Abstract

Malaria remains a major cause of mortality in African children, with no adjunctive treatments currently available to ameliorate the severe clinical forms of the disease. Rosetting, the adhesion of infected erythrocytes (IEs) to uninfected erythrocytes, is a parasite phenotype strongly associated with severe malaria, and hence is a potential therapeutic target. However, the molecular mechanisms of rosetting are complex and involve multiple distinct receptor–ligand interactions, with some similarities to the diverse pathways involved in Plasmodium falciparum erythrocyte invasion. This review summarizes the current understanding of the molecular interactions that lead to rosette formation, with a particular focus on host uninfected erythrocyte receptors including the A and B blood group trisaccharides, complement receptor one, heparan sulphate, glycoporphin A and glycoporphin C. There is strong evidence supporting blood group A trisaccharides as rosetting receptors, but evidence for other molecules is incomplete and requires further study. It is likely that additional host erythrocyte rosetting receptors remain to be discovered. A rosette-disrupting low anti-coagulant heparin derivative is being investigated as an adjunctive therapy for severe malaria, and further research into the receptor–ligand interactions underlying rosetting may reveal additional therapeutic approaches to reduce the unacceptably high mortality rate of severe malaria.

Introduction

Rosetting is a Plasmodium falciparum infected erythrocyte (IE) adhesion phenotype that is associated with severe malaria in sub-Saharan Africa (summarized in Doumbo et al., 2009). It is a form of cell adhesion in which erythrocytes infected with mature, asexual parasites bind to uninfected erythrocytes to form clusters of cells (Fig. 1). Rosetting is a phenotypically variable property, which is common in parasite isolates collected from severe malaria patients, but infrequent in parasites from uncomplicated malaria cases. For culture-adapted P. falciparum isolates, only a subset of parasite lines can be selected in vitro for the rosetting phenotype, and many of the commonly used laboratory strains such as 3D7, rosette poorly or not at all. The relative rarity of rosetting in culture-adapted parasite lines may explain why rosetting is studied infrequently, despite being a virulence-associated phenotype in clinical isolates.

Rosetting can contribute to IE sequestration and microvascular congestion, leading to obstruction to blood flow (Kaul et al., 1991), one of the major pathological events in severe falciparum malaria contributing to inflammation, tissue damage and organ failure (Miller et al., 2002; White et al., 2013). Rosetting also causes membrane changes in uninfected erythrocytes that may contribute to phagocytic removal and anaemia (Uyoga et al., 2012). In Africa, high levels of rosetting occur in parasites sampled from severe malaria patients with all clinical types of disease including cerebral malaria (Carlson et al., 1990; Treutiger et al., 1992; Ringwald et al., 1993; Rowe et al., 1995; Kun et al., 1998; Doumbo et al., 2009), severe malarial anaemia (Newbold et al., 1997; Doumbo et al., 2009) and respiratory distress (Warimwe et al., 2012). Rosette-like clusters of cells have been seen in the microvasculature in histological studies of fatal malaria cases (Dondorp et al., 2004; Barrera et al., 2018). The major Plasmodium species that infect humans are all able to form rosettes (Udomsangpet et al., 1995; Angus et al., 1996; Choivanich et al., 1998; Lowe et al., 1998). However, the link between severity of disease and rosetting is confined to P. falciparum, possibly due to the unique ability to bind both endothelial cells and uninfected erythrocytes simultaneously (Udomsangpet et al., 1992; Adams et al., 2014), such that P. falciparum rosetting IEs are sequestered and not seen in peripheral blood. Recently it has been suggested that rosetting may contribute to anaemia in Plasmodium vivax infections (Marin-Menéndez et al., 2013).

The biological function of rosetting in vivo remains unknown. Rosettes may shield IEs from host immune attack, or close contact with uninfected erythrocytes in rosettes might enhance merozoite invasion (Wahlgren et al., 1989; Deans and Rowe, 2006). However, firm evidence to support either of these hypotheses is lacking. Most rosetting parasite isolates form larger, stronger rosettes with blood group A erythrocytes compared to other blood groups (Carlson and Wahlgren, 1992), and these group A rosettes may shield IEs to reduce antibody binding to parasite variant surface antigens (VSAs) (Moll et al., 2015). Whether this translates
Rosetting receptors on host erythrocytes

A number of different molecules on uninfected erythrocytes have been proposed as receptors for *P. falciparum* rosetting (Fig. 2 and Table 1), and multiple receptor–ligand interactions may contribute to rosetting in any given parasite isolate. Some of the proposed rosetting receptor molecules, including blood group A and B sugars, hepan sulphate (HS)-like molecules and complement receptor one (CR1) are widely accepted as having a role in rosetting, whereas other recent candidates such glycoporin A (GYPA) and glycoporin C (GYPc) are less well-authenticated. However, a close examination of the underlying data shows that in most cases, the evidence is incomplete, as discussed in detail below.

Evidence needed to establish a role for a specific host receptor in rosetting

In order to prove that a particular molecule acts as a host receptor for *P. falciparum* rosetting, a variety of different types of evidence have been provided. Essential data include proof that the molecule in question is found on normal human erythrocytes and that erythrocytes lacking the molecule show reduced/absent rosetting. Direct binding between IEs and/or recombinant parasite adhesion proteins and the receptor molecule should be demonstrated. Ideally, a crystal structure of the parasite adhesion molecule–host receptor complex should show the precise binding interaction site. Supportive evidence includes the ability of antibodies against the receptor or soluble receptor proteins to inhibit rosetting, and biochemical approaches to remove or alter the receptor on erythrocytes. Human genetic evidence can also provide indirect supportive evidence that particular molecules are important in life-threatening malaria. Several putative rosetting receptors have high-frequency polymorphisms in populations from malaria endemic regions that reduce rosetting and are associated with protection against severe malaria and death [reviewed in Rowe et al. (2009a, 2009b)]. These various lines of evidence are summarized below for each potential host rosetting receptor.

Blood group A and B trisaccharides

The most well-validated rosetting receptors are the blood group A and B trisaccharides (Fig. 3). *In vitro* experiments have shown that rosetting parasites have a ‘preference’ for blood groups A, B or AB rather than O (Carlson and Wahlgren, 1992; Udomsangphet et al., 1993; Barragan et al., 2000b; Pipitaporn et al., 2000; Vigan-Womas et al., 2012; Moll et al., 2015). This varies by parasite genotype, with A-preference being the commonest. Clinical isolates from non-O (i.e. A, B or AB) patients show higher levels of rosetting than isolates from group O patients in studies from sub-Saharan Africa (Rowe et al., 1995, 2007) and India (Rout et al., 2012), although the same result was not seen in one Thai
study (Lee et al., 2014). When parasites are cultured in their ‘preferred’ blood group, they form larger, stronger rosettes that are more resistant to disruption by antibodies or chemical agents than in group O cells (Carlson and Wahlgren, 1992; Barragan et al., 2000b; Chung et al., 2016). Enzymatic removal of the terminal sugars (N-acetyl-D-galactosamine for A and D-galactose for B) results in smaller, weaker rosettes, equivalent to those seen in group O erythrocytes (Barragan et al., 2000b). Rosettes do, however, still occur with blood group O erythrocytes (that express the H antigen), and also in Bombay phenotype red cells (Carlson and Wahlgren, 1992; Rowe et al., 1997). This indicates that other red cell surface molecules in addition to the A and B antigens can act as host receptors for rosette formation.

For the blood group A-preferring parasite line, Palo Alto 89F5, direct binding between the VarO PfEMP1 adhesion molecule and the blood group A trisaccharide was shown by Surface Plasmon Resonance (Vigan-Womas et al., 2012). The VarO PfEMP1 variant also binds to the B trisaccharide, but with lower affinity (Vigan-Womas et al., 2012). A crystal structure of the PfEMP1 N-terminal region was obtained and the A-trisaccharide binding site mapped (Vigan-Womas et al., 2012). A recent study suggests that P. falciparum RIFIN molecules may also be able to interact with blood group A sugars to contribute to rosette formation (Goel et al., 2015), although direct RIFIN-A trisaccharide interaction was not shown.

The importance of the A and B antigens in rosetting is emphasized by the fact that the non-O blood groups are associated with increased risk of severe malaria and death compared to O (Rowe et al., 2007; Fry et al., 2008; Tekeste and Petros, 2010; Rout et al., 2012; Malaria Genomic Epidemiology Network, 2014; Ndila et al., 2018; Degarege et al., 2019). Reduced rosetting in blood group O, and therefore reduced microvascular obstruction and reduced downstream pathological effects, is the proposed mechanism for the protective association with group O (Udomsangpetch et al., 1993; Rowe et al., 2007). ABO blood group does not influence parasite burden (Rowe et al., 2007; Degarege et al., 2019), and evidence for an effect of ABO on P. falciparum invasion or other host–parasite interactions is conflicting and requires further study (Chung et al., 2005; Wolofsky et al., 2012; Pathak et al., 2016; Theron et al., 2018). The ABH antigens are known to be present on endothelial cells (Ito et al., 1990) and it is likely, but has not been shown experimentally, that cytoadhesion and overall levels of sequestration of rosetting parasites are enhanced in group A/B/AB patients compared to O.

Despite the progress in identifying the A and B trisaccharides as rosetting receptors and key genetic determinants of host susceptibility to severe malaria, there have been no attempts to develop specific therapies to block P. falciparum interaction with A/B antigens. The PfEMP1-blood group A trisaccharide binding pair described above (Vigan-Womas et al., 2012) remains the most clearly defined molecular interaction between parasite ligand and host receptor in rosetting, and could be used as a starting point to develop rosette-blocking therapeutics. Vigan-Womas et al. did report that the interaction between PfEMP1 and the A and B trisaccharides is indirectly inhibited by heparin (Vigan-Womas et al., 2012), and the development of a heparin-derivative as a potential adjunctive therapy for severe malaria is described below (Leitgeb et al., 2017).

**Complement receptor one (CR1, CD35)**

CR1 is a red cell membrane glycoprotein that regulates complement activation on cell surfaces (Thielen et al., 2018) and carries the Knops Blood Group antigens (Moulds, 2010). In malaria, CR1 plays a role in both rosetting and parasite invasion of erythrocytes (Schmidt et al., 2015). CR1 was first identified as a rosetting receptor from a screen of 23 naturally occurring erythrocyte
A Parasite strains used are not consistent between studies with a wide range of culture-adapted and clinical isolates in use. Results are therefore not necessarily generalizable from single studies.

**Table 1. Summary of host erythrocyte receptors for Plasmodium falciparum rosetting**

| Name                                      | Characteristics                                                                 | Studies*                                                                                           | Comments                                                                                       |
|-------------------------------------------|--------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------|
| **ABO blood group antigens**              |                                                                               |                                                                                                   |                                                                                                |
| Differ based on terminal sugar:           |                                                                               |                                                                                                   |                                                                                                |
| A = N-acetyl-D-galactosamine,             |                                                                               |                                                                                                   |                                                                                                |
| B = D-galactose,                          |                                                                               |                                                                                                   |                                                                                                |
| O (H-antigen) = L-Fucose                   |                                                                               |                                                                                                   |                                                                                                |
| O is a predominant blood group in sub-Saharan Africa |                                                                               |                                                                                                   |                                                                                                |
| Blood group O protects against severe malaria |                                                                               |                                                                                                   |                                                                                                |
| (Rowe et al., 2007; Fry et al., 2008; Tekeste and Petros, 2010; Rout et al., 2012; Malaria Genomic Epidemiology Network, 2014; Ndila et al., 2018; Degarege et al., 2019) |                                                                                                   |                                                                                                |
|                                                                                       |                                                                               |                                                                                                   | Blood group A antigen is the most well-validated host rosetting receptor Both PIEMP1 (Vigan-Womas et al., 2012) and RIFINs (Goel et al., 2015) may interact with A antigen Challenging to manipulate therapeutically |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Complement receptor 1 (CR1)**           | Membrane glycoprotein responsible for regulating the complement system (Thielen et al., 2018) | Rosetting reduced in CR1 deficient erythrocytes (Rowe et al., 1997) Soluble CR1 and CR1 antibodies disrupt rosettes in some parasite isolates (Rowe et al., 1997; Rowe et al., 2000; Vigan-Womas et al., 2012) Essential region mapped to the C3b binding site on CR1 (Rowe et al., 2000) | Further work needed to assess the relative importance of CR1 in rosetting isolates and potential as a therapeutic target Soluble recombinant CR1 has been considered for therapeutic use in humans, e.g. cardiac and renal disease (Li et al., 2006; Reddy et al., 2017) |
| Polymorphisms affect CR1 copy number, molecular weight and sequence (Schmidt et al., 2015) |                                                                               |                                                                                                   |                                                                                                |
| RBC CR1 deficiency protects in medium-high (Cookburn et al., 2004; Sinha et al., 2005; Rout et al., 2011; Panda et al., 2012) but not low malaria transmission areas (Nagayasu et al., 2001; Teeranaipong et al., 2008) |                                                                                                   |                                                                                                |
| CR1 Knops blood group polymorphisms associated with severe malaria (Opi et al., 2018) |                                                                               |                                                                                                   |                                                                                                |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Heparan sulphate (HS)**a                | Glycosaminoglycan Heparin is a highly sulfated form of HS that is only found in mast cells HS is a receptor for P. falciparum sporozoite invasion of hepatocytes (Frevert et al., 1993) | Heparin partially disrupts rosettes in some isolates (Udomsangpetch et al., 1989; Carlson et al., 1992; Rogerson et al., 1994; Rowe et al., 1994; Barragan et al., 1999) Heparinase treatment reported to reduce rosetting in two culture-adapted parasite lines (Barragan et al., 1999) Heparin binds to rosetting IE (Barragan et al., 2000a; Heddini et al., 2001) and to rosette-mediating PIEMP1 (Barragan et al., 2000b; Vogt et al., 2003; Juillerat et al., 2010; Juillerat et al., 2011; Adams et al., 2014) | Limited evidence that HS is present on mature RBCs (Vogt et al., 2004) Further work needed to determine whether HS is present on normal erythrocytes and acts as a rosetting receptor Therapeutic potential due to PIEMP1 binding and rosette disruption. Clinical trials of low anticoagulant heparin ongoing (Leigeb et al., 2017) |
| Glycosaminoglycan Heparin is a highly sulfated form of HS that is only found in mast cells HS is a receptor for P. falciparum sporozoite invasion of hepatocytes (Frevert et al., 1993) |                                                                               |                                                                                                   |                                                                                                |
| HS is a receptor for infected erythrocyte cytoadherence to endothelial cells (Vogt et al., 2003; Adams et al., 2014) |                                                                               |                                                                                                   |                                                                                                |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Chondroitin sulphate (CS)**             | Glycosaminoglycan Receptor for infected erythrocyte placental sequestration in pregnancy malaria (Fried and Duffy, 1996) | Soluble CS did not disrupt rosettes (Rogerson et al., 1994; Rowe et al., 1994) Chondroitinase treatment reduced rosetting in one parasite line only (Barragan et al., 1999) | No evidence that CS is present on mature RBC Minimal evidence for a role in rosetting                                                                 |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **CD36**                                  | Widely distributed membrane protein and scavenger receptor (Silverstein and Febbraio, 2009) | Antibodies disrupt rosettes in single culture-adapted line only (Handunnetti et al., 1992) PIEMP1 variants that mediate rosetting are group A types that do not bind CD36 (Robinson et al., 2003) | Minimal evidence for a widespread role in rosetting                                                                                               |
| Deficiency is common in Africa but not associated with severe malaria (Fry et al., 2009) |                                                                               |                                                                                                   |                                                                                                |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Glycophorin C (GYPC)**                  | Red cell membrane protein responsible for Gerlich blood group (Jaskiewicz et al., 2018) Receptor for merozoite invasion of erythrocytes (Maier et al., 2003) 'Gerlich-negative' blood group common in Melanesians (Patel et al., 2001), but no evidence yet for association with protection against severe malaria | Reduced rosetting with GYPC antibodies and GYPC knockdown RBCs (Niang et al., 2014) single culture-adapted parasite line tested Gerlich-negative erythrocytes formed rosettes normally with five P. falciparum lines (Rowe et al., 1997) Possible role in P. vivax rosetting (Lee et al., 2014) | Further work needed to assess the relative importance of GYPC in P. falciparum rosetting isolates and potential as a therapeutic target                                                                 |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Glycophorin A (GYPA)**                  | Sialoglycoprotein which, along with glycophorin B, constitutes the MNS blood group receptor for merozoite invasion of erythrocytes (Sim et al., 1994) | GYPA-deficient erythrocytes showed reduced rosetting with RIFIN transfected parasites (Goel et al., 2015) GYPA antibodies had no inhibitory effect on rosetting (Lee et al., 2014) (Niang et al., 2014) GYPA null erythrocytes formed rosettes with five culture-adapted P. falciparum lines (Rowe et al., 1997) | Further work needed to assess the relative importance of GYPA in P. falciparum rosetting isolates and potential as a therapeutic target                                                                 |
|                                                                                       |                                                                               |                                                                                                   |                                                                                                |
| **Unknown receptor/s**                    | Possibly carbohydrate or protease-resistant protein | Protease and hexasporase treated erythrocytes capable of forming rosettes (Udomsangpetch et al., 1989; Rowe et al., 1994) | Further work needed to identify novel rosetting receptors                                                                                           |

*Parasite strains used are not consistent between studies with a wide range of culture-adapted and clinical isolates in use. Results are therefore not necessarily generalizable from single studies.

*Many studies included here use heparin instead of/in addition to heparan sulphate.
null mutants, each missing a particular blood group molecule or membrane glycoprotein (Rowe et al., 1997). The only variant to show substantially reduced rosetting with five *P. falciparum* parasite lines was Knops null cells, which are deficient in CR1. Normally, erythrocytes have between 100 and 1000 molecules of CR1 per cell (Wilson et al., 1986), whereas Knops null cells have fewer than 100 molecules per cell (Moulds et al., 1992).

Erythrocytes with fewer than 50 CR1 molecules per cell form rosettes poorly (Rowe et al., 1997), with normal rosetting occurring above a threshold of around 100 molecules per cell (JA Rowe, unpublished data).

Soluble CR1 and CR1 antibodies were shown to inhibit rosetting in some but, not all *P. falciparum* rosetting laboratory strains and clinical isolates, with only monoclonal antibodies (mAbs) that map to the C3b binding site on CR1 being effective inhibitors (Rowe et al., 1997, 2000; Vigan-Womas et al., 2012). A recent paper suggested that the commercially available CR1 mAb E11 that recognizes epitopes outside the C3b binding site (Nickells et al., 1998) may inhibit *P. falciparum* rosetting (Lee et al., 2014), but this was not seen in our hands (Rowe et al., 2000). Further evidence of a role for CR1 in rosetting came from the expression of recombinant PfEMP1 domains in COS-7 cells, which bound to normal erythrocytes but not to CR1-deficient cells (Rowe et al., 1997).

Despite these supportive data, direct binding of IEs to CR1 protein has not been demonstrated, and recombinant rosette-mediating PfEMP1 proteins produced in *E. coli* (Ghumra et al., 2012) do not bind to CR1 in Surface Plasmon Resonance experiments (Tetteh-Quarcoo et al., 2012). This could reflect a genuine lack of interaction between the two molecules, or could be due to technical reasons (e.g. the recombinant CR1 used in experiments was produced in mouse rather than human cells, whereas CR1 glycosylation, which may affect function, is cell-type specific) (Lublin et al., 1986). It is also possible that a serum protein mediates the interaction between PfEMP1 on IEs and CR1 on uninfected erythrocytes, as the original experiments were all performed in the presence of serum (Rowe et al., 1997).

Human genetic studies provide additional support for the importance of CR1 in malaria host–parasite interactions.

Erythrocyte CR1 deficiency is common in some malaria-endemic countries such as Papua New Guinea (Cockburn et al., 2004) and India (Sinha et al., 2009), and is associated with protection against severe malaria in medium to high transmission areas (Cockburn et al., 2004; Sinha et al., 2009; Rout et al., 2011; Panda et al., 2012). However, erythrocyte CR1 deficiency may be detrimental in areas such as Thailand, where malaria transmission is low (Nagayasu et al., 2001; Teeeranaipong et al., 2008). There is also evidence that the CR1 Swain Langley 2 (SI2) Knops blood group polymorphism that is common in African populations (Moulds, 2010) is associated with protection against severe malaria (Thathy et al., 2005; Opie et al., 2018). Red cells carrying the SI2 antigen on CR1 show reduced rosetting (Rowe et al., 1997; Opie et al., 2018), and SI2 may have additional effects on complement activation and regulation (Opie et al., 2018).

Overall, the ability of CR1 mAbs and soluble protein to reverse rosettes suggests that CR1 plays a role in rosetting for some *P. falciparum* isolates. However, further work is needed to fully investigate the molecular interactions between parasite adhesion molecules and CR1, and to explore the potential for CR1 reagents (Li et al., 2006; Reddy et al., 2017) as therapeutic disruptors of rosetting.

**Heparan sulphate and chondroitin sulphate**

The glycosaminoglycans HS and chondroitin sulphate (CS) are found on cell surfaces and in the extracellular matrix of many tissues, and have a role in multiple aspects of the *P. falciparum* life cycle including hepatocyte invasion (Frevert et al., 1993), endothelial cell cytoadherence (Vogt et al., 2003; Adams et al., 2014) and, for CS, placental sequestration (Fried and Duffy, 1996). A number of papers have shown that heparin (which is a highly-sulphated form of HS found only in mast cells) can partially disrupt rosettes in about one-third to one-half of *P. falciparum* clinical isolates in vitro (Udomsangpetch et al., 1989; Carlson et al., 1992; Rogerson et al., 1994; Rowe et al., 1994; Barragan et al., 1999). It was shown that treating erythrocytes with heparinase III, which selectively cleaves HS chains, reduces rosetting in two *P. falciparum* lines (Barragan et al., 1999), and therefore suggested that 'HS-like'
molecules on red cells are receptors for rosetting (Chen et al., 2000). However, there has been only one paper reporting the existence of HS on normal human erythrocytes (Vogt et al., 2004) and we have been unable to confirm this, and unable to detect any rosette-reducing effect of heparinase III in a range of parasite lines (McQuaid and Rowe, unpublished data).

Fluorescently-labelled heparin does bind to the surface of erythrocytes infected with rosetting parasites more than non-rosetting lines (Barragan et al., 2000a; Heddini et al., 2001), and some rosette-mediating PfEMP1 variants bind directly to heparin (Barragan et al., 2000a; Vogt et al., 2003; Juillerat et al., 2010, 2011; Adams et al., 2014). The heparin binding site in the N-terminal region of the varO PfEMP1 variant was mapped onto a crystal structure (Juillerat et al., 2011), and shown to be on the opposite side of the molecule from the erythrocyte binding site (Vigan-Womas et al., 2012). Hence, the rosette-disrupting effect of heparin is not due to direct blocking of receptor binding, but may result from aggregating PfEMP1 monomers and preventing their interaction with erythrocyte receptors (Vigan-Womas et al., 2012). Similarly, for another rosette-mediating PfEMP1 variant IT4var60, site-directed mutagenesis studies of recombinant proteins showed that mutations that disrupt heparin binding are distinct from mutations that disrupt erythrocyte binding, indicating that heparin-like molecules are not the main host rosetting receptor in this case (Angeletti et al., 2013).

Overall, whether HS is present on normal erythrocytes and is a host receptor for rosetting requires further confirmation. HS in present on the luminal surface of microvascular endothelial cells (albeit at a much lower density than on basolateral surfaces) (de Agostini et al., 1990; Stoler-Barak et al., 2014), therefore interactions between IE and endothelial HS (Vogt et al., 2003; Adams et al., 2014) are physiologically relevant and are likely to contribute to cytoadherence and sequestration in vivo.

Despite the uncertainty on the precise role of HS as an erythrocyte rosetting receptor, heparin and other sulphated glycoconjugate compounds have clear potential as adjunctive therapies for severe malaria due to their rosette-disrupting effects (Udomsangphet et al., 1989; Carlson et al., 1992; Rogerson et al., 1994; Rowe et al., 1994; Kyriacou et al., 2007). There are reports of successful heparin treatment in severe malaria (Rampengan, 1991) but its use is not recommended due to a high incidence of bleeding complications (World Health Organisation, 1986). As an alternative, Wahlgren and coworkers have developed a low anti-coagulant heparin derivative, Sevuparin, that reverses rosetting and cytoadherence in some P. falciparum isolates (Leitgeb et al., 2011; Saiwaew et al., 2017) and also blocks merozoite invasion (Leitgeb et al., 2017). Sevuparin has been shown to be safe in adults with uncomplicated malaria (Leitgeb et al., 2017), but has not yet been tested in severe malaria patients.

The evidence for CS as a rosetting receptor is minimal. One report shows that rosetting in the P. falciparum line TM284 was partially inhibited by soluble CS and by chondroitinase ABC treatment of erythrocytes, and that several clinical isolates showed reduced rosetting in the presence of CS (Barragan et al., 1999). However, other studies have found no effect of CS on rosetting in a variety of culture-adapted lines and clinical isolates (Rogerson et al., 1994; Rowe et al., 1994). There is also no convincing evidence that CS is found on the surface of normal human erythrocytes. Overall, current data do not support a role for CS in rosetting.

**CD36**

The membrane glycoprotein CD36 is a scavenger receptor for oxidized lipoproteins and a fatty acid translocase (Silverstein and Febbraio, 2009). It is expressed on a variety of cell types including monocytes, macrophages, platelets, microvascular endothelial cells and adipocytes (Silverstein and Febbraio, 2009), and at low levels on erythrocytes (van Schravendijk et al., 1992). The binding of PfEMP1 (group B and C variants) to CD36 on microvascular endothelial cells plays a major role in P. falciparum sequestration (Baruch et al., 1996; Robinson et al., 2003). Almost all P. falciparum isolates bind to CD36, and increased CD36 binding (Newbold et al., 1997; Ochola et al., 2011) and predominant expression of group B and C PfEMP1 (Kraemer and Smith, 2006; Kyriacou et al., 2006) are associated with uncomplicated malaria.

The role of CD36 in rosetting is less clear. Anti-CD36 mAbs are capable of disrupting rosettes in a single culture-adapted parasite line, Malayan Camp (Handunnetti et al., 1992), but not in a wide range of other laboratory lines or clinical isolates (Udomsangphet et al., 1989; Wahlgren et al., 1992; Rowe et al., 2000; Niang et al., 2014). The PfEMP1 variants identified as parasite rosetting ligands (Rowe et al., 1997; Vigan-Womas et al., 2011; Ghunna et al., 2012) are mostly of the group A type, which do not bind to CD36 (Robinson et al., 2003).

Intriguingly, while CD36 deficiency is fairly common in African populations, large-scale genetic studies have shown that CD36 polymorphisms do not influence severe malaria risk (Fry et al., 2009). There is some evidence that interaction between IEs and CD36 may benefit the host, as CD36 may contribute to innate immune clearance of IEs and platelet-mediated parasite death (McGilvray et al., 2000; McMorrán et al., 2012; Cabrera et al., 2014). Overall, it is unlikely that CD36 is a clinically significant rosetting receptor or a useful therapeutic target in severe malaria (Cabrera et al., 2014).

**Glycoporphin C (GYPC; GPC; CD236)**

GYPC is a red cell membrane glycoprotein that carries the Gerbich blood group antigens (Jaskiewicz et al., 2018). It is a P. falciparum invasion receptor bound by the merozoite protein EBA-140/BAEBL (Maier et al., 2003; Mayer et al., 2006). Recently, two studies have suggested that GYPC is a rosetting receptor for both P. falciparum (Niang et al., 2014) and P. vivax (Lee et al., 2014; Niang et al., 2014). Niang et al. showed that the rosetting of a 3D7-derived P. falciparum laboratory strain (5A-R+) was partially inhibited by a GYPC mAb (clone Ret40f) and by soluble recombinant GYPC (Niang et al., 2014). Furthermore, cultured GYPC knockdown erythrocytes failed to rosette, providing strong evidence that GYPC is an essential rosetting receptor for 5A-R+ parasites (Niang et al., 2014). Other parasite lines or clinical isolates were not tested, therefore the wider role of GYPC in P. falciparum rosetting was not determined.

Lee et al. (2014) focussed mainly on P. vivax, but also assessed the ability of GYPC mAb fragments to inhibit rosette formation in ten P. falciparum clinical isolates from Thailand. A significant decrease in rosetting was reported with GYPC mAb BRIC 40f and by soluble recombinant GYPC (Niang et al., 2014). There is evidence that the binding of two or more uninfected erythrocytes, which helps to identify genuine cell–cell interactions and avoid spurious identification of rosettes due to close packing of cells under the coverslip during microscopy.

For P. vivax, Lee et al. showed that the GYPC mAb reduced the median rosette frequency from 30 to 22% when tested on 11 Thai
isolates, and that GYPC knockdown cultured erythrocytes formed rosettes poorly compared to GYPC-positive control cells (median rosette frequency 6.2 vs. 35.4% in controls, tested on three isolates). *Plasmodium falciparum* isolates were not tested with the GYPC knockdown erythrocytes.

If GYPC is a rosetting receptor, it is possible that the ‘Gerbich-negative’ blood group type, which is common in Melanesian populations (Patel et al., 2001), might influence rosetting. As part of a screen of null blood group erythrocytes with five high-rosetting *P. falciparum* culture-adapted parasite lines, Rowe et al. (1997) tested two donors with the Gerbich-negative blood group (formed by deletion of exon 3 of the GYPC gene on chromosome 2, giving a truncated protein with altered glycosylation). Gerbich-negative erythrocytes formed rosettes normally with the five parasite lines tested. Goel et al. also report normal rosetting of Gerbich-negative erythrocytes from two donors (Goel et al., 2015). The true null phenotype for GYPC, called the Leach phenotype (which arises due to the deletion of exon 3 and exon 4, encoding the transmembrane and cytoplasmic domains, respectively) is rare and has not been tested in rosetting assays to our knowledge.

Taking into account all existing evidence, further investigation of a wider range of parasite lines is needed to determine whether GYPC is an important host receptor for both *P. falciparum* and *P. vivax* rosetting.

**Glycopyrin A (GYPA, GPA, CD235a)**

GYPA is a highly-expressed erythrocyte surface glycoprotein that carries the MNS blood group antigens. It is known to be a receptor for *P. falciparum* erythrocyte invasion (Sim et al., 1994), and polymorphisms in GYPA are associated with resistance to severe malaria (Band et al., 2015; Leffler et al., 2017).

There is some limited evidence to suggest that GYPA may have a role in rosetting. Parasites of the strain FCR3S1.2 transfected with a specific RIFIN gene formed rosettes that were largely dependent on blood group A (Goel et al., 2015). However, rosetting of the RIFIN-transfected parasites was significantly reduced with GYPA null cells from blood group O and B donors, whereas blood group A GYPA null erythrocytes formed rosettes normally. These data suggest that GYPA may have an accessory role for RIFIN-mediated rosetting in the absence of the A antigen (Goel et al., 2015), although whether this applies to rosetting in non-genetically manipulated parasites is unknown.

Despite the above positive evidence, there are no other data supporting a role for GYPA in rosetting. GYPA mAb fragments had no inhibitory effect on rosetting in ten *P. falciparum* and 11 *P. vivax* clinical isolates (Lee et al., 2014), and a GYPA mAb did not inhibit 3D7 SA-R+ rosettes (Niang et al., 2014). Furthermore, GYPA null erythrocytes (MKM cells, lacking both GYPA and glycopyrin B) formed rosettes with five culture-adapted *P. falciparum* lines (Rowe et al., 1997). Overall, existing evidence does not support a major role for GYPA in rosetting, but as with GYPC, further investigation is needed.

**New receptors and new approaches**

None of the receptors described above fully account for the adhesion interactions between infected and uninfected erythrocytes, and it is likely that other host rosetting receptors remain to be identified. There is evidence to suggest that these unknown host receptors are carbohydrates or protein-resistant proteins, because uninfected group O erythrocytes treated with trypsin and other proteases are still able to form rosettes (Udomsangpetch et al., 1989; Rowe et al., 1994).

In order to progress rosetting research, alternative methods are needed. Rosetting experiments with GYPC and CR1 knockdown cultured human red cells derived from CD34+ haematopoietic stem cells have been performed (Lee et al., 2014; Niang et al., 2014), using lentiviral transduction of short hairpin RNA (Be et al., 2010). However, these cultured erythrocytes have a short life-span, limiting their usefulness. The development of immortalized erythroid lines (Kurita et al., 2013; Kanjee et al., 2017; Trakarnsanga et al., 2017; Scully et al., 2019) may overcome this limitation. Nevertheless, attention must be paid to the subtle but real differences between mature erythrocytes and these, still relatively immature, immortalized CD34+ derived cells (Wilson et al., 2016; Dankwa et al., 2017; Trakarnsanga et al., 2017). CRISPR-Cas9 technology (Doudna and Charpentier, 2014) has led to an explosion in the ability to genetically manipulate multiple cell types, including erythrocyte precursors and immortalized haematopoietic lines (Song et al., 2015; Kanjee et al., 2017; Hawksworth et al., 2018; Chung et al., 2019; Scully et al., 2019), potentially giving the opportunity to generate multiple knockout lines for rosetting research. A consistent supply of knockout erythrocytes would allow large-scale screens for new rosetting receptors using cells as close to their normal physiological form as possible, raising exciting prospects for future work.

**Conclusions**

Of the rosetting receptors described over the past 30 years, only the blood group A trisaccharide has been authenticated by a variety of methodological approaches from a range of different investigators. For all other potential rosetting receptors, the evidence remains fragmentary (Table 1) and further research is needed (Table 2). Recent technical advances in genetic manipulation of red cell precursors and immortalised lines should enable reverse genetic studies to bring further clarity to this biologically important topic.

**Financial support.** This work was supported by the Welcome Trust (PhD studentship grant number 108685/Z/15/Z).

**Conflict of interest.** None.

**Ethical standards.** Not applicable.

**Table 2. Key areas for future research on rosetting receptors**

| Area                                                                 | Notes                                                                 |
|----------------------------------------------------------------------|----------------------------------------------------------------------|
| Determine the relative importance of known host erythrocyte receptors |                                                                  |
| Develop screening technologies to identify novel host rosetting receptors |                                                                  |
| Use immortalized erythroid lines for reverse genetic studies in rosetting |                                                                  |
| Develop novel rosette-disrupting adjunctive therapies               |                                                                  |
| Develop in vivo models to test rosette-disrupting adjunctive therapies |                                                                  |

**References**

Adams Y, Kuhnrae P, Higgins MK, Ghumra A and Rowe JA (2014) Rosetting *Plasmodium falciparum*-infected erythrocytes bind to human brain microvascular endothelial cells in vitro, demonstrating a dual adhesion phenotype mediated by distinct *P. falciparum* erythrocyte membrane protein 1 domains. *Infection and Immunity* 82, 949–959.

Albrecht I, Moll K, Blomqvist K, Normark J, Chen Q and Wahlgren M (2011) Var gene transcription and PEmPI expression in the rosetting and cytoadhesive *Plasmodium falciparum* clone FCR3S1.2. *Malaria Journal* 10, 17.

Angetti D, Sandalova T, Wahlgren M and Achour A (2015) Binding of subdomains 1/2 of PEmPI-DB1alphA to heparan sulfate or heparin mediates *Plasmodium falciparum* rosetting. *PLoS ONE* 10, e0118888.

Angus BJ, Thanikkul K, Silamut K, White NJ and Udomsangpetch R (1996) Short report: rosette formation in *Plasmodium ovale* infection. *American Journal of Tropical Medicine and Hygiene* 55, 560–561.
Carlson J, Spencer CC, Kwiatkowski DP and Network MGE (2015) A novel locus of resistance to severe malaria in a region of ancient balancing selection. Nature 526, 253–257.

Barragan A, Spillmann D, Kremsner PG, Wahlgren M and Carlson J (1999) Plasmodium falciparum: molecular background to strain-specific rosette disruption by glycosaminoglycans and sulfated glycoconjugates. Experimental Parasitology 91, 133–143.

Barragan A, Fernandez V, Chen Q, von Euler A, Wahlgren M and Spillmann D (2000a) The Duffy-binding-like domain 1 of Plasmodium falciparum erythrocyte membrane protein 1 (PfEMP1) is a heparan sulfate ligand that requires 12 mers for binding. Blood 95, 3594–3599.

Barragan A, Kremsner PG, Wahlgren M and Carlson J (2000b) Blood group A antigen is a coreceptor in Plasmodium falciparum rosetting, Infection and Immunity 68, 2971–2975.

Bera V, MacCracken JG, Cambier C, Hiscott PS, White VA, Craig AG, Beare NAV, Cuhel LH, Zheng Y, Biddolph SC, Milner DA, Kamiza S, Molyneux ME, Taylor TE and Harding SP (2018) Neurovascular sequestration in paediatric Plasmodium falciparum malaria is visibly clinical in the retina. Elife 7, e32208. doi: 10.7554/elif.e32208.

Baruch DI, Gormley JA, Ma C, Howard RJ and Pasloske BL (1996) Plasmodium falciparum erythrocyte membrane protein 1 is a parasitized erythrocyte receptor for adherence to CD36, thrombospondin, and intercellular adhesion molecule 1. Proceedings of the National Academy of Sciences USA 93, 3497–3502.

Beai K, Brugnara C and Duraisingham MT (2010) In vitro genetic analysis of an erythrocyte determinant of malaria infection. Journal of Infectious Diseases 202, 1722–1727.

Cabrera A, Neculai D and Kain KC (2014) CD36 and malaria: friends or foes? A decade of data provides some answers. Experimental Parasitology 142, 53–64.

Dondorp AM, Pongponratn E and White NJ (2004) Reduced microcirculatory flow in severe falciparum malaria: pathophysiology and electron-microscopic pathology. Acta Tropica 89, 309–317.

Doudna JA and Charpentier E (2014) Genome editing. The new frontier of genome engineering with CRISPR-Cas9. Science 346, 1258096.

Doumbo OK, Thera MA, Koné AK, Raza A, Tempest LJ, Lyke KE, Plowe CV and Rowe JA (2008) High levels of Plasmodium falciparum infection in all clinical forms of severe malaria in African children. American Journal of Tropical Medicine and Hygiene 81, 987–993.

Fleuret U, Sinnis P, Cerami C, Shreffler W, Takas B and Nussenzweig V (1993) Malaria circumsporozoite protein binds to heparan sulfate proteoglycans associated with the surface membrane of hepatocytes. Journal of Experimental Medicine 177, 1287–1298.

Fried M and Duffy PE (1996) Adherence of Plasmodium falciparum to chondroitin sulfate A in the human placenta. Science 272, 1502–1504.

Fry AE, Griffiths MJ, Auburn S, Diakite M, Forton JT, Green A, Richardson A, Wilson J, Jallow M, Siyad-Jof F, Pinder M, Peshu N, Williams TN, Marsh K, Molyneux ME, Taylor TE, Rowan KA and Kwiatkowski DP (2008) Common variation in the ABO glycosyltransferase is associated with susceptibility to severe Plasmodium falciparum malaria. Human Molecular Genetics 17, 567–576.

Fry AE, Ghansa A, Small KS, Palma A, Auburn S, Diakite M, Green A, Campino S, Teo YY, Clark TG, Jeffreys AE, Wilson J, Jallow M, Siyad-Jof F, Pinder M, Griffiths MJ, Peshu N, Williams TN, Newton CR, Marsh K, Molyneux ME, Taylor TE, Koram KA, Oduro AR, Rogers WO, Rowan KA, Sabeti PC and Kwiatkowski DP (2009) Positive selection of a CD36 nonsense variant in sub-Saharan Africa, but no association with severe malaria phenotypes. Human Molecular Genetics 18, 2683–2692.

Ghumra A, Semblat JP, McIntosh RS, Raza A, Rasmussen IB, Braithen R, Johansen FE, Sandlie I, Mongini PK, Rowe JA and Plesch RJ (2008) Identification of residues in the C-terminus domain of parasomic IgM essential for interaction with Plasmodium falciparum erythrocyte membrane protein 1 (PfEMP1). Journal of Immunology 181, 1988–2000.

Ghumra A, Semblat JP, Ataiade R, Kifude C, Adams Y, Claessens A, Anong DN, Bull PC, Fennell C, Arman M, Amambua-Ngwa A, Walther M, Conway DJ, Kasambara L, Doumbo OK, Raza A and Rowe JA (2012) Induction of strain-transcending antibodies against Group A PfEMP1 surface antigens from virulent malaria parasites. PLoS Pathogens 8, e1002665.

Goel S, Palmkvist M, Poll K, Joannin N, Lara P, Ahkouri RR, Moradi N, Öjemalm K, Westman M, Angeletti D, Kjellin H, Lehtio J, Blixt O, Iedstrom L, Gahmbberg CG, Storry JR, Hult AK, Olsson ML, von Heijne G, Nilsson I and Wahlgren M (2013) RIFINs are adhesins implicated in severe Plasmodium falciparum malaria. Nature Medicine 21, 314–317.

Handunnetti SM, van Schravendijk MR, Hasler T, Barnwell JW, Greenwell DE and Howard RJ (1992) Involvement of CD36 on erythrocytes as a rosetting receptor for Plasmodium falciparum-infected erythrocytes. Blood 80, 2097–2104.

Hawksworth J, Satchwell TJ, Meinders M, Daniels DE, Regan F, Thornton NM, Wilson MC, Dobge JG, Streekstra GJ, Trakarnsanga K, Heesom KJ, Anstee DJ, Frayne J and Toye A (2018) Enhancement of red blood cell transfusion compatibility using CRISPR-mediated erythroblast gene editing. EMBO Molecular Medicine 10, e8454. doi: 10.15252/emmm.201708454.
Juillerat A, Pettersson F, Kai O, Shafi J, Obiero J, Chen Q, Barragan A, Li JS, Jagers J and Anderson PA (2006) The use of TP10, soluble complement receptor 1, in cardiopulmonary bypass. Expert Reviews in Cardiovascular Therapy 4, 649–654.

Lowe BS, Mosobo M and Bull PC (1998) All four species of human malaria parasites form rosettes. Transactions of the Royal Society of Tropical Medicine and Hygiene 92, 526.

Lublin DM, Griffith RC and Atkinson JP (1996) Influence of glycosylation on allelic and cell-specific Mr variation, receptor processing, and ligand binding of the human complement C3b/C4b receptor. Journal of Biological Chemistry 261, 5736–5744.

Luginbuhl A, Nikolic M, Beck HP, Wahlgren M and Lutz HU (2007) Complement factor D, albumin, and immunoglobulin G anti-band 3 protein antibodies mimic serum in promoting rosetting of malaria-infected red blood cells. Infection and Immunity 75, 1771–1777.

Maier AG, Duraisingham MT, Reeder JC, Patel SS, Kazura JW, Zimmerman PA and Cowman AF (2003) Plasmodium falciparum erythrocyte invasion through glycoporphin C and selection for Gerbic negativity in human populations. Nature Medicine 9, 87–92.

Malaria Genomic Epidemiology Network (2014) Reappraisal of known malaria resistance loci in a large multicenter study. Nature Genetics 46, 1197–1204.

Martín-Menéndez A, Bardaji A, Martínez-Espinosa FE, Böttó-Menezes C, Lacerda MV, Ortiz J, Cisteró P, Piqueras M, Felger I, Müller I, Ordí J, del Portillo H, Menéndez C, Wahlgren M and Mayor A (2013) Rosetting in Plasmodium vivax: a cytoadhesion phenotype associated with anaemia. PLoS Neglected Tropical Diseases 7, e2155.

Mayor DC, Jiang L, Achur RN, Kakizaki I, Gowda DC and Miller LH (2006) The glycoporphin C N-linked glycan is a critical component of the ligand for the Plasmodium falciparum erythrocyte receptor BAEBL. Proceedings of the National Academy of Sciences USA 103, 2358–2362.

McGivray JD, Sergidis L, Kapus A, Rotstein OD and Kain KC (2000) Nonopsomic monocyte/macrophage phagocytosis of Plasmodium falciparum-parasitized erythrocytes: a role for CD36 in malarial clearance. Blood 96, 3321–3340.

McMorran BJ, Wieczorski L, Drysdale KE, Chan JA, Huang HM, Smith C, Awuondo KO, Mturi N, Tsofa B, Sepulveda N, Clark TG, Band G, Minakawa N, Eapen A, Winkler S, Rathan S, Vala H, Sepulveda N, Clark TG, Band G, Wagner M and Masekwa J (2015) Pfemp1 gene transcription in Plasmodium falciparum: establishment of immortalized human erythroid progenitor cell lines able to produce enucleated red blood cells. Current Opinion in Microbiology 35, 356–364.

Miller LH, Baruch DI, Marsh K and Doumbo OK (2002) The pathological basis of malaria. Nature 415, 673–679.

Moll K, Palmkvist M, Ch‘ng J, Kiwuro MS and Wahlgren M (2015) Evasion of immunity to Plasmodium falciparum: rosettes of blood group A impair recognition of PEMP1. PLoS ONE 10, e0145120.

Moulds JM (2010) The Knops blood-group system: a review. Immunohematology 26, 2–7.

Moulds JM, Moulds JJ, Brown M and Atkinson JP (1992) Antiglobulin testing for CR1-related (Knops/Mcc/Osw/Scw/Slan-Langley/York) blood group antigens: negative and weak reactions are caused by variable expression of CR1. Vox Sang 62, 230–235.

Nagayasu E, Ito M, Akaki M, Nakano Y, Kimura M, Looareesuwan S and Wahlgren M (2007) Merozoite surface antigen 1 and 2 gene transcription in Plasmodium falciparum based on allelic and cell-specific Mr variation, receptor processing, and ligand binding of the human complement C3b/C4b receptor. Journal of Biological Chemistry 282, 13518–13526.

Nagayasu E, Ito M, Akaki M, Nakano Y, Kimura M, Looareesuwan S and Wahlgren M (2001) CR1 density polymorphism on erythrocytes of falciparum malaria patients. Transactions of the Royal Society of Tropical Medicine and Hygiene 95, 526–530.

Newbold C, Warn P, Black G, Berendt A, Craig A, Snow B, Msobo M, Peshu N and Marsh K (1997) Receptor-specific adhesion and clinical
disease in *Plasmodium falciparum*. *American Journal of Tropical Medicine and Hygiene* 57, 389–398.

Niang M, Bei AK, Madnani KG, Pelly S, Dankowa S, Kanjee U, Gunalan K, Amaladoss A, Yeo KP, Bob NS, Malleter B, Duraishing MT and Preiser PR (2014) STEVOR is a *Plasmodium falciparum* erythrocyte binding protein that mediates merozoite invasion and rosetting. *Cell Host & Microbe* 16, 81–86.

Nickells M, Hauhart R, Krych M, Subramanian VB, geoGehegan-Barek K, Niang M, Bei AK, Madnani KG, Pelly S, Dankowa S, Kanjee U, Gunalan K, 10 Fiona McQuaid and J. Alexandra Rowe

A complement receptor one alleles have opposing associations with cerebral malaria adherence is associated with disease outcome. *PLoS ONE* 6, e14741.

Opi DH, Swann O, Macharia A, Uyoga S, Band G, Ndila CM, Harrison E, Patel SS, Mehlotra RK, Kastens W, Mgone CS, Kazura JW and Rampengan TH (1991) Cerebral malaria in children. Comparative study. *Clinical and Experimental Immunology* 88, 112–27.3.

Ochola LB, Siddonno BR, Ocholla H, Nkya S, Kimani EN, Williams TN, Makale JO, Liljander A, Urban BC, Pull PC, Szestak T, Marsh K and Craig AG (2011) Specific receptor usage in *Plasmodium falciparum* cytoadherence is associated with disease outcome. *PLoS ONE* 6, e14741.

Pathak V, Colah R and Ghosh K (2016) Correlation between ‘H’ blood group antigen and *Plasmodium falciparum* invasion. *Annals of Hematology* 95, 1067–1075.

Pipitaporn B, Sueblinving T, Dharmkrong-at A and Udomsangretch R (2000) Rosetting of *Plasmodium falciparum* required multiple components of the uninfected erythrocytes. *Asian Pacific Journal of Allergy and Immunology* 18, 29–35.

Rampengan TH (1991) Cerebral malaria in children. Comparative study between heparin, dexamethasone and placebo. *Paediatrica Indonesia* 31, 59–66.

Reddy YN, Siedlecki AM and Francis JM (2017) Breaking down the complement system: a review and update on novel therapies. *Current Opinion in Nephrology and Hypertension* 26, 123–128.

Riback U, Moll K, Albrecht L, Ahmed Ismail H, Normark J, Flaberg E, Szekely L, Hultenby K, Persson KE, Egwang TG and Wahlgren M (2013) Improved in vitro culture of *Plasmodium falciparum* permits establishment of clinical isolates with preserved multiplication, invasion and rosetting phenotypes. *PLoS ONE* 8, e69781.

Ringwald P, Peyron F, Lepers JP, Rabarison P, Rakotomalala C, Razanamparany M, Rabodonirina M, Roux J and Le Bras J (1993) Parasite virulence factors during falciparum malaria: rosetting, cytoadherence and formation of pigment membrane 1 mediating erythrocyte rosetting. *Infection and Immunity* 61, 5198–5204.

Robinson BA, Welch TL and Smith JD (2003) Widespread functional specialization of *Plasmodium falciparum* erythrocyte membrane protein 1 family members to bind CD36 analysed across a parasite genome. *Molecular Microbiology* 47, 1265–1278.

Rowe JA, Opi DH and Williams TN (2009b) Blood groups and malaria: fresh insights into pathogenesis and identification of targets for intervention. *Expert Opinion in Hematology* 2, 480–487.

Rowe JA, Claessens A, Corrigan RA and Arman M (2009a) Adhesion of *Plasmodium falciparum*-infected erythrocytes to human cells: molecular mechanisms and therapeutic implications. *Expert Reviews in Molecular Medicine* 11, e16.

Rowe JA, Opi DH and Williams TN (2009) Blood groups and malaria: fresh insights into pathogenesis and identification of targets for intervention. *Current Opinion in Hematology* 16, 480–487.

Salvaew S, Sritabil J, Piarkasa N, Keayarsa S, Ruengweareyut R, Utaisin A, Sila P, Niramis R, Udomsangretch R, Charunwatthanpa P, Pongponratn E, Pukrittayakamee S, Leitgeb AM, Wahlgren M, Lee SJ, Day NP, White NJ, Donderup AM and Chotivanich K (2017) Effects of sevuparin on rosette formation and cytoadherence of *Plasmodium falciparum* infected erythrocytes. *PLoS ONE* 12, e0172718.

Schmidt CQ, Kennedy AT and Tham WH (2015) More than just immune evasion: Hijacking complement by *Plasmodium falciparum*. *Molecular Immunology* 67, 71–84.

Scholander C, Tretutijer CG, Hultenby K and Wahlgren M (1996) Novel fibrillar structure confers adhesion property to malaria-infected erythrocytes. *Nature Medicine* 2, 204–208.

Scully EJ, Shabani E, Rangel GW, Gruning C, Kanjee U, Clark MA, Chaand M, Kurnit R, Nakamura Y, Ferreira MU and Duraishing MT (2019) Generation of an immortalized erythroid progenitor cell line from peripheral blood: a model system for the functional analysis of *Plasmodium* spp. invasion. *American Journal of Hematology* 94, 963–974.

Semblat JP, Ghunna A, Czajkowski DM, Wallis R, Mitchell DA, Raza A and Rowe JA (2015) Identification of the minimal binding region of a *Plasmodium falciparum* IgM binding PEMP1 domain. *Molecular and Biochemical Parasitology* 201, 76–82.

Silverstein RL and Feilbrabo M (2009) *CD36*, a scavenger receptor involved in immune, metabolism, angiogenesis, and behavior. *Science Signaling* 2, re3.

Sim BK, Chitsne CS, Wasnioska K, Hadley TJ and Miller LH (1994) Receptor and ligand domains for invasion of erythrocytes by *Plasmodium falciparum*. *Science* 264, 1941–1944.

Sinha S, Jha GN, Anand P, Qidwai T, Pati SS, Mohanty S, Mishra SK, Tyagi PK, Sharma SK, Venkatesh V and Habib S (2009) CR1 levels and gene polymorphisms exhibit differential association with falciparum malaria in regions of varying disease endemicity. *Human Immunology* 70, 245–250.

Song B, Fan Y, He W, Zhu D, Niu X, Wang D, Ou Z, Luo M and Sun X (2015) Improved hematopoietic differentiation efficiency of gene-corrupted beta-thalassemia induced pluripotent stem cells by CRISPR/Cas9 system. *Stem Cells and Development* 24, 1053–1065.

Stevenson L, Huda P, Jeppesen L, Claessens A, Corrigan RA and Arman M (2015a) Investigating the function of Fc-specific binding of IgM to *Plasmodium falciparum* erythrocyte membrane protein 1 mediating erythrocyte rosetting. *Cellular Microbiology* 17, 819–831.

Stevenson L, Laursen A, Cowan GJ, Bandoh B, Barfod L, Cavanagh DR, Andersen GR and Hviid L (2015) e2-Macroglobulin can crosslink multiple *Plasmodium falciparum* erythrocyte membrane protein 1 (PEMP1) molecules and may facilitate adhesion of parasitized erythrocytes. *PLoS Pathogens* 11, e1005022.

Stoler-Barak L, Moussion C, Shezen E, Hatzav M, Sixt M and Alon R (2014) Blood vessels pattern heparan sulfate gradients between their apical and basolateral aspects. *PLoS ONE* 9, e85699.
Teeranaipong P, Ohashi J, Patarapotikul J, Kimura R, Nuchnoi P, Hananantachai H, Naka I, Putapatont C, Jongwutwises S and Tokunaga K (2008) A functional single-nucleotide polymorphism in the CR1 promoter region contributes to protection against cerebral malaria. Journal of Infectious Diseases 198, 1880–1891.

Tekete Z and Petros B (2010) The ABO blood group and Plasmodium falciparum malaria in Awash, Metehara and Zitway areas, Ethiopia. Malaria Journal 9, 280.

Tetteh-Quarcoo PB, Schmidt CQ, Tham WH, Haultart R, Mertens HD, Rowe A, Atkinson JP, Cowman AF, Rowe JA and Barlow PN (2012) Lack of evidence from studies of soluble protein fragments that Knops blood group polymorphisms in complement receptor-type 1 are driven by malaria. PLoS ONE 7, e34820.

Thathy V, Moulds JM, Guyah B, Otieno W and Stoute JA (2005) Complement receptor 1 polymorphisms associated with resistance to severe malaria in Kenya. Malaria Journal 4, 54.

Theron M, Cross N, Cawkill P, Bustamante LY and Rayner JC (2018) An in vitro erythrocyte preference assay reveals that Plasmodium falciparum parasites prefer Type O over Type A erythrocytes. Scientific Reports 8, 8133.

Thielen AJF, Zeerleder S and Wouters D (2018) Consequences of dysregulated complement regulators on red blood cells. Blood Reviews 32, 280–288.

Trakarsanga K, Griffiths RE, Wilson MC, Blair A, Satchwell TJ, Meinders M, Cogan N, Kupzipz S, Kurita R, Nakamura Y, Toye AM, Anstee DJ and Frayne J (2017) An immortalized adult human erythrocyte line facilitates sustainable and scalable generation of functional red cells. Nature Communications 8, 14750.

Treutiger CJ, Hedlund I, Helmby H, Carlson J, Jepsen A, Twumasi P, Kwiatoowski D, Greenwood BM and Wahlgren M (1992) Rosette formation in Plasmodium falciparum isolates and anti-rossette activity of sera from Gambians with cerebral or uncomplicated malaria. American Journal of Tropical Medicine and Hygiene 46, 503–510.

Treutiger CJ, Scholander C, Carlson J, McAdam KP, Raynes JG, Falksveden L and Wahlgren M (1999) Rouleaux-forming serum proteins are involved in the rosetting of Plasmodium falciparum-infected erythrocytes. Experimental Parasitology 93, 215–224.

Udomsangpetch R, Wahlin B, Carlson J, Berzins K, Torii M, Aikawa M, Perlmann P and Wahlgren M (1989) Plasmodium falciparum-infected erythrocytes form spontaneous erythrocyte rosettes. Journal of Experimental Medicine 169, 1835–1840.

Udomsangpetch R, Webster HK, Pattanapanyasat K, Pitchayangkanl S and Thaithong S (1992) Cytoadherence characteristics of rosette-forming Plasmodium falciparum. Infection and Immunity 60, 4483–4490.

Udomsangpetch R, Todd J, Carlson J and Greenwood BM (1993) The effects of hemoglobin genotype and ABO blood group on the formation of rosettes by Plasmodium falciparum-infected red blood cells. American Journal of Tropical Medicine and Hygiene 48, 149–153.

Udomsangpetch R, Thanikkul K, Paktrittayakamnee S and White NJ (1995) Rosette formation by Plasmodium vivax. Transactions of the Royal Society of Tropical Medicine and Hygiene 89, 635–637.

Uyoga S, Skorokhod OA, Opiyo M, Orori EN, Williams TN, Arse P and Schwarze E (2012) Transfer of 4-hydroxynonenal from parasitized to non-parasitized erythrocytes in rosettes. Proposed role in severe malaria anemia. British Journal of Haematology 157, 116–124.

van Schravendijk M, Handunnetti S, Barnwell J and Howard R (1992) Normal human erythrocytes express CD36, an adhesion molecule of monocytes, platelets and endothelial cells. Blood 80, 2105–2114.

Vigan-Womas I, Guillotte M, Le Scam C, Igonet S, Petres S, Juillerat A, Badatur C, Nato F, Schneider A, Lavergne A, Contamin H, Tall A, Baril L, Bentley GA and Mercereau-Puijalon O (2008) An in vivo and in vitro model of Plasmodium falciparum rosetting and autoagglutination mediated by varO, a group A var gene encoding a frequent serotype. Infection and Immunity 76, 5565–5580.

Vigan-Womas I, Guillotte M, Juillerat A, Vallieres C, Levit-Bentley A, Tall A, Baril L, Bentley GA and Mercereau-Puijalon O (2011) Allelic diversity of the Plasmodium falciparum erythrocyte membrane protein 1 entails variant-specific red cell surface epitopes. PLoS ONE 6, e18544.

Vigan-Womas I, Guillotte M, Juillerat A, Hessel A, Raynal B, England P, Cohen JH, Bertrand O, Peyrard T, Bentley GA, Lewit-Bentley A and Mercereau-Puijalon O (2012) Structural basis for the ABO blood group-dependence of Plasmodium falciparum rosetting. PLoS Pathogens 8, e1002781.

Vogt AM, Barragan A, Chen Q, Kirofne F, Spillmann D and Wahlgren M (2003) Heparan sulfate on endothelial cells mediates the binding of Plasmodium falciparum-infected erythrocytes via the D81.1 alpha domain of PHEMP1. Blood 101, 2405–2411.

Vogt AM, Winter G, Wahlgren M and Spillmann D (2004) Heparan sulfate identified on human erythrocytes: a Plasmodium falciparum receptor. Biochemical Journal 381, 593–597.

Wahlgren M, Carlson J, Udomsangpetch R and Perlmann P (1989) Why do Plasmodium falciparum-infected erythrocytes form spontaneous erythrocyte rosettes? Parasitology Today 5, 183–185.

Wang CW and Hviid L (2015) Rifsins, rosetting, and red blood cells. Trends in Parasitology 31, 285–286.

Warimwe GM, Fegan G, Musyoki JN, Newton CR, Opiyo M, Githinji G, Andisi C, Menza F, Kitaso B, Marsh K and Bull PC (2012) Prognostic indicators of life-threatening malaria are associated with distinct parasite variant antigens. Science Translational Medicine 4, 129ra45.

White NJ, Turner GD, Day NP and Dondorp AM (2013) Lethal malaria: Marchiafava and Bignami were right. Journal of Infectious Diseases 208, 192–198.

Wilson JG, Murphy EE, Wong WW, Klickstein LB, Weis JH and Fearon DT (1986) Identification of a restriction fragment length polymorphism by a CR1 cDNA that correlates with the number of CR1 on erythrocytes. Journal of Experimental Medicine 164, 50–59.

Wilson MC, Trakarsanga K, Heesom KJ, Cogan N, Green C, Toye AM, Parsons SF, Anstee DJ and Frayne J (2016) Comparison of the proteome of adult and cord erythrocytes, and changes in the proteome following reticulocyte maturation. Molecular and Cellular Proteomics 15, 1938–1946.

Wolofsky KT, Ayi K, Branch DR, Hult AK, Olsson ML, Liles WC, Cserti-Gazdewich CM and Kain KC (2012) ABO blood groups influence macrophage-mediated phagocytosis of Plasmodium falciparum-infected erythrocytes. PLoS Pathogens 8, e1002942.

World Health Organisation (1986) Severe and complicated malaria. World Health Organization Malaria Action Programme. Transactions of the Royal Society of Tropical Medicine and Hygiene 80(suppl.), 3–50.

Yam XY, Niang M, Madmani KG and Preiser PR (2017) Three is a crowd – new insights into rosetting in Plasmodium falciparum. Trends in Parasitology 33, 309–320.