AMPK acts as a molecular trigger to coordinate glutamatergic signals and adaptive behaviours during acute starvation

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Abstract The stress associated with starvation is accompanied by compensatory behaviours that enhance foraging efficiency and increase the probability of encountering food. However, the molecular details of how hunger triggers changes in the activity of neural circuits to elicit these adaptive behavioural outcomes remains to be resolved. We show here that AMP-activated protein kinase (AMPK) regulates neuronal activity to elicit appropriate behavioural outcomes in response to acute starvation, and this effect is mediated by the coordinated modulation of glutamatergic inputs. AMPK targets both the AMPA-type glutamate receptor GLR-1 and the metabotropic glutamate receptor MGL-1 in one of the primary circuits that governs behavioural response to food availability in *C. elegans*. Overall, our study suggests that AMPK acts as a molecular trigger in the specific starvation-sensitive neurons to modulate glutamatergic inputs and to elicit adaptive behavioural outputs in response to acute starvation.

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Introduction

Most organisms are faced with unpredictable fluctuations in their natural environment that often lead to periods of limited food resources. Their ability to adapt to these changes in resource availability is critical for survival and is often a driving force in evolution (Gray et al., 2004; Wang et al., 2005). When resources are scarce, pathways associated with energy conservation at both the organismal and the cellular levels become activated, and these are often complemented by behavioural modifications that simultaneously enhance foraging efficiency (Wang et al., 2005; Ashrafi, 2006).

The mammalian central nervous system (CNS) integrates internal and external cues that signal energy demand and availability and coordinately regulates outputs ranging from energy expenditure to feeding and associated locomotory behaviours (Cone, 2005; Balthasar et al., 2005; Belgardt et al., 2009; Dietrich and Horvath, 2011; Yang et al., 2011; Aponte et al., 2011; Sternson et al., 2013; Dietrich et al., 2015). In mice, the Agouti-related protein (AGRP)- and Pro-opiomelanocortin (POMC)-expressing neurons in the arcuate nucleus of the hypothalamus form a core circuit to regulate food intake and energy expenditure through the modulation of their neuronal activity in response to hormonal signals linked to metabolic status (Cowley et al., 2001; Bewick et al., 2005; Yang et al., 2011). The activity of both of these neuronal populations is mediated by engaging signalling pathways that control the strength and/or plasticity of rapid, excitatory glutamatergic transmission (Bito et al., 2010; Collingridge et al., 2010; Liu et al., 2012), but how energy stress results in changes in neuronal activity to elicit adaptive, or even compulsive behaviours are just now beginning to be elucidated (Dietrich et al., 2015).

The many signalling networks that are triggered throughout the nervous system that mediate the action of small molecules, hormones and nutrients on energy balance are of major interest due to...
their implication in, or treatment of various disorders. One of the key factors of paramount importance for metabolic homeostasis and survival is a highly conserved heterotrimeric protein kinase called AMP-activated protein kinase (AMPK). AMPK is regulated by the ratio of cellular AMP/ATP and by upstream activating kinases (Hardie, 2008; Hardie et al., 2012). It functions as a ‘fuel gauge’ to monitor cellular energy status by inhibiting anabolic pathways and activating catabolic pathways so as to generate sufficient levels of metabolic substrates required to maintain a minimal threshold of basal cellular activities (Hardie, 2008; Hardie et al., 2012; Hardie and Ashford, 2014).

Energy stress has been demonstrated to induce adaptive behaviours in a neuronal AMPK-dependent manner (Lee et al., 2008; Cunningham et al., 2014). Moreover, accumulating evidence has implicated AMPK in the hypothalamic regulation of metabolic rate and food intake behaviour (Kola, 2008; Lopez et al., 2010; Lim et al., 2010; Yang et al., 2011; Schneeberger and Claret, 2012). However, our understanding of how starvation influences adaptive foraging behaviours in an AMPK-dependent manner is still largely unknown, mostly due to the overwhelming complexity of the response in higher animals (Dietrich et al., 2015).

In C. elegans these foraging behaviours are comparatively simple, consisting of a series of forward or backward movements, specific turns, or changes in direction (Gray et al., 2005; Piggott et al., 2011; Chen et al., 2013; Hendricks, 2015). Food availability has been demonstrated to affect various aspects of these key elements in C. elegans locomotion (Sawin et al., 2000; Gray et al., 2005; Chalasani et al., 2007; Flavell et al., 2013). In the absence of food, well-fed animals reverse frequently, a behavioural pattern that reflects a sensory memory of food that is expressed by the navigation circuit and results in efficient exploration of a limited area. In contrast, starvation suppresses reversals and induces forward movement (runs) that allow animals to explore distal areas; a strategy that is referred to as dispersal behaviour or alternatively, distal exploration (Gray et al., 2005).

The simplicity of the neural circuits and locomotory behaviours in conjunction with its amenability to genetic manipulation makes C. elegans an ideal model to investigate the mechanisms through which AMPK regulates neuronal activity and adaptive locomotory behaviour (searching for food) in
response to hunger. Unlike most other organisms studied to date, *C. elegans* mutants that lack all AMPK signalling are viable, but show clear phenotypes when subjected to energy stress (Narbonne and Roy, 2006; 2009). Therefore, using calcium imaging, cell type–specific optogenetic techniques, and classic genetic analysis, we identified and characterized the neural circuit in which AMPK functions as a molecular switch. This circuit includes the AIB and AIY interneurons; two neurons that form one of the primary circuits that dictate appropriate food- and odour-evoked behaviours (Gray et al., 2005; Chalasani et al., 2007). We discovered that AMPK regulates forward locomotion through two distinct mechanisms to ultimately ensure that adaptive foraging behaviours are appropriately triggered during periods of starvation. In the AIB interneurons AMPK regulates the abundance of the α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor GLR-1 in the postsynaptic elements, presumably by phosphorylation of serine 907 and 924 resulting in changes in synaptic strength and specific behavioural outputs. In addition, we also demonstrate that AMPK modulates a key metabotropic glutamate receptor called MGL-1 in the AIY interneuron at both mRNA and protein levels leading to an increase in AIY neuronal activity in starved animals. Together, our results indicate that AMPK acts as a starvation-inducible molecular trigger in the nervous system that modulates glutamatergic neuronal activity by at least two distinct mechanisms to modify behavioural outcomes in response to energy stress.

**Results**

**Neuronal AMPK signalling triggers distal exploratory behaviour in starved animals**

AMPK has been implicated in the regulation of feeding behaviours in higher animals (Minokoshi et al., 2004; Yang et al., 2011) although its essential role in development and cellular homeostasis has made it very difficult to study its role outside this closed circuit. Accordingly, the identification of the neuronal targets of this protein kinase that may be important for these behaviours has been virtually impossible to interrogate. Fortunately, *C. elegans* mutants that completely lack AMPK signalling are viable, thus providing us with an opportunity to investigate how AMPK regulates characteristic foraging behaviours in response to acute periods of starvation.

We and others found that removal of aak-2, the more prominent of two catalytic subunits of AMPK present in *C. elegans*, disrupts most cellular AMPK signalling and affects various aspects of *C. elegans* foraging behaviour (Lee et al., 2008; Cunningham et al., 2014). More specifically, aak-2 mutants exhibit a behavioural profile in response to acute starvation that is more typical of satiated animals, where animals tend to explore locally rather than foraging in more distant locations. This is reflected by the frequency of body bends; which is considered as a representative readout for locomotory movement away from a nutrient-depleted environment in search of new resources, and reversals; a behaviour most often associated with local exploration (Gray et al., 2005). Both of these behaviours are affected in starved aak-2 mutants (Figure 1A). These behaviours appear to be regulated predominantly by aak-2 activity since removal of both catalytic subunits and hence eliminating all AMPK signalling (aak(0)) only modestly enhanced the phenotype of aak-2 mutants in all our behavioural assays suggesting that aak-2 mutants recapitulate a severe loss of AMPK function.

Since aak-2 is a major regulator of foraging behaviour in *C. elegans*, we transferred mid-fourth larval stage (L4) aak-2 mutant larvae to bacteria-free plates and monitored their movements for 20 hr to delineate sub-behaviours underlying aak-2 deficiency. We found that starved aak-2 mutants show defects in their rate of forward locomotion measured in the absence of food, which is compounded by their inability to appropriately suppress reversal behaviour during periods of more prolonged starvation (Figure 1B). Because we observe changes in each of these AMPK-dependent parameters in starved animals, and given the antagonistic relationship between forward and backward locomotion where increased reversal frequency affects the onset and/or duration of forward movement (Burbea et al., 2002; Juo et al., 2007), we chose to assess both behaviours in starved animals throughout our study.

The aak-2-dependent foraging defects we observed in starved animals are not likely to be due to a more general role of AMPK in the regulation of appropriate motor neuron development or function since AMPK mutants were comparable to wild type controls when reversal frequency and forward locomotion were quantified during conditions when food was abundant (Figure 1—figure
Figure 1. Neuronal AMPK regulates distal exploratory behaviour in starved animals. (A) Starved aak-2 mutants display defective transition from local to distal exploration indicated by (i) decreased forward locomotion and (ii) increased reversal rate during acute bouts of starvation. These phenotypes are not dependent on aak-1 (n>40), (one-way ANOVA **p<0.001, ***p<0.0001). (B) AMPK mutants display persistent decreased forward locomotion rate (i) and increased reversal frequency (ii) during periods of acute starvation (n>10), Error bars represent ± SEM (Student’s t-test, *p<0.05, **p<0.001, ***p<0.0001). (C) aak-2 reconstitution within the nervous system using the pan-neural [Punc-119::aak-2] transgene rescues the defective distal exploratory behaviour in starved aak-2 mutants while its reconstitution within the body wall muscle using [Punc-54::aak-2] transgene does not improve the distal exploratory behaviour of aak-2 mutants (n>15), (one-way ANOVA *p<0.05, **p<0.001). (D,E,F) aak-2 is highly expressed throughout the nervous system. Scale bars are 40 μm. In the box and whisker plots (A,C) the central line is the median, the edges of the box are the 25th and 75th percentiles, and the whiskers extend to the most extreme data points.

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The following source data and figure supplements are available for figure 1:

**Source data 1.** Locomotory behaviour in AMPK mutant animals.
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**Source data 2.** Food and stimuli-related behaviours in AMPK mutants.
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**Source data 3.** Locomotory behaviour upon depletion of aak-2 with 1535 RNAi.
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**Figure supplement 1.** AMPK function is not required for other food-related behaviours rather than distal exploratory behaviour.
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**Figure supplement 2.** Reduced forward locomotion rate in aak-2 mutants is not a consequence of developmental defect in their nervous system.
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Moreover, aak-2 mutants displayed a normal fleeing speed in response to a mechanical stimulus that was applied to the posterior (Figure 1-figure supplement 1D). In addition, we observed that aak-2 (RNAi) performed at the late L2 stage, when the majority of neurons have already been generated, resulted in a reduction in forward locomotion compounded with an increase in reversal frequency in starved animals. This suggests that the reduced locomotory speed is unlikely to be the consequence of a general defect in motor function or neuromuscular development (Figure 1-figure supplement 2).

To discern whether aak-2 is specifically required in neurons, muscle, or both tissues to modulate distal exploratory behaviours we used tissue-specific promoters to drive aak-2 expression and test its ability to correct the behavioural defects typical of AMPK mutants. While an aak-2 cDNA (Narbonne and Roy, 2009) expressed under the control of a pan neuronal promoter [Punc-119::aak-2] rescued the distal exploratory defect of aak-2 mutants, aak-2 mutants continued to exhibit prolonged local exploration when the same aak-2 cDNA was reconstituted in body wall muscle using [Punc-54::aak-2] transgene (Figure 1C). Consistent with these results, we observed that a rescuing translational fusion reporter [Paak-2::aak-2::GFP] was broadly expressed throughout the nervous system (Figure 1D,E,F). These data are consistent with previous observations indicating that AMPK is required in the nervous system to regulate locomotory behaviour in response to food availability (Lee et al., 2008; Cunningham et al., 2014).

AMPK is not required for all starvation-related behaviours

The ability of AMPK mutants to appropriately respond to starvation could result from a global AMPK-dependent defect in their nervous system rendering them incapable of responding to multiple cues in addition to starvation. However, this seems unlikely since two other food-related behaviours namely, basal slowing response and enhanced slowing response, which are triggered in well-fed and starved animals that are reintroduced to food, respectively, remain unaffected in AMPK mutants. In its natural habitat, a well-fed animal would be more likely to risk exploring distant locations for high quality food sources, whereas a starved animal would be less likely to stray far from a recently discovered vital food supply. These paradigms regulate behavioural plasticity in response to starvation in C. elegans and are mediated by distinct dopaminergic and serotonergic signalling (Sawin et al., 2000). When we re-introduced well-fed or starved aak-2 mutants to food we did not detect any significant difference in either the basal slowing response or the enhanced slowing response (Figure 1-figure supplement 1B,C) suggesting that AMPK is not involved in the behavioural plasticity that occurs after re-introduction of animals to food and that the neural circuitry that mediates such behavioural plasticity is presumably intact and functional in animals that lack AMPK.

AAK-2 is required in both the AIY and the AIB interneurons to mediate the transition from local to distal exploration in response to starvation

Since pan neuronal aak-2 expression rescued the distal exploration defect of aak-2 mutants, we next sought to identify the individual neurons that require aak-2 activity to trigger this behaviour. The neural circuitry that dictates these simple behavioural outcomes has been well described (White et al., 1976; 1986; Chalfie et al., 1985; Wicks et al., 1996; Gray et al., 2005). In particular, five pairs of command interneurons are required for the control of coordinated movement. The PVC and AVB interneurons are primarily required for the initiation of forward movement, while the AVA, AVD, and AVE interneurons control reversals (White et al., 1976; Chalfie et al., 1985; Wicks et al., 1996; Gray et al., 2005). Sensory inputs and serotonergic signalling together contribute to the behavioural changes that occur following removal from food (Gray et al., 2005). Moreover, several studies have underscored a critical role for both the AIY and the AIB interneurons, which are independently required to suppress and enhance reversals, respectively, resulting in longer or shorter durations of forward movement (Tsailik and Hobert, 2003; Wakabayashi et al., 2004; Gray et al., 2005; Chalasani et al., 2007; Luo et al., 2014). The RIM interneurons have also been determined as regulators of reversal frequency (Gray et al., 2005; Gordus et al., 2015).

To map the precise sites of AMPK function in its regulation of distal exploratory behaviour, we introduced aak-2 in different subsets of neurons using neuronal sub-type-specific promoters: tph-1 (serotonergic neurons) (Cunningham et al., 2012), glr-1 (command interneurons) (Zheng et al., 1999), che-2 (chemosensory neurons) (Gray et al., 2005), ttx-3 (AIY) (Chalasani et al., 2007), npr-9
(AIB) (Piggott et al., 2011), tdc-1 (RIM) (Cunningham et al., 2012) and rig-3 (AVA) (Marvin et al., 2013). Reconstitution of AMPK (aak-2) using the glr-1, tdc-1, ttx-3, npr-9 promoters was sufficient to partially rescue the defective distal exploration typical of aak-2 mutants suggesting that AMPK functions within these neurons to phosphorylate targets involved in regulating these behavioural responses. Conversely, introducing AMPK (aak-2) in the sensory neurons, the serotonergic neurons, or the command interneurons involved in triggering reversals failed to correct the defect (Figure 2A). As AMPK expression within the GLR-1-expressing neurons partially rescued the locomotory defect of aak-2 mutants, it is very likely that AMPK may be independently required in the GLR-1-expressing neurons AIB and RIM, although this does not exclude a role for aak-2 in other GLR-1-expressing neurons.

The A1Y and AIB interneurons receive synapses from the AWC sensory neurons and act in parallel to suppress and enhance reversals, respectively, in response to food availability (Figure 2—figure supplement 1A,B) (Chalasani et al., 2007). The AIB interneurons relay their signals to the RIM and AVA, which in turn send their outputs to the muscles (Figure 2—figure supplement 1C) (Gordus et al., 2015). As an AMPK expression within the A1Y, AIB and RIM partially restored the defective distal exploration of starved aak-2 mutants, it is likely that AMPK functions in multiple neurons in a parallel circuit to regulate locomotory behaviour in response to starvation.

When we expressed aak-2 in both the A1Y and the AIB interneurons, we completely restored the defective distal exploration of aak-2 mutants, while expression of aak-2 within the RIM and the AIB interneurons, or in RIM and A1Y, did not improve the distal exploration defect of starved aak-2 mutants beyond the rescue observed in AMPK mutant animals expressing aak-2 within the AIB or the A1Y interneurons alone (Figure 2B). The RIM neurons form chemical and electrical synapses with AIB and A1Y, respectively (White et al., 1986). Of note, it has been shown that chemical synapses from the RIM neurons enhance the variability in response to odour in the AWC-AIB-RIM olfactory circuit (Gordus et al., 2015). Therefore, aak-2 expression within the RIM neuron might influence this variability, whereby the expression of aak-2 within the AIB and RIM neurons biases the circuit towards either a more probabilistic or reliable output. Taken together, our data highlight a critical role of AMPK in the A1Y and AIB interneurons to regulate the appropriate transition from local to distal exploration in animals subjected to acute starvation.

**AMPK functions within the AIB and A1Y interneurons to integrate sensory signals in response to starvation**

AMPK mutants demonstrate reduced forward movement accompanied by an increased frequency of reversals. If these defects disrupt distal exploratory foraging behaviour then starved aak-2 mutants could also be defective in finding food at a distance. We therefore placed starved animals on plates with a point food source (E. coli) and monitored the time required for each animal to encounter the bacteria. We noted that aak-2 mutants were substantially less efficient than wild type controls in their ability to track to the food source (Figure 2C). However, from this assay alone, we were unable to distinguish if this defect is a consequence of a deficiency in their ability to sense food or to integrate the detection of nutritional signals to appropriately modify locomotory behaviour since we also observed a significant defect in the ability of AMPK mutants to chemotaxis toward the attractant isoamyl alcohol (IAA) (Figure 2D). However, since the expression of aak-2 within the sensory neurons failed to restore the defective distal exploratory behaviour in aak-2 mutants, while this defect was fully rescued by driving aak-2 expression specifically within the AIB and A1Y interneurons, we conclude that AMPK is critical for the integration of sensory cues and not with chemosensation per se (Figure 2D). Taken together, our results indicate that AMPK is required in the AIB and A1Y interneurons to trigger appropriate exploratory behaviour in response to acute starvation.

**AMPK functions upstream of mgl-1, glr-1 and eat-4 to modulate distal exploration**

To position how and where AMPK functions within the A1Y and AIB interneurons to modulate the transition between local and distal exploration, we performed optogenetic experiments to manipulate AIB and A1Y activity in starved aak-2 mutants (Kocabas et al., 2012). ChR2-mediated depolarization of the A1Y interneurons in aak-2 mutant animals resulted in a significant decrease in reversal frequency that was persistent throughout the illumination time. In contrast, A1Y activation in WT
animals slightly altered the forward locomotion and reversal rate which is consistent with previous studies demonstrating the activation of the AIY interneurons in starved animals (Gray et al., 2005; Flavell et al., 2013) (Figure 3A, Figure 4—figure supplement 2A).
Figure 3. AMPK regulates glutamatergic signalling. (A, B) ChR2-mediated activation of AIY interneurons (A), or ARCH-mediated silencing of AIB interneurons (B) in starved aak-2 mutants grown in the presence of all-trans retinal (ATR) induces distal exploration by suppressing reversals and inducing forward locomotion (n>15). Error bars represent ± SEM (two-way ANOVA *p<0.05, ***p<0.0001). (C, D, E) Compromised eat-4, glr-1 or mgl-1 function rescues the defective exploratory behaviour of starved aak-2 mutants by inducing forward locomotion (i) and suppressing reversals (ii) (n>20), (one-way ANOVA *p<0.05, **p<0.001, ***p<0.0001). In the box and whisker plots (C, D, E) the central line is the median, the edges of the box are the 25th and 75th percentiles, and the whiskers extend to the most extreme data points.

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Source data 1. Optogenetics and epistatic analysis.
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Source data 2. The analysis of locomotory behaviour of potential AMPK phospho targets.
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Source data 3. Potential AMPK phosphorylation targets expressed within the AIB and AIY interneurons.
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Figure supplement 1. glr-1, eat-4 and mgl-1 are epistatic to aak-2.
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In a complementary experiment, we assessed the behaviour of starved aak-2 mutants following inactivation of AIB induced by the light-gated chloride pump archaerhodopsin-3 (Arch) (Kocabas et al., 2012). The Arch-induced hyperpolarization of AIB was sufficient to suppress reversals in the aak-2 mutants, while also promoting normal distal exploratory behaviour during starvation. In contrast, Arch-induced AIB inactivation in WT animals altered their locomotory behaviour slightly which can be explained by inactivation of AIB during acute bouts of starvation as the locomotory behaviour of starved animals has been shown to remain unchanged upon AIB ablation (Gray et al., 2005) (Figure 3B). We interpret these observations to indicate that although AMPK activity is required within the AIB and AIY interneurons for appropriate distal exploration in response to acute starvation, it does so without negatively impacting normal neurotransmission, neuronal development or connectivity within the interneurons.

Having identified the interneurons in which aak-2 is required to suppress reversals, promote forward locomotion, and eventually trigger distal exploration, we then set out to identify AMPK targets within these interneurons that mediate its effects in response to starvation. Using bioinformatic tools we scanned the C. elegans proteome for proteins that possessed consensus AMPK phosphorylation sites and were known to be expressed in the AIB or AIY interneurons, providing us with a list of potential neuronal phosphorylation targets of AMPK (Figure 3—source data 3).

Among the candidates that we identified through our bioinformatic analysis GLR-1, EAT-4, and MGL-1 emerged as highly predicted phosphorylation targets of AMPK (Figure 3C,D,E, Figure 3—figure supplement 1A). eat-4 encodes an ortholog of a mammalian brain-specific sodium-dependent inorganic phosphate co-transporter I (BNPI), and is required for glutamatergic neurotransmission in the AIB (Lee et al., 1999; Piggott et al., 2011). Interestingly, we found that eat-4 is epistatic to AMPK function during the response to acute starvation; single eat-4 mutants behave like starved WT animals by decreasing reversal frequency in contrast to the aak-2 mutants that exhibit an increased number of reversals. The aak-2; eat-4 double mutants behave more like starved WT (or eat-4) animals than aak-2 mutants (Figure 3C) indicating that eat-4 acts downstream of, or in parallel to AMPK, in the genetic pathway controlling this starvation-induced behaviour.

The AMPA-type GluR GLR-1 is expressed in both the command interneurons and in some motor neurons, and is implicated in memory formation and the behavioural responses to light nose touch and to sensory cues such as food (Hart et al., 1995; Rose et al., 2003; Chalasani et al., 2007). Moreover, GLR-1 function is required for glutamatergic activity in the AIB (Chalasani et al., 2007). Similar to eat-4, mutations in glr-1 suppress the defective distal exploratory behaviour of AMPK mutants suggesting that like eat-4, glr-1 is epistatic to aak-2 (Figure 3D). Taken together, these results indicate that blocking the glutamatergic inputs to the AIB through the elimination of glr-1, or blocking its glutamatergic neurotransmission by disrupting eat-4, can compensate for the behavioural defects in starved aak-2 mutants. Our data are therefore consistent with aak-2 acting upstream of, or in parallel to these two glutamatergic effectors to modulate neuronal activity in the AIB interneuron during periods of acute starvation.

As initially determined in mammals, the metabotropic glutamate receptors (mGluRs) also have an important neuromodulatory role in glutamatergic transmission within the C. elegans nervous system (Dillon et al., 2006, 2015). mgl-1 encodes one of the three metabotropic glutamate receptors in C. elegans and is expressed throughout the AIY interneurons. Based on sequence analysis, MGL-1 is a group II metabotropic glutamate receptor that acts as both pre- and postsynaptic detectors of glutamate to reduce neuronal excitation at least partly by inhibition of adenylyl cyclase activity (Dillon et al., 2015; Niswender and Conn, 2010). Previous studies have identified mgl-1 as an essential component for the appropriate regulation of fat accumulation (Greer et al., 2008) and the starvation responses mediated by AIY (Kang and Avery, 2009). Our genetic analyses indicate that mutations in mgl-1 could also significantly, but not completely, suppress the prolonged local exploration typical of aak-2 mutants whereby a compromise of mgl-1 in the aak-2 background results in reduced reversals and increased distal runs (Figure 3E). These results suggest that AMPK may negatively regulate MGL-1 activity in the AIY in response to starvation.

Our data are consistent with AMPK acting as a potential modulator of glutamatergic inputs in both the AIB and the AIY, but it cannot distinguish whether AMPK affects glutamatergic inputs in a subset of glutamatergic neurons (i.e., the AIB and AIY) or in all glutamatergic neurons. Furthermore, if AMPK does have a specific role in the AIB and the AIY, then what determines its functionality in this subset of AMPK-sensitive neurons? To test if AMPK affects other mediators of glutamatergic
inputs we assessed whether AMPK compromise could modify the phenotypes associated with: nmr-1, which encodes a NMDA-type ionotropic glutamate receptor subunit; the glutamate-gated chloride channels avr-14 and mgl-2 which encodes a group I metabotropic glutamate receptor (Chalasani et al., 2007; Piggott et al., 2011; Dillon et al., 2015). Double mutant combinations with aak-2 did not modify any of the aak-2 phenotypes we tested (Figure 3—figure supplement 1B) suggesting that aak-2 does not affect glutamatergic transmission generally, but rather it seems to only affect a subset of glutamatergic neurons critical for starvation-dependent functions and/or behaviours.

**Parallel and opposing function of AMPK in the modulation of AIB and AIY outputs**

Given the epistatic relationship between aak-2 and genes involved in glutamatergic signalling we wanted to clarify if AMPK regulates the AIB and AIY neural activity through these targets. We monitored calcium transients using the genetically-encoded calcium sensor G-CaMP individually in both interneurons as a proxy for neuronal activity (Figure 4A,B,C,D,E,F). We observed a decrease in AIY activity in starved aak-2 mutants that was rescued by AIY-specific aak-2 expression, consistent with aak-2 regulating AIY activity/response during starvation (Figure 4A,B,C,G, Figure 4—figure supplement 1A). Previous studies have shown that calcium spikes in the AIY interneurons correlate positively with forward runs and negatively with the initiation of reversals with gradual increases preceding forward run initiation (Flavell et al., 2013; Luo et al., 2014). Our results suggest that the decreased AIY activity in starved aak-2 mutants at least partially contributes to their defective distal exploratory behaviour. To further investigate whether the increased reversal rate correlates with decreased AIY activity in starved aak-2 mutants, we monitored the calcium levels in the AIY interneuron of freely behaving animals. The AIY calcium levels were consistently higher upon the termination of reversals, and during the subsequent forward runs with gradual increase preceding forward run initiation suggesting that the calcium peaks in the AIY correlate with the suppression of reversals and the observed acute increase in forward speed (Figure 4—figure supplement 2A). Furthermore, consistent with our previous results, we found that starved aak-2 mutants display increased reversal frequency and shorter durations of forward locomotion, both of which correlated with decreased calcium influx and associated AIY activity (Figure 4—figure supplement 2C,E,F).

The epistatic relationship between mgl-1 and aak-2 in distal exploration was further corroborated by the total integrated fluorescence intensity demonstrated by aak-2; mgl-1 animals. These readings resemble both starved WT and/or mgl-1 mutants, reinforcing our data indicating that mgl-1 is epistatic to aak-2 function in the AIY interneuron during acute starvation, where MGL-1 potentially acts as a postsynaptic receptor in the AIY (Figure 4H—figure supplement 1A). AIY-ablated animals are defective in transitioning between local and distal exploration (Gray et al., 2005), these results reveal an additional, albeit critical, role of AMPK within the AIY interneurons to modulate AIY neuronal activity autonomously, thus contributing to the behavioural transition associated with the decision to explore more distal environments when animals are subjected to starvation.

Unlike the AIY interneurons, activity in AIB increased in starved aak-2 mutants and could be rescued by AIB-specific aak-2 expression (Figure 4D,E,F,I). Calcium peaks in the AIB have been shown to correlate with reversal frequency (Piggott et al., 2011; Gordus et al., 2015), but during our recording the animals are restrained therefore we presume that each calcium peak is representative of a reversal attempt. Given the increased calcium levels in the AIB interneurons in starved aak-2 mutants, our data indirectly suggest that the increased reversal frequency and the abnormally prolonged local exploration is at least partly a consequence of increased AIB neuronal activity. To further examine if the increased AIB activity in starved aak-2 mutants correlates with their increased reversal frequency, we examined the behaviour of freely moving animals during acute starvation. The calcium peaks in the AIB were well correlated with reversal frequency, further suggesting that the increased reversal frequency in starved aak-2 mutants is at least partially due to the increased AIB activity (Figure 4—figure supplement 2B,D,G,H).

The genetic requirement for aak-2 in the AIB interneurons to modulate neuronal activity, along with its epistatic relationship with eat-4 and glr-1 in the modulation of locomotory behaviour, are consistent with at least two models: During acute starvation aak-2 might regulate excitatory synaptic inputs onto the AIB interneurons through modulation of glr-1 activity, or aak-2 might affect glutamatergic neurotransmission from the AIB interneurons through, or in parallel with eat-4. To distinguish...
Figure 4. AMPK regulates AIB and AIY neuronal activity. (A,B,C,D,E,F) A sample ΔF/F₀ plot showing spontaneous changes in the AIY and AIB neuronal activity in starved individuals during the course of a 65 and 120 s imaging window, respectively. These plots are the basis of the data in G, H, I and J. The heatmaps show normalized G-CaMP1 and G-CaMP3 traces in AIY and AIB, respectively in multiple animals. (G) The average total integrated

Figure 4 continued on next page

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fluorescence intensity \((\Delta F/F^0)\) over the course of a 65 s window. A1Y neuronal activity is reduced in starved aak-2 mutants and this reduction in the A1Y neuronal activity is restored by targeted expression of aak-2 within the A1Y interneurons using the A1Y-specific [Ptx-3::aak-2] transgene (n>10). (H) Removal of mgl-1 in aak-2 mutants rescues the reduced A1Y neuronal activity observed in starved aak-2 mutants (n>10). (I) The average total integrated fluorescence intensity \((\Delta F/F^0)\) in AIB is increased upon removal of aak-2 in starved animals, and this increase is reversed by expression of aak-2 within the AIB using the AIB-specific [Pnpr-9::aak-2] transgene (n>10). (J) Removal of glr-1, but not eat-4, restores the increased AIB neuronal activity in starved aak-2 mutants (n>10). Error bars represent \(\pm SEM\) (one-way ANOVA **p<0.0001).

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The following source data and figure supplements are available for figure 4:

**Source data 1.** Calcium imaging in the AIB and A1Y interneurons in WT, aak-2, mgl-1, glr-1 and eat-4 and double mutants.
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**Source data 2.** Calcium imaging in freely WT and aak-2 behaving animals.
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**Figure supplement 1.** Heat maps showing A1Y and AIB spontaneous neuronal activity in starved animals.
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**Figure supplement 2.** A1Y and AIB spontaneous neuronal activity in starved freely behaving animals.
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between these possibilities, we monitored calcium transients in aak-2; glr-1 and aak-2; [Pnpr-9::eat-4 (RNAi)] double mutants. Of note, we observed that although blocking the AIB neurotransmission in aak-2; [Pnpr-9::eat-4 (RNAi)] animals resulted in normal distal exploratory behaviour during starvation (Figure 3C), the total integrated signal was significantly higher compared to both WT and [Pnpr-9::eat-4 (RNAi)] animals (Figure 4J, Figure 4—figure supplement 1B). This suggests that aak-2 acts independently of eat-4 to modulate neural activity in the AIB. Taken together, our genetic analyses indicate that AMPK is required within the A1Y and the AIB interneurons to modulate their neuronal activity through its direct or indirect regulation of the glutamatergic receptors GLR-1 and MGL-1 during starvation-induced exploratory behaviour.

**AMPK controls GLR-1 abundance in the neuronal postsynaptic membranes**

Increased GLR-1-mediated glutamatergic inputs have been shown to bias the locomotory circuit toward reversals (Zheng et al., 1999). Consistent with this, overexpression of GLR-1 specifically within the A1B interneurons increases reversal frequency and turning rate in the absence of food (Chalasani et al., 2007). Because the abundance of GLR-1 within the AIB influences reversal behaviour, we tested whether the increased reversal frequency in aak-2 mutants is a consequence of changes in GLR-1 levels. We quantified glr-1 mRNA levels in WT and aak-2 mutants under well-fed and starved conditions and we did not detect any differences (Figure 5—figure supplement 1A). However, we did notice that total GLR-1 protein levels in starved WT animals were reduced and this effect was less pronounced in starved aak-2 mutants (Figure 5—figure supplement 1B) suggesting that AMPK may modulate GLR-1 protein levels in starved animals either at the level of synthesis or stability.

Studies have demonstrated that disruption of GLR-1 endocytosis results in increased GLR-1 abundance in the postsynaptic elements which results in increased synaptic strength and reversal frequency (Burbea et al., 2002; Juo et al., 2007). Given the epistatic relationship between aak-2 and glr-1 we next asked if the increased reversal frequency in starved aak-2 mutants occurs as a result of decreased GLR-1 turnover linked to a change in its post-translational regulation due to the absence of AMPK. To examine this possibility, we used a previously described chimaeric receptor tagged with the green fluorescent protein (GLR-1::GFP) that localizes to discrete punctate structures and that can be used to visualize central glutamatergic synapses in living animals (Burbea et al., 2002). We measured the density of GLR-1::GFP-containing puncta in the AIB neuronal process in a strain that expresses GLR-1::GFP driven by the AIB-specific promoter npr-9. Notably, we observed that the GLR-1 levels in the AIB neuronal process in the nerve ring where the AIB interneurons form their synapses (Figure 5A) was significantly greater in aak-2 mutants than WT animals during starvation (Figure 5B,C). Moreover, this difference could be rescued by introducing AMPK specifically within
the AIB interneurons (Figure 5C), highlighting the importance of AMPK in modulating GLR-1 abundance within the AIB interneurons.

Our data suggest that AMPK regulates the GLR-1 abundance in the AIB interneurons to potentially modulate synaptic inputs and consequently affect reversal behaviour. But aak-2 mutants have additional behavioural defects that may also manifest due to inappropriate glutamatergic signalling.

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Figure 5. AMPK regulates GLR-1 abundance in the AIB neuronal process and the ventral nerve cord. (A) Schematic representation showing AIB interneurons that extend their processes into the nerve ring. (B,C) Representative images of [Pnpr-9::glr-1::gfp] in the AIB neuronal processes in starved WT and aak-2 mutants (B). GLR-1::GFP level is significantly increased in the AIB neuronal processes in the aak-2 mutants compared to WT animals under starvation condition. Targeted expression of aak-2 within the AIB interneurons rescues the increased GLR-1::GFP level in starved aak-2 mutants (C). At least 3 [Pnpr-9::glr-1::gfp] transgenic lines were separately examined for this experiment (n>20), Error bars represent ± SEM (one-way ANOVA ***p<0.0001). Scale bars are 10 µm. (D,E,F) Representative images of [Pglr-1::glr-1::gfp] in the ventral nerve cord (D). GLR-1::GFP level (E) and puncta width (F) are significantly increased in the ventral nerve cord of starved aak-2 mutants compared to WT animals. (n>20), Error bars represent ± SEM (Student’s 2-tailed t test ***p<0.0001). Scale bars are 10 µm.

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The following source data and figure supplement are available for figure 5:

Source data 1. Measurement of GLR-1 abundance in the AIB neuronal process and ventral nerve cord of WT and aak-2 mutants.
DOI: 10.7554/eLife.16349.023

Source data 2. The extent of GLR-1 and EAT-4 colocalization in WT and aak-2 mutants.
DOI: 10.7554/eLife.16349.024

Figure supplement 1. AMPK regulates GLR-1 abundance by affecting steady state protein levels.
DOI: 10.7554/eLife.16349.025
AMPK acts with UNC-11/AP180 to regulate GLR-1 levels within the postsynaptic elements and consequently modulate synaptic strength

Mammalian AMPA receptors are removed from postsynaptic membranes by clathrin-mediated endocytosis (Carroll et al., 1999; Man et al., 2000). In C. elegans, the ubiquitylation of conserved cytoplasmic lysine residues on GLR-1 constitutes an endocytic signal to remove the receptors from postsynaptic elements which subsequently targets them for degradation; a process that is mediated by the unc-11/AP180 clathrin adaptin protein (Burbea et al., 2002). To further investigate if AMPK modulates GLR-1 abundance by affecting endocytosis in response to starvation, we measured GLR-1 abundance in unc-11 mutants that are unable to internalize GLR-1. The GLR-1 levels in aak-2; unc-11 double mutants were indistinguishable from those found in either unc-11 single mutants (Figure 6A, B, C). Similarly, the increased GLR-1 levels and size/width of GLR-1 puncta measured in the aak-2 mutants was not further enhanced when all four cytoplasmic lysine residues required for GLR-1 ubiquitylation and endocytosis were mutated to arginine [GLR-1(4KR)::GFP], further suggesting that aak-2 acts in a linear pathway with an unc-11/clathrin-mediated mechanism to regulate GLR-1 endocytosis (Figure 6A, B, C).

Activation of AMPK during periods of starvation could result in the regulation of GLR-1 abundance by its direct phosphorylation through its consensus phosphorylation sites. To determine the importance of these sites we used the [Pnpr-9::glr-1::gfp] fusion protein as a template to generate a GLR-1::GFP variant with non-phosphorylatable AMPK sites. The cytoplasmic domain of GluR has already been described as an important phosphoregulatory target, mostly by kinases such as PKA and PKC (Roche et al., 1996). Since two conserved AMPK phosphorylation sites were identified within the cytoplasmic domain of GLR-1 (Figure 5—figure supplement 1f), we mutated both S907 and S924 to non-phosphorylatable alanine residues to evaluate how these sites contribute to the stability of GLR-1 within the AIB neuronal process during periods of starvation. Wild type animals bearing the compound (S907A, S924A) variant transgene demonstrated defective exploratory behaviour that was comparable to aak-2 mutants (Figure 6D). To further examine if the GLR-1 receptors that accumulate in AMPK mutants act as functional synaptic receptors we examined the locomotory behaviour of eat-4; glr-1 double mutants expressing the non-phosphorylatable variant (S907A, S924A). We noticed that the increased reversal frequency rate accompanied by reduced forward locomotion in animals expressing the (S907A, S924A) variant was largely suppressed in the absence of eat-4 indicating that the compromise of AMPK signalling results in an increased abundance of functional synaptic GLR-1 receptors (Figure 6D). Furthermore, the GLR-1 (S907A, S924A) variant accumulated in the AIB neuronal process and this accumulation was not enhanced upon removal of unc-11 (Figure 6E), further suggesting that AMPK regulates GLR-1 abundance directly by phosphorylating the protein to mediate UNC-11/AP180-dependent GLR-1 endocytosis in response to starvation.
Elevated glutamatergic transmission within the command interneurons is associated with a hyper-reversal phenotype and a dramatic decrease in the duration of forward movement (Zheng et al., 1999). It would therefore be intuitive that an increase in synaptic strength, or input, would result in a consequential increase in reversal frequency. Although we were unable to directly examine the role of AMPK in the regulation of synaptic strength, taken together, our data suggest that AMPK

Figure 6. AMPK directly regulates GLR-1 abundance through endocytic pathway. (A) Representative images of [Pglr-1::glr-1::gfp] in the ventral nerve cord in starved WT, aak-2, unc-11 and aak-2; unc-11 mutants. Scale bars are 10 μm. (B,C) Similar to starved aak-2 mutants, GLR-1::GFP level (B) and puncta width (C) are increased upon removal of unc-17 or mutation of four lysines required for GLR-1 ubiquitination and endocytosis [Pglr-1::glr-1(4KR)::gfp]. Increased GLR-1::GFP level and enhanced puncta width are not further enhanced upon depletion of aak-2 in GLR-1 endocytosis defective mutants (n>25), Error bars represent ± SEM (one-way ANOVA ***p<0.0001). (D) Mutating AMPK phosphorylation sites S907 and S924 to non-phosphorylatable alanine residues present in the cytoplasmic domain of GLR-1 (S907A, S924A) results in increased reversal frequency compounded with reduced forward locomotion and this defect can be rescued by introducing eat-4 mutations (n>10), (one-way ANOVA *p<0.05, **p<0.001, ***p<0.0001). In the box and whisker plots the central line is the median, the edges of the box are the 25th and 75th percentiles, and the whiskers extend to the most extreme data points. (E) Expression of the non-phosphorylatable variant of [Pnpr-9::glr-1(S907A, S924A)::gfp] results in increased GLR-1::GFP level in the AIB neuronal process and it is not further increased upon disruption of endocytosis in unc-11 mutants. At least 3 transgenic lines expressing non-phosphorylatable variant of GLR-1 (S907A, S924A) were separately examined for this experiment (n>15), Error bars represent ± SEM (one-way ANOVA ***p<0.0001).

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The following source data is available for figure 6:

Source data 1. Analysis of GLR-1 abundance in endocytosis defective mutants.
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modulates synaptic strength by regulating GLR-1 abundance in the postsynaptic puncta to bias the behavioural readout in response to starvation.

**AMPK modulates MGL-1 in the mRNA and protein levels in the AIY interneurons to modify distal exploratory behaviour in response to starvation**

The results of our epistasis analysis between *mgl-1* and *aak-2* are consistent with *mgl-1* acting downstream of *aak-2* to regulate the transition from local to distal exploration in parallel with *glr-1* in response to starvation. Therefore, to further explore the mechanism by which AMPK regulates MGL-1 activity in AIY, we determined the fluorescence intensity of a [*Pmgl-1::mgl-1::GFP*] translational fusion protein in both well-fed and starved animals. Starved animals all showed reduced MGL-1 levels, which was completely reversed in *aak-2* mutants (*Figure 7A,B, Figure 7—figure supplement 1A*). Moreover, expressing *aak-2* specifically within the AIY interneurons was sufficient to restore MGL-1 levels to those of starved WT animals suggesting that AMPK activity modulates MGL-1 abundance in the AIY in response to starvation.

To determine how AMPK affected the MGL-1 abundance in the AIY we first measured the mRNA levels in well-fed and starved animals using semi quantitative RT-PCR, while in parallel we also monitored glc-3 levels which are expressed in the AIY where it acts as a glutamate-gated chloride channel required for local search behaviour (*Chalasani et al., 2007*). Although we did not observe any change in the levels of glc-3 mRNA in well-fed or starved animals, we did observe an slight increase in *mgl-1* mRNA levels in well-fed and a more significant increase in starved *aak-2* mutants (*Figure 7—figure supplement 1B,C*) suggesting that AMPK regulates MGL-1 levels by either blocking its transcription or affecting the stability of *mgl-1* transcripts. The former possibility is less plausible given the short time delay that occurs between the response to food deprivation and the consequent behavioural changes. Since we did not detect any significant change in the *mgl-1* mRNA levels in WT animals, it is likely that AMPK controls the abundance of *mgl-1* transcripts in the AIY and possibly other *mgl-1*-expressing neurons in response to starvation, typical of its function in adapting to energy stress. These results however do not exclude the possible role of AMPK in regulating MGL-1 protein levels directly through phosphorylation, as has been described for some ion channels (*Lang and Föller, 2014*), although this would be in addition to its observed impact on *mgl-1* transcript abundance.

To further investigate this possibility, we mutated the conserved potential AMPK phosphorylation site in MGL-1 (*Figure 7—figure supplement 1D*) to examine if AMPK also regulates MGL-1 through direct phosphorylation. We noted that mutating serine 234 to alanine (S234A) resulted in a modest, but nevertheless significant reduction in forward locomotion (*Figure 7C*), although this mutation did not result in increased MGL-1 levels (*Figure 7—figure supplement 1E*). This suggests that AMPK regulates *mgl-1* activity by impinging on both mRNA and protein function. The AMPK-mediated phosphorylation of serine 234 in MGL-1 may be required for the modulation of distal exploratory behaviour independent of its regulation of MGL-1 levels. We currently cannot exclude that this post-translational modification could affect some aspect of MGL-1 receptor function. Taken together, our results are consistent with a role for AMPK in modulating *mgl-1* function in the AIY interneurons whereby *mgl-1* activity is attenuated in response to starvation-induced AMPK signalling resulting in increased AIY activity and the consequent suppression of reversals.

Our results so far suggest that increased GLR-1 and MGL-1 abundance in the AIB and AIY interneurons, respectively, cause the defective distal exploration in starved *aak-2* animals. To further test whether indeed this may be the case we overexpressed GLR-1 and MGL-1 in the AIB and AIY interneurons, respectively, in WT animals to verify if this misexpression could recapitulate the defective distal exploration of starved *aak-2* mutants. The overexpression of MGL-1 in the AIY resulted in a phenotype that was quite similar to that seen in starved *aak-2* mutants whereas overexpression of GLR-1 in the AIB resulted in a more modest, but nevertheless significant, defect in transition between local to distal exploration. This difference can be explained by the fact that the overexpressed GLR-1 is most likely still targeted for endocytosis and degradation while AMPK is present and active, while in *aak-2* mutants there is a defect in GLR-1 endocytosis allowing it to accumulate to functionally critical levels in the puncta. Nevertheless, the overexpression of either of these transgenes results in a behavioural defect which phenocopies that of starved *aak-2* mutants (*Figure 8—figure supplement 1*).
Discussion

In most free-living organisms energy stress is often accompanied by characteristic compensatory behaviours that are triggered to increase the probability of encountering a food source (Wang et al., 2005; Gray et al., 2005; Dietrich et al., 2015). The spectrum of these responses however varies dramatically and naturally reflects both the environment and the physiology of the organism. Humans are characteristically hypersensitive and/or rapidly irritable during periods of acute food deprivation providing a familiar example of how a collection of neural circuits become reproducibly activated or blocked in response to this specific acute stress, culminating in highly

Figure 7. AMPK regulates the abundance of MGL-1 levels, while direct phosphorylation may affect MGL-1 function. (A,B) Starvation reduces [MGL-1::GFP] levels significantly in the AIY [Pttx-3::RFP] of WT animals and to a lesser extent in aak-2 mutants (AIY neuron is shown in the circle). Increased [MGL-1::GFP] levels in starved aak-2 mutants can be reversed by targeted expression of aak-2 in the AIY interneurons using [Pttx-3::aak-2] transgene (n>15), Error bars represent ± SEM (two-way ANOVA ***p<0.0001). Scale bars are 20 μm. (C) Mutating the consensus AMPK phosphorylation site at serine 234 to a non-phosphorylatable alanine in MGL-1 (S234A) resulted in a reduced forward locomotion rate that is not further reduced in aak-2; mgl-1 (S234A) double mutants (i) (n>10), (one-way ANOVA *p<0.05, **p<0.001). In the box and whisker plots the central line is the median, the edges of the box are the 25th and 75th percentiles, and the whiskers extend to the most extreme data points.

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The following source data and figure supplement are available for figure 7:

Source data 1. Analysis of MGL-1 abundance in WT and aak-2 mutants.
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Figure supplement 1. AMPK regulates MGL-1 by modulating its steady state mRNA levels while also affecting MGL-1 function through direct phosphorylation.
DOI: 10.7554/eLife.16349.030
predictable behavioural responses. These behavioural changes are achieved in part through synaptic plasticity, which is required for adaptation to varying circumstances in an experience-dependent manner (Bito, 2010; Collingridge et al., 2010; Liu et al., 2012). During our work we have taken advantage of a common physiological response to an acute environmental stress in order to delineate the genetic pathway, the corresponding neuronal circuitry, and the biochemical targets that drive adaptive exploratory behaviours in response to starvation in C. elegans.

AMPK function within the AWC-AIB-AIY circuitry is required for modification of distal exploratory behaviour in response to acute starvation

We and others found that the disruption of AMPK signalling resulted in marked changes in exploratory behaviour partly by reduced forward locomotion (Lee et al., 2008; Cunningham et al., 2014). However, we also noticed that starved aak-2 animals execute frequent reversals, which might contribute significantly to their defective distal exploration given the antagonistic relationship between the well-characterized forward and backward locomotory circuits. We were able to improve forward locomotion by the targeted expression of aak-2 in the AIB and AIY interneurons, while consequently reducing the abnormally frequent reversal behaviour typical of AMPK mutants, consistent with the involvement of AIB and AIY in regulating both reversal frequency and the duration of forward movement (TsaliK and Hobert, 2003; Wakabayashi et al., 2004). Notably, unlike aak-2 expression in the sensory neurons, its expression within the AIB and AIY interneurons also improved the chemotaxis defect of aak-2 mutants suggesting that AMPK may be critical for the integration of sensory signals received by AIB and AIY interneurons.

A recent study showed that aak-2 compromise mimics the effect of elevated serotonin on movement (Cunningham et al., 2014). The AIB and AIY interneurons synapse directly or indirectly onto the command interneurons to control whether C. elegans moves forward or backward (Gray et al., 2005; Chalasani et al., 2007; Piggott et al., 2011; Chen et al., 2013). Moreover, these neurons have been identified as potential targets of serotonergic signalling to mediate the effects of serotonin on locomotion (Harris et al., 2009). It is likely therefore that aak-2 acts within the AIB and AIY interneurons to modulate the effects of serotonergic signalling on the resulting movement rather than functioning directly within the serotonergic neurons per se.

AMPK acts as a molecular trigger in the AWC-AIB-AIY circuitry to modify their neuronal activity and the resulting behavioural outputs

The very short time delay that occurs between the response to the lack of food and the observed behaviours makes it unlikely that the effects of AMPK within the AIB and AIY interneurons are mediated by transcription. Alternatively, AMPK has been demonstrated to act directly upon ion channels and gap junctions (Hallows et al., 2000; Lang and Föller, 2014). Our calcium imaging data are consistent with a model whereby starvation-induced AMPK activation leads to the inhibition of neuronal activity within the AIB, while promoting activity in the AIY interneuron, thereby altering behavioural outputs in a coordinated manner.

How then can we account for the AMPK-dependent starvation-induced changes in calcium levels within the AIB and AIY interneurons and the effects on their respective neural activity? Neuronal activity results from the interplay between synaptic excitation and inhibition. In the brain, excitation is mainly achieved through glutamatergic transmission (Bito, 2010; Collingridge et al., 2010; Liu et al., 2012). Remarkably, our results delineate a novel role for AMPK in the modulation of neuronal activity in response to starvation through the simultaneous regulation of glutamatergic inputs mediated by two different receptors: GluR/GLR-1 in the AIB and MGL-1 in the AIY. Of note, unlike mgl-1 null mutations that resulted in the partial rescue of locomotory behaviour of aak-2 mutants, we noticed a complete rescue upon removal of eat-4 or glr-1. As eat-4 is the only identified vesicular glutamate transporter in C. elegans, inhibition of eat-4 is expected to result in a complete inactivation of both mgl-1- and glr-1-dependent outputs that are both potentially dependent on glutamate release. In the case of glr-1, AIY may require either direct or indirect synaptic inputs from an additional glr-1-expressing neuron(s), or alternatively may signal to a glr-1-expressing neuron(s) so that eliminating glr-1 not only results in reduced AIB activity, but also directly or indirectly blocks AIY synaptic inputs or outputs.
AMPK coordinates glutamatergic signals through distinct mechanisms

In addition to the plethora of processes that are regulated downstream of AMPK activation, AMPK has been shown to modulate synaptic aging (Samuel et al., 2014). In mice, starvation increases the activity of the AgRP neurons by affecting their firing rate, ultimately inducing intense food-seeking behaviour and increased feeding (Dietrich et al., 2015). The enhanced firing rate of the AgRP neurons has been shown to be largely dependent on glutamatergic inputs mediated by glutamatergic ionotropic AMPA and NMDA receptors (Bito, 2010; Liu et al., 2012). AMPK modulates this food intake behaviour through the regulation of glutamate release in both mice and in C. elegans, although the precise mechanism has not been elucidated in either case (Yang et al., 2011; Cunningham et al., 2012).

Our data demonstrate that AMPK (aak-2) regulates the GLR-1 and MGL-1 abundance postsynaptically. The AWC is sensitive to food availability and is inhibited during starvation, resulting in the inactivation of AIB and the activation of AY (Gray et al., 2005; Chalasani et al., 2007). Our data demonstrate that the loss of AMPK signalling results in an increase in AIB neuronal activity and a concomitant decrease in AY activity. This may be rationalized by the fact that C. elegans neurons release neurotransmitter in a graded fashion. Like vertebrate rod and cone photoreceptors which show tonic activity in the dark, and are hyperpolarized or depolarized by light, and removal of light respectively, AWC olfactory neurons have been shown to display basal activity in the absence of odour, but are inhibited or stimulated by odour or odour removal, respectively (Chalasani et al., 2007). Therefore, the increased GLR-1 and MGL-1 abundance in the AIB and AY interneurons could potentially result in responses of longer duration and increased strength upon tonic neurotransmitter release at rest, or upon activation of AWC consequently increasing reversal frequency and shortening runs (Figure 8).

In C. elegans an increase in glutamatergic inputs mediated by GLR-1 results in an enhanced reversal frequency making this simple behaviour a faithful determinant of increased synaptic strength (Zheng et al., 1999; Burbea et al., 2002; Chalasani et al., 2007). We provide several lines of evidence that demonstrate that AMPK regulates GLR-1 abundance, by affecting its ubiquitylation status during conditions of acute starvation. First, the total GLR-1 level is decreased upon starvation. Second, the amplitude and size/width of GLR-1 puncta that are two determinants of GLR-1 abundance are increased in the AIB neuronal process in the ventral nerve cord in aak-2 mutants. Third, this elevated GLR-1 accumulation cannot be further enhanced upon the disruption of the various processes required to remove GLR-1 from the membrane suggesting that AMPK and the ubiquitin-mediated internalization of GLR-1 may therefore function in a linear genetic pathway. Fourth, mutating the AMPK phosphorylation sites in GLR-1 results in increased GLR-1 abundance and defective distal exploratory behaviour. Finally, this defect is comparable to that observed in aak-2 mutants, while crossing the GLR-1 (S907A, S924A) variant into the aak-2 mutant background did not further exacerbate the distal exploratory behavioural defect. Therefore, consistent with its well-described role in the mouse where it regulates glutamatergic neurotransmission (Yang et al., 2011), AMPK is required to regulate the abundance of GLR-1 in specific glutamatergic neurons. In doing so it modulates glutamatergic inputs and synaptic strength to elicit appropriate behavioural outcomes in response to starvation.

AMPAR receptors are either inserted or removed from postsynaptic membranes in an activity-dependent manner leading to the potentiation or depression of synaptic transmission, respectively (Carroll et al., 1999). Our findings might be further extended to implicate AMPK in the control of synaptic plasticity that underlies learning and memory as the appropriate regulation of glutamate receptor abundance is critical for long-term memory through habituation in C. elegans (Rose et al., 2003). Any increase in GluR abundance, and hence signalling, that arises in AMPK-compromised animals is very likely to result in irregular plasticity and abnormal behavioural outcomes, consistent with what is observed in starved AMPK mutants.

In addition to its regulation of GLR-1 abundance in the AIB, we show that AMPK also regulates glutamatergic inputs by regulating MGL-1 in the AY. A key function of group II mGlu receptors is to modulate neuronal excitability and synaptic transmission at least partly by inhibiting adenylate cyclase (Dillon et al., 2006; Niswender and Conn, 2010). Group II mGlu receptors are required in both cognitive and emotional processes, and have been linked to various neuropsychiatric conditions, including anxiety, stress-related disorders, schizophrenia and substance misuse.
In mammals, activation of mGluR II induces long term depression at GABAergic synapses in the cochlear nucleus magnocellularis neurons where they have been shown to modulate synaptic plasticity (Lu, 2007).

In C. elegans, mgl-1 regulates a systemic starvation response and fat accumulation (Greer et al., 2008; Kang et al., 2009). Consistent with the inhibitory role of group II mGlu receptors in the vertebrate nervous system, mgl-1 was recently demonstrated to coordinate environmental sensory cues to modulate the activity of the pharyngeal neural network (Dillon et al., 2015). Although some studies have indicated that metabotropic glutamate receptors can be phosphoregulated by kinases such as PKC (Kim et al., 2005; Ko et al., 2012), our work has uncovered an AMPK-mediated modulation of the MGL-1 in response to acute food deprivation in C. elegans. This effect is at least partly achieved by influencing the regulation of mgl-1 mRNA levels in starved animals. Considering the brief delay between stimulus and response, it is most likely that AMPK mediates its effect on mgl-1 transcript levels through the phosphorylation of key regulators of mgl-1 mRNA stability as described previously (Yun et al., 2005). Therefore, in addition to its well-described role in the regulation of

**Figure 8.** AMPK coordinates neuronal activity of the AIB and AIY interneurons by regulating their glutamatergic synaptic inputs in response to acute starvation. The schematic showing the proposed role of AMPK in the AIB and AIY interneurons for regulation of appropriate transition between local to distal exploration during starvation conditions. Green and red colors show depolarized and hyperpolarized neurons, respectively. AMPK regulates GLR-1-mediated synaptic inputs into AIB interneurons by directly phosphorylating GLR-1 and subsequently modulating its endocytosis and potentially its degradation leading to decreased AIB activity in starved animals. AMPK also regulates the MGL-1-dependent synaptic inputs into the AIY interneurons, not only through its effects on mgl-1 mRNA levels but also by direct phosphorylation of MGL-1 protein. These functions of AMPK within the AIB and AIY interneurons collectively result in the transition between local exploration that occurs in well-fed animals to distal exploration that occurs in starved animals, thus allowing them to explore their environment more extensively for energy resources. Legends to figure supplements

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The following source data and figure supplement are available for figure 8:

**Source data 1.** glr-1 and mgl-1 over-expression data.

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**Figure supplement 1.** Overexpression of glr-1 and mgl-1 in the AIB and AIY, respectively results in the similar defect in distal locomotory behaviour as aak-2 mutants.

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(Niswender and Conn, 2010). In mammals, activation of mGluR II induces long term depression at GABAergic synapses in the cochlear nucleus magnocellularis neurons where they have been shown to modulate synaptic plasticity (Lu, 2007).
ionic channels by affecting their degradation, AMPK may regulate its downstream neuronal effectors by affecting mRNA stability to regulate protein levels.

In summary, our findings suggest a simple model, in which AMPK coordinately regulates the activity of the AIB and AIY interneurons by modulating their glutamatergic synaptic inputs through two distinct mechanisms. AMPK modulates glutamatergic inputs in both interneurons by impinging on AMPA-type glutamate receptor GLR-1 and metabotropic glutamate receptor MGL-1 which consequently regulate distinct downstream behavioural outputs. The most salient feature of our study is our demonstration that AMPK acts as a molecular switch in the nervous system that is most likely not limited to the circuitries engaged in the regulation of feeding and locomotory behaviours. Given the conserved AMPK phosphorylation sites in GLR-1 and MGL-1, we speculate that AMPK regulates the activity of neural circuits in various organisms at least partly by affecting these two receptor types to modulate a wide range of neuronal outputs that respond to physiological or developmental contexts involving various stresses. Alternatively, its starvation-inducible or pharmacological activation could provide a highly effective, non-invasive means of modifying behavioural outputs.

Materials and methods

Strains

*Caenorhabditis elegans* strains were maintained under standard conditions (*Brenner*, 1974). The Bristol isolate (N2) was used as wild type. The following alleles were used in this study: aak-1 (tm1944), aak-2(ok524), eat-4(ky5), glr-1 (n2461), lite-1(ec314), unc-11(e48), mgl-1(tm1811), osm-6 (p811), avr-14(ad1305), nmr-1(ak4), mgl-2(tm355), sid-1(pk3321). Please see Supplementary file 1 for the complete list of strains used in this study.

Plasmid constructs and transgenes

Genomic aak-2 and mgl-1 and their 2 kb and 3 kb upstream sequences were amplified and cloned upstream of GFP to generate [P*aak-2::aak-2::gfp*] and [P*mgl-1::mgl-1::gfp*]. unc-119, unc-54, glr-1, tph-1, rig-3, ttx-3, npr-9, che-2 promoters were amplified from genomic DNA and were cloned upstream of aak-2 cDNA (*Narbonne and Roy*, 2009) to drive aak-2 expression globally throughout the nervous system, body wall muscle and in different neuronal subtypes. GLR-1::GFP was amplified from *nuIs24* strain and was inserted downstream of npr-9 promoter to generate [P*npr-9::glr-1::gfp*]. The resulting constructs were injected at 10–50 ng/µl and either Punc-122::gfp, Pelt-2::gfp, Pmyo-2::gfp or Punc-122::dsred were used as the co-injection marker. To generate the GLR-1 variants including (S907A) and (S924A), PCR-mediated site-directed mutagenesis was performed using Gene-Tailor site-directed mutagenesis (Invitrogen, Carlsbad, CA) on [P*npr-9::glr-1::gfp*] and transgenic lines were generated after microinjecting plasmids at 50 ng/µl.

We used CRISPR/Cas9 to mutagenize the AMPK phosphorylation site serine 234 to alanine in mgl-1 as described previously (*Paix et al.*, 2014).

Quantitative analysis of exploratory behaviour

Well-fed adult animals were scored for exploratory behaviour following 2 hr starvation as described previously (*Gray et al.*, 2005; *Calhoun et al.*, 2015). For our time course experiment mid-L4 animals were scored for exploratory behaviour either immediately after removal from food or following longer durations off food. To minimize the behavioural modifications caused by variability in their environment, animals were grown on the similar size and amount of bacterial patches (*Calhoun et al.*, 2015). All body bends and reversals were scored by eye, by an investigator blind to the genotype of the animals.

L4 animals of sid-1(pk3321); uls69 [pCFJ90 (pmyo-2::mCherry) + unc-119p::sid-1] and aak-2 (ok524); sid-1(pk3321); uls69 [pCFJ90 (pmyo-2::mCherry) + unc-119p::sid-1] which are nervous system RNAi hypersensitive were transferred to the RNAi plates seeded with overnight cultures of HT115 *E. coli* clones expressing double stranded RNAi targeting glr-1, glr-2, eat-4, mod-1, ser-2, inx-7, unc-7, inx-19, gar-2, gcy-1, flip-1, mgl-1, npr-11. The exploratory behaviour of the resultant progeny was monitored upon reaching to adulthood.
**Locomotory behaviour in the presence of food**

The basal and enhanced slowing response was measured by transferring well-fed animals to plates with no bacteria, and transferring them to an assay plate with a thin layer of bacteria immediately or after 30 min off food as described previously (Sawin et al., 2000).

**Chemotactic analysis**

Chemotaxis assays towards a bacterial lawn or isoamyl alcohol (IAA) were performed on 10 cm nematode growth medium (NGM) plates incubated at room temperature (22°C) overnight as previously described. Animal behaviour was scored by eye, by an investigator blind to the genotype of the animal. 10 and 300 animals were used for each chemotaxis assay, either toward the bacterial lawn and IAA, respectively (Hart, 2006).

**Optogenetics**

*C. elegans* grown on NGM plates supplied with 5 μM all-trans retinal (ATR) were starved for 2 hr and their exploratory behaviour was monitored on retinal-free NGM plates. A 1 min pulse of blue (480 nm) or yellow light (540 nm) was delivered from an Arc lamp (EXFO) by a 10× objective (Zeiss M2Bio) to the head of an individual animal to activate ChR2 or Arch, respectively as described previously (Piggott et al., 2011; Kocabas et al., 2012). In order to eliminate intrinsic phototaxis responses, we used lite-1 mutants that are defective in UV light phototaxis. lite-1 mutants did not show any obvious defect in the locomotory behaviour in our assays. To depolarize the AY interneurons or hyperpolarize the AIB interneurons, we used previously described strains expressing channelrhodopsin-2 (ChR2) or archaerhodopsin-3 (Arch), respectively (Kocabas et al., 2012).

**In vivo imaging**

For the calcium imaging adult transgenic animals expressing [Pnpr-9::G-CaMP3+ Pnpr-9::DsRed2B] (Piggott et al., 2011) or [Ptx-3::G-CaMP1+ unc-122::GFP] (Chalasani et al., 2007) were transferred to a 10% agarose pad following 2 hr starvation. A coverslip was then applied and the calcium transients were monitored in the cell body or neuronal process of the AIB and AY interneurons, respectively as previously described (Lemieux et al., 2015). Briefly, to measure calcium transients in the AIB interneurons fluorescence images (green channel: G-CaMP3, red channel: DsRed2B) were acquired sequential over 120 s window at a frequency of 2 Hz through a 63X objective on a Zeiss microscope. To measure calcium transients in the AIB interneurons fluorescence images (Green channel) were acquired from the neuronal process in the nerve ring over a 65 s window at a frequency of 3 Hz through a 63X objective. The resulting time-point series obtained from the fluorescence images for the AIB and AY were then analyzed using ImageJ (RRID:SCR_001935).

To calculate the baseline values (F<sub>b</sub>) averages of global minima for 5–10 frames over the duration of each time-lapse sequence were obtained and the difference between a given each time point's fluorescence and F<sub>b</sub> (ΔF) was normalized by dividing ΔF to F<sub>b</sub> to plot ΔF/F<sub>b</sub> for each individual. In our analyses, we report the total integrated signal of all the observed peaks during the 65 s or 120 s recording window that consists of an unambiguously defined initiation of a spontaneous transient in the neural process or cell body for the AY and AIB, respectively. To measure the total integrated fluorescence the ΔF/F for each time point were summed over each time-lapse sequence to obtain a measurement for each time series sequence as described previously (Lemieux et al., 2015).

Calcium imaging in freely behaving animals was performed on a Zeiss microscope equipped with a Hamamatsu camera as previously described (Luo et al., 2014; Flavell et al., 2013). To prevent behavioural responses to blue light, the imaging was performed in the lite-1 background (Flavell et al., 2013). Animals were placed on 10% agarose pads to slow down their locomotion and the pad was sealed in a small chamber to prevent evaporation. The elapsed time from placing animals on the agarose pads to initiation of imaging was <5 min. Fluorescence time-lapse imaging was performed over 90 s at a frequency of 2 Hz using a 10x objective by an experimenter blind to the genotype. The images were processed as described before (Lemieux et al., 2015; Luo et al., 2014).

**Microscopy**

All images were acquired using a Zeiss AX10 microscope equipped with a Hamamatsu camera as previously described (Burbea et al., 2002). Starved GLR-1::GFP-expressing strains were immobilized
with 10 mM levamisole. Images were captured and processed using AxioVision software. Maximum intensity projections of Z series stacks were obtained. GLR-1 levels were calculated using imageJ and normalized relative to WT. Puncta widths were estimated as the peak width at half-maximal amplitudes as described previously (Burbea et al., 2002). At least three transgenic lines were examined for these experiments and all the imaging and subsequent quantification was performed by an experimenter who was blind to the genotype.

The extent of signal overlaps between GLR-1 and the presynaptic marker EAT-4 was indicated by imaging transgenic animals expressing GLR-1::GFP and EAT-4::mCherry. Maximum intensity projections of sequential images from fluorescent green and red filters were obtained. The local maximum in each punctum was used to determine the spatial position of the GFP and mCherry peaks. The percentage of GLR-1 puncta co-localizing with a EAT-4 punctum was assessed by the fraction of GLR-1 puncta peaks that were less than 1 μm (or the width of an average punctum) from a presynaptic peak as previously described (Burbea et al., 2002).

For quantification of MGL-1 abundance in the AIY, the fluorescence intensity was measured in at least 3 transgenic lines expressing [Pmgl-1::mgl-1::gfp] which were starved for two hours. The exposure time was adjusted in the well-fed wild-type background and images were acquired by an experimenter who was blind to the genotype. Pixel intensity was measured in ImageJ (NIH) by calculating the mean pixel intensity for the entire region of interest.

### Western blot

To compare GLR-1 levels, proteins were extracted from well-fed and starved WT and aak-2 mutants by sonication in lysis buffer (50 mM Hepes pH7.5, 150 mM NaCl, 10% glycerol, 1% Triton X-100, 1.5 mM MgCl2, 1 mM EDTA and protease inhibitors). Protein concentration was then determined by NanoDrop 2000c spectrophotometer (Thermo Scientific, Waltham, MA). A similar amount of protein for each condition was then subjected to 8% SDS-PAGE and then transferred to a nitrocellulose membrane (Bio-Rad, Canada) and blotted by anti-GFP antibody (RRID:AB_162553) as described previously (Xie and Roy, 2015).

### Semi-qRT-PCR

RNA extraction was performed in well-fed and starved animals as described previously (Lu and Roy. Similar quantities of RNA from each sample were used to generate comparable amplicon levels for each gene tested. PCR was performed using gene-specific primers for each query gene for 15 cycles with ProtoScript M-MuLV Taq RT-PCR Kit (NEB, E6400S).

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References
Aponte Y, Atasoy D, Sternson SM. 2011. AGRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. Nature Neuroscience 14:351–355. doi: 10.1038/nn.2739
Ashrafi K. 2006. Mapping out starvation responses. Cell Metabolism 3:235–236. doi: 10.1016/j.cmet.2006.03.002
Balthasar N, Dalgaard LT, Lee CE, Yu J, Funahashi H, Williams T, Ferreira M, Tang V, McGovern RA, Kenny CD, Christiansen LM, Edelstein E, Choi B, Boss O, Aschenasi C, Zhang CY, Mountjoy K, Kishi T, Elmquist JK, Lowell BB. 2005. Divergence of melanocortin pathways in the control of food intake and energy expenditure. Cell 123:493–505. doi: 10.1016/j.cell.2005.08.035
Belgardt BF, Okamura T, Brüning JC. 2009. Hormone and glucose signalling in POMC and AgRP neurons. Journal of Physiology 587:5305–5314. doi: 10.1113/jphysiol.2009.179192
Bewick GA, Gardiner JV, Dhillo WS, Kent AS, Webster Z, Ghatei MA, Bloom SR. 2005. Postembryonic ablation of AgRP neurons in mice leads to a lean, hypophagic phenotype. FASEB Journal 19:1680–1682. doi: 10.1096/fj.04-3434fje
Bito H. 2010. The chemical biology of synapses and neuronal circuits. Nature Chemical Biology 6:560–563. doi: 10.1038/nchembio.408
Brenner S. 1974. The genetics of Caenorhabditis elegans. Genetics 77:71–94.
Burbea M, Dreier L, Dittman JS, Grunwald ME, Kaplan JM. 2002. Ubiquitin and AP180 regulate the abundance of GLR-1 glutamate receptors at postsynaptic elements in C. elegans. Neuron 35:107–120. doi: 10.1016/S0896-6273(02)00749-3
Calhoun AJ, Tong A, Pokala N, Fitzpatrick JA, Sharpee TO, Chalasani SH. 2015. Neural mechanisms for evaluating environmental variability in Caenorhabditis elegans. Neuron 86:428–441. doi: 10.1016/j.neuron.2015.03.026
Carroll RC, Lissin DV, von Zastrow M, Nicoll RA, Malenka RC. 1999. Rapid redistribution of glutamate receptors contributes to long-term depression in hippocampal cultures. Nature Neuroscience 2:454–460. doi: 10.1038/8123
Chalasani SH, Chronis N, Tsunozaki M, Gray JM, Ramot D, Goodman MB, Bargmann CI. 2007. Dissecting a circuit for olfactory behaviour in Caenorhabditis elegans. Neuron 450:63–70. doi: 10.1038/nature06292
Chalfie M, Sulston JE, White JG, Southgate E, Thomson JN, Brenner S. 1985. The neural circuit for touch sensitivity in Caenorhabditis elegans. Journal of Neuroscience 5:956–964.
Chen Z, Hendrickx M, Cornils A, Maier W, Alcedo J, Zhang Y. 2013. Two insulin-like peptides antagonistically regulate aversive olfactory learning in C. elegans. Neuron 77:572–585. doi: 10.1016/j.neuron.2012.11.025
Collingridge GL, Peineau S, Howland JG, Wang YT. 2010. Long-term depression in the CNS. Nature Reviews Neurosciences 11:459–473. doi: 10.1038/nrn2867
Cone RD. 2005. Anatomy and regulation of the central melanocortin system. Nature Neuroscience 8:571–578. doi: 10.1038/nn1455
Cowley MA, Smart JL, Rubinstein M, Cerdán MG, Diana S, Horvath TL, Cone RD, Low MJ. 2001. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. Nature 411:480–484. doi: 10.1038/35078085
Cunningham KA, Bouagoun AD, Barros AG, Lin L, Malard L, Romano-Silva MA, Ashrafi K. 2014. Loss of a neural AMP-activated kinase mimics the effects of elevated serotonin on fat, movement, and hormonal secretions. PLoS Genetics 10:e1004394. doi: 10.1371/journal.pgen.1004394
Cunningham KA, Hua Z, Srinivasan S, Liu J, Lee BH, Edwards RH, Ashrafi K. 2012. AMP-activated kinase links serotonergic signaling to glutamate release for regulation of feeding behavior in C. elegans. Cell Metabolism 16:113–121. doi: 10.1016/j.cmet.2012.05.014
Dietrich MO, Horvath TL. 2011. Synaptic plasticity of feeding circuits: hormones and hysteresis. Cell 146:863–865. doi: 10.1016/j.cell.2011.08.031
Dietrich MO, Zimmer MR, Bober J, Horvath TL. 2015. Hypothalamic Agrp neurons drive stereotypic behaviors beyond feeding. Cell 160:1222–1232. doi: 10.1016/j.cell.2015.02.024
Ahmadi and Roy. eLife 2016;5:e16349. DOI: 10.7554/eLife.16349

Dillon J, Franks CJ, Murray C, Edwards RJ, Calahorro F, Ishihara T, Katsuiri I, Holden-Dye L, O’Connor V. 2015. Metabotropic glutamate receptors: Modulators of context-dependent feeding behaviour in C. elegans. The Journal of Biological Chemistry 290:15052–15065. doi: 10.1074/jbc.M114.606608

Dillon J, Hopper NA, Holden-Dye L, O’Connor V. 2006. Molecular characterization of the metabotropic glutamate receptor family in Caenorhabditis elegans. Biochemical Society Transactions 34:942–948. doi: 10.1042/BST03409442

Flavell SW, Pokala N, Macosko EZ, Albrecht DR, Larsch J, Bargmann CI. 2013. Serotonin and the neuropeptide PDF initiate and extend opposing behavioral states in C. elegans. Cell 154:1023–1035. doi: 10.1016/j.cell.2013.08.001

Gordus A, Pokala N, Levy S, Flavell SW, Bargmann CI. 2015. Feedback from network states generates variability in a probabilistic olfactory circuit. Cell 161:215–227. doi: 10.1016/j.cell.2015.02.018

Gray JM, Hill JJ, Bargmann CI. 2005. A circuit for navigation in Caenorhabditis elegans. PNAS 102:3184–3191. doi: 10.1073/pnas.0409009101

Gray JV, Petsko GA, Johnston GC, Ringe D, Singer RA, Werner-Washburne M. 2004. “Sleeping beauty”: quiescence in Saccharomyces cerevisiae. Microbiology and Molecular Biology Reviews 68:187–206. doi: 10.1128/MMBR.68.2.187-206.2004

Greer ER, Pérez CL, Van Gilst MR, Lee BH, Ashrafi K. 2008. Neuronal and molecular dissection of a C. elegans sensory circuit that regulates fat and feeding. Cell Metabolism 8:118–131. doi: 10.1016/j.cmet.2008.06.005

Hallows KR, Raghuram V, Kemp BE, Witters LA, Foskett JK. 2000. Inhibition of cystic fibrosis transmembrane conductance regulator by novel interaction with the metabolic sensor AMP-activated protein kinase. Journal of Clinical Investigation 106:1711–1721. doi: 10.1172/JCI9622

Hardie DG, Ashford ML. 2014. AMPK: regulating energy balance at the cellular and whole body levels. Physiology 29:99–107. doi: 10.1152/physiol.00050.2013

Hardie DG, Ross FA, Hawley SA. 2012. AMPK: a nutrient and energy sensor that maintains energy homeostasis. Nature Reviews Molecular Cell Biology 13:251–262. doi: 10.1038/nrm3311

Harden DE. 2008. AMPK: a key regulator of energy balance in the single cell and the whole organism. International Journal of Obesity 32:S7–S12. doi: 10.1038/ijo.2008.116

Harris GP, Hapiak VM, Wragg RT, Miller SB, Hughes LJ, Hobson RJ, Steven R, Bamber B, Komuniecki RW. 2009. Three distinct amine receptors operating at different levels within the locomotory circuit are each essential for the serotonergic modulation of chemosensation in Caenorhabditis elegans. Journal of Neuroscience 29:1446–1456. doi: 10.1523/JNEUROSCI.4585-08.2009

Hart A. 2006. Behavior. In: The C. elegans Research Community (Eds). WormBook. doi: 10.1895/wormbook.1.87.1

Hart AC, Sims S, Kaplan JM. 1995. Synaptic code for sensory modalities revealed by C. elegans GLR-1 glutamate receptor. Nature 378:82–85. doi: 10.1038/378082a0

Hendricks M. 2015. Neuroecology: tuning foraging strategies to environmental variability. Current Biology 25:R498–R500. doi: 10.1016/j.cub.2015.04.042

Juo P, Harbaugh T, Garriga G, Kaplan JM. 2007. CDK-5 regulates the abundance of GLR-1 glutamate receptors in the ventral cord of Caenorhabditis elegans. Molecular Biology of the Cell 18:3883–3893. doi: 10.1091/mbc.E06-09-0818

Kang C, Avery L. 2009. Systemic regulation of starvation response in Caenorhabditis elegans. Genes & Development 23:12–17. doi: 10.1101/gad.1723409

Kim CH, Braud S, Isaac JT, Roche KW. 2005. Protein kinase C phosphorylation of the metabotropic glutamate receptor mGluR5 on Serine 839 regulates Ca2+ oscillations. Journal of Biological Chemistry 280:25409–25415. doi: 10.1074/jbc.M502644200

Ko SJ, Isozaki K, Kim I, Lee JH, Cho HJ, Sohn SY, Oh SR, Park S, Kim DG, Kim CH, Roche KW. 2012. PKC phosphorylation regulates mGluR5 trafficking by enhancing binding of Siah-1A. Journal of Neuroscience 32:16391–16401. doi: 10.1523/JNEUROSCI.1964-12.2012

Kocabas A, Shen CH, Guo ZV, Ramanathan S. 2012. Controlling interneuron activity in Caenorhabditis elegans to evoke chemotactic behaviour. Nature 490:273–277. doi: 10.1038/nature11431

Kola B. 2008. Role of AMP-activated protein kinase in the control of appetite. Journal of Neuroendocrinology 20:942–951. doi: 10.1111/j.1365-2866.2008.01745.x

Lang F, Föller M. 2014. Regulation of ion channels and transporters by AMP-activated kinase (AMPK). Channels 8:20–28. doi: 10.1461/chana.27423

Lee H, Cho JS, Lambacher N, Lee J, Lee SJ, Lee TH, Gartner A, Koo HS. 2008. The Caenorhabditis elegans AMP-activated protein kinase AAK-2 is phosphorylated by LKB1 and is required for resistance to oxidative stress and for normal motility and foraging behavior. Journal of Biological Chemistry 283:14988–14993. doi: 10.1074/jbc.M709115200

Lee RY, Sawin ER, Chalfie M, Horvitz HR, Avery L. 1999. EAT-4, a homolog of a mammalian sodium-dependent inorganic phosphate cotransporter, is necessary for glutamatergic neurotransmission in caenorhabditis elegans. Journal of Neuroscience 19:159–167.

Lemieux GA, Cunningham KA, Lin L, Mayer F, Werb Z, Ashrafi K. 2015. Kyneunic acid is a nutritional cue that enables behavioral plasticity. Cell 160:119–131. doi: 10.1016/j.cell.2014.12.028

Lim CT, Kola B, Korbonits M. 2010. AMPK as a mediator of hormonal signalling. Journal of Molecular Endocrinology 44:87–97. doi: 10.1677/JME-09-0063
Liu T, Kong D, Shah BP, Ye C, Koda S, Saunders A, Ding JB, Yang Z, Sabatini BL, Lowell BB. 2012. Fasting activation of AgRP neurons requires NMDA receptors and involves spinogenesis and increased excitatory tone. *Neuron* **73**:511–522. doi: 10.1016/j.neuron.2011.11.027

Lu Y, Roy R. 2015. Centrosome/Cell cycle uncoupling and elimination in the endoreplicating intestinal cells of *C. elegans*. *PLoS One* **9**:e110958. doi: 10.1371/journal.pone.0110958

Lu Y. 2007. Endogenous mGluR activity suppresses GABAergic transmission in avian cochlear nucleus magnocellular neurons. *Journal of Neurophysiology* **97**:1018–1029. doi: 10.1152/jn.00883.2006

Luo L, Wen Q, Ren J, Hendricks M, Gershov M, Qin Y, Greenwood J, Soucy ER, Klein M, Smith-Parker HK, Calvo AC, Colón-Ramos DA, Samuel AD, Zhang Y. 2014. Dynamic encoding of perception, memory, and movement in a C. elegans chemotaxis circuit. *Neuron* **82**:1115–1128. doi: 10.1016/j.neuron.2014.05.010

López M, Varela L, Vázquez MJ, Rodríguez-Cuenca S, González CR, Velagapudi VR, Morgan DA, Schoenmakers E, Agassandian K, Lage R, Martinez de Morentin PB, Tovar S, Nogueiras R, Carling D, Lelliott C, Gallego R, Oresic M, Chatterjee K, Saha AK, Rahmouni K et al. 2010. Hypothalamic AMPK and fatty acid metabolism mediate thyroid regulation of energy balance. *Nature Medicine* **16**:1001–1008. doi: 10.1038/nm.2207

Man HY, Lin JW, Ju WH, Ahmadian G, Liu L, Becker LE, Sheng M, Wang YT. 2000. Regulation of AMPA receptor-mediated synaptic transmission by clathrin-dependent receptor internalization. *Neuron* **25**:649–662. doi: 10.1016/S0896-6273(00)81067-3

Marvin JS, Borghuis BG, Tian L, Cichon J, Harnett MT, Akerman J, Gordus A, Renninger SL, Chen TW, Bargmann CI, Orger MB, Schreiter ER, Demb JB, Gan WB, Hires SA, Looger LL. 2013. An optimized fluorescent probe for visualizing glutamate neurotransmission. *Annual Review of Pharmacology and Toxicology* **53**:295–322. doi: 10.1146/annurev.pharmtox.011008.145533

Martens Y, Alquier T, Furukawa N, Kim YB, Lee A, Xue B, Mu J, Foufelle F, Ferre P, Birnbaumer MJ, Stuck BJ, Kahn BB. 2004. AMP-kinase regulates food intake by responding to hormonal and nutrient signals in the hypothalamus. *Nature* **428**:569–574. doi: 10.1038/nature02440

Narbonne P, Roy R. 2006. Regulation of germline stem cell proliferation downstream of nutrient sensing. *Cell Division* **1**:29–38. doi: 10.1186/1742-1088-1-29

Narbonne P, Roy R. 2009. Caenorhabditis elegans dauer need LKB1/AMPK to ration lipid reserves and ensure long-term survival. *Nature* **457**:210–214. doi: 10.1038/nature07536

Niswender CM, Conn PJ. 2010. Metabotropic glutamate receptors: physiology, pharmacology, and disease. *Annual Review of Pharmacology and Toxicology* **50**:295–322. doi: 10.1146/annurev.pharmtox.011008.145533

Paix A, Wang Y, Smith HE, Lee CY, Calídas D, Lu T, Smith J, Schmidt H, Krause MW, Seydoux G. 2014. Scalable and versatile genome editing using linear DNAs with microhomology to Cas9 Sites in Caenorhabditis elegans. *Genetics* **198**:1347–1356. doi: 10.1534/genetics.114.170423

Piggott BJ, Liu J, Feng Z, Wescott SA, Xu ZX. 2011. The neural circuits and synaptic mechanisms underlying motor initiation in C. elegans. *Cell* **147**:922–933. doi: 10.1016/j.cell.2011.08.053

Reche KW, O’Brien RJ, Mammen AL, Bernhardt J, Huganir RL. 1996. Characterization of multiple phosphorylation sites on the AMPA receptor GluR1 subunit. *Neuron* **16**:1179–1188. doi: 10.1016/S0896-6273(00)80144-0

Rose JK, Kaun KR, Chen SH, Rankin CH. 2003. GLR-1, a non-NMDA glutamate receptor homolog, is critical for long-term memory in Caenorhabditis elegans. *Journal of Neuroscience* **23**:9595–9599.

Samuel MA, Voinescu PE, Lilley BN, de Cabo R, Forretz M, Viollet B, Pawlyk B, Sandberg MA, Vavvas DG, Sanes JR. 2014. LKB1 and AMPK regulate synaptic remodeling in old age. *Nature Neuroscience* **17**:1190–1197. doi: 10.1038/nn.3772

Sawin ER, Ranganathan R, Horvitz HR. 2000. C. elegans locomotory rate is modulated by the environment through a dopaminergic pathway and by experience through a serotonergic pathway. *Neuron* **26**:619–631. doi: 10.1016/S0896-6273(00)81199-X

Schneeberger M, Claret M. 2012. Recent insights into the role of hypothalamic AMPK signaling cascade upon metabolic control. *Frontiers in Neuroscience* **6**:185. doi: 10.3389/fnins.2012.00185

Sternson SM, Nicholas Betley J, Cao ZF. 2013. Neural circuits and motivational processes for hunger. *Current Opinion in Neurobiology* **23**:353–360. doi: 10.1016/j.conb.2013.04.006

Tsaklik EL, Hobert O. 2003. Functional mapping of neurons that control locomotory behavior in Caenorhabditis elegans. *Journal of Neurobiology* **56**:178–197. doi: 10.1002/neu.10245

Wakabayashi T, Kitagawa I, Shingai R. 2004. Neurons regulating the duration of forward locomotion in Caenorhabditis elegans. *Neuroscience Research* **50**:103–111. doi: 10.1016/j.neures.2004.06.005

Wang T, Hung CC, Randall DJ. 2005. The comparative physiology of food deprivation: from feast to famine. *Annual Review of Physiology* **68**:223–251. doi: 10.1146/annurev.physiol.68.040104.105739

White JG, Southgate E, Thomson JN, Brenner S. 1976. The structure of the ventral nerve cord of Caenorhabditis elegans. *Philosophical Transactions of the Royal Society B: Biological Sciences* **275**:327–348. doi: 10.1098/rstb.1976.0086

White JG, Southgate E, Thomson JN, Brenner S. 1986. The structure of the nervous system of the nematode Caenorhabditis elegans. *Philosophical Transactions of the Royal Society B: Biological Sciences* **314**:1–340. doi: 10.1098/rstb.1986.0056

Wicks SR, Roehrig CJ, Rankin CH. 1996. A dynamic network simulation of the nematode tap withdrawal circuit: predictions concerning synaptic function using behavioral criteria. *Journal of Neuroscience* **16**:4017–4031.

Xie M, Roy R. 2015. The causative gene in chanarian dorfman syndrome regulates lipid droplet homeostasis in C. elegans. *PLoS Genetics* **11**:e1005284. doi: 10.1371/journal.pgen.1005284

Yang Y, Atasoy D, Su HH, Sternson SM. 2011. Hunger states switch a flip-flop memory circuit via a synaptic AMPK-dependent positive feedback loop. *Cell* **146**:992–1003. doi: 10.1016/j.cell.2011.07.039
Yun H, Lee M, Kim SS, Ha J. 2005. Glucose deprivation increases mRNA stability of vascular endothelial growth factor through activation of AMP-activated protein kinase in DU145 prostate carcinoma. *Journal of Biological Chemistry* **280**:9963–9972. doi: 10.1074/jbc.M412994200

Zheng Y, Brockie PJ, Mellem JE, Madsen DM, Maricq AV. 1999. Neuronal control of locomotion in C. elegans is modified by a dominant mutation in the GLR-1 ionotropic glutamate receptor. *Neuron* **24**:347–361. doi: 10.1016/S0896-6273(00)80849-1