Evidence for Circulation of the Rift Valley Fever Virus among Livestock in the Union of Comoros

Matthieu Roger, Marina Beral, Séverine Licciardi, Miradje Soule, Abdourahime Faharoudine, Coralie Foray, Marie-Marie Olive, Marianne Maquart, Abdouroihamane Soulaimane, Ahmed Madi Kassim, et al.

To cite this version:
Matthieu Roger, Marina Beral, Séverine Licciardi, Miradje Soule, Abdourahime Faharoudine, et al.. Evidence for Circulation of the Rift Valley Fever Virus among Livestock in the Union of Comoros. PLoS Neglected Tropical Diseases, 2014, 8 (7), pp.e3045. 10.1371/journal.pntd.0003045. hal-01274567

HAL Id: hal-01274567
https://hal.univ-reunion.fr/hal-01274567
Submitted on 21 Jun 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Distributed under a Creative Commons Attribution 4.0 International License
Evidence for Circulation of the Rift Valley Fever Virus among Livestock in the Union of Comoros

Matthieu Roger1,2,3,*, Marina Beral1,2,3,*, Séverine Licciardi3,*, Miradje Soulé4, Abdourahime Faharoudine4, Coralie Foray1,2,3, Marie-Marie Olive1,2,3,5, Marianne Maquart1,2,3, Abdouroihame Soulaimane4, Ahmed Madi Kassim4, Catherine Cété-Sossah1,2,3, Eric Cardinale1,2,3

1 Centre de Coopération Internationale en Recherche Agronomique pour le Développement (CIRAD), UMR 15 CMAEE, Sainte Clotilde, La Réunion, France, 2 Institut National de la Recherche Agronomique (INRA), UMR 1309 CMAEE, Sainte Clotilde, La Réunion, France, 3 Centre de Recherche et de Veille sur les Maladies Émergentes dans l’Océan Indien (CRVOI), Plateforme de Recherche CYROI, Sainte Clotilde, La Réunion, France, 4 Vice-Présidence en Charge de l’Agriculture, l’Élevage, la Pêche, l’Industrie, l’Énergie et l’Artisanat, Mdé, Moroni, Union des Comores, 5 Unité de Virologie, Institut Pasteur de Madagascar, Antananarivo, Madagascar

Abstract

Rift Valley fever virus (RVFV) is an arthropod-borne phlebovirus reported to be circulating in most parts of Africa. Since 2009, RVFV has been suspected of continuously circulating in the Union of Comoros. To estimate the incidence of RVFV antibody acquisition in the Comorian ruminant population, 191 young goats and cattle were selected in six distinct zones and sampled periodically from April 2010 to August 2011. We found an estimated incidence of RVFV antibody acquisition of 17.5% (95% confidence interval (CI): [8.9–26.1]) with a significant difference between islands (8.2% in Grande Comore, 72.3% in Mohéli and 5.8% in Anjouan). Simultaneously, a longitudinal entomological survey was conducted and ruminant trade-related information was collected. No RVFV RNA was detected out of the 1,568 blood-sucking caught insects, including three potential vectors of RVFV mosquito species. Our trade survey suggests that there is a continuous flow of live animals from eastern Africa to the Union of Comoros and movements of ruminants between the three Comoro islands. Finally, a cross-sectional study was performed in August 2011 at the end of the follow-up. We found an estimated RVFV antibody prevalence of 19.3% (95% CI: [15.6%–23.0%]). Our findings suggest a complex RVFV epidemiological cycle in the Union of Comoros with probable inter-islands differences in RVFV circulation patterns. Mohéli, and potentially Anjouan, appear to be acting as endemic reservoir of infection whereas RVFV persistence in Grande Comore could be correlated with trade in live animals with the eastern coast of Africa. More data are needed to estimate the real impact of the disease on human health and on the national economy.

Citation: Roger M, Beral M, Licciardi S, Soulé M, Faharoudine A, et al. (2014) Evidence for Circulation of the Rift Valley Fever Virus among Livestock in the Union of Comoros. PLoS Negl Trop Dis 8(7): e3045. doi:10.1371/journal.pntd.0003045

Editor: Brian Bird, Centers for Disease Control and Prevention, United States of America

Received October 8, 2013; Accepted June 11, 2014; Published July 31, 2014

Copyright: © 2014 Roger et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was conducted in the framework of AnimalRisk-Ori, a research program on emerging animal diseases in the Indian Ocean, funded by FEDER POCT (European Union, Regional Council of Reunion and the French government). The study was partially funded by EU grant FP7-261504 EDENext and is catalogued by the EDENext Steering Committee as EDENext000 (http://www.edenext.eu). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: matthieu.roger@cirad.fr

These authors contributed equally to this work.

Introduction

Rift Valley fever (RVF) is an arthropod-borne zoonotic disease caused by a RVF virus (RVFV), a member of the Phlebovirus genus of the family Bunyaviridae [1]. RVFV causes significant morbidity and mortality among sheep, goats, cattle and also affects human. In livestock, abortion storms and high mortality observed among the younger animals cause significant economic losses [2,3]. Humans are usually infected by contact with infectious animal tissues through inhalation or aerosols generated by slaughtering and necropsy [4]. Arthropod vectors play an important role during the onset of epidemic and inter-epidemic periods [5]. In endemic areas, RVFV is maintained in the environment through an enzootic vertebrate-arthropod cycle [6]. RVFV has been isolated from many vectors in the field [7], such as ticks and sand flies which are able to transmit the virus in experimental conditions [8,9]. However, mosquitoes are the main insects involved in the spread of RVFV during epidemics. RVFV has been isolated from at least 40 species of mosquitoes belonging to 8 genera but only some of them are susceptible and able to transmit RVFV under laboratory conditions [10]. RVFV is widely present in Africa and has been spreading to Madagascar and the Arabian Peninsula [11,12]. In 2007, RVF outbreaks were reported in several eastern and southern African countries [13]. A few weeks later, and for the first time, RVFV was detected in the Comoros archipelago following the hospitalization of a young Grande Comorian boy showing symptoms of severe encephalitis [14]. In addition, during the 2008 and 2009 rainy seasons, outbreaks due to RVFV strains imported from mainland Africa were reported in Madagascar causing 59 confirmed human cases and seven deaths [11,15].

In Mayotte, the French overseas territory that belongs to the Comoros archipelago, a retrospective study conducted in 2008 confirmed the presence of the disease with 10 human cases
infected with RVFV strains genetically closely linked to the 2006–2007 Kenyan isolates [16]. It was also found that the Mayotte livestock has been infected by RVFV prior to 2004 [17]. Regarding the Union of Comoros, in 2011, Roger et al. reported widespread exposure of Comorian livestock with 32.8% of animals shown to be RVFV-seropositive without any notifications of clinical signs by the Comorian Animal Health Services. From April 2010 to August 2011, we conducted a livestock longitudinal survey in Grande Comore, Moheli and Anjouan. Our study aimed to detect RVFV-specific antibody acquisitions in cattle and goats. Simultaneously, a longitudinal entomological survey was conducted to describe the diversity of mosquitoes in the study zones and ruminant trade-related information was collected. Our investigations showed that Comoros ruminants acquired RVFV-specific antibodies all along the year and particularly in Moheli during the dry season. Our findings suggest a complex RVFV epidemiological cycle in the Union of Comoros with probable inter-islands differences in RVFV circulation patterns. The disease appears to be endemic in Moheli and potentially Anjouan, but the persistence of the disease in Grande Comore could be correlated with trade in live animals with the eastern coast of Africa.

Materials and Methods

Ethics statement

The research protocol was implemented with the approval of the Vice-Presidency of Agriculture, Fisheries and Environment of the Union of Comoros. No endangered or protected species were involved in the survey. Farmers in each zone gave their verbal consent to be included in the study. Permissions for the blood sample collection were obtained. The animals were bled without suffering. Regarding the trade survey, no personal data were collected, and only information concerning the number of animals travelling from one island to another was taken into account.

Study zones

The Comoros islands form an archipelago of volcanic islands located off the southeastern coast of Africa, east of Mozambique and northwest of Madagascar. The archipelago is divided between the sovereign state of the Union of Comoros composed of three islands named Grande Comore, Moheli, and Anjouan, and the French overseas department of Mayotte. The tropical climate of the Comoros islands is characterized by daytime temperatures around 26°C at sea level, with limited variation during the year, and by annual heavy rainfall (2,679 mm) with two seasons: a humid season from November to April, and a dry season from May to October.

Based on the results of a previous RVFV antibody prevalence study in 2009 [18], six zones were selected in the Union of Comoros (Figure 1). Four zones were selected on the island of Grande Comore: zones 1 and 2 located in the center of the island where low RVFV antibody prevalence was found, and zones 3 and 4 located in the south with high RVFV antibody prevalence [18]. Zones 2 and 4 are located along the coast (0–200 m above sea level (asl.)) where ruminants are mostly goats stall reared or ranging free within and outside villages. Zones 1 and 3 are located at a moderate altitude (500–650 m asl.) where ruminants are mostly cattle reared in pastures (zone 1) or raised in stables in the forest (zone 3).

Zone 5, which was located on the southern coast of the island of Moheli, was selected because of its highest RVFV antibody prevalence observed during the 2009 survey [18]. On this island, cattle are reared in stables on an old coconut plantation. Finally, in zone 6 located close to the airport on Anjouan island, cattle were raised in stables in vegetable production areas.

Animals and sampling

Five ml of whole blood was collected from the jugular vein of goats and cattle in Vacutainer tubes (Becton Dickinson, USA). Samples were allowed to clot at 15°C and serum was separated from whole blood by centrifugation; samples were stored in liquid nitrogen in the field and at −80°C in the laboratory.

Livestock longitudinal survey

The livestock longitudinal survey was conducted in the six separate zones detailed in Figure 1. From 20 to 30 ruminants were individually identified in each zone using ear tags. The number of animals sampled per zone was based on the previous survey, with a RVFV antibody prevalence ranging from 20% to 50% [18] with 70% relative precision [23]. To avoid colostral immunity, animals were selected as follows: cattle were between 10 months and one-year of age, and small ruminants were between three to eight months of age. Animals were sampled monthly from April 2010 to...
August 2010 and every four months from August 2010 to August 2011. The first series of serological tests determined the RVF serological status of each sampled animal. Only RVFV antibody negative animals were included in the livestock longitudinal survey and continued to be sampled until their IgG RVFV antibody positive status and were then excluded from the study. When possible, new ruminants were included in the study to substitute lost, dead or RVFV IgG positive animals.

Livestock cross-sectional survey

The RVFV antibody prevalence based on the different study zones was estimated in August 2011. The sample size was based on the previously estimated prevalence [18] with a relative precision of 20% and a confidence level of 95% giving a required minimum of 385 animals to be collected [23]. Without any particular Comorian livestock census, animals were selected on the farmer’s willingness to cooperate during the study.

Longitudinal entomological survey

Blood-sucking insects were sampled every four months from November 2010 to August 2011 along with the longitudinal serological survey using double-net goat baited traps placed from 4:00 pm to 10:00 am. The sampling was carried out for three consecutive days in the study zones numbered 1, 3, 5 and 6 (Figure 1). No sampling was performed in zones 2 and 4 for logistic reasons.

Environmental data

In order to generate hypotheses on potential associations between the estimated RVFV incidence and prevalence with environmental risk factors for RVF infection, we collated climatic variables [24]. Two remotely-sensed MODerate-resolution Imaging Spectroradiometer (MODIS) data sets were sourced from the National Aeronautics and Space Administration (http://modis.gsfc.nasa.gov/), namely the Daytime Land Surface Temperature (DLST) and the Nighttime Land Surface Temperature (NLST), both with spatial and temporal resolution of 1 km and 8 days. In addition, rainfall data were obtained from the Malaria Early Warning System (MEWS) program, freely available in the MEWS repository (http://iridl.ldeo.columbia.edu/expert/SOURCES/.NOAA/.NCEP/.CPC/.FEWS/.Africa/.TEN-DAY/.RFEv2/.est_prcp/), with a spatial and temporal resolution of 11 km and 10 days respectively. DLST, NLST and rainfall values were extracted within a 5-km radius buffer around each farm corresponding to the maximum daily distance for cattle (grazing and watering). For each sampled animal that became RVFV antibody positive,
MODIS and MEWS data recorded at the time of the seroconversion in the zone concerned were compared with MODIS and MEWS data recorded at the same time in the other zones.

Trade survey
The aim of the trade survey was to estimate the movement of live animals between continental Africa and the Comoros archipelago and among the islands of the archipelago themselves. To date, only approximate figures are known without any quantitative data available [17]. The number of imported ruminants was collected monthly between November 2010 and August 2011 as follows i) the local veterinary authorities provided records of animal movements through the official ports of Moroni (Grande Comore), Fomboni (Moheli), and Mutsamudu (Anjouan), ii) one interviewer per island had the task of identifying undeclared animal arrivals on the coast, either in the field or from information provided by village chiefs.

Laboratory tests
Detection of RVFV antibodies. Sera were first tested by two ELISAs (enzyme-linked immunosorbent assay) the Immunoglobulin (Ig) G IDScreen RVF Competition ELISA (IdVet, France) and the IgM capture ELISA [25]. To confirm their status, each of the RVFV ELISA antibody positive samples and some randomly chosen RVFV ELISA negative samples were tested using the virus neutralizing test (VNT), considered as the gold standard method described by OIE [26]. Briefly, duplicates of two-fold serial dilutions of sera starting from 1:5 were added to 100 TCID50 of Smithburn RVFV in 96-well microtiter plates and incubated for 1 h at 37°C. Next, 100,000 Vero cells were added to each well and the plates were incubated with 5% CO2 for 5–6 days at 37°C. Titers were expressed as the inverse highest dilutions giving 50% of CPE. A positive control serum was included. A serum sample with a titer of 1:10 or higher was considered seropositive.

Morphological identification of insects. Specimens were collected by direct aspiration with a home-made vacuum system and anesthetized with chloroform. Each specimen was morphologically identified by microscopy in the field. Insects were pooled (1 to 10 individuals), per species per trap and per zone and stored in liquid nitrogen in the field and at −80°C in the laboratory. Engorged females were not included in the pool.

Detection of RVFV in insects. The pooled insects were ground up with 400 µl of PBS 1X (Phosphate Buffered Saline) twice for 30 seconds with two 3-mm diameter stainless beads using the TissueLyser system (Loudet, France) and transferred to a 96-well plate. Total RNA was extracted with the Biomek NX robot (Beckman Coulter, USA) using the NucleoSpin 96 Virus kit (Macherey-Nagel, Germany). For RVFV RNA detection, the L-Segment based SYBR-Green real time PCR was used [27,28]. Ten-fold serial dilutions of a Smithburn strain which contained 10⁶ TCID50/ml were used as the standard curve for plate validation.

Data analysis and statistics
All statistics were performed using R.3.0.1 [29]. For both Fisher’s exact test and the Student-t test, a value of P<0.05 was considered significant.

A seroconversion was defined as an animal found with either a positive IgM ELISA result or a positive IgG ELISA result or both following a previous negative RVF ELISA sample result.

Incidence rates and instantaneous risk of infection. The rate of instantaneous risk of infection (Txi) is described as:

\[
T_{xi} = \frac{\text{Number of seroconverted animals}}{\text{Number of animals in an at-risk period}}
\]

The incidence rate was determined using an Access database in the Laser format (available at http://livtools.cirad.fr/) [30–32] by calculating the instantaneous risk of infection (the risk that an animal will be infected in a given period) [30] taking into account the risk of seroconversion, death or lost animals. The number of animals in an at-risk period represents the total number of animals in a susceptible period of RVFV infection.

Incidence and seroprevalence analysis. To assess whether location had an effect on the RVFV antibody prevalence, different zones with specific ecosystems were included in the follow-up study. A Fisher’s exact test was used to compare the difference in incidence of RVFV antibody acquisition and RVFV antibody prevalence between zones and islands. Incidence was analyzed and hypotheses were proposed about relationships with environmental and climate conditions.

Analysis of entomological data. A trapping session with no rain and/or wind was included in the data analysis even if no blood-sucking insects were collected. Student’s t-Test was used for statistical analysis.

Trade survey. The movements of animals recorded between the three islands of the Union of Comoros were mapped, as well as their potential connection with continental Africa, Mayotte and Madagascar.

Results
Livestock longitudinal survey in relation with environmental data
A total of 191 ruminants (88 cattle and 103 goats) were included in the livestock longitudinal survey: 135 animals in Grande Comore, 27 in Moheli and 29 in Anjouan. Detection of RVFV antibodies (IgM and IgG) was performed by ELISA for a total of 849 serum samples over the duration of study.

Table 1 presents by date and per zone the number of animals that acquired RVFV antibodies over the duration of the livestock longitudinal survey. A total of 15 animals out of the 191 sampled acquired RVFV antibody during the study. Each of the 13 RVFV IgG ELISA positive samples were confirmed by VNT. Only one RVFV IgM ELISA positive sample was not confirmed by VNT (July 200, Moheli). This animal was confirmed RVFV IgG ELISA positive and VNT positive four months later in November 2010. Out of the 112 RVFV IgG ELISA negative samples randomly chosen, all were found negative by VNT. RVFV IgM antibodies acquisition was detected in three animals and RVFV IgG antibodies acquisition in 12 animals. Only one RVFV IgM ELISA positive animal in Moheli converted to RVFV IgG antibodies. The two others RVFV IgM ELISA positive ruminants were lost or slaughtered before the next sampling session (Table 1). Nine out of the 15, which acquired RVFV antibody, were recorded in Moheli, five in Grande Comore and one in Anjouan. Nine out of those fifteen occurred during the dry season (six in Moheli, one in Anjouan, two in Grande Comore).

The overall annual incidence of RVFV antibody acquisition for the Union of Comoros was estimated at 17.54% [animal risk time = 91], with a 95% confidence interval (CI) [8.95–26.14]) (Table 2). A significant difference was found when incidence of RVFV antibody acquisition was compared between zones (Fisher exact test, p<0.001) or between islands (Fisher exact test, p<
Table 1. Number of animals that acquired RVFV antibody per study zone during the livestock longitudinal survey, Union of Comoros, April 2010 to August 2011.

| Zone   | Sampling month | 2010 | 2011 | 2012 |
|--------|----------------|------|------|------|
| Grande Comore | April | 15 | 26 | 23 |
|          | May   | 22 | 25 | 23 |
|          | June  | 20 | 18 | 13 |
|          | July  | 18 | 20 | 16 |
|          | August | 16 | 22 | 21 |
| Moheli  | April | 22 | 27 | 25 |
|          | May   | 16 | 10 | 0 |
|          | June  | 8  | 0  | 0  |
|          | July  | 8  | 0  | 0  |
|          | August | 9  | 0  | 0  |
| Anjouan | April | 28 | 30 | 30 |
|          | May   | 22 | 28 | 20 |
|          | June  | 18 | 19 | 16 |
|          | July  | 19 | 19 | 16 |
|          | August | 18 | 18 | 15 |

Insect trapping and RVFV detection

Twelve trapping days were conducted in each of the four zones under study (zones 1, 3, 5 and 6, see Figure 1). Blood-sucking insects were trapped in five out of the twelve trapping days in central Grande Comore (zone 1), in eight trapping days in southern Grande Comore (zone 3), eleven trapping days in Moheli (zone 5), and in seven trapping days in Anjouan (zone 6) (Table 4). Out of the 1,568 blood sucking insects caught with the double-net goat baited trap, 1,548 were identified as mosquitoes and 20 were identified as Stomoxys niger. A total of 1,133 insects were collected in Moheli (zone 5), 291 in Anjouan (zone 6), 108 in southern Grande Comore (zone 3) and 36 in central Grande Comore (zone 1). Although the number of comparisons was not large, the average number of trapped mosquitoes per trapping day per zone was significantly higher in Moheli (average was 113 insects) and Anjouan (average was 42 insects) compared to Grande Comore zones 1 (average was 7 insects) and zone 3 (average was 14 insects) (Table 5).

The diversity and number of blood-sucking insects caught with the double net goat baited trap per trapping day per zone are presented in Table 4. A total of seven genera and 16 species were caught of which 14 could be morphologically identified. Fifteen species out of the 16 caught were collected in Moheli (zone 5), nine species were collected in Anjouan (zone 6), three and eight in central and southern Grande Comore respectively (zone 1 and zone 3). Eighty-seven percent of the total number of insects caught belonged to three species with 52% belonging to two Eretmapodites species (E. quinquevittatus and E. subsimplicipes,) and 35% to Aedes cartroni.

No RVFV RNA was detected in any of the 442 pools tested.
Table 2. Incidence of RVFV antibody acquisition per zone including statistical analysis, Union of Comoros 2011.

| Period                  | Union of Comoros | Grande Comore (GC) | Moheli Zone 5 | Anjouan Zone 6 |
|-------------------------|------------------|-------------------|---------------|---------------|
|                         | Incidence of antibody acquisition |                        |               |               |
| Annual                  | 0.175            | 0.082             | 0.051         | 0.055         | 0.071         | 0.227         | 0.723         | 0.058         |
|                         | 91               | 63                | 18            | 19            | 14            | 9             | 14            | 7             |
| 95% CI                  | [0.089–0.261]    | [0.010–0.155]     | [0.000–0.163] | [0.000–0.152] | [0.000–0.212] | [0.000–0.542] | [0.255–1.000] | [0.000–0.173] |
| Rainy season (November–April) | 0.075            | 0.030             | 0.000         | 0.031         | 0.000         | 0.136         | 0.413         | 0.000         |
|                         | 92               | 64                | 28            | 32            | 23            | 15            | 10            | 20            |
| 95% CI                  | [0.02–0.132]     | [0.000–0.065]     | [0.000–0.150] | [0.000–0.092] | [0.000–0.178] | [0.000–0.325] | [0.010–0.819] | [0.000–0.218] |
| Dry season (May–October) | 0.095            | 0.016             | 0.049         | 0.000         | 0.110         | 0.000         | 0.415         | 0.058         |
|                         | 93               | 62                | 21            | 25            | 9             | 7             | 14            | 17            |
| 95% CI                  | [0.003–0.158]    | [0.000–0.048]     | [0.000–0.325] | [0.000–0.165] | [0.000–0.325] | [0.000–0.439] | [0.080–0.747] | [0.000–0.170] |

Statistical analysis (using the dataset above)

|                          | Comparison of dry season and rainy season incidence per zone [p-value]** | Comparison of the study zones incidence [p-value]** |
|--------------------------|---------------------------------------------------------------|-----------------------------------------------|
|                          | 0.795                                                         | Annual                                       |
|                          | 0.443                                                         | < 0.001                                      |
|                          | 0.428                                                         | Rainy season                                 |
|                          | 1                                                             | < 0.001                                      |
|                          | 0.303                                                         | Dry season                                   |
|                          | 1                                                             | 1                                             |
|                          | 1                                                             | 1                                             |

CI: stands for Confidence Interval, * nrisk represents the number of animals at risk in a susceptible RVFV infection period, ** Fisher exact test, p-value significant if p < 0.05. The incidence rate was determined using an Access database in the Laser format [available at http://livtools.cirad.fr/] [29–31] by calculating the instantaneous risk of infection (the risk that an animal will be infected in a given period) [29] taking into account the risk of seroconversion, death or lost animals. The number of animals in an at-risk period represents the total number of animals in a susceptible period of RVFV infection.

doi:10.1371/journal.pntd.0003045.t002
Trade survey
The study highlighted movements of live ruminants between the three islands of the Union of Comoros, the African mainland, Mayotte and Madagascar (Figure 2). Data recorded by veterinarians and technicians showed movements of live ruminants from i) the east coast of Africa to Union of Comoros and ii) between the three islands of the Union of Comoros (Figure 2A). Animals were observed being landed on beaches without any controls or in secondary “ports” like Chindini in the south of Grande Comore. Figure 2B represents the dynamics of live animal importations in Union of Comoros from May 2010 to July 2011. We recorded up to ten fold more ruminants imported in Grande Comore than in Moheli or Anjouan.

Discussion
Rift Valley fever was detected for the first time in Grande Comore in the human population in 2007 [14] and in livestock in 2009 [18]. Our study demonstrates that RVFV is still circulating in the Union of Comoros despite of the absence of apparent clinical signs in livestock.

Fifteen RVFV seroconversions were observed in the Union of Comoros between 2010 and 2011 giving an overall incidence of RVFV antibody acquisition of 17.5%. These results suggest continuous circulation of RVFV on the three islands. However, significant differences in incidence were observed between islands (p<0.001). The incidence of RVFV antibody acquisition was higher in Moheli (72.3%) than in Anjouan (5.8%) and in Grande Comore (8.2%). This is in accordance with differences in RVFV antibody prevalence between the Union of Comoros islands recorded in 2009 and 2011. In 2011, RVFV antibody prevalence in Anjouan was still below the one in Grande Comore, whereas RVFV antibody prevalence remained the highest in Moheli. However, in Grande Comore and Anjouan RVFV antibody prevalence in 2011 appeared to have decreased whereas in Moheli, RVFV antibody prevalence remained similar to the level recorded in 2009, despite herd replacement estimated at 12% (L. Cavalerie, personal communication). These results suggest the existence of island specific RVF circulation patterns.

Seasonality of the incidence of RVFV antibody acquisition needs to be explored. The Comorian livestock farming characteristics (small herd size and small total number of ruminants) as well as the field issues did not allow a sufficient number of young ruminants (risk too small) reducing the power of the statistical analysis.

No clinical signs were reported in the Union of Comoros during the period of our study, as reported in Madagascar, Tanzania, and Mozambique in recent years [33–35], but the fifteen seroconversions observed suggest that RVFV could be circulating in the Comorian environment thanks to local mosquito-mammalian host cycles even if the numbers of caught mosquitoes were not large nor positive for RVF RNA. Out of the 1,568 blood-sucking insects caught, none were found to be RVFV RNA positive by PCR but in the absence of RVF outbreaks, chances of detecting RVFV in vector populations are known to be very low [36]. In 1978, Bruhnes described 30 mosquito species in the Union of Comoros [22]. Four of them: *Ae. aegypti*, *Ae. fowleri*, *Ae. circumluteolus* and *Cx. quinquefasciatus* are considered as RVFV potential vectors because the virus has been already isolated in these species in the field and because of their capacity to transmit RVFV under laboratory conditions [37–39]. All these species, except *Ae. fowleri*, have been caught at least on one island during our study, suggesting a role for this mosquito species to be involved in the transmission cycle on each of the islands. Five other mosquito species caught during our study, *Ex. quinquevittatus*, *An. arabiensis*, *M. uniformis*, *An. coustani* and *Ae. simpsoni* were previously identified as RVFV RNA positive by PCR in the field [40–44]. *An. coustani* and *Ae. simpsoni* were found RVF RNA carrier for the first time in the Indian Ocean area: respectively in Madagascar in 2011 and in Mayotte in 2009 [43,44]. Thus, some of these mosquito species may play a role in RVFV transmission in the Union of Comoros. Geological inaccessibility, sampling design and climatic conditions likely explain the small number of specimens caught and the heterogeneity of entomological findings between islands [45]. These volcanic islands are characterized by a tropical climate with only slight variations in daily temperatures and abundant rainfalls, which theoretically should enable populations of Culicidae species to persist throughout the year.

| RVFV antibody prevalence | Statistical analysis [p-value] |
|--------------------------|-------------------------------|
|                          | Grande Comore | Moheli | Anjouan |
| Grande Comore            | prevalence | 0.247  |         |
|                          | n | 174    |         |
|                          | 95% CI | [0.183–0.311] |         |
| Moheli                   | prevalence | 0.458  | 0.008*  |
|                          | n | 48     |         |
|                          | 95% CI | [0.317–0.599] |         |
| Anjouan                  | prevalence | 0.207  | 0.683   | 0.013*  |
|                          | n | 53     |         |
|                          | 95% CI | [0.098–0.317] |         |
| Union of Comoros         | prevalence | 0.276  | 0.007** |
|                          | n | 275    |         |
|                          | 95% CI | [0.223–0.329] |         |

CI: stands for Confidence Interval, *Fisher exact test, 2 by 2 comparison, p-value significant if p<0.05, ** Fisher exact test, multiple comparison, p-value significant if p<0.05.

doi:10.1371/journal.pntd.0003045.t003
Nevertheless, each island has its own environmental characteristics, as the age of the three islands decreases westward: Moheli is 2.73 ± 0.20 million years old, Anjouan, 1.18 ± 0.03 million years old, and Grande Comore is 0.13 ± 0.02 million years old [46].

On Moheli and Anjouan, the oldest islands, the landscape includes permanent rivers [47] and, as a result, many artificial and natural breeding mosquito sites exist. Moheli has a wide variety of natural and artificial sites in which mosquitoes can breed all year round [47,48]. The presence of clay, resulting from the decomposition of volcanic soils, ensures the presence of abundant surface water impoundments. It allows the cultivation of irrigated rice hence and favors the development of diversified mosquito populations [47]. A greater number of mosquito species were caught in Moheli (15 species) than in the other islands which is in agreement with Brunhes’ inventory in 1978, including two mosquito species known as RVFV potential vectors. Thus, favorable conditions for RVFV persistence being present a better chance for a possible RVFV cycle involving vectors and animals is suggested.

The abundance of mosquitoes trapped in Anjouan (zone 6) was similar to that in Moheli (zone 5) and three mosquito species known as RVFV potential vector have been caught during our study. However, RVFV antibody prevalence in Anjouan was the lowest and appeared to be decreasing. Moreover in 2011, only one ruminant exhibited a RVFV seroconversion. Anjouan shares some similar environmental characteristics with Moheli that could allow mosquitoes to survive all year round but Anjouan has some characteristics that could limit the circulation of RVFV. For example, the landscape is comprised of hill slopes and irrigated field rice is not cultivated on the island. Ruminants are mainly reared in stalls in the highlands in the eastern part of the island. For that reason, the probability of contact between infected vectors and ruminants may be lower in Anjouan than in Moheli and the maintenance of a vector-ruminant cycle may be harder to get. More investigations in other cattle-rearing areas are thus needed to conclude on RVF circulation in Anjouan.

Incidence of RVFV antibody acquisition and the RVFV antibody prevalence in Grande Comore are hard to explain based only on entomological parameters. Presence of steep slopes with decomposed and highly permeable soils characterize Grande Comore, the youngest island of the country [47]. Surface water is

| Table 4. Diversity and number of blood-sucking insects caught with a double baited net per trapping day and per zone, Union of Comoros, 2011. |
|---------------------------------------------------------------|
| **Genus** | **species** | Grande Comore | Moheli | Anjouan |
|-----------|------------|---------------|--------|--------|
|           |            | Zone 1 | Zone 3 | Zone 5 | Zone 6 | Total number |
| Stomoxis  | niger      | 0      | 0      | 11     | 9      | 20          |
| Aedes     | aegypti    | 1      | 4      | 8      | 15     | 28          |
| Aedes     | cartroni   | 0      | 0      | 504    | 48     | 552         |
| Aedes     | circumlateolus | 0    | 0      | 0      | 10     | 10          |
| Aedes     | simpsoni   | 0      | 20     | 1      | 0      | 21          |
| Aedes     | vittatus   | 0      | 3      | 13     | 6      | 22          |
| Anopheles | arabiensis | 0      | 0      | 2      | 1      | 3           |
| Anopheles | coustani   | 0      | 0      | 4      | 0      | 4           |
| Anopheles | sp         | 0      | 0      | 25     | 0      | 25          |
| Culex     | carleti    | 0      | 1      | 9      | 0      | 10          |
| Culex     | quineuefasciatus | 0   | 1      | 12     | 34     | 47          |
| Culex     | sp         | 0      | 0      | 12     | 0      | 12          |
| Eretmapodites | subsimplicipes/quineuefasciatus | 35 | 77    | 524    | 168    | 804         |
| Uranotaenia | pandani  | 0      | 2      | 6      | 0      | 8           |
| Mansonia  | uniformis  | 0      | 0      | 2      | 0      | 2           |
| Total number of blood-sucking insects caught (effective trapping days*) | 36(5) | 108(8) | 1133(11) | 291(7) | 1568(31) |

*number of effective trapping days, i.e. days with the right climatic conditions (no wind or rain) to catch insects.
doi:10.1371/journal.pntd.0003045.t004

| Table 5. Comparison of average number of mosquitoes caught per trapping day per zone (Student’s t-Test), Union of Comoros, 2011. |
|---------------------------------------------------------------|
| **Genus** | **species** | Grande Comore | Moheli |
|-----------|------------|---------------|--------|
|           |            | Zone 1 | Zone 3 | Zone 5 |
| Anjouan   | Zone 6     | p = 0.005* | p = 0.012* | p = 0.116 |
| Moheli    | Zone 5     | p = 0.020* | p = 0.031* | -        |
| Grande Comore | Zone 3 | p = 0.280 | -      | -        |

* p-value significant if p < 0.05.
doi:10.1371/journal.pntd.0003045.t005
Figure 2. Trade in live animals between the Comoros archipelago, Madagascar and East Africa between 2007 and 2012. Trade in live animals between the Comoros archipelago, Madagascar and East Africa between 2007 and 2012 (Figure 2a) and imported animal dynamics (Figure 2b). Data used to create this map were gathered in surveys, or came from official sources or from personal communications.
doi:10.1371/journal.pntd.0003045.g002
References

1. Dauhney R, Hudson JR, Garnham PC (1931) Enzootic hepatitis or Rift Valley fever. An undescribed virus disease of sheep and cattle from East Africa. The Journal of Pathology and Bacteriology 34: 545–579.

2. Easterday BC (1965) Rift Valley fever. Advances in Veterinary Science 10: 65–127.

3. Rich KM, Wanyoike F (2010) An assessment of the regional and national socio-economic impacts of the 2007 Rift Valley fever outbreak in Kenya. American Journal of Tropical Medicine and Hygiene 83: 52–57.

4. McNutt BM, Russell D, dos Santos I, JH G (1980) RVF in humans in South Africa. South African Medical Journal 38: 803–806.

5. Bird BH, Ksiazek TG, Nichol ST, MacLachlan NJ (2009) Rift Valley fever virus. Veterinary Microbiology 133: 289–293.

6. Zeller HG, Fontenelle D, Traore-Lamizana M, Thioune Y, Dugotte JP (1997) Enzootic activity of Rift Valley fever virus in Senegal. American Journal of Tropical Medicine and Hygiene 56: 265–272.

7. Fontenelle D, Traore-Lamizana M, Diallo M, Thioune Y, Dugotte JP, et al. (1998) New vectors of Rift Valley fever in West Africa. Emerging Infectious Diseases 4: 289–293.

8. Linthicum KJ, Bailey CL, Dohrn DJ, Moulton JR. (1989) Transstadial and horizontal transmission of Rift Valley fever virus in Hyalomma truncatum. American Journal of Tropical Medicine and Hygiene 41: 491–496.

9. Dohrn DJ, Rowton ED, Lawyer PG, O’Guin M, Turell MJ (2000) Laboratory transmission of Rift Valley fever virus by Phlebotomus duboscqi, Phlebotomus papatasii, Phlebotomus Sergentei, and Sergentomyia schwetzii (Diptera: Psychodidae). Journal of Medical Entomology 37: 435–438.

10. APSSA (2008) Avis de l’APSSA sur le risque de propagation de la fièvre de la vallée du Rift (FVR) dans un département et une collectivité départementale français de l’Océan Indien (la Réunion et Mayotte). Maisonn-Allfort, France. 156 p.

11. Andriamandimby SF, Randrianirainavo-Solona J, Lejeune PF, Lejeune EM, Ravo- lolemavana L, Razafitraisea I, Fomenko A, et al. (2010) Rift Valley fever during Rainy Seasons, Madagascar, 2008 and 2009. Emerging Infectious Diseases 16: 963–970.

12. Madani TA, Al-Mazrou YI, Al-Jeffri MH, Mishkhas AA, Al-Rabeah AM, et al. (2003) Rift Valley fever epidemic in Saudi Arabia: epidemiological, clinical, and laboratory characteristics. Clinical Infectious Diseases 37(8):1084–92. Epub 2003 Sep 23.

13. Carroll SA, Reynes J-M, Khristova ML, Andriamandimby SF, Rollin PE, et al. (2011) Genetic evidence for Rift Valley fever outbreaks in Madagascar resulting from virus introductions from the East African mainland rather than enzootic maintenance. Journal of Virology 85: 6162–6167.
