, Zimmermann M, Bond R, Shou J, Li C et al (2013) Prostate cancer-associated mutations in speckle-type POZ protein (SPOP) regulate steroid receptor coactivator 3 protein turnover. Proc Natl Acad Sci USA 110: 6997–7002
Geng C, Rajapakse K, Shah SS, Shou J, Eedunuri VK, Foley C, Fiskus W, Rajendran M, Chew SA, Zimmermann M et al (2014) Androgen receptor is the key transcriptional mediator of the tumor suppressor SPOP in prostate cancer. Cancer Res 74: 5631–5643
Gibbs A, Schwartzman J, Deng V, Alumkal J (2009) Sulforaphane destabilizes the androgen receptor in prostate cancer cells by inactivating histone deacetylase 6. Proc Natl Acad Sci USA 106: 16663–16668
Gleave M, Chi K (2015) Toward predictive signatures of enzalutamide response and resistance. Eur Urol 67: 61–63
Gormally MV, Dexheimer TS, Marisco G, Sanders DA, Lowe C, Matak-Vinkovic D, Michael S, Jadhav A, Rai C, Maloney DJ et al (2014) Suppression of the FOXM1 transcriptional programme via novel small molecule inhibition. Nat Commun 5: 5165
Gottlieb B, Beitel KL, Nadarajah A, Paliouras M, Trifiro M (2012) The androgen receptor gene mutations database: 2012 update. Hum Mutat 33: 887–894
Grasso CS, Wu YM, Robinson DR, Cao X, Dhanasekaran SM, Khan AP, Quist MJ, Jing X, Longiro RJ, Brenner JC et al (2012) The mutational landscape of lethal castration-resistant prostate cancer. Nature 487: 239–243
Gregory CW, He B, Johnson RT, Ford OH, Mohler JL, French FS, Wilson EM (2001) A mechanism for androgen receptor-mediated prostate cancer recurrence after androgen deprivation therapy. Cancer Res 61: 4315–4319
Guilley J, Arlen PM, Madan RA, Tsang KY, Pazdur MP, Skarupa L, Jones J, Poole DJ, Higgins JP, Hodge JW et al (2010) Immunologic and prognostic factors associated with overall survival employing a povxiral-based PSA vaccine in metastatic castrate-resistant prostate cancer. Cancer Immunol Immunother 59: 663–674
Guilley J, Madan RA, Stein WD, Wilkerson J, Dahut WL, Heery CR, Schliom J, Wilding G, DiPaola RS (2013) Effect of PSA-tricom, a pox-viral vaccine in prostate cancer (PCa), on tumor growth rates within 80 days after initiation in nonmetastatic PCa. J Clin Oncol 31(Suppl. 6); abstr 57
Guo CC, Dancer JY, Wang Y, Aparicio A, Navone NM, Troncoso P, Czemik IA (2011) TMRPR52-ERG gene fusion in small cell carcinoma of the prostate. Hum Pathol 42: 11–17
Handratta VD, Vasaitis TS, Njar VC, Gediya K, Kataria R, Chopra P, Newman D Jr, Farquhar R, Guo Z, Qiu Y et al (2009) Novel C-17-heteroaryl steroid CYP17 inhibitors/antiandrogens: synthesis, in vitro biological activity, pharmacokinetics, and antitumor activity in the LAPC4 human prostate xenograft model. J Med Chem 48: 2972–2984
He B, Lanz RB, Fiskus W, Geng C, Yi P, Hartig SM, Rajapakse K, Shou J, Wei L, Shah SS et al (2014) GATA2 facilitates steroid receptor coactivator recruitment to the androgen receptor complex. Proc Natl Acad Sci USA 111: 18261–18266
Heath EI, Hillman DW, Vaishampayan U, Sheng S, Sarkar F, Harper F, Gaskins M, Pitot HC, Tan W, Ivy SP et al (2008) A phase II trial of 17-allylamino-17-demethoxygeldanamycin in patients with hormone-refractory metastatic prostate cancer. Clin Cancer Res 14: 7940–7946
Heath EI, Stein MN, Vaishampayan UN, Antonarakis ES, Liu G, Sheng S, Farrow K, Smith DW, Heilbrun LK (2013) Phase II trial of single-agent ganetespib (STA-9090), a heat shock protein 90 (Hsp90) inhibitor in heavily pretreated patients with metastatic castration-resistant prostate cancer (mCRPC) post docetaxel-based chemotherapy: results of a Prostate Cancer Clinical Trials Consortium (PCCCT) study. J Clin Oncol 31(Suppl.): abstr 5085
Heemers HV, Regan KM, Schmidt LJ, Anderson SK, Ballman KV, Tindall DJ (2009) Androgen modulation of coregulator expression in prostate cancer cells. Mol Endocrinol 23: 572–583
Heitzer E, Auer M, Uiz P, Geigl JB, Speicher MR (2013) Circulating tumor cells and DNA as liquid biopsies. Genome Med 5: 73
Huggins C, Stevens R, Hodges CV (1941) Studies on prostatic cancer: II. The effects of castration on advanced carcinoma of the prostate gland. Arch Surg 43: 209 – 223

James ND, Caty A, Borre M, Zonnenberg BA, Beuzechoc P, Morris T, Phung D, Dawson NA (2009) Safety and efficacy of the specific endothelin-A receptor antagonist ZD4054 in patients with hormone-resistant prostate cancer and bone metastases who were pain free or mildly symptomatic: a double-blind, placebo-controlled, randomised, phase 2 trial. Eur Urol 55: 1112 – 1123

James ND, Caty A, Payne H, Borre M, Zonnenberg BA, Beuzechoc P, McIntosh S, Morris T, Phung D, Dawson NA (2010) Final safety and efficacy analysis of the specific endothelin A receptor antagonist zibotentan (ZD4054) in patients with metastatic castration-resistant prostate cancer and bone metastases who were pain-free or mildly symptomatic for pain: a double-blind, placebo-controlled, randomised Phase II trial. BJU Int 106: 966 – 973

Jin HJ, Zhao JW, Ogden I, Bergan RC, Yu J (2013) Androgen receptor-independent function of FOXL1 in prostate cancer metastasis. Cancer Res 73: 3725 – 3736

Jochems C, Tucker JA, Tsang KY, Madan RA, Dahut WL, Liewehr DJ, Steinberg SM, Gulley JL, Schlojm J (2014) A combination trial of vaccine plus ipilimumab in metastatic castration-resistant prostate cancer patients: immune correlates. Cancer Immunol Immunother 63: 407 – 418

Joosse SA, Gorges TM, Pantel K (2014) Biology, detection, and clinical implications of circulating tumor cells. EMBO Mol Med 7: 1 – 11

Joseph JD, Lu N, Qian J, Sensintaffar J, Shao G, Brigham D, Moon M, Maneval SM, Baskin-Bey E, Heeringa M, Ouatas T (2014) Ipilimumab in metastatic castration-resistant prostate cancer patients: a phase II study evaluating the toxicity and efficacy of single-agent ipilimumab versus placebo after radiotherapy in patients with metastatic castration-resistant prostate cancer that had progressed after docetaxel chemotherapy (CA184-043): a multicentre, randomised, double-blind, phase 3 trial. Lancet Oncol 15: 700 – 712

Lamoureux F, Thomas C, Yin MJ, Kuruma H, Beraldi E, Fazli L, Zoubidei A, Gleave ME (2011a) Clusterin inhibition using OXG-011 synergistically enhances Hsp90 inhibitor activity by suppressing the heat shock response in castrate-resistant prostate cancer. Cancer Res 71: 5838 – 5849

Lamoureux F, Thomas C, Yin MJ, Kuruma H, Fazli L, Gleave ME, Zoubidei A (2011b) A novel HSP90 inhibitor delays castrate-resistant prostate cancer without altering serum PSA levels and inhibits osteoclastogenesis. Clin Cancer Res 17: 2301 – 2313

Lamoureux F, Thomas C, Yin MJ, Fazli L, Zoubidei A, Gleave ME (2014) Suppression of heat shock protein 27 using OXG-427 induces endoplasmic reticulum stress and potentiates heat shock protein 90 inhibitors to delay castrate-resistant prostate cancer. Eur Urol 66: 145 – 155

de Leeuw R, de Comstock C, de Pollutri D, Schwierj MJ, Ciment S, de Mankame T, Ostrander W, Den RB, Kelly WK, Gomella LG et al (2014) Leveraging RB status to define therapy for castrate-resistant prostate cancer. j Clin Oncol 32(Suppl. 4): abstr 96

de Leeuw R, Berman-Booty LD, Schwierj MJ, Ciment S, Den RB, Dicker AP, Kelly WK, Trabulsi EJ, Lallas CD, Gomella LG et al (2015) Novel actions of next-generation taxanes benefit advanced stages of prostate cancer. Clin Cancer Res 21: 795 – 807

Li Y, Alsaagbi M, Fan D, Bova GS, Tewfik AH, Dehm SM (2011) Intragenic rearrangement and altered RNA splicing of the androgen receptor in a cell-based model of prostate cancer progression. Cancer Res 71: 2108 – 2117

Li Y, Hwang TH, Oseth LA, Hauge A, Vessella RL, Schmechel SC, Hirsch B, Beckman KB, Silverstein KA, Dehm SM (2012) AR intragenic deletions linked to androgen receptor splice variant expression and activity in models of prostate cancer progression. Oncogene 31: 4759 – 4767

Lin D, Wyatt AW, Xue H, Wang Y, Dong X, Haegert A, Wu R, Brahmbhatt S, Mo F, Jong L et al (2014) High fidelity patient-derived xenografts for accelerating prostate cancer discovery and drug development. Cancer Res 74: 1272 – 1283

Locke JA, Guns ES, Lubik AA, Adomat HH, Hendy SC, Wood CA, Ettinger SL, Gleave ME, Nelson CC (2008) Androgen levels increase by intratumoral de novo steroidogenesis during progression of castration-resistant prostate cancer. Cancer Res 68: 6407 – 6415

Lorentz D, De Bono JS (2014) Molecular alterations and emerging targets in castration resistant prostate cancer. Eur J Cancer 50: 753 – 764

Lotier J, Fizzi K, Jones RJ, Van den Brande J, Molife RL, Omlin A, James ND, Baskin-Bey E, Heeringa M, Baron B et al (2014) Safety, tolerability and inducible up-regulation of intratumoral androgen biosynthesis and androgen receptor expression in an orthotopic VCaP human prostate cancer xenograft model. Am J Pathol 184: 2163 – 2173

Korpal M, Korn JM, Gao X, Rakiec DP, Ruddy DA, Doshi S, Yuan J, Kovats SG, Kim S, Cooke VG et al (2013) An F876L mutation in androgen receptor confers genetic and phenotypic resistance to MDV3100 (enzalutamide). Cancer Discov 3: 1030 – 1043

Kruczek K, Ratterman M, Toliokz J, Sulo S, Lestimg M, Nabban C (2013) A phase II study evaluating the toxicity and efficacy of single-agent temsirolimus in chemotherapy-naive castration-resistant prostate cancer. Br J Cancer 109: 1711 – 1716

Kwon ED, Drake CG, Scher HI, Fizazi K, Bossi A, van den Eertwegh AJ, Krainer M, Houdée N, Santos R, Mahahmed H et al (2014) Ipilimumab versus placebo after radiotherapy in patients with metastatic castration-resistant prostate cancer that had progressed after docetaxel chemotherapy (CA184-043): a multicentre, randomised, double-blind, phase 3 trial. Lancet Oncol 15: 700 – 712

Lamoureux F, Thomas C, Yin MJ, Kuruma H, Beraldi E, Zoubidei A, Gleave ME (2011a) Clusterin inhibition using OXG-011 synergistically enhances Hsp90 inhibitor activity by suppressing the heat shock response in castrate-resistant prostate cancer. Cancer Res 71: 5838 – 5849

Lamoureux F, Thomas C, Yin MJ, Kuruma H, Fazli L, Gleave ME, Zoubidei A (2011b) A novel HSP90 inhibitor delays castrate-resistant prostate cancer without altering serum PSA levels and inhibits osteoclastogenesis. Clin Cancer Res 17: 2301 – 2313

Lamoureux F, Thomas C, Yin MJ, Fazli L, Zoubidei A, Gleave ME (2014) Suppression of heat shock protein 27 using OXG-427 induces endoplasmic reticulum stress and potentiates heat shock protein 90 inhibitors to delay castrate-resistant prostate cancer. Eur Urol 66: 145 – 155

de Leeuw R, de Comstock C, de Pollutri D, Schierj MJ, Ciment S, de Mankame T, Ostrander W, Den RB, Kelly WK, Gomella LG et al (2014) Leveraging RB status to define therapy for castrate-resistant prostate cancer. j Clin Oncol 32(Suppl. 4): abstr 96

de Leeuw R, Berman-Booty LD, Schwierj MJ, Ciment S, Den RB, Dicker AP, Kelly WK, Trabulsi EJ, Lallas CD, Gomella LG et al (2015) Novel actions of next-generation taxanes benefit advanced stages of prostate cancer. Clin Cancer Res 21: 795 – 807

Li Y, Alsaagbi M, Fan D, Bova GS, Tewfik AH, Dehm SM (2011) Intragenic rearrangement and altered RNA splicing of the androgen receptor in a cell-based model of prostate cancer progression. Cancer Res 71: 2108 – 2117

Li Y, Hwang TH, Oseth LA, Hauge A, Vessella RL, Schmechel SC, Hirsch B, Beckman KB, Silverstein KA, Dehm SM (2012) AR intragenic deletions linked to androgen receptor splice variant expression and activity in models of prostate cancer progression. Oncogene 31: 4759 – 4767

Lin D, Wyatt AW, Xue H, Wang Y, Dong X, Haegert A, Wu R, Brahmbhatt S, Mo F, Jong L et al (2014) High fidelity patient-derived xenografts for accelerating prostate cancer discovery and drug development. Cancer Res 74: 1272 – 1283

Locke JA, Guns ES, Lubik AA, Adomat HH, Hendy SC, Wood CA, Ettinger SL, Gleave ME, Nelson CC (2008) Androgen levels increase by intratumoral de novo steroidogenesis during progression of castration-resistant prostate cancer. Cancer Res 68: 6407 – 6415
anti-tumour activity of the androgen biosynthesis inhibitor ASP9521 in patients with metastatic castration-resistant prostate cancer: multi-centre phase I/II study. Invest New Drugs: 32: 995 – 1004
Lotan TL, Gupta NS, Wang W, Toubaji A, Haffner MC, Chaux A, Hicks JL, Meeker AK, Bieberich CJ, De Marzo AM et al (2011) ERG gene rearrangements are common in prostatic small cell carcinomas. Mod Pathol: 24: 820 – 828
Luo JL, Tan W, Ricono JM, Korchynskyi O, Zhang M, Gonias SL, Cheres DA, Karin M (2007) Nuclear cytokine-activated IKKalpha controls prostate cancer metastasis by repressing Mapsin. Nature: 446: 690 – 694
Madan RA, Gulley JL, Kantoff PW (2013) Demystifying immunotherapy in prostate cancer: understanding current and future treatment strategies. Cancer: 19: 50 – 58
Makkouk A, Weiner CJ (2015) Cancer immunotherapy and breaking immune tolerance: new approaches to an old challenge. Cancer Res: 75: 5 – 10
Mathas S, Mistelli T (2009) The dangers of transcription. Cell: 139: 1047 – 1049
Matsumoto H, Yamamoto Y, Shiota M, Kuruma H, Beraldi E, Matsuya H, Zoubeidi A, Gleave M (2013) Cotargeting androgen receptor and clustin delays castrate-resistant prostate cancer progression by inhibiting adaptive stress response and AR stability. Cancer Res: 73: 5206 – 5217
May KF Jr, Gulley JL, Drake CG, Dranoff G, Kantoff PW (2011) Prostate cancer immunotherapy. Clin Cancer Res: 17: S233 – S238
Mellinghoff IK, Vivanco I, Kwon A, Tran C, Wongvipat J, Sawyers CL (2004) HER2/neu kinase-dependent modulation of androgen receptor function through effects on DNA binding and stability. Cancer Cell: 6: S17 – S27
Mezynski J, Pezaro C, Bianchini D, Zivi A, Sandhu S, Thompson E, Hunt J, Sheridan E, Baikady B, Sarvadikar A et al (2012) Antitumour activity of docetaxel following treatment with the CYP17A1 inhibitor abiraterone: clinical evidence for cross-resistance? Ann Oncol: 23: 2943 – 2947
Michaelson MD, Oudard S, Ou YC, Sengelov L, Saad F, Houede N, Ostler P, Stenzl A, Daugaard C, Jones R et al (2014) Randomized, placebo-controlled, phase III trial of sunitinib plus prednisone versus prednisone alone in progressive, metastatic, castration-resistant prostate cancer. J Clin Oncol: 32: 76 – 82
Mitsiades N, Sung CC, Schultz N, Danila DC, He B, Eedunuri VK, Fleisher M, Sander C, Sawyers CL, Scher HI (2012) Distinct patterns of dysregulated expression of genes involved in androgen synthesis and metabolism in metastatic prostate cancer tumors. Cancer Res: 72: 6142 – 6152
Mizokami A, Koh E, Izumi K, Narimoto K, Takeda M, Honma S, Dai J, Keller ET, Namiki M (2009) Prostate cancer stromal cells and LNCaP cells coordinate activation of the androgen receptor through synthesis of testosterone and dihydrotestosterone from dehydroepiandrosterone. Endocr Relat Cancer: 16: 1139 – 1155
Montgomery RB, Mostaghel EA, Vessella R, Hess DL, Kalhorn TF, Higano CS, True LD, Nelson PS (2008) Maintenance of intratumoral androgens in metastatic prostate cancer: a mechanism for castration-resistant tumor growth. Cancer Res: 68: 4447 – 4454
Mostaghel EA, Page ST, Lin DW, Fazio L, Coleman IM, True LD, Knudsen B, Hess DL, Nelson CC, Matsumoto AM et al (2007) Intraprostatic androgens and androgen-regulated gene expression persist after testosterone suppression: therapeutic implications for castration-resistant prostate cancer. Cancer Res: 67: 5033 – 5041
Mulholland DJ, Tran LM, Li Y, Cai H, Morim A, Wang S, Plassier S, Carraway IP, Huang J, Graebner TG et al (2011) Cell autonomous role of PTEN in regulating castration-resistant prostate cancer growth. Cancer Cell: 19: 792 – 804
 Munuganti RS, Hassona MD, Leblanc E, Frewin K, Singh K, Ma D, Ban F, Hsing M, Adomat H, Lalious N et al (2014) Identification of a potent antiandrogen that targets the BF3 site of the androgen receptor and inhibits enzalutamide-resistant prostate cancer. Chem Biol: 21: 1476 – 1485
Munuganti RS, Hassona MD, Leblanc E, Frewin K, Singh K, Ma D, Ban F, Hsing M, Adomat H, Lalious N et al (2014) Identification of a potent antiandrogen that targets the BF3 site of the androgen receptor and inhibits enzalutamide-resistant prostate cancer. Chem Biol: 21: 1476 – 1485
Myung JK, Banuelos CA, Fernandez JG, Mawji NR, Wang J, Tien AH, Yang YC, Tavakoli I, Haile S, Watt K et al (2013) An androgen receptor N-terminal domain antagonist for treating prostate cancer. J Clin Investig: 123: 2948 – 2960
Nelson JF, Fizazi K, Miller K, Higano C, Moul JW, Akaza H, Morris T, McIntosh S, Pemberton K, Gleave M (2012) Phase 3, randomized, placebo-controlled study of zibotentan (ZD4054) in patients with castration-resistant prostate cancer metastatic to bone. Cancer: 118: 5709 – 5718
Nelson AW, Tilley WD, Neal DE, Carroll JS (2014) Estrogen receptor beta in prostate cancer: friend or foe? Endocr Relat Cancer: 21: 7219 – 7244
Noonan KL, North S, Bitting RL, Armstrong AJ, Ellard SL, Chi KN (2015) Clinical activity of abiraterone acetate in patients with metastatic castration-resistant prostate cancer progressing after enzalutamide. Ann Oncol: 24: 1802 – 1807
Nyquist MD, Li Y, Hwang TH, Manlove LS, Vessella RL, Silverstein KA, Voytas DF, Dehm SM (2013) TALEN-engineered AR gene rearrangements reveal endocrine uncoupling of androgen receptor in prostate cancer. Proc Natl Acad Sci USA: 110: 17492 – 17497
Pacey S, Wilson RH, Walton M, Eatock MM, Hardcastle A, Zetterlund A, Arkenau HT, Moreno-Farre J, Banerji U, Roels B et al (2011) A phase I study of the heat shock protein 90 inhibitor alvespimycin (17-DMAG) given intravenously to patients with advanced solid tumors. Cancer Clin Oncol: 17: 1561 – 1570
Parker C, Nilsson S, Heinrich D, Helle SJ, O’Sullivan JM, Fossa SD, Chodacki A, Wiechno P, Logue J, Seke M et al (2013) Alpha emitter radium-223 and survival in metastatic prostate cancer. N Engl J Med: 369: 213 – 223
Petrylak D, Fizazi K, Stemberg C, Budnik N, De Wit R, Wiechno P, Bellmunt J, Barton D, Fandi A, Jungnelius U et al (2012) A Phase 3 study to evaluate the efficacy and safety of docetaxel and prednisone (DP) with or without lanalolamide (LEN) in patients with castrate-resistant prostate cancer (CRPC): the MAINSAIL trial. esMO conference 2012, LBA24
Pezaro CJ, Omlin A, Lorente D, Nava Rodrigues F, Ferraldeschi R, Bianchini D, Mukherji D, Riisnaes R, Altavilla A, Crespo M et al (2014a) Visceral disease in metastatic prostate cancer: friend or foe? J Clin Oncol: 32: 76 – 82
Pezaro CJ, Omlin A, Lorente D, Nava Rodrigues F, Ferraldeschi R, Bianchini D, Mukherji D, Riisnaes R, Altavilla A, Crespo M et al (2014b) Visceral disease in metastatic prostate cancer. Eur Urol: 65: 270 – 273
Pezaro CJ, Omlin A, Altavilla A, Lorente D, Ferraldeschi R, Bianchini D, Deearmorey D, Parker C, de Bono JS, Attard G (2014b) Activity of cabazitaxel in castration-resistant prostate cancer progressing after docetaxel and next-generation endocrine agents. Eur Urol: 66: 459 – 465
Polkinghorn WR, Parker JS, Lee MX, Kass EM, Spratt DE, Iaquinta PJ, Arora VK, Yen WF, Cai L, Zheng D et al (2013) Androgen receptor signaling regulates DNA repair in prostate cancers. Cancer Discov: 3: 1245 – 1253
Pritchard CC, Morrisse C, Kumar A, Zhang X, Smith C, Coleman I, Salipante SJ, Milbank J, Yu M, Grady WM et al (2014) Complex MSH2 and MSH6 mutations in hypermutated microsatellite unstable advanced prostate cancer. Nat Commun: 5: 4988
Qin J, Lee HJ, Wu SP, Lin SC, Lanz RB, Creighton CJ, deMeayo FJ, Tsai SY, Tsai MJ (2014) Androgen deprivation-induced NCoA2 promotes metastatic and castration-resistant prostate cancer. J Clin Investig: 124: 5013 – 5026
Rathkopf DE, Antonarakis ES, Shore ND, Tuttone R, Alumkal J, Ryan CJ, Saleh M, Hauke R, Chow Maneval E, Scher HI (2013a) ARN-509 in men with metastatic castration-resistant prostate cancer (mCRPC). J Clin Oncol: 31: Suppl (6): abstr 48
Rathkopf DE, Morris MJ, Fox JJ, Danila DC, Slovin SF, Hager JH, Rix PJ, Chow Maneval E, Chen I, Conen M et al (2013b) Phase I study of ARN-509, a novel antiandrogen, in the treatment of castration-resistant prostate cancer. J Clin Oncol: 31: 3525 – 3530
Ravindranathan P, Lee TK, Yang L, Centenera MM, Butler L, Tilley WD, Hsieh JT, Ahn JM, Raj GV (2013) Peptidomimetic targeting of critical androgen receptor-coregulator interactions in prostate cancer. Nat Commun 4: 1923

Rocchi P, Beraldi E, Ettinger S, Fazli L, Vessella RL, Nelson C, Gleave M (2005) Increased Hsp27 after androgen ablation facilitates androgen-independent progression in prostate cancer via signal transducers and activators of transcription 3-mediated suppression of apoptosis. Cancer Res 65: 11083 – 11093

Rodriguez-Vida A, Bianchini D, Van Hemelrijck M, Hughes S, Malik Z, Powles T, Bahal A, Rudman S, Payne H, de Bono J et al (2015) Is there an antiandrogen withdrawal syndrome with enzalutamide? BJU Int 115: 373 – 380

Ryan CJ, Smith MR, de Bono JS, Molina A, Logothetis CJ, de Souza P, Fizazi K, Mainwaring P, Piulats JM, Ng S et al (2013) Abiraterone in metastatic prostate cancer without previous chemotherapy. N Engl J Med 368: 138 – 148

Saad F, Fizazi K, Jinga V, Efstathiou E, Fong PC, Hart LL, Jones R, McDermott R, Rürth M, Suzuki K et al (2015) Orteronel plus prednisone in patients with chemotherapy-naive metastatic castration-resistant prostate cancer (ELM-PC-4): a double-blind, multicentre, phase 3, randomised, placebo-controlled trial. Lancet Oncol 16: 338 – 348

Sahu B, Laakso M, Pihlajamaa P, Ovaska K, Lopes JR, Porola P, Ma G, Korhonen M, Konttinen YT (2012) Increased Hsp27-mediated suppression of apoptosis. Cancer Res 72: 1570 – 1580

Sandhu SK, Schelman WR, Wilding C, Moreno V, Baird RD, Miranda S, Rocchi P, Beraldi E, Ettinger S, Fazli L, Vessella RL, Nelson C, Gleave M (2008) Cluzetin knockdown using the antisense oligonucleotide OGX-011 re-sensitizes docetaxel-refractory prostate cancer PC-3 cells to chemotherapy. BJU Int 102: 389 – 397

Sridhar SS, Freedland SJ, Gleave ME, Higano C, Mulders P, Parker C, Santor O, Saad F (2014) Castration-resistant prostate cancer: from new pathophysiology to new treatment. Eur Urol 65: 289 – 299

Stanbrough M, Bubley GJ, Ross K, Colub TR, Rubin MA, Penning TM, Febo PG, Balk SP (2006) Increased expression of genes converting adrenal androgens to testosterone in androgen-independent prostate cancer. Cancer Res 66: 2815 – 2825

Sun Y, Campisi J, Higano C, Beer TM, Porter P, Coleman I, True L, Nelson PS (2012) Treatment-induced damage to the tumor microenvironment promotes prostate cancer therapy resistance through WNT16B. Nat Med 18: 1359 – 1368

Sun Y, Nelson PS (2012) Molecular pathways: involving microenvironment damage responses in cancer therapy resistance. Clin Cancer Res 18: 4019 – 4025

Tan HL, Sood A, Rahimi HA, Wang W, Gupta N, Hicks J, Mosier S, Coke G, Epstein JJ, Netto GJ et al (2014) Rb loss is characteristic of prostatic small cell neuroendocrine carcinoma. Clin Cancer Res 20: 890 – 903

Tannock IF, de Wit R, Berry WR, Horti J, Pizianska A, Chi KN, Oudard S, Theodore C, James ND, Turesson I et al (2004) Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. N Engl J Med 351: 1502 – 1512

Tannock IF, Fizazi K, Ivanov S, Karlsson CT, Flechon A, Skoneczna I, Orandi F, Gravis C, Matveev V, Babvek S et al (2013) Affiblercept versus placebo in combination with docetaxel and prednisone for treatment of men with metastatic castration-resistant prostate cancer (VENICE): a phase 3, double-blind randomised trial. Lancet Oncol 14: 760 – 768

Taplin M, Chu F, Morrison J, Pili R, Rettig M, Stephenson J, Vogelzang N, Montgomery R (2012) ARMTOR1: safety of galeterone (TOK-001) in a phase 1 clinical trial in chemotherapy naive patients with castration resistant prostate cancer (CRPC). Cancer Res 72: CT – 07

Taylor BS, Schultz N, Hieronymus H, Copalain A, Xiao Y, Carver BS, Arora VK, Kaushik P, Cerami E, Reva B et al (2010) Integrative genomic profiling of human prostate cancer. Cancer Cell 18: 11 – 22

Templeton AJ, Dutoit V, Cathamos R, Rothermundt C, Bartschi D, Droge C, Gautschi O, Borne M, Fecther E, Stenner F et al (2013) Phase 2 trial of single-agent everolimus in chemotherapy-naive patients with castration-resistant prostate cancer (SAKK 08/08). Eur Urol 64: 150 – 158

Thomas C, Lamoureux F, Crafter C, Davies BR, Beraldi E, Fazli L, Kim S, Thaper D, Gleave ME, Zoubedi A (2013) Synergistic targeting of PI3K/AKT pathway and androgen receptor axis significantly delays castration-resistant prostate cancer progression in vivo. Mol Cancer Ther 12: 2342 – 2355

Toren PJ, Kim S, Pham S, Mangalji A, Adomat H, Guns ES, Zoubedi A, Moore W, Gleave ME (2015) Anticancer activity of a novel selective CYP17A1 inhibitor in preclinical models of castrate-resistant prostate cancer. Mol Cancer Ther 14: 59 – 69
The adaptive molecular landscape of CRPC

Alexander W Wyatt & Martin E Gleave

Tran C, Ouk S, Clegg NJ, Chen Y, Watson PA, Arora V, Wongvipat J, Smith-Jones PM, Yoo D, Kwon A et al (2009) Development of a second-generation antiandrogen for treatment of advanced prostate cancer. Science 324: 787 – 790

Wang S, Gao J, Lei Q, Rozengurt N, Pritchard C, Jiao J, Thomas GV, Li G, Roy-Burman P, Nelson PS et al (2003) Prostate-specific deletion of the murine Pten tumor suppressor gene leads to metastatic prostate cancer. Cancer Cell 4: 209 – 221

Wang S, Kollipara KR, Srivastava N, Li R, Ravindranathan P, Hernandez E, Freeman E, Humphries CG, Kapur P, Lotan Y et al (2014) Ablation of the oncogenic transcription factor ERG by deubiquitinase inhibition in prostate cancer. Proc Natl Acad Sci USA 111: 4251 – 4256

Whang YE, Armstrong AJ, Rathmell WK, Godley PA, Kim WY, Pruthi RS, Wallen EM, Crane JM, Moore DT, Grigson G (2013) A phase II study of lapatinib, a dual EGFR and HER-2 tyrosine kinase inhibitor, in patients with castration-resistant prostate cancer. Urol Oncol 31: 82 – 86

Williamson SR, Zhang S, Yao JI, Huang J, Lopez-Beltran A, Shen S, Osunkoya AO, MacLennan GT, Montironi R, Cheng L (2011) ERG-TMPRSS2 rearrangement is shared by concurrent prostatic adenocarcinoma and prostatic small cell carcinoma and absent in small cell carcinoma of the urinary bladder: evidence supporting monoclonal origin. Mod Pathol 24: 1120 – 1127

Wolf DA, Herzinger T, Hermeking H, Blaschke D, Horz W (1993) Transcriptional and posttranscriptional regulation of human androgen receptor expression by androgen. Mol Endocrinol 7: 924 – 936

Wyatt AW, Mo F, Wang K, McConathy B, Brahmbhatt S, Jong L, Mitchell DM, Johnston RL, Haegert A, Li E et al (2014) Heterogeneity in the inter-tumor transcriptome of high risk prostate cancer. Genome Biol 15: 426

Xu J, Wu RC, O’Malley BW (2009) Normal and cancer-related functions of the p160 steroid receptor co-activator (SRC) family. Nat Rev Cancer 9: 615 – 630

Yamamoto Y, Loriot Y, Beraldi E, Zhang F, Wyatt AW, Al Nakouzi N, Mo F, Zhou T, Kim Y, Monia BP et al (2015) Generation 2.5 antisense oligonucleotides targeting the androgen receptor and its splice variants suppress enzalutamide resistant prostate cancer cell growth. Clin Cancer Res 21: 1675 – 1687

Yan J, Erdem H, Li R, Cai Y, Ayala G, Ittmann M, Yu-Lee LY, Tsai SY, Tsai MJ (2008) Steroid receptor coactivator-3/AIB1 promotes cell migration and invasiveness through focal adhesion turnover and matrix metalloproteinase expression. Cancer Res 68: 5460 – 5468

Zhang J, Yang PL, Gray NS (2009) Targeting cancer with small molecule kinase inhibitors. Nat Rev Cancer 9: 28 – 39

Zhang F, Kumano M, Beraldi E, Fazli L, Du C, Moore S, Sorensen P, Zoubeidi A, Gleave ME (2014) Clustering facilitates stress-induced lipidation of LC3 and autophagosome biogenesis to enhance cancer cell survival. Nat Commun 5: 5775

Zhou HJ, Yan J, Luo W, Ayala G, Lin SH, Erdem H, Ittmann M, Tsai SY, Tsai MJ (2005) SRC-3 is required for prostate cancer cell proliferation and survival. Cancer Res 65: 7976 – 7983

Zhu ML, Horbinski CM, Carzotto M, Qian DZ, Beer TM, Kyprianou N (2010) Tubulin-targeting chemotherapy impairs androgen receptor activity in prostate cancer. Cancer Res 70: 7992 – 8002

Zoubeidi A, Zardan A, Beraldi E, Fazli L, Sowery R, Rennie P, Nelson C, Gleave M (2007) Cooperative interactions between androgen receptor (AR) and heat-shock protein 27 facilitate AR transcriptional activity. Cancer Res 67: 10455 – 10465

Zoubeidi A, Chi K, Gleave M (2010a) Targeting the cytoprotective chaperone, clusterin, for treatment of advanced cancer. Clin Cancer Res 16: 1088 – 1093

Zoubeidi A, Zardan A, Wiedmann RM, Locke J, Beraldi E, Fazli L, Gleave ME (2010b) Hsp27 promotes insulin-like growth factor-I survival signaling in prostate cancer via p90Rsk-dependent phosphorylation and inactivation of BAD. Cancer Res 70: 2307 – 2317

Zoubeidi A, Gleave M (2012) Small heat shock proteins in cancer therapy and prognosis. Int J Biochem Cell Biol 44: 1646 – 1656

License: This is an open access article under the terms of the Creative Commons Attribution 4.0 License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.