Research Article

Network Pharmacology Integrated with Transcriptomics Deciphered the Potential Mechanism of *Codonopsis pilosula* against Hepatocellular Carcinoma

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Hepatocellular carcinoma (HCC) is the fourth main reason of cancer-related death. *Codonopsis pilosula* is a commonly used traditional Chinese medicine (TCM) for patients with HCC. However, its potential mechanism for treatment of HCC remains unclear. Here, we used transcriptomics and network pharmacology to explore the potential molecular mechanisms of *Codonopsis pilosula*. In our study, twelve differentially expressed genes (DEGs) (5 upregulated and 7 downregulated) of *Codonopsis pilosula* treating HepG2 cells (a kind of HCC cell) were identified. Among the 12 DEGs, HMOX1 may play an essential role. *Codonopsis pilosula* mainly affects the mineral absorption pathway in HCC. We acquired 2957, 1877, and 255 targets from TCMID, SymMap, and TCMSP, respectively. *Codonopsis pilosula* could upregulate HMOX1 via luteolin, capsaicin, and sulforaphane. Our study provided new understanding of the potential pharmacological mechanisms of *Codonopsis pilosula* in treating HCC and pointed out a direction for further experimental research.

1. Introduction

There are multiple types of primary liver cancer, of which hepatocellular carcinoma (HCC), the fourth leading cause of cancer-related death overall worldwide, is the most predominant type [1, 2]. During the last few decades, HCC incidence has been increasing at a global level [3, 4], and it is estimated that more than 1 million people will die from HCC in 2030 [5, 6]. In addition to surgical treatments, drugs are the key to HCC therapy [7]. Sorafenib has been the global treatment standard for patients with HCC since 2007 [8], but its efficacy is unsatisfactory [9]. As a widely used alternative therapy, traditional Chinese medicine (TCM) can probably prolong the median survival time and improve the overall survival among patients with HCC [10]. Moreover, some TCMs have been reported to have the ability to assist in elevating the efficacy of sorafenib in the treatment of HCC [11–13].

*Codonopsis pilosula*, a kind of TCM, has anticancer activity and is widely used in adjuvant anticancer therapy [14]. A lot of evidence has shown that many ingredients of *Codonopsis pilosula*, such as *Codonopsis pilosula* polysaccharide (CPP) and atractylenolide III (ATL), have anti-HCC effects via different pathways. CPP is one of major active constituents in *Codonopsis pilosula*, and it could inhibit the proliferation and motility of HCC cells through the β-catenin/TCF4 pathway [15]. CPP1a and CPP1c are two water-soluble homogeneous polysaccharides isolated and purified from *Codonopsis pilosula*, and they could induce HepG2 cell apoptosis by upregulating the ratio of Bax/Bcl-2 and activating caspase-3 [16]. ATL, a sesquiterpenoid extracted from *Codonopsis pilosula*, exerts tumor-suppressive functions in liver cancer via the miR-195-5p/FGFR1 signaling axis [17]. However, *Codonopsis pilosula*, as a kind of Chinese herb, is often used as a whole in clinical practice. There are few reports on the mechanisms of *Codonopsis pilosula* in the...
treatment of HCC, and its application is greatly limited. The effects of TCM (or herbs of other nations) are not the sum of all active ingredients. In the mixed system of *Codonopsis pilosula*, new effects may emerge that the single active ingredient does not.

In this study, we integrated transcriptomics and network pharmacology to understand the mechanisms of *Codonopsis pilosula* in treating HCC. The differentially expressed genes (DEGs) of *Codonopsis pilosula* were derived from a previous study (GSE115506) [18]. The effective ingredients of *Codonopsis pilosula* and targets were assayed by TCMID, SymMap, and TCMSP [19–21]. The mechanisms of *Codonopsis pilosula* against HCC were assessed by Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis. Furthermore, we found that *Codonopsis pilosula* may regulate mineral absorption through luteolin, capsaicin, and sulforaphane directly targeting HMOX1.

2. Materials and Methods

2.1. Differentially Expressed Genes Screening. We obtained DEGs of *Codonopsis pilosula* from the GEO database (https://www.ncbi.nlm.nih.gov/geo/) (series: GSE115506; samples: GSM3179695, GSM3179696, GSM3179697, GSM3179698, GSM3179699, and GSM3179700). In GSE115506, total RNA was isolated from HepG2 cells 24 hours after 3mg/mL *Codonopsis pilosula* aqueous extract treatment in vitro. We performed differential analysis by the Limma R packages [22], and the cutoff value for identifying DEG was set to |log2 fold change| >1 and adjusted p-value <0.05.

2.2. Components and Targets Acquisition. The components and targets of *Codonopsis pilosula* were acquired from TCMID (http://www.megabionet.org/tcmid/) [20], SymMap (http://www.symmap.org/) [21], and TCMSP (https://tcmsp.e.com/) databases [19]. They are all integrative databases of traditional Chinese medicine.

2.3. Network Building. We performed the protein-protein interaction (PPI) network analysis using STRING (https://string-db.org/) [23]. The *Codonopsis pilosula*-gene network, protein-protein interaction (PPI) network, and *Codonopsis pilosula*-component-target network were visualized by Cytoscape software [24].

2.4. Functional Enrichment Analysis. We conducted Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis and biological process (BP) of Gene Ontology (GO) analysis by R package clusterProfiler [25].

2.5. Expression Analysis of HMOX1. The expression of HMOX1 in HCC was obtained through UALCAN, which is a comprehensive and interactive web resource for analyzing cancer OMICS data [26]. The expression of HMOX1 after apigenin, luteolin, capsaicin, 4-methylsulfinyl butyl isothiocyanate (sulforaphane), and geniposide treatment was obtained from the GEO database (apigenin series: GSE119552, samples: GSM3377483, GSM3377484, GSM3377485, GSM3377486, GSM3377495, GSM3377496, GSM3377497, and GSM3377498; luteolin series: GSE18740, samples: GSM465440, GSM465441, GSM465442, GSM465443, GSM465444, and GSM465445; capsain series: GSE59727, samples: GSM1442972, GSM1442973, GSM1442974, GSM1442975, GSM1442976, GSM1442977, and GSM1442978; sulforaphane series: GSE28813, samples: GSM713517, GSM713518, GSM713519, GSM713520, GSM713521, GSM713522, GSM713523, and GSM713 524; and geniposide series: GSE85871, samples: GSM2286350, GSM2286351, GSM2286248, GSM2286249, GSM2286361, GSM2286317, GSM2286398, and GSM2286399). In GSE18740, mouse BV-2 microglia were treated with 50 μM luteolin for 24 hours; in GSE59727, rat TRPV1-positive neurons were treated with 10 μM capsaicin for 30 minutes; in GSE119552, MCF-7 cells were treated with 10 μM apigenin for 24 hours; in GSE28813, MCF10A cells were treated with 15 μM sulforaphane for 24 hours; and in GSE85871, MCF-7 cells were treated with 10 μM geniposide for 12 hours. We extracted the expression level of the HMOX1 gene from these expression matrices and compared its significance with the t-test.

2.6. Molecular Docking. The structure of HMOX1 protein was obtained from PDB (https://www.rcsb.org/) [27], and the structures of luteolin, capsaicin, and sulforaphane were acquired from ZINC (https://zinc.docking.org/) [28]. We used AutoDock 4.2 to prepare the PDBQT file and perform molecular docking [29]. Finally, molecular docking maps were visualized through PyMOL.

3. Results

3.1. *Codonopsis pilosula*-Gene Network and PPI Analysis. We identified 12 DEGs (5 upregulated and 7 downregulated) from the GSE115506 data set (Table S1). A volcano plot (Figure 1(a)) and a heatmap (Figure 1(b)) were established to show the distribution of DEGs in HepG2 cells after treating *Codonopsis pilosula*. The cutoff value for identifying DEG was set to |log2 fold change| >1 and the adjusted p-value <0.05. Accordingly, we built a *Codonopsis pilosula*-gene network (Figure 1(c)). In order to further explore the potential association among these DEGs, we performed a PPI network analysis for the 12 DEGs by STRING [23]. The final PPI network includes 11 nodes and 13 edges (Figure 1(d)). Furthermore, we identified HMOX1, an upregulated gene, as the hub gene because it has the highest degree.

3.2. GO and KEGG Analysis. Through the clusterProfiler R package for KEGG enrichment analysis, we found that only 1 pathway was significantly affected (padj <0.05) during *Codonopsis pilosula* treatment of HepG2 cells (Figure 2(a)). HMOX1 (hub gene), MT1F, and MT1G were enriched in mineral absorption. In total, 61 biological processes (GO terms) were notably enriched (padj <0.05) (Table S2). The top 5 biological processes are shown in Figure 2(b). The highly enriched biological processes include responses to
iron ions, cellular responses to copper ions, cellular transition metal ion homeostasis, transition metal ion homeostasis, and response to metal ions. These biological processes and pathways are closely related to the metabolism and homeostasis of metal ions, suggesting that *Codonopsis pilosula* mainly affects the mineral absorption pathway in HCC cells.

3.3. *Codonopsis pilosula* Reverses HMOX1 Expression in HCC

We acquired 2957 targets from TCMID, 1877 targets from SymMap, and 255 targets from TCMSP (Table S3). According to the intersection of these targets and 12 DEGs, we found that HMOX1 is the direct target of *Codonopsis pilosula* in three databases (Figure 3(a)). Our results showed that the expression of HMOX1 was significantly enhanced (Figure 1). Interestingly, HMOX1 was significantly decreased in HCC patients (Figure 3(b)). The abovementioned results suggest that *Codonopsis pilosula* may resist HCC by reversing HMOX1 expression in HCC patients.

3.4. *Codonopsis pilosula* Could Upregulate HMOX1 via Luteolin, Capsaicin, and Sulforaphane

To explore how *Codonopsis pilosula* promotes the expression of HMOX1, we established a *Codonopsis pilosula*-component-target network (Figure 4(a)). The result showed that *Codonopsis pilosula* may directly target HMOX1 through apigenin, luteolin, capsaicin, 4-methylsulfinyl butyl isothiocyanate (sulforaphane), and geniposide. In addition, we detected the expression of HMOX1 after treating with these components (Figures 4(b)–4(f)). Figure 4(b) showed that luteolin could upregulate Hmox1 in BV-2 cells, although the p value is 0.081. Figure 4(c) showed that capsaicin could significantly promote the expression of
Hmox1 in dorsal root ganglia neurons. Figure 4(d) showed that apigenin could not affect the expression of HMOX1 in MCF-7 cells. Figure 4(e) manifested that sulforaphane could dramatically enhance the expression of HMOX1 in MCF10A cells. Figure 4(f) showed that geniposide could not affect the expression of HMOX1 in MCF-7 cells. These results imply that Codonopsis pilosula could upregulate HMOX1 in HCC via luteolin, capsaicin, and sulforaphane.

Figure 2: KEGG and GO analysis. (a) KEGG pathway enrichment of *Codonopsis pilosula* treating HepG2 cells. (b) The top 5 biological process in GO terms of *Codonopsis pilosula* treating HepG2 cells.

Figure 3: *Codonopsis pilosula* could target HMOX1. (a) Among all targets and 12 DEGs of *Codonopsis pilosula*, HMOX1 is the only common gene. (b) HMOX1 expression is significantly decreased in HCC.
3.5. Potential Binding Site between Active Ingredients of Codonopsis pilosula and HMOX1 Protein. For exploring potential interaction between active ingredients of Codonopsis pilosula (luteolin, capsaicin, and sulforaphane) and HMOX1 protein, we predicted the potential binding site of them via molecular docking. As shown in Figure 5(a), luteolin may directly bind ASP-140, LEU-141, GLN-145, ALA-173, SER-174, and ALA-175 to promote the expression level of HMOX1. Capsaicin may combine with HMOX1 by ARG-44, LYS-48, and PHE-95, thus enhancing the
expression level of HMOX1 (Figure 5(b)). Sulforaphane may target HMOX1 via binding PHE-167 and ALA-175, resulting in increased expression of HMOX1 (Figure 5(c)). These results suggest that luteolin, capsaicin, and sulforaphane may promote HMOX1 expression through direct binding.

4. Discussion

TCMs are widely used during HCC treatment in China [30]. As with traditional medicine in other nations, herbal medicines are the main form of TCM [31]. Unlike small molecule drugs, herbal medicines contain many components and possess complex targets. Besides, some studies have revealed that miRNAs of herbal medicines may be ingested by the body and regulate the process of disease [32–34]. Complex components and targets limit the exploration of mechanisms in herbal medicines. Although multiple active components of Codonopsis pilosula were proved to have anti-HCC potential [15–17], the overall mechanism of Codonopsis pilosula is unclear.

In the present study, the Codonopsis pilosula-gene network was built by 12 striking DEGs (Figure 1(c)), and we identified HMOX1 as the hub gene via the PPI network (Figure 1(d)). HMOX1 was significantly enriched in the mineral absorption pathway, and the biological processes of its enrichment are primarily related to the metabolism of metal ions (Figures 2(a) and 2(b)). A study reported that there is a remarkable correlation between mineral absorption pathways and HCC development [35]. Furthermore, metal ion metabolism plays an essential role in the progression and treatment of HCC [36, 37]. Consequently, Codonopsis pilosula is highly likely to treat HCC via targeting HMOX1 to affect the mineral absorption pathway.

HMOX1 (heme oxygenase-1) is a stress-induced enzyme that catalyzes the degradation of heme to carbon monoxide, iron, and biliverdin [38]. The byproducts of HMOX1 enzymatic activity are cytoprotective because of their antioxidant and anti-inflammatory properties, showing that HMOX1 is a potential therapeutic target in many diseases [39]. Our results revealed that HMOX1 was significantly decreased in HCC patients (Figure 3(b)), and Codonopsis pilosula could distinctly enhance the expression of HMOX1 in HepG2 cells (Figure 1), suggesting that Codonopsis pilosula may reverse the expression pattern of HMOX1 in the HCC environment. Interestingly, HMOX1 overexpression could inhibit the growth, migration, and invasion in vivo, as well as higher HMOX1 expression was also associated with favorable disease-free survival of HBV-HCC patients who underwent hepatectomy [40]. These results indicate that Codonopsis pilosula is likely to improve the survival of HCC patients by promoting the expression of HMOX1, and is a potential adjuvant therapy for HCC.

Through the network pharmacology strategies, we built a Codonopsis pilosula-component-target network (Figure 4(a)). The network showed that Codonopsis pilosula may directly target HMOX1 via apigenin, luteolin, capsaicin, sulforaphane, and geniposide. To verify this result, we examined the effect of luteolin on Hmox1 expression in mouse BV-2 microglia (GSE18740), capsaicin on Hmox1 expression in rat TRPV1-positive neurons (GSE59727), apigenin on HMOX1 expression in human breast cancer cells MCF-7 (GSE119552), sulforaphane on HMOX1 expression in human breast epithelial cells MCF10A (GSE28813), and geniposide on HMOX1 expression in human breast cancer cells MCF-7 (GSE85871) (Figures 4(b)–4(f)). The results indicated that luteolin, capsaiacin, and sulforaphane could increase the expression of HMOX1 in vivo, although not in HCC cells. Luteolin, a natural flavonoid, plays multiple roles in the anti-HCC process. Growth inhibition of luteolin on HCC cells is induced via multiple signaling pathways of TGF-β1 pathways, p53 pathways, Fas/Fas-ligand pathways [41], ER stress [42], and AKT/OPN pathway [43]. Besides, a recent study reported that luteolin

![Figure 5: The molecular docking results of luteolin, capsaicin, and sulforaphane. (a) Luteolin may bind to HMOX1 with ASP-140, LEU-141, GLN-145, ALA-173, SER-174, and ALA-175. (b) Capsaicin may combine with HMOX1 by ARG-44, LYS-48, and PHE-95. (c) Sulforaphane may target HMOX1 via PHE-167 and ALA-175.](image-url)
could significantly inhibit HCC growth and cause apoptosis and cell cycle arrest in vitro and significantly suppress HCC growth in vivo via upregulating miR-6809-5p [44]. Capsaicin is a natural vanilloid and may inhibit the growth of SK-Hep-1 hepatocellular carcinoma cells by inducing apoptosis via Bcl-2 downregulation and caspase-3 activation [45]. Moreover, capsaicin could induce apoptosis in HepG2 cells by reducing the levels of xIAP and cIAP1 proteins, which are inhibitors of caspase-3 activation [46]. Interestingly, both luteolin and capsaicin are able to assist sorafenib to produce better anti-HCC therapeutic effects [47, 48]. Sulforaphane, a member of the isothiocyanate family, has exhibited promising inhibitory effects on breast cancer, lung cancer, liver cancer, and other malignant tumors [49]. Some studies revealed that sulforaphane could induce apoptosis [50] and enhance the radiation sensitivity [51] in HCC.

Notably, there are about 200 phytometabolites in *Codonopsis pilosula*, and the main bioactive ingredients include polysaccharides, polyene and polyacetylene glycosides, lignans, phenols, alkaloids, flavonoids, and lactones [52]. Polysaccharides are large-molecule components in *Codonopsis pilosula*, which have a significant inhibitory effect on gastric cancer and lung cancer, in addition to liver cancer [53]. Although luteolin, capsaicin, and sulforaphane are not the most abundant ingredients of *Codonopsis pilosula*, they are essential for understanding the pharmacological effects of *Codonopsis pilosula*. Tang et al. found that *Codonopsis pilosula* may play an antigastic cancer role through luteolin [54], suggesting that luteolin may play an important role in the anticancer effects of *Codonopsis pilosula*. Furthermore, several studies showed that luteolin [55], capsaicin [56], and sulforaphane [57] could target HMOX1 and significantly enhance its expression level. Luteolin, capsaicin, and sulforaphane are components of *Codonopsis pilosula*, but their quantitative studies in *Codonopsis pilosula* are insufficient. It is reported that the content of luteolin in *Codonopsis thalictrifolis* is 0.7% via HPLC [58]. It is important to notice that *Codonopsis thalictrifolis* is not *Codonopsis pilosula*, although they belong to the *Codonopsis* genus, and there may be great differences in chemical composition between them. Nonetheless, as a reference, the data implied that the content of the luteolin in *Codonopsis pilosula* may be less than 0.1% or even 0.01%. The lowest dose of luteolin that has been reported to produce anti-HCC effects in rats is 0.2 mg/kg via intraperitoneal injection [59]. In addition, orally administered luteolin (0.2 mg/kg) could produce anticolon cancer effects in rats [60]. Fuzheng Jiedu Xiaoji formulation (including 15 g of *Codonopsis pilosula*) could inhibit HCC progression in patients [61]. Jian Pi Li Qi Decoction (including 20 g of *Codonopsis pilosula*) could improve the prognosis of patients with HCC [62]. Therefore, it is likely that the effective concentration of luteolin can be reached in the application of *Codonopsis pilosula*. At present, there are no quantitative studies on capsaicin and sulforaphane in *Codonopsis pilosula*. Studies showed that capsaicin (2 mg/kg) [63] and sulforaphane (50 mg/kg) [64] could inhibit the growth of HCC in xenograft mice. Compared with the effective dosage of capsaicin and sulforaphane, the application of *Codonopsis pilosula* (15–20 g) is higher. Consequently, luteolin, capsaicin, and sulforaphane are likely to reach effective concentrations in the clinical application of *Codonopsis pilosula*. These studies suggest that *Codonopsis pilosula* is most likely to exert anti-HCC effects via luteolin, capsaicin, and sulforaphane (Figure 6). In fact, although our study showed that luteolin, capsaicin, and sulforaphane may play roles in the adjuvant treatment of HCC by *Codonopsis pilosula*, they may not be the main active ingredients of *Codonopsis pilosula*. As an herbal medicine, *Codonopsis pilosula* contains a variety of ingredients. An inulin fructan from *Codonopsis pilosula* possessed potential anti-HCC effects (inhibiting proliferation and inducing apoptosis) on Huh-7 and HepG2 cells without side effects on normal cells [65]. In addition, a novel fructose-enriched polysaccharide from *Codonopsis pilosula* inhibited HepG2 cell proliferation and promoted apoptosis [66]. Taken together, apoptosis may be one of the anti-HCC pathways of *Codonopsis pilosula*.

![Figure 6: Schematic diagram of *Codonopsis pilosula* for HCC treatment.](image-url)
5. Conclusions

The present study explored the effects of *Codonopsis pilosula* in the treatment of HCC via transcriptomics and network pharmacology. We revealed the transcriptome changes of HCC cells induced by *Codonopsis pilosula*. In addition, *Codonopsis pilosula* is likely to upregulate HMOX1 directly through luteolin, capsaicin, and sulforaphane, thus affecting the mineral absorption pathway in HCC cells. This study provides clues to comprehend the potential mechanisms of *Codonopsis pilosula* in treating HCC. Of course, these conclusions require further experimental support.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

There are no conflicts of interest.

Acknowledgments

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Supplementary Materials

Tables S1–S3 in the Supplemental files. Table S1: twelve DEGs in HepG2 cells after *Codonopsis pilosula* treatment. Table S2: all significantly enriched biological processes after *Codonopsis pilosula* treatment. Table S3: targets of *Codonopsis pilosula*. (Supplementary Materials)

References

[1] A. J. Craig, J. von Felden, T. Garcia-Lezana, S. Sarcognato, and A. Villanueva, “Tumour evolution in hepatocellular carcinoma,” *Nature Reviews Gastroenterology & Hepatology*, vol. 17, no. 3, pp. 139–152, 2020.

[2] D. Sia, A. Villanueva, S. L. Friedman, and J. M. Llovet, “Liver cancer cell of origin, molecular class, and effects on patient prognosis,” *Gastroenterology*, vol. 152, no. 4, pp. 745–761, 2017.

[3] Z. Liu, Y. Jiang, H. Yuan et al., “The trends in incidence of primary liver cancer caused by specific etiologies: results from the global burden of disease study 2016 and implications for liver cancer prevention,” *Journal of Hepatology*, vol. 70, no. 4, pp. 674–683, 2019.

[4] J. Liu, W. Tang, A. Budhu et al., “A viral exposure signature defines early onset of hepatocellular carcinoma,” *Cell*, vol. 182, no. 2, pp. 317–328, 2020.

[5] A. Villanueva and H. Carcmana, “Hepatocellular carcinoma,” *New England Journal of Medicine*, vol. 380, no. 15, pp. 1450–1462, 2019.

[6] J. C. Nault and A. Villanueva, “Biomarkers for hepatobiliary cancers,” *Hepatology*, vol. 73, no. S1, pp. 115–127, 2021.

[7] F. Foerster and P. R. Galle, “The current landscape of clinical trials for systemic treatment of HCC,” *Cancers*, vol. 13, no. 8, 2021.

[8] L. Rimassa, N. Personeni, C. Czauderna, F. Foerster, and P. Galle, “Systemic treatment of HCC in special populations,” *Journal of Hepatology*, vol. 74, no. 4, pp. 931–943, 2021.

[9] S. Busche, K. John, F. Wandrer et al., “BH3-only protein expression determines hepatocellular carcinoma response to sorafenib-based treatment,” *Cell Death & Disease*, vol. 12, no. 8, p. 736, 2021.

[10] X. Liu, M. Li, X. Wang et al., “Effects of adjuvant traditional Chinese medicine therapy on long-term survival in patients with hepatocellular carcinoma,” *Phytomedicine*, vol. 62, Article ID 152930, 2019.

[11] Y. Yang, M. Sun, W. Yao et al., “Compound kushen injection relieves tumor-associated macrophage-mediated immunosuppression through TNFR1 and sensitizes hepatocellular carcinoma to sorafenib,” *Journal for immunotherapy of cancer*, vol. 8, no. 1, 2020.

[12] W. Lam, Z. Jiang, F. Guan et al., “PHY906(KD018), an adjuvant based on a 1800-year-old Chinese medicine, enhanced the anti-tumor activity of Sorafenib by changing the tumor microenvironment,” *Scientific Reports*, vol. 5, no. 1, p. 9384, 2015.

[13] B. Zhai, F. Hu, H. Yan et al., “Bufalin reverses resistance to sorafenib by inhibiting akt activation in hepatocellular carcinoma: the role of endoplasmic reticulum stress,” *PLoS One*, vol. 10, no. 9, Article ID e0138485, 2015.

[14] L. Ye, Y. Jia, K. Ji et al., “Traditional Chinese medicine in the prevention and treatment of cancer and cancer metastasis,” *Oncology Letters*, vol. 10, no. 3, pp. 1240–1250, 2015.

[15] Y. Zhang, Y. Zhang, and H. Xu, “Effect of *Codonopsis pilosula* polysaccharides on the growth and motility of hepatocellular carcinoma HepG2 cells by regulating β-catenin/TCF4 pathway,” *International Journal of Polymer Science*, vol. 2019, Article ID 7068437, 7 pages, 2019.

[16] R. Bai, W. Li, Y. Li et al., “Cytotoxicity of two water-soluble polysaccharides from *Codonopsis pilosula* Nannf. var. modesta (Nannf.) L.T.Shen against human hepatocellular carcinoma HepG2 cells and its mechanism,” *International Journal of Biological Macromolecules*, vol. 120, pp. 1544–1550, 2018.

[17] L. Sheng, J. Li, N. Li et al., “Atractylenolide III predisposes miR-195-5p/FGFR1 signaling axis to exert tumor-suppressive functions in liver cancer,” *Journal of Food Biochemistry*, vol. 45, no. 5, Article ID e13582, 2021.

[18] P.-H. Ko, C.-W. Huang, H.-H. Chang, E. Y. Chuang, M.-H. Tsai, and L.-C. Lai, “Identifying the functions and biomarkers of *Codonopsis pilosula* and Astragalus membranaceus aqueous extracts in hepatic cells,” *Chinese Medicine*, vol. 14, no. 1, p. 10, 2019.

[19] J. Ru, P. Li, J. Wang et al., “TCMSP: a database of systems pharmacology for drug discovery from herbal medicines,” *Journal of Cheminformatics*, vol. 6, no. 1, p. 13, 2014.

[20] L. Huang, D. Xie, Y. Yu et al., “TCMID 2.0: a comprehensive resource for TCM,” *Nucleic Acids Research*, vol. 46, pp. D1117–D1120, 2018.

[21] Y. Wu, F. Zhang, K. Yang et al., “SymMap: an integrative resource for TCM,” *Nature Reports Gastroenterology & Hepatology*, vol. 12, no. 8, p. 736, 2021.
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[23] D. Szklarczyk, A. L. Gable, D. Lyon et al., “STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets,” Nucleic Acids Research, vol. 47, pp. D607–D613, 2019.

[24] P. Shannon, A. Markiel, O. Ozier et al., “Cytoscape: a software environment for integrated models of biomolecular interaction networks,” Genome Research, vol. 13, no. 11, pp. 2498–2504, 2003.

[25] G. Yu, L.-G. Wang, Y. Han, and Q.-Y. He, “clusterProfiler: an R package for comparing biological themes among gene clusters,” OMICS: A Journal of Integrative Biology, vol. 16, no. 5, pp. 284–287, 2012.

[26] D. S. Chandrashekar, B. Bashel, S. A. H. Balasubramanya et al., “UALCAN: a portal for facilitating tumor subgroup gene expression and survival analyses,” Neoplasia, vol. 19, no. 8, pp. 649–658, 2017.

[27] H. M. Berman, J. Westbrook, Z. Feng et al., “The protein data bank,” Nucleic Acids Research, vol. 28, no. 1, pp. 235–242, 2000.

[28] T. Sterling and J. J. Irwin, “Zinc 15—ligand discovery for replication and accelerates the negative conversion of infected Molecules and modern medicine from natural products,” Genes & Diseases, no. 16, pp. 2785–2791, 2009.

[29] H. Yuan, Q. Ma, L. Ye, and G. Piao, “Traditional medicine function,” of miRNAs from gastrodia elata blume and their potential supportive care for the management of liver cancer: past, no. 1, pp. 1–21, 2015.

[30] X. Liao, Y. Bu, and Q. Jia, “Traditional Chinese medicine as supportive care for the management of liver cancer: past, present, and future,” Genes & Diseases, vol. 7, no. 3, pp. 370–379, 2020.

[31] H. Yuan, Q. Ma, L. Ye, and G. Piao, “The traditional medicine and modern medicine from natural products,” Molecules, vol. 21, no. 5, 2016.

[32] L.-K. Zhou, Z. Zhou, X.-M. Jiang et al., “Absorbed plant MIR2921 in honeysuckle decoction inhibits SARS-CoV-2 replication and accelerates the negative conversion of infected patients,” Cell Discovery, vol. 6, no. 1, p. 54, 2020.

[33] C. Xia, H. Zhou, X. Xu et al., “Identification and investigation of miRNAs from gastodila elata blume and their potential function,” Frontiers in Pharmacology, vol. 11, no. 1477, 2020.

[34] Z. Zhou, X. Li, J. Liu et al., “Honeysuckle-encoded atypical microRNA2911 directly targets influenza A viruses,” Cell Research, vol. 25, no. 1, pp. 39–49, 2015.

[35] Y. Li, R. Chen, J. Yang et al., “Integrated bioinformatics analysis reveals key candidate genes and pathways associated with clinical outcome in hepatocellular carcinoma,” Frontiers in Genetics, vol. 11, p. 814, 2020.

[36] J. Wachsmann and F. Feng, “Molecular imaging and therapy targeting copper metabolism in hepatocellular carcinoma,” World Journal of Gastroenterology, vol. 22, no. 1, pp. 221–231, 2016.

[37] W. Wang, Q. Xie, X. Zhou et al., “Mitofusin-2 triggers mitochondrial Ca2+ influx from the endoplasmic reticulum to induce apoptosis in hepatocellular carcinoma cells,” Cancer Letters, vol. 358, no. 1, pp. 47–58, 2015.

[38] L. L. Dunn, S. M. Y. Kong, S. Tumanov et al., “Hmox1 (heme oxygenase-1) protects against ischemia-mediated injury via stabilization of HIF-1α (Hypoxia-Inducible factor-1α),” Arteriosclerosis, Thrombosis, and Vascular Biology, vol. 41, no. 1, pp. 317–330, 2021.

[39] T. D. Hull, A. Agarwal, and J. F. George, “The mononuclear phagocyte system in homeostasis and disease: a role for heme oxygenase-1,” Antioxidants and Redox Signaling, vol. 20, no. 11, pp. 1770–1788, 2014.

[40] C.-N. Yeh, R.-C. Wu, C.-T. Cheng et al., “HO-1 is a favorable prognostic factor for HBV-HCC patients who underwent hepatectomy,” Cancer Management and Research, vol. 10, pp. 6049–6059, 2018.

[41] S. B. Yee, H. J. Choi, S. W. Chung et al., “Growth inhibition of luteolin on HepG2 cells is induced via p53 and Fas/Fasligand besides the TGF-β pathway,” International Journal of Oncology, vol. 47, no. 2, pp. 747–754, 2015.

[42] Y. Lee and Y. H. Kwon, “Regulation of apoptosis and autophagy by luteolin in human hepatocellular cancer Hep3B cells,” Biochemical and Biophysical Research Communications, vol. 517, no. 4, pp. 617–622, 2019.

[43] E. Im, C. Yeo, and E.-O. Lee, “Luteolin induces caspase-dependent apoptosis via inhibiting the AKT/osteopontin pathway in human hepatocellular carcinoma SK-Hep-1 cells,” Life Sciences, vol. 209, pp. 259–266, 2018.

[44] P.-W. Yang, Z.-Y. Lu, Q. Pan et al., “MicroRNA-6809-5p mediates luteolin-induced anticancer effects against heptoma by targeting flotillin 1,” Phytomedicine, vol. 57, pp. 18–29, 2019.

[45] M.-Y. Jung, H.-J. Kang, and A. Moon, “Capsaicin-induced apoptosis in SK-Hep-1 hepatocarcinoma cells involves Bcl-2 downregulation and caspase-3 activation,” Cancer Letters, vol. 165, no. 2, pp. 139–145, 2001.

[46] S. P. Huang, J. C. Chen, C. C. Wu et al., “Capsaicin-induced apoptosis in human hepatoma HepG2 cells,” Anticancer Research, vol. 29, no. 1, pp. 165–174, 2009.

[47] N. Dai, R. Ye, Q. He, P. Guo, H. Chen, and Q. Zhang, “Capsaicin and sorafenib combination treatment exerts synergistic anti-hepatocellular carcinoma activity by suppressing EGFR and PI3K/Akt/mTOR signaling,” Oncology Reports, vol. 40, no. 6, pp. 3235–3248, 2018.

[48] X. Q. Feng, L. W. Rong, R. X. Wang et al., “Luteolin and sorafenib combination kills human hepatocellular carcinoma cells through apoptosis potentiation and JNK activation,” Oncology Letters, vol. 16, no. 1, pp. 648–653, 2018.

[49] G. Wu, Y. Yan, Y. Zhou et al., “Sulforaphane: expected to become a novel antitumor compound,” Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics, vol. 28, no. 4, pp. 439–446, 2020.

[50] S. B. Kntayya, M. D. Ibrahim, N. Mohd Ain, R. Iori, C. Ioannides, and A. F. Abdull Razis, “Induction of apoptosis and cytotoxicity by isothiocyanate sulforaphene in human hepatocarcinoma HepG2 cells,” Nutrients, vol. 10, no. 6, 2018.

[51] K. Ren, Z. Li, Y. Li, W. Zhang, and X. Han, “Sulforaphene enhances radiosensitivity of hepatocellular carcinoma through suppression of the NF-kB pathway,” Journal of Biochemical and Molecular Toxicology, vol. 31, no. 8, 2017.

[52] X. Zeng, J. Li, X. Lyu, J. Chen, X. Chen, and S. Guo, “Untargeted metabolomics reveals multiple phytometabolites in the agricultural waste materials and medicinal materials of Codonopsis pilosula,” Frontiers of Plant Science, vol. 12, 2022.

[53] J.-Y. He, N. Ma, S. Zhu, K. Komatsu, Z.-Y. Li, and W.-M. Fu, “The genus Codonopsis (Campanulaceae): a review of phytochemistry, bioactivity and quality control,” Journal of Natural Medicines, vol. 69, no. 1, pp. 1–21, 2015.

[54] L. Tang, J. Chen, J. Yin, and M. Fang, “Screening of active components and key targets of radix Codonopsis in the treatment of gastric cancer,” Journal of Chemistry, vol. 2021, Article ID 6056636, 10 pages, 2021.

[55] L. Li, R. Zhou, H. Lv, L. Song, X. Xue, and L. Wu, “Inhibitive effect of luteolin on sevoflurane-induced neurotoxicity through activation of the autophagy pathway by HMOX1,”
[56] K. Magierowska, D. Wojcik, A. Chmura et al., "Alterations in gastric mucosal expression of calcitonin gene-related peptides, vanilloid receptors, and heme oxygenase-1 mediate gastroprotective action of carbon monoxide against ethanol-induced gastric mucosal lesions," *International Journal of Molecular Sciences*, vol. 19, no. 10, 2018.

[57] W. N. Nowak, H. Tahe, J. Markiewicz et al., "Atorvastatin and conditioned media from atorvastatin-treated human hematopoietic stem/progenitor-derived cells show proangiogenic activity in vitro but not in vivo," *MEDIATORS OF INFLAMMATION*, vol. 2019, Article ID 1868170, 15 pages, 2019.

[58] X. Liu, J. Liang, S. Peng, L. Ding, and Y. Gu, "Analysis of luteolin from traditional tibetan medicine of Codonopsis thalictrifolis," *Chinese Journal of Analysis Laboratory*, vol. 28, pp. 263-264, 2009.

[59] K. Balamurugan and J. Karthikeyan, "Evaluation of luteolin in the prevention of N-nitrosodiethylamine-induced hepatocellular carcinoma using animal model system," *Indian Journal of Clinical Biochemistry*, vol. 27, no. 2, pp. 157–163, 2012.

[60] N. H. A. Osman, U. Z. Said, A. M. El-Waseef, and E. S. A. Ahmed, "Luteolin supplementation adjacent to aspirin treatment reduced dimethylhydrazine-induced experimental colon carcinogenesis in rats," *Tumor Biology*, vol. 36, no. 2, pp. 1179–1190, 2015.

[61] X. Yang, Y. Feng, Y. Liu et al., "Fuzheng Jiedu Xiaojie formulation inhibits hepatocellular carcinoma progression in patients by targeting the AKT/CyclinD1/p21/p27 pathway," *PhytoMedicine*, vol. 87, Article ID 153575, 2021.

[62] L. Xu, S. Wang, L. Zhuang et al., "Jian pi Li Qi decoction alleviated postembolization syndrome following transcatheter arterial chemoembolization for hepatocellular carcinoma," *Integrative Cancer Therapies*, vol. 15, no. 3, pp. 349–357, 2016.

[63] C. Xie, G. Liu, M. Li et al., "Targeting TRPV1 on cellular plasticity regulated by Ovol 2 and Zeb 1 in hepatocellular carcinoma," *Biomedicine & Pharmacotherapy*, vol. 118, Article ID 109270, 2019.

[64] J. Wu, J. Han, B. Hou, C. Deng, H. Wu, and L. Shen, "Sulforaphane inhibits TGF-β-induced epithelial-mesenchymal transition of hepatocellular carcinoma cells via the reactive oxygen species-dependent pathway," *Oncology Reports*, vol. 35, no. 5, pp. 2977–2983, 2016.

[65] N. Hu, Z. Gao, P. Cao et al., "Uniform and disperse selenium nanoparticles stabilized by inulin fructans from Codonopsis pilosula and their anti-hepatoma activities," *International Journal of Biological Macromolecules*, vol. 203, pp. 105–115, 2022.

[66] J. Yu, X.-D. Dong, J.-S. Jiao et al., "The inhibitory effects of selenium nanoparticles modified by fructose-enriched polysaccharide from Codonopsis pilosula on HepG2 cells," *Industrial Crops and Products*, vol. 176, Article ID 114335, 2022.