Epimerization-free access to C-terminal cysteine peptide acids, carboxamides, secondary amides, and esters via complimentary strategies†

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C-terminal cysteine peptide acids are difficult to access without epimerization of the cysteine α-stereocenter. Diversification of the C-terminus after solid-phase peptide synthesis poses an even greater challenge because of the proclivity of the cysteine α-stereocenter to undergo deprotonation upon activation of the C-terminal carboxylic acid. We present herein two general strategies to access C-terminal cysteine peptide derivatives without detectable epimerization, diketopiperazine formation, or piperidinylalanine side products.

C-terminal cysteine peptides, including prenylated and farnesylated peptides,1 disulfide linked peptide toxins,2 and insulitontropic peptides,3,4 comprise an important but synthetically challenging class of biologically active peptides. Many of these peptides are modified at the C-terminus. C-terminal modifications such as esters and amides can be critical to maintaining a peptide’s active conformation,5 in vivo activity, and pharmacokinetics;6 therefore, the ability to vary the peptide structure in this location is crucial to drug development efforts.7 Although several methods have been reported for C-terminal functionalization after solid-phase peptide synthesis (SPPS) is complete,8 these approaches either result in epimerization when applied to C-terminal Cys peptides9 or the applicability of the method to C-terminal Cys peptides is not addressed.9,10 While activation of the C-terminal carboxylic acid can induce epimerization via oxazolone formation in most amino acids,11 cysteine is also prone to epimerization via direct deprotonation during its attachment to the resin12 and upon prolonged or repeated exposure to base (i.e., during peptide elongation via Fmoc SPPS).13 Therefore, even the preparation of simple carboxylic acids or carboxamides of C-terminal cysteine peptides can be fraught with contamination by epimerized products,14,15 reducing the overall yield and complicating the purification of the target peptides. A method for the epimerization-free synthesis and subsequent C-terminal modification of C-terminal Cys peptides would be highly impactful.

In this work, we report the first mild and convenient method for the epimerization-free diversification of peptides bearing a C-terminal cysteine.16 Carboxylic acids, primary and secondary amides, and esters are accessed without epimerization or formation of diketopiperazine and piperidinylalanine side products.17 We apply this strategy to the total synthesis of the nicotinic acetylcholine receptor (nAChR) antagonist α-conotoxin IIm.18 Additionally, we include an alternate strategy employing N-deprotected cysteine derivatives as nucleophiles, and we demonstrate its utility via the synthesis of the insect pheromone α-factor.1

In the context of our ongoing efforts toward the synthesis of disulfide-linked α- and µ-conotoxins,19,20 we were concerned about possible epimerization of the C-terminal cysteine during the SPPS. We recently reported a strategy for C-terminal functionalization of non-cysteine peptides involving activation of the methyl-diaminobenzoyl (MeDbz) linker (1 → 2)21 followed by nucleophilic cleavage of the N-acyl urea (MeNbz) group22 to yield various protected (3) or unprotected (4) peptides (Scheme 1).23 If this approach were to prove mild enough to enable

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Scheme 1 Our strategy for C-terminal functionalization of non-Cys terminated peptides.
preparation of challenging C-terminal cysteine peptide derivatives, it would establish the MeNbz group as one of the mildest known activated carbonyl intermediates.\textsuperscript{24} We were report herein the exploitation of this reactivity to access C-terminal cysteine peptide acids, primary and secondary amides, and esters without epimerization. For epimerization-free functionalization of Cys, the attachment of the first amino acid,\textsuperscript{21b} peptide elongation, linker activation, and nucleophilic attack all must occur without epimerization of the unusually acidic\textsuperscript{22} Cys $\alpha$-stereocenter.

We expected that the parent diaminobenzyl group would not be sufficiently activating to cause epimerization during prolonged piperidine exposure. Thus, we sought to establish the stereochemical integrity of the Cys residue under these conditions unequivocally. We selected Cys(Trt) for these experiments because of its extreme tendency toward epimerization.\textsuperscript{14} Thus, we synthesized tripeptide Boc-Ala-Trp[Boc]-Cys(Trt)-MeDbzGly-Wang and exposed it to 20\% piperidine/DMF over 2, 4, and 24 h. The peptides were then cleaved under acidic conditions to afford H-AWC-MeDbz-Gly-OH. As expected, no epimerization was detected immediately following SPPS or a treatment with H-Nb-Cys) crypto-thioester approach.\textsuperscript{26} Importantly, this is the first report of a linker for which no epimerization is detected at a C-terminal Cys(Trt) residue after treatment with 20\% piperidine for 24 h.

With this result in hand, we were poised to evaluate the ability of the activated MeNbz linker to undergo nucleophilic displacement without inducing epimerization of the C-terminal cysteine. We first evaluated epimerization-prone Cys(Trt)-terminated peptides with N and O nucleophiles (5). We began with displacement by ammonia because of its small size and the relatively low pK$_a$ of NH$_3$. We were pleased to observe formation of the target peptide (6, Nuc = NH$_3$) with complete conversion and no detectable epimerization in 54\% isolated yield (Table 1, entry 1). We next evaluated benzylamine, which has a similar pK$_a$ but found that treatment of the activated linker with neat BnNH$_2$ led to 16\% epimerization (entry 2). However, using only 5 equiv. benzylamine in MeCN, the product was formed with no detectable epimerization (entry 3). We next tested neat butylamine, which is slightly more basic, and 8\% epimerization was observed. Reducing the amount of amine and varying the solvent did not improve epimerization in this case (entry 4–6). However, when we reduced the amount of butylamine to 1.1 equiv., we retained reactivity while eliminating epimerization of C-terminal Cys(Trt) (entry 7).

Other commercially available Cys PGs should be less prone to epimerization than Trt. Therefore, we used 5 equiv. BuNH$_2$ (i.e., entry 6 conditions) for the remaining protecting groups. First, we evaluated the Acm group in MeCN (entry 8), finding <1\% epimerization in the formation of the C-terminal Cys(Acm) butylamide. Next, we tested Mob, Bn, SrBu, and tBu with BuNH$_2$ in MeCN. In all cases, no epimerization was detected (entries 9–12). Turning our attention to alcohol nucleophiles, we tested MeOH/KO$i$Bu with Cys(Trt) as a benchmark\textsuperscript{26} and found 42\% epimerization (entry 13). However, in the presence of 5 equiv. Hünig’s base (DIEA) in MeOH, no epimerization was observed (entry 14). Because of the lower conversion in this case, we also investigated the use of a 1 : 1 MeOH/phosphate buffer solvent mixture (pH 8). In this case, complete conversion was observed while maintaining no detectable epimerization (entry 15). Even carboxylic acid derivatives with a C-terminal Cys(Trt) can be difficult to access without epimerization.\textsuperscript{25} Therefore, water was investigated as a nucleophile in the presence of Hünig’s base. In this case, the reaction was slower, but the product acid was observed with <1\% epimerization (entry 16). Thus, all nucleophiles tested react with the activated C terminus without any observed epimerization, regardless of the protecting group on cysteine.

Next, we sought to demonstrate the viability of this cleavage strategy in the context of more complex peptides. Because of our interest in disulfide-linked neuroactive peptides,\textsuperscript{27} we targeted the C-terminal carboxamide $\alpha$-conotoxin ImI (10), a sub-type selective nicotinic acetylcholine receptor antagonist\textsuperscript{20} isolated from the venom of Conus imperialis marine snails (Scheme 2).\textsuperscript{31,32} Both the C-terminal carboxamide and the correct disulfide bond connectivity are important to the bioactivity of $\alpha$-ImI (10).\textsuperscript{31,33} Thus, peptide 7 was activated and cleaved with ammonia to yield the fully protected peptide carboxamide. Acidic removal of protecting groups and HPLC purification gave the reduced peptide 8 in 25\% isolated yield.\textsuperscript{28} The first disulfide was formed in the presence of air in 1\% DMSO in phosphate buffer at pH 8 (peptide 9). Subsequent iodine treatment removed both Acm groups and induced oxidation to form the native conotoxin (10) in 52\% isolated yield over both folding steps. Co-injection of with a commercially available standard confirmed the correct folding.\textsuperscript{26} Alternatively, on-resin folding with iodine could be followed by MeDbz activation, cleavage with NH$_4$I, and side-chain deprotection to afford conotoxin 10 in 43\% isolated yield; however, this approach gives a mixture of folded products, as expected.\textsuperscript{35} We envisioned that situations might exist wherein it would prove advantageous to access the target peptides by using various pre-functionalized cysteine derivatives as nucleophiles for cleavage of MeNbz from the resin.\textsuperscript{27,36} For example, this would avoid the need to screen epimerization for each new derivative, it would enable access to authentic standards for peptides generated using direct C-terminal modification approaches, and it would allow incorporation of a very poor or hindered nucleophile. Recently, C-terminal cysteine peptide acids were synthesized using an N-(2-hydroxy-5-nitrobenzyl)-cysteine (N-Hnb-Cys) crypto-thioester approach.\textsuperscript{27} Although this method enabled access to a challenging Pro-Cys linkage at the C terminus, elevated temperatures and long reaction times were required to generate the C-terminal thioester, leading to undesired side products. The stability of esters to this approach was not determined. We envisioned that application of the cysteine elongation tactic to our MeNbz-based C-terminal modification would lead to a convenient alternative method to access challenging targets at ambient temperature and with short reaction times. However, there was no report of an intermolecular\textsuperscript{37} NCL-like reaction being conducted with the C-terminally linked peptide still attached to the resin.

The elongation approach was tested with H-AWA-MeNbz-Gly-Wang peptides (12 and 13), which were treated with free cysteine,
### Table 1: Evaluation of epimerization during nucleophilic cleavage of the MeNbz group in C-terminal cysteine peptides

| Entry | PG | Nuc-H | Base (5 equiv.) | Solvent | % conversion | Epimerization<sup>b</sup> (% d-X) |
|-------|----|-------|----------------|--------|--------------|-------------------------------|
| 1<sup>a</sup> | Trt | NH<sub>3</sub> | — | DMF | >99 | <1 |
| 2 | Trt | PhCH<sub>2</sub>NH<sub>2</sub> | — | DMF | >99 | 16 |
| 3 | Trt | PhCH<sub>2</sub>NH<sub>2</sub> | — | MeCN | >99 | <1 |
| 4 | Trt | BuNH<sub>2</sub> | — | BuNH<sub>2</sub> | >99 | 8 |
| 5 | Trt | BuNH<sub>2</sub> | — | DMF | >99 | 10 |
| 6 | Trt | BuNH<sub>2</sub> | — | MeCN | >99 | 9 |
| 7<sup>d</sup> | Trt | BuNH<sub>2</sub> | — | MeCN | >99 | <1 |
| 8<sup>e</sup> | Acm | BuNH<sub>2</sub> | — | MeCN | >99 | <1 |
| 9<sup>f</sup> | Mob | BuNH<sub>2</sub> | — | MeCN | >99 | <1 |
| 10 | Trt | BuNH<sub>2</sub> | — | MeCN | >99 | <1 |
| 11 | Sib Bu | BuNH<sub>2</sub> | — | MeCN | >99 | <1 |
| 12 | BuNH<sub>2</sub> | KOBu | MeOH | MeOH | >99 | 42 |
| 13 | Trt | MeOH | DIEA | MeOH | >99 | <1 |
| 14<sup>g</sup> | Trt | MeOH | — | MeOH/Na<sub>2</sub>HPO<sub>4</sub>/NaH<sub>2</sub>P<sub>2</sub>O<sub>7</sub> buffer at pH = 8 | >99 | <1 |
| 16 | Trt | H<sub>2</sub>O | DIEA | H<sub>2</sub>O/MeCN | >99 | 56 |

<sup>a</sup> All reactions were performed on 20 mg of resin containing all L amino acids in 200 µL of solvent at ambient temperature (24 ± 1 °C).<br> <sup>b</sup> All PGs were removed prior to epimerization assay unless otherwise noted.<br> <sup>c</sup> Cys(PG) was intact during epimerization assay.<br> <sup>d</sup> 1.1 equiv. of BuNH<sub>2</sub> was used.<br> <sup>e</sup> 0.7 equiv. KOBu. <sup>f</sup> Reaction was conducted for 3 h. <sup>g</sup> Na<sub>2</sub>HPO<sub>4</sub>/NaH<sub>2</sub>P<sub>2</sub>O<sub>7</sub> buffer at pH = 8.

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H-Cys-OEt, H-Cys-NH<sub>2</sub>, or H-Cys-NHBu in the presence of H<sub>2</sub>O/DIEA/H<sub>2</sub>O/MeCN 56 <1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>O/MeCN 56 <1

5 Trt BuNH<sub>2</sub> 59

3 Trt PhCH<sub>2</sub>NH<sub>2</sub> 352

1

16 Trt H<sub>2</sub>O DIEA H<sub>2</sub>
both the C-terminal ester and the prenyl moiety for bioactivity. Protected des-farnesyl \( \alpha \)-factor was generated by displacement of peptide 19 with cysteine methyl ester. The elongation was conducted in NCL buffer in an effort to reduce cystine-functionalized by-products. However, even under these reducing conditions, cystine-appended \( \alpha \)-factor was still observed. Side-chain deprotection, cystine reduction, and HPLC purification afforded peptide 20 in 41% yield over 3 steps. Alkylation per the reported conditions afforded \( \alpha \)-factor in 9% yield (21).

In summary, we have developed two broadly applicable strategies for the epimerization-free preparation of C-terminal cysteine peptides. The first approach exploits the exceptionally mild activating nature of the N-acyl urea group for the direct diversification of the C terminus. Additionally, an alternative strategy wherein cysteine derivatives serve as the nucleophile in a resin-cleaving elongation reaction was also effective. For both strategies, the target peptides are prepared without observation of either diketopiperazine or piperidinylalanine side products. The utility of these methods was demonstrated in the preparation of the disulfide-linked conotoxin \( \alpha \)-ImI, bearing a C-terminal cysteine carboxamide and the insect pheromone

![Scheme 3](image-url)  
**Scheme 3** Synthesis of \( \alpha \)-factor by cysteine elongation.
z-factor, which possesses a C-terminal cysteine methyl ester. Notably, no previous report has demonstrated successful functionalization of C-terminal cysteine peptides to access carboxylic acids, carboxamides, and other C-terminal derivatives without detectable epimerization of the z-stereocenter.

Conflicts of Interest
The authors declare no conflicts of interest.

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