Enhanced susceptibility to predation in corals of compromised condition

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The marine gastropod, *Coralliophila abbreviata*, is an obligate corallivore that causes substantial mortality in Caribbean *Acropora* spp. Considering the imperiled status of *Acropora cervicornis* and *A. palmata*, a better understanding of ecological interactions resulting in tissue loss may enable more effective conservation strategies. We examined differences in susceptibility of *A. cervicornis* to *C. abbreviata* predation based on coral tissue condition. Coral tissue condition was a strong determinant of snail prey choice, with snails preferring *A. cervicornis* fragments that were diseased or mechanically damaged over healthy fragments. In addition, snails always chose fragments undergoing active predation by another snail, while showing no preference for a non-feeding snail when compared with an undisturbed prey fragment. These results indicate that the condition of *A. cervicornis* prey influenced foraging behavior of *C. abbreviata*, creating a potential feedback that may exacerbate damage from predation in coral populations compromised by other types of disturbance.
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Abstract

The marine gastropod, *Coralliophila abbreviata*, is an obligate corallivore that causes substantial mortality in Caribbean *Acropora* spp. Considering the imperiled status of *Acropora cervicornis* and *A. palmata*, a better understanding of ecological interactions resulting in tissue loss may enable more effective conservation strategies. We examined differences in susceptibility of *A. cervicornis* to *C. abbreviata* predation based on coral tissue condition. Coral tissue condition was a strong determinant of snail prey choice, with snails preferring *A. cervicornis* fragments that were diseased or mechanically damaged over healthy fragments. In addition, snails always chose fragments undergoing active predation by another snail, while showing no preference for a non-feeding snail when compared with an undisturbed prey fragment. These results indicate that the condition of *A. cervicornis* prey influenced foraging behavior of *C. abbreviata*, creating a potential feedback that may exacerbate damage from predation in coral populations compromised by other types of disturbance.

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Introduction

Corallivory is widely understood to have significant, if sometimes underestimated, effects on scleractinian coral populations (Rotjan & Lewis, 2008). Depending on the type and intensity of predation, corallivory may result in positive or negative ecological responses (Cole, Pratchett & Jones, 2008). While, at the community level, moderate predation pressure has sometimes been positively correlated with an increase in species diversity (Menge & Sutherland, 1976), for an individual coral, predation often equates to partial mortality that may compromise physiological processes such as growth (Meesters, Noordeloos & Bak, 1994) and reproductive success (Van Veghel & Bak, 1994). Predation can also be associated with negative indirect effects; if tissue regeneration is incomplete or too slow following predation, spatial competitors such as algae and sponges can colonize the dead areas, potentially introducing a myriad of negative interactions (Bak & Steward-Van Es, 1980). Additionally, some corallivores are known to vector disease (Sussman et al., 2003; Williams & Miller, 2005), and tissue lesions (likely including those from partial predation) have been cited as a pre-requisite to disease transmission in *Acropora cervicornis* (Gignoux-Wolfson, Marks & Vollmer, 2012).

While predation among healthy coral populations can promote a stable equilibrium, within degraded coral populations, the per capita relative impact of corallivory is likely to increase as coral cover decreases, potentially affecting the fitness and recovery of affected coral populations (Jayewardene, Donahue & Birkeland, 2009). This was observed on surveyed reefs in Jamaica where populations of *A. cervicornis* were reduced by the acute disturbance of Hurricane Allen. As a result, predation by the corallivorous gastropod, *Coralliophila abbreviata*, was concentrated...
on remaining individuals resulting in further population declines of *A. cervicornis* rather than recovery (Knowlton, Lang & Keller, 1990).

Abiotic and biotic disturbances such as hurricanes, disease and bleaching not only reduce coral populations, but may affect the ‘condition’ of the remaining colonies yielding broken branches, lesions, compromised immune response (Bak & Criens, 1981), growth (Bak, 1983) and reproduction (Rinkevich & Loya, 1979). Furthermore, studies have shown that stressed corals are more susceptible to ambient predation (Morton, Blackmore & Kwok, 2002; Wolf & Nugues, 2013) suggesting that predation pressure on a focal coral may increase as a result of disturbance-related physiological stress in addition to a potential numerical effect (i.e. higher ratio of predators to surviving prey). Field studies on the impact of predation following disturbances such as Knowlton, Lang & Keller (1990) cannot distinguish the relative influence of diminished coral abundance versus coral condition.

The *Acropora-Coralliophila* relationship provides a good model for evaluating the relative influence of coral condition (i.e. healthy, diseased, damaged, etc.) on predator behavior. Considering the region-wide declines of *Acropora* spp., many populations may exist below a threshold abundance where they are now at heightened vulnerability to predation pressure. *Coralliophila abbreviata* is an obligate corallivore with an asymmetrical preference for *Acropora* spp. prey (Johnston & Miller, 2014) and may thus pose a substantive risk to the recovery of decimated *Acropora* populations. In the Florida Keys, predation by *C. abbreviata* was the most prevalent factor causing tissue loss among remnant populations of *A. palmata*, accounting for approximately 29% of all tissue mortality (Williams & Miller, 2012), and is
recognized as one of the top three proximal threats to the recovery of wild (Bruckner, 2002) and
restored Acropora populations (Johnson et al., 2011) throughout the Caribbean. Currently,
management actions are underway throughout the Caribbean to enhance Acropora population
recovery via conservation and restocking strategies. Understanding the interactions of factors
contributing to the loss of Caribbean acroporids may allow for more effective conservation
strategies for these species. We took advantage of the experimental system offered by in situ
nursery propagation of A. cervicornis to conduct field choice assays testing the hypothesis that
susceptibility to colonization by the corallivore, C. abbreviata, would be enhanced for corals
with disease or mechanical damage relative to an apparently healthy coral.

Materials and Methods
This study was conducted at a coral field nursery operated by the Coral Restoration Foundation
(CRF) located off Tavernier, Florida (24° 59’N, 80° 26’W) from June to September 2013 (under
permit # FKNMS-2013-065). The nursery is located on sand bottom surrounded by seagrass at a
depth of 10 m and provided Acropora cervicornis fragments for use in choice experiments.
Coralliophila abbreviata were collected from Pickles Reef (24° 59’N, 80° 24’W), transported
2.2 km to the nursery and kept in holding cages until use in choice experiments. Snails were held
separately based on the host species from which they were collected: A. cervicornis (n = 38) or
A. palmata (n = 55). Snails were starved for at least one week prior to use in choice trials.

Treatment chambers were made by modifying plastic compartment boxes (40 x 23 x 8 cm). Each
chamber contained two choice arenas with a removable lid, which was secured with cable ties
during trial periods (Fig. 1A). A single choice arena consisted of a Y-maze with the subject snail
staged at one end facing two treatment lanes and a treatment coral loosely secured at the end of each lane. Rectangular cutouts covered with window-screen mesh were at each end of the treatment chamber to facilitate flow. Chambers were oriented relative to predominant currents to allow water flow from the treatment coral toward the snail staging area. To minimize water exchange between treatment lanes and arenas, Velcro felt was glued to the top of the walls of each treatment lane allowing the lid to lay flush with no gaps. Each of ten experimental chambers was secured to a cinder block and set on sand bottom approximately 1 m apart (Fig. 1B).

Three types of prey choice experiments were conducted (Fig. 1C-E): healthy coral versus diseased coral (H v D, \( n = 52 \)), healthy coral versus coral with mechanical damage (H v M, \( n = 58 \)) and healthy coral versus coral with active snail predation (H v P; \( n = 3 \)). Prior to the start of the experiment, healthy fragments were snipped from apparently healthy nursery colonies and the cut surfaces were allowed to heal for at least two weeks in the field in order to yield completely undisturbed, healthy tissue. Disease progression is intermittent in *A. cervicornis* requiring actively diseased samples to be identified immediately at the start of a trial. Thus, diseased branch tips were collected from cultured coral with active tissue loss and snipped immediately prior to trials within an area of already-dead skeleton (i.e. no tissue disturbance), approximately 2 cm from the tissue margin. Fragments with mechanical damage were prepared by snipping branch tips from healthy coral, and, immediately prior to the start of a trial, a 3–5 cm long abrasion was created with a clean, dead *A. cervicornis* branch. This treatment was included to mimic the type of damage inflicted from abrasion during storms or hurricanes. At the end of
any trial, if a snail was actively feeding on a healthy coral, the coral and feeding snail were immediately transferred to serve as a choice in a subsequent H v P trial (Fig. 1E).

_Coralliophila abbreviata_ are known to be gregarious (Bruckner, Bruckner & Williams, 1997); so there is an aspect of prey choice that may be purely social. Thus, we also tested snail preference between a healthy coral fragment versus a solitary snail with no coral (H v S, _n_ = 45). In these trials, a treatment snail was tethered via a small length of twine glued to its shell at the treatment end of one of the Y-maze lanes with a healthy coral at the end of the other lane (Fig. S1).

The position of the healthy coral and treatment coral/snail was alternated between the two lanes in subsequent trials. Following each trial, the chamber was flushed of all sand and debris, and the walls and floors were rigorously wiped down with a brush or a diver’s gloved-hand to reduce carryover of mucus or other potential cues. Trials were pooled among those conducted during daytime (8:00 am to approximately 4:00 pm; ~8 h duration) and nighttime (sunset to a few hours after sunrise; ~16 h duration).

During each trial, subject snails were left alone to choose a treatment lane. If the subject snail did not travel more than 5 cm down a treatment lane or remained in the staging area, the trial was determined as ‘no choice’ and excluded from analyses (_n_ represents only the trials where a choice was made; Table 1; see Table S1 for daytime and nighttime trials separated). The proportion of trials in which a choice was made (i.e. response rate) ranged from 41 to 72% across the three treatments (D, M and S), which is well within the range of response rates reported in
other published Y-maze choice studies using gastropod subjects (range: 27 to 100%; Nakashima, 1995; Avila, 1998; Rilov, Gasith & Benayahu, 2002).

For each treatment except H v S, trials among the two host-source subject snails are pooled for analysis as no difference was found between snails sourced from *A. palmata* and *A. cervicornis*. Additionally, trials are pooled among daytime and nighttime for each treatment as no difference was found in preferences expressed during daytime versus nighttime. Differences in frequencies of choices made between healthy and treatment corals/snail were assessed using a Pearson’s chi-squared test. The variation in activity levels (i.e. proportion of trials in which a choice was made) between daytime versus nighttime trials was analyzed using 2 × 2 contingency tables (Statistica Statistical Software v6.0).

**Results and Discussion**

Coral condition significantly affected prey preference of *Coralliophila abbreviata* snails sourced from *Acropora* spp. host colonies. In 70.8% of the trials, snails preferred corals with either disease \( (df = 1, p < 0.001, \text{Fig. 2A}) \) or mechanical damage \( (df = 1, p = 0.01, \text{Fig. 2A}) \) over apparently healthy corals. Other studies have shown similar results for the Pacific corallivorous snail, *Drupella rugosa*, where snails were attracted to corals stressed by either mechanical damage, low salinity or low water temperature suggesting that corals stressed by additional factors beyond the scope of the present study may manifest a similar enhanced susceptibility to corallivores (Morton, Blackmore & Kwok, 2002; Tsang & Ang, 2015).
It is well known that chemoreception is important in foraging behavior of marine benthic organisms (Kohn, 1961; Hay, 2009). Although the specific mode of attractant to prey has not been studied for *C. abbreviata*, it seems likely that variable chemical cues may underlie their preferences. Abraded coral tissue releases mucus and interstitial content that contains primary metabolites such as proteins and amino acids which may attract consumers (Hay, 2009). The release of mucus and/or secretions by damaged cells were the suggested cause of attractants for increases of the corallivorous snail, *D. rugosa*, to stressed corals in Hong Kong (Morton, Blackmore & Kwok, 2002). Similarly, Kita et al. (2005) showed an increase in ‘feeding-attractant activity’ by the corallivorous snail, *Drupella cornus*, when offered montiporic acids isolated from the prey coral, *Montipora* sp., which are suggested to be expelled with coral mucus. As this study only examined behavioral responses based on short distance cues, further studies should determine a range of distances that snails are able to detect such cues to better infer snail foraging patterns on a reef scale.

A similar mechanism that attracts snails to mechanically damaged corals may apply to diseased coral tissue as it may result in tissue deterioration and the production of excess mucus. However, although mechanically damaged corals used in this study appeared to produce more mucus and expel more interstitial content than diseased corals, snails had a slightly stronger attraction to diseased coral (75% of choices made) than mechanically damaged coral (65.5% of choices made) suggesting there may be something more complex attracting the snails than simple quantity of these exudates. Additionally, as this study did not induce disease for H v D treatment corals, it is possible that an undescribed physiological difference in corals making them more
susceptible to disease may also make them more attractive to snails rather than the diseased condition *per se*.

Though there was only opportunity to conduct limited trials of the H v P treatment (*n* = 3), 100% elicited a choice and 100% chose the fragment under active predation (P) suggesting a strong bias toward prey fragments with active conspecific snail feeding. To ensure that the subject snail was not attracted to the mere presence of a conspecific, H v S trials were conducted and showed no significant preference for the conspecific snail relative to a healthy coral (Fig. 2B). However, the following of conspecific mucus trails has been reported for the marine mud snail, *Ilyanassa obsolete* (Trott & Dimock, 1978) and has been suggested to account for aggregation behavior in the corallivorous snail, *Cyphoma gibbosum* (Gerhart, 1986). Yet, considering that traces of previous mucus trails were removed (see methods), this preference may be the result of damaged coral tissue releasing interstitial attractants (described above) or feeding mucus produced by the feeding snail.

Intraspecific behavioral differences in *C. abbreviata* sourced from *A. palmata* and *A. cervicornis* colonies were evident in H v S trials (Fig. 2B) and in activity level based on time of day (Table S1). Snails sourced from *A. cervicornis* showed a strong preference for healthy coral over a solitary snail with no coral, while snails sourced from *A. palmata* showed no preference for either treatment. Additionally, snails sourced from *A. palmata* were significantly more active at night than during the day (evidenced by proportion of trials in which a choice was made), whereas, snails sourced from *A. cervicornis* showed no difference in activity level between daytime and nighttime trials. Intraspecific behavioral differences have been shown for a number
of marine species (e.g., Stachowicz & Hay, 2000; Crosby & Reese, 2005; Jordaens, Dillen &
Backeljau, 2009). In *C. abbreviata*, behavioral and population structural differences (e.g.
size/age structure, sex ratio, etc.) have been previously documented between acroporid and non-
acroporid host corals (Hayes, 1990; Baums, Miller & Szmant, 2003a; Johnston & Miller, 2007),
despite genetic results showing it to be a single species throughout the Caribbean (Johnston,
Miller & Baums, 2012). To our knowledge, no previous studies have documented intraspecific
behavioral differences of *C. abbreviata* between two *Acropora* spp. host corals as evident in our
results. One explanation may be that *A. cervicornis*-sourced snails were presented with their
native host prey species, whereas *A. palmata*-sourced snails were presented with a non-host prey
species. Some sort of host conditioning may result in differing attractiveness of native versus a
congeneric prey alternative. However, such differences were unexpected since the tissue of *A.
cervicornis* and *A. palmata* are qualitatively similar (thin tissues on perforate skeleton) and have
similar nutritional quality as indicated by C:N ratios (*Ac* = 6.1 ± 0.9, Szmant, Ferrer &
FitzGerald, 1990; *Ap* = 6.3 ± 0.3, Baums, Miller & Szmant, 2003b).

Coral disease and physical damage occur regularly on coral reefs, and, as changing climate is
predicted to bring increases in intensity and/or frequency of strong storms (McWilliams et al.,
2005; Knutson et al., 2010), as well as disease (Harvell et al., 2002), it is crucial to understand
how corals may be directly and indirectly affected by these disturbances. This study highlights
the enhanced vulnerability of remnant coral populations following acute disturbance events such
as storms or disease outbreaks due to corallivore behavioral preferences. The indirect effect of
attracting snail predators to these impacted corals implies ongoing tissue loss from predation
inhibiting potential recovery. Furthermore, there is likely a complex feedback between disease
risk and snail predation as *C. abbreviata* has been shown to vector disease among *A. cervicornis* colonies (Williams & Miller, 2005). Although corallivory by invertebrates is relatively well-documented, corallivore behavior and its potential influence on recovery of threatened or endangered coral populations has received little attention. Understanding such behavioral complexities can aid in epidemiological and predictive modelling of disease dynamics and transmission in *Acropora* spp. populations as well as improved species recovery strategies such as targeting snail removal efforts (Williams et al., 2014) following specific types of disturbance.
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References

Avila C. 1998. Chemotaxis in the nudibranch *Hermissenda crassicornis*: does ingestive condition influence its behaviour in a Y-maze. *Journal of Molluscan Studies* 64:215-222. DOI 10.1093/mollus/64.2.215.

Bak RPM, Steward-Van Es Y. 1980. Regeneration of superficial damage in the scleractinian corals *Agaricia agaricites F. Purpurea* and *Porites astreoides*. *Bulletin of Marine Science* 30:883-887.

Bak R, Criens S. 1981. Survival after fragmentation of colonies of *Madracis mirabilis, Acropora palmata* and *A. cervicornis* (scleractinia) and the subsequent impact of a coral disease. *Proceedings of the 4th International Coral Reef Symposium* 2:221-228.

Bak R. 1983. Neoplasia, regeneration and growth in the reef-building coral *Acropora palmata*. *Marine Biology* 77:221-221.

Baums IB, Miller MW, Szmant AM. 2003a. Ecology of a corallivorous gastropod, *Coralliophila abbreviata*, on two scleractinian hosts. I: Population structure of snails and corals. *Marine Biology* 142:1083-1091. DOI 10.1007/S00227-003-1024-9.

Baums IB, Miller MW, Szmant AM. 2003b. Ecology of a corallivorous gastropod, *Coralliophila abbreviata*, on two scleractinian hosts. II. Feeding, respiration and growth. *Marine Biology* 142:1093-1101. DOI 10.1007/s00227-003-1053-4.

Bruckner AW. 2002. Proceedings of the Caribbean *Acropora* Workshop: potential application of the U.S. Endangered Species Act as a conservation strategy. NOAA Technical Memorandum NMFS-OPR-24, Silver Spring, MD, p 199.

Bruckner RJ, Bruckner AW, Williams EHJ. 1997. Life history strategies of *Coralliophila abbreviata* Lamarck (Gastropoda: Coralliophilidae) on the southeast coast of Puerto Rico. *Proceedings of the 8th International Coral Reef Symposium* 1:627-632.

Cole AJ, Pratchett MS, Jones GP. 2008. Diversity and functional importance of coral-feeding fishes on tropical coral reefs. *Fish and Fisheries* 9:286-307. DOI 10.1111/j.1467-2979.2008.00290.x.

Gerhart DJ. 1986. Gregariousness in the gorgonian-eating gastropod *Cyphoma gibbosum*: tests of several possible causes. *Marine Ecology Progress Series* 31:255-263.

Gignoux-Wolfsohn S, Marks CJ, Vollmer SV. 2012. White Band Disease transmission in the threatened coral, *Acropora cervicornis*. *Scientific Reports* 2:804. DOI 10.1038/srep00804.

Harvell CD, Mitchell CE, Ward JR, Altizer A, Dobson AP, Ostfeld RS, Samuel MD. 2002. Climate warming and disease risks for terrestrial and marine biota. *Science* 296:2158-2162. DOI 10.1126/science.1063699.

Hay ME. 2009. Marine chemical ecology: Chemical signals and cues structure marine populations, communities, and ecosystems. *Annual Review of Marine Science* 1:193-212. DOI 10.1146/annurev.marine.010908.163708.

Hayes JA. 1990. Distribution, movement and impact of the corallivorous gastropod *Coralliophila abbreviata* (Lamarck) on a Panamanian patch reef. *Journal of Experimental Marine Biology and Ecology* 142:25-42.
Jayewardene D, Donahue MJ, Birkeland C. 2009. Effects of frequent fish predation on corals in Hawai'i. *Coral Reefs* 28:499-506. DOI 10.1007/s00338-009-0475-y.

Johnson ME, Lustic C, Bartels E, Baums IB, Gilliam DS, Larson L, Lirman D, Miller MW, Nedimyer K, Schopmeyer S. 2011. Caribbean *Acropora* restoration guide: best practices for propagation and population enhancement. The Nature Conservancy. Arlington, VA.

Johnston L, Miller MW. 2007. Variation in life-history traits of the corallivorous gastropod *Coralliophila abbreviata* on three coral hosts. *Marine Biology* 150: 1215-1225. DOI 10.1007/s00227-006-0422-1.

Johnston L, Miller MW, Baums IB. 2012. Assessment of host-associated genetic differentiation among phenotypically divergent populations of a coral-eating gastropod across the Caribbean. *PLoS ONE* 7:e47630. DOI 10.1371/journal.pone.0047630.

Johnston L, Miller MW. 2014. Negative indirect effects of neighbors on imperiled scleractinian corals. *Coral Reefs* 33:1047-1056. DOI 10.1007/s00338-014-1176-8.

Jordaens K, Dillen L, Backeljau T. 2009. Shell shape and mating behavior in pulmonate gastropods (Mollusca). *Biological Journal of the Linnean Society* 96:306-321. DOI 10.1111/j.1095-8312.2008.01122.x.

Kita M, Kitamura M, Koyama T, Teruya T, Matsumoto H, Nakano Y, Uemura D. 2005. Feeding attractants for the muricid gastropod *Drupella cornus*, a coral predator. *Tetrahedron Letters* 46:8583-8585. DOI 10.1016/j.tetlet.2005.09.182.

Knowlton N, Lang JC, Keller B. 1990. Case study of natural population collapse: post-hurricane predation on Jamaican staghorn corals. *Smithsonian Contributions to the Marine Sciences* 31:1-25.

Knutson TR, McBride JL, Chan J, Emanuel K, Holland G, Landsea C, Held I, Kossin JP, Srivastava A, Sugi M. 2010. Tropical cyclones and climate change. *Nature Geoscience* 3:157-163. DOI 10.1038/ngeo779.

Kohn AJ. 1961. Chemoreception in gastropod molluscs. *American Zoologist* 1:291-308. Available at http://www.jstor.org/stable/3881260.

McWilliams JP, Cote IM, Gill JA, Sutherland WJ, Watkinson AR. 2005. Accelerating impacts of temperature-induced coral bleaching in the Caribbean. *Ecology* 86:2055-2060. Available at http://dx.doi.org/10.1890/04-1657.

Meesters EH, Noordeloos M, Bak RPM. 1994. Damage and regeneration: links to growth in the reef-building coral *Montastrea annularis*. *Marine Ecology Progress Series* 112:119-128.

Menge BA, Sutherland JP. 1976. Species diversity gradients: synthesis of the roles of predation, competition, and temporal heterogeneity. *The American Naturalist* 110:351-369. Available at http://www.jstor.org/stable/2459759.

Morton B, Blackmore G, Kwok CT. 2002. Corallivory and prey choice by *Drupella rugosa* (Gastropoda: Muricidae) in Hong Kong. *Journal of Molluscan Studies* 68:217-223. DOI 10.1093/mollus/68.3.217.

Nakashima Y. 1995. Mucous trail following in 2 intertidal nudibranchs. *Journal of Ethology* 13:125-128. DOI 10.1007/BF02352571.

Rilov G, Gasith A, Benayahu Y. 2002. Effect of an exotic prey on the feeding pattern of a predatory snail. *Marine Environmental Research* 54:85-98. DOI 10.1016/S0141-1136(02)00096-X.

Rinkevich B, Loya Y. 1979. The reproduction of the Red Sea coral *Stylophora pistillata*. I. gonads and planulae. *Marine Ecology Progress Series* 1:133-144.
Rotjan RD, Lewis SM. 2008. Impact of coral predators on tropical reefs. Marine Ecology Progress Series 367:73-91.

Stachowicz JJ, Hay ME. 2000. Geographic variation in camouflage specialization by a decorator crab. The American Naturalist 156:59-71. DOI: 10.1086/303366.

Sussman M, Loya Y, Fine M, Rosenberg E. 2003. The marine fireworm Hermodice carunculata is a winter reservoir and spring-summer vector for the coral-bleaching pathogen Vibrio shiloi. Environmental Microbiology 5:250-255. DOI 10.1046/j.1462-2920.2003.00424.x.

Szmant AM, Ferrer LM, FitzGerald LM. 1990. Nitrogen excretion and O:N ratios in reef corals: evidence for conservation of nitrogen. Marine Biology 104:119-127.

Trott TJ, Dimock RV. 1978. Intraspecific trail following by the mud snail Ilyanassa obsoleta. Marine Behaviour and Physiology 5:91-101. DOI 10.1080/10236247809378526.

Tsang RHL, Ang P Jr. 2015. Cold temperature stress and predation effects on corals: their possible roles in structuring a nonreefal coral community. Coral Reefs 34:97-108. DOI 10.1007/s00338-014-1210-x.

Van Veghel MLJ, Bak RPM. 1994. Reproductive characteristics of the polymorphic Caribbean reef building coral Montastrea annularis. III. Reproduction in damaged and regenerating colonies. Marine Ecology Progress Series 109:229-233.

Williams DE, Miller MW. 2005. Coral disease outbreak: pattern, prevalence and transmission in Acropora cervicornis. Marine Ecology Progress Series 301:119-128.

Williams DE, Miller MW. 2012. Attributing mortality among drivers of population decline in Acropora palmata in the Florida Keys (USA). Coral Reefs 31:369-382. DOI 10.1007/s00338-011-0847-y.

Williams DE, Miller MW, Bright AJ, Cameron CM. 2014. Removal of corallivorous snails as a proactive tool for the conservation of acroporid corals. PeerJ 2:e680. DOI 10.7717/peerj.680.

Wolf AT, Nugues MM. 2013. Synergistic effects of algal overgrowth and corallivory on Caribbean reef-building corals. Ecology 94:1667-1674. Available at http://dx.doi.org/10.1890/12-0680.1.
Figure 1 (on next page)

Photo examples of experimental Y-maze chamber design and treatment coral fragments.

(A) Experimental chamber design. Each chamber has two separate choice arenas, depicted as ‘1’ and ‘2.’ White circles depict the initial staging area for the subject snail. The treatment corals are attached at the far end of each treatment lane. (B) Cages were aligned in the same direction facing into the current. (C) Photo example of a treatment coral fragment with disease, (D) a fragment with mechanical damage and (E) a fragment with active snail predation.
Results of Y-maze trials.

Percent of successful choice assay trials comparing snail preference of (A) healthy (H; white bars) *A. cervicornis* fragments vs. fragments with compromised condition (black bars; diseased [D] or mechanically damaged [M]) and (B) H vs. a conspecific snail (S; grey bars) presented separately for subject snails sourced from *A. cervicornis* and *A. palmata* hosts. Asterisks indicate significant results (Pearson Chi-squared tests, p < 0.05). (The total number of successful trials is given at the base of each bar)
Table 1 (on next page)

Summary of Y-maze trial treatments.

Paired choice experiments testing prey preferences by *Coralliophila abbreviata*. *N* gives the number of successful trials (for the treatment paired with H) with subject snails from each of two host corals, *Acropora cervicornis* (Ac) or *A. palmata* (Ap). The ‘# of no choice trials’ represents additional trials conducted wherein the subject snail did not make a choice.
| Treatment                      | Origin                                                                 | $N$ | # of No Choice Trials |
|--------------------------------|------------------------------------------------------------------------|-----|------------------------|
|                                |                                                                        | Ac  | Ap         | Sum |                   |
| Healthy Coral (H)              | 5 cm branch tip snipped from nearby stock colony and allowed to heal for 2 weeks | -   | -          | -   | -                  |
| Diseased Coral (D)             | 4-9 cm branch tip with active disease snipped from nearby stock colony immediately prior to deployment in trial. Breaks were made on dead skeleton approximately 2 cm below active disease margin | 29  | 23         | 52  | 32                 |
| Mechanically Damaged Coral (M) | Healthy branch tip with 3-5 cm section actively abraded with a dead A. cervicornis branch immediately prior to deployment in a trial | 32  | 26         | 58  | 23                 |
| Solitary Snail (S)             | Snail tethered at end of one treatment lane with no coral               | 28  | 17         | 45  | 65                 |
| Coral with Active Snail Predation (P) | Snail feeding on a healthy fragment from an immediately prior trial | 2   | 1          | 3   | 0                  |