Expression profiling of *prospero* in the *Drosophila* larval chemosensory organ: Between growth and outgrowth

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**Abstract**

**Background:** The antenno-maxillary complex (AMC) forms the chemosensory system of the *Drosophila* larva and is involved in gustatory and olfactory perception. We have previously shown that a mutant allele of the homeodomain transcription factor Prospero (prosVoila1, V1), presents several developmental defects including abnormal growth and altered taste responses. In addition, many neural tracts connecting the AMC to the central nervous system (CNS) were affected. Our earlier reports on larval AMC did not argue in favour of a role of *pros* in cell fate decision, but strongly suggested that *pros* could be involved in the control of other aspect of neuronal development. In order to identify these functions, we used microarray analysis of larval AMC and CNS tissue isolated from the wild type, and three other previously characterised *prospero* alleles, including the V1 mutant, considered as a null allele for the AMC.

**Results:** A total of 17 samples were first analysed with hierarchical clustering. To determine those genes affected by loss of *pros* function, we calculated a discriminating score reflecting the differential expression between V1 mutant and other *pros* alleles. We identified a total of 64 genes in the AMC. Additional manual annotation using all the computed information on the attributed role of these genes in the *Drosophila* larva nervous system, enabled us to identify one functional category of potential Prospero target genes known to be involved in neurite outgrowth, synaptic transmission and more specifically in neuronal connectivity remodelling. The second category of genes found to be differentially expressed between the null mutant AMC and the other alleles concerned the development of the sensory organs and more particularly the larval olfactory system. Surprisingly, a third category emerged from our analyses and suggests an association of *pros* with the genes that regulate autophagy, growth and insulin pathways. Interestingly, EGFR and Notch pathways were represented in all of these three functional categories. We now propose that Pros could perform all of these different functions through the modulation of these two antagonistic and synergetic pathways.

**Conclusions:** The current data contribute to the clarification of the *prospero* function in the larval AMC and show that *pros* regulates different function in larvae as compared to those controlled by this gene in embryos. In the future, the possible mechanism by which Pros could achieve its function in the AMC will be explored in detail.

**Background**

In *Drosophila*, some external sensory organs found in the anterior region of larvae are composed of many neurons and support cells that seem to represent an aggregation of several sensory units. This is the case for the antenno-maxillary complex (AMC) that forms the chemosensory system of the *Drosophila* larva. The chemosensory apparatus of the larval head is formed during embryogenesis [1] and consists essentially of three major sensilla complexes on the cephalic lobe, the dorsal (DO), terminal (TO) and ventral organs (VO), and a series of pharyngeal sensilla [2,3]. While the DO appears to be a mixed smell and taste organ, the TO, VO and pharyngeal sensilla may be exclusively gustatory [4-7].

In previous studies, we described a mutant allele of the transcription factor *prospero* (Voila1, V1) that is
associated with several alterations in both AMC and the CNS [8]. V1 homozygotes die before forming pupae. Surviving larvae remain much smaller than wild-type individuals and are impaired for their response to salt and sucrose [9]. Using a set of previously characterised Volia alleles (prosV) that express different levels of Prospero (Pros) protein, we found that the level of Pros expression detected in the embryonic precursor region of the AMC, was related to the degree of alteration of larval taste [10]. In embryonic and larval AMC, Pros is expressed in the same cell cluster (~50 cells), including neuronal cells (~10 cells) and many accessory cells but no glial cells [8].

The pros gene encodes a transcription factor protein that contains a highly divergent putative homeodomain and a conserved Prospero domain that are both necessary for sequence-specific DNA binding and Prospero nuclear localisation [11-13]. Pros is known to be expressed in neuronal precursor cells [14,15] and participates in cell fate decision in both neuroblasts and sensory organ lineages [16-19]. Pros has been shown to control axonal and dendritic outgrowth [15], glial development [20,21] and to be a key regulator of mitotic activity in embryos [22]. Pros affects several cell cycle genes and can either promote or inhibit them depending on the cellular or the developmental context [23,24]. More recently Choksi et al. [25] showed that in the embryonic nerve cord Pros repressed target genes such as cell cycle genes required for self-renewal, and was also required to activate genes involved in terminal differentiation.

In a previous study on the larval antenno-maxillary complex, we showed that loss of pros function did not alter the mitotic activity or the final number of neurons. By contrast, many neural tracts connecting the AMC to the CNS are affected [8]. Therefore, it is possible that one key role of Pros in the larval AMC is to control the expression of genes involved in neuron-specific development such as axon routing and/or neurite outgrowth. However, as Pros is expressed in non-neuronal cells in the AMC (accessory cells), it is likely that, it regulates genes that are also involved in other functions.

In order to identify the Pros target genes associated with this organ, we performed microarray analysis on larval AMC tissue isolated from the wild type, the V1 mutant and two previously characterised prospero alleles [8], V13 (prosVoila13) and V24 (prosVoila24, see also Table 1). To establish the AMC specificity of these genes, we included analysis of samples from isolated larval CNS for these four alleles.

Our findings indicate that, in this sensory organ, pros is mainly associated with the regulation of genes that are essential for correct routing of neural processes and synaptic transmission. Many of these genes are involved in the development and remodelling of the nervous system during metamorphosis. Interestingly, we also found that loss of pros function induced the misregulation of a subset of genes important for growth, and autophagy. Finally, the possible role of EGFR (the epidermal growth factor receptor) and the N (Notch) pathway in regulating all of these functions is discussed.

Results
The developing AMC and Pros expression
We have previously shown [8] that in the developing AMC, Pros is always expressed in the same cluster of cells. In addition, neither mitotic activity nor apoptosis was observed during the third instar larval stage or at late embryonic stages suggesting that the final number of Pros expressing cells is fixed before the end of embryogenesis [8]. This hypothesis was further confirmed by analysing mitotic activity in the developing wild type AMC (prosVoila14, V14) using an H3p marker. Our results showed that the H3p labelling disappeared completely after the embryonic stages 12-13 [Additional file 1] indicating that additional cells are not provided until the last larval stage. However, some Pros expressing cells grew in size at the LIII stage. Scoring the different Pros+ cell type morphology (Figure 1, Table 2), we found that the wild type larval AMC (TO and DO) was composed of 8 (± 1) large Pros+ cells (most probably accessory cells) and 40 (± 4.1) small Pros+ cells. Among the latter, 10.7 (± 2.8) were neuronal cells [8]. Pros is never expressed in glial cells. Interestingly, pros loss of function affected the axonal pathway in the embryonic AMC, but produced the correct number of neuronal cells [8] and curiously induced an excess of glial cells, which, we suspect, originated from incorrect peripheral glial cells migration. Therefore, if Pros was expressed in the same number of cells in the embryo and larvae and since no additional cell division was seen after stage 13, it is likely that pros is not involved in cell fate choice in the larval AMC.

To better clarify the role of pros in the AMC, we carried out microarray analysis on wild type (V14) and three prospero mutants (V1, V13, V24, see also Table 1), which present different expression levels of Pros [8,10]. V1 is considered as a null pros allele for the larval AMC as no Pros protein is detected in this organ. It presents an abnormal taste response to sucrose and NaCl (indifferent to both substances), and shows an alteration of the neural connections between the AMC and CNS as well as arborisation defects in larval neuromuscular junction [10]. In the V1 larval CNS, Pros is still expressed but at a lower level than in the wild type V14. V1 larval CNS also shows several defects, which include early initiation of cell death and abnormal sub-cellular localization of the Pros protein [Additional file 2: supplemental figure A].
Table 1 Overview of the phenotypes associated with the different prosV alleles.

| Allele | Genotype                                      | Stage of lethality | Larval taste response | Pros expression in AMC | Pros expression in CNS | Axonal routing in AMC |
|--------|-----------------------------------------------|--------------------|-----------------------|-------------------------|------------------------|-----------------------|
| V14    | Wild type (complete pGal4 remobilization)     | Viable             | Normal                | Normal                  | Normal                 | Normal                |
| V13    | Partial pGal4 excision (remaining of 718 bp)  | Young adult < 2 days old | Normal                | Normal                  | altered                | Normal                |
| V24    | Partial pGal4 excision (remaining of 7400 pb) | Pupal              | Intermediate          | Normal                  | altered                | Normal                |
| V1     | Full length pGal4 transposon (12900 pb)       | Larva              | Altered               | absent                  | altered                | Misrouting            |

(redrawn from Guenin et al. [8]).

In the prosV1 (V1) allele, the full length pGal4 transposon is inserted upstream of the pros coding region (-216 bp). prosV14 (V14) results from the correct and total remobilization of the transposon, in this strain the wild type phenotype is restored. In prosV24 (V24) and prosV13 (V13), the pGal4 element has been partially removed, respectively 7400 and 718 bp remain inserted 216 bp upstream the pros start site. The peak of developmental lethality, the taste response of late homozygous 2nd instar larva, Pros expression level in larvae and axonal misrouting are indicated for each prosV allele.

The larval taste response was measured towards 0.1 M sucrose and 0.3 M NaCl concentration that are known to respectively attract or repulse wild type Drosophila. V1 mutants were indifferent to both substances (altered taste response), V24 showed an intermediate response: they were repulsed by NaCl but remained indifferent to sucrose and V13 and V14 present a normal taste response to both substances. The Pros expression pattern is indicated by comparison to the V14 wild type: In the AMC, V1 showed no Pros expression but for the other alleles, Pros expression pattern was similar to V14. In the CNS, all mutant alleles showed a distinct altered expression pattern as compared to the wild type (further descriptions of the Pros pattern are found in the text and in [Additional file 2]).

Figure 1 AMC region from third instar larvae observed by optical microscopy. (A) Bright-field view of the larval AMC region (dorsal view, anterior down), the hooks appear in dark. Cells that constitute the AMC are located on either side of the hooks. (B) 3D reconstruction of AMC (TO +DO), labelled with Pros (red) and Elav (green) that labels neuronal cells. (B'1-3) Zoomed view of a confocal section of the framed region in B showing respectively the Pros (B'1), Pros/Elav (B'2) and Elav (B'3) staining. Anti-Prospero labels two types of Pros expressing cells (Pros+): large (arrowheads in B'1) and small cells (arrows in B'1). Some of the small Pros+ cells express Elav (B'2). Scale bars represent 10 μm.
Table 2 Pros expressing cells in the larval AMC

| Cell types | Large cells | Small cells |
|------------|-------------|-------------|
| Alleles    | Pros+       | Pros+       | Pros+/Elav+ | Elav+       |
| V14        | 8 ± 1       | 409 ± 41    | 10.7 ± 28   | 65.8 ± 14   |
| V1         | 0 ***       | 0 ***       | 0 ***       | 62.3 ± 0.9  |

We have quantified the number of Pros expressing cells (Pros+) and neuronal cells (Elav+) in the third instar larval AMC of wild type (V14) and V1 mutants. We distinguish two types of Pros+ cells on the basis of their size: large and small cells. Some small Pros+ cells express Elav markers and are probably differentiated neurons. In V1 mutants, no more Pros protein is detected in the larval AMC, but the number of neurons remains unchanged.

V13 and V24 are both derived from the V1 allele and result from incomplete excision of the PGa14 transposon, respectively 7400 and 718 bp remain inserted upstream of the pros start site (see also Table 1). A previous study showed that V13 [8] and V24 (personal communication) present a correct structure of the larval AMC and a normal expression level of pros mRNA. No variation was observed in the number of glia or neuron cells and the pattern of Pros expression was similar to that of the wild type V14. Since, V14, V24 and V13 present the same expression pattern in the AMC and have the same genetic background, individual variations (independent of Pros expression), can be more easily eliminated by the use of several but similar fly lines.

In the CNS both alleles overexpress (at different levels) the pros mRNA but present a distinct pattern as regards to the mitotic activity or to the Pros and Elav (neuronal cells marker) labelling [Additional file 2]. In the ventral nerve cord V13 showed a clear hyperplasia due to an excess of neurons [Additional file 2: supplemental figure B]. Finally, mitotic activity, revealed by anti-pHistone-H3 antibody (H3p) [Additional file 2: supplemental figure C] was strongly increased in the Optic Lobes of V24. In conclusion, each of V24 and V13 allele presented a distinct abnormal pattern as compared to the wild type V14. Therefore, we used in the CNS V13 allele only as its Pros expression pattern was previously published.

All these alleles were used to for the transcriptome analysis since they have the same genetic background.

Most specifically, to identify the genes that are misregulated in the AMC, the comparison was made between the null AMC V1 mutant and the other alleles V14, V24, V13 as all three present the same wild type expression pattern in this structure.

It should be mentioned that the role of Pros in the larvae CNS was not investigated in this study. The CNS was used to be compared with AMC (for this only V14, V2 and V13 alleles were selected) and to determine whether the putative candidate genes identified in the AMC could be found in the CNS. For this latest purpose, we decided to limit the microarray analysis in the CNS to the comparison between V14 and V1, since the V13 allele presents an intermediate Pros expression pattern in the CNS. This avoids misinterpretation of the CNS data.

Transcription profile of prosV1 larvae

Expression data of a total of 17 samples were analysed, including both CNS and AMC samples for the four prosV alleles and 2 to 3 independent RNA extractions for each allele. We searched for sets of genes participating in the same biological function (with correlated expression) and differentially expressed between prosV mutants. We used the Discriminating Score (DS, see also Methods section) as the detection method for differential expression, smoothed on the hierarchical clustering tree to detect peaks of correlated genes. This method had the advantage of detecting peaks of optimal size. This size could not be known a priori. A DS score can be assigned to each node in the dendrogram. The node corresponding to the maximum DS score was chosen as the node best fitting the peak.

The results were displayed using TreeView [26]. As it can be seen in the Figure 2A, a cluster of genes differentially expressed in AMC and CNS tissues and in a Pros independent manner was observed. The genes of this cluster (AMC tissue specific signature) were clearly overexpressed in the AMC while the same genes were underexpressed for all alleles in the CNS (Figure 2A).

In the next step, we determined those genes affected by loss of pros function for each organ with the DS. For the AMC, as V13, V14 and V24 have a normal Pros expression pattern in this structure, the DS was calculated for each gene between V1 and all other alleles. For the CNS, since V13 exhibited a distinct pattern [Additional file 2], the DS for each gene was calculated between V1 and the V14 alleles only.

To visualize groups of correlated genes that were differentially expressed between V1 and other alleles, the DS score obtained for the AMC or CNS was plotted alongside the hierarchical clustering and smoothed in a sliding windows of 100 genes. As shown in Figure 2B, different peaks can be detected. Each peak represents co-expressed genes sensitive to Pros expression. To avoid the analysis of non-significant variations, we decided to assess the biological functions of these groups of genes. We therefore searched for significant enrichments of Gene Ontology terms (GO) in each cluster using GoMiner [27]. In the AMC, only 3 peaks (peaks 1-3) could be associated with significant GO functions (Figure 2B peaks 1, 2 and 3). Complete gene lists for peaks 1-3 are presented in [Additional file 3 and 4].

In peak 3, the 26 genes overexpressed in V1 AMC were significantly associated with the over-represented GO term « signal transducer activity » (GO:0004871, p = 0.0008, see also Table 3). Significant enrichments of
the GO term “proteasome complex” (GO:0000502, \( p < 10^{-5} \), see also Table 3) were found for 9 genes in peak 2. All of these genes were underexpressed in \( V1 \) AMC. Peak 1, with the highest DS, was common to AMC and CNS and associated with the over-represented GO term “cell fate commitment” (GO:0045165, \( p = 0.0003 \)). Inside this peak, a cluster of 29 genes overexpressed in \( V1 \) AMC and highly correlated (\( r > 0.9 \), Figure 2C, Table 3) was isolated and we found a cluster of 86 genes overexpressed in \( V1 \) CNS and correlated (\( r > 0.9 \), Figure 2D, see also [Additional file 5: Supplemental Table S1]) in the same peak. Interestingly, among the 86 CNS genes, 28 (represented in red in the Figure 2D) also belonged to the AMC gene list (see also Table 3 and [Additional file 5: Supplemental Table S1]).

Finally, to assess our microarray analysis, we quantified the expression of seven selected genes by Q-PCR. As shown in Table 4, our results were consistent with
the microarray data except for the *hb* (*hunchback*) gene found to be overexpressed in the CNS but not in the AMC. Because the two methods have different sensitivities, the magnitude of the change determined by microarray and real time PCR is not the same. The orientation of changes, however, is identical.

An interesting observation was that we found highly correlated genes responding to Pros variation in both CNS and AMC. This tight correlation in two different tissues could suggest that these genes may be controlled by common transcription factors including Pros. We searched for transcription factor binding sites that were common to these 28 genes. We used a Gibbs sampling method [28] on the -1700 to +300 bp promoter region of these genes. This method allowed the determination of degenerated motifs, described by a position weight matrix (PWM), in a set of sequences by iterative sampling.

We found a motif shown as a Logo [29] in Figure 2E. It notably included a CAGCTG core. This motif shows weak and probably not significant similarities with other Pros motifs previously proposed for *Drosophila*: TAAGNCT [25], CACNNCT [12], TAAGACG [30]. Therefore, additional experiments are now necessary to see whether the motif identified in this study could really bind the Prospero transcription factor in vivo.

### Additional manual annotation to specify the role of pros putative target genes in *Drosophila* larvae

Our microarray analyses showed that peak 1 contained 29 overexpressed genes associated with the GO annotation, “cell fate commitment” (Table 3). These data are not consistent with our previous studies showing that *Pros* is not involved in cell fate determination in the larval AMC [8]. We were intrigued by this discrepancy and therefore we looked more deeply for the function of these genes in the larvae. Interestingly, though most of the genes present in this peak were associated with cell fate determination in embryos, such evidence was mostly missing for the larvae peripheral nervous system (PNS). Thus, it is likely that the GO annotation was mostly deduced from the reported function of these genes in *Drosophila* embryos. Therefore, to more specifically identify the role of these genes in the *Drosophila* larva PNS, we used additional manual annotation.

The first step consisted in compiling all of the information available on the role attributed to each of the 64 genes identified from peaks 1-3, but most specifically in *Drosophila* larvae. The information was collected using Flybase, mutant analysis, associated phenotypes, research articles and microarray data. As much as possible, we selected only data that reported the function of these genes in the larval nervous system and more specifically the sensory system. Out of the 64 genes, we found that 27 had unknown biological functions or had not been

| Genes          | Symbol  | Biological function in larvae (manual annotation) |
|----------------|---------|--------------------------------------------------|
| aEst1**        | CG1031  | Sensory neuron morphogenesis                      |
| Art*           | CG6563  | Not studied in larvae                             |
| Ash2*          | CG6677  | Neurite outgrowth, synapse formation, growth, sensory organ development |
| CG10632*       | CG10632 | Unknown                                           |
| CG10671*       | CG10671 | Unknown                                           |
| CG3021*        | CG3021  | Unknown                                           |
| CG31637*       | CG31637 | Unknown                                           |
| CG31961*       | CG31961 | Unknown                                           |
| CG31731*       | CG31731 | Unknown                                           |
| CG6388         | CG6388  | Neurite outgrowth                                 |
| CG7878*        | CG7878  | Unknown                                           |
| CG8155*        | CG8155  | Unknown                                           |
| DPALT1*        | CG17310 | Neuropeptide biosynthesis                         |
| FK506-6 bp1*   | CG6226  | Autophagy; growth                                 |
| Ftz-F1*        | CG4059  | Autophagy, sensory organ formation, olfaction     |
| Hb*            | CG9786  | Labial segment formation including sense organ    |
| Iap2*          | CG8293  | Autophagy, sensory organ development              |
| Inx3*          | CG1448  | Not studied in larvae                             |
| Keren*         | CG32179 | Autophagy, sensory organ development              |
| Mbo*           | CG6819  | Tracheal system development                        |
| Nak*           | CG10637 | Not studied in larvae                             |
| Nej*           | CG15319 | Synaptic transmission, autophagy                  |
| Notch*         | CG9396  | Neurite outgrowth, nutrient sensing/growth, sense organ formation, olfaction |
| Pelo*          | CG9395  | Not studied in larvae                             |
| Psq*           | CG3638  | Sensory organ development, olfaction              |
| Rac1*          | CG2248  | Neurite outgrowth, sensory organ development       |
| Tollo*         | CG6890  | Synaptogenesis, wing development, immune response |
| TFIIs*         | CG3710  | Not studied in larvae                             |
| Lk6            | CG17342 | Autophagy growth, nutrient sensor mechanism, Proteasome complex (GO:000502, p < 10^-5) |
| Pros 26.4      | CG5289  | Neuronal remodelling, Autophagy                   |
| ProsB2         | CG3329  | Neuronal remodelling, Synaptic transmission, autophagy, sensory organ formation |
| Pros26         | CG4097  | Neuronal remodelling, Synaptic transmission, autophagy |
| Prosα5         | CG18495 | Neuronal remodelling, Synaptic transmission, autophagy |
| Prosα7         | CG1519  | Neuronal remodelling, Synaptic transmission, autophagy |
| ProsM5         | CG10938 | Not studied in larvae                             |
| RPN1           | CG7762  | Neuronal remodelling, Autophagy                   |
| RPN2           | CG11888 | Neuronal remodelling, Autophagy                   |
| RPN5           | CG1100  | Neuronal remodelling, Autophagy                   |

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**Table 3: Genes identified as putative Pros targets and their manual annotation**

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Analysis of the resulting compiled data on larva [Additional file 5: Supplemental Tables S2-S4] revealed that the 37 genes fell into at least one of the following functional categories: (1) neurite outgrowth and/or synaptic transmission; (2) growth, autophagy; (3) sensory organ (mainly olfactory) development. A list of these genes and their associated annotation terms is summarized in Table 3, and a schematic representation is given in Figure 3. As shown in Figure 3, some genes can be associated with two functional classes, and four genes (EGFR, Notch, Ash2 and pros) are associated with the three functional categories: neurite outgrowth, autophagy, and olfactory system development.

Genes involved in neural processes and synaptic transmission are misregulated in the V1 AMC mutant

One of the functional categories deduced from manual annotation associates some putative Pros target genes with neurite outgrowth and/or synaptic transmission (Figure 3). Although synaptic transmission and neurite outgrowth belong to different functional categories, we decided to keep these genes in the same class since many of them are involved in both synaptic transmission and neurite outgrowth.

Mostly, the genes are overexpressed in the larval V1 AMC, (Table 3). It has been reported that the upregulation of most of these candidates inhibits neurite outgrowth in larval neurons (reported phenotypes are shown in [Additional file 5: Supplemental Table S2]). This is clearly the case for EGFR (Epidermal Growth Factor Receptor), N (Notch), bnl (branchless) and Rac1 [31-33] whose overexpression was previously shown to inhibit axon extension in larval neurons. This is also the case for gwl (greatwall), limK1 (lim-kinase 1), Nej (Nejire) or CG6388, Pvr, whose upregulation induces axon pathfinding defects or impaired neurotransmitter release in the larval neuromuscular junction (NMJ) [34-38]. Interestingly, similar axon pathfinding and NMJ defects were observed in the V1 mutant [8].

We have noticed that most of the genes included in this functional class can also drive neural connectivity remodelling in larvae, a process particularly important during metamorphosis. EGFR, Notch, bnl, Rac1 [31-33], and the genes associated with the ubiquitin-proteasome system were all reported to be involved in axon

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Table 3: Genes identified as putative Pros targets and their manual annotation (Continued)

| Genes | Manual Annotation | GO Term |
|-------|------------------|---------|
| Bnl   | CG4608            | Neurite outgrowth |
| CaMKII| CG1495            | Synaptic transmission |
| CG10011| CG10011           | Unknown |
| CG10702| CG10702           | Autophagy |
| CG1088| CG10882           | Unknown |
| CG31714| CG31714          | Unknown |
| CG4839| CG4839           | Unknown |
| CG5790| CG5790           | Unknown |
| CG7536| CG7536           | Unknown |
| CG7800| CG7800           | Unknown |
| CRIS α | CG17520      | Sensory organ development |
| Dok | CG2079         | Sensory organ development |
| EGF | CG10079        | Neurite outgrowth, synapse formation, growth, autophagy sensory organ development, olfaction |
| feo | CG11207       | Mitotic spindle organisation |
| Gek | CG4012        | Actin polymerisation |
| Gwl | CG7719         | Neurite outgrowth, synaptic transmission, mitotic cell cycle |
| InaC | CG6518       | Not studied in larvae |
| Kdr | CG1848        | Neurite outgrowth, synaptic transmission |
| LimK1 | CG1848         | Cell cycle, DNA damage checkpoint |
| Loco | CG5248        | Not studied in larvae |
| Mcr | CG7586       | Olfaction |
| PmK | CG1830        | Not studied in larvae |
| Pvr | CG2222       | Hemocyte formation, dorsal closure, macrochaete formation |
| Rho | CG5638       | Not studied in larvae |
| Toll-6 | CG2500       | Not studied in larvae |

The 64 genes found highly correlated in the peak 1, 2 and 3 are grouped according to their respective GO annotation classes. The most significant classes of genes enriched in our list are “Cell fate commitment”, “proteasome complex” and “signal transduction”. The p value indicates the probability for a given ontology to be associated at random to this cluster. The first 28 genes indicated in bold and by an asterisk share a common DNA motif (CAGCTG) in their promoter and were also found to be differentially expressed between V1 and V14 CNS.

The last column on the right specifies the known biological function (manual annotation) of these genes in the Drosophila larvae (Full references can be found in the main text and in [Additional file 5: Supplemental Tables S2-S4]). This manual annotation allowed the attribution of biological function to 37 genes. Some genes have either never been studied in larvae or their respective functions are currently unknown. By contrast to the GO annotation (mostly deduced from embryos), the use of manual annotation indicates that the dysfunction of pros leads in larvae to the misregulation of genes that mostly deal with neurite outgrowth, growth and autophagy and sensory organ formation (mainly olfactory). All genes were found to be overexpressed in the mutant V1 AMC except for the 9 genes associated with the Proteasome complex GO annotation.

documented in larvae (Table 3). Since the genes encoding the subunit of the proteasome complex are underexpressed and the remaining genes are overexpressed in the V1 AMC, a description of the phenotype generated by the upregulation and downregulation of the corresponding genes in larvae was given [Additional file 5: Supplemental Tables S2-S4]. Though many phenotypes were available for a gene, we selected only those reported for the nervous system and preferentially for the PNS.
Figure 3 Schematic representation of the overlapping function attributed to the AMC putative Pros target genes. The functional categories were established using a manual annotation (the criteria used for this annotation are indicated in the text, see also for further phenotypic description and corresponding references [Additional file 5: Supplemental Tables S2-S4]). The three functional groups identified are represented by three distinct colored sets. The genes located at the intersection between two sets can assume both functions. It should be noted that the genes indicated in black (EGFR, Notch, Ash2 and prosβ2) belong to the three functional groups: neurite outgrowth, sensory organ development, and growth/autophagy.

Table 4 Validation of microarray data using real time PCR.

| Gene | Primers | Microarray | Q-PCR | Microarray | Q-PCR |
|------|---------|------------|-------|------------|-------|
| caps | F S'GCAGCCTTGGATGAAGTTTA 3'  
R S'ATGGGCGACCCCATAGTGC 3' | 1.38 | 0.63 | 3.8 | 2.38 |
| Cdk4 | F S'TACCAACGCACGTGCCACAT 3'  
R S'GGGCGACCCCATAGTGC 3' | 0.95 | 1.3 | 4.99 | 2.5 |
| hb   | F S'CCTTCCAGTCGACAAATG 3'  
R S'ATCCGCAACCGTACTGA 3' | 6.71 | 0.85 | 6.38 | 1.6 |
| lap2 | F S'AAGGACTTGCACCTCACAACAT 3'  
R S'CGTTCGACACAGAACATC 3' | 3.69 | 2.16 | 6.48 | 1.9 |
| nak  | F S'AGGAAGCATACACAAATG 3'  
R S'GGACCGACCCCATACG 3' | 1.75 | 1.36 | 0.97 | 1.95 |
| neq  | F S'ATGATCCATACACATCTCT 3'  
R S'CCTGGATCAACCGGATACTAT 3' | 3.26 | 1.63 | 3.79 | 3.75 |
| Notch| F S'AGACACGTCGCGGAACTCCG 3'  
R S'GGTTCTTGCACTGGTCT 3' | 2.9 | 1.76 | 8.96 | 2.52 |

The relative expression level (V1/V14) of selected genes was measured using the Q-PCR or microarray analysis data. Our results were consistent with the microarray data except for the hb (hunchback) gene found to be overexpressed in the CNS but not in the AMC. The values in gray correspond to the genes found differentially expressed between V1 and V14 in the CNS but not in the AMC using microarray analysis. Accordingly, no significant variation was found for these genes in the AMC, using Q-PCR.

Figure 3 Schematic representation of the overlapping function attributed to the AMC putative Pros target genes. The functional categories were established using a manual annotation (the criteria used for this annotation are indicated in the text, see also for further phenotypic description and corresponding references [Additional file 5: Supplemental Tables S2-S4]). The three functional groups identified are represented by three distinct colored sets. The genes located at the intersection between two sets can assume both functions. It should be noted that the genes indicated in black (EGFR, Notch, Ash2 and prosβ2) belong to the three functional groups: neurite outgrowth, sensory organ development, and growth/autophagy.
extension/retraction, pruning and morphogenesis of larval peripheral sensory neurons [Additional file 5: Supplemental Table S2]. Indeed, though most larval sensory neurons will degenerate during metamorphosis, some persist as neurons and undergo stereotyped pruning of their dendrites and axon terminal branches during early metamorphosis [39].

We were not surprised to find that the genes encoding the different proteasome subunits (Prosα7, Prosβ2, Prosα6, Prosα26) are downregulated in V1 AMC (Figure 3, Table 3). Indeed, the acute regulation of their protein level is a primary determinant of protein turnover and neurotransmission strength [40,41].

Recently, an elegant study of Choksi et al. [25] showed that pros is required for activation of neuronal differentiation genes in embryos and identified N, bnl, LimK1, EGFR and PVR, prosα6 as putative pros targets in embryos. This reinforces our finding suggesting that in the larval AMC, pros plays a crucial role in the modulation of neuronal activity through the control of genes involved in neurotransmission and synaptic plasticity.

**Loss of pros function alters the expression of genes involved in autophagy and growth**

The second functional group that emerges from our analysis includes candidates that play a critical role in the control of autophagy, a process used to provide energy and nutrients during metamorphosis and early adulthood.

The association between Pros and the regulation of autophagy is mainly attested by the upregulation of genes such as CG10702, EGFR, Keren (EGFR ligand), Ftz-F1 (Ftz transcription factor 1), FKS06-bp1 (FK506-binding protein 1), lap2 (Inhibitor of apoptosis 2), nej, Notch and genes associated with proteasome complex (Table 3, Figure 3, see also [Additional file 5: Supplemental Table S3]).

Some of the genes cited above were also found to mediate cell growth (Table 3 and [Additional file 5: Supplemental Table S3]). However, it is not yet clear if the overexpression of these genes systematically inhibits cell growth. For example, the upregulation of LK6 (protein serine/threonine kinase) or FKS06-bp1 leads to either the activation or inhibition of cell growth in a context-dependent way [42-44].

We have already mentioned that Notch and EGFR pathways were involved in neurite outgrowth (see above), interestingly, we found that these two pathways were also associated with both the regulation of autophagy and cell growth control [45,46] (Figure 3), suggesting that Pros could mediate all of these functions through the modulation of these two pathways.

Our finding that pros is associated with the expression of genes involved in growth and or autophagy is consistent with the phenotypic defects observed in V1 homozygote mutants: i) individuals died before reaching puparium formation; ii) surviving larvae and pupae remained much smaller than wild-type individuals; iii) numerous labelled cells were observed in the fat body using PGal4 enhancer trap line V1 [9].

It is interesting to note that many of the genes found in this functional group are directly or indirectly associated with insulin-signalling pathways and more specifically the insulin/TOR (target of rapamycin) pathway, an important mediator of growth, autophagy and nutrient sensing [Additional file 5: Supplemental Table S3].

**Pros and the olfactory system**

Pros was detected both in the terminal (TO: mainly gustatory) and in the dorsal (DO: mainly olfactory) organs of the larval AMC [8]. Accordingly, we found that pros loss of function in the AMC induced the upregulation of all candidate genes (except prosβ2) that were known to be involved in the development of sensory organs (Figure 3, Table 3). Most of our knowledge on the function of these genes came from studies done on adult *Drosophila* sensory organs [Additional file 5: Supplemental Table S4]. For example, it has been reported that mutations in the genes ash2 or ckl1 alpha (Casein kinase II alpha subunit) can elicit supernumerary or ectopic adult sensory organs [47,48]. Similarly, overexpression of lap2 or limK1 induces respectively additional macrochaetes [49] or ectopic glomeruli in adult antennae lobes [36]. The transmembrane receptor Notch and the epidermal growth factor receptor EGFR also seem to play an important role in the organisation, remodelling and function of the olfactory system [Additional file 5: Supplemental Table S4]. This confirms previous observations which showed that they were respectively required for selecting the sensory organ precursor lineages [50,51] and for the development of some of the neurons and cuticular structures of the antenna-maxillary sensory complex [52].

**Discussion**

Pros may regulate genes essential for neurite outgrowth and remodelling

In the AMC, the transcription factor Prospero is expressed in a cluster of cells (composed of neuronal and support cells, but not glial cells) that emerge during embryonic life and are maintained till the end of the larval stages. In embryos, Pros was reported to be involved in cell fate decision and in cell-cycle control. By contrast, our earlier data from the larval AMC rather suggested that pros could assume more restricted functions, such as the control of neuron-specific functions [8]. The present study confirms this hypothesis and shows that in the chemosensory organs dedicated to larval olfactory and gustatory sensing, *prospero* could regulate genes involved in neurite outgrowth and synaptic transmission.
Since pros was clearly shown to control axonal and dendritic outgrowth [53], we cannot exclude the possibility that the connection of pros with several genes that drive synaptic activity could be the indirect consequence of its involvement in neurite outgrowth control. In this respect, it is interesting to mention that a recent study [54] showed that axon targeting of the R7 Drosophila photoreceptor cells to their synaptic partner requires R7-specific transcription factor Prospero. These authors proposed that Pros could promote cell-type-specific expression of sensory receptors and cell-surface proteins regulating synaptic target specificity.

As previously mentioned, some of the genes identified in this functional class are also involved in neural connectivity remodelling. How can this be achieved if the AMC is completely histolysed? In fact, in Drosophila, not all sensory neurons degenerate; Some larval neurons persist and remodel to take on a new role in the adult system [55,56]. During the metamorphosis larval arbors of these neurons are pruned back and new adult-specific arbors are generated through a subsequent period of outgrowth. It seems that the neurites of these persistent larval neurons are used to partly guide axons of adult sensory neurons towards and within the CNS [55]. Therefore, histolysis and remodelling are two processes that are achieved during metamorphosis and could concern distinct neurons.

Does Pros play any role in AMC neuronal remodelling? We cannot actually answer the question. However, it has been previously reported that the insulin and epidermal growth factor signalling pathways, as well as ubiquitin-specific proteases are all required for the regulation of Drosophila neuronal remodelling [57]. Interestingly, all of these components emerge clearly from our analysis.

Actually, no work was done on the Drosophila larvae anterior sense organ in order to check whether some of the sensory neurons (which have also an embryonic origin) persist and remodel to take place in the adult olfactory or gustatory system. Therefore, the question is left open. At least the answers will provide important insights into the mechanisms that govern developmental plasticity in insect nervous systems.

In summary, our data collected from larval AMC and the previous genome wide expression profiling done on embryos [25] confirms that pros is associated with the regulation of neuronal specific genes. In this respect, it is essential to note that except for a few genes (126), most of the Pros target genes identified (~1000) in Choksi et al. [25] were not represented on our microarrays. For this reason, and because our experiments were performed on isolated individual larval tissues, it is not possible to determine whether the genes identified by these authors are specifically expressed in embryos and/or in tissues other than AMC.

Prospero and the insulin pathway
In Drosophila, the insulin/TOR signalling pathway [58] is divided into two branches. The insulin and its downstream effectors P13 and FOXO (forkhead box) represent one branch [59] of this pathway, while the other branch acts through the TOR family of Serine-Threonin kinases [60,61]. It has been shown that the insulin/TOR signalling pathway inhibits autophagy (For review see [58]) and controls growth by regulating ribogenesis and protein biosynthetic capacity [62,63]. Cumbini et al. [63] demonstrated that the TOR pathway is a nutritional checkpoint that participates in the systemic control of larval growth emanating from the Fat body.

Our microarray analysis has revealed a group of highly correlated pros candidate genes (correlation index: 0.9) that are either controlled by the insulin/TOR signalling pathway or are directly involved in the signalling cascade. This is the case for Ash2 [64] which was found to be regulated by TOR signalling. Similarly, FK506-bp1 affects autophagy through the modulation of FOXO [44] and Lk6 was reported to be a direct FOXO Target [62]. Therefore it seems that in the larval AMC, Pros could be associated with growth, autophagy and nutrient sensing through the regulation of genes that are directly or indirectly linked to the insulin/TOR pathway. Interestingly, TOR was found to be differentially expressed in the V1 pros mutant in the CNS [Additional file 5: Supplementary Table S1].

Conclusion
As previously described, loss of pros function in the AMC induced several alterations including axon path-finding defects and abnormal growth and taste responses. This is consistent with our microarray results showing that in the larval AMC, Pros expression is associated with the regulation of genes involved in the control of neurite outgrowth, mediation of growth and autophagy and in the organisation and function of the olfactory system. The mechanism by which all of these functions are achieved by pros in the AMC is presently not known but EGFR and/or Notch pathways could play a central role. Several lines of evidence are in favour of this hypothesis.

1- Four ligands are known to bind EGFR receptor: Keren, Gurken, Spitz, and vein [65]. Two of these were identified as potential targets of Prospero: Keren in both larval AMC and CNS and Gurken (Grk) in the larval CNS only (see Table 3 and [Additional file 5: Supplemental Table S1]). Moreover, Notch and EGFR were identified as putative Pros target genes in both embryos
and larval AMC, indicating that they could play a central role.

2- It has been reported that EGFR signalling is required for the development of some of the neurons and cuticular structures present in the AMC [66,67]. In this respect, it is interesting to point out that EGFR involvement has been reported during the development of mouse gustatory epithelia in the palate and tongue [68].

3- The expression of Notch, EGFR and Pros have been shown to be tightly linked. It has been demonstrated that normal levels of Pros expression in photoreceptor R7 cells in the Drosophila eye require EGFR signalling as well as Notch activation [69,70]. In addition, a recent analysis has shown that in R7 cells, Notch and EGFR cooperate in a complex way to promote pros transcription [71].

Although these data suggest that Notch and EGFR could play a central role in the mechanism by which Prospero carries out its function in the larval AMC, this hypothesis has still to be validated. In the future, it will be of great interest to explore in detail the mechanism by which all of these functions are accomplished by the homeodomain transcription factor Prospero.

Methods

Drosophila strains

All strains were maintained on standard cornmeal and yeast medium at 25°C. The prosV strains used in this study have already been described [72]. Briefly, prosV13 (V13) and prosV24 (V24) derived from the same prosV1 (V1) allele which contains the full length PGal4 transposon inserted upstream (~216 bp) of the pros coding region. The resulting behavioural and developmental anomalies observed in these mutants have been previously reported [8], and are summarised in Table 1. Additional descriptions of the expression pattern of these different alleles in the larval CNS are also provided in the [Additional file 2].

Isolation of AMC and CNS tissue

Around 150 larvae were used to obtain the AMC and CNS samples. The anterior region of the Larva was dissected to isolate CNS and AMC. The AMC region is not a well-defined tissue but is rather constituted by a small group of cells located in front of the hooks. Therefore, to maintain AMC integrity we kept the cuticle around it as well as the hooks.

Immunohistochemistry experiments

Staining experiments were performed as previously described by Guenin et al. [8]. Briefly, isolated larval AMC from embryos at stage 10-17 were incubated with various primary antibodies: MR1A mouse anti-Prospero at 1:4 dilution, rat anti-Elav at 1/1000 (a neuronal marker; provided by A. Giangrande), and rabbit anti-phosphohistone H3 at 1/1000 (a marker for mitotic activity; SIGMA). The following secondary antibodies were used to visualize these primary antibodies: antimouse Cy3 at 1/100 (Sigma); anti-mouse Alexa 594 anti-rat Alexa 488 at 1/400 (Molecular probes, USA); anti-rabbit Alexa 488 at 1/400 (Molecular probes, USA). AMC and CNS were mounted on Vectashield (Vector Laboratories, CA) before inspection under a fluorescence microscope (Leica DMRB) or a confocal microscope (Leica 4SD).

RNA extraction and cDNA labelling

Total RNA from third instar larvae was extracted from isolated AMC and CNS, according to Guenin et al. [8]. Four independent extractions were performed for each sample condition. RNA integrity was checked on denaturing formaldehyde agarose gels. The presence of clear bands corresponding to the 28s and 18s RNA with a 2:1 ratio and the absence of a smear were used to assess the RNA quality. Total RNA (3.0 μg) were treated with RQ1 RNase-free DNase (Promega) and reverse transcribed in presence of 33[P] dATP (Amersham Pharmacia Biotech, Bucks, United Kingdom), Random Primers and the reverse transcriptase (Maloney Murine Leukaemia Virus, Invitrogen).

Microarray experiments

Nylon membrane microarrays provided by the TAGC platform (Marseille-Nice Genopole) were used. They contained 7500 amplification PCR products of unique full length cDNA clones from the Drosophila Gene collection version 1.0 (Berkeley Drosophila Genome project). To verify the quality of spotting on the microarrays and the amount of DNA accessible for each spot, a vector probe (labelled oligonucleotide common to all spotted PCR products) hybridization was performed. Hybridization of 33[P] labelled probes was conducted for 24 h at 68°C in 500 μl of hybridization buffer (5× SSC, 5× Denhardt’s, 0,5% SDS). After 3 washing of 1 h in 500 ml of washing buffer (0,1× SSC, 0,2% SDS) at 68°C, arrays were exposed overnight to phosphor imaging plates which were scanned using a BAS 5000 (Fuji, Raytest, Paris France).

Data processing and analysis

Signal intensities were quantified using ArrayGauge software (V1.3; Fuji, Paris, France). All images were carefully inspected to exclude spots with overestimated intensities due to neighbourhood effects. Artefacts were eliminated by visual inspection. Spots were excluded from the quantification, if they were contaminated by overflowing neighbouring spots or if artefacts are present on the membrane. Overflowing spots were also eliminated. One sample was discarded (CNS V14) because of bad vector signals. The variability due to experimental conditions was eliminated by using a local weighted scattered plot smoother analysis (LOWESS, [73]). The data were then
filtered and only values found to be twice the mean local background value were kept. Correlation coefficients between expression measurements of two identical alleles ranged from 0.74 to 0.93 for the same tissue. Data were then log transformed.

Genes belonging to the same biological function or cell type are known to exhibit correlated expression [74]. Thus we searched for sets of genes with correlated expression that were differentially expressed between mutants. We used a method to detect groups of correlated genes and a statistical method to detect differential expression among these groups. Different methods were able to detect correlated expressions. We chose hierarchical clustering which has the advantage of not fixing a priori the number of classes or the number of genes per class. The statistical method used to detect genes differentially expressed between the wild type and pros mutants was the Discriminating Score [75], which is very similar to the widely used SAM method [76]. As the expected number of genes was not known, we calculated an average DS in a sliding window on the cluster. This allowed us to detect peaks of the optimal size. These peaks corresponded to genes with correlated expression and differentially expressed between mutants.

For the clustering, we used the Cluster program with Pearson correlation distance and average linkage as the aggregation strategy. The results were displayed using TreeView [26]. The DS measures the difference in gene expression between 2 groups of samples. If M1 represents mean expression of a given gene in wild type samples, and M2 the mean expression of the same gene in pros mutant samples, and SD the standard deviation of this gene in all considered samples, DS = (M1-M2)/SD. As V13, V14 and V24 have a normal Pros expression pattern in the AMC, they were considered wild type. The DS between V1 and all of the other alleles for the AMC was calculated for each gene. As V13 and V24 exhibited a distinct pattern in the CNS, for each gene, we calculated a DS between the V1 and V14 allele for the CNS. The score for each gene was then smoothed by calculating the mean score in a sliding window of 100 genes.

Finally, to highlight very tightly correlated genes in the main peak (peak 1) for each tissue, we performed a second hierarchical clustering with the genes of this peak, separately in the CNS and AMC.

The complete dataset is available through the National Center for Biotechnology Information (NCBI), in the Gene Expression Omnibus database http://www.ncbi.nlm.nih.gov/geo/ under the GSE12178 accession number.

Functional Annotation
Functional annotations of gene clusters were performed using GoMiner software [27] and the Gene Ontology database [77]. GoMiner determines significant enrichments of GO terms in a cluster of genes. This is performed by comparing the frequencies of each GO term in the cluster and in the microarray using Fisher’s exact test.

Research of a putative common motif in the promoter of co-expressed genes
Promoting regions of co-expressed genes were collected from the Ensembl ftp site http://www.ensembl.org/index.html. The sequence located from -1700 to +300 bp according to the +1 transcription start site of each gene was extracted. Interspersed and simple repeats were masked. These sequences were searched for a common motif using the Gibbs sampling method [28] available at the RSA Tools website: http://rsat.ulb.ac.be/rsat/. This method allowed to determine degenerated motifs described by a position weight matrix (PWM, a probabilistic model of residue frequencies at each position), in a set of sequences by iterative sampling. The initialisation step of the algorithm selects a random subsequence in each sequence to be searched. The predictive step builds a PWM from all the subsequences except one. The sampling step selects a new subsequence from the excluded sequence using a weighting strategy based on the PWM scores. The predictive and the sampling steps are iterated a given number of times or until convergence. We performed many Gibbs sampling runs as this method is stochastic. The motif found was identified in most of the runs. We used a Logo representation [29] to show the information content of the PWM.

Q-PCR validation
In order to validate the Microarray results, seven genes were selected and their expression levels were quantified by Q-PCR for both prosV1 and prosV14. For each RNA extraction used for microarrays, a sample was collected in order to perform Q-PCR experiments. Reverse transcriptions were done from 2 μg of total RNA as described in Guenin et al. [8]. The selected genes and the corresponding primers, designed using Primer Express™ (Applied Biosystems) parameters, are indicated in Table 4. Q-PCR reactions were performed with 1:10 diluted cDNA in 2× SybrGreen PCR master mix (Applied Biosystems) with each specific primer (See table 4) or control primers (actin 5C F 5’GCCCATC-TACGAGGGTTATGC3’ and actin 5C R 5’CAAATCCGCGACCAGCCAG3’). Signals were measured with ABI Prism 7000™ Sequence Detection System software (Applied Biosystems). All signal thresholds to be compared were standardized with the actin 5C mRNA [8].

Additional file 1: Mitotic Activity in the AMC region, in stage 11 and stage 16 embryos. Wild type embryos from stage 11 to 16 were stained with Pros (red) and H3p (green) which label cells in division. (A)
Mitotic activity is observed until stage 11-12 (arrow) while no more activity is detected from stage 16 embryos (B) in the AMC. Scale bar correspond to 20 μm. Click here for file

Additional file 2: Expression pattern of V24 in the third instar larval CNS. The Pros (A) and Elav (B) expression pattern as well as the mitotic activity (C) of V24 are shown as compared to the previously reported pattern of V14, V13 and V1 alleles (B). Although V24 and V13 present the same expression pattern in the AMC, the situation is different in the CNS. As it can be seen Pros (A) and Elav labeling (B) are distinctive for both alleles in the region delimiting the two hemispheres and the Optic lobes (OLs). As compared to V13, V24 presents a decrease of the staining in this region for these two markers (B). In ventral nerve cord (VNC), V13 shows an important hyperplasia (arrowheads) due to an excess of neurons. In V24, the VNC extremity presents a bifida aspect (arrow). The mitotic activity, revealed by anti-phihstone-H3 (H3p) (C) is strongly increased in V24, especially in the OLs. Click here for file

Additional file 3: List of the CNS genes represented in the peak obtained after the application of the discriminating score between V1 and all other alleles for the AMC samples. Peak 1: Differentially expressed genes involved in cell fate commitment. Peak 2: Differentially expressed genes involved in the proteasome complex. Peak 3: Differentially expressed genes involved in signal transducer activity. Click here for file

Additional file 4: List of the CNS genes represented in the peak obtained after the application of the discriminating score between V1 and V1 4 for the CNS samples. The peak contains differentially expressed genes involved in cell fate commitment. Click here for file

Additional file 5: Supplemental Table S1: Genes found to be differentially expressed between V1 and V14 CNS. We found 86 genes that are highly correlated (coefficient > 0.9). Among the 86 genes, 28 were also found to be overexpressed in the V1 AMC (column on the left, gene names are indicated in bold) and contain the common putative pros DNA motif in their promoter. The S8 genes present in the two columns on the right are specifically overexpressed in V1 CNS as compared to V14 CNS. Supplemental Table S2: Phenotypic data related to the candidate genes involved in neurite outgrowth and/or synaptic transmission. The criteria used for the description of the phenotypes were as follows: (1) if many larval phenotypes were available for a gene, we selected only those observed in the nervous system and preferentially in the peripheral nervous system (PNS). (2) If no larval phenotype was available for a gene, we selected those observed in the embryonic and/or adult PNS. (3) If available, the effect of the upregulation or downregulation of these genes is mentioned respectively for those that are overexpressed or underexpressed in V1 AMC. (4) All studies mentioned were done in Drosophila melanogaster. Supplemental Table S3: Phenotype data related to the candidate genes involved in growth and autophagy. The criteria used for the description of the phenotypes were the same as those for Table S2. Supplemental Table S4: Phenotype data related to the candidate genes involved in sensory organ development and most particularly in olfaction (in bold). The criteria used for the description of the phenotypes were the same as those for Table S2. Click here for file

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Authors’ contributions
LG and FB generated the microarray data and drafted the manuscript. LG carry out immunohistochemistry and Q-PCR experiments. LG, MR and RH performed the biostatistic analysis; MR performed the motif discovery. FB generated the Drosophila larvae data compilation for manual annotation. FB and RH provided direction and oversight of the experiments. FB holds the grant. All authors read, corrected and approved the final manuscript.

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