A deep burn wound is very difficult to treat and usually requires reconstruction using vascularized tissues. Regenerative medicine has begun to adopt new treatments for such nonhealing ulcers, and platelet-rich plasma (PRP) is one of the easiest-to-use methods as it can be harvested from the patient’s own blood and thus completely avoids immune rejection and allergic reaction. We previously reported a preparation protocol for platelet-derived factor concentrate (PFC), which achieved platelet and platelet-derived growth factor (PDGF)-BB concentrations 20 and 200 times higher than those found in whole blood, respectively. Herein, we report the clinical outcomes in 2 patients with critical burn wounds with bone and tendon exposure, who were successfully treated by injection of PFC in combination with hyaluronic acid (HA) scaffolds.

METHODS AND RESULTS

A 52-year-old man received a severe left facial wound after a heat-press injury. He promptly received debridement of the wound (Fig. 1A), and 6 days after, skin grafting into the lateral region, which had a good wound bed, and free perifascial areolar tissue graft into the medial region, where bone was exposed, were performed. Although skin graft survival was confirmed 1 week postoperatively, the perifascial areolar tissue graft was not accepted, and bone (partly becoming sequestrum) became exposed again. Therefore, after approval by the institutional review board, we initiated treatment using PFC, which was prepared according to the previous protocol, and also administered HA scaffolds. Briefly, whole blood was drawn from the patient and centrifuged at 270g for 10 minutes. The isolated PRP layer was centrifuged again at 2300g for 10 minutes to spin down platelets. The platelet-poor plasma was partially removed and replaced with a one-tenth volume of phosphate-buffered saline for noncoagulating PFC. Platelet activators such as thrombin were not added, because they will be released from the surrounding tissue. After the platelets were resuspended, 0.6 mL of PFC and 0.4 mL of non–cross-linked HA (Artz, Seikagaku Corporation; Tokyo, Japan) were mixed and injected into the surface of the ulcer of the medial region through a 30-gauge needle once a week (see video, Supplemental Digital Content 1, which displays the injection technique, http://links.lww.com/PRSGO/A281). After 3 sessions of treatment, healthy granulation tissue and periulcer epithelization were noted (Fig. 1B). The remaining ulcer epithelialized within several weeks, and no recurrence was observed as of 1-year follow-up.

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A 61-year-old woman sustained a burn caused by heated oil to her left hand. She was diagnosed with a superficial second-degree burn of the fourth finger and third-degree burn of the fifth finger. She received debridement for the fifth finger on day 3 after injury, resulting in tendon exposure (Fig. 2A), and we began treatment using PFC–HA. After preparation in the same manner as described above, the weekly injection of 0.3 mL of PFC combined with 0.2 mL of non–cross-linked HA was performed into the wound surface of the fifth finger. After 2 sessions of treatment, the wound had almost completely closed, whereas the fourth finger healed with standard conservative treatment but left a hypertrophic scar (Fig. 2B).

**DISCUSSION**

PDGFs are released during the clotting cascade and acts as an initiator of wound healing. PRP contains various bioactive substances including PDGF, transforming growth factor–beta, and epidermal growth factor; and its beneficial influence on tissue remodeling is attributed to mesenchymal cell recruitment and proliferation resulting in vascularization, extracellular matrix synthesis, and granulation. PRP not only aids in the healing of a greater range of wounds but also shortens the time to healing; however, the therapeutic use of PRP is still controversial because of the lack of standardized protocols and indications.

Standard treatment for deep burn wounds with exposed bone or tendons is vascularized tissue coverage; however, we could successfully treat 2 patients by the injection of PFC with HA scaffolds. Our protocol required 2 or 3 sessions of treatment, as the advantageous effects from PDGFs are most likely to occur when they are repeatedly applied to the wound bed. PFC prepared by our protocol has a higher PDGF-BB concentration (157.9 ng/mL on average) than the PRP concentration in previous reports, and it is also highly aqueous because of the lack of fibrinogen, which is discarded together with platelet-poor plasma. Although fibrin gel may be useful as a carrier of growth factors, it reduces the final concentration, hinders the easy injection of PRP, frequently leads to uncontrollable coagulation, and is not preferable in many clinical situations. In this study, we alternatively used non–cross-linked HA as a scaffold and controlled-release carrier of PFC. We previously demonstrated that HA could be a biological cell scaffold where mesenchymal stem cells could adhere, survive, and proliferate. In addition, HA is as-
sumed to keep PFC around the ulcer and continuously releasing PDGF; as PFC is in a liquid form, its effects must be released slowly. Although further evaluation is necessary, PFC–HA treatment is easy to apply, safe, and an effective choice to heal deep burn wounds that are intractable to conventional conservative treatment, and it may be promising for the treatment of refractory ulcers.

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